

A Synthetic Route to 1,4-Disubstituted Tetrahydro- β -carbolines and Tetrahydropyranoindoles via Ring-Opening/Pictet-Spengler Reaction of Aziridines and Epoxides with Indoles/Aldehydes

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Supporting Information

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1. X-ray crystal structures:

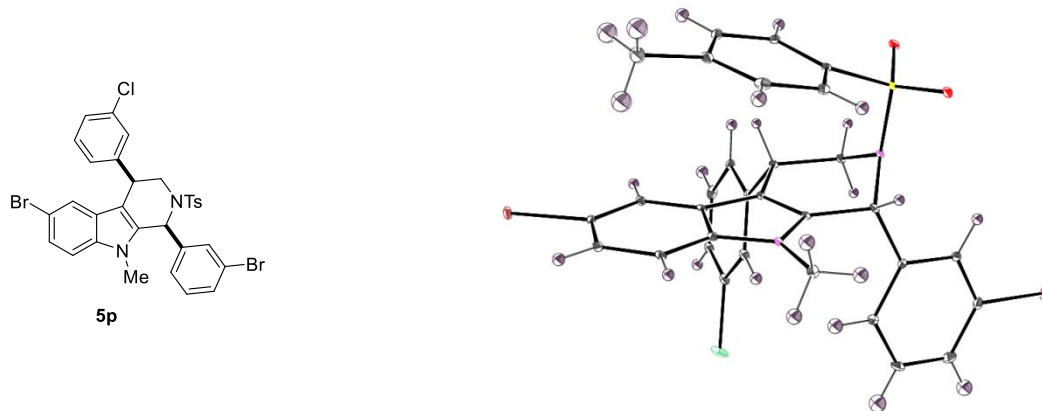


Figure:S1 ORTEP diagram of compound **5p** (50% thermal ellipsoid)

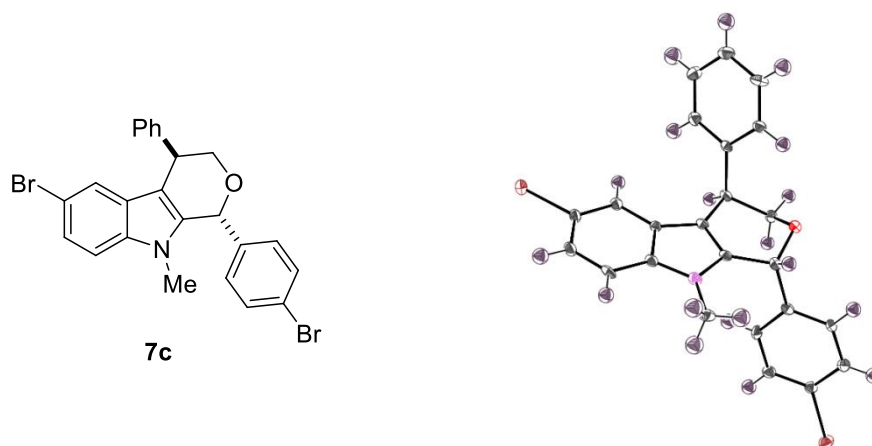


Figure:S2 ORTEP diagram of compound **7c** (50% thermal ellipsoid)

2. X-ray crystallographic analysis of 5p and 7c

The crystals used in the analyses were glued to a glass fiber and mounted on SMART APEX diffractometer. The instrument was equipped with CCD area detector and data were collected using graphite-monochromated Mo K α radiation ($\lambda = 0.71069 \text{ \AA}$) at low temperature (100K). Cell constants were obtained from the least-squares refinement of three-dimensional centroids through the use of CCD recording of narrow ω rotation frames, completing almost all-reciprocal space in the stated θ range. All data were collected with SMART 5.628 and were integrated with the SAINT¹ program. An empirical absorption correction was applied to collect reflections with SADABS² using XPREP.³ The structure was solved using SIR-97⁴ and refined using SHELXL-97.⁵ The space group of the compounds was determined based on the lack of systematic absence and intensity statistics. Full matrix least squares/difference Fourier cycles were performed which located the remaining non-hydrogen atoms. All non-hydrogen atoms were refined with anisotropic displacement parameters. All the hydrogen atoms are fixed by using geometrical constraints using idealized geometries and have been defined isotropically.

Table S1. X-ray crystallographic data and structure refinement

Compound	5p	7c
Formula	C ₃₁ H ₂₅ Br ₂ Cl N ₂ O ₂ S	C ₂₄ H ₁₉ Br ₂ N O
Formula weight	684.86	497.22
CCDC No.	1854875	1854884
Crystal colour, habit	White, Prism	White, Prism
<i>T</i> / K	100(2)	100(2)
Crystal system	Monoclinic	Monoclinic
Space group	<i>P</i> -2 ₁ (no. 4)	<i>P</i> -2 ₁ (no. 4)

$a/\text{\AA}$	10.2533(9)	7.571(3)
$b/\text{\AA}$	29.303(2)	13.464(4)
$c/\text{\AA}$	10.6278(10)	20.373(7)
$\alpha/^\circ$	90	90.00
$\beta/^\circ$	116.562(2)	99.227(10)
$\gamma/^\circ$	90	90.00
$V/\text{\AA}^3$	2856.1(4)	2050.0(12)
Z	4	4
$D_c/\text{g cm}^{-3}$	1.593	1.611
μ/mm^{-1}	3.037	3.969
Reflections measured	28773	16217
Unique reflections	5063	2175
Reflections used	7094	3680
$I > 2\sigma(I)$		
$R_1^a, wR_2^b [I > 2\sigma(I)]$	$R_1 = 0.0503^a$ $wR_2 = 0.0925^b$	$R_1 = 0.0805^a$ $wR_2 = 0.1836^b$
R_1^a, wR_2^b (all data)	$R_1 = 0.0864^a$ $wR_2 = 0.1032^b$	$R_1 = 0.1459^a$ $wR_2 = 0.2111^b$
GOF on F^2	1.013	1.052

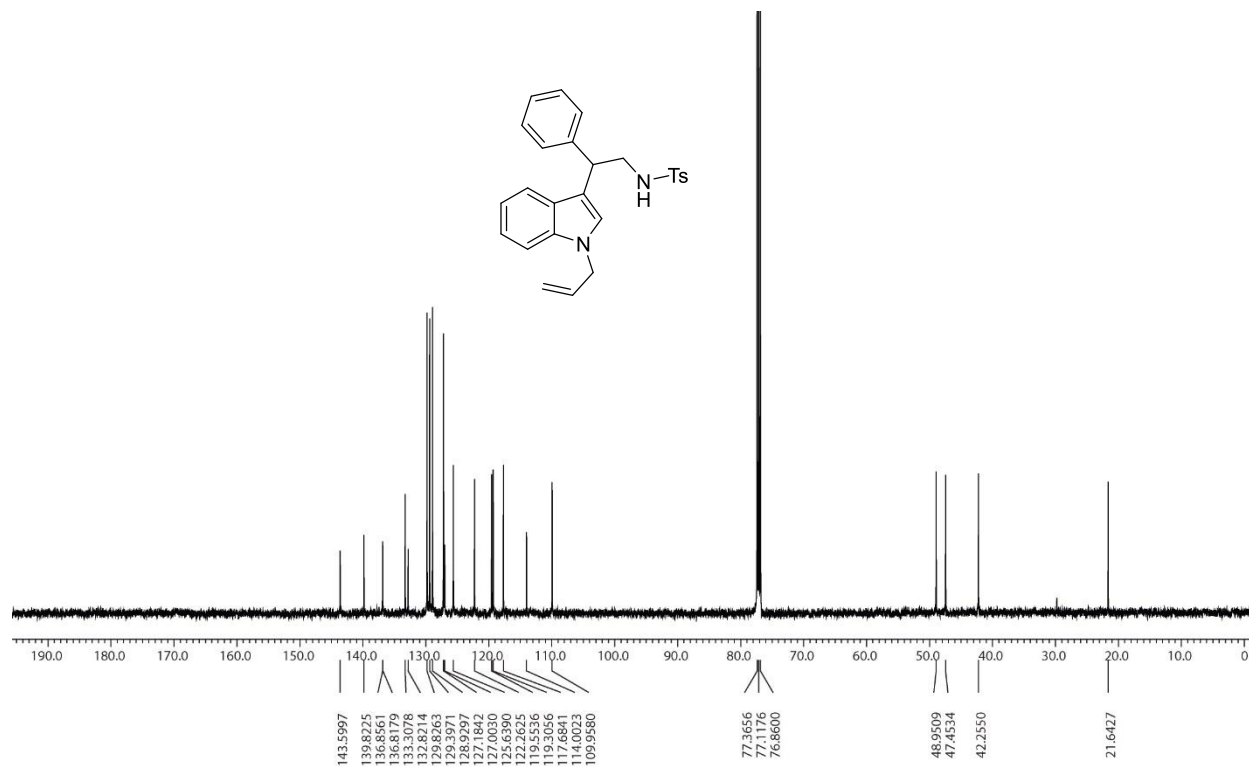
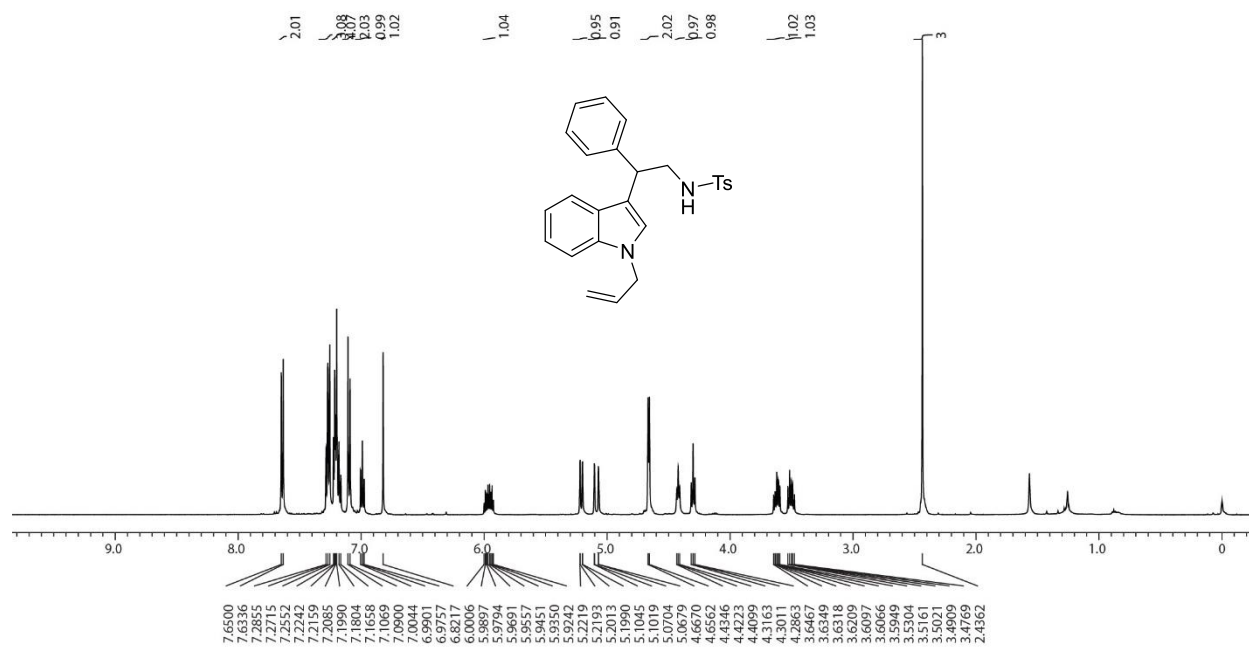
$${}^a R_1 = \Sigma || F_o | - |F_c || / \Sigma |F_o|. \quad {}^b wR_2 = \{ \Sigma [w(|F_o|^2 - |F_c|^2)^2] / \Sigma [w(|F_o|^2)^2] \}^{1/2}$$

3. References

1. SAINT+ 6.02ed.; Bruker AXS, Madison, WI, **1999**.
2. Sheldrick, G. M. SADABS, Empirical Absorption Correction Program, University of Göttingen, Göttingen, Germany, 1997.
3. XPREP, 5.1ed. Siemens Industrial Automation Inc., Madison, WI, **1995**.

4. A. Altomare, M. C. Burla, M. Camalli, G. L. Cascarano, C. Giacovazzo, A. Guagliardi, A. G. Moliterni, G. Polidori, R. Spagna, *J. Appl. Cryst.* **1999**, 32, 115.
5. Sheldrick, G. M. SHELXL-97: Program for Crystal Structure Refinement (University of Göttingen, Göttingen, Germany, 1997).

4. ^1H and $^{13}\text{C}\{^1\text{H}\}$ NMR spectra



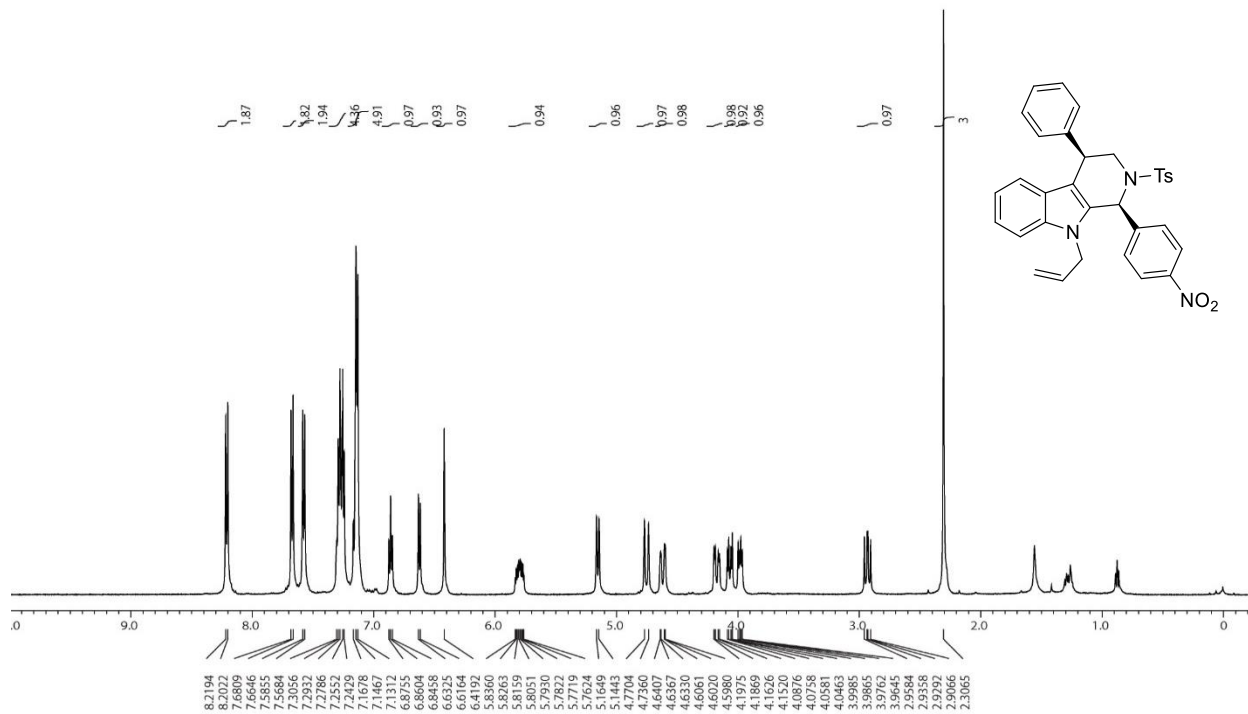


Figure:S5 ^1H NMR spectrum of **5a** (CDCl_3 , 500 MHz)

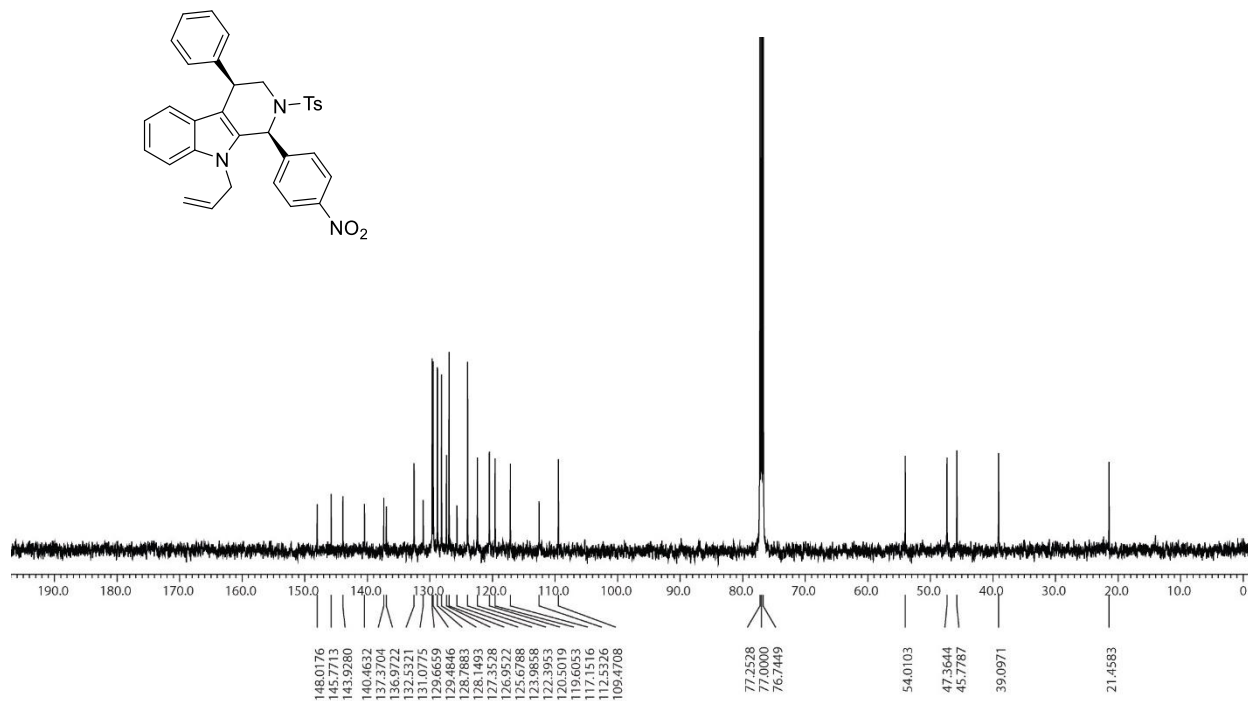


Figure:S6 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **5a** (CDCl_3 , 125 MHz)

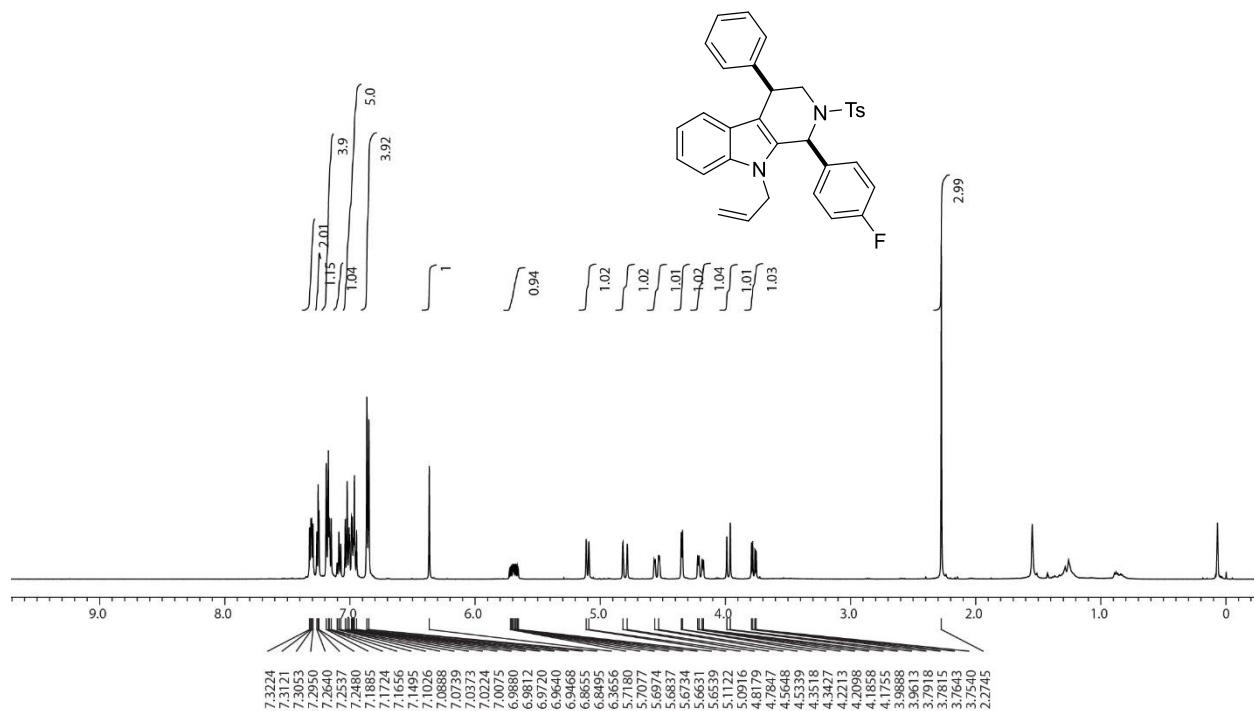


Figure:S7 ^1H NMR spectrum of **5b** (CDCl_3 , 500 MHz)

