

Supporting Information for

Near-Infrared Light Controlled Fluorogenic Labeling of  
Glycoengineered Sialic Acids *in vivo* with Upconverting Photoclick  
Nanoprobe

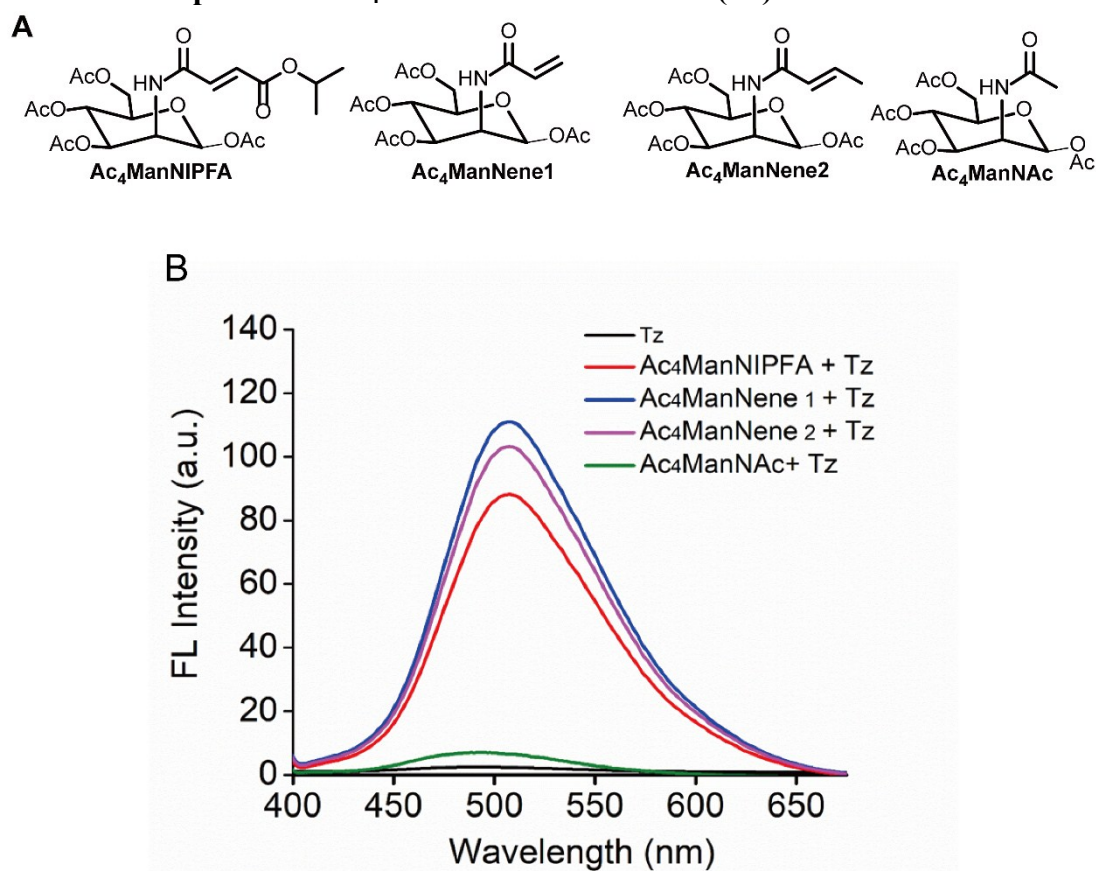
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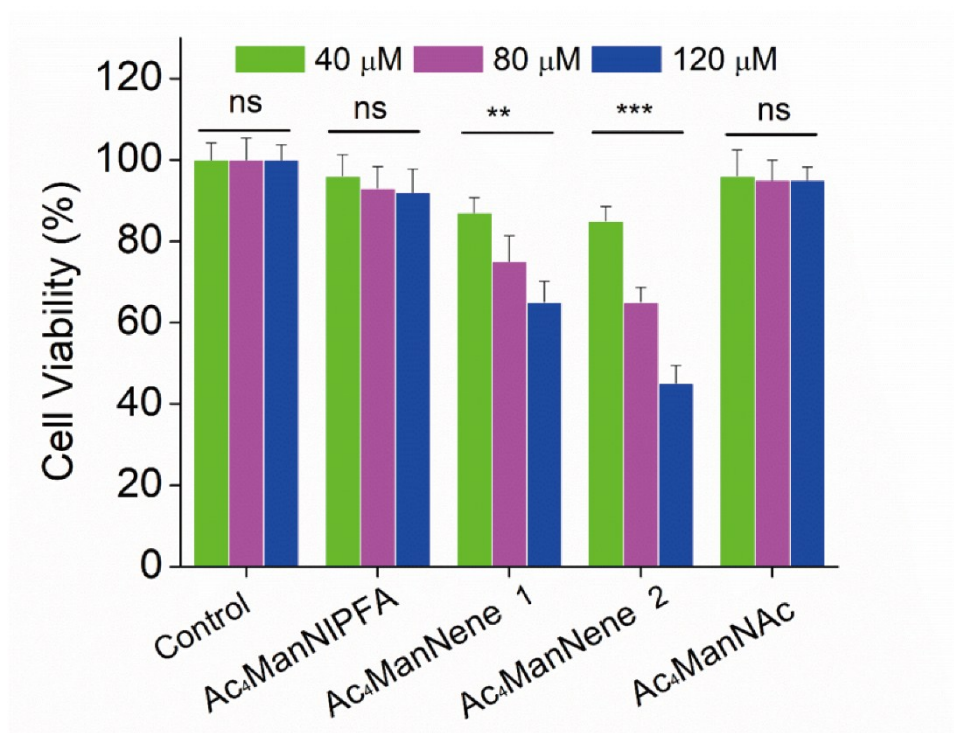
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P.R. China

