

Supporting information for

# Highly-selective H/D exchange reaction of 1,4-dihydropyridines

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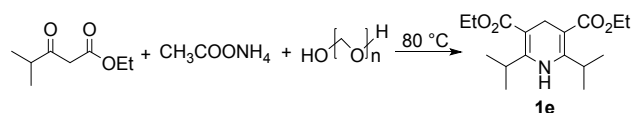
## 1. General information

All reagents were purchased from commercial sources and used without further purification unless otherwise noted. Reactions were monitored by thin-layer chromatography (TLC). Visualization was achieved under a UV lamp (254 nm and 365 nm). Column chromatography was performed using 200-300 mesh silica gels.  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were acquired on 400 and 500 MHz Bruker NMR instruments. NMR chemical shifts were reported in ppm and were referenced to TMS ( $\delta = 0.00$  ppm,  $^1\text{H}$  NMR) or the residual solvent peak for  $\text{CDCl}_3$  ( $\delta = 7.26$  ppm,  $^1\text{H}$  NMR;  $\delta = 77.16$  ppm,  $^{13}\text{C}$  NMR). Following abbreviations are used for multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad. Coupling constants ( $J$ ) are reported in Hertz.

All analytical LC-MS were performed to determine the distribution of hydrogen isotopes of the products on a Shimadzu LC-MS 2020 system equipped with Hadera C18 column (2.1 x 100 mm, 3  $\mu\text{m}$ ; heater set on 40  $^\circ\text{C}$ ) involved a mobile phase of 0.1% formic acid (FA) in water (solvent A) and 0.1% formic acid (FA) in acetonitrile (solvent B) at a flow rate of 0.3 mL/min. All of the samples were tested over the same gradient: from 15 to 55% B in 3 min, then from 55 to 95% B in 7 min, and 95% B for 5 min, 0.1% FA,  $\lambda = 254$  nm.

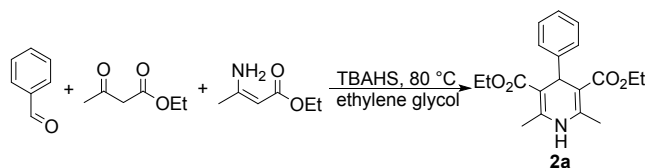
## 2. Synthesis of substrates

**Note:** **1a**, **2k**, **2l**, **2m**, **2n** and **2o** was obtained in commercial source. **1b**, **1c**, **1d**, **1e** and **1f** were synthesized according to reported literature.<sup>1</sup> **2a**, **2b**, **2c**, **2d**, **2e**, **2f**, **2g**, **2h**, **2i**, **2j**, **3a** and **3b** were synthesized according to reported methods.<sup>2-8</sup> The oxidation procedure was performed according to reported literature.<sup>9</sup> Typical synthesis procedures were described as following examples.

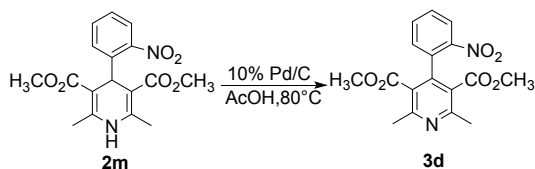


Diethyl 2,6-diisopropyl-1,4-dihydropyridine-3,5-dicarboxylate (**1e**): Ethyl 4-methyl-3-oxopentanoate (1.58 g, 10 mmol), paraformaldehyde (0.15 g, 5 mmol) and ammonium acetate (0.58 g, 7.5 mmol) were added to a 25 mL flask, the resulting mixture was heated to  $80\text{ }^\circ\text{C}$  and stirred for 4 h at this temperature. The mixture was then cooled to room temperature and diluted by ethyl acetate (EA), and washed with brine. The organic layer was dried over anhydrous  $\text{Na}_2\text{SO}_4$ . The solvent was removed under reduced pressure after filtration. A silica gel chromatography was performed with PE/EA = 20:1 to afford a pale yellow solid **1e** (0.76g, 49%).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  5.68 (s, 1H), 4.08 (q,  $J = 7.1$  Hz, 4H), 4.02 (p,  $J = 7.0$  Hz, 2H), 3.18 (s, 2H), 1.20 (t,  $J = 7.1$

Hz, 6H), 1.04 (d,  $J = 7.2$  Hz, 12H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  167.5, 153.4, 97.5, 59.5, 27.5, 25.1, 20.2, 14.4. LRMS (ESI+)  $m/z$ : 310.3 [ $\text{M}+\text{H}^+$ ].



Diethyl 2,6-dimethyl-4-phenyl-1,4-dihydropyridine-3,5-dicarboxylate (**2a**): To a solution of ethyl 3-aminocrotonate (0.65 g, 5 mmol), ethyl acetoacetate (0.65 g, 5 mmol) and the corresponding benzaldehyde (0.53 g, 5 mmol) in ethylene glycol (2 mL) was added TBAHS ( $\text{Bu}_4\text{NHSO}_4$ , 0.2 g, 0.6 mmol). The mixture was heated to 80 °C and stirred for 4 h. After complete consumption of the aldehyde monitored by TLC, the reaction was cooled to room temperature and diluted with EA. The solution was poured into a separatory funnel containing brine and extracted three times with EA. After drying over anhydrous  $\text{Na}_2\text{SO}_4$ , it was filtered and concentrated under reduced pressure. The crude reaction mixture was purified by silica gel chromatography using PE/EA=6:1 as eluent to give **2a** as a pale yellow solid (0.86 g, 52%).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.29 – 7.26 (m, 2H), 7.22 – 7.17 (m, 2H), 7.14 – 7.09 (m, 1H), 6.07 (s, 1H), 4.99 (s, 1H), 4.15 – 4.02 (m, 4H), 2.29 (s, 6H), 1.21 (t,  $J = 7.1$  Hz, 6H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  167.9, 147.9, 144.3, 128.0, 127.9, 126.2, 104.0, 59.8, 39.7, 19.5, 14.3. LRMS (ESI+)  $m/z$ : 330.3 [ $\text{M}+\text{H}^+$ ].



Dimethyl 2,6-dimethyl-4-(2-nitrophenyl)pyridine-3,5-dicarboxylate (**3d**): **2m** (1.73 g, 5 mmol), 10% Pd/C (20% weight of **2m**, 0.35 g) and acetic acid (15 mL) were added to a 50 mL flask. The resulting mixture was heated to 80 °C and stirred for 4 h. After completion of the reaction determined by TLC analysis, Pd/C was filtered off by celite. The filtrate was poured into saturated  $\text{NaHCO}_3$  and extracted with EA for three times. Then the combined organic layers were washed with brine and dried over anhydrous  $\text{Na}_2\text{SO}_4$ . The solvents were removed under reduced pressure and the crude mixture was purified by silica gel chromatography with PE/EA = 10:1 as eluent to afford pure product **3d** as a pale yellow solid (0.86 g, 50%).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  8.18 (dq,  $J = 8.4, 1.8$  Hz, 1H), 7.64 – 7.59 (m, 1H), 7.58 – 7.53 (m, 1H), 7.21 – 7.16 (m, 1H), 3.48 (s, 6H), 2.63 (s, 6H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  167.3, 157.1, 147.7, 145.3, 133.0, 132.1, 130.7, 129.7, 124.9, 124.4, 52.3, 23.7. LRMS (ESI+)  $m/z$ : 345.2 [ $\text{M}+\text{H}^+$ ].

### 3. General procedure 1 for HIE reactions

To a 10 mL Schlenk tube with a magnetic bar was added 1,4-dihydropyridines (0.2 mmol), D<sub>2</sub>O (10 mmol, 181  $\mu$ L), TFA-*d* (23  $\mu$ L, 0.3 mmol (or 154  $\mu$ L, 2 mmol)) and NMP (2 mL). The solution was frozen in liquid nitrogen, then the vessel was evacuated and backfilled with nitrogen following by being warmed to room temperature. The operation was repeated for 3 cycles. The resulting mixture was stirred at 50 °C (or 70 °C) for 24 h (or 48 h), cooled to room temperature, diluted with EA and then washed with saturated NaHCO<sub>3</sub> and brine. The organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuo. The crude product was further purified by silica gel chromatography. Hydrogen isotope distribution of the products was determined by LC-MS. <sup>1</sup>H NMR was performed to determine deuterium incorporation.

## 4. Condition optimization.

### 4.1 Condition optimization for deuteration of Hantzsch ester **1a**

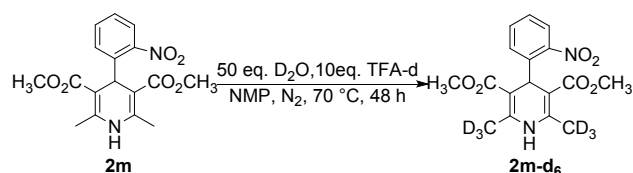
Entry	Variations	D Incorporation <sup>b</sup> (%)	Yield <sup>c</sup> (%)
1	none <sup>a</sup>	85 <sup>d</sup>	53(71 <sup>e</sup> )
2	no TFA- <i>d</i>	0	5
3	TFA	63	54
4	3 eq. TFA- <i>d</i>	82	53
5	RT	38	37
6	70°C	75	68 <sup>e</sup>
7	Air	45	99
8	degassed by N <sub>2</sub> purging	65	57
9	DMF as solvent	74	26
10	DMA as solvent	83	31
11	MeOH as solvent	15	46
12	MeOD as solvent	75	45

<sup>a</sup> standard condition: **1a** (0.2 mmol, 50.7 mg), D<sub>2</sub>O (10 mmol, 181  $\mu$ L), TFA-*d* (0.3 mmol, 23  $\mu$ L), NMP (*N*-Methyl pyrrolidone, 2 mL), N<sub>2</sub>, 50 °C for 24 h. <sup>b</sup> D incorporation determined by <sup>1</sup>H NMR spectroscopy. <sup>c</sup> crude yield determined by <sup>1</sup>H NMR spectroscopy. <sup>d</sup> an average of two parallel reactions. <sup>e</sup> isolated yield.

**Note:** The isolated yield was much higher as all materials were exposed to air during purification process. Absolutely no deuteration and little oxidation occurred when TFA-*d* was omitted, showing acid to be critical for both the D incorporation and oxidation process. Only 5% oxidation product was obtained as no acid was added. No other byproduct was detected during the experiments. We also tested the pH value

of the reaction system under standard conditions. The pH value was 2.42 as all reagents were well mixed and after 24 hours reaction it was 2.43. To our best knowledge, the oxidation of **1a** involved unknown hydrogen transfer under heating and acidic conditions. More deeper studies are needed to get approach to the detailed mechanism. D incorporation did not change even the product was immersed in chloroform-*d* for 3 days longer. To confirm the relationship between oxidation and deuteration, we performed the experiment under standard conditions with **1a'** prepared previously<sup>9</sup>, whereupon 85% D incorporation showed the deuteration and oxidation processes to be independent of one another.

## 4.2 Condition optimization for deuteration of nifedipine **2m**

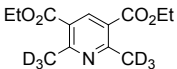


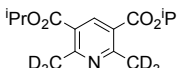
Entry	Variations	D Incorporation <sup>b</sup> (%)
1	1.5 eq. TFA- <i>d</i> , 50 °C, 24 h	24
2	3 eq. TFA- <i>d</i> , 50 °C, 48 h	25
3	6 eq. TFA- <i>d</i>	74
4	none <sup>a</sup>	80 <sup>c</sup>
5	12 eq. TFA- <i>d</i>	80
6	no TFA- <i>d</i> , 24 h	0
7	no D <sub>2</sub> O	15
8	25 eq. D <sub>2</sub> O	47
9	100 eq. D <sub>2</sub> O	76
10	150 eq. D <sub>2</sub> O	80
11	10 eq. TFA	71
12	10 eq. TfOH	68
13	10 eq. BF <sub>3</sub> ·OEt <sub>2</sub>	76
14	20 eq. BF <sub>3</sub> ·OEt <sub>2</sub>	72
15	30 eq. BF <sub>3</sub> ·OEt <sub>2</sub>	71
16	24 h	71
17	150 h	79

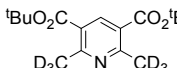
<sup>a</sup> standard condition: **2m** (0.2 mmol, 69.3 mg), D<sub>2</sub>O (10 mmol, 181 μL), TFA-*d* (2 mmol, 154 μL), NMP ( *N*-Methyl pyrrolidone, 2 mL), N<sub>2</sub>, 70 °C for 48 h. <sup>b</sup> D incorporation determined by <sup>1</sup>H NMR spectroscopy. <sup>c</sup> an average of two parallel reactions

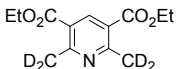
## 5. Deuteration of 1,4-dihydropyridines and derivatives

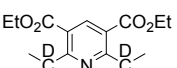
All of the deuteration procedure was done according to general procedure 1 with or without some variations.

 **Diethyl 2,6-bis(methyl-d<sub>3</sub>)pyridine-3,5-dicarboxylate (1a'-d<sub>6</sub>).** **1a** (50.7 mg, 0.2 mmol) and TFA-*d* (23 μL, 0.3 mmol) were used according to general procedure 1 to obtain **1a'-d<sub>6</sub>** as a white solid (36.5 mg, 0.142 mmol, 71%, 85% D). The same result was obtained as **1a'** (50.3 mg, 0.2 mmol) and TFA-*d* (23 μL, 0.3 mmol) were used according to general procedure 1. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.64 (s, 1H), 4.37 (q, *J* = 7.2 Hz, 4H), 2.82 - 2.76 (m, 0.9H), 1.39 (t, *J* = 7.1 Hz, 6H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 166.0, 162.3, 141.0, 123.2, 61.5, 24.9, 24.8, 24.7, 24.5, 24.4, 24.3, 24.2, 24.1, 14.4.

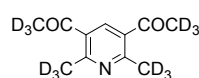
 **Diisopropyl 2,6-bis(methyl-d<sub>3</sub>)pyridine-3,5-dicarboxylate (1b'-d<sub>6</sub>).** **1b** (56.3 mg, 0.2 mmol) and TFA-*d* (23 μL, 0.3 mmol) were used according to general procedure 1 to obtain **1b'-d<sub>6</sub>** as a white solid (40.4 mg, 0.142 mmol, 71%, 77% D). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.59 (s, 1H), 5.24 (hept, *J* = 6.3 Hz, 2H), 2.82 - 2.76 (m, 1.4H), 1.37 (d, *J* = 6.3 Hz, 12H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 165.8, 161.8, 140.9, 123.8, 69.2, 24.9, 24.8, 24.8, 24.7, 24.6, 24.5, 24.4, 24.2, 22.0.

 **Di-tert-butyl 2,6-bis(methyl-d<sub>3</sub>)pyridine-3,5-dicarboxylate (1c'-d<sub>6</sub>).** **1c** (61.8 mg, 0.2 mmol) and TFA-*d* (23 μL, 0.3 mmol) were used according to general procedure 1 to obtain **1c'-d<sub>6</sub>** as a white solid (44.8 mg, 0.144 mmol, 72%, 81% D). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.51 (s, 1H), 2.79 - 2.74 (m, 1.2H), 1.59 (d, *J* = 1.1 Hz, 18H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 165.6, 161.2, 140.9, 124.8, 82.2, 28.3, 25.0, 24.9, 24.8, 24.7, 24.6, 24.5, 24.4, 24.2.

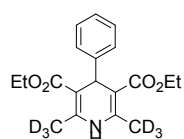
 **Diethyl 2,6-bis(ethyl-1,1-d<sub>2</sub>)pyridine-3,5-dicarboxylate (1d'-d<sub>4</sub>).** **1d** (28.1 mg, 0.1 mmol) and TFA-*d* (12 μL, 0.150 mmol) were used according to general procedure 1 to obtain **1d'-d<sub>4</sub>** as a pale yellow solid (23.2 mg, 0.082 mmol, 82%, 71% D). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.60 (s, 1H), 4.39 (q, *J* = 7.1 Hz, 4H), 3.22-3.13 (m, 1.2H), 1.41 (t, *J* = 7.1 Hz, 6H), 1.29 (d, *J* = 8.5 Hz, 6H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 167.0, 166.2, 141.4, 122.8, 61.6, 30.5, 30.2, 14.4, 14.0, 14.0, 13.9.

 **Diethyl 2,6-bis(propan-2-yl-2-d)pyridine-3,5-dicarboxylate (1e'-d<sub>2</sub>).** **1e** (61.9 mg, 0.2 mmol) and TFA-*d* (23 μL, 0.3 mmol) were used according to general procedure 1 to obtain **1e'-d<sub>2</sub>** as a white solid (46.1 mg, 0.150 mmol, 75%, 19% D). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.43 (s, 1H), 4.37 (q, *J* = 7.1

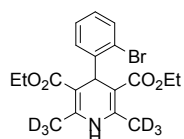
Hz, 4H), 3.86 (hept,  $J = 6.7$  Hz, 1.6H), 1.39 (t,  $J = 7.1$  Hz, 6H), 1.28 (d,  $J = 6.7$  Hz, 12H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  169.4, 166.7, 140.3, 122.2, 61.5, 32.9, 22.3, 22.2, 14.4.



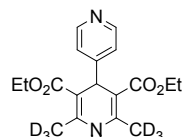
**1,1'-(2,6-bis(methyl-d<sub>3</sub>)pyridine-3,5-diyl)bis(ethan-1-one-2,2,2-d<sub>3</sub>) (1f-d<sub>12</sub>).** **1f** (38.7 mg, 0.2 mmol) and TFA-*d* (23  $\mu\text{L}$ , 0.3 mmol) were used according to general procedure 1 to obtain **1f-d<sub>12</sub>** as a pale yellow solid (26.0 mg, 0.128 mmol, 64%, 71% D, 61% D).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  8.23 (s, 1H), 2.77 – 2.71 (m, 1.7H), 2.62 – 2.57 (m, 2.4H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  199.4, 160.4, 138.0, 130.3, 29.5, 29.4, 29.3, 29.3, 29.2, 29.1, 29.0, 28.8, 24.9, 24.8, 24.7, 24.6, 24.5, 24.4, 24.2.



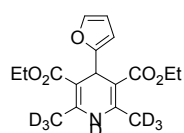
**Diethyl 2,6-bis(methyl-d<sub>3</sub>)-4-phenyl-1,4-dihydropyridine-3,5-dicarboxylate (2a-d<sub>6</sub>).** **2a** (65.9 mg, 0.2 mmol) and TFA-*d* (46  $\mu\text{L}$ , 0.6 mmol) were used according to general procedure 1 to obtain **2a-d<sub>6</sub>** as a white solid (65.1 mg, 0.194 mmol, 97%, 77% D).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.30 – 7.26 (m, 2H), 7.20 (t,  $J = 7.7$  Hz, 2H), 7.11 (td,  $J = 7.0, 1.4$  Hz, 1H), 5.88 (s, 1H), 4.98 (s, 1H), 4.12 – 4.04 (m, 4H), 2.32 – 2.25 (m, 1.4H), 1.21 (t,  $J = 7.1$  Hz, 6H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  167.8, 147.9, 144.1, 128.1, 127.9, 126.2, 104.2, 59.8, 39.7, 19.5, 19.4, 19.4, 19.3, 19.2, 19.1, 19.0, 18.9, 18.7, 14.4.



**Diethyl 4-(2-bromophenyl)-2,6-bis(methyl-d<sub>3</sub>)-1,4-dihydropyridine-3,5-dicarboxylate (2b-d<sub>6</sub>).** **2b** (81.7 mg, 0.2 mmol) and TFA-*d* (154  $\mu\text{L}$ , 2 mmol) were used according to general procedure 1 to obtain **2b-d<sub>6</sub>** as a pale yellow solid (81.2 mg, 0.196 mmol, 98%, 77% D).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.41 (dd,  $J = 8.0, 1.3$  Hz, 1H), 7.37 (dd,  $J = 7.8, 1.8$  Hz, 1H), 7.15 (td,  $J = 7.5, 1.3$  Hz, 1H), 6.93 (ddd,  $J = 7.9, 7.2, 1.7$  Hz, 1H), 6.07 (s, 1H), 5.35 (s, 1H), 4.09 (qd,  $J = 7.1, 4.7$  Hz, 4H), 2.25 – 2.19 (m, 1.4H), 1.19 (t,  $J = 7.1$  Hz, 6H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  167.9, 147.6, 143.9, 143.9, 132.8, 131.7, 127.7, 127.5, 122.7, 104.2, 104.1, 59.8, 39.8, 19.5, 19.4, 19.3, 19.2, 19.2, 19.1, 19.0, 18.9, 18.8, 18.8, 18.7, 18.6, 14.5.



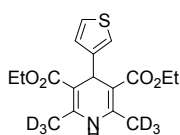
**Diethyl 2,6-bis(methyl-d<sub>3</sub>)-1,4-dihydro-[4,4'-bipyridine]-3,5-dicarboxylate (2c-d<sub>6</sub>).** **2c** (66.1 mg, 0.2 mmol) and  $\text{BF}_3 \cdot \text{OEt}_2$  (494  $\mu\text{L}$ , 4 mmol) were used according to general procedure 1 to obtain **2c-d<sub>6</sub>** as a pale yellow solid (63.9 mg, 0.190 mmol, 95%, 70% D).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  8.42 (d,  $J = 5.1$  Hz, 2H), 7.24 – 7.21 (m, 2H), 6.50 (s, 1H), 5.00 (s, 1H), 4.13 – 4.05 (m, 4H), 2.34 – 2.27 (m, 1.8H), 1.21 (t,  $J = 7.1$  Hz, 6H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  167.3, 156.7, 149.1, 145.3, 145.3, 123.6, 102.7, 102.7, 60.1, 39.6, 19.5, 19.4, 19.3, 19.2, 19.1, 19.1, 18.9, 14.4.



**Diethyl 4-(furan-2-yl)-2,6-bis(methyl-d<sub>3</sub>)-1,4-dihydropyridine-3,5-dicarboxylate-1-d (2d-d).** **2d** (63.9 mg, 0.2 mmol) and TFA-*d* (154  $\mu\text{L}$ , 2 mmol) were used according to general procedure 1 to obtain **2d-d**

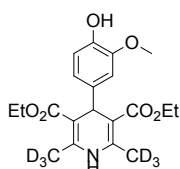


as a white solid (63.5 mg, 0.196 mmol, 98%, 81% D, -ND, 67% D).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.19 (dd,  $J = 1.8, 0.9$  Hz, 0.3H), 6.23 – 6.16 (m, 1H), 6.03 (s, 1H), 5.92 (d,  $J = 3.2$  Hz, 1H), 5.19 (s, 1H), 4.21 – 4.09 (m, 4H), 2.31 – 2.25 (m, 1.1H), 1.25 (t,  $J = 7.1$  Hz, 6H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  167.7, 158.8, 158.7, 145.3, 140.9, 110.1, 109.9, 104.5, 100.8, 59.9, 33.5, 19.5, 19.4, 19.3, 19.2, 19.2, 19.1, 19.0, 18.9, 18.8, 18.8, 18.7, 18.5, 14.4.



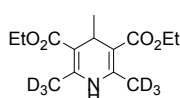
**Diethyl 2,6-bis(methyl-d<sub>3</sub>)-4-(thiophen-3-yl)-1,4-dihydropyridine-3,5-dicarboxylate (2e-d<sub>6</sub>).** **2e** (67.1 mg, 0.2 mmol) and TFA-*d* (154  $\mu\text{L}$ , 2 mmol) were used according to general procedure 1 to obtain **2e-d<sub>6</sub>** as a pale yellow solid (67.3mg, 0.197 mmol, 99%, 82% D).  $^1\text{H}$  NMR (500

MHz,  $\text{CDCl}_3$ )  $\delta$  7.11 (dd,  $J = 5.0, 3.1$  Hz, 1H), 6.98 (dd,  $J = 5.0, 1.3$  Hz, 1H), 6.91 (dd,  $J = 3.3, 1.3$  Hz, 1H), 5.96 (s, 1H), 5.13 (s, 1H), 4.19 – 4.07 (m, 4H), 2.30 – 2.24 (m, 1.1H), 1.24 (t,  $J = 7.1$  Hz, 6H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  167.8, 148.0, 144.6, 144.6, 127.7, 124.7, 120.4, 103.5, 103.5, 77.4, 77.2, 76.9, 59.9, 34.7, 19.4, 19.3, 19.2, 19.2, 19.1, 19.0, 18.9, 18.8, 18.8, 18.7, 18.6, 14.4.



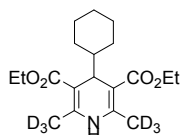
**Diethyl 4-(4-hydroxy-3-methoxyphenyl)-2,6-bis(methyl-d<sub>3</sub>)-1,4-dihydropyridine-3,5-dicarboxylate (2f-d<sub>6</sub>).** **2f** (75.1 mg, 0.2 mmol) and TFA-*d* (154  $\mu\text{L}$ , 2 mmol) were used according to general procedure 1 to obtain **2f-d<sub>6</sub>** as a white solid (75.4 mg, 0.198 mmol, 99%, 80% D).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  6.84 (d,  $J = 1.6$  Hz, 1H), 6.75 – 6.70 (m, 2H),

5.88 (s, 1H), 5.63 – 5.52 (m, 1H), 4.91 (s, 1H), 4.13 – 4.05 (m, 4H), 3.81 (s, 3H), 2.31 – 2.24 (m, 1.2H), 1.23 (t,  $J = 7.1$  Hz, 6H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  167.9, 145.9, 144.0, 143.9, 143.8, 140.2, 120.5, 114.0, 111.0, 104.3, 104.3, 59.8, 55.8, 39.2, 19.5, 19.4, 19.4, 19.3, 19.2, 19.1, 19.0, 19.0, 18.9, 18.8, 18.7, 14.4.



**Diethyl 4-methyl-2,6-bis(methyl-d<sub>3</sub>)-1,4-dihydropyridine-3,5-dicarboxylate (2g-d<sub>6</sub>).** **2g** (53.5 mg, 0.2 mmol) and TFA-*d* (154  $\mu\text{L}$ , 2 mmol) were used according to general procedure 1 to obtain **2g-d<sub>6</sub>** as

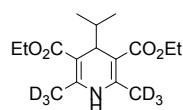
a pale yellow solid (50.8 mg, 0.186 mmol, 93%, 77% D).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  5.74 (s, 1H), 4.27 – 4.13 (m, 4H), 3.84 (q,  $J = 6.5$  Hz, 1H), 2.28 – 2.21 (m, 1.4H), 1.30 (t,  $J = 7.1$  Hz, 6H), 0.98 (d,  $J = 6.5$  Hz, 3H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  168.0, 144.4, 104.8, 104.8, 59.7, 28.6, 22.4, 19.5, 19.4, 19.3, 19.2, 19.2, 19.1, 19.0, 18.9, 18.8, 18.8, 14.6.



**Diethyl 4-cyclohexyl-2,6-bis(methyl-d<sub>3</sub>)-1,4-dihydropyridine-3,5-dicarboxylate (2h-d<sub>6</sub>).** **2h** (67.1 mg, 0.2 mmol) and TFA-*d* (154  $\mu\text{L}$ , 2 mmol) were used according to general procedure 1 to obtain **2h-d<sub>6</sub>** as a pale yellow solid (58.7 mg, 0.172 mmol, 86%, 80% D).  $^1\text{H}$  NMR (400

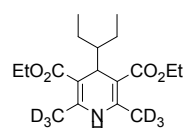
MHz,  $\text{CDCl}_3$ )  $\delta$  6.04 (s, 1H), 4.24 – 4.05 (m, 4H), 3.89 (d,  $J = 5.7$  Hz, 1H), 2.28 – 2.20 (m, 1.2H), 1.65 – 1.58 (m, 2H), 1.57 – 1.46 (m, 3H), 1.26 (t,  $J = 7.1$  Hz, 6H), 1.17 (dq,  $J = 6.0, 3.0$  Hz, 1H), 1.09 – 0.98 (m, 3H), 0.88 (qd,  $J = 13.6, 12.6, 5.1$  Hz, 2H).  $^{13}\text{C}$  NMR (101

MHz, CDCl<sub>3</sub>) δ 168.9, 144.7, 101.9, 59.6, 45.9, 38.4, 28.9, 26.8, 26.7, 19.4, 19.2, 19.0, 18.8, 18.6, 14.5.



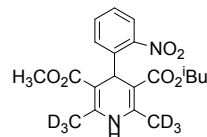
**Diethyl 4-isopropyl-2,6-bis(methyl-d<sub>3</sub>)-1,4-dihydropyridine-3,5-dicarboxylate (2i-d<sub>6</sub>).** 2i (59.1 mg, 0.2 mmol) and TFA-*d* (154 μL, 2 mmol) were used according to general procedure 1 to obtain 2i-d<sub>6</sub> as

a white solid (53.0 mg, 0.176 mmol, 88%, 77% D). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 5.92 (s, 1H), 4.15 (ddq, *J* = 39.2, 10.8, 7.1 Hz, 4H), 3.89 (d, *J* = 5.5 Hz, 1H), 2.28-2.22 (m, 1.4H), 1.56 (pd, *J* = 6.9, 5.4 Hz, 1H), 1.27 (t, *J* = 7.1 Hz, 6H), 0.72 (d, *J* = 6.9 Hz, 6H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 168.9, 144.8, 101.8, 59.7, 38.9, 35.6, 19.3, 19.2, 19.2, 19.1, 19.0, 18.9, 18.8, 18.6, 14.5.



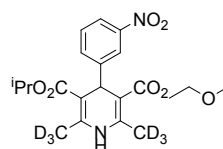
**Diethyl 2,6-bis(methyl-d<sub>3</sub>)-4-(pentan-3-yl)-1,4-dihydropyridine-3,5-dicarboxylate (2j-d<sub>6</sub>).** 2j (64.7 mg, 0.2 mmol) and TFA-*d* (154 μL, 2 mmol) were used according to general procedure 1 to obtain 2j-d<sub>6</sub> as

a white solid (55.4 mg, 0.168 mmol, 84%, 77% D). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 5.70 (s, 1H), 4.21 – 4.09 (m, 5H), 2.27 – 2.21 (m, 1.4H), 1.29 (t, *J* = 7.1 Hz, 6H), 1.14 (ddq, *J* = 20.9, 13.8, 7.1 Hz, 4H), 1.03 (qd, *J* = 6.5, 4.5 Hz, 1H), 0.85 (t, *J* = 7.3 Hz, 6H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 168.9, 144.7, 102.3, 59.7, 50.0, 34.6, 21.3, 19.4, 19.3, 19.2, 19.2, 19.1, 19.0, 18.8, 18.7, 18.6, 14.4, 11.9.



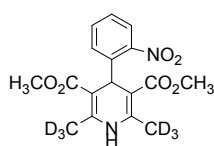
**3-isobutyl 5-methyl 2,6-bis(methyl-d<sub>3</sub>)-4-(2-nitrophenyl)-1,4-dihydropyridine-3,5-dicarboxylate (2k-d<sub>6</sub>).** 2k (77.7 mg, 0.2 mmol) and TFA-*d* (154 μL, 2 mmol) were used according to general procedure 1 to obtain 2k-d<sub>6</sub> as a yellow solid (74.9 mg, 0.190 mmol,

95%, 79% D). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.67 (dd, *J* = 8.1, 1.3 Hz, 1H), 7.51 (dd, *J* = 7.9, 1.5 Hz, 1H), 7.44 (td, *J* = 7.6, 1.3 Hz, 1H), 7.22 (ddd, *J* = 8.5, 7.2, 1.5 Hz, 1H), 6.14 (s, 1H), 5.76 (s, 1H), 3.83 – 3.75 (m, 2H), 3.56 (s, 3H), 2.34 – 2.27 (m, 0.6H), 2.27 – 2.20 (m, 0.6H), 1.87 (dt, *J* = 13.5, 6.7 Hz, 1H), 0.76 (d, *J* = 6.7 Hz, 3H), 0.71 (d, *J* = 6.7 Hz, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 167.8, 167.5, 147.9, 145.1, 145.0, 142.5, 132.9, 131.2, 127.1, 124.1, 103.9, 103.5, 70.5, 51.1, 34.7, 27.6, 19.1.

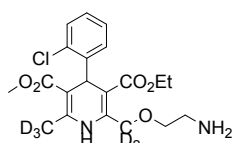


**3-isopropyl 5-(2-methoxyethyl) 2,6-bis(methyl-d<sub>3</sub>)-4-(3-nitrophenyl)-1,4-dihydropyridine-3,5-dicarboxylate (2l-d<sub>6</sub>).** 2l

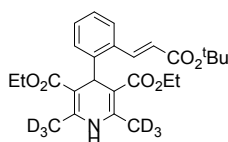
(83.7 mg, 0.2 mmol) and TFA-*d* (154 μL, 2 mmol) were used according to general procedure 1 to obtain 2l-d<sub>6</sub> as a pale yellow solid (79.8 mg, 0.188 mmol, 94%, 68% D). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.12 (t, *J* = 2.0 Hz, 1H), 7.98 (ddd, *J* = 8.2, 2.3, 1.1 Hz, 1H), 7.65 (dt, *J* = 7.7, 1.4 Hz, 1H), 7.36 (t, *J* = 7.9 Hz, 1H), 6.04 (s, 1H), 5.08 (s, 1H), 4.93 (p, *J* = 6.2 Hz, 1H), 4.22 – 4.10 (m, 2H), 3.59 – 3.47 (m, 2H), 3.33 (s, 3H), 2.35 – 2.28 (m, 2H), 1.24 (d, *J* = 6.2 Hz, 3H), 1.07 (d, *J* = 6.2 Hz, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 167.2, 166.7, 150.1, 148.2, 145.3, 144.5, 134.8, 128.7, 123.4, 121.4, 104.0, 103.1, 70.6, 67.4, 63.1, 59.0, 40.1, 22.2, 21.9, 19.6, 19.5, 19.4, 19.4, 19.3, 19.2, 19.1, 19.0.



**Dimethyl 2,6-bis(methyl-d3)-4-(2-nitrophenyl)-1,4-dihydropyridine-3,5-dicarboxylate (2m-d<sub>6</sub>).** **2m** (69.3 mg, 0.2 mmol) and TFA-*d* (154 μL, 2 mmol) were used according to general procedure 1 to obtain **2m-d<sub>6</sub>** as a yellow solid (67.7 mg, 0.192 mmol, 96%, 79% D). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.65 (dd, *J* = 8.1, 1.3 Hz, 1H), 7.50 (dd, *J* = 7.9, 1.5 Hz, 1H), 7.44 (td, *J* = 7.6, 1.4 Hz, 1H), 7.23 (ddd, *J* = 8.4, 7.2, 1.5 Hz, 1H), 6.15 (s, 1H), 5.70 (s, 1H), 3.57 (s, 6H), 2.32 – 2.23 (m, 1.3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 167.7, 147.9, 145.2, 145.2, 142.3, 132.9, 131.1, 127.1, 124.0, 103.6, 103.6, 51.1, 34.6, 19.4, 19.3, 19.2, 19.2, 19.1, 19.0, 18.9, 18.8, 18.8, 18.7, 18.6.



**3-ethyl 5-methyl 2-((2-aminoethoxy)methyl-d2)-4-(2-chlorophenyl)-6-(methyl-d3)-1,4-dihydropyridine-3,5-dicarboxylate (2n-d<sub>5</sub>).** Amlodipine besylate (113.4 mg, 0.2 mmol) and TFA-*d* (154 μL, 2 mmol) were used according to general procedure 1 to obtain **2n-d<sub>5</sub>** as a white solid (79.9 mg, 0.194 mmol, 97%, 80% D, 77% D). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.78 (s, 1H), 7.37 (dd, *J* = 7.8, 1.7 Hz, 1H), 7.20 (dd, *J* = 8.0, 1.4 Hz, 1H), 7.11 (td, *J* = 7.5, 1.4 Hz, 1H), 7.01 (td, *J* = 7.5, 1.7 Hz, 1H), 5.38 (s, 1H), 5.07 (s, 2H), 4.81 – 4.67 (m, 0.5H), 4.07 – 3.96 (m, 2H), 3.68 (hept, *J* = 5.2 Hz, 2H), 3.58 (s, 3H), 3.09 (t, *J* = 4.7 Hz, 2H), 2.33 (d, *J* = 11.0 Hz, 0.6H), 1.16 (t, *J* = 7.2 Hz, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 168.3, 167.4, 145.9, 145.2, 145.1, 145.1, 144.8, 132.4, 131.6, 129.3, 127.5, 127.0, 103.8, 102.2, 102.2, 102.1, 70.0, 68.2, 60.0, 50.9, 40.5, 37.3, 18.7, 14.3.

