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Supporting information

Aldol sensors-inspired fluorescent probes for measuring protein citrullination

Huihong Chen, Hailong Zhao, Lingling Xiang, Haiting Wu, Yunshi Liang, Xin-An Huang,^{*} Jing Zhang^{*}

Artemisinin Research Center, Guangzhou University of Chinese Medicine, 12 Jichang Road, Guangzhou 510405, China.

Email: jingzhang@gzucm.edu.cn; <u>xinanhuang@gzucm.edu.cn</u>

Table of Contents

- 1. Supporting data
- 2. Experimental procedure
- 3. ¹H and ¹³C NMR spectra of compounds
- 4. Reference

• Supporting data

Compound	λ_{ex} (nm)	Quantum yield (Φ F)
Napdialyne	342	0.071
Napdialyne-Cit	319	0.916

Table S1. Quantum yields of Napdialyne and Napdialyne-Cit^{*a*}

^{*a*} The quantum yields of tested compounds were recorded in DMSO with 2-aminopyridine in 0.1 N H_2SO_4 as a standard (λ_{ex} =300 nm, ΦF =0.60).

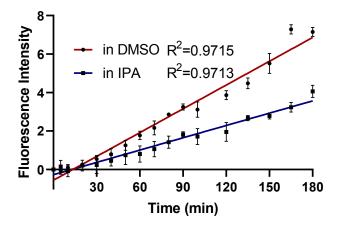


Figure S1. Fluorescence intensity change of reaction mixture over time. Reaction condition: Napdialyne (50 μ M), L-citrulline (250 μ M), 20% trichloroacetic acid in ddH₂O, 50 °C, 0-3 h. The reaction mixture was diluted 10-times with DMSO or IPA before measurement. For DMSO, $\lambda_{ex} = 320$ nm, $\lambda_{em} = 376$ nm; For IPA, $\lambda_{ex} = 317$ nm, $\lambda_{em} = 360$ nm. L-Cit: L-citrulline; IPA: isopropanol. n = 3.

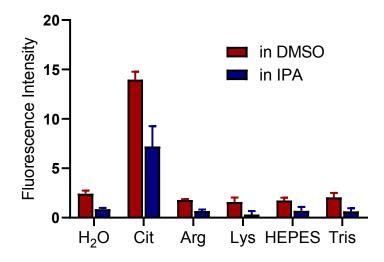


Figure S2. Fluorescence intensity of Napdialyne upon incubation with various analytes. Reaction condition: Napdialyne (50 μ M), analyte (250 μ M), 20% trichloroacetic acid in ddH₂O, 50 °C, 3 h. The reaction mixture was diluted 10-times with DMSO or IPA before measurement. For DMSO, $\lambda_{ex} = 320$ nm, $\lambda_{em} = 376$ nm; For IPA, $\lambda_{ex} = 317$ nm, $\lambda_{em} = 360$ nm. Cit: L-citrulline; Arg: L-arginine; Lys: L-lysine; IPA: isopropanol. n = 2.

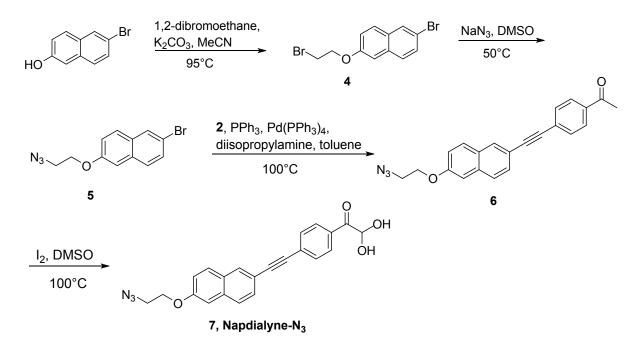


Figure S3. Synthetic routine to probe Napdialyne-N₃.

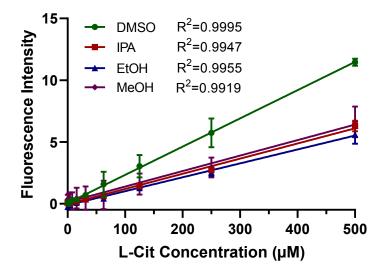


Figure S4. Dose-dependent relationship of fluorescence intensity of Napdialyne-N₃ upon incubation with different concentrations of L-citrulline. Reaction condition: Napdialyne-N₃ (50 μ M), L-citrulline (0.5 to 0 mM, 2-fold dilution), 20% trichloroacetic acid in ddH₂O, 50 °C, 3 h. The reaction mixture was diluted 10-times with different solvents before measurement. For DMSO, $\lambda_{ex} = 320$ nm, $\lambda_{em} = 376$ nm; For IPA, $\lambda_{ex} = 317$ nm, $\lambda_{em} = 360$ nm; For ethanol, $\lambda_{ex} = 315$ nm, $\lambda_{em} = 362$ nm; For methanol, $\lambda_{ex} = 318$ nm, $\lambda_{em} = 360$ nm. L-Cit: L-citrulline; IPA: isopropanol. n = 2.

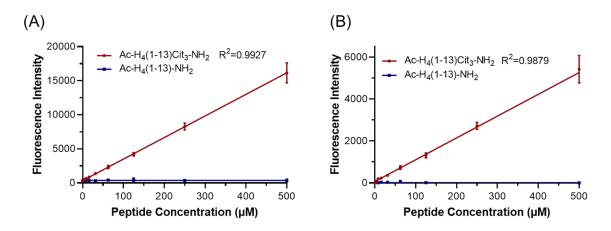


Figure S5. Dose-dependent relationship on fluorescence intensity of probe Napdialyne (A) or probe Napdialyne-N₃ (B) upon incubation with different concentrations of peptides. Reaction condition: probe (50 μ M), peptide (0.5 to 0 mM, 2-fold dilution), 20% trichloroacetic acid in ddH₂O, 50 °C, 3 h. The reaction mixture was diluted 10-times with DMSO before measurement. For Napdialyne, $\lambda_{ex} = 320$ nm, $\lambda_{em} = 376$ nm; For Napdialyne-N₃, $\lambda_{ex} = 317$ nm, $\lambda_{em} = 360$ nm; n = 3. Note that this data were collected on BioTek CytationTM 5 imaging reader.

