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#### Electronic Supporting Information

#### Copper-catalyzed oxidative cleavage of Passerini and Ugi adducts in basic

#### medium yielding α-ketoamides

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#### 1. Optimization studies for oxidative cleavage of Ugi adducts

Table S1. Optimization of the reaction conditions<sup>*a*</sup>



Entry	Base	Additive (mol %)	Solvent	Time (h)	Conversion (%)	<b>2a;</b> yield (%) <sup>c</sup>	<b>4a</b> ; yield (%) <sup>c</sup>
1	NaH		THF	12	5	trace	trace
2	NaH	CuI (5)	THF	2	11	trace	trace
3	NaH	CuI (10)	THF	2	50	35	60
4	NaH	CuI (20)	THF	2	15	5	20
5	NaH	CuI (20)	CH <sub>3</sub> CN	2	20	15	38
6	NaH	CuI (20)	DMF	4	90	50	30
7	KO <sup>t</sup> Bu		THF	12	75	41	52
8	KO <sup>t</sup> Bu	CuI (5)	THF	2	11	trace	trace
9	KO <sup>t</sup> Bu	CuI (10)	THF	2	81	50	67
10	KO <sup>t</sup> Bu	CuI (20)	THF	2	83	65	50
11	KO <sup>t</sup> Bu	CuI (20)	CH <sub>3</sub> CN	0.5	100	81	53
12	KO <sup>t</sup> Bu	CuI (20)	DMSO	4	30	15	35
13	KO <sup>t</sup> Bu	CuI (20)	DMF	4	93	55	45
$14^{b}$	KO <sup>t</sup> Bu	CuI (20)	THF	0.5	5	trace	trace

<sup>*a*</sup>Reaction Conditions: **1a** (0.1 mmol), base (0.3 mmol), CuI in solvent (2.0 mL) at rt under oxygen balloon. <sup>*b*</sup>Under argon. <sup>*c*</sup>Isolated yields.

#### 2. Labeling Experiments



#### a) Preparation of 4-Methoxybenzaldehyde-a-D

4-Methoxybenzaldehyde- $\alpha$ -D was synthesized according to reported procedure.<sup>1</sup> 4-Methoxybenzaldehyde (100.0 mg, 0.734 mmol) and RuHCl(CO)(PPh<sub>3</sub>)<sub>3</sub> (34.9 mg, 0.036 mmol, 5 mol %) were dissolved in toluene (3.0 ml) in an oven-dried screw-cap vial. D<sub>2</sub>O (0.07 ml, 3.670 mmol) was then added and the vial was sparged with argon and capped. The resulting solution was heated to 100 °C and stirred for 30 minutes. On completion of the reaction, the solvent was removed *in vacuo* and crude was purified by column chromatography to afford **4-Methoxybenzaldehyde-\alpha-D** as colourless oil (80.0 mg, 79% yield, 55% D); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): 7.89 – 7.80 (m, 2H), 7.04 – 6.98 (m, 2H), 3.89 (s, 3H); residual formyl proton:  $\delta$  9.89.

#### b) Preparation of the deuterated Passerini adduct D-1d



Equimolar mixture of 4-Methoxybenzaldehyde-α-D (200.0 mg, 1.45 mmol), benzoic acid (194.5 mg, 1.59 mmol) and *tert*-butyl isocyanide (0.16 ml, 1.45 mmol) in water was stirred at room temperature for 12 h. After completion of the reaction (based on TLC), reaction mixture was diluted with ethyl acetate and washed with saturated sodium bicarbonate solution. Aqueous layer was extracted with ethyl acetate and the combined organic layers were dried over anhydrous sodium sulfate followed by evaporation of solvent *in vacuo*. The crude was

purified by silica gel column chromatography to afford the deuterium-labeled Passerini adduct **D-1d** as a white sticky solid (134.0 mg, 27% yield, 65% D); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  8.07 (dt, J = 8.5, 1.6 Hz, 2H), 7.63 – 7.56 (m, 1H), 7.50 – 7.41 (m, 4H), 6.95 – 6.87 (m, 2H), 5.96 (s, 1H), 3.80 (s, 3H), 1.37 (s, 9H); Residual formyl proton:  $\delta$  6.17.

