

Electronic Supplementary Information (ESI)

Novel carbocyclic scaffolds with a 6,5+5,5 ring system, named nor-allodammarane, from the enzymatic reactions of 27-nor-(oxido)squalenes with hopene synthase

Chiaki Nakano, Takumi Watanabe, Mai Minamino and Tsutomu Hoshino*

Department of Applied Biological Chemistry, Faculty of Agriculture, and Graduate School of Science and Technology, Niigata University, Ikarashi 2-8050, Nishi-ku, Niigata, Japan, 950-2181

*Corresponding author: hoshitsu@agr.niigata-u.ac.jp

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1. Enzymatic reaction of racemic mixture of (3*R*, 5*S*)-27-nor-2,3-oxidosqualenes (27-norOXSQ, 18 and 19) with the native SHC.

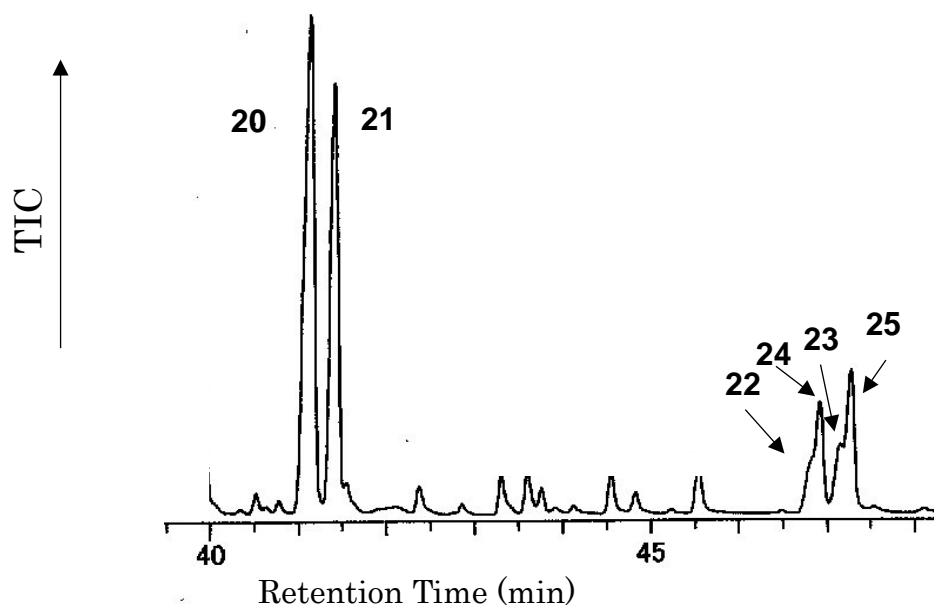
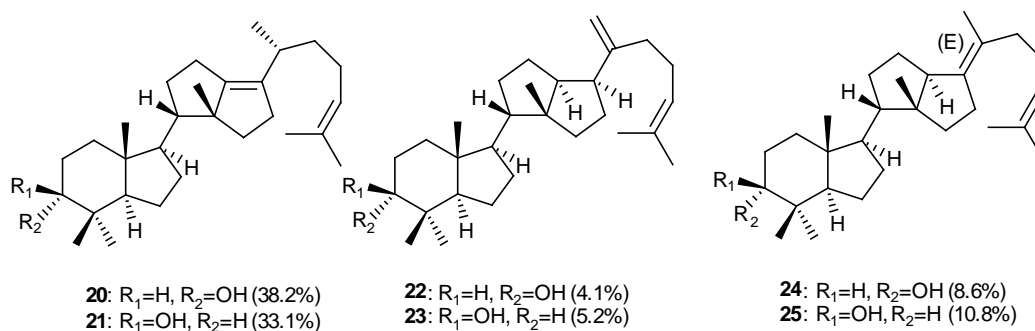


Fig. S1.1. GCMS profile (TIC) of Products 20–25 from 18 and 19

GC-TOFF MS conditions: Inject temp : 280°C; Column temp: 150-280°C (3°C /min); Flow : 1.0 mL/min

Product structures are shown below.



Compound names are as follows: **20**, 3 α -hydroxy-(8*R*,10*S*,14*R*,20*R*)-30-nor-allodammara-13(17),24-diene; **21**, 3 β -hydroxy-(8*R*,10*S*,14*R*,20*R*)-30-nor-allodammara-13(17),24-diene; **22**, 3 α -hydroxy-(8*R*,10*S*,13*S*,14*S*,17*S*)-30-nor-allodammara-20(21),24-diene; **23**, 3 β -hydroxy-(8*R*,10*S*,13*S*,14*S*,17*S*)-30-nor-allodammara-20(21),24-diene; **24**, 3 α -hydroxy-(8*R*,10*S*,13*R*,14*R*)-30-nor-allodammara-(*E*)-17(20),24-diene; and **25**, 3 β -hydroxy-(8*R*,10*S*,13*R*,14*R*)-30-nor-allodammara-(*E*)-17(20),24-diene.

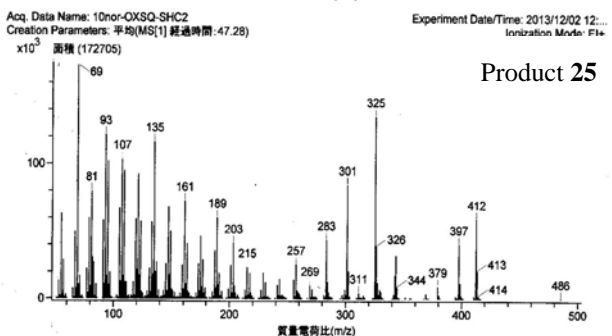
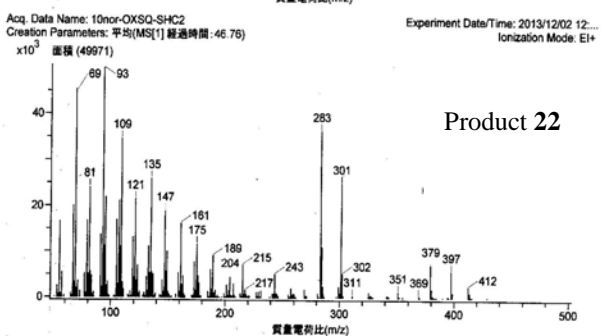
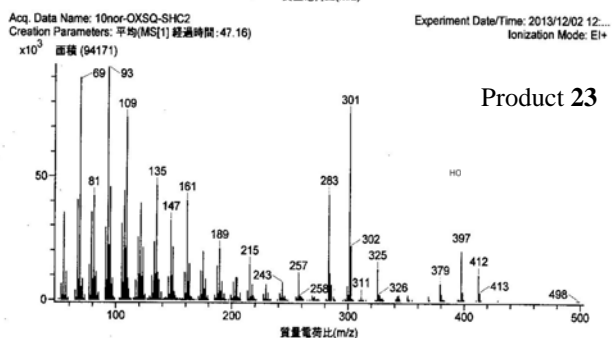
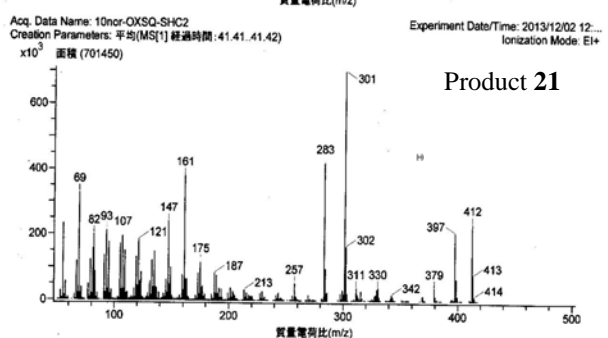
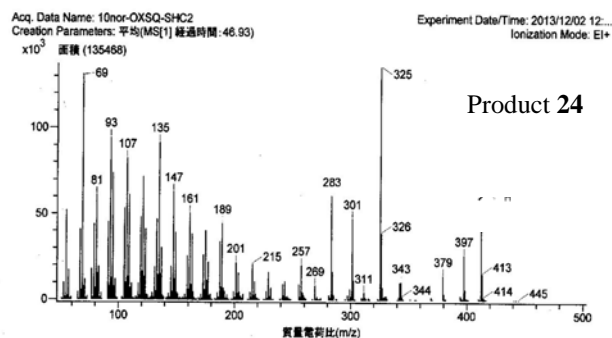
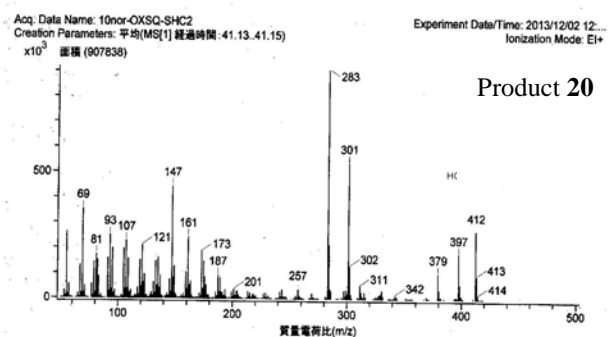


Fig. S1.2. EI-MS spectra of products 20-25

2. Enzymatic reaction of 13a(b) with SHC.

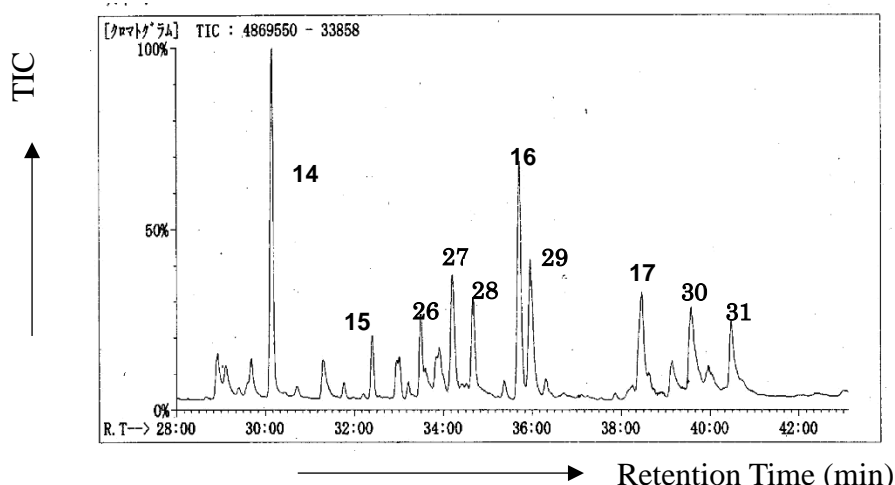
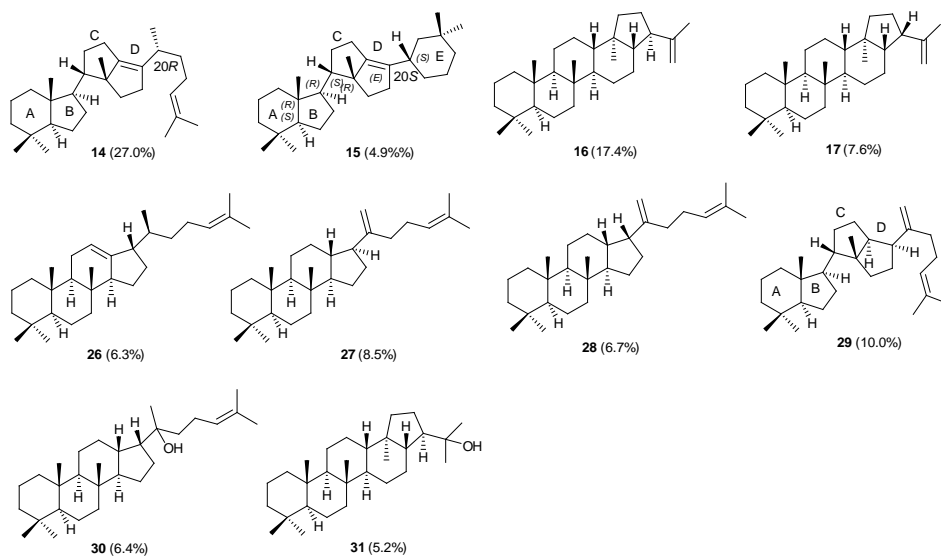


Fig. S2. GC-MS trace (TIC) of the hexane-extract of the incubation mixture of **13a(13b)**.

GC-Q-MS conditions: Inject temp : 280°C; Column temp: 150-280°C (3°C /min); Flow : 1.0 mL/min.

Structures of products **14**, **15**, **16** and **17** were reported in our previous paper (T. Hoshino and S. Ohashi, *Org. Let.*, **2002**, *4*, 2553-2556). However, the stereochemistry at C-20 of **14** and **15** has remained ambiguous. In addition to **14** and **15**, many enzymatic products **26–31** were newly found from the incubation mixture of **13a** and **13b** with the native SHC. Notably, product **29**, which is a key intermediate for predicting the configurations at C-20 of **14** and **15**, was detected in a substantial amount. Product structures are shown below.



Compound names are as follows: **14**, (8*R*,10*S*,14*R*,20*R*)-30-nor-allodammara-13(17),24-diene; **15**, (8*R*,10*S*,14*R*,20*S*)-30-nor-allogammacer-13(17)-ene; **16**, 27-nor-isohopene (21-*epi*-hop-22(29)-ene); **17**, 27-nor-hopene; **26**, (20*S*)-30-nor-isodammara-12(13),24-diene (30-nor-17-*epi*-dammara-12(13),24-diene); **27**, 30-nor-dammara-20(21),24-diene; **28**, 30-nor-isodammara-20(21),24-diene (30-nor-17-*epi*-dammara-20(21),24-diene); **29**, (8*R*,10*S*, 13*S*, 14*S*, 17*S*)-30-nor-allodammara-20(21),24-diene; **30**, 30-nor-20-hydroxy-isodammara-24-ene (17-*epi*-20-hydroxy-dammarene); and **31**, 27-nor-isohopanol (21-*epi*-27-norhopanol).

3. NMR spectra of product **21** in CDCl₃.

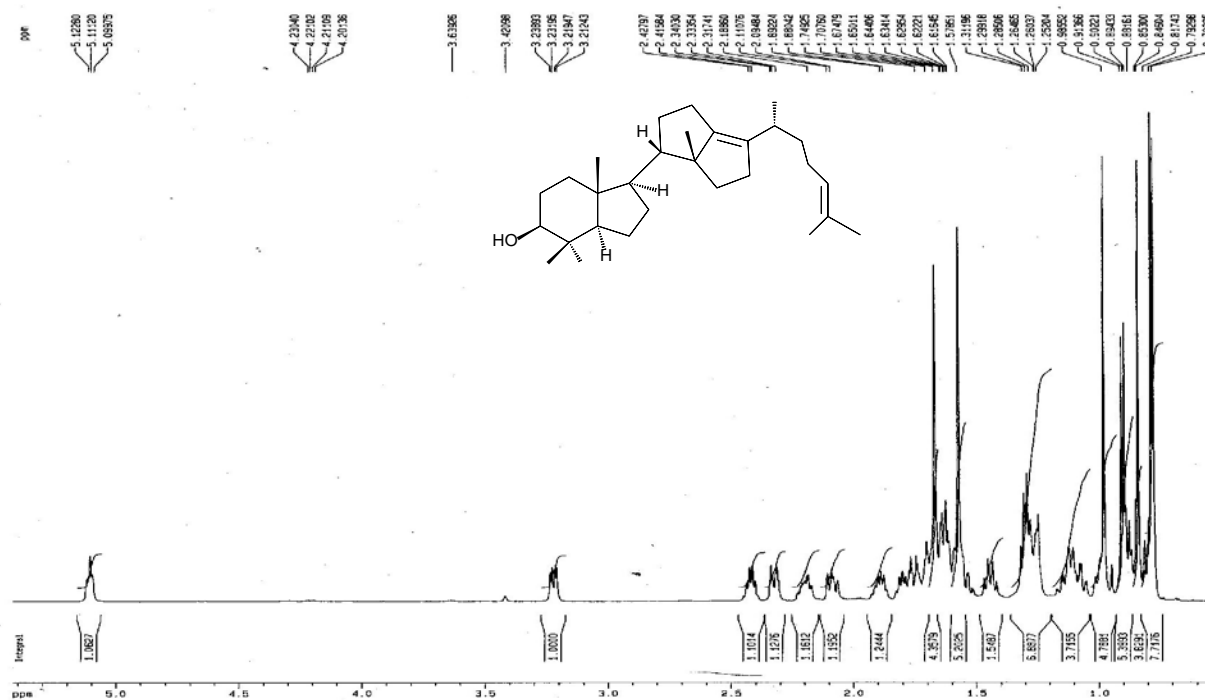


Fig. S3.1. ¹H-NMR spectrum of product **21** in CDCl₃ (600 MHz).

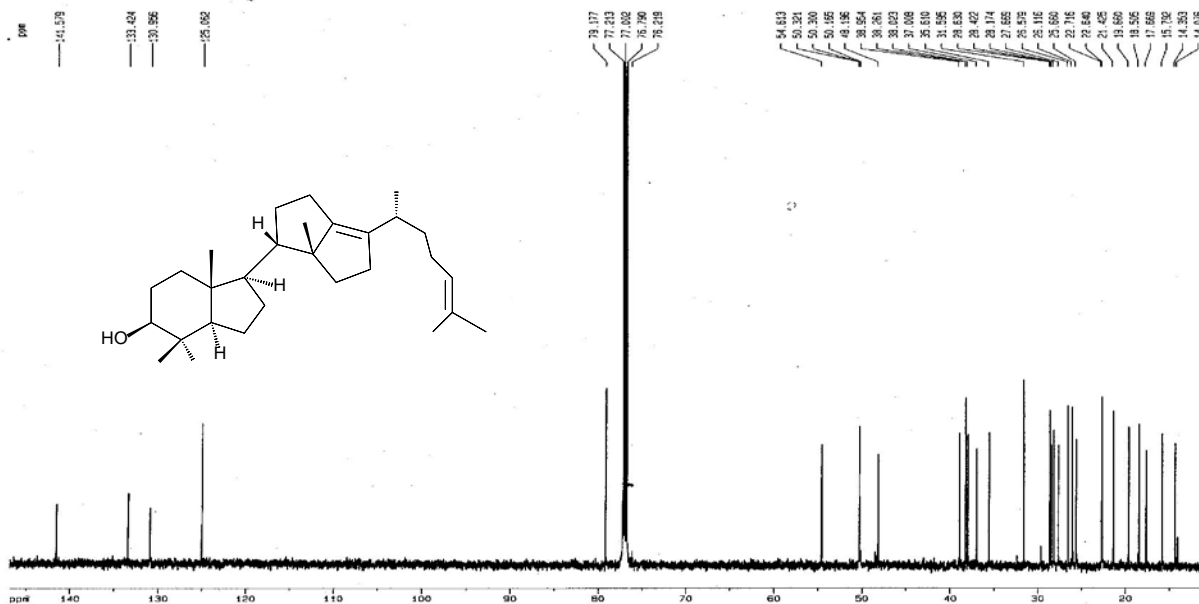


Fig. S3.2. ¹³C-NMR spectrum of product **21** in CDCl₃ (600 MHz) (150 MHz).

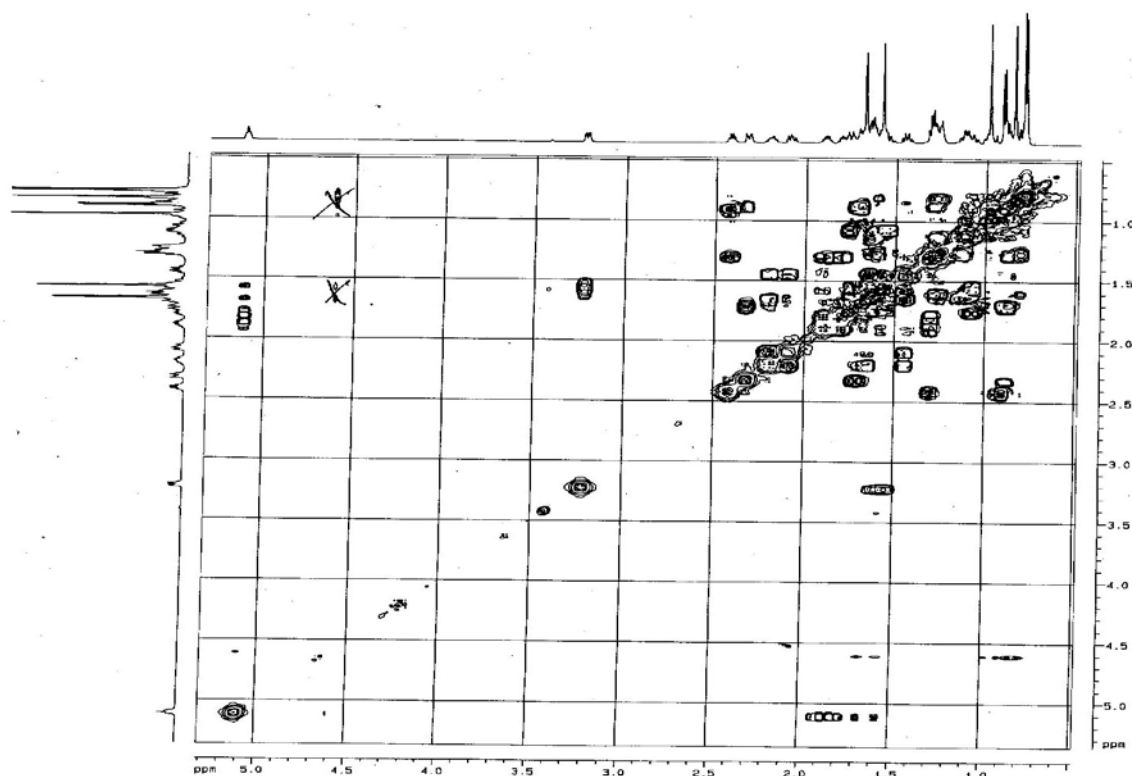


Fig. S3.3. ^1H - ^1H COSY spectrum of product **21** in CDCl_3 .

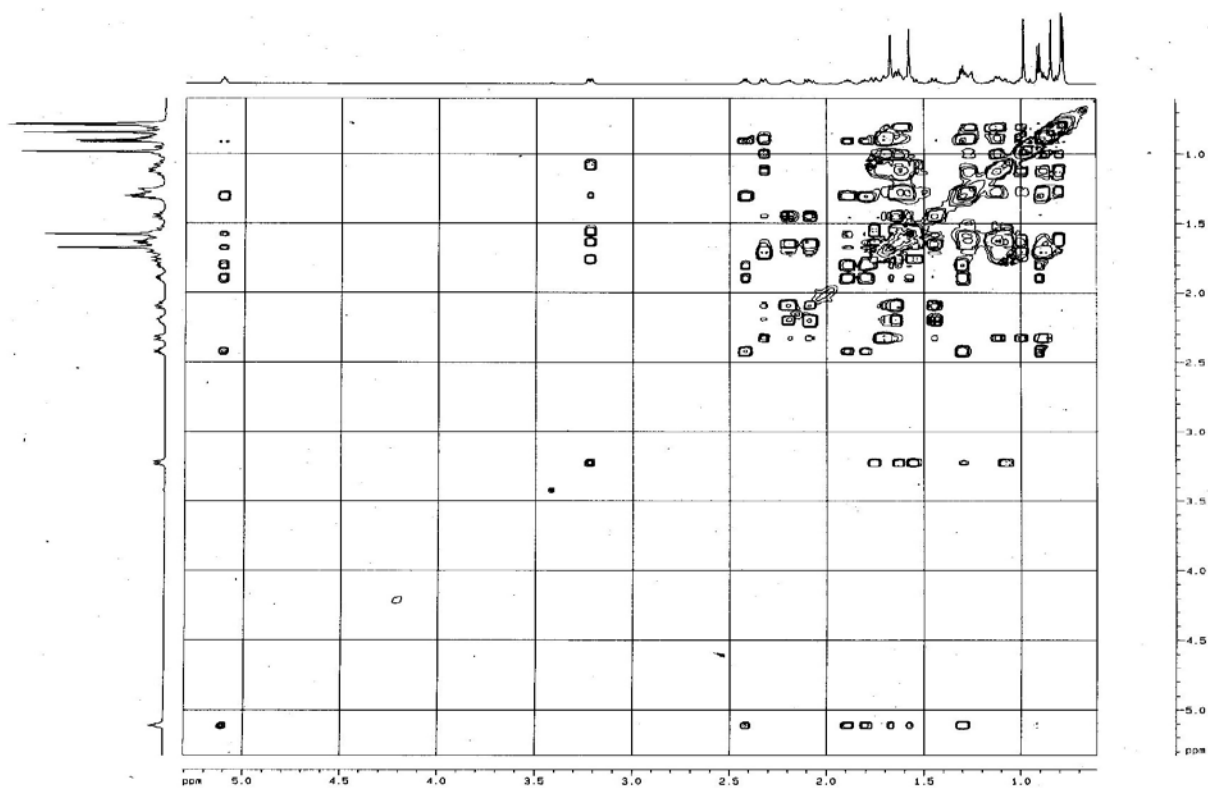


Fig. S3.4. TOCSY spectrum of product **21** in CDCl_3 .

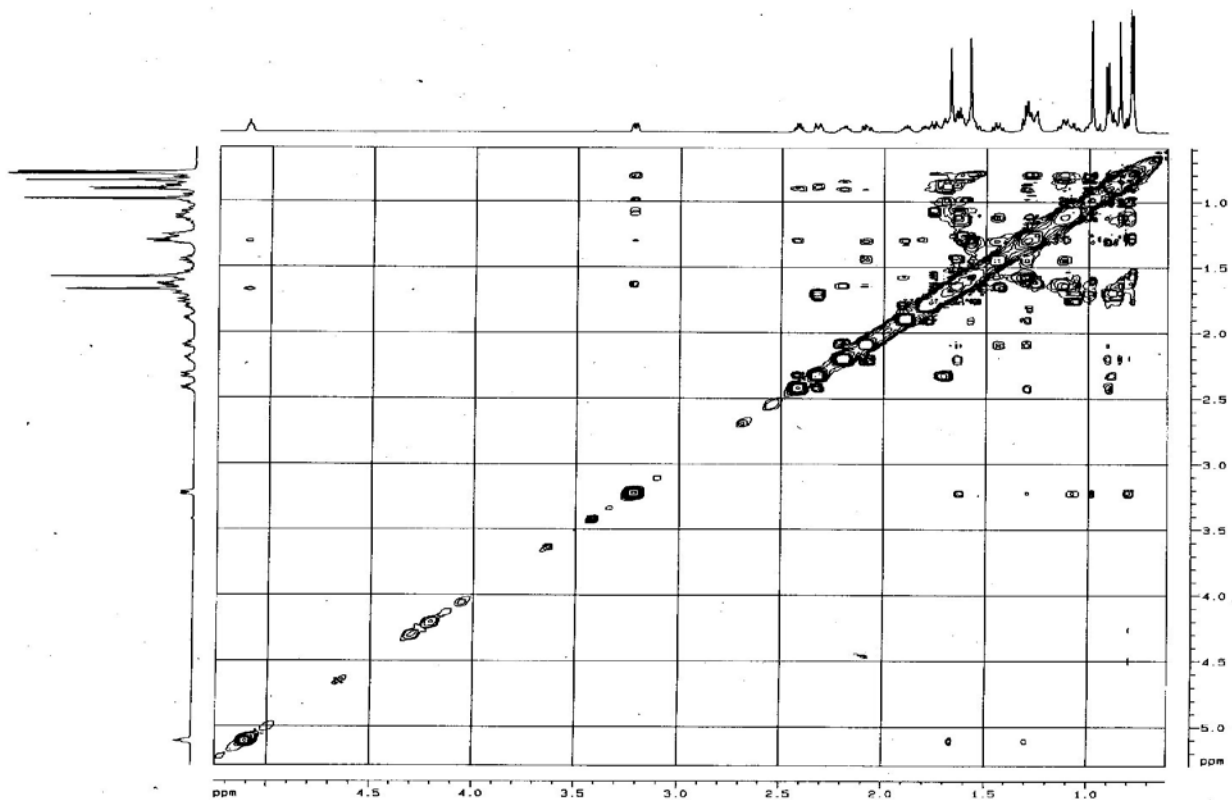


Fig. S3.5. NOESY spectrum of product **21** in CDCl_3 .

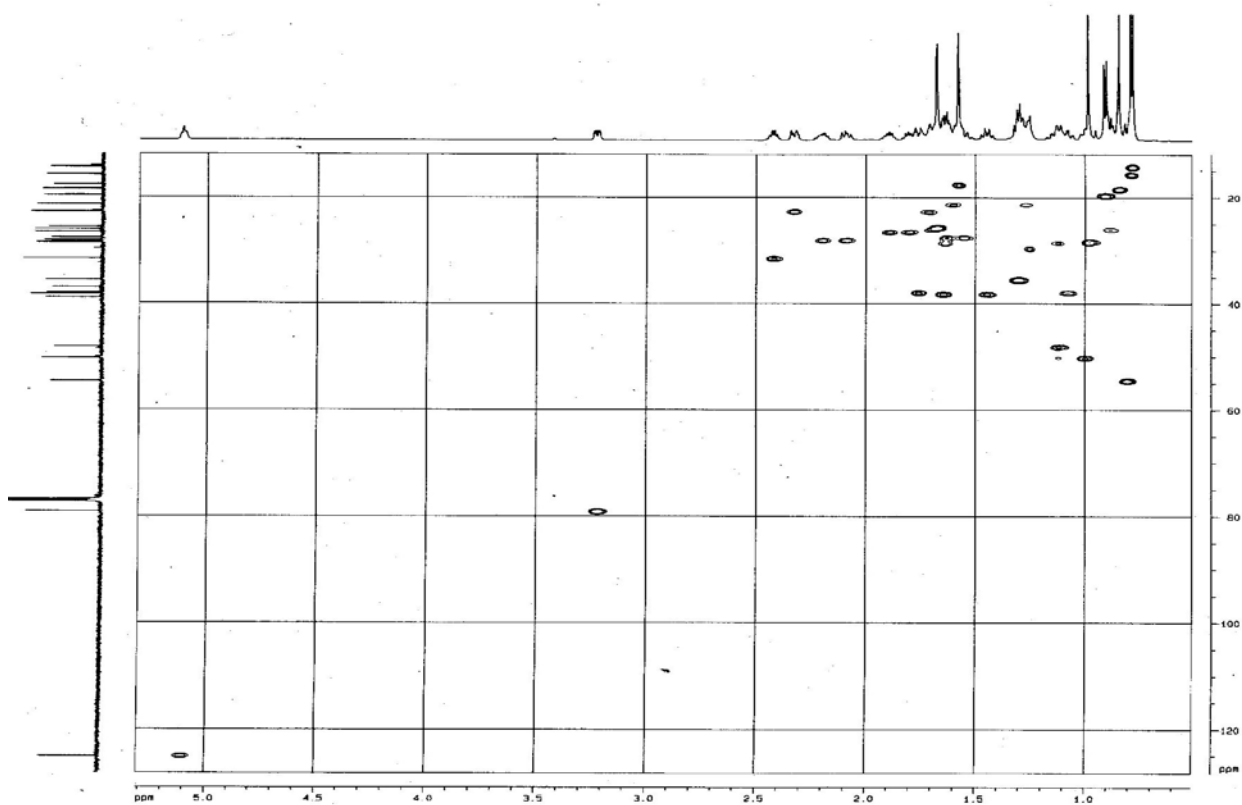


Fig. S3.6. HSQC spectrum of product **21** in CDCl_3 .

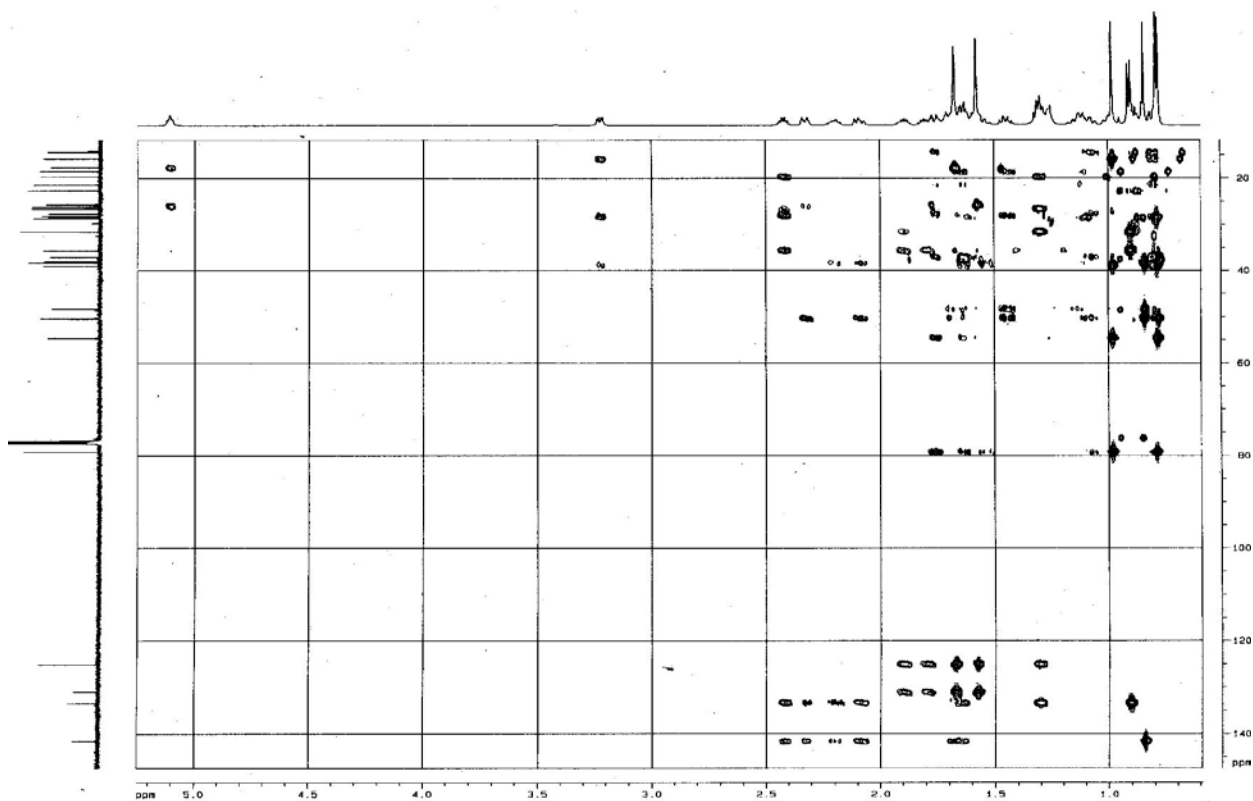
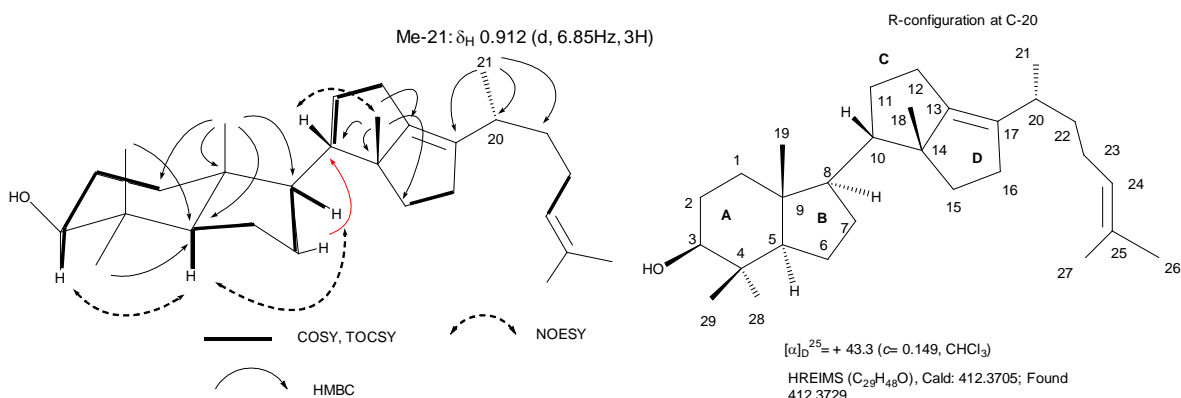


Fig. S3.7. HMBC spectrum of product **21** in CDCl_3 .

