## ELECTRONIC SUPPORTING INFORMATION

## Versatile Functionalization of Polymer Nanoparticles with Carbonate Groups via Hydroxyurethane Linkages

Neha Yadav, Farzad Seidi, Silvano Del Gobbo, Valerio D'Elia,\* Daniel Crespy\*

Department of Materials Science and Engineering, School of Molecular Science and Engineering, Vidyasirimedhi Institute of Science and Technology (VISTEC), Rayong 21210, Thailand **Table S1.** Synthesis of P(GCMA-*co*-MMA) copolymers by miniemulsion copolymerization of GCMA and MMA at 70 °C for 16 h.

Entry	Polymer	Monomer [mol%]		Solid Content	Diameter	
		GCMA	MMA	[%]	D <sub>h</sub> [nm]	PDI
1	P(GCMA)	100	0	63	134	0.21
		(100) <sup>a</sup>	(0) <sup>a</sup>			
2	P(GCMA-co-MMA)	50	50	95	116	0.19
		(63.75) <sup>a</sup>	(36.25) <sup>a</sup>			
3	$P(GCMA_{0.91}\text{-}co\text{-}MMA_{0.9})^{b}$	75	25	72	139	0.20
		(84.47) <sup>a</sup>	(16.63) <sup>a</sup>			

<sup>a</sup> Amount of monomer in mg

<sup>b</sup> Composition determined by <sup>1</sup>H NMR spectroscopy in DMSO-d6.

**Table S2**. Elemental analysis of polymer nanoparticles after reaction with various amines, dialysis, and drying.

Material	Amine/5CCs	С	N	Н	N/C
	[mol%]	[%]	[%]	[%]	(% molar ratio)
P-BA 1	50	54.3	1.3	6.2	2.0
P-BA 3	200	59.7	4.0	6.6	5.9
P-HMDA 2	100	52.4	2.6	7.2	4.2
P-Arg 2	200	54.1	1.4	6.8	2.2
P-Lys 2	200	53.0	1.3	6.2	2.1
P-Alb 1	50ª	52.3	3.5	6.1	5.9
P-Alb 3	200 <sup>[a]</sup>	52.0	3.2	6.1	5.3
P-Dop	200 <sup>[a]</sup>	55.3	4.5	6.6	7.1

<sup>a</sup> wt% ratio of albumin to P(GCM-co-MMA) nanoparticles.



**Figure S1.** <sup>1</sup>H-NMR spectrum of P(GCMA-co-MMA) dissolved in DMSO-d<sub>6</sub> synthesized by miniemulsion polymerization after purification and drying.



**Figure S2.** IR spectra of hydrolyzed copolymer of glycidyl methacrylate (GMA) with methyl methacrylate (MMA) obtained by miniemulsion copolymerization at 70  $^{\circ}$ C for 24 h.







**Figure S3.** <sup>1</sup>H-NMR spectra of GCMA in D<sub>2</sub>O at pD 7 (a), pD 3 (b), and pD 11 (c) Hydrolysis kinetics of GCMA at pD 11 expressed as ratio of NMR signals integrals ( $I_{b+b'}/I_a$ ).



**Figure S4.** IR spectra of P(GCMA-*co*-MMA) at various time intervals in  $D_2O$  after purification and drying of nanoparticles dispersed in a buffer solution at pH 11.









**Figure S5.** IR spectra of purified and dried polymers before and after functionalization of P(GCMA-*co*-MMA) nanoparticles with HMDA (a), arginine (b), L-lysine (c), dopamine (d).



**Figure S6.** IR spectra of PMMA before and after dissolution in DMSO in the presence of benzyl amine.



**Figure S7.** (a) Calibration curve for determining the concentration of benzylamine based on absorbance at 257 nm; (b) UV absorbance spectra of dialysis media of P-BA 3 dispersion (diluted 130 times in water).





(b)







**Figure S8.** Solid state CP/MAS <sup>13</sup>C NMR spectra of purified and dried P(GCMA-co-MMA) nanoparticles (a); P-Dop (b); P-Alb 3; (c) P-BA 3 (d) and P-Lys 2 (e).



