

Supporting information

One-step, rapid fluorescent sensing of fungal viability based on a bioprobe with aggregation-induced emission characteristics

*Xiaoxue Ge,^{†ab} Meng Gao,^{†c} Bo Situ,^{†ab} Weiwei Feng,^{ab} Bairong He,^{ab} Xiaojing He,^{ab}
Shiwu Li,^d Zihao Ou,^{ab} Yiqi Zhong,^{ab} Yahui Lin,^{ab} Xinyi Ye,^{ab} Xiumei Hu,^{ab} Ben Zhong
Tang^{*def} and Lei Zheng^{*ab}*

^aDepartment of Laboratory Medicine, Nanfang Hospital, Southern Medical University, Guangzhou, 510515, China

^bGuangdong Engineering and Technology Research Center for Rapid Diagnostic Biosensors, Nanfang Hospital, Southern Medical University, Guangzhou, 510515, China

^cNational Engineering Research Center for Tissue Restoration and Reconstruction, South China University of Technology, Guangzhou, 510006, China

^dCenter for Aggregation-Induced Emission, State Key Laboratory of Luminescent Materials and Devices, South China University of Technology, Guangzhou, 510640, China

^eDepartment of Chemistry, Hong Kong Branch of Chinese National Engineering Research Center for Tissue Restoration and Reconstruction, The Hong Kong University of Science & Technology, Clear Water Bay, Kowloon, Hong Kong, China

^fHKUST Shenzhen Research Institute, No. 9 Yuexing 1st RD, South Area, Hi-tech Park, Nanshan, Shenzhen, 518057, China

[†] These authors contributed equally to this work.

Corresponding authors: nfyzhenglei@smu.edu.cn (L.Z.), tangbenz@ust.hk (B.Z.T.)

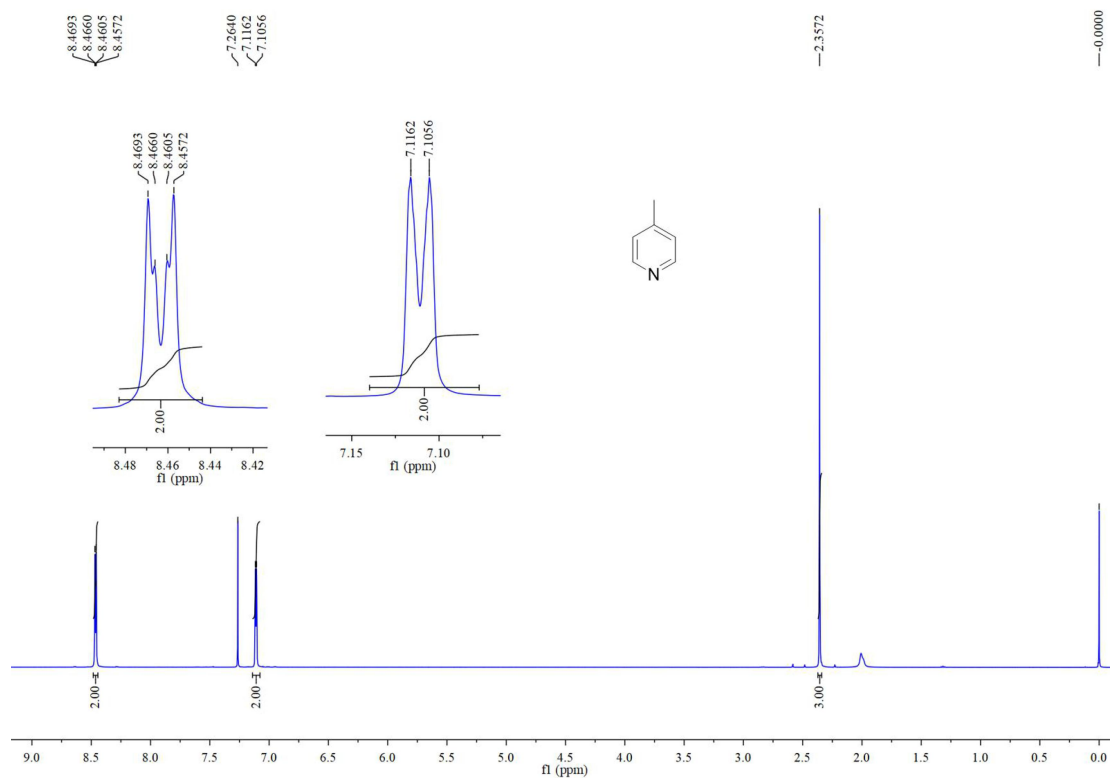


Fig. S1 ^1H NMR of compound **1** (500 MHz, CDCl_3).

