

## Reactions of 2-Aza-21-carbaporphyrin with Aniline Derivatives

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## General methods and instrumentation

Commercial reagents were used without further purification. Solvents were freshly distilled from the appropriate drying agents or purified under nitrogen with the mBraun MBSPS-800 before use. The analytical TLCs were performed with silica gel 60 F254 plates. Column chromatography was performed by using silica gel 60 (200-300 mesh ASTM). The NMR spectra were recorded on a Bruker Avance II spectrometer, operating at 500 MHz for  $^1\text{H}$  and 125 MHz for  $^{13}\text{C}$  or a Bruker Avance II spectrometer operating at 600 MHz for  $^1\text{H}$  and 150 MHz for  $^{13}\text{C}$ . TMS was used as an internal reference for  $^1\text{H}$  and  $^{13}\text{C}$  chemical shifts and  $\text{CDCl}_3$  was used as solvent. Standard pulse programs from the Bruker library were used for homo- and heteronuclear 2D experiments. Mass spectrometry measurements were conducted by using the electrospray ionization technique on a Bruker Daltonics microTOF-Q or Finnigan LCQ Advantage MAX mass spectrometer. Absorption UV/Vis/NIR spectra were recorded by using a Varian Cary 50 Bio and Jasco V-770 spectrophotometers. HPLC separations were carried out by means of Chirex 3010 analytical column (25 cm length, 4.6 mm i.d.) packed with 5 mm silica gel coated with covalently bound (S)-valine and dinitroaniline using Shimadzu chromatographic system or Merck-Hitachi LaChrom series connected to a flow-cell mounted on Jasco J-1500 spectropolarimeter. HPLC-grade hexane was used. The applied dichloromethane was freshly distilled over calcium hydride. Circular dichroism spectra were obtained directly in the flow cell by means of pseudo stopped-flow technique using a Jasco J-1500 spectropolarimeter. Electrochemical measurements were performed by means of Autolab (Metrohm) potentiostat/galvanostat system for dichloromethane solutions with a glassy carbon, a platinum wire, and Ag/AgCl as the working, auxiliary, and reference electrodes, respectively. Tetrabutylammonium hexafluorophosphate was used as a supporting electrolyte. The potentials were referenced with the ferrocene/ferrocenium couple used as an internal standard. Crystal data for **2a** and **3b** were collected at low temperature using Oxford Cryosystem devices on Xcalibur Ruby and Onyx four-circle diffractometers with graphite-monochromated Mo K $\alpha$  radiation ( $\lambda = 0.71073 \text{ \AA}$ ) and Cu K $\alpha$  radiation ( $\lambda = 1.54184 \text{ \AA}$ ). Data reduction and analysis were carried out with the *CrysAlisPRO* programs<sup>1</sup>. The structures were solved by using the SHELXT program<sup>2</sup> and refined by the full-matrix least-squares method on all  $F^2$  data using the SHELXL program.<sup>3</sup> All hydrogen atoms, including those located in the difference density map, were placed in calculated positions and refined as the riding model. For **2a** the SQUEEZE/PLATON procedure<sup>4</sup> was applied for disordered lattice solvent (methanol) molecules. See Table S1 for detailed data.

**Table S1** Crystal data for **2a** and **3b**

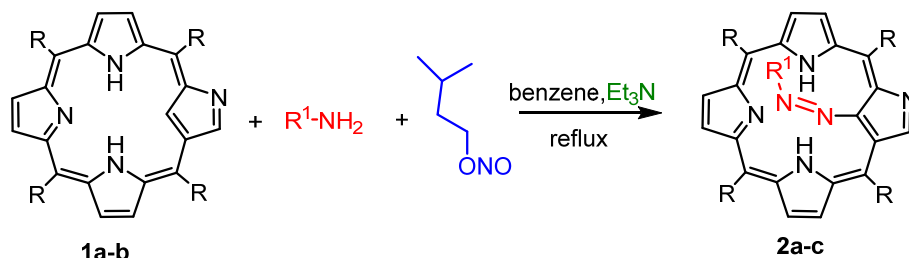
	<b>2a</b>	<b>3b</b>
<b>Chemical formula</b>	C <sub>63</sub> H <sub>58</sub> N <sub>6</sub> O <sub>14</sub> ·0.5(CH <sub>3</sub> OH)	C <sub>65</sub> H <sub>56</sub> N <sub>6</sub> O <sub>6</sub> ·2(C <sub>7</sub> H <sub>8</sub> )
<b>M<sub>r</sub></b>	1139.17	1201.43
<b>Crystal system, space group</b>	Monoclinic, Cc	Monoclinic, P2 <sub>1</sub> /c
<b>Temperature (K)</b>	90	100
<b>a, b, c (Å)</b>	36.594 (2), 7.353 (3), 28.802 (3)	33.740 (2), 12.948(3), 14.823(3)
<b>β (°)</b>	123.87 (3)	100.32 (3)
<b>V (Å<sup>3</sup>)</b>	6435 (3)	6370.7 (6)
<b>Z</b>	4	4
<b>Radiation type</b>	Mo Kα	Cu Kα
<b>μ (mm<sup>-1</sup>)</b>	0.08	0.63
<b>Crystal size (mm)</b>	0.33 × 0.16 × 0.07	0.70 × 0.10 × 0.08
<b>Diffractometer</b>	Xcalibur, Ruby	Xcalibur, Onyx
<b>No. of measured, independent and observed [I &gt; 2σ(I)] reflections</b>	11241, 8011, 5888	41715, 10157, 4894
<b>R<sub>int</sub></b>	0.033	0.137
<b>(sin θ/λ)<sub>max</sub> (Å<sup>-1</sup>)</b>	0.606	0.575
<b>Refinement</b>		
<b>R[F<sup>2</sup> &gt; 2σ(F<sup>2</sup>)], wR(F<sup>2</sup>), S</b>	0.063, 0.170, 1.04	0.068, 0.171, 0.87
<b>No. of reflections</b>	8011	10157
<b>No. of parameters</b>	771	915
<b>No. of restraints</b>	3	54
<b>H-atom treatment</b>	H-atom parameters constrained	H-atom parameters constrained
<b>Δρ<sub>max</sub>, Δρ<sub>min</sub> (e Å<sup>-3</sup>)</b>	0.56, -0.24	0.35, -0.37

**Computational methods**

Density functional theory (DFT) calculations were performed by using the Gaussian 09.E01 program<sup>5</sup>. DFT geometry optimizations were carried out in the unconstrained C<sub>1</sub> symmetry by using the X-ray structure, molecular mechanics, or semiempirical models as starting geometries. DFT geometries were refined to meet standard convergence criteria, and the existence of a local minimum was verified by a normal mode frequency calculation. DFT calculations were performed by using the hybrid B3LYP functional<sup>6-8</sup> functional combined with the 6-31G(d,p) basis set. The electronic spectra were simulated by means of time-dependent density functional theory (TD-DFT) using the Tamm-Dancoff approximation for 50 states. For TD calculations, the polarizable continuum model of solvation was used (PCM, standard dichloromethane parametrization). The electronic transitions and UV/Vis/NIR as well as CD spectra were analyzed by means of the GaussSum program.<sup>9</sup> The transitions were convoluted by Gaussian curves with 2000 cm<sup>-1</sup> half line width.

## Synthesis and characterization

**Synthesis of the precursor.** Starting porphyrins **1a** and **1b** were obtained as described previously.<sup>10</sup>



**1a:** R=3,4,5-(OCH<sub>3</sub>)<sub>3</sub>C<sub>6</sub>H<sub>2</sub>

**1b:** R=4-CH<sub>3</sub>C<sub>6</sub>H<sub>4</sub>

**2a:** R=3,4,5-(OCH<sub>3</sub>)<sub>3</sub>C<sub>6</sub>H<sub>2</sub>, R<sup>1</sup>=2-COOHC<sub>6</sub>H<sub>4</sub>

**2b:** R=4-CH<sub>3</sub>C<sub>6</sub>H<sub>4</sub>, R<sup>1</sup>=2-COOHC<sub>6</sub>H<sub>4</sub>

**2c:** R=4-CH<sub>3</sub>C<sub>6</sub>H<sub>4</sub>, R<sup>1</sup>=2-COOH-4-CH<sub>3</sub>C<sub>6</sub>H<sub>3</sub>

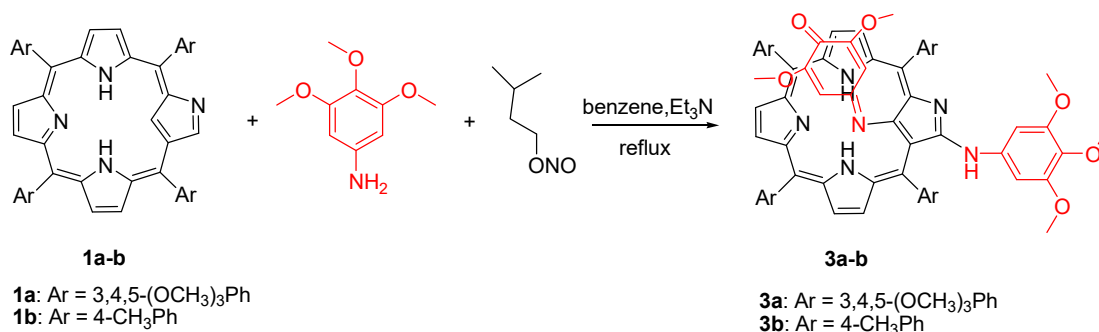
**General method of synthesis of 2.** To a solution of NCP **1** (0.1 mmol) and 2-aminobenzoic acid (0.3 mmol) in 5 ml dry benzene was added triethylamine (100  $\mu$ L, 0.7 mmol) and amyl nitrite (100  $\mu$ L, 0.7 mmol), the mixture was stirred under nitrogen for 2 hours under reflux and then the solvent was removed under vacuum. The residue was passed down a silica gel column with dichloromethane/ethyl acetate solvent as eluent. The first red colored product fraction was collected, followed by removal of solvent to afford desire NCP derivative **2** that was crystallized from dichloromethane solution by addition of hexane. Yields: 85% for **2a**, 80% for **2b**, 65% for **2c**.

Selected data for **2a**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K):  $\delta$  = 1.55 (d, *J* = 8.0 Hz, 1H, ArH), 3.73 (s, 3H, -OCH<sub>3</sub>), 3.96 (s, 6H, -OCH<sub>3</sub>), 3.97 (s, 3H, -OCH<sub>3</sub>), 4.01 (s, 3H, -OCH<sub>3</sub>), 4.08-4.09 (m, 9H, -OCH<sub>3</sub>), 4.13 (s, 3H, -OCH<sub>3</sub>), 4.16 (s, 3H, -OCH<sub>3</sub>), 4.17 (s, 3H, -OCH<sub>3</sub>), 4.19 (s, 3H, -OCH<sub>3</sub>), 5.54 (t, *J* = 7.5 Hz, 1H, ArH), 6.30 (t, *J* = 8.0 Hz, 1H, ArH), 6.86 (s, 1H, ArH), 7.10 (s, 1H, ArH), 7.12 (d, *J* = 8.0 Hz, 1H, ArH), 7.31 (s, 1H, ArH), 7.46 (s, 1H, ArH), 7.61-7.64 (m, 4H), 8.36 (d, *J* = 4.5 Hz, 1H, pyrRH), 8.44 (d, *J* = 4.0 Hz, 1H, pyrRH), 8.69 (d, *J* = 4.0 Hz, 1H, pyrRH), 8.81 (d, *J* = 4.5 Hz, 1H, pyrRH), 8.92 (s, 1H, pyrRH), 9.17 (d, *J* = 4.5 Hz, 2H, pyrRH), 9.80 (s, 1H, -COOH); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 248 K):  $\delta$  = -3.61 (br, 1H, -NH), -2.16 (br, 1H, -NH), 1.40 (d, *J* = 8.0 Hz, 1H, ArH), 3.94 (s, 3H, -OCH<sub>3</sub>), 3.96 (s, 3H, -OCH<sub>3</sub>), 4.00-4.04 (m, 12H, -OCH<sub>3</sub>), 4.12 (s, 3H, -OCH<sub>3</sub>), 4.15-4.20 (m, 12H, -OCH<sub>3</sub>), 4.27 (s, 3H, -OCH<sub>3</sub>), 5.60 (t, *J* = 8.0 Hz, 1H, ArH), 6.35 (t, *J* = 7.5 Hz, 1H, ArH), 6.81 (br, 1H, ArH), 7.11 (d, *J* = 7.5 Hz, 1H, ArH), 7.27 (s, 2H, ArH), 7.33 (s, 1H, ArH), 7.38 (s, 1H, ArH), 7.50 (s, 1H, ArH), 7.96 (d, *J* = 12.0 Hz, 1H, ArH), 8.39 (s, 1H, pyrRH), 8.51 (d, *J* = 4.0 Hz, 1H, pyrRH), 8.69 (s, 1H, pyrRH), 8.86 (s, 1H, pyrRH), 8.93 (s, 1H, pyrRH), 9.23 (s, 1H, pyrRH), 9.26 (s, 1H, pyrRH), 10.27 (s, 1H, COOH); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, 298 K):  $\delta$  = 56.4, 56.47, 56.50, 56.7, 61.2, 61.27, 61.33, 61.37, 110.3, 112.3, 112.7, 112.9, 114.8, 120.9, 122.3, 122.7, 127.2, 127.6, 127.8, 127.9, 129.6, 130.1, 130.2, 130.5, 135.4, 135.6, 135.7, 136.2, 136.4, 136.6, 137.6, 138.1, 138.3, 138.9, 139.6, 140.2, 140.3, 146.7, 151.4, 151.59, 151.64, 151.7, 152.5, 152.7, 157.2, 157.7, 165.6. UV-vis (CHCl<sub>3</sub>):  $\lambda_{max}$ /nm (log  $\epsilon$ ) = 403(4.76), 510(4.54), 740(3.94). ESI-HRMS calc. for [C<sub>63</sub>H<sub>59</sub>N<sub>6</sub>O<sub>14</sub>]<sup>+</sup> (M+H): 1123.4084, Found: 1123.4078.

Selected data for **2b**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K):  $\delta$  = 2.50 (d, *J* = 8.0 Hz, 1H, ArH), 2.65 (s, 3H, -CH<sub>3</sub>), 2.68 (s, 3H, -CH<sub>3</sub>), 2.69 (s, 3H, -CH<sub>3</sub>), 2.71 (s, 6H, -CH<sub>3</sub>), 5.50 (t, *J* = 7.5 Hz, 1H, ArH), 6.24 (t, *J* = 7.5 Hz, 1H, ArH), 7.04 (d, *J* = 7.5 Hz, 1H, ArH), 7.47 (t, *J* = 7.5 Hz, 3H, ArH), 7.56 (t, *J* = 8.0 Hz, 3H, ArH), 7.58 (d, *J* = 7.5 Hz, H, ArH), 7.68 (t, *J* = 7.5 Hz, 4H, ArH), 7.76 (d, *J* = 7.5 Hz, 1H, ArH), 7.90 (d, *J* = 7.0 Hz,

1H, ArH), 8.12 (d,  $J = 7.5$  Hz, 1H, ArH), 8.28 (d,  $J = 7.5$  Hz, 2H, ArH), 8.31 (d,  $J = 4.5$  Hz, 1H, pyrRH), 8.38 (d,  $J = 4.5$  Hz, 1H, pyrRH), 8.56 (d,  $J = 5.0$  Hz, 1H, pyrRH), 8.72 (d,  $J = 4.5$  Hz, 1H, pyrRH), 8.75 (s, 1H, pyrRH), 9.08 (d,  $J = 4.5$  Hz, 1H, pyrRH), 9.13 (d,  $J = 4.5$  Hz, 1H, pyrRH), 9.92 (s, 1H, -COOH);  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ , 248 K):  $\delta = -3.45$  (br, 1H, -NH), -2.35 (br, 1H, -NH), 1.18 (d,  $J = 8.0$  Hz, 1H, ArH), 2.69 (s, 3H, - $\text{CH}_3$ ), 2.73 (s, 3H, - $\text{CH}_3$ ), 2.76 (s, 6H, - $\text{CH}_3$ ), 5.61 (t,  $J = 7.5$  Hz, 1H, ArH), 6.34 (t,  $J = 7.5$  Hz, 1H, ArH), 7.06 (d,  $J = 7.5$  Hz, 1H, ArH), 7.51-7.53 (m, 2H, ArH), 7.76-7.86 (m, 4H, ArH), 7.86 (br, 2H, ArH), 7.94 (br, 1H, ArH), 8.06 (br, 1H, ArH), 8.27 (br, 1H, ArH), 8.39 (br, 1H, ArH), 8.49 (br, 1H, ArH), 8.60 (br, 2H, pyrRH), 8.78 (br, 2H, pyrRH), 8.86 (br, 1H, pyrRH), 9.19 (br, 1H, pyrRH), 9.24 (br, 1H, pyrRH), 10.34 (s, 1H, COOH);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ , 298 K):  $\delta = 21.45, 21.48, 21.51, 21.7, 110.3, 115.55, 115.62, 120.6, 122.2, 122.5, 124.0, 125.8, 126.0, 127.0, 127.4, 127.7, 127.8, 129.0, 129.8, 130.1, 130.2, 132.4, 133.4, 134.0, 134.2, 134.3, 134.7, 135.3, 135.4, 137.1, 137.2, 137.3, 137.7, 137.99, 138.06, 138.15, 138.18, 138.7, 139.4, 140.2, 140.3, 140.4, 146.9, 151.0, 157.1, 157.4, 165.9$ . UV-vis ( $\text{CHCl}_3$ ):  $\lambda_{\text{max}}/\text{nm}$  ( $\log \epsilon$ ) = 394(4.75), 501(4.36), 739(3.88). ESI-HRMS calc. for  $[\text{C}_{55}\text{H}_{43}\text{N}_6\text{O}_2]^+$  (M+H): 819.3442, Found: 819.3455.

Selected data for **2c**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ , 298 K):  $\delta = 1.37$  (br, 1H, ArH), 1.68 (s, 3H, - $\text{CH}_3$ ), 2.65 (s, 3H, - $\text{CH}_3$ ), 2.68 (s, 3H, - $\text{CH}_3$ ), 2.69 (s, 3H, - $\text{CH}_3$ ), 2.70 (s, 3H, - $\text{CH}_3$ ), 5.34 (d,  $J = 8.0$  Hz, 1H, ArH), 6.90 (br, 1H, ArH), 7.47 (t,  $J = 6.5$  Hz, 3H, ArH), 7.54 (d,  $J = 7.5$  Hz, 1H, ArH), 7.58 (d,  $J = 7.5$  Hz, 2H, ArH), 7.68 (t,  $J = 6.5$  Hz, 1H, ArH), 7.76 (d,  $J = 7.5$  Hz, 1H, ArH), 7.90 (d,  $J = 7.5$  Hz, 1H, ArH), 8.12 (d,  $J = 7.5$  Hz, 1H, ArH), 8.26 (d,  $J = 7.5$  Hz, 2H, ArH), 8.29-8.30(m, 1H, ArH), 8.32 (d,  $J = 4.5$  Hz, 1H, pyrRH), 8.39 (d,  $J = 4.5$  Hz, 1H, pyrRH), 8.53 (d,  $J = 4.5$  Hz, 1H, pyrRH), 8.63 (br, 1H, pyrRH), 8.68 (d,  $J = 4.5$  Hz, 1H, pyrRH), 9.06 (d,  $J = 5.0$  Hz, 1H, pyrRH), 9.10 (d,  $J = 4.5$  Hz, 1H, pyrRH), 9.93 (s, 1H, -COOH);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ , 298 K):  $\delta = 20.5, 21.50, 21.54, 21.55, 21.60, 110.4, 120.6, 122.0, 122.3, 127.3, 127.5, 127.6, 127.8, 128.7, 129.0, 130.1, 130.5, 130.8, 134.1, 134.2, 134.4, 134.7, 135.3, 135.5, 137.1, 137.27, 137.32, 137.6, 137.9, 138.0, 138.1, 138.2, 138.3, 138.7, 139.4, 140.2, 140.4, 145.2, 157.3, 157.5, 166.1$ . UV-vis ( $\text{CHCl}_3$ ):  $\lambda_{\text{max}}/\text{nm}$  ( $\log \epsilon$ ) = 394(4.76), 498(4.34), 738(3.81). ESI-HRMS calc. for  $[\text{C}_{56}\text{H}_{45}\text{N}_6\text{O}_2]^+$  (M+H): 833.3599, Found: 833.3604.

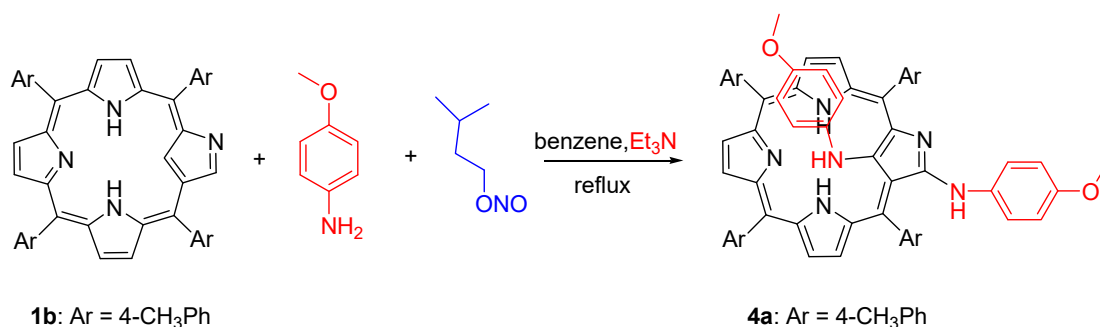


**General method of synthesis of 3.** To a solution of **1a** or **1b** (0.1 mmol) and 3,4,5-trimethoxyaniline (0.33 mmol) in 5 ml of dry benzene was added triethylamine (100  $\mu\text{L}$ , 0.7 mmol) and amyl nitrite (100  $\mu\text{L}$ , 0.7 mmol). The mixture was allowed to react in a sealed tube at 70°C for 2 h. The products were separated by column chromatography (silica gel, DCM/ethyl acetate) and precipitated from DCM by addition of hexane, yielding 52 mg of product **3a** (51%) or 45 mg of **3b** (44%).

Selected data for **3a**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ , 298 K):  $\delta = -1.35$  (br, 1H, -NH), 0.44 (br, 1H, -NH), 0.61 (s, 1H, ArH), 2.01 (s, 3H, - $\text{OCH}_3$ ), 3.05 (s, 3H, - $\text{OCH}_3$ ), 3.68 (s, 6H, - $\text{OCH}_3$ ), 3.77 (s, 3H, - $\text{OCH}_3$ ), 3.91 (s, 3H, - $\text{OCH}_3$ ), 3.95 (s, 3H, - $\text{OCH}_3$ ), 4.02 (s, 6H, - $\text{OCH}_3$ ), 4.04 (s, 10H, - $\text{OCH}_3$ +ArH), 4.10 (br, 3H, - $\text{OCH}_3$ ),

4.14 (s, 3H, -OCH<sub>3</sub>), 4.15 (s, 3H, -OCH<sub>3</sub>), 4.16 (s, 3H, -OCH<sub>3</sub>), 4.21 (s, 3H, -OCH<sub>3</sub>), 6.83 (s, 1H), 6.87 (s, 2H), 7.14 (br, 1H), 7.28 (br, 1H), 7.33 (br, 1H), 7.46 (br, 1H), 7.54 (br, 1H), 7.74 (br, 1H), 8.16 (br, 2H), 8.16 (d, *J* = 3.0 Hz, 1H, pyrH), 8.46 (br, 1H), 8.49 (br, 2H), 8.66 (d, *J* = 3.5 Hz, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, 298 K): δ = 53.6, 55.5, 55.8, 56.3, 56.41, 56.44, 56.46, 56.8, 61.0, 61.2, 61.26, 61.31, 61.5, 95.5, 96.4, 108.1, 111.2, 111.6, 112.0, 112.5, 120.5, 120.6, 122.7, 123.2, 124.5, 125.7, 126.7, 127.1, 127.9, 129.8, 133.0, 134.3, 134.5, 135.5, 136.0, 136.7, 136.9, 137.1, 137.3, 137.7, 137.8, 137.9, 138.2, 139.1, 139.5, 145.9, 146.5, 149.8, 151.59, 151.63, 152.1, 152.8, 153.3, 153.4, 155.1, 156.6, 174.4. UV-vis (CHCl<sub>3</sub>): λ<sub>max</sub>/nm (log ε) = 406(4.83), 435(4.91), 605(4.10). ESI-HRMS calc. for [C<sub>73</sub>H<sub>73</sub>N<sub>6</sub>O<sub>18</sub>]<sup>+</sup> (M+H): 1321.4976, Found: 1321.4985.

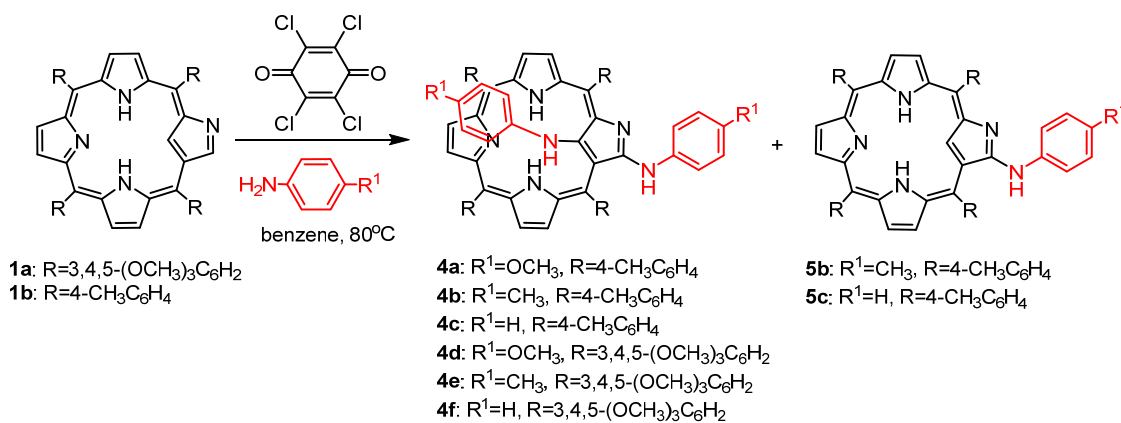
Selected data for **3b**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K): δ = -1.16 (br, 1H, -NH), 0.26 (br, 1H, -NH), 0.60 (s, 1H, ArH), 1.96 (s, 3H, -OCH<sub>3</sub>), 2.59 (s, 3H, -CH<sub>3</sub>), 2.64 (s, 3H, -CH<sub>3</sub>), 2.66 (s, 3H, -CH<sub>3</sub>), 2.74 (s, 3H, -CH<sub>3</sub>), 3.45 (s, 3H, -OCH<sub>3</sub>), 3.56 (s, 6H, -OCH<sub>3</sub>), 3.72 (s, 3H, -OCH<sub>3</sub>), 4.22 (s, 1H, ArH), 6.66 (s, 2H, ArH), 7.10 (s, 1H, ArH), 7.25 (s, 3H), 7.43 (d, *J* = 7.5 Hz, 1H, ArH), 7.50 (d, *J* = 7.5 Hz, 5H, ArH), 7.59 (d, *J* = 7.5 Hz, 1H, ArH), 7.77 (d, *J* = 7.5 Hz, 2H, ArH), 7.90 (d, *J* = 8.0 Hz, 2H, ArH), 8.05 (d, *J* = 7.5 Hz, 1H, ArH), 8.08 (d, *J* = 5.0 Hz, 1H, pyrH), 8.32 (d, *J* = 4.5 Hz, 2H, pyrH), 8.34 (d, *J* = 5.0 Hz, 1H, pyrH), 8.38 (d, *J* = 4.5 Hz, 1H, pyrH), 8.54 (d, *J* = 5.0 Hz, 1H, pyrH); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 233 K): δ = -1.68 (s, 1H, -NH), 0.41 (s, 1H, -NH), 0.54 (s, 1H, ArH), 1.96 (s, 3H, -OCH<sub>3</sub>), 2.63 (s, 3H, -CH<sub>3</sub>), 2.67 (s, 3H, -CH<sub>3</sub>), 2.69 (s, 3H, -CH<sub>3</sub>), 2.78 (s, 3H, -CH<sub>3</sub>), 3.49 (s, 3H, -OCH<sub>3</sub>), 3.56 (br, 6H, -OCH<sub>3</sub>), 3.74 (s, 3H, -OCH<sub>3</sub>), 4.26 (s, 1H, ArH), 7.14 (s, 1H, ArH), 7.45 (d, *J* = 7.5 Hz, 1H), 7.49 (d, *J* = 7.5 Hz, 1H), 7.52-7.60 (m, 6H), 7.64 (d, *J* = 7.5 Hz, 1H), 7.82-7.85 (m, 2H), 7.91 (d, *J* = 7.5 Hz, 1H), 7.94 (d, *J* = 7.5 Hz, 1H, ArH), 7.99 (d, *J* = 7.0 Hz, 1H), 8.09-8.12 (m, 2H), 8.38-8.39 (m, 2H), 8.44 (d, *J* = 4.5 Hz, 1H), 8.47 (d, *J* = 4.5 Hz, 1H), 8.51 (d, *J* = 7.5 Hz, 1H), 8.69 (d, *J* = 4.5 Hz, 1H), 8.74 (d, *J* = 7.5 Hz, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, 298 K): δ = 21.3, 21.5, 53.7, 55.4, 55.7, 60.9, 95.6, 96.3, 107.8, 119.8, 120.0, 123.1, 123.2, 124.5, 125.2, 125.7, 126.8, 127.5, 127.6, 127.7, 127.8, 128.1, 128.5, 132.7, 132.3, 133.7, 134.0, 134.2, 134.4, 135.2, 136.0, 136.8, 137.1, 137.2, 137.3, 137.5, 137.7, 138.0, 138.6, 139.0, 139.1, 139.4, 146.1, 147.6, 150.2, 152.2, 153.1, 153.3, 155.2, 157.0, 174.7. UV-vis (CHCl<sub>3</sub>): λ<sub>max</sub>/nm (log ε) = 401(4.89), 431(4.93), 610(4.16). ESI-HRMS calc. for [C<sub>65</sub>H<sub>57</sub>N<sub>6</sub>O<sub>6</sub>]<sup>+</sup> (M+H): 1017.4334, Found: 1017.4338.



**Synthesis of 4a.** To a solution of NCP **1b** (67 mg, 0.1 mmol) and 4-methoxyaniline (37 mg, 0.3 mmol) in 5 ml of dry benzene was added triethylamine (100 μL, 72.6 mg, 0.7 mmol) and amyl nitrite (100 μL, 87 mg, 0.7 mmol). The mixture was stirred under nitrogen for 3 hours at boiling point of the solution under reflux and then the solvent was removed under vacuum. The residue was passed down a silica gel column with dichloromethane/ethyl acetate solvent as eluent. The red colored product fraction was collected, followed by removal of solvent to afford NCP derivative **4a**. Yield 30 mg (33%).

Selected data for **4a**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K): δ = 2.61 (s, 3H, -CH<sub>3</sub>), 2.62 (s, 3H, -CH<sub>3</sub>), 2.64

(s, 3H, -CH<sub>3</sub>), 2.66 (s, 3H, -CH<sub>3</sub>), 3.23 (s, 3H, -OCH<sub>3</sub>), 3.72 (s, 3H, -OCH<sub>3</sub>), 4.20 (d, *J* = 9.0 Hz, 2H, ArH), 5.60 (d, *J* = 8.5 Hz, 2H, ArH), 6.30 (d, *J* = 9.0 Hz, 2H, ArH), 7.15 (d, *J* = 8.0 Hz, 2H, ArH), 7.46 (d, *J* = 7.5 Hz, 3H, ArH), 7.50 (d, *J* = 8.0 Hz, 3H, ArH), 7.51-7.53 (m, 1H, ArH), 7.63 (d, *J* = 7.5 Hz, 2H, ArH), 7.74-7.78 (m, 1H, ArH+pyrrH), 7.80-7.85 (m, 3H, ArH+pyrrH), 7.96 (d, *J* = 4.5 Hz, 1H, pyrrH), 7.99 (d, *J* = 3.5 Hz, 1H, pyrrH), 8.08 (d, *J* = 5.0 Hz, 1H, pyrrH), 8.13 (d, *J* = 7.5 Hz, 4H, ArH), 8.21 (d, *J* = 4.5 Hz, 1H, pyrrH); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 230 K): δ = 0.64 (br, 2H, -NH), 2.61 (s, 3H, -CH<sub>3</sub>), 2.63 (s, 6H, -CH<sub>3</sub>), 2.64 (s, 3H, -CH<sub>3</sub>), 3.23 (s, 3H, -OCH<sub>3</sub>), 3.73 (s, 3H, -OCH<sub>3</sub>), 4.19 (br, 2H, ArH), 5.55 (br, 2H, ArH), 6.64 (d, *J* = 6.5 Hz, 2H, ArH), 6.74 (s, 1H, ArH), 7.15 (br, 2H, ArH), 7.26 (s, 1H, ArH), 7.46 (br, 3H, ArH), 7.54 (br, 3H, ArH), 7.61 (br, 1H, ArH), 7.67-7.73 (m, 4H, ArH+pyrrH), 7.82-7.98 (m, 5H, ArH+pyrrH), 8.08 (br, 1H, pyrrH), 8.19-8.21 (m, 2H, ArH), 8.36 (br, 1H, pyrrH); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, 298 K): δ = 22.9, 23.0, 56.4, 56.9, 113.3, 114.8, 115.4, 118.3, 119.6, 120.5, 121.8, 123.1, 124.3, 124.8, 128.2, 129.3, 131.6, 132.2, 133.0, 134.3, 135.4, 135.5, 136.4, 137.4, 138.4, 138.5, 138.7, 138.9, 139.3, 139.6, 140.0, 140.3, 140.6, 144.1. UV-vis (CHCl<sub>3</sub>): λ<sub>max</sub>/nm (log ε) = 439(4.88), 501(4.69), 770(3.90). ESI(-)-HRMS calc. for [C<sub>62</sub>H<sub>51</sub>N<sub>6</sub>O<sub>2</sub>]<sup>-</sup> (M-H): 911.4079, Found: 911.4075.



**General method of synthesis of 4:** To a solution of **1** (0.05 mmol) and arylamine (0.25 mmol) in benzene (5 ml) *p*-chloranil (61 mg, 0.25 mmol) was added. The mixture was allowed to react in a sealed tube at 80°C for 1 h, and then the solvent was removed under vacuum. The residue was passed down a silica gel column with 1% methanol in dichloromethane and first weak red band containing **5** was separated from the fraction consisting of major product **4** that was eluted with a dichloromethane/methanol (*v/v* = 100:3) mixture. The solvents were then removed and products were precipitated from DCM solution by addition of hexane and dried *in vacuo*. Yields: 27.5 mg (60%) for **4a**, 29.5 mg (67%) for **4b**, 27.3 mg (64%) for **4c**, 34.7 mg (57%) for **4d**, 30.8 mg (52%) for **4e**, 29.0 mg 50% for **4f**. Product **5** was formed in a minute amount and was characterized only for the reaction products of **1b** with toluidine (**5b**, 3.8 mg, 8%) and aniline (**5c**, 3.8 mg, 10%)

Selected data for **4b**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K): δ = 1.58 (s, 3H), 2.19 (s, 3H), 2.60 (s, 6H), 2.62 (s, 3H), 2.64 (s, 3H), 4.09 (d, *J* = 7.5 Hz, 2H), 5.80 (d, *J* = 7.0 Hz, 2H), 6.86 (d, *J* = 7.5 Hz, 2H), 7.10-7.12 (m, 2H), 7.44-7.49 (m, 6H), 7.63-7.64 (m, 2H), 7.78-7.83 (m, 5H), 8.02-8.03 (m, 2H), 8.10-8.14 (m, 6H), 8.21-8.23 (m, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, 298 K): δ = 20.07, 20.70, 21.47, 21.50, 21.55, 113.63, 117.90, 118.07, 118.25, 122.76, 123.56, 123.69, 126.83, 127.15, 127.80, 129.19, 129.25, 130.16, 130.65, 130.83, 132.77, 133.99, 134.12, 135.02, 135.83, 136.05, 136.94, 137.12, 137.23,



137.50, 137.57, 137.97, 138.42, 138.63, 139.16, 142.47, 149.00. UV-vis (CHCl<sub>3</sub>):  $\lambda_{max}/nm$  (log  $\epsilon$ ) = 440 (4.65), 497 (4.43), 780(3.55). ESI-HRMS calc. for [C<sub>62</sub>H<sub>53</sub>N<sub>6</sub>]<sup>+</sup> (M+H): 881.4332, Found: 881.4324.

Selected data for **4c**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K):  $\delta$  = 2.60 (s, 3H), 2.62 (s, 3H), 2.64 (s, 3H), 2.66 (s, 3H), 4.09 (d, *J* = 8.0 Hz, 2H), 5.85-5.88 (m, 1H), 5.95-5.98 (m, 2H), 6.78-6.81 (m, 1H), 7.05-7.08 (m, 2H), 7.21-7.23 (m, 2H), 7.43-7.46 (m, 3H), 7.49-7.52 (m, 3H), 7.65 (d, *J* = 8.0 Hz, 2H), 7.77-7.80 (m, 3H), 7.85-7.86 (m, 2H), 8.02 (d, *J* = 5.0 Hz, 1H), 8.07 (d, *J* = 5.0 Hz, 1H), 8.13-8.14 (m, 2H), 8.16-8.18 (m, 2H), 8.19-8.22 (m, 2H), 8.26 (d, *J* = 5.0 Hz, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, 298 K):  $\delta$  = 21.49, 21.51, 21.56, 113.94, 117.49, 118.37, 118.51, 118.65, 119.48, 121.32, 122.73, 123.60, 123.80, 124.48, 125.22, 126.55, 126.93, 127.84, 127.89, 128.48, 128.71, 130.22, 130.99, 132.91, 133.95, 134.18, 135.09, 136.21, 136.97, 137.03, 137.43, 137.59, 138.10, 138.16, 138.58, 139.01, 139.04, 140.49, 142.61, 144.46, 148.71, 149.88, 154.92; UV-vis (CHCl<sub>3</sub>):  $\lambda_{max}/nm$  (log  $\epsilon$ ) = 439 (4.89), 508 (4.59), 769 (3.61). ESI-HRMS calc. for [C<sub>60</sub>H<sub>49</sub>N<sub>6</sub>]<sup>+</sup> (M+H): 853.4019, Found: 853.4013.

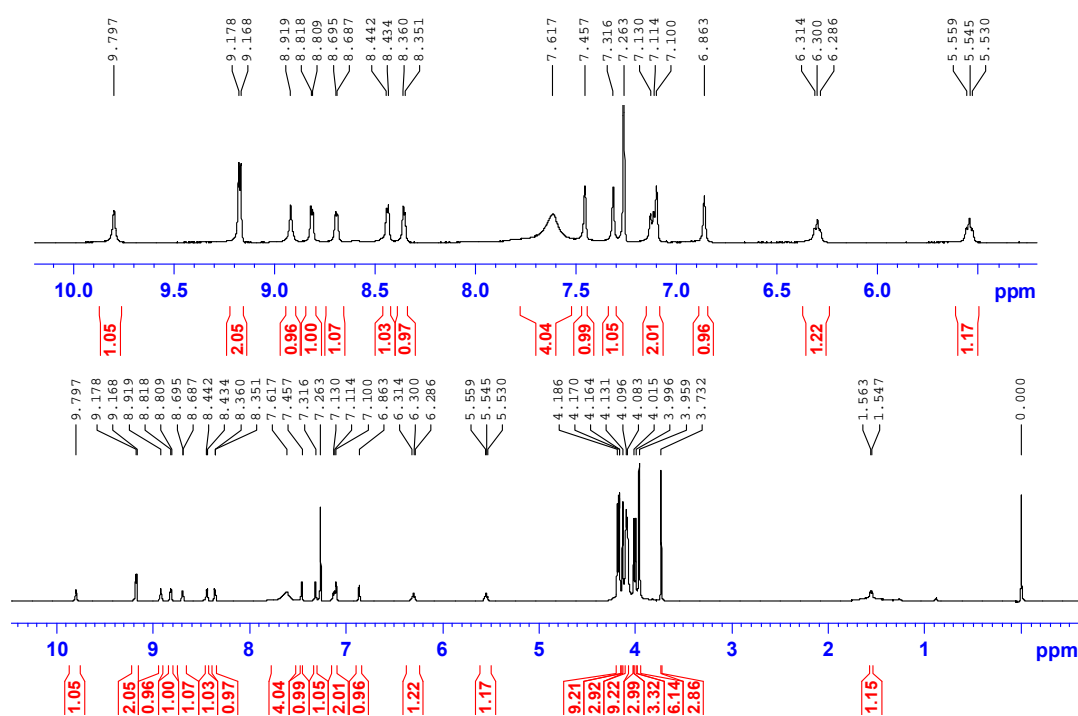
Selected data for **4d**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K):  $\delta$  = 3.26(s, 3H), 3.72 (s, 3H), 3.92 (s, 6H), 3.98 (s, 3H), 4.03 (s, 9H), 4.05 (s, 6H), 4.08 (s, 3H), 4.10 (s, 3H), 4.12 (s, 6H), 4.25 (d, *J* = 8.5 Hz, 2H), 5.65(d, *J* = 8.5Hz, 2H), 6.68 (d, *J* = 8.5 Hz, 2H), 7.09 (s, 1H), 7.18 (s, 1H), 7.19 (s, 1H), 7.34 (d, *J* = 8.5 Hz, 2H), 7.37 (s, 2H), 7.46 (s, 2H), 7.48 (s, 1H), 7.89 (d, *J* = 5.0 Hz, 1H), 7.95 (d, *J* = 5.0 Hz, 1H), 8.06 (d, *J* = 5.0 Hz, 1H), 8.12 (d, *J* = 5.0 Hz, 1H), 8.17 (d, *J* = 5.0 Hz, 1H) 8.30 (d, *J* = 5.0 Hz, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, 298 K):  $\delta$  = 55.00, 55.55, 56.32, 56.41, 56.47, 56.52, 56.61, 56.78, 61.17, 61.24, 61.35, 111.76, 111.86, 111.95, 112.01, 113.56, 113.78, 114.12, 118.10, 119.03, 120.68, 122.79, 123.80, 124.11, 126.79, 127.99, 129.90, 130.87, 131.83, 132.88, 135.41, 136.00, 136.77, 137.40, 138.10, 139.15, 142.27, 149.39, 151.80, 151.93, 153.67, 153.90, 154.68; UV-vis (CHCl<sub>3</sub>):  $\lambda_{max}/nm$  (log  $\epsilon$ ) = 440 (4.84), 509 (4.64), 780 (3.69). ESI-HRMS calc. for [C<sub>70</sub>H<sub>69</sub>N<sub>6</sub>O<sub>14</sub>]<sup>+</sup> (M+H): 1217.4872, Found: 1217.4857.