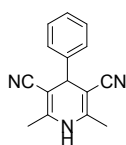


**Diethyl (E)-4-(2-(3-(tert-butoxy)-3-oxoprop-1-en-1-yl)phenyl)-2,6-bis(methyl-d3)-1,4-dihydropyridine-3,5-dicarboxylate (2o-d<sub>6</sub>).** **2o** (91.1 mg, 0.2 mmol) and TFA-*d* (154 μL, 2 mmol) were used according to general procedure 1 to obtain **2o-d<sub>6</sub>** as a white solid (89.5 mg, 0.194 mmol, 97%, 80% D). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.44 (d, *J* = 15.8 Hz, 1H), 7.46 (dd, *J* = 7.9, 1.4 Hz, 1H), 7.40 (dd, *J* = 7.9, 1.3 Hz, 1H), 7.23 (td, *J* = 7.6, 1.4 Hz, 1H), 7.10 (td, *J* = 7.6, 1.4 Hz, 1H), 6.25 (d, *J* = 15.9 Hz, 1H), 5.99 (s, 1H), 5.32 (s, 1H), 4.05 (dq, *J* = 10.8, 7.2 Hz, 2H), 3.93 (dq, *J* = 10.8, 7.1 Hz, 2H), 2.31 – 2.24 (m, 1.2H), 1.53 (s, 9H), 1.13 (t, *J* = 7.1 Hz, 6H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 167.6, 166.8, 148.5, 144.3, 144.3, 143.9, 132.0, 130.6, 130.1, 126.5, 125.5, 120.2, 104.7, 104.6, 80.2, 59.8, 35.8, 28.4, 19.2, 19.1, 19.0, 18.9, 18.8, 18.6, 14.4.

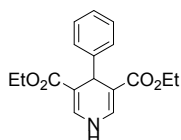
## 6. Controlled experiments

**3c** and **3d** were oxidated according to general procedure **3** and then deuterated according to general procedure 1 using 10 eq. TFA-*d*. Deuteration of **3e**, **3f**, **3g** and **3h** were operated according to general procedure 1 in CH<sub>2</sub>Cl<sub>2</sub> and 1.5eq TFA-*d* was used for each reaction. When CH<sub>2</sub>Cl<sub>2</sub> was used as the solvent, the reaction mixture was degassed by N<sub>2</sub> gently purging the vial for 3 minutes. After 24 h reaction under 50°C,

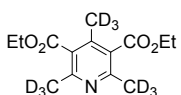
cool to RT.  $\text{CDCl}_3$  was added to dilute the mixture and  $^1\text{H}$  NMR was done without further purification to determine D incorporation. **3e** and **3f** were rarely deuterated after reaction. **3g** was deuterated in 17% D content.



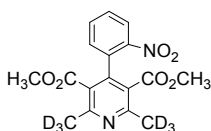
**2,6-dimethyl-4-phenyl-1,4-dihydropyridine-3,5-dicarbonitrile (3a-d<sub>0</sub>).** **3a** (47.1 mg, 0.2 mmol) and TFA-*d* (154  $\mu\text{L}$ , 2 mmol) were used according to general procedure 1. No D incorporation was found. **3a** was 98% recovered.



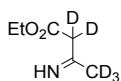
**Diethyl 4-phenyl-1,4-dihydropyridine-3,5-dicarboxylate (3b-d<sub>0</sub>).** **3b** (60.3 mg, 0.2 mmol) and TFA-*d* (154  $\mu\text{L}$ , 2 mmol) were used according to general procedure 1. No D incorporation was found. **3b** was 92% recovered.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.36 – 7.33 (m, 2H), 7.29 – 7.23 (m, 4H), 7.18 – 7.11 (m, 2H), 4.89 (s, 1H), 4.14 – 4.00 (m, 5H), 1.19 (t,  $J$  = 7.2 Hz, 6H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  167.5, 147.1, 134.2, 128.3, 128.0, 126.5, 108.1, 60.1, 37.7, 14.2.



**Diethyl 2,4,6-tris(methyl-d<sub>3</sub>)pyridine-3,5-dicarboxylate (3c-d<sub>9</sub>).** **3c** (53.1 mg, 0.2 mmol) and TFA-*d* (154  $\mu\text{L}$ , 2 mmol) were used according to general procedure 1 to obtain **3c-d<sub>9</sub>** as a colorless oil (53.8 mg, 0.196 mmol, 98%, 70% D, 78% D).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  4.38 (q,  $J$  = 7.1 Hz, 4H), 2.50 – 2.44 (m, 1.8H), 2.24 – 2.20 (m, 0.7H), 1.36 (t,  $J$  = 7.1 Hz, 6H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  168.4, 155.0, 142.2, 127.7, 61.7, 23.0, 22.9, 22.8, 22.7, 22.6, 22.6, 22.5, 22.4, 22.3, 22.2, 22.2, 17.0, 16.9, 16.8, 16.7, 16.7, 16.6, 16.4, 16.3, 16.3, 16.2, 14.3.



**Dimethyl 2,6-bis(methyl-d<sub>3</sub>)-4-(2-nitrophenyl)pyridine-3,5-dicarboxylate (3d-d<sub>6</sub>).** **3d** (68.9 mg, 0.2 mmol) and TFA-*d* (154  $\mu\text{L}$ , 2 mmol) were used according to general procedure 1 to obtain **3d-d<sub>6</sub>** as a white solid (68.0 mg, 0.194 mmol, 97%, 78% D).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  8.18 (dd,  $J$  = 8.1, 1.3 Hz, 1H), 7.62 (td,  $J$  = 7.5, 1.4 Hz, 1H), 7.55 (ddd,  $J$  = 8.9, 7.6, 1.5 Hz, 1H), 7.18 (dd,  $J$  = 7.5, 1.5 Hz, 1H), 3.48 (s, 6H), 2.64 – 2.59 (m, 1.3H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  167.3, 157.1, 147.7, 145.3, 133.0, 132.1, 130.7, 129.7, 124.9, 124.4, 52.3, 23.6, 23.5, 23.5, 23.4, 23.3, 23.2, 23.1, 23.1, 23.0, 22.9, 22.8.



**Ethyl 3-iminobutanoate-2,2,4,4-d<sub>5</sub> (3h'-d<sub>5</sub>).** **3h** (12.9 mg, 0.1 mmol), TFA-*d* (23  $\mu\text{L}$ , 0.3 mmol),  $\text{D}_2\text{O}$  (90  $\mu\text{L}$ , 5 mmol) and  $\text{CH}_2\text{Cl}_2$  (1 mL) were added in a 10 mL schlenk tube.  $\text{N}_2$  purged the headspace of the vial for 3 minutes following by tightly sealed. The reaction mixture was heated at 50°C for 24 h. Cool to room temperature and dilute with 1 mL  $\text{CDCl}_3$  directly for  $^1\text{H}$  NMR.  $^1\text{H}$  NMR (500 MHz, Chloroform-*d*)  $\delta$  4.16 (q,  $J$  = 7.1 Hz, 2H), 3.43 – 3.40 (m, 0.1H), 2.23 (s, 2.3H), 1.24 (t,  $J$  = 7.1 Hz, 3H).

Research on reversibility of HIE reaction. **2m-d<sub>6</sub>** (35.2 mg, 0.1 mmol),  $\text{H}_2\text{O}$  (90  $\mu\text{L}$ , 5 mmol), TFA (75  $\mu\text{L}$ , 1 mmol) and NMP were added in a 10 mL schlenk tube. The mixture

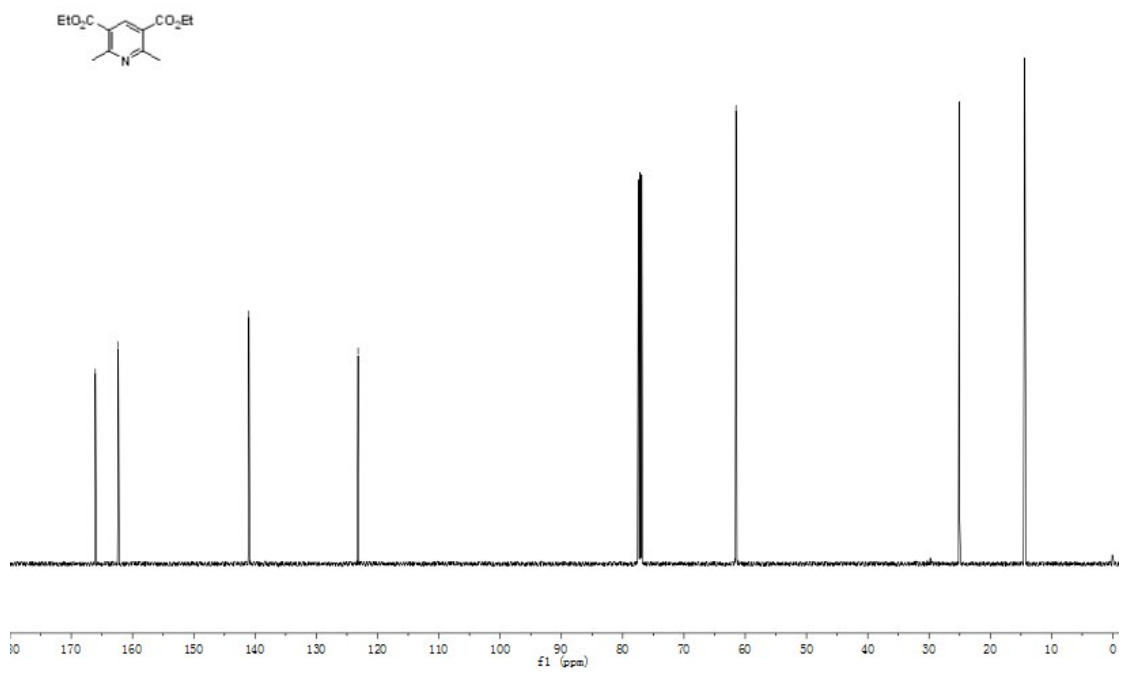
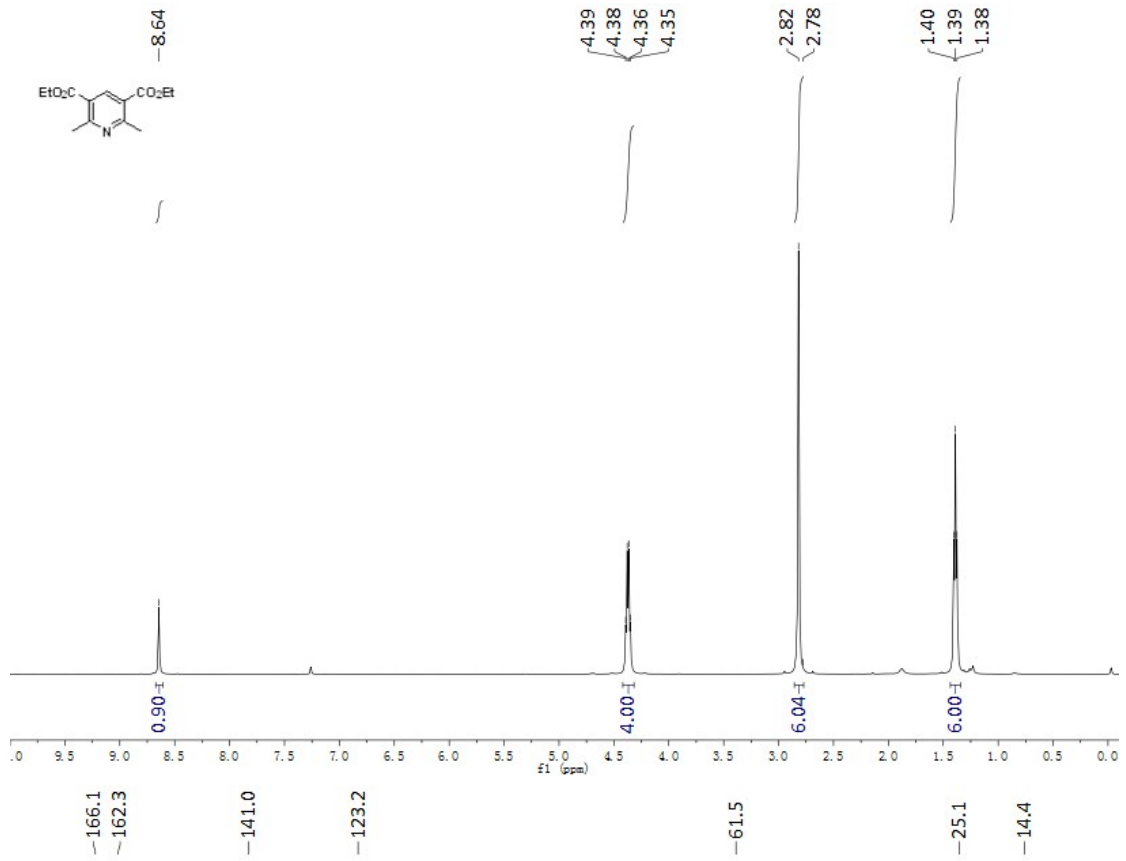
was then treated according to general procedure 1 and reacted at 70°C for 48 h. After general work-up and purification, <sup>1</sup>H NMR was done to show the D incorporation decreased to 12% which meant the HIE reaction was reversible to a great extent. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.67 (dd, *J* = 8.2, 1.3 Hz, 1H), 7.50 (dd, *J* = 7.9, 1.5 Hz, 1H), 7.45 (td, *J* = 8.0, 7.6, 1.4 Hz, 1H), 7.29 – 7.21 (m, 2H), 5.78 (s, 1H), 5.72 (s, 1H), 3.59 (s, 6H), 2.34 (s, 5.3H).

## 7. References

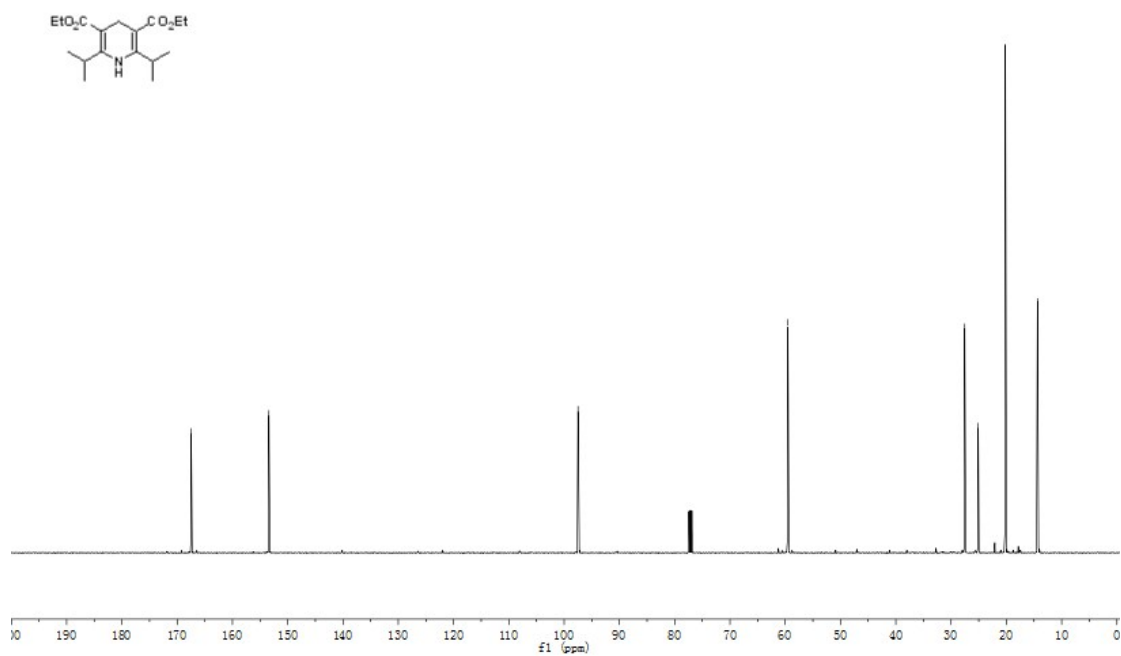
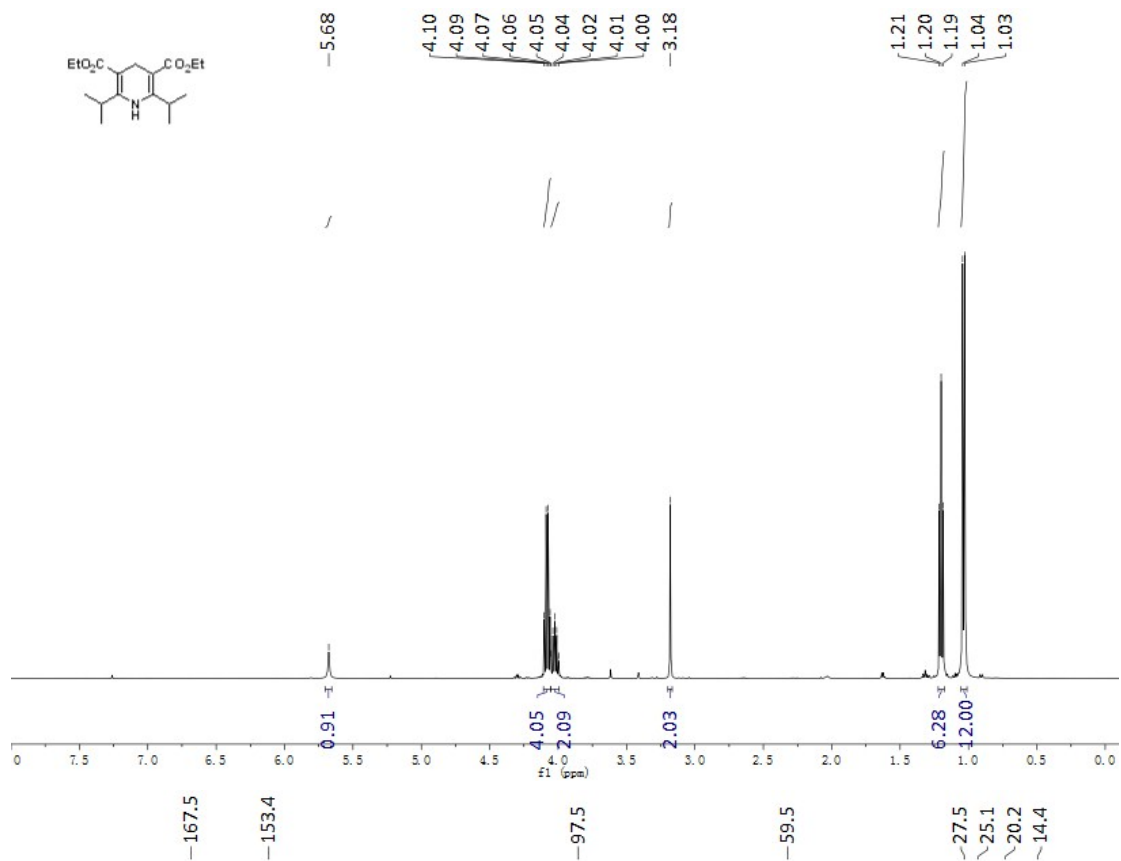
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## 8. NMR spectra

**1a'** (oxidative product of **1a**)

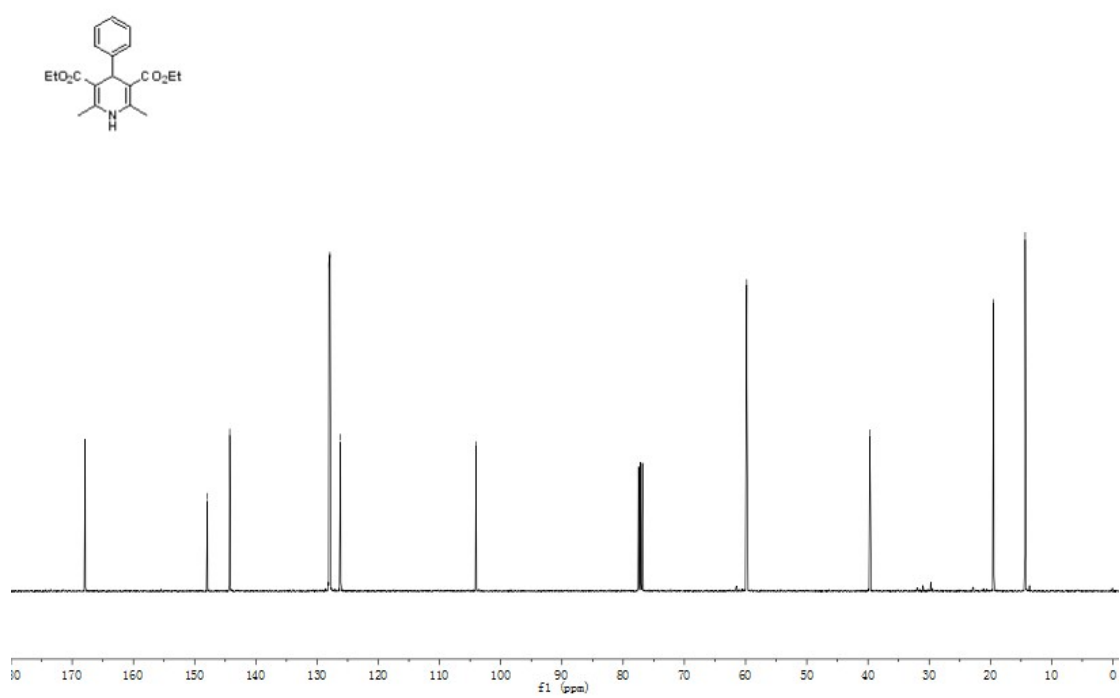
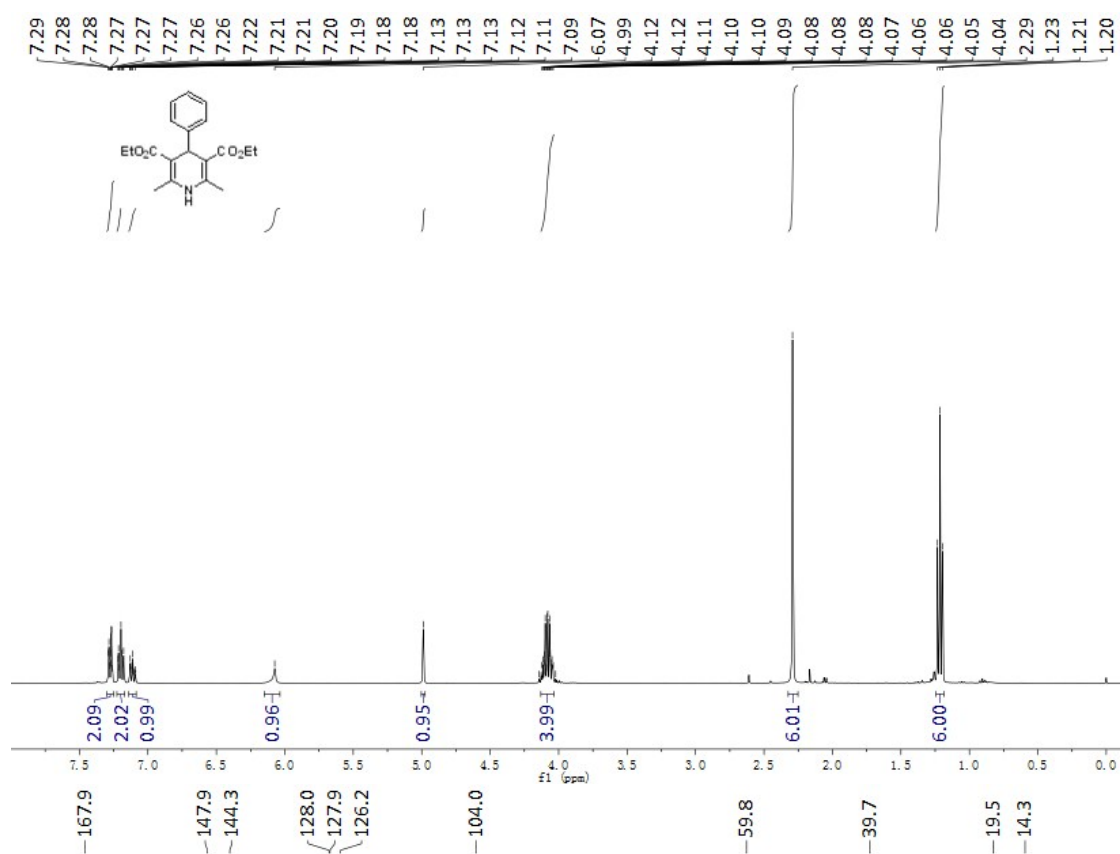


1e

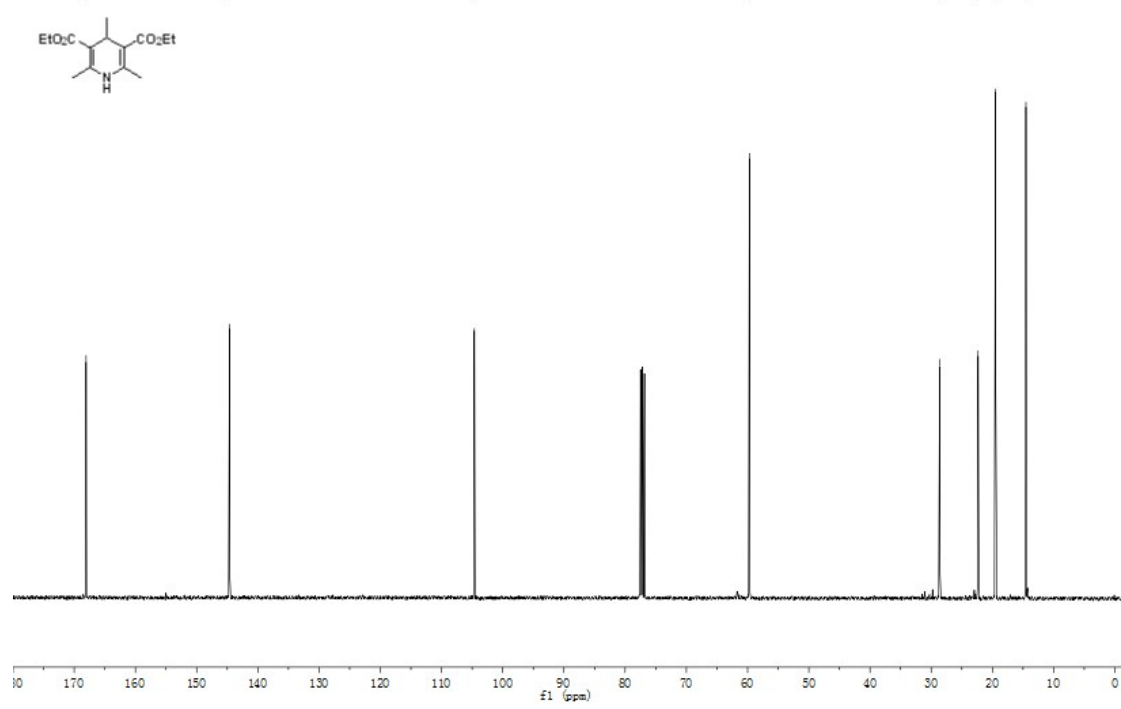
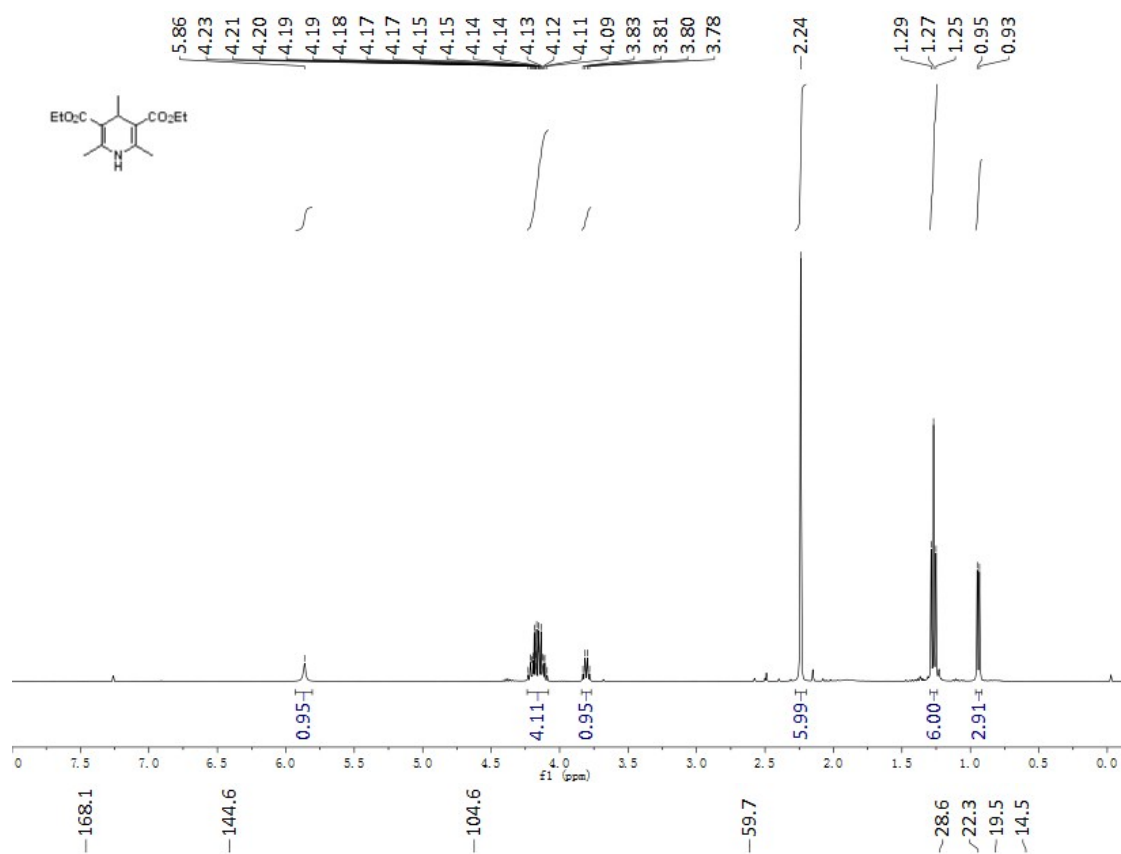




