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Supplementary Information

Key entity of DCAR agonist, phosphatidylinositol mannoside Ac₁PIM₁; Synthesis and its immunomodulatory function

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- **Figure S1**. TLR2-specific NF-κB induction by synthesized compounds in A) HEK-Blue mouse TLR2 cells and B) humanTLR2 cells.
- Figure S2. Antigen presenting cell (APC)-free assay for compounds binding to mCD1d.
- **Figure S3.** Binding assay based on AlphaScreenTM, applied to the synthesized compounds **1**, and **2a**–**c** in comparison with the known ligands.
- **Figure S4**. Proinflammatory cytokine inductions by synthesized compounds in BMDCs. (A) MCP-1 induction. (B) TNF-α induction.

Experimental Section

- 1. Synthesis
- 2. Biology
- 3. NMR spectra

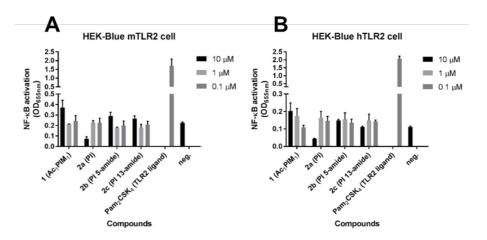


Figure S1. TLR2-specific NF-κB induction by synthesized compounds in A) HEK-Blue mouse and B) human TLR2 cells.

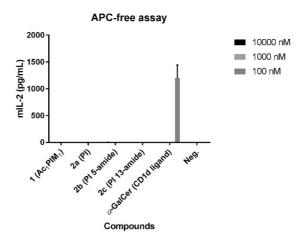


Figure S2. Antigen presenting cell (APC)-free assay^{1, 2} for compounds binding to mCD1d.

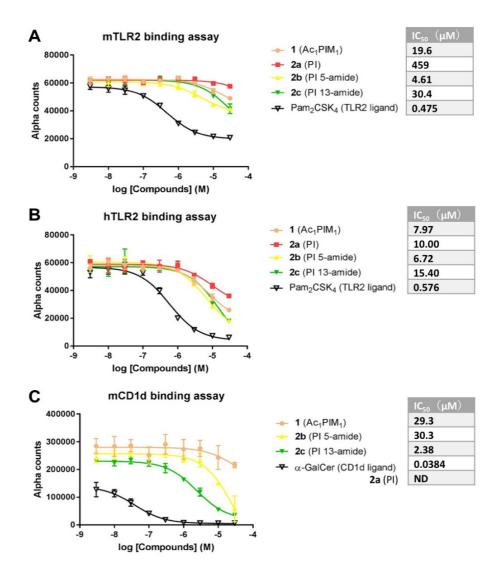


Figure S3. Binding assay based on AlphaScreenTM, applied to the synthesized compounds **1**, and **2a–c** in comparison with the known ligands.^{3,4}

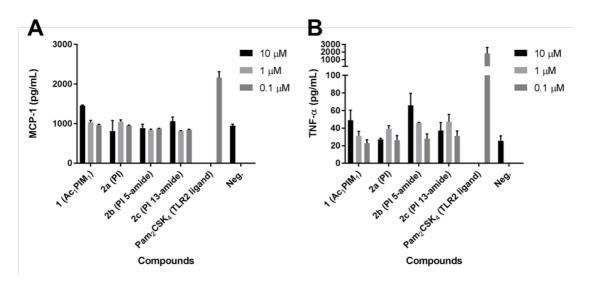


Figure S4. Proinflammatory cytokine inductions by synthesized compounds in BMDCs. (A) MCP-1 induction. (B) TNF- α induction.

1. Synthesis

General procedure:

Nuclear magnetic resonance (¹H NMR, ¹³C NMR, ³¹P NMR) spectra were measured at 25 °C in an indicated solvent with either JEOL ECX 400, ECS 400, JNM-ECZ800R or Bruker AVANCE 800US and analyzed Delta 5.0.5 (JEOL). The proton chemical shifts in CDCl₃ or in mixture solvent (CDCl₃, CD₃OD and D₂O) are reported in parts per million (δ) from tetramethylsilane as an internal standard and coupling constants are in Heltz (Hz). The chemical shifts in other solvent are reported in ppm from the residual proton signal of solvent. The chemical shits for ¹³C NMR are reported in ppm from the internal solvent signal (CDCl₃, δ 77.0). The chemical shits for ³¹P NMR are reported in ppm from the external standard signal (H₃PO₄, δ 0.0). High-resolution mass spectra (HRMS) of synthetic compounds were obtained on an electron spray ionization quadrupole time of flight (ESI-QTOF) mass spectrometer (micrOTOF-QII-HC; BRUKER). Analytical thin layer chromatography (TLC) was performed on Silica gel 60 F₂₅₄ Plates (Merck, 0.25 mm thickness). Silica gel column chromatography was performed using Silica gel 60 N [spherical neutral (Kanto Chemical Co., Inc., 40 – 50 μm)] at medium pressure (2 - 4 kgcm⁻²) using indicated solvent systems. Reagents were purchased from commercial supplier (TCI, nacalai tesque, Wako pure chemical industry, Ltd., Kanto Chemical Co., Inc., Watanabe Chemical ndustries, Ltd.) and were used without further purification. Unless otherwise noted, Non-aqueous reactions were carried out under argon atmosphere. Anhydrous dichloromethane, tetrahydrofuran, N, N-dimethylformamide, methanol and toluene were purchased from Kanto Chemical Co., Inc. Chiral HPLC was performed on a Prominence system (Shimadzu Corporation). Chiral HPLC was carried out by using a CHIRALPAK® AD-H packed column (5 µm, 4.6 x 250 mm) at a flow rate of 1 ml/min.

Synthesis of phosphoryl reagent 9b

To a solution of PCl₃ (148 μ L, 1.70 mmol) and Et₃N (477 μ L, 3.40 mmol) in toluene (3 mL) at 0 °C was added (R)-(+)-5,5',6,6',7,7',8,8'-octahydro-1,1'-bi-2-naphthol (500 mg, 1.70 mmol) under Ar. After stirring for 3 min, to the resulting solution was added Se (148 mg, 1.87 mmol). The mixture was warmed to 110 °C and stirred for 6 h. The reaction was concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with n-hexane-CH₂Cl₂ (3:2) to give **9b** (335 mg, 45%) as a pink solid.

¹H-NMR (400 MHz, CDCl₃) δ : 7.18 (d, J = 4.3 Hz, 1H), 7.16 (d, J = 4.1 Hz, 1H), 7.12 (dd, J = 8.4, 1.8 Hz, 1H), 7.06 (dd, J = 8.4, 2.3 Hz, 1H), 2.89-2.78 (m, 4H), 2.73-2.64 (m, 2H), 2.33-2.25 (m, 2H), 1.85-1.74 (m, 6H), 1.65-1.54 (m, 2H).

¹³C-NMR (100 MHz, CDCl₃) δ: 147.17, 145.36, 138.90, 138.78, 136.85, 136.51, 130.21, 129.98, 126.66, 126.47, 119.12, 118.26, 29.19 (2C), 27.83 (2C), 22.36, 22.34, 22.22, 22.17.

³¹P-NMR (160 MHz, CDCl₃) δ: 71.16.

HRMS (ESI-TOF) calcd for C₂₀H₂₀ClNaO₂PSe [M+Na]⁺ 460.9945, found 460.9944.

Synthesis of phosphoryl reagent 9c

The title compound was synthesized following the procedure for **9b**. (*R*)-3,3'-dibromo-1,1'-bi-2-naphthol (500 mg, 1.13 mmol) was used as starting materials. **9c** (458 mg, 69%) was obtained as a pink solid.

¹H-NMR (400 MHz, CDCl₃) δ : 8.36 (s, 2H), 7.89 (d, J = 8.3 Hz, 2H), 7.54 (t, J = 7.5 Hz, 2H), 7.33 (t, J = 7.6 Hz, 2H), 7.21 (d, J = 8.5 Hz, 2H).

¹³C-NMR (100 MHz, CDCl₃) δ: 134.47, 134.30, 132.40, 132.23, 131.41, 131.30, 129.01, 128.20, 127.66, 127.65, 127.41, 127.37, 127.27, 127.00, 126.93, 125.27, 124.04, 123.83, 114.26, 113.60. ³¹P-NMR (160 MHz, CDCl₃) δ: 64.35.

HRMS (ESI-TOF) calcd for C₂₀H₁₀Br₂ClNaO₂PSe [M+Na]⁺ 608.7530, found 608.7524.

Synthesis of phosphoryl reagent 9d

$$\begin{array}{c} \text{Br} \\ \text{HO} \\ \text{HO} \\ \text{HO} \\ \\ \text{Se, toluene} \end{array} \begin{array}{c} \text{Br} \\ \text{Se} \\ \text{II} \\ \text{O} \\ \text{Br} \\ \end{array}$$

The title compound was synthesized following the procedure for **9b**. (*R*)-(+)-3,3'-dibromo-5,5',6,6',7,7',8,8'-octahydro-1,1'-bi-2-naphthol (500 mg, 1.106 mmol) was used as starting materials. Compound **9d** (584 mg, 89%) was obtained as a pink solid.

¹**H-NMR (400 MHz, CDCl₃)** δ: 7.43 (s, 2H), 2.84-2.79 (m, 4H), 2.59-2.50 (m, 2H), 2.25-2.16 (m, 2H), 1.82-1.75 (m, 6H), 1.63-1.54 (m, 2H).

¹³C-NMR (100 MHz, CDCl₃) δ: 138.03 (2C), 137.86 (2C), 134.17 (2C), 133.90 (2C), 112.40, 112.36, 111.61 (2C), 28.96 (2C), 27.59, 27.55, 22.14, 22.11, 22.02, 21.99.

³¹P-NMR (160 MHz, CDCl₃) δ: 65.01.

HRMS (ESI-TOF) calcd for C₂₀H₁₈Br₂ClNaO₂PSe [M+Na]⁺ 616.8156, found 616.8153.

Synthesis of inositol phosphate derivative 11b

To a stirred solution of **8** (100 mg, 215 μ mol) and DMAP (52.6 mg, 431 μ mol) in CH₂Cl₂ (3 mL) at 0 °C was added **9b** (189 mg, 431 μ mol). After stirring for overnight at the same temperature, mCPBA (70%) (106 mg, 431 μ mol) was added to the reaction mixture. The reaction was extracted with EtOAc

and the extract was washed with H_2O and brine. The organic phase was dried over Na_2SO_4 , filtered and concentrated *in vacuo*. The crude mixture was purified by column chromatography of SiO_2 with toluene-EtOAc (2:1 = in vol. ratio) to afford the mixture of **10b** (1-P) and (3-P) (32.5 mg, 19%) as a white foam.

To a solution of AllocOBt (83.8 mg, 383 μmol) and DMAP (46.7 mg, 383 μmol) in CH₂Cl₂ (1 mL) was added **10b** (**1-P**) and (**3-P**) (32.5 mg, 40.5 μmol). The reaction mixture was stirred for overnight and quenched by an addition of 10% aqueous citric acid. The reaction was extracted with CH₂Cl₂ and washed with brine and dried over Na₂SO₄, filtered and concentrated *in vacuo*. Part of the mixture was analyzed by HPLC [COSMOSIL 5SL-II Packed column, 254 nm, linear gradient of 5–20% (v/v) of IPA in *n*-hexane over 15 min, 1 mL/min] to determine ratio of **11b** (**1-P**):(**3-P**) (77:23). The crude mixture was purified by column chromatography of SiO₂ with toluene-EtOAc (6:1) to give the **11b** (**1-P**) (28.4 mg, 72%) as a white solid.

¹H-NMR (400 MHz, CDCl₃) δ: 7.17 (d, J = 8.5 Hz, 2H), 7.12-7.05 (m, 4H), 7.00-6.85 (m, 2H), 6.81 (d, J = 8.5 Hz, 2H), 6.76 (d, J = 8.5 Hz, 2H), 5.98-5.79 (m, 2H), 5.37 (dd, J = 17.3, 1.3 Hz, 1H), 5.31 (dd, J = 17.3, 1.2 Hz, 1H), 5.28 (dd, J = 10.3, 1.1 Hz, 1H), 5.21 (dd, J = 10.3, 1.1 Hz, 1H), 4.92-4.76 (m, 2H), 4.71 (d, J = 6.5 Hz, 1H), 4.65-4.49 (m, 9H), 4.39 (d, J = 10.5 Hz, 1H), 4.24 (d, J = 10.5 Hz, 1H), 3.97 (t, J = 10.0 Hz, 1H), 3.91 (t, J = 9.8 Hz, 1H), 3.78 (s, 3H), 3.77 (s, 3H), 3.45 (s, 3H), 2.85-2.58 (m, 6H), 2.27-2.20 (m, 2H), 1.81-1.68 (m, 6H), 1.56-1.46 (m, 2H).

¹³C-NMR (100 MHz, CDCl₃) δ: 159.16 (2C), 154.05, 153.92, 146.82 (2C), 138.46, 136.01, 131.40, 131.36, 130.39, 130.21, 130.01, 129.72, 129.43 (2C), 129.35 (2C), 125.72 (2C), 119.12, 119.02, 113.59 (4C), 113.45 (2C), 98.02, 77.98, 75.85, 75.66, 74.93, 74.80, 74.52, 68.81 (2C), 68.69 (2C), 56.11, 55.22 (2C), 29.06, 29.04, 27.84, 27.79, 22.38, 22.33, 22.25, 22.23.

³¹P-NMR (160 MHz, CDCl₃) δ : -0.83.

HRMS (ESI-TOF) calcd for $C_{52}H_{59}NaO_{16}P [M+Na]^+ 993.3433$, found 993.3442.

Synthesis of inositol phosphate derivative 11c

The title compound was synthesized following the procedure for **11b**. **8** (100 mg, 215 µmol) and **9c** (253 mg, 431 µmol) were used as starting materials. Compound **10c (1-P)** and **(3-P)** (53.0 mg, 26%) was obtained as a white form. The ratio of **11c (1-P)**:**(3-P)** (81:19) was analyzed by HPLC [COSMOSIL 5SL-II Packed column, 254 nm, linear gradient of 5–20% (v/v) of IPA in *n*-hexane over 15 min, 1 mL/min]. Compound **11c (1-P)** (51.6 mg, 83%) was obtained as a white solid. **1H-NMR (400 MHz, CDCl₃)** δ : 8.31 (s, 1H), 8.26 (s, 1H), 7.86 (d, J = 8.1 Hz, 1H), 7.78 (d, J = 8.3 Hz, 1H), 7.49 (t, J = 7.4 Hz, 2H), 7.35-7.10 (m, 6H), 6.83-6.79 (m, 2H), 6.59 (d, J = 8.5 Hz, 2H), 6.42 (d, J = 8.5 Hz, 2H), 6.00-5.73 (m, 2H), 5.39 (dd, J = 17.3, 1.3 Hz, 1H), 5.33-5.23 (m, 2H), 5.18 (dd, J = 10.3, 1.1 Hz, 1H), 4.96 (d, J = 6.7 Hz, 1H), 4.89-4.84 (m, 2H), 4.76 (d, J = 6.5 Hz, 1H), 4.74-4.61 (m, 4H), 4.56-4.37 (m, 4H), 4.29 (d, J = 11.0 Hz, 1H), 4.19 (d, J = 11.0 Hz, 1H), 4.09-3.96 (m, 2H), 3.77 (s, 3H), 3.65 (s, 3H), 3.40 (s, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ: 159.15, 158.65, 154.01, 153.96, 143.86, 143.00, 134.44, 133.92, 132.30, 131.99, 131.43, 131.39, 131.28, 131.15, 130.01, 129.86, 129.44 (2C), 128.77 (2C), 127.68, 127.62, 127.37, 127.22, 127.14, 126.98, 126.95, 126.77, 122.78, 122.71, 119.06, 118.90, 114.41, 113.73, 113.58 (2C), 113.16 (2C), 98.37, 79.85, 77.87, 77.78, 76.59, 75.80, 75.04, 74.84, 74.64, 68.83, 68.64, 56.16, 55.21, 55.08.

³¹P-NMR (160 MHz, CDCl₃) δ : 0.19.

HRMS (ESI-TOF) calcd for C₅₂H₄₉Br₂NaO₁₆P [M+Na]⁺ 1141.1017, found 1141.1009.

Synthesis of inositol phosphate derivative 11d

The title compound was synthesized following the procedure for **11b**. **8** (100 mg, 215 µmol) and **9d** (257 mg, 431 µmol) were used as starting materials. Compound **10d** (**1-P**) and (**3-P**) (55.7 mg, 27%) was obtained as a white form. The ratio of **11d** (**1-P**):(**3-P**) (>99:1) was analyzed by HPLC [COSMOSIL 5SL-II Packed column, 254 nm, linear gradient of 5–20% (v/v) of IPA in *n*-hexane over 15 min, 1 mL/min]. Compound **11d** (**1-P**) (45.6 mg, 70%) was obtained as a white solid. **1H-NMR** (**400 MHz, CDCl₃)** δ : 7.40 (s, 1H), 7.32 (s, 1H), 7.23-7.14 (m, 3H), 6.97 (d, J = 8.5 Hz, 1H), 6.82-6.72 (m, 4H), 5.98-5.74 (m, 2H), 5.39-5.23 (m, 3H), 5.19 (d, J = 10.5 Hz, 1H), 4.95-4.76 (m, 4H), 4.68-4.34 (m, 9H), 4.29 (d, J = 11.2 Hz, 1H), 4.08-3.95 (m, 2H), 3.77 (s, 3H), 3.75 (s, 3H), 3.41 (s, 3H), 2.82-2.68 (m, 4H), 2.58-2.44 (m, 2H), 2.23-2.07 (m, 2H), 1.80-1.66 (m, 6H), 1.63-1.39 (m, 2H).

¹³C-NMR (100 MHz, CDCl₃) δ: 159.09, 158.86, 153.98, 153.89, 143.47, 142.17, 137.78, 137.51, 134.02, 133.42, 131.40 (2C), 130.33, 130.01, 129.37 (2C), 129.11 (2C), 127.28, 127.10, 125.23, 120.27, 118.96, 118.76, 113.53 (2C), 113.36 (2C), 112.30, 111.38, 98.41, 79.74, 77.94, 77.58, 76.49, 75.69, 74.77, 74.64, 68.74, 68.65, 68.54, 56.08, 55.69, 55.18, 28.83, 28.71, 27.55 (2C), 22.09 (2C), 21.96 (2C).

³¹P-NMR (160 MHz, CDCl₃) δ: -2.49.

HRMS (ESI-TOF) calcd for $C_{52}H_{57}Br_2NaO_{16}P[M+Na]^+$ 1149.1643, found 1149.1647.

Scheme S1. Synthesis of inositol phosphate 29

Synthesis of inositol phosphate derivative S1b

The solution of **11b** (16.0 mg, 16.5 μ mol), and [CpRu(C₃H₅)(C₉H₆NCOO)]PF₆ (173 μ g, 0.33 μ mol) in MeOH (1 mL) was stirred at 35 °C. After stirring for 3 h, the reaction mixture was concentrated under reduced pressure. The crude residue was purified by column chromatography of SiO₂ with toluene-EtOAc (1:1) to give the **S1b** (16.6 mg, quant.) as a colorless oil.

¹**H-NMR (400 MHz, CDCl₃)** δ : 7.37-7.28 (m, 2H), 7.18-7.00 (m, 6H), 6.87 (d, J = 8.3 Hz, 2H), 6.78 (d, J = 8.1 Hz, 2H), 4.92-4.73 (m, 4H), 4.68-4.33 (m, 3H), 4.28 (d, J = 10.5 Hz, 1H), 3.79-3.71 (m,

8H), 3.57-3.42 (m, 5H), 3.30-3.22 (m, 2H), 2.89-2.57 (m, 6H), 2.27-2.17 (m, 2H), 1.83-1.63 (m, 6H), 1.56-1.40 (m, 2H).

¹³C-NMR (100 MHz, CDCl₃) δ: 159.30, 159.18, 146.93, 146.81, 138.43 (2C), 136.05, 135.66, 130.64, 130.46, 129.76 (2C), 129.70, 129.55 (2C), 129.36, 125.82, 125.78, 118.44, 117.75, 113.87 (2C), 113.75 (2C), 98.78, 80.68, 80.49, 79.80, 79.24, 77.20, 74.74, 74.53, 74.17, 70.84, 70.81 (2C), 56.13, 55.74, 55.25, 29.68, 29.07, 29.03, 27.82, 22.38, 22.30, 22.24, 22.22.

HRMS (ESI-TOF) calcd for C₄₄H₅₁NaO₁₂P [M+Na]⁺ 825.3010, found 825.3011.

Synthesis of inositol phosphate derivative S1c

$$\begin{array}{c} & & & & & & & & & \\ & & & & & & & \\ & & & & & & \\ & & & & & \\ & & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\$$

The title compound was synthesized following the procedure for **S1b**. **11c** (32.5 mg, 29.0 µmol) was used as starting materials. Compound **S1c** (27.4 mg, 99%) was obtained as a colorless oil.

¹H-NMR (400 MHz, CDCl₃) δ: 8.33 (s, 1H), 8.25 (s, 1H), 7.87 (d, J = 8.3 Hz, 1H), 7.78 (d, J = 8.3 Hz, 1H), 7.52 (d, J = 7.4 Hz, 1H), 7.48 (d, J = 7.4 Hz, 1H), 7.38-7.14 (m, 6H), 6.87 (d, J = 8.5 Hz, 2H), 6.71 (d, J = 8.5 Hz, 2H), 6.43 (d, J = 8.5 Hz, 2H), 4.93 (dd, J = 11.7, 6.5 Hz, 2H), 4.86 (d, J = 11.2 Hz, 1H), 4.74 (d, J = 11.0 Hz, 1H), 4.68-4.60 (m, 2H), 4.41 (d, J = 10.8 Hz, 1H), 4.33 (d, J = 11.0 Hz, 1H), 3.84 (t, J = 9.2 Hz, 1H), 3.79 (s, 3H), 3.65 (s, 3H), 3.56-3.47 (m, 3H), 3.43 (s, 3H), 2.36-2.34 (m, 2H).

¹³C-NMR (100 MHz, CDCl₃) δ: 159.26, 158.85, 143.99, 143.05, 134.41, 133.89, 132.28, 131.99, 131.24, 131.17, 130.69, 130.19, 129.74, 129.16, 129.01, 128.99, 128.20, 127.67, 127.62, 127.35, 127.26, 127.09, 126.97, 126.80, 125.27, 122.82, 114.34, 113.84, 113.79, 113.75, 113.71, 113.48, 99.06, 81.33, 80.77, 80.29, 80.04, 74.96, 74.42, 74.11, 70.85, 56.13, 55.25, 55.10.

HRMS (ESI-TOF) calcd for $C_{44}H_{41}Br_2NaO_{12}P[M+Na]^+$ 973.0595, found 973.0587.

Synthesis of inositol phosphate derivative S1d

The title compound was synthesized following the procedure for **S1b**. **11d** (33.2 mg, 29.4 µmol) was used as starting materials. Compound **S1d** (29.4 mg, 96%) was obtained as a colorless oil.