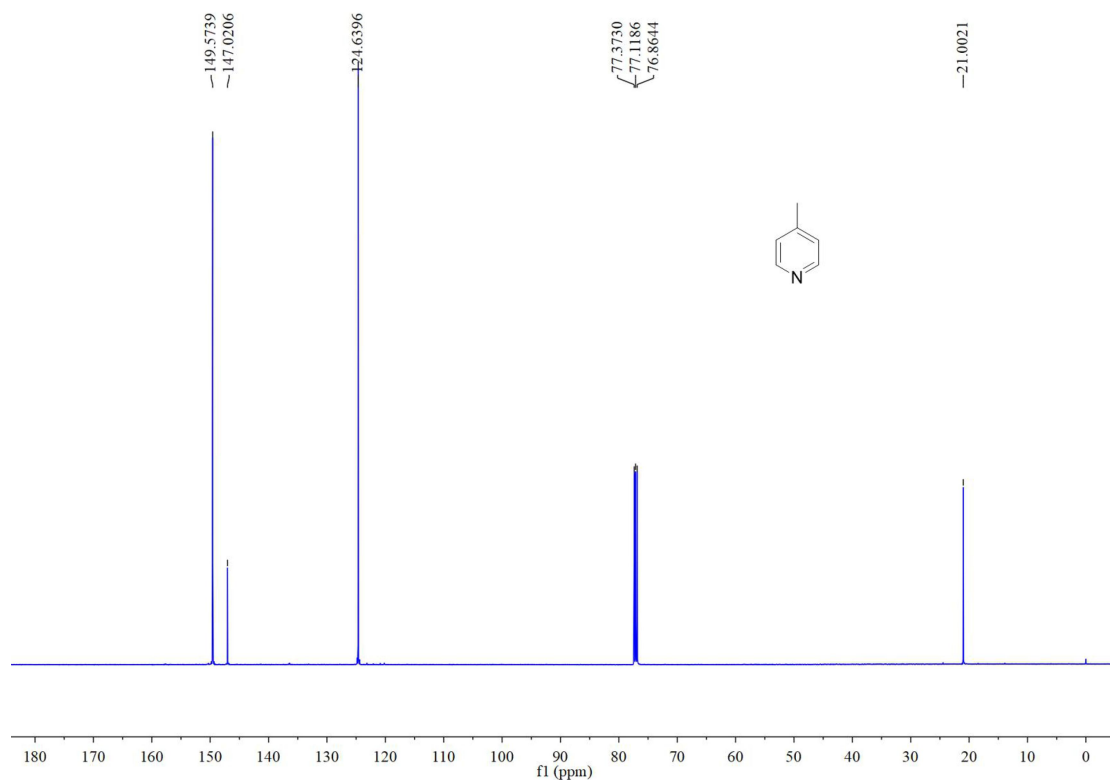


Fig. S2 ^{13}C NMR of compound **1** (125 MHz, CDCl_3).

