Electronic Supplementary Information

Ep7GT, a glycosyltransferase with sugar donor flexibility from *Epimedium pseudowushanense*, catalyzes the 7-*O*-glycosylation of baohuoside

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Time	Solvent A (%)	Solvent B (%)	flow rate
(min)	0.1% formic acid	CH ₃ OH	$(mL min^{-1})$
0.00	90.0	10.0	1.0
10.00	50.0	50.0	1.0
30.00	0.0	100.0	1.0
45.00	0.0	100.0	1.0

Table S1. The HPLC method used in this study

Table S2. Details of the plasmids and strains used in this study

plasmids or E. coli	relevant properties	
strain		
	Plasmids	
pET-28a	pBR322 ori, Kan ^r	
	Primers	
pE-7GT forward	CAAATGGGTCGC <u>GGATCC</u> ATGGGTTCAGAAACTCAT	
pE-7GT reverse	GTGGTGGTGGTG <u>CTCGAG</u> TCAGTGTCCAATAGTTGCAG	
	C	
	Strains	
S0	Transetta (DE3) harboring empty pET-28a	
S1	Transetta (DE3) harboring pE-Ep7GT	



Figure S1. SDS-PAGE of recombinant His₆-Ep7GT purified by affinity chromatography. Lane M: Protein Marker; Lane 1: His-tagged Ep7GT (predicted M.W., 54.1 kDa) purified on Ni Sepharose column chromatography.



Figure S2. Enzymatic reactions catalyzed by crude extracts of recombinant EpGTs with baohuoside (1) as sugar accepter and UDP-glucose as sugar donor. A) EpGT8 (Ep7GT); B) EpGT2; C) EpGT4; D) MS spectra of **1a** and **1** at negative mode.