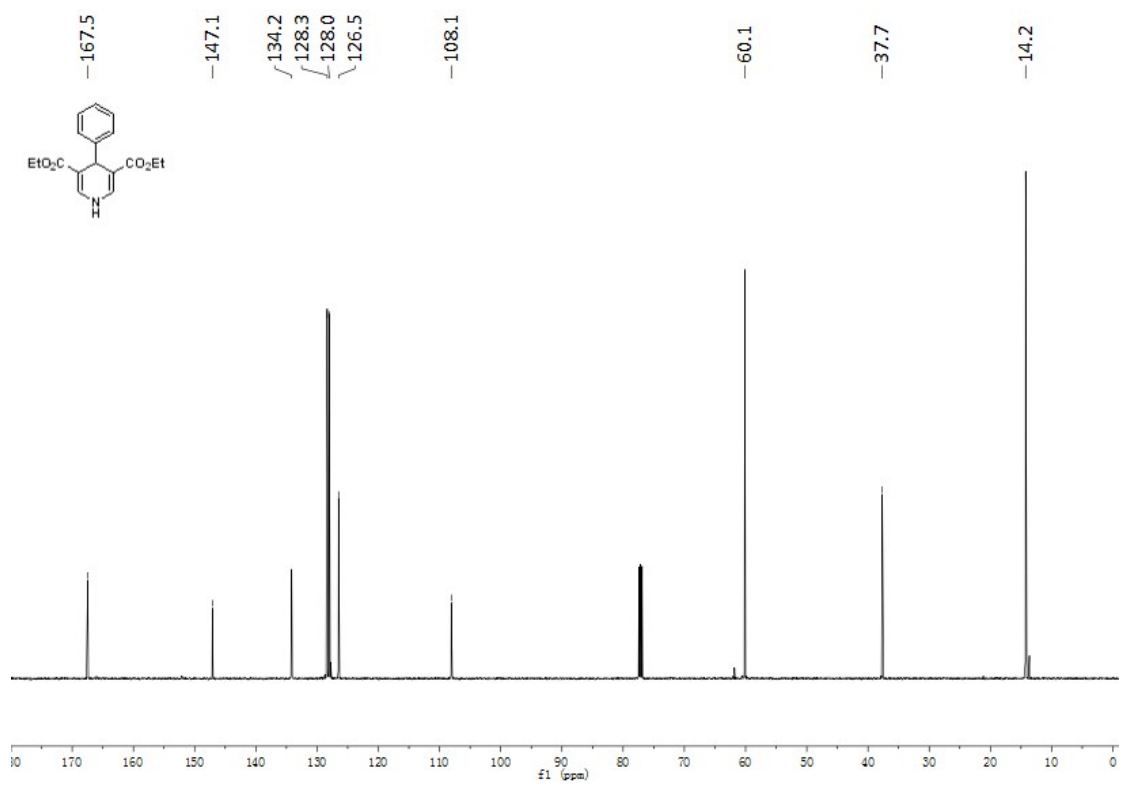
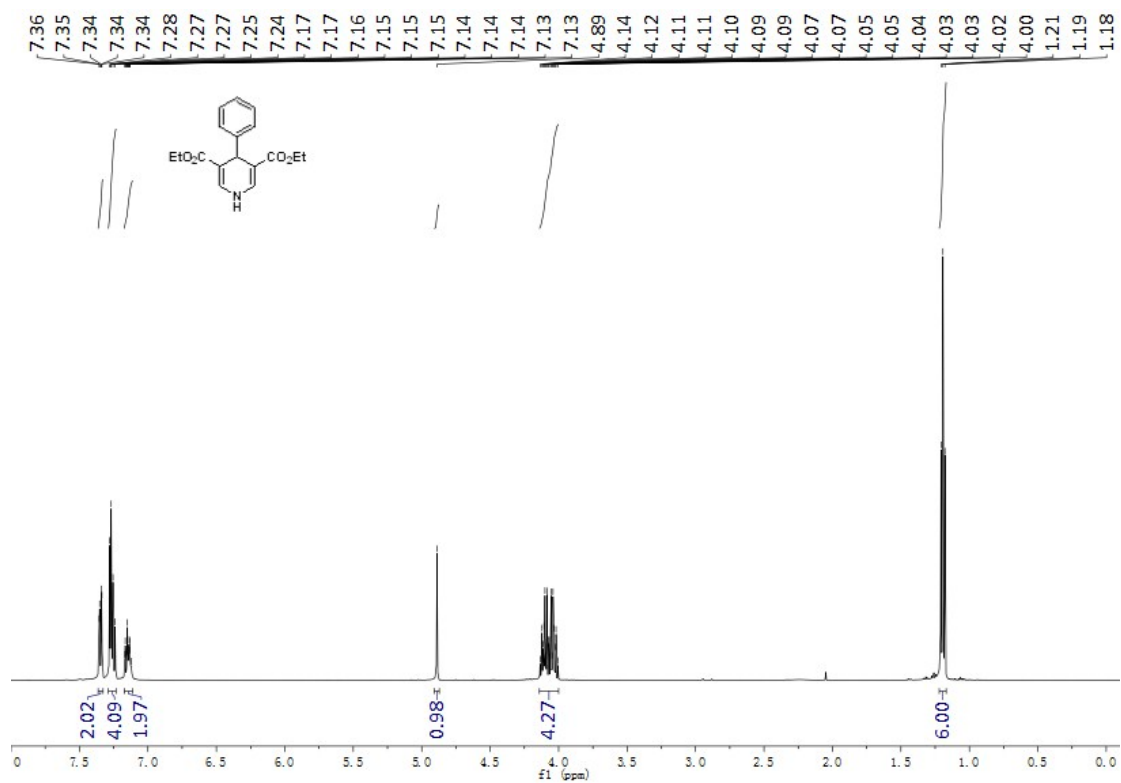
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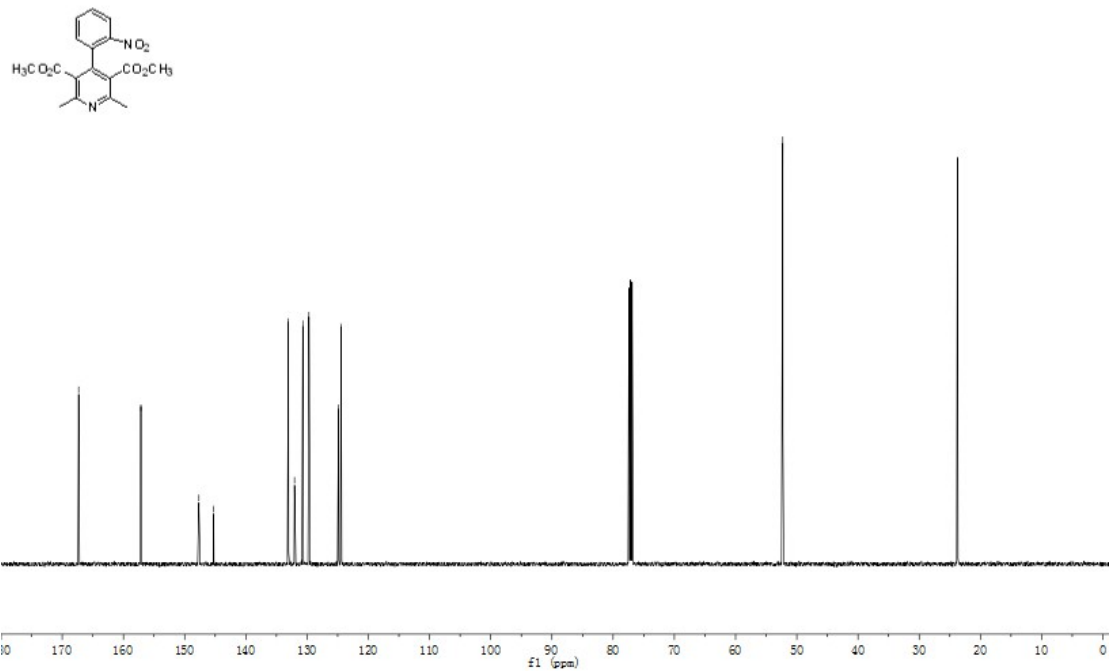
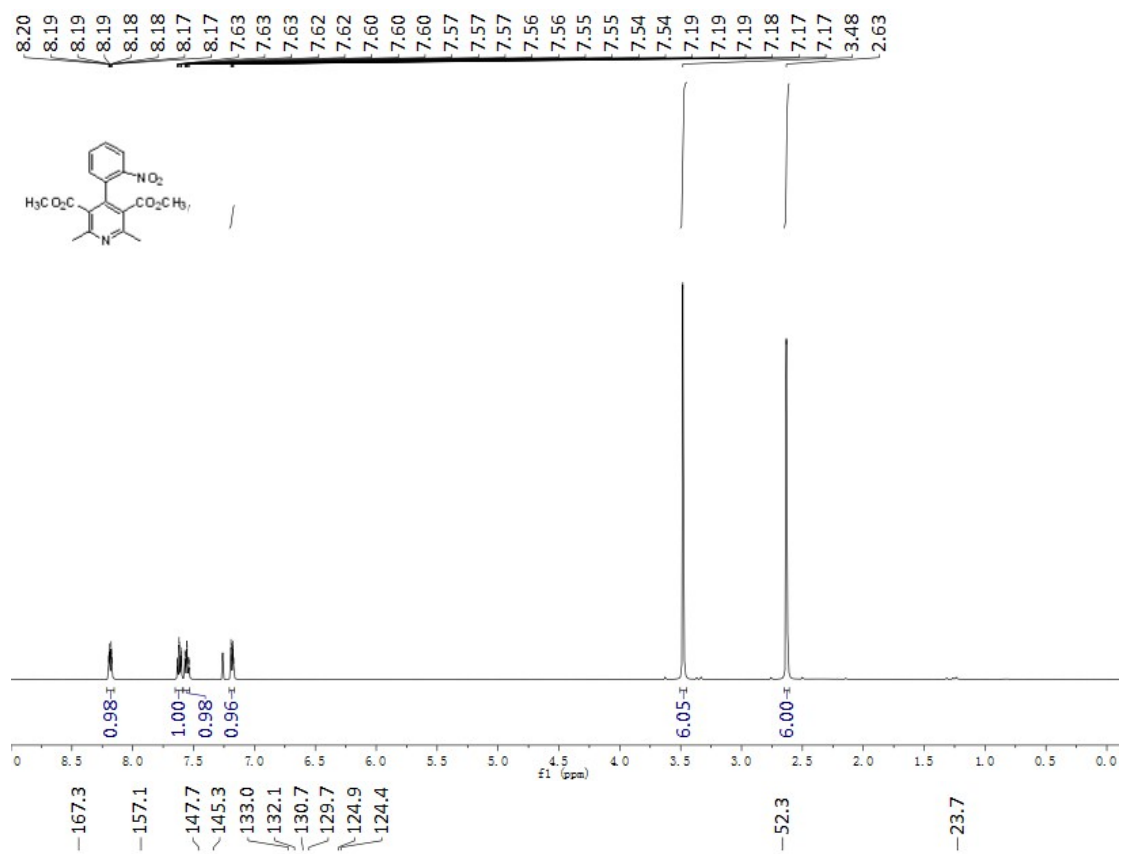
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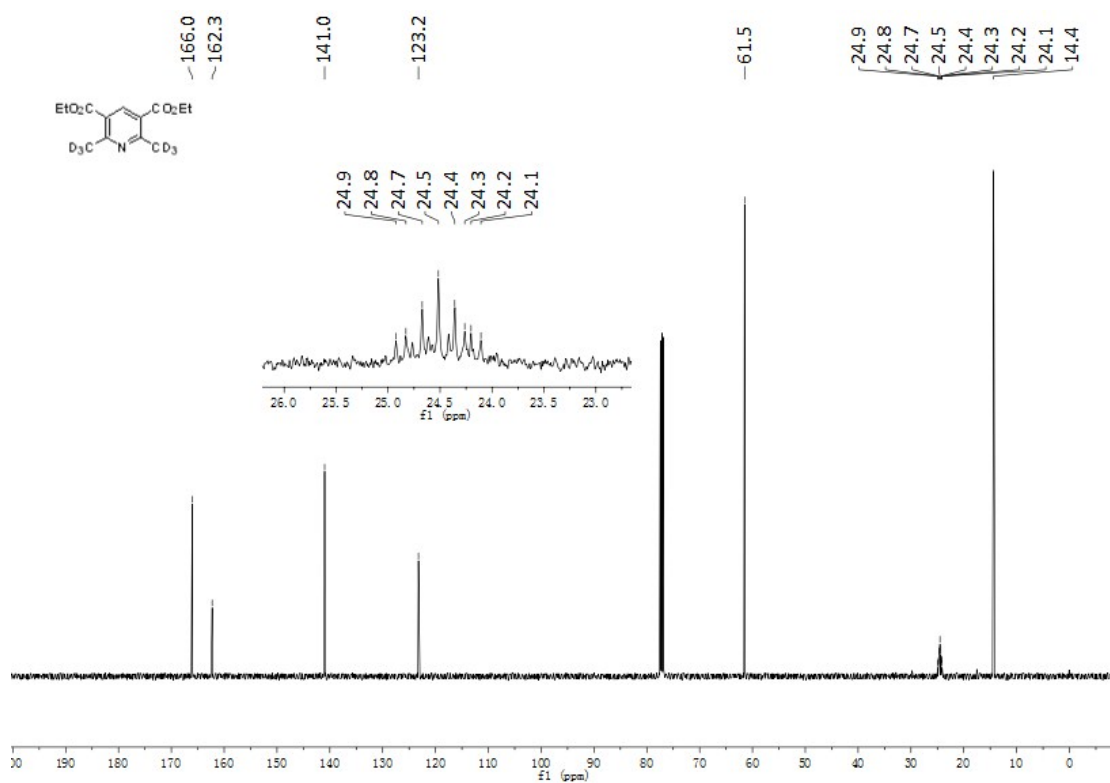
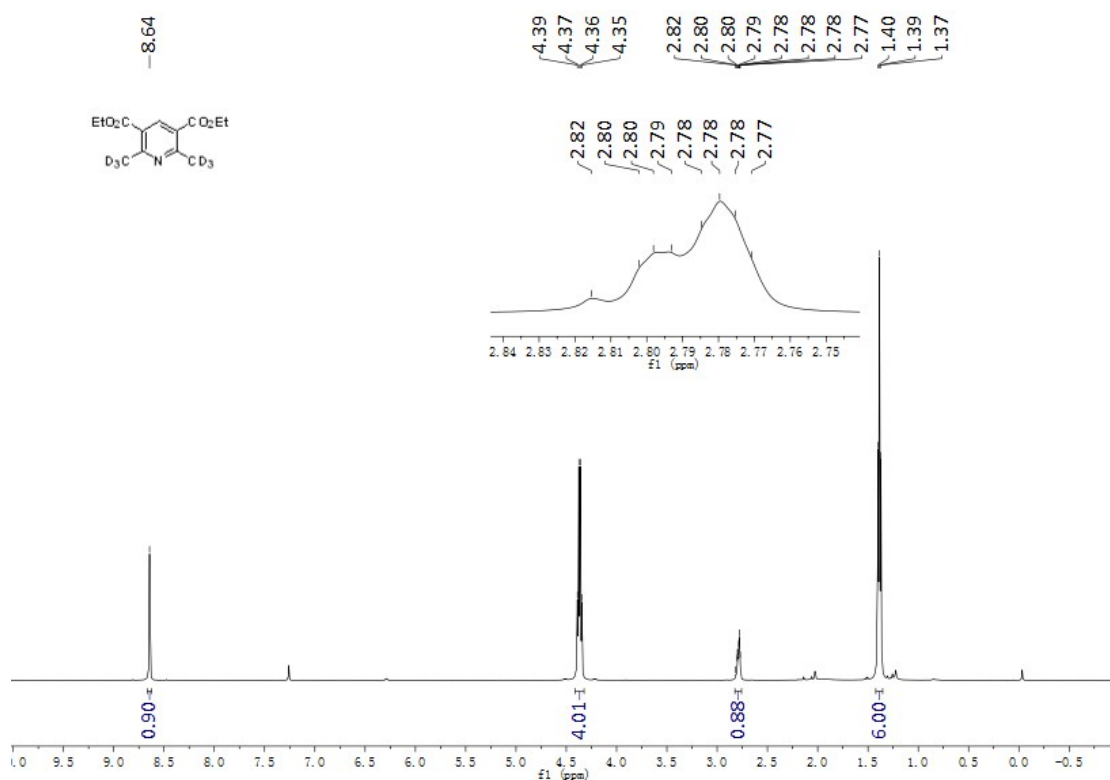
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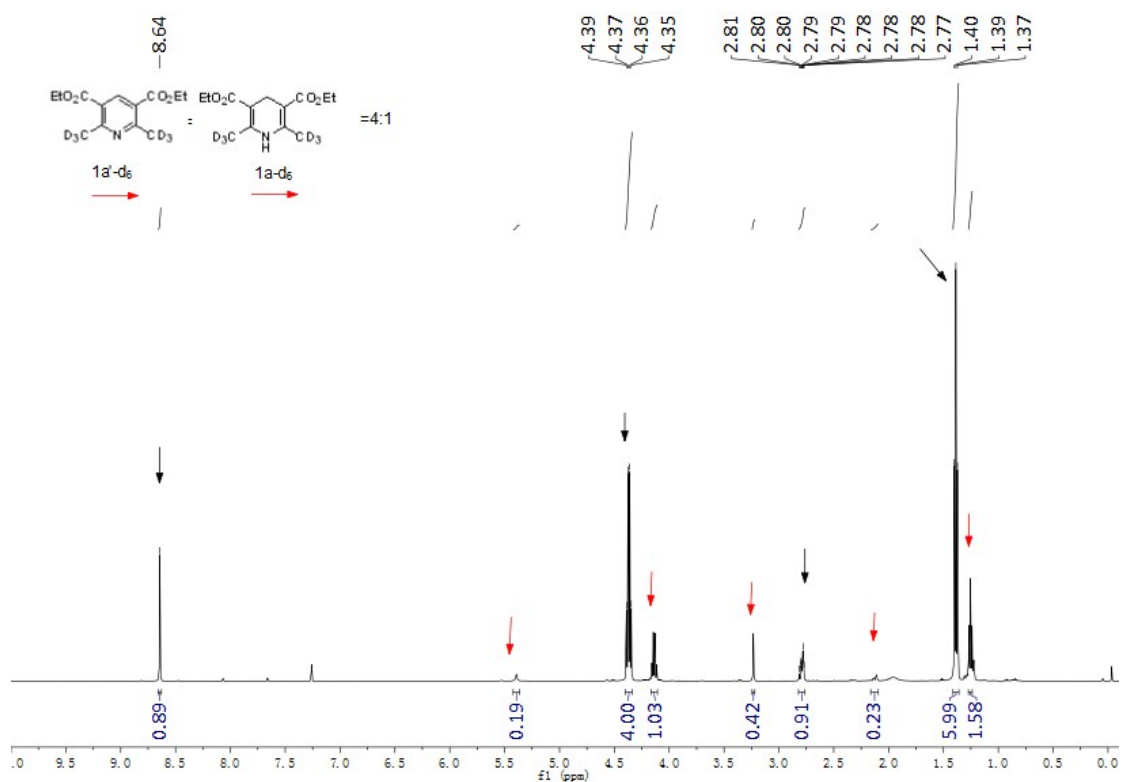
3d



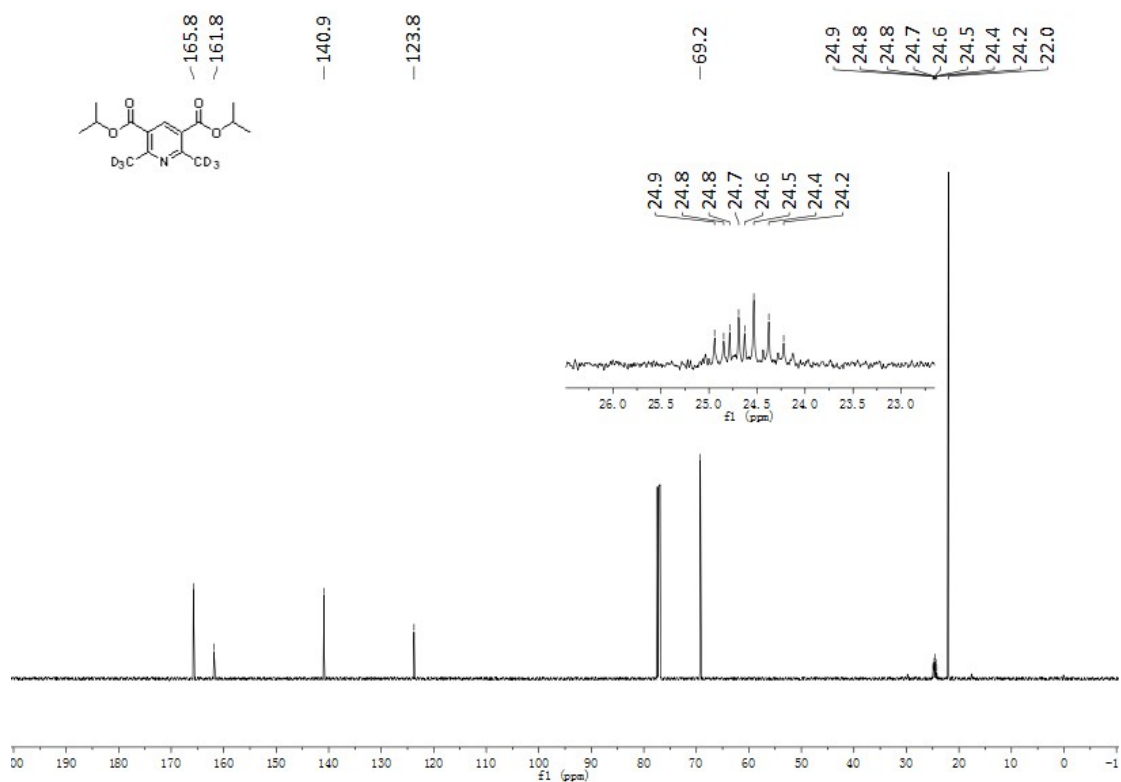
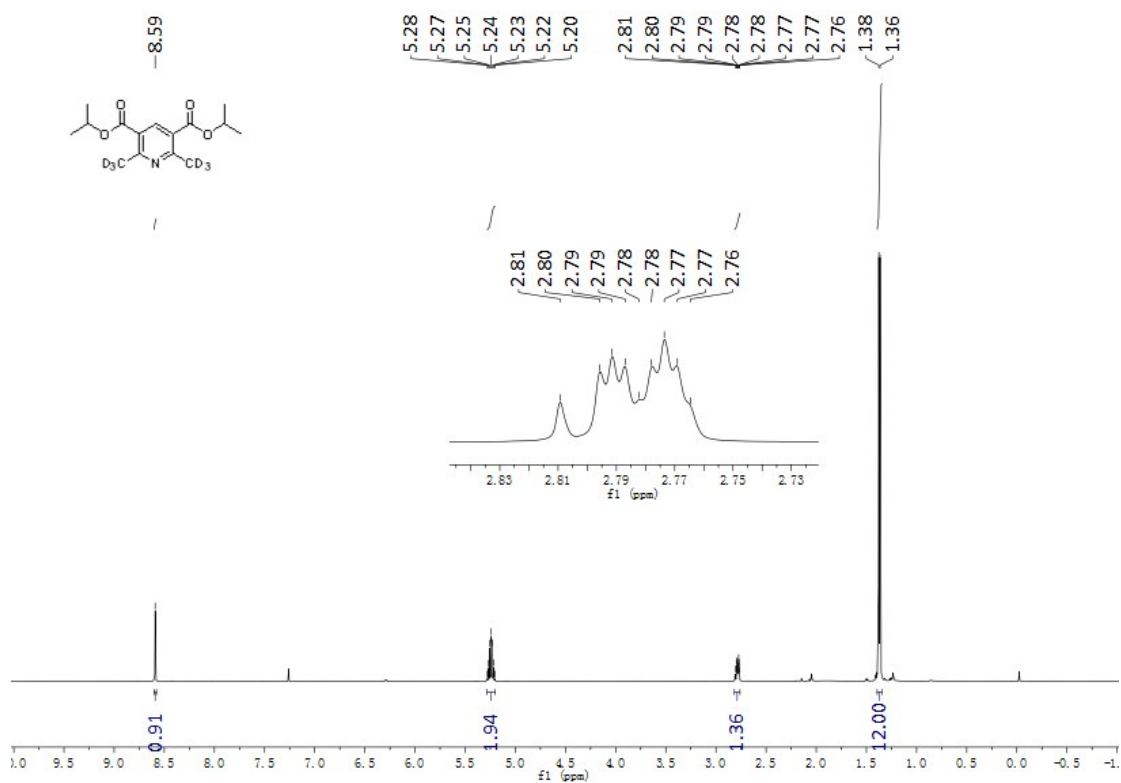
# 1a'-d<sub>6</sub>



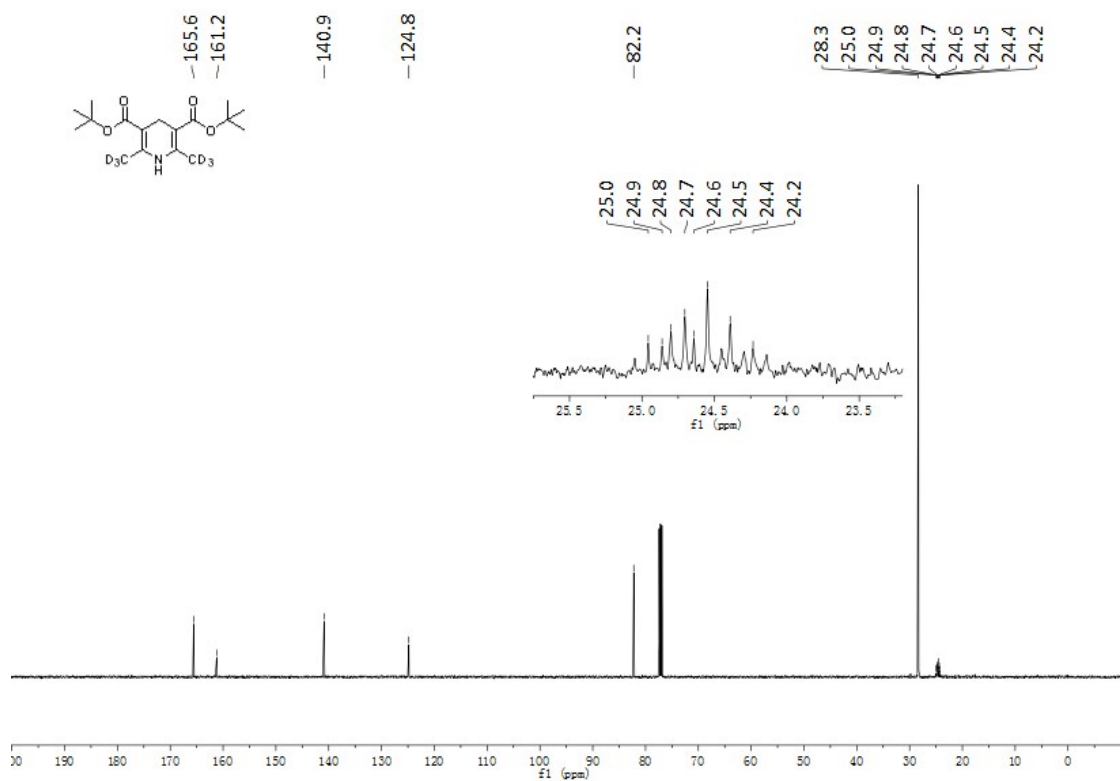
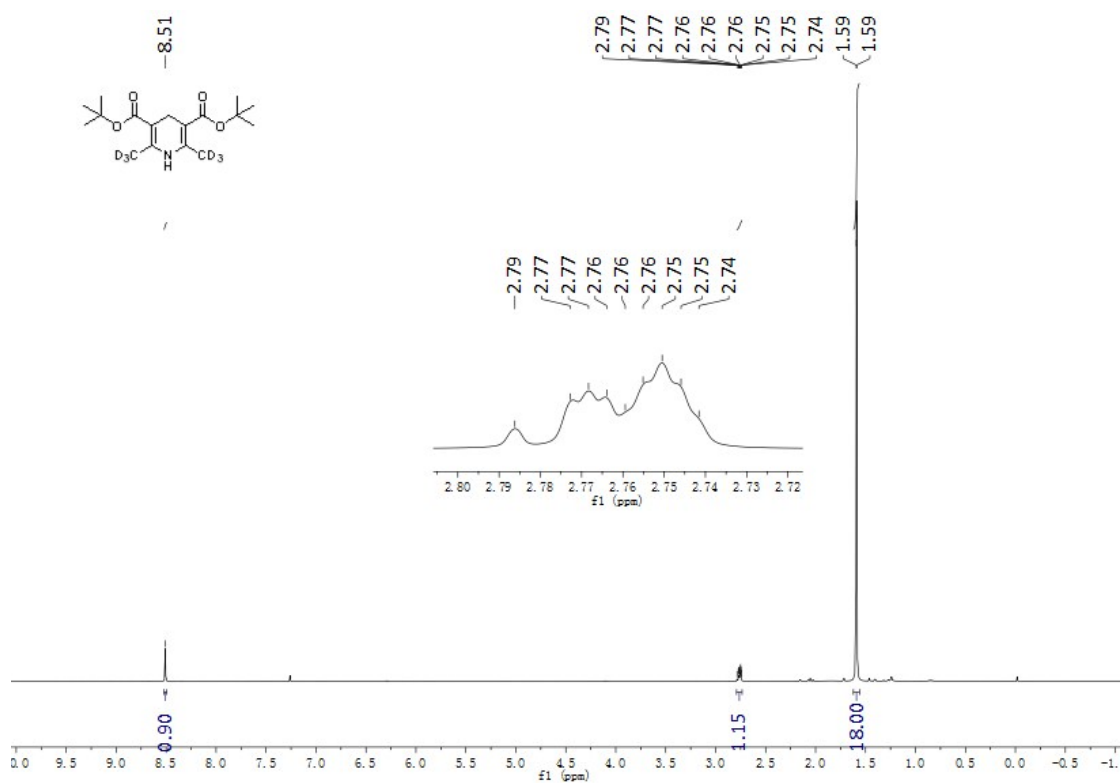
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**1b'-d<sub>6</sub>**

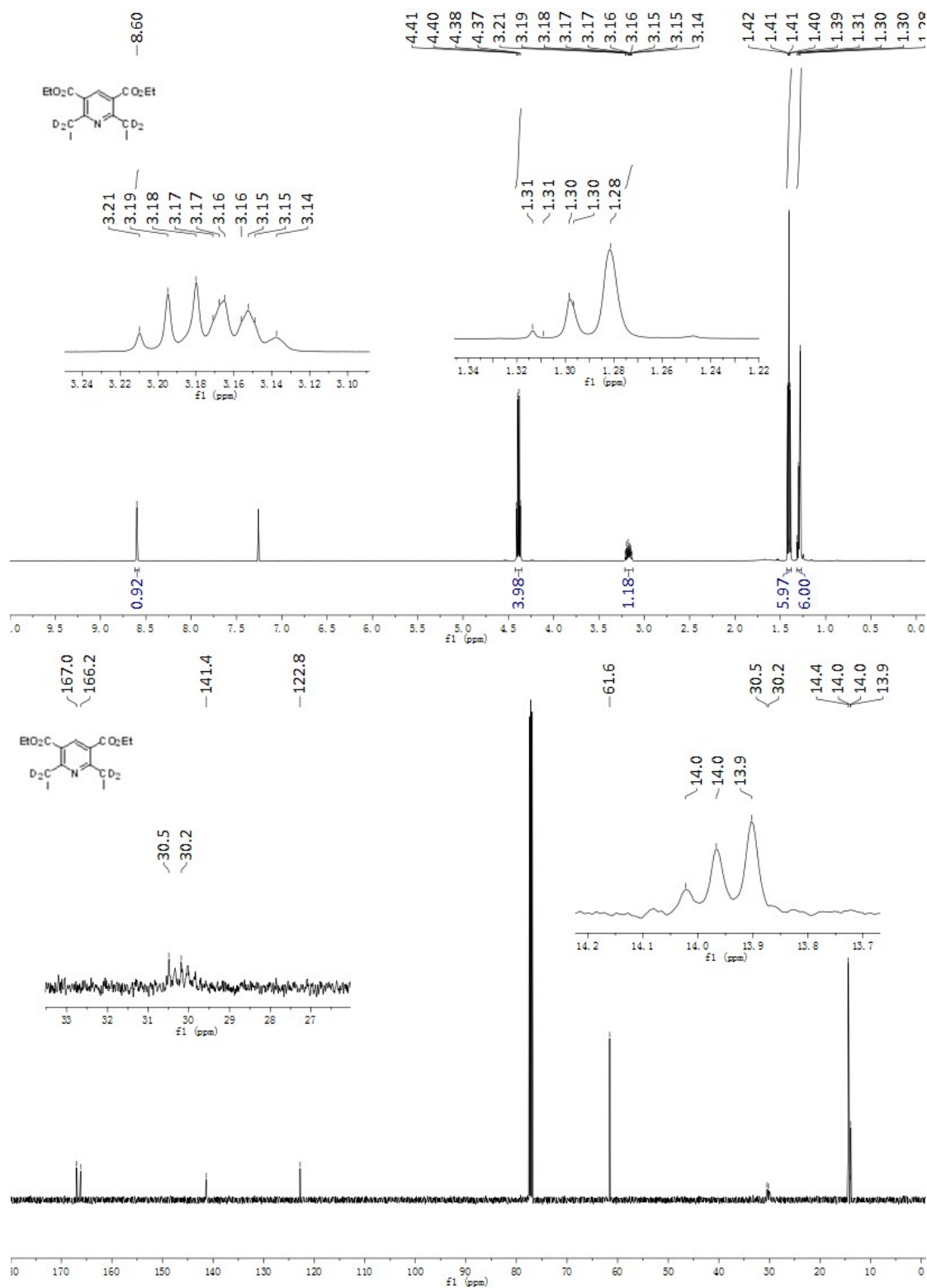


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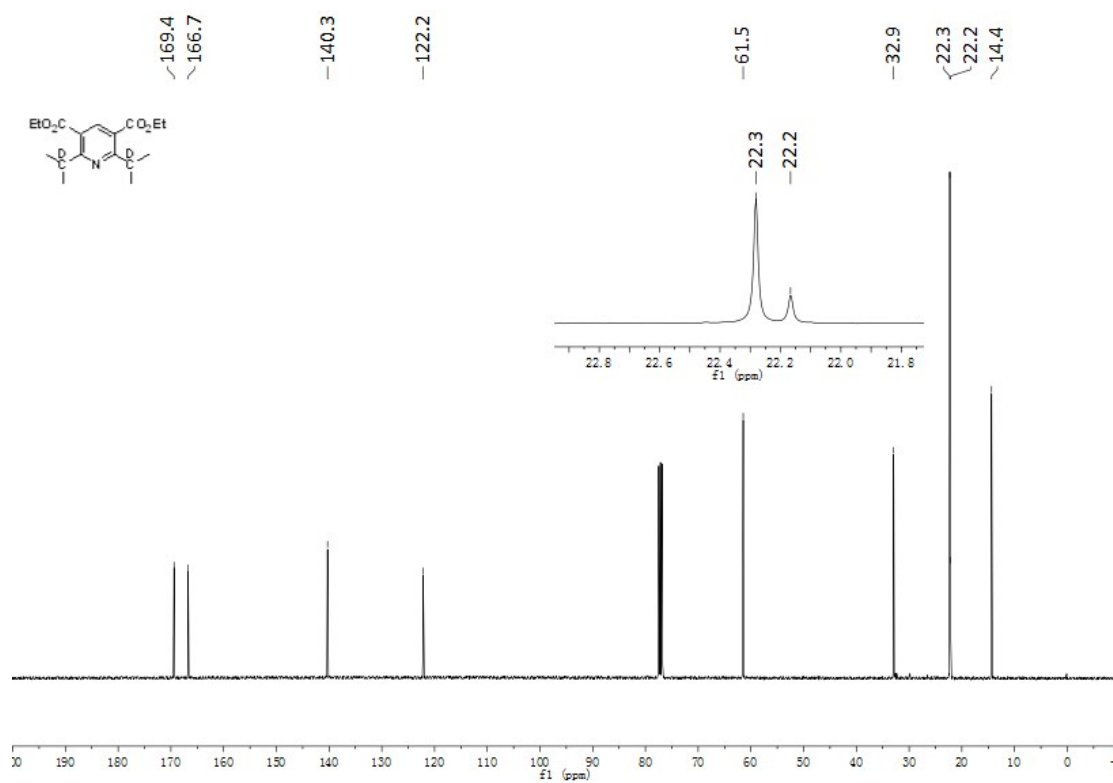
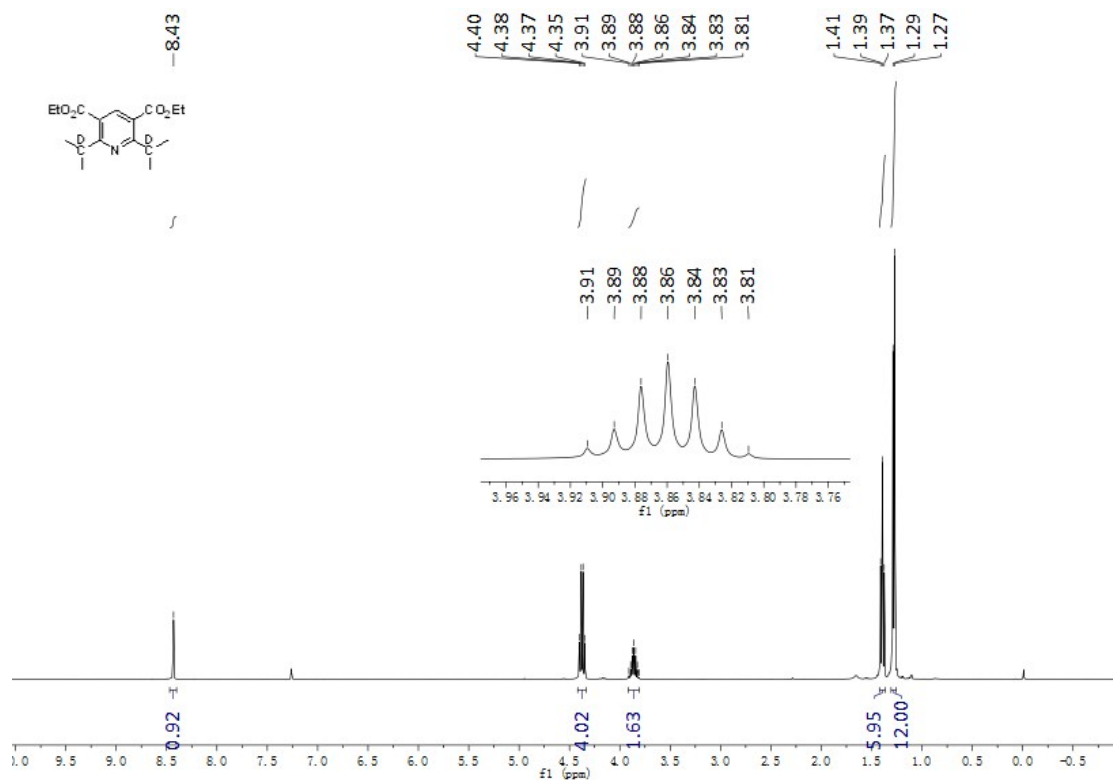




