## Ratiometric fluorescent probe with aggregation-induced emission features for monitoring to HClO in living cell and zebra fish

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## 1. Experiment

**Cell culture.** HeLa cells were seeded in culture dishes in Dulbecco's modified Eagle's medium (DMEM) which was supplemented with 10% fetal bovine serum and 1% penicillin and cultured in a humidified incubator containing 5%  $CO_2$  and 95% air at 37 °C for 24 h. Before imaging, the cells were cultured in an 18 mm glass dish, during which dead cells and cell metabolites were washed away with Phosphate buffer saline (PBS) buffer.

**Cytotoxicity assay.** MTT assay was performed using HeLa cells, which were inoculated into 96-well plate and cultured in a cell culture tank. After the cell attachment was completed, different concentrations (0.0, 5.0, 10.0, 15.0, 20.0  $\mu$ M) of probe **PTZ-HCIO** were added into the 96-well plate and incubated in 5% CO<sub>2</sub> humidified incubator for 24 h. The MTT solution (1.0 mg/mL in PBS) was then added to each well and the cells were incubated in a cell culture tank for another 4 h. Finally, the MTT solution was dumped and DMSO (100.0  $\mu$ L) was added to each well. The absorbance was determined at 490 nm and the cell viability was calculated using the following formula: cell viability = (mean absorbance of test wells - mean absorbance of medium control wells) / (mean absorbance of untreated wells - mean absorbance of medium control wells) × 100%.

**Endogenous HCIO imaging in living HeLa cells.** Endogenous HCIO imaging in living HeLa cells using **PTZ-HCIO**. The first group: Cells were treated with LPS (1  $\mu$ g/mL) and PMA (1  $\mu$ g/mL) for 1 h, then 5  $\mu$ M PTZ-HCIO for additionally 15 min. The second group: Cells were treated with LPS (1  $\mu$ g/mL), PMA (1  $\mu$ g/mL) and ABH (200  $\mu$ M) for 1 h, then 5  $\mu$ M PTZ-HCIO for additionally 15 min. The third group: Cells were treated with LPS (1  $\mu$ g/mL), PMA (1  $\mu$ g/mL) and ABH (200  $\mu$ M) for 1 h, then 5  $\mu$ M PTZ-HCIO for additionally 15 min. The third group: Cells were treated with LPS (1  $\mu$ g/mL), PMA (1  $\mu$ g/mL) and NAC (1 mM) for 1 h, then 5  $\mu$ M PTZ-HCIO for additionally 15 min. Green channel: 515-580 nm (excited at 400 nm). Red channel: 615-750 nm (excited at 435-460 nm).

Fluorescence images of PTZ-HCIO in HeLa cells incubated with different concentrations of HCIO HeLa cells were incubated with PTZ-HCIO (5  $\mu$ M) at 37 °C for 30 min and then further treated with different concentrations of HCIO for 15 min. Green channel: 515-580 nm (excited at 400 nm). Red channel: 615-750 nm (excited at 435-460 nm).

The kinetics of oxidation of the probe PTZ-HClO. Fluorescence spectra of probe PTZ-HClO (5.0  $\mu$ M) in the presence of testing species (200  $\mu$ M). Then test the fluorescence intensity ratio (I<sub>535</sub>/I<sub>670</sub>) of probe PTZ-HClO (5.0  $\mu$ M) in the presence of other strongly oxidizing species. (1) HClO, (2) NO, (3) ONOO<sup>-</sup>, (4) ·OH, (5) <sup>1</sup>O<sub>2</sub>, (6) O<sup>2-</sup>, (7) ROO<sup>-</sup>.

## 2. Spectroscopic Property



**Fig. S1.** Absorption spectra of probe **PTZ-HCIO** (10.0  $\mu$ M) in the presence and absence of (A) HCIO in PBS buffer (pH 7.4, 10.0 mM, containing 20 % CH<sub>3</sub>CN).



**Fig. S2.** The changes of fluorescence intensity ratio ( $I_{535}/I_{670}$ ) of probe **PTZ-HClO** (5.0  $\mu$ M) in the presence and absence of HClO (200.0  $\mu$ M.) at different pH values.



Fig. S3. Time-dependent fluorescence intensity ratio of  $I_{535}/I_{670}$  changes of probe **PTZ-HClO** (5.0  $\mu$ M) with HClO (200.0  $\mu$ M).



Figure S4. The cytotoxicity assay of HeLa cells with different concentrations of PTZ-HCIO.



**Figure S5.** The kinetics of oxidation of the probe **PTZ-HCIO**. **Table. S1** Some fluorescent probes for HCIO.

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Probes	Types	Emission	Detection limit	Ref.
		wavelength /nm		
	Ratiometric	534, 626	15 nM	1
S-S-N	OFF-ON	450	0.76 μΜ	2
S S S S S S S S S S S S S S S S S S S	OFF-ON	588	15.6 nM	3
OHC S	OFF-ON	449.76 nM	513	4
NC_CN SN	OFF-ON	39 nM	613	5

Ratiometric	6.6 nM	524, 586	6
Ratiometric	23 nM	418, 520	7
Ratiometric	25 nM	535, 670	This work

1. W. Chen, G. Li, C. Chen, J. Sheng and L. Yang, *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 2020, **228**, 117724.

2. J.-Y. Wang, J. Qu, H. Zhang, K. Wei and S.-X. Ni, RSC Advances, 2019, 9, 16467-16471.

3. H. Feng, Q. Meng, Y. Wang, C. Duan, W. Cuiping, H. Jia, Z.-Q. Zhang and R. Zhang, *Chemistry - An Asian Journal*, 2018, **13**.

4. H. Song, Y. Zhou, C. Xu, X. Wang, J. Zhang, Y. Wang, X. Liu, M. Guo and X. Peng, *Dyes and Pigments*, 2019, **162**, 160-167.

5. D. Zheng, X. Qiu, C. Liu, X. Jiao, S. He, L. Zhao and X. Zeng, *New Journal of Chemistry*, 2018, **42**, 5135-5141.

6. L. He, Y. Zhang, H. Xiong, J. Wang, Y. Geng, B. Wang, Y. Wang, Z. Yang and X. Song, *Dyes and Pigments*, 2019, **166**, 390-394.

7. P. Hou, S. Chen, G. Liang, H. Li and H. Zhang, *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 2020, **229**, 117866.





Figure S7. <sup>13</sup>C NMR spectrum of PTZ-HCIO in CDCl<sub>3</sub>.



Figure S8. HR-MS spectrum of PTZ-HCIO.



Figure S9. HR-MS spectrum of the reaction product of probe PTZ-HClO with HClO.