Supporting Information

Enantioselective Synthesis of Chiral Multicyclic γ-Lactones *via* Dynamic Kinetic Resolution of Racemic γ-Keto Carboxylic Acids

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1. General Information

Unless otherwise noted, all reagents and solvents were purchased from commercial suppliers and used without further purification. NMR spectra were recorded on Bruker ADVANCE III (400 MHz) spectrometers for ¹H NMR and ¹³C NMR. CD₃OD and CDCl₃ were the solvents used for the NMR analysis, with tetramethylsilane as the internal standard. Chemical shifts were reported upfield to TMS (0.00 ppm) for ¹H NMR and relative to CDCl₃ (77.3 ppm) for ¹³C NMR. Optical rotation was determined using a Perkin Elmer 343 polarimeter. HPLC analysis was conducted on an Agilent 1260 Series instrument. Column Chromatography was performed with silica gel Merck 60 (300-400 mesh). All new products were further characterized by HRMS. A positive ion mass spectrum of sample was acquired on a Thermo LTQ-FT mass spectrometer with an electrospray ionization source.

2. General procedure for the synthesis of β-keto carboxylic acids



Preparation of III according to the literature^{[1][2]}:

To a solution of substituted benzene (I, 75 mmol) and succinic anhydride (50 mmol) dissolved in 100 mL of anhydrous CH_2Cl_2 was slowly added aluminum chloride (100 mmol) at 0 °C. After completion, the reaction mixture was warmed to room temperature and stirred overnight. Then the reaction mixture was cooled to 0 °C and 6N HCl was added carefully. When all aluminum chloride was quenched, the solution was extracted with CH_2Cl_2 three times and the combined organic phase was washed with H_2O and brine and dried over Na₂SO₄, filtered and concentrated under vacuum. Compound II can be used directly without further purification.

 β -benzoyl propionic acid (compound II, 50 mmol) was added to a solution of 0.5M aqueous NaOH (110 mL). Formalin (37%, 4.1 mL) was added to the above solution and the reaction mixture was stirred at room temperature for 1h. Concentrated HCl (32%, 7.2 mL) was added and the solution was stirred overnight. When the reaction

was completed, the mixture was extracted with ethyl acetate three times and washed with brine and dried over Na₂SO₄. The solvent was removed under vacuum and the residue was purified by chromatography on silica gel using hexanes-ethyl acetate (5:1) as eluent to give the target compound III.

Preparation of $V(1a-1o)^{[3]}$:



Compound III (10 mmol) was added slowly to 5 mL concentrated H₂SO₄ at 0 $^{\circ}$ C and then heated to 85 $^{\circ}$ C for 3h. The solution was poured into a beaker containing icewater mixture. The mixture was extracted with DCM for three times, washed with brine and dried over Na₂SO₄. The organic layer was concentrated under vacuum and the residue was purified by chromatography on silica gel using hexanes-ethyl acetate (3:1) and 0.5 % acetic acid as eluent to give the target compound.



To a round bottom flask was added ketone V (10 mmol) and glyoxylic acid (15 mmol). The mixture was stirred at 95 °C for 3h. The reaction mixture was dissolved in acetic acid (15 mL) and water (5 mL). Zinc dust (15 mL) was added over 1h to the solution and the mixture was stirred for additional 3h. The mixture was diluted with ethyl acetate and then filtered through celite. The filtration was extracted with ethyl acetate, washed with brine, dried over Na₂SO₄ and concentrated under vacuum. The residue was purified by chromatography on silica gel using hexanes-ethyl acetate (3:1) and 0.5 % acetic acid as eluent to give the target compound.



2-(1-oxo-2,3-dihydro-1H-inden-2-yl) acetic acid (1a)

White solid, 5 g, 41 % yield; ¹H NMR (400 MHz, CD₃OD) δ 7.75 (d, *J* = 7.6 Hz, 1H), 7.69 (t, *J* = 7.2 Hz, 1H), 7.58 (d, *J* = 7.6 Hz, 1H), 7.45 (t, *J* = 7.4 Hz, 1H), 3.51-3.47 (m, 1H), 3.05 - 2.88 (m, 3H), 2.76 - 2.72 (m, 1H). ¹³C NMR (101 MHz, CD₃OD) δ 193.34, 173.57, 162.03, 135.82, 126.60, 121.05, 120.37, 117.56, 70.03, 42.45, 29.44. ESI-HRMS Calculated for C₁₁H₁₀O₃⁺ ([M+H]⁺): 190.0630, found: 190.0632.

2-(5-fluoro-1-oxo-2,3-dihydro-1*H*-inden-2-yl) acetic acid (1b)

Light yellow solid, 0.48g, 70% yield; ¹H NMR (400 MHz, CD₃OD) δ 7.74 (dd, *J* = 8.5, 5.4 Hz, 1H), 7.26 (dd, *J* = 8.8, 2.1 Hz, 1H), 7.15 (td, *J* = 8.9, 2.3 Hz, 1H), 3.52 - 3.35 (m, 1H), 3.05 - 2.80 (m, 3H), 2.77 - 2.65 (m, 1H). ¹³C NMR (101 MHz, CD₃OD) δ 206.25, 173.86, 167.39 (d, *J*_{C-F} = 254.5 Hz), 157.13 (d, *J*_{C-F} = 10.1 Hz), 132.79, 125.64 (d, *J*_{C-F} = 11.1 Hz), 115.30, 115.06, 112.85(d, *J*_{C-F} = 23.23 Hz), 43.78, 33.98, 32.36, 32.34. ESI-HRMS Calculated for C₁₁H₉FO₃⁺ ([M+H]⁺): 208.0536, found: 208.0533.



2-(5-chloro-1-oxo-2,3-dihydro-1H-inden-2-yl) acetic acid (1c)

Light yellow solid, 0.41g, 71% yield; ¹H NMR (400 MHz, CD₃OD) δ 7.66 (d, *J* = 8.2 Hz, 1H), 7.57 (s, 1H), 7.41 (dd, *J* = 8.2, 1.6 Hz, 1H), 3.46 - 3.40 (m, 1H), 3.03 - 2.81 (m, 3H), 2.76 - 2.70 (m, 1H). ¹³C NMR (101 MHz, CD₃OD) δ 207.23, 172.76, 167.45, 166.23, 157.18, 157.08, 132.79, 125.69, 125.58, 115.30, 115.06, 112.96, 112.73, 43.78, 33.98, 32.36, 32.34. ESI-HRMS Calculated for C₁₁H₉FO₃⁺ ([M+H]⁺): 224.0240, found: 224.0238.



2-(5-bromo-1-oxo-2,3-dihydro-1*H*-inden-2-yl) acetic acid (1d)

Light yellow solid, 0.45g, 68% yield; ¹H NMR (400 MHz, CD₃OD) δ 7.76 (s, 1H), 7.64 - 7.51 (m, 2H), 3.52 -3.36 (m, 1H), 3.03 - 2.80 (m, 3H), 2.80 - 2.65 (m, 1H). ¹³C NMR (101 MHz, CD₃OD) δ 208.13, 172.76, 167.45, 166.23, 157.18, 157.08, 132.79, 125.69, 125.58, 115.30, 115.06, 112.96, 112.73, 43.78, 33.98, 32.36, 32.34. ESI-HRMS Calculated for C₁₁H₉BrO₃⁺ ([M+H]⁺): 267.9735, found: 267.9738.



