

Electronic Supplementary Information for:

**Enzymatically Triggered Chromogenic Cross-Linking Agents
Under Physiological Conditions**

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Table of Contents

Topic	Page
(1) X-ray single crystal structures	S2
(2) Time course of indigogenic reactions selected from Table 1	S4
(3) Indigogenic reactions examined in the presence of oxidants or with tritosomes	S5
(4) pH Profile of β -glucosidase from <i>Agrobacterium</i>	S9
(5) HPLC data from oligomerization studies	S10
(6) Mass spectrometry of the oligomer supernatant	S11
(7) Final concentrations of each ingredient in buffered solutions	S13
(8) Spectral data	S15

(1) X-ray single crystal structures

Summary of the crystal data for 1-acetyl-4-bromo-5-hydroxy-1*H*-indol-3-yl 2,3,4,6-tetra-*O*-acetyl- β -D-glucopyranoside (17)

CCDC registry	1902489
Formula	C ₂₆ H ₃₀ BrNO _{12.50}
Formula Weight (g/mol)	636.42
Crystal Dimensions (mm)	0.130×0.229×0.321
Crystal System	orthorhombic
Space Group	P 21 21 21
Temperature (K)	100(2)
<i>a</i> (Å)	13.3401(8)
<i>b</i> (Å)	14.3756(9)
<i>c</i> (Å)	28.9203(19)
α (°)	90
β (°)	90
γ (°)	90
<i>V</i> (Å ³)	5546.1(6)
Number of reflections to determine final unit cell	9951
Min and Max 2 θ for cell determination (°)	4.398, 51.23
<i>Z</i>	8
<i>F</i> (000)	2624
ρ (g/cm ³)	1.524
λ (Å, Mo K α)	0.71073
μ (mm ⁻¹)	1.550
Max 2 θ for data collection (°)	56.66
Measured fraction of data	0.999
Number of reflections measured	113542
Unique reflections measured	13811
<i>R</i> _{merge}	6.44%
Number of parameters in least-squares	788
<i>R</i> ₁	0.0324
w <i>R</i> ₂	0.0684
<i>R</i> ₁ (all data)	0.0423
w <i>R</i> ₂ (all data)	0.0712

Summary of the crystal data for 1-acetyl-4,6-dibromo-5-hydroxy-1*H*-indol-3-yl 2,3,4,6-tetra-*O*-acetyl- β -D-glucopyranoside (18)

CCDC registry	1902488
Formula	C _{25.50} H ₂₈ Br ₂ NO _{12.50}
Formula Weight (g/mol)	708.31
Crystal Dimensions (mm)	0.072×0.290×0.302
Crystal System	orthorhombic
Space Group	P 21 21 21
Temperature (K)	100
<i>a</i> (Å)	13.4601(5)
<i>b</i> (Å)	14.6763(5)
<i>c</i> (Å)	12.7603 (4)
α (°)	90
β (°)	90
γ (°)	90
<i>V</i> (Å ³)	5670.5(3)
Number of reflections to determine final unit cell	9915
Min and Max 2 θ for cell determination (°)	4.991, 62.78
<i>Z</i>	8
<i>F</i> (000)	2864
ρ (g/cm ³)	1.659
λ (Å, Mo K α)	0.71073
μ (mm ⁻¹)	2.926
Max 2 θ for data collection (°)	66.38
Measured fraction of data	0.999
Number of reflections measured	116273
Unique reflections measured	21707
<i>R</i> _{merge}	4.48%
Number of parameters in least-squares	757
<i>R</i> ₁	0.0297
w <i>R</i> ₂	0.0604
<i>R</i> ₁ (all data)	0.0380
w <i>R</i> ₂ (all data)	0.0627

(2) Time course of indigogenic reactions selected from Table 1

Figure S1 shows the time course of the indigogenic reaction of known indoxyl-glucoside **1** or **45** with β -glucosidase from almonds. The formation of indigoid products was monitored by absorption spectroscopy over the course of 16 h due to the slow reaction rate.

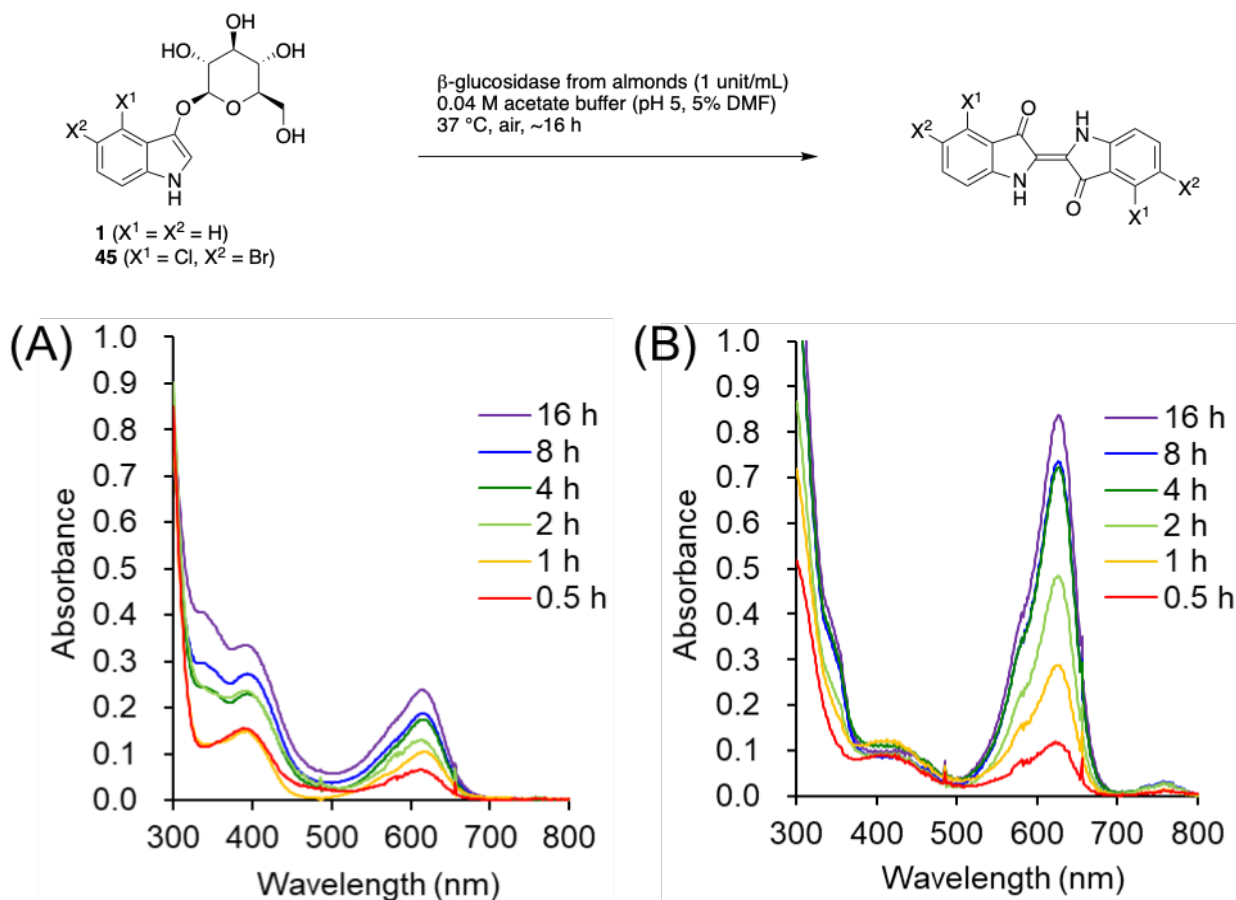


Figure S1. Time course of absorption spectra of indigogenic reactions with β -glucosidase from almonds (1 unit/mL) in 40 mM acetate buffer (pH 5.0, containing 5% DMF) at 37 °C. (A) The reaction of **1** (1 mM). After incubation, the reaction mixture was diluted four times with DMF and used for absorption spectroscopy. (B) The reaction of **45** (1 mM). After incubation, the reaction mixture was diluted ten times with DMF and used for absorption spectroscopy.

(3) Indigogenic reactions examined in the presence of oxidants or with tritosomes

To improve the yield and reaction rate of the indigogenic reaction, addition of oxidizing agents was attempted (Figure S2). However, $K_2Fe(CN)_6/K_3Fe(CN)_6$, peroxidase/ H_2O_2 , 2-(4-iodophenyl)-3-(4-nitrophenyl)-5-phenyl tetrazolium chloride (**S2**, Chart S1), or phenazine methosulfate (**S3**) did not increase the yield of the indigogenic reaction with **1** (Figure S2A). As shown in Figure S2B, the reaction rate of the indigogenic reaction with **45** was not increased by inclusion of $K_2Fe(CN)_6/K_3Fe(CN)_6$ or peroxidase/ H_2O_2 .

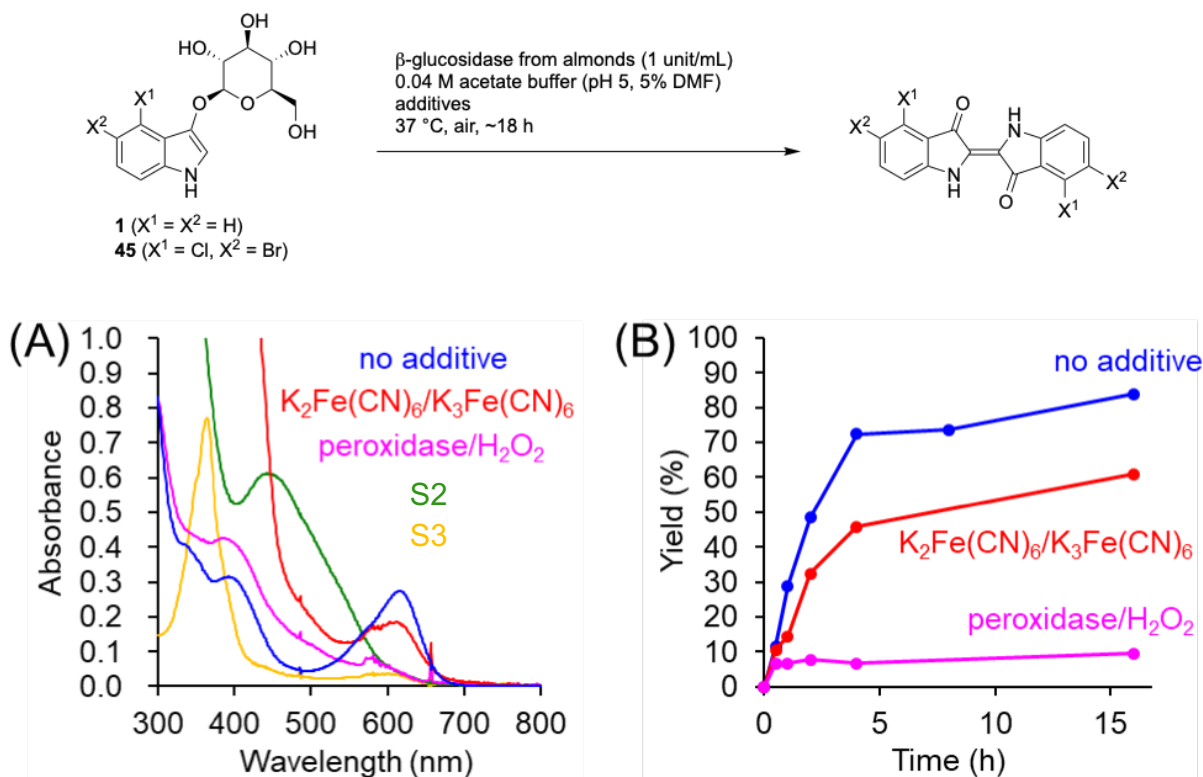


Figure S2. Indigogenic reactions in the presence of oxidizing agents. (A) Absorption spectra from the reaction with **1** (1 mM) without additives or with $K_2Fe(CN)_6$ (3 mM)/ $K_3Fe(CN)_6$ (3 mM), peroxidase (1 unit/mL)/ H_2O_2 (5 mM), **S2** (3 mM), or **S3** (3 mM). After incubation, the reaction mixture was diluted with DMF 4 times (for no additive; $K_2Fe(CN)_6/K_3Fe(CN)_6$; or peroxidase), 12 times (**S2**) or 20 times (**S3**) and used for absorption spectroscopy. (B) Time course of the reaction with **45** (1 mM) without additives, with $K_2Fe(CN)_6$ (3 mM)/ $K_3Fe(CN)_6$ (3 mM), and with peroxidase (1 unit/mL)/ H_2O_2 (5 mM). Yields were estimated from absorption spectroscopy with $\epsilon = 2.00 \times 10^4 \text{ M}^{-1}\text{cm}^{-1}$ reported for 5,5'-dibromo-4,4'-dichloroindigo.⁴⁰

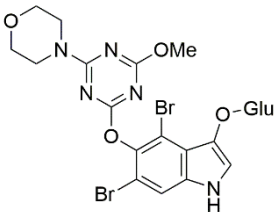
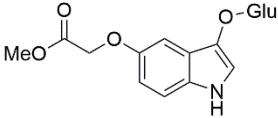
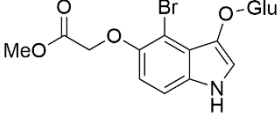
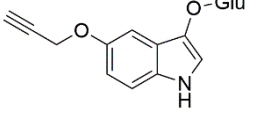
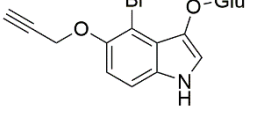
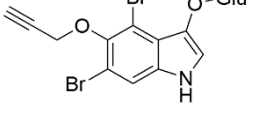
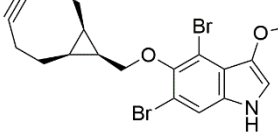
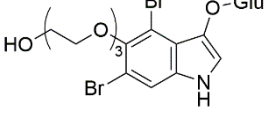


Chart S1. Structures of additives for indigogenic reactions.

A set of indigogenic reactions was examined where oxidants were included along with the β -glucosidase.

Table S1. Indigogenic reactions of indoxyl derivatives.

			Yield (%) ^a		
Entry	Indoxyl	Structure	β -glucosidase from almonds with S2 ^b	tritosomes ^c	tritosomes with catabolic buffer ^d
1	1		Poor	< 1	< 1
2	45		Good	< 1 ^e	8 ^e
3	15		Poor	< 1	< 1
4	16		Poor	8	< 5
5	19		Poor	15	11
6	20		Poor	< 1	< 1

7	21		Poor	< 1	< 1
8	22		Poor	< 1	< 1
9	23		Good	< 1	< 1
10	24		Fair	< 1	< 5
11	25		Poor	< 5	< 5
12	26		Good	< 1	< 1
13	27		Good	< 1	11 (6) ^g
14	30		Poor	< 1	< 1
15	33		Good ^g	— ^f	— ^f

^aThe yield was estimated by absorption spectroscopy with $\epsilon = 1.27 \times 10^4 \text{ M}^{-1}\text{cm}^{-1}$ (DMF/H₂O = 2:1) measured for the parent indigo **4** unless otherwise noted. A yield of <1% implies no color was observed by visual inspection and no absorption was observed spectroscopically. A yield of <5% implies a faint blue color was observed by visual inspection but the absorption was too weak for reliable spectroscopic determination. ^bA mixture of the indoxyl (1 mM), **S2** (3 mM), and β -glucosidase from almonds (1 unit/mL) in 40 mM acetate buffer (pH 5.0, containing 5% DMF) was incubated at 37 °C for 16–19 h. The yield of indigoid compounds was roughly evaluated by

absorption spectroscopy as products from **S2** have a shoulder absorption in the region 600–650 nm. ^cA mixture of the indoxyl (1 mM) and tritosomes (0.13 mg protein/mL) in 20 mM acetate buffer (pH 5.0, containing 5% DMF) was incubated at 37 °C for 16–19 h. ^dA mixture of the indoxyl (1 mM) and tritosomes (0.13 mg protein/mL) in 20 mM acetate buffer (pH 5.0, containing 5% DMF) in the presence of 1x catabolic buffer was incubated at 37 °C for 16–19 h. ^eThe yield was estimated from absorption spectroscopy with $\epsilon = 2.00 \times 10^4 \text{ M}^{-1}\text{cm}^{-1}$ reported for 5,5'-dibromo-4,4'-dichloroindigo.⁴⁰ ^fNot conducted. ^gThe yield was estimated from absorption spectroscopy with $\epsilon = 2.6 \times 10^4 \text{ M}^{-1}\text{cm}^{-1}$ (DMF/H₂O = 2:1) measured for **46**.

Figure S3 shows the effect of the nonionic surfactant tyloxapol (Triton WR1339) over a 1000-fold concentration range on the indigogenic reaction of **45** with β -glucosidase from almonds. The effect was measurable but rather small over the very large range of concentration examined.

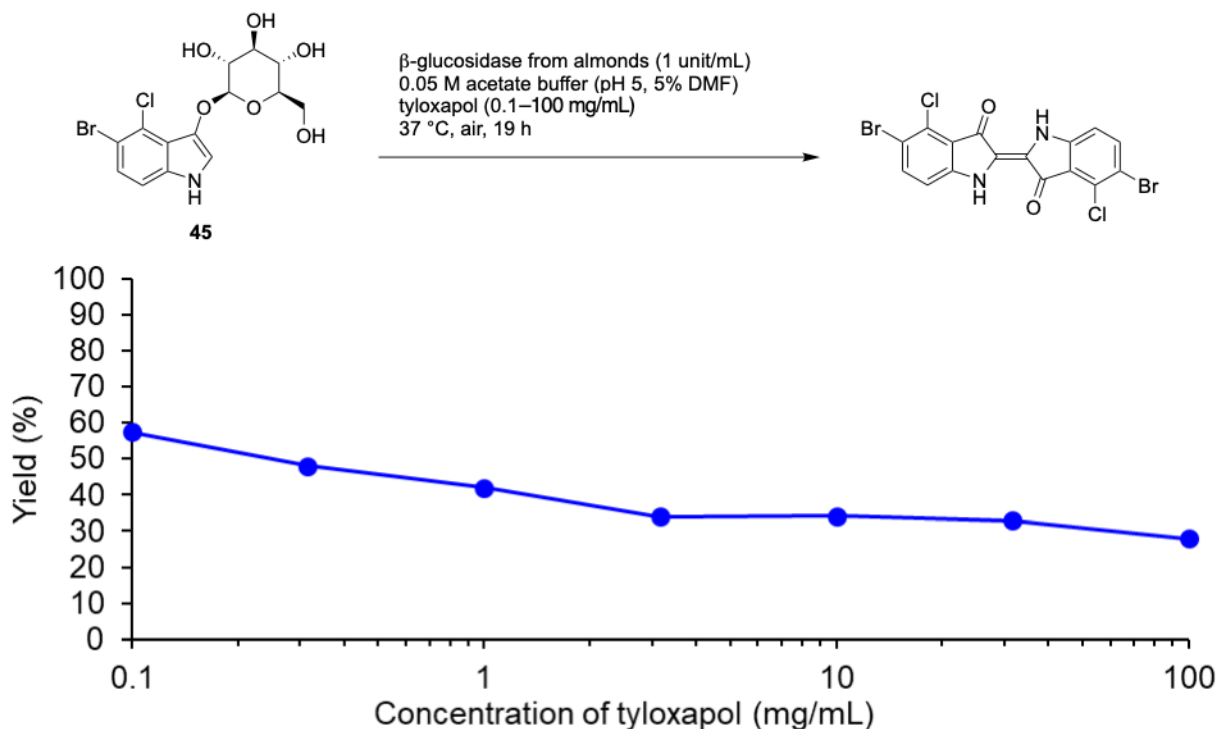


Figure S3. Effect of tyloxapol on the indigogenic reaction of **45** with β -glucosidase from almonds.

(4) pH Profile of β -glucosidase from *Agrobacterium*.

Following a reported procedure⁴³ with slight modification, a solution of 4-nitrophenyl β -D-glucopyranoside (2 mM) in 45 mM phosphate buffers (pH 4.0, 5.0, 6.0, 7.0, 8.0, or 9.0) was incubated at 37 °C for 10 min. A solution of β -glucosidase from *Agrobacterium* in NaPi-NaCl buffer (10 μ L, 1 μ M, pH 7.0, 10 mM NaPi and 50 mM NaCl) was added. The reaction mixture was incubated at 37 °C for 5 min and then quenched by the addition of 1.2 M aqueous NaOH (120, 100, 90, 60, 30, or 20 μ L for the reaction at pH 4.0, 5.0, 6.0, 7.0, 8.0, or 9.0, respectively) at room temperature. The solution was diluted two times with H₂O. The absorption at 400 nm was measured to calculate the activity of the enzyme (Figure S4).

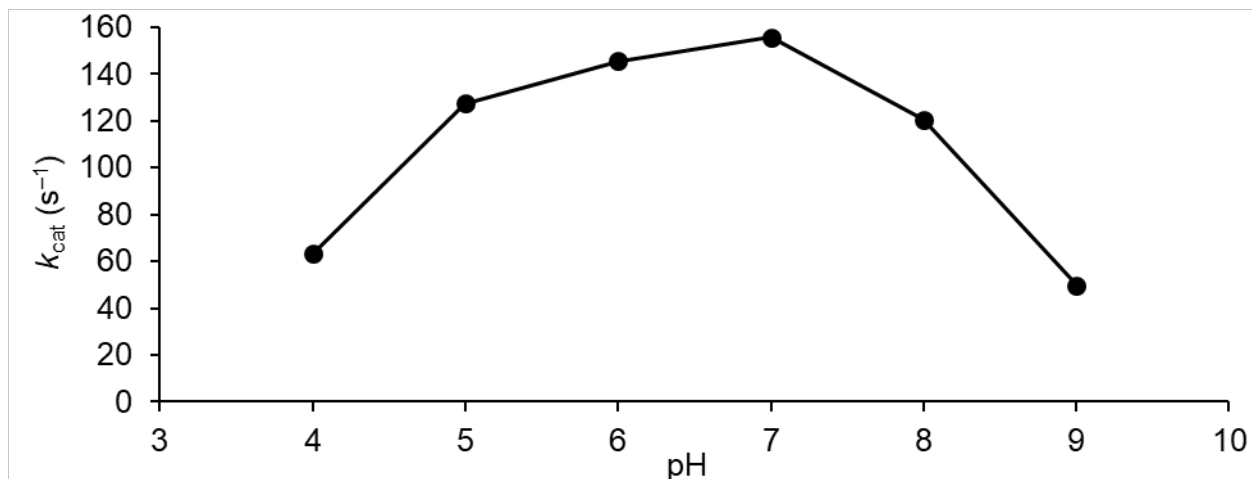


Figure S4. pH profile of β -glucosidase from *Agrobacterium*.

(5) HPLC data from oligomerization studies

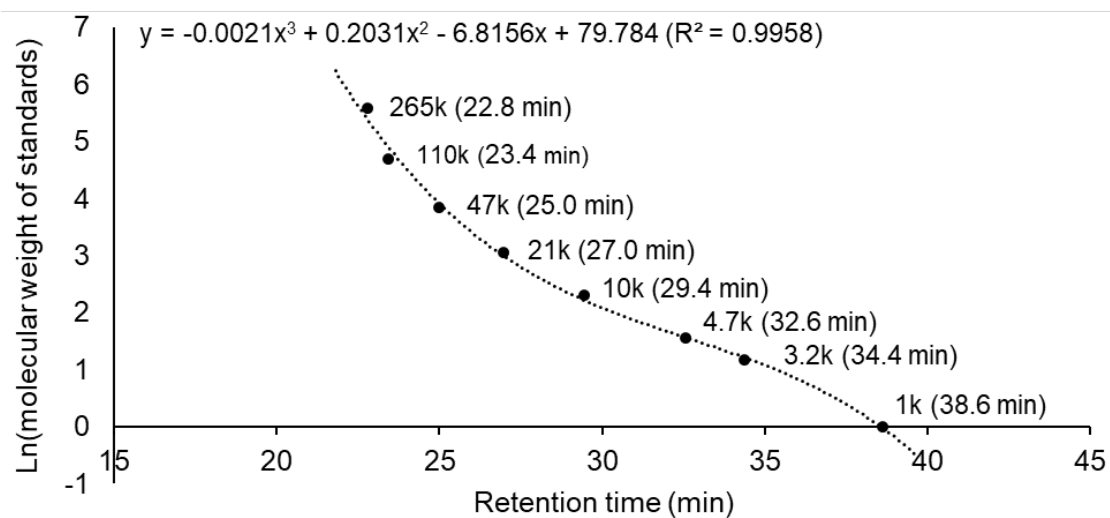


Figure S5. Calibration curve for SEC analysis.

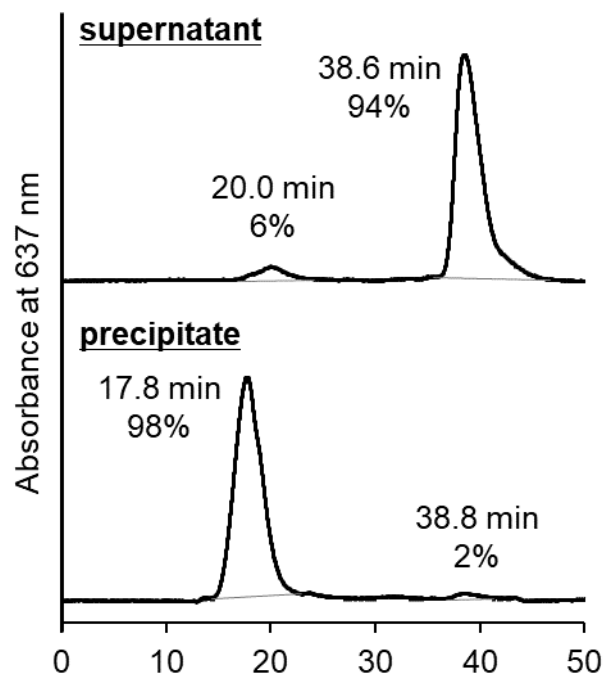


Figure S6. Analytical SEC traces for the supernatant and precipitate samples from 50 μ M of **49**.

(6) Mass spectrometry of the oligomer supernatant

ESI-MS analysis was carried out of the supernatant in the negative-ion mode using MeCN/H₂O (0.1% TFA). The spectrum is shown in Figure S7. Peaks consistent with the monomer and the dimer were detected. The putative monomer and dimer structures are shown in Figure S8.

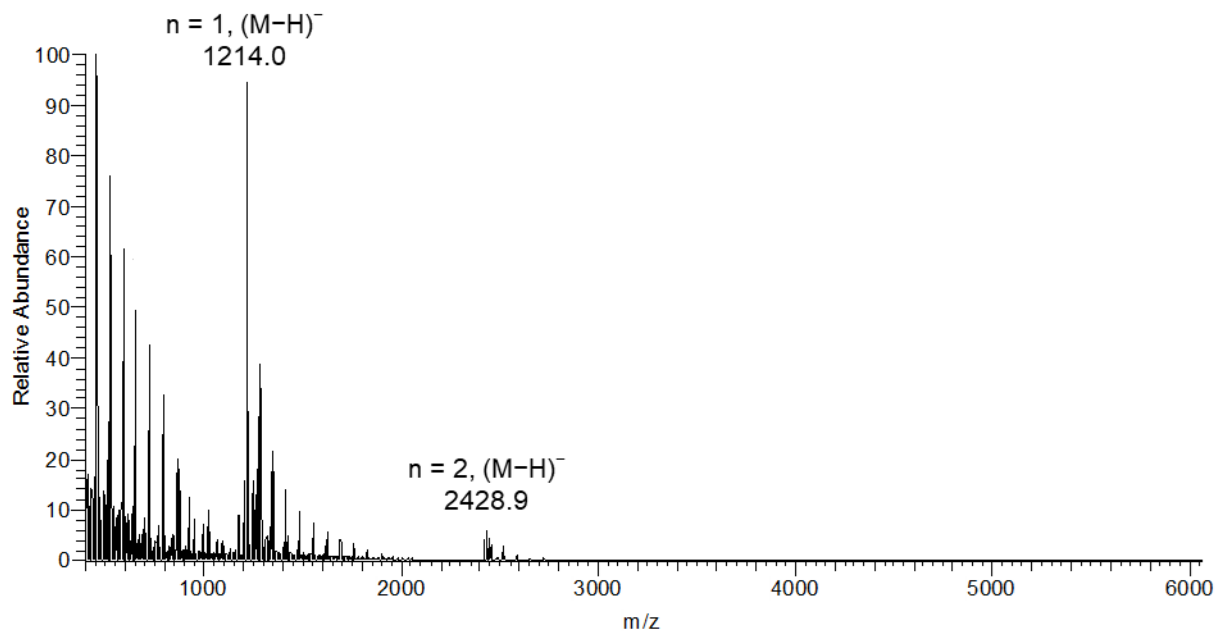


Figure S7. ESI-MS spectrum (negative-ion mode) of the supernatant derived from 300 μ M of **49**.

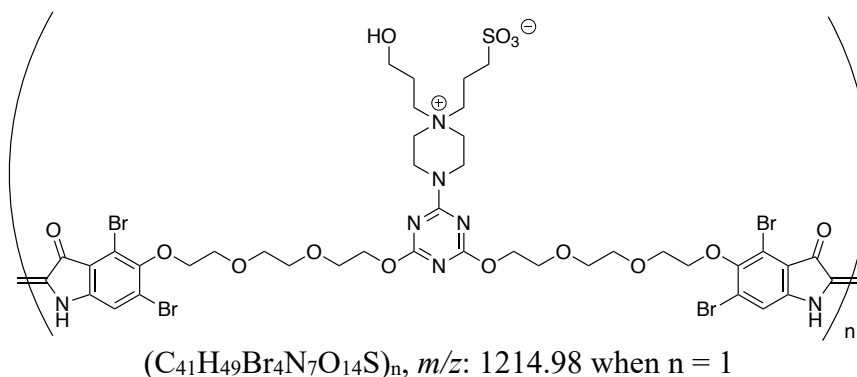


Figure S8. Cyclic monomer and dimer structures where n = 1 and 2, respectively.

MALDI-MS analysis (positive-ion mode) was carried out of the supernatant derived from 300 μ M of **49** using CHCA as a matrix and calibration with peptide standards. The spectrum is shown in Figure S9. The observed peak masses are compared with calculated masses in Table S2.

