Electronic Supplementary Material (ESI) for Polymer Chemistry. This journal is © The Royal Society of Chemistry 2020

1

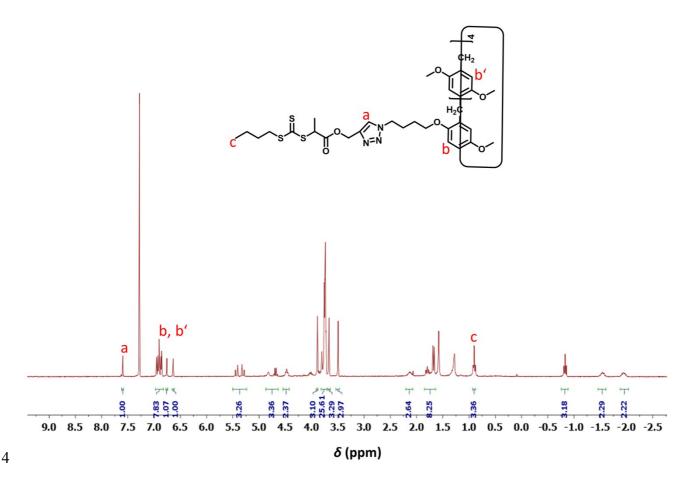
**Supporting Information** 

Degradable polycaprolactone nanoparticles stabilized via supramolecular 2 host-guest interactions with pH-responsive polymer-pillar[5]arene 3 conjugates 4 Peng Wei, <sup>a,b</sup> Fabian H. Sobotta, <sup>a,b</sup> Carolin Kellner, <sup>a,b</sup> Damiano Bandelli, <sup>a,b</sup> Stephanie Höppener, <sup>a,b</sup> Stephanie Schubert, b,c Johannes C. Brendel, a,b Ulrich S. Schubert \*, a,b 6 <sup>a</sup> Laboratory of Organic and Macromolecular Chemistry (IOMC), Friedrich Schiller University Jena, 7 Humboldtstraße 10, 07743 Jena, Germany 8 <sup>b</sup> Jena Center for Soft Matter (JCSM), Friedrich Schiller University Jena, Philosophenweg 7, 07743 9 10 Jena, Germany <sup>c</sup> Institute of Pharmacy and Biopharmacy, Department of Pharmaceutical Technology, Friedrich 11 12 Schiller University Jena, Lessingstrasse 8, 07743 Jena, Germany \*Correspondence to: U. S. Schubert (E-mail: ulrich.schubert@uni-jena.de) 13 14 15 16 17 18 19 20 21

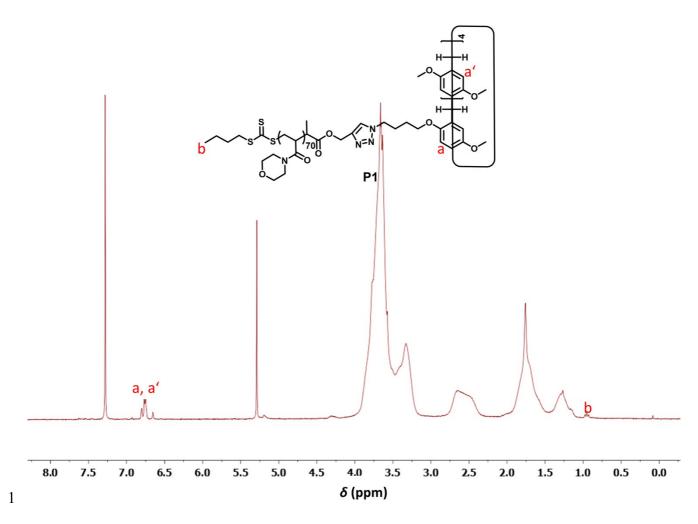
$$\begin{array}{c} DCC \\ DMAP \\ CH_2Cl_2 \\ rt, 24 \ h \end{array}$$

