

Enantiodivergent Reduction of α -Keto Amides Catalyzed by High Valent, Chiral Oxido-Vanadium (V) Complexes

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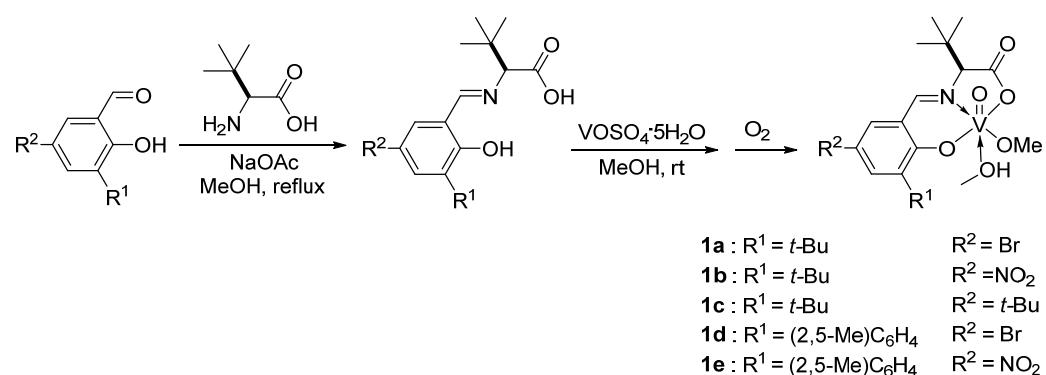
1. General materials and methods:

¹H NMR and ¹³C NMR spectra were recorded on 400 MHz ¹H (100 MHz ¹³C) spectrometers in deutero chloroform with chloroform or deutero methanol with methanol as an internal reference unless otherwise stated. Chemical shifts are reported in ppm (δ). Coupling constants, J , are reported in Hz. The abbreviations s, d, t, pent, quint, sext, dd, ddd, dt, and m stand for the resonance multiplicities singlet, doublet, triplet, pentet, quintet, sextet, doublet of doublets, doublet of doublet of doublets, doublet of triplets, and multiplet, respectively. Mass spectra were recorded with an ionization voltage of 70 or 20 eV unless otherwise stated. Elemental analyses were obtained by the Department of Chemistry, National Tsing Hua University, Taiwan or Department of Photonics, National Chiao Tung University, Taiwan. Fast atom bombardment (FAB) and electrospray ionization (ESI) mass spectra were recorded with data reported in the form m/e (intensity relative to base peak). Analytical TLC was performed on silica gel plates. Visualization was accomplished with UV light (254 nm) or with KMnO₄ staining agents. Column (flash) chromatography was performed using 40-60 μ m silica gel. Analytical high pressure liquid chromatography (HPLC) was performed with a built-in photometric detector ($\lambda = 220$ nm or 254 nm) using a Diacel AS-H, AD-H (0.46 cm \times 25cm). Solvents for HPLC analyses were of spectroscopic grade and filtered before use. All enantiomeric excess determinations for optically enriched products were correlated with the corresponding racemic samples by HPLC analyses on chiral columns. Solvents for extraction and chromatography were reagent grade. Optical rotations are reported as follows: $[\alpha]_D^T$ ($c = g/100mL$, solvent). Toluene, Tetrahydrofuran (THF) and 1,2-Dimethoxyethane (DME) were dried over sodium benzophenone-ketyl intermediate under N₂ atmosphere and distilled before use. Dichloromethane (DCM) were dried over CaH₂. Chlorobenzene was distilled from anhydrous K₂CO₃. 4,4,5,5-Tetramethyl-1,3,2-dioxaborolane (Pinacolborane) and Catecholborane are brought from Acros Organics and are used as such without any further purification. Benzyl isocyanide was purchased from ACROS and used as received. SiCl₄ was purchased from Sigma Aldrich and used as received. All reaction products were isolated as chromatographically pure materials. All the other chemicals are bought from Sigma Aldrich or Acros Organics and are used as such without any purification. Catalysts were synthesized according our previously reported procedure.¹ Compound **3h-j**, **3n** are new compounds which were prepared according to method-b.³

List of Abbreviations:

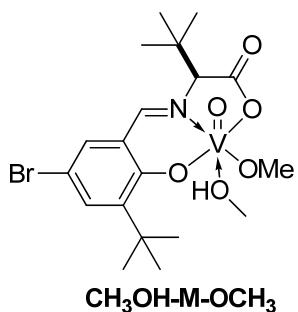
HPLC - high performance liquid chromatography, FT-IR – Fourier transform infrared, HRMS (ESI⁺) – High resolution mass spectra (electrospray ionization), ee - enantiomeric excess, THF-Tetrahydrofuran, TLC - Thin layer chromatography, DME - 1,2 Dimethoxyethane, DCM-Dichloromethane, TBME - *tert*-Butyl methyl ether, HB(pin) - Pinacolborane, HB(Cat) - Catecholborane, Ar – Argon,

2. General Synthetic procedure and characterization data of oxo-vanadium(V) complexes **1a-1e**.¹

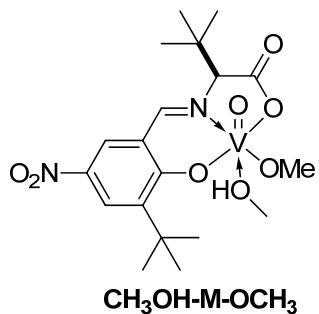


In a 50-mL, two-necked, round-bottomed flask was placed L-*tert*-leucine (5 mmol) and NaOAc-5·H₂O (1.17 g, 10 mmol) in degassed methanol (10 mL). After having been stirred at 60 °C for 10 min to affect their complete dissolution, the reaction mixture was treated dropwise with a solution of respective 2-hydroxy-benzaldehyde derivatives (5 mmol) in degassed methanol (12.5 mL). The reaction mixture became homogeneous by heating at 80 °C for 15 min and then gradually cooled to ambient temperature for 2 h. To the resultant Schiff base was added a solution of vanadyl (IV) sulfate trihydrate (1.08 g, 5 mmol) in degassed methanol (20 mL). Dark green complex started crashing out in 15 min. The reaction mixture was stirred for 2h and then concentrated to half of the original solvent volume. The crude vanadyl(IV) complex collected by filtration was washed sequentially with water (5 × 25 mL) and cold ether (5 × 25 mL) and then dried *in vacuo* to furnish pure vanadyl(IV) catalyst. The corresponding analytically pure oxo-vanadium (V) complexes were obtained by re-crystallization from oxygen-saturated MeOH and were used for asymmetric

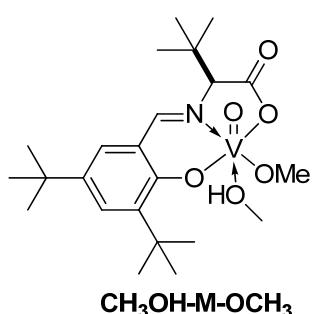
reduction experiments.



Data for complex 1a: **¹H NMR (CD₃OD, 400 MHz)** δ 8.54 (bs, 1H), 7.66 (d, *J* = 2.5 Hz, 1H), 7.63 (d, *J* = 2.5 Hz, 1H), 4.15 (s, 1H), 3.35 (s, OCH₃), 1.45 (s, 9H), 1.20 (s, 9H); **⁵¹V NMR (CD₃OD, 105 MHz)** δ -567.3; **¹³C NMR (CD₃OD, 100 MHz)** δ 167.7, 142.3, 137.0, 136.3, 135.1, 134.7, 123.8, 111.9, 84.7, 49.8, 38.3, 36.3, 29.9, 28.0; **IR (KBr)** 2965 (s), 2913 (m), 2869 (m), 1663 (s), 1615 (s, C=N), 1578 (m), 1548 (m, COO), 1480 (w), 1429 (m), 1368 (m), 1320 (m), 1297 (s), 1181 (m), 1055 (w), 1031 (w), 993 (m, V=O); **M.W. (M: C₁₇H₂₂BrNO₄V)** 434; **MS (ESI)** 450 (**MOH-1⁺**, 100); [α]_D³⁴ 306.53 (*c* 0.1, CH₃OH); **TLC R_f** 0.20 (MeOH/CH₂Cl₂, 1/9); **Anal. Calcd. For [(H₂O)MOH]: C, 43.42; H, 5.36; N, 2.98. Found: C, 43.54; H, 5.13; N, 3.41.**

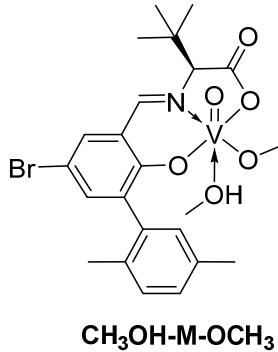


Data for complex 1b: **¹H NMR (CD₃OD, 400 MHz)** δ 8.71 (bs, 1H), 8.54 (d, *J* = 2.6 Hz, 1H), 8.39 (d, *J* = 2.5 Hz, 1H), 4.24 (s, 1H), 3.31 (s, OCH₃), 1.49 (s, 9H), 1.22 (s, 9H); **⁵¹V NMR (CD₃OD, 105 MHz)** δ -549.8, -568.8; **¹³C NMR (CD₃OD, 100 MHz)** δ 168.0, 140.5, 139.1, 130.1, 130.0, 128.4, 127.6, 121.5, 84.9, 38.3, 36.5, 29.7, 28.0; **IR (KBr)** 2965 (w), 2916 (w), 2879 (w), 1627 (m, C=N), 1598 (m, COO), 1509 (w), 1326 (m), 1326 (w), 1225 (w), 1187 (w), 1113 (w), 1034 (w), 990 (w), 927 (w, V=O); **M.W. (M: C₁₇H₂₂N₂O₆V)** 401.3; **MS (ESI)** 850 (**M₂O+H₂O⁺**, 90), 419 (**MOH+H⁺**, 9), 417 (**MOH-1⁺**, 100); [α]_D³⁴ 83.93 (*c* 0.1, CH₃OH); **TLC R_f** 0.30 (MeOH/CH₂Cl₂, 1/4); **Anal. Calcd. For [(H₂O)MOH]: C, 45.57; H, 5.83; N, 6.15.**

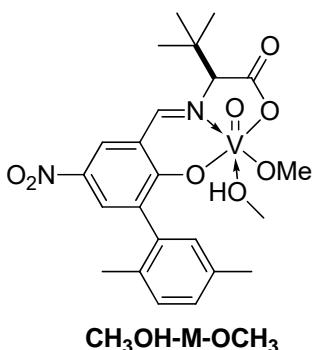


Data for 1c : **¹H NMR (CD₃OD, 400 MHz)** δ 8.60 (bs, 1H), 7.68 (d, *J* = 2.3 Hz, 1H), 7.48 (d, *J* = 2.3 Hz, 1H), 4.14 (s, 1H), 3.35 (s, OCH₃), 1.47 (s, 9H), 1.35 (s, 9H), 1.20 (s, 9H); **⁵¹V NMR (CD₃OD, 105 MHz)** δ -563.2; **¹³C NMR (CD₃OD, 100 MHz)** δ 180.1, 168.9, 161.7, 143.5, 138.6, 132.4, 129.4, 121.9, 84.7, 38.3, 36.3, 35.3, 31.8, 30.3, 28.1; **IR (KBr)** 2958 (m), 2913 (w), 2868 (w), 1699 (m), 1612 (s, C=N), 1559

(w, COO), 1474 (w), 1459 (m), 1436 (w), 1418 (w), 1395 (w), 1363 (w), 1275 (w), 1260 (w), 1210 (w), 1183 (w), 1000 (w, V=O); M.W. (**M**: C₂₁H₃₁NO₄V) 412.4; MS (ESI) 881 (**M**₂O+H₂O+Na⁺, 6), 460 ([H₂OMOCH₃]⁺, 77), 444 ([MOCH₃+H]⁺, 13), 413 (**M**+H⁺, 100); [α]_D³⁴ 262.87 (c 0.1, CH₃OH); TLC **R**_f 0.37 (MeOH/CH₂Cl₂, 1/8); Anal. Calcd. For [(H₂O)**M**O₄]: C, 56.37; H, 7.66; N, 3.13. Found: C, 55.72; H, 7.32; N, 2.71.

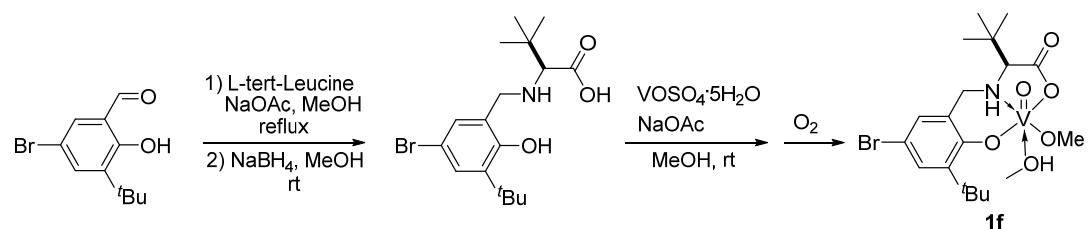


Data for 1d: **¹H NMR (CD₃OD, 400 MHz)**. δ 8.61 (s, 1H), 7.79 (s, 1H), 7.52 (s, 1H), 7.14 (d, *J* = 7.6 Hz, 1H), 7.07 (d, *J* = 7.6 Hz, 1H), 6.97 (s, 1H), 4.17 (s, 1H), 3.33 (s, OCH₃), 2.30 (s, 3H), 2.22 (s, 3H), 1.19 (s, 9H); **⁵¹V NMR (CD₃OD, 105 MHz)** δ -559.3; **¹³C NMR (CD₃OD, 100 MHz)** δ 167.3, 140.4, 137.6, 136.1, 135.9, 135.0, 131.8, 130.7, 129.6, 123.4, 111.4, 84.6, 38.3, 28.0, 21.0, 19.8; **IR (KBr)** 3446 (br, w, NH), 2961 (m), 1687 (m), 1616 (s), 1553 (s), 1432 (s), 1314 (s), 1002 (m, V=O); [α]_D²⁵ 236.43 (c 0.1, CH₃OH); **TLC R**_f 0.28 ('PrOH/Hexanes, 1/5); **HRMS (ESI)** [M+H]⁺ Calcd for C₂₂H₂₆BrNO₅V: 514.0434, found: 514.0434.

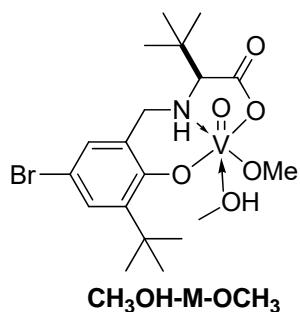


Data for 1e: **¹H NMR (CD₃OD, 400 MHz)** δ 8.79 (bs, 1H), 8.67 (d, *J* = 2.9 Hz, 1H), 8.28 (d, *J* = 2.8 Hz, 1H), 7.21 (d, *J* = 7.8 Hz, 1H), 7.15 (dd, *J* = 1.3, 7.8 Hz, 1H), 7.05 (s, 1H), 3.35 (s, OCH₃), 4.25 (s, 1H), 3.35 (s, OCH₃), 2.36 (s, 3H, CH₃), 2.25 (s, 3H, CH₃), 1.23 (s, 9H); **⁵¹V NMR (105 MHz, CD₃OD)** δ -559.6; **¹³C NMR (100 MHz, CD₃OD)** δ 178.8, 167.8, 166.2, 140.6, 137.2, 136.3, 135.1, 134.3, 132.1, 131.8, 130.8, 130.1, 121.2, 85.0, 38.4, 28.1, 21.0, 19.8; **IR (KBr)** 3005(w), 2975 (w), 1628 (m, C=N), 1599 (m), 1509 (m), 1456 (w), 1434 (w), 1326 (s), 1276 (s), 1260 (m), 1100 (m), 999 (w, V=O); [α]_D³⁴ 150.73 (c 0.1, CH₃OH); **TLC R**_f 0.25 (MeOH/CH₂Cl₂, 1/9). M.W. (**M**:C₂₁H₂₂N₂O₆V,449.4); **MS(ESI)** calcd. For C₂₂H₂₅N₂O₇V+1: 481.1101(**M**+OCH₃+1); found.481.1173 (**M**+OCH₃+1, 100);

3. Synthetic procedure and characterization data of oxo-vanadium(V) complex **1f**.



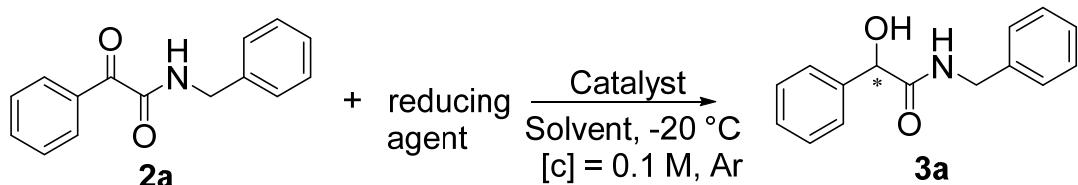
In a 50-mL, two-necked, round-bottomed flask was placed L-tert-leucine (5 mmol) and NaOAc-5·H₂O (643.5 mg, 5.5 mmol) in degassed methanol (10 mL). After having been stirred at 60 °C for 10 min to affect their complete dissolution, the reaction mixture was treated dropwise with a solution of 5-bromo-3-(*tert*-butyl)-2-hydroxybenzaldehyde (5 mmol) in degassed MeOH (12.5 mL). The reaction mixture refluxed at 80 °C for 20 hours and then gradually cooled to ambient temperature. To this mixture sodium borohydride (5 mmol) was added. After 10 mins, under ice bath 1 N aqueous solution of HCl was added to make the pH = 2, white solid was precipitated, washed with distilled water and dried to give compound 5. A solution of vanadyl (IV) sulfate trihydrate (1.08 g, 5 mmol) and NaOAc-5·H₂O (643.5 mg, 5.5 mmol) in degassed methanol (5 mL) was added to compound 5. The reaction mixture was stirred for 4h and then concentrated to half of the original solvent volume. The crude vanadyl(IV) complex collected by filtration was washed sequentially with water (5 × 25 mL) and cold ether (5 × 25 mL) and then dried *in vacuo* to furnish pure vanadyl(IV) catalyst. The corresponding analytically pure oxo-vanadium (V) complexes were obtained by re-crystallization from oxygen-saturated MeOH to obtain dark brown crystals of **1f** in 47% yield.



Data for complex **1f** : ^1H NMR (400 MHz, CD₃OD) δ 7.40 (d, $J = 2.2$ Hz, 1H), 7.26 (d, $J = 2.2$ Hz, 1H), 4.57 (bs, 1H), 3.95 (d, $J = 11.8$ Hz, 1H); 3.73 (d, $J = 11.9$ Hz, 1H), 3.35 (s, OCH₃), 3.28 (s, 1H), 1.48 (s, 9H), 1.21 (s, 9H); ^{13}C NMR (100 MHz, CD₃OD) δ 164.8, 140.4, 130.9, 130.4, 128.4, 126.5, 113.3, 76.0, 53.6, 49.8, 37.2, 36.2, 30.6, 27.2; ^{51}V NMR (105 MHz, CD₃OD) δ -512.0, -525.1; IR 3449(br, w, NH), 2958 (m), 2872 (m), 1654(s), 1463 (m), 1434 (s), 1409 (m), 1340 (m), 1267(s), 1247(s), 1053(s), 970(m, V=O); $[\alpha]_D^{34}$ 562.00 (c 0.1, CH₃OH); TLC R_f 0.1 (MeOH/CH₂Cl₂, 1/10).

4. General reaction procedure for the asymmetric reduction of

N-benzyl- α -keto-amides.

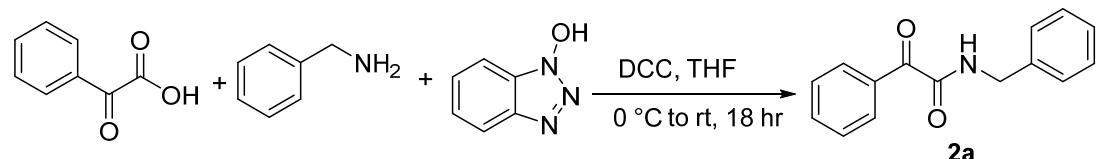


In a dried Schlenk tube was placed *N*-benzyl α -keto-amide (47.9 mg, 0.2 mmol) and vanadyl catalyst (9.4 mg, 0.02 mmol, 10 mol%) in anhydrous toluene (1.75 mL) under argon atmosphere. After having been stirred at ambient temperature for 30 min, the Schlenk tube was cooled to -20 °C for 30 min. A solution of HBpin or HBCat (64 μ L, 0.6 mmol, 3 equiv) in anhydrous toluene (186 μ L) was cooled to -20 °C and then added to the reaction mixture thru a gas-tight microsyringe. After having been complete as checked by TLC, the reaction was quenched by deionized water (2.5 mL) at -20 °C (10 min) and then gradually warmed to ambient temperature for 30 min. The reaction mixture was extracted with EtOAc (10 mL \times 3). The combined organic extracts were dried (MgSO₄, 1-1.2 g), filtered, and evaporated. The crude residue was purified by column chromatography (eluent: ethyl acetate in hexane) on silica gel to provide *N*-benzyl α -hydroxy-amide (46.9 mg, 98% yield).

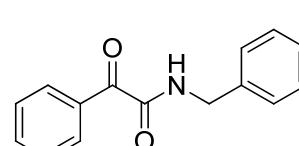
5. General synthetic procedure for starting materials:

5.1 Method (a)²

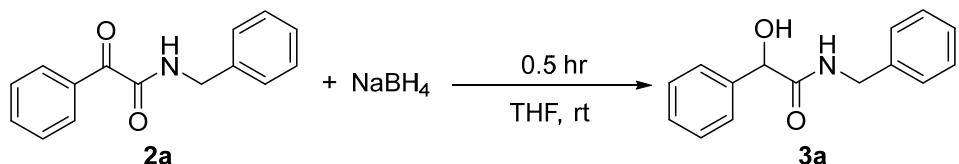
N-Benzyl-2-oxo-2-phenylacetamide (2a)¹



In a 250 mL, two-necked, round bottomed flask was placed phenylglyoxylic (1.5 g, 10.0 mmol, 1.0 equiv), benzylamine (1.1 mL, 10.0 mmol, 1.0 equiv) and *N,N*-dicyclohexylcarbodiimide (DCC, 2.1 g, 10.0 mmol, 1.0 equiv) in 100 mL anhydrous THF. To this reaction mixture was added hydroxybenzotriazole (HOBr, 1.4 g, 10.0 mmol, 1.0 equiv) and stirred at room temperature for 18 hours. The resulting DCU was filtered off with suction filter and the organic filtrate was dried over anhydrous MgSO₄ (3.0 g), and then evaporated to dryness. The crude product was recrystallized from hot diethyl ether to give **2a** (1.93 g, 80% yield).

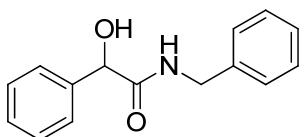
 Data for **2a** : ¹H NMR (CDCl₃, 400 MHz) δ 8.37 (d, *J* = 7.6 Hz, 2H), 7.63 (t, *J* = 7.6 Hz, 1H), 7.49 (t, *J* = 8.0 Hz, 2H), 7.39-7.32 (m, 5H), 4.58 (d, *J* = 6.0 Hz, 2H); ¹³C NMR (CDCl₃, 100 MHz) δ 187.5, 161.6, 137.0, 134.2, 133.1, 131.0, 128.6, 127.7, 127.6, 43.2; TLC R_f 0.22 (EtOAc/Hexane, 1/5); HRMS (ESI) Calcd for C₁₅H₁₃NO₂ : 239.0946, found : 239.0945.

N-Benzyl-2-hydroxy-2-phenylacetamide (3a)¹



In a 25 mL, two-necked, round bottomed flask was placed compound **2a** (224 mg, 0.94 mmol, 1.0 eq.) in anhydrous THF (9.0 mL). Sodium borohydride (36 mg, 0.94 mmol) was then added and the reaction mixture was stirred at ambient temperature for 30 minutes. The reaction mixture was quenched with deionized water (8 mL) and then extracted with ethyl acetate (8 mL × 3). The

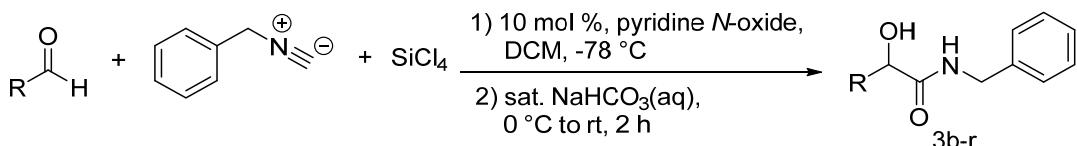
combined extracts were dried over anhydrous MgSO₄, filtered, evaporated. The crude product was purified by column chromatography (ethyl acetate/hexane) on silica gel.



Data for 3a : **¹H NMR (CDCl₃, 400 MHz)** δ 7.39-7.27 (m, 8H), 7.18-7.16 (m, 2H), 6.50 (bs, 1H, NH), 5.05 (d, *J* = 3.2 Hz, 1H), 4.48-4.37 (m, 2H), 3.64 (d, *J* = 3.2 Hz, 1H, OH); **¹³C NMR (CDCl₃, 100 MHz)** δ 172.3, 139.4, 137.4, 128.5, 128.3, 127.4, 126.6, 73.9, 43.1; **TLC R_f** 0.28

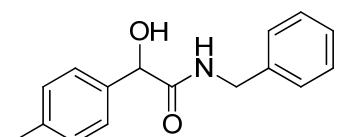
(EtOAc/hecane, 1/2); **HPLC conditions for 3a** : *t_R* 17.04 min (*R*-isomer), 27.90 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₅H₁₅NO₂ : 241.1103, found : 241.1103.

5.2 Method (b) (3b-r):³



In a 100 mL, three-necked, round bottomed flask was placed pyridine *N*-oxide (38 mg, 0.4 mmol, 0.1 equiv) in 12 ml of anhydrous CH₂Cl₂. To this reaction mixture, respective aldehyde (4.0 mmol, 1.0 equiv) was added. Benzyl isocyanide (585 μ L, 4.8 mmol, 1.2 equiv) in 10 mL of CH₂Cl₂ was added dropwise through addition funnel at -78°C , and then silicon tetrachloride (SiCl₄, 504 μ L, 4.4 mmol, 1.1 equiv) was slowly added to the reaction. After the reaction was completed, a sat. aqueous solution of NaHCO₃ (30 mL) was added under ice bath and stirred for two hours. A white precipitate of SiO₂ was formed, and the mixture was filtered through Celite to remove white solid. The resulting filtrate was extracted with CH₂Cl₂ (30 mL \times 3), dried (MgSO₄), and evaporated to give a crude product. The crude product was purified by column chromatography on silica gel with EtOAc/hexanes as eluents. The purified product was recrystallized from hot benzene (20 mL) to give *N*-benzyl- α -hydroxy-amide derivatives in 48–92% yields.

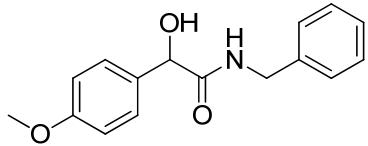
***N*-Benzyl-2-hydroxy-2-*p*-tolyl-acetamide (3b)¹**



Data for 3b : **¹H NMR (CDCl₃, 400 MHz)** δ 7.33-7.28 (m, 5H), 7.20-7.17 (m, 4H), 6.35 (bs, 1H, NH), 5.06 (d, *J* = 3.3 Hz, 1H), 4.52-4.39 (m, 2H), 3.43 (d, *J* = 3.4 Hz, 1H), 2.35 (s, 1H); **¹³C NMR**

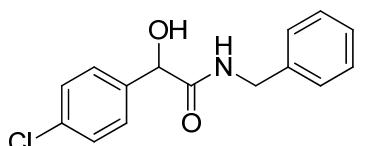
(CDCl₃, 100 MHz) δ 172.6, 138.0, 137.6, 136.4, 129.2, 128.5, 127.4, 127.3, 126.5, 73.8, 43.0, 21.0; **TLC R_f** 0.2 (EtOAc/hexane, 1/4); **HPLC conditions for 3b** : t_R 16.35 min (*R*-isomer), 28.10 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₆H₁₇NO₂ : 255.1059, found : 255.1258.

N-Benzyl-2-hydroxy-2-(4'-methoxyphenyl)-acetamide (3c)¹



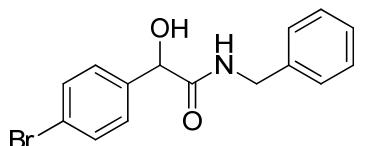
Data for 3c : **¹H NMR (CDCl₃, 400 MHz)** δ 7.32-7.26 (m, 5H), 7.19 (d, *J* = 6.8 Hz, 2H), 6.88 (d, *J* = 8.6 Hz, 2H), 6.47 (bs, 1H, NH), 5.01 (d, *J* = 2.8 Hz, 1H), 4.49-4.37 (m, 2H), 4.38 (s, 1H), 3.54 (d, *J* = 3.2 Hz, 1H, OH); **¹³C NMR (CDCl₃, 100 MHz)** δ 172.3, 159.8, 137.7, 131.5, 128.6, 128.1, 127.5, 114.2, 73.7, 55.2, 43.4; **TLC R_f** 0.075 (EtOAc/hexane, 2/5); **HPLC conditions for 3c** : t_R 12.21 min (*R*-isomer), 21.55 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 85/15, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₆H₁₇NO₃ : 271.1208, found : 271.1211.

N-Benzyl-2-hydroxy-2-(4'-chlorophenyl)-acetamide (3d)¹



Data for 3d : **¹H NMR (CDCl₃, 400 MHz)** δ 8.36 (d, *J* = 8.8 Hz, 2H), 7.46 (d, *J* = 8.5 Hz, 2H), 7.42 (bs, 1H, NH), 7.39-7.29 (m, 5H), 4.56 (d, *J* = 6.0 Hz, 2H); **¹³C NMR (CDCl₃, 100 MHz)** δ 172.2, 137.9, 137.3, 133.9, 128.5, 128.4, 127.8, 127.4, 1327.3, 73.1, 43.0; **TLC R_f** 0.175 (EtOAc/hexane, 1/2); **HPLC conditions for 3d** : t_R 14.30 min (*R*-isomer), 29.31 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₅H₁₄ClNO₂ : 275.0713, found : 275.0716.

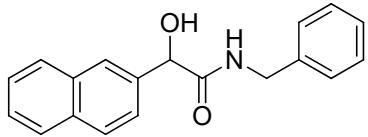
N-Benzyl-2-hydroxy-2-(4'-bromophenyl)-acetamide (3e)¹



Data for 3e : **¹H NMR (CDCl₃, 400 MHz)** δ 7.49 (d, *J* = 8.4 Hz, 2H), 7.36-7.26 (m, 5H), 7.18 (d, *J* = 7.2 Hz, 2H), 6.54 (bs, 1H, NH), 5.03 (d, *J* = 3.6 Hz, 1H), 4.47-4.37 (m, 2H), 3.59 (d, *J* = 3.6 Hz, 1H, OH); **¹³C NMR (CDCl₃, 100 MHz)** δ 171.6, 138.3, 137.4, 131.7, 128.7, 128.3, 127.6, 127.5, 122.5, 73.4, 43.3; **TLC R_f** 0.07 (EtOAc/hexane, 1/4); **HPLC conditions for 3e** : t_R 16.71 min (*R*-isomer), 36.24 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₅H₁₄BrNO₂ : 319.0208, found :

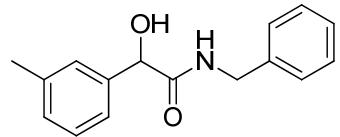
319.0207.

N-Benzyl-2-hydroxy-2-naphthalen-2-yl-acetamide (3f)¹



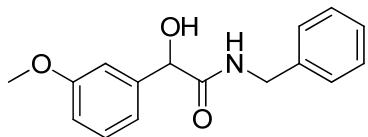
Data for 3f : **¹H NMR (DMSO-d₆, 400 MHz) δ** 8.63 (t, *J* = 6.0 Hz, 1H), 7.92-7.86 (m, 4H), 7.58 (d, *J* = 4.2 Hz, 1H), 7.63-7.47 (m, 2H), 7.27-7.18 (m, 5H), 6.38 (d, *J* = 3.6 Hz, 1H, NH), 5.14 (d, *J* = 3.5 Hz, 1H, OH), 4.34-4.24 (m, 2H); **¹³C NMR (DMSO-d₆, 100 MHz) δ** 172.2, 139.6, 138.9, 132.6, 132.5, 128.2, 127.5, 127.5, 126.7, 126.2, 125.9, 125.3, 124.8 73.7, 41.8; **TLC R_f** 0.05 (EtOAc/hexane, 1/3); **HPLC conditions for 3f :** *t_R* 10.39 min (*R*-isomer), 18.26 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 80/20, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₉H₁₇NO₂ : 291.1259, found : 291.1258.

N-Benzyl-2-hydroxy-2-m-tolyl-acetamide (3g)⁴



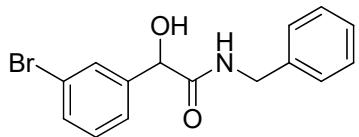
Data for 3g : **¹H NMR (CDCl₃, 400 MHz) δ** 7.33-7.23 (m, 4H), 7.20-7.13 (m, 5H), 6.45 (bs, 1H, NH), 5.02 (d, *J* = 3.2 Hz, 1H), 4.50-4.38 (m, 2H), 3.59 (d, *J* = 3.4 Hz, 1H), 2.34 (s, 1H); **¹³C NMR (CDCl₃, 100 MHz) δ** 172.2, 139.3, 138.5, 137.7, 129.3, 128.6, 127.5, 127.4, 123.8, 74.1, 43.3, 21.3; **TLC R_f** 0.15 (EtOAc/hexane, 1/2); **HPLC conditions for 3g :** *t_R* 13.50 min (*R*-isomer), 22.21 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₆H₁₈NO₂ : 256.1337 ([M+H]⁺), found : 256.1333 ([M+H]⁺).

N-Benzyl-2-hydroxy-2-(3'-methoxyphenyl)-acetamide (3h)



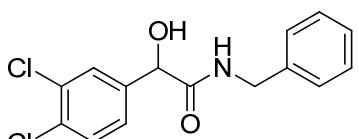
Data for 3h : **¹H NMR (CDCl₃, 400 MHz) δ** 7.32-7.24 (m, 4H), 7.18 (d, *J* = 1.7 Hz, 2H), 7.16 (s, 1H), 6.97 (d, *J* = 7.6 Hz, 1H), 6.93 (t, *J* = 2.2 Hz, 1H), 6.88-6.85 (m, 1H), 6.51 (bs, 1H, NH), 5.01 (d, *J* = 2.8 Hz, 1H), 4.48-4.36 (m, 2H), 3.77 (s, 3H), 3.75 (d, *J* = 3.2 Hz, 1H, OH); **¹³C NMR (CDCl₃, 100 MHz) δ** 172.1, 159.8, 140.9, 137.6, 129.7, 128.6, 127.4, 119.0, 114.3, 111.9, 73.9, 55.1, 43.3; **TLC R_f** 0.18 (EtOAc/hexane, 1/2); **HPLC conditions for 3h :** *t_R* 12.06 min (*R*-isomer), 16.55 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 85/15, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₆H₁₈NO₃ : 272.1286 ([M+H]⁺), found : 272.1288 ([M+H]⁺).

N-Benzyl-2-hydroxy-2-(3'-bromophenyl)-acetamide (3i)



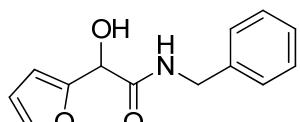
Data for 3i : ¹**H NMR (CDCl₃, 400 MHz) δ** 7.58 (s, 1H), 7.47 (td, *J* = 8.0 Hz, 0.8 Hz, 1H), 7.36-7.18 (m, 7H), 6.49 (bs, 1H, NH), 5.05 (d, *J* = 3.6 Hz, 1H), 4.49-4.40 (m, 2H), 3.51 (d, *J* = 3.7 Hz, 1H); ¹³**C NMR (CDCl₃, 100 MHz) δ** 171.4, 141.5, 137.4, 131.6, 130.2, 129.6, 128.6, 127.6, 127.5, 125.3, 122.7, 73.4, 43.4; **TLC R_f** 0.08 (EtOAc/hexane, 1/3); **HPLC conditions for 3i :** *t_R* 14.02 min (*R*-isomer), 21.79 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₅H₁₅BrNO₂ : 320.0286 ([M+H]⁺), found : 320.0282 ([M+H]⁺).

N-Benzyl-2-hydroxy-2-(3',4'-dichlorophenyl)-acetamide (3j)



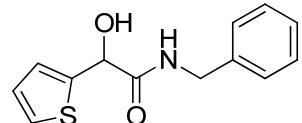
Data for 3j : ¹**H NMR (CDCl₃, 400 MHz) δ** 7.53 (d, *J* = 2.0 Hz, 1H), 7.43 (d, *J* = 8.2 Hz, 1H), 7.35-7.19 (m, 6H), 6.62 (bs, 1H, NH), 5.04 (d, *J* = 3.8 Hz, 1H), 4.47-4.36 (m, 2H), 3.57 (d, *J* = 4.0 Hz, 1H, OH); ¹³**C NMR (CDCl₃, 100 MHz) δ** 171.1, 139.4, 137.3, 132.8, 132.5, 130.5, 128.7, 128.5, 127.7, 127.5, 125.9, 72.8, 43.4; **TLC R_f** 0.08 (hexane/EtOAc, 3/1); **HPLC conditions for 3j :** *t_R* 12.59 min (*R*-isomer), 24.86 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₅H₁₄Cl₂NO₂ : 310.0396 ([M+H]⁺), found : 310.0401 ([M+H]⁺).

N-Benzyl-2-hydroxy-2-furan-2-yl -acetamide (3k)¹



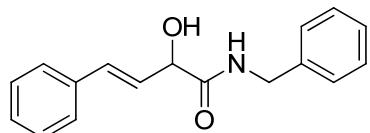
Data for 3k : ¹**H NMR (CDCl₃, 400 MHz) δ** 7.38 (d, *J* = 1.6 Hz, 1H), 7.34-7.22 (m, 5H), 6.70 (bs, 1H, NH), 6.38 (d, *J* = 3.2 Hz, 1H), 6.35 (dd, *J* = 1.8 Hz, 3.2 Hz, 1H), 5.14 (s, 1H), 4.53-4.43 (m, 2H), 3.84 (s, 1H); ¹³**C NMR (CDCl₃, 100 MHz) δ** 169.9, 151.5, 142.9, 137.4, 128.7, 127.6, 127.5, 110.6, 108.7, 67.7, 43.5; **TLC R_f** 0.1 (EtOAc/hexane, 1/2); **HPLC conditions for 3k :** *t_R* 22.06 min (*R*-isomer), 28.14 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₃H₁₄NO₃ : 232.0973 ([M+H]⁺), found : 232.0973 ([M+H]⁺).

N-Benzyl-2-hydroxy-2-thiophen-2-yl -acetamide (3l) ¹



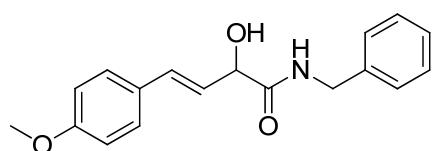
Data for 3l : **¹H NMR (CDCl₃, 400 MHz)** δ 7.34-7.21 (m, 6H), 7.12 (d, J = 3.6 Hz, 1H), 6.98 (dd, J = 3.5 Hz, 5.0 Hz, 1H), 6.53 (bs, 1H, NH), 5.37 (d, J = 4.0 Hz, 1H), 4.53-4.43 (m, 2H), 3.66 (d, J = 4.4 Hz, 1H, OH); **¹³C NMR (CDCl₃, 100 MHz)** δ 171.1, 142.3, 137.4, 128.6, 127.5, 126.8, 126.0, 126.0, 70.1, 43.5; **TLC R_f** 0.075 (EtOAc/hexane, 1/3); **HPLC conditions for 3l :** t_R 18.51 min (*R*-isomer), 27.42 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₃H₁₃NO₂S : 247.0667, found : 247.0667.

2-Hydroxy-4-phenyl-but-3-enoic acid methylamide (3m) ¹



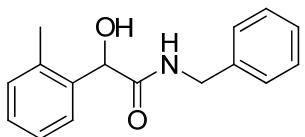
Data for 3m : **¹H NMR (CDCl₃, 400 MHz)** δ 7.40-7.26 (m, 10H), 6.75 (d, J = 15.7 Hz, 1H), 6.54 (bs, 1H, NH), 6.31 (dd, J = 7.0 Hz, 15.8 Hz, 1H), 4.77 (d, J = 6.8 Hz, 1H), 4.55-4.44 (m, 2H), 3.19 (bs, 1H, NH); **¹³C NMR (CDCl₃, 100 MHz)** δ 171.6, 137.6, 135.7, 133.5, 128.7, 128.6, 128.2, 127.7, 127.6, 1267, 126.6, 72.9, 43.6; **TLC R_f** 0.2 (EtOAc/hexane, 2/5); **HPLC conditions for 3m :** t_R 18.40 min (*R*-isomer), 26.94 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₇H₁₇NO₂ : 267.1259, found : 267.1260.

2-Hydroxy-4-(4-methoxy-phenyl)-but-3-enoic acid methylamide (3n)



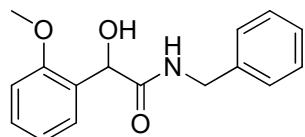
Data for 3n : **¹H NMR (CDCl₃, 400 MHz)** δ 7.36-7.27 (m, 7H), 6.85 (d, J = 8.7 Hz, 1H), 6.69 (d, J = 15.8 Hz, 1H), 6.54 (bs, 1H, NH), 6.15 (dd, J = 7.3 Hz, 15.8 Hz, 1H), 4.73 (dd, J = 3.5 Hz, 7.2 Hz, 1H), 4.54-4.43 (m, 2H), 3.81 (s, 1H), 3.21 (d, J = 3.7 Hz, 1H); **¹³C NMR (CDCl₃, 100 MHz)** δ 171.9, 159.6, 137.6, 133.3, 128.7, 128.5, 127.9, 127.7, 127.6, 124.3, 114.0, 73.1, 55.2, 43.5 ; **TLC R_f** 0.2 (EtOAc/hexane, 1/1); **HPLC conditions for 3n :** t_R 18.32 min (*R*-isomer), 23.28 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 88/12, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₈H₂₀NO₃ : 298.1443 ([M+H]⁺), found : 298.1440 ([M+H]⁺).

N-Benzyl-2-hydroxy-2-*o*-tolyl-acetamide (3o)¹



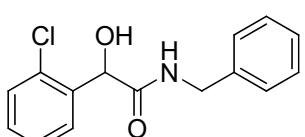
Data for 3o : ¹H NMR (CDCl₃, 400 MHz) δ 7.34-7.27 (m, 4H), 7.24-7.17 (m, 5H), 6.37 (bs, 1H, NH), 5.27 (d, J = 3.0 Hz, 1H), 4.53-4.41 (m, 2H), 3.40 (d, J = 3.0 Hz, 1H), 2.40 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 172.4, 137.7, 137.2, 136.8, 131.1, 128.6, 127.7, 127.6, 127.5, 126.3, 72.2, 43.4, 19.2; TLC R_f 0.16 (EtOAc/hexane, 1/3); HPLC conditions for 3o : t_R 16.11 min (*R*-isomer), 23.57 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 mL/min, λ = 254 nm); HRMS (ESI) Calcd for C₁₆H₁₆NO₂ : 256.1337 ([M+H]⁺), found : 256.1336 ([M+H]⁺).

N-Benzyl-2-hydroxy-2-(2'-methoxyphenyl)-acetamide (3p)¹



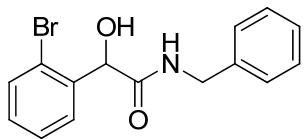
Data for 3p : ¹H NMR (CDCl₃, 400 MHz) δ 7.48 (dd, J = 1.6 Hz, 7.4 Hz, 1H), 7.32-7.25 (m, 4H), 7.14 (d, J = 7.1 Hz, 2H), 7.03 (td, J = 1.6 Hz, 7.5 Hz, 1H), 6.91 (d, J = 8.2 Hz, 1H), 6.90 (bs, 1H, NH), 5.44 (d, J = 5.2 Hz, 1H), 4.53-4.37 (m, 2H), 4.16 (d, J = 5.2 Hz, 1H, OH), 3.81 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 172.7, 155.7, 137.8, 129.2, 128.6, 128.2, 127.4, 127.2, 121.6, 110.8, 68.2, 55.6, 43.6; TLC R_f 0.25 (EtOAc/hexane, 1/2); HPLC conditions for 3p : t_R 19.98 min (*R*-isomer), 25.78 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 88/12, 1.0 mL/min, λ = 254 nm); HRMS (ESI) Calcd for C₁₆H₁₇NO₃ : 271.1208, found : 271.1209.

N-Benzyl-2-hydroxy-2-(2'-chlorophenyl)-acetamide (3q)¹



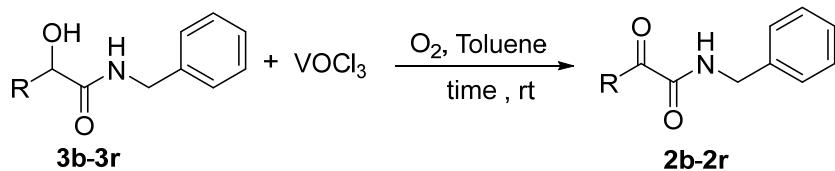
Data for 3q : ¹H NMR (CDCl₃, 400 MHz) δ 7.48 (dd, J = 1.7 Hz, 7.4 Hz, 1H), 7.38 (dd, J = 1.2 Hz, 7.3 Hz, 1H), 7.33-7.26 (m, 5H), 7.18-7.16 (m, 2H), 6.47 (bs, 1H, NH), 5.56 (d, J = 3.6 Hz, H), 4.56-4.40 (m, 2H), 4.03 (d, J = 3.8 Hz, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ 171.5, 137.4, 137.2, 132.6, 129.7, 128.6, 127.5, 127.5, 127.4, 70.2, 43.7; TLC R_f 0.15 (EtOAc/hexane, 1/3); HPLC conditions for 3q : t_R 21.18 min (*R*-isomer), 27.33 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 mL/min, λ = 254 nm); HRMS (ESI) Calcd for C₁₅H₁₅ClNO₂ : 276.0791 ([M+H]⁺), found : 276.0789 ([M+H]⁺).

N-Benzyl-2-hydroxy-2-(2'-bromophenyl)-acetamide (3r)⁴



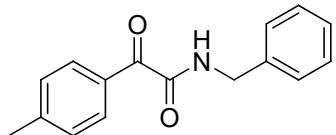
Data for 3r : **¹H NMR (CDCl₃, 400 MHz)** δ 7.55 (dd, *J* = 0.8 Hz, 8.0 Hz, 1H), 7.45 (dd, *J* = 1.7 Hz, 7.7 Hz, 1H), 7.36-7.26 (m, 4H), 7.21-7.16 (m, 3H), 6.56 (bs, 1H, NH), 5.54 (d, *J* = 4.4 Hz, 1H), 4.54-4.39 (m, 2H), 4.13 (d, *J* = 4.4 Hz, 1H, OH); **¹³C NMR (CDCl₃, 100 MHz)** δ 171.4, 138.9, 137.3, 132.9, 130.0, 128.7, 128.2, 127.6, 127.3, 122.8, 72.3, 43.7; TLC R_f 0.15 (EtOAc/hexane, 1/3); **HPLC conditions for 3r :** *t*_R 24.57 min (*R*-isomer), 28.22 min (*S*-isomer) (Chiraldak AD-H, hexane/*i*-PrOH, 92/8, 1.0 mL/min, λ = 254 nm); **HRMS (ESI)** Calcd for C₁₅H₁₄BrNO₂ : 319.0208, found : 319.0207.

N-Benzyl 2-Oxo-Benzeneacetamide Derivatives (2b-r)



In a 100 ml double-neck round bottomed flask was placed respective *N*-benzyl, α -hydroxy-amide derivative (3.0 mmol, 1.0 eq.) in oxygen saturated toluene (30 mL). VOCl_3 (Vanadium(V) oxychloride, 28 l, 0.3 mmol, 0.1 eq.) was added slowly to the reaction mixture and stirred at room temperature. After the completion of reaction, solvent was evaporated under reduced pressure and the crude mixture directly loaded on column chromatography. Ethyl acetate/hexane, 1/10 mixture was used as eluents to get the product and recrystallized from hot diethyl ether to get *N*-benzyl- α - keto-amide derivatives.

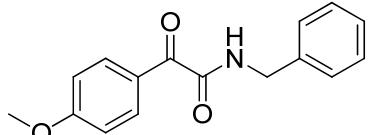
***N*-Benzyl-2-oxo-2-p-tolyl-acetamide (2b)¹**



Data for 2b : ^1H NMR (CDCl_3 , 400 MHz) δ 8.29 (d, $J = 8.0$ Hz, 2H), 7.40 (bs, 1H, NH), 7.38-7.27 (m, 7H), 4.56 (d, $J = 6.0$ Hz, 2H), 2.43 (s, 1H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 187.0, 161.8, 145.6, 137.1, 131.3, 130.7, 129.1, 128.7, 127.7, 127.6, 43.3, 21.8; TLC R_f

0.2 (EtOAc/hexane, 1/10); HRMS (ESI) Calcd for $\text{C}_{16}\text{H}_{15}\text{NO}_2$: 253.1103, found : 253.1102.

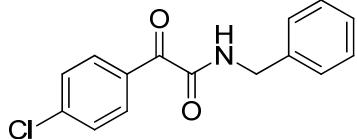
***N*-Benzyl-2-oxo-2-(4'-methoxyphenyl)acetamide (2c)¹**



Data for 2c : ^1H NMR (CDCl_3 , 400 MHz) δ 8.43 (d, $J = 9.2$ Hz, 2H), 7.38 (bs, 1H, NH), 7.37-7.29 (m, 5H), 6.94 (d, $J = 9.2$ Hz, 2H), 4.56 (d, $J = 6.4$ Hz, 2H), 3.89 (s, 1H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 185.4, 164.6, 162.0, 137.2, 133.9, 128.7, 127.8, 127.7,

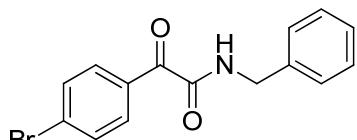
126.3, 113.8, 55.5, 43.3; TLC R_f 0.36 (EtOAc/hexane, 3/1); HRMS (ESI) Calcd for $\text{C}_{16}\text{H}_{15}\text{NO}_3$: 269.1052, found : 269.1050 ($[\text{M}+\text{H}]^+$).

N-Benzyl-2-oxo-2-(4'-chlorophenyl)-acetamide (2d)¹



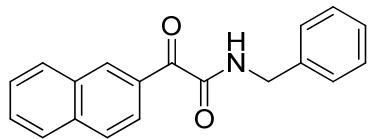
Data for 2d : **¹H NMR (CDCl₃, 400 MHz)** δ 8.36 (d, *J* = 8.5 Hz, 2H), 7.46 (d, *J* = 8.5 Hz, 2H), 7.42 (bs, 1H, NH), 4.56 (d, *J* = 6.0 Hz, 2H); **¹³C NMR (CDCl₃, 100 MHz)** δ 186.0, 161.1, 141.1, 136.9, 132.6, 131.5, 128.7, 127.7, 43.4; **TLC R_f** 0.525 (EtOAc/hexane, 1/3); **HRMS (ESI)** Calcd for C₁₅H₁₂ClNO₂ : 273.0557, found : 273.0556([M+H]⁺).

N-Benzyl-2-oxo-2-(4'-bromophenyl)-acetamide (2e)¹



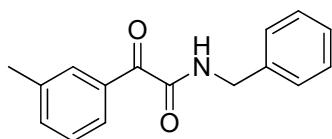
Data for 2e : **¹H NMR (CDCl₃, 400 MHz)** δ 8.28 (d, *J* = 8.8 Hz, 2H), 7.63 (d, *J* = 8.4 Hz, 2H), 7.41 (bs, 1H, NH), 7.41-7.30 (m, 5H), 4.56 (d, *J* = 6.0 Hz, 2H); **¹³C NMR (CDCl₃, 100 MHz)** δ 186.3, 161.0, 136.9, 132.7, 132.0, 131.8, 130.1, 128.8, 127.8, 43.5; **TLC R_f** 0.15 (EtOAc/hexane, 1/10); **HRMS (ESI)** Calcd for C₁₅H₁₂BrNO₂ : 317.0051 found : 317.0051.

N-Benzyl-2-oxo-2-naphthalen-2-yl-acetamide (2f)¹



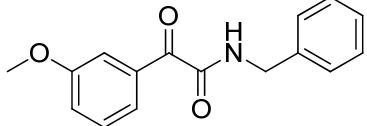
Data for 2f : **¹H NMR (CDCl₃, 400 MHz)** δ 9.25 (s, 1H), 8.21 (dd, *J* = 1.6 Hz, 8.6 Hz, 1H), 8.02 (d, *J* = 8.4 Hz, 1H), 7.90 (d, *J* = 8.8 Hz, 1H), 7.87 (d, *J* = 8.0 Hz, 1H), 7.64 (ddd, *J* = 1.2 Hz, 6.0 Hz, 7.0 Hz, 1H), 7.56 (ddd, *J* = 1.2 Hz, 6.0 Hz, 7.0 Hz, 1H), 7.55 (bs, 1H, NH), 7.40-7.30 (m, 5H), 4.62 (d, *J* = 6.0 Hz, 2H); **¹³C NMR (CDCl₃, 100 MHz)** δ 187.0, 161.7, 137.1, 136.1, 135.0, 132.3, 130.5, 130.3, 129.3, 128.8, 128.3, 127.8, 127.8, 127.7, 126.8, 125.2, 43.5; **TLC R_f** 0.44 (EtOAc/hexane, 1/3); **HRMS (ESI)** Calcd for C₁₉H₁₅NO₂ : 289.1103, found : 289.1103.

N-Benzyl-2-oxo-2-m-tolyl-acetamide (2g)⁴



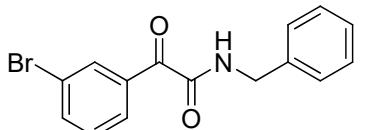
Data for 2g : **¹H NMR (CDCl₃, 400 MHz)** δ 8.18 (s, 1H), 8.16 (s, 1H), 7.44 (d, *J* = 7.6 Hz, 1H), 7.39-7.28 (m, 7H), 4.57 (d, *J* = 6.0 Hz, 2H), 2.42 (s, 1H); **¹³C NMR (CDCl₃, 100 MHz)** δ 187.7, 161.6, 138.3, 137.1, 135.2, 133.3, 131.5, 128.8, 128.4, 128.4, 127.8, 127.8, 43.4, 21.3; **TLC R_f** 0.2 (EtOAc/hexane, 10/1); **HRMS (ESI)** Calcd for C₁₆H₁₆NO₂ : 254.1181 ([M+H]⁺), found : 254.1178 ([M+H]⁺).

N-Benzyl-2-oxo-2-(3'-methoxyphenyl)-acetamide (2h)



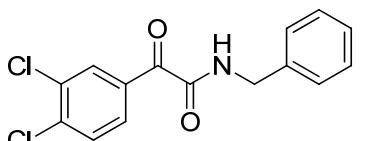
Data for 2h : ^1H NMR (CDCl_3 , 400 MHz) δ 8.02 (dt, $J = 7.6$ Hz, 1.2 Hz, 1H), 7.86 (dd, $J = 1.5$ Hz, 2.6 Hz, 1H), 7.42-7.31 (m, 7H), 7.18 (ddd, $J = 8.2$ Hz, 2.7 Hz, 0.9 Hz, 1H), 4.57 (d, $J = 6.0$ Hz, 2H), 3.86 (s, 3H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 187.2, 161.5, 159.5, 137.0, 134.4, 129.5, 128.8, 127.8, 124.1, 121.5, 114.7, 55.4, 43.4; TLC R_f 0.22 (EtOAc/hexane, 1/6); HRMS (ESI) Calcd for $\text{C}_{16}\text{H}_{16}\text{NO}_3$: 270.1130 ([M+H] $^+$), found : 270.1133 ([M+H] $^+$).

N-Benzyl-2-oxo-2-(3'-bromophenyl)-acetamide (2i)⁶



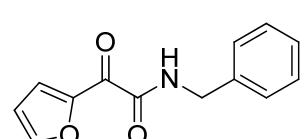
Data for 2i : ^1H NMR (CDCl_3 , 400 MHz) δ 8.51 (s, 1H), 8.34 (dd, $J = 0.8$ Hz, 7.6 Hz, 1H), 7.75 (td, $J = 1.0$ Hz, 7.8 Hz, 1H), 7.39-7.27 (m, 7H, NH), 4.57 (d, $J = 6.0$ Hz, 2H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 186.0, 160.8, 137.2, 136.8, 134.9, 133.9, 130.0, 129.8, 128.8, 127.9, 122.6, 43.5; TLC R_f 0.5 (EtOAc/hexane, 1/3); HRMS (ESI) Calcd for $\text{C}_{15}\text{H}_{13}\text{BrNO}_2$: 318.0127 ([M+H] $^+$), found : 318.0127 ([M+H] $^+$).

N-Benzyl-2-oxo-2-(3',4'-dichlorophenyl)-acetamide (2j)



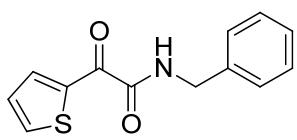
Data for 2j : ^1H NMR (CDCl_3 , 400 MHz) δ 8.54 (d, $J = 1.9$ Hz, 1H), 8.27 (dd, $J = 2.0$ Hz, 8.4 Hz, 1H), 7.56 (d, $J = 8.4$ Hz, 1H), 7.43 (bs, 1H, NH), 7.39-7.29 (m, 5H), 4.56 (d, $J = 6.0$ Hz, 2H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 184.9, 160.6, 139.3, 136.7, 133.1, 133.0, 132.7, 130.6, 130.3, 128.8, 127.9, 127.8, 43.5; TLC R_f 0.35 (EtOAc/hexane, 1/6); HRMS (ESI) Calcd for $\text{C}_{15}\text{H}_{12}\text{Cl}_2\text{NO}_2$: 308.0245 ([M+H] $^+$), found : 308.0241 ([M+H] $^+$).

N-Benzyl-2-oxo-2-furan-2-yl -acetamide (2k)¹



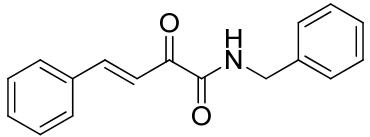
Data for 2k : ^1H NMR (CDCl_3 , 400 MHz) δ 8.22 (d, $J = 3.5$ Hz, 1H), 7.76 (d, $J = 3.5$ Hz, 1H), 7.60 (bs, 1H, NH), 7.37-7.30 (m, 5H), 6.63 (dd, $J = 1.6$ Hz, 3.7 Hz, 1H), 4.54 (d, $J = 6.0$ Hz, 2H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 173.4, 159.9, 149.4, 136.8, 128.8, 127.8, 126.9, 113.1, 43.3; TLC R_f 0.45 (EtOAc/hexane, 1/2); HRMS (ESI) Calcd for $\text{C}_{13}\text{H}_{11}\text{NO}_3$: 229.0739, found : 229.0739.

N-Benzyl-2-oxo-2-thiophen-2-yl -acetamide (2l) ¹



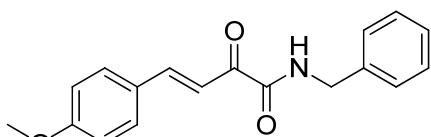
Data for 2l : ¹**H NMR (CDCl₃, 400 MHz)** δ 8.42 (dd, *J* = 1.0 Hz, 3.5 Hz, 1H), 7.84 (dd, *J* = 1.1 Hz, 4.8 Hz, 1H), 7.62 (bs, 1H, NH), 7.38-7.28 (m, 5H), 7.19 (dd, *J* = 4.7 Hz, 4.0 Hz, 1H), 4.56 (d, *J* = 6.0 Hz, 2H); ¹³C NMR (CDCl₃, 100 MHz) δ 178.1, 160.5, 138.6, 138.0, 136.9, 136.7, 128.7, 128.1, 127.8, 127.7, 43.4; TLC R_f 0.44 (EtOAc/hexane, 1/3); HRMS (ESI) Calcd for C₁₃H₁₁NO₂S : 245.0510, found : 245.0509.

2-Oxo-4-phenyl-but-3-enoic acid methylamide (2m) ¹



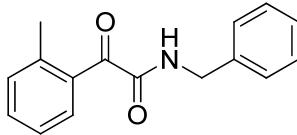
Data for 2m : ¹**H NMR (CDCl₃, 400 MHz)** δ 7.96 (d, *J* = 16.4 Hz, 1H), 7.81 (d, *J* = 16.0 Hz, 1H), 7.68 (dd, *J* = 1.6 Hz, 7.4 Hz, 2H), 7.49 (bs, 1H, NH), 7.47-7.28 (m, 9H), 4.54 (d, *J* = 6.0 Hz, 2H); ¹³C NMR (CDCl₃, 100 MHz) δ 185.3, 161.1, 148.1, 137.0, 134.3, 131.4, 129.1, 129.0, 128.8, 127.8, 127.8, 118.6, 43.5; TLC R_f 0.23 (EtOAc/hexane, 1/10); HRMS (ESI) Calcd for C₁₇H₁₅NO₂ : 265.1103, found : 265.1103.

2-oxo-4-(4-methoxy-phenyl)-but-3-enoic acid methylamide (2n)



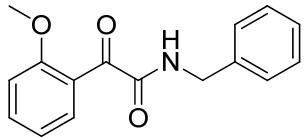
Data for 2n : ¹**H NMR (CDCl₃, 400 MHz)** δ 7.93 (d, *J* = 16.0 Hz, 1H), 7.68 (d, *J* = 16.0 Hz, 1H), 7.64 (dd, *J* = 7.1 Hz, 1.9 Hz, 2H), 7.51 (bs, 1H, NH), 7.37-7.30 (m, 5H), 6.93 (dd, *J* = 6.9 Hz, 1.9 Hz, 2H), 4.54 (d, *J* = 6.1 Hz, 2H), 3.86 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 185.0, 162.5, 161.4, 148.0, 137.1, 131.1, 128.8, 127.8, 127.7, 127.2, 116.5, 114.5, 55.4, 43.5; TLC R_f 0.15 (EtOAc/hexane, 1/6); HRMS (ESI) Calcd for C₁₈H₁₈NO₃ : 296.1286, found : 296.1286 ([M+H]⁺).

N-Benzyl-2-oxo-2-o-tolyl-acetamide (2o) ¹



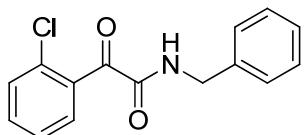
Data for 2o : ¹**H NMR (CDCl₃, 400 MHz)** δ 7.97 (d, *J* = 7.8 Hz, 1H), 7.45 (td, *J* = 1.3 Hz, 8.7 Hz, 1H), 7.39-7.27 (m, 8H), 4.57 (d, *J* = 6.0 Hz, 2H), 2.50 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 191.0, 161.8, 140.1, 137.1, 132.7, 132.6, 132.0, 131.6, 128.8, 127.8, 127.7, 125.3, 43.5, 20.8; TLC R_f 0.45 (EtOAc/hexane, 1/3); HRMS (ESI) Calcd for C₁₆H₁₅NO₂ : 253.1103, found : 253.1102 ([M+H]⁺).

N-Benzyl-2-oxo-2-(2'-methoxyphenyl)-acetamide (2p)¹



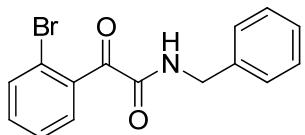
Data for 2p : ¹**H NMR (CDCl₃, 400 MHz)** δ 7.68 (dd, *J* = 2.0 Hz, 7.6 Hz, 2H), 7.41 (ddd, *J* = 1.6 Hz, 7.4 Hz, 8.5 Hz, 1H), 7.38-7.28 (m, 5H), 7.03 (td, *J* = 0.8 Hz, 7.6 Hz, 1H), 6.97 (d, *J* = 8.0 Hz, 2H), 4.56 (d, *J* = 6.0 Hz, 2H), 3.79 (s, 3H); ¹³**C NMR (CDCl₃, 100 MHz)** δ 191.4, 163.0, 159.5, 137.4, 134.6, 131.0, 128.7, 127.8, 127.6, 124.6, 120.7, 111.9, 56.0, 43.3; **TLC R_f** 0.35 (EtOAc/hexane, 1/2); **HRMS (ESI)** Calcd for C₁₆H₁₅NO₃ : 269.1052, found : 269.1050.

N-Benzyl-2-oxo-2-(2'-chlorophenyl)-acetamide (2q)¹



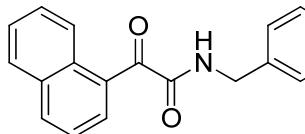
Data for 2q : ¹**H NMR (CDCl₃, 400 MHz)** δ 7.71-7.69 (m, 1H), 7.50-7.44 (m, 3H), 7.39-7.32 (m, 6H, NH), 4.58 (d, *J* = 6.0 Hz, 2H); ¹³**C NMR (CDCl₃, 100 MHz)** δ 189.9, 160.5, 136.8, 133.9, 133.0, 131.2, 130.3, 128.7, 127.8, 127.8, 126.5, 43.6; **TLC R_f** 0.4 (EtOAc/hexane, 1/3); **HRMS (ESI)** Calcd for C₁₅H₁₂ClNO₂ : 273.0557, found : 273.0557.

N-Benzyl-2-oxo-2-(2'-bromophenyl)-acetamide (2r)⁴



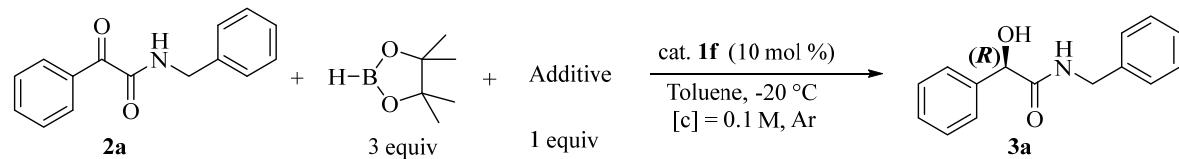
Data for 2r : ¹**H NMR (CDCl₃, 400 MHz)** δ 7.66-7.63 (m, 2H), 7.44-7.30 (m, 8H), 4.57 (d, *J* = 6.4 Hz, 2H); ¹³**C NMR (CDCl₃, 100 MHz)** δ 190.5, 160.1, 136.8, 135.9, 133.45, 132.9, 131.1, 127.9, 127.8, 127.0, 120.8, 43.7; **TLC R_f** 0.4 (EtOAc/hexane, 1/3); **HRMS (ESI)** Calcd for C₁₅H₁₂BrNO₂ : 317.0051, found : 317.0052.