EpGT8 AtUGT73C6	NGSETHQLHALFFPFNAHGHM PM DI ARI FSVRG LKSTI I TTPQHATNI TSK I CRYQNSGLDI KI I T NAFEKNNEPEPI HEVI FPENAOGHM PNVDI ARI LAORG VI I TI VTTPHNAAREKNV I NRALESGLPI NLVO	68 72
EpPF3RT	MSCI SVVKNGTSSI HI AMFPWLPFGHVNPFI QLSNQLAANG YTI SFLTTRDNLPKI KPHNHFPDQI HI VPLDVKPP	76
EpGT2		68
EpGT4 EpGT1	KI OA EKPHI VCI PYPAOGUVDEVSIVE UKELIAKG FHI TEVNEENHRILLE SKOPDALKGVSDE	67
EpGT3	MEKKTI VLYPSPGNCILI SNVELGKLI TKRYPSFSI ALLI TPPPYYTGSTAPY. I DRVSKTNPSI I FHY	68
EpGT5	VSKSSTGPHI LVFPFPAQ <mark>GI</mark> MLPLLDLTHQLALRNLTI TI LVTPKNLSLLSPLLSLYPSI QPLV	64
EpG16 EpG17	NALFGEAKEEKI NVLLVAF SAQGELNPVLRLAKRLACKG LQVTLATTEI VREMKKS KNI DPSTKPTI TI AP	72
EpGT9	ADPNTNOSPPHVALLPSSCMCLTPFLRLAATLSTLN CKITFITHPTVSLAESOL JSRFLFVFP0TPRO	72
EpGT8	LPEPSEEFGLPKGCESVDSLPSRDVLENFFDAVAKLOLEFERILEELHP. DCLVSDVFLPWTND. VACKHGTPRLLFHG	145
AtUGT73C6	VKFPYQEAGLQEGQENNDLLTTMEQITSFFKAVNLLKEPVQNLIEEMSPRPSCLISDNCLSYTSE. I AKKFKIPKILFHG	151
EpPF3RT	PFPFANGQPPPSSAGAPPPADSNSPGLALLQLILATDSLEDEVESQLALIKPDIJIYEFAHYIPA. I ANRLGIKSAFYCV	155
EpG12 EpG14	LPTP PLSSSTPASNG TOVLEELLL. ENNPHYHEALTISISSMIPAPTIDUPCSCSEE.TANOLNIPATYTT IDVIDISA IDVINSANKETVARGILLI ISNDPUVKALVSISATSTP AFTIDUPCSSSPETAROUNIPATYTET	139
EpGT1	FETI POCLPPS DKDATODVPKLCDS VR. KNCLVPF VELVKKLNS SPDVPKVTCI I SDGVNSF GVQ. AAOKLGLPEVQFWI	145
EpGT3	LPAI PFDS. I PTSNSSYRETI I FELLR LSNPHVHEALLTI STYATI P AFI VDFFCFSSLE. LANOLNI PGYFFLT	141
EpGT5	LPFPNNSSLPPGVENAKDLSPI YFIPLIRVIGDLYTPLLQWFQSHPSPP. CAILSDCFLGVTHH. LATQLGIPRIVFSP	141
EpGT0 EpGT7	STQL DEETS DOESD THEN SLDTTFD. CLORE OP QUESTICE OEKRYS CLYNNER PERMAD, VAAQUKTE CAND WI LPALEPENS I PESSISYERTI FELLE. LSNE HVER ALLTI STYATTP. AFL VDFCCFSSLE LANOL NI PGYFLIT	147
EpGT9	FHLLPLDPSTLNSTDP FFLQFETI R QSAHLLSPLLSSSSPPLSA LI TDI TLASAFI PI TASLHLPNYLLFT	143
EpGT8	T SF <mark>FS</mark> LCVANVTREYSVYE SE GETFLL <mark>P</mark> GL <mark>P</mark> D EI KMKKSML <mark>P</mark> SHLG. SKDRFGEMVDRI RDTE	207
AtUGT73C6	MG. CFCLLCVNVLRKNREI LDNLKSD	217
EpPF 3R1 EpGT2	IS ALAVAYHLVPACQPKSVD	220
EpGT4	SOVSUSLEHII PTI HEKTTESF KDN	203
EpGTI	ASACGFNGYLNYRELLKRGLTPLKDESYLTNGFLDNPI DWYPGMP DI RLKDFPSFNRTTDPNDI MLDFLGEEAQNC	221
EpGT3	SGI SFLSI FLHLPTVDQNTTLSFKDM. NTLI DF GVPPIPSSDMF DP VLDR. TDKAYEW VDCAKDFP	207
EpGT5 EpGT6	S GAF ALQI I HSLF QDLPKDDNP	207
EpGT7	SGI SFLSI FLHLPTVDQNTTLSFKDN	207
EpGT9	SSARMLS VCASF PNI DI SSI D EI QI PGSS PVPKS WVPPLLLNA. TNLFTTQFI ENGQKLV	202
EpGT8	VTS YGVLVNS F YELLEP AYADHYRNVLGRRAW, HI GPVSLSSNNI I DKAQRGKKAGI DEHY CLAWINSKEKDSWI YVSF	284
EnPE3RT	KISTONI WAS GELEFA TANDE NEARSONAW. TI OF VS CONVOVDARAEGONS DI DUDE. CLEMEDS NEPCONDITIVE SOSDILI METSKEMSKY ONVIKEOV OKPUVLAGI SI PE PETDDIEDR WESKI GOFAPESKI VVSF	294