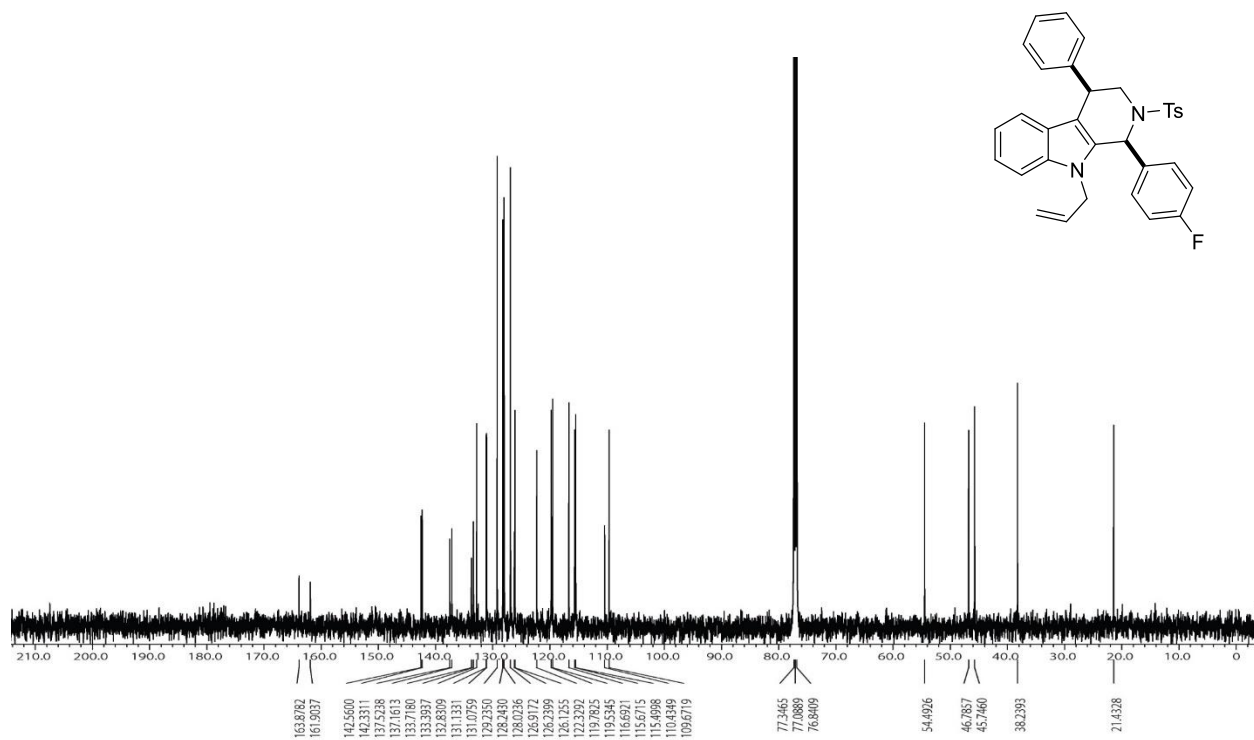


Figure:S8 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **5b** (CDCl_3 , 125 MHz)

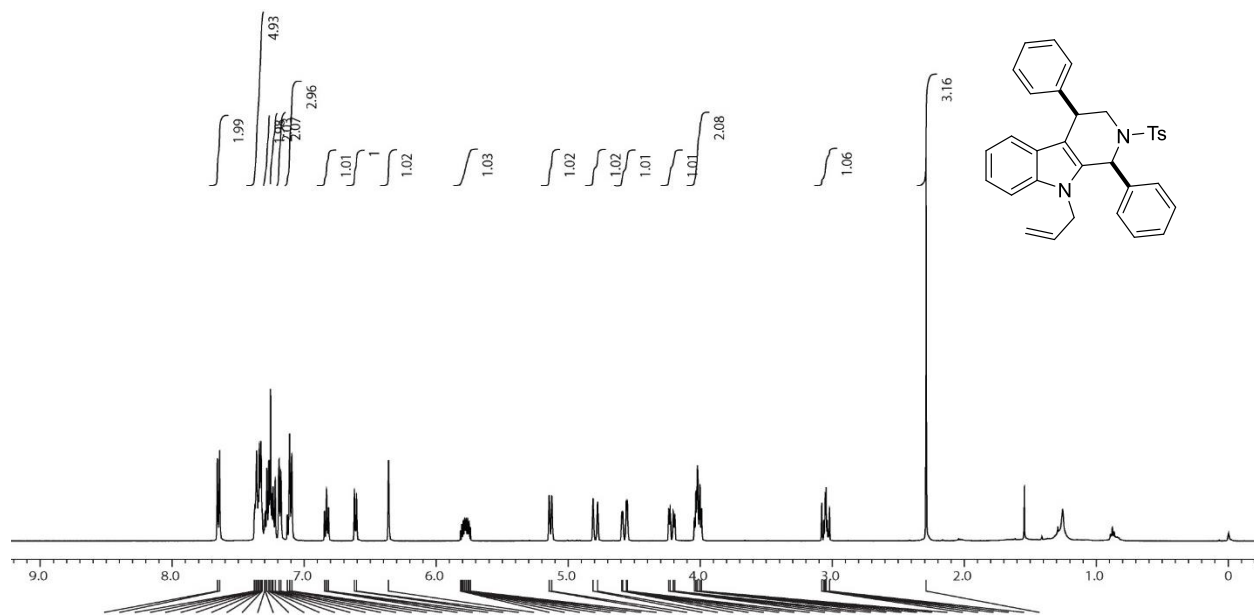


Figure:S9 ^1H NMR spectrum of **5c** (CDCl_3 , 500 MHz)

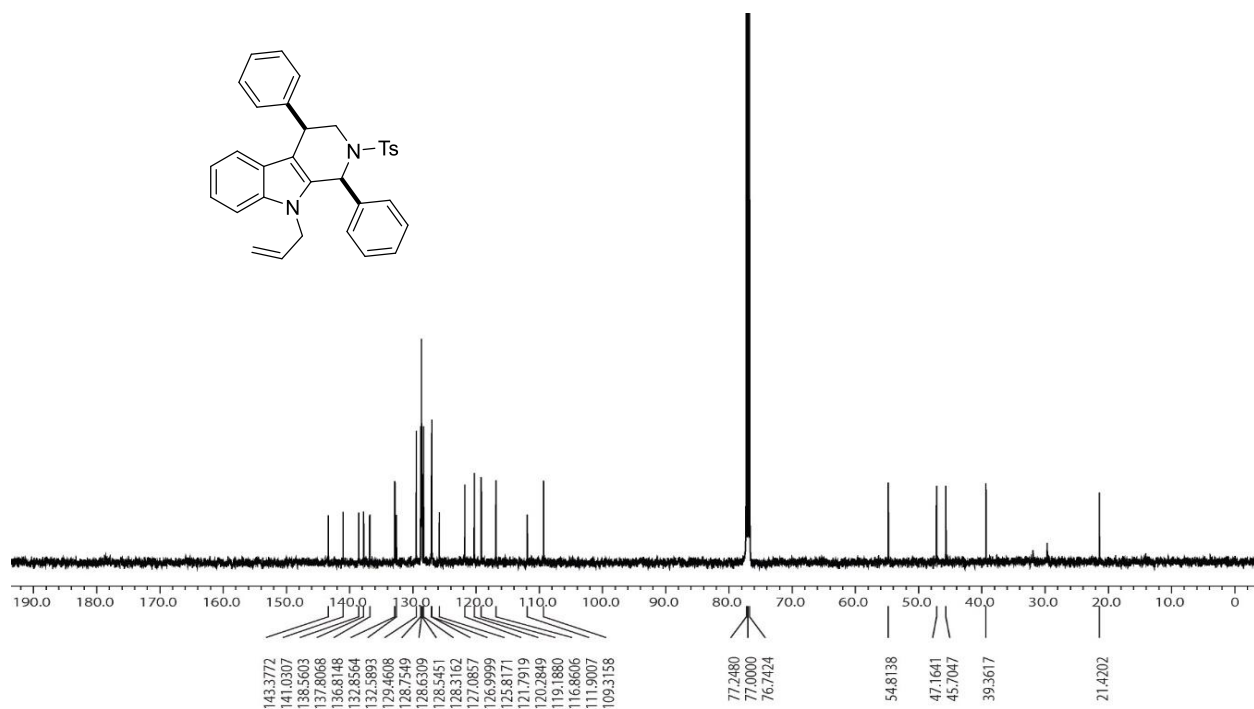


Figure:S10 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **5c** (CDCl_3 , 125 MHz)

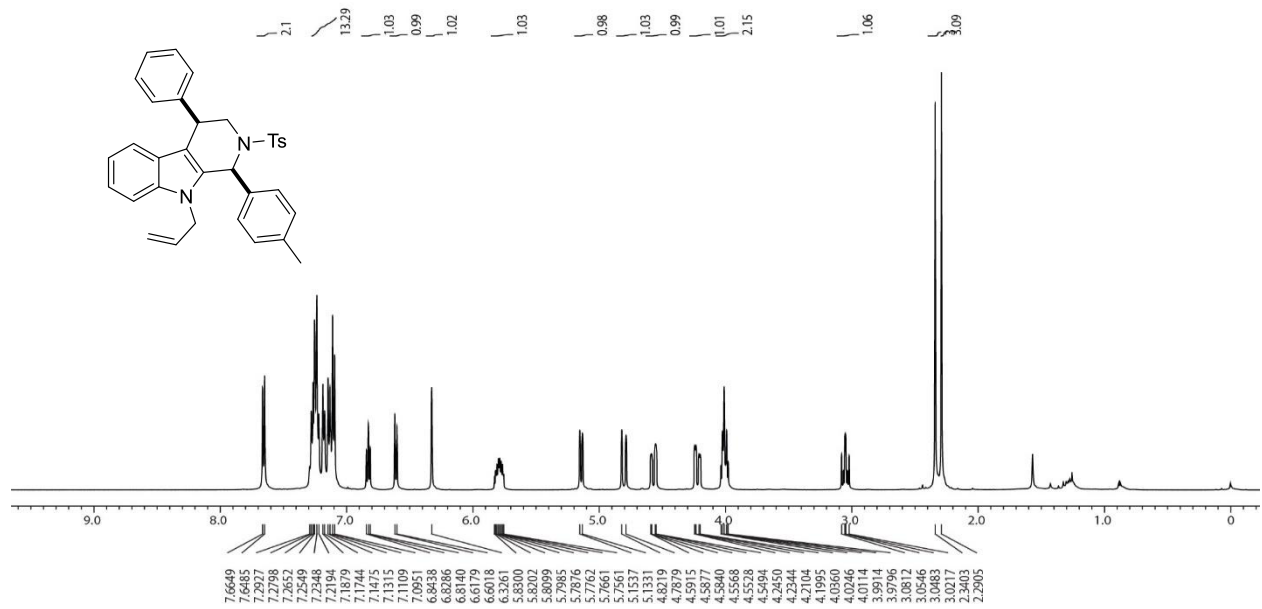


Figure:S11 ¹H NMR spectrum of **5d** (CDCl₃, 500 MHz)

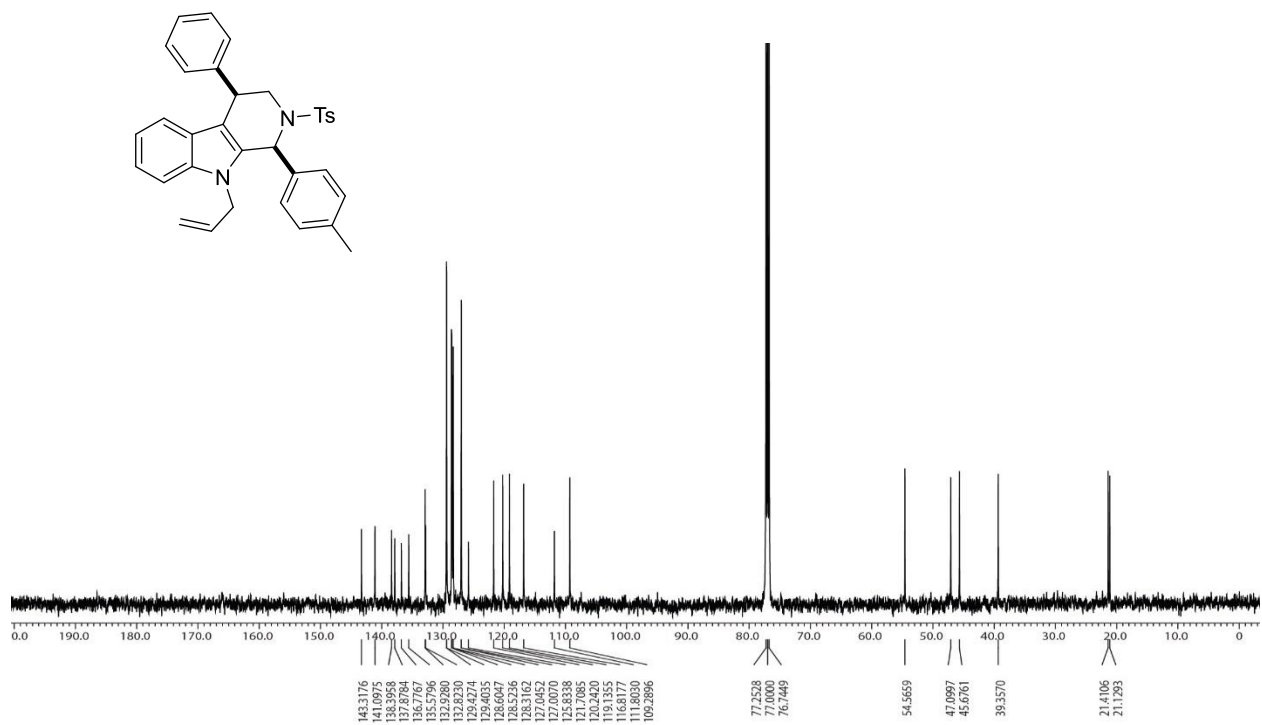
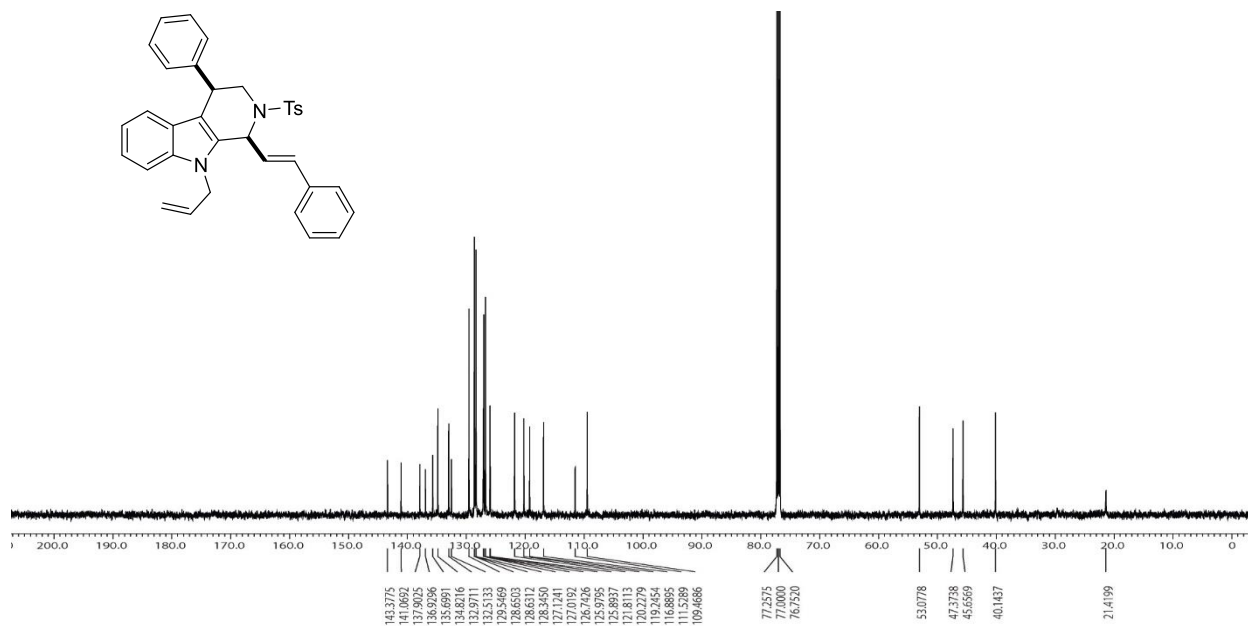
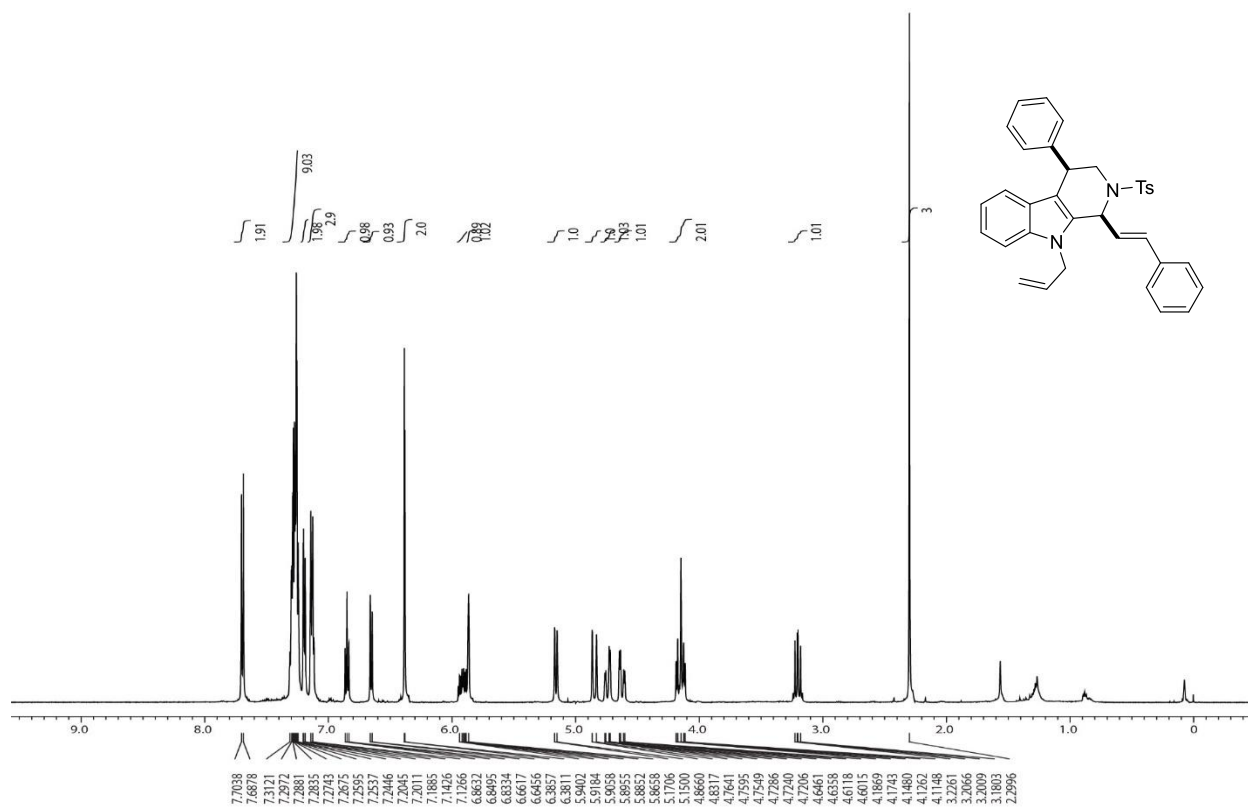


Figure:S12 ¹³C{¹H} NMR spectrum of **5d** (CDCl₃, 125 MHz)



