Electronic supplementary information (ESI)

Copper-based ionic liquid-catalyzed click polymerization of diazides and diynes toward functional polytriazoles for sensing applications

Baixue Li, † Rong Hu, † Anjun Qin *, † and Ben Zhong Tang *, †,‡

^{*†*} State Key Laboratory of Luminescent Materials and Devices, Guangdong Provincial Key Laboratory of Luminescence from Molecular Aggregates, Center for Aggregation-Induced Emission, South China University of Technology, Guangzhou 510640, China

[‡] Department of Chemistry, Hong Kong Branch of Chinese National Engineering Research Center for Tissue Restoration and Reconstruction, Institute for Advanced Study, and Department of Chemical and Biological Engineering, The Hong Kong University of Science & Technology, Clear Water Bay, Kowloon, Hong Kong, China

Table of Contents

Synthesis procedures of monomer 1a-1d.	S4
Synthesis procedures of monomer 2a-2c.	S4
Synthesis procedure of Cu-IL catalyst.	S 6
Cytotoxicity evaluation.	S6
Binding behavior towards pathogens.	S6
Cell imaging.	S 7
Table S1 Effect of solvent on the click polymerization.	S 7
Table S2 Effect of temperature on the click polymerization.	S7
Table S3 Time course of the click polymerization.	S 8
Table S4 Effect of Cu-IL amount on the click polymerization.	S 8
Table S5 Effect of monomer concentration on the click polymerization.	S 8
Fig. S1 WAXD (A) and DSC curves (B) of P1e2c.	S 8
Fig. S2 TGA thermograms of polymers P1a2a-P1e2c. T_d presents the temperat	ure of
5% weight loss.	S9
Fig. S3 FT-IR spectra of 1b (A), 2a (B) and P1b2a (C).	S9
Fig. S4 FT-IR spectra of 1c (A), 2a (B) and P1c2a (C).	S10
Fig. S5 FT-IR spectra of 1d (A), 2a (B) and P1d2a (C).	S10
Fig. S6 FT-IR spectra of 1b (A), 2b (B) and P1b2b (C).	S11
Fig. S7 FT-IR spectra of 1e (A), 2c (B) and P1e2c (C).	S11
Fig. S8 ¹ H NMR spectra of 1b (A), 2a (B) and P1b2a (C) in CDCl ₃ . The solver	nt peaks
are marked with asterisks.	S12
Fig. S9 ¹ H NMR spectra of 1c (A), 2a (B) and P1c2a (C) in CDCl ₃ . The solver	nt peaks
are marked with asterisks.	S13
Fig. S10 ¹ H NMR spectra of 1d (A), 2a (B) and P1d2a (C) in CDCl ₃ . The solver	nt peaks
are marked with asterisks.	S14
Fig. S11 ¹ H NMR spectra of 1b (A), 2b (B) and P1b2b (C) in CDCl ₃ . The solves	nt peaks
are marked with asterisks.	S15
Fig. S12 ¹ H NMR spectra of 1e (A), 2c (B) and P1e2c (C) in DMSO- d_6 . The	solvent
peaks are marked with asterisks.	S16

Fig. S13 ¹³C NMR spectra of 1b (A), 2a (B) and P1b2a (C) in CDCl₃. The solvent

peaks are marked with asterisks.