N-Benzyl-2-oxo-2-naphthalen-1-yl -acetamide (2s)¹



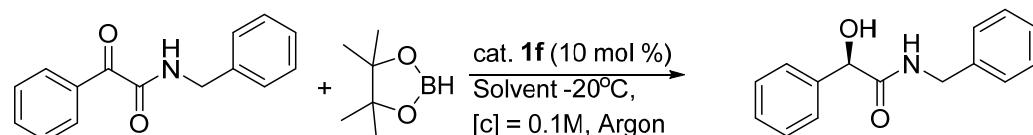
Data for 2s : ¹**H NMR (CDCl₃, 400 MHz)** δ 8.58 (d, *J* = 8.4 Hz, 1H), 8.36 (dd, *J* = 1.2 Hz, 7.2 Hz, 1H), 8.08 (d, *J* = 8.4 Hz, 1H), 7.91 (dd, *J* = 0.8 Hz, 8.8 Hz, 1H), 7.64-7.53 (m, 3H), 7.52 (bs, 1H, NH), 7.44-7.31 (m, 5H), 4.63 (d, *J* = 6 Hz, 2H); ¹³**C NMR (CDCl₃, 100 MHz)** δ 190.1, 161.9, 137.1, 134.6, 133.8, 133.2, 131.1, 129.5, 128.8, 128.6, 128.4, 127.9, 127.8, 126.5, 124.2, 43.7; **TLC R_f** 0.36 (EtOAc/hexane, 1/4); **HRMS (ESI)** Calcd for C₁₉H₁₅NO₂ : 289.1103, found : 289.1101.

6. Table S1: Effect of additives on the asymmetric reduction.



Entry	Reducing Agent	Additive	Time (h)	Yield (%)	Ee (%)
1	PinB–H	—	96	65	88 (<i>R</i>)
2	PinB–H	MeOH	96	40	99 (<i>R</i>)
3	PinB–H	CCl ₃ CH ₂ OH	96	47	91 (<i>R</i>)
4	PinB–H	CF ₃ CH ₂ OH	96	22	96 (<i>R</i>)
5	CatB–H	—	14	99	64 (<i>S</i>)
6	CatB–H	MeOH	14	92	47 (<i>S</i>)
7	CatB–H	CCl ₃ CH ₂ OH	14	86	58 (<i>S</i>)
8	CatB–H	CF ₃ CH ₂ OH	14	84	62 (<i>S</i>)

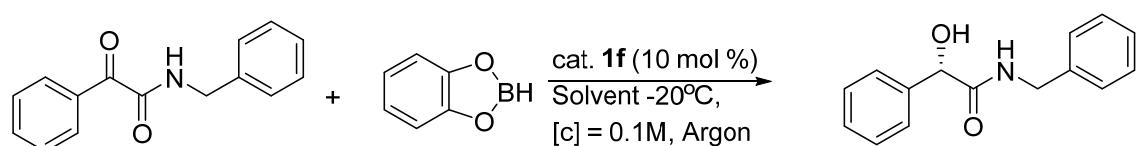
Table S2: Solvent Screening with HBPin:



entry	Solvent	Time(h)	Yield(%)	ee%
1 (final)	toluene	24+24+48	65	88 (<i>R</i>)
2	MeOH	24+24+48	35	52 (<i>R</i>)
3	<i>i</i> -PrOH	24+24+48	43	56 (<i>R</i>)
4	1,4-dioxane	24+24+48	21	36 (<i>R</i>)

5	CH ₃ CN	24+24+48	28	38 (<i>R</i>)
6	DMSO (r.t.)	24+24+48	52	12 (<i>R</i>)
7	DMF	24+24+48	19	24 (<i>R</i>)

Table S3: Solvent Screening with HBCat:



entry	Solvent	Time(h)	Yield(%)	ee%
1 (final)	toluene	12	99	64 (<i>S</i>)
2	CH ₂ Cl ₂	12	97	2 (<i>R</i>)
3	PhCl	14	95	57 (<i>S</i>)
4	THF	12	98	35 (<i>S</i>)
5	<i>t</i> -BuOMe	15	87	4 (<i>R</i>)

Table S4

 mw: 237.26	Reducing Agent	cat. 1f (10 mol %) toluene, temp [c] = 0.1 M, Argon	 mw: 239.27		
				reducing agent	temperature, °C
				Yield, %	ee, %

HBPin	-20/-40	99/0	0/---
HBCat	-20/-40	92/28	2/8 (S)

7. X-ray data of Catalyst 1f:

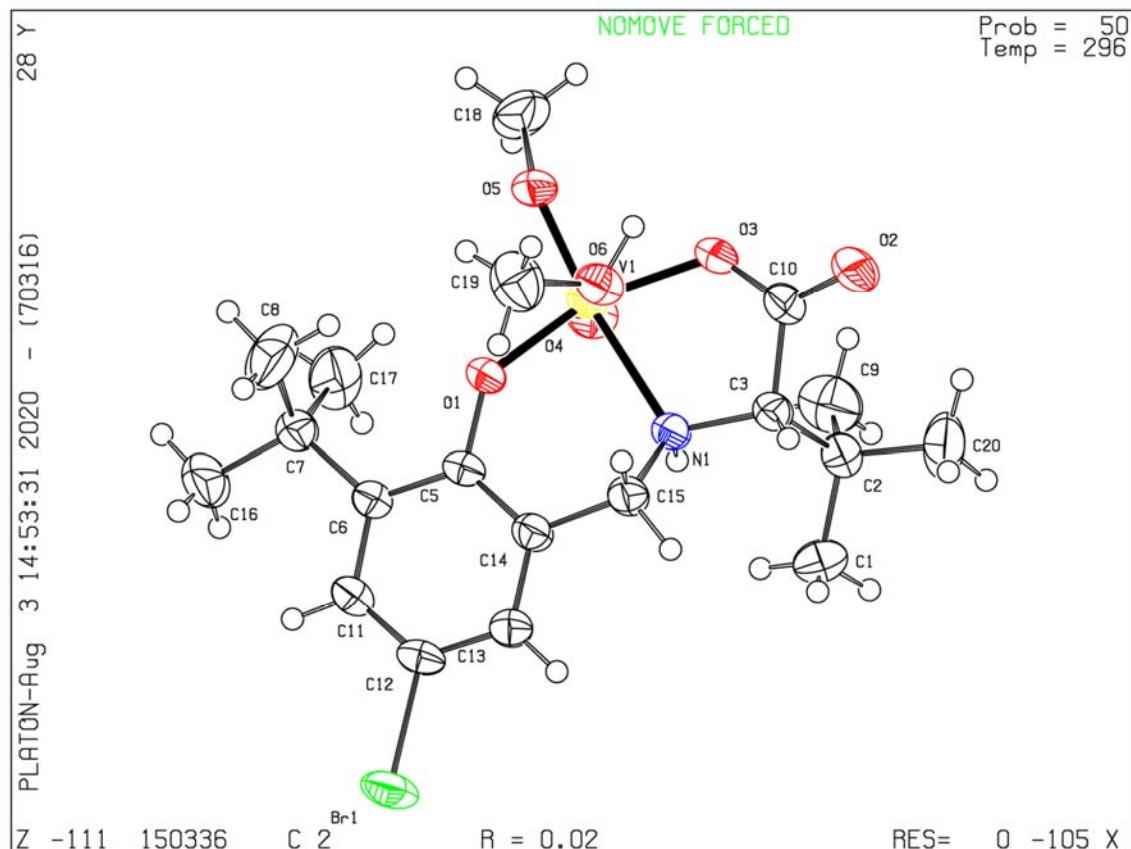


Figure S1. X-ray ORTEP drawing of catalyst **1f** (thermal ellipsoid is scaled to the 50% probability level).

Table 1. Crystal data and structure refinement for 150336.

Identification code	150336
Empirical formula	C ₁₉ H ₃₁ BrN O ₆ V
Formula weight	500.30
Temperature	296(2) K
Wavelength	0.71073 Å

Crystal system	Monoclinic	
Space group	C 2	
Unit cell dimensions	$a = 27.3247(8)$ Å	$\alpha = 90^\circ$.
	$b = 7.3295(2)$ Å	$\beta = 102.472(2)^\circ$.
	$c = 11.7690(3)$ Å	$\gamma = 90^\circ$.
Volume	$2301.43(11)$ Å ³	
Z	4	
Density (calculated)	1.444 Mg/m ³	
Absorption coefficient	2.199 mm ⁻¹	
F(000)	1032	
Crystal size	0.15 x 0.15 x 0.12 mm ³	
Theta range for data collection	1.526 to 26.394°.	
Index ranges	$-34 \leq h \leq 34$, $-9 \leq k \leq 9$, $-14 \leq l \leq 14$	
Reflections collected	10084	
Independent reflections	4260 [R(int) = 0.0237]	
Completeness to theta = 25.242°	99.9 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.9485 and 0.742	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	4260 / 1 / 262	
Goodness-of-fit on F ²	0.558	
Final R indices [I>2sigma(I)]	R1 = 0.0249, wR2 = 0.0711	
R indices (all data)	R1 = 0.0275, wR2 = 0.0762	
Absolute structure parameter	0.013(10)	
Extinction coefficient	n/a	
Largest diff. peak and hole	0.561 and -0.339 e.Å ⁻³	

Datablock: 150336

Bond precision: C-C = 0.0058 Å Wavelength=0.71073
Cell: a=27.3247(8) b=7.3295(2) c=11.7690(3)
alpha=90 beta=102.472(2) gamma=90

Temperature: 296 K

	Calculated	Reported
Volume	2301.43(11)	2301.43(11)
Space group	C 2	C 2
Hall group	C 2y	C 2y
Moiety formula	C19 H31 Br N O6 V	?
Sum formula	C19 H31 Br N O6 V	C19 H31 Br N O6 V
Mr	500.29	500.30
Dx,g cm ⁻³	1.444	1.444
Z	4	4
Mu (mm ⁻¹)	2.199	2.199
F000	1032.0	1032.0
F000'	1032.50	
h,k,lmax	34,9,14	34,9,14
Nref	4727[2556]	4260
Tmin,Tmax	0.726,0.768	0.742,0.948
Tmin'	0.712	
Correction method=	# Reported T Limits: Tmin=0.742	
Tmax=0.948	AbsCorr = MULTI-SCAN	
Data completeness=	1.67/0.90	Theta(max)= 26.394
R(reflections)=	0.0249(3926)	wR2(reflections)= 0.0762(4260)
S =	0.558	Npar= 262

The following ALERTS were generated. Each ALERT has the format
test-name_ALERT_alert-type_alert-level.

Click on the hyperlinks for more details of the test.

🟡 Alert level B

[GOODF01_ALERT_2_B](#) The least squares goodness of fit parameter lies outside the range 0.60 <> 4.00
Goodness of fit given = 0.558

🟢 Alert level C

[PLAT242_ALERT_2_C](#) Low 'MainMol' Ueq as Compared to Neighbors of C2
Check
[PLAT242_ALERT_2_C](#) Low 'MainMol' Ueq as Compared to Neighbors of C7
Check
[PLAT420_ALERT_2_C](#) D-H Without Acceptor N1 --H1 . Please
Check
[PLAT911_ALERT_3_C](#) Missing FCF Refl Between Thmin & STh/L= 0.600 2
Report
[PLAT915_ALERT_3_C](#) No Flack x Check Done: Low Friedel Pair Coverage 79
%

PLAT915_ALERT_3_C	No Flack x Check Done: Low Friedel Pair Coverage %	79
PLAT918_ALERT_3_C	Reflection(s) with I(obs) much Smaller I(calc) . Check	1
PLAT939_ALERT_3_C	Large Value of Not (SHELXL) Weight Optimized S . Check	15.00

• Alert level G

PLAT007_ALERT_5_G	Number of Unrefined Donor-H Atoms	2
Report		
PLAT128_ALERT_4_G	Alternate Setting for Input Space Group C2 Note	I2
PLAT169_ALERT_4_G	The CIF-Embedded .res File Contains AFIX 1 Recds Report	1
PLAT791_ALERT_4_G	Model has Chirality at N1 (Sohnke SpGr) Verify	R
PLAT791_ALERT_4_G	Model has Chirality at C3 (Sohnke SpGr) Verify	S
PLAT794_ALERT_5_G	Tentative Bond Valency for V1 (V) . Info	5.18
PLAT883_ALERT_1_G	No Info/Value for _atom_sites_solution_primary . Do !	Please
PLAT912_ALERT_4_G	Missing # of FCF Reflections Above STh/L= 0.600 Note	8
PLAT913_ALERT_3_G	Missing # of Very Strong Reflections in FCF Note	1
PLAT941_ALERT_3_G	Average HKL Measurement Multiplicity Low	4.0
PLAT965_ALERT_2_G	The SHELXL WEIGHT Optimisation has not Converged Check	Please
PLAT978_ALERT_2_G	Number C-C Bonds with Positive Residual Density. Info	4

0 **ALERT level A** = Most likely a serious problem - resolve or explain
 1 **ALERT level B** = A potentially serious problem, consider carefully
 8 **ALERT level C** = Check. Ensure it is not caused by an omission or oversight
 12 **ALERT level G** = General information/check it is not something unexpected

 1 ALERT type 1 CIF construction/syntax error, inconsistent or missing data
 6 ALERT type 2 Indicator that the structure model may be wrong or deficient
 7 ALERT type 3 Indicator that the structure quality may be low
 5 ALERT type 4 Improvement, methodology, query or suggestion
 2 ALERT type 5 Informative message, check

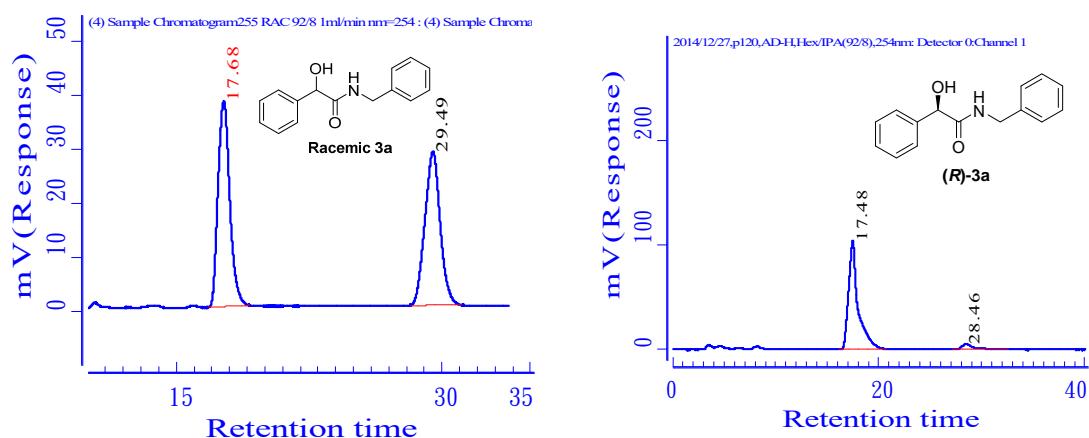
8. References

1. S. S. Weng, M. W. Shen, J. Q. Kao, Y. S. Munot, C. T. Chen, Chiral N-Salicylidene Vanadyl Carboxylate-Catalyzed Enantioselective Aerobic Oxidation of α -Hydroxy Esters and Amides, *Proc. Natl. Acad. Sci. U.S.A.* **2006**, *103*, 3522-3527.
2. S. E.; Denmark, Y. Fan, The First Catalytic, Asymmetric α -Additions of Isocyanides. Lewis-Base-Catalyzed, Enantioselective Passerini-Type Reactions., *J. Am. Chem. Soc.* **2003**, *125*, 7825-

7827.

3. I. Shin, M. R. Lee, J. Lee, M. Jung, W. Lee and J. Yoon, Synthesis of Optically Active Phthaloyl D-Aminoxy Acids from L-Amino Acids or L-Hydroxy Acids as Building Blocks for the Preparation of Aminoxy Peptides, *J. Org. Chem.* **2000**, *65*, 7667.
4. G. Gu, T. Yang, O. Yu, H. Qian, J. Wang, J. Wen, L. Dang and X. Zhang, Enantioselective Iridium-Catalyzed Hydrogenation of α -Keto Amides to α -Hydroxy Amides, *Org. Lett.*, 2017, **19**, 5920–5923.
5. Y.-K. Zhang and B. Wang, European, Synthesis of α -Ketoamides from β -Ketonitriles and Primary Amines: A Catalyst-Free Oxidative Decyanation–Amidation Reaction, *J. Org. Chem.*, 2019, **2019**, 5732–5735.
6. Q. W. Tan, P. Chovatia and M. C. Willis, Copper-Catalysed Synthesis of Alkylidene 2-Pyrrolinone Derivatives from the Combination of α -Keto Amides and Alkynes, *Org. Biomol. Chem.*, 2018, **16**, 7797–7800.

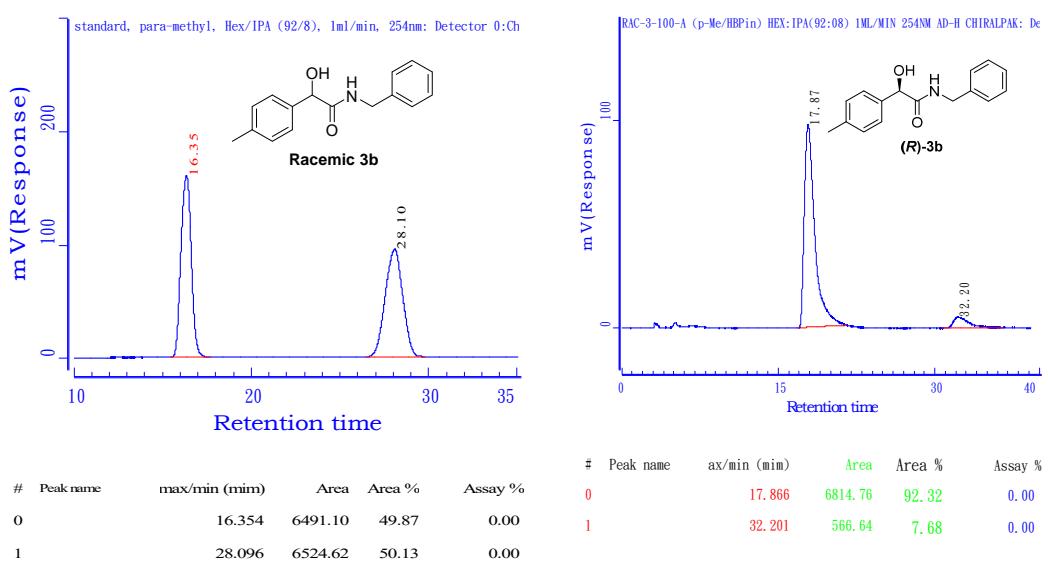
9. HPLC chromatograms:



#	Peak name	x/min (min)	Area	Area %	Assay %
0		17.682	1761.21	50.72	0.00
1		29.485	1711.11	49.28	0.00

#	Peak name	x/min (min)	Area	Area %	Assay %
0		17.484	6668.31	94.05	0.00
1		28.462	421.77	5.95	0.00

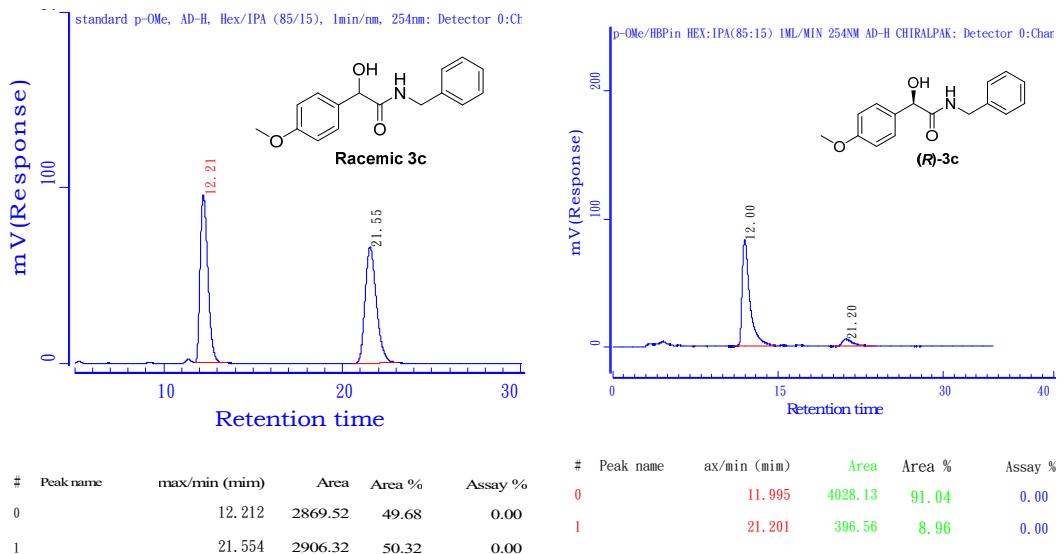
HPLC conditions for **3a** : t_R 17.68 min (major **R**-isomer), 29.49 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, $\lambda = 254$ nm) for 88% ee (**R**)



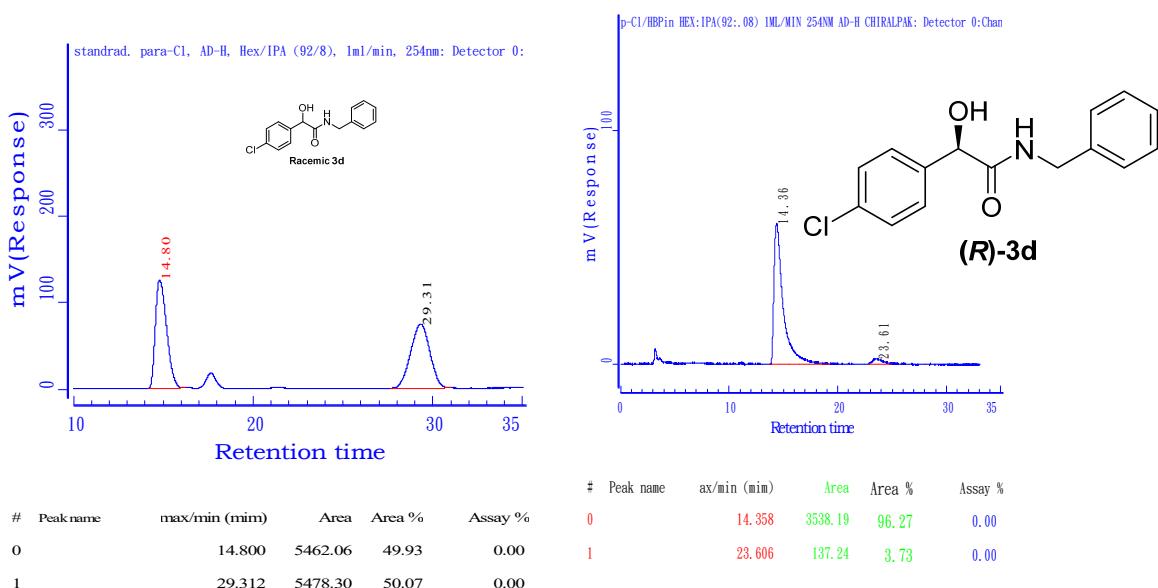
#	Peak name	max/min (min)	Area	Area %	Assay %
0		16.354	6491.10	49.87	0.00
1		28.096	6524.62	50.13	0.00

#	Peak name	x/min (min)	Area	Area %	Assay %
0		17.866	6814.76	92.32	0.00
1		32.201	566.64	7.68	0.00

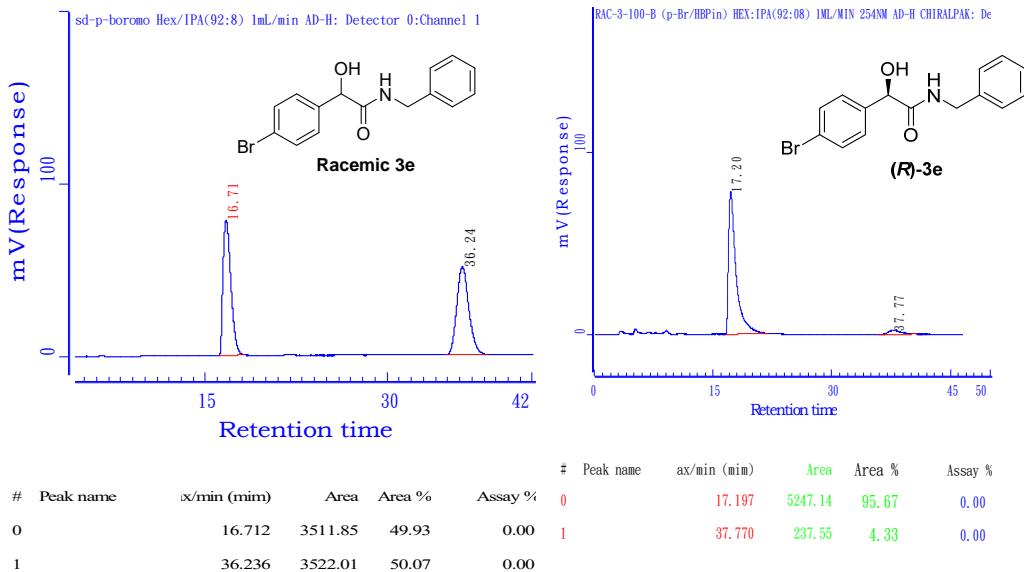
HPLC conditions for **3b** : t_R 16.35 min (major **R**-isomer), 28.10 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, $\lambda = 254$ nm) for 85% ee (**R**).



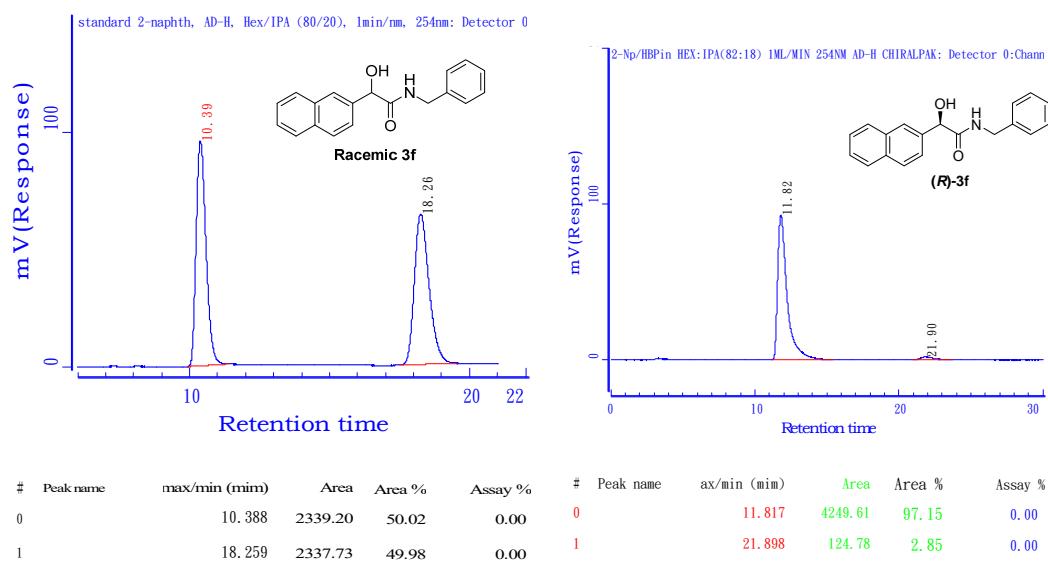
HPLC conditions for **3c** : t_R 12.21 min (major **R**-iosmer), 21.55 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 85/15, 1.0 ml/min, $\lambda = 254$ nm) for 82% ee (**R**).



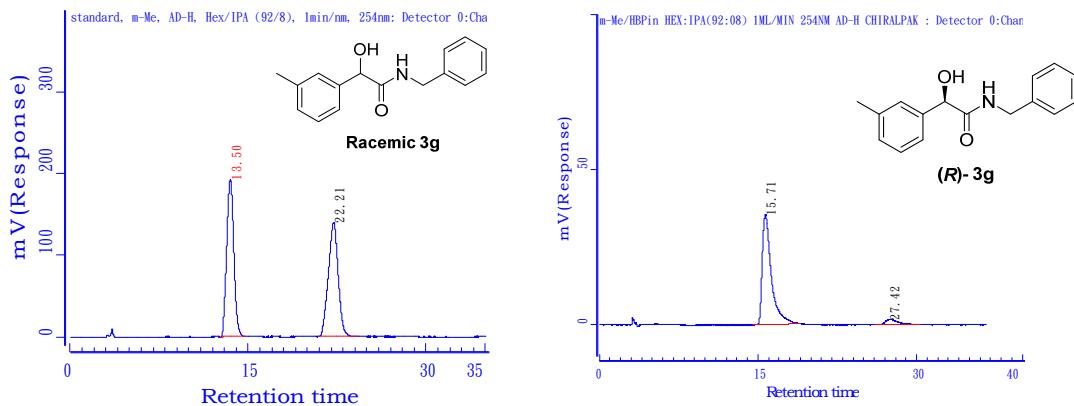
HPLC conditions for **3d** : t_R 14.30 min (major **R**-iosmer), 29.31 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, $\lambda = 254$ nm) for 93% ee (**R**).



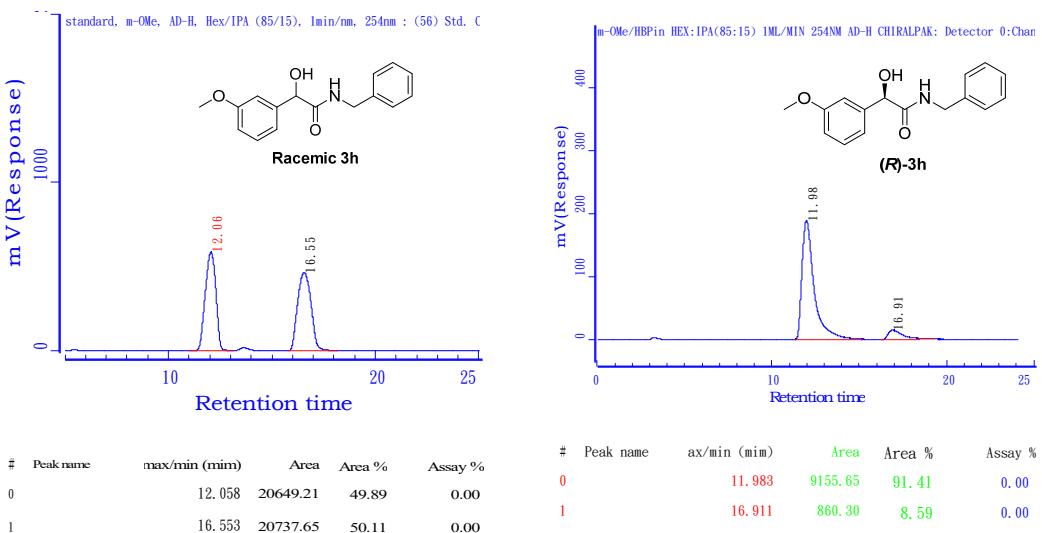
HPLC conditions for **3e** : t_R 16.71 min (major **R**-iosmer), 36.24 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, $\lambda = 254$ nm) for 91% ee (**R**).



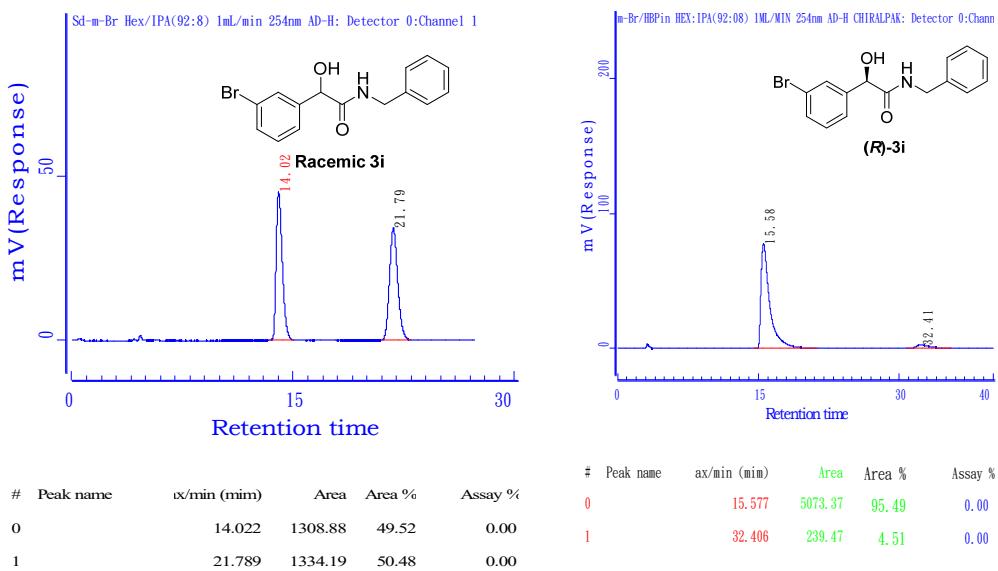
HPLC conditions for **3f** : t_R 10.39 min (major **R**-iosmer), 18.26 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 80/20, 1.0 ml/min, $\lambda = 254$ nm) for 94% ee(**R**).



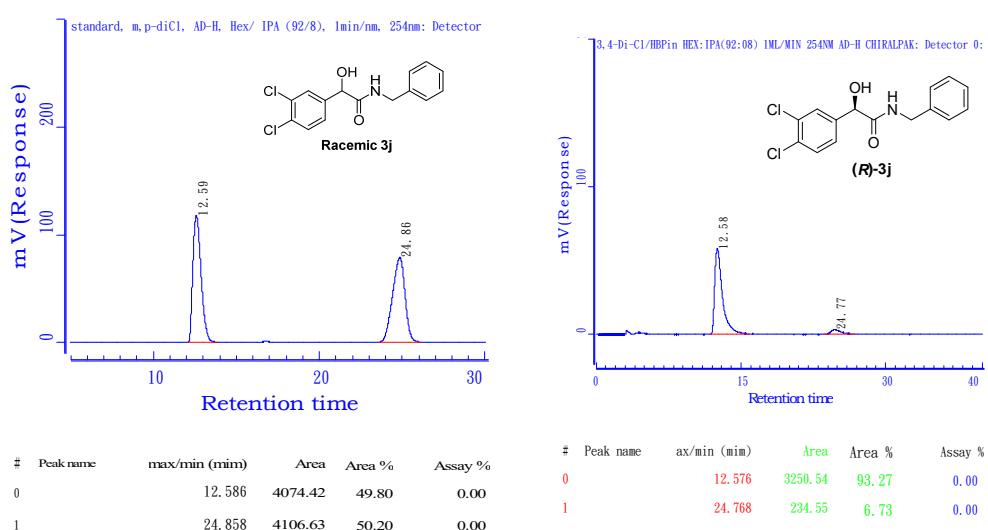
HPLC conditions for **3g** : t_R 13.50 min (major **R**-isomer), 22.21 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 86% ee (**R**).



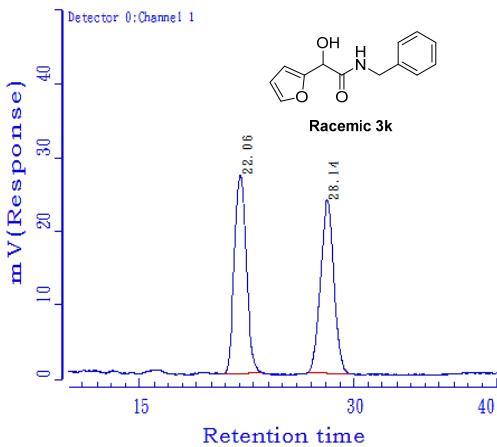
HPLC conditions for **3h** : t_R 12.06 min (major **R**-isomer), 16.55 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 85/15, 1.0 ml/min, λ = 254 nm) for 83% ee (**R**).



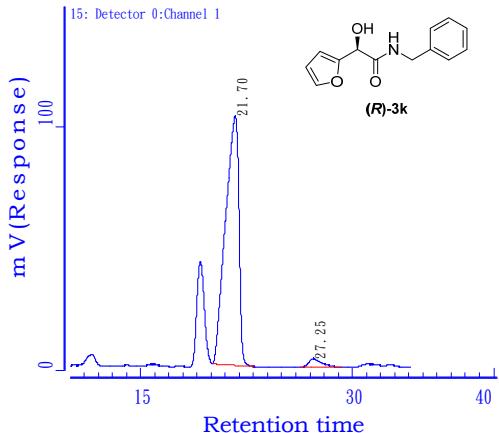
HPLC conditions for **3i** : t_R 14.02 min (major **R**-iosmer), 21.79 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 91% ee (**R**).



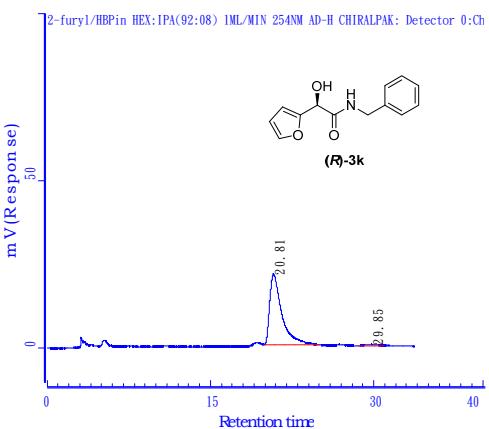
HPLC conditions for **3j** : t_R 12.59 min (major **R**-iosmer), 24.86 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 87% ee.



#	Peak name	x/min (min)	Area	Area %	Assay %
0		22.065	1560.10	50.33	0.00
1		28.144	1539.55	49.67	0.00

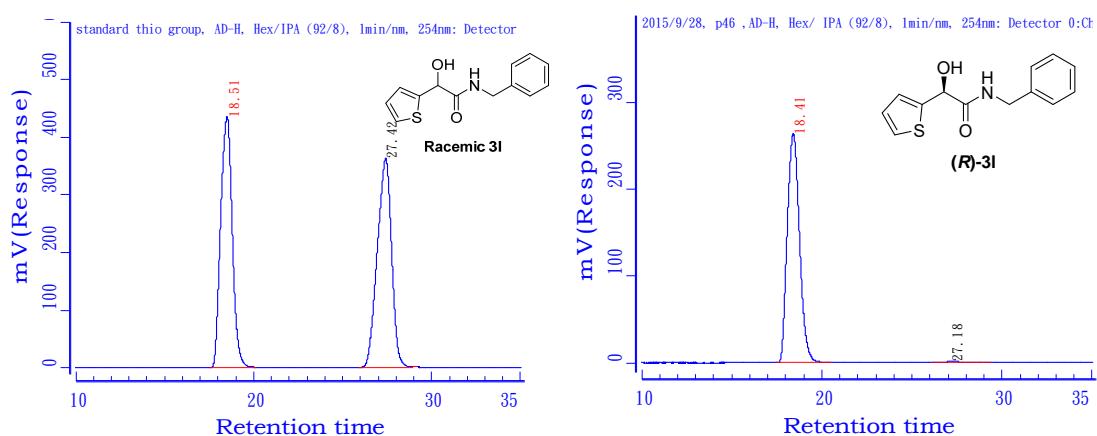


#	Peak name	λ/min (min)	Area	Area %	Assay %
0		21.704	6793.19	96.85	0.00
1		27.245	220.89	3.15	0.00



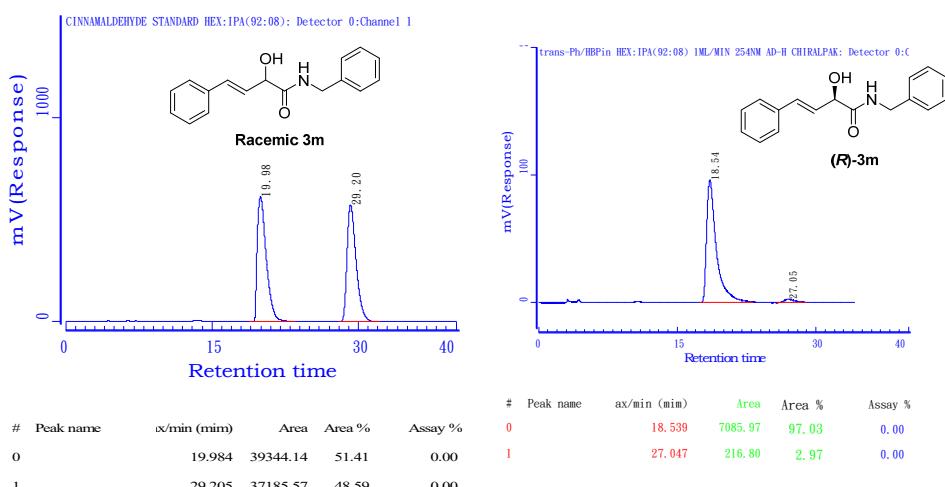
#	Peak name	ax/min (min)	Area	Area %	Assay %
0		20.815	1650.71	98.01	0.00
1		29.852	33.54	1.99	0.00

HPLC conditions for **3k**: t_R 22.06 min (major **R**-iosmer), 28.14 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 94% ee (**R**) and 96% ee (*R*).



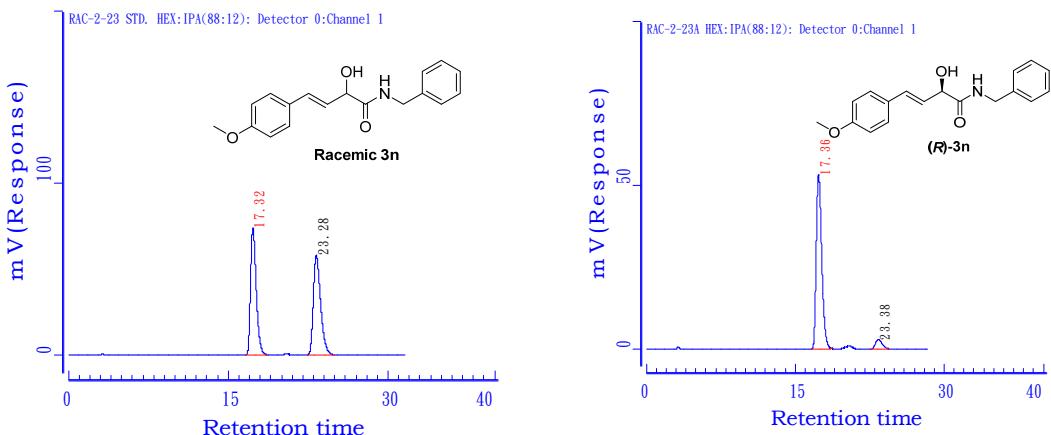
#	Peak name	max/min (min)	Area	Area %	Assay %	#	Peak name	max/min (min)	Area	Area %	Assay %
0		18.508	19638.88	49.29	0.00	0		18.414	12423.07	99.62	0.00
1		27.419	20202.77	50.71	0.00	1		27.179	47.99	0.38	0.00

HPLC conditions for **3l** : t_R 18.51 min (major **R**-iosmer), 27.42 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 99% ee (**R**).



#	Peak name	α/min (min)	Area	Area %	Assay %	#	Peak name	α/min (min)	Area	Area %	Assay %
0		19.984	39344.14	51.41	0.00	0		18.539	7085.97	97.03	0.00
1		29.205	37185.57	48.59	0.00	1		27.047	216.80	2.97	0.00

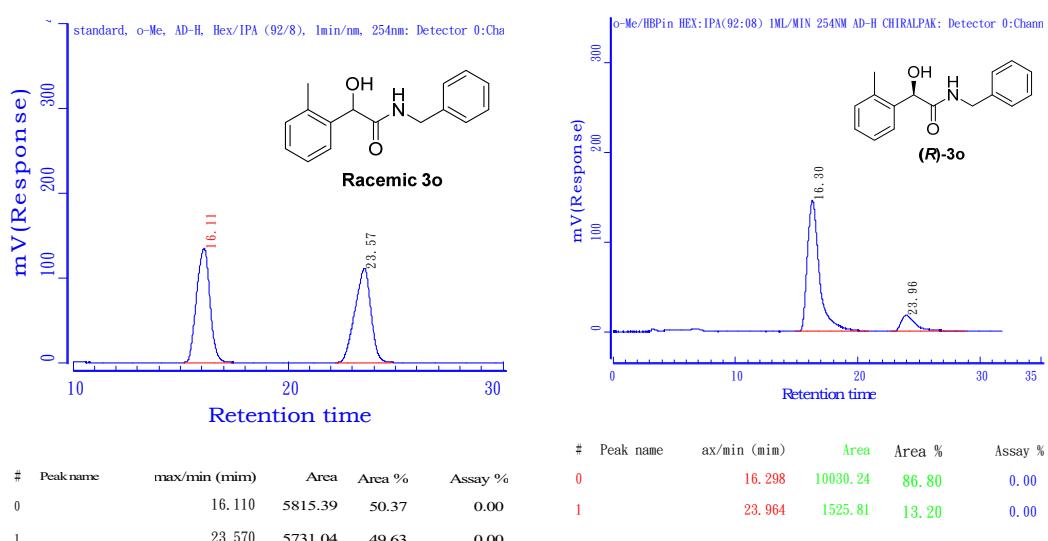
HPLC conditions for **3m** : t_R 19.98 min (major **R**-iosmer), 29.20 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 93% ee (**R**).



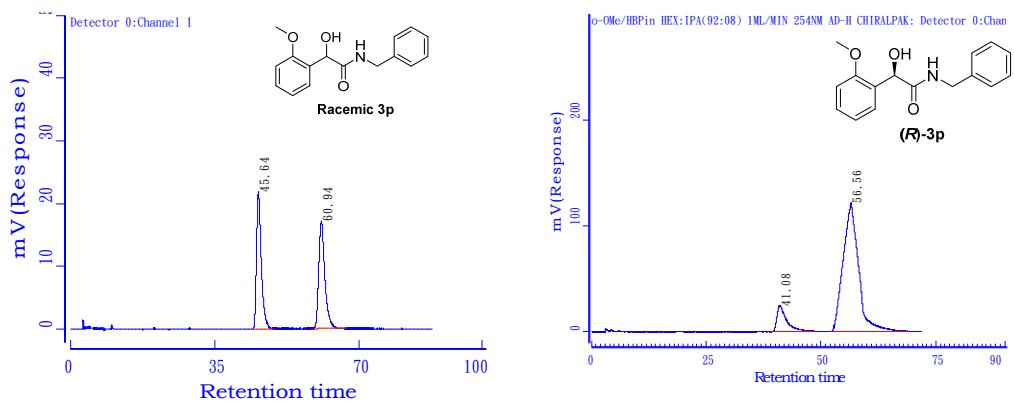
#	Peak name	ax/min (min)	Area	Area %	Assay %
0		17.321	2758.72	49.86	0.00
1		23.284	2774.73	50.14	0.00

#	Peak name	ax/min (min)	Area	Area %	Assay %
0		17.364	2061.16	93.52	0.00
1		23.378	142.73	6.48	0.00

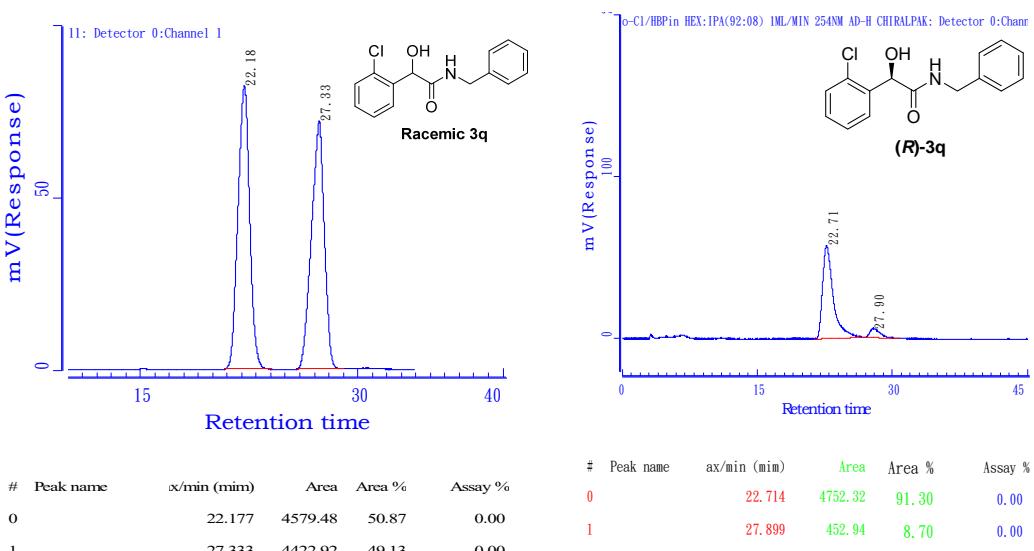
HPLC conditions for **3n** : t_R 18.32 min (major **R**-isomer), 23.28 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 88/12, 1.0 ml/min, λ = 254 nm) for 87% ee.



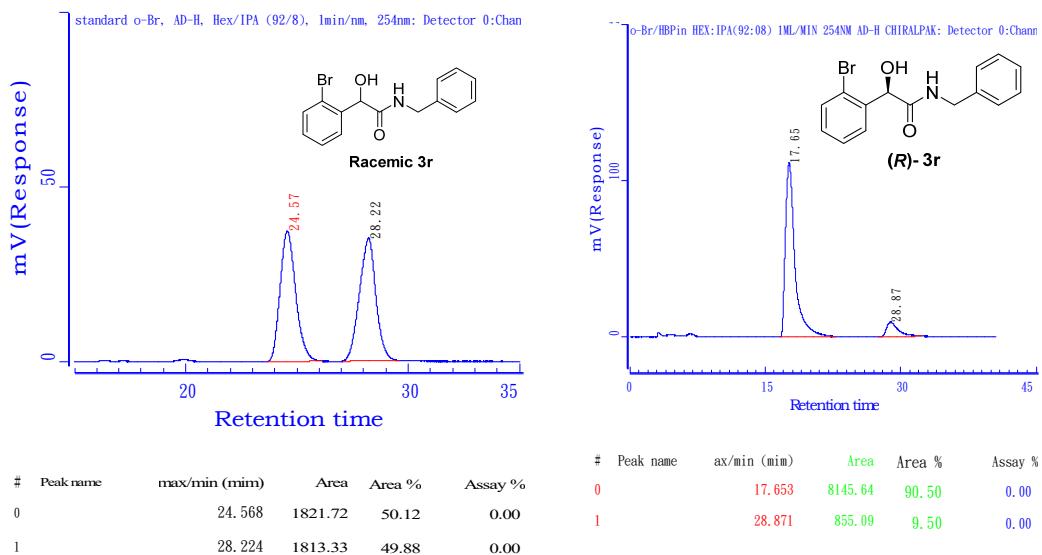
HPLC conditions for **3o** : t_R 16.11 min (major **R**-isomer), 23.57 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 74% ee.



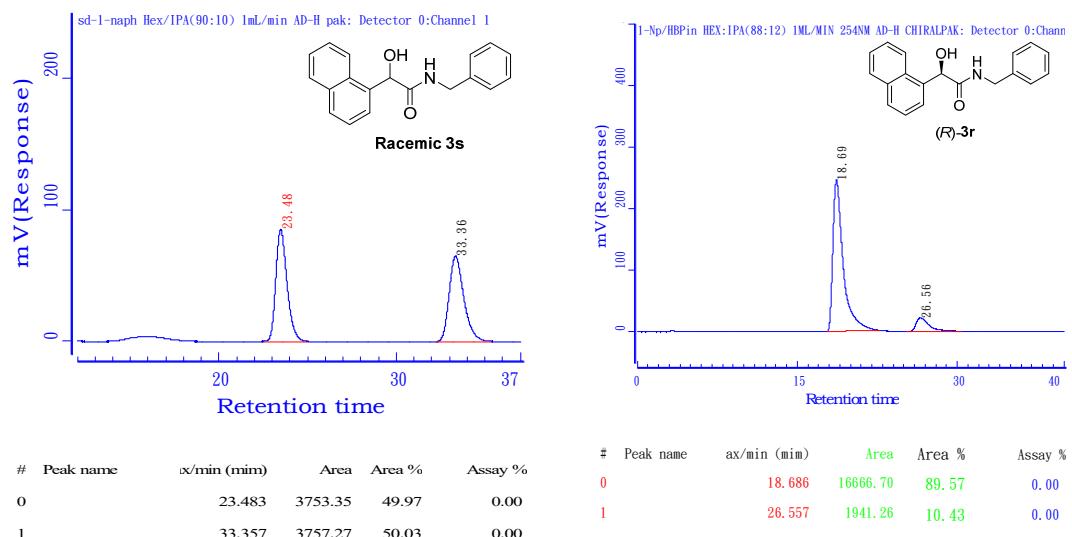
HPLC conditions for **3p** : t_R 16.11 min (**R**-iosmer), 23.57 min (major **S**-isomer)
(Chiraldak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 77% ee.



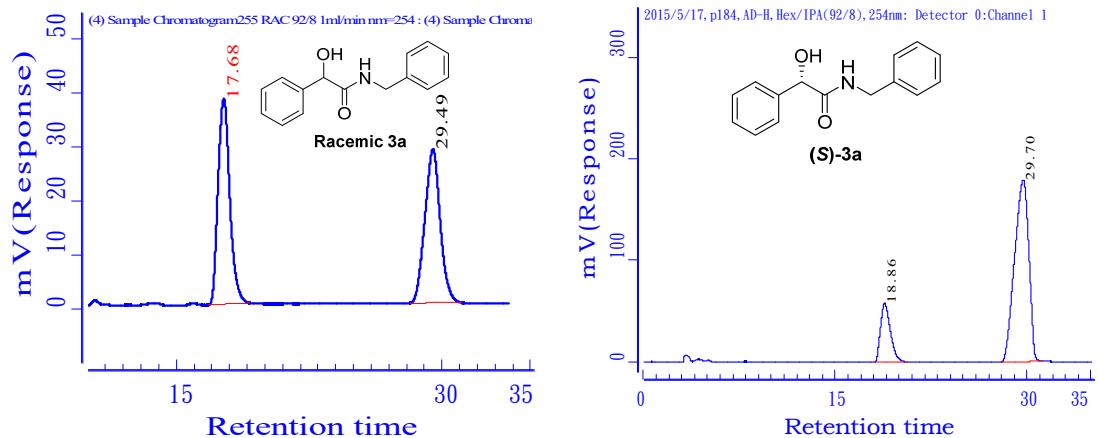
HPLC conditions for **3q** : t_R 22.18 min (major **R**-iosmer), 27.33 min (**S**-isomer)
(Chiraldak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 83% ee.



HPLC conditions for **3r** : t_R 24.57 min (major *R*-isomer), 28.22 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 81% ee.

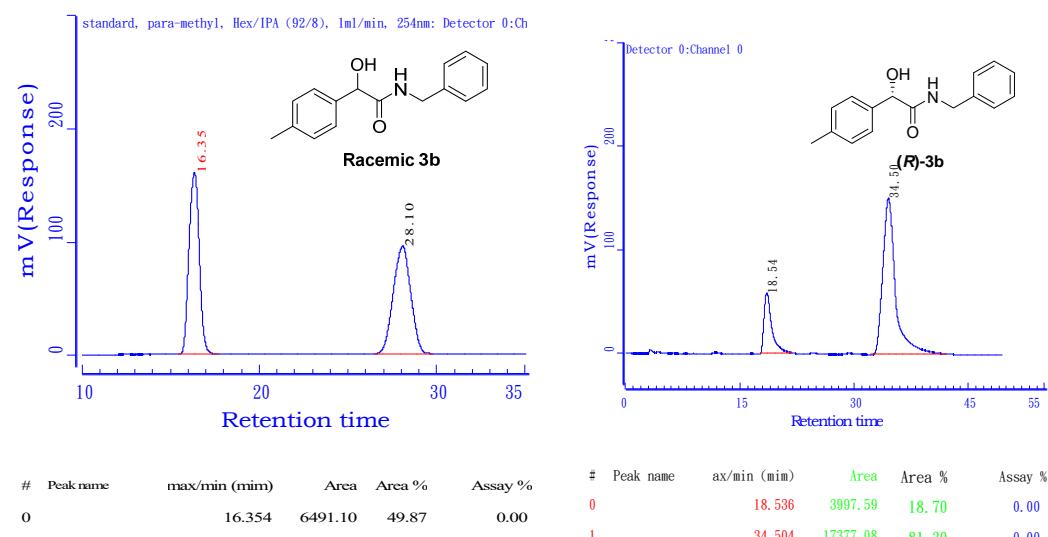


HPLC conditions for **3s** : t_R 23.48 min (major *R*-isomer), 33.36 min (*S*-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 90/10, 1.0 mL/min, λ = 254 nm) for 79% ee.

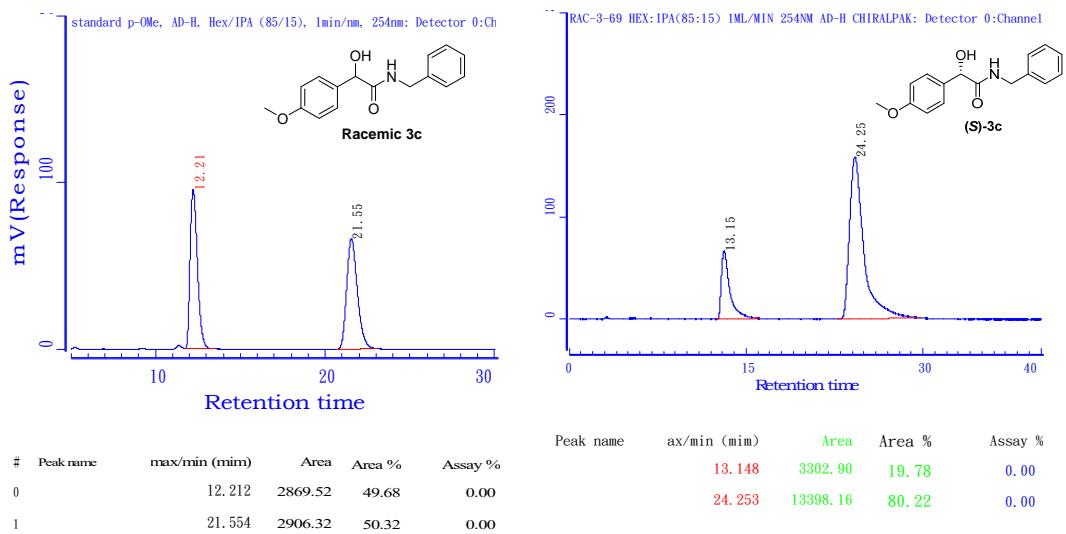


#	Peak name	x/min (min)	Area	Area %	Assay %	#	Peak name	x/min (min)	Area	Area %	Assay %
0		17.682	1761.21	50.72	0.00	0		18.859	3097.18	18.20	0.00
1		29.485	1711.11	49.28	0.00	1		29.701	13919.38	81.80	0.00

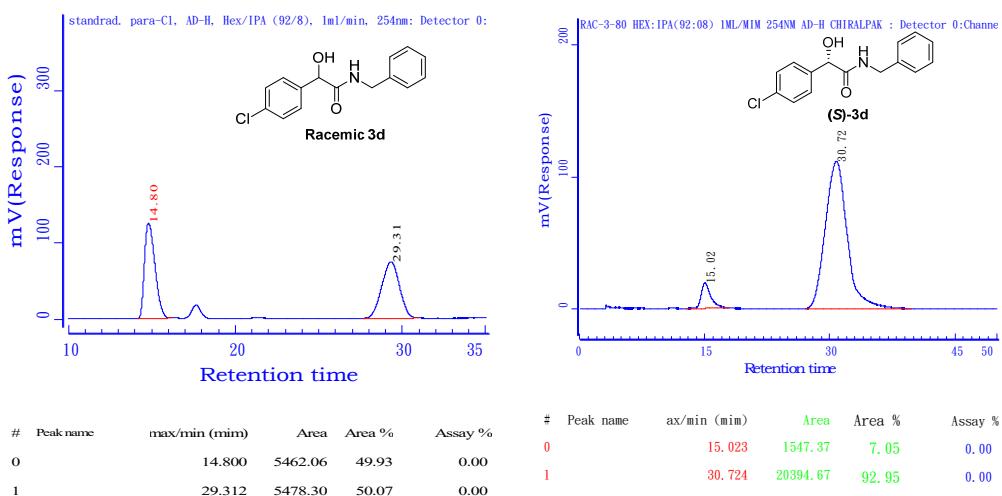
HPLC conditions for **3a** : t_R 17.68 min (**R**-iosmer), 29.49 min (major **S**-isomer)
(Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 64% ee.



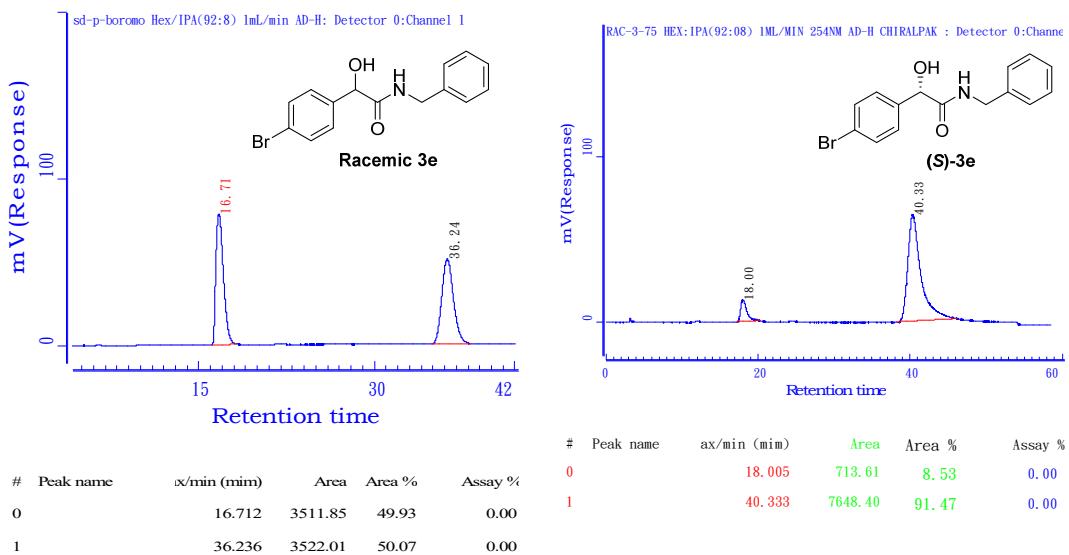
HPLC conditions for **3b** : t_R 16.35 min (**R**-iosmer), 28.10 min (major **S**-isomer)
(Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 63% ee.



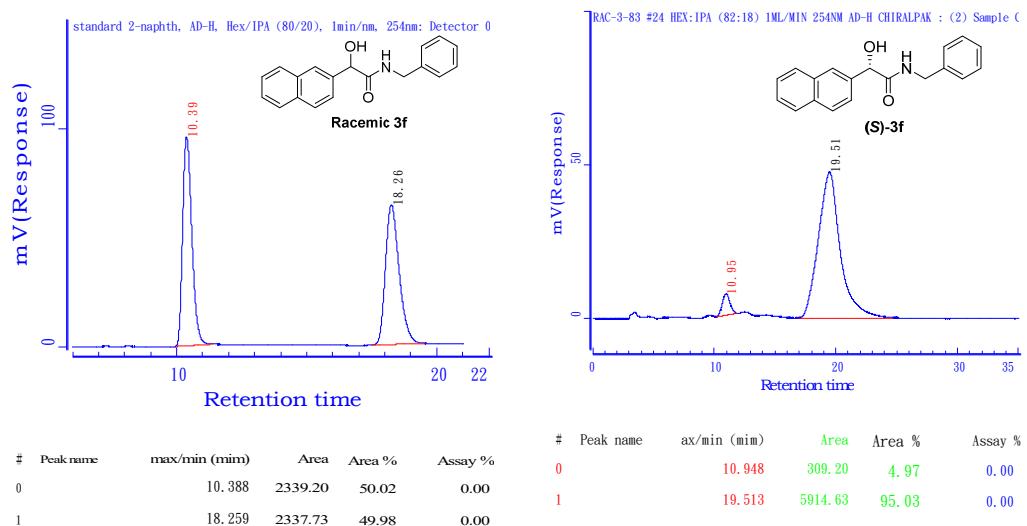
HPLC conditions for **3c** : t_R 12.21 min (**R**-iosmer), 21.55 min (major **S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 85/15, 1.0 ml/min, λ = 254 nm) for 60% ee.



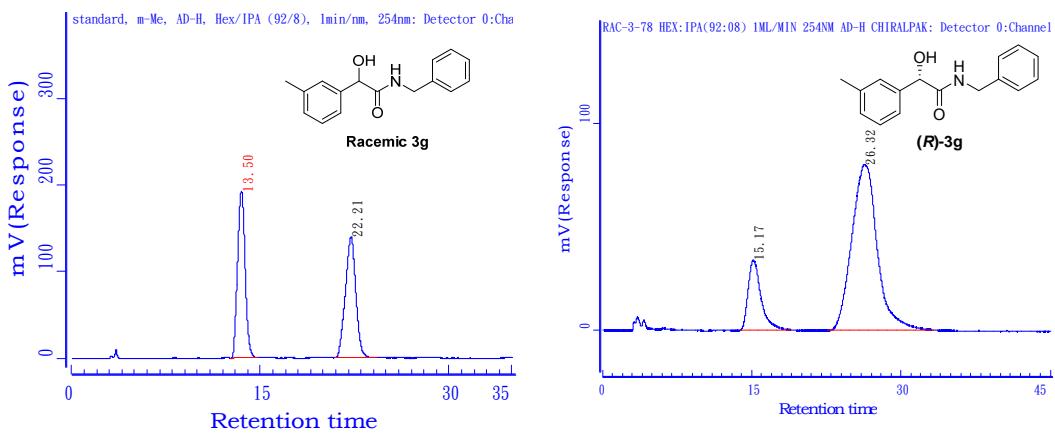
HPLC conditions for **3d** : t_R 14.30 min (**R**-iosmer), 29.31 min (major **S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 86% ee.



HPLC conditions for **3e** : t_R 16.71 min (**R**-iosmer), 36.24 min (major **S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 83% ee.