Figure S9. <sup>1</sup>H (Top) and <sup>13</sup>C (Bottom) NMR spectra of propylene carbonate in  $D_2O$ .



Figure S10. <sup>1</sup>H (Top) and <sup>13</sup>C (Bottom) NMR spectra of L-lysine in  $D_2O$ .



Figure S11. <sup>1</sup>H (Top) and <sup>13</sup>C (Bottom) NMR spectra of L-arginine in  $D_2O$ .



Figure S12. <sup>1</sup>H (Top) and <sup>13</sup>C (Bottom) NMR spectra of 1-Lys in  $D_2O$ .



Figure S13. <sup>1</sup>H (Top) and <sup>13</sup>C (Bottom) NMR spectra of 1-Arg in  $D_2O$ .



Figure S14. <sup>1</sup>H (Top) and <sup>13</sup>C (Bottom) NMR spectra of *N*-Ac-lysine in D<sub>2</sub>O.



**Figure S15.** <sup>1</sup>H (Top) and <sup>13</sup>C (Bottom) NMR spectra of the product of the reaction of *N*-Ac-lysine with PC in presence of TEA ( $D_2O$ ).

Table	<b>S3</b> .	Binding	energies	(eV)	of C	1s	components	evaluated	by	deconvolution	of	high
resolut	ion 2	XPS spect	tra of func	tional	ized l	P(G	CMA-co-MM	(A) nanopa	rtic	les.		

Functional	С-С, С-Н	- <u>C</u> -C=O	C-O, C-N,	-COO-	0= <u>C</u> -N	-O(C=O)O-
Group			C-NH <sub>3</sub> <sup>+</sup>		R-( <u>C</u> =O)-R	-O(C=O)NH
			$C_{\alpha}$ amino acids			-NH- $(C(NH_2)_2)^+$
Compound						
	(C1)	(C2)	(C3)		(C4)	(C5)
P(GCMA-co- MMA)	284.59	285.37	286.85		288.70	290.43
P-BA 1	284.69	285.7	286.7		288.82	290.60
P-HMDA 1	284.7	285.69	286.67		288.81	
P-Dop	284.55	285.57	286.50		288.79	
Arginine	284.58	285.22	286.41	287.25	288.77	
P-Arg 2	284.59	285.26	286.53	287.54	288.90	
P-Lys 2	284.51	285.16	286.34	287.31	288.83	
Albumin	284.76		286.19		287.93	
P-Alb 1	284.66	285.89	286.99		288.53	290.56
P-Alb 3	284.58	285.36	286.52		288.27	290.75

**Table S4.** Binding energies (eV) of N 1s components evaluated by deconvolution of highresolution XPS spectra of functionalized P(GCMA-co-MMA) nanoparticles.

Chemical	-C-NH	O=C-NH	-O(C=O)NH	-NH3 <sup>+</sup>
Environment			-NH-(C(NH <sub>2</sub> ) <sub>2</sub> ) <sup>+</sup>	
	(N1)	(N2)	(N3)	(N4)
Compound				
P(GCMA-co-MMA)				
P-BA 1			399.75	
P-HMDA 1			399.68	401.63
P-Dop			399.87	
Arginine	398.42		399.73	
P-Arg 2			399.23	
P-Lys 2			399.27	
Albumin	398.22	399.72		
P-Alb 1	398.14	399.27	399.94	
P-Alb 3	399.02	399.65	400.30	

Chemical	( <u>O</u> =C)-NH-	-(С= <u>О</u> )ОН	- <u>O</u> -(C=O)- <u>O</u> -
Environment	-O-(C= <u>O</u> )-C-	-O-(C= <u>O</u> )-O-	- <u>O</u> -(C=O)-
	-O-(C= <u>O</u> )-NH-		- <u>O</u> -(C=O)-NH-
			- <u>О</u> Н
Compound	(01)	(02)	(03)
P(GCMA-co-MMA)	531.80	532.61	533.78
P-BA 1	532	2.09	533.48
P-HMDA 1	532	2.01	533.33
P-Dop	532	2.16	533.24
P-Arg 2	532	2.12	533.40
P-Lys 2	532	2.11	533.39
Albumin	530.98	531.94	533.29
P-Alb 1	531.03	531.998	533.66
P-Alb 3	531.10	532.05	533.61

**Table S5.** Binding energies (eV) of O 1s components evaluated by deconvolution of highresolution XPS spectra of functionalized P(GCMA-co-MMA) nanoparticles.



