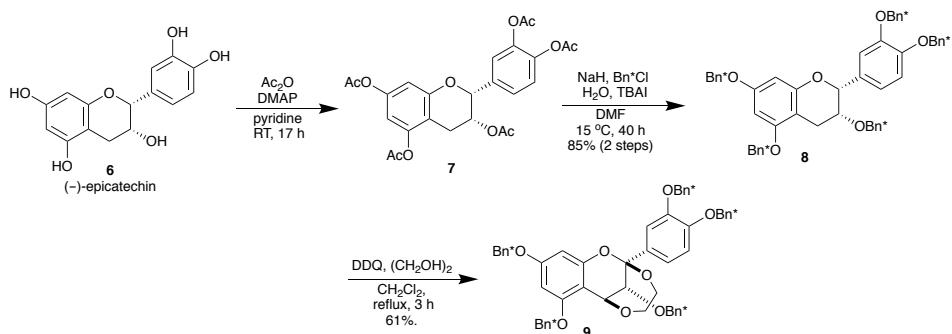


## Supporting Information

### General

All reactions utilizing air- or moisture-sensitive reagents were performed in flame-dried glasswares under an atmosphere of dry argon. Ethereal solvents and dichloromethane (anhydrous; Kanto Chemical Co., Inc.) were purified under argon, using an Organic Solvent Pure Unit (Wako Pure Chemical Industries, Ltd.). For thin-layer chromatography (TLC) analysis, Merck pre-coated plates (TLC silica gel 60 F254, Art 5715, 0.25 mm) were used. Silica-gel preparative thin-layer chromatography (PTLC) was performed using plates prepared from Merck Silica gel 60 PF254 (Art 7747). For flash column chromatography, silica gel 60N (Spherical, neutral, 63–210 µm) from Kanto Chemical was used. Melting point (mp) determinations were performed by using a METTLER TOLEDO MP70 melting point system and are uncorrected. <sup>1</sup>H-, and <sup>13</sup>C-NMR were measured on a Bruker Avance III (600 MHz) spectrometer equipped with the cold probe (CryoProbe Prodigy<sup>TM</sup>) and in the solvent indicated; Chemical shifts (*d*) are expressed in parts per million (ppm) downfield from internal standard (tetramethylsilane 0.00 ppm) or referenced to residual undeuterated solvents as internal standard. All coupling constants (*J*) are reported as hertz (Hz). Splitting patterns are indicated as follows: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad. Infrared (IR) spectra were recorded on a Thermo SCIENTIFIC NICOLET iS5 FT-IR spectrometer. Attenuated total reflectance Fourier transform infrared (ATR-FTIR) spectra were recorded by using Thermo SCIENTIFIC NICOLET iS5 FTIR spectrometer equipped iD5 ATR accessory. High-resolution mass spectra (HRMS) were obtained with Bruker Daltonics micrOTOF-Q II. Optical rotations ([ $\alpha$ ]<sub>D</sub>) were measured on a JASCO P-3000 polarimeter. High performance liquid chromatography (HPLC) analyses were performed on a LC-Net II/ADC controller (JASCO) equipped with a Jasco PU-2080 Plus Intelligent Pump, a Jasco MD-2010 Plus Multiwavelength Detector, a Jasco DG-2080-54 degasser and LG-2080-02 Ternary Gradient Unit. Preparative HPLC separation was performed on a LC-Net II/ADC controller (JASCO) equipped with a Jasco PU-2086 Plus Intelligent Prep Pump, a Jasco UV-1575 UV/Vis Detector and a Jasco DG-2080-54 degasser.

Preparation of acetal **9**



To a solution of **(-)**-epicatechin (**6**) (1.0 g, 3.4 mmol) in pyridine (3 mL) was added  $\text{Ac}_2\text{O}$  (1.95 mL, 20.7 mmol) and DMAP (12.6 mg, 0.103 mmol) at 0 °C and stirred for 17 h at room temperature. The reaction was quenched by adding water and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with 1 M HCl, brine, dried ( $\text{Na}_2\text{SO}_4$ ), and concentrated in vacuo to give penta-acetate **7** as white solid which was used for the next reaction without further purification. To a solution of crude **7** in DMF (20 mL) at 0 °C was added NaH (1.8 g, 63% dispersion in mineral oil, 46 mmol) followed by tetrabutylammonium iodide (TBAI) (260 mg, 0.703 mmol). After stirring for 10 mins at 0 °C,  $\text{C}_6\text{D}_5\text{CD}_2\text{Cl}$  ( $\text{Bn}^*\text{Cl}$ ) (2.1 mL, 18 mmol) was slowly added and the mixture was stirred at 15 °C for 40 h. The reaction was quenched at 0 °C by addition of water and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with brine, dried ( $\text{Na}_2\text{SO}_4$ ) and concentrated in vacuo. The residue was purified by flash column chromatography (hexane/EtOAc = 4/1) to afford **8** (2.4 g, 85%) as a white amorphous foam.

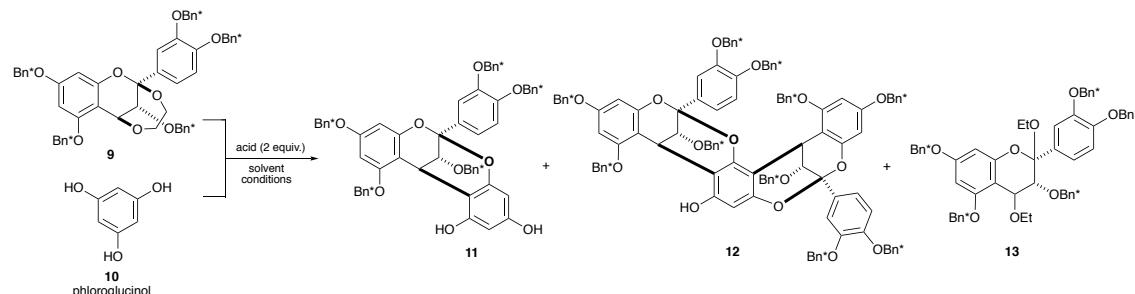
**8:**  $R_f = 0.78$  (EtOAc/hexane = 1/2);  $[\alpha]_D^{20} = -14$  (*c* 0.80,  $\text{CHCl}_3$ );  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  2.80 (dd, *J* = 17.4, 4.2 Hz, 1H), 3.00 (dd, *J* = 17.4, 3.0 Hz, 1H), 3.94 (brs, 1H), 4.97 (s, 1H), 6.27 (d, *J* = 2.4 Hz, 1H), 6.29 (d, *J* = 2.4 Hz, 1H), 6.93 (s, 1H), 6.94 (s, 1H), 7.20 (s, 1H);  $^{13}\text{C}$  NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  27.7, 72.6, 78.3, 93.9, 94.9, 101.7, 114.1, 115.0, 120.0, 126.8–128.4, 132.4, 136.9, 137.0, 137.2, 137.3, 138.0, 148.6, 149.0, 155.8, 158.2, 158.8; IR (neat) 2894, 2272, 2179, 2074, 1616, 1590, 1513, 1492, 1356, 1275, 1236, 1119, 1031, 1014, 1001, 815, 806, 771  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_{50}\text{H}_{10}\text{D}_{35}\text{O}_6$  [ $(\text{M}+\text{H})^+$ ]  $m/z$  776.5407, found  $m/z$  776.5437.

To a solution of **8** (1.0 g, 1.3 mmol) and ethylene glycol (120 mg, 1.93 mmol) in  $\text{CH}_2\text{Cl}_2$  (50 mL) was added DDQ (1.2 g, 5.2 mmol) at room temperature, and then the mixture was refluxed for 3 h. Heating was stopped and reaction was quenched by addition of DMAP (1.27 g, 10.3 mmol) at

0 °C, and the stirring was continued for 30 min. The resulting precipitates were filtered off through a Celite® pad (washed with CH<sub>2</sub>Cl<sub>2</sub>). The filtrate was washed with saturated aqueous NaHCO<sub>3</sub> solution, H<sub>2</sub>O, brine, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated in vacuo. The residue was purified by flash column chromatography (hexane/EtOAc = 2/1) to afford **9** (0.66 g, 61%) as an ivory amorphous foam.

**9:**  $R_f$  = 0.52 (EtOAc/hexane = 1/2);  $[\alpha]_D^{20}$  = -63 (*c* 0.89, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 3.36 (m, 2H), 3.97–3.99 (m, 1H), 4.01 (d, *J* = 1.8 Hz, 1H), 4.06–4.10 (m, 1H), 5.14 (d, *J* = 1.8 Hz, 1H), 6.30 (d, *J* = 1.8 Hz, 1H), 6.33 (d, *J* = 1.8 Hz, 1H), 6.92 (d, *J* = 7.8 Hz, 1H), 7.09 (dd, *J* = 7.8, 1.8 Hz, 1H), 7.19 (d, *J* = 1.8 Hz, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 65.8, 68.1, 70.5, 78.3, 95.3, 95.5, 100.4, 100.8, 114.3, 114.4, 119.6, 126.8–128.4, 135.1, 136.5, 136.8, 137.0, 137.2, 137.3, 148.8, 149.1, 155.1, 159.5, 161.2; IR (neat) 3008, 2920, 2280, 2196, 1616, 1590, 1183, 1162, 1141, 1054, 1032, 819, 754 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>52</sub>H<sub>12</sub>D<sub>35</sub>O<sub>8</sub> [(M+H)<sup>+</sup>] *m/z* 834.5462, found *m/z* 834.5496.

#### Model annulation study with phloroglucinol (**10**)



run	molar ratio of <b>9/10</b>	promotor	solvent (0.5 M)	conditions	<b>11</b> /%	<b>12</b> /%	<b>13</b> /%
1 <sup>a</sup>	1/1	PPTS	EtOH	RT, 24 h	—	—	—
2 <sup>b</sup>	1/1	PPTS	EtOH	reflux, 5 min	49	—	12
3	1/1	PPTS	EtOH	reflux, 0.5 h	50	24	—
4	1/1	CSA	EtOH	RT, 24 h	65	14	16
5	1/1	CSA	dioxane	RT, 24 h	66	8	—
6	1/1	CSA	EtOH, dioxane	RT, 24 h	84	7	—
7	1/1.5	CSA	EtOH, dioxane	RT, 24 h	98	1	—
8	1.5/1	CSA	EtOH, dioxane	RT, 24 h	52	44	—

a) Recovery of acetal **9**. b) 36% of **9** was recovered.

#### *Procedure for entries 1, 4, 5 and 6*

To a solution of acetal **9** (20 mg, 0.024 mmol) and phloroglucinol (**10**) (3.0 mg, 0.024 mmol) in solvent (as mentioned in table, 0.5 M) was added acid (as mentioned in table, 0.048 mmol) at room temperature. After stirring for 24 h at the same temperature, the reaction was diluted with EtOAc and washed with saturated aqueous NaHCO<sub>3</sub> solution, brine, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated in vacuo. The residue was purified by PTLC (hexane/EtOAc = 7/3) to afford the products as mentioned in the table.

*Procedure for entries 2, 3*

To a solution of acetal **9** (20 mg, 0.024 mmol) and **10** (3.0 mg, 0.024 mmol) in ethanol (0.5 mL) was added PPTS (12 mg, 0.048 mmol) and stirred at refluxing temperature as per mentioned in table. It was allowed to cool to room temperature, diluted with EtOAc and washed with saturated aqueous NaHCO<sub>3</sub> solution, brine, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated in vacuo. The residue was purified by PTLC (hexane/EtOAc = 7/3) to afford the product as mentioned in the table.

*Procedure for the entry 7*

To a solution of acetal **9** (20 mg, 0.024 mmol) and **10** (4.5 mg, 0.036 mmol) in mixture of ethanol and 1,4-dioxane (v/v = 1, 0.5 mL) was added CSA (11 mg, 0.048 mmol) at room temperature. After stirring for 24 h at the same temperature, the reaction mixture was diluted with EtOAc and washed with saturated aqueous NaHCO<sub>3</sub> solution, brine, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated in vacuo. The residue was purified by PTLC (hexane/EtOAc = 7/3) to afford **11** (21.0 mg, 98%) and **12** (0.4 mg, 1%).

*Procedure for the entry 8*

To a solution of acetal **9** (29.7 mg, 0.0356 mmol) and **10** (3.0 mg, 0.024 mmol) in mixed solvent ethanol and 1,4-dioxane (0.5 mL, v/v = 1) was added CSA (16.7 mg, 0.0719 mmol) at room temperature. After stirring for 24 h at the same temperature, the reaction mixture was diluted with EtOAc and washed with saturated aqueous NaHCO<sub>3</sub> solution, brine, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated in vacuo. The residue was purified by PTLC (hexane/EtOAc = 7/3) to afford to afford **11** (11.0 mg, 52%) and **12** (17.5 mg, 44%) as amorphous foam.

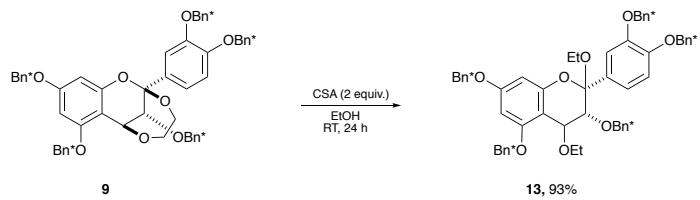
**11:**  $R_f$  = 0.32 (EtOAc/hexane = 3/7);  $[\alpha]_D^{20} = -43$  (c 0.41, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 3.82 (d,  $J$  = 3.0 Hz, 1H), 4.35 (d,  $J$  = 3.0 Hz, 1H), 4.68 (brs, 1H, OH), 6.01 (s, 1H), 6.02 (s, 2H), 6.34 (d,  $J$  = 3.6 Hz, 1H), 6.42 (d,  $J$  = 3.6 Hz, 1H), 6.96–6.98 (m, 2H), 7.24 (d,  $J$  = 1.8 Hz, 1 H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 25.6, 72.4, 95.1, 96.1, 96.3, 98.0, 98.7, 105.1, 106.1, 114.4, 114.7,

120.7, 127.0–128.8, 131.9, 135.0, 136.5, 137.0, 137.1, 137.1, 148.6, 149.6, 152.7, 153.5, 154.5, 155.4, 156.1, 159.1; IR (neat) 3416, 2310, 2180, 1604, 1505, 1419, 1274, 1181, 1136, 1084, 1038, 819, 542 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>56</sub>H<sub>12</sub>D<sub>35</sub>O<sub>9</sub> [(M+H)<sup>+</sup>] *m/z* 898.5415, found *m/z* 898.5382.

**12:** R<sub>f</sub> = 0.55 (EtOAc/hexane = 3/7); [α]<sub>D</sub><sup>20</sup> = -19 (*c* 0.82, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 3.81 (d, *J* = 3.0 Hz, 1H), 3.84 (d, *J* = 3.6 Hz, 1H), 4.37 (d, *J* = 3.0 Hz, 1H), 4.98 (d, *J* = 3.0 Hz, 1H), 5.96 (d, *J* = 1.8 Hz, 1H), 6.15 (s, 1H), 6.23 (d, *J* = 2.4 Hz, 1H), 6.26 (d, *J* = 1.8 Hz, 1H), 6.20 (d, *J* = 2.4 Hz, 1H), 6.91 (s, 1H), 6.95 (d, *J* = 1.8 Hz, 1H), 6.96 (d, *J* = 1.8 Hz, 1H), 7.19–7.22 (m, 2H), 7.25 (d, *J* = 1.2 Hz, 1H), 7.37 (d, *J* = 1.2 Hz, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 25.5, 25.9, 72.4, 72.9, 94.5, 94.9, 95.3, 96.1, 98.2, 98.4, 98.6, 105.0, 105.1, 106.4, 106.7, 114.4, 114.5, 114.5, 115.5, 1210.6, 120.6, 127.0–128.8, 132.5, 132.6, 135.0, 135.6, 136.8, 137.1, 137.2, 137.4, 138.1, 148.5, 141.6, 149.4, 149.6, 152.4, 152.8, 153.4, 143.7, 155.4, 157.6, 158.9, 159.1; IR (neat) 3419, 3011, 2359, 2341, 2275, 2188, 1617, 1508, 1490, 1418, 1273, 1183, 1140, 1085, 1052, 1036, 960, 819, 751, 541 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>106</sub>H<sub>17</sub>D<sub>70</sub>O<sub>15</sub> [(M+H)<sup>+</sup>] *m/z* 1670.0433, found *m/z* 1670.0453.

**13:** R<sub>f</sub> = 0.83 (EtOAc/hexane = 3/7); [α]<sub>D</sub><sup>20</sup> = -27.8 (*c* 1.45, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 0.96 (t, *J* = 4.8 Hz, 3H), 1.01 (t, *J* = 4.8 Hz, 3H), 3.20–3.23 (m, 1H), 3.35–3.42 (m, 2H), 3.50–3.52 (m, 1H), 3.69 (d, *J* = 1.2 Hz, 1H), 4.37 (d, *J* = 1.2 Hz, 1H), 6.28 (d, *J* = 2.4 Hz, 1H), 6.30 (d, *J* = 2.4 Hz, 1H), 6.96 (d, *J* = 7.8 Hz, 1H), 7.15 (d, *J* = 7.8 Hz, 1H) 7.16 (s, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 15.4, 15.7, 58.1, 64.5, 70.5, 94.7, 95.28, 101.2, 103.8, 114.4, 120.9, 127.3–128.3, 133.1, 136.8, 137.0, 137.2, 137.3, 148.6, 148.9, 153.0, 159.8, 160.6; IR (neat) 2912, 2848, 1666, 1650, 1643, 1580, 1538, 1503, 1462, 1454, 1159, 1089, 503 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>54</sub>H<sub>18</sub>D<sub>35</sub>O<sub>8</sub> [(M+H)<sup>+</sup>] *m/z* 864.5931, found *m/z* 864.5890.

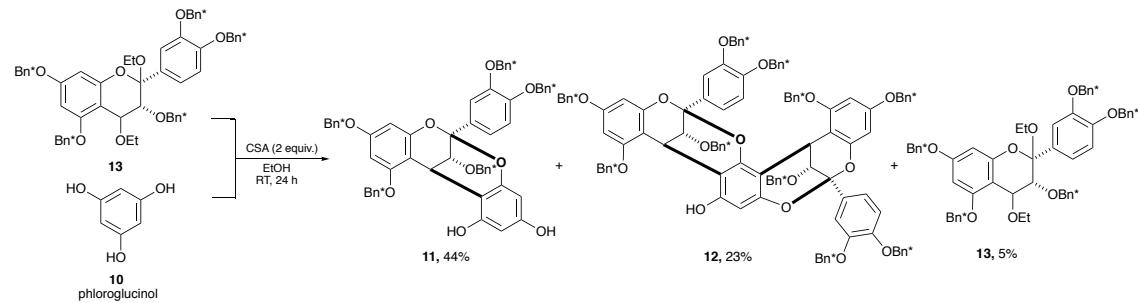
#### Preparation of 2,4-diethoxy flavan **13** from **9**



To a solution of acetal **9** (20 mg, 0.024 mmol) in mixture of ethanol and 1,4-dioxane (v/v = 1, 0.5 mL) was added CSA (11 mg, 0.048 mmol) at room temperature. After stirring for 16 h at the same

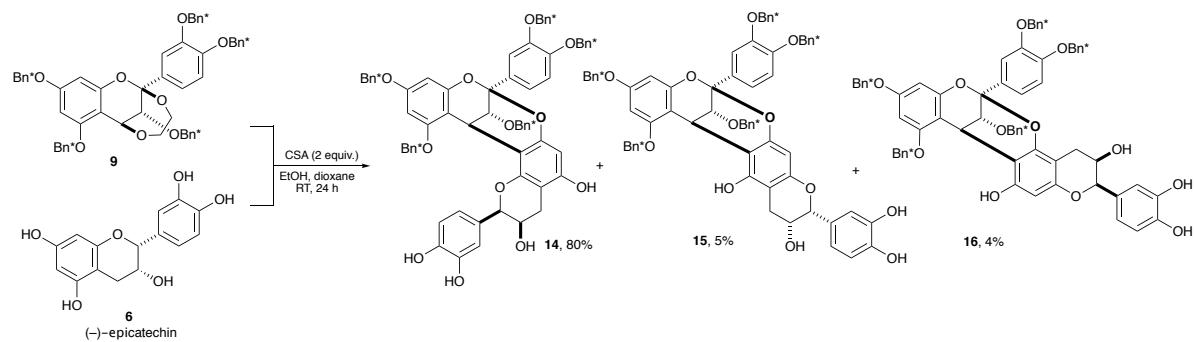
temperature, the reaction mixture was diluted with EtOAc and washed with saturated aqueous NaHCO<sub>3</sub> solution, brine, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated in vacuo. The residue was purified by PTLC (hexane/EtOAc = 8/2) to afford **13** (19.3 mg, 93%).

#### Annulation of 2,4-diethoxy flavan **13** with phloroglucinol (**10**)



To a solution of 2,4-diethoxy flavan **13** (21.6 mg, 0.0248 mmol) and **10** (3.0 mg, 0.024 mmol) in ethanol (0.5 mL) was added CSA (11 mg, 0.048 mmol) at room temperature. After stirring for 24 h at the same temperature, the reaction was diluted with EtOAc and washed with saturated aqueous NaHCO<sub>3</sub> solution, brine, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated in vacuo. The residue was purified by PTLC (hexane/EtOAc = 7/3) to afford to afford **11** (9.7 mg, 44%), **12** (4.5 mg, 23%), and recovered starting material **13** (1.2 mg, 5%).

#### Direct coupling of **9** with (-)-epicatechin (**6**)



To a solution of acetal **9** (50 mg, 0.059 mmol) and (-)-epicatechin **6** (26 mg, 0.090 mmol) in mixture of ethanol and 1,4-dioxane (v/v = 1, 1.25 mL) was added CSA (28 mg, 0.12 mmol) at room temperature. After stirring for 24 h at the same temperature, the reaction was diluted with EtOAc, washed with saturated aqueous NaHCO<sub>3</sub> solution, brine, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated in vacuo. The residue was purified by PTLC (CH<sub>2</sub>Cl<sub>2</sub>/EtOAc = 7/3) to afford dimer **14** (50.8 mg,

80%) as a white amorphous form along with mixture of **15** and **16** which was separated by PTLC ( $\text{CH}_2\text{Cl}_2/\text{EtOAc} = 7/3$ , two runs) giving **15** (3.2 mg, 5%) and **16** (2.7 mg, 4%).

**14:**  $R_f = 0.20$  ( $\text{EtOAc}/\text{CH}_2\text{Cl}_2 = 3/7$ );  $[\alpha]_D^{20} = -59$  ( $c 0.74, \text{CHCl}_3$ );  $^1\text{H}$  NMR (600 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  2.45 (dd,  $J = 16.2, 6.6$  Hz, 1H), 2.90 (dd,  $J = 16.2, 5.4$  Hz, 1H), 3.87 (d,  $J = 6.6$  Hz, 1H), 4.18 (brs, 1H), 4.84 (d,  $J = 2.4$  Hz, 1H), 5.13 (d,  $J = 3.6$  Hz, 1H), 6.08 (s, 1H), 6.16 (d,  $J = 1.8$  Hz, 1H), 6.29 (d,  $J = 1.8$  Hz, 1H), 6.60 (d,  $J = 8.4$  Hz, 1H), 6.62 (d,  $J = 8.4$  Hz, 1H), 6.83 (s, 1H), 7.00 (d,  $J = 8.4$  Hz, 1H), 7.22 (d,  $J = 8.4$  Hz, 1H), 7.26 (s, 1H);  $^{13}\text{C}$  NMR (150 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  27.5, 28.4, 67.9, 75.8, 80.9, 96.5, 96.7, 96.8, 100.8, 103.4, 106.9, 108.0, 116.1, 116.8, 116.8, 121.0, 122.5, 128.2–129.9, 132.2, 134.8, 139.2, 139.3, 139.5, 139.9, 146.5, 146.5, 150.2, 151.3, 153.2, 154.3, 156.2, 157.1, 159.5, 160.8; IR (neat) 3400, 2276, 2116, 1615, 1509, 1494, 1470, 1327, 1183, 1144, 1115, 1095, 1052, 959, 833, 819, 754, 542  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_{65}\text{H}_{20}\text{D}_{35}\text{O}_{12}$   $[(\text{M}+\text{H})^+]$   $m/z$  1062.5884, found  $m/z$  1062.5862.

**15:**  $R_f = 0.28$  ( $\text{EtOAc}/\text{CH}_2\text{Cl}_2 = 3/7$ );  $[\alpha]_D^{20} = -33$  ( $c 0.82, \text{CHCl}_3$ );  $^1\text{H}$  NMR (600 MHz,  $\text{D}_2\text{O}$  in  $\text{CDCl}_3$ )  $\delta$  2.80–2.89 (m, 2H), 3.81 (d,  $J = 3.6$  Hz, 1H), 4.15 (brs, 1H), 4.35 (d,  $J = 3.6$  Hz, 1H), 4.80 (s, 1H), 6.19 (s, 1H), 6.34 (d,  $J = 1.8$  Hz, 1H), 6.42 (d,  $J = 1.8$  Hz, 1H), 6.81 (s, 1H), 6.96 (d,  $J = 7.8$  Hz, 2H), 7.23 (d,  $J = 7.8$  Hz, 1H), 7.26 (2H, overlapped with CH of  $\text{CHCl}_3$ );  $^{13}\text{C}$  NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  25.9, 28.2, 66.8, 72.4, 78.2, 95.2, 96.2, 96.8, 98.7, 101.3, 105.2, 106.3, 113.7, 114.5, 115.5, 119.0, 120.8, 123.0–128.8, 131.1, 132.0, 135.1, 136.5, 137.0, 137.1, 137.1, 143.77, 143.80, 148.6, 149.6, 150.6, 151.9, 153.6, 154.2, 155.4, 159.1; IR (neat) 3411 (br), 2922, 1619, 1600, 1510, 1419, 1275, 1194, 1052, 1124, 1038, 960, 838, 819, 752, 543  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_{65}\text{H}_{20}\text{D}_{35}\text{O}_{12}$   $[(\text{M}+\text{H})^+]$   $m/z$  1062.5884, found  $m/z$  1062.5857.

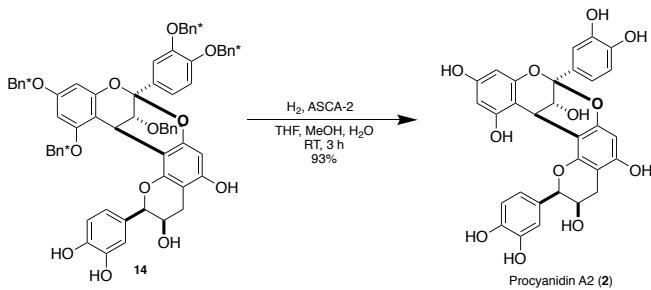
**16:**  $R_f = 0.33$  ( $\text{EtOAc}/\text{CH}_2\text{Cl}_2 = 3/7$ );  $[\alpha]_D^{20} = -51$  ( $c 0.90, \text{CHCl}_3$ );  $^1\text{H}$  NMR (600 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  2.67 (dd,  $J = 16.8, 2.4$  Hz, 1H), 2.73 (dd,  $J = 16.8, 4.8$  Hz, 1H), 3.86 (d,  $J = 3.6$  Hz, 1H), 4.06–4.08 (m, 1H), 4.73 (d,  $J = 3.6$  Hz, 1H), 4.76 (s, 1H), 6.09 (s, 1H), 6.30 (d,  $J = 2.4$  Hz, 1H), 6.35 (d,  $J = 2.4$  Hz, 1H), 6.77 (s, 1H), 6.78 (s, 1H), 6.96 (s, 1H), 7.03 (d,  $J = 8.4$  Hz, 1H), 7.24 (d,  $J = 2.4$  Hz, 1H), 7.28 (dd,  $J = 8.4, 2.4$  Hz, 1H);  $^{13}\text{C}$  NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  27.5, 30.2, 67.8, 72.3, 75.2, 80.3, 80.8, 96.7, 97.5, 99.2, 100.5, 101.6, 107.7, 108.1, 116.0, 116.1, 116.8, 116.9, 120.3, 122.8, 128.9–130.2, 132.9, 134.6, 138.5, 139.2, 139.3, 139.4, 139.4, 146.7, 146.8, 150.3, 151.45, 152.5, 154.7, 155.7, 156.7, 158.4, 161.0; IR (neat) 3412 (br), 2920, 1601, 1508, 1419, 1327, 1274,

1184, 1152, 1084, 1051, 819, 542  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_{65}\text{H}_{20}\text{D}_{35}\text{O}_{12}$   $[(\text{M}+\text{H})^+]$   $m/z$  1062.5884, found  $m/z$  1062.5843.

**General procedure for hydrogenolysis of dimers **14**, **15** and **16**:**

To a solution of dimer (1.0 equiv.) in the presence of ASCA-2 (0.75 equiv.) in a mixture of THF, MeOH and  $\text{H}_2\text{O}$  (v/v/v = 2/2/1, 0.005M) was stirred under  $\text{H}_2$  atmosphere at room temperature. After stirring for 3.0 h, the mixture was carefully filtered through a glass fiber filter (MeOH) under argon atmosphere and the filtrate was evaporated to remove organic solvents and lyophilized to give a crude material, which was further purified by preparative HPLC [Mightysil RP-18GP II, (20 mm f  $\times$  250 mm),  $\lambda$  = 280 nm, 25 °C, MeOH,  $\text{H}_2\text{O}$  = 35/65 containing 0.1% TFA, flow rate 10 mL/min, 20  $\times$  250 mm] and lyophilization to give debenzylated products.

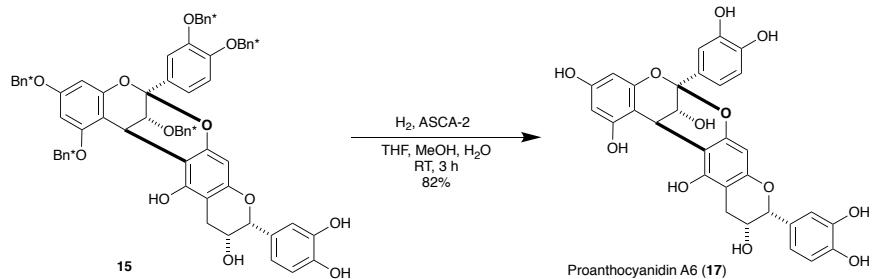
Synthesis of procyanidin A2 (**2**)



Starting from dimer **14** (23 mg, 0.021 mmol) in the presence of ASCA-2 (47 mg, 0.017 mmol), procyanidin A2 (**2**) (11.2 mg, 89%) was obtained as a white amorphous foam.

**2:**  $[\alpha]_{\text{D}}^{20} = +70$  ( $c$  0.10, acetone), [lit.<sup>1</sup>  $[\alpha]_{\text{D}}^{20} = +55.63$  ( $c$  1.08, acetone)];  $^1\text{H}$  NMR (600 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  2.79 (dd,  $J$  = 16.8, 2.4 Hz, 1H), 2.98 (dd,  $J$  = 16.8, 4.8 Hz, 1H), 4.08 (d,  $J$  = 3.6 Hz, 1H), 4.27 (brs, 1H), 4.44 (d,  $J$  = 3.6 Hz, 1H), 4.96 (s, 1H), 6.04 (d,  $J$  = 1.8 Hz, 1H), 6.10 (d,  $J$  = 1.8 Hz, 1H), 6.12 (s, 1H), 6.83 (d,  $J$  = 2.4 Hz, 1H), 6.85 (d,  $J$  = 2.4 Hz, 1H), 7.01 (dd,  $J$  = 7.8, 1.8 Hz, 1H), 7.05 (dd,  $J$  = 7.8, 1.8 Hz, 1H), 7.17 (d,  $J$  = 1.8 Hz, 1H), 7.18 (d,  $J$  = 1.8 Hz, 1H);  $^{13}\text{C}$  NMR (150 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  29.9, 30.1, 68.7, 69.0, 85.4, 97.4, 97.4, 99.0, 101.2, 104.0, 104.9, 107.7, 116.5, 116.6, 116.6, 117.2, 120.7, 121.5, 131.4, 133.2, 146.5, 147.2, 147.6, 147.7, 152.3, 153.1, 155.1, 157.0, 157.6, 159.0; IR (ATR) 3400 (br), 2276, 2116, 1615, 1509, 1494, 1476, 1327, 1183, 1144, 1115, 1052, 959, 838, 819, 754, 542  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_{30}\text{H}_{25}\text{O}_{12}$   $[(\text{M}+\text{H})^+]$   $m/z$  577.1340, found  $m/z$  577.1231.

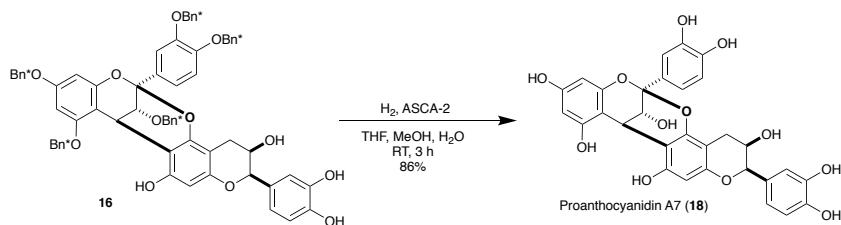
### Synthesis of proanthocyanidin A6 (**17**)



Starting from dimer **15** (20 mg, 0.018 mmol) in the presence of ASCA-2 (41 mg, 0.015 mmol), **17** (9.0 mg, 82%) was obtained as a white amorphous foam.

**17:**  $[\alpha]_{\text{D}}^{20} = +32$  (*c* 0.14, acetone), [lit.<sup>2</sup>  $[\alpha]_{\text{D}}^{21} = +22.4$  (*c* 1.2, acetone)];  $^1\text{H}$  NMR (600 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  2.81 (dd, *J* = 16.8, 3.0 Hz, 1H), 2.96 (dd, *J* = 16.8, 5.4 Hz, 1H), 4.11–4.14 (m, 1H), 4.21 (brs, 1H), 4.31 (d, *J* = 3.6 Hz, 1H), 4.84 (s, 1H), 6.06 (d, *J* = 2.4 Hz, 1H), 6.11 (d, *J* = 2.4 Hz, 1H), 6.14 (s, 1H), 6.78 (d, *J* = 7.8 Hz, 1H), 6.81 (dd, *J* = 8.4, 1.8 Hz, 1H), 6.85 (d, *J* = 8.4 Hz, 1H), 6.99 (d, *J* = 1.8 Hz, 1H), 7.07 (dd, *J* = 8.4, 1.8 Hz, 1H), 7.19 (d, *J* = 1.8 Hz, 1H);  $^{13}\text{C}$  NMR (150 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  30.4, 30.6, 62.4, 68.3, 68.5, 80.8, 97.8, 97.8, 101.4, 103.9, 105.1, 109.7, 116.1, 116.5, 116.7, 116.7, 120.2, 120.8, 132.9, 133.1, 146.5, 146.7, 146.8, 147.7, 152.6, 153.7, 155.3, 156.2, 156.6, 159.0; IR (ATR) 3224 (br), 1606, 1519, 1449, 1352, 1284, 1210, 1106, 969, 867, 820, 793, 714, 654  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_{30}\text{H}_{24}\text{O}_{12}$  ( $[\text{M}-\text{H}]^-$ ) *m/z* 575.1195, found *m/z* 575.1181.