Selected data for **4e**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K):  $\delta$  = 1.64 (s, 3H), 2.23 (s, 3H), 3.92 (s, 3H), 3.93 (s, 3H), 3.98 (s, 3H), 4.03 (s, 9H), 4.05(s, 6H), 4.10 (s, 3H), 4.11 (s, 3H), 4.12 (s, 3H), 4.13 (s, 3H), 4.14-4.16 (m,2H), 5.86 (d, *J* = 8.5 Hz, 2H), 6.94 (d, *J* = 8.0Hz, 2H), 7.11 (s, 1H), 7.18 (s, 1H), 7.19 (s, 1H), 7.32 (d, *J* = 8.5 Hz, 2H), 7.40 (s, 2H), 7.46 (s, 2H), 7.48 (s, 1H), 7.93 (d, *J* = 5.0 Hz, 1H), 8.00 (d, *J* = 5.0 Hz, 1H), 8.12 (d, *J* = 5.0 Hz, 1H), 8.17 (d, *J* = 5.0 Hz, 1H), 8.22 (d, *J* = 5.0 Hz, 1H) 8.34 (d, *J* = 5.0 Hz, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, 298 K):  $\delta$  = 20.16, 20.72, 56.34, 56.47, 56.54, 56.61, 56.78, 61.19, 61.25, 61.36, 111.91, 112.06, 113.86, 117.92, 118.34, 118.71, 121.46, 122.56, 124.19, 124.48, 126.98, 127.26, 128.12, 129.37, 129.89, 130.61, 131.35, 132.54, 135.43, 136.00, 136.02, 136.77, 137.45, 137.68, 138.02, 138.11, 138.28, 138.68, 139.17, 139.86, 142.52, 144.78, 149.37, 151.88, 151.93, 153.92; UV-vis (CHCl<sub>3</sub>):  $\lambda_{max}/nm$  (log  $\epsilon$ ) = 440 (4.71), 506 (4.48), 774 (3.54). ESI-HRMS calc. for [C<sub>70</sub>H<sub>69</sub>N<sub>6</sub>O<sub>12</sub>]<sup>+</sup> (M+H): 1185.4973, Found: 1185.4956.

Selected data for **4f**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 298 K):  $\delta$  = 3.93 (s, 3H), 3.94 (s, 3H), 3.98 (s, 3H), 4.04 (s, 9H), 4.07 (s, 6H), 4.09(s, 3H), 4.11 (s, 3H), 4.13 (s, 3H), 4.14 (s, 3H), 4.15-4.17 (m, 2H), 5.96-5.98(m, 1H), 6.03-6.06 (m, 2H), 6.86-6.89 (m, 1H), 7.12-7.16 (m, 3H), 7.20 (s, 1H), 7.43-7.45 (m, 4H), 7.50 (s, 3H), 7.97 (d, *J* = 5.0 Hz, 1H), 8.02 (d, *J* = 5.0 Hz, 1H), 8.14 (d, *J* = 5.0 Hz, 1H), 8.20 (d, *J* = 5.0 Hz, 1H), 8.27 (d, *J* = 5.0 Hz, 1H) 8.38 (d, *J* = 5.0 Hz, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, 298 K):  $\delta$  =56.33, 56.43, 56.58, 56.62, 56.80, 61.19, 61.26, 61.27, 61.37, 111.80, 111.97, 112.06, 112.12, 114.02, 117.77, 117.95, 118.82, 118.90, 119.96, 120.08, 121.86, 122.46, 124.22, 124.64, 126.68, 127.09, 128.18, 128.90, 130.81, 132.69, 135.34, 135.88, 136.74, 137.25, 137.69, 138.03, 138.23, 138.38, 138.76, 139.23, 142.61, 149.09, 151.84, 151.89, 151.93, 151.97, 153.93. UV-vis (CHCl<sub>3</sub>):  $\lambda_{max}/nm$  (log  $\epsilon$ ) = 442(4.95), 506(4.61), 778(3.68). ESI-HRMS calc. for [C<sub>68</sub>H<sub>65</sub>N<sub>6</sub>O<sub>12</sub>]<sup>+</sup> (M+H): 1157.4655, Found: 1157.4644.

Selected data for **5b**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ , 298 K):  $\delta$  = -4.91(s, 1H, -21-CH), -1.93(br, 2H, 22, 24-NH), 2.29 (s, 3H, - $\text{CH}_3$ ), 2.66 (s, 3H, - $\text{CH}_3$ ), 2.67 (s, 3H, - $\text{CH}_3$ ), 2.68 (s, 3H, - $\text{CH}_3$ ), 3.73 (s, 3H, - $\text{CH}_3$ ), 7.02 (d,  $J$  = 8.0 Hz, 2H, ArH), 7.43-7.45 (m, 2H, ArH), 7.51 (d,  $J$  = 8.0 Hz, 2H, ArH), 7.52 (d,  $J$  = 8.0 Hz, 2H, ArH), 7.60 (d,  $J$  = 8.0 Hz, 2H, ArH), 7.70 (d,  $J$  = 7.5 Hz, 2H, ArH), 8.00 (d,  $J$  = 7.5 Hz, 2H, ArH), 8.05 (d,  $J$  = 7.5 Hz, 2H, ArH), 8.16 (d,  $J$  = 7.5 Hz, 2H, ArH), 8.32 (d,  $J$  = 8.0 Hz, 2H, ArH), 8.41 (d,  $J$  = 4.5 Hz, 1H, pyrRH), 8.45 (d,  $J$  = 4.5 Hz, 1H, pyrRH), 8.49 (d,  $J$  = 4.5 Hz, 1H, pyrRH), 8.51 (d,  $J$  = 4.5 Hz, 1H, pyrRH), 8.58 (d,  $J$  = 4.5 Hz, 1H, pyrRH), 8.69 (d,  $J$  = 4.5 Hz, 1H, pyrRH);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ , 298 K):  $\delta$  = 20.8, 21.53, 21.54, 21.57, 21.60, 101.9, 116.8, 117.0, 118.2, 121.2, 124.7, 125.4, 126.6, 126.9, 127.6, 127.7, 127.9, 128.9, 129.4, 130.1, 131.0, 131.2, 132.8, 134.3, 134.4, 134.7, 136.7, 136.9, 137.1, 137.2, 137.5, 138.1, 138.5, 139.09, 139.15, 139.21, 139.4, 154.0, 156.7. UV-vis ( $\text{CHCl}_3$ ):  $\lambda_{\text{max}}$ /nm ( $\log \epsilon$ ) = 413(4.79), 484(4.93), 652(3.70), 720(3.60). ESI-HRMS calc. for  $[\text{C}_{55}\text{H}_{46}\text{N}_5]^+$  (M+H):776.3748, Found: 776.3748.

Selected data for **5c**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ , 300 K):  $\delta$  = -4.93 (s, 1H, -21-CH), -2.17 (br, 2H, 22, 24-NH), 2.65 (s, 3H, - $\text{CH}_3$ ), 2.66 (s, 3H, - $\text{CH}_3$ ), 2.67 (s, 3H, - $\text{CH}_3$ ), 2.73 (s, 3H, - $\text{CH}_3$ ), 6.91 (tt,  $^3J$  = 7.2 Hz,  $^4J$  = 1.0 Hz 1H, *p*-ArH), 7.21 (m, 2H, ArH), 7.25 (b, 1H, NH), 7.50 (d,  $J$  = 8.4 Hz, 2H, ArH), 7.52 (d,  $J$  = 8.0 Hz, 2H, ArH), 7.55 (d,  $J$  = 7.7 Hz, 2H, ArH), 7.59 (d,  $J$  = 7.7 Hz, 2H, ArH), 7.72 (d,  $J$  = 7.8 Hz, 2H, ArH), 7.99 (d,  $J$  = 7.8 Hz, 2H, ArH), 8.04 (d,  $J$  = 7.9 Hz, 2H, ArH), 8.18 (d,  $J$  = 7.8 Hz, 2H, ArH), 8.32 (d,  $J$  = 7.7 Hz, 2H, ArH), 8.42 (d,  $J$  = 4.7 Hz, 1H, pyrRH), 8.45 (d,  $J$  = 4.4 Hz, 1H, pyrRH), 8.49 (d,  $J$  = 4.9 Hz, 1H, pyrRH), 8.52 (d,  $J$  = 4.5 Hz, 1H, pyrRH), 8.58 (d,  $J$  = 5.0 Hz, 1H, pyrRH), 8.72 (d,  $J$  = 4.8 Hz, 1H, pyrRH);  $^{13}\text{C}$  NMR (125 MHz,  $\text{DCl}_3$ , 300 K):  $\delta$  = 21.5, 21.6, 101.8, 118.2, 121.1, 121.7, 124.7, 125.5, 126.5, 126.9, 127.6, 127.7, 127.9, 128.9, 130.1, 132.9, 134.4, 134.6, 135.2, 136.7, 136.9, 137.2, 137.5, 138.1, 138.4, 139.2, 139.3, 139.4, 154.1, 156.8, 158.1. UV-vis ( $\text{CHCl}_3$ ):@  $\lambda_{\text{max}}$ /nm ( $\log \epsilon$ ) = 276 (4.62) 344sh 414 (4.93), 477 (5.07), 553 sh, 652(3.70), 605 sh, 652 (3.96), 718 (3.90). ESI-HRMS calc. for  $[\text{C}_{54}\text{H}_{44}\text{N}_5]^+$  (M+H $^+$ ): 762.3591, Found: 762.3570.



**Figure S1.**  $^1\text{H}$  NMR spectrum of **2a**, 298 K,  $\text{CDCl}_3$ .

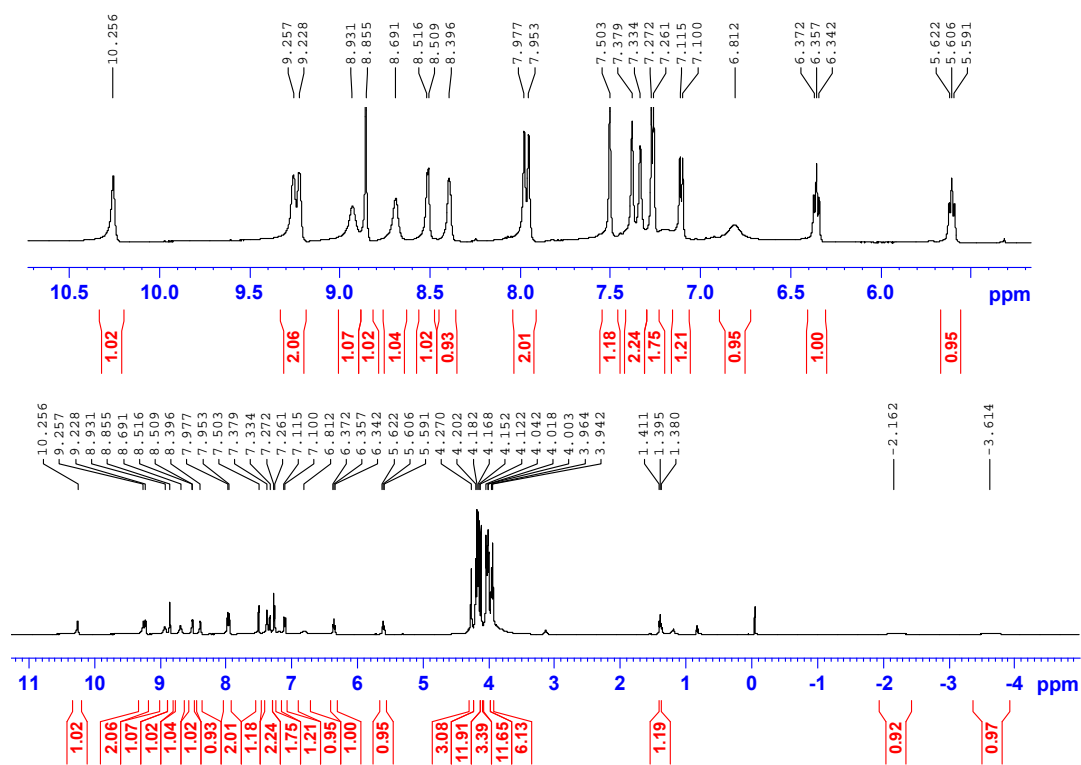


Figure S2. <sup>1</sup>H NMR spectrum of 2a, 248 K, CDCl<sub>3</sub>.

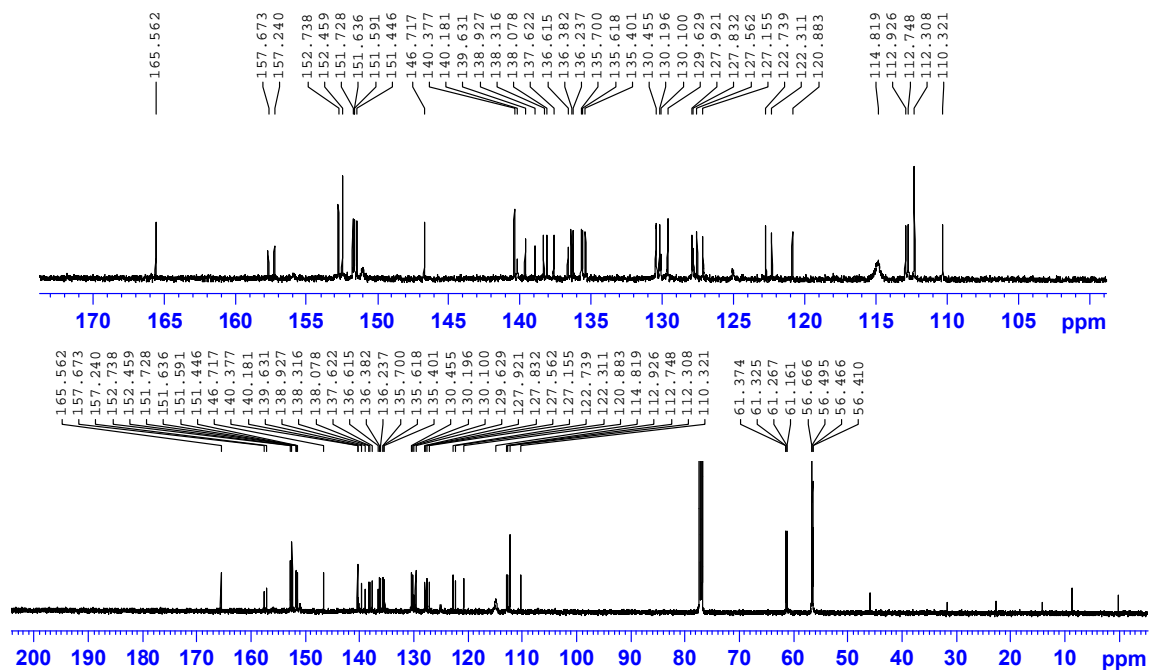
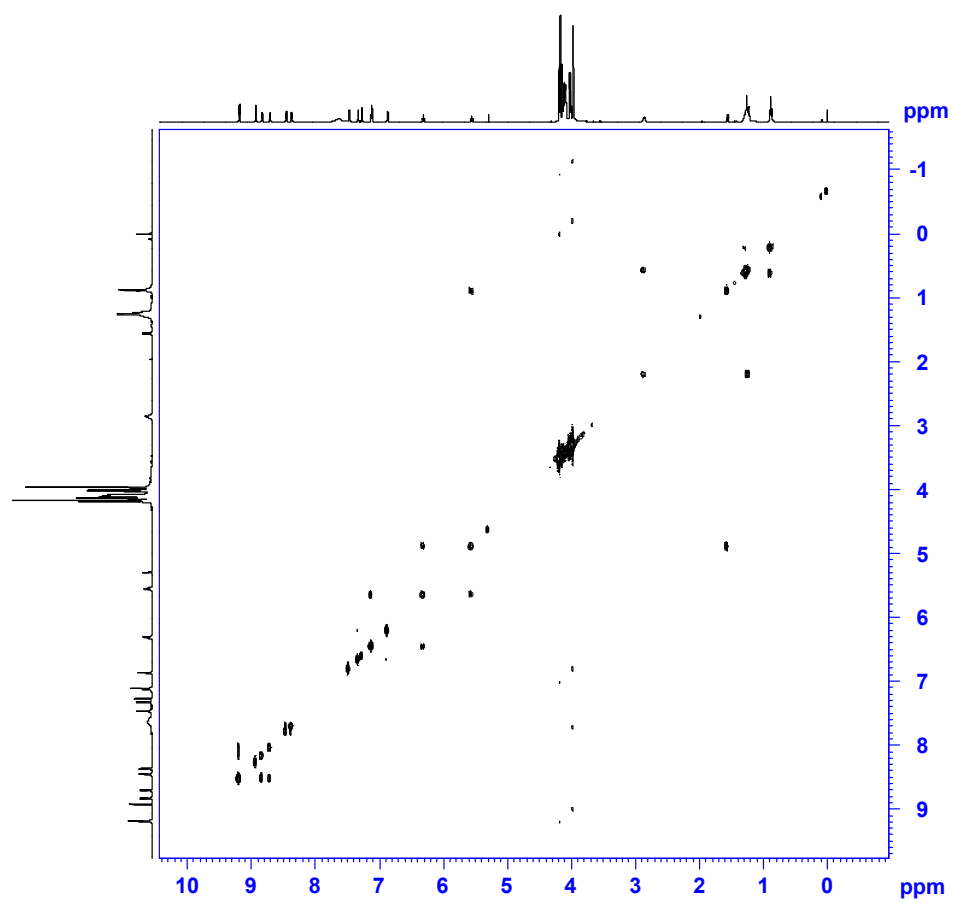


Figure S3. <sup>13</sup>C NMR spectrum of 2a, 298 K, CDCl<sub>3</sub>.



**Figure S4.** COSY spectrum of **2a** (298K).

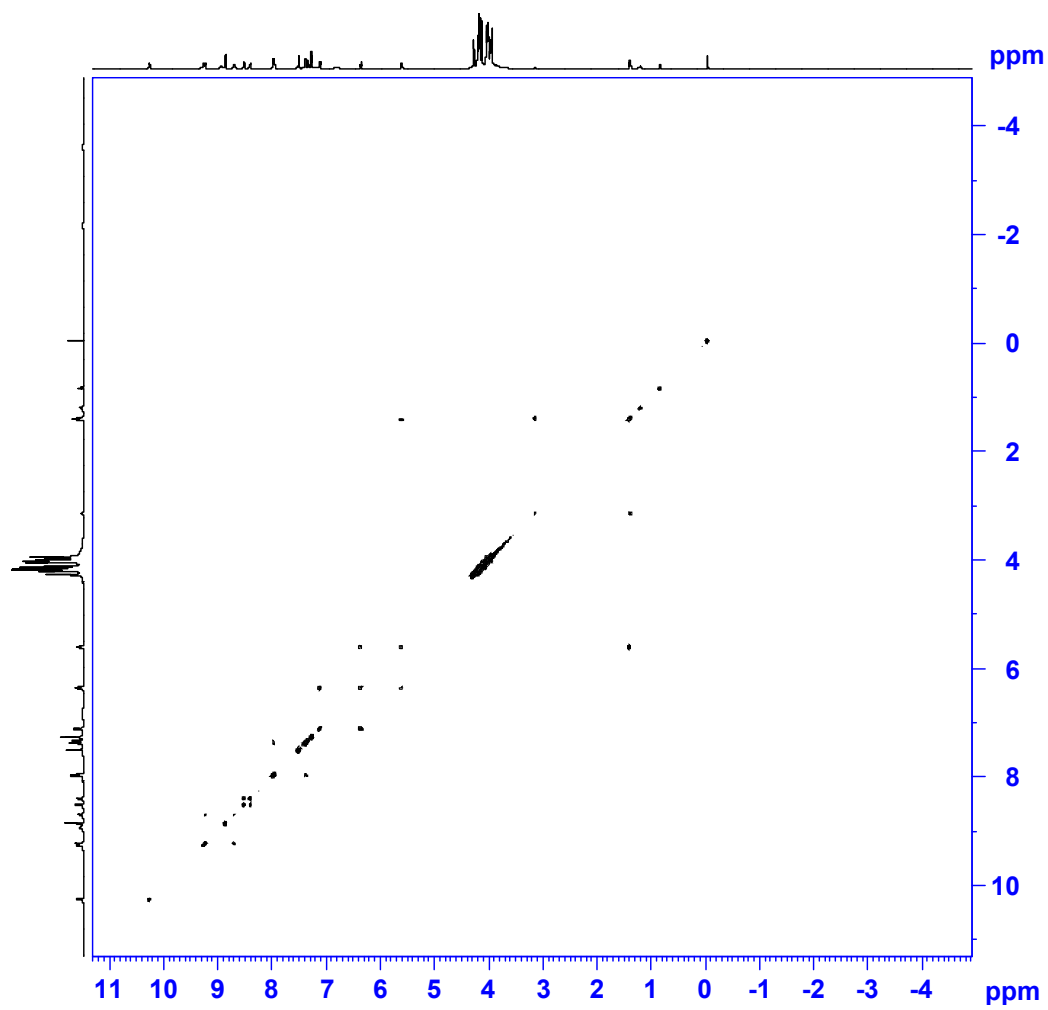


Figure S5. COSY spectrum of 2a (248K).

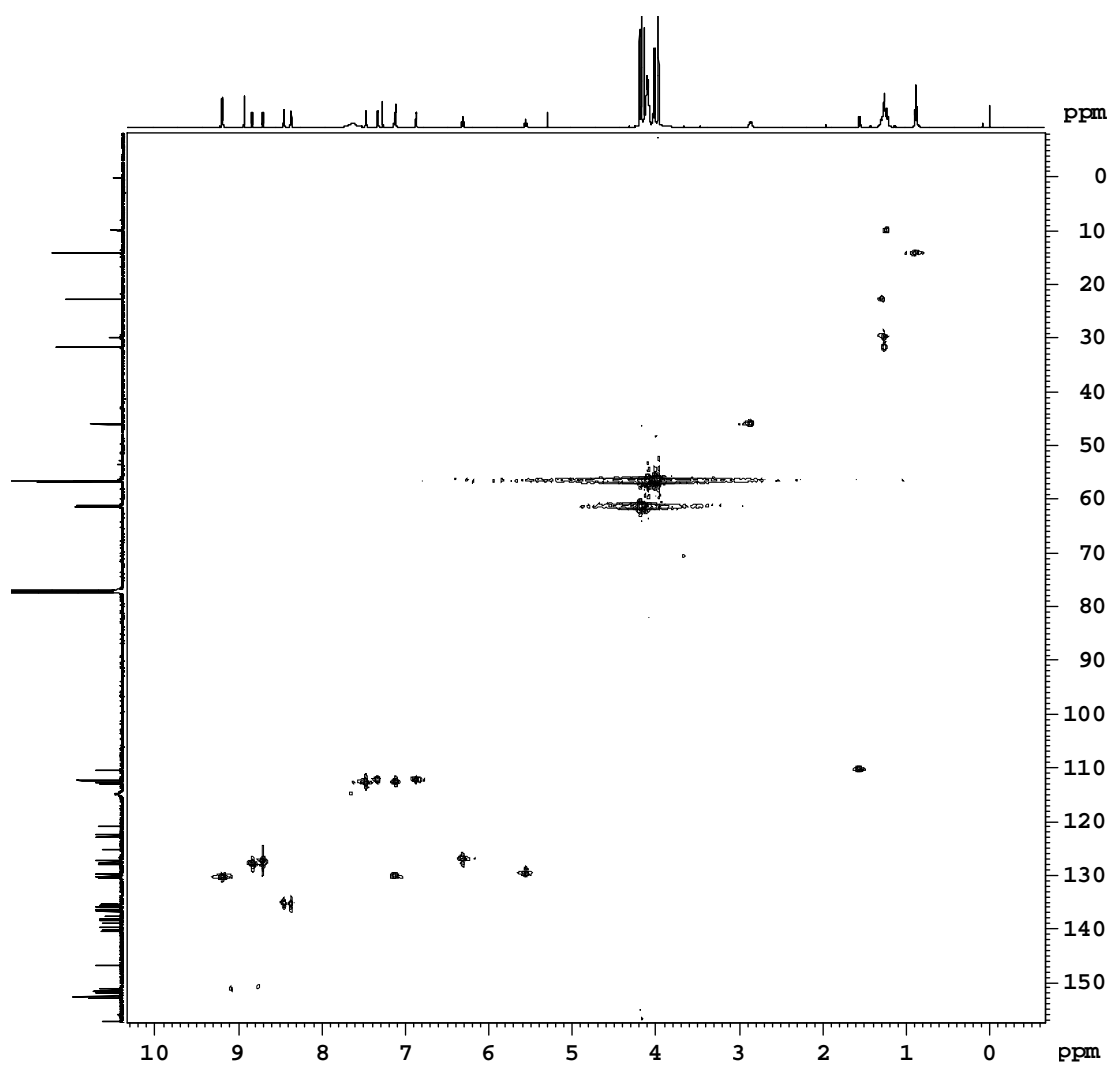


Figure S6. HSQC spectrum of **2a** ( $\text{CDCl}_3$ , 298 K).

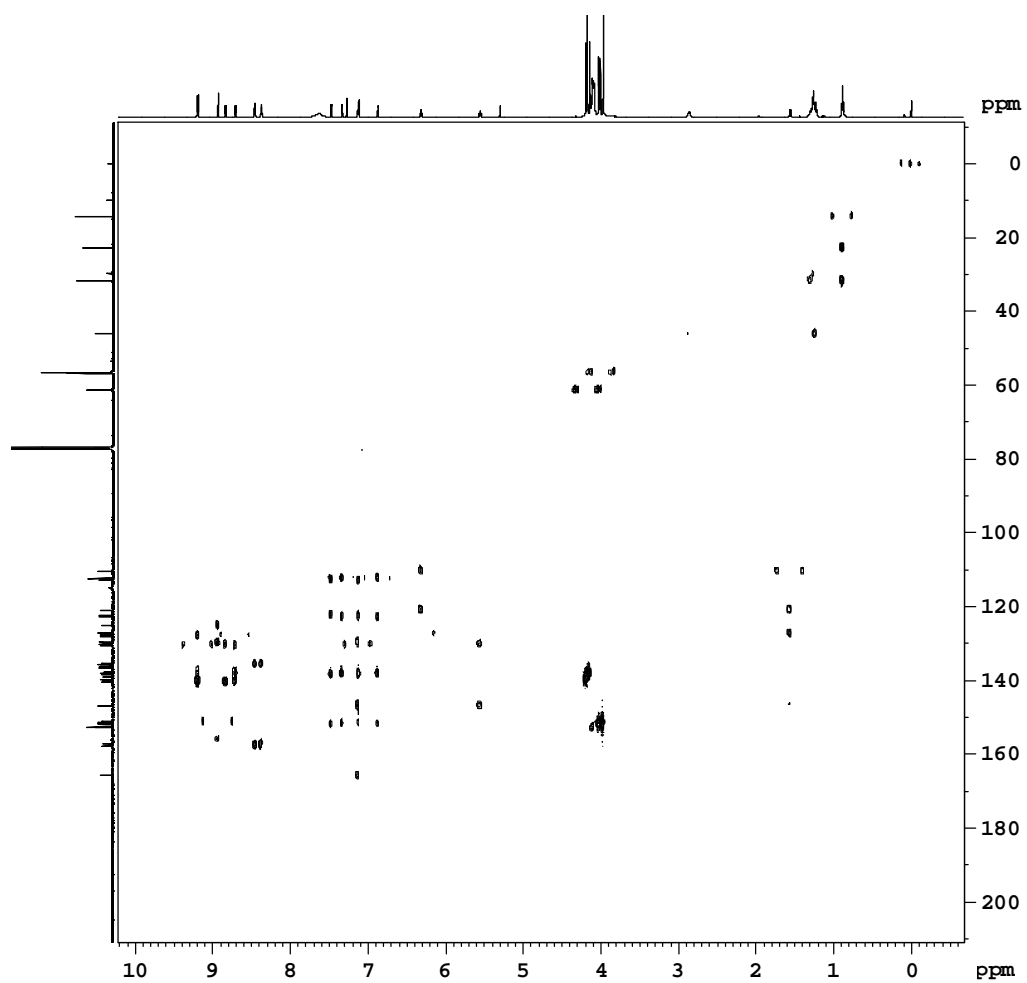


Figure S7. HMBC spectrum of **2a** ( $\text{CDCl}_3$ , 298 K).

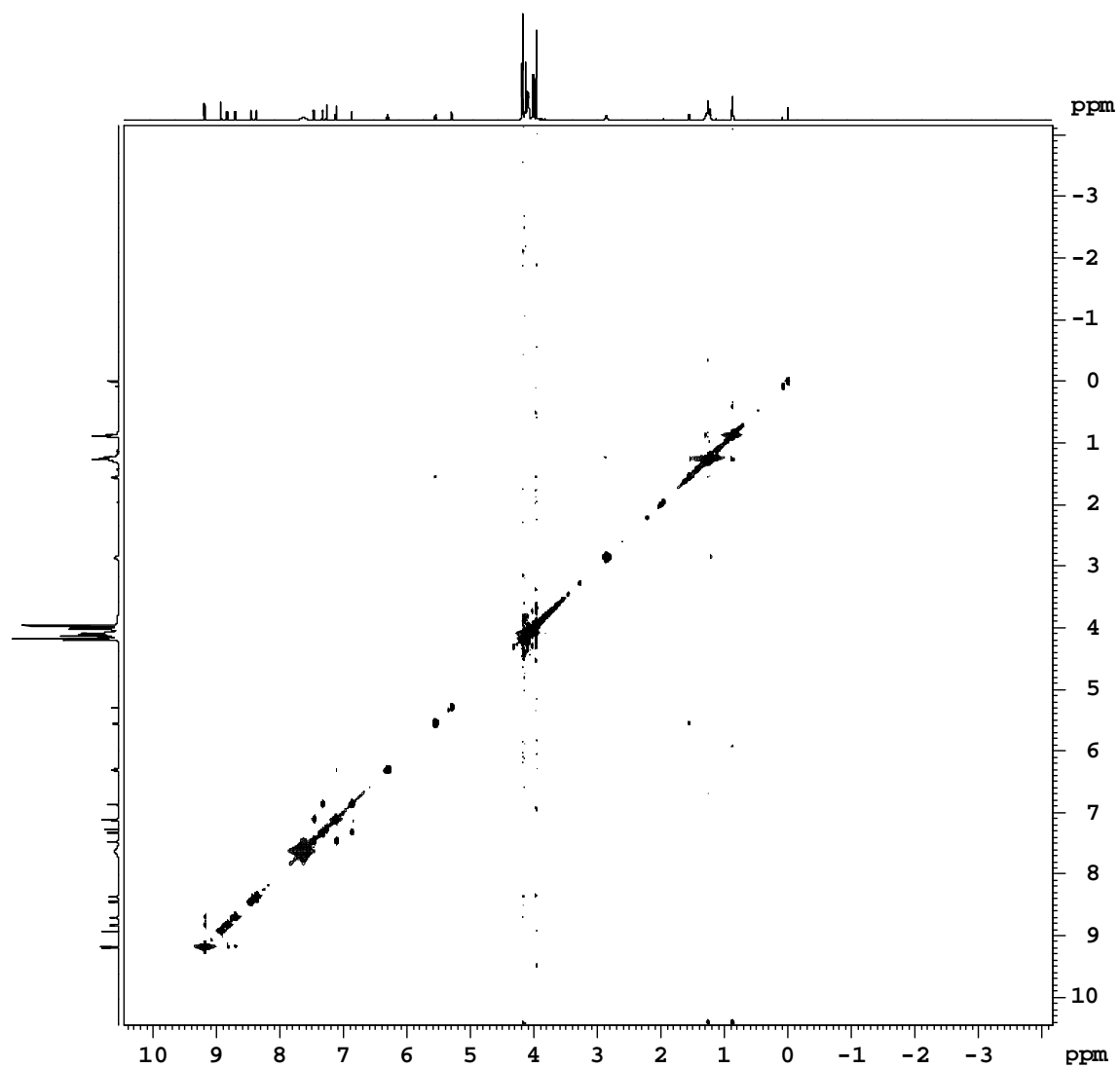


Figure S8. NOESY spectrum of **2a** ( $\text{CDCl}_3$ , 298 K).



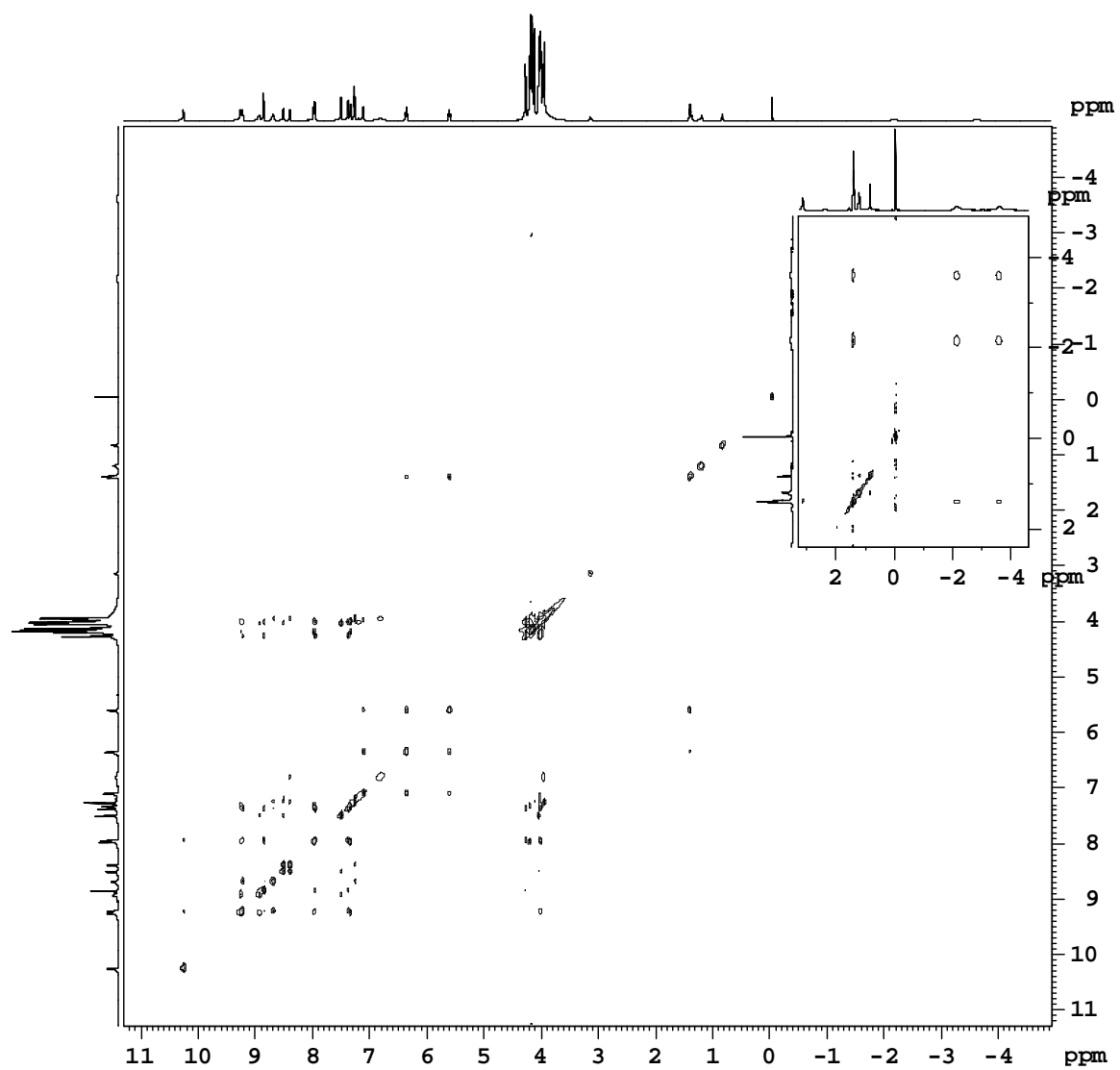


Figure S9. NOESY spectrum of 2a (CDCl<sub>3</sub>, 248 K).

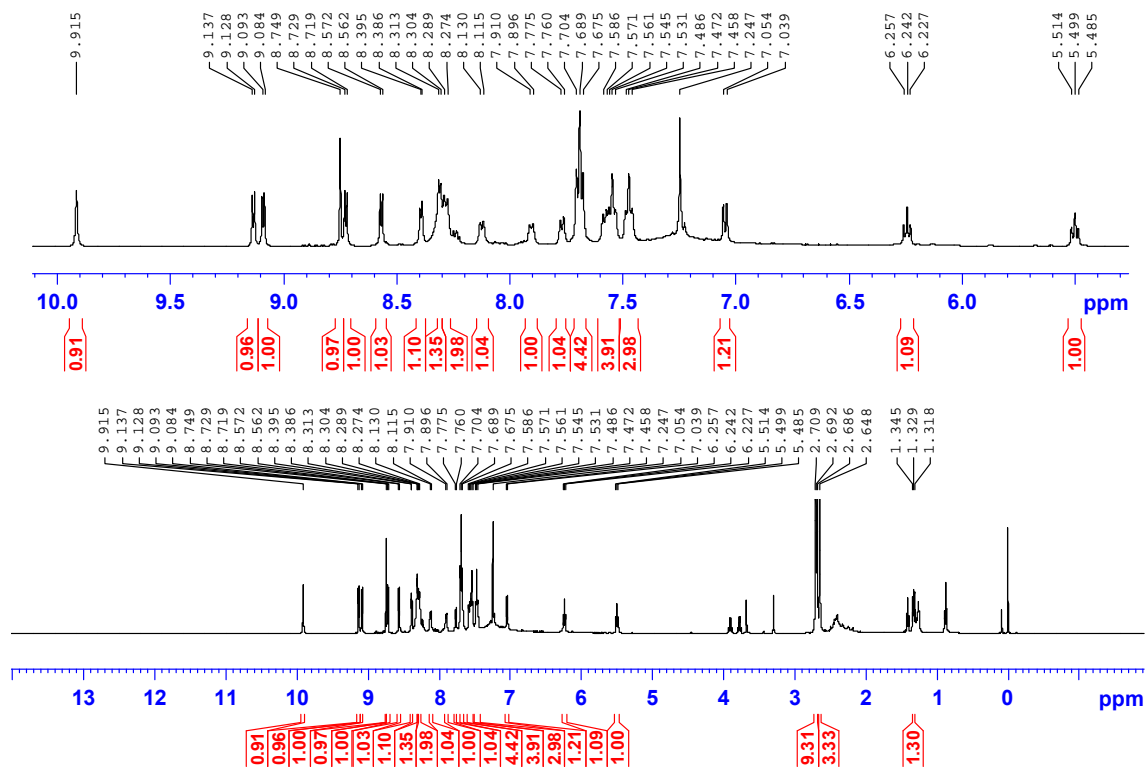


Figure S10. <sup>1</sup>H NMR spectrum of **2b**, 298 K, CDCl<sub>3</sub>.

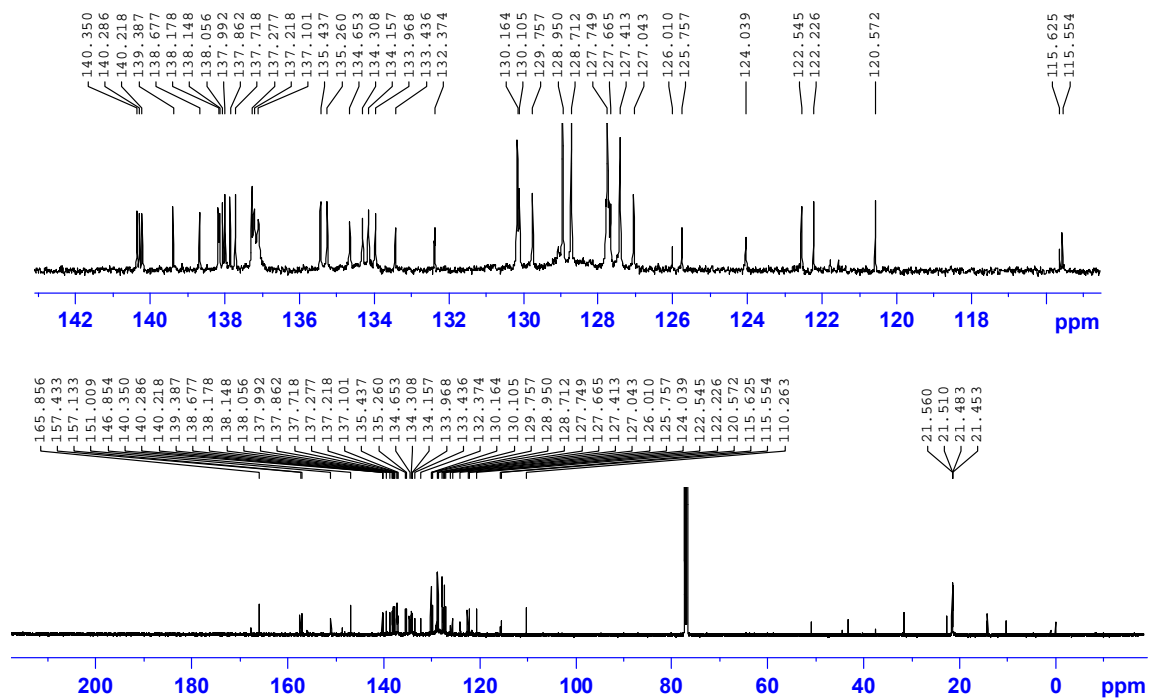
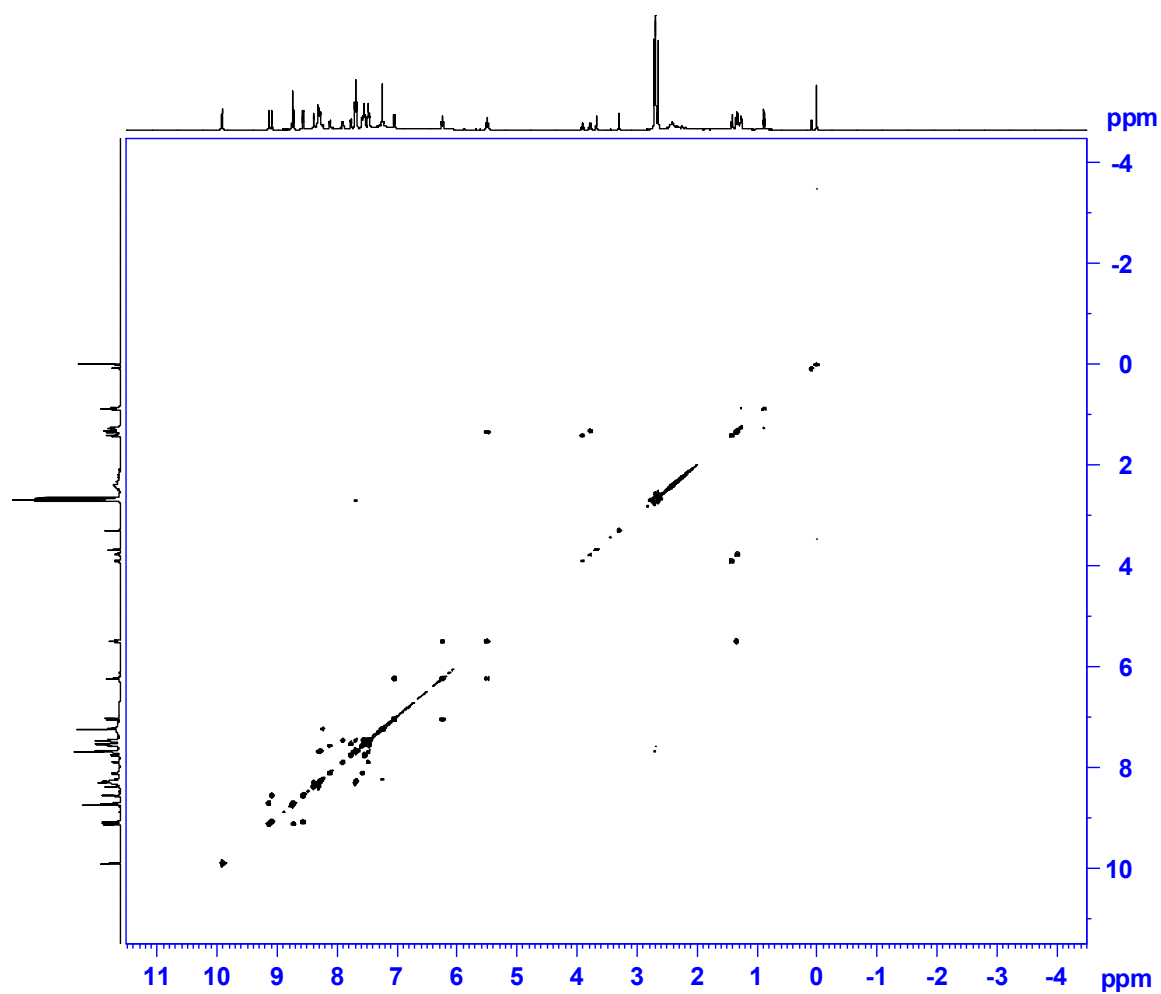


Figure S11. <sup>13</sup>C NMR spectrum of **2b**, 298 K, CDCl<sub>3</sub>.