## 1.Supporting data for the manuscript.

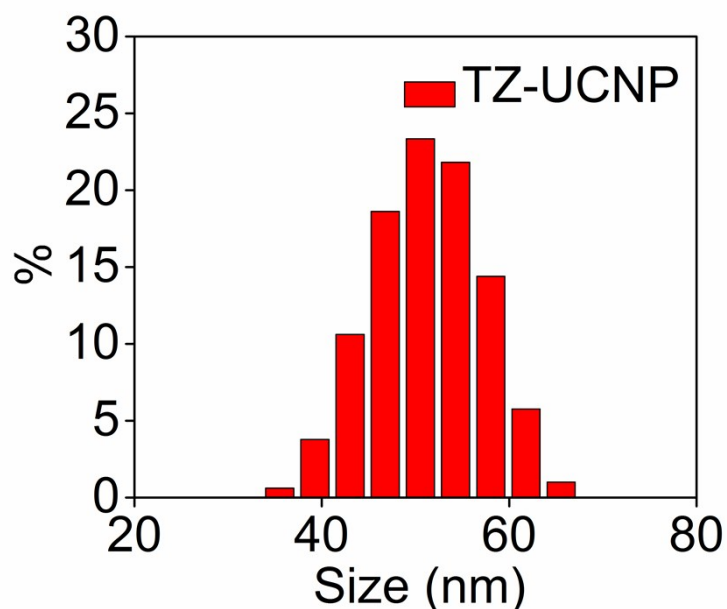
### 1.1 Emission spectra of AC<sub>4</sub>ManNene with tetrazole (Tz).



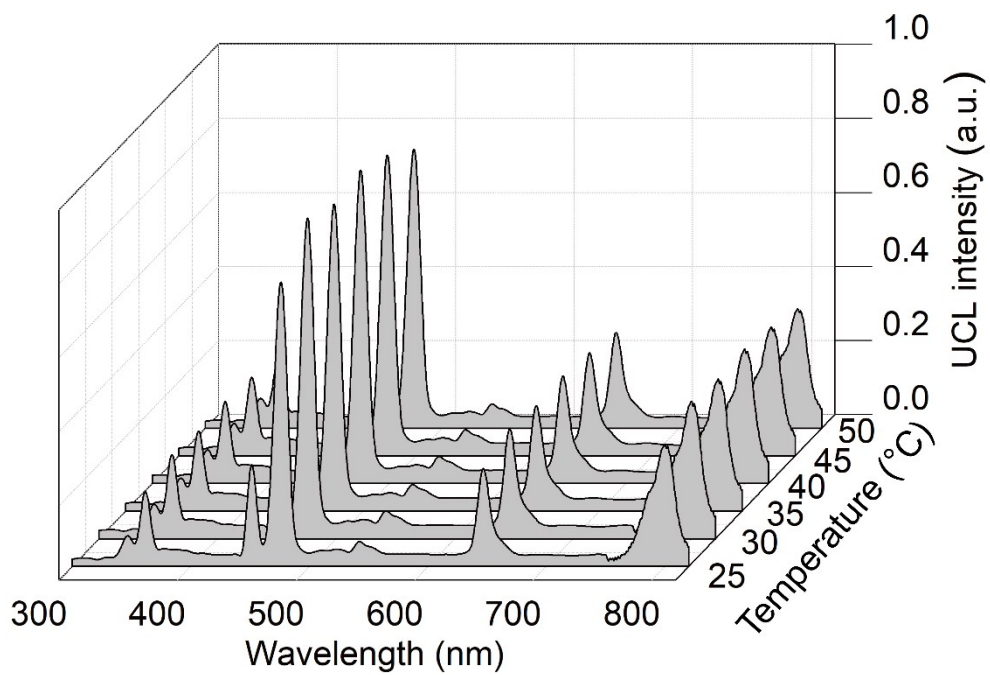
**Fig. S1.** A The chemical structure of Ac<sub>4</sub>ManNIPFA, Ac<sub>4</sub>ManNene 1/2 and Ac<sub>4</sub>ManNAc. B Emission spectra ( $\lambda_{\text{ex}} = 375 \text{ nm}$ ) of Ac<sub>4</sub>ManNIPFA, Ac<sub>4</sub>ManNene 1/2 and Ac<sub>4</sub>ManNAc with tetrazole (Tz) in PBS solution (pH 7.4, 5% DMSO) at a concentration of 10 mM.



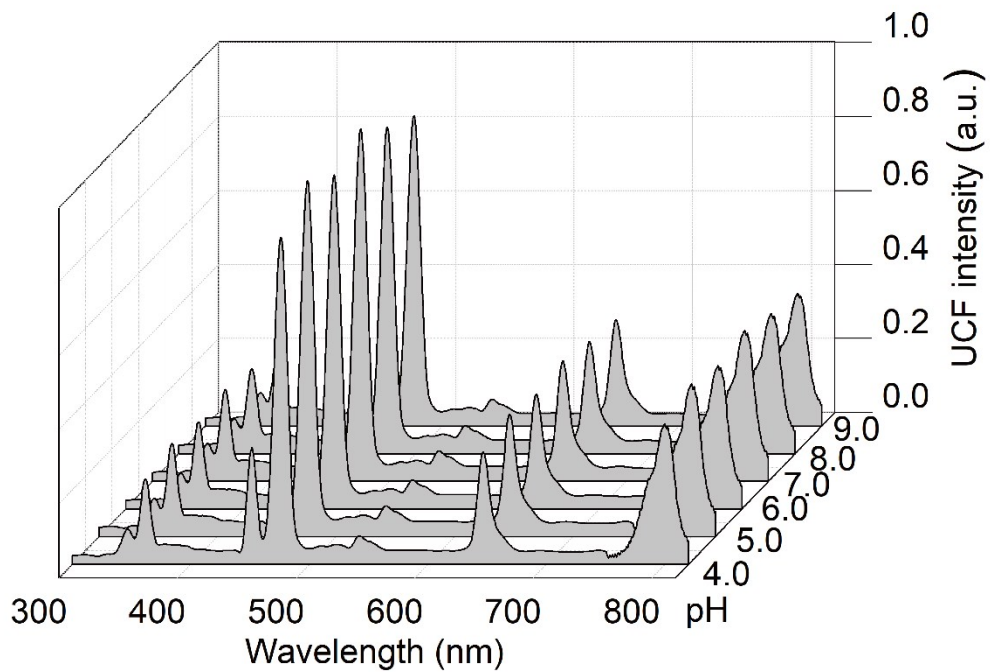
**Fig. S2.** Cell growth analysis after treatment with different alkene sialic acids analogs. A549 cells were grown in the presence of 40 -120 Mm sialic acids analogs (n=5). Data represent mean  $\pm$ SD; \*\*\*p < 0.001, \*\*p < 0.01.



