

## Supporting Information

# Hybrid Vesicles as Intracellular Reactive Oxygen Species and Nitric Oxide Generators

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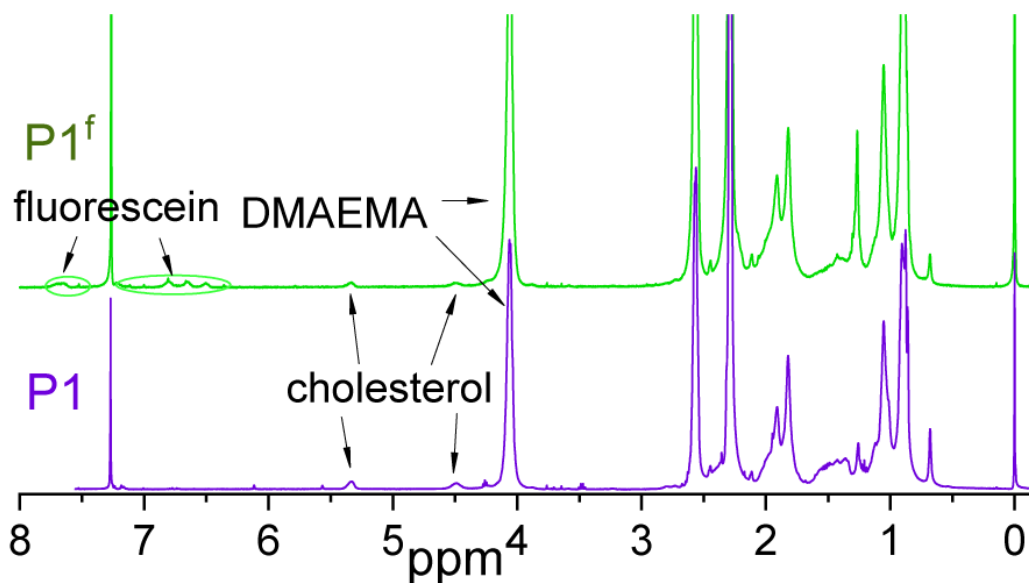
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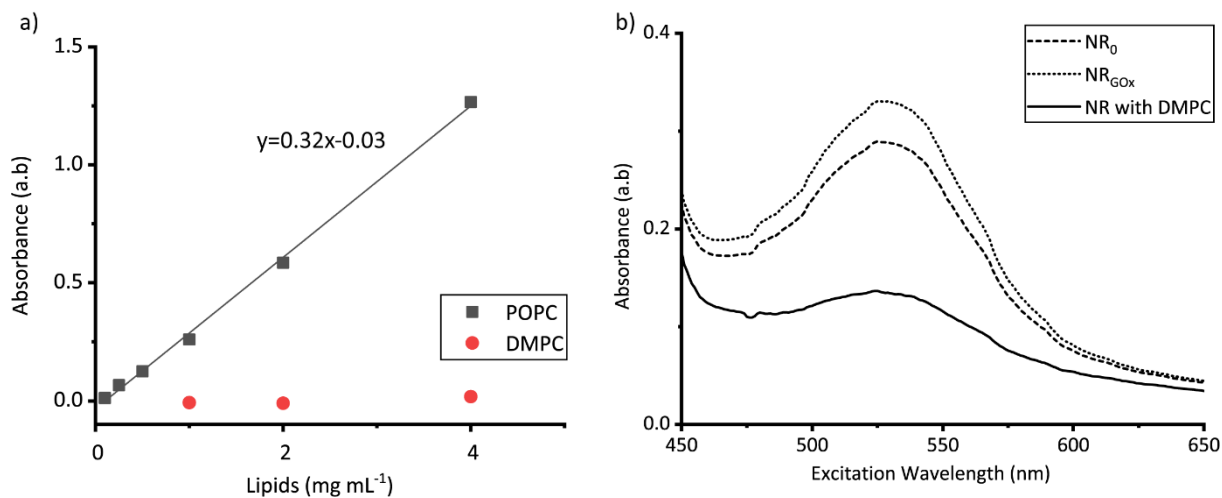
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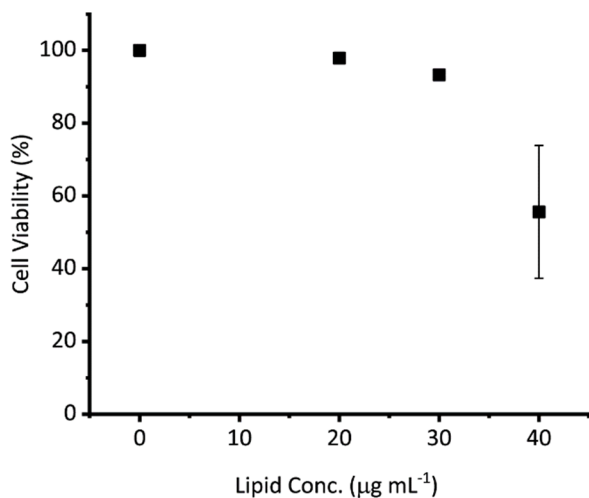
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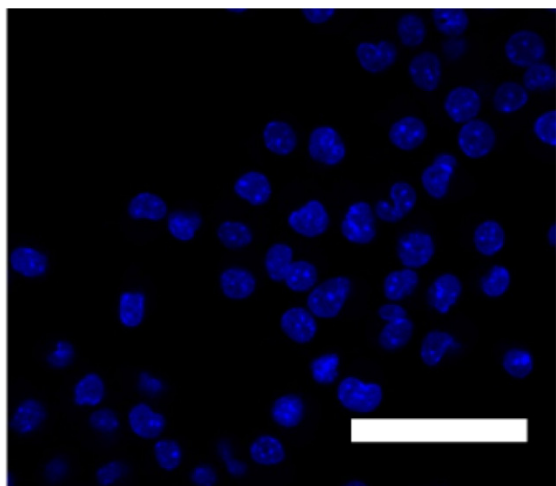
**Figure S1.** <sup>1</sup>H NMR (400 MHz) spectra of the block copolymers P1 and <sup>13</sup>C-P1 in CDCl<sub>3</sub>.