**Figure S12.** COSY spectrum of **2b** (298K).

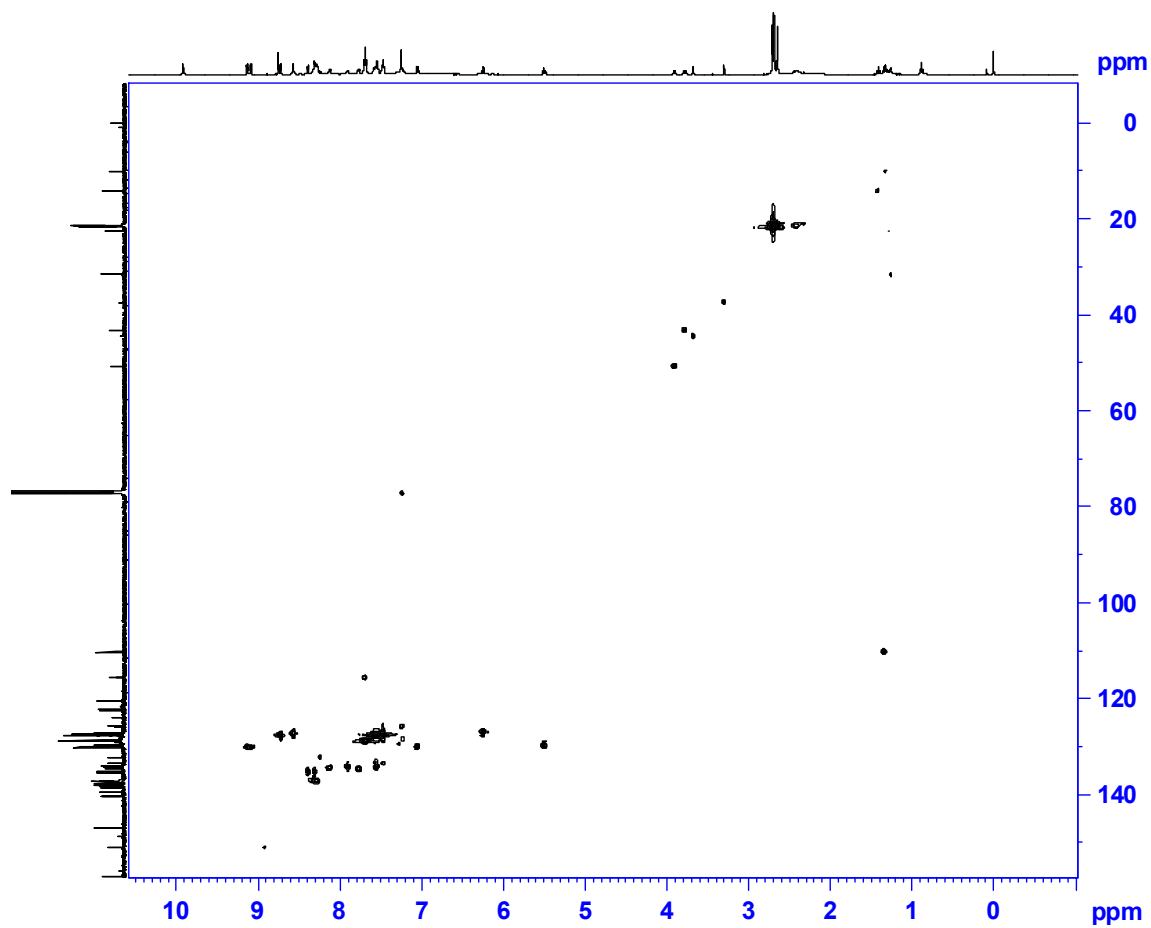


Figure S13. HSQC spectrum of **2b** ( $\text{CDCl}_3$ , 298 K).

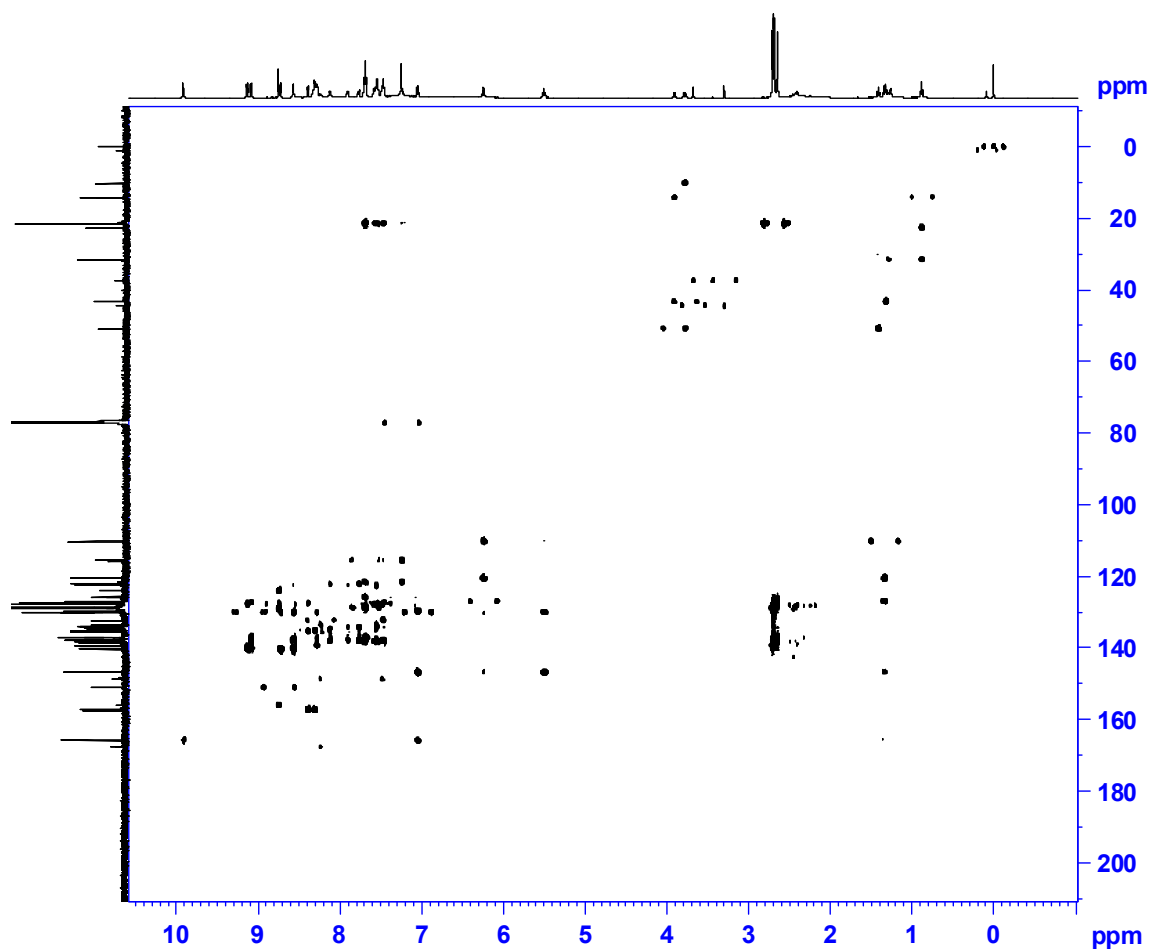


Figure S14. HMBC spectrum of **2b**(CDCl<sub>3</sub>, 298 K).

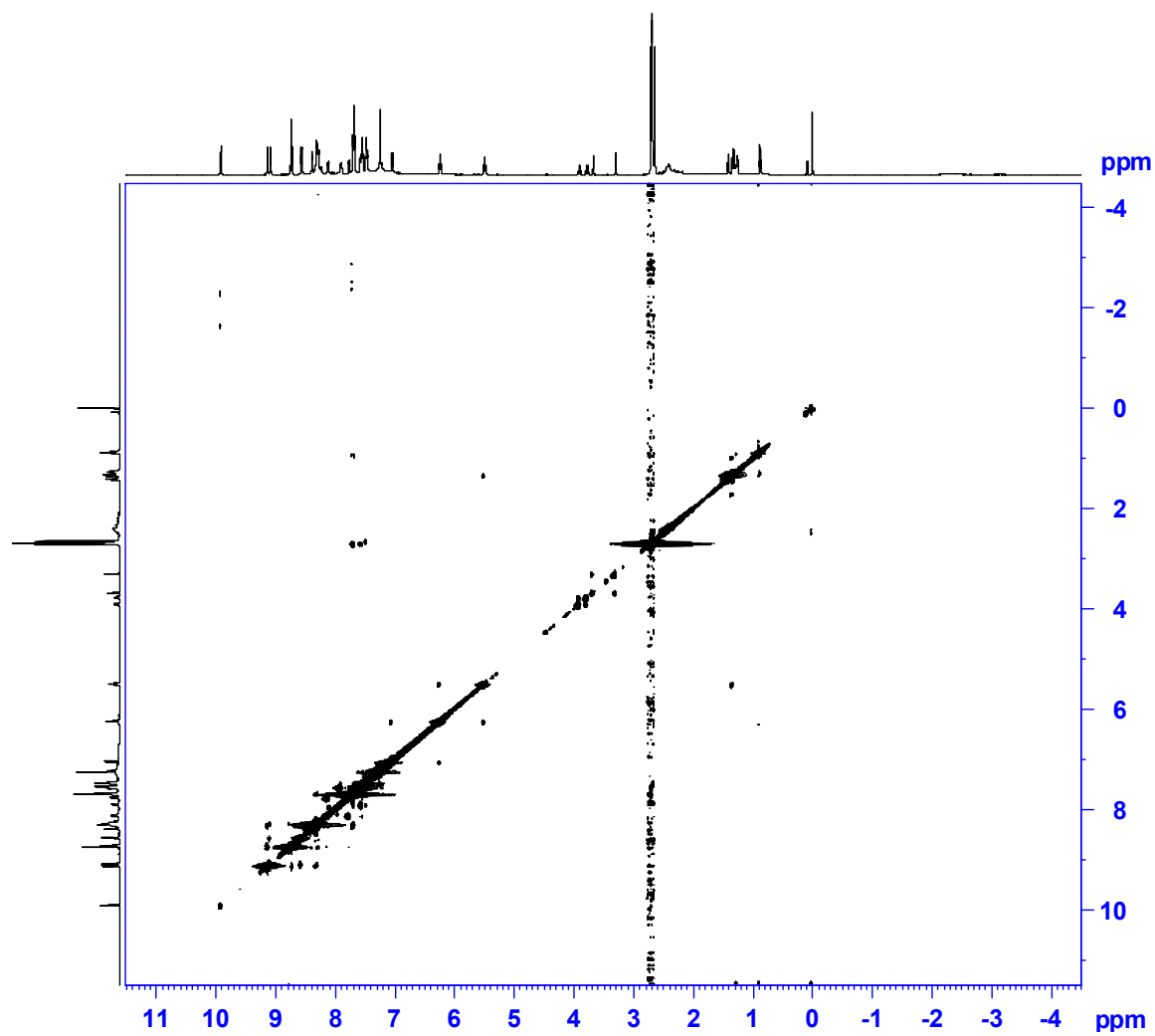


Figure S15. NOESY spectrum of **2b** ( $\text{CDCl}_3$ , 298 K).

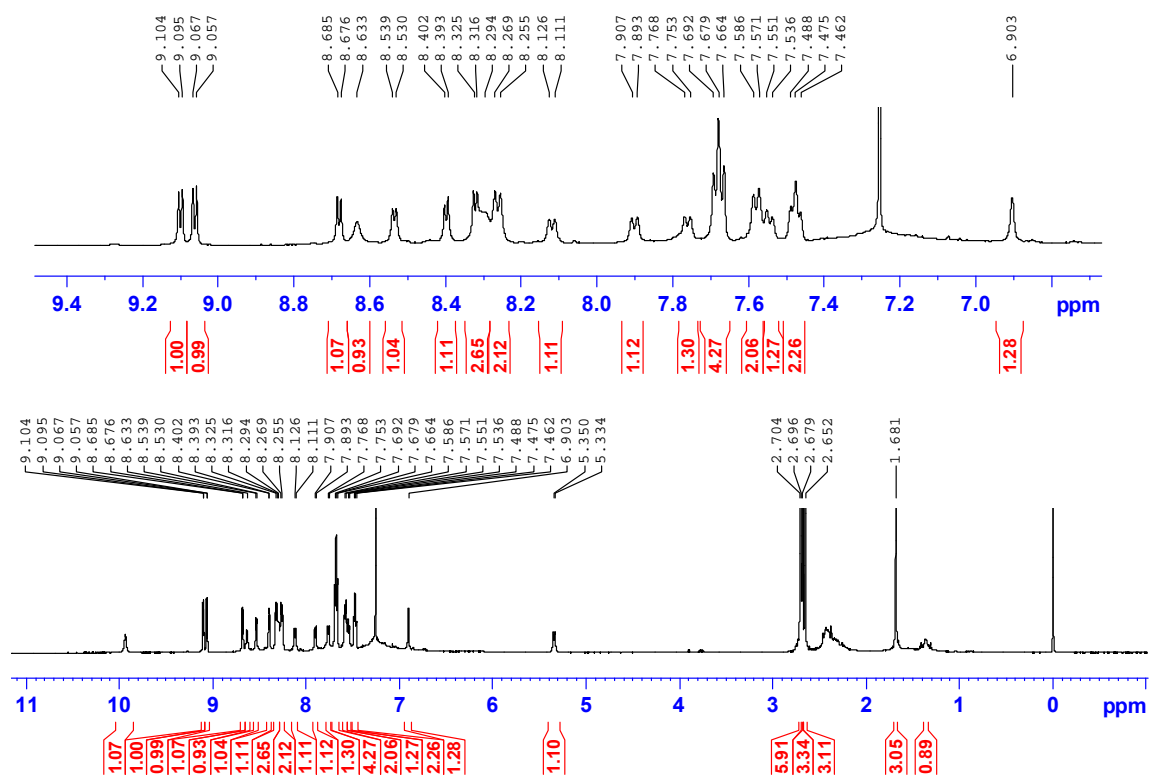


Figure S16.  $^1\text{H}$  NMR spectrum of **2c**, 298 K,  $\text{CDCl}_3$ .

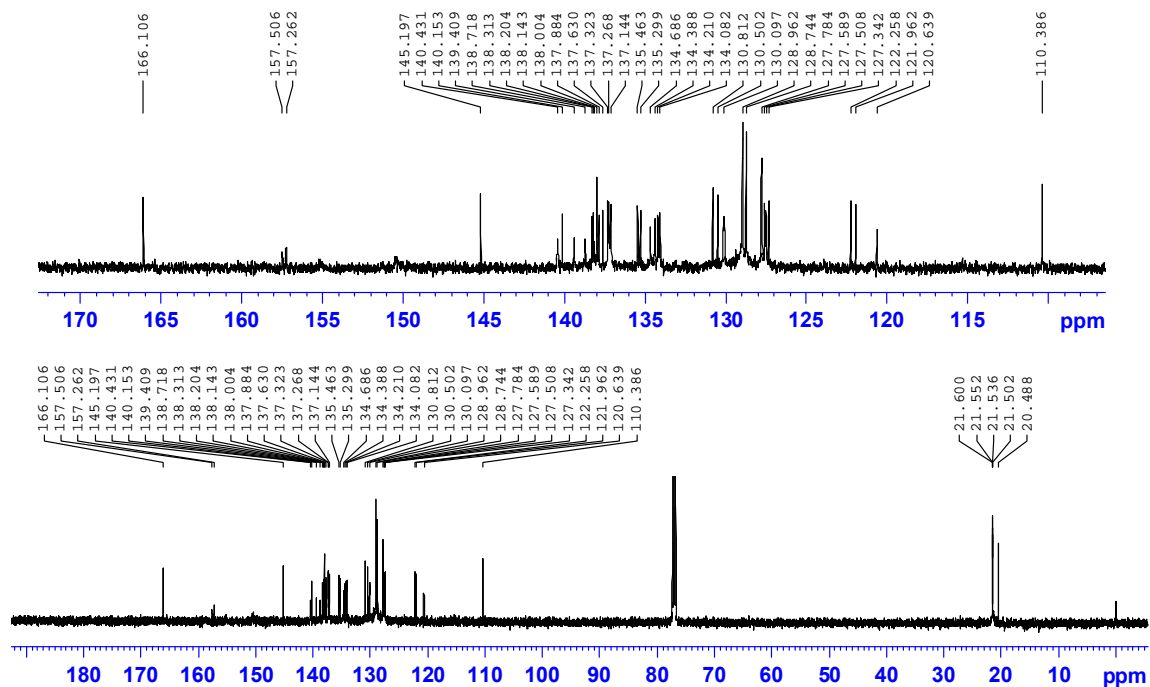


Figure S17.  $^{13}\text{C}$  NMR spectrum of **2c**, 298 K,  $\text{CDCl}_3$ .

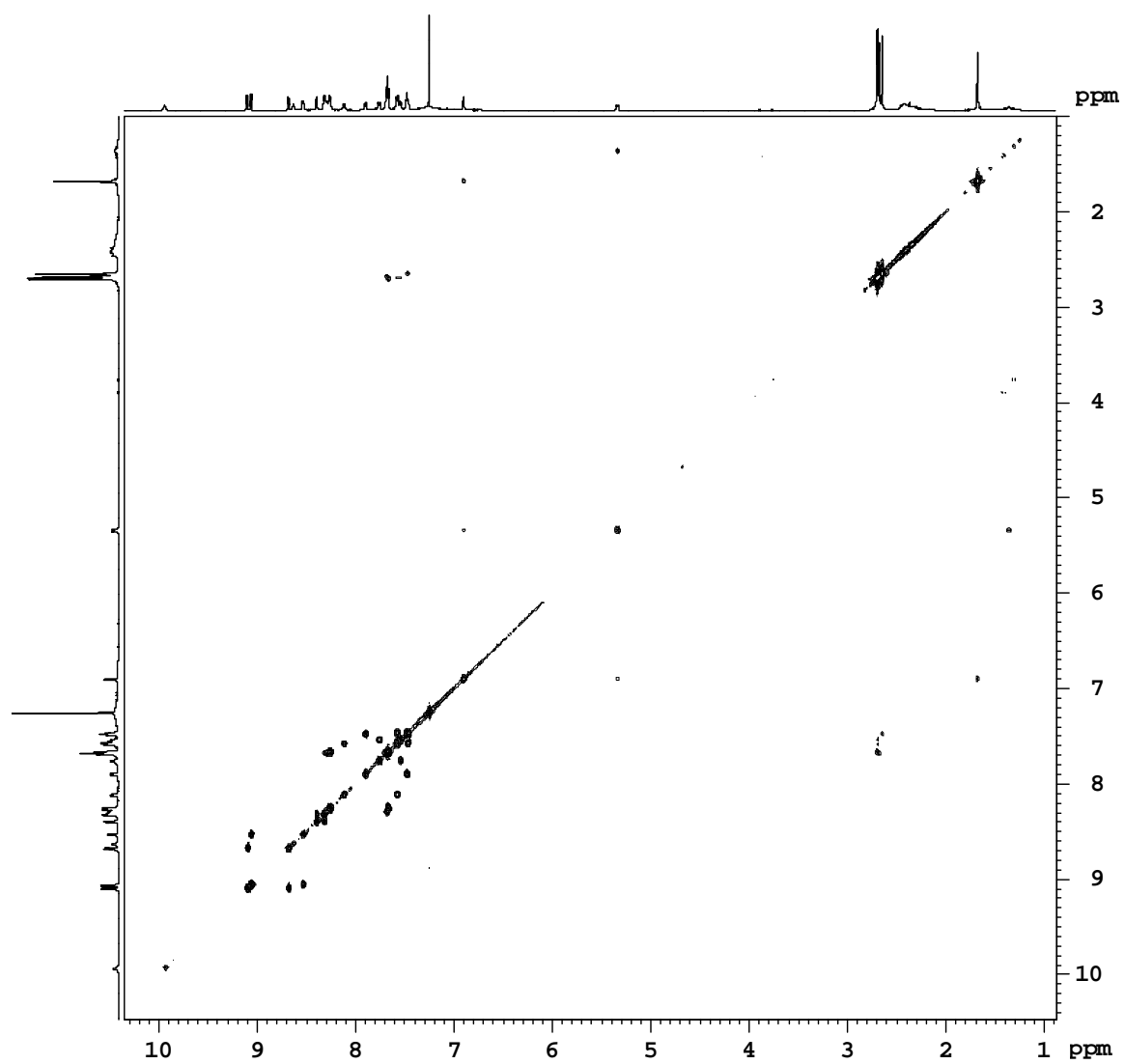


Figure S18. COSY spectrum of 2c (298K).



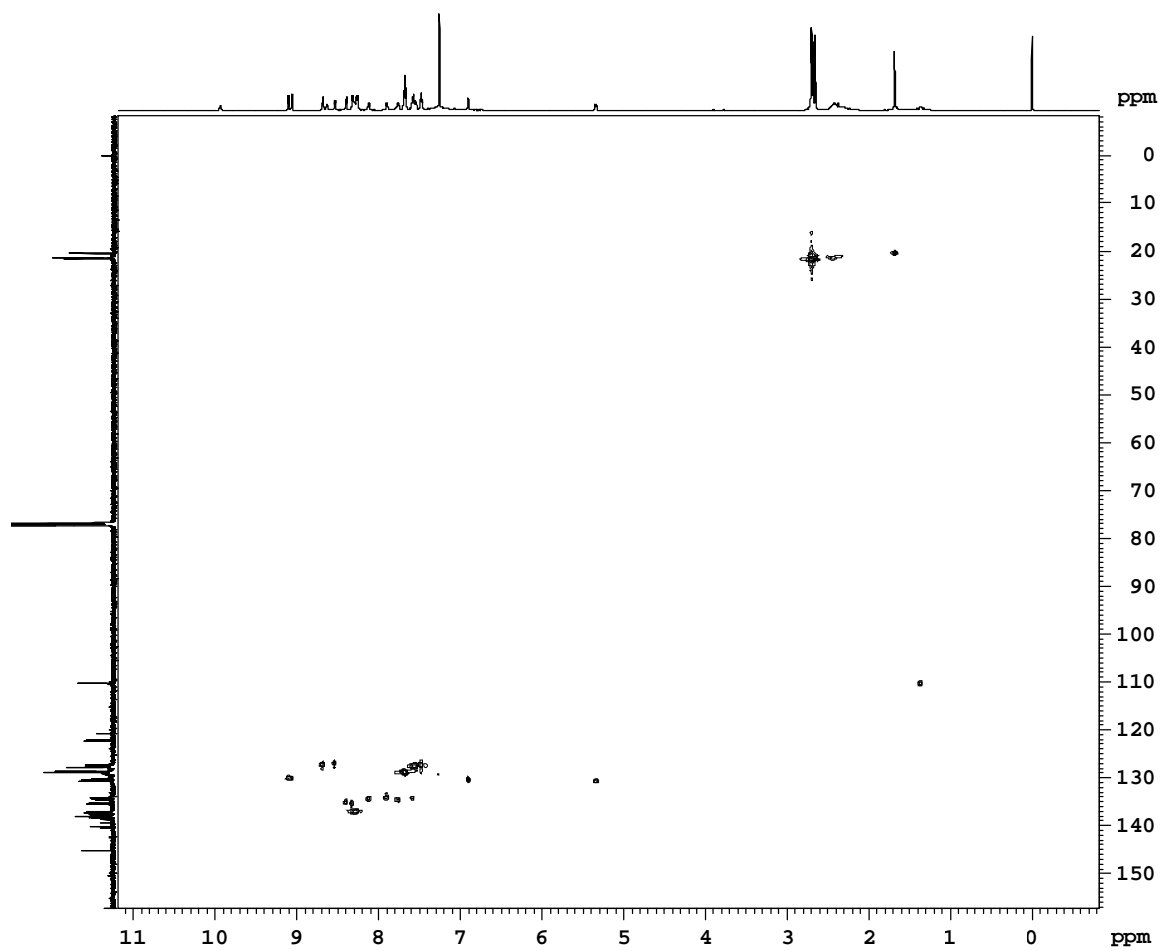


Figure S20. HSQC spectrum of **2c** ( $\text{CDCl}_3$ , 298 K).

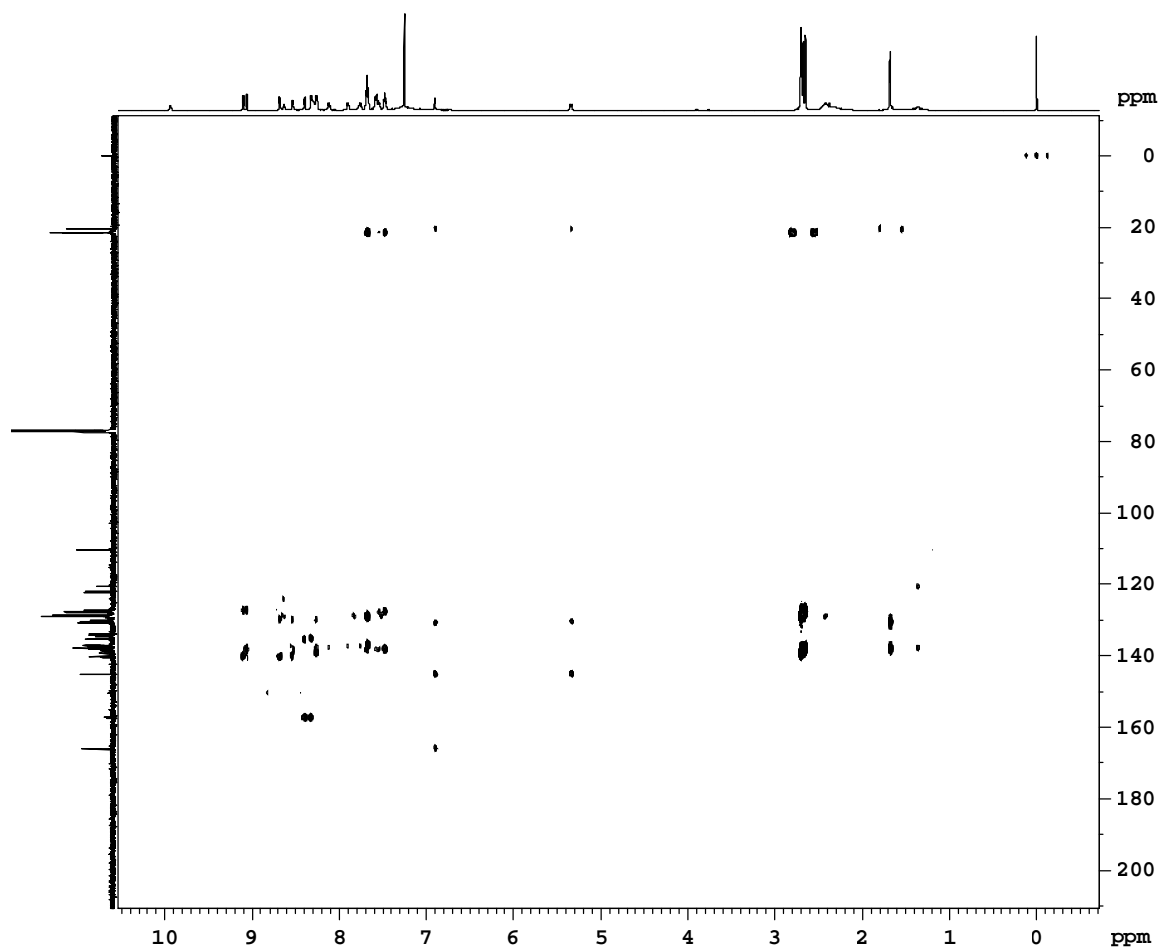


Figure S21. HMBC spectrum of **2c**(CDCl<sub>3</sub>, 298 K).

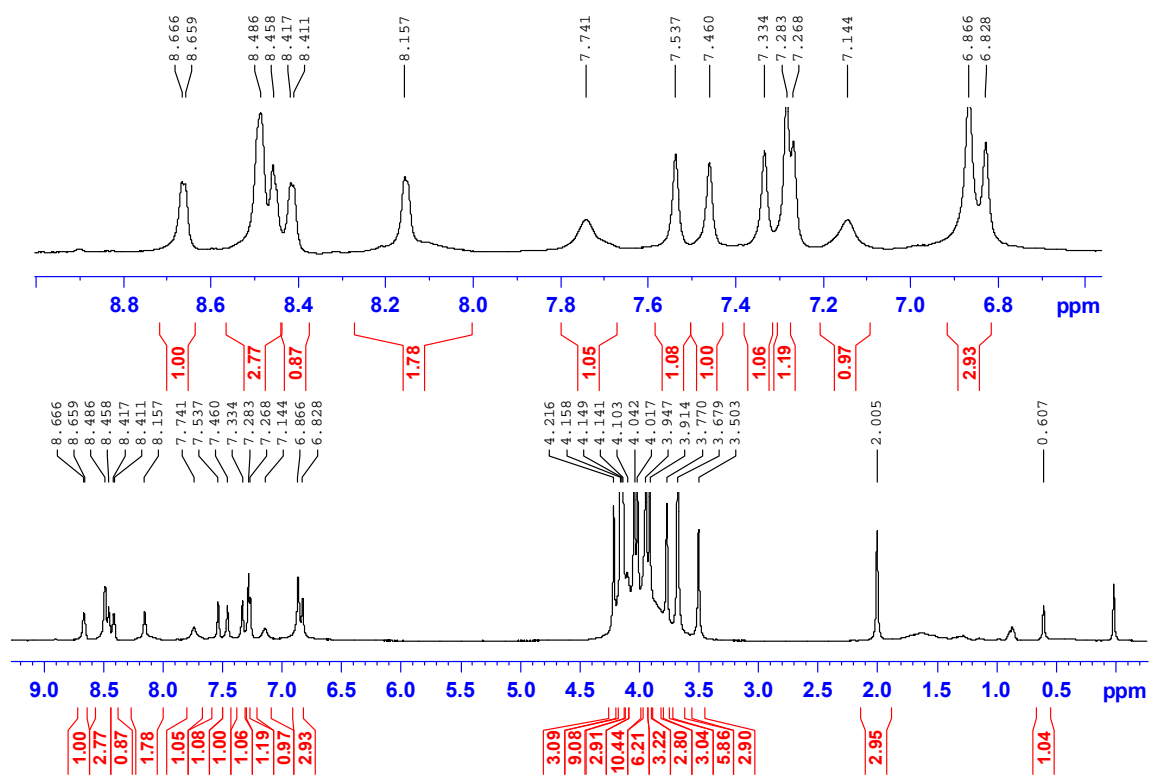


Figure S22.  $^1\text{H}$  NMR spectrum of **3a**, 298 K,  $\text{CDCl}_3$ .

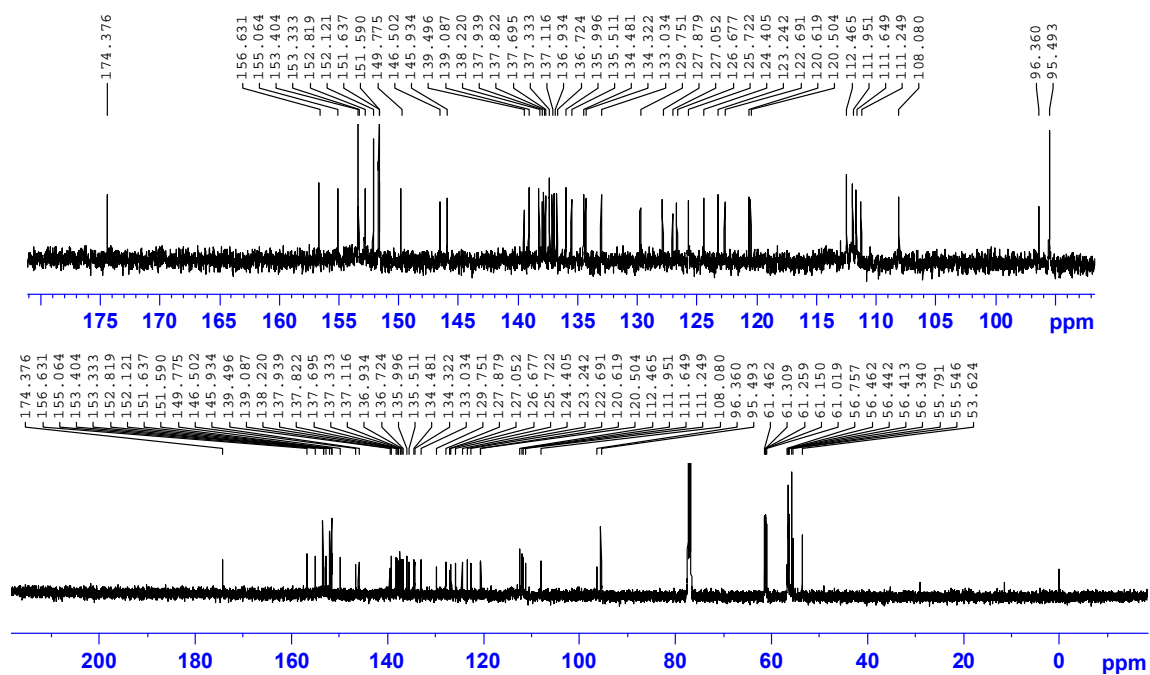


Figure S23.  $^{13}\text{C}$  NMR spectrum of **3a**, 298 K,  $\text{CDCl}_3$ .

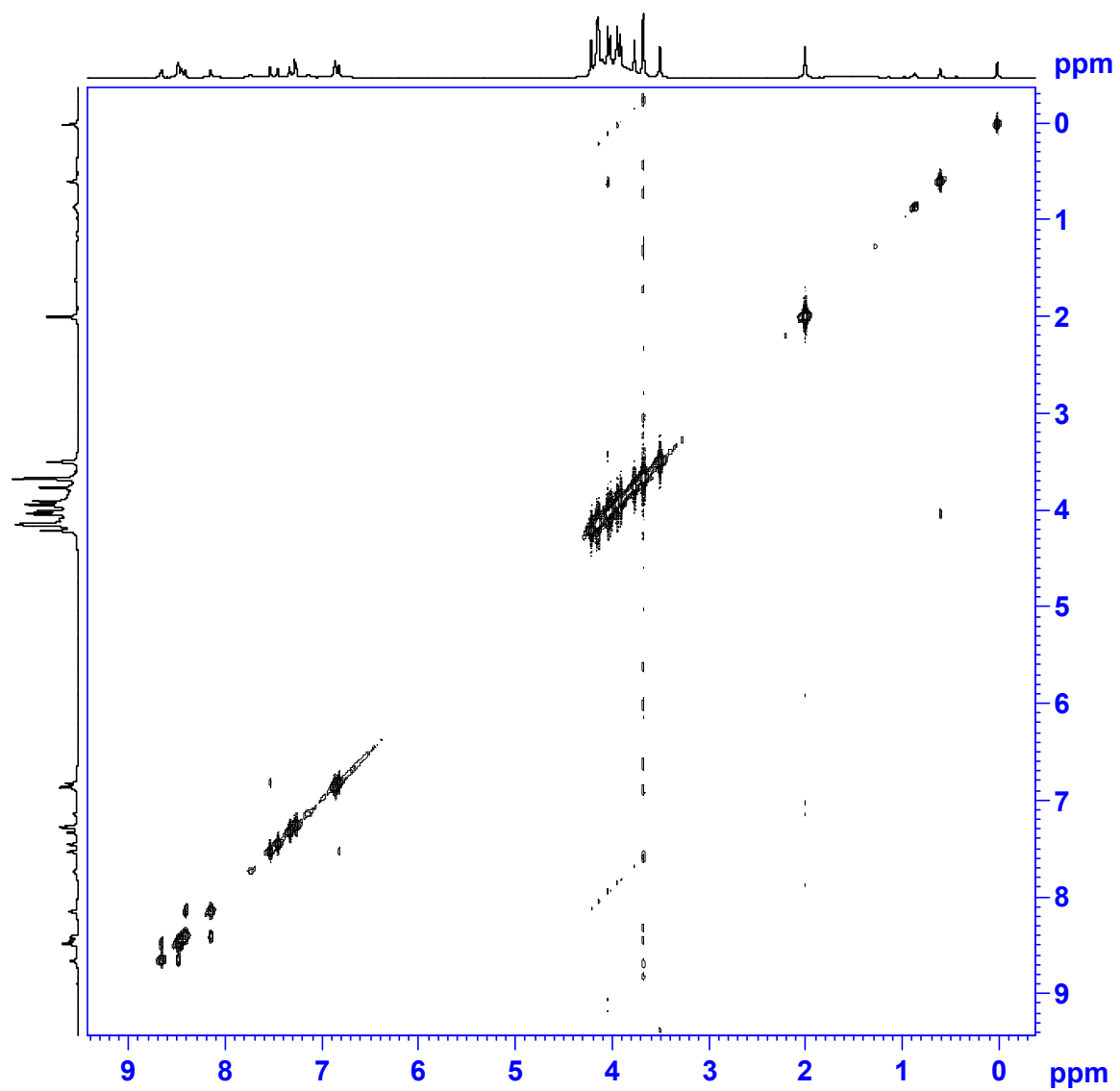


Figure S24. COSY spectrum of 3a (298K).

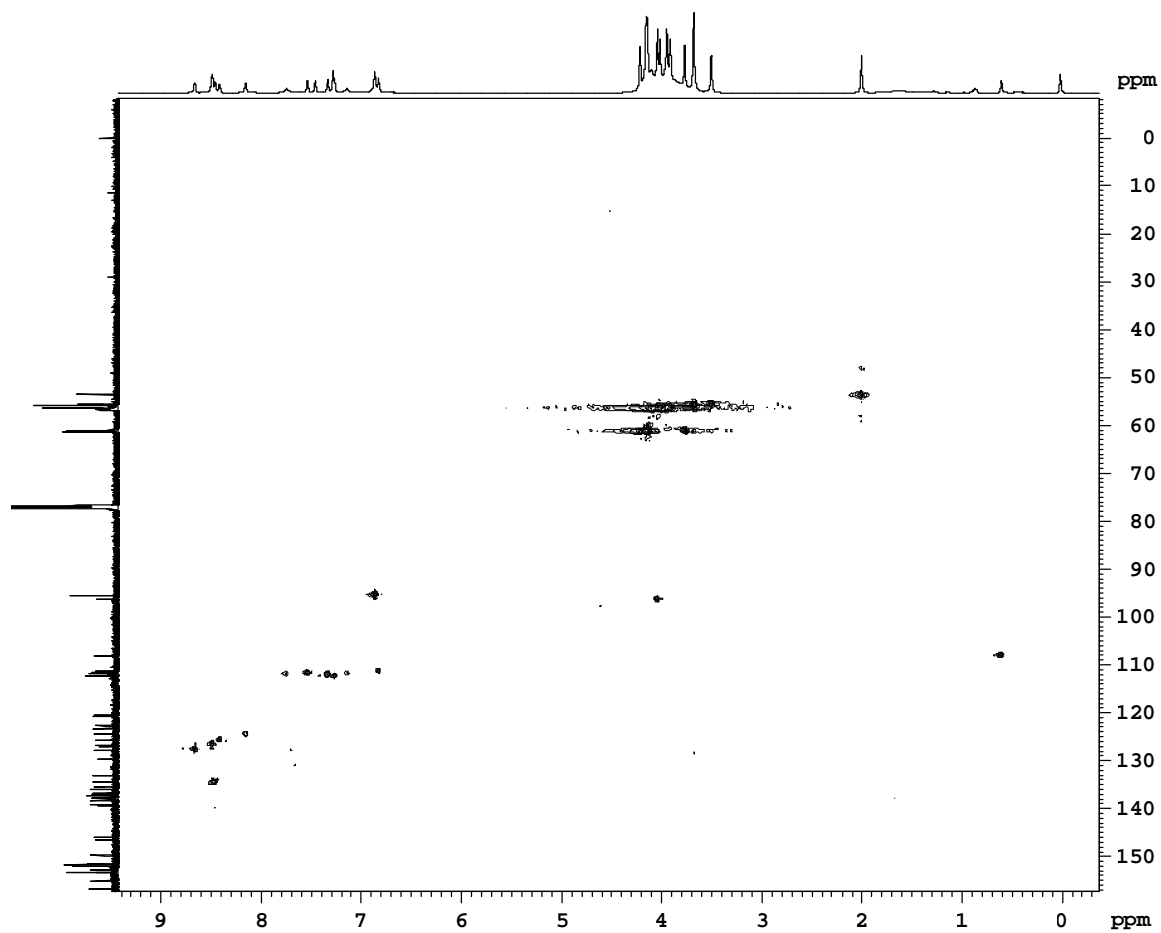


Figure S25. HSQC spectrum of **3a** (CDCl<sub>3</sub>, 298 K).

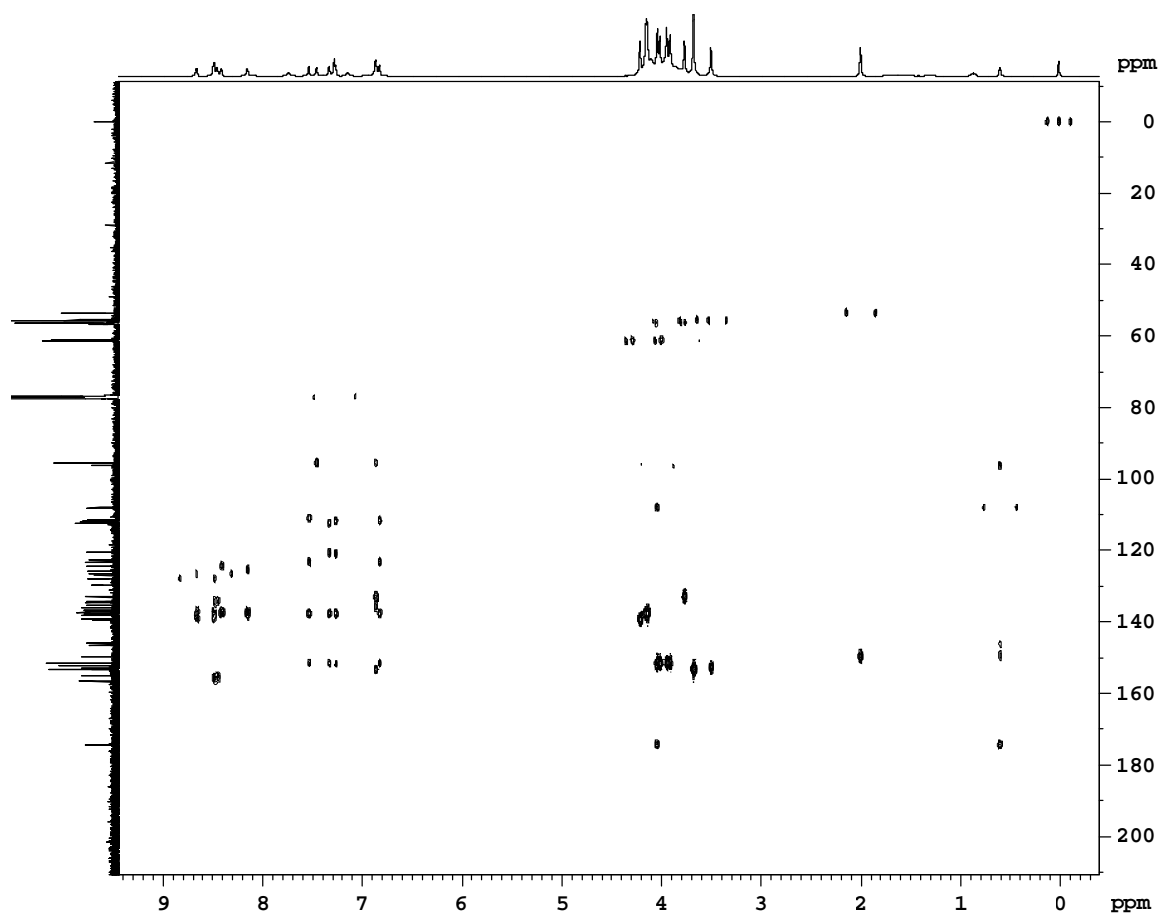


Figure S26. HMBC spectrum of **3a** ( $\text{CDCl}_3$ , 298 K).