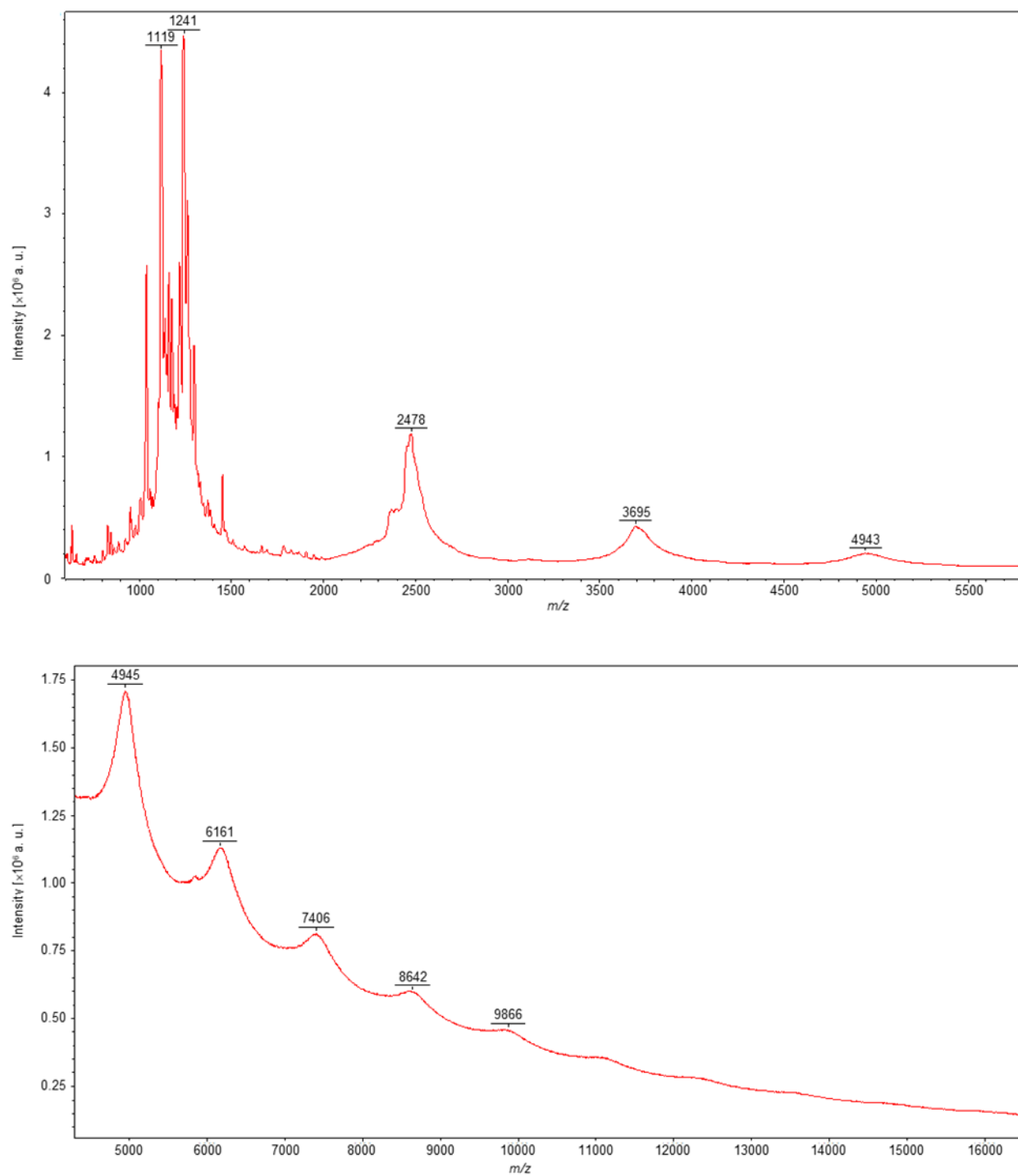


Figure S9. Peaks in the MALDI-MS spectrum of the supernatant derived from 300 μ M of **49**.

Table S2. Comparison of observed and calculated peak masses.

n	Calculated m/z for $[M + Na]^+$	Observed	Difference
1	1238	1241	3
2	2453	2478	25
3	3669	3695	26
4	4884	4943, 4945	59, 61
5	6098	6161	63
6	7315	7406	91
7	8528	8642	114
8	9746	9866	120

(7) Final concentrations of each ingredient in buffered solutions**Table 1****Reactions with β -glucosidase from almonds.**

Ingredient	Final concentration
Indoxyl compound	1 mM
β -Glucosidase from almonds	1 unit/mL
Sodium acetate buffer (pH 5.0)	42.5 mM
DMF	5%

Reactions with β -glucosidase from *Agrobacterium*.

Ingredient	Final concentration
Indoxyl compound	100 μ M
β -Glucosidase from <i>Agrobacterium</i>	200 nM
Sodium phosphate buffer (pH 7.0)	48.2 mM
DMF	2%
NaCl	1 mM
$(NH_4)_2SO_4$	12 mM

Reactions in rat liver homogenate.

Ingredient	Final concentration
Indoxyl compound	100 μ M
Rat liver homogenate	95%
DMF	5%

Reaction of 33 with β -glucosidase from *Agrobacterium* in rat liver homogenate.

Ingredient	Final concentration
33	100 μ M
β -Glucosidase from <i>Agrobacterium</i>	200 nM
Sodium phosphate buffer (pH 7.0)	200 μ M
Rat liver homogenate	96%
DMF	2%
NaCl	1 mM
$(NH_4)_2SO_4$	12 mM

Figure 6
Effect of pH (Figure 6A).

Ingredient	Final concentration
33	100 μ M
β -Glucosidase from <i>Agrobacterium</i>	200 nM
Sodium phosphate buffer (pH 4.0, 5.0, 6.0, 7.0, 8.0, or 9.0)	48.2 mM
DMF	2%
NaCl	1 mM
(NH ₄) ₂ SO ₄	12 mM

Effect of the enzyme concentration (Figure 6B).

Ingredient	Final concentration
33	100 μ M
β -Glucosidase from <i>Agrobacterium</i>	10 nM
Sodium phosphate buffer (pH 4.0, 5.0, 6.0, 7.0, 8.0, or 9.0)	48.6 mM
DMF	2%
NaCl	0.5 mM
(NH ₄) ₂ SO ₄	0.59 mM

Effect of the concentration of 33 (Figure 6C).

Ingredient	Final concentration
33	1.00, 1.78, 3.16, 5.62, 10.0, 17.8, 31.6, or 56.2 μ M)
β -Glucosidase from <i>Agrobacterium</i>	200 nM
Sodium phosphate buffer (pH 7.0)	9.8 mM
DMF	2%
NaCl	50 mM
(NH ₄) ₂ SO ₄	12 mM

Table 2**Oligomerization of 49 in NaPi-NaCl buffer.**

Ingredient	Final concentration
49	300, 100, 50 or 10 μ M
β -Glucosidase from <i>Agrobacterium</i>	200 nM
Sodium phosphate buffer (pH 7.0)	10 mM
NaCl	50 mM
(NH ₄) ₂ SO ₄	12 mM

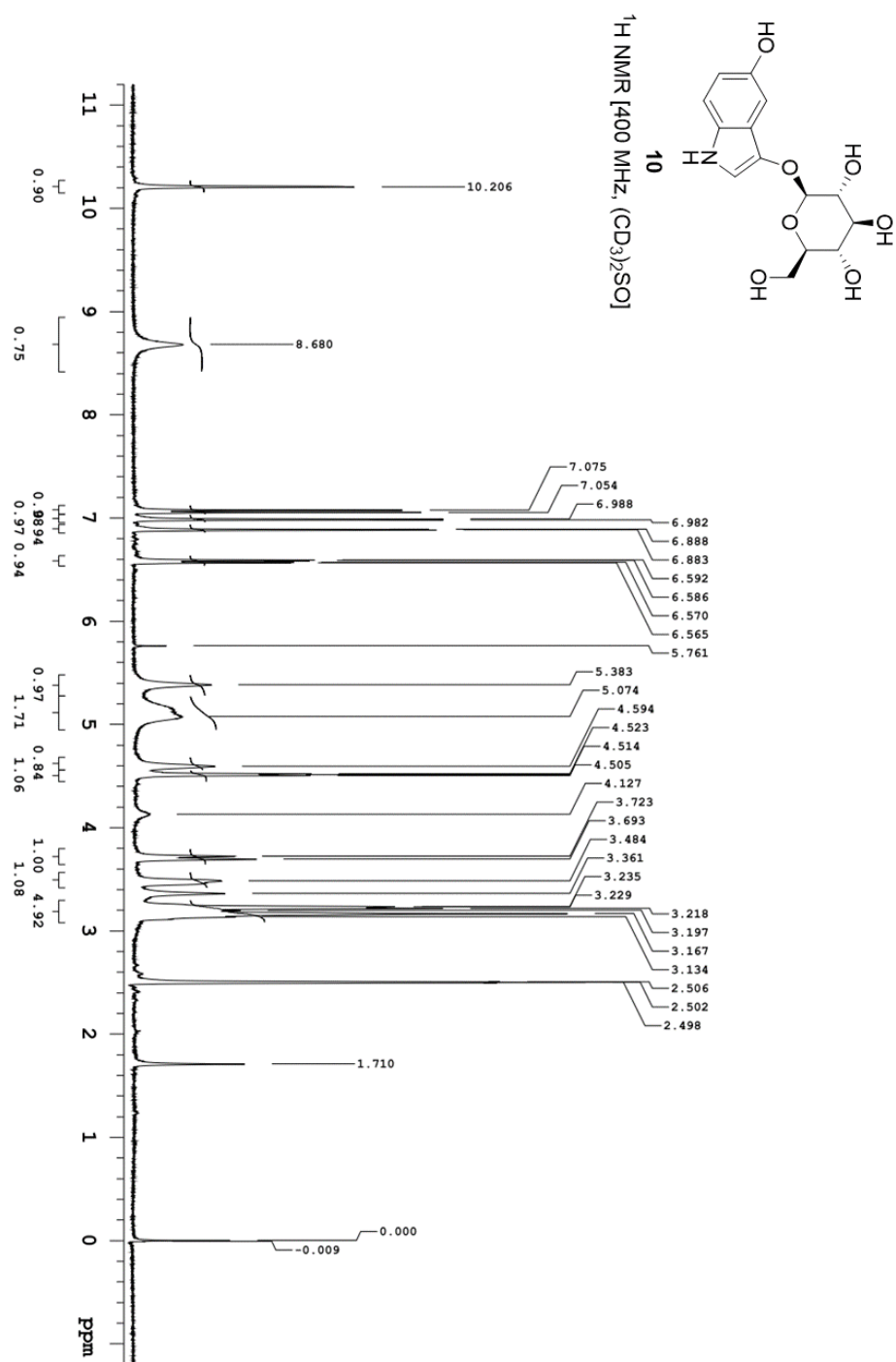
Low-salt oligomerization of 49.

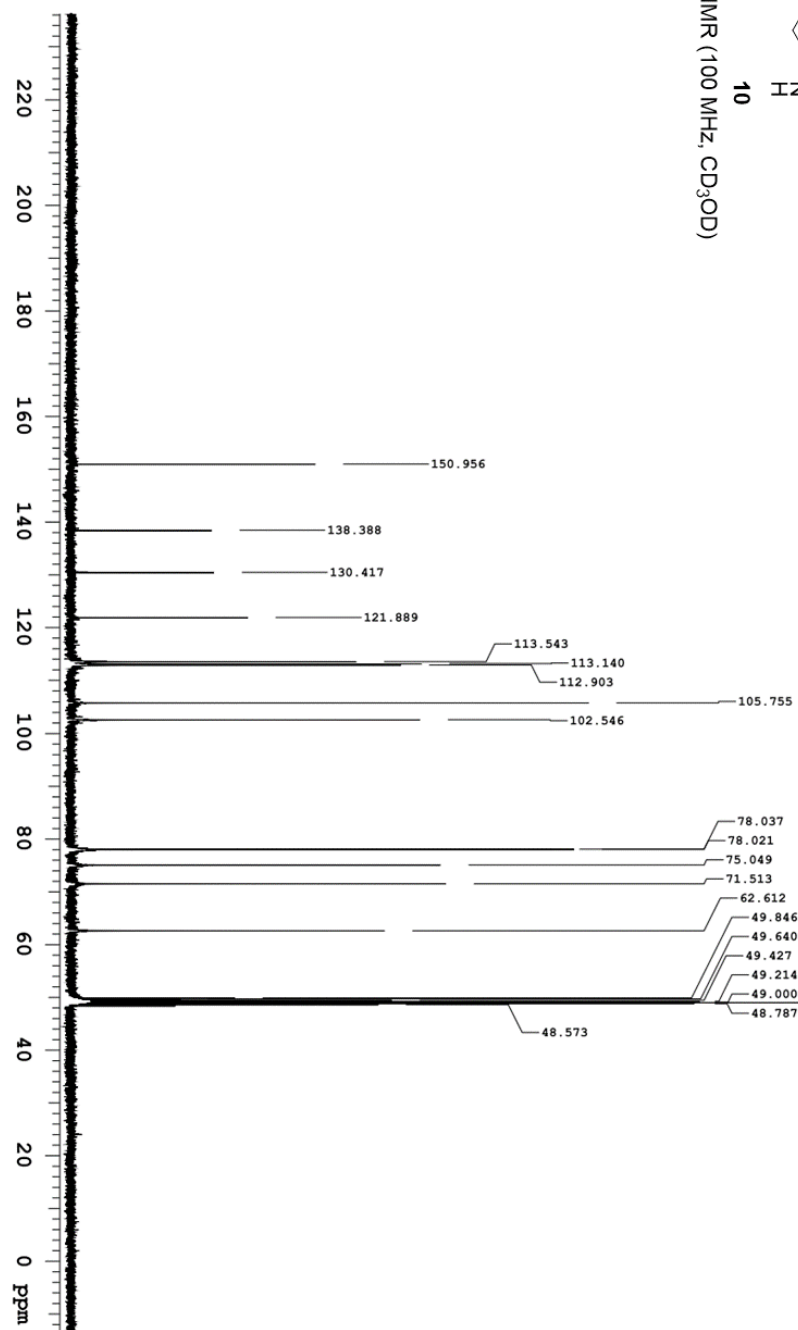
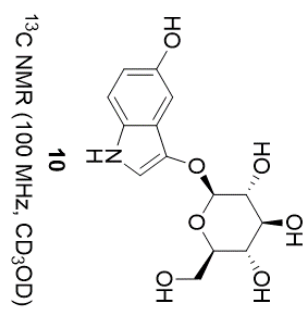
Ingredient	Final concentration
49	300 μ M
β -Glucosidase from <i>Agrobacterium</i>	200 nM
Sodium phosphate buffer (pH 7.0)	9.9 mM
DMF	0.6%
NaCl	1 mM
(NH ₄) ₂ SO ₄	12 mM

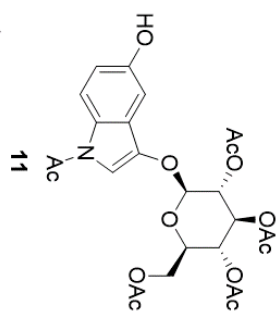
Figure S4**pH Profile of β -glucosidase from *Agrobacterium*.**

Ingredient	Final concentration
4-nitrophenyl β -D-glucopyranoside	2 mM
β -Glucosidase from <i>Agrobacterium</i>	5 nM
Sodium phosphate buffer (pH 4.0, 5.0, 6.0, 7.0, 8.0, or 9.0)	45 mM
NaCl	0.025 mM
(NH ₄) ₂ SO ₄	0.029 mM

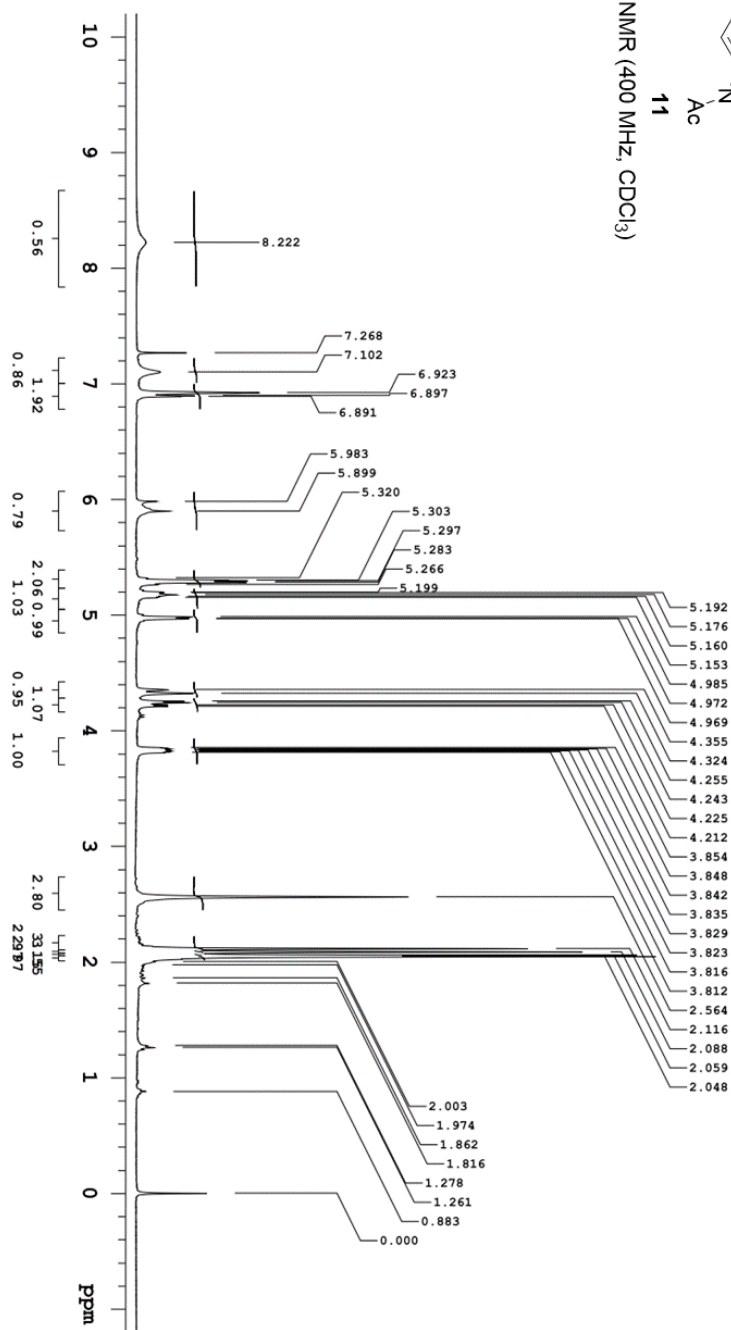
(8) Spectral data

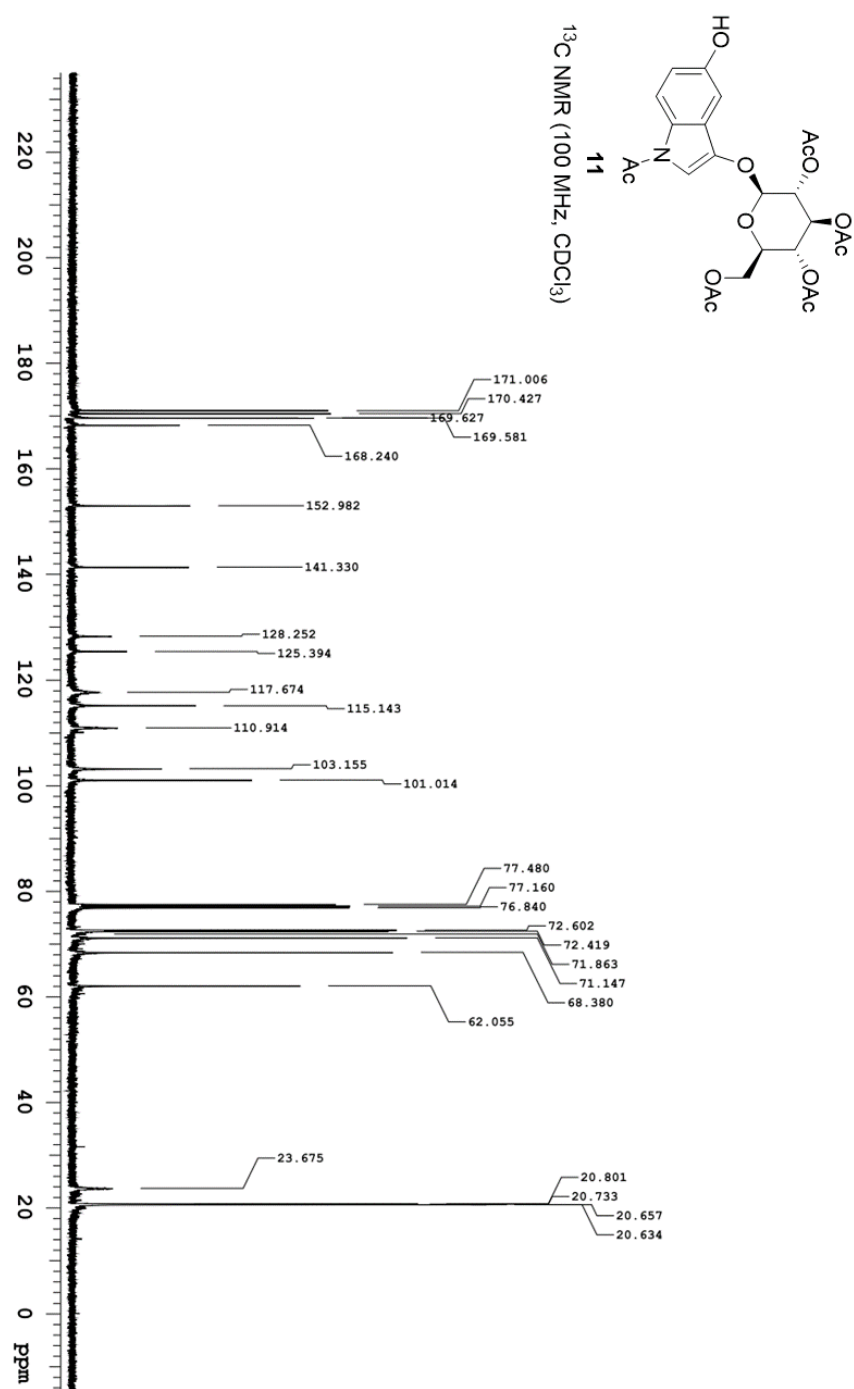


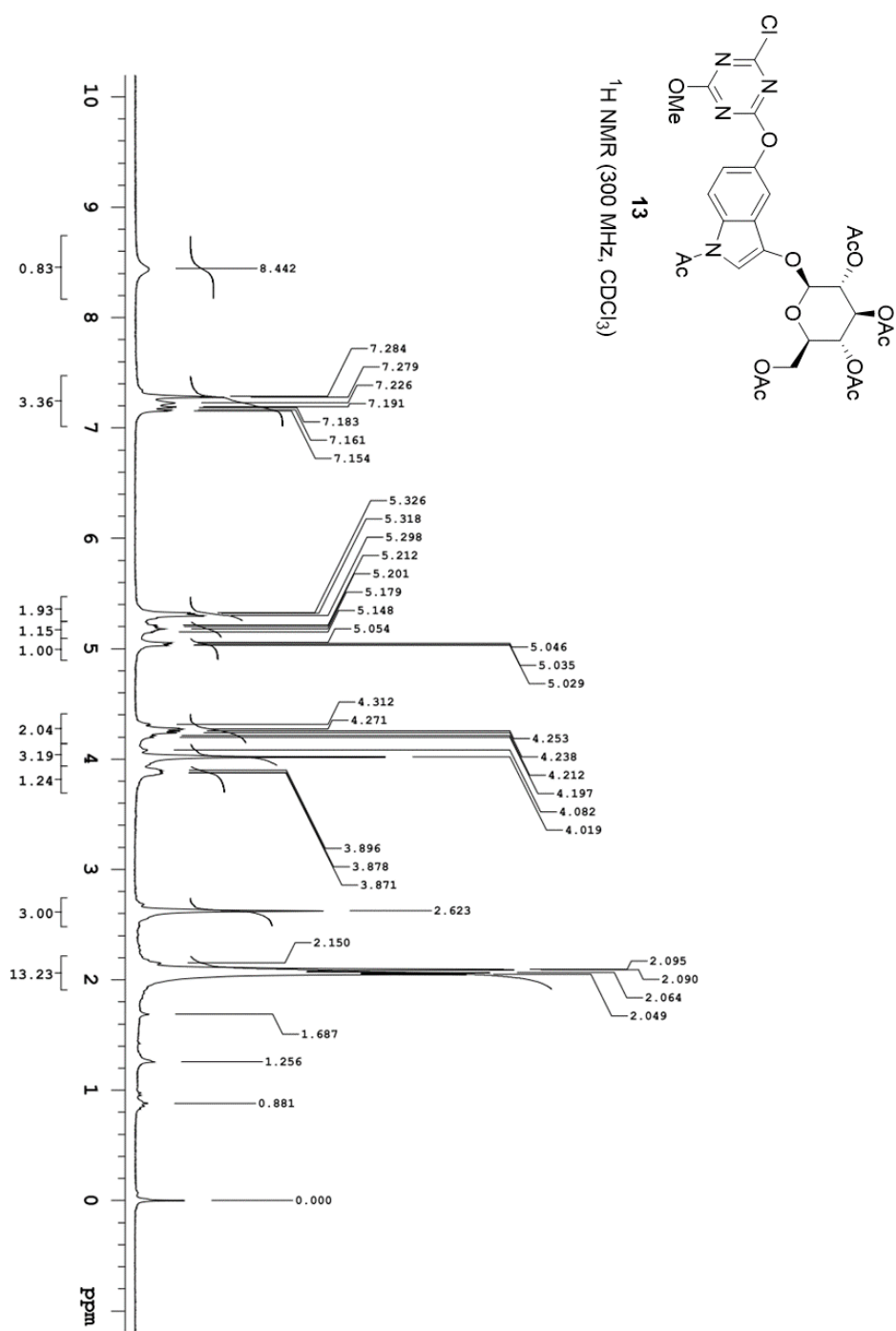


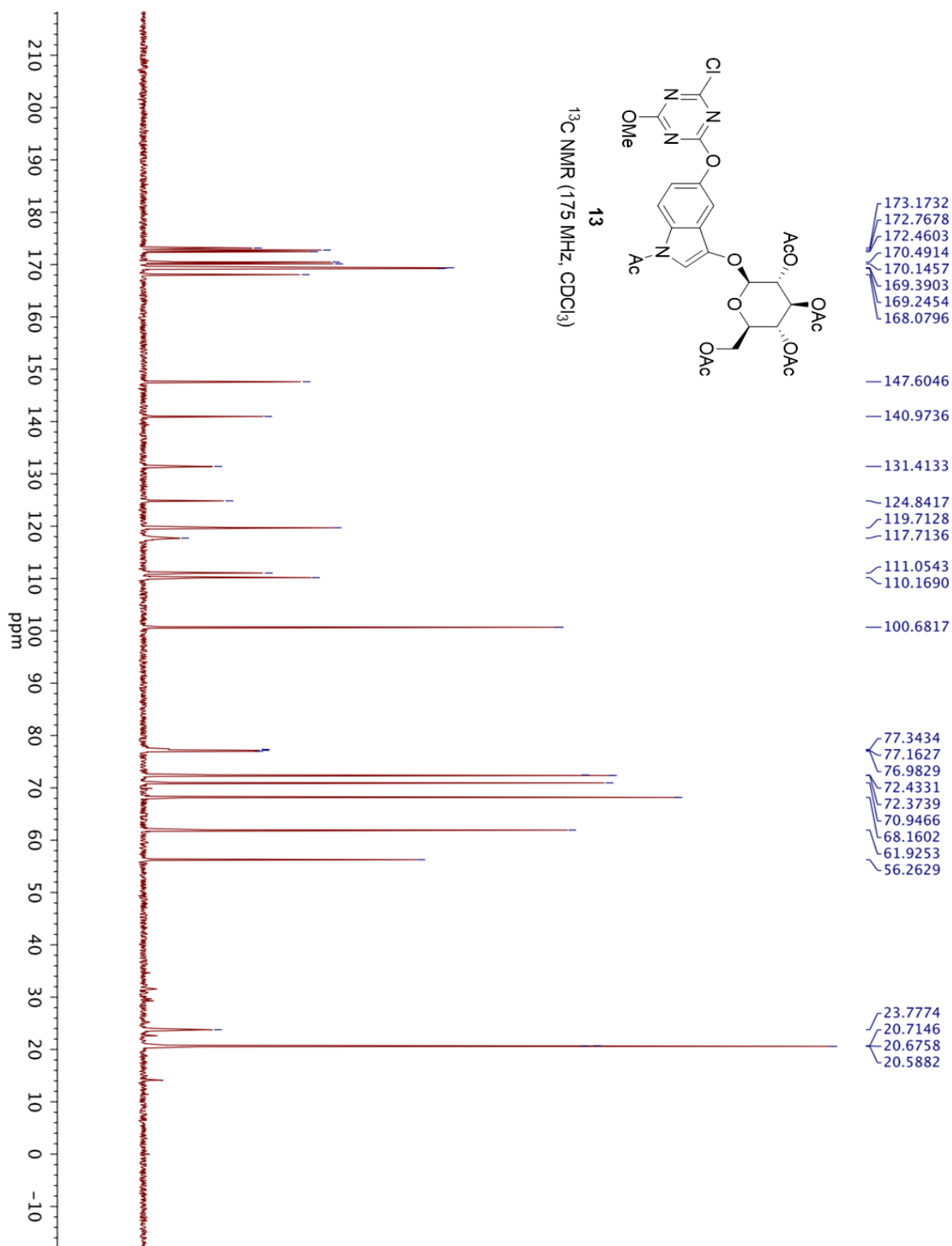


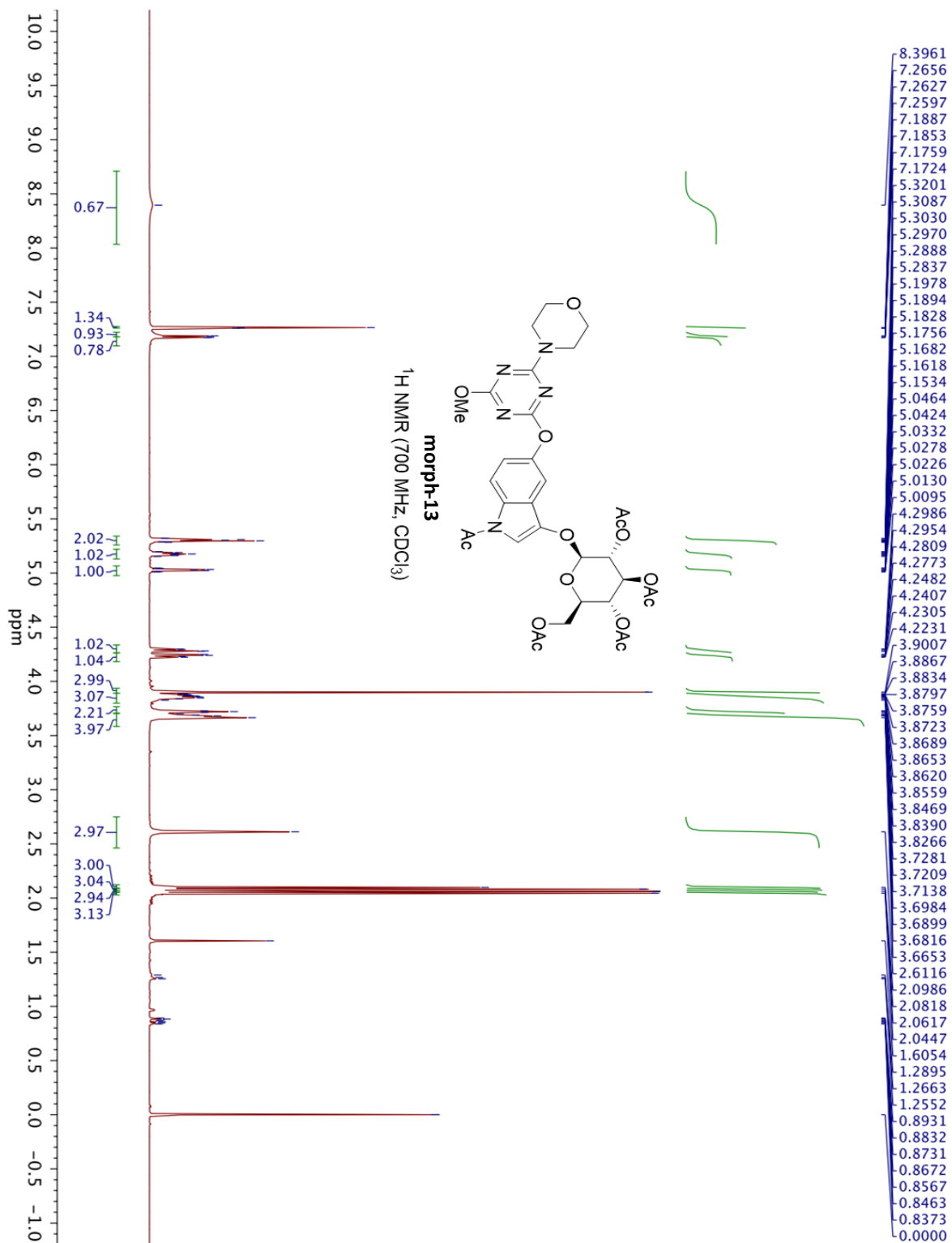
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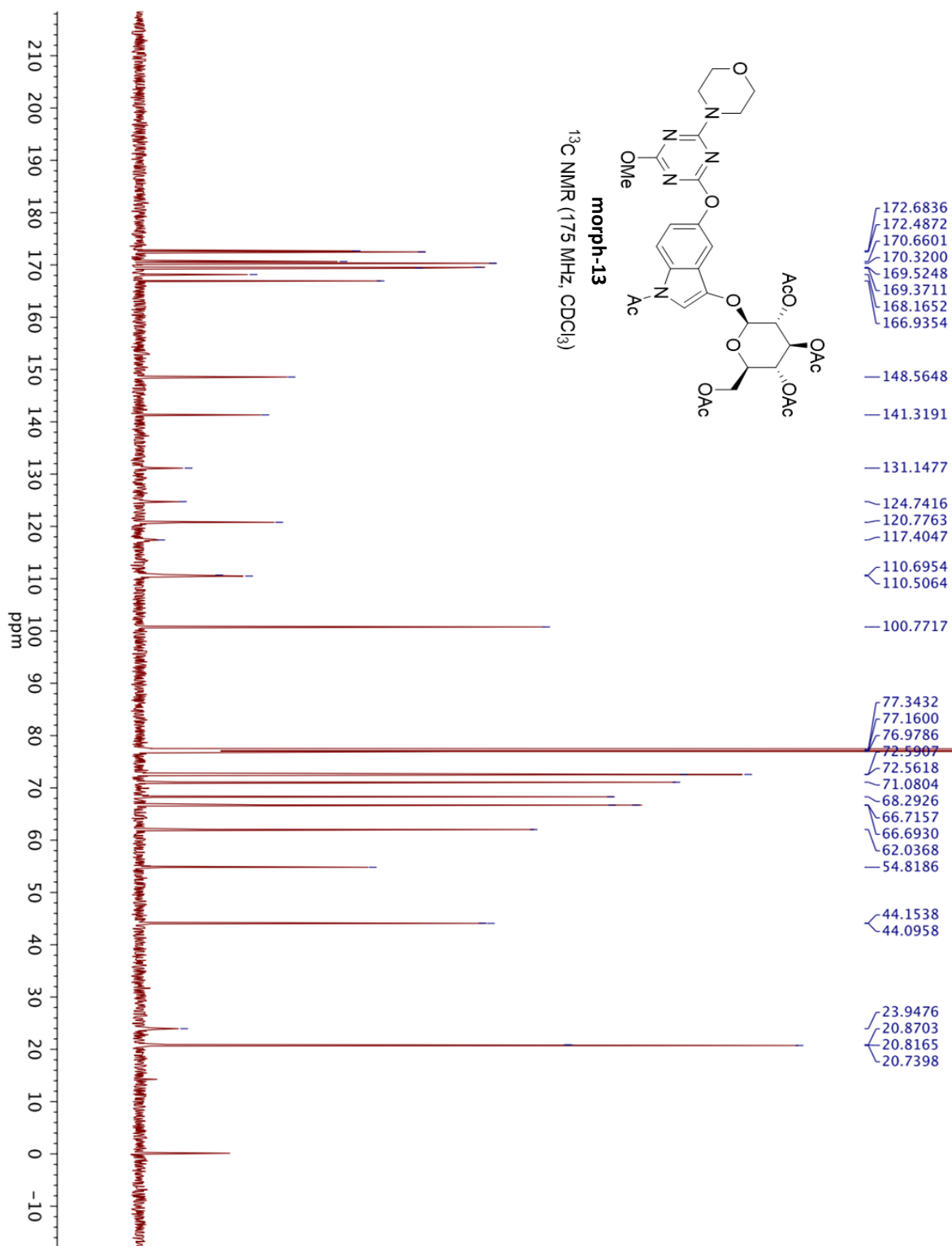