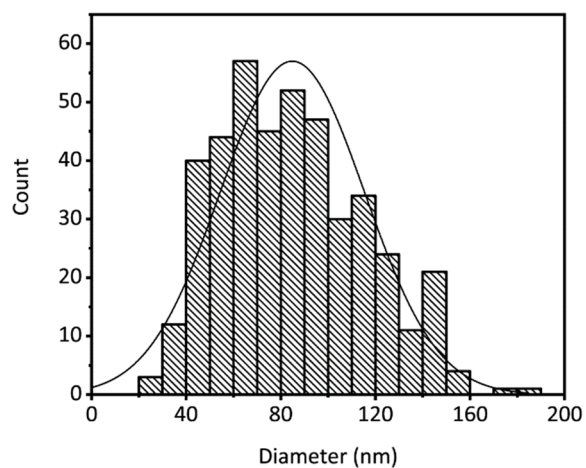
**Figure S2.** Lipid concentration determined by the sulfo-phospho-vanillin reaction. a) Typical concentration curve for (non-extruded) POPC liposomes including the linear fit used to determine unknown lipid amounts in samples, compared to (non-extruded) DMPC liposomes. b) Absorbance spectra of sulfo-phospho-vanillin reacted NR<sub>0</sub>, NR<sub>GOx</sub> and NR containing DMPC instead of POPC after size exclusion chromatography.