- Fig. S14 ¹³C NMR spectra of 1c (A), 2a (B) and P1c2a (C) in CDCl₃. The solvent peaks are marked with asterisks. S18
- Fig. S15 ¹³C NMR spectra of 1d (A), 2a (B) and P1d2a (C) in CDCl₃. The solventpeaks are marked with asterisks.S19
- Fig. S16 ¹³C NMR spectra of 1b (A), 2b (B) and P1b2b (C) in CDCl₃. The solvent peaks are marked with asterisks. S20
- Fig. S17 ¹³C NMR spectra of 1e (A), 2c (B) and P1e2c (C) in DMSO-d6. The solventpeaks are marked with asterisks.S21
- Fig. S18 (A) PL spectra of the P1a2a THF solutions containing different amounts of PA. Polymer concentration: 10 μ M. λ_{ex} : 320 nm. (B) Stern-Volmer plots of I_0/I –1 of P1a2a THF solution versus PA concentration, where I = peak intensity and I_0 = peak intensity at [PA] = 0 μ g mL⁻¹. Inset: the chemical structure of PA. S21
- Fig. S19 (A) PL decay curves of P1e2c aqueous solution at 450 nm in the presence of different amounts of PA. Polymer concentration: 10 μM; λ_{ex}: 340 nm. (B) Normalized absorption spectrum of PA and the PL spectrum of P1e2c aqueous solution.
- Fig. S20 (A) PL decay curves of P1e2c at 450 nm aqueous solution with different amounts of Fe³⁺ ion. Polymer concentration: 10 μ M; λ_{ex} : 340 nm. (B) Normalized absorption spectrum of Fe³⁺ ion and the PL spectrum of P1e2c aqueous solution. S22
- Fig. S21 Cell viability of HeLa cells in the presence of P1e2c with different concentrations.
 S23
- Fig. S22 FT-IR spectra of DEA (A) and Cu-IL (B).S23
- Fig. S23 TGA thermograms of Cu-IL and DEA. T_d presents the temperature of 5%
weight loss.S23

References

S24

Synthesis procedures of monomer 1a-1d.

Synthesis of bis(4-azidophenyl)methane (1a)

$$H_2N + t-BuONO + Me_3SiN_3 + CH_3CN + N_3 + Ia$$

This monomer was prepared according to previously published literature.¹

Synthesis of 4,4'-oxybis(azidobenzene) (1b)

$$H_2N \xrightarrow{O} H_2 + t-BuONO + Me_3SiN_3 \xrightarrow{O \circ C - RT} N_3$$

This monomer was prepared according to previously published literature.¹

FT-IR (KBr disk), v (cm⁻¹): 2121 (-N₃ stretching), 1500, 1307, 1246, 1099, 839, 812, 534, 513. ¹H NMR (500 MHz, CDCl₃), δ (TMS, ppm): 6.99 (Ar-H). ¹³C NMR (125 MHz, CDCl₃), δ (ppm): 154.50, 135.30, 120.55, 120.25.

Synthesis of 1,2-bis(4-azidophenyl)-1,2-diphenylethene (1c)



This monomer was prepared according to previously published literature.²

Synthesis of 1,2-bis(4-(azidomethyl)phenyl)-1,2-diphenylethene (1d)



This monomer was prepared according to previously published procedures.³

Synthesis procedures of monomer 2a-2c.

Synthesis of 2,7-diethynyl-9,9-dihexyl-9H-fluorene (2a)



This monomer was prepared according to our previously published literatures.⁴ FT-IR (KBr disk), v (cm⁻¹): 3303 (\equiv C–H stretching), 2931, 2855, 2107 (C \equiv C stretching), 1462, 888, 823, 647, 605. ¹H NMR (500 MHz, CDCl₃), δ (TMS, ppm): 7.64-7.46 (Ar-H), 3.15 (\equiv CH), 1.95, 1.10, 1.02, 0.77, 0.56. ¹³C NMR (125 MHz, CDCl₃), δ (ppm): 151.11, 141.23, 131.50, 126.63, 121.12, 120.28, 84.86 (\equiv C–), 77.48 (\equiv CH), 55.30, 40.41, 31.66, 29.76, 23.76, 22.63, 14.16.

