Electronic Supplementary Material (ESI) for Organic Chemistry Frontiers. This journal is © the Partner Organisations 2019

# SUPPORTING INFORMATION

# Zinc (II)-Mediated Diastereoselective Passerini Reactions of Biocatalytically Desymmetrized Renewable Inputs

LISA MONI,<sup>a</sup> LUCA BANFI,<sup>a</sup> DANIELE CARTAGENOVA,<sup>a</sup> ANDREA CAVALLI,<sup>b,c</sup> CHIARA LAMBRUSCHINI,<sup>a</sup> ELISA MARTINO,<sup>a</sup> ROMANO V. A. ORRU,<sup>d</sup> EELCO RUIJTER,<sup>d</sup> JORDY M. SAYA,<sup>d</sup> JACOPO SGRIGNANI,<sup>b,c</sup> RENATA RIVA<sup>e\*</sup>

<sup>a</sup> Department of Chemistry and Industrial Chemistry, Università di Genova, via Dodecaneso, 31, 16146 GENOVA, Italy.

<sup>b</sup> Institute for Research in Biomedicine (IRB), Università della Svizzera Italiana (USI), via Vincenzo Vela 6, 6500 BELLINZONA, Switzerland.

- <sup>c</sup> Swiss Institute of Bioinformatics, Quartier Sorge Batiment Genopode, 1015 LAUSANNE, Switzerland.
- <sup>d</sup> Department of Chemistry & Pharmaceutical Sciences, Vrije Universiteit Amsterdam, De Boelelaan 1108, 1081 HZ AMSTERDAM, The Netherlands.
- Department of Pharmacy, Università di Genova, viale Cembrano 4, 16147 GENOVA, Italy.
  [tel.: +39 0103536106; fax: +39 0103536117; E-mail: Renata.Riva@unige.it]

### **Table of contents**

Details on previous desymmetrizations of 2,5-bis(hydroxymethyl)tetrahydrofuran 3 and on determination of absolute configuration

	p. S2
Optimization of the enzymatic acylation of diol <b>3</b>	p. S4
Optimization of Passerini reaction to give <b>7a</b>	p. S7
Mosher's ester analysis to establish the absolute configuration of compounds anti and syn-7a	p. S9
HPLC and NMR determination of enantiomeric excesses and diastereomeric ratios	p. S12
<sup>1</sup> H and <sup>13</sup> C NMR spectra of all new compounds	p. S18

# Details on previous desymmetrizations of 2,5-bis(hydroxymethyl)tetrahydrofuran 3 and on the determination of absolute configuration

The enzymatic monoacylation of **3** was not previously reported. On the other hand, the enzymatic monohydrolysis of diesters of **3**, such as **S1** and **5**, has been previously described.

Naemura et al.<sup>1</sup> reported the monohydrolysis of diacetate **S1** with PLE to give (–)-**S2**, and its conversion into (+)-**S3**. These authors initially determined for (+)-**S3** an ee of 96% by a simple comparison of the polarimetric value with that reported by Jones,<sup>2]</sup> but later corrected it to 51%.<sup>3</sup>

In 1990, Prasad *et al*<sup>4</sup> reported the hydrolysis of dibutyrate **5** with various lipases. The best results were obtained using lipase from *Mucor javanicus* (Fluka). This outcome was confirmed by our optimization, that selected lipase Amano M (again from *Mucor javanicus*) as the best enzyme. The ee reported by Prasad (> 99%) is probably slightly over evaluated, since the reported [ $\alpha$ ]<sub>D</sub> (+16.0) is lower than the one measured by us on a sample of the enantiomer with 99% ee (-17.1). In our hands, we obtained a monobutyrate with 94% ee, and [ $\alpha$ ]<sub>D</sub> = +16.2. It should be noted that Prasad determined the ee by <sup>1</sup>H NMR in the presence of Eu(hfc)<sub>3</sub> and we think that with this method, also due to the fact that in the monobutyrate there are no singlets, the detection limit of the minor enantiomer should be around 3%.

The absolute configuration of (+)-4 and (–)-4 obtained by us was assessed by polarimetric comparison with the value reported by Prasad *et al.* for (+)-4.