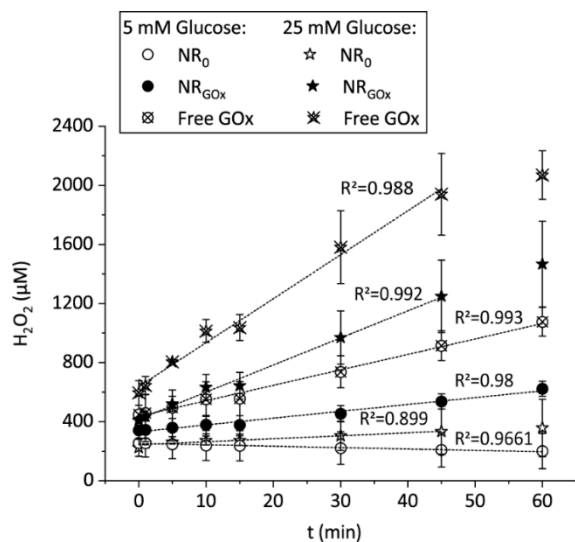
**Figure S3.** Cell viability of RAW 264.7 cells exposed to increasing amounts of NR<sub>0</sub> for 24 h.



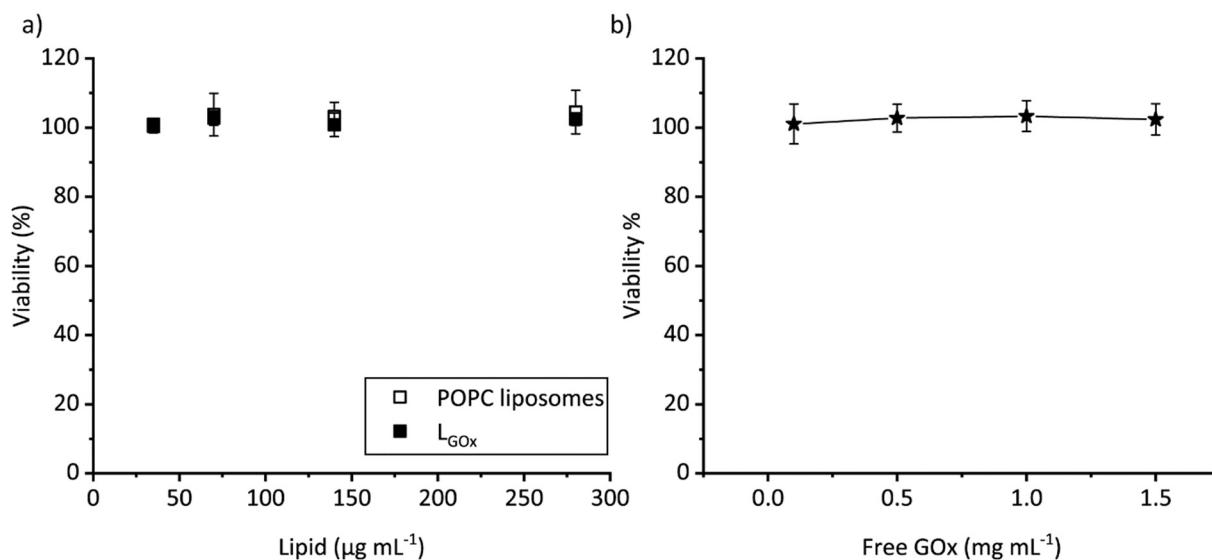
**Figure S4.** Representative CLSM image of fixed and stained RAW 264.7 cells, confirming the absence of red fluorescence when the cells were not exposed to  $^{\text{f}}\text{NR}_o$  (blue: Hoechst stained nuclei). The blue and red channels are overlaid in the same way as in Figure 1d. The scale bar is 50  $\mu\text{m}$ .



**Figure S5.** Size histogram of  $\text{NR}_{\text{GOx}}$  as determined from cryo-EM images using the ImageJ software. The histogram and mean value were calculated using Origin software.