Product 21 from (3S)-27-norOXSQ 21



600 MHz, in CDCl_3 ; the residual solvent peak: δ_{H} 7.26; δ_{C} 77.0 ppm

NO.	^1H	^{13}C	NO.	^1H	^{13}C	NO.	^1H	^{13}C	NO.	^1H	^{13}C
1	1.07(m,1H);1.77(m,1H)	38.02(t)	9	—	37.01(s)	17	—	133.4(s)	25	—	130.9(s)
2	1.55(m);1.62(m)	27.67(t)	10	1.12(m,1H)	48.20(d)	18	0.846(s,3H)	18.50(q)	26	1.675(s,3H)	22.72(q)
3	3.23(dd, J=11.7 Hz;4.2 Hz, 1H)	79.18(d)	11	0.90(m,1H); 1.70(m,1H)	26.11(t)	19	0.783(s,3H)	14.35(q)	27	1.578 (s, 3H)	17.67(q)
4	—	38.95(s)	12	1.70(m,1H); 2.32(m,1H)	22.71(t)	20	2.42(m, 1H)	31.59(d)	28	0.985(s,3H)	28.42(q)
5	0.81(m, 1H)	54.61(d)	13	—	141.6(s)	21	0.912(d, 6.85Hz, 3H)	19.66(q)	29	0.793(s,3H)	15.79(q)
6	1.26(m,1H); 1.60(m, 1H)	21.42(t)	14	—	50.32 (s)	22	1.31 (m, 2H)	35.61(t)			
7	1.12(m,1H); 1.63(m,1H)	28.63(t)	15	1.45(m,1H); 1.63(m, 1H)	38.26(t)	23	1.80(m,1H); 1.89(m,1H)	26.58(t)			
8	1.00 (m, 1H)	50.30(d)	16	2.08(m,1H); 2.19(m,1H)	28.17(t)	24	5.11(brs, 1H)	125.1(d)			

Fig. S3.8. Assignments of NMR data of product **21** in CDCl_3 , HR-EIMS and optical rotation.

4. NMR spectra of product 21 in C₆D₆.

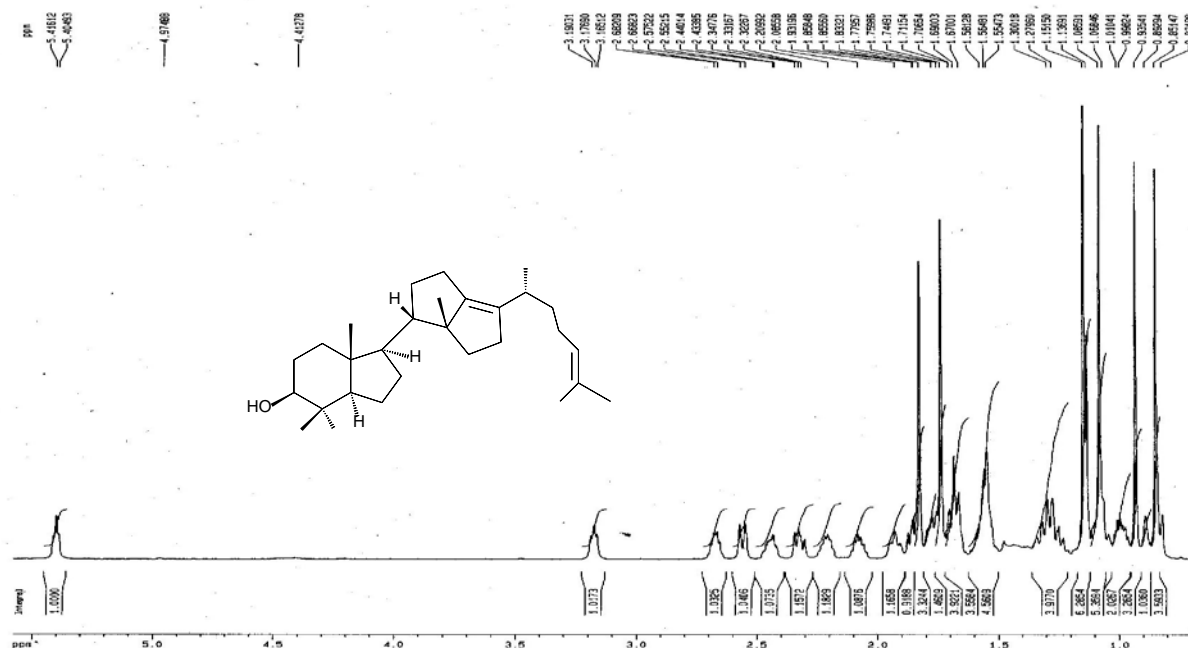


Fig. S4.1. ¹H NMR spectra of product **21** in C₆D₆ (600 MHz).

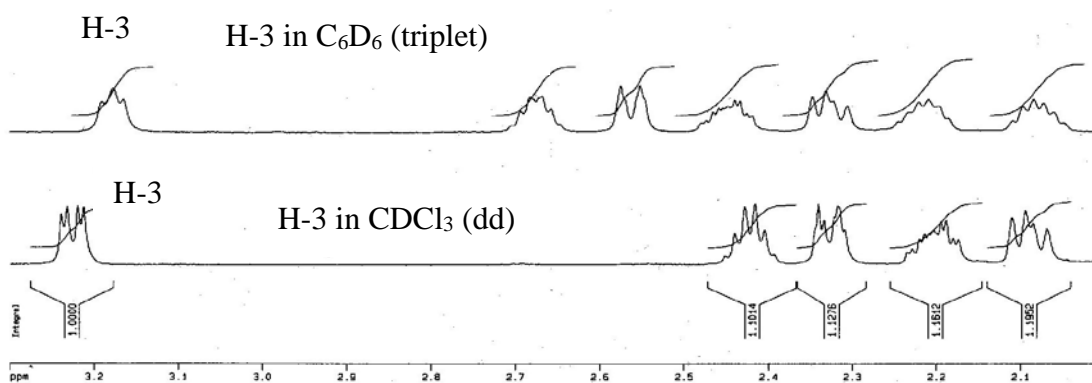


Fig. S4.2. Comparison of ¹H NMR spectra of product **21** between CDCl₃ and C₆D₆ solutions (600 MHz).

The coupling constants of H-3 were compared between the solutions of C₆D₆ and CDCl₃. δ_{H} in C₆D₆: 3.18 (br t, $J=8.0$ Hz); δ_{H} in CDCl₃: 3.23 ((dd, $J=11.7$ Hz and 4.2 Hz). The coupling constants (splitting pattern) in CDCl₃ solution clearly demonstrated that the OH group is β -oriented, although the splitting pattern (triplet) in C₆D₆ suggests that the OH group is α -oriented. This contradictory argument was resolved by the definitive NOE between H-3 and H-5 in the C₆D₆ solution. Consequently, the OH at C-3 of product **1** was credibly determined to be β -oriented. The ¹H-NMR spectra of products **23** and **25** in C₆D₆ solution also showed triplet splitting pattern, but dd splitting in the CDCl₃ solution.

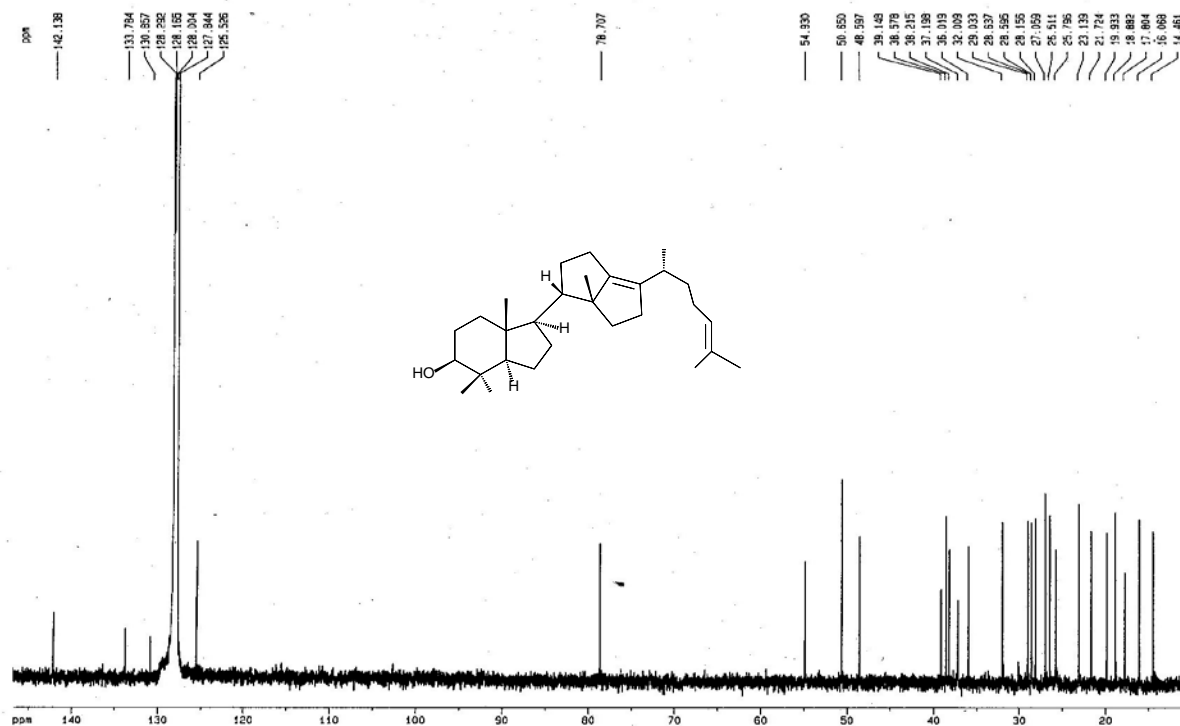


Fig. S4.3. ^{13}C NMR spectrum of product **21** in C_6D_6 (150 MHz).

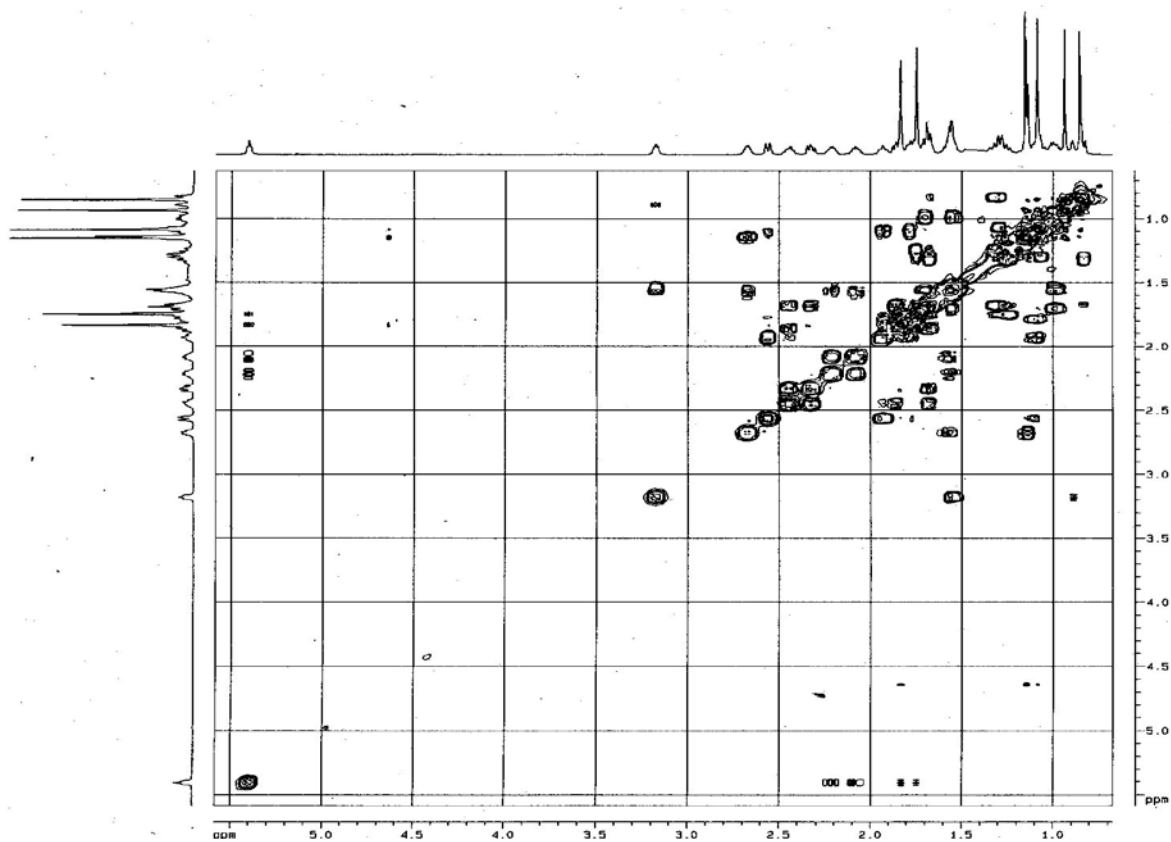


Fig. S4.4. ^1H - ^1H COSY spectrum of product **21** in C_6D_6 .

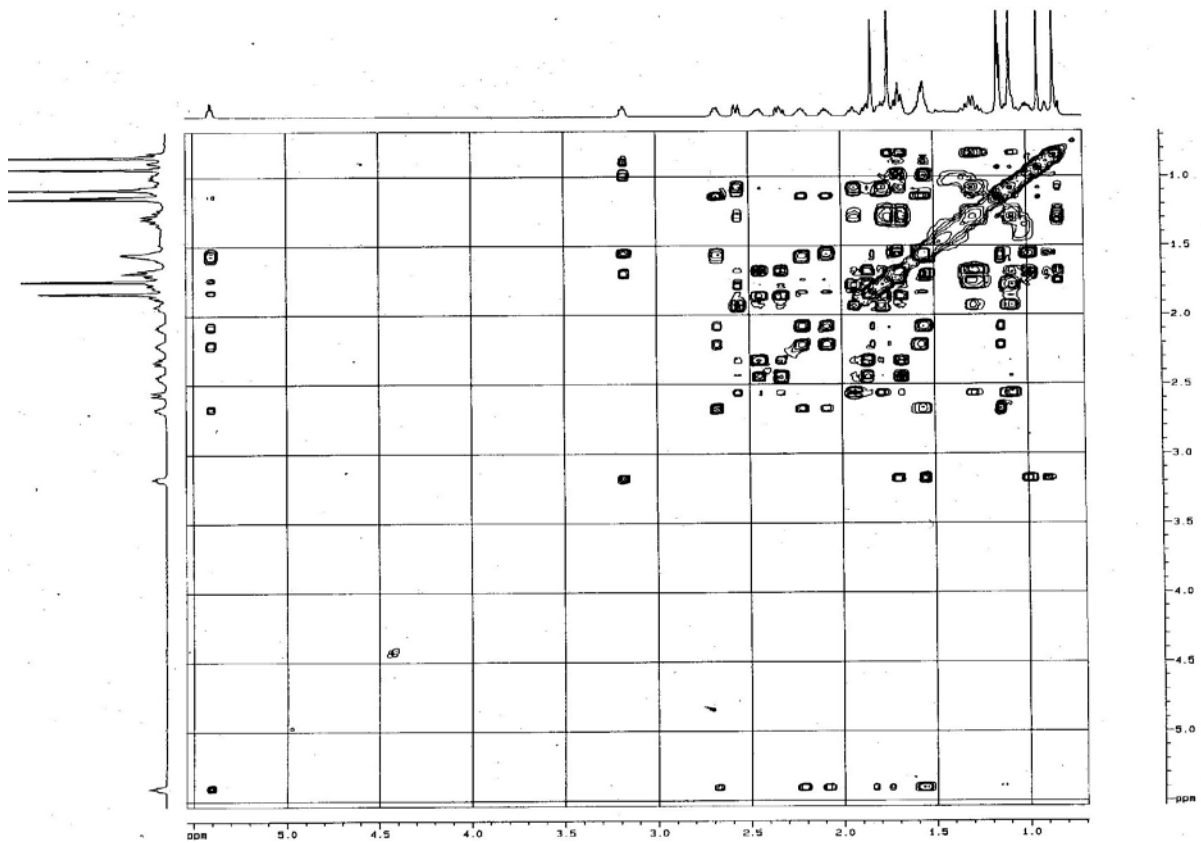


Fig. S4.5. TOCSY spectrum of product **21** in C_6D_6 .

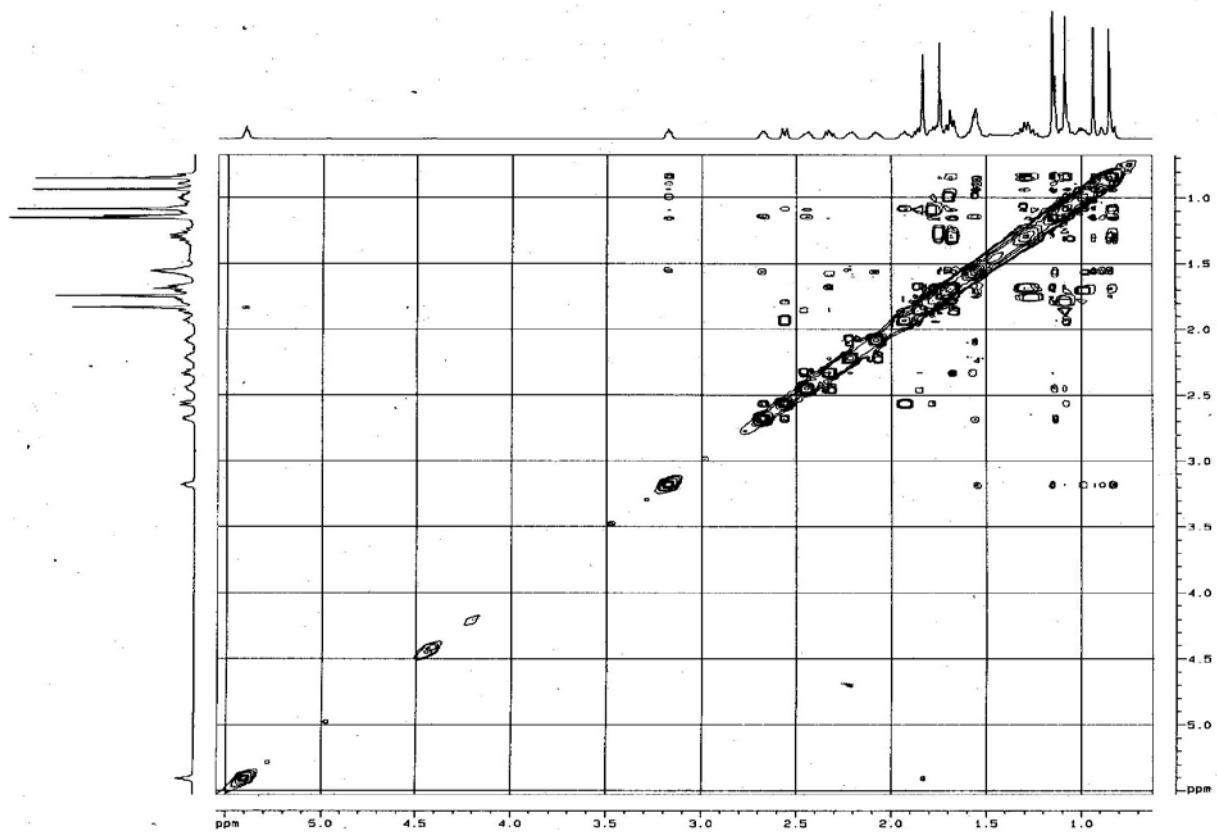


Fig. S4.6. NOESY spectrum of product **21** in C_6D_6 .

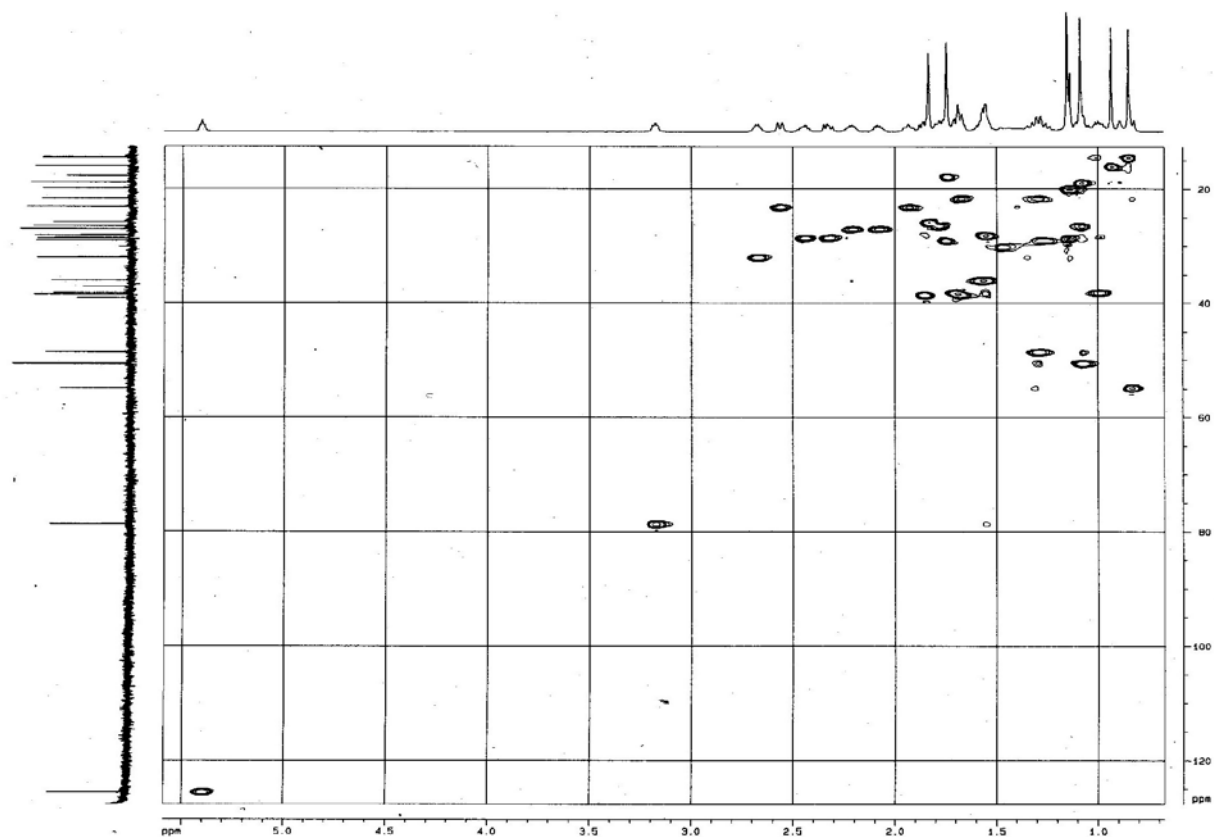


Fig. S4.7. HSQC spectrum of product **21** in C₆D₆.

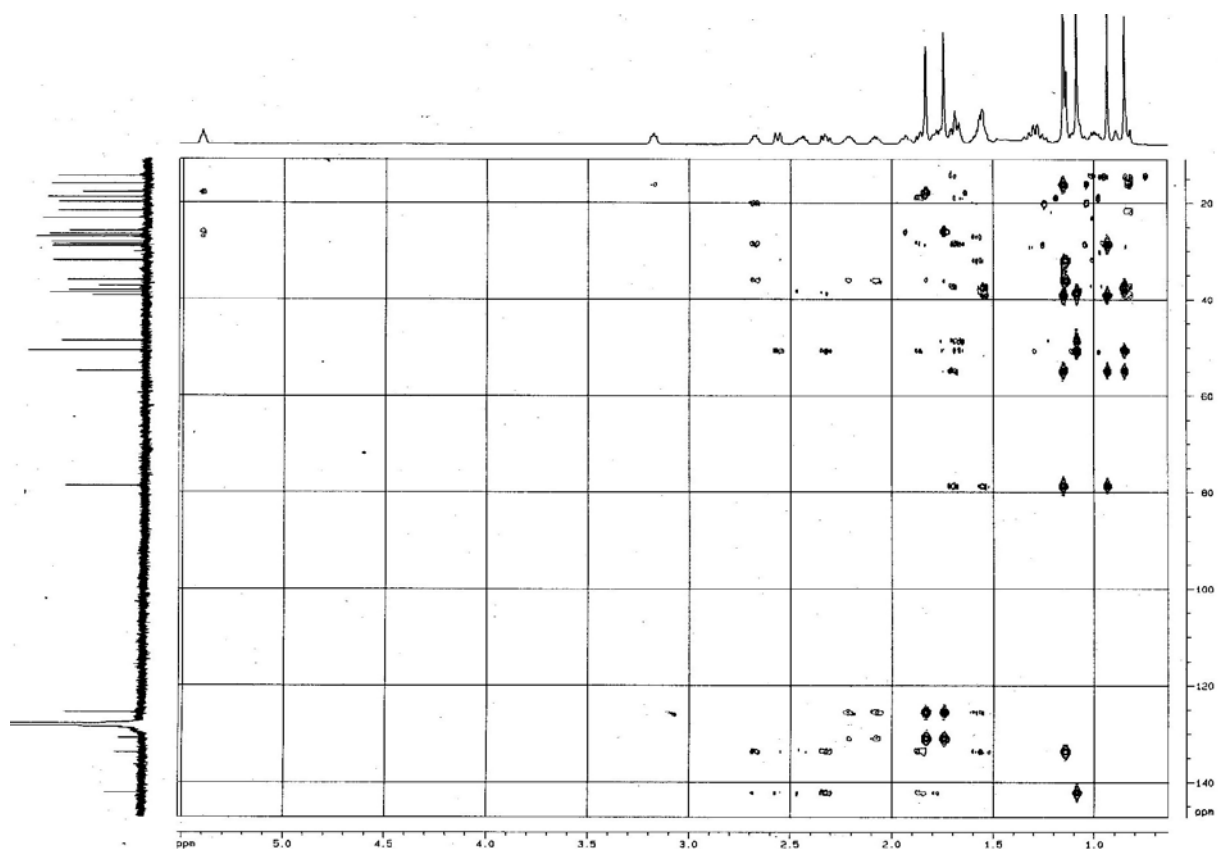
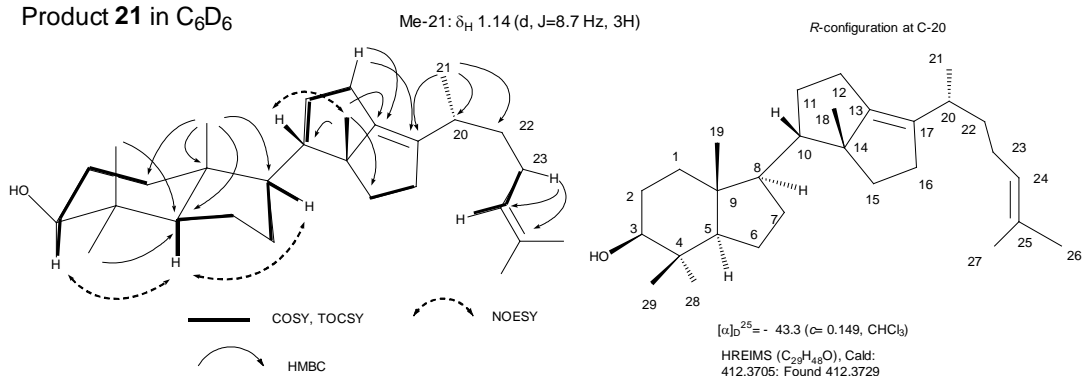


Fig. S4.8. HMBC spectrum of product **21** in C₆D₆.

Product **21** in C₆D₆



600 MHz, in C₆D₆; the residual solvent peak: δ_H 7.26; δ_C 77.0 ppm

NO.	¹ H	¹³ C	NO.	¹ H	¹³ C	NO.	¹ H	¹³ C	NO.	¹ H	¹³ C
1	1.00(m,1H); 1.68(m,1H)	38.21(t)	9	—	37.20(s)	17	—	133.8(s)	25	—	130.9(s)
2	1.54(m, 2H)	28.16(t)	10	1.29(m, 1H)	48.60(d)	18	1.068(s, 3H)	18.88(q)	26	1.833(s, 3H)	25.80(q)
3	3.18(bt, J=6.7 Hz, 1H)	78.71(d)	11	1.26(m, 1H); 1.74(m, 1H)	29.03(t)	19	0.851(s, 3H)	14.46(q)	27	1.745 (s, 3H)	17.80(q)
4	—	39.15(s)	12	1.93(m, 1H); 2.56(bd, J=13.2Hz, 1H)	23.14(t)	20	2.67(m, 1H)	32.01(d)	28	1.151(s, 3H)	28.64(q)
5	0.81 (m, 1H)	54.93(d)	13	—	142.1(s)	21	1.14 (d, J=7.2 Hz, 3H)	19.93(q)	29	0.935(s, 3H)	16.07(q)
6	1.31(m, 1H); 1.67 (m, 1H)	21.73(t)	14	—	50.65 (s)	22	1.55(m, 2H)	36.02(t)			
7	1.09(m, 1H); 1.78(m, 1H)	26.51(t)	15	1.57(m, 1H); 1.85(m, 1H)	38.58(t)	23	2.08(m, 1H); 2.20(m, 1H)	27.06(t)			
8	1.07 (m, 1H)	50.65(d)	16	2.32(m, 1H); 2.44(m, 1H)	28.59(t)	24	5.40(bt, J=6.7 Hz, 1H)	125.5(d)			

Fig. S4.9. Assignments of NMR data of product **21** in C₆D₆, HR-EIMS and optical rotation.

5. NMR spectra of product 23 in C₆D₆.

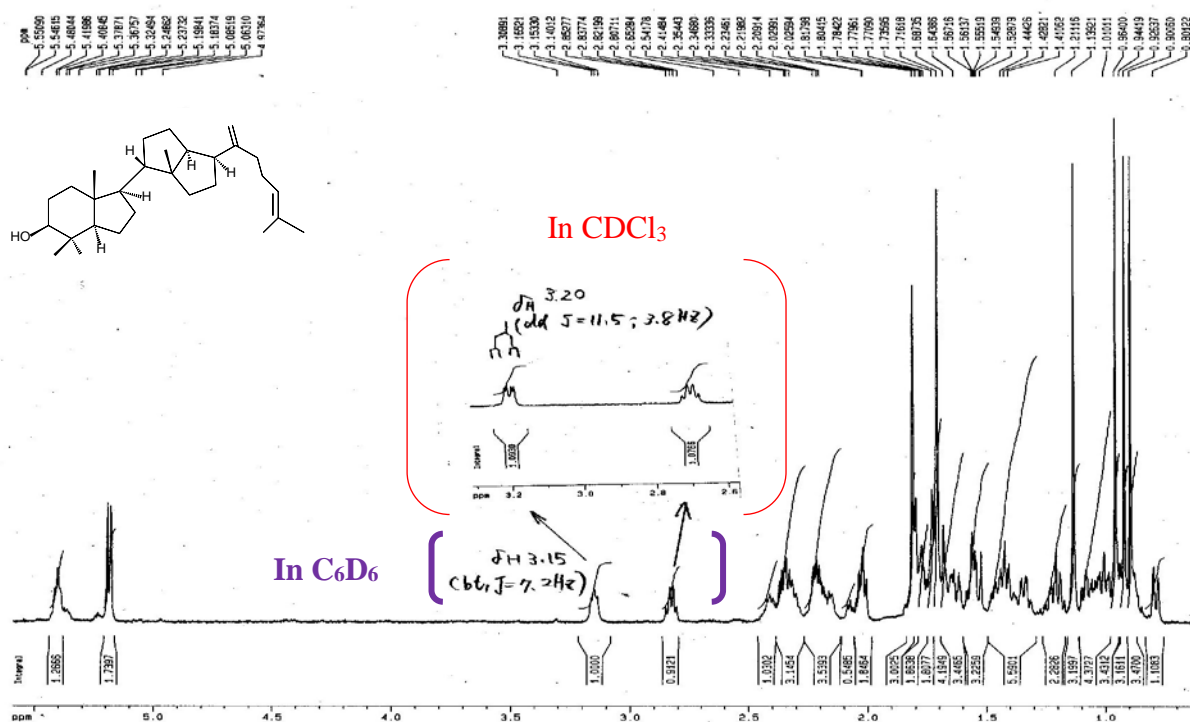


Fig. S5.1. ¹H NMR spectrum of product 23.

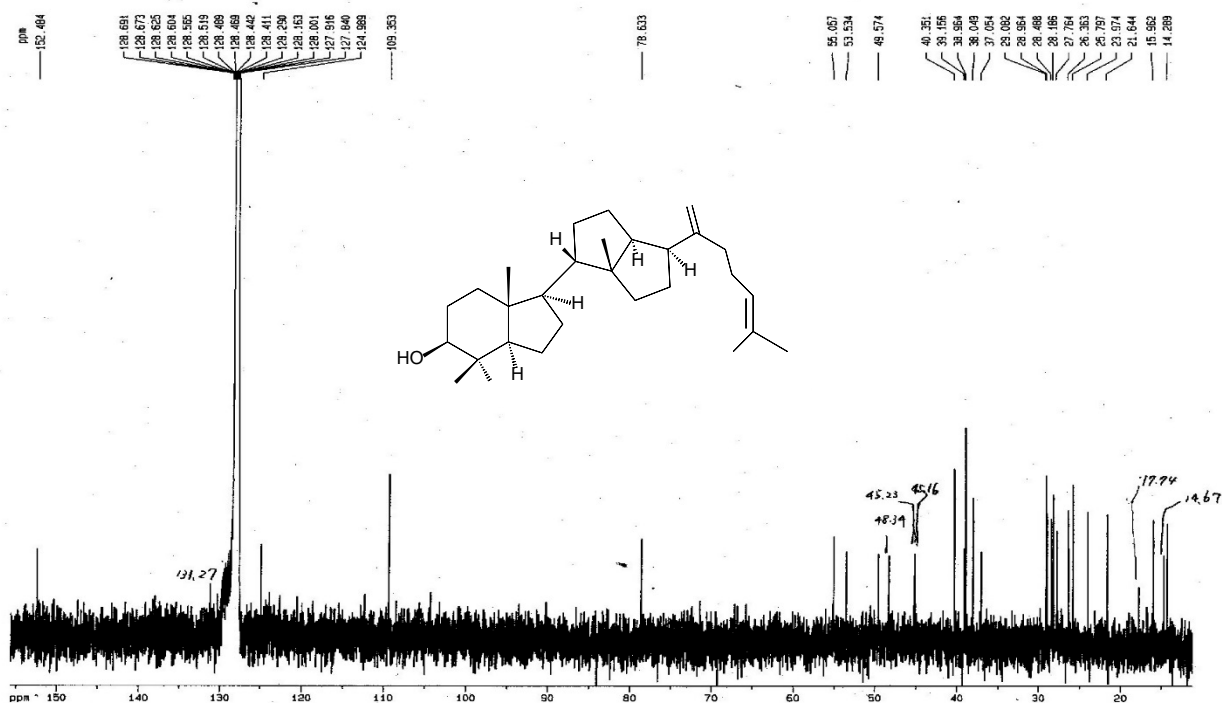


Fig. S5.2. ¹³C NMR spectrum of product 23.

