Cooperative Supramolecular Polymerization of Phosphorescent Alkynyl-Gold(I)–Isocyanide Complexes

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1. Absorption and emission spectroscopy of 2a-b

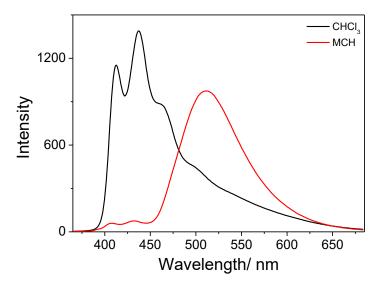


Figure S1. Emission spectra of 2b ($\lambda_{ex} = 350$ nm) in CHCl₃ (black line) and MCH (red line).

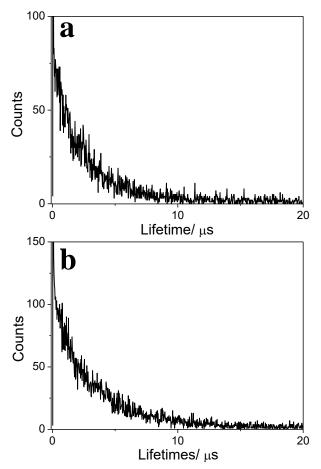


Figure S2. Lifetime measurement of a) 2a and b) 2b (2.00×10^{-5} M in MCH, $\lambda_{ex} = 350$ nm). The lifetime is determined to be 2.94 µs for 2a and 3.08 µs for 2b, suggesting the triplet origin for the emission bands.

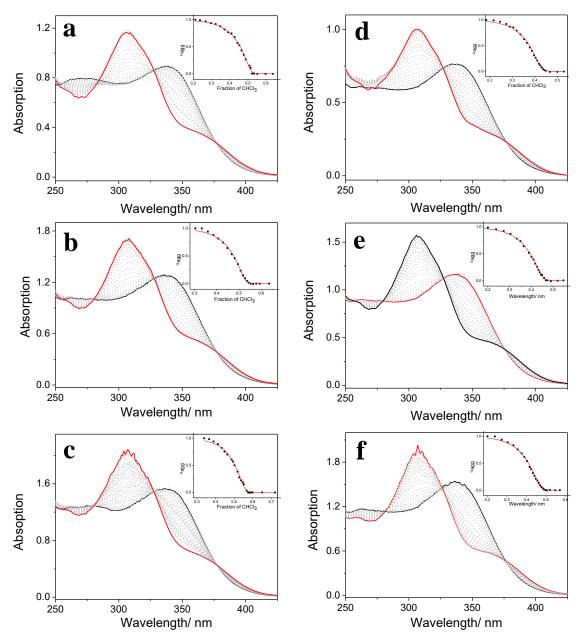


Figure S3. Solvent-dependent UV–Vis spectra (293K) of **2a–b** in MCH at different monomer concentration: a) 1.00×10^{-5} M of **2a**; b) 1.50×10^{-5} M of **2a**; c) 2.00×10^{-5} M of **2a**; d) 1.00×10^{-5} M of **2b**; e) 1.50×10^{-5} M of **2b**; f) 2.00×10^{-5} M of **2b**. Inset: normalized absorbance intensity at 348 nm *versus* CHCl₃ volume, and the corresponding mathematical fitting curves. The quantitative thermodynamic values are shown in Table S1.

2. CD spectroscopy of 2b

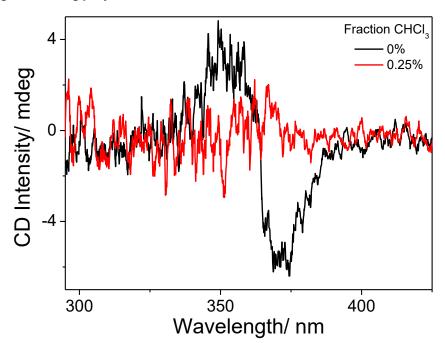


Figure S4. CD spectra of **1a** ($c = 2.00 \times 10^{-4}$ M, 273 K, 1 mm cuvette) at different fraction of CHCl₃: 0.00% (black line) and 0.25% (red line). As can be seen, the CD signal disappears upon adding trace amount of CHCl₃.