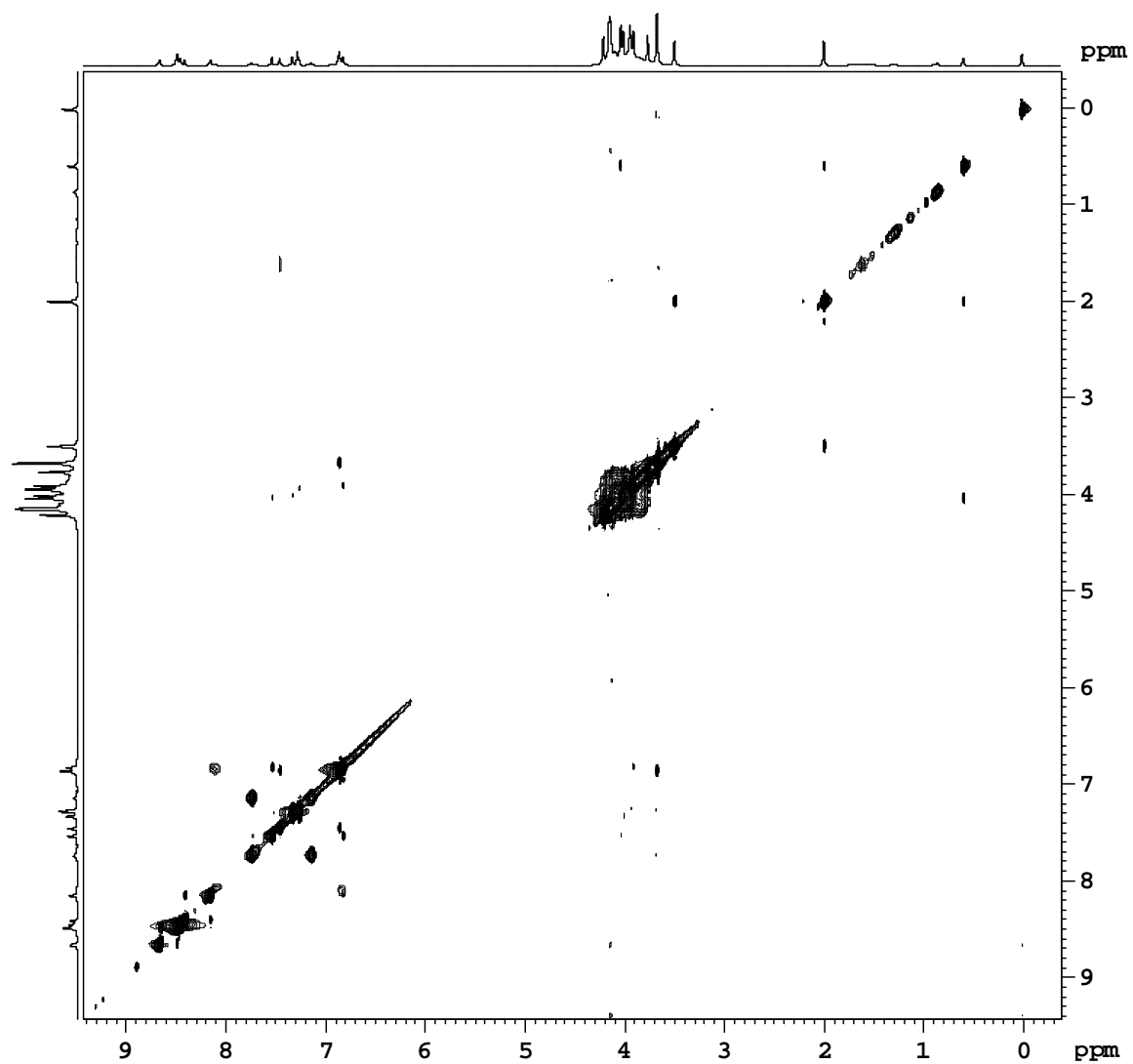


Figure S27. NOESY spectrum of **3a** (CDCl<sub>3</sub>, 298 K).

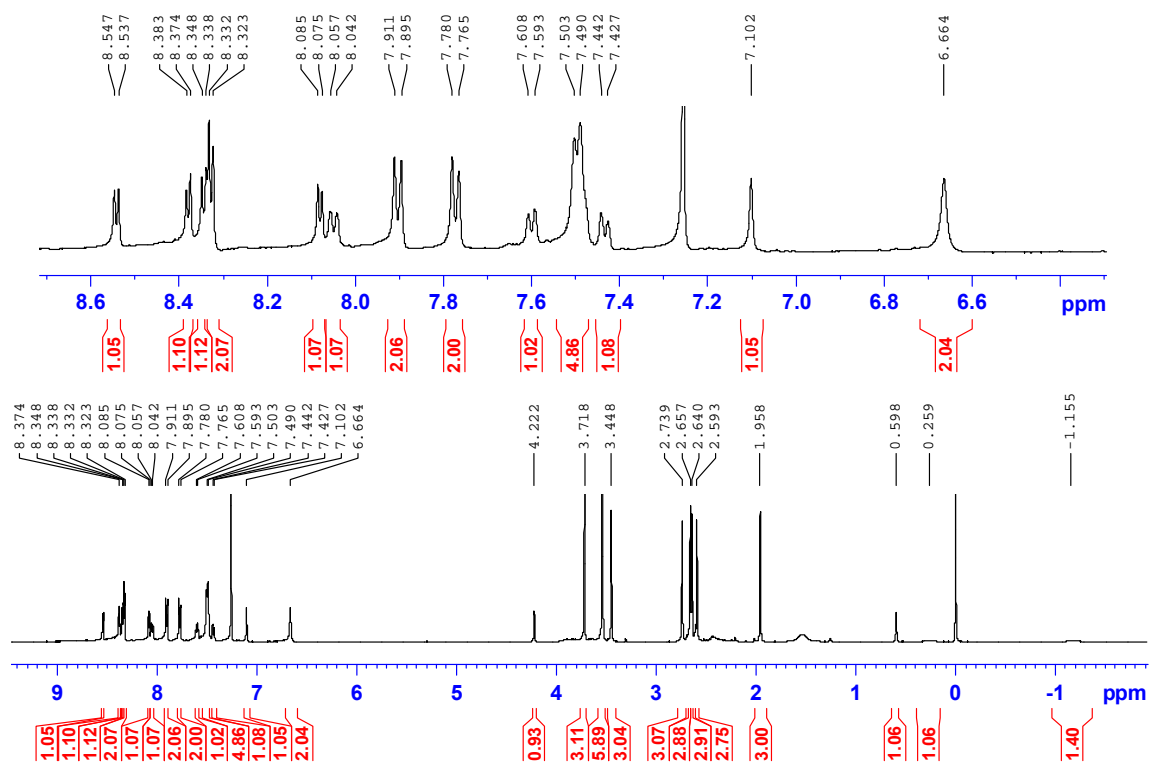


Figure S28.  $^1\text{H}$  NMR spectrum of **3b**, 298 K,  $\text{CDCl}_3$ .

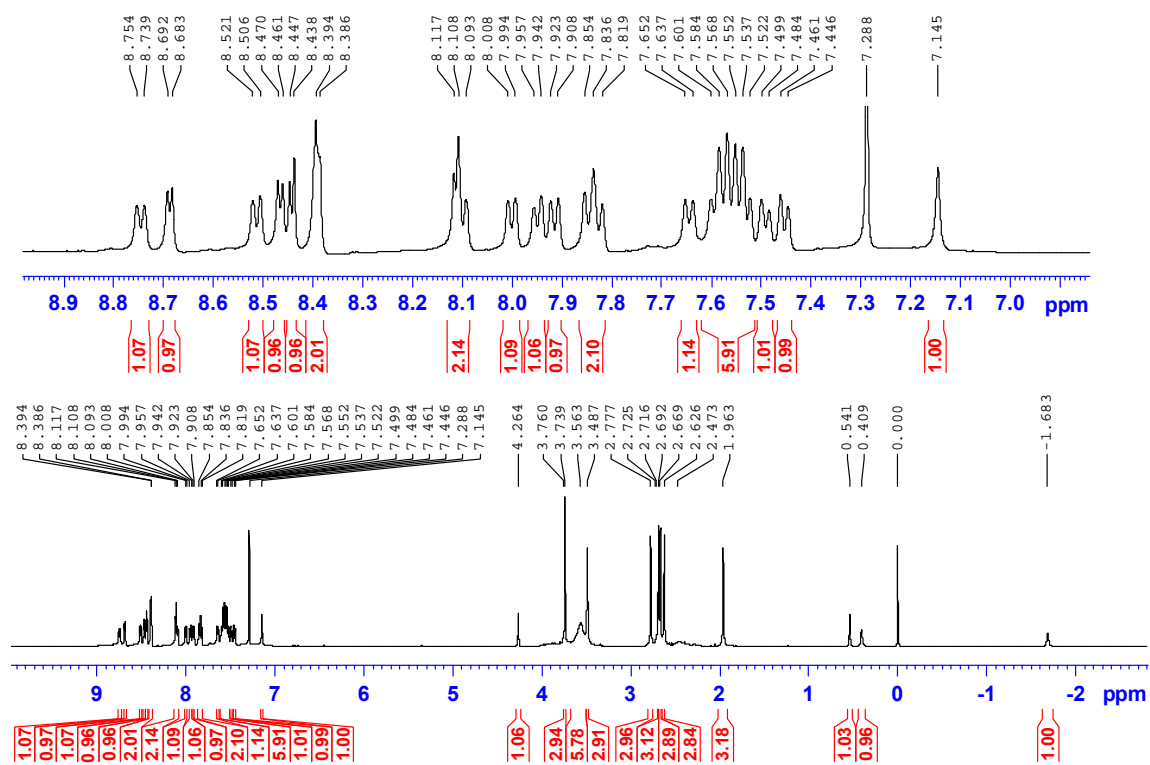
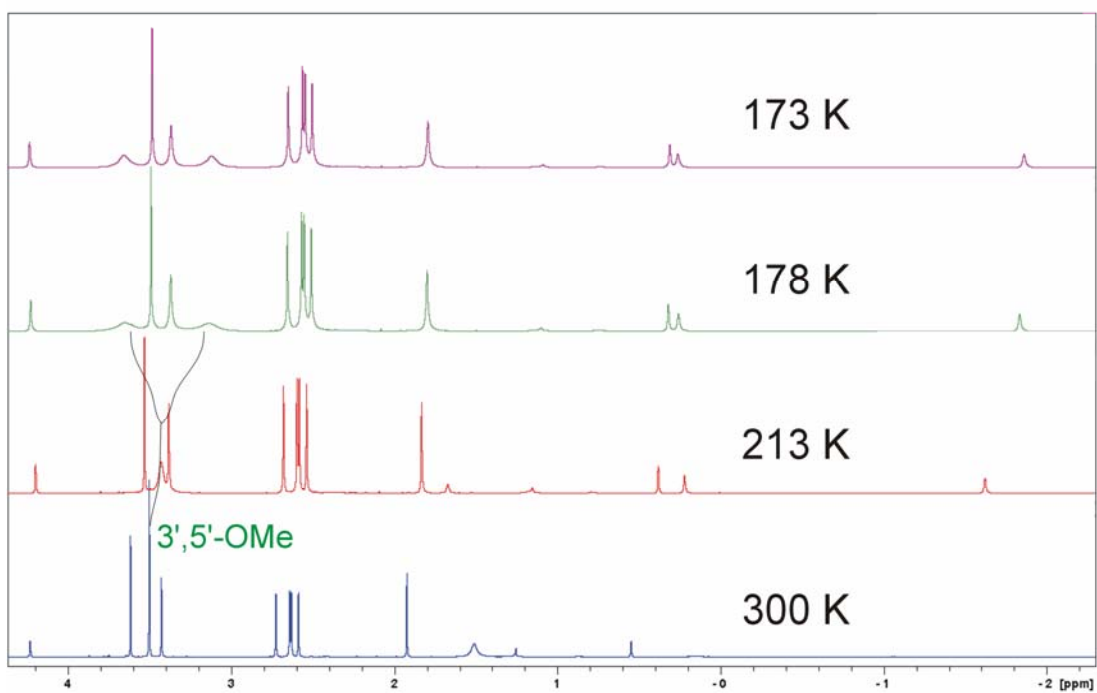
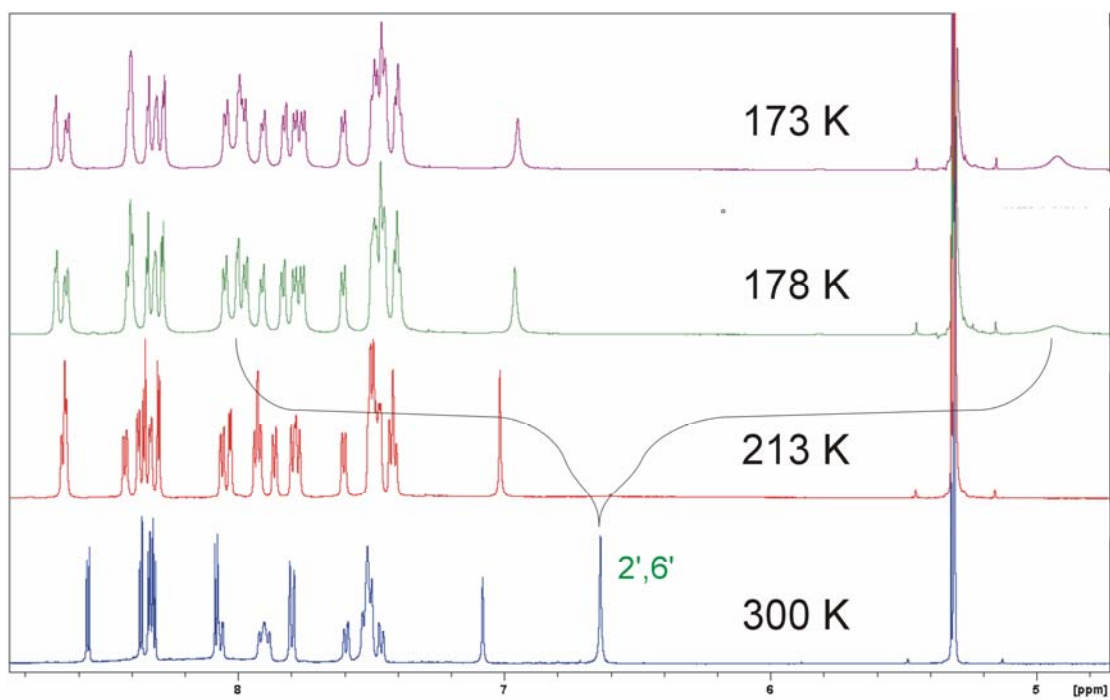


Figure S29.  $^1\text{H}$  NMR spectrum of **3b**, 233 K,  $\text{CDCl}_3$ .





**Figure S29a.** Variable temperature  $^1\text{H}$  NMR spectrum of **3b** (600 MHz,  $\text{CD}_2\text{Cl}_2$ ).

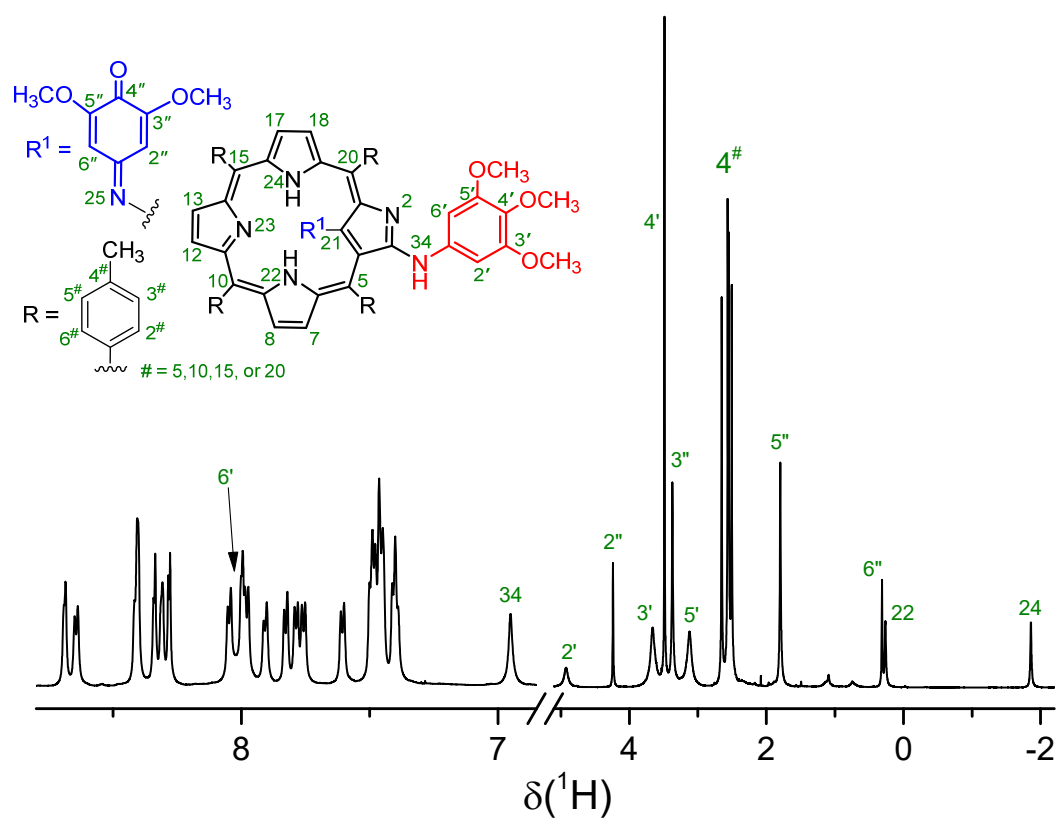


Figure S29b.  $^1\text{H}$  NMR spectrum (600 MHz,  $\text{CD}_2\text{Cl}_2$ , 173 K) of **3b** with a partial assignment of signals.

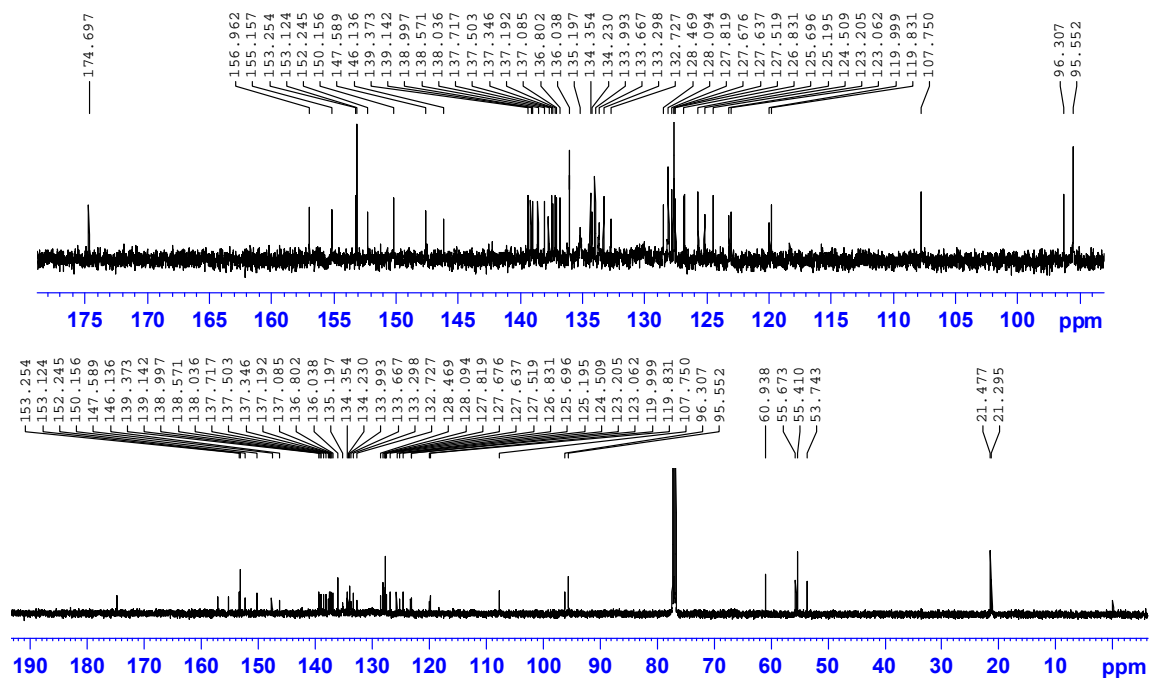


Figure S30.  $^{13}\text{C}$  NMR spectrum of **3b**, 298 K,  $\text{CDCl}_3$ .

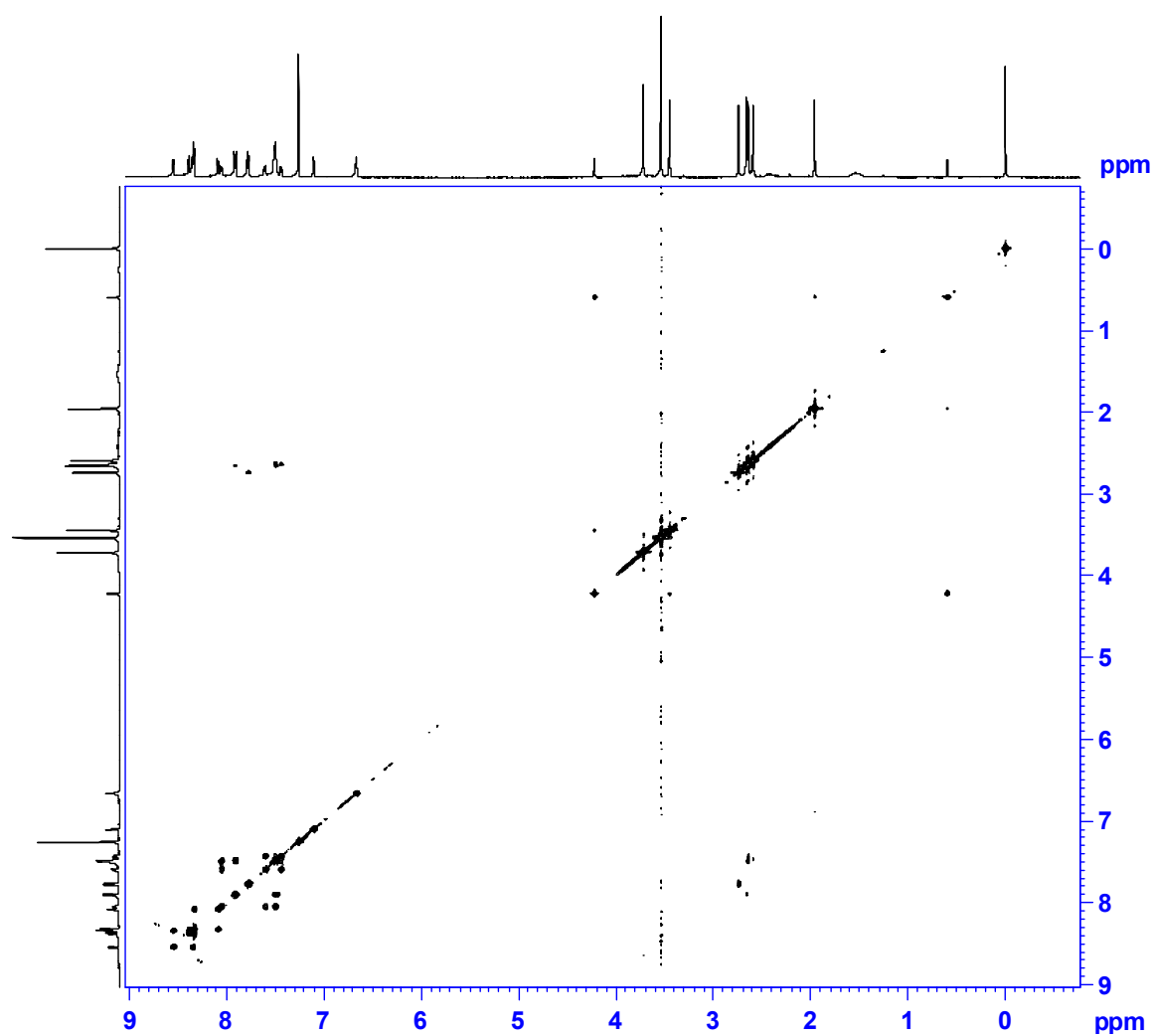


Figure S31. COSY spectrum of **3b** (298K).

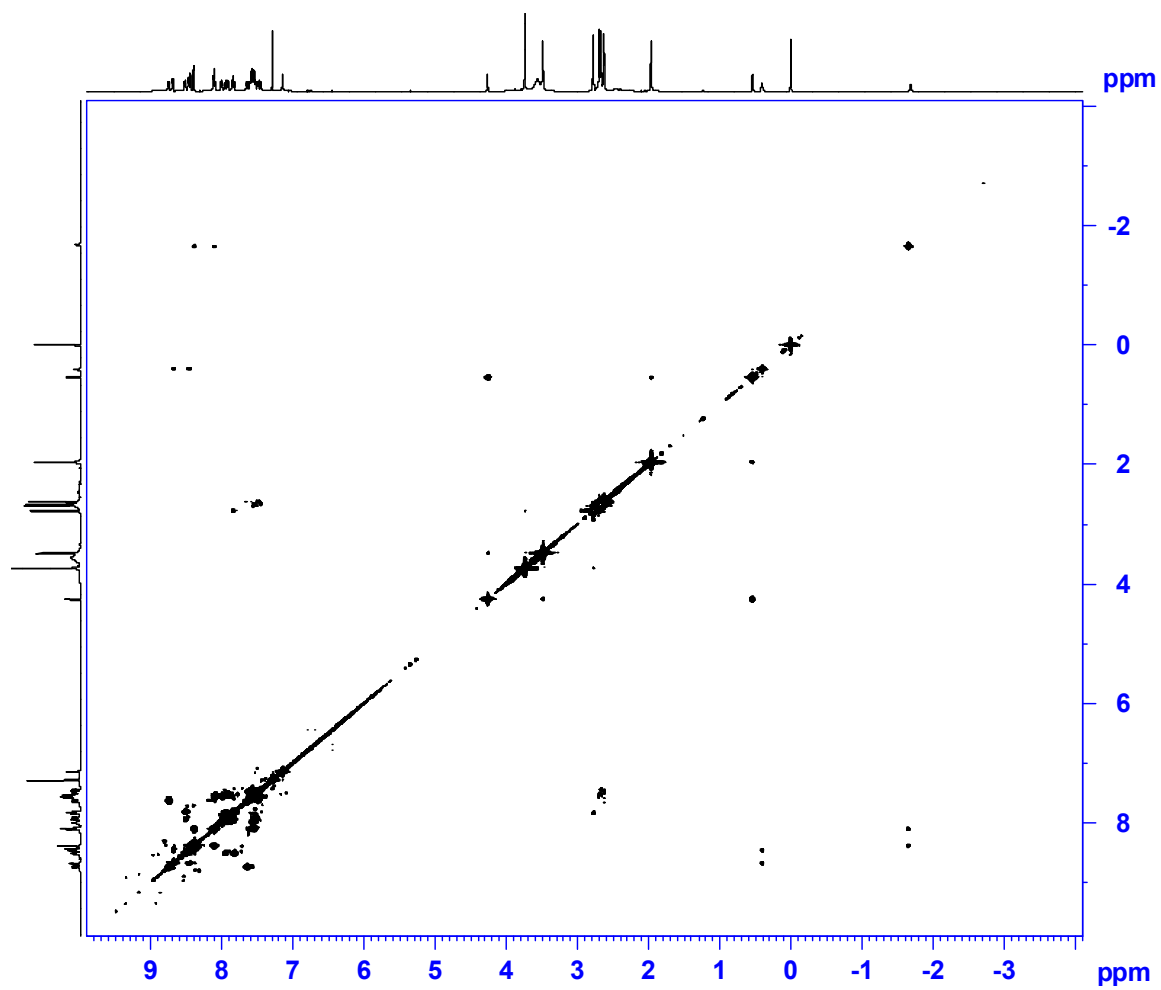


Figure S32. COSY spectrum of **3b** (233K).

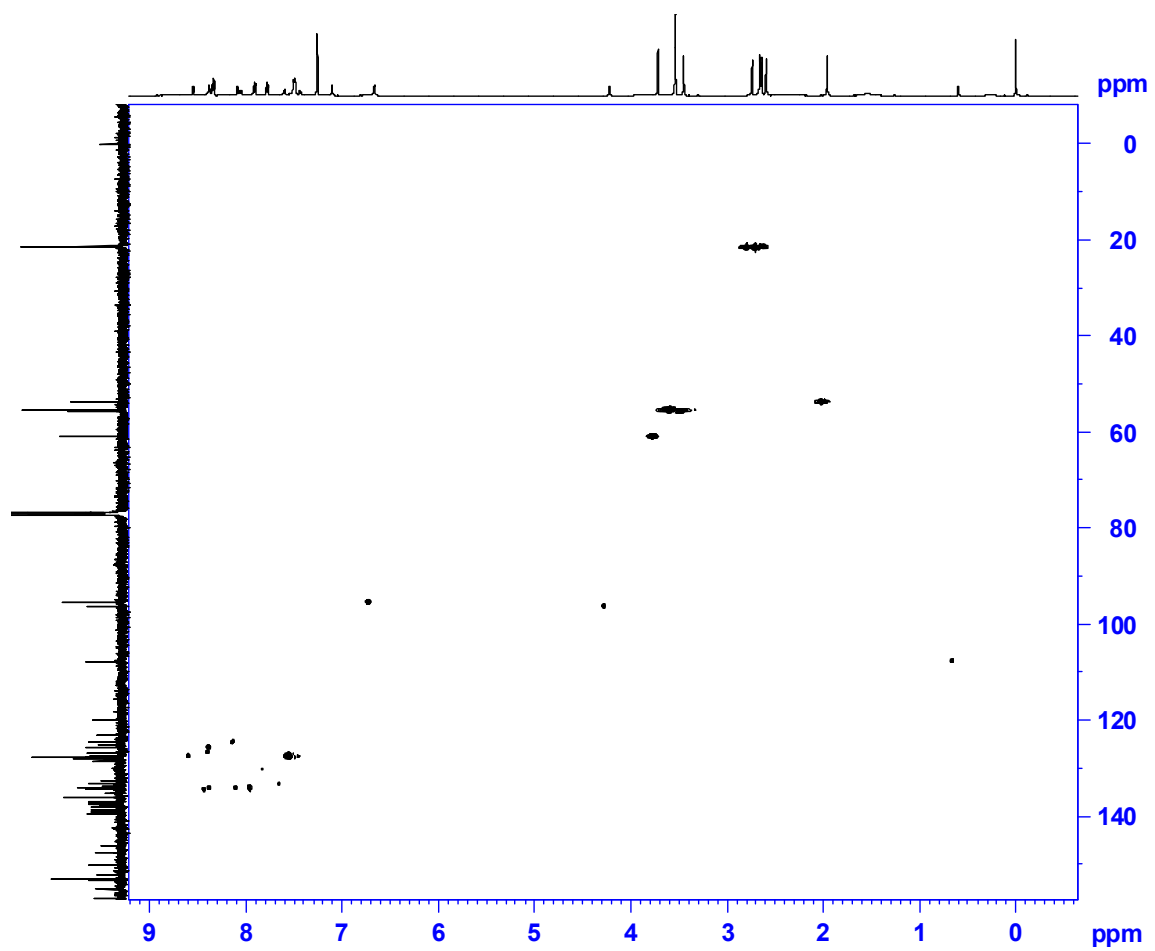


Figure S33. HSQC spectrum of **3b** (CDCl<sub>3</sub>, 298 K).

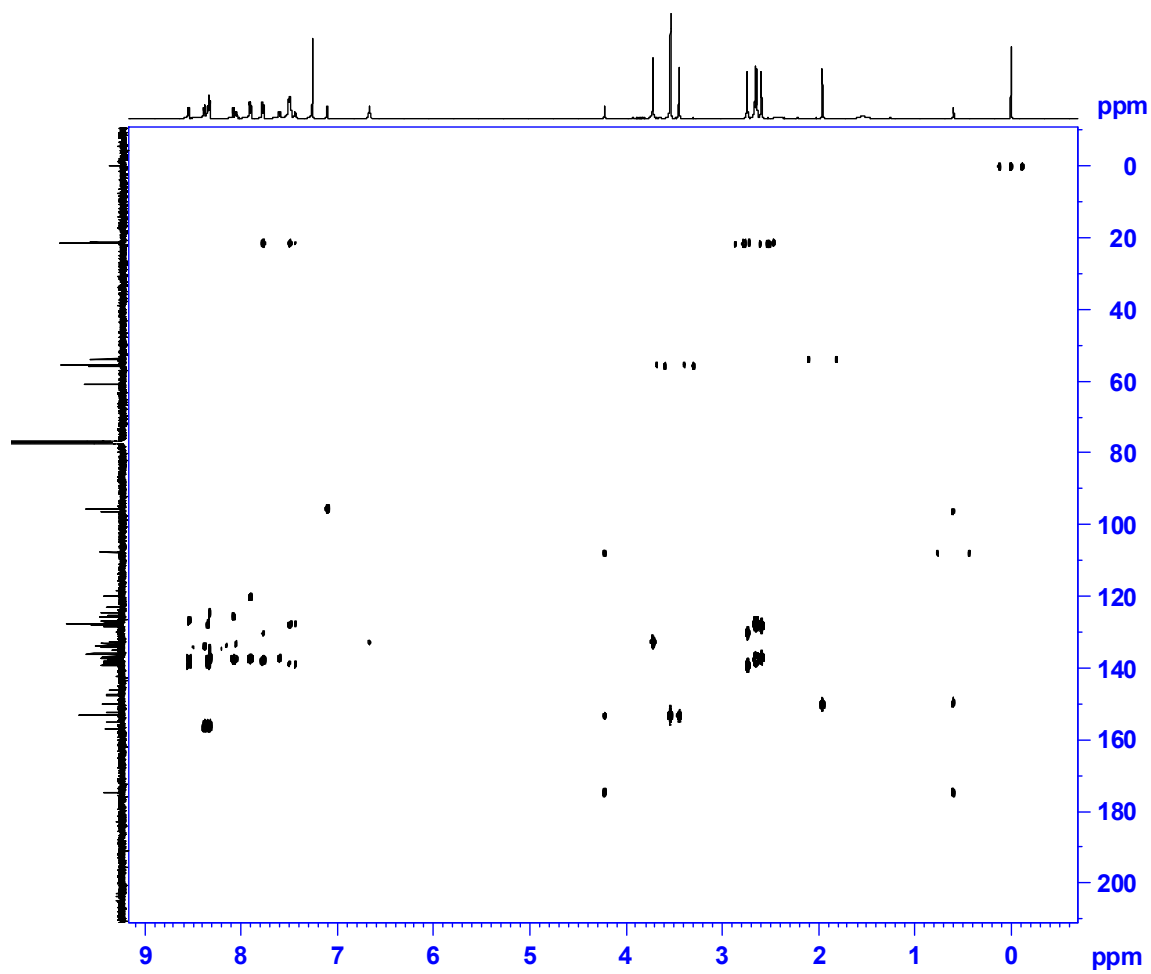


Figure S34. HMBC spectrum of **3b** (CDCl<sub>3</sub>, 298 K).

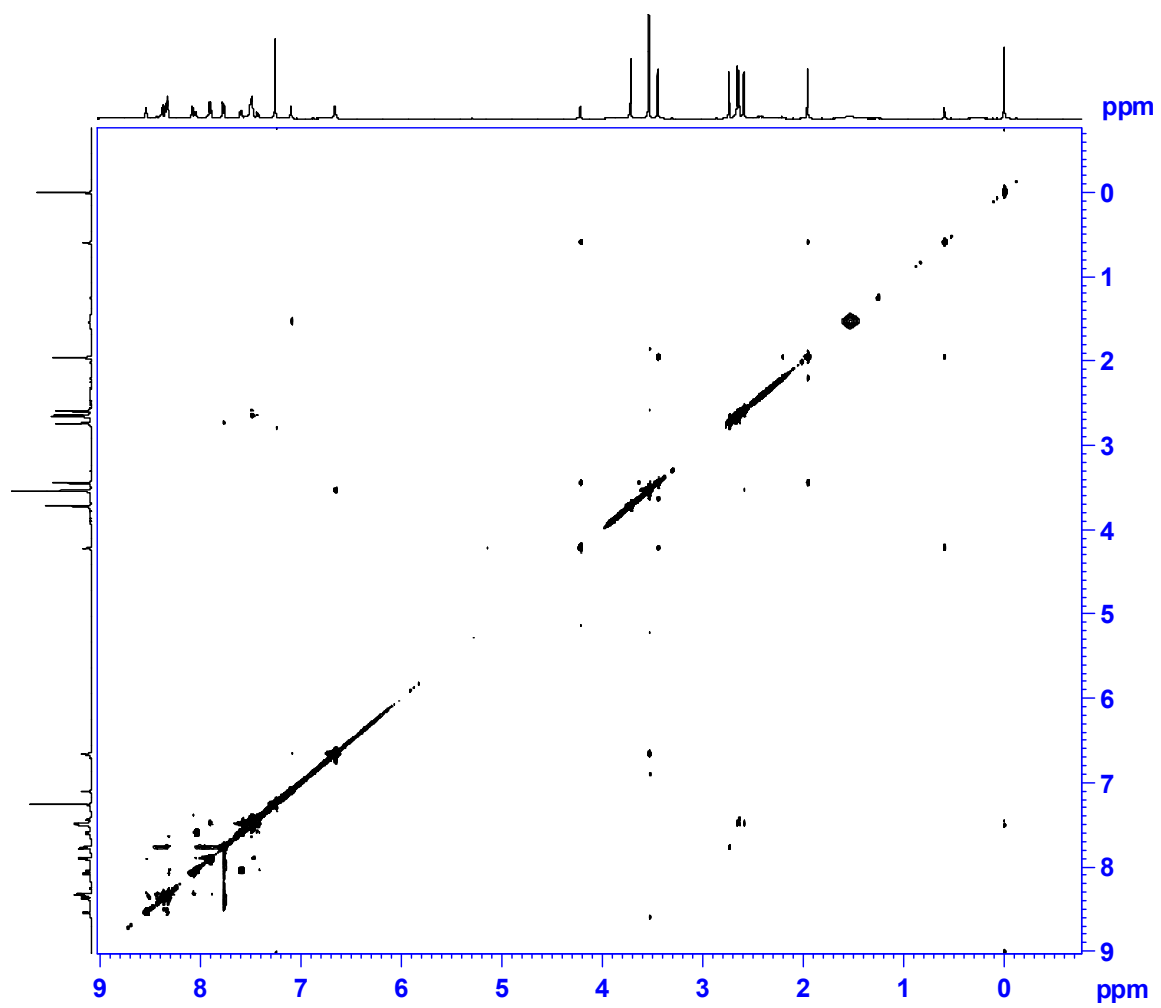


Figure S35. NOESY spectrum of **3b** ( $\text{CDCl}_3$ , 298 K).

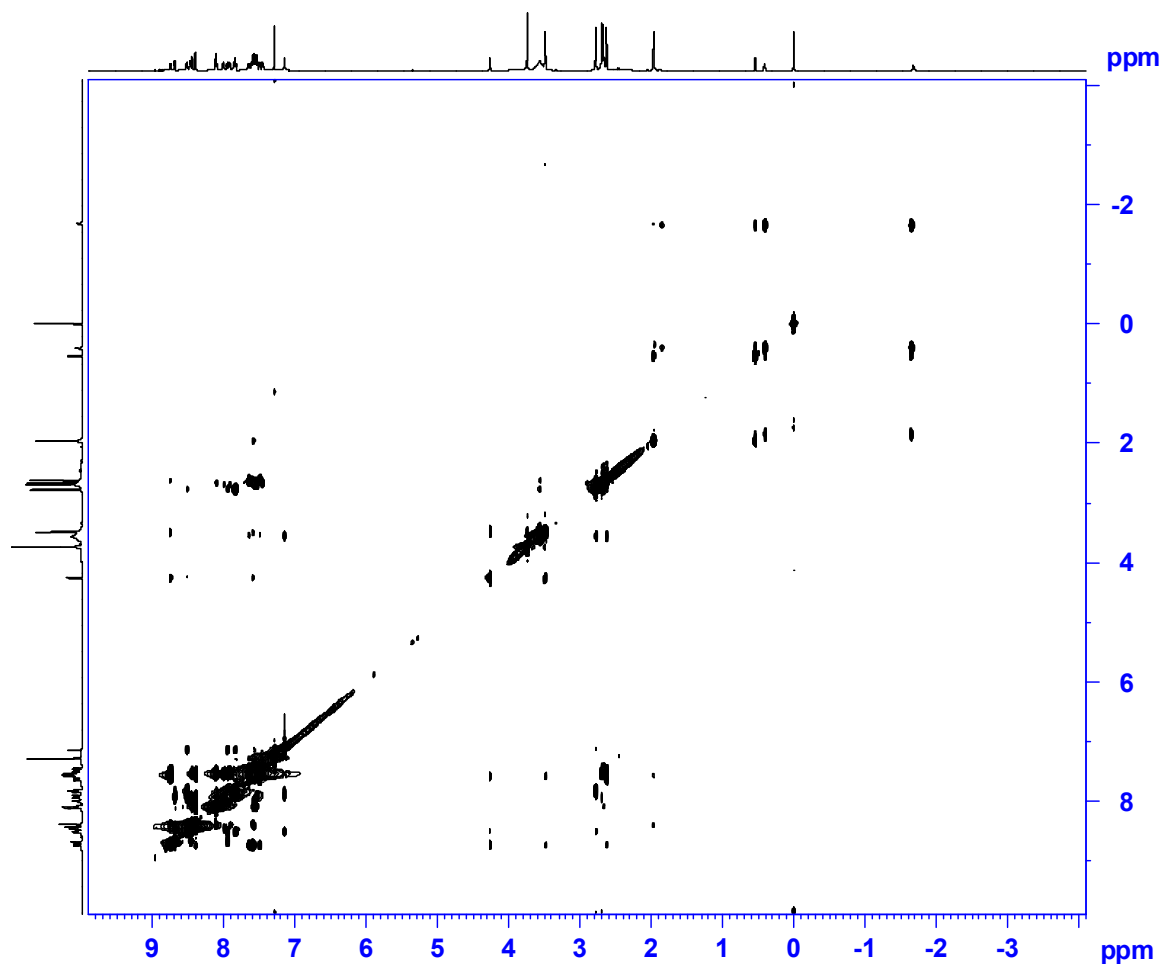


Figure S36. NOESY spectrum of **3b** ( $\text{CDCl}_3$ , 233 K).



