

**Asymmetric organocatalyzed reaction sequence to  
construct bridged bicyclic acetals via multicatalytic process  
involving iminium catalysis and anion-binding catalysis**

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**Supporting Information**

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## A. General information

The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were recorded at 500 MHz or 600 MHz for  $^1\text{H}$  and at 125 MHz for  $^{13}\text{C}$ . The chemical shifts ( $\delta$ ) for  $^1\text{H}$  and  $^{13}\text{C}$  are given in ppm relative to residual signals of the solvents ( $\text{CDCl}_3$  at 7.26 ppm  $^1\text{H}$  NMR, 77.16 ppm  $^{13}\text{C}$  NMR). Coupling constants are given in Hz. The following abbreviations are used to indicate the multiplicity: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet. High-resolution mass spectra (HRMS) were obtained from the Waters Q-ToF Ultima Global. X-ray data were obtained from Zhongke chemical technology service center. Optical rotations are reported as follows:  $[\alpha]_{\text{D}}^{20}$  (c in g per 100 mL, solvent:  $\text{CHCl}_3$ ).

Note: NMR signals containing common solvent contaminants were list.  $\text{H}_2\text{O}$  in  $\text{CDCl}_3$  at 1.56 ppm  $^1\text{H}$  NMR; Ethyl acetate in  $\text{CDCl}_3$  at 2.05 (s), 4.12 (q), 1.26 (t) ppm  $^1\text{H}$  NMR; Dichloromethane in  $\text{CDCl}_3$  at 5.30 (s) ppm  $^1\text{H}$  NMR; Grease in  $\text{CDCl}_3$  at 0.86 (m), 1.26 (br, s) ppm  $^1\text{H}$  NMR; *n*-hexane in  $\text{CDCl}_3$  at 0.88 (t), 1.26 (m) ppm  $^1\text{H}$  NMR.

All the reactions were set up under air and using freshly distilled solvents, without any precautions to exclude moisture, unless otherwise noted open air chemistry on the bench-top. Chromatographic purification of products was accomplished using force-flow chromatography (FC) on silica gel (300-400 mesh). For thin layer chromatography (TLC) analysis throughout this work, Merck pre-coated TLC plates (silica gel 60 GF254, 0.25 mm) were used, using UV light as the visualizing agent and an phosphomolybdic acid or basic aqueous potassium permanganate ( $\text{KMnO}_4$ ) as stain developing solutions. Organic solutions were concentrated under reduced pressure on a Büchi rotary evaporator.

HPLC analyses on chiral stationary phase were performed on a Hitachi Chromaste. Daicel Chiralpak IA, IC, ID, or AD columns with *n*-hexane/*i*-PrOH as the eluent were used. HPLC traces were compared to racemic samples which prepared by mixture of two enantiomeric final products obtained using (*S*) and (*R*) catalysts.

Commercial reagents and solvents were purchased from Sigma Aldrich, Fluka, and Alfa Aesar used as received, without further purification. The 2-hydroxy cinnamaldehydes **1** were prepared according to literature procedures.<sup>1</sup> 4-

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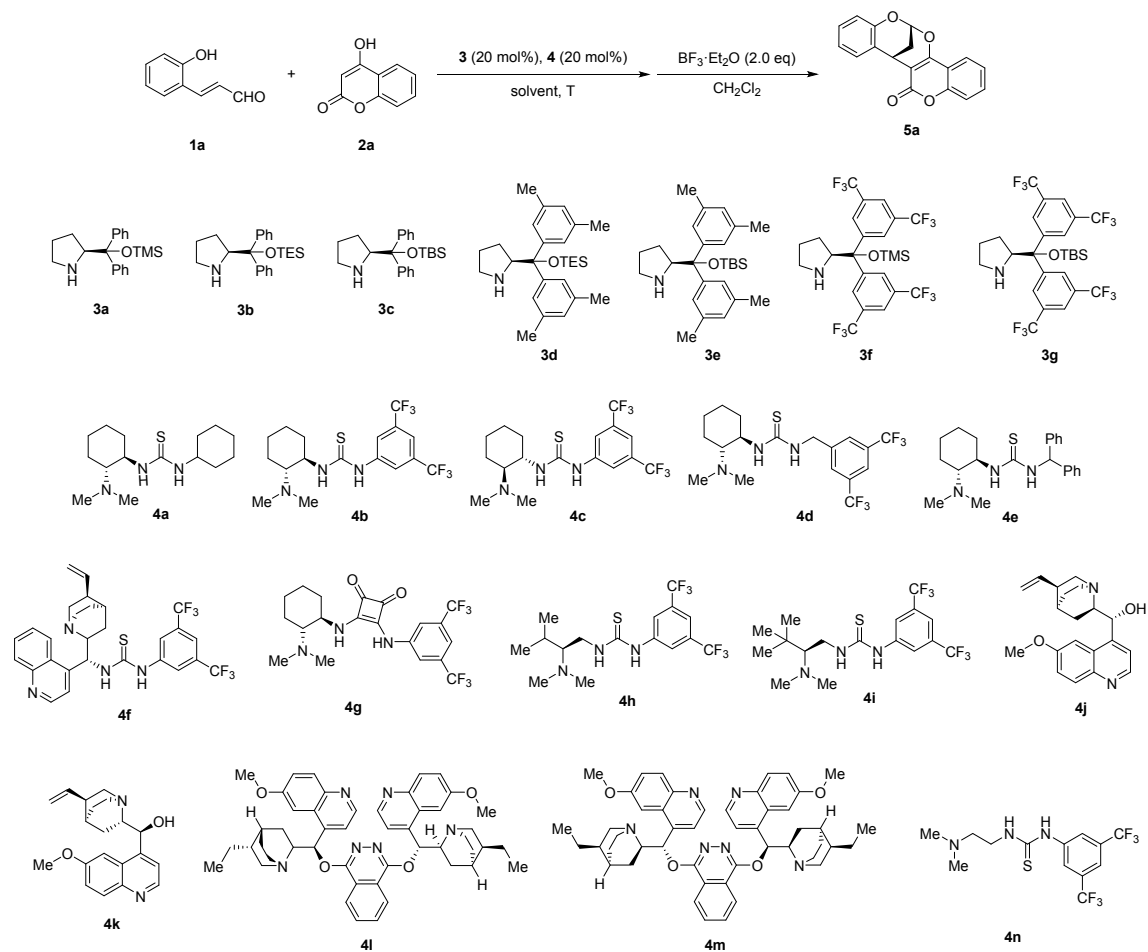
<sup>1</sup> Chen, Y.-H.; Sun, X.-L.; Guan, H.-S.; Liu, Y.-K. *J. Org. Chem.* **2017**, *82*, 4774.

Hydroxycoumarins **2** were prepared according to literature procedures.<sup>2</sup>

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<sup>2</sup> Nolan, K. A.; Doncaster, J. R.; Dunstan, M. S.; Scott, K. A.; Frenkel, A. D.; Siegel, D.; Ross, D.; Barnes, J.; Levy, C.; Leys, D.; Whitehead, R. C.; Stratford, I. J.; Bryce, R. A., *J. Med. Chem.* **2009**, *52*, 7142.

## B. Optimization of the reaction conditions



| entry <sup>[b]</sup> | <b>3</b>  | <b>4</b> | solvent                  | T(°C) | t (h) <sup>[c]</sup> | yield (%) <sup>[d]</sup> | ee (%) <sup>[e]</sup> |
|----------------------|-----------|----------|--------------------------|-------|----------------------|--------------------------|-----------------------|
| 1                    | <b>3a</b> | -        | $\text{CH}_2\text{Cl}_2$ | 25    | 24                   | 45                       | 28                    |
| 2                    | <b>3b</b> | -        | $\text{CH}_2\text{Cl}_2$ | 25    | 24                   | 19                       | 23                    |
| 3                    | <b>3c</b> | -        | $\text{CH}_2\text{Cl}_2$ | 25    | 24                   | 17                       | 30                    |
| 4                    | <b>3d</b> | -        | $\text{CH}_2\text{Cl}_2$ | 25    | 24                   | 23                       | 3                     |
| 5                    | <b>3e</b> | -        | $\text{CH}_2\text{Cl}_2$ | 25    | 24                   | 18                       | 5                     |
| 6                    | <b>3f</b> | -        | $\text{CH}_2\text{Cl}_2$ | 25    | 5                    | 41                       | 77                    |
| 7                    | <b>3g</b> | -        | $\text{CH}_2\text{Cl}_2$ | 25    | 12                   | 27                       | 86                    |
| 8 <sup>[f]</sup>     | <b>3f</b> | -        | $\text{CH}_2\text{Cl}_2$ | 25    | 5                    | 39                       | 71                    |

|           |           |           |                                 |    |    |    |    |
|-----------|-----------|-----------|---------------------------------|----|----|----|----|
| 9 [g]     | <b>3f</b> | -         | CH <sub>2</sub> Cl <sub>2</sub> | 25 | 5  | 64 | 77 |
| 10        | <b>3f</b> | -         | CH <sub>2</sub> Cl <sub>2</sub> | 25 | 5  | 51 | 79 |
| 11        | <b>3f</b> | -         | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 72 | 64 | 95 |
| 12 [g]    | <b>3f</b> | -         | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 20 | 85 | 90 |
| 13 [f]    | <b>3f</b> | <b>4a</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 18 | 73 | 89 |
| 14        | <b>3f</b> | <b>4a</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 18 | 70 | 93 |
| 15        | <b>3f</b> | <b>4b</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 18 | 75 | 97 |
| 16        | <b>3f</b> | <b>4c</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 18 | 62 | 96 |
| 17        | <b>3f</b> | <b>4d</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 18 | 55 | 96 |
| 18        | <b>3f</b> | <b>4e</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 18 | 65 | 97 |
| 19        | <b>3f</b> | <b>4f</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 30 | 75 | 95 |
| 20        | <b>3f</b> | <b>4g</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 30 | 80 | 92 |
| 21        | <b>3f</b> | <b>4h</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 18 | 62 | 96 |
| 22        | <b>3f</b> | <b>4i</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 24 | 68 | 94 |
| 23        | <b>3f</b> | <b>4j</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 24 | 43 | 94 |
| 24        | <b>3f</b> | <b>4k</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 24 | 46 | 95 |
| 25        | <b>3f</b> | <b>4l</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 24 | 68 | 90 |
| 26        | <b>3f</b> | <b>4m</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 24 | 31 | 80 |
| 27        | <b>3f</b> | <b>4b</b> | DCE                             | 0  | 36 | 76 | 96 |
| 28        | <b>3f</b> | <b>4b</b> | CHCl <sub>3</sub>               | 0  | 48 | 73 | 96 |
| 29        | <b>3f</b> | <b>4b</b> | toluene                         | 0  | 48 | 28 | 77 |
| 30        | <b>3f</b> | <b>4b</b> | xylene                          | 0  | 48 | 25 | 80 |
| 31 [h]    | <b>3f</b> | <b>4b</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 25 | 73 | 98 |
| 32 [i]    | <b>3f</b> | <b>4b</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 48 | 68 | 99 |
| 33 [l][h] | <b>3f</b> | <b>4b</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0  | 24 | 68 | 73 |

|                       |           |           |                                 |   |      |    |    |
|-----------------------|-----------|-----------|---------------------------------|---|------|----|----|
| 34 <sup>[h]</sup>     | <b>3f</b> | <b>4n</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0 | 22   | 68 | 95 |
| 35 <sup>[h] [i]</sup> | <b>3f</b> | <b>4b</b> | CH <sub>2</sub> Cl <sub>2</sub> | 0 | >120 | 68 | 86 |

[a] Unless otherwise specified, all reactions were carried out using **1a** (0.10 mmol, 1.0 equiv), **2a** (0.12 mmol, 1.2 equiv) in solvent (0.2 mL) with cat. **3** (20 mol %) and co-cat. **4** (20 mol %) at 0 °C. After full conversion of **1a**, 0.3 mL solvent was added to the solution, then BF<sub>3</sub>•Et<sub>2</sub>O (2.0 eq) was added in one pot. After workup, the mixture was purified by flash chromatography on silica gel to afford **5a**. [b] Entry 1-9 were performed by two steps After full conversion of **1a**, the crude product was obtained by flash chromatography on silica gel. Then the crude product was dissolved in 0.5 mL CH<sub>2</sub>Cl<sub>2</sub>, 1.0 eq BF<sub>3</sub>•Et<sub>2</sub>O was added to the solution and allowed to stir at 25 °C. [c] For the first step. [d] Isolated yield of **5a**. [e] Determined by HPLC analyses of isolated compound **5a** on chiral stationary phases. [f] Use BA (0.2 eq) as additive. [g] Use DIPEA (0.2 eq) as additive. [h] Reaction in solvent (0.5 mL). [i] Reaction in solvent (1.0 mL). [j] Use **3f** (10 mmol %) and **4b** (10 mmol %).

TMS = trimethylsilyl

DCE = 1,2-dichloroethane

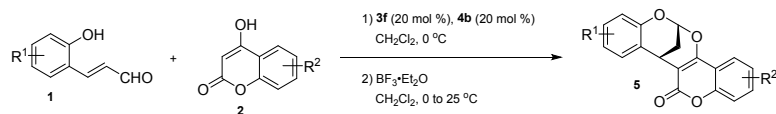
TES = triethylsilyl

BA = benzoic acid

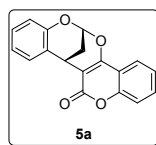
TBS = (1,1-dimethylethyl)dimethylsilyl

DIPEA = N,N-diisopropylethylamine

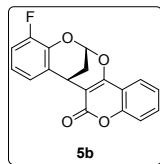
## C. Scope of the reaction



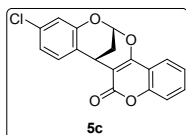
**General procedure:** A glass vial equipped with a magnetic stirring bar was charged with **1** (0.10 mmol, 1.0 equiv), catalyst **3f** (0.02 mmol, 0.2 equiv) and catalyst **4b** (0.02 mmol, 0.2 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (0.5 mL) and the resulting solution was stirred for about 10 min at 0 °C. Then **2** (0.12 mmol, 1.2 equiv) was added. After full conversion of first step, BF<sub>3</sub>•Et<sub>2</sub>O (2.0 eq) was added to the reaction mixture in situ and the reaction mixture was allowed to stir at 25 °C. After completion of the reaction, the reaction mixture was purified by flash chromatography on silica gel to give product **5** for NMR and HPLC analysis. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >20:1.



**5a** was obtained as a white solid 21.2 mg in 73% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.81 (d, *J* = 7.8 Hz, 1H), 7.52-7.46 (m, 2H), 7.30-7.23 (m, 2H), 7.18 – 7.11 (m, 1H), 6.94 (t, *J* = 8.0 Hz, 2H), 6.40 (d, *J* = 1.6 Hz, 1H), 4.29 (s, 1H), 2.38 – 2.31 (m, 1H), 2.28 – 2.22 (m, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.7, 157.8, 152.5, 150.5, 132.1, 128.6, 128.5, 125.5, 124.2, 123.0, 122.1, 116.8, 116.4, 115.1, 106.5, 93.2, 25.5, 24.6 ppm. **HRMS:** [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>13</sub>O<sub>4</sub><sup>+</sup> 293.0808; found: 293.0809. **[α]<sub>D</sub><sup>20</sup>** -67.13 (*c* = 0.32 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 16.01 min, *t*<sub>minor</sub> = 18.69 min, **ee** = **98%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >20:1.

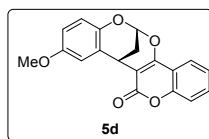


**5b** was obtained as a white solid 15 mg in 48% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.84-7.78 (m, 1H), 7.57 – 7.47 (m, 1H), 7.31-7.24 (m, 3H), 6.99 – 6.91 (m, 1H), 6.91-6.83 (m, 1H), 6.48 (d, *J* = 1.8 Hz, 1H), 4.33 (s, 1H), 2.39 – 2.33 (m, 1H), 2.32 – 2.25 (m, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>). **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.3, 157.5, 152.3, 151.9, 149.9, 138.6(d, *J*<sub>CF</sub> = 11.0 Hz), 132.1, 127.9, 124.1 (d, *J*<sub>CF</sub> = 163.0 Hz), 123.3(d, *J*<sub>CF</sub> = 3.5 Hz), 121.8(d, *J*<sub>CF</sub> = 7.0 Hz), 116.6, 115.2(d, *J*<sub>CF</sub> = 17.8 Hz), 114.7, 105.8, 92.6, 25.0, 24.0 (d, *J*<sub>CF</sub> = 2.7 Hz) ppm. **<sup>19</sup>F NMR** (400 MHz, CDCl<sub>3</sub>) δ -135.7 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>12</sub>FO<sub>4</sub><sup>+</sup> 311.0714; found:311.0711. **[α]<sub>D</sub><sup>20</sup>** -35.45 (*c* = 0.67 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH= 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 15.95 min, *t*<sub>minor</sub> = 15.05 min, **ee** = **99%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >**20:1**.

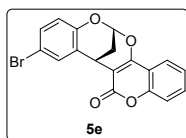


**5c** was obtained as a white solid 20 mg in 61% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.81 (d, *J* = 7.5 Hz, 1H), 7.55 – 7.47 (m, 1H), 7.41 (d, *J* = 8.1 Hz, 1H), 7.28 (d, *J* = 7.9 Hz, 2H), 6.97 – 6.88 (m, 2H), 6.38 (d, *J* = 1.5 Hz, 1H), 4.27 (s, 1H), 2.31 (d, *J* = 13.4 Hz, 1H), 2.26 (d, *J* = 13.4 Hz, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.6, 157.7, 152.5, 151.2, 133.7, 132.3, 129.3, 124.3, 124.1, 123.0, 122.4, 116.9, 116.9, 115.0, 106.2, 93.0, 25.3, 24.2 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>12</sub>ClO<sub>4</sub><sup>+</sup> 327.0419; found:327.0420. **[α]<sub>D</sub><sup>20</sup>** -34.06 (*c* = 1.67 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH= 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 12.61 min, *t*<sub>minor</sub> = 11.75 min, **ee** = **99%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR,

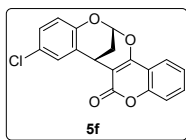
***dr* >20:1.**



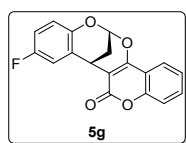
**5d** was obtained as a white solid 15.4 mg in 48% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.81 (d, *J* = 7.8 Hz, 1H), 7.54 – 7.43 (m, 1H), 7.30-7.22 (m, 2H), 7.11 (d, *J* = 7.7 Hz, 1H), 6.90 (t, *J* = 7.9 Hz, 1H), 6.77 (d, *J* = 8.1 Hz, 1H), 6.51 (d, *J* = 1.4 Hz, 1H), 4.31 (s, 1H), 3.87 (s, 3H), 2.36 (d, *J* = 13.4 Hz, 1H), 2.24 (d, *J* = 13.4 Hz, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.5, 157.7, 152.3, 147.8, 139.5, 131.9, 126.3, 124.0, 122.9, 121.9, 120.3, 116.6, 114.9, 110.5, 106.2, 92.8, 55.9, 25.0, 24.2 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>19</sub>H<sub>15</sub>O<sub>5</sub><sup>+</sup> 323.0914; found: 323.0915. [α]<sub>D</sub><sup>20</sup> -30.94 (*c* = 1.25 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 34.49 min, *t*<sub>minor</sub> = 31.84 min, **ee** = **97%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, ***dr* >20:1**.



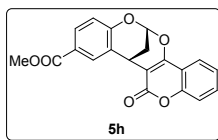
**5e** was obtained as a white solid 25.6 mg in 69% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.80 (d, *J* = 7.9 Hz, 1H), 7.61 (d, *J* = 1.8 Hz, 1H), 7.51 (dd, *J* = 8.2, 7.4 Hz, 1H), 7.32 – 7.20 (m, 3H), 6.80 (d, *J* = 8.6 Hz, 1H), 6.38 (s, 1H), 4.24 (s, 1H), 2.30 (d, *J* = 13.5 Hz, 1H), 2.24 (d, *J* = 13.5 Hz, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.3, 157.6, 152.3, 149.5, 132.1, 131.2, 130.8, 127.3, 124.1, 122.8, 118.0, 116.7, 114.7, 114.1, 105.7, 92.8, 24.9, 24.2 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>12</sub>BrO<sub>4</sub><sup>+</sup> 370.9913; found: 370.9918. [α]<sub>D</sub><sup>20</sup> 66.15 (*c* = 1.67 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 19.55 min, *t*<sub>minor</sub> = 17.85 min, **ee** = **86%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, ***dr* >20:1**.



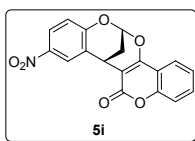
**5f** was obtained as a white solid 18.1 mg in 55% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.81 (d, *J* = 7.9 Hz, 1H), 7.51 (t, *J* = 7.8 Hz, 1H), 7.47 (d, *J* = 1.9 Hz, 1H), 7.34 – 7.23 (m, 2H), 7.10 (dd, *J* = 8.7, 1.8 Hz, 1H), 6.86 (d, *J* = 8.6 Hz, 1H), 6.38 (s, 1H), 4.25 (s, 1H), 2.31 (d, *J* = 13.5 Hz, 1H), 2.25 (d, *J* = 13.5 Hz, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.5, 157.8, 152.6, 149.2, 132.4, 128.5, 128.2, 127.0, 127.0, 124.3, 123.0, 117.7, 116.9, 115.0, 105.9, 93.1, 25.2, 24.5 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>12</sub>ClO<sub>4</sub><sup>+</sup> 327.0419; found: 327.0415. **[α]<sub>D</sub><sup>20</sup>** 32.33 (*c* = 1.5 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 18.48 min, *t*<sub>minor</sub> = 16.16 min, **ee** = **91%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >**20:1**.



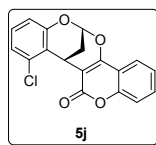
**5g** was obtained as a white solid 19.6 mg in 63% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.83 – 7.74 (m, 1H), 7.54 – 7.45 (m, 1H), 7.30–7.23 (m, 2H), 7.20 (dd, *J* = 8.2, 2.7 Hz, 1H), 6.37 (d, *J* = 1.7 Hz, 1H), 4.24 (d, *J* = 1.6 Hz, 1H), 2.31 (dt, *J* = 13.4, 2.2 Hz, 1H), 2.24 (dt, *J* = 13.5, 2.5 Hz, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.6, 158.5, 157.9, 156.6, 152.5, 146.5, 132.3, 126.7(d, *J*<sub>CF</sub> = 7.8 Hz), 124.3, 123.0, 117.3(d, *J*<sub>CF</sub> = 8.1 Hz), 116.9, 115.2(d, *J*<sub>CF</sub> = 47.6 Hz), 115.0, 105.9, 93.2, 25.2, 24.7 ppm. **<sup>19</sup>F NMR** (400 MHz, CDCl<sub>3</sub>) δ -121.6 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>12</sub>FO<sub>4</sub><sup>+</sup> 311.0714; found: 311.0716. **[α]<sub>D</sub><sup>20</sup>** -39.76 (*c* = 1.58 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 16.72 min, *t*<sub>minor</sub> = 15.53 min, **ee** = **97%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >**20:1**.



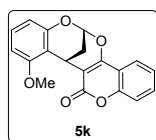
**5h** was obtained as a white solid 21.6 mg in 59% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 3/1). **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.16 (d, *J* = 2.1 Hz, 1H), 7.86 (dd, *J* = 8.5, 2.1 Hz, 1H), 7.81 (dd, *J* = 8.2, 1.5 Hz, 1H), 7.53 – 7.49 (m, 1H), 7.32 – 7.26 (m, 2H), 6.96 (d, *J* = 8.5 Hz, 1H), 6.43 (q, *J* = 1.9 Hz, 1H), 4.34 (d, *J* = 1.8 Hz, 1H), 3.88 (s, 3H), 2.37 – 2.25 (m, 2H) ppm. **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 166.5, 161.3, 157.3, 154.3, 152.4, 132.2, 130.3, 130.0, 125.3, 124.1, 124.1, 122.8, 116.7, 116.4, 114.7, 105.9, 92.9, 52.0, 25.1, 24.4 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>20</sub>H<sub>15</sub>O<sub>6</sub><sup>+</sup> 351.0863; found: 351.0861. [α]<sub>D</sub><sup>20</sup> 5.76 (*c* = 1.67 in CHCl<sub>3</sub>) The enantiomeric excess was determined by HPLC using Chiralpak IA column [*n*-hexane/*i*-PrOH = 70/30, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 25.15 min, *t*<sub>minor</sub> = 11.96 min, **ee** = **97%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >**20:1**.



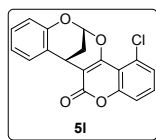
**5i** was obtained as a white solid 12 mg in 36% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 3/1). **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.41 (d, *J* = 2.7 Hz, 1H), 8.06 (dd, *J* = 9.0, 2.7 Hz, 1H), 7.82 (dd, *J* = 8.2, 1.5 Hz, 1H), 7.59 – 7.50 (m, 1H), 7.30 (dd, *J* = 7.7, 6.0 Hz, 2H), 7.03 (d, *J* = 9.0 Hz, 1H), 6.47 (d, *J* = 1.9 Hz, 1H), 4.40 (d, *J* = 1.8 Hz, 1H), 2.40 – 2.29 (m, 2H) ppm. **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 157.3, 155.7, 152.4, 142.3, 132.5, 126.2, 124.4, 124.3, 124.2, 122.8, 116.9, 116.9, 114.5, 105.3, 92.9, 24.8, 24.4 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>12</sub>NO<sub>6</sub><sup>+</sup> 338.0659; found: 338.0656. [α]<sub>D</sub><sup>20</sup> -45.55 (*c* = 0.75 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IA column [*n*-hexane/*i*-PrOH = 65/35, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 14.11 min, *t*<sub>minor</sub> = 12.17 min, **ee** = **99%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >**20:1**.



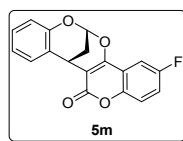
**5j** was obtained as a white solid 20 mg in 61% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.82 (dd,  $J$  = 7.9, 1.4 Hz, 1H), 7.54 – 7.47 (m, 1H), 7.31 – 7.23 (m, 2H), 7.11 – 7.01 (m, 2H), 6.85 (dd,  $J$  = 7.7, 1.5 Hz, 1H), 6.36 (dd,  $J$  = 3.7, 1.8 Hz, 1H), 4.86 (d,  $J$  = 1.6 Hz, 1H), 2.26 (dd,  $J$  = 5.0, 2.3 Hz, 2H) ppm. **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>)  $\delta$  160.8, 158.1, 152.5, 151.9, 133.2, 132.2, 128.5, 123.9, 123.7, 123.4, 122.8, 116.6, 115.1, 114.7, 105.0, 92.3, 25.5, 21.9 ppm. **HRMS:** [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>12</sub>ClO<sub>4</sub><sup>+</sup> 327.0419; found: 327.0416. The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min],  $\lambda$  = 210 nm,  $t_{major}$  = 36.76 min,  $t_{minor}$  = 31.54 min, **ee** = 6%. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >20:1.



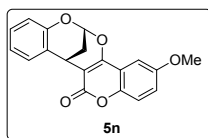
**5k** was obtained as a white solid 18 mg in 56% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 5/1). **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.86 – 7.76 (m, 1H), 7.55 – 7.41 (m, 1H), 7.26 – 7.20 (m, 2H), 7.09 (t,  $J$  = 8.2 Hz, 1H), 6.55 (dd,  $J$  = 18.6, 8.2 Hz, 2H), 6.35 (d,  $J$  = 1.8 Hz, 1H), 4.79 (d,  $J$  = 1.7 Hz, 1H), 3.90 (s, 3H), 2.29 – 2.14 (m, 2H) ppm. **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>)  $\delta$  160.9, 158.2, 157.0, 152.4, 151.2, 131.8, 128.2, 123.8, 122.7, 116.5, 115.0, 114.2, 109.0, 106.4, 104.4, 92.5, 56.1, 25.4, 18.1 ppm. **HRMS:** [M+H]<sup>+</sup> *calcd.* For C<sub>19</sub>H<sub>15</sub>O<sub>5</sub><sup>+</sup> 323.0914; found: 323.0917. The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 75/25, 1 mL/min],  $\lambda$  = 210 nm,  $t_{major}$  = 26.28 min,  $t_{minor}$  = 18.99 min, **ee** = 23%. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >20:1.



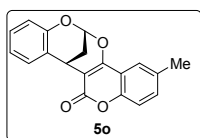
**5l** was obtained as a white solid 16.7 mg in 50% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.52 – 7.44 (m, 1H), 7.34 (t, *J* = 8.1 Hz, 1H), 7.30 – 7.24 (m, 1H), 7.22 – 7.12 (m, 2H), 6.94 (dd, *J* = 11.5, 4.3 Hz, 2H), 6.43 (dd, *J* = 3.9, 1.9 Hz, 1H), 4.32 (dd, *J* = 4.7, 2.7 Hz, 1H), 2.32 (ddd, *J* = 13.4, 2.7, 2.1 Hz, 1H), 2.25 – 2.19 (m, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 160.4, 158.0, 153.8, 150.3, 131.2, 130.8, 128.4, 128.4, 127.7, 125.0, 122.0, 116.3, 116.0, 113.2, 107.2, 92.6, 24.5, 24.5 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>12</sub>ClO<sub>4</sub><sup>+</sup> 327.0419; found: 327.0422. **[α]<sub>D</sub><sup>20</sup>** 50.98 (*c* = 1.25 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 14.89 min, *t*<sub>minor</sub> = 17.69 min, **ee** = **95%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >**20:1**.



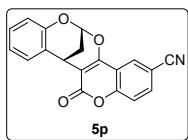
**5m** was obtained as a white solid 21 mg in 68% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.52 – 7.43 (m, 2H), 7.29 – 7.10 (m, 3H), 6.94 (t, *J* = 7.9 Hz, 2H), 6.39 (d, *J* = 1.6 Hz, 1H), 4.28 (d, *J* = 1.4 Hz, 1H), 2.38 – 2.32 (m, 1H), 2.28 – 2.21 (m, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.3, 159.8, 157.9, 157.0, 150.5, 148.6(d, *J*<sub>CF</sub> = 2.1 Hz), 128.6(d, *J*<sub>CF</sub> = 2.3 Hz), 125.2, 122.3, 119.7(d, *J*<sub>CF</sub> = 24.6 Hz), 118.5(d, *J*<sub>CF</sub> = 8.3 Hz), 116.5, 116.1(d, *J*<sub>CF</sub> = 9.0 Hz), 108.9(d, *J*<sub>CF</sub> = 25.5 Hz), 107.3, 93.3, 25.4, 24.6 ppm. **<sup>19</sup>F NMR** (400 MHz, CDCl<sub>3</sub>) δ -117.0 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>12</sub>FO<sub>4</sub><sup>+</sup> 311.0714; found: 311.0718. **[α]<sub>D</sub><sup>20</sup>** -33.54 (*c* = 1.75 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 13.83 min, *t*<sub>minor</sub> = 16.99 min, **ee** = **92%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >**20:1**.



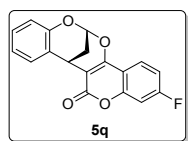
**5n** was obtained as a white solid 21 mg in 68% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.48 (d, *J* = 7.5 Hz, 1H), 7.23 (d, *J* = 2.9 Hz, 1H), 7.20 (d, *J* = 9.0 Hz, 1H), 7.17-7.12 (m, 1H), 7.07 (dd, *J* = 9.0, 3.0 Hz, 1H), 6.94 (t, *J* = 7.7 Hz, 2H), 6.40 (d, *J* = 1.8 Hz, 1H), 4.29 (d, *J* = 1.6 Hz, 1H), 3.85 (s, 3H), 2.38 – 2.31 (m, 1H), 2.27 – 2.21 (m, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.6, 157.3, 155.9, 150.3, 146.8, 128.4, 128.3, 125.3, 122.0, 120.4, 117.8, 116.2, 115.2, 106.5, 104.2, 93.0, 55.8, 25.2, 24.4 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>19</sub>H<sub>15</sub>O<sub>5</sub><sup>+</sup> 323.0914; found: 323.0912. **[α]<sub>D</sub><sup>20</sup>** 16.49 (*c* = 1.83 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 23.89 min, *t*<sub>minor</sub> = 30.70 min, **ee** = **91%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, ***dr* > 20:1**.



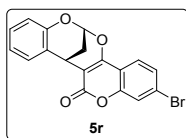
**5o** was obtained as a white solid 17.4 mg in 56% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.61 (s, 1H), 7.48 (dd, *J* = 7.4, 1.3 Hz, 1H), 7.29 (dd, *J* = 8.4, 1.6 Hz, 1H), 7.19 – 7.11 (m, 2H), 6.93 (t, *J* = 8.2 Hz, 2H), 6.38 (d, *J* = 1.9 Hz, 1H), 4.28 (d, *J* = 1.7 Hz, 1H), 2.39 (s, 3H), 2.36 – 2.31 (m, 1H), 2.26 – 2.21 (m, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.7, 157.5, 150.4, 150.3, 133.7, 132.9, 128.4, 128.2, 125.4, 122.4, 121.9, 116.4, 116.2, 114.6, 106.2, 93.0, 25.3, 24.4, 20.9 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>19</sub>H<sub>15</sub>O<sub>4</sub><sup>+</sup> 307.0965; found: 307.0969. **[α]<sub>D</sub><sup>20</sup>** -7.33 (*c* = 1.42 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 18.69 min, *t*<sub>minor</sub> = 22.43 min, **ee** = **97%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, ***dr* > 20:1**.



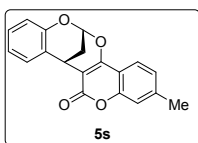
**5p** was obtained as a white solid 15.2 mg in 48% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 5/1). **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>). δ 8.16 (d, *J* = 1.9 Hz, 1H), 7.75 (dd, *J* = 8.6, 2.0 Hz, 1H), 7.45 (d, *J* = 7.4 Hz, 1H), 7.36 (d, *J* = 8.6 Hz, 1H), 7.17 (td, *J* = 7.8, 1.6 Hz, 1H), 6.96 (t, *J* = 7.5 Hz, 2H), 6.43 (d, *J* = 1.8 Hz, 1H), 4.28 (d, *J* = 1.7 Hz, 1H), 2.49 – 2.30 (m, 1H), 2.33 – 2.20 (m, 1H) ppm. **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 160.1, 156.2, 154.4, 150.2, 134.8, 128.7, 128.4, 127.9, 124.6, 122.2, 118.0, 117.7, 116.5, 115.9, 108.2, 107.9, 93.3, 25.1, 24.4 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>19</sub>H<sub>12</sub>NO<sub>4</sub><sup>+</sup> 318.0761; found: 318.0763. **[α]<sub>D</sub><sup>20</sup>** 60.17 (*c* = 0.75 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IA column [*n*-hexane/*i*-PrOH = 75/25, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 15.87 min, *t*<sub>minor</sub> = 21.07 min, **ee** = **97%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >**20:1**.



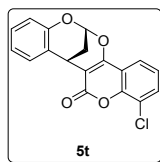
**5q** was obtained as a white solid 21.6 mg in 69% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.85 – 7.76 (m, 1H), 7.47 (d, *J* = 7.5 Hz, 1H), 7.15 (t, *J* = 7.7 Hz, 1H), 7.05–6.89 (m, 4H), 6.39 (s, 1H), 4.26 (s, 1H), 2.34 (d, *J* = 13.4 Hz, 1H), 2.24 (d, *J* = 13.4 Hz, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 165.8, 163.8, 161.5, 157.5, 153.7, 150.5, 128.6(*d*, *J*<sub>CF</sub> = 3.6 Hz), 125.4, 124.9(*d*, *J*<sub>CF</sub> = 10.3 Hz), 122.3, 116.5, 112.5(*d*, *J*<sub>CF</sub> = 22.8 Hz), 111.8, 105.5, 104.4(*d*, *J*<sub>CF</sub> = 25.7 Hz), 93.3, 25.5, 24.5 ppm. **<sup>19</sup>F NMR** (400 MHz, CDCl<sub>3</sub>) δ -105.1 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>12</sub>FO<sub>4</sub><sup>+</sup> 311.0714; found: 311.0717. **[α]<sub>D</sub><sup>20</sup>** -50.03 (*c* = 2.08 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 13.09 min, *t*<sub>minor</sub> = 16.88 min, **ee** = **82%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >**20:1**.



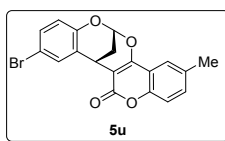
**5r** was obtained as a white solid 31 mg in 82% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.66 (dd,  $J$  = 8.2, 5.2 Hz, 1H), 7.50 – 7.42 (m, 2H), 7.38 (dd,  $J$  = 8.4, 1.3 Hz, 1H), 7.19 – 7.12 (m, 1H), 6.99 – 6.89 (m, 2H), 6.38 (d,  $J$  = 1.6 Hz, 1H), 4.26 (d,  $J$  = 1.4 Hz, 1H), 2.37 – 2.30 (m, 1H), 2.24 (dt,  $J$  = 13.4, 2.3 Hz, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>)  $\delta$  161.0, 157.4, 152.6, 150.4, 128.6, 127.7, 126.0, 125.2, 124.1, 122.3, 120.0, 116.5, 114.2, 110.2, 106.7, 93.3, 25.4, 24.6 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>12</sub>BrO<sub>4</sub><sup>+</sup> 370.9913; found: 370.9916. **[ $\alpha$ ]<sub>D</sub><sup>20</sup>** -86.13 ( $c$  = 0.41 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min],  $\lambda$  = 210 nm,  $t_{major}$  = 15.51 min,  $t_{minor}$  = 20.97 min, **ee** = **82%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >**20:1**.



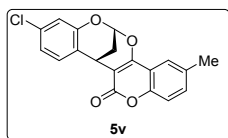
**5s** was obtained as a white solid 16.1 mg in 51% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.67 (d,  $J$  = 8.4 Hz, 1H), 7.48 (d,  $J$  = 7.5 Hz, 1H), 7.18-7.11 (m, 1H), 7.10-7.03 (m, 2H), 6.94 (t,  $J$  = 8.0 Hz, 2H), 6.37 (d,  $J$  = 1.8 Hz, 1H), 4.27 (d,  $J$  = 1.5 Hz, 1H), 2.41 (s, 3H), 2.33 (dt,  $J$  = 13.3, 2.3 Hz, 1H), 2.27 – 2.20 (m, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>)  $\delta$  161.9, 157.9, 152.6, 150.5, 143.3, 128.6, 128.4, 125.7, 125.4, 122.6, 122.1, 116.9, 116.4, 112.6, 105.5, 93.2, 25.5, 24.6, 21.9 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>19</sub>H<sub>15</sub>O<sub>4</sub><sup>+</sup> 307.0965; found: 307.0966. **[ $\alpha$ ]<sub>D</sub><sup>20</sup>** -64.90 ( $c$  = 1.25 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min],  $\lambda$  = 210 nm,  $t_{major}$  = 20.90 min,  $t_{minor}$  = 29.97 min, **ee** = **89%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >**20:1**.



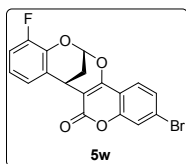
**5t** was obtained as a white solid 14.3 mg in 44% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.71 (dd, *J* = 8.0, 1.2 Hz, 1H), 7.54 (dd, *J* = 7.9, 1.2 Hz, 1H), 7.48 (dd, *J* = 7.5, 1.1 Hz, 1H), 7.22 – 7.11 (m, 2H), 6.98 – 6.90 (m, 2H), 6.40 (d, *J* = 1.7 Hz, 1H), 4.30 (d, *J* = 1.6 Hz, 1H), 2.41 – 2.32 (m, 1H), 2.25 (dt, *J* = 13.4, 2.5 Hz, 1H). **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 160.3, 157.2, 150.2, 148.0, 132.2, 128.4, 128.4, 125.0, 124.1, 122.1, 121.6, 121.3, 116.4, 116.3, 106.8, 93.1, 25.1, 24.4 ppm. **HRMS:** [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>12</sub>ClO<sub>4</sub><sup>+</sup> 327.0419; found: 327.0416. **[α]<sub>D</sub><sup>20</sup>** -104.00 (*c* = 1.17 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 20.60 min, *t*<sub>minor</sub> = 23.01 min, **ee** = **82%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** > **20:1**.



**5u** was obtained as a white solid 14 mg in 36% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.61 (dd, *J* = 5.0, 1.7 Hz, 2H), 7.31 (dd, *J* = 8.5, 1.6 Hz, 1H), 7.23 (dd, *J* = 8.6, 2.4 Hz, 1H), 7.18 (d, *J* = 8.4 Hz, 1H), 6.80 (d, *J* = 8.6 Hz, 1H), 6.37 (d, *J* = 1.8 Hz, 1H), 4.24 (d, *J* = 1.7 Hz, 1H), 2.40 (s, 3H), 2.33 – 2.26 (m, 1H), 2.26 – 2.20 (m, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.7, 157.8, 150.7, 149.8, 134.1, 133.4, 131.4, 131.1, 127.6, 122.7, 118.2, 116.7, 114.6, 114.4, 105.8, 93.0, 25.2, 24.5, 21.1 ppm. **HRMS:** [M+H]<sup>+</sup> *calcd.* For C<sub>19</sub>H<sub>14</sub>BrO<sub>4</sub><sup>+</sup> 385.0070; found: 385.0072. **[α]<sub>D</sub><sup>20</sup>** 42.69 (*c* = 1.00 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 90/10, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 43.09 min, *t*<sub>minor</sub> = 37.95 min, **ee** = **85%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** > **20:1**.

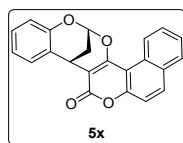


**5v** was obtained as a white solid 20 mg in 59% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.60 (s, 1H), 7.40 (d, *J* = 8.1 Hz, 1H), 7.31 (dd, *J* = 8.4, 1.5 Hz, 1H), 7.17 (d, *J* = 8.4 Hz, 1H), 6.98 – 6.88 (m, 2H), 6.37 (d, *J* = 1.8 Hz, 1H), 4.25 (d, *J* = 1.5 Hz, 1H), 2.40 (s, 3H), 2.34 – 2.27 (m, 1H), 2.27 – 2.21 (m, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.8, 157.6, 151.2, 150.7, 134.1, 133.6, 133.3, 129.3, 124.2, 122.6, 122.3, 116.8, 116.7, 114.6, 106.1, 93.0, 25.3, 24.2, 21.1 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>19</sub>H<sub>14</sub>ClO<sub>4</sub><sup>+</sup> 341.0575; found: 341.0572. **[α]<sub>D</sub><sup>20</sup>** -26.69 (*c* = 1.5 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 90/10, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 25.91 min, *t*<sub>minor</sub> = 24.59 min, **ee** = **99%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, ***dr* > 20:1**.

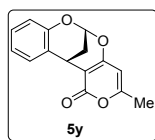


**5w** was obtained as a white solid 18 mg in 46% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.67 (d, *J* = 8.5 Hz, 1H), 7.46 (d, *J* = 1.7 Hz, 1H), 7.41 (dd, *J* = 8.5, 1.8 Hz, 1H), 7.25 (d, *J* = 7.7 Hz, 1H), 7.00 – 6.92 (m, 1H), 6.92–6.84 (m, 1H), 6.47 (dd, *J* = 3.7, 1.9 Hz, 1H), 4.31 (s, 1H), 2.39 – 2.33 (m, 1H), 2.27 (ddd, *J* = 13.6, 3.0, 2.2 Hz, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 160.7, 157.2, 152.5, 127.6, 127.6, 126.1, 124.0, 123.3(d, *J*<sub>CF</sub> = 3.7 Hz), 121.9(d, *J*<sub>CF</sub> = 6.9 Hz), 119.9, 115.3(d, *J*<sub>CF</sub> = 17.8 Hz), 113.7, 110.0, 106.1, 92.6, 24.9, 24.1 ppm. **<sup>19</sup>F NMR** (400 MHz, CDCl<sub>3</sub>) δ -135.6 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>11</sub>BrFO<sub>4</sub><sup>+</sup> 388.9819; found: 388.9820. **[α]<sub>D</sub><sup>20</sup>** -38.12 (*c* = 1.33 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IA column [*n*-hexane/*i*-PrOH = 90/10, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 19.73 min, *t*<sub>minor</sub> = 14.82 min, **ee** = **96%**. The

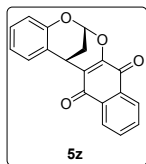
diastereomeric ratio was determined by  $^1\text{H}$  NMR, **dr** >20:1.



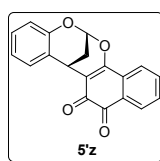
**5x** was obtained as a white solid 20 mg in 58% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 10/1).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  9.26 (d,  $J$  = 8.8 Hz, 1H), 7.92 (d,  $J$  = 9.0 Hz, 1H), 7.84 (t,  $J$  = 8.7 Hz, 1H), 7.65 (t,  $J$  = 7.8 Hz, 1H), 7.58 – 7.49 (m, 2H), 7.40 (d,  $J$  = 8.9 Hz, 1H), 7.16 (t,  $J$  = 7.7 Hz, 1H), 6.96 (t,  $J$  = 8.7 Hz, 2H), 6.60 (s, 1H), 4.41 (s, 1H), 2.40 (d,  $J$  = 13.3 Hz, 1H), 2.31 (d,  $J$  = 13.3 Hz, 1H) ppm.  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ )  $\delta$  161.3, 161.1, 153.1, 150.2, 133.7, 130.8, 128.9, 128.8, 128.5, 128.3, 128.3, 126.5, 125.8, 125.4, 122.0, 117.1, 116.2, 108.6, 106.4, 92.9, 24.7, 24.5 ppm. HRMS:  $[\text{M}+\text{H}]^+$  calcd. For  $\text{C}_{22}\text{H}_{15}\text{O}_4^+$  343.0965; found: 343.0961.  $[\alpha]_{\text{D}}^{20}$  68.06 ( $c$  = 1.33 in  $\text{CHCl}_3$ ). The enantiomeric excess was determined by HPLC using Chiralpak IC column [ $n$ -hexane/ $i$ -PrOH = 80/20, 1 mL/min],  $\lambda$  = 210 nm,  $t_{\text{major}}$  = 31.32 min,  $t_{\text{minor}}$  = 41.45 min, **ee** = 95%. The diastereomeric ratio was determined by  $^1\text{H}$  NMR, **dr** >20:1.



**5y** was obtained as a white solid 13 mg in 51% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.52 – 7.32 (m, 1H), 7.19 – 7.08 (m, 1H), 7.00 – 6.83 (m, 2H), 6.17 (d,  $J$  = 1.8 Hz, 1H), 5.83 (s, 1H), 4.11 (s, 1H), 2.23 (dt,  $J$  = 13.2, 2.3 Hz, 1H), 2.16 (s, 3H), 2.14 – 2.08 (m, 1H) ppm.  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ )  $\delta$  163.4, 162.3, 161.0, 150.5, 128.5, 128.3, 125.9, 122.1, 116.4, 103.8, 99.9, 93.0, 25.4, 24.0, 20.1 ppm. HRMS:  $[\text{M}+\text{H}]^+$  calcd. For  $\text{C}_{15}\text{H}_{13}\text{O}_4^+$  257.0808; found: 257.0804.  $[\alpha]_{\text{D}}^{20}$  -140.09 ( $c$  = 0.67 in  $\text{CHCl}_3$ ). The enantiomeric excess was determined by HPLC using Chiralpak IC column [ $n$ -hexane/ $i$ -PrOH = 80/20, 1 mL/min],  $\lambda$  = 210 nm,  $t_{\text{major}}$  = 15.41 min,  $t_{\text{minor}}$  = 18.39 min, **ee** = 72%. The diastereomeric ratio was determined by  $^1\text{H}$  NMR, **dr** >20:1.



**5z** was obtained as a yellow solid 6.7 mg in 22% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 8/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 8.07 (t, *J* = 6.7 Hz, 2H), 7.74 – 7.63 (m, 2H), 7.40 (d, *J* = 7.6 Hz, 1H), 7.14 (t, *J* = 7.7 Hz, 1H), 6.92 (dd, *J* = 14.8, 7.6 Hz, 2H), 6.38 (s, 1H), 4.45 (s, 1H), 2.35 - 2.25 (m, 1H), 2.17 – 2.10 (m, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 182.6, 179.1, 152.5, 150.7, 134.1, 133.4, 131.6, 130.8, 128.7, 128.4, 126.5, 126.2, 125.0, 124.2, 121.8, 116.5, 92.9, 24.4, 23.3 ppm. **HRMS:** [M+H]<sup>+</sup> *calcd.* For C<sub>19</sub>H<sub>13</sub>O<sub>4</sub><sup>+</sup> 305.0808; found: 305.0809. **[α]<sub>D</sub><sup>20</sup>** -192.8 (*c* = 0.50 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 45.30 min, *t*<sub>minor</sub> = 22.72 min, **ee** = **86%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >**20:1**.

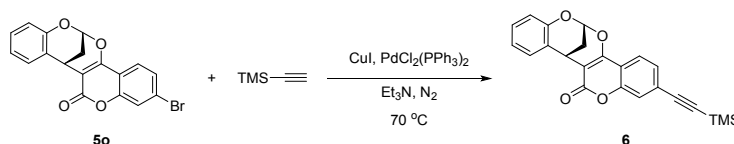


**5'z** was obtained as an orange solid 20 mg in 65% yield for two steps after column chromatography on silica gel (petroleum ether/ethyl acetate = 7/1). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 8.02 (d, *J* = 7.4 Hz, 1H), 7.84 (d, *J* = 7.6 Hz, 1H), 7.63 (t, *J* = 7.3 Hz, 1H), 7.49 (t, *J* = 7.4 Hz, 1H), 7.41 (d, *J* = 7.3 Hz, 1H), 7.14 (t, *J* = 7.5 Hz, 1H), 6.93 (d, *J* = 6.6 Hz, 2H), 6.42 (s, 1H), 4.34 (d, *J* = 31.2 Hz, 1H), 2.31 (d, *J* = 13.2 Hz, 1H), 2.22 (d, *J* = 13.2 Hz, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 179.1, 176.8, 160.5, 150.1, 135.0, 131.2, 131.1, 129.8, 129.1, 128.6, 128.3, 125.2, 124.6, 122.0, 118.4, 116.2, 93.5, 25.0, 23.0 ppm. **HRMS:** [M+H]<sup>+</sup> *calcd.* For C<sub>19</sub>H<sub>13</sub>O<sub>4</sub><sup>+</sup> 305.0808; found: 305.0806. **[α]<sub>D</sub><sup>20</sup>** -138.74 (*c* = 0.83 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH/dichloromethane = 75/20/5, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 22.81 min, *t*<sub>minor</sub> = 24.36 min, **ee** = **87%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >**20:1**.