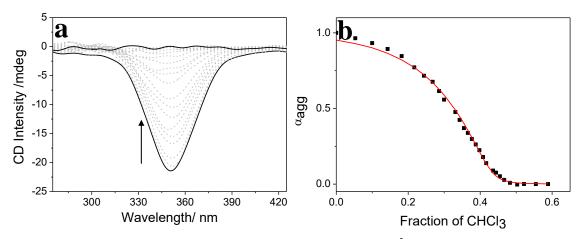


Figure S5. a) Solvent-dependent CD spectra of **2b** ($c = 1.50 \times 10^{-5}$ M in MCH, 293K, 10 mm cuvette). b) Normalized CD intensity of **2b** at 348 nm *versus* CHCl₃ volume fraction (from 0.00 to 0.59). The critical chloroform volume fraction (46.0%) is consistent with that acquired from UV–Vis experiments under the same conditions (46.2%).

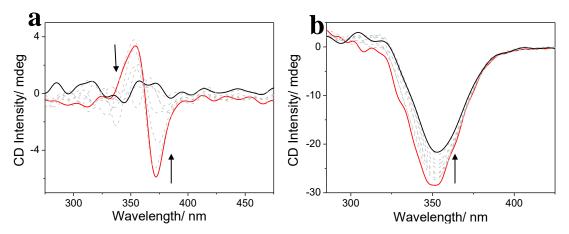


Figure S6. a) Solvent-dependent CD spectra of a) 1a; and b) 2b (2.00 × 10⁻⁴ M in MCH, 1 mm cuvette). Arrows indicate the spectral changes upon increasing toluene fraction (a: from 0% to 8%, and b: from 0% to 100%) The CD signal for 1a disappears upon adding small amount of toluene, suggesting the complete breakup of supramolecular polymeric structure.
On the contrary, the CD signals for 2b maintain even in pure toluene, reflecting the sufficient stability of the self-assembled helical structure.

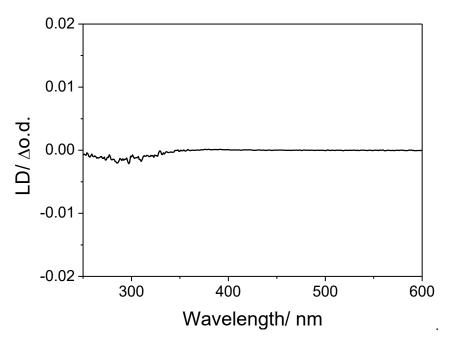


Figure S7. LD spectrum of 2a (1.50 × 10⁻⁵ M in MCH, 293K). As can be seen, no LD signal is detected for the CD-active samples, demonstrating the measured CD signals are real to reflect the supramolecular chirality.

3. ¹H NMR measurements of 2a-b

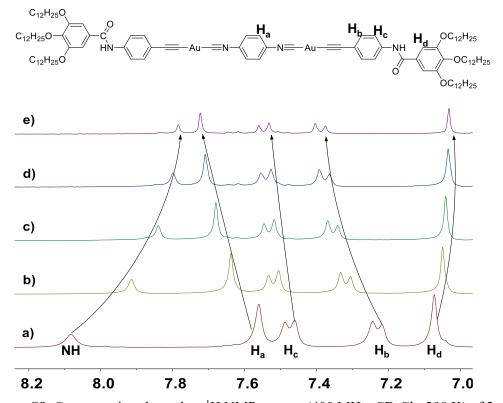


Figure S8. Concentration-dependent ¹H NMR spectra (400 MHz, CD₂Cl₂, 298 K) of 2a: a) 20.0 mM; b) 10.0 mM; c) 5.00 mM; d) 2.50 mM; e) 1.25 mM. Upon increasing the monomer concentration from 0.26 to 20 mM, the amide N–*H* protons undergo significant downfield shifts from 7.78 to 8.08 ppm (Δδ: –0.30 ppm), while the aromatic protons on 2a shift upfield (Δδ: 0.16, 0.15, and 0.07 ppm for H_a, H_b, and H_c, respectively). The phenomena illustrate the prominent roles of both hydrogen bonding and π–π stacking interactions for intermolecular aggregation process.