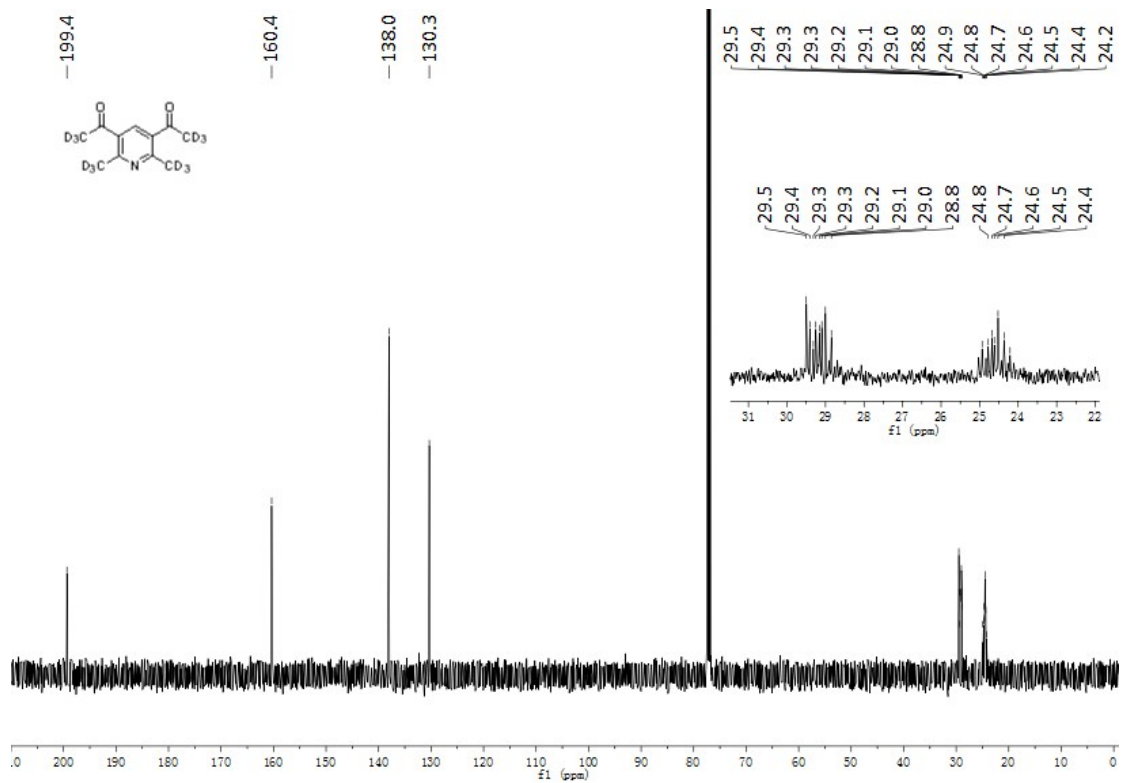
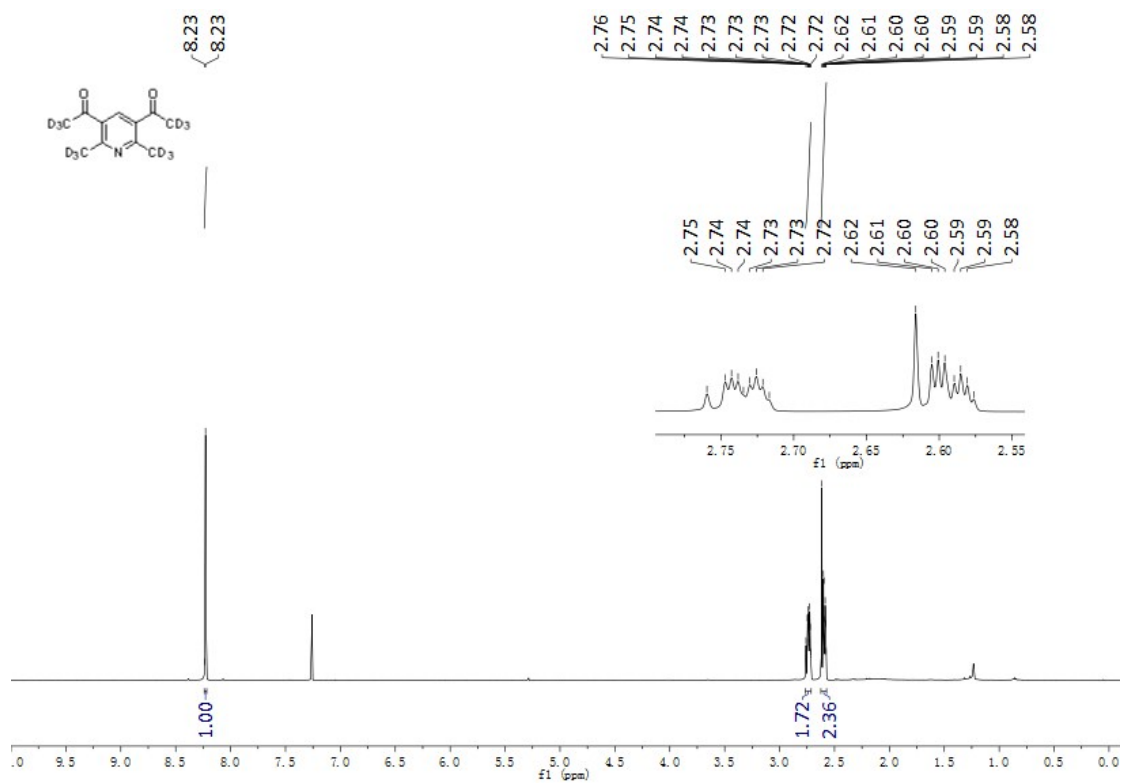
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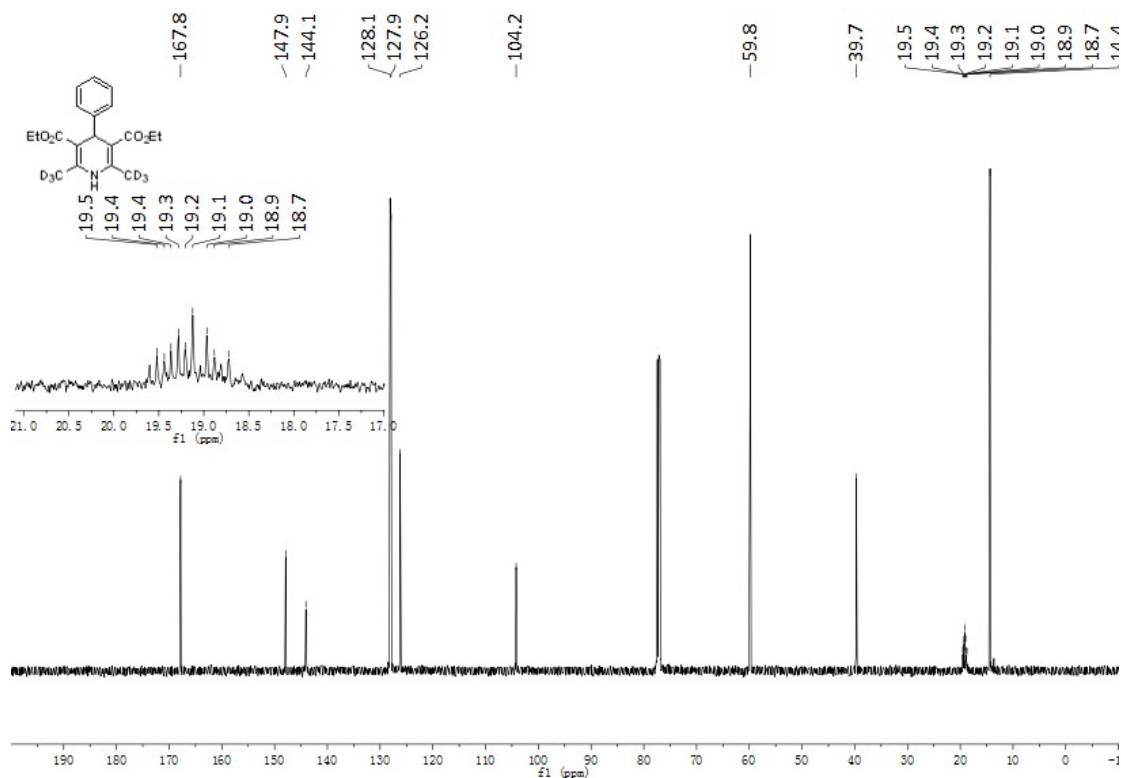
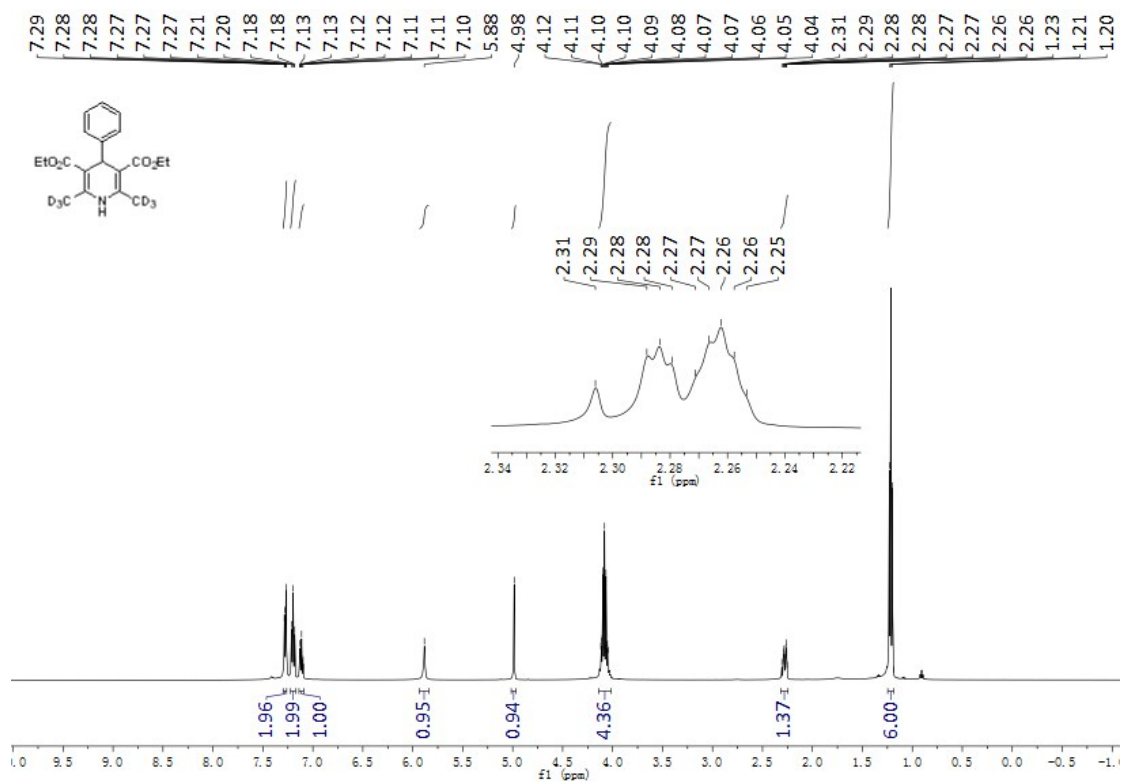
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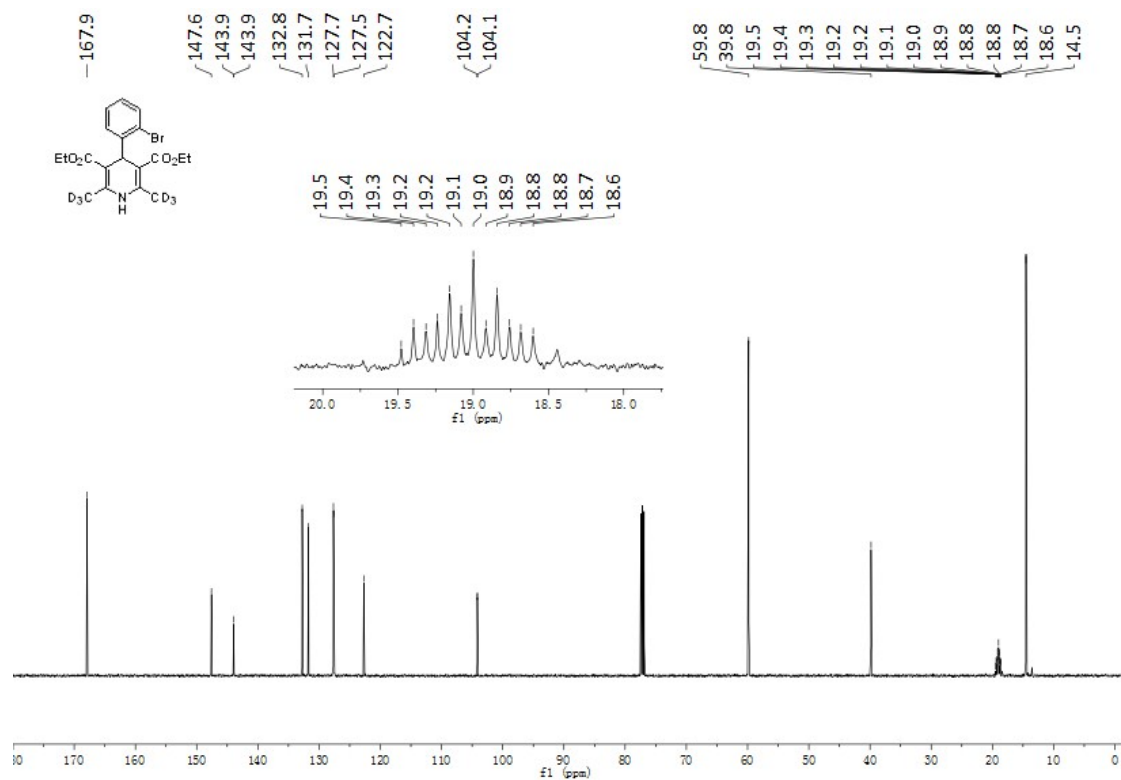
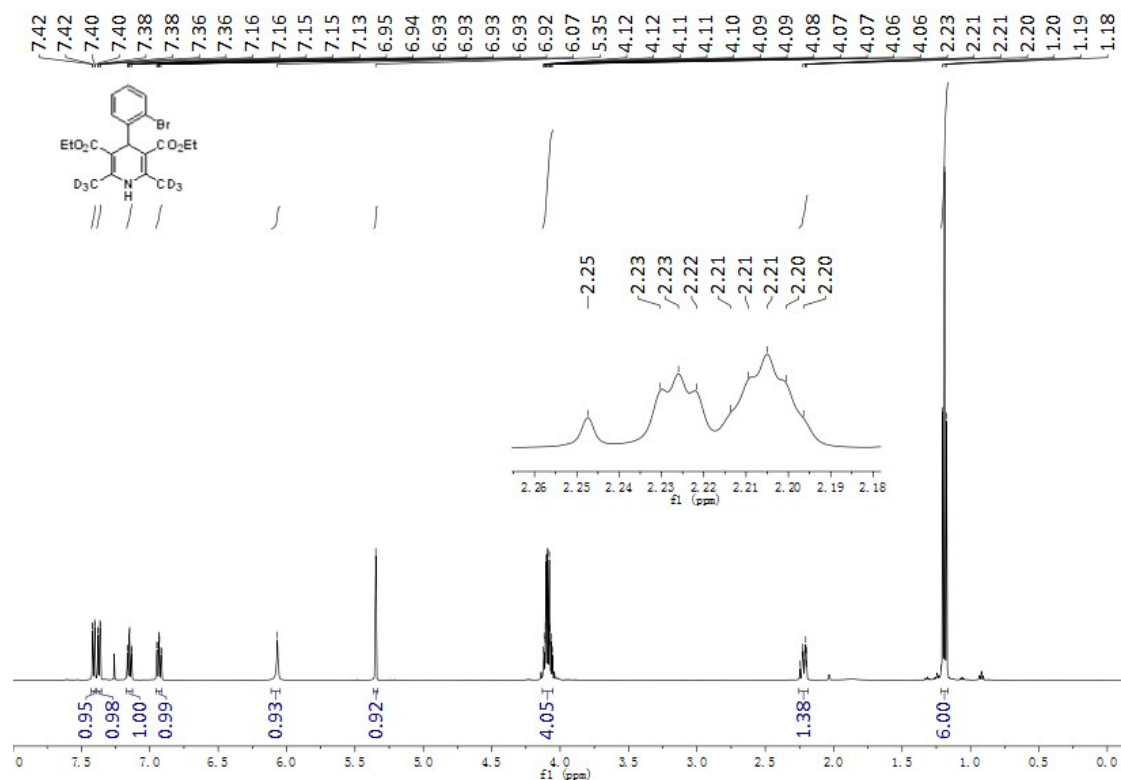
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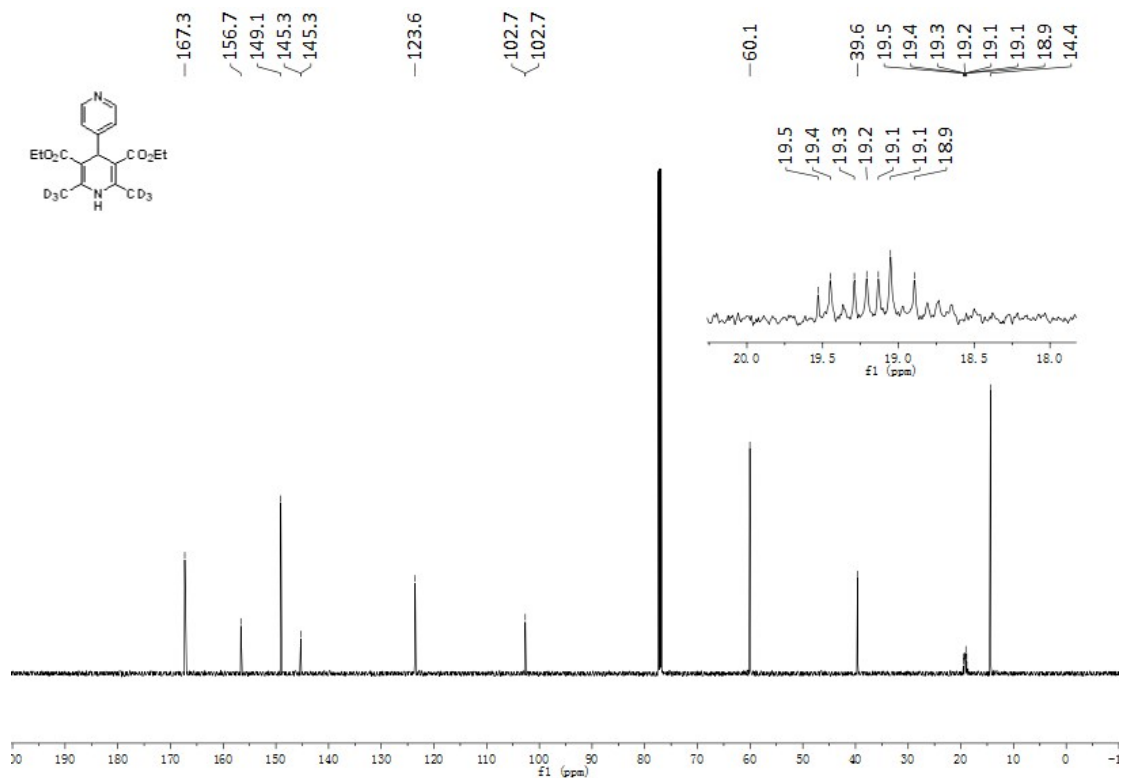
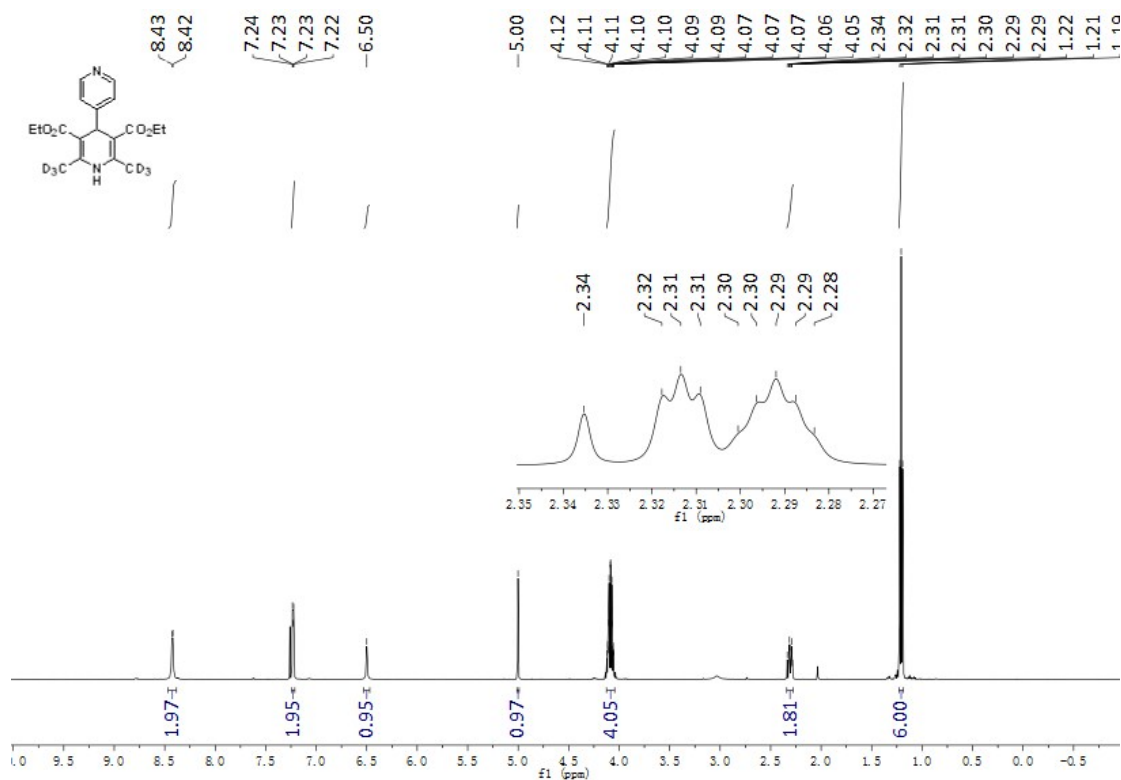
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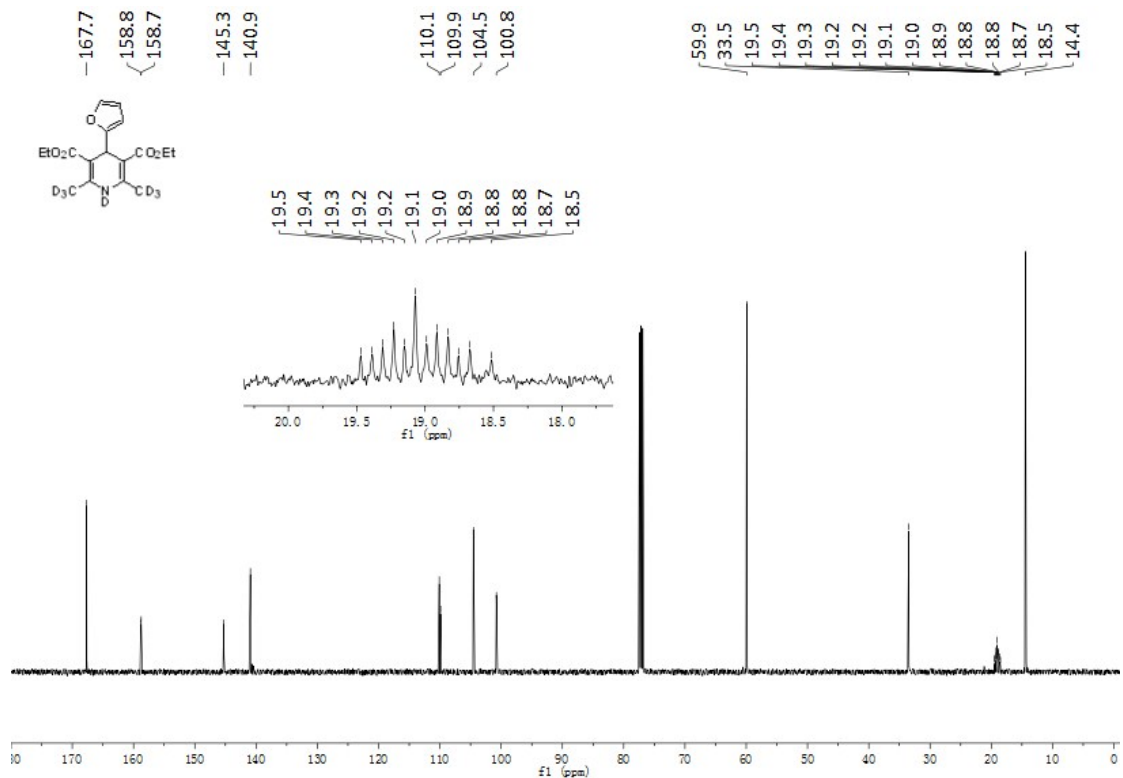
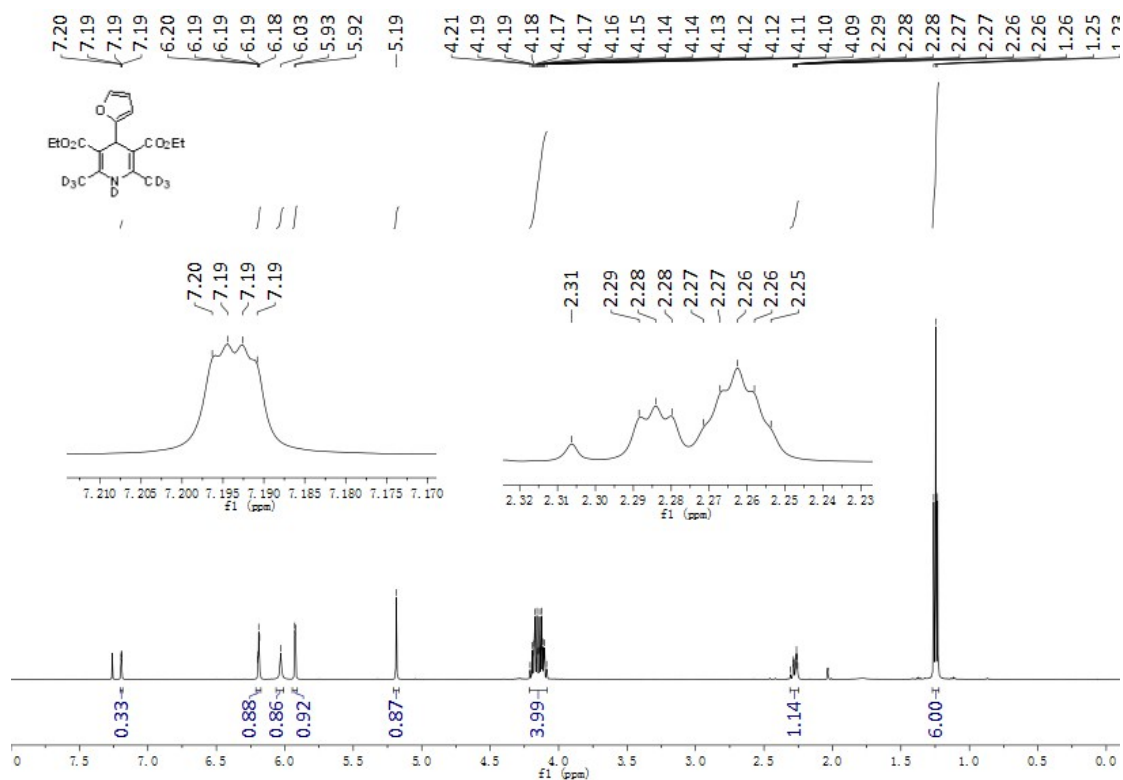
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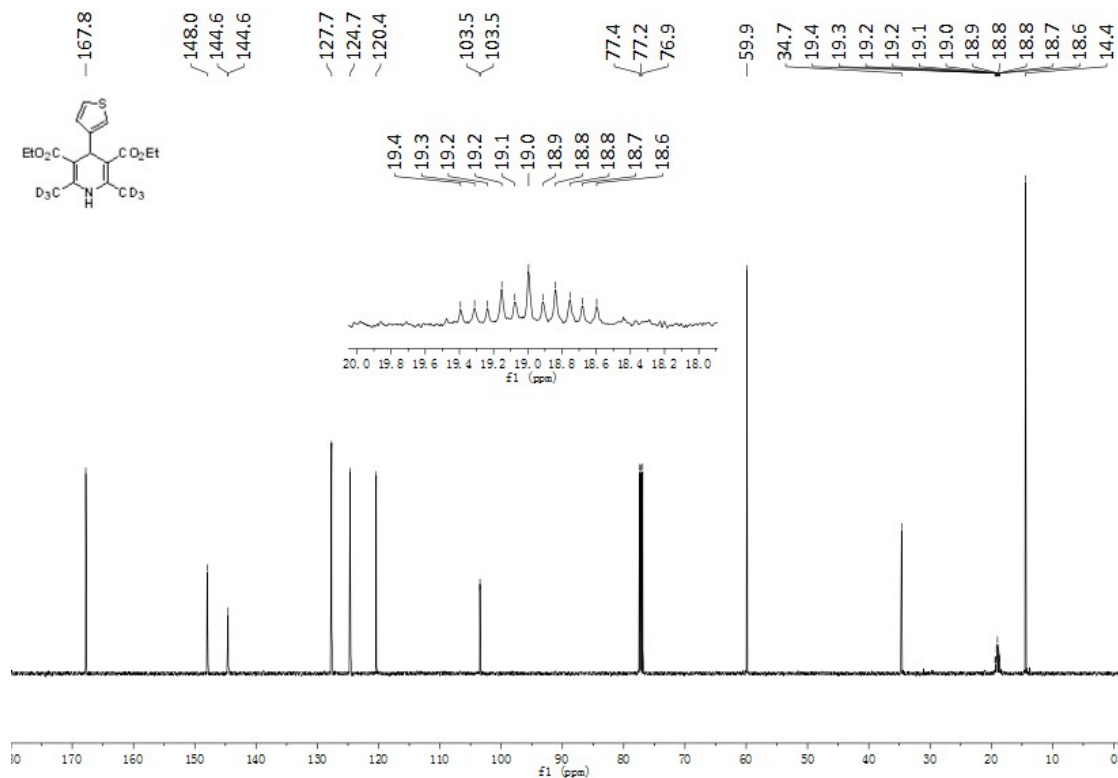
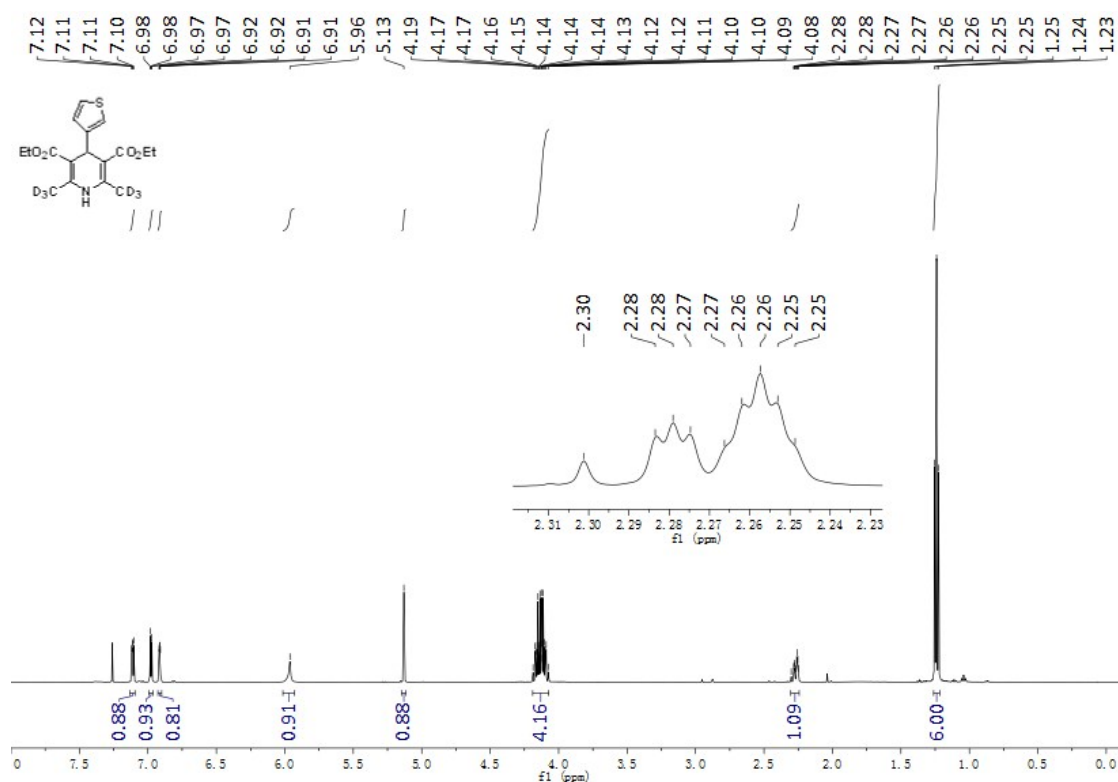
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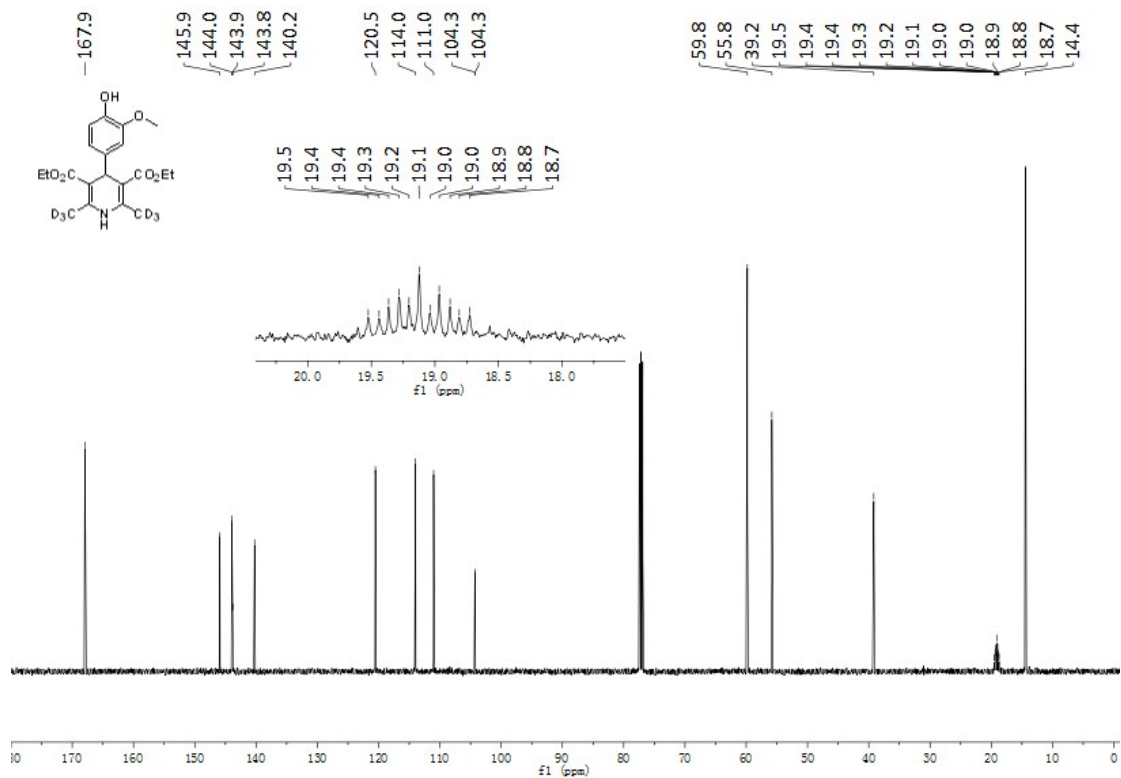
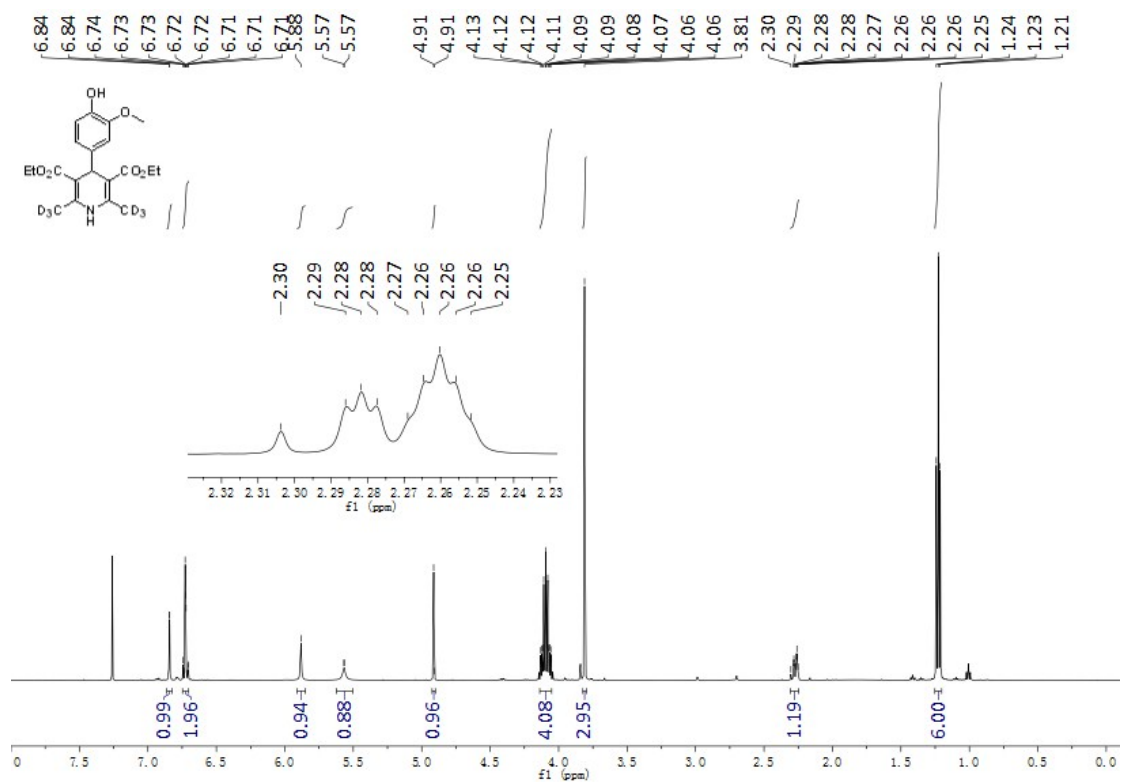