### Synthesis of proanthocyanidin A7 (**18**)

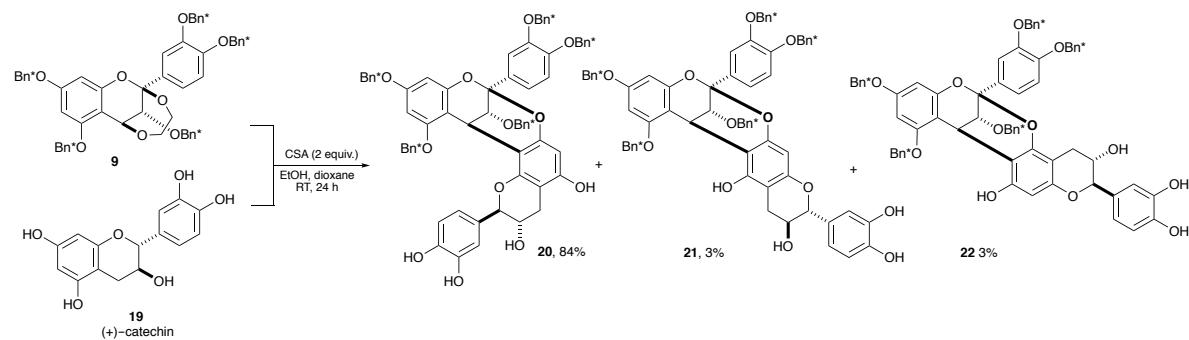


Starting from dimer **16** (19.3 mg, 0.0181 mmol) in the presence of ASCA-2 (39 mg, 0.014 mmol), **18** (8.9 mg, 86%) was obtained as a white amorphous foam.

**18:**  $[\alpha]_{\text{D}}^{20} = +36$  (*c* 0.15, acetone), [lit.<sup>2</sup>  $[\alpha]_{\text{D}}^{21} = +35.9$  (*c* 1.2, acetone)];  $^1\text{H}$  NMR (600 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  2.80 (dd, *J* = 16.8, 3.0 Hz, 1H), 3.01 (dd, *J* = 16.8, 4.2 Hz, 1H), 4.14 (d, *J* = 3.6 Hz, 1H), 4.21 (brs, 1H), 4.32 (d, *J* = 3.6 Hz, 1H), 4.87 (1H, overlapped with HOD), 6.03 (d, *J* = 2.4

Hz, 1H), 6.09 (d,  $J$  = 2.4 Hz, 1H), 6.14 (s, 1H), 6.77 (d,  $J$  = 8.4 Hz, 1H), 6.81 (dd,  $J$  = 8.4, 1.8 Hz, 1H), 6.86 (d,  $J$  = 8.4 Hz, 1H), 6.86 (d,  $J$  = 8.4 Hz, 1H), 6.97 (d,  $J$  = 1.8 Hz, 1H), 7.11 (dd,  $J$  = 7.8, 1.8 Hz, 1H), 7.22 (d,  $J$  = 1.8 Hz, 1H);  $^{13}\text{C}$  NMR (150 MHz, CD<sub>3</sub>OD)  $\delta$  27.9, 28.2, 65.6, 66.3, 78.6, 95.2, 96.3, 96.5, 99.1, 99.6, 103.0, 107.3, 113.8, 114.3, 114.5, 118.0, 118.6, 130.6, 130.9, 144.3, 144.4, 144.6, 145.4, 150.4, 151.1, 152.8, 154.2, 154.7, 156.8; IR (ATR) 3291 (br), 2565, 2187, 1608, 1519, 1443, 1281, 1197, 1078, 1027, 1010, 972, 880, 821, 784, 621 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>30</sub>H<sub>25</sub>O<sub>12</sub> ([M+H]<sup>+</sup>) *m/z* 577.1340, found *m/z* 577.1327.

Direct coupling of **9** with (+)-catechin (**19**)



To a solution of acetal **9** (50 mg, 0.059 mmol) and (+)-catechin (**19**) (26 mg, 0.090 mmol) in mixture of ethanol and 1,4-dioxane (v/v = 1, 1.25 mL) was added CSA (28 mg, 0.12 mmol) at room temperature. After stirring for 24 h at the same temperature, the reaction mixture was diluted with ethyl acetate, washed with saturated aqueous NaHCO<sub>3</sub> solution and brine, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated in vacuo. The residue was purified by PTLC (CH<sub>2</sub>Cl<sub>2</sub>/EtOAc = 7/3) to afford dimer **20** (53.0 mg, 84%) as a white amorphous foam along with mixture of **21** and **22**, which was further separated by PTLC (CH<sub>2</sub>Cl<sub>2</sub>/EtOAc = 7/3, two runs), giving **21** (1.8 mg, 3%) and **22** (1.8 mg, 3%).

**20:**  $R_f$  = 0.22 (EtOAc/CH<sub>2</sub>Cl<sub>2</sub> = 3/7);  $[\alpha]_D^{20} = -74$  (*c* 0.60, CHCl<sub>3</sub>);  $^1\text{H}$  NMR (600 MHz, CD<sub>3</sub>OD)  $\delta$  2.50 (dd,  $J$  = 16.8, 4.2 Hz, 1H), 2.64 (dd,  $J$  = 16.8, 4.8 Hz, 1H), 3.90 (brs, 1H), 4.07 (d,  $J$  = 4.2 Hz, 1H), 4.73 (d,  $J$  = 4.2 Hz, 1H), 5.15 (d,  $J$  = 2.4 Hz, 1H), 6.06 (s, 1H), 6.20 (s, 1H), 6.32 (s, 1H), 6.42 (d,  $J$  = 8.4 Hz, 1H), 6.60 (s, 1H), 6.61 (d,  $J$  = 8.4 Hz, 1H), 7.02 (d,  $J$  = 8.4 Hz, 1H) 7.23 (d,  $J$  = 8.4 Hz, 1H), 7.26 (s, 1H);  $^{13}\text{C}$  NMR (150 MHz, CD<sub>3</sub>OD)  $\delta$  26.6, 27.4, 68.6, 75.7, 82.7, 96.1, 96.6, 100.8, 103.0, 106.8, 108.0, 115.3, 116.1, 116.8, 116.9, 119.9, 112.5, 128.2–130.0, 133.4, 134.9, 139.26, 139.28, 139.3, 139.6, 139.6, 146.6, 146.9, 150.2, 151.3, 152.9, 153.0, 153.7,

153.7, 156.3, 157.1, 157.1, 159.4, 160.9; IR (neat) 3412(br), 2310, 1610, 1508, 1447, 1420, 1274, 1182, 1149, 1116, 1052, 1039, 819, 754 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>65</sub>H<sub>20</sub>D<sub>35</sub>O<sub>12</sub> [(M+H)<sup>+</sup>] *m/z* 1062.5884, found *m/z* 1062.5845.

**21:** *R*<sub>f</sub> = 0.25 (EtOAc/CH<sub>2</sub>Cl<sub>2</sub> = 3/7); [α]<sub>D</sub><sup>20</sup> = -34 (*c* 1.4, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD) δ 2.55 (dd, *J* = 16.2, 7.2 Hz, 1H), 2.80 (dd, *J* = 16.2, 4.8 Hz, 1H), 3.86 (d, *J* = 3.6 Hz, 1H), 4.00 (brs, 1H), 4.50 (d, *J* = 3.0 Hz, 1H), 4.59 (d, *J* = 7.20 Hz, 1H), 6.06 (s, 1H), 6.35 (d, *J* = 2.4 Hz, 1H), 6.37 (d, *J* = 2.4 Hz, 1H), 6.70 (dd, *J* = 8.4, 2.4 Hz, 1H), 6.76 (d, *J* = 8.4 Hz, 1H), 6.83 (d, *J* = 2.4 Hz, 1H), 6.98 (d, *J* = 8.4 Hz, 1H), 7.22 (dd, *J* = 8.4, 1.8 Hz, 1H), 7.26 (d, *J* = 1.8 Hz, 1H); <sup>13</sup>C NMR (150 MHz, CD<sub>3</sub>OD) δ 27.8, 29.1, 69.2, 72.4, 75.0, 83.5, 96.8, 97.7, 97.9, 100.7, 104.6, 107.4, 107.8, 116.0, 116.1, 116.8, 117.0, 120.6, 122.6, 128.9–130.3, 132.9, 134.4, 137.8, 139.0, 139.2, 139.3, 139.3, 147.1, 147.1, 140.2, 151.4, 152.6, 153.6, 155.6, 156.6, 156.4, 157.7, 161.1; IR (neat) 3402 (br), 2310, 1618, 1596, 1488, 1404, 1273, 1184, 1120, 1052, 1039, 819, 747 cm<sup>-1</sup>; HRMS (ESI) calcd C<sub>65</sub>H<sub>20</sub>D<sub>35</sub>O<sub>12</sub> [(M+H)<sup>+</sup>] *m/z* 1062.5884, found *m/z* 1062.5852.

**22:** *R*<sub>f</sub> = 0.38 (EtOAc/CH<sub>2</sub>Cl<sub>2</sub> = 3/7); [α]<sub>D</sub><sup>20</sup> = -46 (*c* 1.1, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD) δ 2.59 (dd, *J* = 16.8, 7.8 Hz, 1H), 2.74 (dd, *J* = 16.8, 4.8 Hz, 1H), 3.85 (d, *J* = 3.6 Hz, 1H), 3.93 (brs, 1H), 4.66 (d, *J* = 7.2 Hz, 1H), 4.71 (d, *J* = 3.6 Hz, 1H), 6.05 (s, 1H), 6.35 (d, *J* = 2.4 Hz, 1H), 6.36 (d, *J* = 2.4 Hz, 1H), 6.66 (dd, *J* = 8.4, 1.8 Hz, 1H), 6.73 (d, *J* = 7.8 Hz, 1H), 6.77 (d, *J* = 1.8 Hz, 1H), 6.99 (d, *J* = 8.4 Hz, 1H), 7.28 (dd, *J* = 8.4, 1.8 Hz, 1H), 7.31 (d, *J* = 1.8 Hz, 1H); <sup>13</sup>C NMR (150 MHz, CD<sub>3</sub>OD) δ 27.5, 28.6, 69.1, 72.2, 72.3, 75.2, 83.6, 96.8, 97.6, 98.9, 100.6, 102.3, 107.6, 108.0, 115.8, 116.1, 116.9, 117.0, 120.6, 122.7, 128.9–130.2, 132.9, 134.6, 138.4, 139.1, 139.2, 139.3, 139.4, 147.1, 150.3, 151.4, 152.1, 154.8, 155.7, 156.2, 158.5, 161.0; IR (neat) 3419 (br), 2315, 1601, 1508, 1419, 1328, 1270, 1184, 1152, 1038, 819, 542 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>65</sub>H<sub>20</sub>D<sub>35</sub>O<sub>12</sub> [(M+H)<sup>+</sup>] *m/z* 1062.5884, found *m/z* 1062.5876.

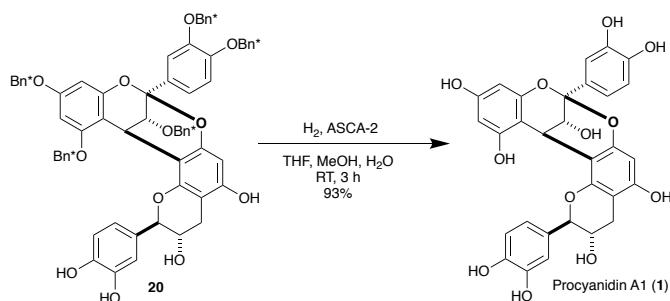
#### General procedure for hydrogenolysis of annulation products **20**, **21** and **22**

To a solution of dimers (1.0 equiv.) in the presence of ASCA-2 (0.75 equiv.) in a mixture of THF, MeOH and H<sub>2</sub>O (v/v/v = 2/2/1, 0.005 M) was stirred under H<sub>2</sub> atmosphere at room temperature. After stirring for 3.0 h, the mixture was carefully filtered through a glass fiber filter (MeOH) under argon atmosphere and the filtrate was evaporated to remove organic solvents and lyophilized to give a crude material, which was further purified by preparative HPLC [Mightysil RP-18GP II, (20 mm f × 250 mm), *l* = 280 nm, 25 °C, MeOH, H<sub>2</sub>O = 35/65 containing 0.1% TFA,

flow rate 10 mL/min, 20 x 250 mm] and lyophilization to give products a white amorphous compounds.

### *Procyanidin A1 (1)*

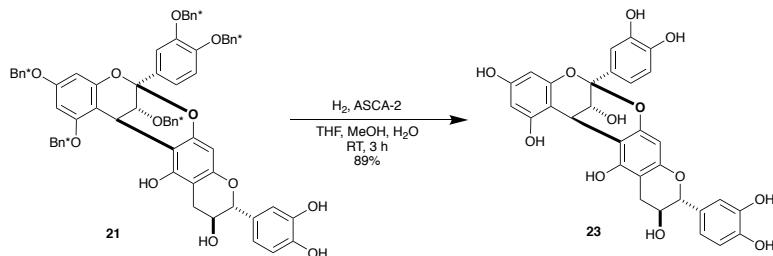
Starting from dimer **20** (20 mg, 0.018 mmol) in the presence of ASCA-2 (41 mg, 0.015 mmol), procyanidin A<sub>1</sub> (**1**) (10.1 mg, 93%) was obtained as a white amorphous foam.



**1:**  $[\alpha]_D^{20} = +78$  (*c* 0.57, acetone) [lit.<sup>3</sup>  $[\alpha]_D^{26} = +62.9$  (*c* 1.4, acetone)]; <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD) δ 2.60 (dd, *J* = 16.8, 8.4 Hz, 1H), 2.97 (dd, *J* = 16.8, 6.6 Hz, 1H), 4.10 (d, *J* = 3.6 Hz, 1H), 4.16–4.19 (m, 1H), 4.26 (d, *J* = 3.6 Hz, 1H), 4.76 (d, *J* = 7.8 Hz, 1H), 5.98 (d, *J* = 2.4 Hz, 1H), 6.09 (d, *J* = 2.4 Hz, 1H), 6.11 (s, 1H), 6.83–6.84 (m, 3H), 6.94 (s, 1H), 7.04 (dd, *J* = 8.4, 2.4 Hz, 1H), 7.16 (d, *J* = 2.4 Hz, 1H); <sup>13</sup>C NMR (150 MHz, CD<sub>3</sub>OD) δ 29.9, 30.1, 68.7, 67.0, 85.4, 97.4, 97.4, 99.0, 101.2, 104.0, 104.9, 107.7, 116.5, 116.6, 116.6, 117.2, 120.7, 121.5, 131.4, 133.2, 146.5, 147.2, 147.6, 147.7, 152.3, 153.1, 155.1, 157.0, 157.6, 159.0; IR (ATR) 3412 (br), 2232, 1610, 1508, 1508, 1447, 1420, 1327, 1274, 1182, 1149, 1116, 1052, 1039, 960, 819, 754, 543 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>30</sub>H<sub>25</sub>O<sub>12</sub> ([M+H]<sup>+</sup>) *m/z* 577.1340, found *m/z* 577.1318.

### *Doubly-linked dimeric proanthocyanidin 23*

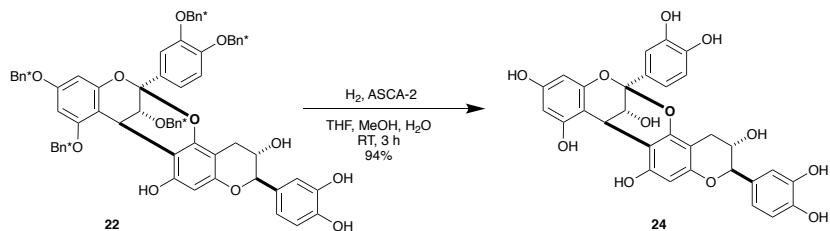
Starting from regiosomeric annulation product **21** (24 mg, 0.023 mmol) in the presence of ASCA-2 (50 mg, 0.018 mmol), **23** (11.7 mg, 89%) was obtained as a white amorphous foam.



**23:**  $[\alpha]_D^{20} = +9$  ( $c$  0.1, acetone), [lit.<sup>1</sup>  $[\alpha]_D^{20} +10.128$  ( $c$  1.05, acetone)];  $^1\text{H}$  NMR (600 MHz, CD<sub>3</sub>OD)  $\delta$  2.65 (dd,  $J = 16.2, 7.2$  Hz, 1H), 2.84 (dd,  $J = 16.2, 5.4$  Hz, 1H), 4.05–4.09 (m, 1H), 4.14 (d,  $J = 2.4$  Hz, 1H), 4.31 (d,  $J = 2.4$  Hz, 1H), 4.70 (d,  $J = 6.6$  Hz, 1H), 6.05 (s, 1H), 6.06 (s, 1H), 6.11 (s, 1H), 6.71 (dd,  $J = 7.8, 1.8$  Hz, 1H), 6.78 (d,  $J = 8.4$  Hz, 1H), 6.83 (d,  $J = 1.8$  Hz, 1H), 6.85 (d,  $J = 8.4$  Hz, 1H), 7.08 (dd,  $J = 8.4, 1.8$  Hz, 1H), 7.19 (d,  $J = 1.8$  Hz, 1H);  $^{13}\text{C}$  NMR (150 MHz, CD<sub>3</sub>OD)  $\delta$  28.7, 30.6, 68.5, 69.3, 83.4, 97.5, 97.8, 101.4, 104.4, 105.1, 109.6, 115.8, 116.5, 116.7, 117.0, 120.4, 120.8, 133.0, 133.1, 146.5, 147.0, 147.1, 152.9, 153.4, 155.3, 156.0, 156.2, 159.1; IR (ATR) 3280 (br), 1606, 1519, 1441, 1342, 1280, 1170, 1103, 1081, 1003, 970, 867, 818, 780, 626 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>30</sub>H<sub>23</sub>O<sub>12</sub> ([M–H]<sup>-</sup>) *m/z* 575.1195, found *m/z* 575.1173.

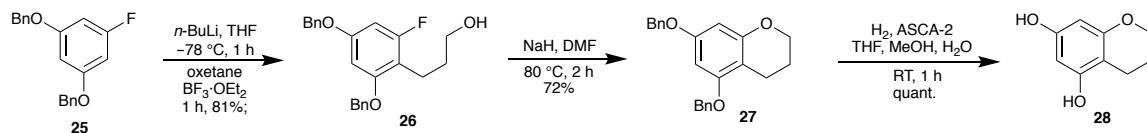
#### Doubly-linked dimeric proanthocyanidin **24**

Starting from regioisomeric annulation product **22** (28 mg, 0.027 mmol) in the presence of ASCA-2 (58 mg, 0.021 mmol), **24** (14.5 mg, 94%) was obtained as a white amorphous foam.



**24:**  $[\alpha]_D^{20} = +33$  ( $c$  0.15, acetone);  $^1\text{H}$  NMR (600 MHz, CD<sub>3</sub>OD)  $\delta$  2.68 (dd,  $J = 16.2, 7.2$  Hz, 1H), 2.80 (dd,  $J = 16.2, 5.4$  Hz, 1H), 4.03–4.06 (m, 1H), 4.13 (d,  $J = 3.0$  Hz, 1H), 4.31 (d,  $J = 3.0$  Hz, 1H), 4.73 (d,  $J = 6.6$  Hz, 1H), 6.06 (s, 1H), 6.11 (s, 1H), 6.12 (s, 1H), 6.70 (d,  $J = 8.4$  Hz, 1H), 6.76 (d,  $J = 8.4$  Hz, 1H), 6.78 (s, 1H), 6.86 (d,  $J = 8.4$  Hz, 1H), 7.12 (d,  $J = 8.4$  Hz, 1H), 7.22 (s, 1H);  $^{13}\text{C}$  NMR (150 MHz, CD<sub>3</sub>OD)  $\delta$  28.3, 30.3, 68.5, 69.1, 83.6, 97.5, 98.3, 98.6, 101.4, 102.5, 105.2, 109.4, 115.6, 116.5, 116.7, 117.0, 120.5, 120.9, 133.0, 133.1, 146.5, 147.0, 147.07, 147.70, 152.2, 153.4, 155.0, 156.1, 156.9, 159.1; IR (ATR) 3299 (br), 1608, 1517, 1441, 1283, 1177, 1120, 1082, 1008, 971, 872, 818, 781 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>30</sub>H<sub>23</sub>O<sub>12</sub> ([M–H]<sup>-</sup>) *m/z* 575.1195, found *m/z* 575.1163.

**Synthesis of model substrate 28**



To a solution of fluorobenzene **25** (1.0 g, 3.2 mmol) in THF (8 mL) was added *n*-BuLi (2.2 mL, 1.58 M hexane solution, 3.5 mmol) at  $-78\text{ }^{\circ}\text{C}$ . After stirring for 1 h, a solution of oxetane (0.25 mL, 3.84 mmol) and  $\text{BF}_3\text{-OEt}_2$  (0.48 mL, 3.88 mmol) in THF (10 mL) were successively added at  $-78\text{ }^{\circ}\text{C}$ . The stirring was continued for 30 min and the reaction was stopped by adding saturated aqueous  $\text{NH}_4\text{Cl}$  solution. The mixture was extracted with EtOAc (x3), and the combined extracts were washed with brine, dried ( $\text{Na}_2\text{SO}_4$ ) and concentrated in vacuo. The residue was purified by flash column chromatography (hexane/EtOAc = 5:1) to afford alcohol **26** (961 mg, 81%) as a white solid.

**26:**  $R_f = 0.51$  (EtOAc/hexane = 2/3); mp 73–75 °C (hexane/EtOAc),  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  1.68 (brs, 1H), 1.77–1.82 (m, 2H), 2.73 (t,  $J = 6.6$  Hz, 2H), 3.56 (t,  $J = 5.4$  Hz, 2H), 5.00 (s, 2H), 5.03 (s, 2H), 7.33–7.36 (m, 2H), 7.38–7.42 (m, 8H);  $^{13}\text{C}$  NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  18.3, 18.3, 32.4, 61.9, 70.5, 70.8, 94.6, 94.8, 96.5, 96.6, 110.1, 110.3, 127.5, 127.7, 128.3, 128.4, 128.8, 128.9, 136.5, 136.6, 158.3, 158.4, 158.4, 158.5, 161.5, 163.1; IR (neat) 3358 (br), 2925, 2862, 1498, 1586, 1452, 1374, 1357, 1164, 1101, 1064, 1036, 1003, 984, 818, 802, 692  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_{23}\text{H}_{24}\text{FO}_3$  [(M+H) $^+$ ]  $m/z$  367.1704, found  $m/z$  367.1592.

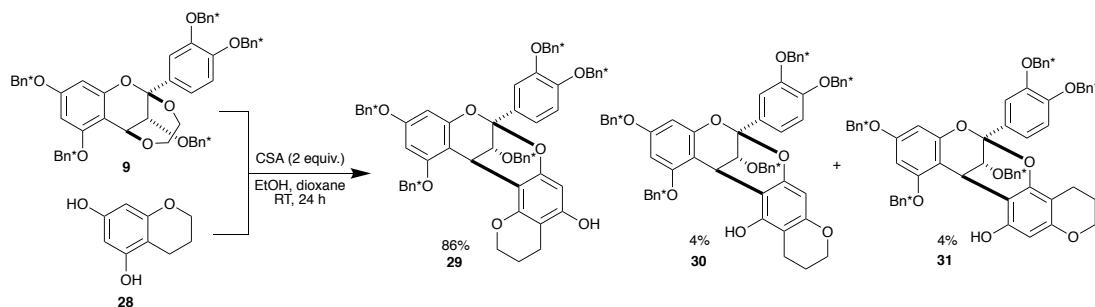
To a suspension of NaH (63% dispersion in mineral oil, 75 mg, 2.0 mmol) in DMF (13 mL) was added alcohol **26** (480 mg, 1.31 mmol) in DMF (13 mL) at 0 °C. After stirring at 80 °C for 2 h, the reaction was quenched with slow addition of water at 0 °C and extracted with EtOAc (x3). The combined extracts were washed with brine, dried ( $\text{Na}_2\text{SO}_4$ ) and concentrated in vacuo. The residue was purified by flash column chromatography (hexane/EtOAc = 3:1) to afford pyran **27** (326 mg, 72%) as off-white solid.

**27:**  $R_f = 0.76$  (EtOAc/hexane = 2/3); mp 84–85 °C (hexane/EtOAc),  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  1.95–1.99 (m, 2H), 2.67 (t,  $J = 6.6$  Hz, 2H), 4.15 (t,  $J = 5.4$  Hz, 2H), 5.00 (s, 2H), 5.02 (s, 2H), 6.14 (d,  $J = 2.4$  Hz, 1H), 6.22 (d,  $J = 2.4$  Hz, 1H), 7.32–7.35 (m, 2H), 7.38–7.44 (m, 8H);  $^{13}\text{C}$  NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  19.2, 22.1, 70.0, 70.3, 93.2, 94.9, 104.6, 127.3, 127.7, 127.9, 128.1, 128.7, 128.7, 137.2, 137.4, 156.3, 157.9, 158.5; IR (neat) 3064, 2954, 2883, 1613, 1594, 1583, 1498, 1468, 1436, 1376, 1228, 1127, 1065, 1035, 910, 808, 765, 744  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_{23}\text{H}_{23}\text{O}_3$  [(M+H) $^+$ ]  $m/z$  347.1641, found  $m/z$  347.1636.

To a solution of pyran **27** (321 mg, 0.926 mmol) in the presence of ASCA-2 (624 mg, 0.223 mmol) in a mixture of THF, MeOH and H<sub>2</sub>O (v/v/v = 2/2/1, 50 mL) was stirred under H<sub>2</sub> atmosphere at room temperature. After stirring for 1 h, the mixture was carefully filtered through a glass fiber filter (MeOH) under argon atmosphere. The filtrate was evaporated, and the residue was purified by recrystallization (EtOAc) to give phenol **28** (152 mg, quant.) as ivory solid.

**28:**  $R_f = 0.18$  (EtOAc/hexane = 2/3); mp 178–179 °C (lit.<sup>4</sup> 176 °C), <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD) δ 1.89–1.93 (m, 2H), 2.54 (t,  $J = 6.2$  Hz, 2H), 4.09 (t,  $J = 5.4$  Hz, 2H), 5.78 (d,  $J = 2.4$  Hz, 1H), 5.90 (d,  $J = 2.4$  Hz, 1H); <sup>13</sup>C NMR (150 MHz, CD<sub>3</sub>OD) δ 20.7, 24.2, 68.1, 96.7, 96.7, 103.6, 158.1, 158.1, 158.2, 158.2, 158.38, 158.40; IR (ATR) 3352, 3217, 1618, 1518, 1472, 1442, 1385, 1275, 1189, 1046, 1011, 948, 875, 802 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>9</sub>H<sub>11</sub>O<sub>3</sub> [(M+H)<sup>+</sup>] *m/z* 167.0702, found *m/z* 167.0707.

#### Annulation of **9** with model substrate **28**



To a solution of acetal **9** (50 mg, 0.059 mmol) and **28** (15 mg, 0.090 mmol) in mixture of ethanol and 1,4-dioxane (v/v=1, 1.25 mL) was added CSA (28 mg, 0.12 mmol) at room temperature. After stirring for 24 h at the same temperature, the reaction mixture was diluted with EtOAc and washed with saturated aqueous NaHCO<sub>3</sub> solution, brine, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated in vacuo. The residue was purified by PTLC (hexane/EtOAc = 3/2) to afford dimer **29** (48.2 mg, 86%) as a white amorphous foam along with mixture of **30** and **31** which was separated by PTLC (hexane/CH<sub>2</sub>Cl<sub>2</sub>/EtOAc = 65/30/5, three runs) giving **30** (2.3 mg, 4%) and **31** (2.0 mg, 4%) as a gummy solid.

**29:**  $R_f = 0.65$  (EtOAc/hexane = 1/1);  $[\alpha]_D^{20} = -36$  (*c* 0.84, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 1.77–1.83 (m, 2H), 2.47–2.54 (m, 2H), 3.65–3.74 (m, 2H), 3.85 (d,  $J = 3.0$  Hz, 1H), 4.71 (s, 1H), 4.98 (d,  $J = 3.0$  Hz, 1H), 5.97 (s, 1H), 6.25 (s, 1H), 6.35 (s, 1H), 6.96 (d,  $J = 8.4$  Hz, 1H), 7.03 (d,  $J = 8.4$  Hz, 1H), 7.24 (s, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 19.0, 21.6, 21.7, 25.1, 65.9,

73.2, 94.7, 94.9, 98.6, 103.0, 105.6, 106.6, 114.4, 114.5, 120.6, 125.5–128.4, 129.2, 132.6, 136.8, 137.2, 137.2, 137.5, 137.5, 138.1, 148.5, 149.3, 151.0, 152.8, 153.2, 153.9, 157.6, 158.9; IR (neat) 3419 (br), 2938, 2276, 1615, 1490, 1453, 1490, 1441, 1419, 1327, 1273, 1201, 1123, 1098, 1051, 960, 909, 818, 730, 542 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>59</sub>H<sub>16</sub>D<sub>35</sub>O<sub>9</sub> [(M+H)<sup>+</sup>] *m/z* 938.5724, found *m/z* 938.5742.

**30:** *R*<sub>f</sub> = 0.68 (EtOAc/hexane = 1/1); [α]<sub>D</sub><sup>20</sup> = -24 (*c* 0.53, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 1.89–1.91 (m, 2H), 2.51–2.61 (m, 2H), 3.82 (d, *J* = 3.6 Hz, 1H), 4.03–4.09 (m, 2H), 4.37 (d, *J* = 3.6 Hz, 1H), 6.06 (s, 1H), 6.33 (s, 1H), 6.42 (s, 1H), 6.97 (d, *J* = 9.0 Hz, 1H), 7.03 (s, 1H), 7.24–7.26 (m, 2H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 19.3, 22.1, 25.9, 66.6, 72.4, 95.2, 96.1, 96.6, 98.6, 104.9, 105.2, 105.3, 114.4, 114.5, 120.7, 127.1–128.7, 132.1, 135.1, 136.5, 137.0, 137.1, 137.1, 148.6, 149.5, 150.1, 151.3, 153.6, 155.2, 155.4, 159.1; IR (neat) 3418 (br), 3010, 2355, 1619, 1597, 1507, 1490, 1450, 1273, 1201, 1120, 1083, 1052, 1038, 959, 909, 819, 731, 541 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>59</sub>H<sub>16</sub>D<sub>35</sub>O<sub>9</sub> [(M+H)<sup>+</sup>] *m/z* 938.5724 found *m/z* 938.5700.

**31:** *R*<sub>f</sub> = 0.68 (EtOAc/hexane = 1/1); [α]<sub>D</sub><sup>20</sup> = -59 (*c* 0.74, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 1.85–1.86 (m, 2H), 2.43–2.54 (m, 2H), 3.81 (d, *J* = 3.6 Hz, 1H), 4.00–4.08 (m, 2H), 4.39 (d, *J* = 3.6 Hz, 1H), 6.04 (s, 1H), 6.34 (s, 1H), 6.43 (s, 1H), 6.80 (s, 1H), 6.99 (d, *J* = 8.4 Hz, 1H), 7.26 (s, 1H), 7.28 (d, *J* = 8.4 Hz, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 19.1, 22.0, 25.6, 66.5, 72.3, 95.1, 96.0, 98.4, 98.5, 103.3, 105.2, 105.5, 114.3, 114.7, 120.9, 126.9, 127.1–129.2, 132.2, 135.0, 136.5, 137.1, 137.09, 137.13, 148.5, 149.4, 149.5, 151.8, 153.5, 155.1, 155.5, 159.1; IR (neat) 3424 (br), 2936, 2228, 1618, 1597, 1508, 1491, 1419, 1328, 1274, 1194, 1153, 1130, 1052, 976, 909, 731, 543 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>59</sub>H<sub>16</sub>D<sub>35</sub>O<sub>9</sub> [(M+H)<sup>+</sup>] *m/z* 938.5724, found *m/z* 938.5755.

#### References:

1. H. Lou, Y. Yamazaki, T. Sasaki, M. Uchida, H. Tanaka and S. Oka, *Phytochemistry*, 1999, **51**, 297.
2. S. Morimoto, G.-I. Nonaka and I. Nishioka, *Chem. Pharm. Bull.*, 1987, **35**, 4717.
3. G.-I. Nonaka, S. Morimoto, J.-I. Kinjo, T. Nohara and I. Nishioka, *Chem. Pharm. Bull.*, 1987, **35**, 149.
4. S. Belapure, Z. G. Beamer, J. E. Bartmess and Shawn R. Campagna, *Tetrahedron*, 2011, **67**, 9265.

## Computational studies

Initial conformer search for **32** and **33** were carried out with Spartan'18 program (Wavefunction, Inc. Irvine, CA) using the MMFF molecular mechanics model, respectively. Structures of each conformers, keeping only those that are within 15 kJ/mol of the lowest energy conformer, were optimized by the DFT method at the  $\omega$ B97X-D/6-31G(d) level of theory. Further accurate calculations were carried out with Gaussian16 program.<sup>5</sup> The structures of **32**, **33** were optimized by the DFT method at the  $\omega$ B97X-D/6-311G(d, p) level of theory. The frequency analyses gave thermodynamic parameters with/without an imaginary wavenumber. The calculations were carried out by the IEFPCM (ethanol) method.

## Reference

5. M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. V. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. J. Bearpark, J. J. Heyd, E. N. Brothers, K. N. Kudin, V. N. Staroverov, T. A. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. P. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman, and D. J. Fox, Gaussian 16, Revision B.01, Gaussian, Inc., Wallingford CT, 2016.

