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Supporting Information

Fused azole-thiazolines via one-pot cyclization of functionalized N-heterocyclic carbene precursors

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Table of Contents

Characterization Data	S2
Formation of SCN-propyl-pyrazole	S21
NMR and HRMS spectra	S23
Selected X-ray crystallographic data	S93



The bromopropyl-imidazolium salt I1 (388 mg, 1.00 mmol) and KSCN (107 mg, 1.10 mmol) were suspended in CH_3CN (50 mL) and stirred overnight before the solvent was removed under reduced pressure. CH_2Cl_2 (120 mL) was added to dissolve the

product and the resulting suspension was filtered and dried to

afford the product as a yellowish solid. Yield: 337 mg, 0.92 mmol, 92%. ¹H NMR (300 MHz, CDCl₃): δ 9.98 (s, 1 H, NCHN), 8.24 (s, 1 H, Imi–H), 7.18 (s, 1 H, Imi–H), 6.93 (s, 2 H, Ar–H), 4.85 (t, ³*J*(H,H) = 7 Hz, 2 H, NCH₂), 3.20 (t, ³*J*(H,H) = 7 Hz, 2 H, SCH₂), 2.54 (m, ³*J*(H,H) = 7 Hz, 2 H, CH₂), 2.26 (s, 3 H, CH₃), 1.99 (s, 6 H, CH₃). ¹³C {¹H} NMR (101 MHz, CDCl₃): 142.1 (NCHN), 138.6, 134.8, 131.2, 130.6, 124.13, 124.07 (Ar–C), 112.8 (SCN), 49.0 (NCH₂), 31.5, 31.4 (CH₂), 21.7, 18.4 (CH₃). HRMS (ESI) m/z calcd. for C₁₆H₂₀N₃S [M – Br]⁺ 286.1371; found, 286.1372.



The thiocyanatopropyl-imidazolium salt II1 (37 mg, 0.10 mmol) and a NaOH aqueous solution (1 M, 10 μ L) were mixed in CH₃CN (10 mL) and stirred for 1 h before KSCN (49 mg, 0.50 mmol) was added. The reaction mixture was stirred overnight and the solvent was removed under reduced pressure.

CH₂Cl₂ (80 mL) was added and the resulting solution was filtered. Removal of the solvent from the filtrate yielded the product as an off-white solid. Yield: 28 mg, 0.09 mmol, 87%. ¹H NMR (500 MHz, CDCl₃): δ 7.87 (s, 1 H, Imi–H), 7.12 (s, 1 H, Imi–H), 6.93 (s, 2 H, Ar–H), 4.59 (br-s, 2 H, NCH₂), 3.45 (br-s, 2 H, SCH₂), 2.50 (br-s, 2 H,

CH₂), 2.25 (s, 3 H, CH₃), 1.95 (s, 6 H, CH₃). ¹³C {¹H} NMR (126 MHz, CDCl₃): 142.8 (NCN), 142.2, 135.7, 130.4, 129.0, 125.5, 122.0 (Ar–C), 47.3 (NCH₂), 26.4 (SCH₂), 22.4 (CH₂), 21.5, 17.8 (CH₃), SCN⁻ signal could not be detected. Anal. Calcd. for $C_{16}H_{19}N_3S_2$: C, 60.53; H, 6.03; N, 13.24. Found: C, 60.15; H, 5.79; N, 13.22. HRMS (ESI) m/z calcd. for $C_{15}H_{19}N_2S$ [M – SCN]⁺ 259.1263; found, 259.1269.



This compound was prepared in analogy to **II1** from **I2** (374 mg, 1.00 mmol) and KSCN (486 mg, 5.00 mmol) as a pinkish solid. Yield: 326 mg, 0.99 mmol, 99%. ¹H NMR (400 MHz, CDCl₃ with 5 drops of CD₃CN): δ 9.35 (br-s, 1 H, NCHN), 8.01 (br-s, 1 H, Imi–H), 7.25 (br-s, 1 H, Imi–H), 6.95 (br-s, 2 H, Ar–H), 4.94 (br-s, 2 H, NCH₂), 3.73 (br-s, 2 H, SCH₂), 2.27 (s,

3 H, CH₃), 2.02 (s, 6 H, CH₃). ¹³C{¹H} NMR (101 MHz, CDCl₃ with 5 drops of CD₃CN): 141.8 (NCHN), 138.1, 134.7, 130.9, 130.2, 124.5, 124.4 (Ar–C), 111.7 (SCN), 50.0 (NCH₂), 34.4 (SCH₂), 21.4, 17.8 (CH₃), SCN⁻ signal could not be detected. HRMS (ESI) m/z calcd. for C₁₅H₁₈N₃S [M – SCN]⁺ 272.1216; found, 272.1224.



Compound 2 was prepared in analogy to 1 from II2 (33 mg, 0.10 mmol), NaOH aqueous solution (1M, 10 μ L) and KSCN (48 mg, 0.50 mmol) as a white solid. Yield: 28 mg, 0.09 mmol, 92%. ¹H NMR (500 MHz, *d*₄-Methanol): δ 7.81 (s, 1 H, Imi–H), 7.58 (s, 1 H, Imi–H), 7.13 (s, 2 H, Ar–H), 4.76 (t, ³*J*(H,H) = 8 Hz, 2 H,

NCH₂), 4.36 (t, ³*J*(H,H) = 8 Hz, 2 H, SCH₂), 2.36 (s, 3 H, CH₃), 2.13 (s, 6 H, CH₃). ¹³C{¹H} NMR (126 MHz, *d*₄-Methanol): 153.8 (NCN), 143.1, 136.1 (Ar–C), 133.6 (SCN⁻), 131.8, 131.0, 128.7, 121.9 (Ar–C), 51.1 (NCH₂), 39.1 (SCH₂), 21.2, 17.5 (CH₃). Anal. Calcd. for C₁₅H₁₇N₃S₂: C, 59.37; H, 5.65; N, 13.85. Found: C, 59.33; H, 5.57; N, 13.73. HRMS (ESI) m/z calcd. for C₁₄H₁₇N₂S [M – SCN]⁺ 245.1107; found, 245.1115.



A mixture of *N*-mesityl imidazole (931 mg, 5.00 mmol) and 1,4dibromobutane (12 mL) was stirred at 90 °C for one day. The excess 1,4-dibromobutane was recovered by vacuum distillation, and CH_2Cl_2 was added to the residue. The resulting suspension was filtered through Celite. The filtrate was dried and washed with diethyl ether (5 × 10 mL) to afford the product as a white solid.

Yield: 1.55 g, 3.85 mmol, 77%. ¹H NMR (300 MHz, CDCl₃): δ 10.48 (s, 1 H, NCHN), 7.71 (s, 1 H, Imi–H), 7.17 (s, 1 H, Imi–H), 7.01 (s, 2 H, Ar–H), 4.85 (t, ³*J*(H,H) = 7 Hz, 2 H, NCH₂), 3.51 (t, ³*J*(H,H) = 7 Hz, 2 H, SCH₂), 2.34 (s, 3 H, CH₃), 2.26–2.18 (m, 2 H, CH₂), 2.08 (s, 6 H, CH₃), 2.06–2.01 (m, 2 H, CH₂). ¹³C{¹H} NMR (75 MHz, CDCl₃): 140.6 (NCHN), 136.9, 133.7, 130.3, 129.3, 123.6, 123.0 (Ar–C), 48.6 (NCH₂), 32.4, 28.8, 28.7 (CH₂), 20.7, 17.2 (CH₃). HRMS (ESI) m/z calcd. for C₁₆H₂₂BrN₂ [M – Br]⁺ 321.0961; found, 321.0968.



This compound was prepared in analogy to **II1** from **I3** (137 mg, 0.34 mmol) and KSCN (35 mg, 0.36 mmol) as a yellowish solid. Yield: 118 mg, 0.31 mmol, 91%. ¹H NMR (300 MHz, CDCl₃): δ 9.96 (s, 1 H, NCHN), 8.16 (s, 1 H, Imi–H), 7.18 (s, 1 H, Imi–H), 6.94 (s, 2 H, Ar–H), 4.74 (t, ³*J*(H,H) = 7 Hz, 2 H, NCH₂), 3.11 (t, ³*J*(H,H) = 7 Hz, 2 H, SCH₂), 2.28 (s, 3 H, CH₃), 2.23–2.20 (m, 2

H, CH₂), 2.00 (s, 6 H, CH₃), 1.95–1.89 (m, 2 H, CH₂). ¹³C {¹H} NMR (75 MHz, CDCl₃): 141.8 (NCHN), 137.9, 134.7, 131.1, 130.4, 124.2, 124.0 (Ar–C), 112.9 (SCN), 49.7 (NCH₂), 33.6, 29.1, 26.8 (CH₂), 21.6, 18.1 (CH₃). HRMS (ESI) m/z calcd. for $C_{17}H_{22}N_3S [M - Br]^+$ 300.1529; found, 300.1540.



Compound **3** was prepared in analogy to **1** from **II3** (148 mg, 0.39 mmol), NaOH aqueous solution (1 M, 39 μ L) and KSCN (189 mg, 1.95 mmol) as an off-white solid. Yield: 124 mg, 0.37 mmol, 96%. ¹H NMR (300 MHz, CDCl₃): δ 8.04 (s, 1 H, Imi–H), 7.17 (s, 1 H, Imi–H), 6.86 (s, 2 H, Ar–H), 4.57 (br-s,

2 H, NCH₂), 2.89 (br-s, 2 H, SCH₂), 2.17 (br-s, 5 H, CH₂ and CH₃), 2.01 (br-s, 2 H, CH₂), 1.83 (s, 6 H, CH₃). ¹³C{¹H} NMR (126 MHz, CDCl₃): 144.5 (NCN), 141.5, 134.6 (Ar–C), 134.3 (SCN⁻), 130.9, 129.8, 127.2, 123.5 (Ar–C), 52.5 (NCH₂), 33.8 (SCH₂), 31.0, 26.1 (CH₂), 21.2, 17.7 (CH₃). HRMS (ESI) m/z calcd. for C₁₆H₂₁N₂S [M – SCN]⁺ 273.1420; found, 273.1425.



1,2-Bis(bromomethyl)benzene (1.85 g, 7.00 mmol) was heated at 100 °C until it became molten and then *N*-mesityl imidazole (559 mg, 3.00 mmol) was added. The reaction mixture was stirred at 100 °C overnight. After cooling it to ambient temperature,

dichloromethane (2 mL) was added. The resulting solution was

added to diethyl ether (150 mL) dropwise and a white solid was obtained which was washed with more diethyl ether (4 × 100 mL). The residue was then washed with acetone (2 × 20 mL). Removal of the solvent from the combined acetone washing afforded the product as a white solid. Yield: 293 mg, 0.65 mmol, 22%. ¹H NMR (300 MHz, d_6 -DMSO): δ 9.59 (s, 1 H, NCHN), 8.04 (s, 1 H, Imi–H), 7.99 (s, 1 H, Imi–H), 7.59–7.33 (m, 4 H, Ar–H), 7.15 (s, 2 H, Ar–H), 5.73 (s, 2 H, NCH₂), 4.95 (s, 2 H, BrCH₂), 2.33 (s, 3 H, CH₃), 2.04 (s, 6 H, CH₃). ¹³C {¹H} NMR (75 MHz, d_6 -DMSO): 140.2 (NCHN), 137.9, 136.6, 134.2, 132.7, 131.4, 131.1, 129.6, 129.5, 129.2, 124.2, 123.5 (Ar–C, two are coincident), 49.3 (NCH₂), 31.7 (BrCH₂), 20.5, 17.0 (CH₃). HRMS (ESI) m/z calcd. for C₂₀H₂₂BrN₂ [M – Br]⁺ 369.0961; found, 369.0962.