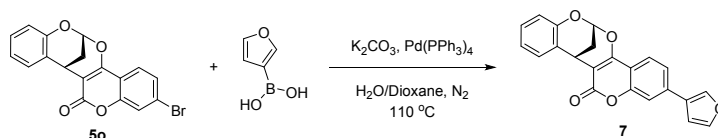
## D. Synthetic transformations

### (8*R*,14*S*)-4-((trimethylsilyl)ethynyl)-1*H*,14*H*-8,14-methanobenzo[7,8][1,3]dioxocino [5,4-*c*]chromen-1-one (**6**)



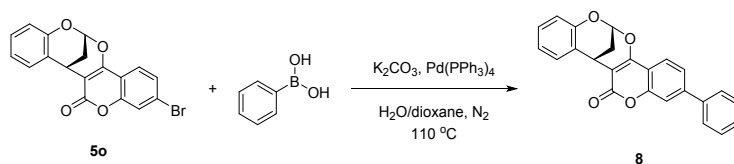
A round Schlenk flask equipped with a stirring bar was charged with **5o** (25 mg, 0.07 mmol), trimethylsilyl acetylene (15  $\mu$ L, 0.11 mmol) and 1 mL of freshly distilled dry triethylamine. A mixture of [(PPh<sub>3</sub>)<sub>2</sub>PdCl<sub>2</sub>] (1.5 mg, 0.002 mmol), and CuI (0.5 mg, 0.003 mmol) was added under N<sub>2</sub>. Then the suspension was allowed to stir at 70 °C. After completion of the reaction, it was allowed to cool down to room temperature and then was diluted with ethyl acetate, washed with saturated aqueous NH<sub>4</sub>Cl solution and brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 10:1) to give the desired product **8**. (17 mg, 65%). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.76 – 7.68 (m, 1H), 7.47 (dd, *J* = 7.5, 1.5 Hz, 1H), 7.36–7.29 (m, 2H), 7.19–7.12 (m, 1H), 6.99 – 6.90 (m, 2H), 6.43 – 6.33 (m, 1H), 4.28 (d, *J* = 1.7 Hz, 1H), 2.37 – 2.30 (m, 1H), 2.28 – 2.19 (m, 1H), 0.33 – 0.20 (m, 9H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>)  $\delta$  161.4, 157.3, 152.1, 150.5, 128.7, 128.6, 127.8, 127.0, 125.3, 122.8, 122.2, 119.9, 116.5, 115.1, 106.9, 103.5, 98.5, 93.3, 25.4, 24.6, 0.0 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>23</sub>H<sub>21</sub>O<sub>4</sub>Si<sup>+</sup> 389.1204; *found*: 389.1207. [ $\alpha$ ]<sub>D</sub><sup>20</sup> -0.25 (*c* = 1.08 in CHCl<sub>3</sub>). The diastereomeric ratio was determined by <sup>1</sup>H NMR, *dr* >20:1.

**(8*R*,14*S*)-4-(furan-3-yl)-1*H*,14*H*-8,14-methanobenzo[7,8][1,3]dioxocino[5,4-*c*]chromen-1-one (7)**



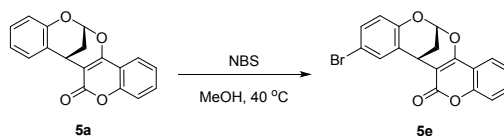
A round Schlenk flask equipped with a stirring bar was charged with **5o** (25 mg, 0.07 mmol), phenylboronic acid (9 mg, 0.08 mmol), Pd(PPh<sub>3</sub>)<sub>4</sub> (3.1 mg, 0.003 mmol), and K<sub>2</sub>CO<sub>3</sub> (20 mg, 0.15 mmol). The component solvent of dioxane and H<sub>2</sub>O (1 mL, 1:1 v/v) was added under N<sub>2</sub> at room temperature. Then the reaction mixture was moved to stir at 110 °C. After the reaction was completed, it was allowed to cool down to room temperature and was concentrated under reduced pressure. Then the residue was diluted with ethyl acetate, washed with saturated aqueous NH<sub>4</sub>Cl solution and brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 8:1) to give the desired product **7**. (13 mg, 54%). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.84 – 7.75 (m, 2H), 7.53 – 7.46 (m, 2H), 7.42 – 7.34 (m, 2H), 7.20-7.10 (m, 1H), 7.00-6.90 (m, 2H), 6.74 – 6.68 (m, 1H), 6.40 (d, *J* = 1.8 Hz, 1H), 4.29 (d, *J* = 1.7 Hz, 1H), 2.38 – 2.31 (m, 1H), 2.29 – 2.22 (m, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.5, 157.5, 152.8, 150.3, 144.2, 139.8, 136.5, 128.4, 128.3, 125.3, 125.2, 123.2, 122.0, 121.6, 116.2, 113.5, 113.2, 108.5, 105.8, 93.0, 25.3, 24.4 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>22</sub>H<sub>15</sub>O<sub>5</sub><sup>+</sup> 359.0914; *found*: 359.0916. [ $\alpha$ ]<sub>D</sub><sup>20</sup> -0.54 (*c* = 1.08 in CHCl<sub>3</sub>). The diastereomeric ratio was determined by <sup>1</sup>H NMR, *dr* >20:1.

**(8*R*,14*S*)-4-phenyl-1*H*,14*H*-8,14-methanobenzo[7,8][1,3]dioxocino[5,4-*c*]chromen-1-one (8)**



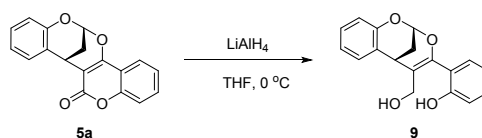
A round Schlenk flask equipped with a stirring bar was charged with **5o** (23 mg, 0.06 mmol), phenylboronic acid (9.2 mg, 0.07 mmol), Pd(PPh<sub>3</sub>)<sub>4</sub> (2.9 mg, 0.002 mmol), and K<sub>2</sub>CO<sub>3</sub> (19 mg, 0.14 mmol). The component solvent of dioxane and H<sub>2</sub>O (1 mL, 1:1 v/v) was added under N<sub>2</sub> at room temperature. Then the reaction mixture was moved to stir at 110 °C. After the reaction was completed, it was allowed to cool down to room temperature and was concentrated under reduced pressure. The residue was diluted with ethyl acetate, washed with saturated aqueous NH<sub>4</sub>Cl solution and brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and concentrated under reduced pressure. Then the residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 8:1) to give the desired product **6**. (18.4 mg, 81%). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.86 (d, *J* = 8.2 Hz, 1H), 7.60 (d, *J* = 7.4 Hz, 2H), 7.55 – 7.43 (m, 5H), 7.40 (t, *J* = 7.3 Hz, 1H), 7.16 (t, *J* = 7.2 Hz, 1H), 6.95 (t, *J* = 7.5 Hz, 2H), 6.41 (d, *J* = 1.4 Hz, 1H), 4.31 (s, 1H), 2.36 (d, *J* = 13.4 Hz, 1H), 2.26 (d, *J* = 13.4 Hz, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.8, 157.7, 152.9, 150.5, 145.3, 139.4, 129.3, 128.7, 128.6, 128.5, 127.4, 125.6, 123.3, 123.1, 122.2, 116.4, 115.0, 114.0, 106.3, 93.2, 25.5, 24.6 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>24</sub>H<sub>17</sub>O<sub>4</sub><sup>+</sup> 369.1121; found: 369.1125. **[α]<sub>D</sub><sup>20</sup>** -3.70 (*c* = 1.50 in CHCl<sub>3</sub>). The diastereomeric ratio was determined by <sup>1</sup>H NMR, ***dr* >20:1**.

**(8*R*,14*S*)-12-bromo-1*H*,14*H*-8,14-methanobenzo[7,8][1,3]dioxocino[5,4-*c*]chromen-1-one (5e)**



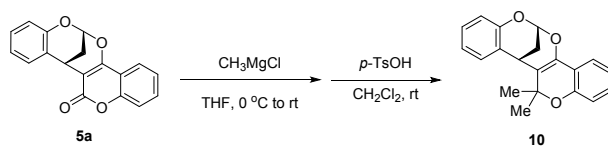
To a solution of **5a** (20 mg, 0.068 mmol) in CH<sub>3</sub>OH (1 mL) was added N-bromosuccinimide (1.2 equiv.), then the reaction mixture was allowed to stir at 40 °C. Until the starting material **5a** was consumed completely, the solvent was removed under vacuum and the crude product was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate = 8:1) to obtain **5e** as a white solid. (18 mg, 71%). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.80 (dd, *J* = 7.9, 1.2 Hz, 1H), 7.61 (d, *J* = 2.3 Hz, 1H), 7.55 – 7.48 (m, 1H), 7.32 – 7.20 (m, 3H), 6.81 (d, *J* = 8.6 Hz, 1H), 6.38 (d, *J* = 1.8 Hz, 1H), 4.24 (d, *J* = 1.7 Hz, 1H), 2.33 – 2.27 (m, 1H), 2.24 (ddd, *J* = 13.5, 2.9, 2.3 Hz, 1H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 161.5, 157.8, 152.5, 149.7, 132.4, 131.4, 131.0, 127.5, 124.3, 123.0, 118.2, 116.9, 114.9, 114.4, 105.9, 93.1, 25.2, 24.5 ppm. **HRMS**: [M+H]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>12</sub>BrO<sub>4</sub><sup>+</sup> 370.9913; found: 370.9910. **[α]<sub>D</sub><sup>20</sup>** 79.49 (*c* = 1.50 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 19.89 min, *t*<sub>minor</sub> = 18.21 min, **ee** = 97%. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >20:1.

**2-((2*R*,6*S*)-5-(hydroxymethyl)-6*H*-2,6-methanobenzo[*d*][1,3]dioxocin-4-yl)phenol**  
**(9)**



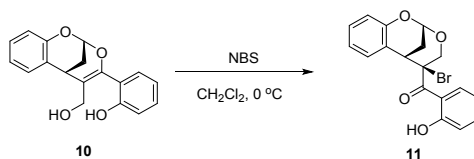
To a magnetically stirred suspension of  $\text{LiAlH}_4$  (20 mg, 0.52 mmol) in 0.5 mL of anhydrous tetrahydrofuran (THF), was added dropwise a solution of **5a** (30 mg, 0.10 mmol) in anhydrous tetrahydrofuran (0.5 mL) at  $0\text{ }^\circ\text{C}$ . After the reaction was completed, the mixture was added dropwise a small amount of 15 % KOH solution. Then the white suspension was filtered, the filtrate was evaporated and the crude product was purified by column chromatography on a silica gel (petroleum ether/ethyl acetate = 2/1) to get the desired product **9**. (20 mg, 66%).  **$^1\text{H}$  NMR** (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.26 – 7.22 (m, 2H), 7.21-7.14 (m, 2H), 6.98 (d,  $J$  = 8.0 Hz, 1H), 6.94-6.84 (m, 3H), 6.60 (s, 1H), 6.12 (d,  $J$  = 1.7 Hz, 1H), 4.04 (s, 2H), 3.73 (s, 1H), 2.29 (ddd,  $J$  = 13.0, 3.1, 1.9 Hz, 1H), 2.13 (dt,  $J$  = 13.1, 2.2 Hz, 1H) ppm.  **$^{13}\text{C}$  NMR** (125 MHz,  $\text{CDCl}_3$ )  $\delta$  154.4, 151.9, 145.6, 131.0, 130.6, 128.2, 127.5, 126.9, 121.3, 120.3, 120.1, 117.0, 116.4, 116.2, 92.5, 61.9, 28.3, 26.1 ppm. **HRMS**:  $[\text{M}-\text{H}]^-$  *calcd.* For  $\text{C}_{18}\text{H}_{15}\text{O}_4$  295.0976; found: 295.0978.  $[\alpha]_{\text{D}}^{20}$  -27.06 ( $c$  = 1.08 in  $\text{CHCl}_3$ ). The diastereomeric ratio was determined by  $^1\text{H}$  NMR, *dr* >20:1.

**(8*R*,14*S*)-1,1-dimethyl-1*H*,14*H*-8,14-methanobenzo[7,8][1,3]dioxocino[5,4-*c*]chromene (10)**



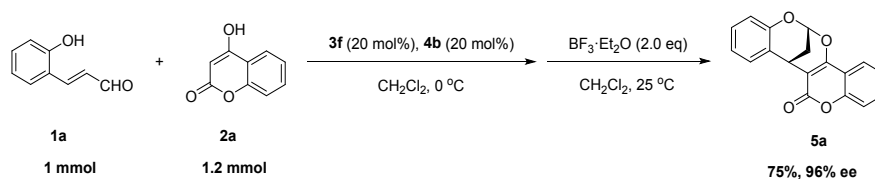
To a stirred solution of **5a** (18 mg, 0.062 mmol) in 0.5 mL anhydrous of tetrahydrofuran (THF), a solution of methyl magnesium chloride (3 M in tetrahydrofuran, 206  $\mu\text{L}$ , 0.62 mmol) was added at 0 °C under  $\text{N}_2$ . Then the reaction mixture was allowed to stir at room temperature. After stirring for about 30 min (TLC monitoring), the reaction mixture was poured into water. The organic phase was separated and the aqueous phase was extracted with ethyl acetate. The combined organic phases were washed with water and brine, dried with  $\text{Na}_2\text{SO}_4$ , and concentrated in vacuo. The residue was purified by chromatography on silica gel (petroleum ether/ethyl acetate = 3/1). The obtained product was dissolved in  $\text{CH}_2\text{Cl}_2$  (0.5 mL), then  $p\text{-TsOH}$  (11 mg, 0.062 mmol) was added to the solution. The reaction mixture was allowed to stir at 25 °C until the reaction was completed. The product was purified by chromatography on silica gel (petroleum ether/ethyl acetate = 50/1) to afford the desired product **10**. (11 mg, 60% for two steps).  **$^1\text{H}$  NMR** (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.42 (dd,  $J$  = 7.7, 1.5 Hz, 1H), 7.19 – 7.09 (m, 3H), 6.95 (d,  $J$  = 7.9 Hz, 1H), 6.92–6.85 (m, 2H), 6.76 (d,  $J$  = 8.0 Hz, 1H), 6.15 (d,  $J$  = 1.7 Hz, 1H), 3.51 (s, 1H) 2.18 (ddd,  $J$  = 12.9, 3.1, 2.0 Hz, 1H), 2.06 (dt,  $J$  = 12.9, 2.5 Hz, 1H), 1.65 (s, 3H), 1.26 (s, 3H) ppm.  **$^{13}\text{C}$  NMR** (125 MHz,  $\text{CDCl}_3$ )  $\delta$  152.7, 151.6, 139.2, 129.5, 127.9, 127.2, 126.4, 121.7, 120.7, 120.7, 118.4, 116.5, 116.1, 112.8, 91.4, 79.1, 27.1, 26.8, 26.7, 26.6 ppm. **HRMS**:  $[\text{M}+\text{H}]^+$  *calcd.* For  $\text{C}_{20}\text{H}_{19}\text{O}_3^+$  307.1329; found: 307.1325.  $[\alpha]_{\text{D}}^{20}$  -37.52 ( $c$  = 0.92 in  $\text{CHCl}_3$ ). The diastereomeric ratio was determined by  $^1\text{H}$  NMR, ***dr* >20:1**.

**(5-bromo-5,6-dihydro-4H-2,6-methanobenzo[d][1,3]dioxocin-5-yl)(2-hydroxyphenyl)methanone (11)**



To a stirred solution of **10** (38 mg, 0.13 mmol) in 1.0 mL CH<sub>2</sub>Cl<sub>2</sub>, was added bromosuccinimide (27.4 mg, 0.15 mmol) at 0 °C. After the full conversion of **10**, The product was purified by chromatography on silica gel (petroleum ether/ethyl acetate = 25/1) to afford the desired product **11** (42 mg, 86%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 11.65 (s, 1H), 8.47 (d, *J* = 8.3 Hz, 1H), 7.56 – 7.49 (m, 1H), 7.37 (dd, *J* = 7.6, 1.2 Hz, 1H), 7.33 – 7.28 (m, 1H), 7.14 – 7.07 (m, 1H), 7.04 - 6.99 (m, 1H), 6.99 – 6.93 (m, 2H), 5.60 (s, 1H), 4.51 (dd, *J* = 12.2, 1.0 Hz, 1H), 4.12 (s, 1H), 3.81 (d, *J* = 12.1 Hz, 1H), 2.11 (d, *J* = 13.8 Hz, 1H), 2.00 (ddd, *J* = 13.9, 4.1, 1.7 Hz, 1H) ppm. <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 198.2, 163.6, 154.9, 136.7, 132.0, 131.2, 129.8, 120.5, 120.2, 119.8, 118.4, 116.1, 115.7, 91.6, 64.3, 63.7, 37.2, 28.4 ppm. HRMS: [M+Na]<sup>+</sup> *calcd.* For C<sub>18</sub>H<sub>15</sub>BrNaO<sub>4</sub><sup>+</sup> 397.0046; found: 397.0048. [α]<sub>D</sub><sup>20</sup> -90.51 (*c* = 0.67 in CHCl<sub>3</sub>). The diastereomeric ratio was determined by <sup>1</sup>H NMR, *dr* >20:1.

## E. 1 mmol scale

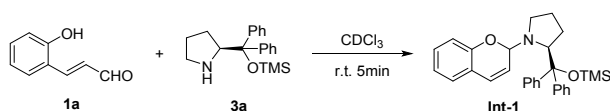


A glass vial equipped with a magnetic stirring bar was charged with **1a** (1.0 mmol, 1.0 equiv), catalyst **3f** (0.2 mmol, 0.2 equiv) and catalyst **4b** (0.2 mmol, 0.2 equiv) in  $\text{CH}_2\text{Cl}_2$  (5.0 mL) and the resulting solution was stirred for about 10 min at  $0\text{ }^\circ\text{C}$ . Then **2a** (1.2 mmol, 1.2 equiv) was added. After full conversion of first step,  $\text{BF}_3 \cdot \text{Et}_2\text{O}$  (2.0 eq) was added to the reaction mixture in situ and the reaction mixture was allowed to stir at  $25\text{ }^\circ\text{C}$ . After completion of the reaction, the reaction mixture was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate = 8/1) to give product **5a** 220 mg with 75% yield and 96% ee.

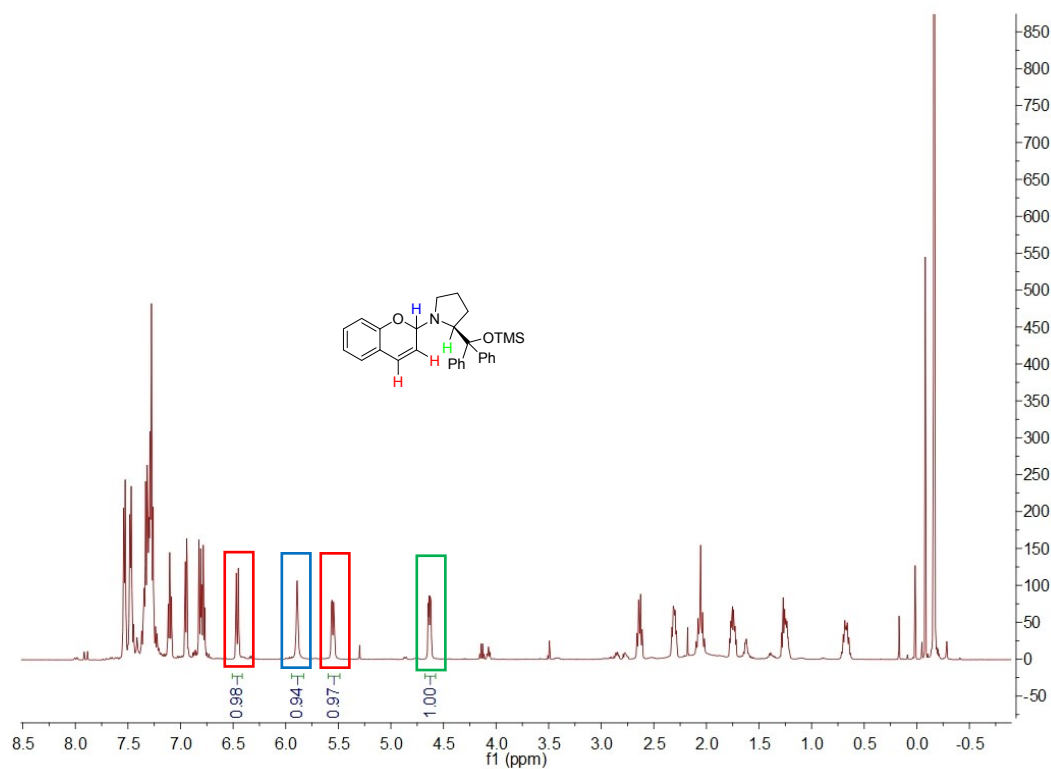
## F. NMR and Control Experiments

### NMR experiments

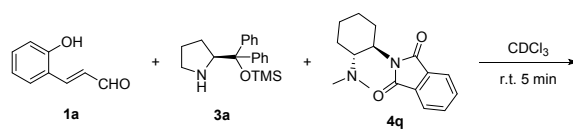
1)



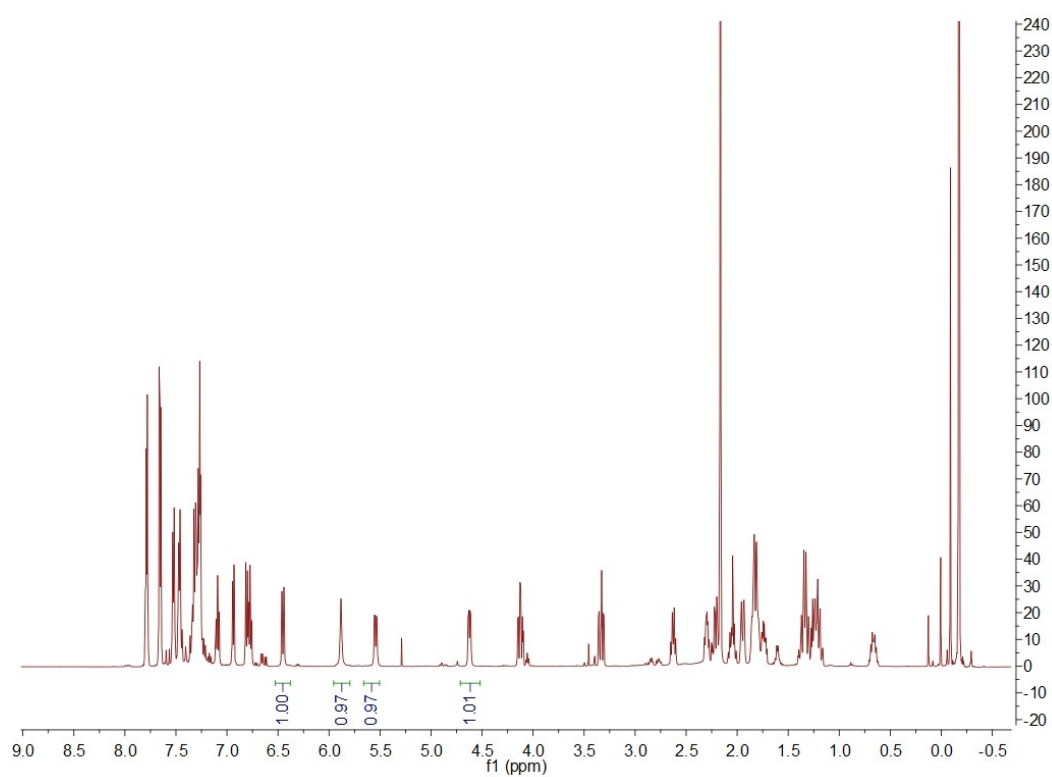
General procedure: To a solution of catalyst **3a** (0.05 mmol) in  $\text{CDCl}_3$  (0.4 ml) was added **1a** (0.05 mmol), and the mixture was allowed at room temperature for about 5 min. Then the system was monitored by  $^1\text{H}$  NMR (500 MHz). The result showed that a high concentration of aminoral **Int-1** was generated quickly.



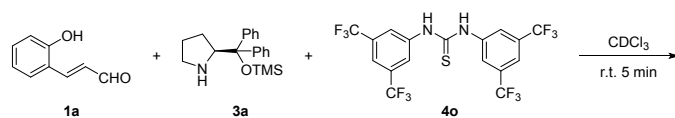
2)



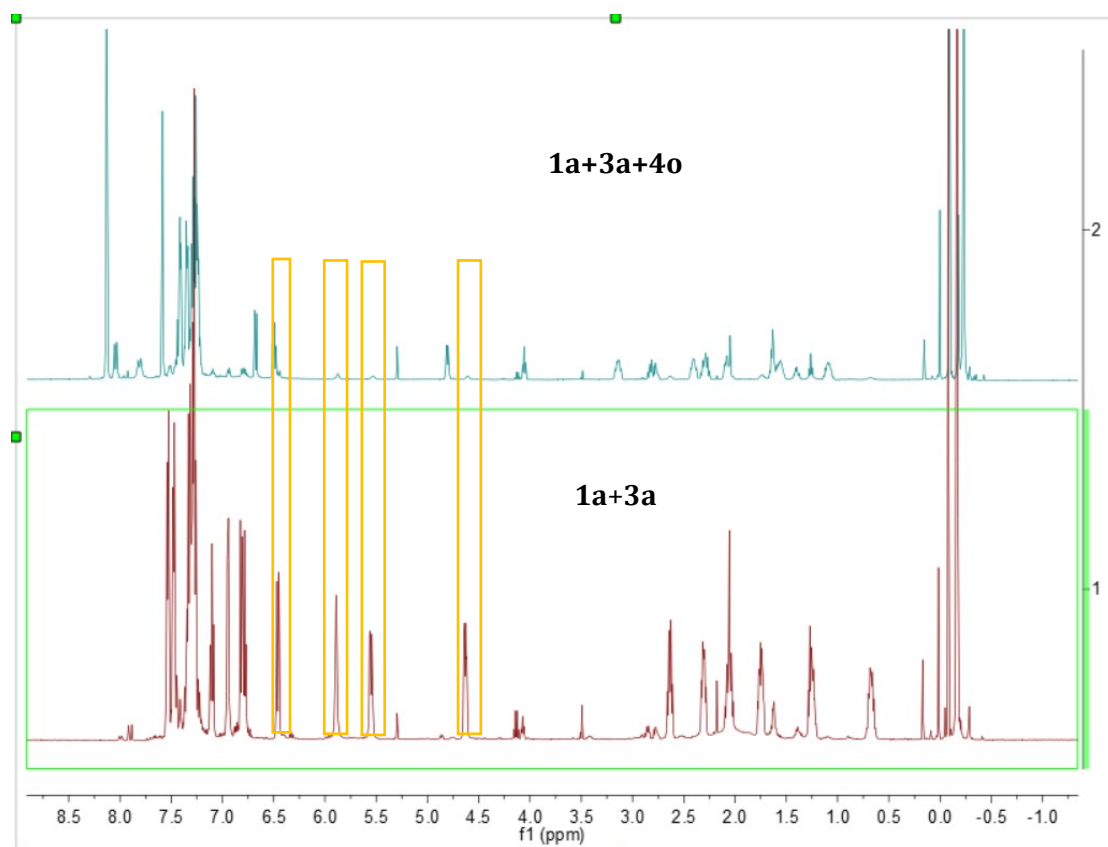
General procedure: To a solution of catalyst **3a** (0.05 mmol) and **4q** (0.05 mmol) in  $\text{CDCl}_3$  (0.4 ml) was added **1a** (0.05 mmol), and the mixture was allowed at room temperature for about 5 min. Then the system was monitored by  $^1\text{H}$  NMR (500 MHz). The result showed that tertiary amine has little effect on **Int-1**.



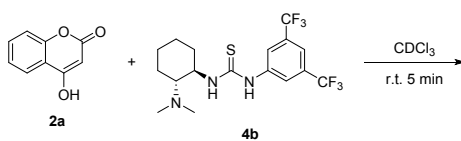
3)



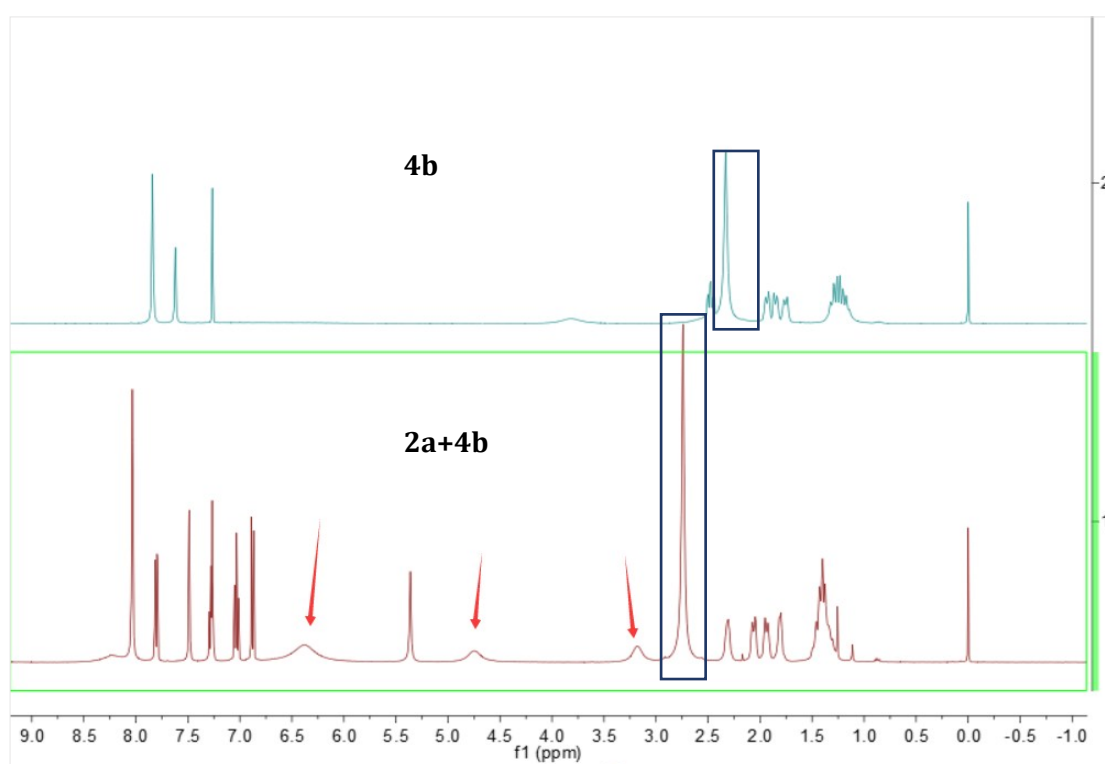
General procedure: To a solution of catalyst **3a** (0.05 mmol) and **4o** (0.05 mmol) in  $\text{CDCl}_3$  (0.4 ml) was added **1a** (0.05 mmol), and the mixture was allowed at room temperature for about 5 min. Then the system was monitored by  $^1\text{H}$  NMR (500 MHz). The result showed that thiourea has significant effect on **Int-1**. Addition of thiourea results in disappearance of **Int-1**.



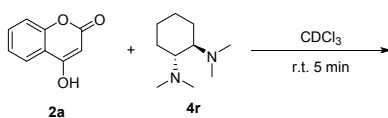
4)



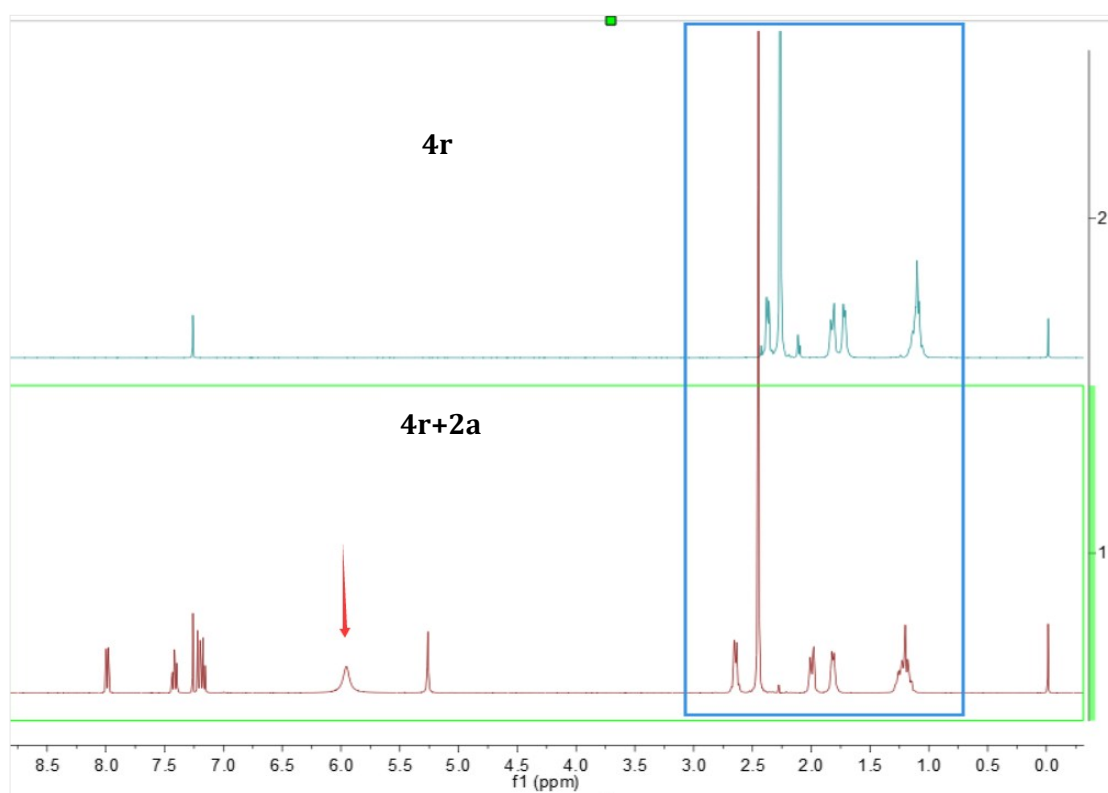
General procedure: To a solution of **2a** (0.05 mmol) in  $\text{CDCl}_3$  (0.5 ml) was added catalyst **4b** (0.05 mmol), the solution was changed immediately from turbidity to clarification. Then the system was monitored by  $^1\text{H}$  NMR (400 MHz). A dramatic proton shift of catalyst **4b** showed that **4b** has great interaction with **2a**.



5)

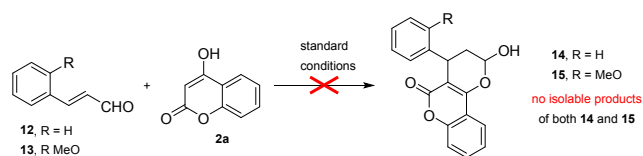


General procedure: To a solution of **2a** (0.05 mmol) in  $\text{CDCl}_3$  (0.5 ml) was added catalyst **4r** (0.05 mmol), the solution was also changed immediately from turbidity to clarification. Then the system was monitored by  $^1\text{H}$  NMR (400 MHz). A similar dramatic proton shift of catalyst **4r** showed that amine has significant effect on **2a**.



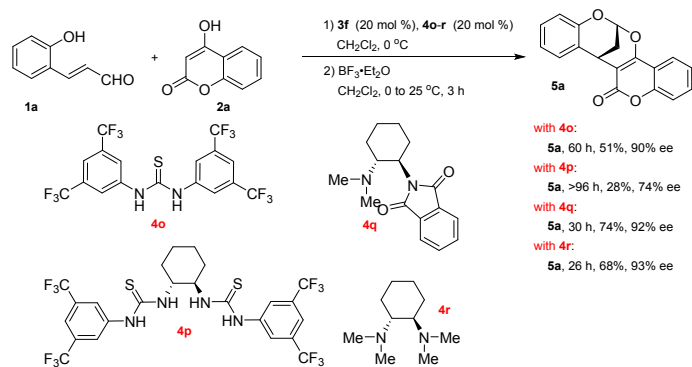
## Control experiments:

1)



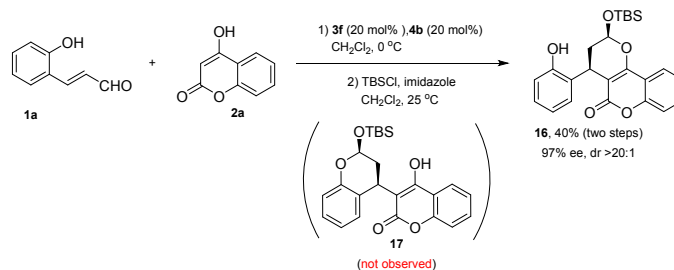
These results clearly indicated the important role of the phenolic hydroxyl group of substrates **1** in the developed multicatalytic system.

2)



These results emphasized the crucial role of bifunctional thiourea-tertiary amine catalysts for the superior efficiency of this multicatalytic system. It should be noted that base cocatalysts should be beneficial for stereochemical induction in this multicatalytic system.

3)



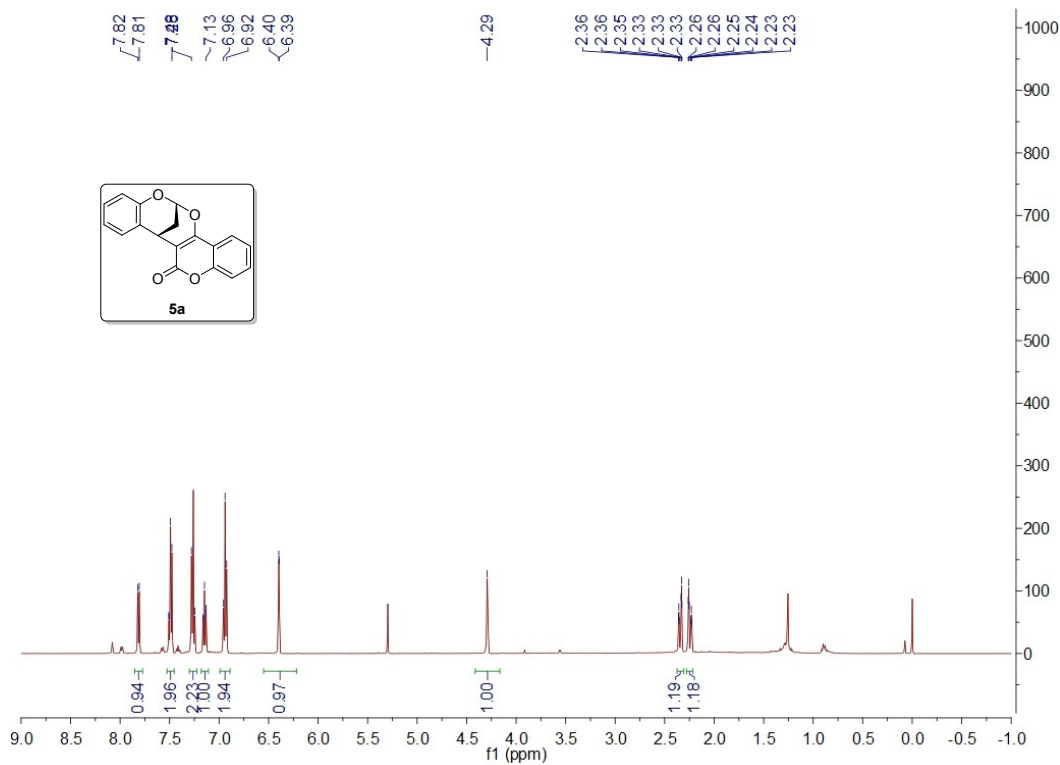
The results demonstrated the reaction intermediate of the Michael addition step.

Experiment procedure of **16** :

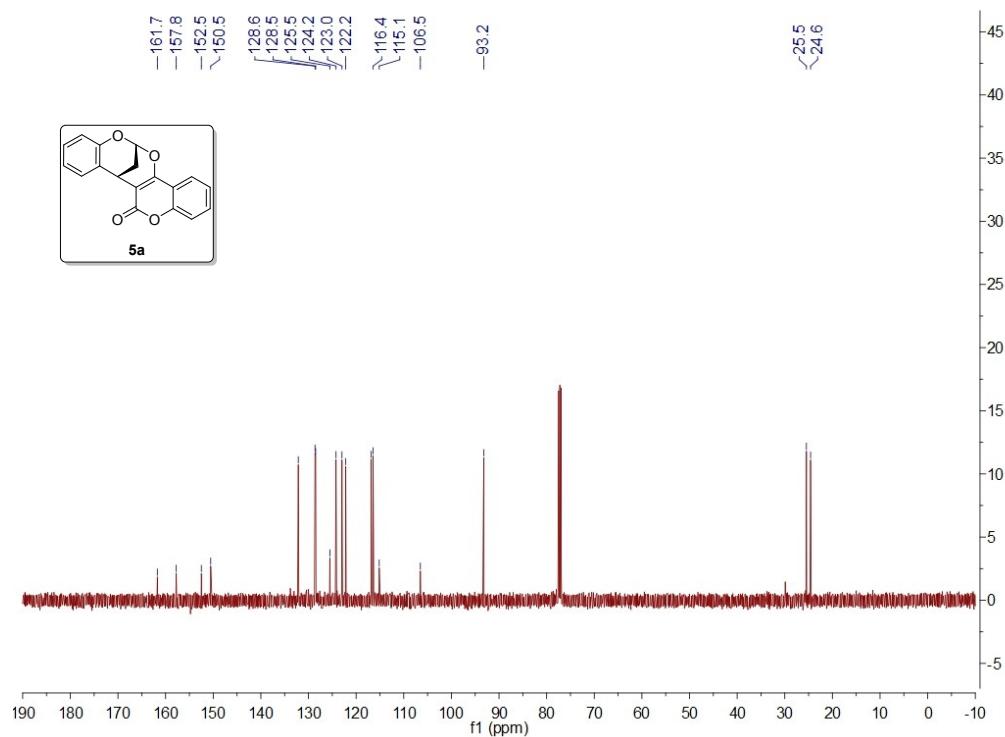
A glass vial equipped with a magnetic stirring bar was charged with **1** (0.10 mmol, 1.0 equiv), catalyst **3e** (0.02 mmol, 0.2 equiv) and catalyst **4b** (0.02 mmol, 0.2 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (0.5 mL) and the resulting solution was stirred for about 10 min at 0 °C. Then **2** (0.12 mmol, 1.2 equiv) was added. After full conversion of **1**, the crude product was purified by chromatography on silica gel (dichloromethane/ methanol = 20/1). The crude product was dissolved with CH<sub>2</sub>Cl<sub>2</sub>, then imidazole (0.3 mmol, 3.0 eq) and *tert*-butyldimethylchlorosilane (0.03 mmol, 3.0 eq) was added to the solution. The reaction mixture was allowed to stir at 25 °C until the reaction was completed. Then water was dropped to the mixture. The organic phase was separated and the aqueous phase was extracted with ethyl acetate. The combined organic phases were washed with water and brine, dried with Na<sub>2</sub>SO<sub>4</sub>, and concentrated in vacuo. The residue was purified by chromatography on silica gel (petroleum ether/ethyl acetate = 20/1) to afford compound **12** (17 mg, 40% for two steps). **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 8.73 (s, 1H), 7.74 (dd, *J* = 7.9, 1.2 Hz, 1H), 7.56 – 7.47 (m, 1H), 7.33 (d, *J* = 8.3 Hz, 1H), 7.22 (dd, *J* = 16.6, 8.4 Hz, 2H), 7.08 (d, *J* = 7.8 Hz, 1H), 6.93 (dd, *J* = 7.5, 4.8 Hz, 2H), 5.71 (t, *J* = 2.4 Hz, 1H), 4.80 (d, *J* = 10.2 Hz, 1H), 2.72 (ddd, *J* = 13.6, 10.5, 2.7 Hz, 1H), 2.16 (d, *J* = 14.7 Hz, 1H), 0.85 (s, 12H), 0.21 (s, 3H), 0.12 (s, 3H) ppm. **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 164.5, 160.4, 152.5, 151.0, 131.8, 129.4, 128.6, 123.8, 123.5, 122.5, 121.7, 118.5, 116.8, 116.2, 108.9, 90.7, 34.9, 26.2, 25.5, 18.3, -4.9, -5.5 ppm. **HRMS**: [M+Na]<sup>+</sup> *calcd.* For C<sub>24</sub>H<sub>28</sub>NaO<sub>5</sub>Si<sup>+</sup> 447.1598; found: 447.1596. **[α]<sub>D</sub><sup>20</sup>** -39.91 (*c* = 1.33 in CHCl<sub>3</sub>). The enantiomeric excess was determined by HPLC using Chiralpak IC column [*n*-hexane/*i*-PrOH = 80/20, 1 mL/min], λ = 210 nm, *t*<sub>major</sub> = 10.44 min, *t*<sub>minor</sub> = 9.55 min, **ee** = **97%**. The diastereomeric ratio was determined by <sup>1</sup>H NMR, **dr** >**20:1**.