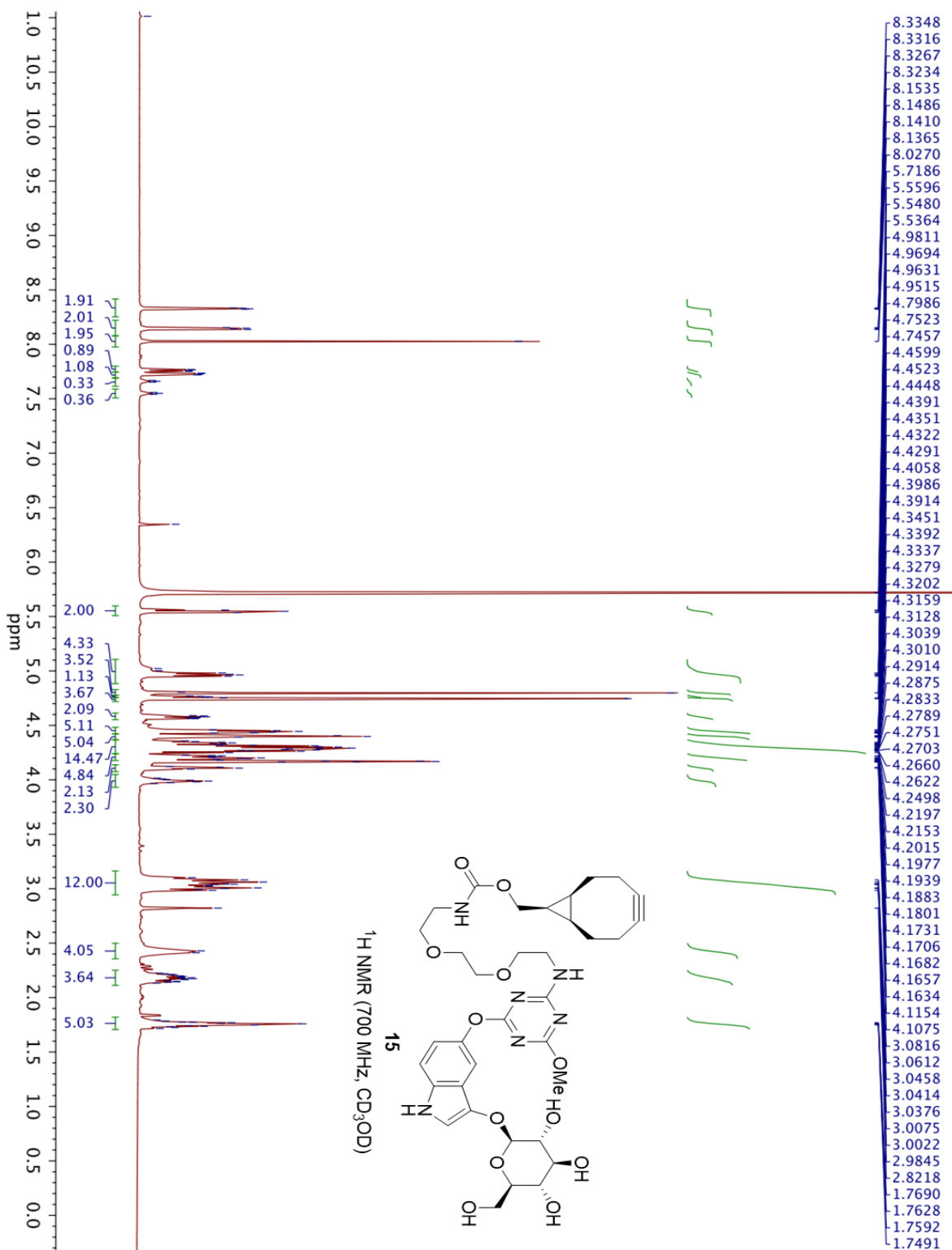


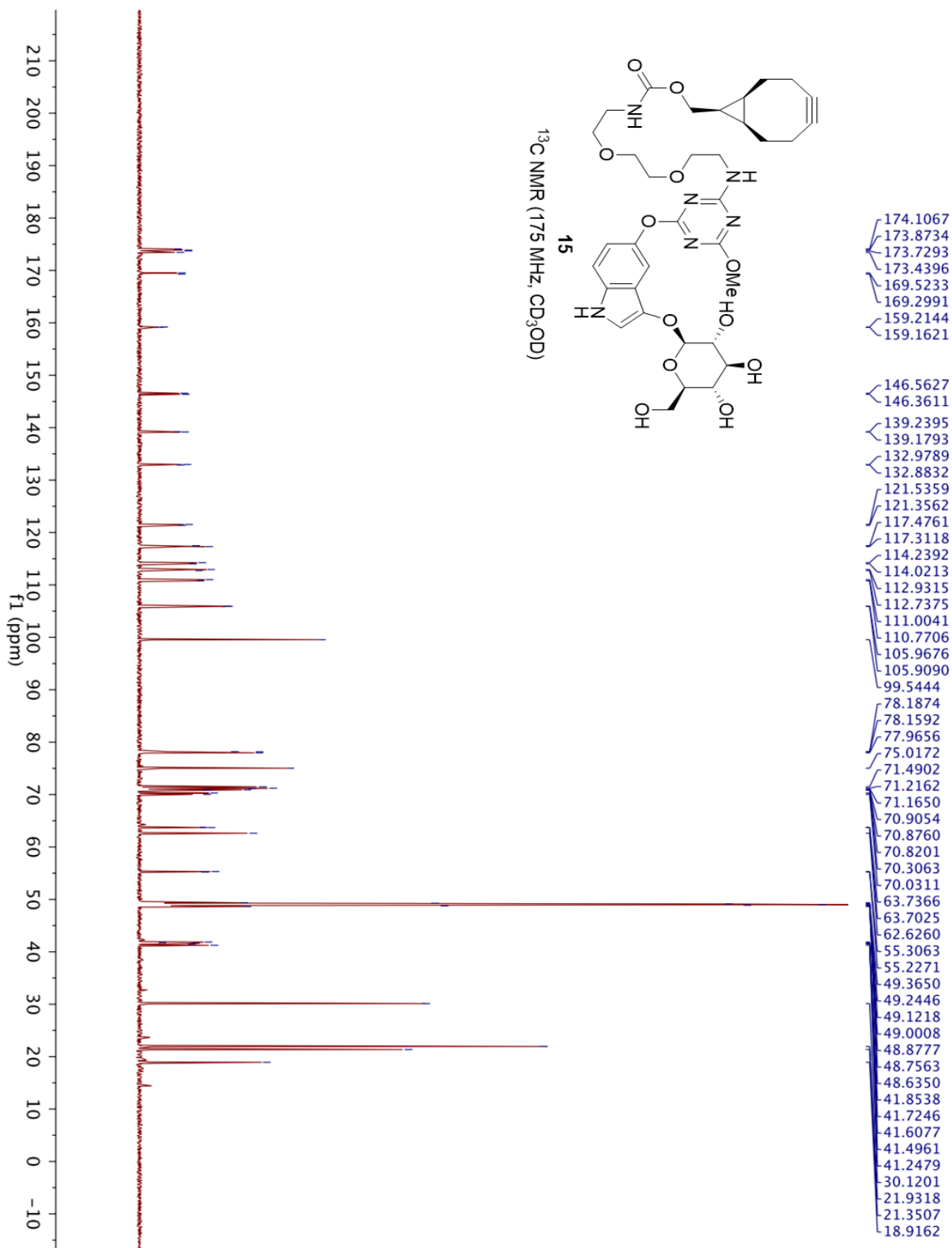


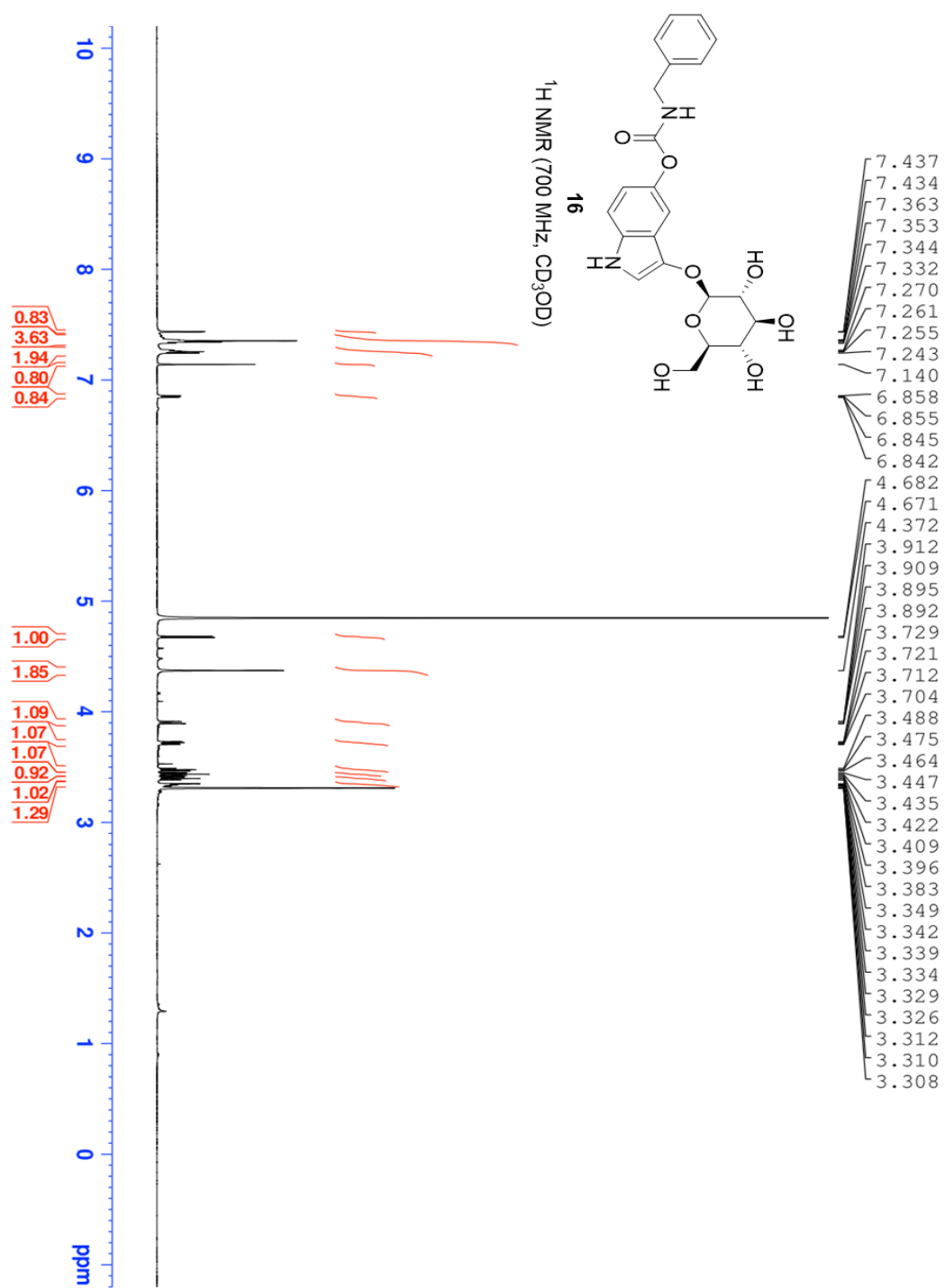


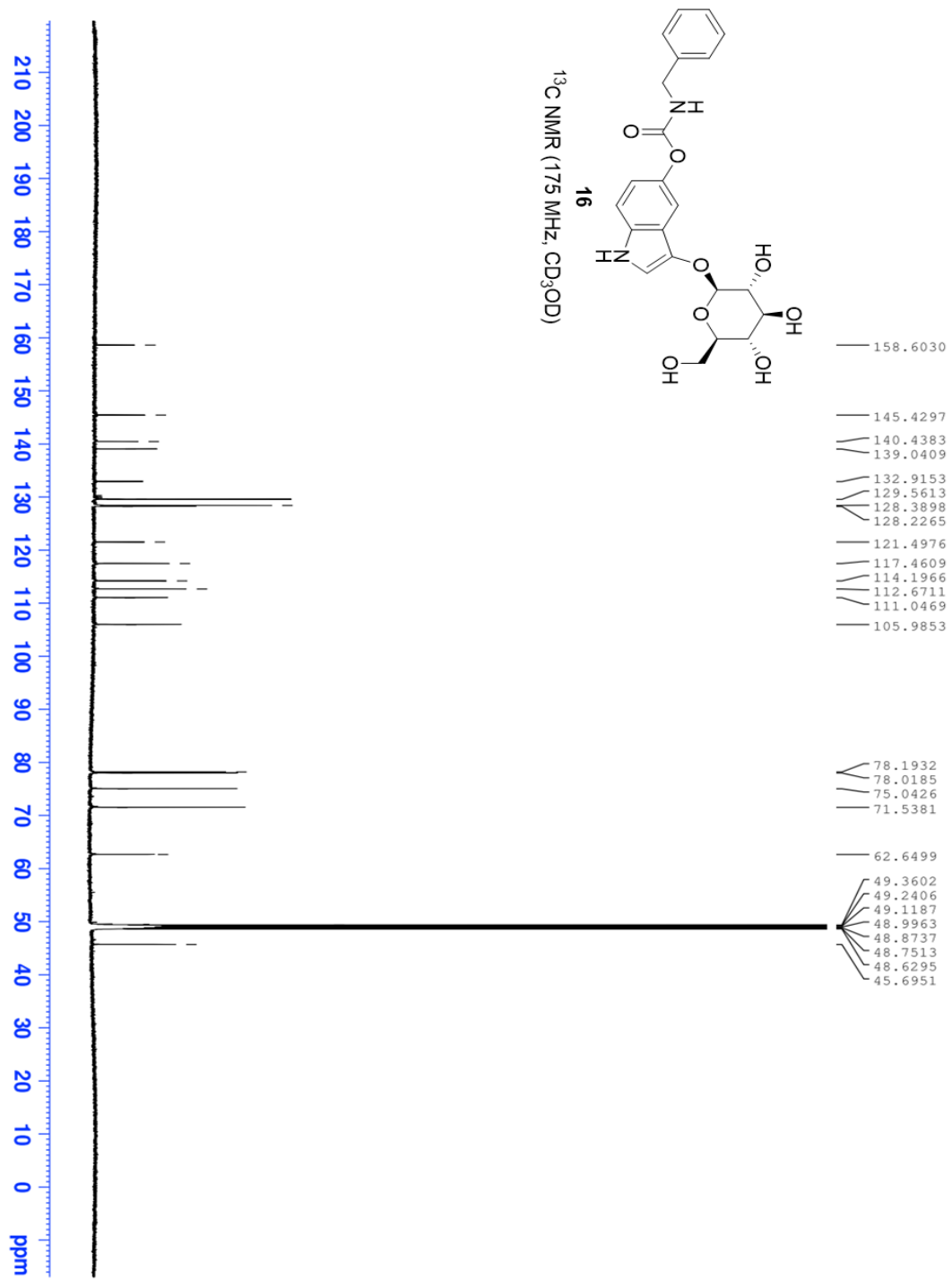


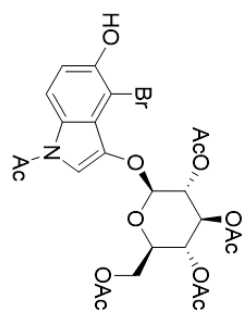




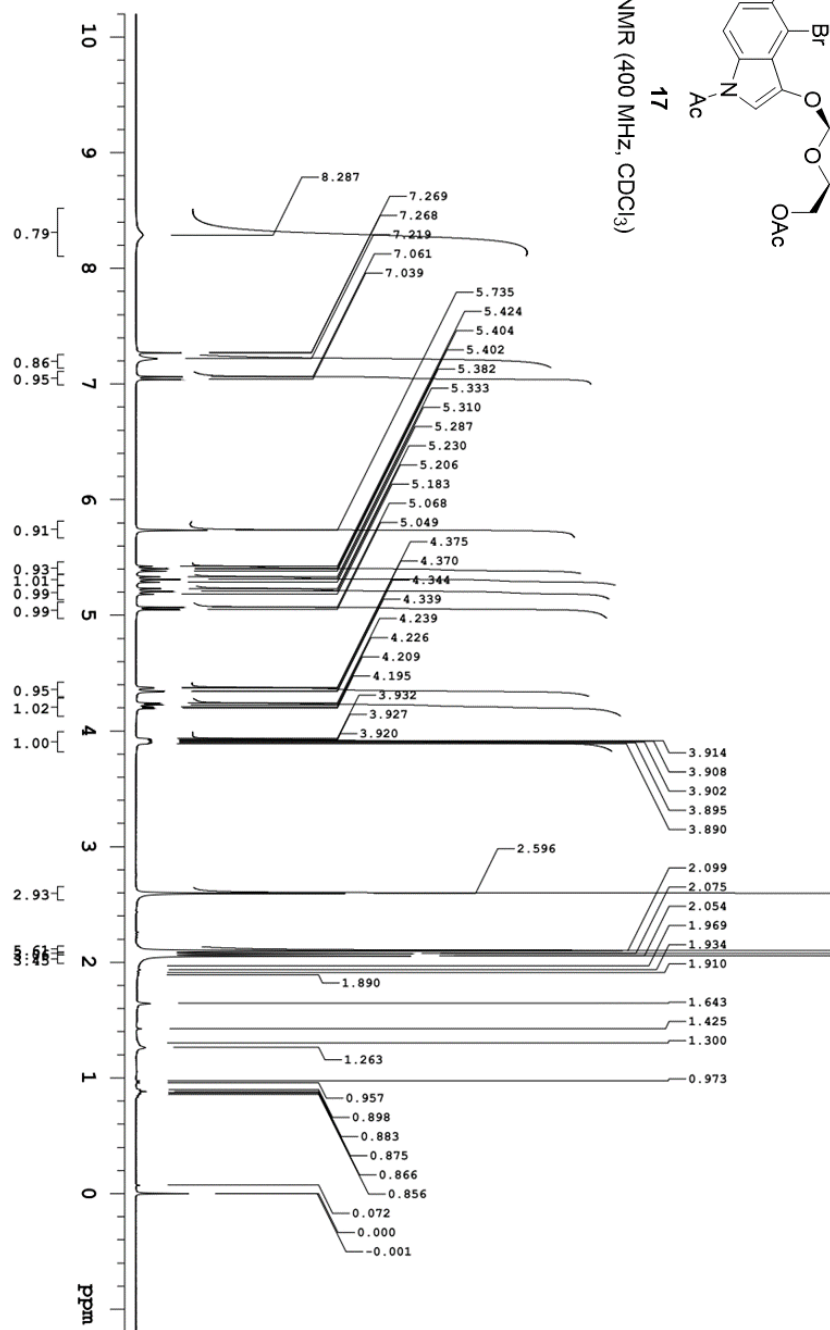


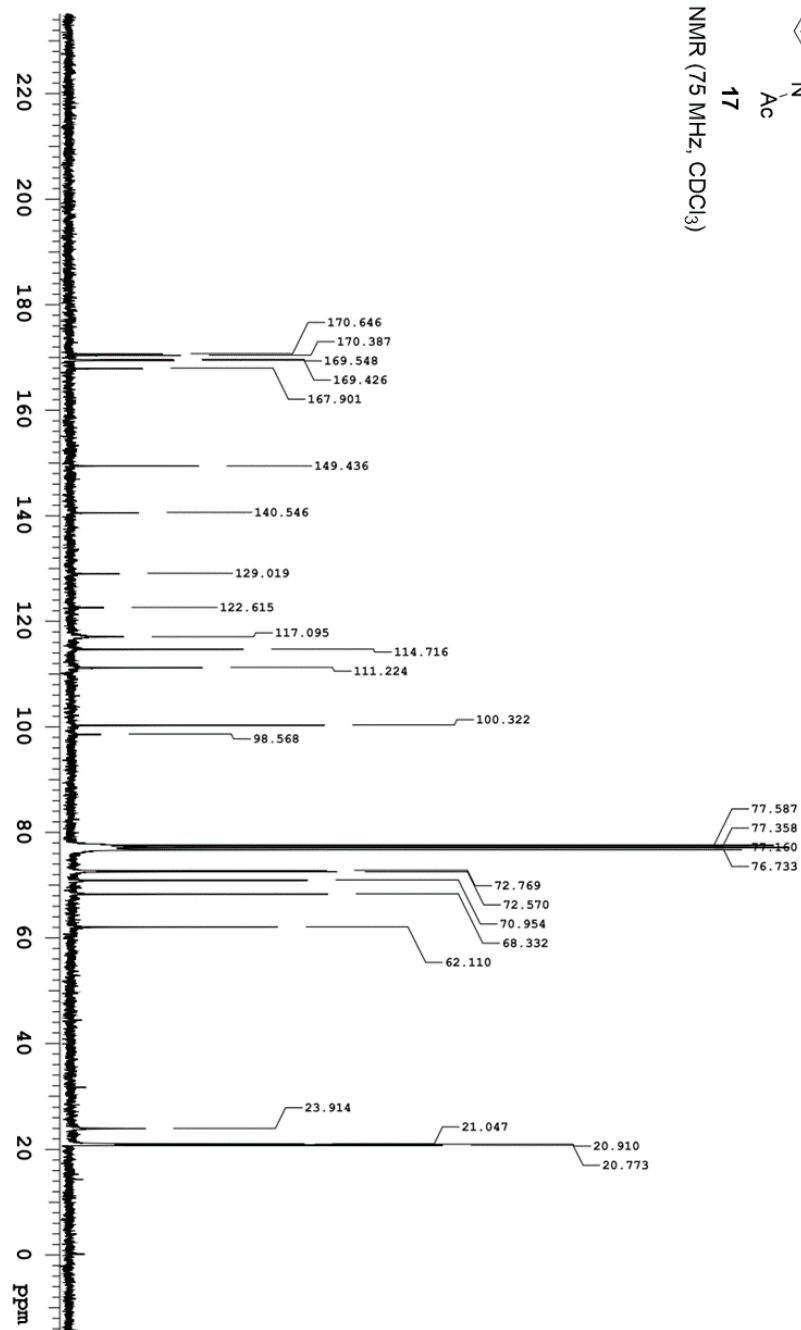
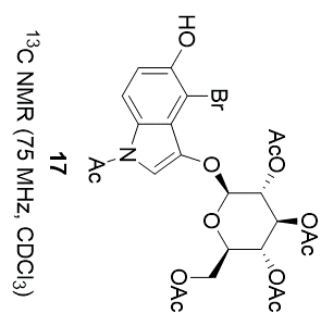


