

Contents

General methods.....	2
Experimental procedures	3
Glycosylations.....	12
References:.....	26
NMR spectra.....	27

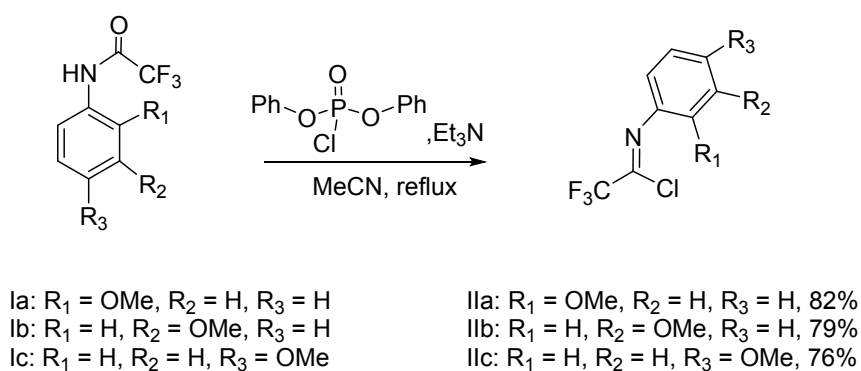
General methods

Chemicals were used as purchased without further purification except 2-methoxyaniline, which was vacuum distilled prior to use. Dry solvents were obtained from a purification solvent system. All reactions of air and moisture sensitive materials were performed under nitrogen atmosphere, with all glassware pre-dried in an oven at 200 °C overnight or flame dried prior to use. Columns were packed with pre-neutralized silica gel LC-60A (40 – 63 μ M, pre-neutralized from the manufacturer to a pH of 6.0 to 8.0). TLC analysis was performed on plates (Merck 60, F254), the detection was by examination under UV light (254 nm) and by charring with 10% sulfuric acid in ethanol. ^1H NMR and ^{13}C NMR spectra were acquired using a 500 MHz Avance III HD equipped with a cryogenically cooled 5 mm observe probe optimized for ^{13}C . Chemical shifts are reported in parts per million (ppm) relative to residual solvents signals ($\delta = 7.26$ for ^1H NMR and 77.16 for ^{13}C NMR) as the internal standard. ^{19}F NMR spectra were recorded on a 500 MHz instrument using trifluoroacetic acid ($\delta = -76.55$ ppm, CDCl_3 solution) in a capillary served as an internal standard. High-resolution mass spectral (HRMS) data were obtained by MALDI – MS using a Solarix XR 7 T ESI/MALDI – FT – ICR MS instrument. Optical rotation data were obtained on an Anton-Paar Polarimeter. All melting points are uncorrected.

Experimental procedures

Synthesis of *N*-substituted imidoyl chlorides

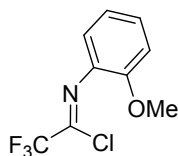
To the solution of a corresponding 2,2,2-trifluoro-*N*-phenylacetamide^[1] in MeCN (*c* = 1.3 M) diphenyl phosphoryl chloride was added (2 equiv) followed by the addition of Et₃N (2 equiv) and the reaction mixture was refluxed (85 °C) overnight. Subsequently, the reaction mixture was cooled down to room temperature, ethyl acetate was added, and the precipitate was filtered off. The filtrate was concentrated in *vacuo* and the crude product was distilled under reduced pressure to afford the desired products.^[2]



Scheme 1. *N*-substituted imidoyl chloride synthesis.

***N*-(2-methoxyphenyl)-2,2,2-**

trifluoroacetimidoyl chloride (IIa)



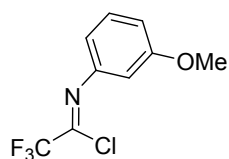
Yield: 82%, 7.10 g

¹H NMR (500 MHz, CDCl₃): δ 7.24 (ddd, *J* = 8.3, 7.4, 1.7 Hz, 1H), 7.02-6.96, (m, 2H), 6.93 (dd, *J* = 7.7, 1.7 Hz, 1H), 3.83 (s, 3H).

¹³C NMR{¹H NMR} (126 MHz, CDCl₃): δ 149.4, 134.2 (*q* = 42.7 Hz), 133.3, 128.1, 120.7, 120.4, 116.9 (*q* = 277.1 Hz), 112.0, 55.8.

The NMR values in accordance with the literature.^[1]

***N*-(3-methoxyphenyl)-2,2,2-trifluoroacetimidoyl chloride (IIb)**



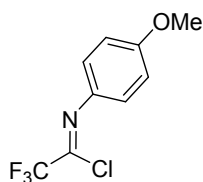
Yield: 79%, 6.85 g

¹H NMR (500 MHz, CDCl₃): δ 7.24-7.14 (m, 2H), 6.98, (ddd, *J* = 7.9, 2.0, 0.7 Hz, 1H), 6.72 (ddd, *J* = 8.3, 2.5, 0.8 Hz, 1H), 3.75 (s, 3H).

¹³C NMR{¹H NMR} (126 MHz, CDCl₃): δ 160.4, 144.8, 132.3 (*q*, *J* = 43.1 Hz), 130.1, 128.1, 120.7, 120.4, 117.0 (*q*, *J* = 277.0 Hz), 113.1, 112.7, 106.4, 79.9, 55.3.

The NMR values in accordance with the literature.^[1]

***N*-(4-methoxyphenyl)-2,2,2-trifluoroacetimidoyl chloride (IIc)**



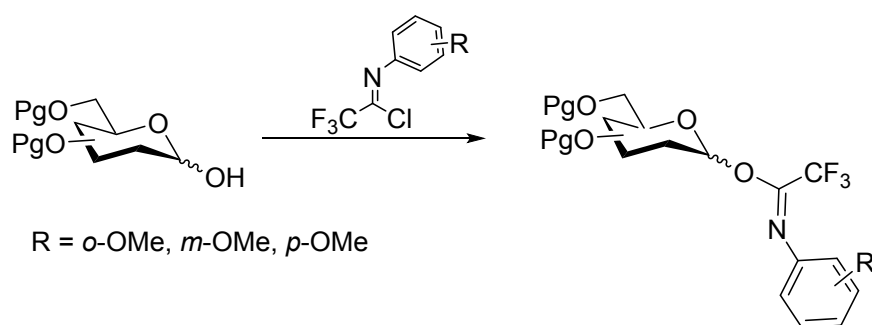
Yield: 76%, 2.32 g

¹H NMR (500 MHz, CDCl₃) δ 7.36-7.31 (m, 2H), 6.99-6.95 (m, 2H), 3.84 (s, 3H).

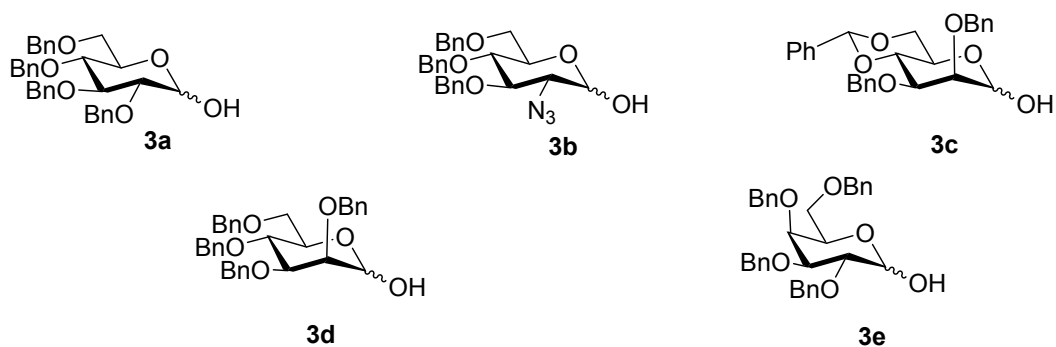
^{13}C NMR{ ^1H NMR} (126 MHz, CDCl_3) δ 159.7, 135.6, 128.1 (q, $J = 42.9$ Hz), 124.5, 120.5, 117.2 (q, $J = 276.5$ Hz), 55.6.

The NMR values in accordance with the literature. ^[1]

Synthesis of *N*-(methoxyphenyl) trifluoroacetimidate donors



List of hemiacetals used in this study:



General procedure A

To the solution of hemiacetal (3a-e) in CH_2Cl_2 carbonate (2 equiv) was added (see table) followed by the addition of *N*-phenyl-2,2,2-trifluoroacetimidoyl chloride (2 equiv) (IIa-c) and the reaction mixture was stirred at room temperature overnight unless otherwise stated. The solids were filtered, and the filtrate was concentrated in *vacuo*. The crude products were purified by column chromatography to afford desired PTFA donors.

General procedure B

To the solution of hemiacetal (3a-e) in DMF DABCO (1,4-diazabicyclo[2.2.2]octane) (4 equiv) was added followed by the addition of *N*-phenyl trifluoroacetimidoyl chloride (1.1 equiv). When TLC analysis indicated the full conversion of starting material, the reaction mixture was concentrated in *vacuo*, diluted with diethyl ether, and washed with NaHCO_3 (10% aq. solution) and water (3x), respectively. The organic phase was dried over MgSO_4 , filtered and concentrated in *vacuo*. The crude products were purified by column chromatography to afford desired PTFA donors.

General procedure C

To the solution of hemiacetal (3a-e) in CH_2Cl_2 Et_3N (6.7 equiv) was added. The reaction mixture was stirred for 10 min, and then a solution ($c = 0.3 \text{ M}$) of *N*-phenyl trifluoroacetimidoyl chloride (2 equiv) was added via syringe pump (1 mL/h). When TLC analysis indicated full conversion of starting material, the reaction mixture was washed with NaHCO_3 (conc. aq. solution) and brine, respectively. The organic phase was dried over MgSO_4 , filtered, and concentrated in *vacuo*. The crude products were purified by column chromatography to afford desired PTFA donors.

General procedure D

To the solution of hemiacetal (3a-e) in pyridine (6.7 equiv) *N*-phenyl trifluoroacetimidoyl chloride (2 equiv) was added. When TLC analysis indicated full conversion of starting material, the reaction mixture was diluted with EtOAc and washed with water. The organic phase was dried over MgSO_4 , filtered, and concentrated in *vacuo*. The crude products were purified by column chromatography to afford desired PTFA donors.