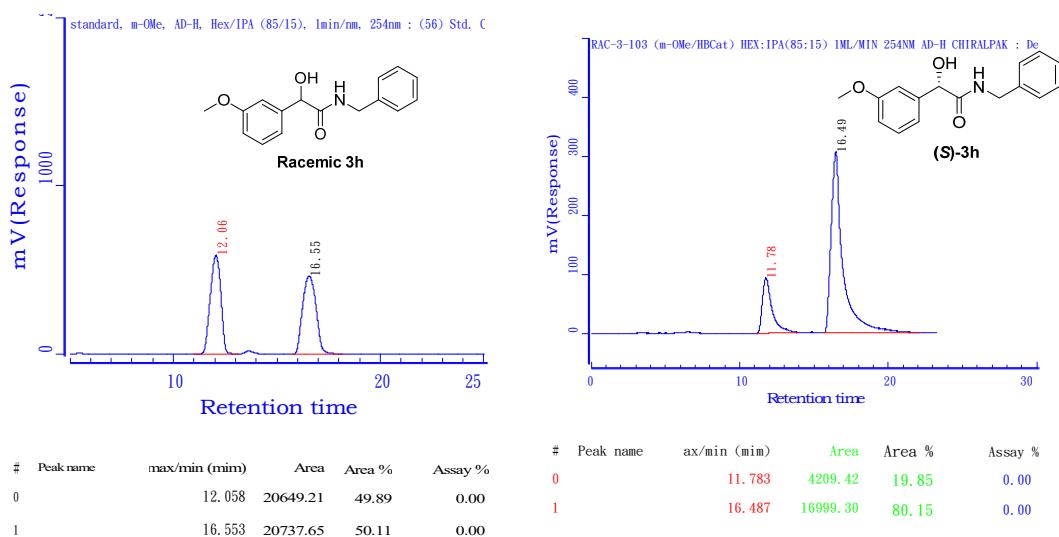


HPLC conditions for **3f** : t_R 10.39 min (**R**-iosmer), 18.26 min (major **S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 80/20, 1.0 ml/min, λ = 254 nm) for 90% ee.



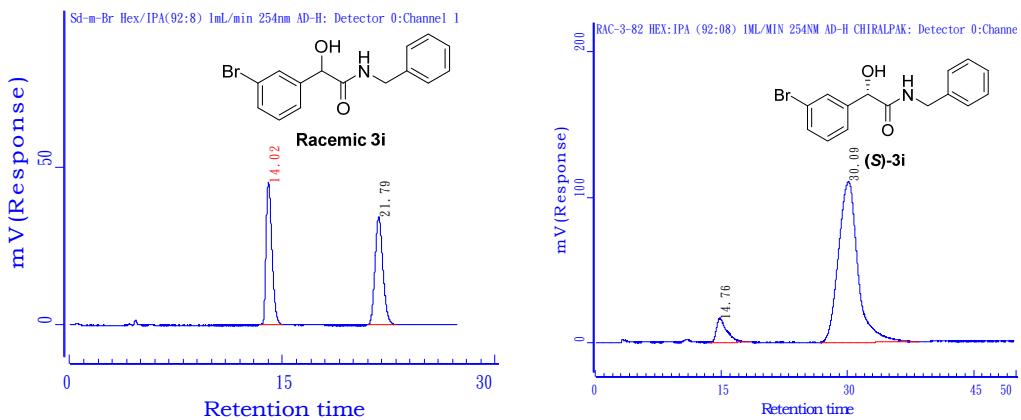
#	Peak name	max/min (min)	Area	Area %	Assay %	#	Peak name	ax/min (min)	Area	Area %	Assay %
0		13.501	8072.94	49.88	0.00	0		15.174	3087.54	17.68	0.00
1		22.210	8112.04	50.12	0.00	1		26.317	14375.21	82.32	0.00

HPLC conditions for **3g** : t_R 13.50 min (**R**-iosmer), 22.21 min (major **S**-isomer) (Chiraldak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 65% ee.



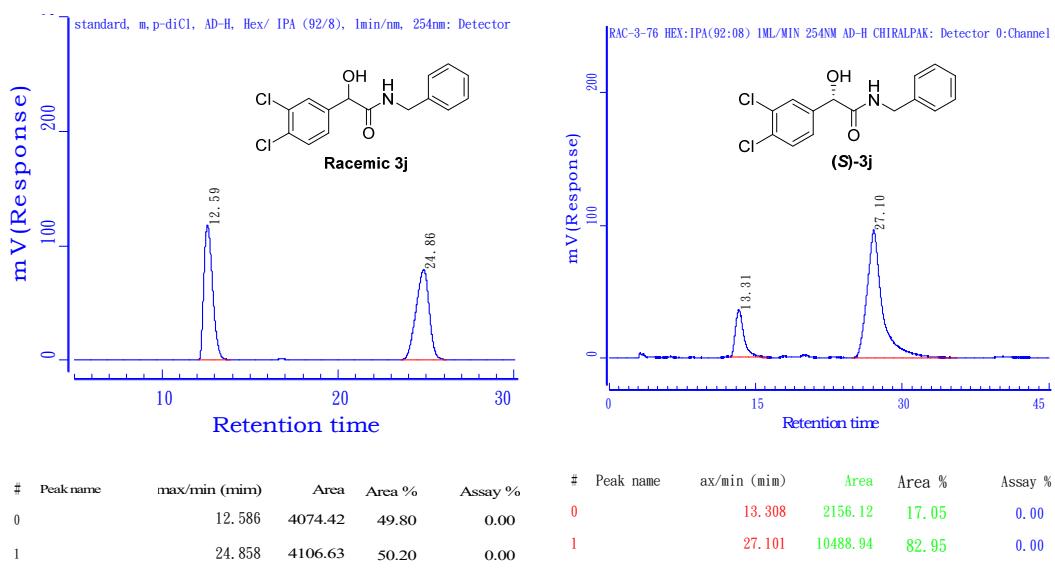
#	Peak name	max/min (min)	Area	Area %	Assay %	#	Peak name	ax/min (min)	Area	Area %	Assay %
0		12.058	20649.21	49.89	0.00	0		11.783	4209.42	19.85	0.00
1		16.553	20737.65	50.11	0.00	1		16.487	16999.30	80.15	0.00

HPLC conditions for **3h** : t_R 12.06 min (**R**-iosmer), 16.55 min (major **S**-isomer) (Chiraldak AD-H, hexane/*i*-PrOH, 85/15, 1.0 ml/min, λ = 254 nm) for 60% ee (**S**).



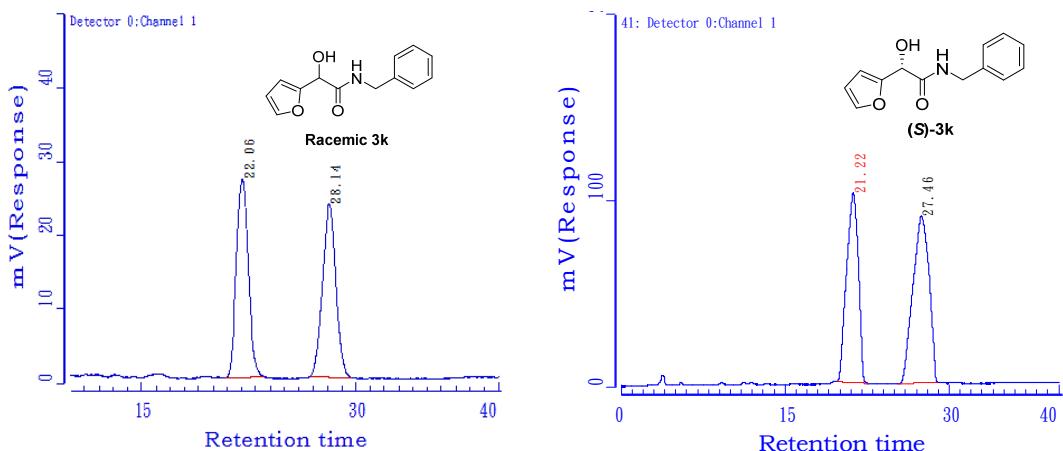
#	Peak name	ax/min (min)	Area	Area %	Assay %	#	Peak name	ax/min (min)	Area	Area %	Assay %
0		14.022	1308.88	49.52	0.00	0		14.759	1422.15	7.40	0.00
1		21.789	1334.19	50.48	0.00	1		30.090	17803.85	92.60	0.00

HPLC conditions for **3i** : t_R 14.02 min (**R**-iosmer), 21.79 min (major **S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 85% ee (**R**).



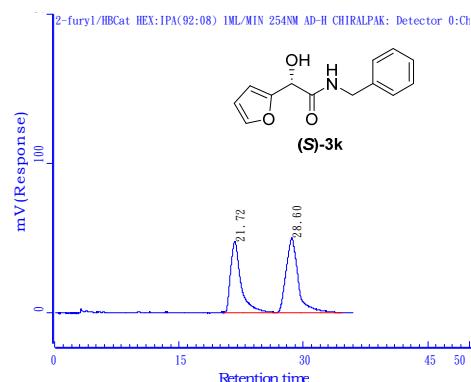
#	Peak name	max/min (min)	Area	Area %	Assay %	#	Peak name	ax/min (min)	Area	Area %	Assay %
0		12.586	4074.42	49.80	0.00	0		13.308	2156.12	17.05	0.00
1		24.858	4106.63	50.20	0.00	1		27.101	10488.94	82.95	0.00

HPLC conditions for **3j** : t_R 12.59 min (**R**-iosmer), 24.86 min (major **S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 66% ee.



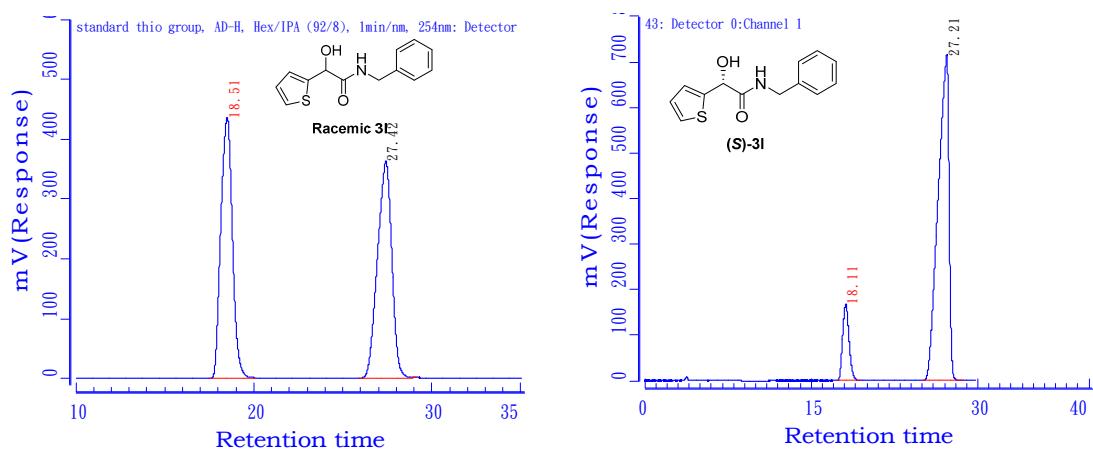
#	Peak name	α/min (min)	Area	Area %	Assay %
0		22.065	1560.10	50.33	0.00
1		28.144	1539.55	49.67	0.00

#	Peak name	α/min (min)	Area	Area %	Assay %
0		21.221	7651.67	45.28	0.00
1		27.455	9247.44	54.72	0.00

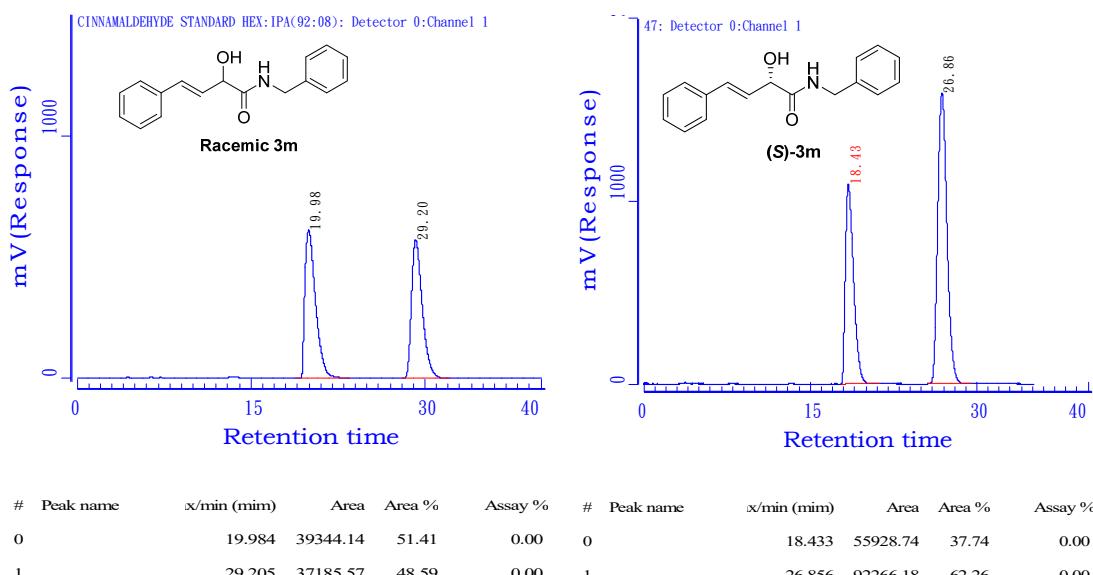


#	Peak name	α/min (min)	Area	Area %	Assay %
0		21.723	4076.55	44.75	0.00
1		28.603	5032.30	55.25	0.00

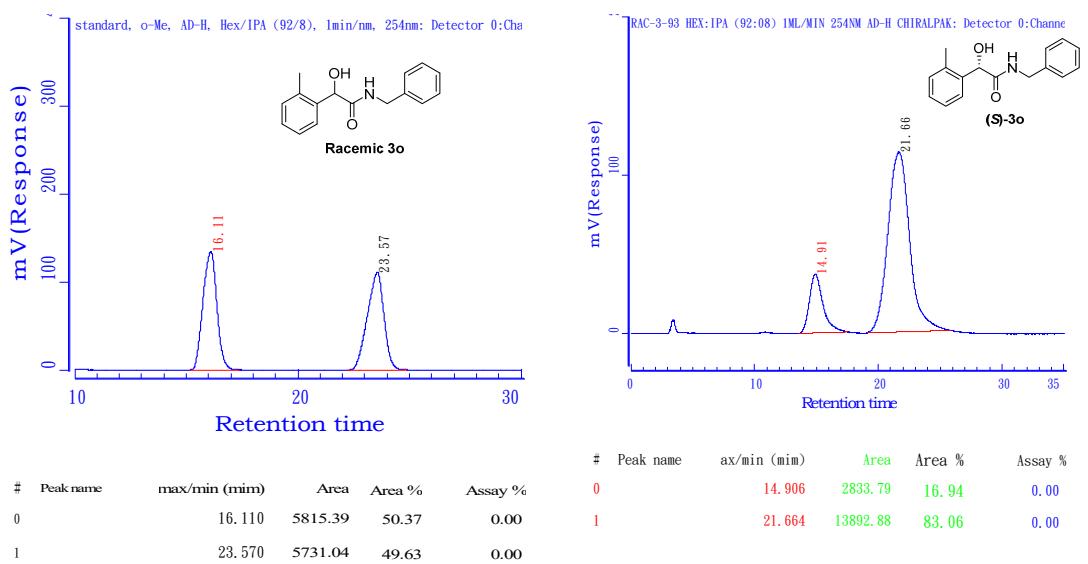
HPLC conditions for **3k** : t_R 22.06 min (**R**-iosmer), 28.14 min (major **S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, $\lambda = 254$ nm) for 10% ee (*S*) and 11%ee (*S*).



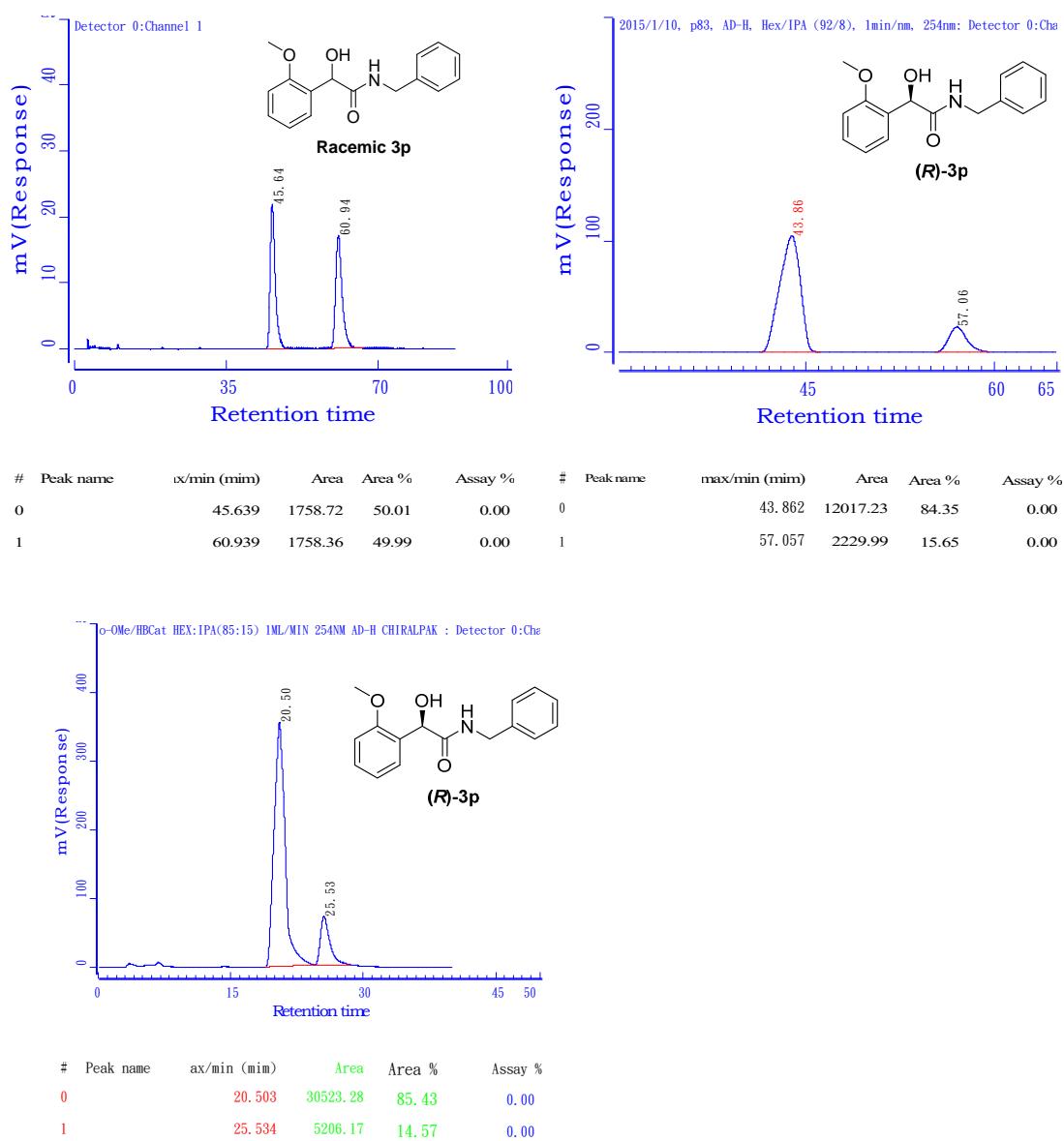
HPLC conditions for **3n** : t_R 18.51 min (**R**-iosmer), 27.42 min (major **S**-isomer)
(Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, $\lambda = 254$ nm) for 75% ee.



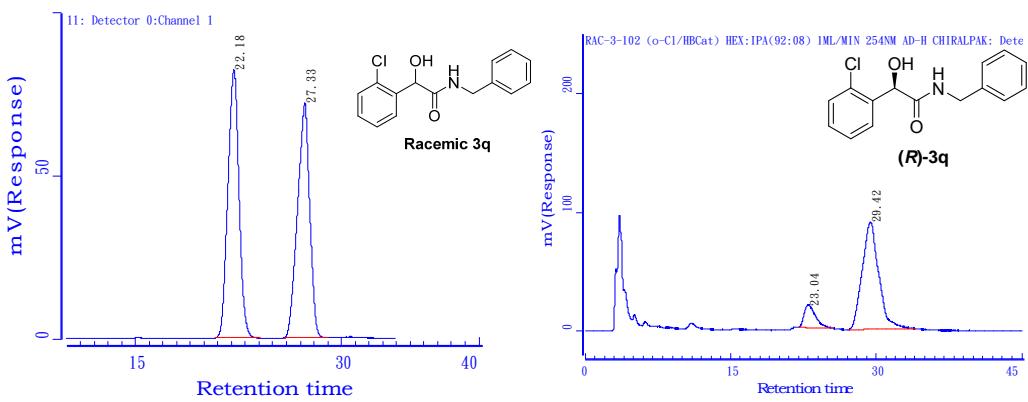
HPLC conditions for **3m** : t_R 19.98 min (**R**-iosmer), 29.20 min (major **S**-isomer)
(Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, $\lambda = 254$ nm) for 25% ee.



HPLC conditions for **3o** : t_R 16.11 min (**R**-iosmer), 23.57 min (major **S**-isomer) (Chiraldak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 66% ee.

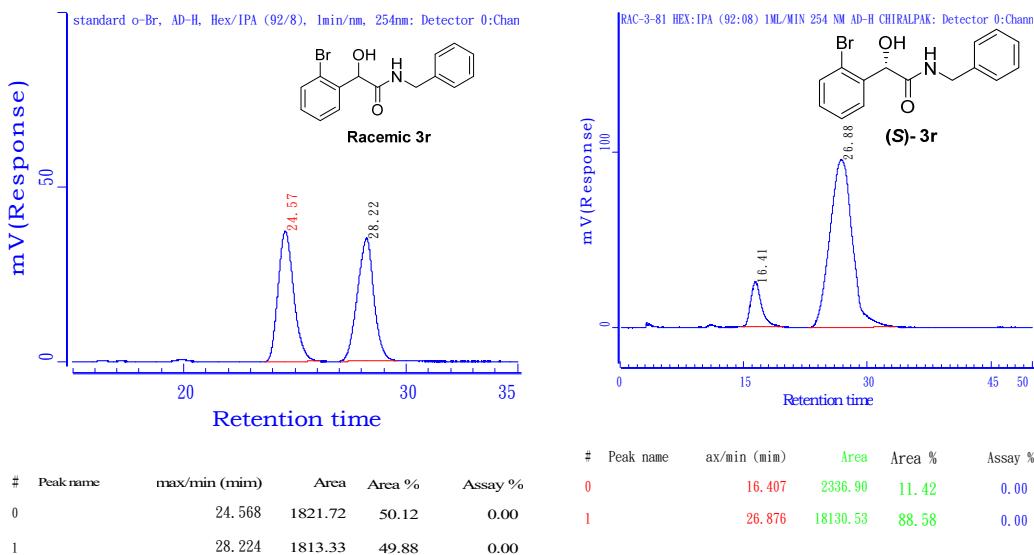


HPLC conditions for **3p** : t_R 43.86 min (major **R**-isomer), 57.06 min (**S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, $\lambda = 254$ nm) for 69% ee (**R**) and 71%ee (**R**).

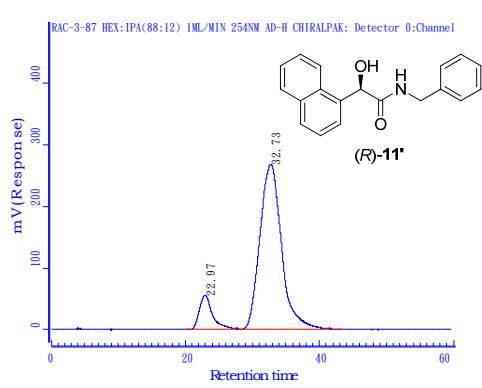
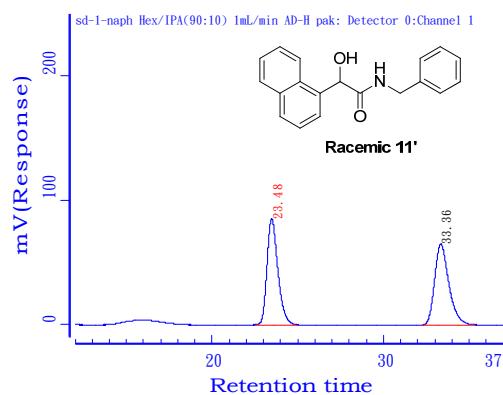


#	Peak name	ax/min (min)	Area	Area %	Assay %	#	Peak name	ax/min (min)	Area	Area %	Assay %
0		22.177	4579.48	50.87	0.00	0		23.044	1568.23	12.76	0.00
1		27.333	4422.92	49.13	0.00	1		29.423	10721.67	87.24	0.00

HPLC conditions for **3q** : t_R 22.18 min (**R**-iosmer), 27.33 min (major **S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 75% ee.

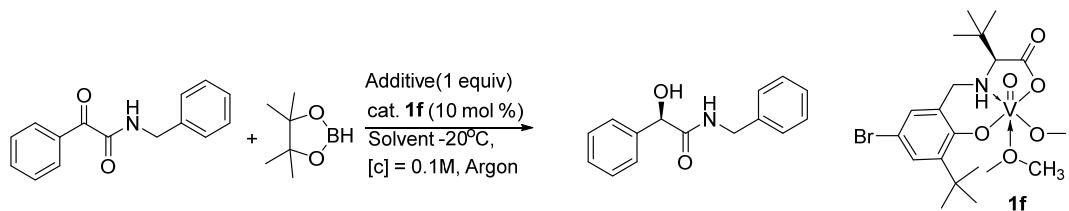


HPLC conditions for **3r** : t_R 24.57 min (**R**-iosmer), 28.22 min (major **S**-isomer) (Chiralpak AD-H, hexane/*i*-PrOH, 92/8, 1.0 ml/min, λ = 254 nm) for 77% ee.



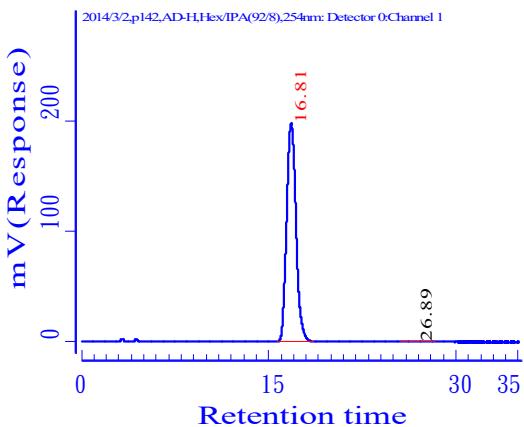
HPLC conditions for **11'** : t_R 23.48 min (*R*-isomer), 33.36 min (major *S*-isomer)
 (Chiralpak AD-H, hexane/*i*-PrOH, 90/10, 1.0 mL/min, $\lambda = 254$ nm) for 78% ee.

Table S1

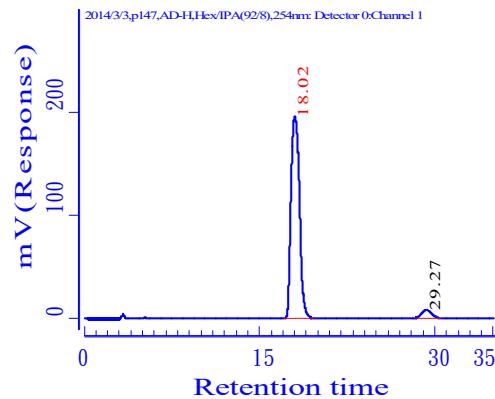


Entry	Reducing Agent	Additive	Time (h)	Yield (%)	Ee (%)
1	PinB–H	—	96	65	88 (R)
2	PinB–H	MeOH	96	40	99 (R)
3	PinB–H	CCl ₃ CH ₂ OH	96	47	91 (R)
4	PinB–H	CF ₃ CH ₂ OH	96	22	96 (R)
5	CatB–H	—	14	99	64 (S)
6	CatB–H	MeOH	14	92	47 (S)
7	CatB–H	CCl ₃ CH ₂ OH	14	86	58 (S)
8	CatB–H	CF ₃ CH ₂ OH	14	84	62 (S)

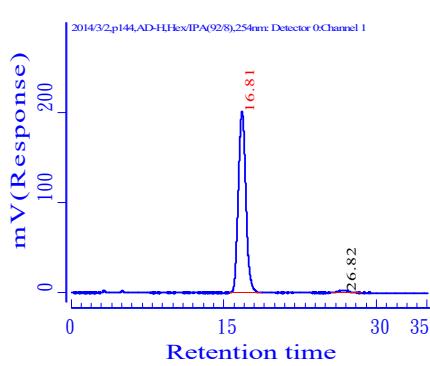
Entry 2 Table S1



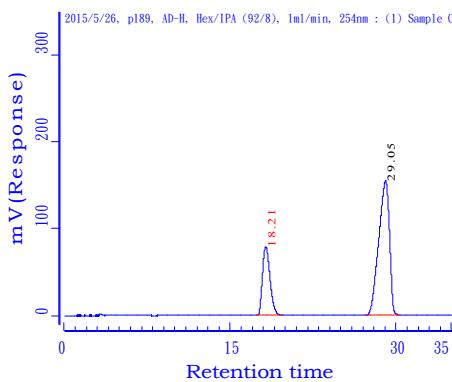
Entry 3 Table S1



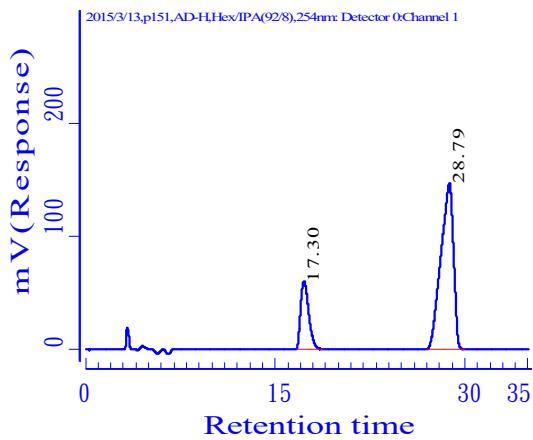
Entry 4 Table S1



Entry 6 Table S1



Entry 7 Table S1



Entry 8 Table S1

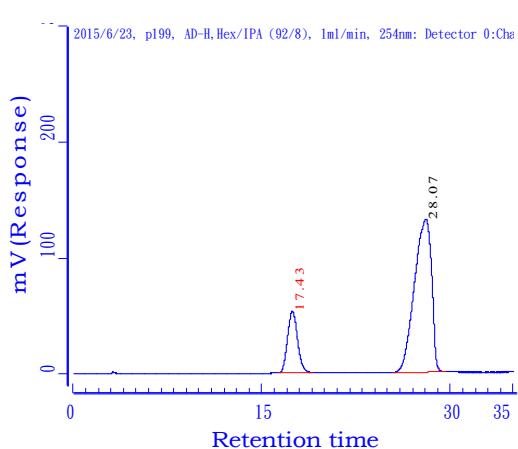
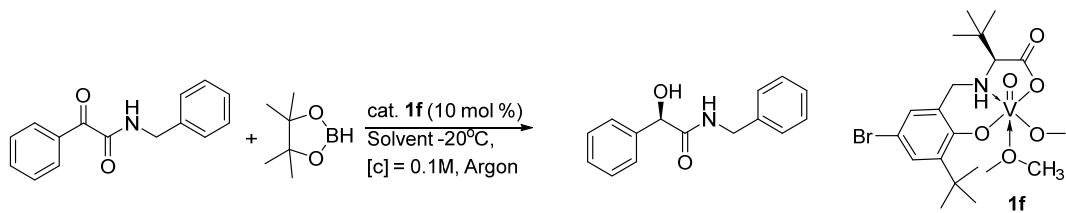
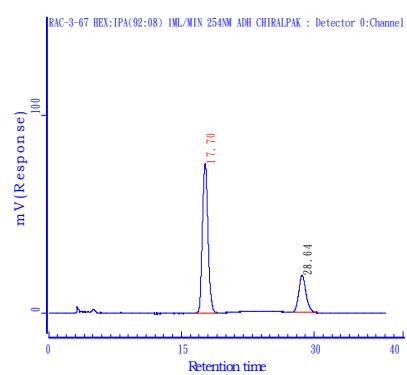


Table S2



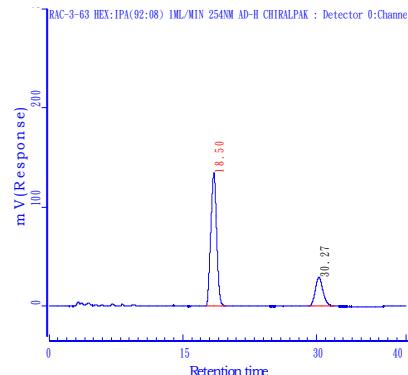
entry	Solvent	Time(h)	Yield(%)	ee%
1 (final)	toluene	24+24+48	65	88 (<i>R</i>)
2	MeOH	24+24+48	35	52 (<i>R</i>)
3	<i>i</i> -PrOH	24+24+48	43	56 (<i>R</i>)
4	1,4-dioxane	24+24+48	21	36 (<i>R</i>)
5	CH ₃ CN	24+24+48	28	38 (<i>R</i>)
6	DMSO (r.t.)	24+24+48	52	12 (<i>R</i>)
7	DMF	24+24+48	19	24 (<i>R</i>)

Entry 2 Table S2



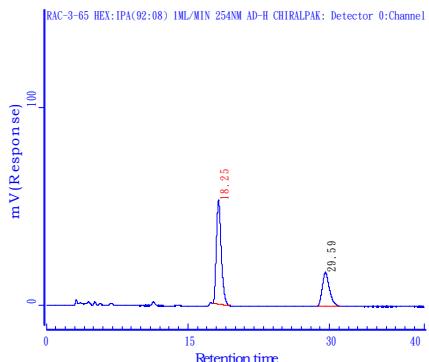
#	Peak name	x/min (min)	Area	Area %	Assay %
0		17.696	3321.98	75.88	0.00
1		28.641	1055.82	24.12	0.00

Entry 3 Table S2

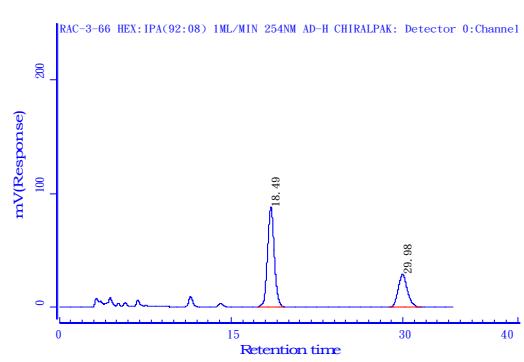


#	Peak name	x/min (min)	Area	Area %	Assay %
0		18.502	6143.29	78.02	0.00
1		30.272	1730.47	21.98	0.00

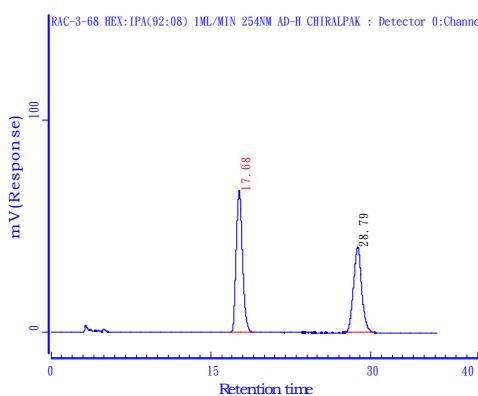
Entry 4 Table S2



Entry 5 Table S2



Entry 6 Table S2



Entry 7 Table S2

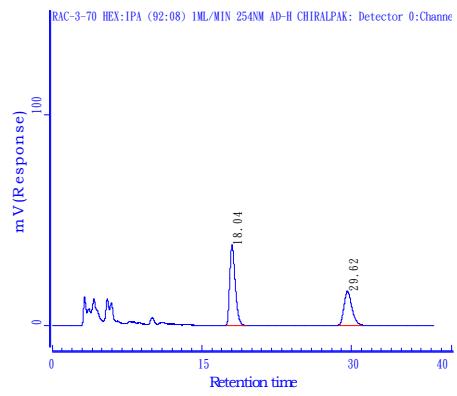
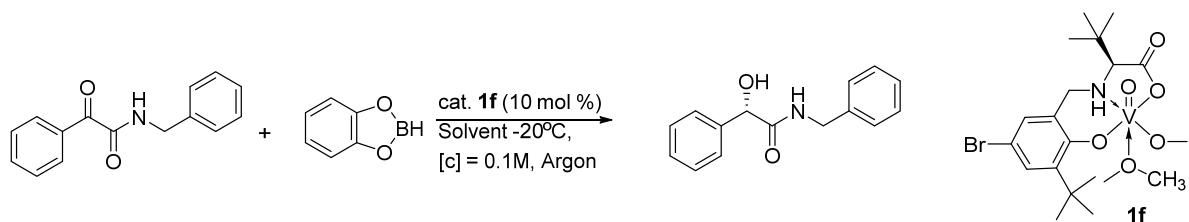
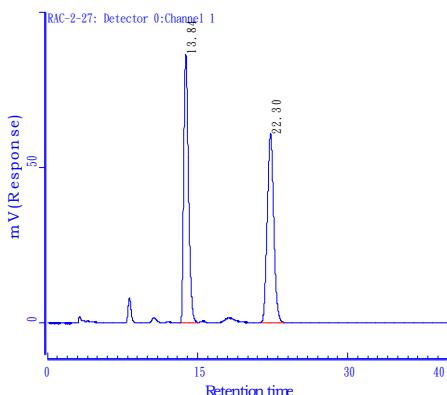


Table S3

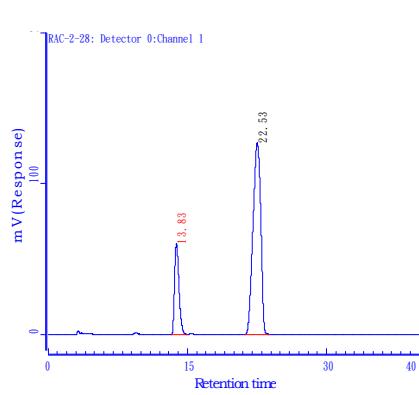


entry	Solvent	Time(h)	Yield(%)	ee%
1 (final)	toluene	12	99	64 (S)
2	CH ₂ Cl ₂	12	97	2 (R)
3	PhCl	14	95	57 (S)
4	THF	12	98	35 (S)
5	<i>t</i> -BuOMe	15	87	4 (R)

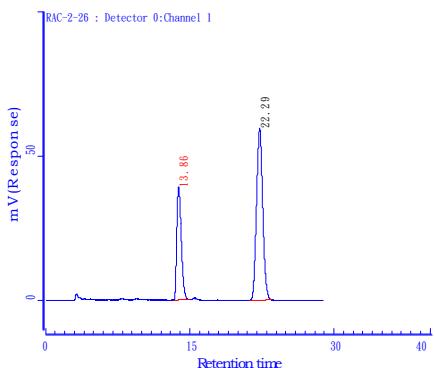
Entry 2 Table S3



Entry 3 Table S3

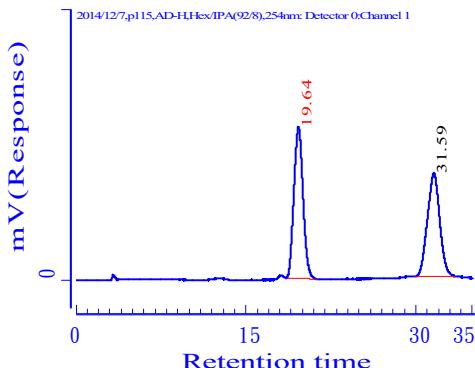


Entry 4 Table S3



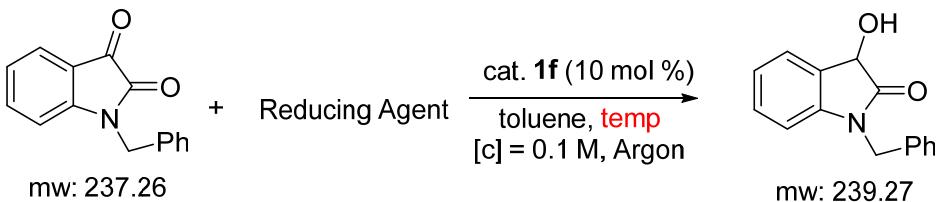
#	Peak name	x/min (min)	Area	Area %	Assay %
0		13.860	1277.85	32.42	0.00
1		22.289	2663.32	67.58	0.00

Entry 5 Table S3

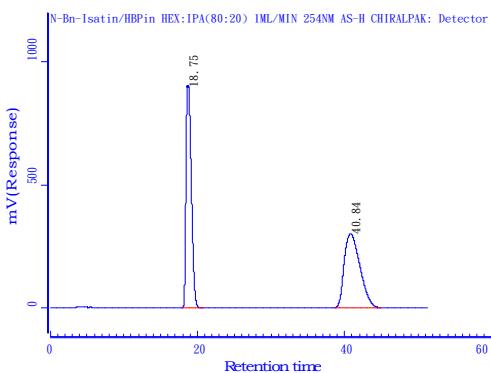


#	Peak name	x/min (min)	Area	Area %	Assay %
0		19.642	3133.66	52.16	0.00
1		31.586	2873.65	47.84	0.00

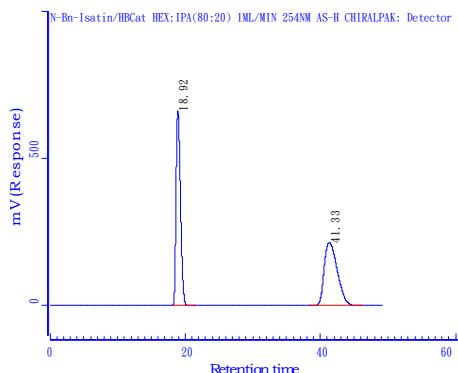
Table S4



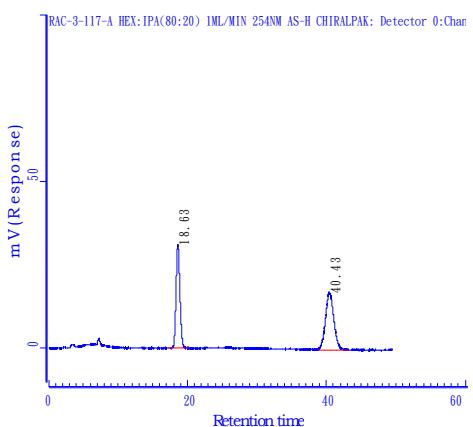
reducing agent	temperature, °C	Yield, %	ee, %
HBPin	-20/-40	99/0	0/---
HBCat	-20/-40	92/28	2/8 (S)



#	Peak name	ax/min (min)	Area	Area %	Assay %
0		18.753	45594.00	50.36	0.00
1		40.842	44942.66	49.64	0.00

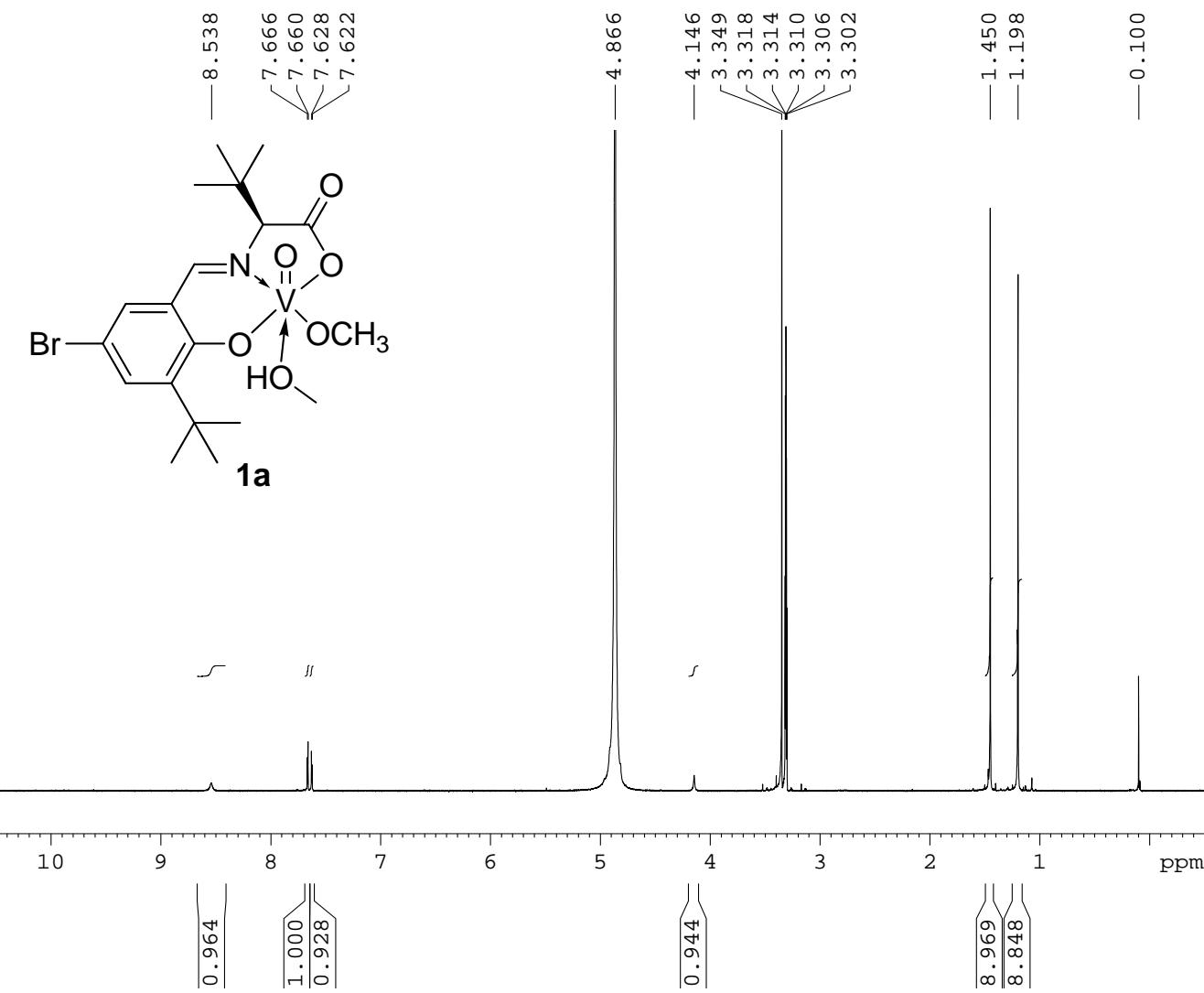


#	Peak name	ax/min (min)	Area	Area %	Assay %
0		18.917	30101.72	50.96	0.00
1		41.334	28965.59	49.04	0.00



#	Peak name	ax/min (min)	Area	Area %	Assay %
0		18.625	1223.93	46.08	0.00
1		40.425	1432.16	53.92	0.00

7. ^1H and ^{13}C NMR spectra

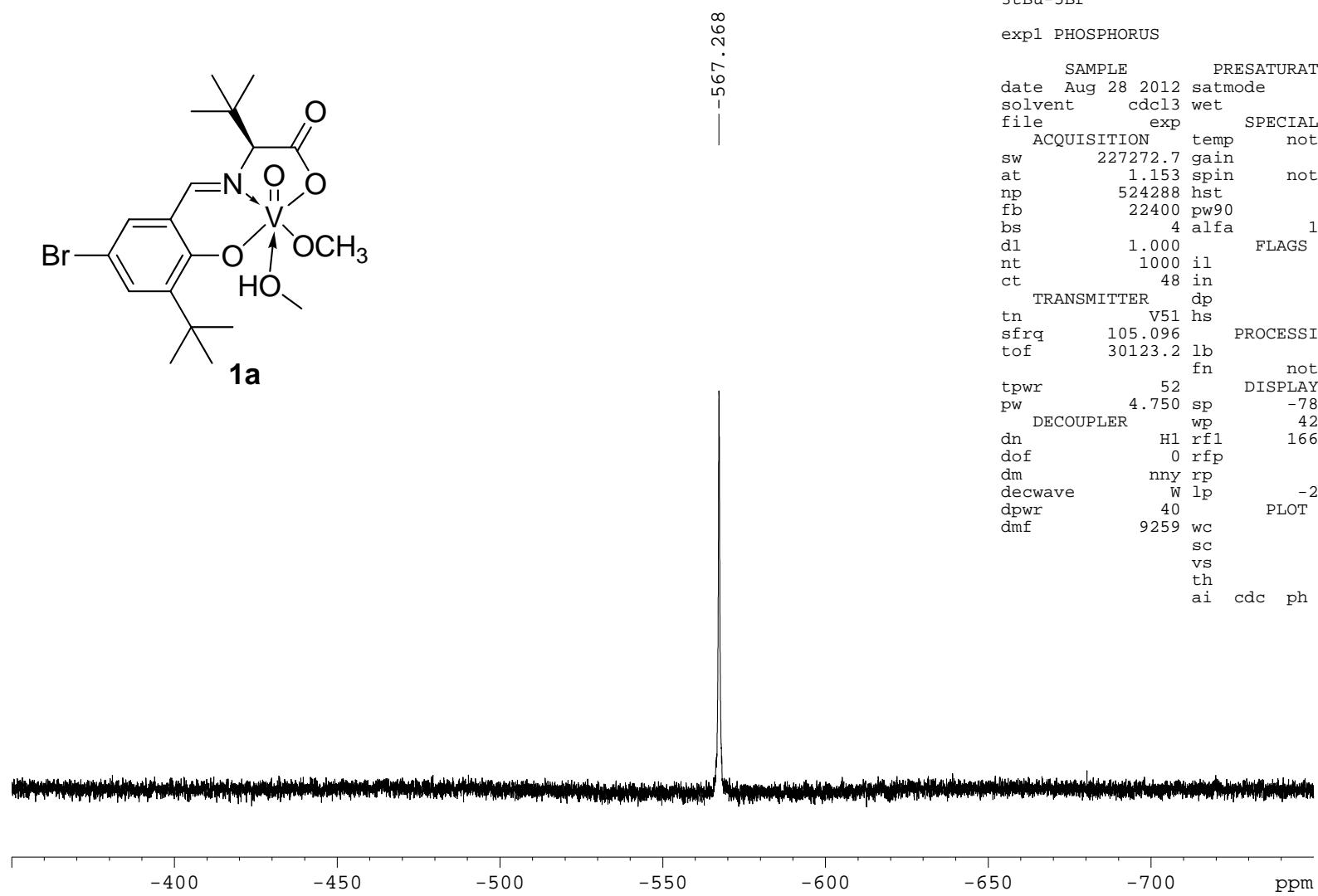
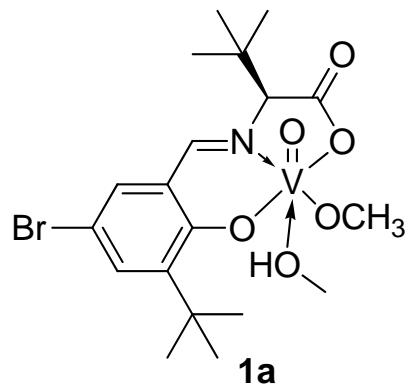


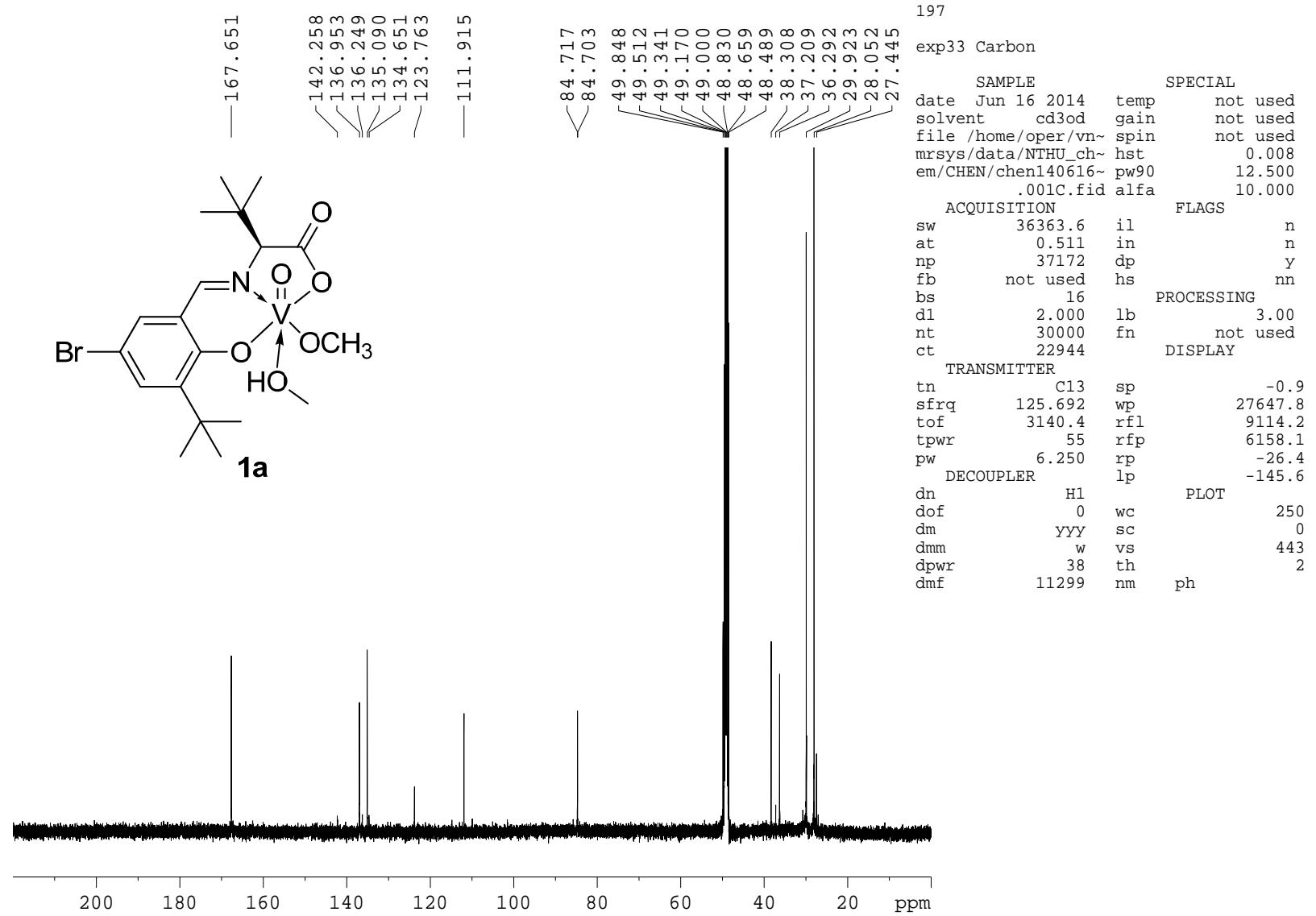
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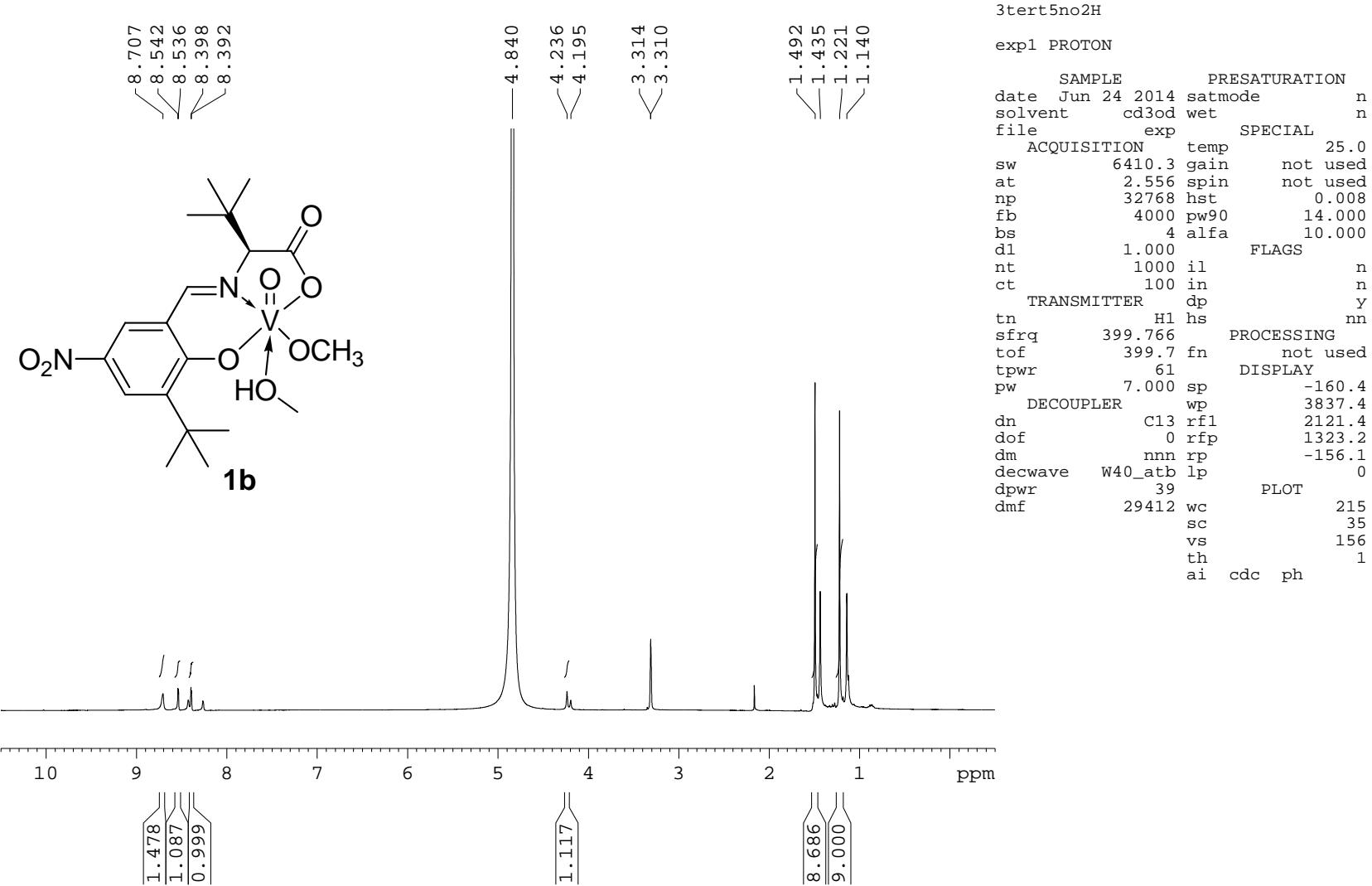
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EXPNO         31
PROCNO        1
Date_        20120829
Time         16.01
INSTRUM      spect
PROBHD       5 mm DUL 13C-1
PULPROG      zg30
TD           32768
SOLVENT      MeOD
NS            8
DS            0
SWH          6410.256 Hz
FIDRES       0.195625 Hz
AQ            2.5559540 sec
RG            4
DW           78.0000 usec
DE            6.00 usec
TE           300.0 K
D1           2.00000000 sec
TD0           1

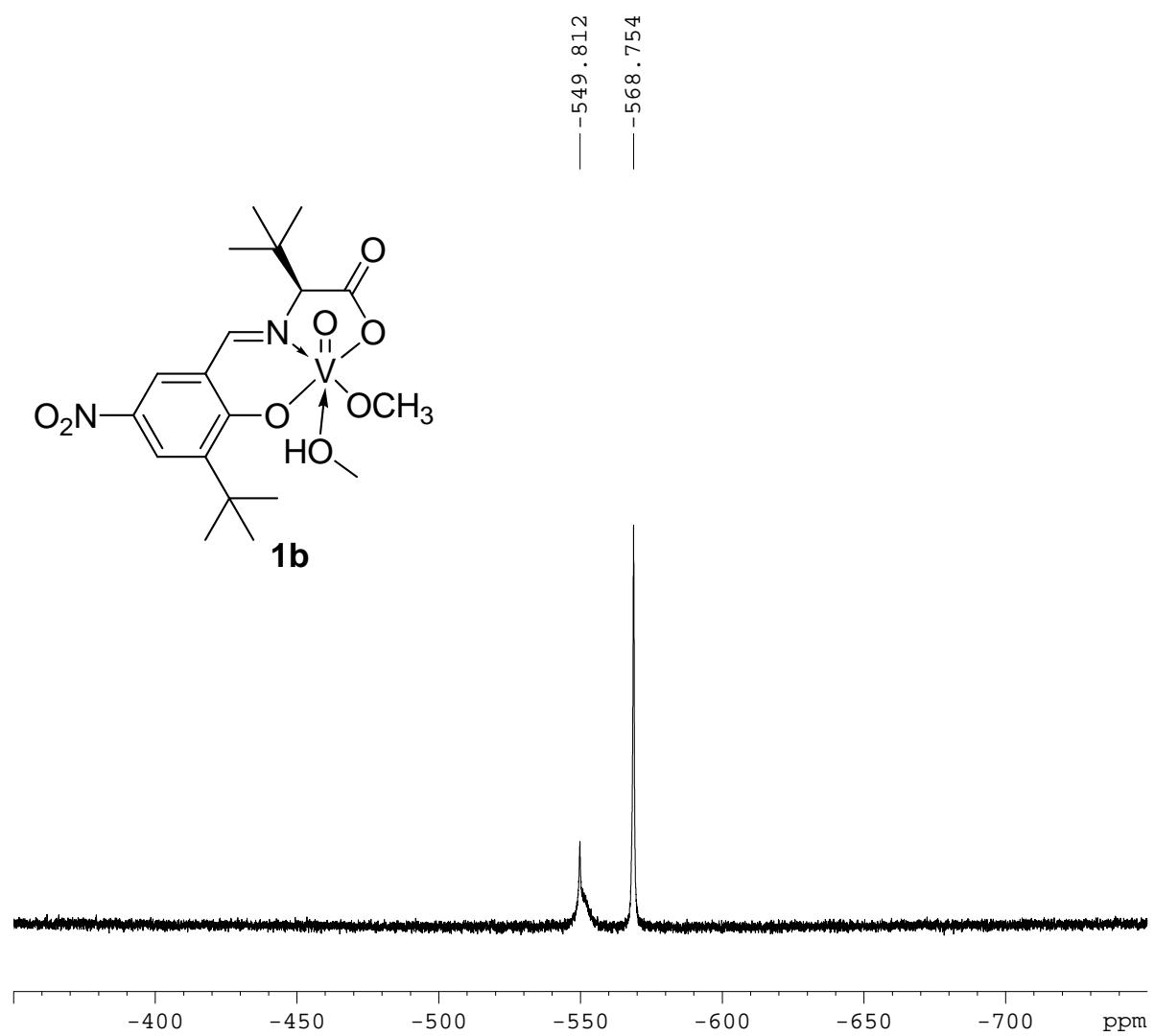
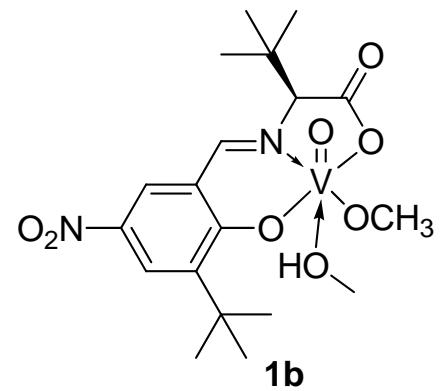
===== CHANNEL f1 =====
NUC1           1H
P1            10.00 usec
PL1           -2.40 dB
SFO1        400.1528010 MHz
SI             16384
SF           400.1500067 MHz
WDW           EM
SSB            0
LB            0.00 Hz
GB            0
PC            1.00