¹H-NMR (400 MHz, CDCl₃) δ : 7.41 (s, 1H), 7.34-7.29 (m, 3H), 7.28-7.14 (m, 2H), 7.06 (d, J = 8.8 Hz, 1H), 6.90-6.84 (m, 2H), 6.77 (d, J = 8.8 Hz, 1H), 4.90 (s, 2H), 4.82 (t, J = 10.7 Hz, 1H), 4.76 (t, J = 11.0 Hz, 1H), 4.63-4.52 (m, 2H), 4.48-4.38 (m, 1H), 4.22 (d, J = 11.2 Hz, 1H), 3.85-3.80 (m, 2H), 3.79 (s, 3H), 3.76 (s, 3H), 3.60-3.51 (m, 3H), 3.42 (s, 3H), 3.24 (s, 1H), 2.84-2.69 (m, 3H), 2.58-2.47 (m, 3H), 2.22-2.08 (m, 2H), 1.81-1.39 (m, 8H).

¹³C-NMR (100 MHz, CDCl₃) δ: 159.23, 159.15, 137.77, 137.56, 137.12, 134.01, 133.43, 130.75, 130.63, 129.73, 129.71, 129.34, 129.32, 129.02, 128.21, 127.31, 127.26, 125.27, 113.84, 113.81, 113.78, 112.21, 111.48, 99.11, 81.23, 80.62, 80.22, 80.16, 74.67, 74.38, 73.89, 70.74, 56.10, 55.25, 28.86, 28.76, 27.57, 22.12, 22.01.

HRMS (ESI-TOF) calcd for $C_{44}H_{49}Br_2NaO_{12}P$ [M+Na]⁺ 981.1221, found 981.1215.

Synthesis of inositol phosphate derivative S2b

To a solution of **S1b** (16.6 mg, 20.7 μ mol) in DMF (1 mL) at room temperature were added TBSCl (31.2 mg, 207 μ mol) and imidazole (16.9 mg, 248 μ mol). The mixture was warmed up to 60 °C. After stirring for 1 d, methanol was added to the reaction, and then extracted with Et₂O and washed with brine. The organic phase was dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with toluene-EtOAc (10:1) to give **S2b** (2.73 mg, 11%)

as a colorless oil.

¹H-NMR (400 MHz, CDCl₃) δ: 7.42-7.36 (m, 2H), 7.19 (d, J = 8.5 Hz, 2H), 7.13 (d, J = 8.5 Hz, 2H), 7.04-6.97 (m, 2H), 6.82 (d, J = 8.8 Hz, 2H), 6.71 (d, J = 8.5 Hz, 2H), 5.39-5.30 (m, 2H), 5.04-4.82 (m, 3H), 4.75-4.61 (m, 1H), 4.44-4.28 (m, 3H), 3.79 (s, 3H), 3.74 (s, 3H), 3.64 (t, J = 9.0 Hz, 2H), 3.50 (s, 3H), 3.43 (t, J = 8.9 Hz, 1H), 2.96-2.51 (m, 4H), 2.06-1.94 (m, 2H), 1.77-1.60 (m, 10H), 0.82 (s, 9H), 0.78 (s, 9H), 0.10 (s, 3H), -0.03 (s, 3H), -0.17 (s, 3H), -0.18 (s, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ: 158.49, 158.13, 146.80 (2C), 138.21, 138.14, 131.28, 130.96, 130.22, 130.01, 129.56, 128.84 (3C), 127.37 (3C), 125.85, 118.44, 118.40, 113.18 (2C), 113.07 (2C), 97.90, 81.59, 80.06, 77.84, 77.20, 74.80, 74.66, 74.61, 73.20, 56.06, 55.20, 55.13, 31.92, 29.68, 29.51, 29.31, 29.06, 27.77, 25.95 (2C), 25.89 (2C), 22.68, 22.41, 22.25, 22.10, -0.01, -3.98, -4.33, -4.90.

HRMS (ESI-TOF) calcd for C₅₆H₇₉NaO₁₂PSi₂ [M+Na]⁺ 1053.4740, found 1053.4736.

Synthesis of inositol phosphate derivative S2c

The title compound was synthesized following the procedure for **S2b**. **S1c** (27.4 mg, 28.8 μmol) was used as starting materials. Compound **S2c** (7.46 mg, 22%) was obtained as a colorless oil.

¹H-NMR (400 MHz, CDCl₃) δ: 8.23 (s, 1H), 8.14 (s, 1H), 7.80 (d, J = 7.7 Hz, 1H), 7.62 (d, J = 7.7 Hz, 1H), 7.43 (t, J = 7.0 Hz, 1H), 7.28-7.23 (m, 8H), 7.13 (t, J = 7.0 Hz, 1H), 6.94 (d, J = 7.7 Hz, 2H), 6.85 (d, J = 8.2 Hz, 2H), 4.72-4.69 (m, 1H), 4.58-4.45 (m, 4H), 4.34-4.28 (m, 3H), 3.89-3.83 (m, 4H), 3.80 (s, 3H), 3.75-3.70 (m, 1H), 3.56-3.50 (m, 1H), 3.41 (s, 3H), 3.28-3.21 (m, 1H), 0.74 (s, 9H), 0.70 (s, 9H), -0.09 (s, 3H), -0.17 (s, 3H), -0.21 (s, 3H), -0.81 (s, 3H).

¹³C-NMR (100 MHz, CDCl₃) 8: 159.47, 159.30, 148.73, 147.63, 133.81, 132.81, 132.60, 129.82 (3C), 129.67 (3C), 129.46, 129.28, 127.11, 126.99, 126.82, 126.78, 126.68, 126.48, 126.10, 125.98, 124.87, 122.98, 122.25, 116.78, 113.91 (2C), 113.77 (2C), 111.37, 95.01, 81.79, 80.97, 80.79, 79.85, 75.38, 72.14, 71.49, 70.94, 55.90, 55.35, 55.26, 26.11 (3C), 25.76 (3C), 18.69, 18.00, -3.12, -3.72, -4.78, -

HRMS (ESI-TOF) calcd for $C_{56}H_{69}Br_2NaO_{12}PSI_2$ [M+Na]⁺ 1201.2324, found 1201.2325.

Synthesis of inositol phosphate derivative S2d

The title compound was synthesized following the procedure for **S2b**. **S1d** (29.4 mg, 30.6 μmol) was used as starting materials. Compound **S2d** (10.01 mg, 28%) was obtained as a colorless oil.

¹**H-NMR (400 MHz, CDCl₃)** δ: 7.40 (s, 1H), 7.23-7.11 (m, 3H), 6.96 (d, J = 7.9 Hz, 2H), 6.83 (d, J = 7.7 Hz, 2H), 6.64 (d, J = 8.4 Hz, 2H), 5.06-4.93 (m, 2H), 4.87 (t, J = 10.5 Hz, 1H), 4.69 (t, J = 11.2 Hz, 1H), 4.56 (d, J = 8.6 Hz, 2H), 4.30 (dt, J = 37.5, 12.5 Hz, 2H), 3.87-3.78 (m, 5H), 3.74 (s, 3H), 3.66-3.56 (m, 2H), 3.50 (s, 3H), 2.84-1.68 (m, 16H), 0.85 (s, 9H), 0.75 (s, 9H), 0.14 (s, 3H), 0.00 (s, 3H), -0.18 (s, 3H), -0.27 (s, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ: 158.26, 158.10, 137.62, 137.35 (2C), 136.82, 133.79 (2C), 133.28 (2C), 131.32, 131.07, 128.45 (3C), 127.45 (3C), 113.24, 113.06 (2C), 113.00 (2C), 111.14, 98.35, 81.27, 80.44, 77.67, 77.20, 74.79, 74.55, 74.37, 73.17, 56.24, 55.13 (2C), 29.65, 28.88, 28.61, 27.52, 27.26, 26.01, 25.90 (6C), 22.16, 21.98, 18.07, 17.91, -4.05, -4.09, -4.40, -4.76.

HRMS (**ESI-TOF**) calcd for $C_{56}H_{77}Br_2NaO_{12}PSI_2 [M+Na]^+ 1209.2950$, found 1209.2955.

Synthesis of inositol phosphate derivative S3 from S2b

To a suspension of NaH (60% oil dispersion) (0.275 mg, 6.90 μ mol) in THF (0.5 mL) at 0 °C was added benzyl alcohol (0.785 μ L, 7.59 μ mol). After stirring for 30 min, the reaction was added to the solution of **S2b** (2.37 mg, 2.30 μ mol) in THF (0.5 mL) at 0 °C. The reaction mixture was warmed up to room temperature, and stirred for 1.5 h. The reaction was cooled to 0 °C, and then quenched with 10% aqueous citric acid. The mixture was extracted with ethyl acetate. The organic solution was washed with sat. NaHCO₃ aq. and brine, dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with toluene-EtOAc (15:1) to give the compound **S3** (1.40 mg, 64%) as a colorless oil.

¹H NMR spectra was consistent with literature data.⁵

Synthesis of inositol phosphate derivative S3 from S2c

The title compound was synthesized following the procedure for **S3** from **S2b**. **S2c** (5.95 mg, 5.04 µmol) was used as starting materials. Compound **S3** (2.31 mg, 48%) was obtained as a colorless oil. ¹H NMR spectra was consistent with literature data.⁵

Synthesis of inositol phosphate derivative S3 from S2d

The title compound was synthesized following the procedure for **S3** from **S2b**. **S2d** (4.61 mg, 3.88 μmol) was used as starting materials. Compound **S3** (1.96 mg, 53%) was obtained as a colorless oil. ¹H NMR spectra was consistent with literature data.⁵

Synthesis of mannose derivative S4

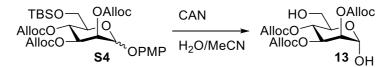
To a solution of 12^6 (3.37 g, 8.41 mmol), Et₃N (5.91 mL, 42.1 mmol) and DMAP (514 mg, 4.21 mmol) in CH₂Cl₂ (20 mL) was added AllocCl (4.47 mL, 42.1 mmol). The resulting mixture was stirred at room temperature for overnight. The reaction was quenched by an addition of sat. NaHCO₃ aq. and extracted with CHCl₃. Organic layer was washed with 10% citric acid aq. and brine and dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with *n*-hexane-EtOAc (5:1) to give S4 (3.06 g, 56%) as a colorless oil.

¹H-NMR (400 MHz, CDCl₃) δ : 7.04 (td, J = 6.3, 4.0 Hz, 2H), 6.81 (td, J = 6.3, 4.0 Hz, 2H), 5.98-5.85 (m, 3H), 5.47 (d, J = 1.8 Hz, 1H), 5.43-5.20 (m, 9H), 4.67-4.62 (m, 6H), 4.02 (ddd, J = 10.1, 5.2, 2.7 Hz, 1H), 3.80-3.71 (m, 5H), 0.87-0.85 (m, 9H), 0.02 (s, 3H), 0.01 (s, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ : 155.39, 154.22, 153.91, 149.87 (2C), 131.31, 131.22, 131.05, 119.19, 118.96, 118.89, 118.18 (2C), 114.59 (2C), 96.41, 73.15, 72.74, 71.65, 70.31, 69.06, 68.93, 68.81, 62.20, 55.61, 25.77 (3C), 18.19, -5.49 (2C).

HRMS (ESI-TOF) calcd for C₃₁H₄₄NaO₁₃Si [M+Na]⁺ 675.2443, found 675.2451.

Synthesis of mannose derivative 13



To a solution of **S4** (2.25 g, 3.45 mmol) in MeCN/H₂O (4:1) (30 mL) at 0 °C was added CAN (5.67 g, 10.3 mmol) and stirred for overnight. The reaction was poured into water and extracted with EtOAc. Organic layer was washed with brine and dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with toluene-EtOAc (1:1) to give **13** (640 mg, 43%) as a colorless oil.

¹**H-NMR (400 MHz, CDCl₃)** δ : 5.99-5.87 (m, 3H), 5.41-5.23 (m, 9H), 5.14 (t, J = 10.1 Hz, 1H), 4.66-4.62 (m, 6H), 4.09 (ddd, J = 9.9, 4.3, 2.5 Hz, 1H), 3.85 (d, J = 3.8 Hz, 1H), 3.80-3.70 (m, 2H), 2.64 (t, J = 6.5 Hz, 1H).

¹³C-NMR (100 MHz, CDCl₃) δ: 154.26 (2C), 153.79, 131.24, 131.02, 130.96, 119.44, 119.36, 118.96, 91.93, 73.19, 72.51, 70.43, 70.07, 69.17 (2C), 68.96, 61.23.

HRMS (ESI-TOF) calcd for $C_{18}H_{24}NaO_{12}$ [M+Na]⁺ 455.1160, found 455.1169.

Synthesis of mannose derivative S5

To a solution of **13** (1.70 g, 3.93 mmol) and TBSCl (652 mg, 4.33 mmol) in CH₂Cl₂ (50 mL) was added imidazole (293 mg, 4.33 mmol). The resulting mixture was stirred at room temperature for overnight. The reaction was quenched by an addition of water and extracted with EtOAc. Organic layer was washed with brine and dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with toluene-EtOAc (3:1) to give **S5** (1.69 g, 79%) as white solid.

¹H-NMR (400 MHz, CDCl₃) δ : 5.97-5.86 (m, 3H), 5.40-5.23 (m, 8H), 5.20 (dd, J = 3.3, 1.9 Hz, 1H), 5.10 (t, J = 10.1 Hz, 1H), 4.64-4.61 (m, 6H), 4.12-4.08 (m, 1H), 4.05 (d, J = 4.3 Hz, 1H), 3.84-3.72 (m, 2H), 0.90-0.87 (m, 9H), 0.07 (s, 3H), 0.06 (s, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ: 154.24, 153.97, 153.79, 131.31, 131.17, 131.10, 119.11, 119.01, 118.77, 91.76, 73.23, 72.93, 71.16, 70.52, 68.95, 68.83, 68.78, 62.72, 25.87 (3C), 18.41, -5.41, -5.49.

HRMS (ESI-TOF) calcd for C₂₄H₃₈NaO₁₂Si [M+Na]⁺ 569.2025, found 569.2033.

Synthesis of mannose derivative 5

To the solution of the $\bf S5$ (356 mg, 651 µmol) and ClC(NPh)CF₃ (87.9 µL, 542 µmol) in CH₂Cl₂ (5 mL) was added DMAP (66.2 mg, 542 µmol). After stirring for 6 h, the reaction mixture was purified by column chromatography of SiO₂ with toluene-EtOAc (9:1) to give $\bf 5$ (419 mg, quant.) as a colorless oil.

¹H-NMR (400 MHz, CDCl₃) δ: 7.34-7.28 (m, 2H), 7.14-7.10 (m, 1H), 6.83 (d, J = 7.6 Hz, 2H), 6.38 (s, 1H), 5.97-5.87 (m, 3H), 5.40-5.26 (m, 9H), 4.66-4.62 (m, 6H), 4.02-3.95 (m, 1H), 3.81 (d, J = 3.6 Hz, 2H), 0.89 (s, 9H), 0.06 (s, 3H), 0.05 (s, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ: 154.00, 153.78, 153.72, 142.89, 131.20, 131.11, 130.89, 128.80 (2C), 124.62, 119.31 (3C), 119.12, 118.97, 73.69, 72.82, 71.16, 69.47 (2C), 69.23 (2C), 69.04, 68.95 (2C), 61.96, 25.76, 18.21 (3C), -5.41, -5.45.

HRMS (**ESI-TOF**) calcd for C₃₂H₄₂F₃NNaO₁₂Si [M+Na]⁺ 740.2321, found 740.2327.

Synthesis of inositol phosphate derivative 15

To a suspension of NaH (60% oil dispersion) (229 mg, 5.71 mmol) in THF (15 mL) at 0 °C was added allyl alcohol (435 μ L, 6.40 mmol). After stirring for 30 min, the reaction was added to the solution of **14**⁷ (2.00 g, 2.29 mmol) in THF (35 mL) at 0 °C. The reaction mixture was warmed up to room temperature, and stirred for 1.5 h. The reaction was cooled to 0 °C, and then quenched with 10%

aqueous citric acid. The mixture was extracted with ethyl acetate. The organic solution was washed with sat. NaHCO₃ aq. and brine, dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with *n*-hexane-EtOAc (5:1) to give the compound **15** (1.10 g, 68%) as a colorless oil.

¹**H-NMR (400 MHz, CDCl₃)** δ : 7.32-7.26 (m, 4H), 6.85 (t, J = 8.6 Hz, 4H), 6.01-5.77 (m, 4H), 5.35-5.14 (m, 8H), 4.83-4.75 (m, 4H), 4.70 (t, J = 10.1 Hz, 2H), 4.55-4.44 (m, 4H), 4.36 (dd, J = 12.3, 5.4 Hz, 1H), 4.32-4.27 (m, 2H), 4.19 (dd, J = 12.6, 5.4 Hz, 1H), 4.15-4.11 (m, 1H), 4.07 (dd, J = 12.6, 5.4 Hz, 1H), 3.90 (t, J = 9.6 Hz, 1H), 3.84 (t, J = 9.5 Hz, 1H), 3.80 (s, 3H), 3.79 (s, 3H), 3.44 (s, 3H), 3.29-3.23 (m, 2H).

¹³C-NMR (100 MHz, CDCl₃) δ: 159.18, 159.04, 135.16, 134.57, 132.47, 132.39, 130.87, 130.69, 129.85 (2C), 129.49 (2C), 118.12, 118.00, 116.82, 116.39, 113.73 (2C), 113.57 (2C), 97.48, 82.70, 80.92, 79.58, 79.28, 77.83, 75.59, 75.25, 74.53, 73.73, 71.35, 68.20, 68.10, 55.81, 55.25, 55.23.

³¹P-NMR (160 MHz, CDCl₃) δ: -1.02.

HRMS (ESI-TOF) calcd for $C_{36}H_{49}NaO_{12}P$ [M+Na]⁺ 727.2854, found 727.2861.

Synthesis of inositol phosphate derivative 16

To a stirred mixture of **15** (900 mg, 1.28 mmol) in CH₂Cl₂ (34.5 mL) and pH 7 phosphate buffer (3.83 mL) was added DDQ (870 mg, 3.83 mmol). After stirring for 4 h, the reaction mixture was extracted with CHCl₃ and washed with sat. NaHCO₃ aq. and brine, dried over Na₂SO₄, filtrated and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with toluene-EtOAc (1:1) to give PMB group deprotected compound (560 mg, 94 %) as a colorless oil. To a PMB group deprotected compound (560 mg, 1.20 mmol) and AllocOBt (1.31 g, 6.00 mmol) in CH₂Cl₂ (15 mL) was added DMAP (733 mg, 6.00 mmol) and the mixture was stirred for 4 h. The reaction was quenched by addition of 10% citric acid aq. and extracted with CHCl₃. The organic layer was washed with brine, dried over Na₂SO₄, filtrated and concentrated *in vacuo*. The crude mixture was purified by column chromatography of SiO₂ with toluene-EtOAc (3:1) to give **16** (422 mg, 56 %) as

a colorless oil.

¹H-NMR (400 MHz, CDCl₃) δ: 5.98-5.74 (m, 6H), 5.40-5.15 (m, 13H), 5.12 (dd, J = 10.4, 1.5 Hz, 1H), 4.78 (s, 2H), 4.67-4.63 (m, 3H), 4.61 (dt, J = 5.8, 1.3 Hz, 1H), 4.58-4.50 (m, 4H), 4.37-4.31 (m, 2H), 4.19-4.11 (m, 3H), 3.94 (ddt, J = 12.9, 5.5, 1.4 Hz, 1H), 3.45 (t, J = 9.5 Hz, 1H), 3.40 (s, 3H), 3.34 (dd, J = 10.2, 2.1 Hz, 1H).

¹³C-NMR (100 MHz, CDCl₃) δ: 154.27, 154.21, 134.16, 133.80, 132.17, 132.09, 131.45, 131.33, 119.02, 118.94, 118.36, 118.25, 117.30, 117.06, 97.35, 78.08, 76.53, 76.17, 75.22, 73.62, 72.65, 72.62, 70.97, 68.73, 68.61, 68.38, 68.31, 55.92.

³¹P-NMR (160 MHz, CDCl₃) δ: -1.04.

HRMS (ESI-TOF) calcd for C₂₈H₄₁NaO₁₄P [M+Na]⁺ 655.2126, found 655.2135.

Synthesis of 17

The solution of **16** (775 mg, 1.23 mmol) in CH₂Cl₂ (15 mL) was cooled to 0 °C. To the solution was added TFA (15 mL) at 0 °C and stirred for 1h. The solvent removed under reduced pressure and added toluene then concentrated *in vacuo* to give the de-2-O-MOM compound (629 mg, 87%). The de-2-O-MOM compound (124 mg, 211 µmol) and **5** (313 mg, 435 µmol) were co-evaporated with toluene three times and dried highly under vacuum. The residue was dissolved with Et₂O (0.5 mL) and CH₂Cl₂ (0.5 mL) under Ar. To the mixture was added MS4A (100 mg) and then TMSOTf (19.1 µL, 106 µmol). The mixture was stirred at room temperature for 13 h. The reaction was quenched with Et₃N (400 µL) and then the mixture was concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with toluene-EtOAc (3:1) to give **17** (112 mg, 48%) as a colorless oil.

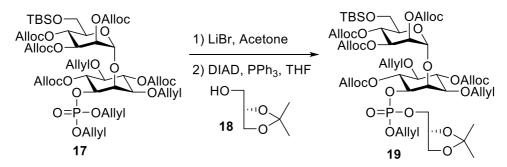
¹H-NMR (400 MHz, CDCl₃) δ: 6.00-5.74 (m, 9H), 5.38-5.10 (m, 22H), 4.67-4.58 (m, 12H), 4.54-4.50 (m, 4H), 4.45-4.40 (m, 2H), 4.27-4.23 (m, 1H), 4.20-4.15 (m, 1H), 4.14-4.12 (m, 2H), 3.96-3.90

(m, 1H), 3.73 (dd, J = 11.6, 3.7 Hz, 1H), 3.67 (dd, J = 11.4, 2.0 Hz, 1H), 3.48 (t, J = 9.5 Hz, 1H), 3.36 (dd, J = 10.1, 2.2 Hz, 1H), 0.88 (s, 9H), 0.03 (s, 3H), 0.01 (s, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ: 153.89, 153.85, 153.78, 153.73, 153.38, 134.11, 133.63, 132.20, 132.13, 132.02, 131.95, 131.46, 131.39, 131.21, 128.97, 128.17, 125.24, 118.99, 118.69, 118.48, 118.23, 117.49, 117.14, 98.36, 77.72, 76.41, 75.96, 75.69, 75.46, 74.01, 73.49, 73.30, 72.59, 71.46, 70.98, 69.58, 68.76, 68.73, 68.71, 68.69, 68.65, 68.56, 68.43, 61.72, 25.81, 18.25 (3C), -5.51, -5.56. ³¹P-NMR (160 MHz, CDCl₃) δ: -1.50.