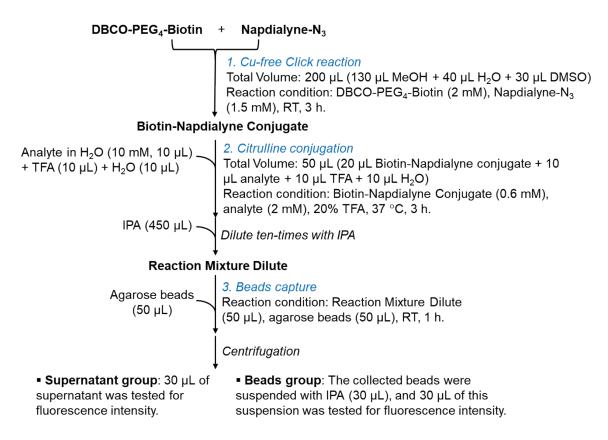


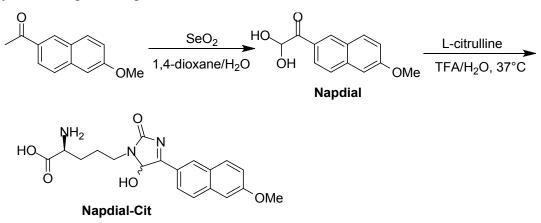
Figure S6. Work-flow of the three-step capture assay by probe Napdialyne-N₃.

• Experimental procedure

Methods and materials

All chemicals were used as received unless otherwise stated. L-citrulline was obtained from Bide Pharmatech Ltd. (Shanghai, China). The Peptides were custom-synthesized from GL Biochem Ltd. (Shanghai, China). The streptavidin agarose resin (catalog No. 20347) was purchased from Thermo Scientific (USA). The 96-well plate (black, flat, not treated) was purchased from Corning® (USA). ¹H NMR and ¹³C NMR spectra were measured on a Bruker 400 NMR spectrometer. Proton chemical shifts of NMR spectra were calibrated with TMS as internals reference. ESI-MS and HRMS spectral data were recorded on Quadrupole LC/MS 6120 and TripleTOFTM 5600⁺. Fluorescence analysis was performed on Thermo ScientificTM VarioskanTM LUX microplate reader unless otherwise stated.

1. Synthesis of probe Napdial

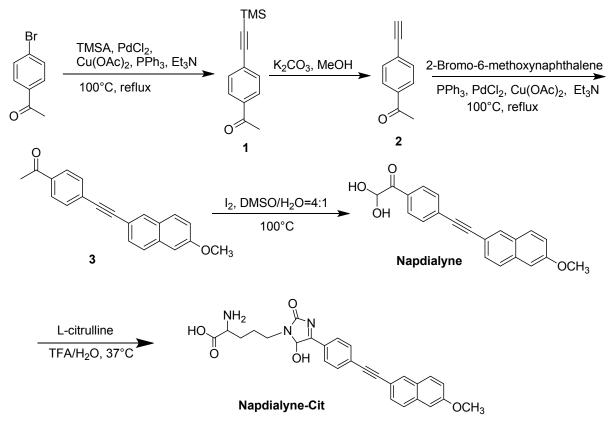


A solution of selenium dioxide (278 mg, 2.5 mmol) in 1,4-dioxane (5 mL) and H₂O (750 μL) was heated to 55°C until the selenium dioxide was dissolved. Then 2-acetyl-6-methoxunaphthalene (500 mg, 2.5 mmol) was added in. The reaction mixture was refluxed overnight. After cooling, the precipitate was filtered off, and the filtrate was concentrated. The crude was purified by silica gel column chromatography (petroleum ether : ethyl acetate = 10 : 1), and compound **Napdial** was obtained as a yellow solid (370 mg, 70% yield). ¹H NMR (400 MHz, CDCl₃) δ 9.75 (s, 0.5H), 8.81 (s, 0.5H), 8.63 (s, 0.5H), 8.11 (t, *J* = 8.4 Hz, 1H), 7.90 (d, *J* = 8.8 Hz, 1H), 7.82 (d, *J* = 8.8 Hz, 1H), 7.24 (dd, *J* = 2.4, 8.8 Hz, 1H), 7.17 (t, *J* = 2.5 Hz, 1H), 6.09 (s, 0.5H), 3.97 (s, 3H); ¹³C NMR (101 MHz, d₆-DMSO) δ 196.33, 193.78, 190.55, 188.11, 160.58, 160.14, 159.94, 138.05, 137.63, 137.45, 133.51, 132.19, 131.92, 131.80, 129.32, 128.98, 127.90, 127.73, 127.66, 127.43, 127.28, 125.84, 125.61, 125.53, 120.28, 119.95, 106.75, 106.53, 91.25, 89.57, 56.00, 55.89; HRMS: (ESI) [M+H-H₂O]⁺ calc. for C₁₃H₁₁O₃, 215.0708, observed 215.0707.

To a solution of L-citrulline (41 mg, 0.23 mmol) in H₂O/TFA (1:1, 4 mL) was added Napdial (50 mg, 0.23 mmol). After being stirred under 37 °C for 10 hours, the reaction mixture was concentrated. The obtained crude was purified by reverse-phase silica gel column chromatography (H₂O : MeOH = 2 : 1) to afford **Napdial-Cit** as a white solid (26 mg, 43% yield). ¹H NMR (400 MHz, d₆-DMSO) δ 7.87 (d, *J* = 8.4 Hz, 2H), 7.80 (s, 1H), 7.35 (d, *J* =

2.4 Hz, 1H), 7.28 (d, J = 8.8 Hz, 1H), 7.23 (dd, J = 2.4, 9.2 Hz, 1H), 5.26 (d, J = 3.6 Hz, 1H), 3.88 (s, 3H), 3.59 (m, 2H), 2.71 (m, 1H), 1.67 (m, 4H); ¹³C NMR (101 MHz, d₆-DMSO) δ 173.1, 171.2, 158.2, 157.1, 135.0, 129.9, 129.1, 128.7, 128.3, 127.8, 125.4, 119.7, 106.4, 64.3, 64.2, 55.7, 53.1, 52.8, 28.1, 27.9, 23.4, 23.0; HRMS: (ESI) [M+H]⁺ calc. for C₁₉H₂₂N₃O₅, 372.1559, observed 372.1562.