Prasad and coworkers have established the absolute configuration of (+)-(2*S*,5*R*)-**4** by its conversion into (–)(2*R*,5*S*)-**S3**.<sup>4</sup> In turn, the absolute configuration of (–)(2*R*,5*S*)-**S3** was previously demonstrated by Jones,<sup>2</sup> who reported the chemoenzymatic synthesis of (–)-**S3** (42% ee) through hydrolytic desymmetrization of diester **S4**. In order to determine the absolute configuration, he converted (–)-**S3** into known (+)-**S6**. This was done by epimerization at the  $\alpha$  position of the ester, followed by LiAlH<sub>4</sub> reduction. The mixture of (+)-**S6** and *meso* **3** was separated and (+)-**S6** was recognized as (*S*,*S*) by polarimetric comparison with the enantiomerically pure compound previously described.<sup>5</sup>

(+)-**S6** was indeed chemically correlated to (–)-**S7**.<sup>5</sup> Moreover, (–)-**S7** was obtained by enantioretentive oxidation of (–)-(R)-2-hexanol.<sup>6</sup> Note that in the paper by Mihailovic et al.<sup>6</sup> the formula of (–)(R,R)-**S7** is erroneously depicted. However, since it derives from (R)-2-hexanol, it must necessarily have the (R,R) configuration.

The absolute configuration of (+)(S,S)-S6 was also confirmed by Jung et al., who obtained it in 8 steps starting from D-glucosamine.<sup>7</sup>

- (1) K. Naemura, N. Takahashi, H. Chikamatsu, Chem. Lett. 1988, 17, 1717-1720.
- (2) J. B. Jones, R. S. Hinks, P. G. Hultin, *Can. J. Chem.* **1985**, 63, 452-456.
- (3) K. Naemura, R. Fukuda, N. Takahashi, M. Konishi, Y. Hirose, Y. Tobe, *Tetrahedron: Asymm.* 1993, 4, 911-918.
- (4) H. Estermann, K. Prasad, M. J. Shapiro, O. Repic, G. E. Hardtmann, J. J. Bolsterli, M. D. Walkinshaw, Tetrahedron Lett. 1990, 31, 445-448.
- (5) M. Nakazaki, K. Naemura, M. Makimura, A. Matsuda, T. Kawano, Y. Ohta, J. Org. Chem. 1982, 47, 2429-2435.

- (6) M. L. Mihailović, R. I. Mamuzić, L. Žigić-Mamuzić, J. Bošnjak, Ž. Čeković, *Tetrahedron* **1967**, 23, 215-226.
- (7) M. E. Jung, I. D. Trifunovich, A. W. Sledeski, *Heterocycles* **1993**, *35*, 273-280.



#### Optimization of the enzymatic acylation of diol 3

**General note to the two following tables:** "As usual for desymmetrization reactions of this type, conversion is defined as the percentage of acylated hydroxy groups. Thus, conversion = ([S1] + [S2]/2)/[starting 3] or  $([5]^*2 + [4])/[\text{starting 3}]^*2$ . Therefore, for example, at 100% conversion, all 3 is converted into diacetate S2, whereas, at 50% conversion, the amounts of diol 3 and diacetate S2 are equal and the yield of S1 is highest. Pushing the reaction at conversions higher than 50% is known to increase the ee of monoester, at the expense of the yield, thanks to a kinetic resolution, which follows the first enantioselective monoacylation. For this reason, it is important to correlate e.e. with conversion. On the other hand, the yield of monoester depends on conversion (it is maximum at 50% conversion), but also on substrate selectivity. In other words, ideally, at 50% conversion one should have 100% monoester and no diol and no diester, because monoester is a poorer substrate than diol. In the reality, substrate selectivity is never so high, and thus, at 50% conversion, some starting diol and some diester are detected. The analytical yield is therefore very useful, during optimization, in order to have an idea of substrate selectivity.

Initially we investigated the enzymatic acetylation of **3**. After a preliminary screening, lipase Amano PS immobilized on celite was selected for further optimization.