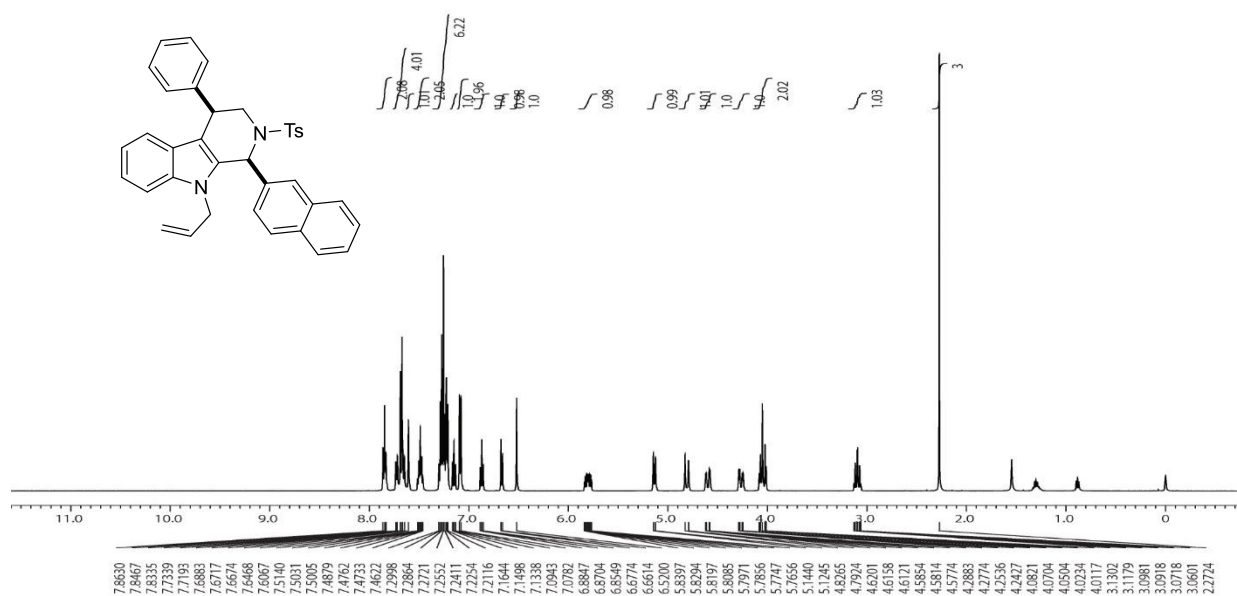


Figure:S15 ¹H NMR spectrum of **5f** (CDCl₃, 500 MHz)

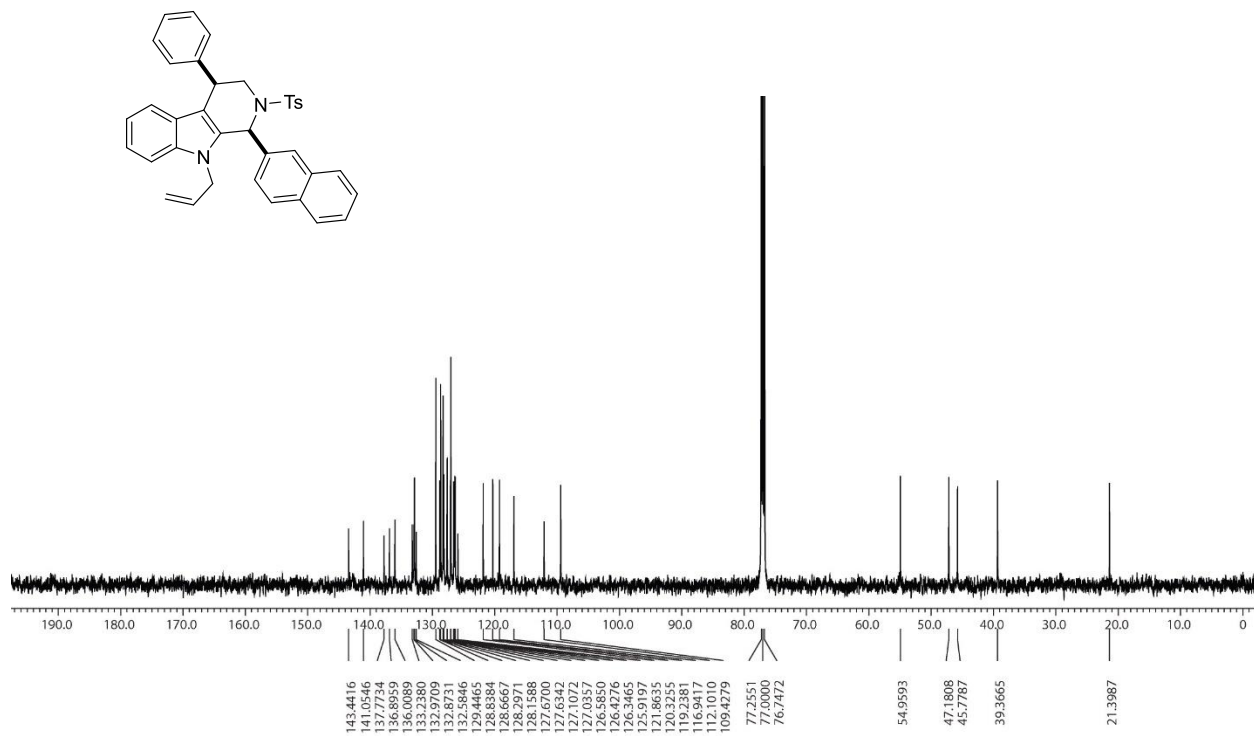


Figure:S16 ¹³C{¹H} NMR spectrum of **5f** (CDCl₃, 125 MHz)

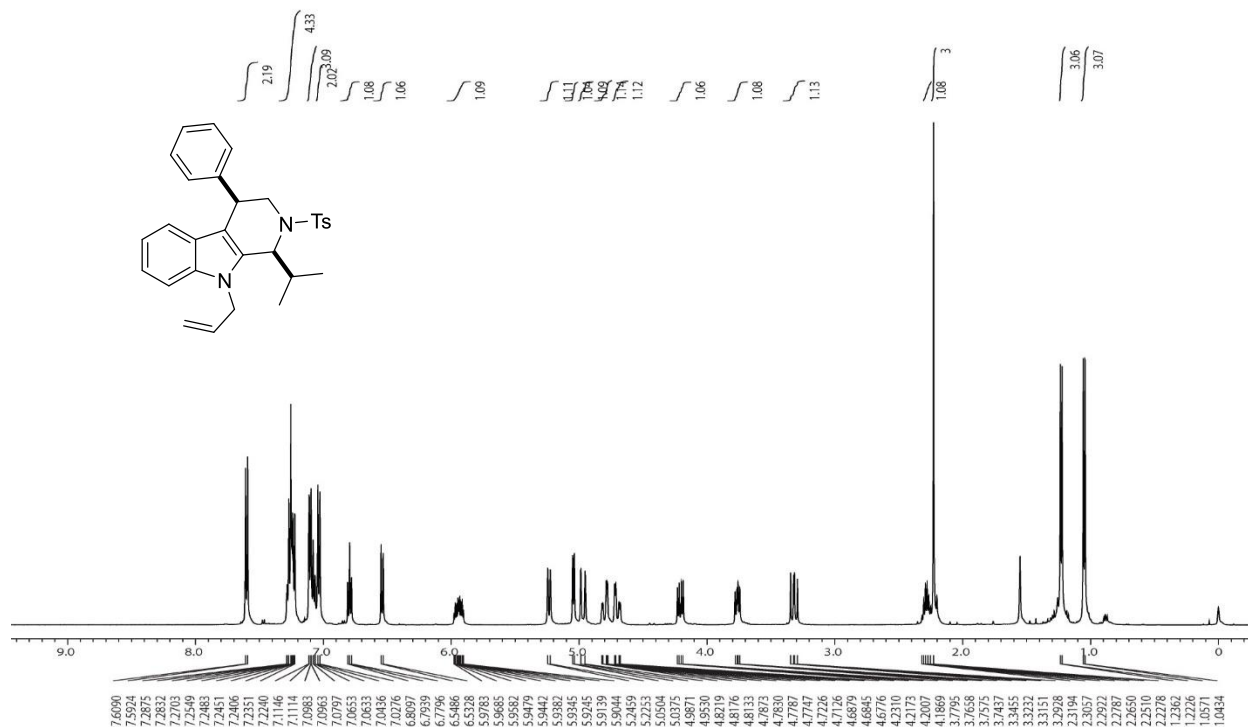


Figure:S17 ¹H NMR spectrum of **5g** (CDCl₃, 500 MHz)

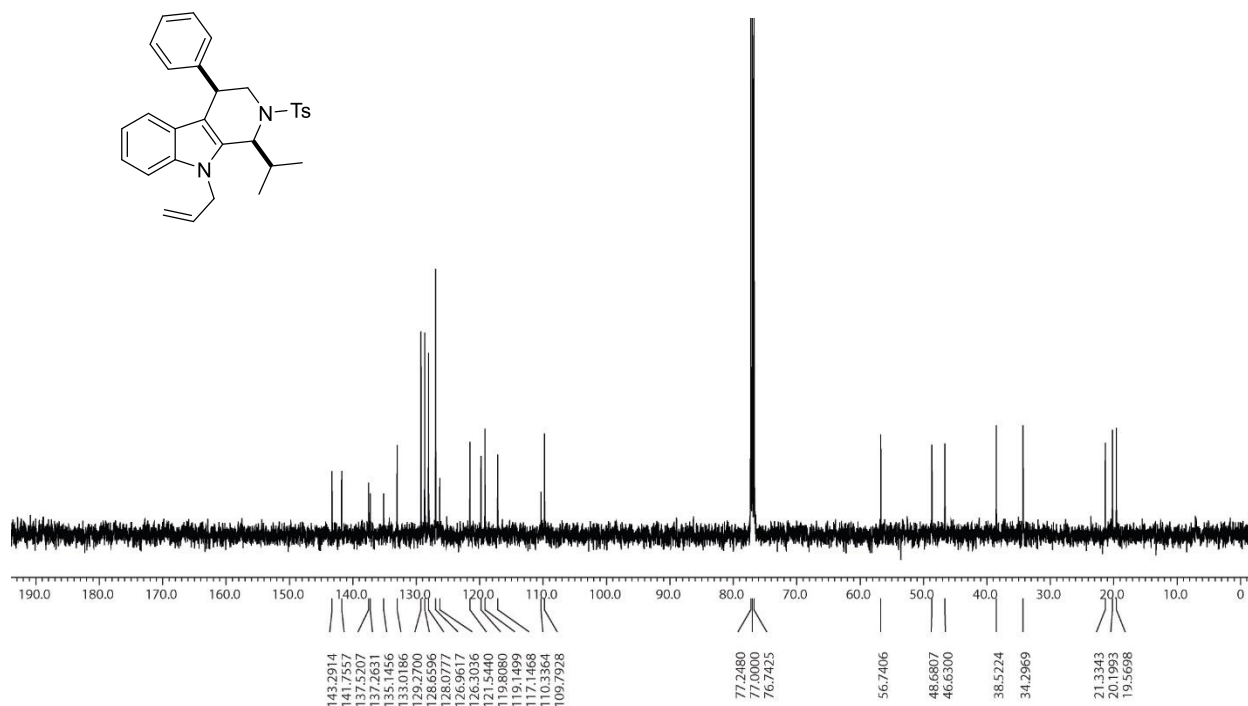


Figure:S18 ¹³C {¹H} NMR spectrum of **5g** (CDCl₃, 125 MHz)

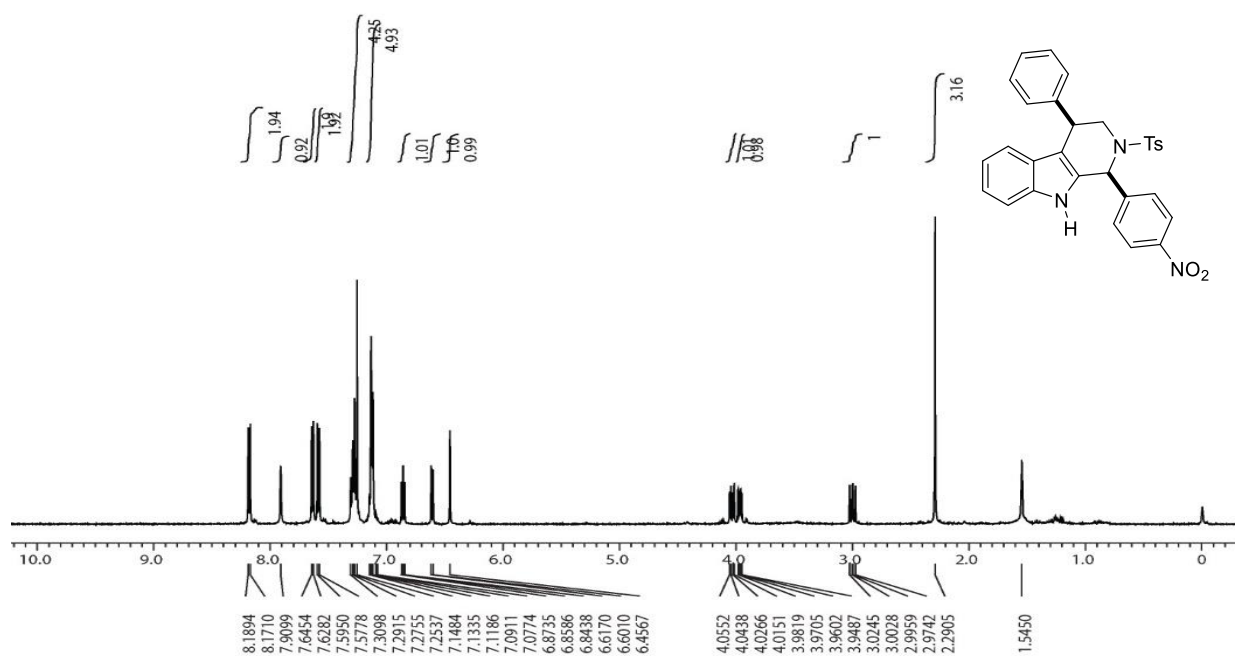


Figure:S19 ¹H NMR spectrum of 5h (CDCl₃, 500 MHz)

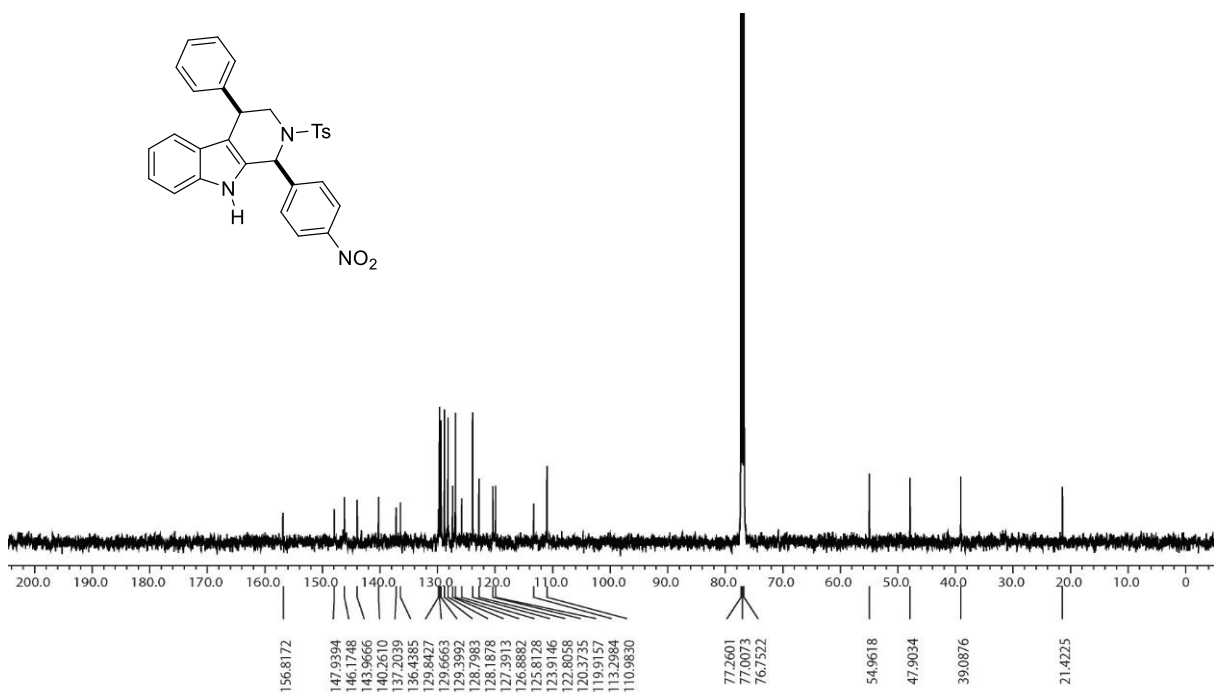


Figure:S20 ¹³C{¹H} NMR spectrum of 5h (CDCl₃, 125 MHz)

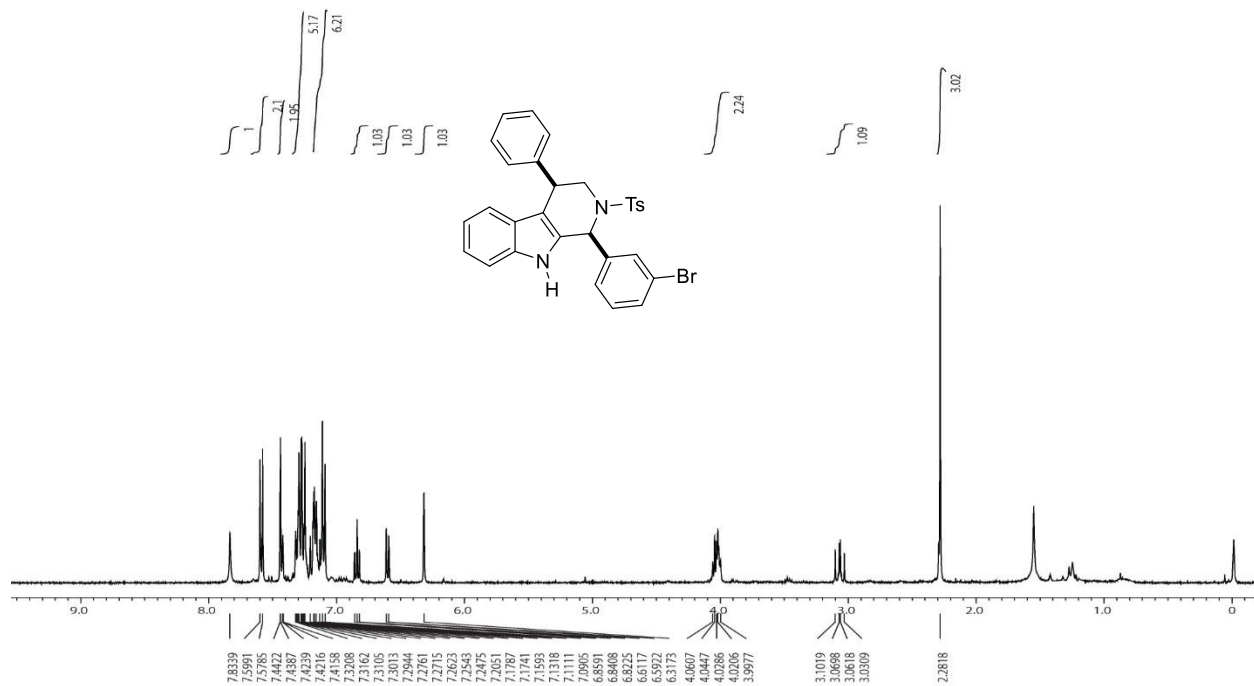


Figure:S21 ¹H NMR spectrum of **5i** (CDCl₃, 500 MHz)

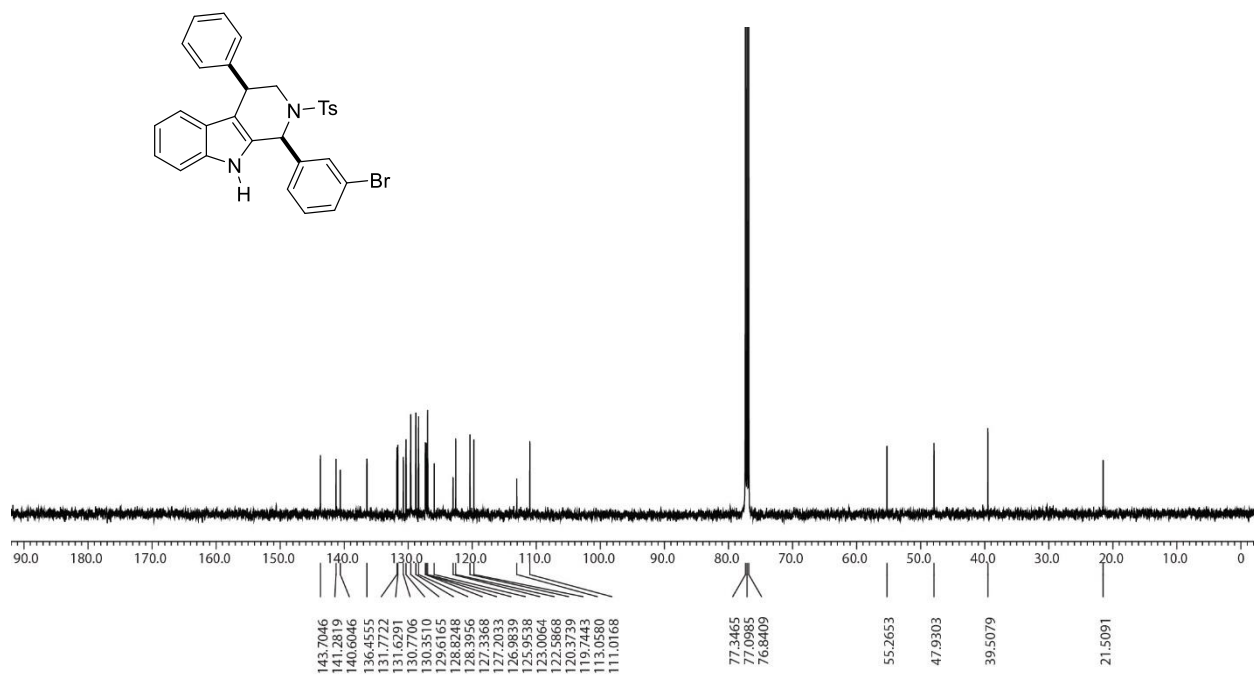


Figure:S22 ¹³C{¹H} NMR spectrum of **5i** (CDCl₃, 125 MHz)