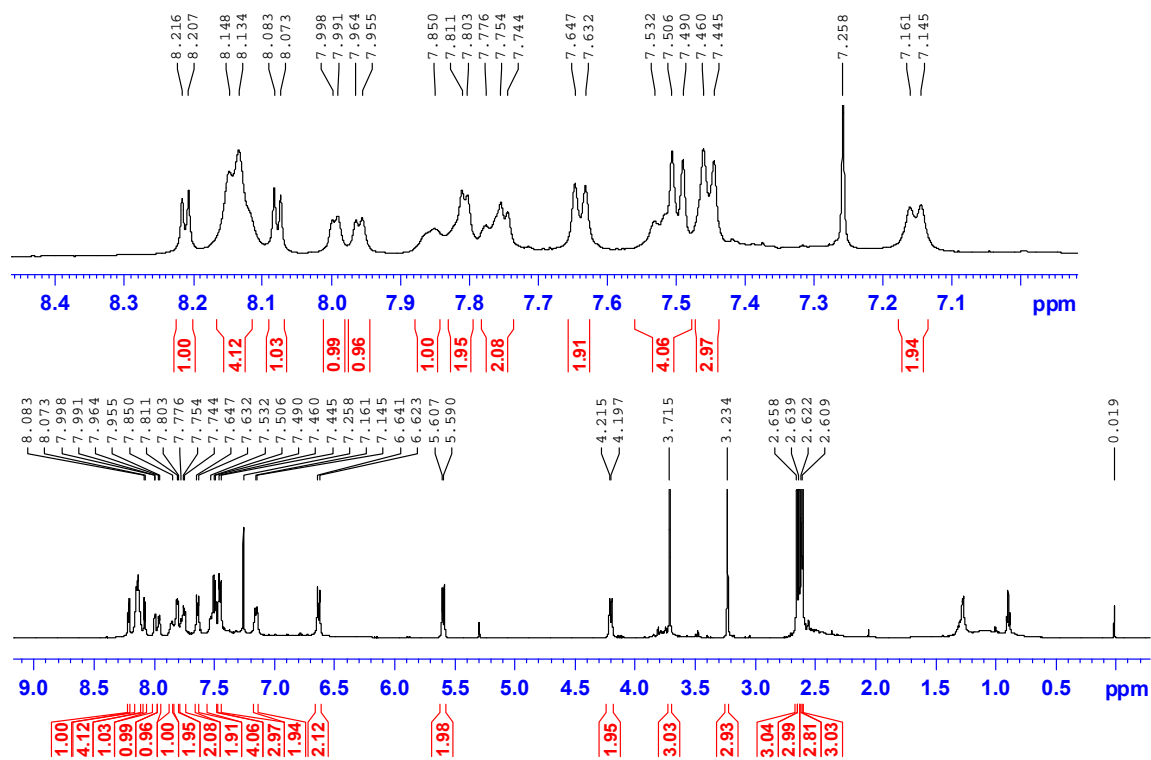


Figure S37.  $^1\text{H}$  NMR spectrum of **4a**, 298 K,  $\text{CDCl}_3$ .

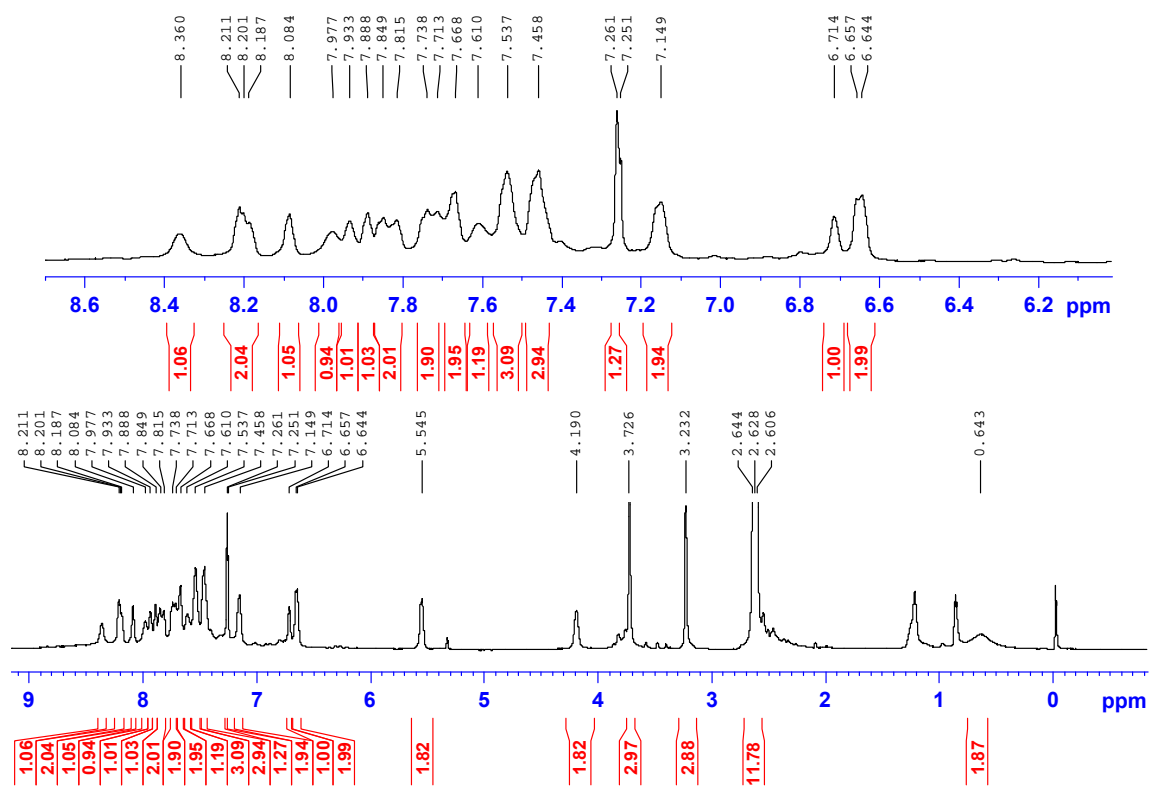


Figure S38.  $^1\text{H}$  NMR spectrum of **4a**, 230 K,  $\text{CDCl}_3$ .

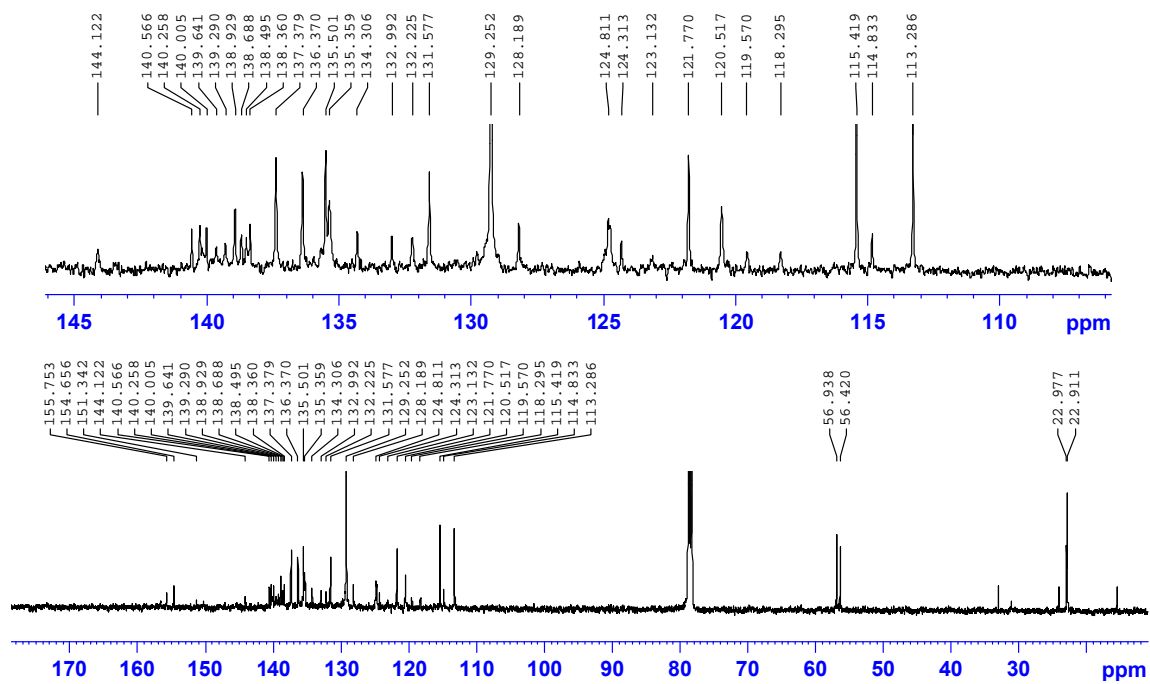
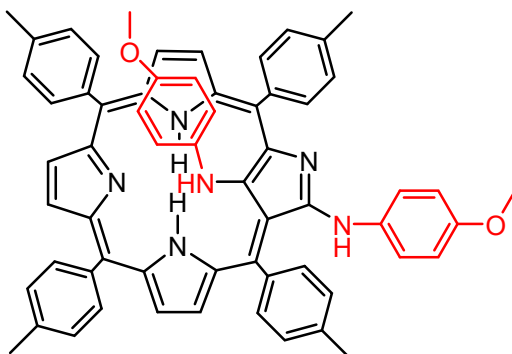


Figure S39.  $^{13}\text{C}$  NMR spectrum of **4a**, 298 K,  $\text{CDCl}_3$ .



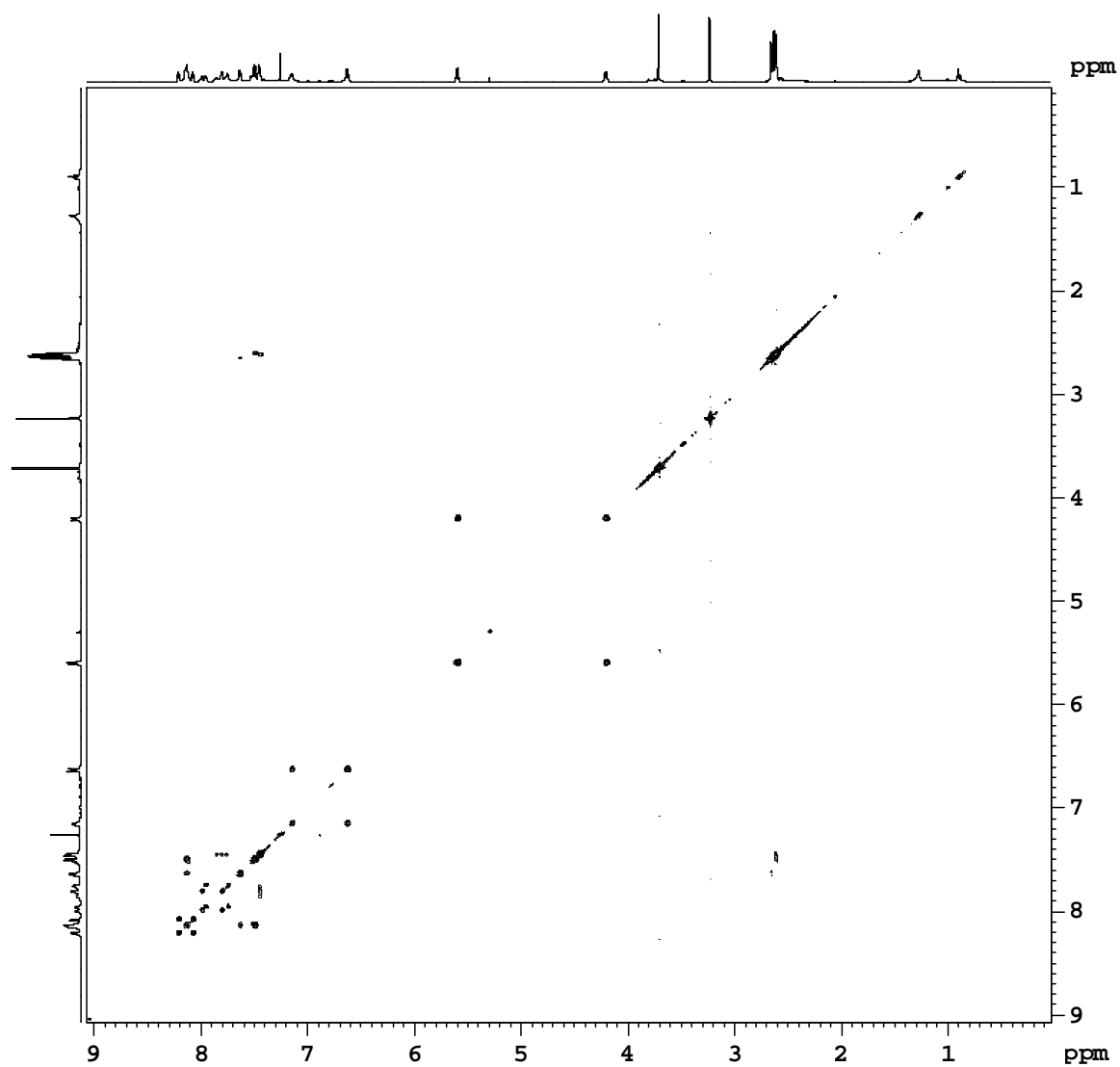
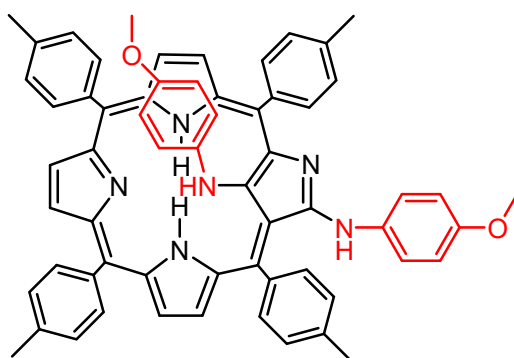


Figure S40. COSY spectrum of 4a (298K).



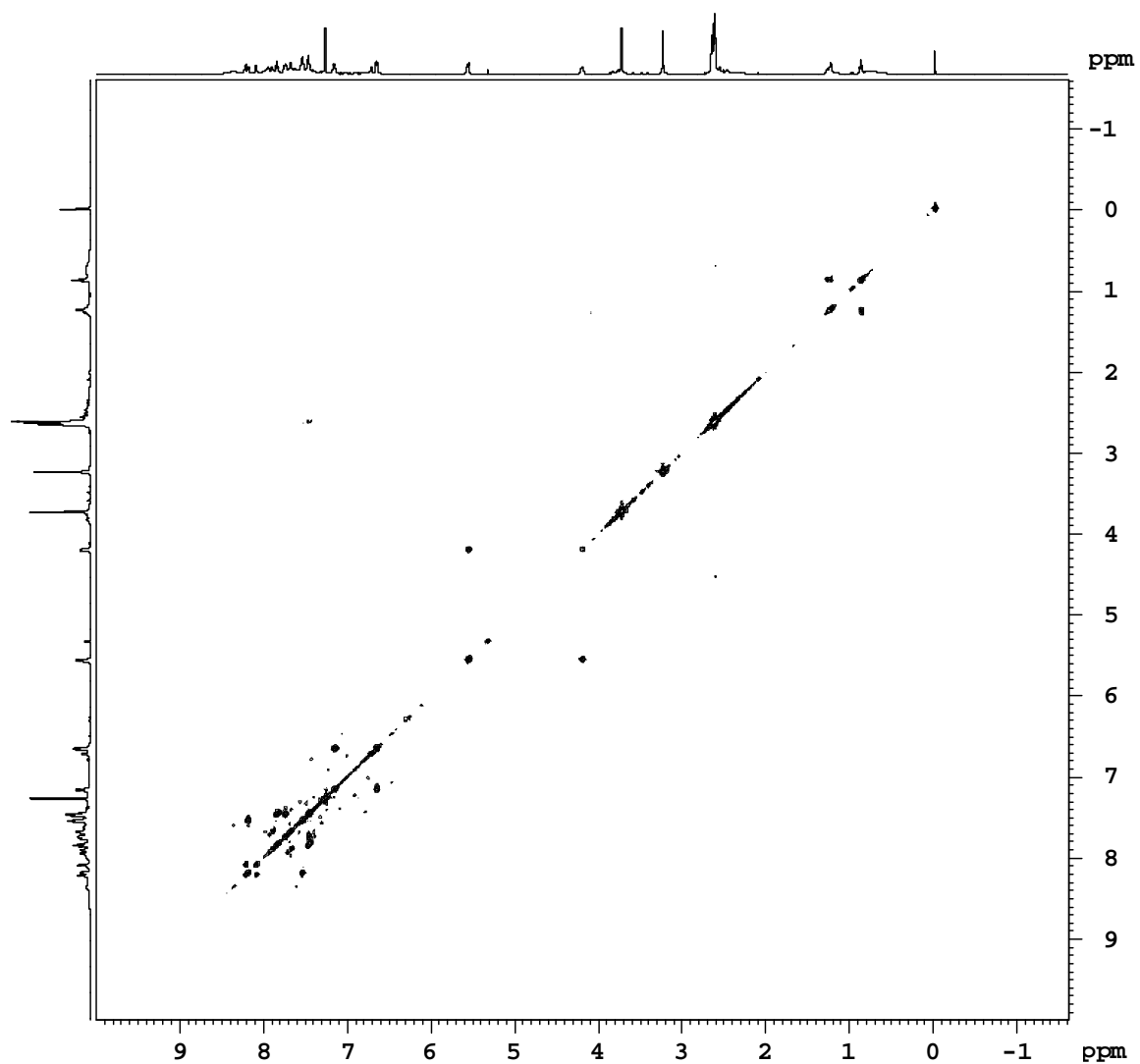
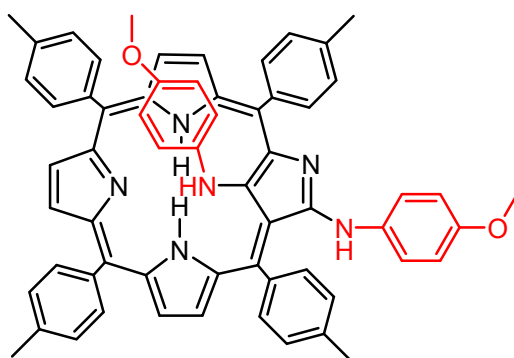


Figure S41. COSY spectrum of 4a (230K).



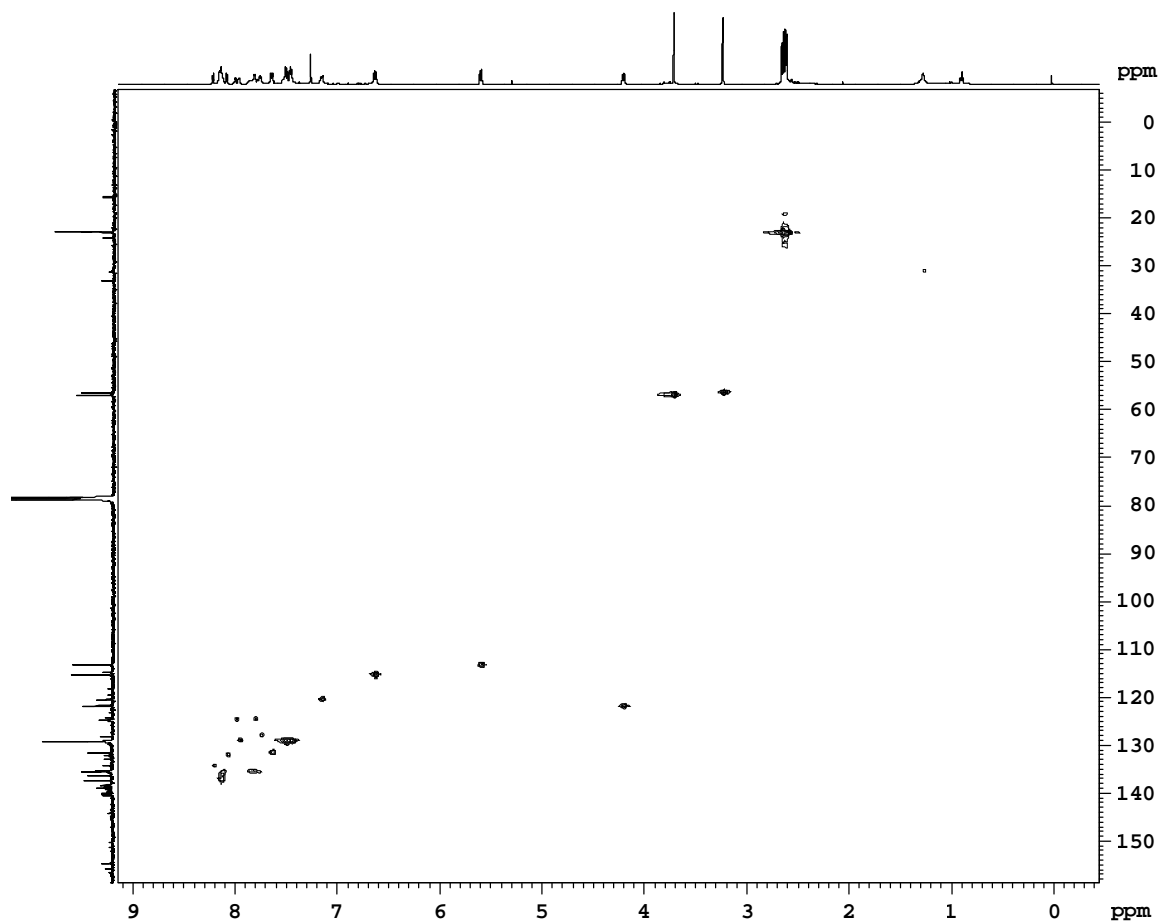
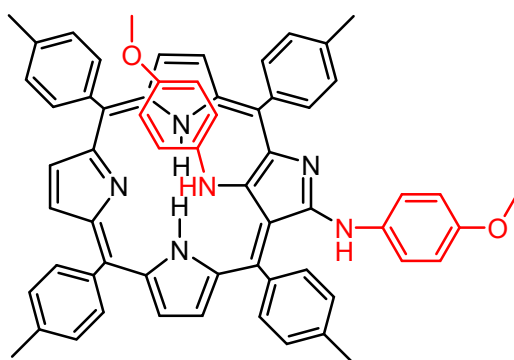


Figure S42. HSQC spectrum of **4a** (CDCl<sub>3</sub>, 298 K).



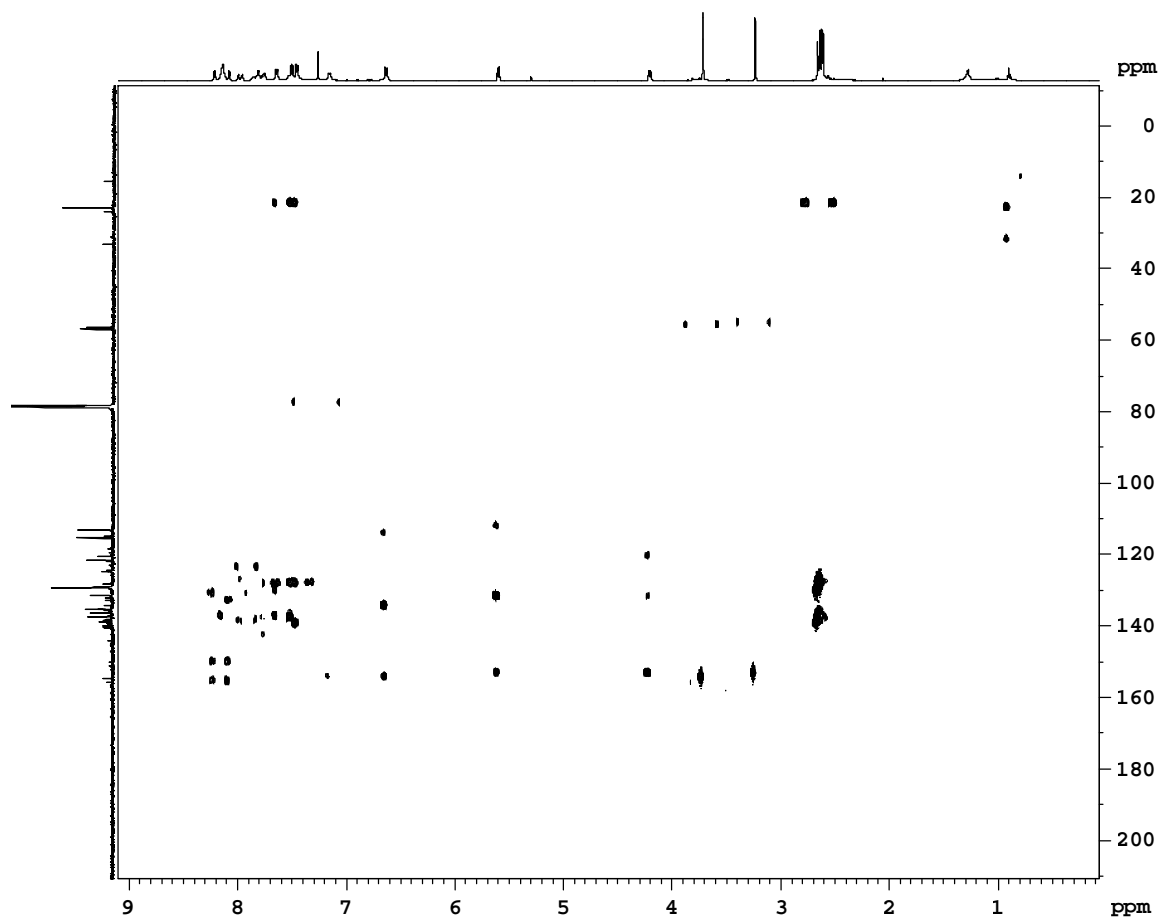
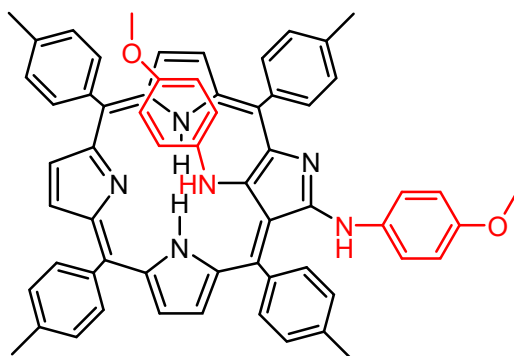


Figure S43. HMBC spectrum of **4a** (CDCl<sub>3</sub>, 298 K).



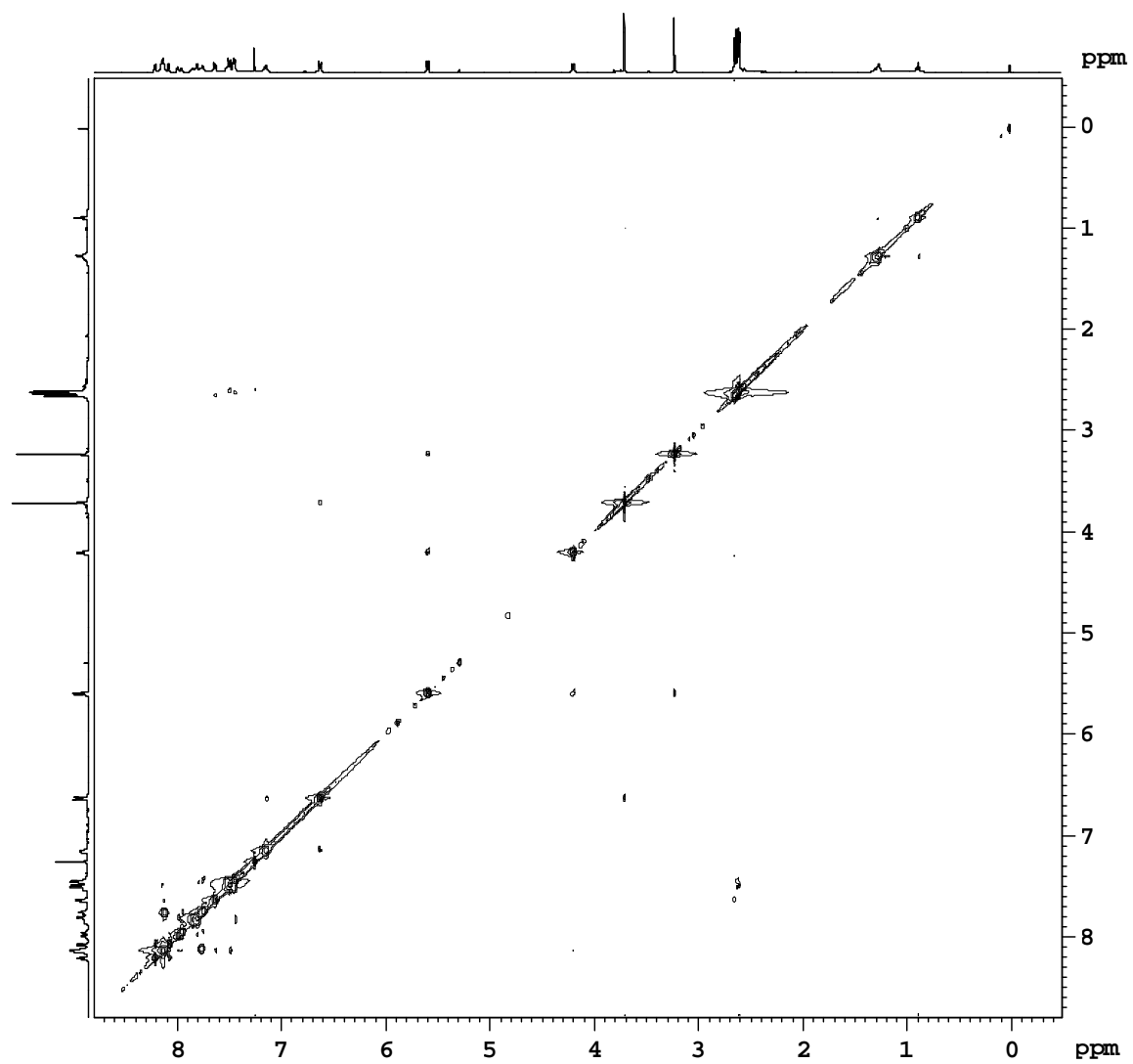
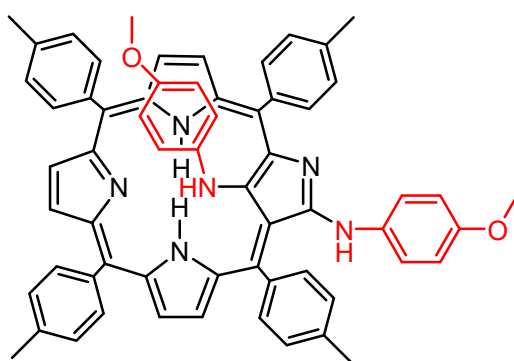


Figure S44. NOESY spectrum of **4a** (CDCl<sub>3</sub>, 298 K).



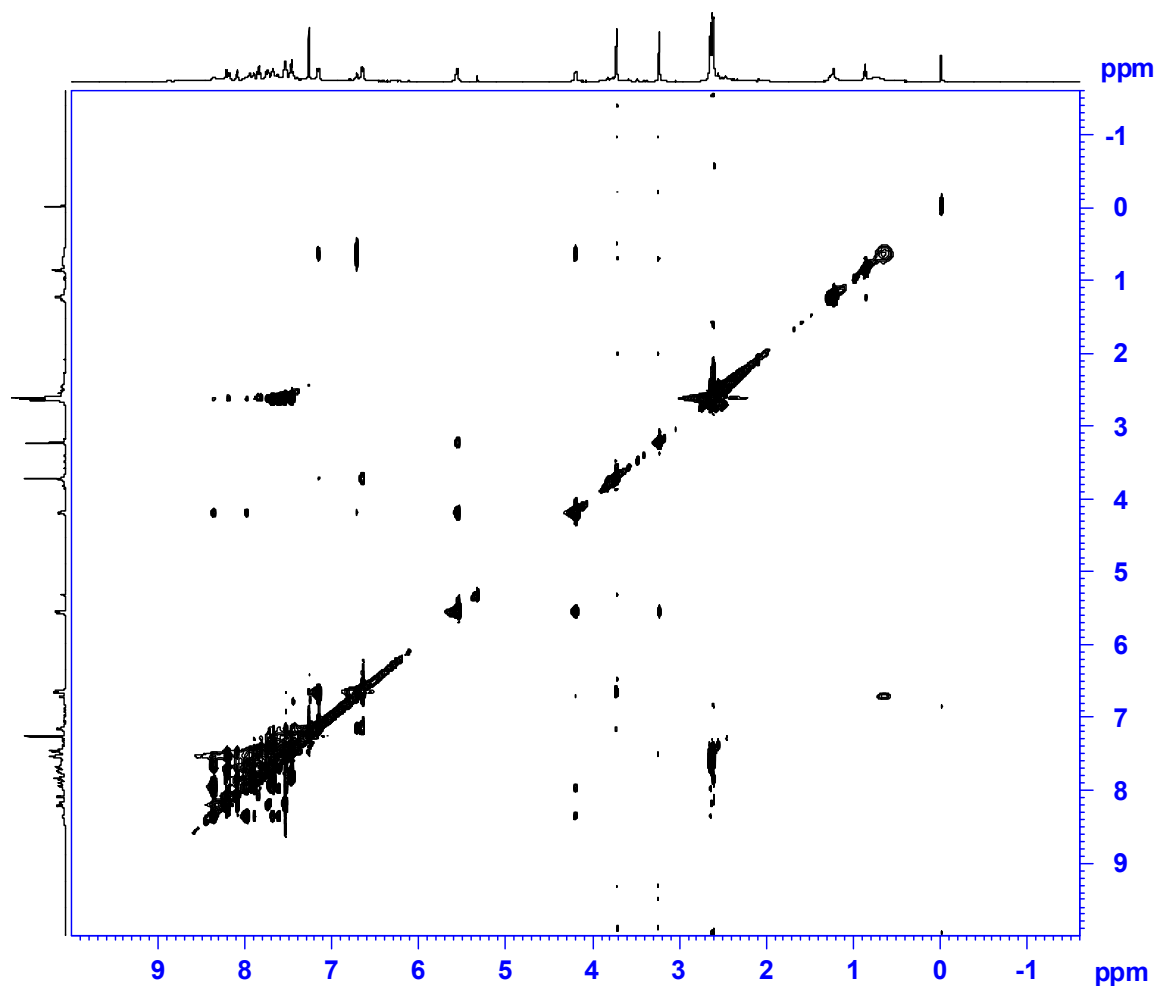
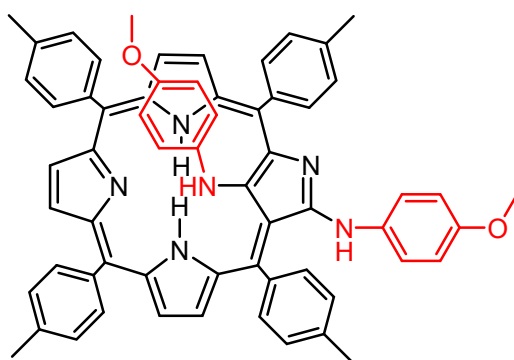


Figure S45. NOESY spectrum of **4a** (CDCl<sub>3</sub>, 230 K).





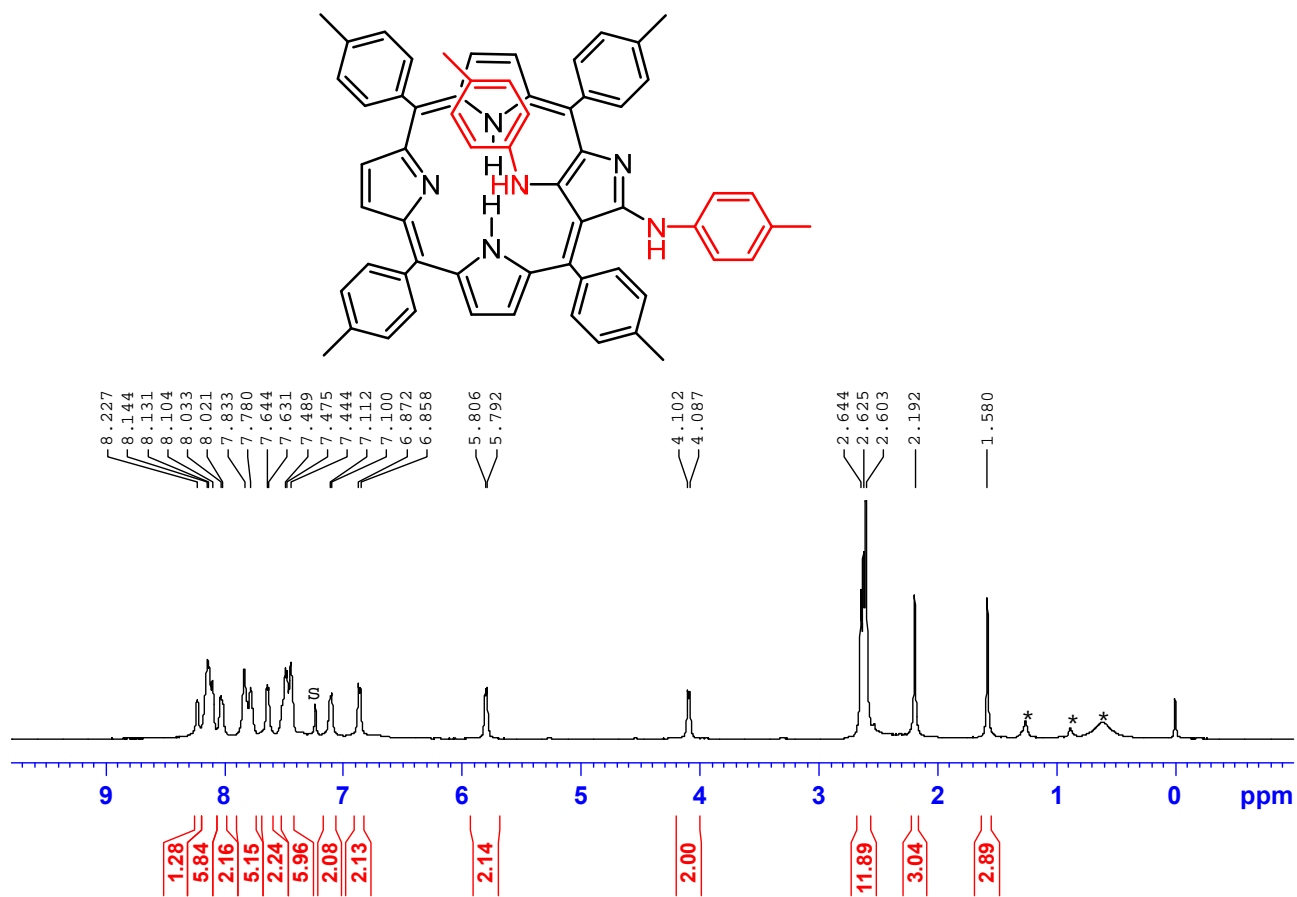


Figure S46. <sup>1</sup>H NMR spectrum of **4b**, 298 K, CDCl<sub>3</sub>.

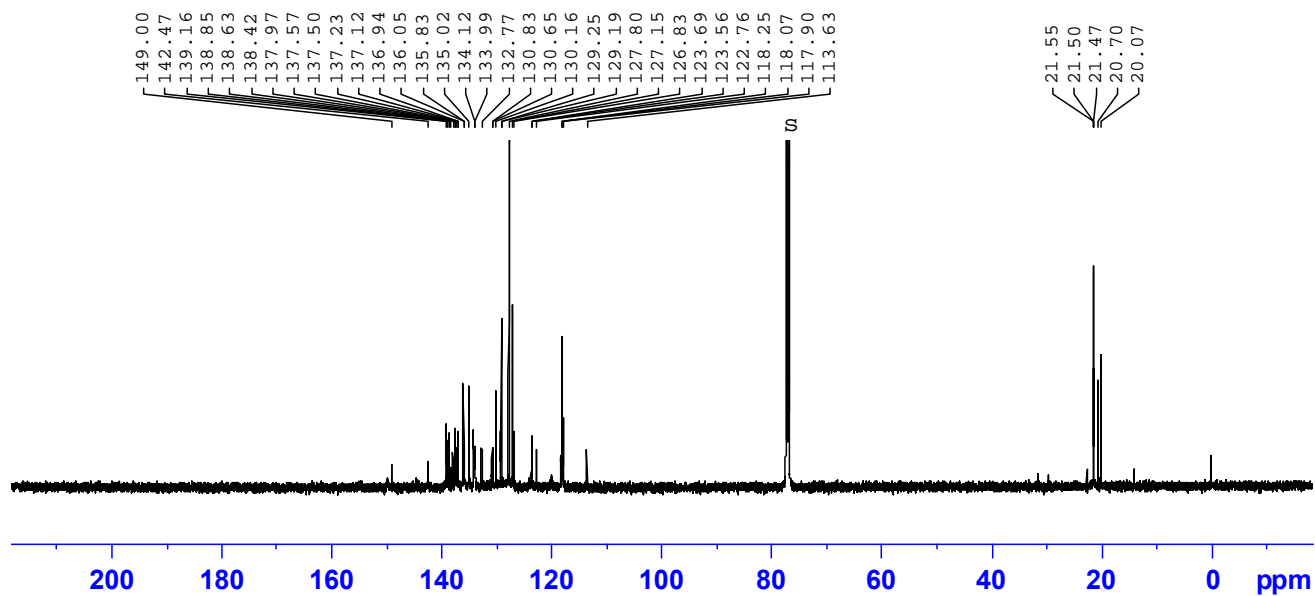


Figure S47. <sup>13</sup>C NMR spectrum of **4b**, 298 K, CDCl<sub>3</sub>.