HRMS (ESI-TOF) calcd for C₅₀H₇₃NaO₂₄PSi [M+Na]⁺ 1139.3891, found 1139.3888.

Synthesis of 19



To the solution of 17 (127 mg, 114 μ mol) in acetone (4 mL) was added LiBr (29.7 mg, 342 μ mol) and refluxed for overnight. After the removal of solvent, the crude mixture was purified by column chromatography of SiO₂ with toluene:EtOAc (5:1) to CHCl₃:MeOH (6:1) to give lithium salt. The desire product was diluted with EtOAc and washed with sat. KHSO₄ aq. and brine. The organic layer was dried over Na₂SO₄, filtrated and concentrated *in vacuo* to obtain proton form. The resulting phosphoric acid, (S)-(+)-2,2-Dimethyl-1,3-dioxolane-4-methanol 18 (42.2 μ L, 342 μ mol) and PPh₃ (89.7 mg, 342 μ mol) were dissolved with THF (1 mL) under Ar. To the mixture was added DIAD (62.5 μ L, 342 μ mol) and stirred for overnight. The reaction mixture was concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with toluene-EtOAc (3:1) to give 19 (69.9 mg, 51%) as white solid.

¹H-NMR (400 MHz, CDCl₃) δ : 6.00-5.74 (m, 8H), 5.39-5.33 (m, 7H), 5.30-5.21 (m, 11H), 5.20-5.11 (m, 4H), 4.74-4.53 (m, 12H), 4.46-4.38 (m, 2H), 4.29-3.90 (m, 9H), 3.83-3.71 (m, 2H), 3.67 (dd, J = 11.6, 1.9 Hz, 1H), 3.48 (td, J = 9.6, 2.9 Hz, 1H), 3.37-3.35 (m, 1H), 1.41 (s, 3H), 1.34-1.33 (m, 3H), 0.88 (s, 9H), 0.02 (s, 3H), 0.01 (s, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ: 153.97, 153.92, 153.82, 153.76, 153.43, 134.10, 133.65, 132.16, 132.00, 131.50, 131.45, 131.41, 131.20, 119.06, 119.04, 118.74, 118.72, 118.66, 118.43, 117.42, 117.19, 109.84, 98.63, 77.66, 76.39, 75.94, 75.61, 75.48, 74.66, 73.86, 73.51, 73.37, 72.62, 71.40, 71.01, 69.55, 68.86, 68.81, 68.74, 68.71, 68.58, 68.01, 65.97, 65.73, 61.64, 26.70, 25.82 (3C), 25.20, 18.26, -5.52, -5.58.

³¹P-NMR (160 MHz, CDCl₃) δ : -1.68.

HRMS (ESI-TOF) calcd for $C_{53}H_{79}Na_2O_{26}PSi [M+2Na]^{2+} 618.2075$, found 618.2076.

Synthesis of S6

To the solution of 19 (69.9 mg, 58.7 μ mol) in THF (6 mL) was added the mixture of pyridine (94.5 μ L, 1.17 mmol) and HF pyridine (70% HF) (145 μ L) and stirred for 2 d. After an addition of EtOAc, the mixture was quenched by addition of sat. NaHCO₃ aq. and extracted with EtOAc. The organic layer was dried over Na₂SO₄, filtrated and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with toluene-EtOAc (1:1) to give de-TBS compound (34.1 mg, 54%) as a colorless oil. The solution of de-TBS compound (34.1 mg, 31.7 μ mol) in CH₂Cl₂ (5 mL) was cooled to 0 °C. To the solution was added TFA (382 μ L) at 0 °C and stirred for 4 h. The reaction was quenched by addition of sat. NaHCO₃ aq. and extracted with AcOEt. The organic layer was washed with brine, dried over Na₂SO₄, filtrated and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with CHCl₃-MeOH (30:1) to give **S6** (30.0 mg, 91%) as a colorless oil.

¹H-NMR (400 MHz, CDCl₃) δ: 6.00-5.74 (m, 8H), 5.38-5.11 (m, 22H), 4.70-4.41 (m, 14H), 4.24-3.86 (m, 8H), 3.71-3.58 (m, 4H), 3.50 (t, J = 9.6 Hz, 1H), 3.39 (td, J = 10.2, 2.2 Hz, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 154.11, 154.04, 154.00, 153.92, 153.40, 134.01, 133.53, 131.95, 131.42, 131.29, 131.24, 131.01, 130.94, 119.42, 119.36, 119.22, 119.10, 119.01, 118.77, 117.71, 117.25, 97.91, 77.20, 76.89, 75.47, 73.77, 72.65, 72.57, 71.76, 71.10, 70.37, 70.33, 69.65, 69.23, 69.16, 69.09 (2C), 68.99, 68.95 (2C), 68.89, 68.76, 62.76, 60.94.

³¹P-NMR (160 MHz, CDCl₃) δ : -0.88.

HRMS (ESI-TOF) calcd for $C_{44}H_{61}Na_2O_{26}P [M+2Na]^{2+} 541.1487$, found 541.1493.

Synthesis of 3

To a solution of **S6** (13.6 mg, 13.1 μmol), palmitic acid (16.8 mg, 65.7 μmol) and EDC•HCl (12.6 mg, 65.7 μmol) in CH₂Cl₂ (1 mL) was added DMAP (8.02 mg, 65.7 μmol). The resulting mixture was stirred at room temperature for overnight. The reaction was quenched by an addition of sat. KHSO₄ aq. and extracted with CHCl₃. Organic layer was washed with brine and dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with toluene-EtOAc (10:1) to give **3** (11.7 mg, 51%) as white solid.

¹H-NMR (400 MHz, CDCl₃) δ : 5.99-5.75 (m, 8H), 5.39-5.11 (m, 23H), 4.66-4.07 (m, 24H), 3.98 (dd, J = 12.9, 5.3 Hz, 1H), 3.53-3.46 (m, 1H), 3.43-3.37 (m, 1H), 2.41-2.28 (m, 6H), 1.62-1.59 (m, 6H), 1.33-1.23 (m, 72H), 0.88 (t, J = 6.8 Hz, 9H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.55, 173.20, 172.86, 154.01, 153.89, 153.85, 153.33, 153.31, 134.13, 133.54, 132.11, 132.05, 131.45, 131.29, 131.22, 119.17, 119.13, 119.05, 118.86, 118.72, 118.61, 117.80, 117.26, 98.73, 77.62, 77.23, 76.36, 75.95, 75.57, 75.41, 73.66, 72.86, 72.50, 71.62, 69.37 (2C), 69.06, 68.99, 68.88, 68.80 (3C), 68.71 (2C), 65.94, 61.68, 34.18, 34.10, 34.01, 31.93 (3C), 29.72 (12C), 29.67 (6C), 29.54 (3C), 29.38 (3C), 29.33 (3C), 29.17 (3C), 24.85 (2C), 24.70, 22.70 (3C), 14.13 (3C).

³¹P-NMR (160 MHz, CDCl₃) δ: -1.87.

HRMS (ESI-TOF) calcd for $C_{92}H_{151}Na_2O_{29}P [M+2Na]^{2+} 898.4932$, found 898.4940.

Synthesis of Ac₁PIM₁ 1

To a stirred solution of **3** (21.8 mg, 12.5 μ mol) in acetone (1 mL) was added LiBr (10.8 mg, 125 μ mol) and refluxed for 6 h. After the removal of solvent, the crude mixture was purified by column chromatography of SiO₂ (toluene:EtOAc = 1:1 to CHCl₃:MeOH = 6:1) to give a lithium salt. The salt was dissolved in CH₂Cl₂:MeOH = 1:1 containing 1% TFA (vol/vol) (1 mL). [CpRu(C₃H₅)(C₉H₆NCOO)]PF₆ (6.53 mg, 12.5 μ mol) was added to the reaction mixture. After stirring for 1 h, the solvent was removed *in vacuo*. The crude mixture was purified by column chromatography of SiO₂ with CHCl₃-MeOH (10:1 to 1:1) and metal scavenger SiliaMetS® DMT with CHCl₃-MeOH (10:1 to 1:1) and SiO₂ with CHCl₃-MeOH (10:1 to 1:1) to give **1** (2.15 mg, 14%) as a white solid. The purity of **1** was 99% (measured by quantitative NMR using CHBr3 as an internal standard).

¹H-NMR (800 MHz, CDCl₃:CD₃OD:D₂O = 65:35:8) δ: 5.25 (d, J = 4.7 Hz, 1H), 5.10-5.09 (m, 1H), 4.42 (dd, J = 12.1, 2.7 Hz, 1H), 4.34 (dd, J = 11.7, 1.6 Hz, 1H), 4.26-4.24 (m, 1H), 4.22-4.21 (m, 1H), 4.18 (dd, J = 12.1, 7.4 Hz, 1H), 4.15-4.13 (m, 1H), 4.06-4.02 (m, 1H), 4.02-4.00 (m, 1H), 3.99-3.94 (m, 2H), 3.74 (dd, J = 9.4, 3.9 Hz, 1H), 3.72 (t, J = 9.4 Hz, 1H), 3.65 (t, J = 9.8 Hz, 1H), 3.57 (t, J = 9.8 Hz, 1H), 3.47 (dd, J = 10.2, 2.3 Hz, 1H), 3.26 (t, J = 9.0 Hz, 1H), 2.36 (t, J = 7.8 Hz, 2H), 2.34 (t, J = 7.4 Hz, 2H), 2.30 (td, J = 7.8, 1.8 Hz, 2H), 1.63-1.56 (m, 6H), 1.33-1.22 (m, 72H), 0.88 (t, J = 7.0 Hz, 9H).

¹³C-NMR (200 MHz, CDCl₃:CD₃OD:D₂O = 65:35:8) δ : 175.15, 174.29, 173.97, 101.75, 78.99, 76.51, 74.55, 72.60, 72.29, 70.73, 70.59, 70.48, 70.44, 70.11, 67.09, 63.62, 63.54, 62.90, 34.18, 34.06, 33.86, 31.86 (3C), 29.67 (3C), 29.64 (6C), 29.58 (3C), 29.55 (3C), 29.52 (3C), 29.36 (3C), 29.28 (3C), 29.17 (3C), 29.10 (3C), 24.89, 24.80, 24.73, 22.58 (3C), 13.86 (3C).

³¹P-NMR (160 MHz, CDCl₃:CD₃OD:D₂O = 65:35:8) δ : -0.16.

HRMS (**ESI-TOF**) calcd for $C_{63}H_{118}O_{19}P$ [M-H]⁻ 1209.8019, found 1209.8010.

Synthesis of glycerolipid derivative 24a

To a solution of (R)-3-(4-methoxybenzyloxy)propane-1,2-diol **20**⁸ (108 mg, 500 μmol), palmitic acid (385 mg, 1.50 mmol) and EDC•HCl (288 mg, 1.50 mmol) in CH₂Cl₂ (3 mL) was added DMAP (18.3 mg, 150 μmol). The resulting mixture was stirred at room temperature for overnight. The reaction was quenched by an addition of sat. KHSO₄ aq. and extracted with CHCl₃. Organic layer was washed with brine and dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with *n*-hexane-EtOAc (6:1) to give **24a** (345 mg, 95%) as white solid.

¹H-NMR (400 MHz, CDCl₃) δ: 7.23 (d, J = 8.5 Hz, 2H), 6.87 (d, J = 8.5 Hz, 2H), 5.26-5.19 (m, 1H), 4.47 (dd, J = 19.1, 11.8 Hz, 2H), 4.33 (dd, J = 12.1, 3.8 Hz, 1H), 4.17 (dd, J = 12.0, 6.3 Hz, 1H), 3.81 (s, 3H), 3.55 (d, J = 5.1 Hz, 2H), 2.33-2.25 (m, 4H), 1.64-1.51 (m, 4H), 1.33-1.18 (m, 48H), 0.88 (t, J = 6.8 Hz, 6H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.25, 172.97, 159.23, 129.64, 129.19 (2C), 113.68 (2C), 72.84, 69.92, 67.77, 62.59, 55.07, 34.22, 34.01, 31.87 (2C), 29.64 (3C), 29.61 (3C), 29.58 (2C), 29.53, 29.43 (2C), 29.40, 29.31 (2C), 29.23 (2C), 29.20, 29.05 (2C), 29.01, 24.88, 24.81, 22.62 (2C), 14.04 (2C). HRMS (ESI-TOF) calcd for C₄₃H₇₆NaO₆ [M+Na]⁺ 711.5534, found 711.5538.

Synthesis of glycerolipid derivative 22b

To a solution of (R)-3-(4-methoxybenzyloxy)propane-1,2-diol 20^8 (425 mg, 2.00 mmol), palmitic acid (513 mg, 2.00 mmol) and EDC•HCl (383 mg, 2.00 mmol) in CH₂Cl₂ (10 mL) was added DMAP (24.4 mg, 200 µmol). The resulting mixture was stirred at room temperature for overnight. The reaction was quenched by an addition of sat. KHSO₄ aq. and extracted with CHCl₃. Organic layer was washed with

brine and dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with *n*-hexane-EtOAc (2:1 to EtOAc only) to give **22b** (83.0 mg, 35%) as white solid.

¹H-NMR (400 MHz, CDCl₃) δ : 7.25 (d, J = 8.8 Hz, 2H), 6.88 (d, J = 8.8 Hz, 2H), 5.29 (s, 1H), 4.48 (s, 2H), 4.20-4.09 (m, 2H), 4.05-3.98 (m, 1H), 3.80 (s, 3H), 3.52 (dd, J = 9.3, 4.4 Hz, 1H), 3.45 (dd, J = 9.5, 6.1 Hz, 1H), 2.56 (d, J = 4.4 Hz, 1H), 2.32 (t, J = 7.6 Hz, 2H), 1.65-1.57 (m, 2H), 1.25-1.25 (m, 22H), 0.88 (t, J = 6.6 Hz, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.91, 159.33, 129.71, 129.38 (2C), 113.82 (2C), 73.12, 70.52, 68.87, 65.32, 55.22, 34.11, 31.89, 29.65 (2C), 29.63, 29.61 (2C), 29.57, 29.43, 29.32, 29.22, 29.09, 24.87, 22.65, 14.08.

HRMS (ESI-TOF) calcd for C₂₇H₄₆NaO₅ [M+Na]⁺ 473.3237, found 473.3241.

Synthesis of glycerolipid derivative 22c

The title compound was synthesized following the procedure for **22b**. (R)-3-(4-methoxybenzyloxy)propane-1,2-diol **20**⁸ (106 mg, 500 μ mol) and compound **21c**³ (136 mg, 500 μ mol) were used as starting materials. Compound **22c** (97.8 mg, 33%) was obtained as a white solid.

¹H-NMR (400 MHz, CDCl₃) δ: 7.25 (d, J = 8.8 Hz, 2H), 6.89 (dd, J = 6.6, 2.2 Hz, 2H), 4.49 (s, 2H), 4.20-4.09 (m, 2H), 4.02 (td, J = 9.6, 5.0 Hz, 1H), 3.81 (s, 3H), 3.53 (dd, J = 9.5, 4.1 Hz, 1H), 3.46 (dd, J = 9.5, 6.1 Hz, 1H), 3.24 (q, J = 6.8 Hz, 2H), 2.52 (d, J = 4.4 Hz, 1H), 2.39-2.25 (m, 2H), 2.19 (q, J = 7.6 Hz, 2H), 1.51-1.44 (m, 2H), 1.34-1.22 (m, 16H), 1.15 (t, J = 7.6 Hz, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.77, 159.11, 129.64, 129.21 (2C), 129.11, 113.62 (2C), 72.92, 70.47, 68.55, 65.26, 55.06, 39.33, 33.94, 29.50, 29.42, 29.27, 29.26, 29.18, 29.08, 29.01, 28.88, 26.72, 24.68, 9.83.

HRMS (ESI-TOF) calcd for C₂₆H₄₃NNaO₆ [M+Na]⁺ 488.2983, found 488.2992.

Synthesis of glycerolipid derivative 24b

To a solution of **22b** (83.0 mg, 184 μmol), **23b**³ (100 mg, 368 μmol) and EDC•HCl (70.6 mg, 368 μmol) in CH₂Cl₂ (3 mL) was added DMAP (4.50 mg, 36.8 μmol). The resulting mixture was stirred at room temperature for overnight. The reaction was quenched by an addition of sat. KHSO₄ aq. and extracted with CHCl₃. Organic layer was washed with brine and dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with *n*-hexane-EtOAc (2:1) to give **24b** (112 mg, 87%) as white solid.

¹H-NMR (400 MHz, CDCl₃) δ: 7.23 (d, J = 8.4 Hz, 2H), 6.87 (d, J = 8.5 Hz, 2H), 5.92-5.82 (m, 1H), 5.21 (dt, J = 10.3, 4.6 Hz, 1H), 4.46 (dd, J = 15.7, 11.6 Hz, 2H), 4.38 (dd, J = 12.0, 3.7 Hz, 1H), 4.15 (dd, J = 11.9, 5.9 Hz, 1H), 3.80 (s, 3H), 3.56 (d, J = 5.3 Hz, 2H), 3.35-3.19 (m, 2H), 2.36 (t, J = 7.1 Hz, 2H), 2.28 (t, J = 7.6 Hz, 2H), 2.12 (t, J = 7.6 Hz, 2H), 1.86-1.79 (m, 2H), 1.65-1.52 (m, 4H), 1.35-1.19 (m, 38H), 0.88 (t, J = 6.7 Hz, 6H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.43, 173.24, 172.57, 159.29, 129.51, 129.28 (2C), 113.76 (2C), 72.93, 70.41, 67.73, 62.36, 55.17, 38.50, 36.68, 34.02, 31.85, 31.82, 31.52, 29.62 (2C), 29.58 (2C), 29.55, 29.51, 29.44, 29.42, 29.31, 29.28 (3C), 29.25, 29.21, 29.05, 25.68, 24.81, 24.51, 22.62, 14.04. HRMS (ESI-TOF) calcd for C₄₂H₇₃NNaO₇ [M+Na]⁺ 726.5279, found 726.5287.

Synthesis of glycerolipid derivative 24c

The title compound was synthesized following the procedure for **24b**. **22c** (177 mg, 380 μ mol) and palmitic acid (292 mg, 1.14 mmol) were used as starting materials. Compound **24c** (230 mg, 86%) was obtained as a white solid.

¹H-NMR (400 MHz, CDCl₃) δ : 7.23 (d, J = 8.5 Hz, 2H), 6.87 (d, J = 8.6 Hz, 2H), 5.41 (s, 1H), 5.26-5.19 (m, 1H), 4.47 (dd, J = 18.6, 11.7 Hz, 2H), 4.32 (dd, J = 11.8, 3.8 Hz, 1H), 4.17 (dd, J = 11.8, 6.5

Hz, 1H), 3.81 (s, 3H), 3.55 (dd, J = 5.2, 1.0 Hz, 2H), 3.24 (dd, J = 13.1, 7.0 Hz, 2H), 2.37-2.24 (m, 4H), 2.19 (q, J = 7.6 Hz, 2H), 1.64-1.56 (m, 4H), 1.51-1.45 (m, 2H), 1.33-1.20 (m, 38H), 1.16 (t, J = 7.6 Hz, 3H), 0.88 (t, J = 6.8 Hz, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.64, 173.42, 173.14, 159.28, 129.73, 129.28 (2C), 113.79 (2C), 72.94, 69.99, 67.88, 62.69, 55.25, 39.51, 34.32, 34.08, 31.91, 29.80, 29.68 (2C), 29.65 (3C), 29.63, 29.47 (3C), 29.40, 29.35 (2C), 29.28, 29.27, 29.23, 29.07 (2C), 26.89, 24.94, 24.84, 22.67, 14.12, 9.94. HRMS (ESI-TOF) calcd for C₄₂H₇₃NNaO₇ [M+Na]⁺ 726.5279, found 726.5287.

Synthesis of glycerolipid derivative 25a

To a solution of **24a** (46.7 mg, 67.8 μmol) in CH₂Cl₂ (2 mL) and pH 7 phosphate buffer (220 μL) was added DDQ (30.8 mg, 136 μmol). The resulting mixture was stirred at room temperature for overnight. The reaction was quenched by an addition of sat. NaHCO₃ aq. and extracted with CHCl₃. Organic layer was washed with brine and dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with toluene-EtOAc (6:1) to give **25a** (29.4 mg, 76%) as white solid.

¹**H-NMR (400 MHz, CDCl₃)** δ : 5.04-4.99 (m, 1H), 4.25 (dd, J = 12.0, 4.4 Hz, 1H), 4.16 (dd, J = 12.0, 5.7 Hz, 1H), 3.66 (d, J = 5.2 Hz, 2H), 2.26 (q, J = 7.9 Hz, 4H), 1.60-1.51 (m, 4H), 1.27-1.14 (m, 48H), 0.81 (t, J = 6.7 Hz, 6H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.80, 173.44, 72.08, 61.99, 61.51, 34.27, 34.09, 31.91 (2C), 29.68 (4C), 29.67 (2C), 29.64 (4C), 29.61 (2C), 29.46 (2C), 29.34 (2C), 29.26 (2C), 29.10, 29.07, 24.93, 24.87, 22.67 (2C), 14.10 (2C).

HRMS (ESI-TOF) calcd for $C_{35}H_{68}NaO_5$ [M+Na]⁺ 591.4959, found 591.4963.

Synthesis of glycerolipid derivative 25b

The title compound was synthesized following the procedure for **25a**. **24b** (30.0 mg, 42.6 μ mol) and DDQ (19.3 mg, 85.2 μ mol) were used as starting materials. Compound **25b** (23.3 mg, 94%) was obtained as a white solid.

¹**H-NMR (400 MHz, CDCl₃)** δ: 5.72-5.68 (m, 1H), 5.10-5.05 (m, 1H), 4.32-4.05 (m, 2H), 3.84-3.80 (m, 1H), 3.69 (dd, *J* = 12.5, 5.4 Hz, 1H), 3.51-3.09 (m, 2H), 2.39-2.26 (m, 4H), 2.16-2.11 (m, 2H), 1.91-1.69 (m, 2H), 1.61-1.54 (m, 4H), 1.31-1.18 (m, 38H), 0.85 (t, *J* = 6.8 Hz, 6H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.84, 173.66, 172.50, 72.94, 62.16, 61.22, 38.04, 36.75, 34.10, 31.91, 31.87, 31.07, 29.68 (3C), 29.64 (3C), 29.60, 29.55, 29.48, 29.46, 29.34, 29.31, 29.28, 29.27, 29.10, 25.68, 25.09, 24.87, 22.67, 22.66, 14.11 (2C).

HRMS (ESI-TOF) calcd for C₃₄H₆₅NNaO₆ [M+Na]⁺ 606.4704, found 606.4708.

Synthesis of glycerolipid derivative 25c

The title compound was synthesized following the procedure for **25a**. **24c** (193 mg, 274 μ mol) and DDQ (125 mg, 549 μ mol) were used as starting materials. Compound **25c** (145 mg, 90%) was obtained as a white solid.