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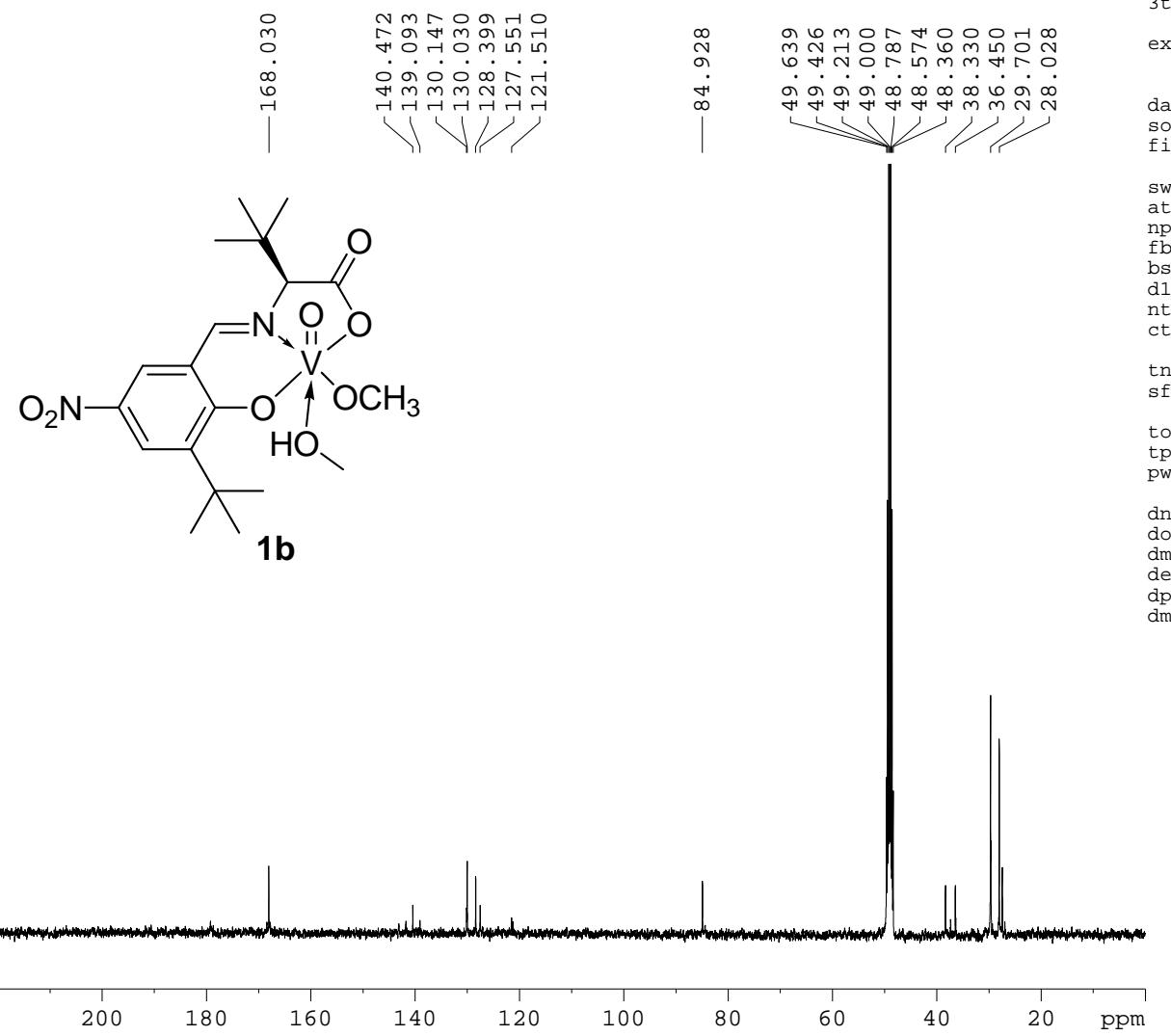
NO₂tertV

exp2 PHOSPHORUS

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SAMPLE      PRESATURATION
date  Apr 12 2014 satmode      n
solvent    cd3od wet          n
file       exp          SPECIAL
ACQUISITION temp      not used
sw        227272.7 gain      50
at         0.577 spin      not used
np        262144 hst       0.008
fb        22400 pw90      9.500
bs          4 alfa     10.000
d1        1.000           FLAGS
nt        1000 il          n
ct         164 in          n
TRANSMITTER dp          y
tn        V51 hs          nn
sfrq     105.088          PROCESSING
tof      22089.8 lb      3.00
          fn      not used
tpwr      52           DISPLAY
pw        4.750 sp      -78862.8
DECOUPLER   wp      42058.8
dn         H1 rrf1     174712.3
dof        0 rfp          0
dm        nny rp      -106.7
decwave    W lp      -923.0
dpwr      40           PLOT
dmf       8889 wc      215
          sc          0
          vs      2517
          th      14
ai       cdc ph

```



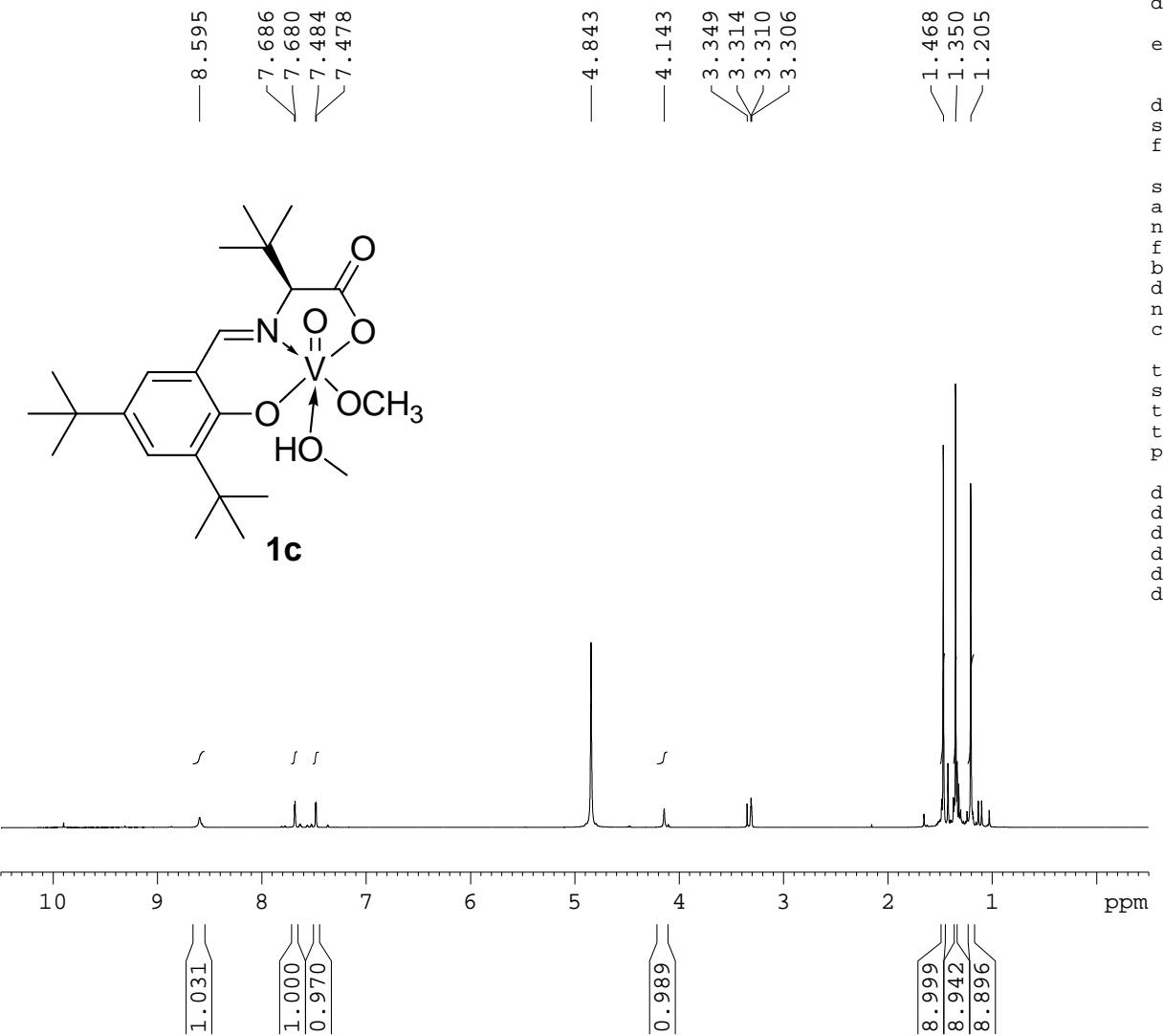
3tert5no2C

exp33 CARBON

```

SAMPLE             PRESATURATION
date   Jun 24 2014 satmode      n
solvent  cd3od wet           n
file    exp          SPECIAL
ACQUISITION temp       25.0
sw     25510.2 gain        30
at      1.285 spin      not used
np      65536 hst         0.008
fb     17000 pw90        9.800
bs        4 alfa       10.000
d1      1.000          FLAGS
nt     100000 il          n
ct      988 in           n
TRANSMITTER dp          y
tn      C13 hs          nn
sfrq    100.532          PROCESSING
                      lb       3.00
tof     1530.7 fn      not used
tpwr     60          DISPLAY
pw      4.900 sp        -0.9
DECOUPLER   wp       22114.3
dn        H1 rrf1      6483.4
dof        0 rfp        4925.5
dm        YYY rp        -90.2
decwave    W lp        14.0
dpwr      41          PLOT
dmf     10695 wc        250
                      sc        0
                      vs        405
                      th        5
nm      cdc      ph

```

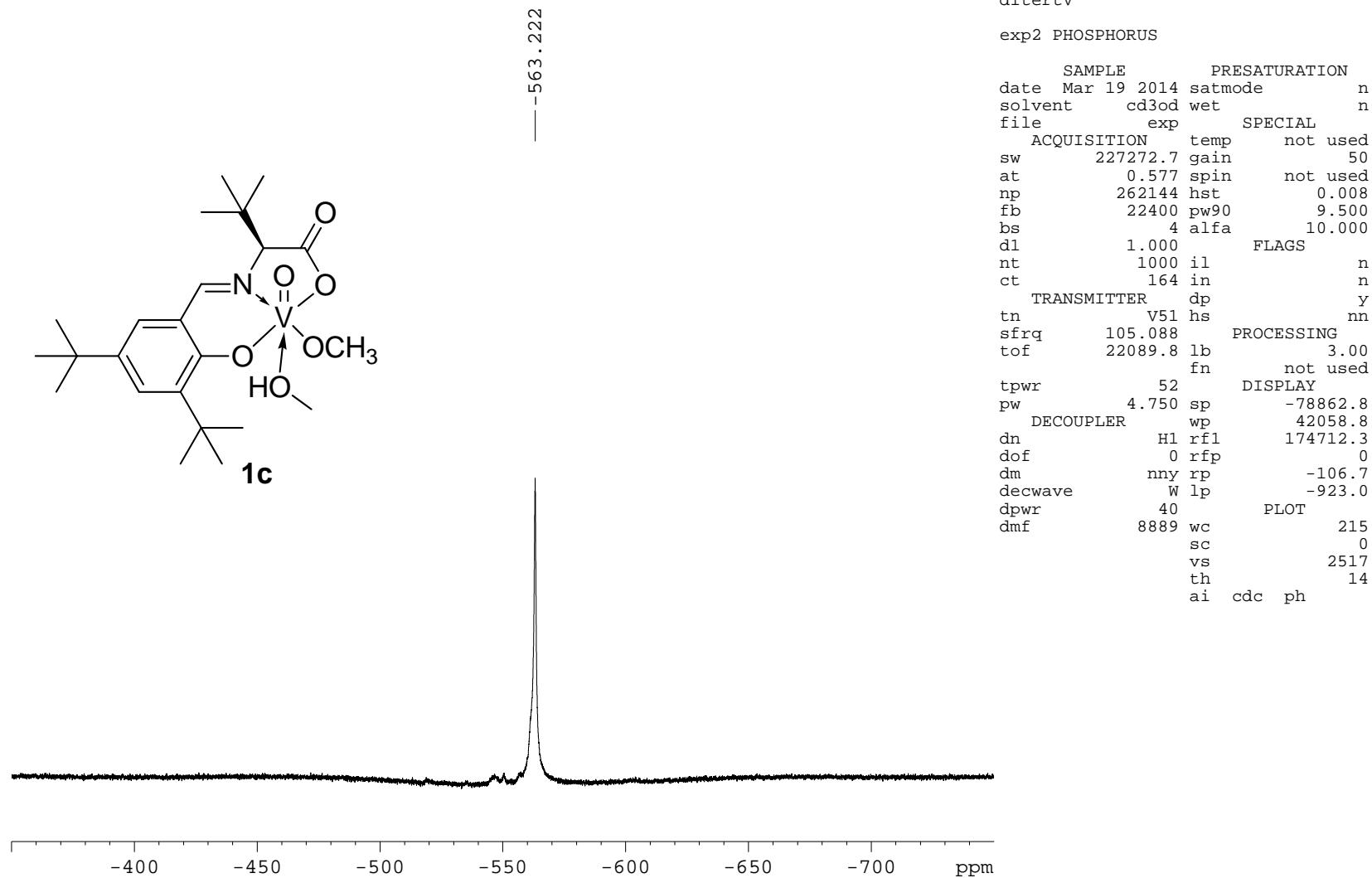
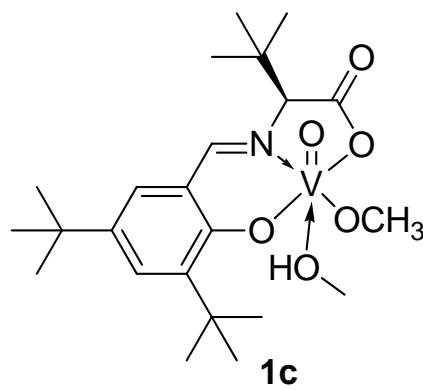


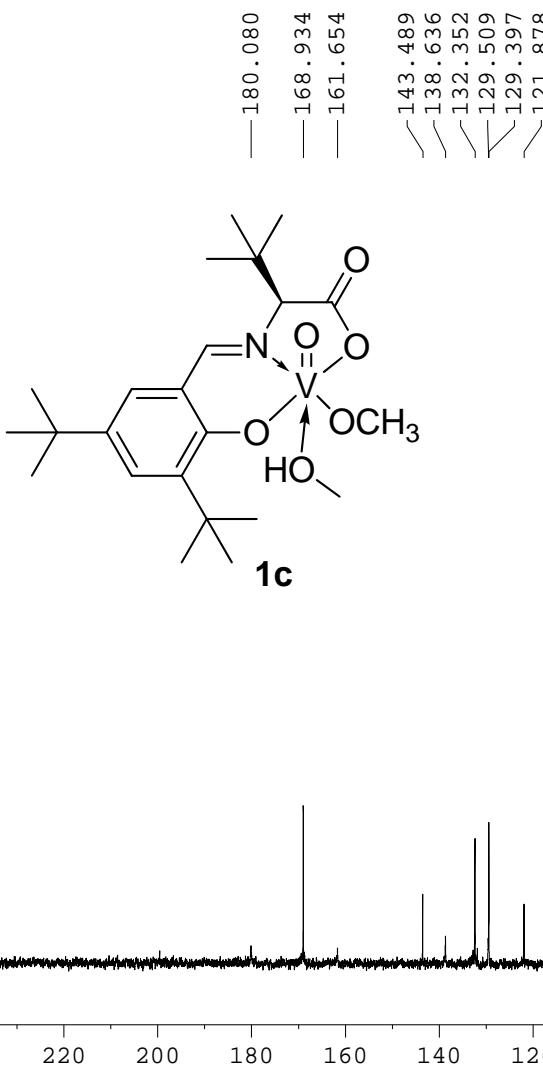
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diterth
exp33 PROTON

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date Jun 25 2014 satmode      n
solvent cd3od wet          n
file     exp      SPECIAL
      ACQUISITION temp    not used
sw       6410.3 gain   not used
at        2.556 spin   not used
np       32768 hst    0.008
fb        4000 pw90   14.000
bs         4 alfa   10.000
d1       1.000      FLAGS
nt       1000 il     n
ct        104 in     n
      TRANSMITTER dp      y
tn           H1 hs    nn
sfrq      399.766      PROCESSING
tof       399.7 fn    not used
tpwr       61      DISPLAY
pw       7.000 sp    -199.9
      DECOUPLER wp     4197.3
dn        C13 rf1   2121.4
dof        0 rfp   1323.2
dm        nnn rp    -155.5
decwave   W40_atb lp    2.9
dpwr       39      PLOT
dmf      29412 wc    215
      sc     35
      vs     945
      th     15
      ai cdc ph

```



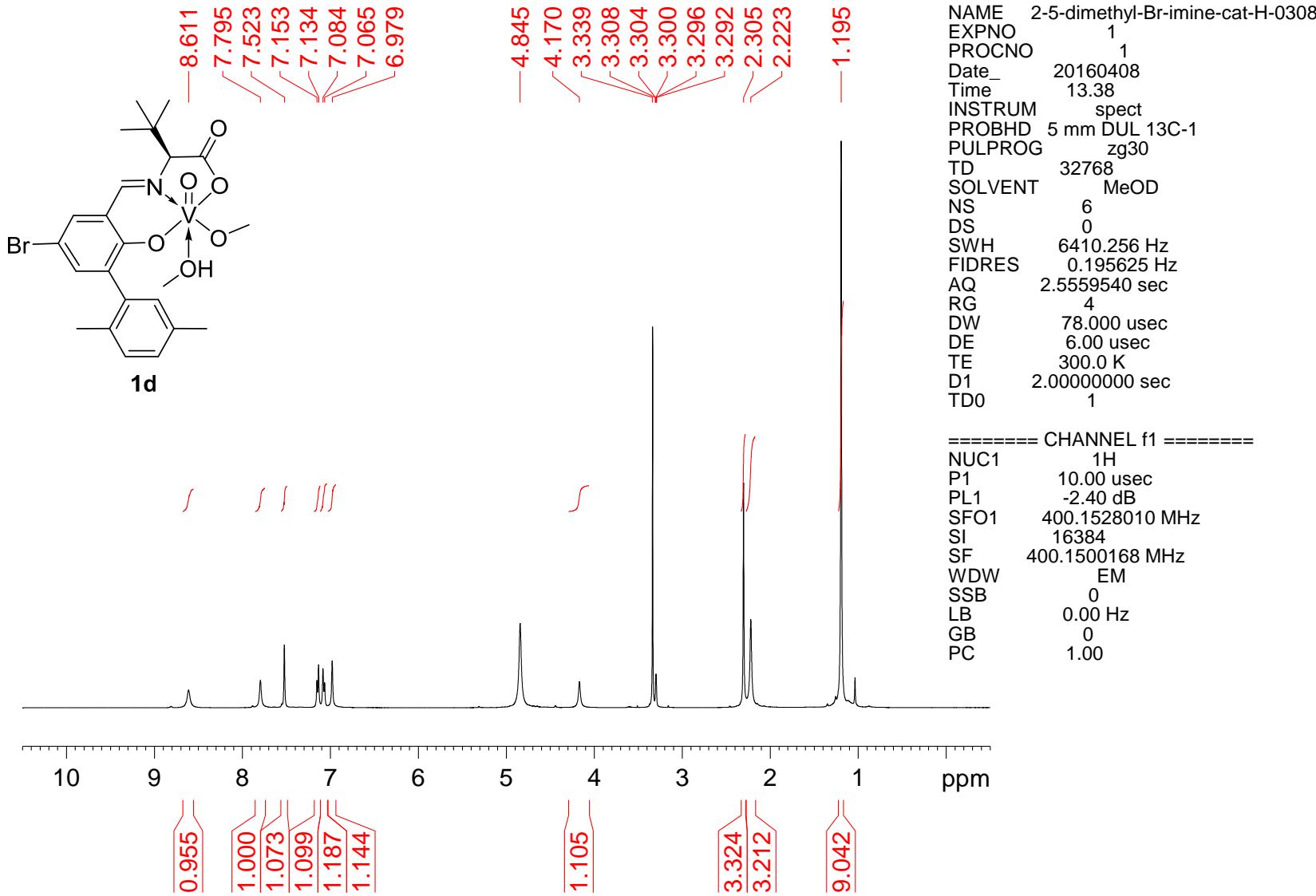


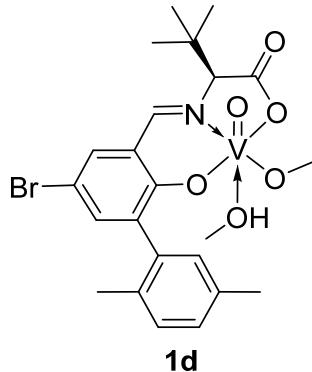
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ditertC
exp33 CARBON

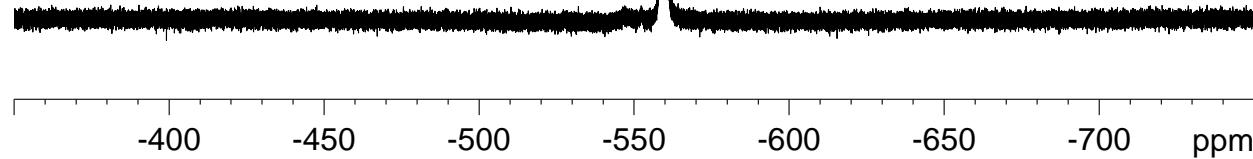
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date Jun 25 2014 satmode      n
solvent cd3od wet          n
file   exp      SPECIAL
      ACQUISITION temp    not used
sw     25510.2 gain     30
at      1.285 spin    not used
np      65536 hst      0.008
fb     17000 pw90     9.800
bs       4 alfa    10.000
d1      1.000           FLAGS
nt     100000 il      n
ct      1036 in      n
      TRANSMITTER dp      y
tn      C13 hs      nn
sfrq   100.532           PROCESSING
                  lb      3.00
tof     1530.7 fn      not used
tpwr    60           DISPLAY
pw     4.900 sp      -0.9
      DECOUPLER  wp      22114.3
dn      H1 rrf1     6480.3
dof     0 rfp      4925.5
dm      YYY rp      -91.9
decwave W lp      27.2
dpwr    41           PLOT
dmf    10695 wc      250
                  sc      0
                  vs      333
                  th      7
                  nm cdc ph

```



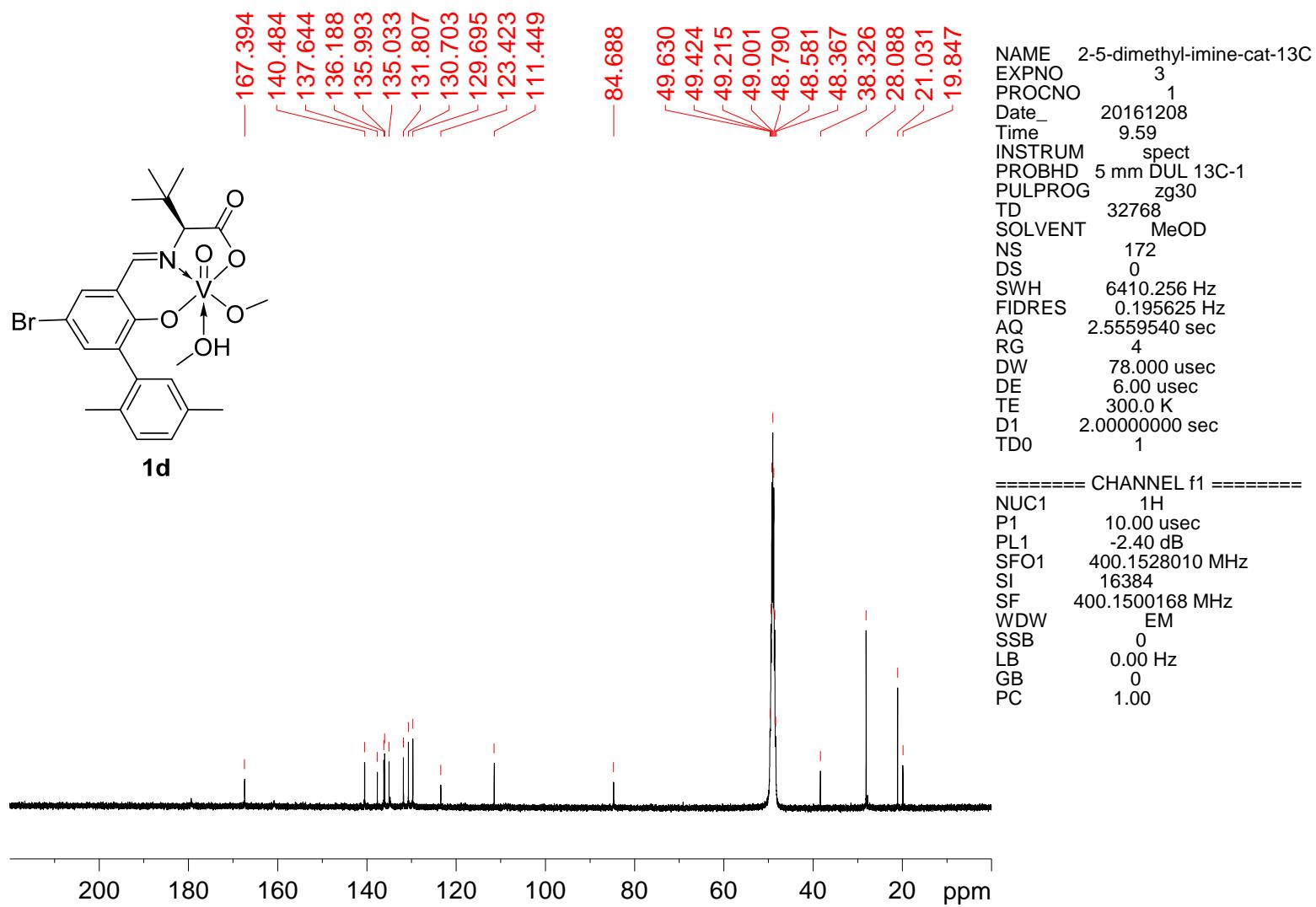


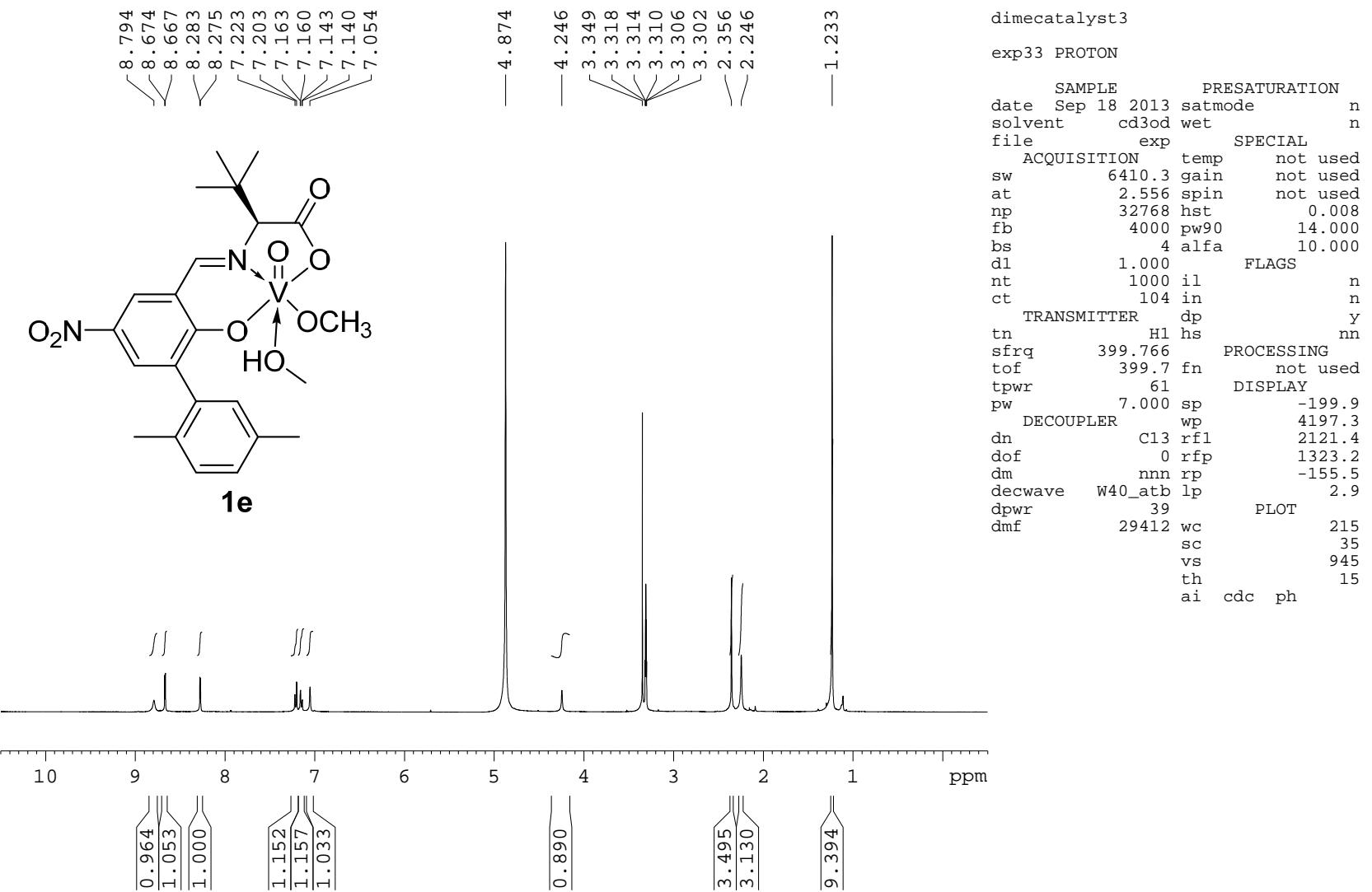
—559.384

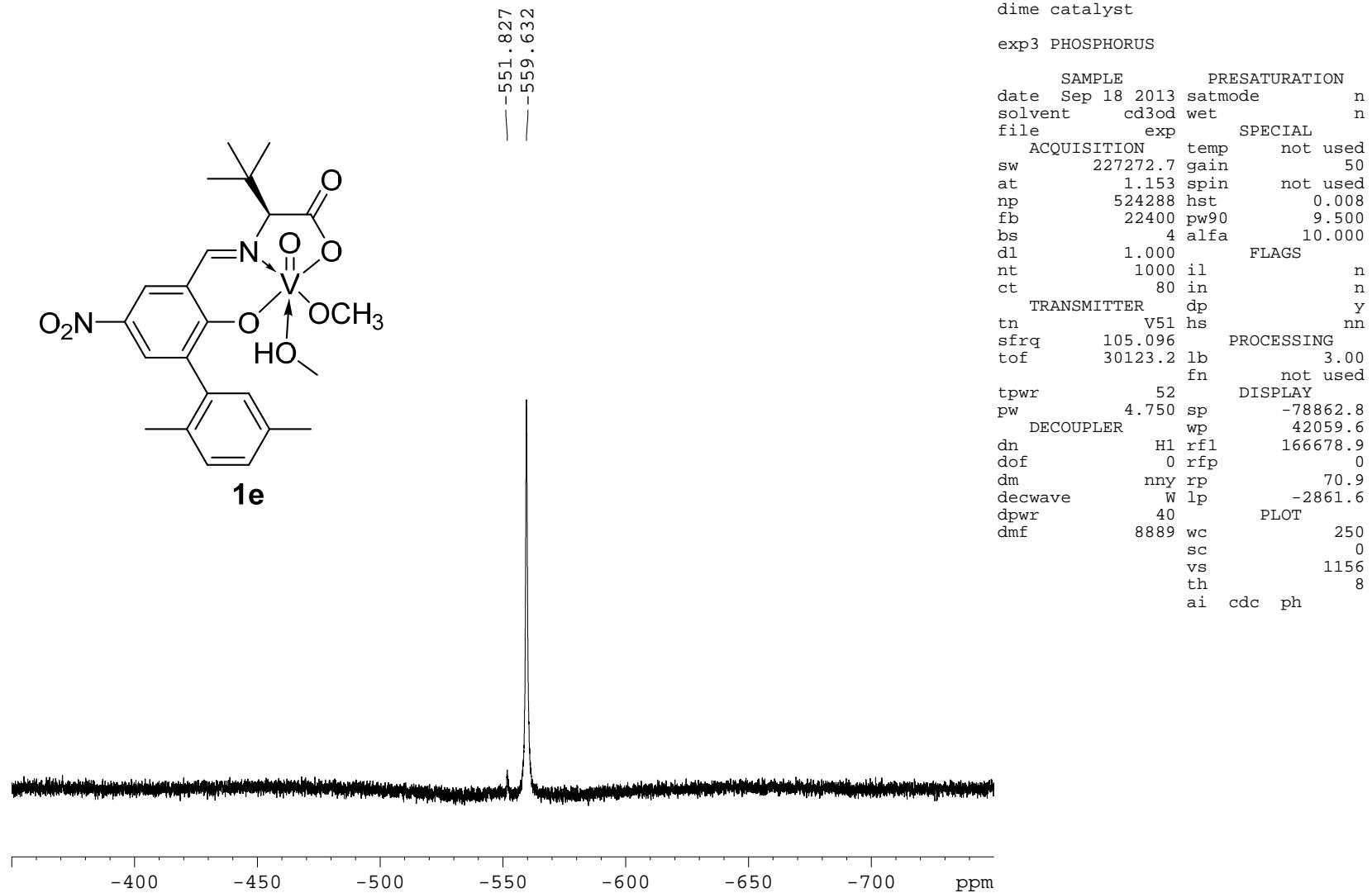
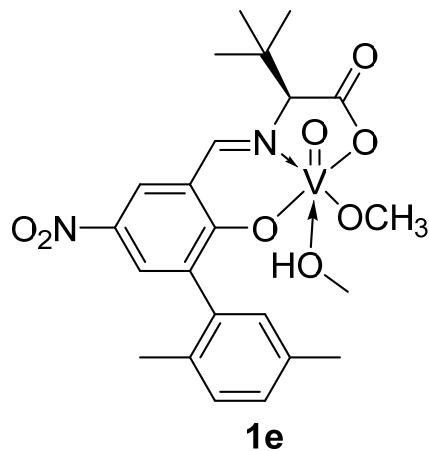


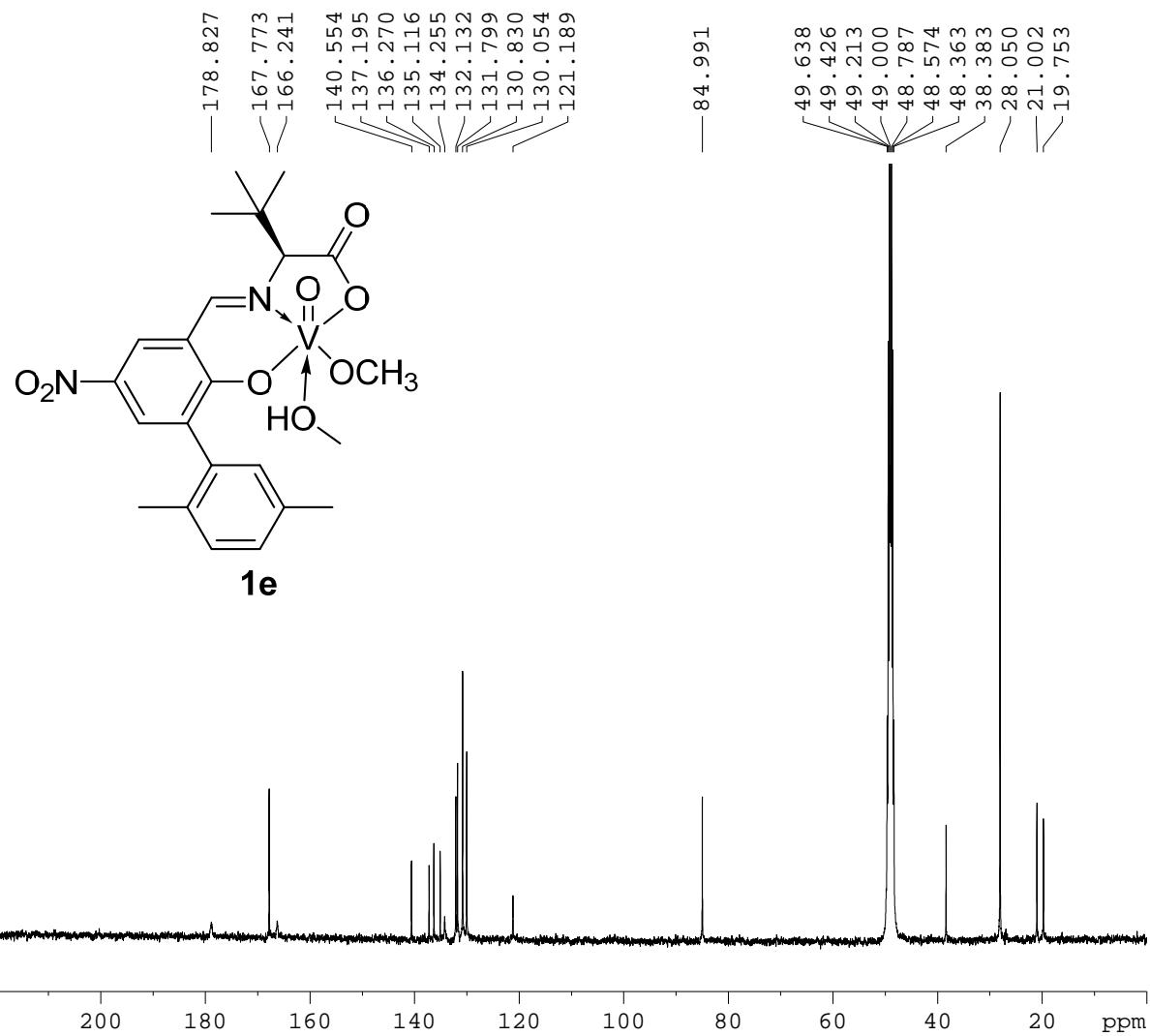
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exp2 PHOSPHORUS	PRESATURATION	
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data May 22 2016	wet	n
solvent cd3od	temp	
file exp	gain	50
ACQUISITION	spin	
sw 227272.7	hst	
at 1.153	pw90	0.008
np 524288	alfa	9.500
fb 22400		10.000
bs 4	FLAGS	
d1 1.000	il	n
nt 1000	in	n
ct 128	dp	y
TRANSMITTER	hs	nn
tn V51	PROCESSING	
sfrq 105.096	lb	2.00
tof 30123.2	fn	not used
tpwr 52	DISPLAY	
pw 4.750	sp	-78862.8
DECOUPLER	wp	42059.6
dn H1	rfl	166830.6
dof 0	rfp	0
dm nny	rp	-123.8
dmm w	lp	-2861.6
dpwr 40	PLOT	
dmf 9662	wc	250
	sc	0
	vs	3610
	th	20
	ai	
	cdc	
	ph	









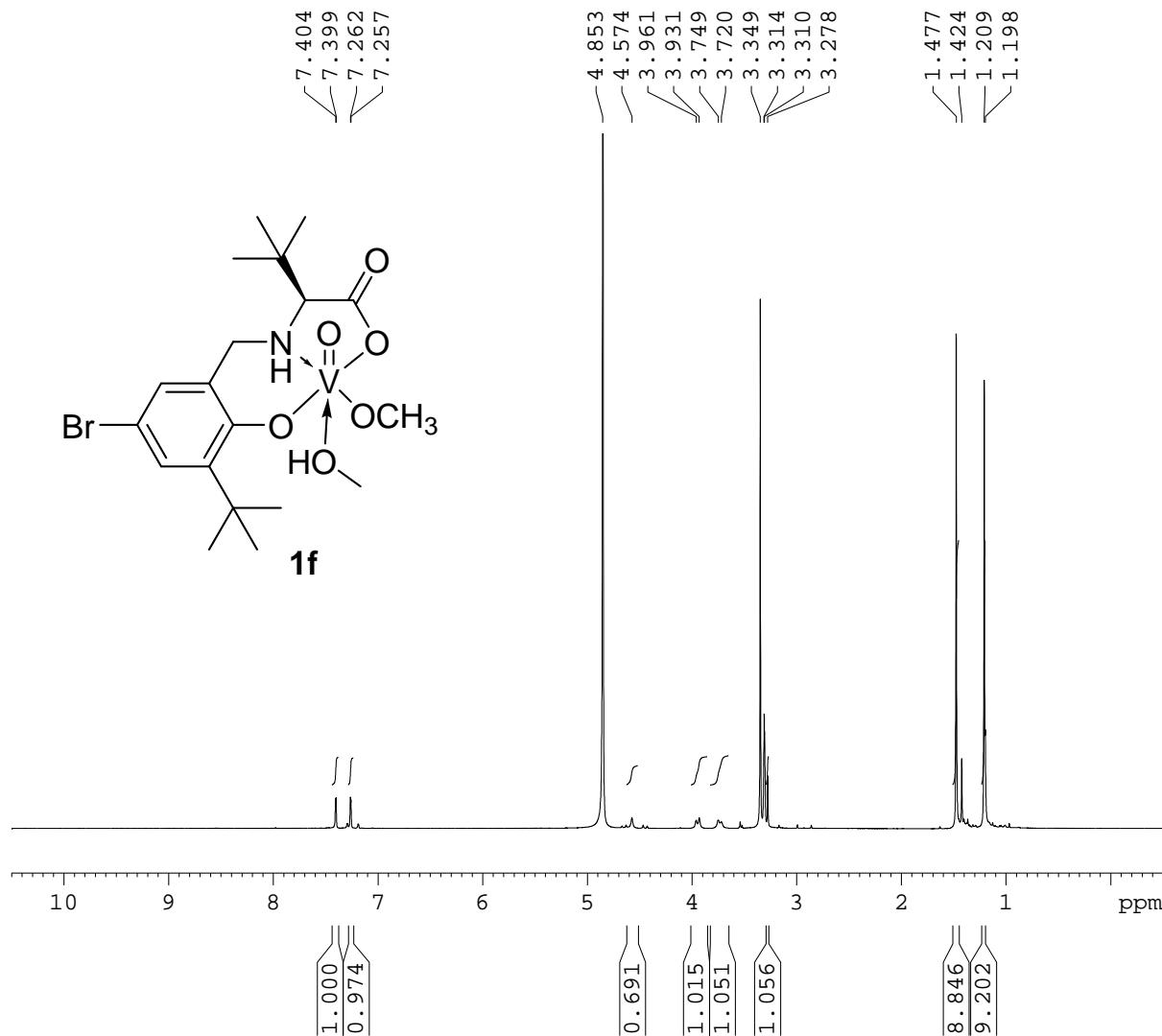
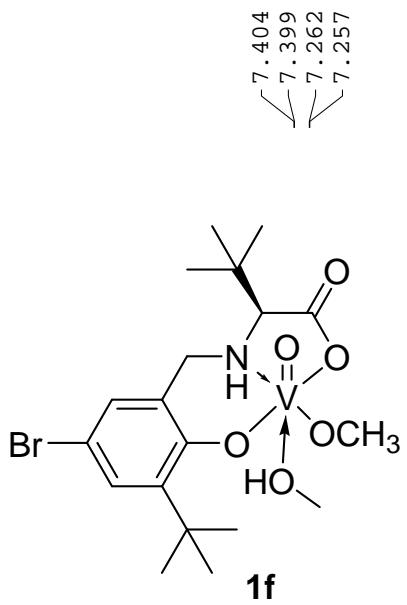
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25diMe5no2carbasalateC

exp1 CARBON

      SAMPLE      PRESATURATION
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solvent cd3od wet          n
file   exp      SPECIAL
      ACQUISITION temp    not used
sw     25510.2 gain     30
at      1.285 spin    not used
np      65536 hst      0.008
fb      17000 pw90     9.800
bs        4 alfa    10.000
d1      1.000
nt     100000 il       n
ct      4760 in       n
      TRANSMITTER dp      y
tn      C13 hs      nn
sfrq   100.532
      PROCESSING
      3.00
tof     1530.7 fn      not used
tpwr    60
      DISPLAY
      -0.6
pw      4.900 sp      22114.3
      DECOUPLER   wp      1697.8
dn      H1 rrf1
dof     0 rfp      0
dm      YYY rp      -94.9
decwave W lp      -4.5
dpwr    41
      PLOT
dmf    10695 wc      250
      sc      0
      vs      400
      th      5
      nm cdc ph

```



3tbu5brreducedH

exp1 PROTON

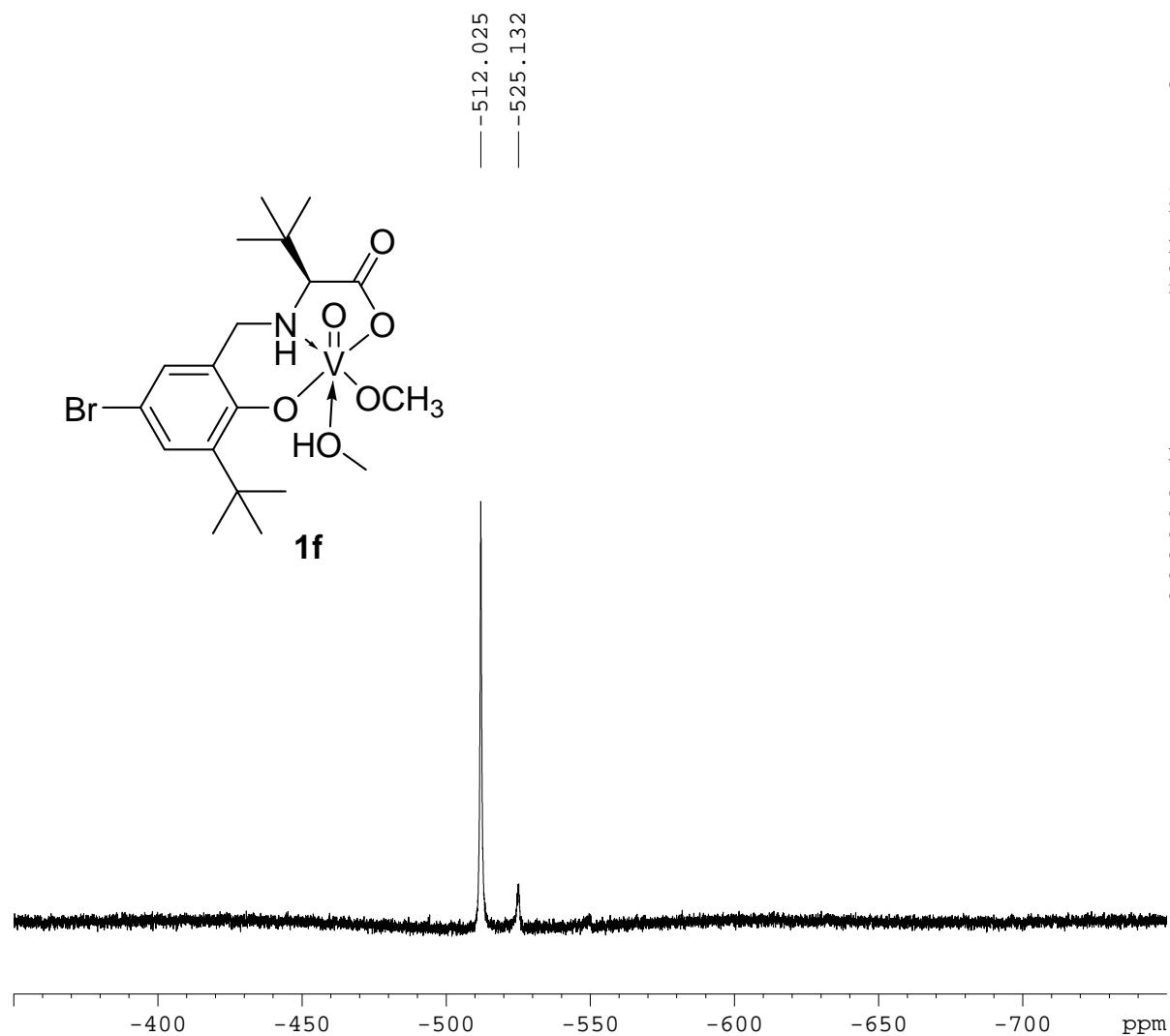
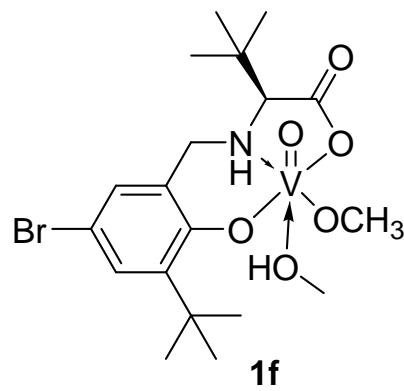
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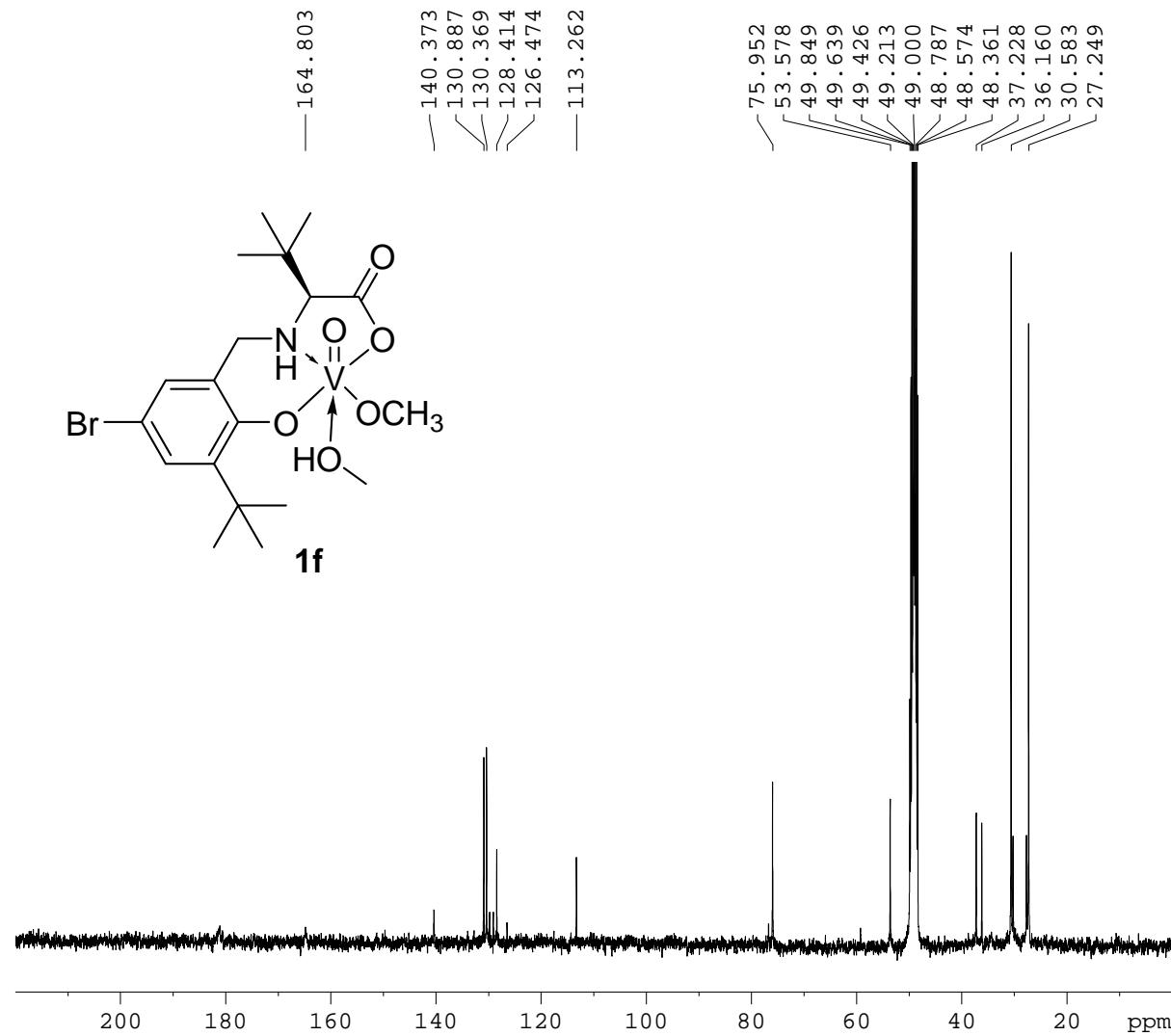
SAMPLE      PRESATURATION
date Jun 26 2014 satmode      n
solvent    cd3od wet          n
file       exp      SPECIAL
ACQUISITION temp      not used
sw        6410.3 gain      not used
at         2.556 spin      not used
np        32768 hst       0.008
fb        4000 pw90      14.000
bs         4 alfa       10.000
d1        1.000          FLAGS
nt        1000 il       n
ct         64 in       n
TRANSMITTER dp       y
tn          H1 hs      nn
sfrq     399.766      PROCESSING
tof       399.7 fn      not used
tpwr      61          DISPLAY
pw        7.000 sp      -160.4
DECOUPLER wp       3837.4
dn        C13 rf1      2121.0
dof       0 rfp       1323.2
dm       nnn rp      -158.1
decwave   W40_atb lp      0
dpwr      39          PLOT
dmf      29412 wc      215
sc        35          35
vs        500         500
th        5           5
ai cdc ph

```

exp33 PHOSPHORUS

SAMPLE PRESATURATION
date Mar 30 2014 satmode n
solvent cd3od wet n
file exp SPECIAL
ACQUISITION temp not used
sw 227272.7 gain 50
at 1.153 spin not used
np 524288 hst 0.008
fb 22400 pw90 9.500
bs 4 alfa 10.000
d1 1.000 FLAGS
nt 1000 il n
ct 192 in n
TRANSMITTER dp y
tn V51 hs nn
sfrq 105.096 PROCESSING
tof 30123.2 lb 3.00
fn not used
tpwr 52 DISPLAY
pw 4.750 sp -78862.8
DECOUPLER wp 42059.6
dn H1 rfp 166678.9
dof 0 rfp 0
dm nny rp 125.2
decwave W lp -2861.6
dpwr 40 PLOT
dmf 8889 wc 250
sc 0
vs 1832
th 8
ai cdc ph





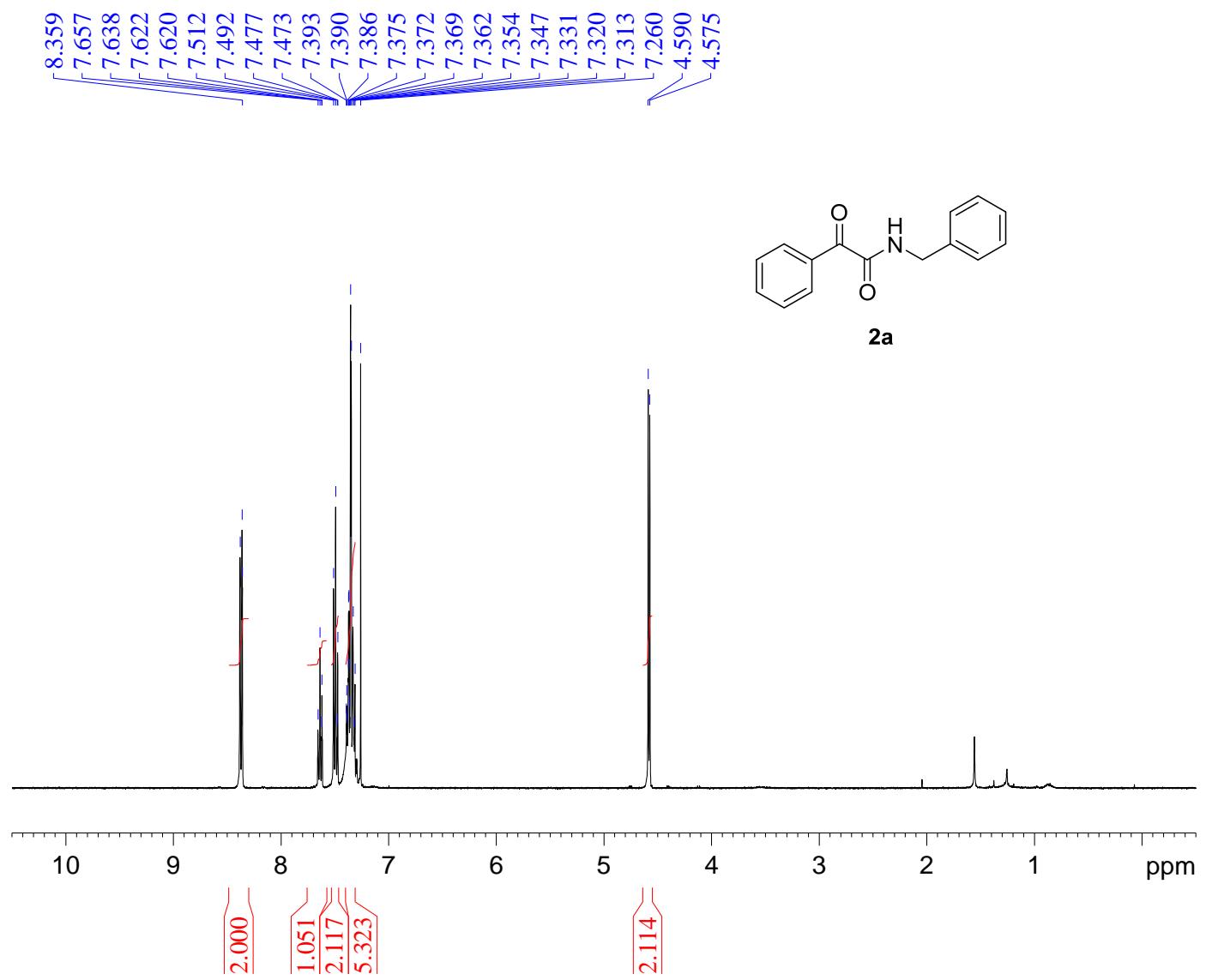
3tbu5brreducedC

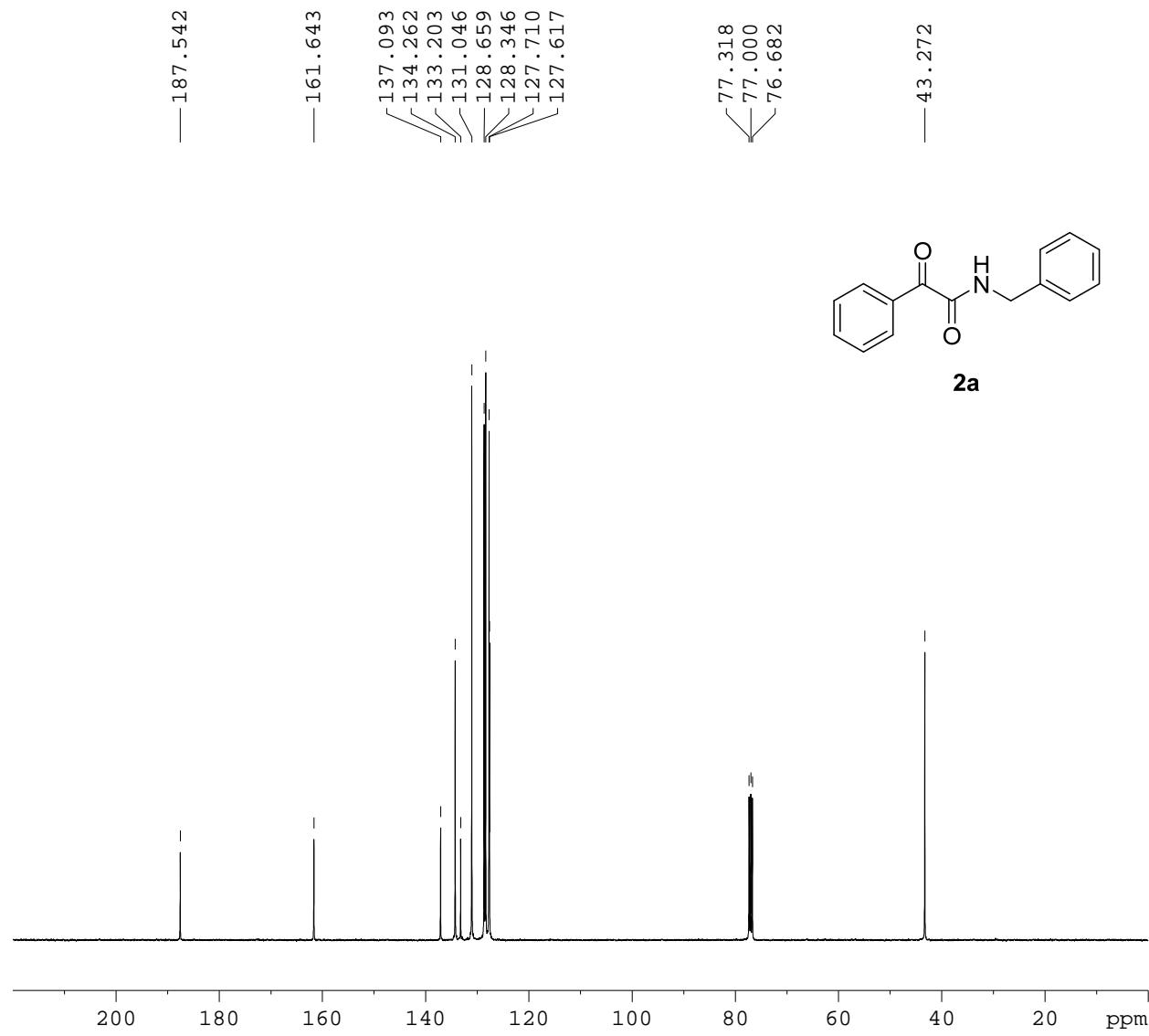
exp33 CARBON