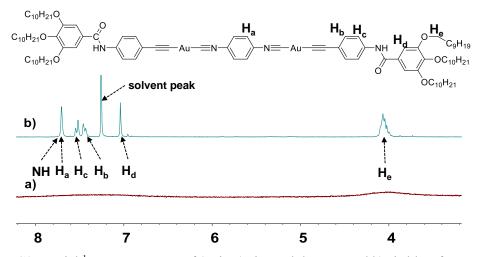


Figure S9. Partial ¹H NMR spectra of 2b in a) d₁₂-cyclohexane and b) d-chloroform at 2.00 mM. 2b shows broadened and indiscernible signals, indicative of the formation of high-molecular-weight polymeric structure.

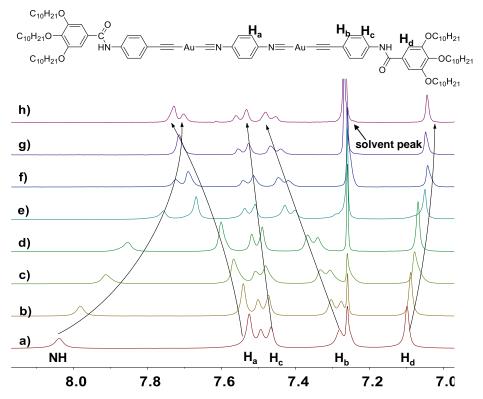


Figure S10. Concentration-dependent ¹H NMR spectra (400 MHz, CDCl₃, 298 K) of **2b**: a) 20.0 mM; b) 16.0 mM; c) 11.4 mM; d) 8.00 mM; e) 4.00 mM; f) 2.67 mM; g) 2.00 mM; h) 1.25 mM. Upon increasing the monomer concentration from 1.25 to 20 mM, the amide N–*H* protons undergo significant downfield shifts from 7.70 to 8.04 ppm ($\Delta\delta$: –0.34 ppm), while the aromatic protons on **2b** shift upfield ($\Delta\delta$: 0.19, 0.19, and 0.06 ppm for H_a, H_b, and H_c, respectively). The phenomena illustrate the prominent roles of both hydrogen bonding and π – π stacking interactions for intermolecular aggregation process.

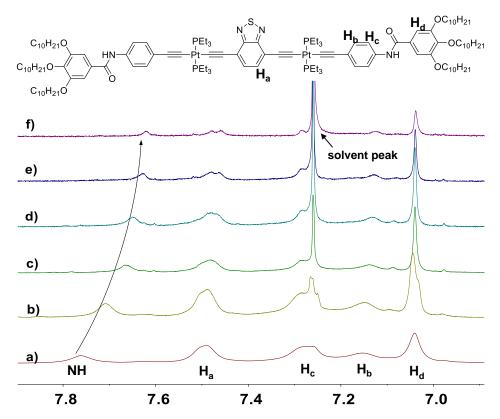


Figure S11. Concentration-dependent ¹H NMR spectra (400 MHz, CDCl₃, 298 K) of 1a: a) 40.0 mM; b) 20.0 mM; c) 10.0 mM; d) 5.00 mM; e) 2.50 mM; f) 1.25 mM. No aromatic signal changes occur for 1a, while the amide N–H resonances exhibit slight downfield shift (from 7.62 to 7.71 ppm, Δδ: -0.09 ppm). It suggests that only hydrogen bonds play critical role, while π–π stacking is absent for the 1a self-assembly process.