This compound was prepared in analogy to **II1** from **I4** (293 mg, 0.65 mmol) and KSCN (316 mg, 3.25 mmol) as a palepink spongy solid. Yield: 233 mg, 0.57 mmol, 88%. ¹H NMR (500 MHz, CDCl₃): δ 9.45 (s, 1 H, NCHN), 7.57 (s, 1 H, Imi–H), 7.54–7.43 (m, 4 H, Ar–H), 7.16 (s, 1 H, Imi–H), 6.99 (s, 2 H, Ar–H), 5.99 (s, 2 H, NCH₂), 4.52 (s, 2 H, SCH₂), 2.32 (s, 3 H, CH₃), 2.08 (s, 6 H, CH₃). ¹³C{¹H} NMR (126 MHz, CDCl₃): 142.2 (NCHN), 138.2, 135.6, 134.9, 133.1, 132.3, 131.6, 131.3, 130.8, 130.6, 124.0, 123.6 (Ar–C, two are coincident), 112.5 (SCN), 51.6 (NCH₂), 36.1 (SCH₂), 21.7, 18.3 (CH₃), SCN⁻ signal could not be detected. HRMS (ESI) m/z calcd. for $C_{21}H_{22}N_3S$ [M – SCN]⁺ 348.1529; found, 348.1525.



Compound 4 was prepared in analogy to 1 from II4 (89 mg, 0.22 mmol), NaOH aqueous solution (1 M, 22 μ L) and KSCN (107 mg, 1.10 mmol) as an off-white solid. Yield: 80 mg, 0.21 mmol, 95%. ¹H NMR (500 MHz, CDCl₃): δ 8.32 (d, ³*J*(H,H) = 2 Hz, 1 H, Imi–H), 7.77–7.75 (m, 1 H, Ar–H), 7.41–7.40 (m, 2 H, Ar–H), 7.35–7.33 (m, 1 H, Ar–H), 7.07 (d, ³*J*(H,H) =

2 Hz, 1 H, Imi–H), 6.97 (s, 2 H, Ar–H), 6.07 (s, 2 H, NCH₂), 4.75 (s, 2 H, SCH₂), 2.32 (s, 3 H, CH₃), 1.92 (s, 6 H, CH₃). ¹³C{¹H} NMR (126 MHz, CDCl₃): 144.1 (NCN), 142.6, 136.8, 135.9, 133.8, 131.5, 131.3, 130.6, 130.4, 129.5, 129.1, 127.5, 122.0 (Ar–C), 53.5 (NCH₂), 32.9 (SCH₂), 21.8, 18.2 (CH₃), SCN⁻ signal could not be detected. HRMS (ESI) m/z calcd. for $C_{20}H_{21}N_2S$ [M – SCN]⁺ 321.1420; found, 321.1424.



This compound was prepared in analogy to **II1** from **I5** (396 mg, 1.00 mmol) and KSCN (486 mg, 5.00 mmol) as a pale-pink solid. Yield: 349 mg, 0.99 mmol, 99%. ¹H NMR (300 MHz, CDCl₃): δ 10.23 (s, 1 H, NCHN), 7.98 (d, ${}^{3}J(H,H) = 8$ Hz, 1 H, Ar–H), 7.64–7.28 (m, 8 H, Ar–H), 5.79 (s, 2 H, NCH₂Ph), 5.12 (br-s, 2 H, NCH₂), 3.78 (br-s, 2 H, SCH₂). ${}^{13}C{}^{1}H}$ NMR (75 MHz, CDCl₃): 143.2 (NCHN), 132.7, 132.3, 131.6, 130.0, 129.9, 129.0, 128.3, 128.1, 114.5, 113.9 (Ar–C), 112.3 (SCN), 52.5 (NCH₂Ph), 47.5 (NCH₂), 34.1 (CH₂), SCN⁻ signal could not be detected. HRMS (ESI) m/z calcd. for C₁₇H₁₆N₃S [M – SCN]⁺ 294.1059; found, 294.1064.



Compound **5** was prepared in analogy to **1** from **II5** (232 mg, 0.66 mmol), NaOH aqueous solution (1 M, 66 μ L) and KSCN (321 mg, 3.30 mmol) as an off-white solid. Analytically pure product was obtained by recrystallization from a concentrated MeOH solution. Yield: 184 mg, 0.56 mmol, 85%. ¹H NMR (500

MHz, *d*₄-Methanol): δ 7.80–7.79 (m, 1 H, Ar–H), 7.73–7.71 (m, 1 H, Ar–H), 7.60–7.54 (m, 2 H, Ar–H), 7.50–7.42 (m, 5 H, Ar–H), 5.58 (s, 2 H, NCH₂Ph), 4.77 (t, ${}^{3}J$ (H,H) = 8 Hz, 2 H, NCH₂), 4.34 (t, ${}^{3}J$ (H,H) = 8 Hz, 2 H, SCH₂). ${}^{13}C$ { ${}^{1}H$ } NMR (126 MHz, *d*₄-Methanol): 161.1 (NCN), 137.8, 133.6 (Ar–C), 133.5 (SCN⁻), 131.8, 130.5, 130.4, 130.1, 127.4, 126.9, 113.8, 113.4 (Ar–C), 51.9 (NCH₂Ph), 48.0 (NCH₂), 39.1 (CH₂). Anal. Calc. for C₁₇H₁₅N₃S₂: C, 62.74; H, 4.65; N, 12.91. Found: C, 62.37; H, 4.59; N, 12.71. HRMS (ESI) m/z calcd. for C₁₆H₁₅N₂S [M – SCN]⁺ 267.0950; found, 267.0947. A mixture of bromopropyl-benzimidazolium bromide (41 mg, 0.10 mmol) and KSCN (49 mg, 0.50 mmol) was suspended in CH₃CN (10 mL) and stirred overnight. An aqueous NaOH solution (1 M, 10 µL) was subsequently added and the suspension was



stirred for another 12 h. Dichloromethane (50 mL) was added and the resulting solution was filtered. Removal of the solvent yielded the product as a yellowish solid. The yellowish solid was further purified by column chromatography using silica gel as a stationary phase and by eluting with a mixture of CH_2Cl_2/CH_3OH

(95:5 v/v) to obtain the pure compound. Yield: 32 mg, 0.10 mmol, 95%. ¹H NMR (500 MHz, CD₃CN): δ 7.74 (d, 1H, ³*J*(H,H) = 8 Hz, Ar–H), 7.67 (d, 1H, ³*J*(H,H) = 8 Hz, Ar–H), 7.55 (m, 2H, Ar–H), 7.39–7.35 (m, 5H, Ar–H), 5.49 (s, 2H, NCH₂Ph), 4.44 (t, 2H, ³*J*(H,H) = 6 Hz, NCH₂), 3.55 (t, 2H, ³*J*(H,H) = 6 Hz, SCH₂), 2.53 (q, 2H, ³*J*(H,H) = 6 Hz, CH₂). ¹³C{¹H} NMR (125 MHz, CD₃CN): 151.6 (NCN), 134.1, 134.0, 132.3, 130.0, 129.7, 128.6, 127.2, 126.6, 112.6, 112.5 (Ar–C), 49.5 (NCH₂Ph), 44.3 (NCH₂), 27.3 (SCH₂), 21.8 (CH₂), SCN⁻ signal could not be detected. HRMS (ESI) m/z calcd. for C₁₇H₁₇N₂S [M – SCN]⁺ 281.1107; found, 281.1107.



Compound **I7** was synthesized in analogy to **I3** from *N*-benzyl benzimidazole (1.04 g, 5.00 mmol) and 1,4-dibromobutane (12 mL) as a yellowish solid. Yield: 1.42 g, 3.35 mmol, 67%. ¹H NMR (300 MHz, CDCl₃): δ 11.38 (s, 1 H, NCHN), 7.76 (d, ³*J*(H,H) = 8 Hz, 1 H, Ar–H), 7.61–7.47 (m, 5 H, Ar–H), 7.33–7.30 (m, 3 H, Ar–H), 5.86 (s, 2 H, NCH₂Ph), 4.72 (br-s, 2

H, NCH₂), 3.47 (t, ³*J*(H,H) = 6 Hz, 2 H, BrCH₂), 2.25 (br-s, 2 H, CH₂), 2.02 (br-s, 2 H, CH₂). ¹³C{¹H} NMR (75 MHz, CDCl₃): 143.4 (NCHN), 133.3, 132.0, 131.8, 130.0,

129.8, 129.0, 127.9, 114.6, 113.8 (Ar–C), 52.2, 47.6 (NCH₂), 33.5 (BrCH₂), 29.8, 28.5 (CH₃). HRMS (ESI) m/z calcd. for C₁₈H₂₀BrN₂ [M – Br]⁺ 343.0804; found, 343.0808.



Compound 7 was prepared in analogy to **6** from **I7** (42 mg, 0.10 mmol), KSCN (49 mg, 0.50 mmol), NaOH aqueous solution (1 M, 10 μ L) as a pale brown solid. Yield: 35 mg, 0.10 mmol, 98%. ¹H NMR (400 MHz, CD₃CN): δ 7.87 (d, ³*J*(H,H) = 8 Hz, 1 H, Ar–H), 7.77 (d, ³*J*(H,H) = 8 Hz, 1 H, Ar–H), 7.68–7.58 (m, 2

H, Ar–H), 7.39–7.32 (m, 5 H, Ar–H), 5.72 (s, 2 H, NCH₂Ph), 4.64 (t, ${}^{3}J$ (H,H) = 6 Hz, 2 H, NCH₂), 3.25 (t, ${}^{3}J$ (H,H) = 6 Hz, 2 H, SCH₂), 2.31–2.27 (m, 2 H, CH₂), 2.07–2.02 (m, 2 H, CH₂). ${}^{13}C$ {¹H} NMR (101 MHz, CD₃CN): 153.9 (NCN), 134.9, 134.2 (Ar–C), 132.9 (SCN⁻), 130.1, 129.7, 129.5, 128.5, 128.0, 127.9, 114.0, 113.7 (Ar–C), 51.2, 48.9 (NCH₂), 34.6, 31.1, 25.7 (CH₂). HRMS (ESI) m/z calcd. for C₁₈H₁₉N₂S [M – SCN]⁺ 295.1263; found, 295.1273.



Compound **8** was prepared in analogy to **6** from **I8** (167 mg, 0.50 mmol), KSCN (243 mg, 2.50 mmol), NaOH aqueous solution (1 M, 50 μL) as a pale-yellow solid. Yield: 128 mg, 0.49 mmol, 97%.

¹H NMR (400 MHz, CD₃CN): δ 7.72–7.68 (m, 2 H, Ar–H), 7.58–7.55 (m, 2 H, Ar–H), 4.40 (t, ³*J*(H,H) = 8 Hz, 2 H, NCH₂), 3.79 (s, 3 H, CH₃), 3.53 (t, ³*J*(H,H) = 8 Hz, 2 H, SCNCH₂), 2.50 (m, ³*J*(H,H) = 7 Hz, 2 H, CH₂). ¹³C{¹H} NMR (75 MHz, CD₃CN): 151.3 (NCN) 133.7, 132.9, 127.0, 126.4, 112.3, 112.2 (Ar–C), 44.1 (NCH₂), 32.0 (CH₃), 27.1 (SCH₂), 21.9 (CH₂), SCN⁻ signal could not be detected. Anal. Calc. for $C_{12}H_{13}N_3S_2$: C, 54.72; H, 4.98; N, 15.95. Found: C, 54.83; H, 4.67; N, 15.63. HRMS (ESI) m/z calcd. for $C_{11}H_{13}N_2S$ [M – SCN]⁺ 205.0794; found, 205.0796.