2-(5-methyl-1-oxo-2,3-dihydro-1*H*-inden-2-yl) acetic acid (1e)

White solid, 0.4g, 75% yield; ¹H NMR (400 MHz, CD₃OD) δ 7.58 (d, *J* = 7.9 Hz, 1H), 7.34 (s, 1H), 7.26-7.17 (m, 1H), 3.41 - 3.35 (m, 1H), 2.97 - 2.91 (m, 1H), 2.89 - 2.83 (m, 2H), 2.67 - 2.61 (m, 1H), 2.43 (s, 3H). ¹³C NMR (101 MHz, CD₃OD) δ 207.78, 174.04, 154.52, 146.44, 133.80, 128.36, 126.66, 123.05, 43.71, 34.17, 32.31, 20.68. ESI-HRMS Calculated for C₁₂H₁₂O₃⁺ ([M+H]⁺):204.0786, found: 204.0784.



2-(6-methyl-1-oxo-2,3-dihydro-1H-inden-2-yl) acetic acid (1f)

White solid, 0.43g, 74% yield; ¹H NMR (400 MHz, CD₃OD) δ 7.66 - 7.14 (m, 3H), 3.41 - 3.34 (m, 1H), 2.98 - 2.81 (m, 3H), 2.71 - 2.64 (m, 1H), 2.39 (s, 3H). ¹³C NMR (101 MHz, CD₃OD) δ 208.62, 208.27, 174.03, 151.34, 137.32, 136.28, 136.04, 135.15, 127.35, 126.09, 123.01, 120.56, 43.91, 43.54, 34.19, 34.15, 32.08, 31.28, 19.67, 16.36. ESI-HRMS Calculated for C₁₂H₁₂O₃⁺ ([M+H]⁺):204.0786, found: 204.0785.



2-(1-oxo-1,2,3,4-tetrahydronaphthalen-2-yl) acetic acid (1g)

White solid, 0.38g, 40% yield; ¹H NMR (400 MHz, CD₃OD) δ 7.99 - 7.84 (m, 1H), 7.52 - 7.48 (m, 1H), 7.42 - 7.23 (m, 2H), 3.18 - 3.10 (m, 1H), 3.06 - 2.93 (m, 2H), 2.86 (dd, *J* = 16.6, 5.9 Hz, 1H), 2.47 (dd, *J* = 16.6, 6.4 Hz, 1H), 2.27 - 2.21 (m, 1H), 2.06 - 1.90 (m, 1H). ¹³C NMR (101 MHz, CD₃OD) δ 199.12, 174.76, 144.51, 133.30, 131.96, 128.67, 126.65, 126.24, 44.68, 34.37, 28.97, 28.80. ESI-HRMS Calculated for C₁₂H₁₂O_{3⁺} ([M+H]⁺): 204.0786, found: 204.0788.



2-(6-methoxy-1-oxo-1,2,3,4-tetrahydronaphthalen-2-yl) acetic acid (1h)

White solid, 0.51g, 50% yield; ¹H NMR (400 MHz, CD₃OD) δ 8.12 - 7.68 (m, 1H), 6.95 - 6.64 (m, 2H), 3.86 (s, 3H), 3.19 - 3.02 (m, 1H), 3.02 - 2.77 (m, 3H), 2.50 - 2.35 (m, 1H), 2.33 - 2.13 (m, 1H), 2.10 - 1.85 (m, 1H). ¹³C NMR (101 MHz, CD₃OD) δ 202.03, 178.81, 167.98, 151.14, 133.08, 129.32, 116.98, 116.09, 58.56, 48.32, 38.39, 33.09, 32.97. ESI-HRMS Calculated for C₁₃H₁₄O₄⁺ ([M+H]⁺): 234.0892, found: 234.0891.



2-(7-methoxy-1-oxo-1,2,3,4-tetrahydronaphthalen-2-yl) acetic acid (1i)

White solid, 0.48g, 49% yield; ¹H NMR (400 MHz, CD₃OD) δ 7.43 (d, *J* = 2.8 Hz, 1H), 7.21 (d, *J* = 8.4 Hz, 1H), 7.09 (dd, *J* = 8.4, 2.8 Hz, 1H), 3.80 (s, 3H), 3.09 - 2.81 (m, 4H), 2.47 (dd, *J* = 16.5, 6.4 Hz, 1H), 2.28 - 2.16 (m, 1H), 2.03 - 1.87 (m, 1H). ¹³C NMR (101 MHz, CD₃OD) δ 199.31, 174.82, 156.94, 133.01, 132.93, 126.68, 118.14,

114.23, 54.82, 44.13, 34.26, 28.21, 22.33. ESI-HRMS Calculated for $C_{13}H_{14}O_4^+$ ([M+H]⁺): 234.0892, found: 234.0889.



2-(5-methoxy-1-oxo-1,2,3,4-tetrahydronaphthalen-2-yl) acetic acid (1j)

White solid, 0.48g, 48% yield; ¹H NMR (400 MHz, CD₃OD) δ 7.43 (d, *J* = 2.8 Hz, 1H), 7.21 (d, *J* = 8.4 Hz, 1H), 7.09 (dd, *J* = 8.4, 2.8 Hz, 1H), 3.80 (s, 3H), 3.10 - 2.78 (m, 4H), 2.47 (dd, *J* = 16.5, 6.4 Hz, 1H), 2.27 - 2.15 (m, 1H), 2.02 - 1.87 (m, 1H). ¹³C NMR (101 MHz, CD₃OD) δ 199.00, 174.76, 158.42, 136.91, 132.72, 129.87, 120.96, 109.10, 54.42, 44.58, 34.42, 29.21, 27.98. ESI-HRMS Calculated for C₁₃H₁₄O₄⁺ ([M+H]⁺): 234.0892, found: 234.0890.



2-(4-oxochroman-3-yl) acetic acid (1k)

White solid, 0.45g, 40% yield; ¹H NMR (400 MHz, CD₃OD) δ 7.82 (dd, *J* = 7.9, 1.7 Hz, 1H), 7.54 - 7.50 (m, 1H), 7.12 - 6.92 (m, 2H), 4.61 (dd, *J* = 11.2, 5.4 Hz, 1H), 4.33 (dd, *J* = 12.3, 11.3 Hz, 1H), 3.30 - 3.21 (m, 1H), 2.81 (dd, *J* = 17.2, 5.1 Hz, 1H), 2.48 (dd, *J* = 17.2, 7.3 Hz, 1H). ¹³C NMR (101 MHz, CD₃OD) δ 193.34, 173.57, 162.03, 135.82, 126.60, 121.05, 120.37, 117.56, 70.03, 42.45, 29.44. ESI-HRMS Calculated for C₁₁H₁₀O_{4⁺} ([M+H]⁺): 206.0579, found: 206.0576.