```

SAMPLE      PRESATURATION
date Jun 26 2014 satmode      n
solvent cd3od wet          n
file   exp      SPECIAL
ACQUISITION temp      not used
sw      25510.2 gain      30
at      1.285 spin      not used
np      65536 hst       0.008
fb      17000 pw90      9.800
bs      4 alfa      10.000
d1      1.000
nt      100000 il       n
ct      15396 in       n
TRANSMITTER dp       y
tn      C13 hs      nn
sfrq    100.532
PROCESSING
tof      1530.7 fn      not used
tpwr    60
DISPLAY
pw      4.900 sp      -0.9
DECOUPLER wp      22114.3
dn      H1 rfl      6478.8
dof     0 rfp      4925.5
dm      YYY rp      -90.0
decwave W lp      11.6
dpwr    41
PLOT
dmf     10695 wc      250
sc      0
vs      2252
th      13
nm cdc ph

```





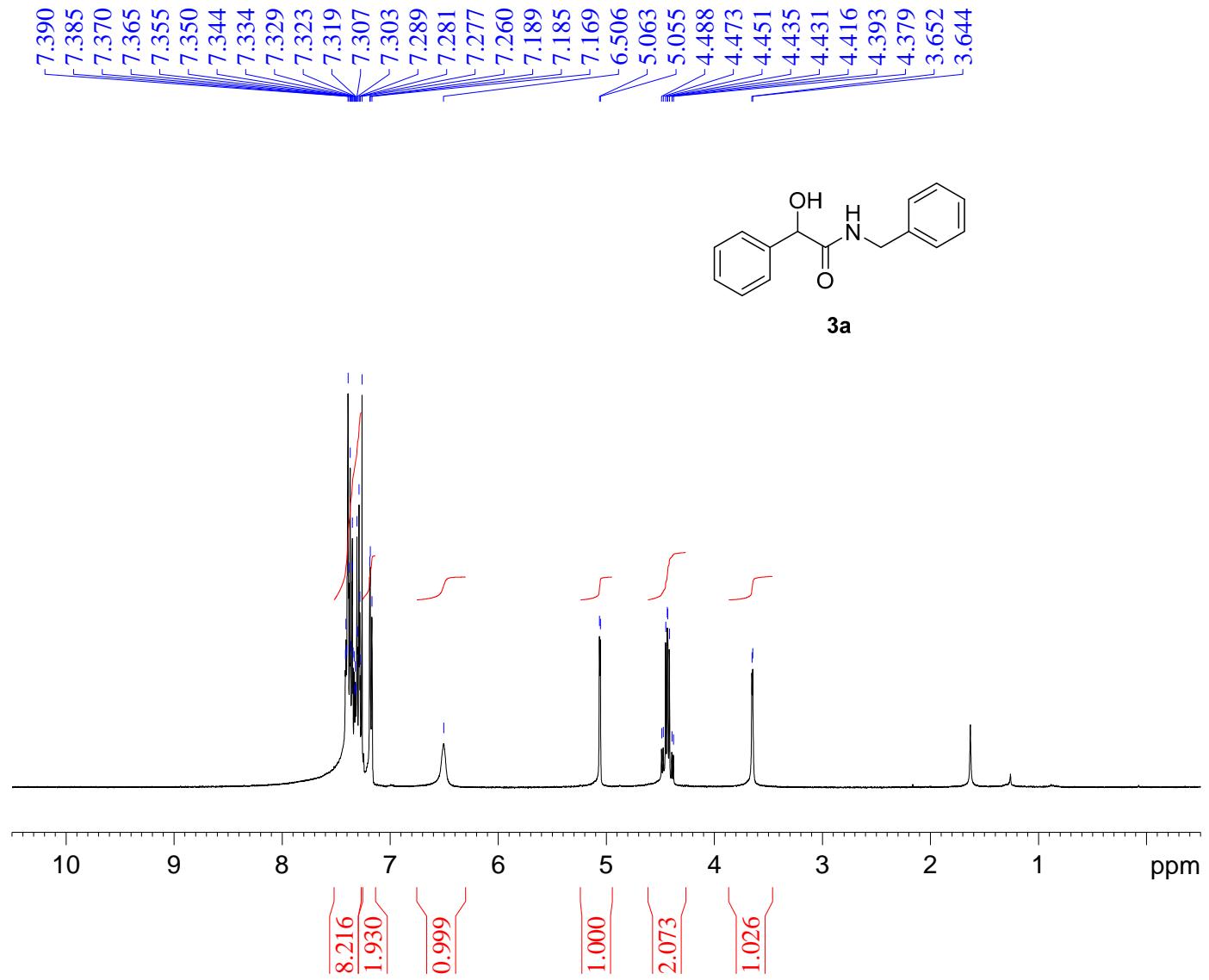
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NAME           13C
EXPNO          2
PROCNO         1
Date_ 20150110
Time   19.41
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD      65536
SOLVENT   CDCl3
NS       1363
DS        0
SWH     22727.273 Hz
FIDRES   0.346791 Hz
AQ      1.4418420 sec
RG        57
DW      22.000 usec
DE       6.00 usec
TE      300.0 K
D1      2.0000000 sec
d11     0.0300000 sec
DELTA    1.89999998 sec
TDO      1

===== CHANNEL f1 =====
NUC1           13C
P1            9.70 usec
PL1          -0.50 dB
SFO1      100.6288660 MHz

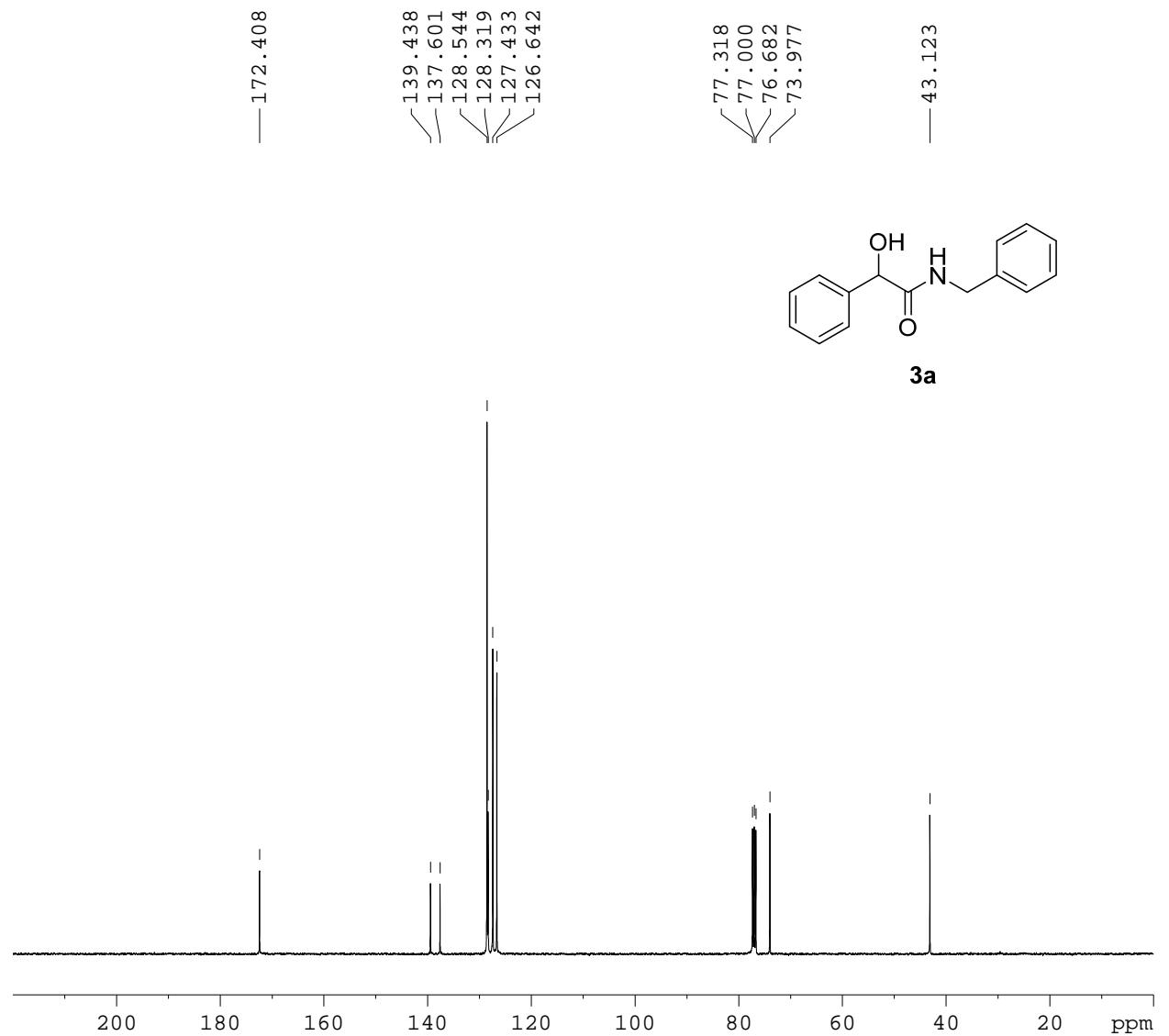
===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2           1H
PCPD2        90.00 usec
PL2          -2.40 dB
PL12         15.10 dB
PL13         18.10 dB
SFO2      400.1516010 MHz
SI            32768
SF      100.6178159 MHz
WDW           EM
SSB            0
LB            3.00 Hz
GB            0
PC         1.00

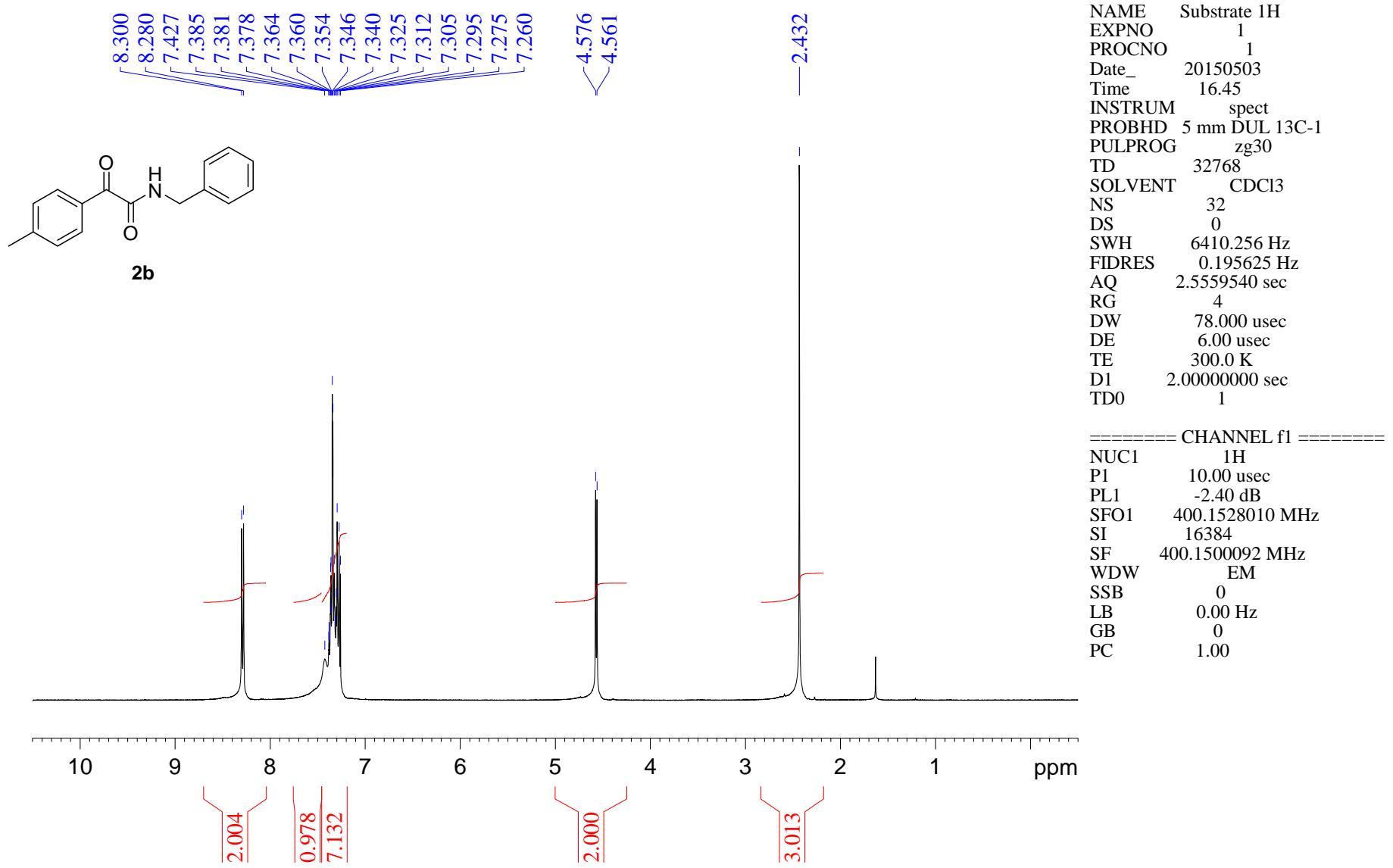
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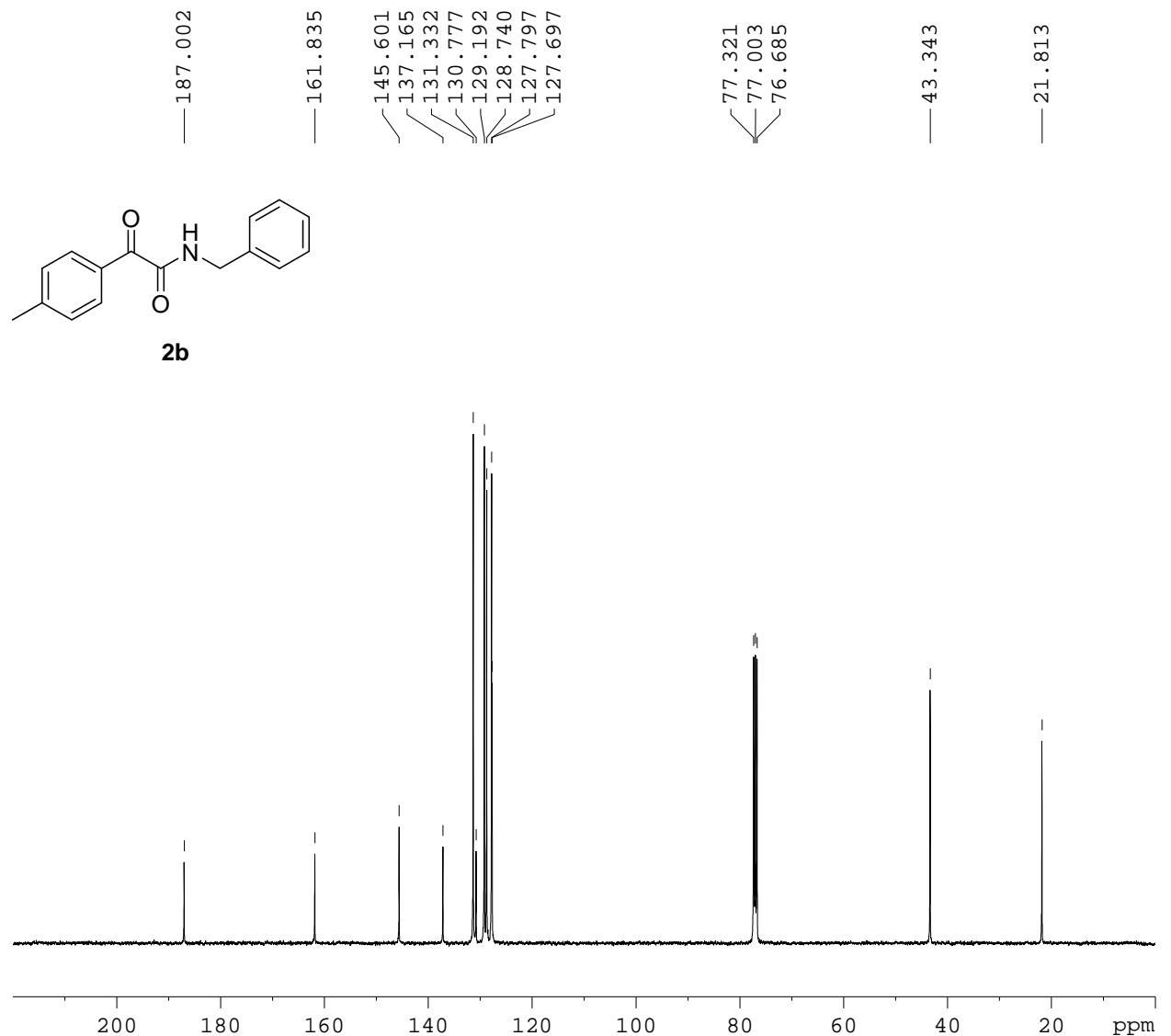


NAME	Substrate
EXPNO	7
PROCNO	1
Date_	20150702
Time	18.27
INSTRUM	spect
PROBHD	5 mm DUL
PULPROG	13C-1 zg30
TD	32768
SOLVENT	CDCl3
NS	48
DS	0
SWH	6410.256 Hz
FIDRES	0.195625 Hz
AQ	2.5559540 sec
RG	4
DW	78.000 usec
DE	6.00 usec
TE	300.0 K
D1	2.0000000 sec
TD0	1

===== CHANNEL f1 =====
NUC1 1H
P1 10.00 usec
PL1 -2.40 dB
SFO1 400.1528010 MHz
SI 16384
SF 400.1500088 MHz
WDW EM
SSB 0
LB 0.00 Hz
GB 0
PC 1.00



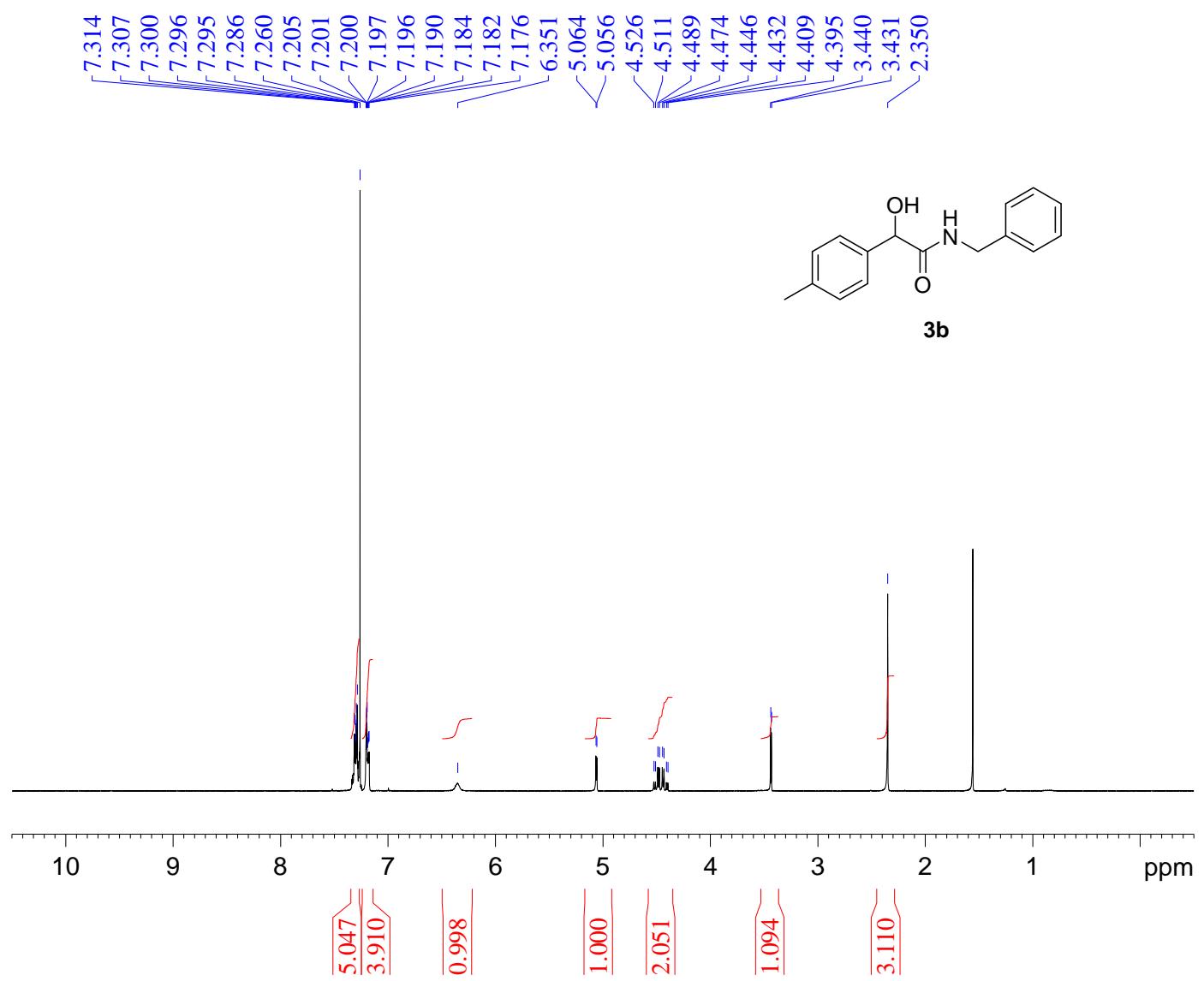




NAME Substrate 13C
 EXPNO 1
 PROCNO 1
 Date_ 20150503
 Time 16.53
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgppg30
 TD 65536
 SOLVENT CDCl3
 NS 1512
 DS 0
 SWH 22727.273 Hz
 FIDRES 0.346791 Hz
 AQ 1.4418420 sec
 RG 57
 DW 22.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 DELTA 1.8999999 sec
 TDO 1

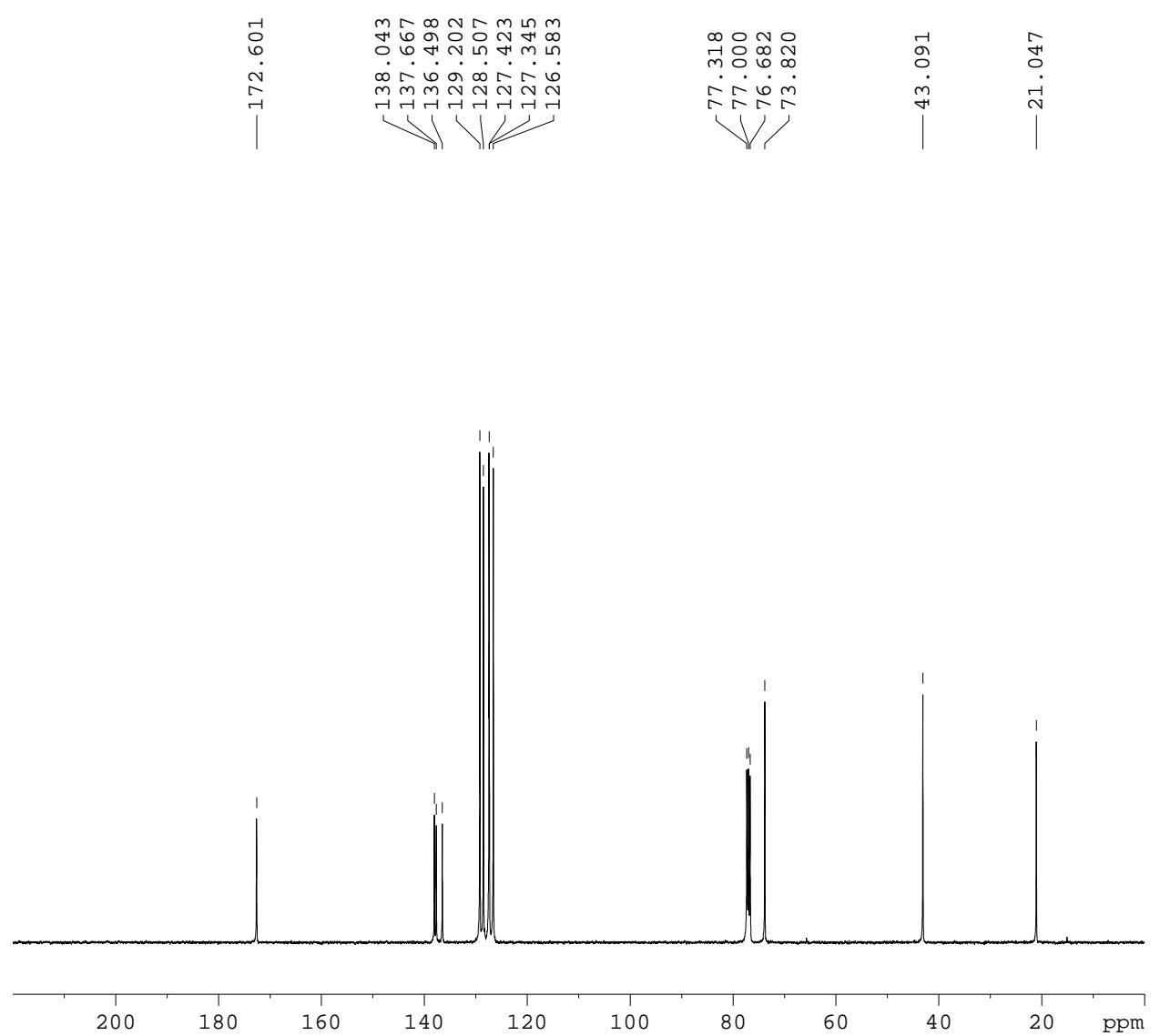
===== CHANNEL f1 =====
 NUC1 13C
 P1 9.70 usec
 PL1 -0.50 dB
 SFO1 100.6288660 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 -2.40 dB
 PL12 15.10 dB
 PL13 18.10 dB
 SFO2 400.1516010 MHz
 SI 32768
 SF 100.6178076 MHz
 WDW EM
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.00



NAME 20151029
EXPNO 1
PROCNO 1
Date_ 20151029
Time 13.48
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 32768
SOLVENT CDCl₃
NS 36
DS 0
SWH 6410.256 Hz
FIDRES 0.195625 Hz
AQ 2.5559540 sec
RG 4
DW 78.000 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
TD0 1

===== CHANNEL f1 ======
NUC1 1H
P1 10.00 usec
PL1 -2.40 dB
SFO1 400.1528010 MHz
SI 16384
SF 400.1500088 MHz
WDW EM
SSB 0
LB 0.00 Hz
GB 0
PC 1.00



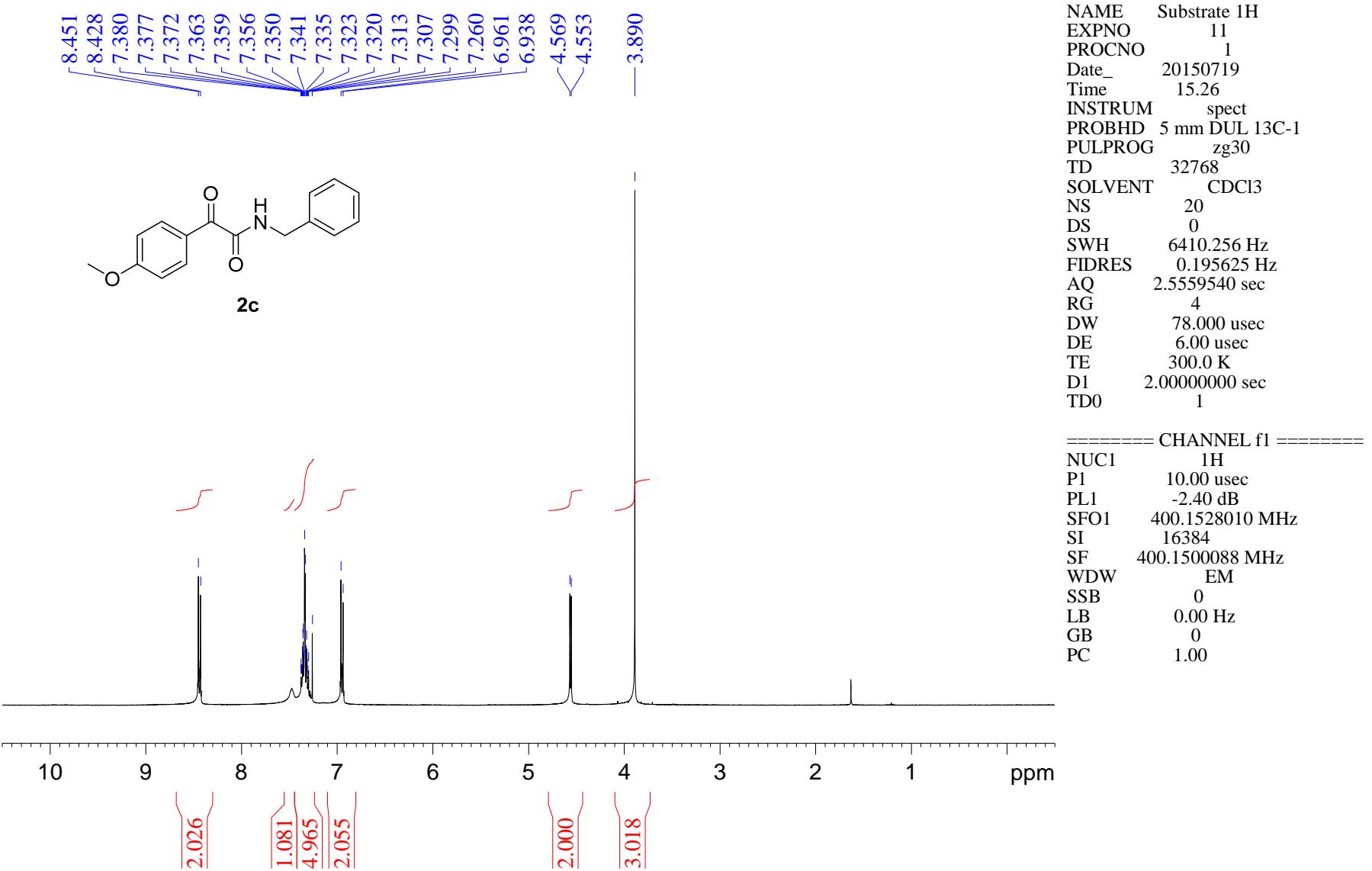
NAME 13C
 EXPNO 6
 PROCNO 1
 Date_ 20150321
 Time 17.40
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 1520
 DS 0
 SWH 22727.273 Hz
 FIDRES 0.346791 Hz
 AQ 1.4418420 sec
 RG 57
 DW 22.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 DELTA 1.8999998 sec
 TDO 1

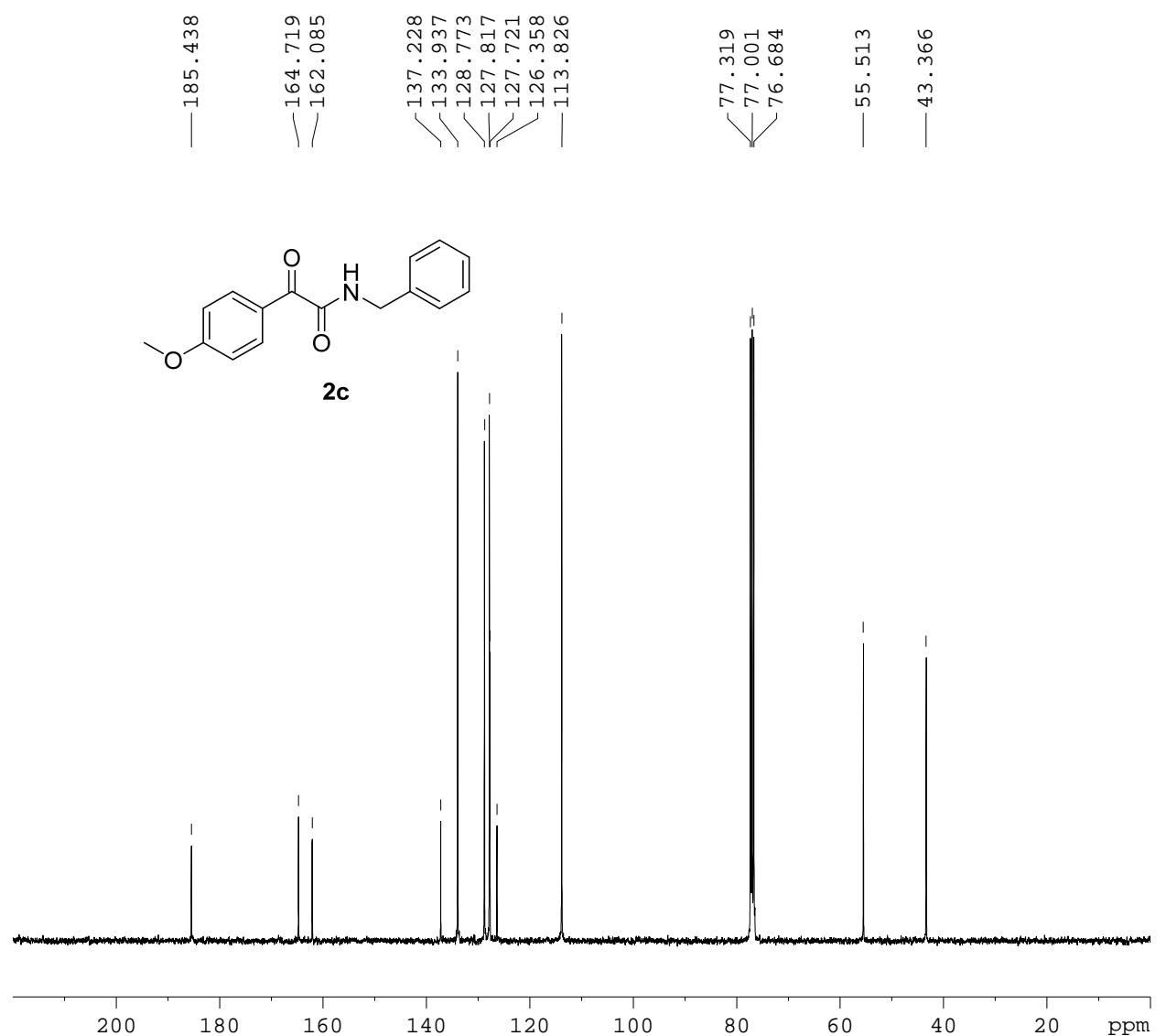
===== CHANNEL f1 =====

NUC1 13C
 P1 9.70 usec
 PL1 -0.50 dB
 SFO1 100.6288660 MHz

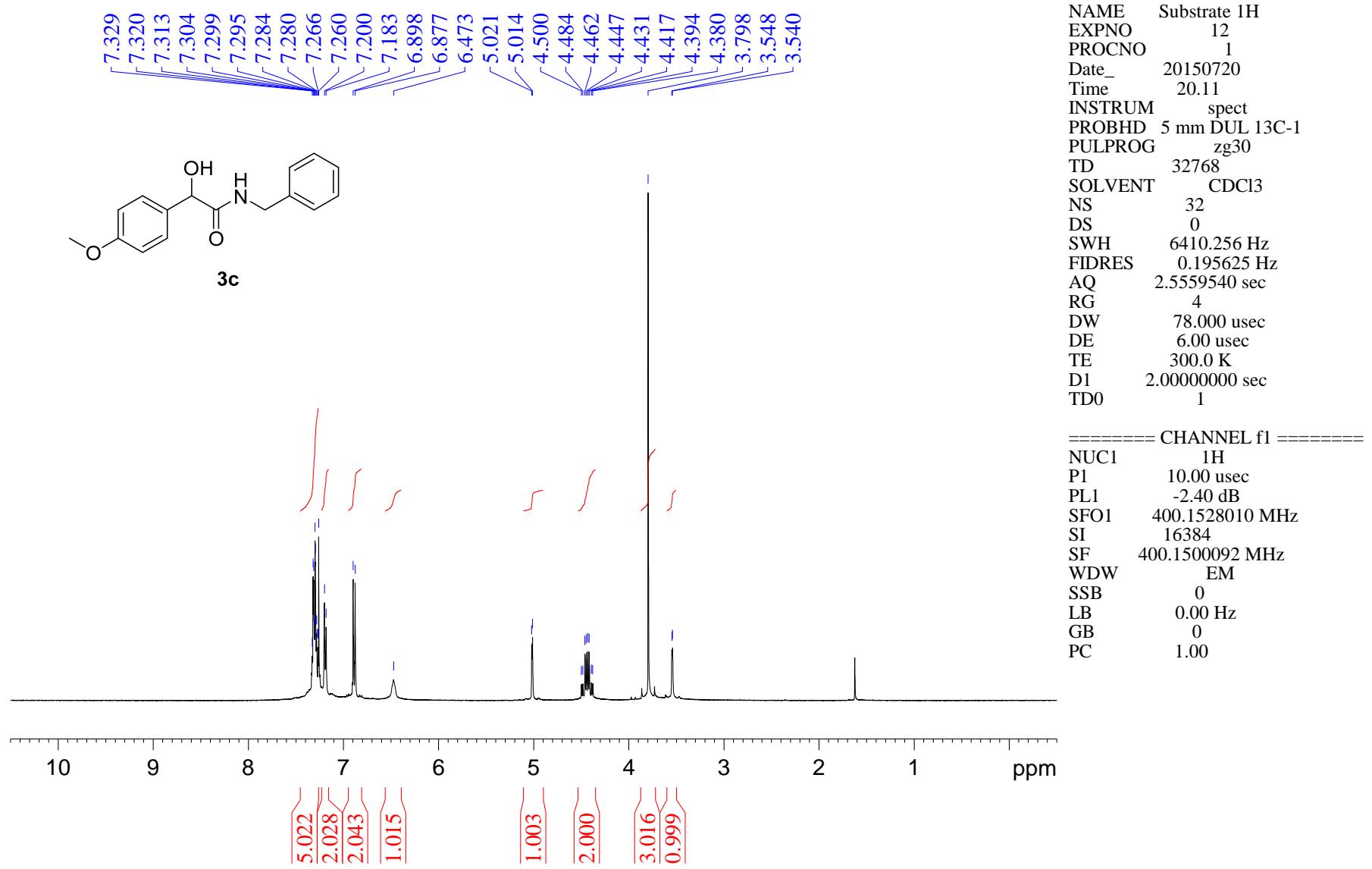
===== CHANNEL f2 =====

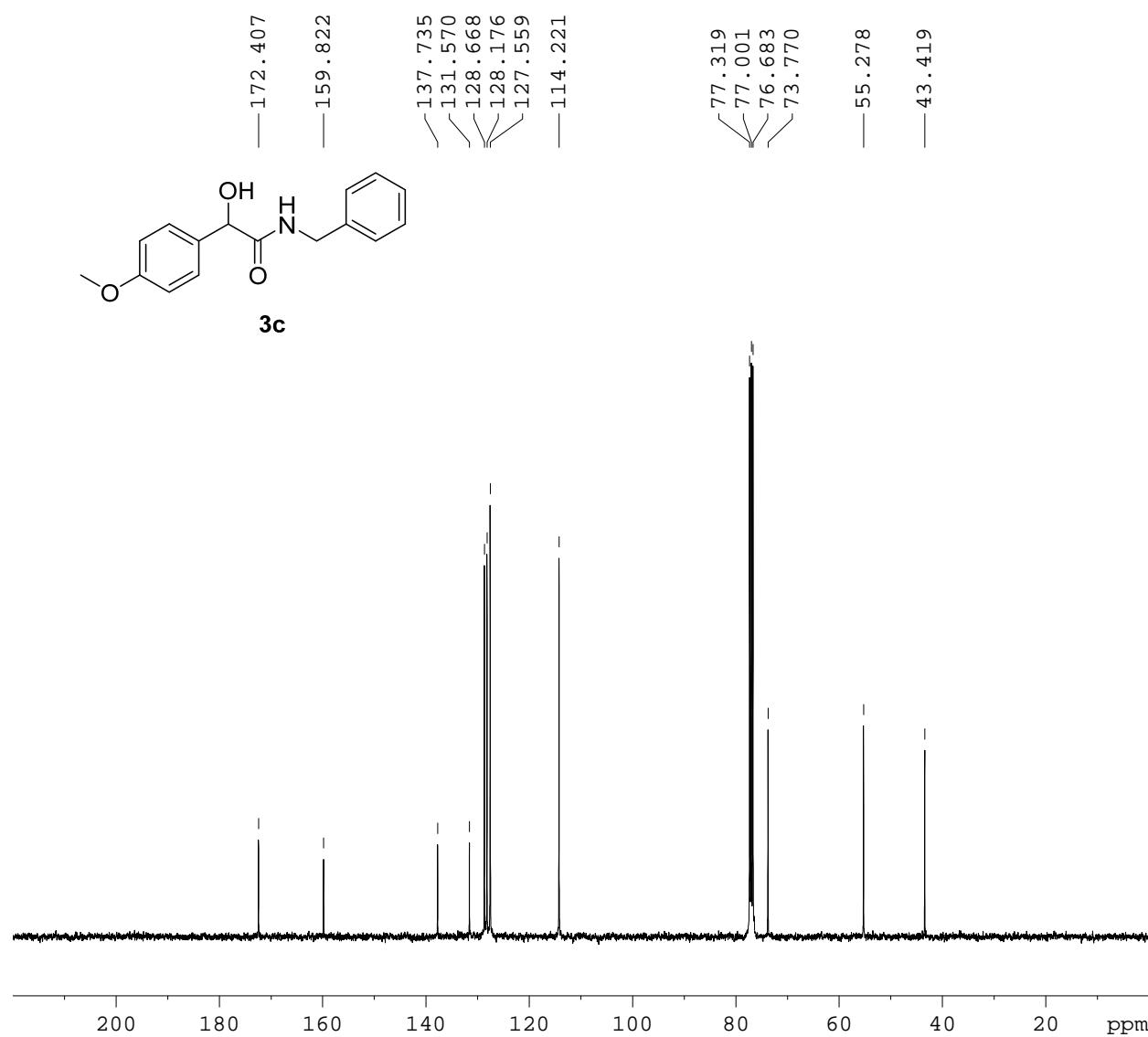
CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 -2.40 dB
 PL12 15.10 dB
 PL13 18.10 dB
 SFO2 400.1516010 MHz
 SI 32768
 SF 100.6178151 MHz
 WDW EM
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.00

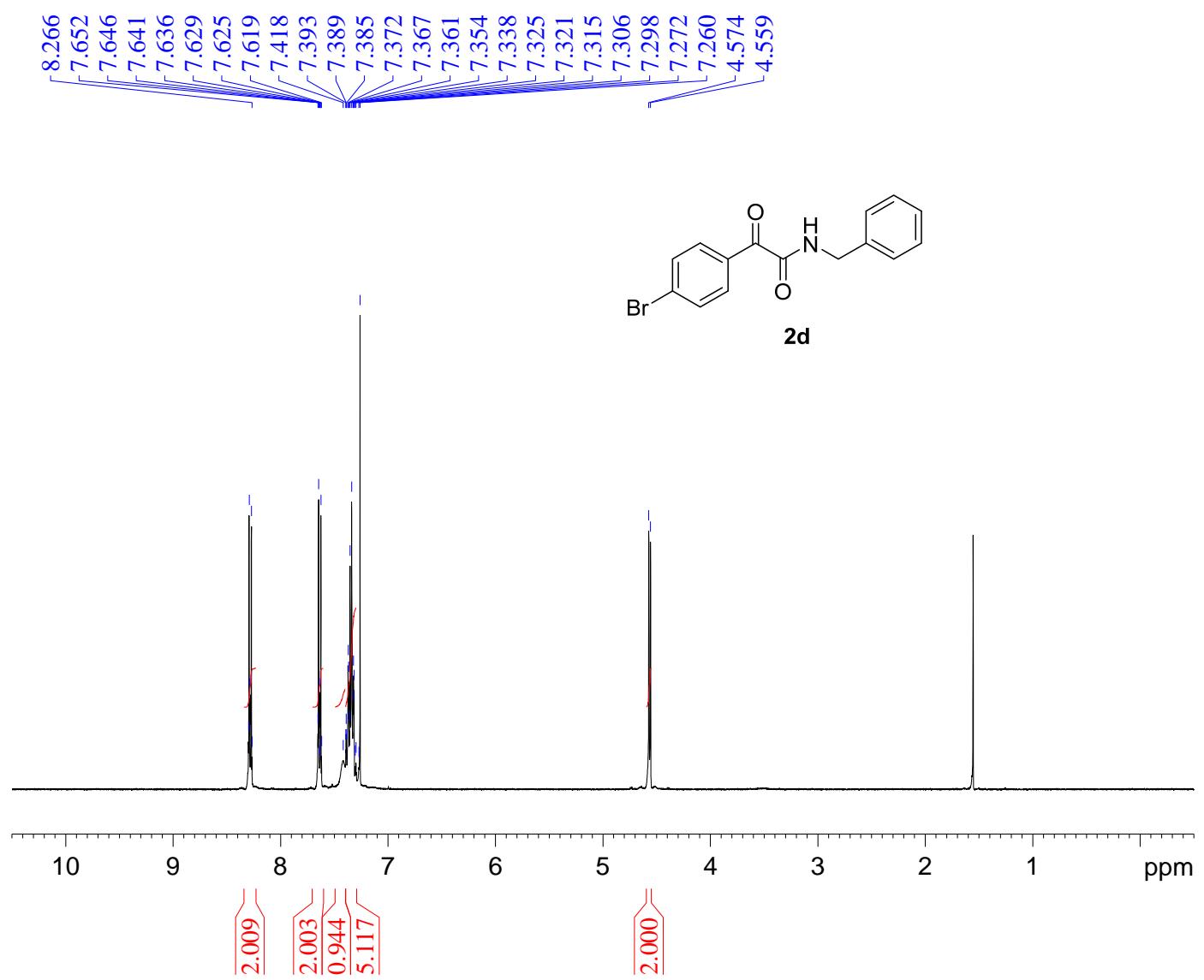




NAME	Substrate	¹³ C
EXPNO		11
PROCNO		1
Date_	20150719	
Time	15.31	
INSTRUM	spect	
PROBHD	5 mm DUL	13C-1
PULPROG	zgpg30	
TD	65536	
SOLVENT	CDCl ₃	
NS	1191	
DS	0	
SWH	22727.273	Hz
FIDRES	0.346791	Hz
AQ	1.4418420	sec
RG	57	
DW	22.000	usec
DE	6.00	usec
TE	300.0	K
D1	2.00000000	sec
d11	0.03000000	sec
DELTA	1.89999998	sec
TDO	1	
===== CHANNEL f1 =====		
NUC1	¹³ C	
P1	9.70	usec
PL1	-0.50	dB
SFO1	100.6288660	MHz
===== CHANNEL f2 =====		
CPDPRG2	waltz16	
NUC2	¹ H	
PCPD2	90.00	usec
PL2	-2.40	dB
PL12	15.10	dB
PL13	18.10	dB
SFO2	400.1516010	MHz
SI	32768	
SF	100.6178033	MHz
WDW	EM	
SSB	0	
LB	3.00	Hz
GB	0	
PC	1.00	



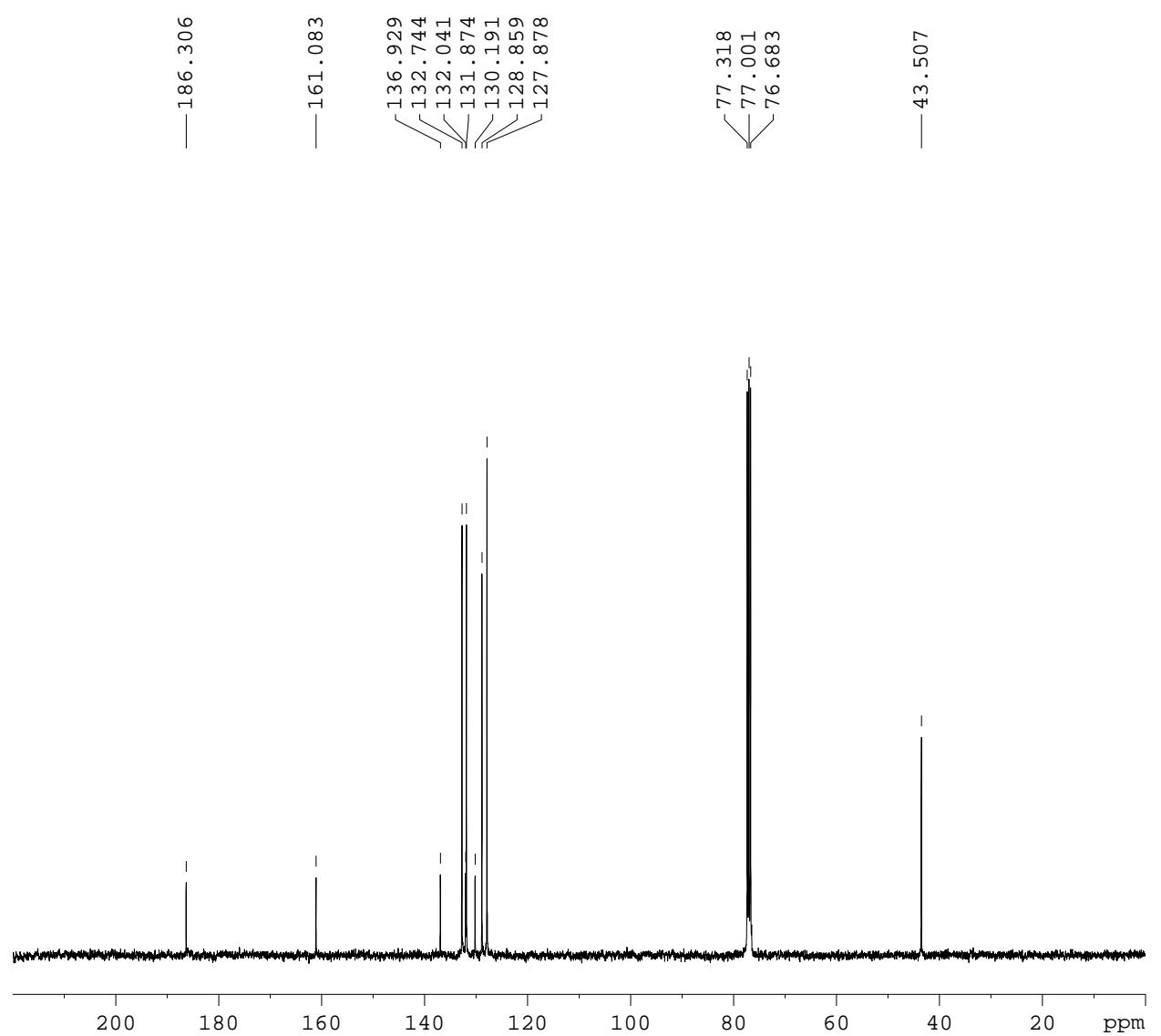




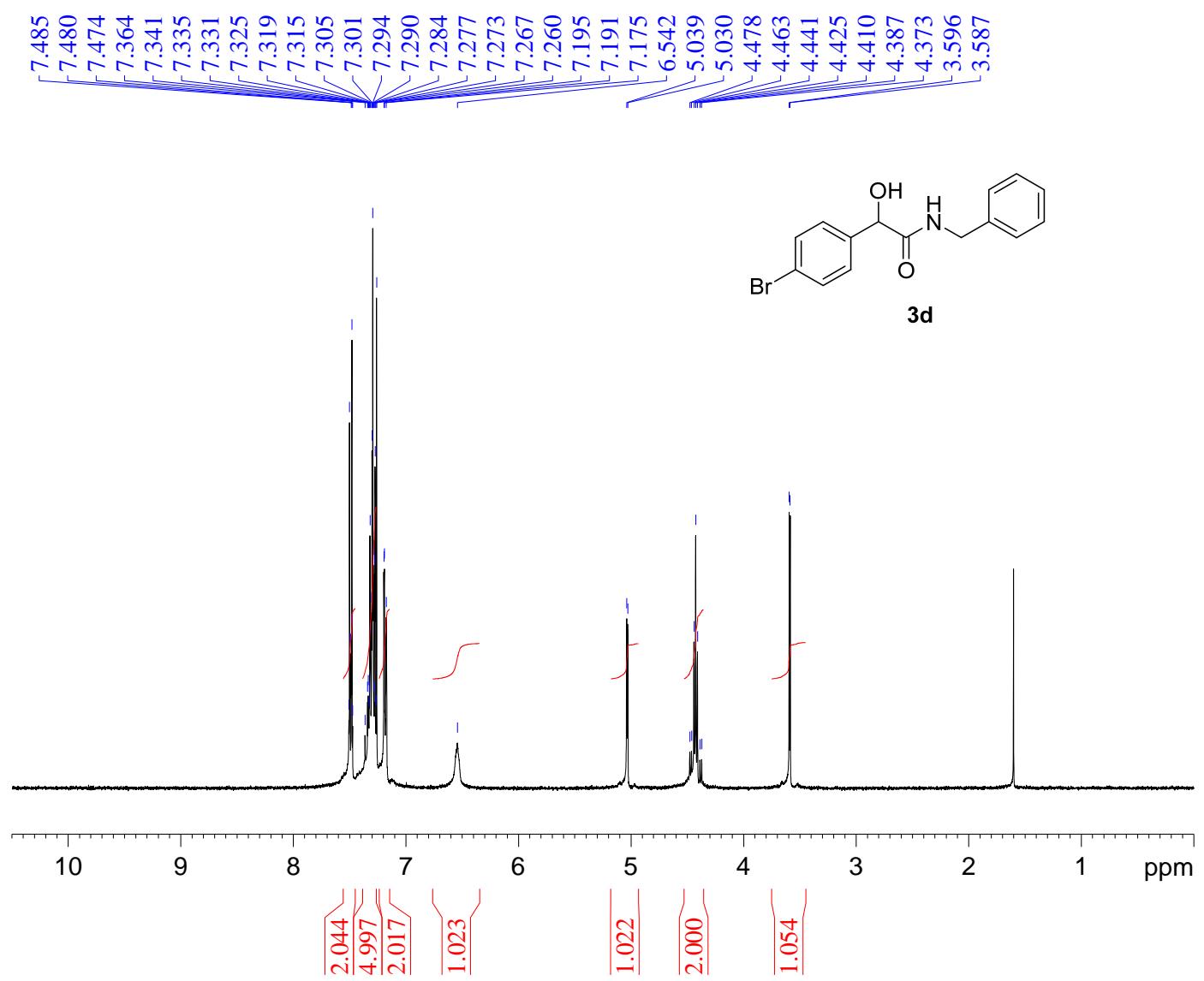
NAME Substrate 1H
EXPNO 14
PROCNO 1
Date_ 20150723
Time 20.25
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 32768
SOLVENT CDCl₃
NS 44
DS 0
SWH 6410.256 Hz
FIDRES 0.195625 Hz
AQ 2.5559540 sec
RG 4
DW 78.000 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
TD0 1

===== CHANNEL f1 ======

Parameter	Value
NUC1	1H
P1	10.00 usec
PL1	-2.40 dB
SFO1	400.1528010 MHz
SI	16384
SF	400.1500088 MHz
WDW	EM
SSB	0
LB	0.00 Hz
GB	0
PC	1.00

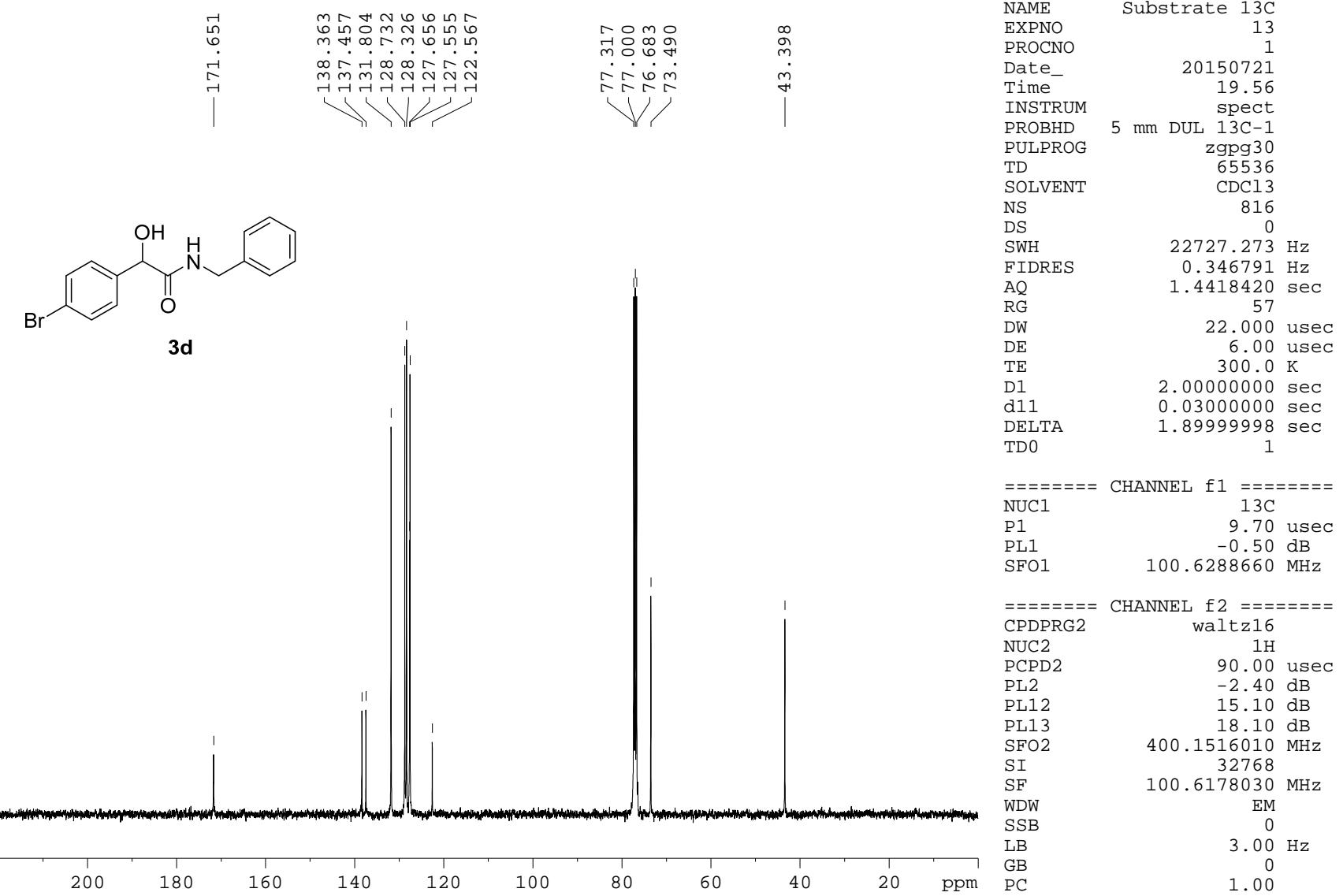


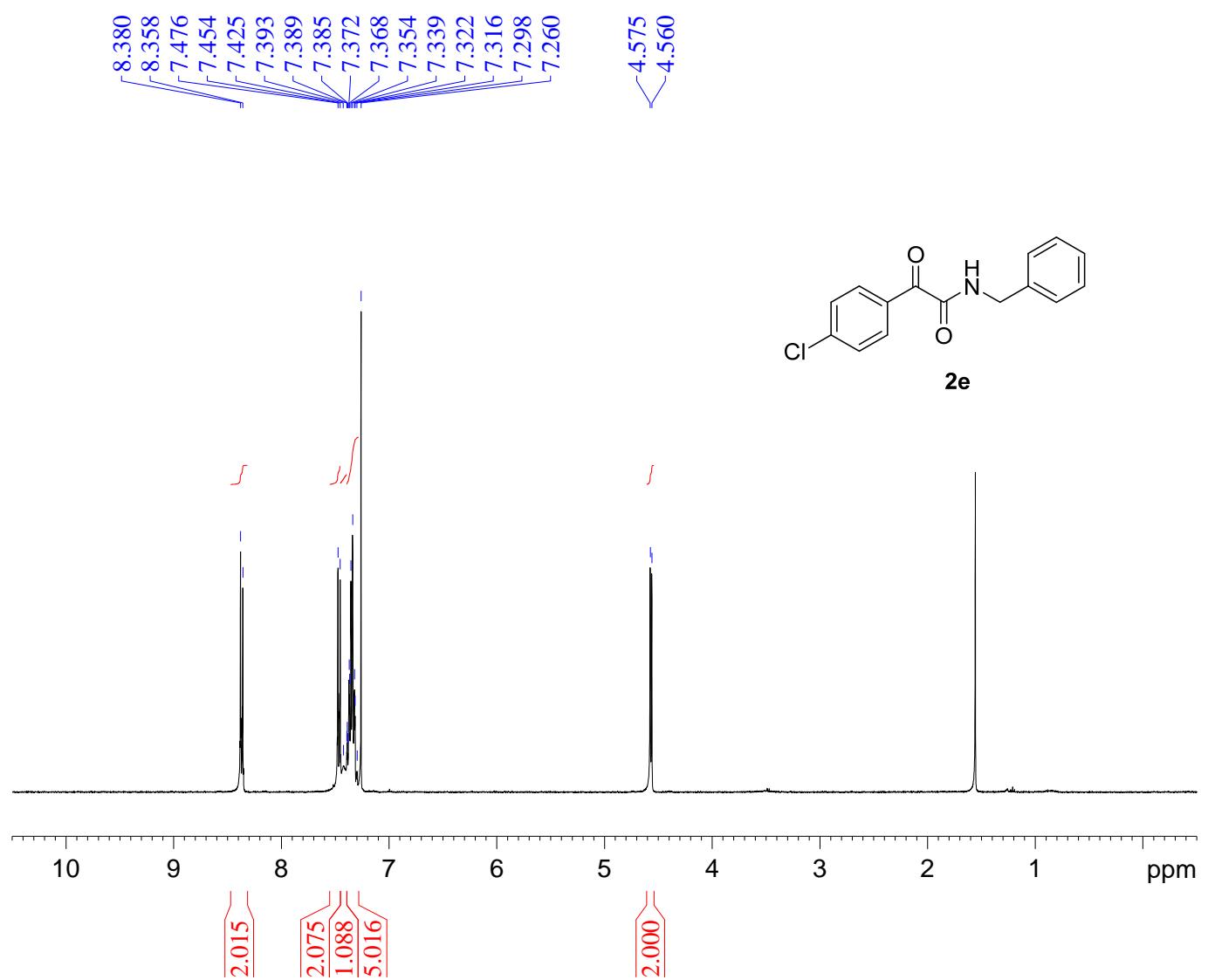
NAME	Substrate	¹³ C
EXPNO		14
PROCNO		1
Date_	20150723	
Time	20.31	
INSTRUM	spect	
PROBHD	5 mm DUL	¹³ C-1
PULPROG	zgpg30	
TD	65536	
SOLVENT	CDC13	
NS	496	
DS	0	
SWH	22727.273	Hz
FIDRES	0.346791	Hz
AQ	1.4418420	sec
RG	57	
DW	22.000	usec
DE	6.00	usec
TE	300.0	K
D1	2.00000000	sec
d11	0.03000000	sec
DELTA	1.89999998	sec
TDO	1	
===== CHANNEL f1 =====		
NUC1	13C	
P1	9.70	usec
PL1	-0.50	dB
SFO1	100.6288660	MHz
===== CHANNEL f2 =====		
CPDPRG2	waltz16	
NUC2	1H	
PCPD2	90.00	usec
PL2	-2.40	dB
PL12	15.10	dB
PL13	18.10	dB
SFO2	400.1516010	MHz
SI	32768	
SF	100.6178020	MHz
WDW	EM	
SSB	0	
LB	3.00	Hz
GB	0	
PC	1.00	



NAME Substrate 1H
 EXPNO 13
 PROCNO 1
 Date_ 20150721
 Time 19.52
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 32768
 SOLVENT CDCl₃
 NS 16
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.195625 Hz
 AQ 2.5559540 sec
 RG 4
 DW 78.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 TD0 1

===== CHANNEL f1 ======
 NUC1 1H
 P1 10.00 usec
 PL1 -2.40 dB
 SFO1 400.1528010 MHz
 SI 16384
 SF 400.1500088 MHz
 WDW EM
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00



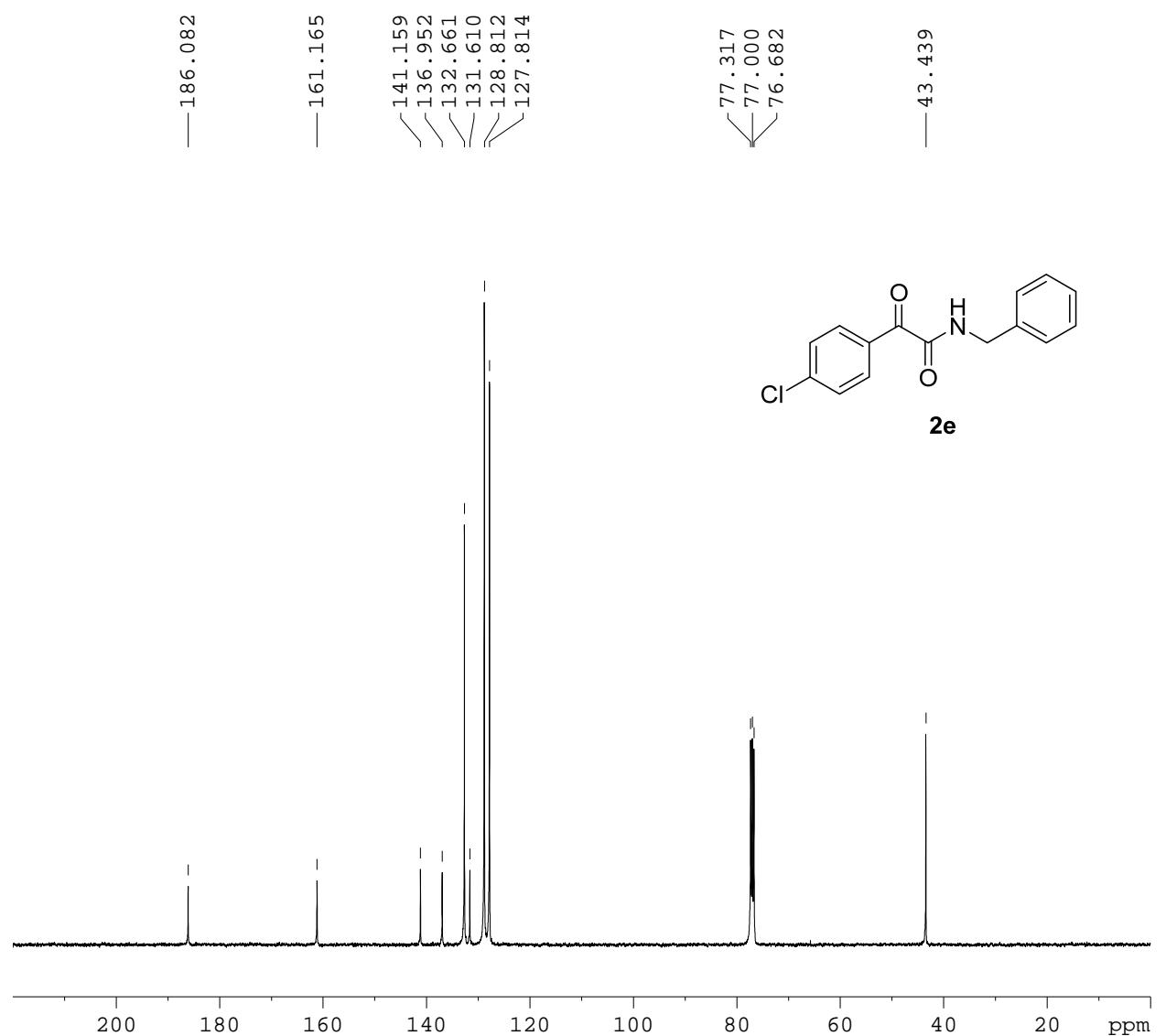


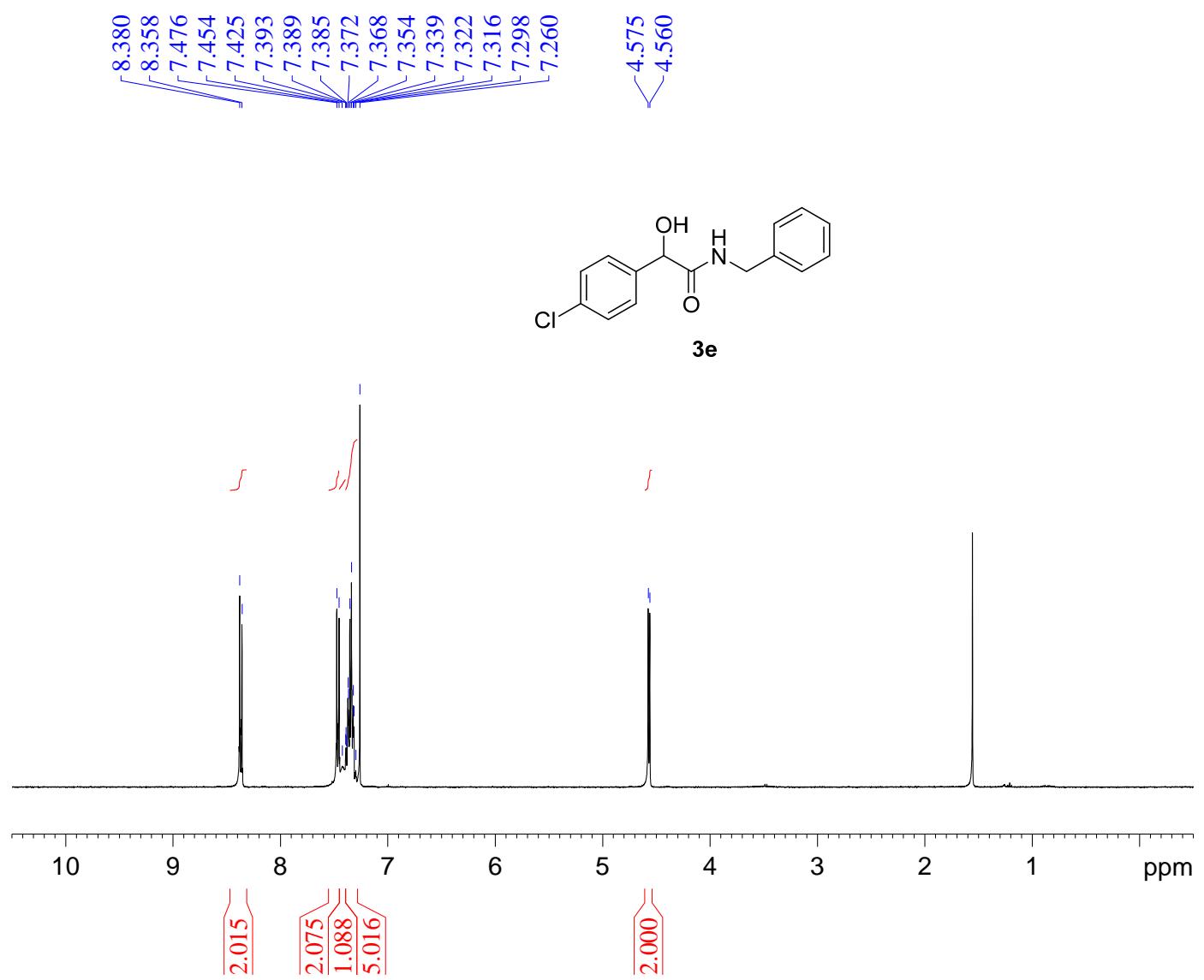
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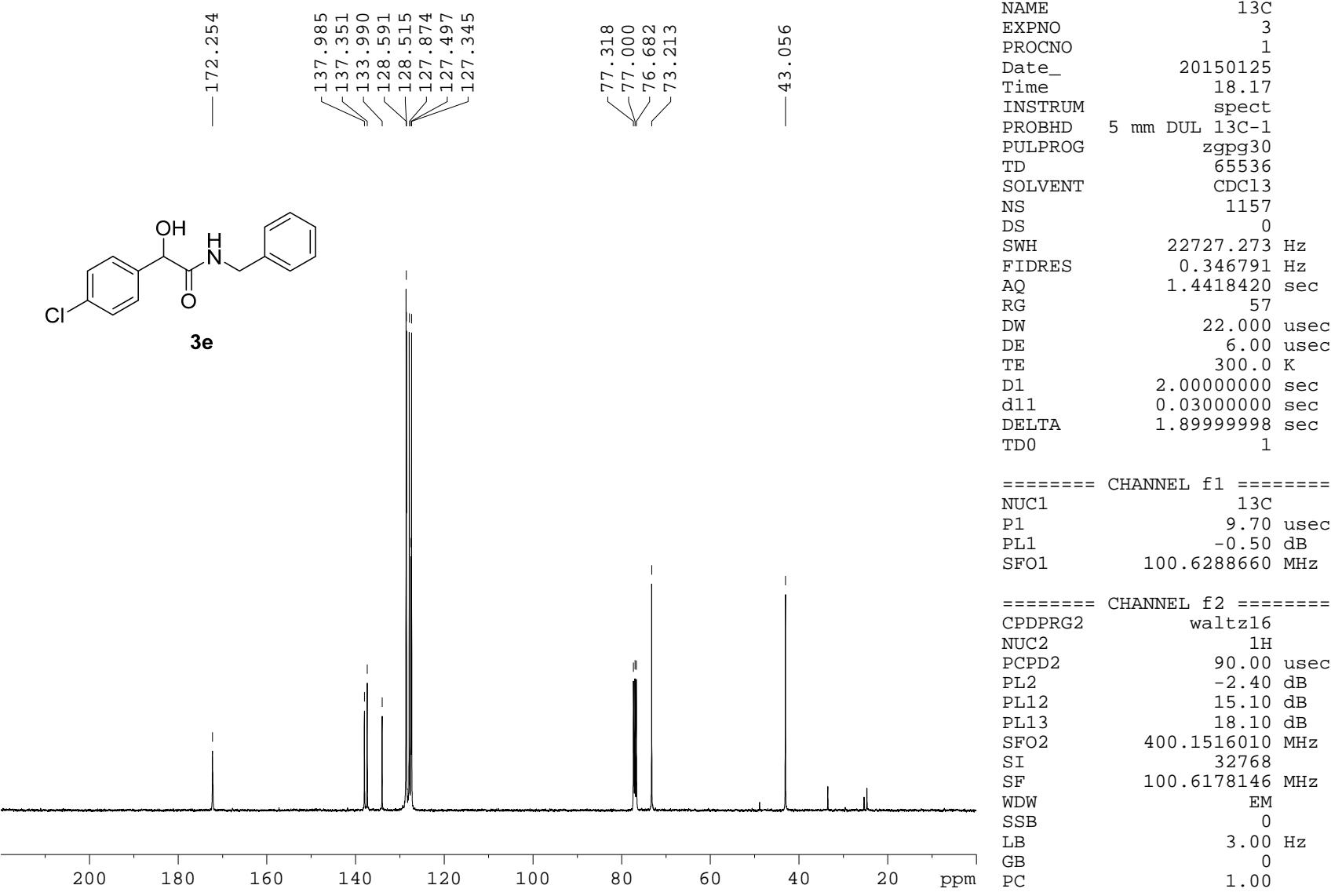
NAME      20151029
EXPNO     2
PROCNO    1
Date_   20151029
Time   13.54
INSTRUM spect
PROBHD  5 mm DUL 13C-1
PULPROG zg30
TD      32768
SOLVENT  CDCl3
NS       71
DS        0
SWH     6410.256 Hz
FIDRES  0.195625 Hz
AQ      2.5559540 sec
RG        4
DW      78.000 usec
DE       6.00 usec
TE      300.0 K
D1   2.0000000 sec
TD0        1

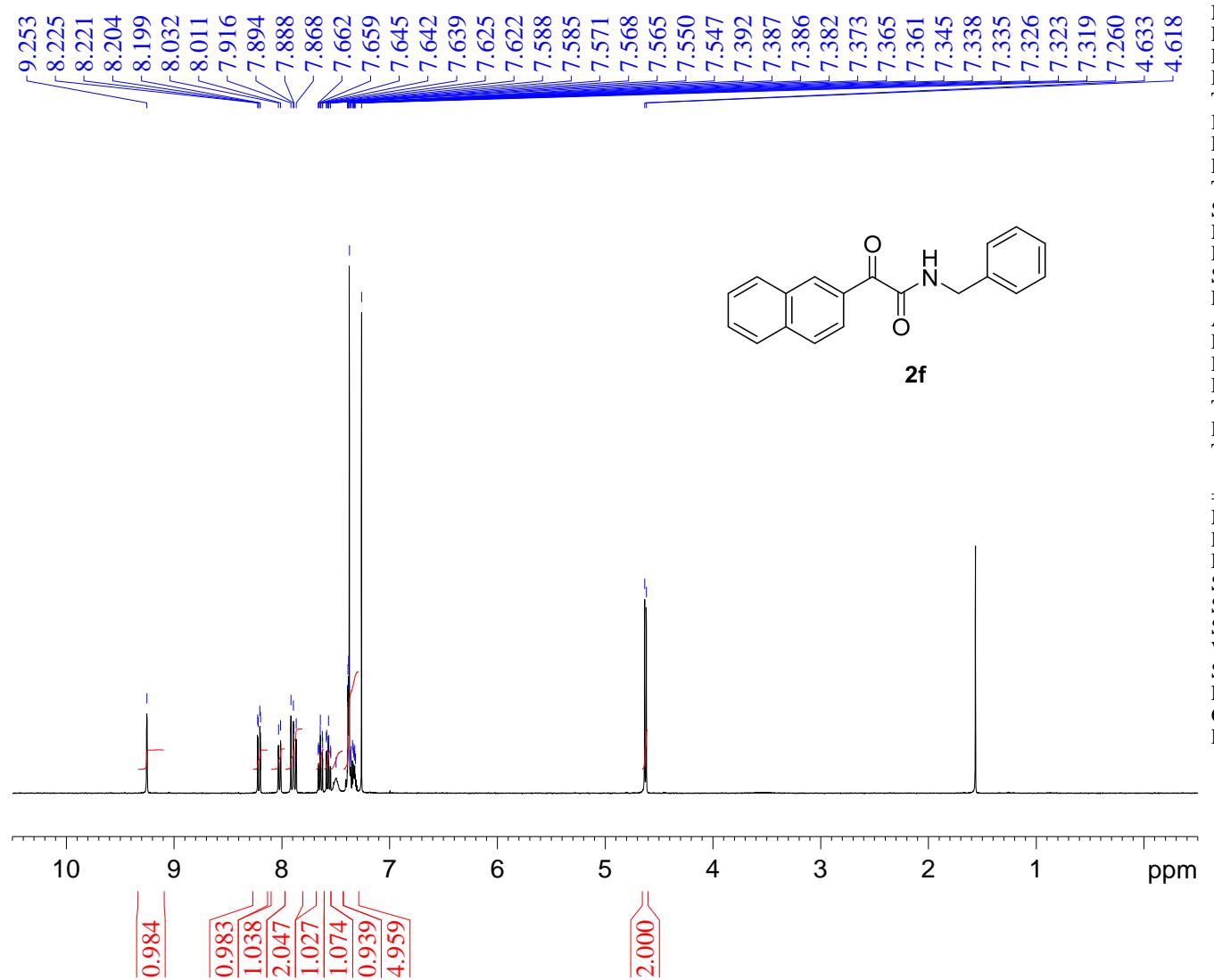
===== CHANNEL f1 =====
NUC1      1H
P1       10.00 usec
PL1      -2.40 dB
SFO1    400.1528010 MHz
SI       16384
SF      400.1500088 MHz
WDW      EM
SSB       0
LB      0.00 Hz
GB       0
PC       1.00