Compound **9** was prepared in analogy to **6** from **I9** (113 mg, 0.30 mmol), KSCN (146 mg, 1.50 mmol), NaOH aqueous solution (1 M, 30 μ L) as a pale-yellow solid. Yield: 89 mg, 0.29 mmol, 97%. ¹H NMR (400 MHz, CDCl₃): δ 7.65–7.63 (m, 1 H, Ar–H),

7.56–7.54 (m, 1 H, Ar–H), 7.45–7.38 (m, 2 H, Ar–H), 4.55 (t, ${}^{3}J(H,H) = 6$ Hz, 2 H, NCH₂), 3.99 (d, ${}^{3}J(H,H) = 7$ Hz, 2 H, NCH₂), 3.61 (t, ${}^{3}J(H,H) = 6$ Hz, 2 H, SCH₂), 2.57 (m, ${}^{3}J(H,H) = 6$ Hz, 2 H, CH₂), 2.22 (m, ${}^{3}J(H,H) = 7$ Hz, 1 H, CH), 0.91 (d, ${}^{3}J(H,H) = 7$ Hz, 6 H, CH₃). ${}^{13}C\{{}^{1}H\}$ NMR (101 MHz, CDCl₃): 150.0 (NCN), 133.0, 131.9 (Ar–C), 131.2 (SCN⁻), 127.8, 126.9, 126.3, 112.1 (Ar–C), 53.3, 44.0 (NCH₂), 28.8, 27.1, 21.9 (CH and CH₂), 20.5 (CH₃). HRMS (ESI) m/z calcd. for C₁₄H₁₉N₂S [M – SCN]⁺ 247.1263; found, 247.1271.



Compound **I10** was synthesized in analogy to **I3** from *N*methylpropionato benzimidazole (1.02 g, 5.00 mmol) and 1,3-dibromopropane (10 mL) as a yellowish solid. Yield: 1.36 g, 3.35 mmol, 67%. ¹H NMR (400 MHz, CDCl₃): δ 10.79 (s, 1 H, NCHN), 7.83–7.78 (m, 2 H, Ar–H), 7.51–7.49 (m, 2 H, Ar–H), 4.82 (t, ³*J*(H,H) = 7 Hz, 2 H, NCH₂), 4.73 (t, ${}^{3}J(H,H) = 7$ Hz, 2 H, NCH₂), 3.47 (s, 3 H, CH₃), 3.42 (t, ${}^{3}J(H,H) = 7$ Hz, 2 H, BrCH₂), 3.10 (t, ${}^{3}J(H,H) = 7$ Hz, 2 H, COCH₂), 2.54 (m, ${}^{3}J(H,H) = 7$ Hz, 2 H, CH₂). ${}^{13}C{}^{1}H{}$ NMR (101 MHz, CDCl₃): δ 170.9 (CO), 143.3 (NCHN), 131.42, 131.37, 127.62, 127.57, 113.9, 113.5 (Ar–C), 52.6, 46.2, 43.4, 33.7, 32.2 (CH₂), 30.1 (CH₃). Anal. Calcd. for C₁₄H₁₈Br₂N₂O₂: C, 41.41; H, 4.47; N, 6.90. Found: C, 41.96; H, 4.79; N, 7.02. HRMS (ESI) m/z calcd. for C₁₄H₁₈BrN₂O₂ [M – Br]⁺ 325.0546; found, 325.0545.



Compound **10** was prepared in analogy to **6** from **I10** (406 mg, 1.00 mmol), KSCN (486 mg, 5.00 mmol), NaOH aqueous solution (1 M, 100 μ L) as an off-white solid. Yield: 329 mg, 0.98 mmol, 98%. ¹H NMR (500 MHz, CDCl₃): δ 7.74 (d, ³*J*(H,H) = 7 Hz, 1 H, Ar–H), 7.70 (d, ³*J*(H,H) = 7 Hz,

1 H, Ar–H), 7.51–7.46 (m, 2 H, Ar–H), 4.68 (t, ${}^{3}J(H,H) = 7$ Hz, 2 H, NCH₂), 4.60 (t, ${}^{3}J(H,H) = 7$ Hz, 2 H, NCH₂), 3.79 (t, ${}^{3}J(H,H) = 7$ Hz, 2 H, SCH₂), 3.62 (s, 3 H, CH₃), 3.00 (t, ${}^{3}J(H,H) = 7$ Hz, 2 H, COCH₂), 2.64 (m, ${}^{3}J(H,H) = 7$ Hz, 2 H, CH₂). ${}^{13}C\{{}^{1}H\}$ NMR (126 MHz, CDCl₃): δ 171.0 (CO), 150.7 (NCN), 133.3, 131.6, 127.2, 126.7, 112.42, 112.38 (Ar–C), 53.0, 44.4, 42.0, 32.9, 27.7 (CH₂), 22.0 (CH₃), SCN⁻ signal could not be detected. HRMS (ESI) m/z calcd. for C₁₄H₁₇N₂O₂S [M – SCN]⁺ 277.1005; found, 277.1013.



Compound **I11** was synthesized in analogy to **I3** from phthalazine (651 mg, 5.00 mmol) and 1,3-dibromopropane (10 mL) as a pale-yellow solid. Yield: 1.49 g, 4.48 mmol,

90%. ¹H NMR (300 MHz, CDCl₃): δ 11.95 (s, 1 H, NCH), 10.00 (s, 1 H, NCH), 8.82 (d, ³*J*(H,H) = 8 Hz, 1 H, Ar–H), 8.62 (d, ³*J*(H,H) = 8 Hz, 1 H, Ar–H), 8.36 (t, ³*J*(H,H) = 8 Hz, 1 H, Ar–H), 8.25 (t, ³*J*(H,H) = 8 Hz, 1 H, Ar–H), 5.21 (t, ³*J*(H,H) = 7 Hz, 2 H, NCH₂), 3.53 (t, ³*J*(H,H) = 7 Hz, 2 H, BrCH₂), 2.74 (m, ³*J*(H,H) = 7 Hz, 2 H, CH₂). ¹³C {¹H} NMR (75 MHz, CDCl₃): 155.2 (NCH), 152.5 (NCH), 140.1, 136.9, 131.7, 129.0, 128.5, 128.4 (Ar–C), 63.0 (NCH₂), 32.8 (BrCH₂), 29.6 (CH₂). HRMS (ESI) m/z calcd. for C₁₁H₁₂BrN₂ [M – Br]⁺ 251.0178; found, 251.0181.



Compound 11 was prepared in analogy to 6 from I11 (33 mg, 0.10 mmol), KSCN (49 mg, 0.5 mmol), NaOH aqueous solution (1 M, 10 μ L) as a dark-brown solid. Yield: 24 mg, 0.09 mmol, 92%. ¹H NMR (300 MHz, CDCl₃): δ 8.36 (d,

 ${}^{3}J(H,H) = 7$ Hz, 1 H, Ar–H), 8.12 (s, 1 H, NCH), 7.77–7.63 (m, 3 H, Ar–H), 4.29 (t, ${}^{3}J(H,H) = 7$ Hz, 2 H, NCH₂), 2.74 (t, ${}^{3}J(H,H) = 7$ Hz, 2 H, SCH₂), 2.21 (m, ${}^{3}J(H,H) =$ 7 Hz, 2 H, CH₂). ${}^{13}C\{{}^{1}H\}$ NMR (75 MHz, CDCl₃): 159.9 (NCH), 138.4, 133.6, 132.2, 130.1, 128.4, 127.2, 126.6 (Ar–C), 50.3 (NCH₂), 36.5 (SCH₂), 28.7 (CH₂), SCN⁻ signal could not be detected. HRMS (ESI) m/z calcd. for C₁₁H₁₁N₂S [M – SCN]⁺ 203.0637; found, 203.0643.



Compound **I12** was synthesized in analogy to **I3** from 1-benzyl triazole (796 mg, 5.00 mmol) and 1,3-dibromopropane (10 mL) as a yellowish solid. Yield: 1.17 g, 3.25 mmol, 65%. ¹H NMR (400 MHz, CDCl₃): δ 11.60 (s, 1 H, NCHN), 9.09 (s, 1 H, NCHN), 7.54–7.51 (m, 2 H, Ar–H), 7.36–7.34 (m, 3 H, Ar–H),

5.68 (s, 2 H, NCH₂Ph), 4.72 (t, ³*J*(H,H) = 7 Hz, 2 H, NCH₂), 3.47 (t, ³*J*(H,H) = 7 Hz, 2 H, BrCH₂), 2.60 (m, ³*J*(H,H) = 7 Hz, 2 H, CH₂). ¹³C{¹H} NMR (101 MHz, CDCl₃): 144.9 (NCHN), 143.7 (NCHN), 132.4, 130.3, 130.1, 129.9 (Ar–C), 57.1, 48.0 (NCH₂), 32.7 (BrCH₂), 29.8 (CH₂). HRMS (ESI) m/z calcd. for C₁₂H₁₅BrN₃ [M – Br]⁺ 280.0444; found, 280.0446.



Compound **12** was prepared in analogy to **6** from **I12** (36 mg, 0.10 mmol), KSCN (49 mg, 0.5 mmol), NaOH aqueous solution (1 M, 10 μ L) as a dark-brown solid. Yield: 23 mg, 0.08 mmol, 80%. ¹H NMR (300 MHz, *d*₆-DMSO): δ 9.16 (s,

1 H, NCHN), 7.41–7.34 (m, 5 H, Ar–H), 5.43 (s, 2 H, NCH₂Ph), 4.31 (t, ${}^{3}J$ (H,H) = 6 Hz, 2 H, NCH₂), 3.50 (t, ${}^{3}J$ (H,H) = 6 Hz, 2 H, SCH₂), 2.31 (m, ${}^{3}J$ (H,H) = 6 Hz, 2 H, CH₂). ${}^{13}C$ {¹H} NMR (75 MHz, *d*₆-DMSO): 149.1 (NCN), 145.0 (NCHN), 132.8 (SCN⁻), 129.6, 128.9, 128.7, 128.4 (Ar–C), 53.0, 44.9 (NCH₂), 26.6 (SCH₂), 20.5 (CH₂). HRMS (ESI) m/z calcd. for C₁₂H₁₄N₃S [M – SCN]⁺ 232.0903; found, 232.0911.

N-phenyl pyrazole (721 mg, 5.00 mmol) and a CH₃CN (10 mL) solution of NaI (749



mg, 5.00 mmol) was added to 1,3-dibromopropane (10 mL). The mixture was stirred at 90 °C for one day. The excess 1,3dibromopropane was recovered by vacuum distillation, and

CH₂Cl₂ was added to the residue. The resulting suspension was filtered through Celite. The filtrate was dried and washed with ethyl acetate (3 × 10 mL) to afford the product as a red oil. Yield: 688 mg, 1.75 mmol, 35%. ¹H NMR (400 MHz, CDCl₃): δ 8.75 (d, ³*J*(H,H) = 3 Hz, 1 H, NCH), 8.26 (d, ³*J*(H,H) = 3 Hz, 1 H, NCH), 7.80–7.73 (m, 5 H, Ar–H), 7.08 (t, ³*J*(H,H) = 3 Hz, 1 H, CH), 4.63 (t, ³*J*(H,H) = 6 Hz, 2 H, NCH₂), 3.41 (t, ³*J*(H,H) = 6 Hz, 2 H, BrCH₂), 2.38 (m, ³*J*(H,H) = 6 Hz, 2 H, CH₂). ¹³C {¹H} NMR (101 MHz, CDCl₃): δ 139.9, 139.7 (NCH), 134.0, 132.1, 131.8, 128.6 (Ar–C), 110.4 (CH), 50.7 (NCH₂), 32.3 (BrCH₂), 29.5 (CH₂). HRMS (ESI) m/z calcd. for C₁₂H₁₄BrN₂ [M – I]⁺ 265.0335; found, 265.0336.