General procedure E

To the solution of hemiacetal (3a-e) in DMF NaH (60% dispersion in mineral oil) (1.1 equiv) was added. The reaction mixture was stirred for 10 min followed by the addition of *N*-phenyl trifluoroacetimidoyl chloride (1.1 equiv). When TLC analysis indicated full conversion of starting material, the reaction mixture was diluted with ethyl acetate and washed with water (3x). The organic phase was dried over MgSO_4 , filtered, and concentrated in *vacuo*. The crude products were purified by column chromatography to afford desired PTFA donors.

Entry	Hemiacetal	Electrophile (2a-c)	General procedure	Base	Solvent, (concentration) [M]	Time [h]	α/β^a (Yield) [%]
1	3a	2a	B	DABCO	DMF, (c = 0.35)	2	β only (91)
2	3a	2a	A	K ₂ CO ₃	CH ₂ Cl ₂ , (c = 0.15)	24	β only (82)
3	3a	2a	A	Cs ₂ CO ₃	CH ₂ Cl ₂ , (c = 0.15)	24	β only (77)
4	3a	2a	C	Et ₃ N	CH ₂ Cl ₂ , (c = 0.1)	2	β only (67)
5	3a	2a	A	K ₂ CO ₃	CH ₂ Cl ₂ , (c = 0.05)	48	β only (74)
6	3a	2a	D	-	pyridine (c = 0.1)	24	Nr
7	3a	2a	C	Et ₃ N	CH ₂ Cl ₂ , (c = 0.1) ^b	2	β only (74)
8	3a	2a	C	Et ₃ N ^c	CH ₂ Cl ₂ , (c = 0.1) ^b	1	β only (71)
9	3a	2a	C	Et ₃ N ^d	CH ₂ Cl ₂ , (c = 0.1) ^b	24	nr
10	3a	2a	A	K ₂ CO ₃	CH ₂ Cl ₂ , (c = 0.15) ^b	8	β only (82)
11	3a	2a	A	DBU	CH ₂ Cl ₂ , (c = 0.1) ^b	24	nr
12	3a	2a	A	NaH	CH ₂ Cl ₂ , (c = 0.1) ^a	24	nr
13	3a	2b	A	K ₂ CO ₃	CH ₂ Cl ₂ , (c = 0.15)	24	β only ^f
14	3a	2b	B	DABCO	DMF, (c = 0.35)	2	β only ^f
15	3a	2c	A	K ₂ CO ₃	CH ₂ Cl ₂ , (c = 0.15)	24	β only ^e
16	3a	2c	B	DABCO	DMF, (c = 0.35)	1.5	β only ^f
17	3a	2c	C	Et ₃ N	CH ₂ Cl ₂ , (c = 0.1) ^b	1	β only ^f
18	3b	2a	B	DABCO	DMF, (c = 0.35)	4	β only (84)
19	3b	2a	A	K ₂ CO ₃	CH ₂ Cl ₂ , (c = 0.15)	24	β only (76)
20	3c	2a	B	DABCO	CH ₂ Cl ₂ , (c = 0.05)	48	1:1 (74)
21	3c	2a	A	K ₂ CO ₃ ^e	CH ₂ Cl ₂ , (c = 0.05)	48	1:1 (66)
22	3d	2a	A	DBU	CH ₂ Cl ₂ , (c = 0.1) ^b	24	nr
23	3d	2a	E	NaH	DMF, (c = 0.1) ^b	24	nr
24	3d	2a	B	DABCO	DMF, (c = 0.35)	2	β only (88)
25	3d	2a	A	K ₂ CO ₃	CH ₂ Cl ₂ , (c = 0.15)	24	β only (73)
26	3d	2a	A	Cs ₂ CO ₃	CH ₂ Cl ₂ , (c = 0.15)	6	β only (81)
27	3d	2a	C	Et ₃ N	CH ₂ Cl ₂ , (c = 0.1)	24	β only (65)
28	3d	2a	A	K ₂ CO ₃	CH ₂ Cl ₂ , (c = 0.15) ^b	4 h	β only (69)
29	3d	2a	A	DIPEA	CH ₂ Cl ₂ , (c = 0.1)	6 h	β only (72)
30	3d	2a	B	DABCO	DMF, (c = 0.35) ^b	1	β only (66)
31	3e	2a	B	DABCO	DMF, (c = 0.35)	2	β only (76)

^a Anomeric ratio of the crude mixture

^b Reflux conditions.

^c TBAI (10% mol) was added.

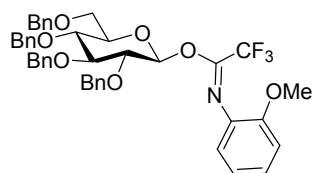
^d DMAP (10% mol) was added.

^e Sonication bath was used.

^f Product decomposed on the pre-neutralized silica and basic alumina.

Table 1. *N*-substituted trifluoroacetimidate synthesis.

2,3,4,6-tetra-*O*-benzyl- β -D-glucopyranosyl *N*-(2-methoxyphenyl)-2,2,2-trifluoroacetimidate (D1)



Off-white amorphous solid.

Yield: 91% (4.64 g)

¹H NMR (500 MHz, CDCl₃) δ 7.31-7.26 (m, 10H), 7.25 – 7.19 (m, 8H), 7.12 - 7.09 (m, 2H), 6.99 (td, J = 8.0, 1.7 Hz, 1H), 6.82 – 6.77 (m, 2H), 6.75 – 6.71 (m, 1H), 4.87 (d, J = 12.1 Hz, 1H), 4.85 (d, J = 11.0 Hz, 1H), 4.76 (d, J = 10.9 Hz, 2H), 4.71 (d, J = 10.8 Hz, 1H), 4.58 (d, J = 12.1 Hz, 1H), 4.50 (d, J = 11.9 Hz, 2H), 3.77 – 3.56 (m, 10H).

¹³C NMR{¹H NMR} (126 MHz, CDCl₃) δ 149.0, 138.6, 138.1 (C x 2), 138.0, 132.9, 128.7 - 127.5 (C x 22), 125.2, 120.6, 120.5, 111.2, 97.4, 84.6, 81.4, 81.1, 77.3, 75.8, 75.2, 75.0, 73.6, 68.2, 55.6.

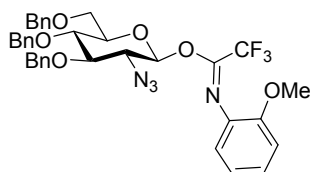
¹⁹F NMR{¹H NMR} (470 MHz, CDCl₃) δ -69.3.

HRMS (MALDI) m/z : [M+Na⁺] Calcd for C₄₃H₄₂F₃NO₇Na⁺ 764.2806; found 764.2811.

$[\alpha]_D^{25}$ = +45.3 (c = 1.02 CHCl₃)

Melting point: 80-82 °C

2-azido-2-deoxy-3,4,6-tri-*O*-benzyl- β -D-glucopyranosyl *N*-(2-methoxyphenyl)-2,2,2-trifluoroacetimidate (D2)



Yellowish oil.

Yield: 84%, 1.13 g

¹H NMR (500 MHz, CDCl₃) δ 7.38 – 7.24 (m, 13H), 7.20 – 7.14 (m, 3H), 7.08 – 7.03 (m, 1H), 6.88 – 6.84 (m, 2H), 6.79 (dd, J = 7.5, 1.3 Hz, 1H), 4.87 (d, J = 11.0 Hz, 1H), 4.81 (d, J = 10.8 Hz, 1H), 4.78 (d, J = 11.0 Hz, 1H), 4.60 (d, J = 12.0 Hz, 1H), 4.55 (d, J = 10.9 Hz, 1H), 4.52 (d, J = 12.1 Hz, 1H), 3.85 – 3.25 (m, 10H).

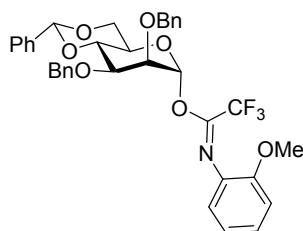
^{13}C NMR{ ^1H NMR} (126 MHz, CDCl_3) δ 149.0, 138.0, 137.9, 131.6, 129.2, 128.6 - 127.9 (C x 16), 125.4 (x 2), 120.6 (x 2), 111.2, 96.0, 83.1, 77.3, 76.0, 75.8, 75.2, 73.6, 68.0, 65.6, 55.7.

^{19}F NMR{ ^1H NMR} (470 MHz, CDCl_3) δ -69.5.

HRMS (MALDI) m/z : $[\text{M}+\text{Na}^+]$ Calcd for $\text{C}_{43}\text{H}_{42}\text{F}_3\text{NO}_7\text{Na}^+$ 699.2401; found 699.2393.

$[\alpha]_D^{25} = +33.6$ (c = 1.00 CHCl_3)

2,3-di-*O*-benzyl-4,6-*O*-benzylidene- α -D-mannopyranosyl *N*-(2-methoxyphenyl)-2,2,2-trifluoroacetimidate (D3 α)



Yellowish oil.

Yield: 74% (α/β , 1/1 anomeric mixture), 0.63 g (α anomer)

^1H NMR (500 MHz, CDCl_3) δ 7.44 (dd, $J = 7.7, 1.6$ Hz, 2H), 7.35 – 7.23 (m, 9H), 7.22 – 7.17 (m, 4H), 7.01 (td, $J = 8.0, 1.6$ Hz, 1H), 6.83 (td, $J = 7.6, 1.2$ Hz, 1H), 6.80 (dd, $J = 8.2, 1.0$ Hz, 1H), 6.66 (d, $J = 7.4$ Hz, 1H), 5.58 (s, 1H), 4.76 (d, $J = 12.1$ Hz, 1H), 4.67 (bs, 2H), 4.59 (d, $J = 12.0$ Hz, 1H), 4.28 - 4.21 (m, 2H), 3.98 – 3.73 (m, 5H), 3.65 (s, 3H).