4. Macroscopic gelation behaviors of 2a-b

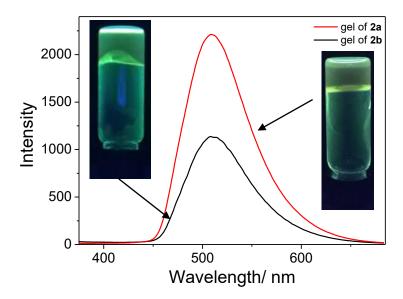


Figure S12. Emission spectra of **2a** (red line) and **2b** (black line) in the gel state (10.0 mM in MCH/CHCl₃, 95 : 5 (v/v), $\lambda_{ex} = 350$ nm). Inset pictures: gels under 365 nm UV lamp.

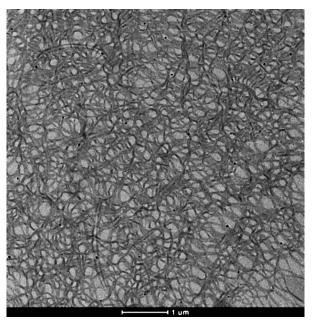


Figure S13. TEM image of 2b (copper grid, drop-casting from 0.40 mM solution in MCH/CHCl₃(95 : 5, v/v)). One-dimensional fibres are observed for 2b, laying the basis for the formation of three-dimensional supramolecular gels.

5. Light-harvesting behavior of 2b/1b

Determination of fluorescence resonance energy transfer (FRET) efficiency: spectral overlap integral $J(\lambda)$ between the emission spectra of donor and absorption spectra of acceptor is calculated according to Eq. S2:

$$J(\lambda) = \int_0^\infty f_D(\lambda) \varepsilon_A(\lambda) \lambda^4 d\lambda \qquad (Eq. S1)$$

In this equation, λ and $\varepsilon_A(\lambda)$ are the wavelength (cm) and molar extinction coefficient of FRET acceptor (**1b**) at the selected wavelength, respectively. $f_D(\lambda)$ is the fraction of fluorescent intensity of FRET donor (**2b**). The spectra overlap between the emission spectrum of **2b** and the extinction spectrum of **1b** is shown in Figure 5a in the main text. The overlap integral $J(\lambda)$ is calculated to be $1.99 \times 10^{12} \text{ M}^{-1} \text{ cm}^{-1} \text{ nm}^{3}$.

FRET efficiency (Φ_{ET}) is calculated according to Eq. S3:^{S1}

$$\phi_{ET} = 1 - I_{DA} / I_D \qquad (Eq. S2)$$

In this equation, I_{DA} and I_D are the emission intensities of FRET donor with and without the presence of FRET acceptor, respectively.

Energy transfer rate constant ($K_{\rm ET}$) is calculated according to Eq. S4:^{S2}

$$\phi_{ET} = K_{ET} / (K_{ET} + \tau_D^{-1})$$
 (Eq. S3)

In this equation, τ_D is the emission lifetime of donor **2b** in MCH.

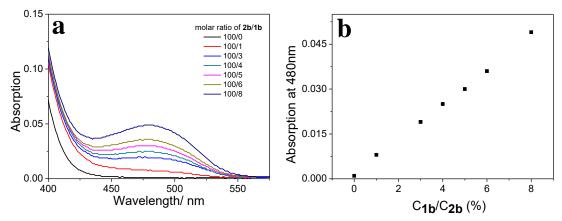


Figure S14. a) UV–Vis spectra of 2b with the presence of different amount of 1b. b) Absorbance at 480 nm displays the linear relationship upon increasing the amount of 1b. The result suggests the absence of ground-state interaction between the antenna (2b) and acceptor (1b) units.