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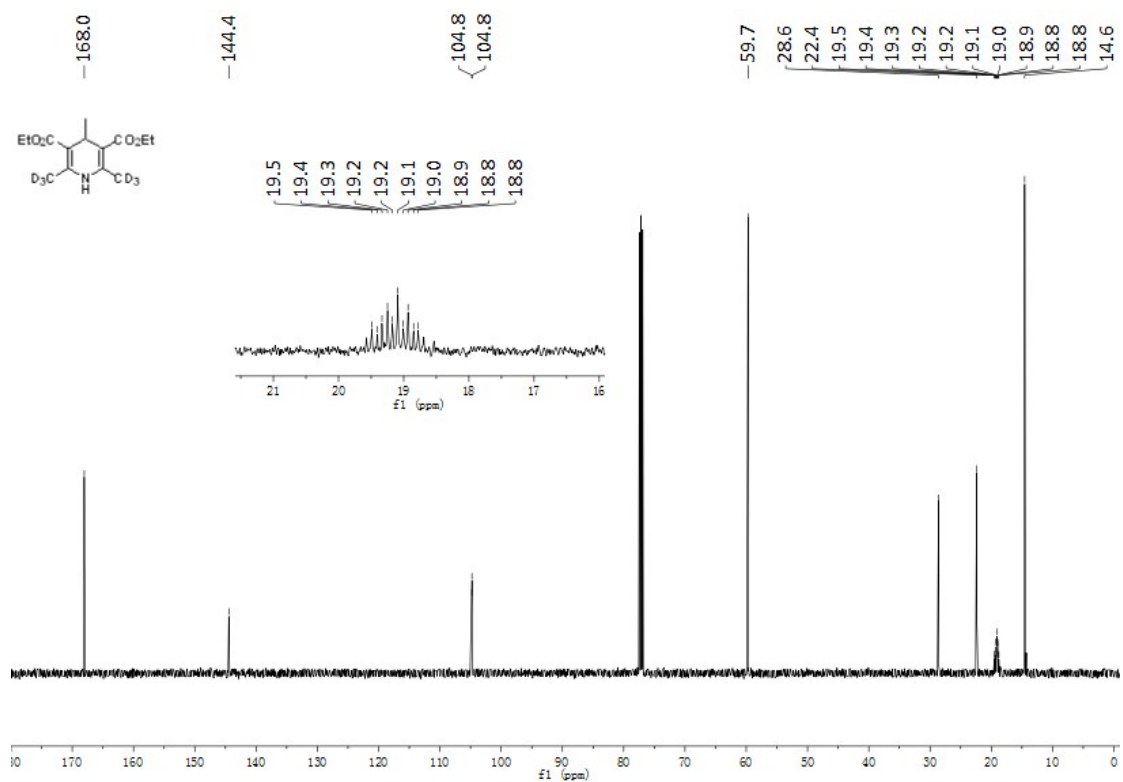
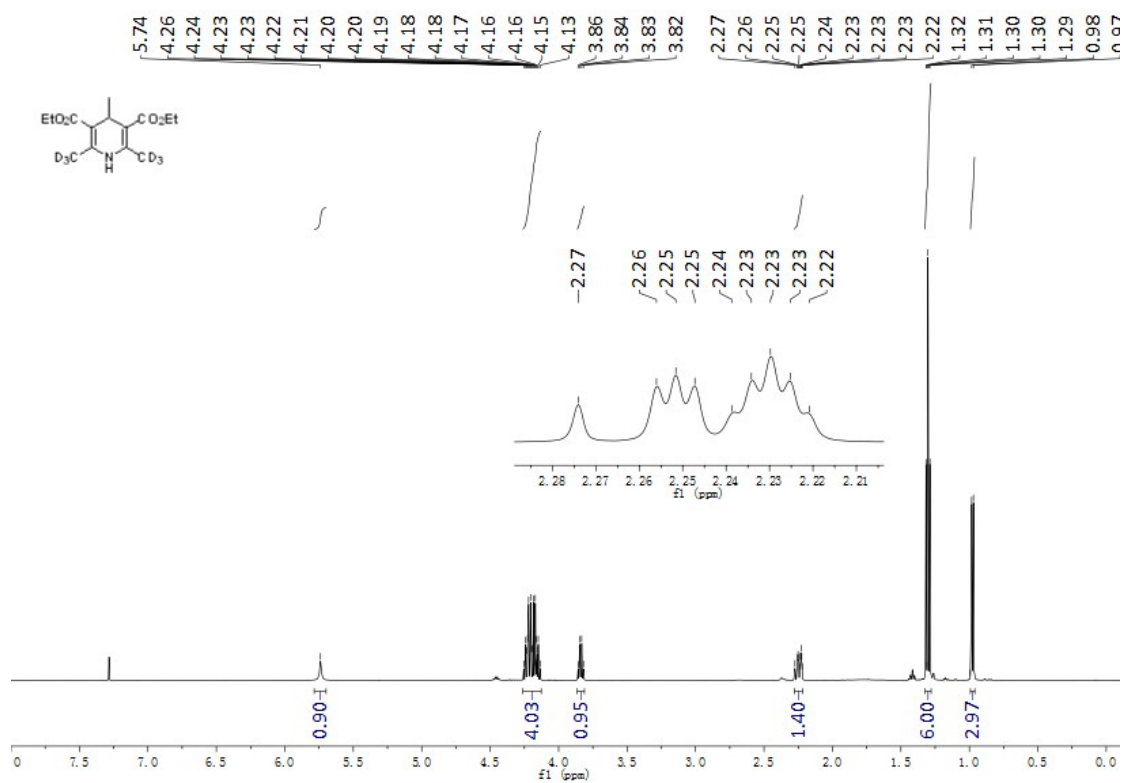




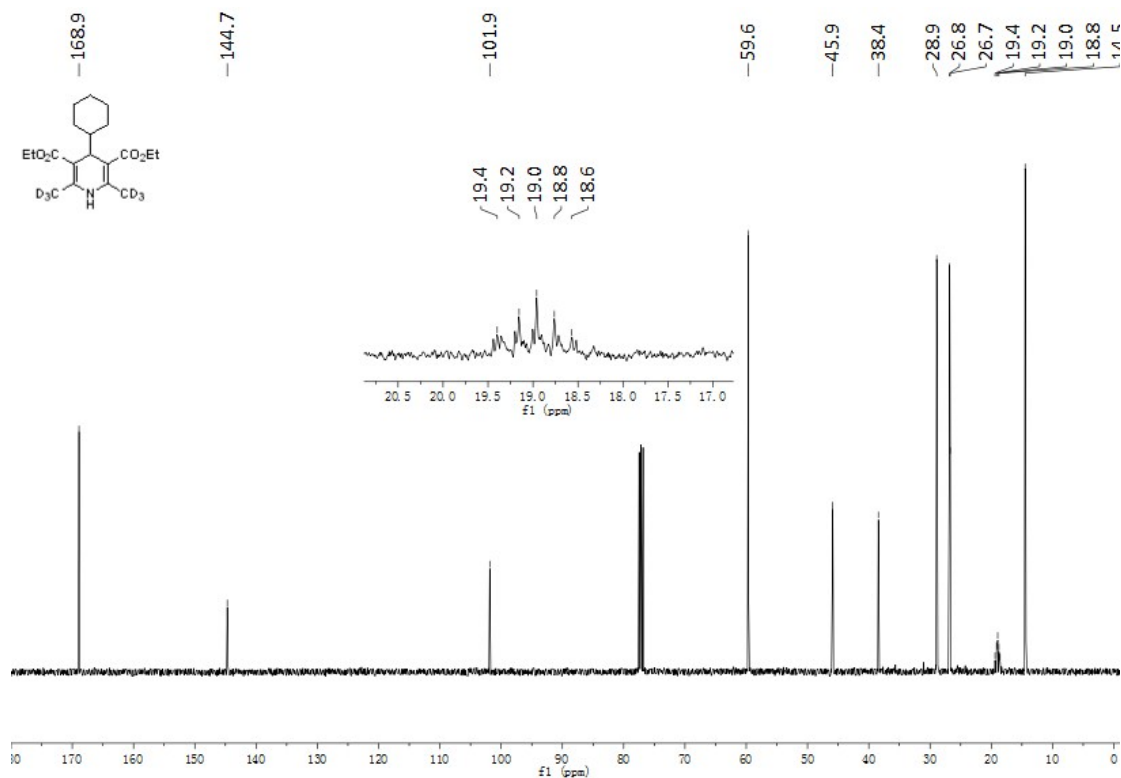
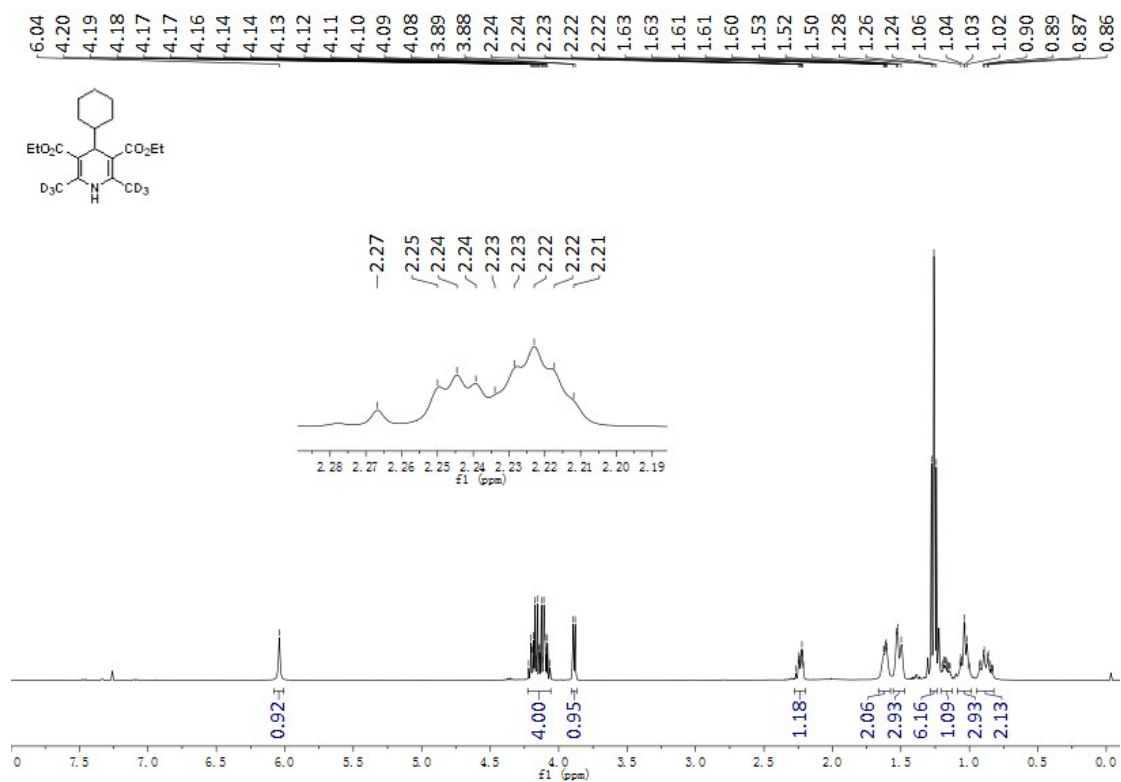
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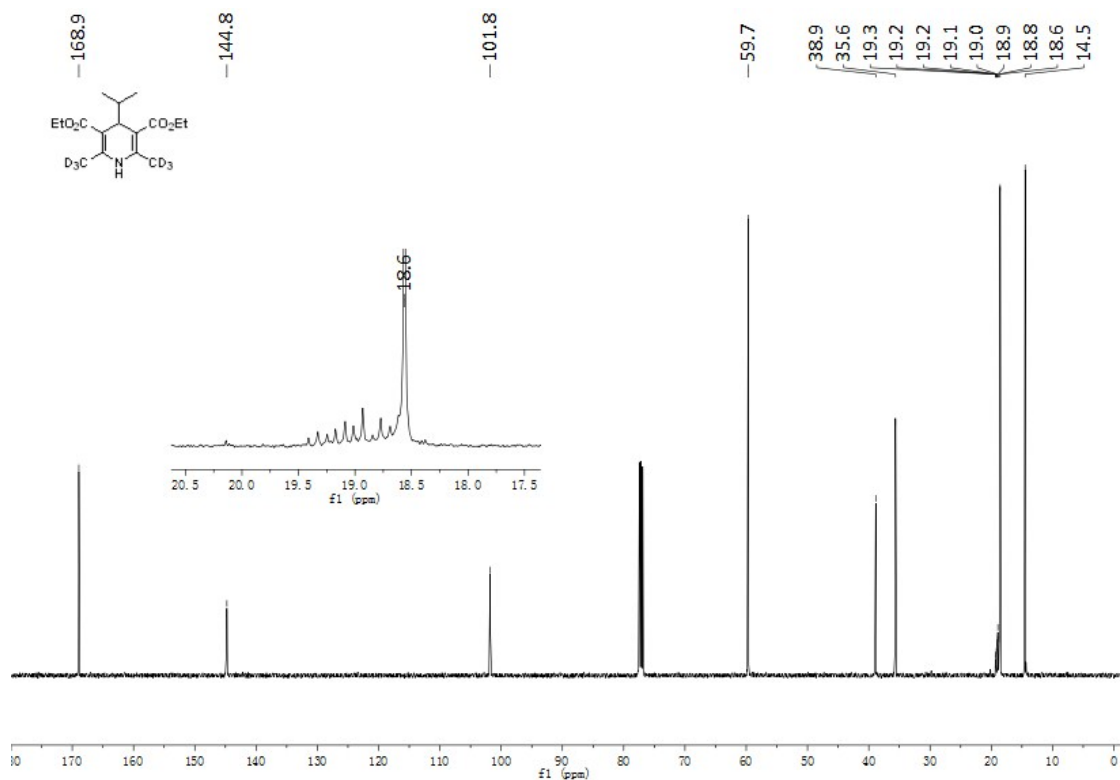
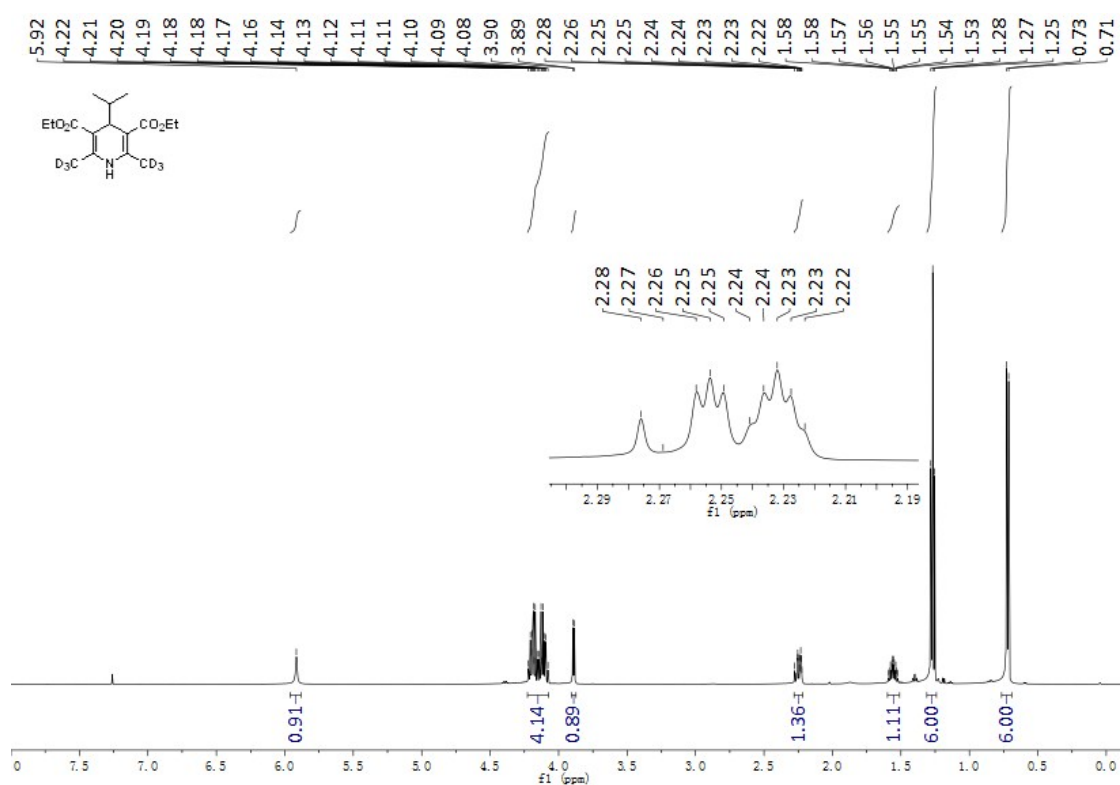
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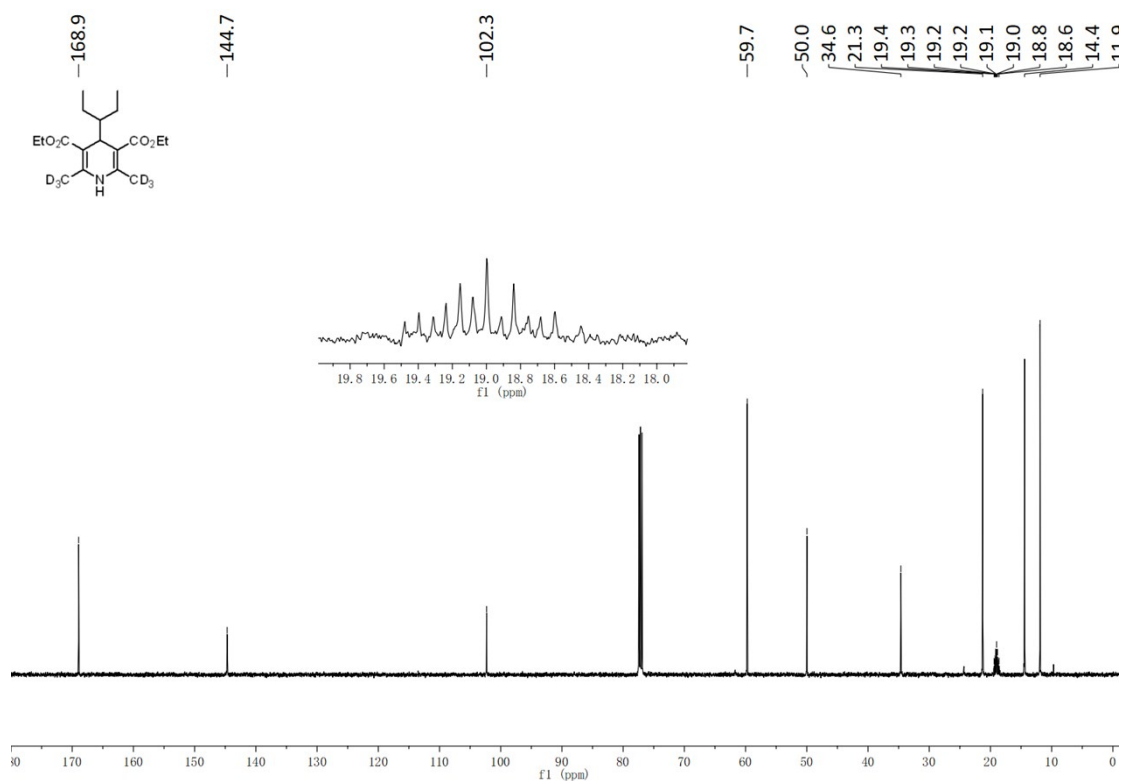
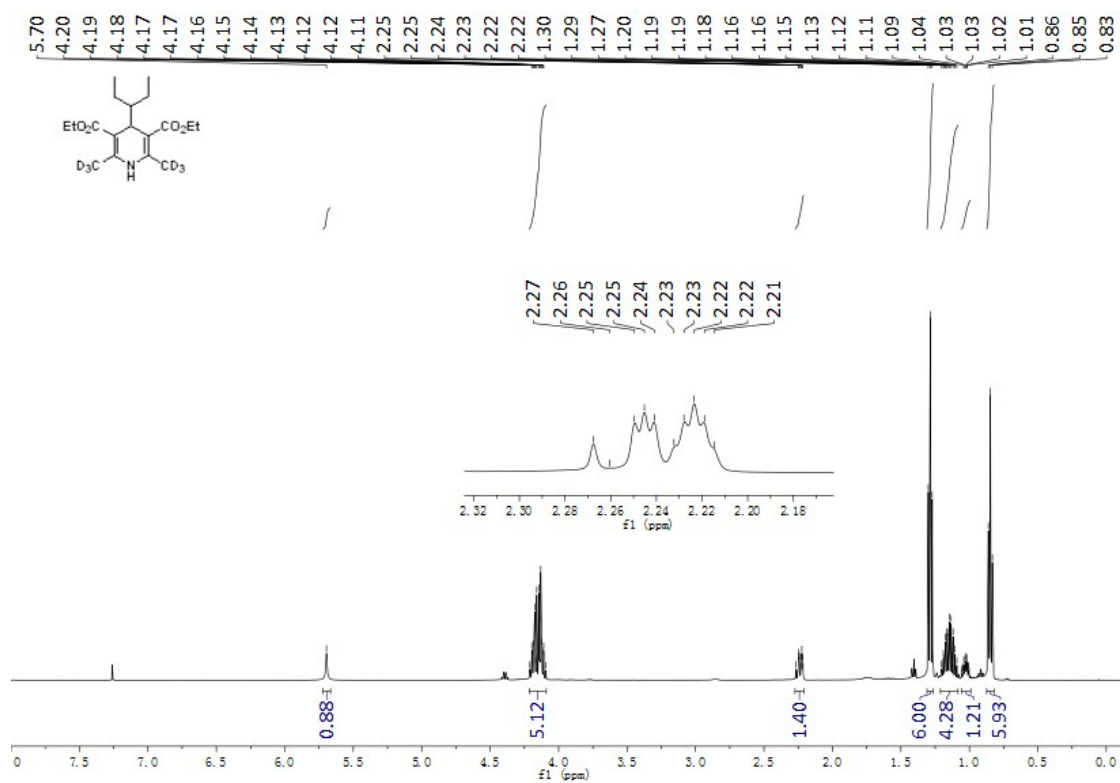
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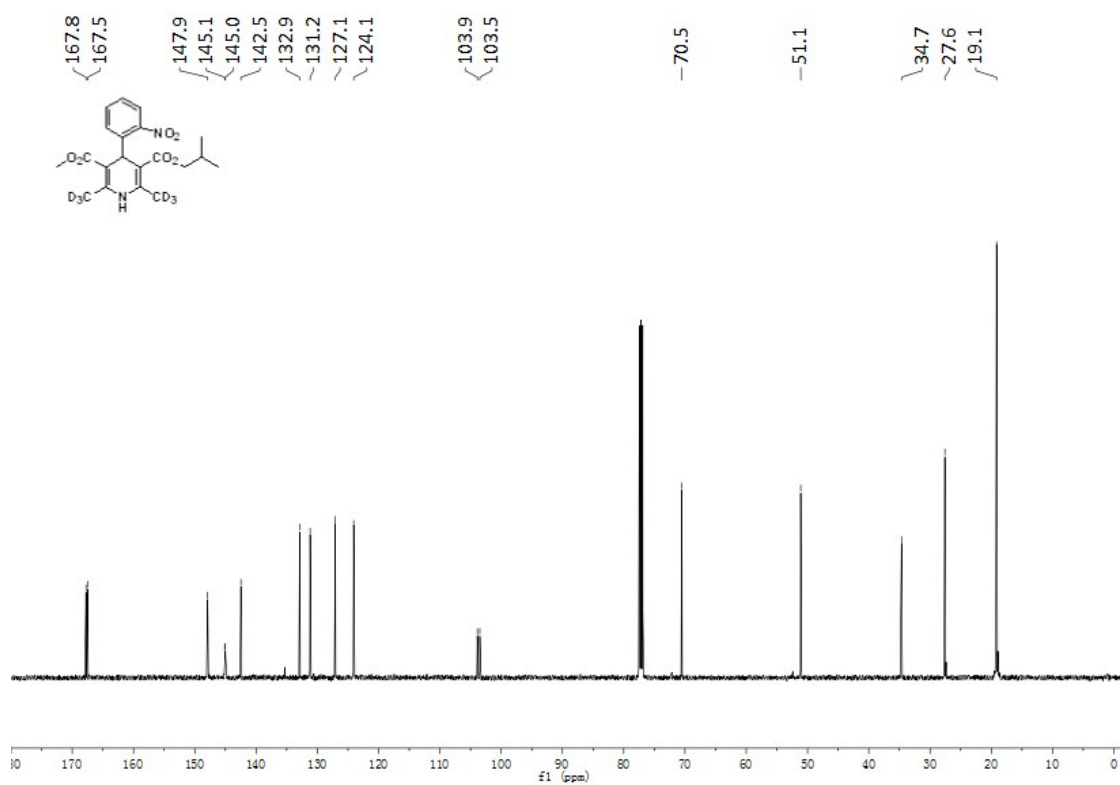
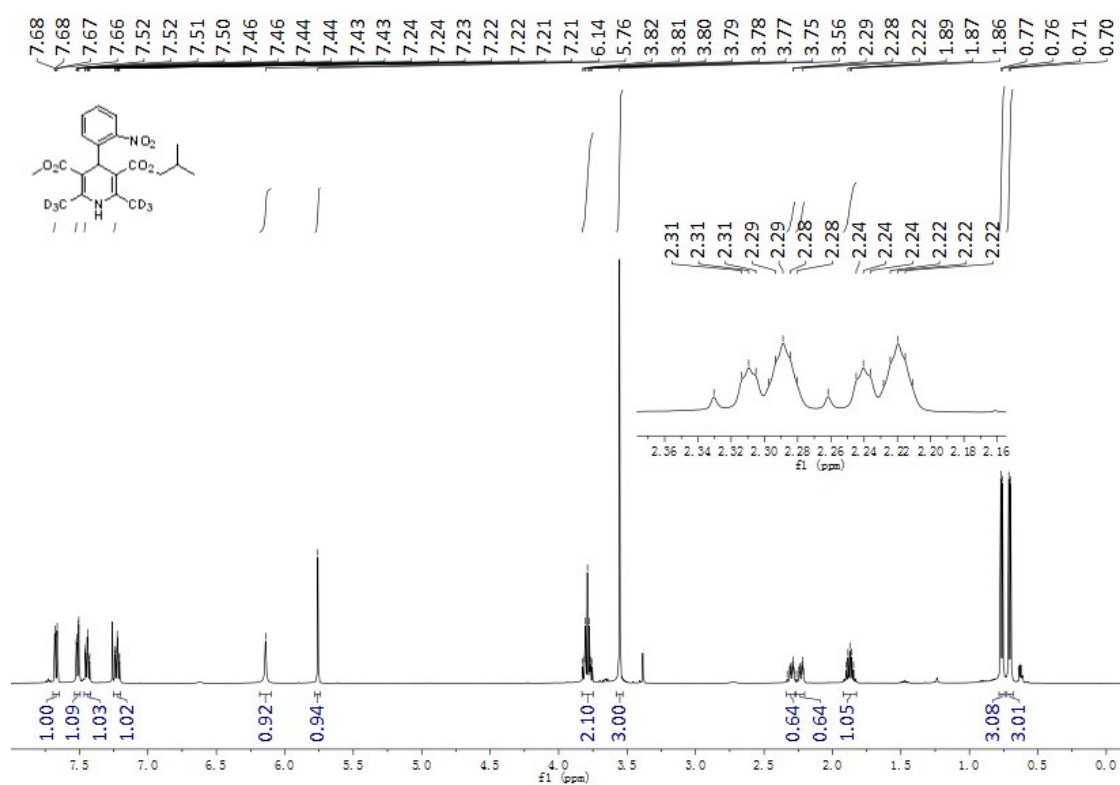
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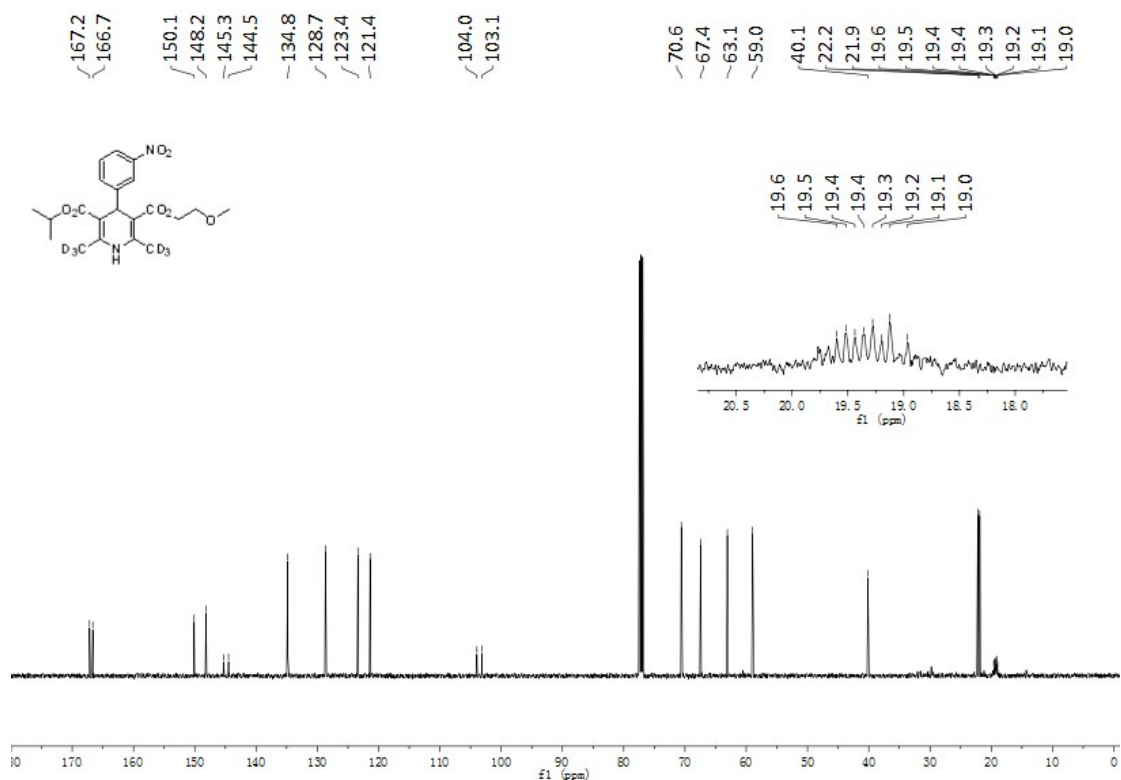
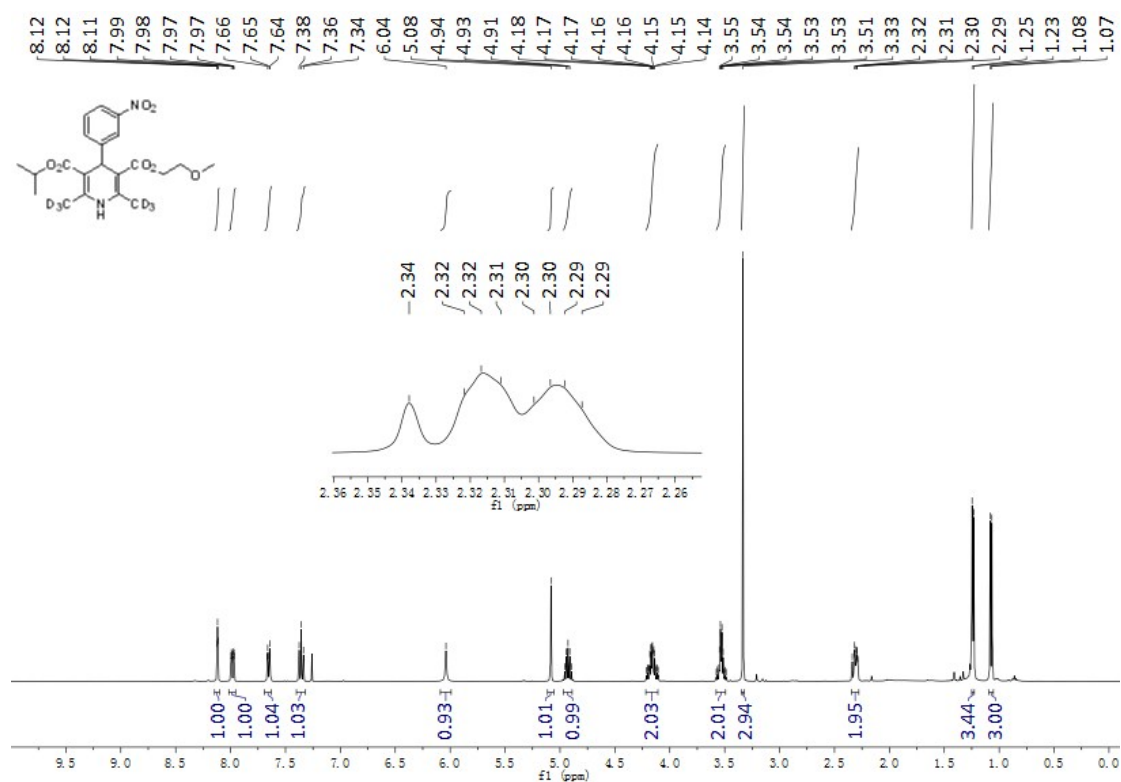
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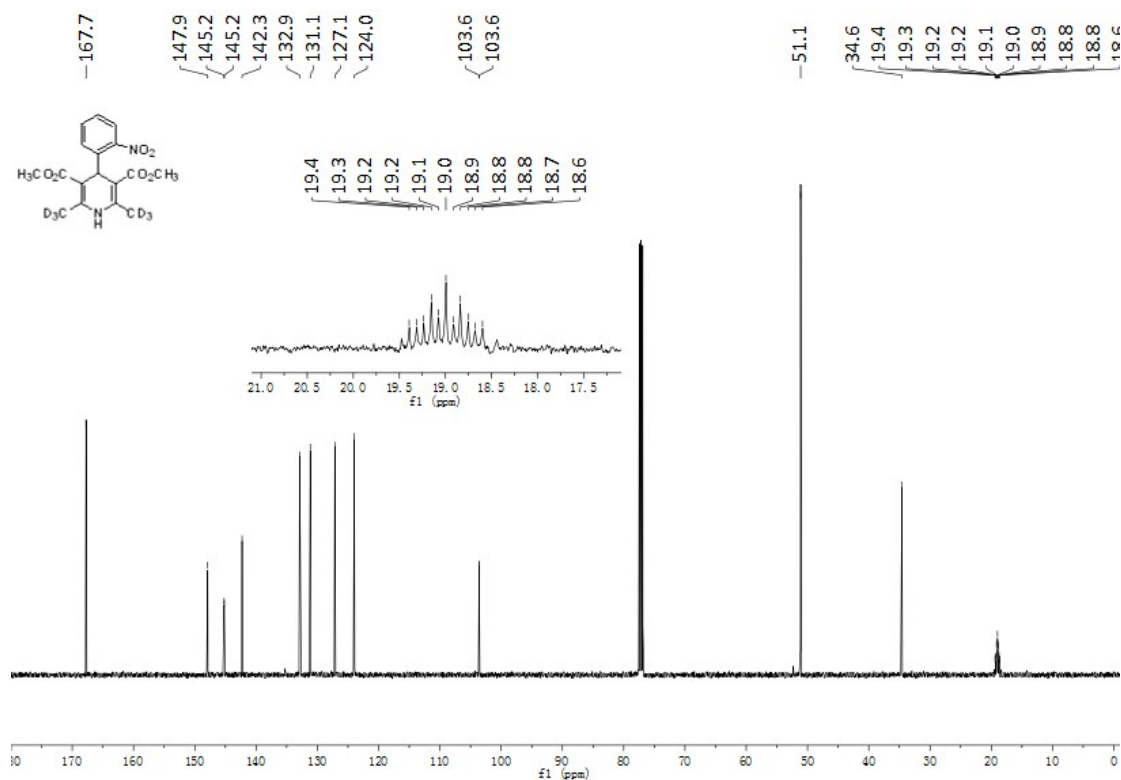
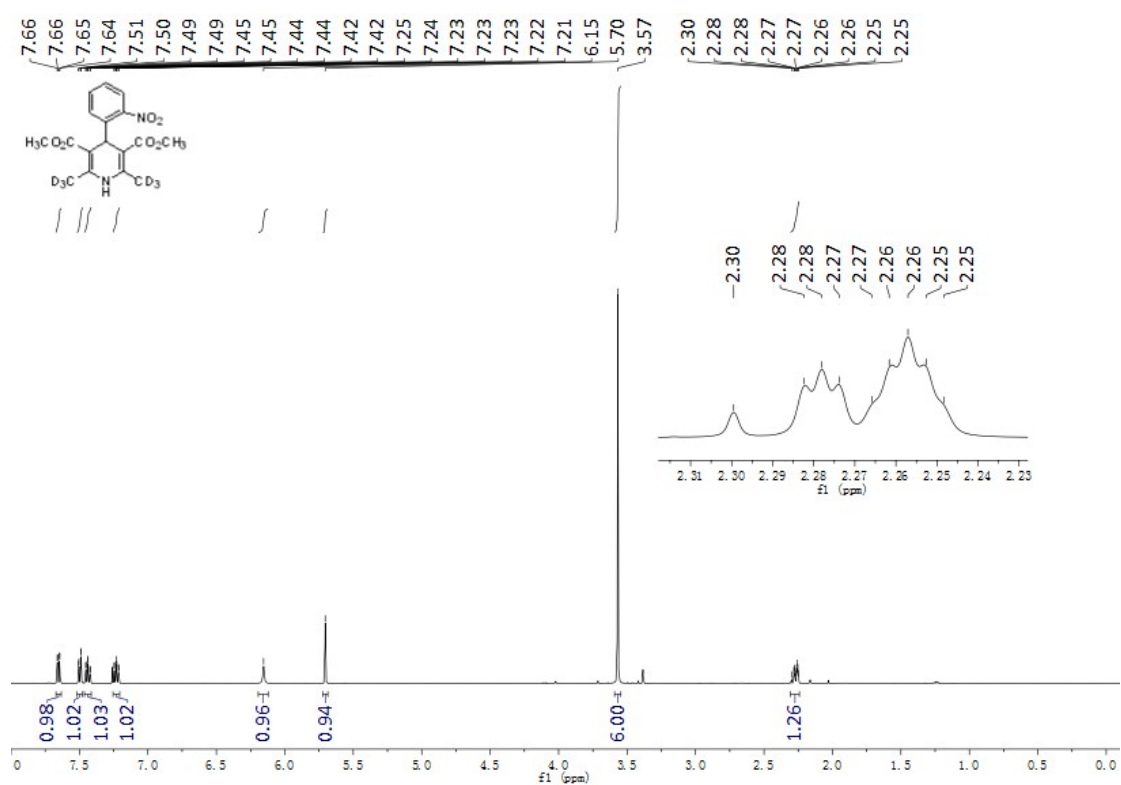
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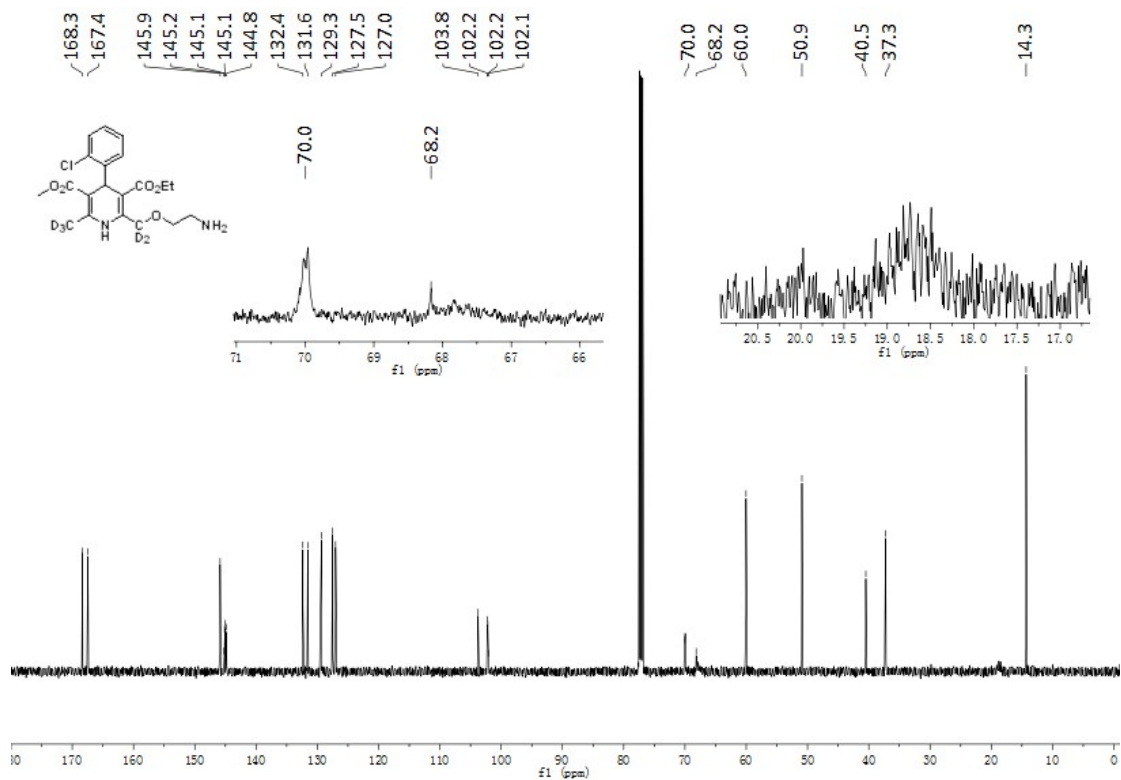
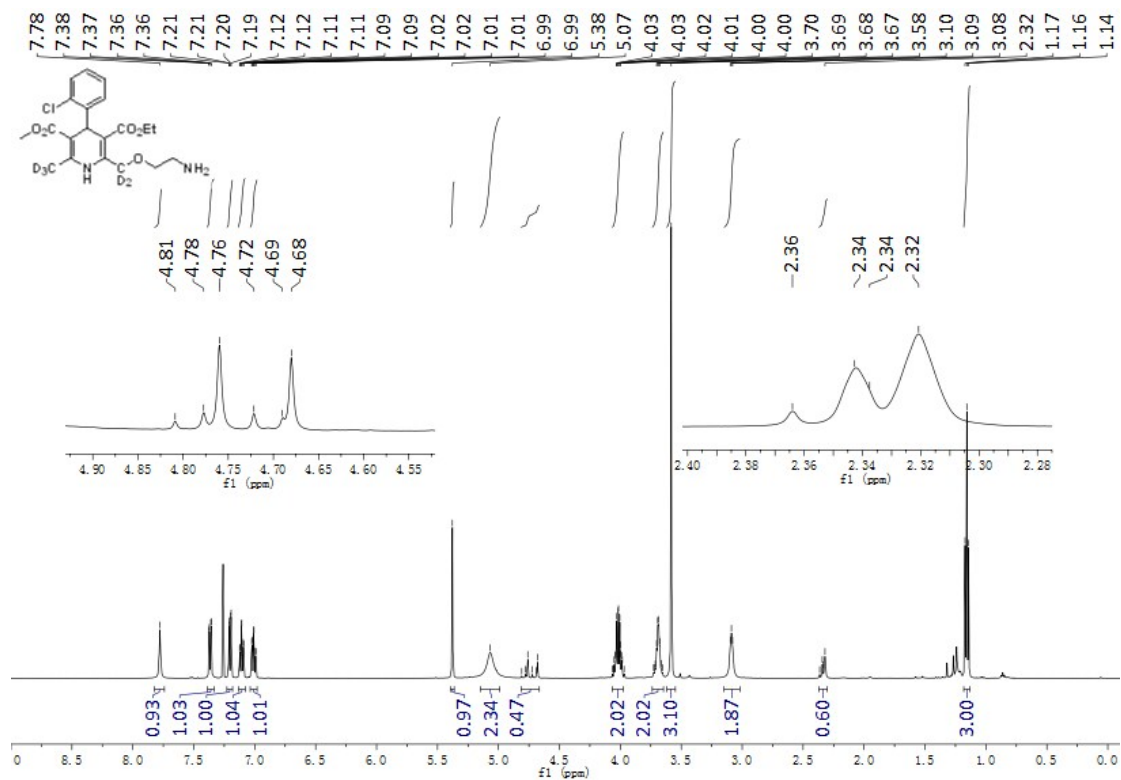