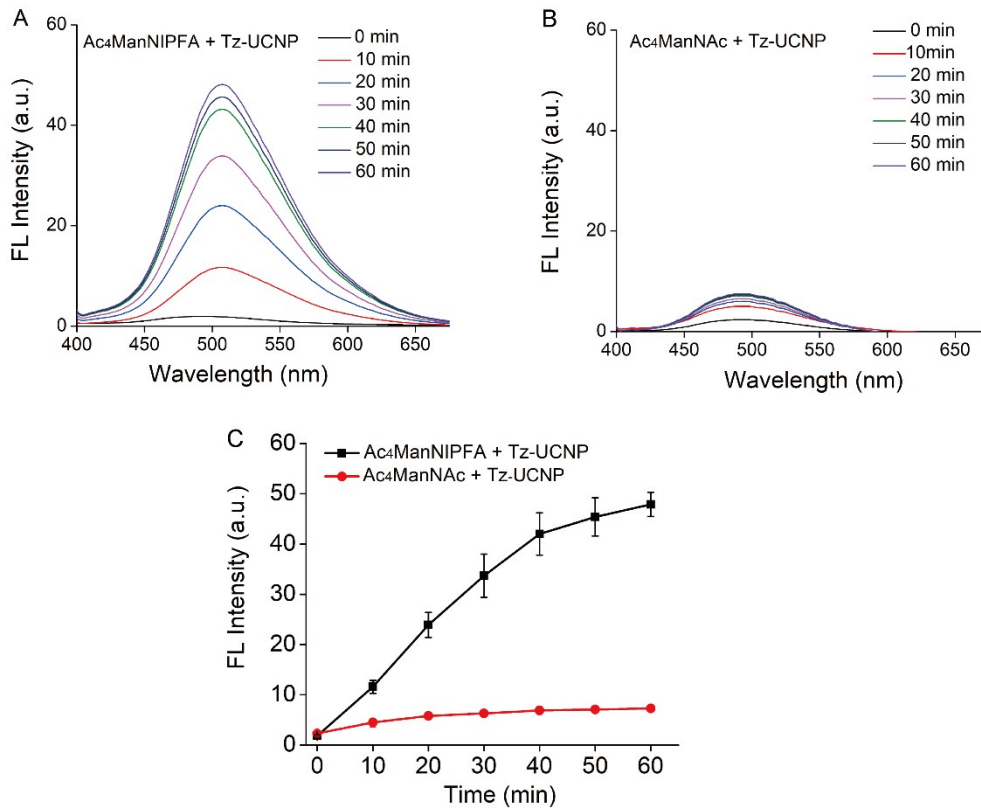
**Fig. S3.** Dynamic light scattering of Tz-UCNP