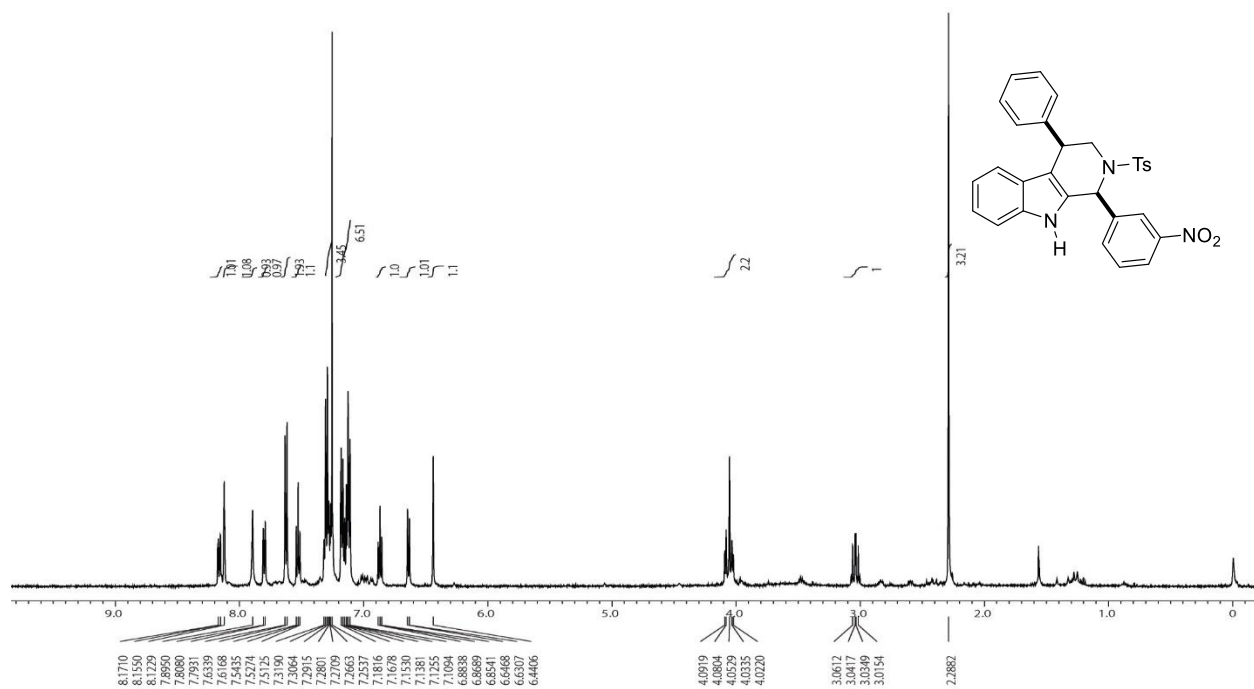


Figure:S23 ¹H NMR spectrum of **5j** (CDCl₃, 500 MHz)

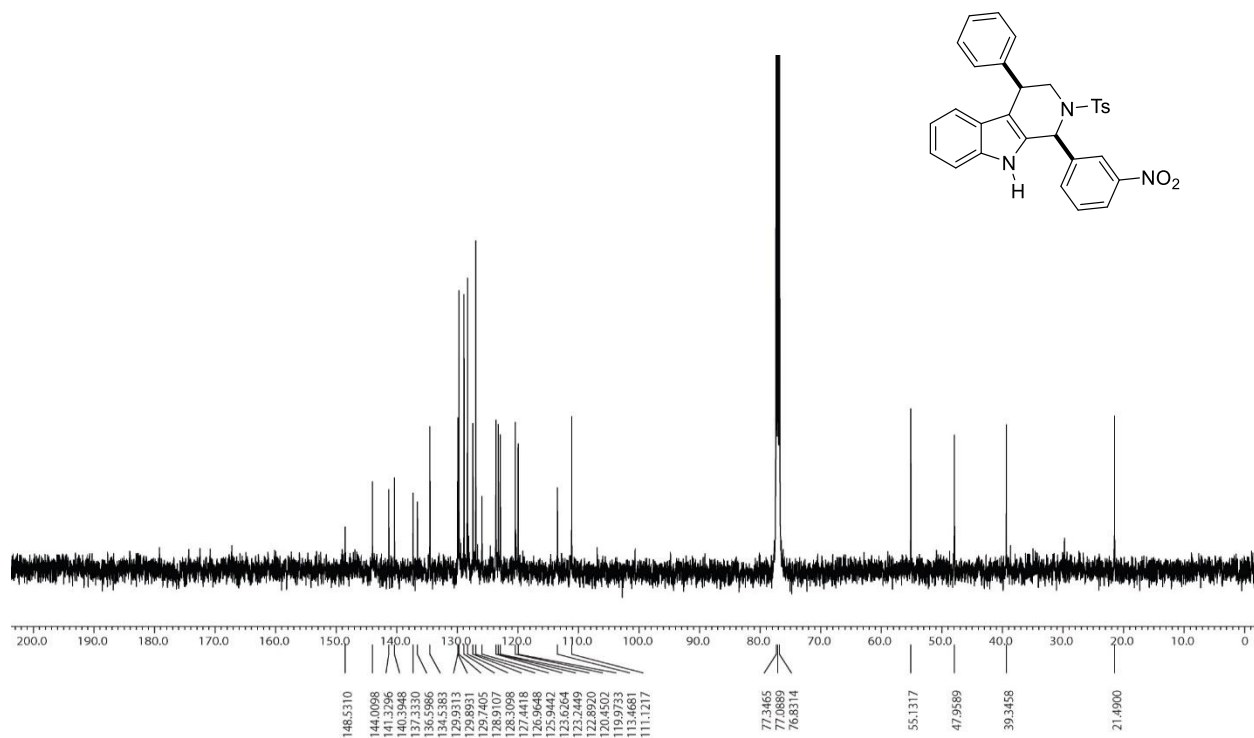


Figure:S24 ¹³C{¹H} NMR spectrum of **5j** (CDCl₃, 125 MHz)

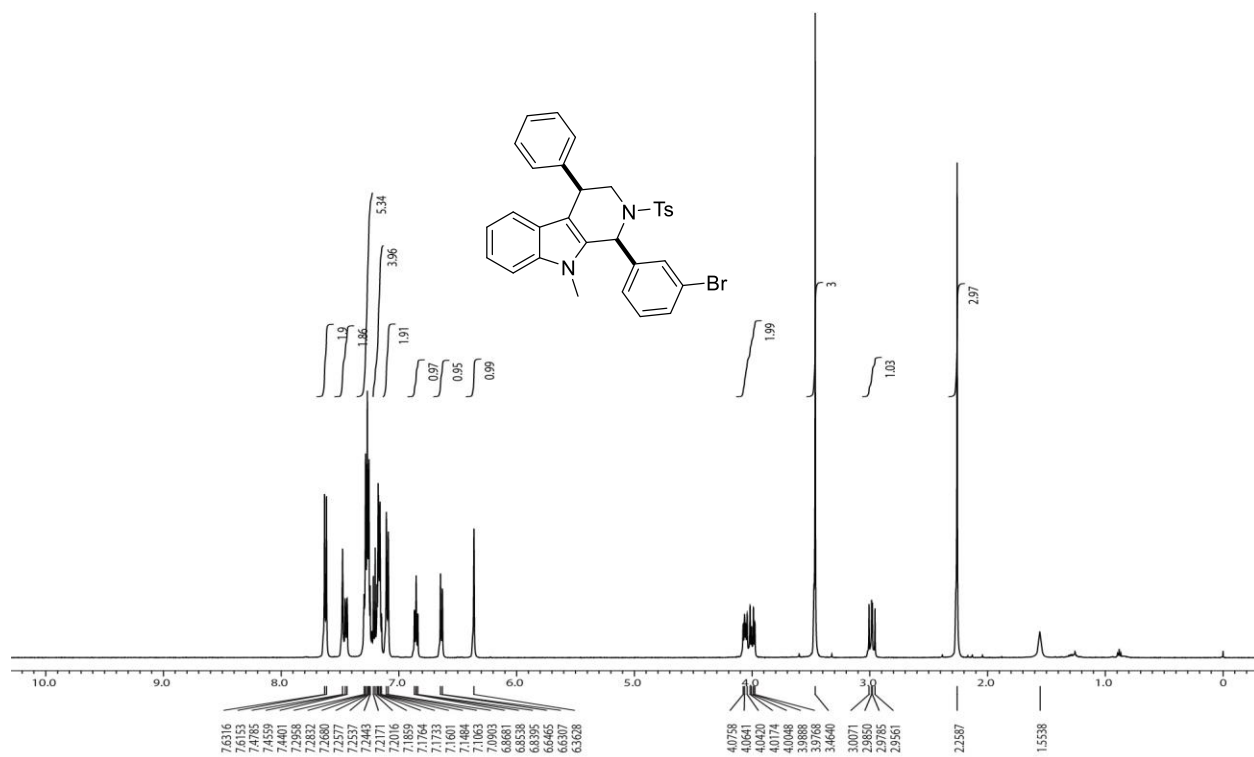


Figure:S25 ¹H NMR spectrum of **5k** (CDCl₃, 500 MHz)

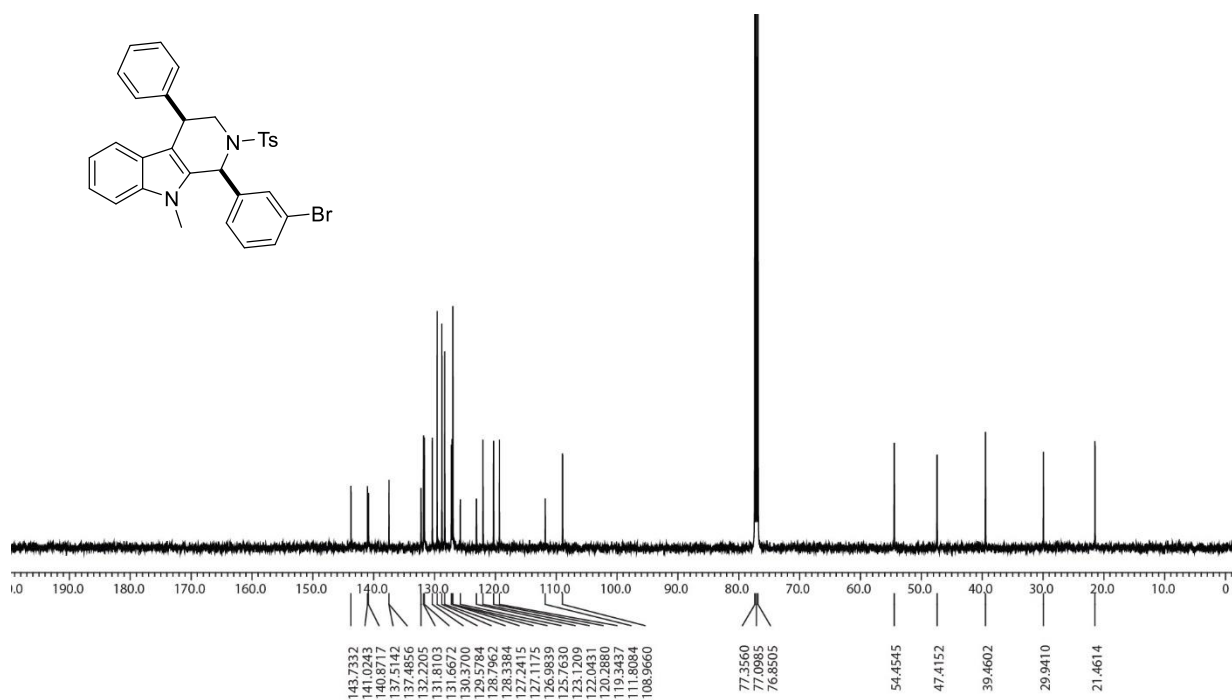


Figure:S26 ¹³C{¹H} NMR spectrum of **5k** (CDCl₃, 125 MHz)

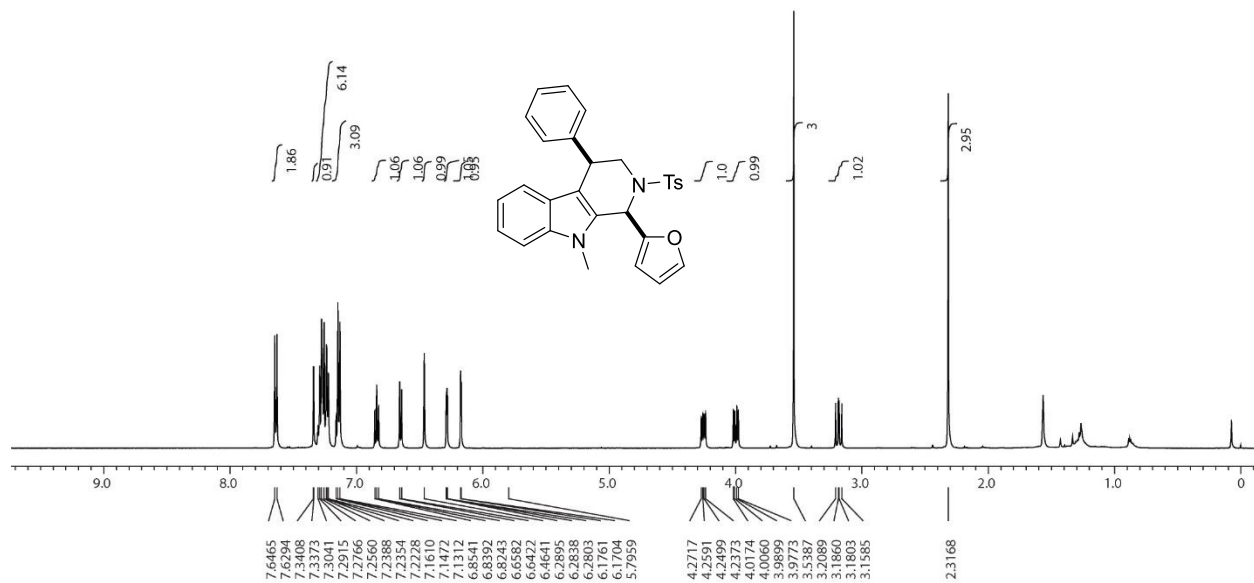


Figure:S29 ¹H NMR spectrum of **5m** (CDCl₃, 500 MHz)

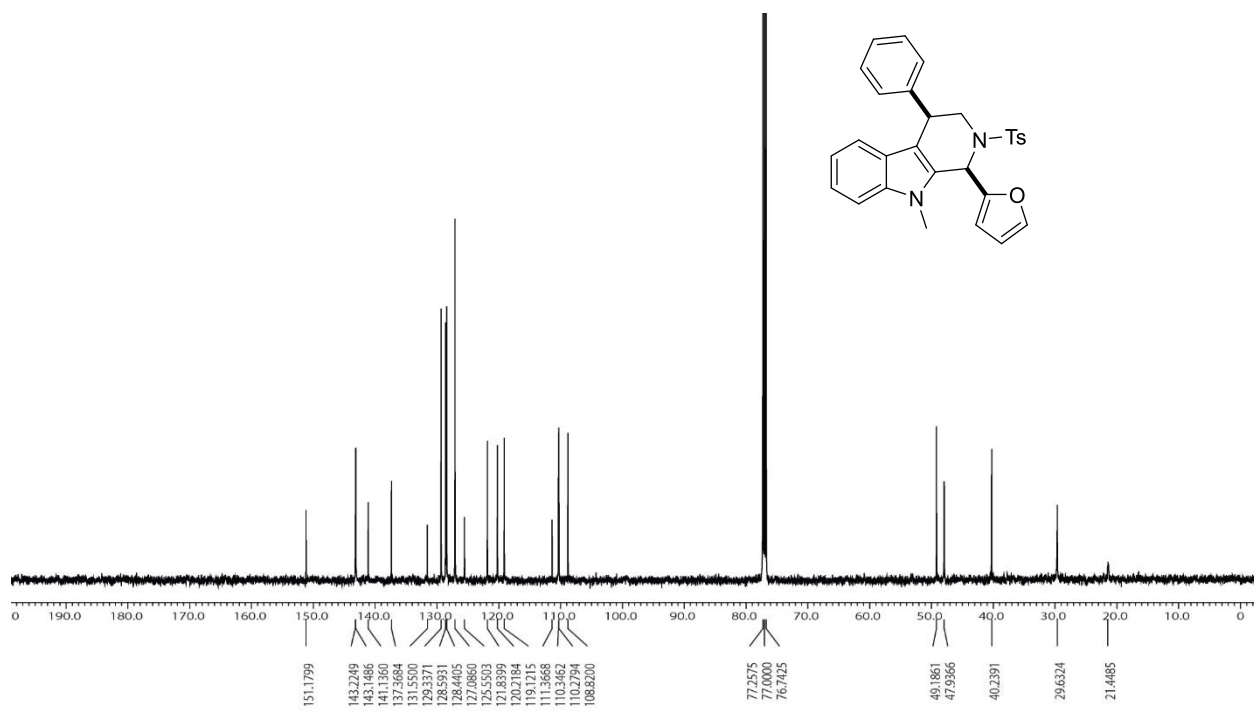


Figure:S30 ¹³C{¹H} NMR spectrum of **5m** (CDCl₃, 125 MHz)

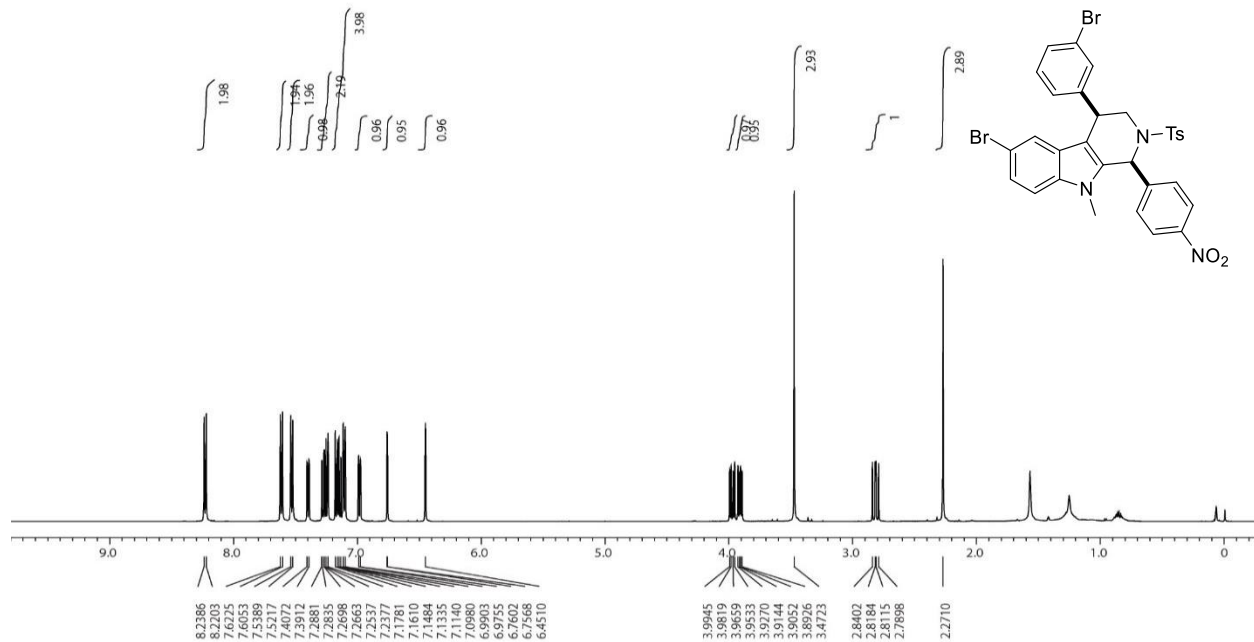


Figure:S33 ^1H NMR spectrum of **5o** (CDCl_3 , 500 MHz)