#### c) Preparation of <sup>18</sup>O-labeled benzaldehyde



<sup>18</sup>O-labeled benzaldehyde was synthesized according to reported procedure.<sup>2</sup> Sodium (0.05 g, 2.17 mmol) was added to <sup>18</sup>O-labeled water (98% H<sub>2</sub><sup>18</sup>O, 0.75 mL) in a flask followed by the addition of benzyl chloride (0.5 mL, 4.34 mmol). The mixture was heated to 95 °C and then heated at reflux for 48 h with continuous stirring. The product was purified by column chromatography to yield the <sup>18</sup>O-labeled benzyl alcohol (0.2 g, 1.8 mmol, 83% yield). To a solution of the <sup>18</sup>O-labeled benzyl alcohol (0.2 g, 1.8 mmol) in anhydrous dichloromethane (20.0 mL) under nitrogen, Dess-Martin periodinane (0.99 g, 2.34 mmol) was added at 0 °C, and the resulting mixture was stirred at room temperature for 30 minutes. On completion of the reaction (progress monitored by TLC analysis), the reaction was quenched by the slow addition of NaHCO<sub>3</sub>/Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> solution, and the mixture was vigorously stirred for 30 minutes. The aqueous layer was extracted with dichloromethane and the combined organic layers were washed with brine and dried with anhydrous Na<sub>2</sub>SO<sub>4</sub>. The solvent was removed under reduced pressure to afford the <sup>18</sup>O-labeled benzaldehyde in quantitative yield.

#### d) Preparation of <sup>18</sup>O-labeled benzoic acid



<sup>18</sup>O-labeled benzoic acid was synthesized according to reported procedure.<sup>3</sup>  $\alpha, \alpha, \alpha$ trichlorotoluene (2.5 g, 12.5 mmol) and H<sub>2</sub><sup>18</sup>O (1.0 g, 50.0 mmol) were heated at 120 °C in a sealed tube for 24 h. The reaction mixture was concentrated *in vacuo* to remove excess water and HCl, then a solution of NaOH (0.15 M, 75 mL) was added to the crude mixture. The aqueous phase was washed with ethyl acetate, acidified with an aqueous HCl (1 N) solution and extracted with dichloromethane. The combined organic layers were dried upon anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered, concentrated *in vacuo* to afford <sup>18</sup>O-enriched benzoic acid (1.5 g, yield: 99%) as white solid.

#### e) Procedure for the synthesis of <sup>18</sup>O-labeled Passerini adduct <sup>18</sup>O-1b.



Equimolar mixture of <sup>18</sup>O-labeled benzaldehyde (200.00 mg, 1.85 mmol), <sup>18</sup>O-labeled benzoic acid (233.29 mg, 1.85 mmol) and *tert*-butyl isocyanide (0.21 mL, 1.85 mmol) in anhydrous dichloromethane was stirred at room temperature for 12 h under nitrogen. After completion of the reaction (based on TLC), reaction mixture was diluted with dichloromethane and washed with saturated sodium bicarbonate solution. Aqueous layer was extracted with dichloromethane and the combined organic layers were dried over anhydrous sodium sulfate followed by evaporation of solvent *in vacuo*. The crude was purified by silica

gel column chromatography to afford the <sup>18</sup>O-labeled Passerini adduct <sup>18</sup>O-1b (240.00 mg, 40.8%). Isotopic distribution amounted to 70.92%  $^{18}O/^{18}O/^{18}O$  and 29.08%  $^{18}O/^{18}O/^{16}O$  respectively (Figure S1).