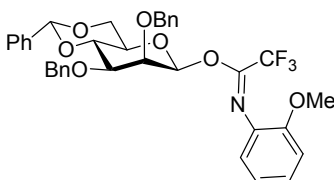
^{13}C NMR{ ^1H NMR} (126 MHz, CDCl_3) δ 149.0, 138.4, 137.7, 137.6, 132.9, 129.1, 128.5 - 127.8 (x 15), 126.2, 125.4, 120.7, 120.5, 111.2, 101.7, 96.1 ($J_{\text{C-H}} = 178.5$ Hz), 78.4, 75.7, 74.8, 73.6, 73.5, 68.6, 66.8, 55.6.

^{19}F NMR{ ^1H NMR} (470 MHz, CDCl_3) δ -69.8.

HRMS (MALDI) m/z : $[\text{M}+\text{Na}^+]$ Calcd for $\text{C}_{43}\text{H}_{42}\text{F}_3\text{NO}_7\text{Na}^+$ 672.2180; found 672.2175.

$[\alpha]_D^{25} = -1.8$ (c = 0.82 CHCl_3)

2,3-di-*O*-benzyl-4,6-*O*-benzylidene- β -D-mannopyranosyl *N*-(2-methoxyphenyl)-2,2,2-trifluoroacetimidate (D3 β)



Yellowish oil.

Yield: 74% (α/β , 1/1 anomeric mixture), 0.63 g (β anomer)

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.47 – 7.41 (m, 4H), 7.37 – 7.22 (m, 10H), 7.05 (td, $J = 8.1, 1.6$ Hz, 1H), 6.89 – 6.72 (m, 2H), 6.70 (d, $J = 7.5$ Hz, 1H), 5.58 (s, 1H), 4.93 (d, $J = 11.9$ Hz, 1H), 4.86 (d, $J = 11.9$ Hz, 1H), 4.74 (d, $J = 12.3$ Hz, 1H), 4.60 (d, $J = 12.3$ Hz, 1H), 4.30 – 4.15 (m, 4H), 3.95 – 3.86 (m, 1H), 3.77 – 3.59 (m, 5H).

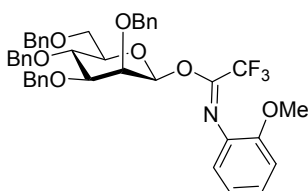
$^{13}\text{C NMR}\{^1\text{H NMR}\}$ (126 MHz, CDCl_3) δ 148.9, 138.3, 138.1, 137.5, 132.7, 129.1, 128.7 – 127.7 (C x 15), 126.2, 125.5, 120.7, 120.5, 111.3, 101.7, 96.5 ($J_{\text{C-H}} = 163.2$ Hz), 78.4, 78.1, 75.1, 74.8, 72.9, 68.4, 68.3, 55.7.

$^{19}\text{F NMR}$ (470 MHz, CDCl_3) δ -69.4.

HRMS (MALDI) m/z : $[\text{M}+\text{K}^+]$ Calcd for $\text{C}_{43}\text{H}_{42}\text{F}_3\text{NO}_7\text{K}^+$ 688.1919; found 688.1913.

$[\alpha]_D^{25} = +33.6$ ($c = 1.00$ CHCl_3)

2,3,4,6-tetra-*O*-benzyl- β -D-mannopyranosyl *N*-(2-methoxyphenyl)-2,2,2-trifluoroacetimidate (D4)



Colourless oil.

Yield: 88%, 0.94 g

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.41 (d, $J = 6.4$ Hz, 2H), 7.29 – 7.27 (m, 5H), 7.25 – 7.19 (m, 11H), 7.16 – 7.11 (m, 2H), 7.01 (td, $J = 7.9, 1.6$ Hz, 1H), 6.82 – 6.78 (m, 2H), 6.65 (d, $J = 7.5$ Hz, 1H), 4.93 (d, $J = 12.2$ Hz, 1H), 4.83 (d, $J = 10.4$ Hz, 1H), 4.80 (d, $J = 11.4$ Hz, 1H), 4.61 – 4.48 (m, 5H), 4.15 (bs, 1H), 3.77 – 3.69 (m, 3H), 3.70 – 3.52 (m, 6H).

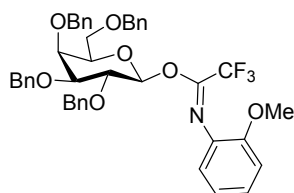
$^{13}\text{C NMR}\{^1\text{H NMR}\}$ (126 MHz, CDCl_3) δ 149.0, 138.5, 138.4, 138.3, 138.1, 133.0, 128.6 – 127.7 (C x 24), 125.2, 120.64, 111.2, 96.4 ($J_{\text{C-H}} = 162.6$ Hz), 82.0, 75.2, 74.4, 74.1, 73.6, 72.9, 72.1, 69.1, 55.6.

$^{19}\text{F NMR}\{^1\text{H NMR}\}$ (470 MHz, CDCl_3) δ -69.3.

HRMS (MALDI) m/z : $[\text{M}+\text{Na}^+]$ Calcd for $\text{C}_{43}\text{H}_{42}\text{F}_3\text{NO}_7\text{Na}^+$ 764.2806; found 764.2798.

$[\alpha]_D^{25} = +17.5$ ($c = 1.00$ CHCl_3)

2,3,4,6-tetra-*O*-benzyl- β -D-galactopyranosyl *N*-(2-methoxyphenyl)-2,2,2-trifluoroacetimidate (D5)



Yellowish oil.

Yield: 76%, 0.71 g

¹H NMR (500 MHz, CDCl₃) δ 7.31 – 7.18 (m, 18H), 7.18 – 7.14 (m, 3H), 6.95 (td, J = 8.0, 1.6 Hz, 1H), 6.77-6.72 (m, 2H), 6.69 – 6.64 (m, 1H), 4.87 (d, J = 11.2 Hz, 1H), 4.78 (d, J = 10.6 Hz, 1H), 4.81 – 4.75 (m, 1H), 4.73 (d, J = 10.7 Hz, 1H), 4.70 – 4.61 (m, 1H), 4.54 (d, J = 11.5 Hz, 1H), 4.38 (d, J = 11.7 Hz, 1H), 4.32 (d, J = 11.7 Hz, 1H), 3.98 (bs, 1H), 3.88-3.84 (m, 1H), 3.6 – 3.31 (m, 8H).

¹³C NMR (126 MHz, CDCl₃) δ 149.0, 138.6, 138.4, 138.3, 138.0, 133.2, 128.6 – 127.7 (C x 23), 125.0, 120.5, 111.2, 97.5, 82.2, 78.3, 75.5, 74.9, 74.5, 73.6, 73.4, 73.2, 68.3, 55.6.

¹⁹F NMR (470 MHz, CDCl₃) δ -70.0.

HRMS (MALDI) m/z : [M+Na⁺] Calcd for C₄₃H₄₂F₃NO₇Na⁺ 764.2806; found 764.2799.

$[\alpha]_D^{25}$ = +42.7 (c = 0.98 CHCl₃)

Glycosylations

General procedure A

Under an inert atmosphere, to the solution of the donor (1.2 equiv) and acceptor (1 equiv) in the corresponding solvent ($c = 0.05$ M) (see table) activator was added (10 mol% with respect to donor). The reaction mixture was monitored with TLC. Upon completion, the reaction was quenched with Et_3N . The solids were filtered over Celite, and the filtrate was concentrated in *vacuo*. The crude NMR was recorded in order to determine the anomers ratio. The crude products were purified by column chromatography to afford the products.

General procedure B

Under an inert atmosphere, to the solution of the donor (1.2 equiv) and acceptor (1 equiv) in the corresponding solvent ($c = 0.05$ M) (see table) acid scavenger (2,4,6-tri-*tert*-butylpyrimidine) (0.5 equiv), was added followed by the addition of activator (10 mol% with respect to donor). The reaction mixture was monitored with TLC. Upon completion, the reaction was quenched with Et_3N . The solids were filtered over Celite, and the filtrate was concentrated in *vacuo*. The crude NMR was recorded in order to determine the anomers ratio. The crude products were purified by column chromatography to afford the products.

General procedure C

Under an inert atmosphere, to the solution of donor (1.2 equiv) in the corresponding solvent ($c = 0.05$ M) (see table) activator (10 mol% with respect to donor) was added followed by the addition of acceptor (1 equiv). The reaction mixture was monitored with TLC. Upon completion, the reaction was quenched with Et_3N . The solids were filtered over Celite, and the filtrate was concentrated in *vacuo*. The crude NMR was recorded in order to determine the anomers ratio. The crude products were purified by column chromatography to afford the products.

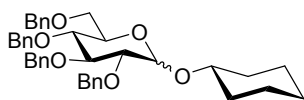
General procedure D

Under an inert atmosphere, to the solution of acceptor (1.0 equiv) in the corresponding solvent ($c = 0.05$ M) (see table) activator (10 mol% with respect to donor) was added followed by the slow addition (via syringe pump - 1 mL/1h) of donor (1.2 equiv) in 1 mL of solvent. The reaction mixture was monitored with TLC. Upon completion, the reaction was quenched with Et_3N . The solids were filtered over Celite, and the filtrate was concentrated in *vacuo*. The crude NMR was recorded in order to determine the anomers ratio. The crude products were purified by column chromatography to afford the products.

Glycosylation yields provided for the General procedure A: $\text{Fe}(\text{OTf})_3$ was used as a catalyst and diethyl ether as a solvent.

Cyclohexyl 2,3,4,6-tetra-*O*-

benzyl-D-glucopyranoside (1)



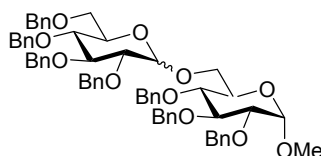
Yield: 95%, 87 mg

1 α : ^1H NMR (500 MHz, CDCl_3) δ 7.40 – 7.23 (m, 18H), 7.20 – 7.12 (m, 2H), 5.01 (d, J = 10.9 Hz, 1H), 4.98 (d, J = 3.7 Hz, 1H), 4.86 – 4.80 (m, 2H), 4.77 (d, J = 12.1 Hz, 1H), 4.54 – 4.44 (m, 2H), 4.02 (t, J = 9.3 Hz, 1H), 3.90–3.88 (m, 1H), 3.77 (dd, J = 10.6, 3.8 Hz, 1H), 3.67 – 3.62 (m, 2H), 3.59 – 3.54 (m, 2H), 1.99 – 1.72 (m, 3H), 1.58 – 1.16 (m, 7H).