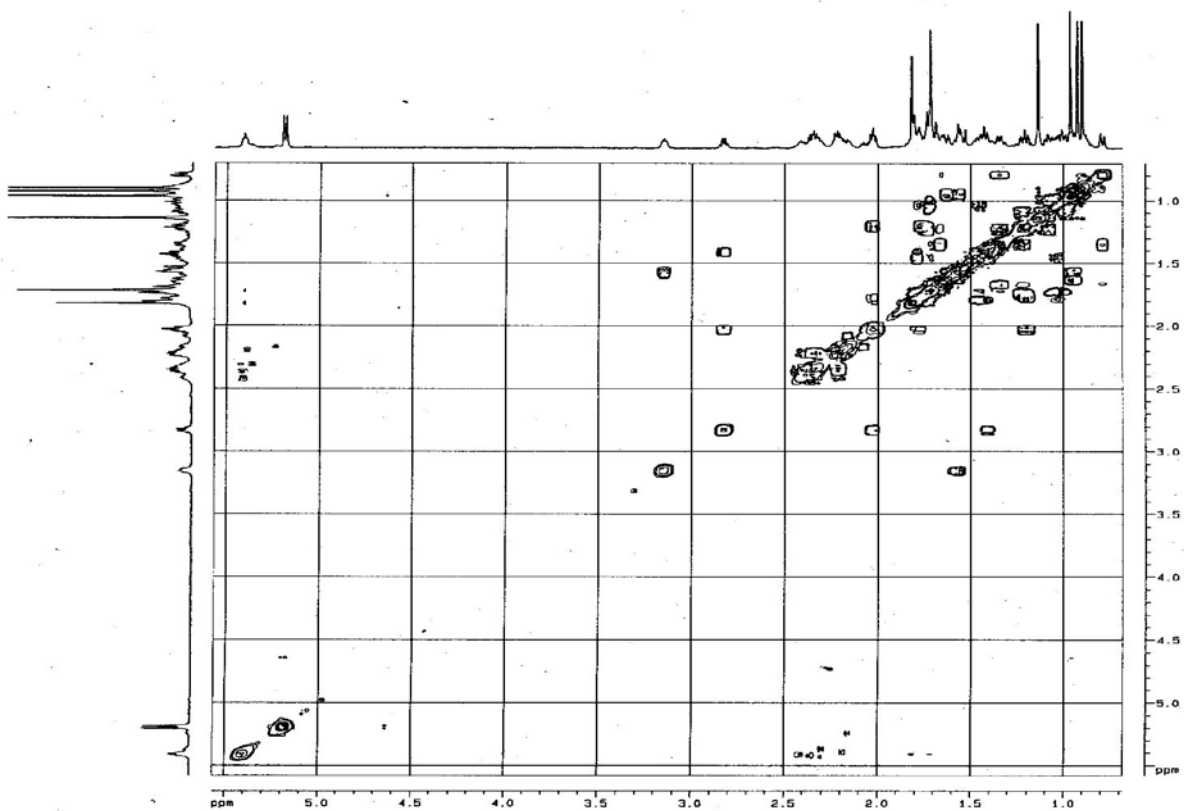


Fig. S5.3. COSY spectrum of product **23**.

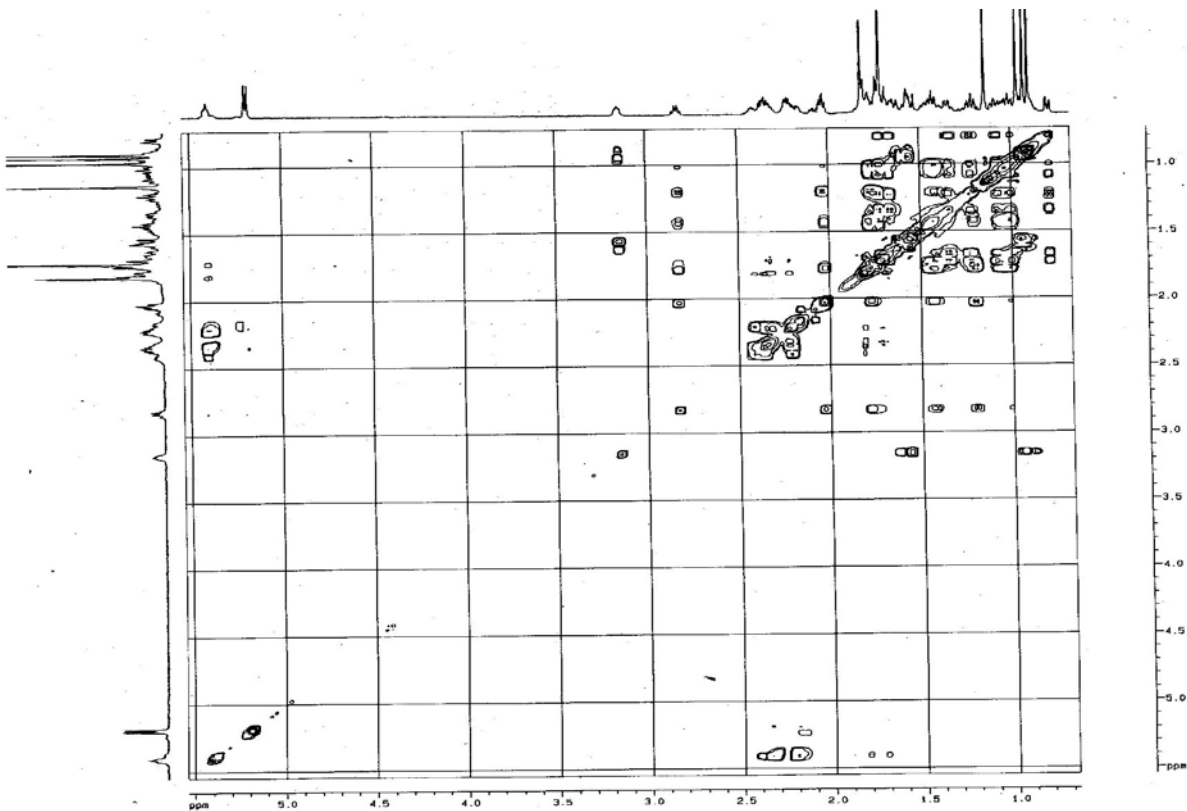


Fig. S5.4. TOCSY spectrum of product **23**

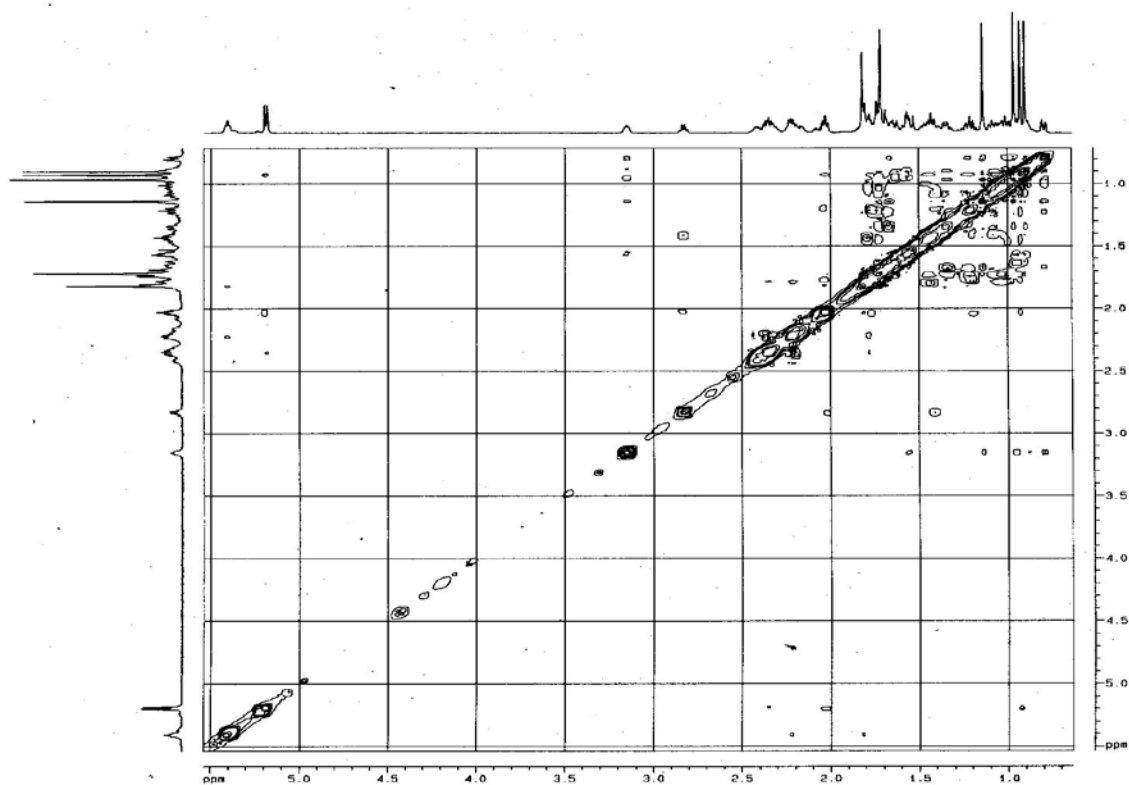


Fig. S5.5. NOESY spectrum of product 23.

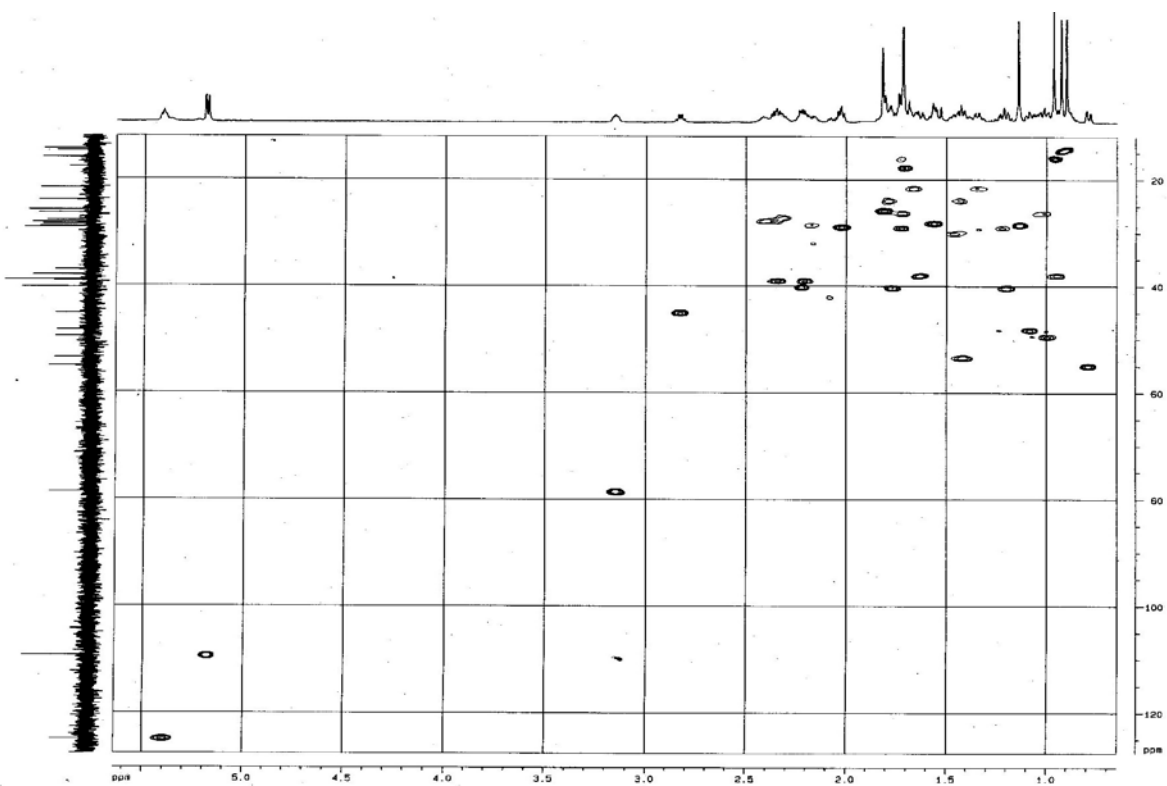


Fig. S5.6. HSQC spectrum of product 23.

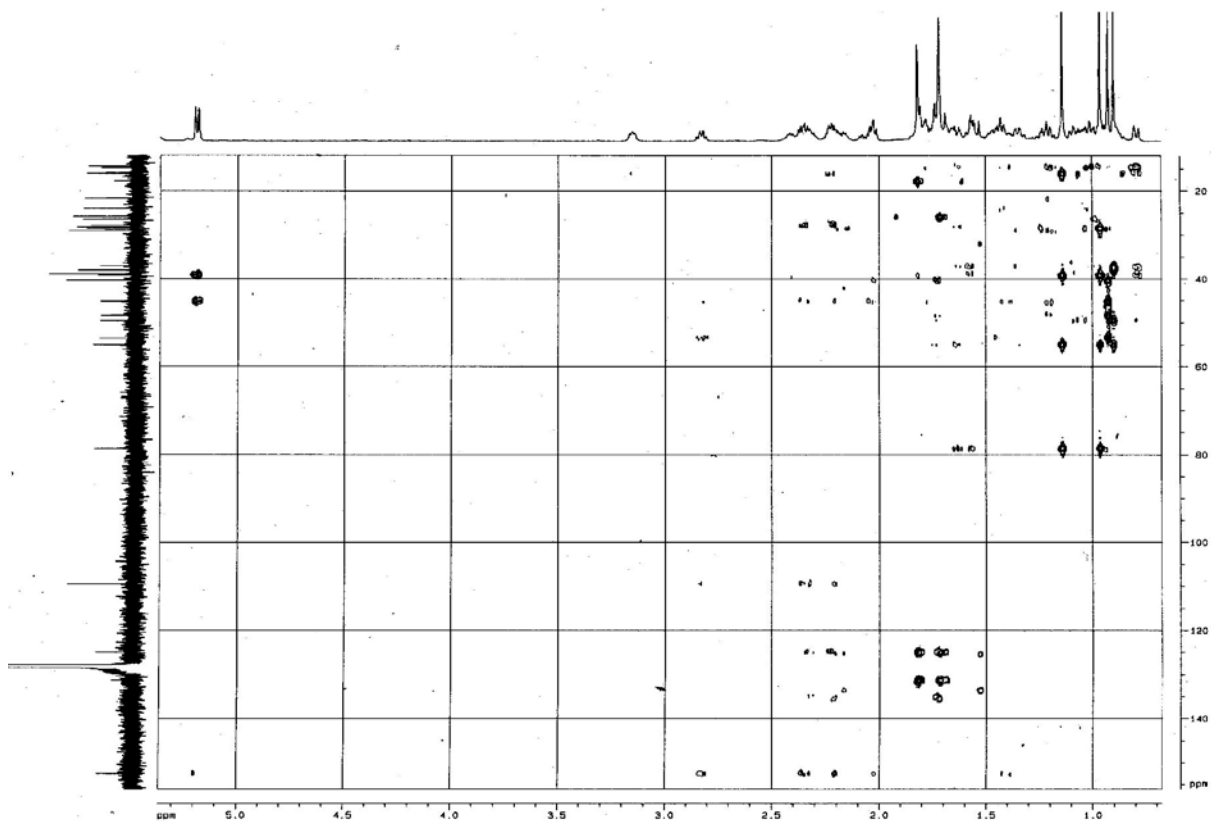
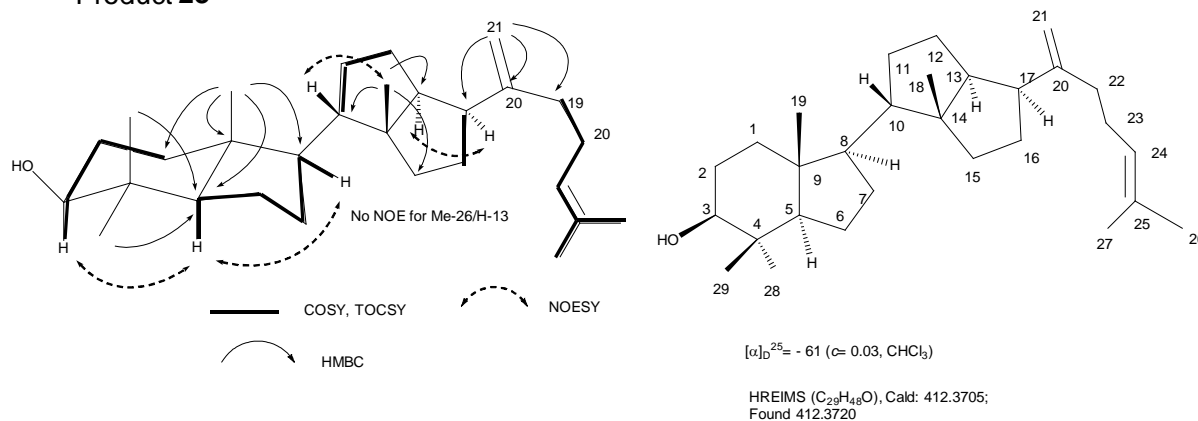


Fig. S5.7. HMBC spectrum of product **23**.

Product 23



600 MHz, in C_6D_6 ; the residual solvent peak: δ_{H} 7.28; δ_{C} 128.0 ppm

NO.	^1H	^{13}C	NO.	^1H	^{13}C	NO.	^1H	^{13}C	NO.	^1H	^{13}C
1	0.94(m,1H);1.62(m,1H)	38.05(t)	9	—	37.05(s)	17	2.83(dt, J=9.0 Hz, 1H)	45.16(d)	25	—	131.3(s)
2	1.56 (m, 2H)	28.19(t)	10	1.08(m, 1H)	48.34(d)	18	0.926(s, 3H)	14.67(q)	26	1.818(s, 3H)	25.80(q)
3	3.15 (br t, J=7.2 Hz, 1H)	78.63(d)	11	1.02(m, 1H); 1.72(m,1H)	26.36(t)	19	0.901(s, 3H)	14.29(q)	27	1.716 (s, 3H)	17.74(q)
4	—	39.16(s)	12	1.43(m,1H); 1.78(m,1H)	23.97(t)	20	—	152.5(s)	28	1.14(s,3H)	28.49(q)
5	0.794(dd, J=11.8; 2.1 Hz, 1H)	55.06(d)	13	1.42 (m, 1H)	53.53(d)	21	5.18 (s, 1H); 5.20(s, 1H)	109.3(t)	29	0.964(s,3H)	15.97(q)
6	1.34(m,1H); 1.67(m, 1H)	21.64(t)	14	—	45.23 (s)	22	2.21(m); 2..33(m)	38.96(t)			
7	1.22(m, 1H); 1.73(m, 1H)	29.08(t)	15	1.20(m,1H); 1.77(m, 1H)	40.35(t)	23	2.36(m,1H); 2.41(m,1H)	27.76(t)			
8	1.00 (m, 1H)	49.57(d)	16	2.03(m,2H)	28.96(t)	24	5.41(t, J=6.8 Hz, 1H)	125.0(d)			

Fig. S5.8. Assignments of NMR data of product **23** in C_6D_6 , HR-EIMS and optical rotation.

Remark; In the C_6D_6 solution, the H-3 showed br triplet splitting pattern, but in the CDCl_3 solution, H-3 showed the dd splitting pattern (δ_{H} 3.20, dd, $J=11.5$ and 3.8 Hz, see Fig. S6.1), thus strongly indicating that the OH at C-3 is positioned in a β -orientation. A definitive NOE for H-3/H-5 further supported β -OH orientation at C-3 in the NOESY spectrum (C_6D_6 solution). (See Fig. S6.5).

6. NMR spectra of product 25 in C₆D₆.

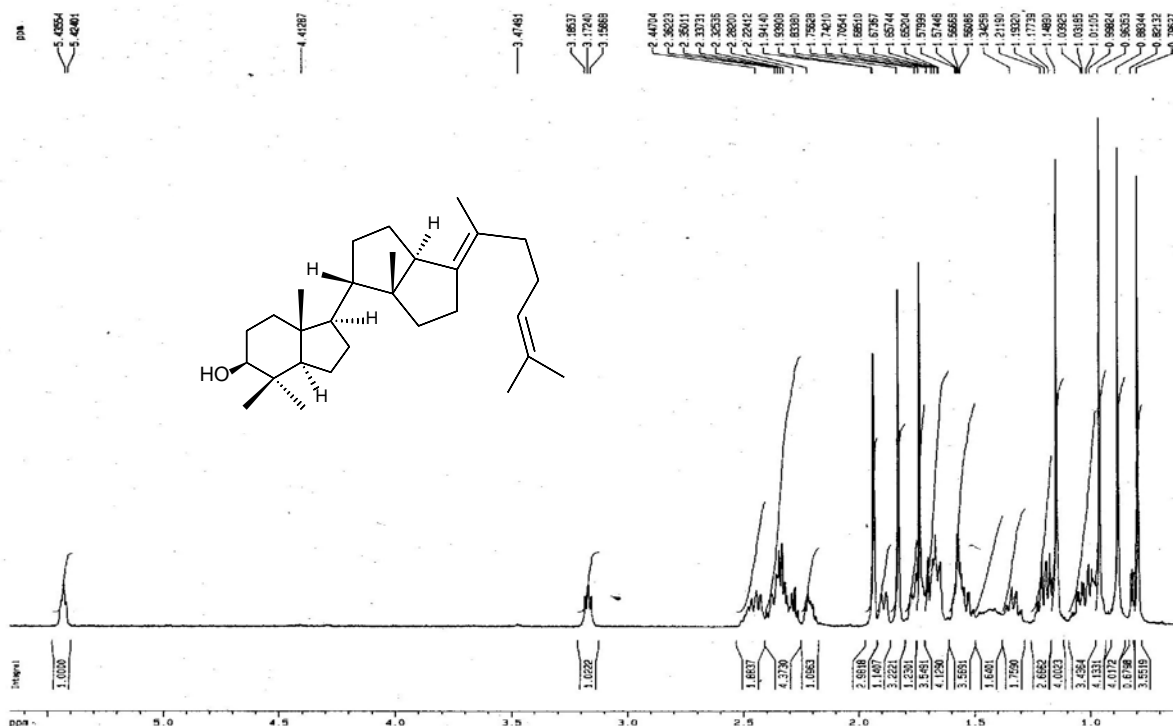


Fig. S6.1. ¹H NMR spectrum of product **25**

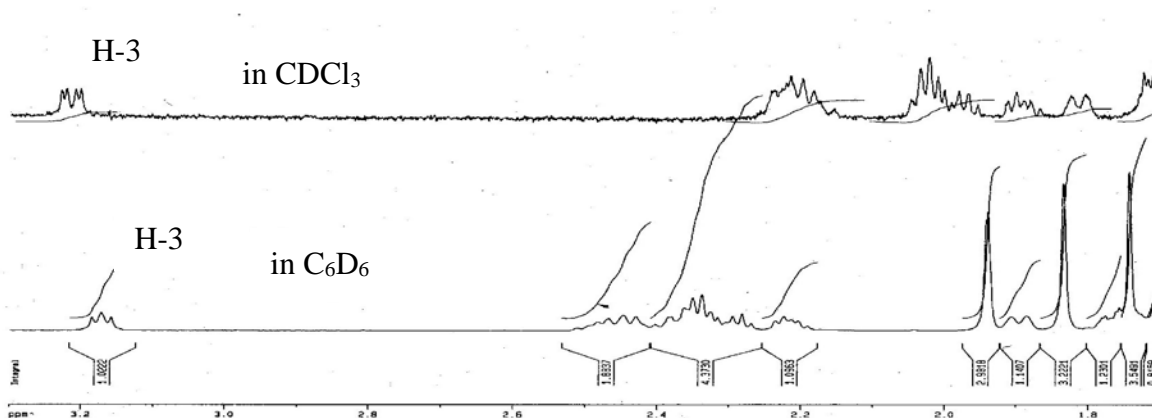


Fig. S6.2. Comparison of ¹H NMR spectra of product **25** between CDCl₃ and C₆D₆ solutions (600 MHz).

The coupling constants of H-3 were compared between the solutions of C₆D₆ and CDCl₃.

δ_{H} in C₆D₆: 3.17 (br t, $J=8.0$ Hz); δ_{H} in CDCl₃: 3.21 (dd, $J=11.7$ Hz and 4.3 Hz). The coupling constants (splitting pattern) in CDCl₃ solution clearly demonstrated that the OH group is β -oriented, although the splitting pattern (triplet) in C₆D₆ suggests that the OH group is α -oriented. This contradictory argument was resolved by the definitive NOE between H-3 and H-5 in the C₆D₆ solution. Consequently, the OH at C-3 of product **25** was credibly determined to be β -oriented. The ¹H-NMR spectra of products **21** and **23** in C₆D₆ solution also triplet splitting pattern, but dd splitting in CDCl₃.