EpGT2	KA. RGLI VNTFESLEPRALKAI TOGLSAPNG. PTPPVYCI GPVI SP SNQNASKPA CLO <mark>MU</mark> DKQPRK <mark>SV</mark> VFLCF	276
EpGT4	KS. QGI I VNSFDSLEPRALKAVRDGLSVPDA. PTPPVYCI GPLI ATDDRSGTEAGSGQAS. CLEWDTQPSKSVPLCF	283
EpGT1 EpGT3	LNATATI I NTEDDLE QEVLDAT SSKEPETY, TI GPLSVLSSNEPKSDI NSLRSNLWKEDTDCLKWI DEKEPDSMI YVNY KS. POCIVINSEOTTOPPATKAT TOCI SVINS. PTPDI VPLCPLI ASEDDSCGE AGCI CEVP. CLEWIDT OPSDB	299
EpGT5	S VS WGVVLNT FELLES VLOHWKS	275
EpGT6	DKLKWVLVNSFNELENDVVESNVGLSPVRAI GPLVFSLLLGDQESD DVGVDNWKPDETCI K <mark>WU</mark> DEKPPS <mark>V</mark> I VVSF	292
EpGT7 EpGT9	KS. RGVI VNSFOTLEPRALKAI TDGLSVPNS. PTPPI YPI GPLI AS EAGI GKVP. CLEWLDLOPSRSWVFLCF	277
Lpon/		2/2
EpGT8		350
AtUGT73C6	GSI CNLPLS OLLELGLEES ORP I WIRG. WEKYKELVEWFSES OF EDRI ODROLLI KONSPONLI S	363
EpPF3RT	GSQDVLSKEQITELVLCLEESGVPENAVLK	353
EpG12 EpGT4	OSNGAF SKAQNKEI ANGLEKS GORDLWVR. SNGAF SKAQNKEI ANGLEKS GORDLWVR.	343
EpGTI	GS VTVNS EKDLI EF AVGLANTKLPELWI VR. PDVVNGDS AAL POEF MEI TGRGET SN. WCPCDOVIJS	365
EpGT3	GSNGLFSGAQLKEI AI GLEKSGQRELWVVRSPPTSDESSKGI AATPEPDLDALLPDGFLERTKERGLVVKSWAPGVEVLK	363
EpGT5	GSQTVLNNKOVEQLALCI EQS GTREI WCAKEP	345
EpGT7	SITE SALE ALCENSE AND LEVEN AND A SALE AND A	357
EpGT9	GSRTANSREQIRELGDCLVRSGCKFLWVKDKKVDKEDEEGVEEVLGSELNDKVKEKCLVVKSVVDCEKIUN	344
EpGT8	HPAI GGE VTECGWNSI LE GVS AGVPMI TWPLF AEGEMNEALI TQVNNI GWKVGVERWSDWTEQG HVLVTKETVKKVVN	428
AtUG173C6 EpPE3RT	HPS VGOPLIFIC GWAS ILD GITAGLPMLIWPLF AD PENEKUWULLKVGVS AV KEVNKWGEBERI GVLVD KEVKAVE	443
EpGT2	KESVGGEVTICGWSVLEAVIAGVPWGWPLVAECRLNRVLLVEEWVALSMEE. RKDG. FVTAEEVEKKVR	413
EpGT4	NESLGGF VTHCGWNS TLEALLAGI PNVAWPLYAEQRI NRVLLVEEI KVALSMEE GVDG F VT AQE VEKKVR	432
EpGT1	HPSVGGFLTHCGWSTLBSISSGVPVVWPFFAEQCINCRVACHWGIGVEIDN. NVKRDEVEKIVR	431
EpGT5	REAVOOR VIECOWSI VERVAAVENVAMELDAERE DATI LVDHIGVARVCE G PRTVPDSGELAO	433
EpGT6	HQAI TCF I TECEWS LLE TVTCGVPVI AF POWTOPITNAKL VEDVF RVGVRFQCD	429
EpGT7	KEAVGOF VTECOWS VLEAVCAGVP WAWPLDAE ORI NRVVI VEELKLALSMVE	427
EPO19		410
EpGT8	OLNAT FECEELENRARM KGLARKAVKEDOSSETDITELEELHHAATLG	478
AtUGT73C6	ELNGESDDAKERRRAKELGESAHKAVEEG <mark>GSS</mark> HSNI TFLLQDI NQLAQSN	494
EpPF3RT	SLAVEVDG. EVGKEI RGNHAKLRDMLLDKETQSGYLEQVLEELEKLAKGV	473
EpGT2 EpGT4	GLWES, EGGS VI RORI VKLSDEANOAT LEAGSS VNALVINL VES WKL	458 477
EpGT1	EVNEG. EEGKKMRTKALDWKEKVFNACKEGGSSYKNFDRFVKDVLKLSN.	479
EpGT3	GLNES. EEGNLI KERVVNLSGEAKAAVDKDGSSEKSLSDLADLWKTGNRDS	483
EpGT5 EpGT6	AF WKS VS DKS LEKVRI EDLCKAAAGAI AN. SSEGDF VKALCEL VVVDG. PNSRELKRALELKEAARKAVADGGSSDRNI OFFTDETS VDDS	454 477
EpGT7	GLNES. EEGNLI KERVVNLSGEAKAAVDKDGSSEKSLSDLADLWKTGNRDS	477
EpGT9	EMNGD EKLKAEAARVREEARKAVGSGGSSNGGL/KCLO/WNKKGHHD	463