## G. NMR spectra and HPLC traces

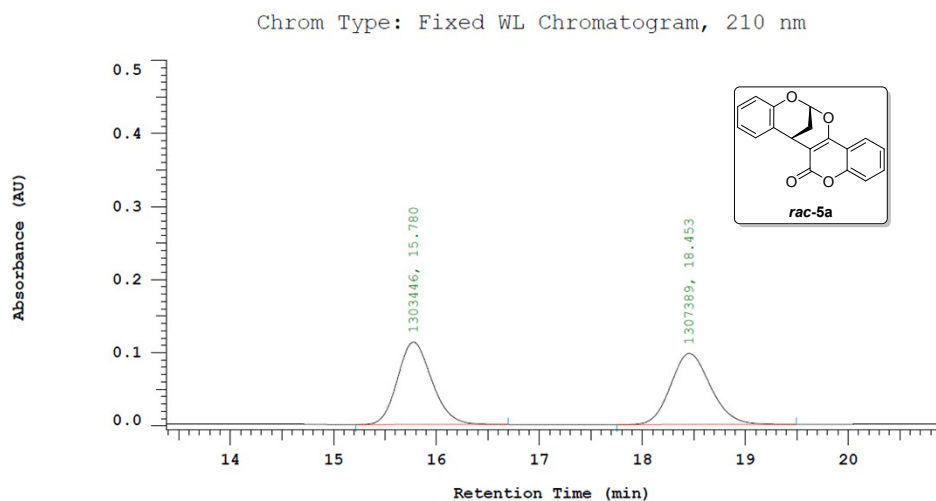
The  $^1\text{H}$  NMR spectrum of 5a (500 MHz,  $\text{CDCl}_3$ )



The  $^{13}\text{C}$  NMR spectrum of 5a (125 MHz,  $\text{CDCl}_3$ )



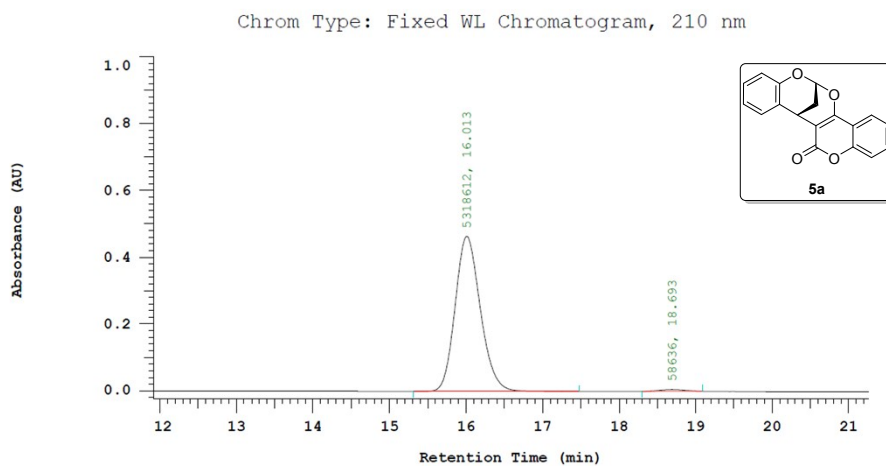
The HPLC of racemic 5a



Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 15.780 | 1303446 | 49.924  | BB |
| 2   | 18.453 | 1307389 | 50.076  | BB |
|     |        | 2610835 | 100.000 |    |

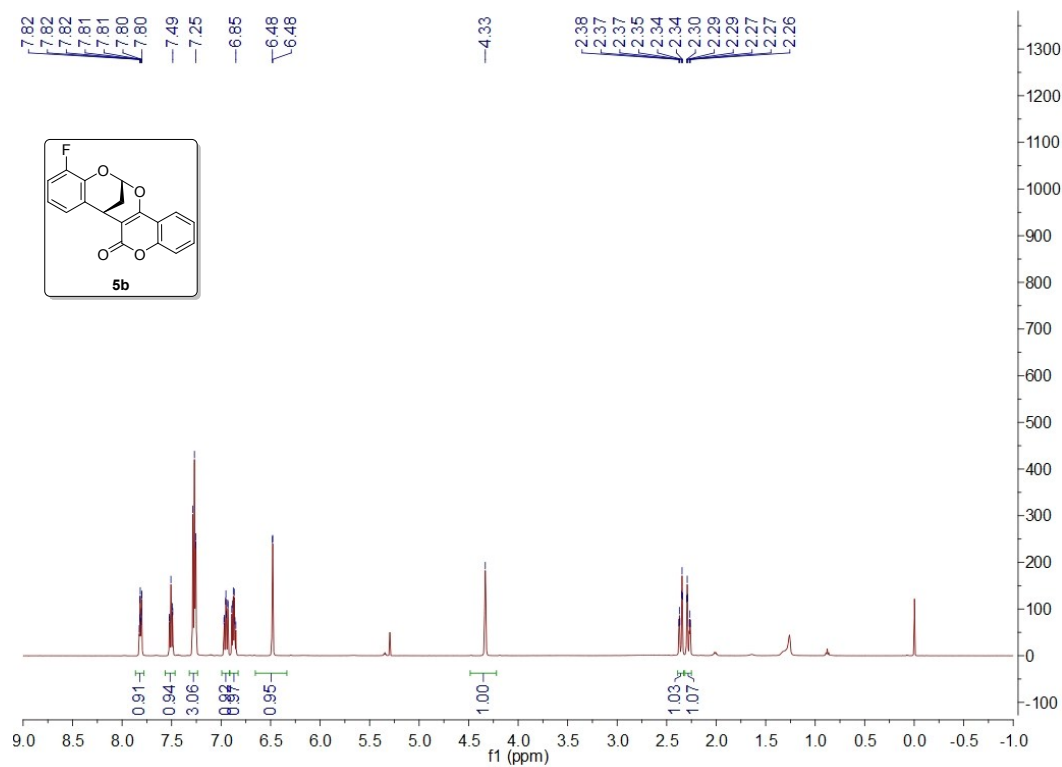
## The HPLC of chiral 5a



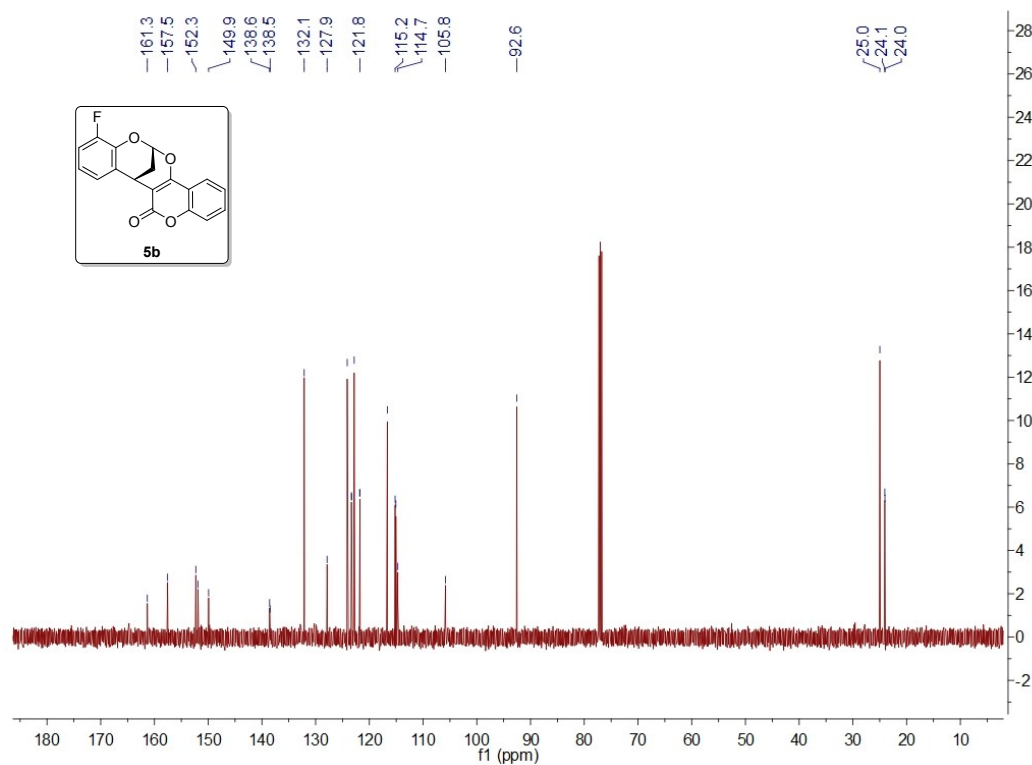
Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 16.013 | 5318612 | 98.910  | BB |
| 2   | 18.693 | 58636   | 1.090   | BB |
|     |        | 5377248 | 100.000 |    |

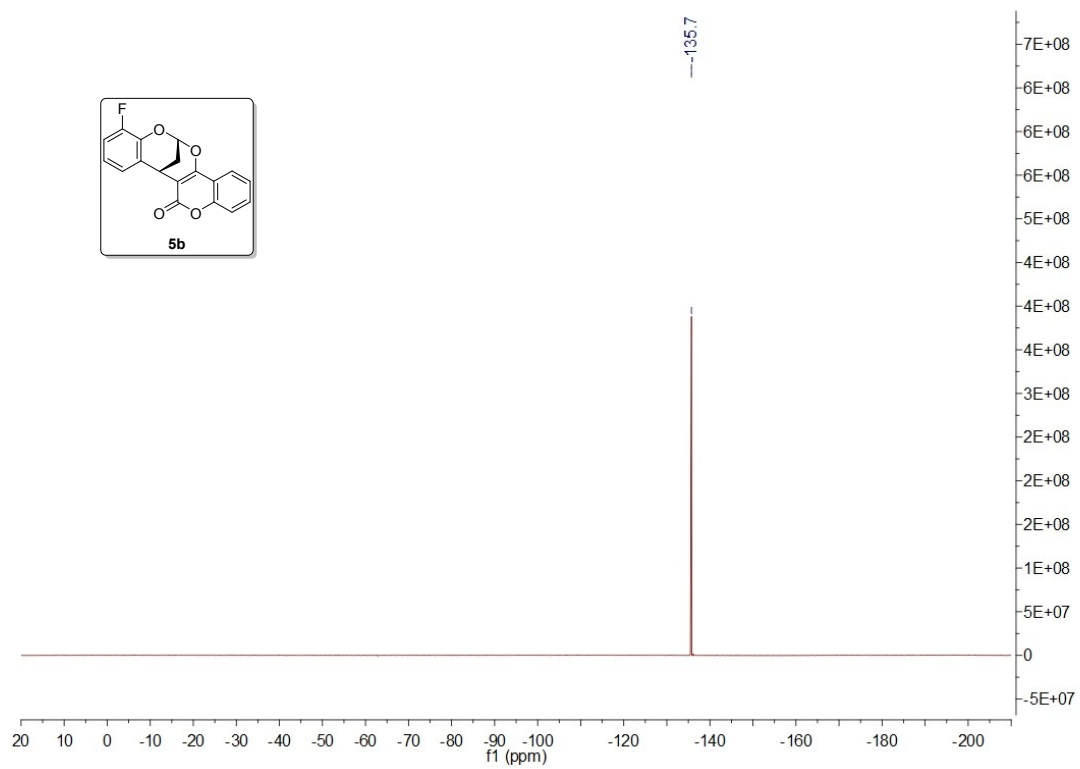
## The <sup>1</sup>H NMR spectrum of 5b (500 MHz, CDCl<sub>3</sub>)



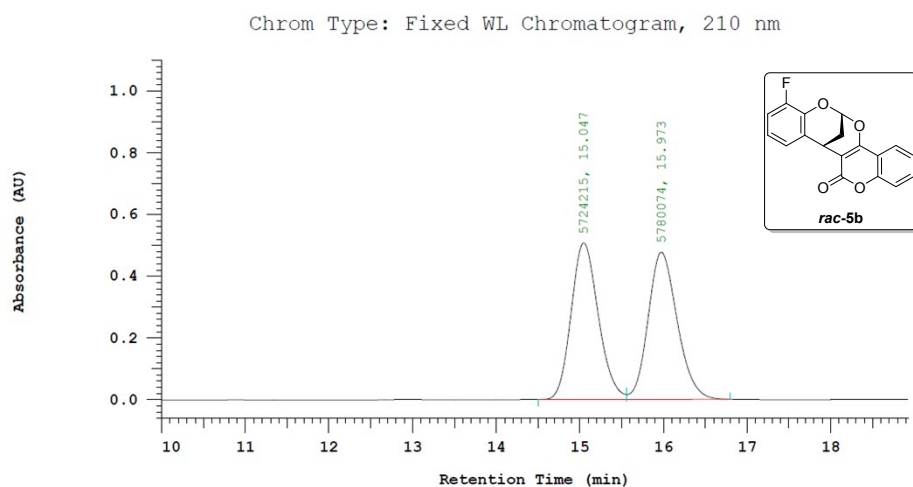
The <sup>13</sup>C NMR spectrum of 5b (125 MHz, CDCl<sub>3</sub>)



The <sup>19</sup>F NMR spectrum of 5b (400 MHz, CDCl<sub>3</sub>)



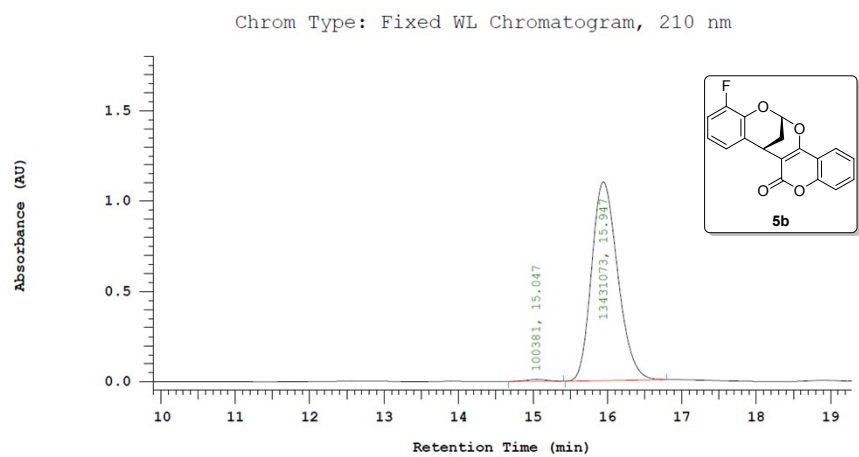
## The HPLC of racemic 5b



Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 15.047 | 5724215  | 49.757  | BV |
| 2   | 15.973 | 5780074  | 50.243  | VB |
|     |        | 11504289 | 100.000 |    |

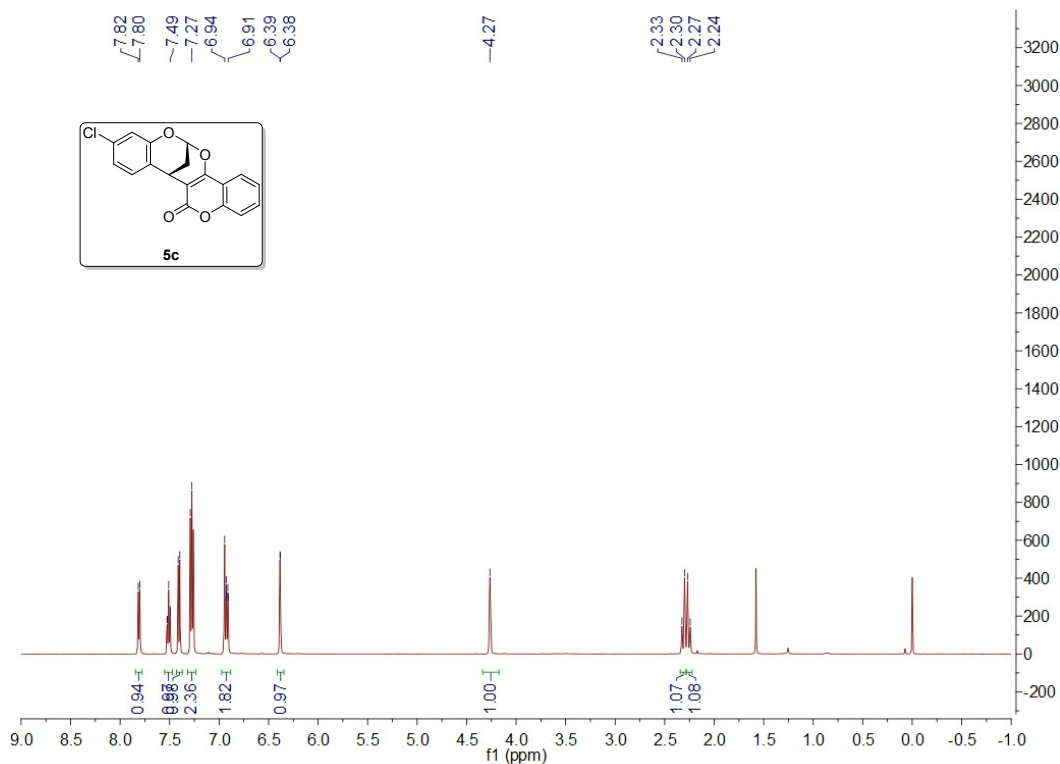
## The HPLC of chiral 5b



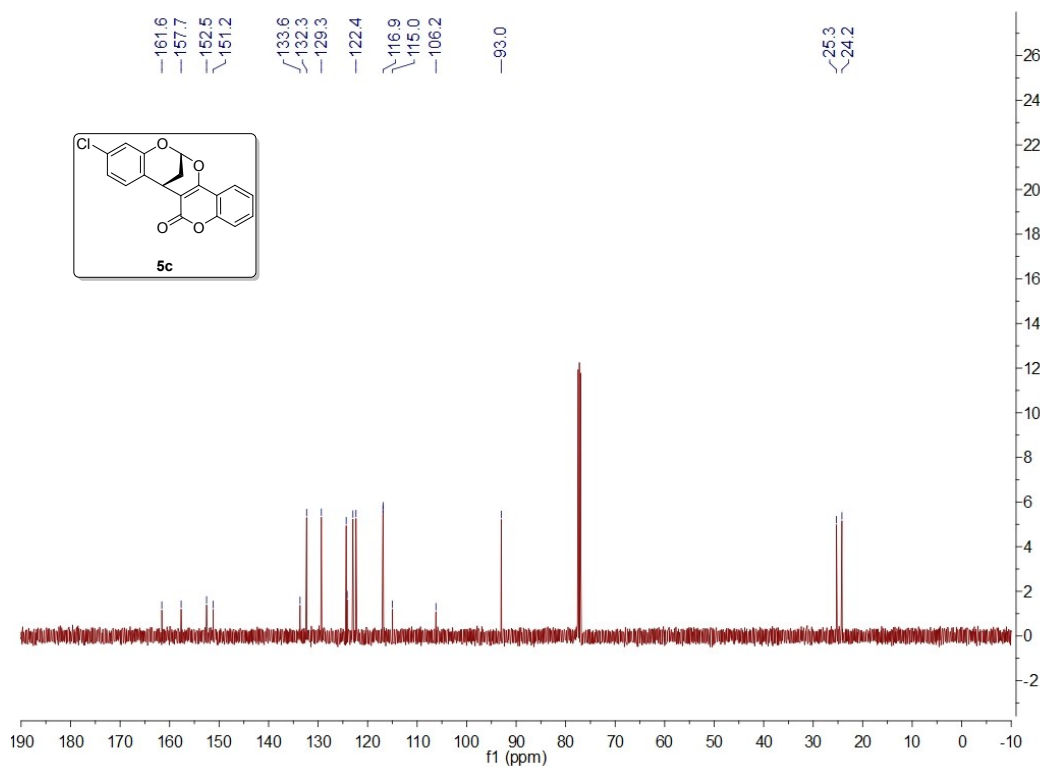
Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 15.047 | 100381   | 0.742   | BB |
| 2   | 15.947 | 13431073 | 99.258  | BB |
|     |        | 13531454 | 100.000 |    |

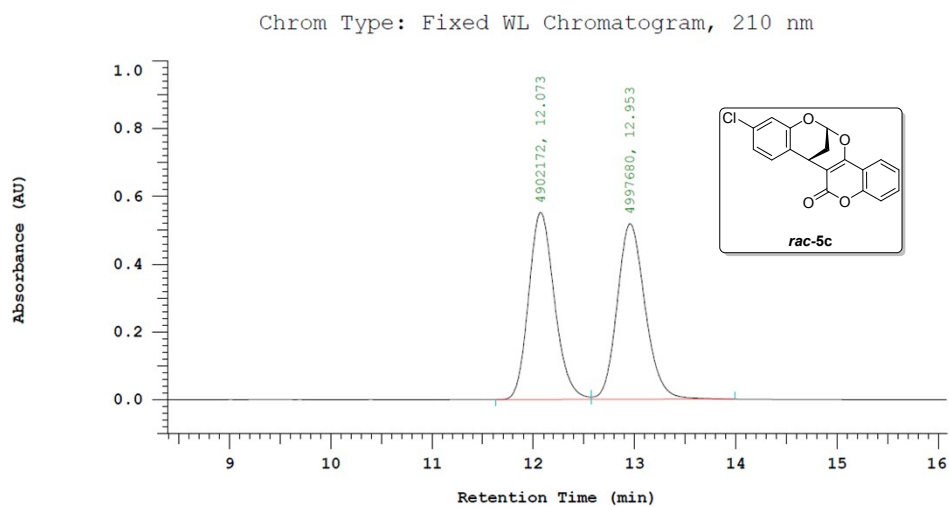
The  $^1\text{H}$  NMR spectrum of 5c (500 MHz,  $\text{CDCl}_3$ )



The  $^{13}\text{C}$  NMR spectrum of 5c (125 MHz,  $\text{CDCl}_3$ )



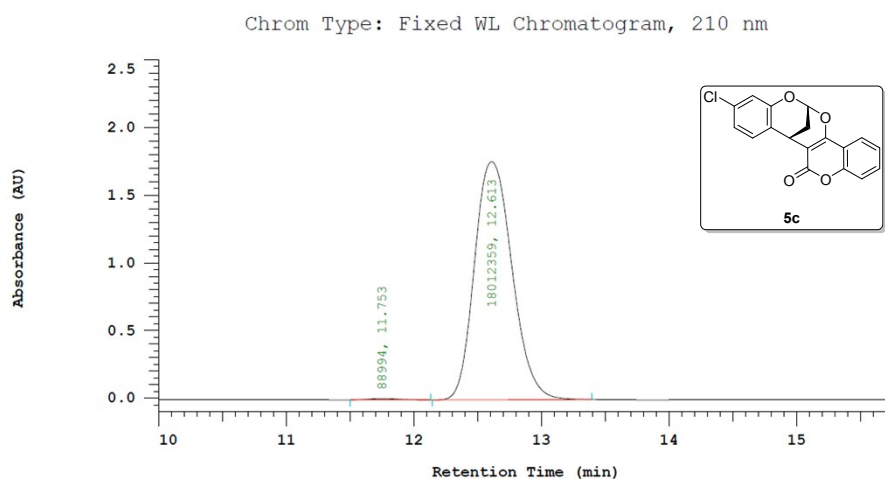
## The HPLC of racemic 5c



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 12.073 | 4902172 | 49.518  | BV |
| 2   | 12.953 | 4997680 | 50.482  | VB |
|     |        | 9899852 | 100.000 |    |

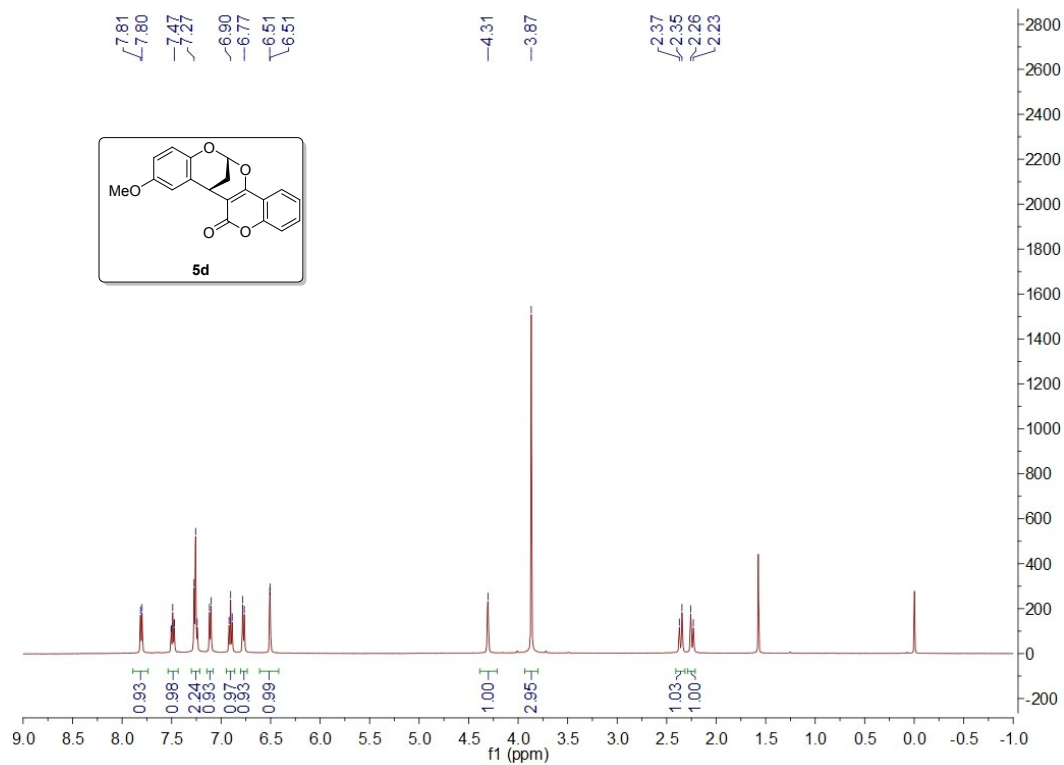
## The HPLC of chiral 5c



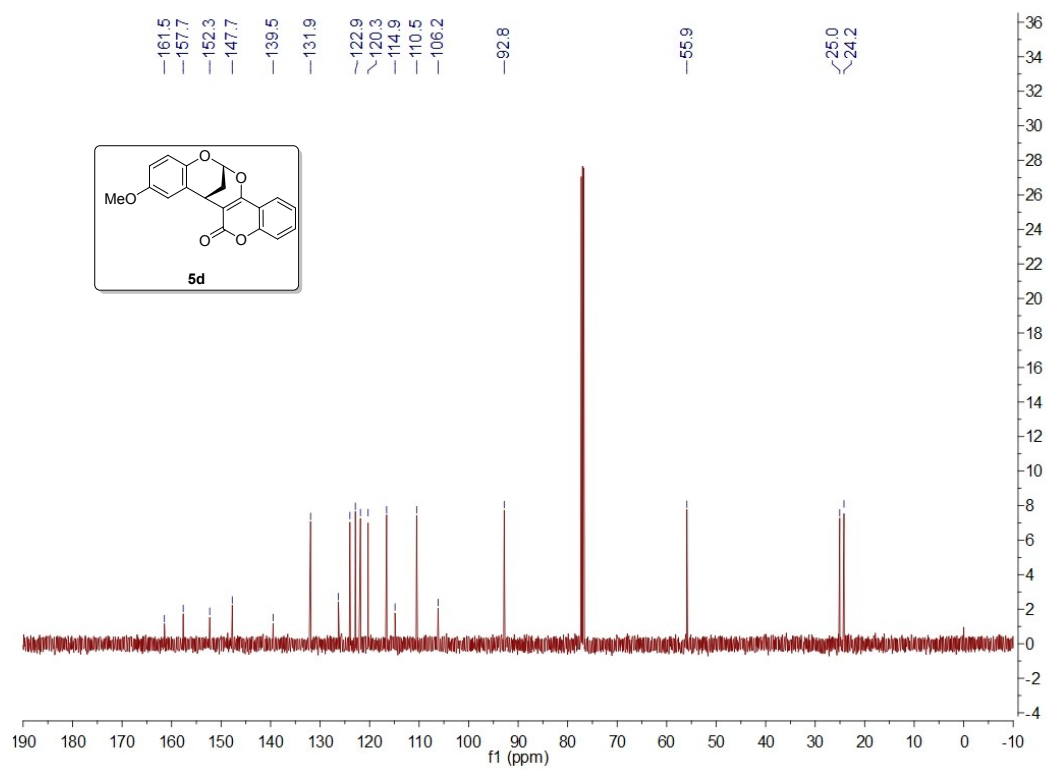
Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 11.753 | 88994    | 0.492   | BB |
| 2   | 12.613 | 18012359 | 99.508  | BB |
|     |        | 18101353 | 100.000 |    |

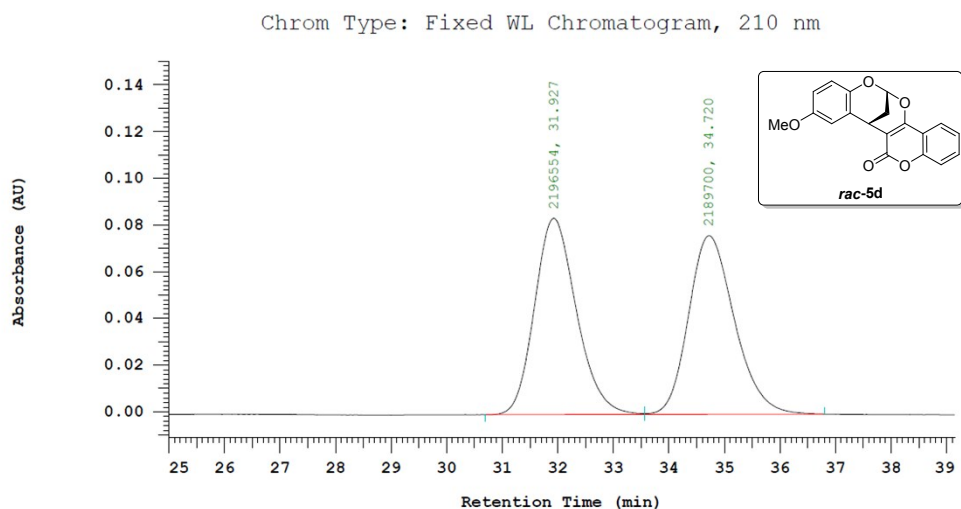
## The <sup>1</sup>H NMR spectrum of 5d (500 MHz, CDCl<sub>3</sub>)



### The <sup>13</sup>C NMR spectrum of 5d (125 MHz, CDCl<sub>3</sub>)



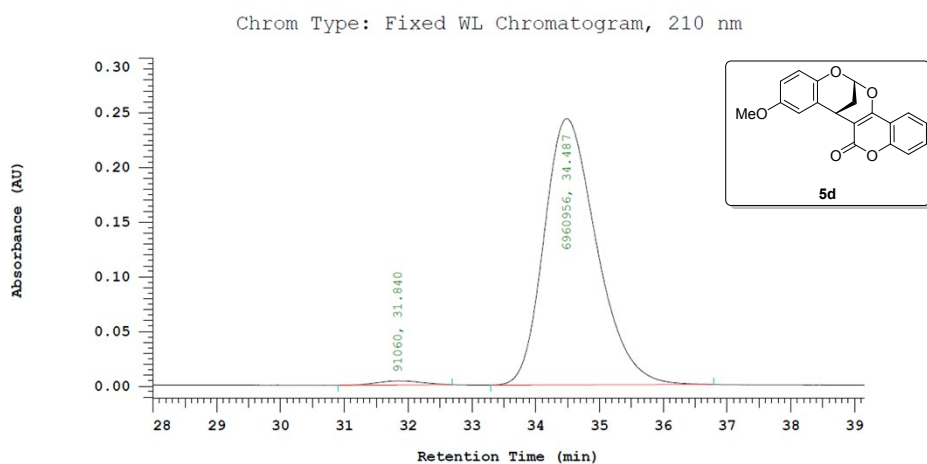
### The HPLC of racemic 5d



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 31.927 | 2196554 | 50.078  | BV |
| 2   | 34.720 | 2189700 | 49.922  | VB |
|     |        | 4386254 | 100.000 |    |

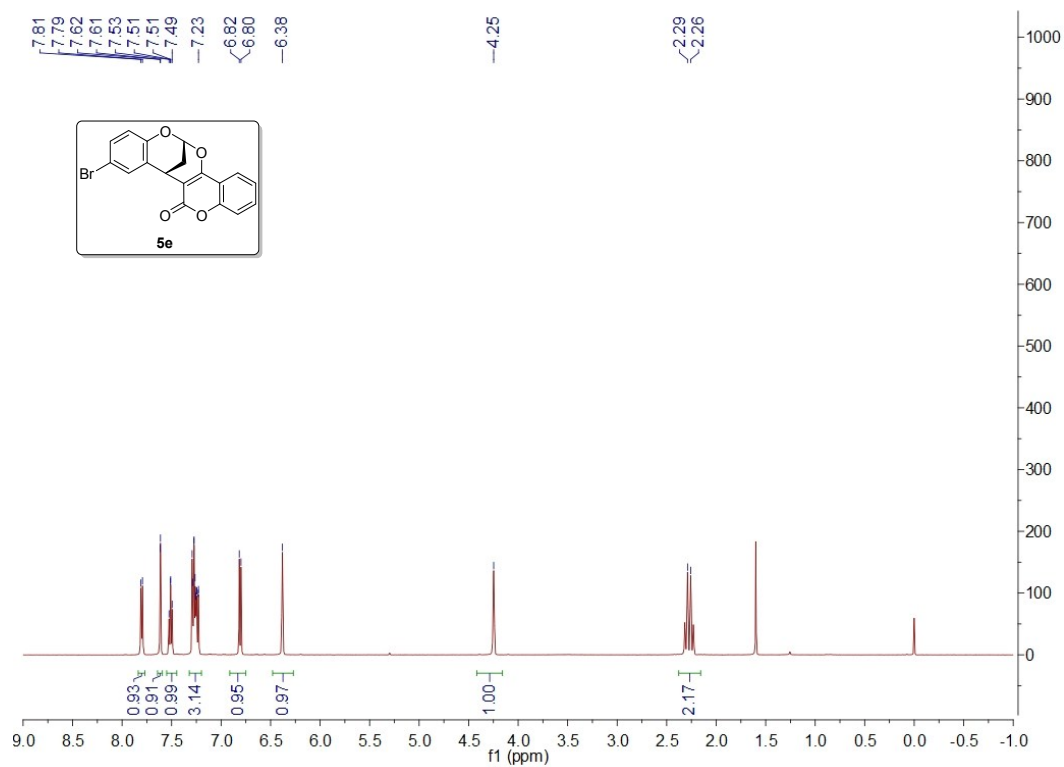
## The HPLC of chiral 5d



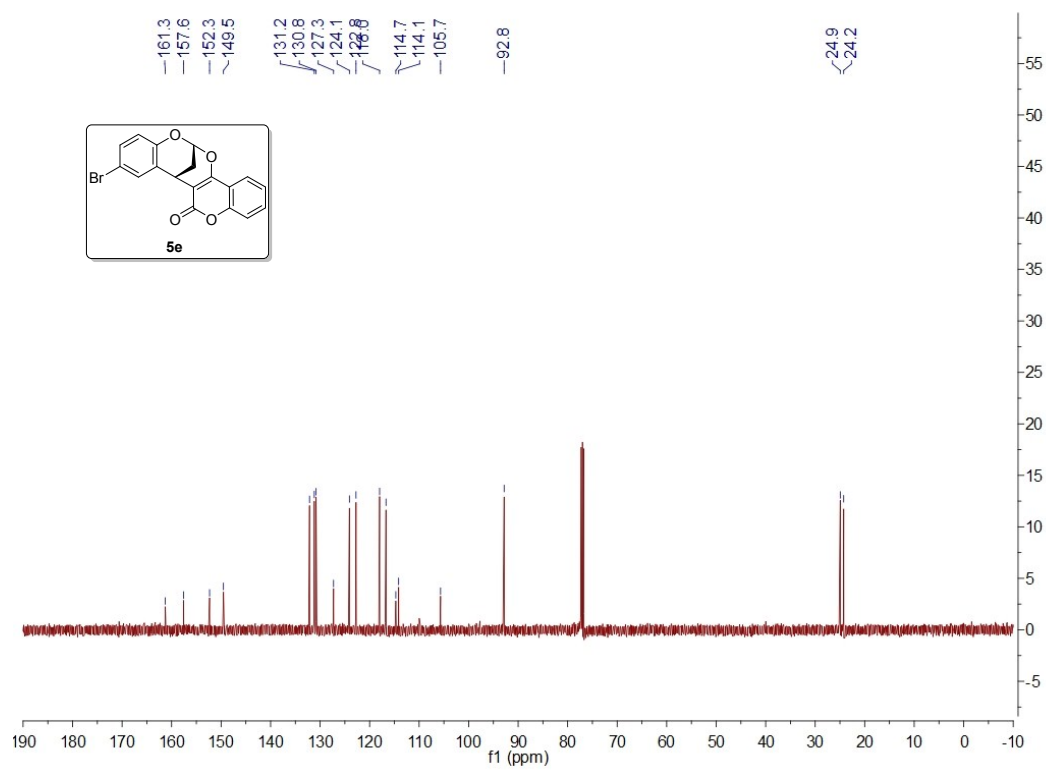
Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 31.840 | 91060   | 1.291   | BB |
| 2   | 34.487 | 6960956 | 98.709  | BB |
|     |        | 7052016 | 100.000 |    |

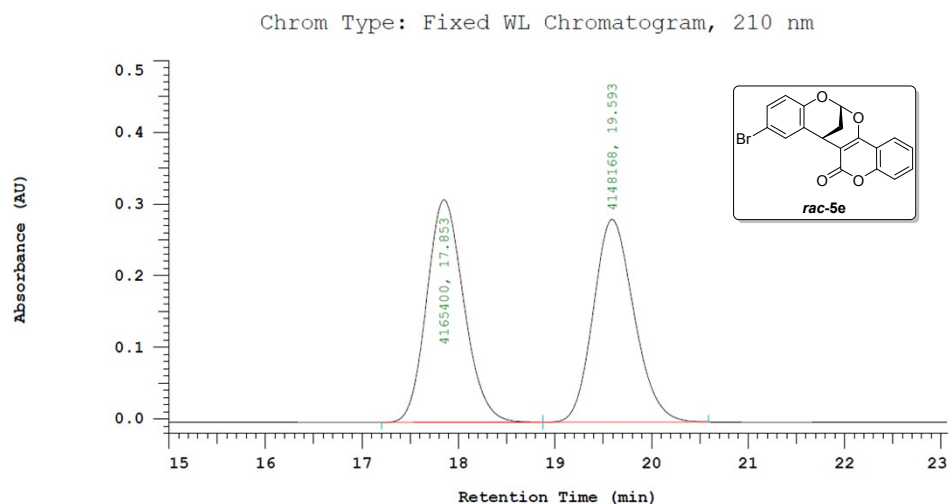
## The <sup>1</sup>H NMR spectrum of 5e (500 MHz, CDCl<sub>3</sub>)



### The <sup>13</sup>C NMR spectrum of 5e (125 MHz, CDCl<sub>3</sub>)



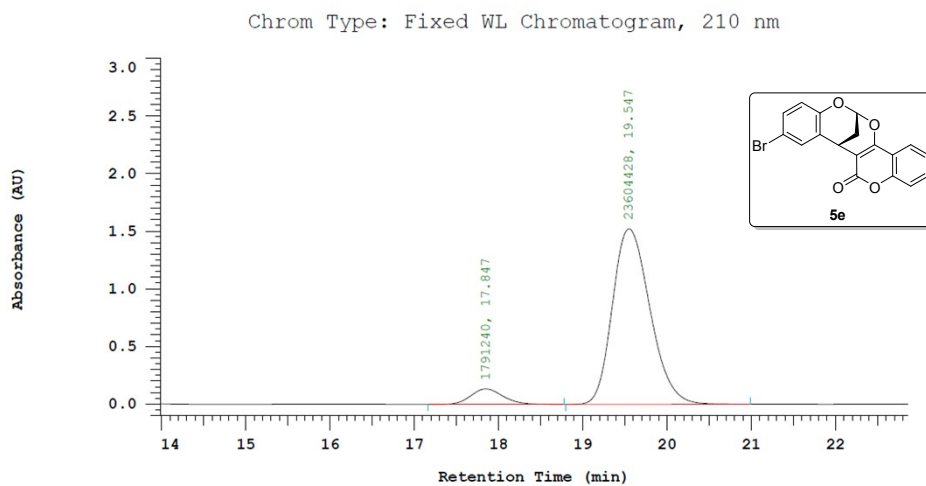
### The HPLC of racemic 5e



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 17.853 | 4165400 | 50.104  | BB |
| 2   | 19.593 | 4148168 | 49.896  | BB |
|     |        | 8313568 | 100.000 |    |

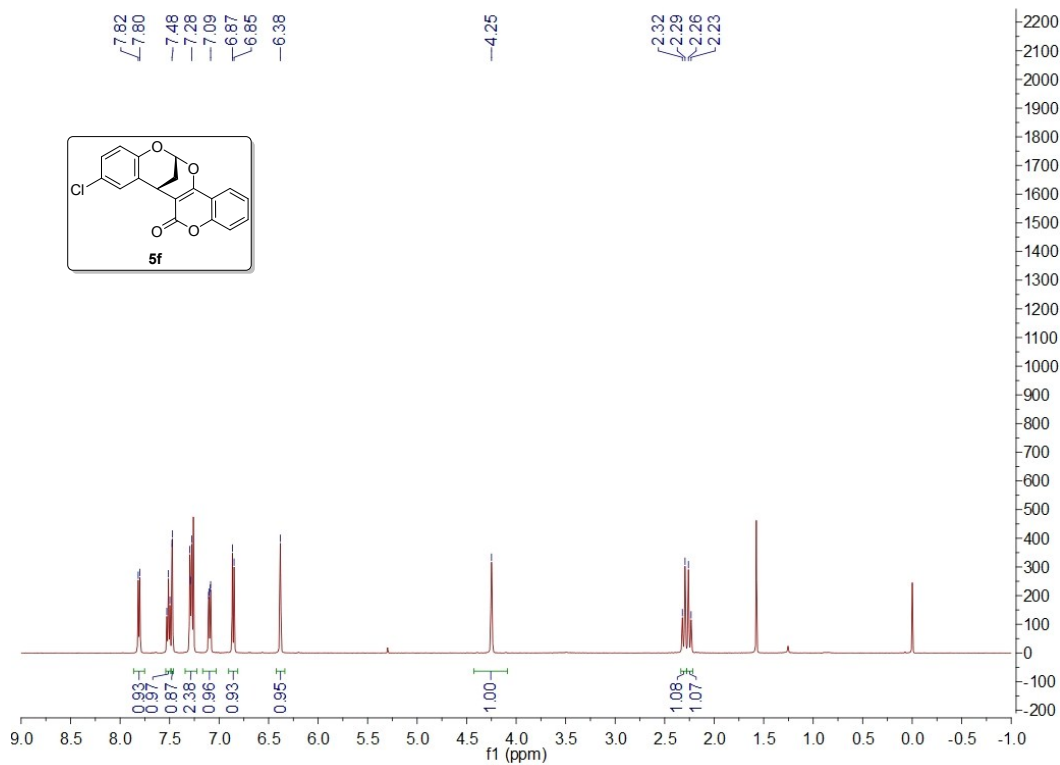
## The HPLC of chiral 5e



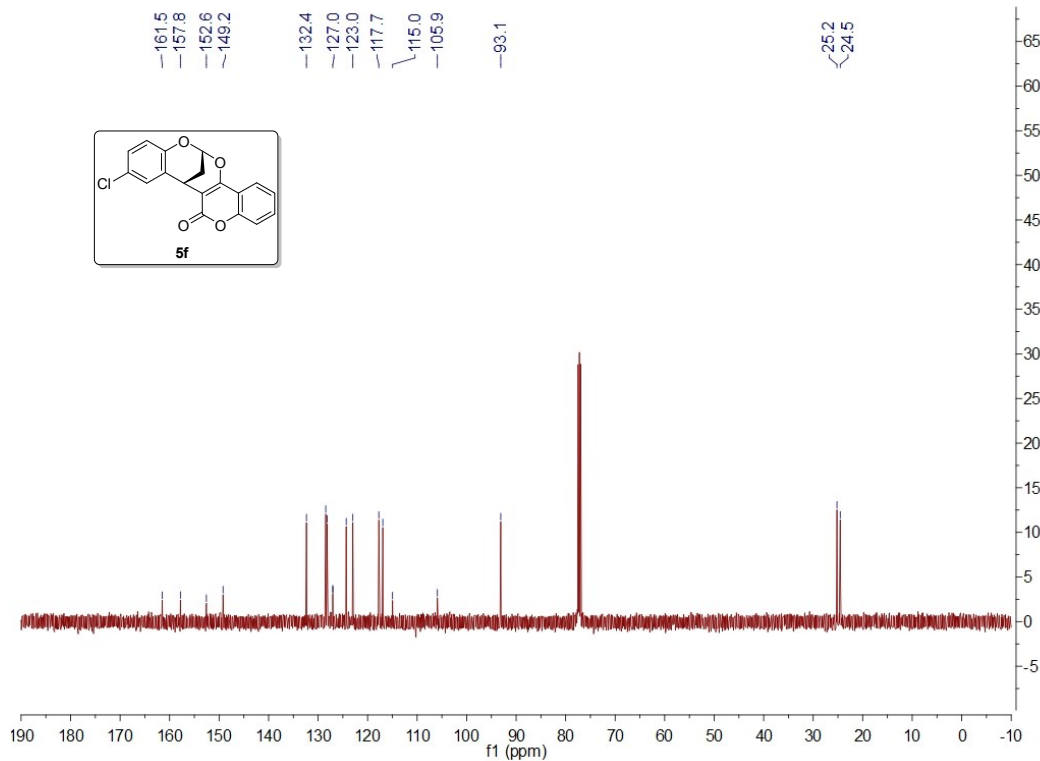
Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 17.847 | 1791240  | 7.053   | BB |
| 2   | 19.547 | 23604428 | 92.947  | BB |
|     |        | 25395668 | 100.000 |    |

## The <sup>1</sup>H NMR spectrum of 5f (500 MHz, CDCl<sub>3</sub>)

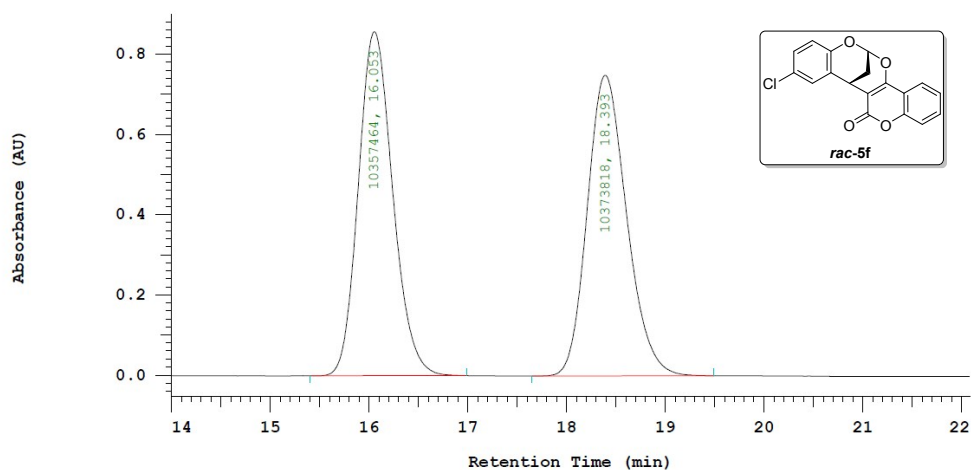


### The <sup>13</sup>C NMR spectrum of 5f (125 MHz, CDCl<sub>3</sub>)



### The HPLC of racemic 5f

Chrom Type: Fixed WL Chromatogram, 210 nm



Chrom Type: Fixed WL Chromatogram, 210 nm

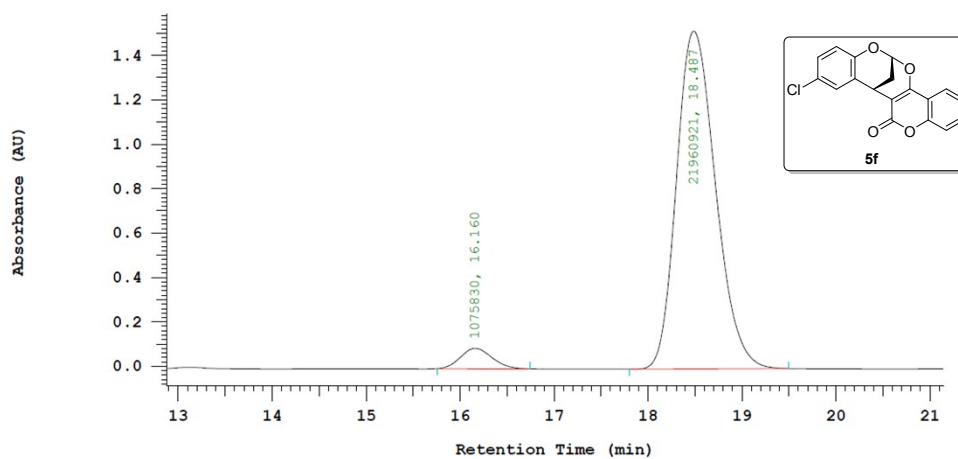
Peak Quantitation: AREA

Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 16.053 | 10357464 | 49.961  | BB |
| 2   | 18.393 | 10373818 | 50.039  | BB |
|     |        | 20731282 | 100.000 |    |

## The HPLC of chiral 5f

Chrom Type: Fixed WL Chromatogram, 210 nm



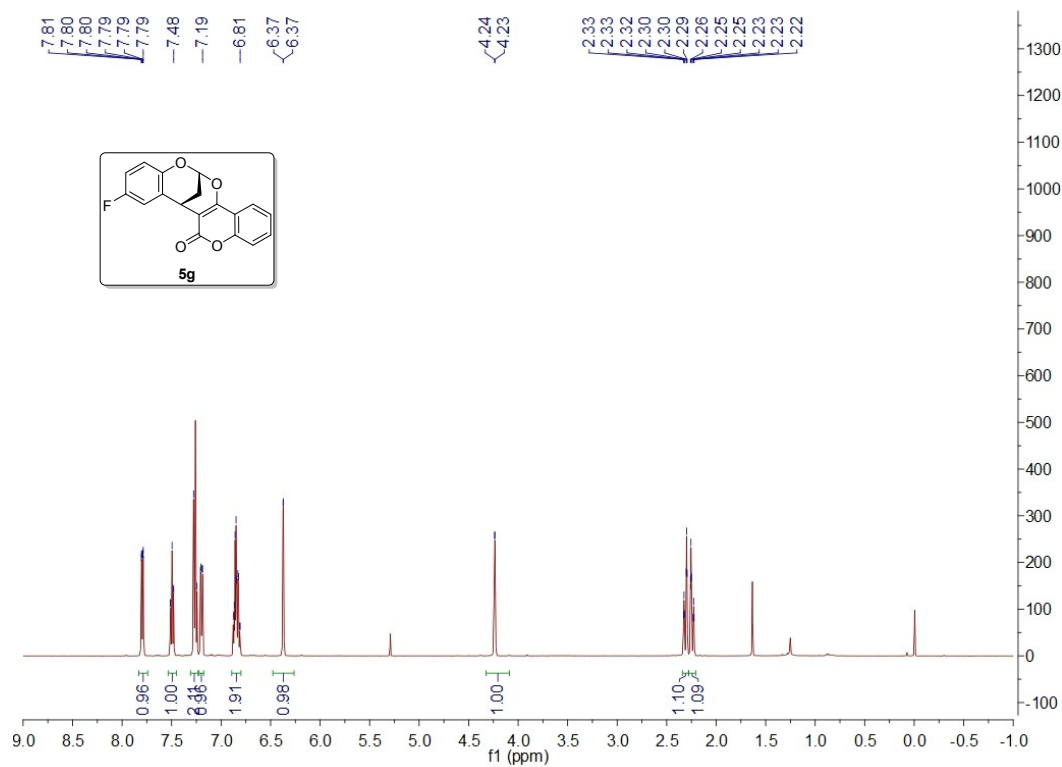
Chrom Type: Fixed WL Chromatogram, 210 nm

Peak Quantitation: AREA

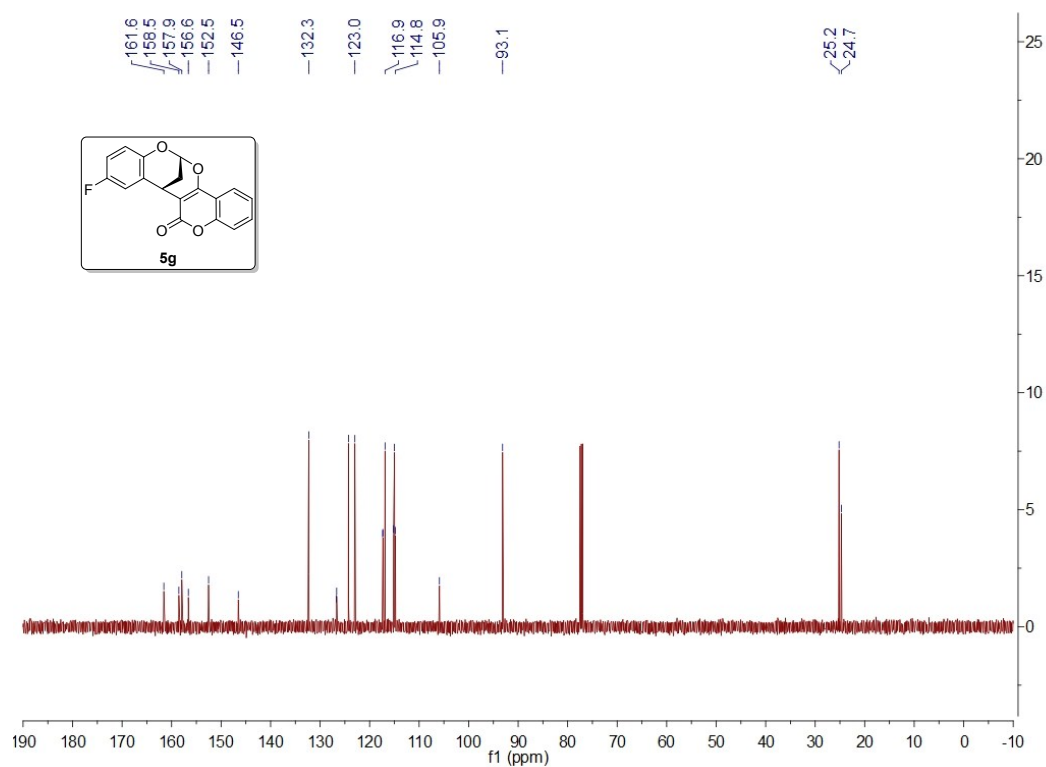
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 16.160 | 1075830  | 4.670   | BB |
| 2   | 18.487 | 21960921 | 95.330  | BB |
|     |        | 23036751 | 100.000 |    |