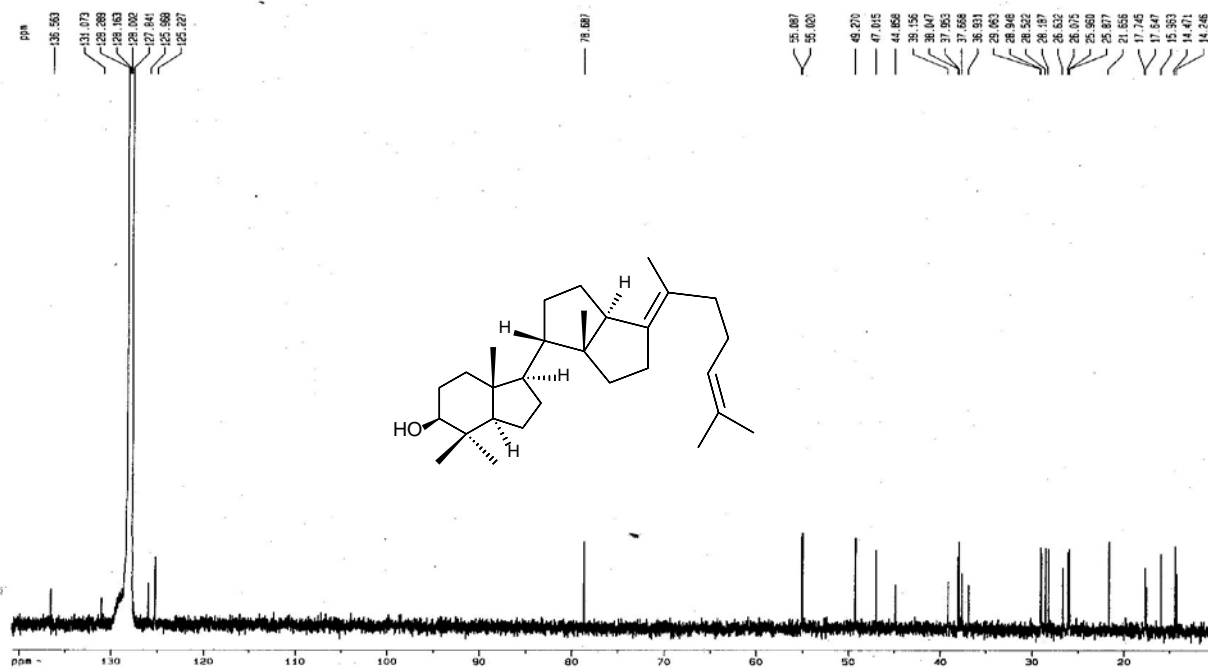


Fig. S6.3. ^{13}C NMR spectrum of product 25

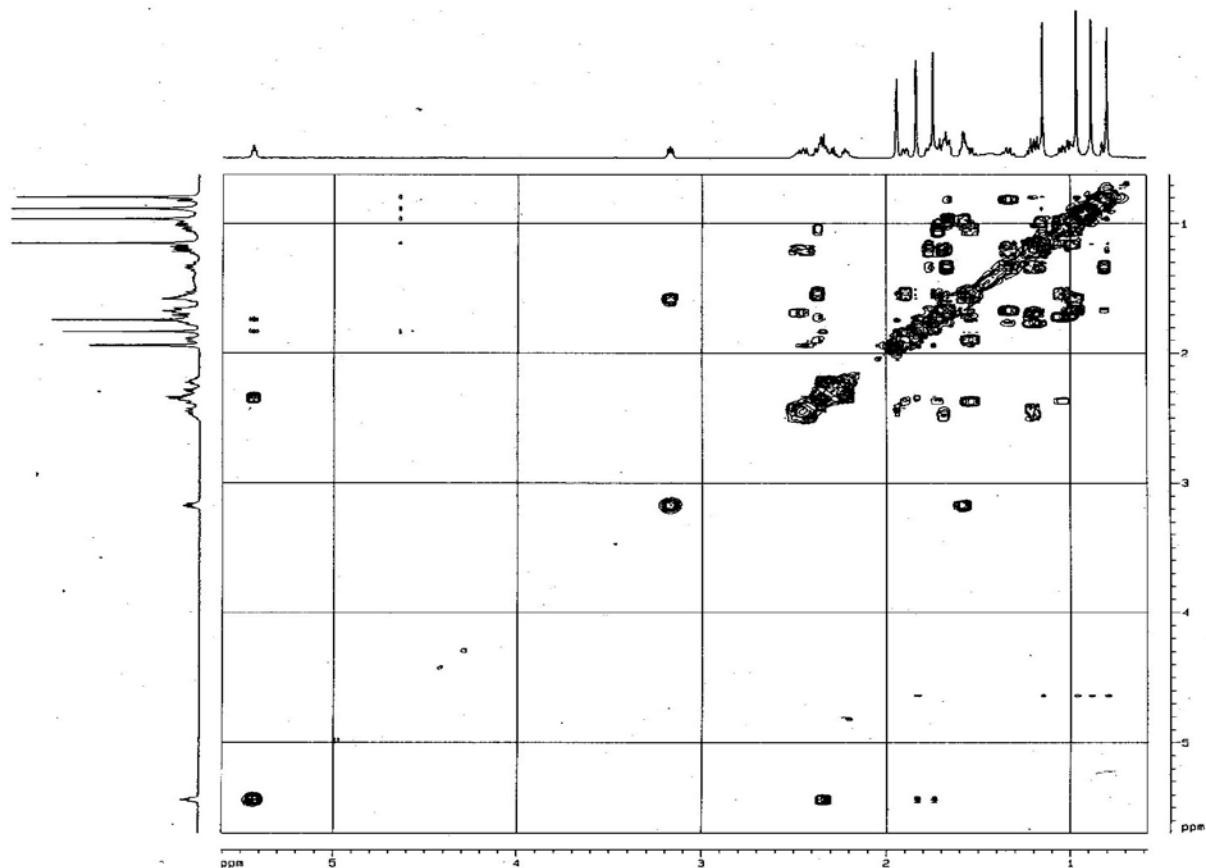


Fig. S6.4. COSY spectrum of product 25

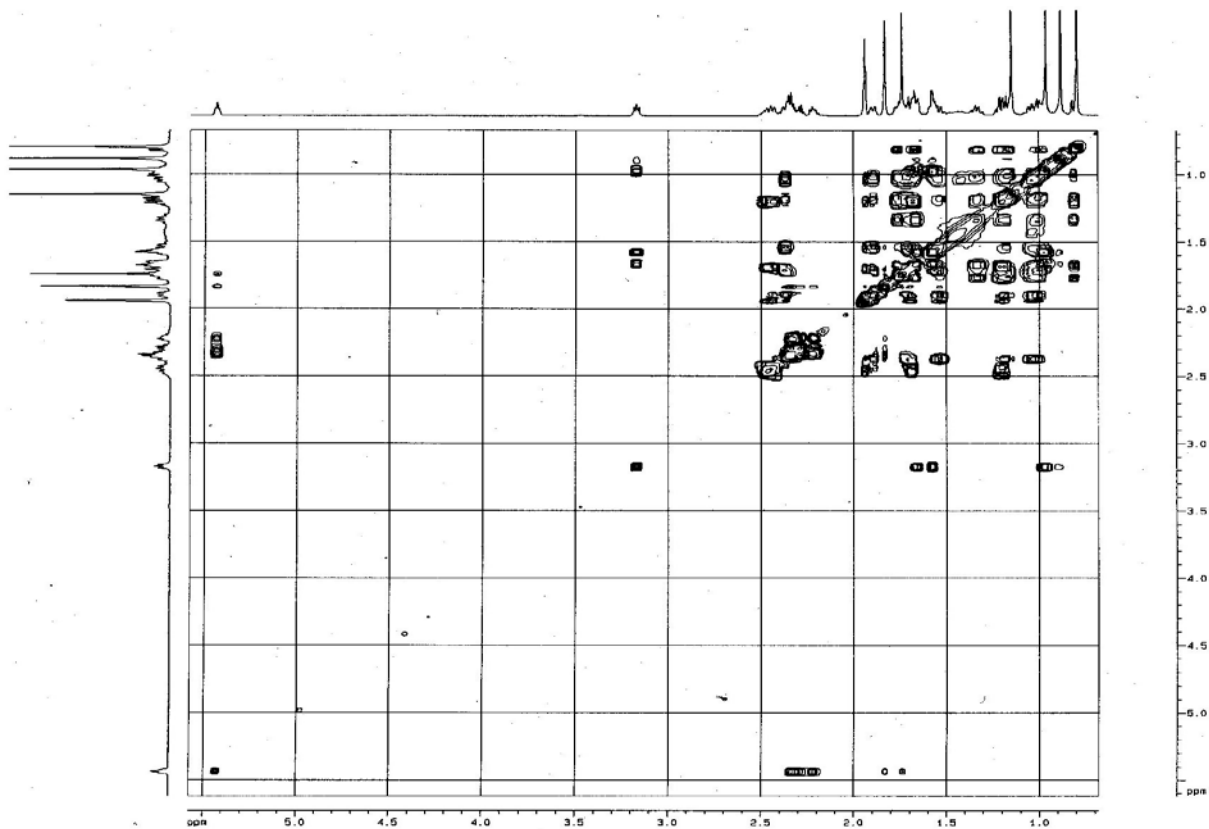


Fig. S6.5. TOCSY spectrum of product 25

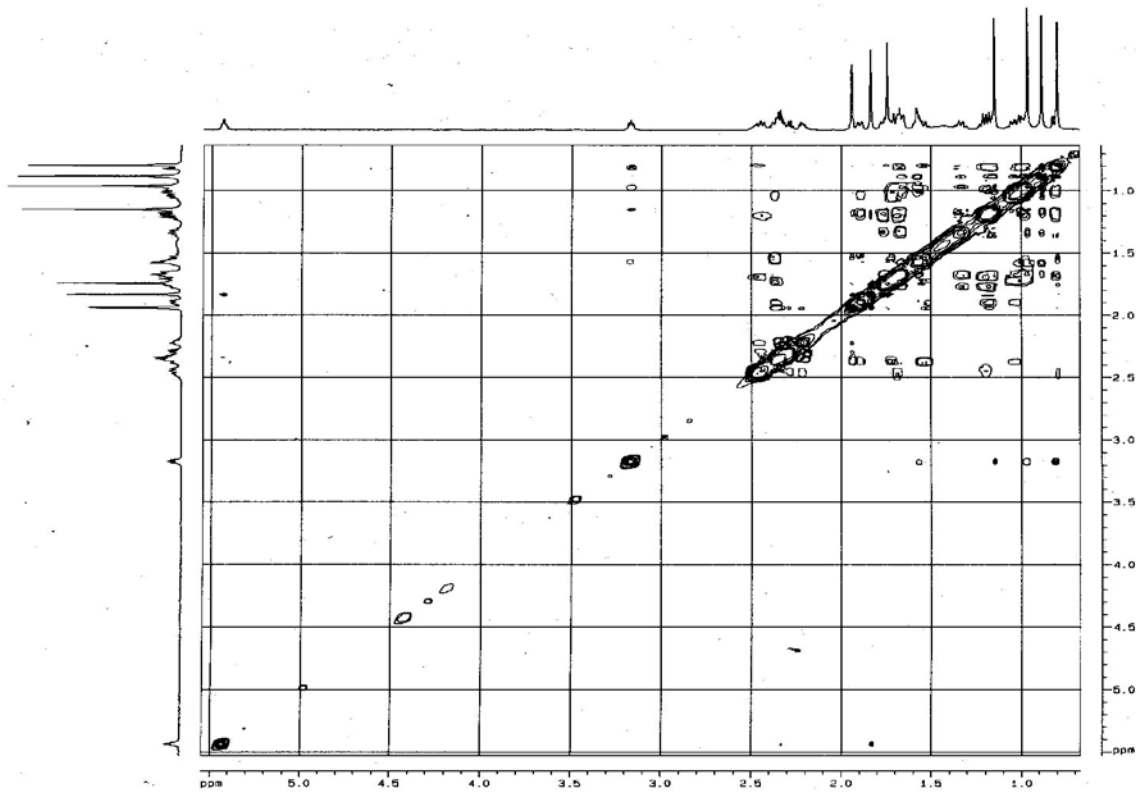


Fig. S6.6. NOESY spectrum of product 25

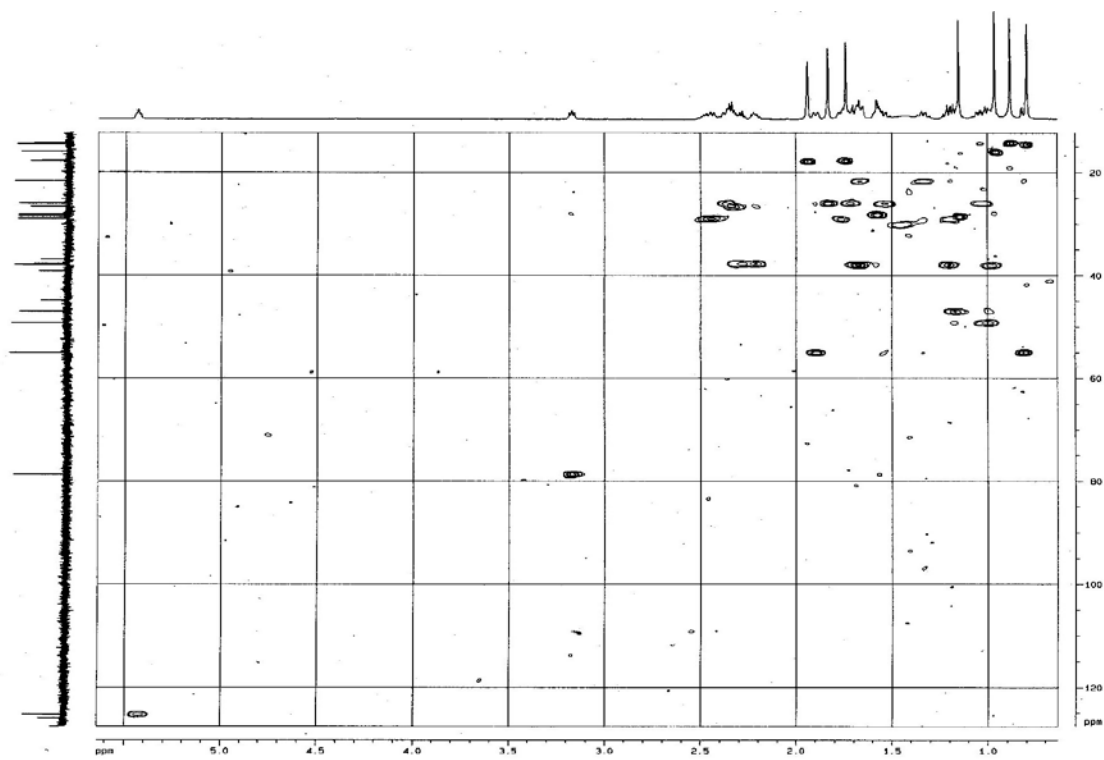


Fig. S6.7. HSQC spectrum of product 25

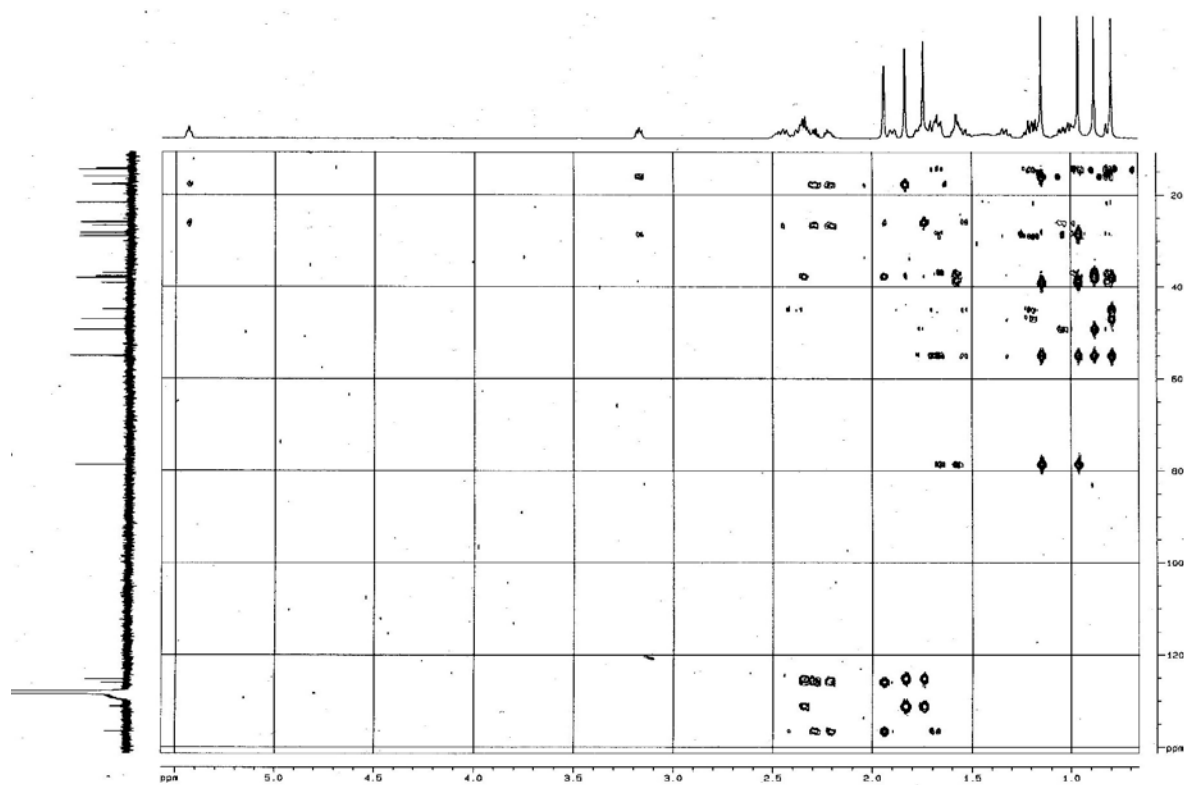
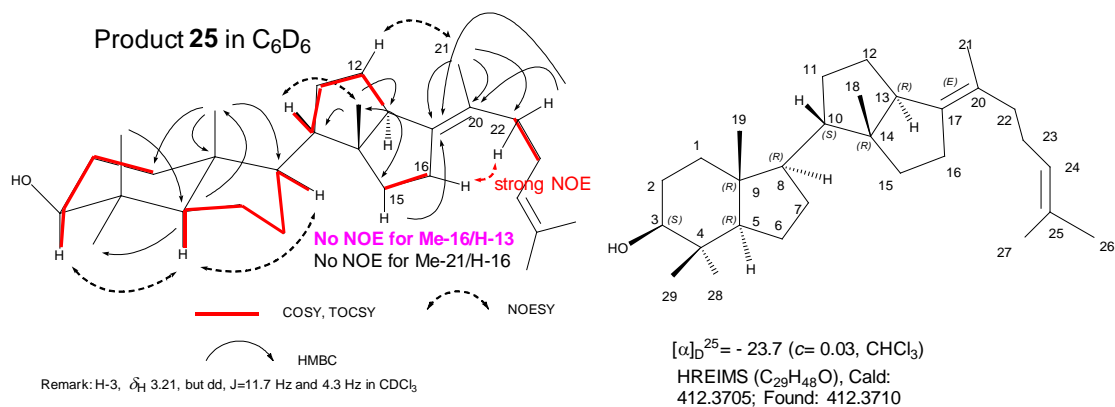


Fig. S6.8. HMBC spectrum of product 25

Fig. S6.9. Assignments of NMR data of product **25** in C₆D₆, HR-EIMS and optical rotation.



600 MHz, in C₆D₆; the residual solvent peak: δ_{H} 7.28; δ_{C} 128.0 ppm

NO.	¹ H	¹³ C	NO.	¹ H	¹³ C	NO.	¹ H	¹³ C	NO.	¹ H	¹³ C
1	0.97(m,1H);1.68(m,1H)	37.95(t)	9	—	36.93(s)	17	—	136.6(s)	25	—	131.0(s)
2	1.57(m,2H)	28.19(t)	10	1.16 (m, 1H)	42.01(d)	18	0.796(s,3H)	14.47(q)	26	1.834 (s, 3H)	25.96(q)
3	3.17 (bt, J=8.0 Hz, 1H)	78.69(d)	11	1.02(m, 1H); 1.72(m,1H)	25.88(t)	19	0.883(s,3H)	14.24(q)	27	1.742(s,3H)	17.65(q)
4	—	39.16(s)	12	1.55(m,1H); 2.35(m,1H)	26.07(t)	20	—	125.9(s)	28	1.489(s,3H)	28.52(q)
5	0.82(m, 1H)	55.09(d)^a	13	1.90 (bd, J=13.0 Hz, 1H)	55.02(d)^a	21	1.940 (bs, 3H)	17.74(q)	29	0.963(s,3H)	15.93(q)
6	1.33(m,1H); 1.67(m, 1H)	21.65(t)	14	—	44.85 (s)	22	2.21(m, 1H);2.30(m, 1H)	37.68(t)			
7	1.20(m, 1H); 1.77(m, 1H)	29.06(t)	15	1.20(m,1H); 1.68(m, 1H)	38.05(t)	23	2.35 (m, 2H)	26.93(t)			
8	1.00(m, 1H)	49.27(d)	16	2.45(m,2H)	28.95(t)	24	5.43(bd, J=6.7 Hz, 1H)	125.2(d)			

a) The assignment of C-5 and C-13 may be exchangeable.

Remark. The H-3 showed br triplet splitting patten (Figs. S14.2) in the C₆D₆ solution. However, in the CDCl₃ solution, H-3 showed the dd splitting pattern (δ_{H} 3.21, d, $J=11.7$ Hz and 4.3 Hz), thus strongly indicating that the OH at C-3 is β -oriented. The finding of a clear NOE in the C₆D₆ solution (Fig. S14.6) further supported the β -orientation of 3-OH.

7. EIMS and NMR data of product 26

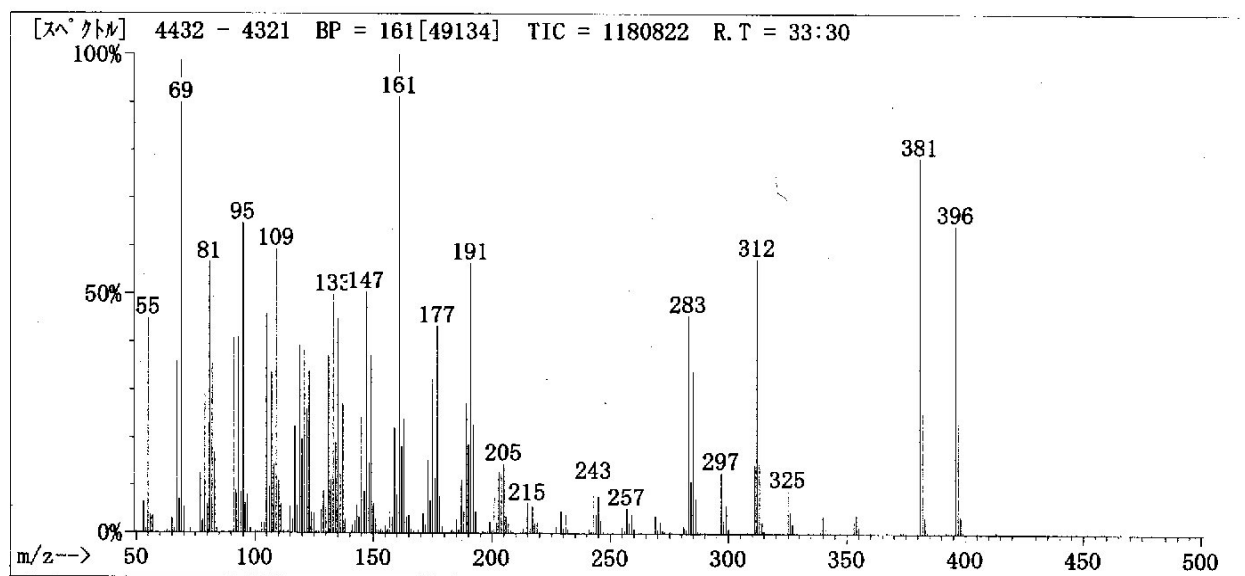


Fig. S7.1. EIMS of product 26.

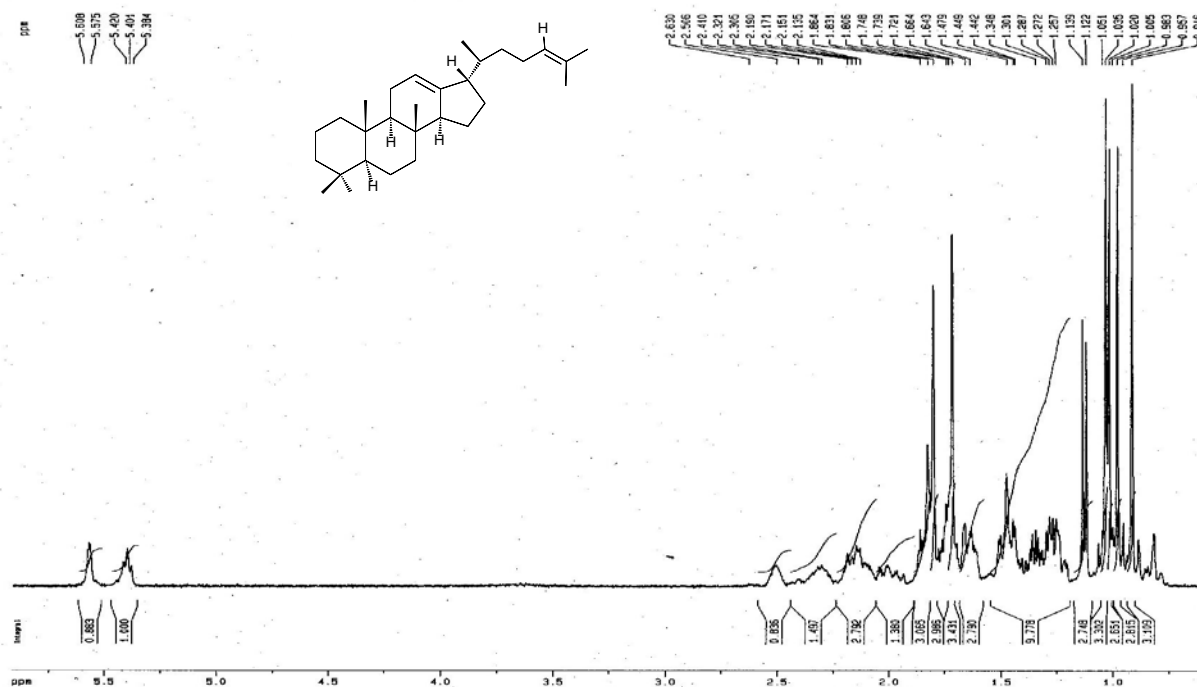


Fig. S7.2.1. ^1H NMR spectrum of product 26 (400 MHz, C_6D_6).

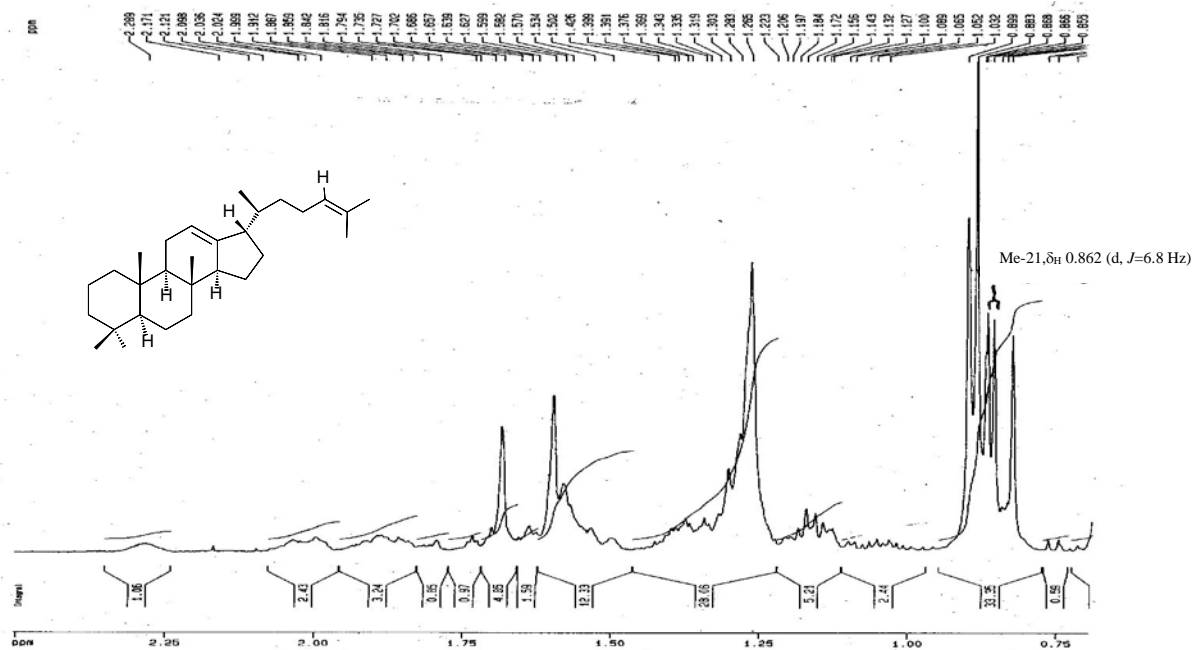


Fig. S7.2.2. ^1H NMR spectrum (expanded region) of product **26** (400 MHz, CDCl_3)

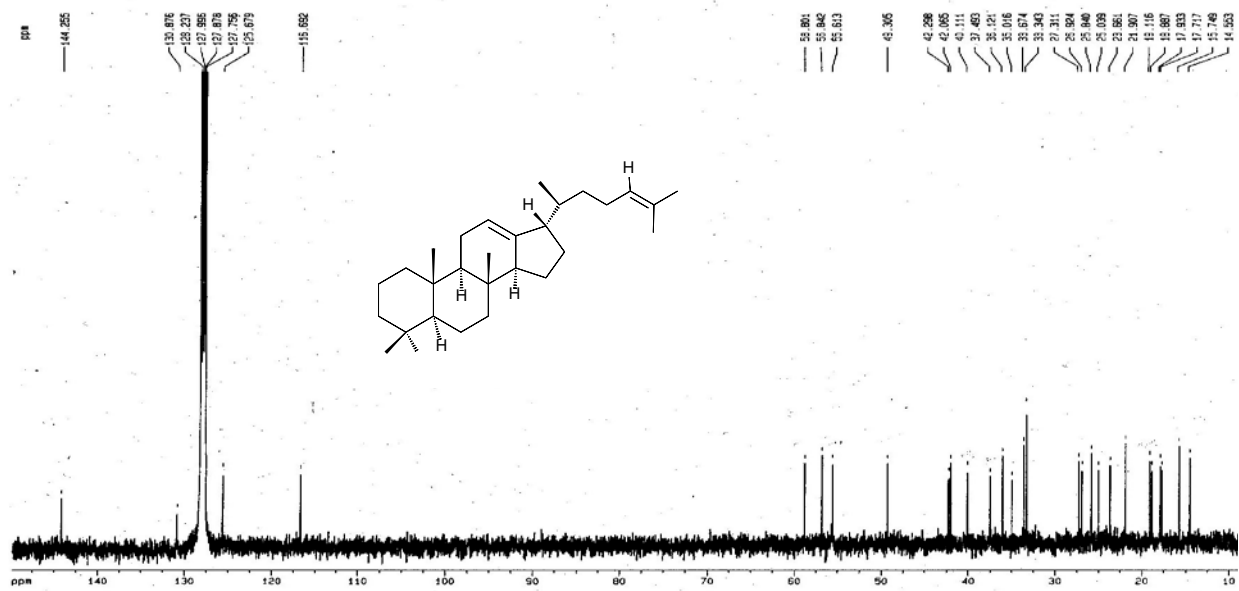


Fig. S7.3. ^{13}C NMR of product **26** (100 MHz, C_6D_6).

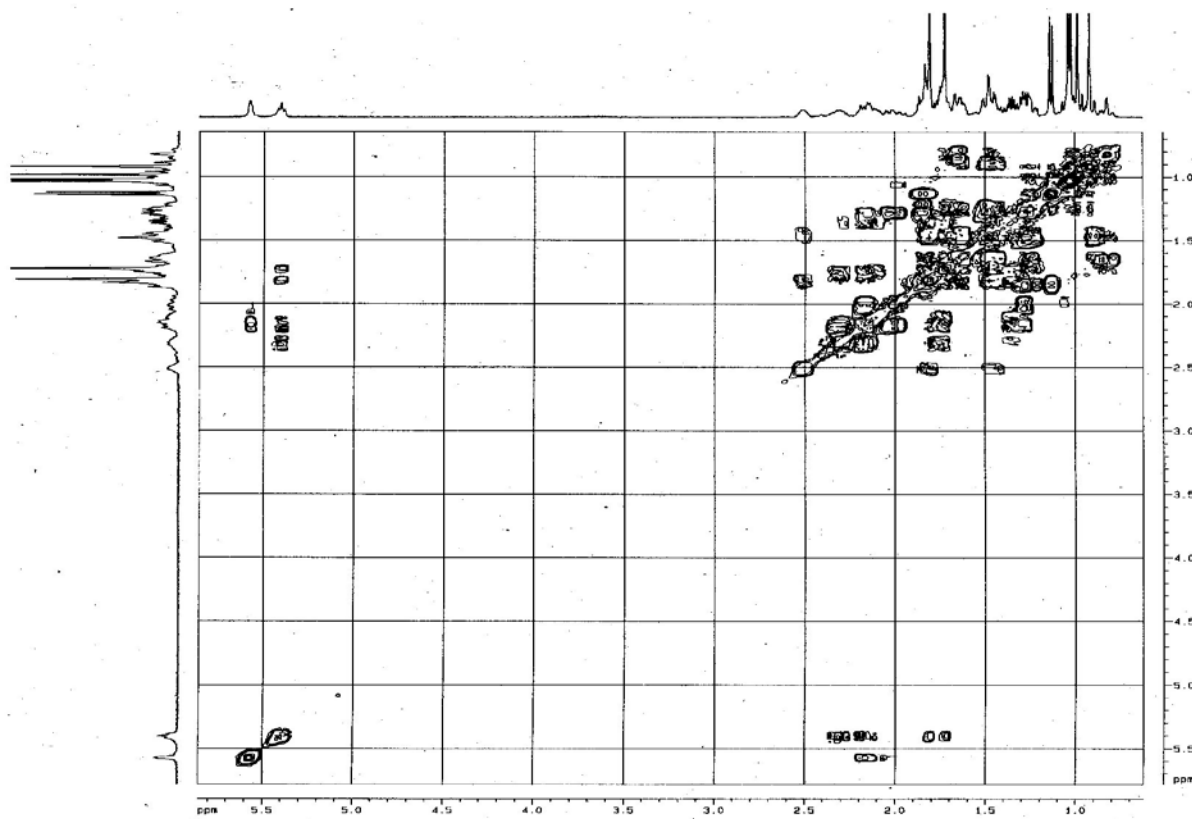


Fig. S7.4. ^1H - ^1H COSY of product **26** (400 MHz, C_6D_6).

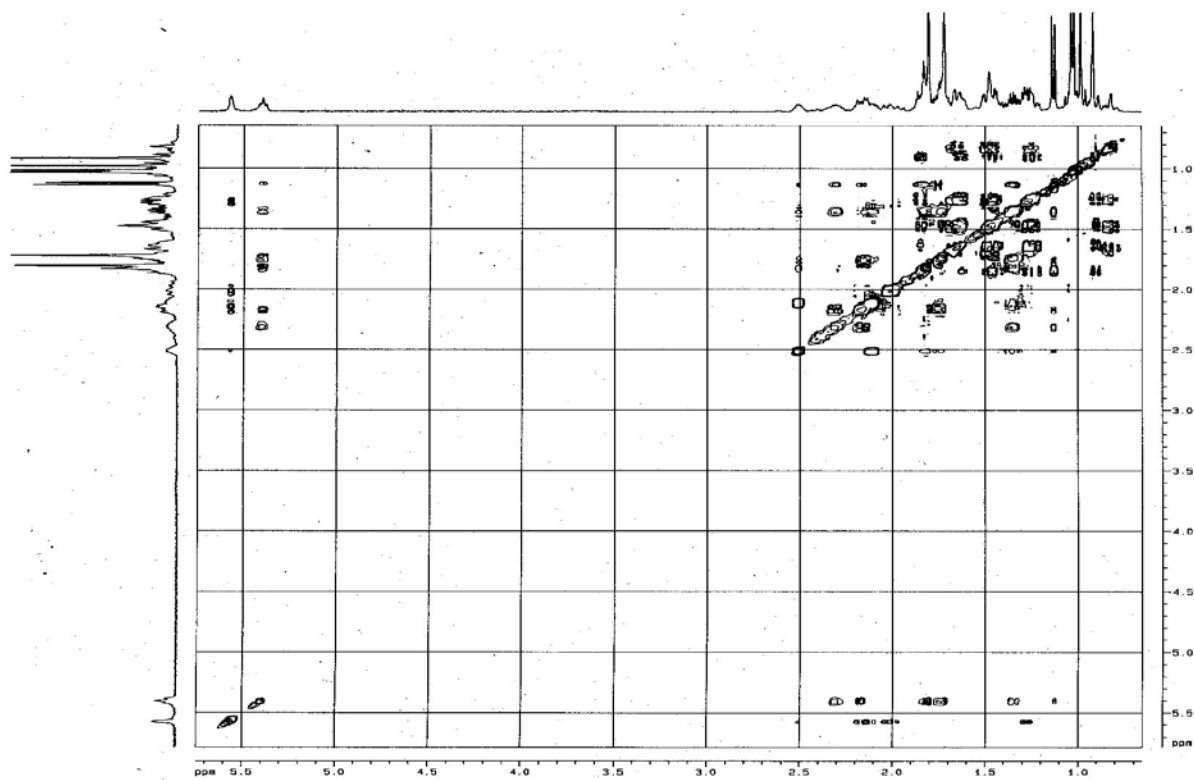


Fig. S7.5. TOCSY of Product **26** (400 MHz, C_6D_6).

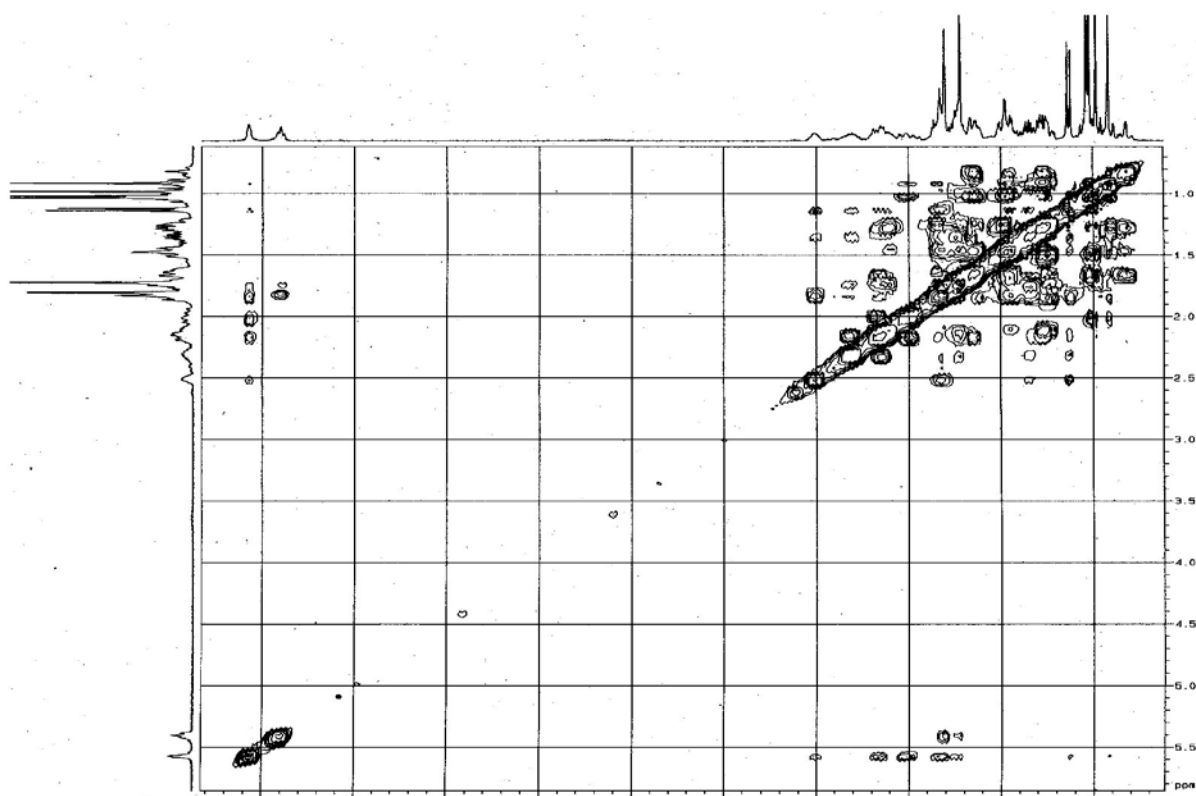


Fig. S7.6. NOESY of Product 26 (400 MHz, C₆D₆).

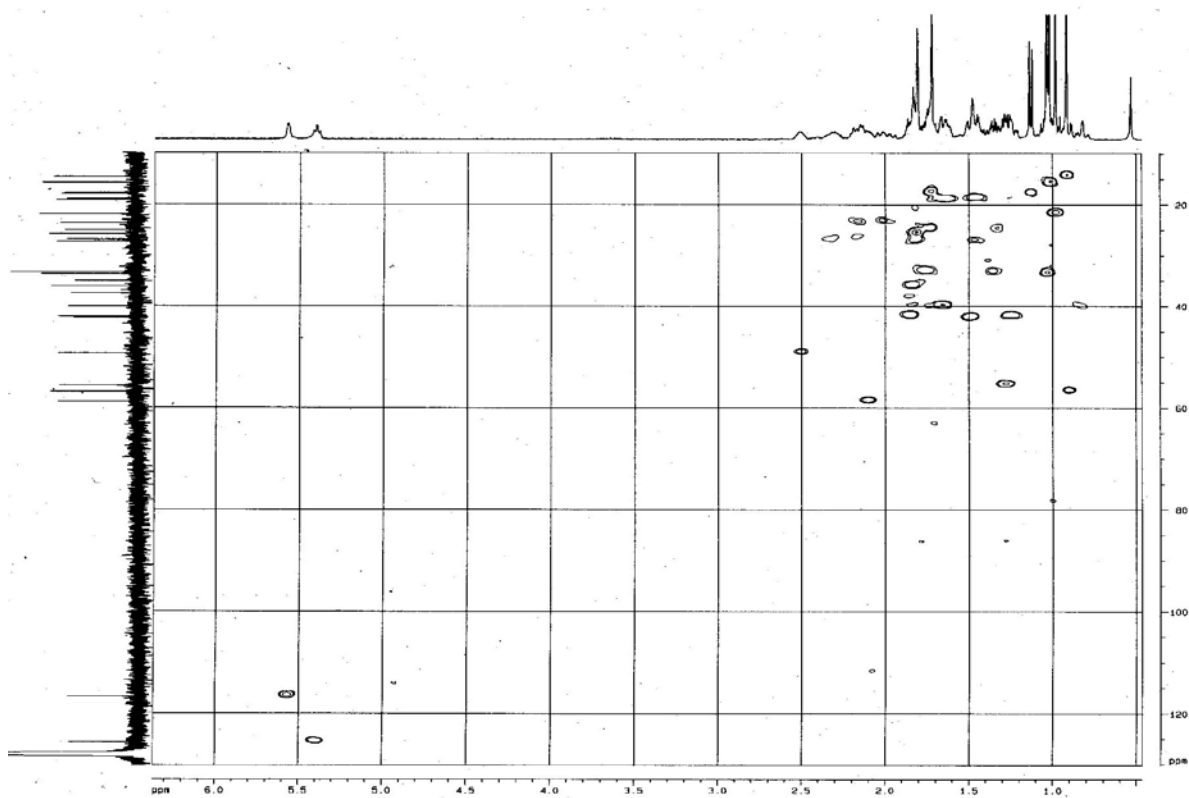


Fig. S7.7. HSQC of Product 26 (400 MHz, C₆D₆).

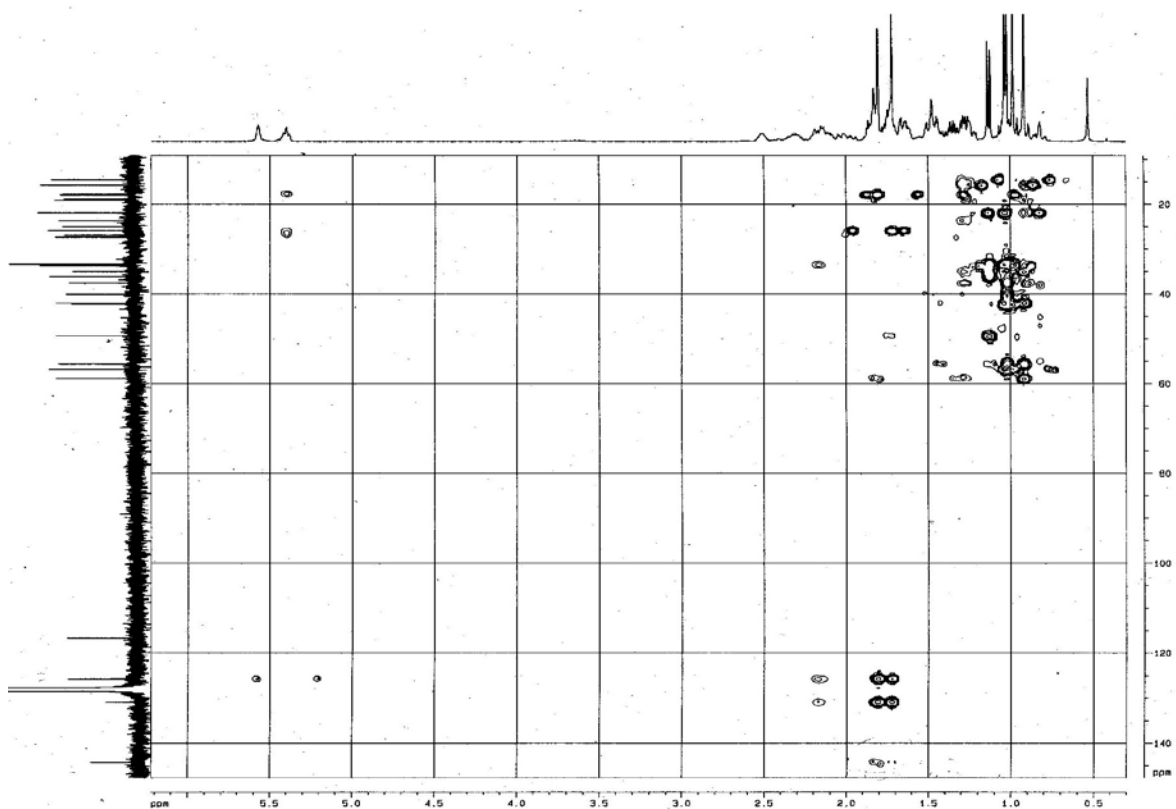
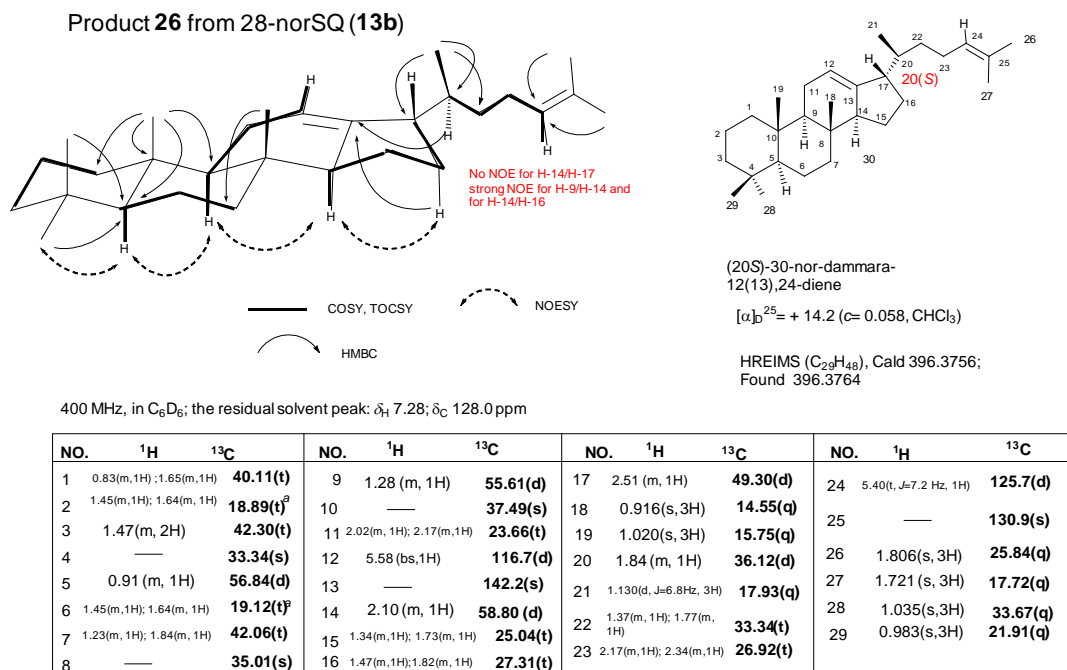


Fig. S7.8. HMBC of Product **26** (400 MHz, C₆D₆).