**Fig. S4.** Tz-UCNP luminescence spectra at different temperatures (25°C -50°C) PBS buffer.

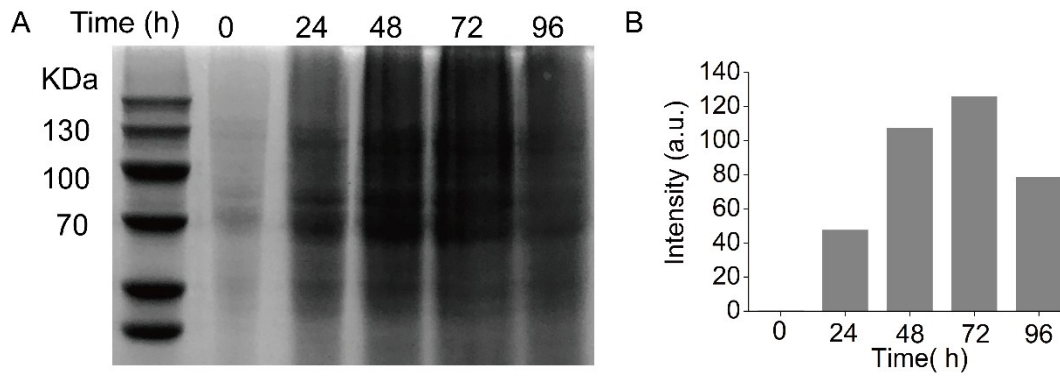


**Fig. S5.** Tz-UCNP luminescence spectra at different pH (4.0-9.0) PBS buffer.

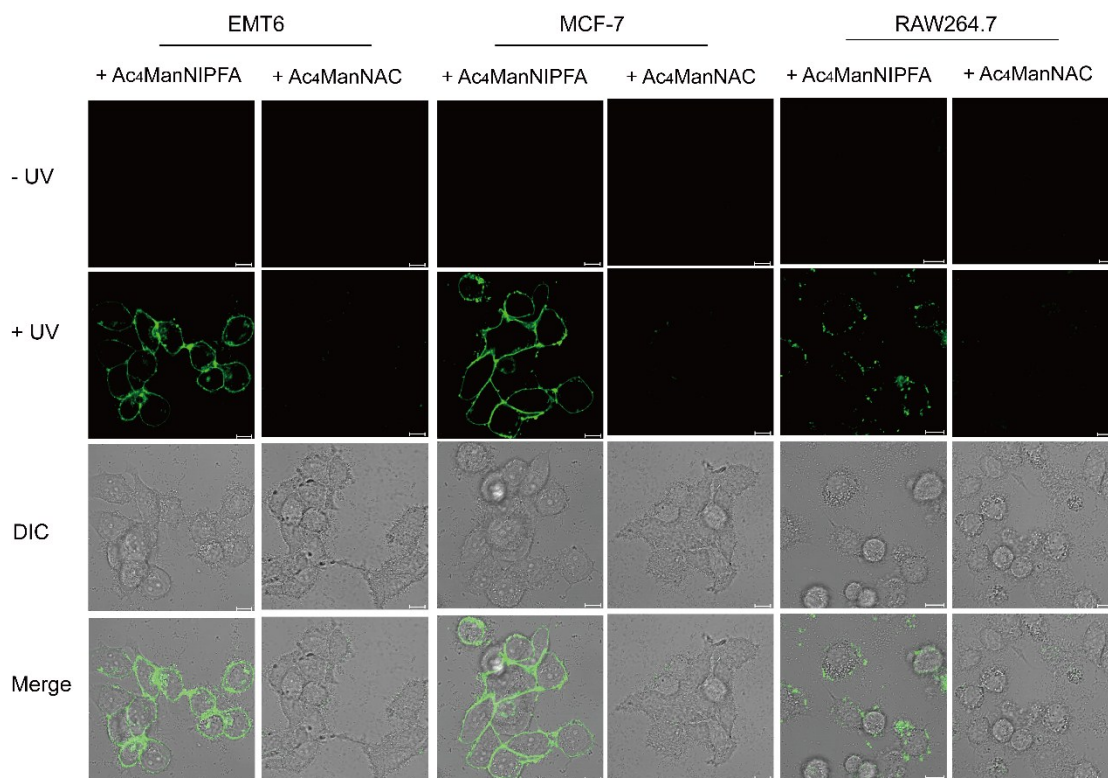


**F**

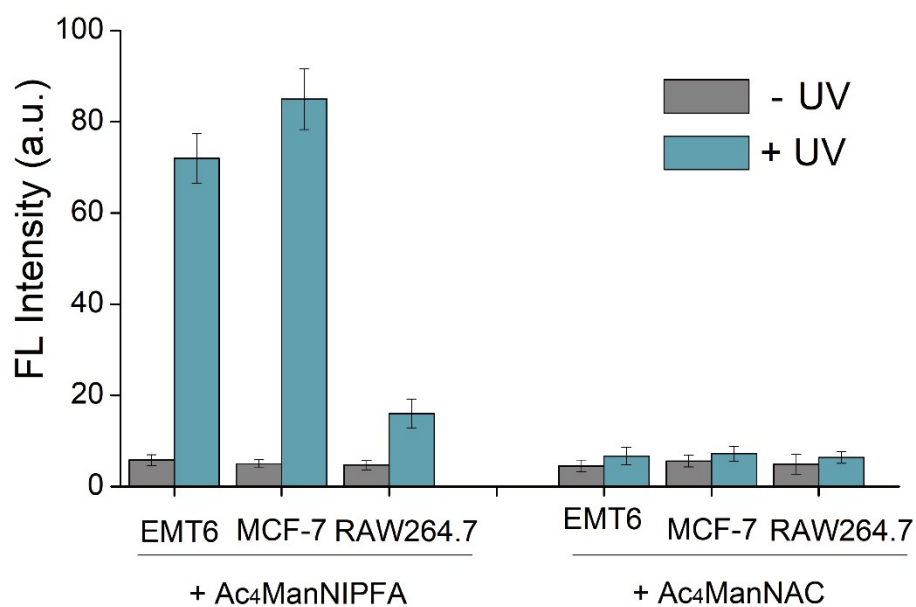
**ig. S6.** A and B. Emission spectra (c,  $\lambda_{ex} = 375$  nm) of AC<sub>4</sub>ManNIPFA, with Tz-UCNP in PBS solution (pH 7.4, 5% DMSO). C. Data statistics of A and B.



**Fig. S7** A. Western blot analysis of lysates from A549 cells A549 cells were pre-incubated with Ac<sub>4</sub>ManNIPFA for 0, 24, 48, 72 and 96 h. B. Data statistics of A

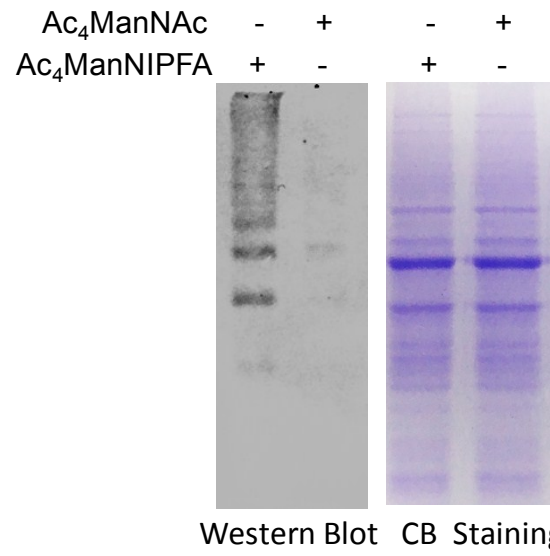


**Fig . S8.** Confocal images of Tz (30  $\mu$ M) labeling on the Ac<sub>4</sub>ManNIPFA (40  $\mu$ M) and Ac<sub>4</sub>ManNac (40  $\mu$ M) pretreated EMT6, MCF-7 and RAW264.7 cells in the absence and presence of UV irradiation for 10 minutes. Scale bar = 10  $\mu$ m.

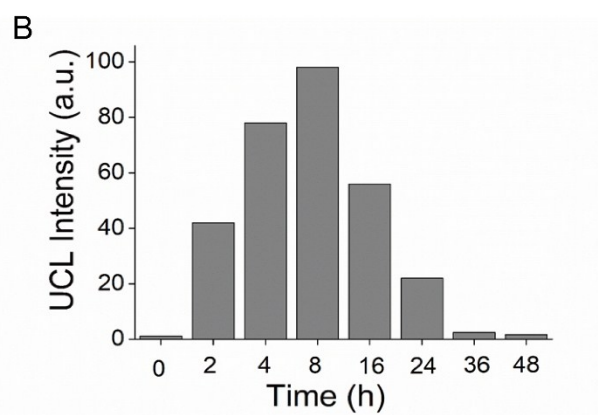
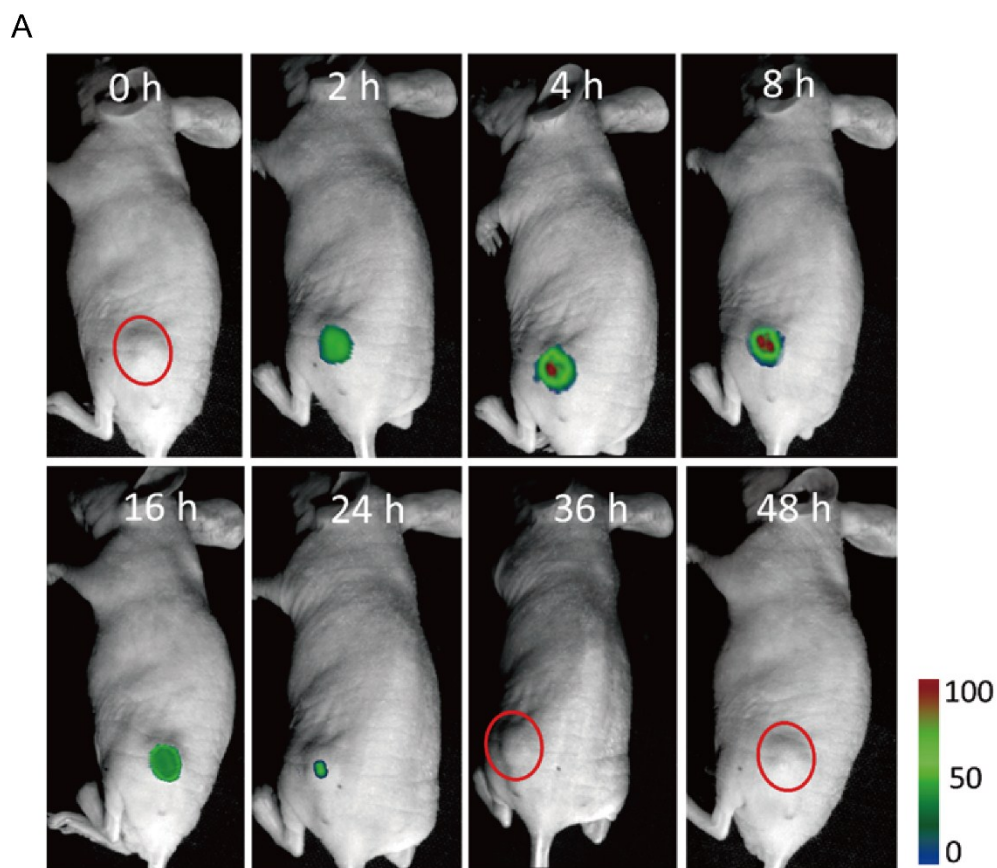