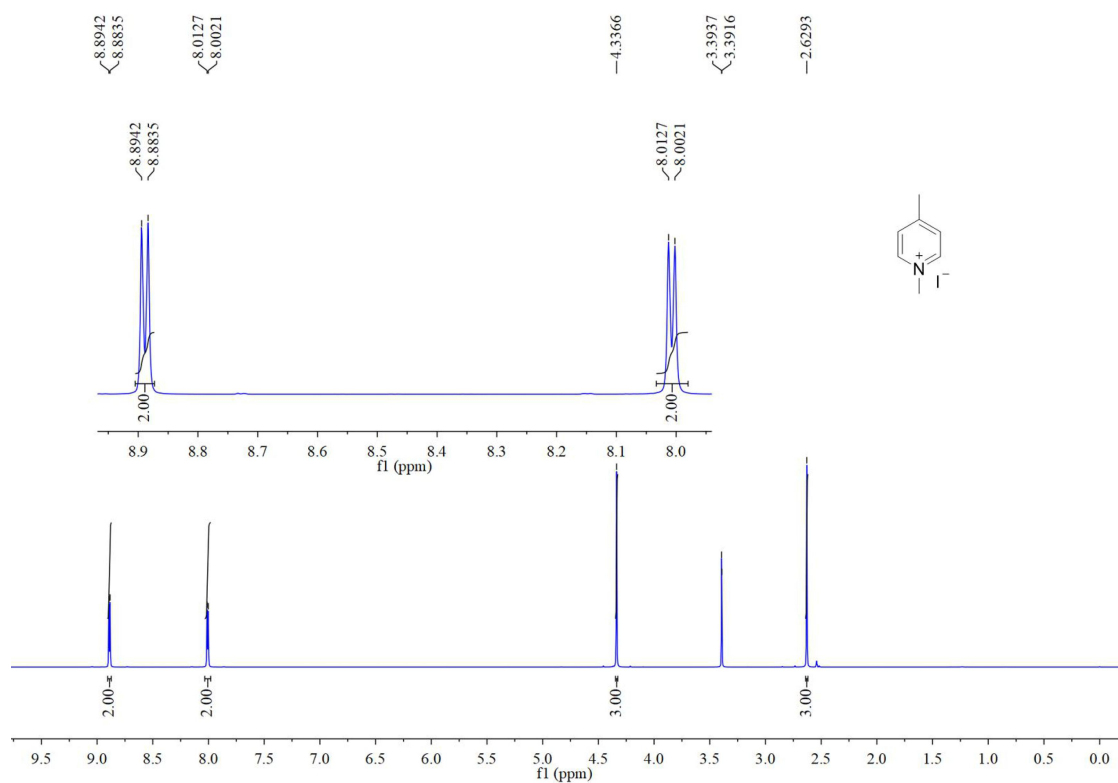


Fig. S3 ^1H NMR of compound **2** (600 MHz, $\text{DMSO-}d_6$).

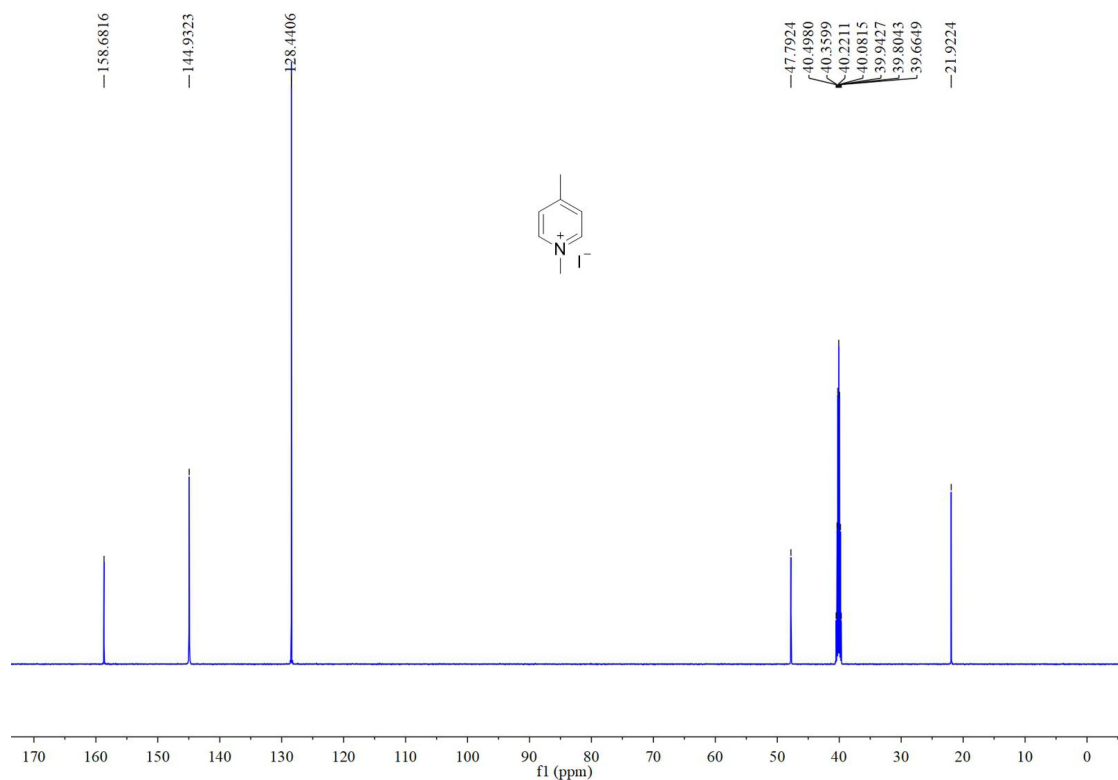


Fig. S4 ^{13}C NMR of compound **2** (150 MHz, $\text{DMSO-}d_6$).