Me-27: δ_H=0.862 (d, J=6.8 Hz) in CDCl₃, which is indicative of 18-S stereochemistry.

Fig. S7.9. Assignments of NMR data of product **26** in C₆D₆, HR-EIMS and optical rotation.

8. EIMS and NMR data of product 27.

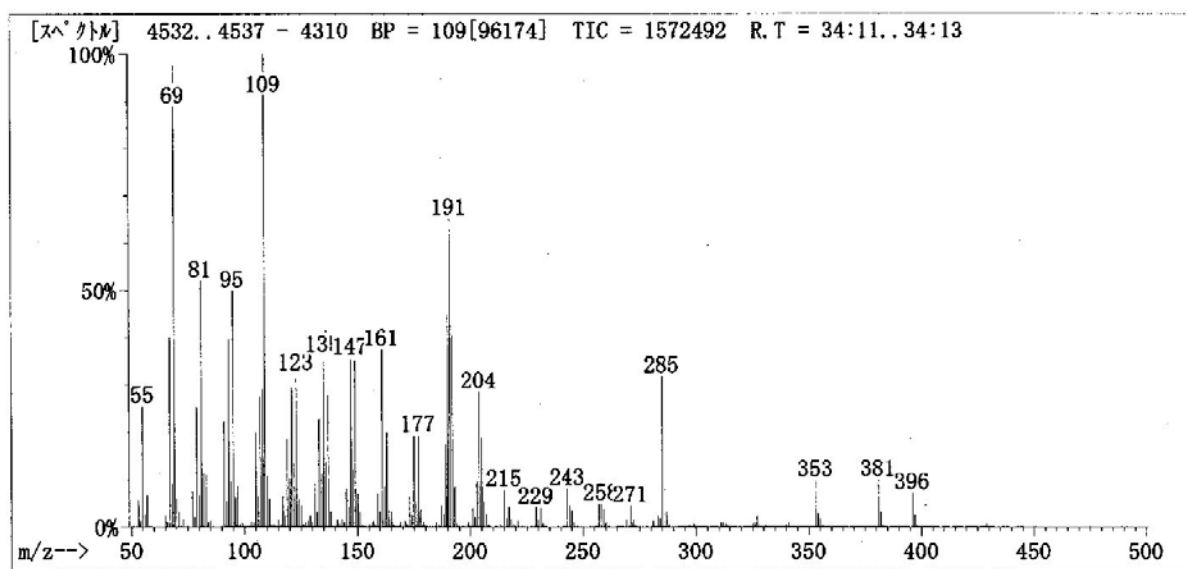


Fig. S8.1. EIMS of product 27.

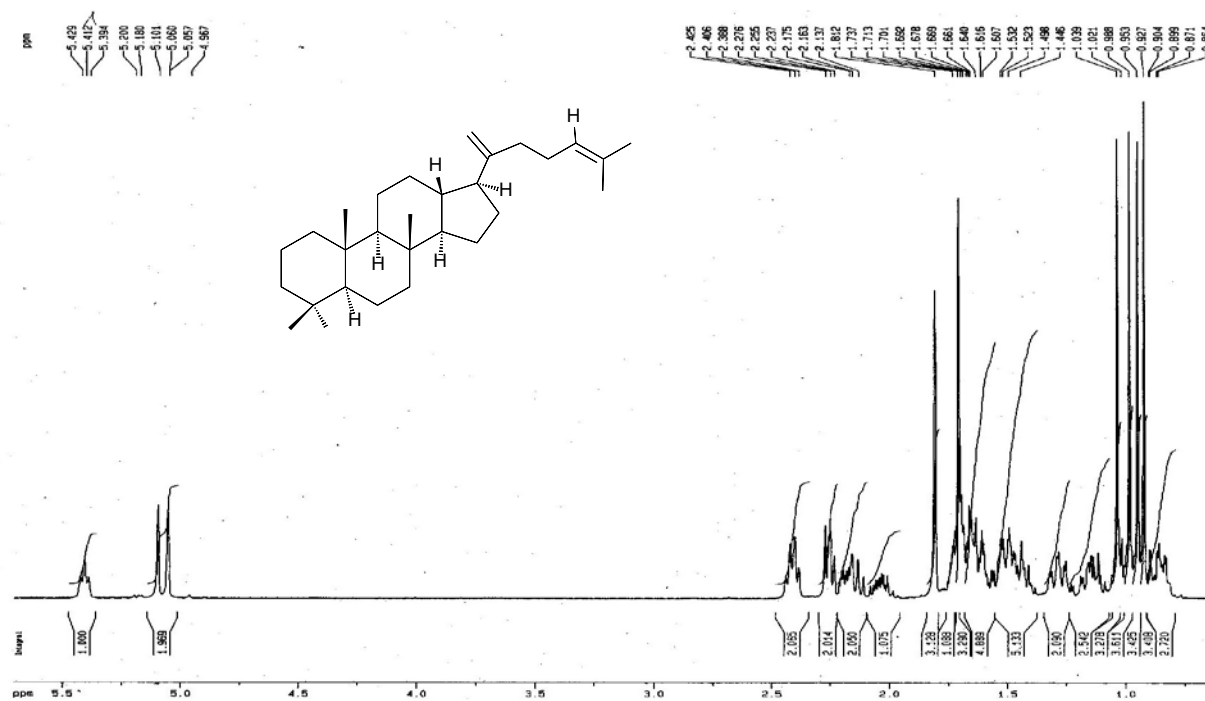


Fig. S8.2. ^1H NMR of Product 27 (400 MHz, C_6D_6).

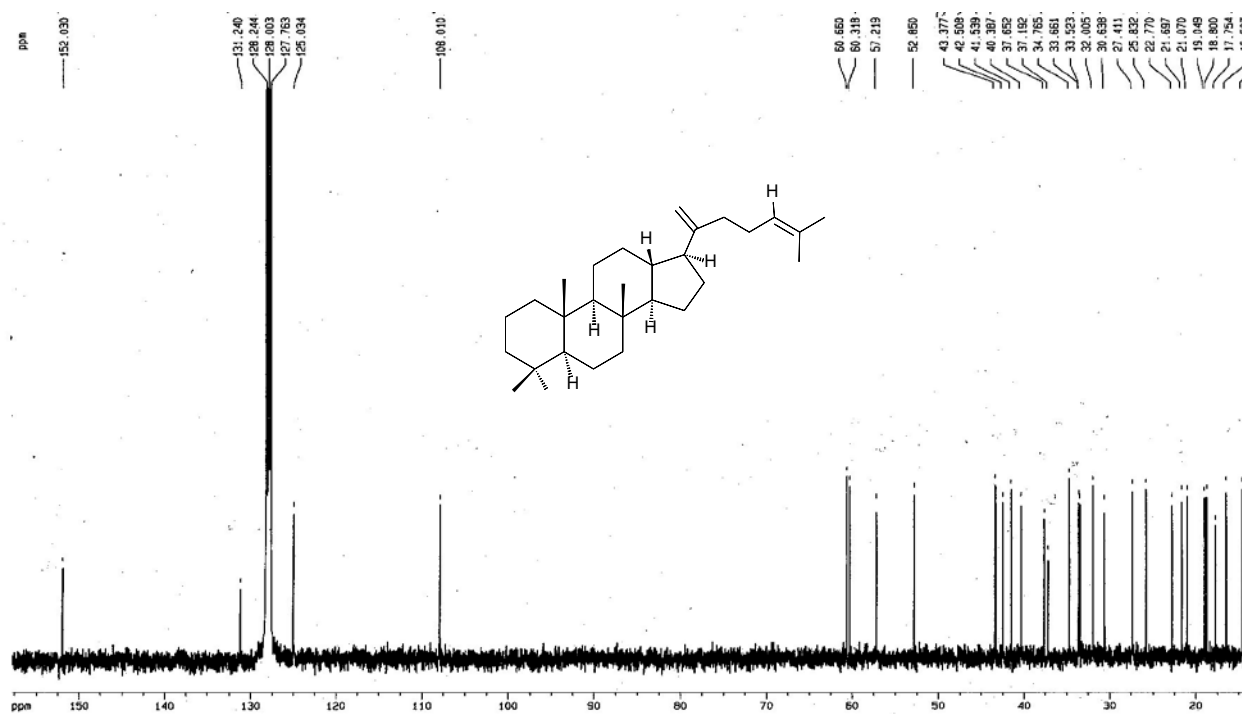


Fig. S8.3. ^{13}C NMR of Product 27 (100 MHz, C_6D_6).

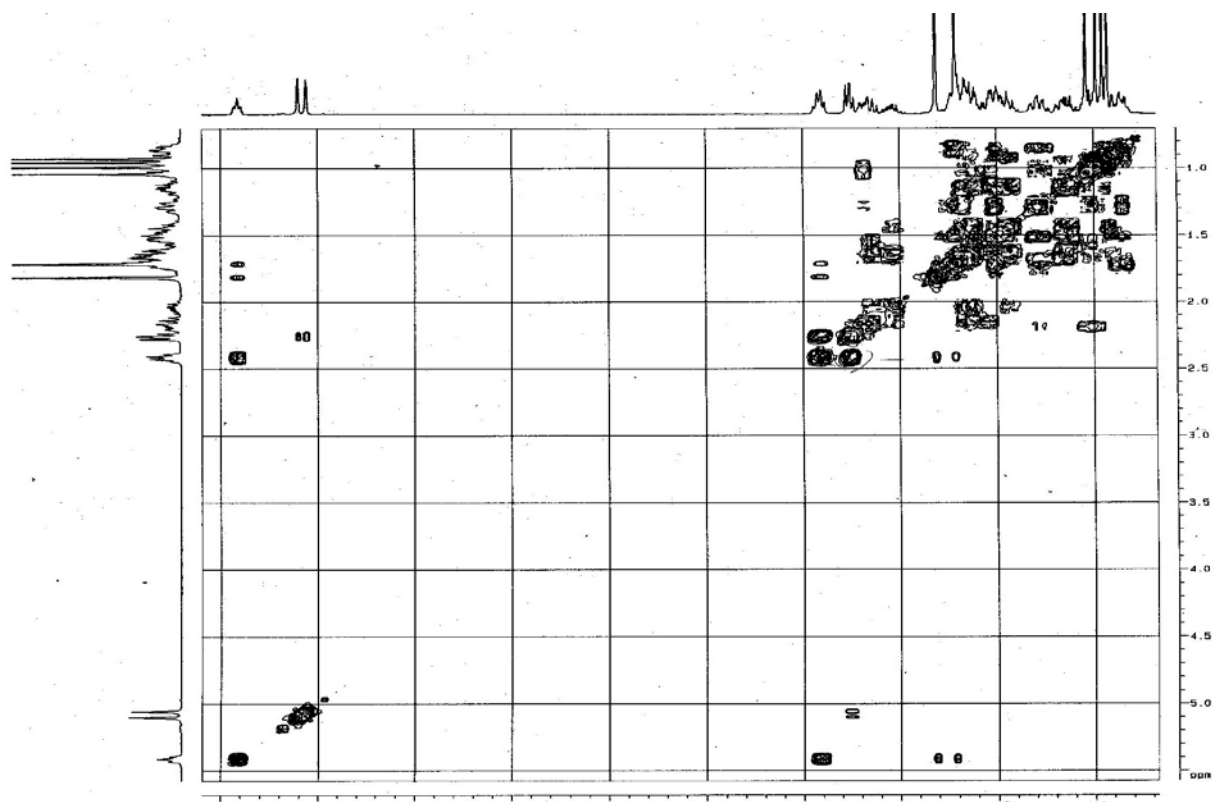


Fig. S8.4. ^1H - ^1H COSY of product 27 (400 MHz, C_6D_6).

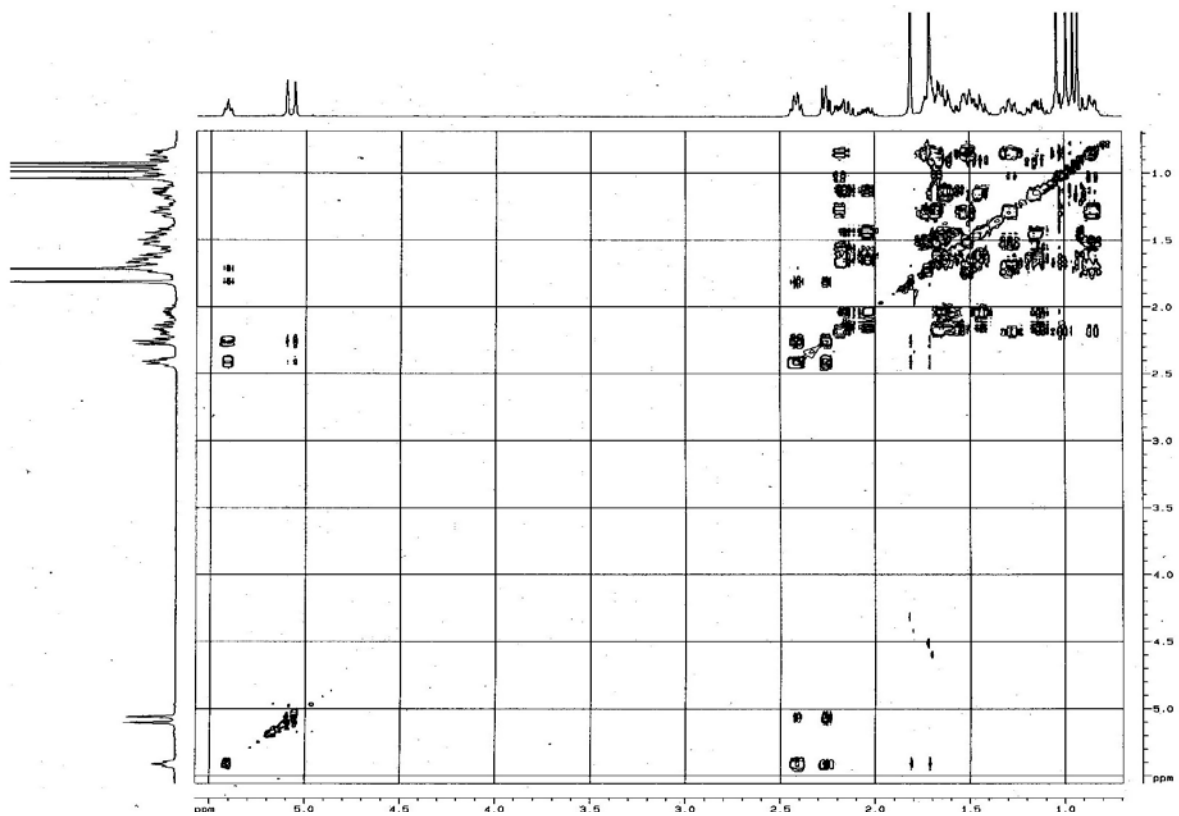


Fig. S8.5. TOCSY of **27** (400 MHz, C₆D₆).

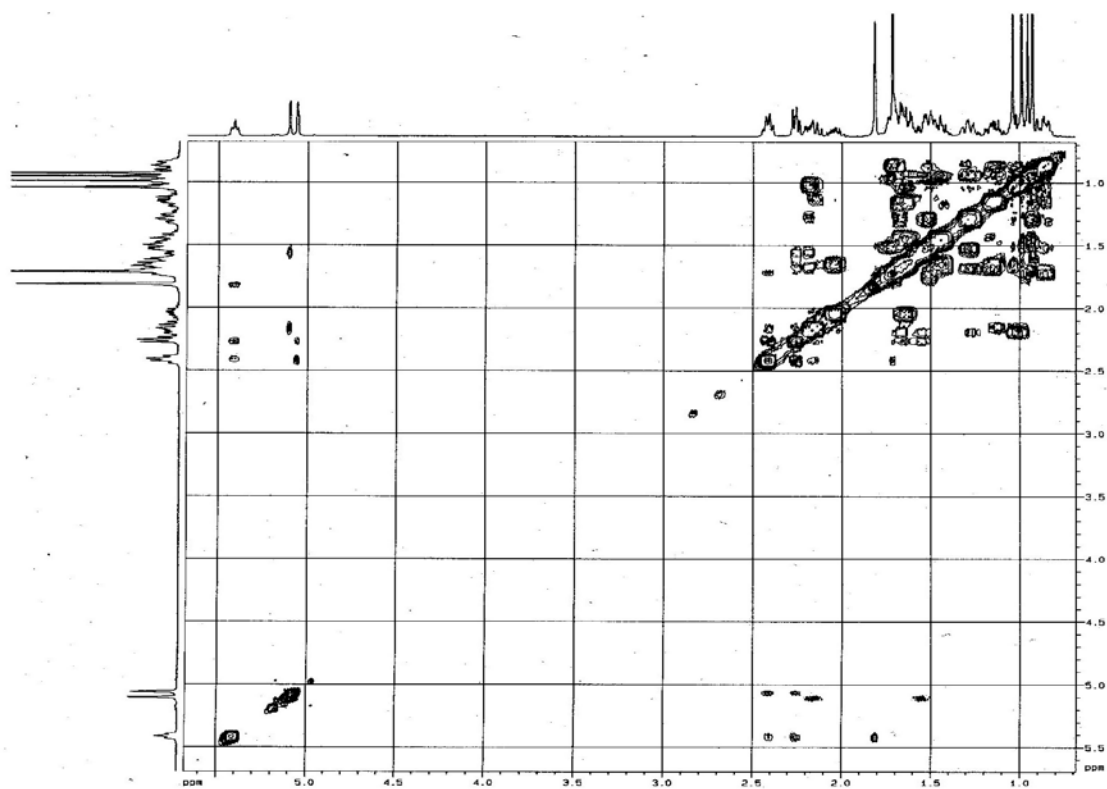


Fig. S8.6. NOESY of **27** (400 MHz, C₆D₆).

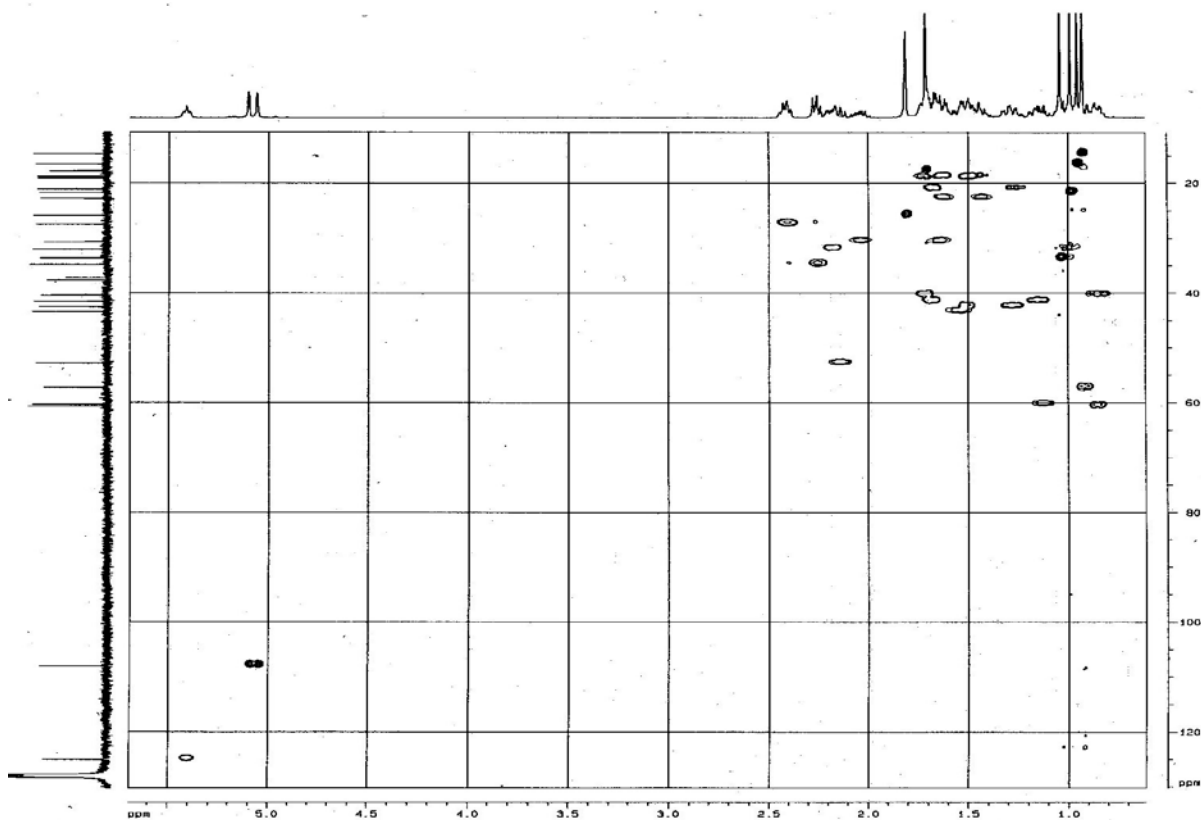


Fig. S8.7. HSQC of **27** (400 MHz, C_6D_6).

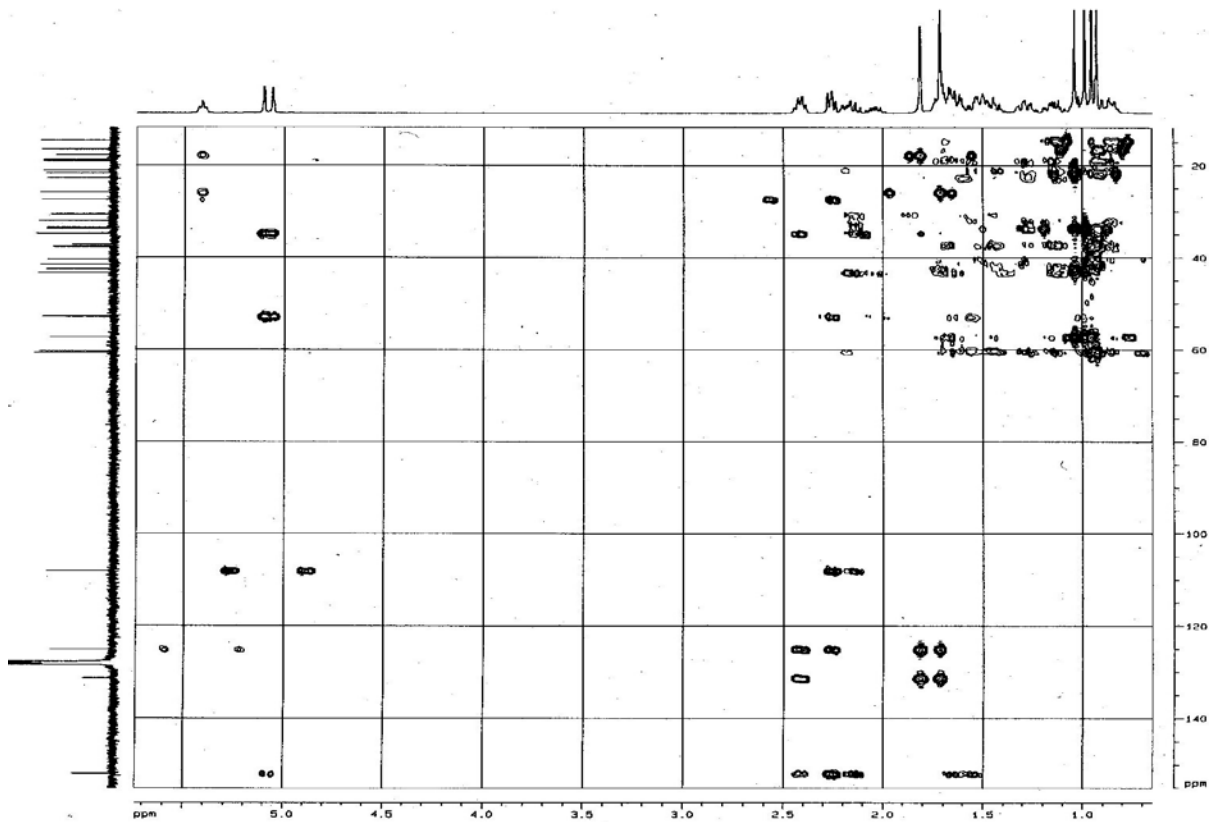


Fig. S8.8. HMBC of **27** (400 MHz, C_6D_6).

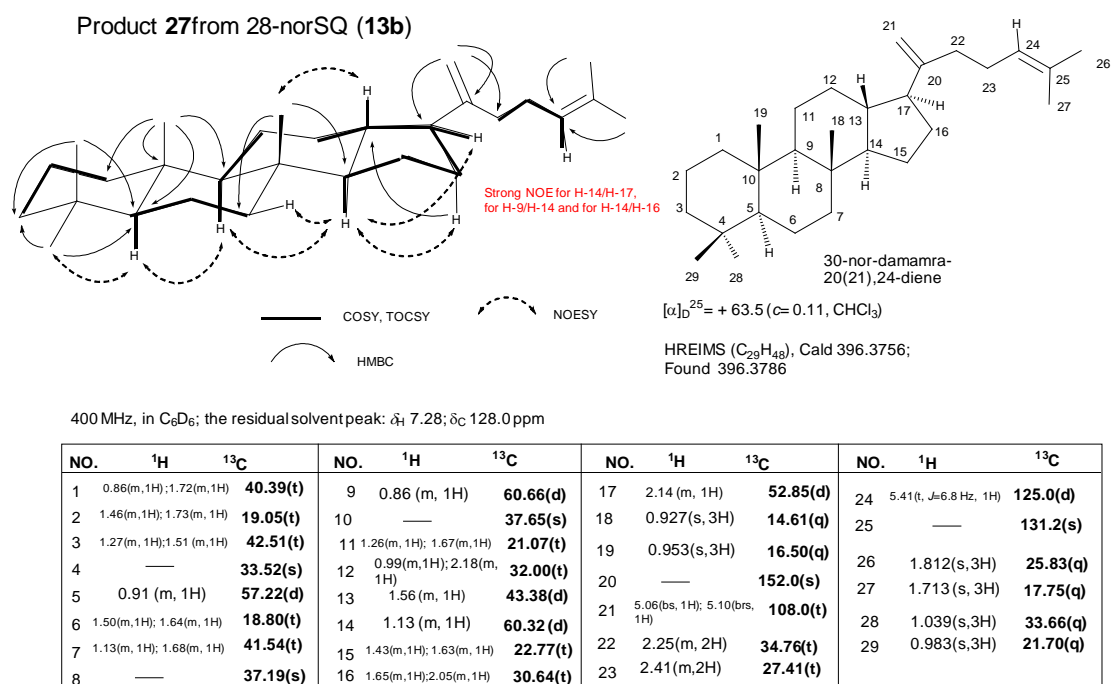


Fig. S8.9. Assignments of NMR data of product **27** in C_6D_6 , HR-EIMS and optical rotation.

9. EIMS and NMR data of product 28.

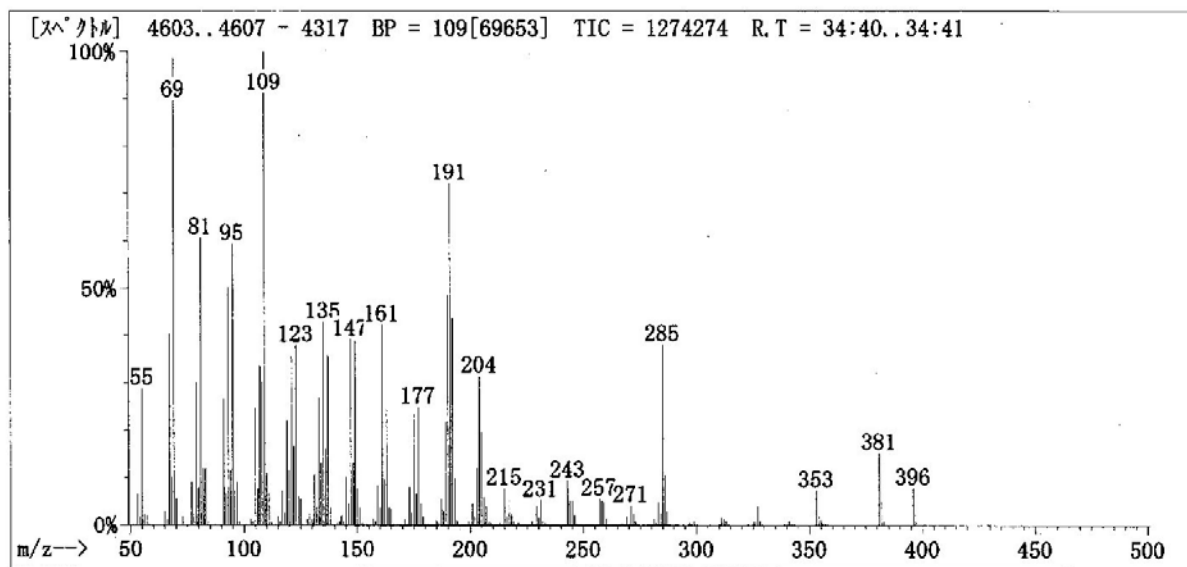


Fig. S9.1. EIMS of product 28.

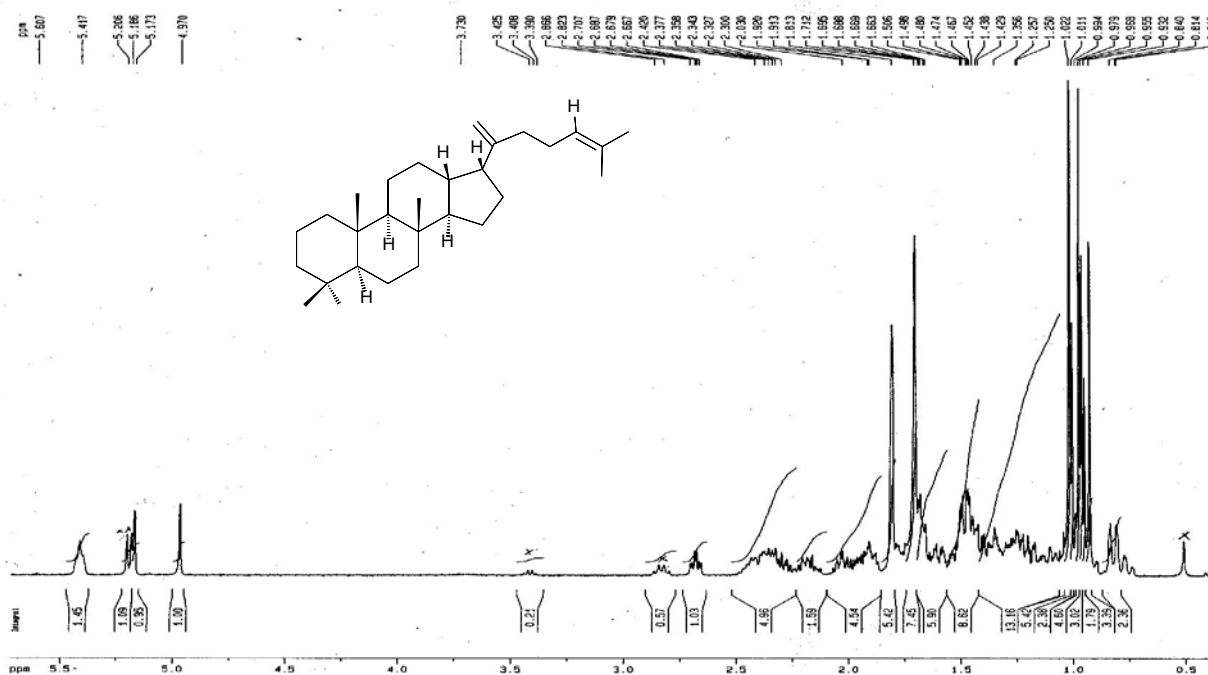


Fig. S9.2. ¹H NMR of Product 28 (400 MHz, C₆D₆).

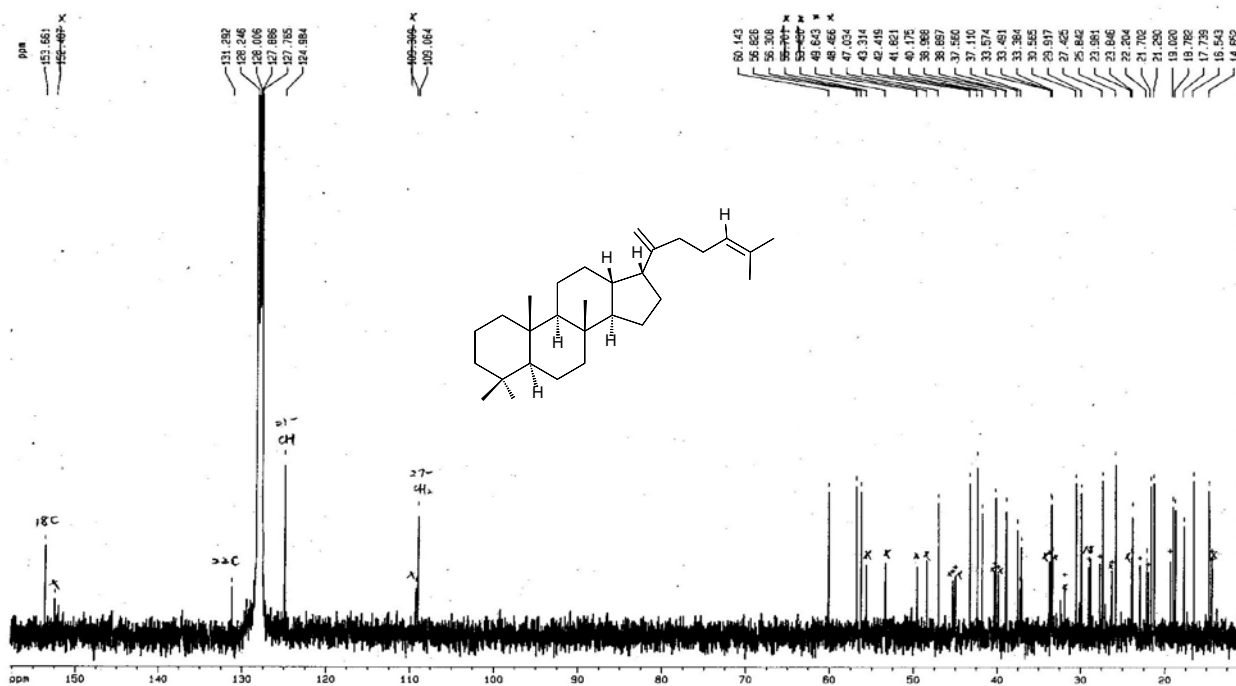


Fig. S9.3. ^{13}C NMR of Product 28 (100 MHz, C_6D_6).