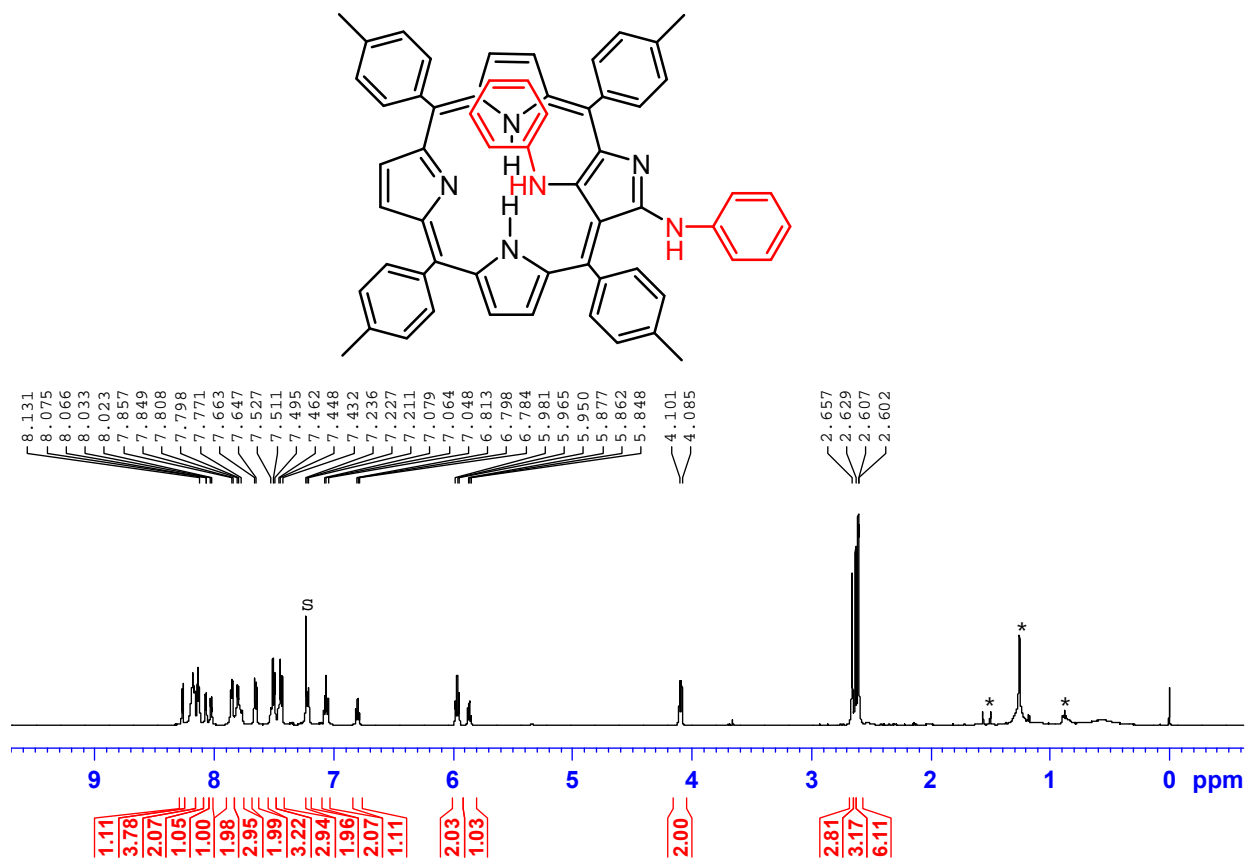


Figure S48.  $^1\text{H}$  NMR spectrum of **4c**, 298 K,  $\text{CDCl}_3$ .

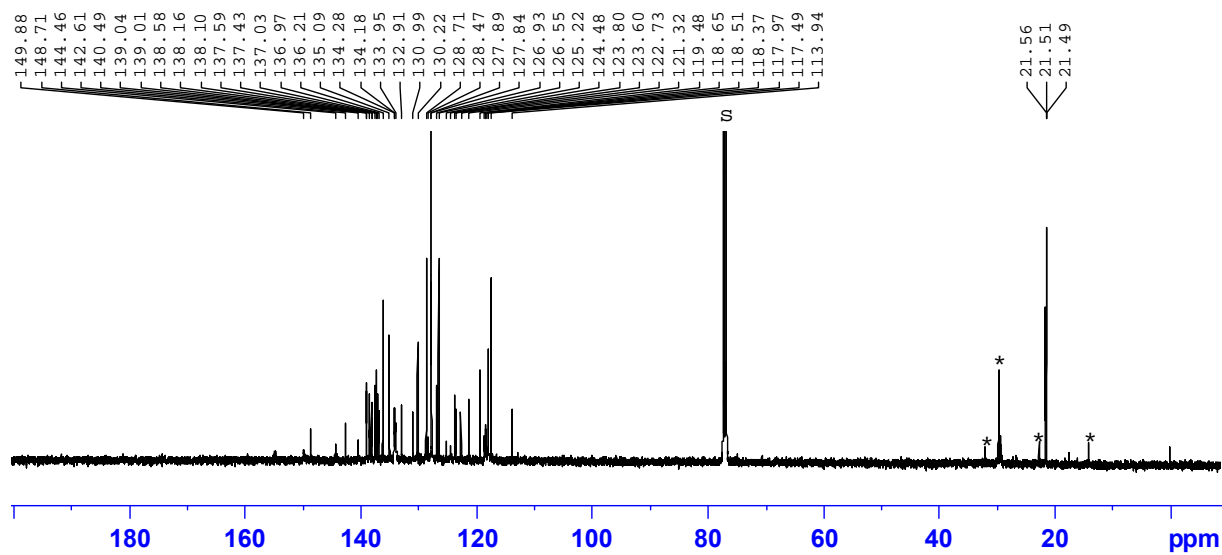


Figure S49.  $^{13}\text{C}$  NMR spectrum of **4c**, 298 K,  $\text{CDCl}_3$ .

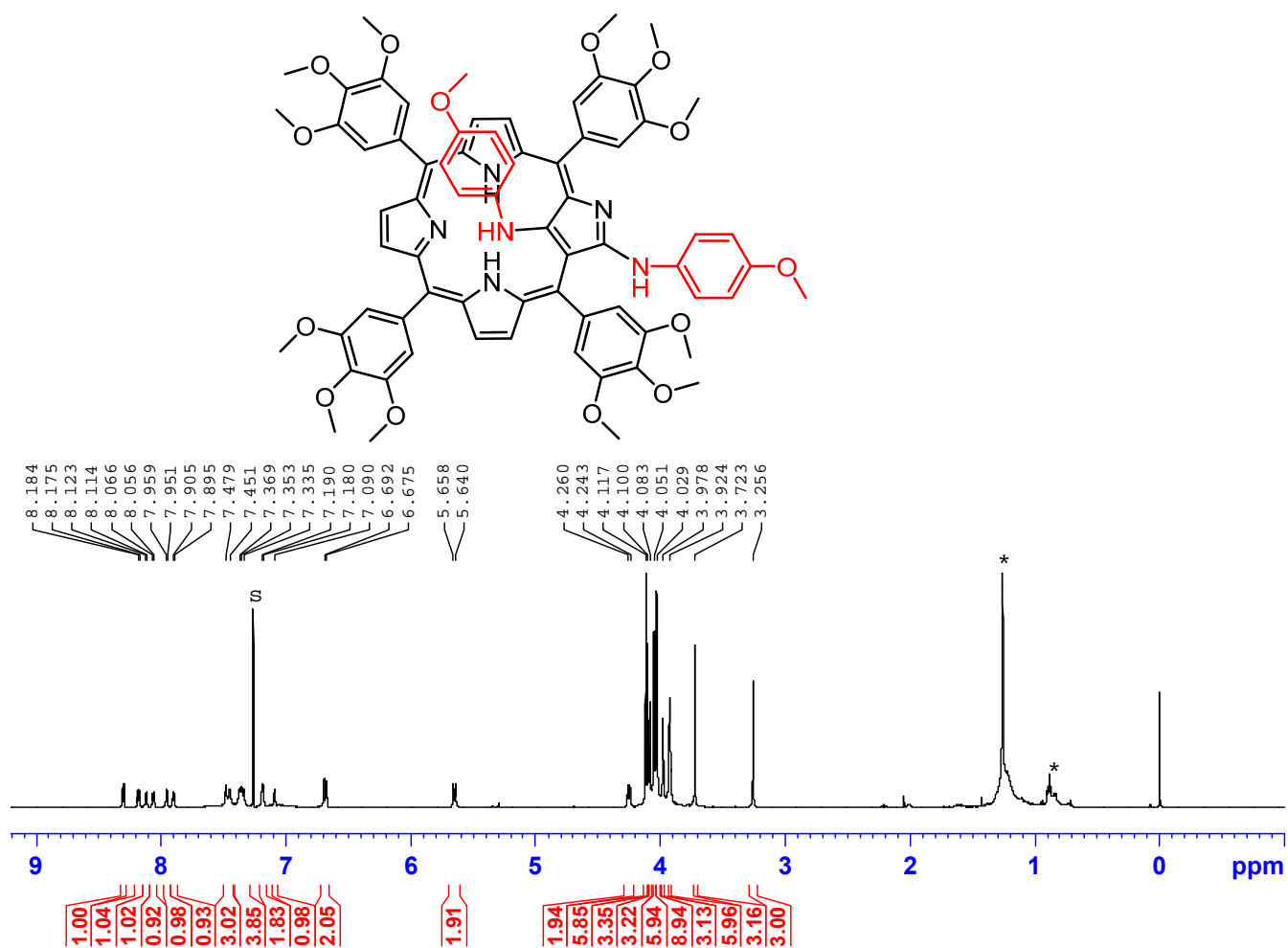


Figure S50. <sup>1</sup>H NMR spectrum of **4d**, 298 K, CDCl<sub>3</sub>.

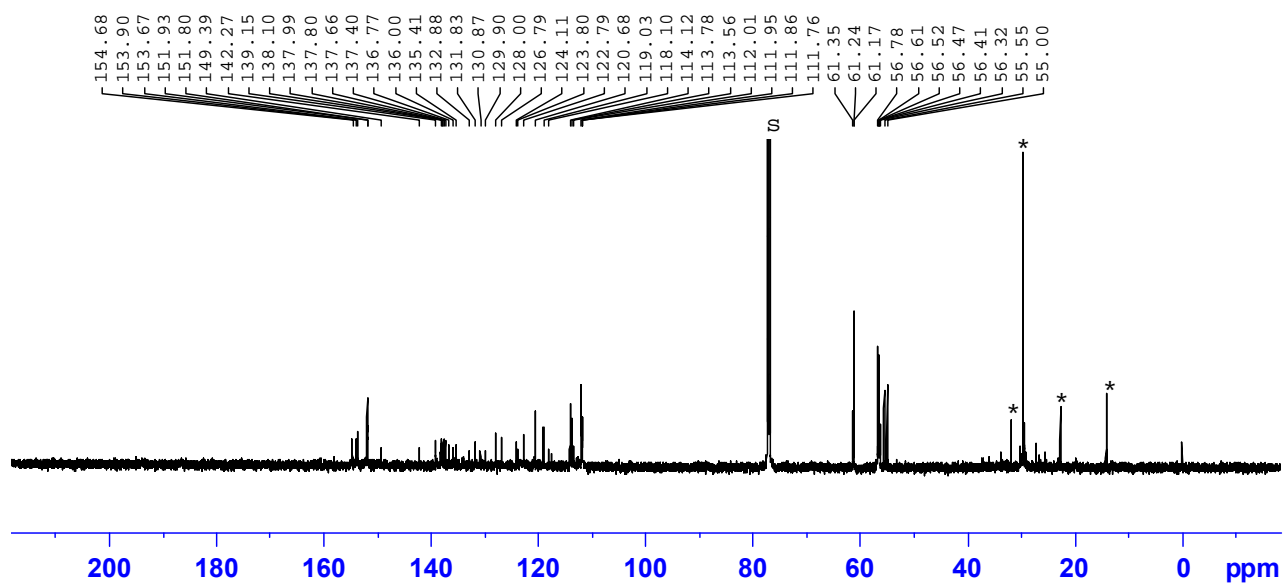


Figure S51. <sup>13</sup>C NMR spectrum of **4d**, 298 K, CDCl<sub>3</sub>.

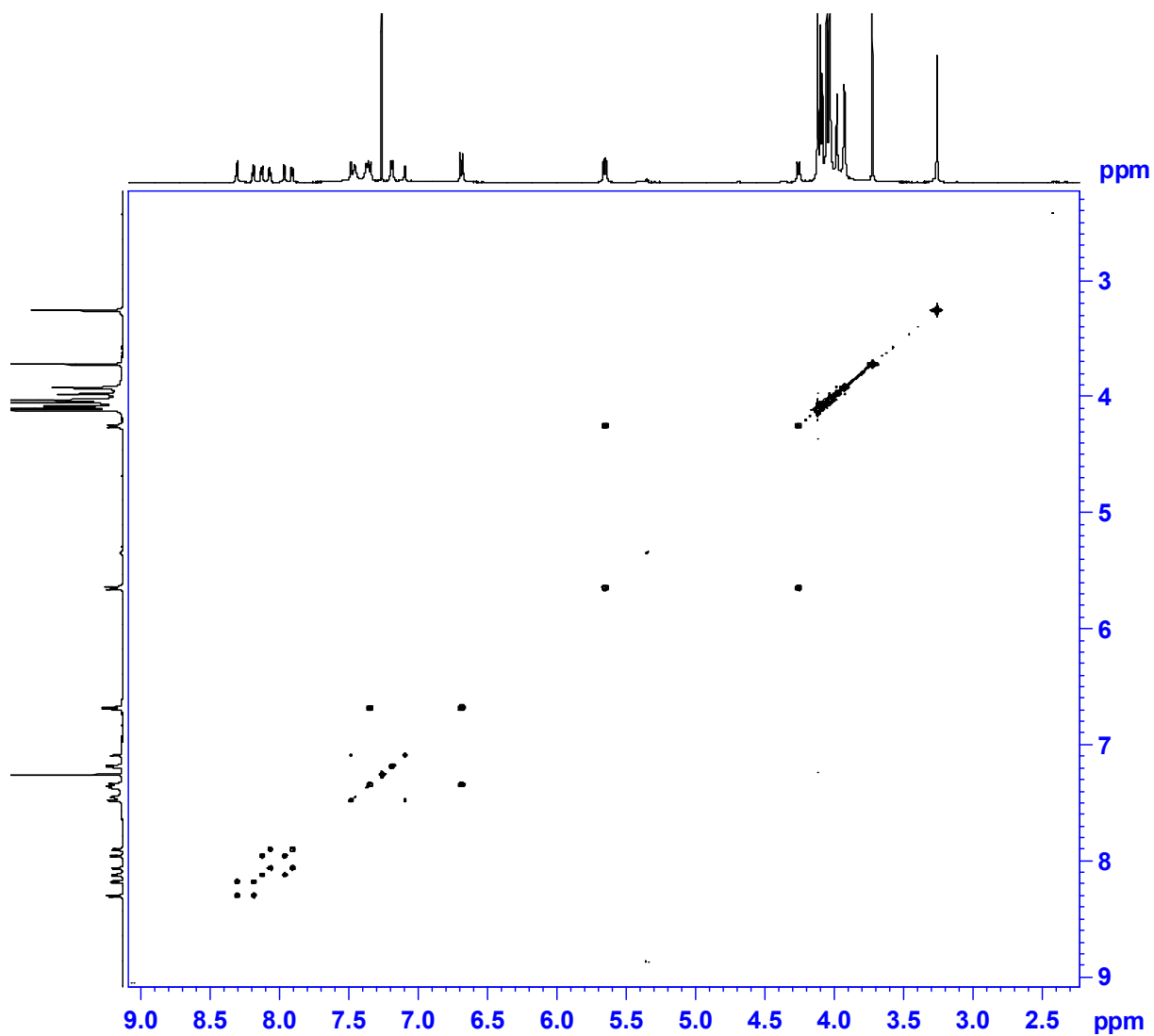
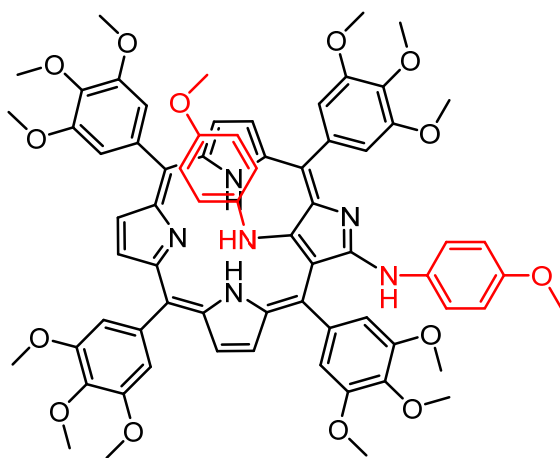


Figure S52. COSY spectrum of 4d (CDCl<sub>3</sub>, 298 K).



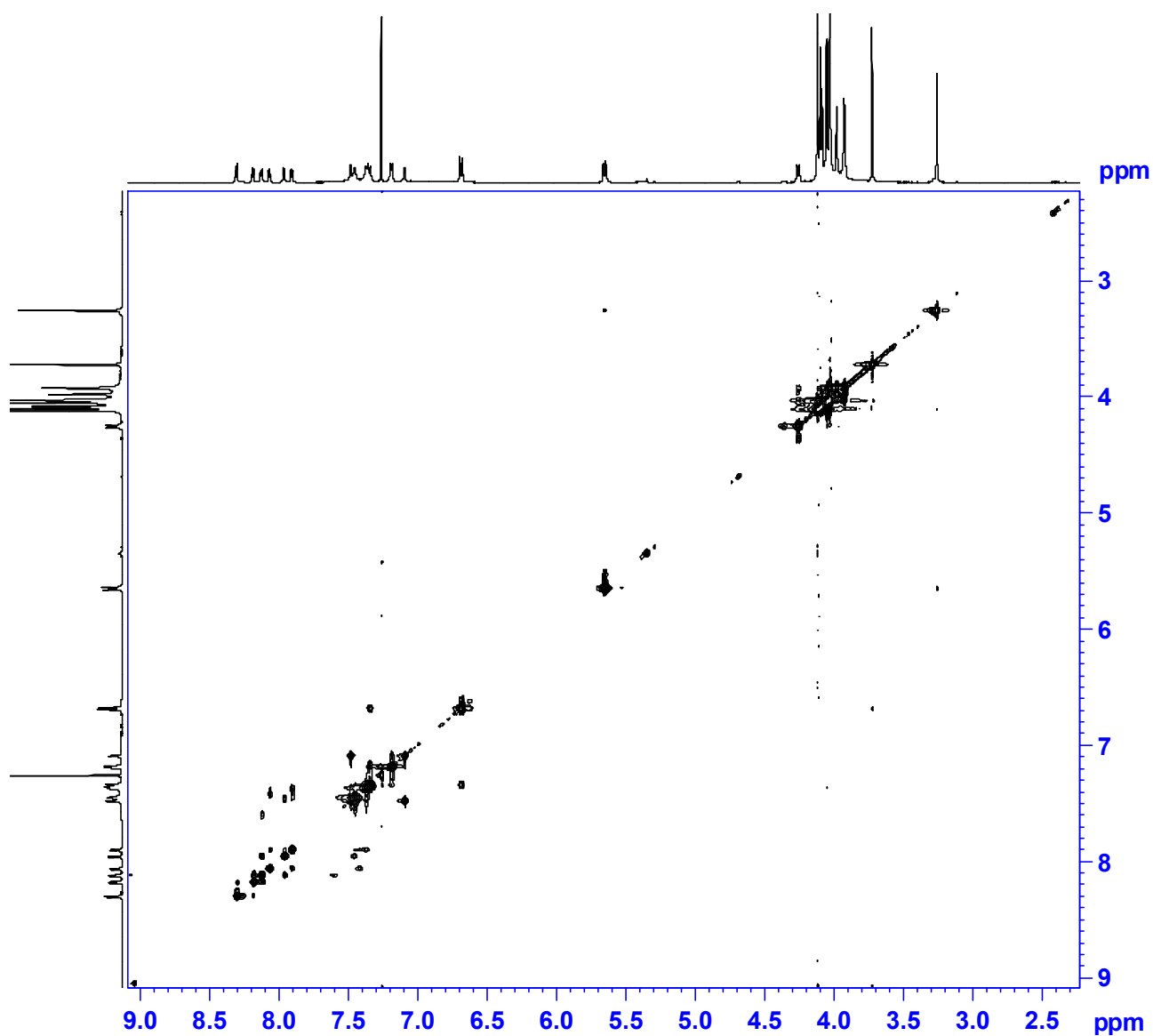
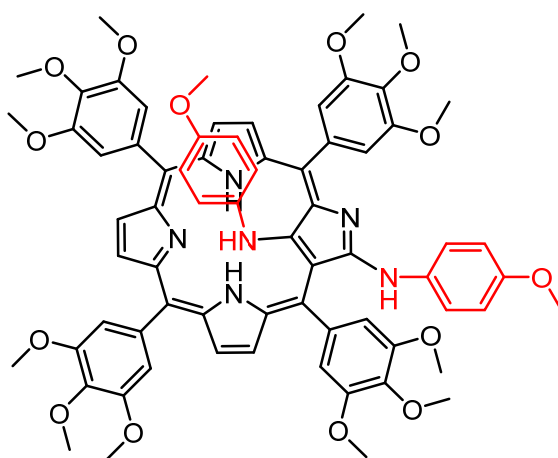


Figure S53. NOESY spectrum of **4d** (CDCl<sub>3</sub>, 298 K).



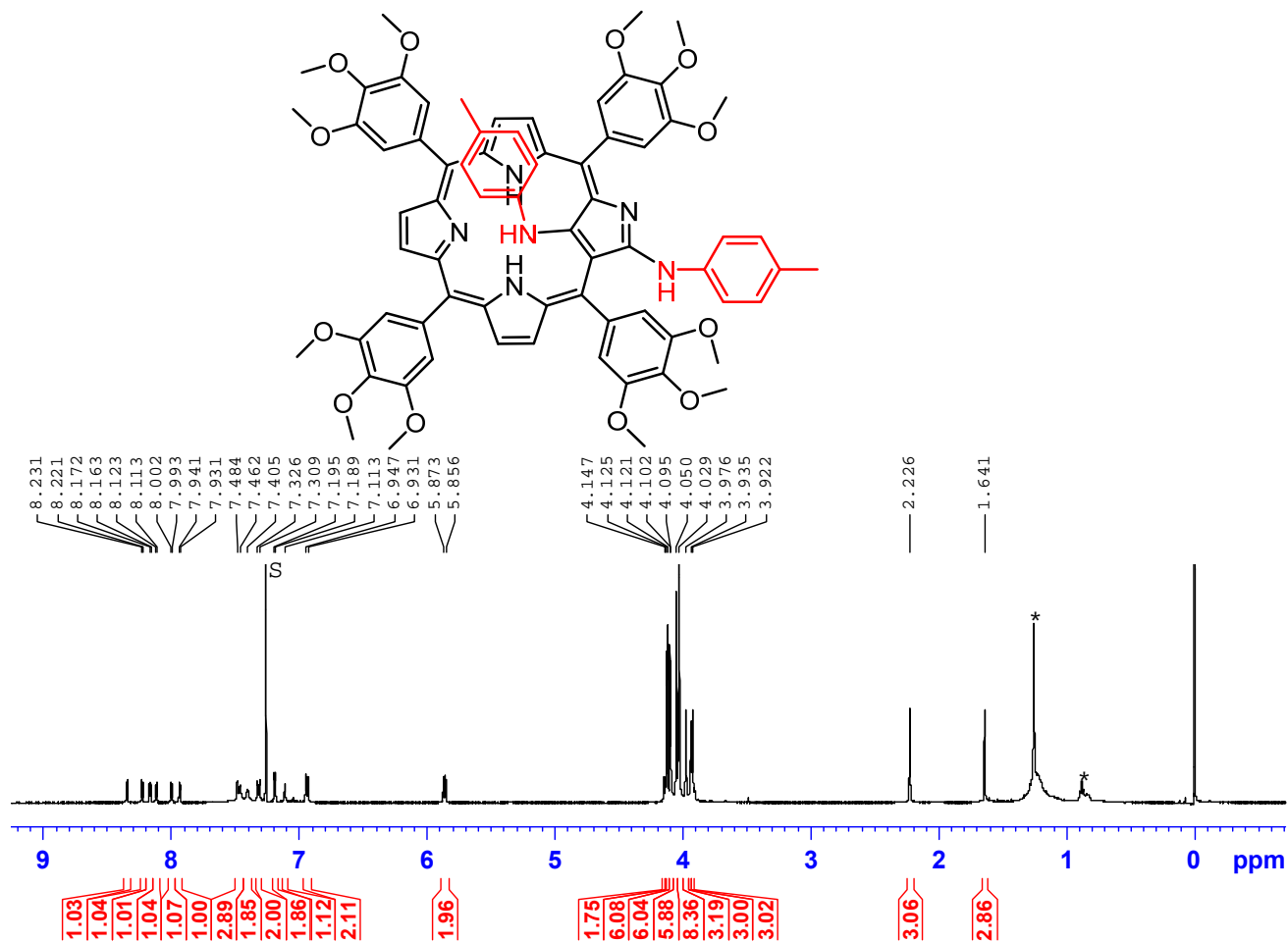


Figure S54. <sup>1</sup>H NMR spectrum of 4e, 298 K, CDCl<sub>3</sub>.

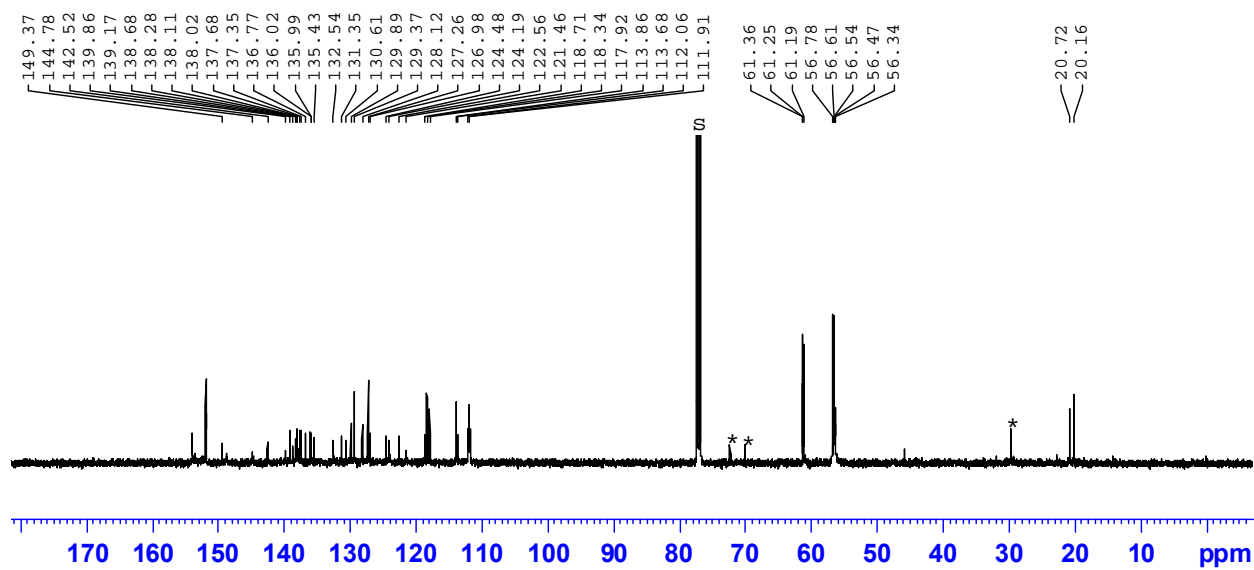


Figure S55. <sup>13</sup>C NMR spectrum of 4e, 298 K, CDCl<sub>3</sub>.

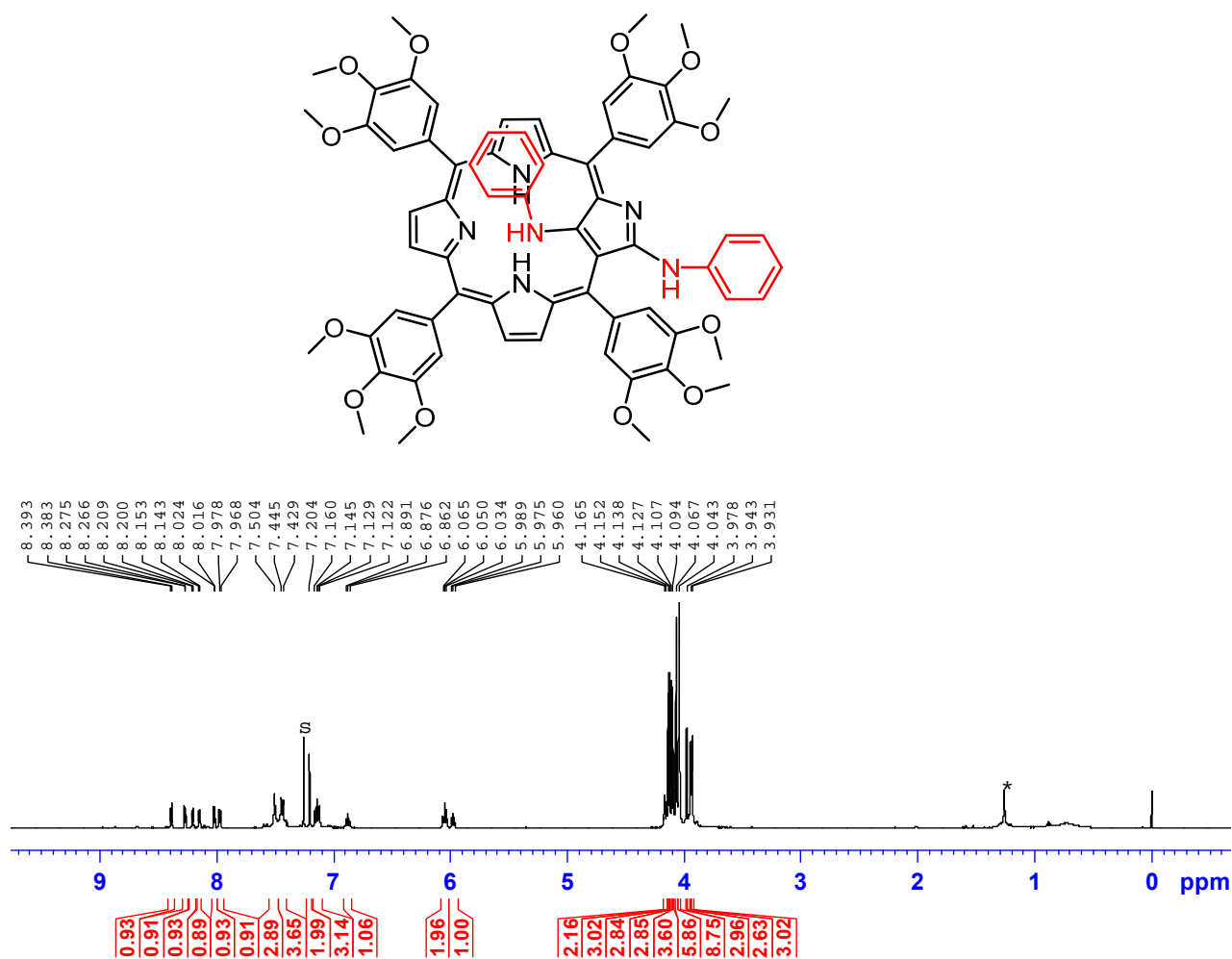


Figure S56.  $^1\text{H}$  NMR spectrum of **4f**, 298 K,  $\text{CDCl}_3$ .

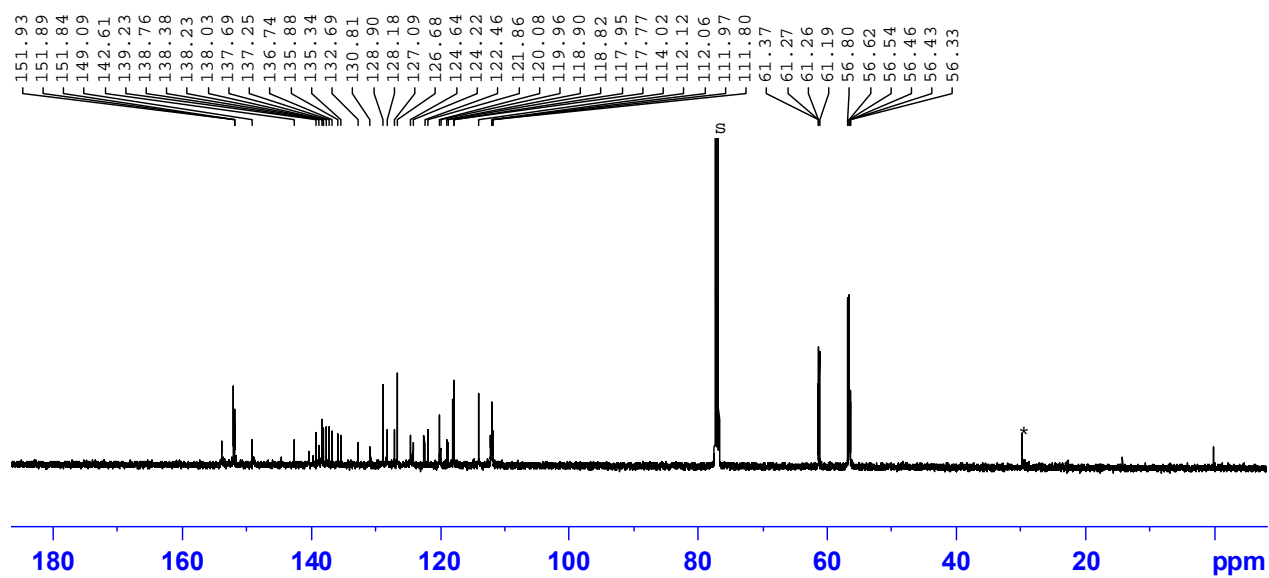


Figure S57.  $^{13}\text{C}$  NMR spectrum of **4f**, 298 K,  $\text{CDCl}_3$ .

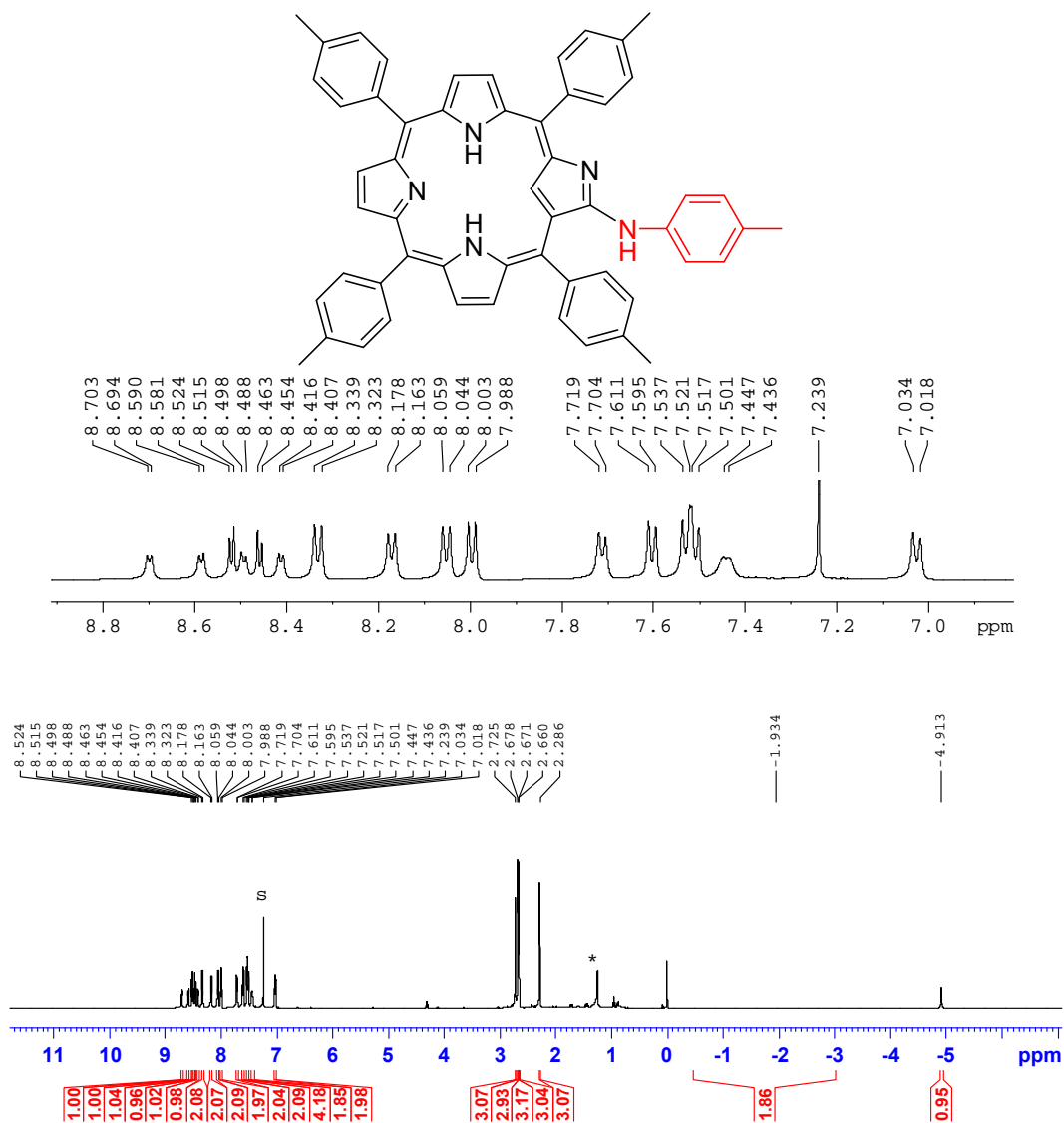


Figure S58.  $^1\text{H}$  NMR spectrum of **5b**, 298 K,  $\text{CDCl}_3$ .

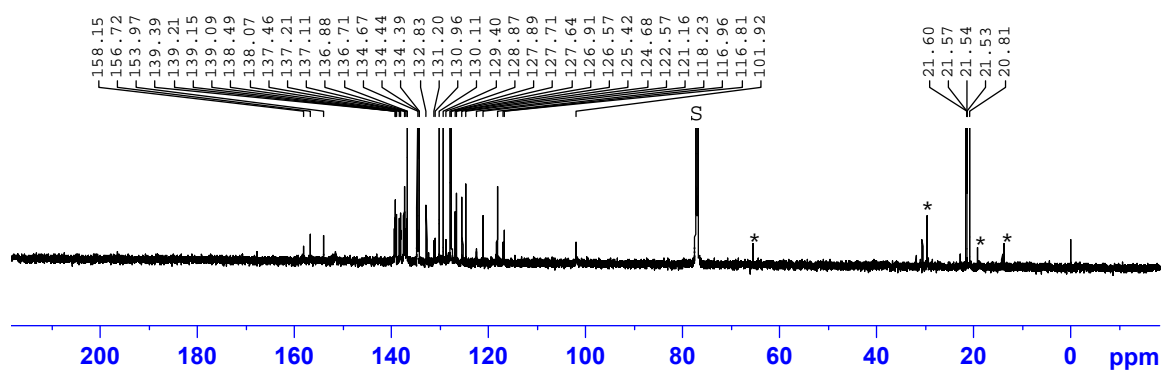


Figure S59.  $^{13}\text{C}$  NMR spectrum of **5b**, 298 K,  $\text{CDCl}_3$ .



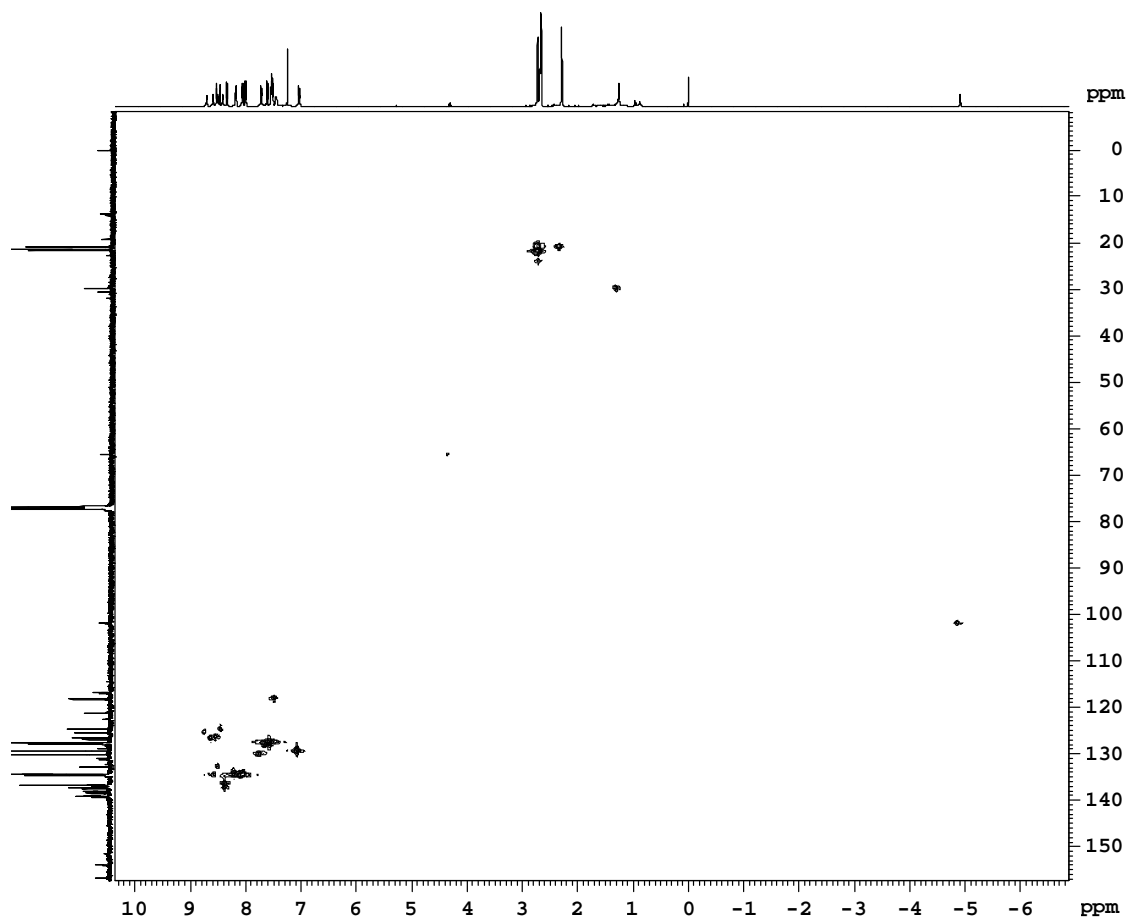
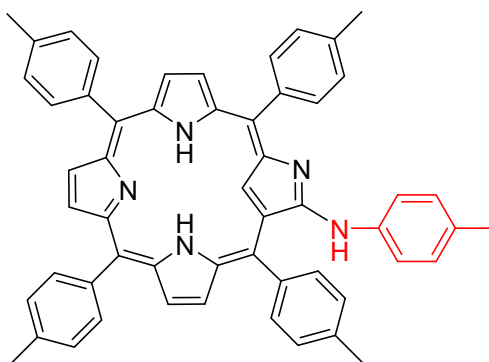


Figure S60. HSQC spectrum of **5b** (CDCl<sub>3</sub>, 298 K).