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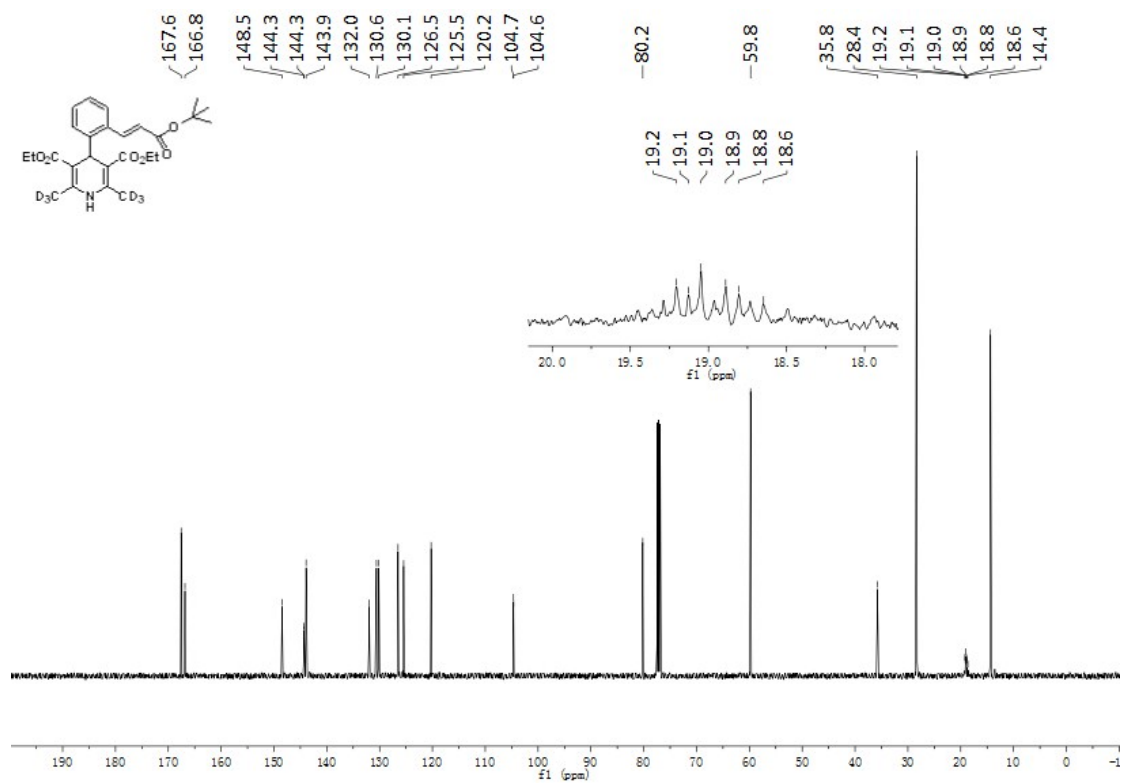
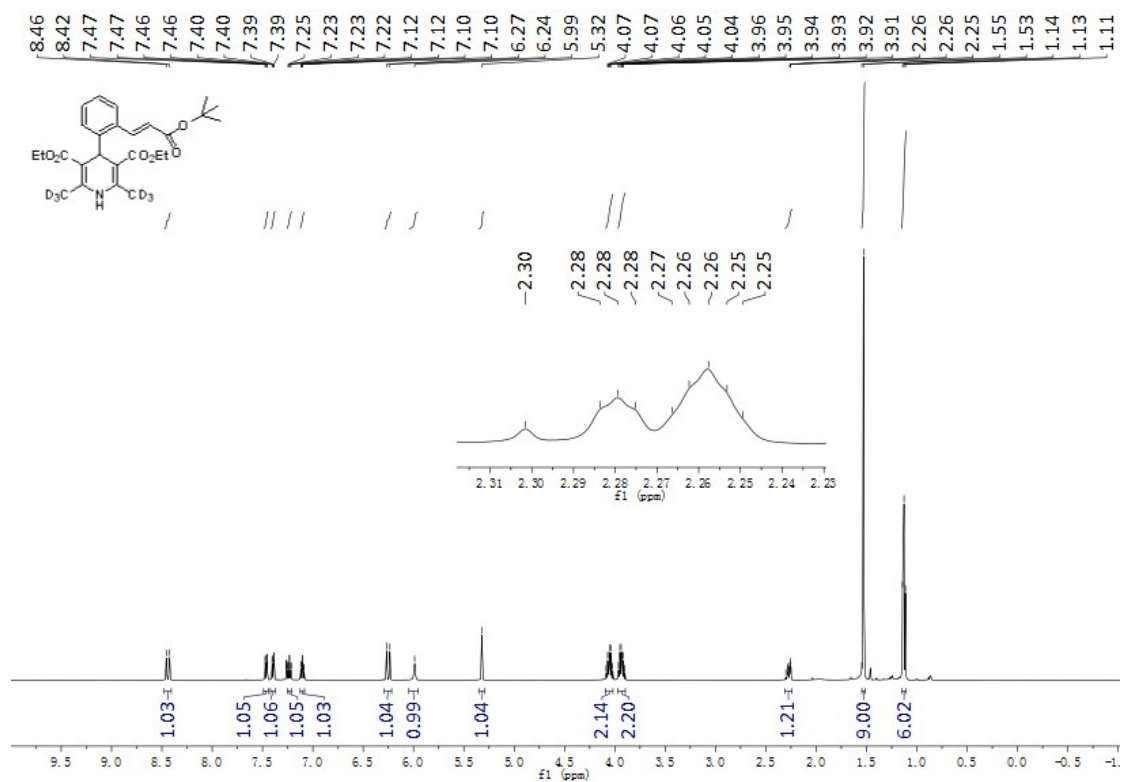




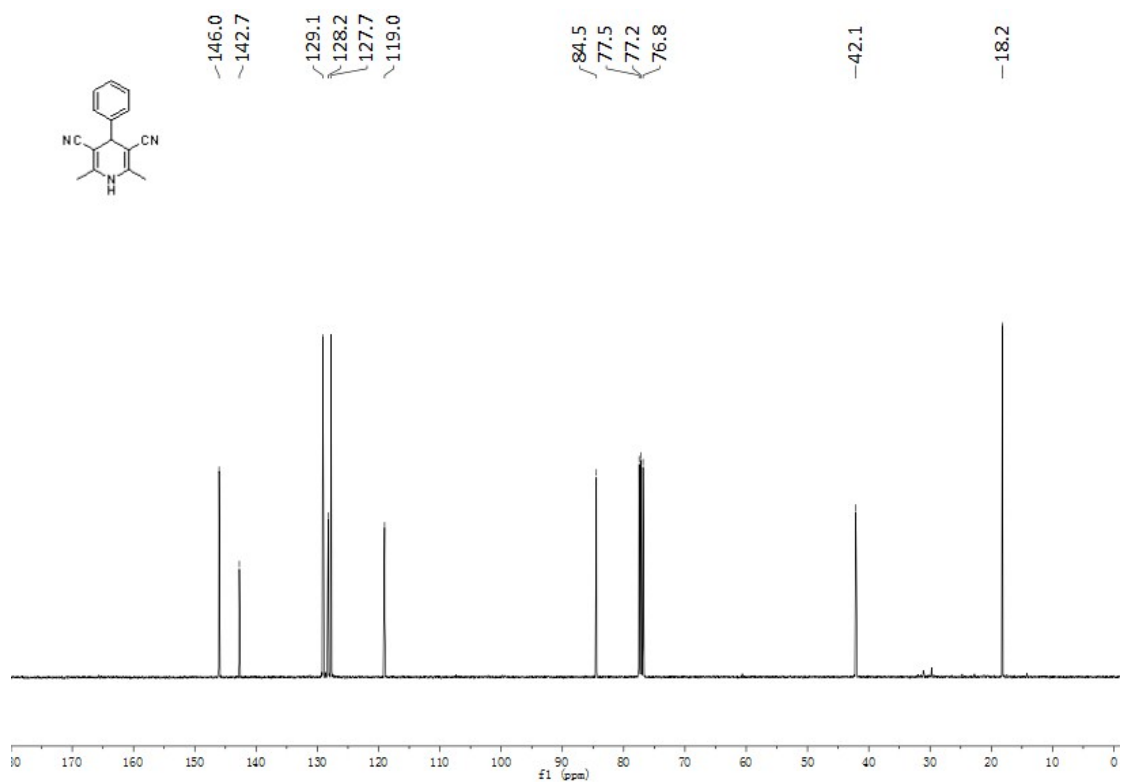
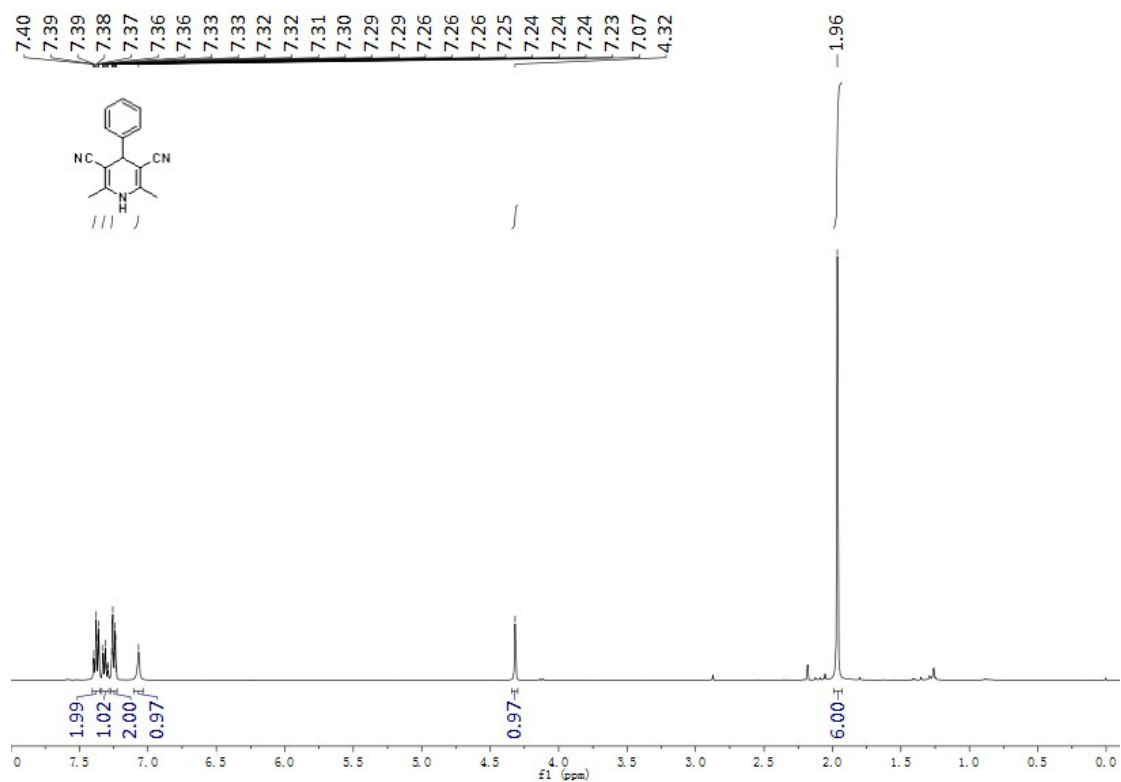
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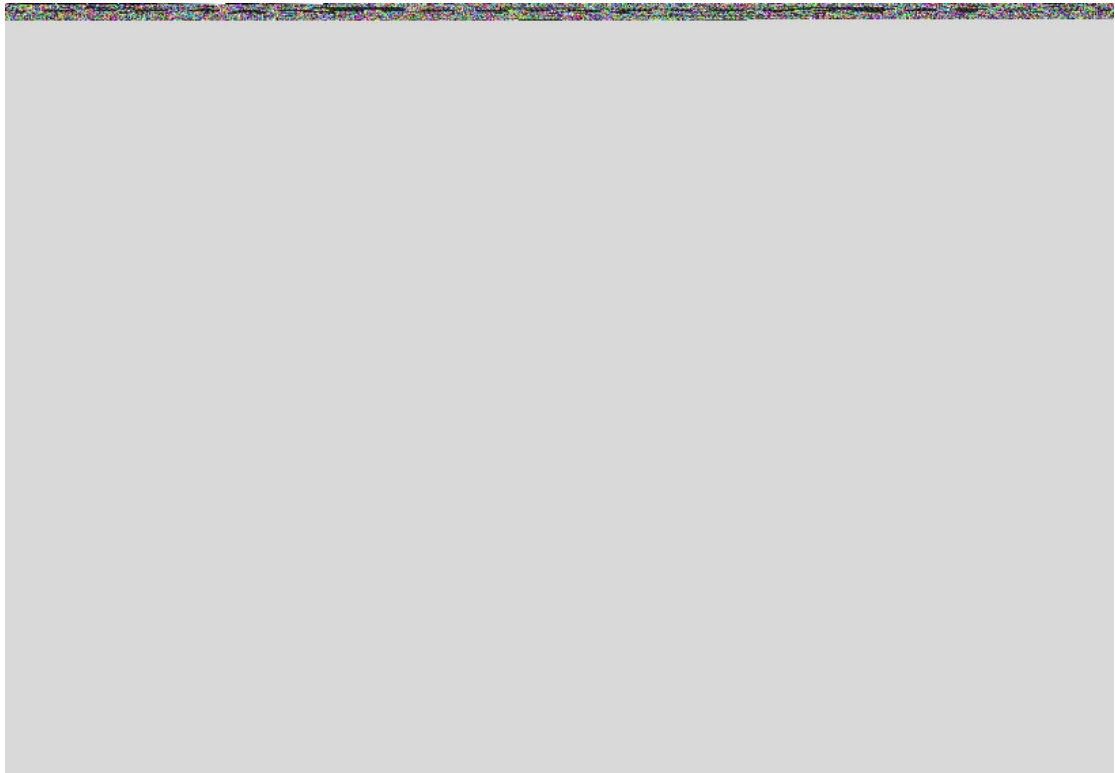
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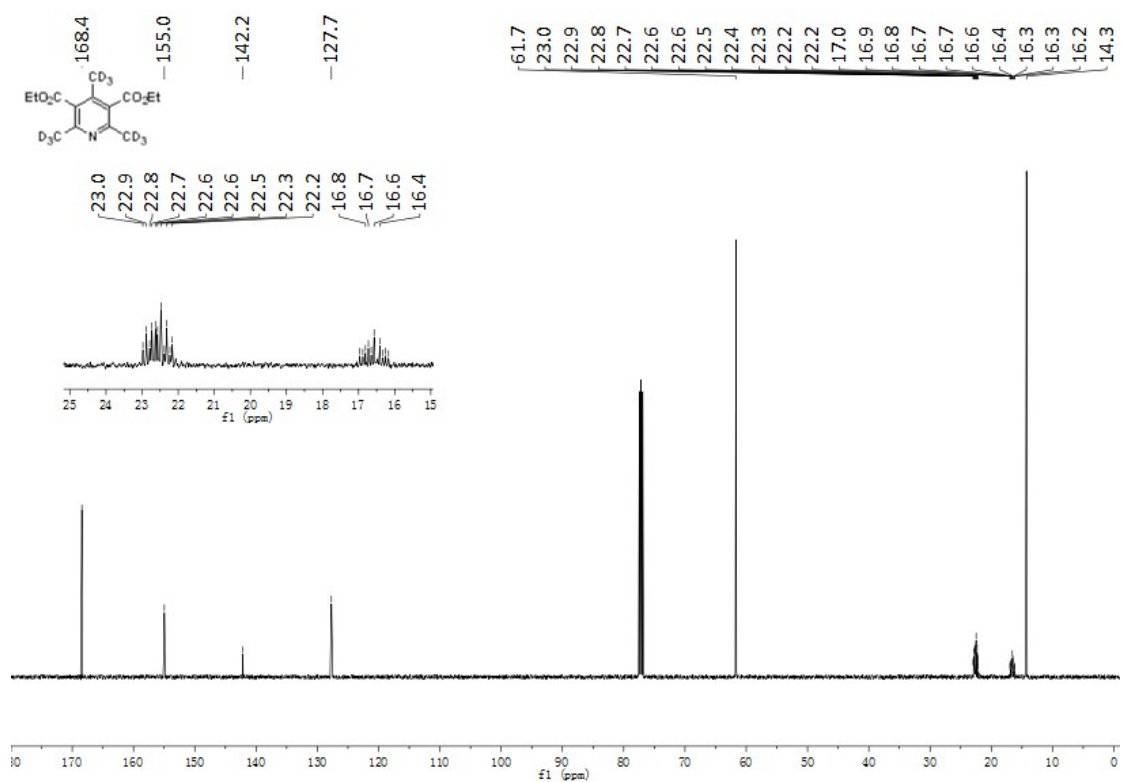
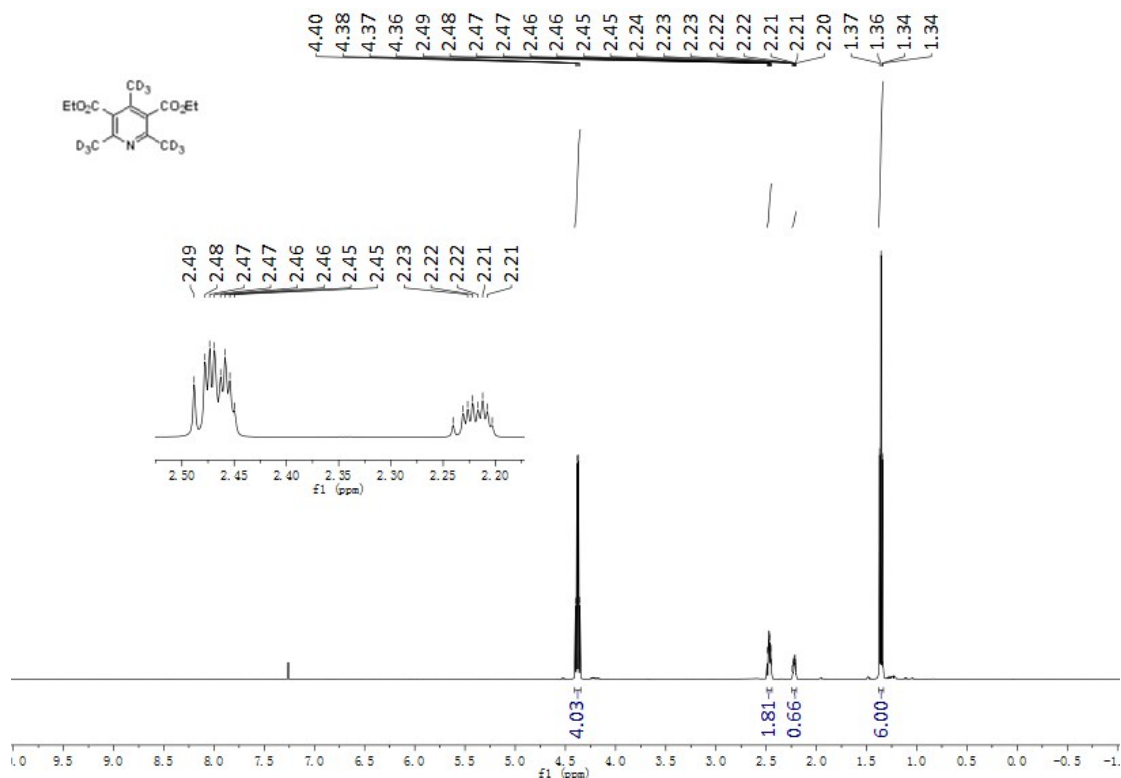
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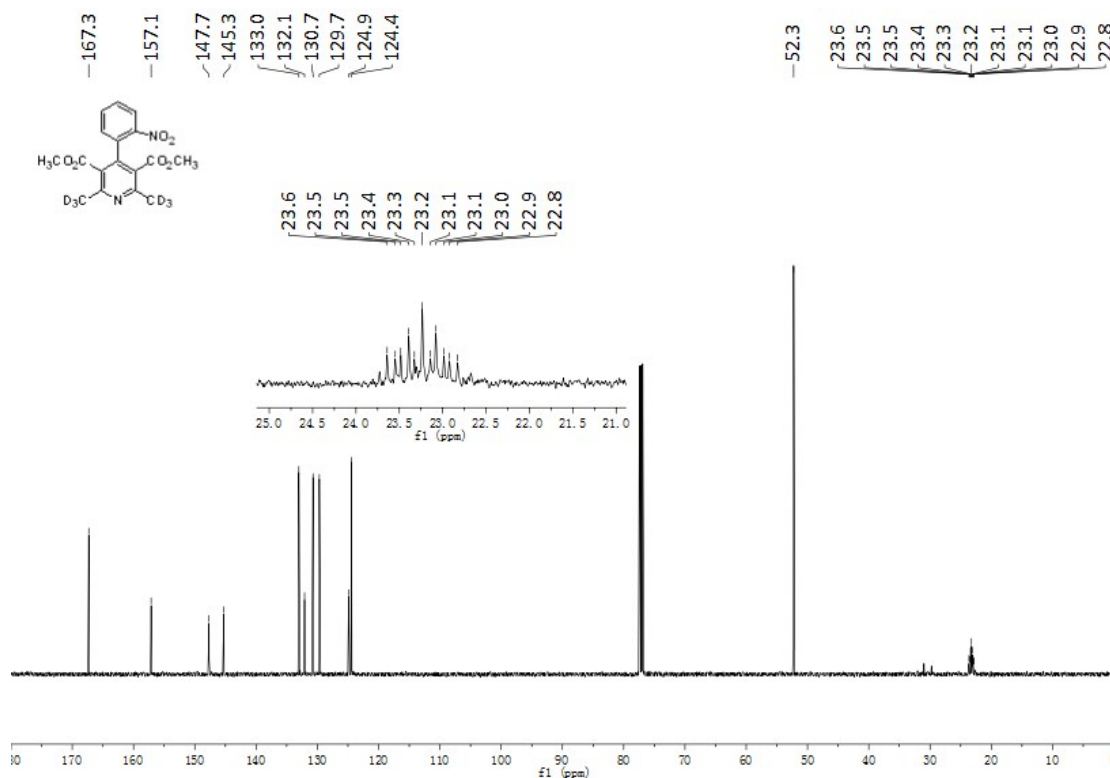
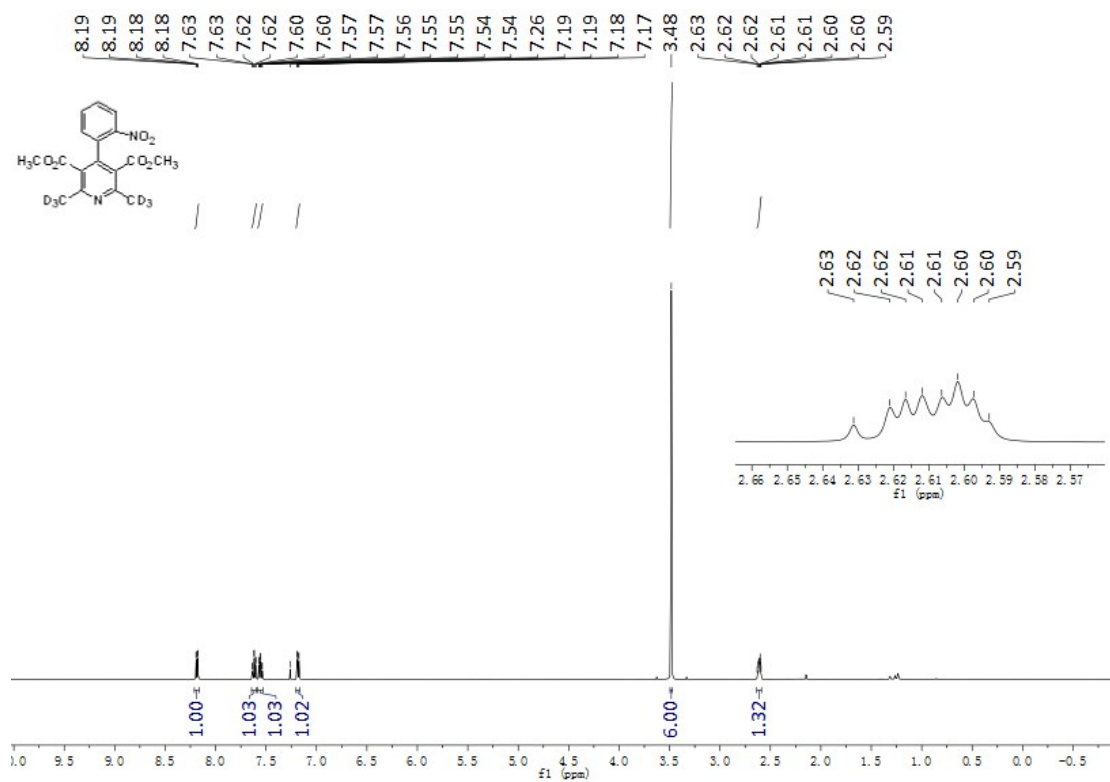
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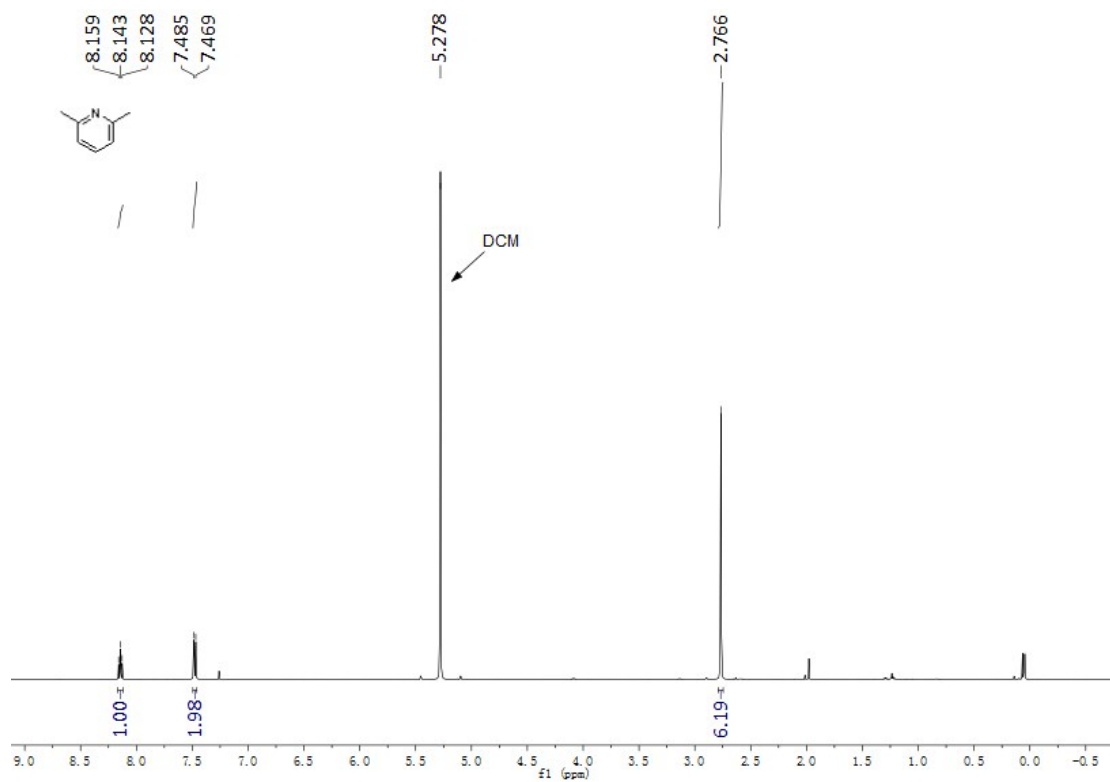
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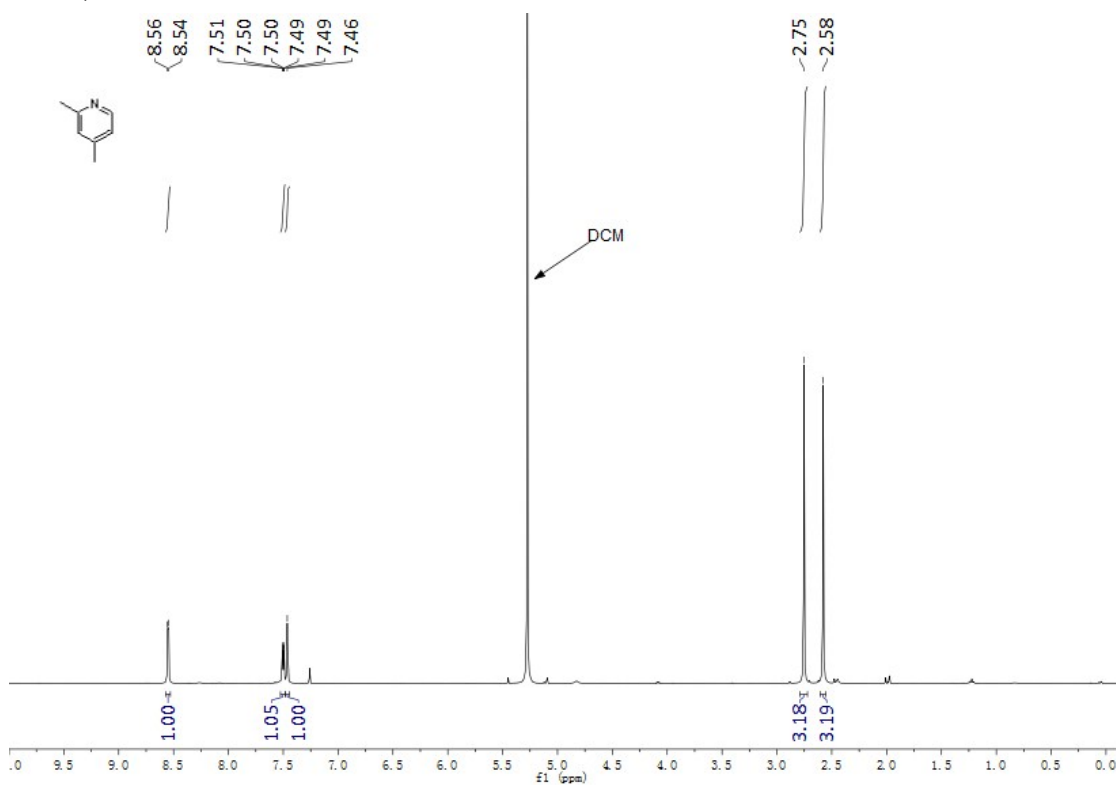
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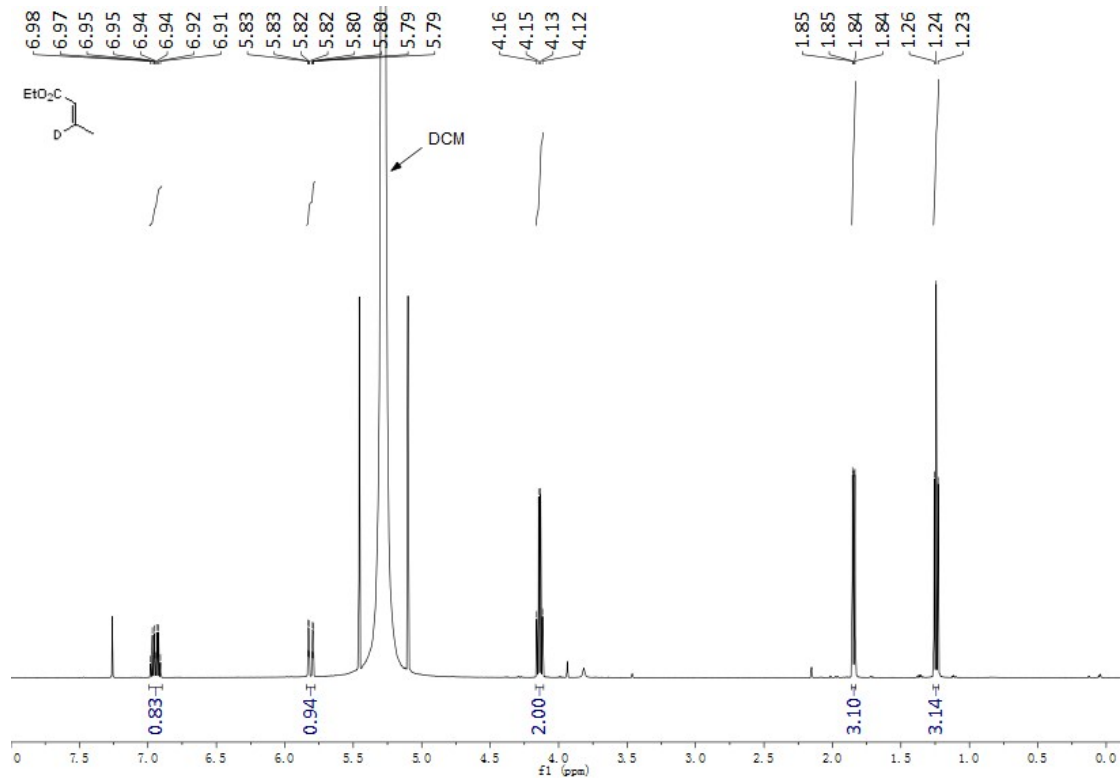
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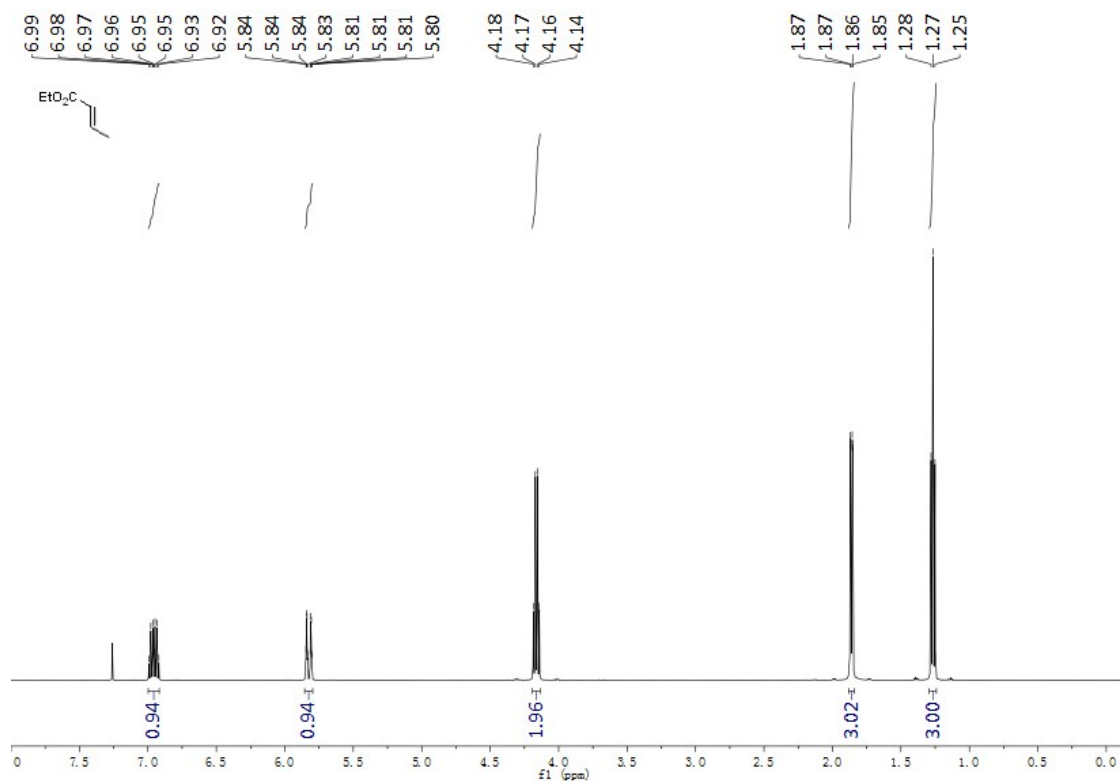
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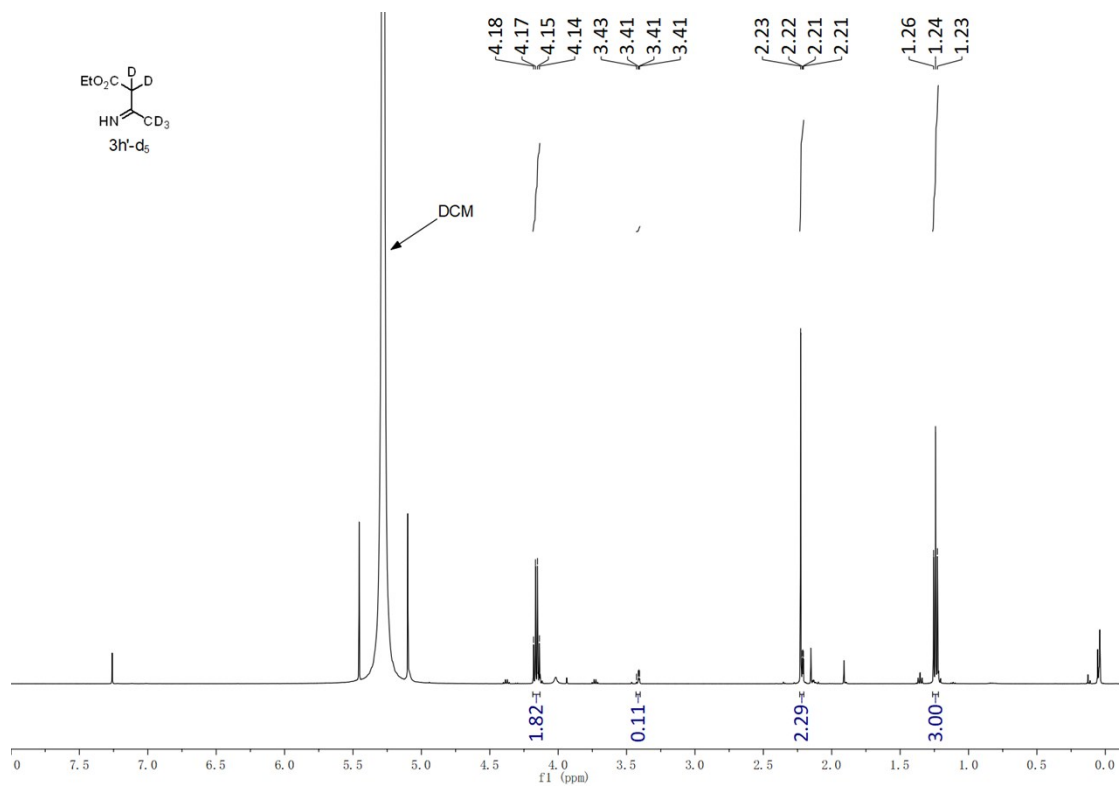