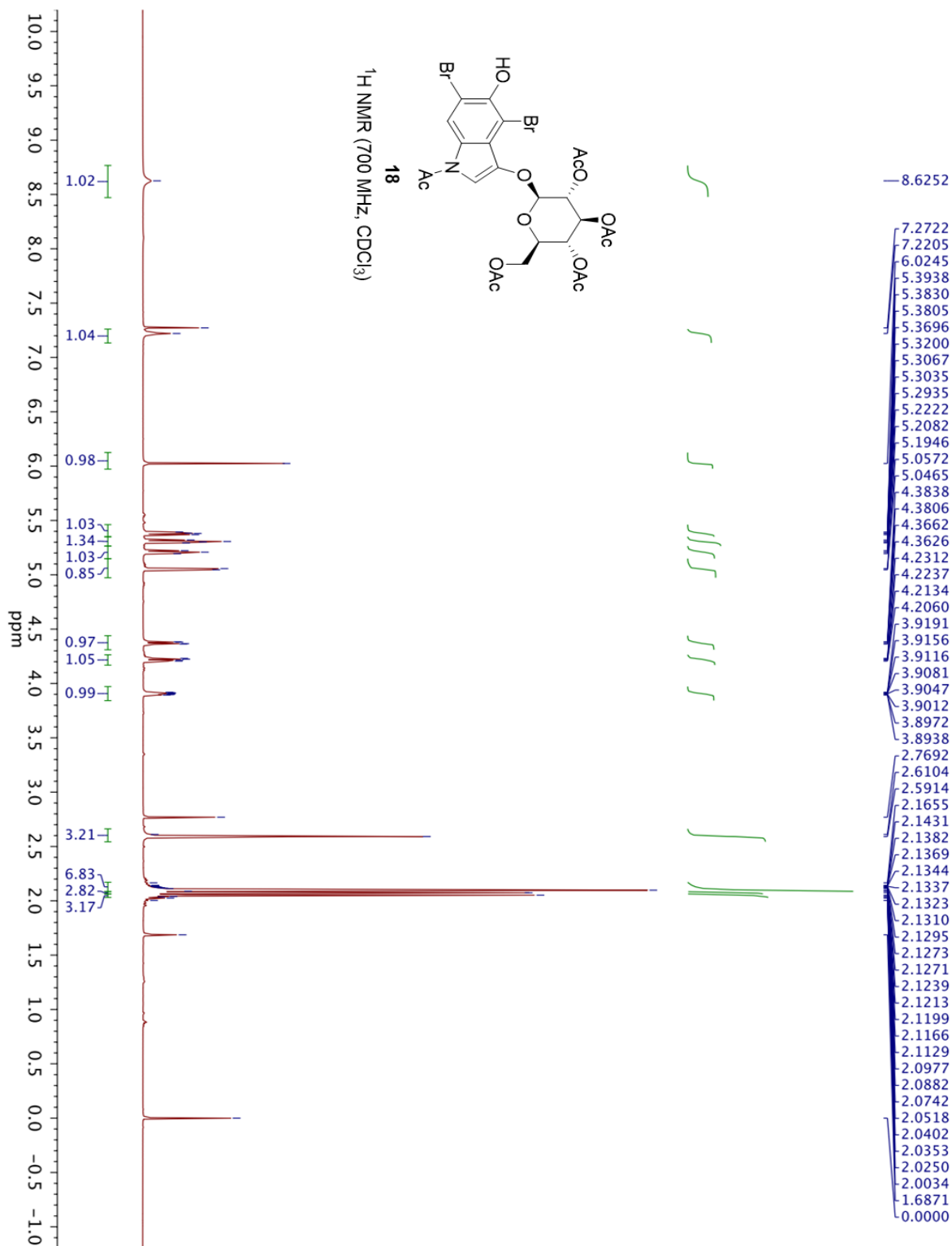


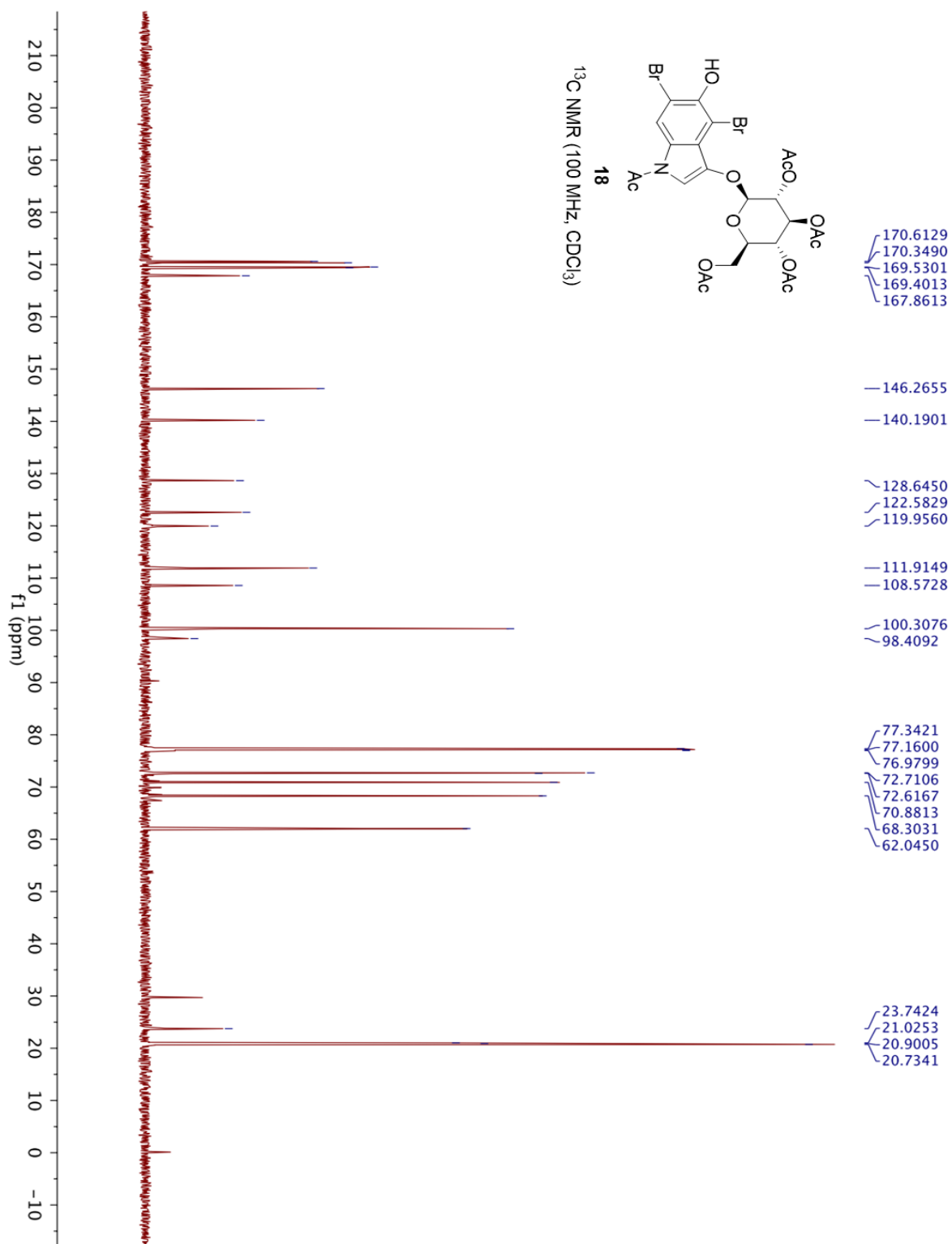


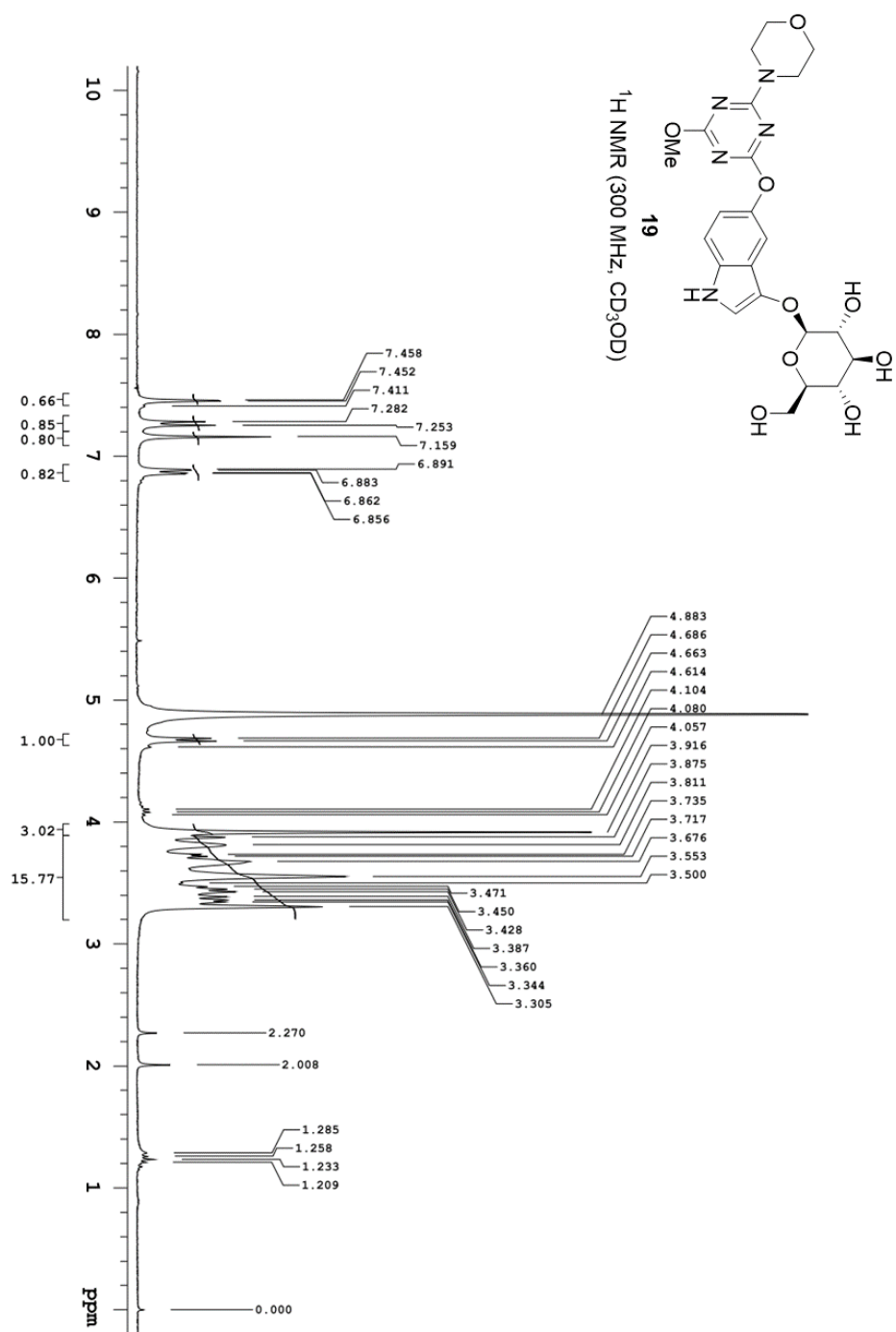
¹H NMR (400 MHz, CDCl₃)