		HO	н			(S) OH		AcC			c		
		3			(2R,5S	)- <b>S2</b>			S	1			
Entry	Enzyme	Acyl donor <sup>a</sup>	Solvent	Enzyme quantity	Conc.	Temp.	Time	Conv (%) <sup>b</sup>	% <b>3</b> ℃	% <b>S2</b> °	% <b>S1</b> ℃	Analytical yield <sup>d</sup>	ee <sup>e</sup>
1	Amano PS- Imm	vinyl acetate	THF	0.2 g / g <b>3</b>	0.2 M	rt	6.5 h	68	4	56	40	53%	85%
2	Amano PS- Imm	vinyl acetate	THF	0.2 g / g <b>3</b>	0.2 M	0 °C	4.5 h	41	26	66	8	56%	88%
3	Amano PS- Imm	vinyl acetate	THF	0.2 g / g <b>3</b>	0.2 M	0 °C	4.5 h	41	26	66	8	56%	88%
4	Amano PS- Imm	phenyl acetate	THF	0.2 g / g <b>3</b>	0.2 M	0 °C	8 h	54	16	60	24	54%	86%
5	Amano PS- Imm	vinyl acetate	THF	0.2 g / g <b>3</b>	0.05 M	rt	8 h	46	23	62	15	58%	85%
6	Amano PS- Imm	vinyl acetate	<i>i</i> Pr <sub>2</sub> O	0.2 g / g <b>3</b>	0.05 M	0 °C	3.5 h	54	14	64	22	58%	90%
7	Amano PS- Imm	isopropenyl acetate	<i>i</i> Pr <sub>2</sub> O	0.2 g / g <b>3</b>	0.05 M	0 °C	6.5 h	56	10	68	22	63%	88%

<sup>a)</sup> 5 Equivalents. <sup>b)</sup> Defined as ([**S1**] + [**S2**]/2)/[starting **3**]. <sup>c)</sup> Molar ratios, determined by <sup>1</sup>H NMR on the crude product, which contained only **3**, **S2** and **S1**. <sup>d)</sup> Analytical yield based on <sup>1</sup>H-NMR with 2,5-dimethylfuran as internal standard. <sup>e)</sup> Determined by HPLC on a chiral stationary phase.

Both ees and yields were unsatisfactory. The low yield is due to a poor substrate selectivity. At 50% conversion, substantial amounts of **3** and **S1** were present, as shown in the Table. However, the use of *i*Pr<sub>2</sub>O as solvent showed a slight improvement in ee.

We then passed on to explore the related butyrylation reaction.

			Lipase Amano									
		но, 🔨 он	PS-Imm	nPrCO <sub>2</sub>	(S)	он	<i>n</i> PrC	0 <sub>2</sub>	$\int \int o c$	:O <i>n</i> Pr		
			vinyl butyrate		(s) 0 ~							
		3	(0.5 equiv.)	(	2R,5S) <b>-4</b>			ų	5			
Entry	Additive	Solvent	Enzyme	Conc.	Temp.	Time	Conv (%) <sup>b</sup>	% 3°	% 4°	% 5°	Analytical	ee <sup>e</sup>
			quantity								yield <sup>d</sup>	
1	none	THF	0.2 g / g <b>3</b> f	0.2 M	rt	5 h	63	12	50	38	30%	95%
2	none	THF	0.2 g / g <b>3</b> <sup>f</sup>	0.2 M	0° C	4 h	53	13	68	19	65%	93%
3	none	<i>i</i> Pr <sub>2</sub> O	0.2 g / g <b>3</b> <sup>f</sup>	0.2 M	rt	2.2 h	68	3	58	39	53%	97%
4	none	<i>i</i> Pr <sub>2</sub> O	0.2 g / g <b>3</b> <sup>f</sup>	0.2 M	0 °C	4 h	37	28	70	2	67%	94%
5	none	<i>i</i> Pr <sub>2</sub> O	0.2 g / g <b>3</b> <sup>f</sup>	0.2 M	0 °C	5.5 h	50	8	84	8	80%	96%
6	none	<i>i</i> Pr <sub>2</sub> O	0.2 g / g <b>3</b> <sup>f</sup>	0.2 M	0 °C	7 h	65	0	70	30	70% <sup>e</sup>	98%
7	none	<i>i</i> Pr <sub>2</sub> O	0.2 g / g <b>3</b> <sup>f</sup>	0.2 M	–15 °C	7 h	32	38	60	2	54%	92%
8	none	<i>i</i> Pr <sub>2</sub> O + 5% THF	0.2 g / g <b>3</b> <sup>f</sup>	0.1 M	0 °C	30 h	21	60	38	2	31%	90%
9	none	<i>i</i> Pr <sub>2</sub> O + 20% THF	0.4 g / g <b>3</b> f	0.1 M	0 °C	15 h	45	20	70	10	61%	94%
10	none	<i>i</i> Pr <sub>2</sub> O + 20% CH <sub>2</sub> Cl <sub>2</sub>	0.4 g / g <b>3</b> f	0.1 M	0 °C	62 h	59	3	76	21	70% <sup>e</sup>	99%
11	mol. sieves <sup>a</sup>	<i>i</i> Pr <sub>2</sub> O + 20% CH <sub>2</sub> Cl <sub>2</sub>	0.4 g / g <b>3</b> <sup>f</sup>	0.1 M	0 °C	20 h	59	3	76	21	72% <sup>e</sup>	99%
12	mol. sieves <sup>a</sup>	<i>i</i> Pr <sub>2</sub> O + 20% CH <sub>2</sub> Cl <sub>2</sub>	0.4 g / g <b>3</b> f	0.1 M	0 °C	14 h	50	6	88	6	82% <sup>e</sup>	96%
<sup>a)</sup> 5 Equ	uivalents. <sup>b)</sup> Defi	ned as ([ <b>5</b> ]*2 + [ <b>4</b> ])/[star	ting <b>3</b> ]*2. <sup>c)</sup> Molar ra	atios, dete	rmined by <sup>*</sup>	<sup>1</sup> H NMR on	the crude proc	duct, whic	h containe	ed only <b>3</b> , 4	<b>1</b> and <b>5</b> . <sup>d)</sup> A	nalytical
yield ba	ased on <sup>1</sup> H-NMF	R with 2,5-dimethylfuran	as internal standar	rd. <sup>e)</sup> Deter	mined by H	IPLC on a o	chiral stationar	y phase. <sup>f</sup>	<sup>f)</sup> 1 g of th	is supporte	ed catalyst	
corresp	onds to 0.223 g	of native Amano PS. T	herefore 0.2 g corr	esponds to	o 0.045 g o	f enzyme, a	nd 0.4 g corre	sponds to	0.089 g (	8.9% w/w)	).	