2-(5-oxo-6,7,8,9-tetrahydro-5H-benzo[7]annulen-6-yl) acetic acid (11)

White solid, 0.39g, 38% yield; ¹H NMR (400 MHz, CD₃OD) δ 7.60 (dd, J = 8.0,

1.4 Hz, 1H), 7.43 - 7.39 (m, 1H), 7.31 - 7.23 (m, 2H), 3.41 - 3.32 (m, 1H), 3.18 - 3.05 (m, 1H), 3.02 - 2.84 (m, 2H), 2.52 - 2.47 (m, 1H), 2.15 - 2.12 (m, 1H), 1.99 - 1.94 (m, 1H), 1.73 - 1.50 (m, 2H). ¹³C NMR (101 MHz, CD₃OD) δ 206.39, 174.56, 142.37, 139.29, 131.33, 129.69, 128.01, 125.97, 45.69, 35.61, 32.98, 29.42, 25.47. ESI-HRMS Calculated for C₁₃H₁₄O₃⁺ ([M+H]⁺): 218.0943, found: 218.0941.

3. General procedure for asymmetric hydrogenation of γ-keto carboxylic acids



A suspension of γ -keto carboxylic acid (0.16 mmol), **Cat** (0.0032 mmol), 5:2 HCO₂H/Et₃N (1 mL) were stirred under N₂ at 60 °C for 12 h until completion according to TLC detection. 5.0 mL 2N HCl was added to the reaction, the mixture was then extracted with ethyl acetate (3×5mL) three times, washed with brine, dried over Na₂SO₄ and concentrated. The desired product was purified by silica gel chromatography (hexanes: ethyl acetate = 1:1). The enantioselectivity of the products was determined by HPLC analysis. The racemic samples of diastereomeric mixtures of **2a-2l** for HPLC analysis were prepared following literature procedures.



(3aR,8bS)-3,3a,4,8b-tetrahydro-2H-indeno[1,2-b]furan-2-one (2a)

White solid, 25 mg, 90% yield, 98% ee, dr = 20:1; $[\alpha]_D^{20} = +100$ (c = 0.985, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak AD-H column, hexane: isopropanol = 85:15; flow rate = 0.8 mL/min; UV detection at 210 nm; t_R = 10.9 min (major), 12.0 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.48 (d, *J* = 7.4 Hz, 1H), 7.39 - 7.26 (m, 3H), 5.90 (d, *J* = 7.1 Hz, 1H), 3.48 - 3.21 (m, 2H), 3.04 - 2.73

(m, 2H), 2.47 - 2.27 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 176.97, 142.51, 138.76, 130.01, 127.60, 126.39, 125.37, 87.70, 37.93, 37.35, 35.70. ESI-HRMS Calculated for C₁₁H₁₀O₂⁺ ([M+H]⁺): 174.0681, found: 174.0680.



(3aR,8bS)-6-fluoro-3,3a,4,8b-tetrahydro-2H-indeno[1,2-b]furan-2-one (2b)

Light yellow solid, 27.5 mg, 90% yield, 98% ee, dr = 20:1; $[\alpha]_D^{20} = +84.8$ (c = 1.00, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak AD-H column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; t_R = 17.8 min (major), 19.2 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.54 - 7.37 (m, 1H), 7.09 - 6.85 (m, 2H), 5.84 (d, *J* = 7.1 Hz, 1H), 3.43 - 3.39 (m, 1H), 3.32 (dd, *J* = 16.6, 8.5 Hz, 1H), 2.91 - 2.85 (m, 2H), 2.41 (dd, *J* = 4.0, 8.0 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 176.67, 164.17 (d, *J*_{C-F} = 249.47), 145.17(d, *J*_{C-F} = 8.1 Hz), 134.62, 127.83 (d, *J*_{C-F} = 9.1 Hz), 115.09 (d, *J*_{C-F} = 23.2 Hz), 112.15 (d, *J*_{C-F} = 22.2 Hz), 86.77, 38.02, 38.00, 37.92, 35.68. ESI-HRMS Calculated for C₁₁H₉FO₂⁺ ([M+H]⁺): 192.0587, found: 192.0585.



(3a*R*,8b*S*)-6-chloro-3,3a,4,8b-tetrahydro-2*H*-indeno[1,2-b]furan-2-one (2c)

Light yellow solid, 30.6 mg, 92% yield, 99% ee, dr = 20:1; $[\alpha]_D^{20} = +73$ (c = 0.6, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak IB column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; t_R = 28.0 min (major), 30.8 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, *J* = 8.0 Hz, 1H), 7.29 (d, *J* = 1.8 Hz, 2H), 5.84 (d, *J* = 7.1 Hz, 1H), 3.46 - 3.35 (m, 1H), 3.31 (dd, *J* = 16.6, 8.4 Hz, 1H), 2.95 - 2.85 (m, 2H), 2.39 (dd, *J* = 18.2, 5.6 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 144.49, 137.31, 128.05, 127.47, 125.56, 86.72, 37.78, 37.65, 35.54.

ESI-HRMS Calculated for C₁₁H₉ClO₂⁺ ([M+H]⁺): 208.0291, found: 208.0292.



(3aR,8bS)-6-bromo-3,3a,4,8b-tetrahydro-2H-indeno[1,2-b]furan-2-one (2d)

Light yellow solid, 35.8 mg, 89% yield, 99% ee, dr=20:1; $[\alpha]_D^{20} = +62$ (c = 0.5, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak IB column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; t_R = 29.6 min (major), 32.8 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.43 (d, *J* = 4.9 Hz, 2H), 7.35 (d, *J* = 8.5 Hz, 1H), 5.83 (d, *J* = 7.1 Hz, 1H), 3.43 - 3.36 (m, 1H), 3.31 (dd, *J* = 10.7, 5.8 Hz, 1H), 2.96 - 2.87 (m, 2H), 2.40 (dd, *J* = 13.4, 4.8 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 176.51, 144.78, 137.81, 130.92, 128.59, 127.80, 124.26, 86.76, 37.72, 37.57, 35.49. ESI-HRMS Calculated for C₁₁H₉BrO₂⁺ ([M+H]⁺): 251.9786, found: 251.9783.



(3a*R*,8b*S*)-6-methyl-3,3a,4,8b-tetrahydro-2*H*-indeno[1,2-b]furan-2-one (2e)

White solid, 27.7 mg, 92% yield, 98% ee, dr = 20:1; $[\alpha]_D^{20} = +90.3$ (c = 1.00, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralcel OD column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; t_R = 23.3 min (minor), 24.3 min (major). ¹H NMR (400 MHz, CDCl₃) δ 7.36 (d, *J* = 7.7 Hz, 1H), 7.16 - 6.97 (m, 2H), 5.86 (d, *J* = 7.0 Hz, 1H), 3.42 - 3.31 (m, 1H), 3.32 - 3.21 (m, 1H), 2.93 - 2.86 (m, 2H), 2.43 - 2.37 (m, 1H), 2.36 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 177.06, 142.80, 140.14, 135.96, 128.53, 126.08, 125.84, 87.62, 37.85, 37.57, 35.81, 21.50. ESI-HRMS Calculated for C₁₂H₁₂O₂⁺ ([M+H]⁺): 188.0837, found: 188.0838.



(3aR,8bS)-7-methyl-3,3a,4,8b-tetrahydro-2*H*-indeno[1,2-b]furan-2-one (2f)

White solid, 27.3 mg, 91% yield, 99% ee, dr = 20:1; $[\alpha]_D^{20} = +105.7$ (c = 1.00, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak AD-H column, hexane: isopropanol = 95: 5; flow rate = 1.0 mL/min; UV detection at 210 nm; t_R = 14.9 min (major), 20.3 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.29 (s, 1H), 7.16 (s, 2H), 5.86 (d, *J* = 7.2 Hz, 1H), 3.38 - 3.34 (m, 1H), 3.27 (dd, *J* = 16.1, 8.4 Hz, 1H), 2.87 (m, 2H), 2.40 (dd, *J* = 15.4, 9.7 Hz, 1H), 2.36 (s, 3H).¹³C NMR (101 MHz, CDCl₃) δ 177.03, 139.39, 138.90, 137.44, 130.96, 126.77, 125.06, 87.72, 37.60, 37.52, 35.73, 21.21. ESI-HRMS Calculated for C₁₂H₁₂O₂⁺ ([M+H]⁺): 188.0837, found: 188.0835.