2. Synthesis of probe Napdialyne



The synthesis of probe Napdialyne was based on a combination of previous reports with modifications.^{1, 2}

A solution of 4'-bromoacetophenone (1.0 g, 5.02 mmol), triphenylphosphine (65 mg, 0.25 mmol), palladium chloride (9 mg, 0.05 mmol), cupric acetate (9.1 mg, 0.05 mmol), trimethylsilylacetylene (TMSA, 1078 μ L, 7.63 mmol) in trimethylamine (25 mL) was refluxed under 100°C for 1.5 h. After cooling, the reaction mixture was mixed with water, and extracted with CH₂Cl₂ twice. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated on a rotary evaporator. The obtained crude was further purified by silica gel column chromatography (petroleum ether : ethyl acetate = 30 : 1) to yield compound **1** as a yellow oil (1.087 g, quantitative yield). ¹H NMR (400 MHz, CDCl₃) δ 7.90 (d, *J* = 8.4 Hz, 2H), 7.55 (d, *J* = 8.4 Hz, 2H), 2.60 (s, 3H), 0.26 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 197.32, 136.52, 132.20, 128.27, 128.09, 104.20, 98.22, 26.73, 0.00; HRMS: (ESI) [M+H]⁺ calc. for C₁₃H₁₇OSi, 217.1049, observed 217.0987.

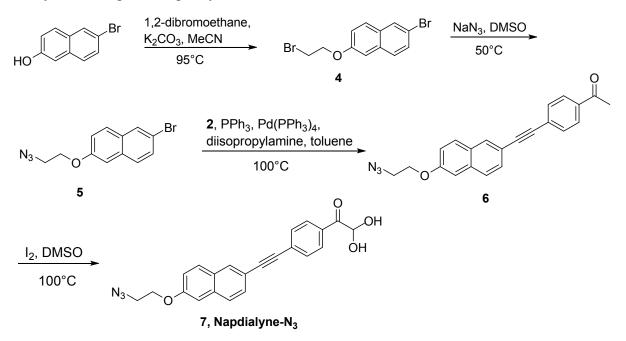
A solution of compound 1 (1.0 g, 5.04 mmol) and K_2CO_3 (690 mg, 5.04 mmol) in MeOH (40 mL) was stirred at room temperature for 15 min. Then the reaction mixture was diluted with water, and extracted with CH_2Cl_2 twice. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated on a rotary evaporator to get

compound **2** as a yellow solid (666 mg, quantitative yield). ¹H NMR (400 MHz, CDCl₃) δ 7.90 (d, J = 8.8 Hz, 2H), 7.58 (d, J = 8.4 Hz, 2H), 3.25 (s, 1H), 2.61 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 197.25, 136.75, 132.28, 128.18, 126.91, 82.76, 80.45, 26.63; HRMS: (ESI) [M+H]⁺ calc. for C₁₀H₉O, 145.0653, observed 145.0602.

A solution of compound **2** (250 mg, 1.74 mmol), triphenylphosphine (23 mg, 0.087 mmol), palladium chloride (15.5 mg, 0.087 mmol), cupric acetate (16 mg, 0.087 mmol), and 2-bromo-6-methoxynaphthalene (1650 mg, 6.96 mmol) in triethylamine (25 mL) was refluxed under 100°C for 2 h. After cooling, the reaction mixture was diluted with water, and extracted with CH₂Cl₂ twice. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated on a rotary evaporator. The obtained crude was purified by Al₂O₃ column chromatography (petroleum ether : ethyl acetate = 20 : 1) to yield compound **3** as a white solid (195 mg, 39% yield). ¹H NMR (400 MHz, CDCl₃) δ 8.01 (s, 1H), 7.97 (d, *J* = 8.4 Hz, 2H), 7.74 (dd, *J* = 4.8, 8.8 Hz, 2H), 7.65 (d, *J* = 8.4 Hz, 2H), 7.57 (d, *J* = 8.4 Hz, 1H), 7.19 (dd, *J* = 2.4, 8.8 Hz, 1H), 7.14 (d, *J* = 2.0 Hz, 1H), 3.94 (s, 3H), 2.63 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 197.37, 158.57, 136.02, 134.42, 131.69, 131.63, 129.44, 128.88, 128.45, 128.43, 128.30, 126.96, 119.58, 117.47, 105.84, 93.49, 88.38, 55.37, 26.62; HRMS: (ESI) [M+H]⁺ calc. for C₂₁H₁₇O₂, 301.1229, observed 301.1230.

A solution of compound **3** (200 mg, 0.67 mmol) and iodine (338 mg, 1.33 mmol) in a mixture of DMSO and H₂O (20 mL, DMSO/H₂O = 4 /1) was refluxed under 100°C for 2 h. After cooling, the reaction mixture was diluted with H₂O, extracted with CH₂Cl₂ for three times. The combined organic layers were washed sequentially with Na₂S₂O₃ aqueous solution (1 mol/L, 20 mL) and brine, dried over anhydrous Na₂SO₄ and concentrated on a rotary evaporator. The crude was stirred with CH₂Cl₂ to give a white slurry, which was filtered to give the final compound **Napdialyne** as a light yellow solid (130 mg, 62% yield). ¹H NMR (400 MHz, d₆-DMSO) δ 8.17 (s, 1H), 8.13 (d, *J* = 8.4 Hz, 2H), 7.88 (t, *J* = 8.8 Hz, 2H), 7.72 (d, *J* = 8.0 Hz, 2H), 7.60 (d, *J* = 9.6 Hz, 1H), 7.38 (d, *J* = 2.0 Hz, 1H), 7.24 (dd, *J* = 2.4, 8.8 Hz, 1H), 6.86 (d, *J* = 7.2 Hz, 2H), 5.68 (t, *J* = 7.2 Hz, 1H), 3.90 (s, 3H); ¹³C NMR (101 MHz, d₆-DMSO) δ 195.97, 158.94, 134.74, 133.49, 132.01, 131.71, 130.21, 129.98, 128.98, 128.49, 127.71, 127.65, 120.04, 116.94, 106.62, 93.63, 90.03, 88.95, 55.83; HRMS: (ESI) [M+H-H₂O]⁺ calc. for C₂₁H₁₅O₃, 315.1021, observed 315.1005.