```





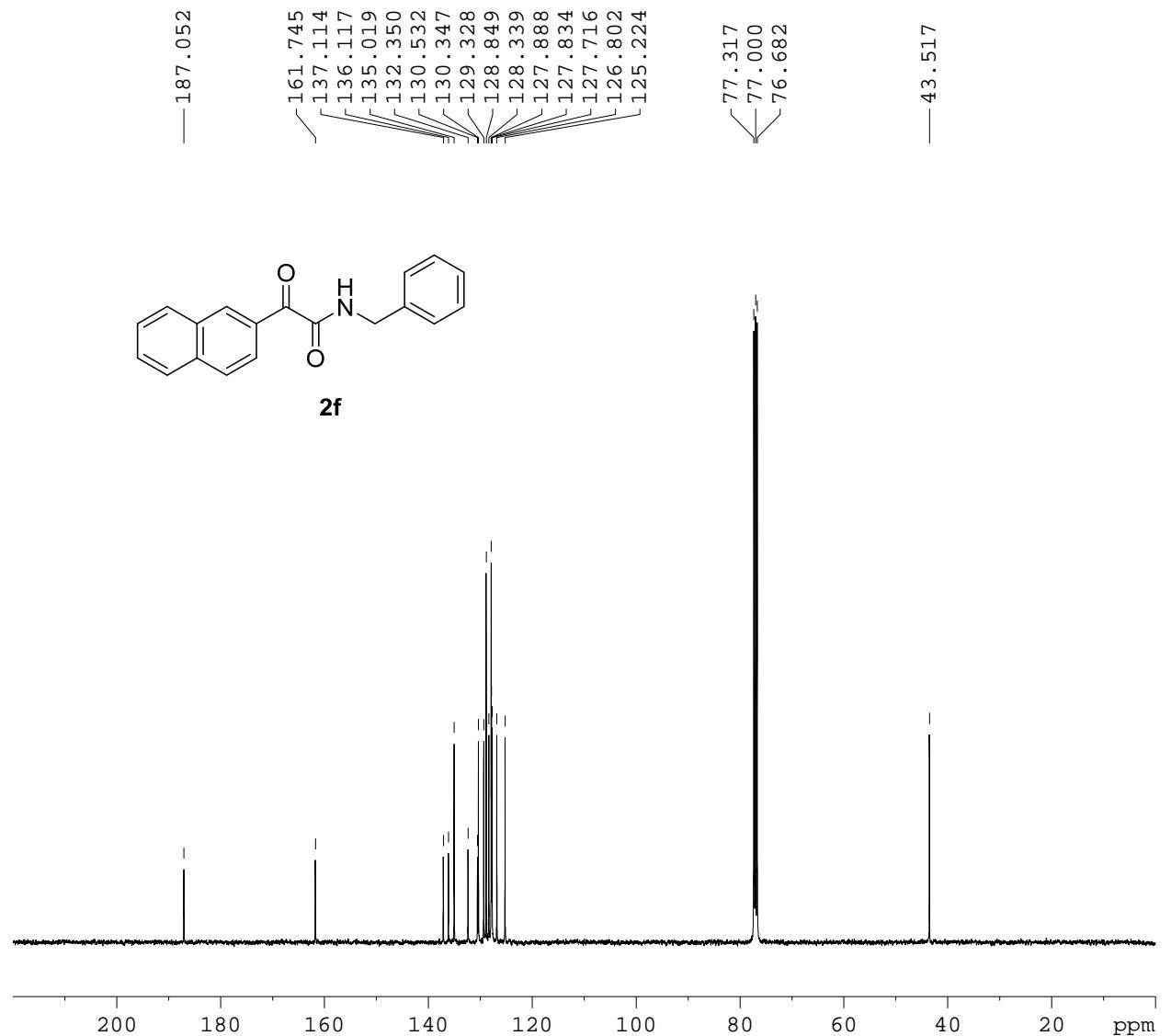


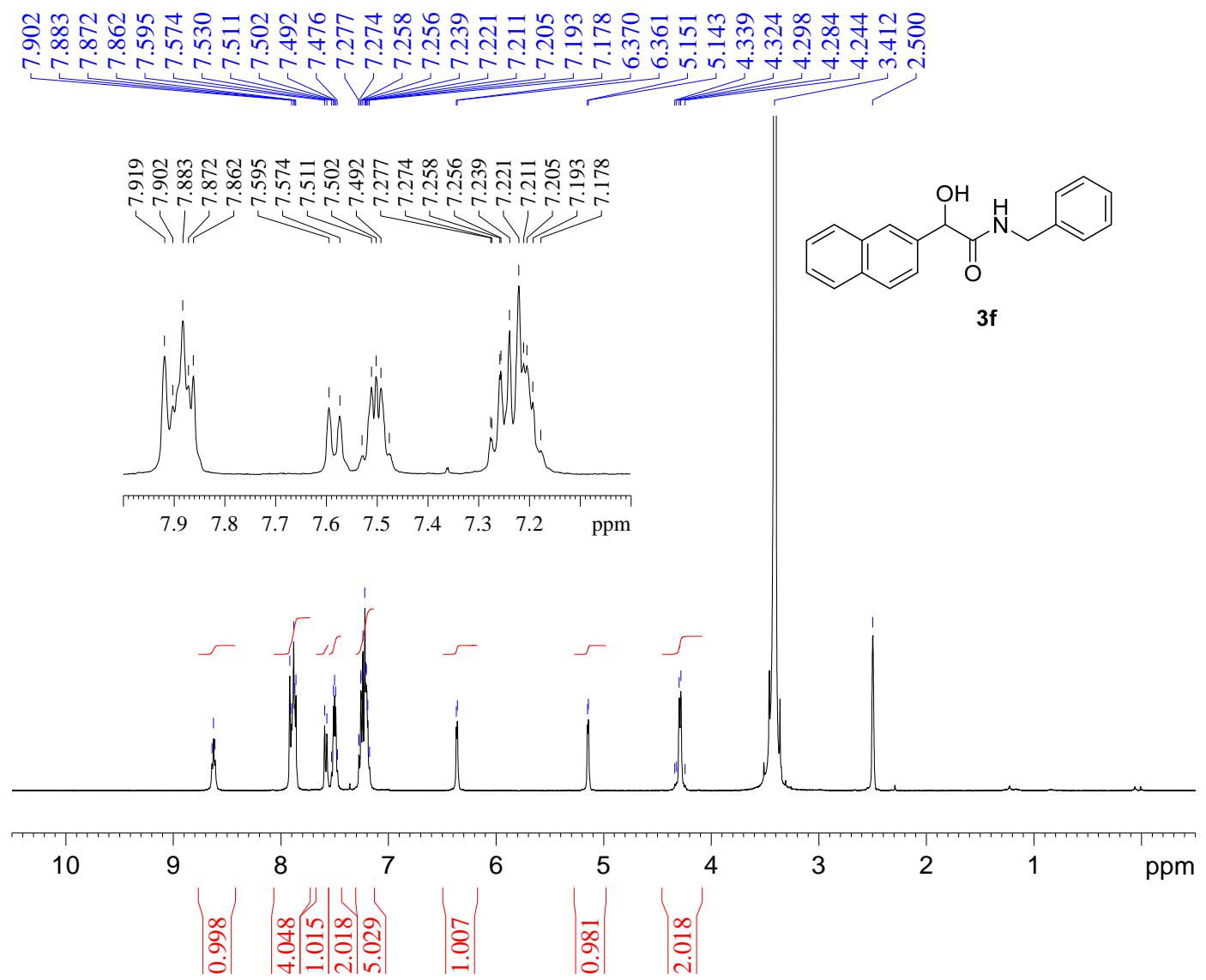


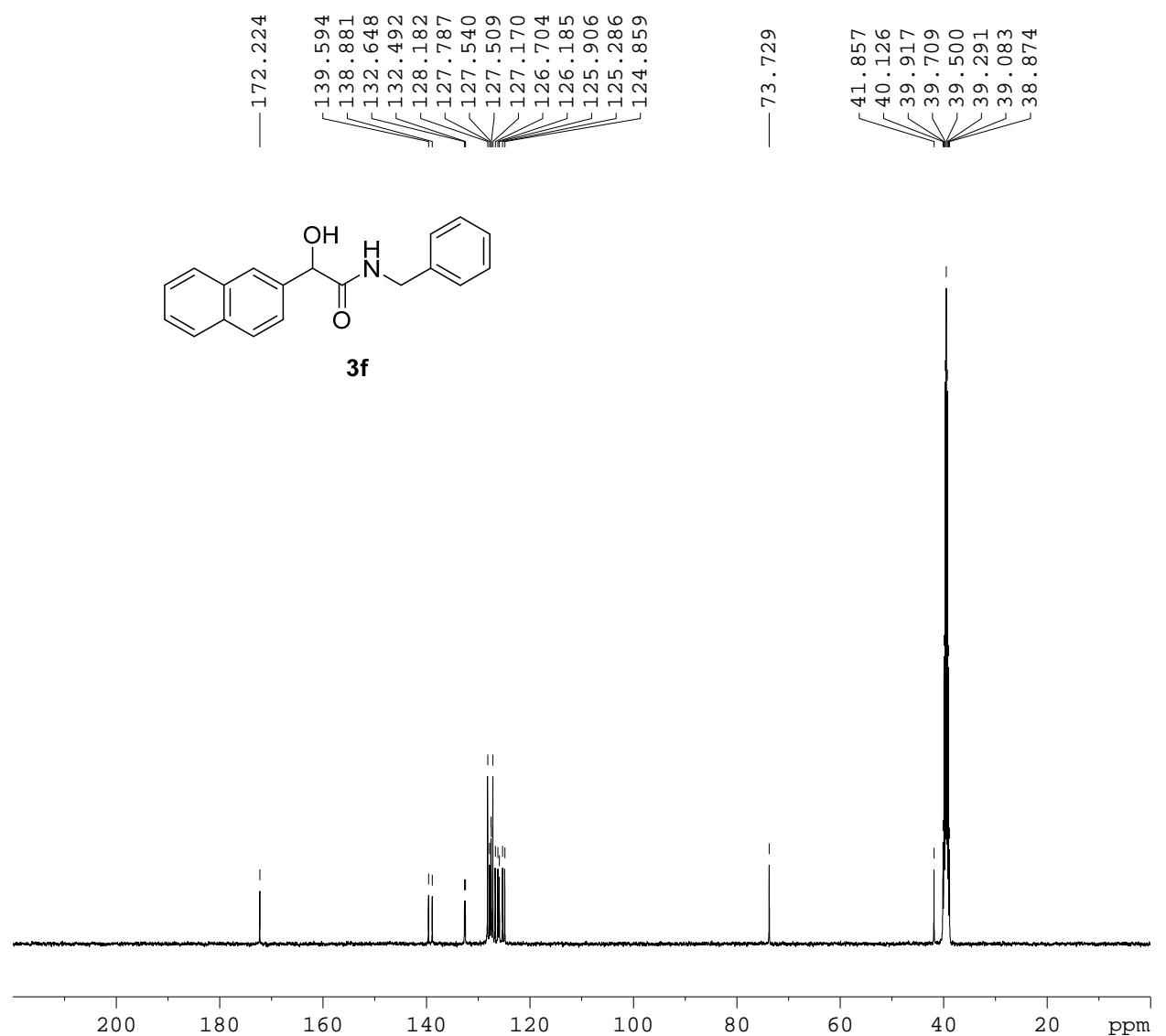
NAME Substrate 1H
 EXPNO 4
 PROCNO 1
 Date_ 20150613
 Time 16.55
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 32768
 SOLVENT CDCl₃
 NS 44
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.195625 Hz
 AQ 2.5559540 sec
 RG 4
 DW 78.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 TD0 1

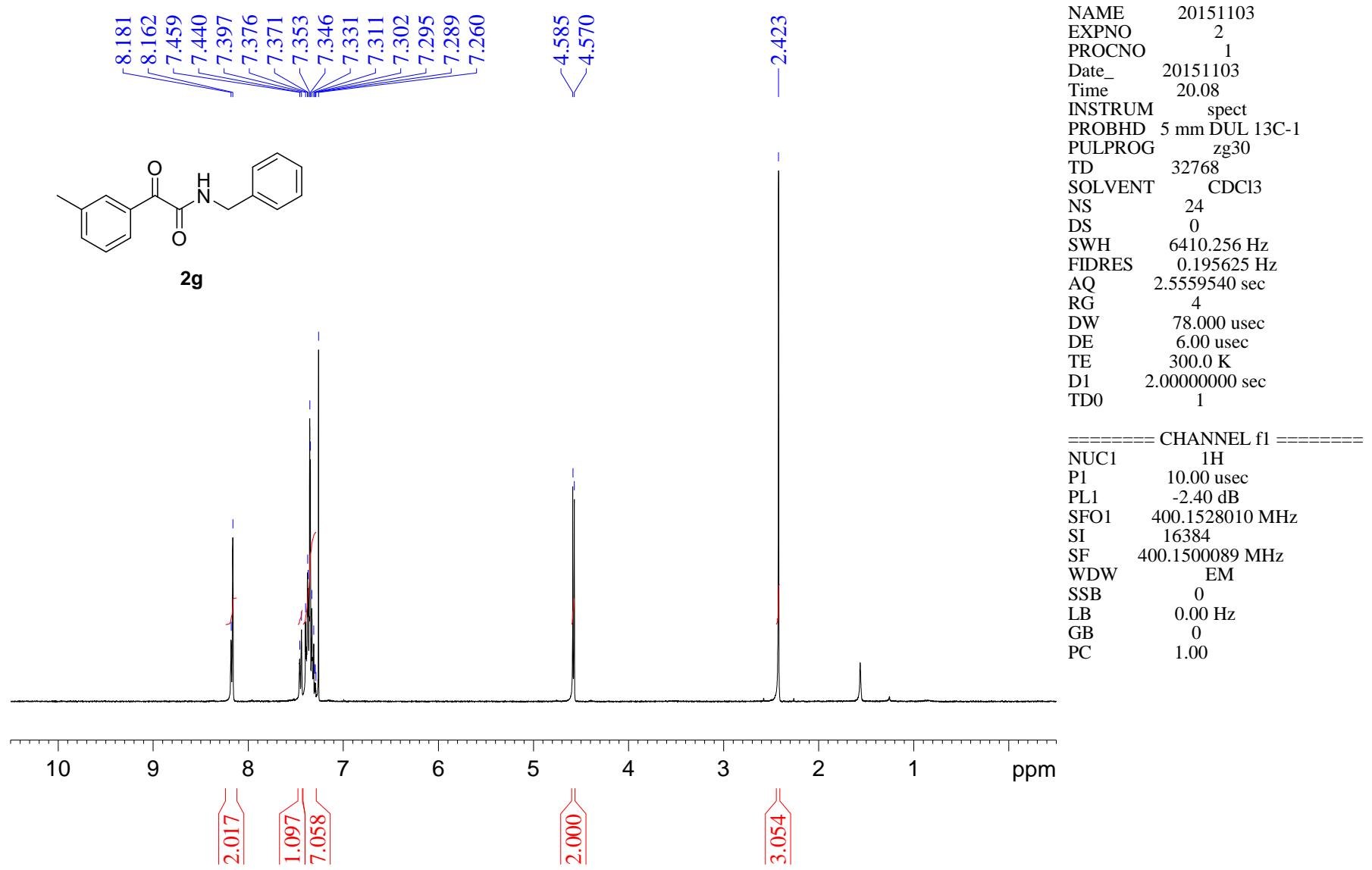
===== CHANNEL f1 =====

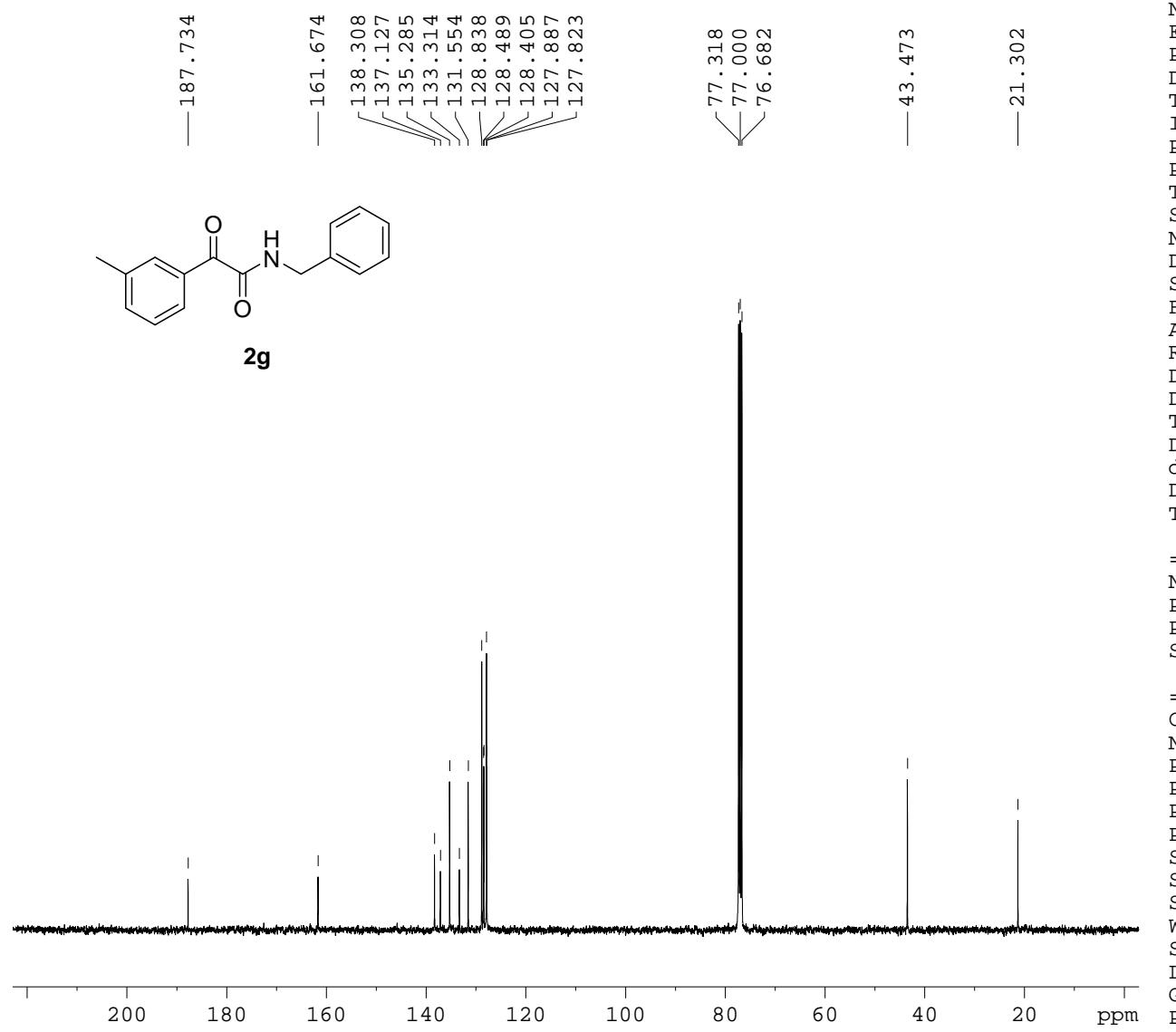
NUC1 1H
 P1 10.00 usec
 PL1 -2.40 dB
 SFO1 400.1528010 MHz
 SI 16384
 SF 400.1500088 MHz
 WDW EM
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00

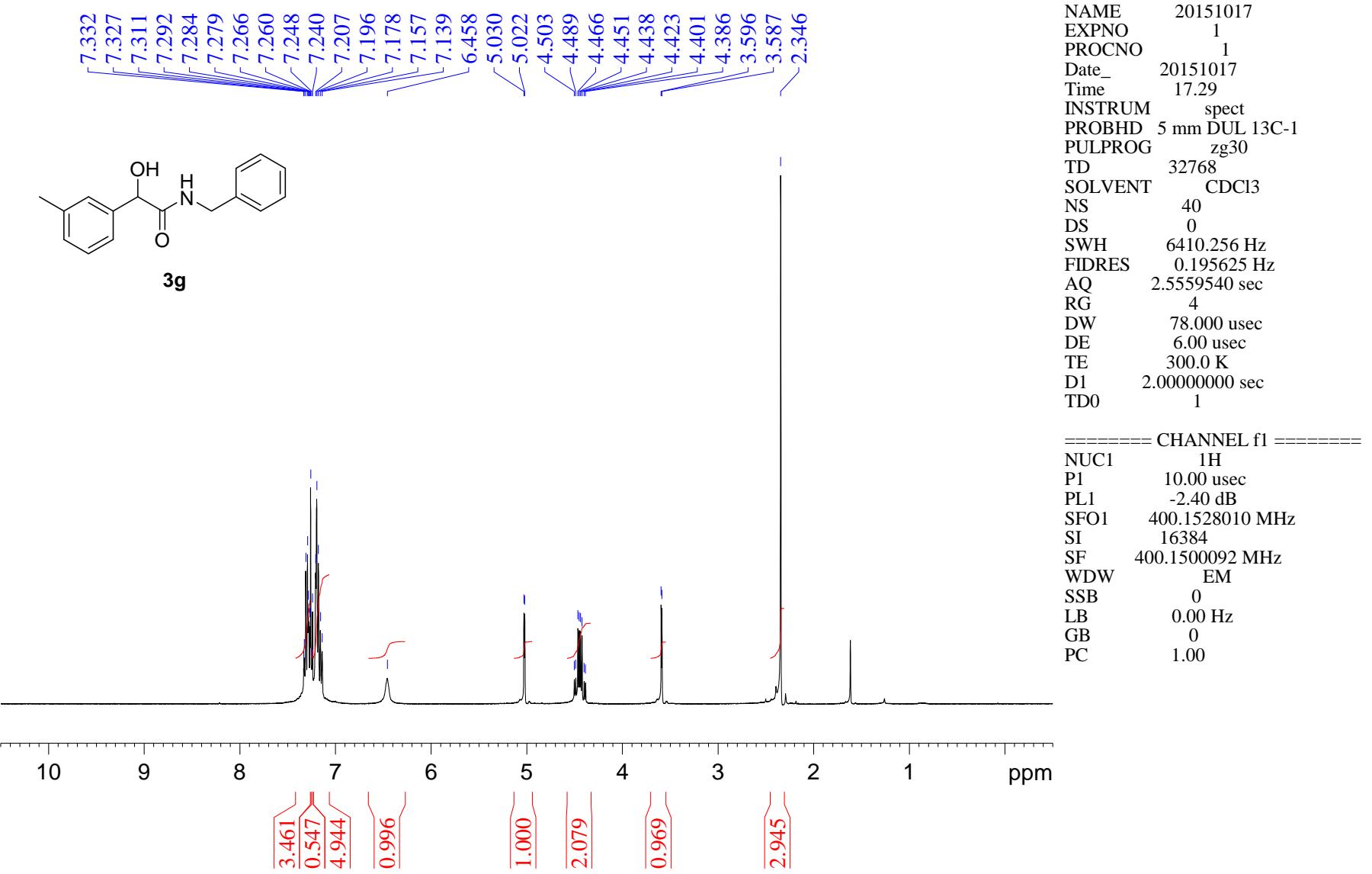




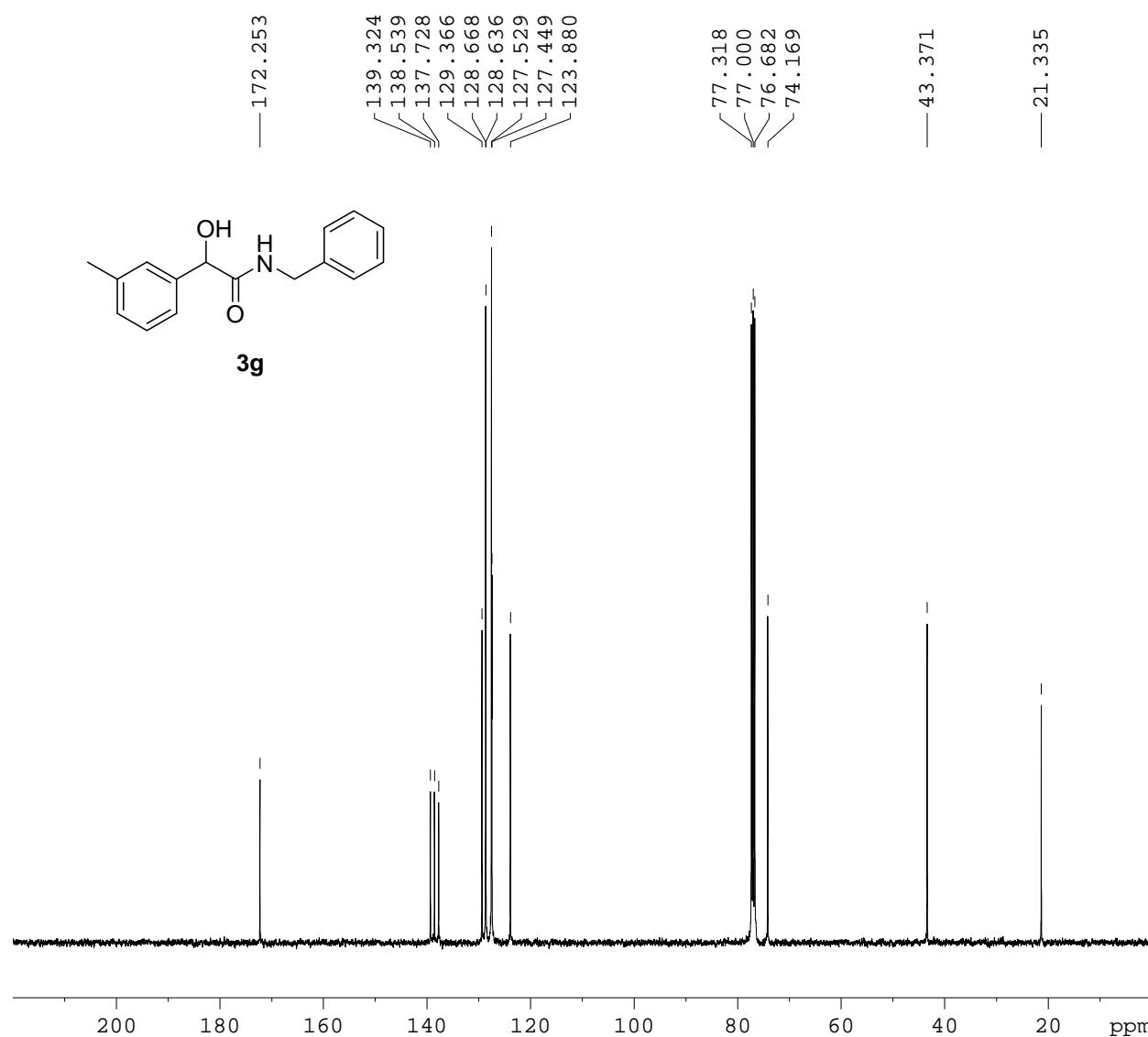








S100



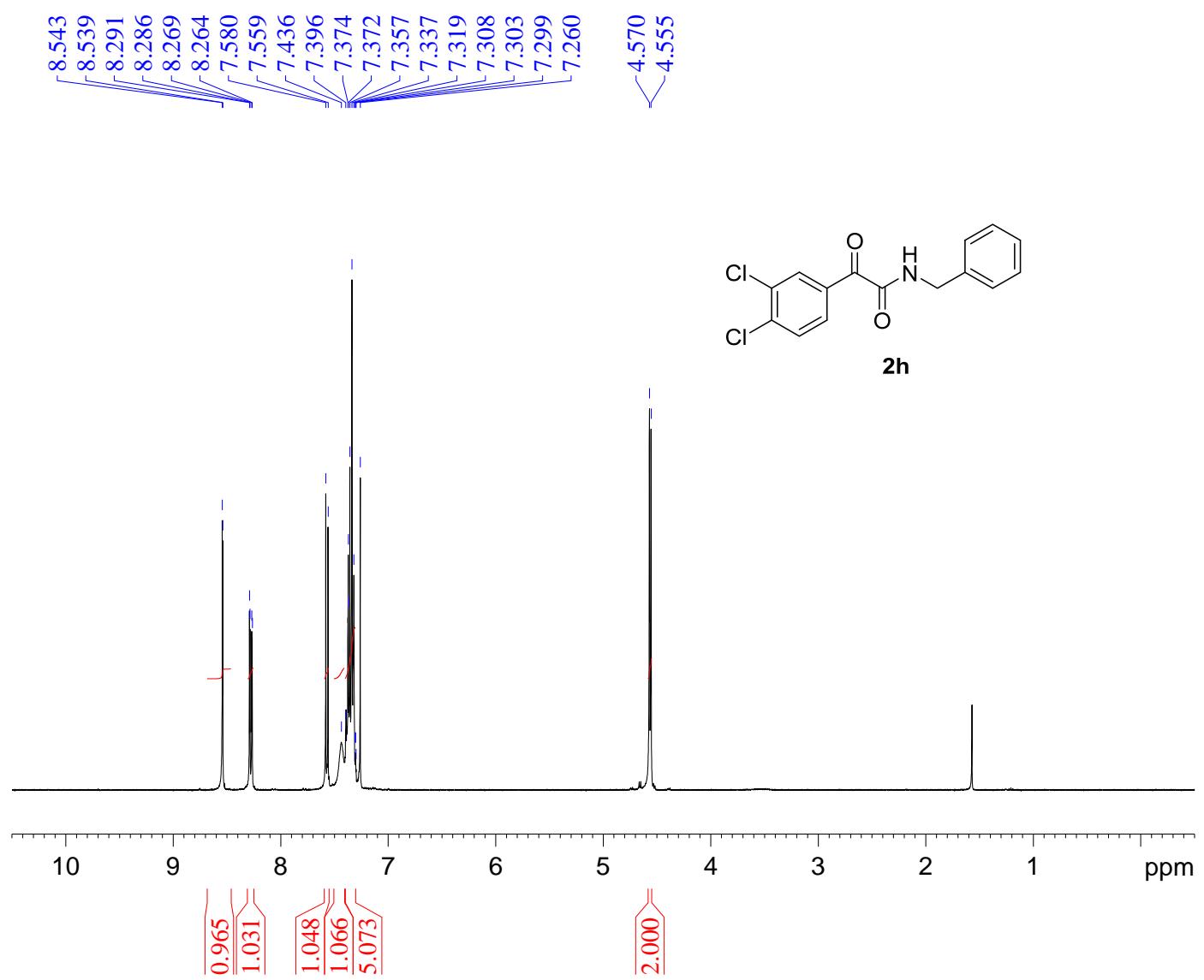
NAME Substrate 13C
 EXPNO 23
 PROCNO 1
 Date_ 20151018
 Time 16.11
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgppg30
 TD 65536
 SOLVENT CDCl3
 NS 1116
 DS 0
 SWH 22727.273 Hz
 FIDRES 0.346791 Hz
 AQ 1.4418420 sec
 RG 57
 DW 22.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 DELTA 1.8999998 sec
 TDO 1

===== CHANNEL f1 =====

NUC1 13C
 P1 9.70 usec
 PL1 -0.50 dB
 SFO1 100.6288660 MHz

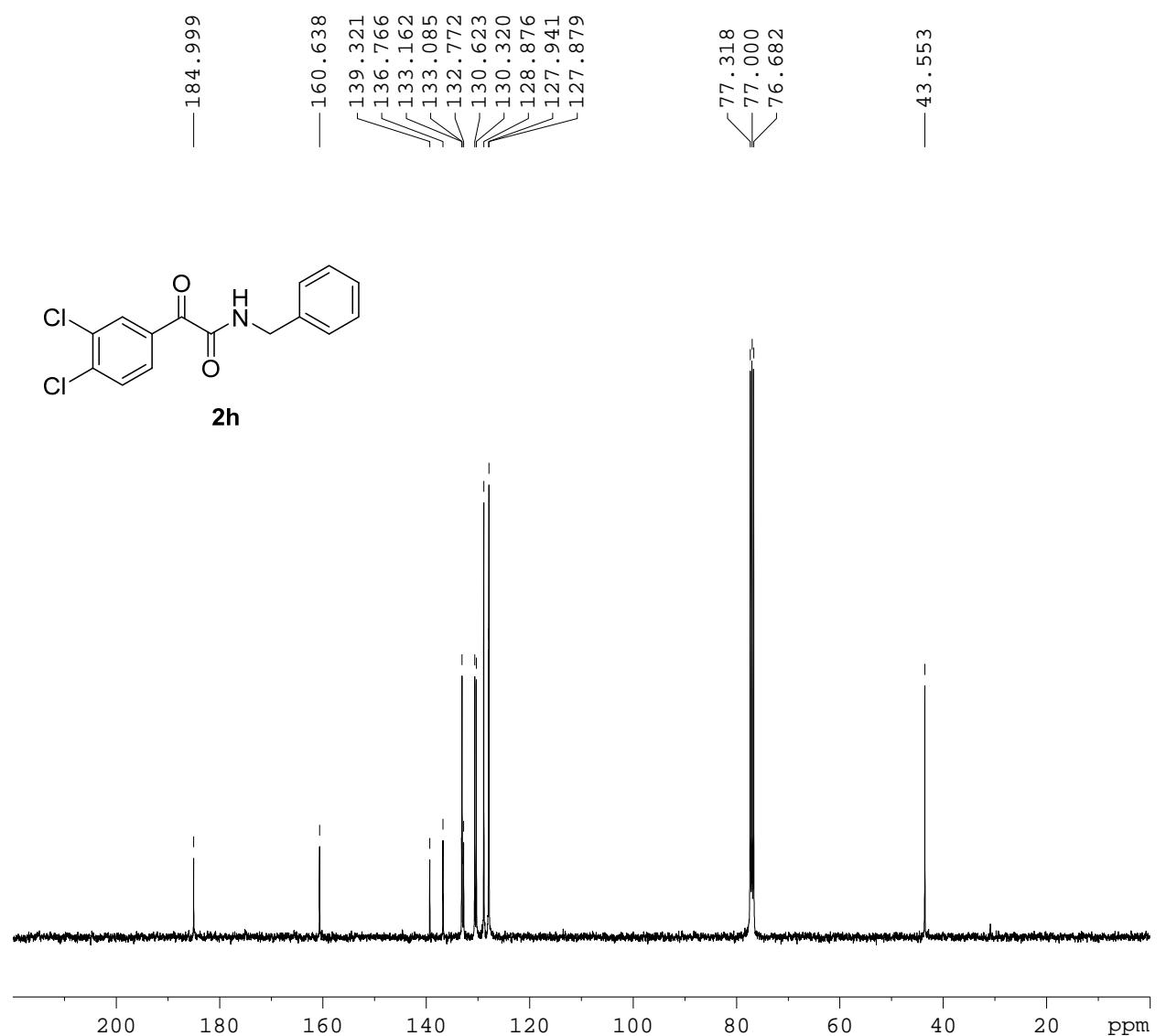
===== CHANNEL f2 =====

CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 -2.40 dB
 PL12 15.10 dB
 PL13 18.10 dB
 SFO2 400.1516010 MHz
 SI 32768
 SF 100.6178044 MHz
 WDW EM
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.00



NAME 20151207
 EXPNO 1
 PROCNO 1
 Date_ 20151207
 Time 9.50
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 32768
 SOLVENT CDCl₃
 NS 32
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.195625 Hz
 AQ 2.5559540 sec
 RG 4
 DW 78.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 TD0 1

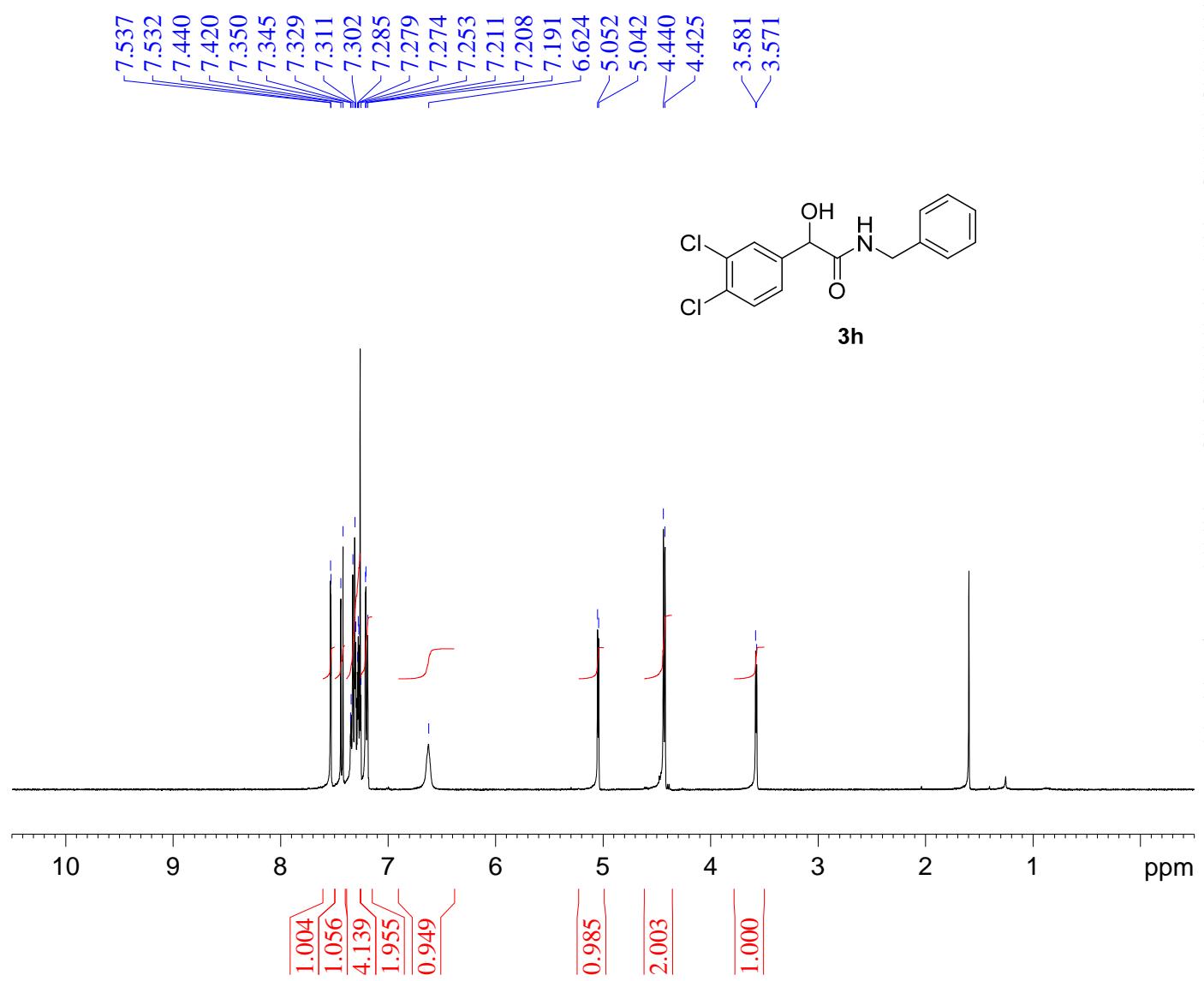
===== CHANNEL f1 =====
 NUC1 1H
 P1 10.00 usec
 PL1 -2.40 dB
 SFO1 400.1528010 MHz
 SI 16384
 SF 400.1500089 MHz
 WDW EM
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00



NAME 20151208
 EXPNO 2
 PROCNO 1
 Date_ 20151208
 Time 21.22
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 666
 DS 0
 SWH 22727.273 Hz
 FIDRES 0.346791 Hz
 AQ 1.4418420 sec
 RG 57
 DW 22.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 TDO 1

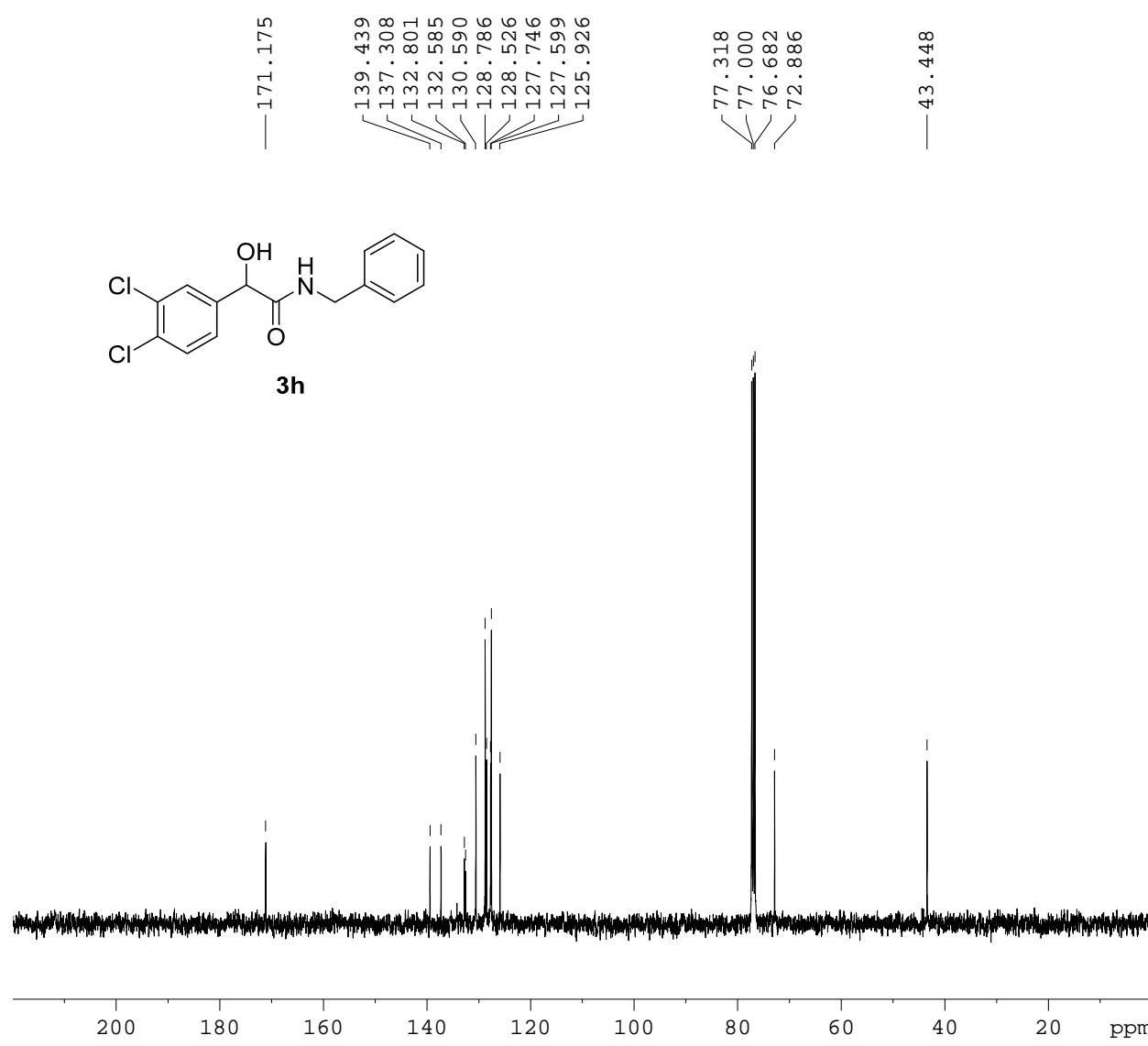
===== CHANNEL f1 =====
 NUC1 13C
 P1 9.70 usec
 PL1 -0.50 dB
 SFO1 100.6288660 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 -2.40 dB
 PL12 15.10 dB
 PL13 18.10 dB
 SFO2 400.1516010 MHz
 SI 32768
 SF 100.6178028 MHz
 WDW EM
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.00



NAME 20151209
EXPNO 1
PROCNO 1
Date_ 20151209
Time 20.04
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 64
DS 0
SWH 6410.256 Hz
FIDRES 0.195625 Hz
AQ 2.5559540 sec
RG 4
DW 78.000 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
TD0 1

===== CHANNEL f1 ======
NUC1 1H
P1 10.00 usec
PL1 -2.40 dB
SFO1 400.1528010 MHz
SI 16384
SF 400.1500088 MHz
WDW EM
SSB 0
LB 0.00 Hz
GB 0
PC 1.00



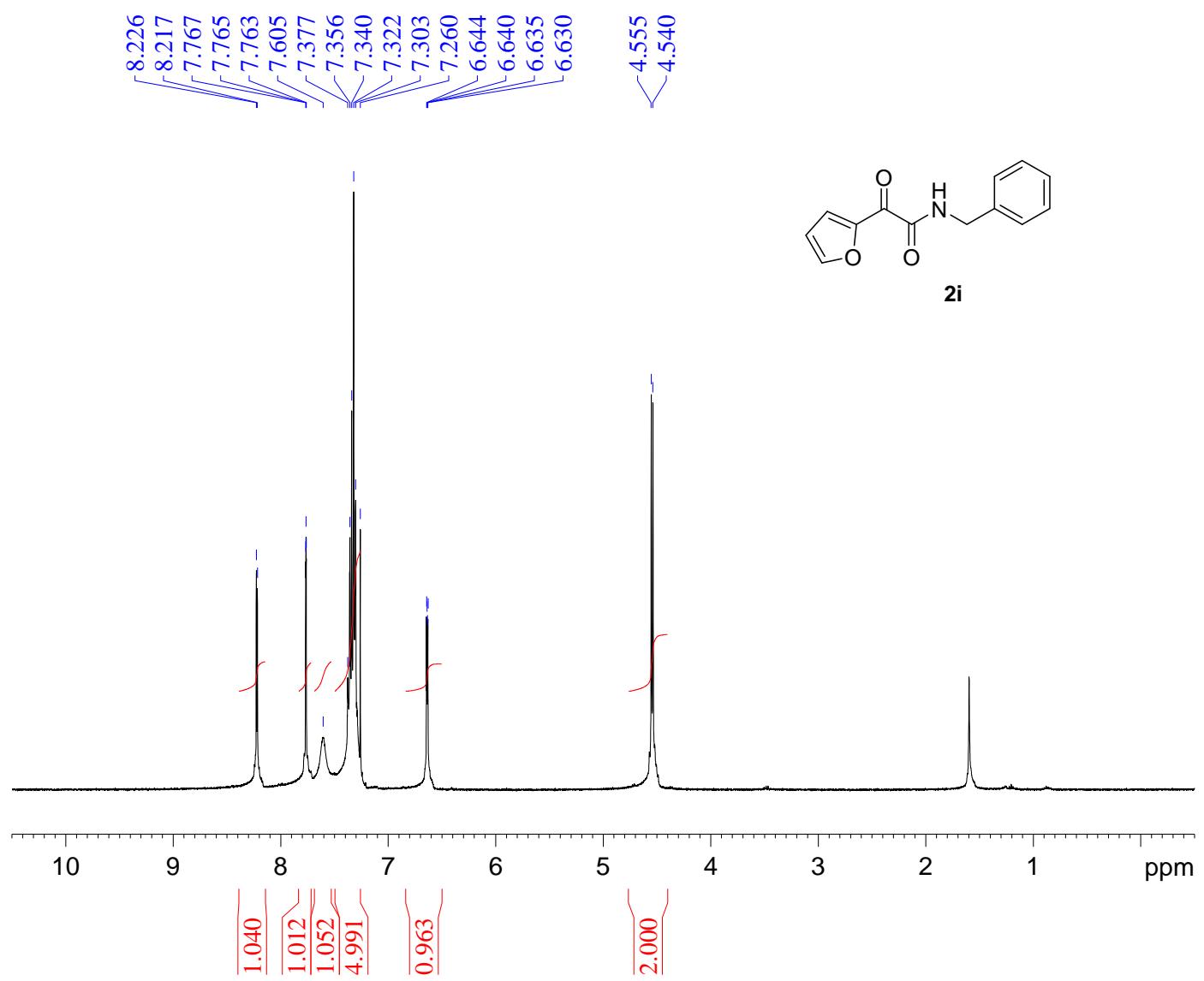
NAME 20151209
 EXPNO 2
 PROCNO 1
 Date_ 20151209
 Time 20.13
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgppg30
 TD 65536
 SOLVENT CDCl3
 NS 100
 DS 0
 SWH 22727.273 Hz
 FIDRES 0.346791 Hz
 AQ 1.4418420 sec
 RG 57
 DW 22.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 TDO 1

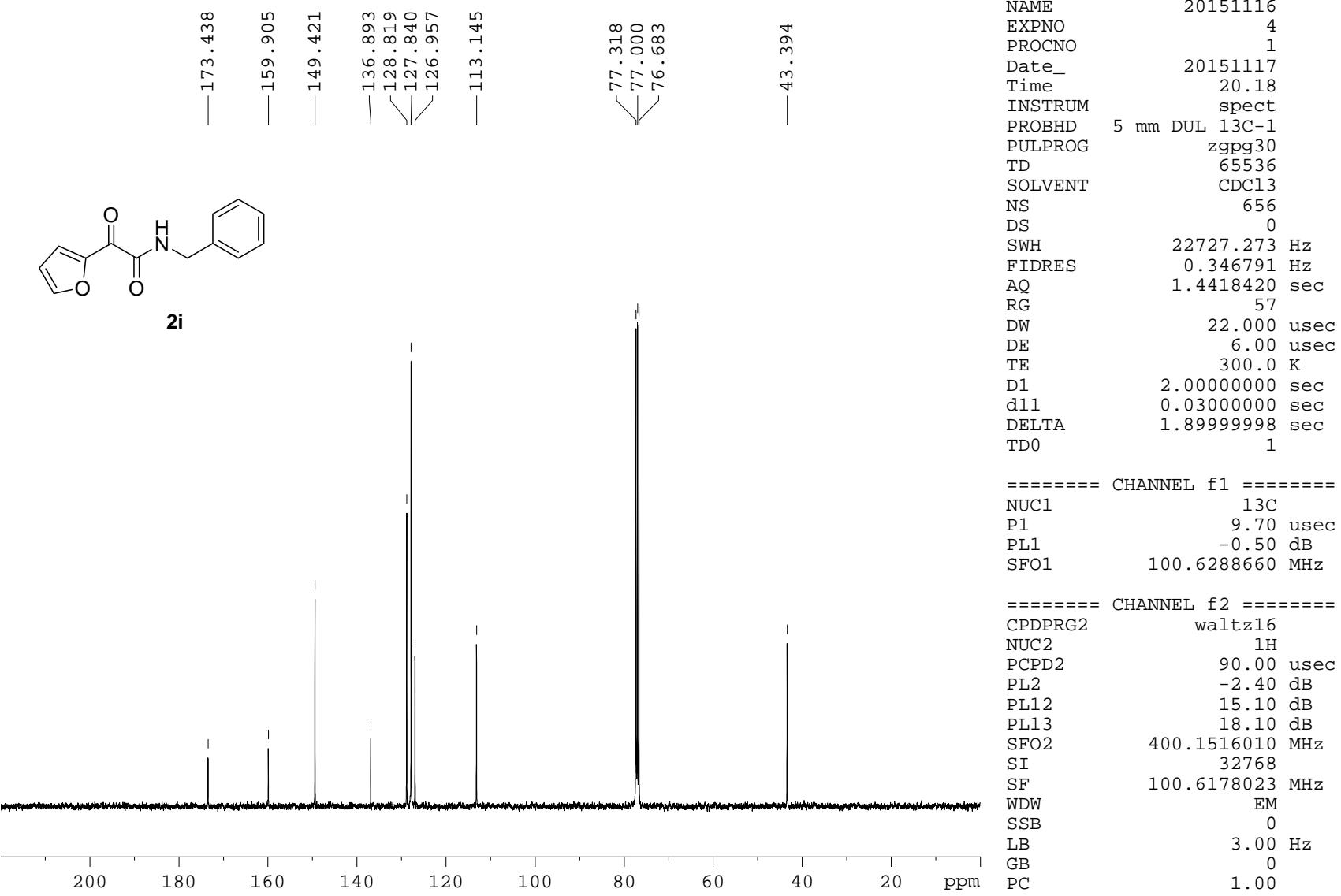
===== CHANNEL f1 =====

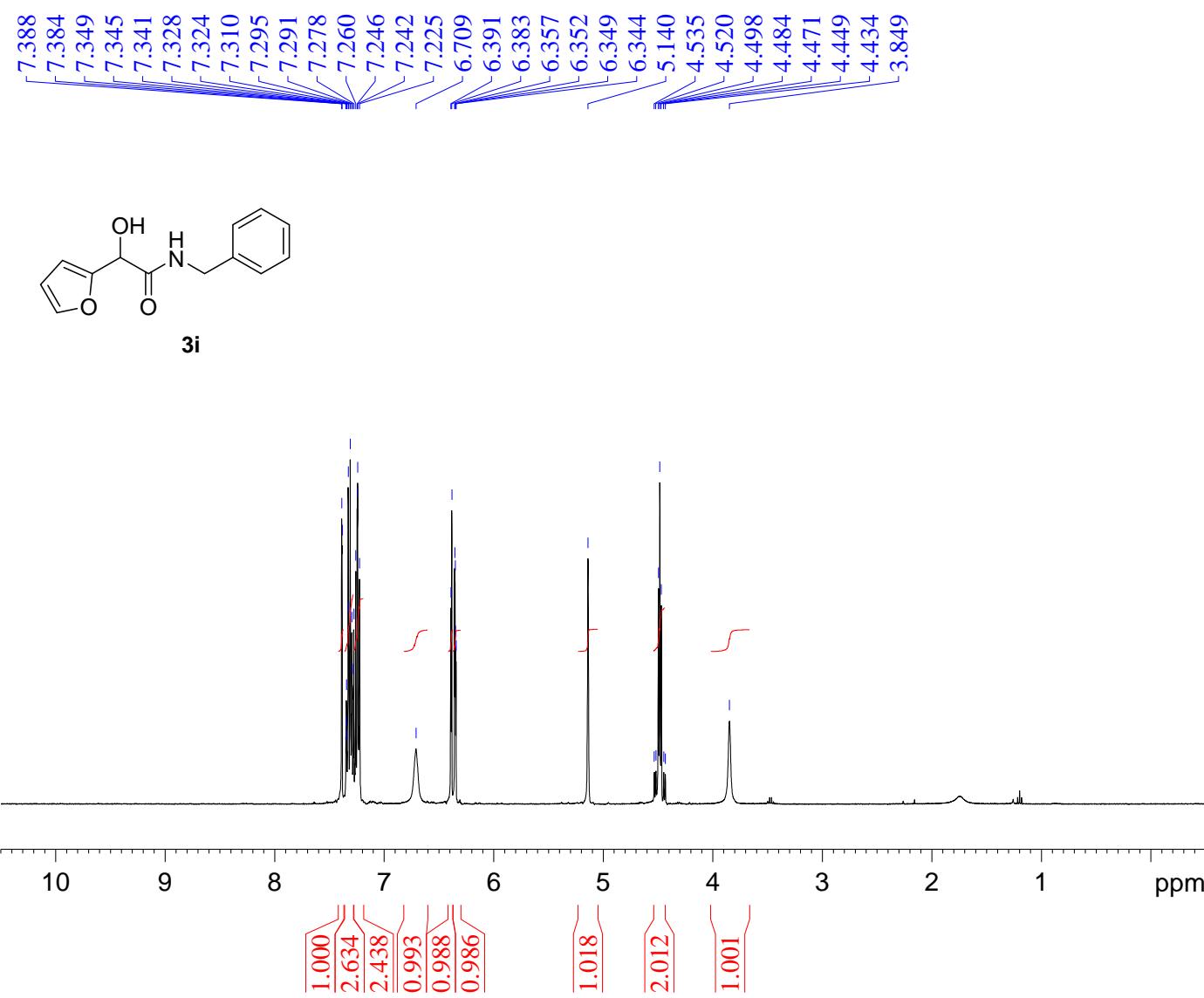
NUC1 13C
 P1 9.70 usec
 PL1 -0.50 dB
 SFO1 100.6288660 MHz

===== CHANNEL f2 =====

CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 -2.40 dB
 PL12 15.10 dB
 PL13 18.10 dB
 SFO2 400.1516010 MHz
 SI 32768
 SF 100.6178021 MHz
 WDW EM
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.00



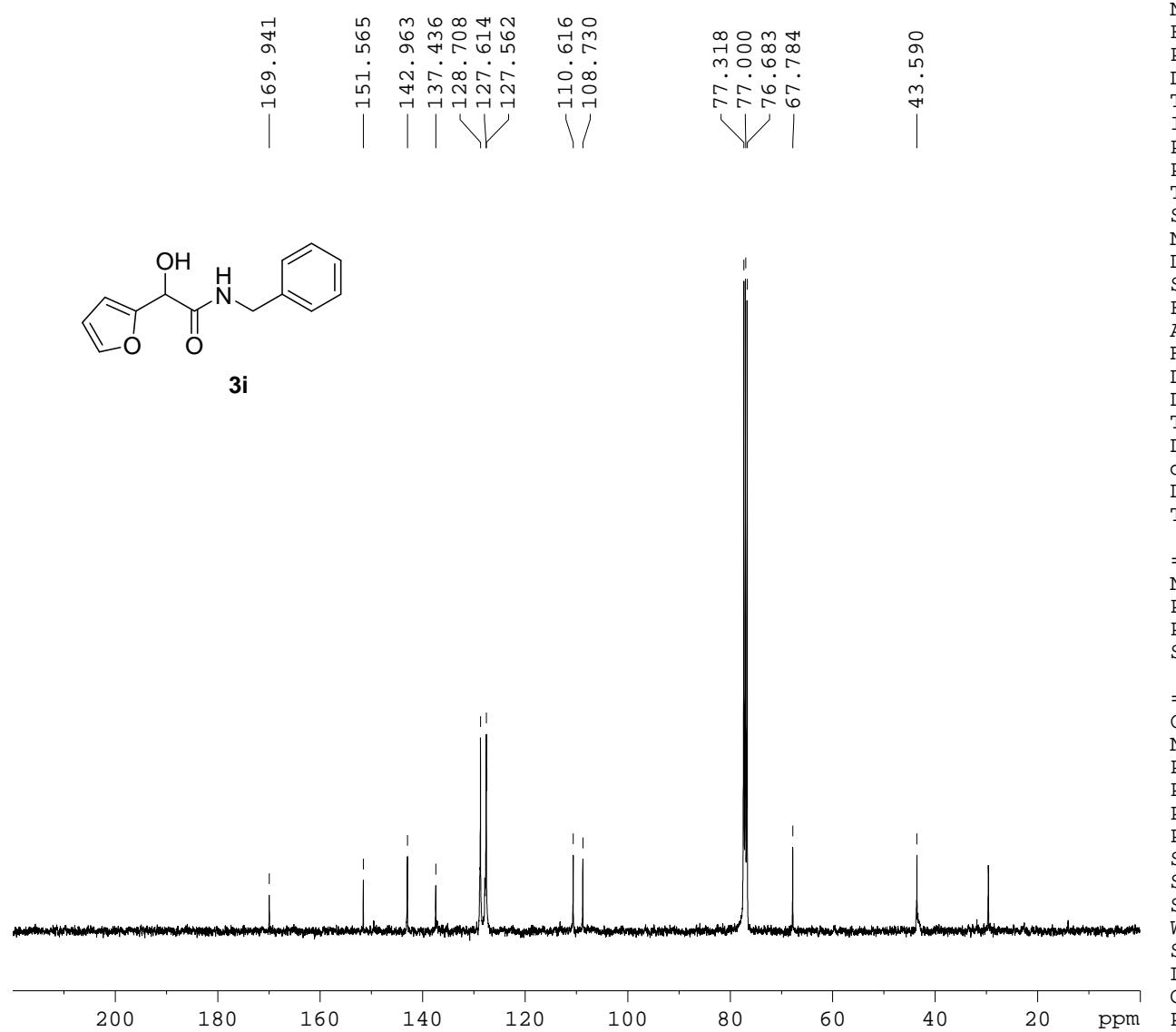




NAME	20160216
EXPNO	3
PROCNO	1
Date_	20150608
Time	18.28
INSTRUM	spect
PROBHD	5 mm DUL 13C-1
PULPROG	zg30
TD	32768
SOLVENT	CDCl ₃
NS	31
DS	0
SWH	6410.256 Hz
FIDRES	0.195625 Hz
AQ	2.5559540 sec
RG	4
DW	78.000 usec
DE	6.00 usec
TE	300.0 K
D1	2.0000000 sec
TD0	1