Compound **13** was prepared in analogy to **6** from **I13** (39 mg, 0.10 mmol), KSCN (49 mg, 0.5 mmol) and NaOH aqueous solution (1 M, 10 μ L) as an orange oil. Yield: 24 mg, 0.09

mmol, 87%. ¹H NMR (400 MHz, CDCl₃): δ 8.10 (d, ³*J*(H,H) = 3 Hz, 1 H, NCH), 7.86–7.83 (m, 2 H, Ar–H), 7.65–7.60 (m, 3 H, Ar–H), 6.70 (d, ³*J*(H,H) = 3 Hz, 1 H, CH), 4.41 (t, ³*J*(H,H) = 6 Hz, 2 H, NCH₂), 3.46 (t, ³*J*(H,H) = 6 Hz, 2 H, SCH₂), 2.55 (m, ³*J*(H,H) = 6 Hz, 2 H, CH₂). ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 146.3 (NCH), 137.0 (NCS), 133.1, 132.4, 131.3, 128.9 (Ar–C), 107.6 (CH), 47.8 (NCH₂), 25.0 (SCH₂), 23.6 (CH₂), SCN⁻ signal could not be detected. HRMS (ESI) m/z calcd. for $C_{12}H_{13}N_2S$ [M – SCN]⁺ 217.0794; found, 217.0791.



Compound **10** (34 mg, 0.10 mmol) was mixed with an aqueous NaOH solution (1 M, 100 μ L) in CH₃CN (5 mL), and the reaction mixture was stirred at 70 °C overnight. After the solvent

was removed in vacuum, the product **14** was washed out by diethyl ether (5 × 10 mL) and dried as a white solid. Yield: 18 mg, 0.10 mmol, 97%. ¹H NMR (300 MHz, CDCl₃): δ 7.59 (d, ³*J*(H,H) = 7 Hz, 1 H, Ar–H), 7.21–7.19 (m, 3 H, Ar–H), 4.17 (t, ³*J*(H,H) = 7 Hz, 2 H, NCH₂), 3.22 (t, ³*J*(H,H) = 7 Hz, 2 H, SCH₂), 2.45 (m, ³*J*(H,H) = 7 Hz, 2 H, CH₂). ¹³C{¹H} NMR (75 MHz, CDCl₃): δ 147.6 (NCN), 141.6, 135.7, 123.5, 122.5, 118.0, 108.7 (Ar–C), 43.2 (NCH₂), 26.4 (SCH₂), 23.7(CH₂). HRMS (ESI) m/z calcd. for C₁₀H₁₀N₂S [M + H]⁺ 191.0637; found, 191.0642. Same as in literature.¹



This compound was synthesized in analogy to **I4** from 1,2bis(bromomethyl)benzene (1.85 g, 7.00 mmol) and *N*methylpropionato imidazole (154 mg, 1.00 mmol) as a yellow solid. The yellow solid was further purified by column chromatography using silica gel as a stationary phase and by eluting with a

CH₂Cl₂/CH₃OH mixture (90:10 v/v) to obtain the pure compound.Yield: 163 mg, 0.39 mmol, 39%. ¹H NMR (400 MHz, CD₃CN): δ 9.64 (br, 1 H, NCHN), 7.72–7.66 (m, 1

H, Imi–H), 7.55–7.53 (m, 1 H, Imi–H), 7.47 (br, 1 H, Ar–H), 7.37 (br, 3 H, Ar–H), 5.71 (s, 2 H, NCH₂), 4.81 (s, 2 H, BrCH₂), 4.48–4.45 (m, 2H, NCH₂), 3.60 (s, 3 H, CH₃), 2.99–2.96 (m, 2H, COCH₂). ¹³C{¹H} NMR (101 MHz, CD₃CN): δ 171.5 (CO), 137.92, 137.87 (Ar–C), 133.2 (NCHN), 132.2, 131.3, 130.7, 130.4, 123.9, 123.2 (Ar–C), 52.6, 50.5, 46.0, 34.8 (CH₂), 32.2 (CH₃). HRMS (ESI) m/z calcd. for C₁₅H₁₈BrN₂O₂ [M – Br]⁺ 337.0546; found, 337.0545.



Compound **I15** (84 mg, 0.20 mmol) and KSCN (97 mg, 1.00 mmol) were suspended in CH₃CN (10 mL) and stirred overnight. An aqueous solution of NaOH (1 M, 220 μ L) was subsequently added, and the resulting mixture was stirred at ambient temperature for 4 h

and then heated at 70 °C overnight. After the solvent was removed in vacuum, the product **15** was washed out with ether (5 × 10 mL) and dried as a white solid. The white solid was purified by recrystallization from CHCl₃. Yield: 32 mg, 0.16 mmol, 80%. Single crystal was obtained by slow evaporation of a saturated solution in CH₂Cl₂/diethyl ether. ¹H NMR (500 MHz, CDCl₃): δ 7.38–7.32 (m, 2 H, Ar–H), 7.30–7.29 (m, 2 H, Imi–H), 6.97–6.96 (m, 2 H, Ar–H), 5.21 (s, 2 H, NCH₂), 4.29 (s, 2 H, SCH₂). ¹³C {¹H} NMR (126 MHz, CDCl₃): δ 139.9 (NCN), 138.1, 134.7, 130.3, 129.8, 129.5, 128.9, 128.5, 122.6 (Ar–C), 50.9 (NCH₂), 32.7 (SCH₂). Anal. Calcd. for C₁₁H₁₀N₂S: C, 65.32; H, 4.98; N, 13.85. Found: C, 65.48; H, 5.06; N, 13.83. HRMS (ESI) m/z calcd. for C₁₁H₁₀N₂S [M + H]⁺ 203.0637; found, 203.0639.



Compound I16 was synthesized in analogy to I3 from 1methylpropionato triazole (776 mg, 5.00 mmol) and 1,3dibromopropane (10 mL) as a vellowish and sticky solid. Yield: 696 mg, 1.95 mmol, 39%. ¹H NMR (300 MHz, CDCl₃): δ 11.31 (s, 1 H, NCHN), 9.11 (s, 1 H, NCHN), 4.85 (t, ${}^{3}J(H,H) = 6$ Hz, 2 H, NCH₂), 4.75 (t, ${}^{3}J$ (H,H) = 6 Hz, 2 H, NCH₂), 3.69 (s, 3 H, CH₃), 3.54 (t, ${}^{3}J$ (H,H) $= 6 \text{ Hz}, 2 \text{ H}, \text{BrCH}_2), 3.13 \text{ (t, } {}^{3}J(\text{H},\text{H}) = 6 \text{ Hz}, 2 \text{ H}, \text{COCH}_2), 2.65 \text{ (m, } {}^{3}J(\text{H},\text{H}) = 6 \text{ Hz}, 3 \text{ Hz}$ 2 H, CH₂). ¹³C {¹H} NMR (75 MHz, CDCl₃): δ 170.9 (CO), 144.9, 144.5 (NCHN), 53.2, 49.1, 48.0, 33.0, 32.8 (CH₂), 30.0 (CH₃). HRMS (ESI) m/z calcd. for C₉H₁₅BrN₃O₂ [M

- Br]⁺ 276.0342; found, 276.0346.



Compound 16 was prepared in analogy to 15 from I16 (71 mg, 0.2 mmol), KSCN (97 mg, 1.00 mmol) and a NaOH aqueous solution (1 M, 220 μ L) as a yellowish oil. Yield: 27 mg, 0.19 mmol, 94%. ¹H NMR

 $(400 \text{ MHz}, \text{CDCl}_3)$: δ 8.06 (s, 1 H, NCHN), 4.10 (t, ${}^{3}J(\text{H},\text{H}) = 6 \text{ Hz}, 2 \text{ H}, \text{NCH}_2), 3.13$ $(t, {}^{3}J(H,H) = 6 Hz, 2 H, SCH_{2}), 2.31 (m, {}^{3}J(H,H) = 6 Hz, 2 H, CH_{2}). {}^{13}C{}^{1}H} NMR$ (101 MHz, CD₃CN): δ 146.4 (NCN), 144.0 (NCHN), 44.0 (NCH₂), 26.4 (SCH₂), 23.9 (CH₂). Anal. Calcd. for C₅H₇N₃S: C, 42.53; H, 5.00; N, 29.76. Found: C, 42.90; H, 5.04; N, 30.00. HRMS (ESI) m/z calcd. for $C_5H_7N_3S$ [M + H]⁺ 142.0433; found, 142.0432.

using chromatography Compound purified column (Rf: 0.05. 17 was



hexane/EA/CH2Cl2: 1/2/1,) and isolated as a white solid. Yield: 17 mg, 0.08 mmol, 31%. ¹H NMR (400 MHz, CDCl₃): δ 7.87 (d, ³J(H,H) = 8 Hz, 1 H, Ar-H), 7.44–7.37 (m, 3 H, Ar-H),

4.54–4.48 (m, 1 H, NCH₂), 4.21–4.14 (m, 1 H, NCH₂), 3.50–3.46 (m, 1 H, SCH₂), 3.27–3.15 (m, 2 H, SCH₂ and CH₂), 2.47–2.40 (m, 1 H, CH₂). ¹³C{¹H} NMR (126 MHz, CDCl₃): δ 148.8 (NCN), 142.7, 135.0, 126.1, 124.8, 122.2, 110.7 (Ar–C), 46.1, 43.8, 14.2 (CH₂). Anal. Calcd. for C₁₀H₁₀N₂OS: C, 58.23; H, 4.89; N, 13.58. Found: C, 58.44; H, 5.08; N, 13.73. HRMS (ESI) m/z calcd. for C₁₀H₁₀N₂OS [M + H]⁺ 207.0587; found, 207.0588.



Compound **18** was purified using column chromatography (Rf: 0.33, hexane/EA/CH2Cl2: 1/2/1,) and isolated as a white solid. Yield: 14 mg, 0.06 mmol, 24%. ¹H NMR (500 MHz, CD₃CN): δ

7.82 (d, ${}^{3}J(H,H) = 8$ Hz, 1H, Ar–H), 7.52 (d, ${}^{3}J(H,H) = 8$ Hz, 1H, Ar–H), 7.48 (t, ${}^{3}J(H,H) = 8$ Hz, 1H, Ar–H), 7.41 (t, ${}^{3}J(H,H) = 8$ Hz, 1H, Ar–H), 4.32 (t, ${}^{3}J(H,H) = 6$ Hz, 2H, NCH₂), 3.67–3.64 (m, 2H, SCH₂), 2.77–2.73 (m, 2H, CH₂). ${}^{13}C\{{}^{1}H\}$ NMR (125 MHz, CD₃CN): δ 149.4 (NCN), 142.1, 135.0, 126.3, 125.0, 121.9, 112.4 (Ar–C), 51.8, 43.9, 22.6 (CH₂). Anal. Calcd. for C₁₀H₁₀N₂O₂S: C, 54.04; H, 4.54; N, 12.60. Found: C, 53.85; H, 4.72; N, 12.46. HRMS (ESI) m/z calcd. for C₁₀H₁₀N₂O₂S [M + H]⁺ 223.0536; found, 223.0532.

Scheme S1. Formation of SCN-propyl-pyrazole.





Compound **I19** was synthesized in analogy to **I3** from *N*methylpropionato pyrazole (771 mg, 5.00 mmol) and 1,3dibromopropane (10 mL) as a colorless oil. Yield: 1.42 g, 4.00 mmol, 80%. ¹H NMR (400 MHz, CDCl₃): δ 8.79 (d, ³*J*(H,H) = 3 Hz, 1 H, NCH), 8.73 (d, ³*J*(H,H) = 3 Hz, 1 H,

NCH), 6.74 (t, ${}^{3}J(H,H) = 3$ Hz, 1 H, CH), 5.02–4.99 (m, 4 H, NCH₂), 3.61 (s, 3 H, CH₃), 3.53 (t, ${}^{3}J(H,H) = 6$ Hz, 2 H, BrCH₂), 3.17 (t, ${}^{3}J(H,H) = 6$ Hz, 2 H, COCH₂), 2.53 (m, ${}^{3}J(H,H) = 6$ Hz, 2 H, CH₂). ${}^{13}C{}^{1}H$ NMR (101 MHz, CDCl₃): δ 170.9 (CO), 138.98, 138.88 (NCH), 108.9 (CH), 53.1, 49.7, 46.7, 34.5, 32.8 (CH₂), 29.7 (CH₃). HRMS (ESI) m/z calcd. for C₁₀H₁₆BrN₂O₂ [M – Br]⁺ 275.0390; found, 275.0389.