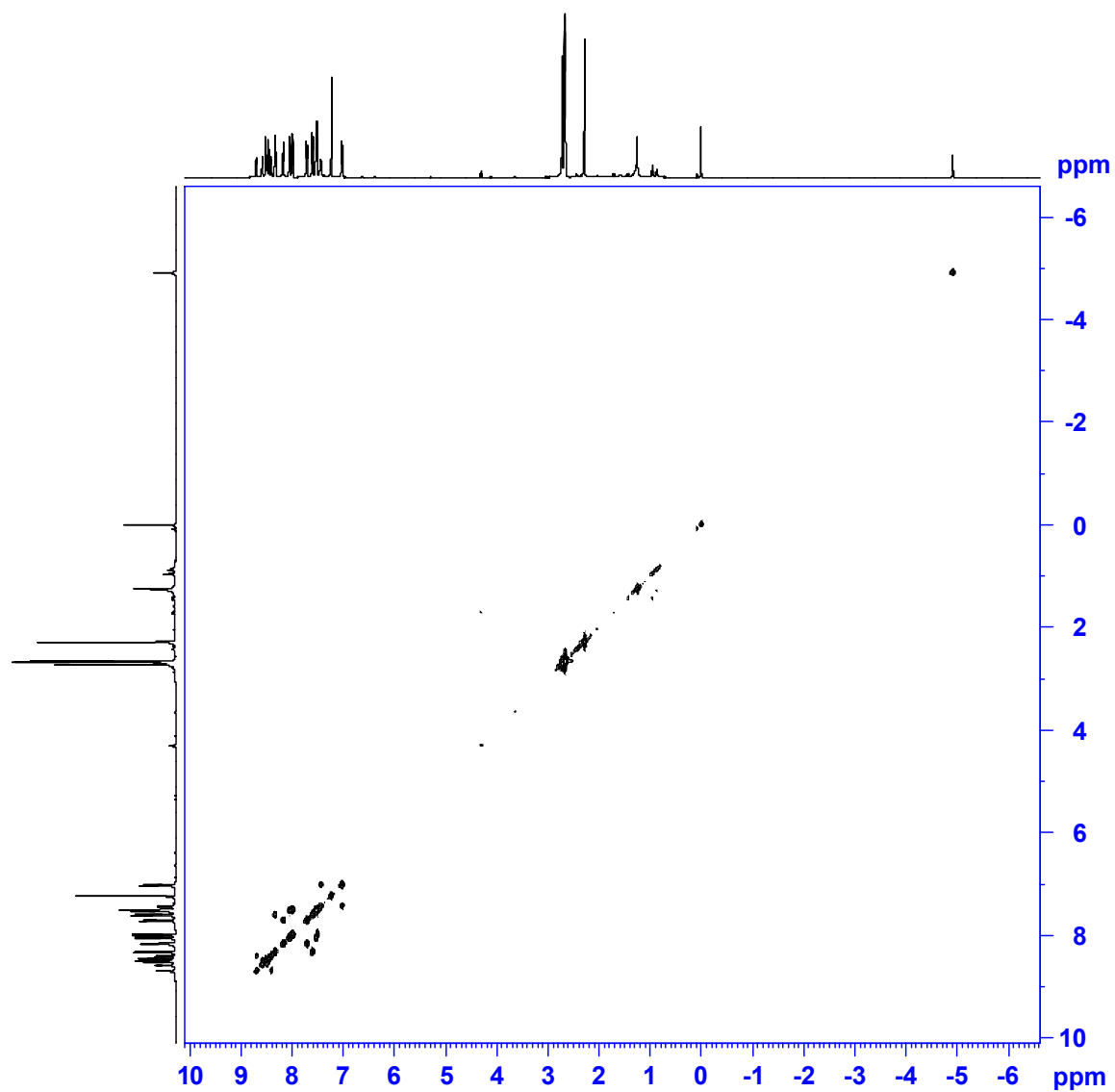
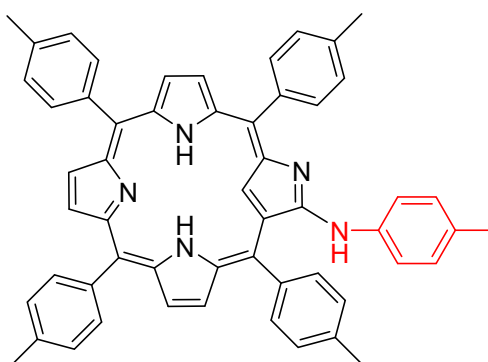


Figure S61. COSY spectrum of **5b** (CDCl<sub>3</sub>, 298 K).



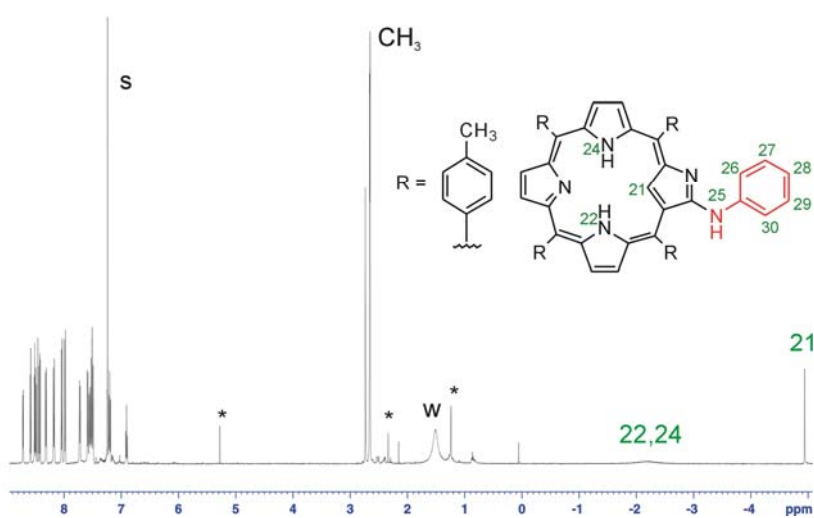
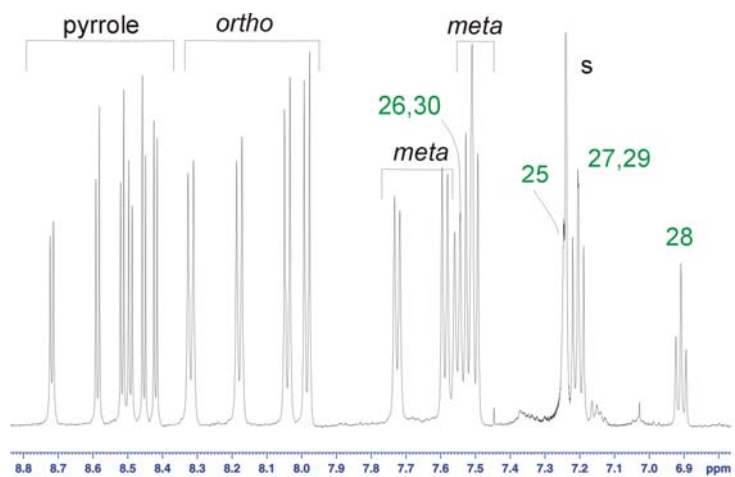


Figure S62.  $^1\text{H}$  NMR spectrum (500 MHz,  $\text{CDCl}_3$ , 300 K) of **5c**.

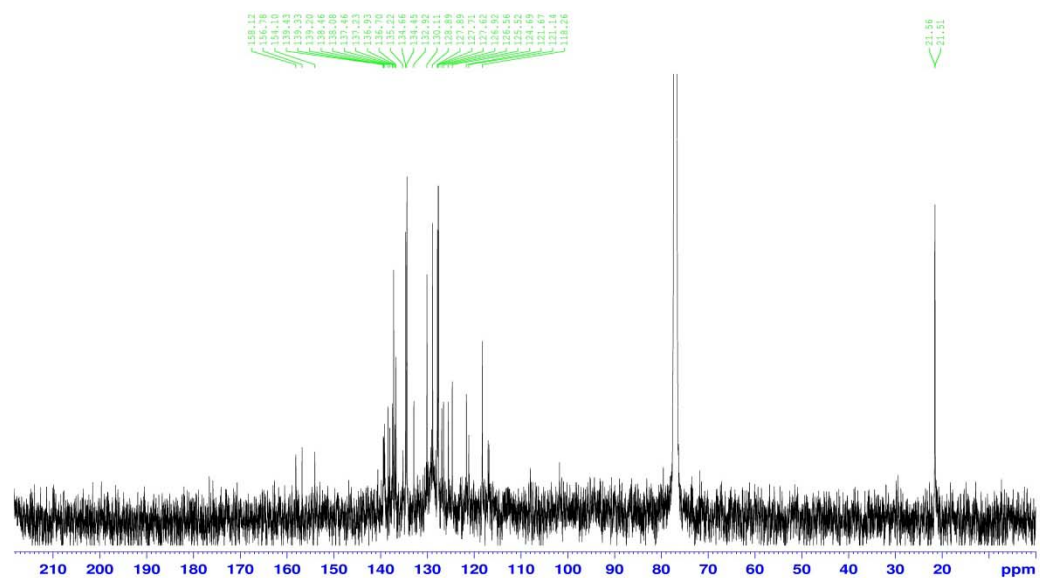


Figure S63.  $^{13}\text{C}$  NMR (126 MHz, 300 K,  $\text{CDCl}_3$ ) for **5c**.

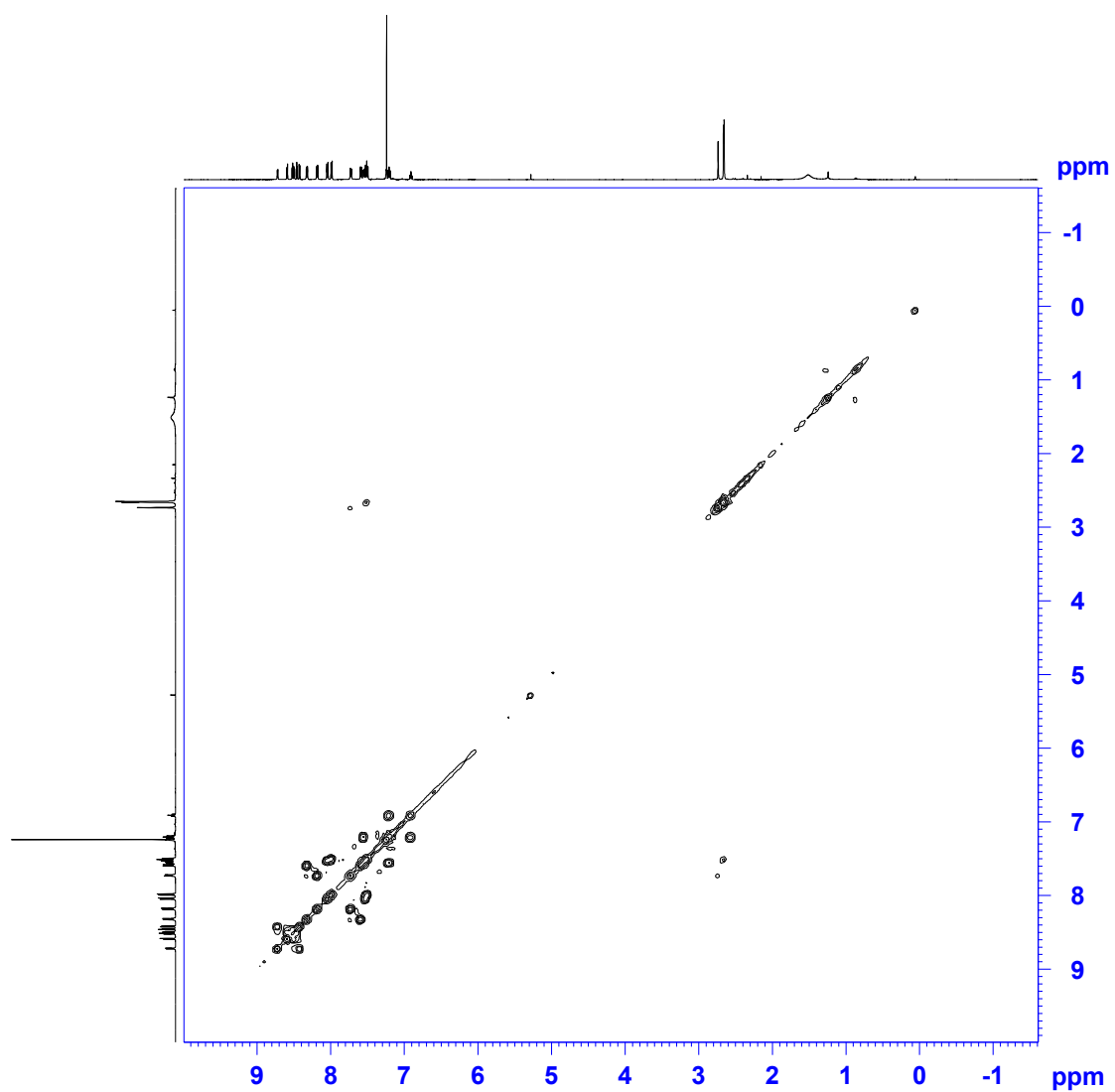
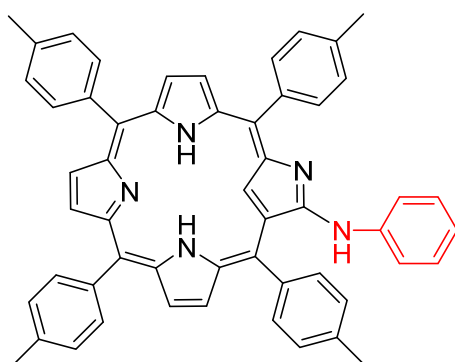


Figure S64. COSY spectrum (600 MHz, 300 K CDCl<sub>3</sub>) of **5c**.



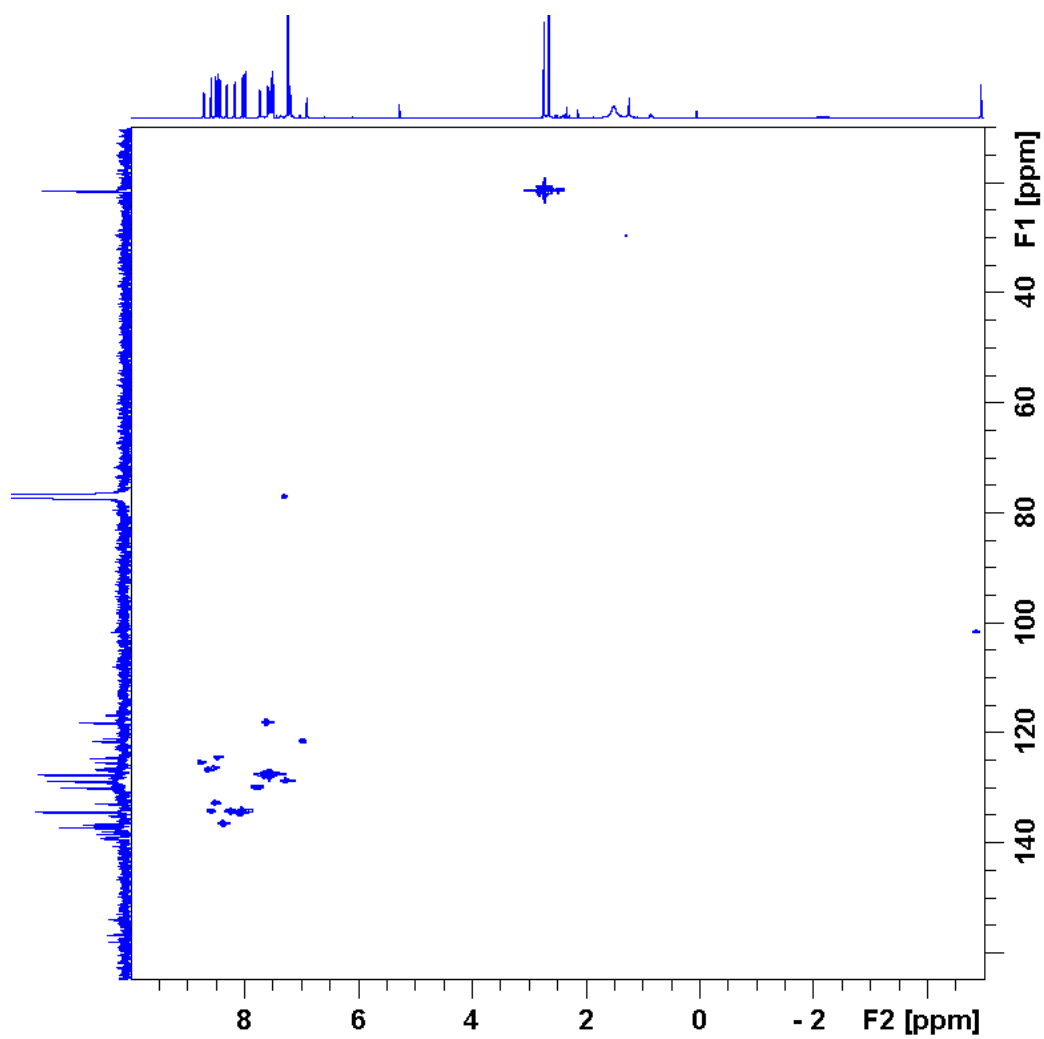
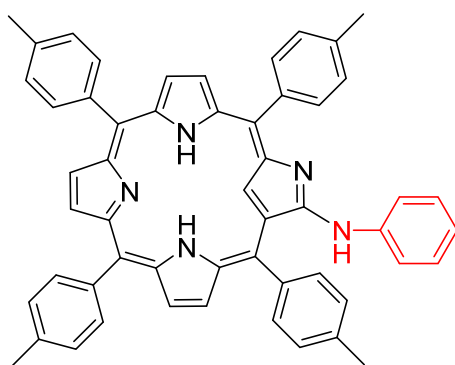
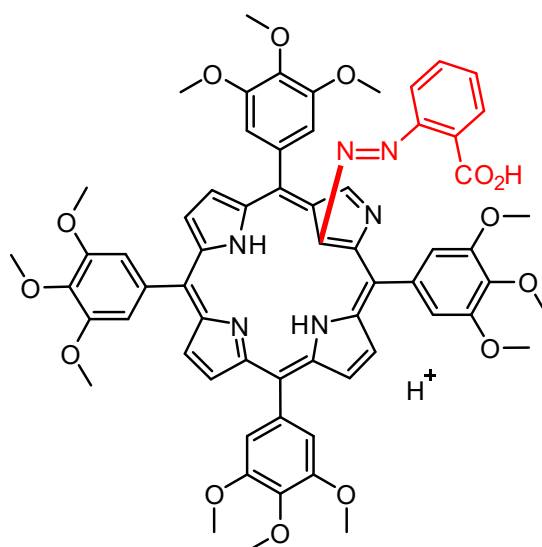


Figure S65. HSQC spectrum (300 K CDCl<sub>3</sub>) of 5c.





Chemical Formula: C<sub>63</sub>H<sub>59</sub>N<sub>6</sub>O<sub>14</sub><sup>+</sup>  
Exact Mass: 1123.4084

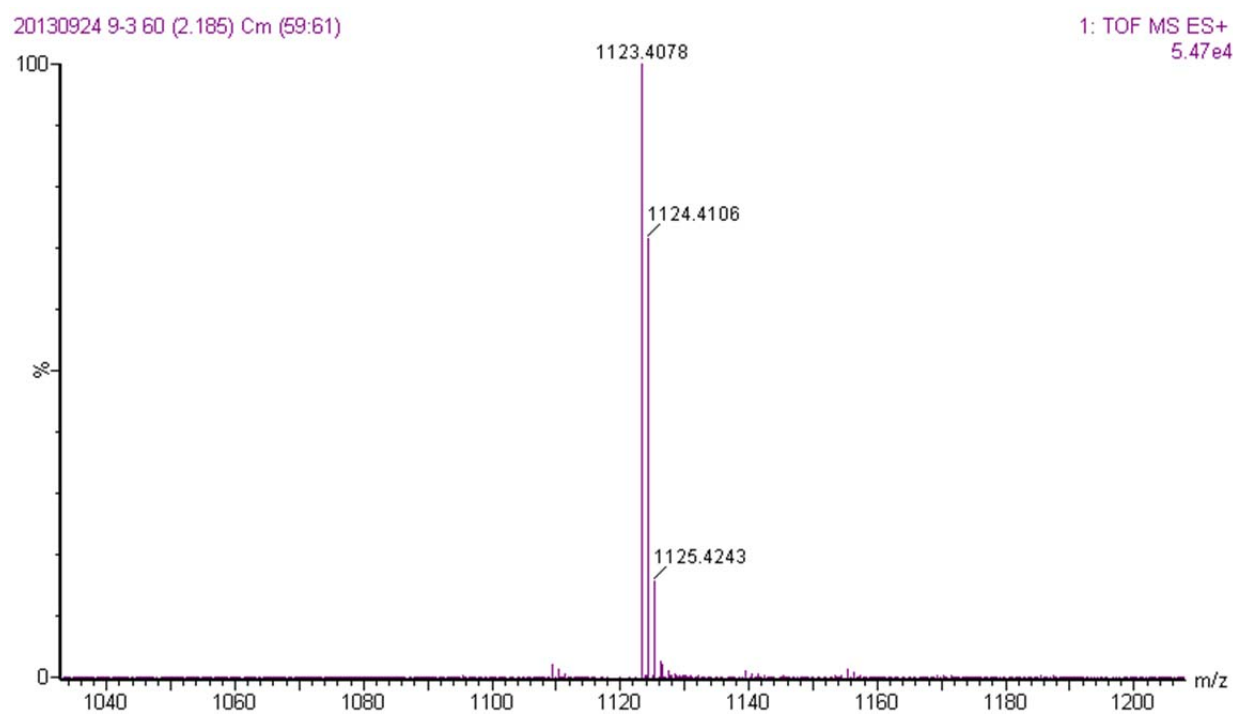
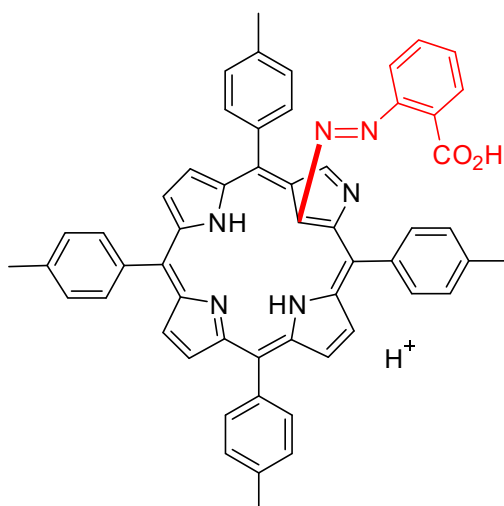


Figure S66. HRMS spectrum of 2a.



Chemical Formula:  $C_{55}H_{43}N_6O_2^+$   
Exact Mass: 819.3442

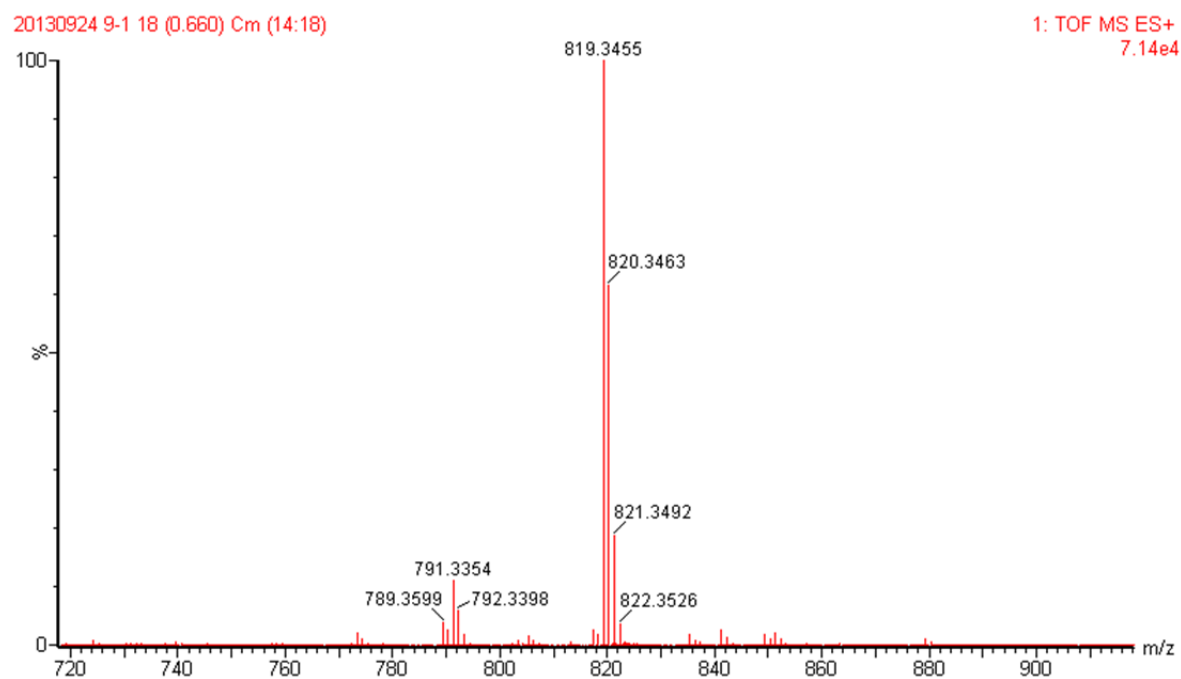
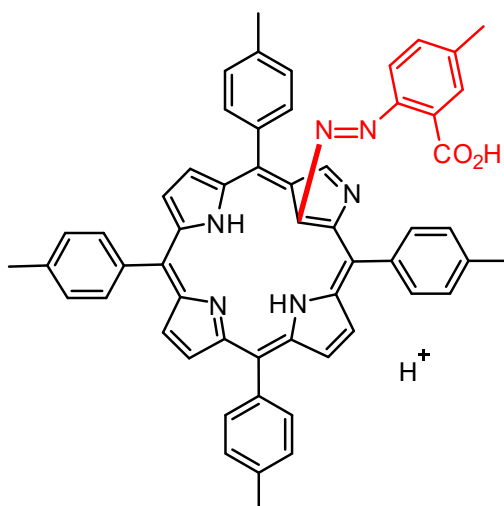


Figure S67. HRMS spectrum of **2b**.



Chemical Formula:  $C_{56}H_{45}N_6O_2^+$   
Exact Mass: 833.3599

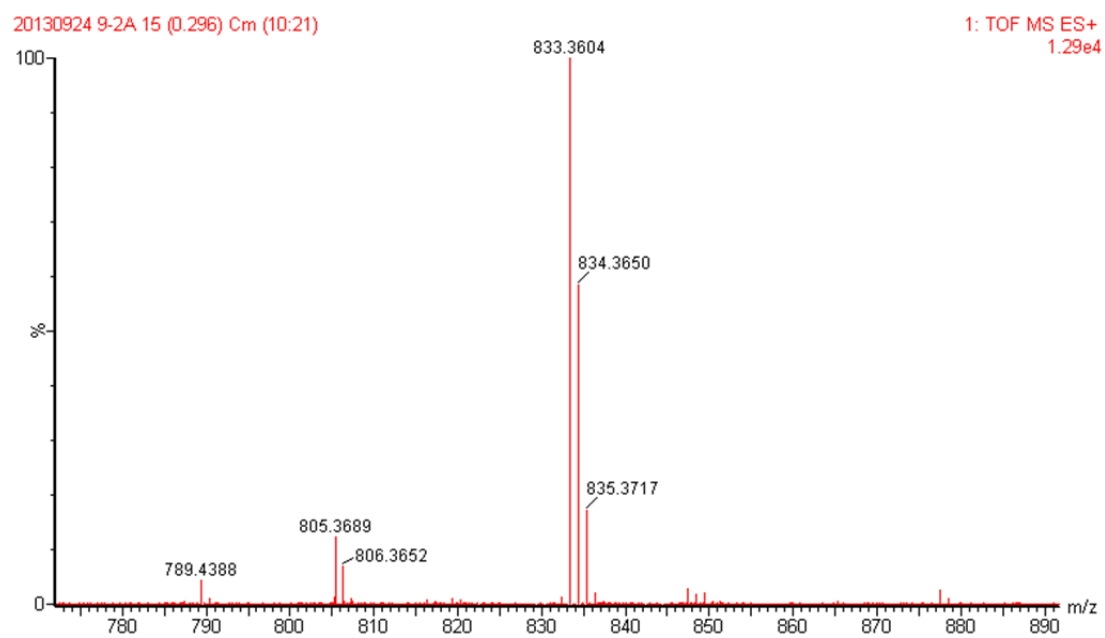
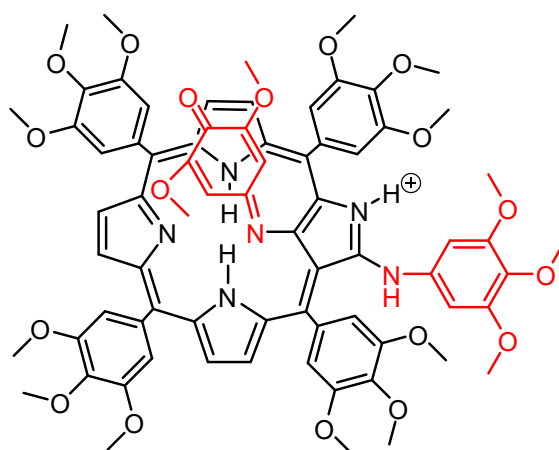


Figure S68. HRMS spectrum of **2c**.





Chemical Formula:  $C_{73}H_{73}N_6O_{18}^+$   
Exact Mass: 1321.4976

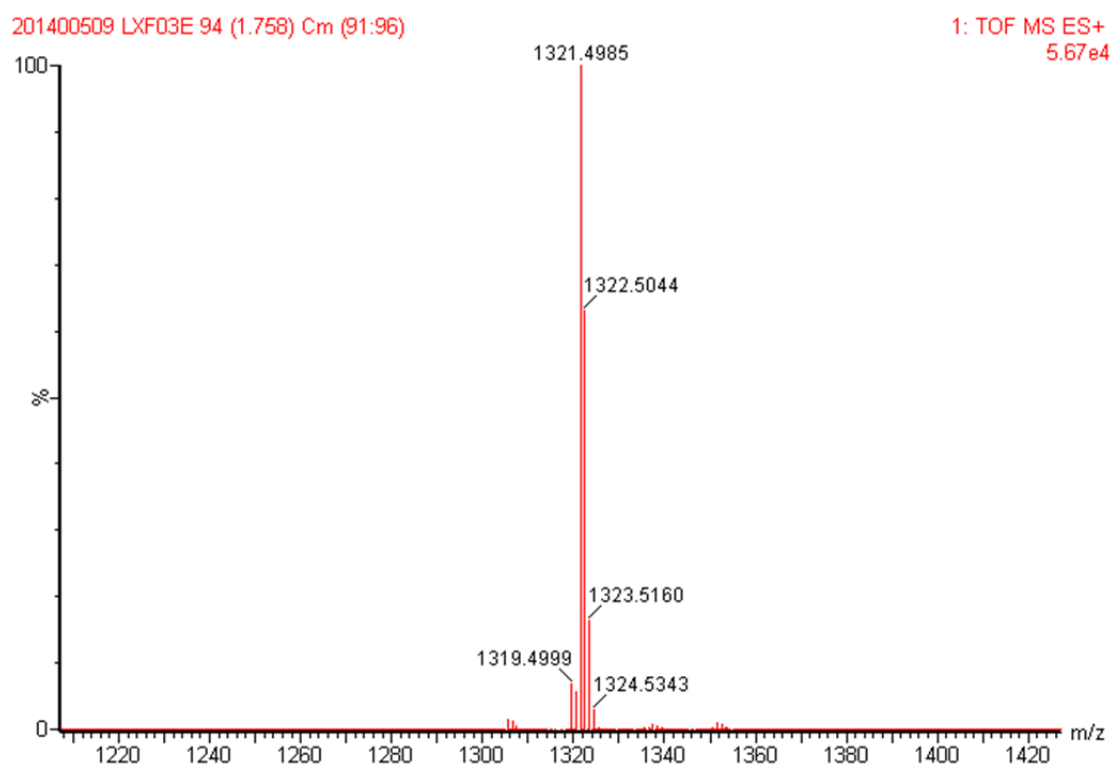
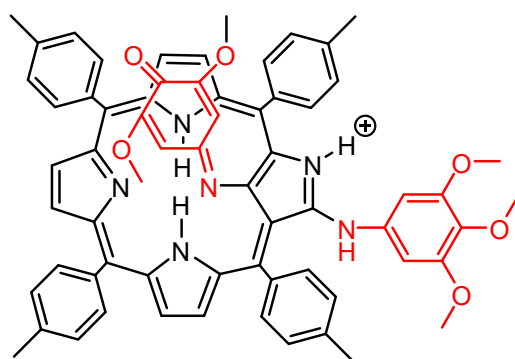


Figure S69. HRMS spectrum of **3a**.



Chemical Formula:  $C_{65}H_{57}N_6O_6^+$   
Exact Mass: 1017.4334

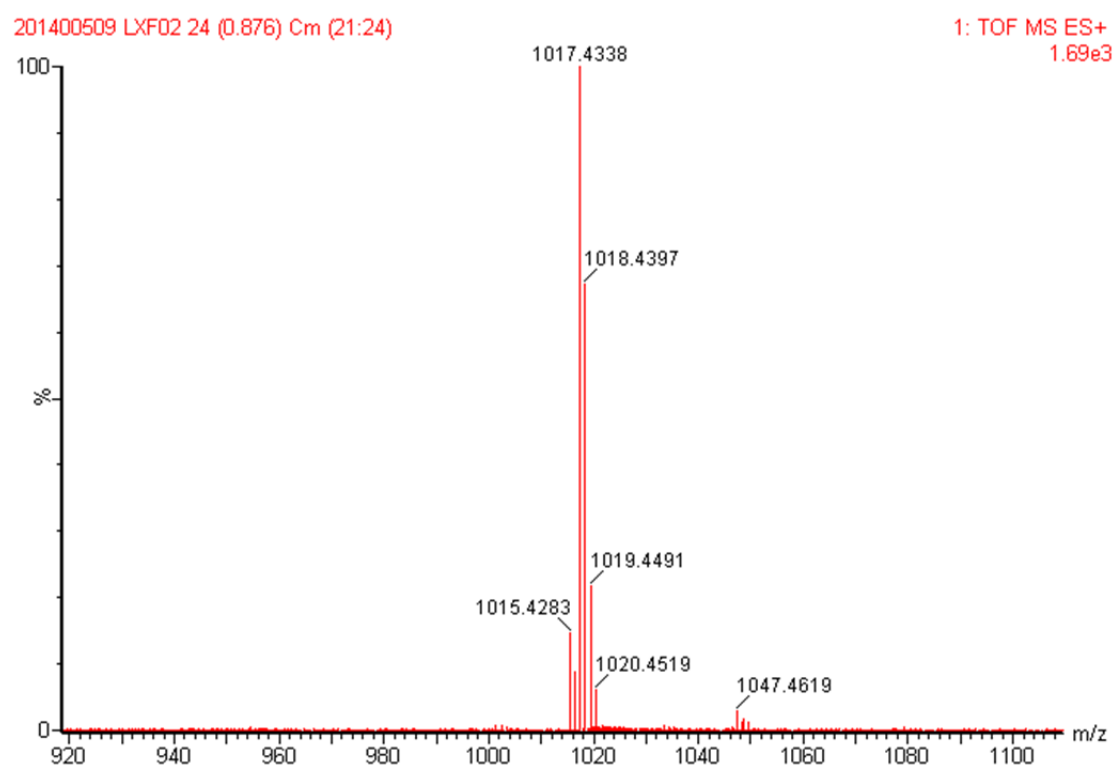
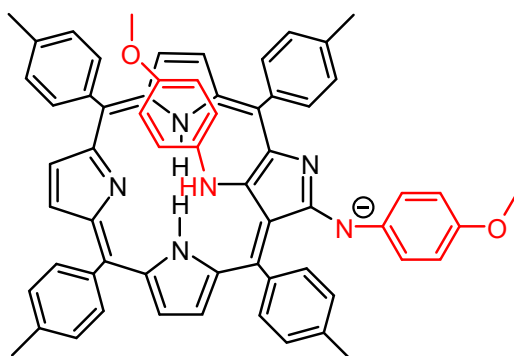


Figure S70. HRMS spectrum of **3b**.



Chemical Formula:  $C_{62}H_{51}N_6O_2^-$

Exact Mass: 911.4079

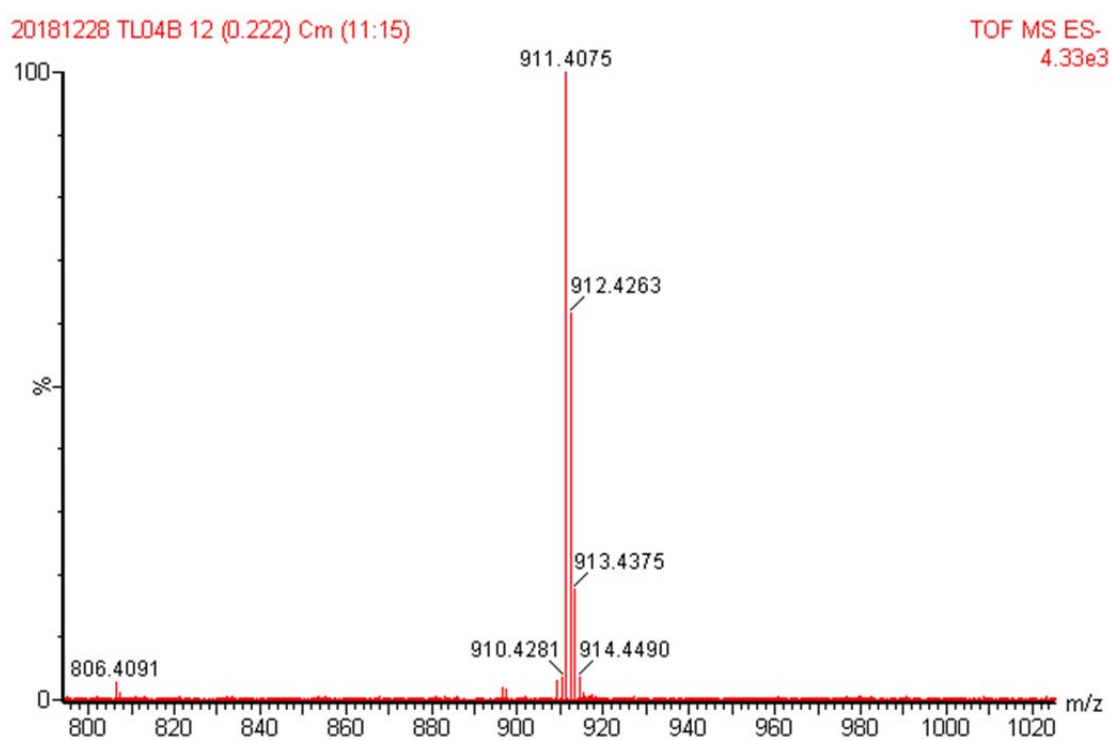
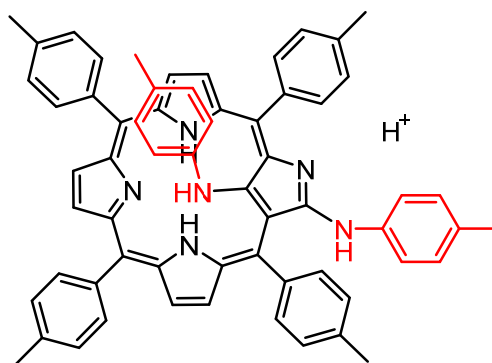


Figure S71. HRMS spectrum of 4a.



$C_{62}H_{53}N_6$   
Exact Mass: 881.4332

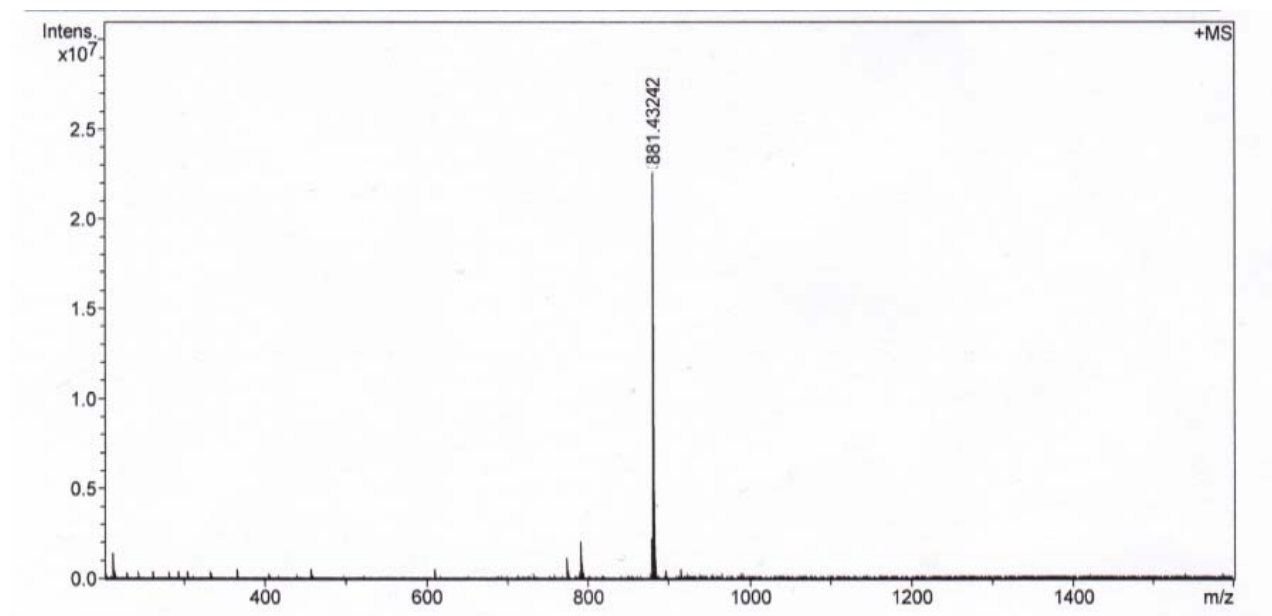
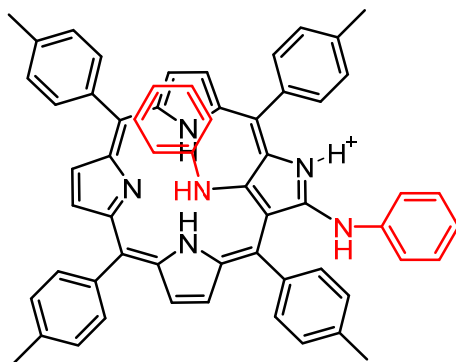


Figure S72. HRMS spectrum of **4b**.



Chemical Formula: C<sub>60</sub>H<sub>49</sub>N<sub>6</sub><sup>+</sup>  
Exact Mass: 853.4013

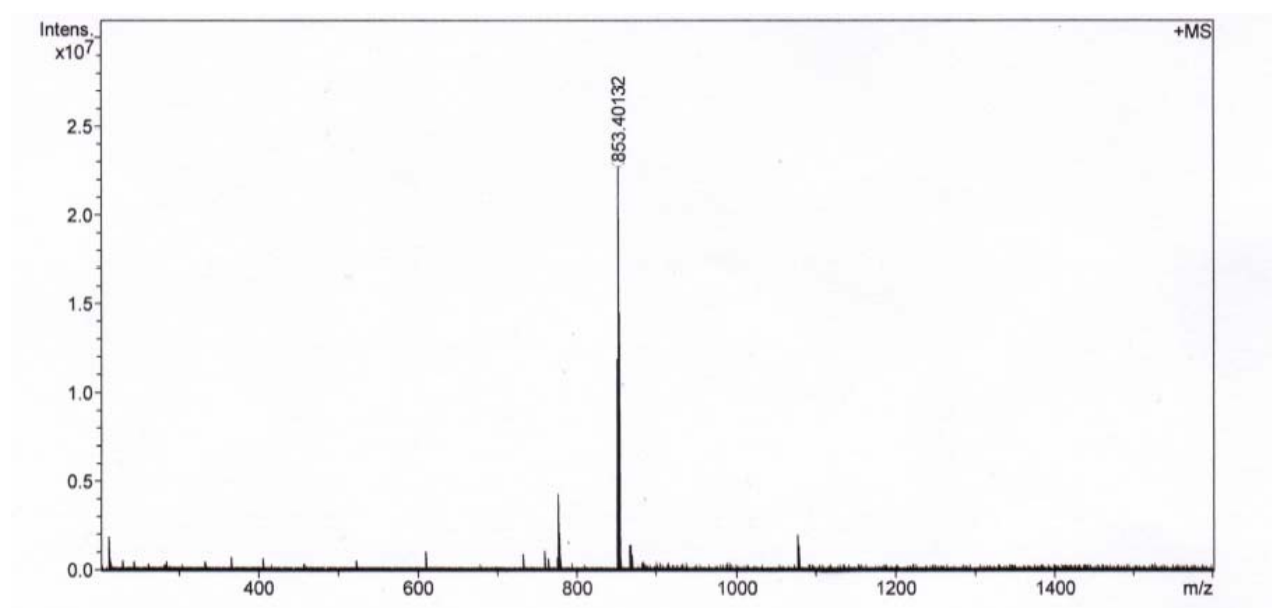
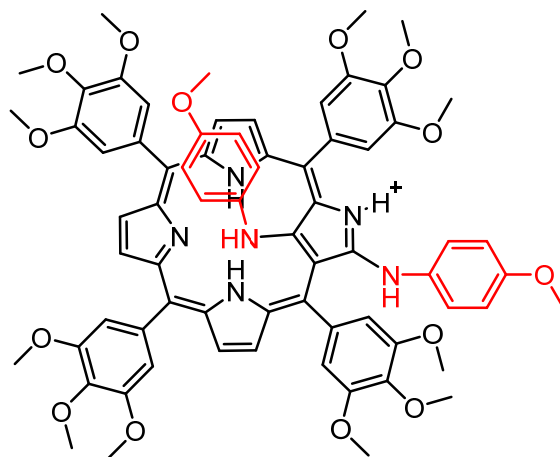


Figure S73. HRMS spectrum of 4c.