1 β : ^1H NMR (500 MHz, CDCl_3) δ 7.43 – 7.23 (m, 18H), 7.21 – 7.14 (m, 2H), 5.00 (d, J = 10.9 Hz), 4.93 (d, J = 10.8 Hz), 4.84 (d, J = 10.9 Hz), 4.76 (d, J = 11.1 Hz, 1H), 4.70 (d, J = 10.9 Hz, 1H), 4.64 – 4.55 (m, 3H), 4.51 (d, J = 7.9 Hz, 1H), 3.80 – 3.71 (m, 2H), 3.69 – 3.61 (m, 2H), 3.56 (t, J = 9.3 Hz, 1H), 3.49 – 3.40 (m, 2H), 2.03 – 1.88 (m, 2H), 1.85 – 1.71 (m, 2H), 1.54 – 1.14 (6H).

The NMR values in accordance with literature. ^[3]

Methyl 2,3,4-tri-*O*-benzyl-6-*O*-(2,3,4,6-tetra-*O*-benzyl-D-glucopyranosyl)- α -D-glucopyranoside (2)



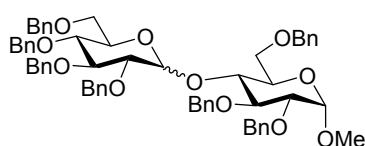
Yield: 96%, 147 mg

2 α : ^1H NMR (500 MHz, CDCl_3) δ 7.38 – 7.20 (m, 33H), 7.17 – 7.10 (m, 2H), 4.99 (d, J = 3.9 Hz, 1H), 4.96 (d, J = 11.6 Hz, 1H), 4.92 (d, J = 11.3 Hz, 1H), 4.90 (d, J = 10.9 Hz, 1H), 4.81 (d, J = 11.1 Hz, 1H), 4.80 (d, J = 10.6 Hz, 1H), 4.77 – 4.71 (m, 2H), 4.66 – 4.55 (m, 5H), 4.54 (d, J = 3.6 Hz, 1H), 4.49 – 4.37 (m, 2H), 4.08 – 4.03 (m, 1H), 3.92 – 3.52 (m, 10H), 3.46 – 3.43 (m, 1H), 3.34 (s, 3H).

2 β : ¹H NMR (500 MHz, CDCl₃) δ 7.37 – 7.15 (m, 35H), 4.98 (d, J = 11.5 Hz, 1H), 4.96 (d, J = 11.0 Hz, 1H), 4.90 (d, J = 10.6 Hz, 1H), 4.80 (d, J = 10.9 Hz, 1H), 4.82 – 4.75 (m, 3H), 4.70 (d, J = 10.9 Hz, 1H), 4.65 – 4.61 (m, 2H), 4.60 – 4.45 (m, 5H), 4.34 (d, J = 7.9 Hz, 1H), 4.20 (m, 1H), 3.99 (d, J = 9.3 Hz, 1H), 3.84 (dd, J = 10.5, 3.0, 1H), 3.77 – 3.41 (m, 9H), 3.31 (s, 3H).

The NMR values in accordance with literature. ^[4]

Methyl 2,3,6-tri-*O*-benzyl-4-*O*-(2,3,4,6-tetra-*O*-benzyl-D-glucopyranosyl)- α -D-glucopyranoside (3)



Yield: 76%, 105

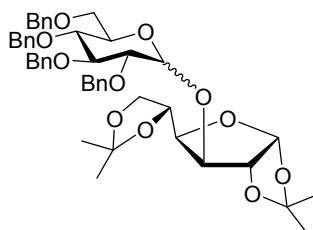
mg

3 α : ¹H NMR (500 MHz, CDCl₃) δ 7.32 – 7.18 (m, 33H), 7.11 – 7.07 (m, 2H) 5.68 (d, J = 3.6 Hz, 1H), 5.02 (d, J = 11.5 Hz, 1H), 4.85 (d, J = 10.9 Hz, 1H), 4.83 – 4.74 (m, 3H), 4.66 (d, J = 12.1 Hz, 1H), 4.61 – 4.46 (m, 7H), 4.40 (d, J = 10.8 Hz, 1H), 4.25 (d, J = 12.1 Hz, 1H), 4.08 (t, J = 9.1 Hz, 1H), 4.04 (t, J = 9.0 Hz, 1H), 3.92 (t, J = 9.3 Hz, 1H), 3.87 – 3.80 (m, 2H), 3.71 – 3.69 (m, 1H), 3.67 – 3.61 (m, 2H), 3.60 (dd, J = 9.3, 3.6 Hz, 1H), 3.50 – 3.45 (m, 2H), 3.39 – 3.34 (m, 4H)

3 β : ¹H NMR (500 MHz, CDCl₃) δ 7.43 – 7.38 (m, 2H), 7.33 – 7.15 (m, 33H), 5.10 (d, J = 11.2 Hz, 1H), 4.83 (d, J = 10.9 Hz, 1H), 4.80 – 4.71 (m, 6H), 4.63 – 4.52 (m, 4H), 4.43 (d, J = 12.1 Hz, 1H), 4.39 – 4.34 (m, 3H), 3.92 (t, J = 9.4 Hz, 1H), 3.86 – 3.80 (m, 2H), 3.71 (dd, J = 11.0, 1.9 Hz, 1H), 3.61 – 3.55 (m, 2H), 3.50 (dd, J = 11.0, 4.7 Hz, 1H), 3.51 – 3.43 (m, 3H), 3.38 – 3.30 (m, 4H), 3.25 (ddd, J = 10.0, 4.7, 1.9 Hz, 1H)

The NMR values in accordance with literature. ^[5]

1,2:5,6-di-*O*-isopropylidene-3-*O*-(2,3,4,6-tetra-*O*-benzyl-D-glucopyranosyl)- α -D-glucofuranose (4)



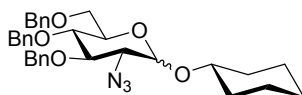
Yield: 76%, 93 mg

4 α : ¹H NMR (500 MHz, CDCl₃) δ 7.34 – 7.26 (m, 17H), 7.20 – 7.16 (m, 3H), 5.86 (d, J = 3.6 Hz, 1H), 5.25 (d, J = 3.5 Hz, 1H), 4.99 (d, J = 11.1 Hz, 1H), 4.87 – 4.81 (m, 3H), 4.75 (d, J = 12.0 Hz, 1H), 4.74 (m, 2H), 4.64 – 4.47 (m, 5H), 4.25 (d, J = 2.7 Hz, 1H), 4.17 (dd, J = 8.0, 2.7 Hz, 1H), 4.08 – 4.07 (m, 1H), 3.96 (t, J = 9.3 Hz, 1H), 3.84 – 3.82 (m, 1H), 3.74 (m, 2H), 3.65 – 3.61 (m, 2H), 1.52 (s, 3H), 1.44 (s, 3H), 1.29 (s, 3H), 1.25 (s, 3H).

4 β : ¹H NMR (500 MHz, CDCl₃) δ 7.38 – 7.24 (m, 20H), 5.97 (d, J = 3.3 Hz, 1H), 4.88 (d, J = 10.6 Hz, 1H), 4.87 (d, J = 10.5 Hz, 1H), 4.70 (d, J = 11.3 Hz, 1H), 4.65 (d, J = 12.0 Hz, 1H), 4.64 (d, J = 10.0 Hz, 1H), 4.62 (m, 1H), 4.56 (d, J = 3.3 Hz, 1H), 4.55–4.53 (m, 2H), 4.51 (d, J = 11.2 Hz, 1H), 4.32 – 4.296 (m, 3H), 4.15 (d, J = 3.3 Hz, 1H), 4.09 – 4.00 (m, 2H), 3.87 (dd, J = 3.3 Hz, 1H), 3.85 – 3.65 (m, 4H), 1.35 (s, 3H), 1.33 (s, 3H), 1.30 (s, 3H), 1.29 (s, 3H).

The NMR values in accordance with literature. [6]

Cyclohexyl 2-azido-2-deoxy-3,4,6-tri-*O*-benzyl-D-glucopyranoside (**5**)



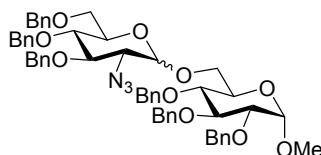
Yield: 80%, 72 mg

5 α : ¹H NMR (500 MHz, CDCl₃) δ 7.39 – 7.25 (m, 14H), 7.16 – 7.14 (m, 1H), 5.08 (d, J = 3.6 Hz, 1H), 4.90 – 4.84 (m, 2H), 4.80 (d, J = 10.9 Hz, 1H), 4.63 (d, J = 12.1 Hz, 1H), 4.51 – 4.42 (m, 2H), 4.02 (dd, J = 10.3, 8.8 Hz, 1H), 3.92 – 3.88 (m, 1H), 3.76 (dd, J = 10.8, 3.7 Hz, 1H), 3.70 (dd, J = 10.2, 8.8 Hz, 1H), 3.64 (dd, J = 10.7, 2.0 Hz, 1H), 3.62 (dt, J = 9.1, 4.0 Hz, 1H), 3.29 (dd, J = 10.3, 3.6 Hz, 1H), 1.94 – 1.80 (m, 2H), 1.78 – 1.66 (m, 2H), 1.54 – 1.47 (m, 1H), 1.46 – 1.36 (m, 2H), 1.32 – 1.19 (m, 3H).

5 β : ¹H NMR (500 MHz, CDCl₃) δ 7.39 – 7.25 (m, 14H), 7.16 – 7.14 (m, 1H), 5.05 (d, J = 3.6 Hz, 1H), 4.90 – 4.84 (m, 2H), 4.81 (d, J = 10.7 Hz, 1H), 4.65 (d, J = 12.1 Hz, 1H), 4.50 (dd, J = 10.5 Hz, 1H), 4.48 (dd, J = 10.9 Hz, 1H), 4.02 (dd, J = 10.3, 8.8 Hz, 1H), 3.92 (dt, J = 10.1, 2.8 Hz, 1H), 3.76 (dd, J = 10.7, 3.7 Hz, 1H), 3.70 (dd, J = 10.1, 8.8 Hz, 1H), 3.64 (dd, J = 10.7, 2.1 Hz, 1H), 3.62 (dt, J = 9.1, 4.0 Hz, 1H), 3.27 (dd, J = 10.2, 3.6 Hz, 1H), 1.94 – 1.80 (m, 2H), 1.78 – 1.66 (m, 2H), 1.54 – 1.47 (m, 1H), 1.46 – 1.36 (m, 2H), 1.32 – 1.19 (m, 3H).