**3g**

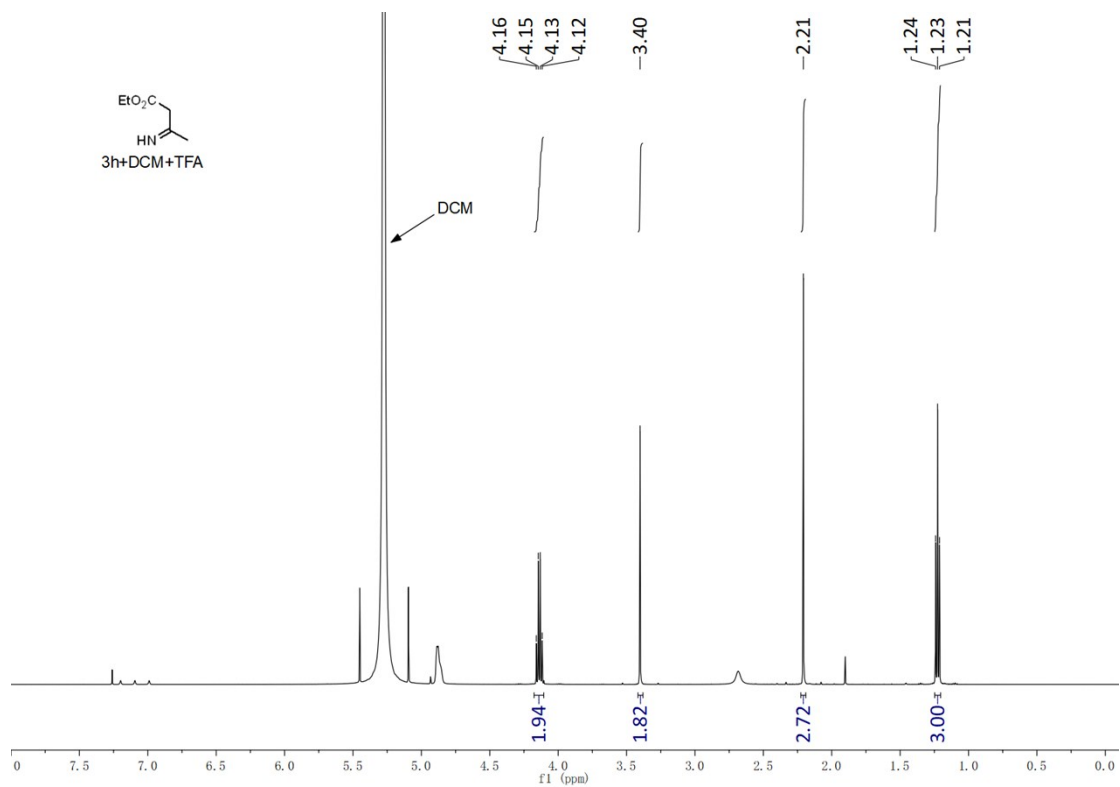




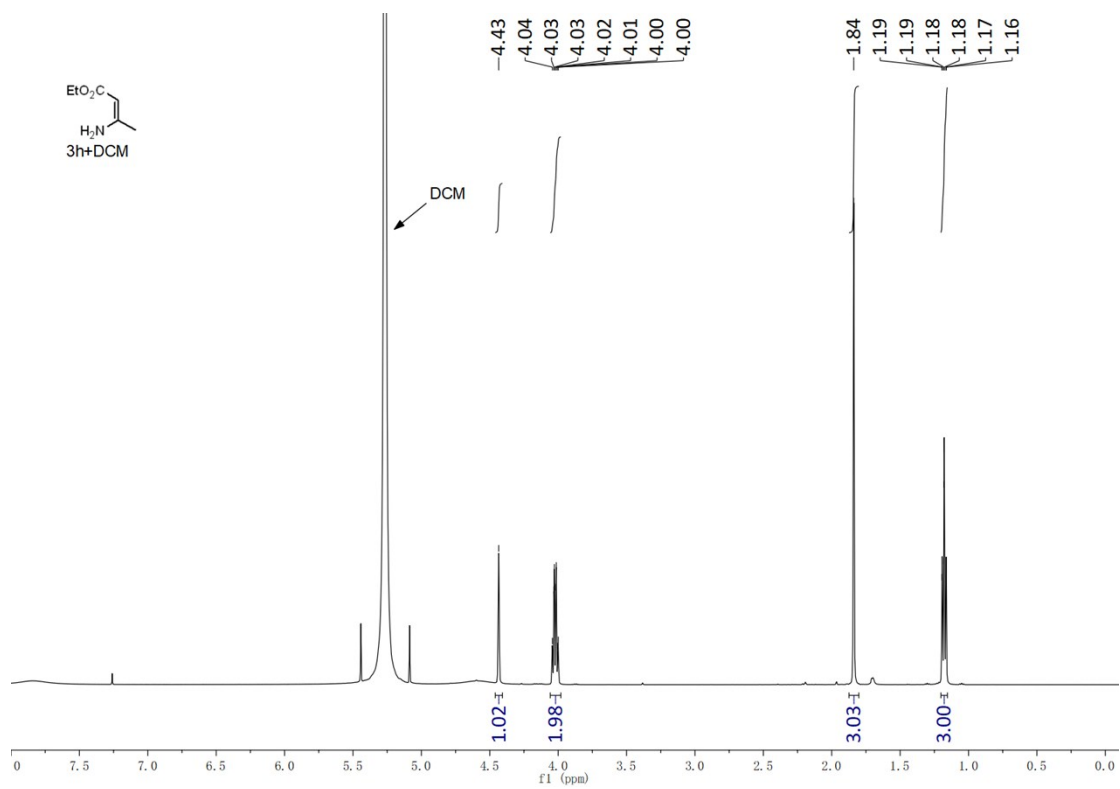
### 3h'-d<sub>5</sub>(crude)



### 3h + CH<sub>2</sub>Cl<sub>2</sub>+TFA

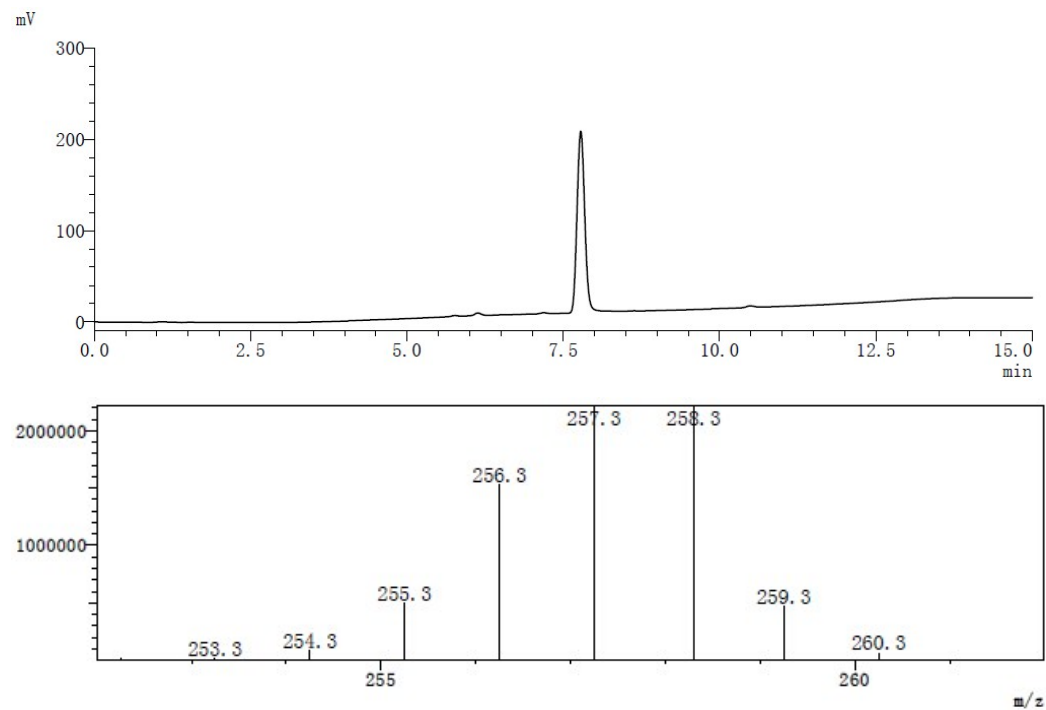


3h + CH<sub>2</sub>Cl<sub>2</sub>

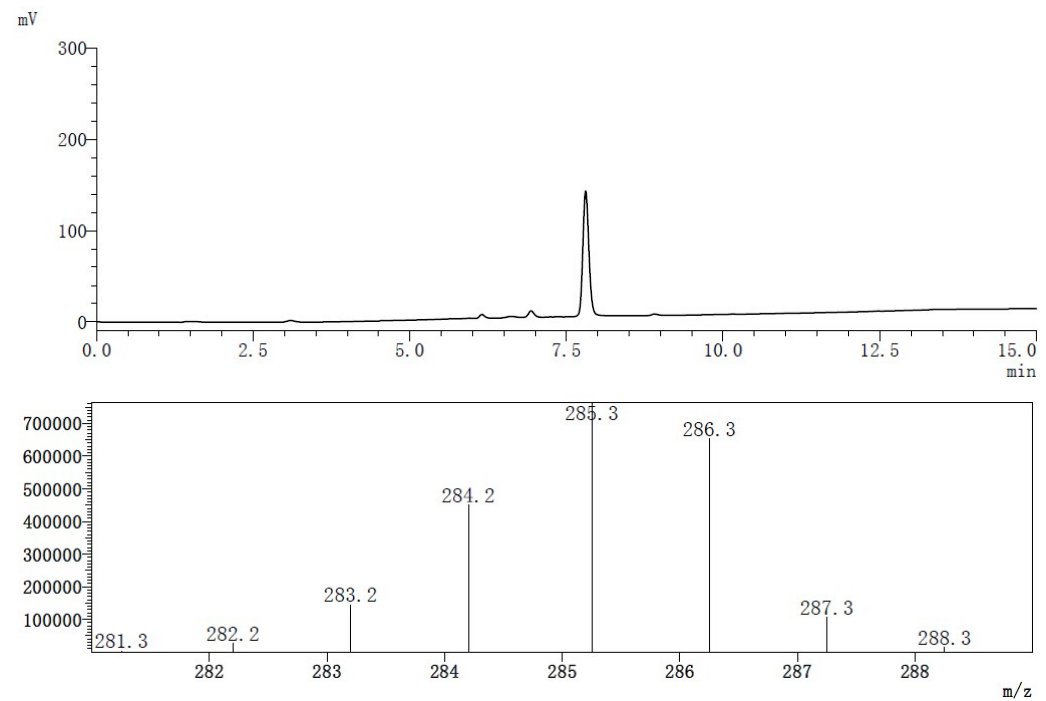


## 9. LC-MS spectra

### 1a'-d<sub>6</sub>

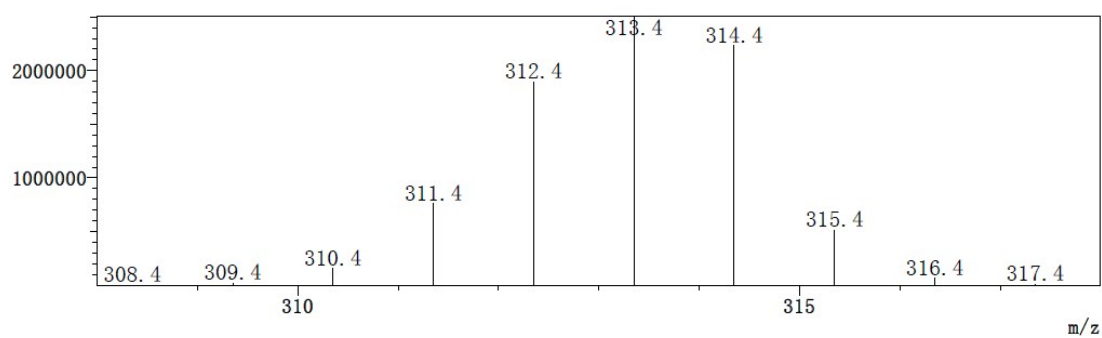
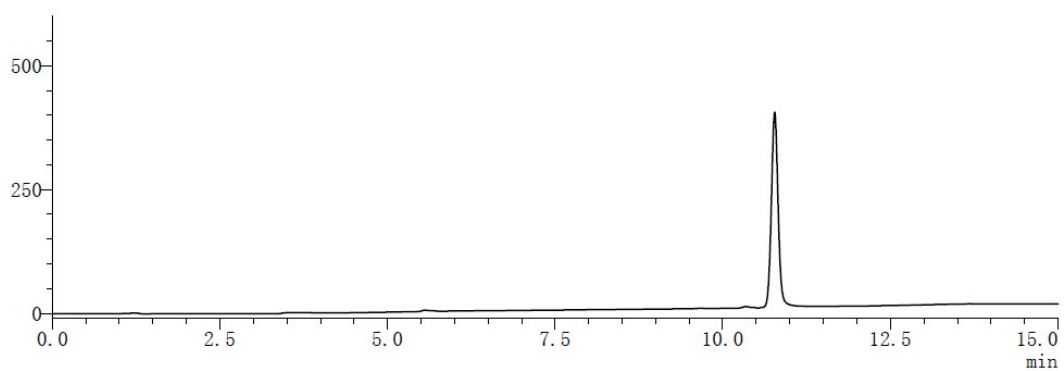


### 1b'-d<sub>6</sub>



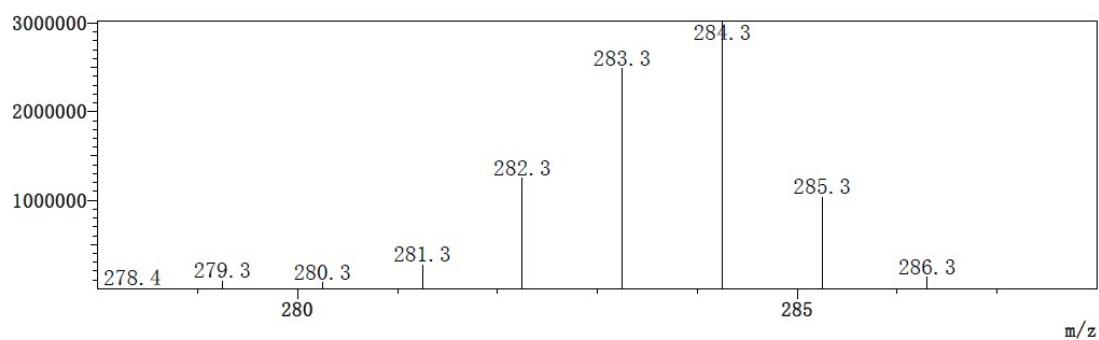
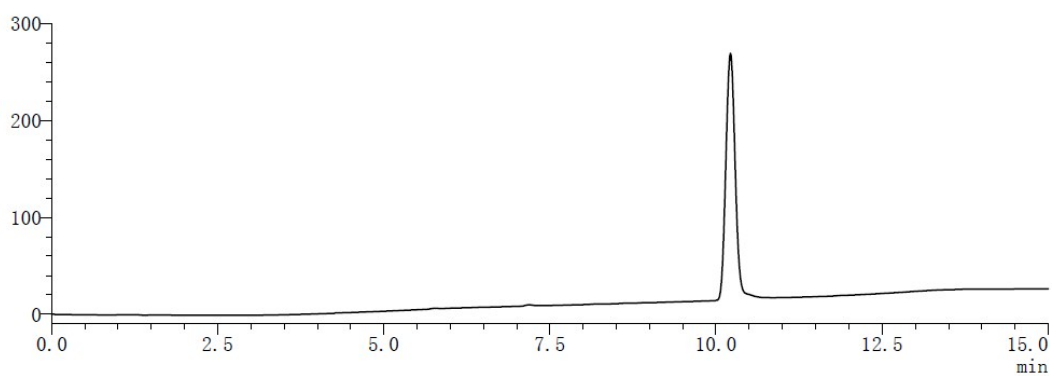
**1c'-d<sub>6</sub>**

mV

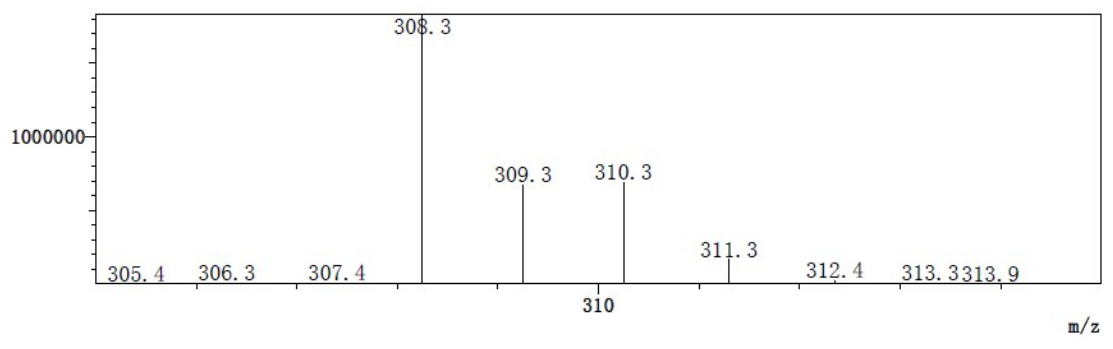
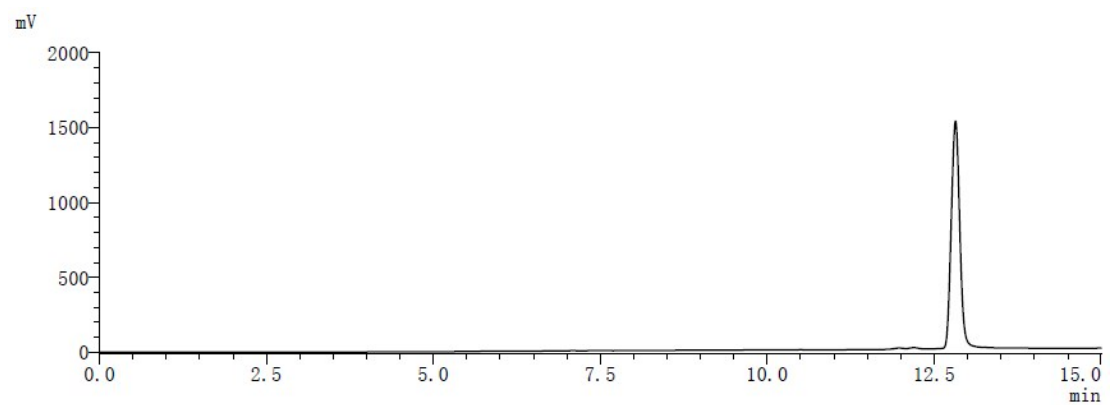


**1d'-d<sub>4</sub>**

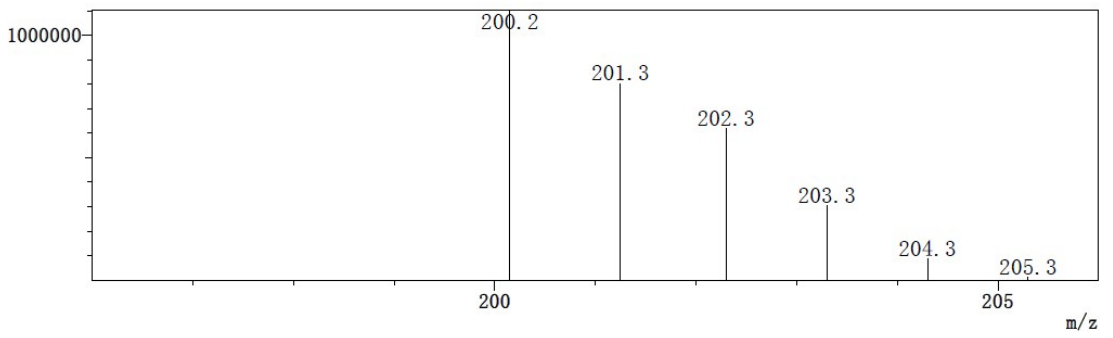
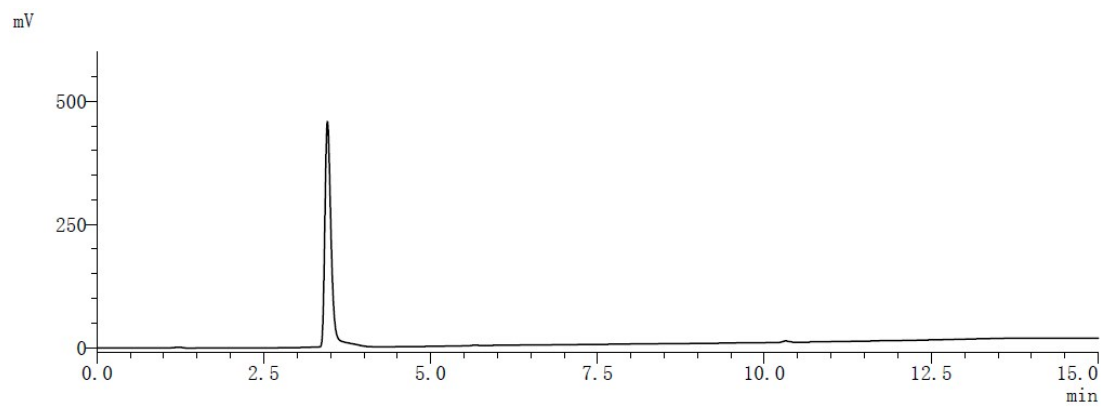
mV



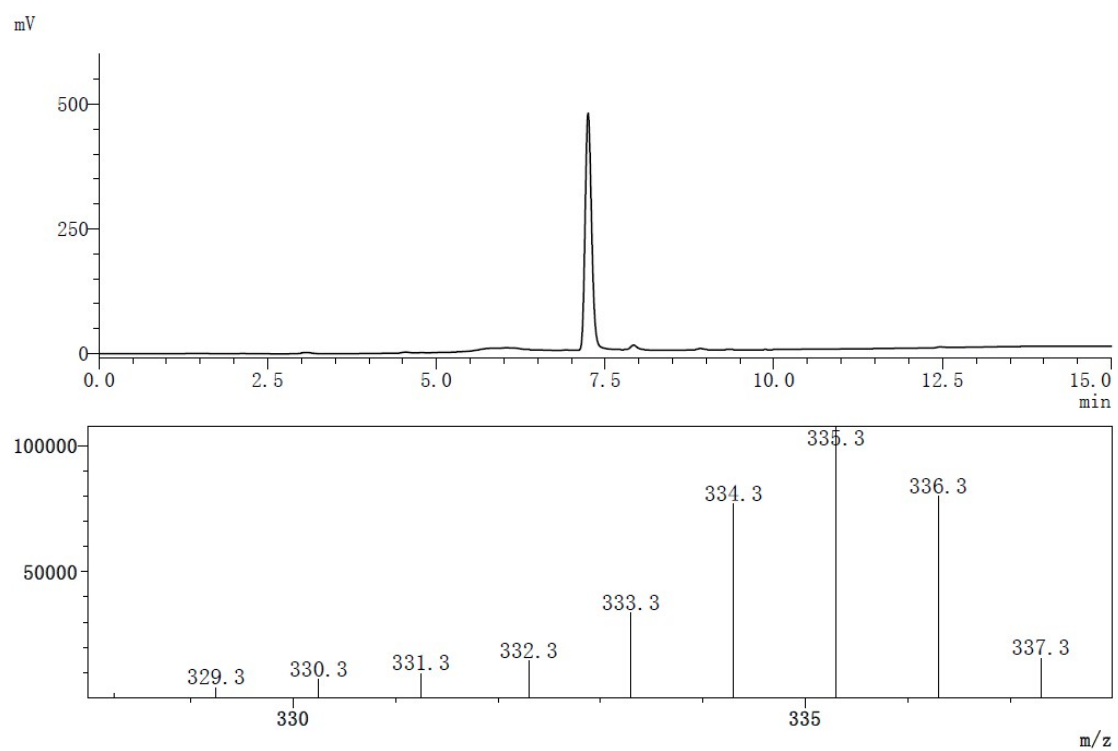
**1e'-d<sub>2</sub>**



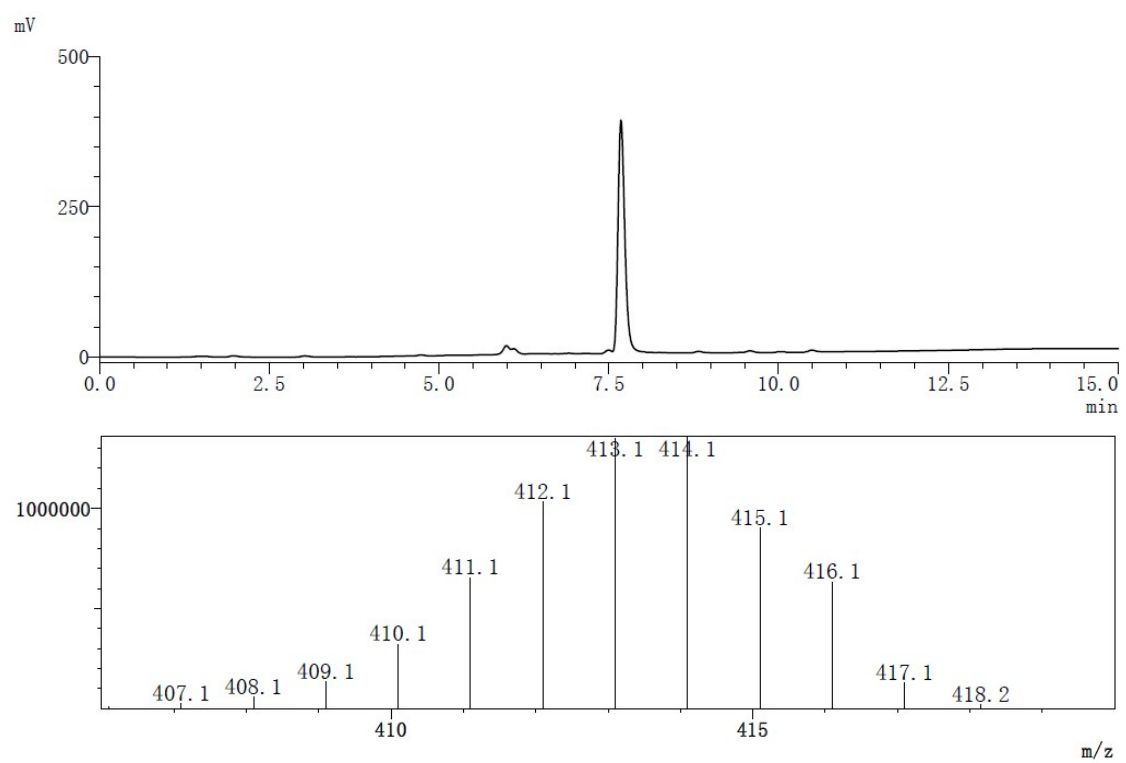
**1f'-d<sub>12</sub>**



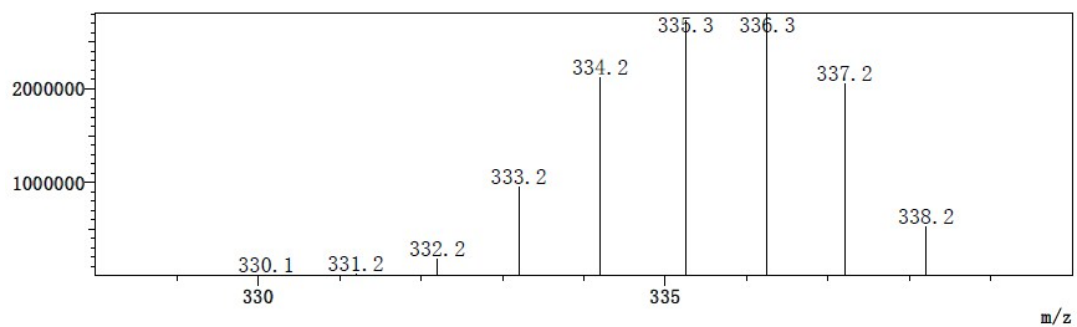
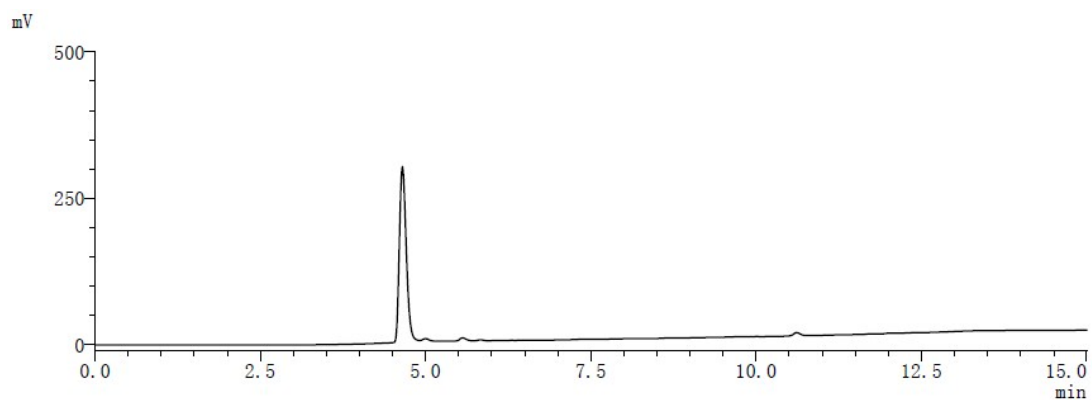
### 2a-d<sub>6</sub>