Figure S3. Multiple alignment of the amino acid sequences of EpGT8 (renamed Ep7GT), AtUGT78D1 (GenBank accession number NM_102790, from *Arabidopsis thaliana*) and EpPF3RT (GenBank accession number MG264429, from *E. pseudowushanense*) and other EpGTs.



Figure S4. Ep7GT-catalyzed glycosylation of 8-prenylkaempferol (**3**) with different sugar donors. A) UDP-glucose; B) UDP-galactose; C) UDP-*N*-acetylglucosamine; D) UDP-xylose; E) MS spectra of **3** and the main products at positive mode.



Figure S5. Ep7GT-catalyzed glycosylation of anhydroicaritin (4) with different sugar donors. A) Control group; B) UDP-glucose, C) TDP-glucose; D) MS spectra of the main products at positive mode.



Figure S6. Ep7GT-catalyzed glycosylation of baohuoside II (**5**) with different sugar donors. A) Control group; B) UDP-glucose, C) UDP-*N*-acetylglucosamine; D) UDP-xylose; E) MS spectra of the main products at positive mode.



Figure S8. ¹³C NMR spectrum of icariin (1a) in DMSO- d_6





Figure S10. ¹³C NMR spectrum of 1b in DMSO-d₆



Figure S11. HSQC spectrum of 1b in DMSO- d_6



Figure S12. HMBC spectrum of 1b in DMSO- d_6



Figure S14. ¹³C NMR spectrum of 1c in DMSO-*d*₆



Figure S15. HSQC spectrum of 1c in DMSO- d_6



Figure S16. HMBC spectrum of 1c in DMSO- d_6



Figure S18. ¹³C NMR spectrum of 2a in DMSO- d_6



Figure S19. HSQC spectrum of 2a in DMSO- d_6



Figure S20. HMBC spectrum of 2a in DMSO-d₆





Figure S22. ¹³C NMR spectrum of 2b in DMSO-*d*₆



Figure S23. HMBC spectrum of 2b in DMSO- d_6



Figure S24. ¹H NMR spectrum of 2c in Methanol- d_4



Figure S26. HSQC spectrum of 2c in Methanol- d_4



Figure S27. HMBC spectrum of 2c in Methanol- d_4



Figure S28. The HRESIMS spectrum of 1b







Figure S30. The apparent K_m values of recombinant Ep7GT for baohuoside (A), kaempferol (B), UDP-glucose (C), UDP-*N*-acetylglucosamine (D) and UDP-xylose (E).

Table S3. The apparent K_m , K_{cat} and K_{cat}/K_m values of recombinant Ep7GT for different sugar donors with 1 as the accepter.

Sugar donors	K_m (μ M)	$K_{cat}(S^{-1})$	K_{cat}/K_m (S ⁻¹ mM ⁻¹)
UDP-glucose	146.9	0.13	0.88
UDP-N-acetylglucosamine	348.0	0.014	0.04
UDP-xylose	541.3	0.038	0.07



Figure S31. The linear regression models and the regression equations of the external standard method established for the quantitative analysis of baohuoside (A) and icariin (B).



Figure S32. Exploring the catalytic reversibility of Ep7GT when **1a** and UDP were used as substrate.



Figure S33. Time course assay of 1a with whole cell catalyst of engineered *E. coli*.