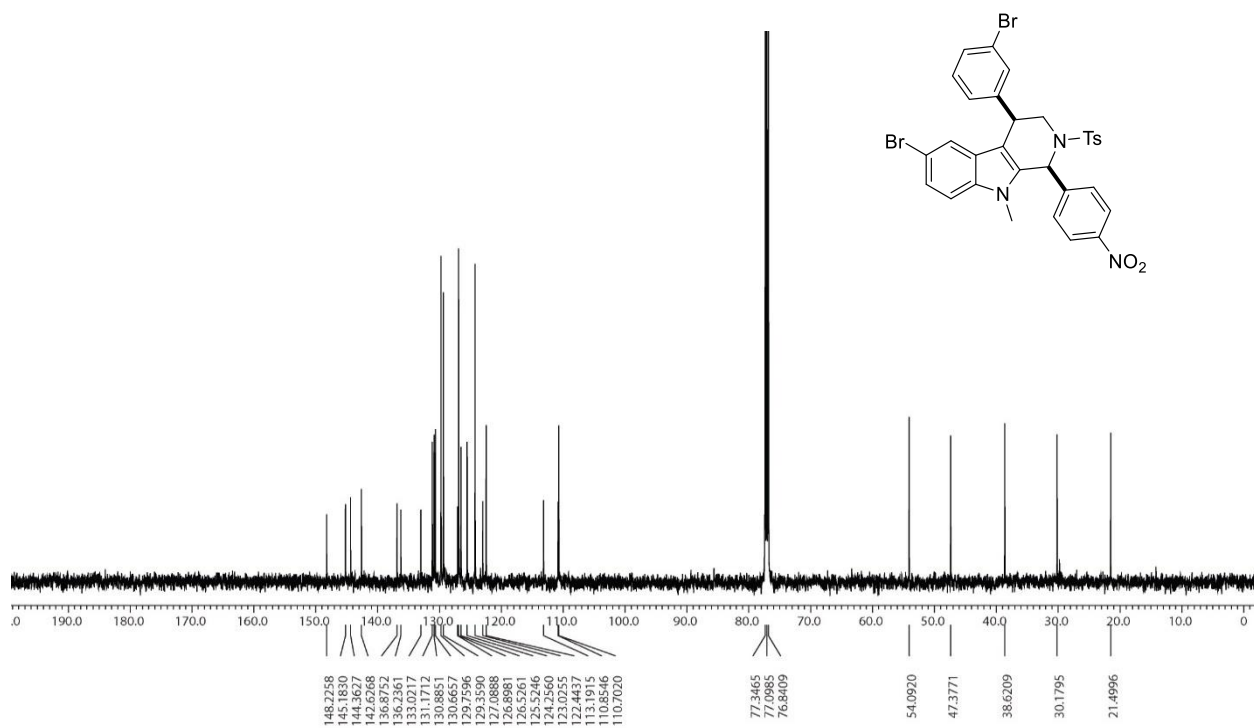


Figure:S34 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **5o** (CDCl_3 , 125 MHz)

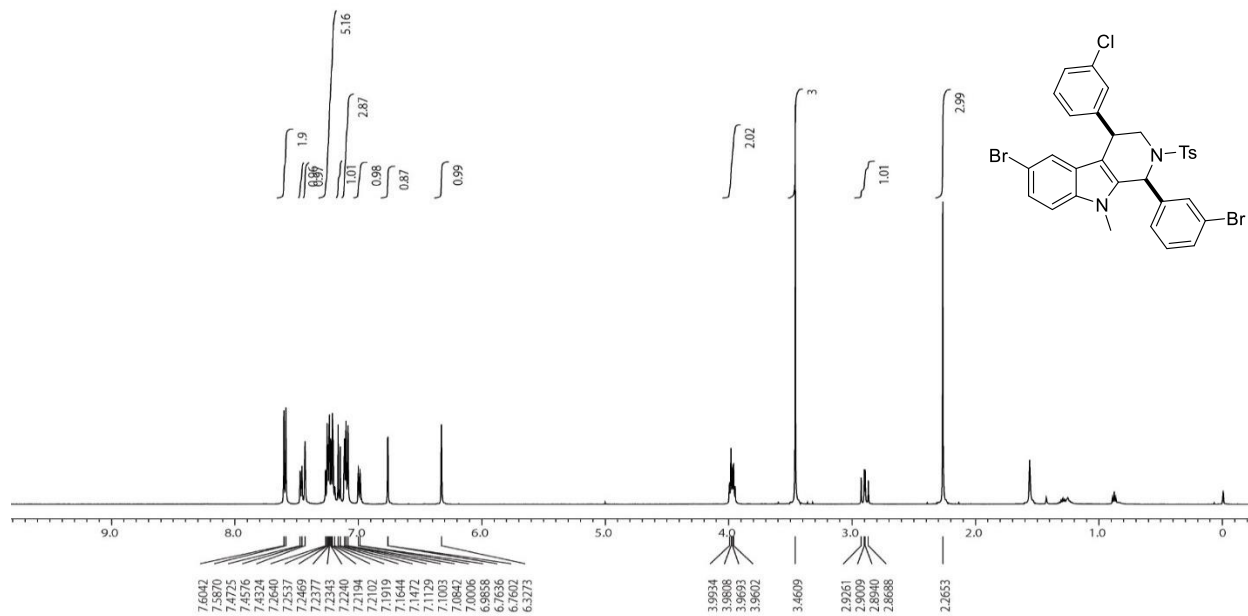


Figure:S35 ^1H NMR spectrum of **5p** (CDCl_3 , 500 MHz)

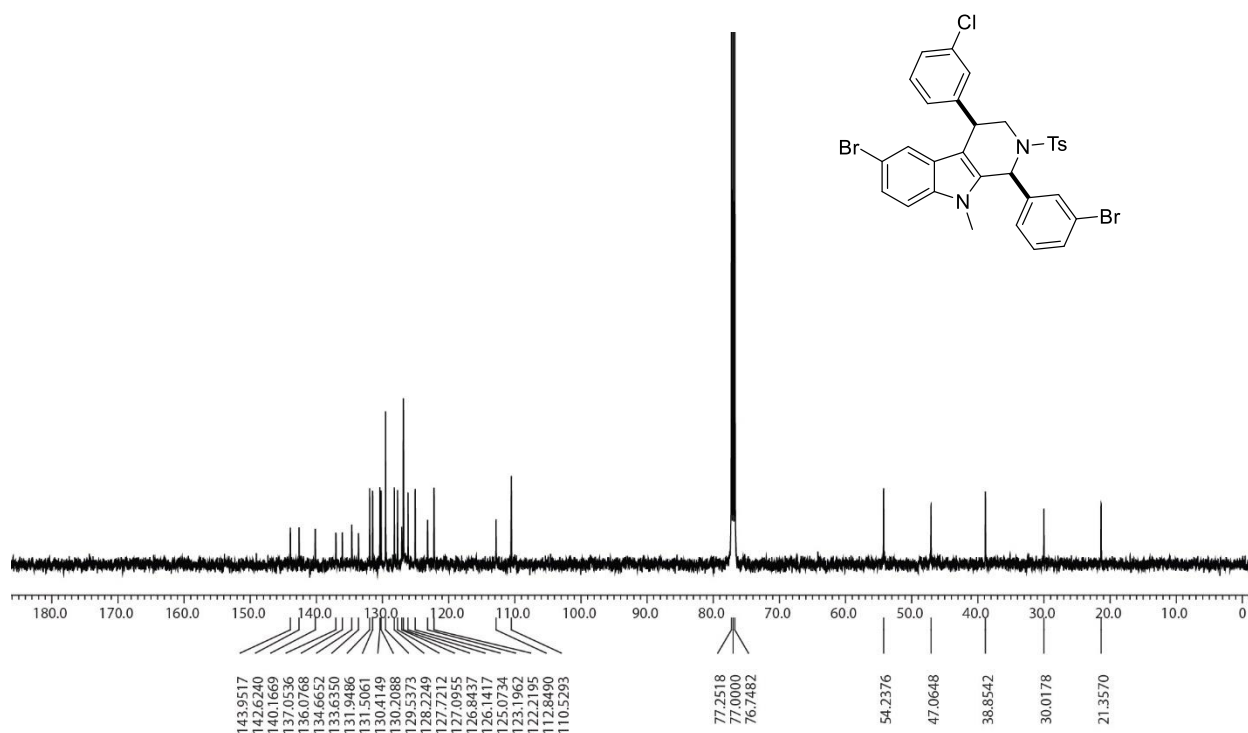


Figure:S36 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **5p** (CDCl_3 , 125 MHz)

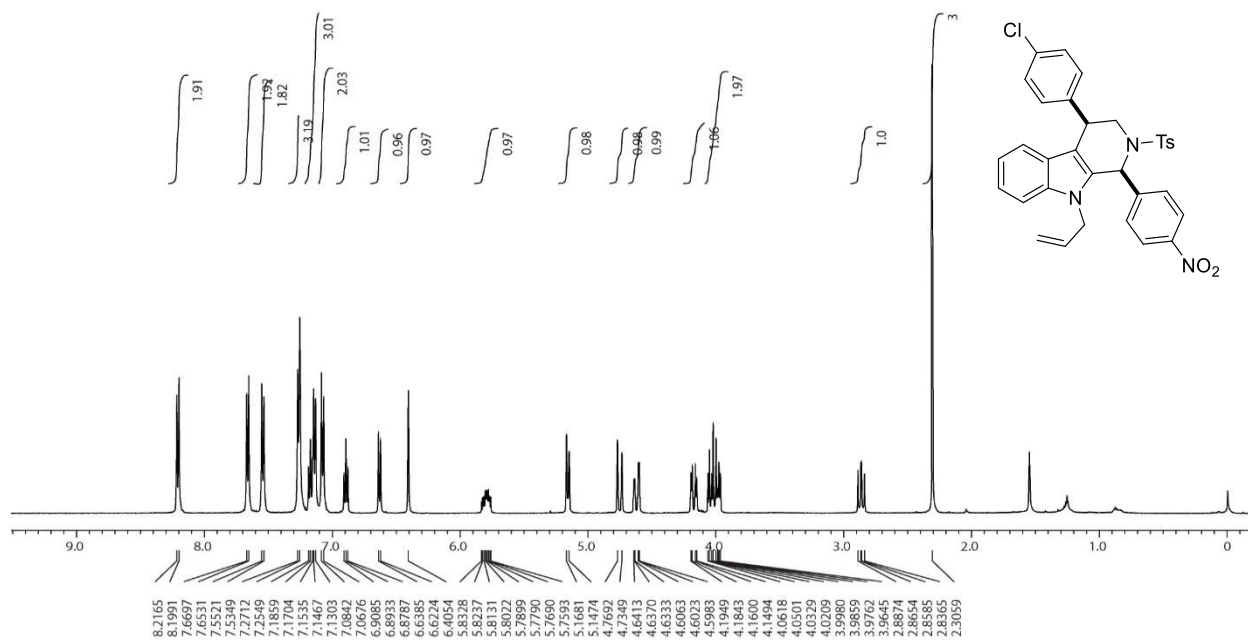


Figure:S37 ^1H NMR spectrum of **5q** (CDCl_3 , 500 MHz)

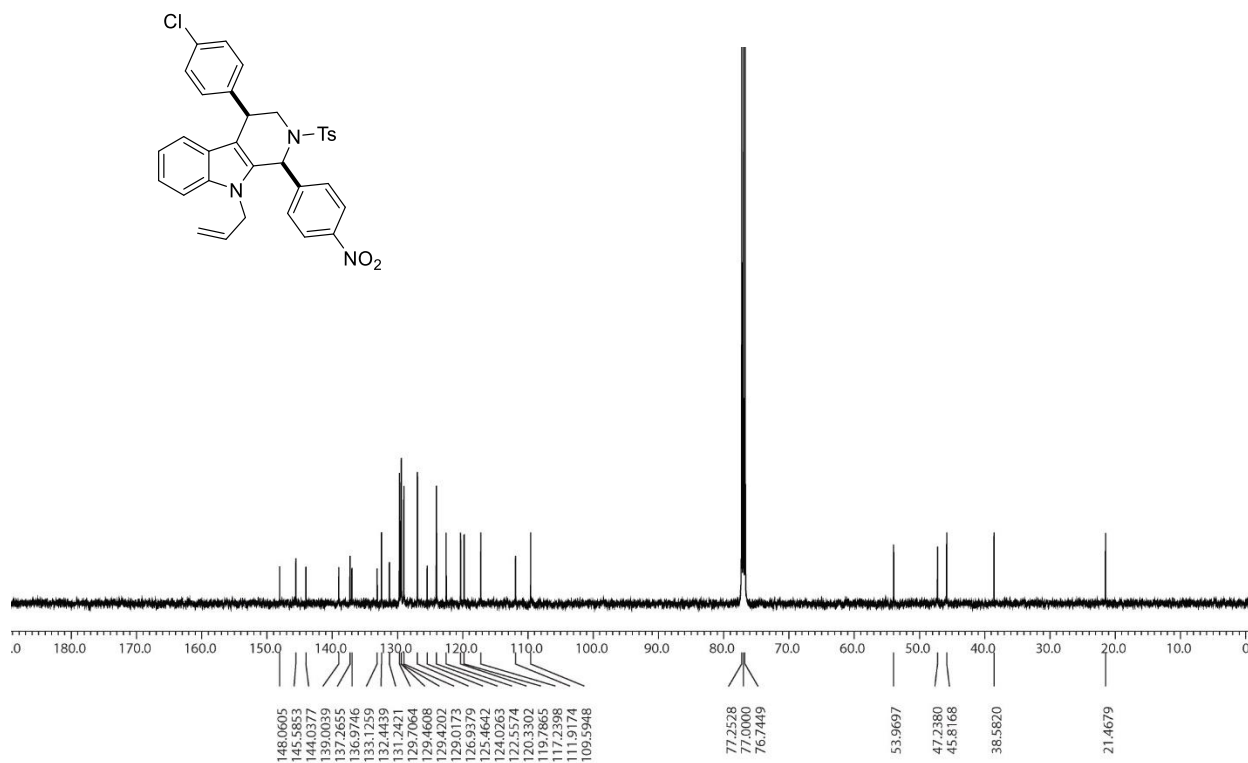
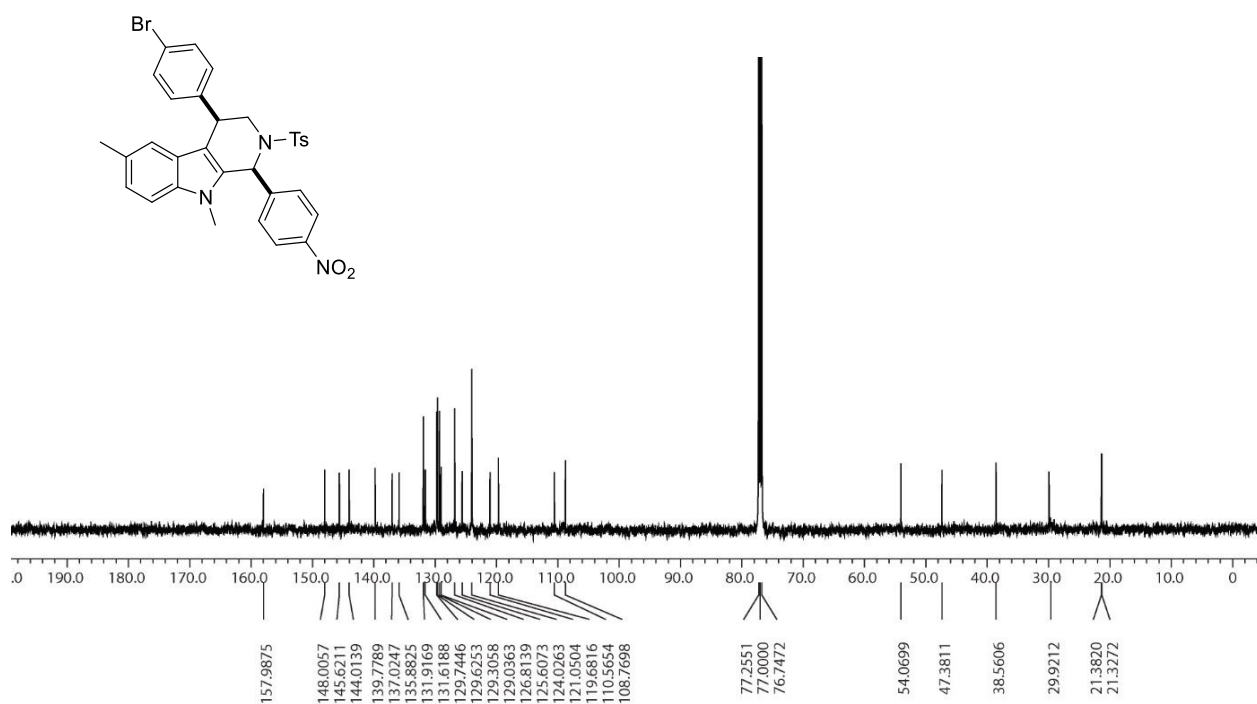
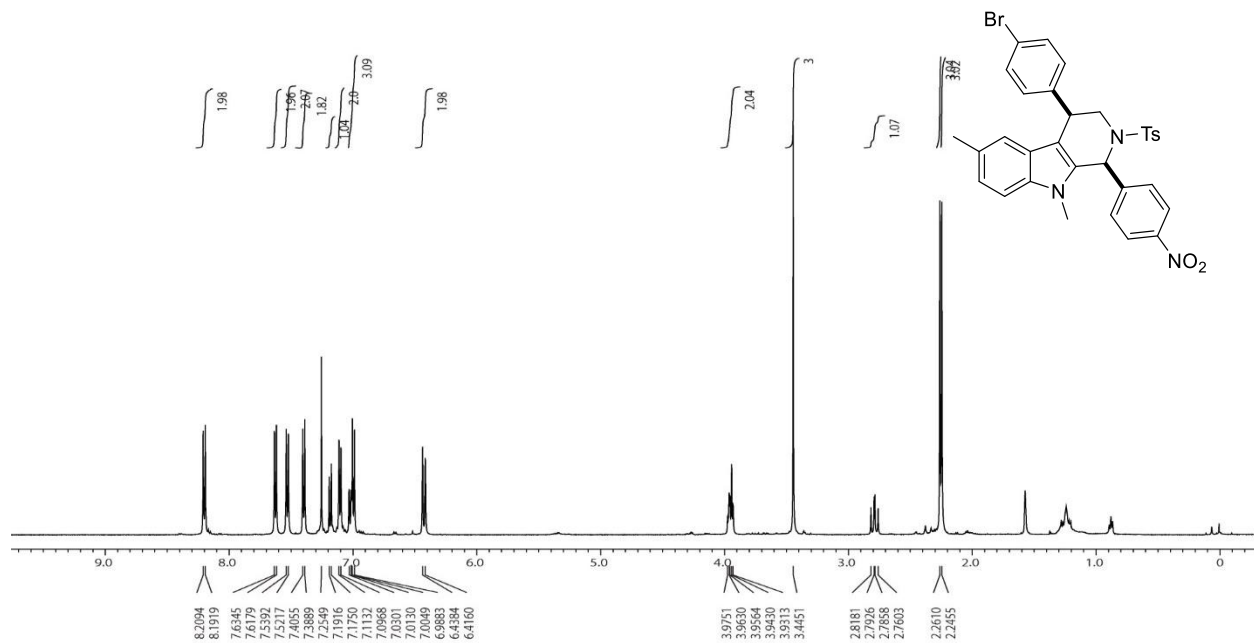


Figure:S38 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **5q** (CDCl_3 , 125 MHz)



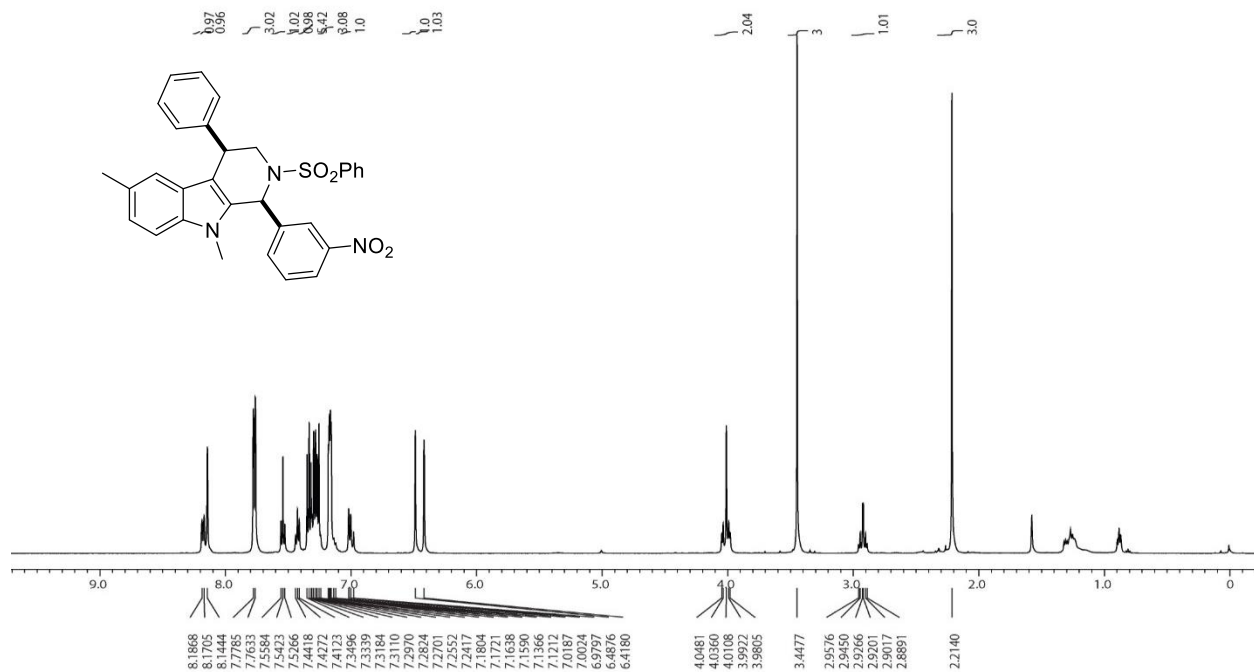


Figure:S41 ¹H NMR spectrum of **5s** (CDCl₃, 500 MHz)