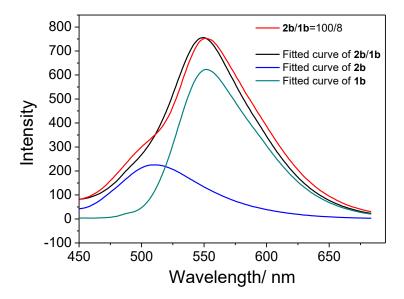


Figure S15. Peak-differentiation-imitating spectra of the emission spectra of 2b/1b. Herein 2b/1b = 100 : 8 is taken as an example. The quantitative energy-transfer efficiency (Φ_{ET}) is calculated to 69.0%.

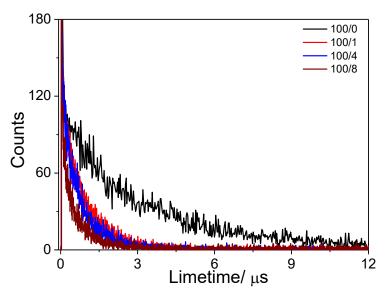


Figure S16. Lifetime decay of 2b with the presence of different amount of 1b. Upon varying the molar ratio of 2b/1b from 100 : 0 to 100 : 8, the averaged lifetime of 2b decreases from 3.08 µs to 0.53 µs.

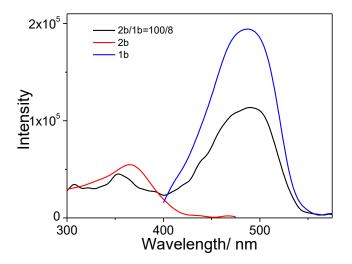
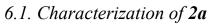


Figure S17. Excitation spectra of complex 2b/1b ($\lambda_{em} = 580$ nm), and the individual species.

- 6. Structural characterization of **2a-b**
 - $\begin{array}{c} 7.55\\ 7.55\\ 7.52\\$ - ~~~~ 12.74 20.78 99.86 18.32 2.00 3.99 3.95 4.02 4.19 ò 5 8.0 7.5 7.0 6.5 5.5 4.5 4.0 1.5 1.0 6.0 5.0 3.5 3.0 2.5 2.0 Figure S18. ¹H NMR spectrum (400 MHz, CD₂Cl₂, 298 K) of 2a.



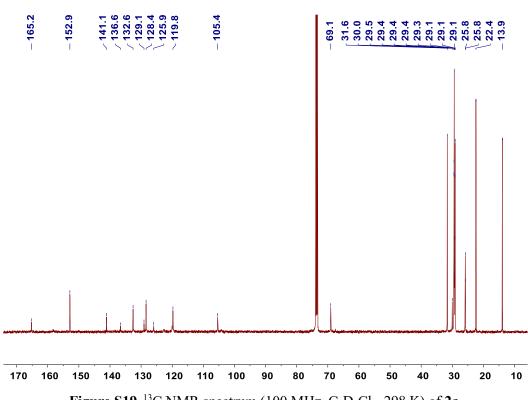


Figure S19. ¹³C NMR spectrum (100 MHz, C₂D₂Cl₄, 298 K) of 2a.

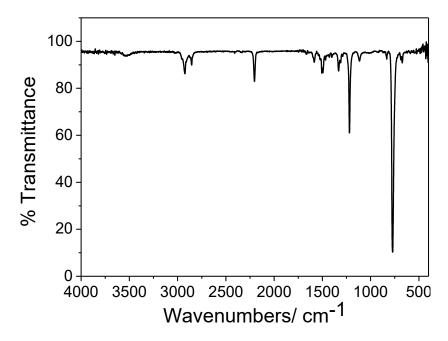


Figure S20. FT–IR spectrum of 2a. The IR band at 2200 cm⁻¹ can be assigned to the vibration peaks of C=C unit.