To a solution of L-citrulline (56 mg, 0.318 mmol) in H₂O/TFA (1:1, 8 mL) was added Napdialyne (100 mg, 0.318 mmol). After being stirred under 37 °C for 10 hours, the reaction mixture was concentrated. The obtained crude was recrystallized with MeOH, and the obtained solid (unpure) was dissolved in MeOH and further precipitated from petroleum ether. And the obtained solid was further mixed with CH₂Cl₂ to give a white slurry, which was filtered to afford **Napdialyne-Cit** as a white solid (46 mg, 31% yield). ¹H NMR (400 MHz, d₆-DMSO) δ 8.12 (s, 1H), 7.86 (t, *J* = 8.4 Hz, 2H), 7.63 (d, *J* = 8.0 Hz, 2H), 7.58 (dd, *J* = 1.2, 8.0 Hz, 1H), 7.34 (m, 3H), 7.21 (dd, *J* = 2.4, 9.2 Hz, 1H), 5.28 (s, 1H), 3.90 (s, 3H), 3.54 (m, 2H), 2.67 (s, 1H), 1.55 (m, 4H); ¹³C NMR (101 MHz, d₆-DMSO) δ 173.03, 170.26, 158.71, 157.04, 134.71, 134.66, 134.55, 132.41, 131.68, 129.89, 129.00, 128.51, 127.71, 123.43, 119.97, 106.58, 91.19, 88.89, 63.90, 63.71, 55.82, 54.24, 53.94, 28.44, 23.61, 23.46; HRMS: (ESI) [M+H]⁺ calc. for C₂₇H₂₆N₃O₅, 472.1872, observed 472.1859. 3. Synthesis of probe Napdialyne-N₃



A solution of 6-bromo-2-naphthol (2.0 g, 8.97 mmol), K₂CO₃ (1240 mg, 8.97 mmol) and 1,2dibromoethane (3863 µL, 44.83 mmol) in acetonitrile (30 mL)was refluxed for 8 h under 95°C. After cooling to room temperature, the reaction mixture was mixed with water, and extracted with CH₂Cl₂ twice. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated on a rotary evaporator. The obtained crude was further purified by silica gel column chromatography (petroleum ether : ethyl acetate = 10 : 1) to yield compound **4** as a yellow oil (1.46 g, 50% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.92 (s, 1H), 7.67 (d, *J* = 8.8 Hz, 1H), 7.59 (d, *J* = 8.8 Hz, 1H), 7.51 (dd, *J* = 1.4, 8.8 Hz, 1H), 7.19 (dd, *J* = 2.4, 9.2 Hz, 1H), 7.09 (d, *J* = 2.4 Hz, 1H), 4.40 (t, *J* = 6.4 Hz, 2H), 3.70 (t, *J* = 6.4 Hz, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 156.31, 132.85, 130.27, 129.78, 129.68, 128.75, 128.41, 119.75, 117.44, 107.03, 67.85, 28.89; HRMS: (ESI) [M+H]⁺ calc. for C₁₂H₁₁BrO, 328.9177, observed 328.9129.

A solution of compound **4** (750 mg, 2.29 mmol) and sodium azide (164 mg, 2.52 mmol) in a mixture of DMSO (10 mL) was heated under 50°C for 2 h. After cooling to room temperature, the reaction mixture was diluted with H₂O, and extracted with CH₂Cl₂ for three times. The combined organic layers were washed brine, dried over anhydrous Na₂SO₄ and concentrated on a rotary evaporator to get compound **5** as a white solid (890 mg, quantitative yield).¹H NMR (400 MHz, CDCl₃) δ 7.92 (s, 1H), 7.68 (d, *J* = 9.2 Hz, 1H), 7.60 (d, *J* = 8.8 Hz, 1H), 7.52 (dd, *J* = 2.0, 8.8 Hz, 1H), 7.21 (dd, *J* = 2.4, 9.2 Hz, 1H), 7.10 (d, *J* = 2.4 Hz, 1H), 4.24 (t, *J* = 5.2 Hz, 2H), 3.68 (t, *J* = 5.2 Hz, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 156.45, 132.83, 130.24, 129.75, 129.68, 128.72, 128.40, 119.73, 117.40, 106.79, 66.95, 50.09; HRMS: (ESI) [M+H]⁺ calc. for C₁₂H₁₁BrN₃O, 292.0085, observed 292.0023.

A solution of compound **2** (500 mg, 3.47 mmol), tetrakis(triphenylphosphine) palladium (320 mg, 0.278 mmol), and compound **5** (1210 mg, 4.16 mmol) in toluene (20 mL) and diisopropylamine (20 mL) was refluxed under 100°C for 5 h. After cooling to room temperature, the reaction mixture was diluted with water, and extracted with CH_2Cl_2 twice. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated on a rotary evaporator. The obtained crude was purified by silica gel column chromatography (petroleum ether : DCM = 1 : 1) to yield compound **6** as a yellow solid (440 mg, 36% yield). ¹H NMR (400 MHz, CDCl₃) δ 8.02 (s, 1H), 7.97 (d, *J* = 8.0 Hz, 2H), 7.77 (d, *J* = 8.8 Hz, 1H), 7.73(d, *J* = 8.8 Hz, 1H), 7.65 (d, *J* = 8.0 Hz, 2H), 7.58 (d, *J* = 8.4 Hz, 1H), 7.24 (dd, *J* = 2.4, 2.4 Hz, 1H), 7.14 (d, *J* = 2.0 Hz, 1H), 4.29 (t, *J* = 5.2 Hz, 2H), 3.69 (t, *J* = 4.8 Hz, 2H), 2.63 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 197.36, 157.13, 136.07, 134.21, 131.69, 131.65, 129.69, 129.01, 128.69, 128.35, 128.31, 127.01, 119.55, 117.86, 106.84, 93.33, 88.54, 66.98, 50.09, 26.63; HRMS: (ESI) [M+H]⁺ calc. for C₂₂H₁₈N₃O₂, 356.1399, observed 356.1394.

A solution of compound **6** (130 mg, 0.366 mmol) and iodine (186 mg, 0.732 mmol) in a mixture of DMSO and H₂O (20 mL, DMSO/H₂O = 4 /1) was refluxed under 100 °C for 4 h. After cooling to room temperature, the reaction mixture was diluted with H₂O, and extracted with CH₂Cl₂ for three times. The combined organic layers were washed sequentially with Na₂S₂O₃ aqueous solution (1 mol/L, 20 mL) and brine. Then it was dried over anhydrous Na₂SO₄, and concentrated on a rotary evaporator. The obtained crude was mixed with CH₂Cl₂ to give a white slurry, which was filtered to give the final compound **7** (Napdialyne-N₃) as a light yellow solid (130 mg, 65% yield). ¹H NMR (400 MHz, d₆-DMSO) δ 8.19 (s, 1H), 8.14 (d, *J* = 8.4 Hz, 2H), 7.91 (d, *J* = 9.2 Hz, 1H), 7.89 (d, *J* = 8.4 Hz, 1H), 7.73 (d, *J* = 8.8 Hz, 2H), 7.62 (dd, *J* = 1.6, 8.4 Hz, 1H), 7.44 (d, *J* = 2.4 Hz, 1H), 7.24 (dd, *J* = 2.4, 8.8 Hz, 1H), 6.87 (d, *J* = 7.2 Hz, 2H), 5.67 (t, *J* = 7.2 Hz, 1H), 4.33 (t, *J* = 4.8 Hz, 2H), 3.75 (t, *J* = 4.8 Hz, 2H); ¹³C NMR (101 MHz, d₆-DMSO) δ 195.97, 157.49, 134.60, 133.51, 132.03, 131.72, 130.24, 130.19, 129.06, 128.65, 127.85, 127.56, 119.95, 117.17, 107.65, 93.54, 90.04, 89.04, 67.41, 49.97; HRMS: (ESI) [M+H-H₂O]⁺ calc. for C₂₂H₁₆N₃O₃, 370.1192, observed 370.1149.