Synthesis of 2,7-diethynyl-9,9-dioctyl-9H-fluorene (2b)



This monomer was prepared according to our previously published procedures.⁴

Synthesis of 2,5-diethynyl-1,4-phenylene tetraethyl bis(phosphate) (2c)



This monomer was prepared according to our previously published literatures.⁴⁻⁶

FT-IR (KBr disk), v (cm⁻¹): 3188 (\equiv C–H stretching), 2984, 2913, 2107 (C \equiv C stretching), 1493, 1392, 1282, 1268, 1189, 1164, 1051, 953, 892, 786, 746, 629, 521, 481, 463. ¹H NMR (500 MHz, DMSO- d_6), δ (TMS, ppm): 7.42 (Ar-H), 4.73 (\equiv CH), 4.21, 1.28. ¹³C NMR (125 MHz, DMSO- d_6), δ (ppm): 147.51, 124.51, 116.26, 88.53 (\equiv C–), 77.10 (\equiv CH), 64.89, 15.86.

Synthesis procedure of Cu-IL catalyst.



This Cu-IL was prepared according to previously published literature.^{7,8}

Cytotoxicity evaluation.

HeLa cells were seeded in 96-well plate with 8×10^3 cells per well. After 24 h of culture, the medium was replaced by a series of different concentrations of P1e2c from 0 μ M to 64 μ M (100 μ L well⁻¹). 24 h later, the medium was then exchanged by 100 μ L of fresh medium containing 10 μ L of MTT solution in each well and incubated at 37 °C for 4 h, and then 100 μ L of DMSO was added to dissolve the purple crystals. The absorption was recorded by a microplate reader at 570 nm after shaking for 2 min. The cell viability ratio (VR) was evaluated according to the following equation:

$$VR = \frac{A}{A_0} \times 100\%$$

Where A_0 is the absorbance of cells without any drugs, and A is the absorbance of cells incubated with Ple2c.

Binding behavior towards pathogens.

Preparation of bacterial solutions. A single colony of *S. aureus* on a solid Nutrient Broth (NB) agar plate was transferred to 10 mL of liquid Luria-Bertani (LB) culture medium

and grew at 37 °C for 12 h. Bacteria were harvested by centrifuging (7100 rpm for 1 min) and washed by phosphate buffer saline (PBS, 10 mM, pH = 7.4) for three times. The supernatant was discarded and the remaining *S. aureus* was resuspended in PBS solution, and diluted to an optical density of 1.0 at 600 nm ($OD_{600} = 1.0$). As for *E. coli*, except that the culture medium was replaced by LB medium (with 10 µM Amp) other experimental conditions and operations were totally the same as that of *S. aureus*.

100 μ L of bacterial solution (OD₆₀₀ = 1.0) was added into PBS solution containing P1e2c (10 μ M), followed by incubated at 37 °C for 20 min. Individual aliquots of 3 μ L of the pre-prepared mixed suspensions were added to clean glass slides followed by slightly covering coverslips for immobilization. The specimens were then examined by confocal laser scanning microscopy using a 405 nm laser.

Cell imaging.

HeLa cells were incubated with P1e2c (10 μ M) in Dulbecco's modified Eagle medium (DMEM) containing 10% fetal bovine serum (FBS) for 1 h, 4 h and 12 h respectively, followed with the incubation of Lysotracker for 30 min. Then the species were observed on CLSM. 405 nm laser for P1e2c, and 488 nm for Lysotracker.

Table ST Effect of solvent on the check polymenzation					
Entry	Solvent	Yield (%)	${M_{ m w}}^b$	D^b	
1	DMF	94	77 900	2.71	
2	Toluene	11	2600	1.05	
3	1,4-Dioxane	14	2700	1.07	
4	THF	Trace			

Table S1 Effect of solvent on the click polymerization^a

^{*a*} Carried out at 50 °C for 4 h under nitrogen, [Cu-IL]/[**1a**] = 0.1, [**1a**] = [**2a**] = 0.05 M. ^{*b*} Estimated by advanced polymer chromatography (APC) using THF as an eluent on the basis of a polystyrene (PS) calibration, M_w = weight-average molecular weight, polydispersity index (D) = M_w/M_n , M_n = number-average molecular weight.