In general, the reaction was more enantioselective than the corresponding acetylation. Di-*iso*-propyl ether gave higher rates and better enantioselectivity than THF. The enantiomeric excess tends to increase when conversion goes beyond 50%, because of favorable kinetic resolution of intermediate **4** during its conversion into **5**. The reaction of entry 6 was nearly perfect, but it was poorly reproducible in

terms of rate on upscaling, probably because of the presence of variable amounts of water and of poor solubility of diol **3** in di-*iso*-propyl ether. Therefore, conditions of entry 11 are the ideal ones, because the reaction times are more reproducible.

The optimized conditions use 400 mg of supported enzyme (corresponding to 89 mg of received Amano PS) per gram of substrate (8.9% w/w). It should be noted that the work-up is very simple. By filtration on a sintered funnel one can recover the supported enzyme, which showed no loss in activity upon three times reuse. The mother liquors, on evaporation to dryness, give crude monobutyrate contaminated with some diol **3** and dibutyrate **5**, depending on conversion.

# Optimization of Passerini reaction to give 7a

Entry	Prep.	Solvent	Additive	Mode of	Temp	Time	Yield % <sup>c</sup>	Yield	dr
	of <b>6</b> ª		(equiv.)	addition <sup>b</sup>	(° C)	(min)		determination	(anti:syn) <sup>d</sup>
1	Α	THF/CH <sub>2</sub> Cl <sub>2</sub> 1:1	none	D	20	40	93	Isolated	59:41
2	Α	THF/CH <sub>2</sub> Cl <sub>2</sub> 1:1	none	D	-78	205	27	NMR	58:42
3	Α	THF/CH <sub>2</sub> Cl <sub>2</sub> 1:1	MgCl <sub>2</sub> (1)	D	20	30	77	NMR	65:35
4	Α	THF/CH <sub>2</sub> Cl <sub>2</sub> 1:1	MgBr <sub>2</sub> (1)	D	20	90	48	NMR	63:37
5	Α	THF/CH <sub>2</sub> Cl <sub>2</sub> 1:1	CuBr (1)	D	20	30	-	NMR	-
6	Α	THF/CH <sub>2</sub> Cl <sub>2</sub> 1:1	CuBr <sub>2</sub> (1)	D	20	60	9	NMR	58:42
7	Α	THF/CH <sub>2</sub> Cl <sub>2</sub> 1:1	BF <sub>3</sub> .Et <sub>2</sub> O (1)	D	20	40	13	NMR	61:39
8	Α	THF/CH <sub>2</sub> Cl <sub>2</sub> 1:1	Yb(OTf) <sub>3</sub> (1)	D	20	90	5	NMR	57:43
9	Α	THF/CH <sub>2</sub> Cl <sub>2</sub> 1:1	Ti(O <i>i</i> Pr) <sub>4</sub> (1)	D	20	40	50	NMR	68:32
10	Α	THF/CH <sub>2</sub> Cl <sub>2</sub> 1:1	Zn(OTf) <sub>2</sub> (1)	D	20	90	47	NMR	57:43