¹H-NMR (400 MHz, CDCl₃) δ : 5.47-5.35 (m, 1H), 5.11-5.06 (m, 1H), 4.32 (dd, J = 12.0, 4.6 Hz, 1H), 4.24 (dd, J = 12.0, 5.6 Hz, 1H), 3.76-3.70 (m, 2H), 3.24 (td, J = 6.8, 6.1 Hz, 2H), 2.34 (q, J = 7.9 Hz, 4H), 2.19 (q, J = 7.6 Hz, 2H), 1.66-1.58 (m, 4H), 1.52-1.46 (m, 2H), 1.33-1.21 (m, 38H), 1.16 (t, J = 7.6 Hz, 3H), 0.88 (t, J = 6.8 Hz, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.77, 173.65, 173.41, 72.07, 62.00, 61.49, 39.49, 34.27, 34.07, 31.91, 29.80, 29.68 (3C), 29.64 (3C), 29.60, 29.46, 29.42, 29.39, 29.34, 29.31, 29.26, 29.21, 29.14, 29.07, 29.02, 26.84, 24.93, 24.83, 22.67, 14.11, 9.93.

Synthesis of inositol phosphate derivative 26

To a suspension of NaH (60% oil dispersion) (27.6 mg, 689 μ mol) in THF (7 mL) at 0 °C was added allyl alcohol (52.6 μ L, 773 μ mol). After stirring for 30 min, the reaction was added to the solution of 14⁷ (241 mg, 276 μ mol) in THF (3 mL) at 0 °C. The reaction mixture was warmed up to room temperature, and stirred for 1.5 h. The reaction was cooled to 0 °C, and then quenched with 10% aqueous citric acid. The mixture was extracted with ethyl acetate. The organic solution was washed with sat. NaHCO₃ aq. and brine, dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with *n*-hexane-EtOAc (5:1). The resulting product was dissolved with acetone (3 mL). To the solution was added LiBr (120 mg, 1.38 mmol) and refluxed for 6 h. After the removal of solvent, the crude mixture was purified by column chromatography of SiO₂ with toluene-EtOAc (5:1), then CH₂Cl₂-MeOH (6:1) to give lithium salt. The desire product was diluted with EtOAc and washed with sat. KHSO₄ aq. and brine. The organic layer was dried over Na₂SO₄, filtrated and concentrated *in vacuo* to obtain proton form 26 (138 mg, 75%) as a colorless oil.

¹H-NMR (400 MHz, CDCl₃) δ: 9.82 (s, 1H), 7.33-7.27 (m, 4H), 6.87-6.82 (m, 4H), 6.01-5.78 (m, 3H), 5.34-5.30 (m, 1H), 5.29-5.22 (m, 2H), 5.19-5.14 (m, 3H), 4.83-4.66 (m, 6H), 4.48-4.45 (m, 2H), 4.34-4.28 (m, 3H), 4.19-4.15 (m, 1H), 4.11-4.04 (m, 2H), 3.89 (t, J = 9.6 Hz, 1H), 3.83 (t, J = 9.5 Hz, 1H), 3.79 (s, 3H), 3.76 (s, 3H), 3.44 (s, 3H), 3.27-3.20 (m, 2H).

¹³C-NMR (100 MHz, CDCl₃) δ: 159.12, 159.08, 135.16, 134.52, 132.37, 132.30, 130.87, 130.49, 129.83, 129.75, 117.86, 116.74, 116.27, 113.65, 113.56, 97.47, 82.50, 80.82, 79.68, 79.61, 79.17, 77.88, 77.82, 75.49, 75.39, 74.49, 73.80, 71.31, 68.00, 67.94, 55.71, 55.19, 55.14.

³¹P-NMR (160 MHz, CDCl₃) δ : 0.69.

HRMS (**ESI-TOF**) calcd for C₃₃H₄₄NaO₁₂P [M+Na]⁺ 663.2576, found 663.2584.

Synthesis of inositol phospholipid derivative 4a

To the solution of **26** (50.1 mg, 75.4 μ mol), **25a** (85.9 mg, 151 μ mol) and PPh₃ (39.6 mg, 151 mmol) in THF (750 μ L) was added DIAD (27.6 μ L, 151 μ mol) and stirred for overnight. The reaction mixture was concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with *n*-hexane-EtOAc (1:3) to give **4a** (60.5 mg, 66%) as white solid.

¹H-NMR (400 MHz, CDCl₃) δ: 7.32-7.26 (m, 4H), 6.87-6.83 (m, 4H), 6.00-5.77 (m, 3H), 5.35-5.14 (m, 6H), 5.10-4.94 (m, 1H), 4.81-4.76 (m, 4H), 4.68 (dd, J = 10.3, 3.4 Hz, 2H), 4.55-4.41 (m, 2H), 4.37 (dd, J = 13.0, 4.9 Hz, 1H), 4.31-3.92 (m, 9H), 3.88 (t, J = 8.5 Hz, 1H), 3.83 (t, J = 8.5 Hz, 1H), 3.80 (s, 3H), 3.79 (s, 3H), 3.45-3.39 (m, 3H), 3.30-3.23 (m, 2H), 2.32-2.23 (m, 4H), 1.62-1.52 (m, 4H), 1.33-1.19 (m, 48H), 0.88 (t, J = 6.7 Hz, 6H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.08, 172.74, 159.17, 159.06, 135.11, 134.56, 132.28, 132.22, 130.85, 130.59, 129.81, 129.45, 129.32, 118.28, 118.16, 116.77, 116.39, 113.71, 113.60, 113.56, 97.46, 82.68, 80.91, 79.28, 77.96, 75.56, 75.15, 74.50, 73.67, 71.38, 69.23, 68.41, 65.56, 65.23, 61.61, 55.78, 55.22, 55.19, 34.06, 33.92, 31.89 (2C), 29.67 (6C), 29.63 (6C), 29.47 (2C), 29.33 (2C), 29.27 (2C), 29.09, 29.06, 24.78 (2C), 22.66 (2C), 14.09 (2C).

³¹P-NMR (160 MHz, CDCl₃) δ : -0.59.

HRMS (ESI-TOF) calcd for $C_{68}H_{111}NaO_{16}P [M+Na]^+ 1273.7502$, found 1273.7496.

Synthesis of inositol phospholipid derivative 4b

The title compound was synthesized following the procedure for **4a**. compounds **26** (65.0 mg, 97.8 μ mol) and **25b** (114 mg, 196 μ mol) were used as starting materials. Compound **4b** (70.6 mg, 59%) was obtained as a white solid.

¹H-NMR (400 MHz, CDCl₃) δ: 7.30-7.27 (m, 4H), 6.88-6.84 (m, 4H), 6.27-6.04 (m, 1H), 5.93-5.84 (m, 3H), 5.34-5.05 (m, 7H), 4.82-4.76 (m, 4H), 4.70-4.64 (m, 2H), 4.53-4.46 (m, 2H), 4.39-4.00 (m, 10H), 3.98-3.77 (m, 8H), 3.43 (s, 3H), 3.40-3.13 (m, 4H), 2.41-2.24 (m, 4H), 2.18-2.10 (m, 2H), 1.86-1.76 (m, 2H), 1.64-1.56 (m, 4H), 1.33-1.21 (m, 38H), 0.88 (t, *J* = 6.5 Hz, 6H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.31, 173.27, 172.24, 159.19, 159.12, 135.06, 134.51, 130.81, 130.57, 130.53, 129.82, 129.79, 129.47, 129.34, 118.42, 118.26, 116.86, 116.80, 116.41, 113.72, 113.61, 97.42, 82.74, 80.90, 79.26, 78.04, 75.58, 74.50, 73.65, 71.39, 68.39, 67.70, 65.62, 64.81, 61.43, 61.43, 55.74, 55.23, 55.20, 38.22, 36.69, 33.94, 31.88, 31.84, 31.06, 29.65 (2C), 29.61, 29.58, 29.55, 29.53, 29.49, 29.45, 29.42, 29.36, 29.31, 29.28, 29.25, 29.10, 29.08, 25.74, 24.78, 24.21, 22.65, 22.63, 14.08 (2C).

³¹P-NMR (160 MHz, CDCl₃) δ: -0.64.

HRMS (ESI-TOF) calcd for C₆₇H₁₀₈NNaO₁₇P [M+Na]⁺ 1252.7247, found 1252.7253.

Synthesis of inositol phospholipid derivative 4c

The title compound was synthesized following the procedure for **4a**. Compounds **26** (65.0 mg, 97.8 μ mol) and **25c** (114 mg, 196 μ mol) were used as starting materials. Compound **4c** (115 mg, 95%) was obtained as a white solid.

¹**H-NMR (400 MHz, CDCl₃)** δ: 7.31-7.28 (m, 4H), 6.87-6.83 (m, 4H), 6.00-5.77 (m, 3H), 5.35-5.24 (m, 3H), 5.22-5.14 (m, 3H), 5.12-5.07 (m, 1H), 4.80-4.77 (m, 4H), 4.68 (dd, J = 10.3, 4.4 Hz, 2H), 4.52-4.42 (m, 2H), 4.39-4.01 (m, 10H), 3.97-3.71 (m, 8H), 3.45-3.40 (m, 3H), 3.29-3.20 (m, 5H), 2.36-2.12 (m, 6H), 1.63-1.56 (m, 4H), 1.50-1.46 (m, 2H), 1.30-1.23 (m, 38H), 1.15 (t, J = 7.6 Hz, 3H), 0.88 (t, J = 6.7 Hz, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.63, 173.28, 172.67, 159.11, 159.02, 135.05, 134.49, 132.21,

132.13, 130.80, 130.52, 129.72, 129.38, 129.26, 118.21, 116.72, 116.29, 113.64 (2C), 113.53, 113.50, 97.37, 82.63, 80.85, 79.22, 75.47, 75.08, 74.42, 73.61, 71.98, 71.33, 69.27, 68.22, 62.09, 61.47, 61.17, 55.70, 55.15, 55.10, 39.41, 34.19, 33.98, 31.81, 29.64, 29.58 (3C), 29.54 (3C), 29.40 (2C), 29.31, 29.25 (2C), 29.19 (2C), 29.13, 29.06, 28.98, 28.93, 26.81, 24.74, 24.69, 22.57, 14.02, 9.87.

³¹P-NMR (160 MHz, CDCl₃) δ : -0.67.

HRMS (ESI-TOF) calcd for $C_{67}H_{108}NNaO_{17}P$ [M+Na]⁺ 1252.7247, found 1252.7255.

Synthesis of inositol phospholipid derivative S7a

The solution of **4a** (60.5 mg, 49.8 μmol) in CH₂Cl₂ (1 mL) was cooled to 0 °C. To the solution was added trifluoroacetic acid (1 mL) and stirred for 1h. The solvent removed under reduced pressure and added toluene then concentrated *in vacuo*. The residue was purified by column chromatography of SiO₂ with CHCl₃-MeOH (9:1) to give **S7a** (55.5 mg, quant.) as a colorless oil.

¹H-NMR (400 MHz, CDCl₃) δ: 6.02-5.89 (m, 3H), 5.40-5.16 (m, 7H), 4.65-4.57 (m, 2H), 4.39-4.12 (m, 10H), 4.07-4.01 (m, 1H), 3.93 (td, *J* = 9.5, 4.6 Hz, 1H), 3.29-3.25 (m, 1H), 3.19 (td, *J* = 9.3, 4.2 Hz, 1H), 2.35-2.28 (m, 4H), 1.62-1.57 (m, 4H), 1.29-1.23 (m, 48H), 0.88 (t, *J* = 6.8 Hz, 6H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.50, 173.25, 134.95, 134.05 (2C), 118.83, 118.27, 117.30, 81.51, 81.47, 78.27, 78.23, 73.92, 73.90, 71.60, 71.58, 71.48, 71.36, 61.83, 61.78, 34.12, 33.99, 31.90 (2C), 29.68 (6C), 29.66 (2C), 29.64 (4C), 29.48 (2C), 29.34 (2C), 29.28 (2C), 29.11, 29.07, 24.79 (2C), 22.67 (2C), 14.09 (2C).

³¹P-NMR (160 MHz, CDCl₃) δ: -1.44.

HRMS (ESI-TOF) calcd for $C_{50}H_{91}NaO_{13}P$ [M+Na]⁺ 953.6090, found 953.6082.

Synthesis of inositol phospholipid derivative S7b

The title compound was synthesized following the procedure for **S7a**. Compound **4b** (70.6 mg, 57.4 µmol) was used as starting materials. Compound **S7b** (41.3 mg, 76%) was obtained as a colorless oil.

¹H-NMR (400 MHz, CDCl₃) δ: 6.02-5.89 (m, 3H), 5.41-5.17 (m, 7H), 4.66-4.57 (m, 2H), 4.39-4.10 (m, 10H), 4.03 (t, J = 9.5 Hz, 1H), 3.93 (td, J = 9.5, 6.6 Hz, 1H), 3.33-3.23 (m, 3H), 3.16 (td, J = 9.3, 4.2 Hz, 1H), 2.44-2.37 (m, 2H), 2.31 (t, J = 7.5 Hz, 2H), 2.15 (t, J = 7.7 Hz, 2H), 1.87-1.79 (m, 2H), 1.62-1.58 (m, 4H), 1.25-1.25 (m, 38H), 0.88 (t, J = 6.7 Hz, 6H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.70, 173.43, 172.49, 135.02, 134.11, 132.12, 118.70, 118.14, 117.22, 81.73, 79.10, 78.46, 73.98, 71.67, 71.26, 70.91, 69.79, 68.90, 68.09, 65.71, 61.55, 38.34, 36.73, 33.97, 31.89, 31.85, 31.15, 29.66 (3C), 29.62 (2C), 29.59, 29.55, 29.49, 29.45, 29.35, 29.32 (2C), 29.28, 29.25, 29.09, 25.75, 24.79, 24.59, 22.65 (2C), 14.08 (2C).

³¹P-NMR (160 MHz, CDCl₃) δ : -0.80.

HRMS (**ESI-TOF**) calcd for C₄₉H₈₈NNaO₁₄P [M+Na]⁺ 968.5835, found 968.5842.

Synthesis of inositol phospholipid derivative S7c

The title compound was synthesized following the procedure for **S7a**. **4c** (115 mg, 93.3 μmol) were used as starting materials. Compound **S7c** (59.9 mg, 68%) was obtained as a colorless oil.

¹H-NMR (400 MHz, CDCl₃) δ: 6.01-5.90 (m, 3H), 5.40-5.17 (m, 7H), 4.65-4.58 (m, 2H), 4.38-4.32 (m, 5H), 4.26-4.11 (m, 5H), 4.06-4.01 (m, 1H), 3.93 (td, J = 9.5, 5.0 Hz, 1H), 3.28-3.14 (m, 4H), 2.35-2.29 (m, 4H), 2.20 (q, J = 7.6 Hz, 2H), 1.63-1.57 (m, 4H), 1.50-1.46 (m, 2H), 1.30-1.24 (m, 38H),

1.15 (t, J = 7.6 Hz, 3H), 0.88 (t, J = 6.8 Hz, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ: 173.78, 173.40, 173.06, 134.99 (2C), 134.10, 118.70, 118.14, 117.20, 81.64, 79.11, 78.39, 73.93, 71.61, 71.38, 71.23, 69.34, 68.95, 68.27, 65.72, 61.73, 39.49, 34.10, 33.95, 31.87, 29.72, 29.65 (3C), 29.62, 29.60, 29.56, 29.45, 29.40, 29.38, 29.30 (3C), 29.25, 29.20, 29.13, 29.04, 28.99, 26.82, 24.77, 24.73, 22.64, 14.07, 9.90.

³¹P-NMR (160 MHz, CDCl₃) δ : -0.96.

HRMS (ESI-TOF) calcd for C₄₉H₈₈NNaO₁₄P [M+Na]⁺ 968.5835, found 968.5839.

Synthesis of PI 2a

To a solution of S7a (55.5 mg, 59.6 μmol) and 1,3-dimethylbarbituric acid (55.8 mg, 358 μmol) in CH₂Cl₂/MeOH/H₂O (10/10/3) (5 mL) was added Pd(PPh₃)₄ (68.9 mg, 59.6 μmol). The resulting mixture was stirred at room temperature for 4 h. The reaction was concentrated under reduced pressure. The crude mixture was purified by column chromatography of SiO₂ with CHCl₃-MeOH (10:1 to 1:1) and metal scavenger SiliaMetS® DMT with CHCl₃-MeOH (10:1 to 1:1) and SiO₂ with CHCl₃-MeOH (10:1 to 1:1) to give PI 2a (13.5 mg, 28%) as white solid. The purity of 2a was 99% (measured by quantitative NMR using CHBr3 as an internal standard).

¹H-NMR (800 MHz, CDCl₃:CD₃OD:D₂O = 70:40:6) δ: 5.25-5.24 (m, 1H), 4.42 (dd, J = 11.9, 2.8 Hz, 2H), 4.20-4.16 (m, 1H), 4.08-3.99 (m, 2H), 3.89-3.85 (m, 1H), 3.74 (t, J = 9.6 Hz, 1H), 3.62 (t, J = 9.6 Hz, 1H), 3.42 (dd, J = 10.1, 2.8 Hz, 1H), 3.24 (t, J = 9.2 Hz, 1H), 2.33 (t, J = 7.3 Hz, 2H), 2.30 (t, J = 7.8 Hz, 2H), 1.61-1.56 (m, 4H), 1.31-1.22 (m, 48H), 0.87 (t, J = 6.9 Hz, 6H).

¹³C-NMR (200 MHz, CDCl₃:CD₃OD:D₂O = 70:40:6) δ : 173.88, 173.57, 76.19, 74.02, 72.06, 71.30, 71.08, 70.81, 70.15, 63.21, 62.49, 33.77, 33.64, 31.45 (2C), 29.22 (4C), 29.19 (2C), 29.17 (2C), 29.11 (2C), 29.07 (2C), 28.92 (2C), 28.87 (2C), 28.69 (2C), 28.66 (2C), 24.46, 24.40, 22.18 (2C), 13.44 (2C).

³¹P-NMR (160 MHz, CDCl₃:CD₃OD:D₂O = 70:40:6) δ : 0.03.

HRMS (ESI-TOF) calcd for $C_{41}H_{78}O_{13}P$ [M-H]⁻ 809.5195, found 809.5186.

Synthesis of PI derivative 2b

The title compound was synthesized following the procedure for **2a**. **S7b** (41.3 mg, 43.7 μmol) were used as starting materials. Compound **2b** (13.0 mg, 34%) was obtained as a colorless oil. The purity of **2b** was 98% (measured by quantitative NMR using CHBr3 as an internal standard).

¹H-NMR (800 MHz, CDCl₃:CD₃OD:D₂O = 70:40:6) δ: 5.26-5.23 (m, 1H), 4.40 (dd, J = 12.2, 3.0 Hz, 1H), 4.20-4.17 (m, 2H), 4.05 (t, J = 5.5 Hz, 2H), 3.92-3.88 (m, 1H), 3.75 (t, J = 9.4 Hz, 1H), 3.63 (t, J = 9.6 Hz, 1H), 3.43 (dd, J = 9.9, 3.0 Hz, 1H), 3.25 (t, J = 9.4 Hz, 1H), 3.21 (td, J = 7.1, 2.8 Hz, 2H), 2.46-2.33 (m, 2H), 2.31 (t, J = 7.6 Hz, 2H), 2.17 (t, J = 7.8 Hz, 2H), 1.83-1.78 (m, 2H), 1.61-1.56 (m, 4H), 1.30-1.24 (m, 38H), 0.89-0.86 (m, 6H).

¹³C-NMR (200 MHz, CDCl₃:CD₃OD:D₂O = 70:40:6) δ: 175.68, 174.93, 174.02, 77.02, 74.93, 73.02, 72.30, 72.01, 71.79, 71.51, 64.35, 63.21, 38.95, 36.88, 34.50, 32.41, 32.39, 31.89, 30.18 (3C), 30.09, 30.04, 30.02, 29.89, 29.83 (2C), 29.81 (2C), 29.63, 26.44, 25.31, 24.86, 23.13 (2C), 14.39 (2C).

³¹P-NMR (240 MHz, CDCl₃:CD₃OD:D₂O = 70:40:6) δ : 0.06.

HRMS (ESI-TOF) calcd for $C_{40}H_{75}O_{14}P [M-H]^{-} 824.4931$, found 824.4935.

Synthesis of PI derivative 2c

The title compound was synthesized following the procedure for 2a. S7c (59.9 mg, 63.3 μmol) were

used as starting materials. Compound **2c** (16.2 mg, 31%) was obtained as a colorless oil. The purity of **2c** was 98% (measured by quantitative NMR using CHBr3 as an internal standard).

¹H-NMR (800 MHz, CDCl₃:CD₃OD:D₂O = 70:40:6) δ: 5.27-5.21 (m, 1H), 4.47-4.39 (m, 1H), 4.20-4.14 (m, 2H), 4.07-3.97 (m, 2H), 3.93-3.86 (m, 1H), 3.74 (t, J = 9.5 Hz, 1H), 3.62 (t, J = 9.7 Hz, 1H), 3.43 (dd, J = 9.6, 2.3 Hz, 1H), 3.24 (t, J = 9.3 Hz, 1H), 3.15 (t, J = 7.2 Hz, 2H), 2.35-2.27 (m, 4H), 2.18 (q, J = 7.6 Hz, 2H), 1.61-1.55 (m, 4H), 1.50-1.43 (m, 2H), 1.33-1.21 (m, 38H), 1.12 (t, J = 7.6 Hz, 3H), 0.86 (t, J = 6.7 Hz, 3H).

¹³C-NMR (200 MHz, CDCl₃:CD₃OD:D₂O = 70:40:6) δ: 176.21, 174.92, 174.62, 77.03, 74.91, 73.00, 72.27, 72.05, 71.77, 71.09, 64.30, 63.48, 40.04, 34.75, 34.60, 32.45, 30.21 (2C), 30.17 (2C), 30.08, 30.05, 30.00 (2C), 29.92 (2C), 29.88 (2C), 29.82, 29.64 (2C), 27.47, 25.44, 25.34, 23.18, 14.44, 10.43.

HRMS (ESI-TOF) calcd for C₄₀H₇₅O₁₄P [M-H]⁻ 824.4931, found 824.4940.

³¹P-NMR (160 MHz, CDCl₃:CD₃OD:D₂O = 70:40:6) δ : 2.09.

2. Biology

DCAR Reporter Assay

2B4-NFAT-GFP reporter cells expressing DCAR + FcR γ were prepared as previously described⁹. For stimulation of reporter cells, compounds were diluted in isopropanol (Nakalai tesque) and added on 96-well plates (Corning) at 20 μ L/well.¹⁰ After evaporation of the solvent, the cells were stimulated with plate-coated lipids. Anti-flag tag antibody (Wako) was dissolved in NaHCO₃ buffer (10 μ g/ml) and 50 μ l of the antibody solution were incubated in DOTAP (Avanti) coated 96-well plate wells for 2 h at 37 °C.