**Fig . S9.** Fluorescence analysis of EMT6, MCF-7 and RAW264.7, cells treated with Ac<sub>4</sub>ManNIPFA and Ac<sub>4</sub>ManNac (40  $\mu$ M).

## 1.2 Supporting data for mice mode.

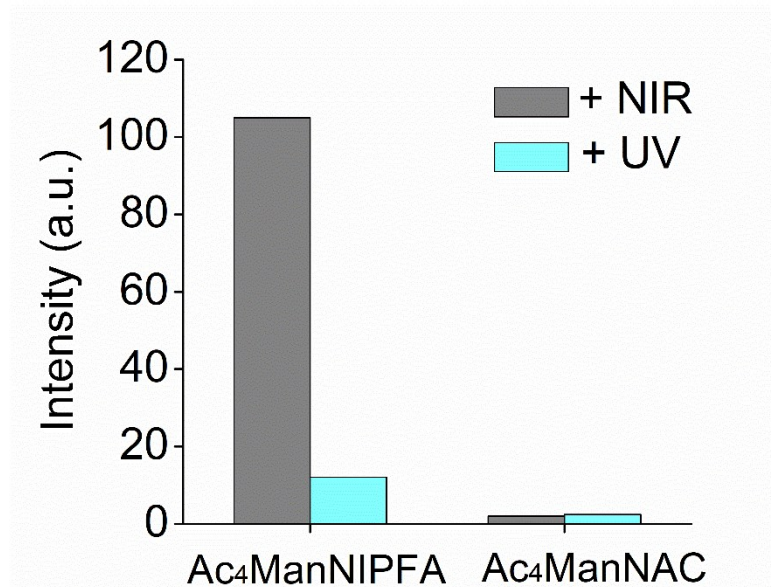


**Fig. S10.** Western blot analysis of lysates from A549 tumor treated with AC<sub>4</sub>ManNIPFA (3 mg/kg) and Ac<sub>4</sub>ManNAc (3 mg/kg).

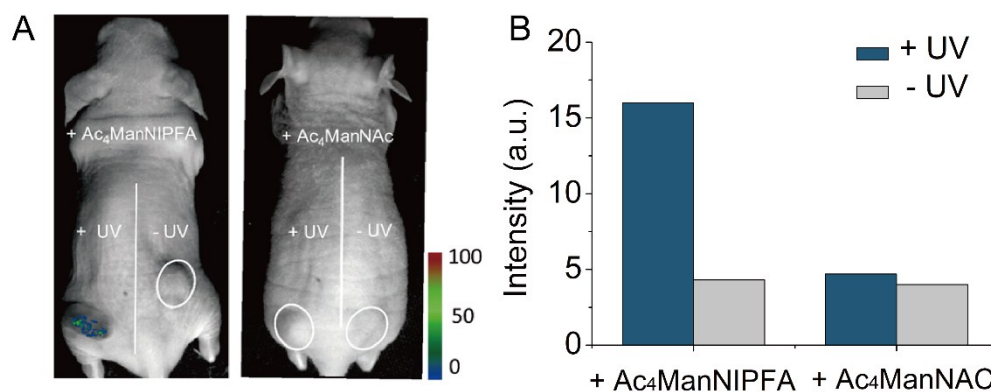


**Fig. S11.** A. Fluorescence images were acquired after different time of incubation the probe Tz-UCNP,  $\lambda_{\text{exc}} = 980 \text{ nm}$ .  $\lambda_{\text{em}} = 800\text{-}810 \text{ nm}$  30 mg/kg of Tz-UCNP was injected into the tail vein of the above mouse. B. Data statistics of A.