## Cartesian coordinates of optimized **32**

C	-2.093630	-0.070093	-0.213986	H	2.803562	-0.354996	1.464336
C	-1.061662	-1.145845	-0.217478	H	4.023538	0.174724	0.305344
C	-1.775494	1.243065	-0.070772	C	2.094437	1.141666	0.072290
H	-2.540952	2.009530	-0.041596	H	2.172950	1.908215	0.844699
C	0.339818	-0.626166	-0.225827	H	2.428892	1.597734	-0.864618
C	0.661554	0.692973	-0.051514	C	-0.411657	1.608130	0.026673
O	-3.320390	-0.535352	-0.333385	O	-0.069240	2.865457	0.184747
H	-3.976103	0.170258	-0.296848	H	-0.836190	3.447323	0.236489
O	1.222930	-1.584971	-0.374738	C	-1.268974	-2.081169	1.000080
C	2.624487	-1.224744	-0.482336	H	-2.273105	-2.500974	0.971225
H	2.807893	-0.984876	-1.532502	H	-1.137978	-1.524696	1.929944
H	3.159748	-2.134166	-0.221668	H	-0.540356	-2.889048	0.960434
C	2.965142	-0.061895	0.423137	H	-1.202408	-1.735651	-1.129820

## Summary of the frequency analysis for **32**

Calculation Type = FREQ  
Calculation Method = RwB97XD  
Basis Set = 6-311G(d,p)  
Charge = 1  
Spin = Singlet  
Solvation = scrf=(iefpcm,solvent=ethanol)

E(RwB97XD) = -614.36585 Hartree  
 RMS Gradient Norm = 1.4978e-05 Hartree/Bohr  
 Imaginary Freq = 0  
 Dipole Moment = 4.6155345 Debye  
 Polarizability (?) = 151.22033 a.u.  
 Point Group = C1  
 Job cpu time: 0 days 1 hours 26 minutes 5.3 seconds.  
 Thermo Tab Data Section:  
 Imaginary Freq = 0  
 Temperature = 298.15 Kelvin  
 Pressure = 1 atm  
 Frequencies scaled by = 1  
 Electronic Energy (EE) = -614.36585 Hartree  
 Zero-point Energy Correction = 0.221483 Hartree  
 Thermal Correction to Energy = 0.232946 Hartree  
 Thermal Correction to Enthalpy = 0.23389 Hartree  
 Thermal Correction to Free Energy = 0.18442 Hartree  
 EE + Zero-point Energy = -614.14437 Hartree  
 EE + Thermal Energy Correction = -614.13291 Hartree  
 EE + Thermal Enthalpy Correction = -614.13196 Hartree  
 EE + Thermal Free Energy Correction = -614.18143 Hartree  
 E (Thermal) = 146.176 kcal/mol  
 Heat Capacity (Cv) = 45.593 cal/mol-kelvin  
 Entropy (S) = 104.119 cal/mol-kelvin

Opt Tab Data Section:  
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 Maximum force = 4.1e-05 Converged  
 RMS force = 6e-06 Converged  
 Maximum displacement = 0.001944 Not converged  
 RMS displacement = 0.000456 Converged  
 Predicted energy change = -3.480319e-08 Hartree

### Cartesian coordinates of optimized 33

C 1.740369	-1.014863	-0.253705	H -3.091003	0.094471	-1.423158
C 1.866015	0.470482	-0.233087	H -4.074240	0.811351	-0.145033
C 0.550220	-1.643192	-0.098750	C -1.966202	1.295234	-0.015983
H 0.460480	-2.721039	-0.069472	H -1.956488	2.116404	-0.738583
C 0.550421	1.175405	-0.196366	H -2.126294	1.731416	0.975486
C -0.654288	0.549370	-0.051073	C -0.630067	-0.869408	0.033593
O 2.901634	-1.626538	-0.397016	C 2.745603	0.914571	0.961869
H 2.802702	-2.584879	-0.368530	H 3.720255	0.434754	0.891855
O -1.712033	-1.572593	0.219303	H 2.273422	0.631353	1.904138
C -2.985027	-0.913431	0.460854	H 2.875254	1.995065	0.936337
H -3.034291	-0.709617	1.532746	H 2.369971	0.774168	-1.158148
H -3.726536	-1.664192	0.201618	O 0.688675	2.484088	-0.297156
C -3.111133	0.346909	-0.359649	H -0.154902	2.944627	-0.226515

### Summary of the frequency analysis for 33

Calculation Type = FREQ  
 Calculation Method = RwB97XD

Basis Set = 6-311G(d,p)  
Charge = 1  
Spin = Singlet  
Solvation = scrf=(iefpcm,solvent=ethanol)  
E(RwB97XD) = -614.36427 Hartree  
RMS Gradient Norm = 9.608e-06 Hartree/Bohr  
Imaginary Freq = 0  
Dipole Moment = 1.7212947 Debye  
Polarizability (?) = 151.787 a.u.  
Point Group = C1  
Job cpu time: 0 days 1 hours 23 minutes 41.3 seconds.

Thermo Tab Data Section:

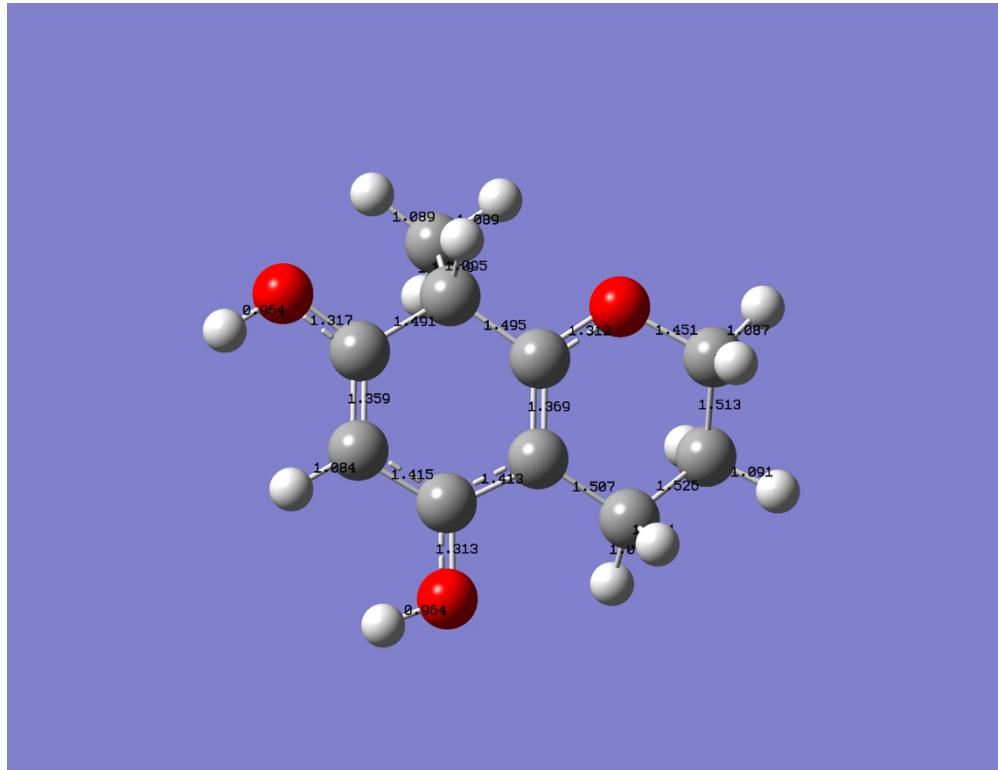
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Thermal Correction to Energy = 0.232853 Hartree  
Thermal Correction to Enthalpy = 0.233797 Hartree  
Thermal Correction to Free Energy = 0.18423 Hartree  
EE + Zero-point Energy = -614.14296 Hartree  
EE + Thermal Energy Correction = -614.13142 Hartree  
EE + Thermal Enthalpy Correction = -614.13048 Hartree  
EE + Thermal Free Energy Correction = -614.18004 Hartree  
E (Thermal) = 146.117 kcal/mol  
Heat Capacity (Cv) = 45.828 cal/mol-kelvin  
Entropy (S) = 104.323 cal/mol-kelvin

Opt Tab Data Section:

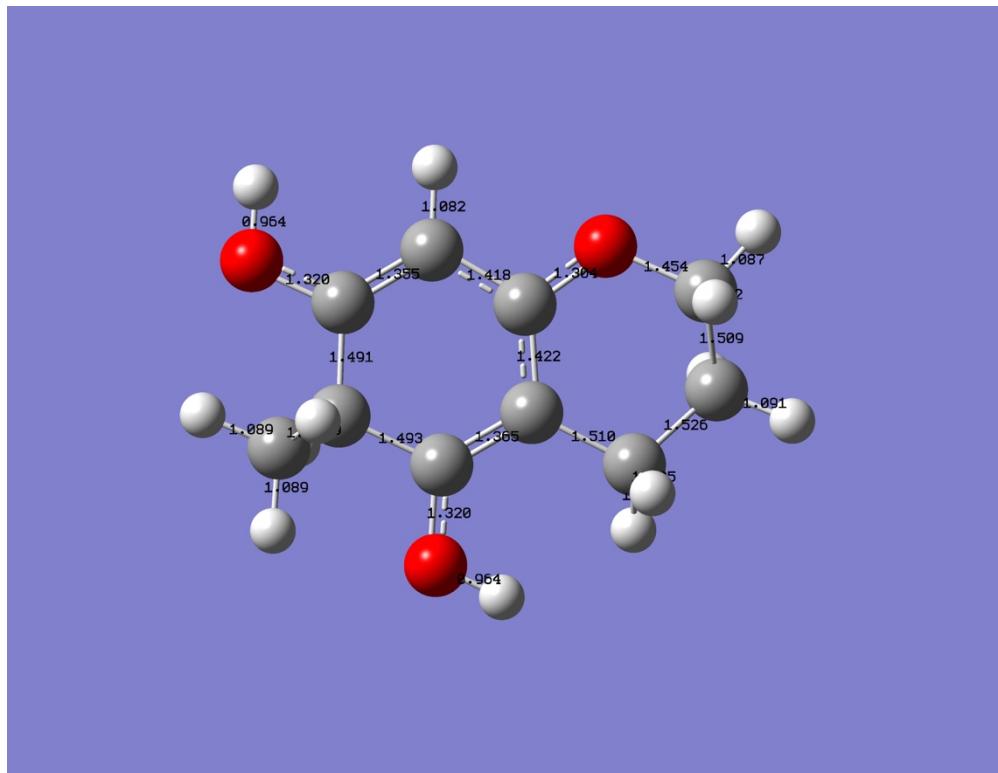
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Optimized structures of **32** and **33**

Intermediate **32**

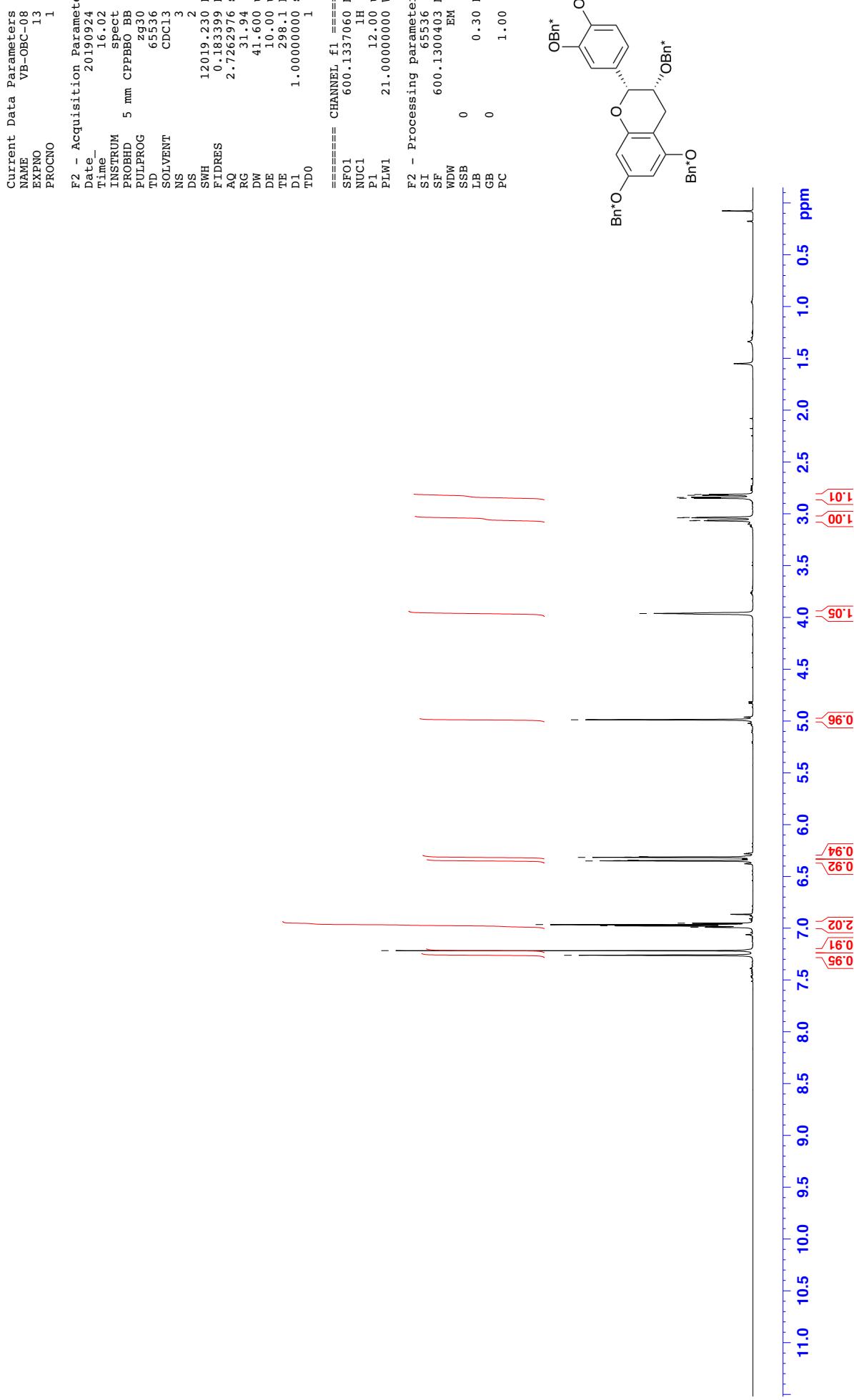


Intermediate **33**

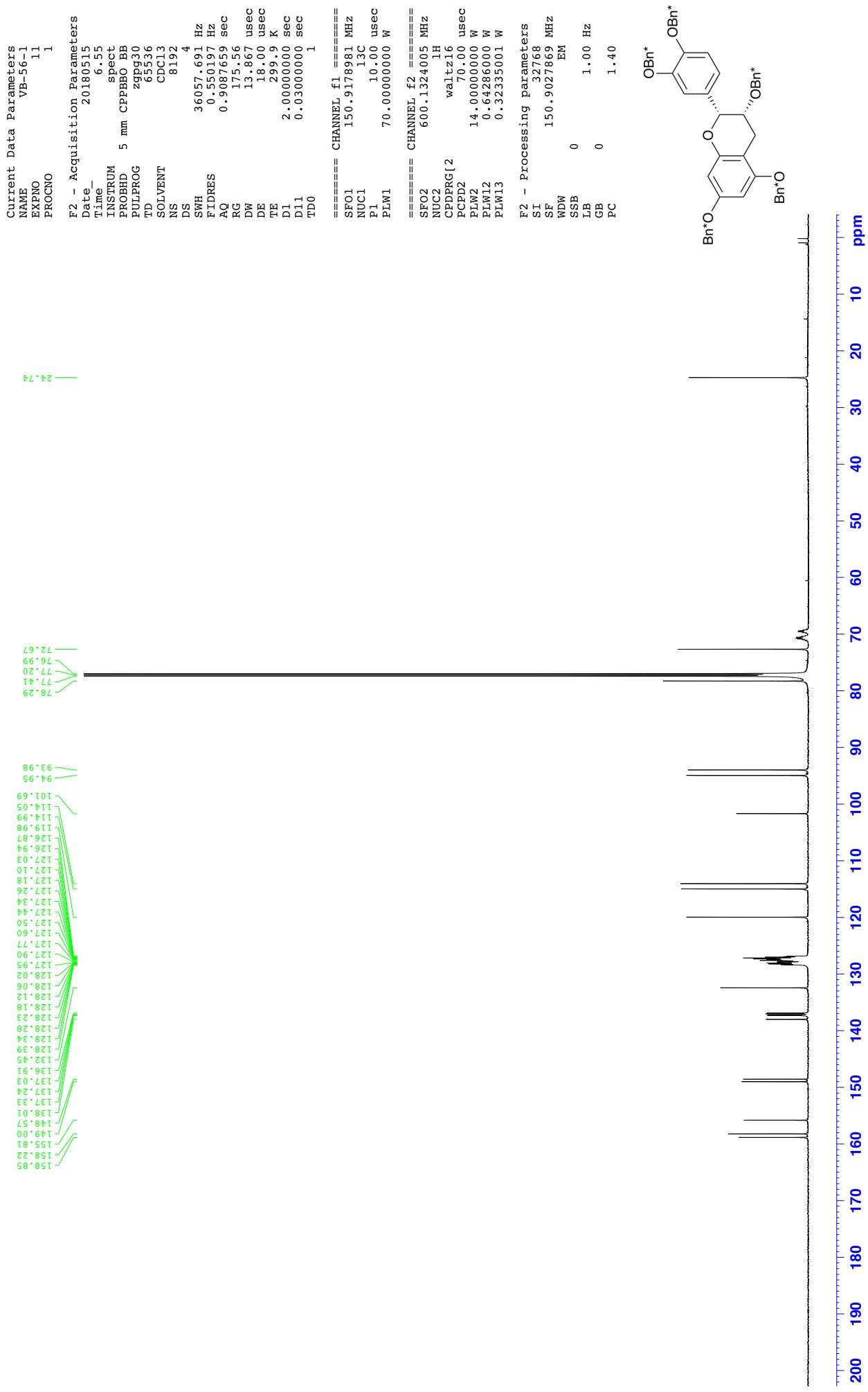




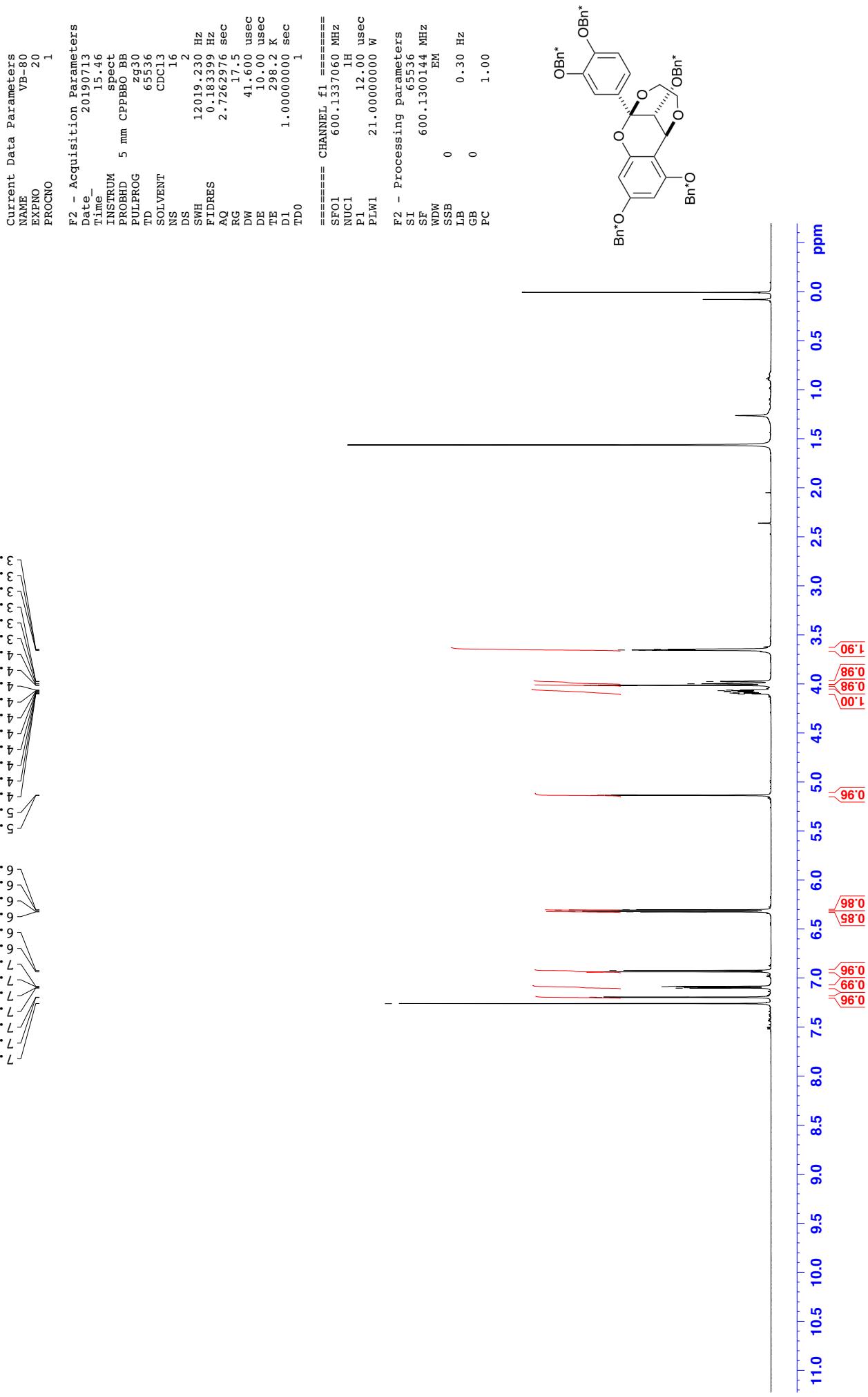
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<sup>13</sup>C NMR of 8 (600MHz, CDCl<sub>3</sub>)

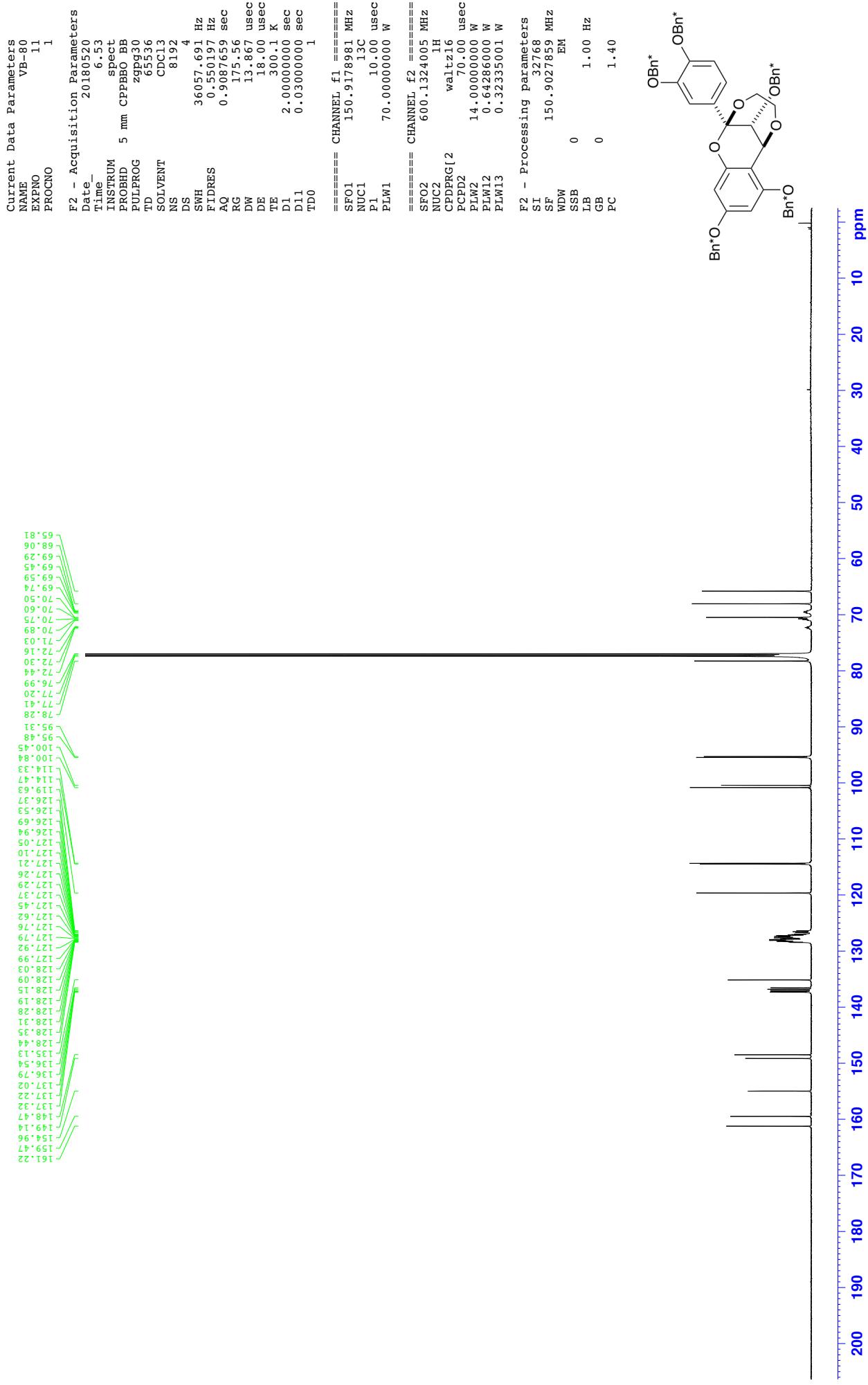


<sup>1</sup>H NMR of 9 (600MHz, CDCl<sub>3</sub>)

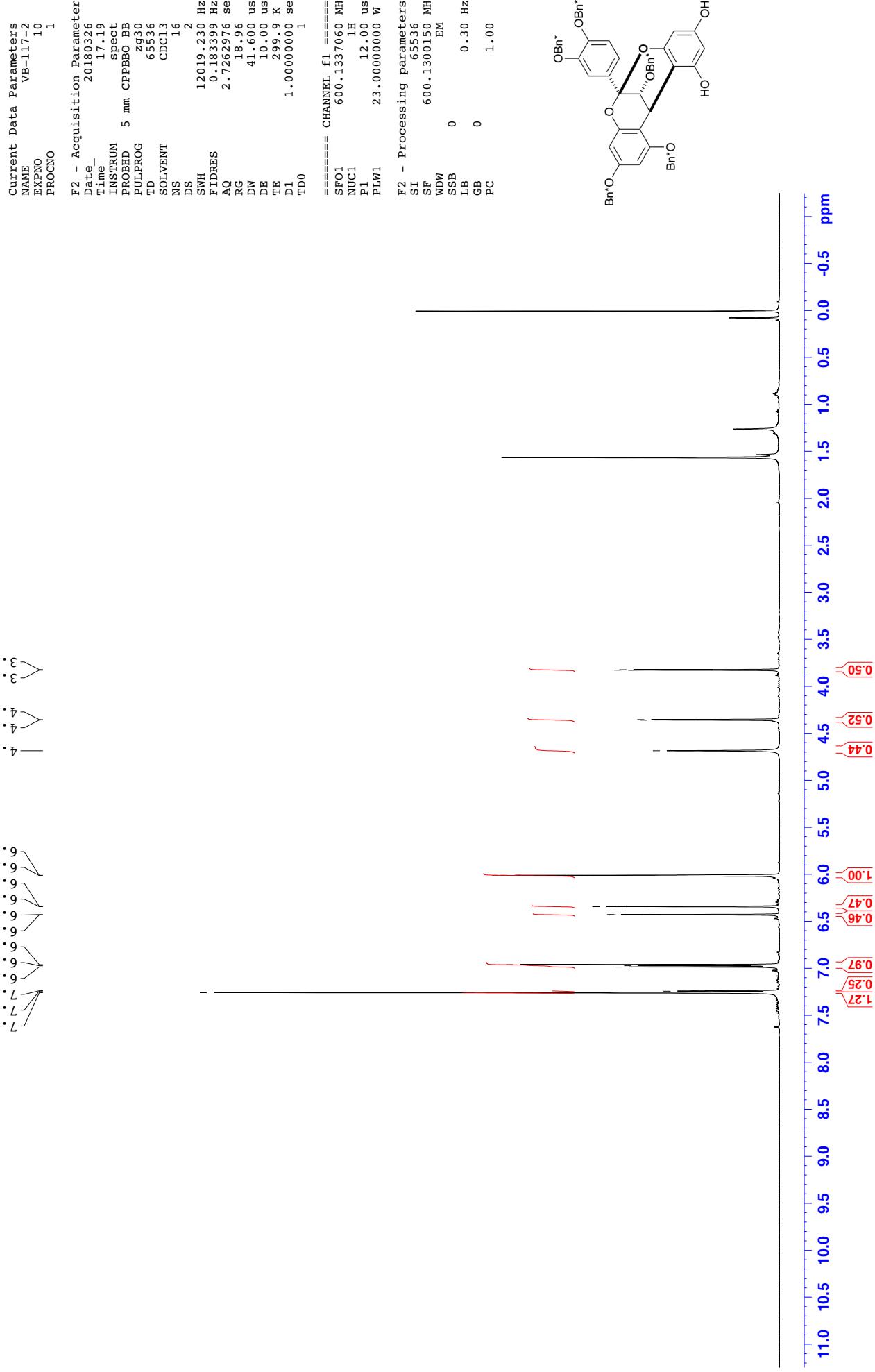




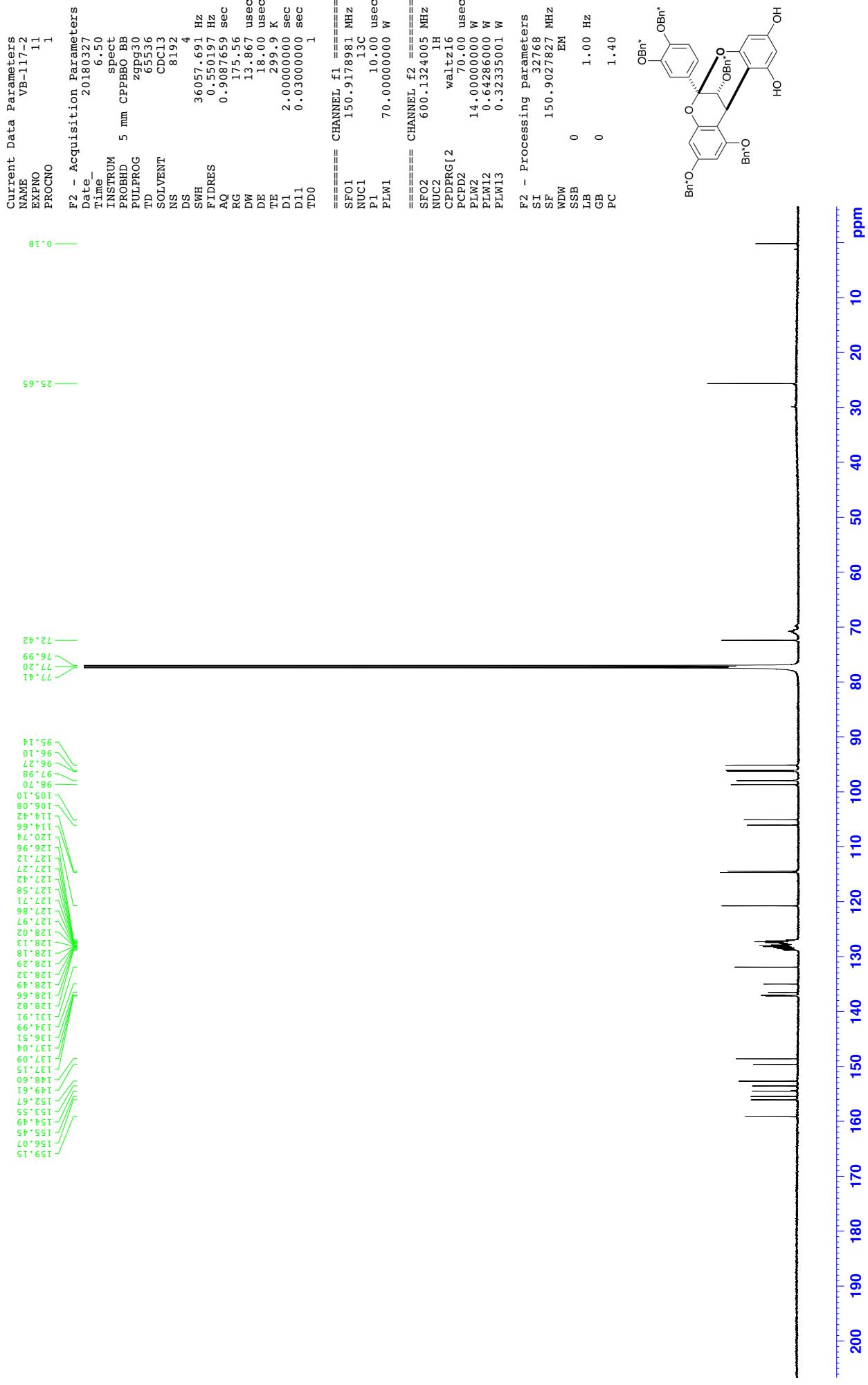
<sup>13</sup>C NMR of 9 (600MHz, CDCl<sub>3</sub>)



<sup>1</sup>H NMR of 11 (600MHz, CDCl<sub>3</sub>)



<sup>13</sup>C NMR of 11 (150MHz, CDCl<sub>3</sub>)





<sup>1</sup>H NMR of 12 (600MHz, CDCl<sub>3</sub>)