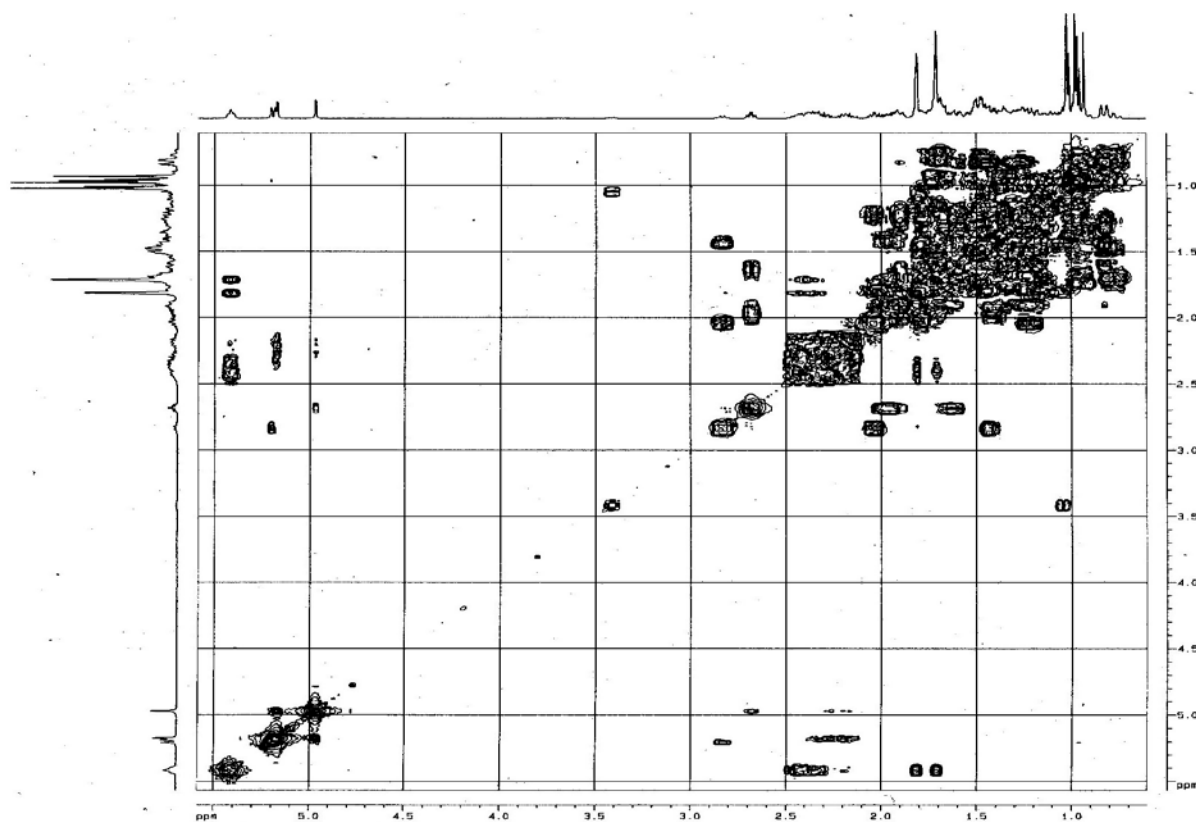


Fig. S9.4. ^1H - ^1H COSY of product 28 (400 MHz, C_6D_6).

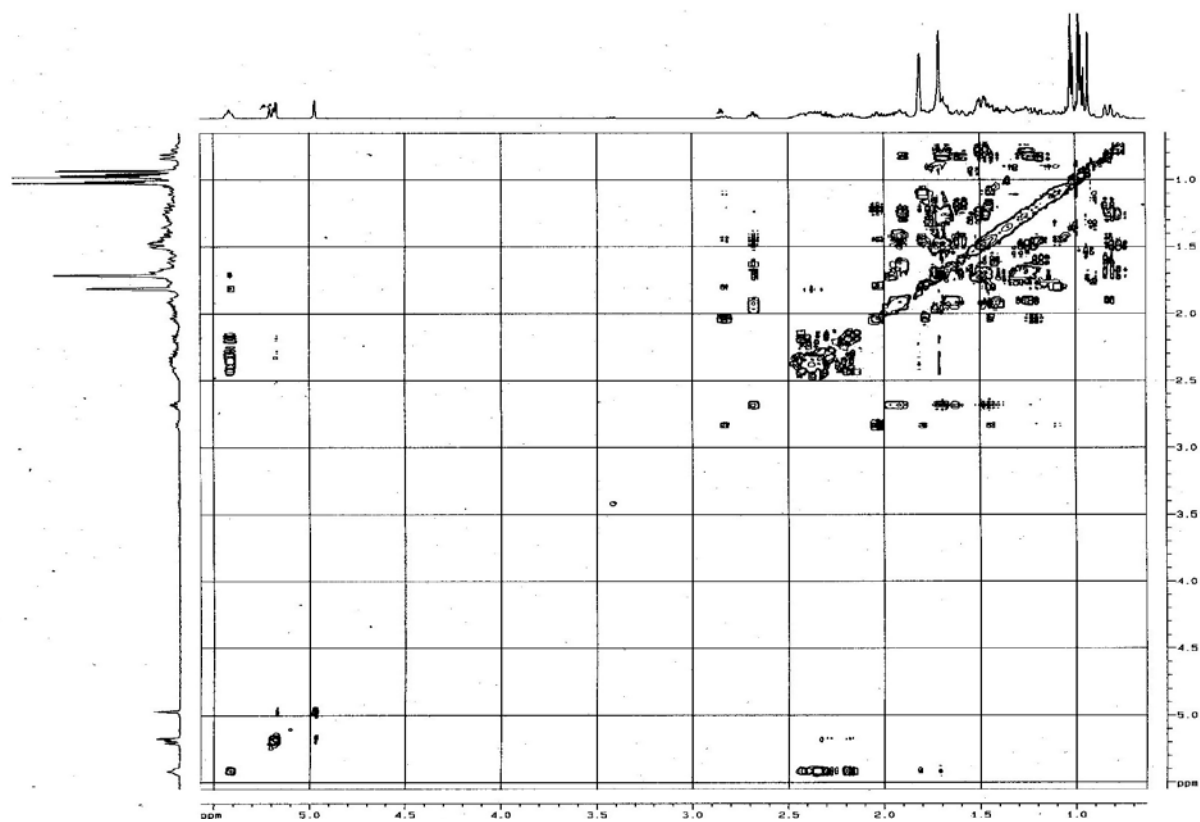


Fig. S9.5. TOCSY of **28** (400 MHz, C₆D₆).

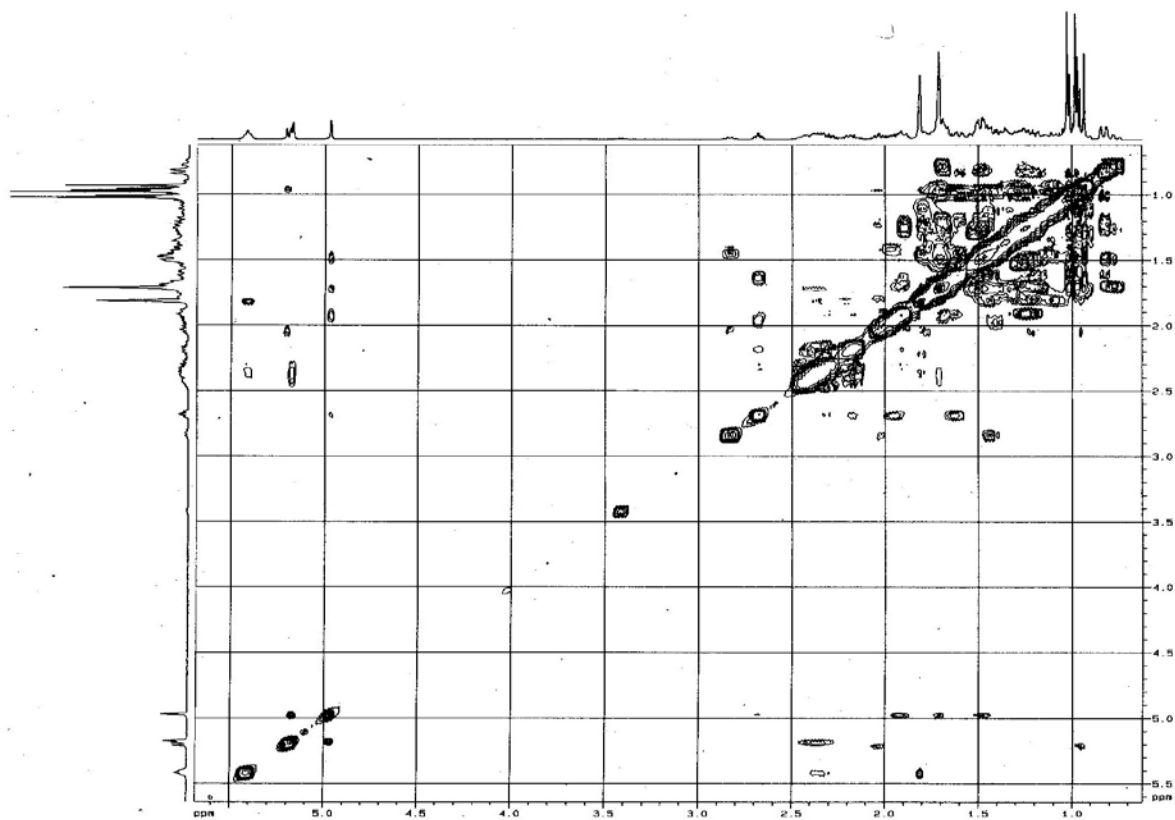


Fig. S9.6. NOESY of **28** (400 MHz, C₆D₆).

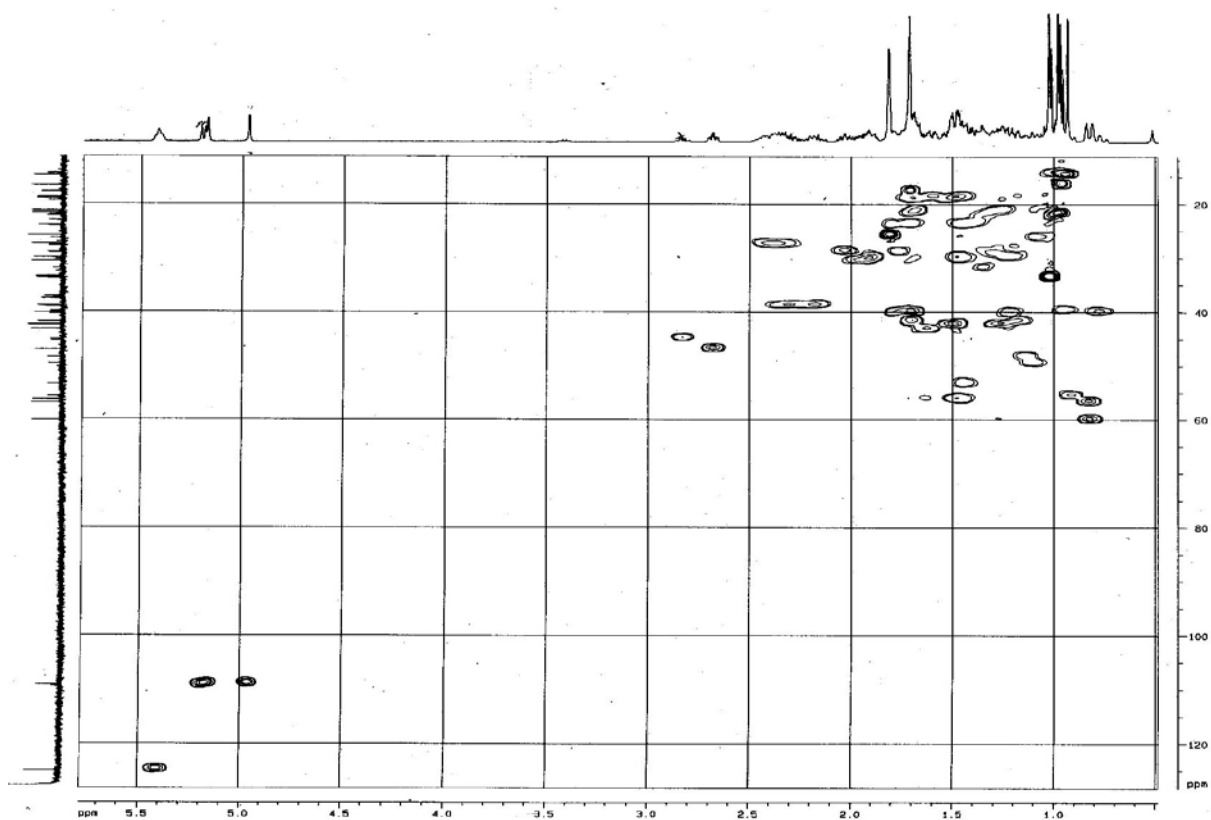


Fig. S9.7. HSQC of **28** (400 MHz, C₆D₆).

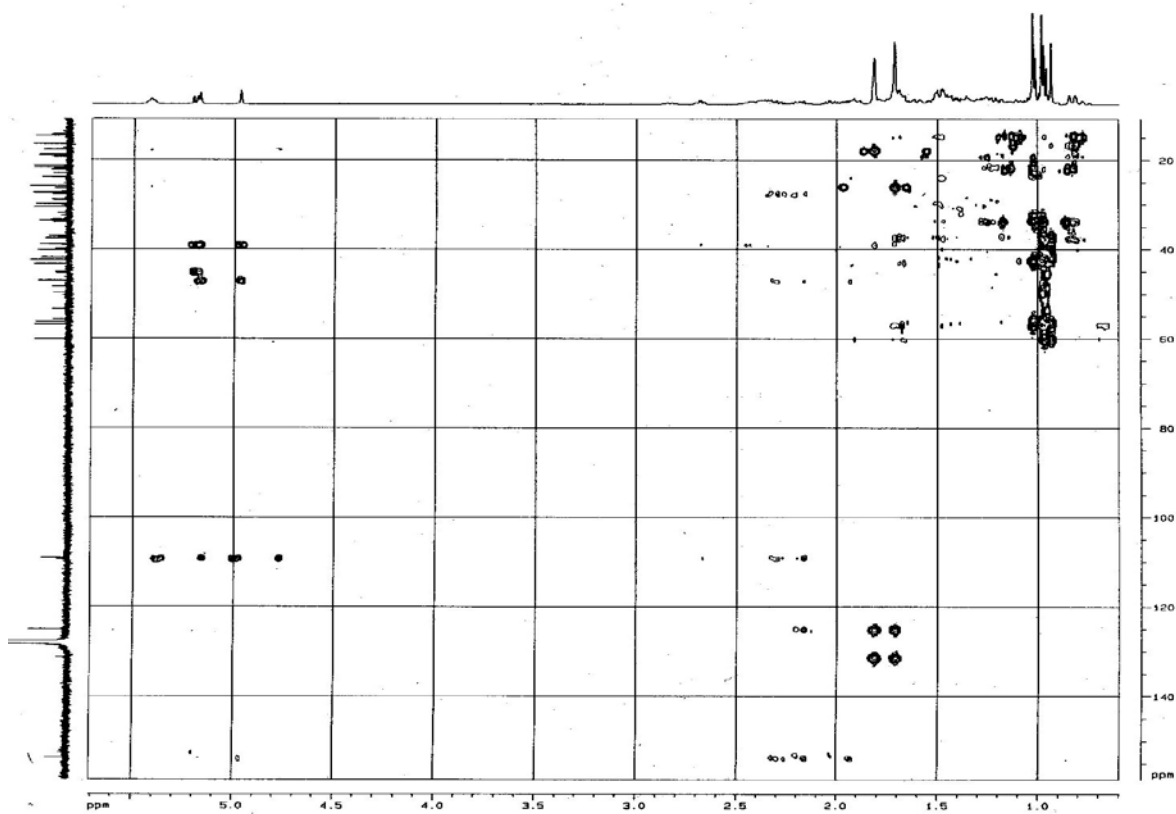
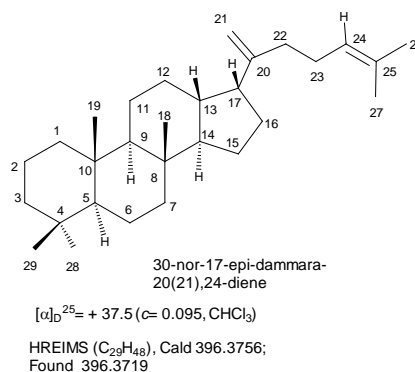
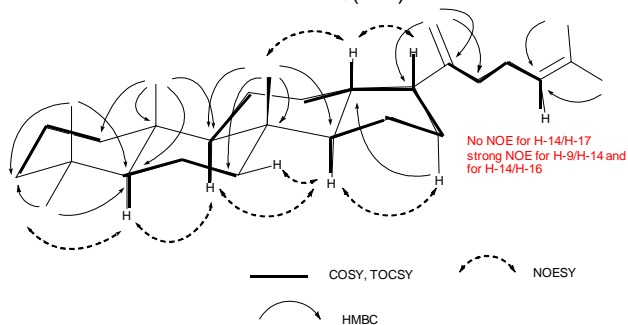


Fig. S9.8. HMBC of **28** (400 MHz, C₆D₆).

Product **28** from 28-norSQ (**13b**)



400 MHz, in C_6D_6 ; the residual solvent peak: δ_H 7.28; δ_C 128.0 ppm

NO.	^1H	^{13}C	NO.	^1H	^{13}C	NO.	^1H	^{13}C	NO.	^1H	^{13}C
1	0.78(m,1H);1.70(m,1H)	40.18(t)	9	0.82 (m, 1H)	60.14(d)	17	2.68 (dt, $J=3.2, 8.0\text{Hz}$, 1H)	47.03(d)	24	5.42(t, $J=6.8\text{Hz}$, 1H)	125.0(d)
2	1.47(m,1H); 1.69(m, 1H)	19.02(t)	10	—	37.56(s)	18	0.932(s,3H)	14.65(q)	25	—	131.3(s)
3	1.27(m, 1H);1.49 (m,1H)	42.42(t)	11	1.26(m, 1H); 1.67(m,1H)	21.29(t)	19	0.968(s,3H)	16.54(q)	26	1.813(s,3H)	25.84(q)
4	—	33.49(s)	12	1.22(m,1H);1.91(m, 1H)	29.92(t)	20	—	153.7(s)	27	1.712 (s, 3H)	17.74(q)
5	0.93 (m, 1H)	56.83(d)	13	1.62 (m, 1H)	43.31(d)	21	4.97(bs, 1H); 5.17(bvs, 1H)	109.0(t)	28	1.022(s,3H)	33.57(q)
6	1.43(m,1H); 1.57(m, 1H)	18.78(t)	14	1.47 (m, 1H)	56.31 (d)	22	2.17(m, 1H);2.32(m, 1H)	38.90(t)	29	0.979(s,3H)	21.70(q)
7	1.16(m, 1H); 1.69(m, 1H)	41.82(t)	15	1.43(m,1H); 1.76(m, 1H)	23.84(t)	23	2.38(m,2H)	27.42(t)			
8	—	37.11(s)	16	1.68(m,1H);1.98(m, 1H)	30.56(t)						

Fig. S9.9. Assignments of NMR data of product **28** in C_6D_6 , HR-EIMS and optical rotation.

10. EIMS and NMR data of product 29.

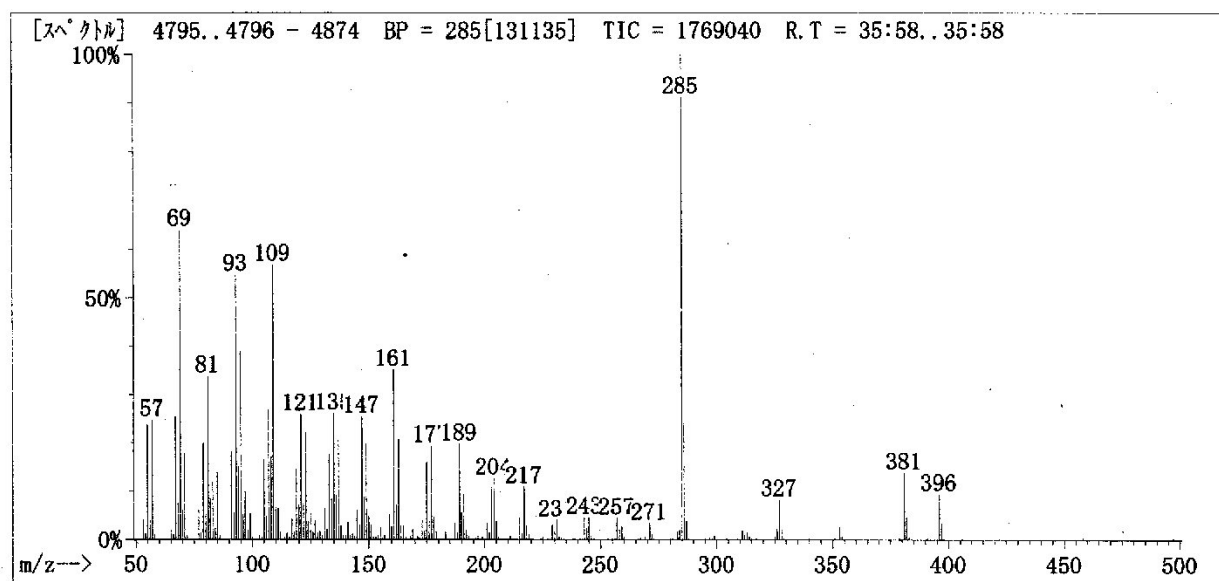


Fig. S10.1. EIMS of product 29.

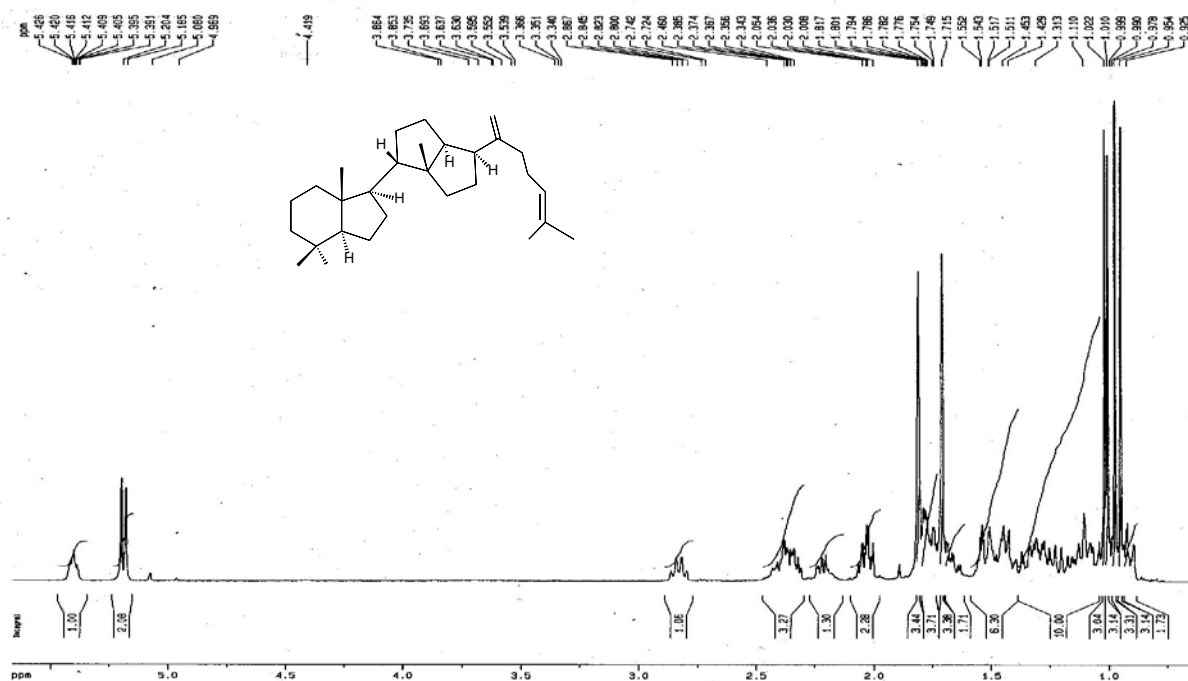


Fig. S10.2. ^1H NMR of Product 29 (400 MHz, C_6D_6).

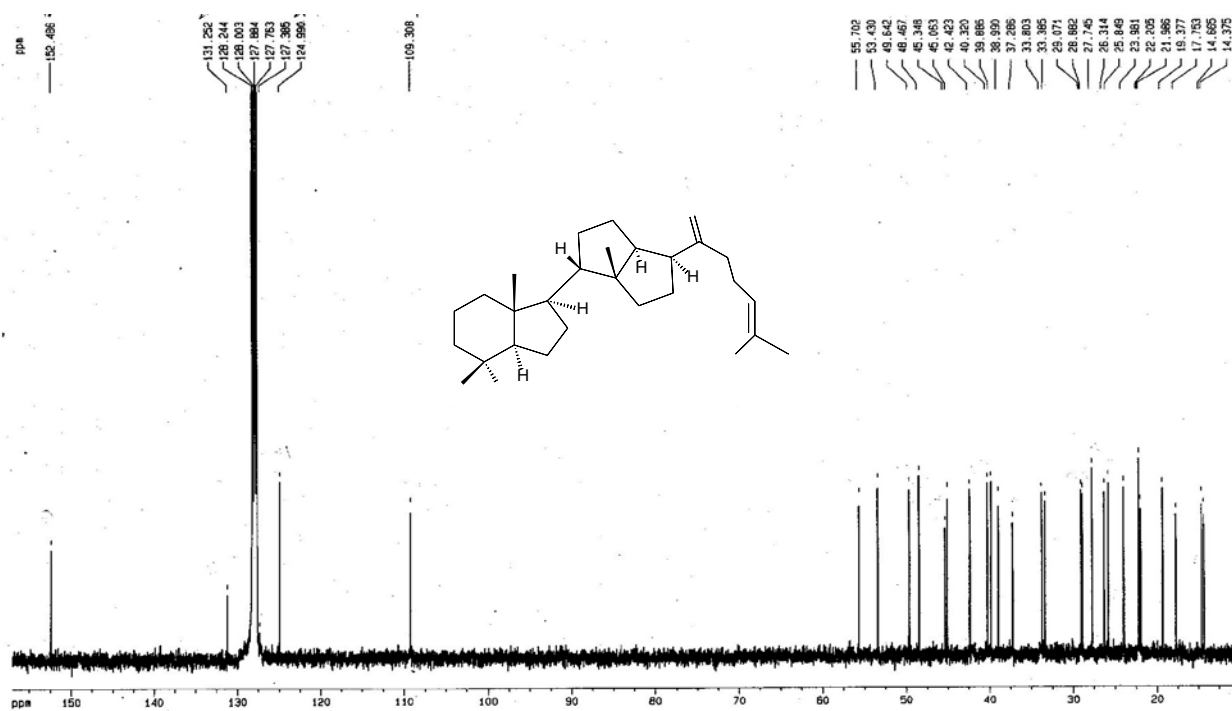


Fig. S10.3. ^{13}C NMR of Product 29 (100 MHz, C_6D_6).

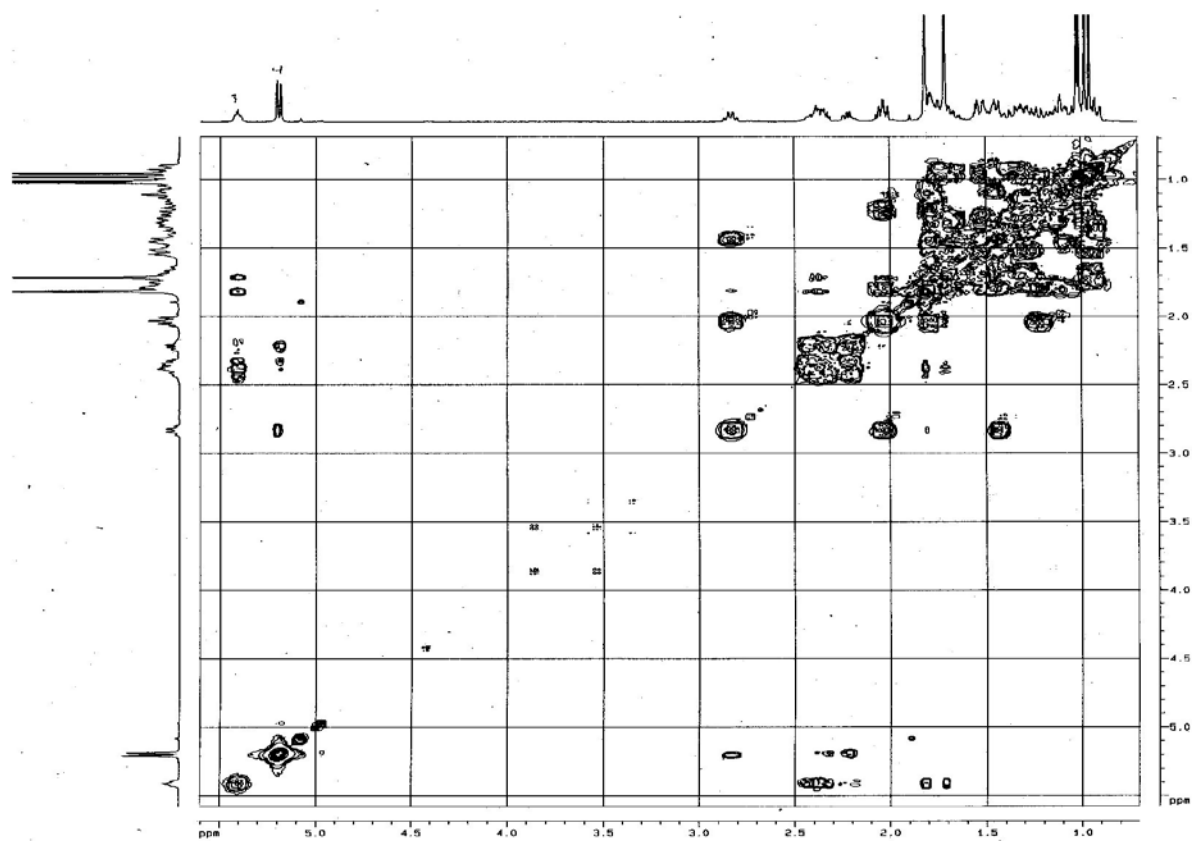


Fig. S10.4. ^1H - ^1H COSY of product 29 (400 MHz, C_6D_6).

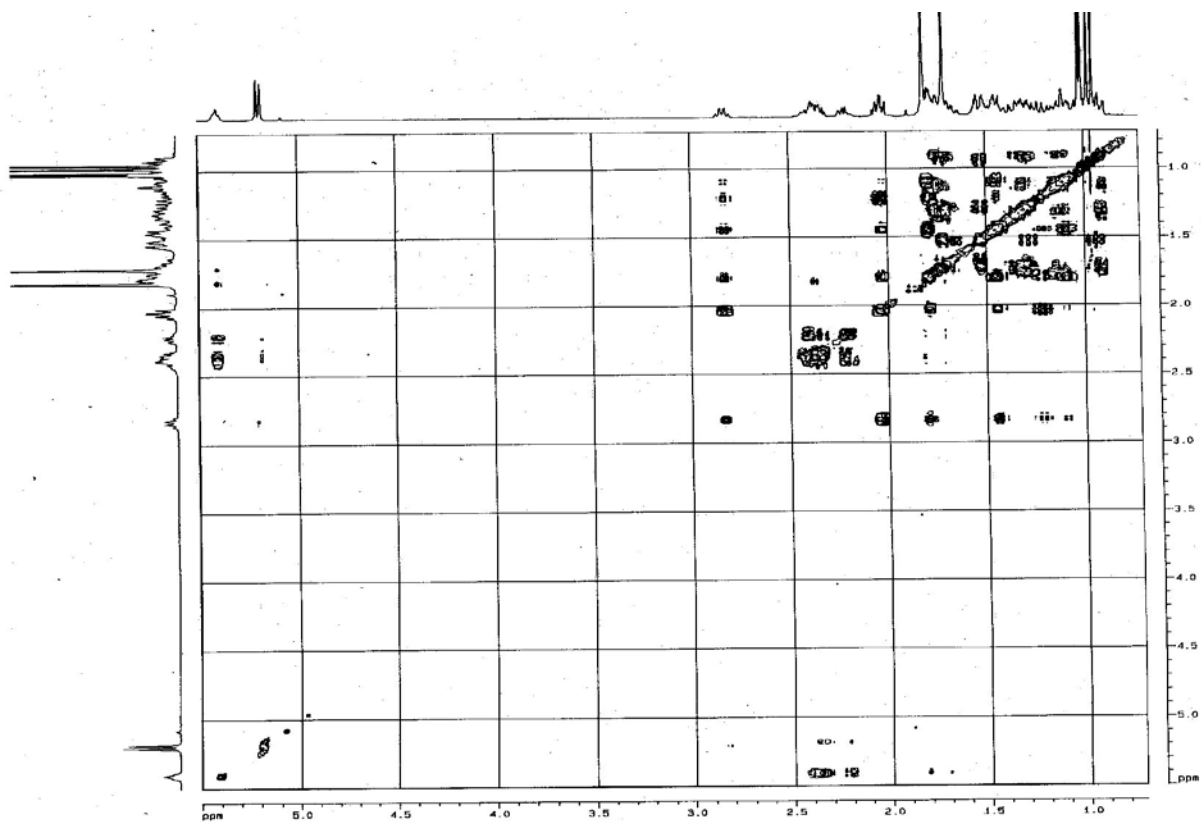


Fig. S10.5. TOCSY of **29** (400 MHz, C_6D_6).

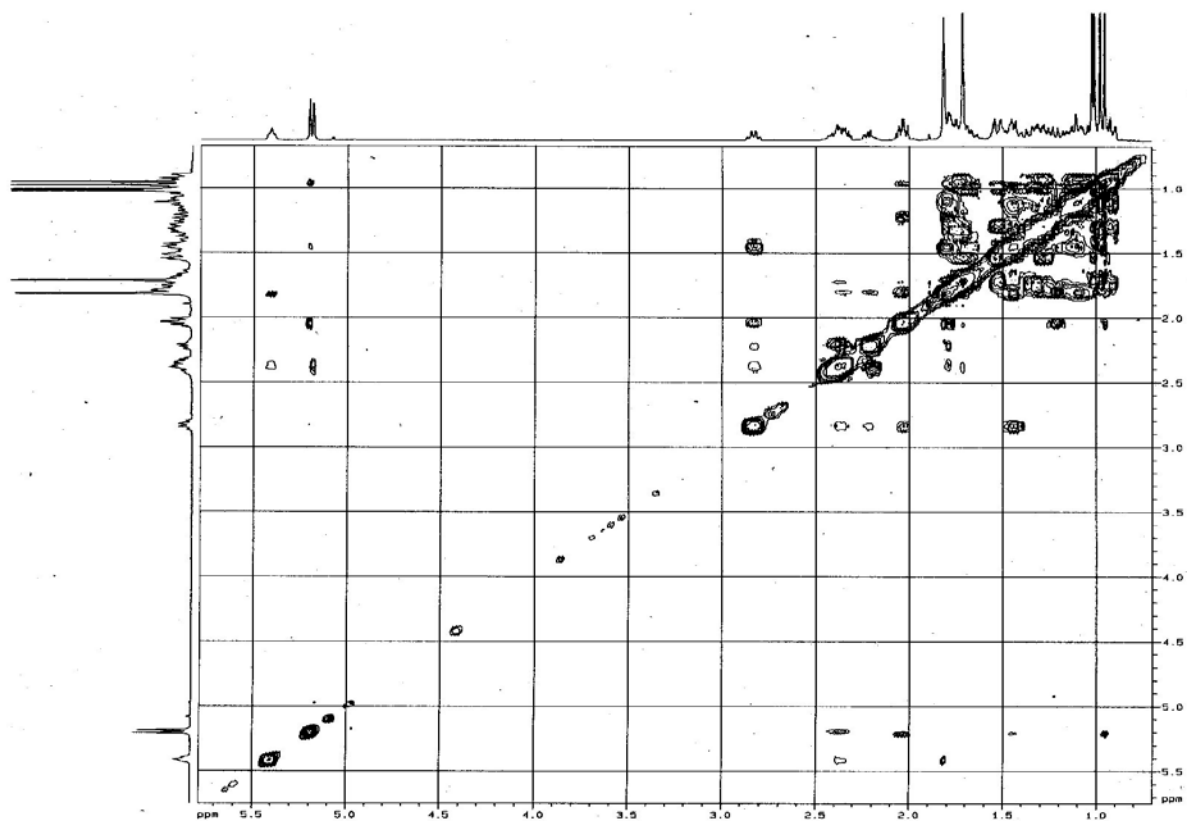


Fig. S10.6. NOESY of **29** (400 MHz, C_6D_6).