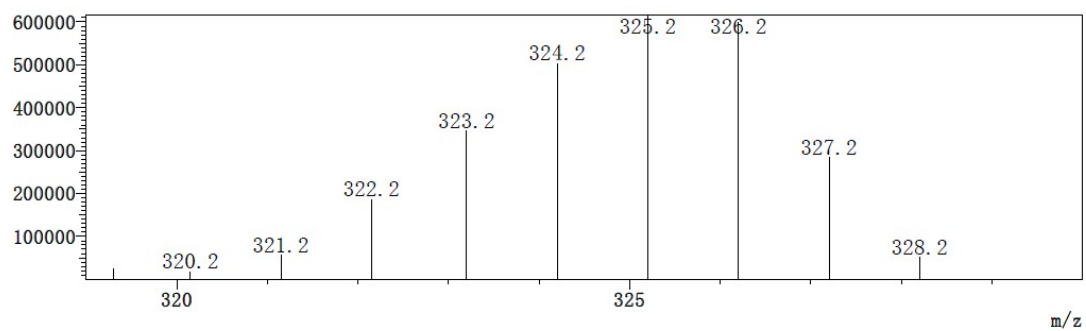
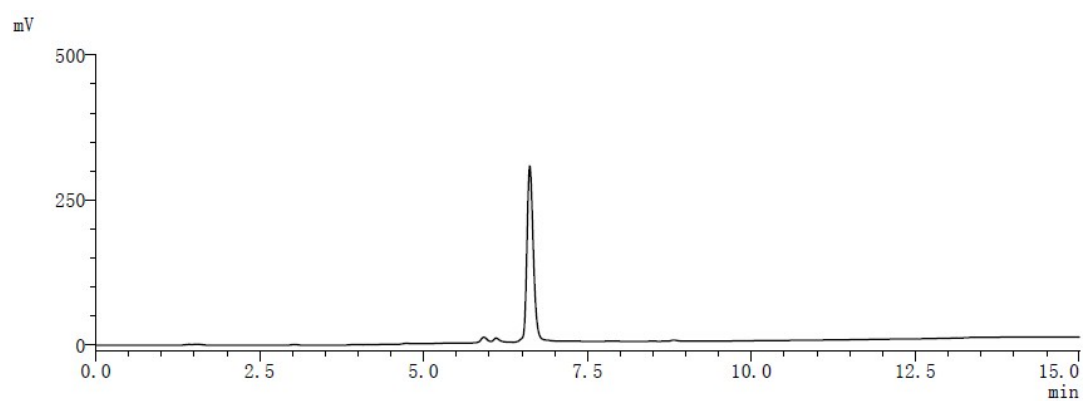
### 2b-d<sub>6</sub>



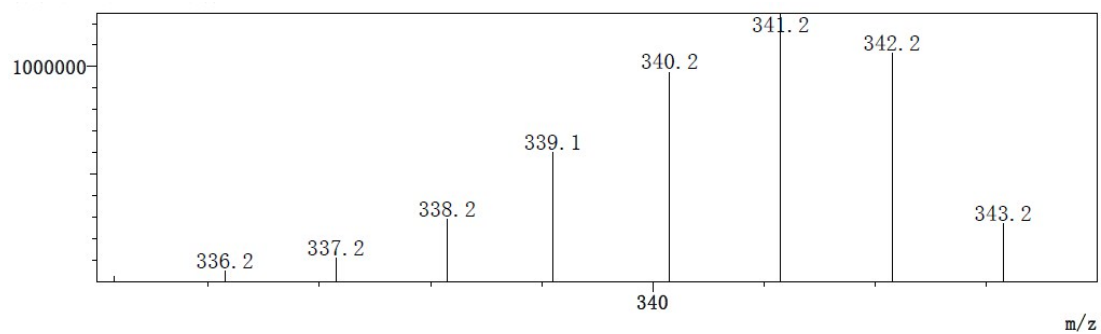
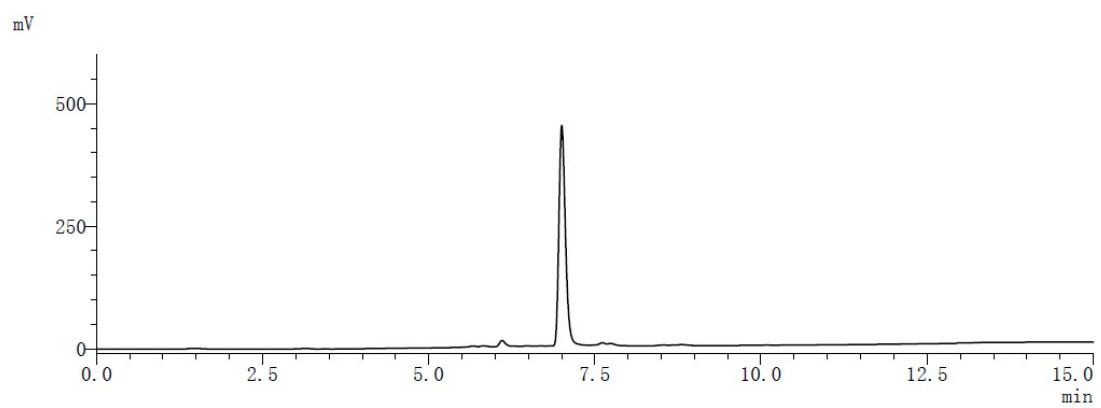
### 2c-d<sub>6</sub>



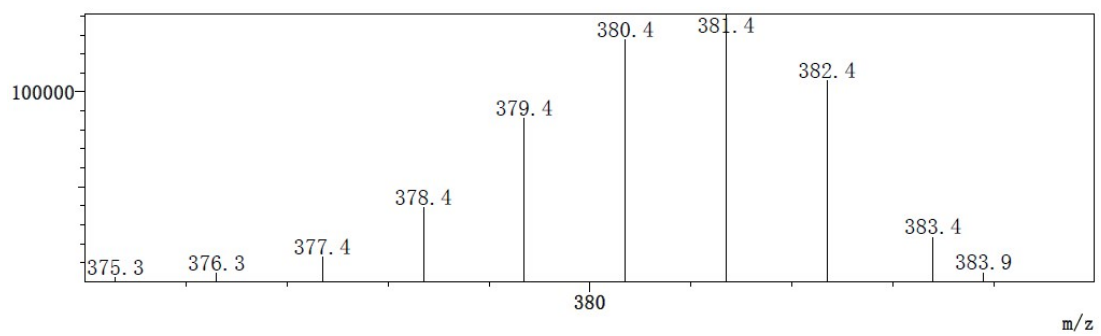
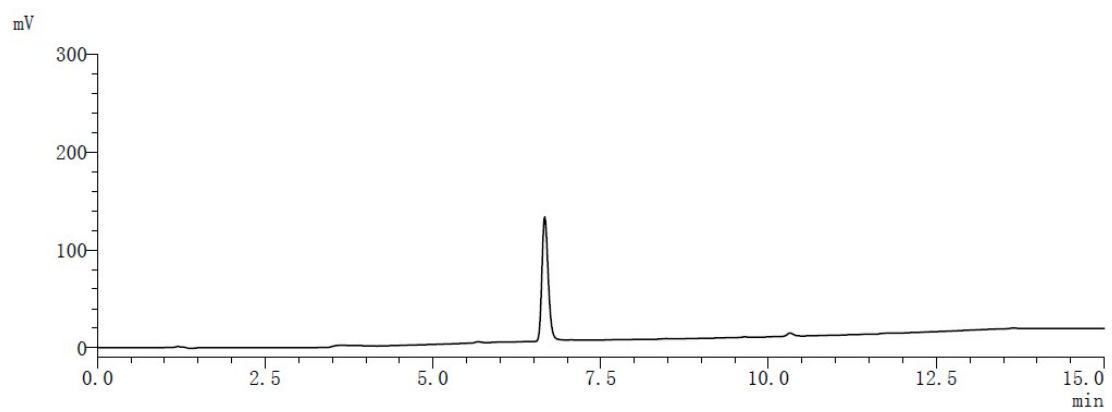
### 2d-d<sub>7</sub>



### 2e-d<sub>6</sub>



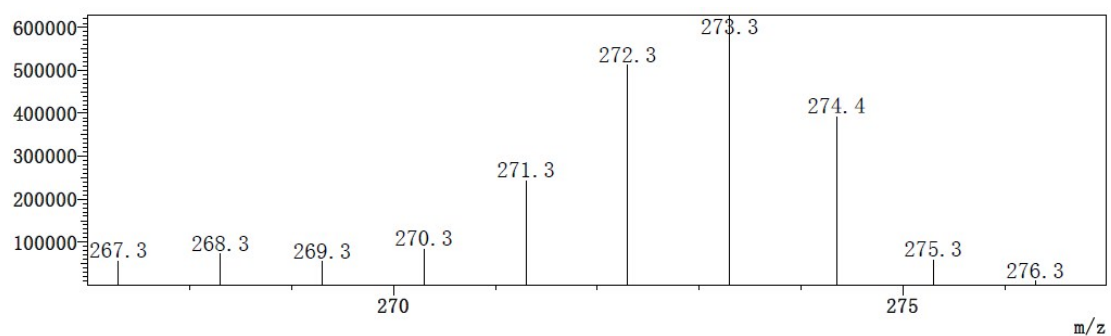
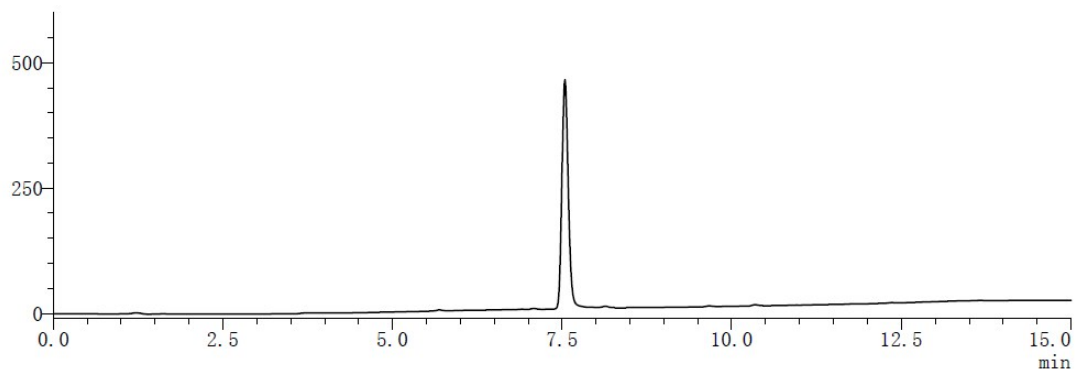
### 2f-d<sub>6</sub>





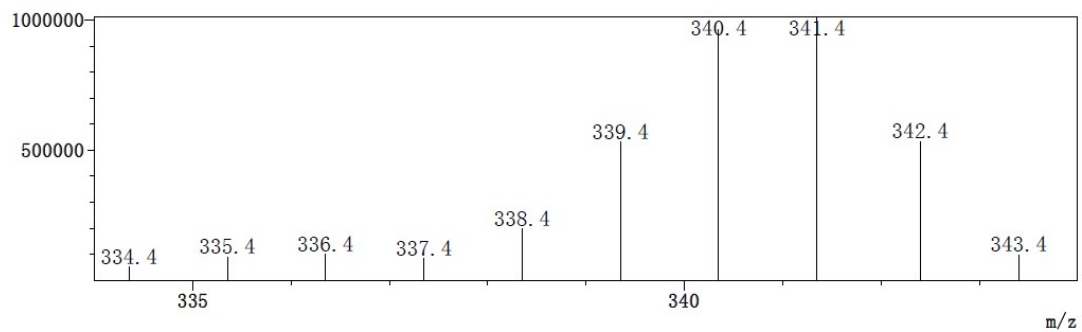
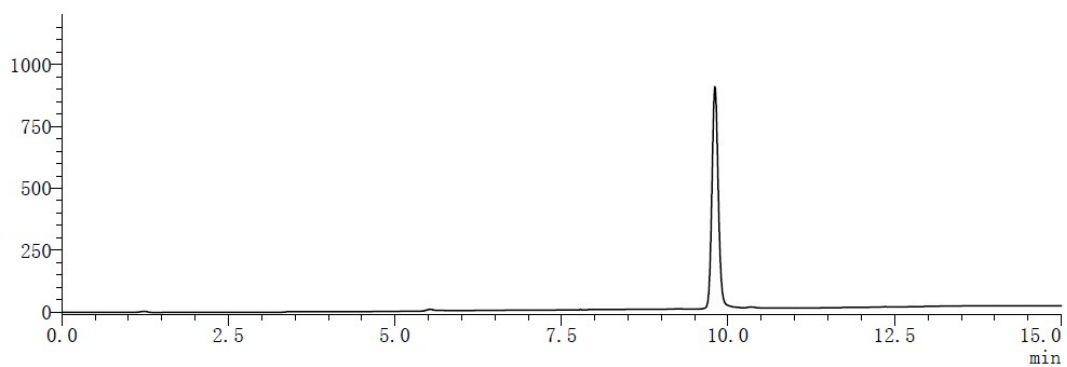
### 2g-d<sub>6</sub>

mV

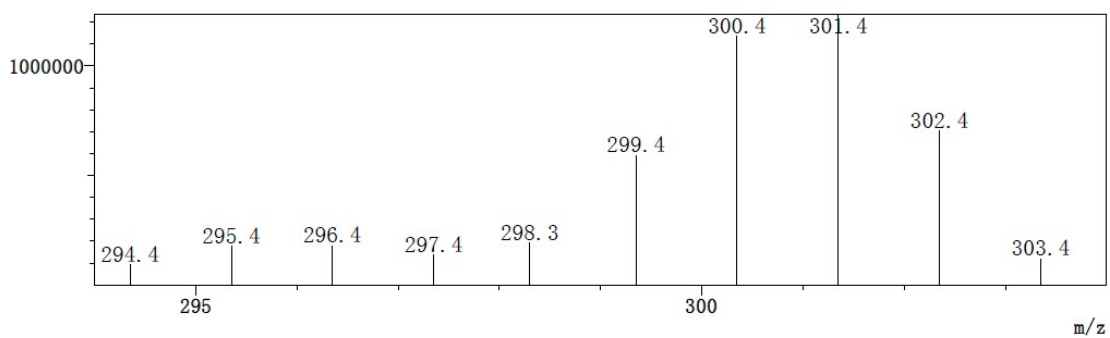
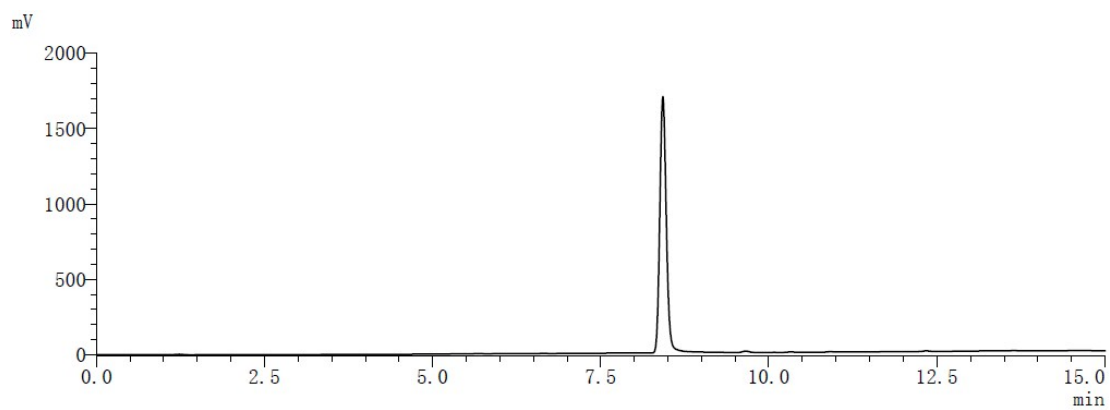


### 2h-d<sub>6</sub>

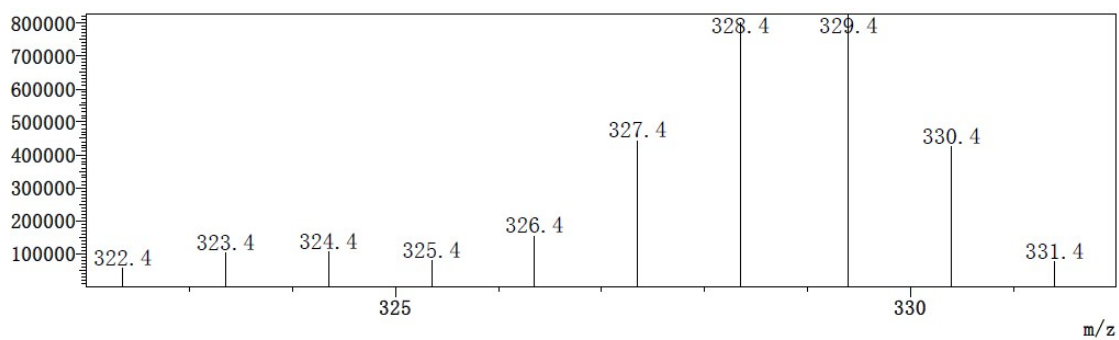
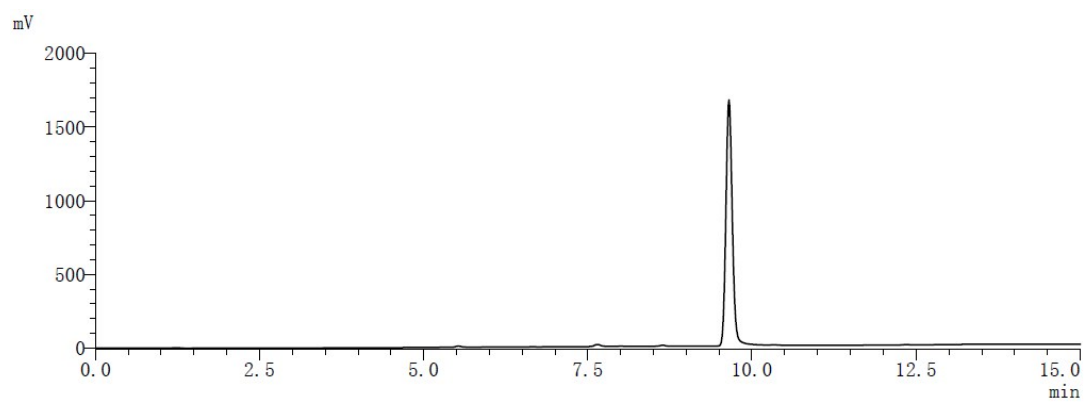
mV



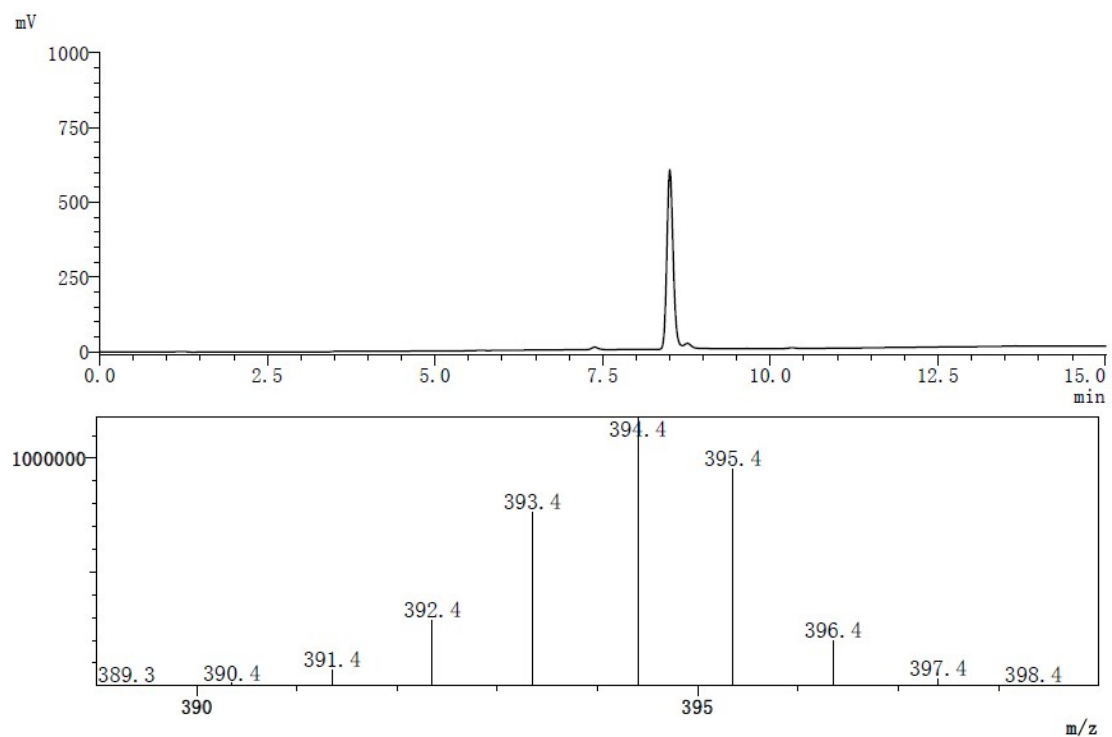
### 2i-d<sub>6</sub>



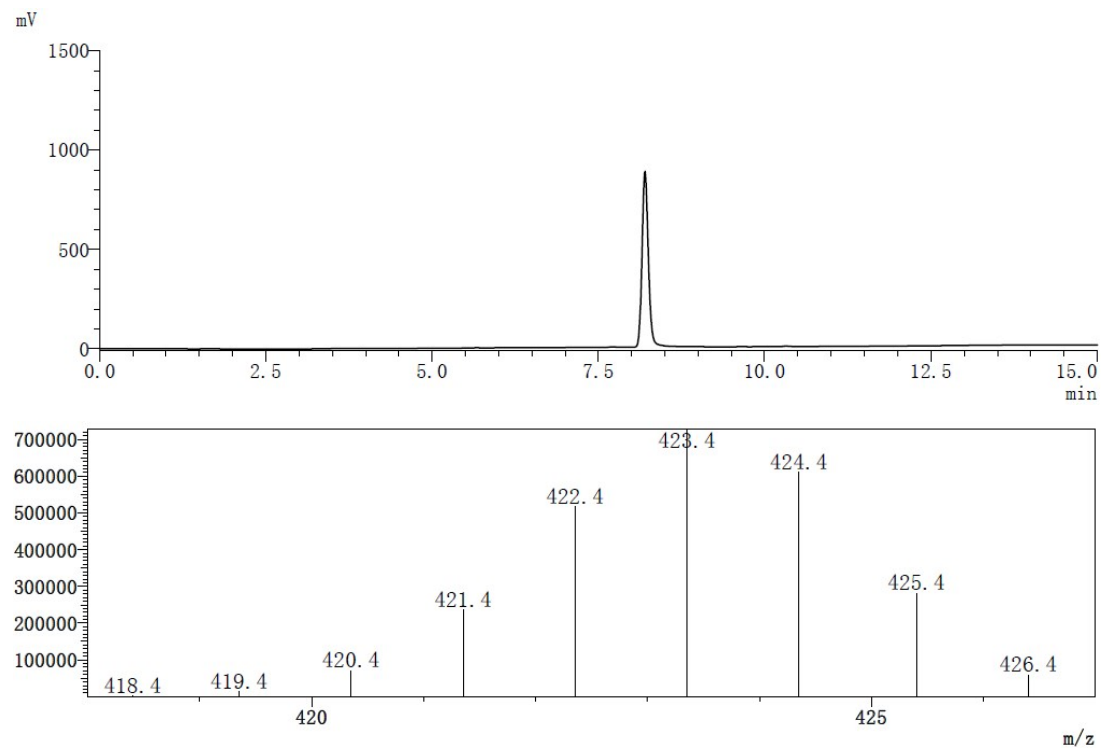
### 2j-d<sub>6</sub>



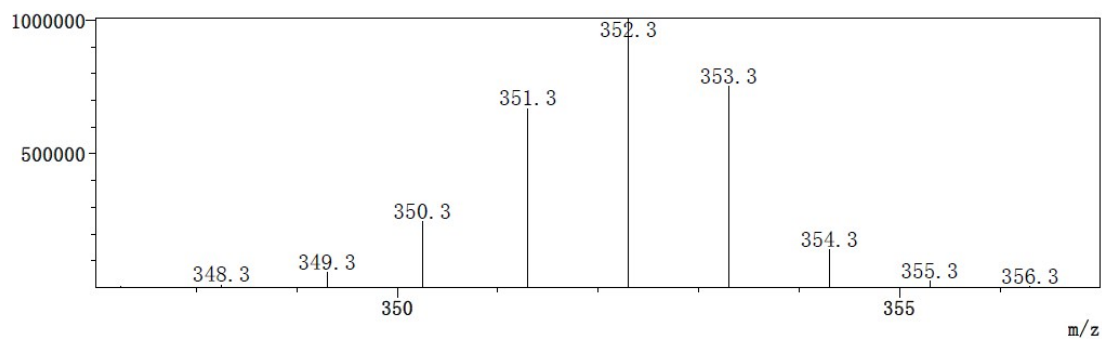
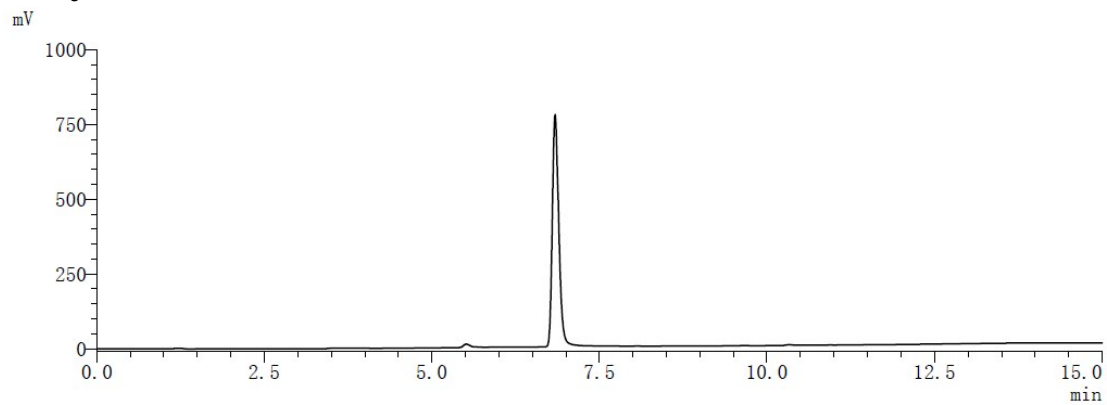
### 2k-d<sub>6</sub>



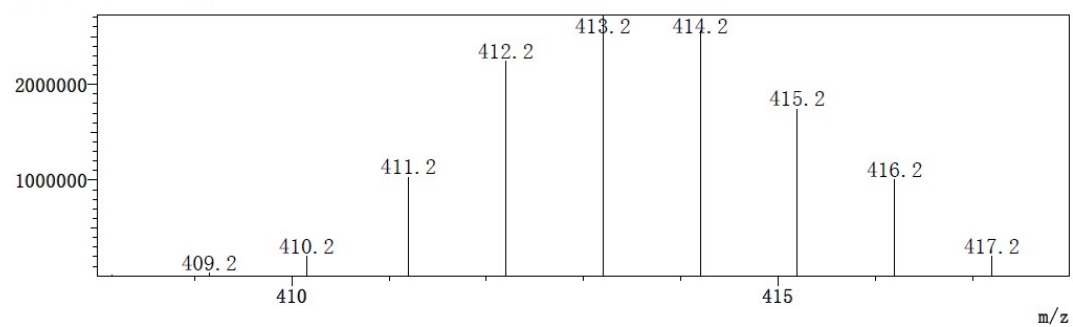
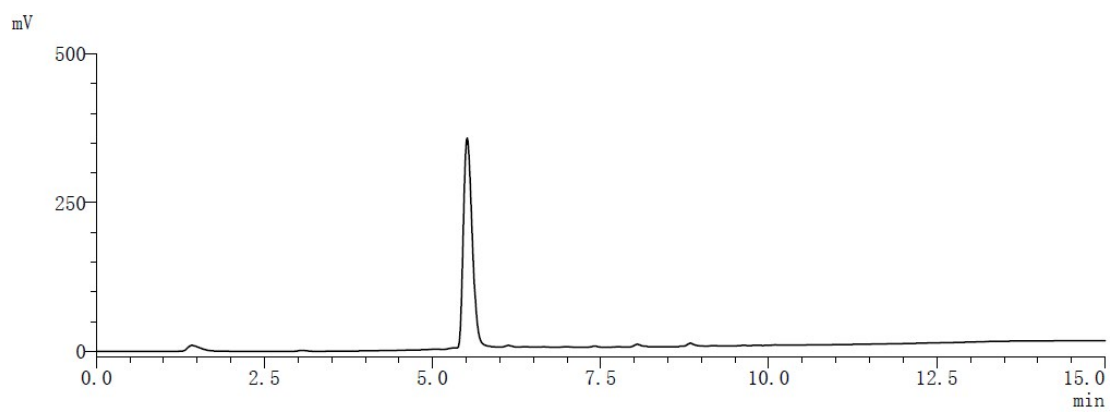
### 2l-d<sub>6</sub>



### 2m-d<sub>6</sub>

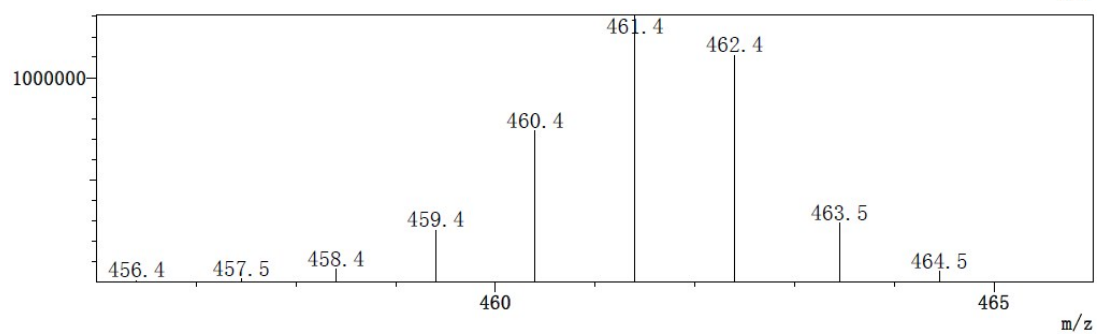
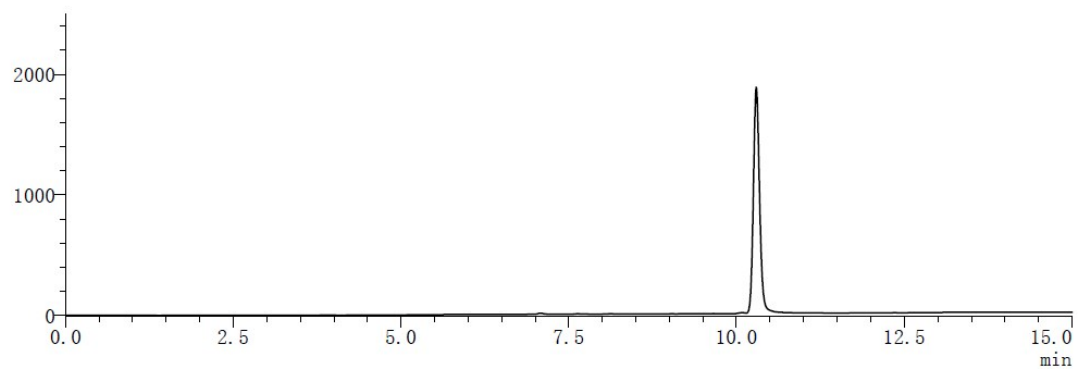


### 2n-d<sub>5</sub>



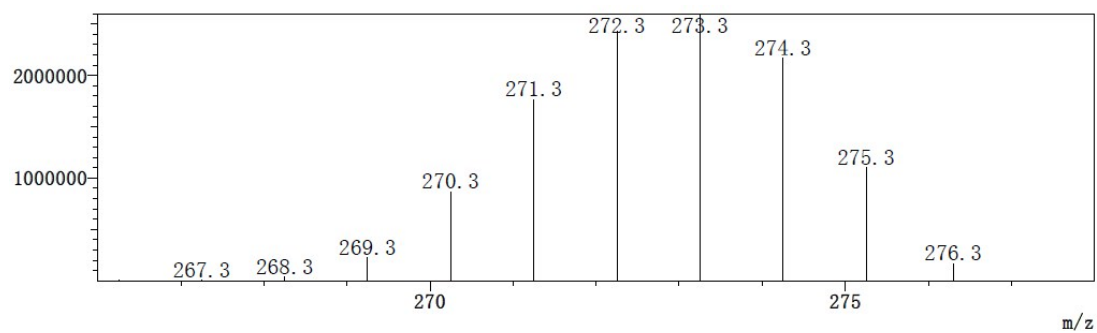
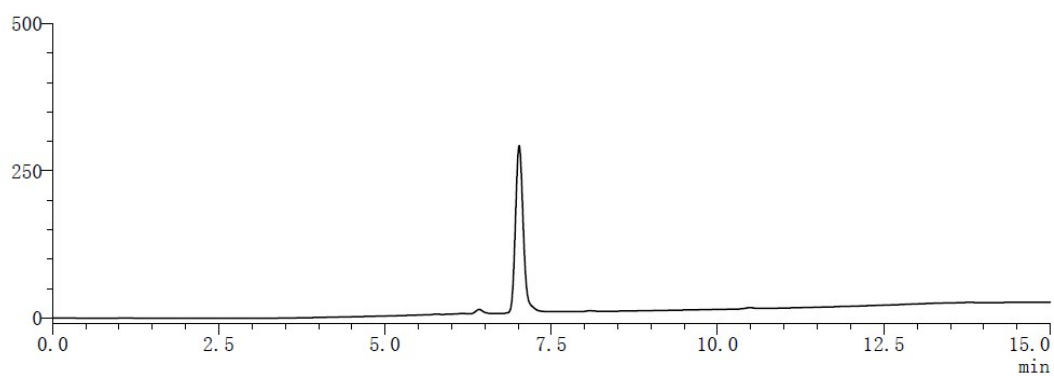
**2o-d<sub>6</sub>**

mV



**3c-d<sub>9</sub>**

mV



### 3d-d<sub>6</sub>