11	Α	THF/CH <sub>2</sub> Cl <sub>2</sub> 1:1	ZnCl <sub>2</sub> (1)	D	20	60	59	NMR	63:37
12	Α	THF/CH <sub>2</sub> Cl <sub>2</sub> 1:1	Znl <sub>2</sub> (1)	D	20	60	85	NMR	64:36
13	Α	THF/CH <sub>2</sub> Cl <sub>2</sub> 1:1	Znl <sub>2</sub> (1)	D	-78	120	36	NMR	65:35
14	Α	THF/CH <sub>2</sub> Cl <sub>2</sub> 1:1	ZnBr <sub>2</sub> (1)	D	20	40	60	NMR	71:29
16	В	toluene	none	D	20	60	69	NMR	62:38
17	В	toluene	ZnBr <sub>2</sub> (1)	D	20	60	61	NMR	72:28
18	В	trifluorotoluene	none	D	20	60	74	NMR	60:40
19	В	trifluorotoluene	ZnBr <sub>2</sub> (1)	D	20	60	80	NMR	72:28
20	В	CH <sub>2</sub> Cl <sub>2</sub>	none	D	20	60	99	NMR	59:41
21	В	CH <sub>2</sub> Cl <sub>2</sub>	ZnBr <sub>2</sub> (1)	D	20	60	32	NMR	64:36
22	В	CHCl <sub>3</sub>	none	D	20	75	78	NMR	63:37
23	В	CHCI <sub>3</sub>	ZnBr <sub>2</sub> (1)	D	20	60	60	NMR	64:36
24	В	THF	none	D	20	60	77	NMR	62:38
25	В	THF	ZnBr <sub>2</sub> (1)	D	20	60	56	NMR	71:29
26	В	Et <sub>2</sub> O	none	D	20	60	74	NMR	62:38

Entry	Prep.	Solvent	Additive	Mode of	Temp	Time	Yield % <sup>c</sup>	Yield	dr
	of <b>6</b> ª		(equiv.)	addition <sup>b</sup>	(° C)	(min)		determination	(anti:syn) <sup>d</sup>
27	В	Et <sub>2</sub> O	ZnBr <sub>2</sub> (1)	D	20	60	78	NMR	72:28
28	В	dimethyl carbonate	none	D	20	60	99	NMR	63:37
29	В	dimethyl carbonate	ZnBr <sub>2</sub> (1)	D	20	90	48	NMR	65:35
30	В	CH₃CN	none	D	20	60	68	NMR	56:44
31	В	CH₃CN	ZnBr <sub>2</sub> (1)	D	20	90	56	NMR	62:38
32	В	MeOH	none	D	20	270	30	NMR	63:37
33	В	MeOH	ZnBr <sub>2</sub> (1)	D	20	240	46	NMR	73:27
34	В	<i>i</i> Pr <sub>2</sub> O	none	D	20	90	94	NMR	62:38
35	В	<i>i</i> Pr <sub>2</sub> O	ZnBr <sub>2</sub> (1)	D	20	90	85	NMR	76:24
36	В	<i>i</i> Pr <sub>2</sub> O/THF 9:1	ZnBr <sub>2</sub> (1)	D	20	90	80	NMR	72:28
37	В	<i>i</i> Pr <sub>2</sub> O	ZnBr <sub>2</sub> (0.4)	D	20	120	85	NMR	76:24
38	В	<i>i</i> Pr <sub>2</sub> O	ZnBr <sub>2</sub> (0.4)	E	20	180	57	NMR	72:28
39	В	<i>i</i> Pr <sub>2</sub> O	ZnBr <sub>2</sub> (0.4)	F	20	180	68	NMR	80:20
40	В	<i>i</i> Pr <sub>2</sub> O	ZnBr <sub>2</sub> (0.4)	G	20	180	88	NMR	79:21
41	В	<i>i</i> Pr <sub>2</sub> O	ZnBr <sub>2</sub> (0.4)	G	20	180	77	isolated	79:21
42	С	<i>i</i> Pr <sub>2</sub> O	ZnBr <sub>2</sub> (0.4)	G	20	180	82	isolated	81:19