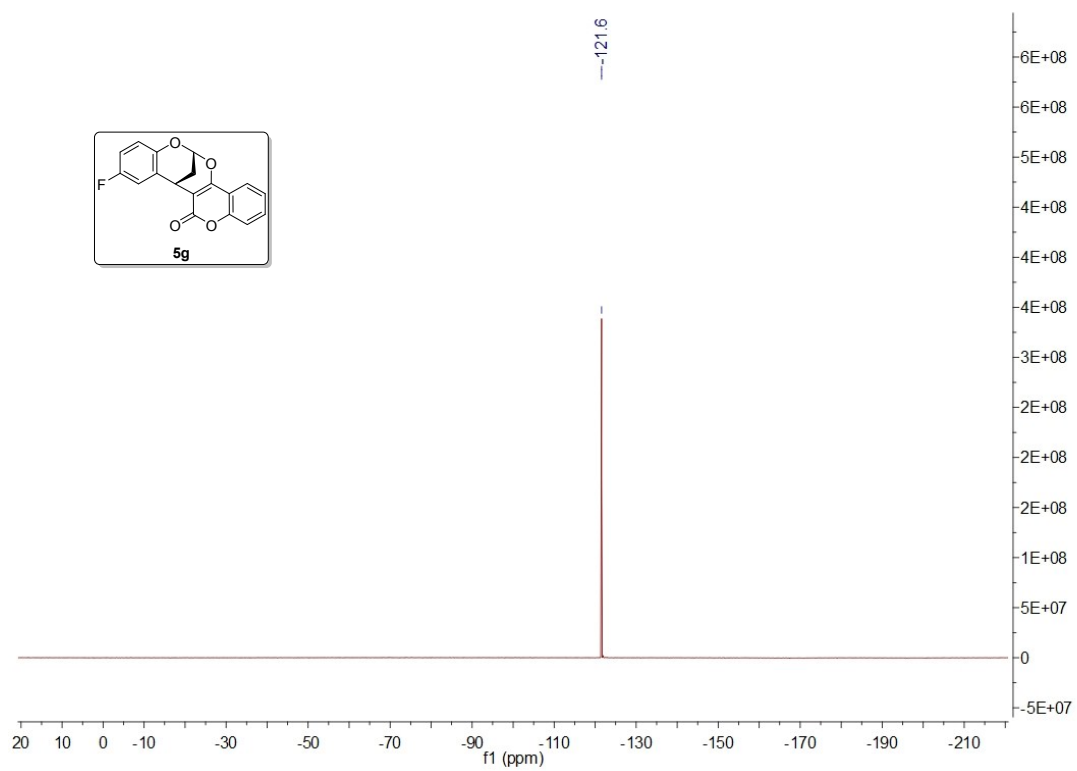
## The <sup>1</sup>H NMR spectrum of 5g (500 MHz, CDCl<sub>3</sub>)



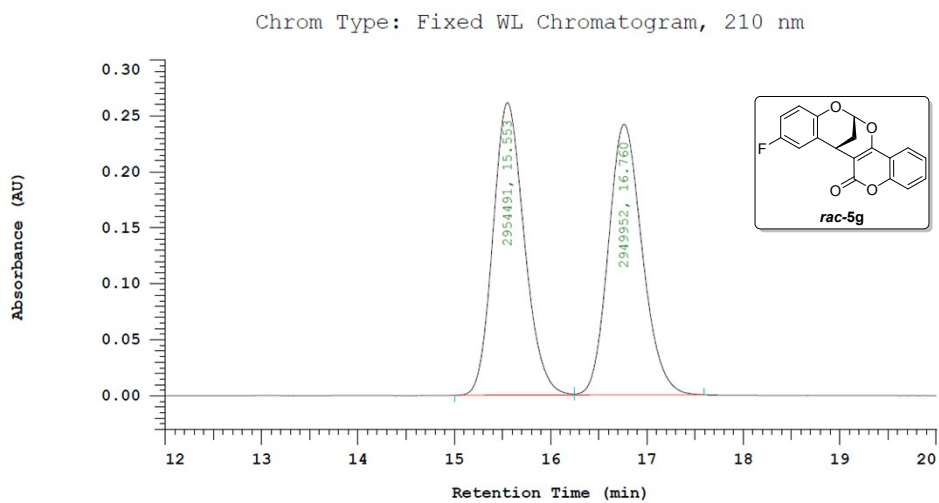
The <sup>13</sup>C NMR spectrum of 5g (125 MHz, CDCl<sub>3</sub>)



The <sup>19</sup>F NMR spectrum of 5g (400 MHz, CDCl<sub>3</sub>)



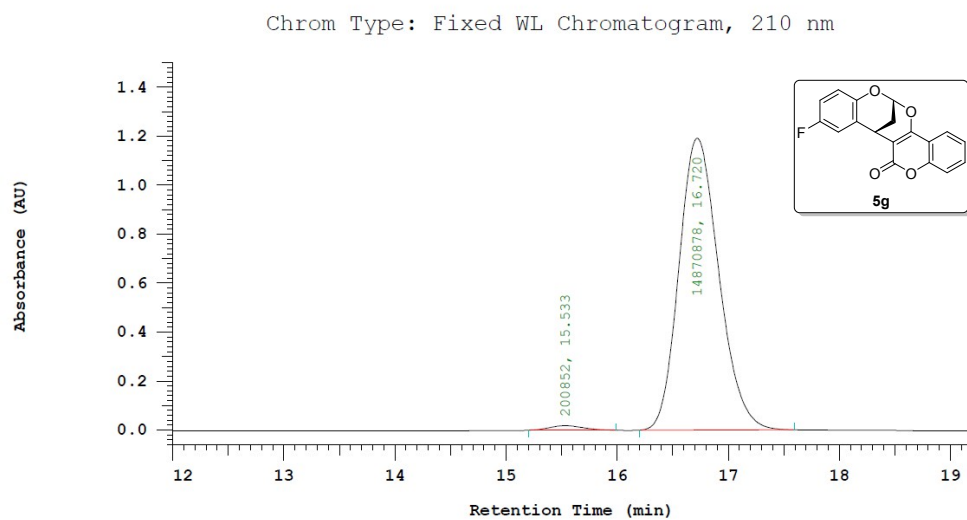
## The HPLC of racemic 5g



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 15.553 | 2954491 | 50.038  | BV |
| 2   | 16.760 | 2949952 | 49.962  | VB |
|     |        | 5904443 | 100.000 |    |

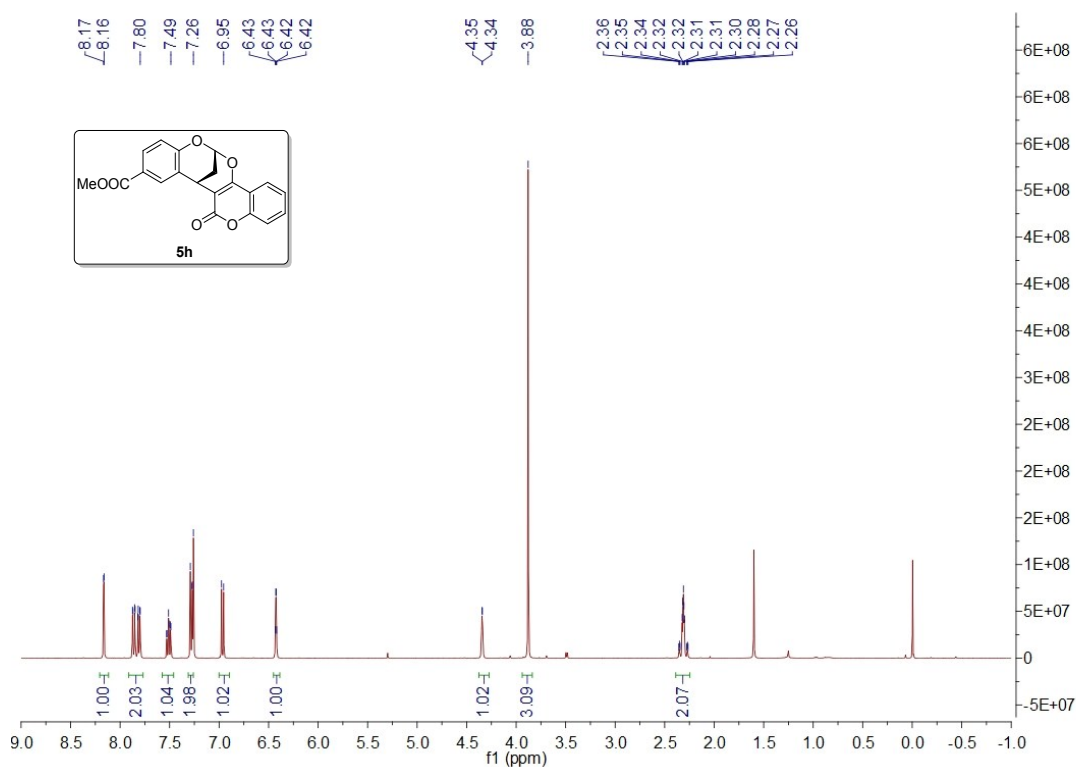
## The HPLC of chiral 5g



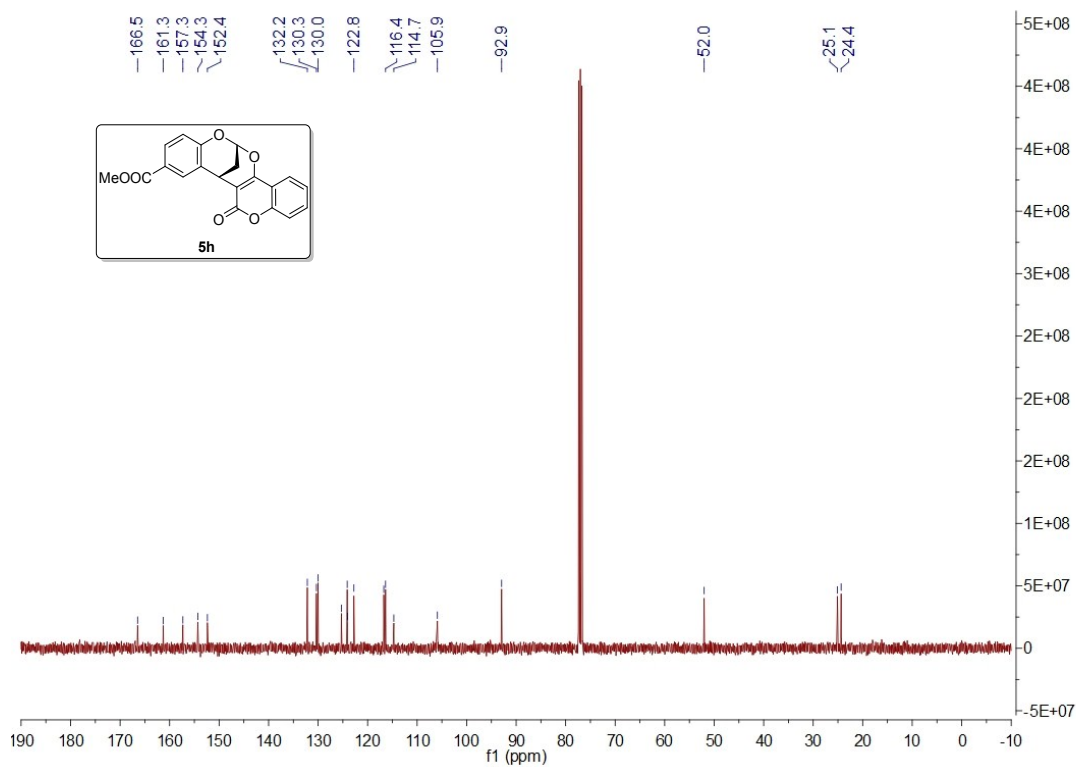
Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 15.533 | 200852   | 1.333   | BB |
| 2   | 16.720 | 14870878 | 98.667  | BB |
|     |        | 15071730 | 100.000 |    |

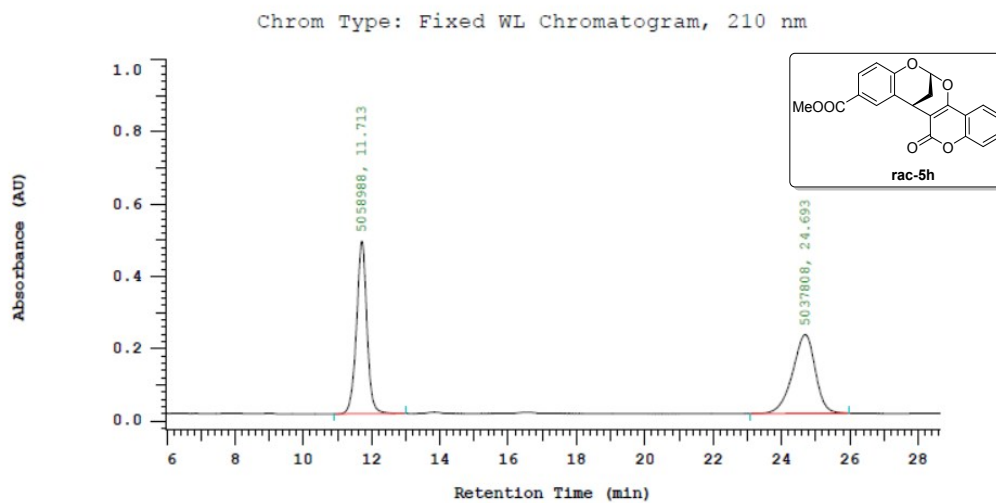
The  $^1\text{H}$  NMR spectrum of 5h (400 MHz,  $\text{CDCl}_3$ )



The  $^{13}\text{C}$  NMR spectrum of 5h (100 MHz,  $\text{CDCl}_3$ )



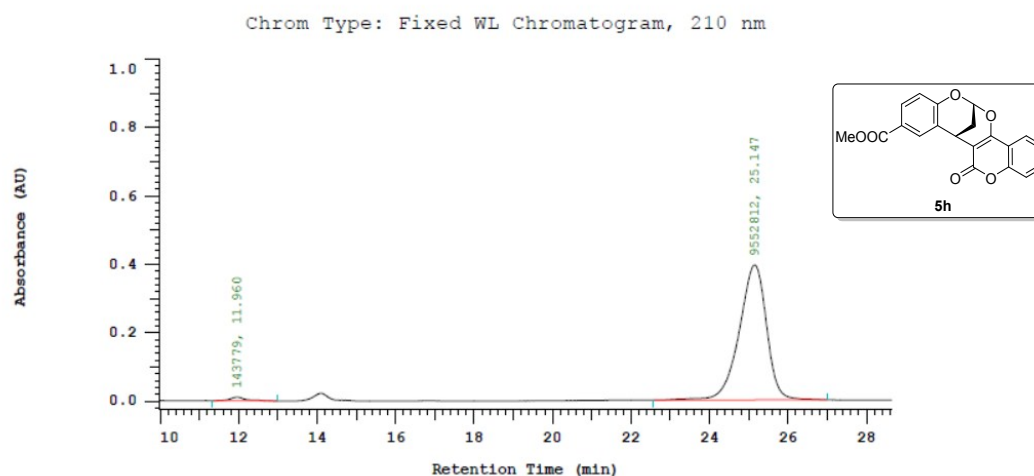
## The HPLC of racemic 5h



Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 11.713 | 5058988  | 50.105  | BB |
| 2   | 24.693 | 5037808  | 49.895  | BB |
|     |        | 10096796 | 100.000 |    |

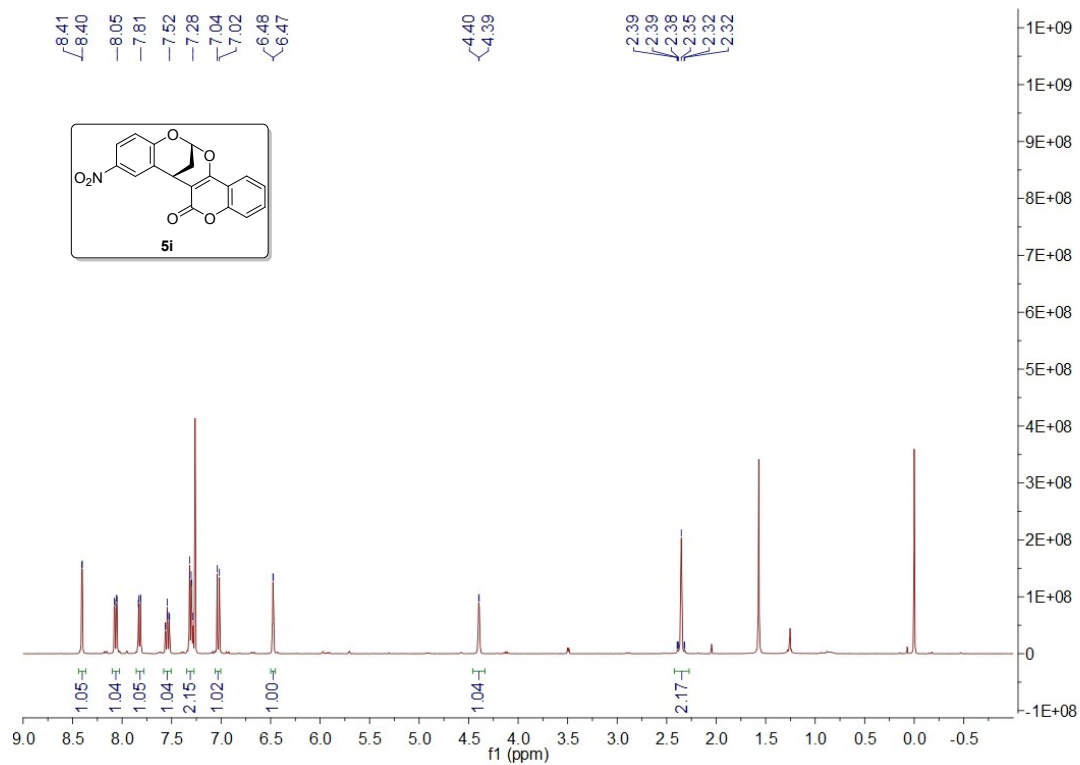
## The HPLC of chiral 5h



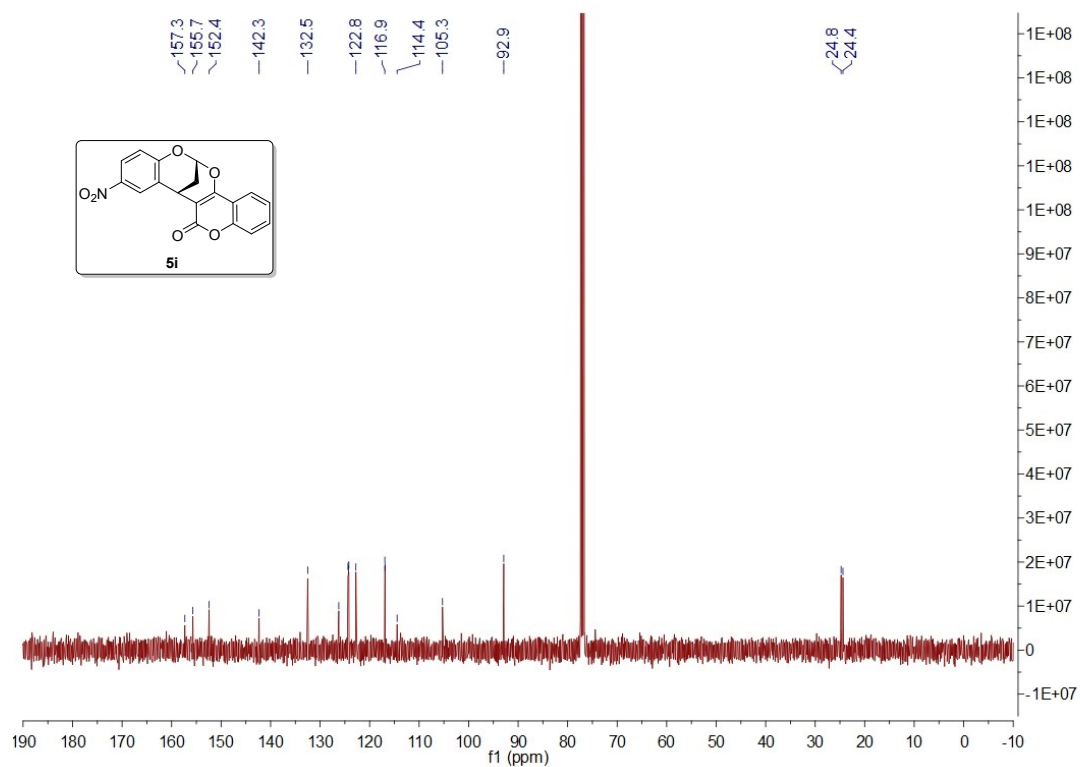
Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 11.960 | 143779  | 1.483   | BB |
| 2   | 25.147 | 9552812 | 98.517  | BB |
|     |        | 9696591 | 100.000 |    |

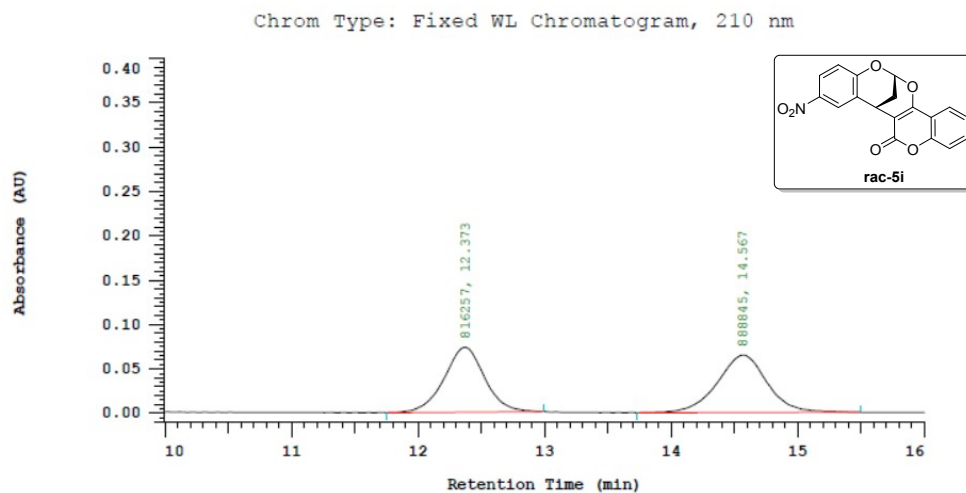
## The $^1\text{H}$ NMR spectrum of 5i (400 MHz, $\text{CDCl}_3$ )



### The <sup>13</sup>C NMR spectrum of **5i** (100 MHz, CDCl<sub>3</sub>)



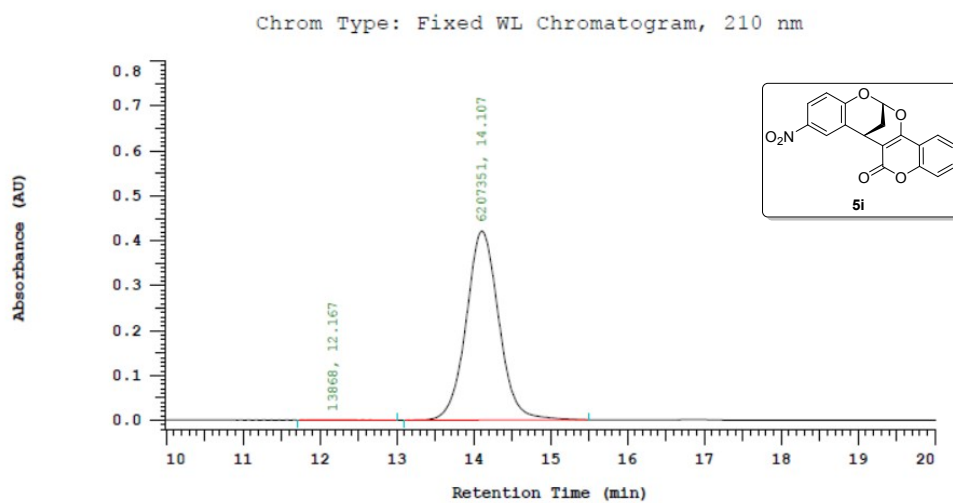
### The HPLC of racemic **5i**



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 12.373 | 816257  | 47.871  | BB |
| 2   | 14.567 | 888845  | 52.129  | BB |
|     |        | 1705102 | 100.000 |    |

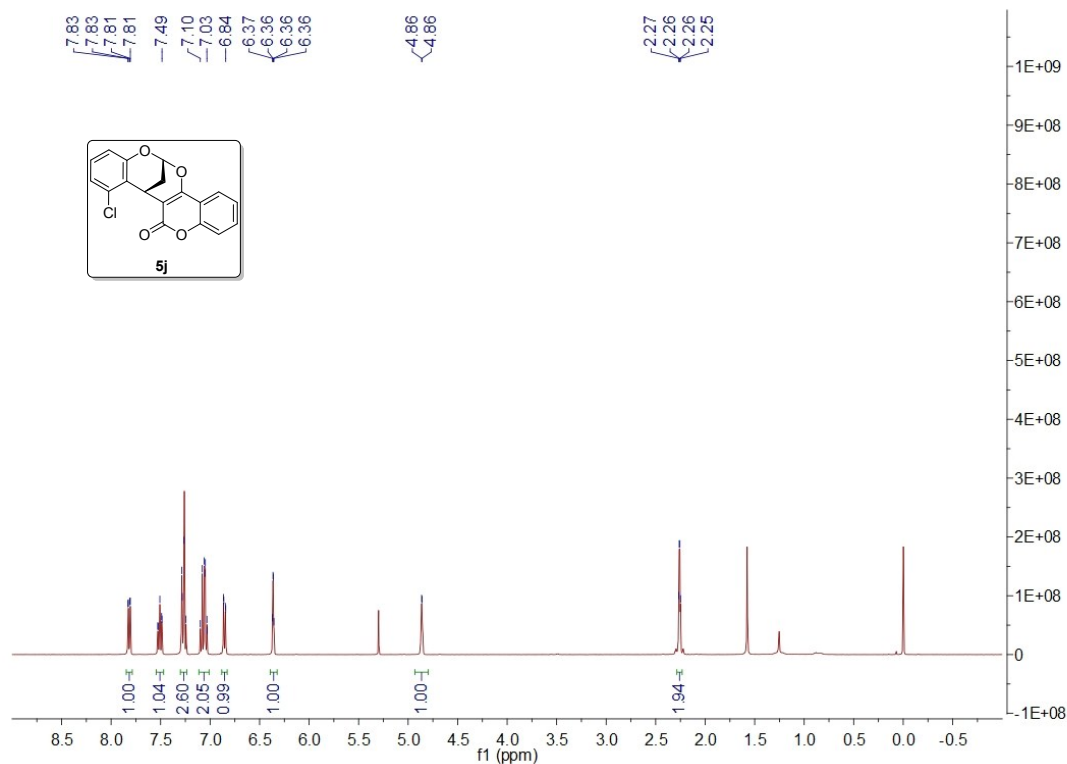
## The HPLC of chiral 5i



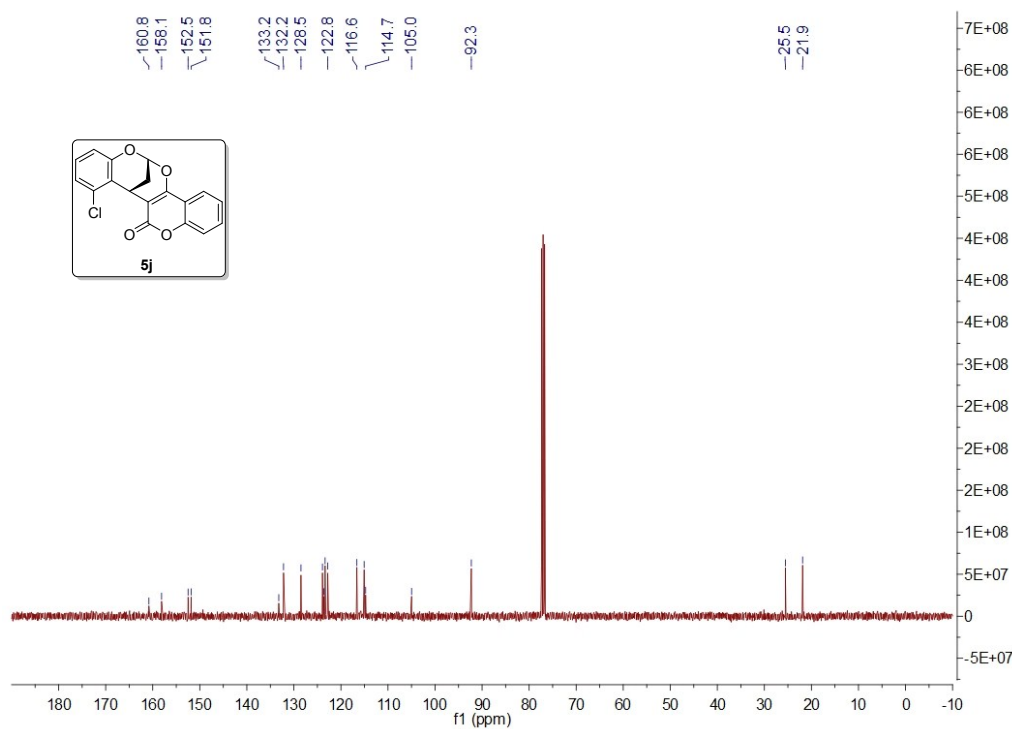
Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 12.167 | 13868   | 0.223   | BB |
| 2   | 14.107 | 6207351 | 99.777  | BB |
|     |        | 6221219 | 100.000 |    |

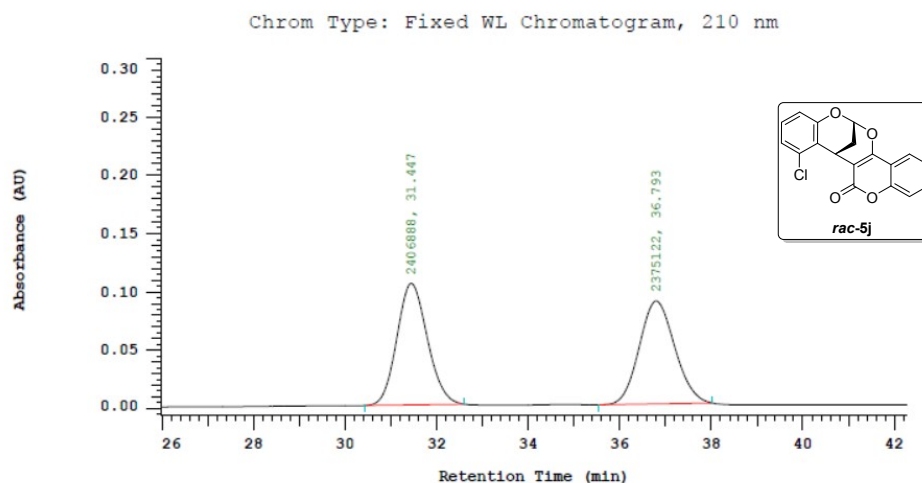
The  $^1\text{H}$  NMR spectrum of 5j (400 MHz,  $\text{CDCl}_3$ )



The  $^{13}\text{C}$  NMR spectrum of 5j (100 MHz,  $\text{CDCl}_3$ )



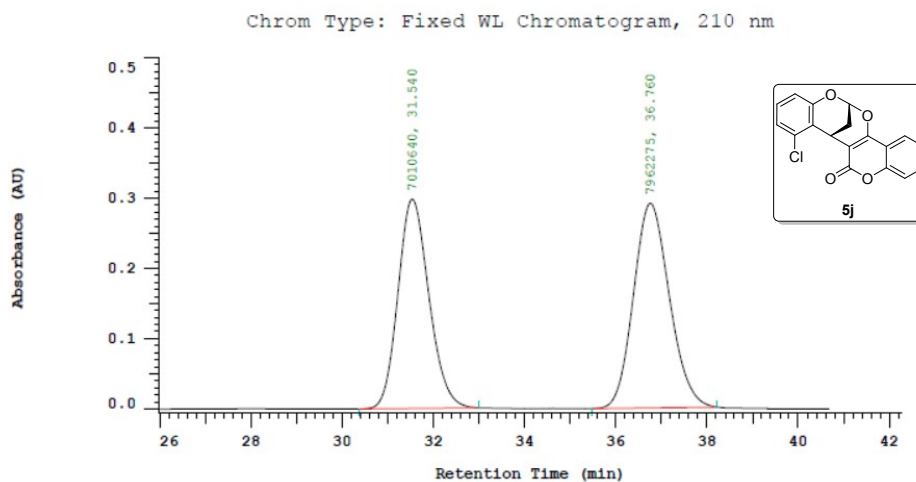
## The HPLC of racemic 5j



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 31.447 | 2406888 | 50.332  | BB |
| 2   | 36.793 | 2375122 | 49.668  | BB |
|     |        | 4782010 | 100.000 |    |

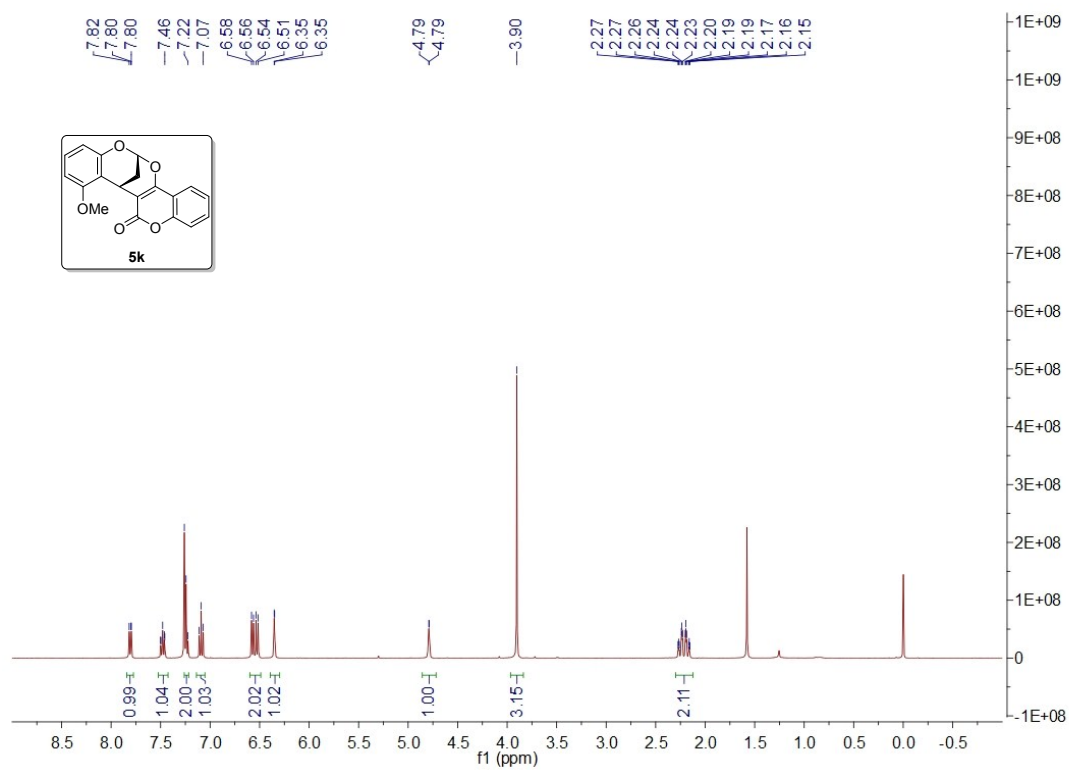
## The HPLC of chiral 5j



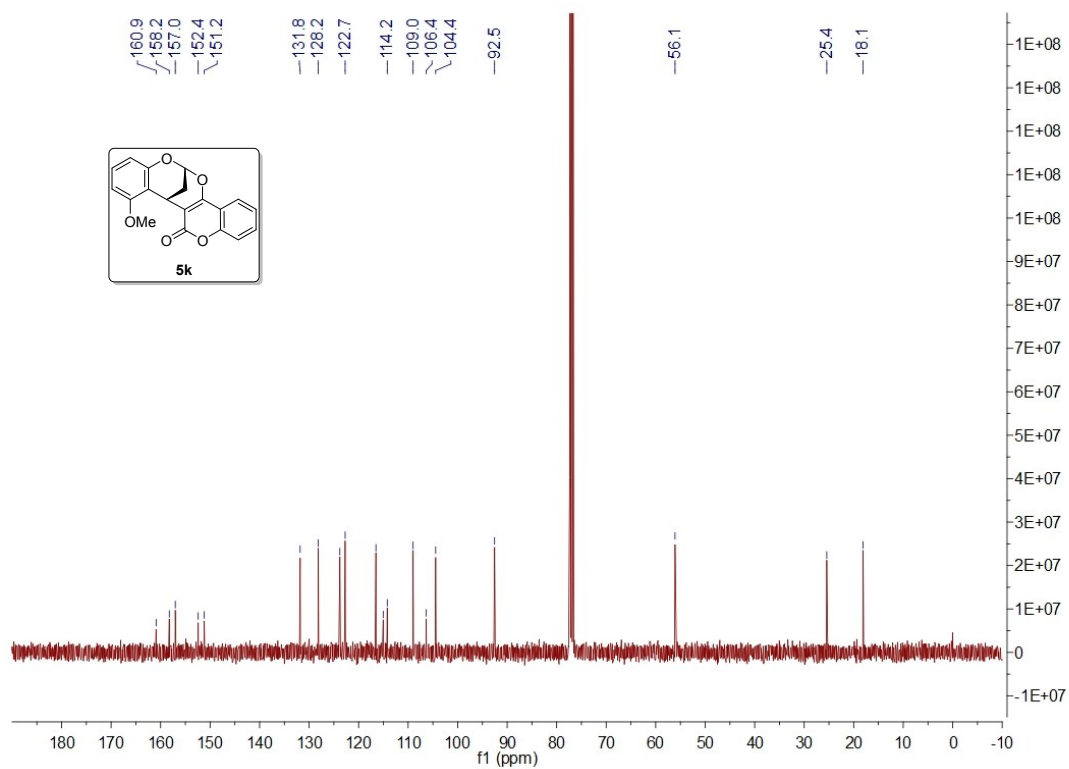
Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 31.540 | 7010640  | 46.822  | BB |
| 2   | 36.760 | 7962275  | 53.178  | BB |
|     |        | 14972915 | 100.000 |    |

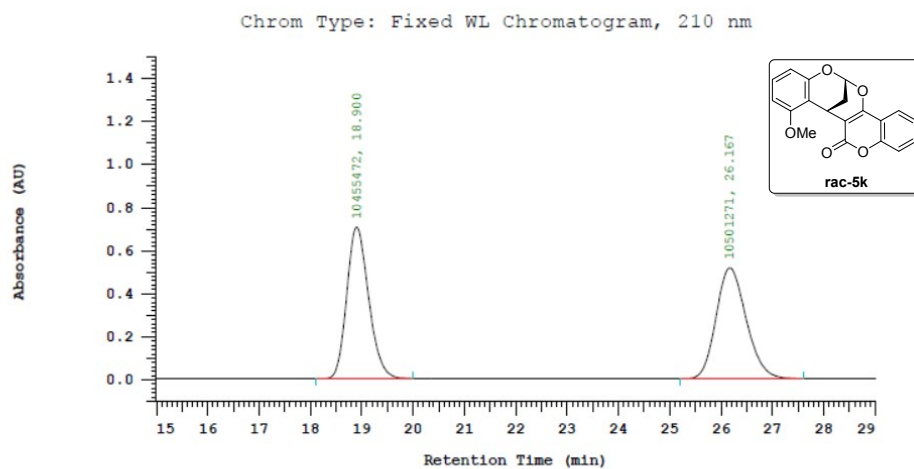
The  $^1\text{H}$  NMR spectrum of 5k (400 MHz,  $\text{CDCl}_3$ )



The  $^{13}\text{C}$  NMR spectrum of 5k (100 MHz,  $\text{CDCl}_3$ )



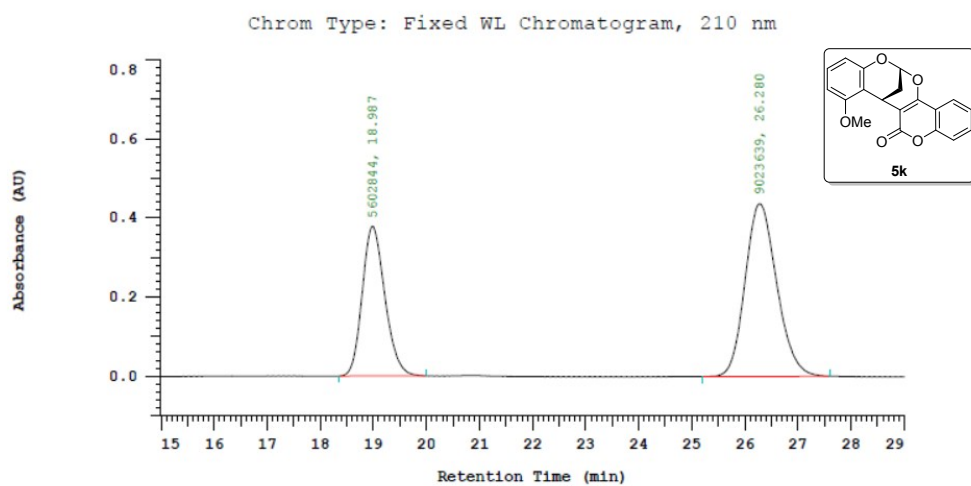
## The HPLC of racemic 5k



Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 18.900 | 10455472 | 49.891  | BB |
| 2   | 26.167 | 10501271 | 50.109  | BB |
|     |        | 20956743 | 100.000 |    |

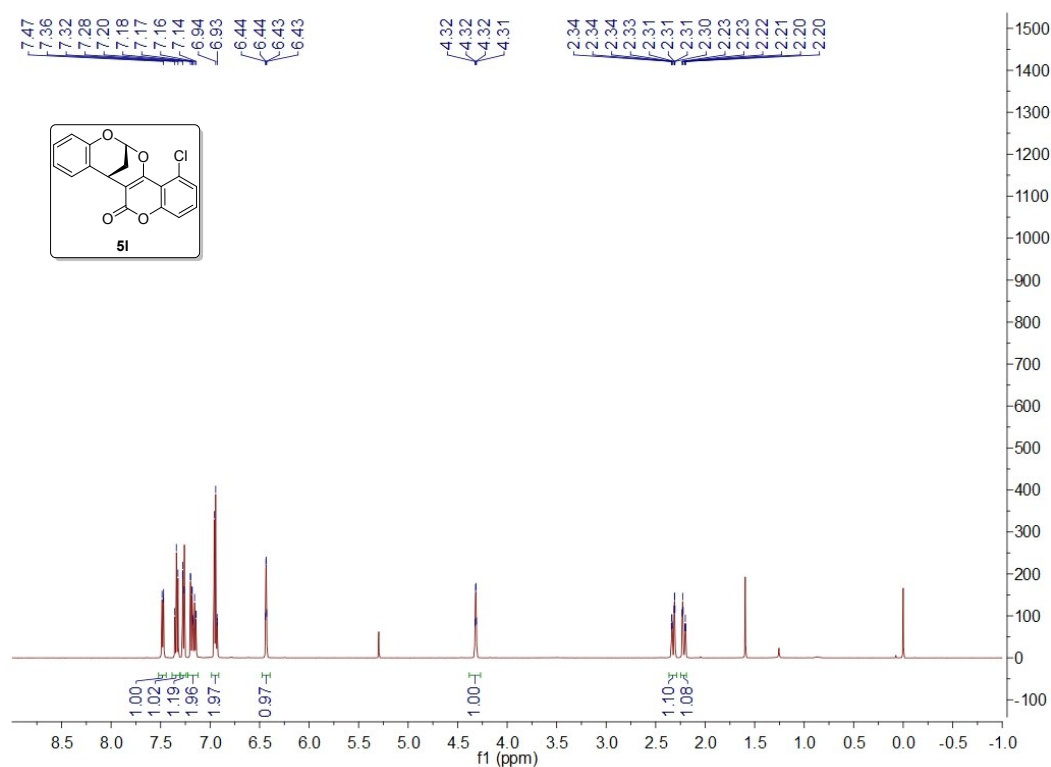
## The HPLC of chiral 5k



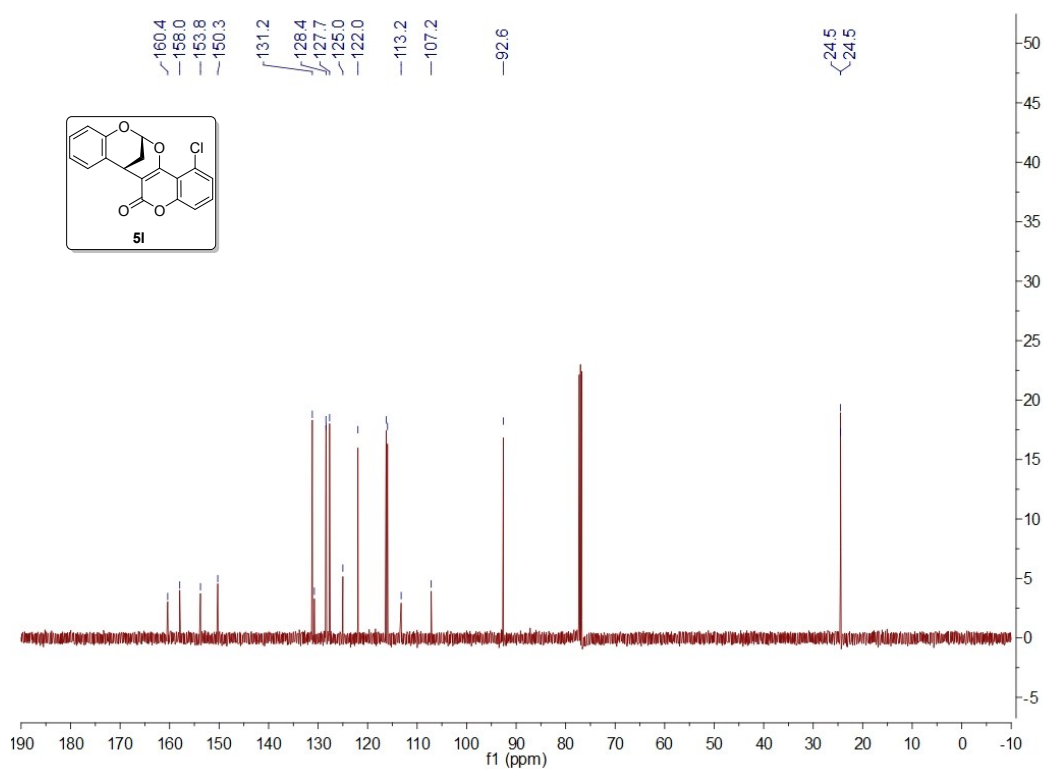
Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 18.987 | 5602844  | 38.306  | BB |
| 2   | 26.280 | 9023639  | 61.694  | BB |
|     |        | 14626483 | 100.000 |    |

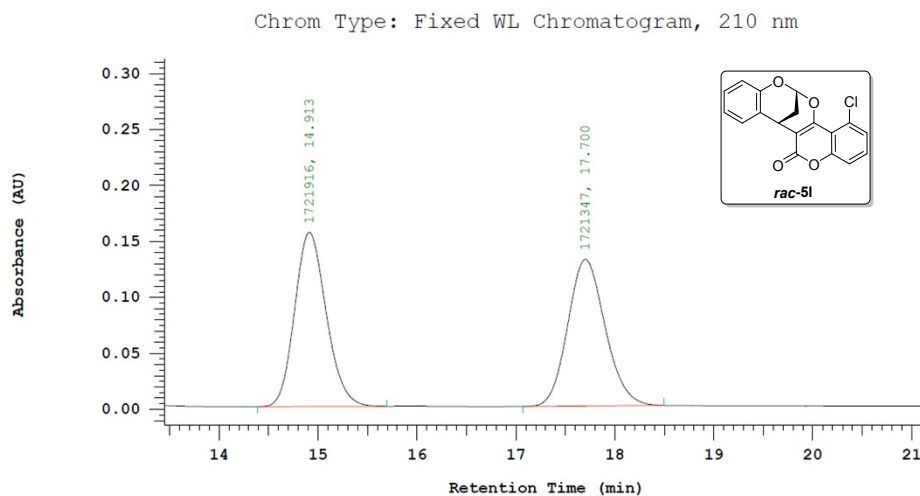
### The $^1\text{H}$ NMR spectrum of 5l (500 MHz, $\text{CDCl}_3$ )



### The $^{13}\text{C}$ NMR spectrum of 5l (125 MHz, $\text{CDCl}_3$ )



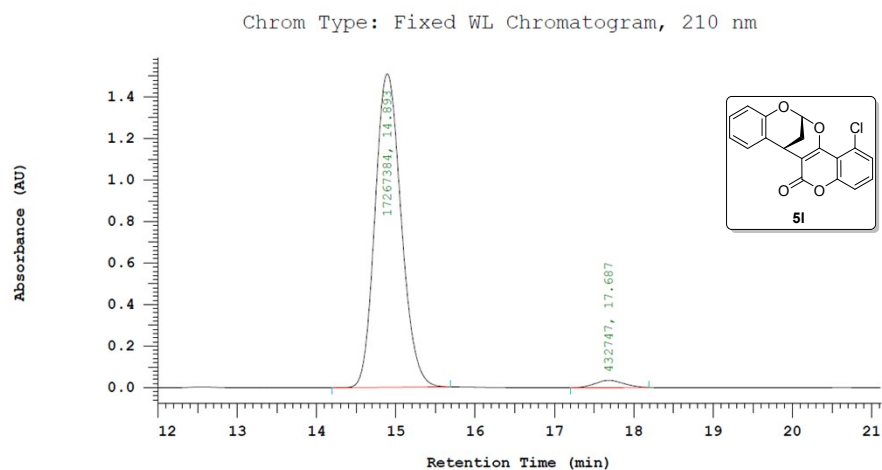
## The HPLC of racemic 5l



Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 14.913 | 1721916 | 50.008  | BB |
| 2   | 17.700 | 1721347 | 49.992  | BB |
|     |        | 3443263 | 100.000 |    |