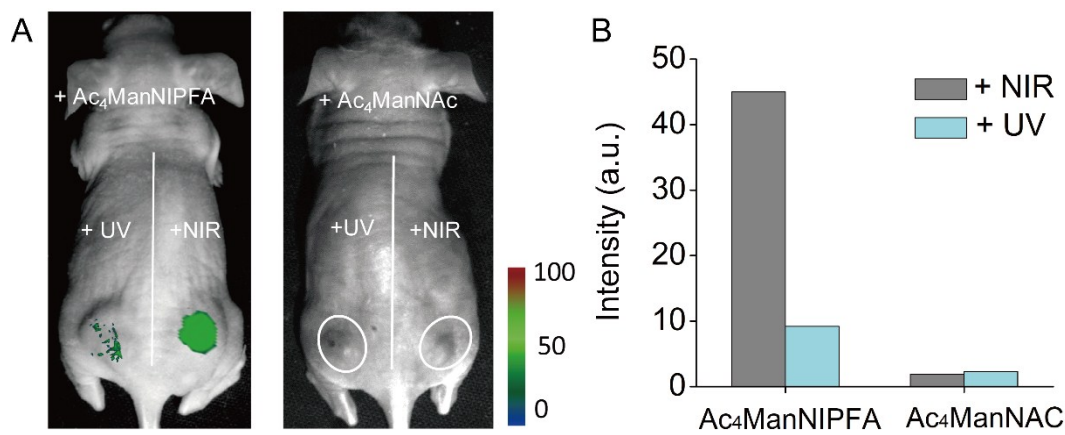




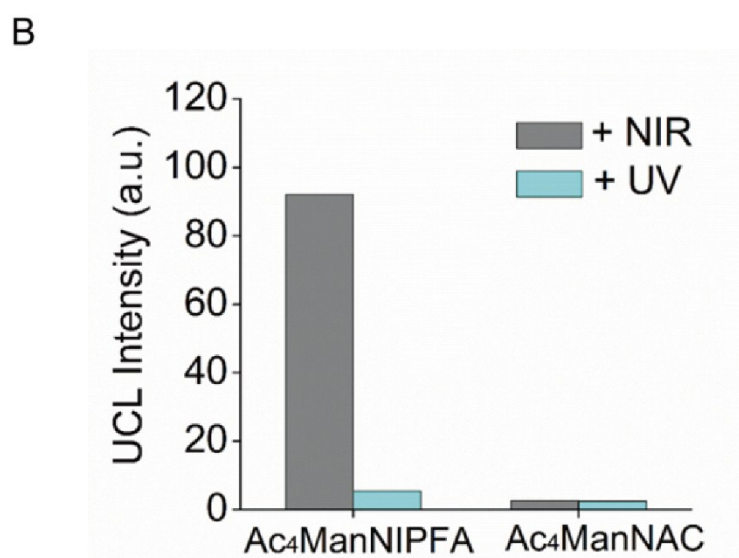
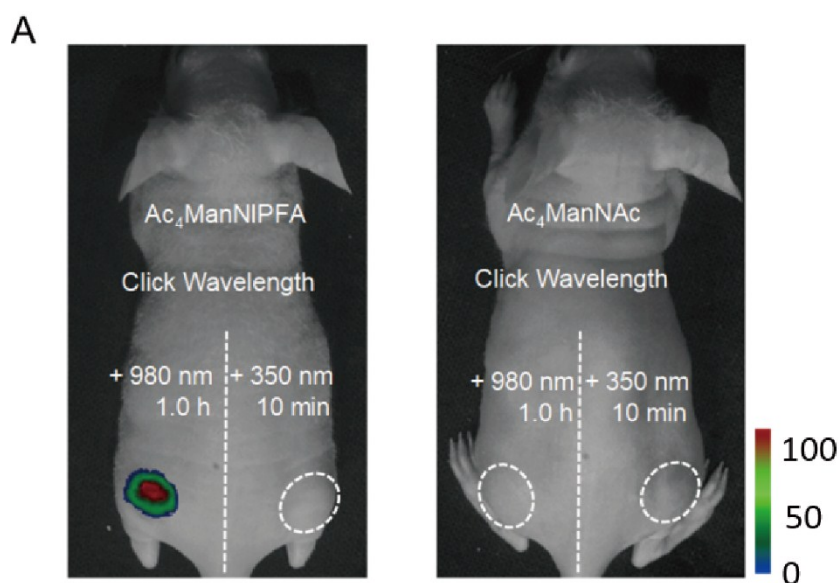
**Fig. S12.** Data statistics of fluorogenic labeling results of sialic acids in living mice.,  $\lambda_{ex} = 980 \text{ nm}$ .  $\lambda_{em} = 505 - 525 \text{ nm}$ . Left: irradiation with UV light 350 nm for 10 min; right irradiation with NIR light 980 nm ( $1.0 \text{ w/cm}^2$ ) for 1 h.



**Fig. S13.** A. Fluorogenic labeling results of sialic acids in living mice.  $\text{Ac}_4\text{ManNIPFA}/\text{Ac}_4\text{ManNAC}$  (3 mg/kg) was intratumoral injected to tumor tissues, once a day for 4 days and then 3 mg/kg of Tz was injected into tumor tissues of the above mouse. Fluorescence images were acquired after the left of side irradiation with UV light 350 nm for 10 min.,  $\lambda_{ex} = 980 \text{ nm}$ .  $\lambda_{em} = 505 - 525 \text{ nm}$ . B. Data statistics of A.



**Fig. S14.** A. Fluorogenic labeling results of sialic acids in living mice.,  $\lambda_{ex} = 980$  nm.  $\lambda_{em} = 505 - 525$  nm. Left: irradiation with UV light 350 nm for 10 min; Right: irradiation with NIR light 980 nm (1.0 w/cm<sup>2</sup>) for 10 min. B. Data statistics of A..



**Fig. S15** A. Fluorescence images of UCNPs in living mice that were treated with Ac<sub>4</sub>ManNIPFA injection with nanoparticle. (3 mg/kg of Ac<sub>4</sub>ManNIPFA and Ac<sub>4</sub>ManNAC were injected into the tail vein of A549 tumor-bearing mice once a day for 4 days, then 30 mg/kg of Tz-UCNP was injected into the tail vein of the above mouse. Fluorescence images were acquired after 48 hours of incubation the probe of Tz-UCNP.  $\lambda_{\text{ex}} = 980 \text{ nm}$  ( $1.0 \text{ w/cm}^2$ ),  $\lambda_{\text{em}} = 800 - 810 \text{ nm}$  Left: irradiation with NIR light ( $1.0 \text{ w/cm}^2$ ) for 1.0 h; Right: irradiation with UV light for 10 min. B. Data statistics of A.