(3a*R*,9b*S*)-3a,4,5,9b-tetrahydronaphtho[1,2-b]furan-2(3*H*)-one (2g)

White solid, 26.1 mg, 87% yield, 99% ee, dr = 20:1; $[\alpha]_D^{20} = +37.4$ (c = 0.5, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak AD-H column, hexane: isopropanol = 95: 5; flow rate = 1.0 mL/min; UV detection at 210 nm; t_R = 19.6 min (minor), 23.0 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.50 - 7.42 (m, 1H), 7.27 - 7.29 (m, 2H), 7.22 - 7.13 (m, 1H), 5.44 (d, *J* = 5.9 Hz, 1H), 2.93 (dd, *J* = 17.4, 8.1 Hz, 1H), 2.87 - 2.81 (m, 1H), 2.80 - 2.69 (m, 2H), 2.43 (dd, *J* = 17.4, 2.2 Hz, 1H), 1.93 - 1.90 (m, 1H), 1.70 - 1.56 (m, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 176.37, 137.51, 131.11, 130.93, 128.92, 128.77, 126.73, 78.52, 36.43, 34.74, 27.77, 25.15. ESI-HRMS Calculated for C₁₂H₁₂O₂⁺ ([M+H]⁺): 188.0837, found: 188.0839.



(3a*R*,9b*S*)-7-methoxy-3a,4,5,9b-tetrahydronaphtho[1,2-b]furan-2(3*H*)-one (2h)

White solid, 31.4 mg, 91% yield, 99% ee, dr = 20:1; $[\alpha]_D^{20} = +20.3$ (c = 0.3, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak AD column, hexane: isopropanol = 90: 10; flow rate = 1.0 mL/min; UV detection at 210 nm; t_R = 14.8 min (minor), 16.6 min (major). ¹H NMR (400 MHz, CDCl₃) δ 7.37 (d, *J* = 8.5 Hz, 1H), 6.83 (dd, *J* = 8.5, 2.5 Hz, 1H), 6.67 (d, *J* = 2.2 Hz, 1H), 5.40 (d, *J* = 5.7 Hz, 1H), 3.81 (s, 3H), 2.92 (dd, *J* = 17.4, 8.0 Hz, 1H), 2.89 - 2.78 (m, 1H), 2.76 - 2.66 (m, 2H), 2.44 - 2.39 (m, 1H), 1.90 - 1.87 (m, 1H), 1.67 - 1.57 (m, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 176.48, 159.81, 139.14, 132.29, 123.41, 113.16, 113.03, 78.59, 55.31, 36.67, 34.81, 28.15, 25.07. ESI-HRMS Calculated for C₁₃H₁₄O₃⁺ ([M+H]⁺): 218.0943, found: 218.0944.



(3aR,9bS)-8-methoxy-3a,4,5,9b-tetrahydronaphtho[1,2-b]furan-2(3H)-one (2i)

White solid, 31.4 mg, 90% yield, 99% ee, dr = 20:1; $[\alpha]_D^{20} = +47.2$ (c = 1.00, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak AD-H column, hexane: isopropanol = 95: 5; flow rate = 1.0 mL/min; UV detection at 210 nm; t_R = 28.4 min (major), 29.9 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.08 (d, *J* = 8.5 Hz, 1H), 6.97 (d, *J* = 2.6 Hz, 1H), 6.86 (dd, *J* = 8.4, 2.7 Hz, 1H), 5.40 (d, *J* = 6.0 Hz, 1H), 3.80 (s, 3H), 2.92 (dd, *J* = 17.4, 8.1 Hz, 1H), 2.83 - 2.72 (m, 2H), 2.72 - 2.60 (m, 1H), 2.42 (dd, *J* = 17.4, 2.5 Hz, 1H), 1.92 - 1.89 (m, 1H), 1.69 - 1.54 (m, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 176.38, 158.19, 132.00, 129.76, 129.52, 116.15, 114.37, 78.69, 55.42, 36.21, 34.67, 26.89, 25.43. ESI-HRMS Calculated for C₁₃H₁₄O₃⁺ ([M+H]⁺): 218.0943, found: 218.0941.



(3a*R*,9b*S*)-6-methoxy-3a,4,5,9b-tetrahydronaphtho[1,2-b]furan-2(3*H*)-one (2j)

White solid, 32 mg, 92% yield, 99% ee, dr = 20:1; $[\alpha]_D^{20} = +25.3$ (c = 0.8, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak AD-H column, hexane: isopropanol = 95: 5; flow rate = 1.0 mL/min; UV detection at 210 nm; t_R = 25.3 min (major), 37.8 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.24 (d, *J* = 8.0 Hz, 1H), 7.08 (d, *J* = 7.7 Hz, 1H), 6.83 (d, *J* = 8.1 Hz, 1H), 5.42 (d, *J* = 5.8 Hz, 1H), 3.84 (s, 3H), 2.99 (dt, *J* = 17.4, 4.4 Hz, 1H), 2.90 (dd, *J* = 17.3, 7.9 Hz, 1H), 2.78 - 2.67 (m, 1H), 2.50 -2.38 (m, 2H), 1.96 - 1.90 (m, 1H), 1.61 - 1.50 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 176.48, 156.64, 132.08, 127.04, 126.71, 122.79, 109.85, 78.53, 55.49, 36.27, 34.35, 24.32, 20.88. ESI-HRMS Calculated for C₁₃H₁₄O₃⁺ ([M+H]⁺): 218.0943, found: 218.0942.



(3aS,9bS)-3a,9b-dihydro-4H-furo[3,2-c]chromen-2(3H)-one (2k)

White solid, 25.9 mg, 85% yield, 96% ee, dr = 20:1; $[\alpha]_D^{20} = +62.5$ (c = 1.00, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak AD-H column, hexane: isopropanol = 95: 5; flow rate = 1.0 mL/min; UV detection at 210 nm; t_R = 27.5 min (major), 38.3 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.41 (dd, *J* = 7.7, 1.4 Hz, 1H), 7.33 - 7.28 (m, 1H), 7.04 - 7.00 (m, 1H), 6.92 (d, *J* = 8.3 Hz, 1H), 5.49 (d, *J* = 6.3 Hz, 1H), 4.21 (dd, *J* = 11.5, 4.3 Hz, 1H), 3.85 - 3.80 (m, 1H), 3.05 - 2.98 (m, 1H), 2.87 (dd, *J* = 17.7, 8.3 Hz, 1H), 2.46 (dd, *J* = 17.7, 4.2 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 175.43, 155.10, 131.44, 130.75, 121.97, 118.49, 117.45, 74.29, 64.85,

33.61, 31.17. ESI-HRMS Calculated for $C_{11}H_{10}O_3^+$ ([M+H]⁺): 190.0630, found: 190.0631.