4. Fluorescence measurement

4.1 Fluorescence measurement using probe Napdial

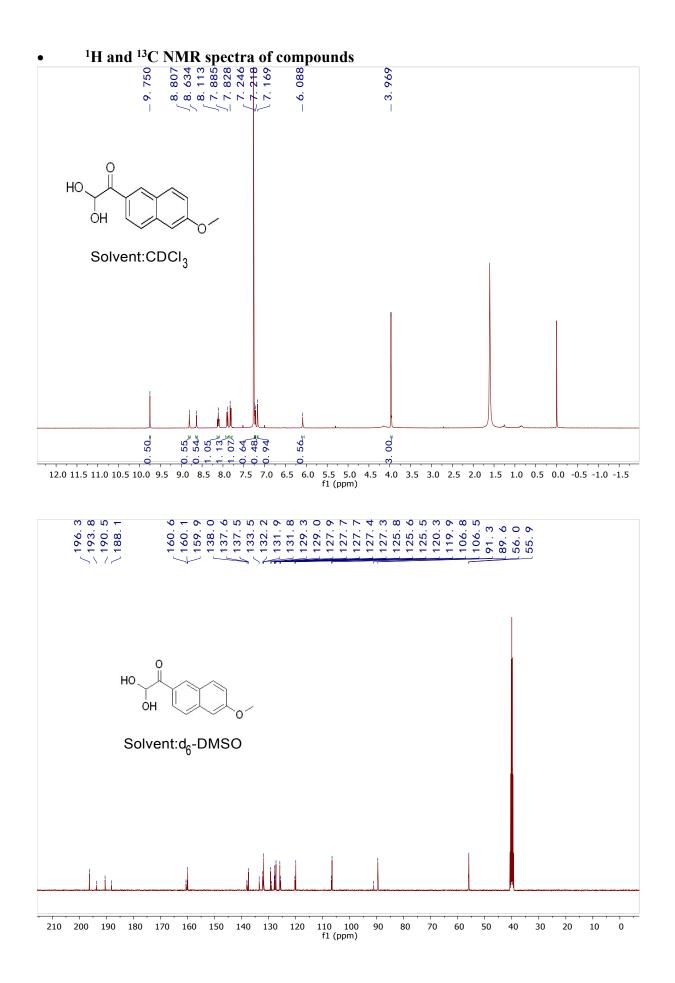
For a typical experiment, a solution of Napdial, L-citrulline (2-fold dilution) and trichloroacetic acid (40 μ L) in ddH₂O (160 μ L) was incubated at 50 °C for 3 h in 96-well plates. The final concentration for Napdial was 100 μ M. Then 10 μ L of reaction mixture was taken out, and diluted with MeOH (90 μ L) before measurement.

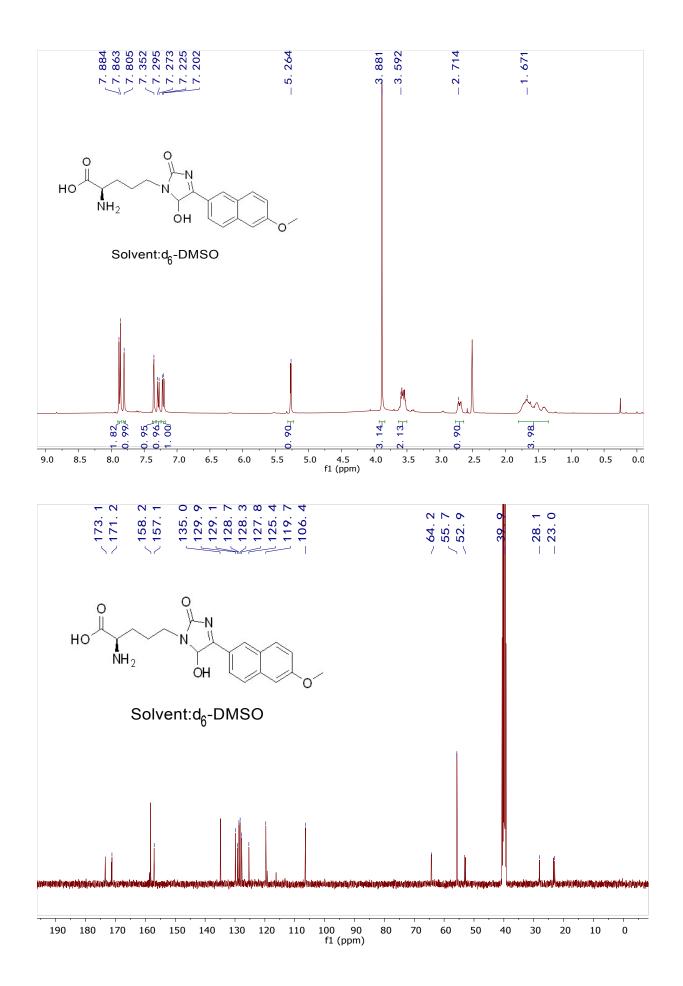
4.2 Fluorescence measurement using probe Napdialyne