Table S2 Effect of temperature on the click polymerization^{*a*}

Entry	<i>T</i> (°C)	Yield (%)	$M_{ m w}{}^b$	D^b
1	30	63	6800	1.42
2	40	90	45 500	2.49
3 ^{<i>c</i>}	50	94	77 900	2.71

^{*a*} Carried out in DMF for 4 h under nitrogen, [Cu-IL]/[1a] = 0.1, [1a] = [2a] = 0.05 M. ^{*b*} Estimated by APC using THF as an eluent on the basis of a PS calibration, M_w = weight-average molecular weight, $D = M_w/M_n$, M_n = number-average molecular weight. ^{*c*} Data taken from Table S1, entry 1.

Table S3 Time course of the click polymerization^a

Entry	<i>t</i> (h)	Yield (%)	${M_{ m w}}^b$	D^b
1	1	57	6300	1.33
2	2	90	59 900	2.47
3	3	88	76 900	2.53
4^c	4	94	77 900	2.71

^{*a*} Carried out in DMF at 50 °C under nitrogen, [Cu-IL]/[**1a**] = 0.1, [**1a**] = [**2a**] = 0.05 M. ^{*b*} Estimated by APC using THF as an eluent on the basis of a PS calibration, M_w = weight-average molecular weight, $D = M_w/M_n$, M_n = number-average molecular weight. ^{*c*} Data taken from Table S2, entry 3.

Table S4 Effect of Cu-IL amount on the click polymerization^a

Entry	Cu-IL (mol%)	Yield (%)	${M_{ m w}}^b$	D^b
1 ^c	10.0	90	59 900	2.47
2	5.0	92	77 800	2.67
3	2.5	22	4400	1.22
a Corried out	in DME at 50 °C for 2 h under	\mathbf{r} nitrogen $[1_{2}] = [2_{2}]$	-0.05 M ^b Eatis	moted by ADC

^{*a*} Carried out in DMF at 50 °C for 2 h under nitrogen, [1a] = [2a] = 0.05 M. ^{*b*} Estimated by APC using THF as an eluent on the basis of a PS calibration, $M_w =$ weight-average molecular weight, $D = M_w/M_n$, $M_n =$ number-average molecular weight. ^{*c*} Data taken from Table S3, entry 2.

Table S5 Effect of monomer concentration on the click polymerization^{*a*}

Entry	[M] (mol/L)	Yield (%)	$M_{ m w}{}^b$	D^b
1^c	0.050	92	77 800	2.67
2	0.040	94	39 700	2.76
3	0.033	84	14 700	1.98
4	0.025	28	4700	1.22

^{*a*} Carried out in DMF at 50 °C for 2 h under nitrogen, [Cu-IL]/[1a] = 0.05, [1a] = [2a]. ^{*b*} Estimated by APC using THF as an eluent on the basis of a PS calibration, M_w = weight-average molecular weight, $D = M_w/M_n$, M_n = number-average molecular weight. ^{*c*} Data taken from Table S4, entry 2.





Fig. S2 TGA thermograms of polymers P1a2a-P1e2c. T_d presents the temperature of 5% weight loss.



Fig. S3 FT-IR spectra of 1b (A), 2a (B) and P1b2a (C).



Fig. S4 FT-IR spectra of 1c (A), 2a (B) and P1c2a (C).



Fig. S5 FT-IR spectra of 1d (A), 2a (B) and P1d2a (C).



Fig. S6 FT-IR spectra of 1b (A), 2b (B) and P1b2b (C).



Fig. S7 FT-IR spectra of 1e (A), 2c (B) and P1e2c (C).



Fig. S8 ¹H NMR spectra of **1b** (A), **2a** (B) and P**1b2a** (C) in CDCl₃. The solvent peaks are marked with asterisks.



Fig. S9 ¹H NMR spectra of **1c** (A), **2a** (B) and P**1c2a** (C) in CDCl₃. The solvent peaks are marked with asterisks.



Fig. S10 ¹H NMR spectra of **1d** (A), **2a** (B) and P**1d2a** (C) in CDCl₃. The solvent peaks are marked with asterisks.