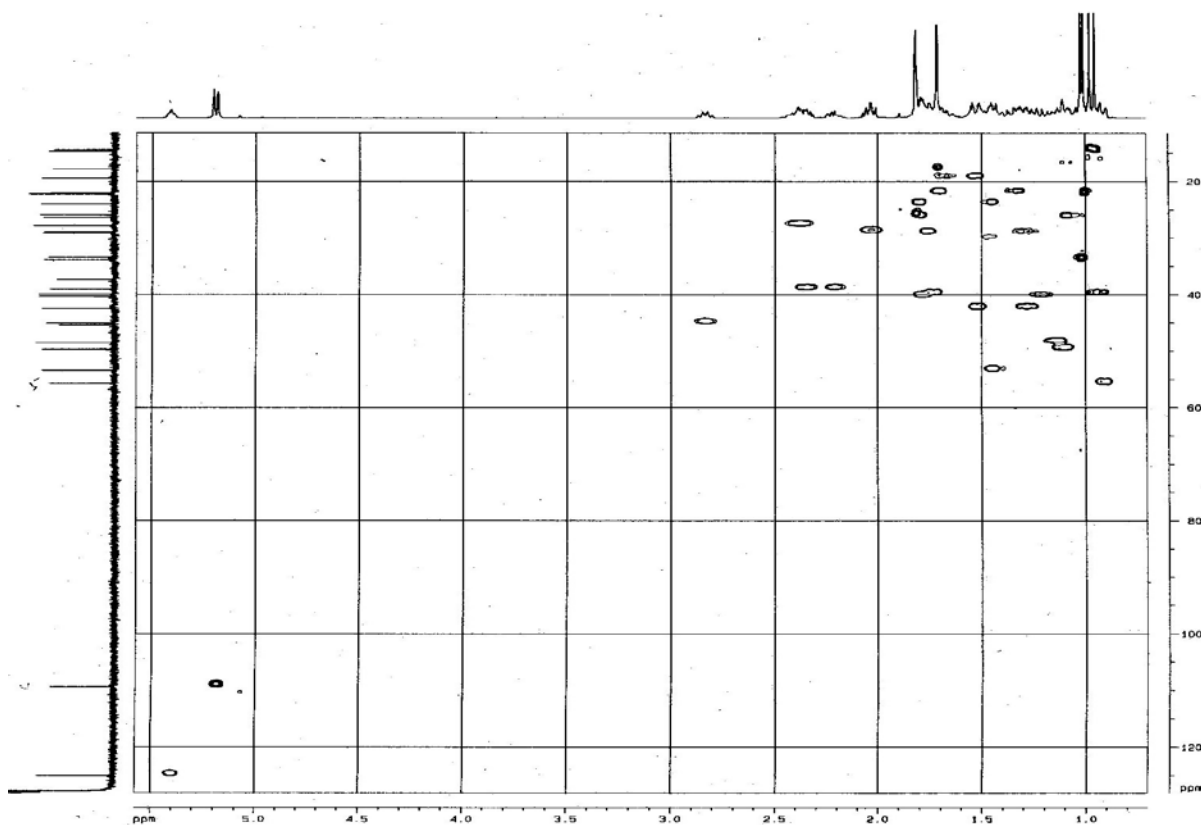


Fig. S10.7. HSQC of **29** (400 MHz, C₆D₆).

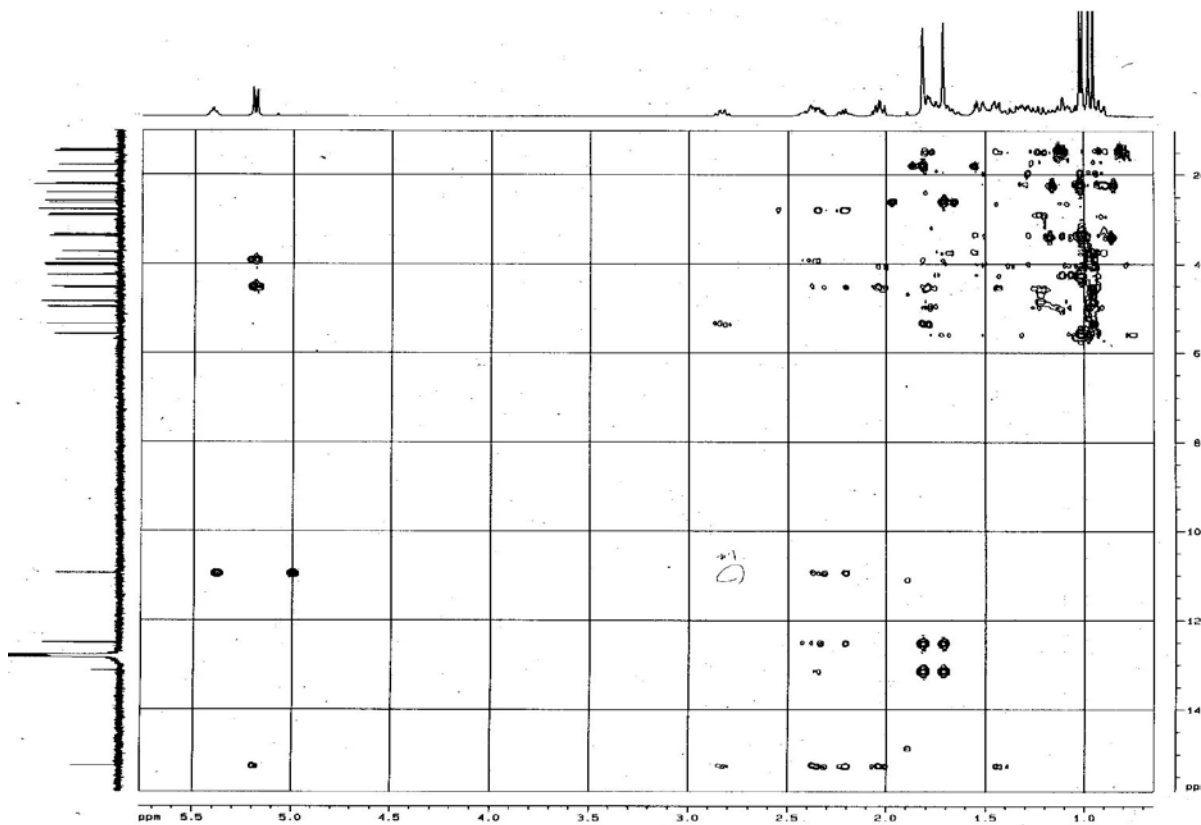
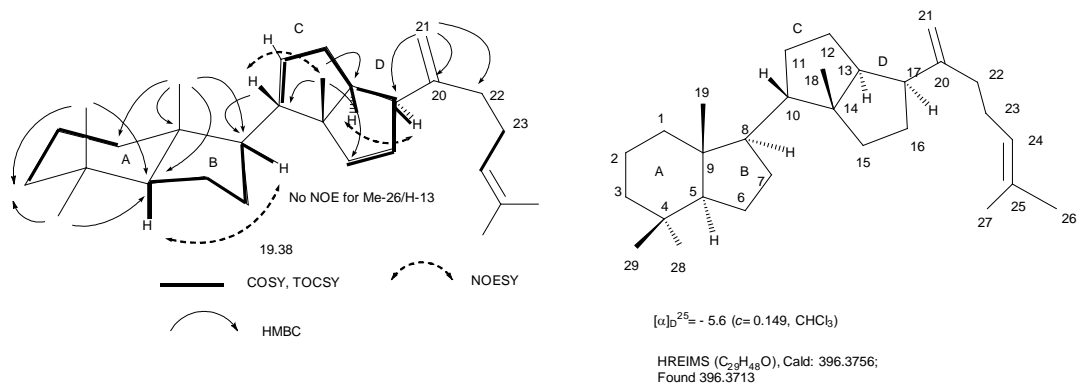


Fig. S10.8. HMBC of **29** (400 MHz, C₆D₆).

Product **29** from 27-norSQ **13a**



400 MHz, in C_6D_6 ; the residual solvent peak: δ_{H} 7.28; δ_{C} 128.0 ppm

NO.	^1H	^{13}C	NO.	^1H	^{13}C	NO.	^1H	^{13}C	NO.	^1H	^{13}C
1	0.94(m,1H);1.71(m,1H)	39.89(t)	9	—	37.28(s)	17	2.83(dt, J=9.0 Hz, 1H)	45.06(d)	25	—	131.3(s)
2	1.54 (m, 1H);1.67(m, 1H)	19.38(t)	10	1.14(m, 1H)	48.47(d)	18	0.954(s,3H)	14.66(q)	26	1.817(s,3H)	25.85(q)
3	1.27 (m, 1H); 1.52(m,1H)	42.42(t)	11	1.07(m, 1H); 1.78(m,1H)	26.31(t)	19	0.978(s,3H)	14.37(q)	27	1.715 (s, 3H)	17.75(q)
4	—	33.38(s)	12	1.45(m,1H); 1.80(m,1H)	23.98(t)	20	—	152.5(s)	28	1.022(s,3H)	33.80(q)
5	0.91(m, 1H)	55.70(d)	13	1.45 (m, 1H)	53.43(d)	21	5.18 (1H, s); 5.20(s, 1H)	109.3(t)	29	1.010(s,3H)	22.20(q)
6	1.34(m,1H); 1.71(m, 1H)	21.98(t)	14	—	45.35(s)	22	2.21(m); 2.34(m)	38.99(t)			
7	1.30(m, 1H); 1.75(m, 1H)	29.07(t)	15	1.22(m,1H); 1.77(m, 1H)	40.32(t)	23	2.36(m,2H)	27.74(t)			
8	1.10 (m, 1H)	49.64(d)	16	2.03(m,2H)	28.88(t)	24	5.41(t, J=6.8 Hz, 1H)	125.0(d)			

Fig. S10.9. Assignments of NMR data of product **29** in C_6D_6 , HR-EIMS and optical rotation.

11. EIMS and NMR data of product 30.

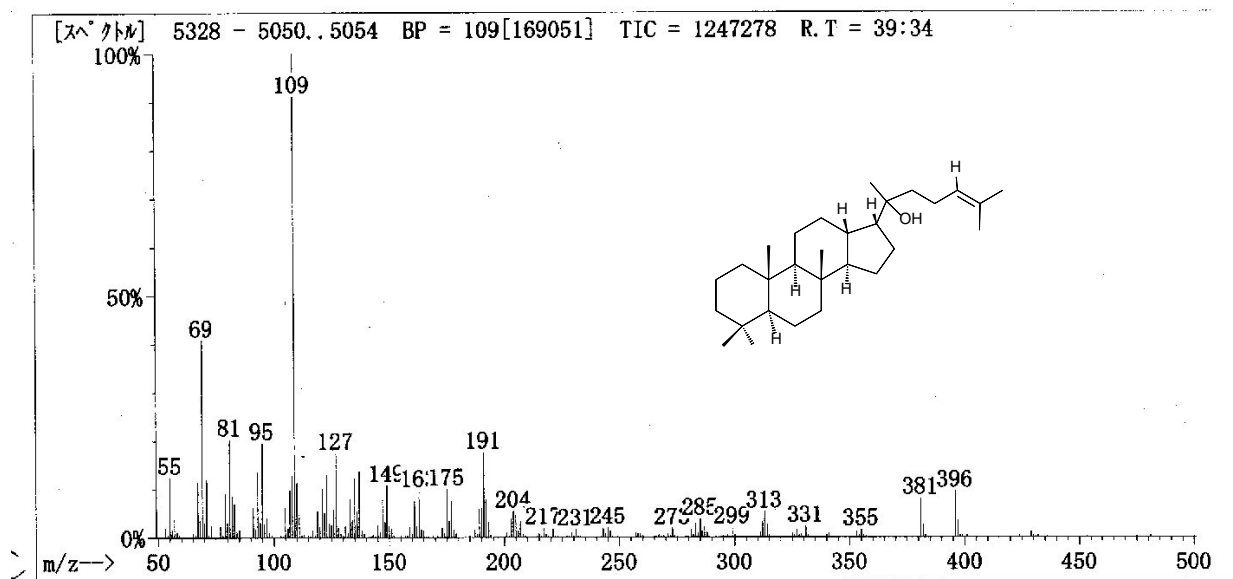


Fig. S11.1. EIMS of product 30.

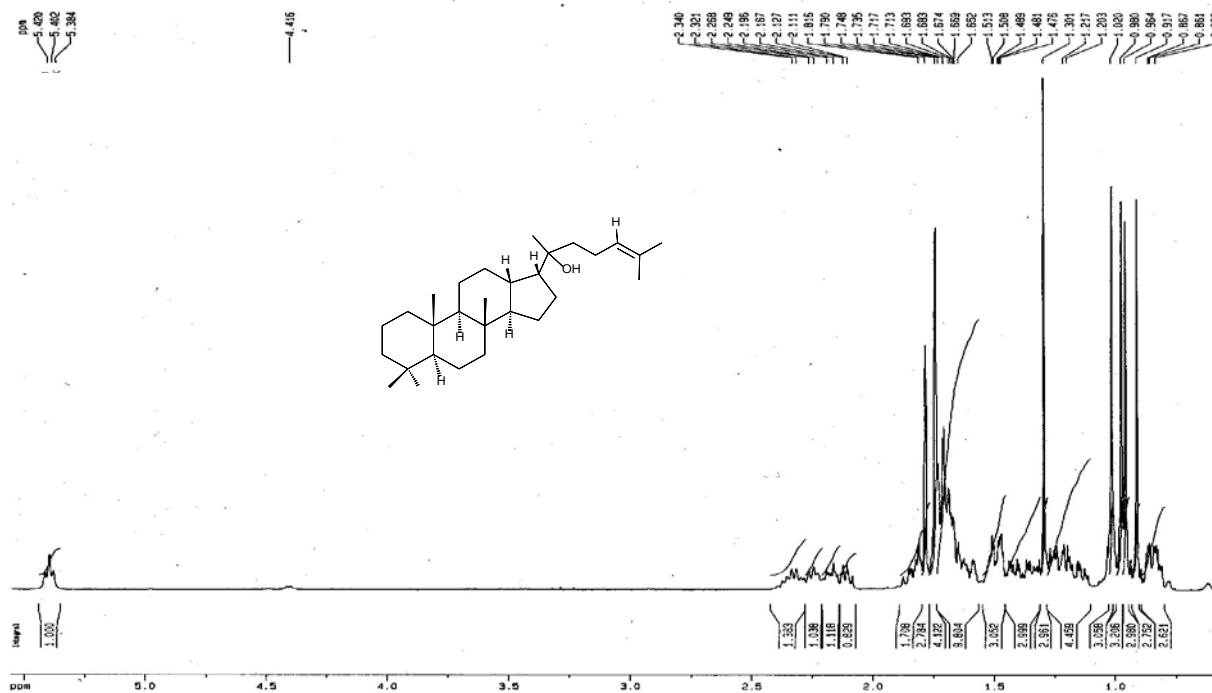


Fig. S11.2. ¹H NMR of Product 30 (400 MHz, C₆D₆).

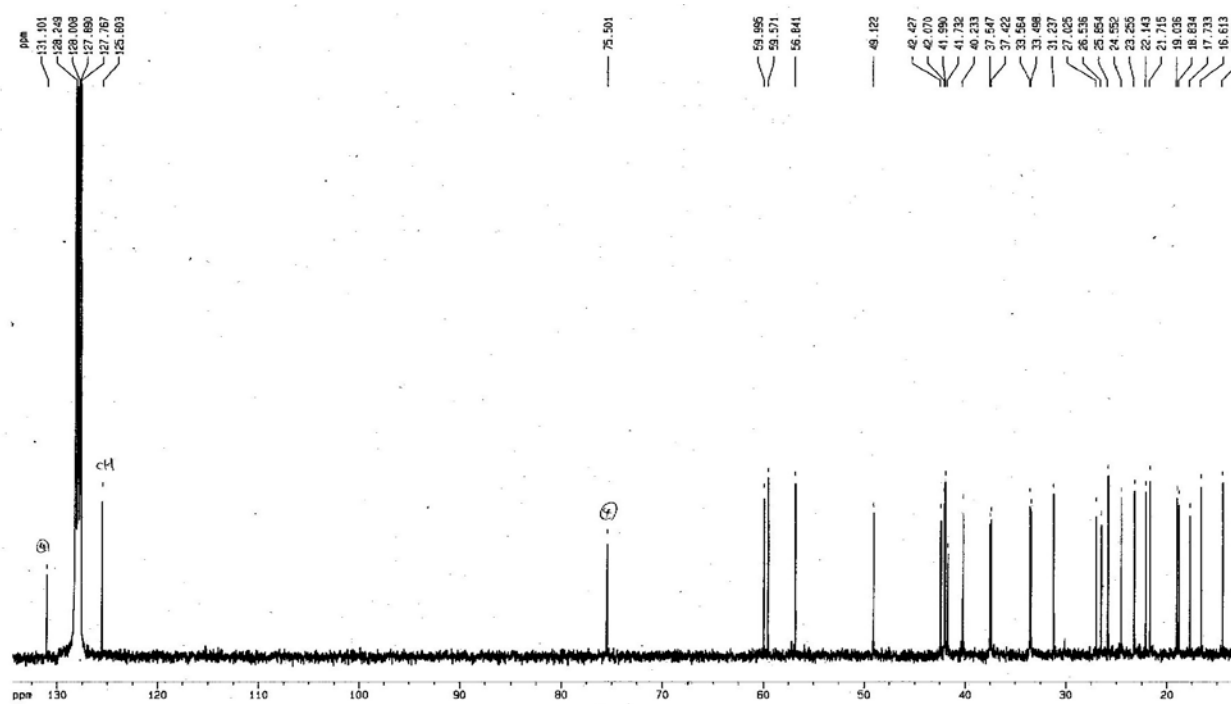


Fig. S11.3. ^{13}C NMR of Product **30** (100 MHz, C_6D_6).

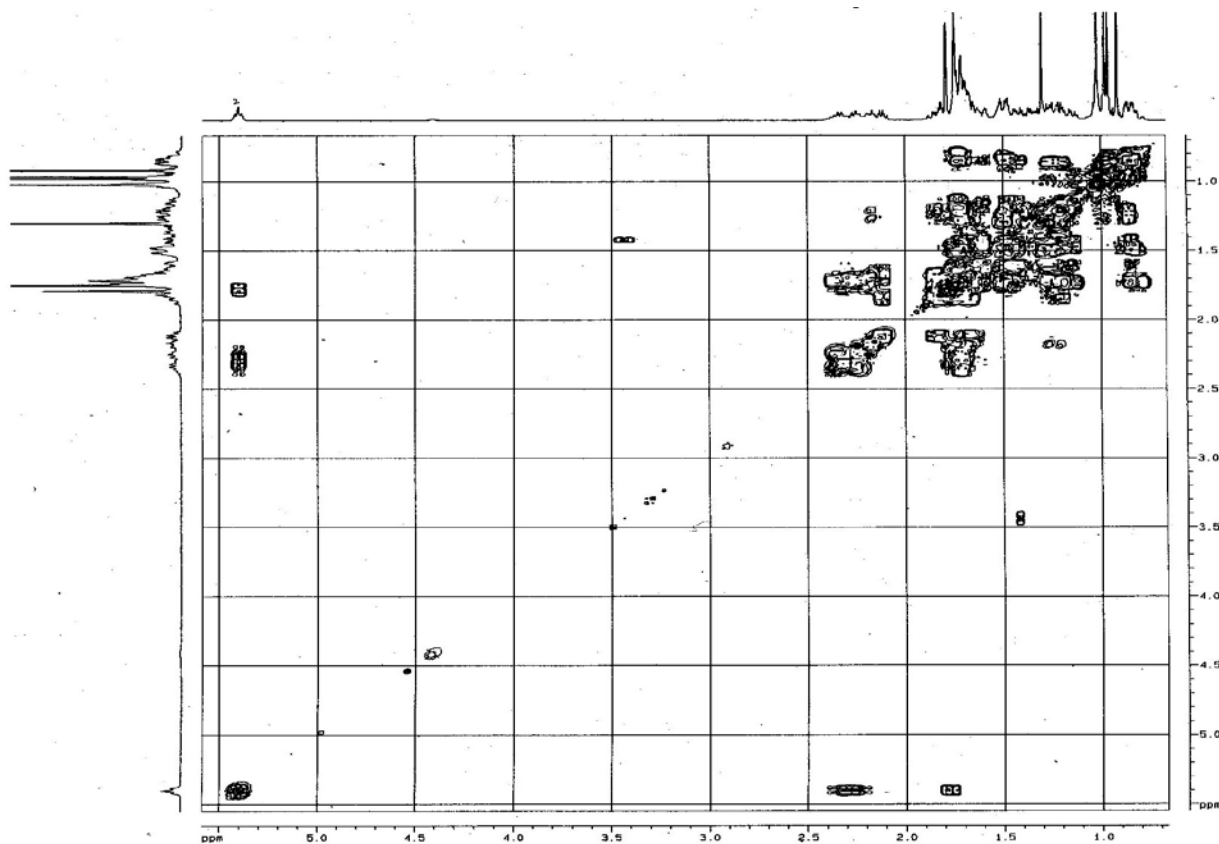


Fig. S11.4. ^1H - ^1H COSY of product **30** (400 MHz, C_6D_6).

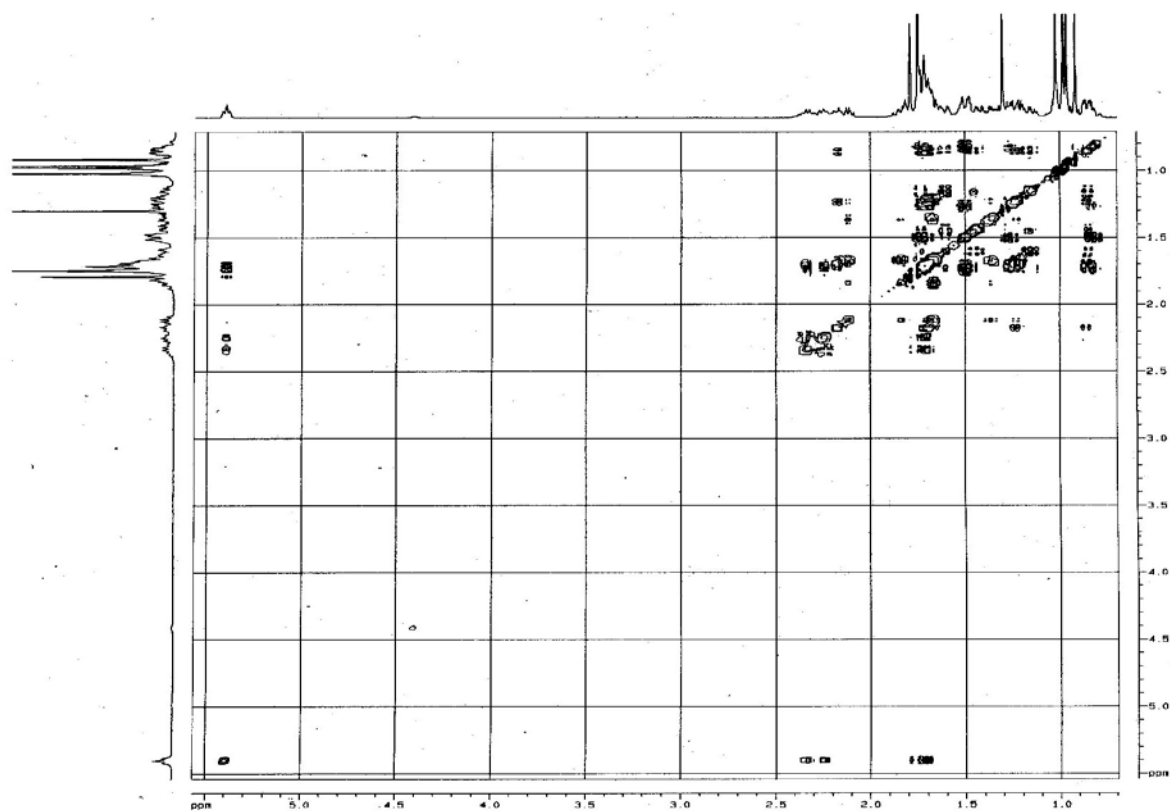


Fig. S11.5. TOCSY of **30** (400 MHz, C₆D₆).

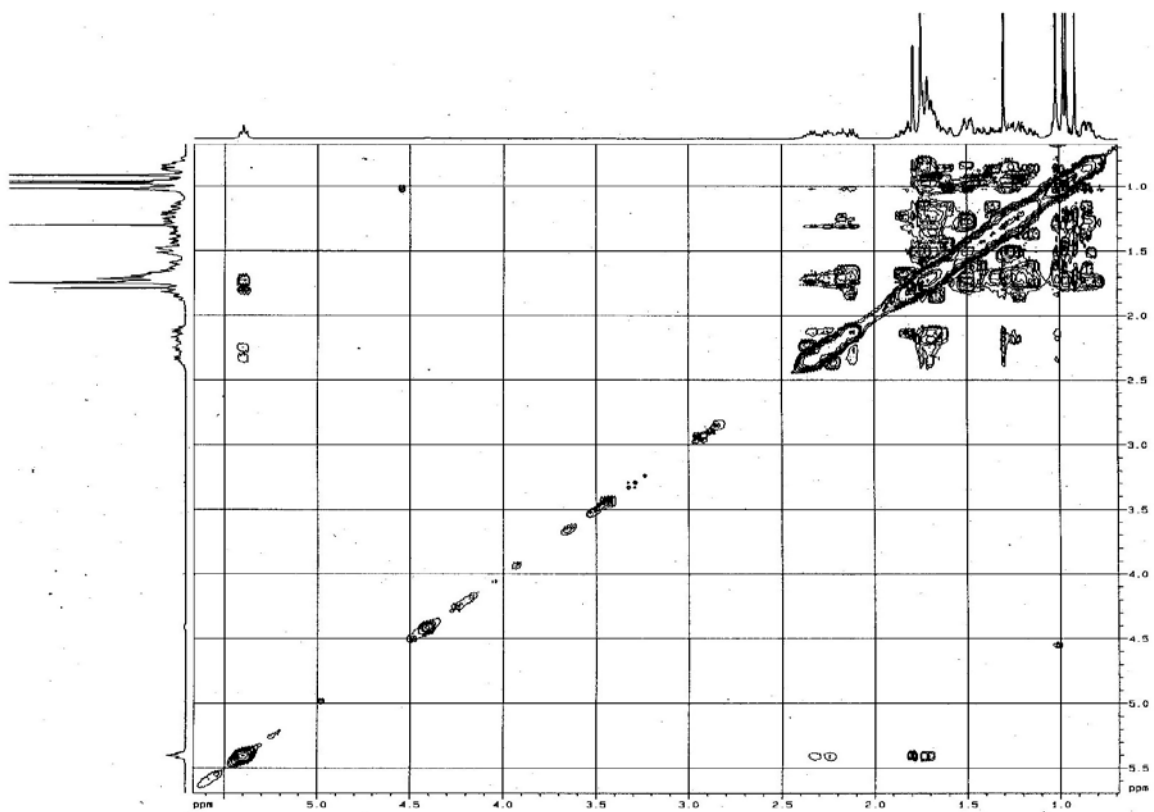


Fig. S11.6. NOESY of **30** (400 MHz, C₆D₆).

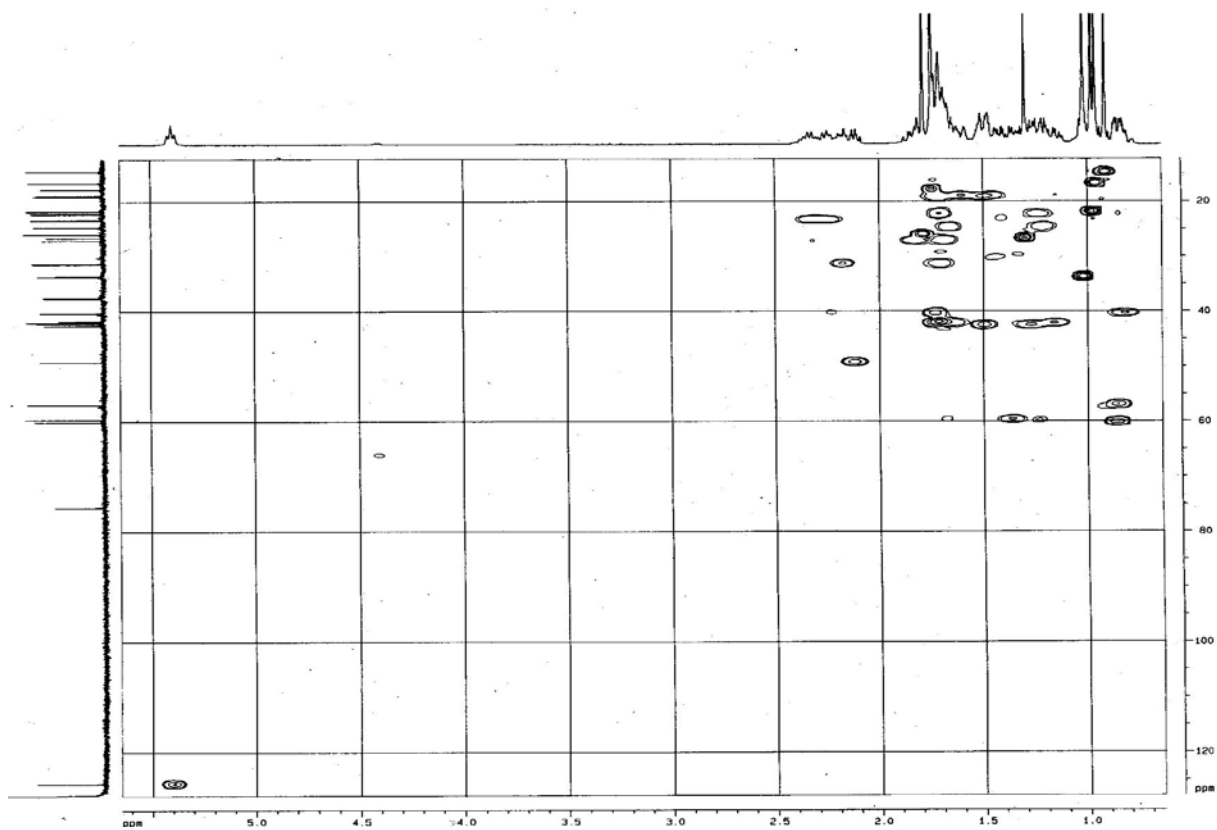


Fig. S11.7. HSQC of **30** (400 MHz, C₆D₆).

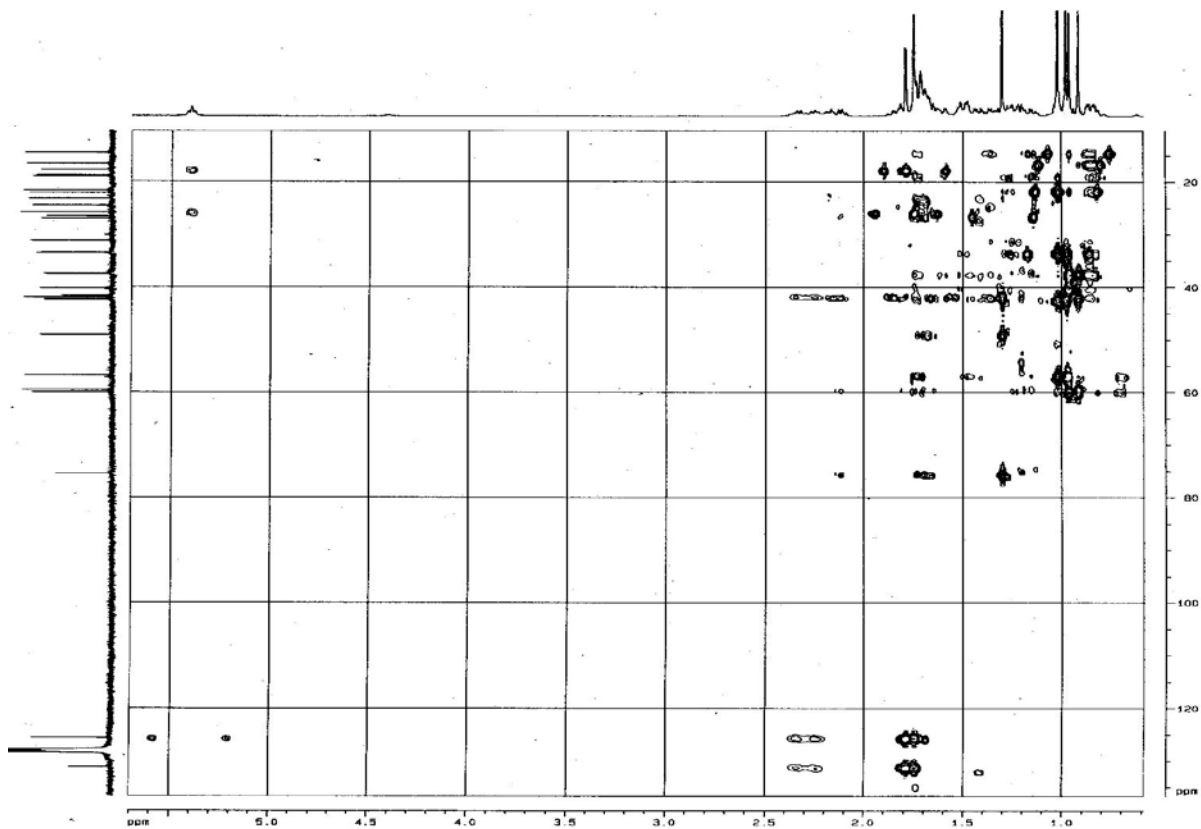


Fig. S11.8. HMBC of **30** (400 MHz, C₆D₆).

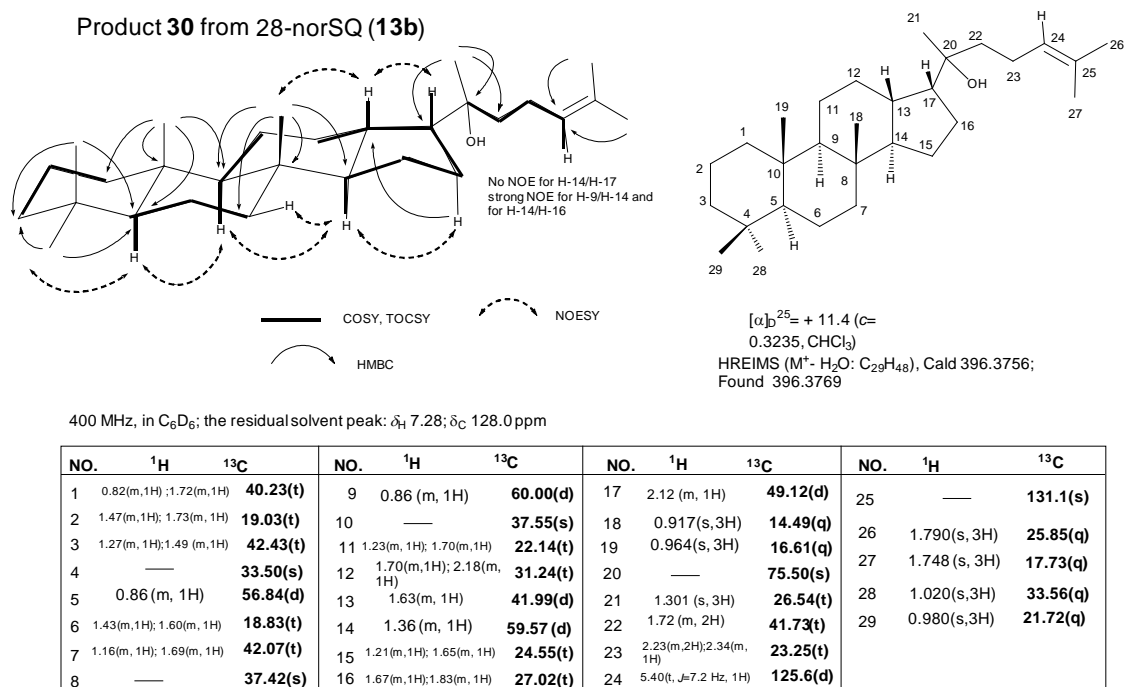


Fig. S11.9. Assignments of the NMR data of product **30** in C₆D₆, HR-EIMS and optical rotation.