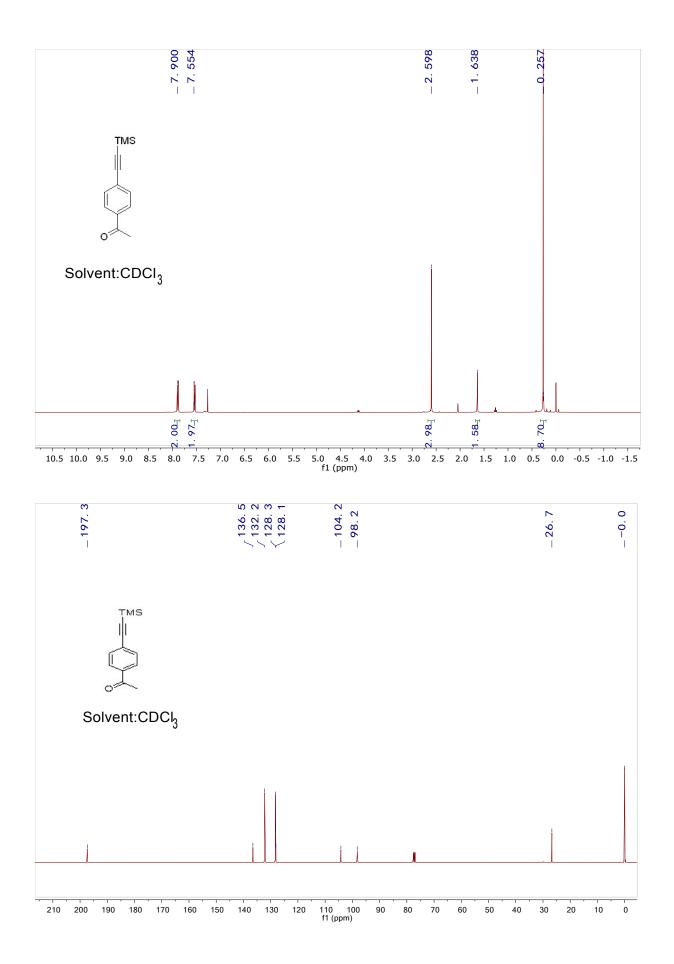
For a typical experiment, a solution of Napdialyne (10 μ L), L-citrulline (10 μ L, 2-fold dilution) and trichloroacetic acid (40 μ L) in ddH₂O (140 μ L) to incubated at 50 °C for 3 h in 96-well plates. The final concentration for Napdialyne was 50 μ M. Then 10 μ L of reaction mixture was taken out, and diluted with different solvents (90 μ L) before measurement.

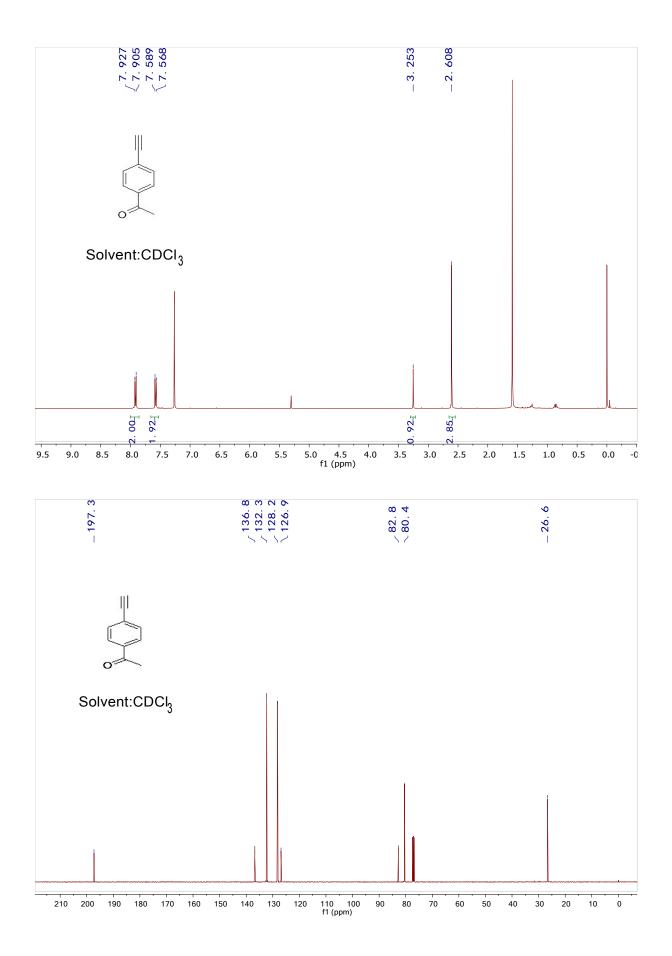
4.3 Fluorescence measurement using Napdialyne-N3 with DBCO-PEG4-biotin

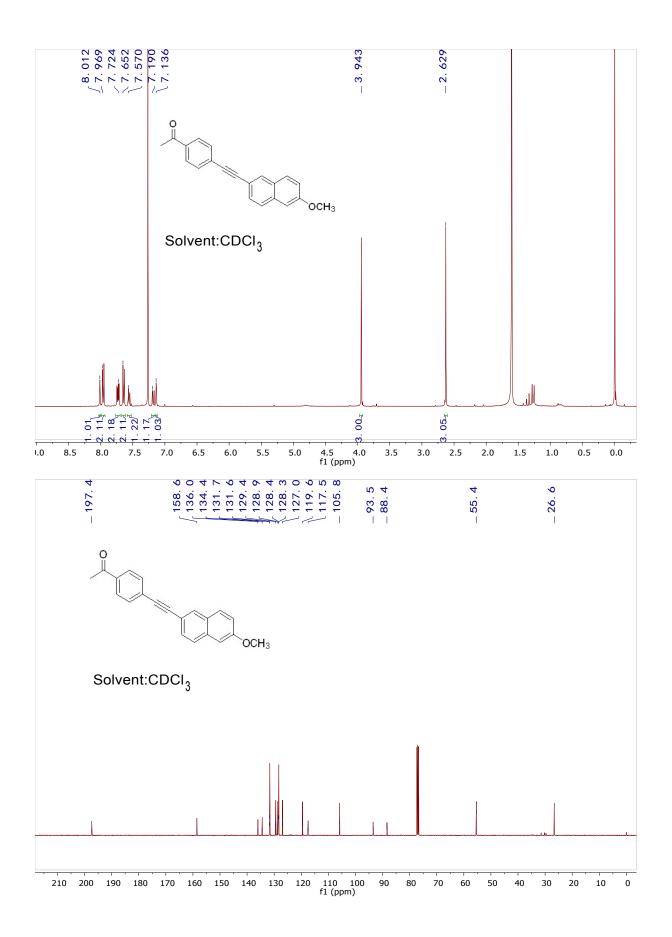
For a typical experiment, a solution of DBCO-PEG₄-Biotin (40 μ L, 10 mM in H₂O) and Napdialyne-N₃ (30 μ L, 10 mM in DMSO) in MeOH (130 μ L) was rotated at room temperature for 3 h in 1.5 mL centrifuge tube. Then 20 μ L of this reaction solution was taken out, and mixed with 10 μ L of analyte, 10 μ L of TFA and 10 μ L of H₂O. The mixture was allowed to incubate at 37 °C for 3 h. Then it was diluted 10-times with IPA (450 μ L). After that, 50 μ L of this dilute was taken out, and mixed with agarose beads (50 μ L). The mixture was rotated at room temperature for 1 h, and centrifuged to separate liquid and beads. 30 μ L of the supernatant was analyzed by fluorescence measurement, and designed as "supernatant" group. The beads were further diluted with 30 μ L of IPA, and 30 μ L of this IPA dilute was analyzed by fluorescence measurement, and designed as "beads" group.