Fig. S11 ¹H NMR spectra of **1b** (A), **2b** (B) and P**1b2b** (C) in CDCl₃. The solvent peaks are marked with asterisks.



Fig. S12 ¹H NMR spectra of 1e (A), 2c (B) and P1e2c (C) in DMSO- d_6 . The solvent peaks are marked with asterisks.



Fig. S13 ¹³C NMR spectra of 1b (A), 2a (B) and P1b2a (C) in CDCl₃. The solvent peaks are marked with asterisks.



Fig. S14 ¹³C NMR spectra of **1c** (A), **2a** (B) and P**1c2a** (C) in CDCl₃. The solvent peaks are marked with asterisks.



Fig. S15 ¹³C NMR spectra of 1d (A), 2a (B) and P1d2a (C) in CDCl₃. The solvent peaks are marked with asterisks.



Fig. S16 ¹³C NMR spectra of 1b (A), 2b (B) and P1b2b (C) in CDCl₃. The solvent peaks are marked with asterisks.



Fig. S17 ¹³C NMR spectra of 1e (A), 2c (B) and P1e2c (C) in DMSO- d_6 . The solvent peaks are marked with asterisks.



Fig. S18 (A) PL spectra of the P**1a2a** THF solutions containing different amounts of PA. Polymer concentration: 10 μ M. λ_{ex} : 320 nm. (B) Stern-Volmer plots of $I_0/I - 1$ of P**1a2a** THF solution versus PA concentration, where I = peak intensity and $I_0 =$ peak intensity at [PA] = 0 μ g mL⁻¹. Inset: the chemical structure of PA.



Fig. S19 (A) PL decay curves of P1e2c aqueous solution at 450 nm in the presence of different amounts of PA. Polymer concentration: 10 μ M; λ_{ex} : 340 nm. (B) Normalized absorption spectrum of PA and the PL spectrum of P1e2c aqueous solution.



Fig. S20 (A) PL decay curves of P1e2c at 450 nm aqueous solution with different amounts of Fe³⁺ ion. Polymer concentration: 10 μ M; λ_{ex} : 340 nm. (B) Normalized absorption spectrum of Fe³⁺ ion and the PL spectrum of P1e2c aqueous solution.



Fig. S21 Cell viability of HeLa cells in the presence of P1e2c with different concentrations.



Fig. S22 FT-IR spectra of DEA (A) and Cu-IL (B).



Fig. S23 TGA thermograms of Cu-IL and DEA. T_d presents the temperature of 5% weight loss.

References

- (1) Y. Zhao and T. M. Swager, Eur. J. Org. Chem., 2015, 2015, 4593-4597.
- (2) J. Wang, J. Mei, E. Zhao, Z. Song, A. Qin, J. Z. Sun and B. Z. Tang, *Macromolecules*, 2012, 45, 7692–7703.
- (3) A. Qin, L. Tang, J. W. Y. Lam, C. K. W. Jim, Y. Yu, H. Zhao, J. Z. Sun and B. Z. Tang, *Adv. Funct. Mater.*, 2009, **19**, 1891–1900.
- (4) E. Zhao, H. Li, J. Ling, H. Wu, J. Wang, S. Zhang, J. W. Y. Lam, J. Z. Sun, A. Qin and B. Z. Tang, *Polym. Chem.*, 2014, 5, 2301–2308.
- (5) S. S. L. Corre, M. Berchel, H. C. Gourvès, J. P. Haelters and P. A. Jaffrès, *Beilstein J. Org. Chem.*, 2014, **10**, 1166–1196.
- (6) R. Hu, R. Ye, J. W. Y. Lam, M. Li, C. W. T. Leung and B. Z. Tang, *Chem.—Asian J.*, 2013, 8, 2436–2445.
- (7) B. Mohan, H. Kang and K. H. Park, Inorg. Chem. Commun., 2013, 35, 239-241.
- (8) H. D. Pratt III, A. J. Rose, C. L. Staiger, D. Ingersoll and T. M. Anderson, *Dalton Trans.*, 2011, 40, 11396–11401.