This compound was prepared in analogy to **II1** from **I19** (356 mg, 1.00 mmol) and KSCN (486 mg, 5.00 mmol) as a pinkish oil. Yield: 291 mg, 0.93 mmol, 93%. ¹H NMR (400 MHz, CD₃CN): δ 8.28 (d, ³*J*(H,H) = 3 Hz, 1 H, NCH), 8.26 (d, ³*J*(H,H) = 3 Hz, 1 H, NCH), 6.78 (t,

 ${}^{3}J(H,H) = 3$ Hz, 1 H, CH), 4.70 (t, ${}^{3}J(H,H) = 6$ Hz, 2 H, NCH₂), 4.62 (t, ${}^{3}J(H,H) = 6$

Hz, 2 H, NCH₂), 3.64 (s, 3 H, CH₃), 3.15 (t, ${}^{3}J$ (H,H) = 6 Hz, 2 H, SCNCH₂), 3.02 (t, ${}^{3}J$ (H,H) = 6 Hz, 2 H, COCH₂), 2.41 (m, ${}^{3}J$ (H,H) = 6 Hz, 2 H, CH₂). ${}^{13}C$ {¹H} NMR (101 MHz, CD₃CN): δ 171.3 (CO), 139.1, 138.5 (NCH), 113.0 (SCN), 109.0 (CH), 52.8, 49.1, 46.5, 33.8, 30.9 (CH₂), 30.2 (CH₃), SCN⁻ signal could not be detected. HRMS (ESI) m/z calcd. for C₁₁H₁₆N₃O₂S [M – SCN]⁺ 254.0958; found, 254.0965.



Compound **II19** (81 mg, 0.26 mmol) was dissolved in CH_3CN and an aqueous solution of NaOH (1 M, 286 μ L) was added. The suspension was stirred at ambient temperature for four hours and then heated at 80 °C

overnight. After the solvent was removed in vacuum, the product was washed out by ether (5 × 10 mL) as a colorless oil (41 mg, 0.25 mmol, 95%). ¹H NMR (400 MHz, CDCl₃): δ 7.56 (d, ³*J*(H,H) = 3 Hz, 1 H, NCH), 7.45 (d, ³*J*(H,H) = 3 Hz, 1 H, NCH), 6.30 (t, ³*J*(H,H) = 3 Hz, 1 H, CH), 4.37 (t, ³*J*(H,H) = 6 Hz, 2 H, NCH₂), 2.91 (t, ³*J*(H,H) = 6 Hz, 2 H, SCNCH₂), 2.42 (m, ³*J*(H,H) = 6 Hz, 2 H, CH₂). ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 140.0, 130.6 (NCH), 112.4 (SCN), 106.8 (CH), 49.7 (NCH₂), 31.5 (SCH₂), 31.0 (CH₂). Anal. Calcd. for C₇H₉N₃S: C, 50.28; H, 5.42; N, 25.13. Found: C, 49.92; H, 5.31; N, 25.49. HRMS (ESI) m/z calcd. for C₇H₉N₃S [M + H]⁺ 168.0590; found, 168.0596.



Figure S1. ¹H NMR and ¹³C{¹H} NMR spectrum of compound II1 in CDCl₃.

	N	Mass S	Spectru	mS	Smar	tForm	ula Report		
Analysis Info							Acquisition Date	1/22/2	020 10:18:55 AM
Analysis Name	D:\Data\Chen	n\2020 San	nples\20200	1\012	2\CS-II1-	1.d	63		
Method	YCH-50-500.	m	85				Operator	defaul	t user
Sample Name	CS-II1						Instrument / Ser#	micrO	TOF-Q II 10269
Comment	A/P Huynh Ha	an Vinh							
Acquisition Par	rameter								
Source Type	ESI		Ion Polarity		Pos	itive	Set Nebulizer	r	2.0 Bar
Focus	Not active		Set Capillan	1	450	0 V 0	Set Dry Heat	er	200 °C
Scan Begin	50 m/z		Set End Pla	te Offs	et -50	v	Set Dry Gas		6.0 l/min
Scan End	800 m/z		Set Collision	Cell F	RF 200	.0 Vpp	Set Divert Va	lve	Waste
Meas.m/z # I	Formula	m/z	err [ppm]	rdb	e ⁻ Con	f N-Rule			
286 1371 1 (C 16 H 20 N 3 S	286 1372	04	85	even	ok			



Figure S2. HR-MS (ESI) data of II1.



Figure S3. ¹H NMR and ¹³C{¹H} NMR spectrum of compound 1 in CDCl₃.

	Mas	s Spectrum Sn	nartForm	nula Report	
Analysis Info				Acquisition Date 1/	10/2020 2:31:13 PM
Analysis Name	D:\Data\Chem\2020	0 Samples\202001\0109\C	S1.d		
Method	YCH-50-500.m			Operator de	efault user
Sample Name	CS1			Instrument / Ser# m	icrOTOF-Q II 10269
Comment	A/P Huynh Han Vin	h			
Acquisition Par	ameter				
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	800 m/z	Set Collision Cell RF	200.0 Vpp	Set Divert Valve	Waste

Meas. m/z # Formula m/z err [ppm] rdb e⁻ Conf N-Rule 259.1263 1 C 15 H 19 N 2 S 259.1263 -2.1 7.5 even ok



Figure S4. HR-MS (ESI) data of 1.



Figure S5. ¹H NMR and ¹³C{¹H} NMR spectrum of compound II2 in CDCl₃.

Mass Spectrum SmartFormula Report									
Analysis Info							Acquisition Date	1/10/2020	0 2:54:50 PM
Analysis Name	D:\Data\Chen	n\2020 Sar	nples\20200	1\010)\CSII2.d				
Method	YCH-50-500.	m	•				Operator	default us	ser
Sample Name	CSII2						Instrument / Ser#	micrOTO	F-Q II 10269
Comment	A/P Huynh Ha	an Vinh							
Acquisition Pa	rameter								
Source Type	ESI		Ion Polarity		Positi	ve	Set Nebulizer		2.0 Bar
Focus	Not active		Set Capillar	y	4500	v	Set Dry Heate	er	200 °C
Scan Begin	50 m/z		Set End Pla	te Offse	et -500 '	V	Set Dry Gas		6.0 l/min
Scan End	800 m/z		Set Collision	n Cell R	F 200.0	Vpp	Set Divert Val	ve	Waste
Meas.m/z #	Formula	m/z	err [ppm]	rdb	e ⁻ Conf	N-Rule			
272.1224 1	C 15 H 18 N 3 S	272.1216	-2.9	8.5	even	ok			

Intens. x104 1.5 1.5



Figure S6. HR-MS (ESI) data of II2.



Figure S7. ¹H NMR and ¹³C{¹H} NMR spectrum of compound 2 in CD₃OD.

	Ν	Aass S	Spectru	m S	Sm	artF	Formu	ula Repo	rt	
Analysis Info								Acquisition Da	ate 1/10	0/2020 2:43:48 PM
Analysis Name	D:\Data\Chen	n\2020 San	nples\20200	1\010	9\CS	2.d				
Method	YCH-50-500.	m						Operator	defa	ault user
Sample Name	CS2							Instrument / S	er# mic	rOTOF-Q II 10269
Comment	A/P Huynh Ha	an Vinh								
Acquisition Par	rameter									
Source Type	ESI		Ion Polarity			Positiv	/e	Set Neb	ulizer	2.0 Bar
Focus	Not active		Set Capillar	1		4500	/	Set Dry I	leater	200 °C
Scan Begin	50 m/z		Set End Pla	e Offs	et	-500 \	'	Set Dry	Gas	6.0 l/min
Scan End	800 m/z		Set Collision	Cell F	۲F	200.0	Vpp	Set Dive	rt Valve	Waste
Aleas.m/z # I	Formula	m/z	err [ppm]	rdb	e (Conf	N-Rule			
245.1115 1 0	C 14 H 17 N 2 S	245.1107	-3.4	7.5	eve	n	ok			



Figure S8. HR-MS (ESI) data of 2.



Figure S9. ¹H NMR and ${}^{13}C{}^{1}H$ NMR spectrum of compound I3 in CDCl₃.

Analysis Info				Acquisition Date	1/22/2020 10:24:28 AM
Analysis Name	D:\Data\Chem\2020 Sa	mples\202001\0122\C	S-13.d		
Method	YCH-50-500.m			Operator	default user
Sample Name	CS-13			Instrument / Ser#	micrOTOF-Q II 10269
Comment	A/P Huynh Han Vinh				
Acquisition Par	ameter				
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heate	er 200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	800 m/z	Set Collision Cell RF	200.0 Vpp	Set Divert Va	ve Source

Mass Spectrum SmartFormula Report

 Meas. m/z
 #
 Formula
 m/z
 err [ppm]
 rdb
 e⁻ Conf
 N-Rule

 321.0968
 1
 C 16 H 22 Br N 2
 321.0961
 -2.3
 6.5
 even
 ok



Figure S10. HR-MS (ESI) data of I3.



Figure S11. ¹H NMR and ¹³C{¹H} NMR spectrum of compound II3 in CDCl₃.

	Ν	Aass S	Spectru	m S	Sm	nartF	Form	ula Report		
Analysis Info								Acquisition Date	1/1	0/2020 2:50:46 PM
Analysis Name	D:\Data\Chen	n\2020 San	nples\20200	1\010	9103	SII3-1.c	1			
Method	YCH-50-500.	m						Operator	def	ault user
Sample Name	CSII3							Instrument / Ser#	mic	rotof-Q II 10269
Comment	A/P Huynh Ha	an Vinh								
Acquisition Par	rameter									
Source Type	ESI		Ion Polarity			Positiv	/e	Set Nebulize	er	2.0 Bar
Focus	Not active		Set Capillar	Y		4500	V	Set Dry Hea	ter	200 °C
Scan Begin	50 m/z		Set End Pla	te Offs	et	-500 \	1	Set Dry Gas		6.0 l/min
Scan End	800 m/z		Set Collision	I Cell F	۲F	200.0	Vpp	Set Divert Va	alve	Source
Meas.m/z # I	Formula	m/z	err [ppm]	rdb	e	Conf	N-Rule			
300.1540 1 0	C 17 H 22 N 3 S	300.1529	-3.8	8.5	eve	en	ok			



Figure S12. HR-MS (ESI) data of II3.



Figure S13. ¹H NMR and ¹³C{¹H} NMR spectrum of compound 3 in CDCl₃.

	Mas	s Spectrum Sn	nartForn	nula Report	
Analysis Info				Acquisition Date	1/22/2020 10:30:19 AM
Analysis Name	D:\Data\Chem\2020) Samples\202001\0122\C	S-3.d		
Method	YCH-50-500.m			Operator	default user
Sample Name	CS-3			Instrument / Ser#	micrOTOF-Q II 10269
Comment	A/P Huynh Han Vin	h			
Acquisition Par	ameter				
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heate	er 200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	800 m/z	Set Collision Cell RF	200.0 Vpp	Set Divert Val	ve Waste

Meas.m/z # Formula m/z err [ppm] rdb e⁻ Conf N-Rule 273.1425 1 C 16 H 21 N 2 S 273.1420 -2.0 7.5 even ok



Figure S14. HR-MS (ESI) data of 3.



Figure S15. ¹H NMR and ¹³C{¹H} NMR spectrum of compound I4 in d_6 -DMSO.
	N	lass S	Spectru	m S	martF	ormu	Ila Report		
Analysis Info							Acquisition Date	1/22/2020 11	12:18 AM
Analysis Name	D:\Data\Chem	12020 Sam	ples\20200	\0122	CS-I4.d		1999 - 19		
Method	YCH-50-500.r	n	84				Operator	default user	
Sample Name	CS-I4						Instrument / Ser#	micrOTOF-Q	II 10269
Comment	A/P Huynh Ha	an Vinh							
Acquisition Pa	rameter								
Source Type	ESI		Ion Polarity		Positive	е	Set Nebulizer	2.0 E	lar
Focus	Not active		Set Capillary		4500 V		Set Dry Heate	er 200 °	C
Scan Begin	50 m/z		Set End Plat	e Offset	-500 V		Set Dry Gas	6.0 1/	min
Scan End	800 m/z		Set Collision	Cell RF	400.0 \	/pp	Set Divert Val	ve Was	e
Meas.m/z #	Formula	m/z	err [ppm]	rdb	e ⁻ Conf	N-Rule	:		
369.0962 1	C 20 H 22 Br N 2	369.0961	-0.3	10.5	even	ok			



Figure S16. HR-MS (ESI) data of I4.