$$\begin{array}{c} Cu_2SO_4 \circ 5H_2O \\ NaVc \\ N \circ NaVc \\$$

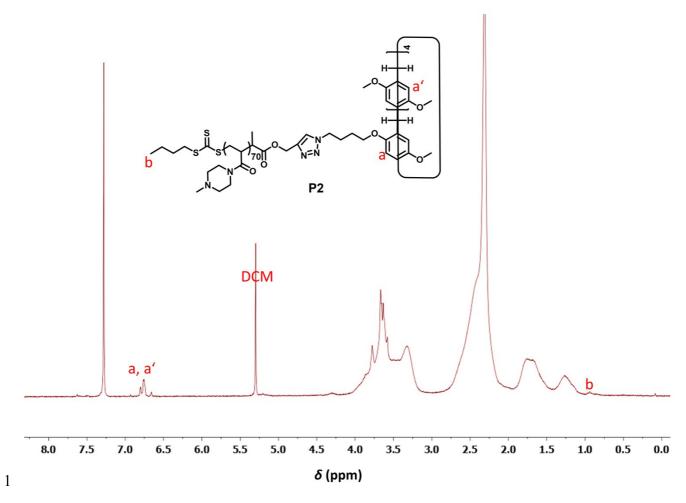
2 Scheme S1 Synthetic route of pillar[5]arene modified CTA.



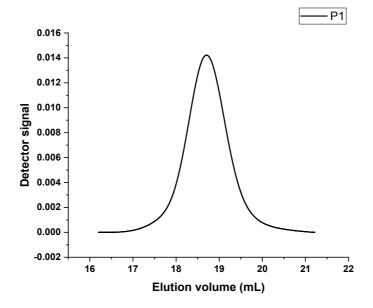
**Figure S1** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) spectrum of the pillar[5]arene modified chain transfer agent 6 (CTA).



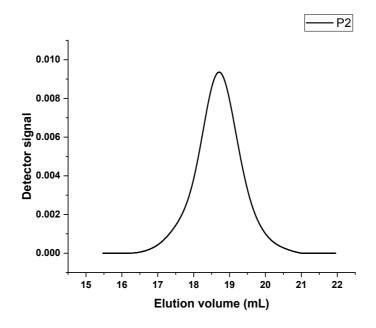
2 Figure S2 <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) spectrum of P1 (PNAM).



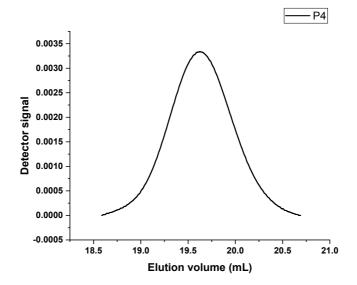
**Figure S3** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) spectrum of **P2** (PNAMP).



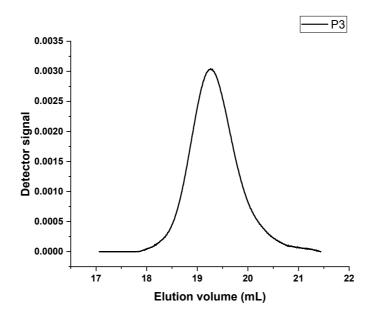
**Figure S4** SEC trace of **P1** (PNAM).



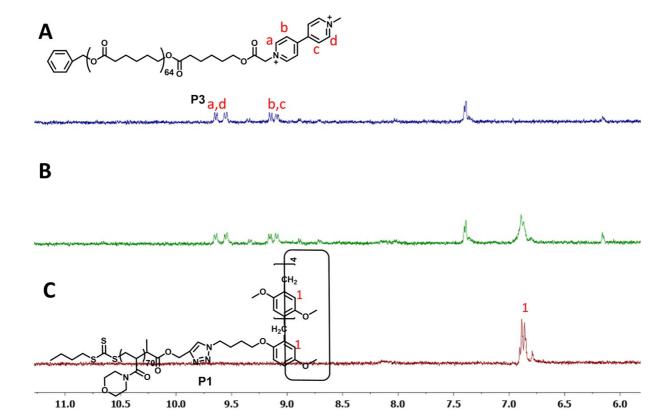
**Figure S5** SEC trace of **P2** (PNAMP).