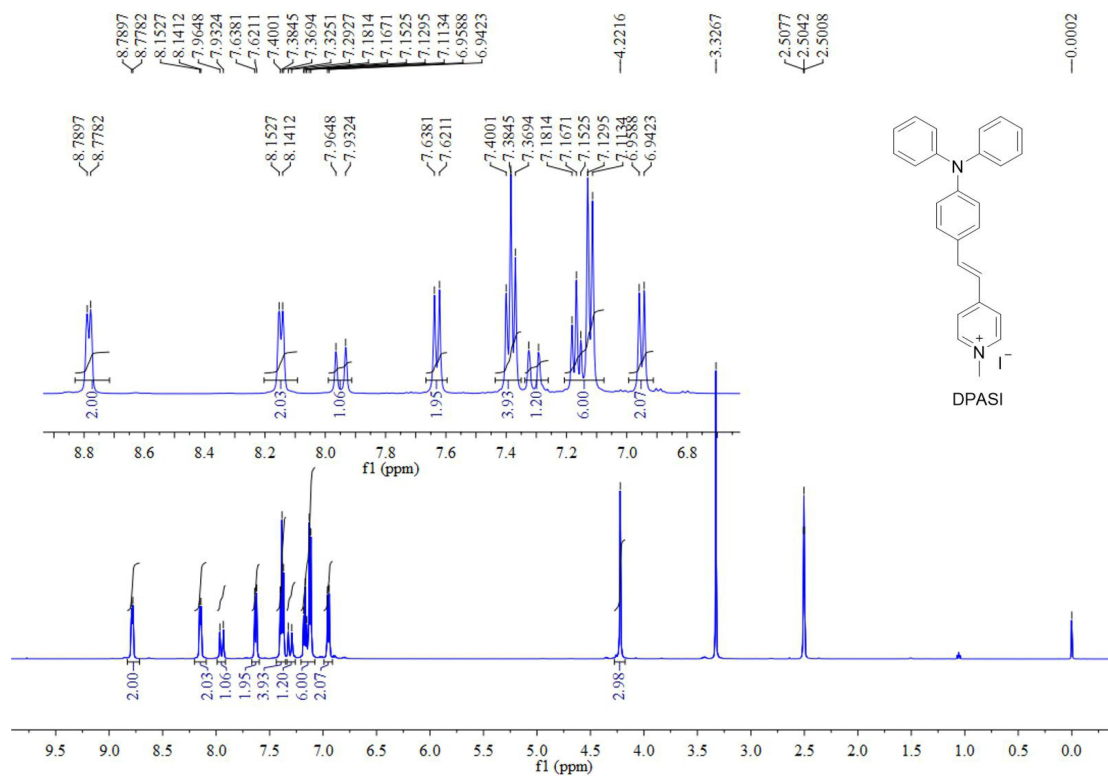


Fig. S5 ^1H NMR of DPASI (500 MHz, $\text{DMSO-}d_6$).

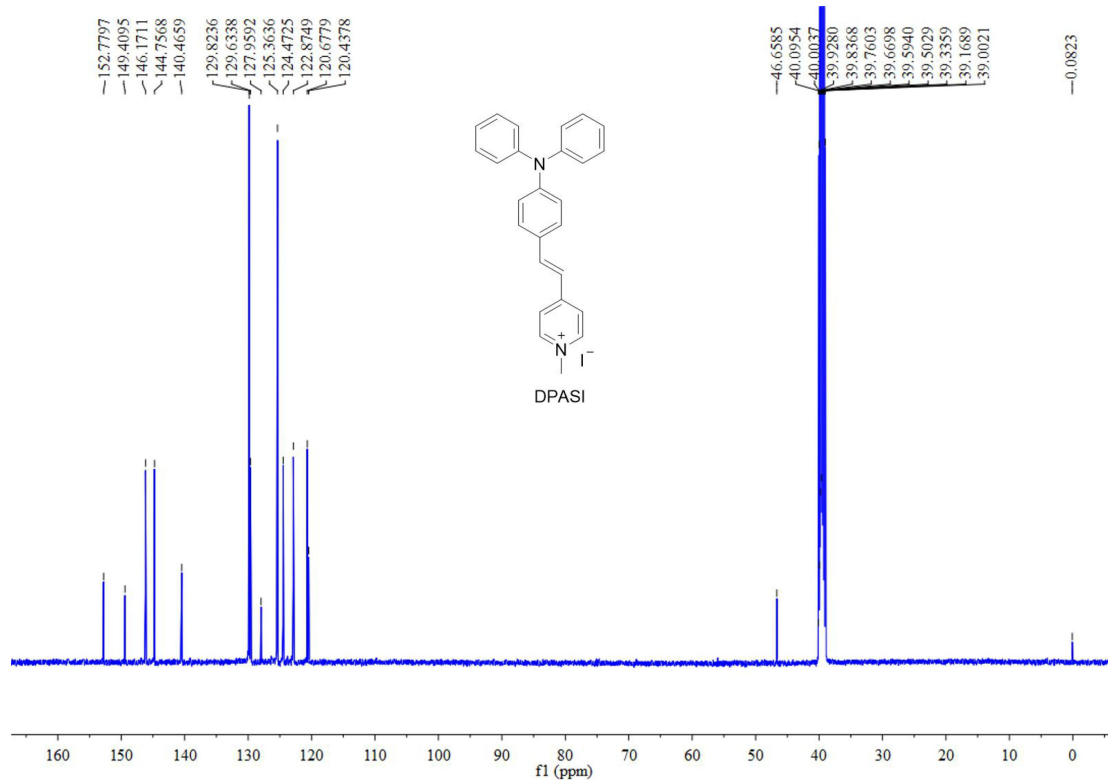


Fig. S6 ^{13}C NMR of DPASI (125 MHz, $\text{DMSO-}d_6$).