<sup>a)</sup> **A**: oxidation was performed with TEMPO/PhI(OAc)<sub>2</sub> system, and the following Passerini was carried out in a one-pot manner. Yield is determined from alcohol **4**. **B**: oxidation was performed with TEMPO/PhI(OAc)<sub>2</sub> system, and the aldehyde purified by chromatography. Yield is determined from aldehyde **6**; **C**: oxidation was performed with Swern methodology, the aldehyde purified by chromatography, and thoroughly dried on molecular sieves. Yield is determined from aldehyde **6**. <sup>b)</sup> **D**: all reagents added together in a short time; **E**: isocyanide added slowly (during 2 h) to the mixture of aldehyde, ZnBr<sub>2</sub> and carboxylic acid; **F**: aldehyde added slowly (during 2 h) to the mixture of isocyanide and ZnBr<sub>2</sub>. <sup>c)</sup> Overall yield of the two diastereomers. NMR yield was determined by <sup>1</sup>H NMR of the crude in the presence of 2,5-dimethylfuran as internal standard. The precision of the method was assessed by preparing a series of solutions by mixing the standard with variable amounts of pure product **7a** and examining them at <sup>1</sup>H NMR. By comparing the real molar % of **7a** in these mixtures (determined by weight) and the ratios determined by NMR we obtained errors always < 3%. <sup>d)</sup> Determined by <sup>1</sup>H-NMR analysis on the crude product, by integration of the signals of *CH*-OAc of the two diastereomers.

The conditions of entry 42, compared to those of entries 40 or 41 were thus selected by us as the best ones. Despite, at first sight, entry 40 gave a better yield, the conditions of entry 42 bring about a better reproducibility and a cleaner crude. While the 81:19 diastereomeric ratio seems not excellent, we must consider that we started from poor dr of 59:41 under the standard Passerini conditions.



## Mosher's ester analysis to establish the absolute configuration of compounds anti and syn-7a

As shown in the Scheme above, both diastereomers of alcohols **9** were converted into the corresponding Mosher's esters **S9a,b** and **S9c,d** by reaction with (R) or (S) Mosher's chloride.

The method by Mosher is based on the chemical shifts of the protons of the groups bound to the stereogenic centers. According to the expected preferred conformations, the anisotropic shielding effect of the phenyl group is exerted on one of the two groups. The table below reports the most diagnostic chemical shift for the four diastereomers **S9a-d**.

	chemical shifts (CDCl <sub>3</sub> )								
Signal	Compound S9a	Compound S9b	Compound S9c	Compound S9d					
(CH <sub>3</sub> ) <sub>3</sub> C	1.24	1.22	1.20	1.29					
NH	6.19	5.91	5.63	5.93					
H-2'	4.31	4.40	4.27	4.34					

In particular, the chemical shifts of the isocyanide derived portion is fully in agreement with the proposed absolute configuration. For the tetrahydrofuran portion we examined the H-2' proton. While for **S9a** and **S9b** also these chemical shifts are in line with the expected effect, for **S9c** and **S9d** the effect seems opposite. However, in this fragment, prediction of the shielding effects is more difficult, because also the *p*-methoxyphenyl group could provoke shielding effects, which clearly depend on the overall conformation of the molecule.