The NMR values in accordance with the literature. [7]

Methyl 2,3,4-tri-*O*-benzyl-6-*O*-(2-azido-2-deoxy-3,4,6-tri-*O*-benzyl-D-glucopyranosyl)- α -D-glucopyranoside (6)



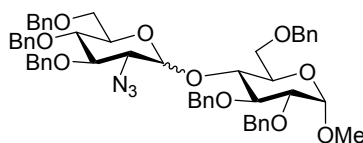
Yield: 83%, 119 mg

6 α ^1H NMR (500 MHz, CDCl_3) δ 7.40 – 7.24 (m, 28H), 7.17 – 7.14 (m, 1H), 5.00 (d, J = 3.6 Hz, 1H), 4.96 (d, J = 10.9 Hz, 1H), 4.92 (d, J = 11.2 Hz), 4.87 – 4.83 (m, 2H), 4.82 – 4.78 (m, 2H), 4.76 (d, J = 2.7 Hz, 1H), 4.65 (d, J = 12.1 Hz, 1H), 4.61 – 4.55 (m, 3H), 4.51 (d, J = 11.2 Hz, 1H), 4.40 (d, J = 12.1 Hz, 1H), 4.05 (t, J = 9.4 Hz, 1H), 3.94 (dd, J = 10.2, 8.6 Hz, 1H), 3.85 (dd, J = 11.3, 4.5 Hz, 1H), 3.79 – 3.72 (m, 2H), 3.71 – 3.65 (m, 2H), 3.63 (dd, J = 10.8, 3.3 Hz, 1H, H-6'), 3.59 – 3.50 (m, 3H), 3.36 (s, 3H), 3.33 (dd, J = 10.3, 3.6 Hz, 1H).

6 β : ^1H NMR (500 MHz, CDCl_3) δ 7.40 – 7.24 (m, 28H), 7.17 – 7.14 (m, 1H), 4.97 (d, J = 11.0 Hz, 1H), 4.94 (d, J = 11.2 Hz, 1H), 4.88 (d, J = 10.5 Hz, 1H), 4.85 (d, J = 10.9 Hz, 1H), 4.81 – 4.76 (m, 3H), 4.83 (d, J = 12.1 Hz, 1H), 4.64 (d, J = 3.6 Hz, 1H, H-1), 4.57 (d, J = 12.1 Hz, 1H), 4.53 (d, J = 12.2 Hz, 1H), 4.18 (d, J = 7.8 Hz, 1H), 4.12 (dd, J = 10.9, 1.6 Hz, 1H), 4.00 (dd, J = 9.6, 8.7 Hz, 1H), 3.81 – 3.79 (m, 1H), 3.70 – 3.62 (m, 3H), 3.57 – 3.53 (m, 1H), 3.52 (dd, J = 9.6, 8.7 Hz, 1H), 3.51 (dd, J = 9.6, 3.6 Hz, 1H), 3.45 – 3.39 (m, 2H), 3.3 (s, 3H), 3.32 (m, 1H).

The NMR values in accordance with the literature. [8]

Methyl 2,3,6-tri-*O*-benzyl-4-*O*-(2-azido-2-deoxy-3,4,6-tri-*O*-benzyl-D-glucopyranosyl)- α -D-glucopyranoside (7)



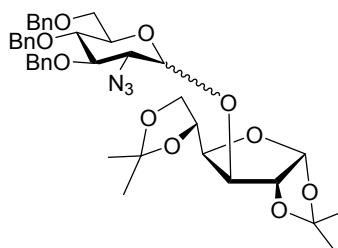
Yield: 84%, 119 mg

7 α : ¹H NMR (500 MHz, CDCl₃) δ 7.38 – 7.16 (m, 16H), 7.13 – 7.07 (m, 14H), 5.69 (d, J = 3.7 Hz, 1H), 5.06 (d, J = 10.8 Hz, 1H), 4.88 – 4.74 (m, 3H), 4.77 – 4.70 (m, 2H), 4.61 (d, J = 12.1 Hz, 1H), 4.56 (d, J = 3.6 Hz, 1H), 4.52 – 4.46 (m, 3H), 4.43 (d, J = 10.8 Hz, 1H), 4.23 (d, J = 12.1 Hz, 1H), 4.06 (t, J = 9.1 Hz, 1H), 3.87 (t, J = 9.3 Hz, 1H), 3.82 (dd, J = 10.2, 8.3 Hz, 1H), 3.75 – 3.72 (m, 1H), 3.71 (dd, J = 11.1, 4.5 Hz, 1H), 3.65 – 3.60 (m, 3H), 3.54 (dd, J = 9.6, 3.6 Hz, 1H), 3.50 (dd, J = 10.7, 2.5 Hz, 1H), 3.36 (s, 3H), 3.30 (dd, J = 10.8, 1.6 Hz, 1H), 3.25 (dd, J = 10.2, 3.9 Hz, 1H).

7 β : ¹H NMR (500 MHz, CDCl₃) δ 7.38 – 7.16 (m, 15H), 7.14 – 7.17 (m, 15H), 5.00 (d, J = 11.5 Hz, 1H), 4.83 (d, J = 11.6 Hz, 1H), 4.81 – 4.72 (m, 4H), 4.64 (d, J = 12.1 Hz, 1H), 4.60 – 4.56 (m, 2H), 4.53 (d, J = 10.9 Hz, 1H), 4.45 (d, J = 12.0 Hz, 1H), 4.40 – 4.32 (m, 2H), 4.21 (d, J = 8.0 Hz, 1H), 3.95 (t, J = 9.3 Hz, 1H), 3.91 (dd, J = 11.2, 3.6 Hz, 1H), 3.88 (t, J = 9.3 Hz, 1H), 3.75 (dt, J = 10.1, 2.5 Hz, 1H), 3.69 (dd, J = 10.9, 2.0 Hz, 1H), 3.63 – 3.56 (m, 2H), 3.51 – 3.44 (m, 2H), 3.35 (s, 3H), 3.30 (dd, J = 9.9, 8.1 Hz, 1H), 3.22 (t, J = 9.4 Hz, 1H), 3.14 (ddd, J = 9.9, 4.4, 1.9 Hz, 1H).

The NMR values in accordance with the literature.^[9]

1,2:5,6-di-*O*-isopropylidene-3-*O*-(2-azido-2-deoxy-3,4,6-tri-*O*-benzyl-D-glucopyranosyl)- α -D-glucopyranoside (8)



Colourless oil

Yield: 72%, 58 mg

8 α : ¹H NMR (500 MHz, CDCl₃) δ 7.31 – 7.26 (m, 7H), 7.25 – 7.20 (m, 6H), 7.09 – 7.06 (m, 2H), 5.80 (d, J = 3.6 Hz, 1H), 5.21 (d, J = 3.6 Hz, 1H), 4.84 – 4.76 (m, 2H), 4.73 (d, J = 10.6 Hz, 1H), 4.60 – 4.52 (m, 3H), 4.45 – 4.41 (m, 2H), 4.21 (d, J = 2.7 Hz, 1H), 4.10 (dd, J = 8.6, 6.2 Hz, 1H), 4.00 (dd, J = 8.6, 2.7 Hz, 1H), 3.90 (dd, J = 8.6, 5.6 Hz, 1H), 3.87 – 3.79 (m, 1H), 3.73 (ddd, J = 9.9, 3.9, 2.0 Hz, 1H), 3.69 (dd, J = 10.6, 3.8 Hz, 1H), 3.66 – 3.60 (m, 2H), 3.43 (dd, J = 10.3, 3.6 Hz, 1H), 1.41 (s, 3H), 1.34 (s, 3H), 1.28 (s, 3H), 1.17 (s, 3H).

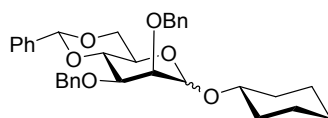
8 α : ¹³C NMR {¹H NMR} (126 MHz, CDCl₃) δ 137.8, 137.7, 137.6, 128.6–127.9 (C x 15), 112.0, 109.3, 105.2, 98.1, 83.8, 81.9, 81.5, 80.3, 80.1, 78.2, 75.6, 75.3, 73.6, 71.9, 71.6, 68.6, 67.3, 64.0, 26.8, 26.8, 26.2, 25.3.

8 β : ¹H NMR (500 MHz, CDCl₃) δ 7.30 – 7.19 (m, 13H), 7.14 – 7.11 (m, 2H), 5.91 (d, J = 3.8 Hz, 1H), 4.80 (d, J = 10.8 Hz, 1H), 4.77 – 4.69 (m, 3H), 4.56– 4.44 (m, 5H), 4.38 – 4.33 (m, 1H), 4.32 – 4.27 (m, 3H), 3.99 (dd, J = 6.3, 4.4 Hz, 2H), 3.71 – 3.55 (m, 4H), 3.38 – 3.27 (m, 2H), 1.44 (s, 3H), 1.35 (s, 3H), 1.27 (s, 3H), 1.23 (s, 3H).

8 β : ¹³C NMR {¹H NMR} δ 137.0, 136.8, 136.7, 127.9 – 126.9 (C x 15), 111.0, 107.6, 104.6, 99.3, 81.9, 81.8, 79.7, 79.3, 76.5, 74.5, 74.0, 74.1, 72.6, 72.3, 71.8, 67.7, 65.2, 65.0, 25.8, 25.5, 25.3, 24.3.

HRMS (MALDI) m/z : [M+Na⁺] Calcd for C₃₉H₄₇N₃O₁₀Na⁺ 740.3154; found 740.3159.