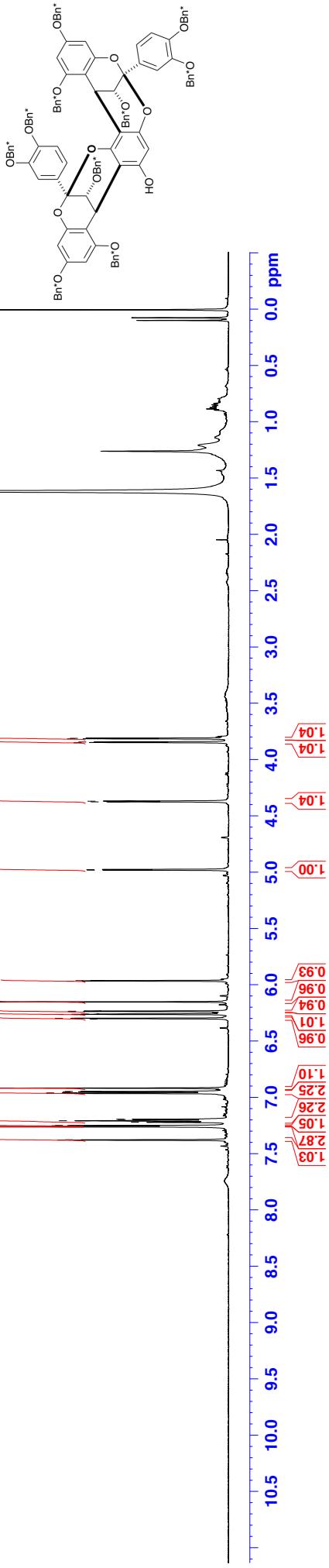
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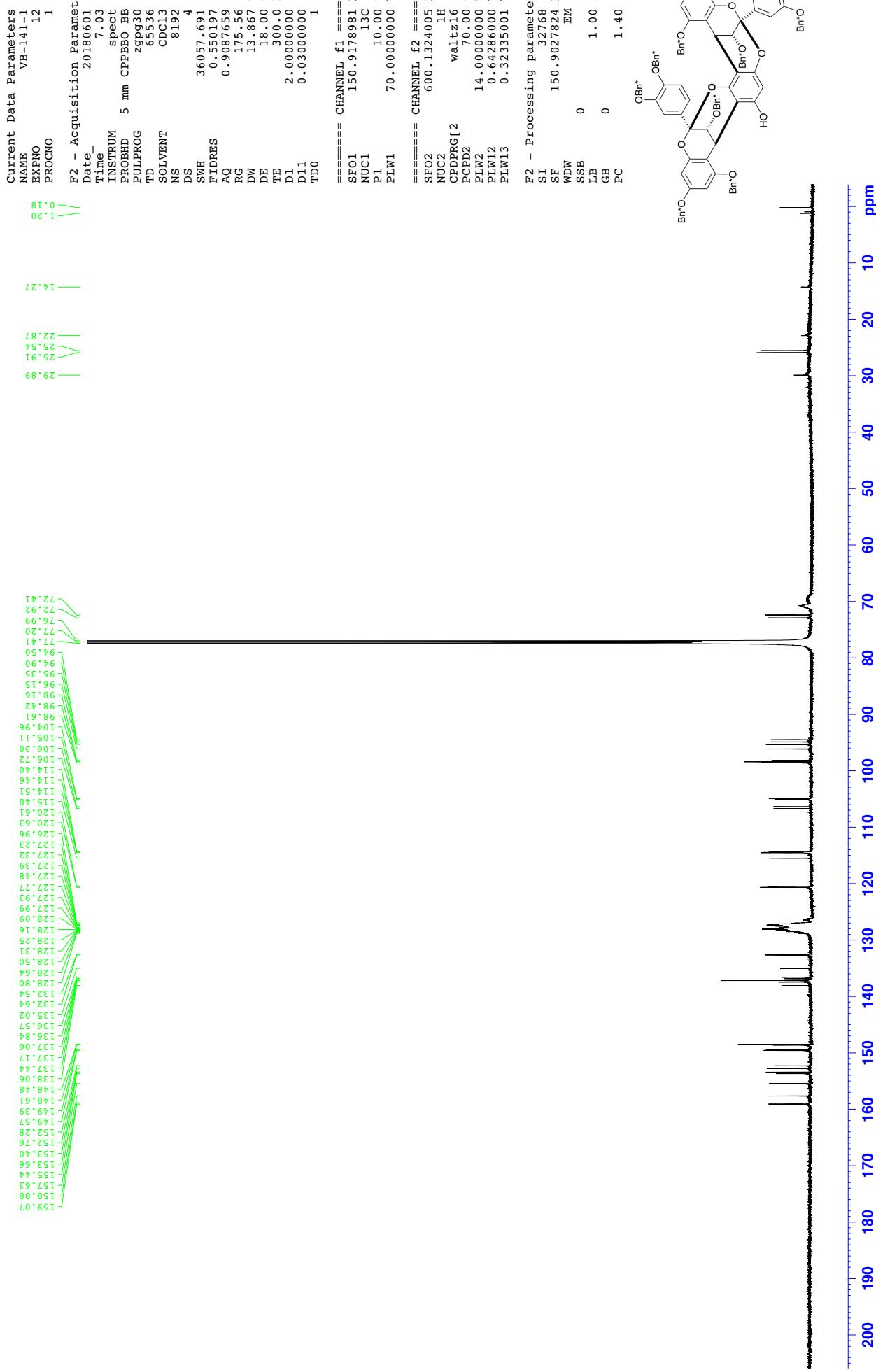
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DE 10.00 usec  
TE 300.0 K  
D1 1.0000000 sec  
TDO 1

===== CHANNEL f1 =====  
SFO1 600.1337060 MHz  
NUC1 1H  
P1 12.00 usec  
PLW1 23.00000000 W

F2 - Processing parameters  
SI 65536  
SF 600.1300146 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

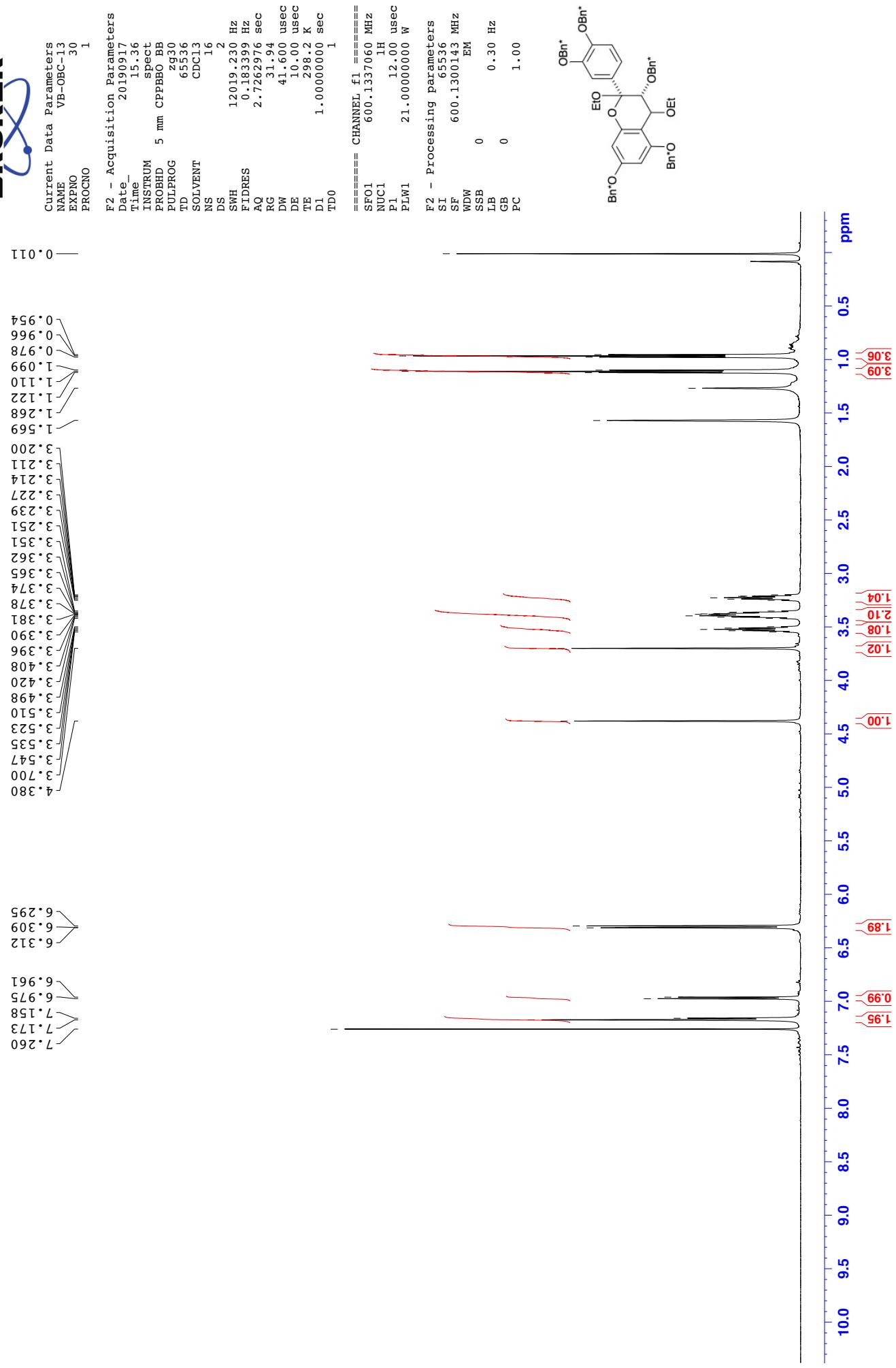


<sup>13</sup>C NMR of 12 (150MHz CDCl<sub>3</sub>)

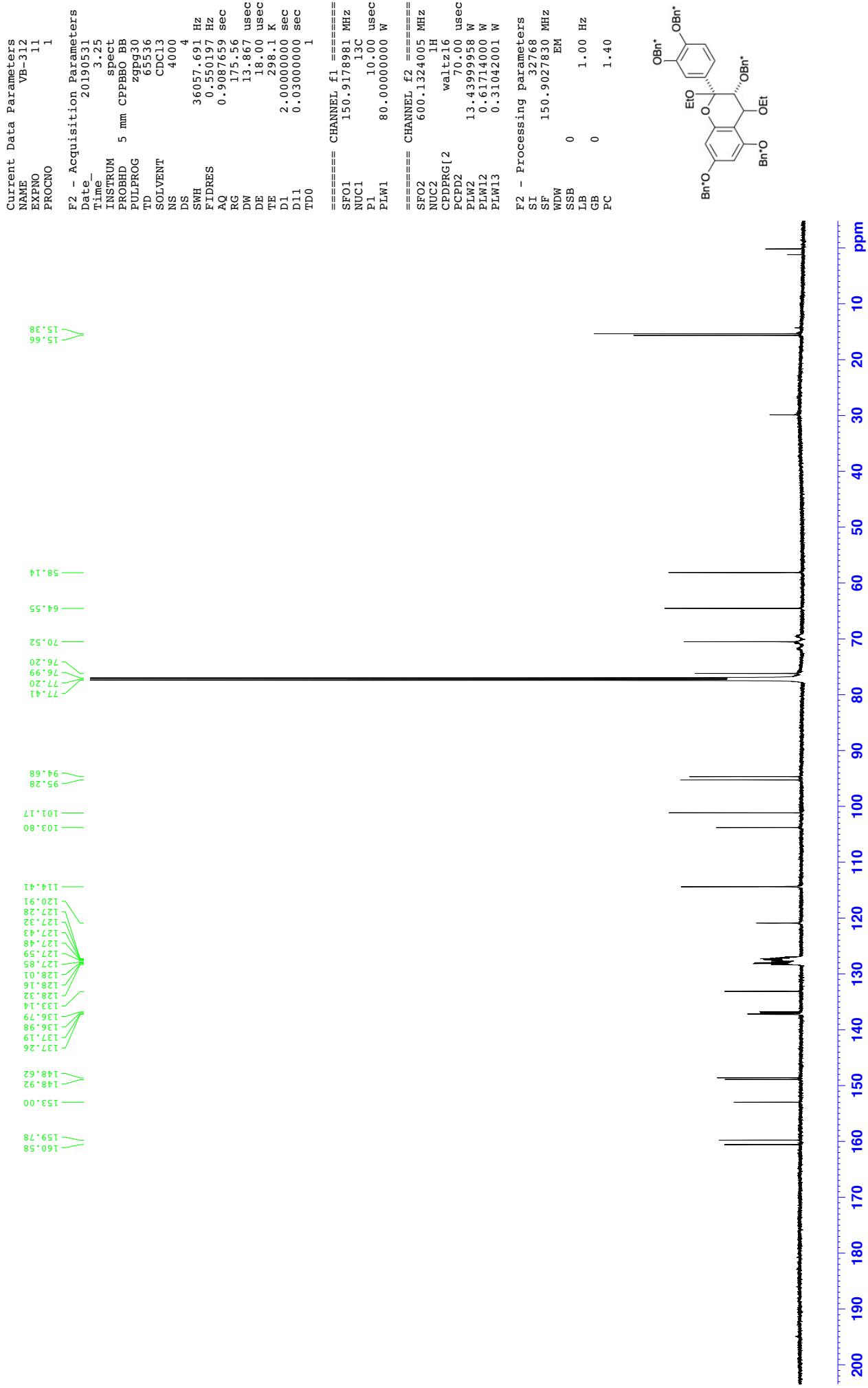




## 1H NMR of 13 (600MHz, CDCl3)



<sup>13</sup>C NMR of 13 (150MHz, CDCl<sub>3</sub>)





<sup>1</sup>H NMR of 14 (600MHz, CD<sub>3</sub>OD)

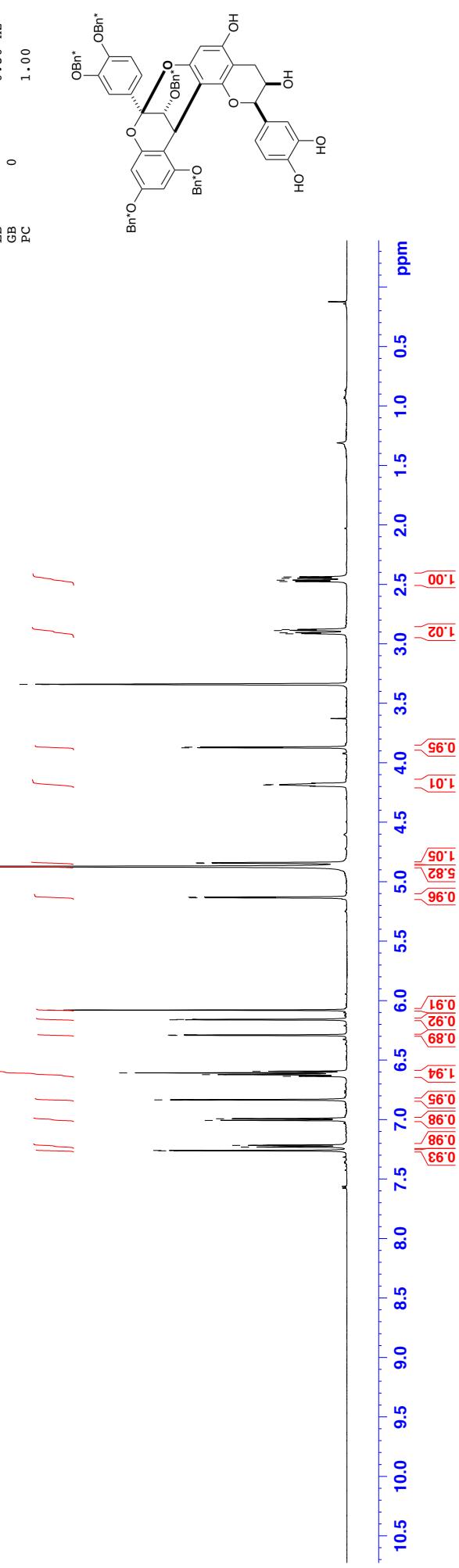
7.262  
7.231  
7.217  
7.007  
6.993  
6.935  
6.636  
6.608  
6.594  
6.289  
6.152  
6.122  
5.136  
4.845  
4.841  
4.186  
4.183  
3.874  
3.868  
3.342  
2.915  
2.888  
2.879  
2.476  
2.465  
2.449  
2.438

Current Data Parameters  
NAME VB-19-03  
EXPNO 10  
PROCNO 1

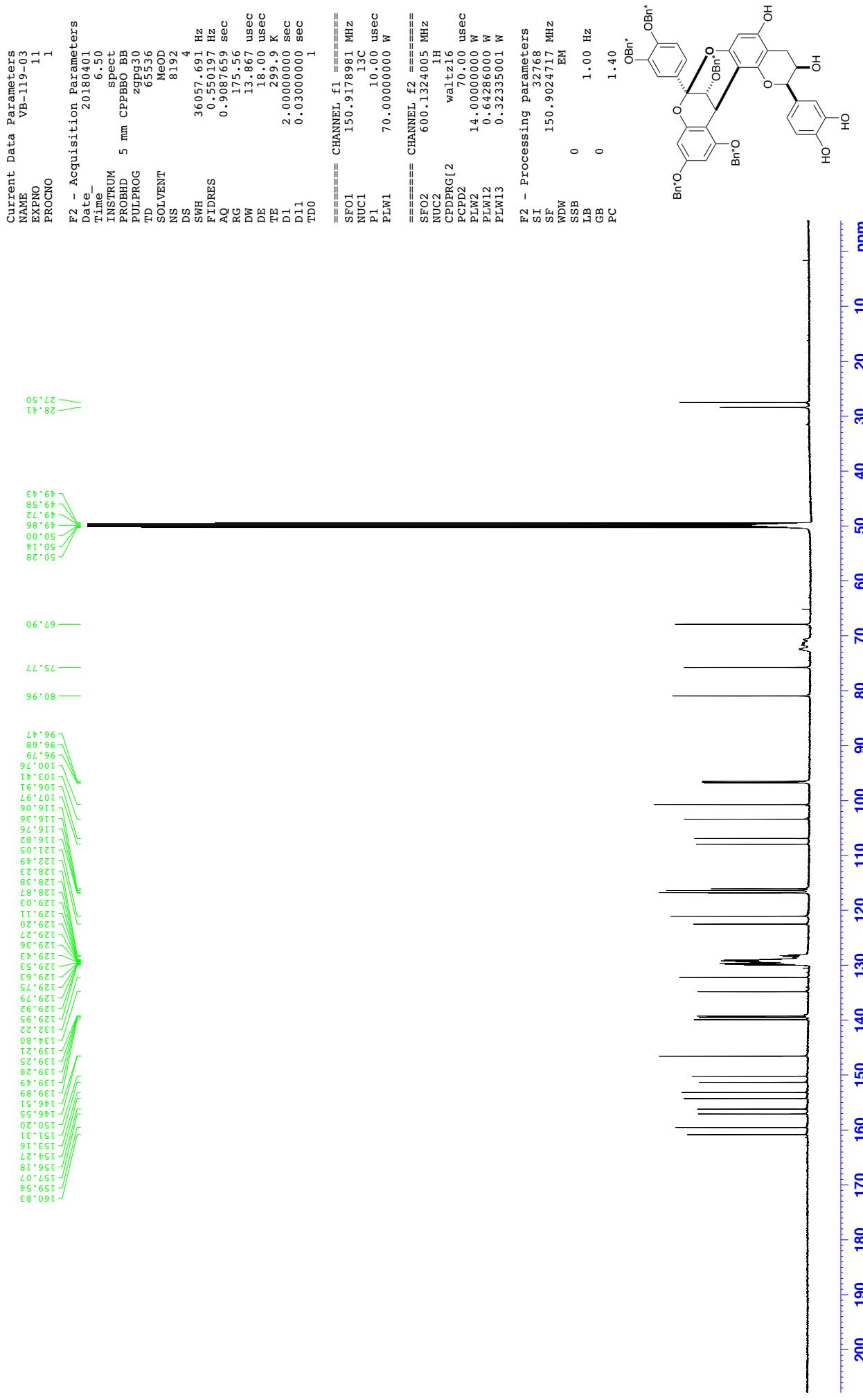
F2 - Acquisition Parameters  
Date 20180330  
Time 17.09  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 16  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.183399 Hz  
AQ 2.762976 sec  
RG 31.94  
DW 41.600 usec  
DE 10.00 usec  
TE 300.0 K  
D1 1.0000000 sec  
TDO 1

===== CHANNEL f1 =====  
SFO1 600.1337060 MHz  
NUC1 1H  
P1 12.00 usec  
PLW1 23.00000000 W

F2 - Processing parameters  
SI 65536  
SF 600.1299925 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



<sup>13</sup>C NMR of 14 (150MHz, CD<sub>3</sub>OD)





<sup>1</sup>H NMR of 15 (600MHz, 5%D<sub>2</sub>O in CDCl<sub>3</sub>)

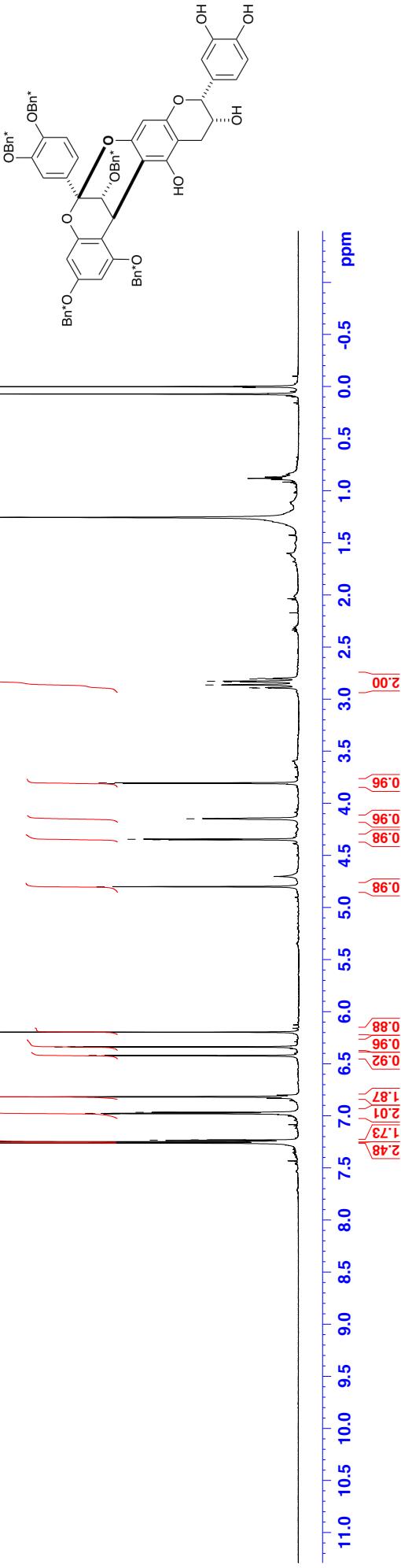
7.260  
7.247  
7.234  
7.230  
6.978  
6.965  
6.817  
6.420  
6.339  
6.336  
6.196  
4.801  
4.351  
4.148  
3.810  
3.804  
2.894  
2.865  
2.835  
2.807  
2.799

Current Data Parameters  
NAME VB-186-2D20  
EXPNO 20  
PROCNO 1

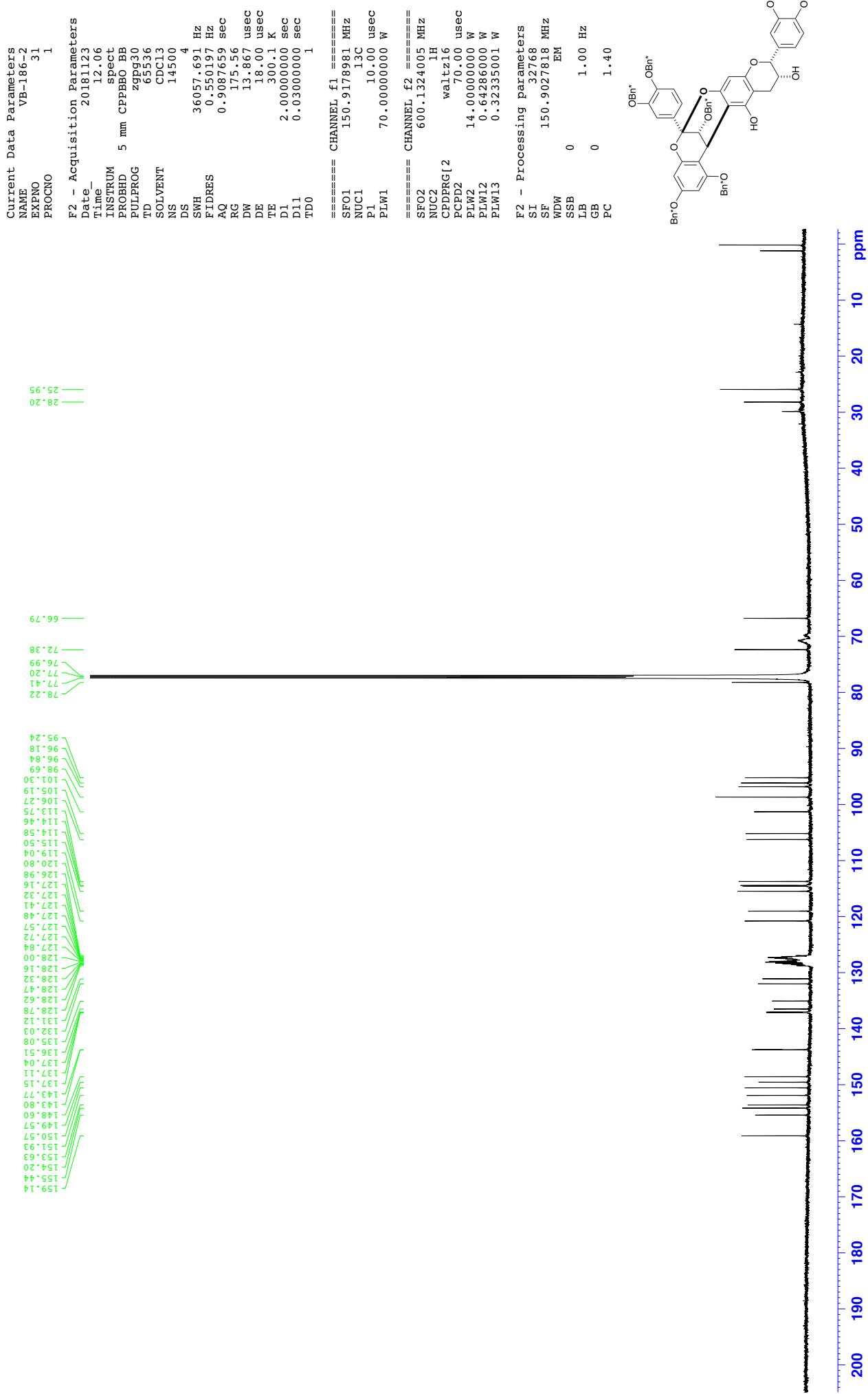
F2 - Acquisition Parameters  
Date 20181130  
Time 20.04  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zg30  
TD 65536  
SOLVENT CDCl<sub>3</sub>  
NS 50  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.1833399 Hz  
AQ 2.7562976 sec  
RG 31.94  
DW 41.600 usec  
DE 10.00 usec  
TE 300.1 K  
D1 1.0000000 sec  
TDO 1

===== CHANNEL f1 =====  
SFO1 600.1337060 MHz  
NUC1 1H  
P1 12.00 usec  
PLW1 23.00000000 W

F2 - Processing parameters  
SI 65536  
SF 600.1300144 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



<sup>13</sup>C NMR of 15 (150MHz, CDCl<sub>3</sub>)





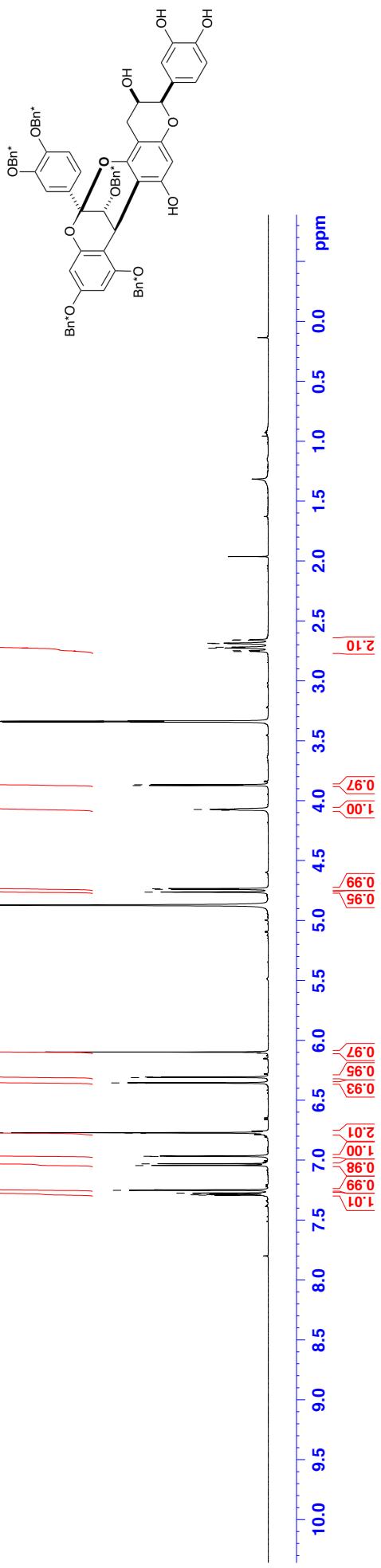
<sup>1</sup>H NMR of 16 (600MHz, CD<sub>3</sub>OD)

7.293  
7.289  
7.275  
7.279  
7.252  
7.248  
7.045  
7.030  
6.968  
6.966  
6.774  
6.771  
6.358  
6.354  
6.310  
6.306  
6.098  
  
4.872  
4.763  
4.740  
4.073  
4.068  
3.873  
3.867  
3.342  
3.340  
3.337  
2.755  
2.747  
2.727  
2.719  
2.689  
2.685  
2.661  
2.657

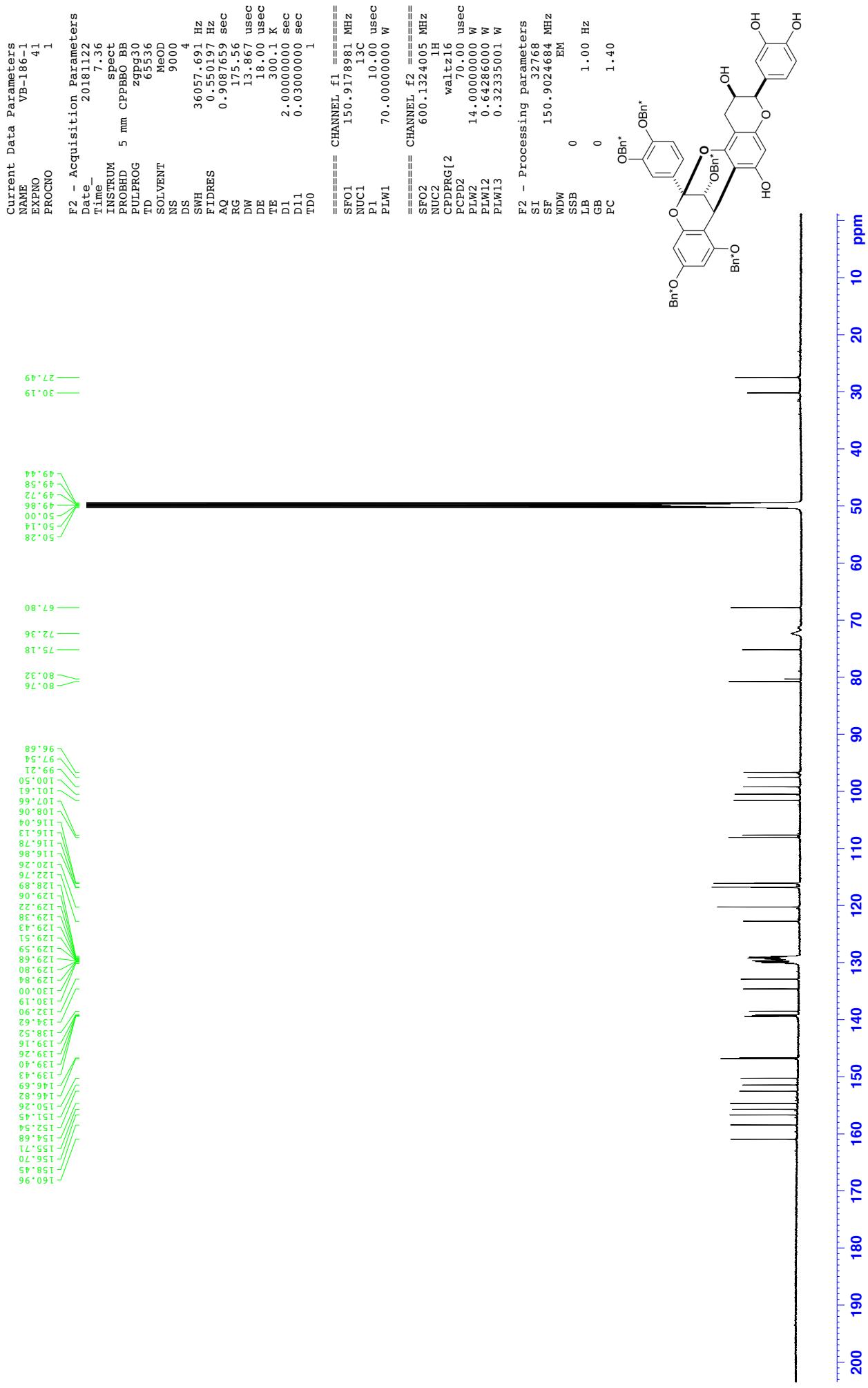
Current Data Parameters  
NAME VB-186-1  
EXPNO 40  
PROCNO 1

F2 - Acquisition Parameters  
Date 20181121  
Time 20.25  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 16  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.1833399 Hz  
AQ 2.7262976 sec  
RG 31.94  
DW 41.600 usec  
DE 10.00 usec  
TE 300.1 K  
D1 1.0000000 sec  
TDO 1

===== CHANNEL f1 ======  
SFO1 600.1337060 MHz  
NUC1 1H  
P1 12.00 usec  
PLW1 23.00000000 W  
  
F2 - Processing parameters  
SI 65536  
SF 600.1299934 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