2 Figure S6 SEC trace of P4 (PCL).



5 Figure S7 SEC trace of P3 (viologen-PCL).



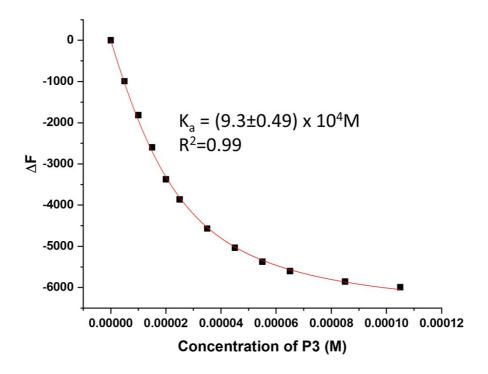
2 Figure S8 <sup>1</sup>H NMR (acetone-d6, 300 MHz) spectra of A) 4 mM P3 (viologen-PCL), B) 1/1 mixture

 $\delta$  (ppm)

3 of P3 (viologen-PCL) and P1 (PNAM), C) 4 mM P1 (PNAM) measured at 25 °C.

1

4



**Figure S9** Change of fluorescence intensity (**P2**) with increasing concentration of **P3**. The red line represents a corresponding exponential fit from which the association constant  $K_a$  can be calculated according to:  $\Delta F = (\Delta F \infty / [H]_0)(0.5[G]_0 + 0.5([H]_0 + 1/Ka) - (0.5([G]_0^2 + (2[G]_0(1/Ka - [H]_0)) + (1/Ka + [H]_0)^2)^{0.5}))$ .  $\Delta F$  is the difference in fluorescence intensity at 240 nm at  $[H]_0$  (**P2**),  $\Delta F \infty$  is the difference in fluorescence intensity at 240 nm when **P2** is completely complexed.  $[H]_0$  is the fixed initial concentration of **P2** and  $[G]_0$  is the initial concentration of **P3**.

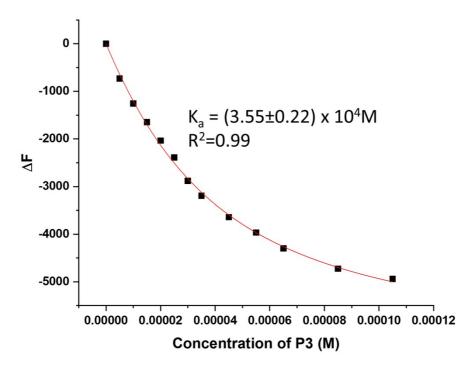
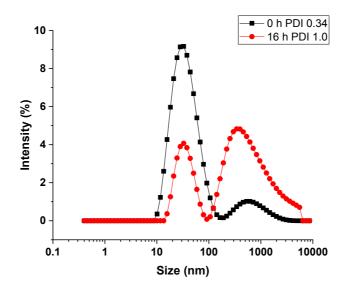


Figure S10 Change of fluorescence intensity (P1) with increasing concentration of P3. The red line represents a corresponding exponential fit from which the association constant  $K_a$  can be calculated according to:  $\Delta F = (\Delta F \infty/[H]_0)(0.5[G]_0 + 0.5([H]_0 + 1/Ka) - (0.5([G]_0^2 + (2[G]_0(1/Ka - [H]_0)) + (1/Ka + [H]_0)^2)^{0.5}))$ .  $\Delta F$  is the difference in fluorescence intensity at 240 nm at  $[H]_0$  (P1),  $\Delta F \infty$  is the difference in fluorescence intensity at 240 nm when P1 is completely complexed.  $[H]_0$  is the fixed initial concentration of P1 and  $[G]_0$  is the initial concentration of P3.



2 Figure S11 Size distribution (intensity weighted) of the particles formed by P1+P3 after preparation

3 (0 h) and after 16 h measured by DLS in PBS.