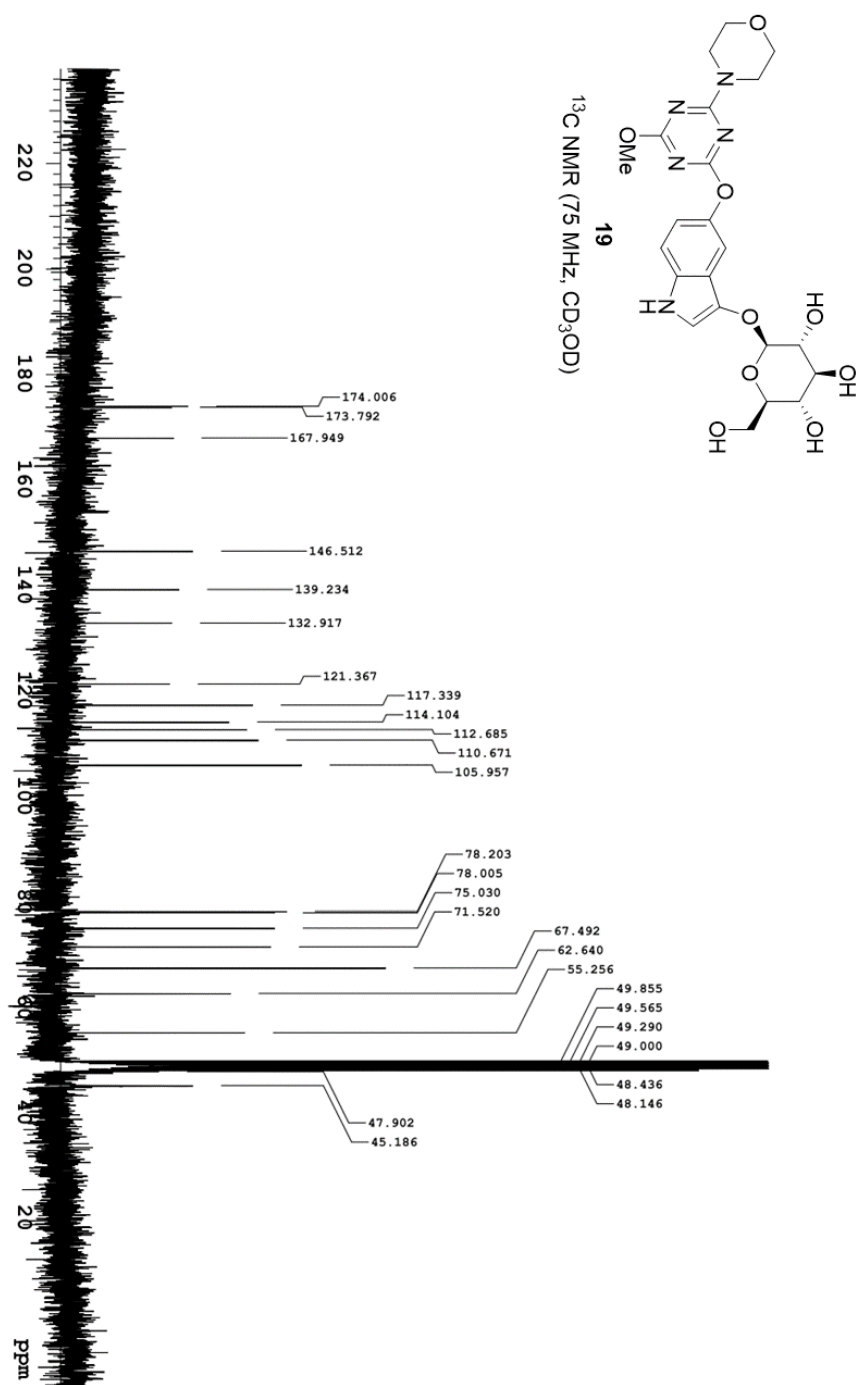


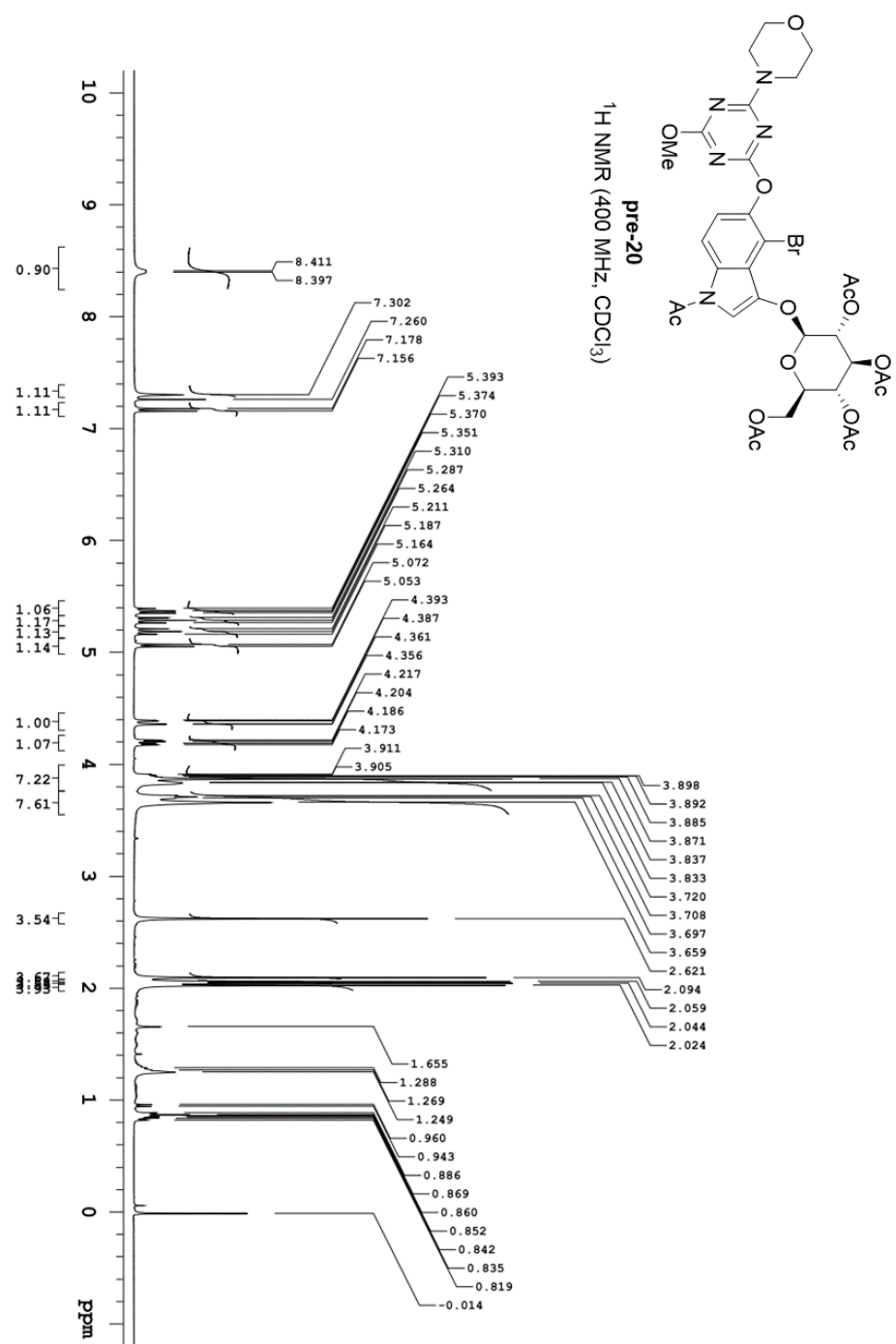


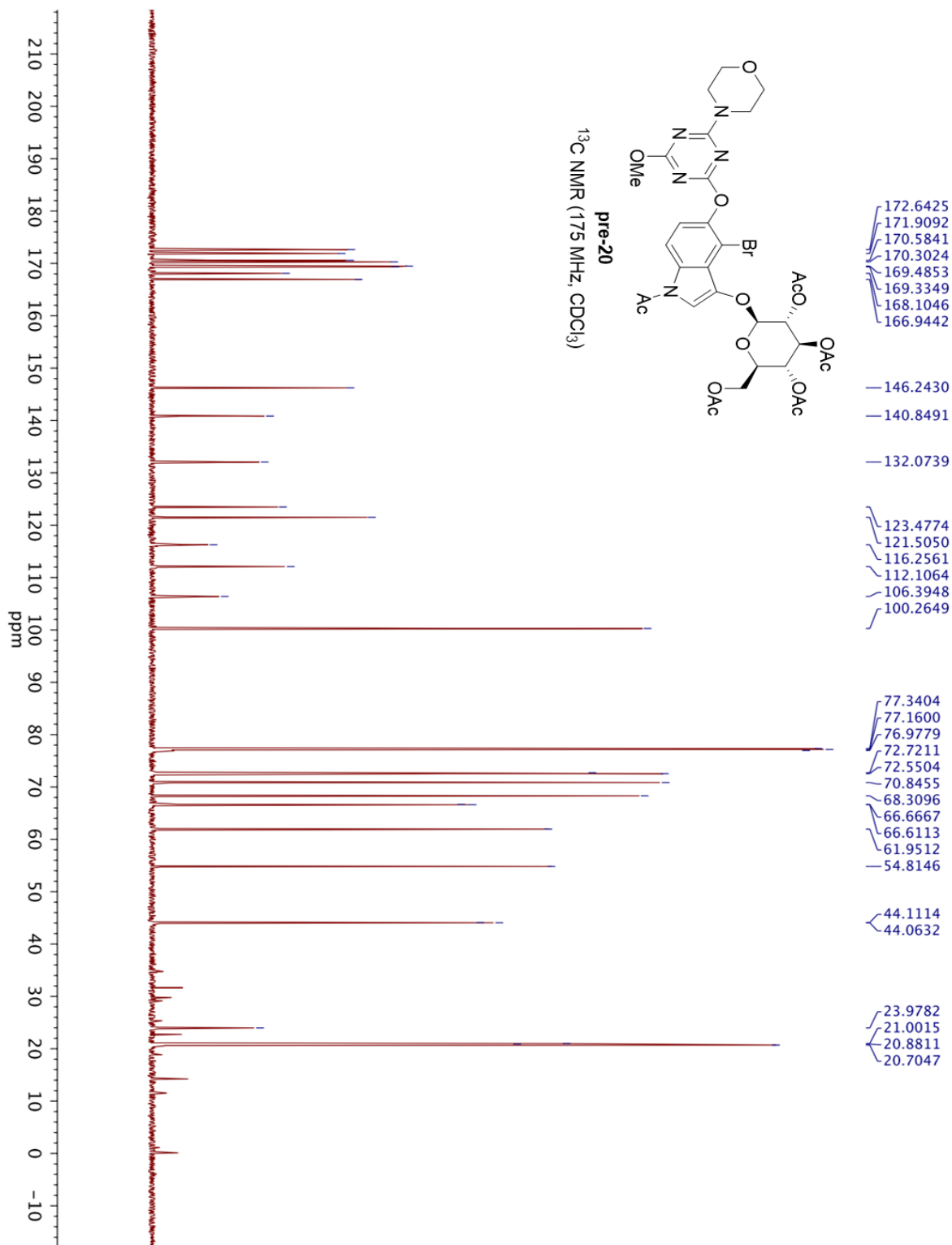


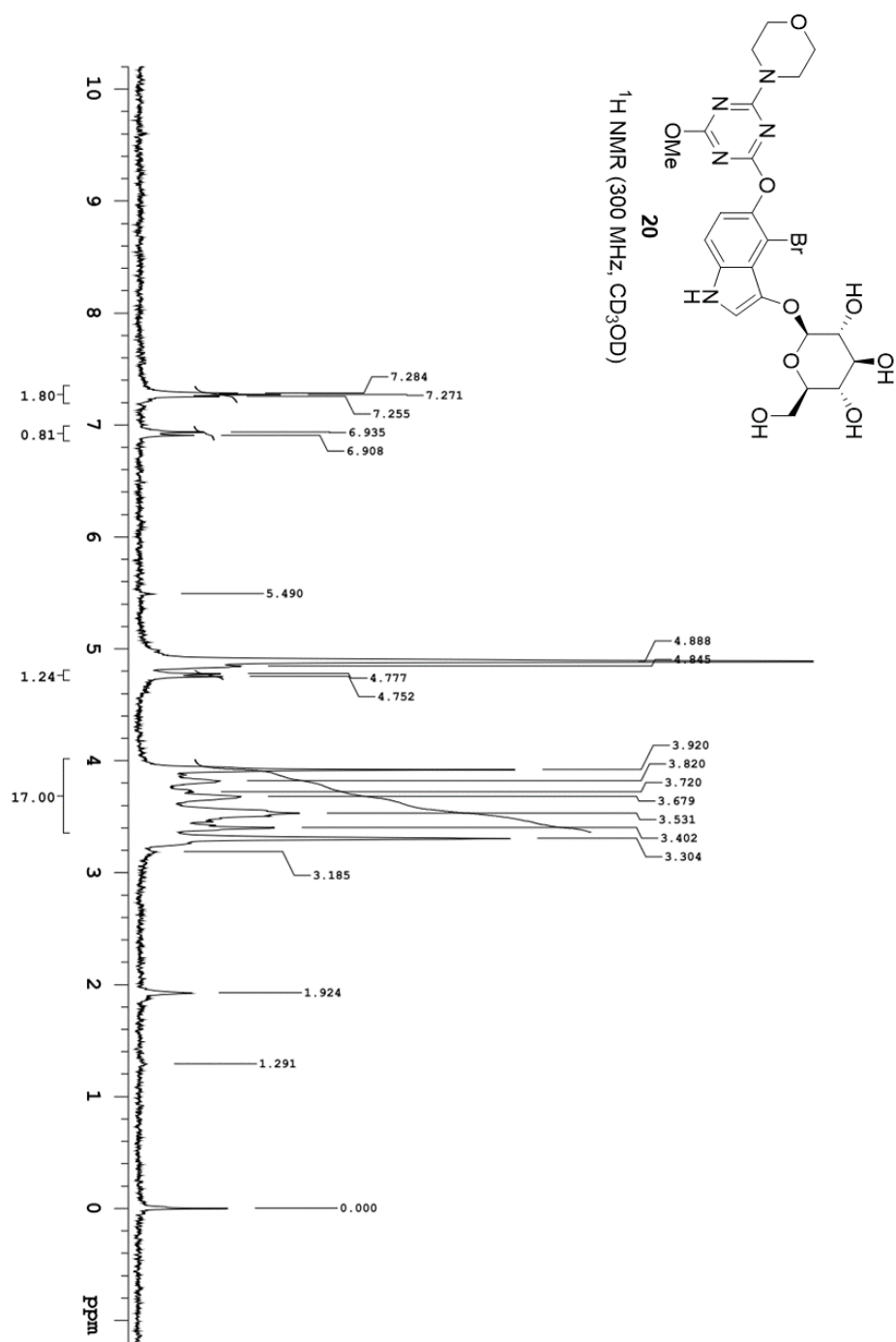


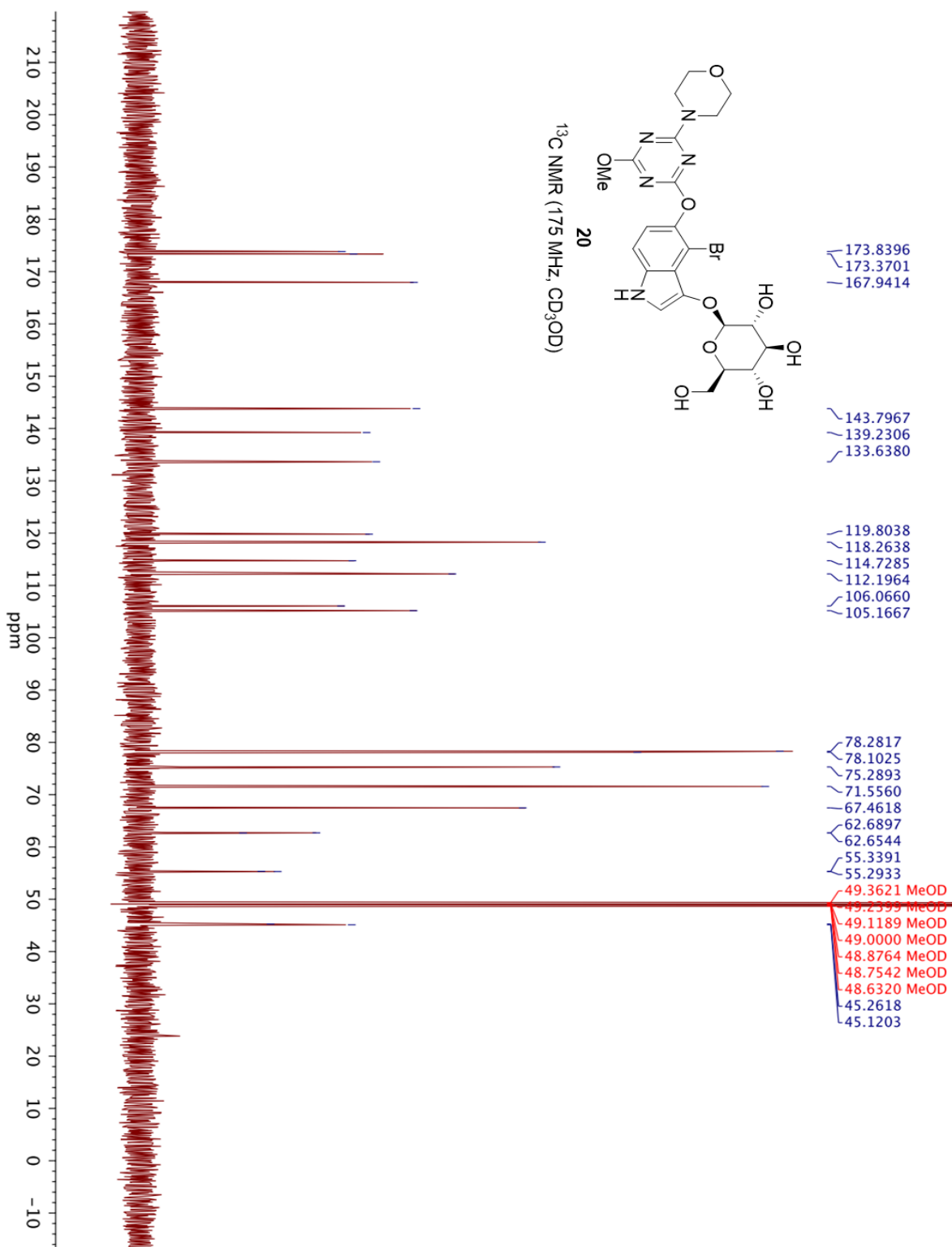


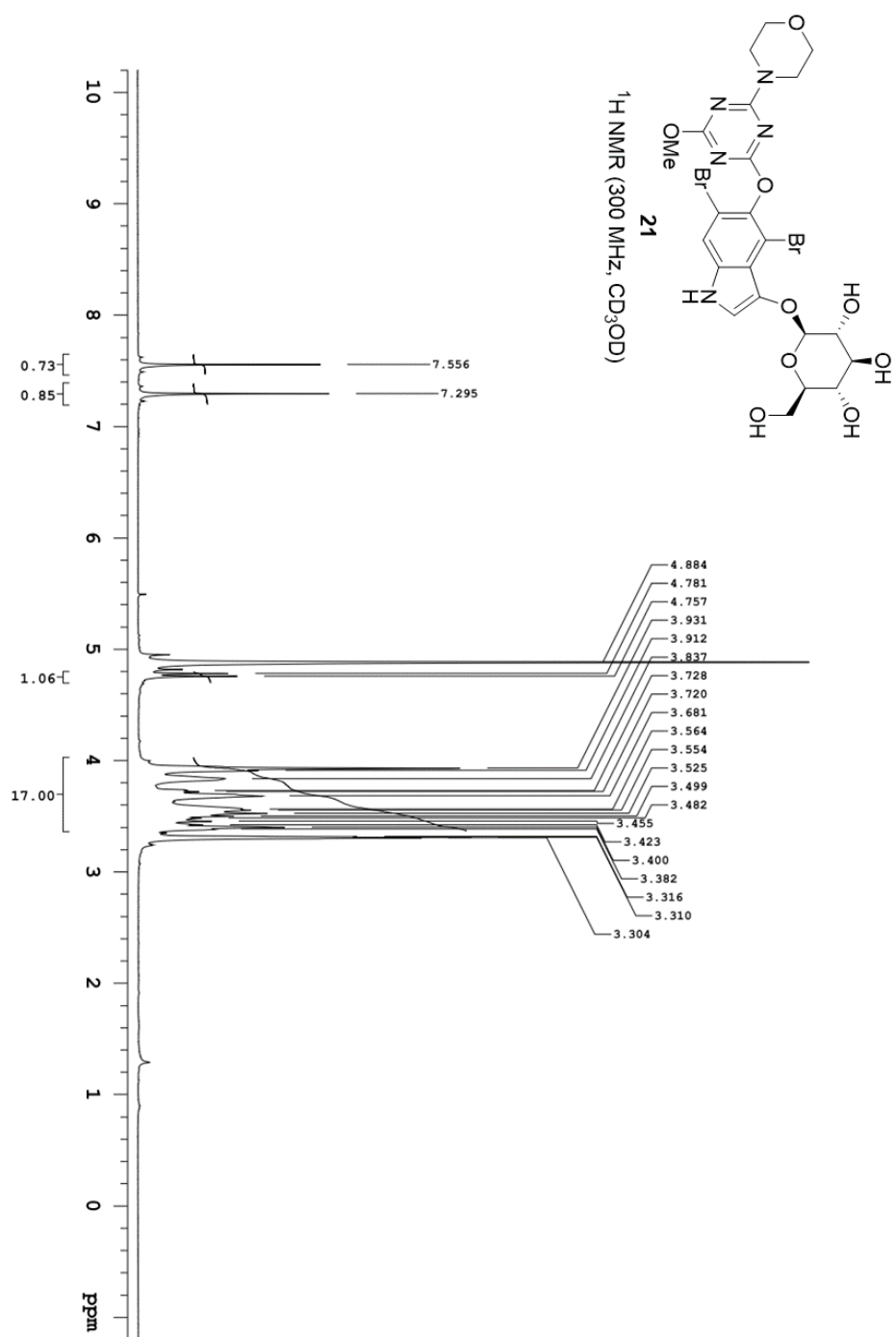


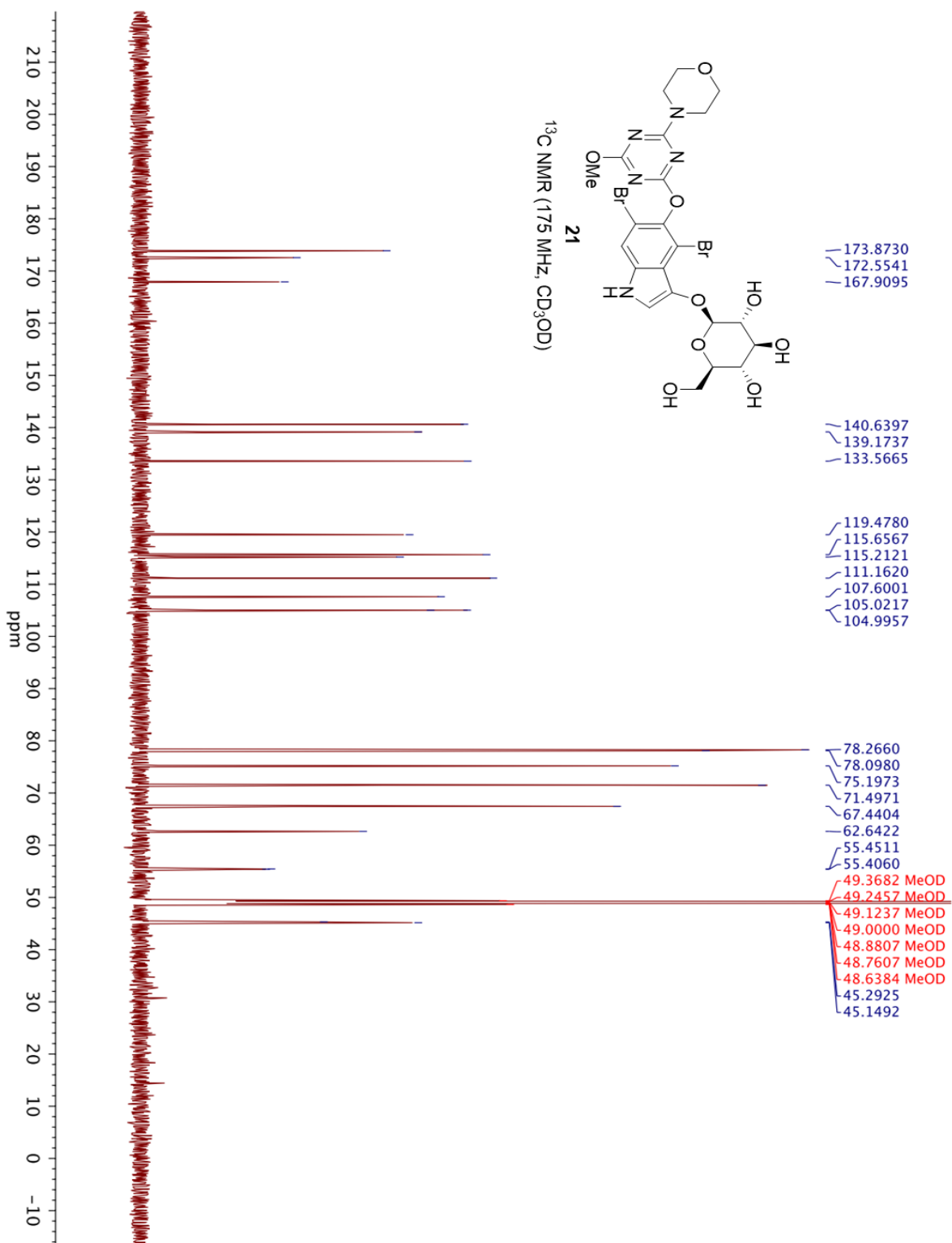


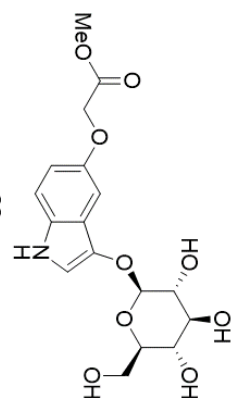




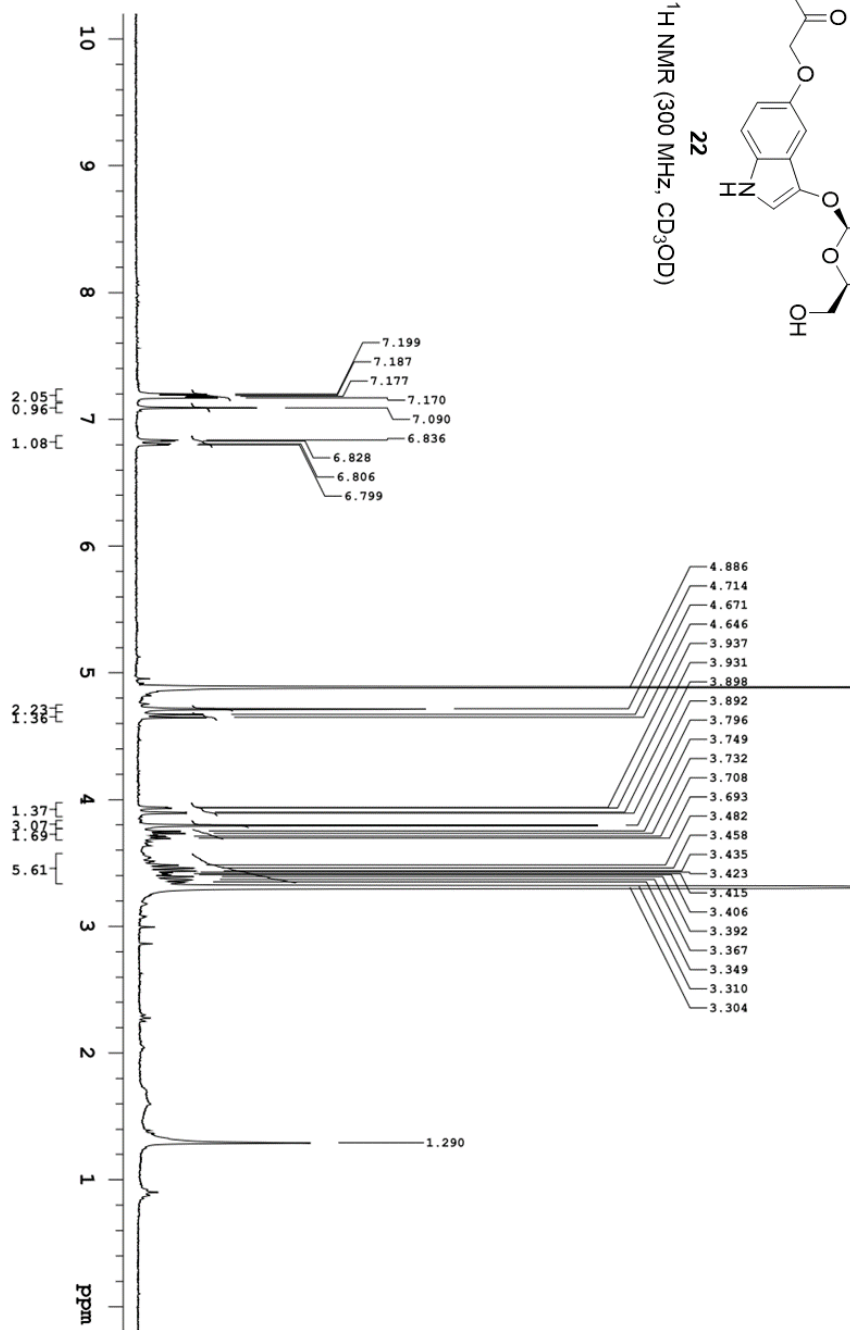


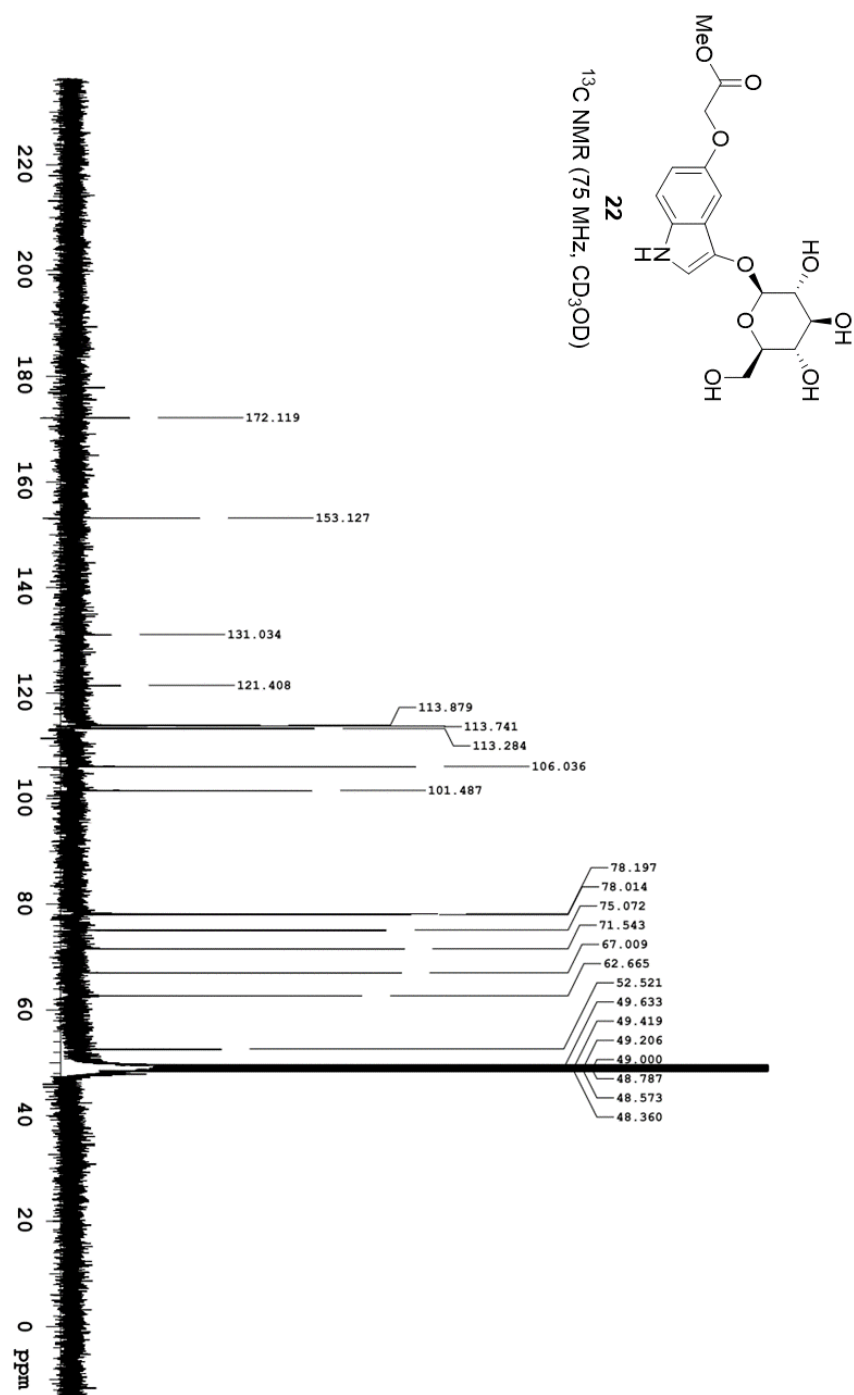


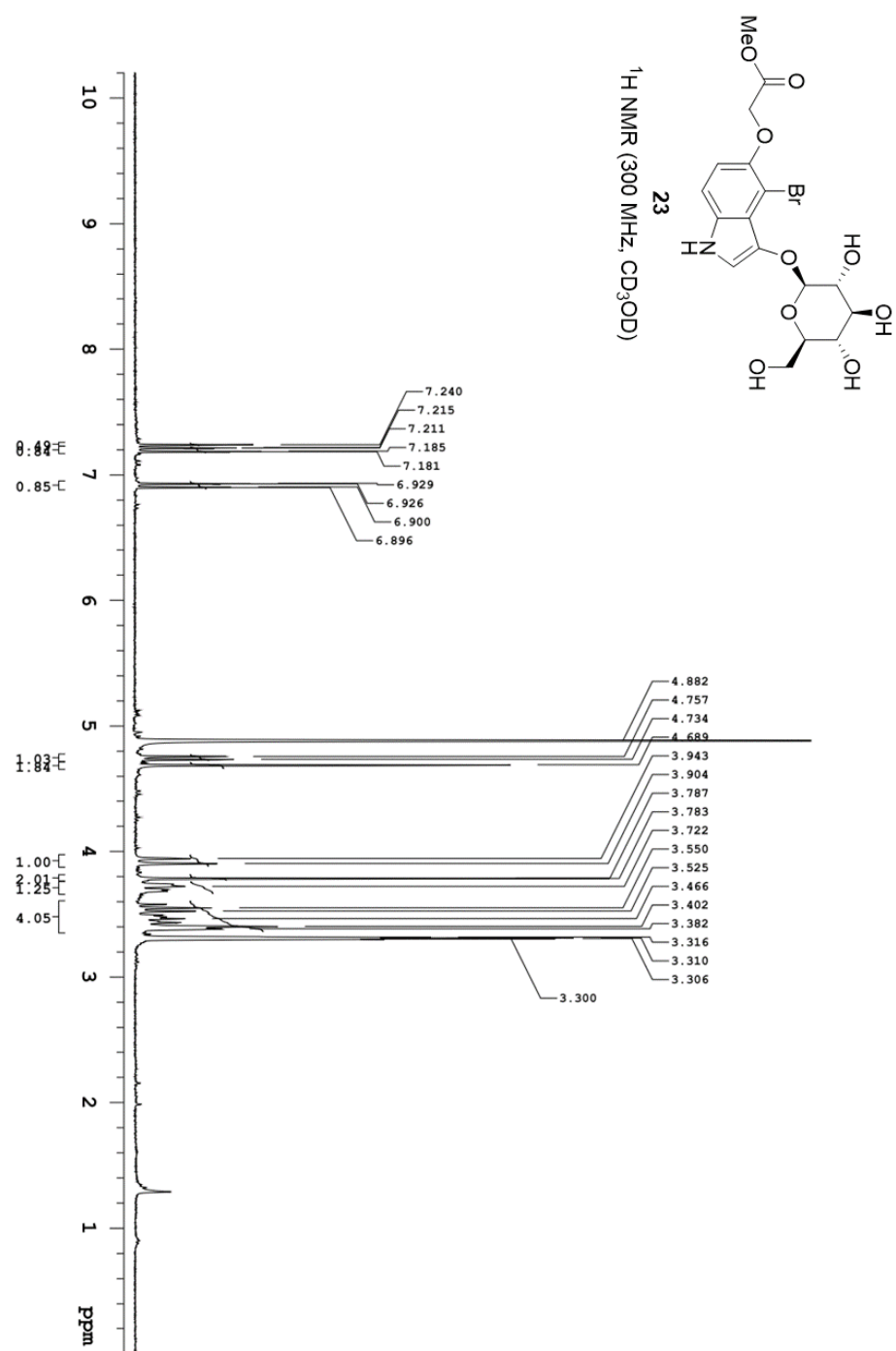


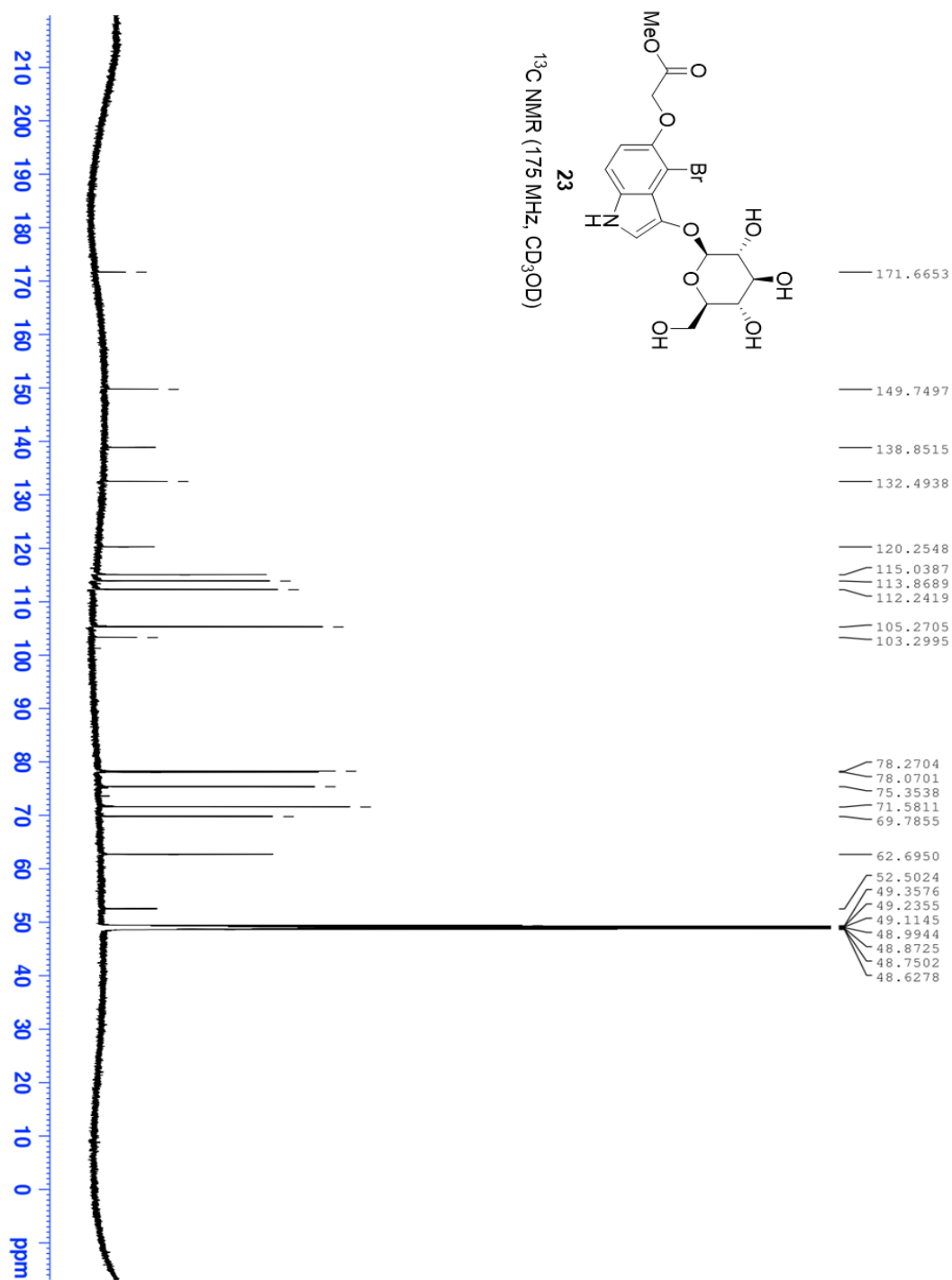


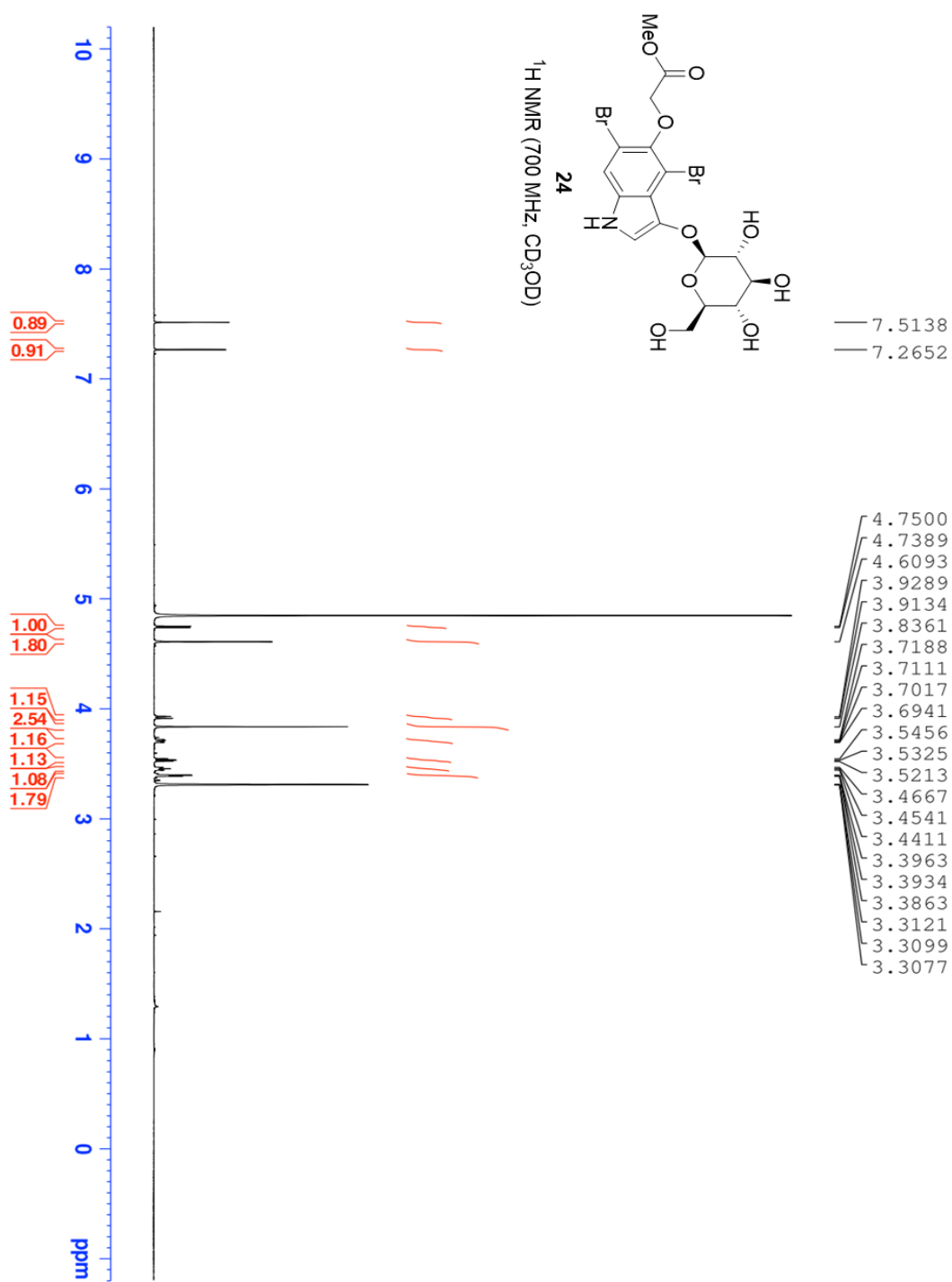
¹H NMR (300 MHz, CD₃OD)

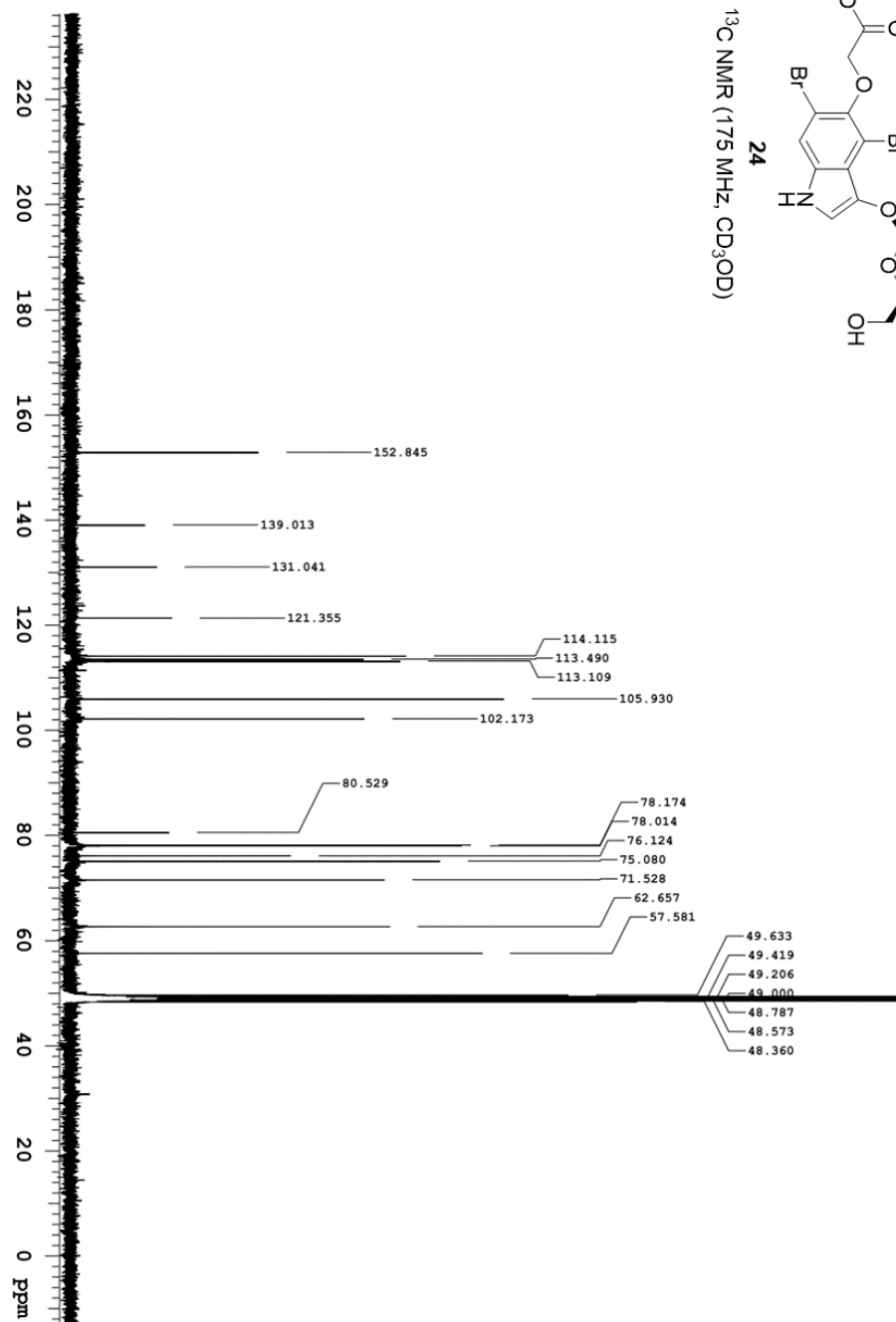
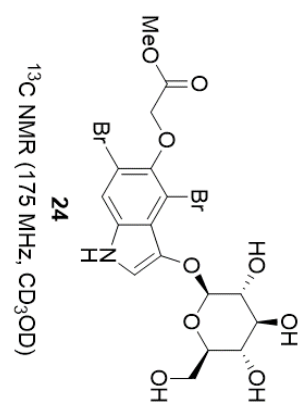


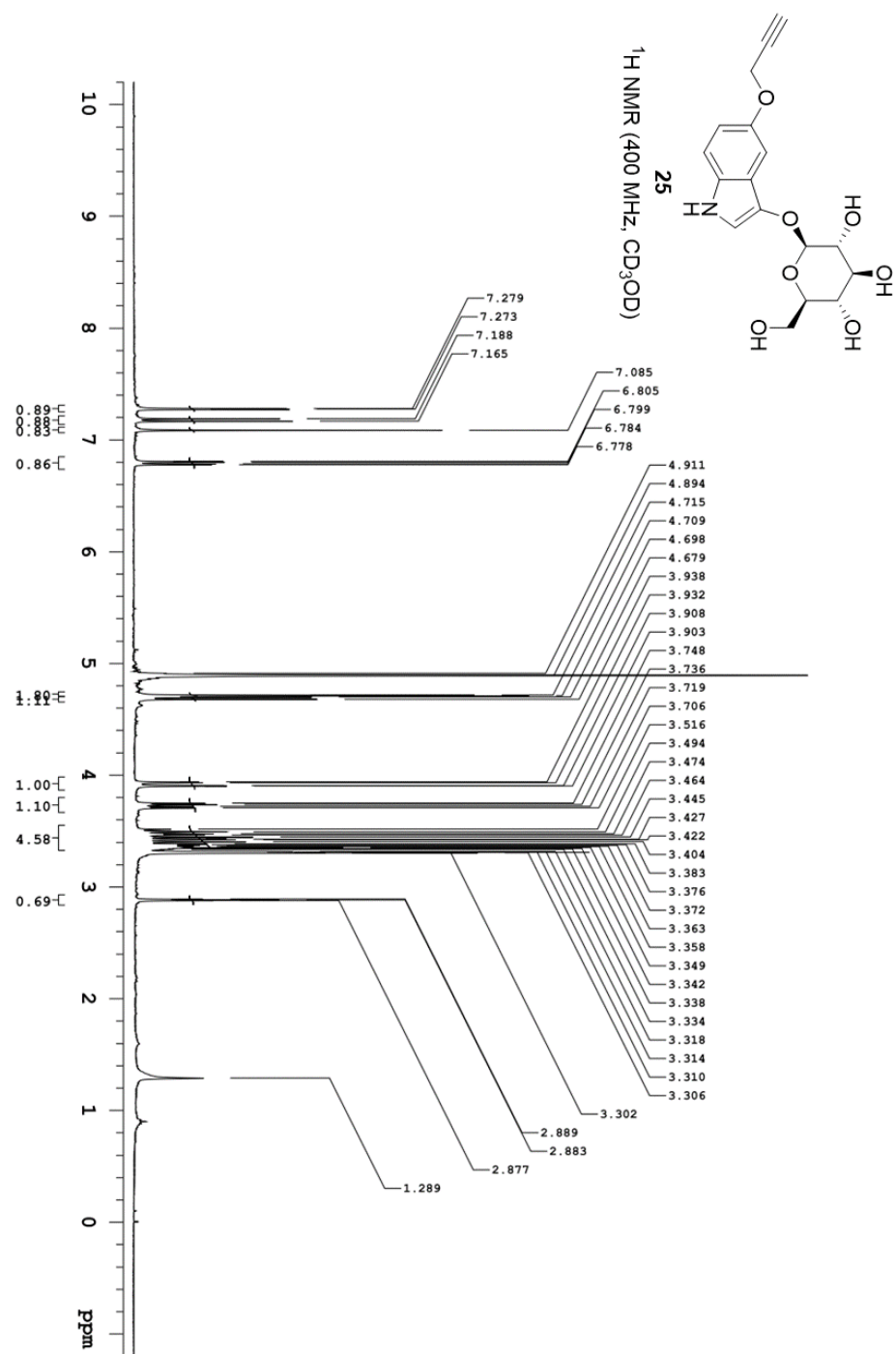


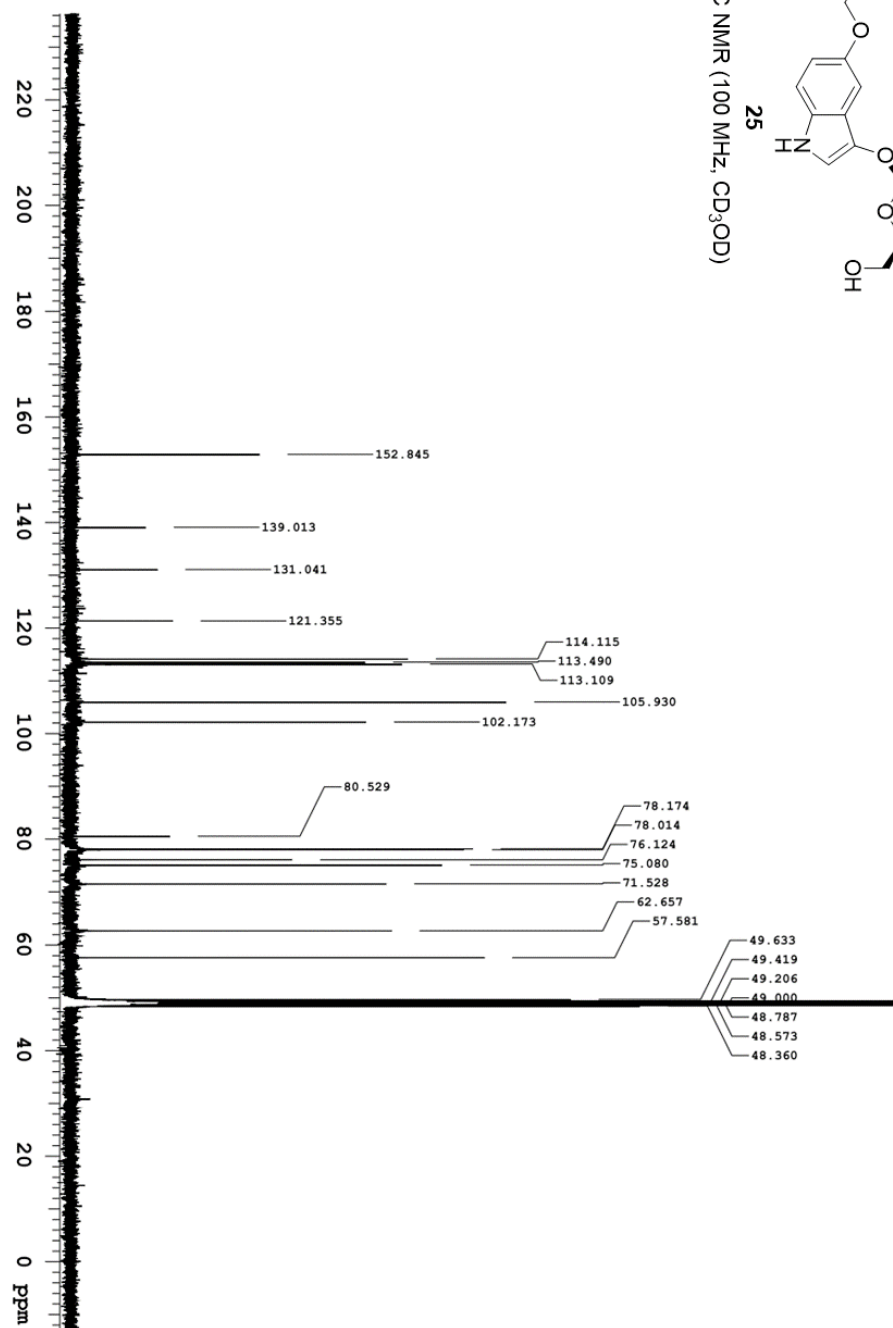
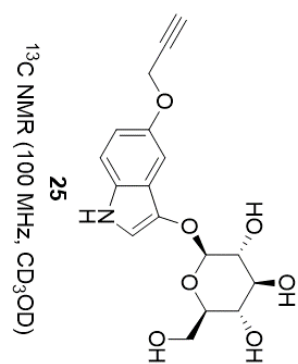