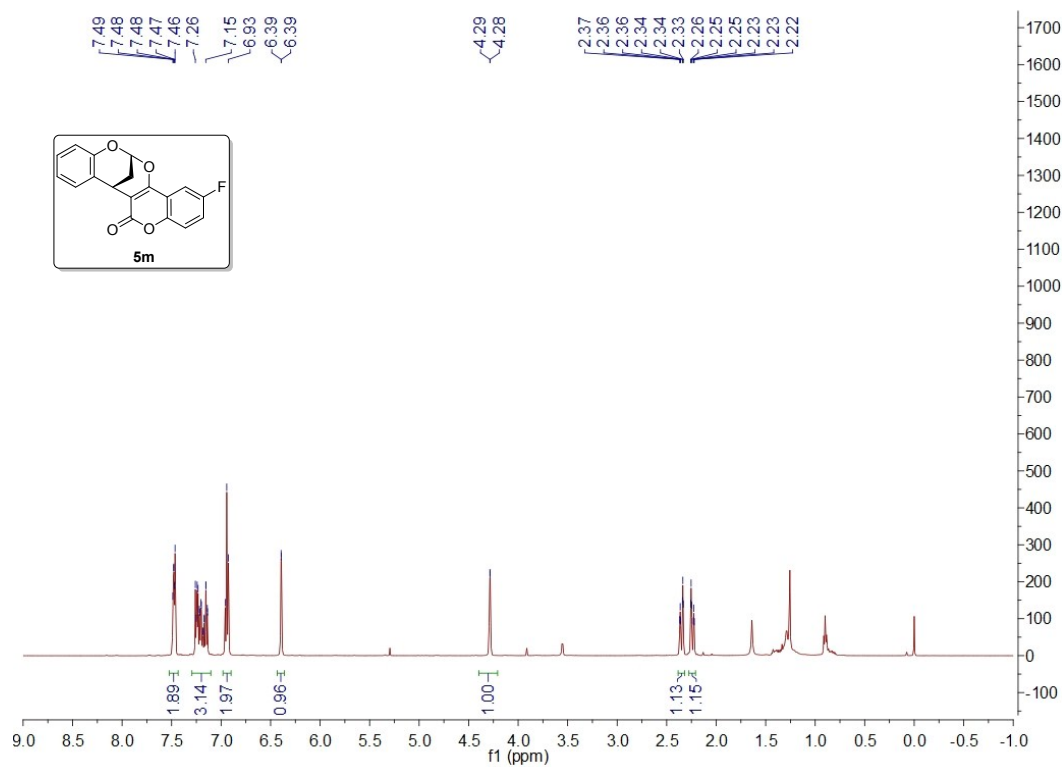
## The HPLC of chiral 5l



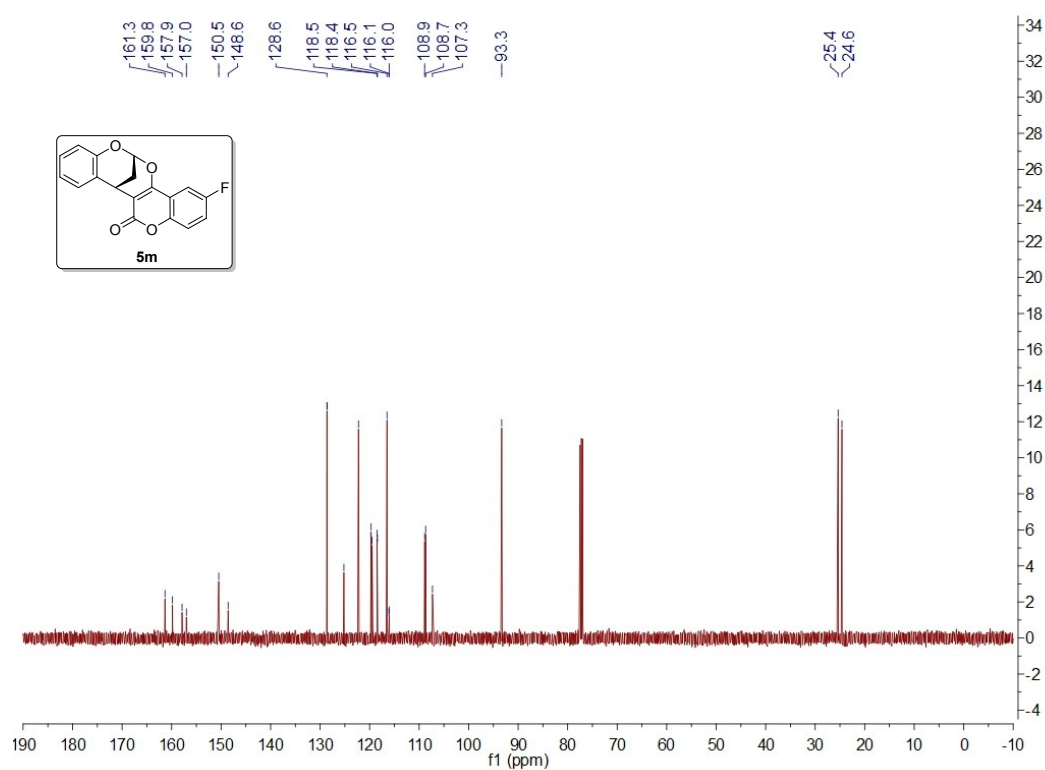
Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 14.893 | 17267384 | 97.555  | BB |
| 2   | 17.687 | 432747   | 2.445   | BB |
|     |        | 17700131 | 100.000 |    |

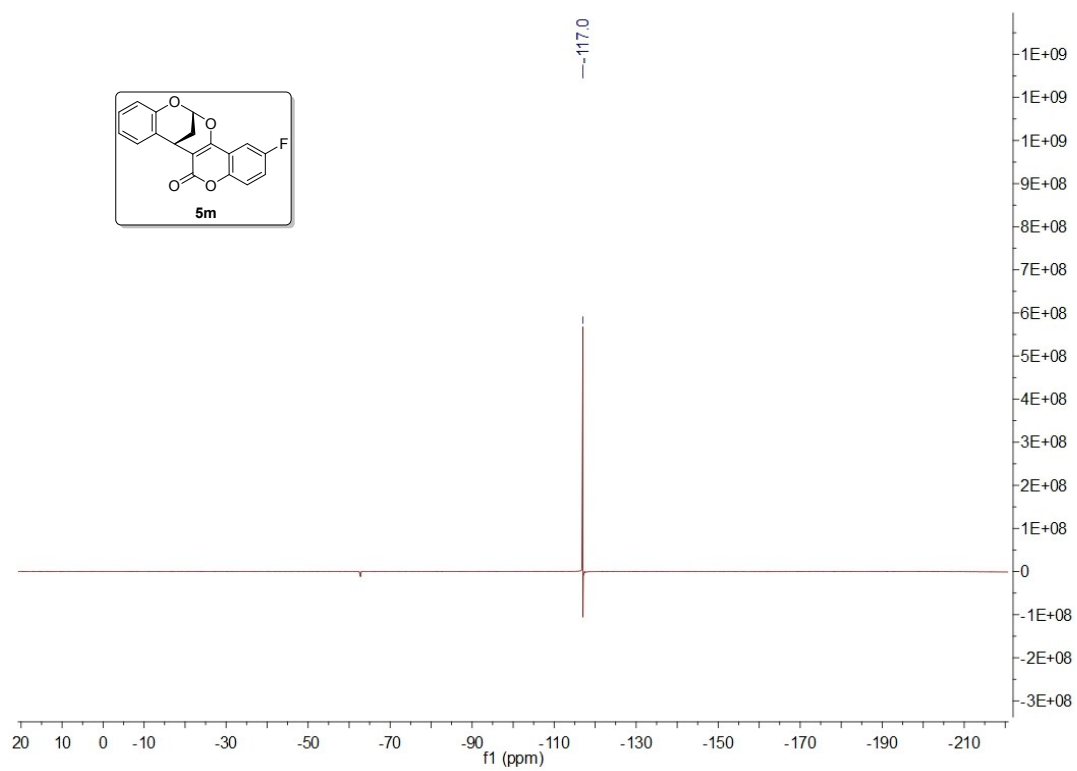
## The <sup>1</sup>H NMR spectrum of 5m (500 MHz, CDCl<sub>3</sub>)



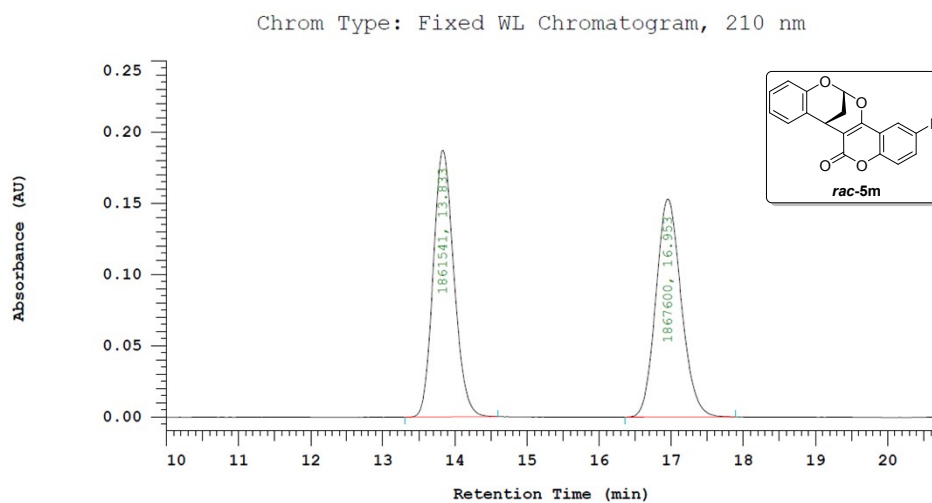
The <sup>13</sup>C NMR spectrum of 5m (125 MHz, CDCl<sub>3</sub>)



The <sup>19</sup>F NMR spectrum of 5m (400 MHz, CDCl<sub>3</sub>)



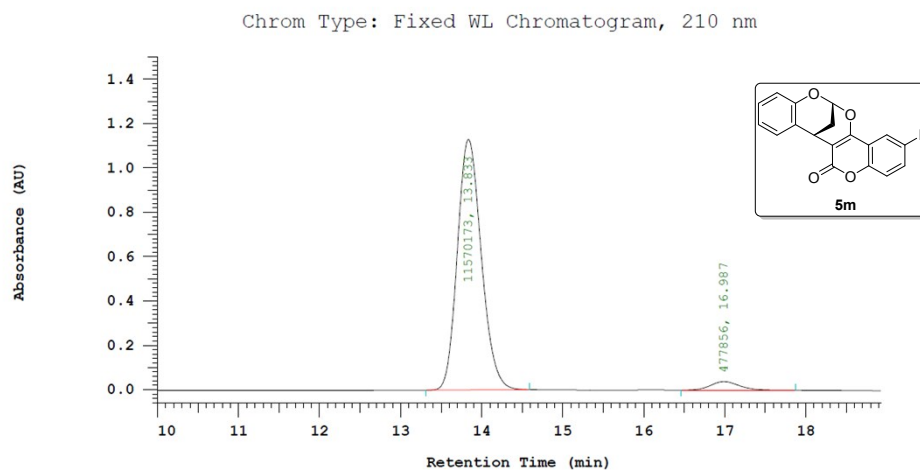
## The HPLC of racemic 5m



Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 13.833 | 1861541 | 49.919  | BB |
| 2   | 16.953 | 1867600 | 50.081  | BB |
|     |        | 3729141 | 100.000 |    |

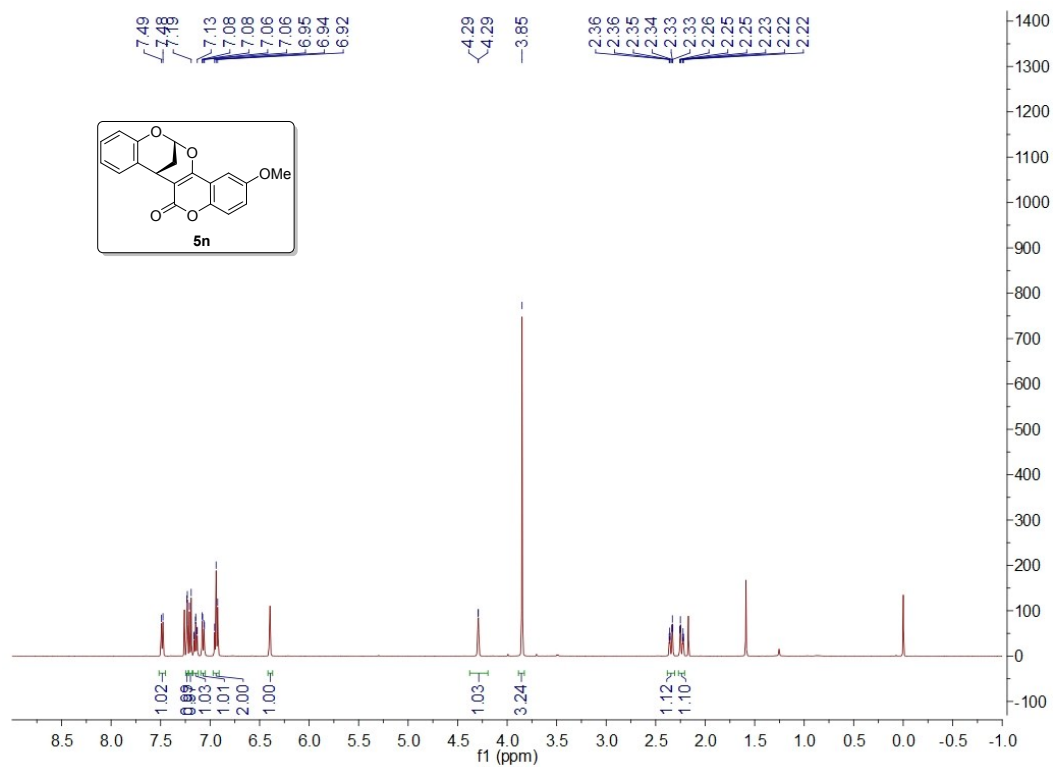
## The HPLC of chiral 5m



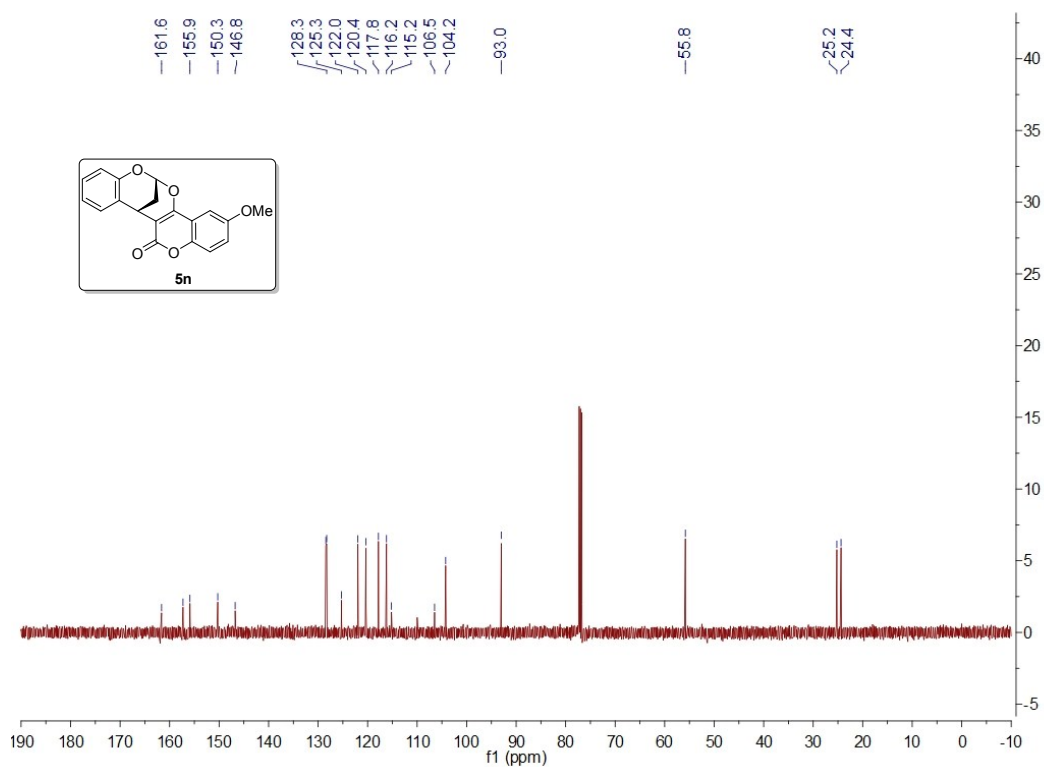
Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 13.833 | 11570173 | 96.034  | BB |
| 2   | 16.987 | 477856   | 3.966   | BB |
|     |        | 12048029 | 100.000 |    |

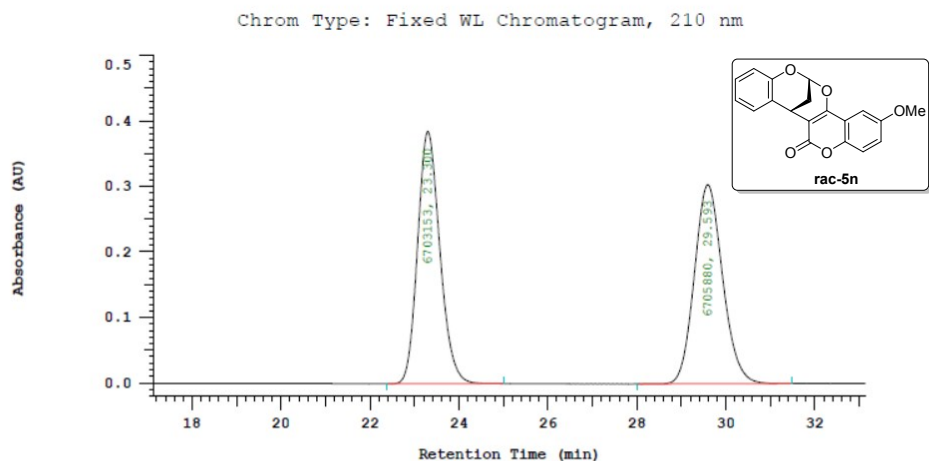
## The <sup>1</sup>H NMR spectrum of 5n (500 MHz, CDCl<sub>3</sub>)



### The <sup>13</sup>C NMR spectrum of 5n (125 MHz, CDCl<sub>3</sub>)



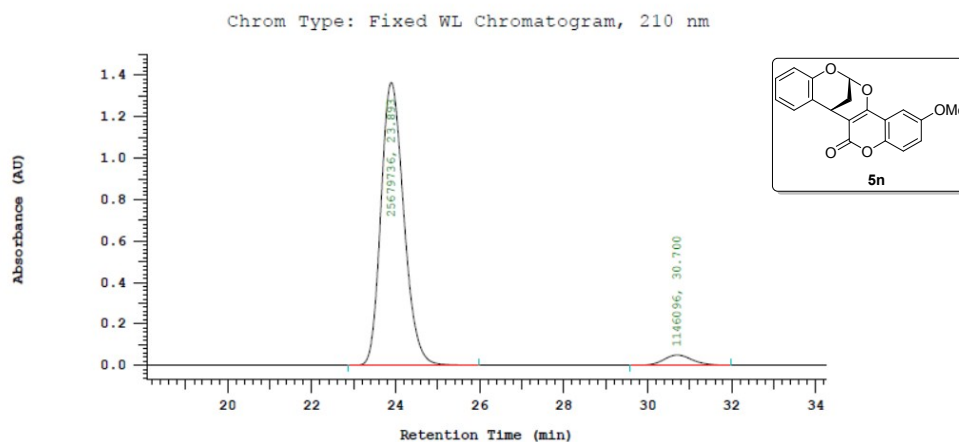
### The HPLC of racemic 5n



Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 23.300 | 6703153  | 49.990  | BB |
| 2   | 29.593 | 6705880  | 50.010  | BB |
|     |        | 13409033 | 100.000 |    |

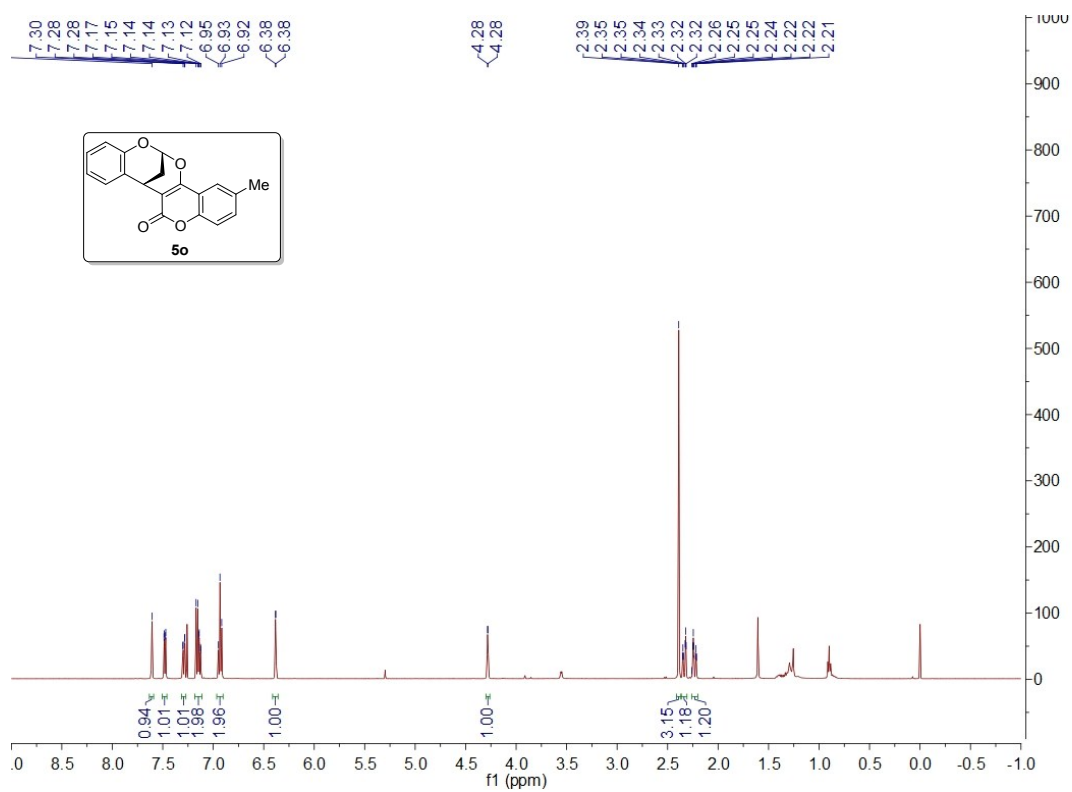
## The HPLC of chiral 5n



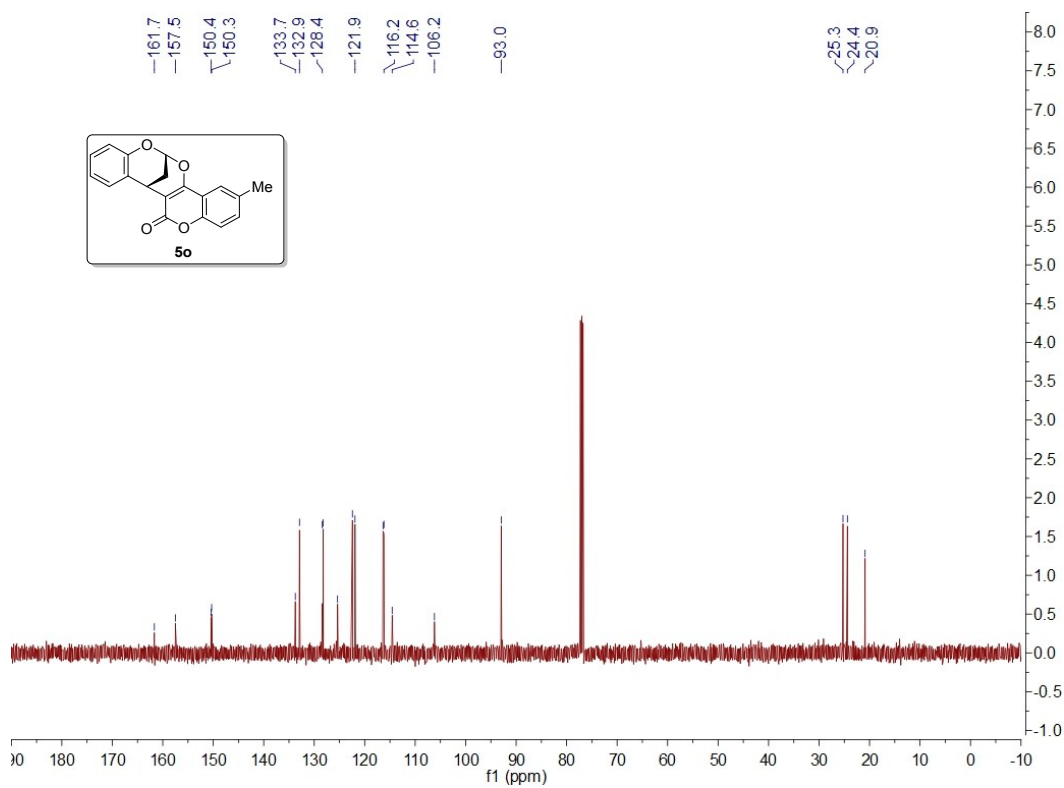
Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 23.893 | 25679736 | 95.728  | BB |
| 2   | 30.700 | 1146096  | 4.272   | BB |
|     |        | 26825832 | 100.000 |    |

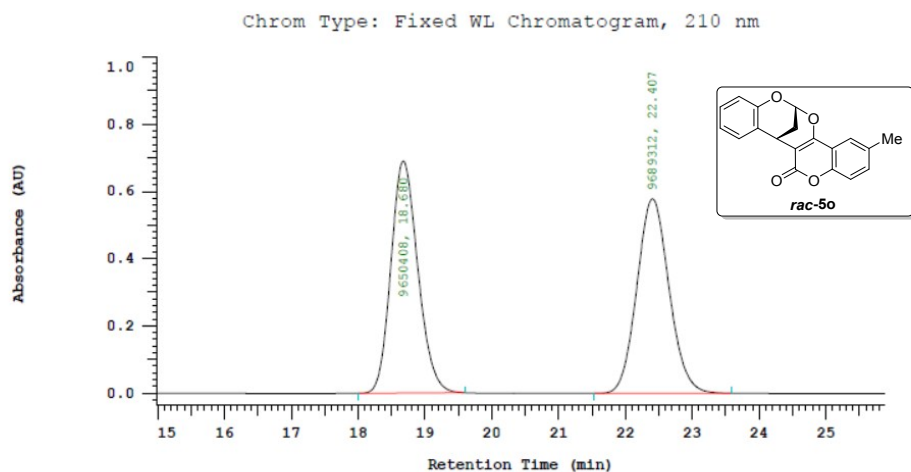
## The $^1\text{H}$ NMR spectrum of 5o (500 MHz, $\text{CDCl}_3$ )



### The <sup>13</sup>C NMR spectrum of 5o (125 MHz, CDCl<sub>3</sub>)



### The HPLC of racemic 5o

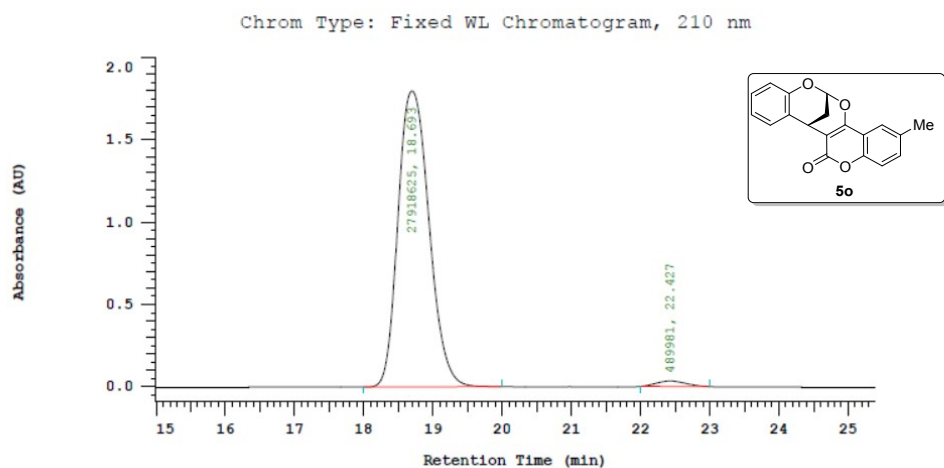


Chrom Type: Fixed WL Chromatogram, 210 nm

Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 18.680 | 9650408  | 49.899  | BB |
| 2   | 22.407 | 9689312  | 50.101  | BB |
|     |        | 19339720 | 100.000 |    |

## The HPLC of chiral 5o

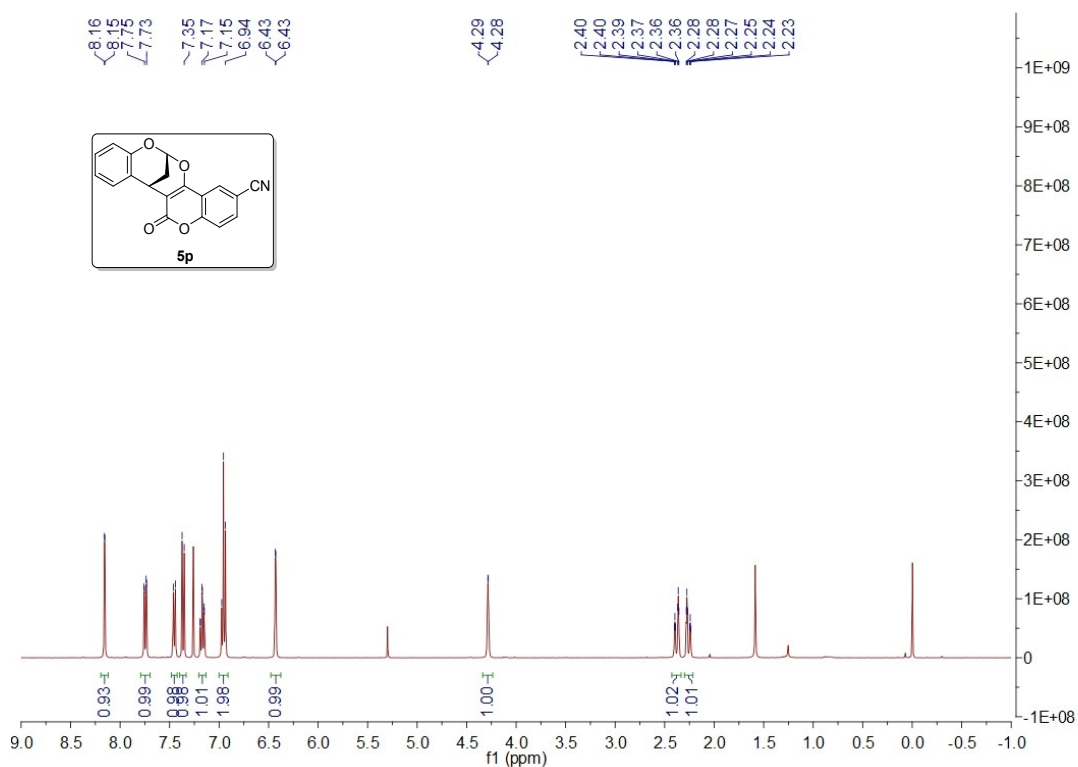


Chrom Type: Fixed WL Chromatogram, 210 nm

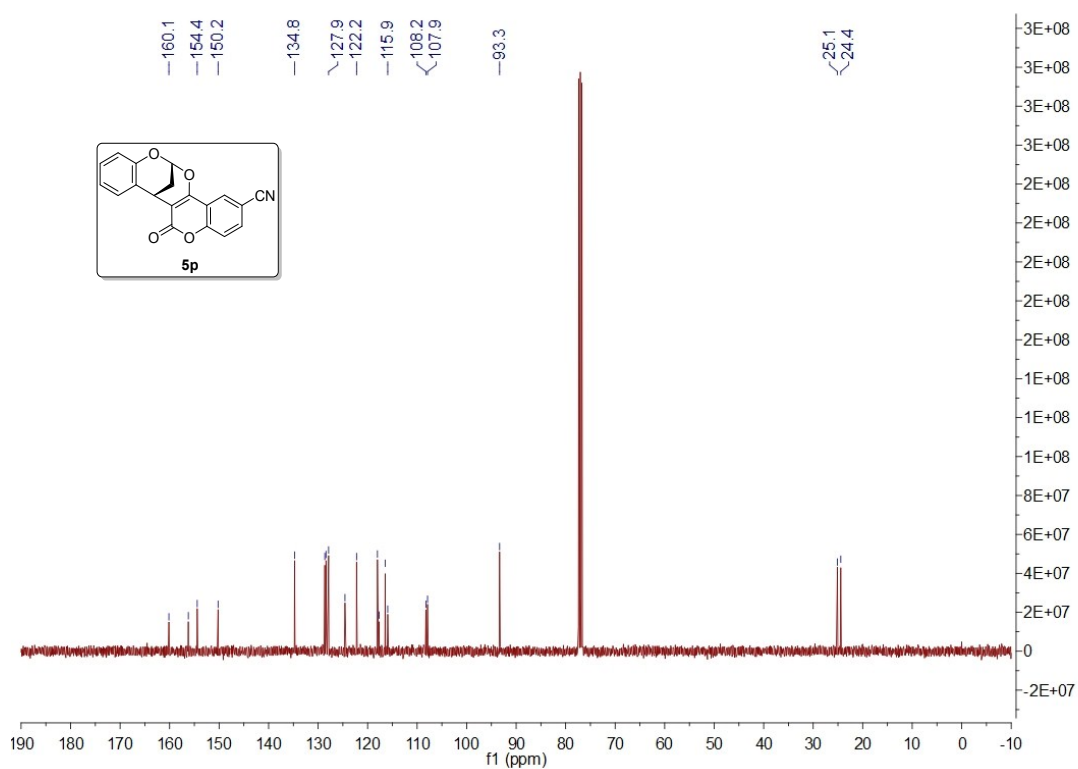
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 18.693 | 27918625 | 98.275  | BB |
| 2   | 22.427 | 489981   | 1.725   | BB |
|     |        | 28408606 | 100.000 |    |

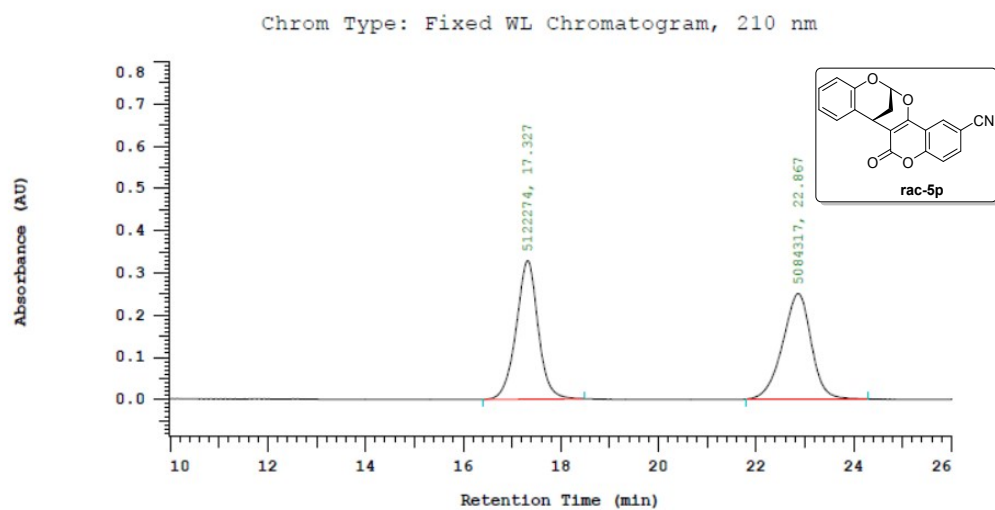
### The $^1\text{H}$ NMR spectrum of 5p (400 MHz, $\text{CDCl}_3$ )



### The $^{13}\text{C}$ NMR spectrum of 5p (100 MHz, $\text{CDCl}_3$ )



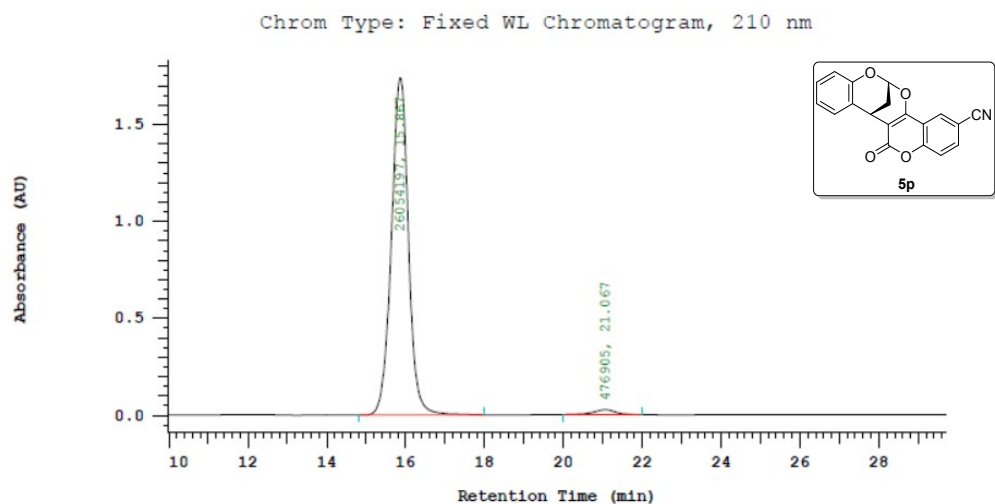
## The HPLC of racemic 5p



Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 17.327 | 5122274  | 50.186  | BB |
| 2   | 22.867 | 5084317  | 49.814  | BB |
|     |        | 10206591 | 100.000 |    |

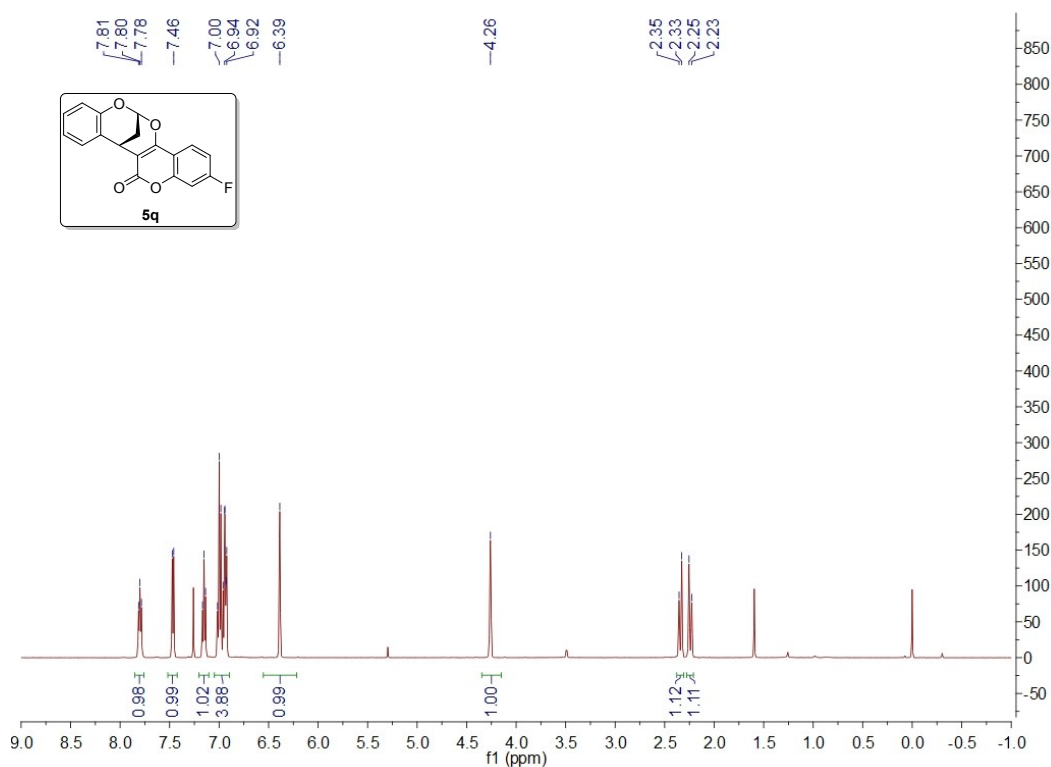
## The HPLC of chiral 5p



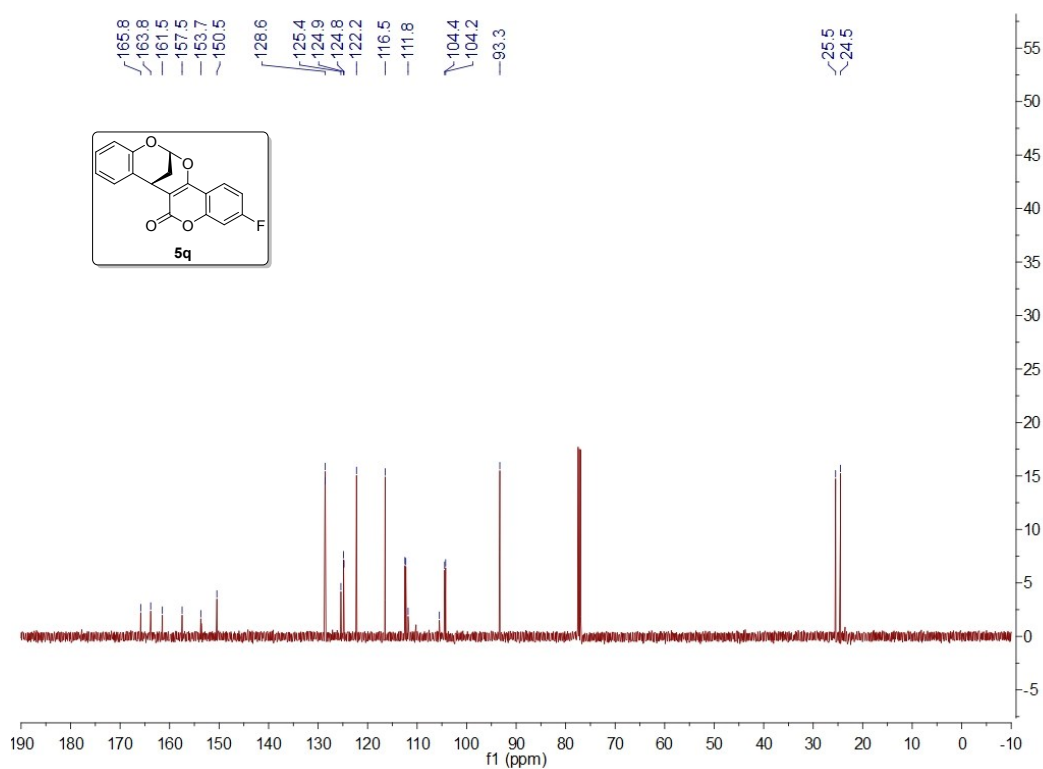
Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 15.867 | 26054197 | 98.202  | BB |
| 2   | 21.067 | 476905   | 1.798   | BB |
|     |        | 26531102 | 100.000 |    |

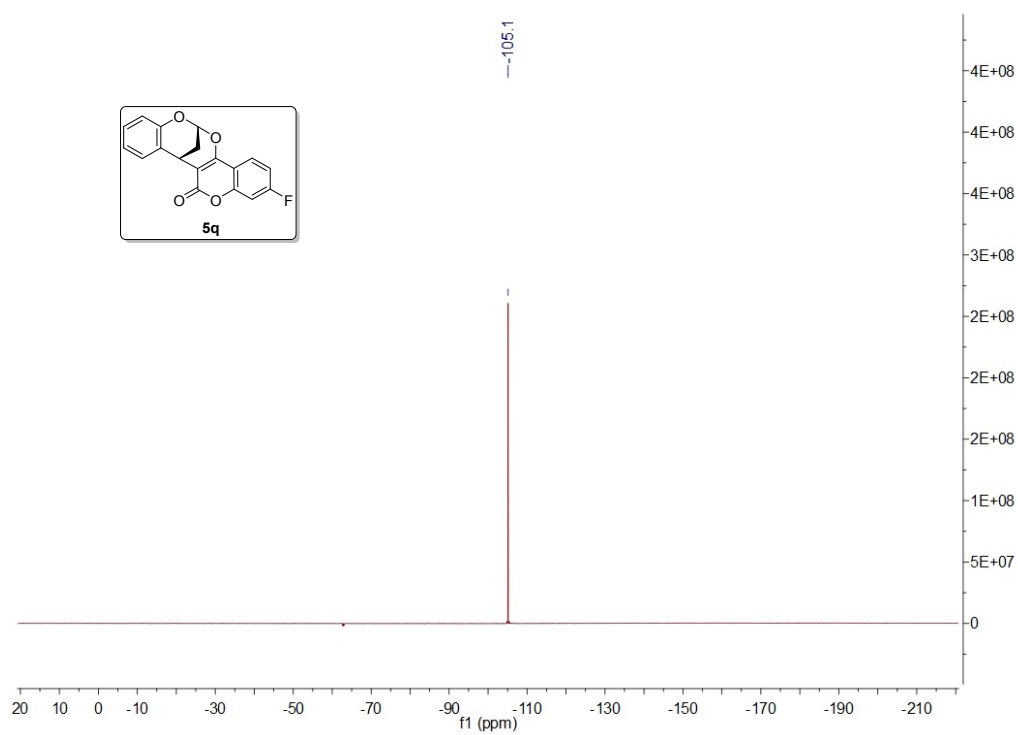
**The  $^1\text{H}$  NMR spectrum of 5q (500 MHz,  $\text{CDCl}_3$ )**



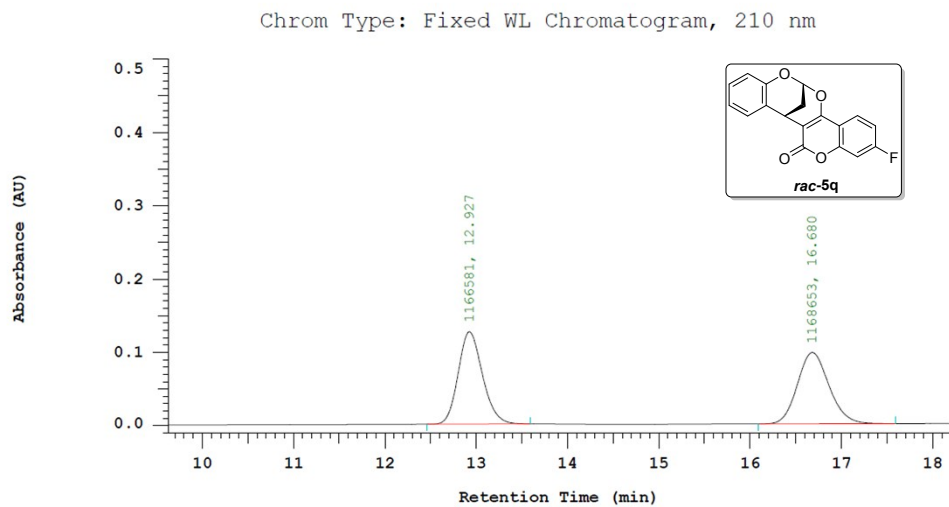
**The  $^{13}\text{C}$  NMR spectrum of 5q (125 MHz,  $\text{CDCl}_3$ )**



**The  $^{19}\text{F}$  NMR spectrum of 5q (400 MHz,  $\text{CDCl}_3$ )**



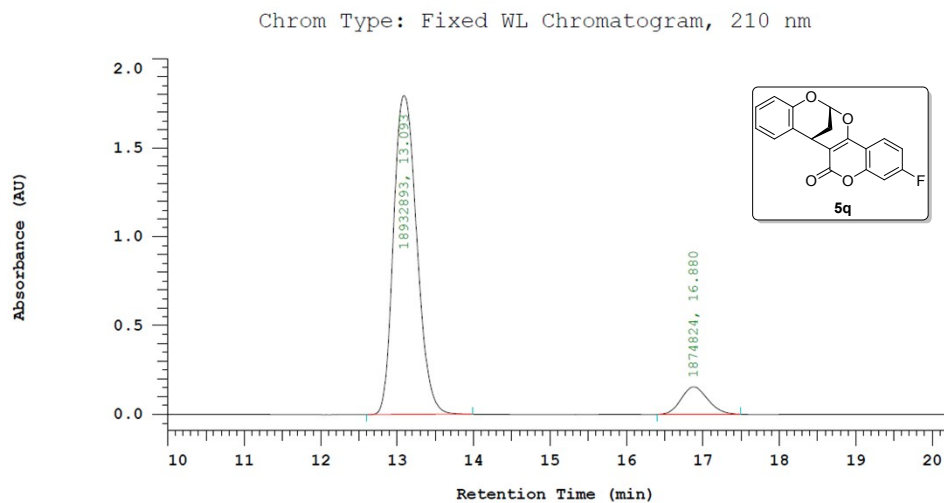
## The HPLC of racemic 5q



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 12.927 | 1166581 | 49.956  | BB |
| 2   | 16.680 | 1168653 | 50.044  | BB |
|     |        | 2335234 | 100.000 |    |