Figure S17. ¹H NMR and ¹³C{¹H} NMR spectrum of compound II4 in CDCl₃.

	Ν	Aass S	Spectru	ım S	martF	ormu	ula Report		
Analysis Info							Acquisition Date	1/22/20	020 11:19:09 AM
Analysis Name	D:\Data\Chen	n\2020 Sar	nples\20200	1\0122\	CS-II4.d				
Method	YCH-50-500.	m					Operator	default	user
Sample Name	CS-II4						Instrument / Ser#	micrOT	OF-Q II 10269
Comment	A/P Huynh Ha	an Vinh							
Acquisition Par	ameter								
Source Type	ESI		Ion Polarity		Positive	e	Set Nebulizer	2	2.0 Bar
Focus	Not active		Set Capillar	y	4500 V		Set Dry Heate	er	200 °C
Scan Begin	50 m/z		Set End Pla	te Offset	-500 V		Set Dry Gas		6.0 l/min
Scan End	800 m/z		Set Collision	1 Cell RF	200.0	/pp	Set Divert Va	lve	Waste
vleas.m/z # l	Formula	m/z	err [ppm]	rdb	e [_] Conf	N-Rule			
348,1525 1 0	C 21 H 22 N 3 S	348.1529	1.0	12.5	even	ok			



Figure S18. HR-MS (ESI) data of II4.



Figure S19. ¹H NMR and ¹³C{¹H} NMR spectrum of compound 4 in CDCl₃.

		r	Aass S	Spectru	Im S	Smar	tForm	nula Report		
Analysis In	fo							Acquisition Date	1/10/2020 3	3:14:31 PM
Analysis Na	me	D:\Data\Cher	n\2020 Sar	nples\20200	1\010	9\CS4.d		85		
Method		YCH-50-500.	m	85				Operator	default user	
Sample Na	me	CS4						Instrument / Ser#	micrOTOF-0	Q II 10269
Comment		A/P Huynh Ha	an Vinh							
Acquisition	۱P	arameter								
Source Type		ESI		Ion Polarity		Pos	sitive	Set Nebulizer	2.0) Bar
Focus		Not active		Set Capillar	y	450	0 V 0	Set Dry Heat	er 200	0°C
Scan Begin		50 m/z		Set End Pla	te Offse	et -50	0 V 0	Set Dry Gas	6.0) l/min
Scan End		800 m/z		Set Collision	n Cell R	F 200	.0 Vpp	Set Divert Va	lve Wa	aste
vleas. m/z	#	Formula	m/z	err [ppm]	rdb	e ⁻ Co	nf N-Ru	le		
321.1424	1	C 20 H 21 N 2 S	321.1420	-1.4	11.5	even	a <u>altais</u> a	ok		







Figure S21. ¹H NMR and ¹³C{¹H} NMR spectrum of compound II5 in CDCl₃.

	Ν	/lass S	Spectru	m S	mart	Form	ula Report		
Analysis Info							Acquisition Date	1/10/2	020 3:04:59 PM
Analysis Name	D:\Data\Chen	12020 San	nples\20200	1\0109	CSII5.d		<i></i>		
Method	YCH-50-500.r	n					Operator	default	tuser
Sample Name	CSII5						Instrument / Ser#	micrO	TOF-Q II 10269
Comment	A/P Huynh Ha	an Vinh							
Acquisition Par	rameter								
Source Type	ESI		Ion Polarity		Posi	tive	Set Nebulizer		2.0 Bar
Focus	Not active		Set Capillan	/	4500	v	Set Dry Heat	er	200 °C
Scan Begin	50 m/z		Set End Pla	te Offse	t -500	v	Set Dry Gas		6.0 l/min
Scan End	800 m/z		Set Collision	Cell R	F 200.	0 Vpp	Set Divert Va	lve	Waste
Meas.m/z # I	Formula	m/z	err [ppm]	rdb	e ⁻ Con	N-Rule	É		
294.1064 1 0	C 17 H 16 N 3 S	294.1059	-1.6	11.5	even	ok			



Figure S22. HR-MS (ESI) data of II5.



Figure S23. ¹H NMR and ¹³C $\{^{1}H\}$ NMR spectrum of compound 5 in CD₃OD.

	N	lass S	Spectru	m S	marth	orm	ula Report		
Analysis Info							Acquisition Date	1/10/20	20 3:19:52 PM
Analysis Name	D:\Data\Chen	n\2020 San	nples\20200	1\0109	CS5.d		•		
Method	YCH-50-500.	n	•				Operator	default	user
Sample Name	CS5						Instrument / Ser#	micrOT	OF-Q II 10269
Comment	A/P Huynh Ha	an Vinh							
Acquisition Pa	rameter								
Source Type	ESI		Ion Polarity		Positiv	e	Set Nebulizer		2.0 Bar
Focus	Not active		Set Capillary	1	4500 \	/	Set Dry Heate	er	200 °C
Scan Begin	50 m/z		Set End Pla	e Offset	-500 V		Set Dry Gas		6.0 l/min
Scan End	800 m/z		Set Collision	Cell RF	200.0	Vpp	Set Divert Va	lve	Waste
Meas.m/z #	Formula	m/z	err [ppm]	rdb	e ^C Onf	N-Rule			
267.0947 1	C 16 H 15 N 2 S	267.0950	1.3	10.5	even	ok			





1H AMX500 chans07Feb20-CS-6-1H



Figure S25. ¹H NMR and ${}^{13}C{}^{1}H$ NMR spectrum of compound 6 in CD₃CN.

	N	Mass S	Spectru	m S	martF	ormu	ula Report		
Analysis Info							Acquisition Date	1/10/2020 3:22	:51 PM
Analysis Name	D:\Data\Chen	n\2020 Sar	nples\20200	1\0109	CS6.d				
Method	YCH-50-500.	m					Operator	default user	
Sample Name	CS6						Instrument / Ser#	micrOTOF-Q II	10269
Comment	A/P Huynh Ha	an Vinh							
Acquisition Pa	rameter								
Source Type	ESI		Ion Polarity		Positiv	е	Set Nebulizer	2.0 Ba	r
Focus	Not active		Set Capillar	/	4500 V		Set Dry Heate	er 200 °C	:
Scan Begin	50 m/z		Set End Pla	e Offset	-500 V		Set Dry Gas	6.0 l/m	in
Scan End	800 m/z		Set Collision	Cell RF	200.0	Vpp	Set Divert Val	ve Waste	
Aleas.m/z #	Formula	m/z	err [ppm]	rdb	e ⁻ Conf	N-Rule			
281.1107 1	C 17 H 17 N 2 S	281.1107	-0.1	10.5	even	ok			



Figure S26. HR-MS (ESI) data of 6.



Figure S27. ¹H NMR and ¹³C $\{^{1}H\}$ NMR spectrum of compound I7 in CDCl₃.

	Mas	s Spectrum Sn	nartForn	nula Report	
Analysis Info				Acquisition Date	1/22/2020 12:07:28 PM
Analysis Name	D:\Data\Chem\2020	0 Samples\202001\0122\C	S-17-1.d		
Method	YCH-50-500.m			Operator	default user
Sample Name	CS-17			Instrument / Ser#	micrOTOF-Q II 10269
Comment	A/P Huynh Han Vir	h			
Acquisition Par	ameter				
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	800 m/z	Set Collision Cell RF	200.0 Vpp	Set Divert Valv	e Waste

Meas. m/z # Formula m/z err [ppm] rdb e⁻Conf N-Rule 343.0808 1 C 18 H 20 Br N 2 343.0804 -1.1 9.5 even ok



Figure S28. HR-MS (ESI) data of I7.



Figure S29. ¹H NMR and ¹³C $\{^{1}H\}$ NMR spectrum of compound 7 in CD₃CN.

	Ν	/ass S	Spectru	m S	SmartF	orm	ula Report		
Analysis Info							Acquisition Date	2/6/20)20 2:39:51 PM
Analysis Name	D:\Data\Chen	n\2020 Sar	nples\20200	2\0206	CS-7.d		50646 • LO 140363560000 0006000		
Method	YCH-50-500.	n	•				Operator	defaul	lt user
Sample Name	CS-7						Instrument / Ser#	micrO	TOF-Q II 10269
Comment	A/P Huynh Ha	an Vinh							
Acquisition Pa	rameter								
Source Type	ESI		Ion Polarity		Positiv	е	Set Nebulizer	5-1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1	2.0 Bar
Focus	Not active		Set Capillar	/	4500 \	1	Set Dry Heate	er	200 °C
Scan Begin	50 m/z		Set End Pla	te Offse	t -500 V		Set Dry Gas		6.0 l/min
Scan End	800 m/z		Set Collision	Cell R	F 100.0	Vpp	Set Divert Va	lve	Waste
Meas.m/z #	Formula	m/z	err [ppm]	rdb	e ⁻ Conf	N-Rule			
295.1273 1	C 18 H 19 N 2 S	295.1263	-3.3	10.5	even	ok			



Figure S30. HR-MS (ESI) data of 7.



Figure S31. ¹H NMR and ¹³C $\{^{1}H\}$ NMR spectrum of compound 8 in CD₃CN.

9 <u>7</u>		nuoo c	peoure		Jinaru	Unit	ala report		
Analysis Info							Acquisition Date	2/6/202	20 2:46:48 PM
Analysis Name Method Sample Name Comment	D:\Data\Chen YCH-50-500.r CS-8 A/P Huynh Ha	n\2020 Sar m an Vinh	nples\20200	2\020	6\CS-8.d		Operator Instrument / Ser#	default micrOT	user OF-QII 10269
Acquisition Pa	rameter								
Source Type Focus Scan Begin Scan End	ESI Not active 50 m/z 800 m/z		lon Polarity Set Capillar Set End Pla Set Collision	y te Offs n Cell F	Positi 4500 et -500 V RF 100.0	ve V V Vpp	Set Nebulizer Set Dry Heate Set Dry Gas Set Divert Va	er Ive	2.0 Bar 200 °C 6.0 I/min Waste
Meas. m/z # 205.0796 1	Formula C 11 H 13 N 2 S	m/z 205.0794	err [ppm] -1.1	rdb 6.5	e [—] Conf even	N-Rule ok			

Mass Spectrum SmartFormula Report



Figure S32. HR-MS (ESI) data of 8.



Figure S33. ¹H NMR and ¹³C{¹H} NMR spectrum of compound 9 in CDCl₃.

Analysis Info				Acquisition Date	1/10/2020 3:28:56 PM
Analysis Name	D:\Data\Chem\2020	Samples\202001\0109\C	S9-1.d		
Method	YCH-50-500.m			Operator	default user
Sample Name	CS9			Instrument / Ser#	micrOTOF-Q II 10269
Comment	A/P Huynh Han Vinh				
Acquisition Par	ameter				
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heate	er 200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	800 m/z	Set Collision Cell RF	200.0 Vpp	Set Divert Val	ve Waste

Mass Spectrum SmartFormula Report

Meas. m/z # Formula m/z err [ppm] rdb e Conf N-Rule 247.1271 1 C 14 H 19 N 2 S 247.1263 -2.9 6.5 even ok



Figure S34. HR-MS (ESI) data of 9.



Figure S35. ¹H NMR and ¹³C{¹H} NMR spectrum of compound I10 in CDCl₃.