Ac₁PIM₂

Ac₁PIM₂ was purified from *M. bovis* BCG as previous described.¹¹

HEK- Blue Reporter Assay

The induction of NF- κ B in a reporter gene assay was quantified using HEK-Blue mTLR2 and hTLR2 cells (Invivogen). HEK-Blue cells were cultured in DMEM (Nacalai tesque) supplemented with 10% fetal bovine serum (FBS; Biowest), 1% penicillin–streptomycin (Gibco) and 100 μ g/mL Normocin. The cells were seeded into 96-well plates (1.0 x10⁵ cells/well), and ligands were added with the indicated concentrations before incubation at 37 °C for 24 h. SEAP was measured using a HEK-Blue Detection (Invivogen).

APC (Antigen Presenting Cell)-free Assay for CD1d-Lipid Binding 1,2

Initially, 96-well microplates (multiwell plate 96F, Sumitomo Bakelite Co., Ltd.) were coated with mouse CD1d:Ig fusion protein (0.25 μ g/well, BD Biosciences) in PBS (100 μ L) at 37 °C for 24 h. After washing with PBS, various concentrations of compounds in PBS containing 1% DMSO and 0.005% TritonX-100 were added and incubated at 37 °C for 24 h. After washing with PBS, 2E10 NKT hybridoma cells¹² (2.5 × 10⁵ cells/well) were added and cultured at 37 °C for 48 h. IL-2 in the supernatant were measured by ELISA kit (Affymetrix). 2E10 NKT hybridoma cells were a kind gift of Prof. Kazuya Iwabuchi (Kitasato University).

Binding Studies

The AlphaScreen assay (PerkinElmer Life Science) was used to determine receptor (mouse TLR2, human TLR2 and mouse CD1d) -ligand interactions.^{3,4} For analysis of mouse TLR2-ligand interaction, mouse TLR2-Fc (R&D systems, final concentration 10 nM) was mixed with Pam2CSK4-biotin (InvivoGen, final concentration 10 nM), in PBS containing 0.05 % human serum albumin. After 45 min, compounds (concentration range 3 nM-30 μM) was added and incubation was continued for another 45 min. Next, 10 μg/mL streptavidin donor beads and 10 μg/mL Protein-A acceptor beads were added and incubated for 45 min. Samples were measured at 680 nm in EnSpireTM Alpha (PerkinElmer) or Spark 10M (Tecan Ltd.). Data were analyzed using Graphpad Prism (GraphPad Software Inc.).

For analysis of human TLR2-ligand interaction, human TLR2-His tag (R&D systems, final concentration 15 nM) was mixed with Pam2CSK4-biotin (InvivoGen, final concentration 15 nM), in PBS containing 0.05 % human serum albumin. After 45 min, compounds (concentration range 3 nM-30 μM) was added and incubation was continued for another 45 min. Next, 10 μg/mL streptavidin donor beads and 10 μg/mL Ni chelate acceptor beads were added and incubated for 45 min. Samples were measured at 680 nm in EnSpireTM Alpha (PerkinElmer) or Spark 10M (Tecan Ltd.). Data were analyzed using Graphpad Prism (GraphPad Software Inc.).

For analysis of mouse CD1d-ligand interaction, mouse CD1d-IgG (BD Biosciences, final concentration 5 nM) was mixed with 10 μg/mL anti-mouse IgG acceptor beads in PBS containing 0.005% Tween 20. After 1 h, Biotinyl-PE (Avanti, final concentration 2 mM) and compounds (concentration range 3 nM-30 μM) was added to the plate. After incubation at 37 °C for 18 h, 10 μg/mL streptavidin donor beads was added and incubation was continued for another 1 h. Samples were measured at 680 nm in EnSpireTM Alpha (PerkinElmer) or Spark 10M (Tecan Ltd.). Data were analyzed using Graphpad Prism (GraphPad Software Inc.).

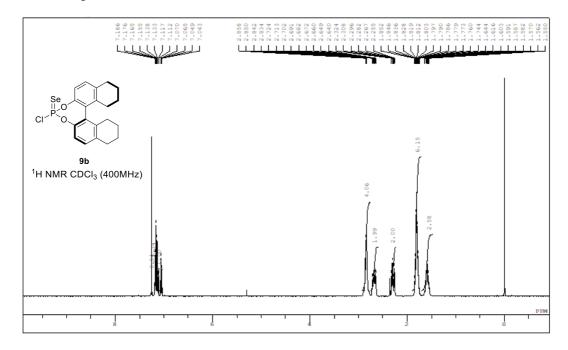
Inflammatory cytokine induction assay

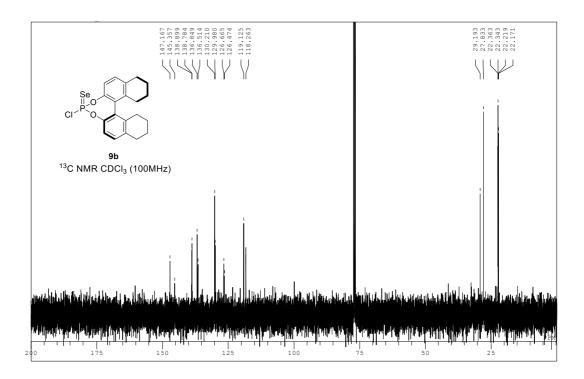
Mouse macrophages (RAW264.7 cells) were purchased from ATCC (TIB-71). These cells were

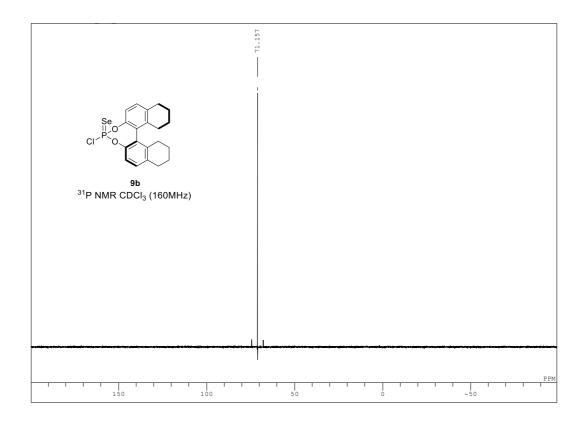
cultured as monolayers in RPMI-1640 (Nacalai Tesque) supplemented with 10% fetal bovine serum (FBS; Biowest), and 1% penicillin–streptomycin (Gibco). The cells were seeded into 96-well plates $(5.0 \text{ x} 10^4 \text{ cells/well})$, and ligands were added with the indicated concentrations before incubation at 37 °C for 24 h. Cytokine release was measured using an ELISA kit (Affymetrix).

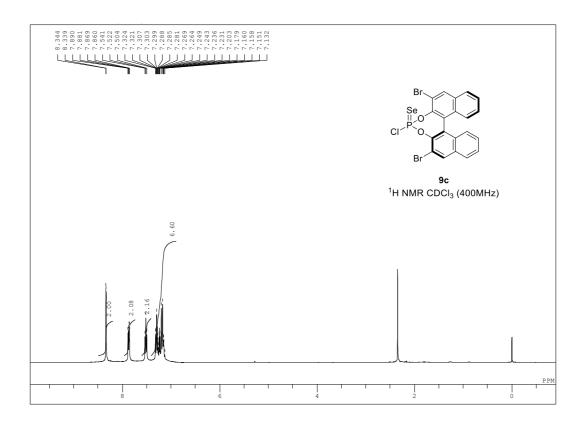
Bone marrow cells were flushed from the tibias and femurs of C57BL/6J mice (Charles River) with PBS. For BMDCs differentiation, the cell suspension was cultured at a density of 1.0 x 10⁶ cells/mL in RPMI-1640 (Nacalai Tesque) supplemented with 10% fetal bovine serum (FBS; Biowest), 1% penicillin–streptomycin (Gibco) and 20 ng/mL GM-CSF. On day 3, fresh medium containing GM-CSF was added and on day 5 one half of the medium was renewed. BMDCs were harvested on day 8 and seeded into 96-well plates (2.0 x10⁵ cells/well), and ligands were added with the indicated concentrations before incubation at 37 °C for 24 h. Cytokine release was measured using an ELISA kit (Affymetrix).