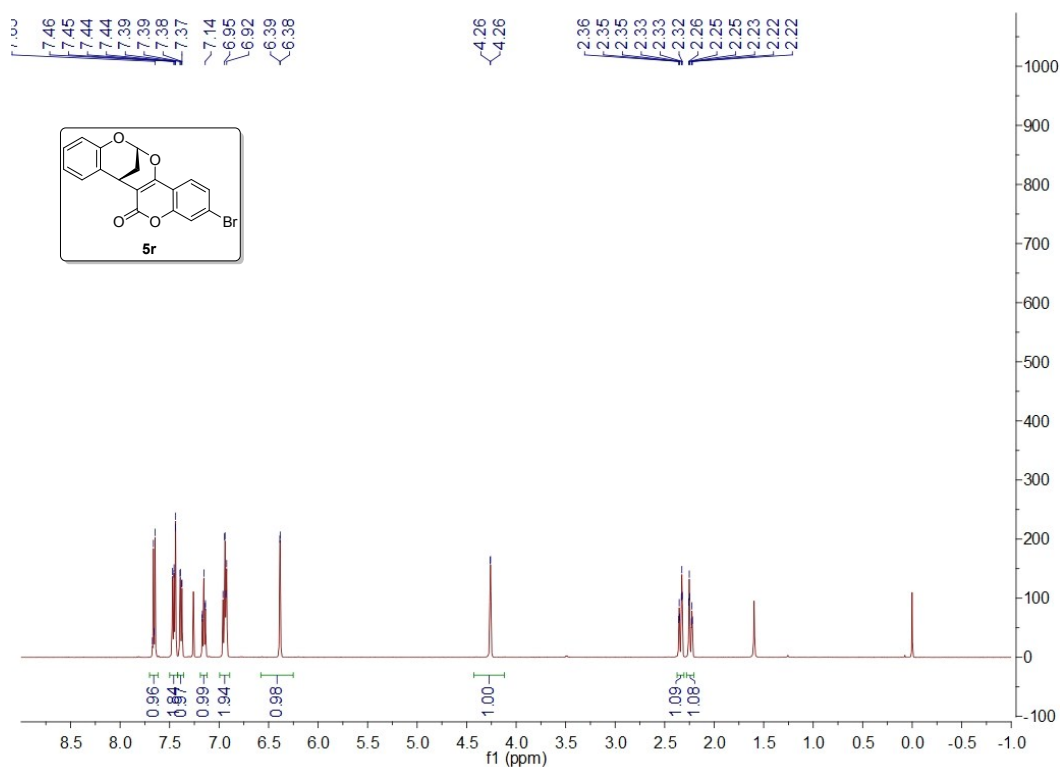
## The HPLC of chiral 5q



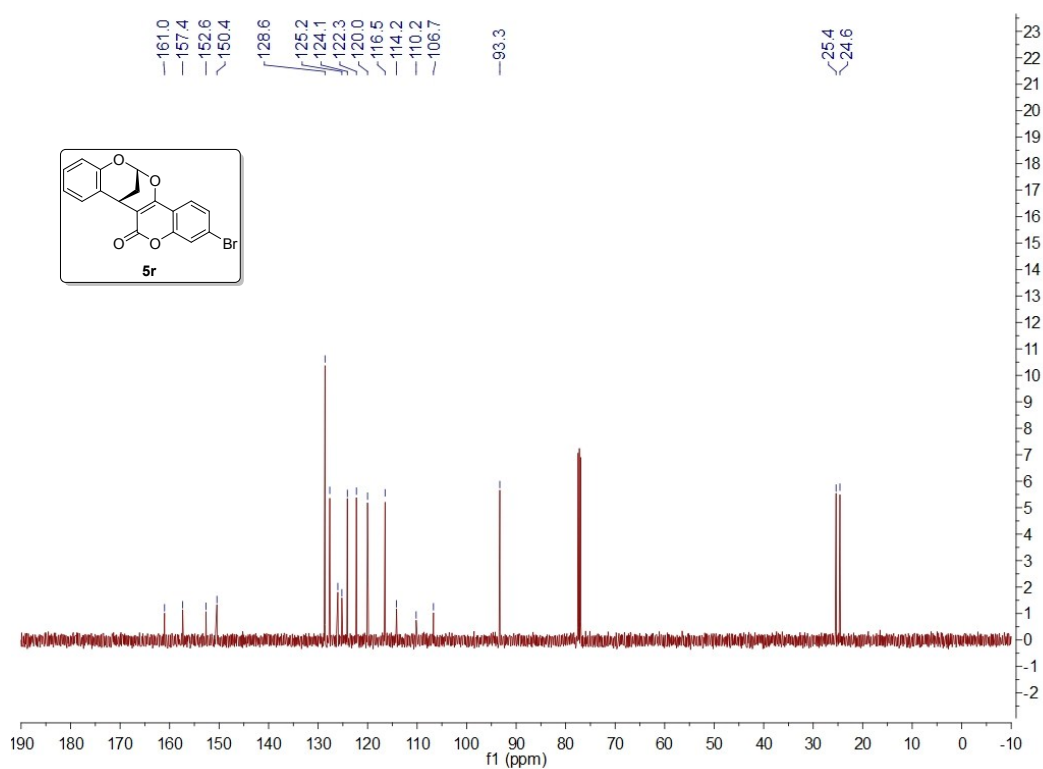
Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 13.093 | 18932893 | 90.990  | BB |
| 2   | 16.880 | 1874824  | 9.010   | BB |
|     |        | 20807717 | 100.000 |    |

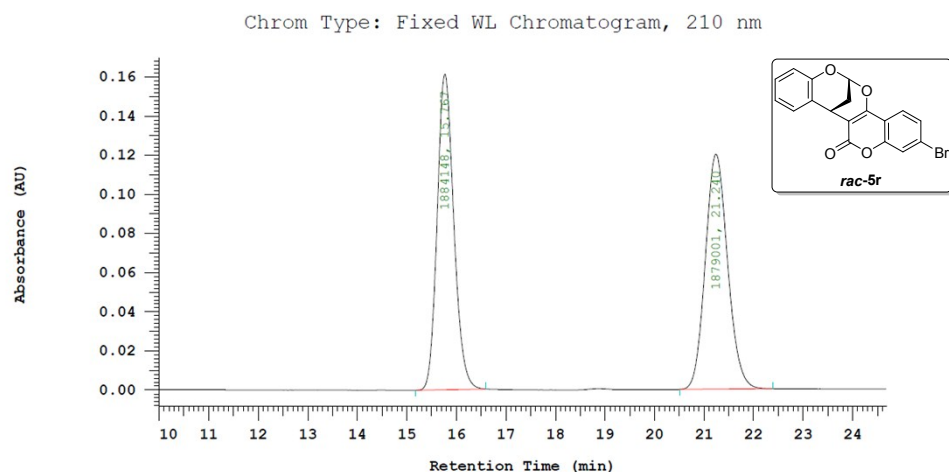
**The  $^1\text{H}$  NMR spectrum of 5r (500 MHz,  $\text{CDCl}_3$ )**



**The  $^{13}\text{C}$  NMR spectrum of 5r (125 MHz,  $\text{CDCl}_3$ )**



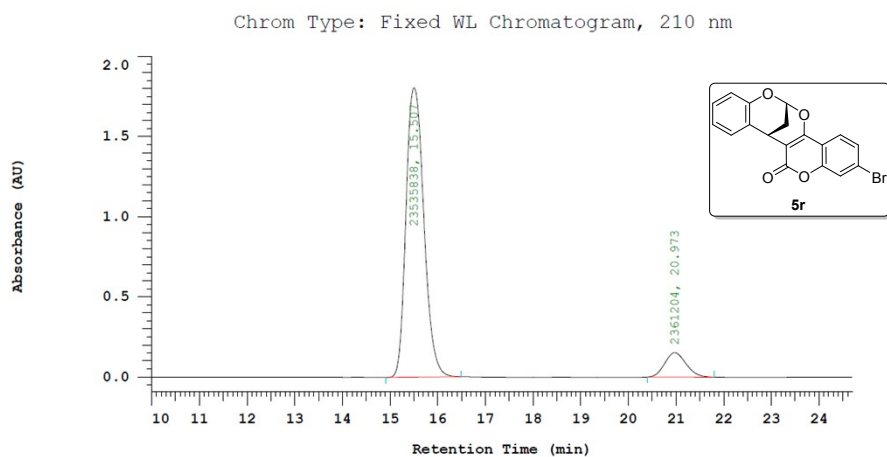
## The HPLC of racemic 5r



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 15.767 | 1884148 | 50.068  | BB |
| 2   | 21.240 | 1879001 | 49.932  | BB |
|     |        | 3763149 | 100.000 |    |

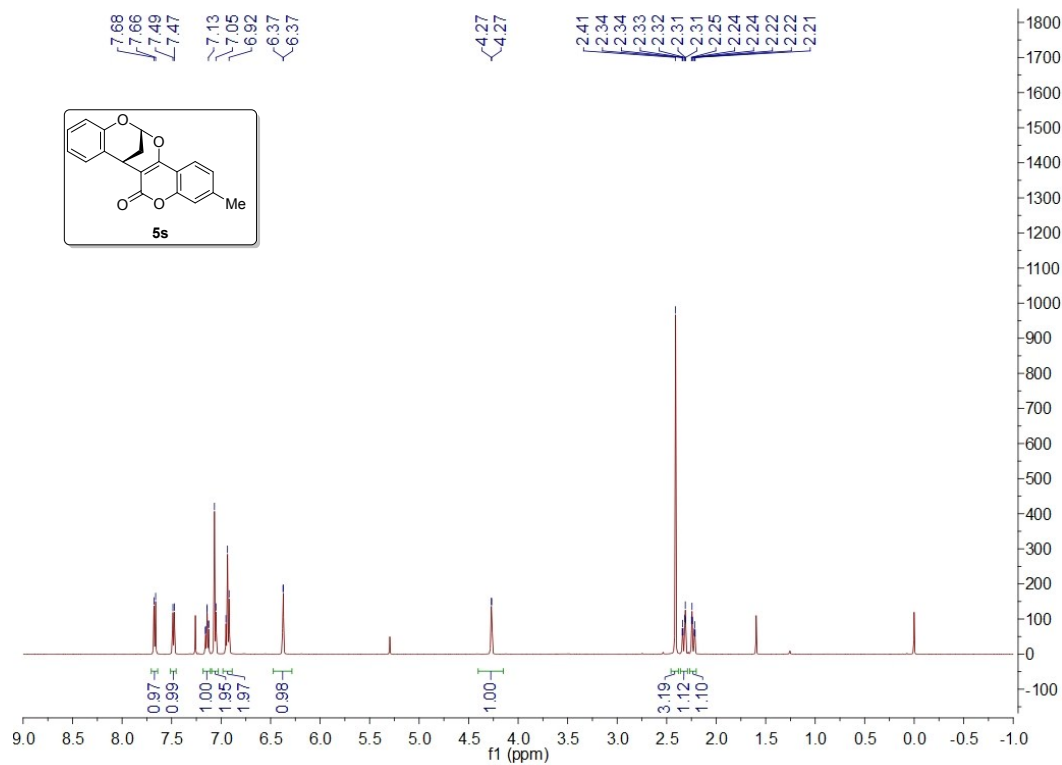
## The HPLC of chiral 5r



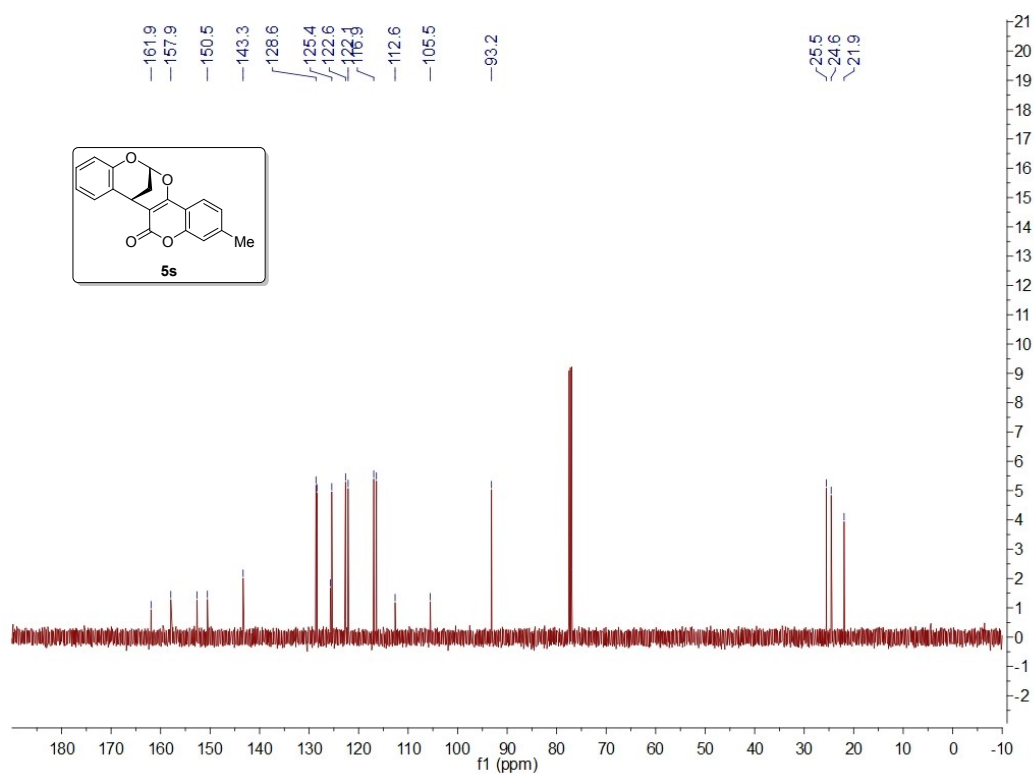
Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 15.507 | 23535838 | 90.882  | BB |
| 2   | 20.973 | 2361204  | 9.118   | BB |
|     |        | 25897042 | 100.000 |    |

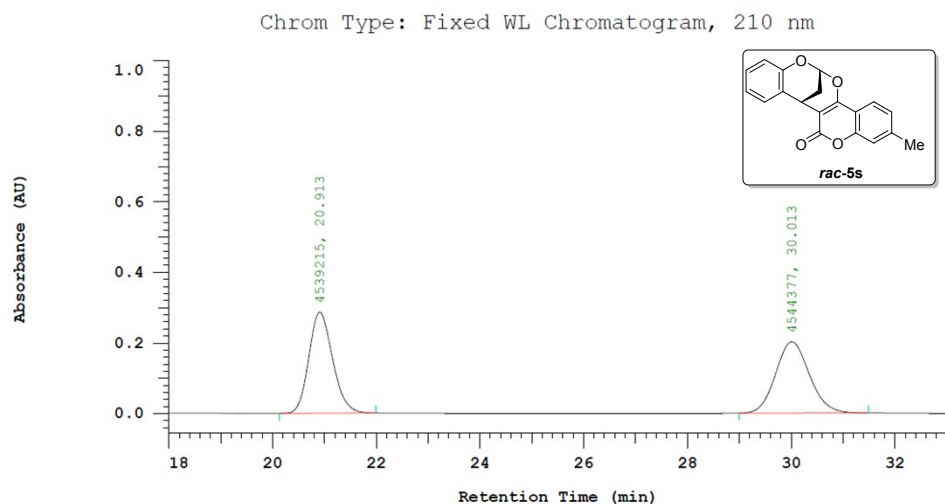
## The <sup>1</sup>H NMR spectrum of 5s (500 MHz, CDCl<sub>3</sub>)



### The <sup>13</sup>C NMR spectrum of 5s (125 MHz, CDCl<sub>3</sub>)



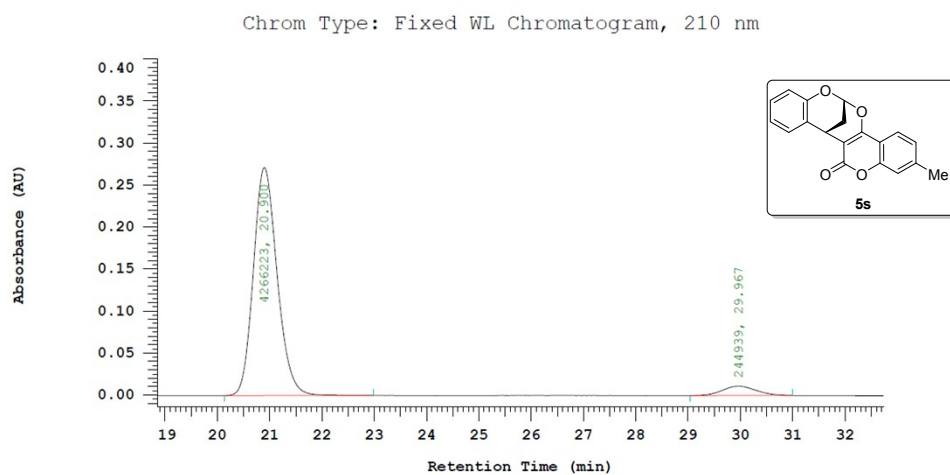
### The HPLC of racemic 5s



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 20.913 | 4539215 | 49.972  | BB |
| 2   | 30.013 | 4544377 | 50.028  | BB |
|     |        | 9083592 | 100.000 |    |

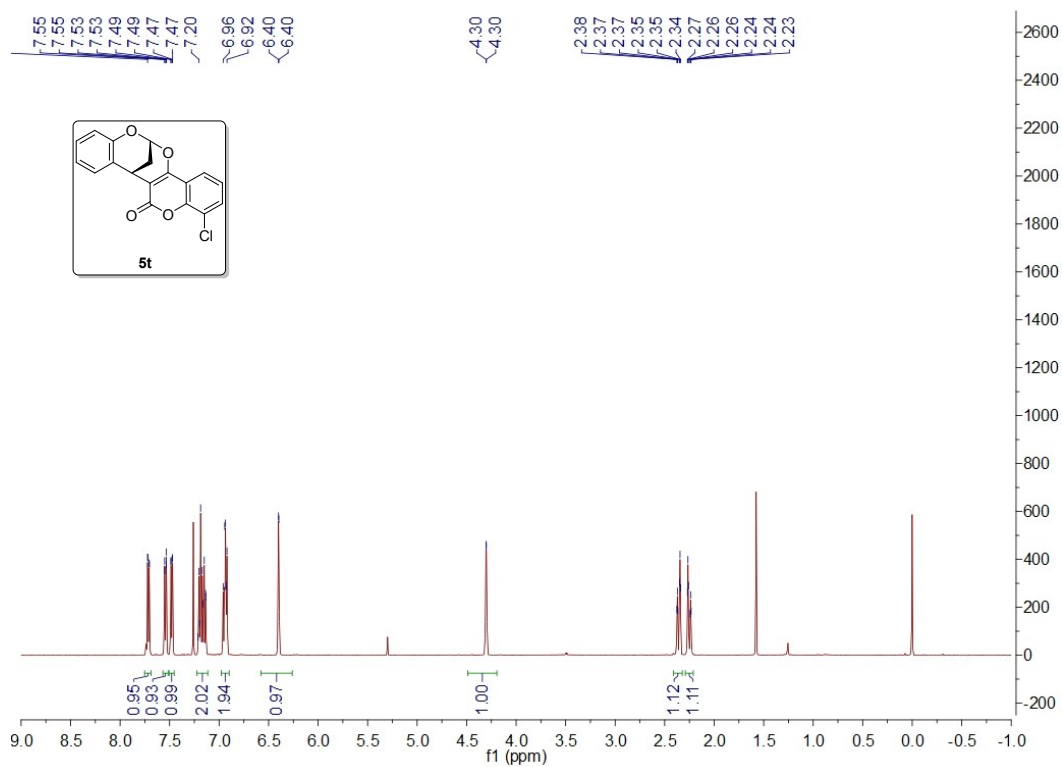
## The HPLC of chiral 5s



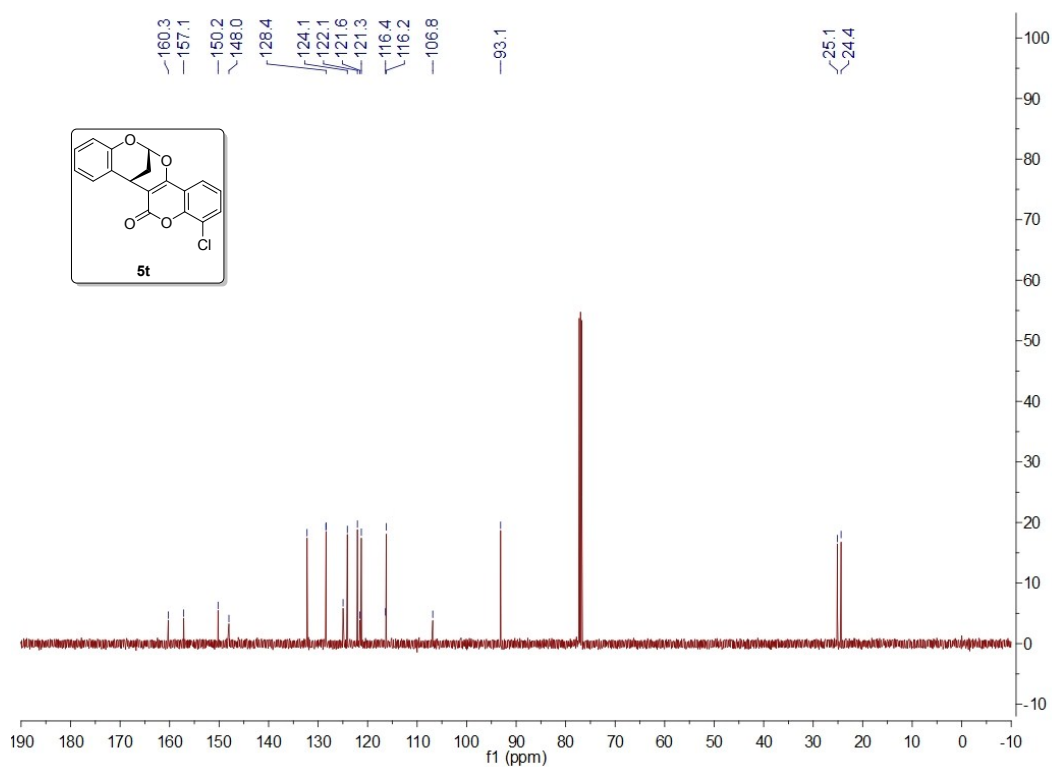
Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 20.900 | 4266223 | 94.570  | BB |
| 2   | 29.967 | 244939  | 5.430   | BB |
|     |        | 4511162 | 100.000 |    |

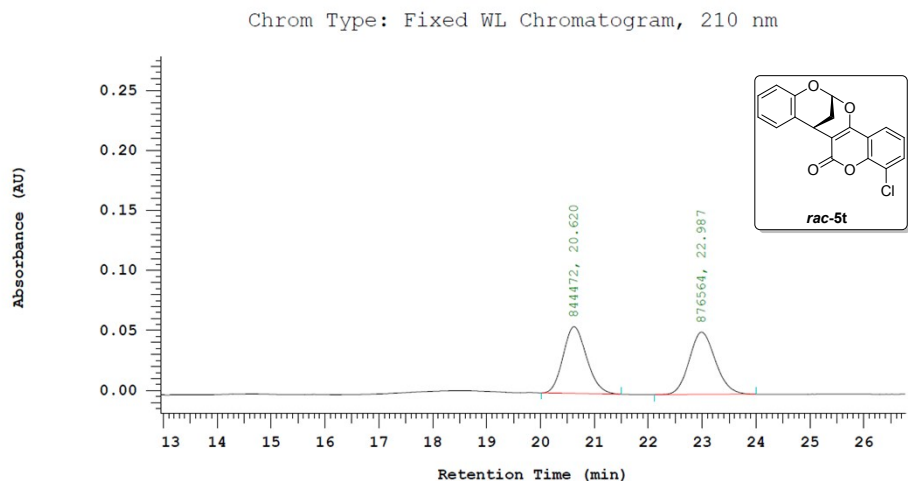
## The <sup>1</sup>H NMR spectrum of 5t (500 MHz, CDCl<sub>3</sub>)



### The <sup>13</sup>C NMR spectrum of 5t (125 MHz, CDCl<sub>3</sub>)



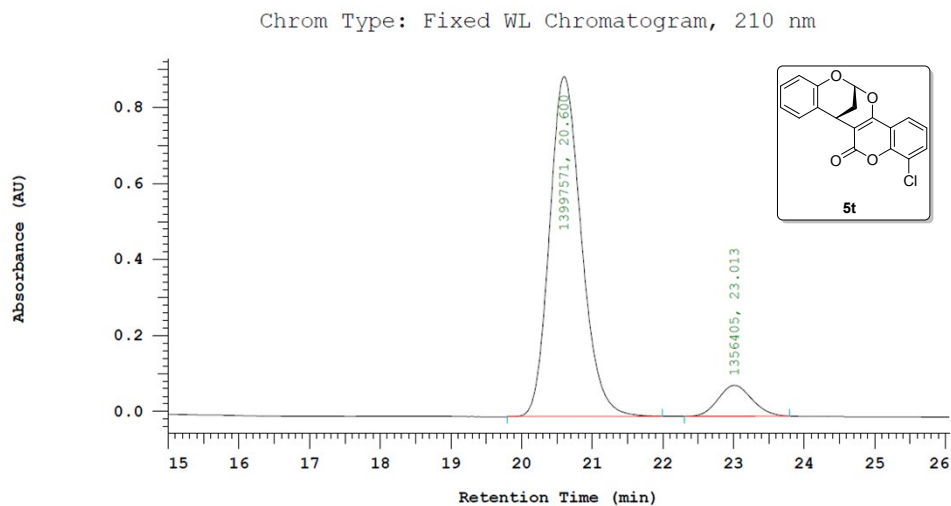
### The HPLC of racemic 5t



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 20.620 | 844472  | 49.068  | BB |
| 2   | 22.987 | 876564  | 50.932  | BB |
|     |        | 1721036 | 100.000 |    |

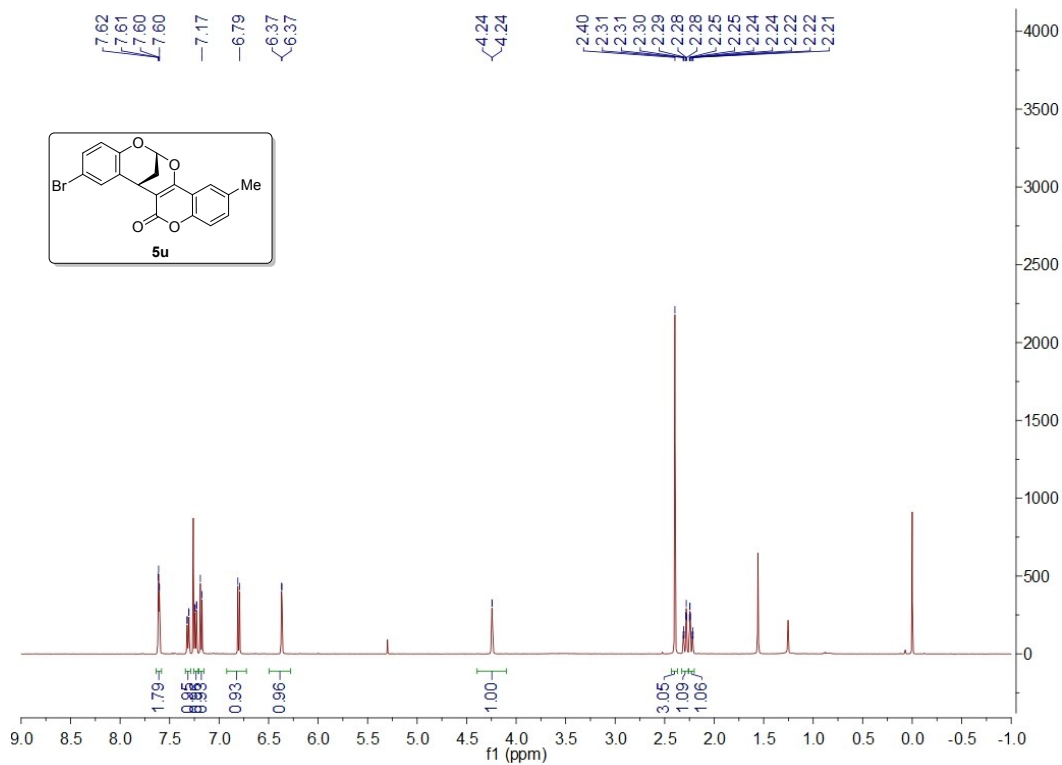
## The HPLC of chiral 5t



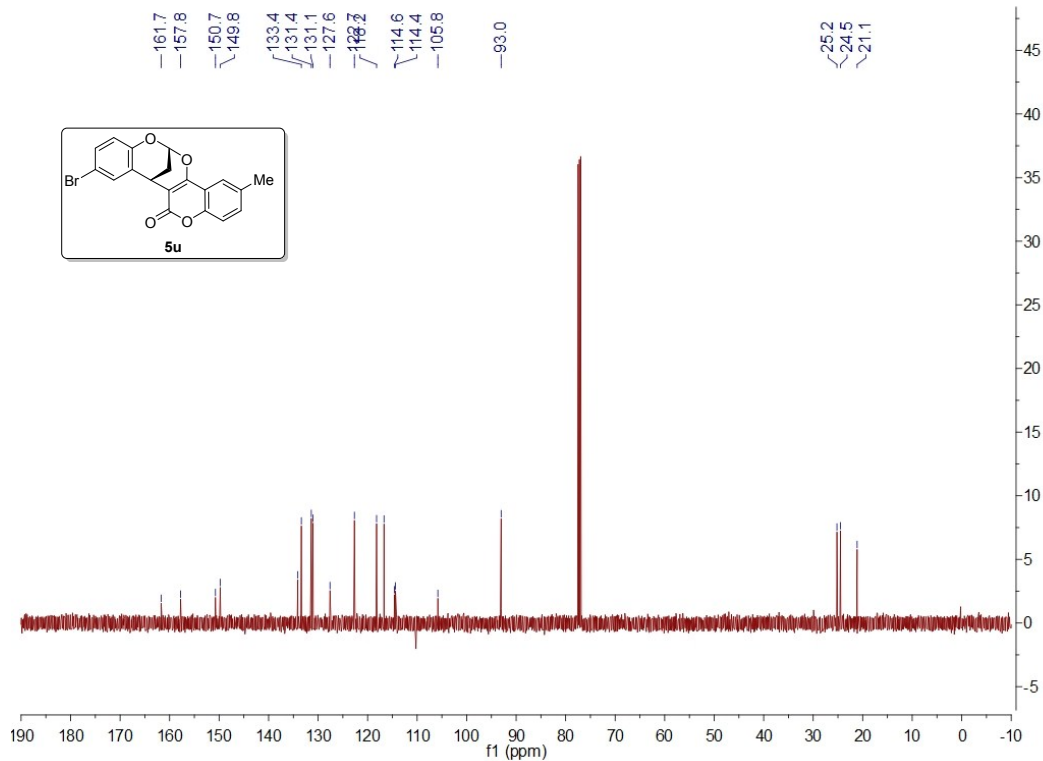
Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 20.600 | 13997571 | 91.166  | BB |
| 2   | 23.013 | 1356405  | 8.834   | BB |
|     |        | 15353976 | 100.000 |    |

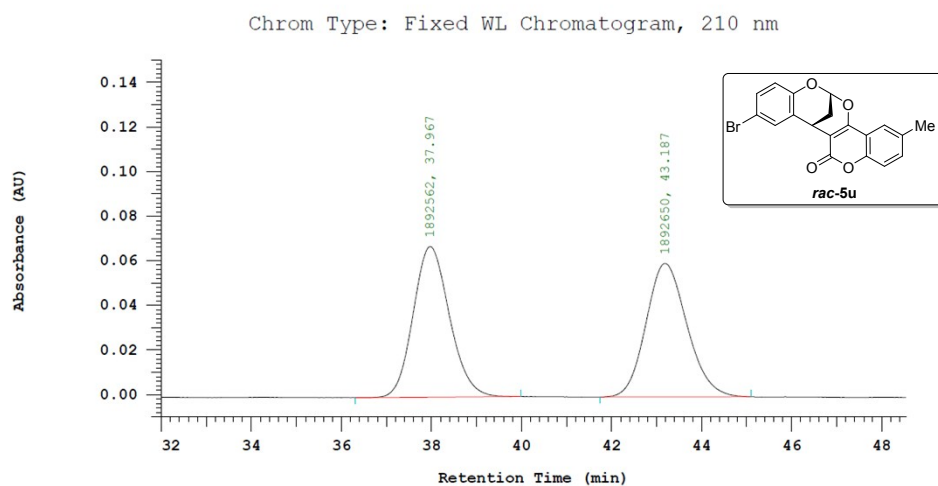
## The <sup>1</sup>H NMR spectrum of 5u (500 MHz, CDCl<sub>3</sub>)



### The <sup>13</sup>C NMR spectrum of 5u (125 MHz, CDCl<sub>3</sub>)



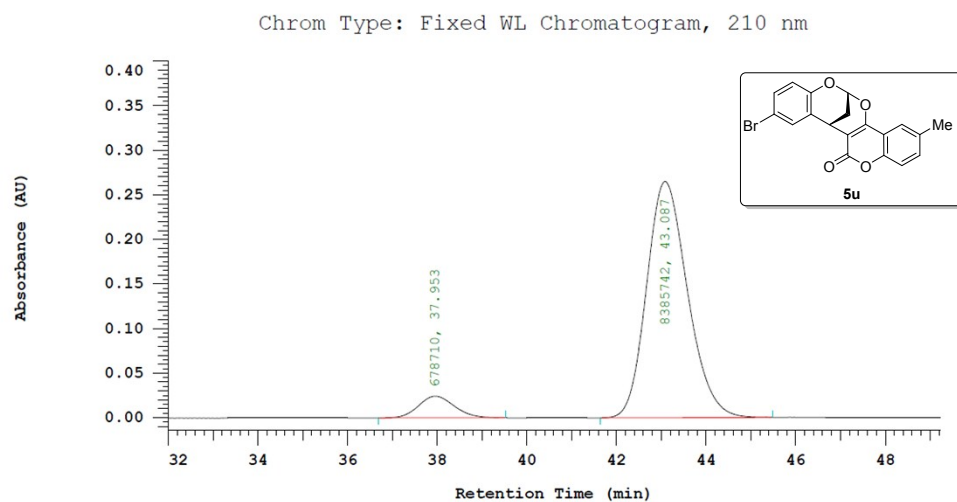
### The HPLC of racemic 5u



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 37.967 | 1892562 | 49.999  | BB |
| 2   | 43.187 | 1892650 | 50.001  | BB |
|     |        | 3785212 | 100.000 |    |

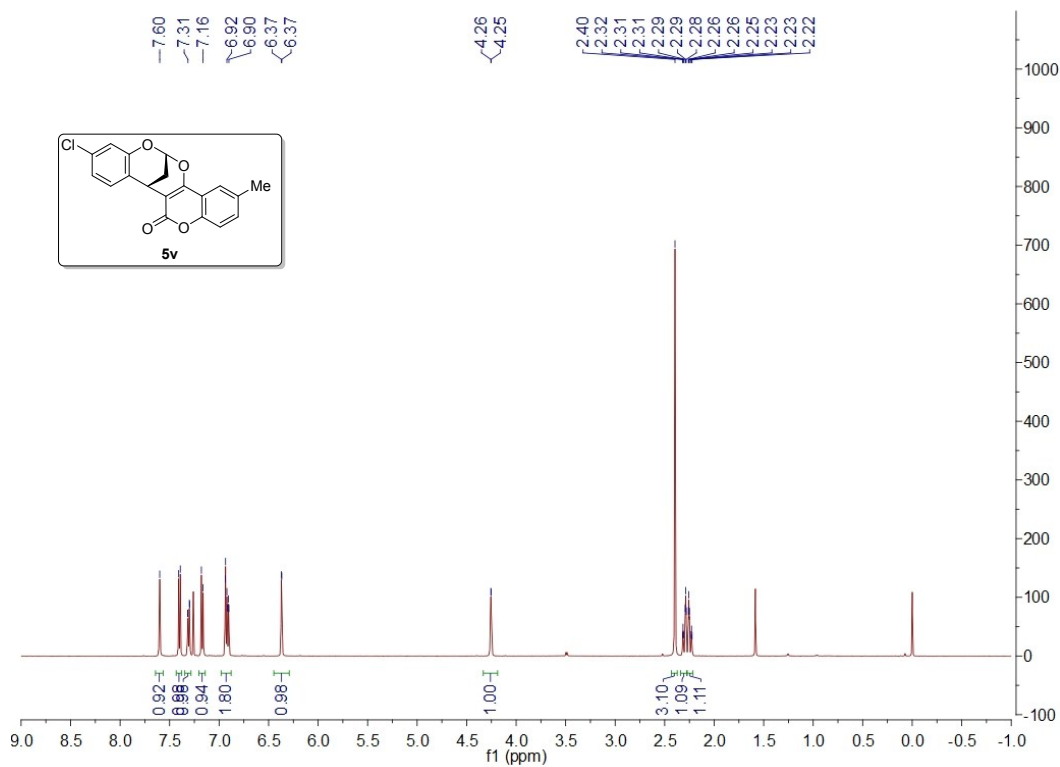
## The HPLC of chiral 5u



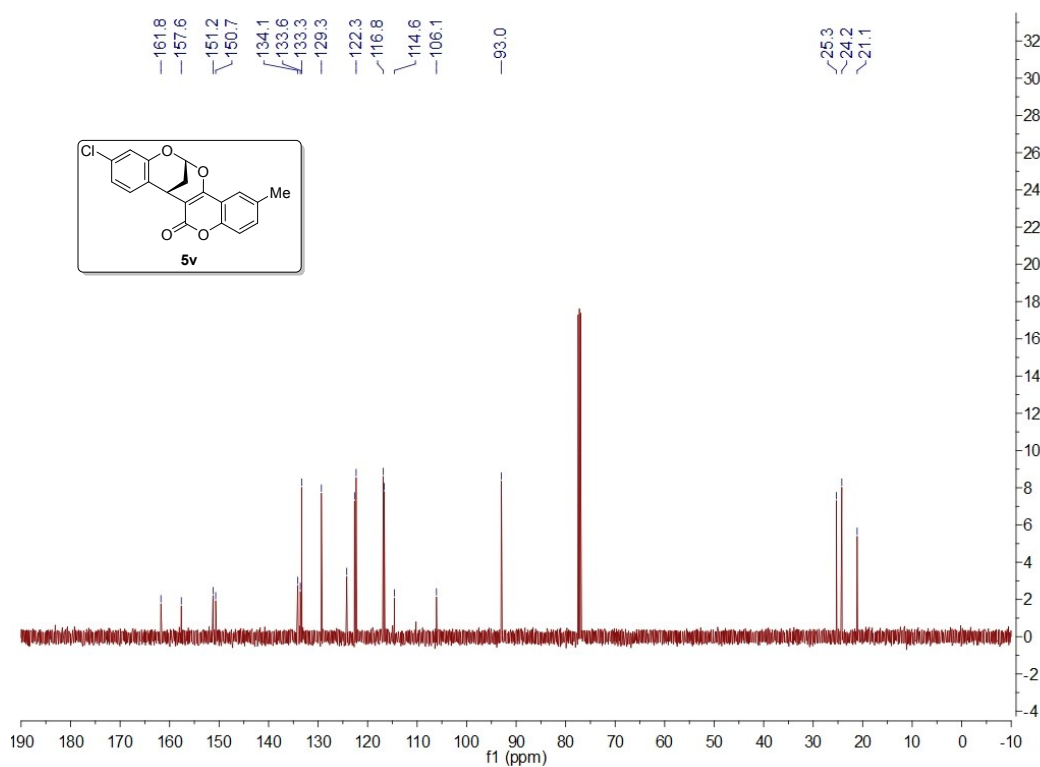
Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 37.953 | 678710  | 7.488   | BB |
| 2   | 43.087 | 8385742 | 92.512  | BB |
|     |        | 9064452 | 100.000 |    |

## The <sup>1</sup>H NMR spectrum of 5v (500 MHz, CDCl<sub>3</sub>)

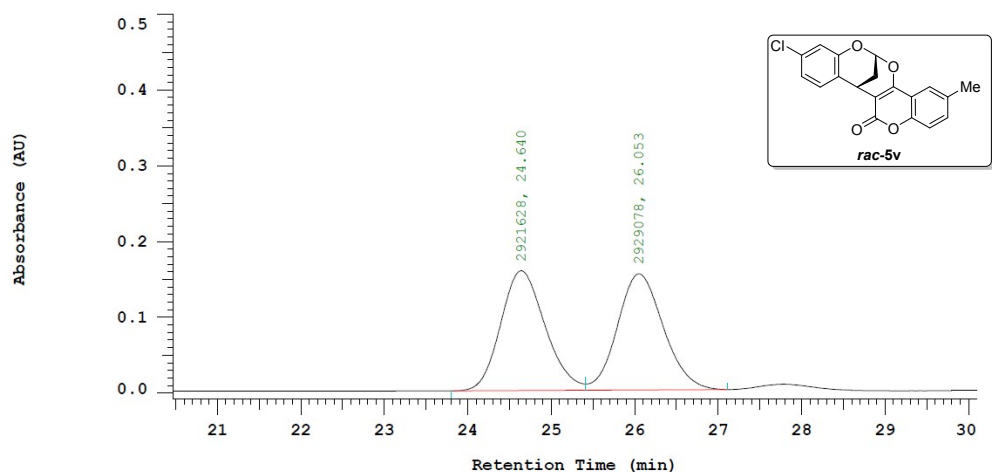


The <sup>13</sup>C NMR spectrum of 5v (125 MHz, CDCl<sub>3</sub>)



The HPLC of racemic 5v

Chrom Type: Fixed WL Chromatogram, 210 nm



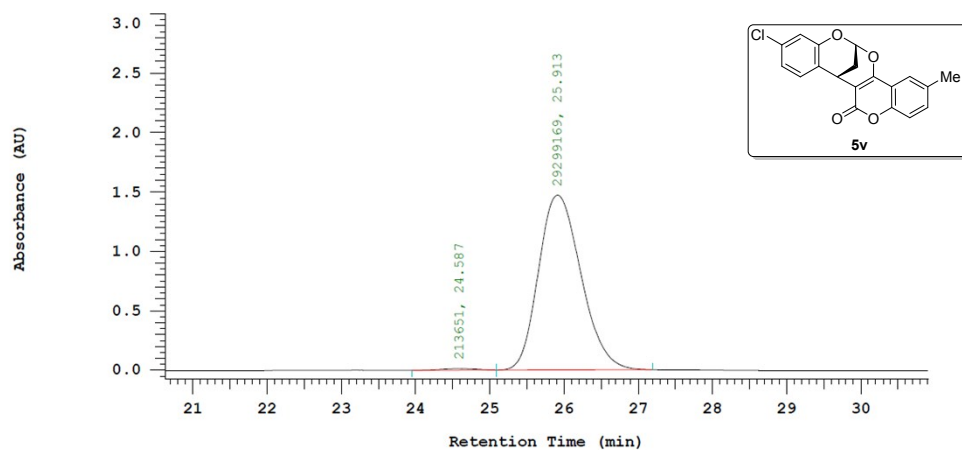
Chrom Type: Fixed WL Chromatogram, 210 nm

Peak Quantitation: AREA  
Calculation Method: AREA%

| No.     | RT     | Area    | Area %  | BC |
|---------|--------|---------|---------|----|
| 1       | 24.640 | 2921628 | 49.936  | BV |
| 2       | 26.053 | 2929078 | 50.064  | VB |
| 5850706 |        |         | 100.000 |    |

## The HPLC of chiral 5v

Chrom Type: Fixed WL Chromatogram, 210 nm

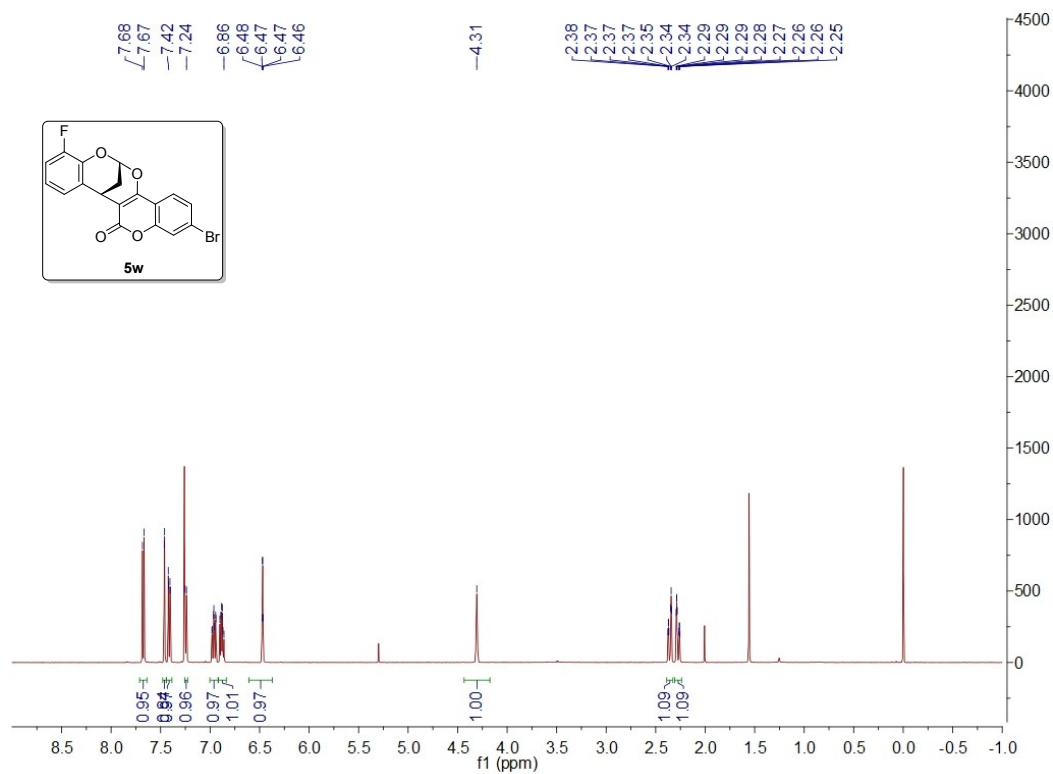


Chrom Type: Fixed WL Chromatogram, 210 nm

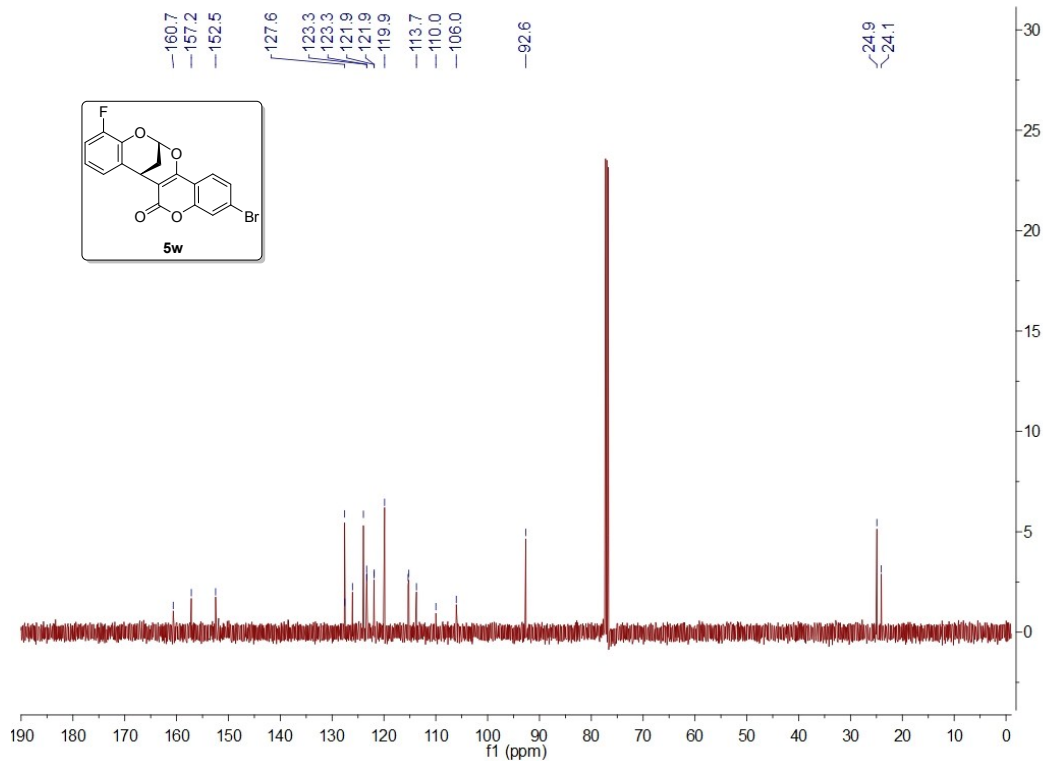
Peak Quantitation: AREA  
Calculation Method: AREA%

| No.      | RT     | Area     | Area %  | BC |
|----------|--------|----------|---------|----|
| 1        | 24.587 | 213651   | 0.724   | BV |
| 2        | 25.913 | 29299169 | 99.276  | VB |
| 29512820 |        |          | 100.000 |    |

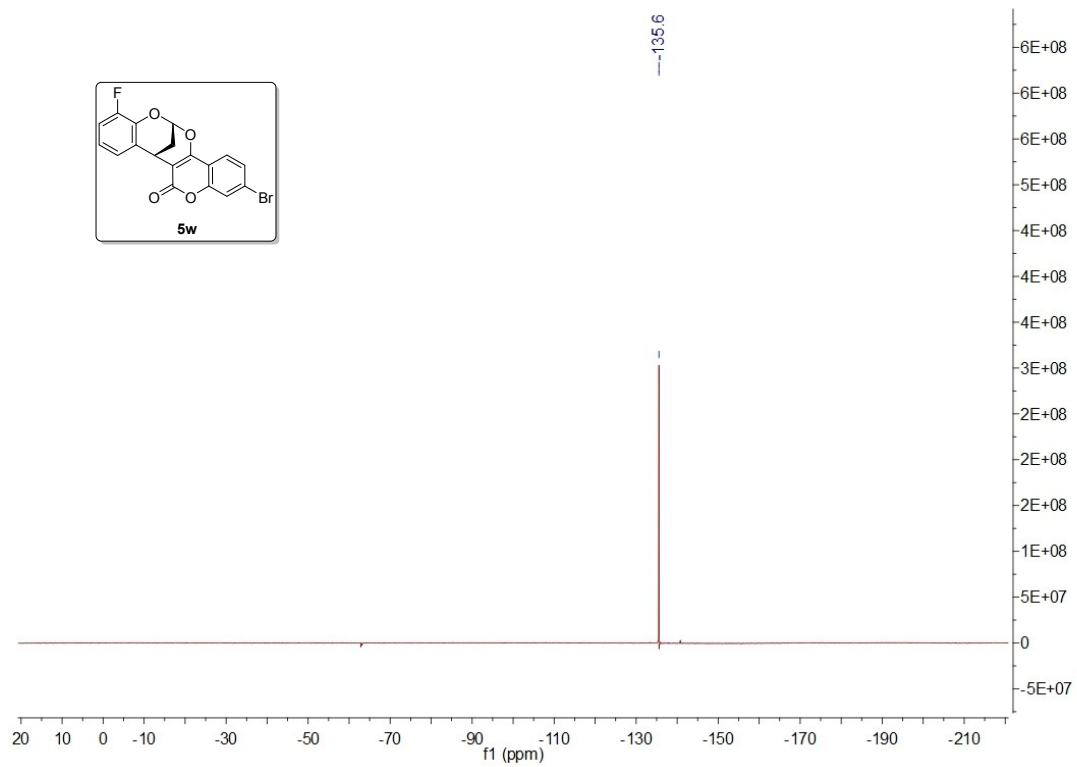
## The <sup>1</sup>H NMR spectrum of 5w (500 MHz, CDCl<sub>3</sub>)