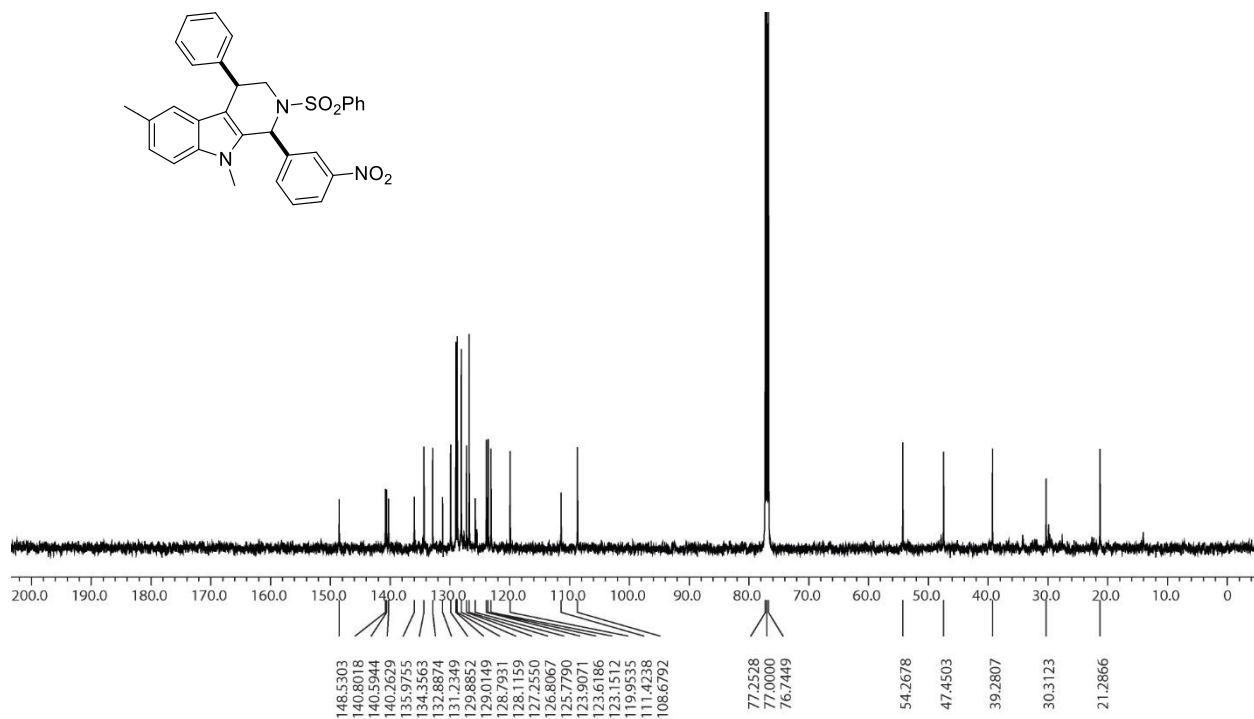


Figure:S42 ¹³C{¹H} NMR spectrum of **5s** (CDCl₃, 125 MHz)

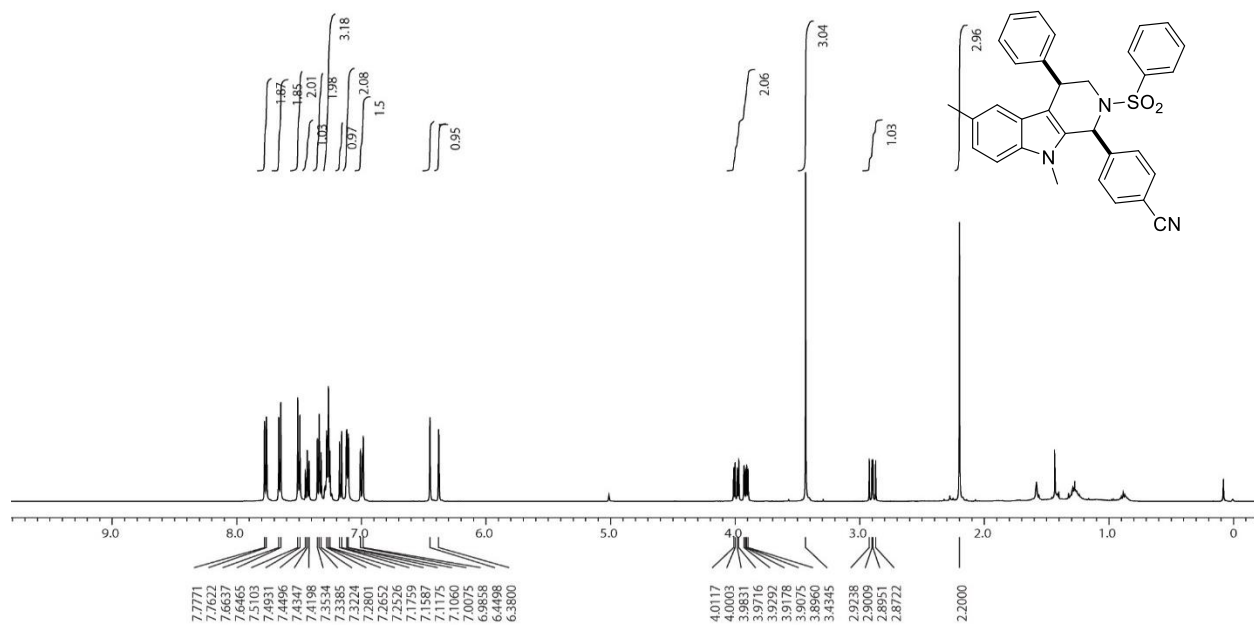


Figure:S43 ¹H NMR spectrum of **5t** (CDCl₃, 500 MHz)

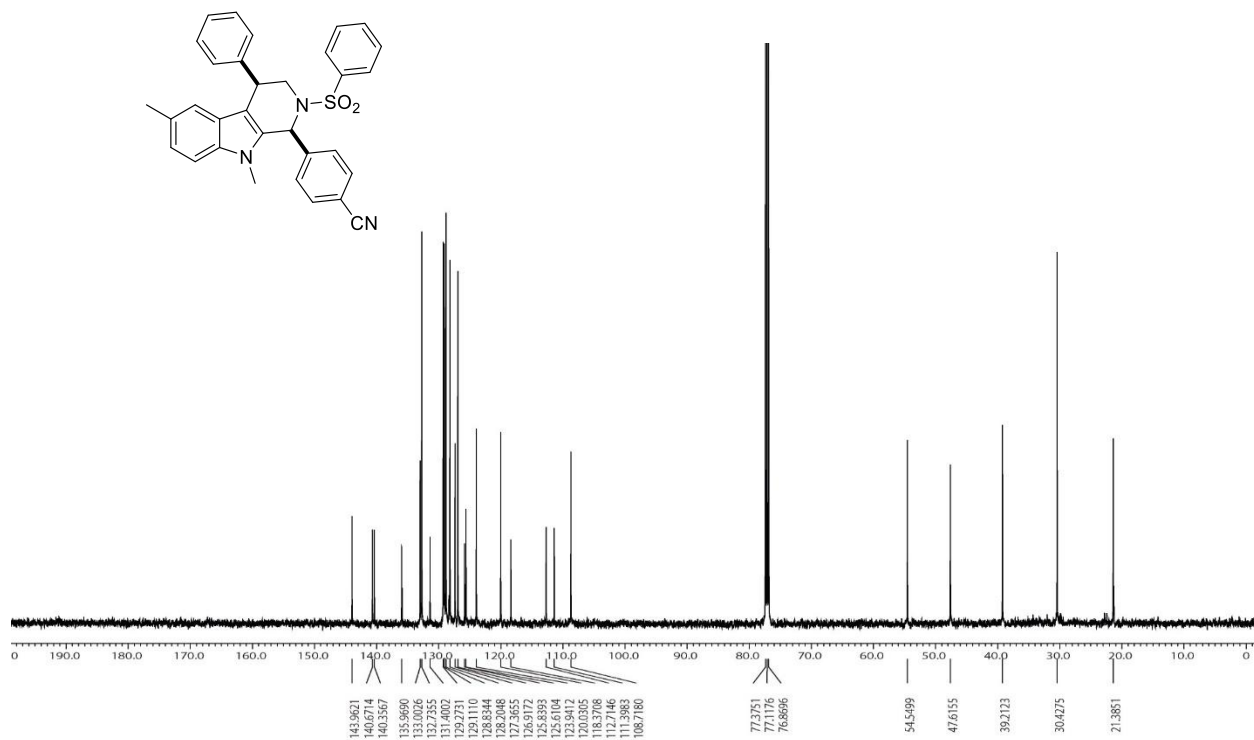


Figure:S44 ¹³C{¹H} NMR spectrum of **5t** (CDCl₃, 125 MHz)

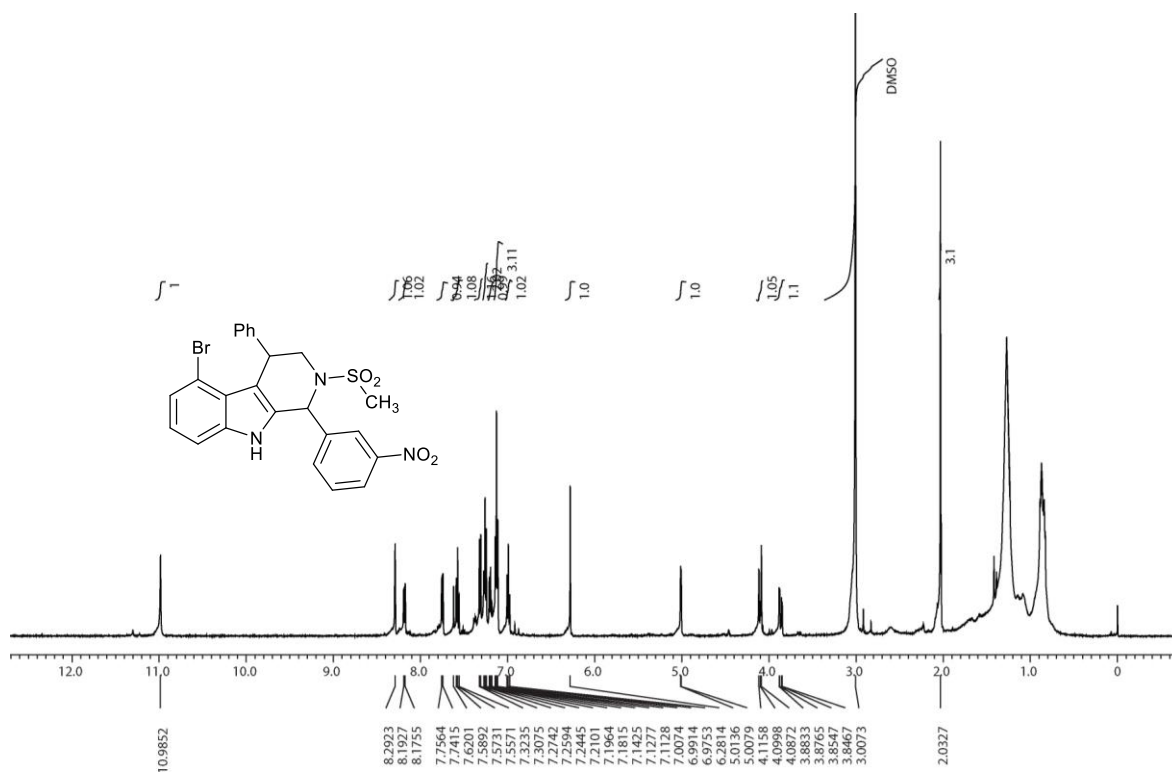


Figure:S45 ^1H NMR spectrum of **5u** (CDCl₃ + few drops of DMSO, 400 MHz)

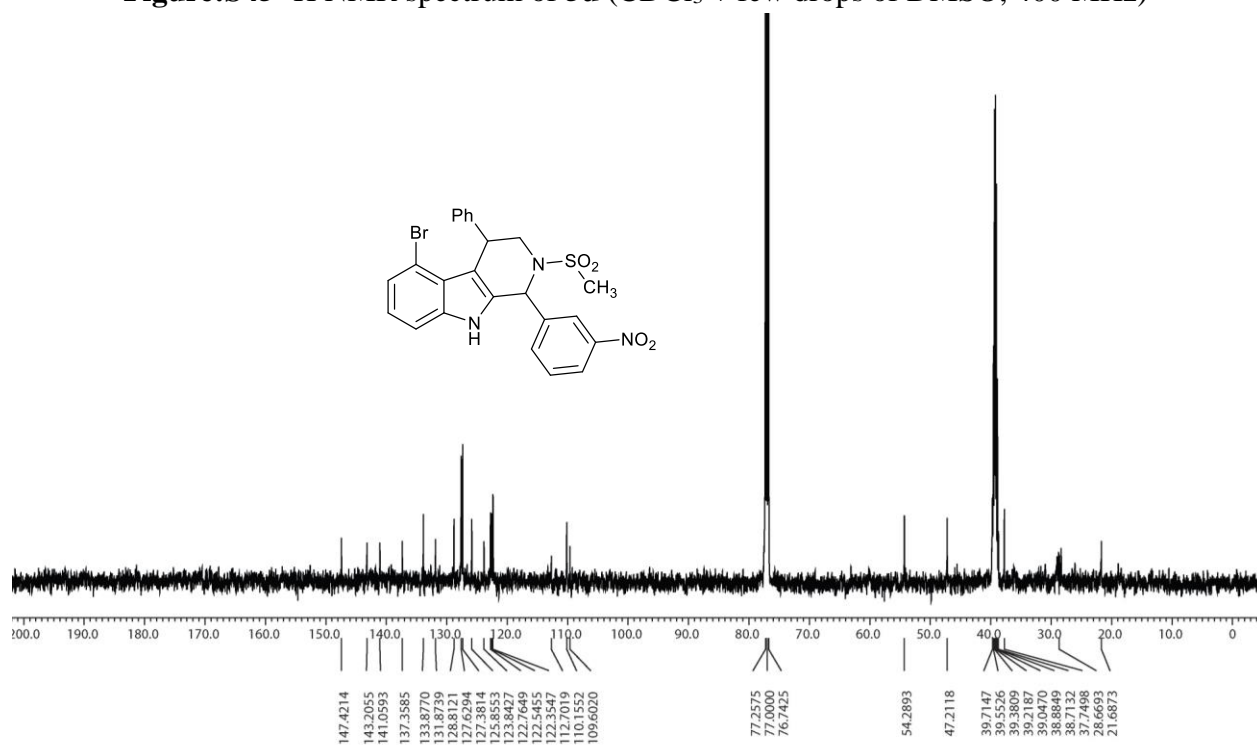


Figure:S46 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **5u** (CDCl₃, 100 MHz)

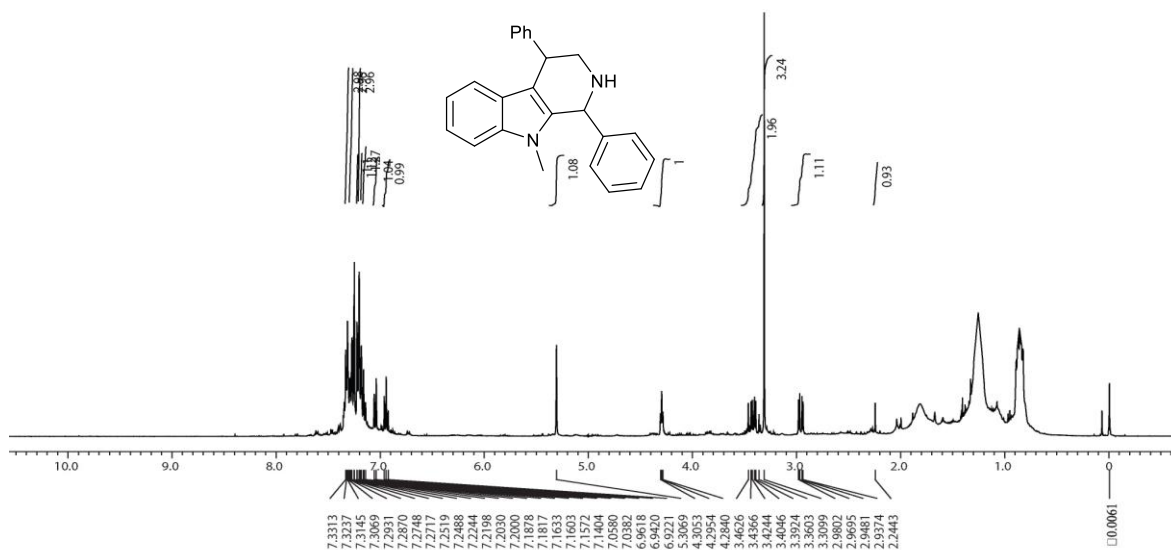


Figure:S47 ¹H NMR spectrum of **5v** (CDCl₃, 400 MHz)

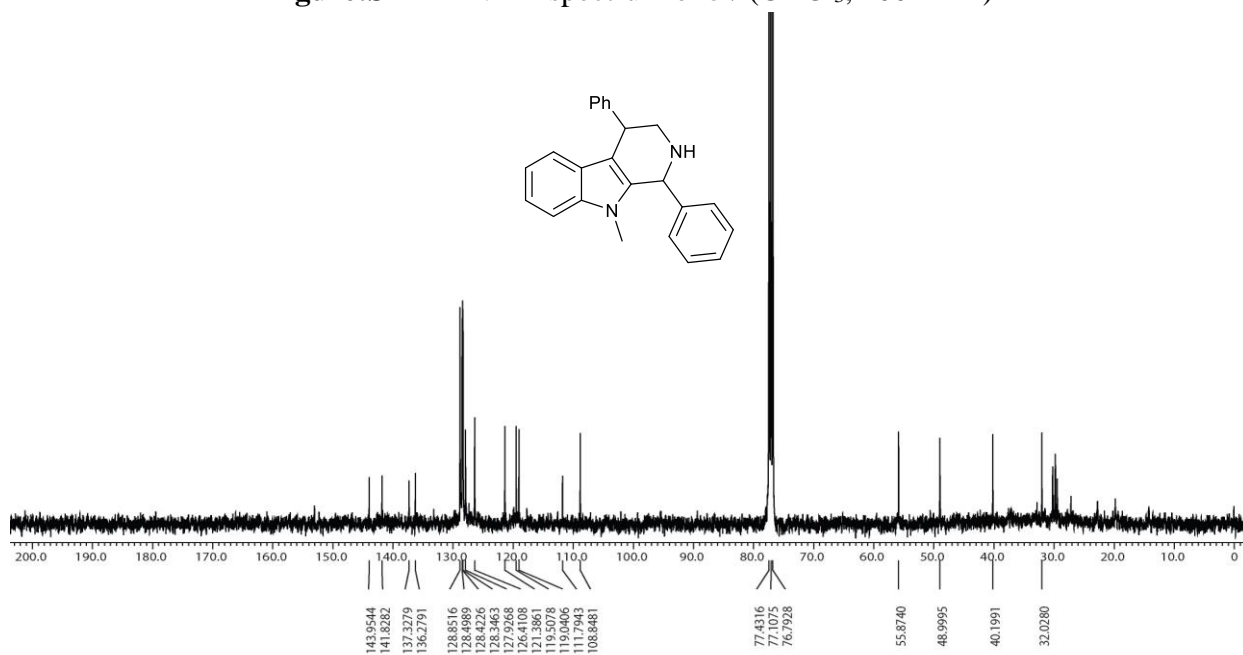


Figure:S48 ¹³C {¹H} NMR spectrum of **5v** (CDCl₃, 100 MHz)