## 2. Supporting experimental section.

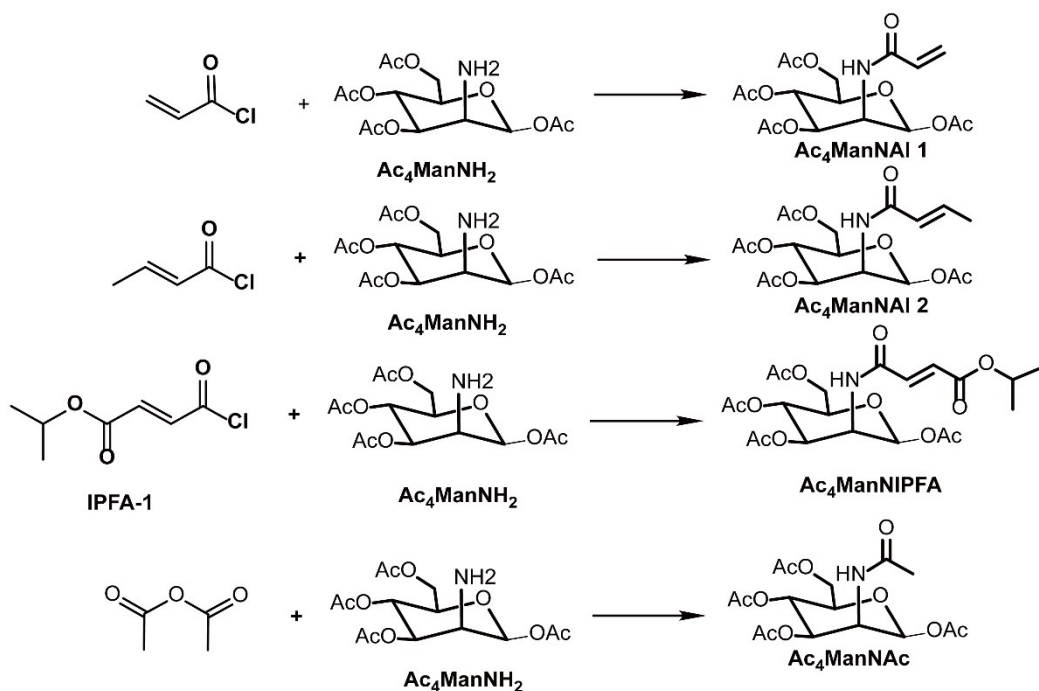
### 2.1 Materials

Y(CH<sub>3</sub>COOH)<sub>3</sub>, Yb(CH<sub>3</sub>COOH)<sub>3</sub>, Tm(CH<sub>3</sub>COOH)<sub>3</sub>, N-Hydroxysuccinimide (NHS), N, N-dimethylformamide (DMF), Dimethyl sulfoxide (DMSO) and 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDC) were purchased from Aladdin Reagent, Ltd. (Shanghai, China); CCK-8 was obtained from Dojindo (Japan). Dulbecco's modified Eagles medium (DMEM) cell culture medium, fetal bovine serum, streptomycin, and penicillin were purchased from Thermo Fisher Scientific Co., Ltd. (China). All chemicals used in this study were of analytical and reagents were used without further purification. All aqueous solutions were prepared using ultrapure deionized water (DI water), which was obtained through a Millipore Milli-Q water purification system (Billerica, USA) and had an electric resistance > 18.2 MΩ.

### 2.2 Instruments.

The <sup>1</sup>H NMR spectra were recorded on a Bruker Ultrashield 400 Plus NMR spectrometer. All chemical shifts are reported in the standard δ notation of parts per million. The UV-vis absorption spectra were recorded on a UV-vis spectrometer (Lambda 35 UV-vis spectrometer, Perkin-Elmer, USA) at room temperature. The fluorescence spectrum was measured using a LS-55 fluorescence spectrophotometer (Perkin-Elmer, USA). The upconversion fluorescence spectra of upconversion nanoparticles were recorded using a 1.0 W/cm<sup>2</sup> 980 nm diode CW laser (Changchun New Industries Optoelectronics Tech. Co., Ltd.) as the excitation source. The TEM images were collected on a field emission high resolution 2100 F transmission electron microscope (JEOL, Japan) operating at an acceleration voltage of 200 kV. The confocal fluorescence images of cells were collected on a ZEISS LSM 710 META confocal microscope. A 40× objective was used for image capturing. The cell viability was measured using a microplate reader (Infinite 200, TECAN, Switzerland). Quantitative data were expressed as mean ± SD. P values were calculated by two-tailed Student's test (\*\*\*p < 0.001, \*\*p < 0.01, or \*p < 0.05). P < 0.05 was considered statistically

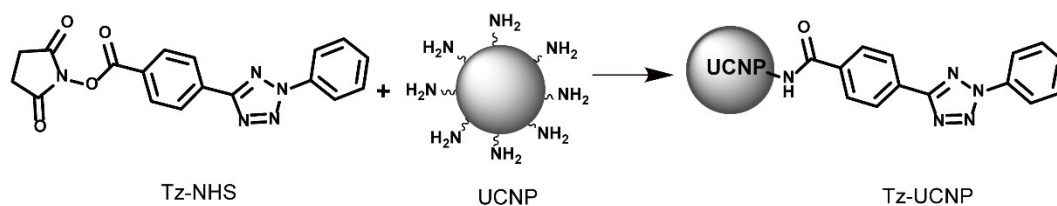
significant. All statistical analyses were carried out using GraphPad Prism Software (Version 6.0, GrapPad Software, San Diego, CA).



**Scheme S1.** Synthetic procedures of the Ac<sub>4</sub>ManNIPFA, AC<sub>4</sub>ManNene 1/2 and Ac<sub>4</sub>ManNAc.

### 2.3 Synthesis of the Ac<sub>4</sub>ManNIPFA.

Ac<sub>4</sub>ManNH<sub>2</sub> was produced according to the previous literature. Ac<sub>4</sub>ManNH<sub>2</sub> (34.8 mg, 0.1 mmol), IPFA (88 mg, 0.5 mmol) were dissolved in dry methylene chloride (10 mL) under nitrogen. The mixture was stirred for 12 h at room temperature. The reaction mixture was collected with methylene chloride solution and dried with Na<sub>2</sub>SO<sub>4</sub>, and concentrated. The crude product was purified over a silica gel column chromatograph with methanol/methylene chloride (1:20) as the eluent to afford the pure product as the Ac<sub>4</sub>ManNIPFA (30.3 mg, 62.2%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): 6.15 (d, *J* = 4.0 Hz, 1H), 6.08 (d, *J* = 8.0 Hz 1H), 5.36 (d, *J* = 4 Hz, 1H), 4.30 (t, *J* = 4 Hz, 1H), 4.06 (t, *J* = 4 Hz, 2H), 2.96 (s, 1H), 2.89 (s, 1H), 2.19 (s, 3H), 2.12 (s, 3H), 2.07 (s, 3H), 2.00 (s, 3H), 1.32 (s, 3H), 1.31 (s, 3H) ESI-MS (*m/z*): calcd for C<sub>21</sub>H<sub>29</sub>NO<sub>12</sub>, 487.46 [M<sup>+</sup>Na<sup>+</sup>2H]; found, 512