**2-**(*tert*-butylamino)-2-(oxo-<sup>18</sup>*O*)-1-phenylethyl benzoate-<sup>18</sup>*O*<sub>2</sub> (<sup>18</sup>O-1b): White solid (240.0 mg, 41%); m.p : 140 °C;  $R_f = 0.32$  (30% EtOAc in hexane); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  8.12 – 8.06 (m, 2H), 7.64 – 7.57 (m, 1H), 7.55 – 7.44 (m, 4H), 7.43 – 7.34 (m, 3H), 6.22 (s, 1H), 5.99 (s, 1H), 1.37 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  167.3, 164.8, 135.9, 133.5, 129.7, 129.4, 128.9, 128.7, 128.6, 127.4, 76.0, 51.6, 28.7; HRMS (ESI): calcd. for  $C_{19}H_{22}N^{18}O_3$  [M+H]<sup>+</sup>: 318.1727, found: 318.1717.

#### Figure S1: HRMS Spectrum of <sup>18</sup>O-1b



#### f) Procedure for the oxidative cleavage of <sup>18</sup>O-1b.



To the solution of <sup>18</sup>O-1b (193.0 mg, 0.608 mmol) in dry THF was added KO'Bu (204.7 mg, 1.82 mmol) and CuI (23.1 mg, 20 mol %) at room temperature and the reaction vessel was flushed with O<sub>2</sub>. The resulting reaction mixture was stirred at room temperature. After completion of the reaction (based on TLC) in 10 minutes, the reaction mixture was quenched with water and the crude product was extracted with ethyl acetate. The organic layer was dried over anhydrous sodium sulfate, concentrated in vacuo and the crude was purified by silica gel column chromatography to afford  $\alpha$ -ketoamide <sup>18</sup>O-2b (34.0 mg, 27%). The aqueous layer was acidified with HCl solution up to pH 2-3, followed by extraction with dichloromethane. The organic layer was dried over anhydrous sodium sulfate, concentrated in vacuo to afford the <sup>18</sup>O-labeled benzoic acid <sup>18</sup>O-2b' (29.0 mg, 38%). HRMS analysis revealed that the  $\alpha$ -ketoamide <sup>18</sup>O-2b (having only C-1 oxygen labeled) has m/z = 208.1217, while the <sup>18</sup>O-labeled benzoic acid has m/z = 127.9792. This indicates that the oxygen at C-2 position of the  $\alpha$ -ketoamide <sup>18</sup>O-2b is coming from molecular oxygen. On the contrary, this experiment also confirmed the incorporation of only one <sup>16</sup>O oxygen atom in the  $\alpha$ -ketoamide <sup>18</sup>O-2b. For  $\alpha$ -ketoamide <sup>18</sup>O-2b, isotopic distribution amounted to 90.9% <sup>16</sup>O/<sup>18</sup>O and 9.09% <sup>16</sup>O/<sup>16</sup>O (Figure S2). For the acid <sup>18</sup>O-2b', isotopic distribution amounted to 51.02% <sup>18</sup>O/<sup>18</sup>O, 23.98% <sup>18</sup>O/<sup>16</sup>O and 25% <sup>16</sup>O/<sup>16</sup>O (Figure S3).



Figure S2: HRMS Spectrum of <sup>18</sup>O-2b

Figure S3: HRMS Spectrum of <sup>18</sup>O-2b'



#### 3. Kinetic Isotope Effect (KIE) measurement by independent reactions



The aerobic oxidative cleavage of substrate **1d** and **D-1d** (65% D-enriched) were carried out in parallel under the standard reaction conditions to study the kinetic isotopic effect (KIE). To the solutions of **1d** (100.0 mg, 0.29 mmol) and **D-1d** (100.0 mg, 0.29 mmol) in dry THF, KO'Bu (98.6 mg, 0.87 mmol) and CuI (11.1 mg, 0.058 mmol) were added at room temperature and the reaction vessel was flushed with O<sub>2</sub>. The resulting reaction mixtures were stirred at room temperature. KIE value was determined by comparison of rates of formation of  $\alpha$ -ketoamide by LCMS analysis. Aliquots (25 µL) were periodically removed to provide the following conversions as determined by LCMS analysis. K<sub>H</sub>/K<sub>D</sub> was calculated to be 3.14.