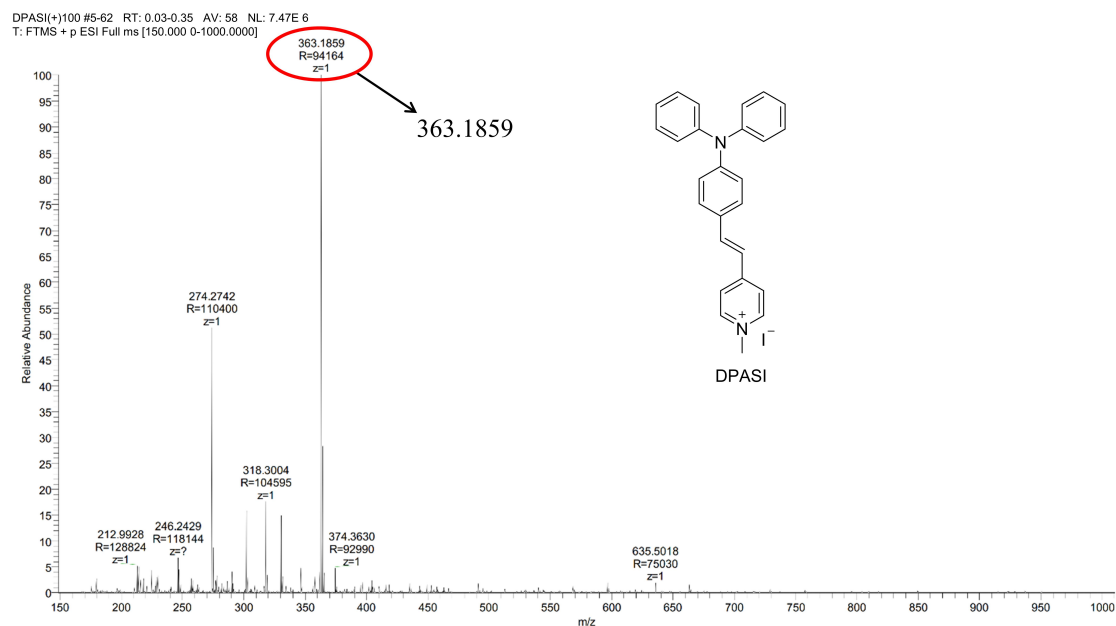


Fig. S7 High resolution mass spectra of DPASI.

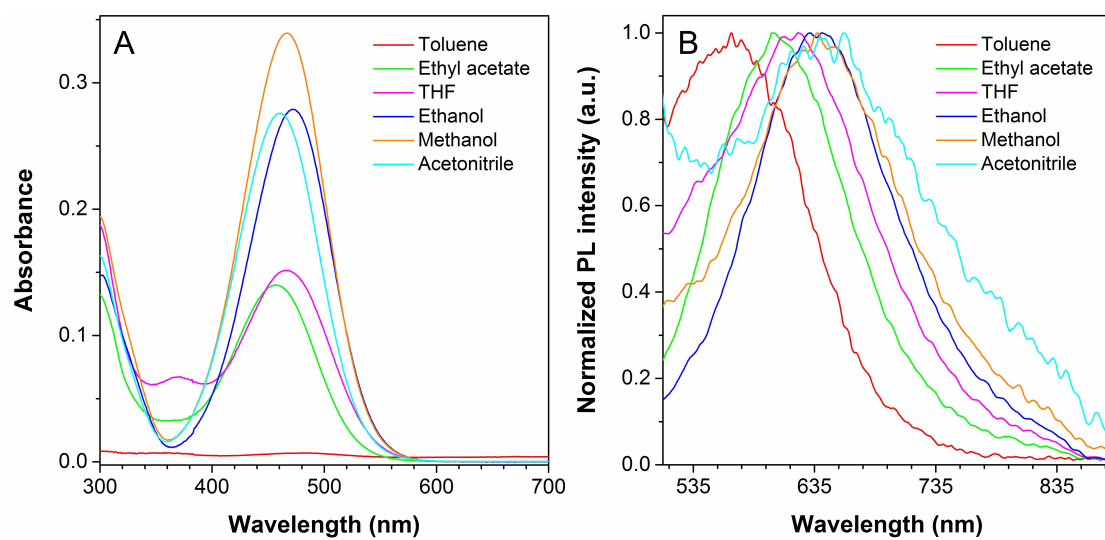


Fig. S8 (A) UV-vis absorption spectra of DPASI (10 μ M) in solvents with different polarities. (B) PL spectra of DPASI (10 μ M) in solvents with different polarities. λ_{ex} = 467 nm.