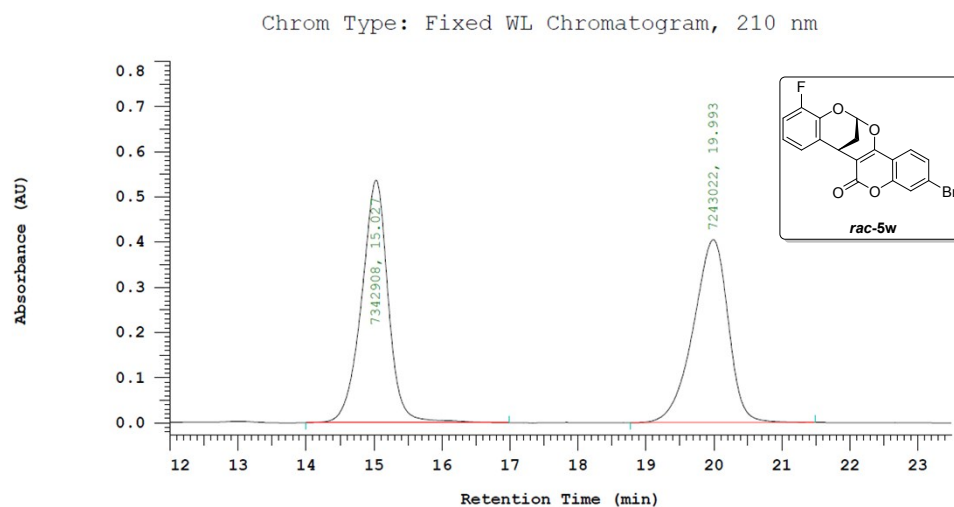
The <sup>13</sup>C NMR spectrum of 5w (125 MHz, CDCl<sub>3</sub>)



The <sup>19</sup>F NMR spectrum of 5w (400 MHz, CDCl<sub>3</sub>)



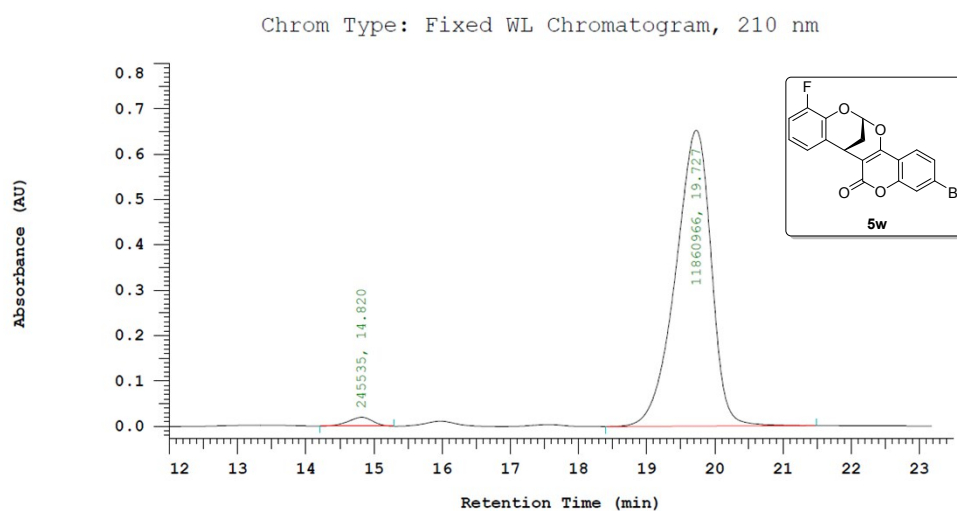
## The HPLC of racemic 5w



Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 15.027 | 7342908  | 50.342  | BB |
| 2   | 19.993 | 7243022  | 49.658  | BB |
|     |        | 14585930 | 100.000 |    |

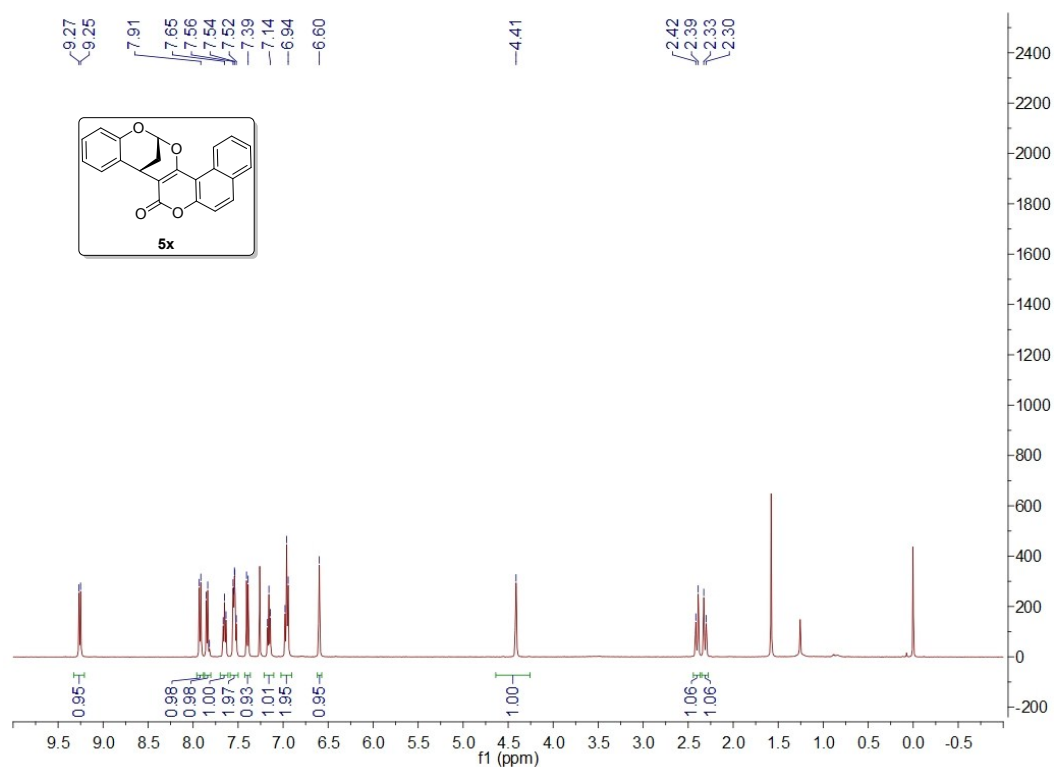
## The HPLC of chiral 5w



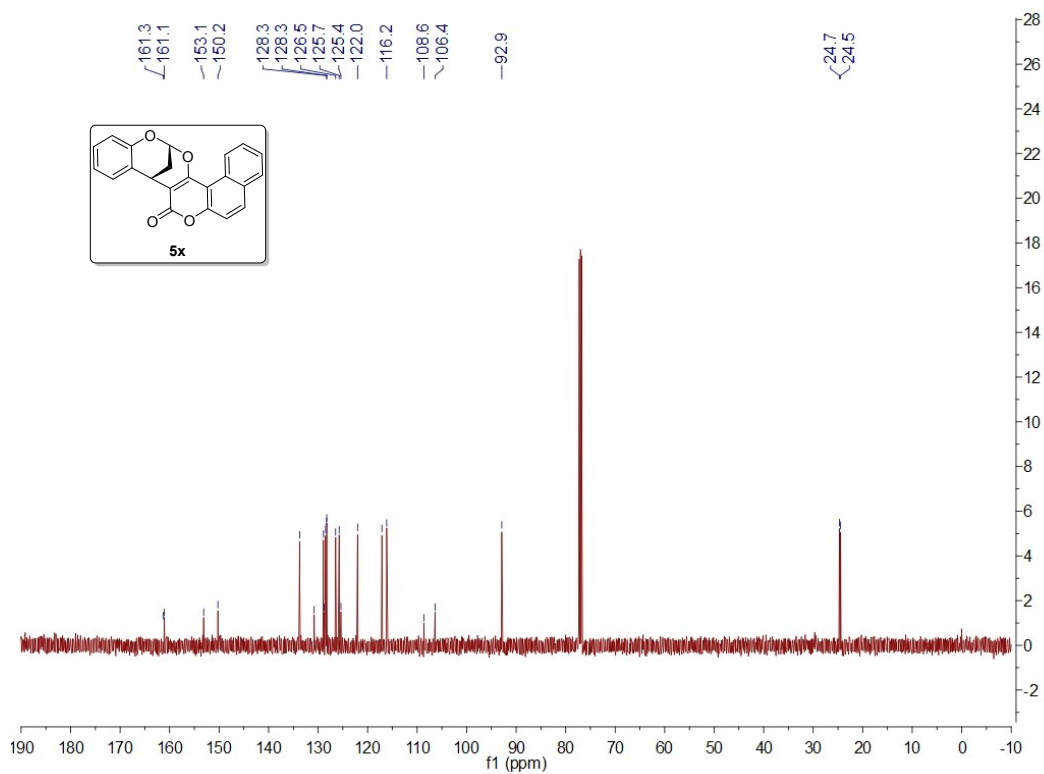
Chrom Type: Fixed WL Chromatogram, 210 nm  
Peak Quantitation: AREA  
Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 14.820 | 245535   | 2.028   | BB |
| 2   | 19.727 | 11860966 | 97.972  | BB |
|     |        | 12106501 | 100.000 |    |

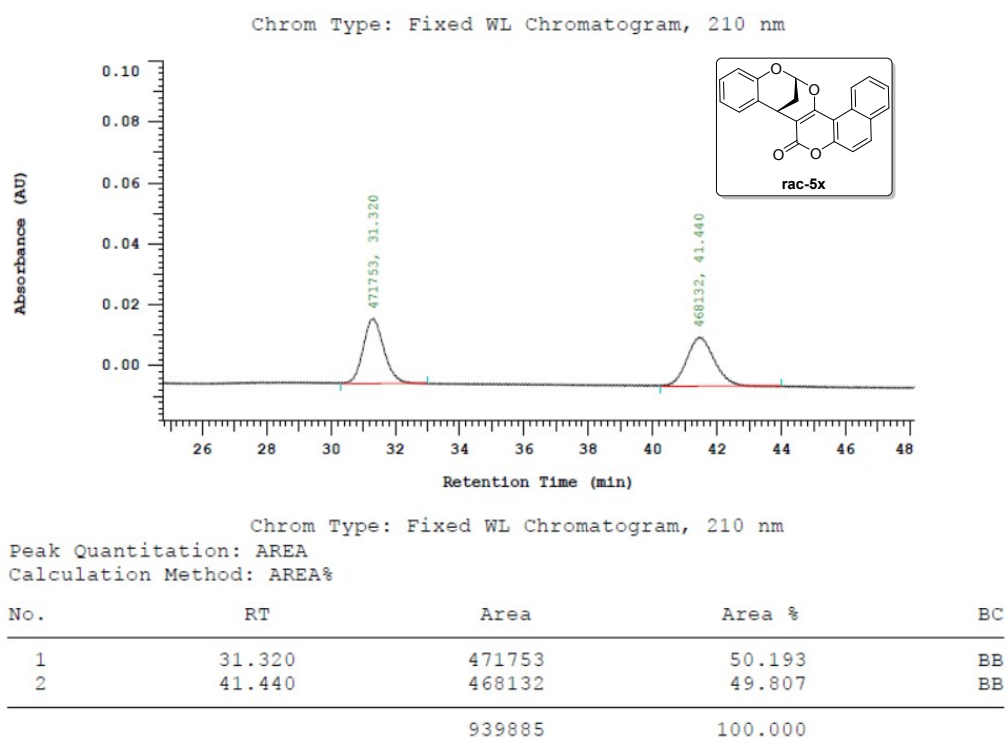
**The  $^1\text{H}$  NMR spectrum of 5x (500 MHz,  $\text{CDCl}_3$ )**



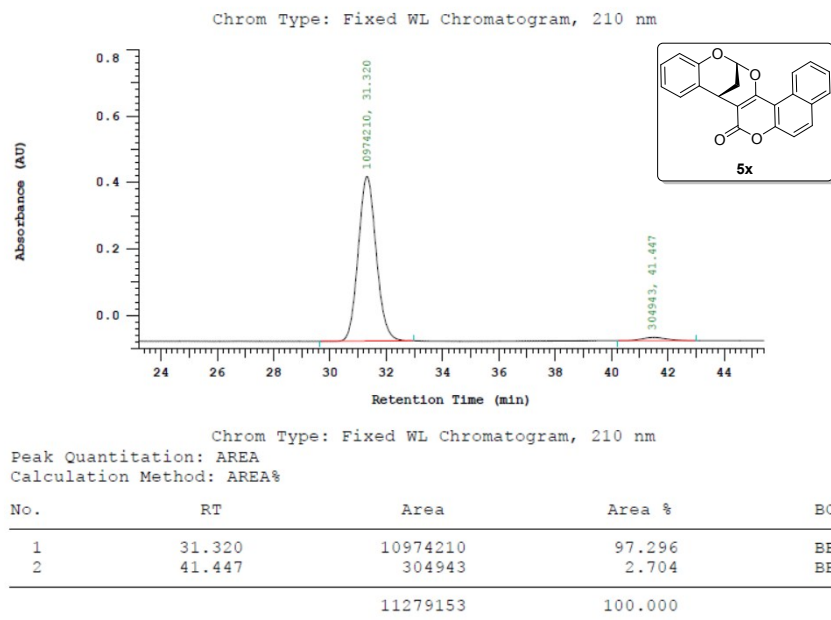
**The  $^{13}\text{C}$  NMR spectrum of 5x (125 MHz,  $\text{CDCl}_3$ )**



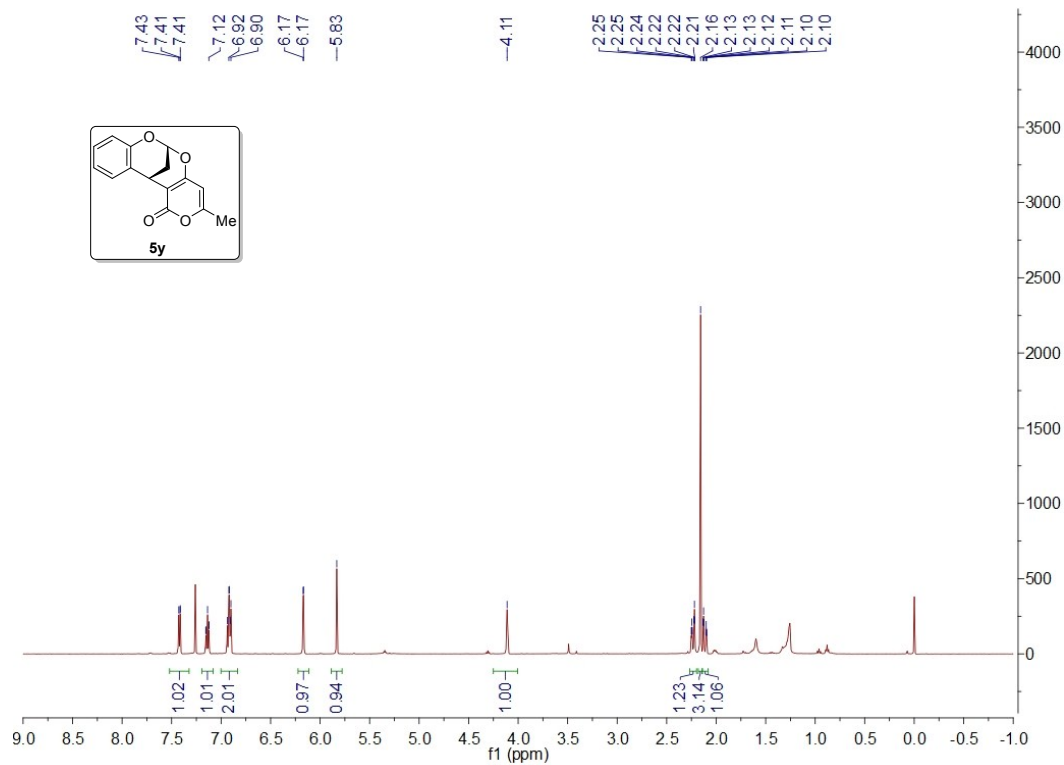
## The HPLC of racemic 5x



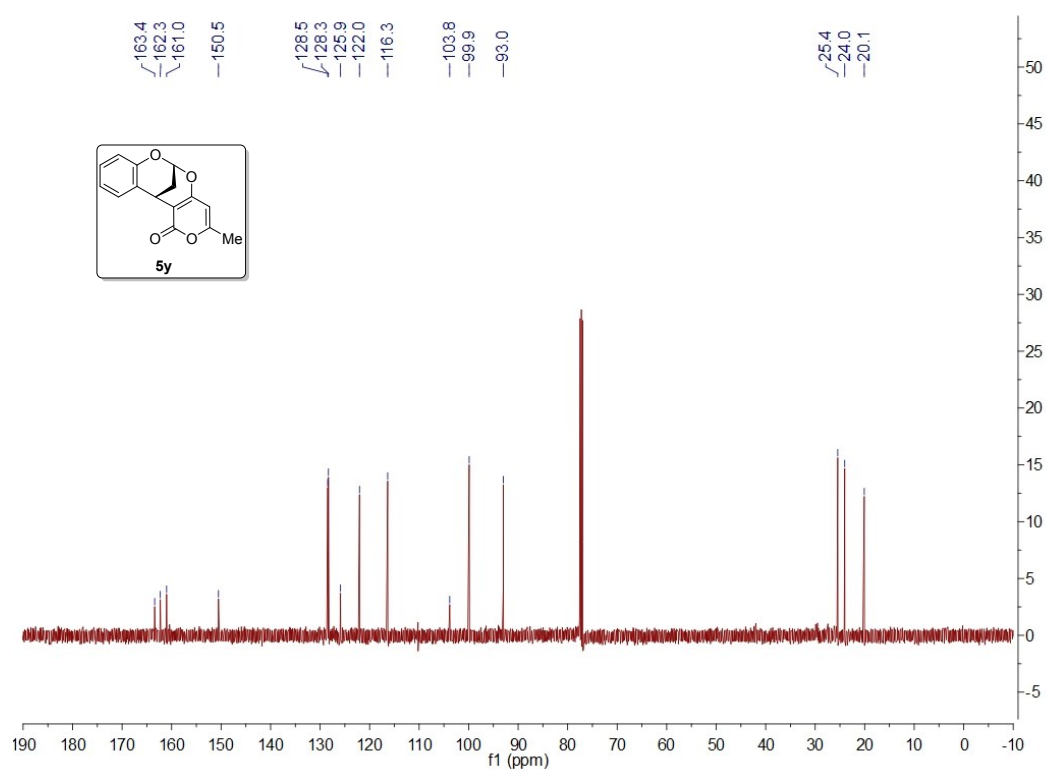
## The HPLC of chiral 5x



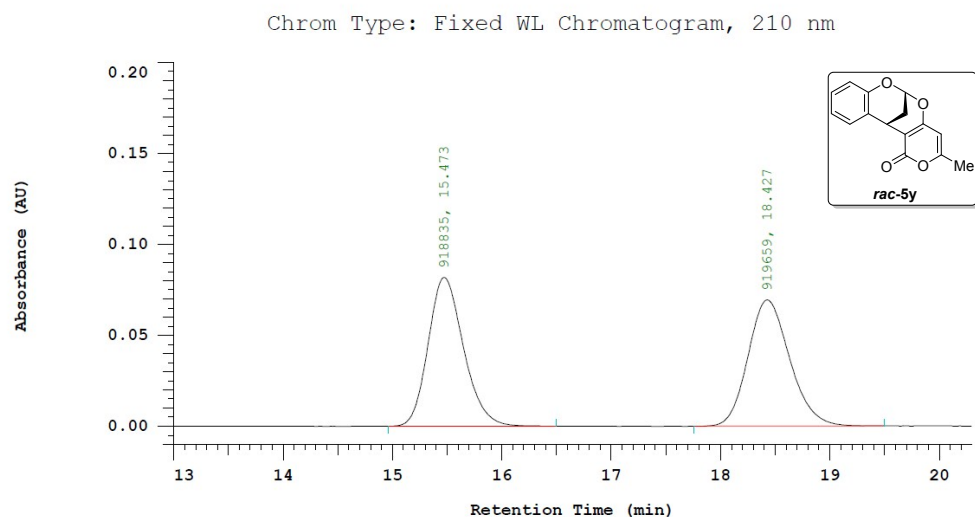
## The <sup>1</sup>H NMR spectrum of 5y (500 MHz, CDCl<sub>3</sub>)



The <sup>13</sup>C NMR spectrum of 5y (125 MHz, CDCl<sub>3</sub>)



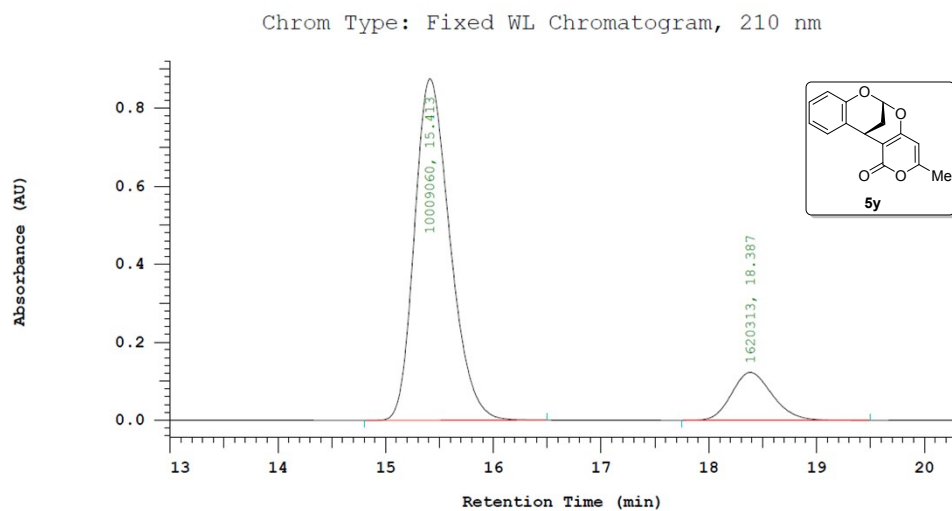
The HPLC of racemic 5y



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 15.473 | 918835  | 49.978  | BB |
| 2   | 18.427 | 919659  | 50.022  | BB |
|     |        | 1838494 | 100.000 |    |

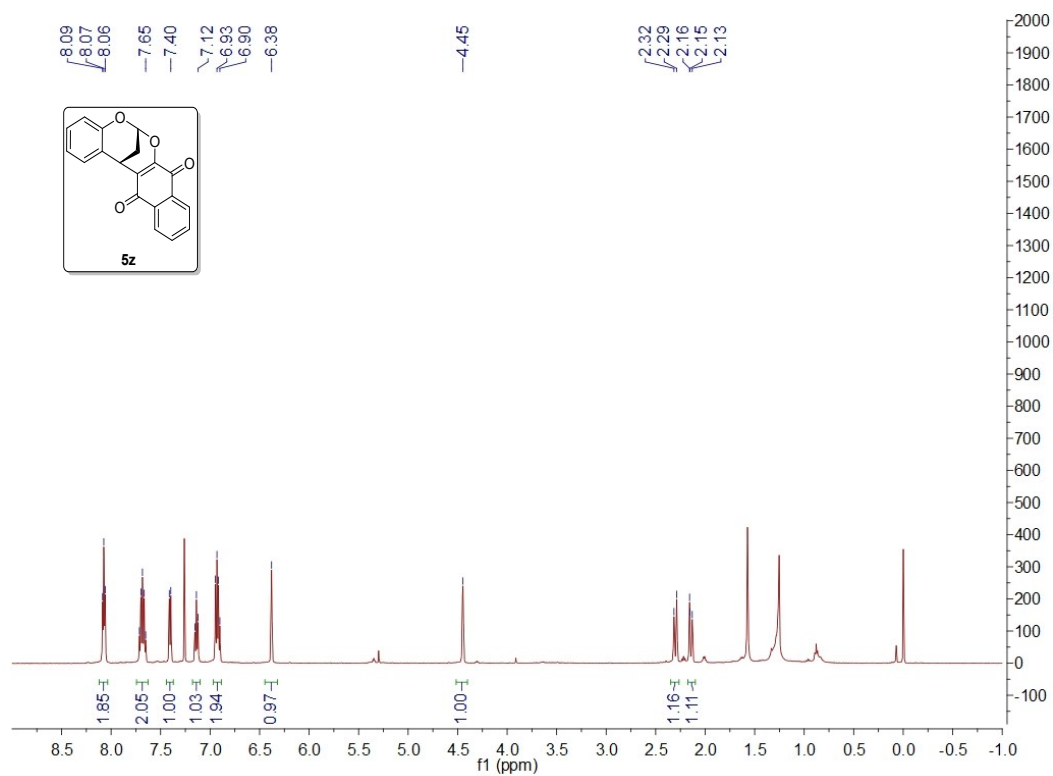
## The HPLC of chiral 5y



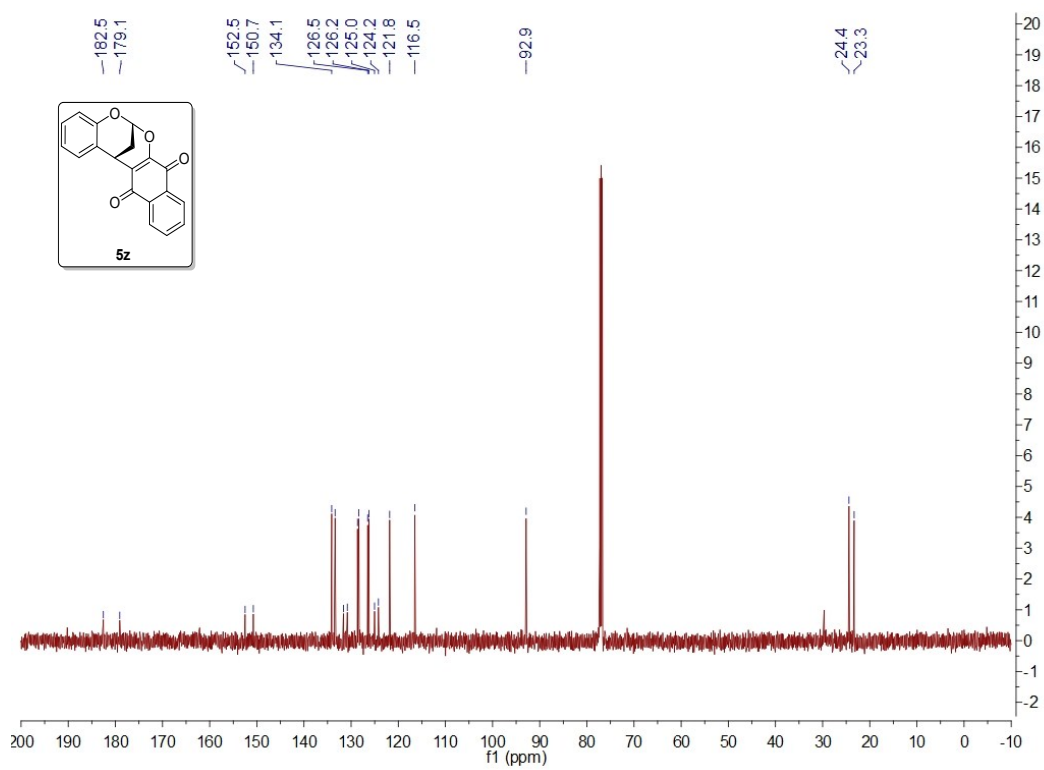
Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 15.413 | 10009060 | 86.067  | BB |
| 2   | 18.387 | 1620313  | 13.933  | BB |
|     |        | 11629373 | 100.000 |    |

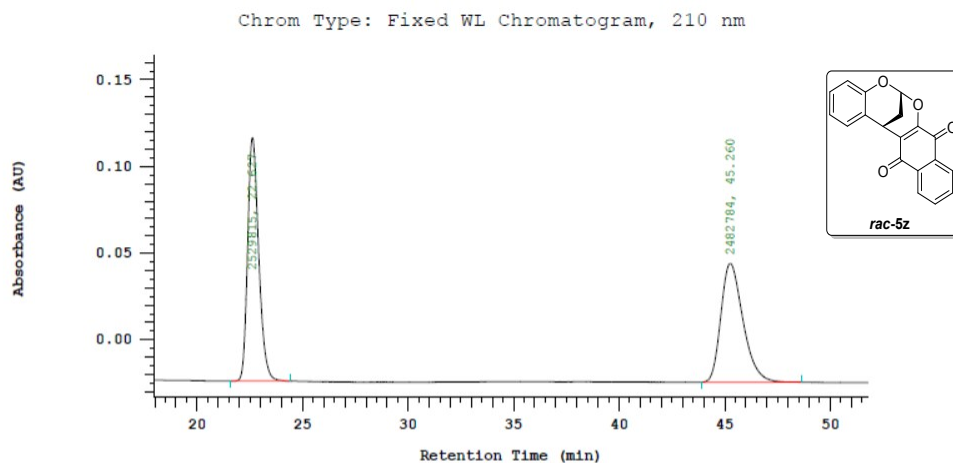
## The <sup>1</sup>H NMR spectrum of 5z (500 MHz, CDCl<sub>3</sub>)



The <sup>13</sup>C NMR spectrum of 5z (125 MHz, CDCl<sub>3</sub>)



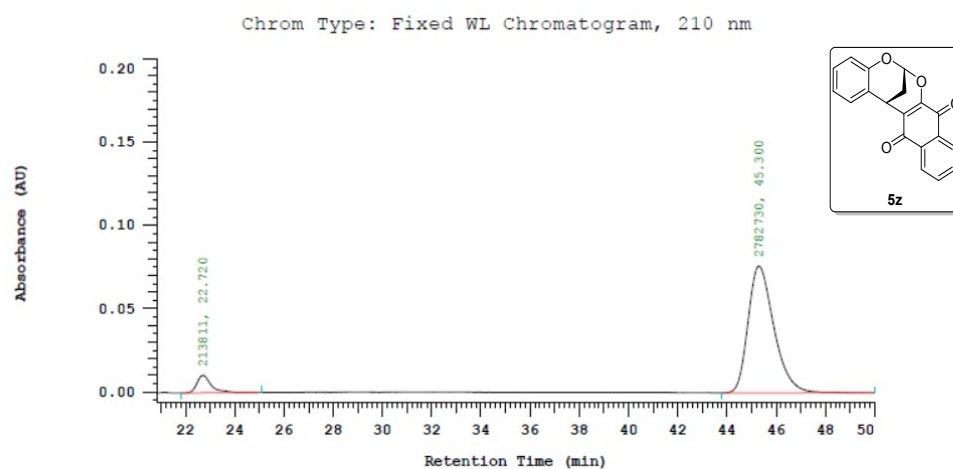
The HPLC of racemic 5z



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 22.627 | 2529815 | 50.469  | BB |
| 2   | 45.260 | 2482784 | 49.531  | BB |
|     |        | 5012599 | 100.000 |    |

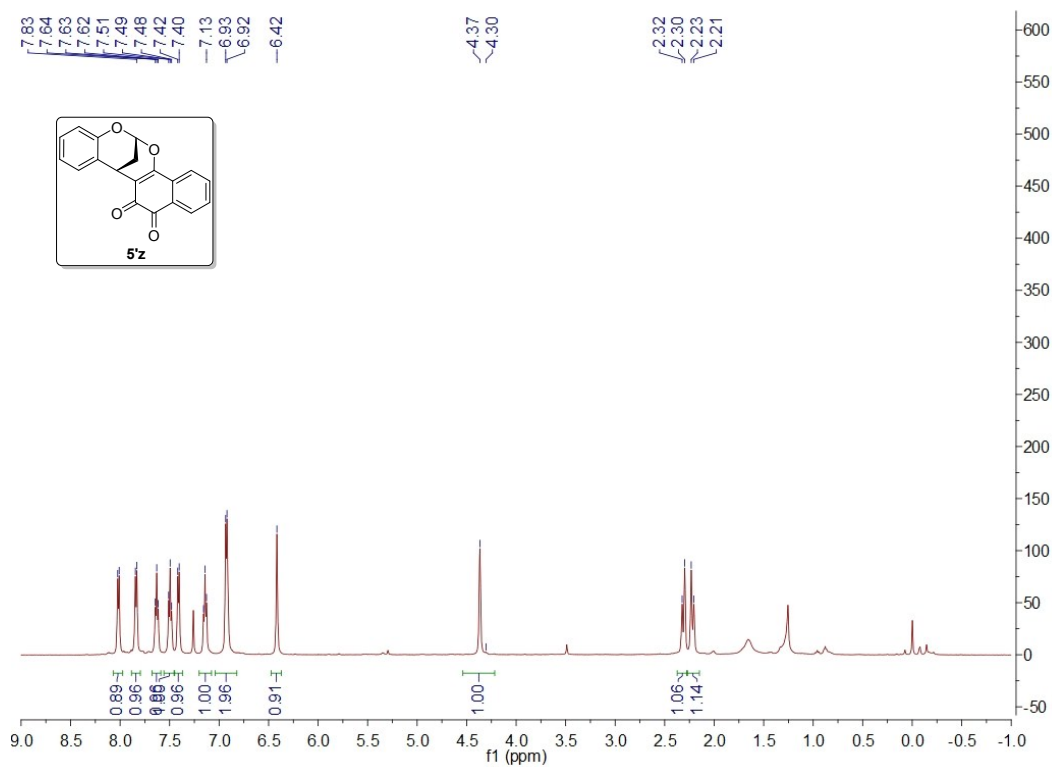
## The HPLC of chiral 5z



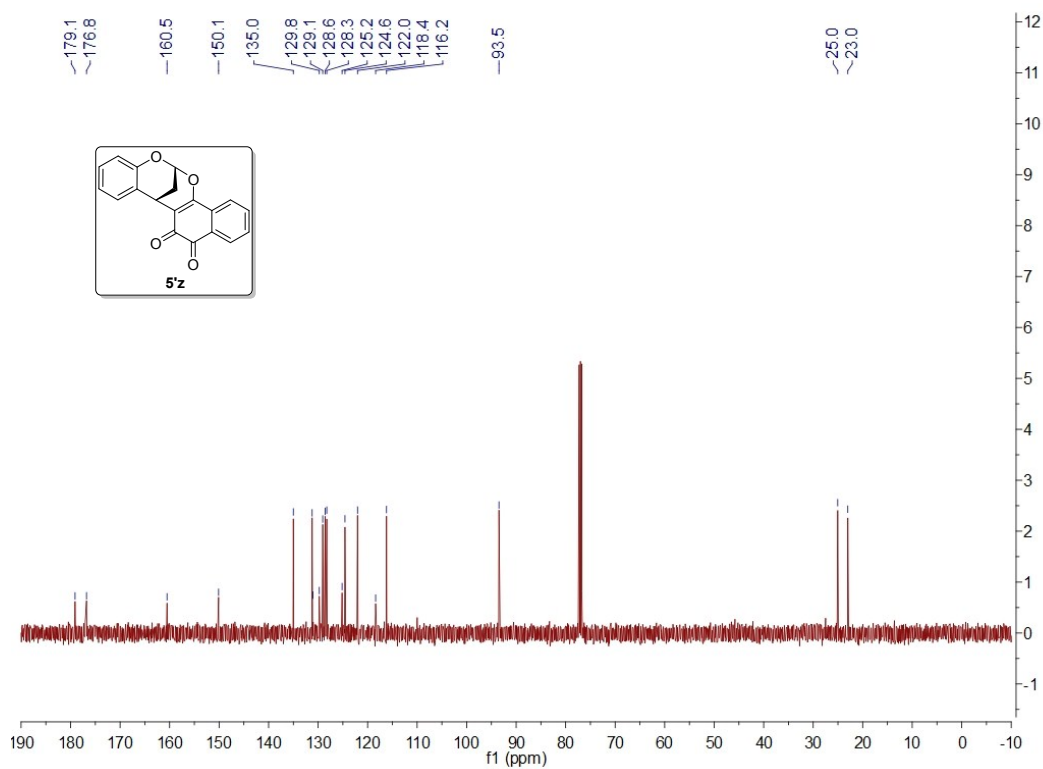
Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 22.720 | 213811  | 7.135   | BB |
| 2   | 45.300 | 2782730 | 92.865  | BB |
|     |        | 2996541 | 100.000 |    |

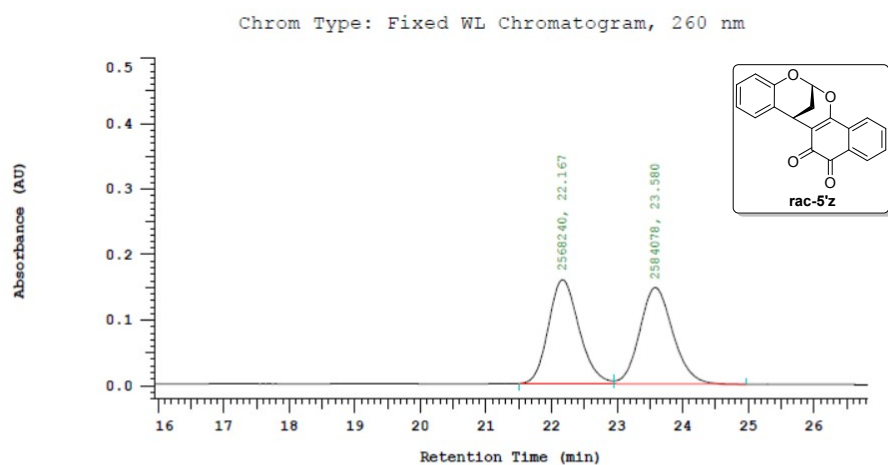
### The $^1\text{H}$ NMR spectrum of 5'z (500 MHz, $\text{CDCl}_3$ )



### The $^{13}\text{C}$ NMR spectrum of 5'z (125 MHz, $\text{CDCl}_3$ )



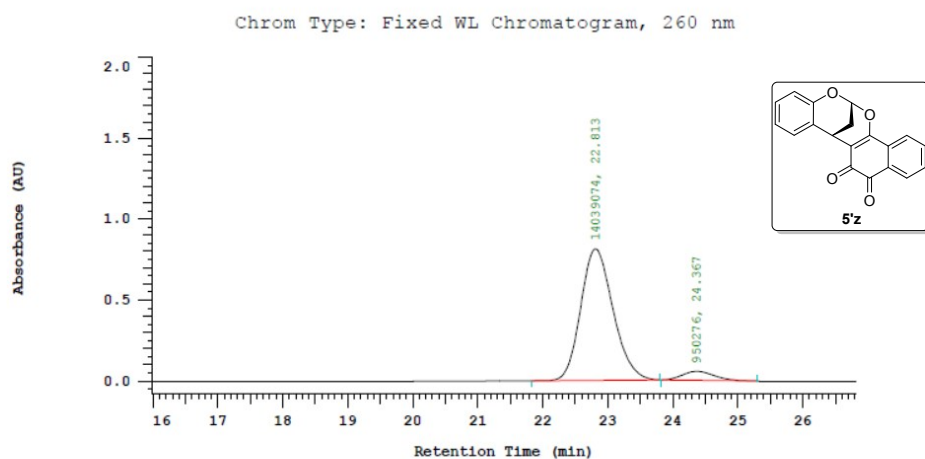
## The HPLC of racemic 5'z



Chrom Type: Fixed WL Chromatogram, 260 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 22.167 | 2568240 | 49.846  | BV |
| 2   | 23.580 | 2584078 | 50.154  | VB |
|     |        | 5152318 | 100.000 |    |

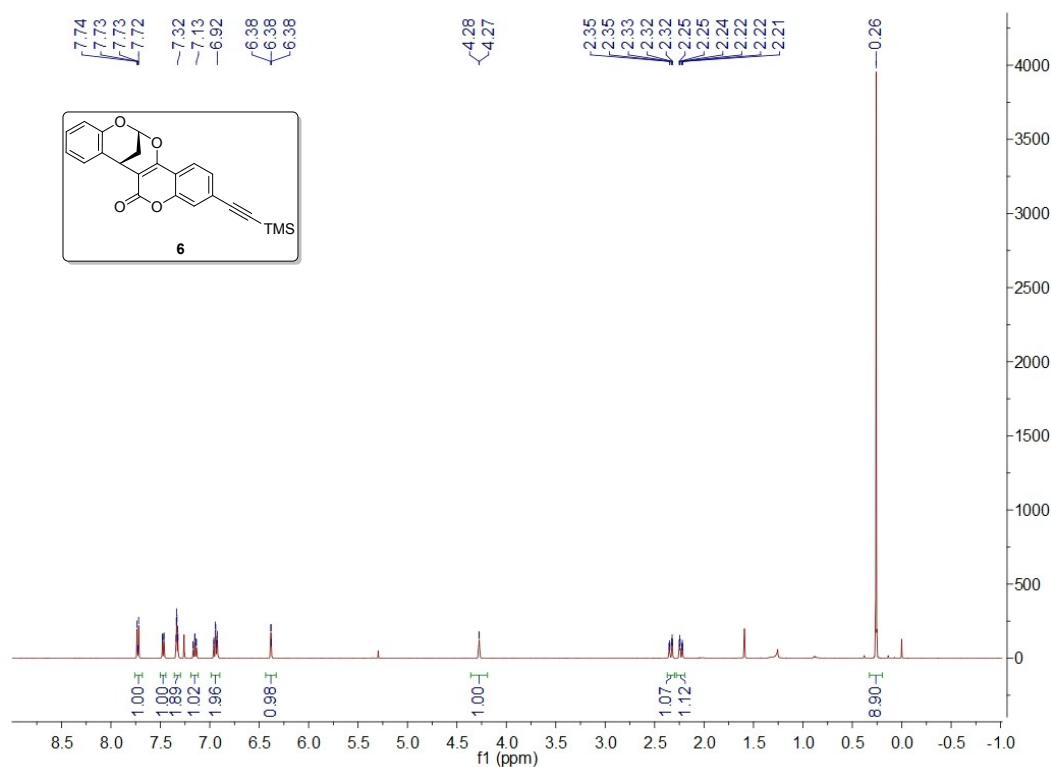
## The HPLC of chiral 5'z



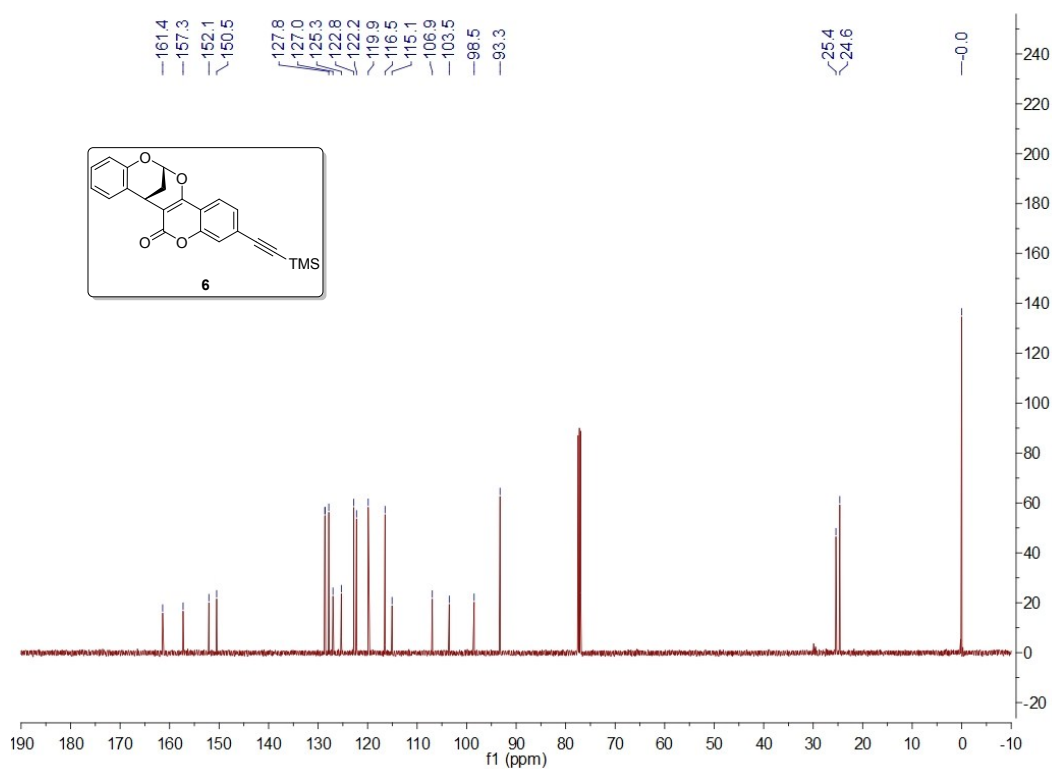
Chrom Type: Fixed WL Chromatogram, 260 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 22.813 | 14039074 | 93.660  | BB |
| 2   | 24.367 | 950276   | 6.340   | BB |
|     |        | 14989350 | 100.000 |    |

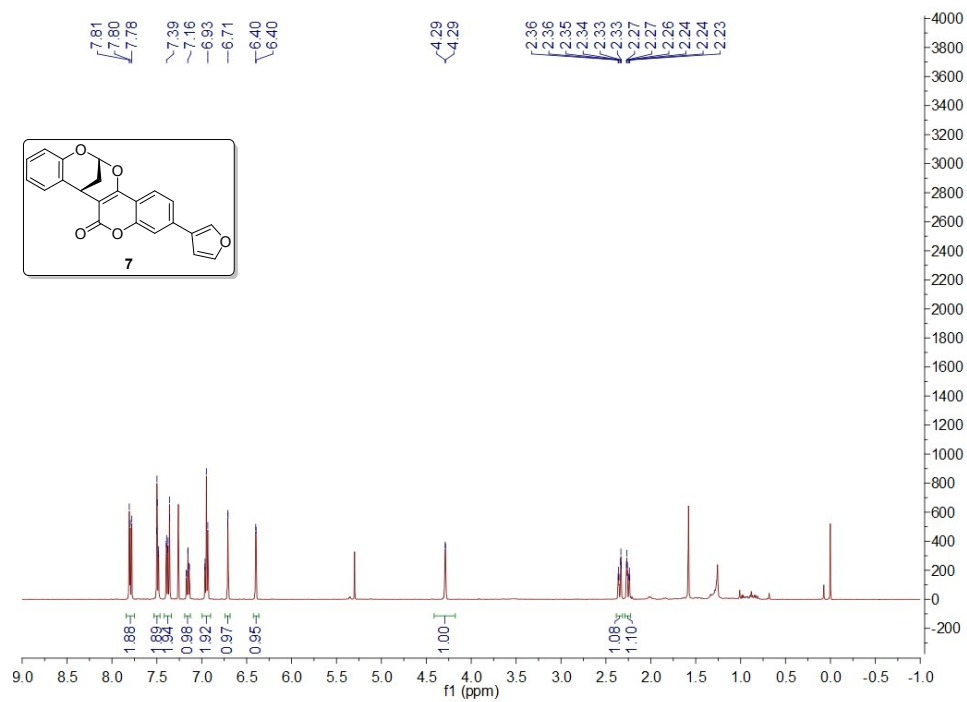
**The  $^1\text{H}$  NMR spectrum of 6 (500 MHz,  $\text{CDCl}_3$ )**



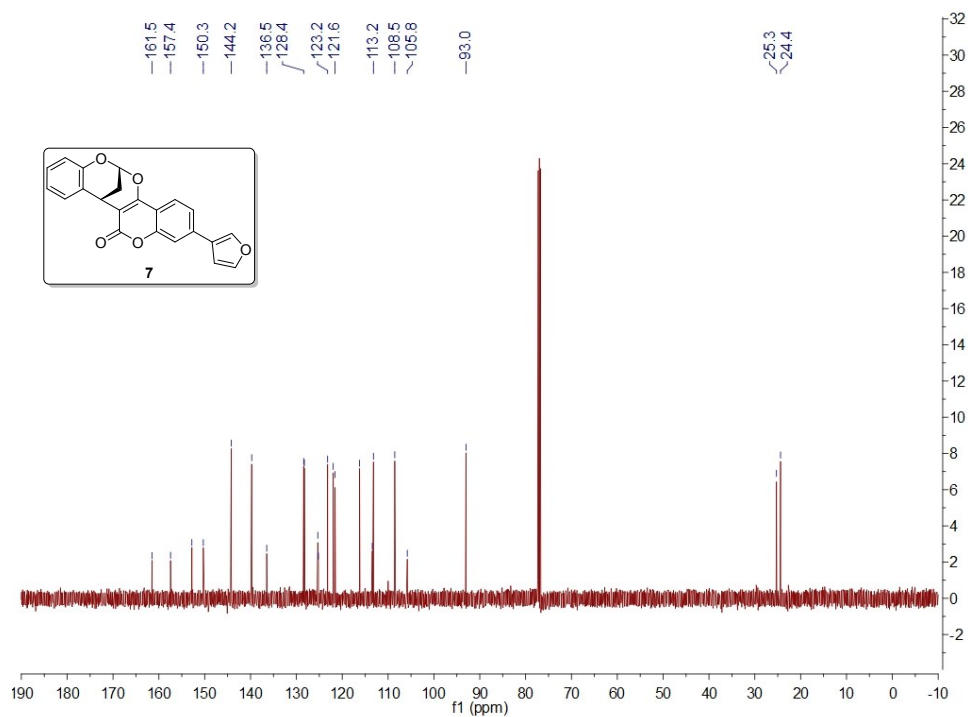
**The  $^{13}\text{C}$  NMR spectrum of 6 (125 MHz,  $\text{CDCl}_3$ )**