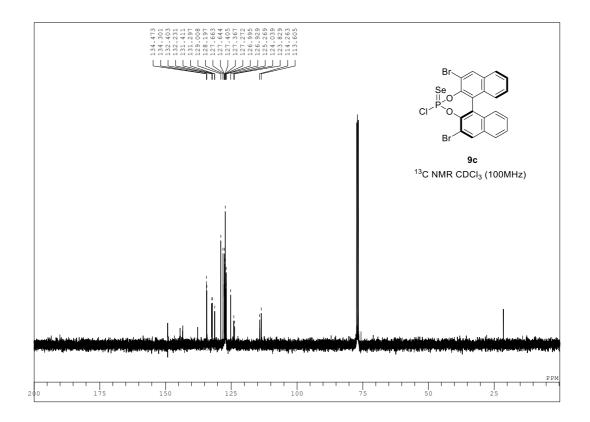
3. NMR spectra

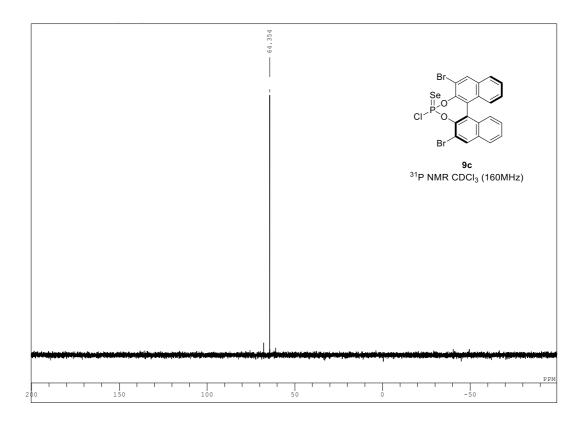


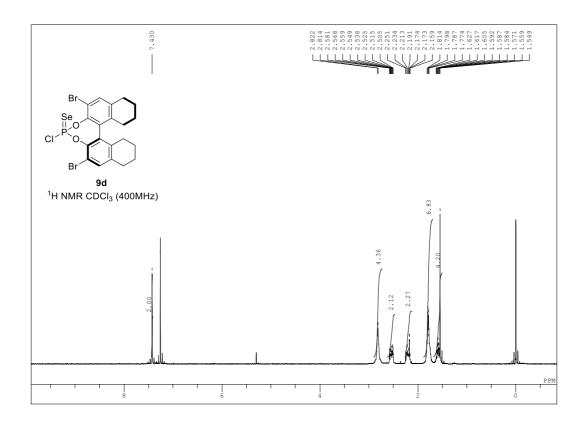


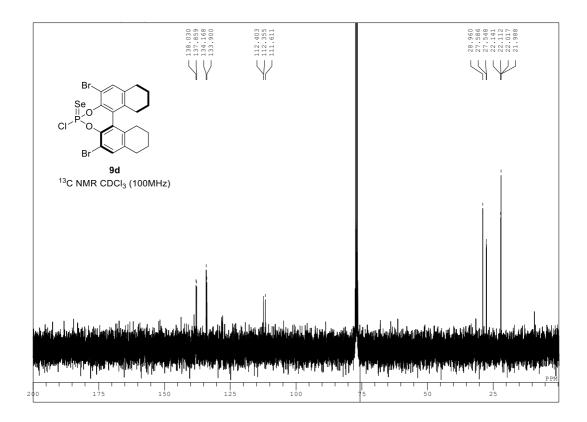


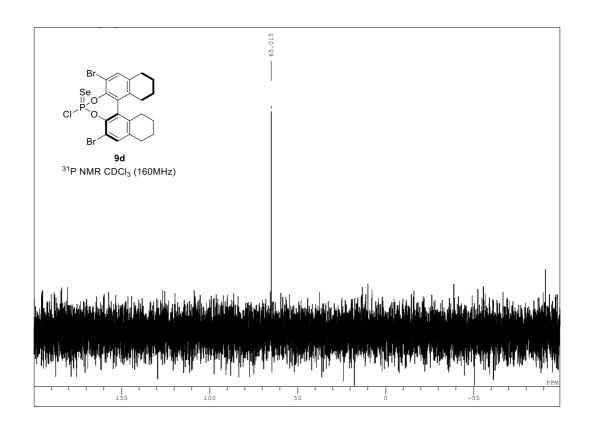


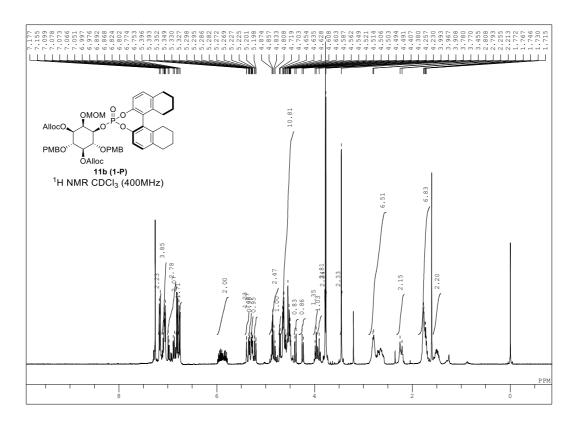


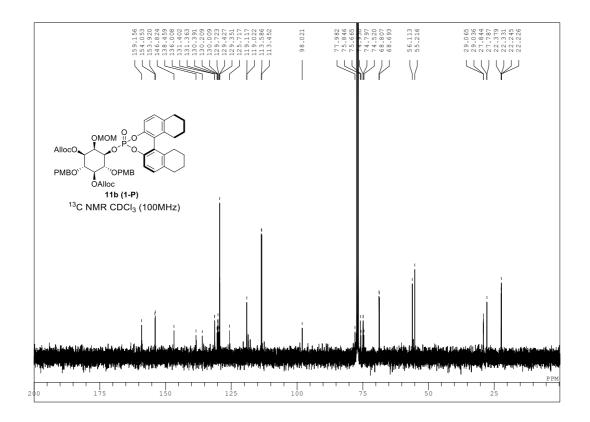


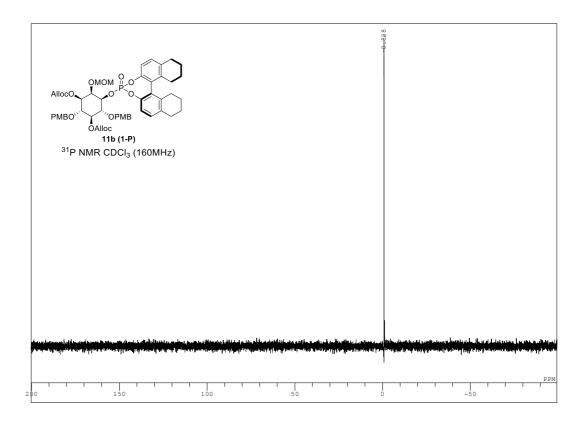


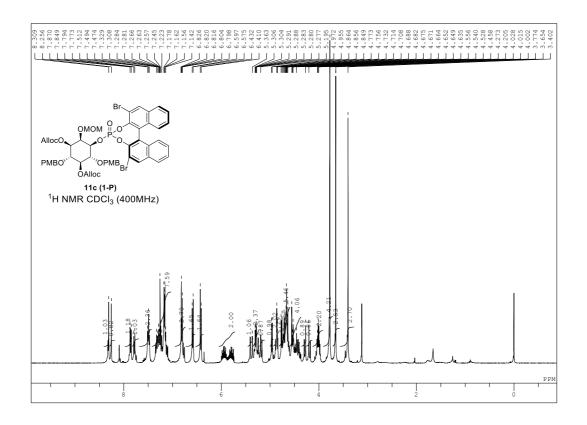


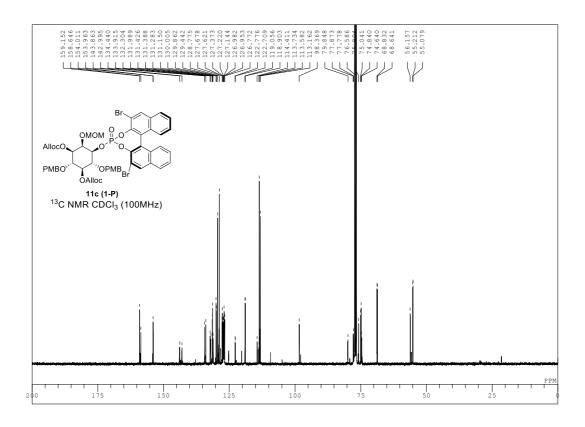


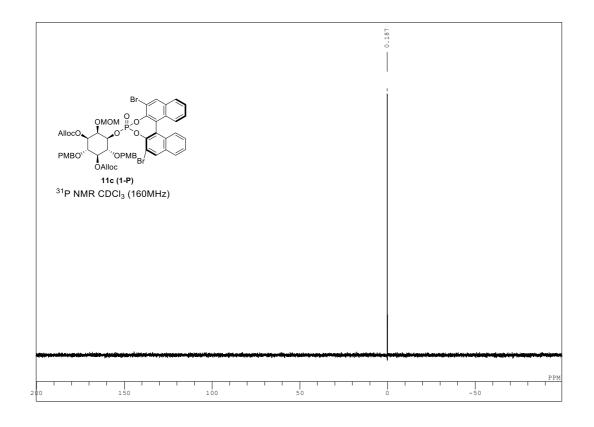


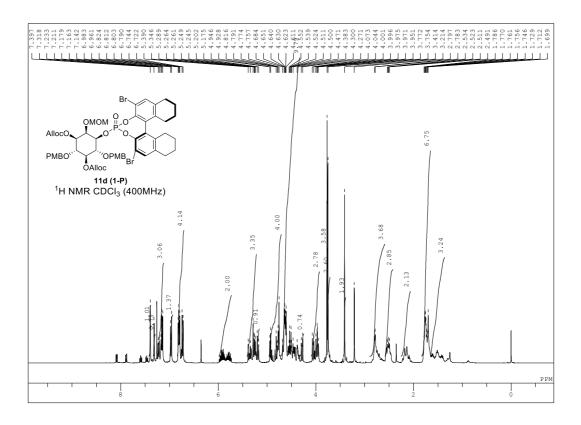


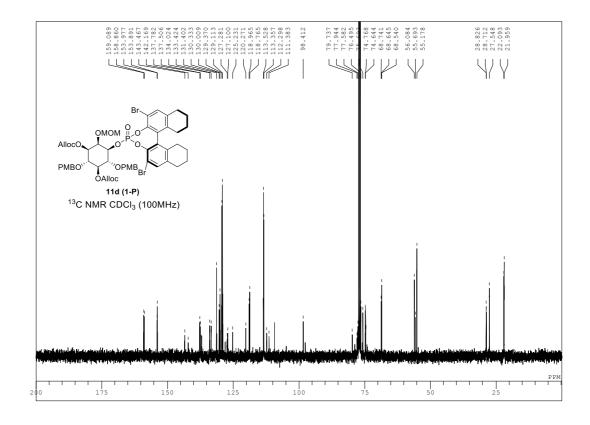


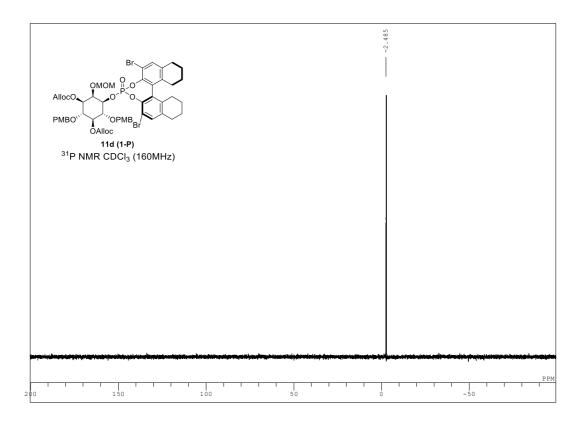


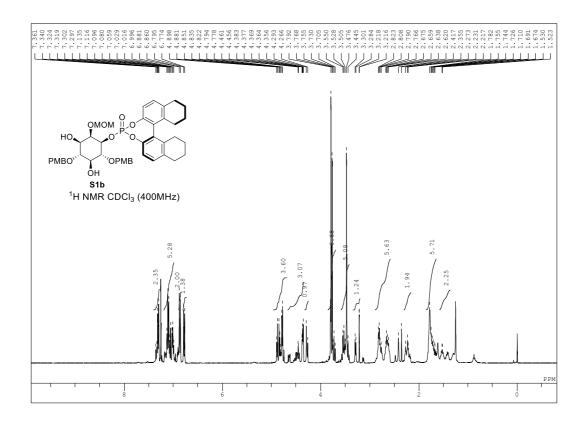


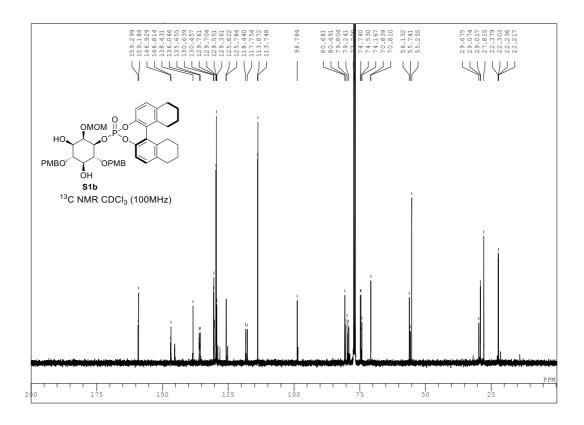


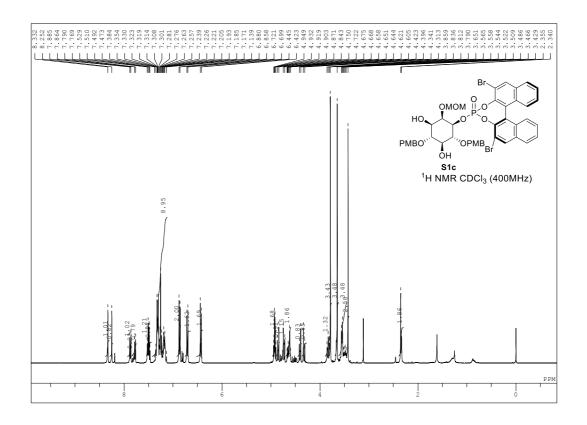


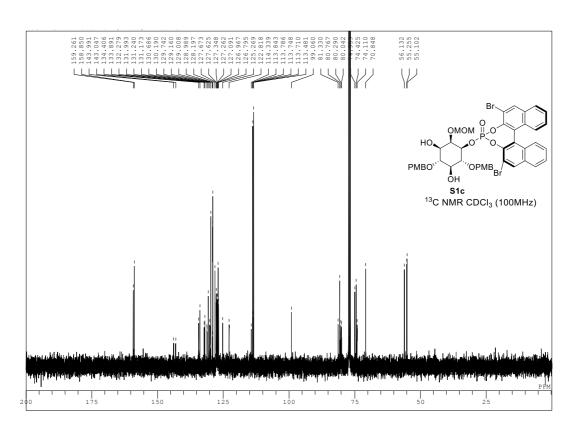


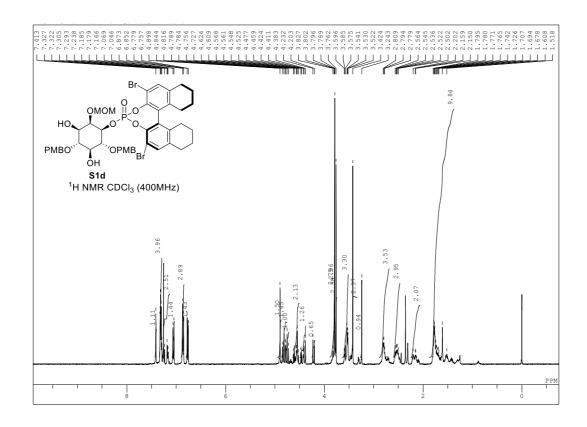


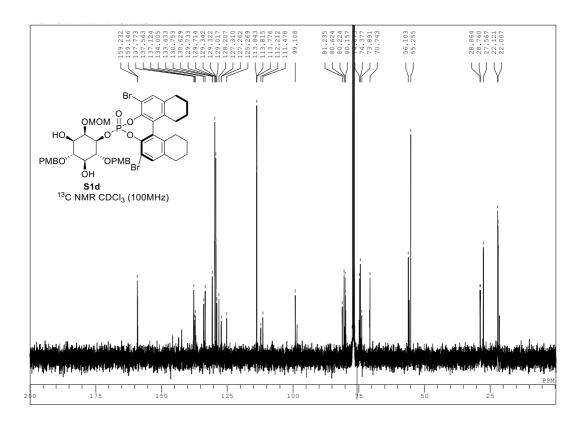


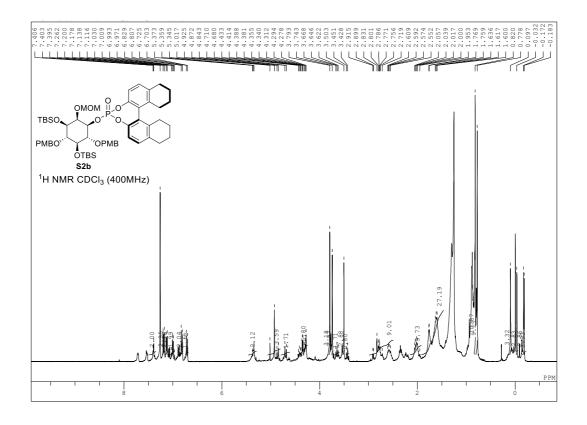


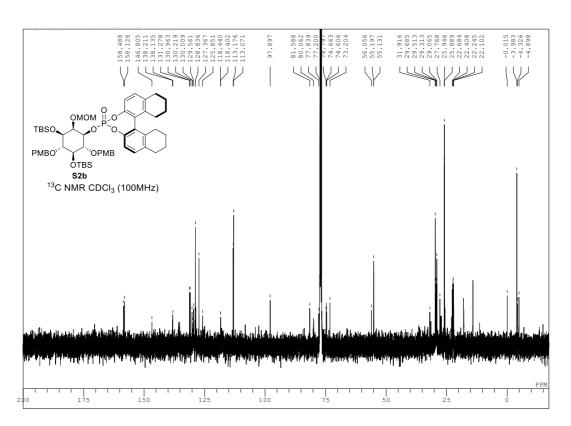


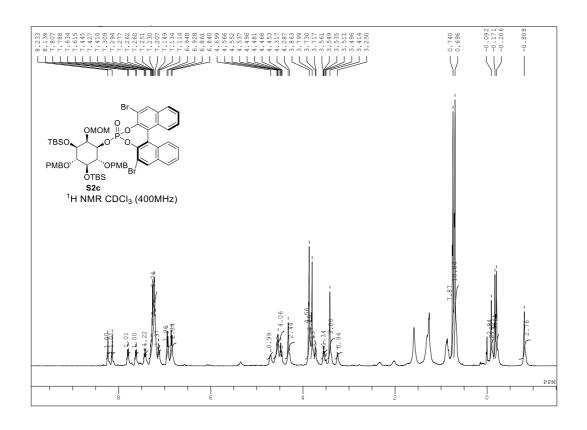


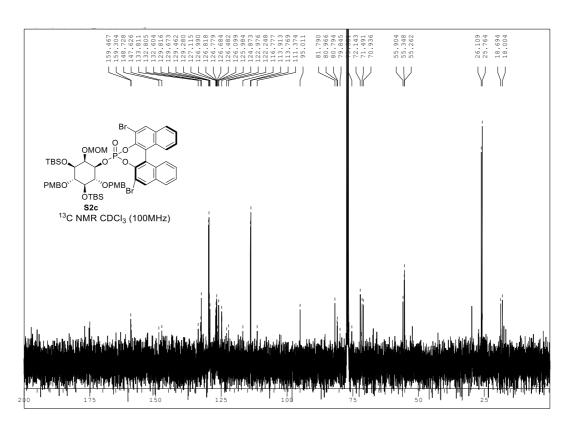


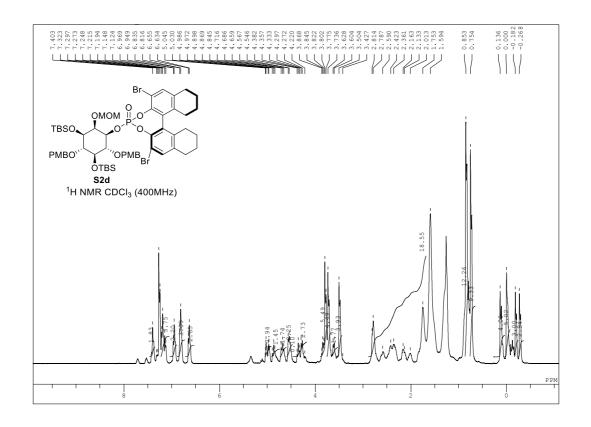


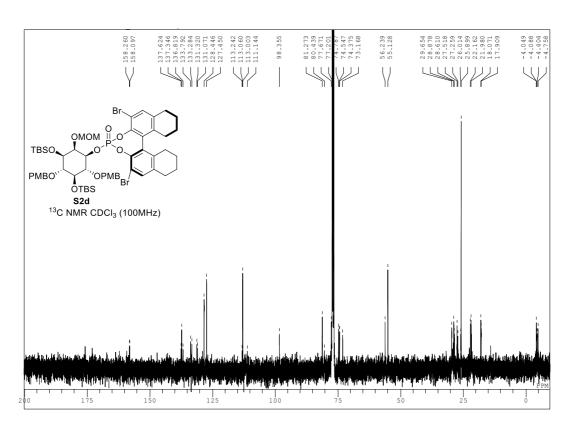


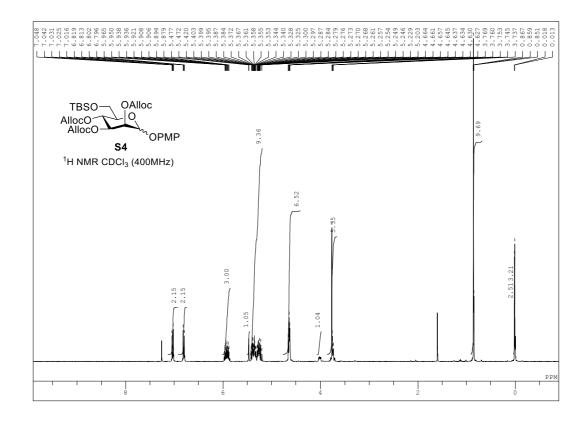


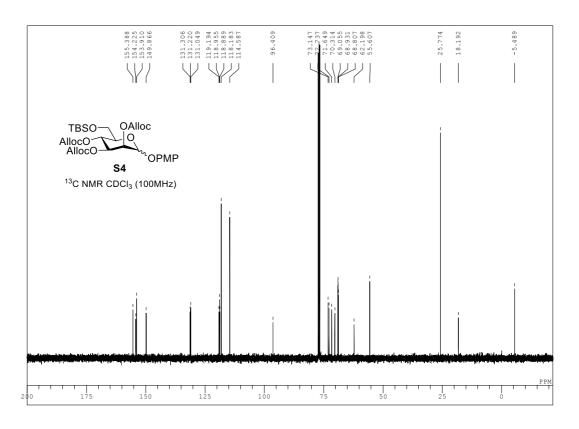


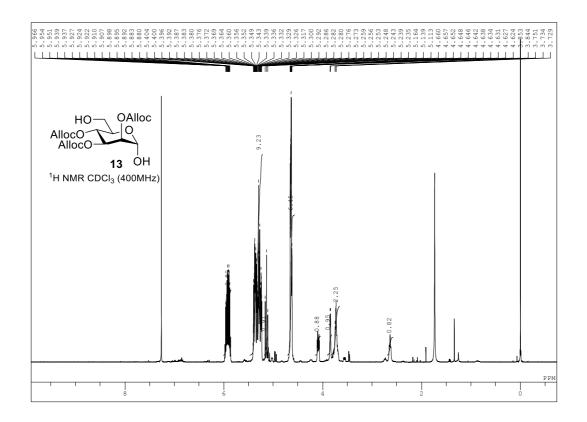


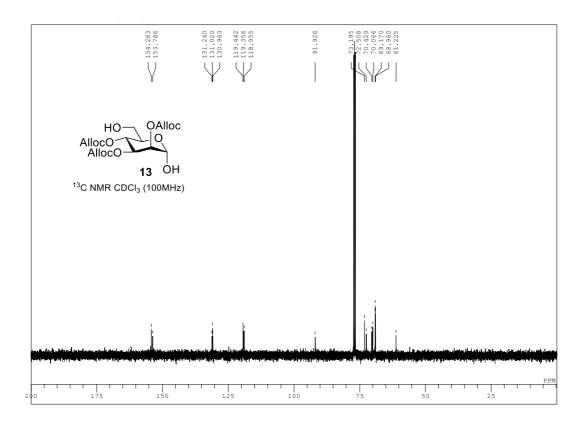


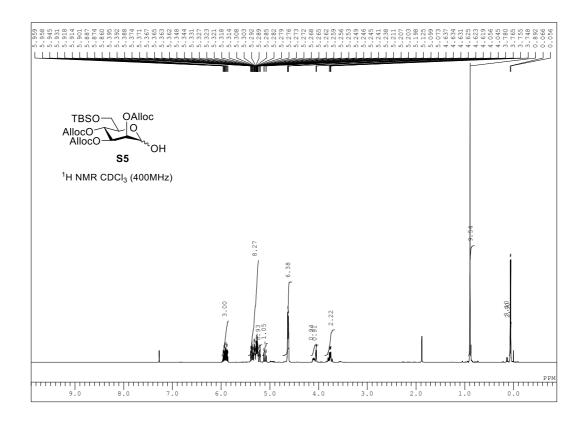


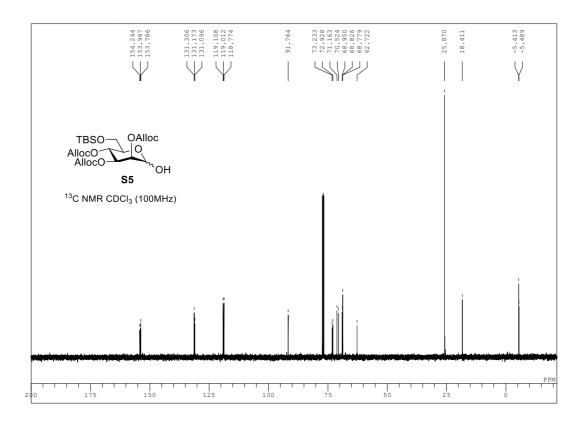


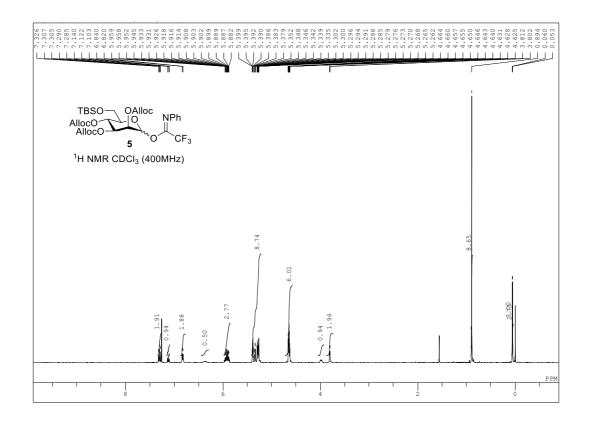


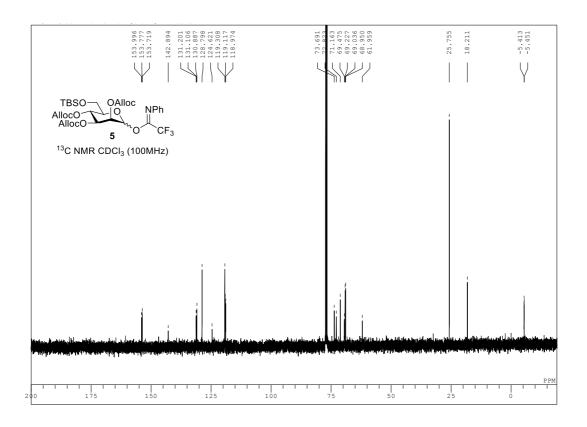