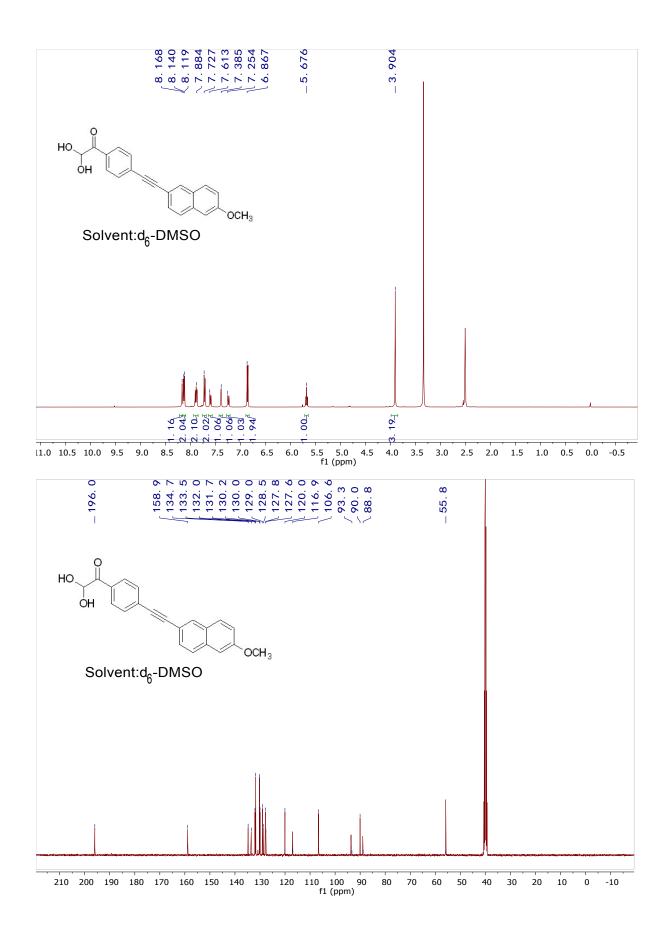


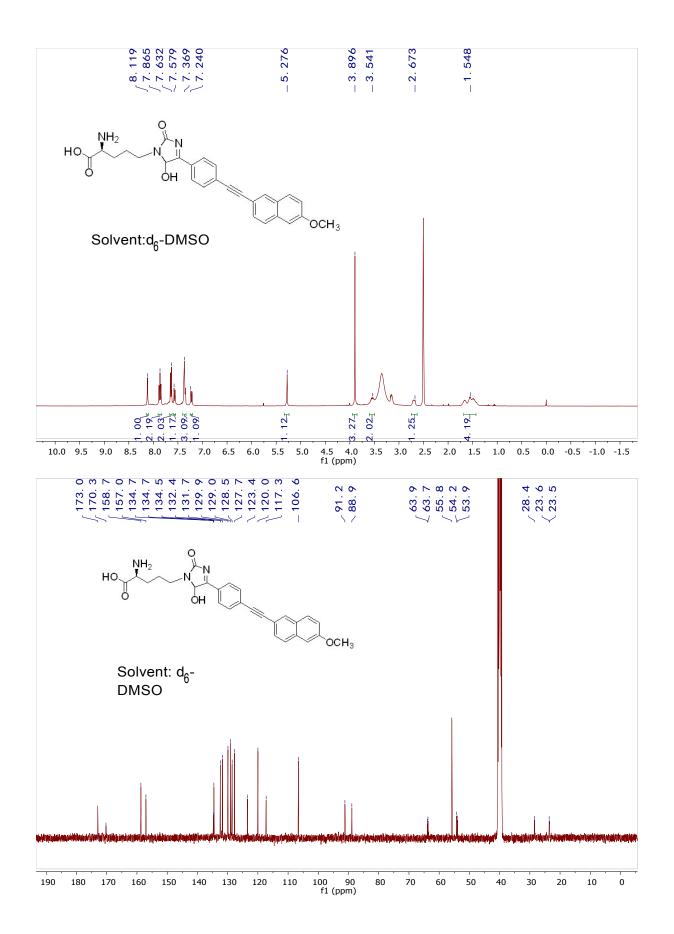


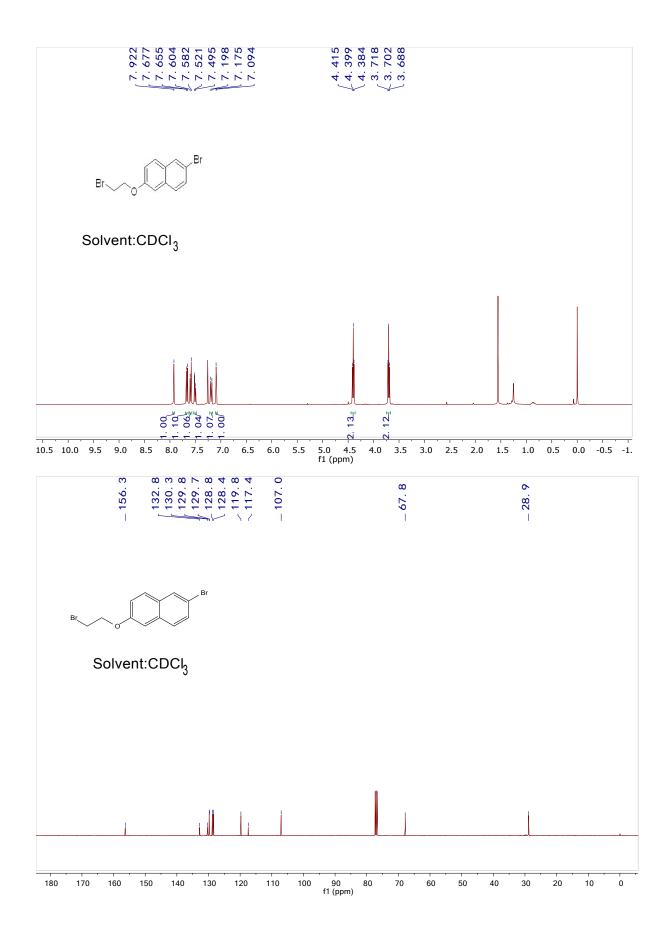


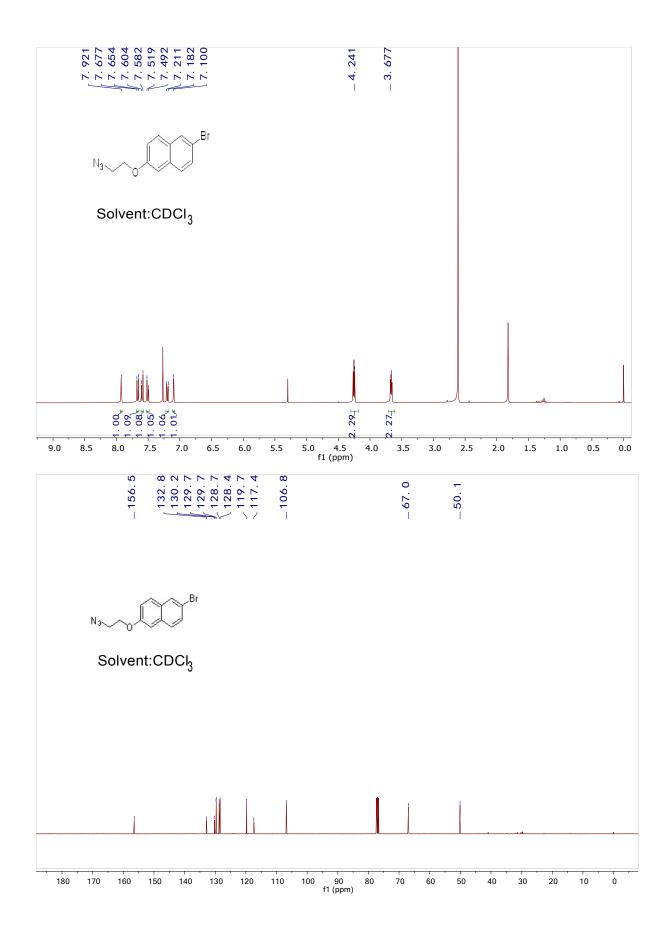