<i>t</i> (min)	1	3	5	10
1d (%)	13.67	14.09	15.79	18.93
<b>D-1d</b> <sup>#</sup> (%)	3.71	4.52	5.08	5.58

**Table S2.** % Formation vs time table.

<sup>#</sup>The product formation was calibrated by multiplying with 0.65 taking into the account that only 65% substrate was D-labeled.



Figure S4. Kinetic Isotope Effect

#### 4. References:

- E. S. Isbrandt, J. K. Vandavasi, W. Zhang, M. P. Jamshidi and S. G Newman, *Synlett*. 2017, 28, 2851.
- C. Du, X. Wang, S. Jin, H. Shi, Y. Li, Y. Pang, Y. Liu, M. Cheng, C. Guo and Y. Liu, Asian J. Org. Chem. 2016, 5, 755.
- C.-H. Lei, L. Zhao, D.-X. Wang, J. Zhu and M.-X Wang, Org. Chem. Front. 2014, 1, 909.

# 5. Copies of <sup>1</sup>H and <sup>13</sup>C NMR Spectra

Figure S5: <sup>1</sup>H NMR of compound 1a



# Figure S7: <sup>1</sup>H NMR of compound 1b



# Figure S9: <sup>1</sup>H NMR of compound 1c



# Figure S11: <sup>1</sup>H NMR of compound 1d



# Figure S13: <sup>1</sup>H NMR of compound 1e



# Figure S15: <sup>1</sup>H NMR of compound 1f



Figure S16: <sup>13</sup>C NMR of compound 1f



# Figure S17: <sup>1</sup>H NMR of compound 1g



# Figure S19: <sup>1</sup>H NMR of compound 1h



Figure S20: <sup>13</sup>C NMR of compound 1h



# Figure S21: <sup>1</sup>H NMR of compound 1i





Figure S23: <sup>1</sup>H NMR of compound 1j



# Figure S25: <sup>1</sup>H NMR of compound 1k



#### Figure S27: <sup>1</sup>H NMR of compound 11





#### Figure S29: <sup>1</sup>H NMR of compound 1m



#### Figure S31: <sup>1</sup>H NMR of compound 1n

88.88 88.15 88.15 88.15 88.15 88.15 88.15 88.15 88.15 77.55 77.75 77.75 77.55 77.75 77.55 77.75 77.55 77.75 77.55 77.75 77.55 77.75 77.55 77.75 77.55 77.75 77.55 77.75



#### Figure S32: <sup>13</sup>C NMR of compound 1n



# Figure S33: <sup>1</sup>H NMR of compound 2a



# Figure S35: <sup>1</sup>H NMR of compound 2b



# Figure S37: <sup>1</sup>H NMR of compound 2c



# Figure S39: <sup>1</sup>H NMR of compound 2d



# Figure S41: <sup>1</sup>H NMR of compound 2e







# Figure S43: <sup>1</sup>H NMR of compound 2f



# Figure S45: <sup>1</sup>H NMR of compound 2g



# Figure S47: <sup>1</sup>H NMR of compound 2h





# Figure S49: <sup>1</sup>H NMR of compound 2i



# Figure S51: <sup>1</sup>H NMR of compound 2j



# Figure S53: <sup>1</sup>H NMR of compound 2k



#### Figure S55: <sup>1</sup>H NMR of compound 2k'


Figure S57: <sup>1</sup>H NMR of compound 2l'