Here we report the complete <sup>1</sup>H NMR data of Mosher esters

**S9a**. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C): *δ* = 7.68-7.60 (2 H, m, aromatics), 7.46-7.38 (3 H, aromatics), 6.85 (4 H, s, aromatics of anisyl), 6.19 (1 H, br s, NH), 5.18 (1 H, d, J = 6.7 Hz, CHOC=O), 4.36-4.27 (2 H, m, 2'-H, 5'-H), 3.97 (1 H, dd, J = 9.9, 3.9 Hz, CHOAn), 3.82 (1 H, dd, J = 9.9, 4.5 Hz, CHOAn), 3.78 (3 H, s, CH<sub>3</sub>O), 3.62 (3 H, q, J = 0.9 Hz, CH<sub>3</sub>O Mosher), 2.00-1.77 (4 H, m, 3'-H, 4'-H), 1.24 (9 H, s, C(CH<sub>3</sub>)<sub>3</sub>)).

**S9b**. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C):  $\delta$  = 7.61-7.54 (2 H, m, aromatics), 7.42-7.35 (3 H, aromatics), 6.81 (4 H, s, aromatics of anisyl), 5.91 (1 H, br s, NH), 5.43 (1 H, d, J = 4.5 Hz, CHOC=O), 4.40 (dt, J = 4.5, 6.9 Hz, 2'-H), 4.35-4.26 (1 H, m, 5'-H), 3.88, 3.86 (2 H, AB part of ABX system, J<sub>AB</sub> = 9.8, J<sub>AX</sub> = 5.4, J<sub>BX</sub> = 4.5 Hz, CH<sub>2</sub>OAn), 3.77 (3 H, s, CH<sub>3</sub>O), 3.55 (3 H, q, J = 0.9 Hz, CH<sub>3</sub>O Mosher), 2.06-1.95 (3 H, m, 3-H, 4-H), 1.90-1.78 (1 H, m, 3-H, 4-H), 1.22 (9 H, s, C(CH<sub>3</sub>)<sub>3</sub>)).

**S9c**. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C):  $\delta$  = 7.63-7.55 (2 H, m, aromatics), 7.44-7.35 (3 H, aromatics), 6.80 (4 H, s, aromatics of anisyl), 5.64 (1 H, br s, NH), 5.23 (1 H, d, J = 6.6 Hz, CHOC=O), 4.34-4.22 (2 H, m, 2'-H, 5'-H); 3.88, 3.85 (2 H, AB part of ABX system, J<sub>AB</sub> = 9.8, J<sub>AX</sub> = 4.8, J<sub>BX</sub> = 5.7 Hz, CH<sub>2</sub>OAn), 3.76 (3 H, s, CH<sub>3</sub>O), 3.55 (3 H, q, J = 0.9 Hz, CH<sub>3</sub>O Mosher), 2.16-1.98 (3 H, m, 3-H, 4-H), 1.89-1.72 (1 H, m, 3-H, 4-H), 1.20 (9 H, s, C(CH<sub>3</sub>)<sub>3</sub>)).

**S9d**. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 25 °C):  $\delta$  = 7.61-7.54 (2 H, m, aromatics), 7.45-7.33 (3 H, aromatics), 6.81 (4 H, s, aromatics of anisyl), 5.93 (1 H, br s, NH), 5.24 (1 H, d, J = 4.6 Hz, CHOC=O), 4.34 (dt, J = 4.6, 6.5 Hz, 2'-H), 4.29-4.18 (1 H, m, 5'-H), 3.83, 3.80 (2 H, AB part of ABX system, J<sub>AB</sub> = 9.8, J<sub>AX</sub> = 4.7, J<sub>BX</sub> = 6.1 Hz, CH<sub>2</sub>OAn), 3.77 (3 H, s, CH<sub>3</sub>O), 3.53 (3 H, q, J = 0.9 Hz, CH<sub>3</sub>O Mosher), 2.07-1.83 (3 H, m, 3-H, 4-H), 1.74-1.61 (1 H, m, 3-H, 4-H), 1.29 (9 H, s, C(CH<sub>3</sub>)<sub>3</sub>)).