Cyclohexyl 2,3-di-*O*-benzyl-4,6-*O*-benzylidene-D-mannopyranoside (9)



Yield: 87%, 77 mg (starting from α donor)

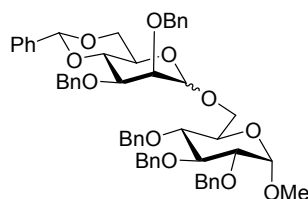
Yield: 83%, 73 mg (starting from β donor)

9 α : ¹H NMR (500 MHz, CDCl₃) δ 7.54 – 7.49 (m, 2H), 7.43 – 7.25 (m, 13H), 5.59 (s, 1H), 4.90 (d, J = 1.2 Hz, 1H), 4.86 (d, J = 11.9 Hz, 1H), 4.66 (d, J = 11.8 Hz, 1H), 4.35 – 4.19 (m, 2H), 4.05 – 4.00 (m, 1H), 3.92 – 3.83 (m, 2H), 3.78 (m, 1H), 3.55–3.44 (m, 1H), 1.88 – 1.79 (m, 1H), 1.75–1.68 (m, 3H), 1.54–1.50 (m, 2H), 1.42–1.12 (m, 4H)

9 β : ¹H NMR (500 MHz, CDCl₃) δ 7.54 – 7.49 (m, 2H), 7.43 – 7.25 (m, 13H), 5.65 (s, 1H), 5.00 (d, J = 12.3 Hz, 1H), 4.89 (d, J = 12.5 Hz, 1H), 4.66 (d, J = 12.5 Hz, 1H), 4.60 (d, J = 12.5 Hz, 1H), 4.55 (s, 1H), 4.30 (dd, J = 10.3, 4.9 Hz, 1H), 4.22 (t, J = 9.6 Hz, 1H), 3.95 (t, J = 10.4 Hz, 1H), 3.88 (d, J = 3.1 Hz, 1H), 3.72 – 3.66 (m, 1H), 3.58 (dd, J = 9.9, 3.3 Hz, 1H), 3.34–3.30 (m, 1H), 1.89 – 1.80 (m, 1H), 1.75–1.68 (m, 3H), 1.54–1.50 (m, 2H), 1.42–1.12 (m, 4H).

The NMR values in accordance with the literature. ^[10]

Methyl 2,3,4-tri-*O*-benzyl-6-*O*-(2,3-di-*O*-benzyl-4,6-*O*-benzylidene-D-mannopyranosyl)- α -D-glucopyranoside (10)



Yield: 81%, 125 mg (starting from α donor)

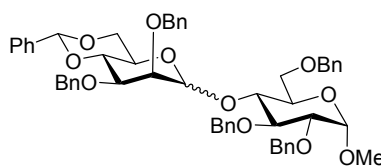
Yield: 79%, 122 mg (starting from β donor)

10 α : ^1H NMR (500 MHz, CDCl_3) δ 7.50 – 7.48 (m, 2H), 7.45 – 7.15 (m, 28H), 5.52 (s, 1H), 5.00 (d, J = 10.1 Hz, 1H), 4.86 (d, J = 12.1 Hz, 1H), 4.85 (d, J = 1.2 Hz, 1H), 4.84 – 4.73 (m, 4H), 4.67 (d, J = 12.8 Hz, 1H), 4.66 (d, J = 12.1 Hz, 1H), 4.65 (d, J = 11.9 Hz, 1H), 4.59 (d, J = 3.2 Hz, 1H), 4.50 (d, J = 10.1 Hz, 1H), 4.25 – 4.23 (m, 1H), 4.19 – 4.11 (m, 1H), 3.97 – 3.96 (m, 1H), 3.90 (dd, J = 9.4, 2.6 Hz, 1H), 3.76 – 3.68 (m, 4H), 3.67 – 3.65 (m, 1H), 3.59 (dd, J = 11.0, 1.2 Hz, 1H), 3.44 – 3.34 (m, 2H), 3.30 (s, 3H).

10 β : ^1H NMR (500 MHz, CDCl_3) δ 7.50 – 7.48 (m, 2H), 7.45 – 7.15 (m, 28H), 5.58 (s, 1H), 5.09 (d, J = 10.9 Hz, 1H), 4.96 (d, J = 12.1 Hz, 1H), 4.89 (d, J = 12.0 Hz, 1H), 4.87 – 4.75 (m, 3H), 4.74 – 4.58 (m, 4H), 4.54 (d, J = 11.5 Hz, 1H), 4.29 (dd, J = 10.4, 4.2 Hz, 1H), 4.20 (t, J = 9.5 Hz, 1H), 4.11 – 3.99 (m, 3H), 3.91–3.89 (m, 1H), 3.75 (d, J = 3.2 Hz, 1H), 3.69 (d, J = 2.9 Hz, 1H), 3.55 – 3.44 (m, 4H), 3.33 (s, 3H), 3.22 – 3.18 (m, 1H).

The NMR values in accordance with the literature. ^[10]

Methyl 2,3,6-tri-*O*-benzyl-4-*O*-(2,3-di-*O*-benzyl-4,6-*O*-benzylidene-D-mannopyranosyl)- α -D-glucopyranoside (11)



Yield: 73%, 130 mg (starting from α donor)

Yield: 77%, 136 mg (starting from β donor)

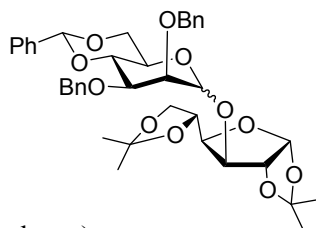
11 α : ^1H NMR (500 MHz, CDCl_3) δ 7.54 – 7.50 (m, 2H) 7.49–7.15 (m, 28H), 5.61 (s, 1H), 5.30 (d, J = 1.4 Hz, 1H), 5.11 (d, J = 11.5 Hz, 1H), 4.80 (d, J = 12.2 Hz, 1H), 4.66 (d, J = 12.0 Hz, 1H), 4.66–4.53 (m, 5H), 4.53 (d, J = 11.9 Hz, 1H), 4.42 (d, J = 11.9 Hz, 1H), 4.30–4.16 (m, 2H), 4.14–4.09 (m, 1H), 3.94 (dd, J = 9.9, 3.2 Hz, 1H), 3.91–3.67 (m, 8H), 3.53 (dd, J = 9.4, 3.5 Hz, 1H), 3.40 (s, 3H).

11 β : ^1H NMR (500 MHz, CDCl_3) δ 7.54 – 7.50 (m, 2H) 7.49–7.15 (m, 28H), 5.54 (s, 1H), 5.08 (d, J = 10.9 Hz, 1H), 4.90 – 4.77 (m, 3H), 4.77 (d, J = 12.4 Hz, 1H), 4.70 – 4.49 (m, 4H), 4.40 (s, 1H), 4.31 (d,

$J = 12.0$ Hz, 1H), 4.17 – 4.01 (m, 2H), 3.96 – 3.83 (m, 2H), 3.67 (d, $J = 3.0$ Hz, 1H), 3.65 – 3.61 (m, 1H), 3.60 – 3.44 (m, 3H), 3.43 (s, 3H), 3.36 (dd, $J = 9.8, 3.0$ Hz, 1H), 3.09 (td, $J = 9.8, 4.9$ Hz, 1H).

The NMR values in accordance with literature.^[10]

1,2:5,6-di-*O*-isopropylidene-3-*O*-(2,3-di-*O*-benzyl-4,6-*O*-benzylidene-D-mannopyranosyl)- α -D-glucofuranose



Yield: 69%, 86 mg (starting from α donor)

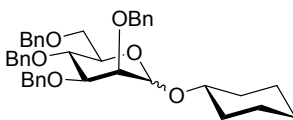
Yield: 67%, 84 mg (starting from β donor)

12 α : ^1H NMR (500 MHz, CDCl_3) δ 7.55 – 7.25 (m, 15H), 5.84 (d, $J = 3.6$ Hz, 1H), 5.67 (s, 1H), 5.21 (d, $J = 1.5$ Hz, 1H), 4.80 (d, $J = 12.1$ Hz, 1H), 4.76 (d, $J = 12.2$ Hz, 1H), 4.60 (d, $J = 12.2$ Hz, 1H), 4.53 (d, $J = 3.6$ Hz, 1H), 4.36–4.26 (m, 3H), 4.14–4.00 (m, 4H), 3.99–3.76 (m, 4H), 1.52 (s, 3H), 1.43 (s, 3H), 1.35 (s, 3H), 1.32 (s, 3H).

12 β : ^1H NMR (500 MHz, CDCl_3) δ 7.53 – 7.25 (m, 15H), 5.89 (d, $J = 3.9$ Hz, 1H), 5.63 (s, 1H), 4.87 (d, $J = 12.0$ Hz, 1H), 4.81 (d, $J = 11.9$ Hz, 1H), 4.75 (d, $J = 12.4$ Hz, 1H), 4.63 (d, $J = 12.0$ Hz, 1H), 3.86–3.83 (m, 1H), 3.61 (dd, $J = 9.7, 3.0$ Hz, 1H), 3.35–3.27 (m, 1H), 1.50 (s, 3H), 1.41 (s, 3H), 1.34 (s, 3H), 1.30 (s, 3H).

The NMR values in accordance with the literature.^[11]

Cyclohexyl 2,3,4,6-tetra-*O*-benzyl-D-mannopyranoside (13)



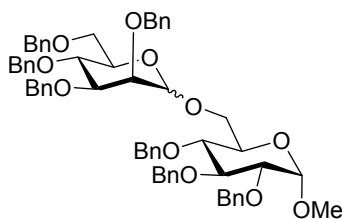
Yield: 88%, 84 mg

13 α : ^1H NMR (500 MHz, CDCl_3) δ 7.40–7.15 (m, 20 H), 5.00 (d, $J = 1.9$ Hz, 1H), 4.88 (d, $J = 10.7$ Hz, 1H), 4.80–4.62 (m, 4H), 4.55–4.43 (m, 2H), 3.98 (dd, $J = 9.3, 4.5$ Hz, 1H), 3.93 (dd, $J = 9.8, 3.0$, 1H), 3.88–3.82 (m, 1H), 3.80–3.77 (m, 2H), 3.73 (dd, $J = 9.3, 2.2$ Hz, 1H), 3.72 – 3.70 (m, 1H), 3.87–3.68 (m, 1H), 1.86 – 1.13 (m, 10H).