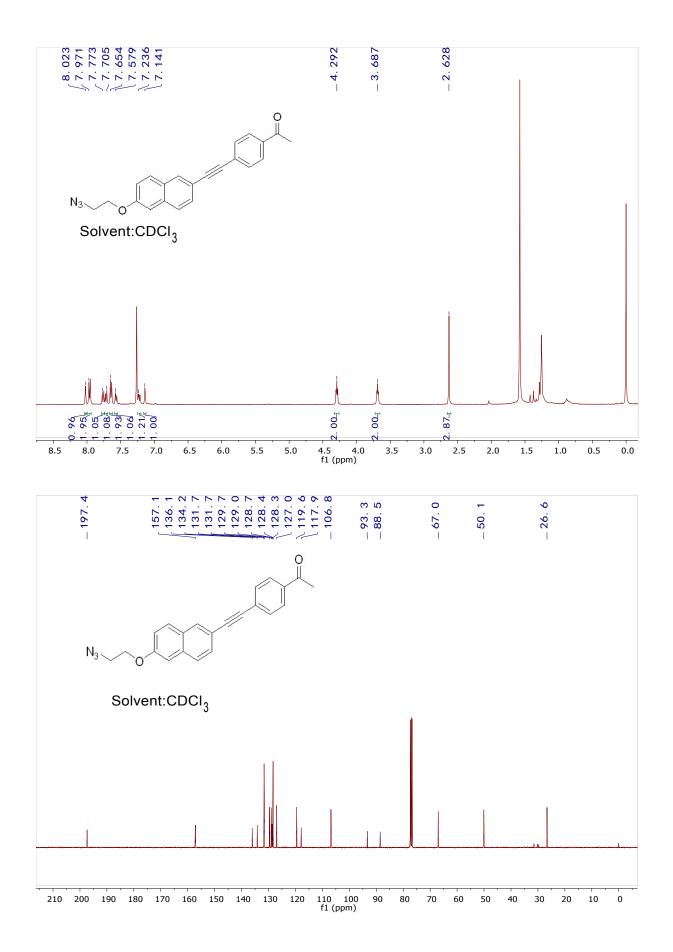


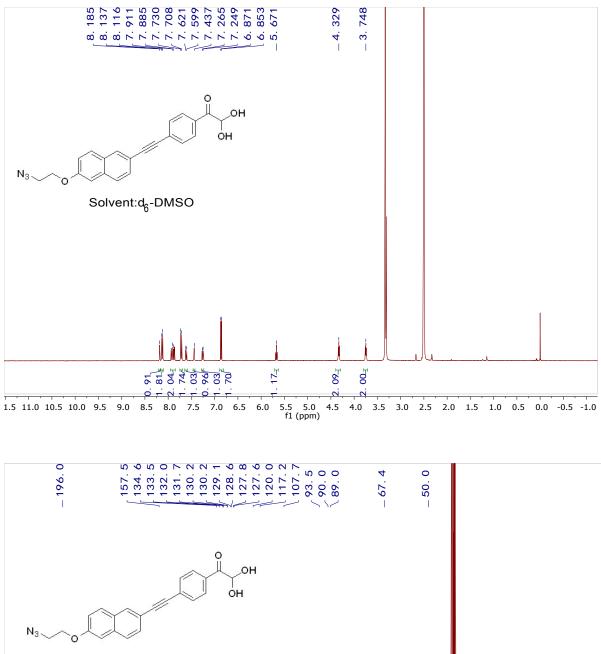


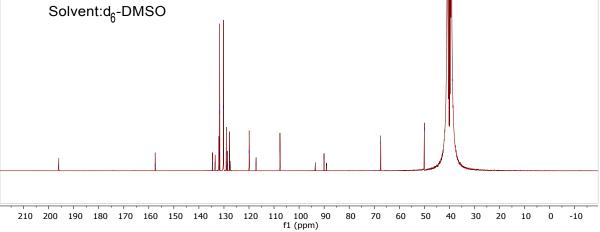












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