(3a*R*,10b*S*)-3,3a,4,5,6,10b-hexahydro-2*H*-benzo[6,7]cyclohepta[1,2-b]furan-2-one (2l)

White solid, 22.6 mg, 70% yield, 98% ee, dr = 20:1; $[\alpha]_D^{20} = -14.4$ (c = 1.00, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak AD-H column, hexane: isopropanol = 95:5; flow rate = 1.0 mL/min; UV detection at 210 nm; t_R = 12.1 min (minor), 13.2 min (major). ¹H NMR (400 MHz, CDCl₃) δ 7.36 - 7.27 (m, 3H), 7.18 - 7.10 (m, 1H), 5.86 (d, *J* = 9.2 Hz, 1H), 2.87 - 2.73 (m, 2H), 2.70 - 2.63 (m, 2H), 2.10 (dd, *J* = 17.8, 10.7 Hz, 1H), 1.72 - 1.68 (m, 3H), 1.03 - 0.87 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 176.67, 135.67, 135.46, 128.65, 128.20, 126.90, 123.24, 81.18, 36.37, 34.06, 30.22, 27.99, 22.85. ESI-HRMS Calculated for C₁₃H₁₄O₂⁺ ([M+H]⁺): 202.0994, found: 202.0995.

4. Gram scale reaction



A suspension of **1a** (2.5 g, 13.15 mmol), **cat.** C (0.082 g, 0.1315 mmol), 5:2 HCO_2H/Et_3N (10 mL) were stirred under N₂ at 60 °C for until completion according to TLC detection. 10 mL 2N HCl was added to the reaction, the mixture was then extracted with ethyl acetate (3×15 mL) three times, washed with brine, dried over Na₂SO₄ and concentrated. The crude product was purified by silica gel chromatography (hexanes:

ethyl acetate = 1:1), affording desired product in 85% yield with 98% ee.

5. Synthesis of (+)-GR24 and (+)-epi-GR24^[4]

To a solution of **2a** (1.1 g, 6.3 mmol) and methyl formate (0.82 mL, 9.45 mmol) in anhydrous THF (15 mL) was added potassium *tert*-butoxide (0.85 g, 7.56 mmol) in small portions at 0 $^{\circ}$ C under nitrogen. The reaction mixture was stirred at room temperature until completion. THF was removed in vacuo. The resulting solid was resolved in 20 mL anhydrous DMF under N₂. To this solution was added bromobutenolide (1.67 g, 9.45 mmol) and the reaction mixture was stirred overnight. The reaction was quenched with saturated aqueous ammonium chloride (20 mL). The reaction mixture was diluted with ethyl acetate (50 mL) and washed with water (3×30 mL). The organic extract was then washed with brine, dried over Na₂SO₄, and the solvent removed under vacuum. The residue was purified by column chromatography, affording pure (+)-GR24 and (+)-*epi*-GR24.



(+)-epi-GR24

White Solid, 0.65 g, 35% yield, $[\alpha]_D^{20} = +292$ (c = 0.50, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.50-7.49 (m, 1H), 7.35-7.23 (m, 1H), 6.99 (s, 1H), 6.21 (s, 1H), 5.96 (d, *J* = 8.0 Hz, 1H), 3.97-3.92 (m, 1H), 3.42 (dd, *J* = 16.9, 9.3 Hz, 1H), 3.10 (dd, *J* = 16.9, 3.1 Hz, 1H), 2.03 (t, *J* = 1.4 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 171.42, 170.40, 151.29, 142.69, 141.25, 138.76, 135.76, 130.05, 127.45, 126.34, 125.28, 113.26, 100.79, 86.00, 38.79, 37.42, 29.69, 10.74.



(+)-GR24

White solid, 0.65 g, 35% yield, $[\alpha]_D^{20} = +451$ (c = 0.50, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.51-7.49 (m, 2H), 7.35 (m, 3H), 7.00 (s, 1H), 6.21 (s, 1H), 5.95 (d, *J* = 7.9 Hz, 1H), 3.97-3.92 (m, 1H), 3.44 (dd, *J* = 16.9, 9.4 Hz, 1H), 3.11 (dd, *J* = 16.9, 3.2 Hz, 1H), 2.03 (t, *J* = 1.4 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.44, 170.37, 151.27, 142.66, 141.16, 138.82, 135.83, 130.04, 127.49, 126.42, 125.18, 113.14, 100.71, 85.99, 38.85, 37.31, 10.73.

6. Reference:

- (1) R. Z. Guo et al. Bioorg. Org. Lett., 2016, 18, 504-507;
- (2) F. Meneghetti et al. Eur. J. Org. Chem. 2015, 4907–4912;
- (3) L. J. Bromhead et al., Aust. J. Chem. 2015, 68, 1221-1227;
- (4) H. Malik et al. Tetrahedron, 2010, 66, 7198-7203.

7. NMR spectra





210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)









S22



S23











210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)



























S41



8. HPLC spectra of 2a-2l

Data File E:\DATA\XZC\XZC-4-159\XZC-4-159-2 2017-07-04 10-30-15\031-0201.D Sample Name: XZC-4-159-RAC

Acg. Operator :	SYSTEM		Seg. Line :	2		
Acq. Instrument :	1260HPLC-VWD		Location :	Vial 31		
Injection Date :	7/4/2017 10:41:48 AM	1	Inj :	1		
		1	inj Volume :	3.000 µl		
Acq. Method :	E:\DATA\XZC\XZC-4-1	59\XZC-4-159	9-2 2017-07-0	04 10-30-1	5\VWD-AD(1-2)-85-3	15-
Lost showed .	0.8ML-3UL-210NM-20M3	IN.M Nature System				
Last changed : Analysis Method :	F.\DATA\Y7C\Y7C_4_1	1 DY SISIEM 591777-4-150	2-2 2017-07-0	14 10-30-1	5)WWD-AD(1-2)-85-	15-
Andrysis neared .	0.8ML-3UL-210NM-20M	IN.M (Sequer	ce Method)	J4 10 J0 1.	5,700-AD(1-2)-05	1.0
Last changed :	4/8/2019 11:19:47 AM	I by SYSTEM				
	(modified after load	ling)				
Additional Info :	Peak(s) manually int	tegrated	2 2047 07 04 40 20	45004.0004.00		
	engur-210 nm (EXDATAV20V20	-4108020-4108	2 2017-07-04 10-30	F15031-0201.D)		
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Dilution	: 1.0000					
Do not use Multipl	ier & Dilution Facto	or with ISTI)s			
Signal I. UUD 1 ×	Norrelangth - 210 mm					
Signal I: YWDI A,	wavelengun=210 nm					
Peak RetTime Type	Width Area	Height	Area			
# [min]	[min] [mAU*s]	[mAU]	÷			
	-					
1 10.923 BV	0.2170 275.72559	19.03780	50.9647			
2 12.022 MM	0.2582 265.28693	17.12114	49.0353			
Totals :	541.01251	36.15894				
	*** End of F	======================================				
	2					
HPLC-VWD 4/8/2019	11:19:54 AM SYSTEM				Page 1 of 1	