13 β : ^1H NMR (500 MHz, CDCl_3) δ 7.40-7.15 (m, 20 H), 4.91 (d, J = 12.2 Hz, 1H), 4.82 (d, J = 11.9 Hz, 1H), 4.81 (d, J = 10.9 Hz, 1H), 4.60 (d, J = 11.8 Hz, 1H), 4.51 (d, J = 10.5 Hz, 1H), 4.50 – 4.42 (m, 2H), 4.35 (d, J = 11.9 Hz, 1H), 4.29 (d, J = 10.5 Hz, 1H), 3.82 – 3.64 (m, 5H), 3. – 3.70 (m, 1H), 3.43-3.35 (m, 1H), and 1.86 - 1.13 (m, 10H).

The NMR values in accordance with the literature. ^[4]

Methyl 2,3,4-tri-*O*-benzyl-6-*O*-(2,3,4,6-tetra-*O*-benzyl-D-mannopyranosyl)- α -D-glucopyranoside (14)



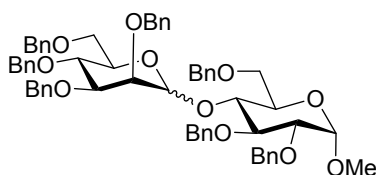
Yield: 83%, 114 mg

14 α : ^1H NMR (500 MHz, CDCl_3) δ 7.40 – 7.11 (m, 35H), 4.98 (d, J = 10.8 Hz, 1H), 4.96 (d, J = 1.8 Hz, 1H), 4.86 (dd, J = 11.0, 7.3 Hz, 2H), 4.82 (dd, J = 11.5, 6.1 Hz, 2H), 4.73 (d, J = 12.1 Hz, 1H), 4.70 (d, J = 11.1 Hz, 1H), 4.65 (d, J = 10.9 Hz, 1H), 4.63 - 4.54 (m, 4H), 4.50 (dd, J = 11.0, 2.5 Hz, 2H), 4.43 (d, J = 12.1 Hz, 1H), 3.99 (t, J = 9.5 Hz, 2H), 3.88 – 3.74 (m, 3H), 3.72 – 3.55 (m, 5H), 3.45 (dd, J = 9.6, 3.6 Hz, 1H), 3.42 (t, J = 9.3 Hz, 1H), 3.30 (s, 3H).

14 β : ^1H NMR (500 MHz, CDCl_3) δ 7.40 – 7.11 (m, 35H), 5.01 (d, J = 10.9 Hz, 1H), 4.94 (d, J = 12.0 Hz, 1H), 4.86 (d, J = 10.8 Hz, 1H), 4.83-4.81 (m, 2H), 4.81 (d, J = 11.0 Hz, 1H), 4.77 (d, J = 11.5 Hz, 1H), 4.66 (d, J = 11.6 Hz, 1H), 4.58 - 4.56 (m, 2H), 4.55 – 4.48 (m, 2H), 4.51 (dd, J = 11.0, 1H), 4.46 (d, J = 11.9 Hz, 1H), 4.18 (dd, J = 10.0, 1.2 Hz, 2H), 4.09 – 4.04 (m, 2H), 3.85 – 3.65 (m, 5H), 3.51 (dd, J = 9.3, 2.0 Hz, 1H), 3.49 – 3.35 (m, 4H), 3.33 (s, 3H).

The NMR values in accordance with the literature. ^[12]

Methyl 2,3,6-tri-*O*-benzyl-4-*O*-(2,3,4,6-tetra-*O*-benzyl-D-mannopyranosyl)- α -D-glucopyranoside (15)



Yield: 76%, 105 mg

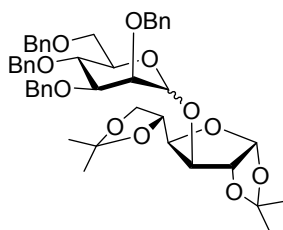
15 α : ^1H NMR (500 MHz, CDCl_3) δ 7.36 – 7.11 (m, 35H), 5.29 (d, J = 1.8 Hz, 1H), 5.11 (d, J = 11.9 Hz, 1H), 4.83 (d, J = 11.2 Hz, 1H), 4.68 (d, J = 12.2 Hz, 1H), 4.63 – 4.51 (m, 7H), 4.50 (d, J = 10.9

Hz, 1H), 4.44 (dd, $J = 12.0, 2.3$ Hz, 2H), 4.31 (d, $J = 12.1$ Hz, 1H), 4.21 (d, $J = 12.1$ Hz, 1H), 3.96 (t, $J = 9.4$ Hz, 1H), 3.88 – 3.77 (m, 3H), 3.74 – 3.61 (m, 5H), 3.65 (dd, $J = 10.8, 4.8$ Hz, 1H), 3.56 (dd, $J = 10.8, 1.8$ Hz, 1H), 3.54 (dd, $J = 9.6, 3.6$ Hz, 1H), 3.38 (s, 3H).

15 β : ¹H NMR (500 MHz, CDCl₃) δ 7.36 – 7.15 (m, 35H), 5.15 (d, $J = 11.5$ Hz, 1H), 4.86 (d, $J = 10.9$ Hz, 1H), 4.83 – 4.79 (m, 2H), 4.78 – 4.72 (m, 2H), 4.59 (d, $J = 12.2$ Hz, 1H), 4.57 (s, 1H), 4.58 (d, $J = 9.5$ Hz, 1H), 4.51 (d, $J = 10.9$ Hz, 1H), 4.49 – 4.42 (m, 2H), 4.40 (d, $J = 12.2$ Hz, 1H), 4.41 (s, 1H), 4.39 – 4.30 (m, 2H), 3.93 – 3.89 (m, 2H), 3.88 (t, $J = 9.6$ Hz, 1H), 3.74 – 3.70 (m, 1H), 3.69 (d, $J = 2.9$ Hz, 1H), 3.67 (dd, $J = 11.2, 1.8$ Hz, 1H), 3.58 – 3.55 (m, 2H), 3.54 (dd, $J = 11.3, 5.4$ Hz, 1H), 3.47 – 3.40 (m, 1H), 3.37 (s, 3H), 3.30 – 3.25 (m, 2H).

The NMR values in accordance with the literature. ^[13]

1,2:5,6-di-*O*-isopropylidene-3-*O*-(2,3,4,6-tetra-*O*-benzyl-D-mannopyranosyl)- α -D-glucofuranose (16)



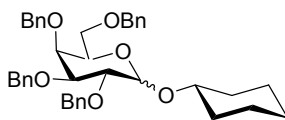
Yield: 71%, 89 mg

16 α : ¹H NMR (500 MHz, CDCl₃) δ 7.41-7.20 (m, 18H), 7.19-7.14 (m, 2H), 5.81 (d, $J = 3.6$ Hz, 1H), 5.25 (s, 1H), 4.92 (d, $J = 10.6$ Hz, 1H), 4.76 (d, $J = 12.2$ Hz, 1H), 4.72-4.60 (m, 3H), 4.60- 4.49 (m, 4H), 4.28-4.26 (m, 1H), 4.07-3.92 (m, 5H), 3.85-3.76 (m, 5H), 1.51 (s, 3H), 1.43 (s, 3H), 1.36 (s, 3H), 1.25 (s, 3H).

16 β : ¹H NMR (500 MHz, CDCl₃) δ 7.41-7.20 (m, 20H), 5.91 (d, $J = 3.6$ Hz, 1H), 4.90 (d, $J = 12.1$ Hz, 1H), 4.88 (d, $J = 10.6$ Hz, 1H), 4.79 (d, $J = 11.9$ Hz, 1H), 4.74 (d, $J = 12.1$ Hz, 1H), 4.60- 4.54 (m, 3H), 4.52 (d, $J = 11.9$ Hz, 1H), 4.49 (s, 1H), 4.48-4.44 (m, 2H), 4.35 (d, $J = 2.9$ Hz, 1H), 4.29 (dd, $J = 5.1, 3.1$ Hz, 1H), 4.13- 4.09 (m, 1H), 4.08-4.04 (m, 1H), 3.96 (t, $J = 9.6$ Hz, 1H), 3.87 (d, $J = 2.8$ Hz, 1H), 3.83-3.76 (m, 2H), 3.53 (dd, $J = 9.6, 2.9$ Hz, 1H), 3.43 – 3.41 (m, 1H), 1.50 (s, 3H), 1.39 (s, 3H), 1.33 (s, 3H), 1.25 (s, 3H).

The NMR values in accordance with the literature. ^[14]

Cyclohexyl 2,3,4,6-tetra-*O*-benzyl-D-galactopyranoside (17)



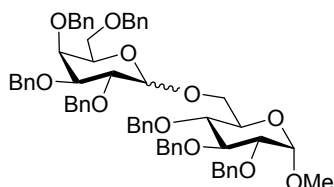
Yield: 86%, 80 mg

17 α : ^1H NMR (500 MHz, CDCl_3) δ 7.41 – 7.22 (m, 20H), 5.01 (d, J = 3.5 Hz, 1H), 4.95 (d, J = 11.5 Hz, 1H), 4.84 (d, J = 11.6 Hz, 1H), 4.78 (d, J = 12.0 Hz, 1H), 4.73 (d, J = 11.6 Hz, 1H), 4.67 (d, J = 11.9 Hz, 1H), 4.57 (d, J = 11.6 Hz, 1H), 4.48 (d, J = 11.5 Hz, 1H), 4.40 (d, J = 11.8 Hz, 1H), 4.07 (t, J = 6.5 Hz, 1H), 4.04 (dd, J = 9.9, 3.6 Hz, 1H), 3.99 – 3.94 (m, 2H), 3.57 – 3.50 (m, 3H), 1.94 – 1.33 (m, 10H).

17 β : ^1H NMR (500 MHz, CDCl_3) δ 7.41 – 7.22 (m, 20H), 4.97 (d, J = 10.9 Hz, 1H), 4.90 (d, J = 11.7 Hz, 1H), 4.77 – 4.72 (m, 2H), 4.70 (d, J = 11.8 Hz, 1H), 4.63 (d, J = 11.7 Hz, 1H), 4.45 (d, J = 7.7 Hz, 1H), 4.44 – 4.37 (m, 2H), 3.87 (d, J = 3.0 Hz, 1H), 3.80 (dd, J = 9.8, 7.7 Hz, 1H), 3.72 – 3.63 (m, 1H), 3.60 – 3.55 (m, 2H), 3.53 – 3.47 (m, 2H), 1.94 – 1.33 (m, 10H).