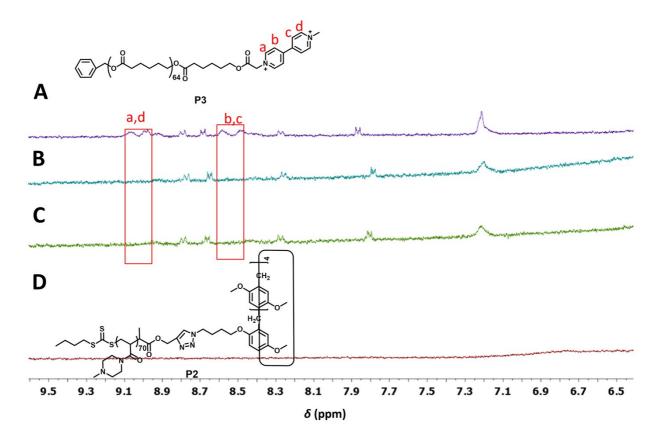


Figure S12 <sup>1</sup>H NMR (D<sub>2</sub>O, 300 MHz) spectra of A) 4 mM P3 (viologen-PCL), B) 1/1 mixture of P3 (viologen-PCL) and P2 (PNAMP), C) 1/1 mixture of P3 (viologen-PCL) and P2 (PNAMP) in PBS, D) 4 mM P2 (PNAMP) measured at 25 °C. The additionally visible signals belong to small impurities (< 5%, see also Figure S8) of free viologen salts, which are intensified due to their comparably good solubility and could not fully be removed by the purification steps.

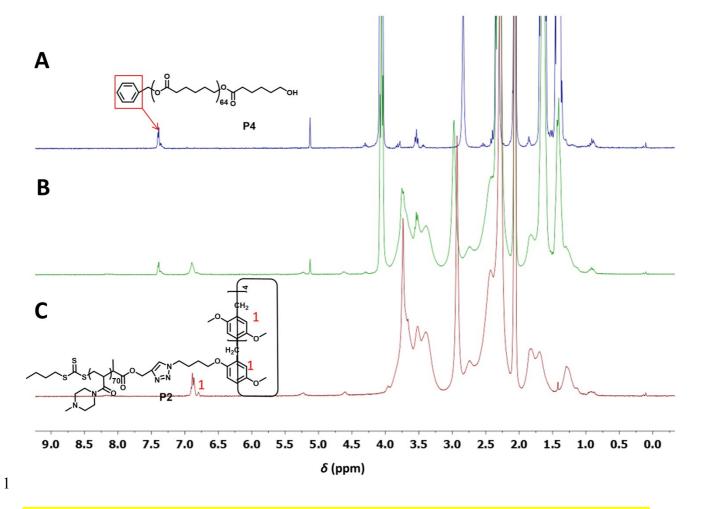
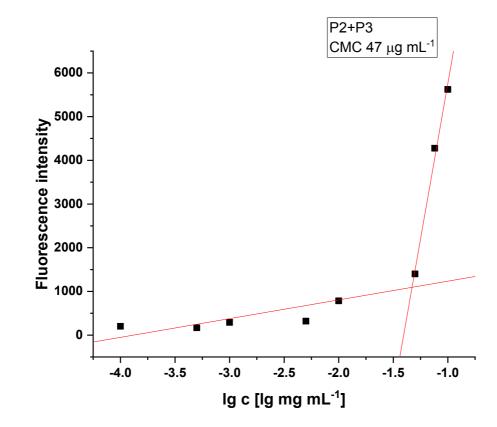


Figure S13 <sup>1</sup>H NMR (acetone-d6, 300 MHz) spectra of A) 4 mM P4 (PCL), B) 1/1 mixture of P4

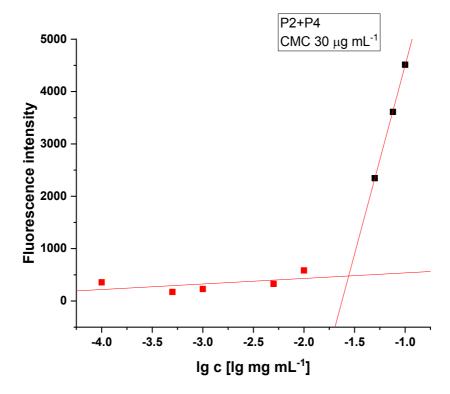
3 (PCL) and **P2** (PNAMP), C) 4 mM **P2** (PNAMP) measured at 25 °C.