12. EIMS and NMR data of product 31.

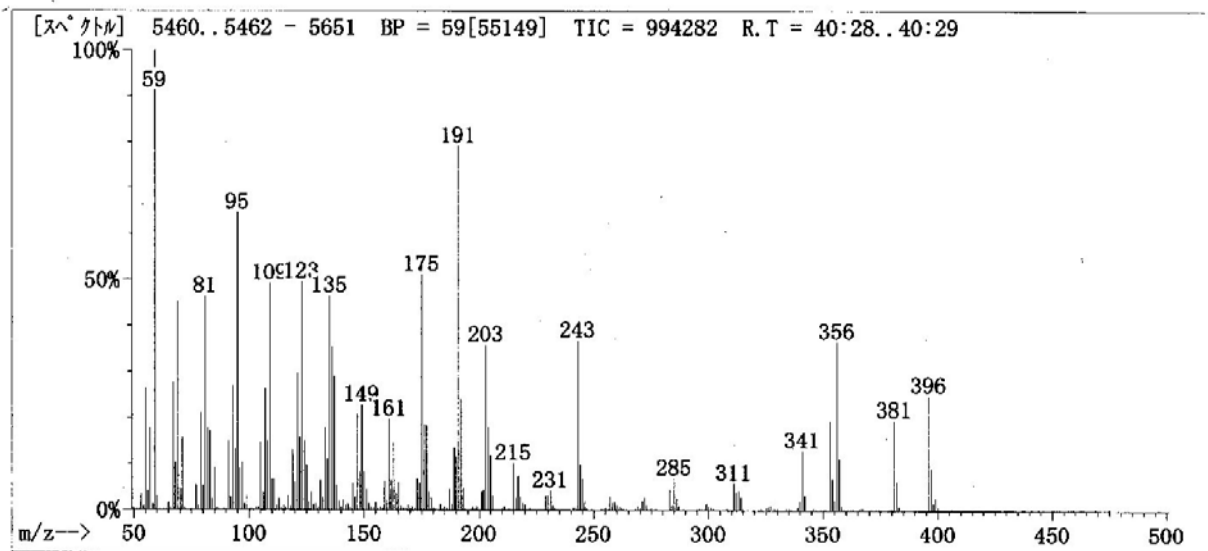


Fig. S12.1. EIMS of product 31.

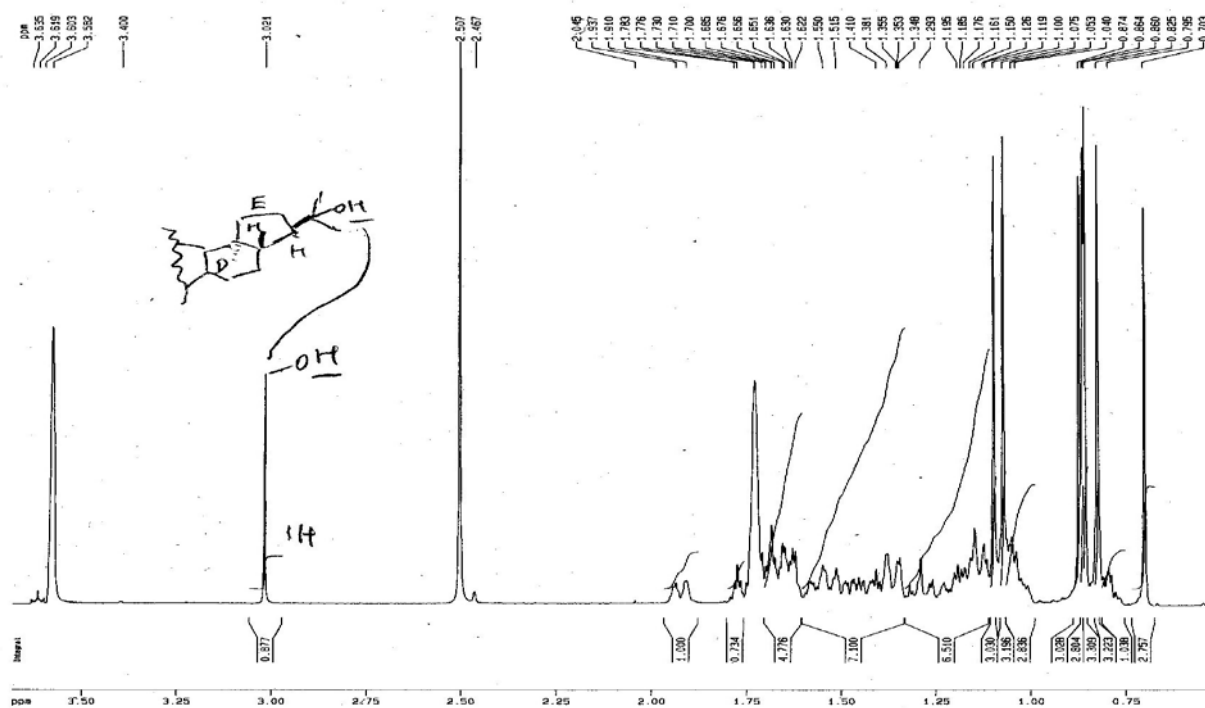


Fig. S12.2. ¹H NMR (whole spectrum) of Product 31 (400 MHz, THF d₈).

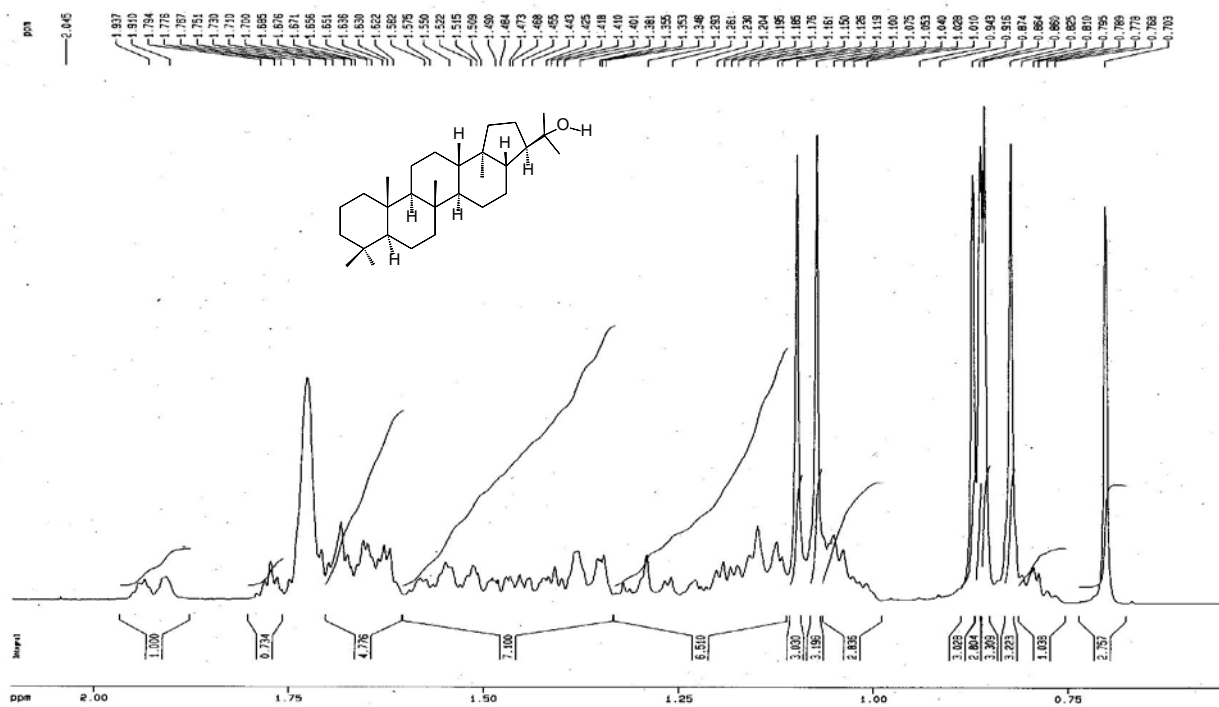


Fig. S12.3. ¹H NMR (expanded region) of Product **31** (400 MHz, THF d₈).

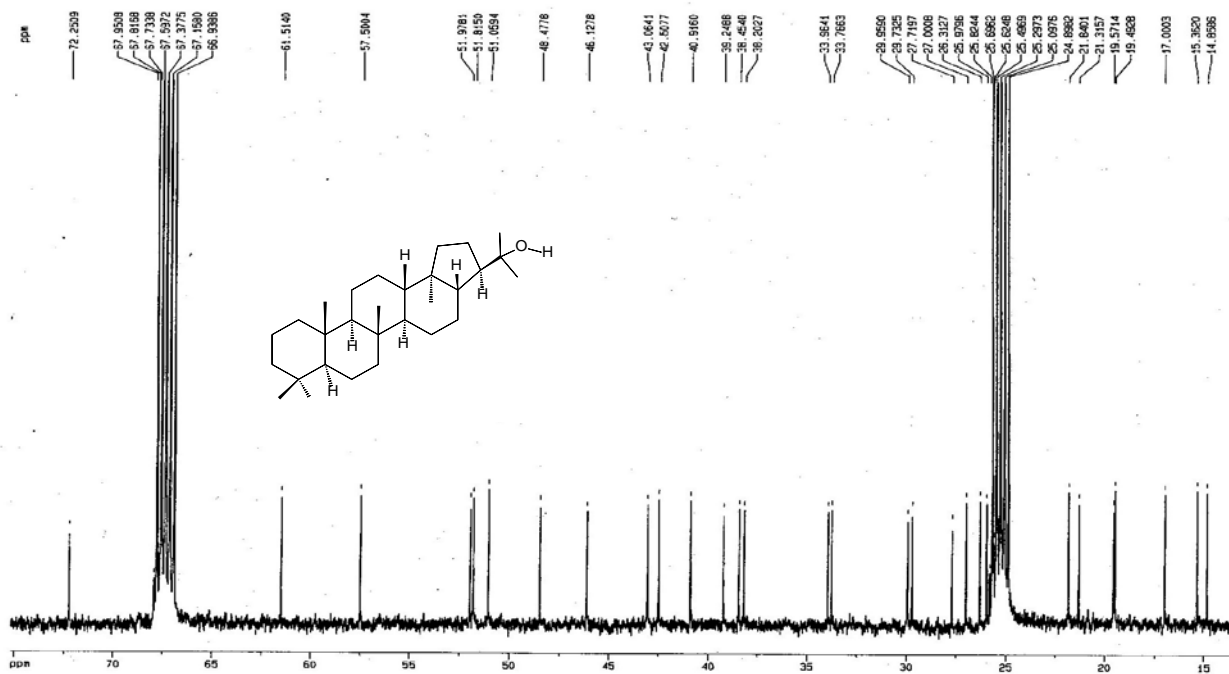


Fig. S12.4. ¹³C NMR of Product **31** (400 MHz, THF d₈).

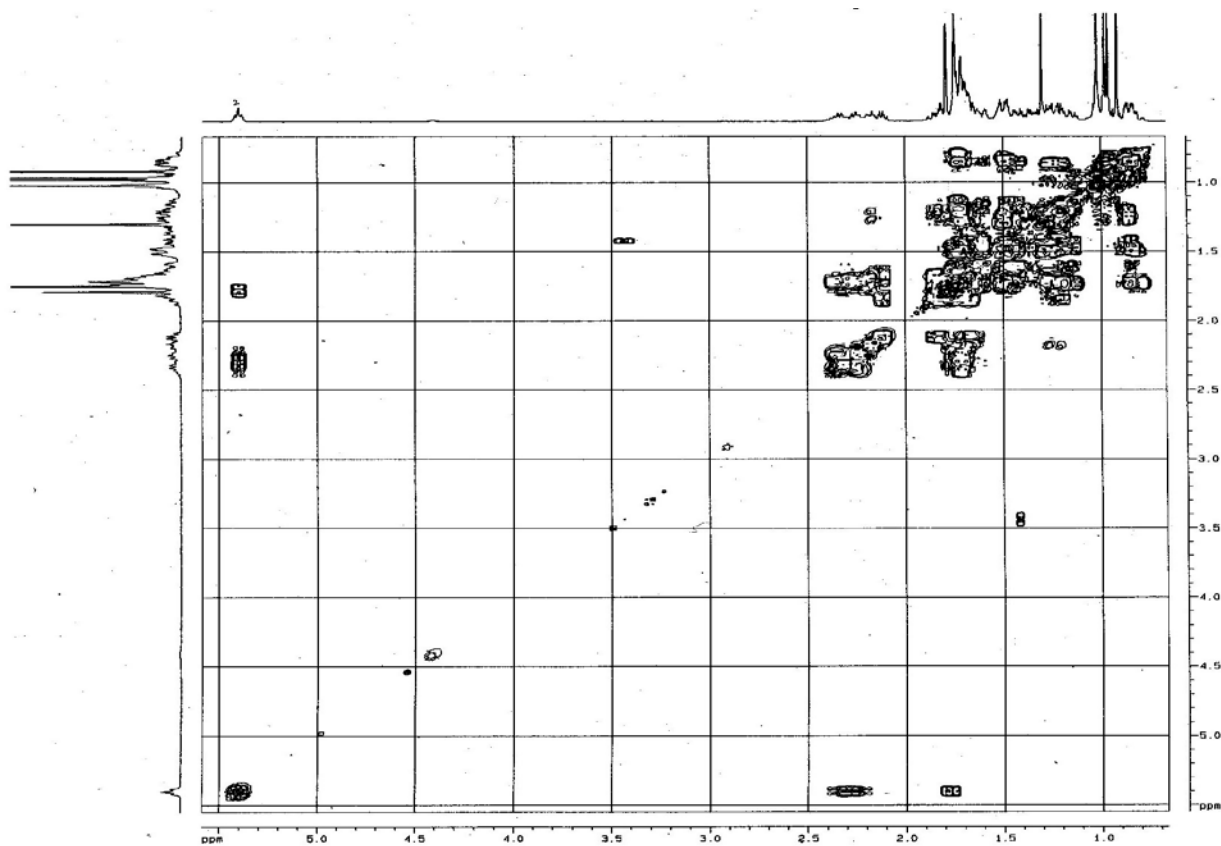


Fig. S12.5. ^1H - ^1H COSY of product **31** (400 MHz, THF d_8).

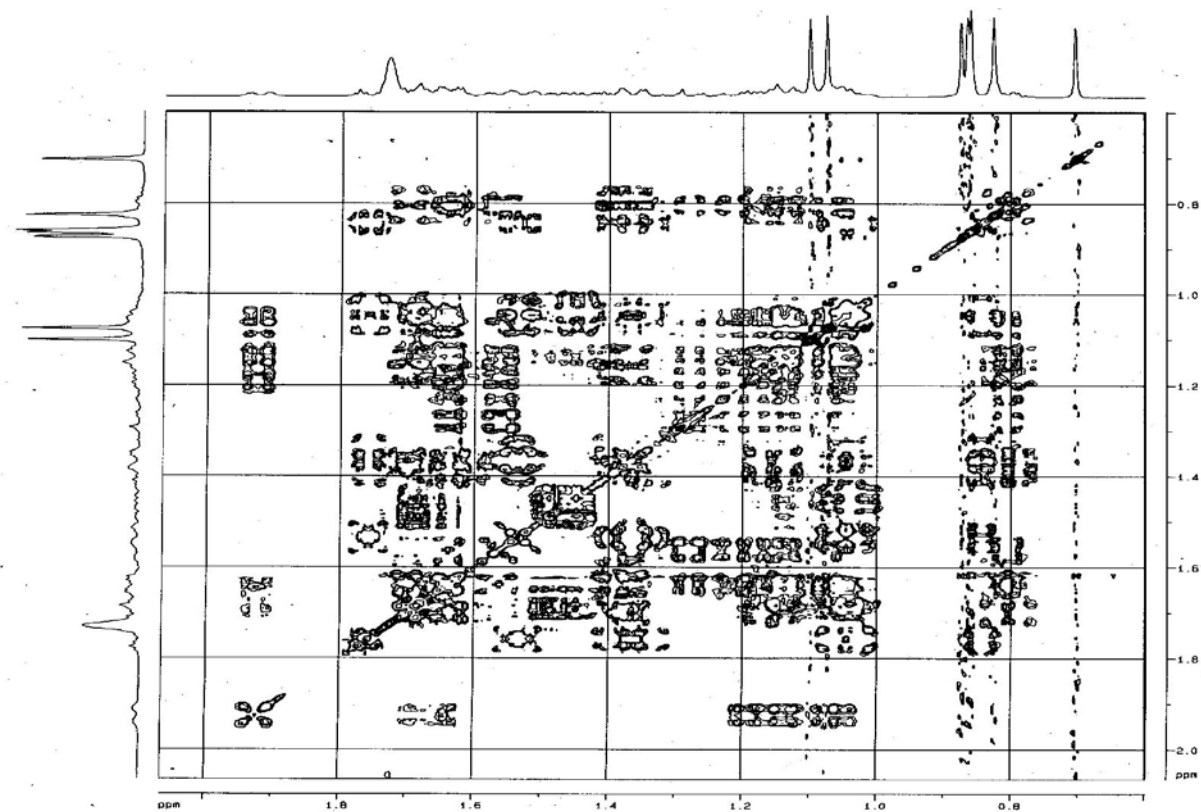


Fig. S12.6. TOCSY of **31** (400 MHz, THF d_8).

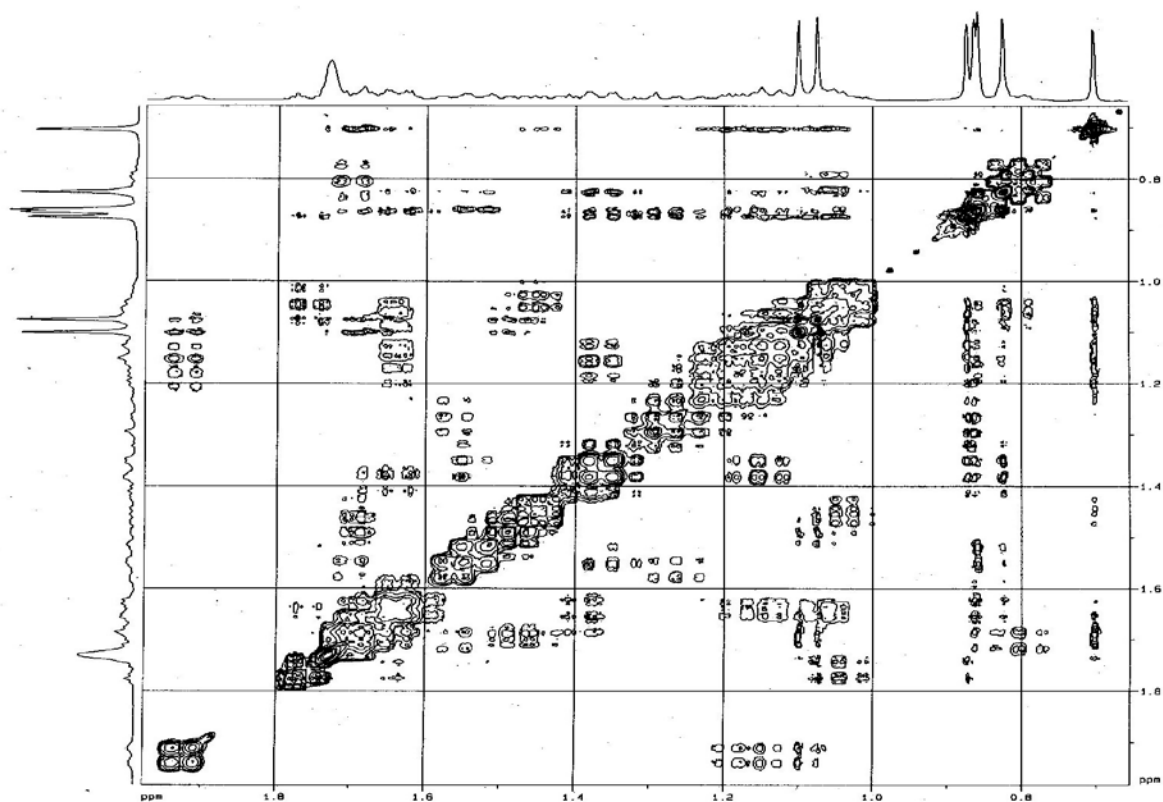


Fig. S12.7. NOESY of **31** (400 MHz, THF *d*₈).

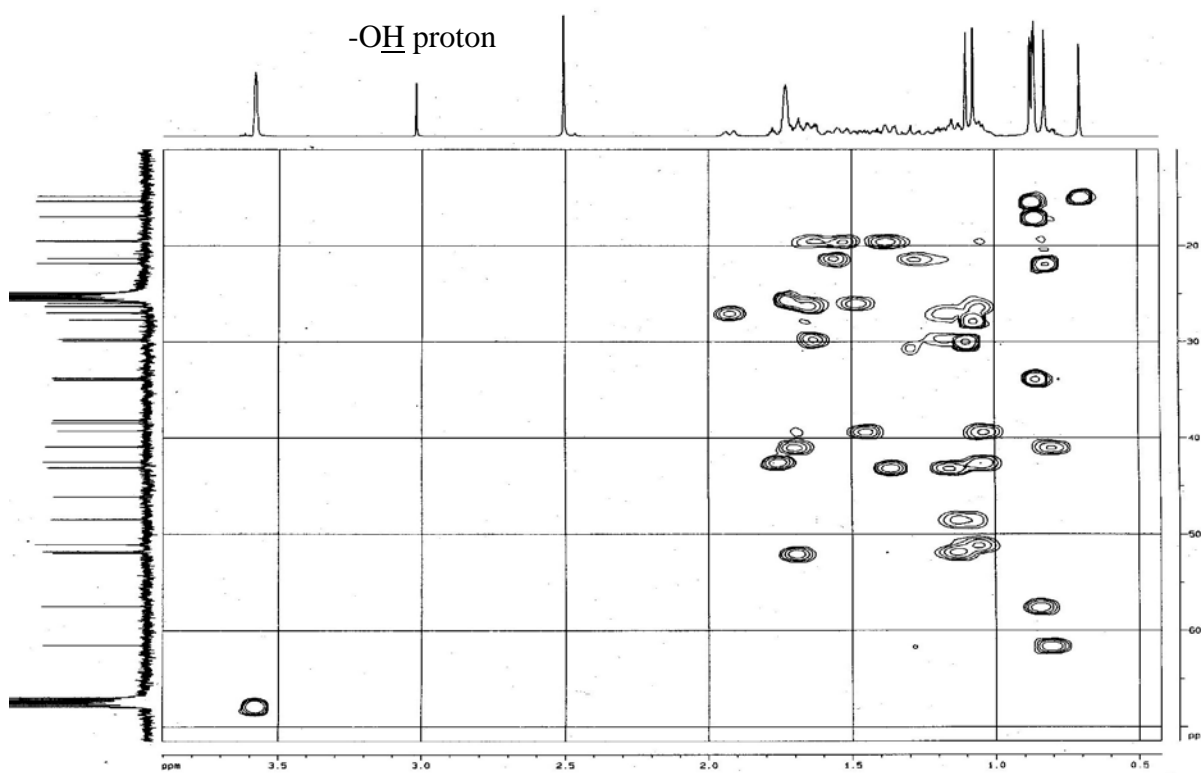


Fig. S13.8. HSQC (whole spectrum) of **31** (400 MHz, THF *d*₈).

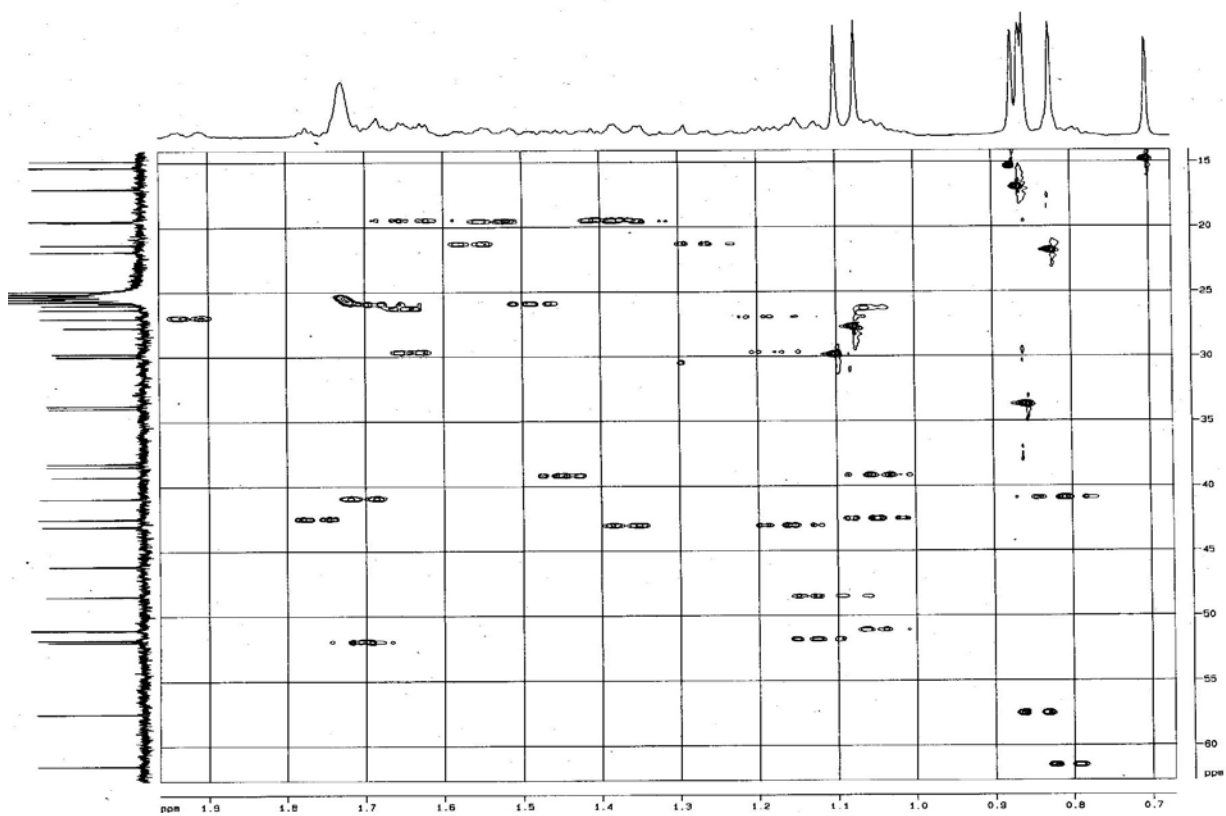


Fig. S12.9. HSQC (Expanded region) of **31** (400 MHz, THF d_8).

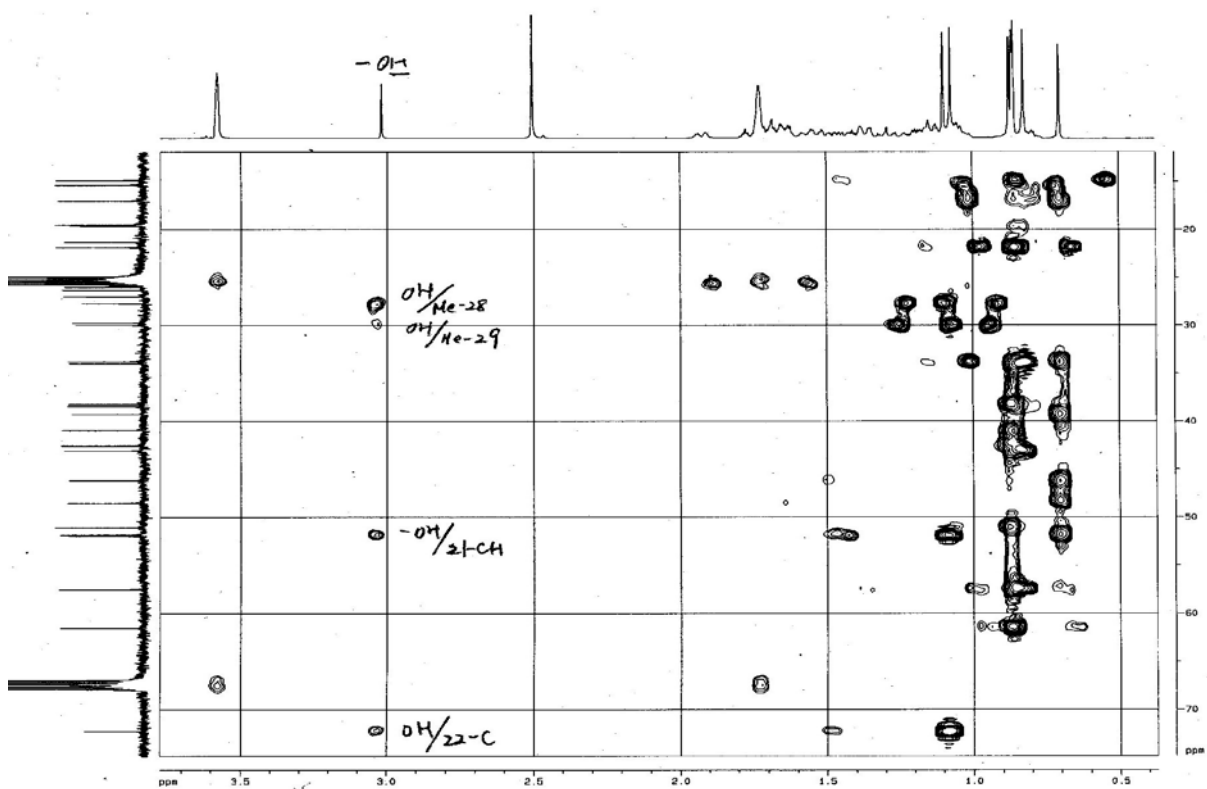


Fig. S12.10. HMBC (whole spectrum) of **31** (400 MHz, THF d_8).

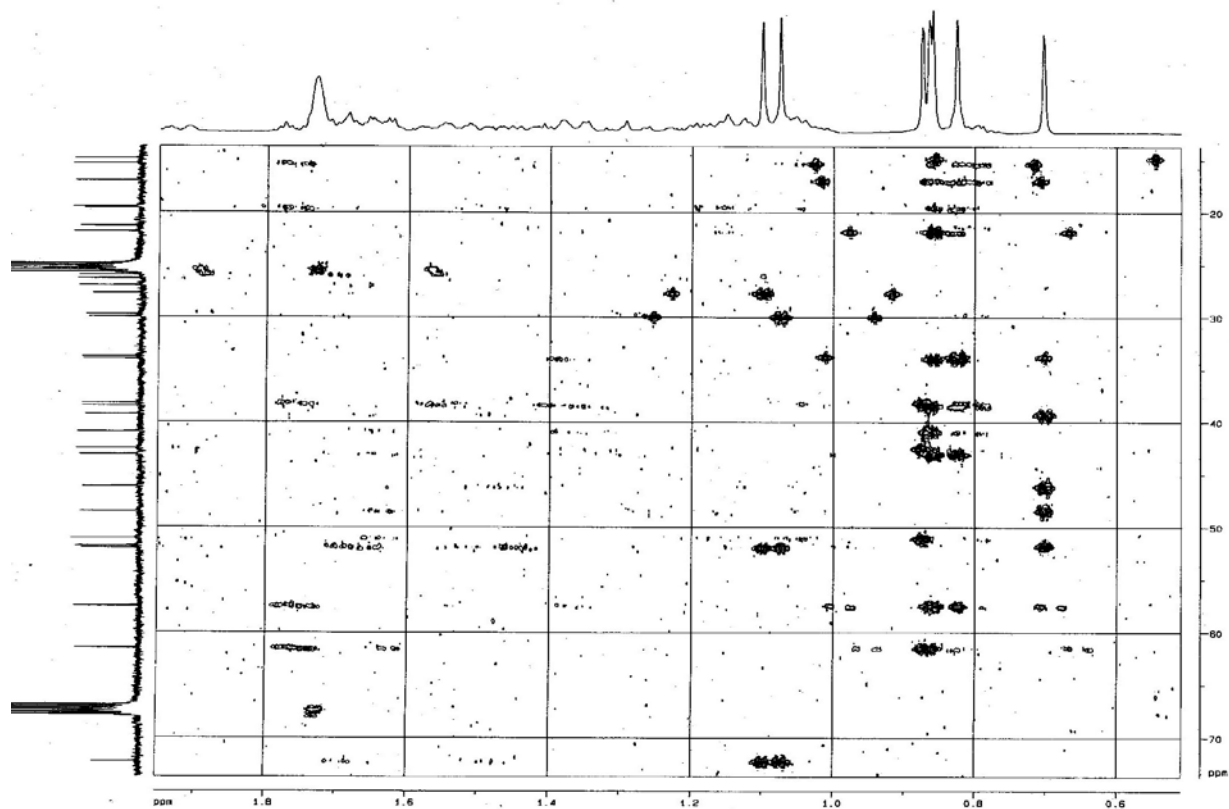
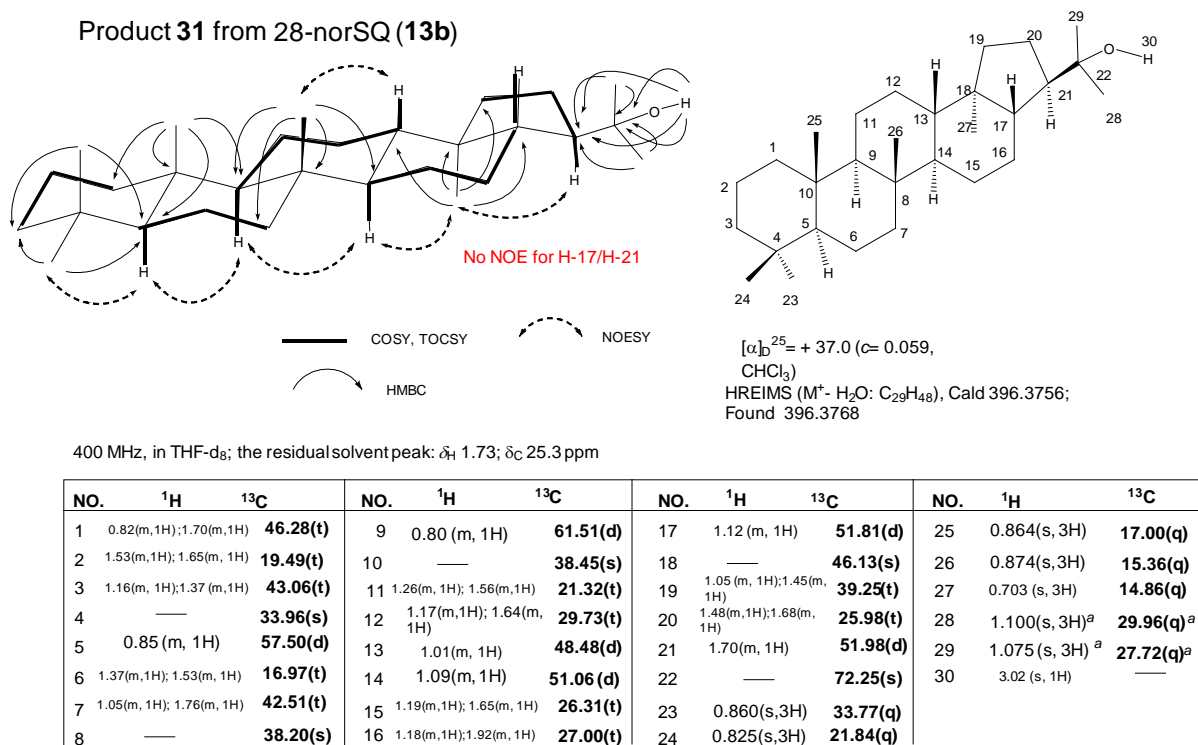


Fig. S12.11. HMBC (expanded region) of **31** (400 MHz, THF d_8).



a: exchangeable

Fig. S12.12. Assignments of the NMR data of product **41** in THF d_8 , HR-EIMS and optical rotation.