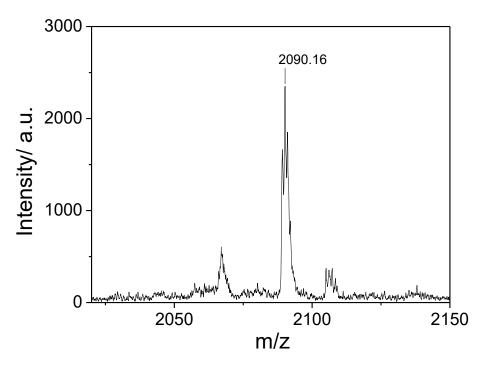
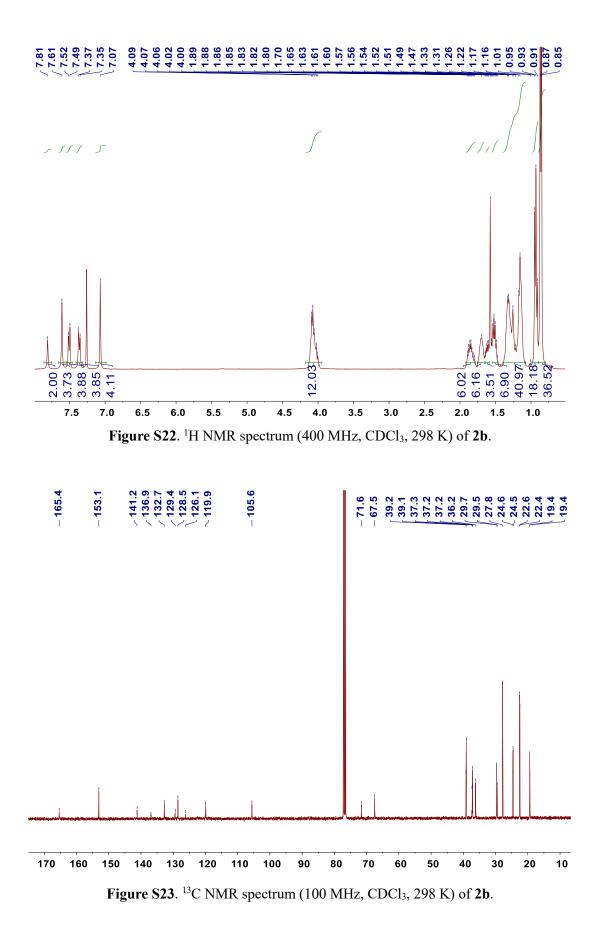


Figure S21. MALDI-TOF mass spectrum of 2a.

6.2. Characterization of 2b



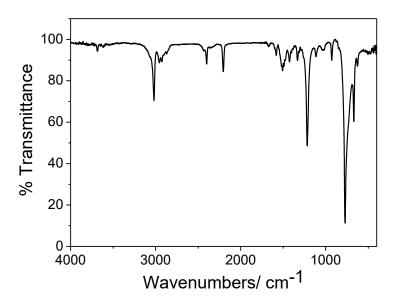


Figure S24. FT–IR spectrum of 2b. Two IR bands are observed at 2200 cm⁻¹ and 2399 cm⁻¹. They are assigned to the vibration peaks of C=C and N=C units, respectively.

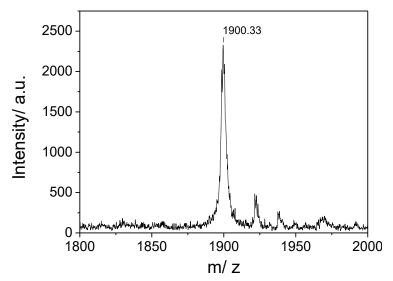


Figure S25. MALDI-TOF mass spectrum of 2b.

REFERENCES:

S1. Z. Xu, S. Peng, Y.-Y. Wang, J.-K. Zhang, A. I. Lazar and D.-S. Guo, *Adv. Mater.*, 2016, **28**, 7666.

S2. Y. Liu, S. Li, K. Li, Y. Zheng, M. Zhang, C. Cai, C. Yu, Y. Zhou and D. Yan, *Chem. Commun.*, 2016, **52**, 9394.