2 Figure S14 Fluorescence intensity of Nile Red at various concentrations of P2+P3. The CMC was

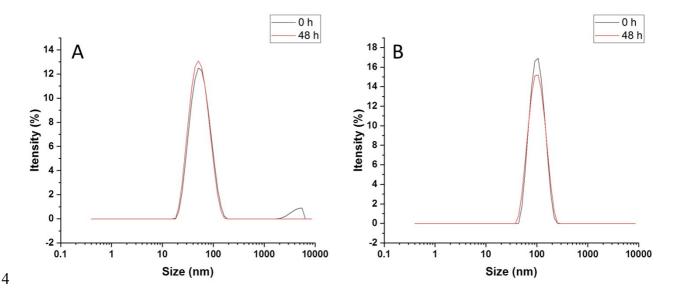
3 determined as the intersection of the corresponding linear fits (red lines).

1



2 Figure S15 Fluorescence intensity of Nile Red at various concentrations of P2+P4. The CMC was

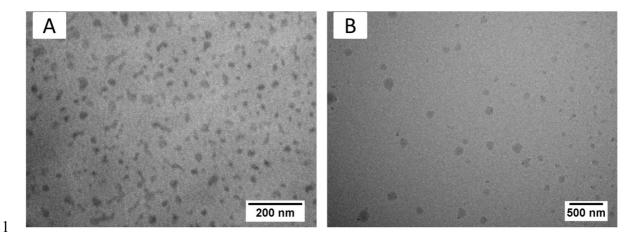
3 determined as the intersection of the corresponding linear fits (red lines).



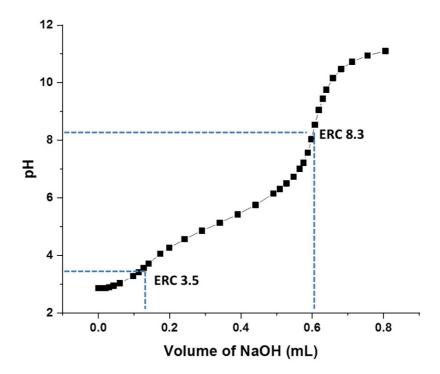
5 Figure S16 Size distribution (intensity weighted) of the particles formed by A) P2+P3 and B) P2+P4

6 in cell culture medium after 48 h.

1

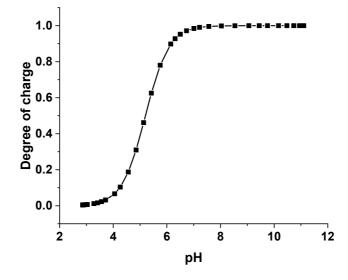


2 Figure S17 TEM images of A) P3 and B) P4.

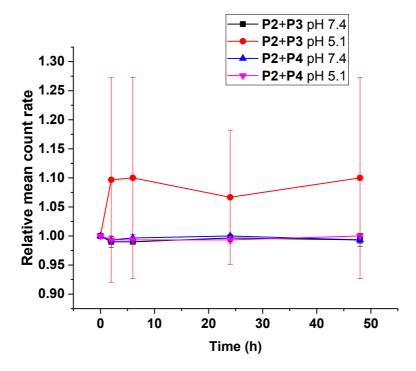


7 Figure S18 Titration of P2 (1 mg mL<sup>-1</sup>) at 25 °C with NaOH (0.1 M) after acidification with HCl

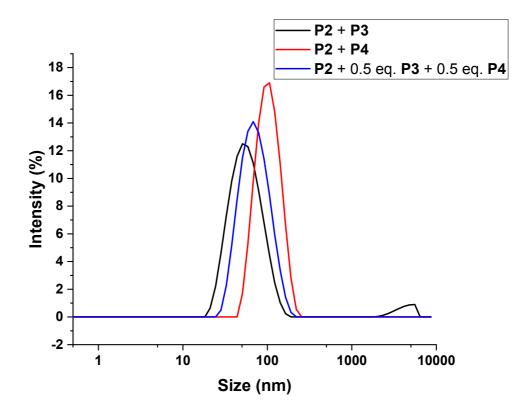
8 (0.1 M). NaCl (0.1 M) was added to simulate physiological conditions.



- 2 Figure S19 Degree of charge of PNAMP (P2) segments as a function of pH; the dashed line
- 3 corresponds to the behavior of a dilute **P2** solution (intrinsic  $pK_a$  5.2).



- 5 Figure S20: Relative mean count rate of the respective dispersions at different pH at 37 °C
- 6 determined by DLS.



2 Figure S21: Size distributions (intensity weighted) of the respective nanoparticles measured by DLS.