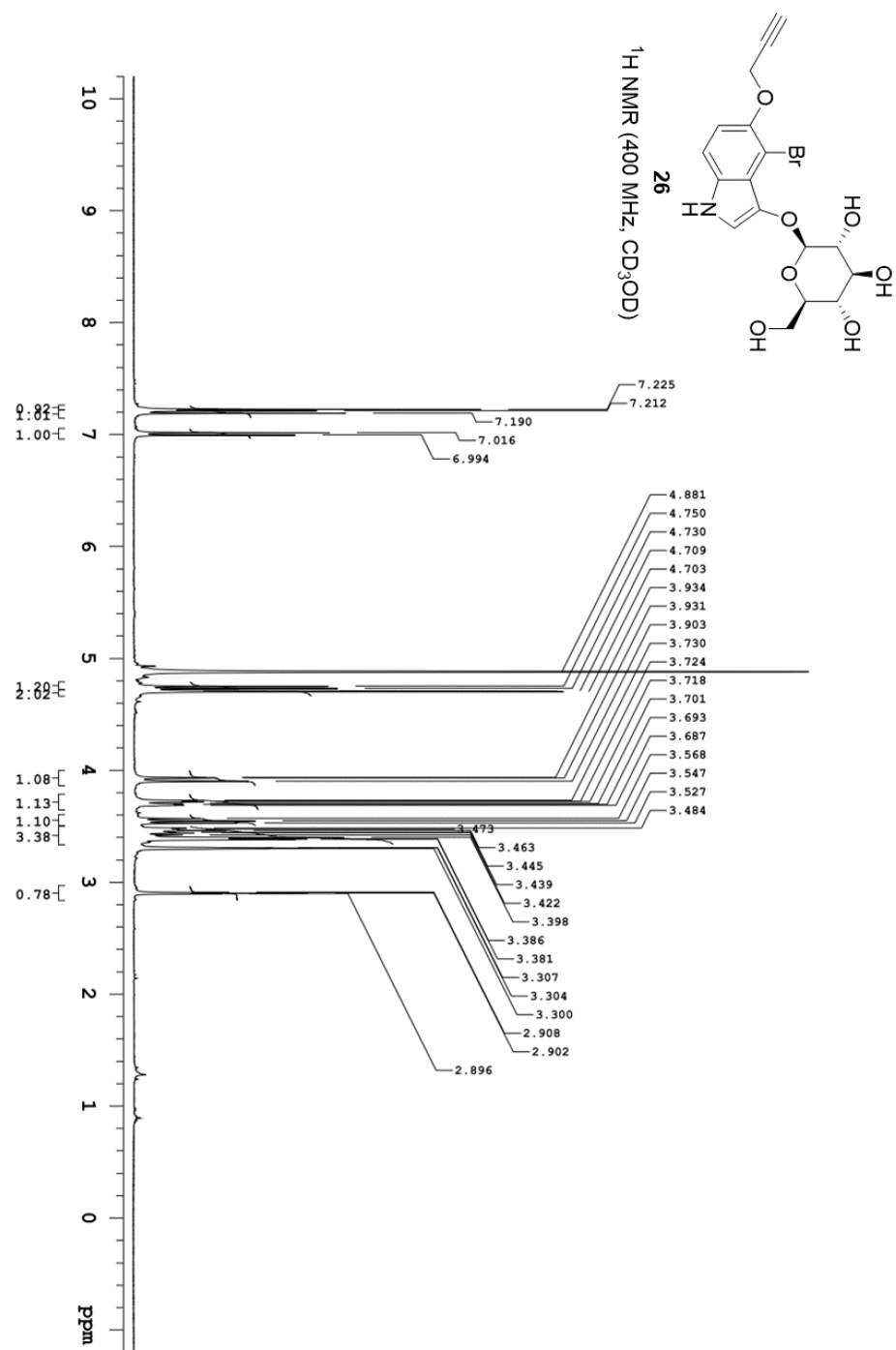


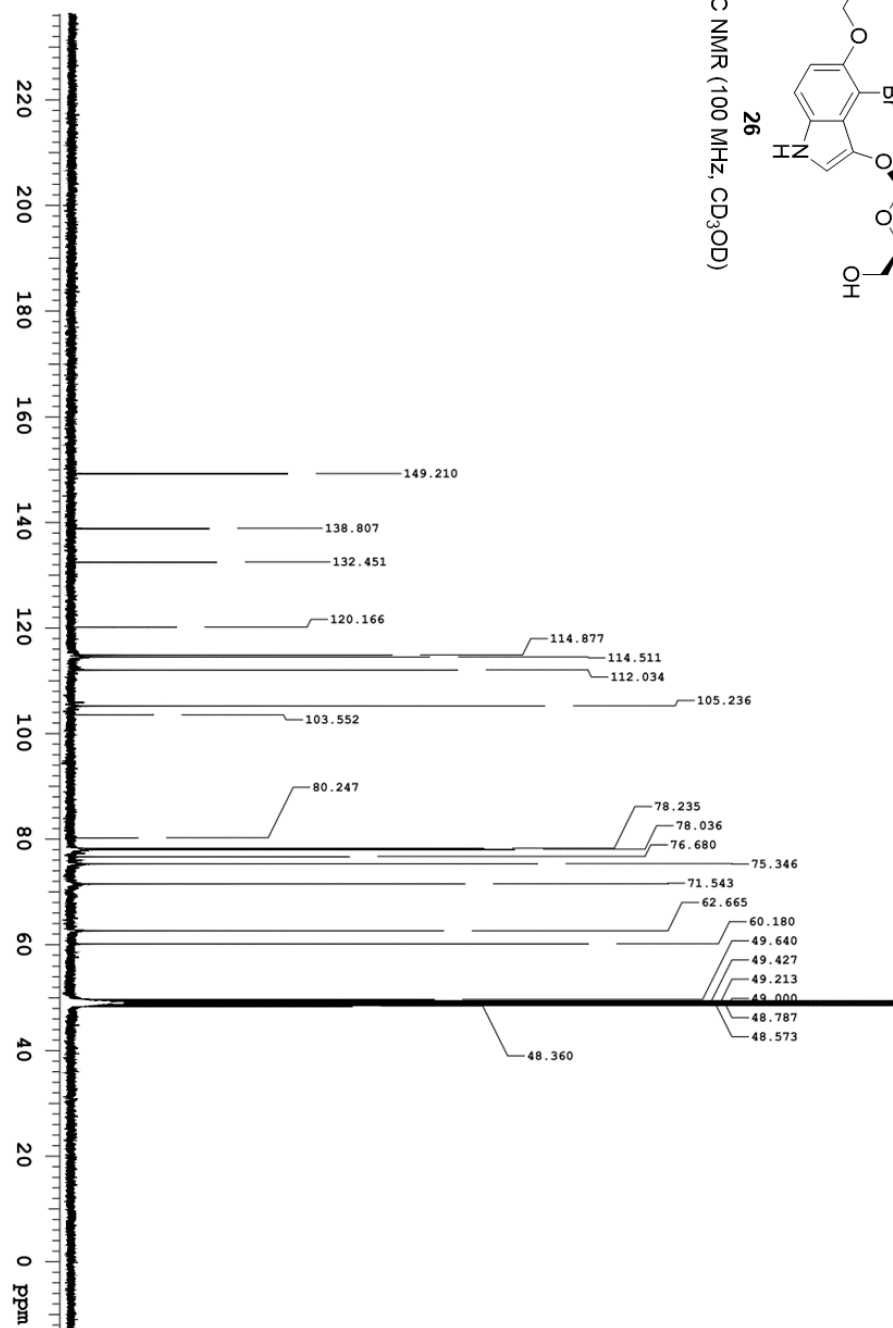
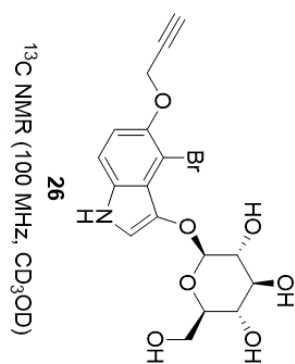