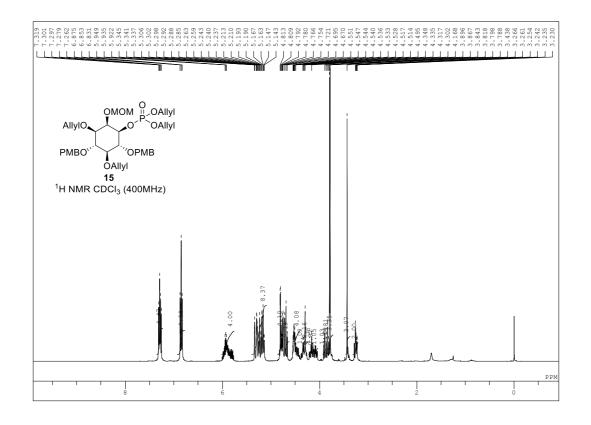


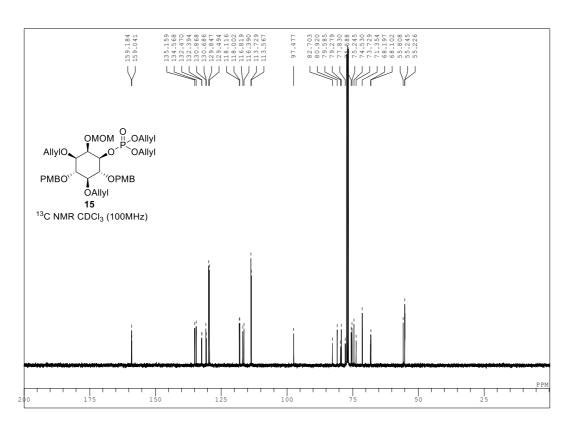


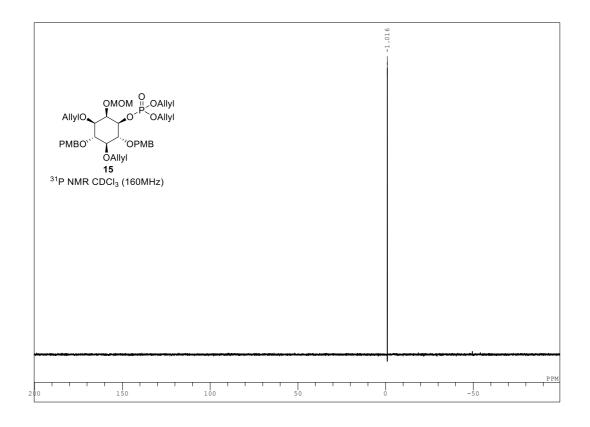


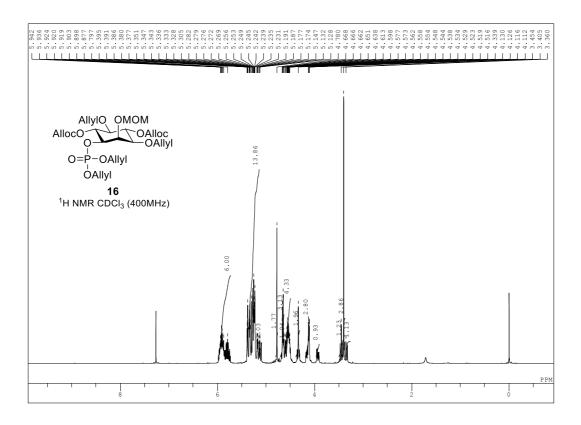


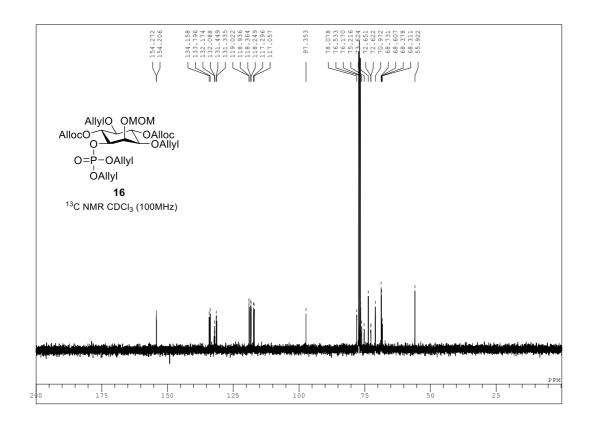


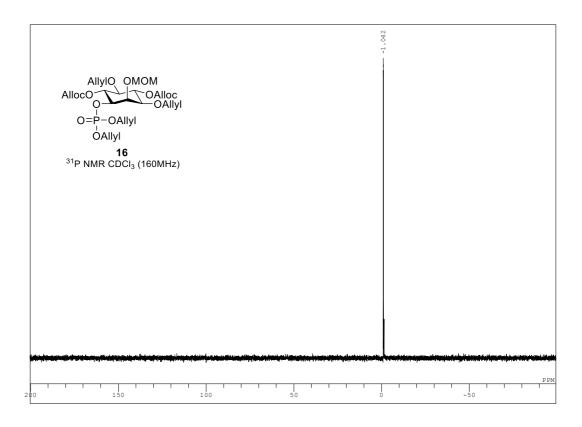


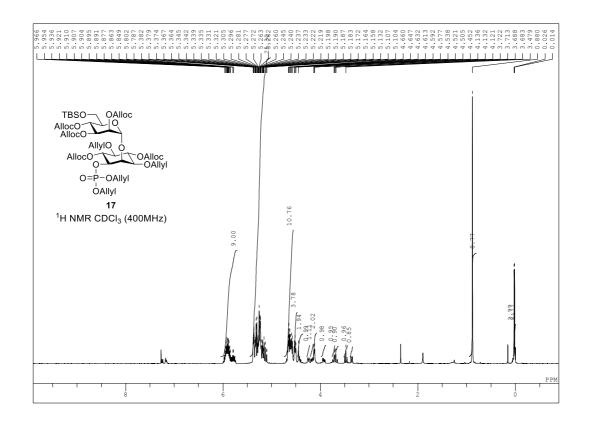


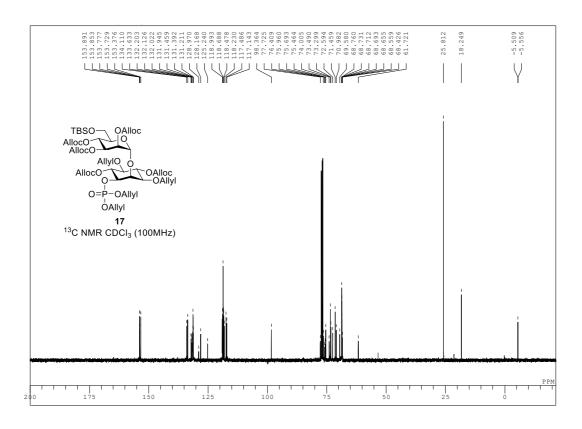


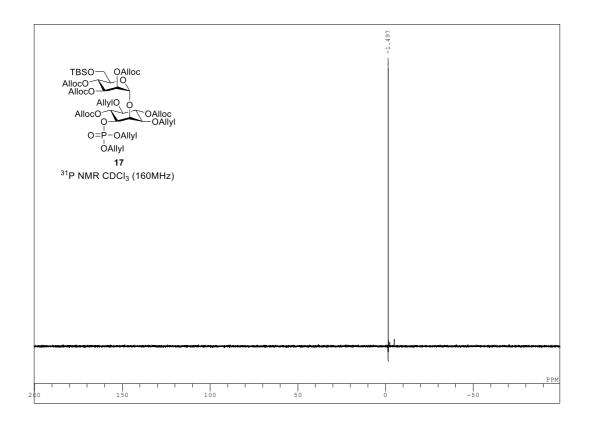


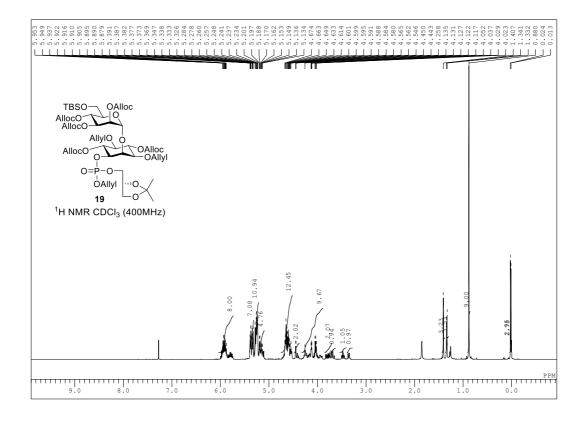


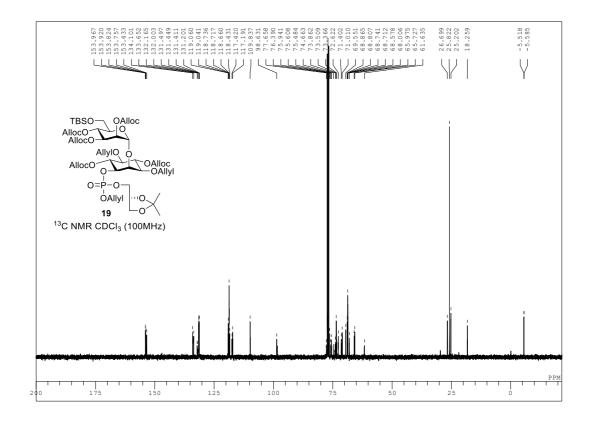


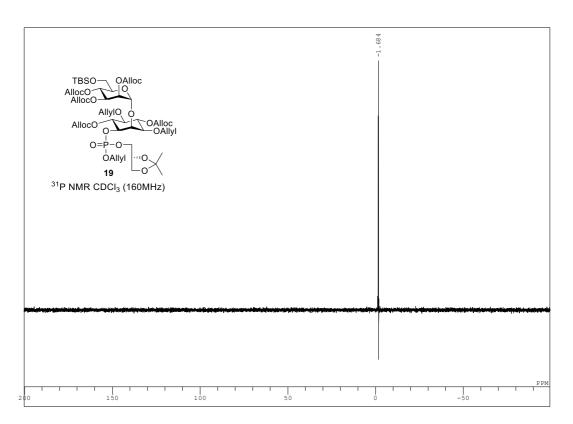


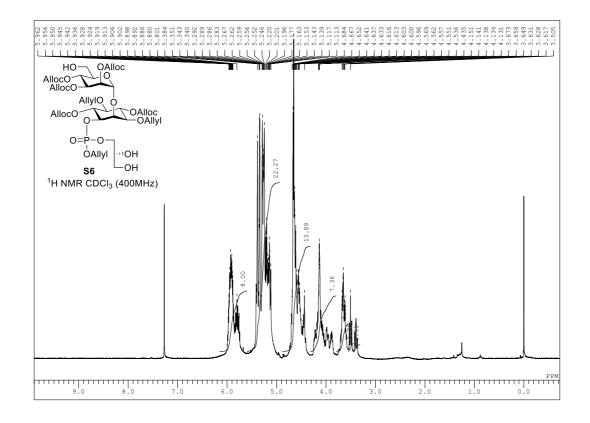


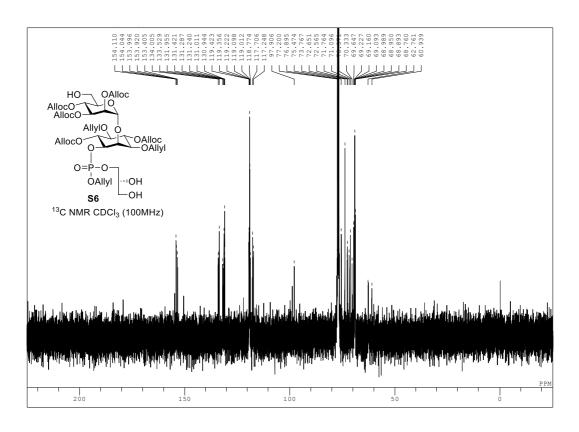


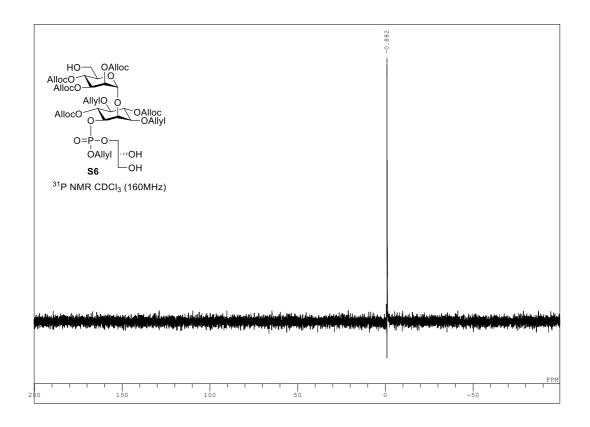


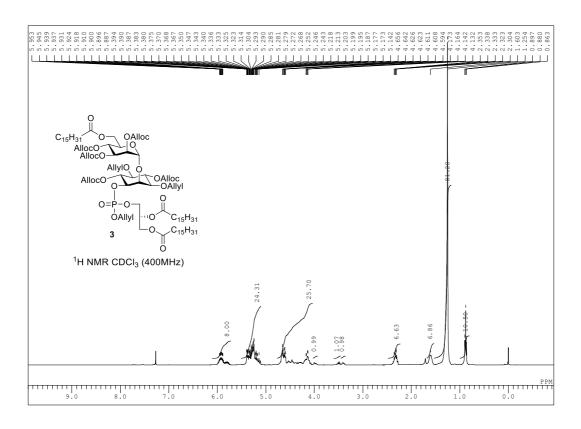


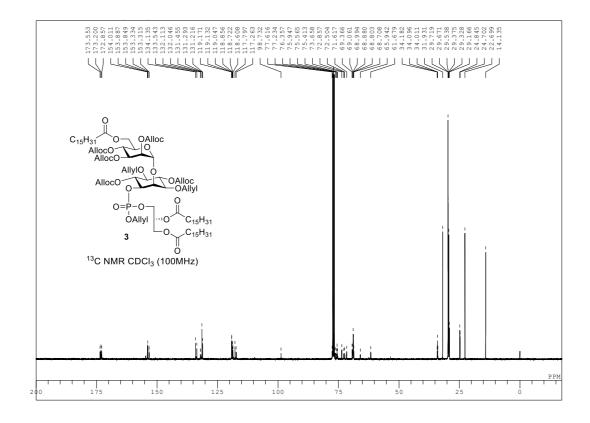


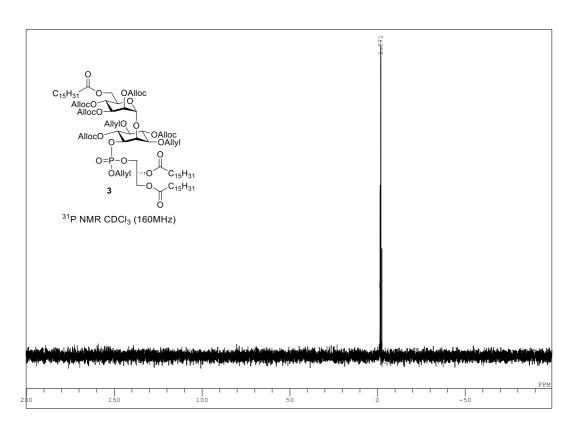


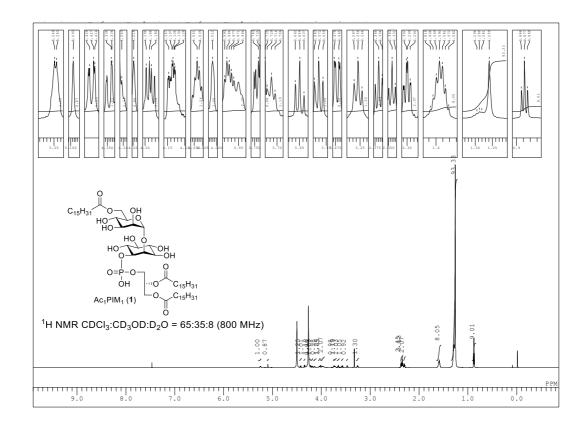


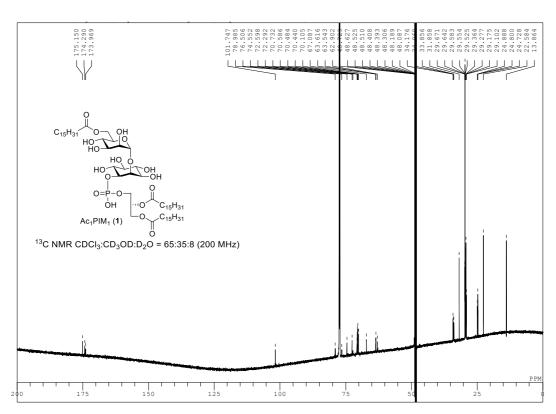


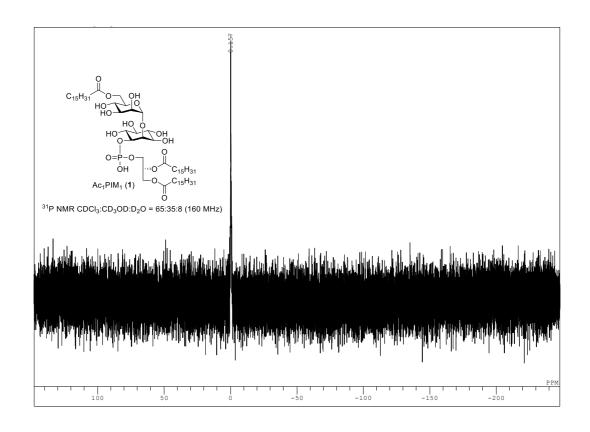


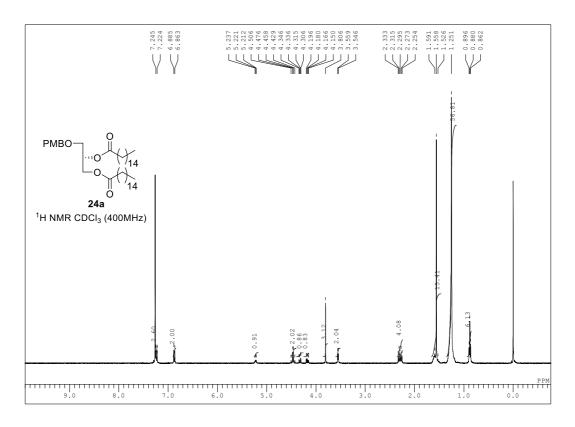


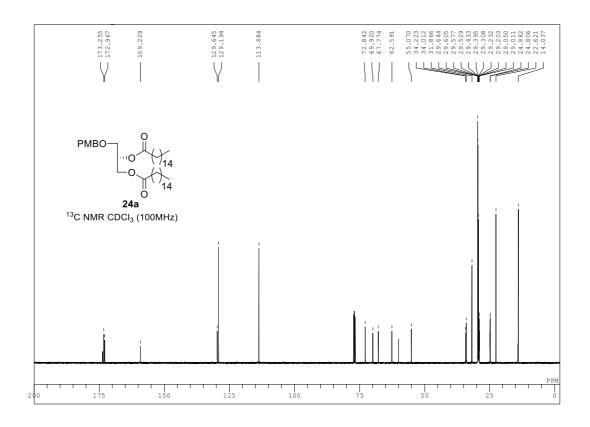


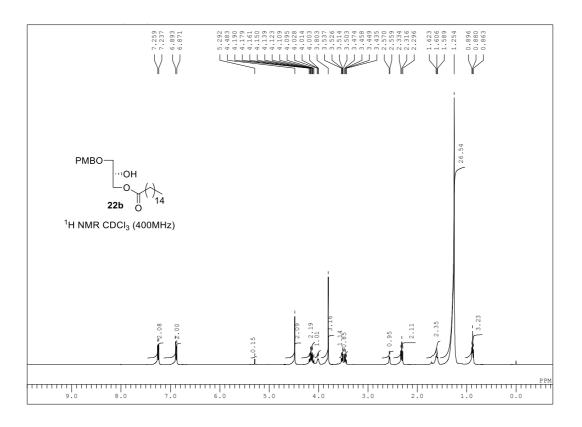


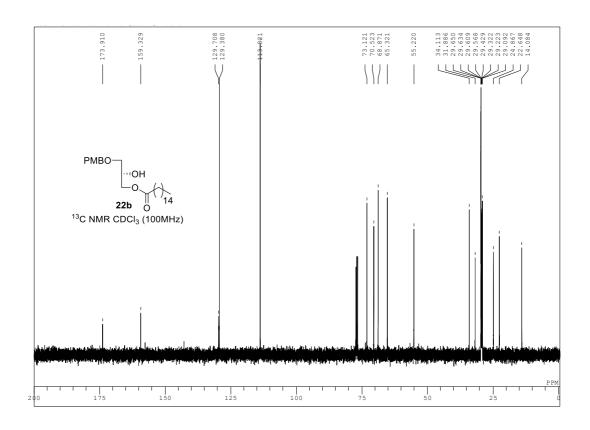


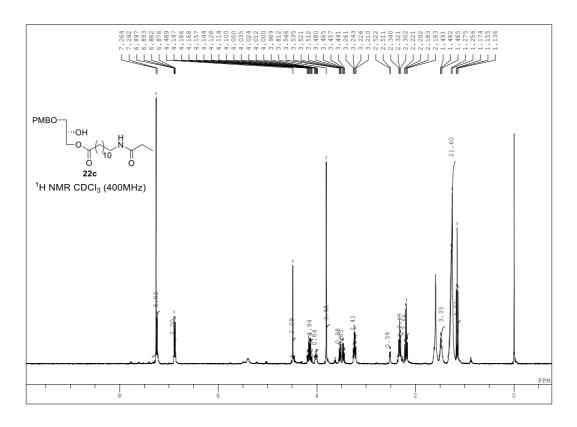


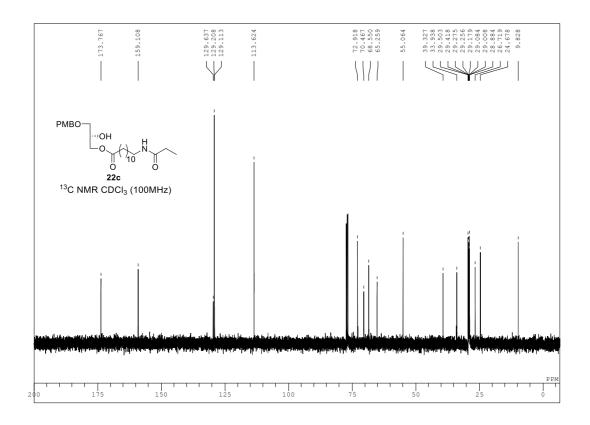


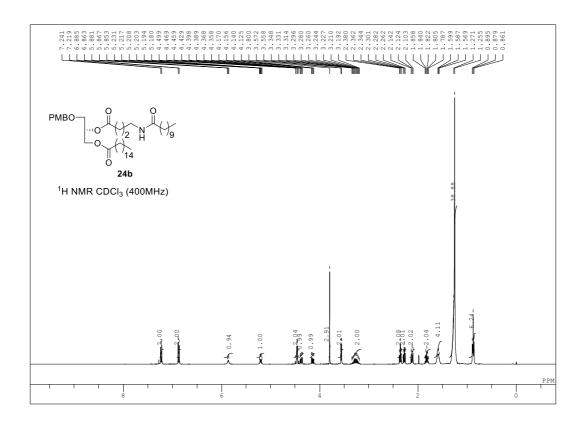


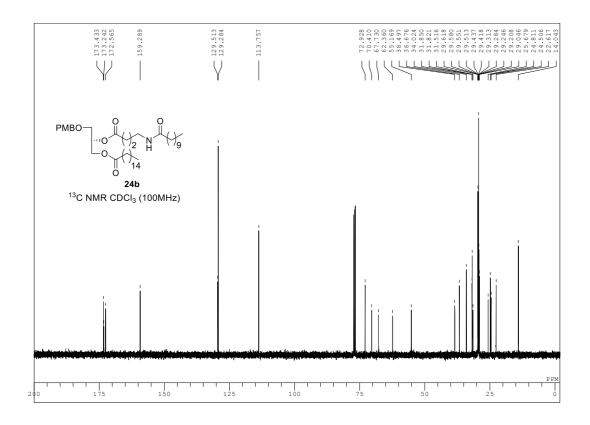


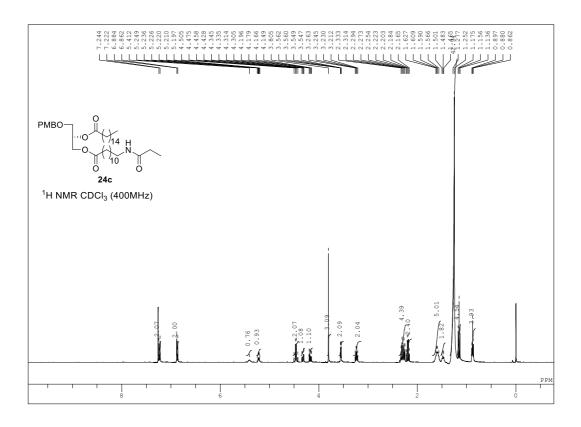


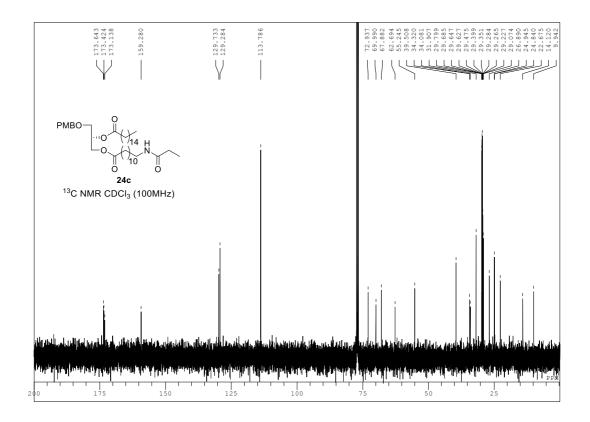


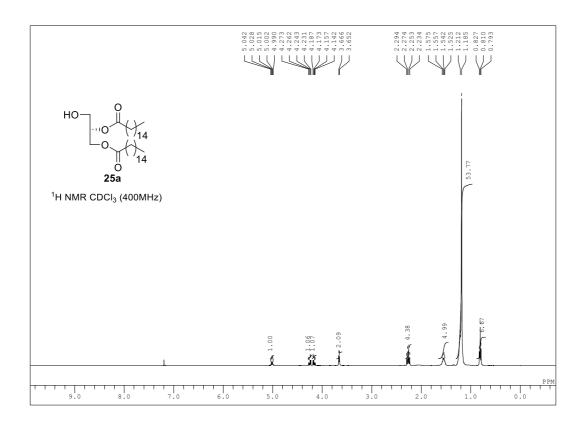


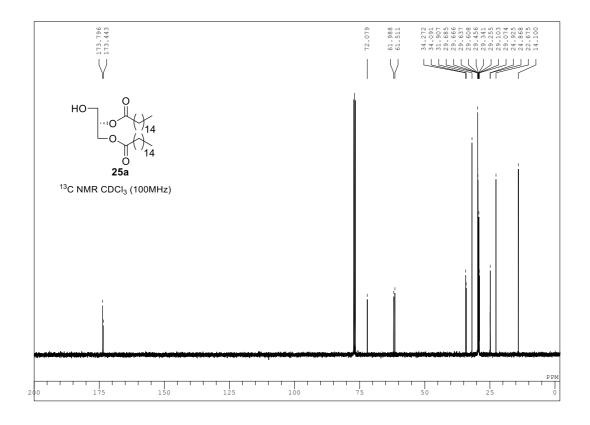


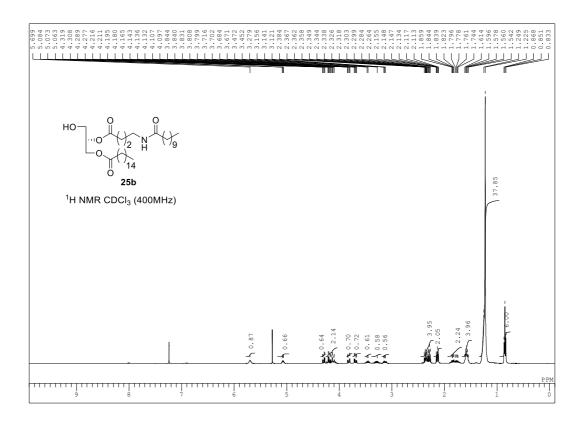