===== CHANNEL f1 =====

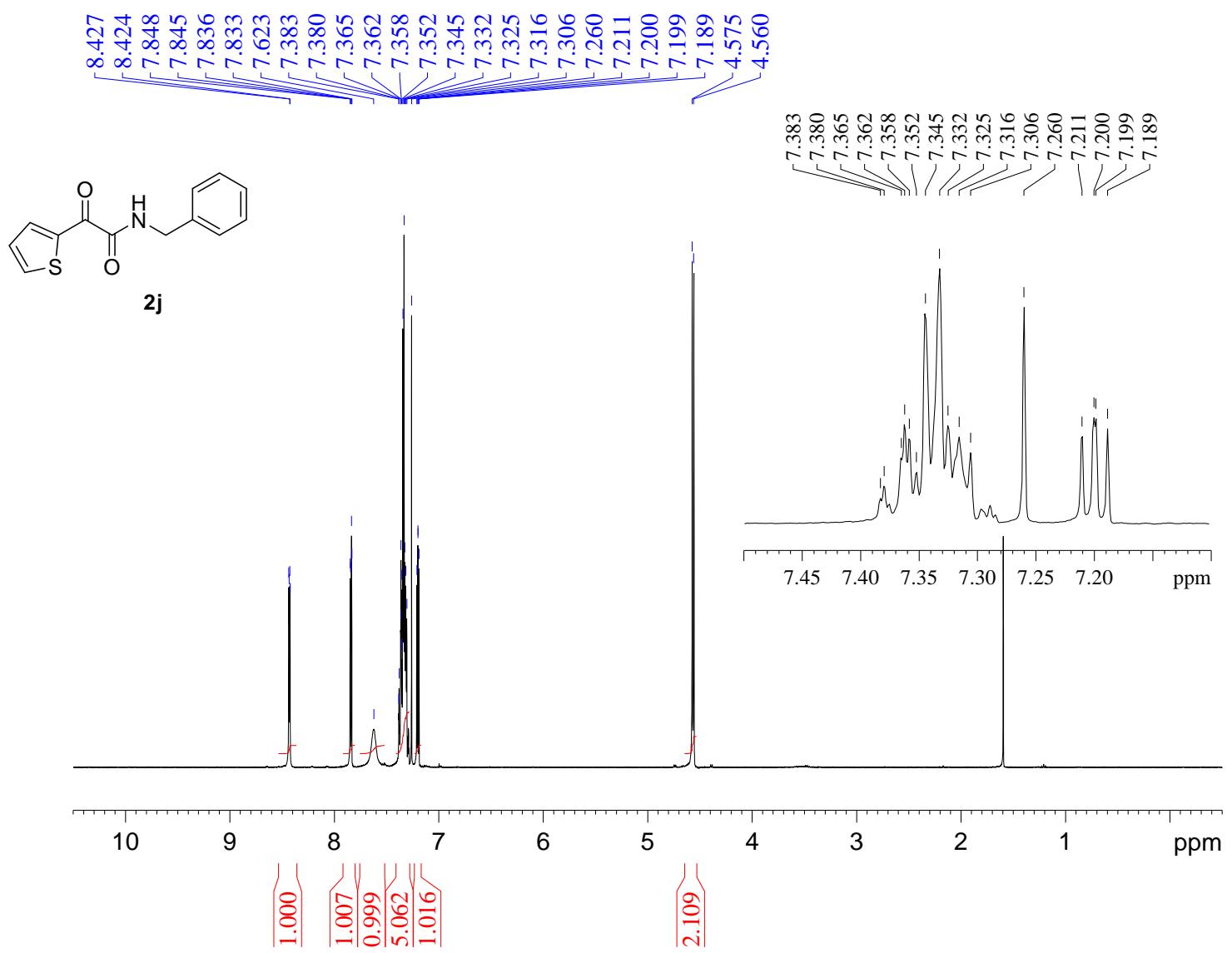
NUC1	1H
P1	10.00 usec
PL1	-2.40 dB
SFO1	400.1528010 MHz
SI	16384
SF	400.1500088 MHz
WDW	EM
SSB	0
LB	0.00 Hz
GB	0
PC	1.00



NAME 20160216
 EXPNO 1
 PROCNO 1
 Date_ 20160216
 Time 18.59
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgppg30
 TD 65536
 SOLVENT CDCl3
 NS 960
 DS 0
 SWH 22727.273 Hz
 FIDRES 0.346791 Hz
 AQ 1.4418420 sec
 RG 57
 DW 22.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 DELTA 1.8999999 sec
 TDO 1

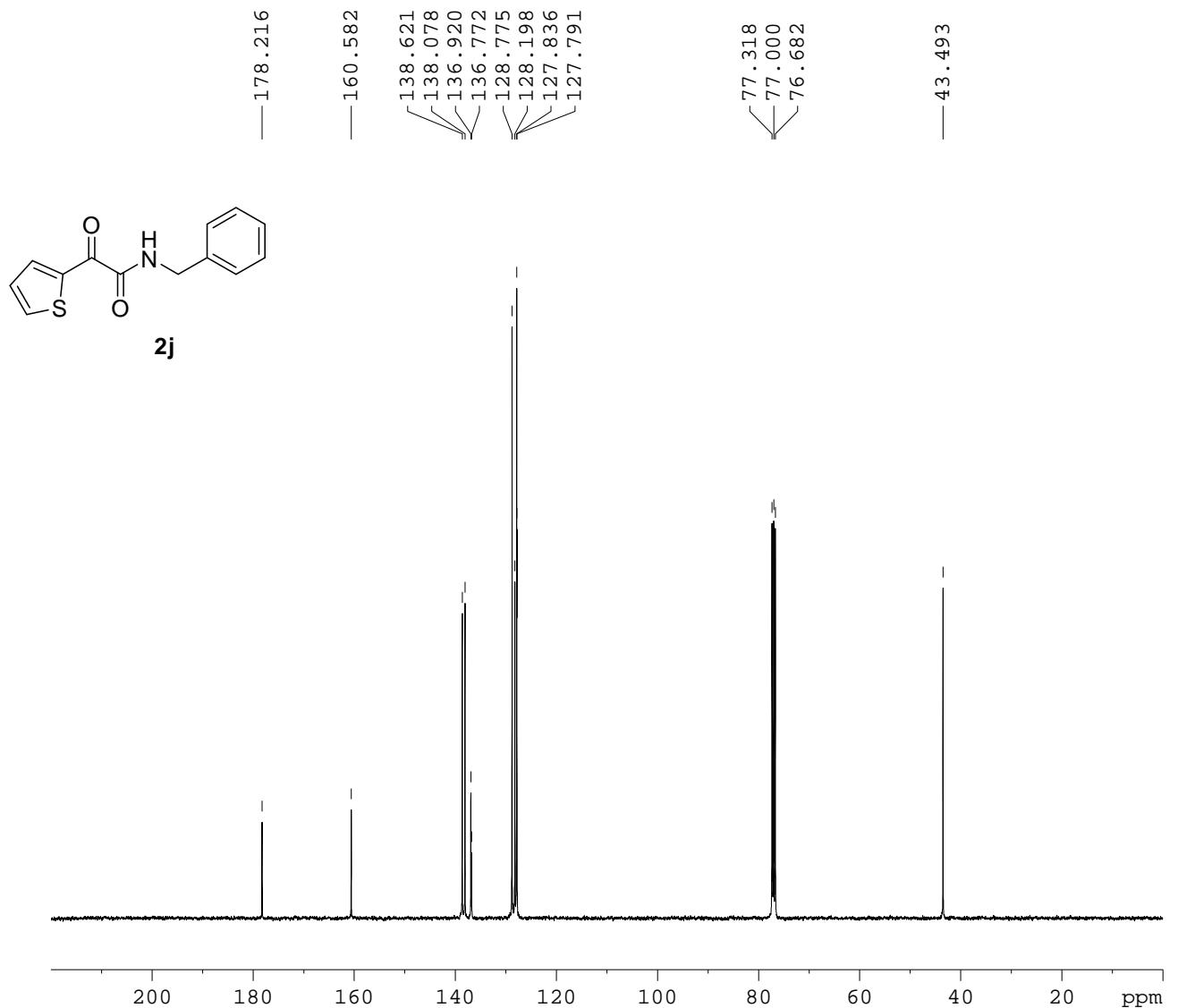
===== CHANNEL f1 =====
 NUC1 13C
 P1 9.70 usec
 PL1 -0.50 dB
 SFO1 100.6288660 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 -2.40 dB
 PL12 15.10 dB
 PL13 18.10 dB
 SFO2 400.1516010 MHz
 SI 32768
 SF 100.6178033 MHz
 WDW EM
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.00



NAME Substrate 1H
 EXPNO 2
 PROCNO 1
 Date_ 20150530
 Time 18.10
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 32768
 SOLVENT CDCl₃
 NS 32
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.195625 Hz
 AQ 2.5559540 sec
 RG 4
 DW 78.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 TD0 1

===== CHANNEL f1 ======
 NUC1 1H
 P1 10.00 usec
 PL1 -2.40 dB
 SFO1 400.1528010 MHz
 SI 16384
 SF 400.1500088 MHz
 WDW EM
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00

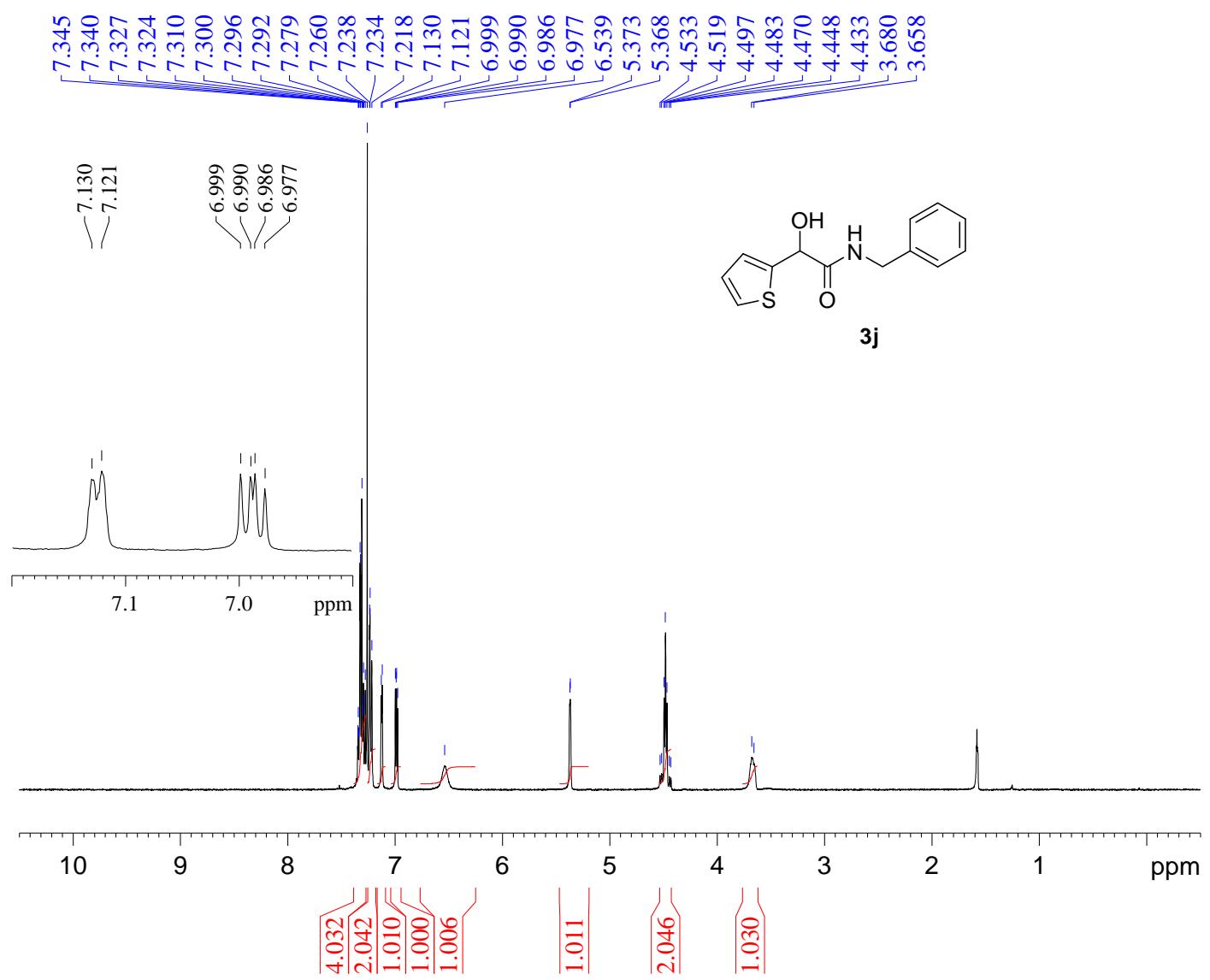


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NAME          Substrate 13C
EXPNO         2
PROCNO        1
Date_        20150530
Time         18.16
INSTRUM      spect
PROBHD      5 mm DUL 13C-1
PULPROG     zgppg30
TD           65536
SOLVENT      CDCl3
NS            1467
DS             0
SWH          22727.273 Hz
FIDRES       0.346791 Hz
AQ           1.4418420 sec
RG            57
DW           22.000 usec
DE            6.00 usec
TE           300.0 K
D1           2.0000000 sec
d11          0.03000000 sec
DELTA        1.89999998 sec
TDO           1

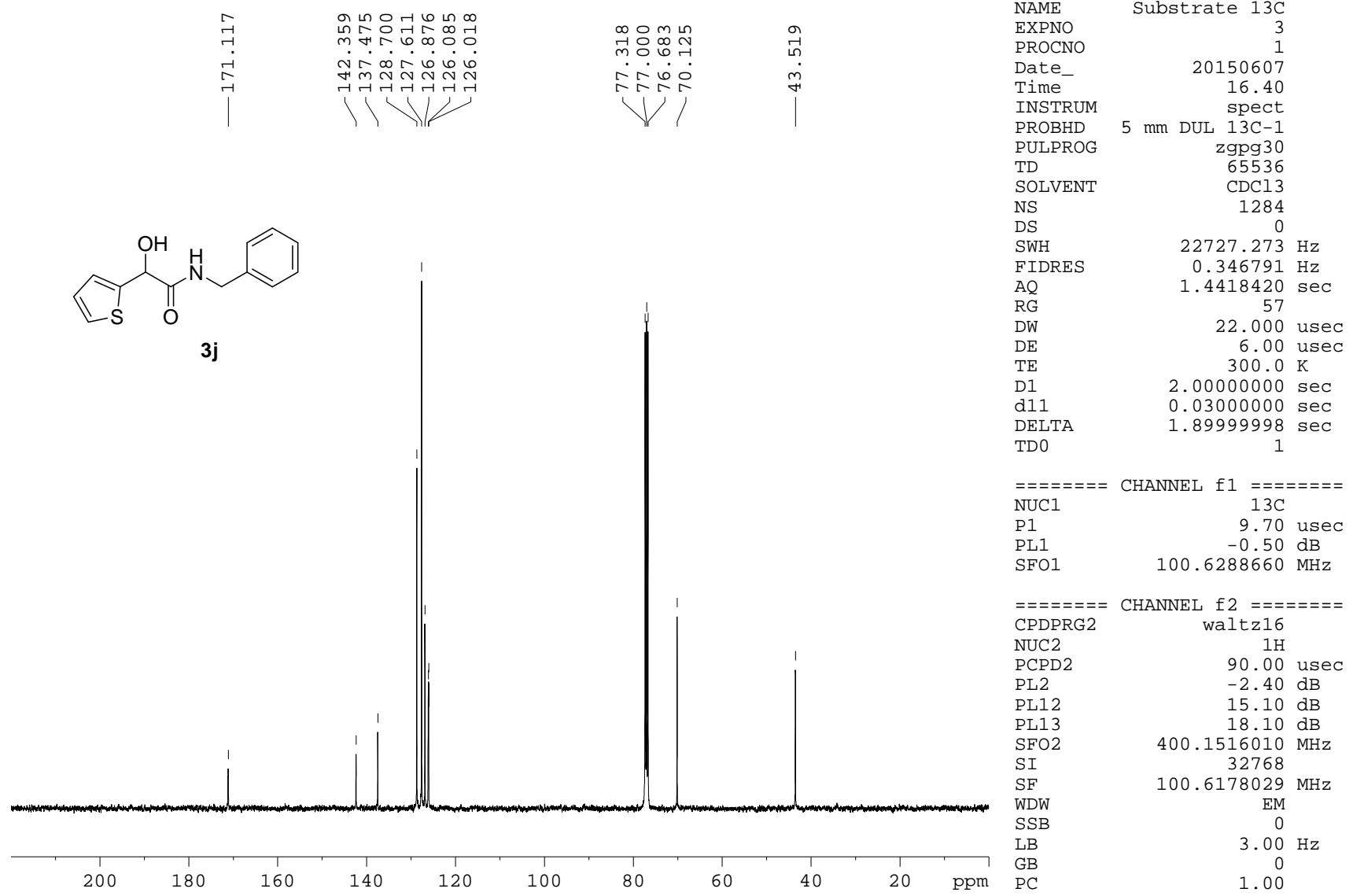
===== CHANNEL f1 =====
NUC1           13C
P1            9.70 usec
PL1           -0.50 dB
SFO1        100.6288660 MHz

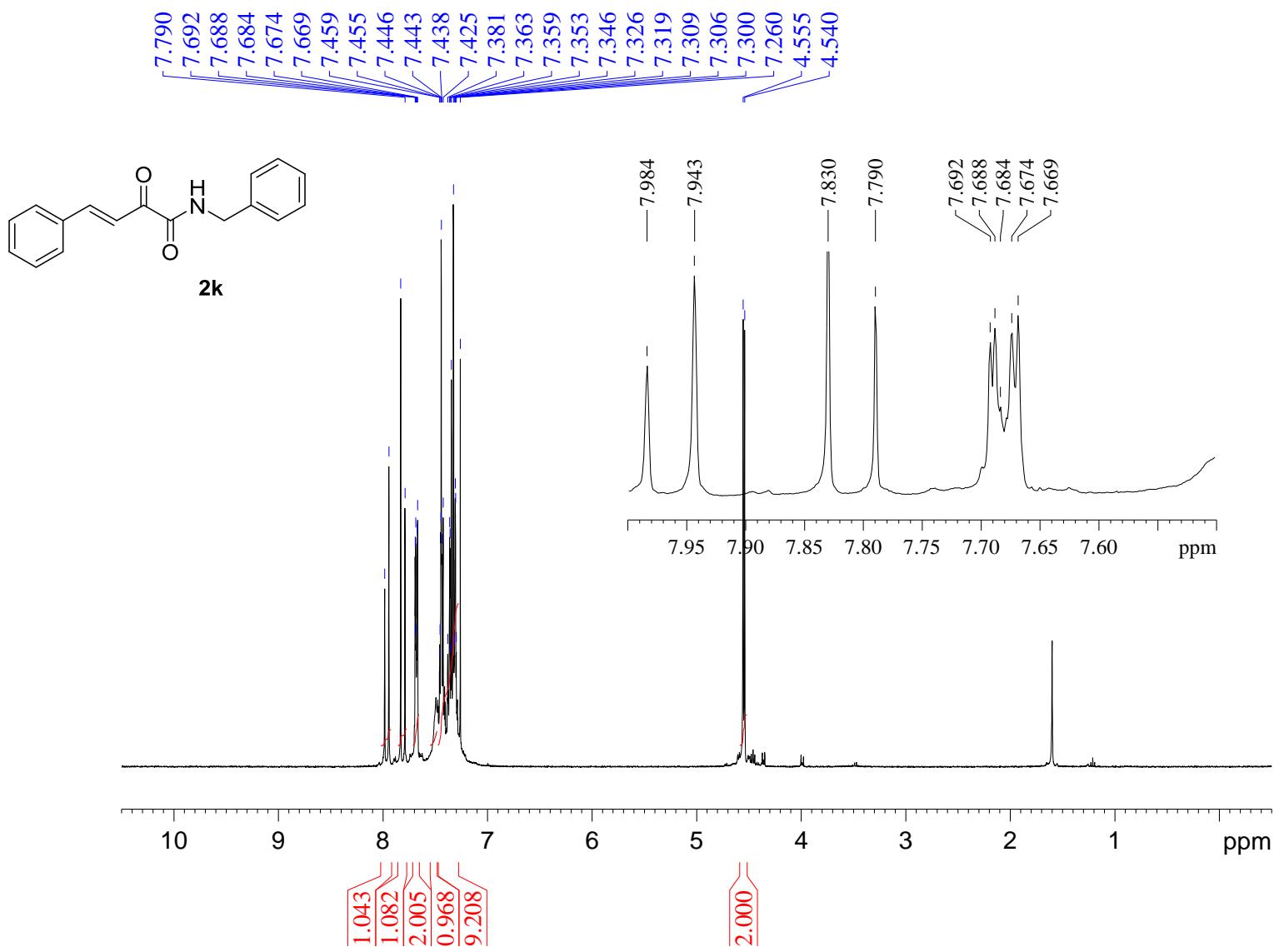
===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2           1H
PCPD2         90.00 usec
PL2            -2.40 dB
PL12          15.10 dB
PL13          18.10 dB
SFO2        400.1516010 MHz
SI            32768
SF           100.6178066 MHz
WDW            EM
SSB             0
LB            3.00 Hz
GB             0
PC            1.00
  
```

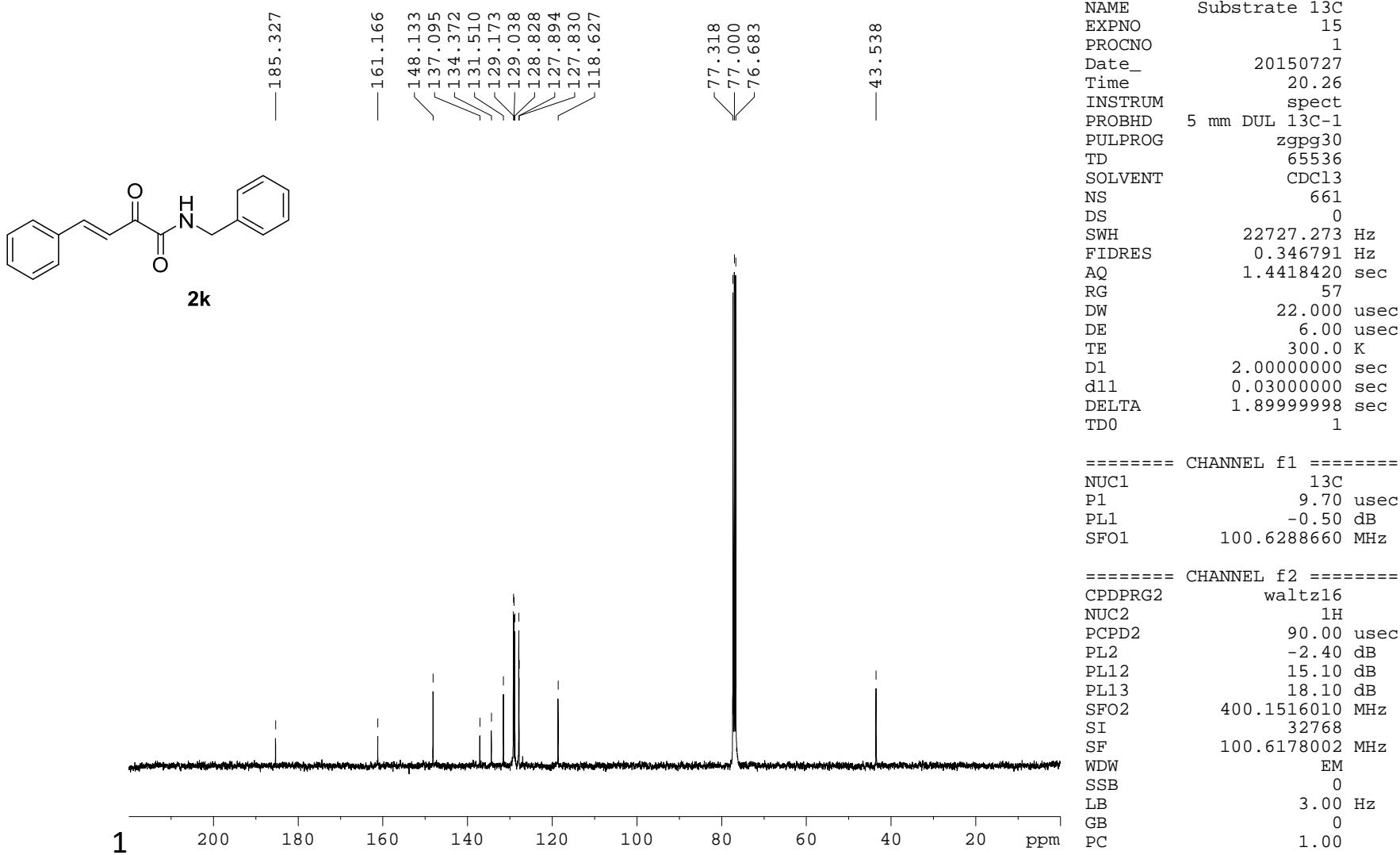


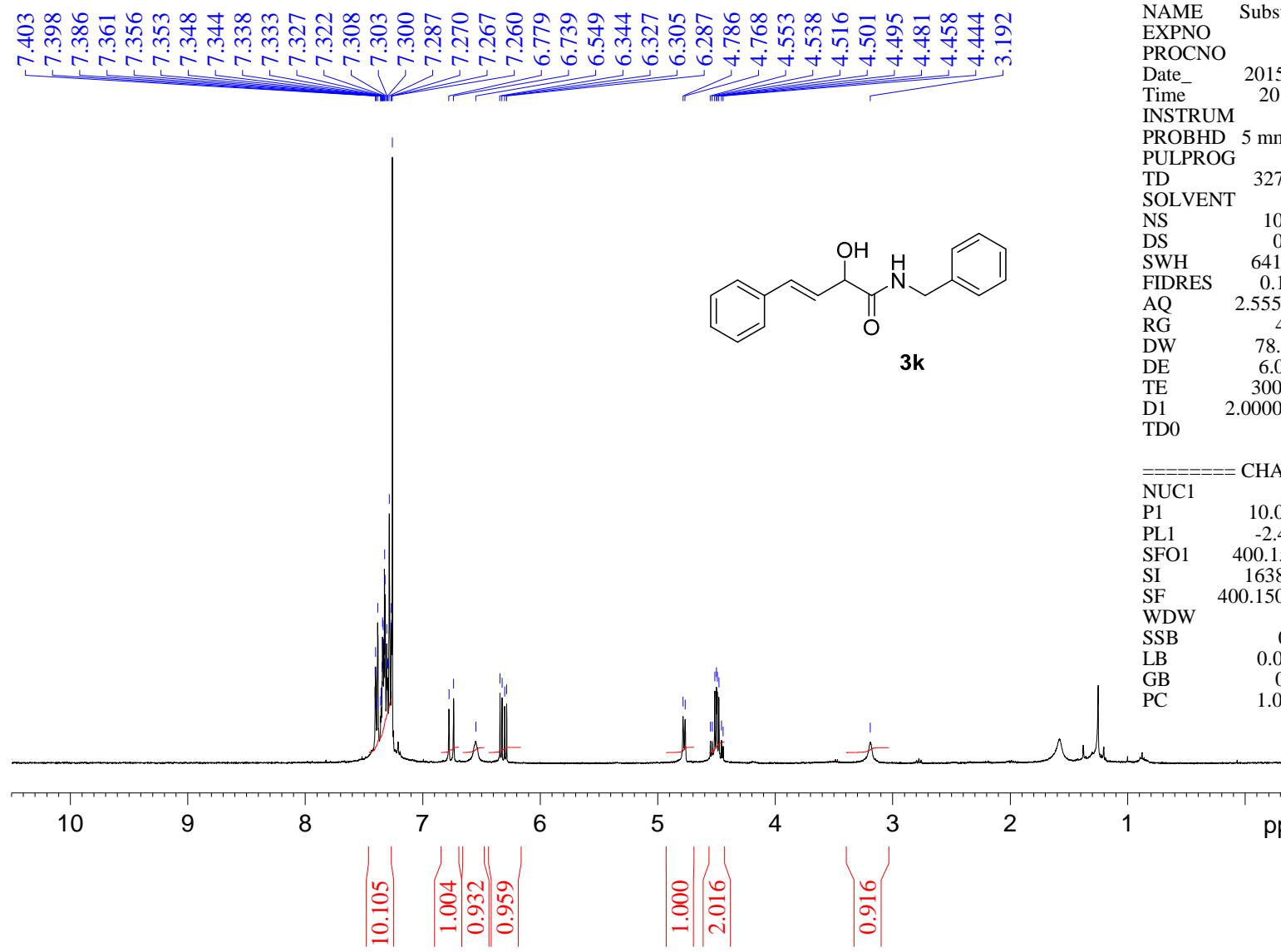
NAME Substrate 1H
 EXPNO 3
 PROCNO 1
 Date_ 20150607
 Time 16.32
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 32768
 SOLVENT CDCl₃
 NS 48
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.195625 Hz
 AQ 2.5559540 sec
 RG 4
 DW 78.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 TD0 1

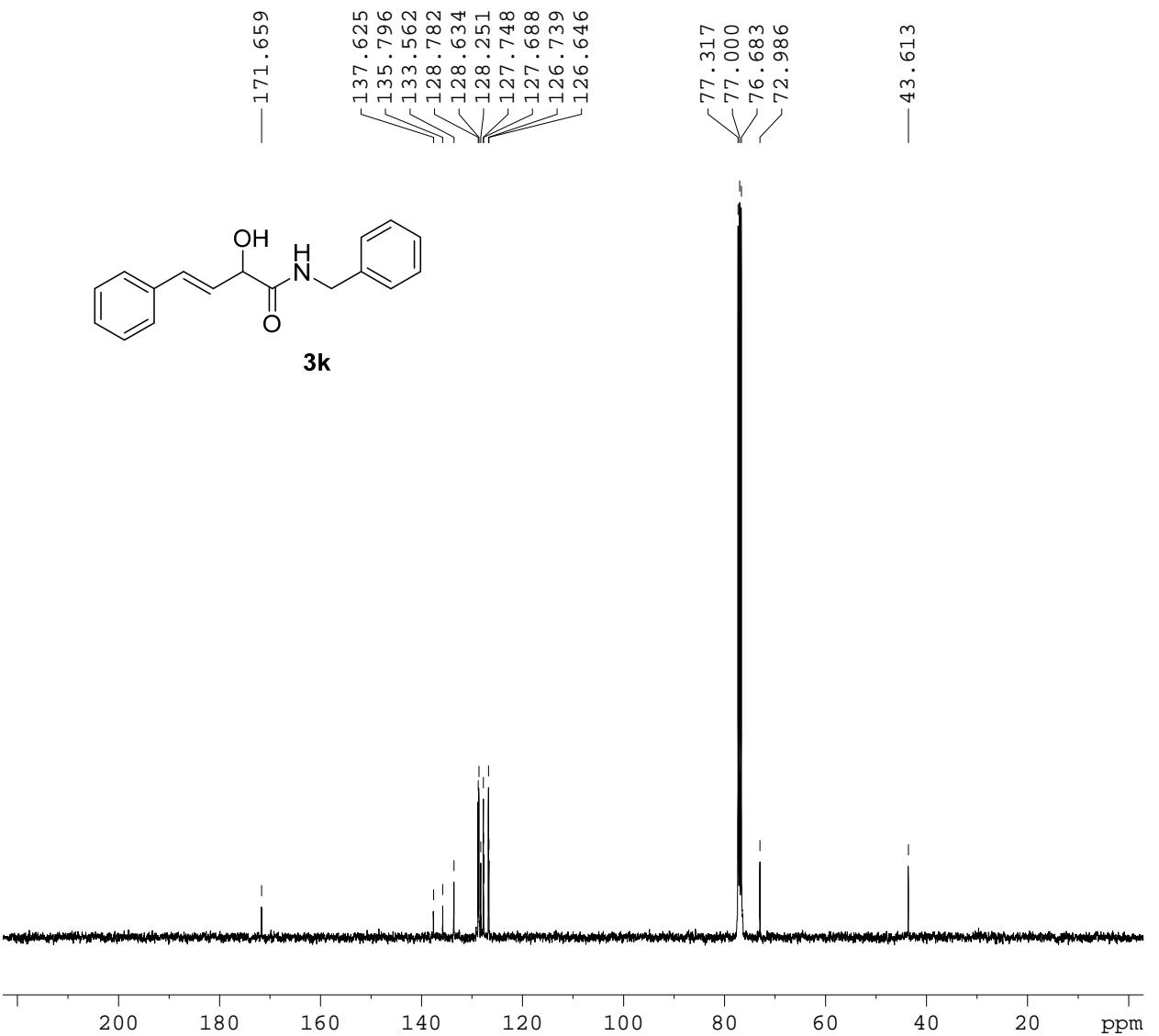
 ===== CHANNEL f1 ======
 NUC1 1H
 P1 10.00 usec
 PL1 -2.40 dB
 SFO1 400.1528010 MHz
 SI 16384
 SF 400.1500088 MHz
 WDW EM
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00











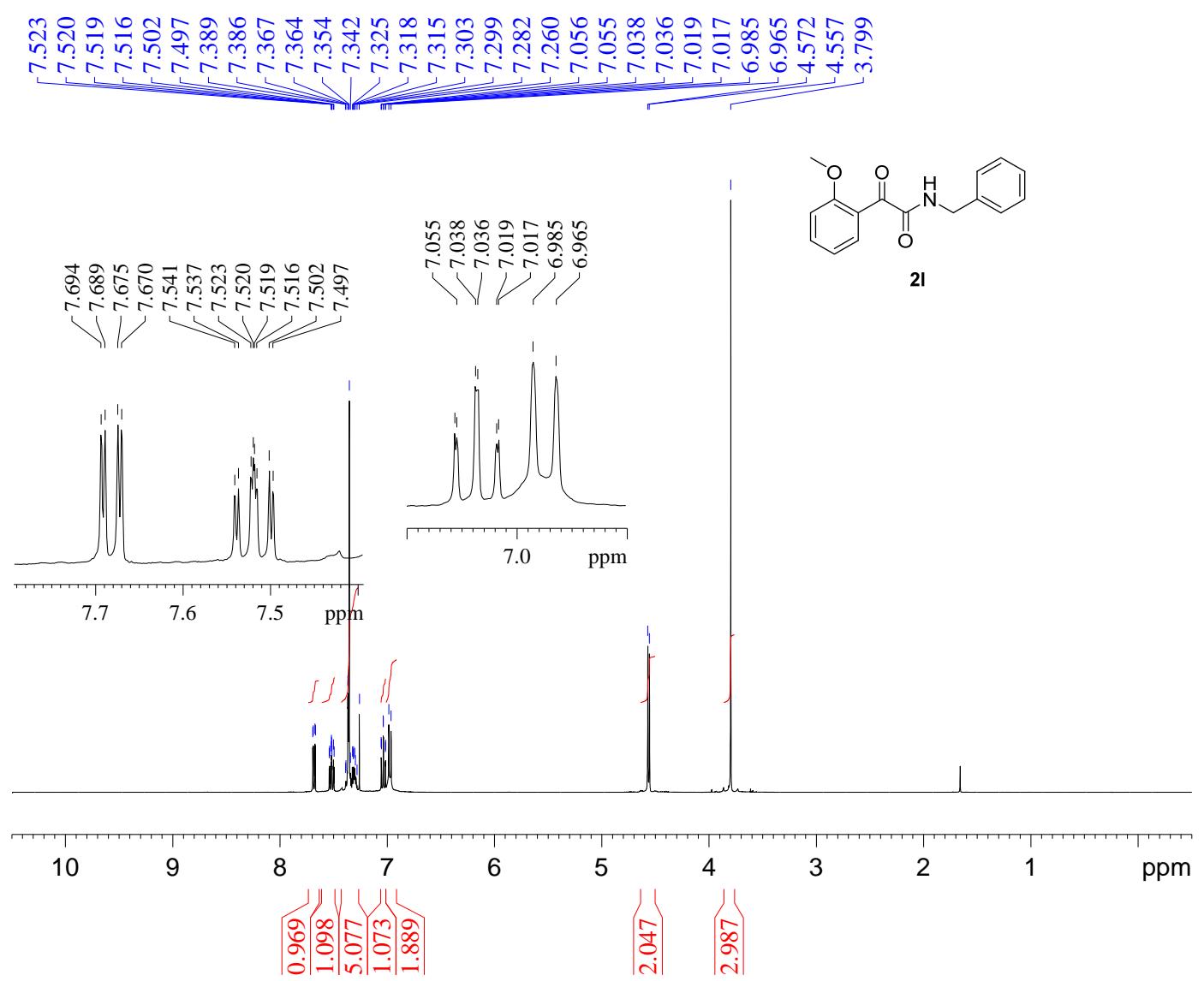
```

NAME           Substrate 13C
EXPNO          16
PROCNO         1
Date_        20150729
Time         20.20
INSTRUM       spect
PROBHD      5 mm DUL 13C-1
PULPROG      zgpg30
TD             65536
SOLVENT        CDC13
NS              723
DS              0
SWH            22727.273 Hz
FIDRES       0.346791 Hz
AQ            1.4418420 sec
RG              57
DW             22.000 usec
DE              6.00 usec
TE              300.0 K
D1            2.00000000 sec
d11           0.03000000 sec
DELTA        1.89999998 sec
TD0                 1

===== CHANNEL f1 =====
NUC1            13C
P1              9.70 usec
PL1            -0.50 dB
SFO1        100.6288660 MHz

===== CHANNEL f2 =====
CPDPRG2        waltz16
NUC2            1H
PCPD2         90.00 usec
PL2            -2.40 dB
PL12           15.10 dB
PL13           18.10 dB
SFO2        400.1516010 MHz
SI              32768
SF            100.6178001 MHz
WDW                EM
SSB                  0
LB              3.00 Hz
GB                  0
PC              1.00