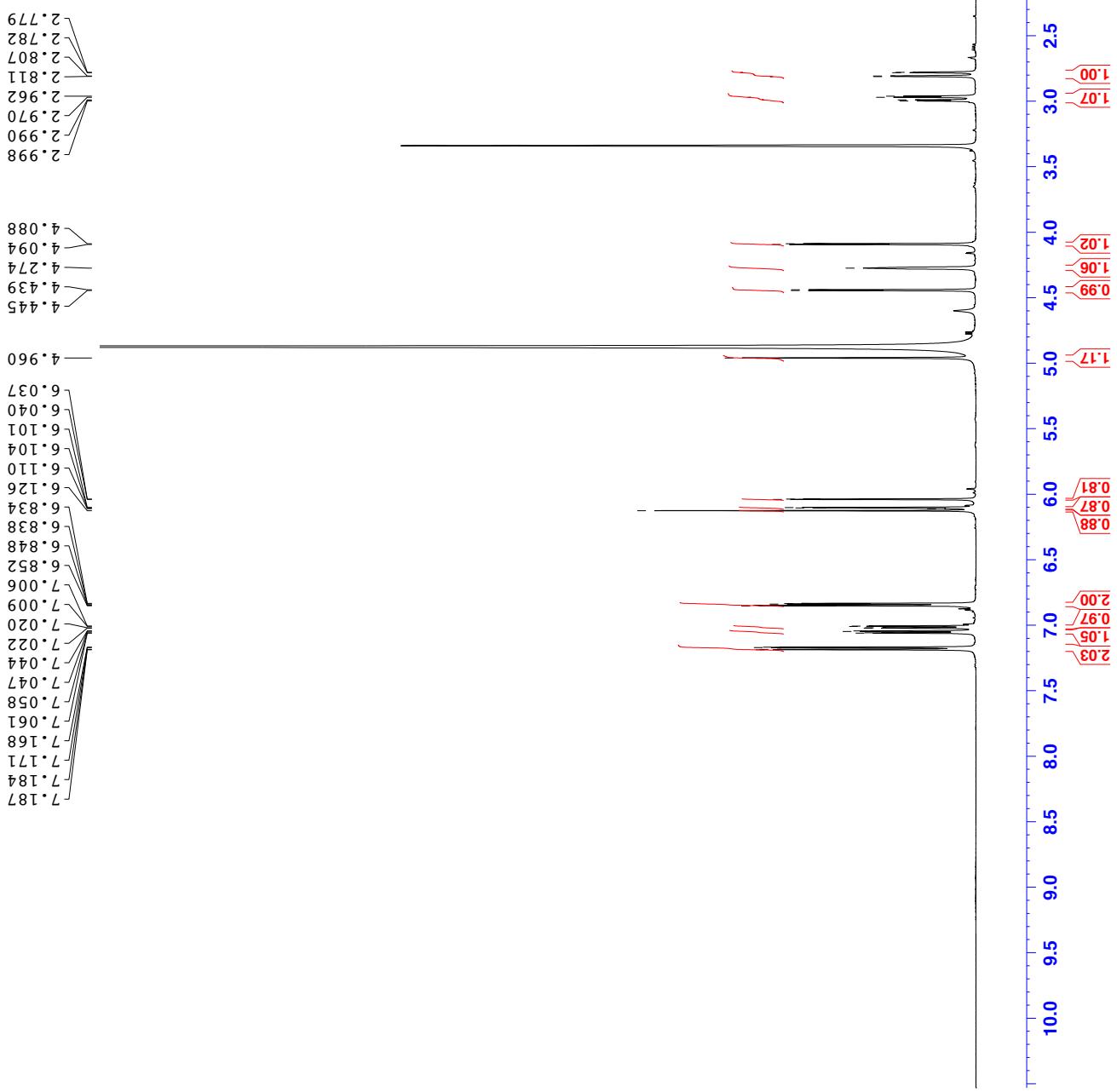


**<sup>13</sup>C NMR of 16 (150MHz, CD<sub>3</sub>OD)**





<sup>1</sup>H NMR of 2 (600MHz, CD<sub>3</sub>OD)



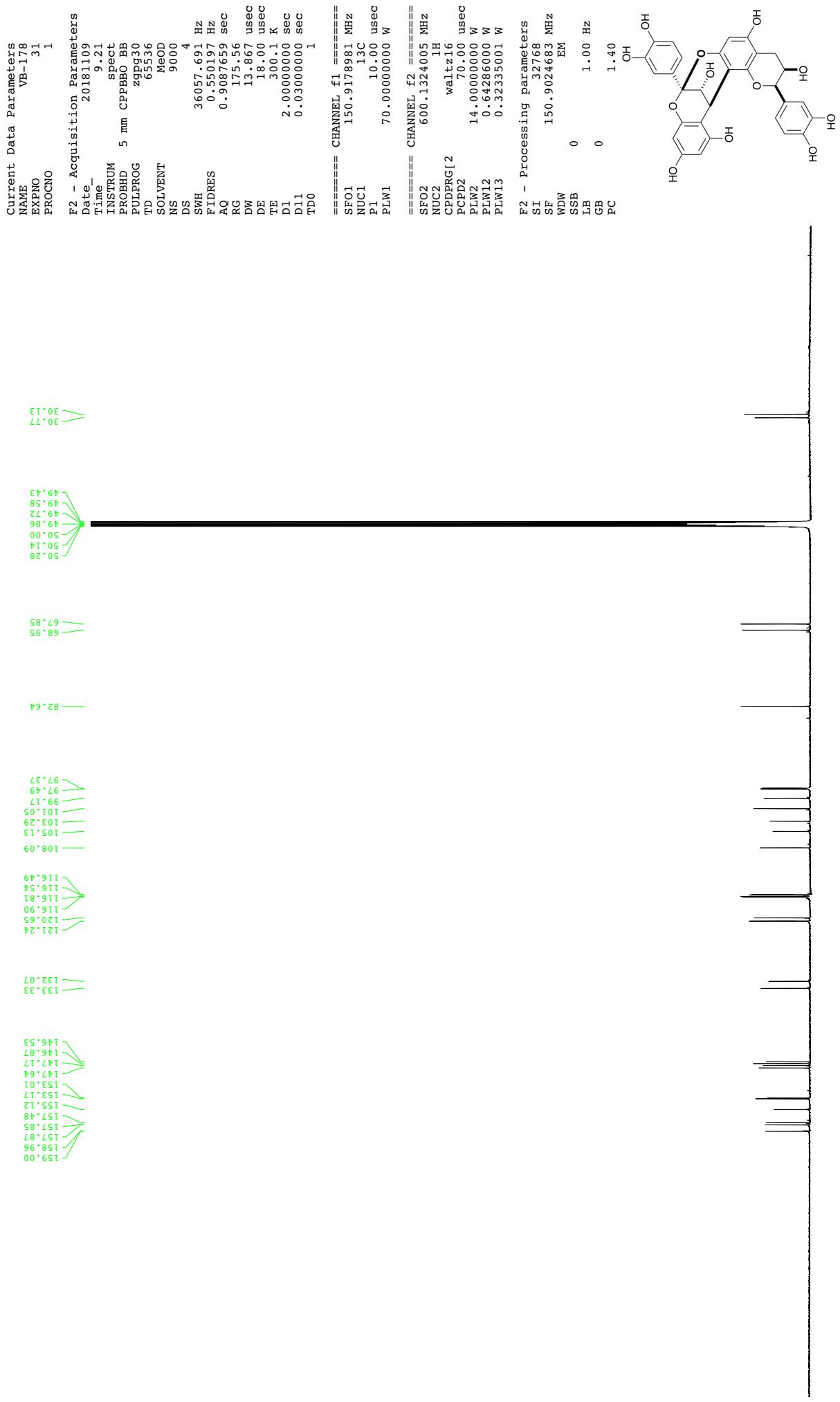
Current Data Parameters  
NAME VB-178  
EXPNO 30  
PROCNO 1

F2 - Acquisition Parameters  
Date 20181108  
Time 10.16  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 50  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.183399 Hz  
AQ 2.7562976 sec  
RG 31.94  
DW 41.600 usec  
DE 10.00 usec  
TE 300.1 K  
D1 1.0000000 sec  
TDO 1

===== CHANNEL f1 =====  
SFO1 600.1337060 MHz  
NUC1 1H  
P1 12.00 usec  
PLW1 23.00000000 W  
  
F2 - Processing parameters  
SI 65536  
SF 600.1299940 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



# <sup>1</sup>H NMR of 2 (150MHz, CD<sub>3</sub>OD)





<sup>1</sup>H NMR of 17 (600MHz, CD<sub>3</sub>OD)

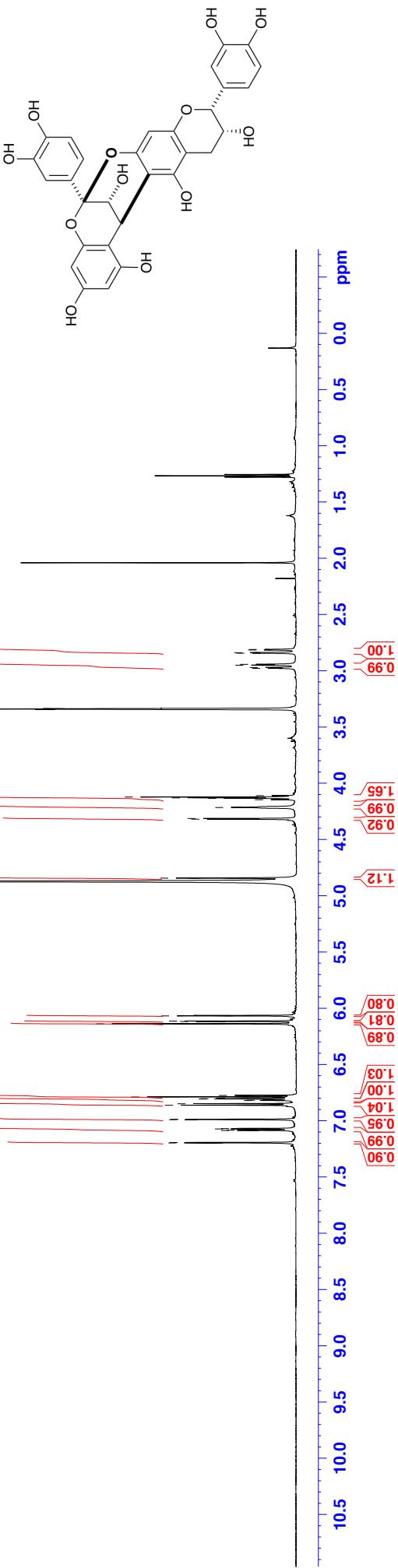
2.811  
2.816  
2.839  
2.844  
2.943  
2.950  
2.971  
2.978  
4.110  
4.123  
4.129  
4.134  
4.146  
4.215  
4.313  
4.319  
4.843  
6.064  
6.068  
6.114  
6.117  
6.138  
6.774  
6.788  
6.803  
6.806  
6.817  
6.820  
6.848  
6.862  
6.987  
6.990  
7.069  
7.072  
7.082  
7.193  
7.197

Current Data Parameters  
NAME VB-295  
EXPNO 11  
PROCNO 1

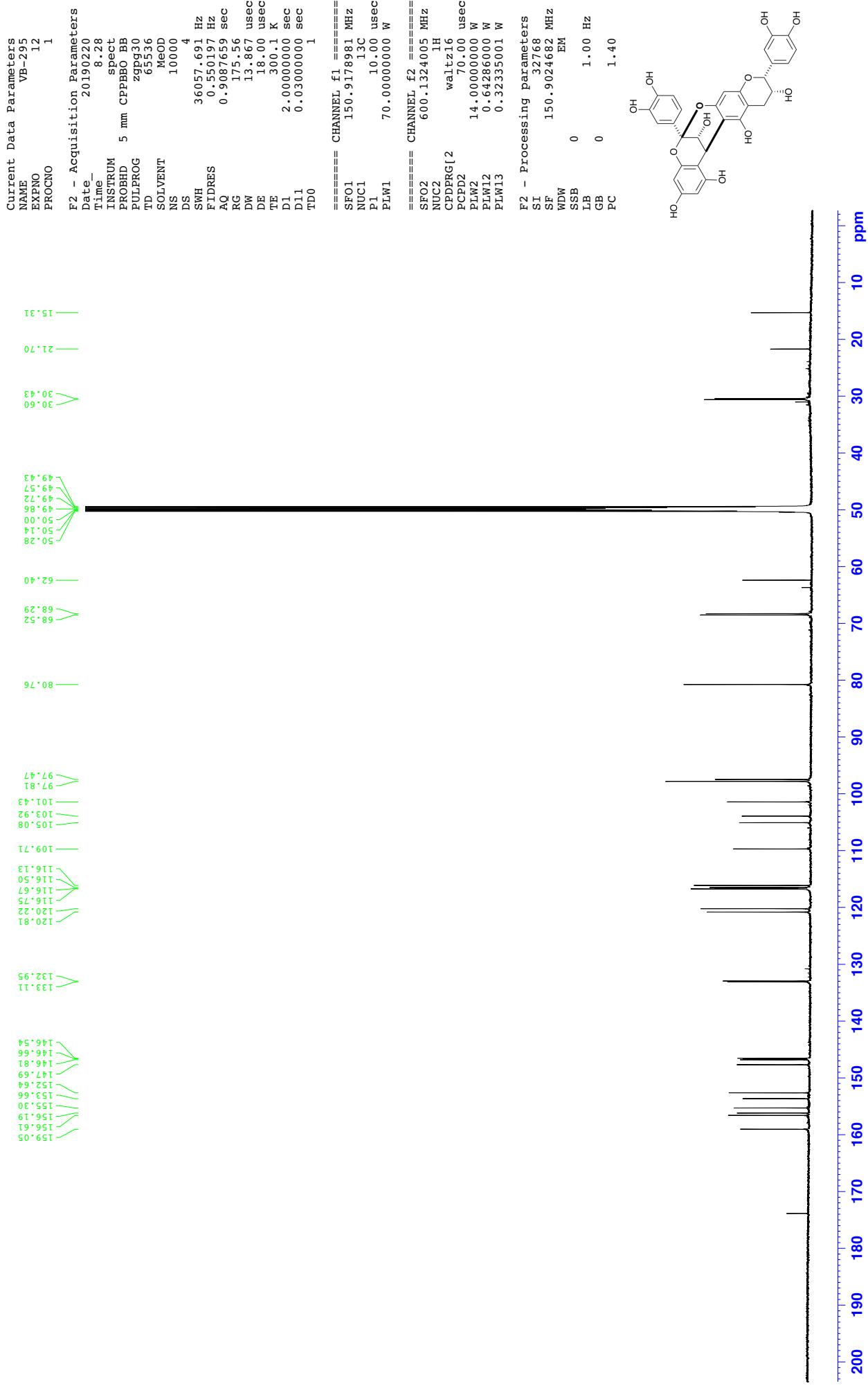
F2 - Acquisition Parameters  
Date 20190220  
Time 0.09  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 50  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.1833399 Hz  
AQ 2.7262976 sec  
RG 31.94  
DW 41.600 usec  
DE 10.00 usec  
TE 300.1 K  
D1 1.0000000 sec  
TDO 1

===== CHANNEL f1 ======  
SFO1 600.1337060 MHz  
NUC1 1H  
P1 12.00 usec  
PLW1 23.0000000 W

F2 - Processing parameters  
SI 65536  
SF 600.1299933 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

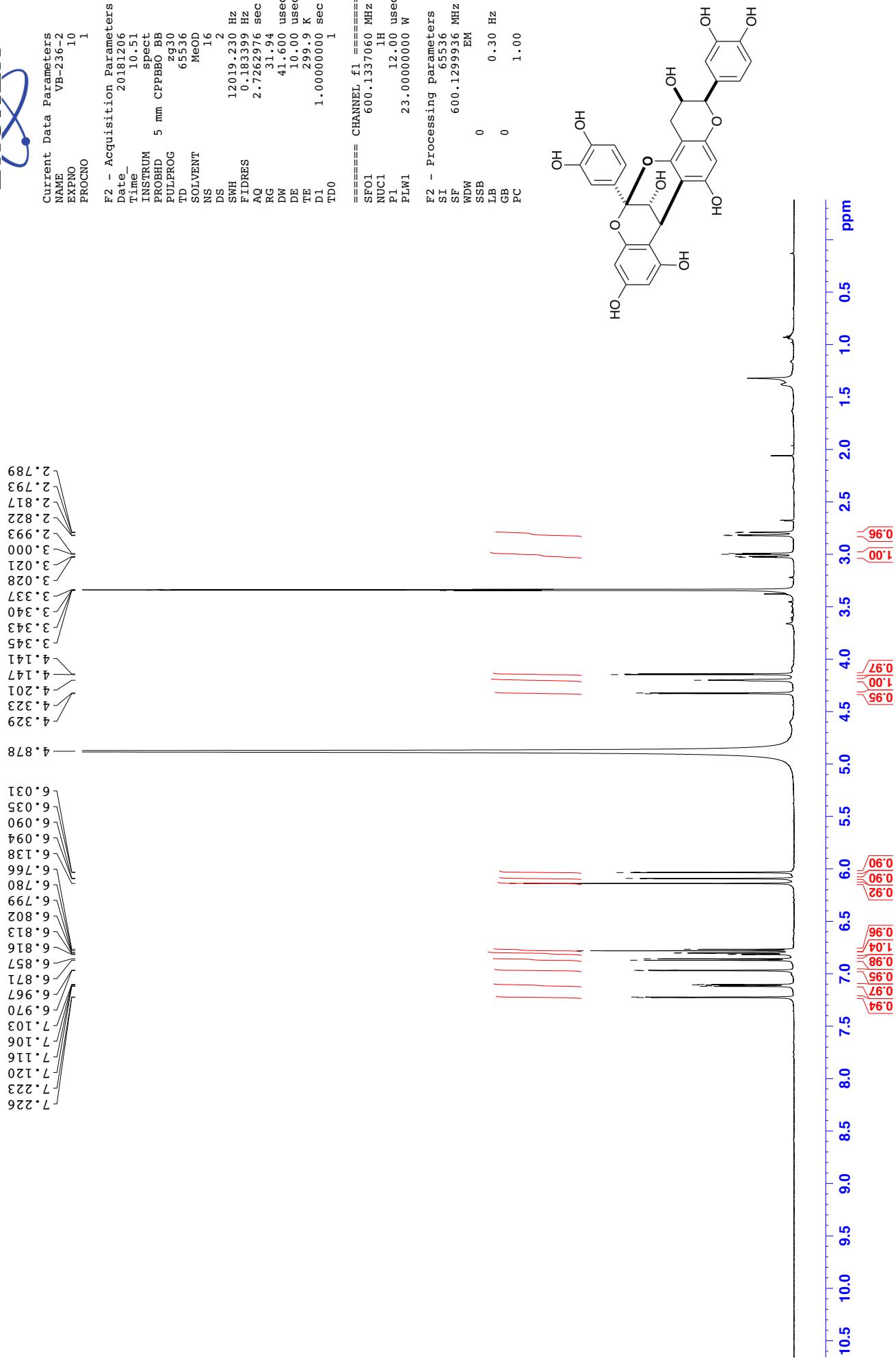


<sup>13</sup>C NMR of 17 (150MHz, CD<sub>3</sub>OD)



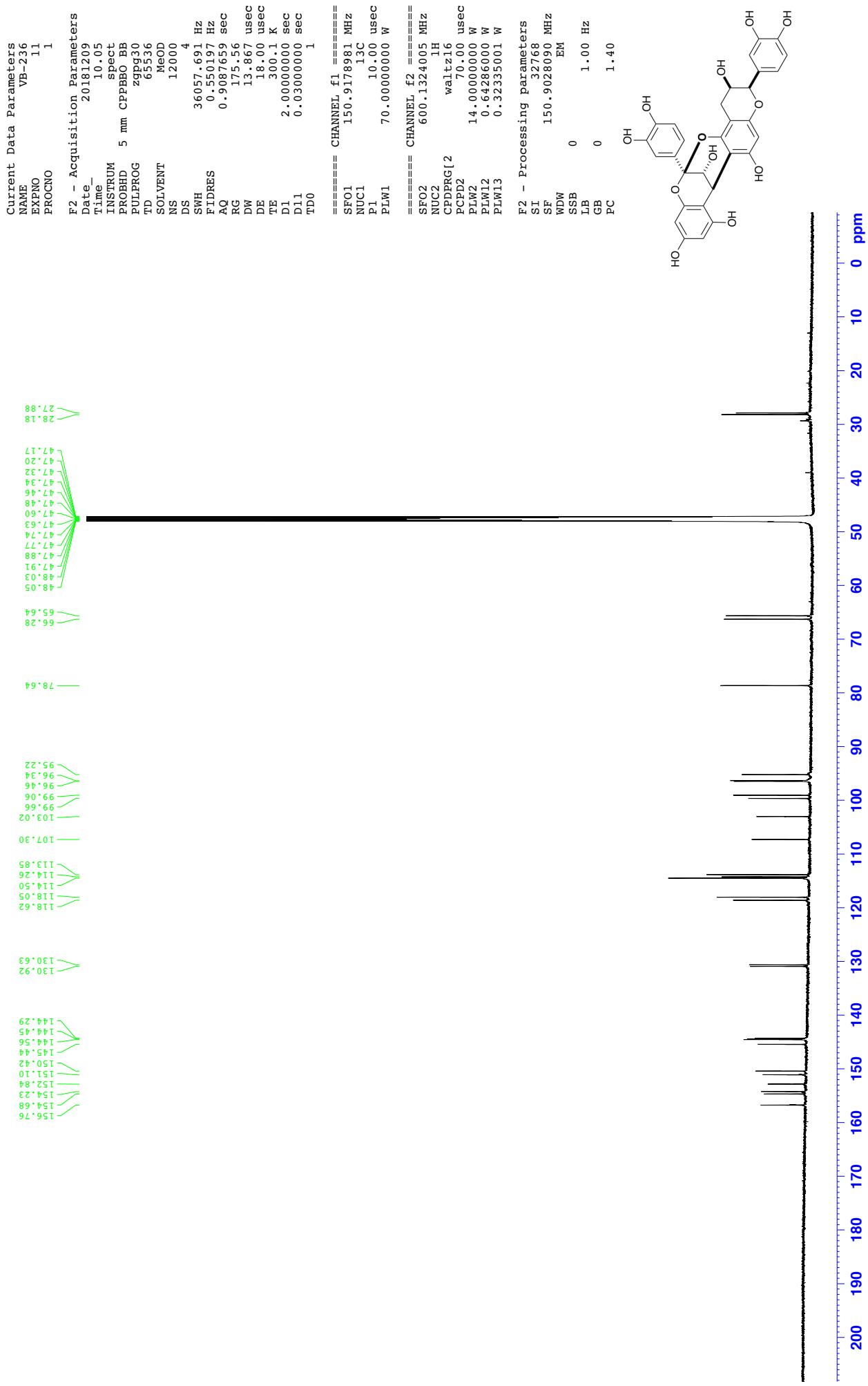


<sup>1</sup>H NMR of 18 (600MHz, CD<sub>3</sub>OD)



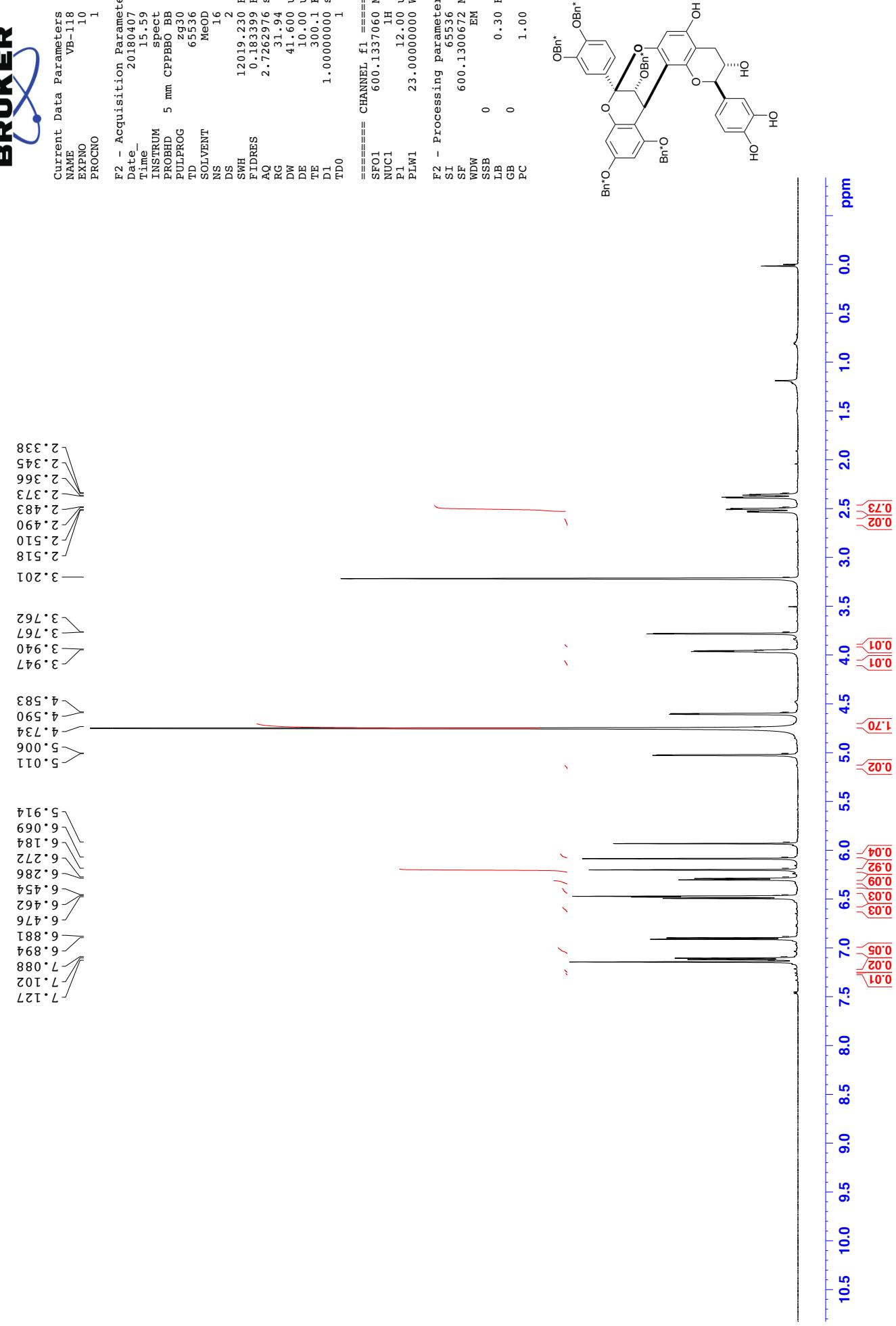


## <sup>13</sup>C NMR of 18 (150MHz, CD<sub>3</sub>OD)

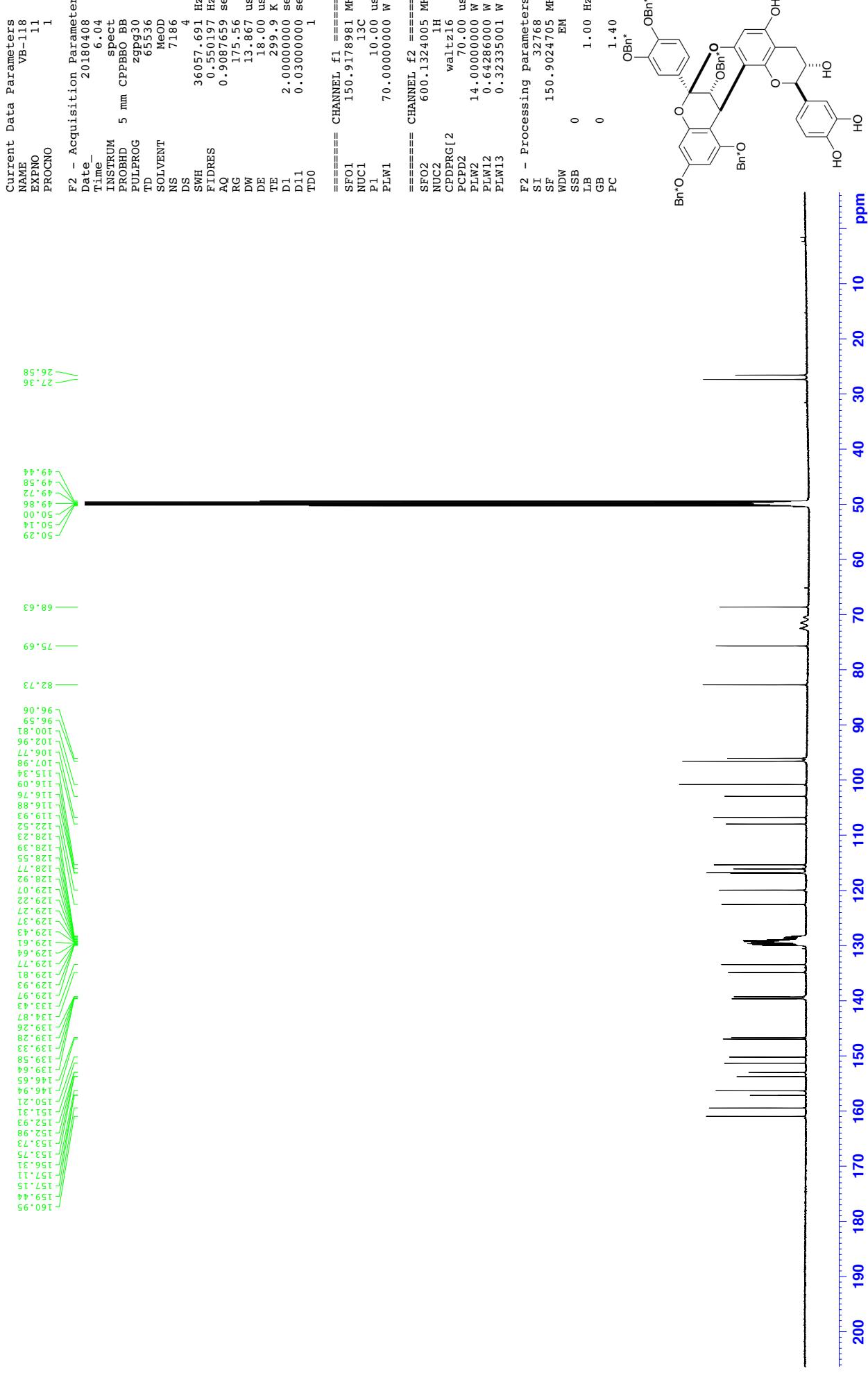




<sup>1</sup>H NMR of 20 (600MHz, CD<sub>3</sub>OD)

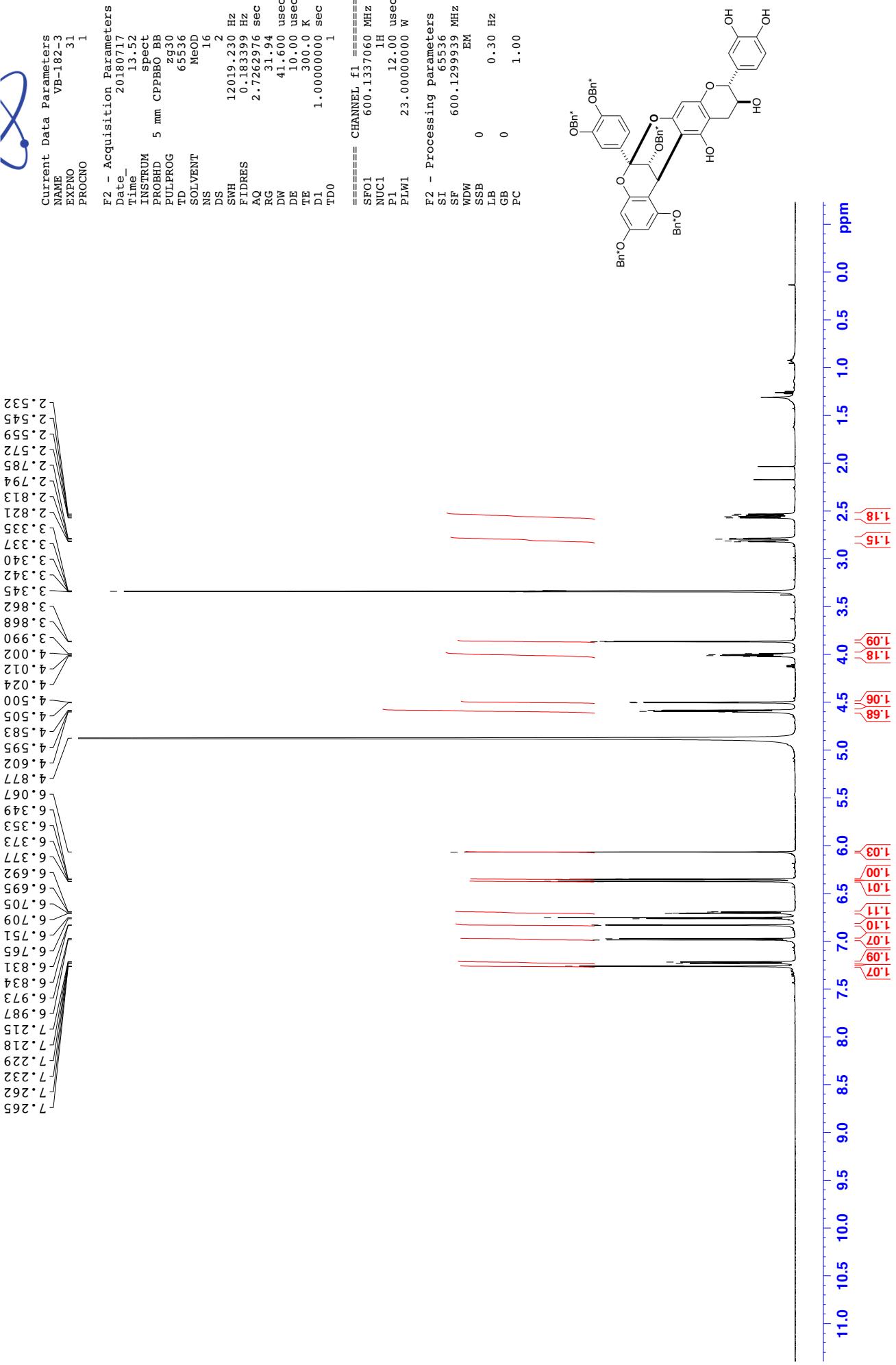


<sup>13</sup>C NMR of 20 (600MHz, CD<sub>3</sub>OD)

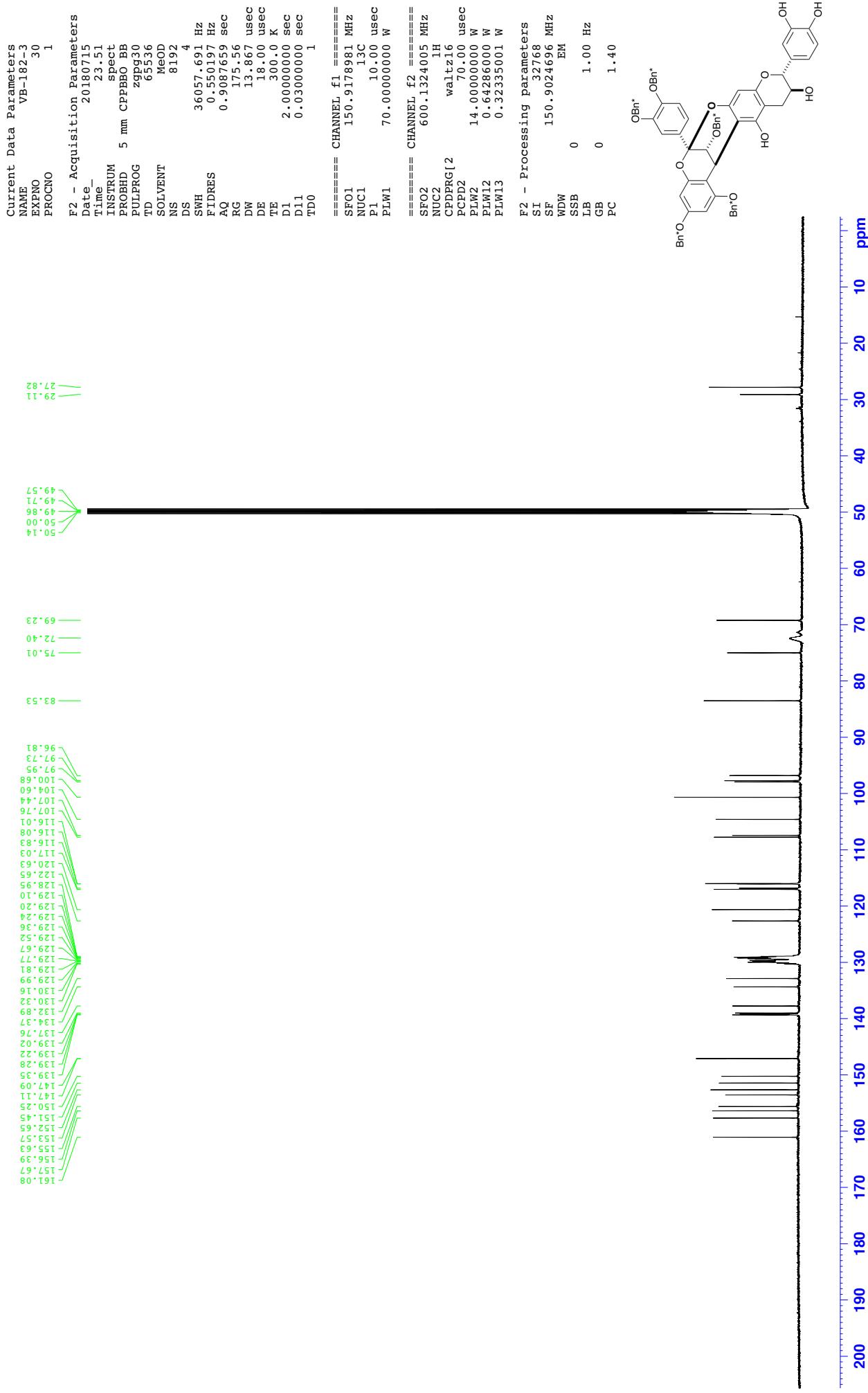




<sup>1</sup>H NMR of 21 (600MHz, CD<sub>3</sub>OD)