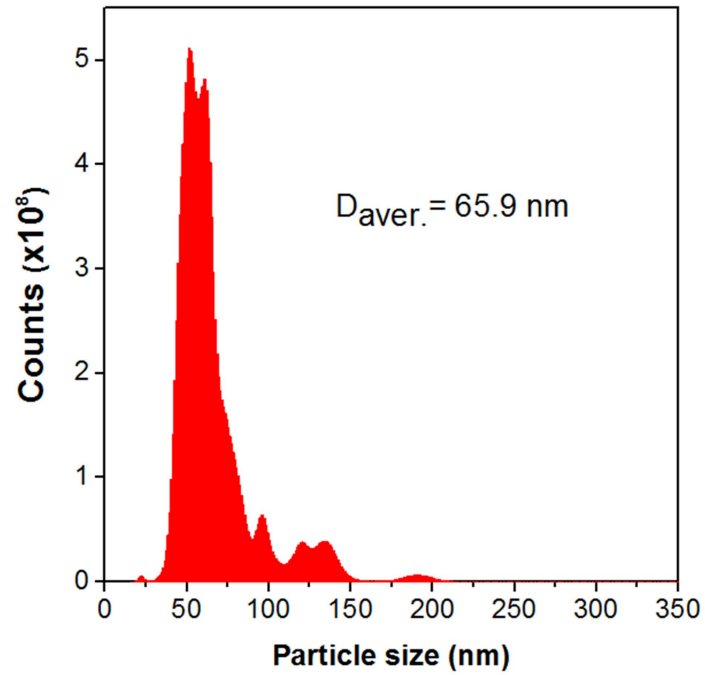


Fig. S9 Particle size of aggregates of DPASI (10 μ M) formed in phosphate buffer.

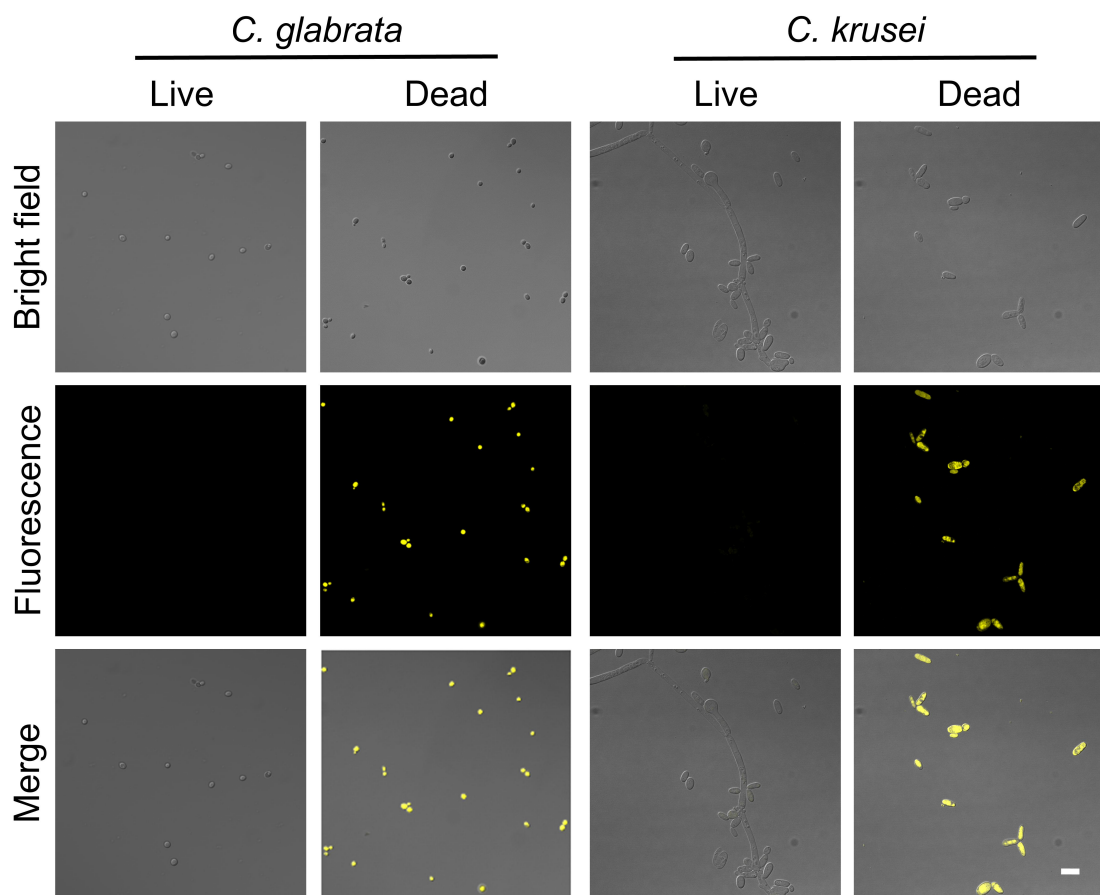


Fig. S10 CLSM images of *C. glabrata* and *C. krusei* stained with 10 μ M DPASI. λ_{ex} = 488 nm; Scale bar = 10 μ m.