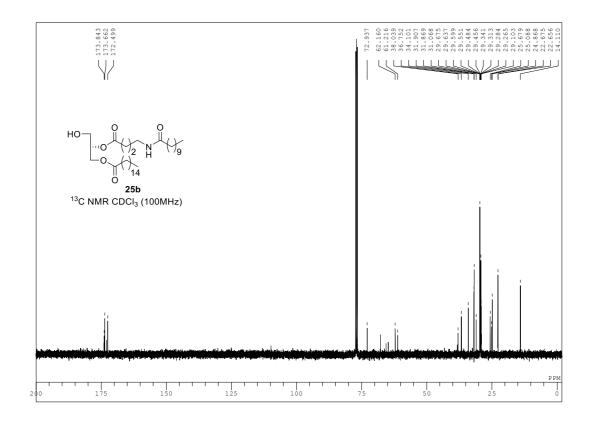


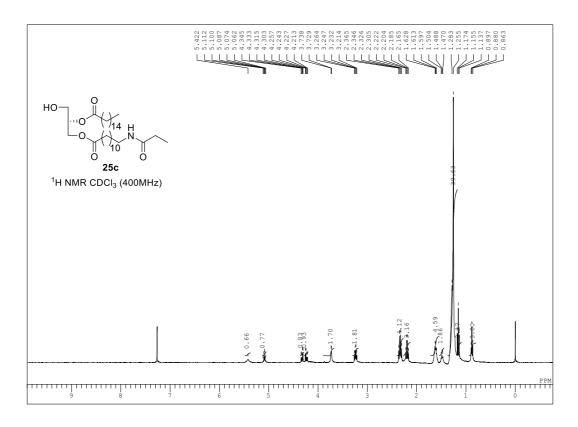


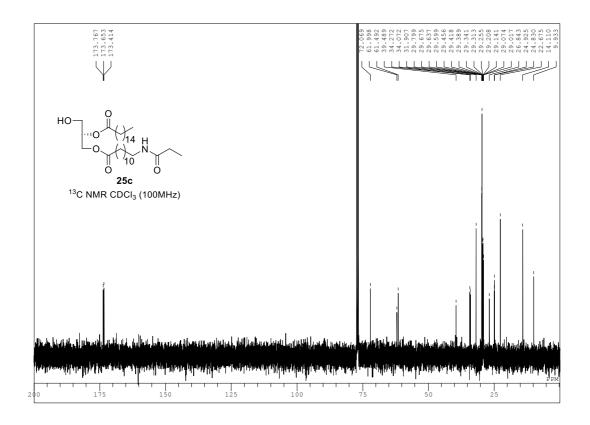


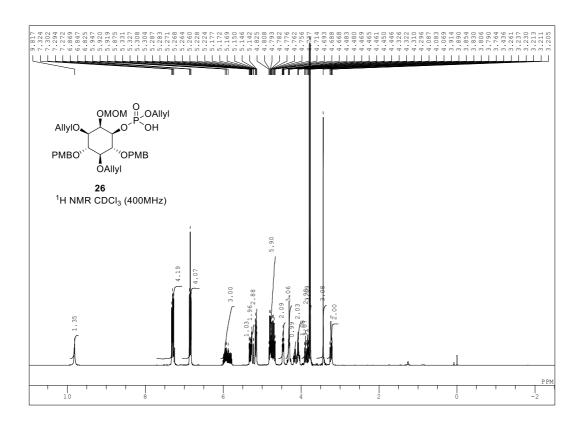


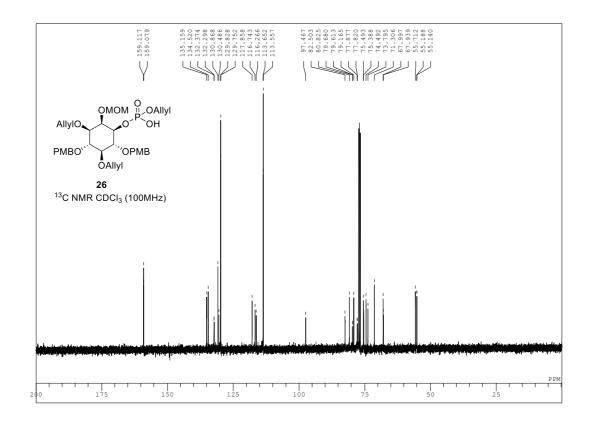


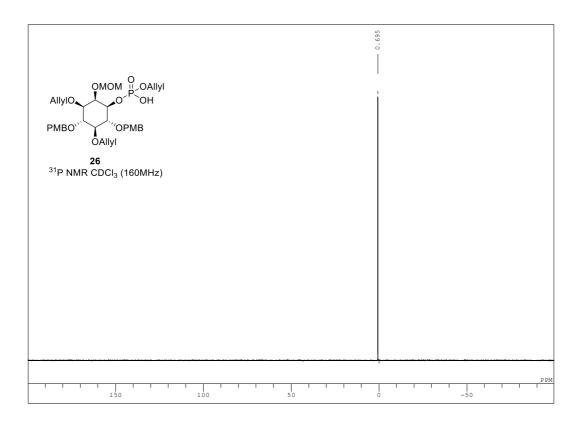


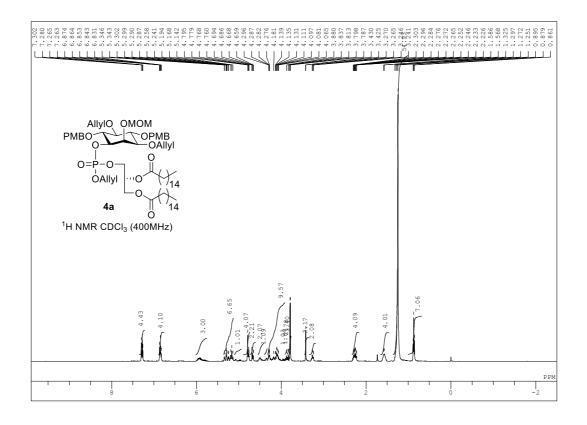


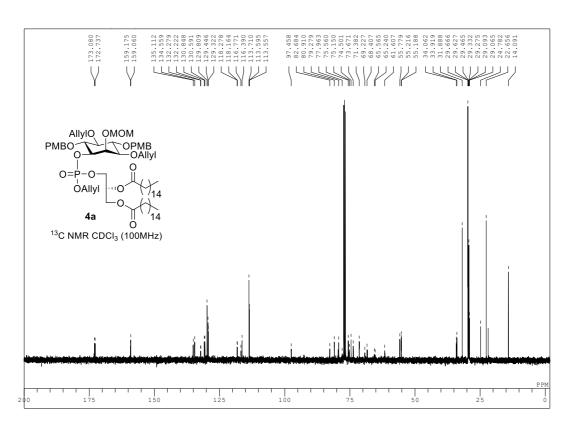


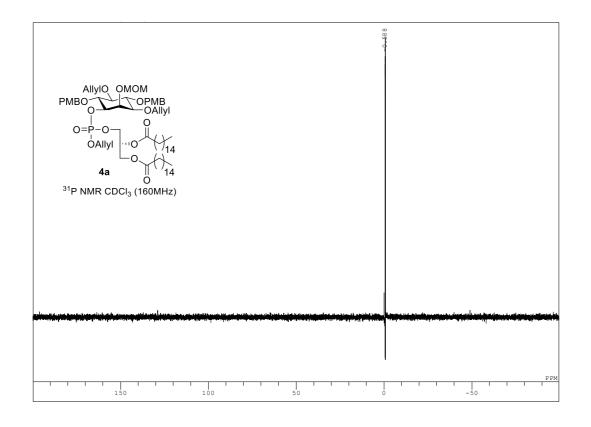


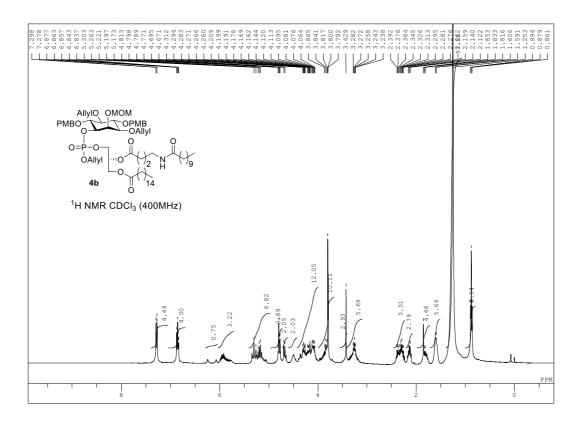


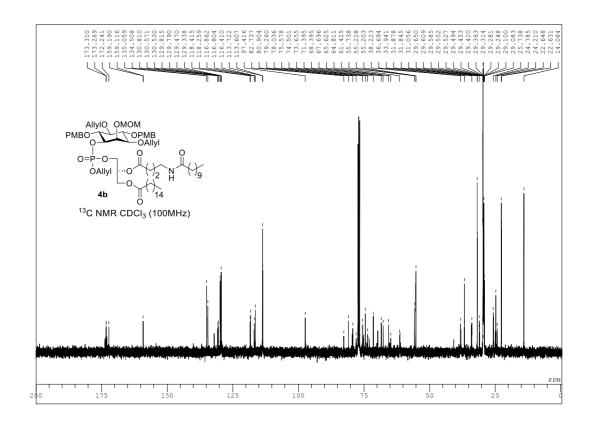


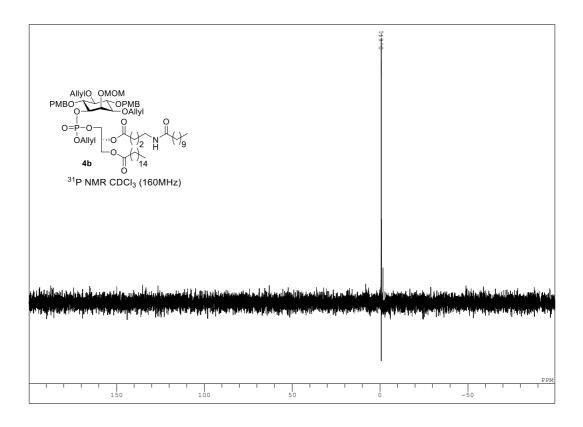


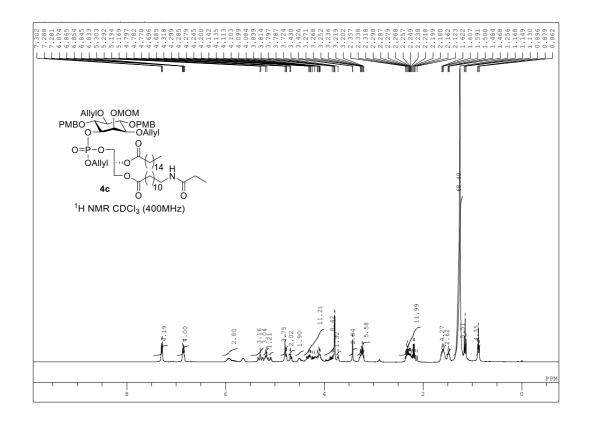


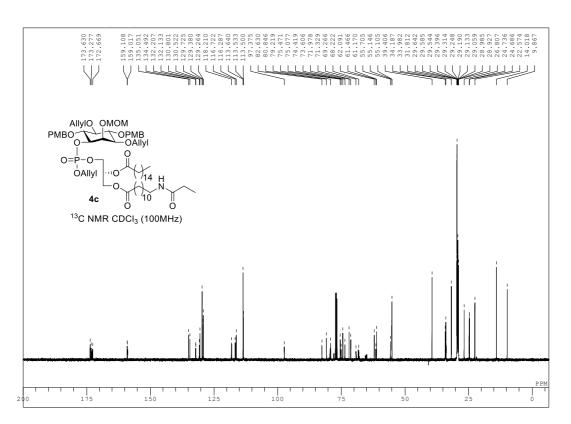


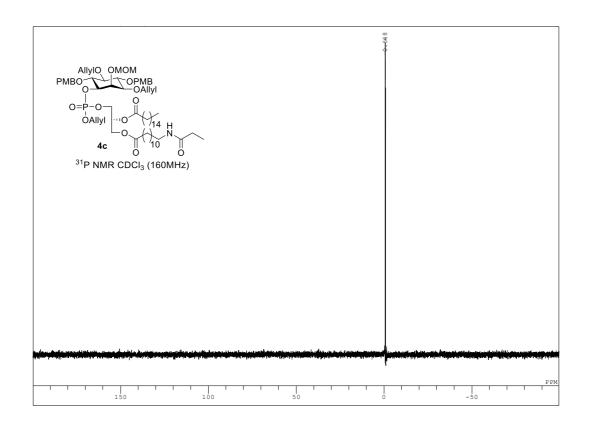


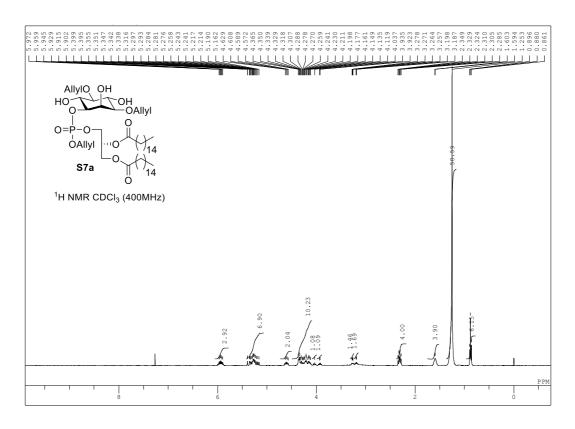


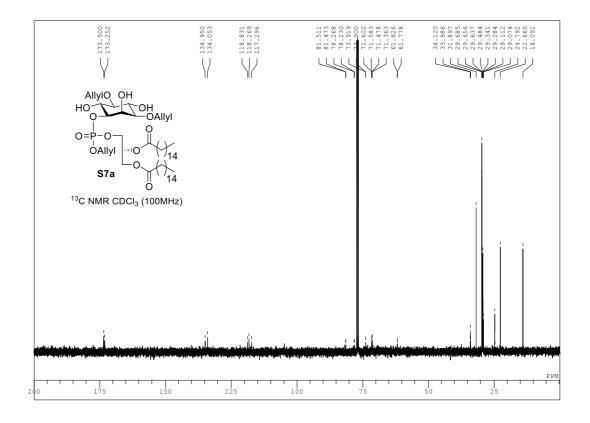


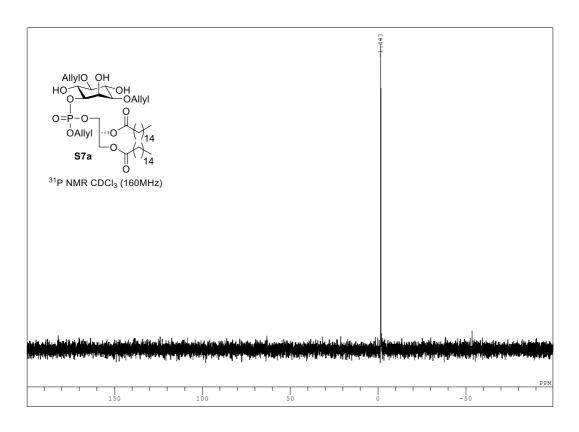


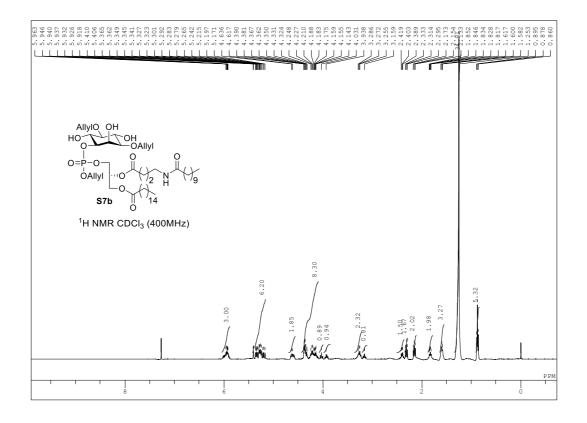


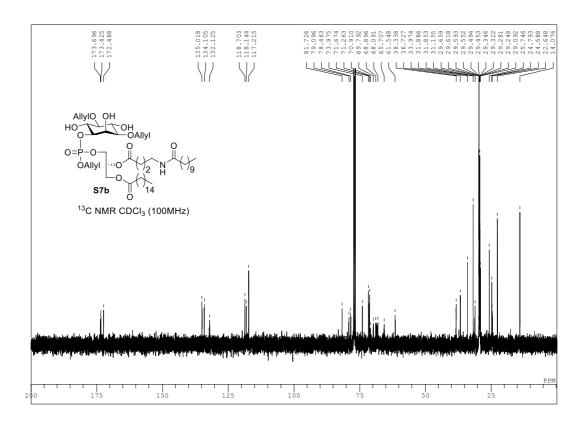


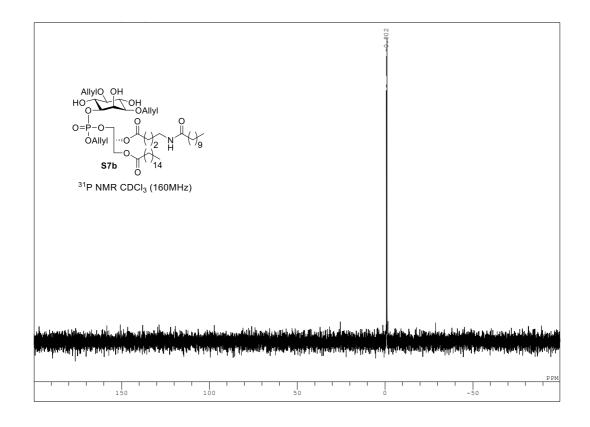


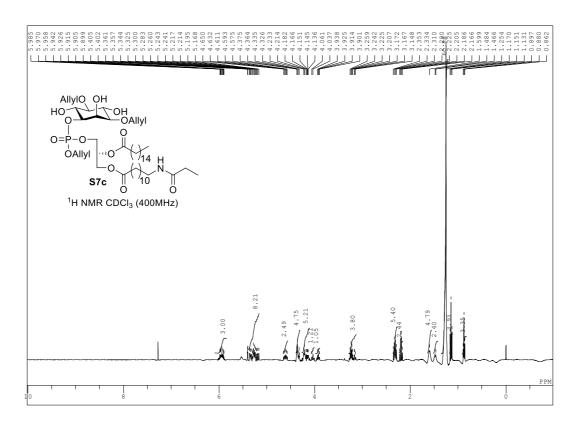


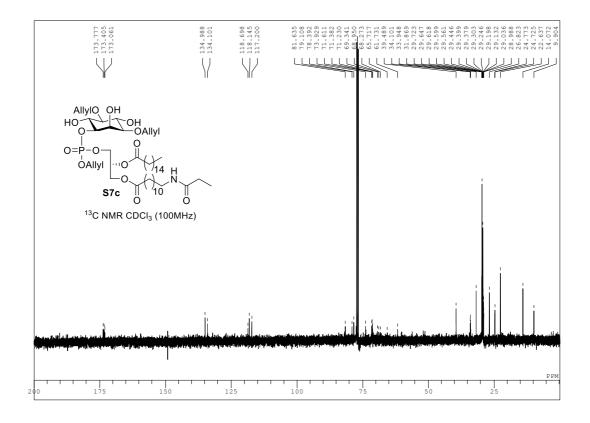


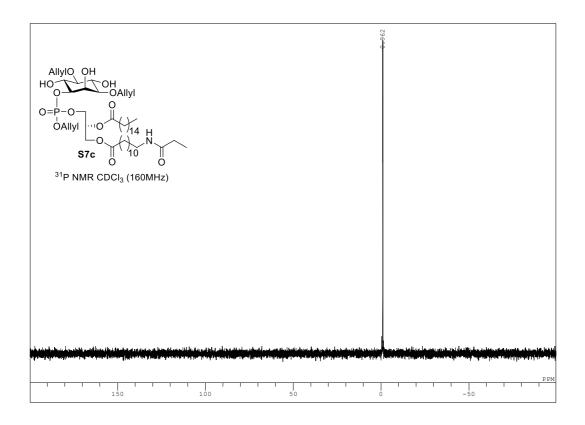


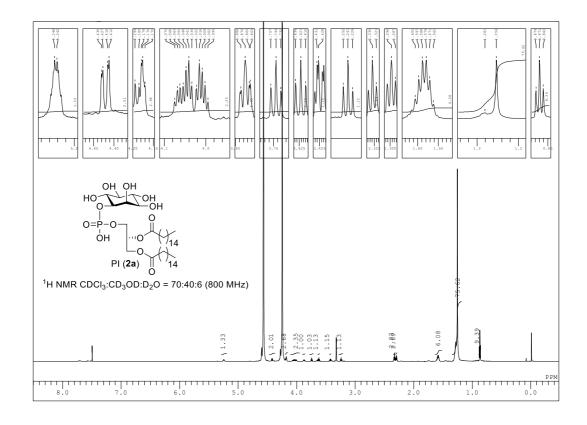


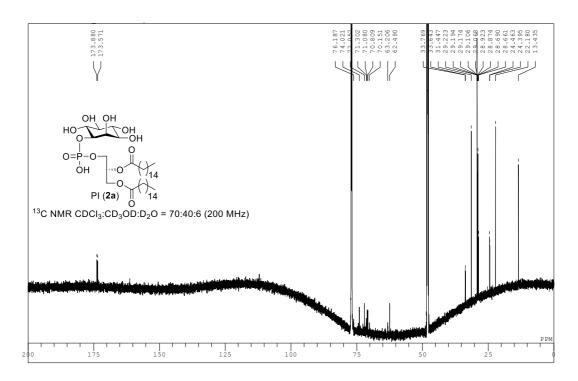


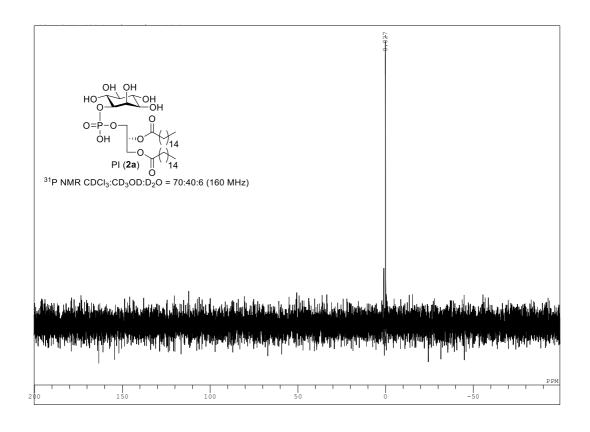


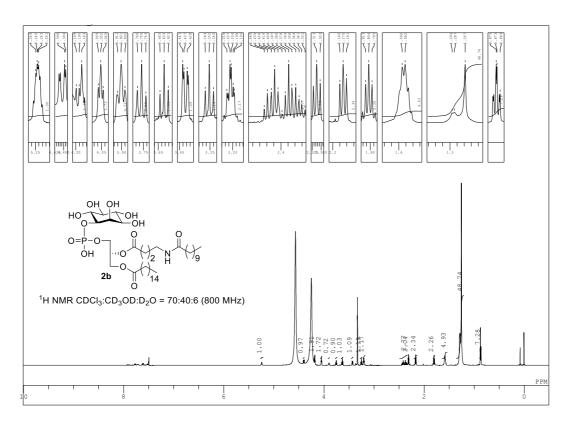


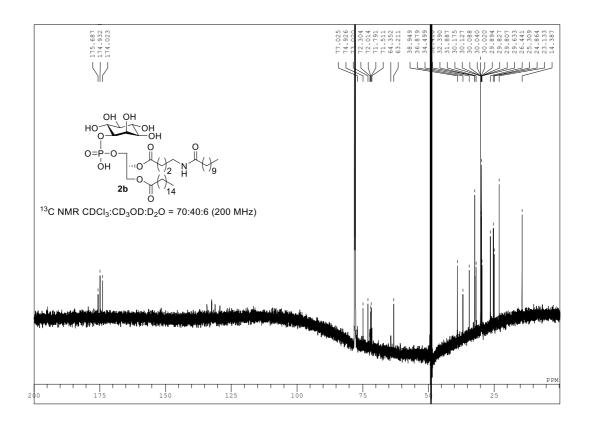


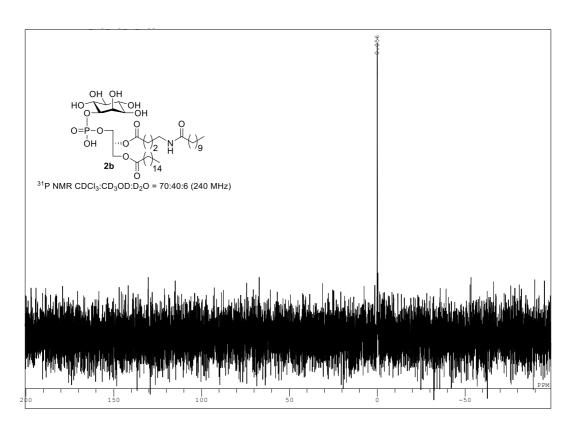


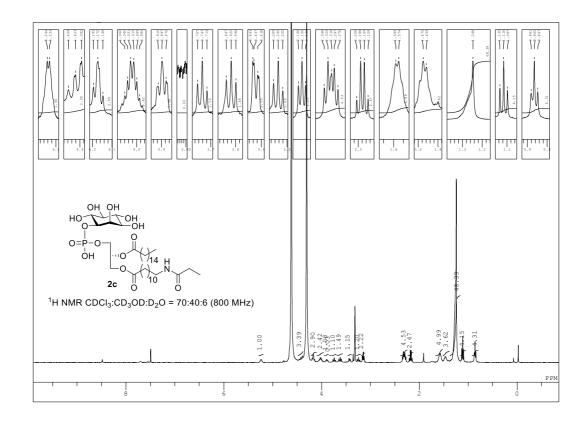


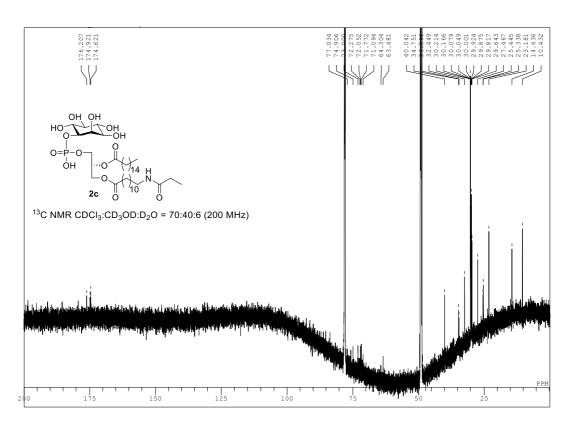


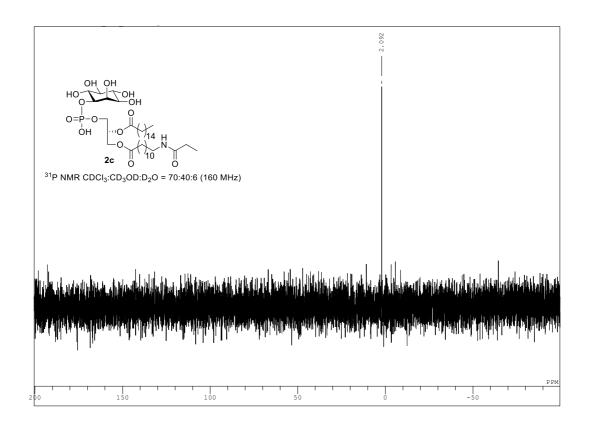












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