<sup>13</sup>C NMR of 21 (150MHz, CD<sub>3</sub>OD)





<sup>1</sup>H NMR of 22 (600MHz, CD<sub>3</sub>OD)

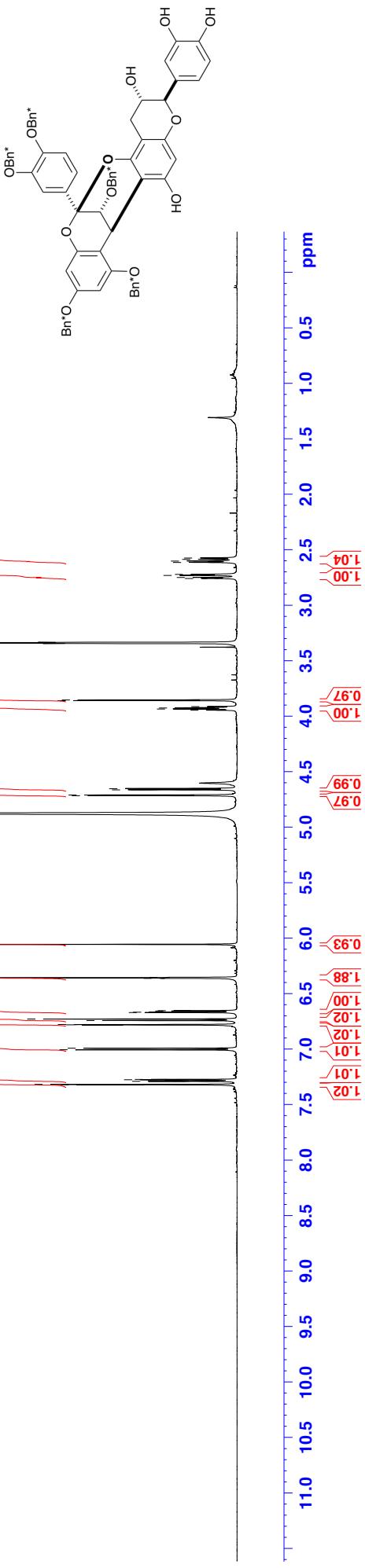
7.321  
7.318  
7.290  
7.287  
7.273  
7.276  
7.206  
6.992  
6.782  
6.779  
6.742  
6.670  
6.656  
6.653  
6.633  
6.358  
6.354  
6.057  
4.877  
4.716  
4.664  
3.946  
3.935  
3.926  
3.860  
3.854  
2.758  
2.750  
2.731  
2.723  
2.601  
2.586  
2.574  
2.574  
2.566  
2.562  
2.550  
2.544  
2.531  
2.514  
2.501  
2.496  
2.490  
2.484  
2.478  
2.472  
2.466  
2.460  
2.454  
2.448  
2.442  
2.436  
2.430  
2.424  
2.418  
2.412  
2.406  
2.400  
2.394  
2.388  
2.382  
2.376  
2.370  
2.364  
2.358  
2.354  
2.350  
2.344  
2.338  
2.332  
2.326  
2.320  
2.314  
2.308  
2.302  
2.296  
2.290  
2.284  
2.278  
2.272  
2.266  
2.260  
2.254  
2.248  
2.242  
2.236  
2.230  
2.224  
2.218  
2.212  
2.206  
2.200  
2.194  
2.188  
2.182  
2.176  
2.170  
2.164  
2.158  
2.152  
2.146  
2.140  
2.134  
2.128  
2.122  
2.116  
2.110  
2.104  
2.098  
2.092  
2.086  
2.080  
2.074  
2.068  
2.062  
2.056  
2.050  
2.044  
2.038  
2.032  
2.026  
2.020  
2.014  
2.008  
2.002  
2.000  
1.994  
1.988  
1.982  
1.976  
1.970  
1.964  
1.958  
1.952  
1.946  
1.940  
1.934  
1.928  
1.922  
1.916  
1.910  
1.904  
1.900  
1.894  
1.888  
1.882  
1.876  
1.870  
1.864  
1.858  
1.852  
1.846  
1.840  
1.834  
1.828  
1.822  
1.816  
1.810  
1.804  
1.800  
1.794  
1.788  
1.782  
1.776  
1.770  
1.764  
1.758  
1.752  
1.746  
1.740  
1.734  
1.728  
1.722  
1.716  
1.710  
1.704  
1.700  
1.694  
1.688  
1.682  
1.676  
1.670  
1.664  
1.658  
1.652  
1.646  
1.640  
1.634  
1.628  
1.622  
1.616  
1.610  
1.604  
1.600  
1.594  
1.588  
1.582  
1.576  
1.570  
1.564  
1.558  
1.552  
1.546  
1.540  
1.534  
1.528  
1.522  
1.516  
1.510  
1.504  
1.500  
1.494  
1.488  
1.482  
1.476  
1.470  
1.464  
1.458  
1.452  
1.446  
1.440  
1.434  
1.428  
1.422  
1.416  
1.410  
1.404  
1.400  
1.394  
1.388  
1.382  
1.376  
1.370  
1.364  
1.358  
1.352  
1.346  
1.340  
1.334  
1.328  
1.322  
1.316  
1.310  
1.304  
1.300  
1.294  
1.288  
1.282  
1.276  
1.270  
1.264  
1.258  
1.252  
1.246  
1.240  
1.234  
1.228  
1.222  
1.216  
1.210  
1.204  
1.200  
1.194  
1.188  
1.182  
1.176  
1.170  
1.164  
1.158  
1.152  
1.146  
1.140  
1.134  
1.128  
1.122  
1.116  
1.110  
1.104  
1.100  
1.094  
1.088  
1.082  
1.076  
1.070  
1.064  
1.058  
1.052  
1.046  
1.040  
1.034  
1.028  
1.022  
1.016  
1.010  
1.004  
1.000

Current Data Parameters  
NAME VB-182-2  
EXPNO 31  
PROCNO 1

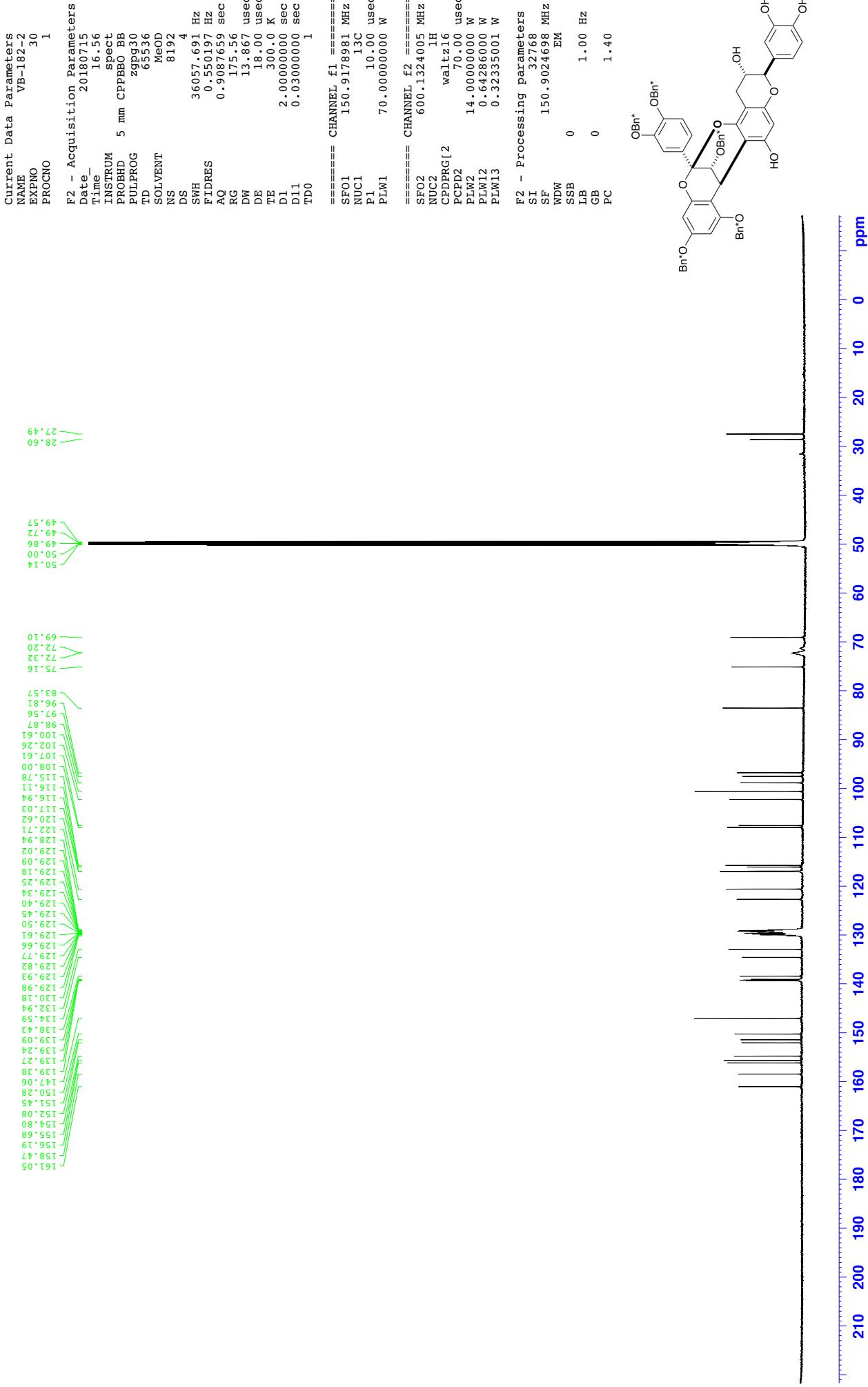
F2 - Acquisition Parameters  
Date 20180717  
Time 13.47  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 16  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.183399 Hz  
AQ 2.762976 sec  
RG 31.94  
DW 41.600 usec  
DE 10.00 usec  
TE 300.0 K  
D1 1.0000000 sec  
TDO 1

===== CHANNEL f1 ======  
SFO1 600.1337060 MHz  
NUC1 1H  
P1 12.00 usec  
PLW1 23.00000000 W

F2 - Processing parameters  
SI 65536  
SF 600.1299938 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

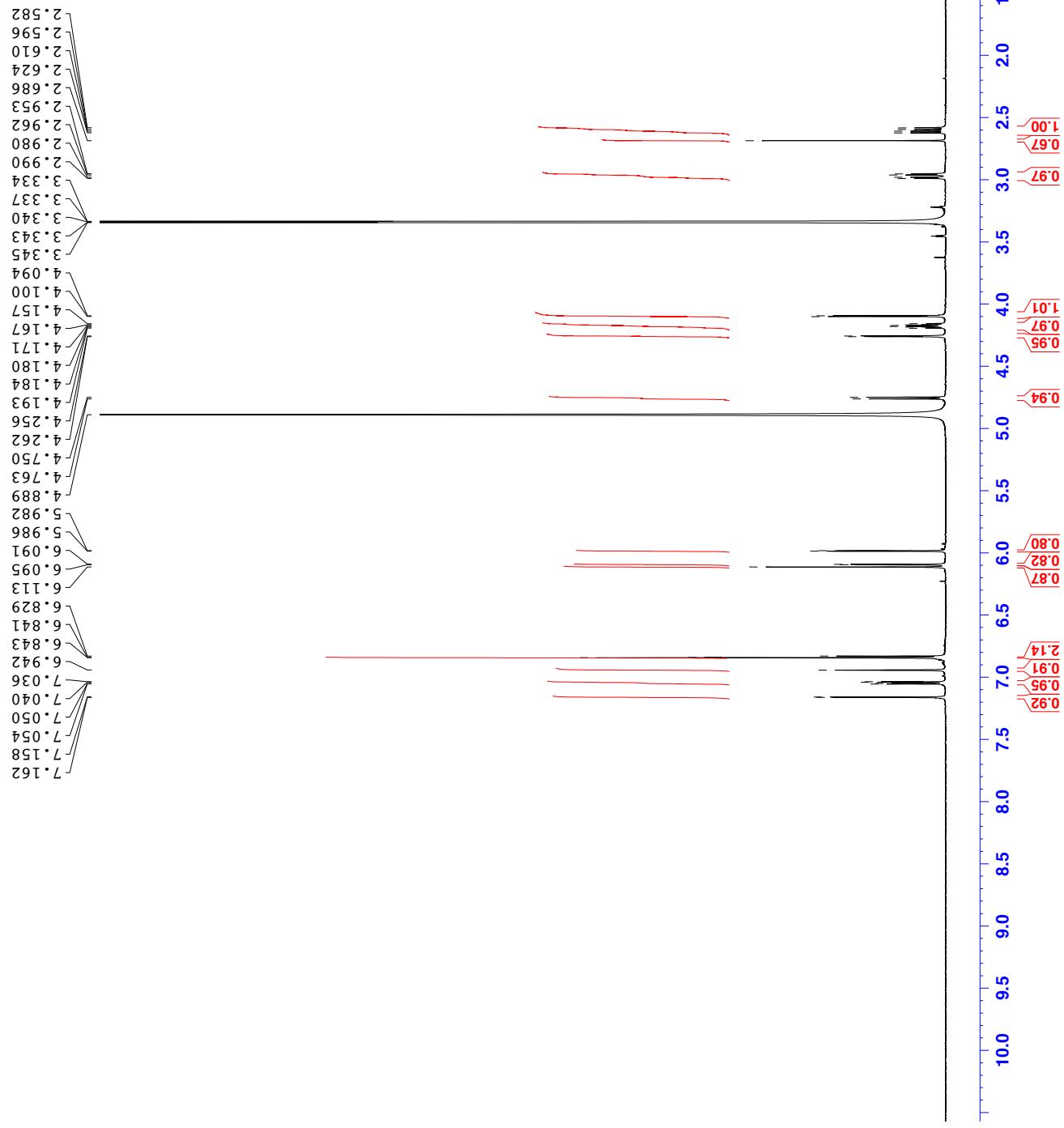


<sup>13</sup>C NMR of 22 (150MHz, CD<sub>3</sub>OD)





<sup>1</sup>H NMR of 1 (600MHz, CD<sub>3</sub>OD)



Current Data Parameters  
NAME VB-81  
EXPNO 20  
PROCNO 1

F2 - Acquisition Parameters  
Date 20190712  
Time 23.13  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 16  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.183399 Hz  
AQ 2.7562976 sec  
RG 31.94  
DW 41.600 usec  
DE 10.00 usec  
TE 298.1 K  
D1 1.0000000 sec  
TDO 1

===== CHANNEL f1 =====  
SFO1 600.1337060 MHz  
NUC1 1H  
P1 12.00 usec  
PLW1 21.00000000 W

F2 - Processing parameters  
SI 65536  
SF 600.1299934 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

<sup>13</sup>C NMR of 1 (150MHz, CD<sub>3</sub>OD)



```

Current Data Parameters
NAME          VB-81
EXPNO         11
PROCNO        1

F2 - Acquisition Parameters
Date         20180408
Time         12.52
INSTRUM      spect
PROBID       5 mm CPPBBO BB
PULPROG     zgpg30
TD           65536
T1            4
SWH          36057.691 Hz
FIDRES      0.9987659 sec
AQ           0.250197 Hz
RG           175.56
DW           13.867 usec
DE           18.00 usec
TE           300.1 K
D1           2.0000000 sec
D11          0.03000000 sec
TD0          1

===== CHANNEL f1 =====
SFO1        150.9178981 MHz
NUC1         13C
P1           10.00 usec
PLW1        70.00000000 W

===== CHANNEL f2 =====
SFO2        600.1324005 MHz
NUC2         1H
CPDPKG[1]  2
PCPD2       14.00000000 usec
PLW2        0.64286000 W
PLW12       0.32335001 W
PLW13       0.32335001 W

F2 - Processing parameters
SI            32768
SF          150.9024672 MHz
WDW           EM
SSB            0
LB            1.00 Hz
GB            0
PC           1.40

```

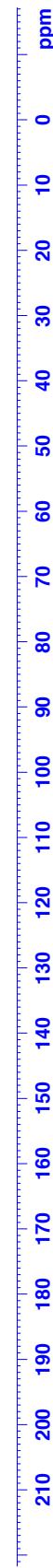
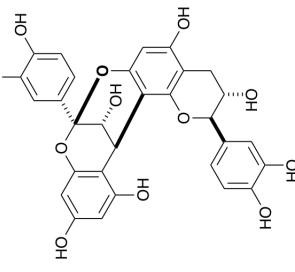
-29.89  
30.10

49.58  
49.72  
49.76  
50.00  
50.14  
50.28

68.69  
68.97

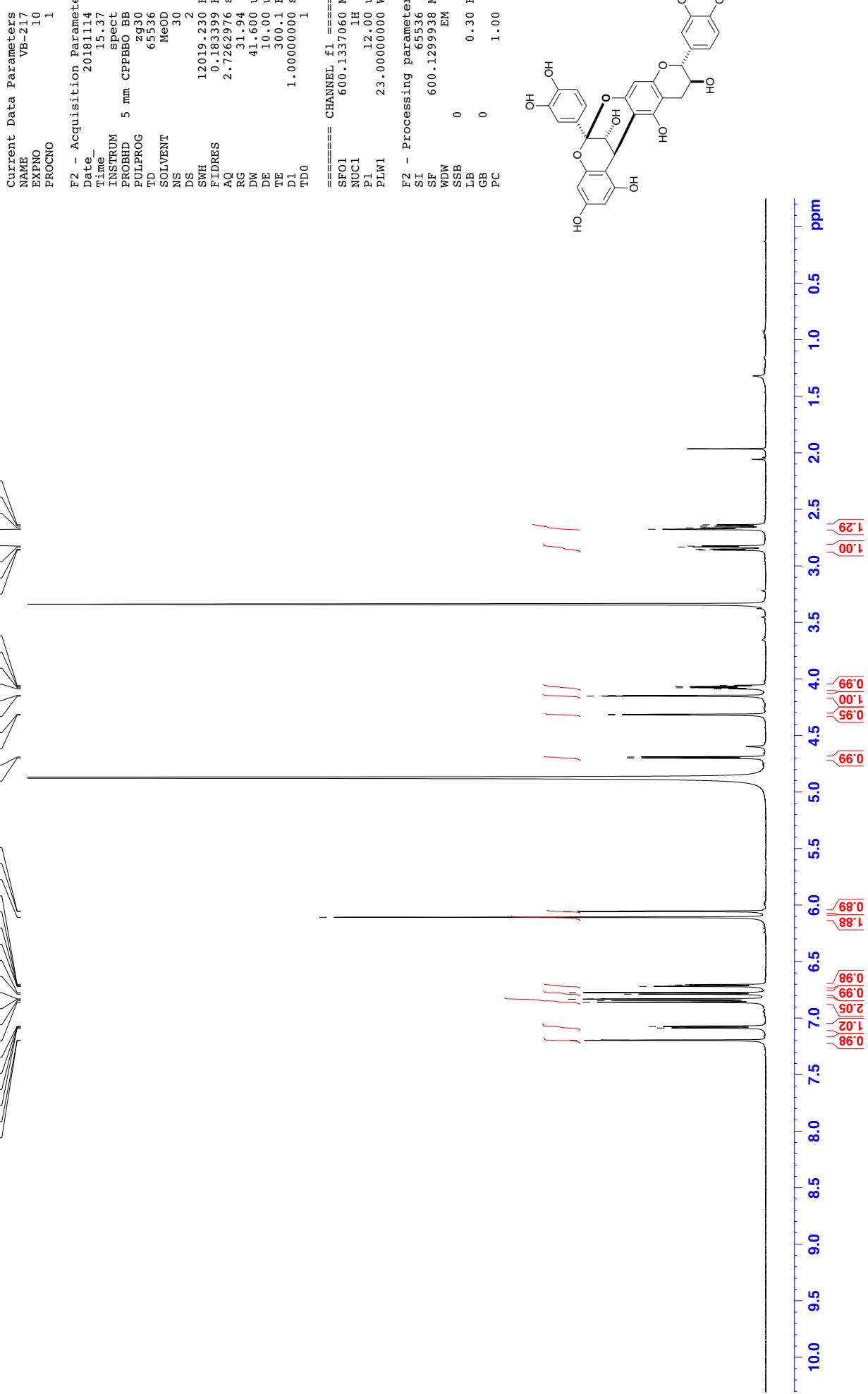
85.41

97.42  
99.04  
101.20  
104.01  
104.90  
107.68  
116.48  
116.58  
117.19  
117.69  
120.69  
121.55  
131.44  
133.19  
146.53  
147.23  
147.66  
147.96  
152.29  
153.04  
153.08  
155.11  
156.99  
157.03  
157.65  
158.94  
159.04



**BRUKER**

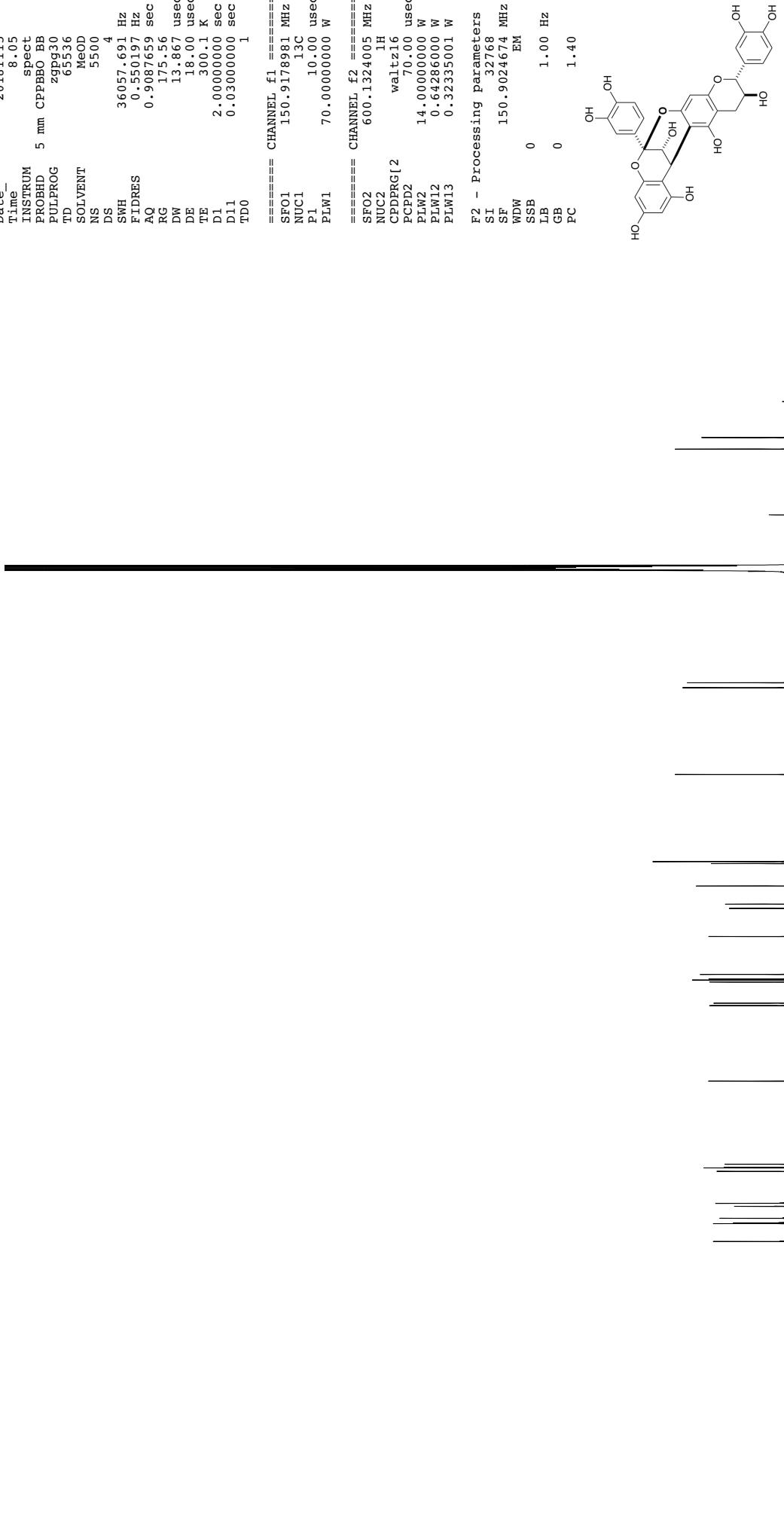
<sup>1</sup>H NMR of 23 (600MHz, CD<sub>3</sub>OD)





<sup>13</sup>C NMR of 23 (150MHz, CD<sub>3</sub>OD)

Current Data Parameters  
NAME VB-217  
EXPNO 11  
PROCNO 1  
  
F2 - Acquisition Parameters  
Date 20181115  
Time 8.05  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zgppg30  
TD 65536  
SOLVENT MeOD  
NS 5500  
DS 4  
SWH 36057.691 Hz  
FIDRES 0.9987659 sec  
AQ 0.250197 Hz  
RG 175.56  
DW 13.867 usec  
DE 18.00 usec  
TE 300.1 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TD0 1  
  
===== CHANNEL f1 ======  
SFO1 150.9178981 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 70.00000000 W  
  
===== CHANNEL f2 ======  
SFO2 600.1324005 MHz  
NUC2 1H  
CPDPKG[ 2  
PCPD2 14.00000000 usec  
PLW2 0.64286000 W  
PLW12 0.32335001 W  
PLW13 0.32335001 W  
  
F2 - Processing parameters  
SI 32768  
SF 150.9024674 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40





<sup>1</sup>H NMR of 24 (600MHz, CD<sub>3</sub>OD)

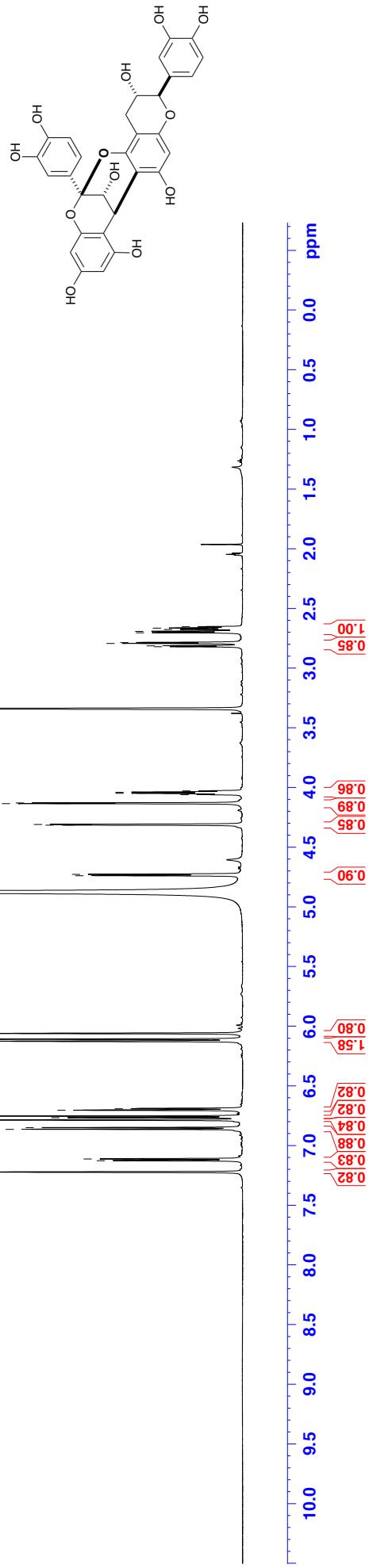
2.877  
4.113  
4.130  
4.135  
4.151  
4.160  
4.049  
4.039  
4.029  
3.415  
3.400  
4.030  
4.021  
2.793  
2.785  
2.703  
2.691  
2.664  
2.653

Current Data Parameters  
NAME VB-216  
EXPNO 10  
PROCNO 1

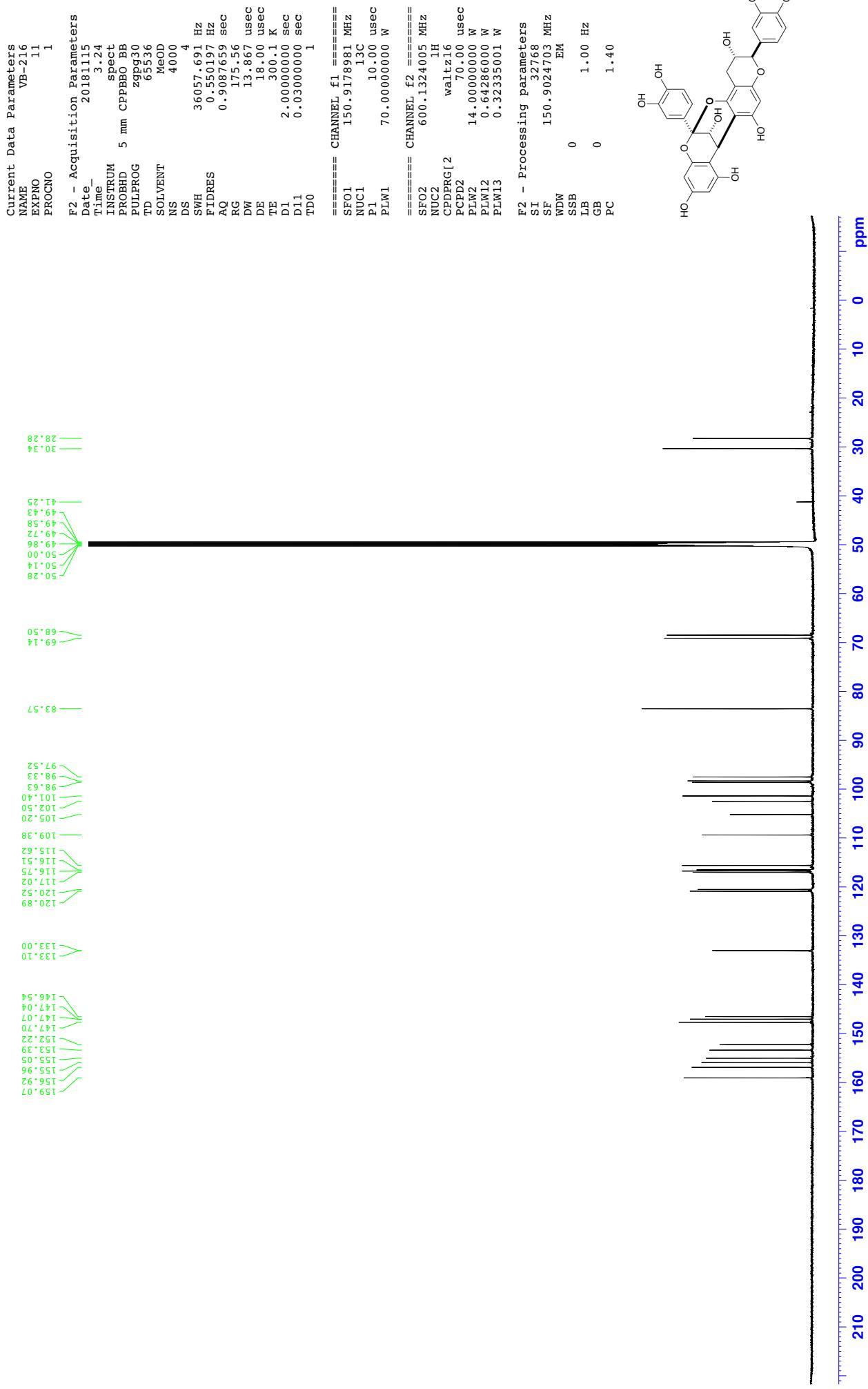
F2 - Acquisition Parameters  
Date 20181114  
Time 15.30  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zg30  
TD 65536  
SOLVENT MeOD  
NS 30  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.183399 Hz  
AQ 2.7262976 sec  
RG 31.94  
DW 41.600 usec  
DE 10.00 usec  
TE 300.0 K  
D1 1.0000000 sec  
TDO 1

===== CHANNEL f1 =====  
SFO1 600.1337060 MHz  
NUC1 1H  
P1 12.00 usec  
PLW1 23.00000000 W

F2 - Processing parameters  
SI 65536  
SF 600.1299942 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

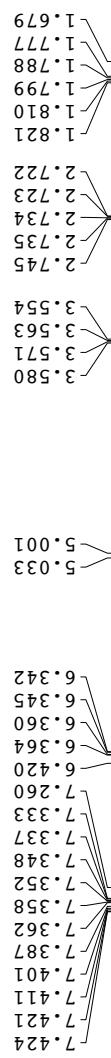


<sup>13</sup>C NMR of 24 (150MHz, CD<sub>3</sub>OD)





<sup>1</sup>H NMR of 26 (600MHz, CDCl<sub>3</sub>)