Chemical Formula:  $C_{70}H_{69}N_6O_{14}^+$   
Exact Mass: 1217.4866

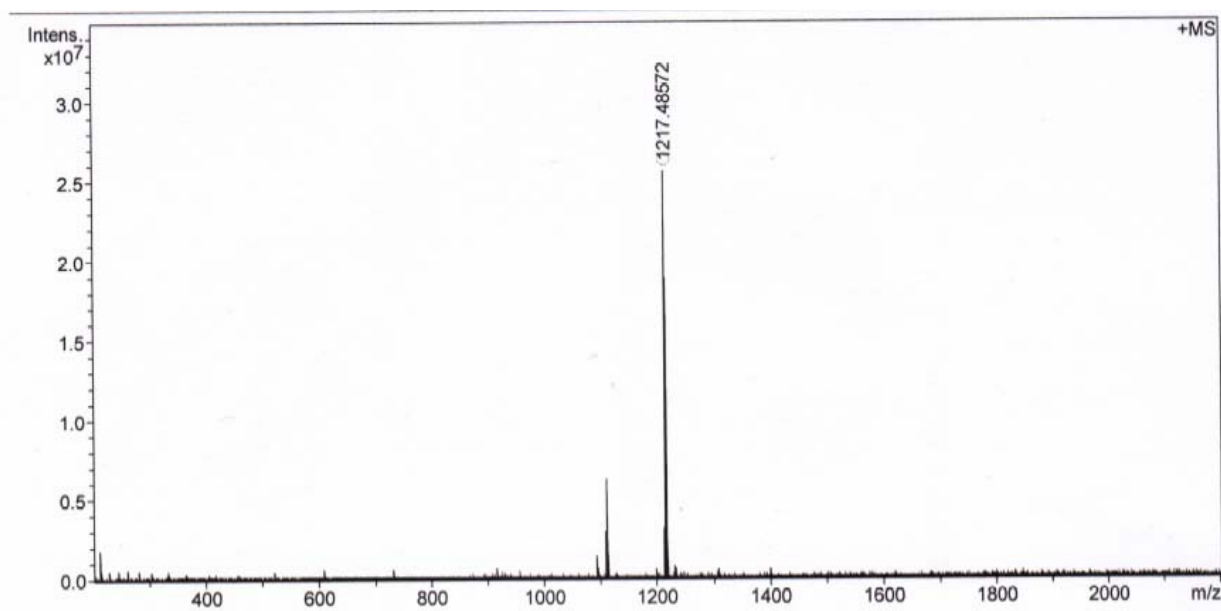
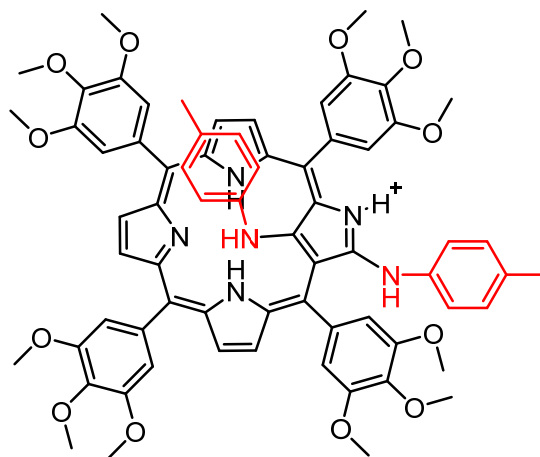


Figure S74. HRMS spectrum of **4d**.



Chemical Formula:  $C_{70}H_{69}N_6O_{12}^+$   
Exact Mass: 1185.4968

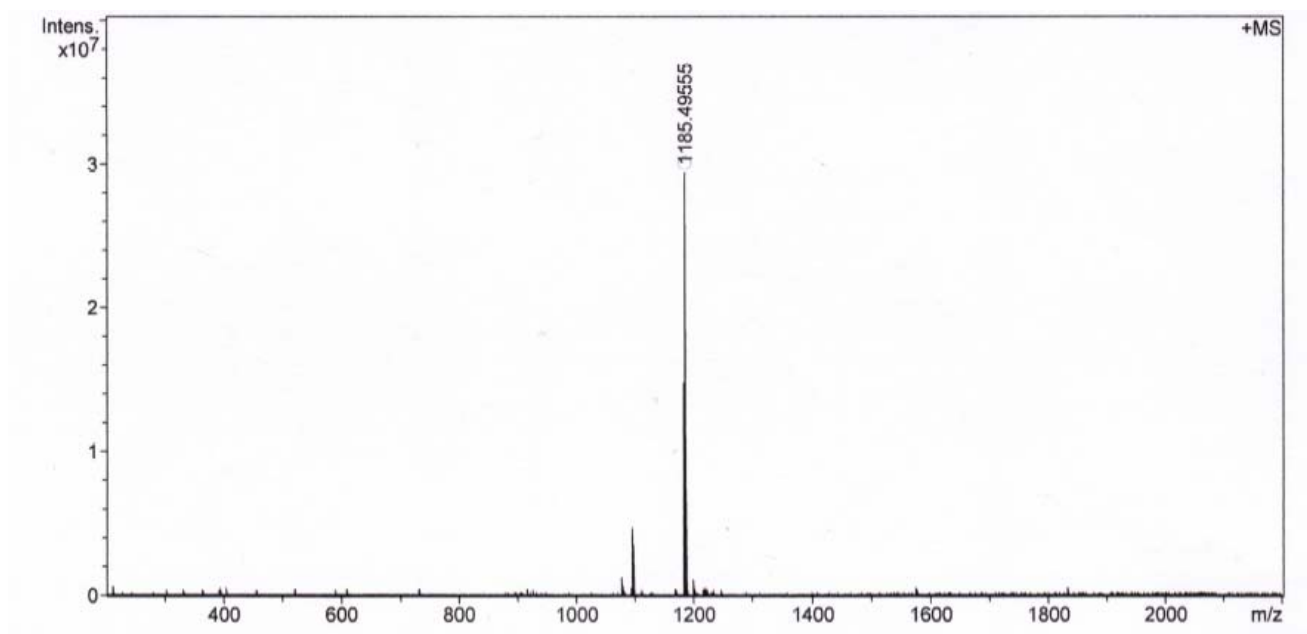
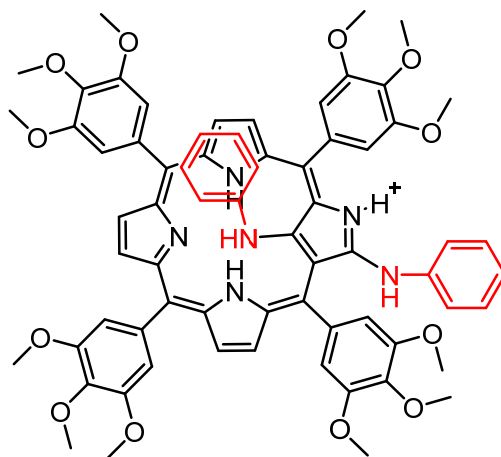


Figure S75. HRMS spectrum of 4e.



Chemical Formula:  $C_{68}H_{65}N_6O_{12}^+$   
Exact Mass: 1157.4655

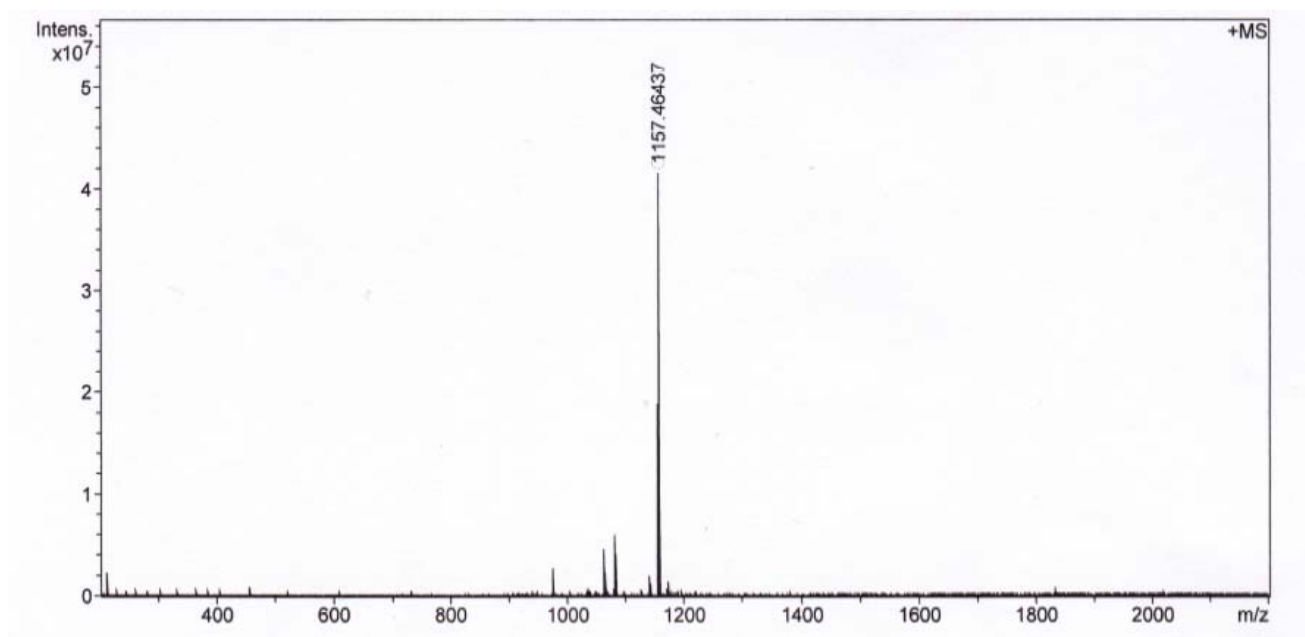
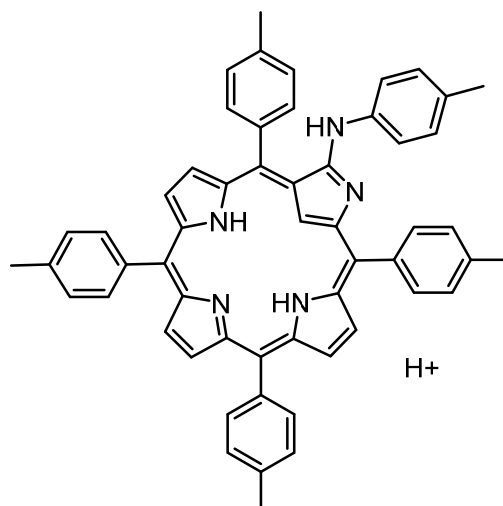


Figure S76. HRMS spectrum of **4f**.





Chemical Formula:  $C_{55}H_{46}N_5^+$   
Exact Mass: 776.3748

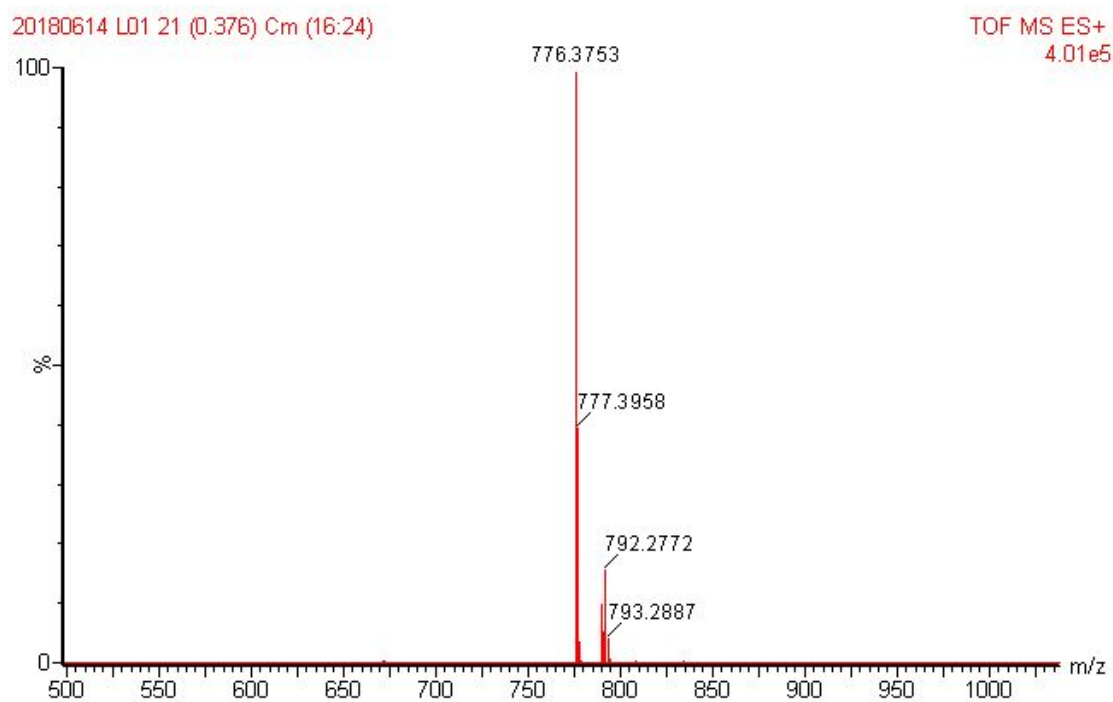
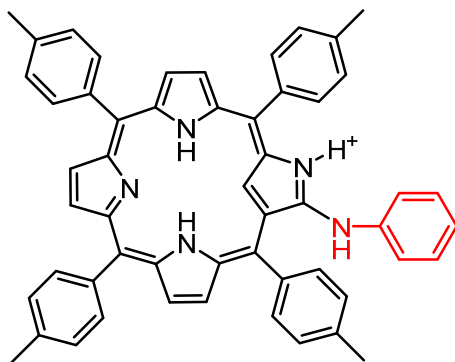
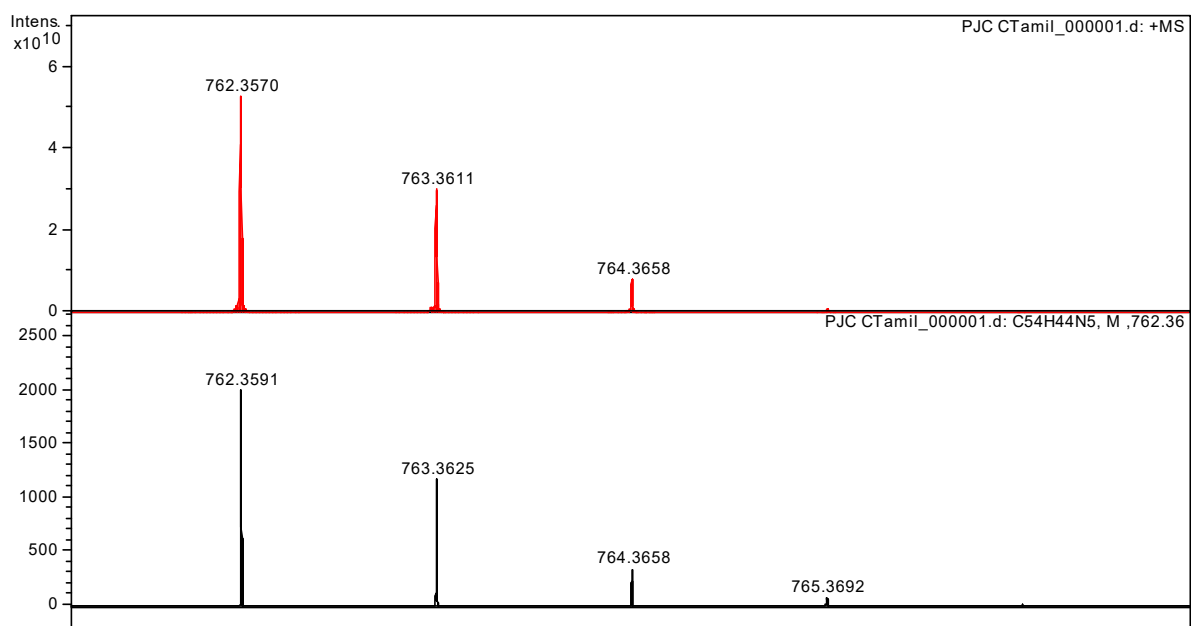


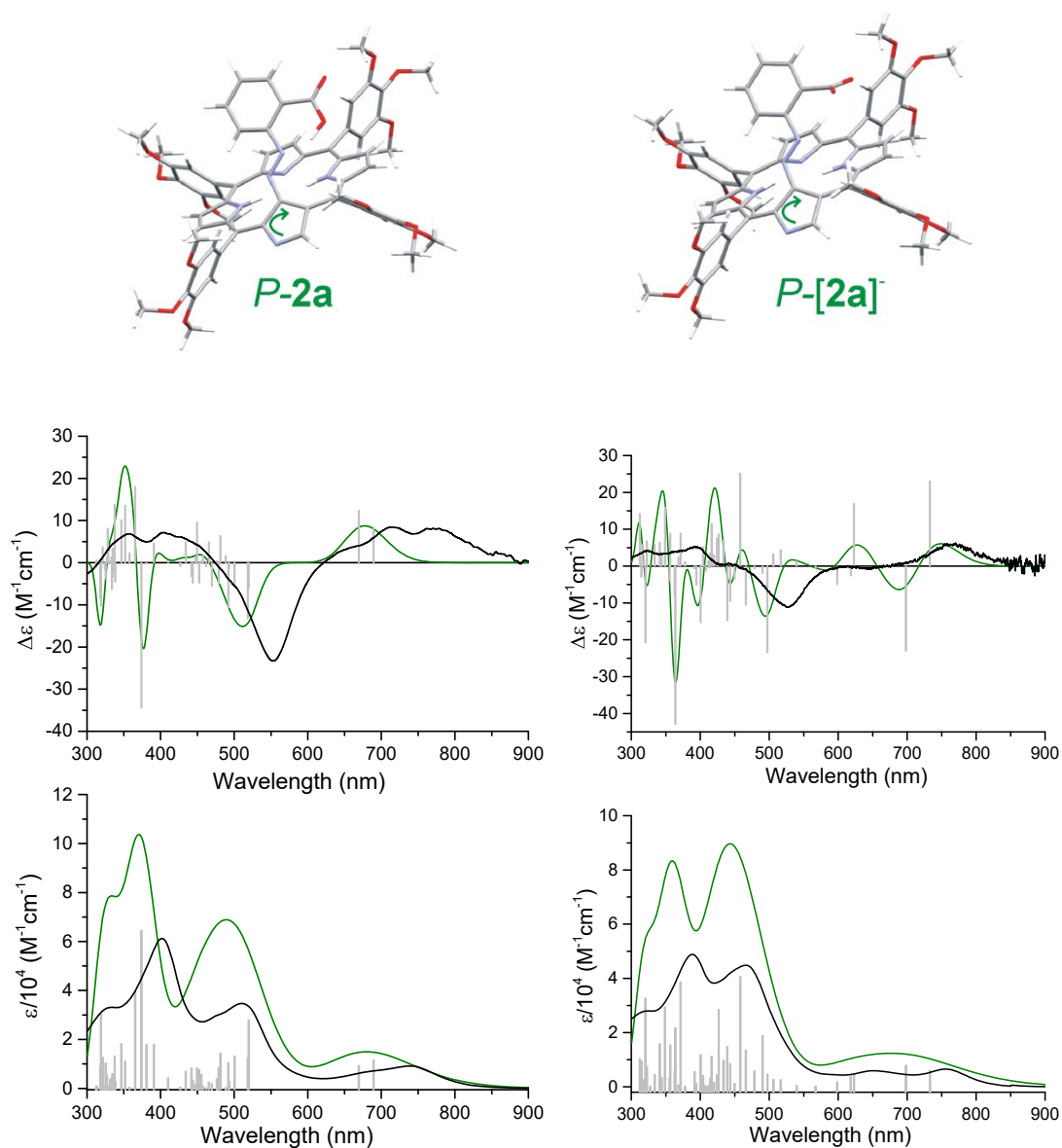
Figure S77. HRMS spectrum of **5b**.



Chemical Formula:  $C_{54}H_{44}N_5^+$   
Exact Mass: 762.3591



**Figure S78.** HRMS spectrum of **5c**. Top, experimental; bottom, simulated isotope pattern.



**Figure S79.** Experimental (black line) and calculated (green line) CD (top) and UV-vis-NIR (bottom) spectra of *P-2a* (left column) and *P-[2a]<sup>-</sup>* (right column). The gray sticks represent oscillator strengths or rotational strengths of the calculated transitions. The column headings are the DFT-optimized structures of the molecules for which spectra were simulated with definitions of their absolute configurations marked with curved arrows.

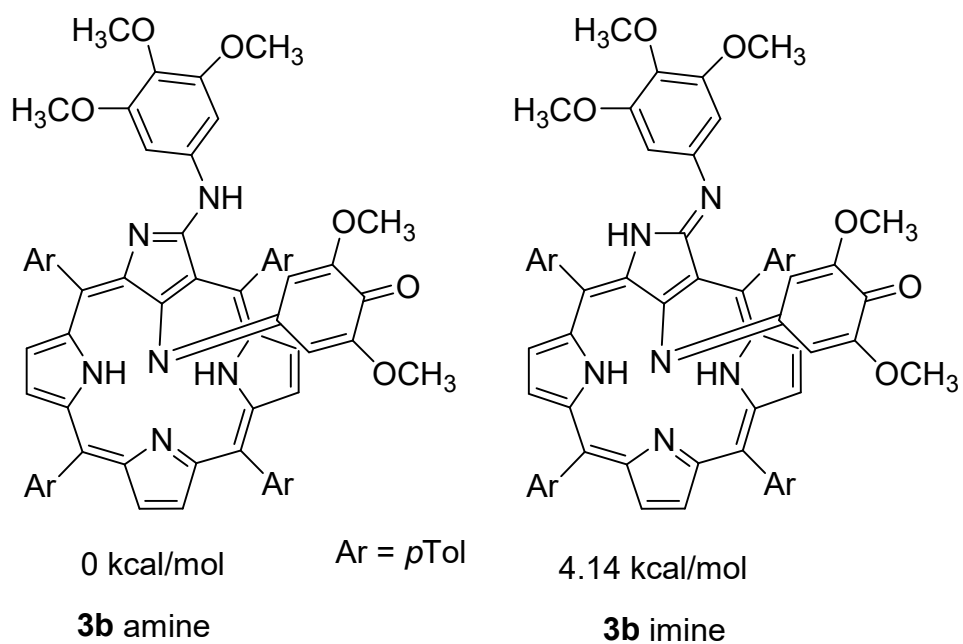


Figure S80. Tautomers of **3b**.

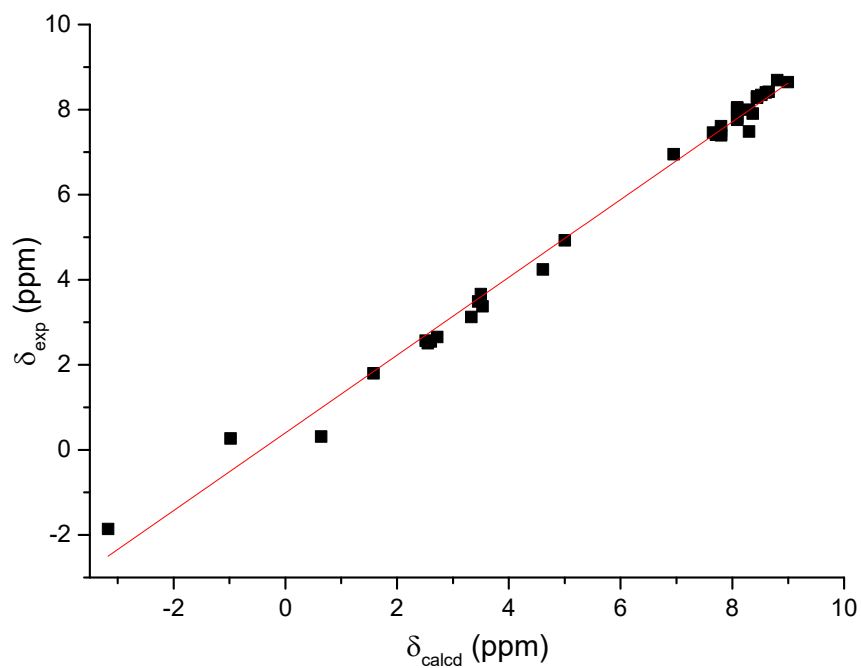
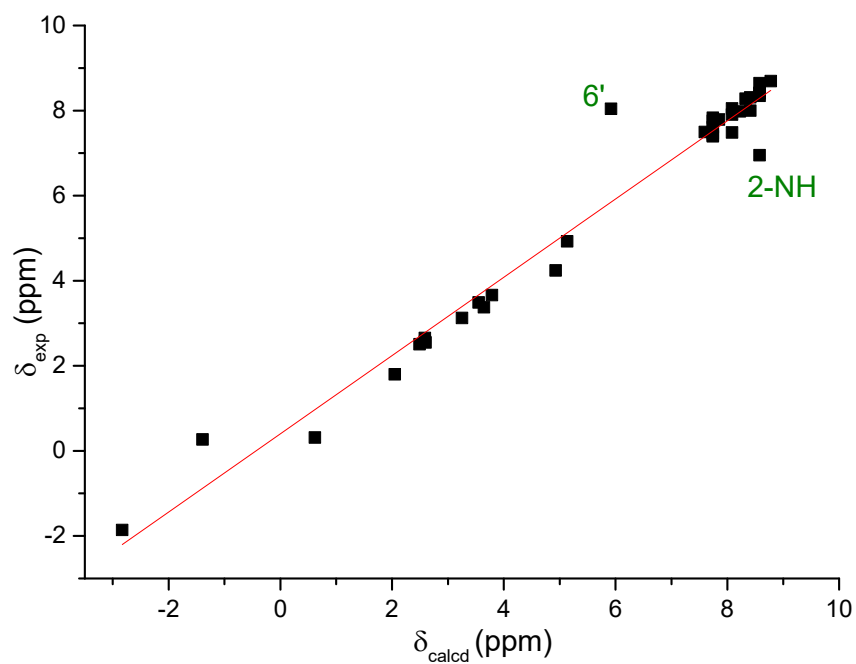
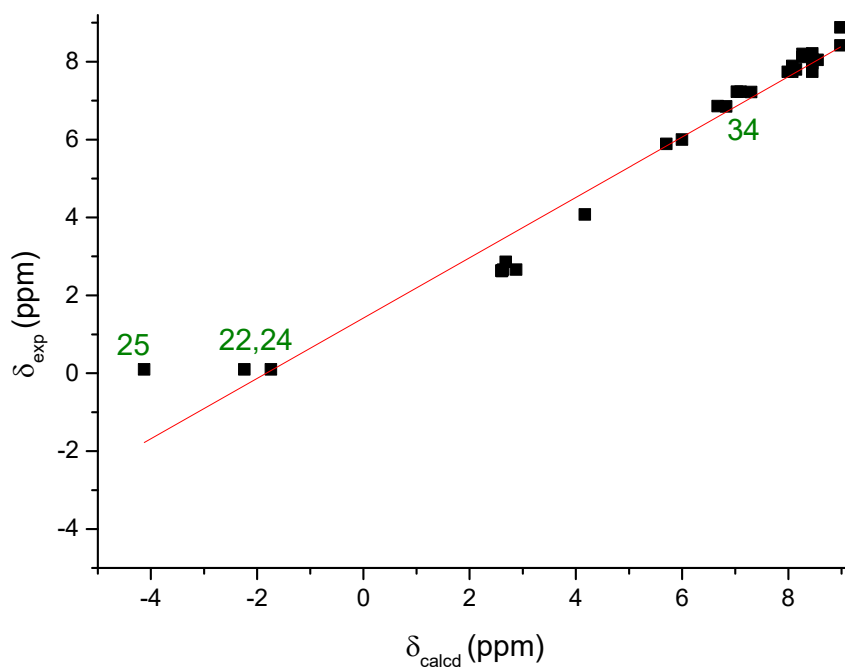


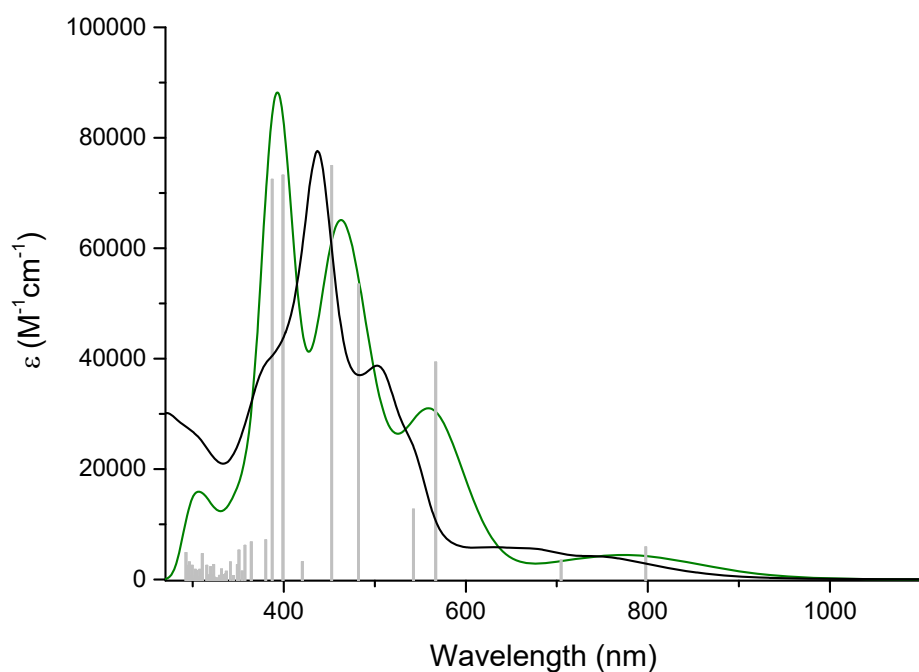
Figure S81. Correlation of experimental (600 MHz,  $\text{CD}_2\text{Cl}_2$ , 173 K) and GIAO-calculated chemical shifts for the amine tautomer of **3b**.



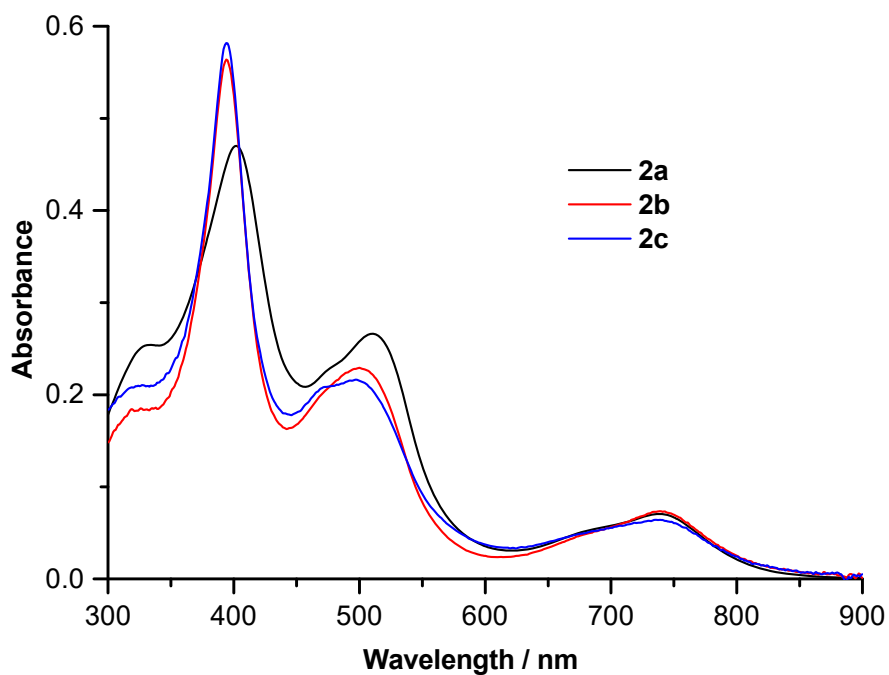
**Figure S82.** Correlation of experimental (600 MHz,  $\text{CD}_2\text{Cl}_2$ , 173 K) and GIAO-calculated chemical shifts for the imine tautomer of **3b**.



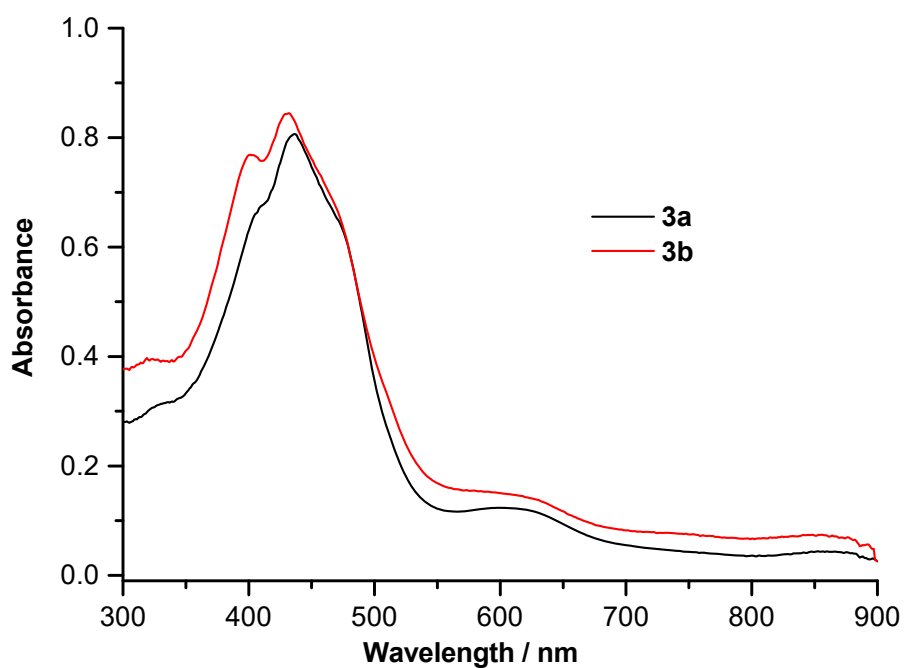
**Figure S83.** Correlation of experimental (600 MHz,  $\text{CD}_2\text{Cl}_2$ , 240 K) and GIAO-calculated chemical shifts for the amine tautomer of **4c**.



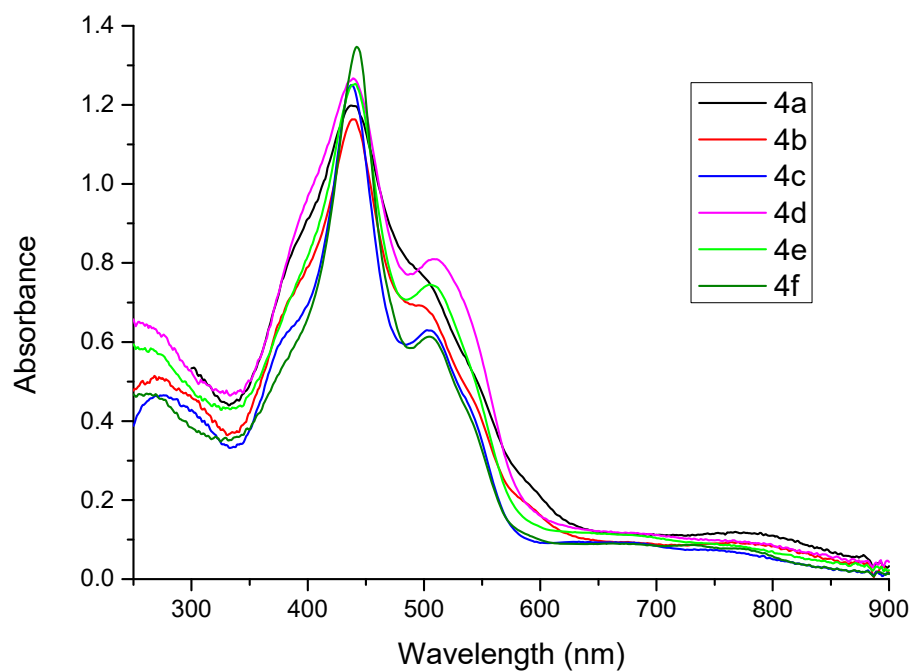
**Figure S84.** Experimental (black line, DCM) and TD DFT simulated (green line) optical spectra of **4c**. The gray sticks represent a histogram consisting of calculated transitions with oscillator strengths multiplied by  $10^5$  for scaling.



**Figure S85.** UV-vis-NIR absorption spectra ( $\text{CHCl}_3$ , 298 K) of 21-aryazo-NCP derivatives **2a-c**.



**Figure S86.** UV-vis-NIR absorption spectra ( $\text{CHCl}_3$ , 298 K) of 3-arylamino-21-quinoneimino-NCP derivatives **3a-b**.

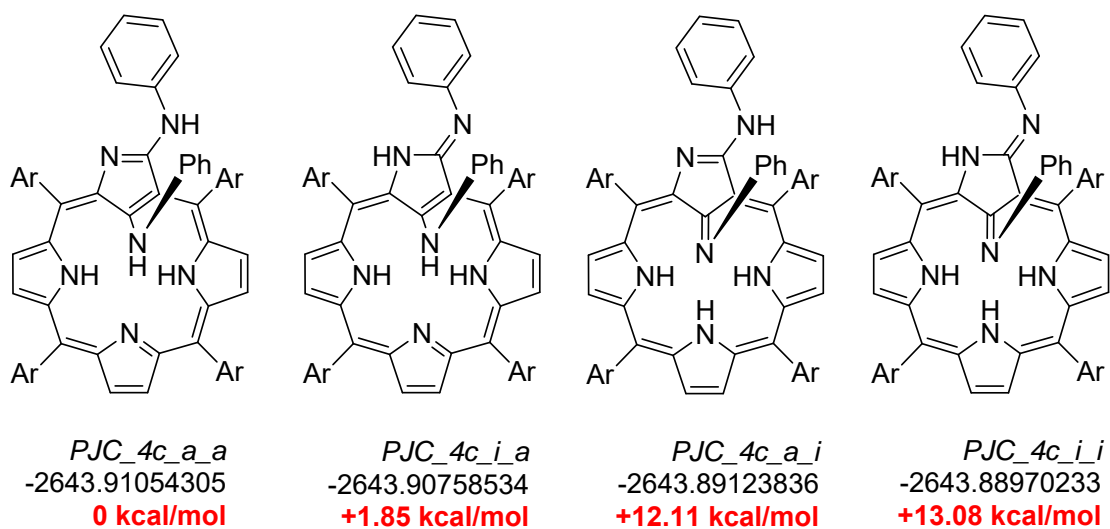


**Figure S87.** UV-vis-NIR absorption spectra ( $\text{CHCl}_3$ , 298 K) of 3,21-bis(arylamine)NCP derivatives **4a-f**.

**Table S2.** Computational details for structures discussed in the paper. Optimizations were performed at the PCM(CH<sub>2</sub>Cl<sub>2</sub>)/B3LYP/6-31G(d,p) level of theory.

Structure	Code <sup>[a]</sup>	SCF E <sup>[b]</sup>	ZPV <sup>[c]</sup>	lowest freq. <sup>[d]</sup>	G <sup>[e]</sup>	HOMO	LUMO	HLG
		a.u.	a.u.	cm <sup>-1</sup>	a.u.	eV	eV	eV
<b>2a</b>	PJC_2a	-3817.16252546	1.110974	6.70	-3816.051551	-5.24	-3.06	2.18
<b>[2a]<sup>-</sup></b>	PJC_2a_neg	-3816.65615019	1.097942	9.27	-3815.558208	-4.61	-2.43	2.18
<b>3b amine</b>	PJC_3b_a	-3290.52006418	1.064930	9.53	-3289.455135	-4.72	-2.91	1.81
<b>3b imine</b>	PJC_3b_i	-3290.51346583	1.064070	7.69	-3289.449396	-4.74	-3.03	1.71
<b>4c a/a<sup>[f]</sup></b>	PJC_4c_a_a	-2643.91054305	0.922494	11.67	-2642.988049	-4.59	-2.52	2.07
<b>4c i/a<sup>[f]</sup></b>	PJC_4c_i_a	-2643.89123836	0.922388	11.15	-2642.968850	-4.18	-2.48	1.70
<b>4c a/i<sup>[f]</sup></b>	PJC_4c_a_i	-2643.90758534	0.922029	11.04	-2642.985556	-4.60	-2.42	2.18
<b>4c i/i<sup>[f]</sup></b>	PJC_4c_i_i	-2643.88970233	0.921707	11.26	-2642.967996	-4.33	-2.51	1.82

[a] Optimized geometry available as <code>.pdb file. [b] Electronic energy. [c] Zero-point vibrational energy. [d] Lowest vibrational frequency. [e] Gibbs free energy. [f] Tautomers: a/a, 3,21-amine; i/a, 3-imine-21-amine; a/i, 3-amine-21-imine; i/i, 3,21-imine.





## Reference List

1. Rigaku Oxford Diffraction (2015). CrysAlis PRO. Rigaku Oxford Diffraction, Wrocław, Poland.
2. G. M. Sheldrick, *Acta Cryst.* 2015, **A71**, 3.
3. G. M. Sheldrick, *Acta Cryst.* 2015, **C71**, 3.
4. A. L. Spek, *Acta Cryst.*, 2015, **C71**, 9.
5. Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, Jr., J. A.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, J. M.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, Ö.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. Gaussian 09 Revision B.01.
6. A. D. Becke, *Phys.Rev.A*, 1988, **38**, 3098.
7. A. D. Becke, *J.Chem.Phys.*, 1993, **98**, 5648.
8. C. Lee, W. Yang, and R. G. Parr, *Phys.Rev.B*, 1988, **37**, 785.
9. N. M. O'Boyle, A. L. Tenderholt, and K. M. Langner, *J.Comp.Chem.*, 2008, **29**, 839.
10. G. R. Geier, III, D. M. Haynes, and J. S. Lindsey, *Org.Lett.*, 1999, **1**, 1455.