**Scheme S2.** Synthesis of the Tz-UCNP

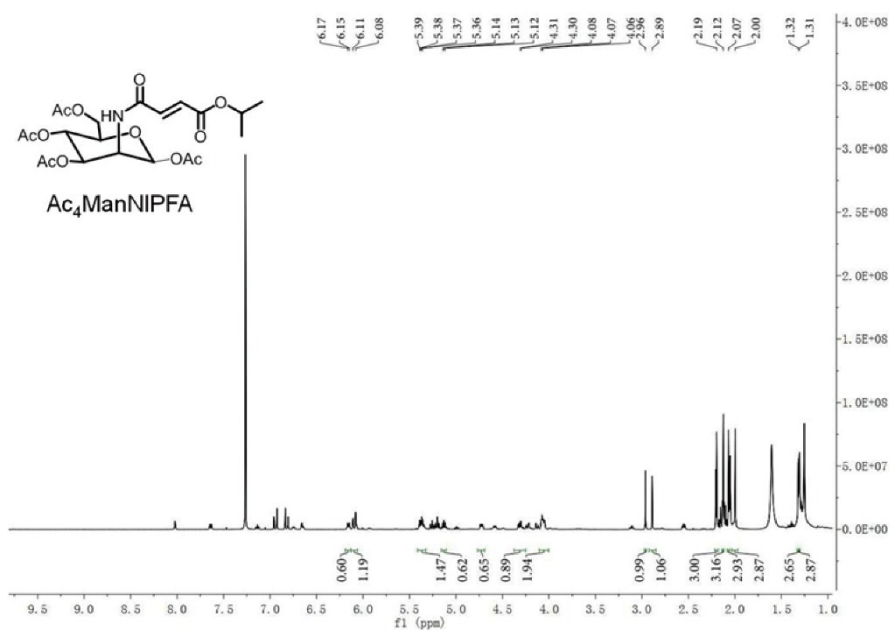
#### 2.4 Synthesis of the UCNP (PEI-Capped NaYF<sub>4</sub>: Yb<sup>3+</sup>, Tm<sup>3+</sup>).

Water dispersed NaYF<sub>4</sub>:Yb<sup>3+</sup>, Tm<sup>3+</sup> nanoparticles were prepared by one step hydrothermal method. Typically, PEI (branched 25 kDa, 1 g), 1 mmol of NaCl, and 1 mmol of Y(CH<sub>3</sub>COOH)<sub>3</sub> (0.5 m), Yb (CH<sub>3</sub>COOH)<sub>3</sub> (0.5 m), and Tm(CH<sub>3</sub>COOH)<sub>3</sub> (0.1 m) with a molar ratio of 79.8:20:0.2 were dissolved in 20 mL of ethanol and stirred for 30 min. Then, 10 mL of ethanol containing 5.5 mmol of NH<sub>4</sub>F was added to the above solution and stirred for another 30 min. After that, the obtained mixture was transferred into a 50 mL stainless teflon-lined autoclave. The autoclave was then sealed and kept at 190 °C for 24 h. After that, the auto clave was cooled to room temperature naturally. The precipitation was then collected by centrifugation (7000 g, 5 min) and washed with ethanol and deionized (DI) water several times and then dried over vacuum at 60 °C for 12 h to afford the as-prepared NaYF<sub>4</sub>: Yb<sup>3+</sup>, Tm<sup>3+</sup> coated with PEI as a white powder.

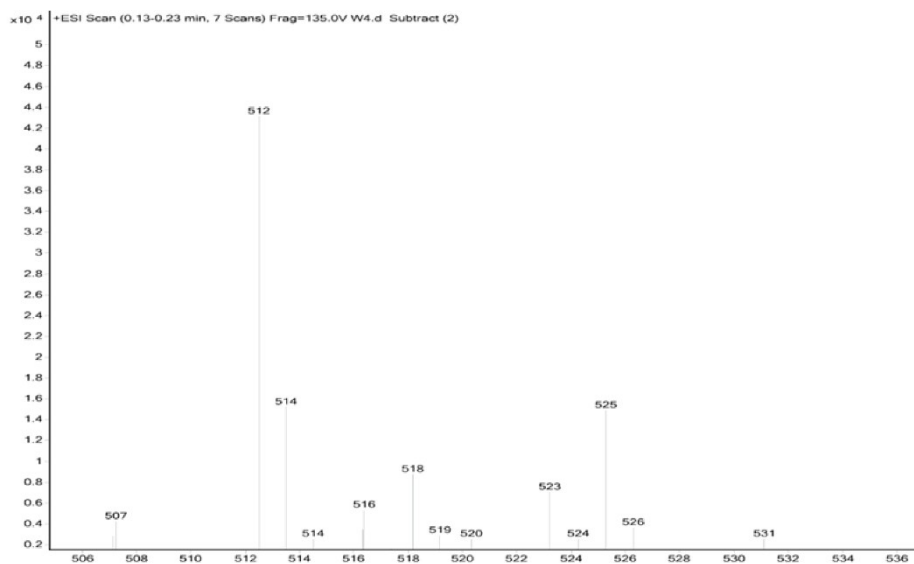
#### 2.5 Cell culture and microscopy.

A549, MCF-7, RAW264.7 and EMT6 cells were cultured in Dulbecco's modified Eagles medium (DMEM). All cell lines were supplemented with 10% fetal bovine serum and 100 U/ml 1% penicillin/streptomycin antibiotics and maintained at 37°C in a 100% humidified atmosphere containing 5% CO<sub>2</sub>. Cell density was determined using a hemocytometer before experimentation. Images were acquired on a ZEISS LSM 710 META confocal microscope. A 40× objective was used to capture images. Images were processed and analyzed using the LSM software.

#### 2.6 <sup>1</sup>H NMR and MS spectra.



400 MHz-<sup>1</sup>H-NMR (CDCl<sub>3</sub>) spectrum of Ac<sub>4</sub>ManNIPFA



ESI-MS spectrum of Ac<sub>4</sub>ManNIPFA