Data File E:\DATA\XZC\XZC-4-159\XZC-4-159-3 2017-07-04 11-12-30\097-1101.D Sample Name: XZC-4-159-6 ------Acq. Operator : SYSTEM Acq. Instrument : 1260HPLC-VWD Seq. Line : 11 Location : Vial 97 Injection Date : 7/4/2017 2:30:48 PM Inj: 1 Inj Volume : 3.000 µl : E:\DATA\XZC\XZC-4-159\XZC-4-159-3 2017-07-04 11-12-30\VWD-AD(1-2)-85-15-Acg. Method 0.8ML-3UL-210NM-20MIN.M Last changed : 7/4/2017 11:12:30 AM by SYSTEM Analysis Method : E:\DATA\XZC\XZC-4-159\XZC-4-159-3 2017-07-04 11-12-30\VWD-AD(1-2)-85-15-0.8ML-3UL-210NM-20MIN.M (Sequence Method) Last changed : 4/8/2019 11:18:14 AM by SYSTEM (modified after loading) Additional Info : Peak (s) manually integrated VWD1A, Wavelength=210 nm (E:\DATAVZCVZC-4-159\/ZC-4-159\/3 2017-07-0411-12-30\/097-1101.D) mAU 0 700 600 · 500 10.934 400 · 300 -200 -100 12.062 0 10 12 å min Area Percent Report Sorted By Signal : Multiplier 1.0000 : 1.0000 Dilution : Do not use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=210 nm Peak RetTime Type Width Area Height Area [min] [mAU*s] [mAU] * # [min] 1 10.934 BB 0.2208 5685.89355 388.29538 98.7844 2 12.062 BB 0.2554 69.96854 4.07334 1.2156 5755.86209 392.36872 Totals : *** End of Report *** Page 1 of 1 1260HPLC-VWD 4/8/2019 11:18:16 AM SYSTEM

Data File E:\DATA\XZC\DATA-2\XZC-20180314-2 2018-03-14 14-46-08\052-0301.D Sample Name: XZC-pF-RAC-1 -----Acq. Operator : SYSTEM Acq. Instrument : 1260HPLC-VWD Seq. Line : 3 Location : Vial 52 Injection Date : 3/14/2018 3:38:36 PM Inj: 1 Inj Volume : 2.000 µl Acq. Method : E:\DATA\XZC\DATA-2\XZC-20180314-2 2018-03-14 14-46-08\VWD-AD(1-2)-95-5-1ML-2UL-210NM-40MIN.M Last changed : 3/14/2018 2:46:08 PM by SYSTEM Analysis Method : E:\DATA\XZC\DATA-2\XZC-20180314-2 2018-03-14 14-46-08\VWD-AD(1-2)-95-5-1ML-2UL-210NM-40MIN.M (Sequence Method) Last changed : 4/8/2019 10:58:20 AM by SYSTEM (modified after loading) Additional Info : Peak(s) manually integrated VWD1A, Wavelength=210 nm (E\DATAVZC\DATA-2VZC-201803142 201803-14 14-46-08/052-0301.D) mAU ,0 O 175 150 · 125 -100 -75 · 19.981 18.997 **5**0 · 25 -0 25 7.5 10 12.5 15 17.5 20 min Area Percent Report Sorted By Signal : Multiplier 1.0000 : 1.0000 Dilution : Do not use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=210 nm Peak RetTime Type Width Area Height Area [min] [mAU*s] [mAU] ÷ # [min] # [min] [min] [min] [____] 1 18.997 BV 0.4186 1336.70410 49.26241 48.8079 2 19.981 VB 0.3899 1402.00256 54.53653 51.1921 2738.70667 103.79893 Totals : *** End of Report *** Page 1 of 1

1260HPLC-VWD 4/8/2019 10:58:25 AM SYSTEM

_____ Acq. Operator : Seq. Line : 6 Acq. Instrument : Instrument 2 Location : Vial 34 Injection Date : 10/29/2018 10:26:43 PM Inj: l Inj Volume : 5.000 µl : D:\DATA\XZC\XZC-KET0 ACID\XZC-ACID-1029-1 2018-10-29 20-45-25\DAD-AD(1-6)-Acq. Method 95-5-1ML-5UL-210-35MIN.M Last changed : 10/29/2018 8:46:43 PM Analysis Method : D:\METHOD\XZC\VWD-OD(1-6)-90-10-1ML-5UL-210NM-40MIN.M Last changed : 10/30/2018 12:00:56 PM (modified after loading) Additional Info : Peak(s) manually integrated DAD1 C, Sig=210 4 Ref=off (DADATAWZCWZC-KETO ACIDWZC-ACID-1029-1 2018-10-29 20-45-25'034-0601.D) mAU *,*,0 \cap 800 19891.0 600 400 200 193013 202 Û 17 23 18 22 24 16 19 20 21 -----Area Percent Report ------Signal Sorted Bv : Multiplier : 1.0000 1.0000 Dilution : Use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 C, Sig=210,4 Ref=off Peak RetTime Type Width Height Area Area * - | -- -- - | 1 17.896 MF 0.6084 1.98916e4 544.92200 99.0360 2 19.202 FM 0.2470 193.61253 13.06445 0.9640 Totals : 2.00852e4 557.98644 Page 1 of 2 Instrument 1 10/30/2018 12:01:00 PM

Data File D:\DATA\XZC\XZC-KET0 ACID\XZC-ACID-1029-1 2018-10-29 20-45-25\034-0601.D

Sample Name: XZC-acid-pF

Data File E:\DATA\XZC\DATA-2\XZC-20180314-2 2018-03-14 14-46-08\053-1001.D Sample Name: XZC-pCl-RAC-2 -----Acq. Operator : SYSTEM Acq. Instrument : 1260HPLC-VWD Seq. Line : 10 Location : Vial 53 Injection Date : 3/14/2018 7:24:35 PM Inj: 1 Inj Volume : 2.000 µl Acg. Method : E:\DATA\XZC\DATA-2\XZC-20180314-2 2018-03-14 14-46-08\VWD-IB(1-6)-95-5-1ML-2UL-210NM-40MIN.M Last changed : 3/14/2018 3:52:29 PM by SYSTEM Analysis Method : E:\DATA\XZC\DATA-2\XZC-20180314-2 2018-03-14 14-46-08\VWD-IB(1-6)-95-5-1ML-2UL-210NM-40MIN.M (Sequence Method) Last changed : 4/8/2019 11:00:51 AM by SYSTEM (modified after loading) Additional Info : Peak(s) manually integrated VWD1A, Wavelength=210 nm (E:\DATAVZC\DATA-2VZC-2018031422018-03-1414-46-08'053-1001.D) mAU 175 \cap 0 150 · CI 125 -100 -R SFI SALA 75 · **5**0 · 25 -0 <u>30</u> 5 10 15 25 20 min Area Percent Report Sorted By Signal : Multiplier 1.0000 : 1.0000 Dilution : Do not use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=210 nm Peak RetTime Type Width Area Height Area [min] [mAU*s] [mAU] * # [min] " [min] [min] [mao"3] [mao] " 1 29.822 MF 0.6802 2671.78833 65.46633 45.3150 2 30.809 FM 0.7901 3224.25220 68.01778 54.6850 5896.04053 133.48411 Totals : *** End of Report *** Page 1 of 1 1260HPLC-VWD 4/8/2019 11:01:09 AM SYSTEM