**Figure S6.** Time-dependent  $H_2O_2$  production in low- (5 mM) and high- (25 mM) glucose media for  $NR_0$ ,  $NR_{GOx}$ , and free GOx. The dashed lines represent the linear fits for each curve.

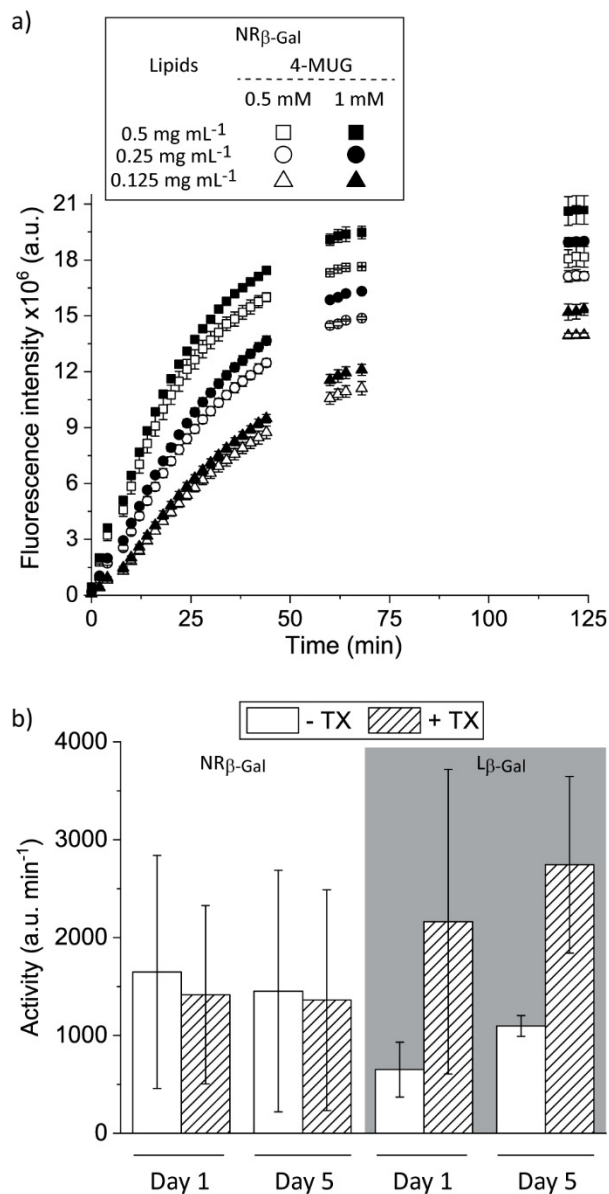


**Figure S7.** Viability of RAW 264.7 macrophages exposed to glucose-free media for 2 h to a) empty POPC liposomes and GOx-loaded POPC liposomes ( $L_{GOx}$ ) at concentrations similar to  $NR_{GOx}$ , and b) free GOx at concentrations up to 5× higher than the determined encapsulated amount of GOx in  $NR_{GOx}$ . Following on, the cells were incubated for 24 h in high-glucose media, and the viability was assessed using the CCK-8 kit. Data represent mean values  $\pm$  StD ( $n = 3$ ).