#### HPLC and NMR determination of enantiomeric excesses and diastereomeric ratios

Colonna Chirale Chiral Pak AD 250x4,6mm senza precolonn a, Campione GPO3 racemo (Conc.: 400ug/ml Exane/isoPrOH 83:17) flusso 0,5ml/min, T= 26°C term.ON, vinj=20ul,Dad 226nm, isocr. A=Esano=95% B=isoPrOH=5%



Sorted By		:	Signal	
Multiplier		:	1.0000	
Dilution		:	1.0000	
Use Multiplier	&	Dilution	Factor with	ISTDs

Signal 1: DAD1 A, Sig=226,4 Ref=360,100

Peak #	RetTime [min]	Туре	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.335	BV	0.4515	1.49606e4	502.27441	49.9223
2	20.681	VB	0.4740	1.50072e4	466.87311	50.0777

Colonna Chirale ChiralPak AD 250x4,6mm senza precolonna , Campione AC\_003(Conc.:200ug/ml Exane:IsoPOH 9-1)) flu sso 0,5ml/min, T= 26°C term.ON, vinj=20ul, Dad 226nm, isocr A=Esano B=isoprOH, A=95%, B=5%

Injection Date	• 1/7/2006 8·12·01 MM
Sample Name	: AC 003 Tocation : Vial 1
Acg. Operator	: AeV( Lisa)
Acq. Instrument	: stanza306new Inj Volume : 20 µl
Acq. Method	: C:\HPCHEM\1\METHODS\CHIRAL.M
Last changed	: 1/7/2006 8:25:33 AM by AeV( Lisa)
	(modified after loading)
Analysis Method	: C:\HPCHEM\1\METHODS\CHIRAL.M
Last changed	: 1/7/2006 8:42:34 AM by AeV( Lisa)
	(modified after loading)
DAD1 A, Sig	=226,4 Ref=450,100 (LISA\AC_00300.D)
mAU -	0 0
	500°
400 -	Matter in the second
	ľ
350 -	
300	
1 1	
1	
250 -	
] ]	
200 -	
150	
400	
50	₹ A

mir

Area Percent Report

Sorted By	:	Signal	
Multiplier	:	1.0000	
Dilution	:	1.0000	
Use Multiplier a	Dilution	Factor with	ISTDs

Signal 1: DAD1 A, Sig=226,4 Ref=450,100

Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	00
1	22.574	MM	0.5072	52.75705	1.73360	0.3370
2	23.989	MM	0.6087	1.56006e4	427.17075	99.6630

(-)-4 from enzymatic butyrylation of 3

#### **RACEMIC 4**

#### Anti-ent-10



Peak #	RetTime [min]	Туре	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	7.137	MM	0.2922	33.96866	1.93753	1.1328
2	9.382	MM	0.3429	2964.68018	144.10350	98.8672



12 5.11 5.10 5.09 5.08 5.07 5.06 5.05 5.04 5.03 5.02 5.01 5.00 4.99 4.98 4.97 4.96 4.95 fl (ppm)



IS 5.180 5.175 5.170 5.165 5.160 5.155 5.150 5.145 5.140 5.135 5.130 5.125 5.120 5.115 5.110 5.105 5.100 5.095 5.090 5.085 5.080 5.075 f1 (com)

-4.579 -4.568 -4.557 -4.557 -4.557 -4.557 -4.5519 -4.450 -4.496 -4.496 -4.457





5.240 5.180 5.170 f1 (ppm) 5.160 5.130 5.230 5.220 5.210 5.200 5.190 5.150 5.140 5.120





anti**-7**f

4.50

85.50

4.47

4.46 4.45

4.48

4.52 4.50 f1 (ppm) 4.64 4.62 4.60 4.58 4.56 4.54 4.48 4.46 4.44 4.42 4.40 4.38



20 6.18 6.16 6.14 6.12 6.10 6.08 6.06 6.04 6.02 6.00 5.98 5.96 5.94 5.92 5.90 5.88 5.86 5.84 5.82 5.80 5.78 5.76 5.74 f1 (ppm)



) 5.59 5.58 5.57 5.56 5.55 5.54 5.53 5.52 5.51 5.50 5.49 5.48 5.47 5.46 5.45 5.44 5.43 5.42 5.41 5.4( f1 (ppm)





6.30 6.28 6.26 6.24 6.22 6.20 6.18 6.16 6.14 6.12 6.10 6.08 6.06 6.04 6.02 6.00 5.98 5.96 5.94 5.92 5.90 5.88 5.84 fl (ppm)



52 6.50 6.48 6.46 6.44 6.42 6.40 6.38 6.36 6.34 6.32 6.30 6.28 6.26 6.24 6.22 6.20 6.18 6.16 6.14 6.12 ft (ppm)



# <sup>1</sup>H and <sup>13</sup>C NMR spectra of all new compounds







































































































