**Figure S16.** High resolution XPS spectra of P(GCMA-co-MMA) nanoparticles in the C 1s (a) and O 1s (b) spectral regions with components obtained by deconvolution.





**Figure S17.** High resolution XPS spectra of P-BA 1 (a-c), P-HMDA 1, (d-f) P-Dop 1, (g-i) P-Arg 2, (j-l) and P-Lys 2 (m-o) in the C 1s, N 1s and O 1s spectral regions with components obtained by deconvolution. The O 1s fittings for P-Dop and P-Arg 2 include a minor component indicated by \* that could be due to the presence of inorganic impurities.<sup>1</sup>



**Figure S18.** High resolution XPS spectra of pure albumin (a-c) and of P-Alb-1 (d-f) and P-Alb-3 (g-i) samples in the C 1s, N 1s and O 1s spectral regions with components obtained by deconvolution.



**Figure S19.** High resolution XPS spectra of arginine in the C 1s (a), N 1s (b) and O1s (c) spectral regions with components obtained by deconvolution.

**Table S6.** Atomic ratio of nitrogen to carbon measured by XPS for nanoparticles functionalized

 with several amines and albumin.

Entry	Amine/5CCs	N/C
	[mol%]	(% molar ratio)
P-BA 1	50	3.0
P-HMDA 1	20	3.4
P-Dop	200	5.6
P-Arg 2	200	2.7
P-Lys 2	200	2.2
P-Alb 1	50ª	7.7

<sup>a</sup> wt% albumin to P(GCMA-co-MMA) nanoparticles.



**Figure S20.** SEM micrographs of purified and dried P/Dop nanoparticles before catalysis (a); after catalysis (b); particle size of P-Dop determined from SEM measurements before catalysis (c).



**Figure S21.** <sup>1</sup>H NMR spectrum of the products of the reaction between styrene oxide and  $CO_2$  to afford styrene carbonate. The reaction is catalyzed by TBAI (Table 2, Entry 1). Additional peaks in the spectrum arise from residual CHCl<sub>3</sub> in the CDCl<sub>3</sub> (7.24 ppm), H<sub>2</sub>O in CDCl<sub>3</sub> (1.6 ppm), TMS standard (0.0 ppm), and TBAI (3.26, 1.70, 1.35 and 0.9 ppm).



**Figure S22.** <sup>1</sup>H NMR spectrum of the product of the reaction between styrene oxide and CO<sub>2</sub> to afford styrene carbonate. The reaction is catalyzed by TBAI and 50 mg P-Dop (Table 2, Entry 4). Additional peaks in the spectrum arise from residual CHCl<sub>3</sub> in CDCl<sub>3</sub> (7.24 ppm), H<sub>2</sub>O in CDCl<sub>3</sub> (1.6 ppm), TMS standard (0.0 ppm), and TBAI (3.26, 1.70, 1.35 and 0.9 ppm).



**Figure S23.** <sup>1</sup>H NMR spectrum of the product of the reaction between styrene oxide and CO<sub>2</sub> to afford styrene carbonate. The reaction is catalyzed by TBAI and 100 mg P-Dop (Table 2, Entry 5). Additional peaks in the spectrum arise from residual CHCl<sub>3</sub> in CDCl<sub>3</sub> (7.24 ppm), H<sub>2</sub>O in CDCl<sub>3</sub> (1.6 ppm), TMS standard (0.0 ppm), and TBAI (3.26, 1.70, 1.35 and 0.9 ppm).