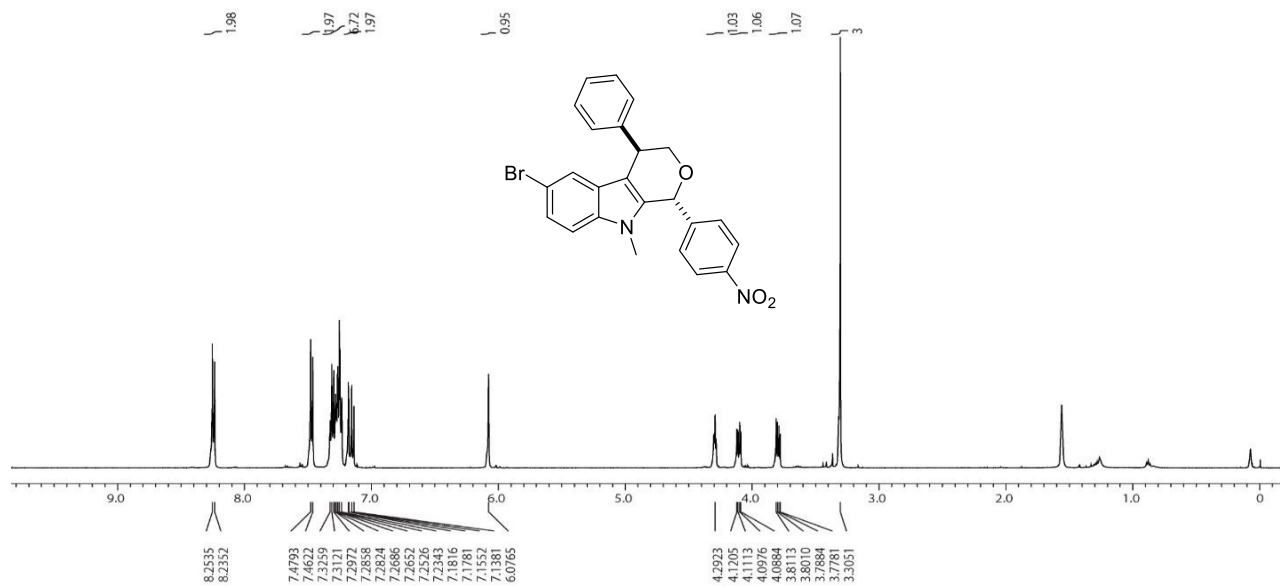


Figure:S49 ^1H NMR spectrum of **7a** (CDCl_3 , 500 MHz)

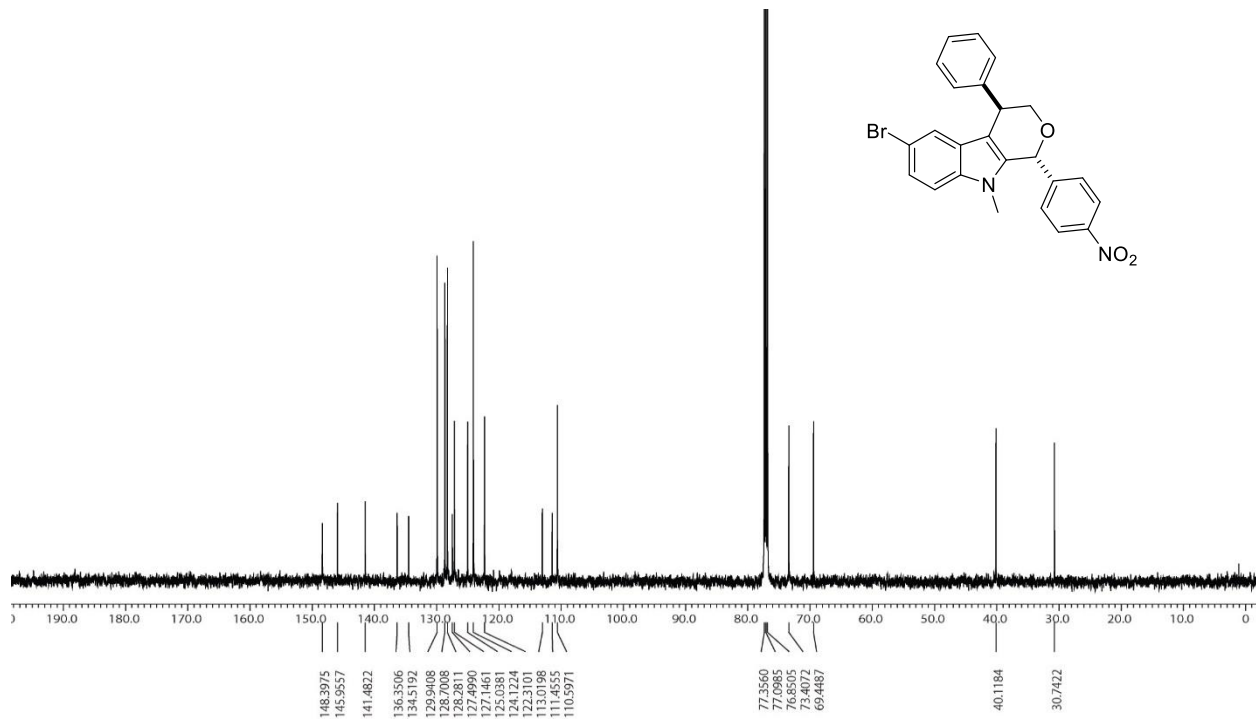


Figure:S50 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **7a** (CDCl_3 , 125 MHz)

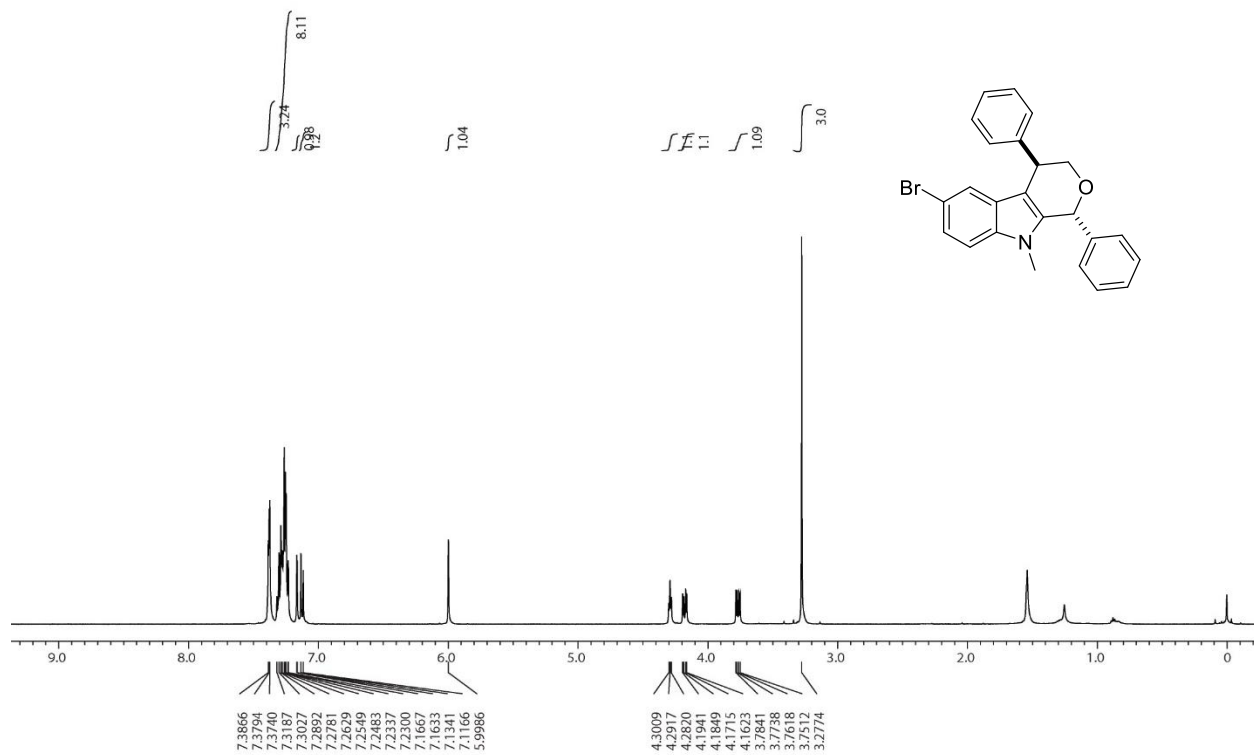


Figure:S51 ¹H NMR spectrum of **7b** (CDCl₃, 500 MHz)

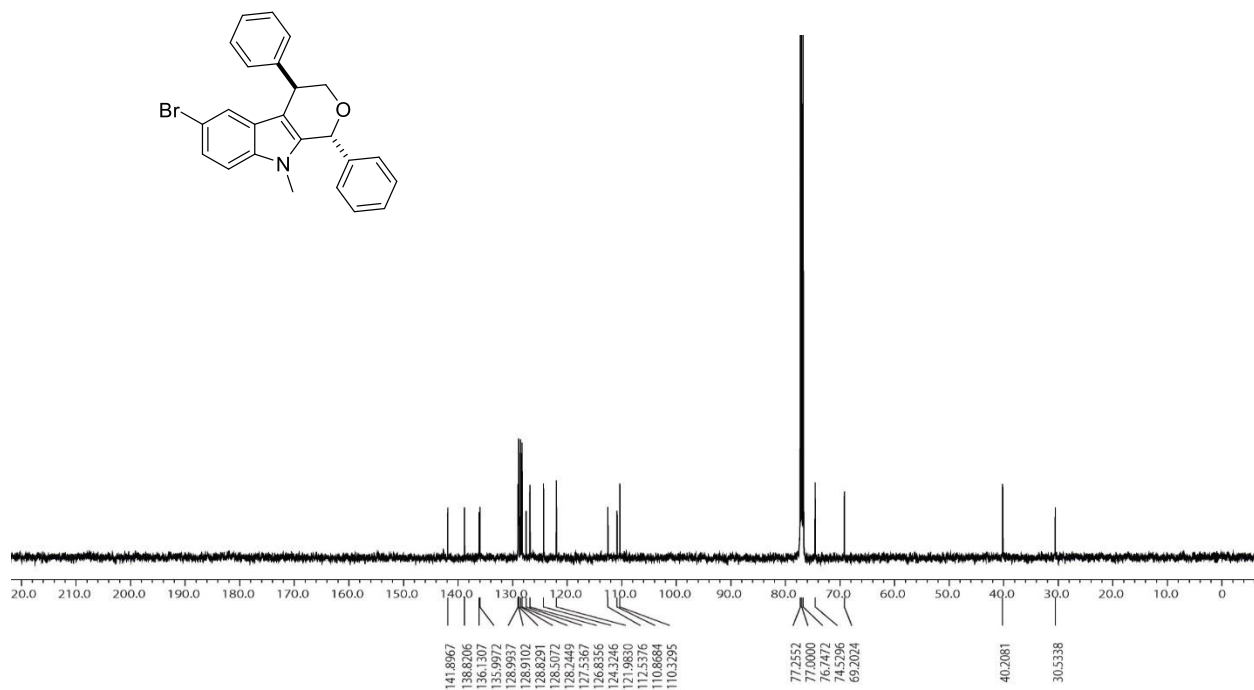


Figure:S52 ¹³C{¹H} NMR spectrum of **7b** (CDCl₃, 125 MHz)

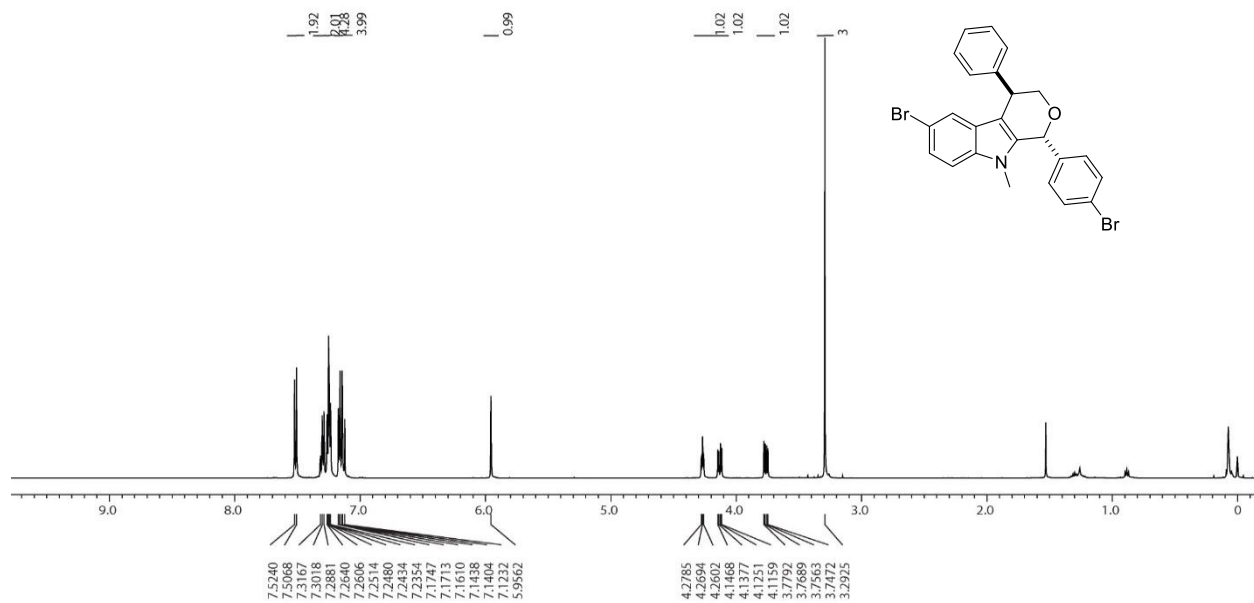


Figure:S53 ^1H NMR spectrum of **7c** (CDCl_3 , 500 MHz)

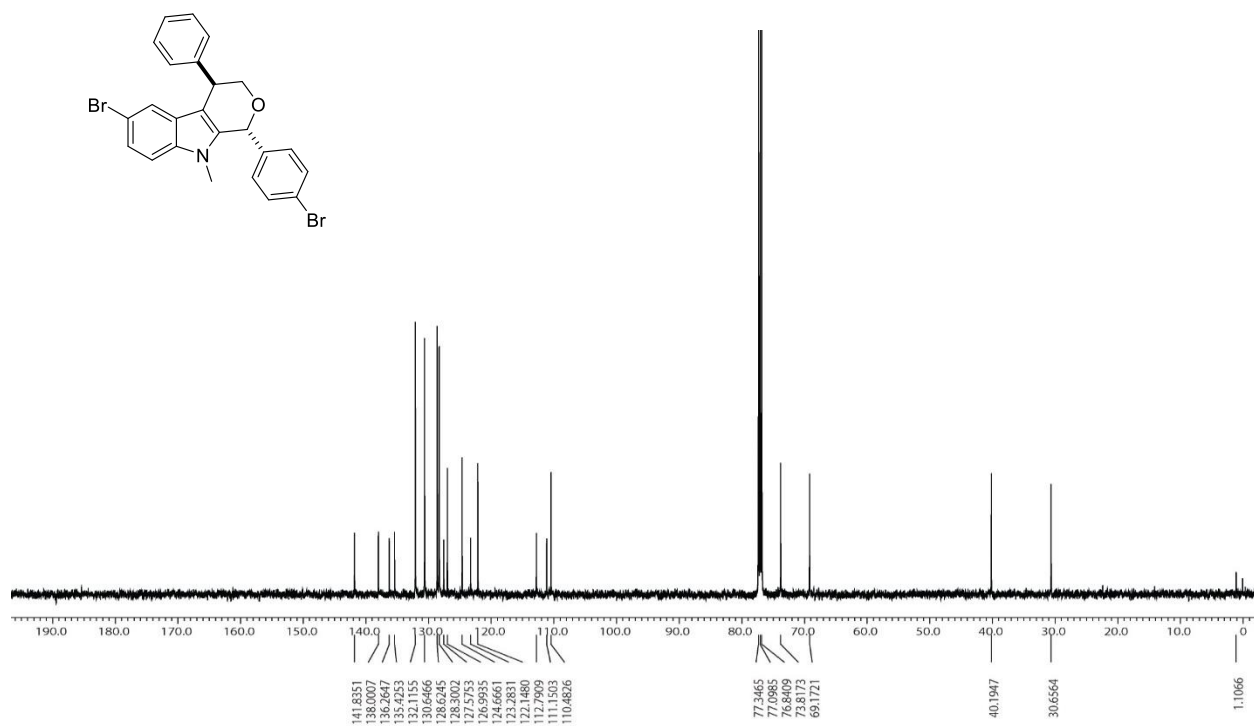


Figure:S54 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **7c** (CDCl_3 , 125 MHz)

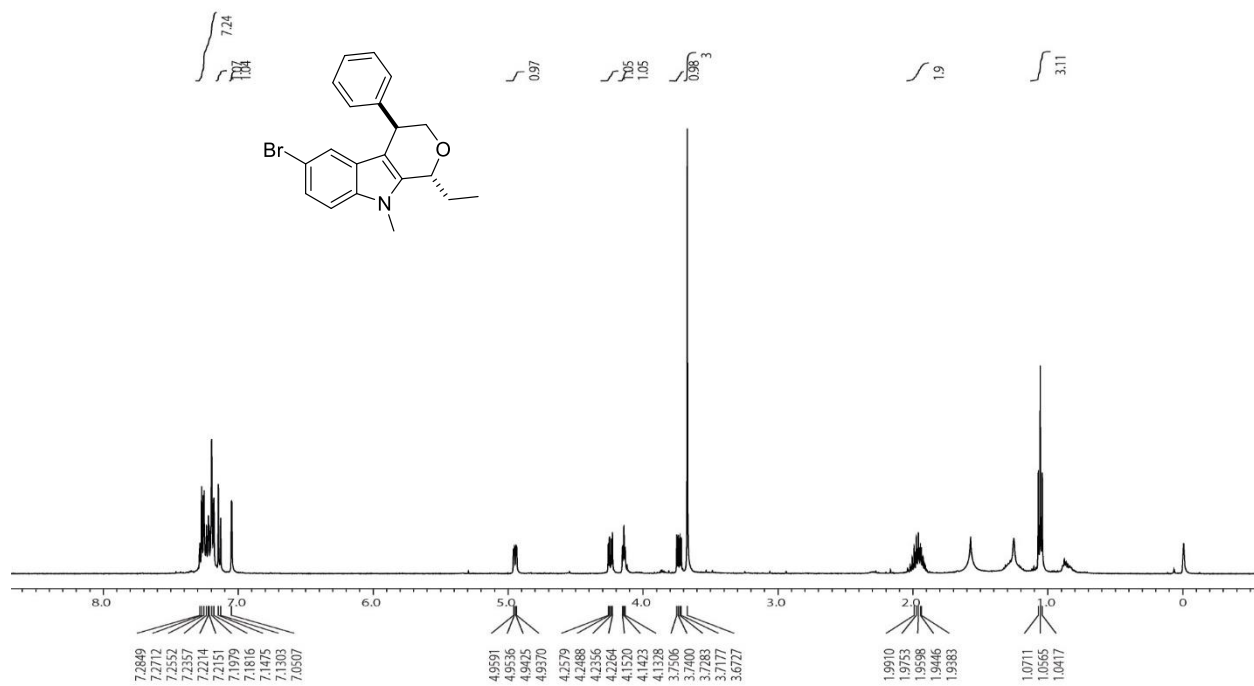


Figure:S55 ^1H NMR spectrum of **7d** (CDCl_3 , 500 MHz)

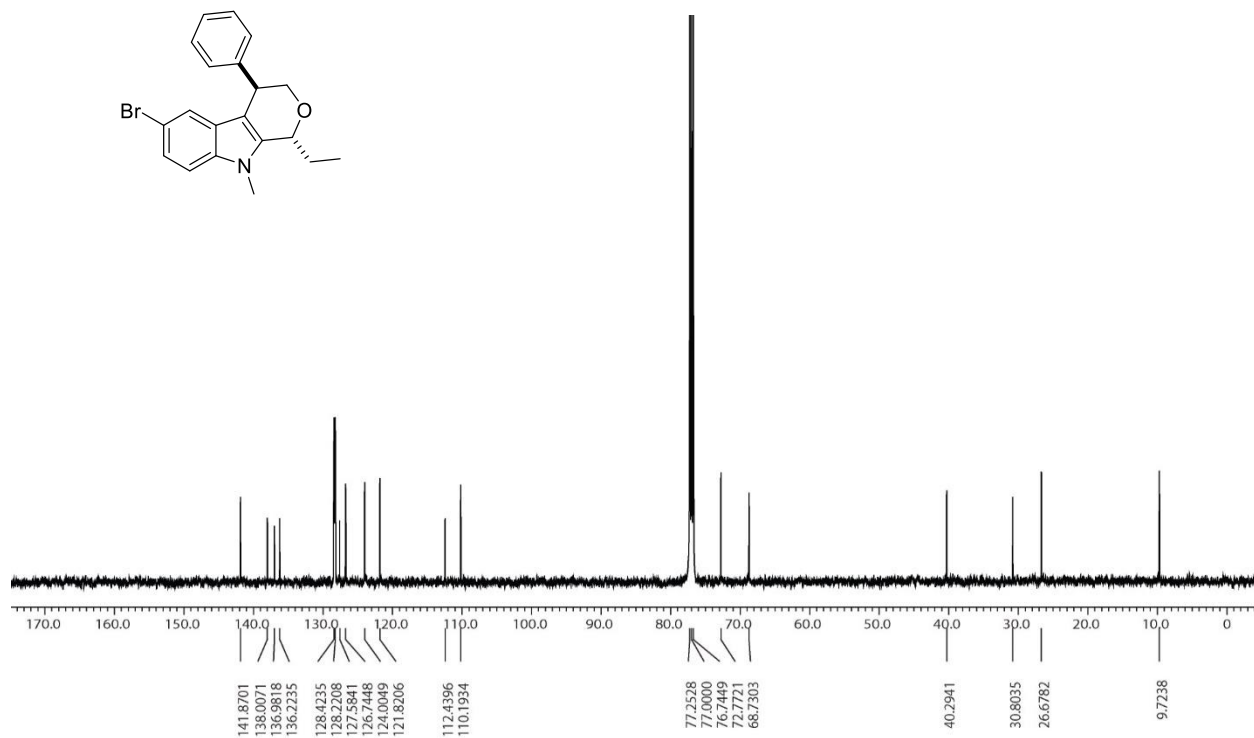


Figure:S56 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **7d** (CDCl_3 , 125 MHz)