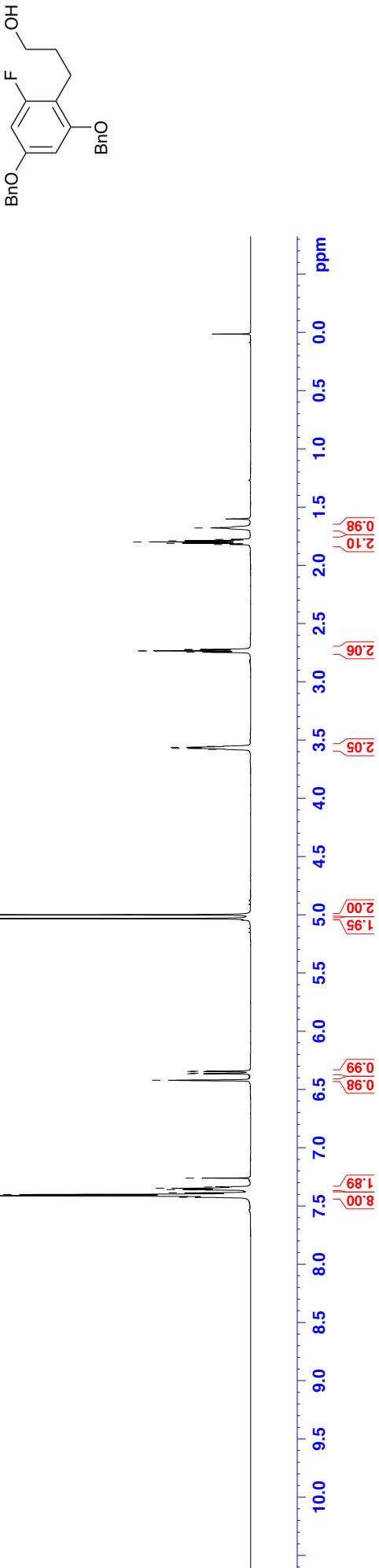


Current Data Parameters  
NAME VB-297  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date 20190227  
Time 12.09  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zg30  
TD 65536  
SOLVENT CDC13  
NS 16  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.183399 Hz  
AQ 2.7622976 sec  
RG 17.5  
DW 41.600 usec  
DE 10.00 usec  
TE 300.1 K  
D1 1.0000000 sec  
TDO 1

===== CHANNEL f1 ======  
SFO1 600.1337060 MHz  
NUC1 1H  
P1 12.00 usec  
PLW1 23.00000000 W

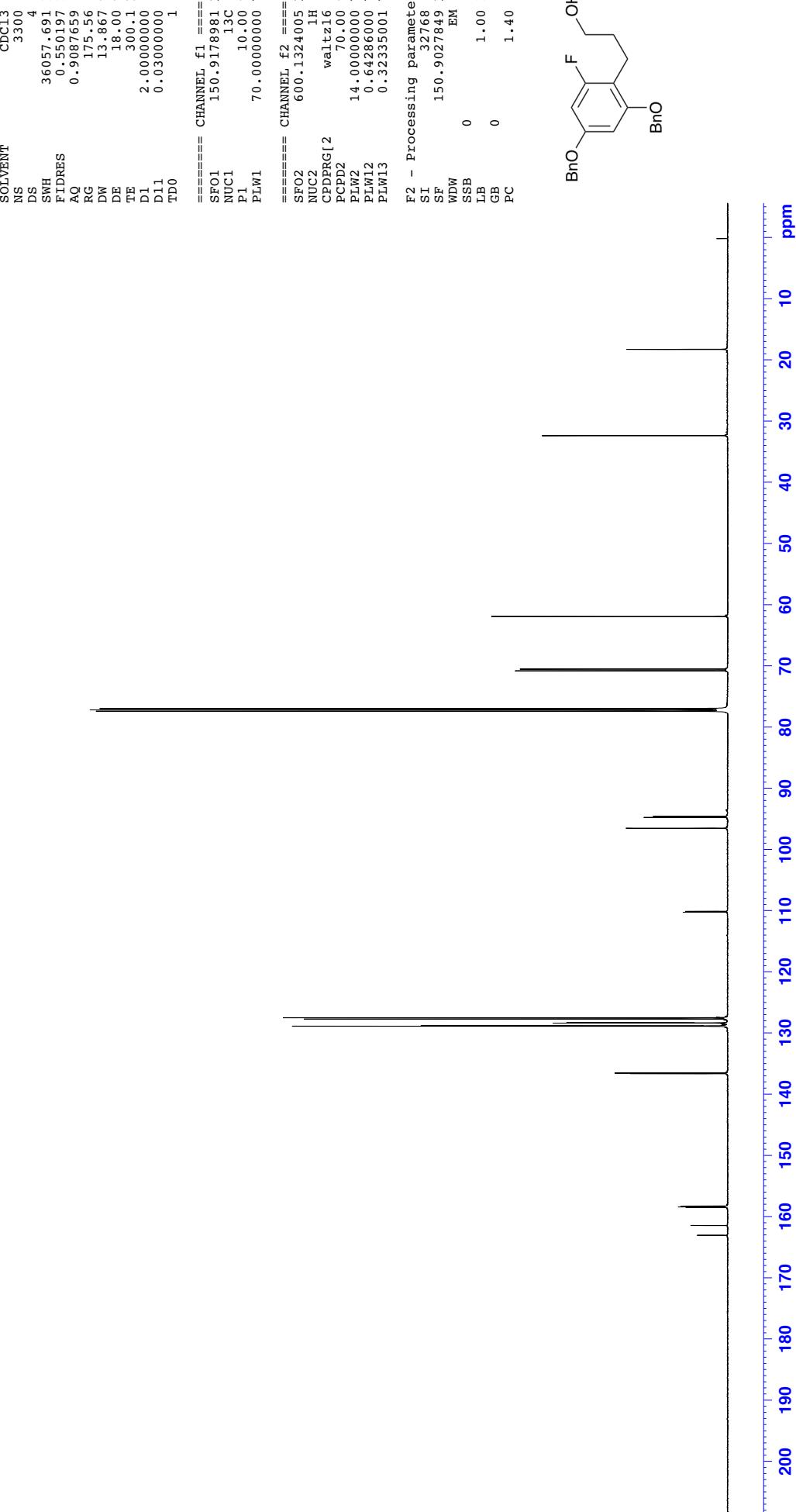
F2 - Processing parameters  
SI 65536  
SF 600.1300146 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00





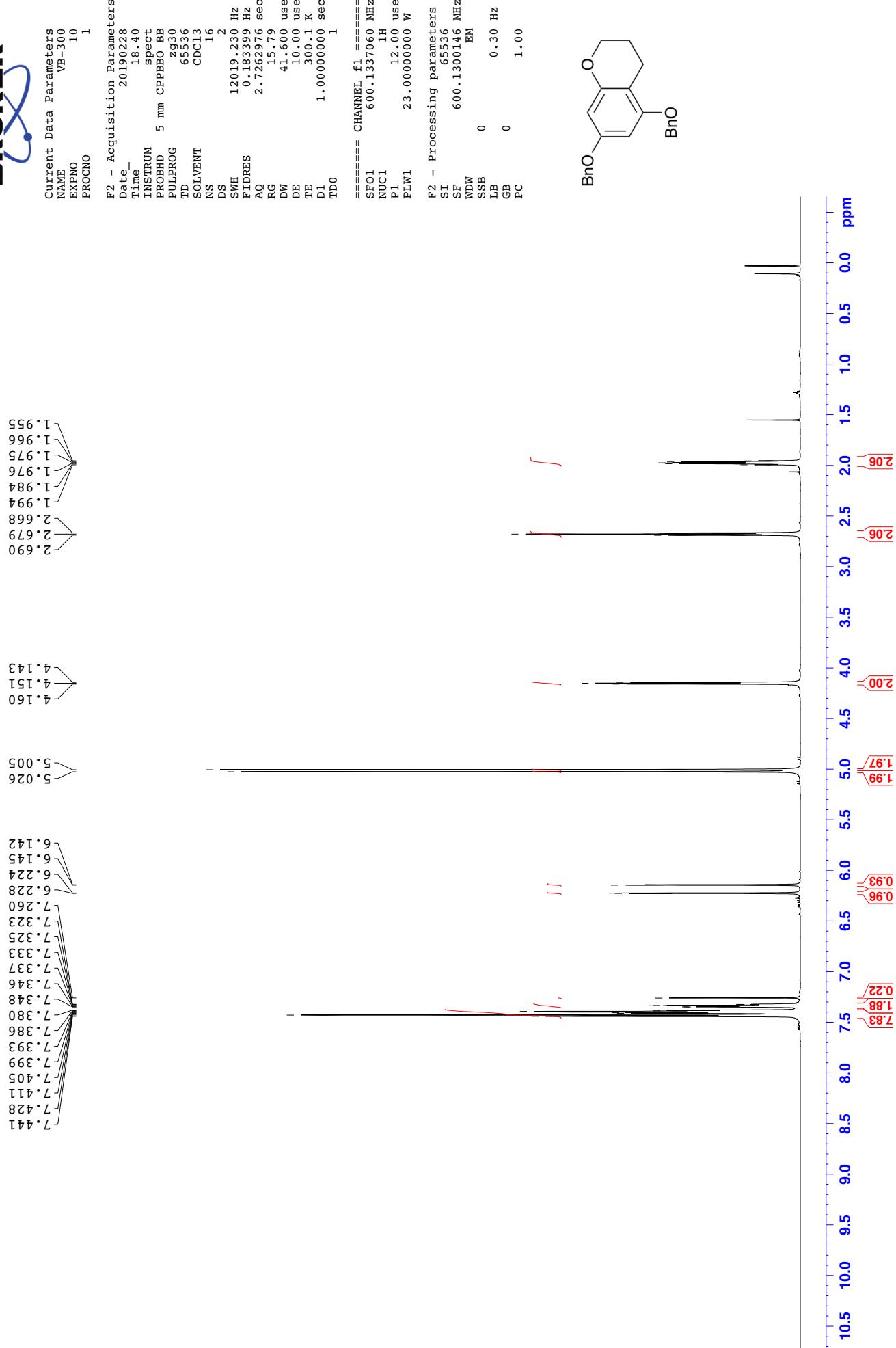
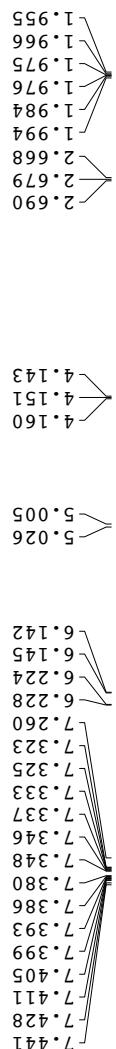
<sup>13</sup>C NMR of 26 (150MHz, CDCl<sub>3</sub>)

Current Data Parameters  
NAME VB-297  
EXPNO 11  
PROCNO 1  
  
F2 - Acquisition Parameters  
Date 20190228  
Time 2.48  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zgpg30  
TD 65536  
SOLVENT CDC13  
NS 3300  
DS 4  
SWH 36057.691 Hz  
FIDRES 0.9987659 sec  
AQ 0.550197 Hz  
RG 175.56  
DW 13.867 usec  
DE 18.00 usec  
TE 300.1 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TD0 1  
  
===== CHANNEL f1 ======  
SFO1 150.9178981 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 70.00000000 M  
  
===== CHANNEL f2 ======  
SFO2 600.1324005 MHz  
NUC2 1H  
CPDPKG[ 2 waltz16  
PCPD2 14.00000000 usec  
PLW2 0.64286000 W  
PLW12 0.32335001 W  
PLW13 0.32335001 W  
  
F2 - Processing parameters  
SI 32768  
SF 150.9027849 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40



**BRUKER**

**$^1\text{H}$  NMR of 27 (600MHz,  $\text{CDCl}_3$ )**



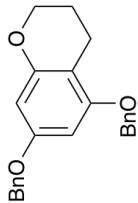
<sup>13</sup>C NMR of 27 (150MHz, CDCl<sub>3</sub>)



Current Data Parameters  
NAME VB-300  
PRONO 11  
PROCNO 1

F2 - Acquisition Parameters  
Date 20190303  
Time 2.48  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zgpg30  
TD 65536  
SOLVENT CDC13  
NS 3300  
DS 4  
SWH 36057.691 Hz  
FIDRES 0.9987659 sec  
AQ 0.550197 Hz  
RG 175.56  
DW 13.867 usec  
DE 18.00 usec  
TE 300.1 K  
D1 2.0000000 sec  
D11 0.0300000 sec  
TD0 1

===== CHANNEL f1 ======  
SFO1 150.9178981 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 70.0000000 A  
===== CHANNEL f2 ======  
SFO2 600.1324005 MHz  
NUC2 1H  
CPDPKG[ 2 waltz16  
PCPD2 14.0000000 usec  
PLW2 0.64286000 W  
PLW12 0.32335001 W  
PLW13  
F2 - Processing parameters  
SI 32768  
SF 150.9027882 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40



19.17  
22.13  
22.48

66.60  
70.03  
70.27  
76.99  
77.20  
77.41

93.23  
94.92

104.62

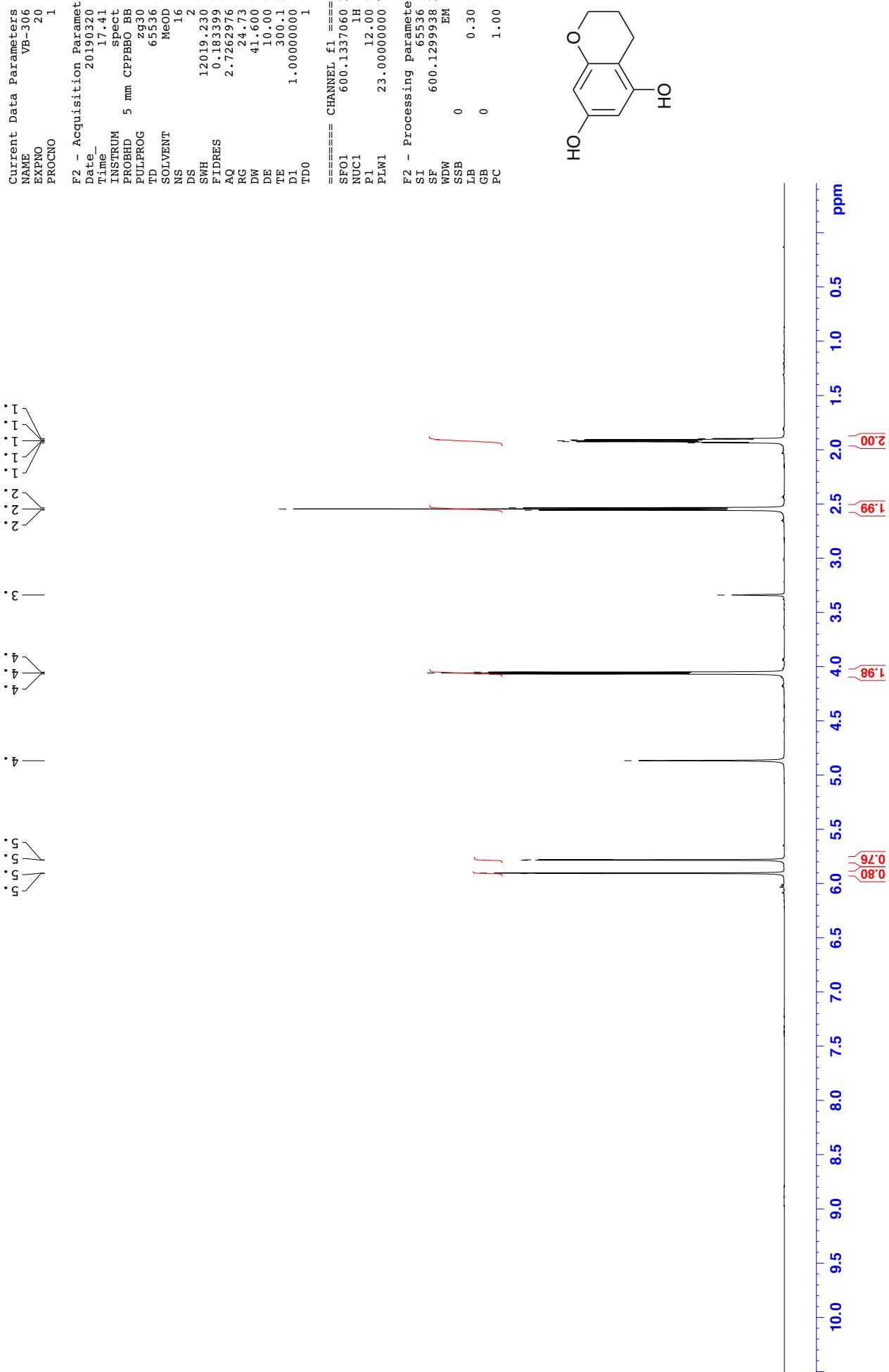
127.31  
127.73  
127.95  
128.08  
128.67  
128.73  
128.74  
137.40

156.31  
157.91  
158.48





<sup>1</sup>H NMR of 28 (600MHz, CD<sub>3</sub>OD)



**<sup>13</sup>C NMR of 28 (150MHz, CD<sub>3</sub>OD)**



Current Data Parameters  
NAME VB-306  
EXPNO 1  
PROCNO 1  
  
F2 - Acquisition Parameters  
Date 20190321  
Time 17.02  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zgpg30  
TD 65536  
SOLVENT MeOD  
NS 2200  
DS 4  
SWH 36057.691 Hz  
FIDRES 0.9987659 sec  
AQ 175.56  
RG 13.867 usec  
DE 18.00 usec  
TE 300.1 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TD0 1  
  
===== CHANNEL f1 ======  
SFO1 150.9178981 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 70.0000000 W  
  
===== CHANNEL f2 ======  
SFO2 600.1324005 MHz  
NUC2 1H  
CPDPKG[ 2 waltz16  
PCPD2 14.0000000 usec  
PLW2 0.64286000 W  
PLW12 0.32335001 W  
PLW13  
  
F2 - Processing parameters  
SI 32768  
SF 150.9024737 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

20.72  
24.21

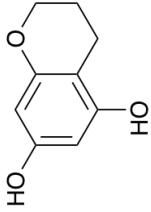
49.33  
49.28  
49.27  
49.26  
49.25  
49.24  
49.23  
50.00  
50.14  
50.28

68.08

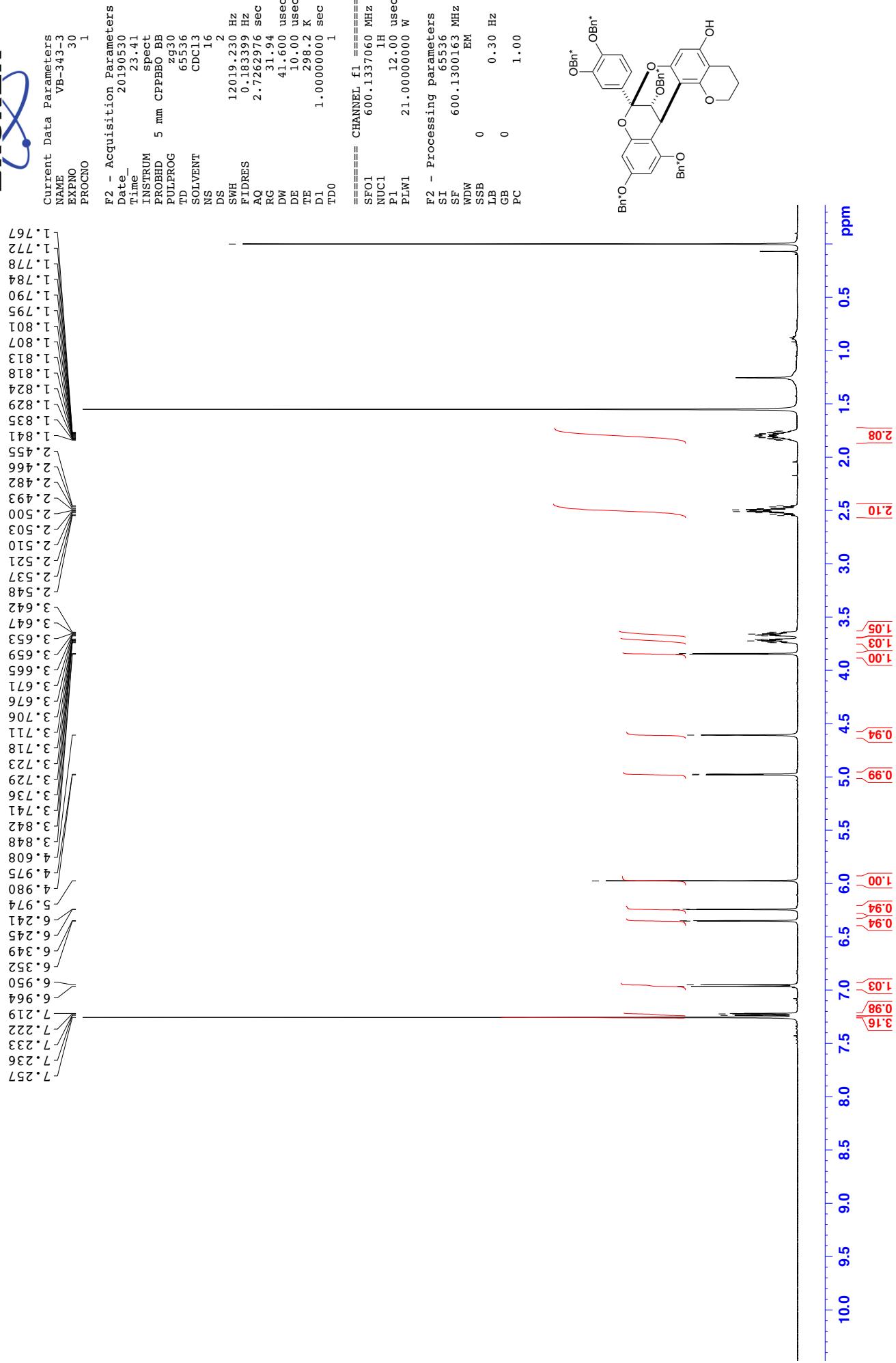
96.70  
96.72

103.61

158.06  
158.10  
158.17  
158.20  
158.23  
158.38  
158.40

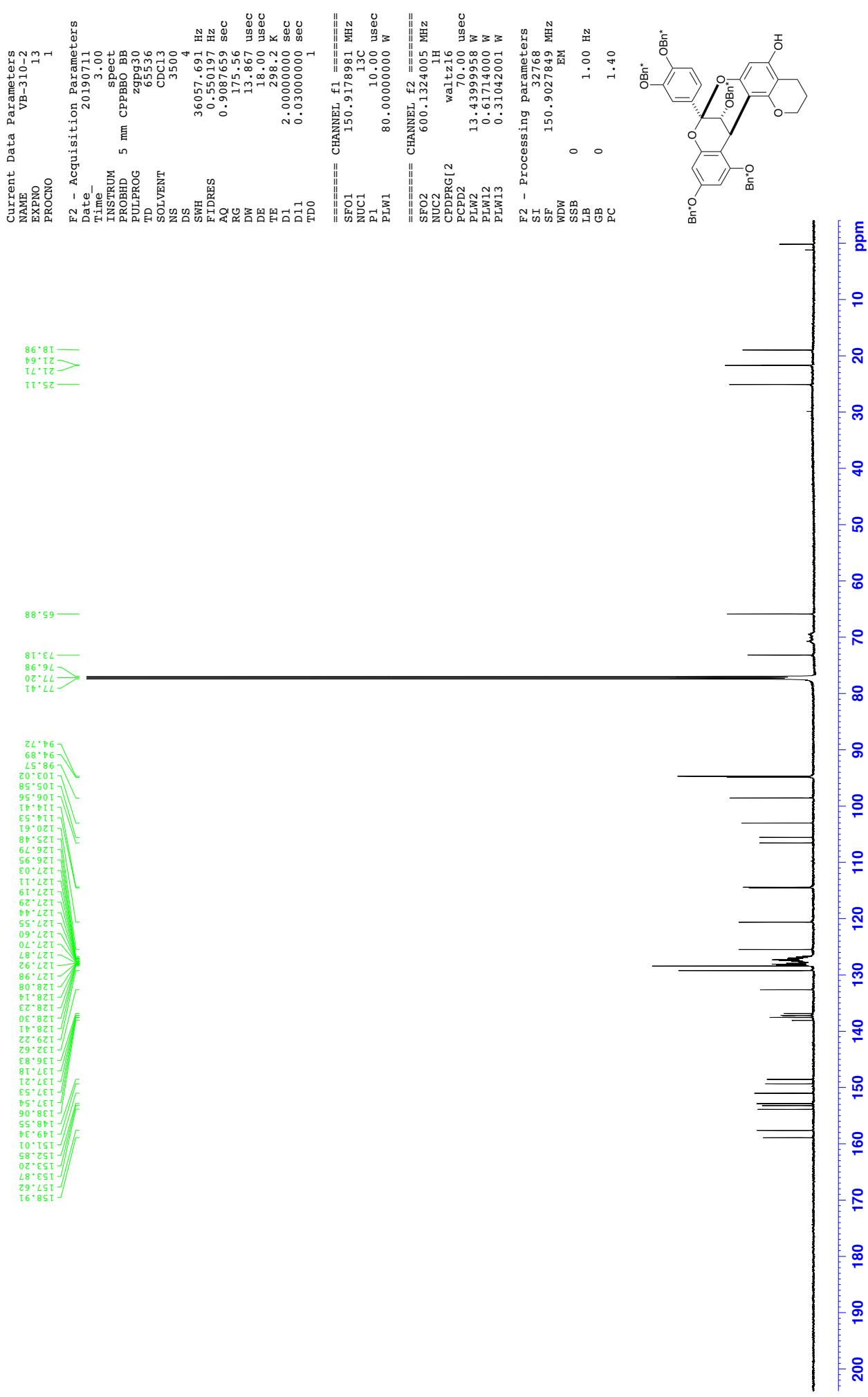


<sup>1</sup>H NMR of 29 (600MHz, CDCl<sub>3</sub>)





<sup>13</sup>C NMR of 29 (150MHz, CDCl<sub>3</sub>)



<sup>1</sup>H NMR of 30 (600MHz, CDCl<sub>3</sub>)

**BRUKER**

Current Data Parameters  
NAME VB-310-A  
EXPNO 30  
PROCNO 1

F2 - Acquisition Parameter  
Date 20190709  
Time 12:20  
INSTRUM spect  
PROBHD 5 mm CPPBBO BB  
PULPROG zg30  
TD 65536  
SOLVENT CDCl<sub>3</sub>  
NS 16  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.183339 Hz  
AQ 2.7262976 sec  
RG 31.94  
DW 41.600 us  
DE 10.00 us  
TE 298.2 K  
D1 1.00000000 sec  
TDO 1

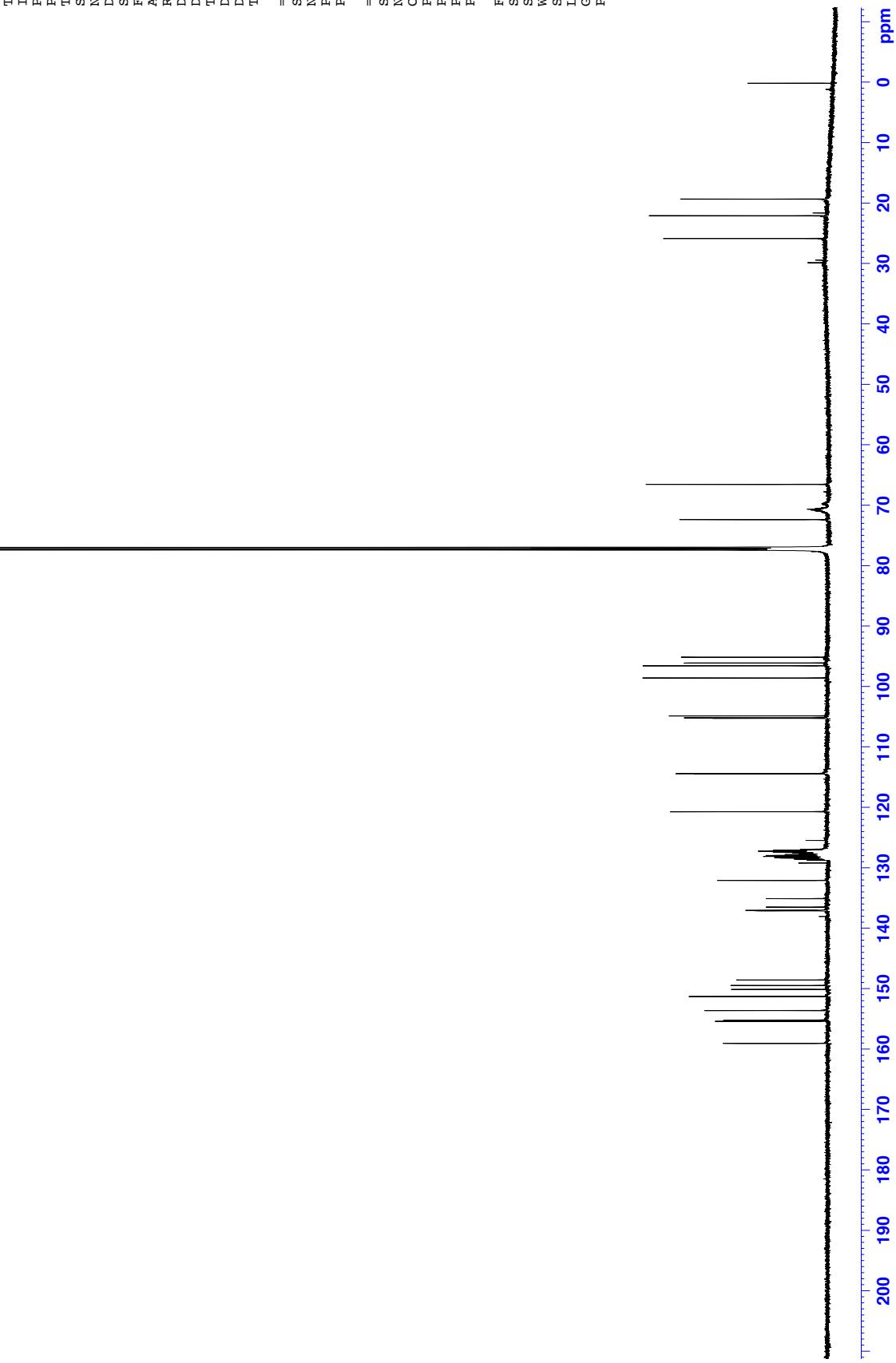
===== CHANNEL f1 =====  
SFO1 600.1337000 MHz  
NUC1 <sup>1</sup>H  
P1 12.00 us  
PLW1 21.00000000 W

F2 - Processing Parameter  
SI 65536  
SF 600.1300145 MHz  
WDW EM  
SSB 0  
LB 0  
GB 0  
PC 1.00



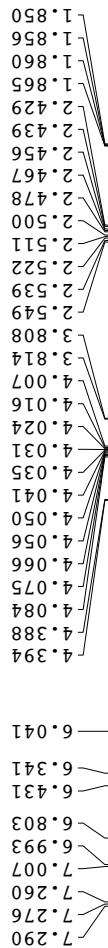
<sup>13</sup>C NMR of 30 (150MHz, CDCl<sub>3</sub>)

Current Data Parameters  
NAME VB-310-IA  
EXPNO 31  
PROCNO 1  
  
F2 - Acquisition Parameters  
Date 20190710  
Time 7.05  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zgpg30  
TD 65536  
SOLVENT CDC13  
NS 4000  
DS 4  
SWH 36057.691 Hz  
FIDRES 0.9987659 sec  
AQ 0.550197 Hz  
RG 175.56  
DW 13.867 usec  
DE 18.00 usec  
TE 298.1 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TDO 1  
  
===== CHANNEL f1 ======  
SFO1 150.9178981 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 80.0000000 W  
  
===== CHANNEL f2 ======  
SFO2 600.1324005 MHz  
NUC2 1H  
CPDPKG[ 2  
PCPD2 13.4399958 W  
PLW2 0.61714000 W  
PLW12 0.31042001 W  
PLW13  
  
F2 - Processing parameters  
SI 32768  
SF 150.9027838 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40





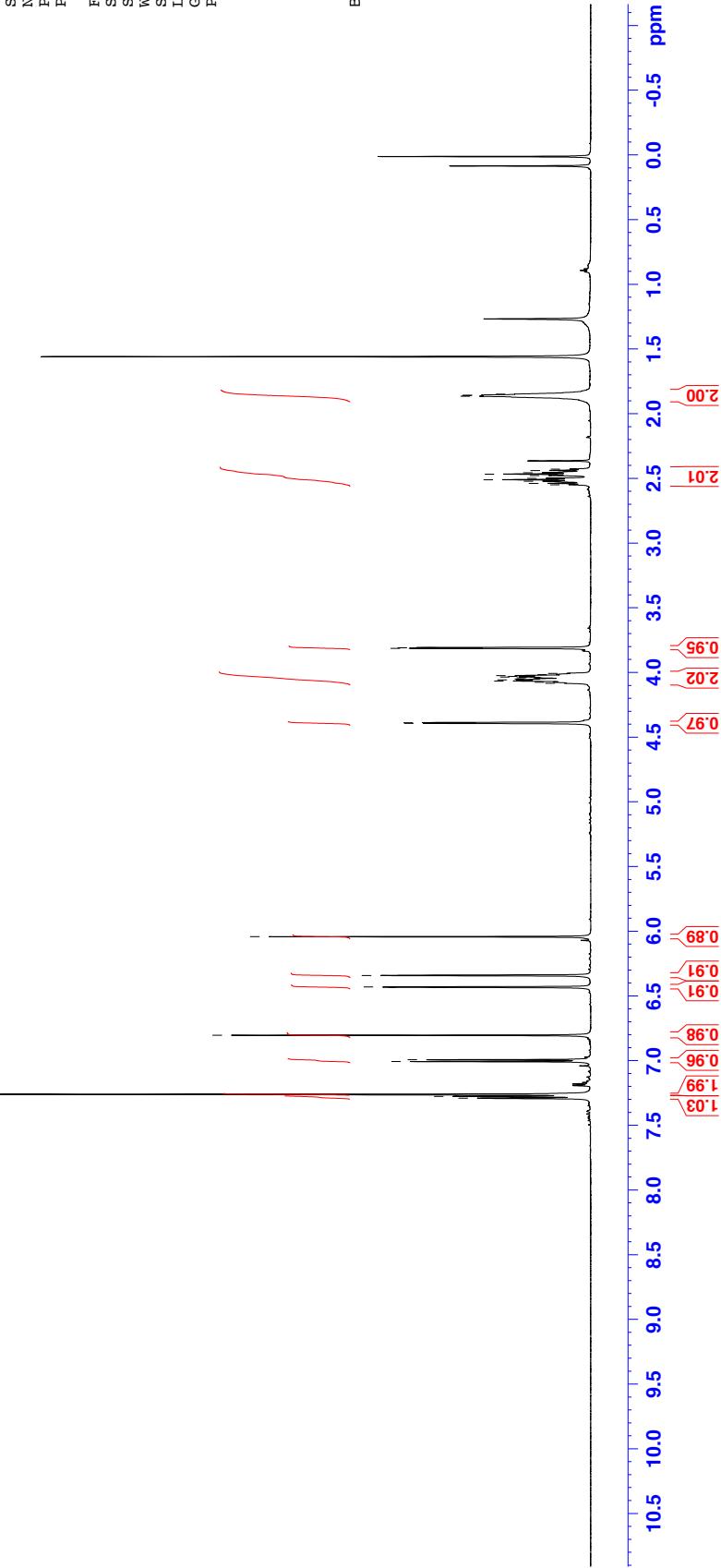
<sup>1</sup>H NMR of 31 (600MHz, CDCl<sub>3</sub>)