**Figure S24.** <sup>1</sup>H NMR spectrum of the product of the reaction between styrene oxide and CO<sub>2</sub> to afford styrene carbonate. The reaction is catalyzed by TBAI and recycled (1<sup>st</sup> time) P-Dop (Table 2, Entry 6). Additional peaks in the spectrum arise from residual CHCl<sub>3</sub> in CDCl<sub>3</sub> (7.24 ppm), H<sub>2</sub>O in CDCl<sub>3</sub> (1.6 ppm), TMS standard (0.0 ppm), and TBAI (3.26, 1.70, 1.35 and 0.9 ppm).



**Figure S25.** <sup>1</sup>H NMR spectrum of the product of the reaction between styrene oxide and CO<sub>2</sub> to afford styrene carbonate. The reaction is catalyzed by TBAI and recycled ( $2^{nd}$  time) P-Dop (Table 2, Entry 7). Additional peaks in the spectrum arise from residual CHCl<sub>3</sub> in CDCl<sub>3</sub> (7.24 ppm), H<sub>2</sub>O in CDCl<sub>3</sub> (1.6 ppm), TMS standard (0.0 ppm), and TBAI (3.26, 1.70, 1.35 and 0.9 ppm).



**Figure S26.** <sup>1</sup>H NMR spectrum of the product of the reaction between styrene oxide and CO<sub>2</sub> to afford styrene carbonate. The reaction is catalyzed by TBAI and recycled ( $3^{rd}$  time) P-Dop (Table 2, Entry 8). Additional peaks in the spectrum arise from residual CHCl<sub>3</sub> in CDCl<sub>3</sub> (7.24 ppm), H<sub>2</sub>O in CDCl<sub>3</sub> (1.6 ppm), TMS standard (0.0 ppm), and TBAI (3.26, 1.70, 1.35 and 0.9 ppm).



**Figure S27.** <sup>1</sup>H NMR spectrum of the product of the reaction between 1-hexene oxide and CO<sub>2</sub> to afford 1-hexene carbonate. The reaction is catalyzed by TBAI and P-Dop (Table 2, Entry 9). Additional peaks in the spectrum arise from residual CHCl<sub>3</sub> in CDCl<sub>3</sub> (7.24 ppm), TMS standard (0.0 ppm), and TBAI (3.26, 1.35 and 0.9 ppm).



**Figure S28** <sup>1</sup>H NMR spectrum of the product of the reaction between epichlorohydrin and  $CO_2$  to afford epichlorohydrin carbonate. The reaction is catalyzed by TBAI and P-Dop (Table 2, Entry 10). Additional peaks in the spectrum arise from TMS standard (0.0 ppm) and TBAI (3.26, 1.35 and 0.9 ppm).



**Figure S29.** <sup>1</sup>H NMR spectrum of the product of the reaction between styrene oxide and CO<sub>2</sub> to afford styrene carbonate catalyzed by TBAI and dopamine (Table 2, Entry 11). Additional peaks in the spectrum arise from residual CHCl<sub>3</sub> in CDCl<sub>3</sub> (7.24 ppm), H<sub>2</sub>O in CDCl<sub>3</sub> (1.6 ppm), TMS standard (0.0 ppm), and TBAI (3.26, 1.70, 1.35 and 0.9 ppm).

![](_page_44_Figure_0.jpeg)

**Figure S30.** <sup>1</sup>H NMR spectrum of the product of the reaction between styrene oxide and CO<sub>2</sub> to afford styrene carbonate catalyzed by TBAI and catechol (Table 2, Entry 12). Additional peaks in the spectrum arise from residual CHCl<sub>3</sub> in CDCl<sub>3</sub> (7.24 ppm), H<sub>2</sub>O in CDCl<sub>3</sub> (1.6 ppm), TMS standard (0.0 ppm), and TBAI (3.26, 1.70, 1.35 and 0.9 ppm).

## Reference

1. Chu, D. H.; Vinoba, M.; Bhagiyalakshmi, M.; Hyun, B., II; Nam, S. C.; Yoon, Y.; Kim, S. H.; Jeong, S. K., CO2 mineralization into different polymorphs of CaCO3 using an aqueous-CO2 system. *RSC Advances* **2013**, *3*, 21722.