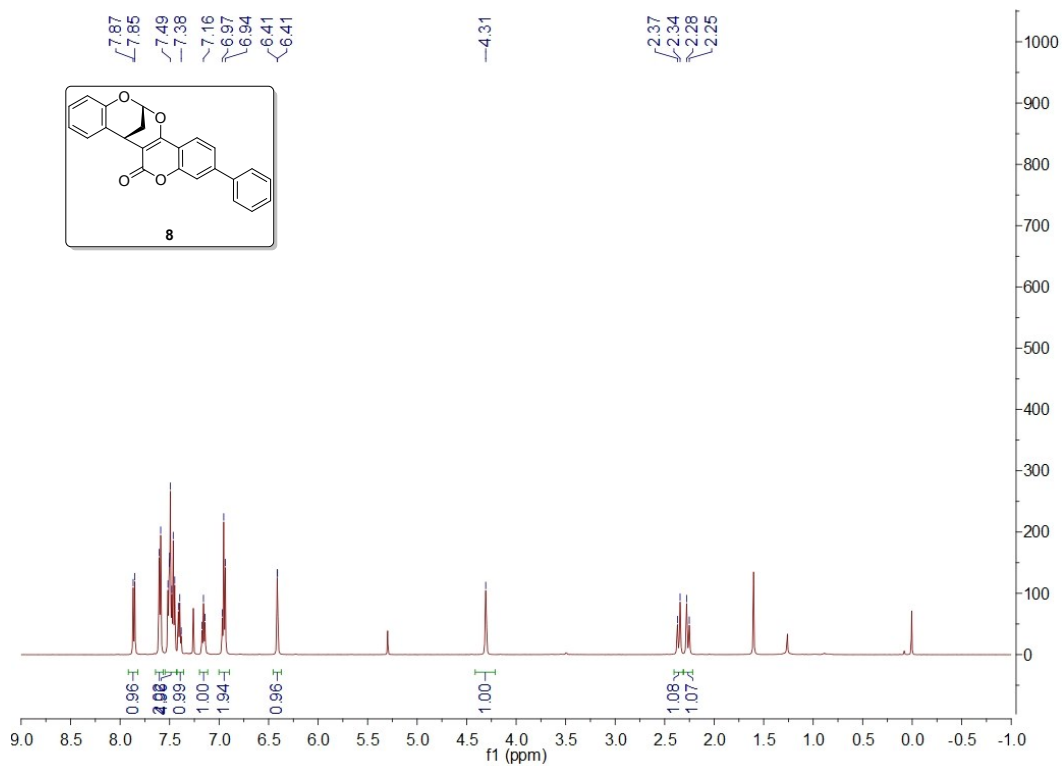
**The  $^1\text{H}$  NMR spectrum of 7 (500 MHz,  $\text{CDCl}_3$ )**



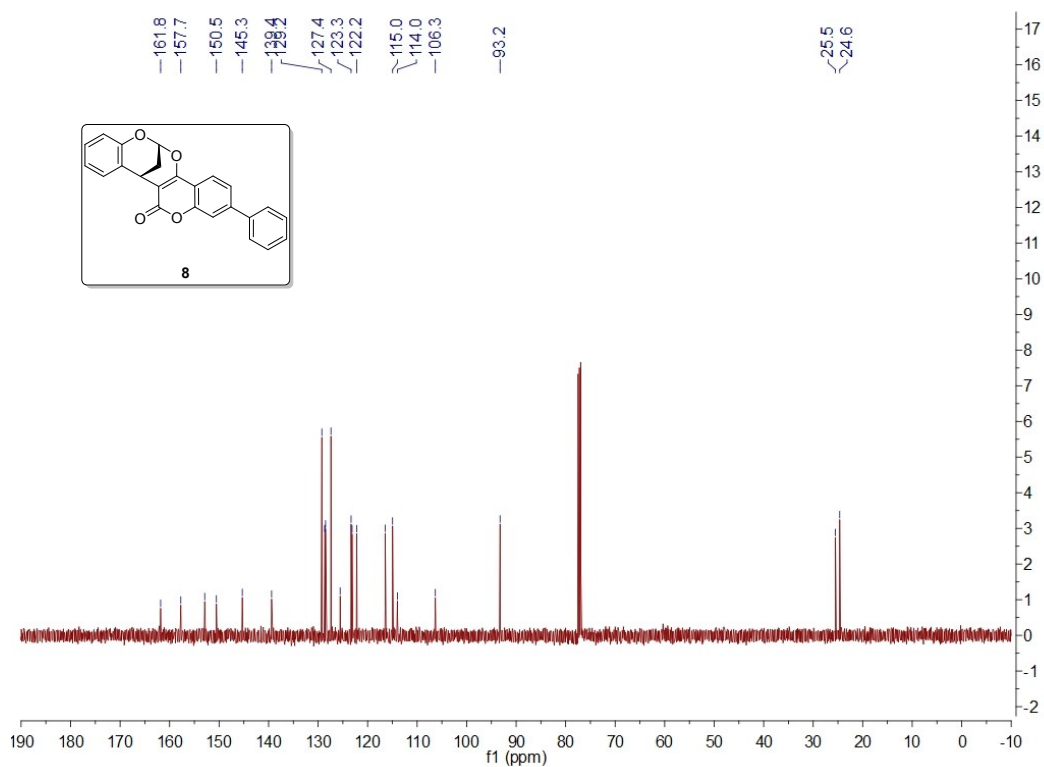
The <sup>13</sup>C NMR spectrum of 7 (125 MHz, CDCl<sub>3</sub>)



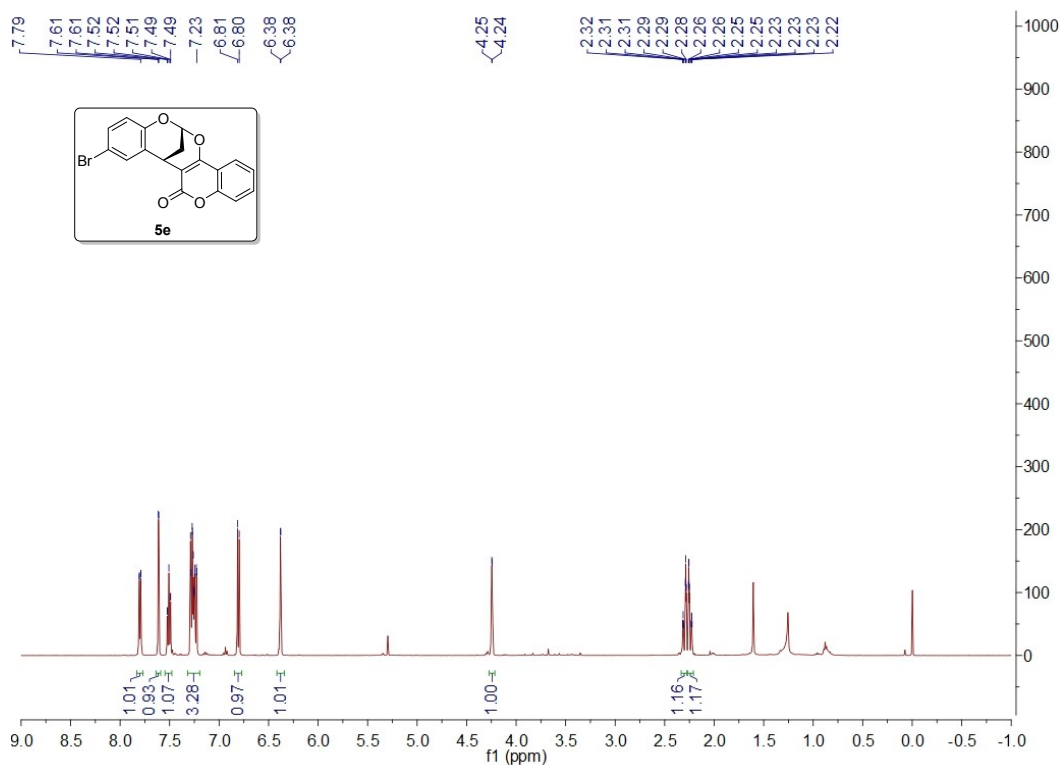
The  $^1\text{H}$  NMR spectrum of **8** (500 MHz,  $\text{CDCl}_3$ )



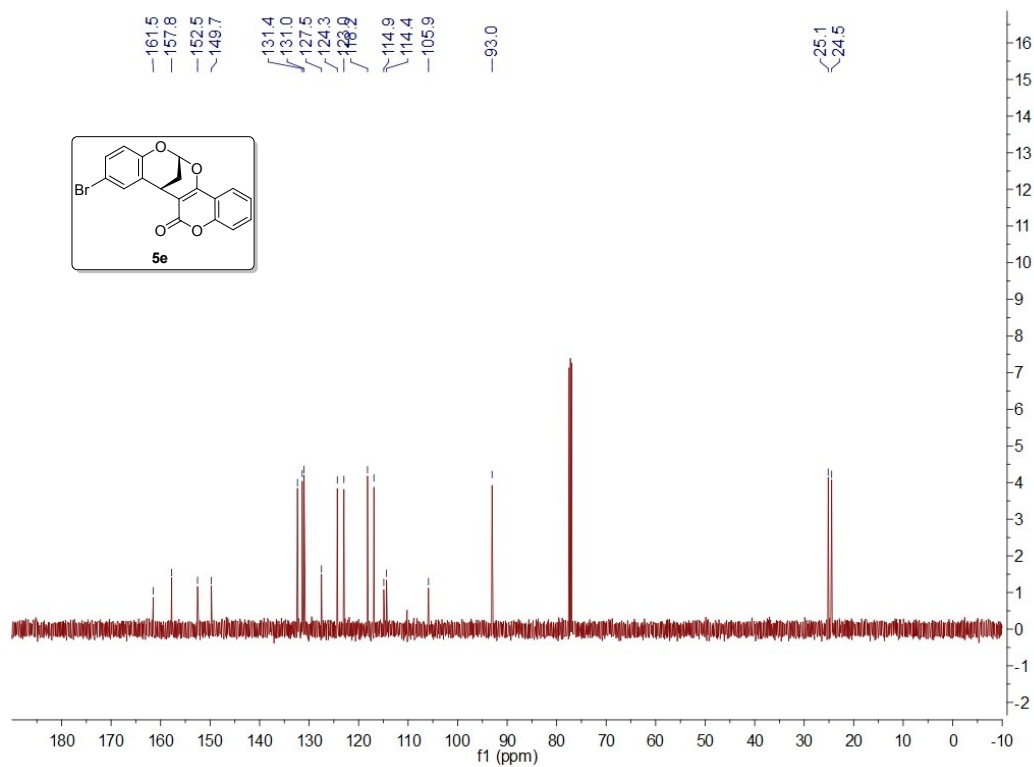
The  $^{13}\text{C}$  NMR spectrum of **8** (125 MHz,  $\text{CDCl}_3$ )



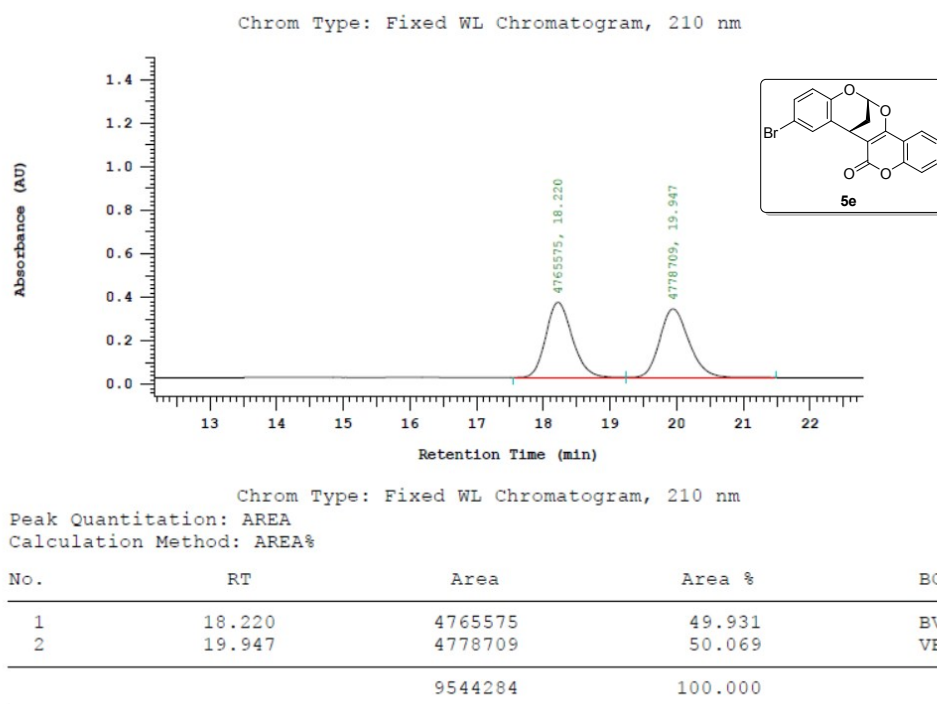
**The  $^1\text{H}$  NMR spectrum of 5e (500 MHz,  $\text{CDCl}_3$ )**



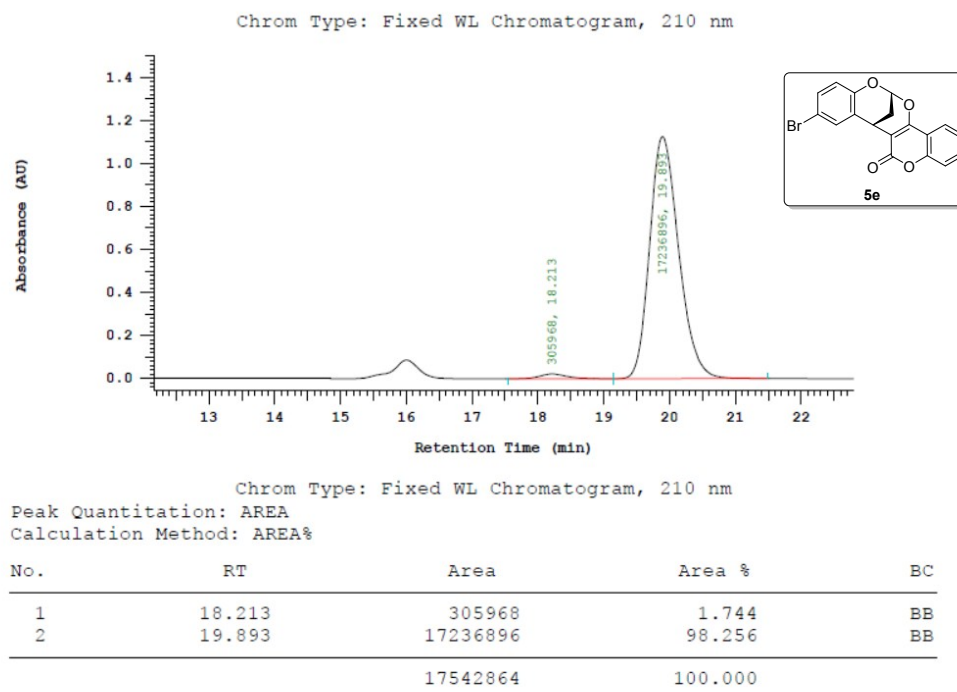
**The  $^{13}\text{C}$  NMR spectrum of 5e (125 MHz,  $\text{CDCl}_3$ )**



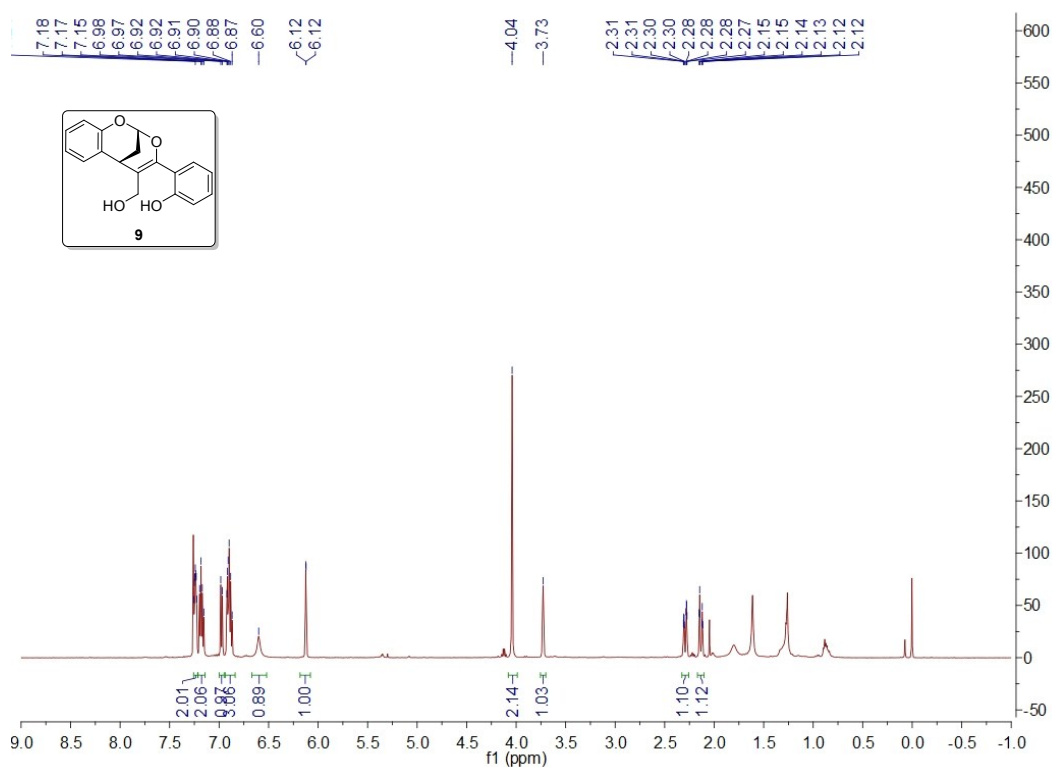
## The HPLC of racemic 5e



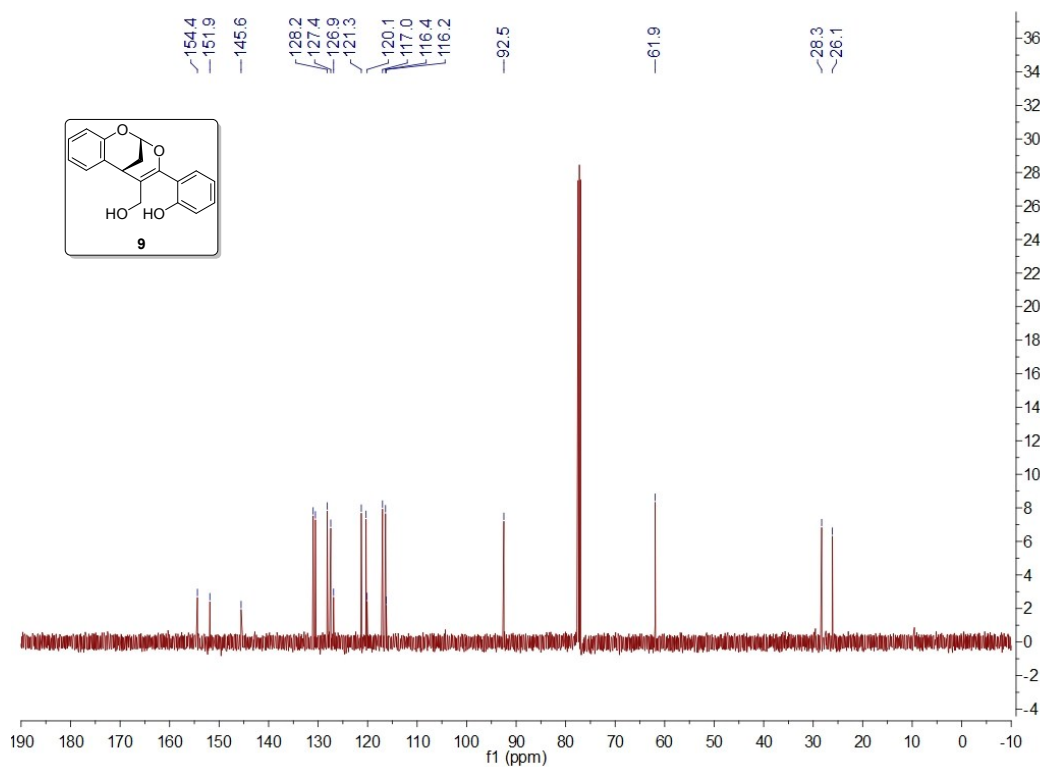
## The HPLC of chiral 5e



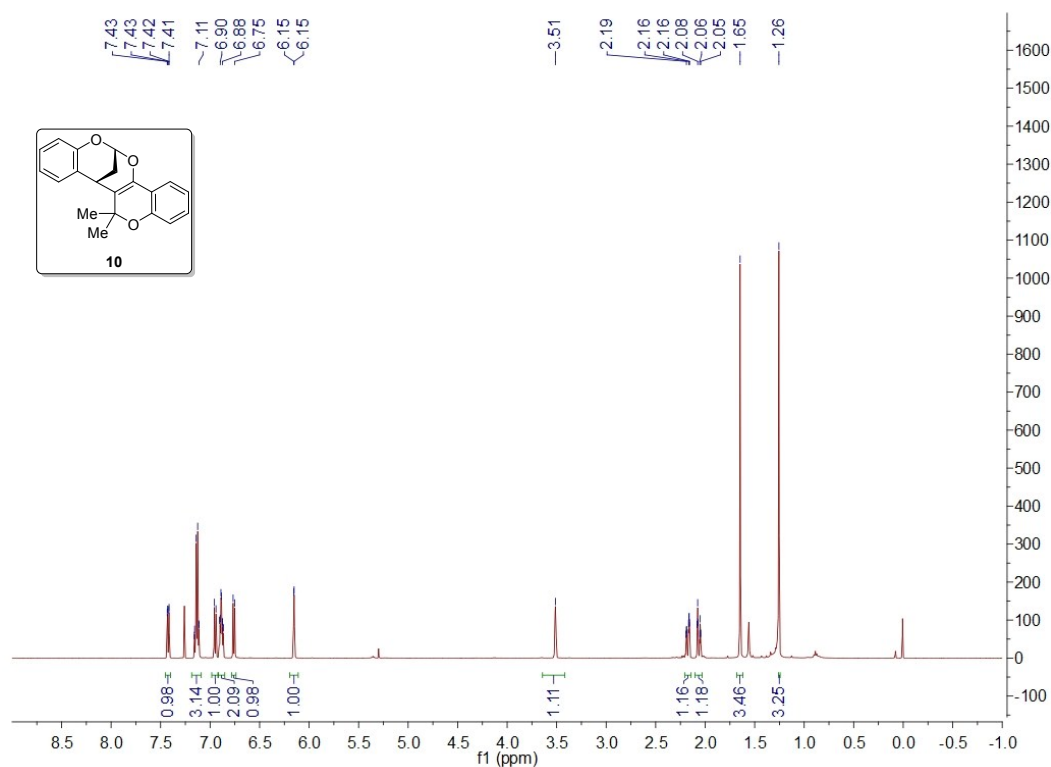
The  $^1\text{H}$  NMR spectrum of **9** (500 MHz,  $\text{CDCl}_3$ )



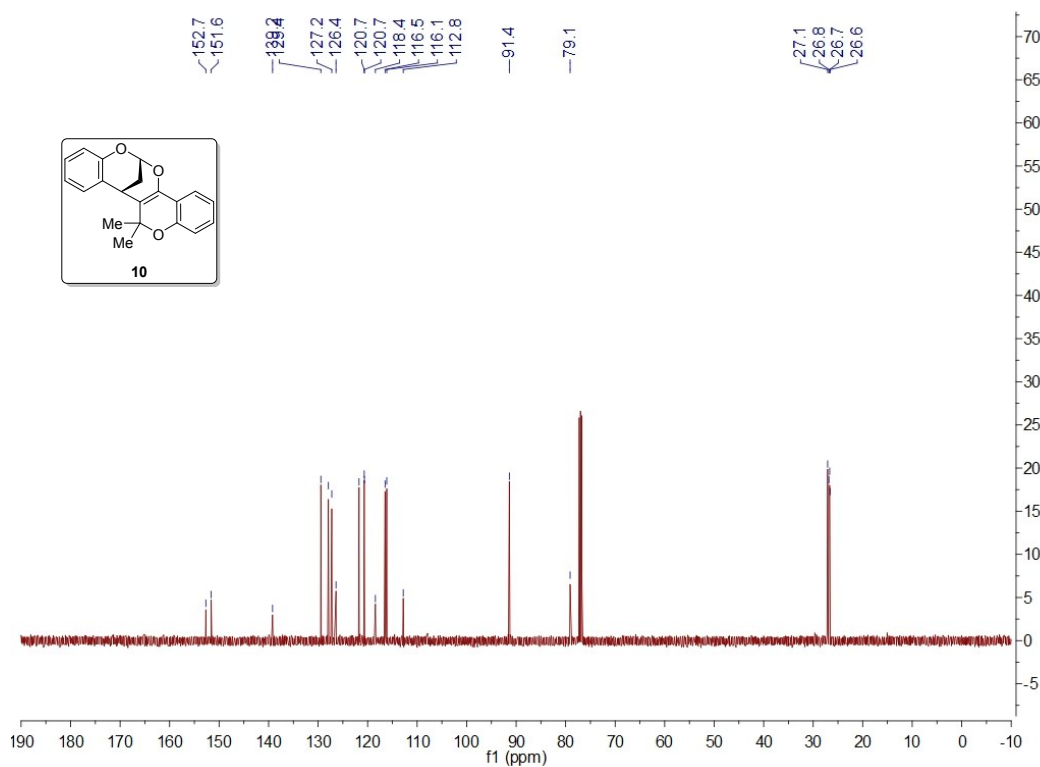
The  $^{13}\text{C}$  NMR spectrum of **9** (125 MHz,  $\text{CDCl}_3$ )



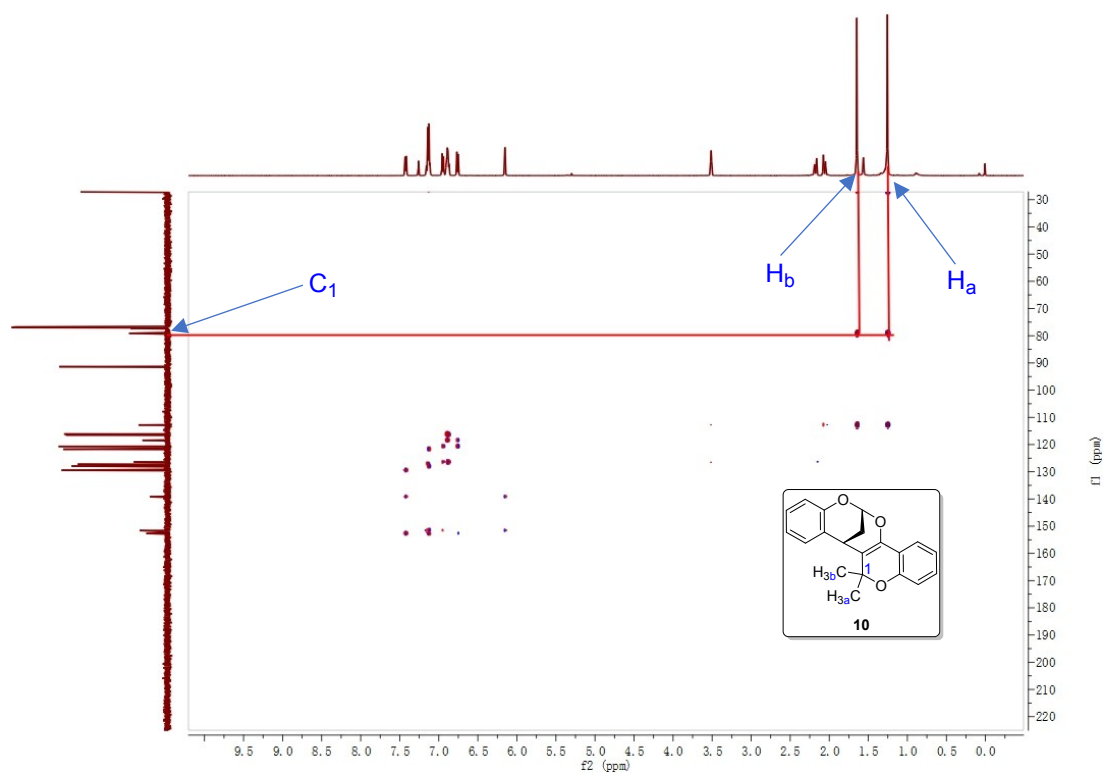
The  $^1\text{H}$  NMR spectrum of 10 (500 MHz,  $\text{CDCl}_3$ )



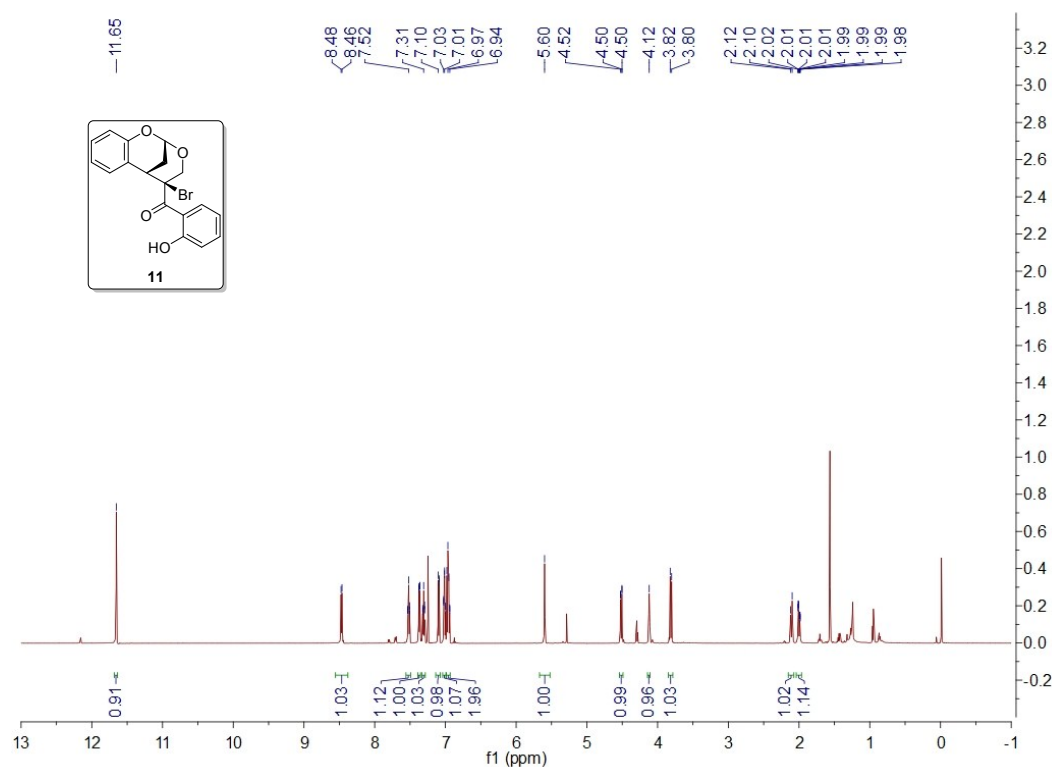
The  $^{13}\text{C}$  NMR spectrum of 10 (125 MHz,  $\text{CDCl}_3$ )



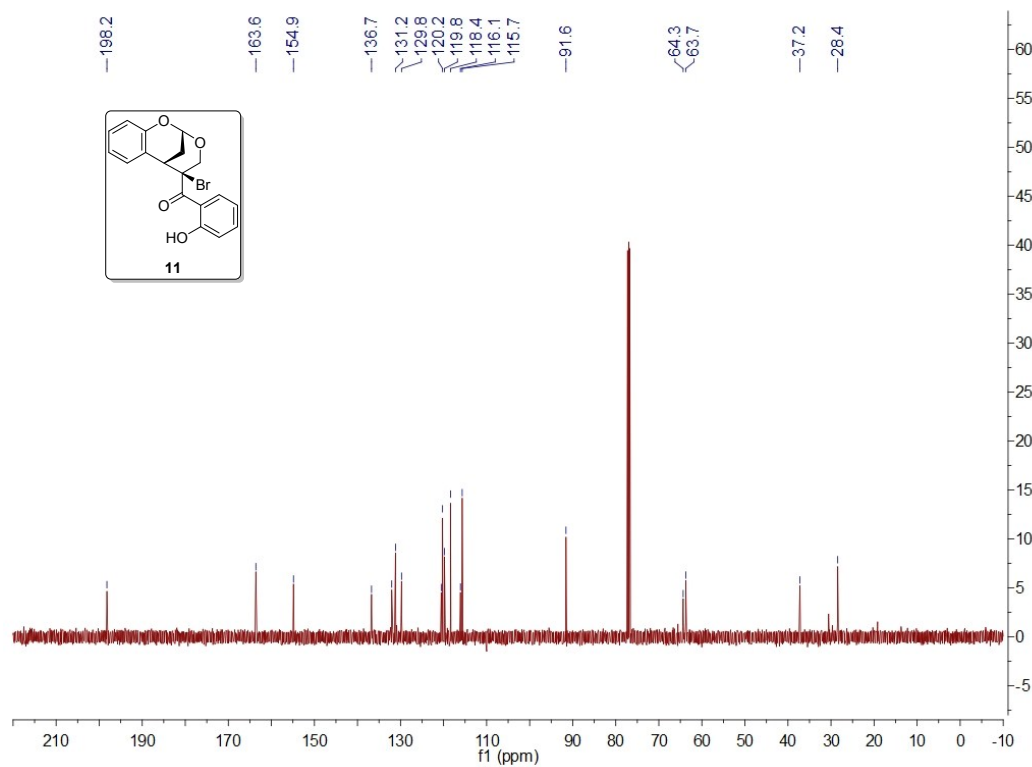
The HMBC spectrum of **10** (500 MHz, CDCl<sub>3</sub>)



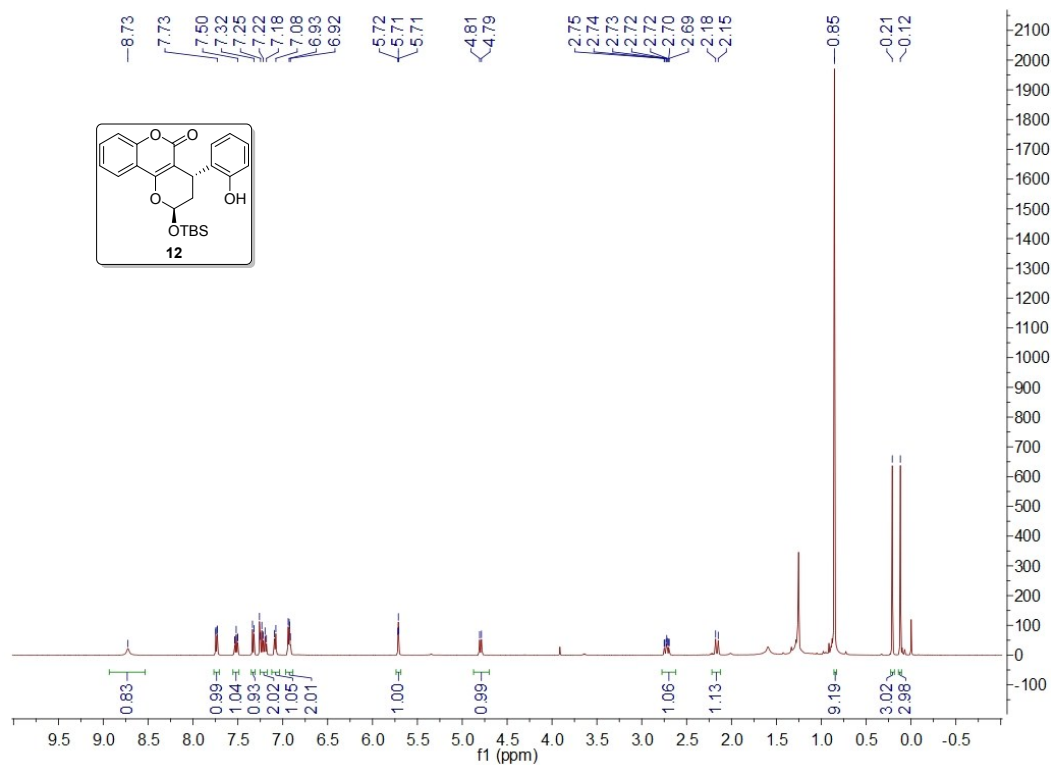
**The  $^1\text{H}$  NMR spectrum of 11 (600 MHz,  $\text{CDCl}_3$ )**



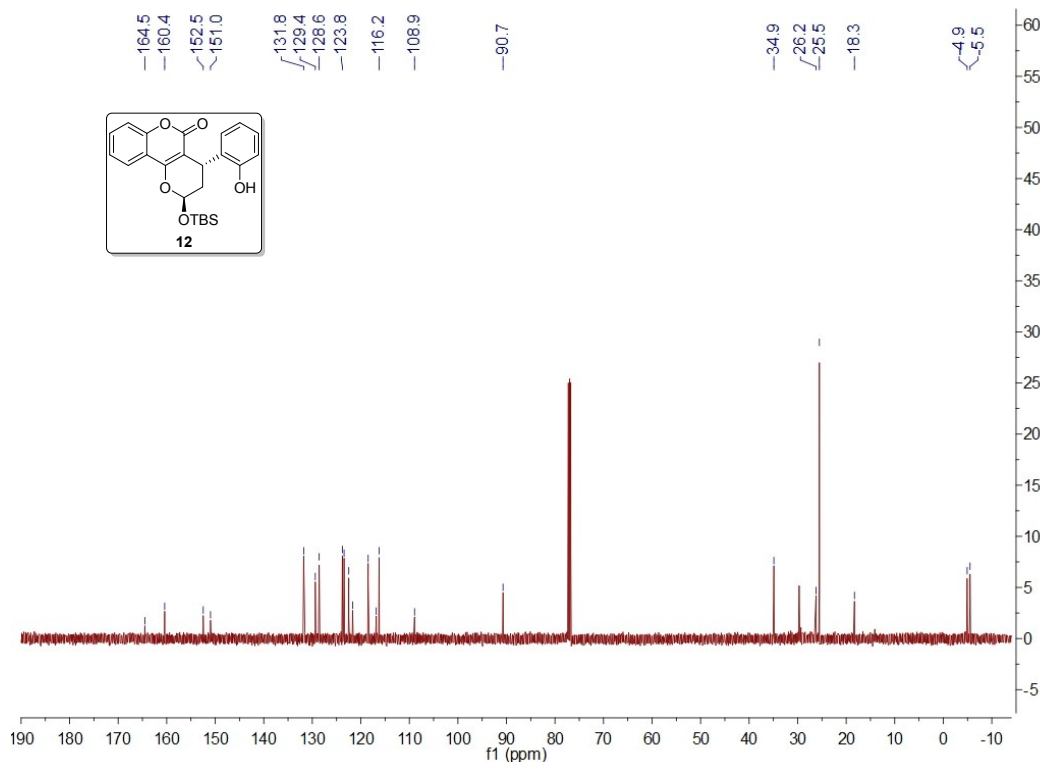
**The  $^{13}\text{C}$  NMR spectrum of 11 (125 MHz,  $\text{CDCl}_3$ )**



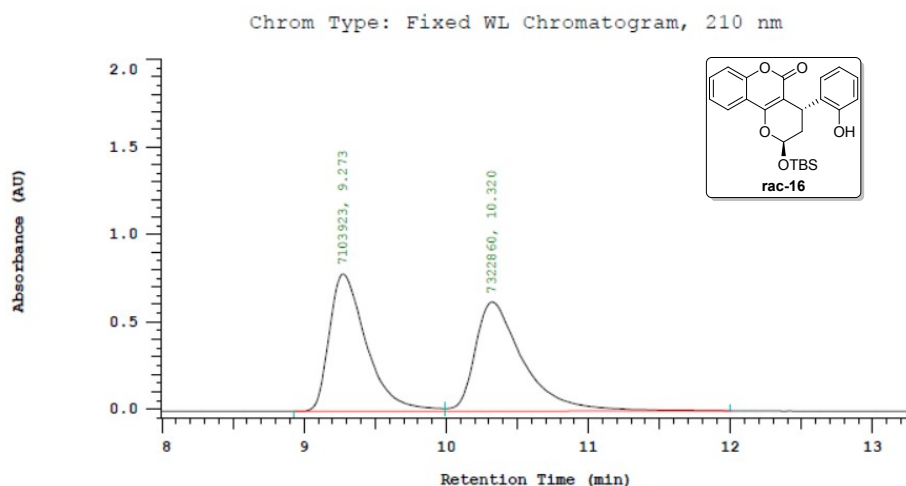
### The $^1\text{H}$ NMR spectrum of 16 (500 MHz, $\text{CDCl}_3$ )



### The $^{13}\text{C}$ NMR spectrum of 16 (125 MHz, $\text{CDCl}_3$ )



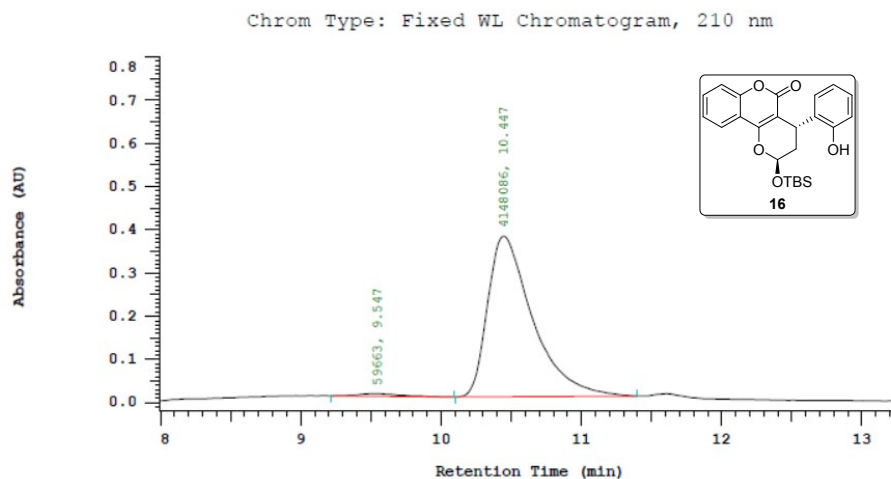
### The HPLC of racemic 16



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area     | Area %  | BC |
|-----|--------|----------|---------|----|
| 1   | 9.273  | 7103923  | 49.241  | BV |
| 2   | 10.320 | 7322860  | 50.759  | VB |
|     |        | 14426783 | 100.000 |    |

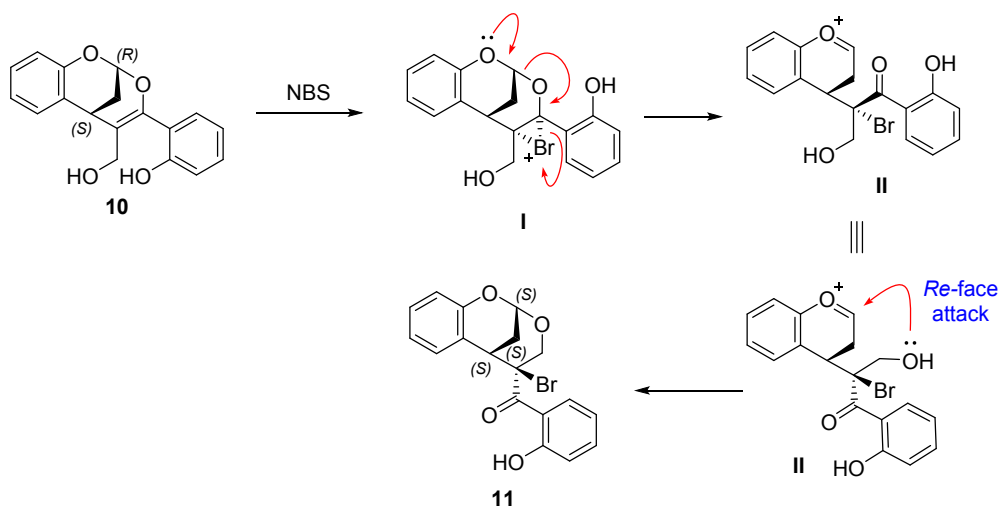
## The HPLC of chiral 16



Chrom Type: Fixed WL Chromatogram, 210 nm  
 Peak Quantitation: AREA  
 Calculation Method: AREA%

| No. | RT     | Area    | Area %  | BC |
|-----|--------|---------|---------|----|
| 1   | 9.547  | 59663   | 1.418   | BB |
| 2   | 10.447 | 4148086 | 98.582  | BB |
|     |        | 4207749 | 100.000 |    |

## H. Proposed mechanism for the preparation of **11**

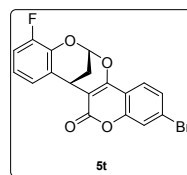
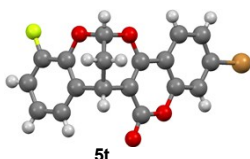


The reaction of the enantiopure dihydroxy compound **10** with NBS formed the electrophilic bromonium ion **I**, which could be converted into oxocarbenium ion **II** via electron transfer. Subsequently, the intramolecular *Re*-face attack of the aliphatic hydroxyl to the oxocarbenium ion moiety gave the new bicyclic acetal compound **11** with the proposed absolute configuration.

# I. Single crystal X-Ray diffraction data

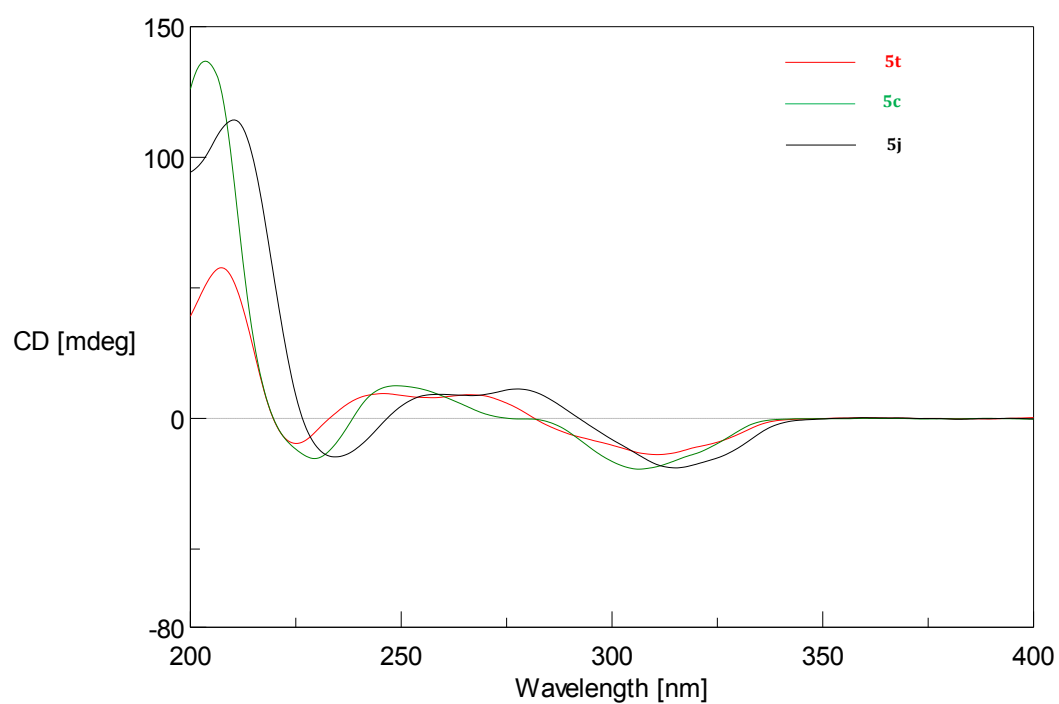
[CCDC 1959362 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via [www.ccdc.cam.ac.uk/data\\_request/cif](http://www.ccdc.cam.ac.uk/data_request/cif).]

Absolute configuration of **5t**



|                                                               |                 |                                 |              |
|---------------------------------------------------------------|-----------------|---------------------------------|--------------|
| Bond precision:                                               | C-C = 0.0063 Å  | Wavelength=1.54184              |              |
| Cell:                                                         | a=9.2599(2)     | b=12.5261(3)                    | c=12.6868(4) |
|                                                               | alpha=90        | beta=90                         | gamma=90     |
| Temperature:                                                  | 293 K           |                                 |              |
|                                                               | Calculated      | Reported                        |              |
| Volume                                                        | 1471.55(7)      | 1471.54(6)                      |              |
| Space group                                                   | P 21 21 21      | P 21 21 21                      |              |
| Hall group                                                    | P 2ac 2ab       | P 2ac 2ab                       |              |
| Moiety formula                                                | C18 H10 Br F O4 | ?                               |              |
| Sum formula                                                   | C18 H10 Br F O4 | C18 H10 Br F O4                 |              |
| Mr                                                            | 389.16          | 389.17                          |              |
| Dx, g cm-3                                                    | 1.757           | 1.757                           |              |
| Z                                                             | 4               | 4                               |              |
| Mu (mm-1)                                                     | 4.115           | 4.115                           |              |
| F000                                                          | 776.0           | 776.0                           |              |
| F000'                                                         | 775.60          |                                 |              |
| h,k,lmax                                                      | 11,14,15        | 11,14,15                        |              |
| Nref                                                          | 2631[ 1524]     | 2633                            |              |
| Tmin,Tmax                                                     | 0.641,0.636     | 0.688,1.000                     |              |
| Tmin'                                                         | 0.581           |                                 |              |
| Correction method= # Reported T Limits: Tmin=0.688 Tmax=1.000 |                 |                                 |              |
| AbsCorr = MULTI-SCAN                                          |                 |                                 |              |
| Data completeness=                                            | 1.73/1.00       | Theta(max)= 67.195              |              |
| R(reflections)=                                               | 0.0320( 2427)   | wR2(reflections)= 0.0803( 2633) |              |
| S =                                                           | 1.063           | Npar= 218                       |              |

ECD analysis of compound **5c**, **5j** and **5t**



ECD spectra were recorded in methanol on JASCO J-815 spectropolarimeter (JASCO Corporation, Tokyo, Japan).

The nearly identical ECD spectra of **5t**, **5c** and **5j** indicate that they display the same stereochemistry.