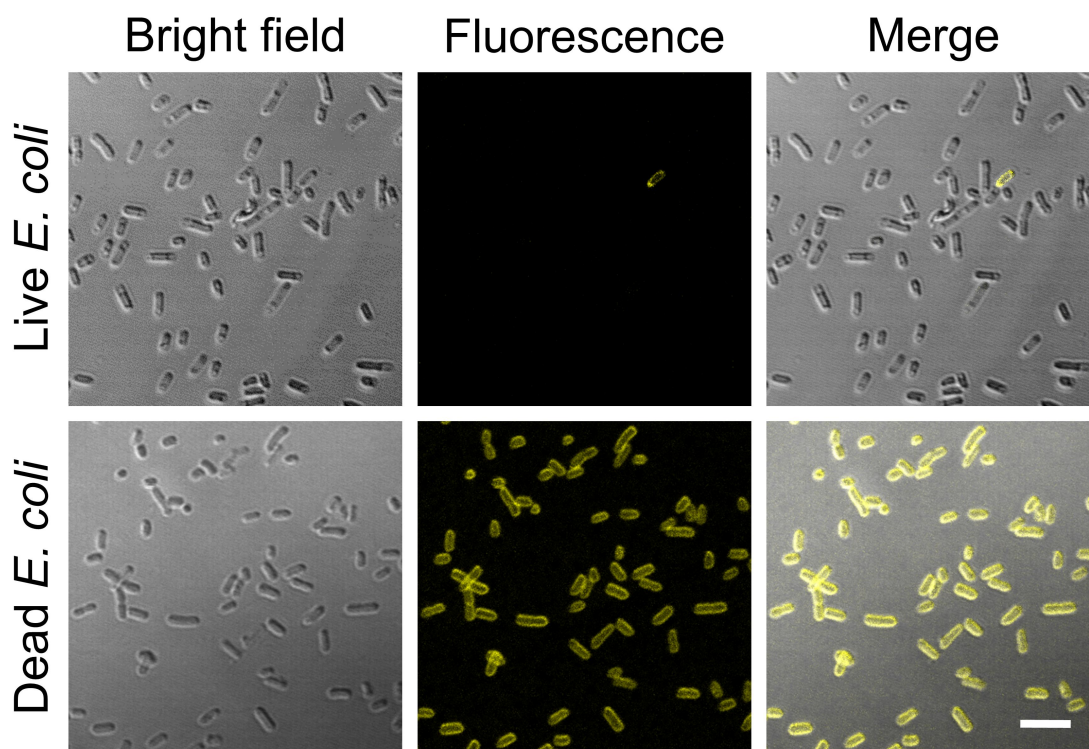


Fig. S11 CLSM images of live and dead (treated with 75% ethanol for 15 minutes) *E. coli* stained with $10\ \mu\text{M}$ DPASI. $\lambda_{\text{ex}} = 488\ \text{nm}$; Scale bar = $5\ \mu\text{m}$.

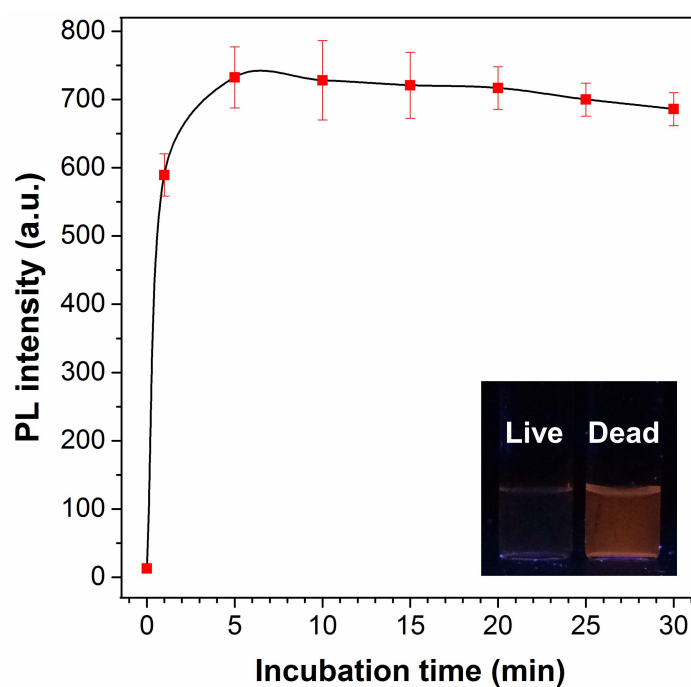


Fig. S12 Change in the PL intensity of DPASI in *C. albicans* suspensions versus different incubation time. $[\text{DPASI}] = 10\ \mu\text{M}$; $\lambda_{\text{ex}} = 467\ \text{nm}$. Inset: Fluorescence photograph of DPASI in *C. albicans* suspensions taken under 365 nm UV irradiation. Error bars indicate the standard error of mean (\pm SEM).