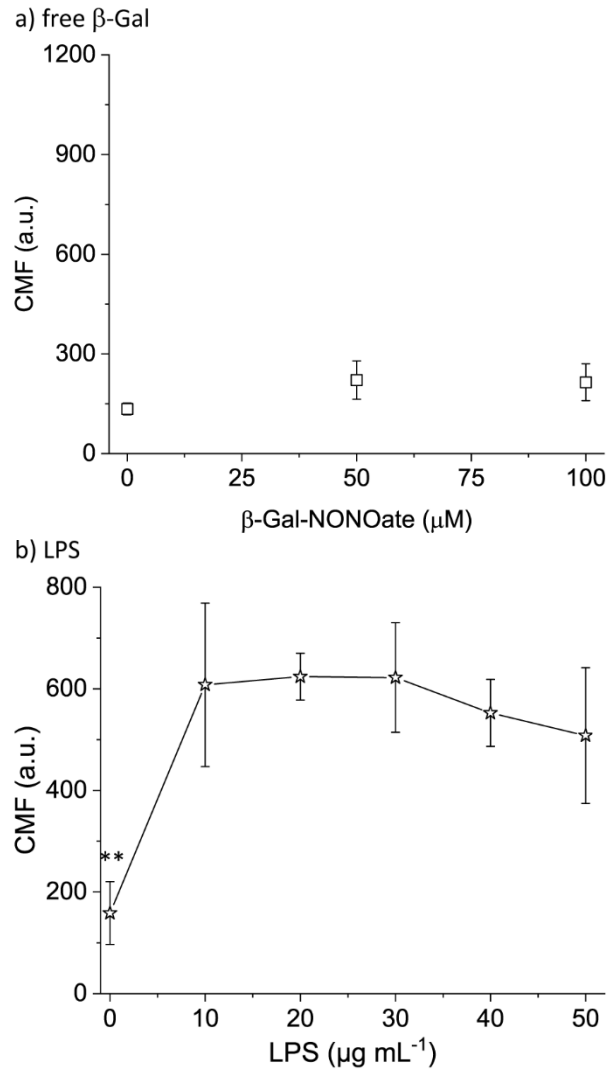
**Table S1. N<sub>o</sub>, NR<sub>GOx</sub> and NR<sub>β-Gal</sub> characterization**

Vesicle type	Size (nm) <sup>a)</sup>	PDI <sup>b)</sup>	LE (%) <sup>c)</sup>	Usage
NR <sub>o</sub>	206 ± 36	0.20	--	Dosing in RAW 264.7 macrophages
	437 ± 117	0.25	--	Controls for NR <sub>β-Gal</sub> in RAW 264.7 macrophages and primary macrophages Hybrid vesicle imaging Controls for the intracellular activity of NR <sub>GOx</sub> in RAW 264.7 macrophages
NR <sub>GOx</sub>	445±66	0.27	~35	All the experiments involving NR <sub>GOx</sub>
L <sub>GOx</sub>	225±8	0.09	NA	Liposome control experiments
NR <sub>β-Gal</sub>	240 ± 65	0.28	~6	All the experiments involving NR <sub>β-Gal</sub> beside imaging
	440 ± 97	0.27	N/A	Hybrid vesicle imaging
L <sub>β-Gal</sub>	218 ± 7	0.13	NA	Liposome control experiments

<sup>a)</sup>Hydrodynamic diameter assessed by DLS; <sup>b)</sup>Polydispersity index; <sup>c)</sup>Enzyme loading efficiency



**Figure S8. NR $\beta$ -Gal activity:** a) Conversion kinetics of 0.5 mM (open symbols) and 1 mM (closed symbols) 4-MUG into fluorescent 4-MUB by NR $\beta$ -Gal within 120 min. Three different NR $\beta$ -Gal concentrations were used. Data represent mean  $\pm$  StD. ( $n = 3$ ). b) Slopes obtained from the enzymatic reaction profiles of NR $\beta$ -Gal and L $\beta$ -Gal as prepared (day 1) and after 5 days incubation at 37  $^{\circ}$ C using 0.5 mM 4-MUG in the presence or absence of Triton X (TX 0.1 wt%). L $\beta$ -Gal represents  $\beta$ -Gal loaded POPC liposomes.



**Figure S9.** a) CMF of RAW 264.7 cells exposed to  $\beta$ -Gal in solution followed by incubation with different concentrations of  $\beta$ -Gal-NONOate. b) CMF of RAW 264.7 cells stimulated with increasing amounts of LPS for 19 h. In a) and b), the intracellular NO was determined using DAF-FM and flow cytometry as the read-out method. (n = 3, \*\*p < 0.01)