The NMR values in accordance with the literature. ^[7]

Methyl 2,3,4-tri-*O*-benzyl-6-*O*-(2,3,4,6-tetra-*O*-benzyl-D-galactopyranosyl)- α -D-glucopyranoside (18)



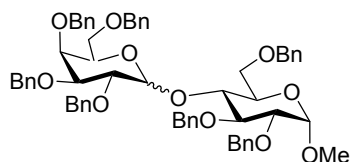
Yield: 81%, 116 mg

18 α : ^1H NMR (500 MHz, CDCl_3) δ 7.39 – 7.20 (m, 35H), 5.00 (d, J = 3.5 Hz, 1H), 4.95 (d, J = 11.5 Hz, 1H), 4.95 (d, J = 10.9 Hz, 1H), 4.84 (d, J = 11.0 Hz, 1H), 4.81 – 4.78 (m, 2H), 4.73 – 4.66 (m, 4H), 4.65 – 4.54 (m, 2H), 4.54 (d, J = 5.3 Hz, 1H), 4.51 (d, J = 3.5 Hz, 1H), 4.44 (d, J = 11.8 Hz, 1H), 4.37 (d, J = 11.9 Hz, 1H), 4.04 (dd, J = 5.9, 3.5 Hz, 1H), 3.99 – 3.87 (m, 4H), 3.80 – 3.71 (m, 3H), 3.60 (t, J = 9.3 Hz, 1H), 3.55 – 3.48 (m, 2H), 3.40 (dd, J = 6.0, 3.6 Hz, 1H), 3.32 (s, 3H).

18 β : ^1H NMR (500 MHz, CDCl_3) δ 7.39 – 7.20 (m, 35H), 4.99 – 4.92 (m, 3H), 4.83 (d, J = 10.9 Hz, 1H), 4.80 – 4.72 (m, 5H), 4.68 (d, J = 12.1 Hz, 1H), 4.60 – 4.57 (m, 2H), 4.52 (d, J = 11.2 Hz, 1H), 4.45 (d, J = 12.0 Hz, 1H), 4.41 (d, J = 11.6 Hz, 1H), 4.32 (d, J = 7.6 Hz, 1H), 4.15 (dd, J = 10.8, 2.0 Hz, 1H), 3.99 (t, J = 9.2 Hz, 1H), 3.93 – 3.81 (m, 3H), 3.65 – 3.45 (m, 7H), 3.31 (s, 3H).

The NMR values in accordance with the literature. ^[3]

Methyl 2,3,6-tri-*O*-benzyl-4-*O*-(2,3,4,6-tetra-*O*-benzyl-D-galactopyranosyl)- α -D-glucopyranoside (19)



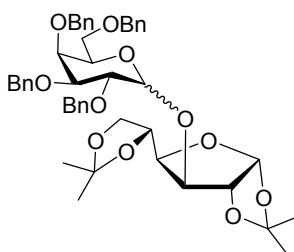
Yield: 77%, 102 mg

19 α : ^1H NMR (500 MHz, CDCl_3) δ 7.38 – 7.14 (m, 35H), 5.77 (d, J = 3.8 Hz, 1H), 4.99 (d, J = 11.6 Hz, 1H), 4.88 (d, J = 11.5 Hz, 1H), 4.81 (d, J = 11.5 Hz, 1H), 4.74 – 4.63 (m, 4H), 4.61 (d, J = 12.1 Hz, 1H), 4.58 (d, J = 3.6 Hz, 1H), 4.56 – 4.51 (m, 3H), 4.44 (d, J = 12.1 Hz, 1H), 4.33 (d, J = 11.6 Hz, 1H), 4.21 (d, J = 11.6 Hz, 1H), 4.09 (t, J = 9.2 Hz, 1H), 4.03 – 3.90 (m, 3H), 3.88 – 3.81 (m, 2H), 3.83 (dd, J = 10.3, 2.7 Hz, 1H), 3.70 (dd, J = 10.6, 4.7 Hz, 1H), 3.65 (m, 1H), 3.55 (dd, J = 9.6, 3.5 Hz, 1H), 3.48 (m, 1H), 3.43 (m, 1H), 3.36 (s, 3H).

19 β : ^1H NMR (500 MHz, CDCl_3) δ 7.38 – 7.14 (m, 35H), 5.03 (d, J = 10.5 Hz, 1H), 4.98 (d, J = 11.5 Hz, 1H), 4.83 – 4.78 (m, 2H), 4.75 – 4.71 (m, 2H), 4.70 – 4.65 (m, 2H), 4.62 (d, J = 12.1 Hz, 1H), 4.55 – 4.54 (m, 1H), 4.54 – 4.51 (m, 2H), 4.36 (d, J = 11.9 Hz, 1H), 4.32 (d, J = 12.1 Hz, 1H), 4.29 (d, J = 7.8 Hz, 1H), 4.22 (d, J = 11.8 Hz, 1H), 3.91 – 3.85 (m, 2H), 3.83 – 3.79 (m, 2H), 3.73 (dd, J = 9.7, 7.7 Hz, 1H), 3.60 (ddd, J = 10.0, 3.4, 2.0 Hz, 1H), 3.53 (t, J = 8.7 Hz, 1H), 3.50 – 3.44 (m, 2H), 3.38 – 3.35 (m, 1H), 3.34 (s, 3H), 3.33 – 3.28 (m, 2H).

The NMR values in accordance with the literature. ^[15]

1,2:5,6-di-*O*-isopropylidene-3-*O*-(2,3,4,6-tetra-*O*-benzyl-D-galactopyranosyl)- α -D-glucofuranose (20)



Yield: 64%, 78 mg

20 α : ¹H NMR (500 MHz, CDCl₃) δ 7.37 – 7.16 (m, 20H), 5.79 (d, J = 3.6 Hz, 1H), 5.12 (d, J = 3.7 Hz, 1H), 4.87 (d, J = 11.6 Hz, 1H), 4.78 (d, J = 11.5 Hz, 1H), 4.70 (d, J = 11.6 Hz, 1H), 4.66 (d, J = 11.7 Hz, 1H), 4.64 (d, J = 12.1 Hz, 1H), 4.63 (d, J = 3.6 Hz, 1H), 4.49 (d, J = 11.9 Hz, 1H), 4.42 (d, J = 12.2 Hz, 1H), 4.39 – 4.38 (m, 1H), 4.35 (d, J = 11.8 Hz, 1H), 4.09 – 3.95 (m, 5H), 3.88 – 3.82 (m, 2H), 3.75 (m, 1H), 3.52 (m, 1H), 3.40 (m, 1H), 1.40 (s, 3H), 1.35 (s, 3H), 1.17 (s, 3H), 1.12 (s, 3H).

20 β : ¹H NMR (500 MHz, CDCl₃) δ 7.37 – 7.16 (m, 20H), 5.68 (d, J = 3.8 Hz, 1H), 4.95 (d, J = 11.9 Hz, 1H), 4.83 (d, J = 11.6 Hz, 1H), 4.72 – 4.68 (m, 2H), 4.66 (d, J = 11.9 Hz, 1H), 4.55 (d, J = 12.1 Hz, 1H), 4.47 (d, J = 10.9 Hz, 1H), 4.46 – 4.44 (m, 2H), 4.43 (d, J = 7.9 Hz, 1H), 4.39 – 4.30 (m, 5H), 4.00 – 3.92 (m, 3H), 3.87 – 3.75 (m, 2H), 3.65 (dd, J = 9.7, 7.6 Hz, 1H), 1.35 (s, 3H), 1.32 (s, 3H), 1.14 (s, 3H), 1.09 (s, 3H).

The NMR values in accordance with the literature. ^[16]

References:

- [1] J. Ohtaka, T. Sakamoto, Y. Kikugawa, *Tetrahedron Lett.* **2009**, *50*, 1681 - 1683.
- [2] Y. Sato, K. Koguro, S. Mitsui, Patent WO, 2005/035484.
- [3] M. Kreuzer, J. Thiem, *Carbohydr. Res.* **1986**, *149*, 347 - 361.
- [4] Y. Ito, T. Ogawa, M. Numata, M. Sugimoto, *Carbohydr. Res.* **1990**, *202*, 165 - 175.
- [5] M. Koshiha, N. Suzuki, R. Arihara, T. Tsuda, H. Nambu, S. Nakamura, S. Hashimoto, *Chem. Asian. J.* **2008**, *3*, 1664 – 1677.
- [6] R. R. Schmidt, J. J. Michel, *Carbohydr. Chem.* **1985**, *4*, 141 – 169.
- [7] P. Weng, D. Crich, *Org. Lett.* **2017**, *19*, 2402 – 2405.
- [8] A. Marra, F. Gauffeny, P. Sinay, *Tetrahedron* **1991**, *47*, 5149 – 5150.
- [9] T. Tsuda, S. Nakamura, S. Hashimoto, *Tetrahedron* **2004**, *60*, 10711 – 10737.
- [10] H. Nagai, K. Sasaki, S. Matsumura, K. Toshima, *Carbohydr. Res.* **2005**, *340*, 337 - 353.
- [11] K. S. Kim, J. H. Kim, Y. J. Lee, Y. J. Lee, J. Park, *J. Am. Chem. Soc.* **2001**, *123*, 8477 – 8481.
- [12] S. Hotha, S. Kashyap, *J. Am. Chem. Soc.* **2006**, *128*, 9620 – 9621.
- [13] Y. Zhang, P. Wang, N. Song, M. Li, *Carbohydr. Res.* **2013**, *381*, 101-111.
- [14] D. Crich, and O. Vinogradova, *J. Am. Chem. Soc.* **2007**, *129*, 11756 - 11765.
- [15] H. M. Nguyen, Y. Chen, S. G. Duron, D. Y. Gin, *J. Am. Chem. Soc.* **2001**, *123*, 8766 – 8772.
- [16] E. J. Grayson, S. J. Ward, A. L. Hall, P. M. Rendle, D. P. Gamblin, A. S. Batsanov, B. G. Davis, *J. Org. Chem.* **2005**, *70*, 9740 – 9754.

NMR spectra