	Ma	ss Speo	ctrum	Sm	nartFor	mula	Report		
Analysis Info						Acq	uisition Date	1/22/2020 11:39:	31 AM
Analysis Name	D:\Data\Chem\20	20 Samples\2	202001\01	22\CS	S-I10.d	41/01/2017			
Method	YCH-50-500.m	_				Ope	erator	default user	
Sample Name	CS-I10					Inst	rument / Ser#	micrOTOF-Q II 1	0269
Comment	A/P Huynh Han V	inh							
Acquisition Par	ameter								
Source Type	ESI	Ion Po	olarity		Positive		Set Nebulizer	2.0 Bar	
Focus	Not active	Set C	apillary		4500 V		Set Dry Heate	r 200 °C	
Scan Begin	50 m/z	Set E	nd Plate Of	fset	-500 V		Set Dry Gas	6.0 l/min	
Scan End	800 m/z	Set C	ollision Cell	RF	200.0 Vpp		Set Divert Val	ve Waste	
Meas.m/z # F	Formula	m/z	err [ppm]	rdb	e ⁻ Conf	N-Rule			
325.0545 1 0	C 14 H 18 Br N 2 O 2	325.0546	0.5	6.5	even	ok			



Figure S36. HR-MS (ESI) data of I10.



Figure S37. ¹H NMR and ¹³C{¹H} NMR spectrum of compound 10 in CDCl₃.

Analysis Info				Acquisition Date	1/10/2020 3:34:49 PM
Analysis Name	D:\Data\Chem\2020) Samples\202001\0109\C	S10.d		
Method	YCH-50-500.m			Operator	default user
Sample Name	CS10			Instrument / Ser#	micrOTOF-Q II 10269
Comment	A/P Huynh Han Vin	h			
Acquisition Par	ameter				
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heate	er 200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	800 m/z	Set Collision Cell RF	100.0 Vpp	Set Divert Val	ve Waste

-

Meas. m/z	#	Formula	m/z	err [ppm]	rdb	e ⁻ Conf	N-Rule
277.1013	1	C 14 H 17 N 2 O 2 S	277.1005	-2.8	7.5	even	ok

. .



Figure S38. HR-MS (ESI) data of 10.



Figure S39. ¹H NMR and ${}^{13}C{}^{1}H$ NMR spectrum of compound I11 in CDCl₃.

	Mas	s Spectrum Sn	nartForm	nula Report	
Analysis Info				Acquisition Date 1/	22/2020 10:37:53 AM
Analysis Name	D:\Data\Chem\2020	Samples\202001\0122\C	S-I11.d		
Method	YCH-50-500.m			Operator de	efault user
Sample Name	CS-I11			Instrument / Ser# m	icrOTOF-Q II 10269
Comment	A/P Huynh Han Vin	h			
Acquisition Par	ameter				
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	800 m/z	Set Collision Cell RF	200.0 Vpp	Set Divert Valve	Waste

 Meas. m/z
 #
 Formula
 m/z
 err [ppm]
 rdb
 e⁻ Conf
 N-Rule

 251.0181
 1
 C 11 H 12 Br N 2
 251.0178
 -1.0
 6.5
 even
 ok



Figure S40. HR-MS (ESI) data of I11.



Figure S41. ¹H NMR and ¹³C $\{^{1}H\}$ NMR spectrum of compound 11 in CDCl₃.

	<u> </u>	Mass S	spectru	m	Smar	Form	ula Report		
Analysis Info							Acquisition Date	1/22/2020 1	0:45:00 AM
Analysis Name	D:\Data\Chen	n\2020 San	nples\20200	1\012	2\CS-11.	b			
Method	YCH-50-500.	m					Operator	default user	
Sample Name	CS-11						Instrument / Ser#	micrOTOF-0	QII 10269
Comment	A/P Huynh Ha	an Vinh							
Acquisition Pa	rameter								
Source Type	ESI		Ion Polarity		Pos	tive	Set Nebulizer	2.0	Bar
Focus	Not active		Set Capillar	4	450	v	Set Dry Heate	er 200	O°C
Scan Begin	50 m/z		Set End Pla	te Offs	et -500	V	Set Dry Gas	6.0	l/min
Scan End	800 m/z		Set Collision	n Cell F	RF 200	0 Vpp	Set Divert Val	ve Wa	iste
Meas.m/z #	Formula	m/z	err [ppm]	rdb	e ⁻ Cont	N-Rule			
203.0643 1	C11H11N2S	203.0637	-2.5	7.5	even	ok			



Figure S42. HR-MS (ESI) data of 11.



Figure S43. ¹H NMR and ¹³C{¹H} NMR spectrum of compound I12 in CDCl₃.

	Mas	s Spectrum Sn	nartForm	nula Report	
Analysis Info				Acquisition Date 1	/22/2020 11:44:43 AM
Analysis Name	D:\Data\Chem\2020	0 Samples\202001\0122\C	S-I12.d		
Method	YCH-50-500.m	annen han en		Operator o	lefault user
Sample Name	CS-I12			Instrument / Ser# r	nicrOTOF-Q II 10269
Comment	A/P Huynh Han Vin	h			
Acquisition Par	ameter				
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	800 m/z	Set Collision Cell RF	200.0 Vpp	Set Divert Valve	e Waste

 Meas.m/z
 #
 Formula
 m/z
 err [ppm]
 rdb
 e⁻ Conf
 N-Rule

 280.0446
 1
 C 12 H 15 Br N 3
 280.0444
 -0.9
 6.5
 even
 ok



Figure S44. HR-MS (ESI) data of I12.



Figure S45. ¹H NMR and ¹³C{¹H} NMR spectrum of compound 12 in d_6 -DMSO.

	Mas	s Spectrum Sn	nartForn	nula Report	
Analysis Info				Acquisition Date 1/10	/2020 3:38:05 PM
Analysis Name	D:\Data\Chem\2020) Samples\202001\0109\C	S12.d		
Method	YCH-50-500.m			Operator defa	ult user
Sample Name	CS12			Instrument / Ser# micr	OTOF-Q II 10269
Comment	A/P Huynh Han Vin	h			
Acquisition Par	ameter				
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	800 m/z	Set Collision Cell RF	100.0 Vpp	Set Divert Valve	Waste

Meas. m/z # Formula m/z err [ppm] rdb e⁻ Conf N-Rule 232.0911 1 C 12 H 14 N 3 S 232.0903 -3.5 7.5 even ok



Figure S46. HR-MS (ESI) data of 12.



Figure S47. ¹H NMR and ${}^{13}C{}^{1}H$ NMR spectrum of compound I13 in CDCl₃.

	Mas	s Spectrum Sn	nartForm	nula Report	
Analysis Info				Acquisition Date	1/22/2020 10:54:55 AM
Analysis Name	D:\Data\Chem\2020	Samples\202001\0122\C	S-I13.d		
Method	YCH-50-500.m			Operator	default user
Sample Name	CS-I13			Instrument / Ser#	micrOTOF-Q II 10269
Comment	A/P Huynh Han Vin	h			
Acquisition Par	ameter				
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heate	r 200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	800 m/z	Set Collision Cell RF	200.0 Vpp	Set Divert Valv	ve Waste

Meas.m/z # Formula m/z err [ppm] rdb e⁻Conf N-Rule 265.0336 1 C 12 H 14 Br N 2 265.0335 -0.5 6.5 even ok



Figure S48. HR-MS (ESI) data of I13.



Figure S49. ¹H NMR and ¹³C $\{^{1}H\}$ NMR spectrum of compound 13 in CDCl₃.

Analysis Info				Acquisition Date	1/22/2020 10:59:48 AM
Analysis Name	D:\Data\Chem\2020	Samples\202001\0122\C	S-13.d		
Method	YCH-50-500.m			Operator	default user
Sample Name	CS-13			Instrument / Ser#	micrOTOF-Q II 10269
Comment	A/P Huynh Han Vin	h			
Acquisition Par	ameter		14-14 - 11-14		
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heate	er 200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	800 m/z	Set Collision Cell RF	200.0 Vpp	Set Divert Va	ve Waste

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Figure S50. HR-MS (ESI) data of 13.



Figure S51. ¹H NMR and ¹³C $\{^{1}H\}$ NMR spectrum of compound 14 in CDCl₃.
	N	lass S	Spectru	m	Smar	Form	ula Report		
Analysis Info							Acquisition Date	1/10/2020) 3:43:32 PM
Analysis Name	D:\Data\Chen	1\2020 San	nples\20200	1\010	9\CS14.c	ľ			
Method	YCH-50-500.r	n	•				Operator	default us	ser
Sample Name	CS14						Instrument / Ser#	micrOTO	F-Q II 10269
Comment	A/P Huynh Ha	an Vinh							
Acquisition Pa	rameter								
Source Type	ESI		Ion Polarity		Pos	tive	Set Nebulizer		2.0 Bar
Focus	Not active		Set Capillary	1	450	v	Set Dry Heate	er	200 °C
Scan Begin	50 m/z		Set End Plat	e Offs	et -500	v	Set Dry Gas		6.0 l/min
Scan End	800 m/z		Set Collision	Cell F	RF 60.0	Vpp	Set Divert Va	lve	Waste
vleas.m/z #	Formula	m/z	err [ppm]	rdb	e ⁻ Cont	N-Rule			
191.0642 1	C 10 H 11 N 2 S	191.0637	-2.2	6.5	even	ok			



Figure S52. HR-MS (ESI) data of 14.



Figure S53. ¹H NMR and ¹³C $\{^{1}H\}$ NMR spectrum of compound I15 in CD₃CN.

	Ma	ss Spe	ectrum	Sm	nartFor	mula	Report		
Analysis Info						Acq	uisition Date	1/22/20	20 12:00:10 PM
Analysis Name	D:\Data\Chem\20	20 Samples	202001\01	22\C	S-I15.d				
Method	YCH-50-500.m					Ope	erator	default	user
Sample Name	CS-I15					Inst	rument / Ser#	micrOT	OF-Q II 10269
Comment	A/P Huynh Han V	inh							
Acquisition Pa	rameter								
Source Type	ESI	lon	Polarity		Positive		Set Nebulizer		2.0 Bar
Focus	Not active	Set	Capillary		4500 V		Set Dry Heate	er	200 °C
Scan Begin	50 m/z	Set	End Plate Of	fset	-500 V		Set Dry Gas		6.0 l/min
Scan End	800 m/z	Set	Collision Cell	RF	200.0 Vpp		Set Divert Va	lve	Waste
Meas.m/z #	Formula	m/z	err [ppm]	rdb	e ⁻ Conf	N-Rule			
337 0545 1	C 15 H 18 Br N 2 O 2	337.0546	0.3	7.5	even	ok			



Figure S54. HR-MS (ESI) data of I15.

1H AMX500 chans04Feb20-CS-15-1H



Figure S55. ¹H NMR and ¹³C{¹H} NMR spectrum of compound 15 in CDCl₃.

Analysis Info				Acquisition Date	1/10/2020 3:47:43 PM
Analysis Name	D:\Data\Chem\2020) Samples\202001\0109\C	S15.d		
Method	YCH-50-500.m	•		Operator	default user
Sample Name	CS15			Instrument / Ser#	micrOTOF-Q II 10269
Comment	A/P Huynh Han Vin	h			
Acquisition Par	ameter				
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	800 m/z	Set Collision Cell RF	60.0 Vpp	Set Divert Valv	e Waste

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Figure S56. HR-MS (ESI) data of 15.



Figure S57. ¹H NMR and ¹³C{¹H} NMR spectrum of compound I16 in CDCl₃.

	Mas	s Spectrum Sn	nartForn	nula Report		
Analysis Info				Acquisition Date	1/22/2020 11:48:20 AM	
Analysis Name	e D:\Data\Chem\2020 Samples\202001\0122\CS-I16.d					
Method	YCH-50-500.m	92		Operator	default user	
Sample Name	CS-I16			Instrument / Ser#	micrOTOF-Q II 10269	
Comment	A/P Huynh Han Vin	h				
Acquisition Par	ameter					
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar	
Focus	Not active	Set Capillary	4500 V	Set Dry Heate	r 200 °C	
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min	
Casa Lad	800 m/z	Set Collision Cell RF	200.0 Vpp	Set Divert Val	ve Waste	

Meas. m/z # Formula m/z err [ppm] rdb e⁻ Conf N-Rule 276.0346 1 C 9 H 15 Br N 3 O 2 276.0342 -1.3 3.5 even ok



Figure S58. HR-MS (ESI) data of I16.



