Supporting Information

A Trustworthy CpG Nanoplatform for Highly Safe and Efficient Cancer Photothermal Combined Immunotherapy

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Fig. S1 TEM images of Pd(5) (a) and Pd(30) (b). (c) TEM image of negative staining of Pd(30)-CpG by using uranyl acetate. The scale bar in a, b and c was 50 nm. The hydrodynamic sizes of Pd(5) and Pd(5)-CpG (d), and Pd(30) and Pd(30)-CpG (e), respectively. (f) Raman spectra of CpG, PVP, Pd NSs and Pd-CpG in the range of 200 to 450 cm⁻¹, laser power (632 nm, 0.66 mW).



Fig. S2 HAADF-STEM-EDX mapping images of Pd-CpG.



Fig. S3 (a, b) Typical fluorescence spectra of FAM-labeled CpG, Pd NSs modified with FAM-labeled CpG and the unloaded FAM-labeled CpG. (c) Standard working curve of FAM-labeled CpG relative to fluorescence intensity. The loading stability of Pd(5)-CpG (d) and Pd(30)-CpG (e) in different media (n=3 for each test). (f, g) UV–Vis–NIR absorption spectra of Pd-CpG, Pd NSs and CpG.



Fig. S4 The *in vitro* IR thermal images of PBS, CpG, Pd(30), Pd(30)-CpG, Pd(5) and Pd(5)-CpG upon 808 nm laser irradiation at different time intervals. The power density of 808 nm laser was 0.15 W cm^{-2} and the concentration of Pd was 25 ppm.



Fig. S5 Routine analysis of blood of mice treated with Pd(5) and Pd(30) after 7 days (n=3 for each group). Mean values and error bars are defined as mean and S.D., respectively.



Fig. S6 The biodistributions of Pd (% injected dose (ID) of Pd per gram of tissue) in main tissues at 7 days after intravenous administration of Pd(5) (a) and Pd(30) (b) (n=3 for each group). Mean values and error bars are defined as mean and S.D., respectively.



Fig. S7 Flow cytometry analysis of CD3⁺ T cells, CD4SP (CD3⁺ CD4⁺ CD8⁻)/CD8SP (CD3⁺ CD4⁻ CD8⁺), CD4⁺/CD8⁺ Naïve T cells (CD44⁻ CD62L⁺), Memory T cells (CD44⁺ CD62L⁺), Effector T cells (CD44⁺ CD62L⁻) from the spleen of mice (n=3 for each group).



Fig. S8 Flow cytometry analysis of CD3⁺ T cells, CD4SP (CD3⁺ CD4⁺ CD8⁻)/CD8SP (CD3⁺ CD4⁻ CD8⁺), CD4⁺/CD8⁺ Naïve T cells (CD44⁻ CD62L⁺), Memory T cells (CD44⁺ CD62L⁺), Effector T cells (CD44⁺ CD62L⁻) from the lymph node of mice (n=3 for each group).



Fig. S9 Flow cytometry analysis of the maturation B cells from lymph nodes, blood and spleen of mice (n=3 for each group).



Fig. S10 Flow cytometry analysis of macrophages and neutrophils from the spleen of mice (n=3 for each group).



Fig. S11 Flow cytometry analysis of macrophages and neutrophils from the bone marrow of mice (n=3 for each group).



Fig. S12 Flow cytometry analysis of dendritic cell (DC) maturation after various treatments. The DC cells were stained with anti-CD80 and anti-CD86 (n=3 for each group). *p<0.05.



Fig. S13 Comparison of the TNF- α (a) and IL-6 (b) releases in DCs culture stimulated by PBS, CpG(PS), Pd(5) and Pd(5)–CpG(PS) without or with NIR irradiation (808 nm, 0.15 W cm⁻², 5 min), respectively.



Fig.S14 The activation of CD8⁺ T cells stimulated by matured DCs. DCs were first incubated with supernatant from Pd(5)-CpG(PS)+NIR treated B16F10-OVA (or other controls). BMDCs were then subjected to an *in vitro* presentation assay using CFSE-labeled CD8⁺ T cells (n=4 for each group).



Fig. S15 Pharmacokinetics studies of Pd(5) and Pd(5)-CpG(PS). The blood circulation curves of intravenously injected Pd(5) (a) and Pd(5)-CpG(PS) (b) (n = 5). The half-lives (t1/2) for Pd(5) and Pd(5)-CpG(PS) were calculated to be ~21 h and ~24 h, respectively.



Fig. S16 The biodistributions of Pd (% injected dose (ID) of Pd per gram of tissue) in main tissues and tumors in 1, 2, 4 and 6 days after intravenous administration of Pd(5) (a) and Pd(5)-CpG(PS) (b). Mean values and error bars are defined as mean and S.D., respectively (n=5).



Fig. S17 Urine and feces excretion of Pd(5) and Pd(5)-CpG(PS) (n=3).



Fig. S18 The temperature increase profiles of tumor tissues in mice during laser irradiation. The power density of 808 nm laser was 0.15 W cm⁻².



Fig. S19 Optical photos of mice before and after treatment at different treated groups.



Fig. S20 PTT effect of Pd(5)-CpG on RAW 264.7 cells. The laser power density of 808 nm was 0.15 W cm⁻².



Fig. S21 The matured APCs (CD11c⁺MHC II⁺) both in percentage (b) and number (c) in tumor after different treatments (n=4 for each group).



Fig. S22 CD11c⁺ IL12p40⁺ cells (a and b), CD3⁺ CD8⁺ cells (c and d) and IFN γ CD8⁺ cells (e and f) from the spleen of mice (n=4 for each group).



Fig. S23 CD11c⁺ IL12p40⁺ cells (a and b), CD3⁺ CD8⁺ cells (c and d) and IFN γ CD8⁺ cells (e and f) from the blood of mice (n=4 for each group).



Fig. S24 Flow cytometry analysis of macrophage (a and b) from the spleen of mice (n=4 for each group).



Fig. S25 Concentrations of TNF- α (a) and IL-6 (b) in the sera of different groups treated mice (Control, Pd(5)+NIR and Pd(5)-CpG(PS)+NIR).



Fig. S26 Flow cytometry analysis of MDSCs (CD11b⁺Gr-1⁺), M2 macrophages (Arg1⁺F4/80⁺) and Treg cells (CD4⁺FOXP3⁺) in tumor after Pd(5)+NIR and Pd(5)-CpG(PS)+NIR treatment (n=4 for each group).