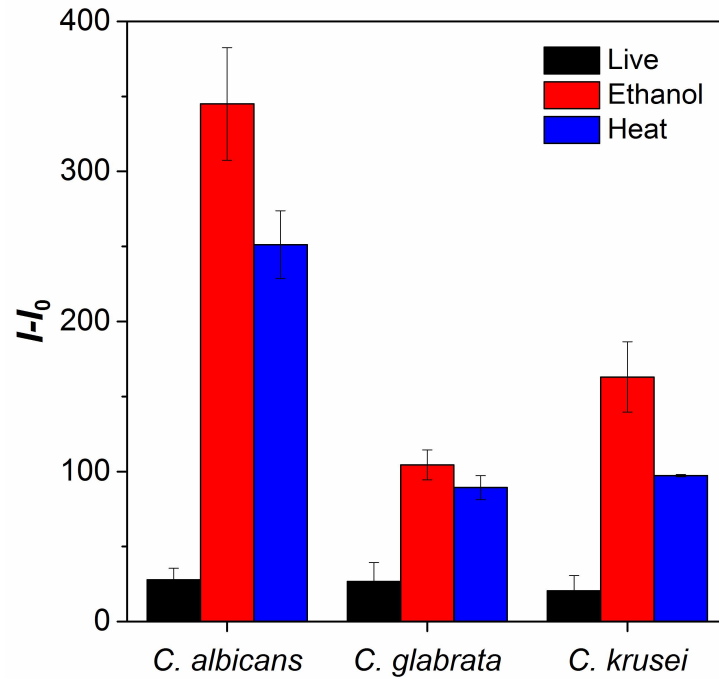


Fig. S13 Fluorescence intensity of *Candida* suspensions with different treatment followed by staining with DPASI (10 μ M); λ_{ex} = 467 nm. Error bars indicate the standard error of mean (\pm SEM).

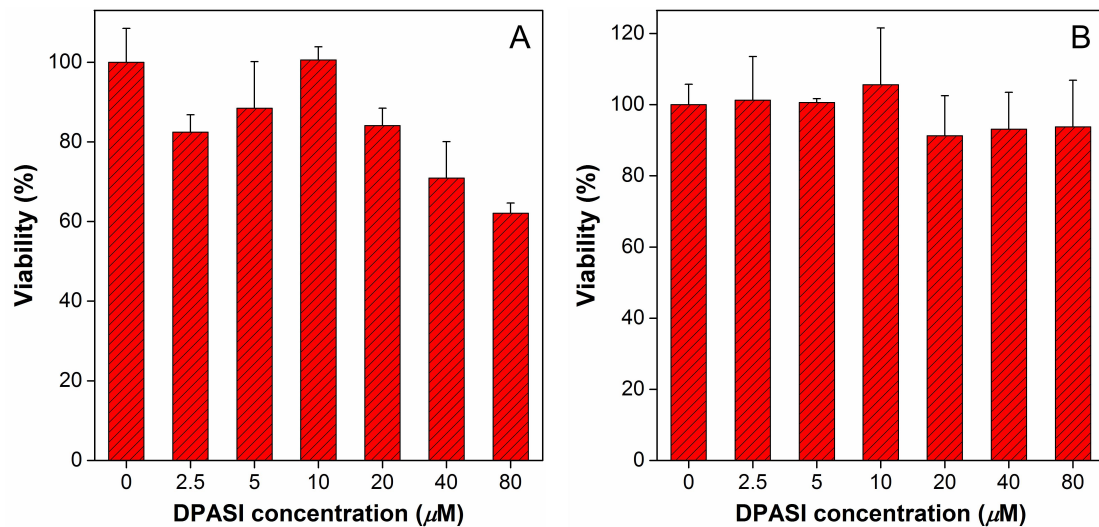


Fig. S14 Viability of *C. glabrata* (A) and *C. krusei* (B) in the presence of different concentrations of DPASI determined by counting after 2-hours or 1-hour incubation, respectively. Error bars indicate the standard error of mean (\pm SEM).

Additional supporting movie

Movie S1. Representative 3D image of the boxed *Candida* cell in Fig. 5 stained by DAPI, Calcofluor White Staining, and DPASI under SIM excited by 405 nm and 488 nm laser.