### Figure S59: <sup>1</sup>H NMR of compound 2m'



### Figure S61: <sup>1</sup>H NMR of compound 2n



### Figure S63: <sup>1</sup>H NMR of compound 20



# Figure S65: <sup>1</sup>H NMR of compound 2p



### Figure S67: <sup>1</sup>H NMR of compound 2q



### Figure S69: <sup>1</sup>H NMR of compound 2r



### Figure S71: <sup>1</sup>H NMR of compound 2s



### Figure S73: <sup>1</sup>H NMR of compound 2t



### Figure S75: <sup>1</sup>H NMR of compound 2u



### Figure S77: <sup>1</sup>H NMR of compound 2v



### Figure S79: <sup>1</sup>H NMR of compound 2w



#### Figure S81: <sup>1</sup>H NMR of compound 2x



### Figure S83: <sup>1</sup>H NMR of compound 2y



### Figure S85: <sup>1</sup>H NMR of compound 2z



### Figure S87: <sup>1</sup>H NMR of compound 2aa



### Figure S89: <sup>1</sup>H NMR of compound 2ab



### Figure S91: <sup>1</sup>H NMR of compound 2ac



### Figure S93: <sup>1</sup>H NMR of compound 2ad



### Figure S95: <sup>1</sup>H NMR of compound 2ae



### Figure S97: <sup>1</sup>H NMR of compound 3a



#### Figure S99: <sup>1</sup>H NMR of compound 3b



### Figure S101: <sup>1</sup>H NMR of compound 3c



### Figure S103: <sup>1</sup>H NMR of compound 3d



### Figure S105: <sup>1</sup>H NMR of compound 3e



### Figure S107: <sup>1</sup>H NMR of compound 3f



### Figure S109: <sup>1</sup>H NMR of compound 3g







### Figure S113: <sup>1</sup>H NMR of compound 3i



Figure S115: <sup>1</sup>H NMR of compound 3j



### Figure S117: <sup>1</sup>H NMR of compound 3k



#### Figure S119: <sup>1</sup>H NMR of compound 31





#### Figure S121: <sup>1</sup>H NMR of compound 3m



## Figure S123: <sup>1</sup>H NMR of compound 3n



## Figure S125: <sup>1</sup>H NMR of compound 30



### Figure S127: <sup>1</sup>H NMR of compound 3p


# Figure S129: <sup>1</sup>H NMR of compound 3q







### Figure S133: <sup>1</sup>H NMR of compound 3s



### Figure S135: <sup>1</sup>H NMR of compound 3t







### Figure S139: <sup>1</sup>H NMR of compound 4a



## Figure S141: <sup>1</sup>H NMR of compound 4b







### Figure S143: <sup>1</sup>H NMR of compound 4c



### Figure S145: <sup>1</sup>H NMR of compound 4d



### Figure S147: <sup>1</sup>H NMR of compound 5a







### Figure S149: <sup>1</sup>H NMR of compound 5b







### Figure S153: <sup>1</sup>H NMR of compound 6a



### Figure S155: <sup>1</sup>H NMR of compound 6b



### Figure S157: <sup>1</sup>H NMR of compound 6c



80 70 60 50

40 30 20 10 0 -10

130 120 110 100 90 f1 (ppm)

160 150

140

210

200

190 180

170

### Figure S159: <sup>1</sup>H NMR of compound 8a



### Figure S161: <sup>1</sup>H NMR of compound 9b



### Figure S163: <sup>1</sup>H NMR of compound 10



### Figure S165: <sup>1</sup>H NMR of compound 11a



### Figure S167: <sup>1</sup>H NMR of compound 11b







#### Figure S171: <sup>1</sup>H NMR of compound 11d



### Figure S173: <sup>1</sup>H NMR of compound 11e



#### Figure S175: <sup>1</sup>H NMR of compound 11f



#### Figure S177: <sup>1</sup>H NMR of 4-Methoxybenzaldehyde-α-D