Figure S59. ¹H NMR and ¹³C{¹H} NMR spectrum of compound 16 in CDCl₃.

Analysis Info				Acquisition Date	1/10/2020 3:51:10 PM	
Analysis Name	D:\Data\Chem\202	0 Samples\202001\0109\C	S16.d			
Method	YCH-50-500.m			Operator	default user	
Sample Name	CS16			Instrument / Ser# micrOTOF-Q II 102		
Comment	A/P Huynh Han Vinh					
Acquisition Par	ameter					
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar	
Focus	Not active	Set Capillary	4500 V	Set Dry Heate	er 200 °C	
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min	
Scan End	800 m/z	Set Collision Cell RF	60.0 Vpp	Set Divert Val	ve Waste	

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Figure S60. HR-MS (ESI) data of 16.



Figure S61. ¹H NMR and ¹³C $\{^{1}H\}$ NMR spectrum of compound 17 in CDCl₃.

	M	ass Sp	pectrur	n Si	martFo	ormu	Ila Report		
Analysis Info							Acquisition Date	1/10/2020 3:56:1	4 PM
Analysis Name	D:\Data\Chem\	2020 Samp	les\202001\	0109\	CS17.d				
Method	YCH-50-500.m	3 1 3					Operator	default user	
Sample Name	CS17						Instrument / Ser#	micrOTOF-Q II 1	0269
Comment	A/P Huynh Han Vinh								
Acquisition Pa	arameter								
Source Type	ESI	lo	on Polarity		Positive		Set Nebulizer	2.0 Bar	
Focus	Not active	S	et Capillary		4500 V		Set Dry Heate	er 200 °C	
Scan Begin	50 m/z	S	et End Plate	Offset	-500 V		Set Dry Gas	6.0 l/min	
Scan End	800 m/z	S	et Collision (cell RF	60.0 Vpp		Set Divert Val	ve Waste	
Meas.m/z #	Formula	m/z	err [ppm]	rdb	e ⁻ Conf	N-Rule	9		
207.0588 1	C10H11N2OS	207.0587	-0.8	6.5	even	o	k		



Figure S62. HR-MS (ESI) data of 17.

1H AMX500 chans04Feb20-CS-18-1H



220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 ppm Figure S63. ¹H NMR and ¹³C {¹H} NMR spectrum of compound **18** in CD₃CN.

	Mass Spectrum SmartFormula Report								
Analysis Info	nalysis Info				Acc	Acquisition Date 1/10		10/2020 4:00:20 PM	
Analysis Name Method Sample Name	D:\Data\Chem\2020 Samples\202001\0109\CS18.d YCH-50-500.m CS18				Ope	Operator default user		er 	
Comment	A/P Huynh Han Vinh						admontr odni		4 10200
Acquisition Pa	rameter								
Source Type Focus Scan Begin Scan End	ESI Not active 50 m/z 800 m/z	lon Sei Sei	Polarity t Capillary t End Plate O t Collision Ce	ffset II RF	Positive 4500 V -500 V 60.0 Vpp		Set Nebulizer Set Dry Heate Set Dry Gas Set Divert Val	er 22 Ve V	200 °C 200 °C 3.0 l/min Waste
Meas.m/z # 223.0532 1	Formula C 10 H 11 N 2 O 2 S	m/z 223.0536	err [ppm] 1.8	rdb 6.5	e Conf even	N-Rule ok			



Figure S64. HR-MS (ESI) data of 18.



Figure S65. ¹H NMR and ${}^{13}C{}^{1}H$ NMR spectrum of compound I19 in CDCl₃.

Analysis Info				Acquisition Date	1/22/2020 11:52:21 AM
Analysis Name	D:\Data\Chem\2020 Si	amples\202001\0122\C	S-I19.d		
Method	YCH-50-500.m			Operator	default user
Sample Name	CS-I19			Instrument / Ser#	micrOTOF-Q II 10269
Comment	A/P Huynh Han Vinh				
Acquisition Par	ameter				
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heate	er 200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	800 m/z	Set Collision Cell RF	200.0 Vpp	Set Divert Va	ve Waste

Meas. m/z	#	Formula	m/z	err [ppm]	rdb	e Conf	N-Rule
275.0389	1	C 10 H 16 Br N 2 O 2	275.0390	0.2	3.5	even	ok



Figure S66. HR-MS (ESI) data of I19.



Figure S67. ¹H NMR and ¹³C{¹H} NMR spectrum of compound II19 in CD₃CN.

	Mas	s Spectrum Sn	nartForm	nula Report	
Analysis Info				Acquisition Date 1	1/10/2020 3:10:42 PM
Analysis Name	D:\Data\Chem\2020 Samples\202001\0109\CSII19.d			1640-310 • CONTENTS (1754-1710-174-363)	
Method	YCH-50-500.m			Operator o	lefault user
Sample Name	CSII19	SII19			nicrOTOF-Q II 10269
Comment	A/P Huynh Han Vin	h			
Acquisition Par	ameter				
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	800 m/z	Set Collision Cell RF	200.0 Vpp	Set Divert Valv	e Waste

 Meas. m/z
 # Formula
 m/z
 err [ppm]
 rdb
 e⁻ Conf
 N-Rule

 254.0965
 1
 C 11 H 16 N 3 O 2 S
 254.0958
 -2.8
 5.5
 even
 ok



Figure S68. HR-MS (ESI) data of II19.



Figure S69. ¹H NMR and ¹³C $\{^{1}H\}$ NMR spectrum of compound 19 in CDCl₃.

	Mas	s Spectrum Sn	nartForr	nula Report	
Analysis Info				Acquisition Date 2/6	0/2020 3:02:37 PM
Analysis Name	D:\Data\Chem\2020 Samples\202002\0206\CS-19-1.d				
Method	YCH-50-500.m			Operator de:	fault user
Sample Name	CS-19			Instrument / Ser# mid	crOTOF-QII 10269
Comment	A/P Huynh Han Vin	h			
Acquisition Par	ameter				
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	2.0 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	6.0 l/min
Scan End	500 m/z	Set Collision Cell RF	50.0 Vpp	Set Divert Valve	Waste

 Meas. m/z
 #
 Formula
 m/z
 err [ppm]
 rdb
 e⁻Conf
 N-Rule

 168.0596
 1
 C 7 H 10 N 3 S
 168.0590
 -3.7
 4.5
 even
 ok



Figure S70. HR-MS (ESI) data of 19.

X-ray Diffraction Studies. Single crystals of 2, 4, 5, 12, 15 were obtained by slow evaporation of a concentrated solution in CH₂Cl₂/diethyl ether or CH₂Cl₂/hexane. Xray data for them were collected with a Bruker AXS SMART APEX diffractometer, using Mo- or Cu-K_{α} radiation with the SMART suite of Programs.² Data were processed and corrected for Lorentz and polarization effects with SAINT,³ and for absorption effect with SADABS.⁴ Structural solution and refinement were carried out with the SHELXTL suite of programs.⁵ The structure was solved by direct methods to locate the heavy atoms, followed by difference maps for the light, non-hydrogen atoms. All non-hydrogen atoms were generally given anisotropic displacement parameters in the final model. All H-atoms were put at calculated positions. A summary of the most important crystallographic given Table S1. data is in

	2	4	5
formula	$C_{15}H_{17}N_3S_2$	$C_{21}H_{21}N_3S_2$	$C_{17}H_{15}N_3S_2$
fw	303.44	379.53	325.44
color, habit	yellow, block	colorless, block	colorless, block
cryst size [mm]	0.60 x 0.30 x 0.15	0.16 x 0.14 x 0.12	0.46 x0.26 x0.20
temp [K]	100(2)	223(2)	100(2)
crystsyst	orthorhombic	monoclinic	monoclinic
space group	Pbca	<i>P</i> 21/c	P21/n
<i>a</i> [Å]	10.5462(6)	8.1553(5)	5.3591(3)
<i>b</i> [Å]	11.8138(6)	16.6915(10)	19.3576(11)
<i>c</i> [Å]	24.1532(13)	14.5815(9)	15.0546(8)
α [deg]	90.00	90.00	90.00
β [deg]	90.00	102.2050(10)	96.0410(10)
γ [deg]	90.00	90.00	90.00
V[Å ³]	3009.3(3)	1940.0(2)	1553.08(15)
Ζ	8	4	4
$D_{\rm c} [{\rm g}{\rm cm}^{-3}]$	1.340	1.299	1.392
radiation used	Μο Κα	Μο Κα	Μο Κα
μ [mm ⁻¹]	0.347	0.284	0.342
θ range [deg]	1.69-27.50	1.88-27.50	2.10-27.48
no. of unique data	20165	13597	10876
max., min. transmn	0.9498, 0.8188	0.9667, 0.9560	0.9348, 0.8586
final R indices	$R_1 = 0.0467,$	$R_1 = 0.0591$	R1 = 0.0391,
$[I > 2\sigma(I)]$	$wR_2 = 0.1138$	$wR_2 = 0.1310$	wR2 = 0.1026
<i>R</i> indices (all data)	$R_1 = 0.0517,$	$R_1 = 0.0925$	R1 = 0.0437,
	$wR_2 = 0.1167$	$wR_2 = 0.1459$	wR2 = 0.1057
goodness-of-fit	1.096	1.043	1.057
Largest diff. peak	0.542/-0.304	0.308/-0.211	0.361/-0.216

 Table S1. Selected X-ray crystallographic data for complexes 2, 4, 5, 12, 15.

Ta	ble	S1	(cont.)
			· · ·

	12	15
formula	$C_{13}H_{14}N_4S_2$	$C_{11}H_{10}N_2S$
fw	290.40	202.27
color, habit	red, rod	colorless, thin plate
cryst size [mm]	0.60 x0.36 x0.30	0.20 x 0.16 x 0.04
temp [K]	223(2)	100(2)
crystsyst	orthorhombic	monoclinic
space group	P2(1)2(1)2(1)	P21/n
<i>a</i> [Å]	9.035(5)	11.0937(7)
<i>b</i> [Å]	11.727(7)	6.0127(4)
<i>c</i> [Å]	13.103(8)	13.9608(10)
α [deg]	90.00	90.00
β [deg]	90.00	93.459(5)
γ[deg]	90.00	90.00
$V[Å^3]$	1388.4(14)	929.53(11)
Ζ	4	4
$D_{\rm c} [{\rm g}{\rm cm}^{-3}]$	1.389	1.445
radiation used	Μο Κα	Cu Ka
μ [mm ⁻¹]	0.375	2.716
θ range [deg]	2.33-27.50	4.949-70.072
no. of unique data	9140	7913
max., min. transmn	0.8959, 0.8064	0.7533, 0.5482
final R indices	R1 = 0.0369,	$R_1 = 0.0556,$
$[I \ge 2\sigma(I)]$	wR2= 0.0893	$wR_2 = 0.1304$
R indices (all data)	R1 = 0.0391,	$R_1 = 0.0809,$
	wR2 = 0.0906	$wR_2 = 0.1447$
goodness-of-fit	1.071	1.024
Largest diff. peak	0.333/-0.176	0.540/-0.420

References

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- Ghabbour, H.-K. Fun, *Molecules* **2016**, *21*, 12–23.
- S2 SMART, version 5.628; Bruker AXS Inc.: Madison, WI, 2001.
- S3 SAINT+, version 6.22a; Bruker AXS Inc.: Madison, WI, 2001.
- S4 G. W. Sheldrick, *SADABS* version 2.10; University of Göttingen: Göttingen, Germany, 2001.
- S5 SHELXTL, version 6.14; Bruker AXS Inc.: Madison, WI, 2000.