Data File D:\DATA\XZC\XZC-KET0 ACID\XZC-ACID-1029-1 2018-10-29 20-45-25\023-1201.D Sample Name: XZC-acid-pCl _____ Acq. Operator : Seq. Line : 12 Acq. Instrument : Instrument 2 Location : Vial 23 Injection Date : 10/30/2018 2:07:48 AM Inj: l Inj Volume : 5.000 µl Acq. Method : D:\DATA\XZC\XZC-KET0 ACID\XZC-ACID-1029-1 2018-10-29 20-45-25\DAD-IB(1-2)-95-5-1ML-5UL-210-35MIN.M Last changed : 10/29/2018 9:14:44 PM Analysis Method : D:\METHOD\LWD\DAD-AD(1-6)-95-5-1ML-3UL-ALL-10MIN.M Last changed : 10/30/2018 12:04:06 PM (modified after loading) Additional Info : Peak(s) manually integrated DAD1 C, Sig=210 4 Ref=off (DADATAWZCWZC-KETO ACIDWZC-ACID-1029-1 2018-10-29 20-45-25'023-1201.D) mAU .0 250 28.008 CI 200 150 100 50 Û 28 32 34 22 26 30 24 min -----Area Percent Report ------Sorted Bv : Signal Multiplier : : 1.0000 1.0000 Dilution Use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 C, Sig=210,4 Ref=off Peak RetTime Type Width Height Area Area # [min] [min] [mAU*s] [mAU] % 1 28.008 BB 0.7630 1.08725e4 208.29680 100.0000 1.08725e4 208.29680 Totals :

Instrument 2 10/30/2018 12:04:09 PM

Data File E:\DATA\XZC\DATA-2\XZC-20180314-2 2018-03-14 14-46-08\054-1101.D Sample Name: XZC-pBr-RAC-2 -----Acq. Operator : SYSTEM Acq. Instrument : 1260HPLC-VWD Seq. Line : 11 Location : Vial 54 Injection Date : 3/14/2018 8:05:19 PM Inj: 1 Inj Volume : 2.000 µl : E:\DATA\XZC\DATA-2\XZC-20180314-2 2018-03-14 14-46-08\VWD-IB(1-6)-95-5-Acg. Method 1ML-2UL-210NM-40MIN.M Last changed : 3/14/2018 3:52:29 PM by SYSTEM Analysis Method : E:\DATA\XZC\DATA-2\XZC-20180314-2 2018-03-14 14-46-08\VWD-IB(1-6)-95-5-1ML-2UL-210NM-40MIN.M (Sequence Method) Last changed : 4/8/2019 11:04:40 AM by SYSTEM (modified after loading) Additional Info : Peak(s) manually integrated VWD1A, Wavelength=210 nm (E:\DATAVZC\DATA-2VZC-201803142201803141446-080541101.D) mAU 140 Ο 120 Rr 100 80 -60 32,845 31,805 40 20 o - 20 5 10 15 20 25 30 зŚ min Area Percent Report Sorted By Signal : Multiplier 1.0000 : 1.0000 Dilution : Do not use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=210 nm Peak RetTime Type Width Area Height Area * [min] [mAU*s] [mAU] # [min] # [min] [min] [mA0^3] [mA0] % 1 31.805 BV 0.6424 1402.03259 33.69883 43.3821 2 32.845 VB 0.7539 1829.79211 35.40828 56.6179 3231.82471 69.10712 Totals : *** End of Report *** Page 1 of 1 1260HPLC-VWD 4/8/2019 11:04:46 AM SYSTEM

Data File D:\DATA\XZC\XZC-KET0 ACID\XZC-ACID-1029-1 2018-10-29 20-45-25\024-1301.D Sample Name: XZC-acid-pBr _____ Acq. Operator : Seq. Line : 13 Acq. Instrument : Instrument 2 Location : Vial 24 Injection Date : 10/30/2018 2:43:47 AM Inj: l Inj Volume : 5.000 µl Acq. Method : D:\DATA\XZC\XZC-KET0 ACID\XZC-ACID-1029-1 2018-10-29 20-45-25\DAD-IB(1-2)-95-5-1ML-5UL-210-35MIN.M Last changed : 10/29/2018 9:14:44 PM Analysis Method : D:\METHOD\XZC\VWD-OD(1-6)-90-10-1ML-5UL-210NM-40MIN.M Last changed : 10/30/2018 2:17:17 PM (modified after loading) Additional Info : Peak(s) manually integrated DAD1 C, Sig=210 4 Ref=off (DADATA%ZC%ZC-KETO ACID%ZC-ACID-1029-1 2018-10-29 20-46-25/024-1301.D) mAU .O 800 Br 600 29.675 400 200 ۵ 28 30 27 29 31 32 33 34 26 min -----Area Percent Report ------Sorted Bv : Signal Multiplier : 1.0000 : 1.0000 Dilution Use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 C, Sig=210,4 Ref=off Peak RetTime Type Width Height Area Area # [min] [min] [mAU*s] [mAU] % * 1 29.675 BB 0.8708 2.53935e4 421.53577 100.0000 2.53935e4 421.53577 Totals :

Instrument 1 10/30/2018 2:17:21 PM

Data File D:\DATA\XZC\XZC-DATA-3\XZC-2019-0303-1 2019-03-03 15-04-35\001-0301.D Sample Name: xzc-pMe-rac



Instrument 1 3/3/2019 7:00:07 PM

Data File D:\DATA\XZC\XZC-DATA-3\XZC-2019-0227-2 2019-02-27 20-35-46\042-0401.D Sample Name: xzc-pMe



Instrument 1 3/3/2019 6:58:11 PM

Data File D:\DATA\XZC\XZC-DATA-3\XZC-2019-0306-1 2019-03-06 19-38-20\031-0201.D Sample Name: XZC-mMe-RAC



Instrument 1 3/7/2019 2:20:01 PM

Data File D:\DATA\XZC\XZC-DATA-3\XZC-2019-0306-1 2019-03-06 19-38-20\032-0301.D Sample Name: XZC-mMe



Instrument 1 3/7/2019 2:22:34 PM



Instrument 1 3/13/2019 3:29:18 PM

Data File D:\DATA\XZC\XZC-DATA-3\XZC-2019-0303-3 2019-03-03 19-19-48\012-0301.D Sample Name: XZC-hex

_____ Acq. Operator : Seq. Line : 3 Acq. Instrument : Instrument 1 Location : Vial 12 Injection Date : 3/3/2019 8:03:25 PM Inj: l Inj Volume : 5.000 µl Acq. Method : D:\DATA\XZC\XZC-DATA-3\XZC-2019-0303-3 2019-03-03 19-19-48\VWD-AD(1-2)-95-5 -1ML-5UL-210NM-30MIN.M Last changed : 3/2/2019 8:06:09 PM Analysis Method : D:\METHOD\GUAN YUQING\LONGJIAO\DAD-0J(1-6)-99-1-0.5ML-5UL-ALL-60MIN.M Last changed : 3/13/2019 3:30:05 PM (modified after loading) Additional Info : Peak(s) manually integrated W/D1_A, Wavelength=210 nm (D:\DATAXZCWZC-DATA3XZC-2019-0303-3 2019-03-03 19-19-48/012-0301.D) mAU 1400 - \cap 1200 10.00 800 600 19.625 400 ,3878¹ 200 23085 Û 15 10 Ś. 20 25 -----Area Percent Report ------Sorted Bv : Signal Multiplier : 1.0000 : 1.0000 Dilution Use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=210 nm Peak RetTime Type Width Height Area Area 1 19.625 BB 0.4099 9240.15527 339.09265 99.6035 2 23.085 MM 0.3791 36.78712 1.46427 0.3965 Totals : 9276.94239 340.55692