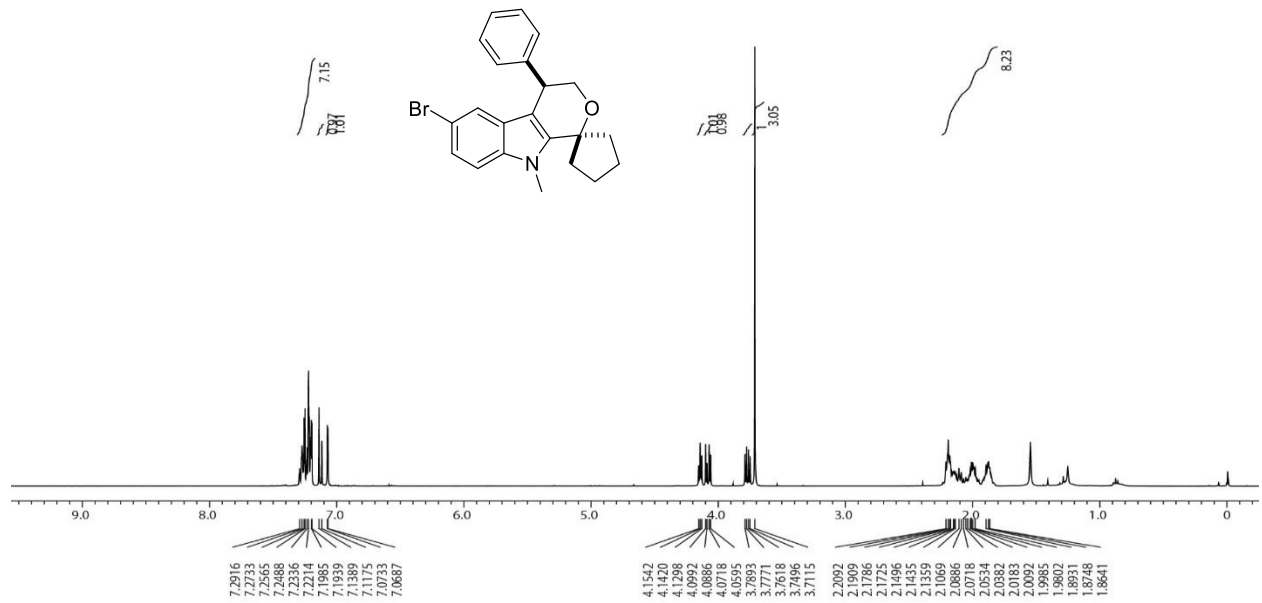


Figure:S57 ^1H NMR spectrum of **7e** (CDCl_3 , 500 MHz)

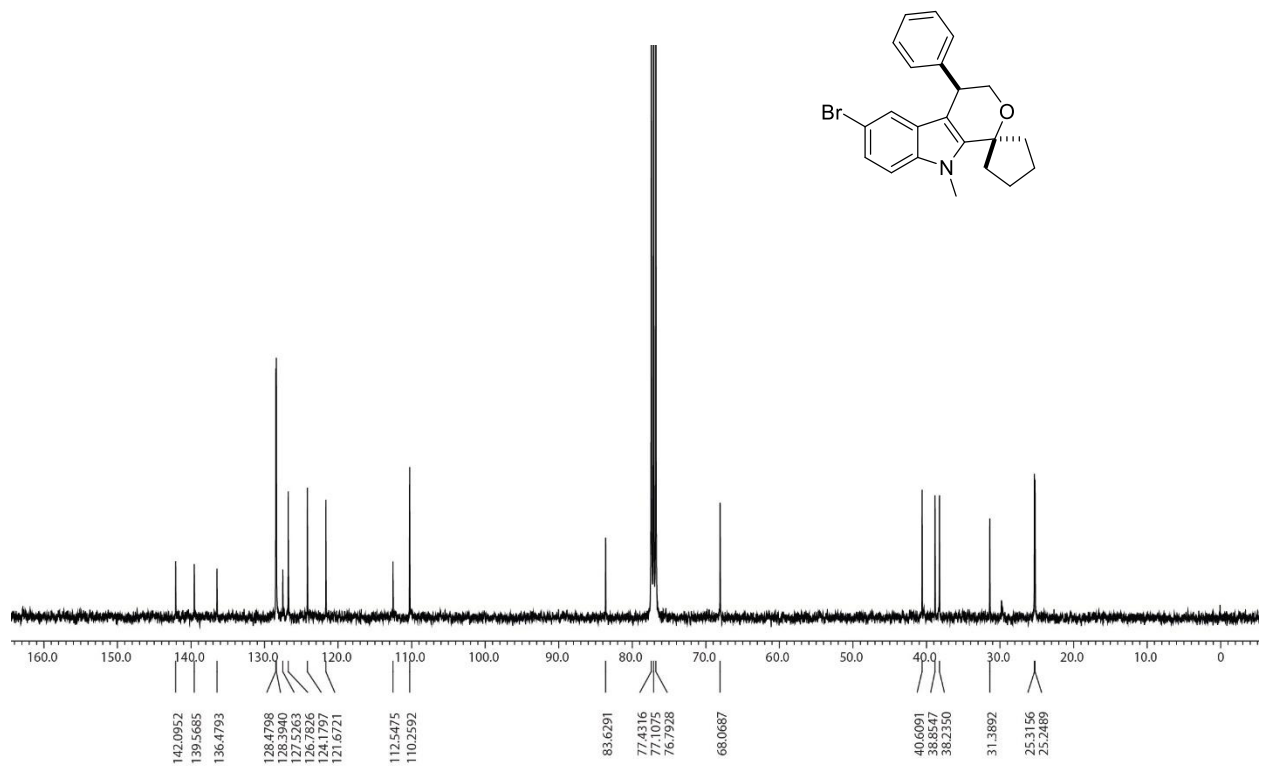


Figure:S58 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **7e** (CDCl_3 , 125 MHz)

5. Selected HPLC chromatogram for *ee* determination:

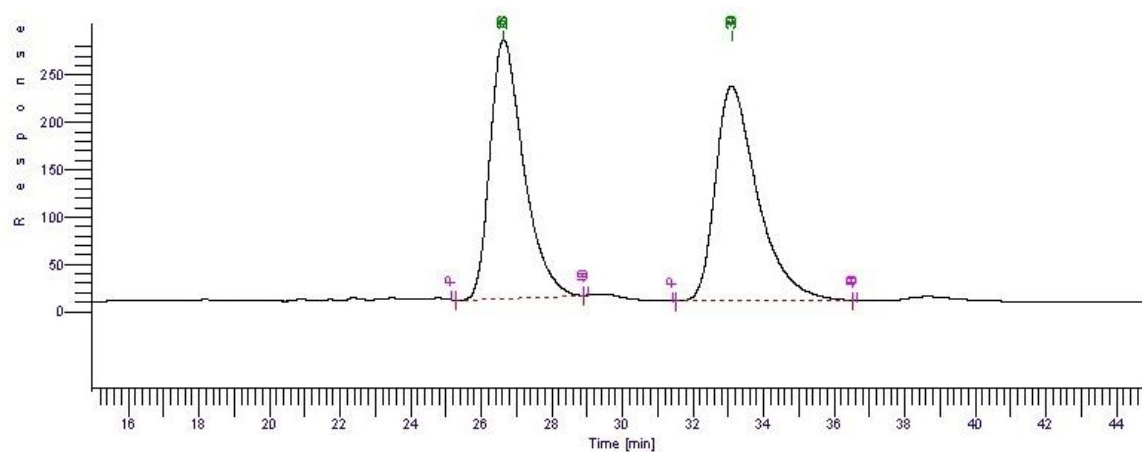


Figure:S59 HPLC chromatogram of racemic compound **5a** (AD-H column; 90:10 Hexane–Isopropanol; 1.0 mL min⁻¹)

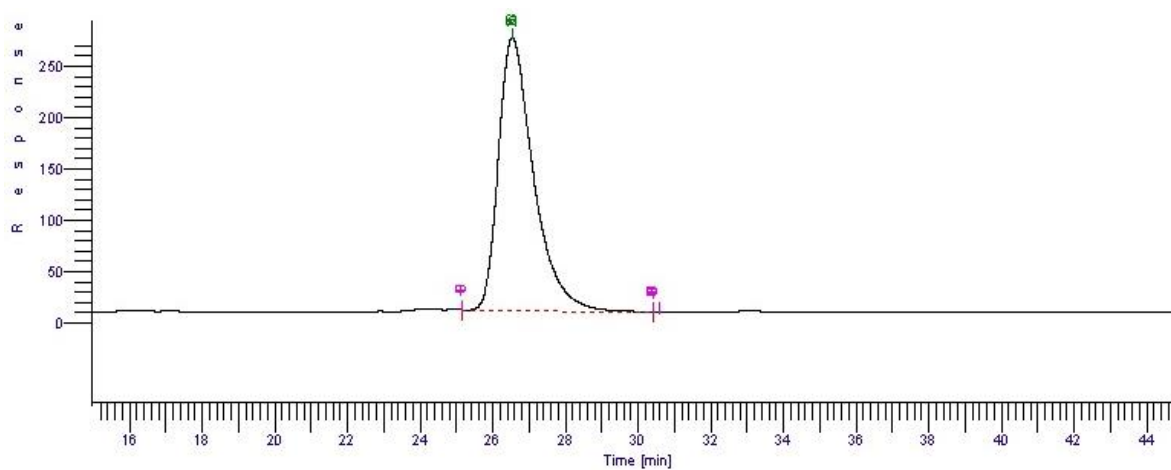


Figure:S60 HPLC chromatogram of chiral compound **5a** (>99% *ee*; AD-H column; 90:10 Hexane–Isopropanol; 1.0 mL min⁻¹)

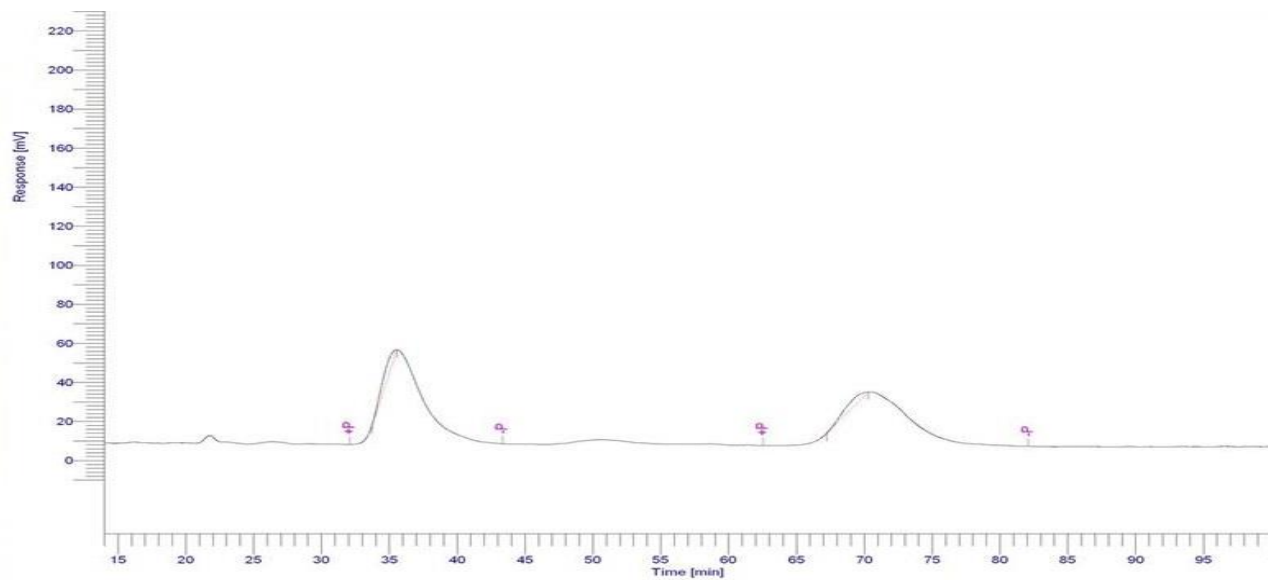


Figure:S61 HPLC chromatogram of racemic compound **5k** (OD-H column; 95:5 Hexane–Isopropanol; 1.0 mL min⁻¹)

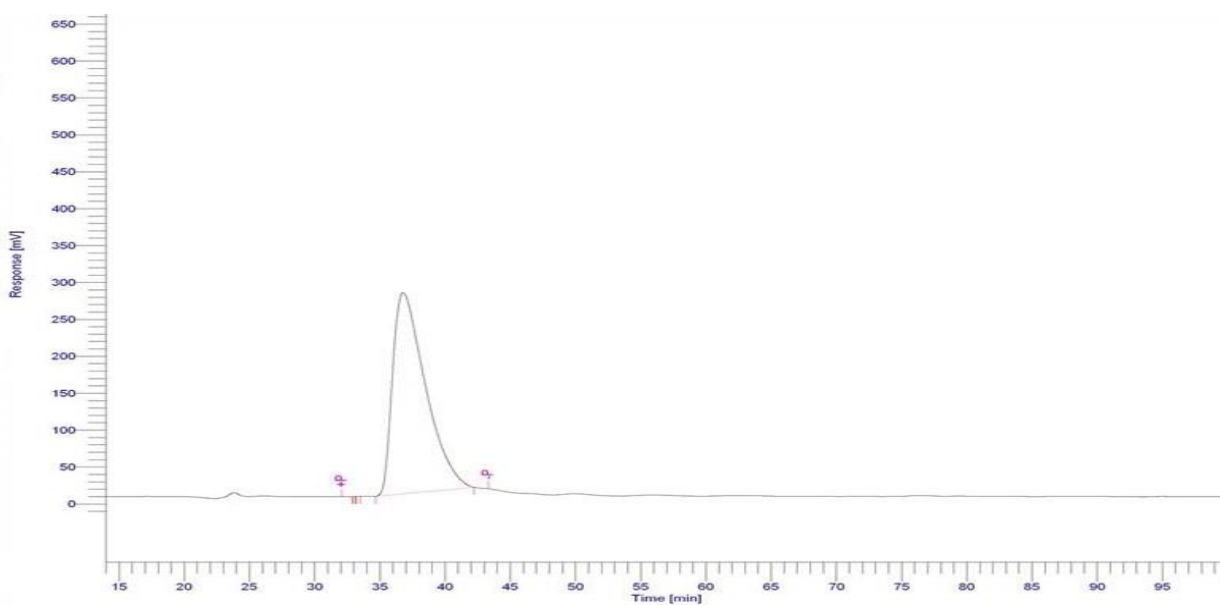


Figure:S562 HPLC chromatogram of chiral compound **5k** (OD-H column; 95:5 Hexane–Isopropanol; 1.0 mL min⁻¹)

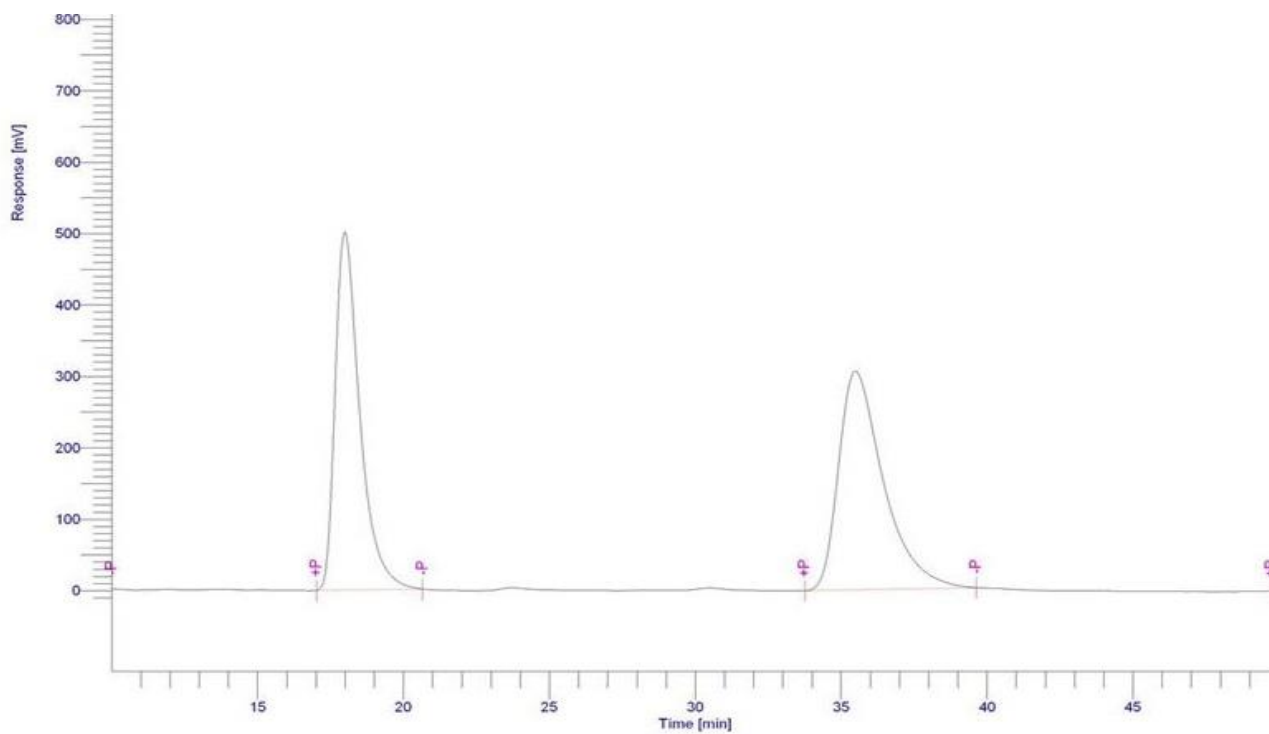


Figure:S63 HPLC chromatogram of racemic compound **5m** (AD-H column; 90:10 Hexane–Isopropanol; 1.0 mL min⁻¹)

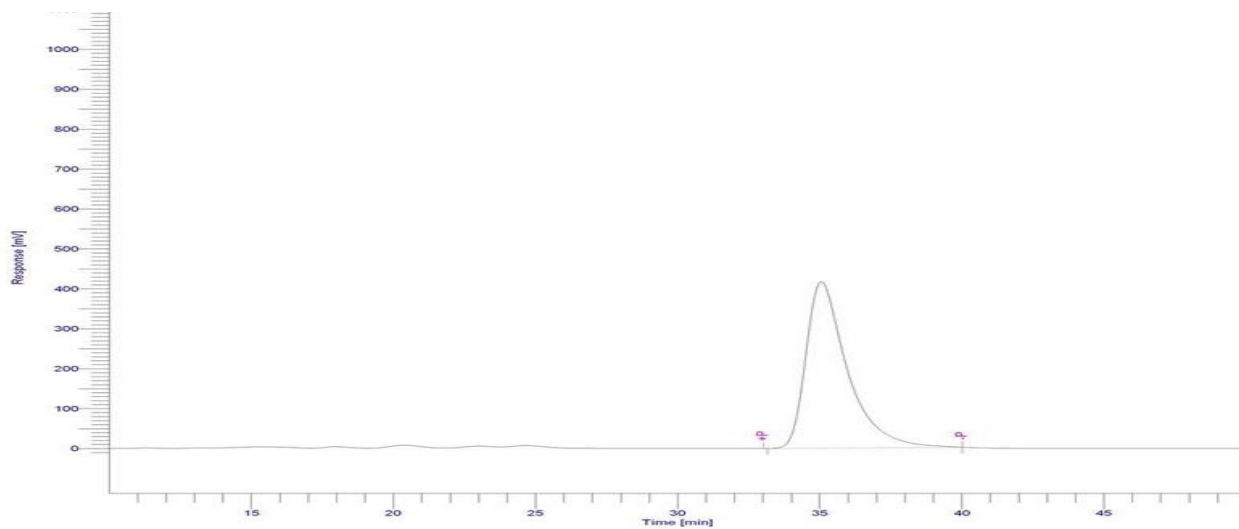


Figure:S64 HPLC chromatogram of chiral compound **5m** (AD-H column; 90:10 Hexane–Isopropanol; 1.0 mL min⁻¹)