Current Data Parameters  
NAME VB-310-1B  
EXPNO 30  
PROCNO 1

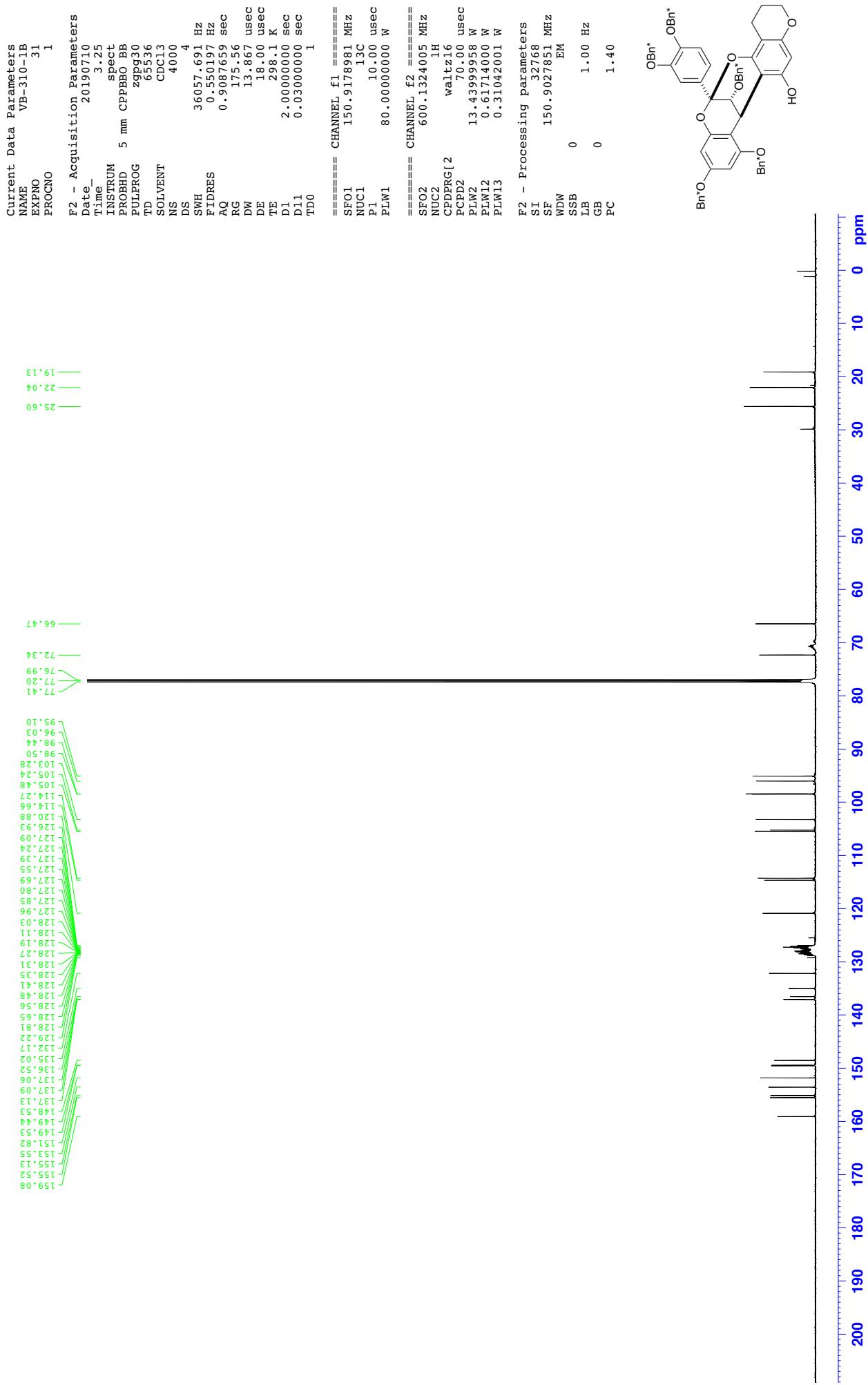
F2 - Acquisition Parameters  
Date 20190709  
Time 12.15  
INSTRUM spect  
PROBID 5 mm CPPBBO BB  
PULPROG zg30  
TD 65536  
SOLVENT CDC13  
NS 16  
DS 2  
SWH 12019.230 Hz  
FIDRES 0.1833399 Hz  
AQ 2.7262976 sec  
RG 31.94  
DW 41.600 usec  
DE 10.00 usec  
TE 298.1 K  
D1 1.0000000 sec  
TDO 1

===== CHANNEL f1 =====  
SFO1 600.1337060 MHz  
NUC1 1H  
P1 12.00 usec  
PLW1 21.00000000 W  
  
F2 - Processing parameters  
SI 65536  
SF 600.1300148 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

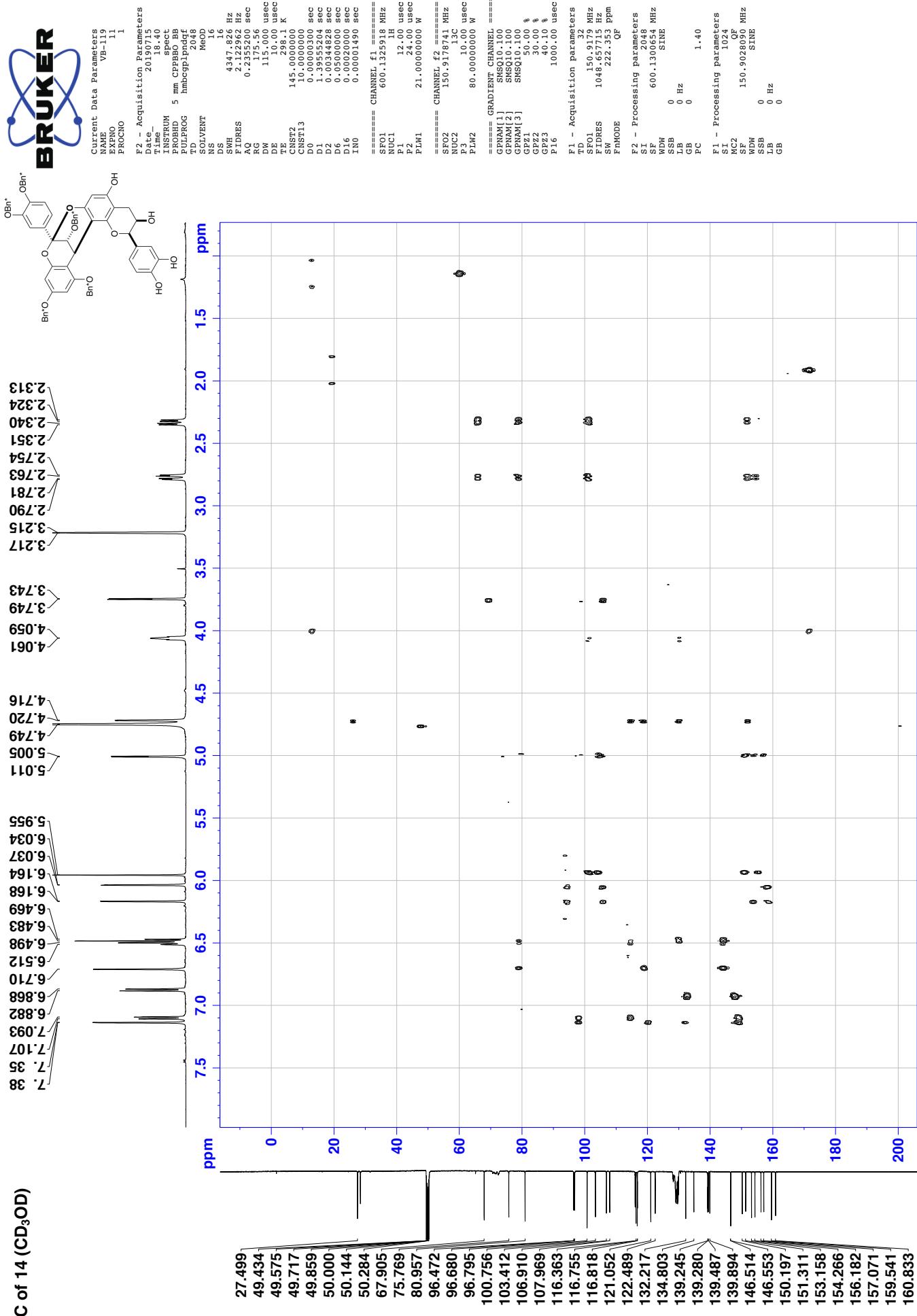




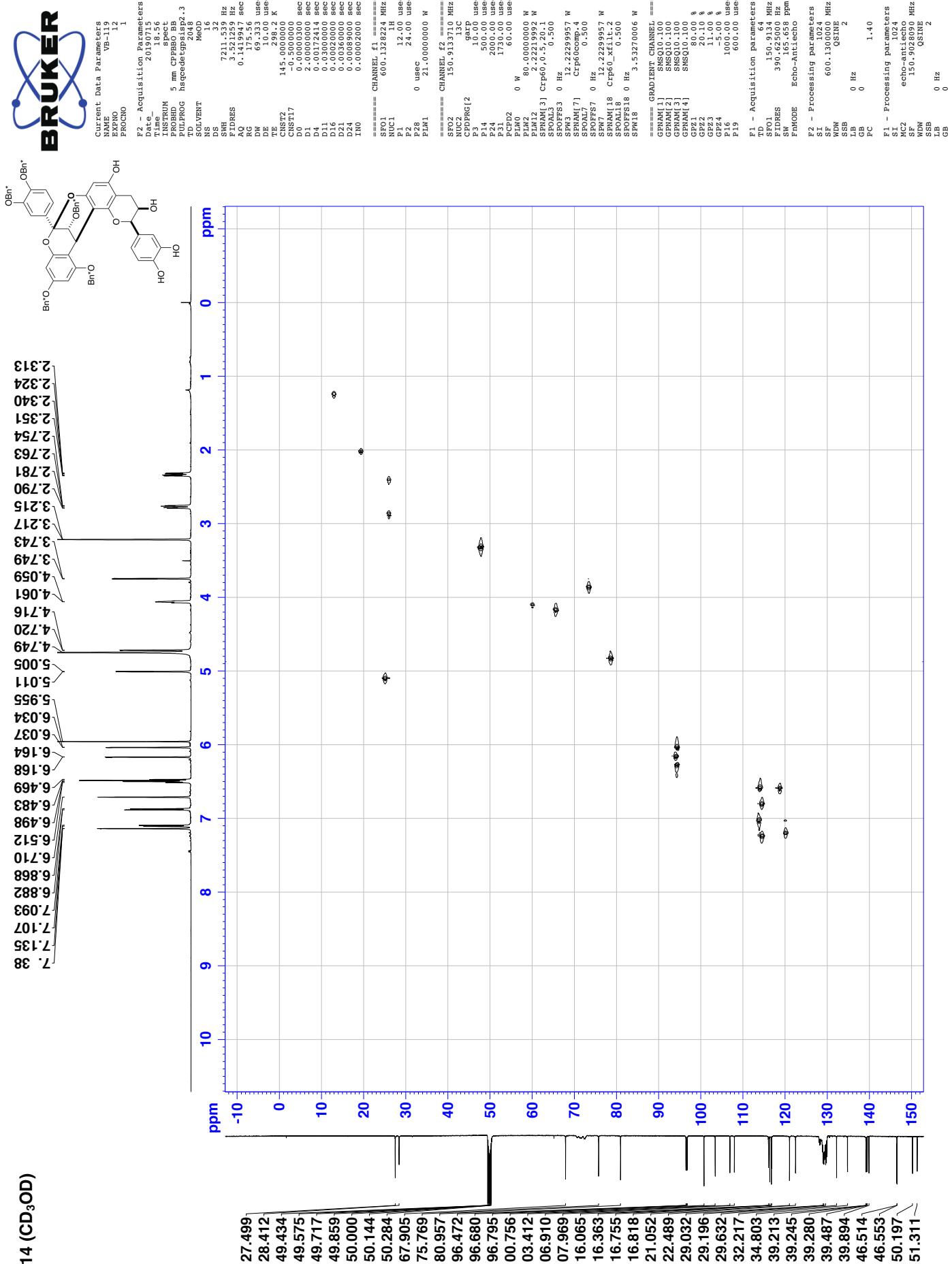
<sup>13</sup>C NMR of 31 (150MHz, CDCl<sub>3</sub>)

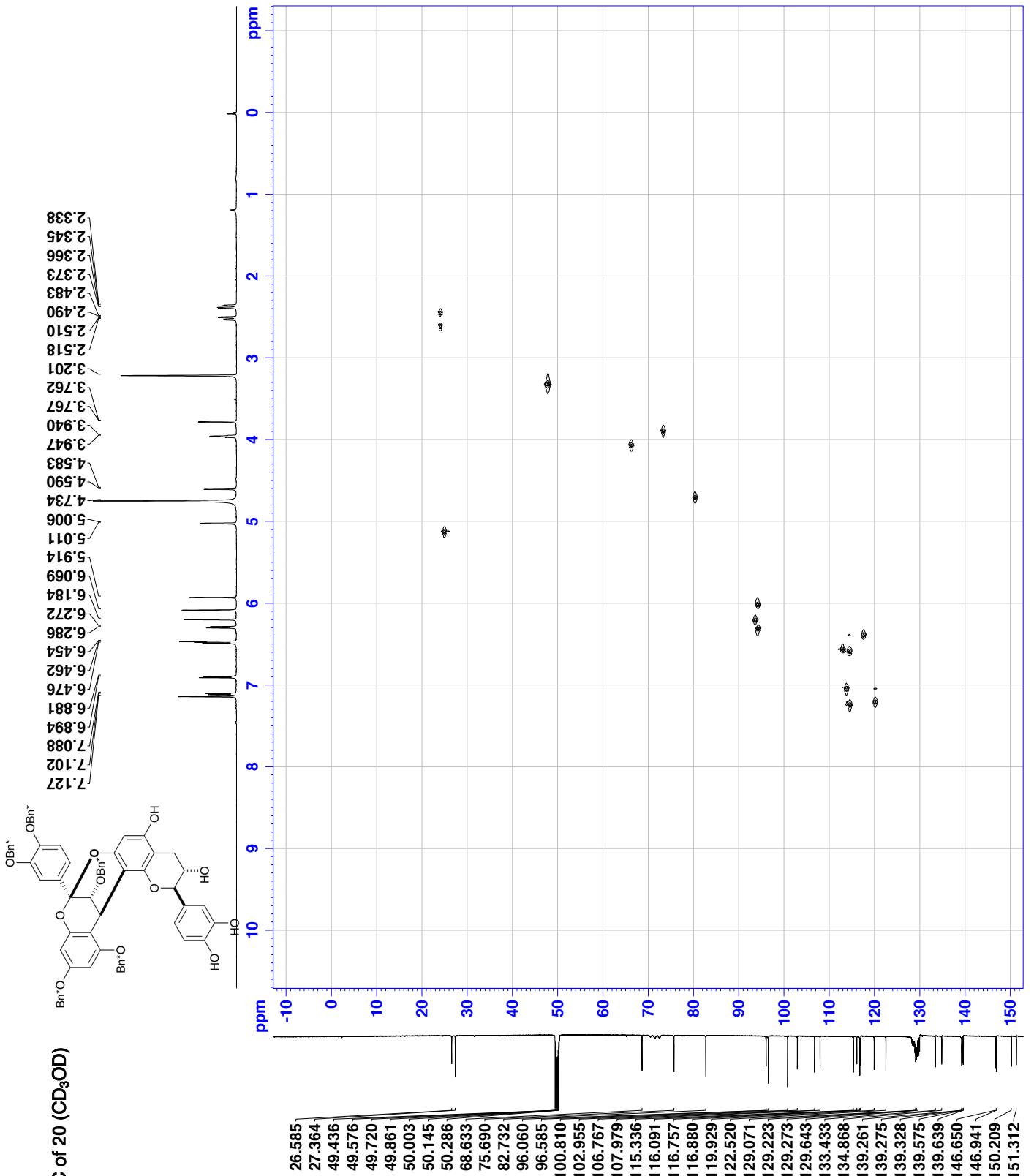


**HMBC of 14 ( $\text{CD}_3\text{OD}$ )**



HSQC of 14 ( $\text{CD}_3\text{OD}$ )

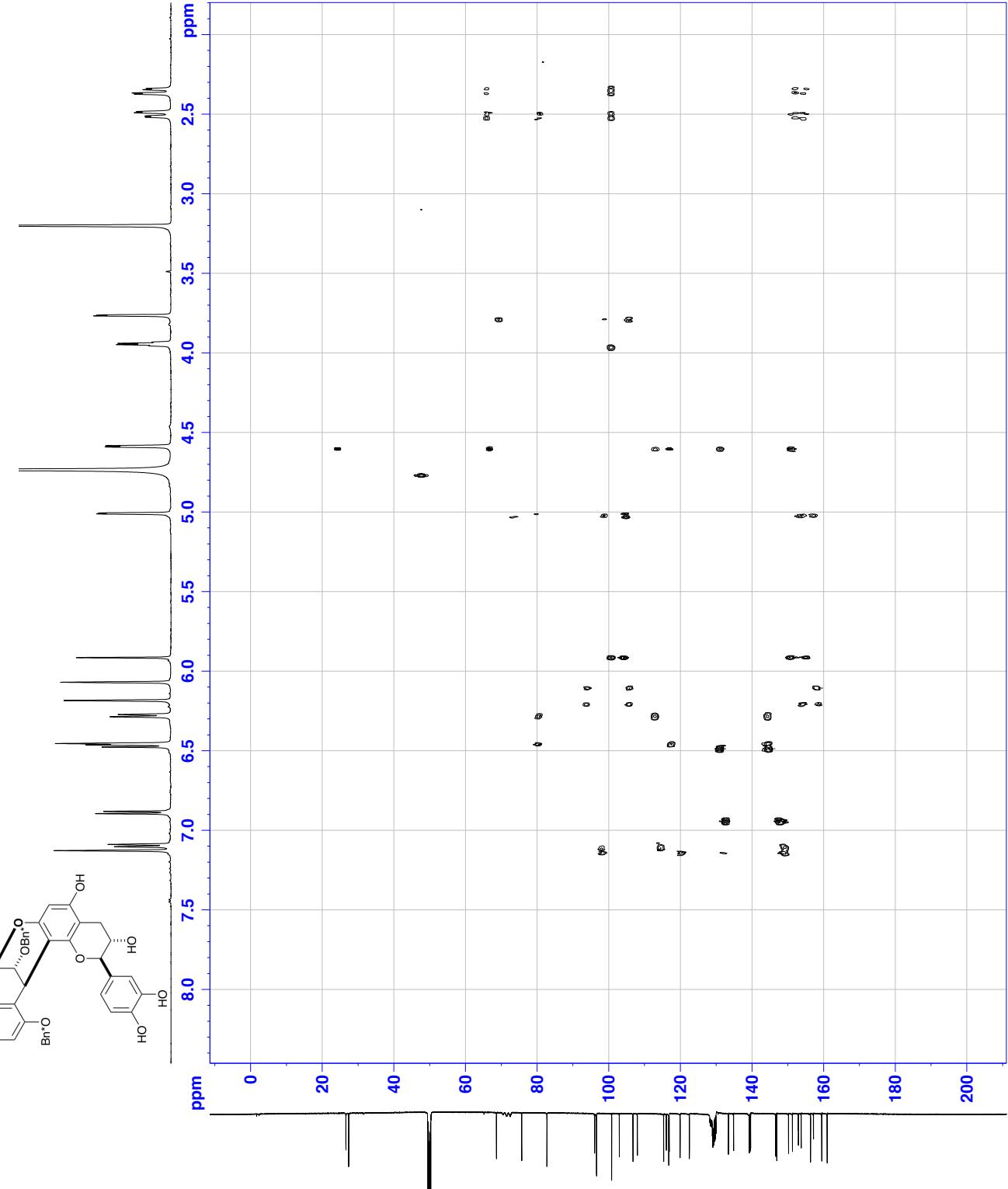




### HSQC of 20 (CD<sub>3</sub>OD)



**HMBC of 20 (CD<sub>3</sub>OD)**

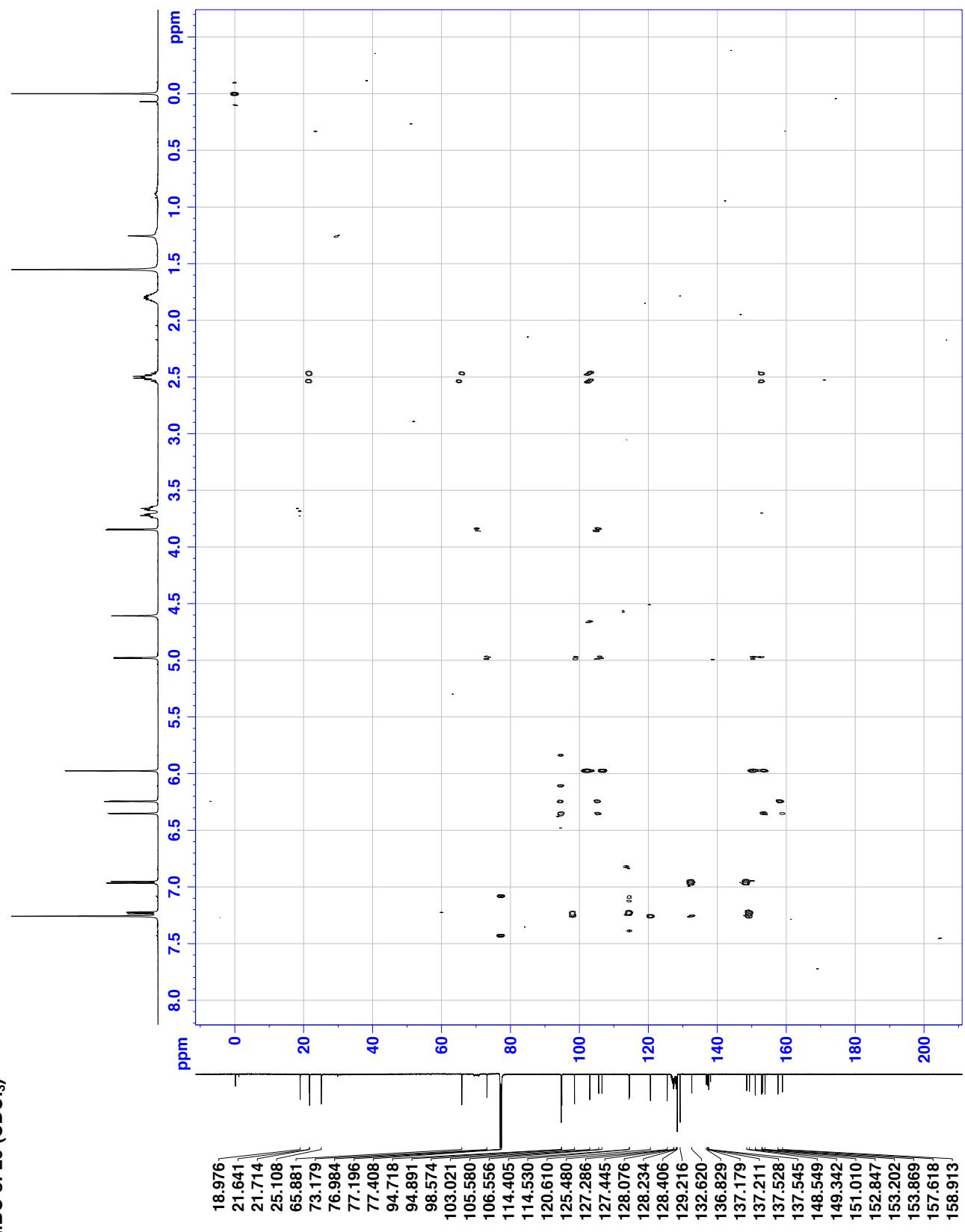


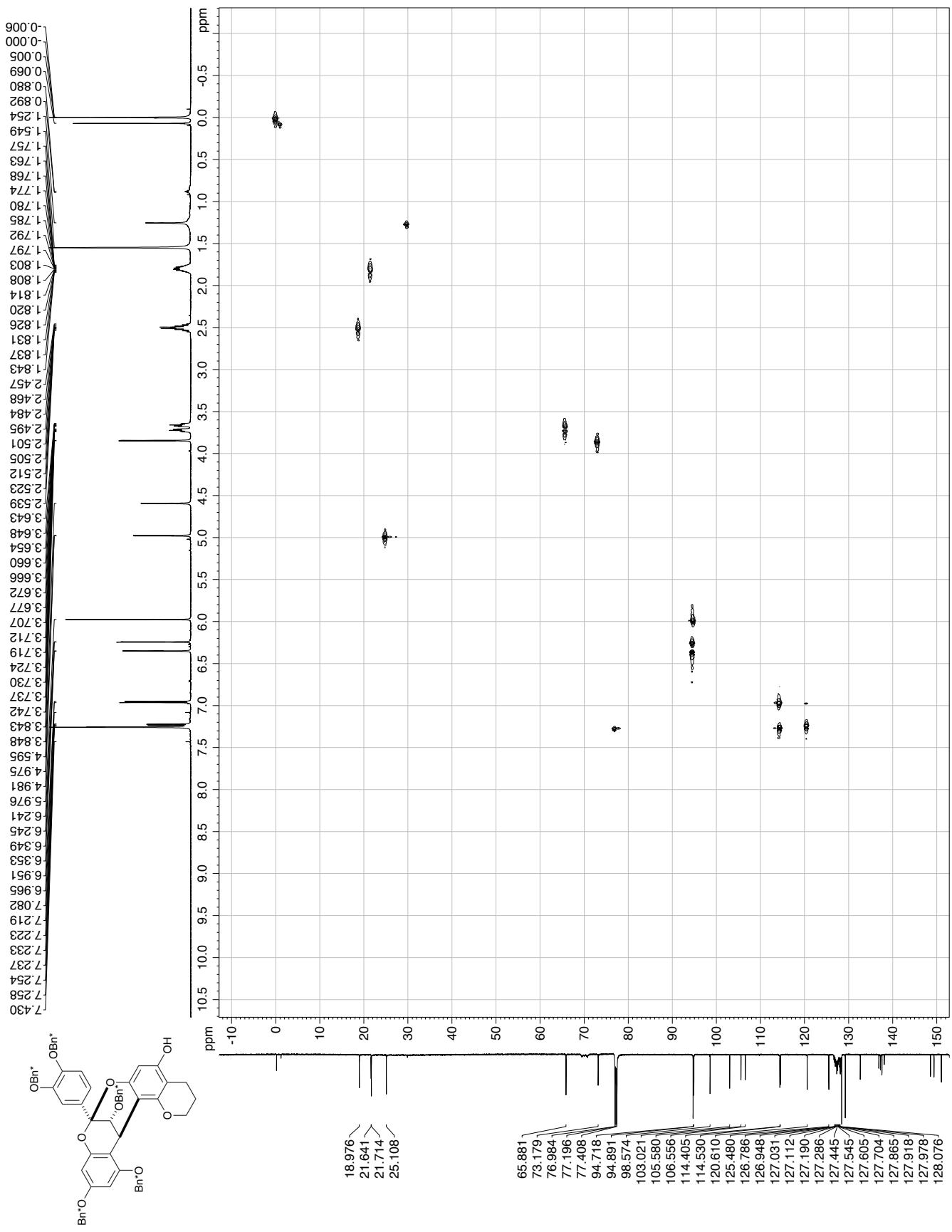
```

Current Data Parameters
NAME          VB-118
EXPNO         32
PROCNO        1
P1 - Acquisition parameters
Date-        2016/07/15
Time-        11:13:23
INSTRUM      spect
PROBHD      5 mm CPB60 BB
PULPROG     hmbcgrnddrf
TD            2048
SOLVENT       Meod
NS             16
DS            16
SWH           4000.000 Hz
FIDRES       1.953125 Hz
AQ            0.256000 sec
RG            175.56
DW            125.000 usec
DE            10.00 usec
TE            298.2 K
CPSV         CNST13
CNST         145.000000
D1            0.0000300 sec
D2            1.3750200 sec
D3            0.0344828 sec
D6            0.0000000 sec
T1            1.0000000 sec
IN0           0.00004450 sec
IM0           0.0000000 sec
SW0           4000.000 Hz
SF01          600.1331444 MHz
NUC1          NUC1
P1            12.00 usec
P2            24.00 usec
PLW01         21.0000000 W
=====
SFO2          150.9179741 MHz
NUC2          13C
P3            10.00 usec
PLW2          80.0000000 W
=====
P1 - Gradient CHANNEL =====
GPRA01[1]    SWSQ10.100
GPRA01[2]    SWSQ10.100
GPRA01[3]    SWSQ10.100
GPZ1          35.00 %
GPZ2          40.00 %
GPZ3          100.00 usec
P1_6          1000.00 usec
F1 - Acquisition parameters
TD            32
SFO1          104.8657715 Hz
FIDRES       222.333 ppm
F1NODF       QF
F2 - Processing parameters
SI            2048
SF            600.1300632 MHz
WDW           SINE
SSB           0
LB            0 Hz
GB            0
PC            1.40
P1 - Processing parameters
SI            1024
MC2           104
SF            150.3920800 MHz
WDW           SINE
SSB           0 Hz
LB            0
GB            0

```

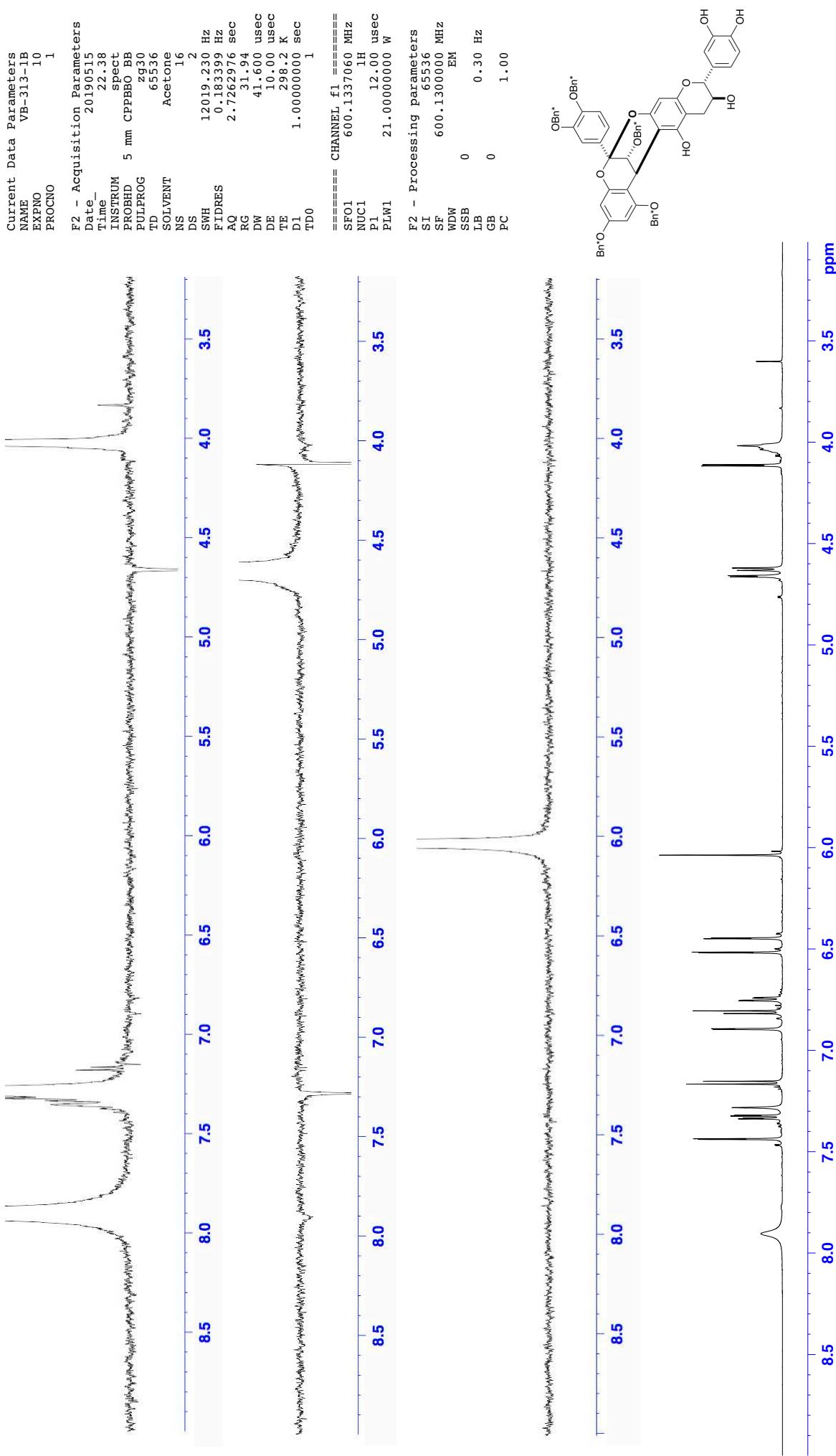
**HMBC of 20 (CD<sub>3</sub>OD)**

HMBC of 29 (CDCl<sub>3</sub>)

**HSQC of 29 (CDCl<sub>3</sub>)**










Current Data Parameters  
 NAME VB-313-IA  
 EXPNO 20  
 PROCN0 1

F2 - Acquisition Parameters  
 Date 20190517  
 Time 19.22  
 INSTRUM spect  
 PROBID 5 mm CPPBBO BB  
 PULPROG zg30  
 TD 65536  
 SOLVENT Acetone  
 NS 16  
 DS 2  
 SWH 12019.230 Hz  
 FIDRES 0.183399 Hz  
 FIDRES 2.7622976 sec  
 AQ 18.96  
 RG 41.600 usec  
 DW 10.00 usec  
 DE 298.2 K  
 D1 1.0000000 sec  
 TDO 1

===== CHANNEL f1 =====  
 SFO1 600.1337060 MHz  
 NUC1 1H  
 P1 12.00 usec  
 PLW1 21.00000000 W

F2 - Processing parameters  
 SI 65536  
 SF 600.1300000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