Instrument 2 3/13/2019 3:30:19 PM



Instrument 1 3/3/2019 3:41:12 PM



Instrument 1 3/3/2019 3:45:50 PM



Instrument 1 4/8/2019 4:53:09 PM

Data File D:\DATA\GUAN YUQING\LJ-113-1\LJ-113-1(2) 2019-03-27 15-58-33\012-1701.D Sample Name: xzc-2MeO _____ Acq. Operator : Seq. Line : 17 Acq. Instrument : Instrument 1 Location : Vial 12 Injection Date : 3/27/2019 11:03:35 PM Inj: l Inj Volume : 5.000 µl : D:\DATA\GUAN YUQING\LJ-113-1\LJ-113-1(2) 2019-03-27 15-58-33\VWD-AD(1-2)-95 Acq. Method -5-1ML-5UL-210NM-40MIN.M Last changed : 7/10/2018 8:21:32 PM Analysis Method : D:\METHOD\GUAN YUQING\LONGJIAO\DAD-OD(1-2)-99-1-1ML-5UL-ALL-60MIN.M Last changed : 3/28/2019 9:07:09 AM (modified after loading) Additional Info : Peak(s) manually integrated W/D1 A, Wavelength=210 nm (D:\DATAGUAN YUQING\LJ-113-1\LJ-113-1(2) 2019-03-27 15-58-33\012-1701.D) mAU Ο 200 MeO 150 1.^{5210,15} 100 4 50 Û 25 15 20 30 10 _____ Area Percent Report ------Sorted Bv : Signal : Multiplier 1.0000 1.0000 Dilution Use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=210 nm Peak RetTime Type Width Height Area Area # [min] [min] [mAU*s] [mAU] % 1 28.404 MM 0.6931 3210.74707 77.20502 100.0000 3210.74707 77.20502 Totals :

Instrument 2 3/28/2019 9:08:12 AM



Instrument 1 3/11/2019 3:05:55 PM

Data File D:\DATA\XZC\XZC-DATA-3\XZC-2019-0310-1 2019-03-10 11-01-33\042-0401.D Sample Name: XZC-4Me0 _____ Acq. Operator : Seq. Line : 4 Acq. Instrument : Instrument 1 Location : Vial 42 Injection Date : 3/10/2019 12:06:31 PM Inj: l Inj Volume : 5.000 µl : D:\DATA\XZC\XZC-DATA-3\XZC-2019-0310-1 2019-03-10 11-01-33\VWD-AD(1-2)-95-5 Acq. Method -1ML-5UL-210NM-40MIN.M Last changed : 7/10/2018 8:21:32 PM Analysis Method : D:\METHOD\GUAN YUQING\LONGJIA0\VWD-OD(1-6)-99-1-0.5ML-5UL-254NM-60MIN.M Last changed : 3/11/2019 3:07:15 PM (modified after loading) Additional Info : Peak(s) manually integrated W/D1_A, Wavelength=210 nm (D\DATAXZCWZC-DATA3\XZC-2019-0310-1 2019-03-10 11-01-33\042-0401.D) mAU С 800 600 ÓMe 400 25.351 200 ٥ 20 10 15 з'n -----Area Percent Report ------Sorted Bv : Signal Multiplier : 1.0000 : 1.0000 Dilution Use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=210 nm Peak RetTime Type Width Height Area Area # [min] [min] [mAU*s] [mAU] * 1 25.351 BB 0.5319 1.08740e4 308.75864 99.6236 2 37.885 MM 0.6678 41.07934 1.02520 0.3764 Totals : 1.09150e4 309.78384

Instrument 1 3/11/2019 3:07:20 PM

Data File D:\DATA\LG\201910\YCC-ET-AD 2019-10-28 14-42-55\002-0901.D Sample Name: CYZ-RAC-2



Instrument 1 10/29/2019 8:37:52 AM



Instrument 1 3/13/2019 3:19:26 PM

Data File D:\DATA\XZC\XZC-DATA-3\XZC-2019-0310-1 2019-03-10 12-50-51\002-0601.D Sample Name: XZC-hep-rac-1 _____ Acq. Operator : Seq. Line : 6 Acq. Instrument : Instrument 1 Location : Vial 2 Injection Date : 3/10/2019 3:08:02 PM Inj: l Inj Volume : 5.000 µl Acq. Method : D:\DATA\XZC\XZC-DATA-3\XZC-2019-0310-1 2019-03-10 12-50-51\VWD-AD(1-2)-95-5 -1ML-5UL-210NM-40MIN.M Last changed : 7/10/2018 8:21:32 PM Analysis Method : D:\METHOD\GUAN YUQING\LONGJIA0\VWD-OD(1-6)-99-1-0.5ML-5UL-254NM-60MIN.M Last changed : 3/11/2019 3:10:27 PM (modified after loading) Additional Info : Peak(s) manually integrated W/D1_A, Wavelength=210 nm (D\DATAXZCWZC-DATA3\XZC-2019-0310-1 2019-03-10 12-50-51\002-0601.D) mAU 800 600 400 13.351 200 0 10 12 _____ Area Percent Report ------Sorted Bv : Signal Multiplier : : 1.0000 1.0000 Dilution Use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=210 nm Peak RetTime Type Width Height Area Area # [min] [min] [mAU*s] [mAU] % 1 12.226 BB 0.2426 4202.47021 262.70895 50.1776 2 13.351 BV 0.3258 4172.71436 198.83009 49.8224 Totals : 8375.18457 461.53905

Instrument 1 3/11/2019 3:10:33 PM

Data File D:\DATA\LYH\LYH-4-674\LYH-4-674-2-P-TBU 2019-03-12 21-14-55\004-0701.D Sample Name: XZC-hep _____ Acq. Operator : Seq. Line : 7 Acq. Instrument : Instrument 1 Location : Vial 4 Injection Date : 3/13/2019 12:40:44 AM Inj: 1 Inj Volume : 5.000 µl Acq. Method : D:\DATA\LYH\LYH-4-674\LYH-4-674-2-P-TBU 2019-03-12 21-14-55\VWD-AD(1-2)-95-5-1ML-5UL-210NM-40MIN.M Last changed : 7/10/2018 8:21:32 PM Analysis Method : D:\METHOD\ZX\VWD-AD(1-2)-90-10-0.5ML-3UL-254NM-60MIN.M Last changed : 3/13/2019 3:10:10 PM (modified after loading) Additional Info : Peak(s) manually integrated W/D1_A, Wavelength=210 nm (D:\DATALYH\LYH-4674\LYH-4674\LYH-46742-P-TBU 2019-03-12 21-14-55'004-0701.D) mAU 800 600 400 13 22 4 200 (P.S. ₫ ŝ 0 10 12 _____ Area Percent Report ------Sorted Bv : Signal Multiplier : : 1.0000 1.0000 Dilution Use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=210 nm Peak RetTime Type Width Height Area Area # [min] [min] [mAU*s] [mAU] % 1 12.101 MM 0.2222 129.35524 9.70152 2.0625 2 13.224 BV 0.2909 6142.51025 321.56558 97.9375 Totals : 6271.86549 331.26710

Instrument 1 3/13/2019 3:12:01 PM