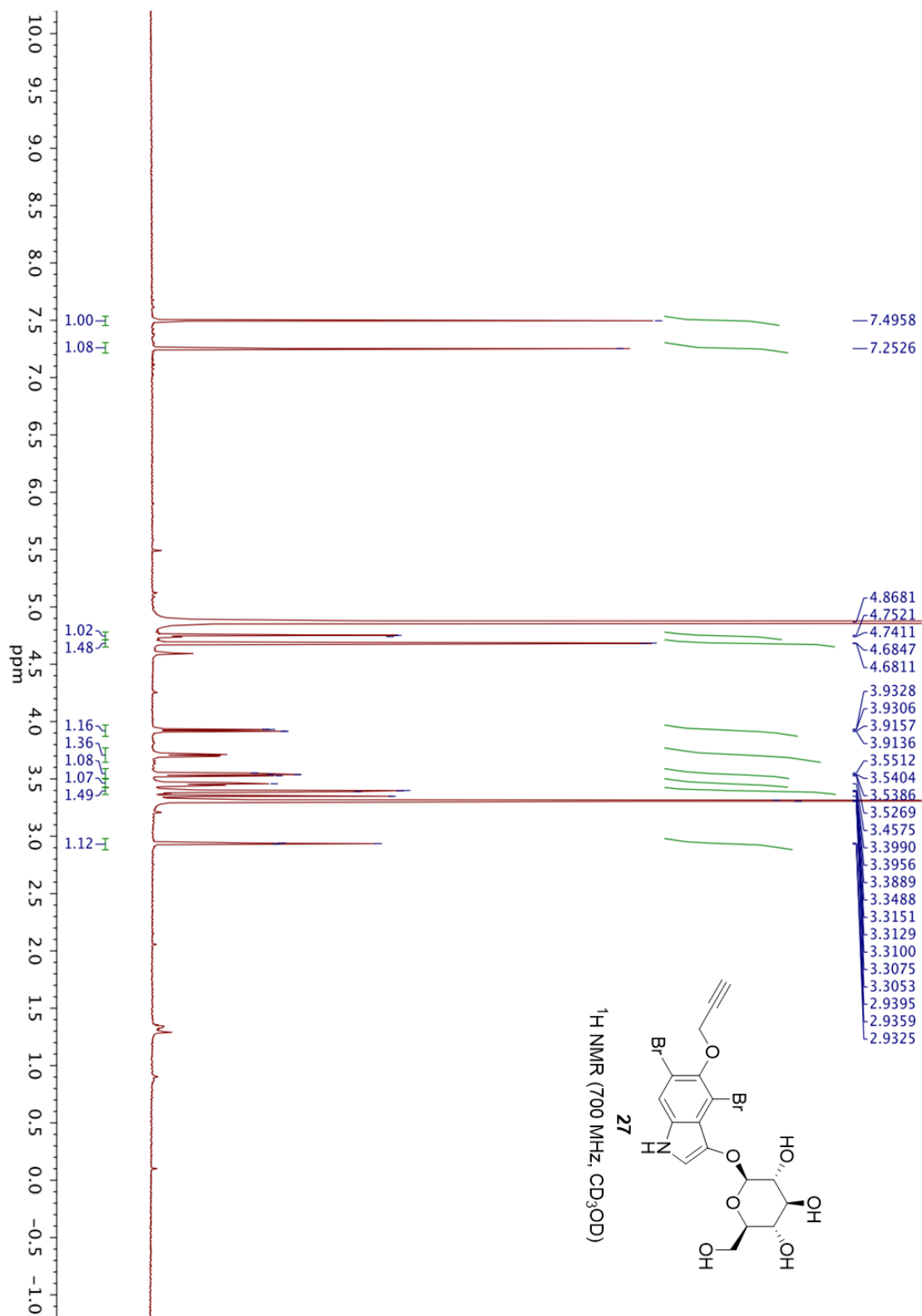


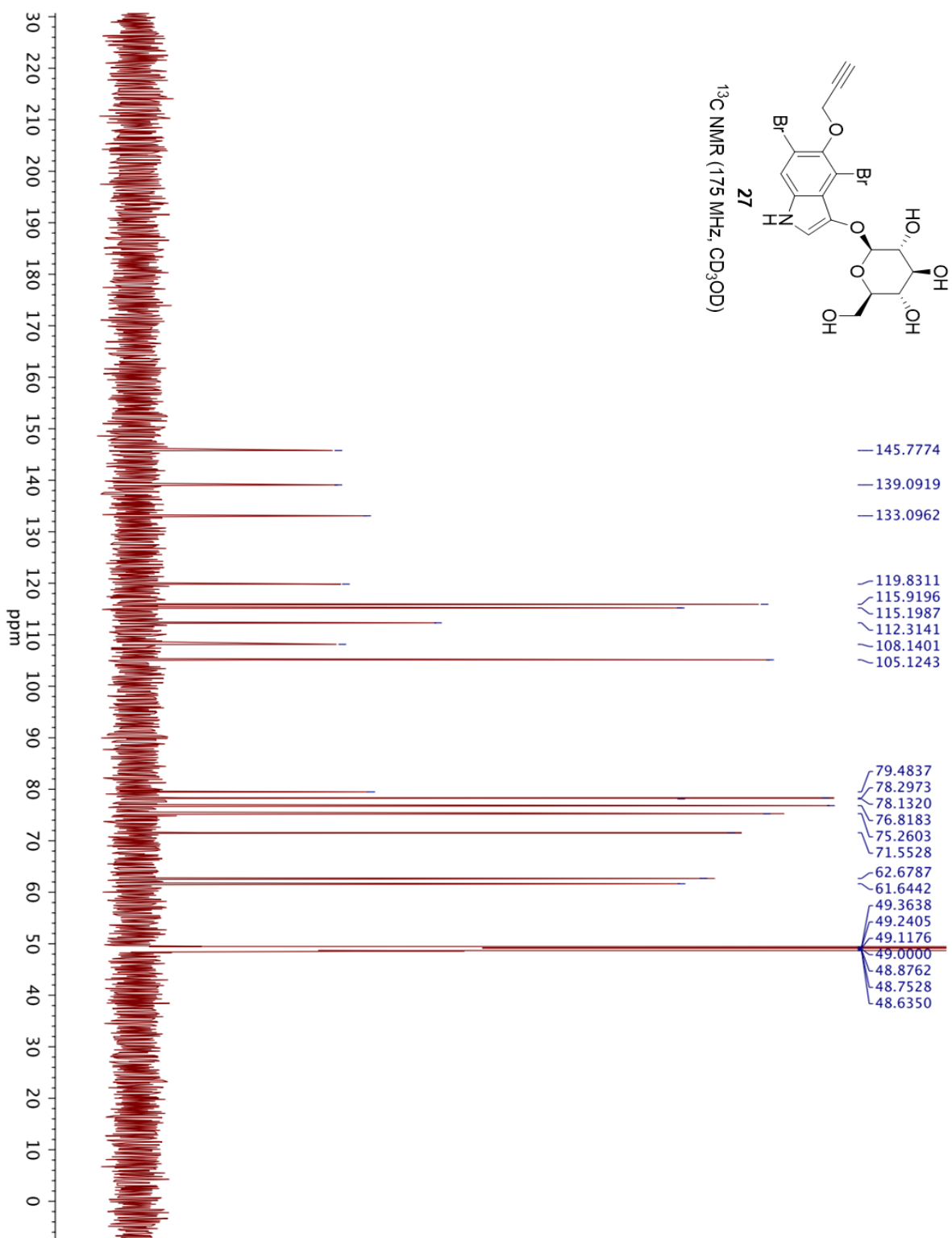


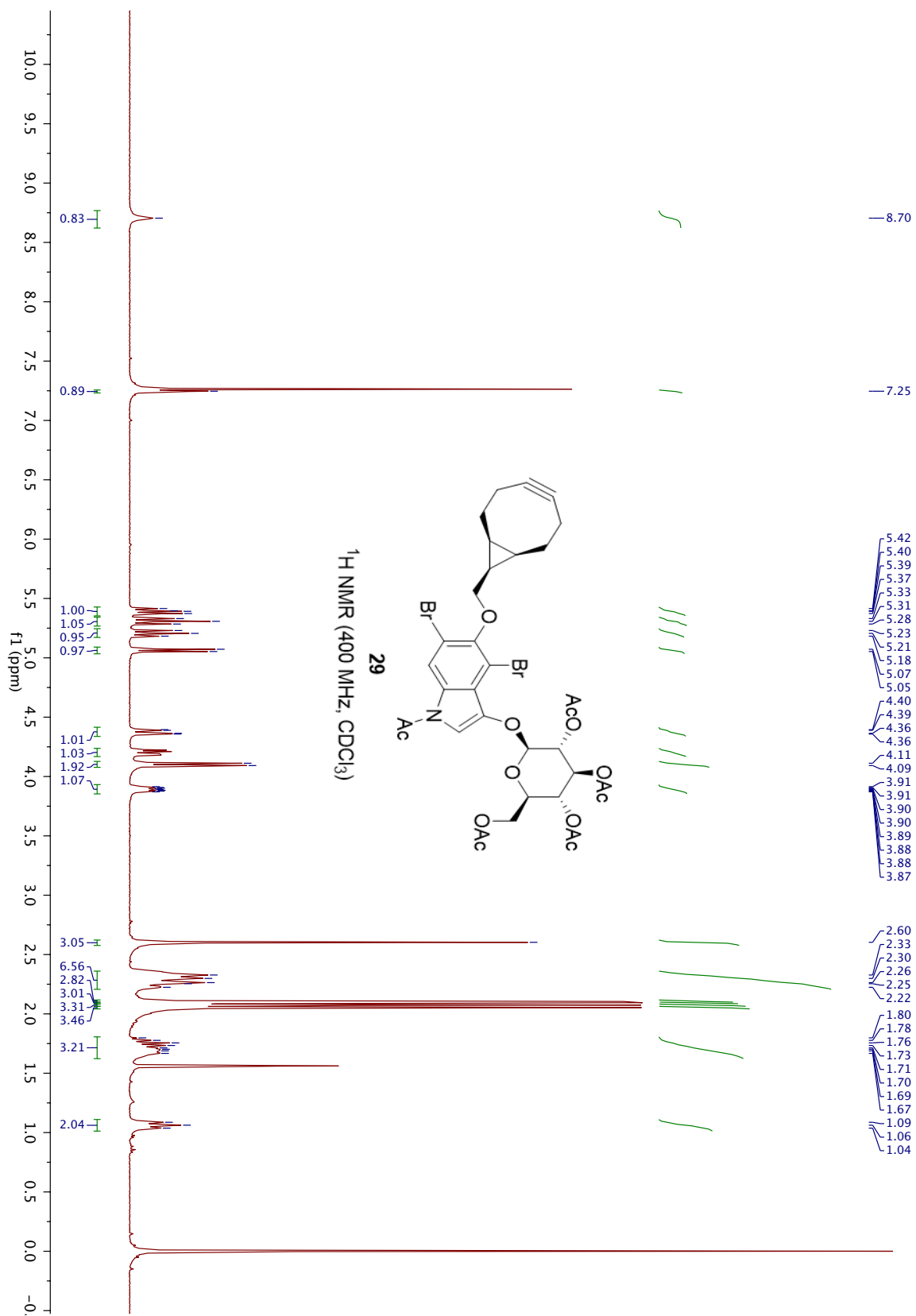


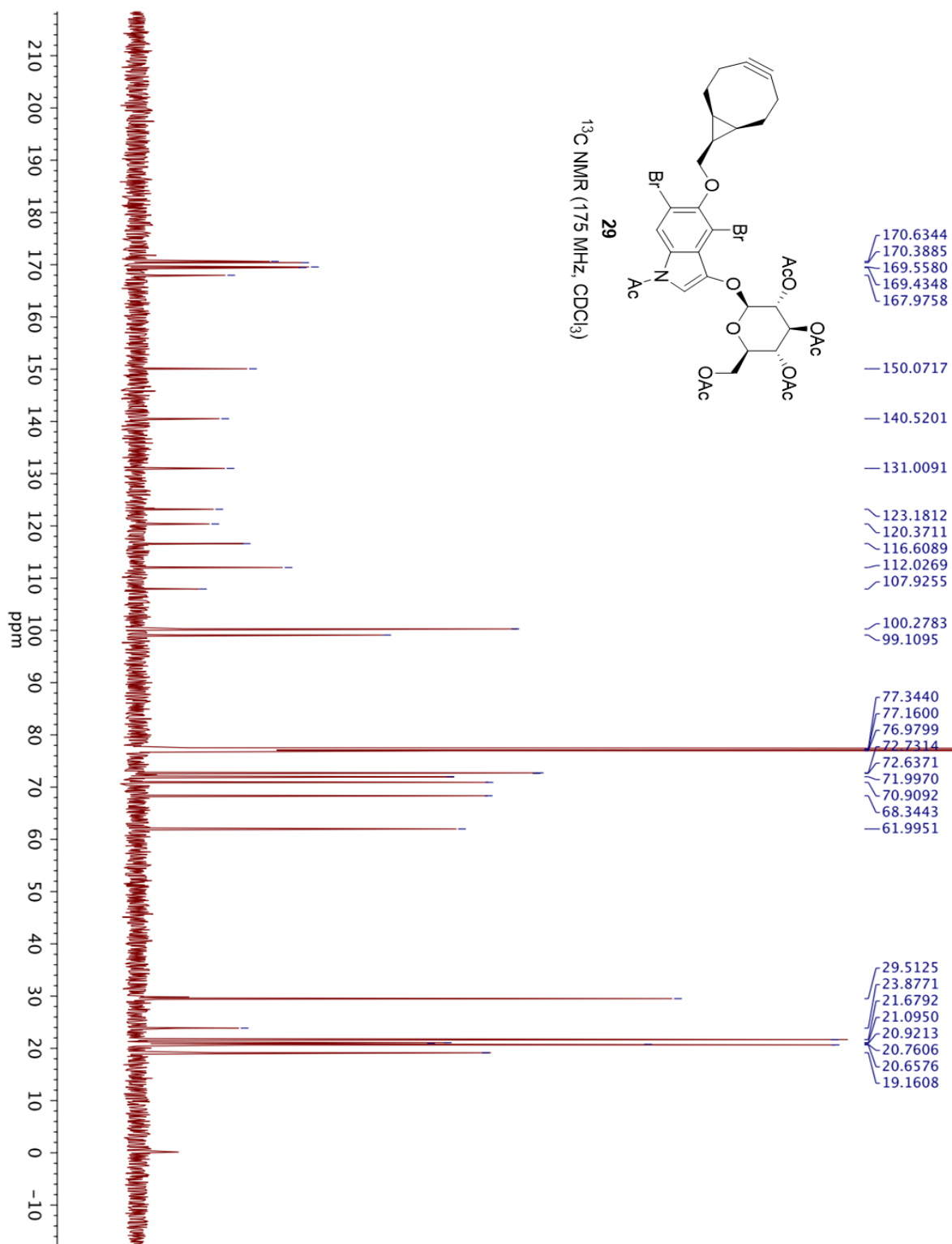


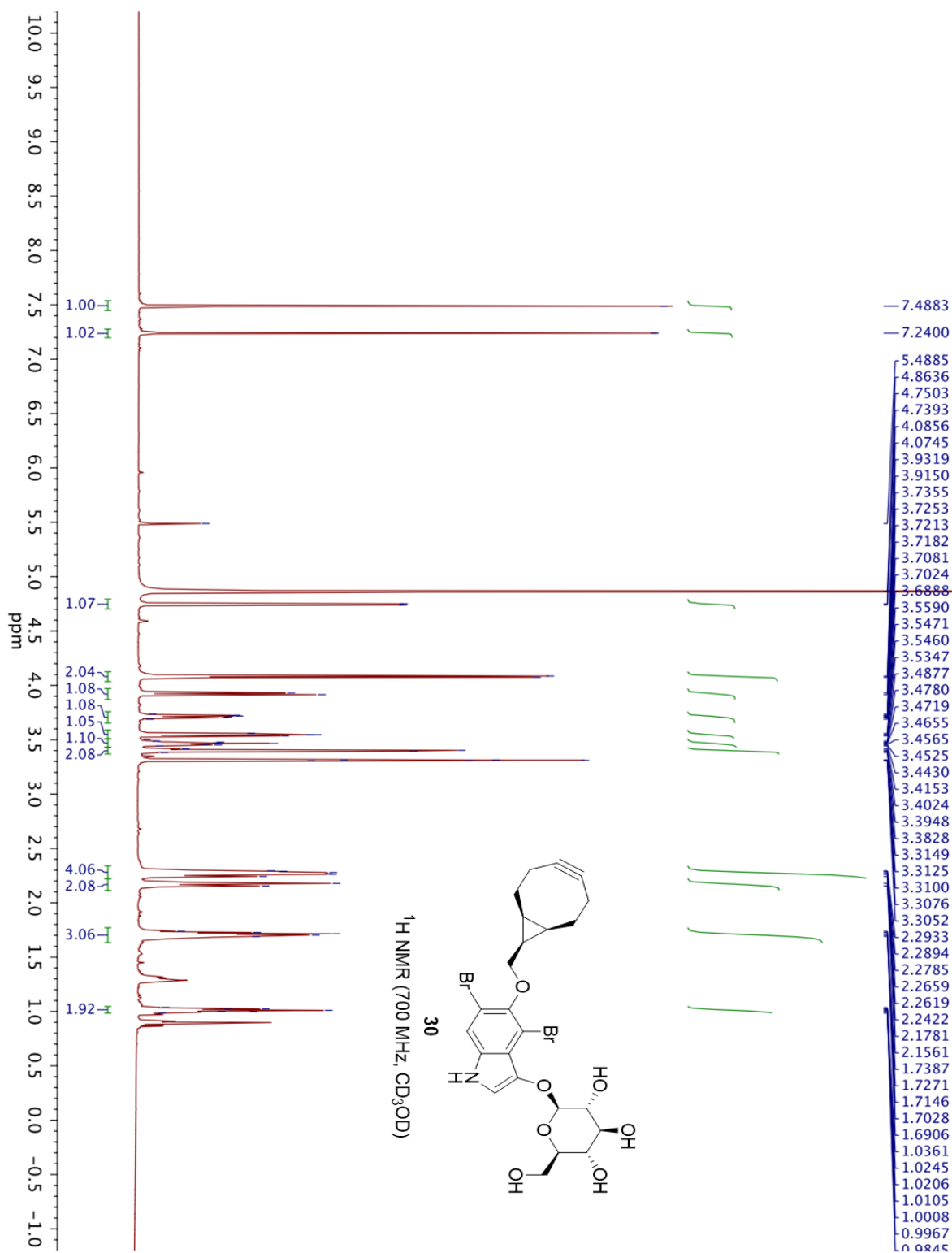


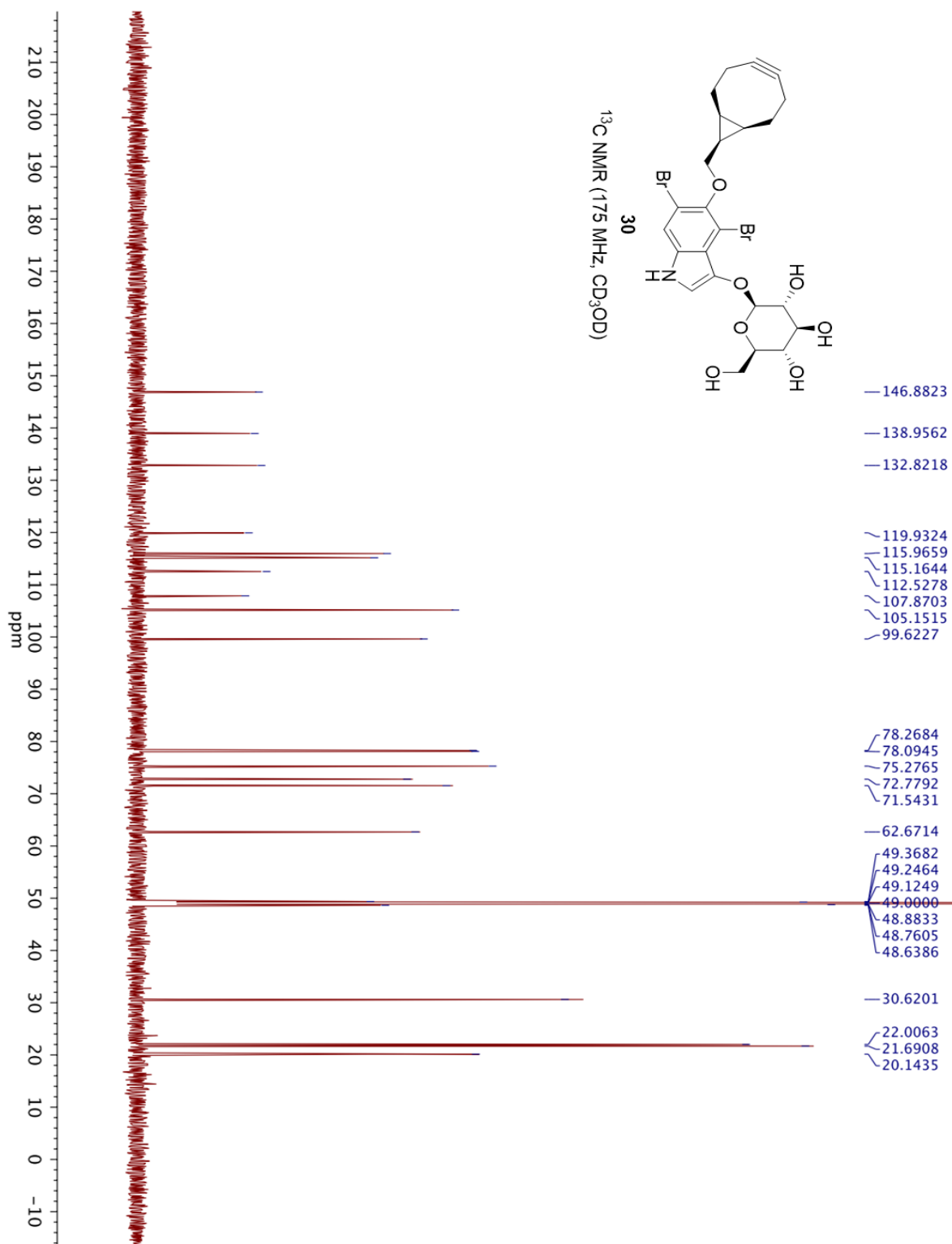


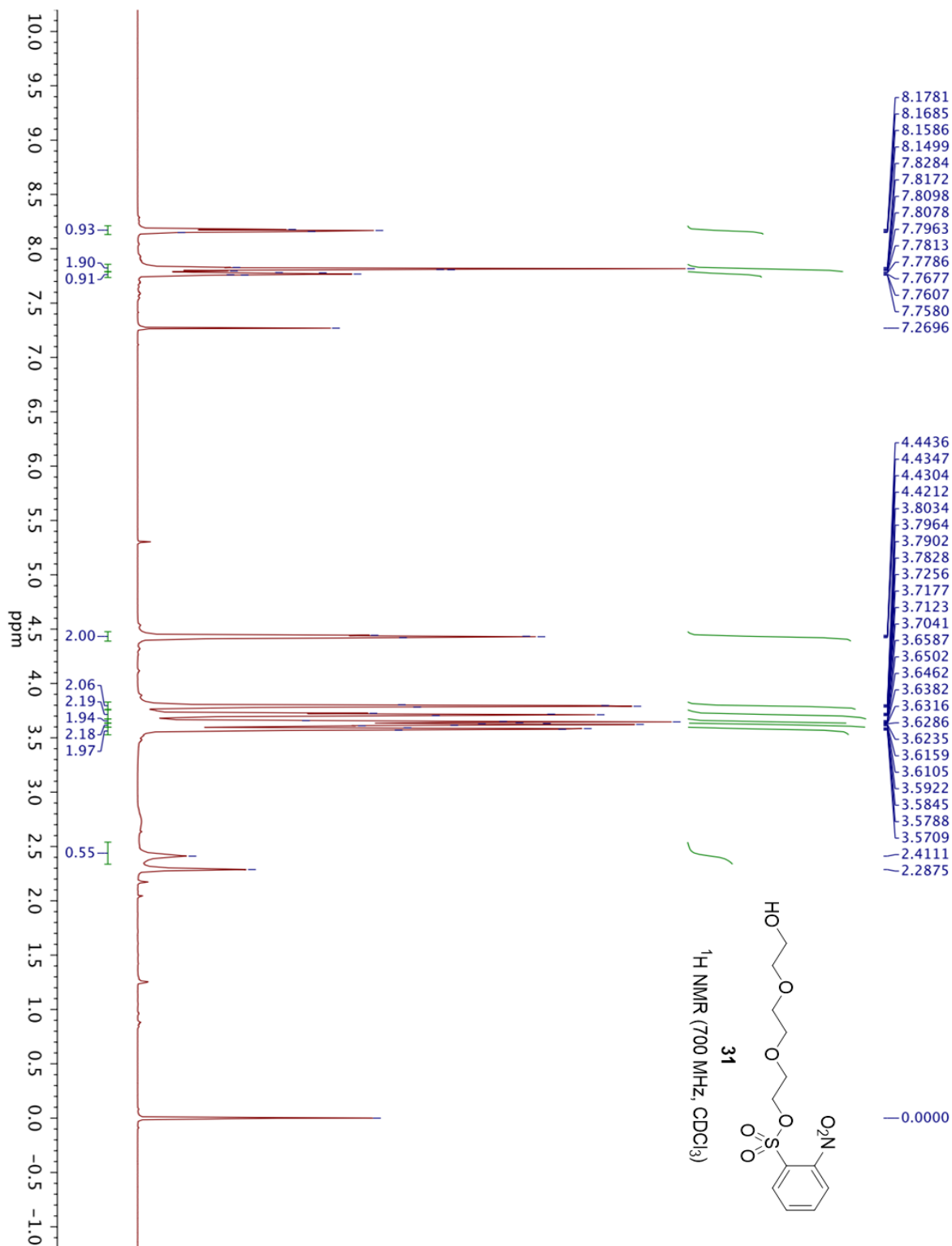


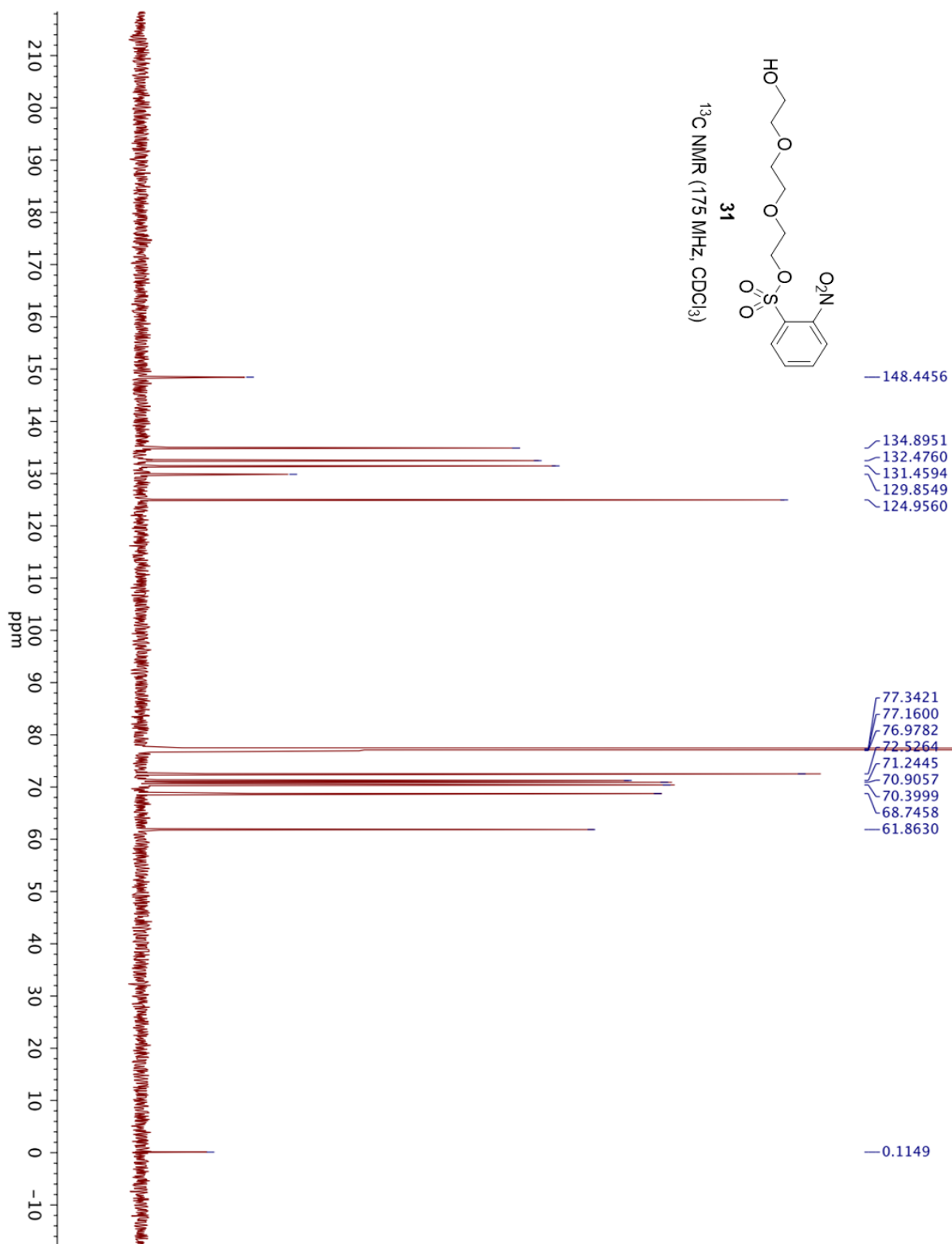


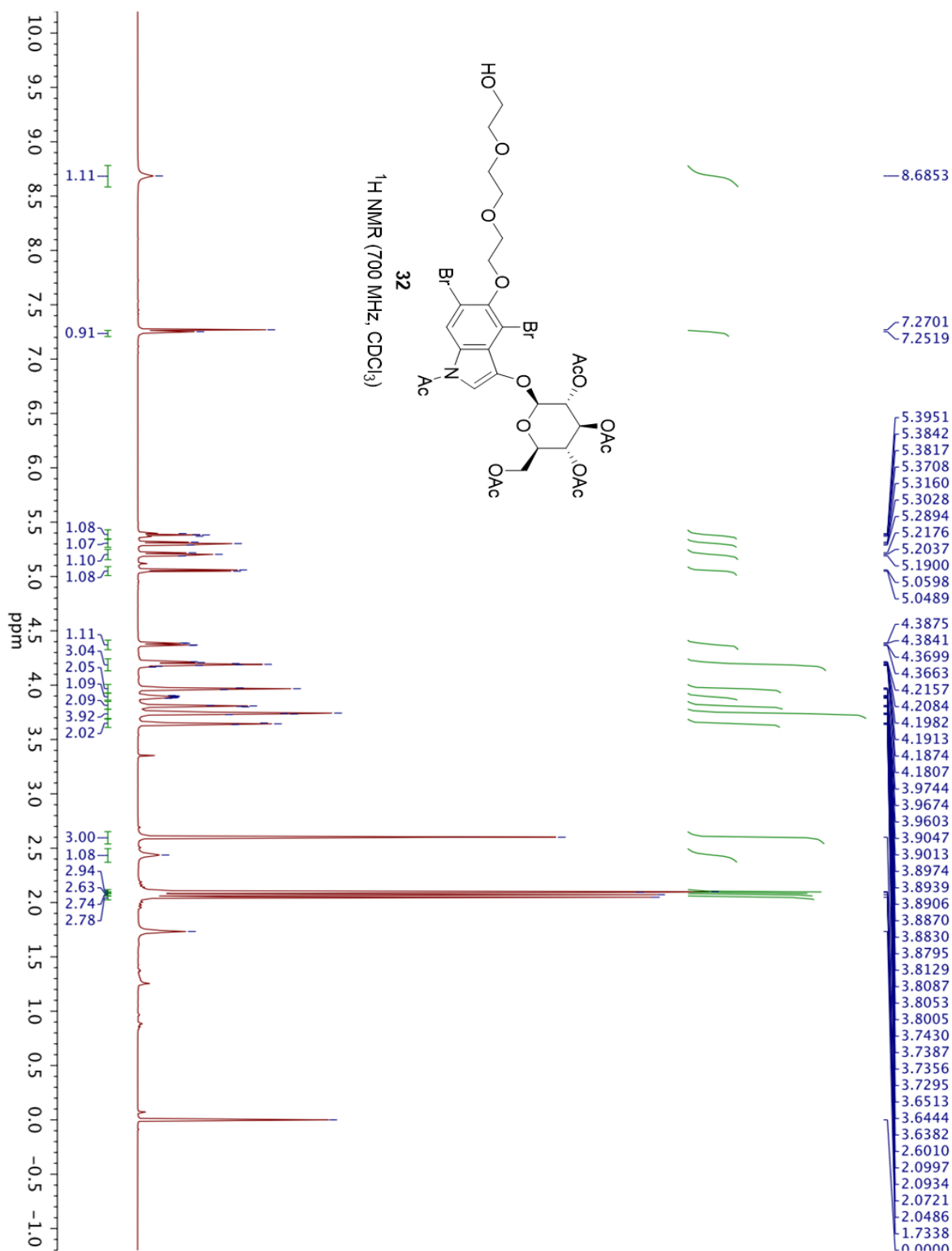


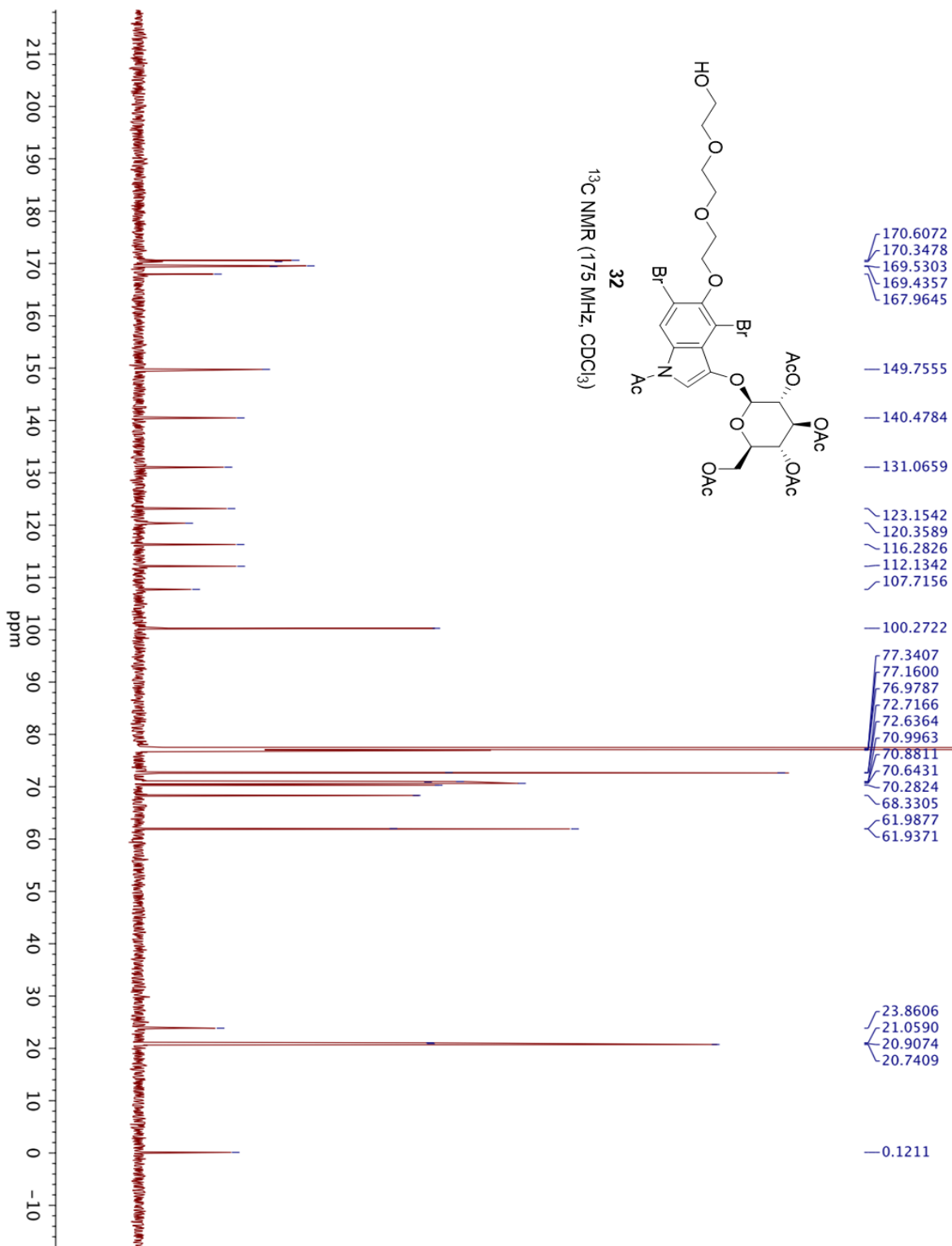


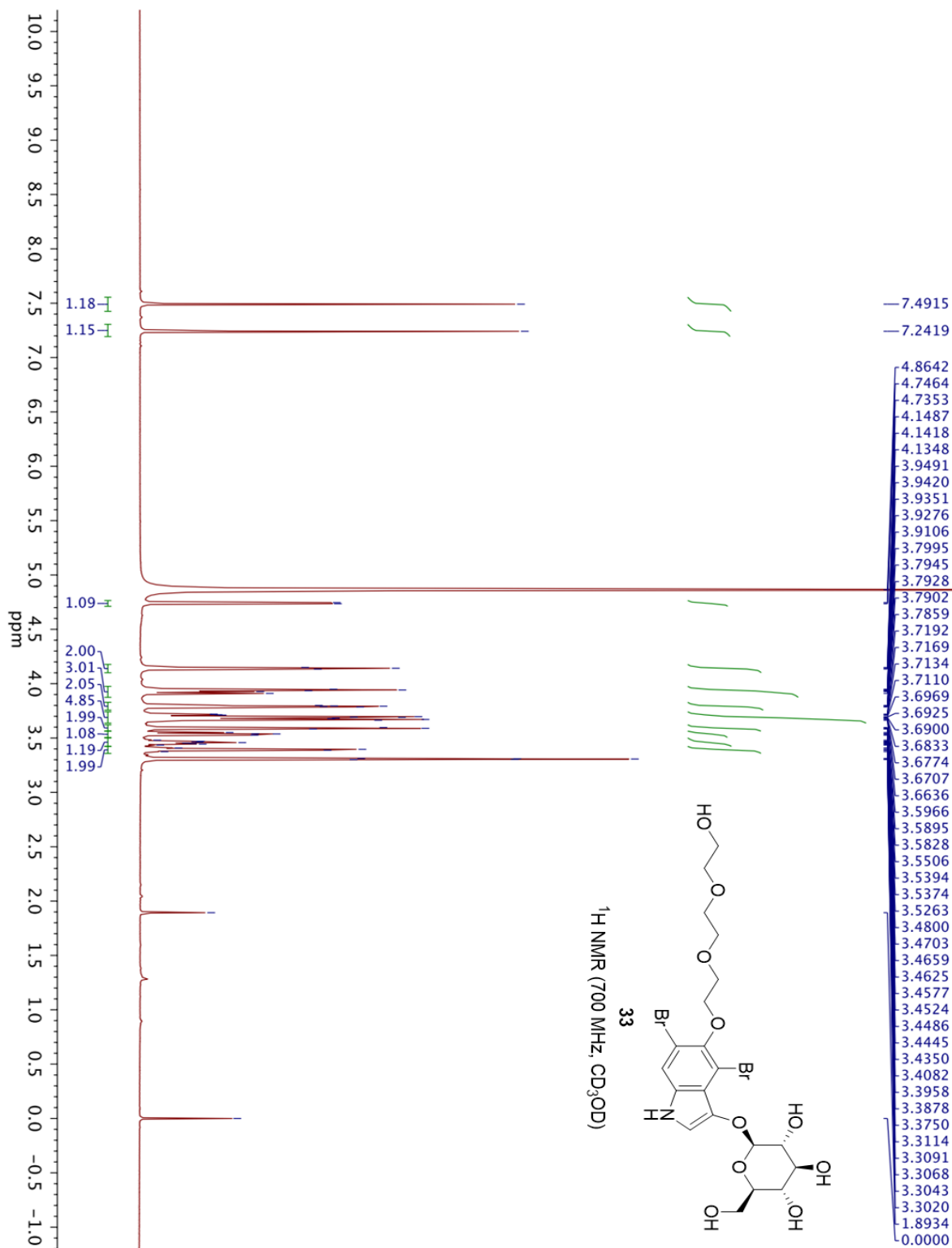


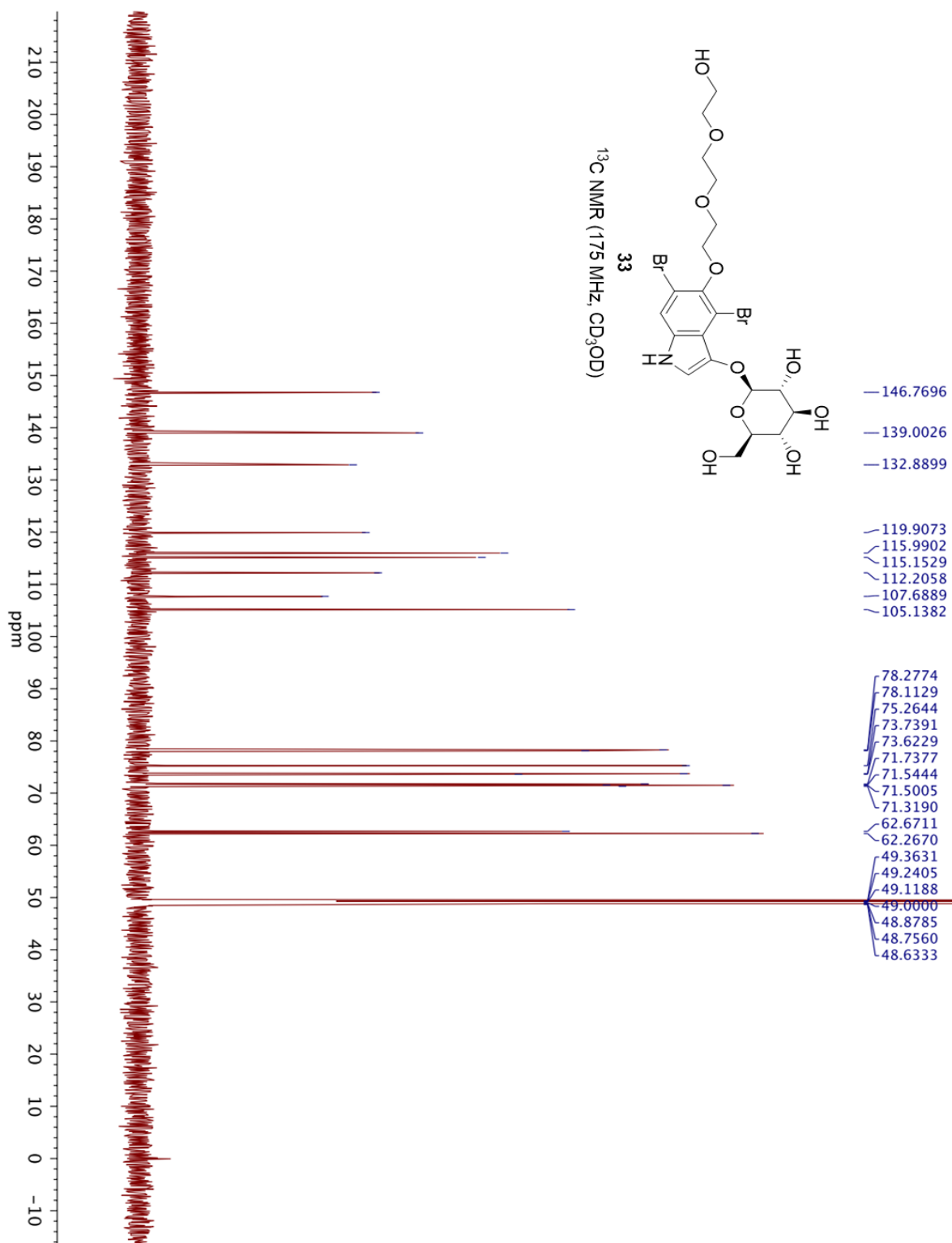


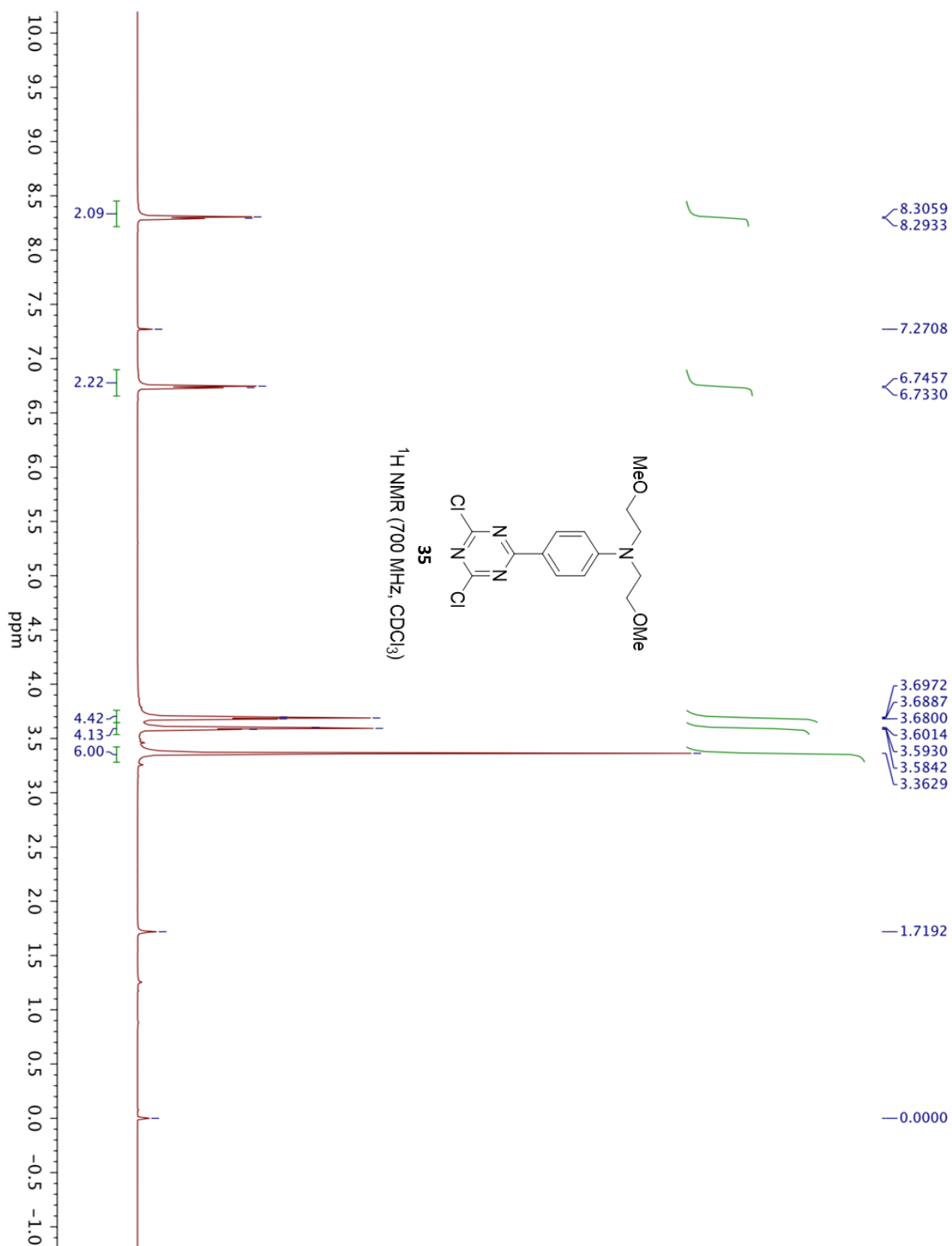


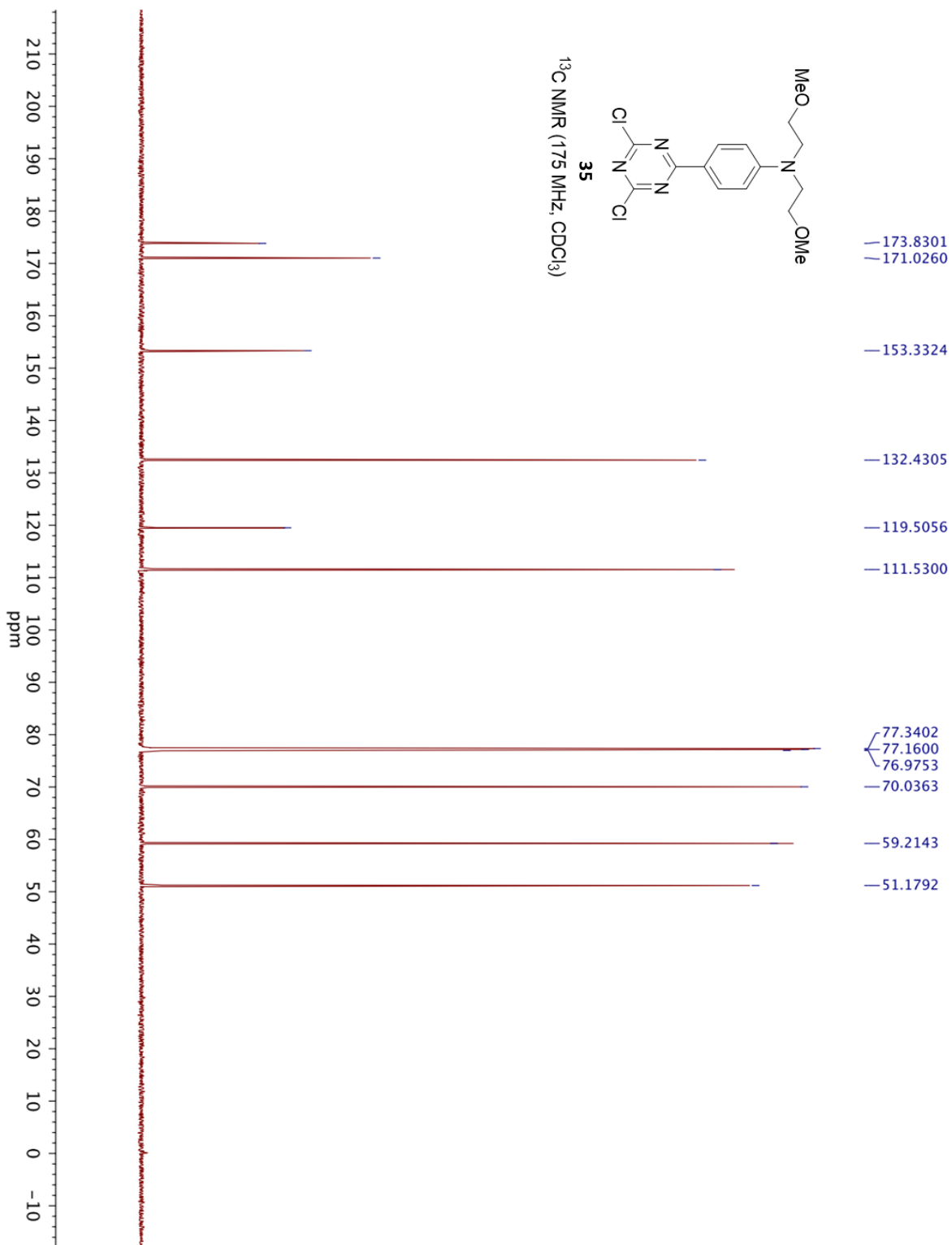


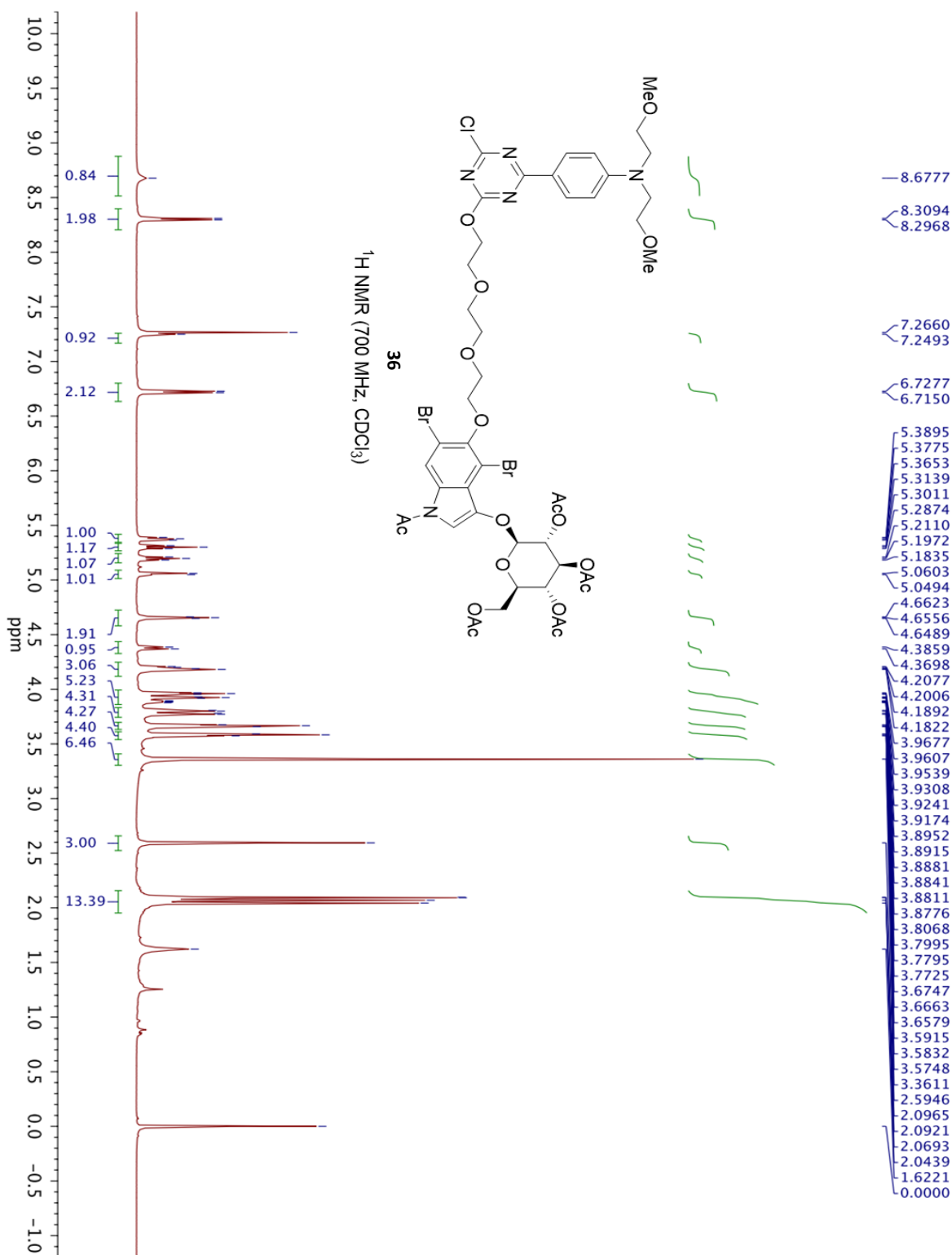


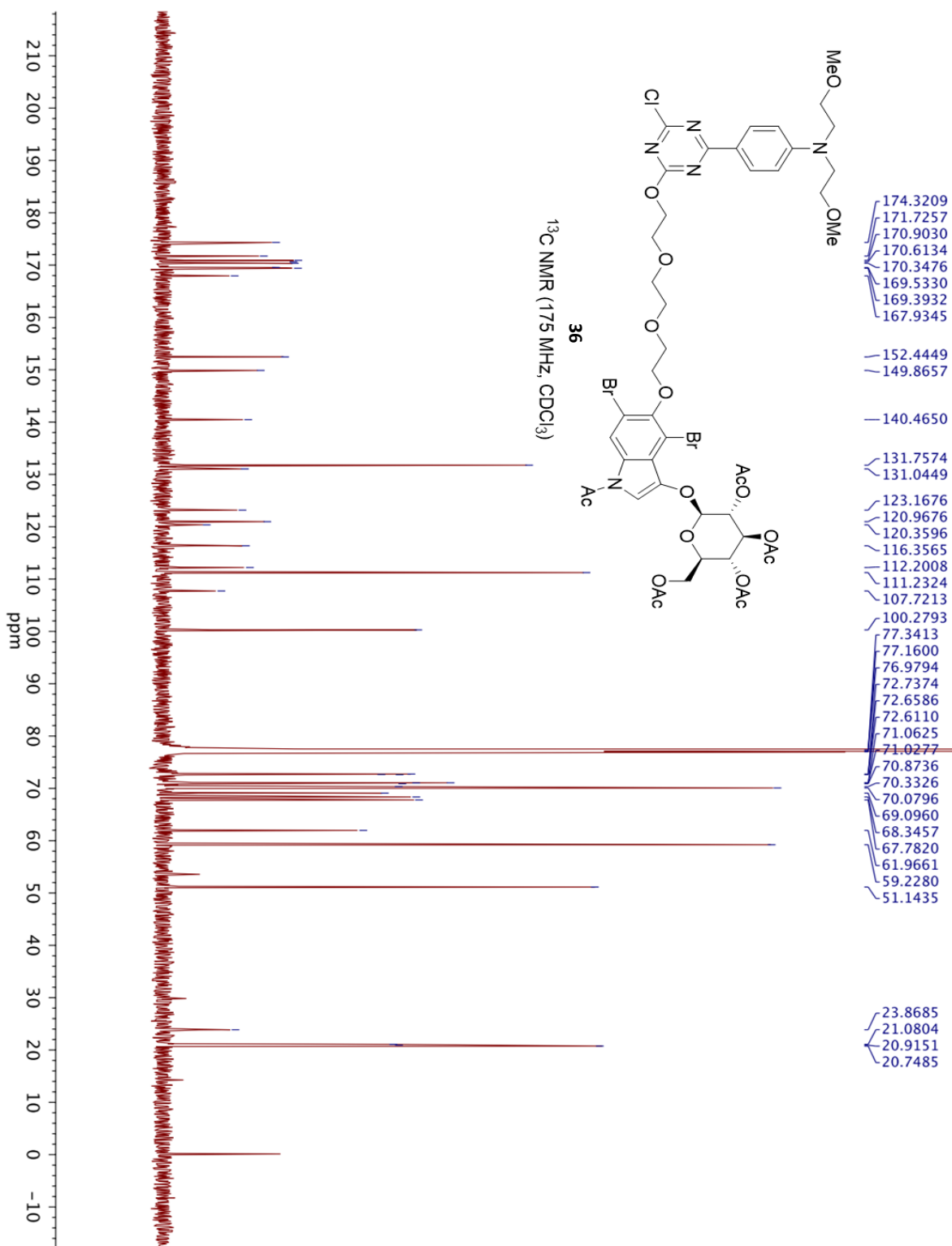


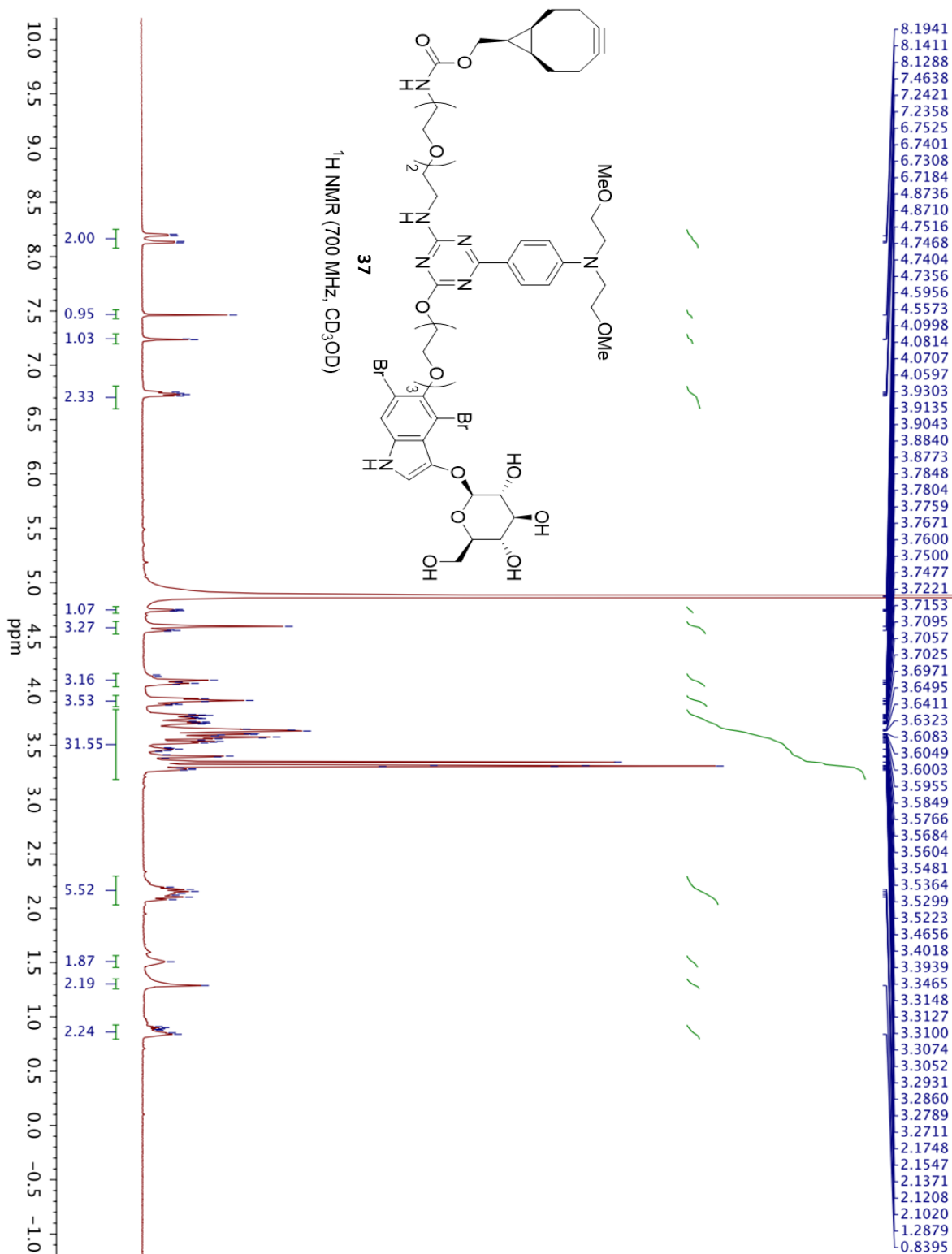


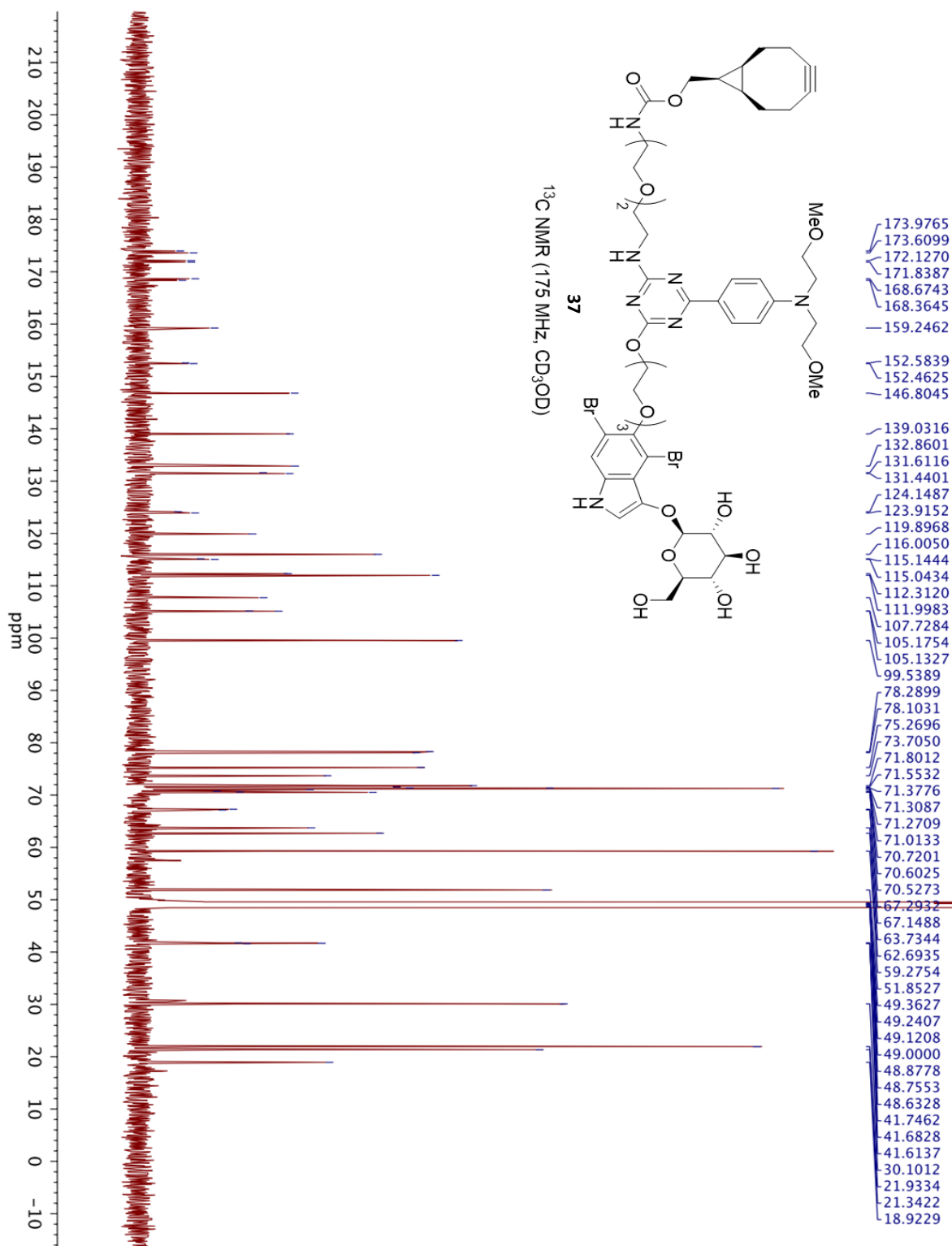


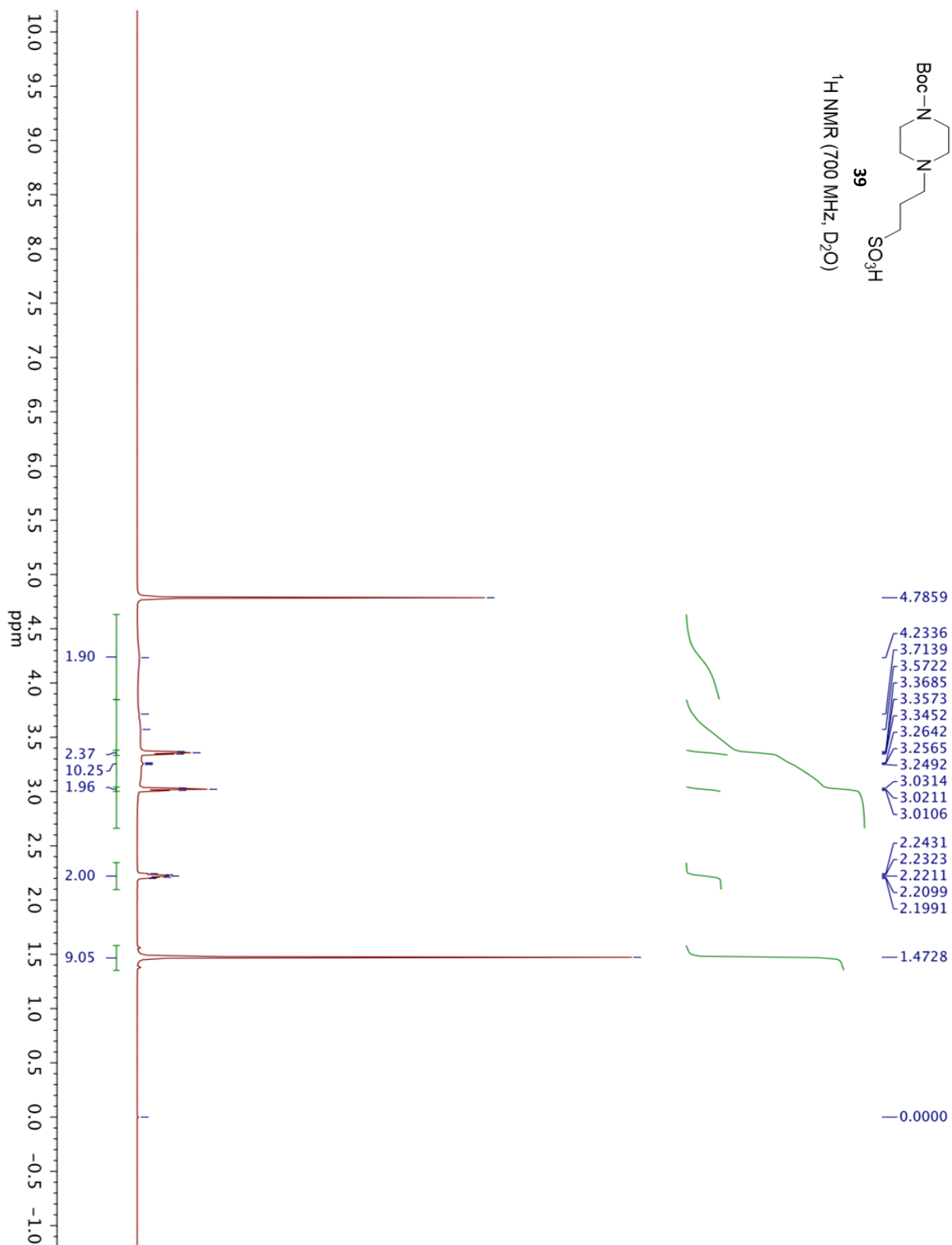
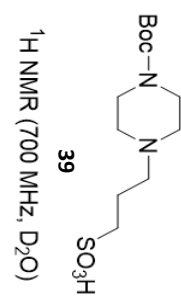


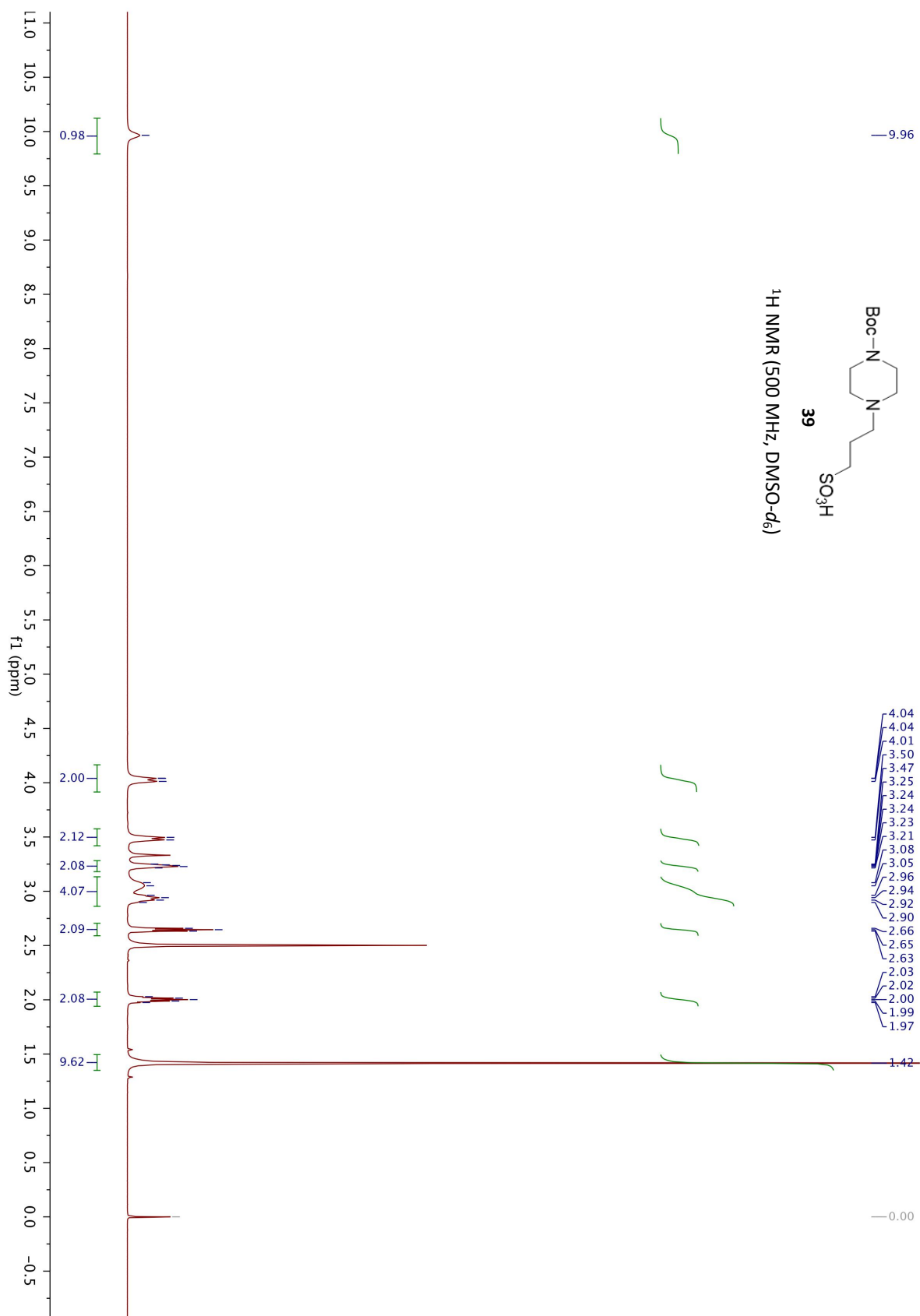


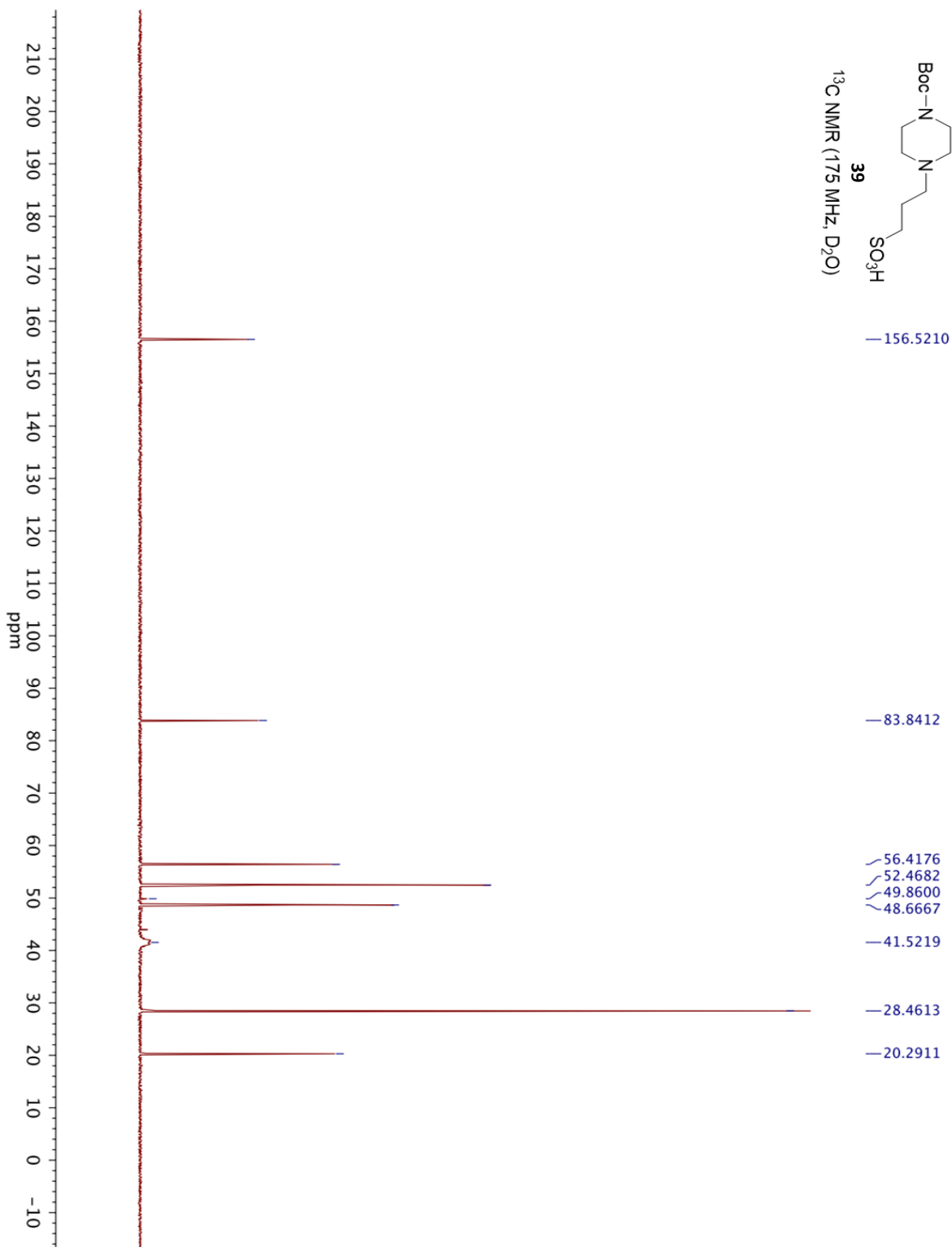
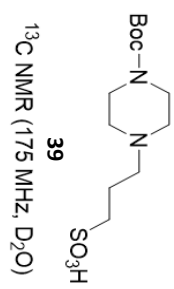


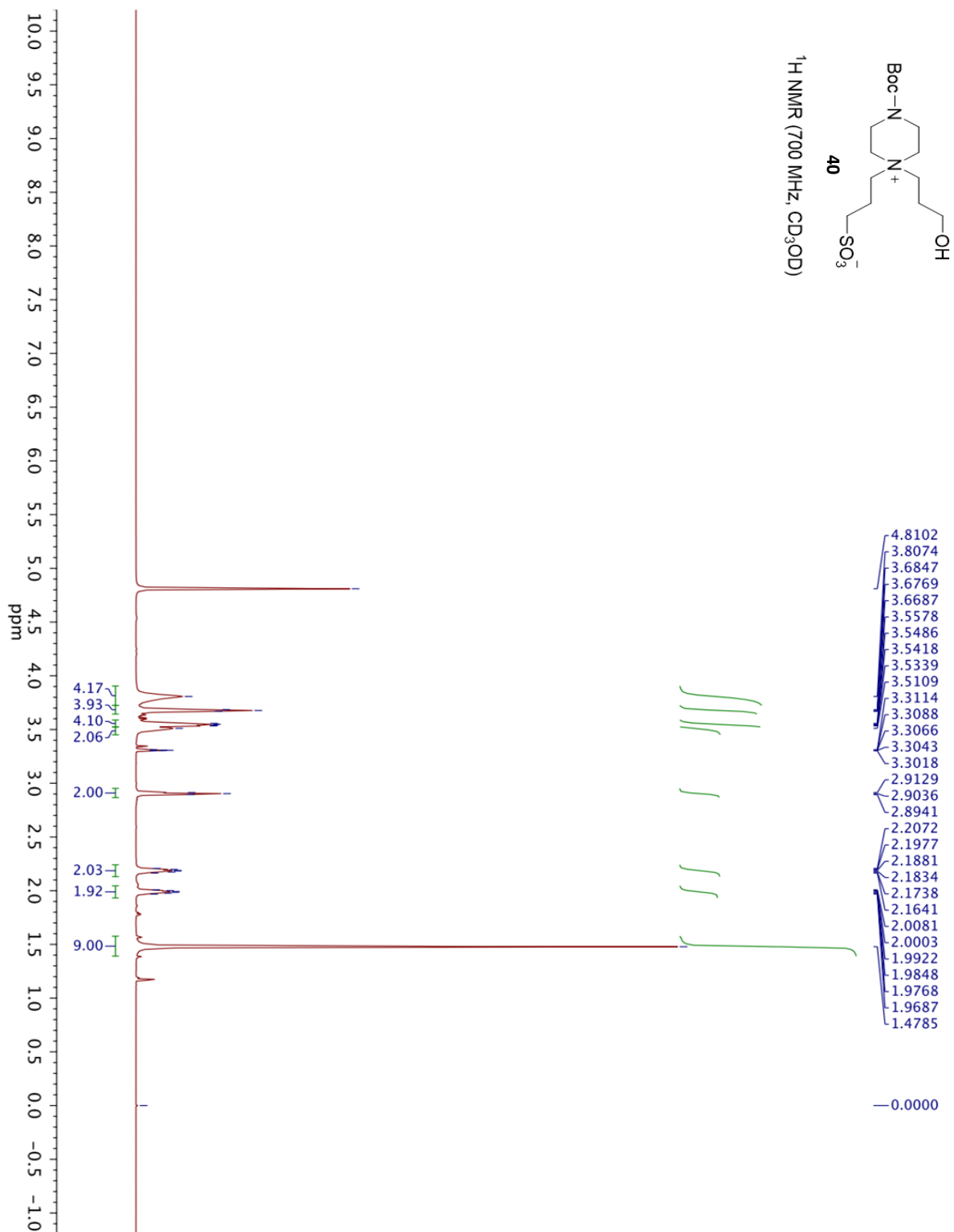


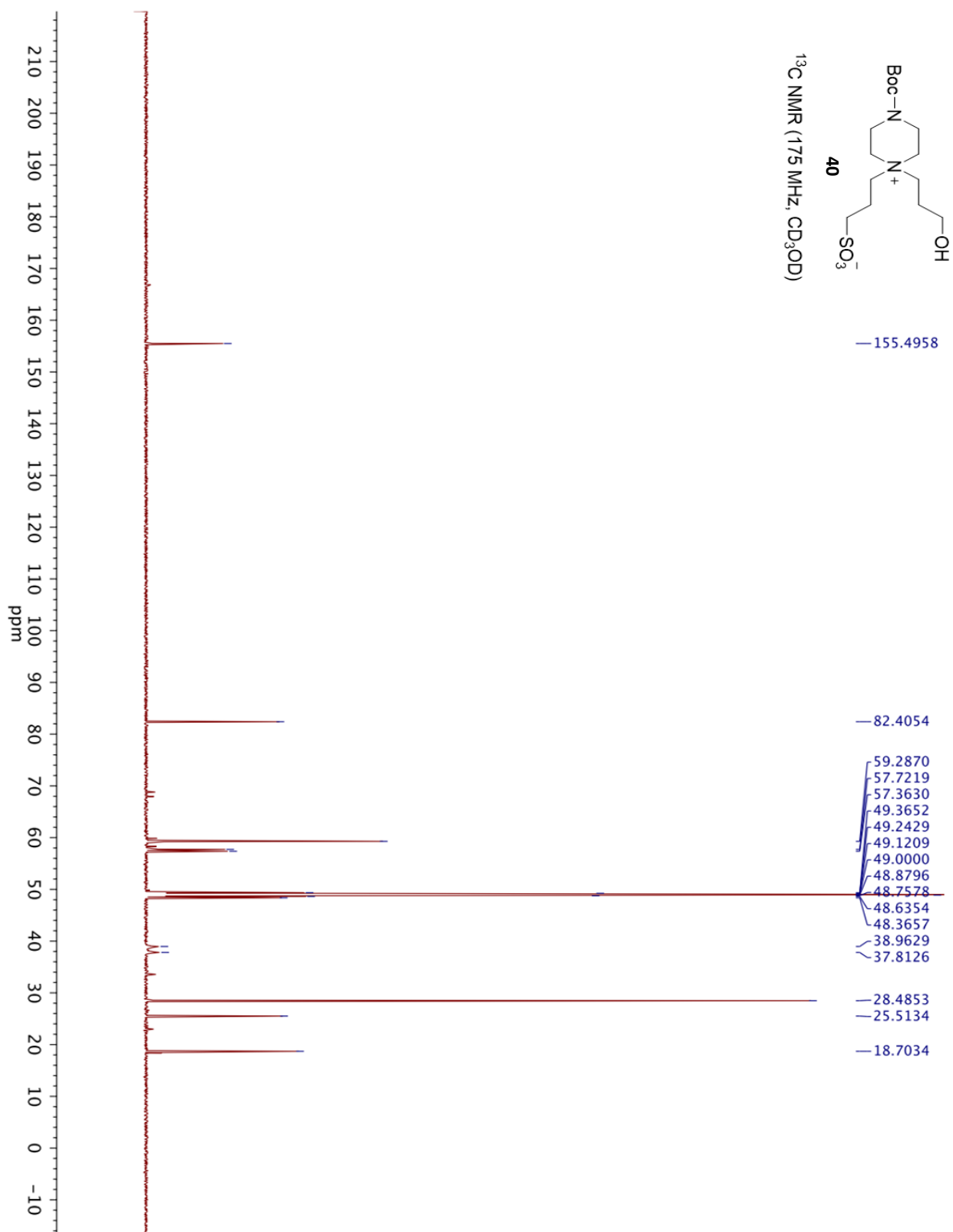


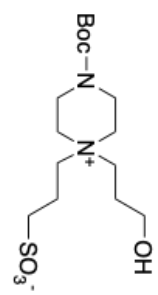












40
¹H NMR (700 MHz, DMSO-d₆)