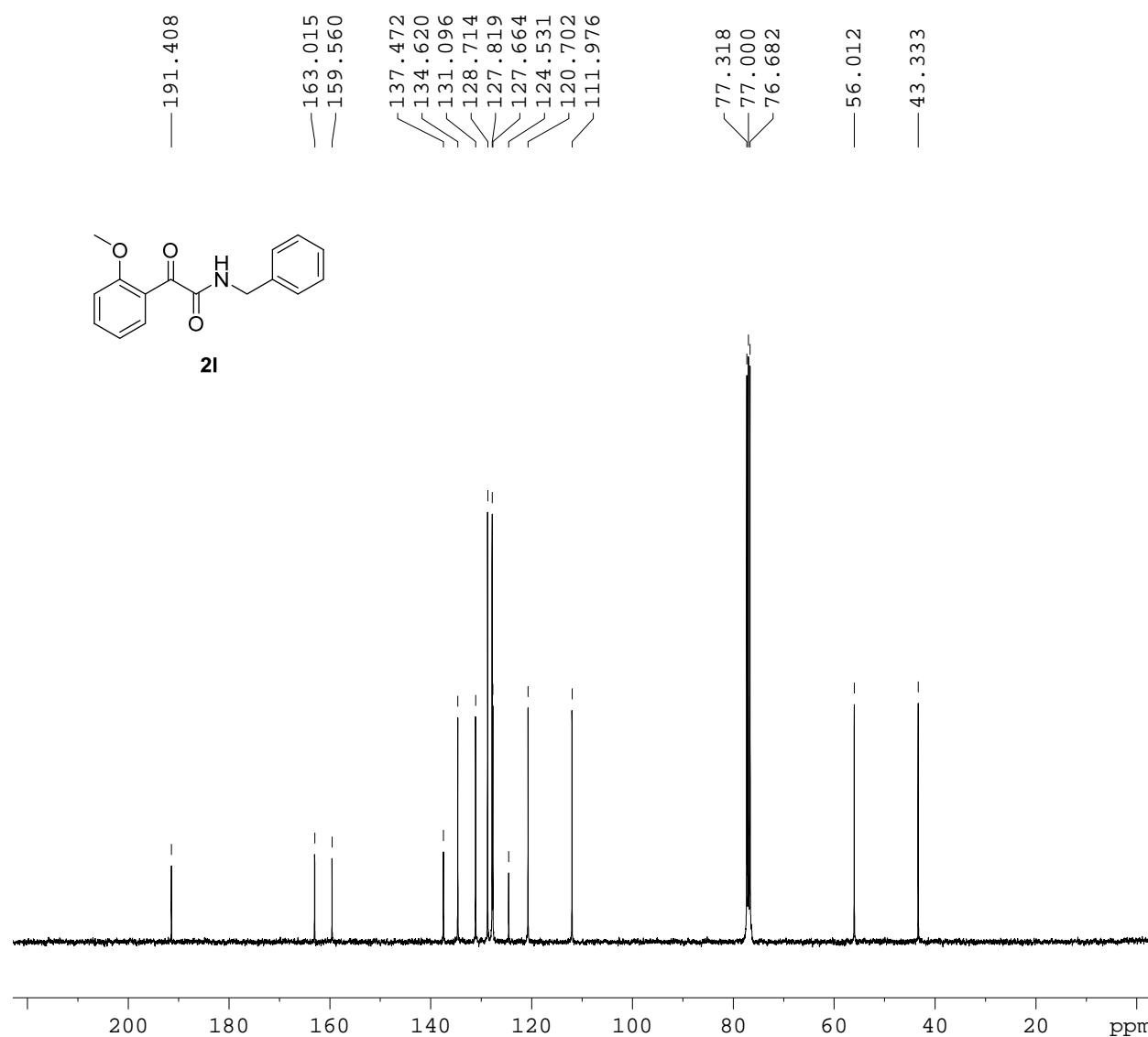
```



NAME	Substrate 1H
EXPNO	10
PROCNO	1
Date_	20150719
Time	14.05
INSTRUM	spec
PROBHD	5 mm DUL 13C-1
PULPROG	zg30
TD	32768
SOLVENT	CDCl ₃
NS	32
DS	0
SWH	6410.256 Hz
FIDRES	0.195625 Hz
AQ	2.5559540 sec
RG	4
DW	78.000 usec
DE	6.00 usec
TE	300.0 K
D1	2.0000000 sec
TD0	1

===== CHANNEL f1 =====

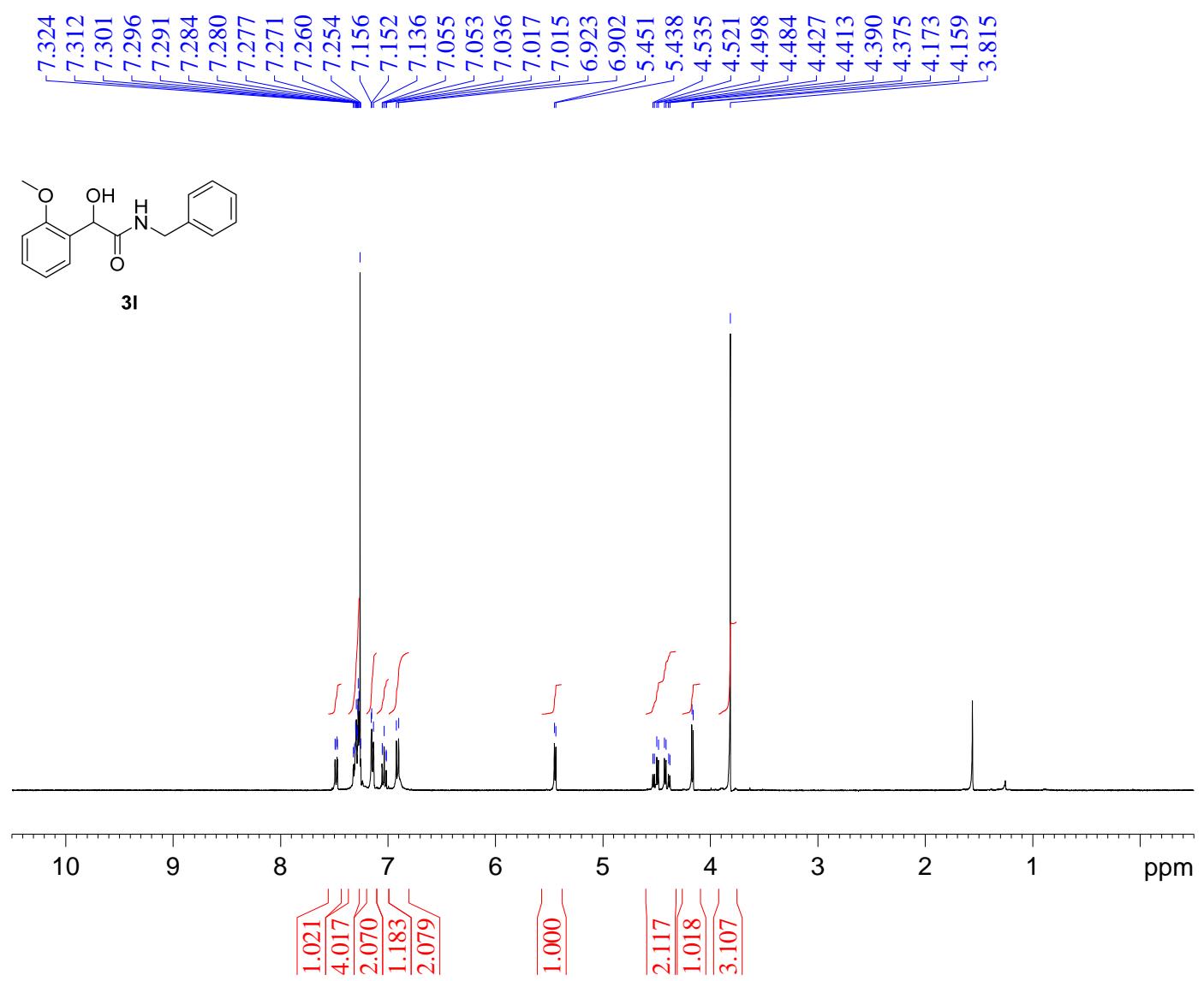
NUC1	1H
P1	10.00 usec
PL1	-2.40 dB
SFO1	400.1528010 MHz
SI	16384
SF	400.1500088 MHz
WDW	EM
SSB	0
LB	0.00 Hz
GB	0
PC	1.00



NAME Substrate 13C
 EXPNO 10
 PROCNO 1
 Date_ 20150719
 Time 14.09
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgppg30
 TD 65536
 SOLVENT CDCl3
 NS 1239
 DS 0
 SWH 22727.273 Hz
 FIDRES 0.346791 Hz
 AQ 1.4418420 sec
 RG 57
 DW 22.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 DELTA 1.8999999 sec
 TDO 1

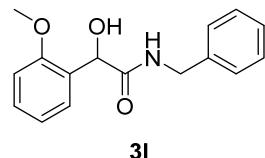
===== CHANNEL f1 =====
 NUC1 13C
 P1 9.70 usec
 PL1 -0.50 dB
 SFO1 100.6288660 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 -2.40 dB
 PL12 15.10 dB
 PL13 18.10 dB
 SFO2 400.1516010 MHz
 SI 32768
 SF 100.6178033 MHz
 WDW EM
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.00

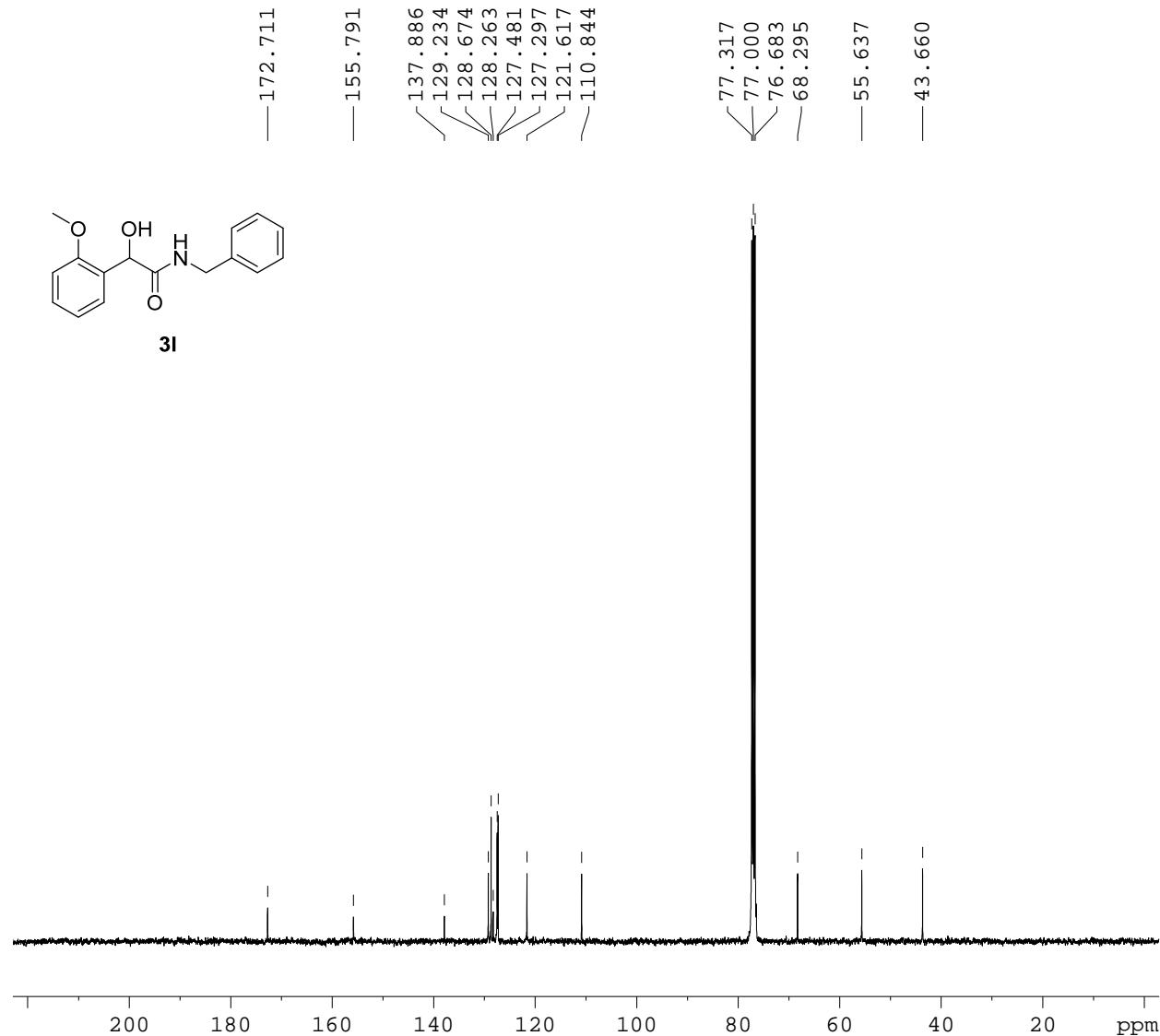


NAME Substrate 1H
 EXPNO 9
 PROCNO 1
 Date_ 20150718
 Time 12.28
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 32768
 SOLVENT CDCl₃
 NS 40
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.195625 Hz
 AQ 2.5559540 sec
 RG 4
 DW 78.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 TD0 1

===== CHANNEL f1 ======
 NUC1 1H
 P1 10.00 usec
 PL1 -2.40 dB
 SFO1 400.1528010 MHz
 SI 16384
 SF 400.1500088 MHz
 WDW EM
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00



3



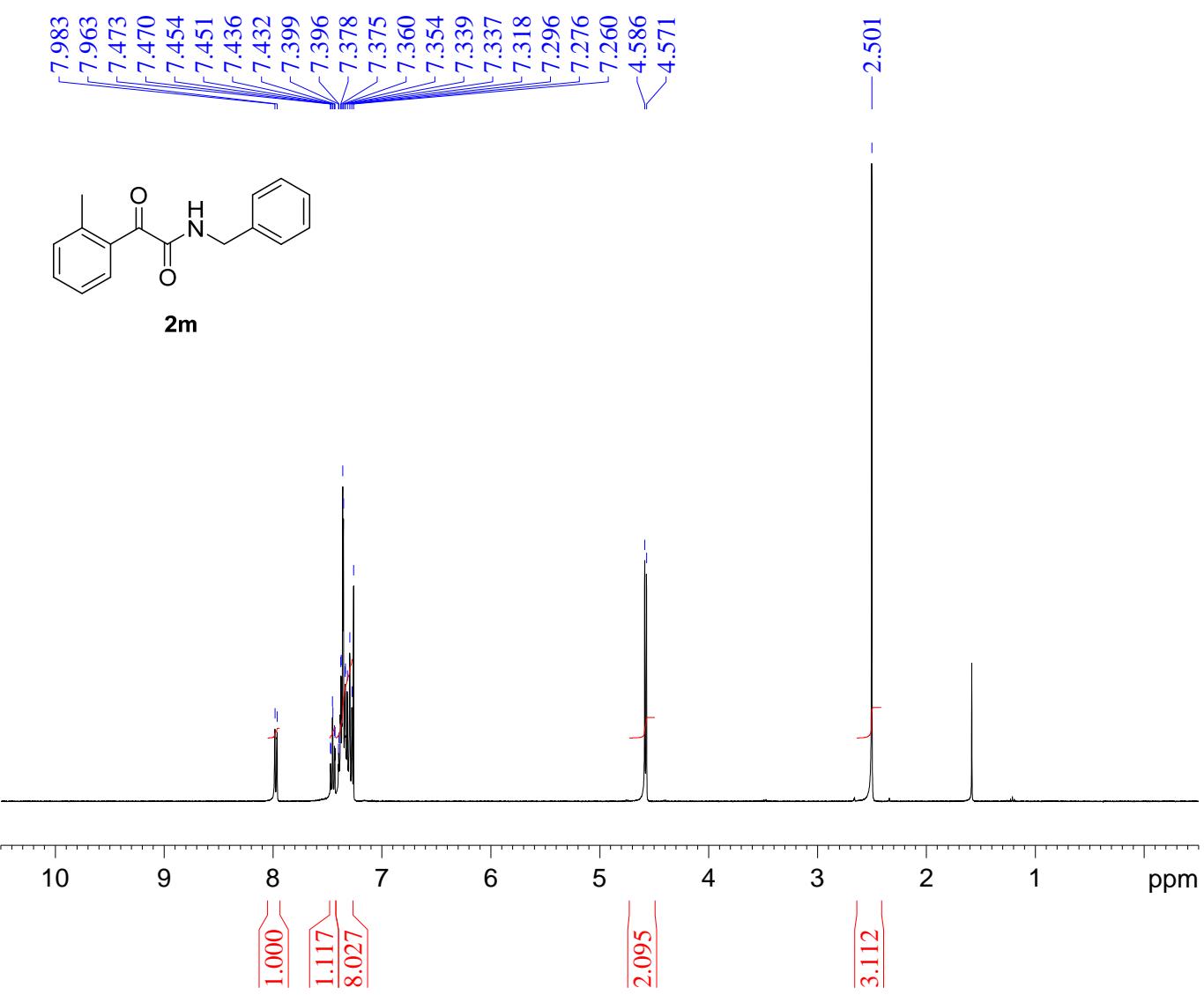
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NAME           Substrate 13C
EXPNO          9
PROCNO         1
Date_        20150718
Time         12.36
INSTRUM      spect
PROBHD      5 mm DUL 13C-1
PULPROG     zpgpg30
TD            65536
SOLVENT      CDC13
NS            2359
DS             0
SWH           22727.273 Hz
FIDRES       0.346791 Hz
AQ            1.4418420 sec
RG             57
DW            22.000 usec
DE             6.00 usec
TE            300.0 K
D1            2.00000000 sec
d11           0.03000000 sec
DELTA         1.89999998 sec
TD0            1

===== CHANNEL f1 =====
NUC1           13C
P1              9.70 usec
PL1            -0.50 dB
SFO1          100.6288660 MHz

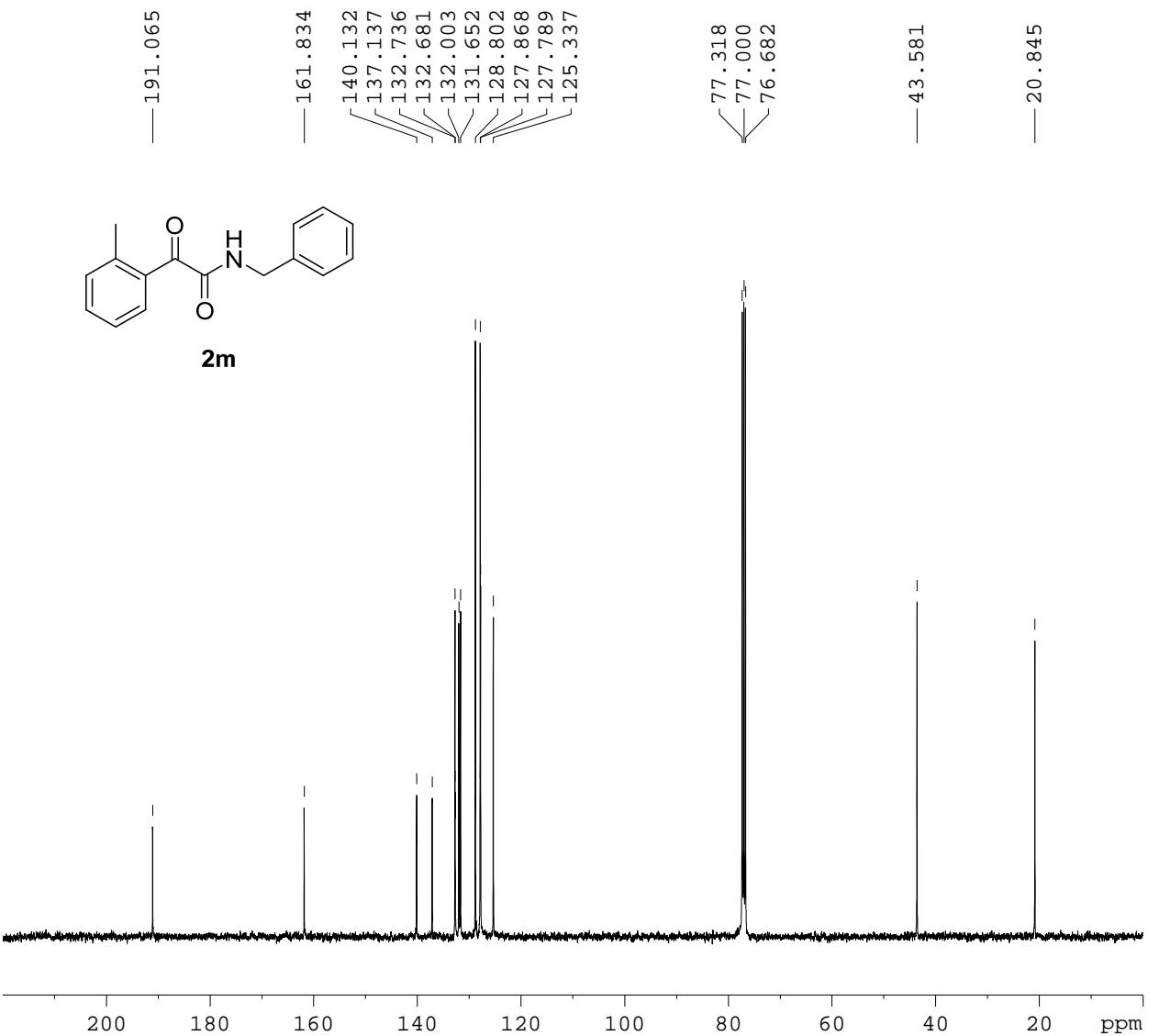
===== CHANNEL f2 =====
CPDPRG2        waltz16
NUC2            1H
PCPD2          90.00 usec
PL2            -2.40 dB
PL12           15.10 dB
PL13           18.10 dB
SFO2          400.1516010 MHz
SI              32768
SF          100.6177996 MHz
WDW             EM
SSB              0
LB              3.00 Hz
GB              0
PC              1.00

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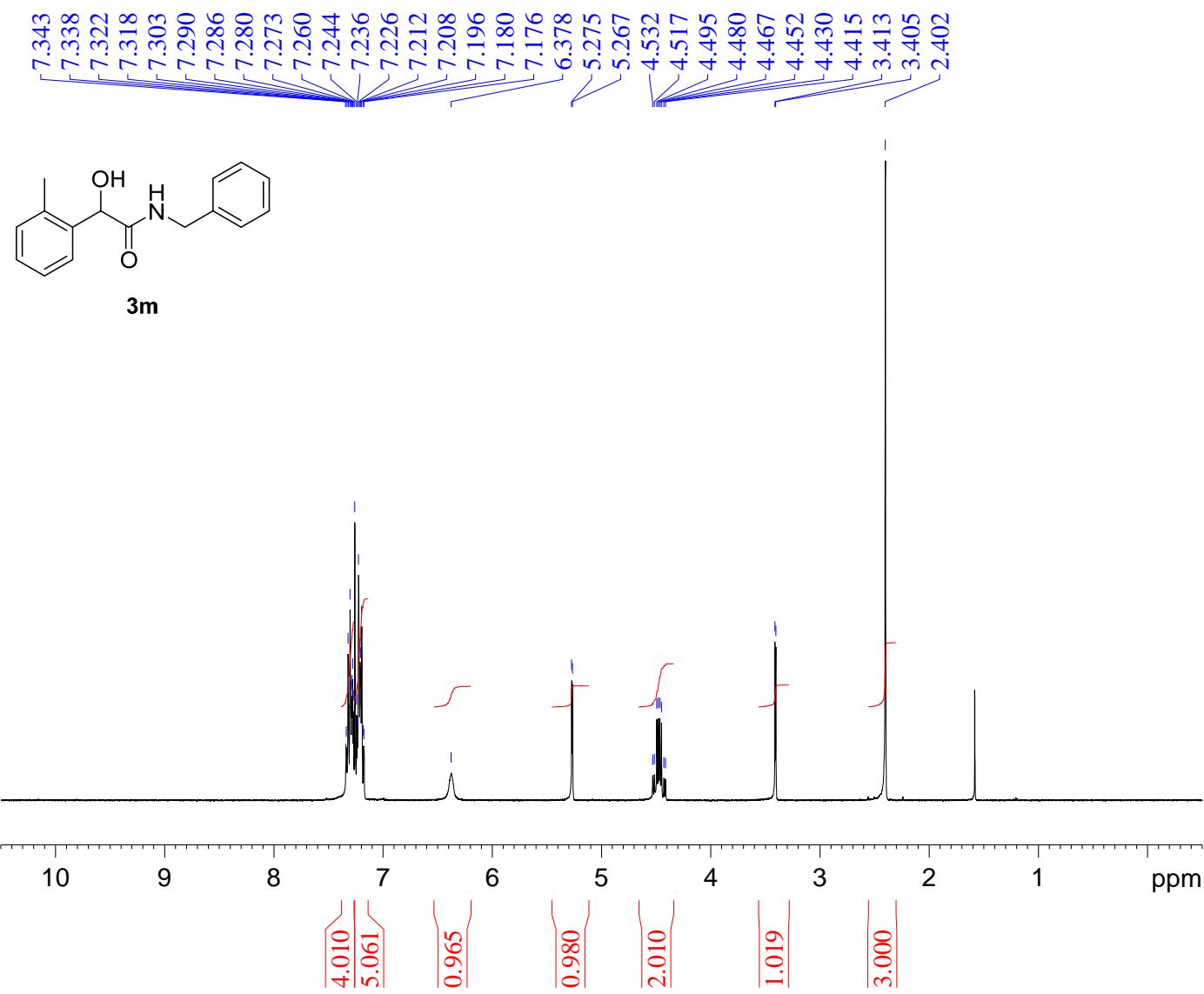


NAME 20151115
EXPNO 1
PROCNO 1
Date_ 20151115
Time 20.05
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 0
SWH 6410.256 Hz
FIDRES 0.195625 Hz
AQ 2.5559540 sec
RG 4
DW 78.000 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 10.00 usec
PL1 -2.40 dB
SFO1 400.1528010 MHz
SI 16384
SF 400.1500089 MHz
WDW EM
SSB 0
LB 0.00 Hz
GB 0
PC 1.00



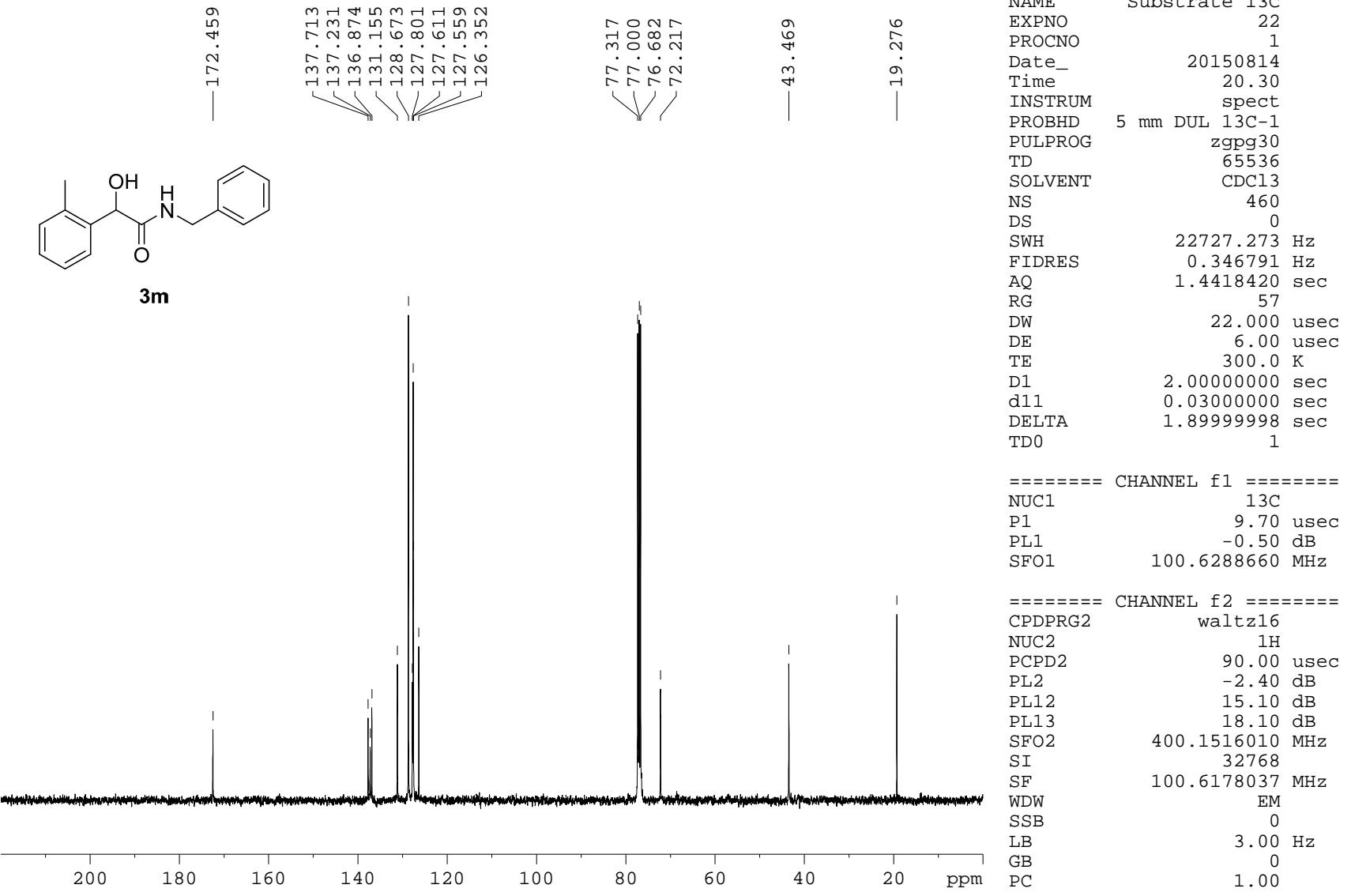
NAME	20151115
EXPNO	2
PROCNO	1
Date_	20151115
Time	20.12
INSTRUM	spect
PROBHDL	5 mm DUL 13C-1
PULPROG	zgpg30
TD	65536
SOLVENT	CDC13
NS	756
DS	0
SWH	22727.273 Hz
FIDRES	0.346791 Hz
AQ	1.4418420 sec
RG	57
DW	22.000 usec
DE	6.00 usec
TE	300.0 K
D1	2.00000000 sec
d11	0.03000000 sec
DELTA	1.89999998 sec
TD0	1
===== CHANNEL f1 =====	
NUC1	13C
P1	9.70 usec
PL1	-0.50 dB
SFO1	100.6288660 MHz
===== CHANNEL f2 =====	
CPDPKG2	waltz16
NUC2	1H
PCPD2	90.00 usec
PL2	-2.40 dB
PL12	15.10 dB
PL13	18.10 dB
SFO2	400.1516010 MHz
SI	32768
SF	100.6178050 MHz
WDW	EM
SSB	0
LB	3.00 Hz
GB	0
PC	1.00

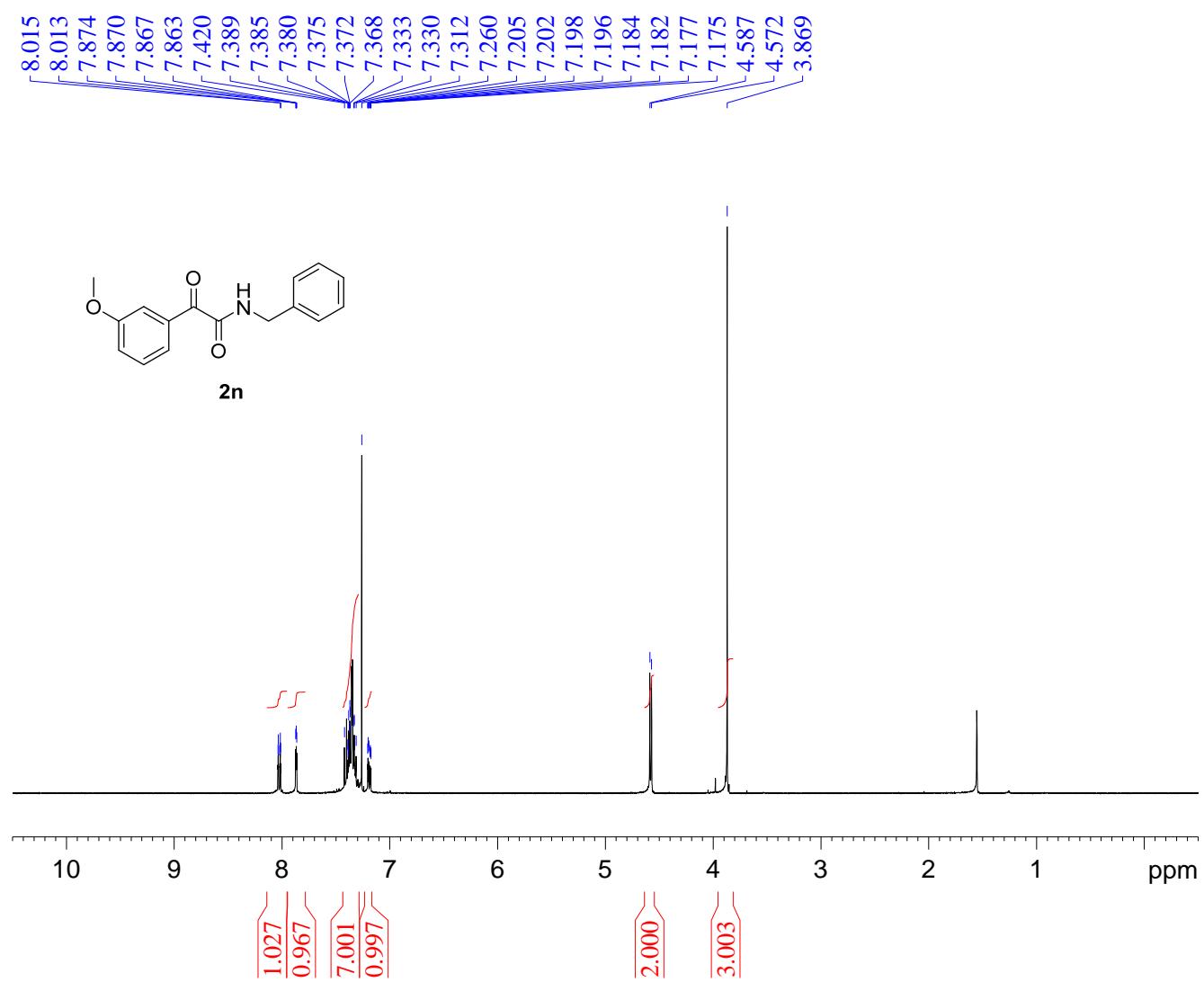


NAME	20160114
EXPNO	2
PROCNO	1
Date_	20160114
Time	20.27
INSTRUM	spect
PROBHD	5 mm DUL 13C-1
PULPROG	zg30
TD	32768
SOLVENT	CDCl ₃
NS	13
DS	0
SWH	6410.256 Hz
FIDRES	0.195625 Hz
AQ	2.5559540 sec
RG	4
DW	78.000 usec
DE	6.00 usec
TE	300.0 K
D1	2.0000000 sec
TD0	1

===== CHANNEL f1 =====

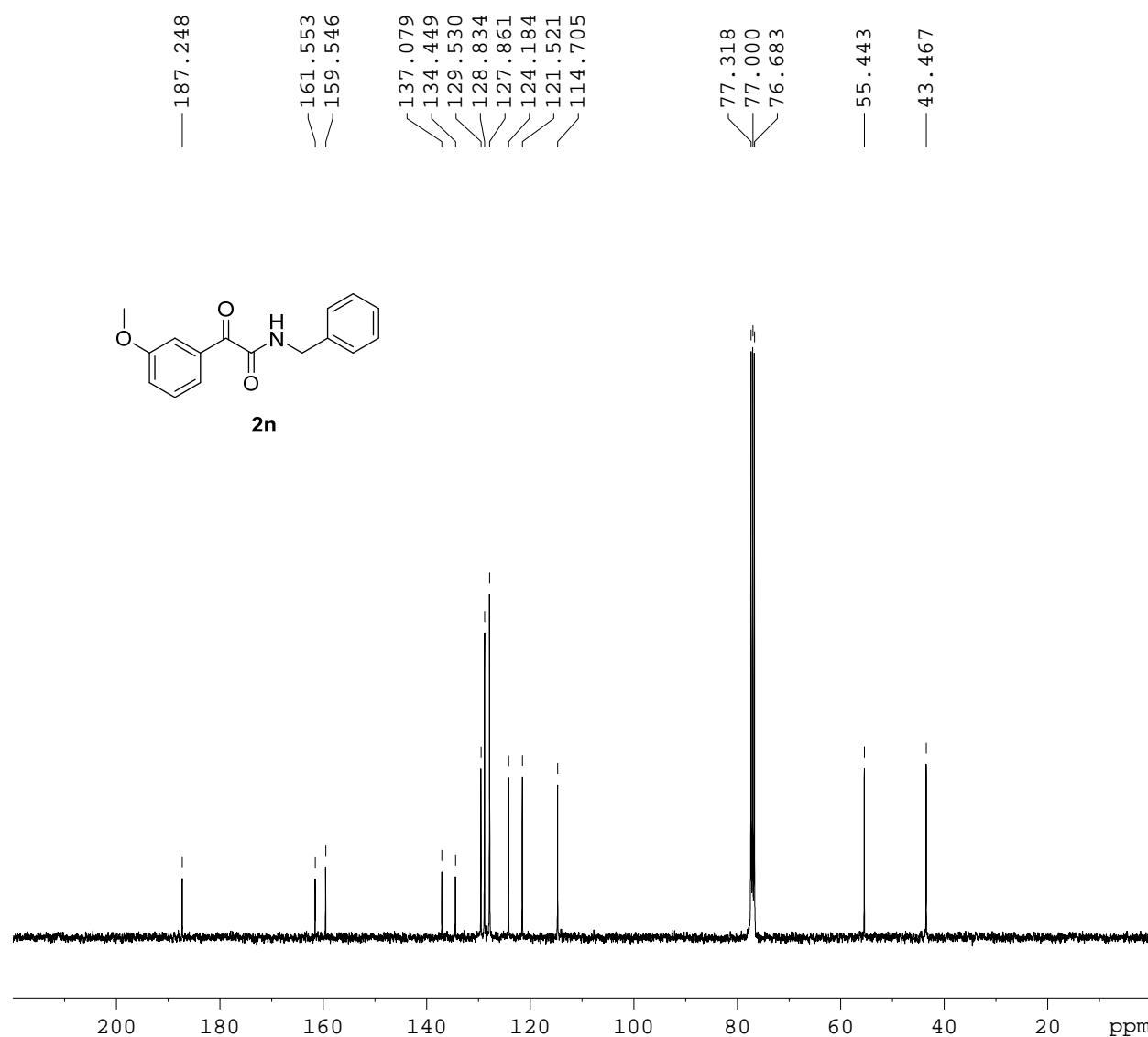
NUC1	1H
P1	10.00 usec
PL1	-2.40 dB
SFO1	400.1528010 MHz
SI	16384
SF	400.1500091 MHz
WDW	EM
SSB	0
LB	0.00 Hz
GB	0
PC	1.00



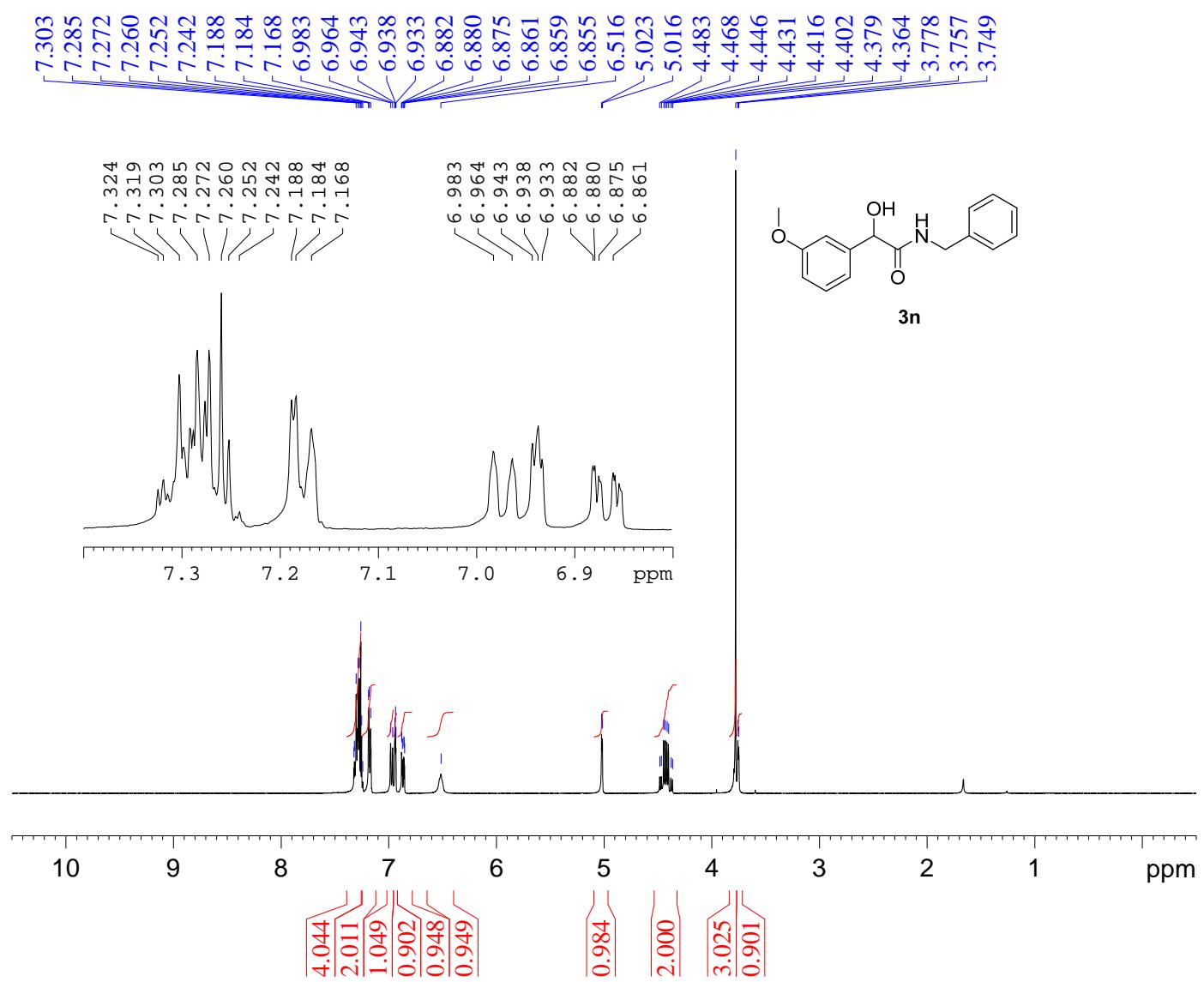


NAME 20151125
 EXPNO 1
 PROCNO 1
 Date_ 20151125
 Time 19.08
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 32768
 SOLVENT CDCl₃
 NS 30
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.195625 Hz
 AQ 2.5559540 sec
 RG 4
 DW 78.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 10.00 usec
 PL1 -2.40 dB
 SFO1 400.1528010 MHz
 SI 16384
 SF 400.1500089 MHz
 WDW EM
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00

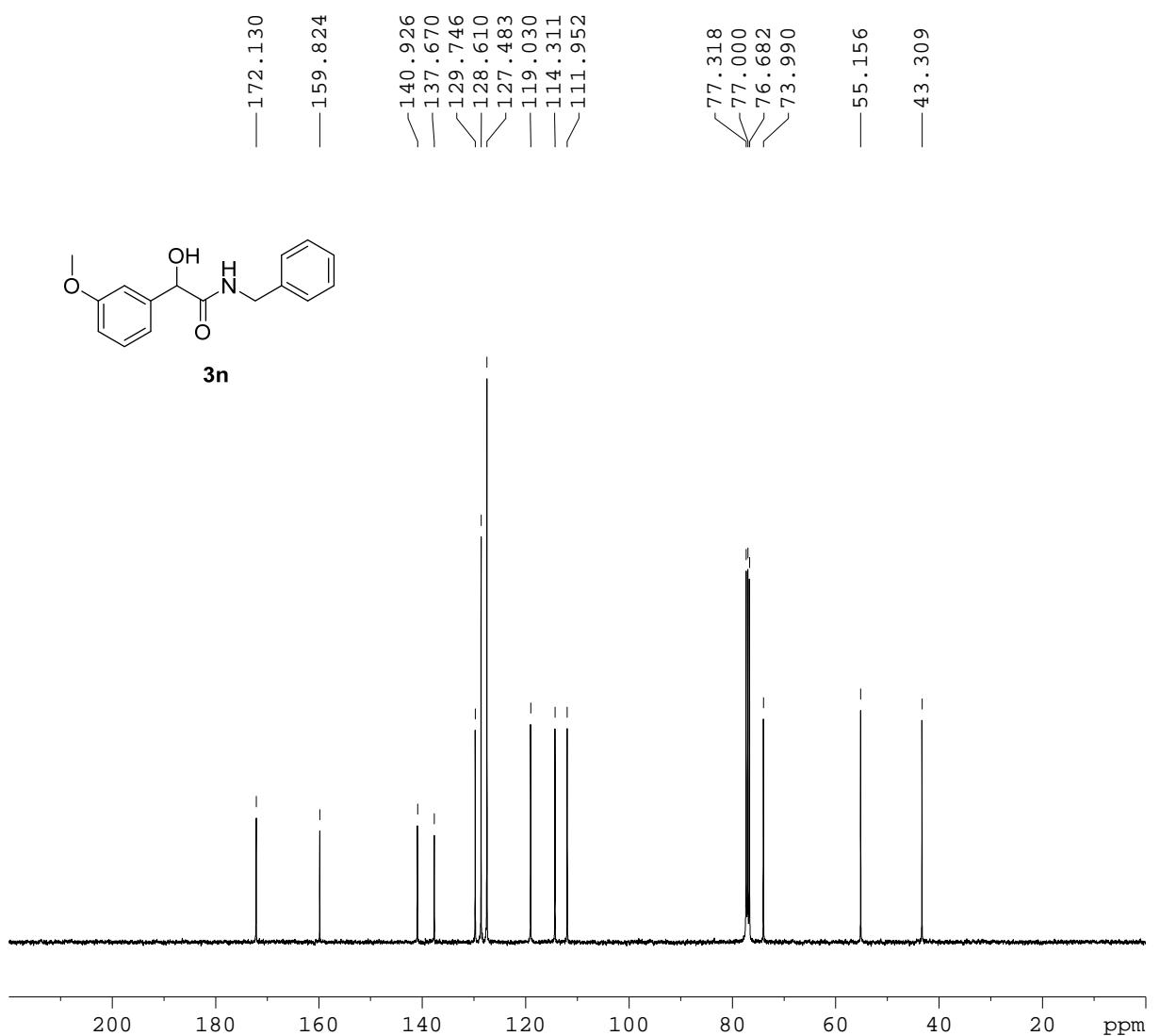


NAME	20151125
EXPNO	2
PROCNO	1
Date_	20151125
Time	19.38
INSTRUM	spect
PROBHD	5 mm DUL 13C-1
PULPROG	zgpg30
TD	65536
SOLVENT	CDCl ₃
NS	492
DS	0
SWH	22727.273 Hz
FIDRES	0.346791 Hz
AQ	1.4418420 sec
RG	57
DW	22.000 usec
DE	6.00 usec
TE	300.0 K
D1	2.0000000 sec
d11	0.03000000 sec
DELTA	1.89999998 sec
T0D0	1
===== CHANNEL f1 =====	
NUC1	13C
P1	9.70 usec
PL1	-0.50 dB
SFO1	100.6288660 MHz
===== CHANNEL f2 =====	
CPDPGR2	waltz16
NUC2	1H
PCPD2	90.00 usec
PL2	-2.40 dB
PL12	15.10 dB
PL13	18.10 dB
SFO2	400.1516010 MHz
SI	32768
SF	100.6178017 MHz
WDW	EM
SSB	0
LB	3.00 Hz
GB	0
PC	1.00



NAME	20151116
EXPNO	1
PROCNO	1
Date_	20151116
Time	20.08
INSTRUM	spect
PROBHD	5 mm DUL
PULPROG	13C-1 zg30
TD	32768
SOLVENT	CDCl3
NS	16
DS	0
SWH	6410.256 Hz
FIDRES	0.195625 Hz
AQ	2.5559540 sec
RG	4
DW	78.000 usec
DE	6.00 usec
TE	300.0 K
D1	2.0000000 sec
TD0	1

===== CHANNEL f1 =====
NUC1 1H
P1 10.00 usec
PL1 -2.40 dB
SFO1 400.1528010 MHz
SI 16384
SF 400.1500089 MHz
WDW EM
SSB 0
LB 0.00 Hz
GB 0
PC 1.00



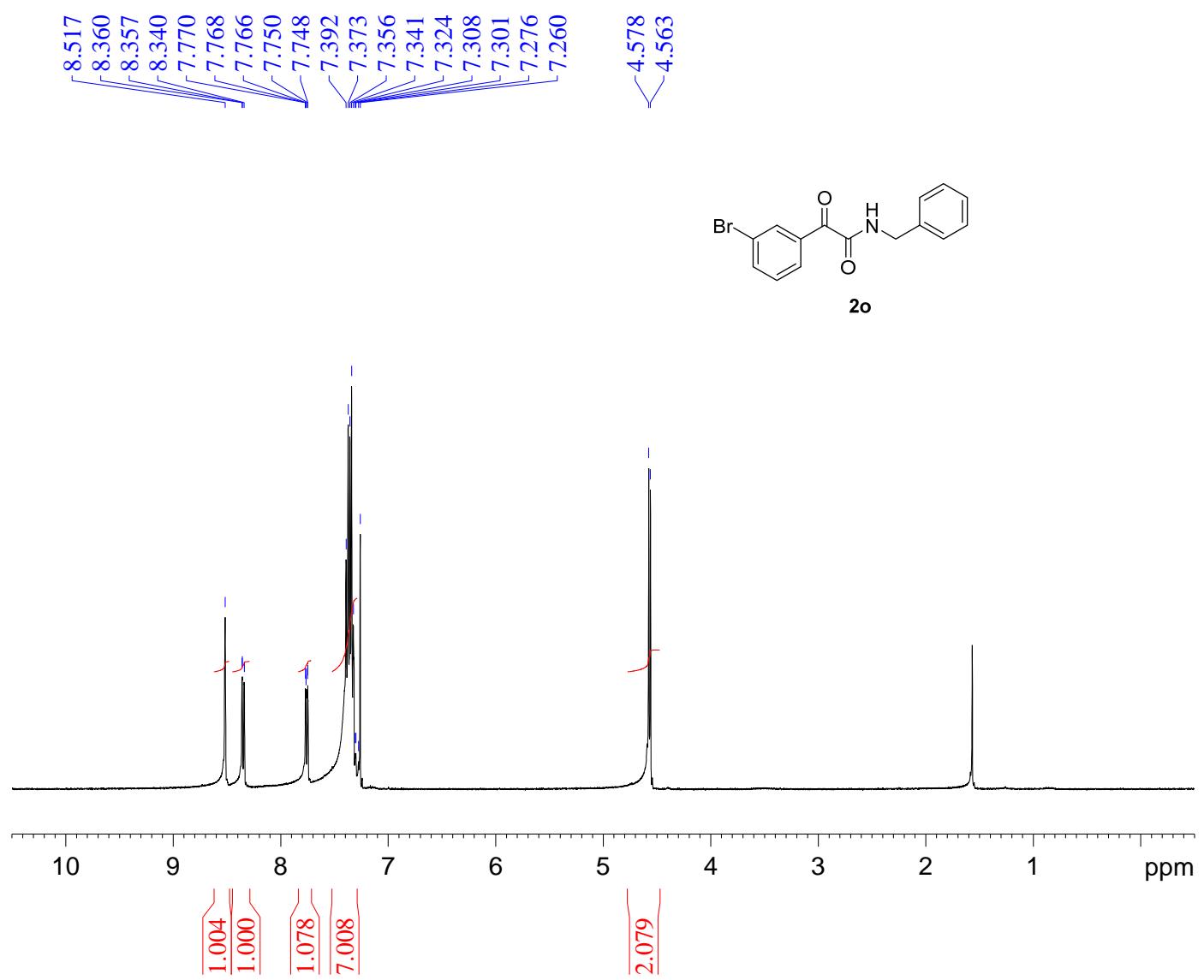
NAME 20151116
EXPNO 2
PROCNO 1
Date_ 20151116
Time 20.15
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 748
DS 0
SWH 22727.273 Hz
FIDRES 0.346791 Hz
AQ 1.4418420 sec
RG 57
DW 22.000 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.8999998 sec
TDO 1

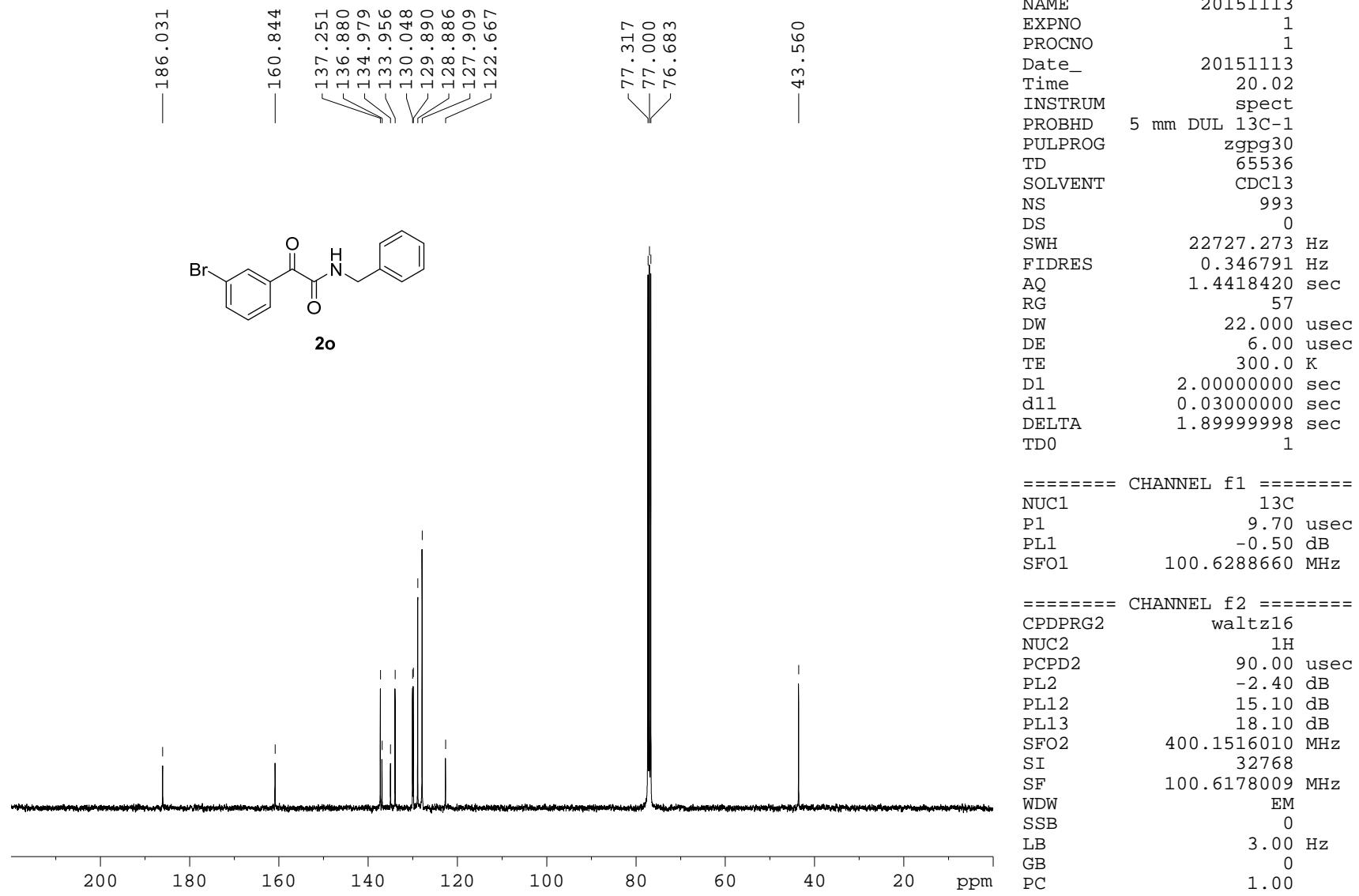
===== CHANNEL f1 ======

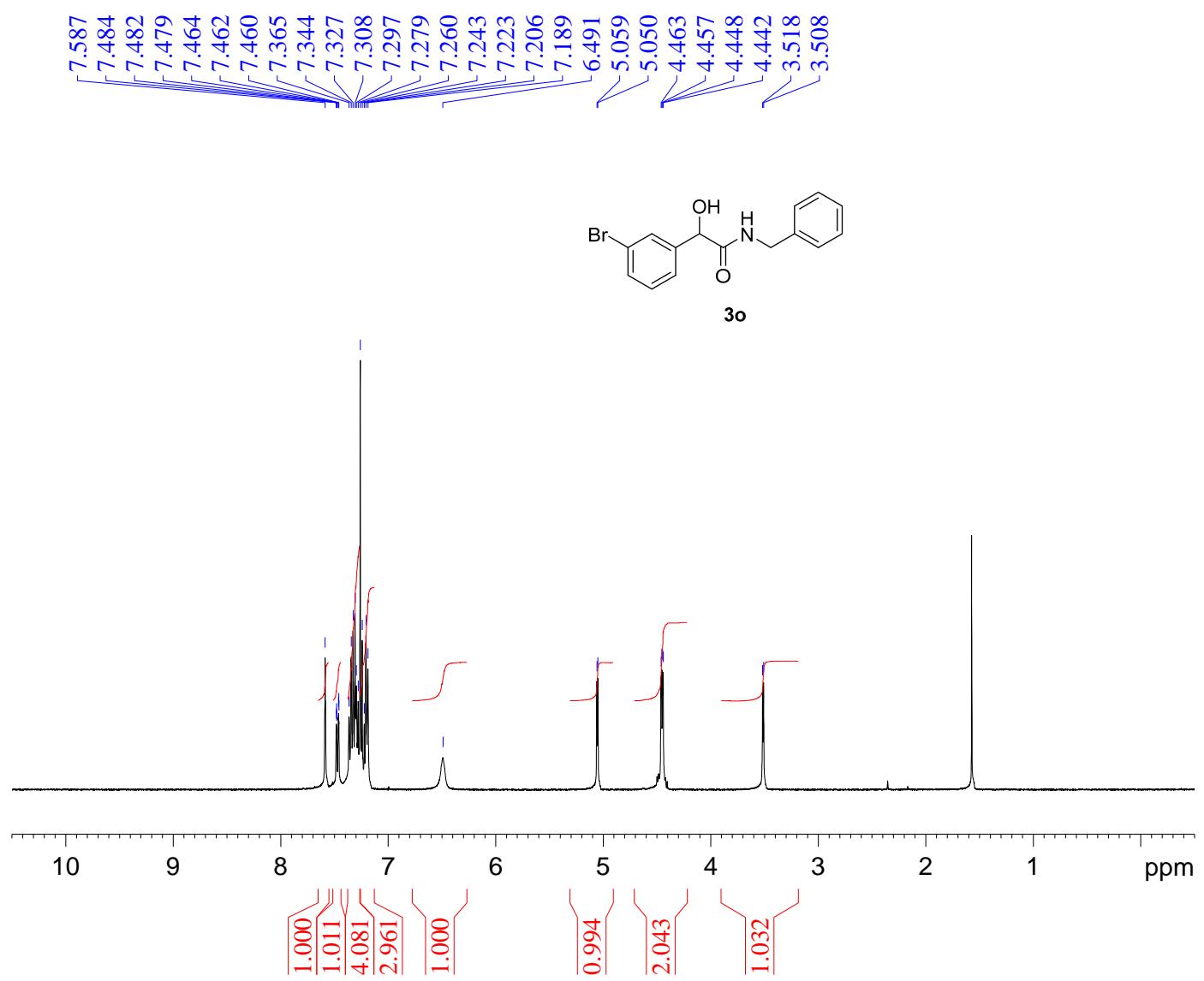
NUC1 13C
P1 9.70 usec
PL1 -0.50 dB
SFO1 100.6288660 MHz

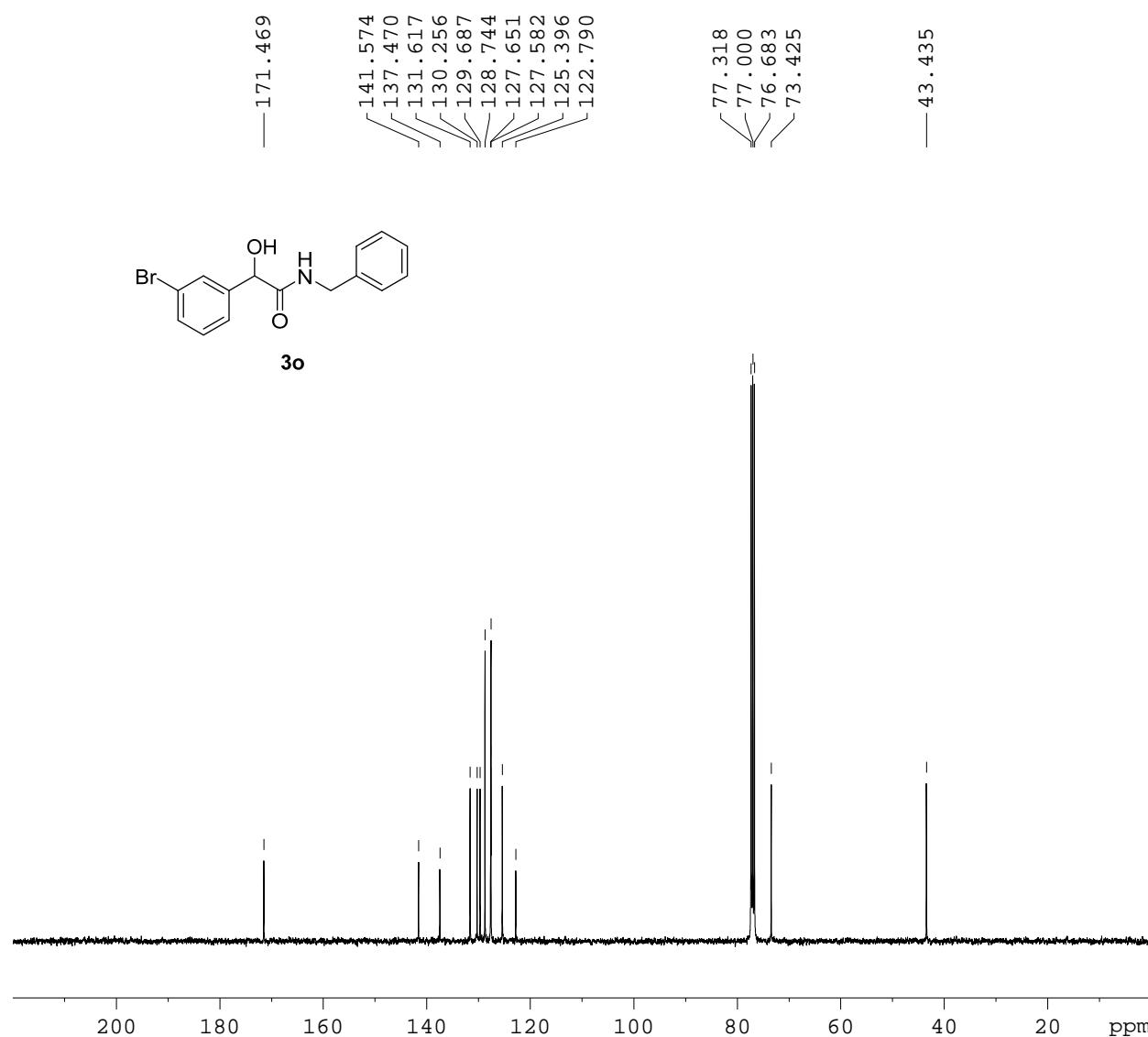
===== CHANNEL f2 ======

CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PL2 -2.40 dB
PL12 15.10 dB
PL13 18.10 dB
SFO2 400.1516010 MHz
SI 32768
SF 100.6178066 MHz
WDW EM
SSB 0
LB 3.00 Hz
GB 0
PC 1.00









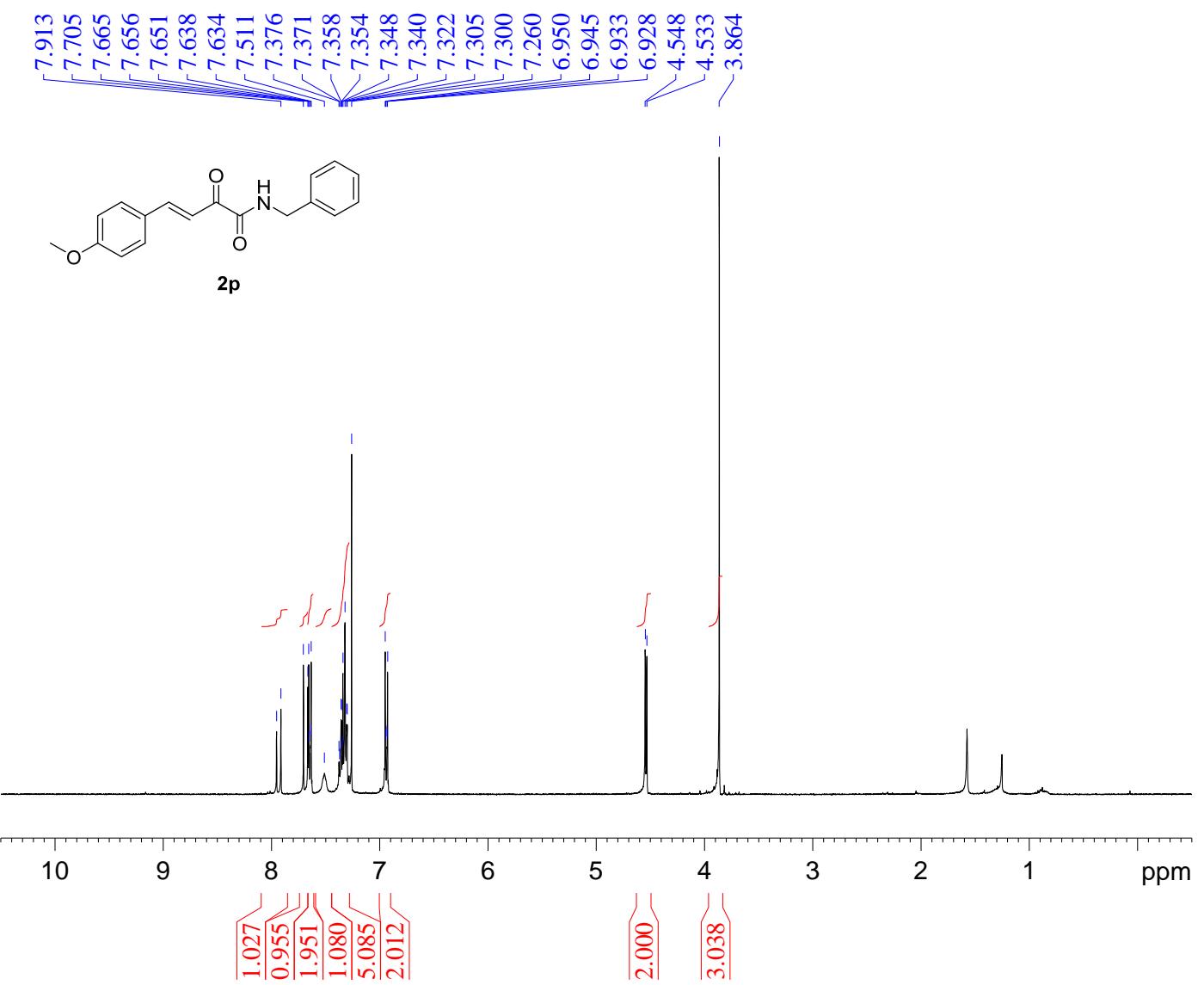
NAME 20151114
 EXPNO 1
 PROCNO 1
 Date_ 20151114
 Time 20.09
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 752
 DS 0
 SWH 22727.273 Hz
 FIDRES 0.346791 Hz
 AQ 1.4418420 sec
 RG 57
 DW 22.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 TDO 1

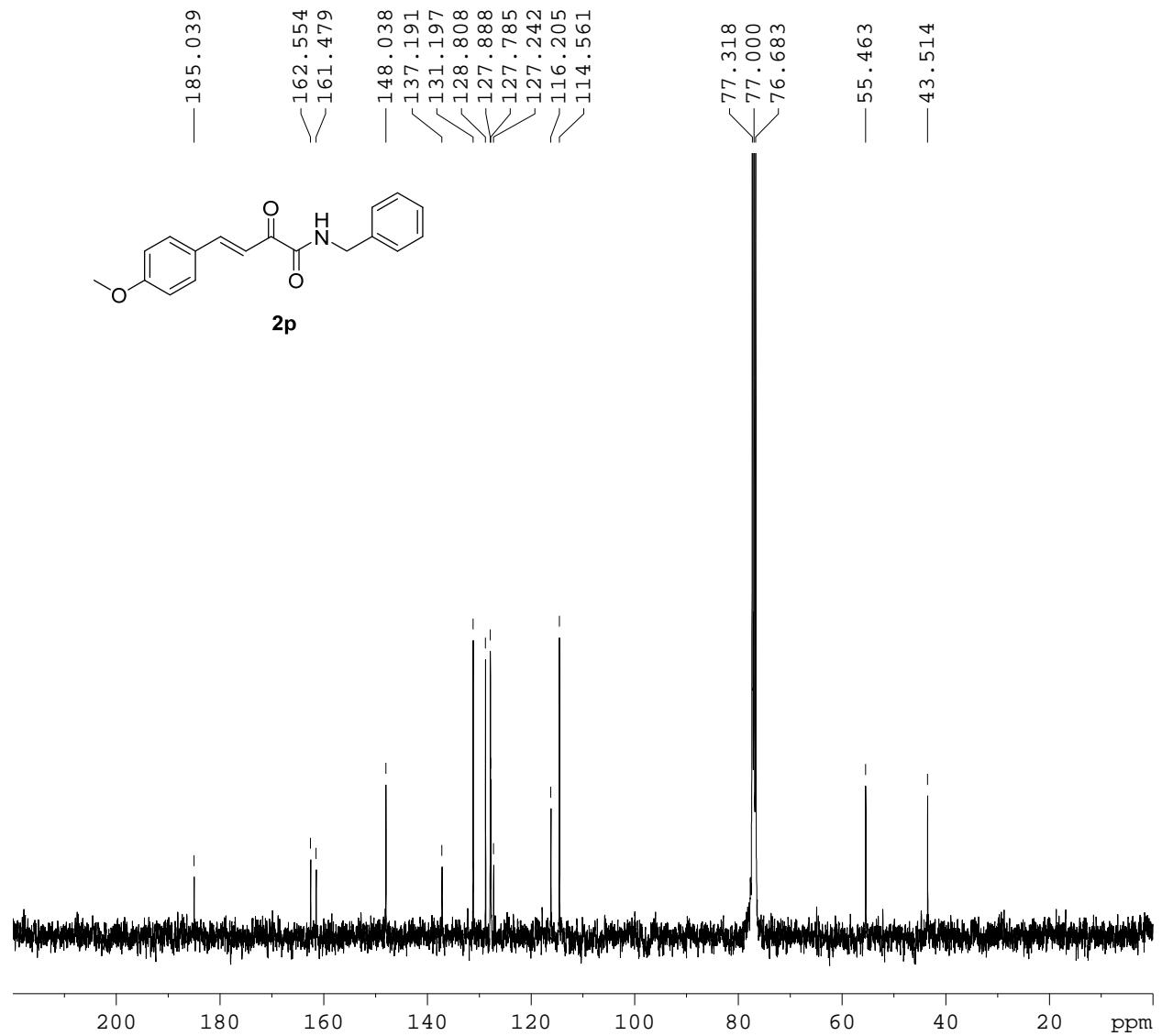
===== CHANNEL f1 =====

NUC1 13C
 P1 9.70 usec
 PL1 -0.50 dB
 SFO1 100.6288660 MHz

===== CHANNEL f2 =====

CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 -2.40 dB
 PL12 15.10 dB
 PL13 18.10 dB
 SFO2 400.1516010 MHz
 SI 32768
 SF 100.6178022 MHz
 WDW EM
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.00





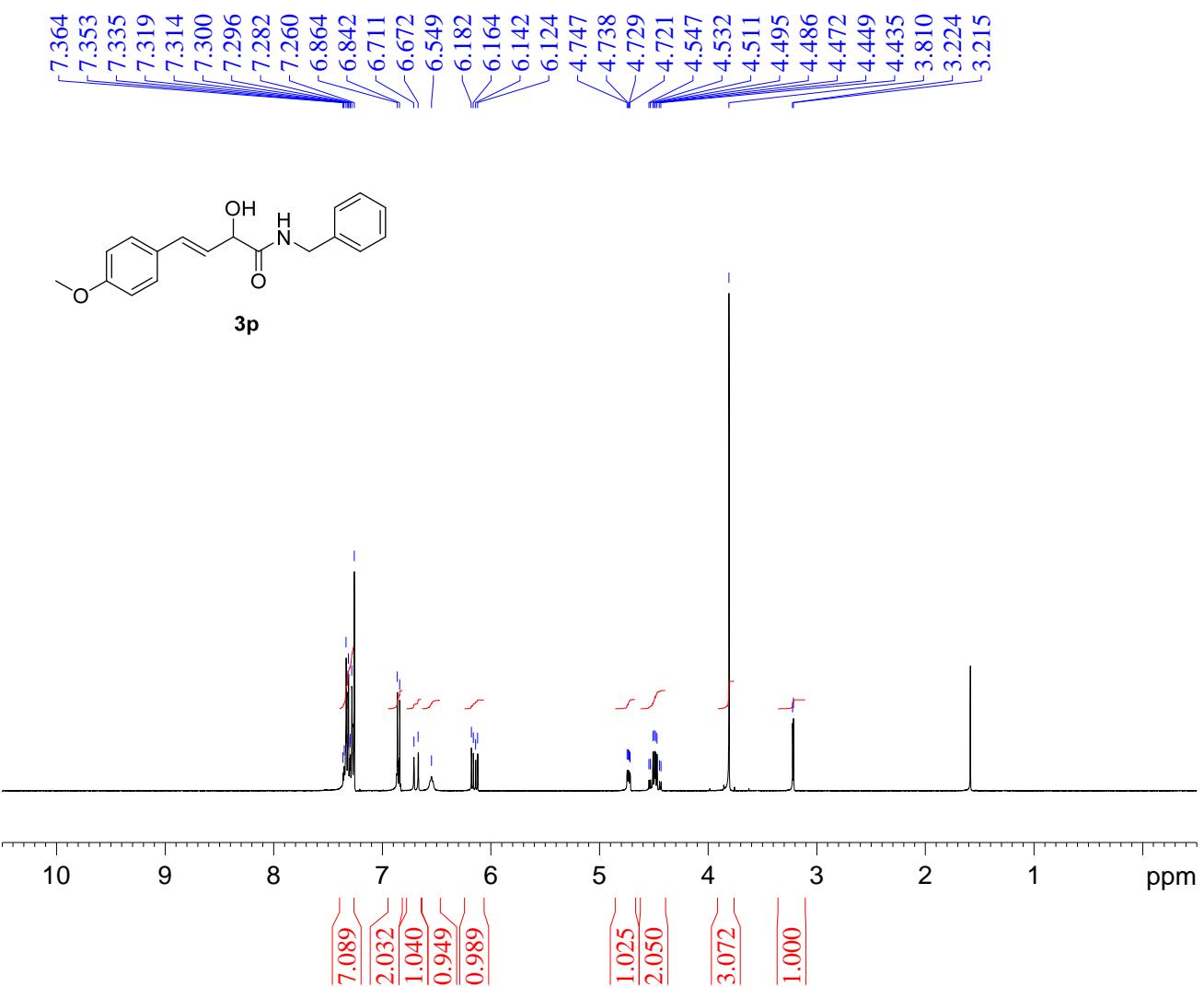
```

NAME          20151213
EXPNO         2
PROCNO        1
Date_         20151213
Time          19.05
INSTRUM      spect
PROBHD      5 mm DUL 13C-1
PULPROG     zgpg30
TD           65536
SOLVENT      CDC13
NS            786
DS             0
SWH          22727.273 Hz
FIDRES      0.346791 Hz
AQ           1.4418420 sec
RG            57
DW           22.000 usec
DE            6.00 usec
TE            300.0 K
D1           2.000000000 sec
d11          0.03000000 sec
DELTA        1.89999998 sec
TD0           1

===== CHANNEL f1 =====
NUC1          13C
P1            9.70 usec
PL1          -0.50 dB
SFO1        100.6288660 MHz

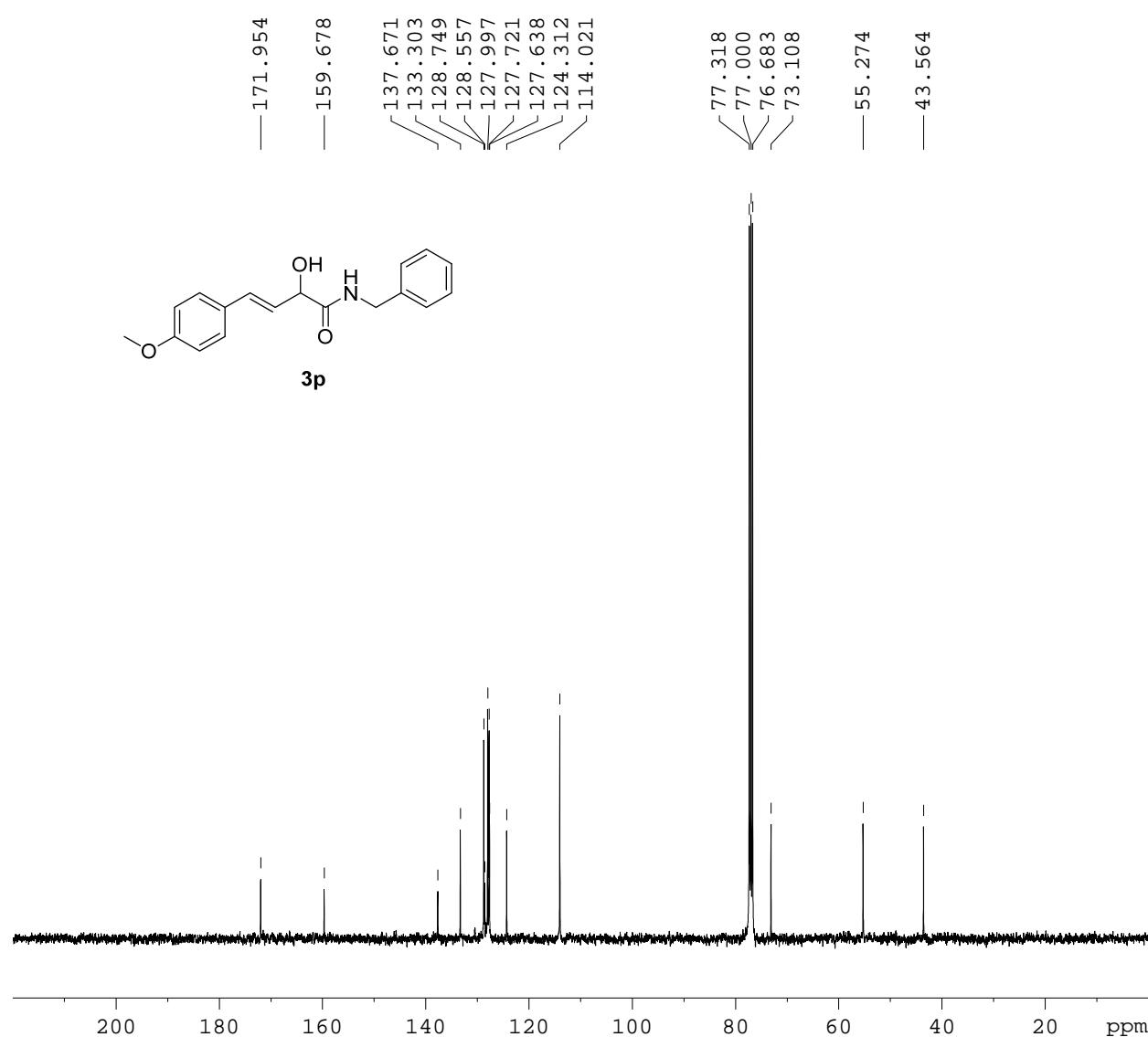
===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2          1H
PCPD2        90.00 usec
PL2          -2.40 dB
PL12         15.10 dB
PL13         18.10 dB
SFO2        400.1516010 MHz
SI            32768
SF           100.6177996 MHz
WDW           EM
SSB             0
LB            3.00 Hz
GB             0
PC           1.00

```



NAME 20151210
 EXPNO 1
 PROCNO 1
 Date_ 20151210
 Time 15.11
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 32768
 SOLVENT CDCl₃
 NS 38
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.195625 Hz
 AQ 2.5559540 sec
 RG 4
 DW 78.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 TD0 1

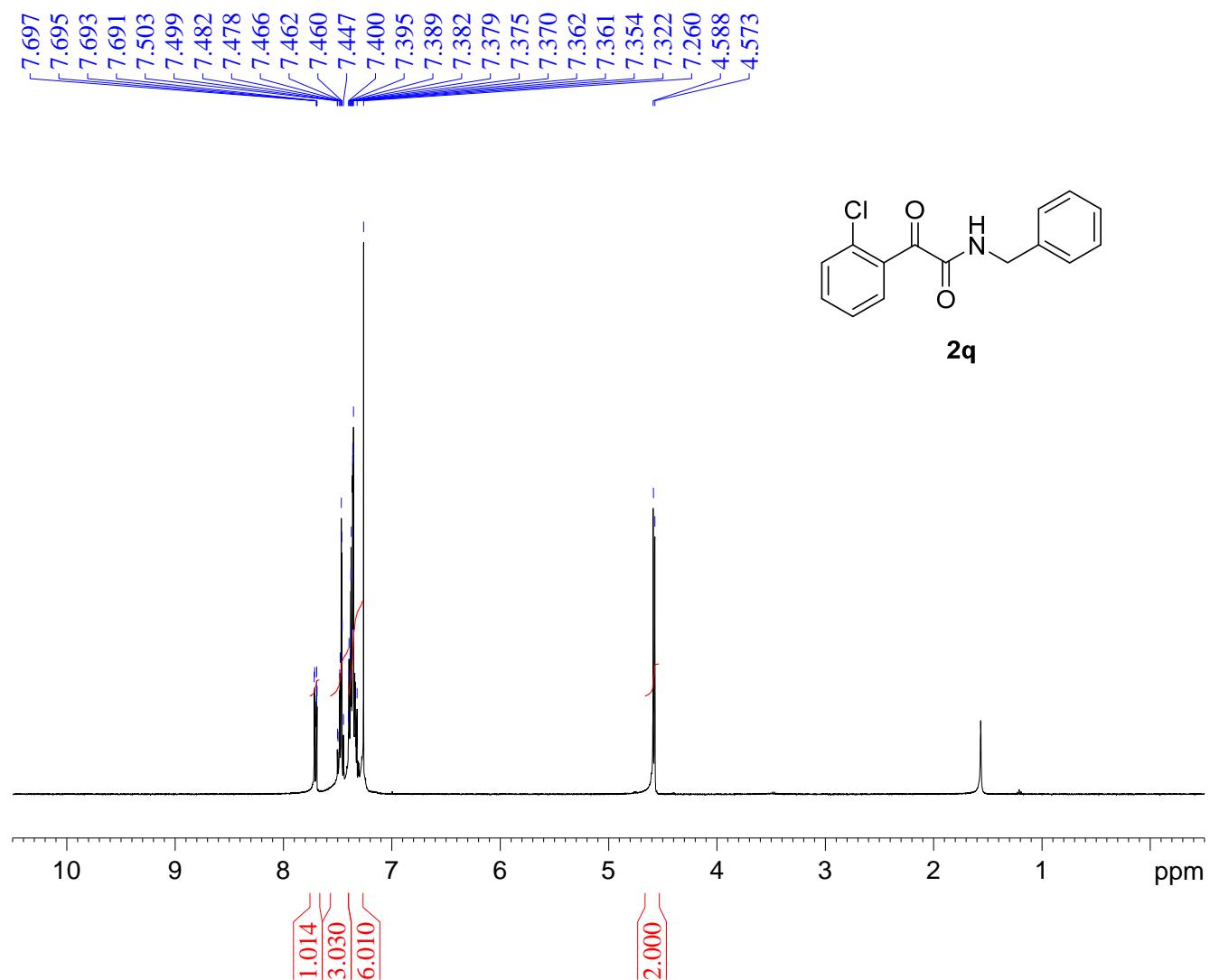
===== CHANNEL f1 ======
 NUC1 1H
 P1 10.00 usec
 PL1 -2.40 dB
 SFO1 400.1528010 MHz
 SI 16384
 SF 400.1500091 MHz
 WDW EM
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00

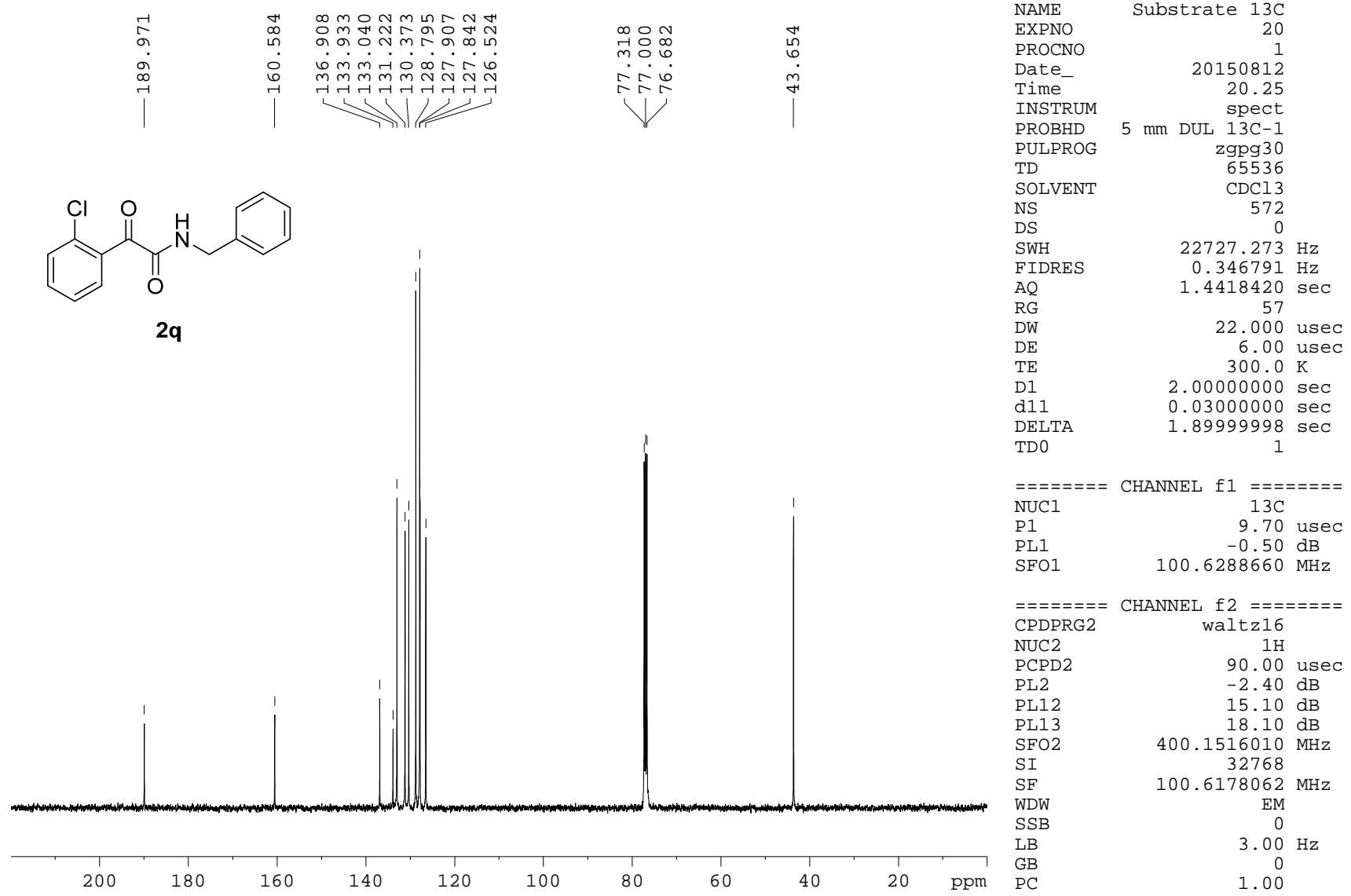


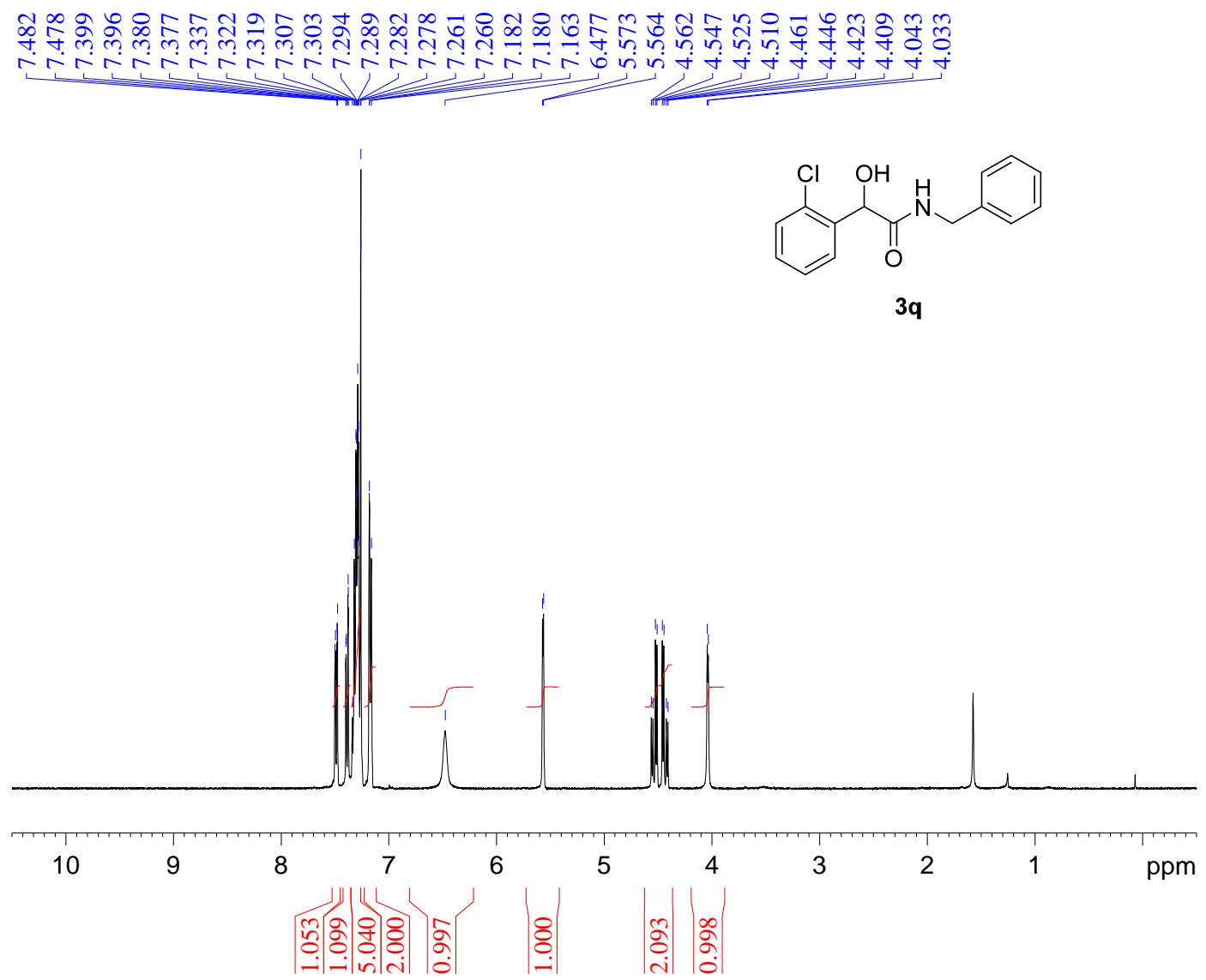
NAME 20151210
 EXPNO 2
 PROCNO 1
 Date_ 20151210
 Time 21.23
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 712
 DS 0
 SWH 22727.273 Hz
 FIDRES 0.346791 Hz
 AQ 1.4418420 sec
 RG 57
 DW 22.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 T0D 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.70 usec
 PL1 -0.50 dB
 SFO1 100.6288660 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 -2.40 dB
 PL12 15.10 dB
 PL13 18.10 dB
 SFO2 400.1516010 MHz
 SI 32768
 SF 100.6178015 MHz
 WDW EM
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.00

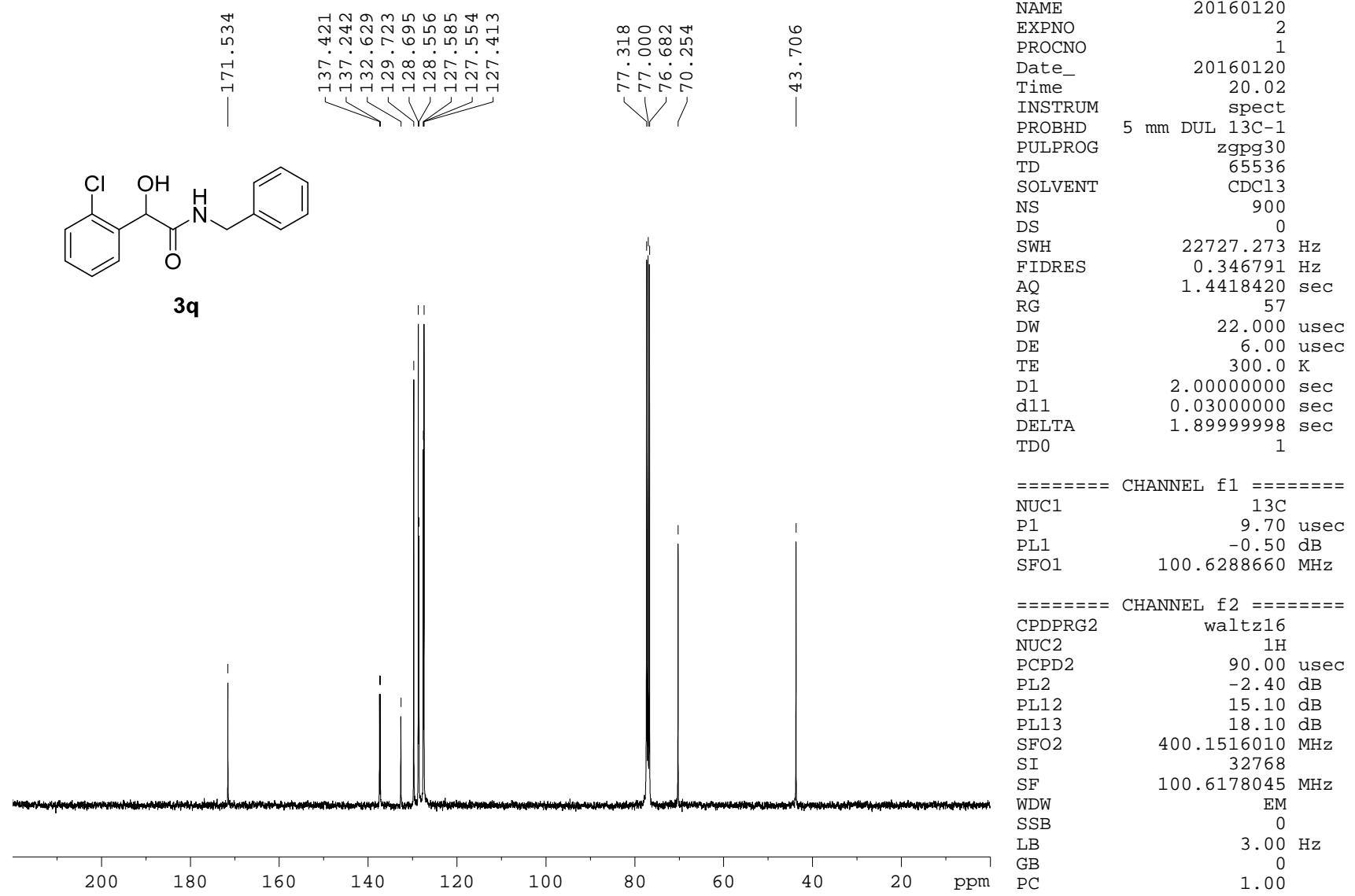


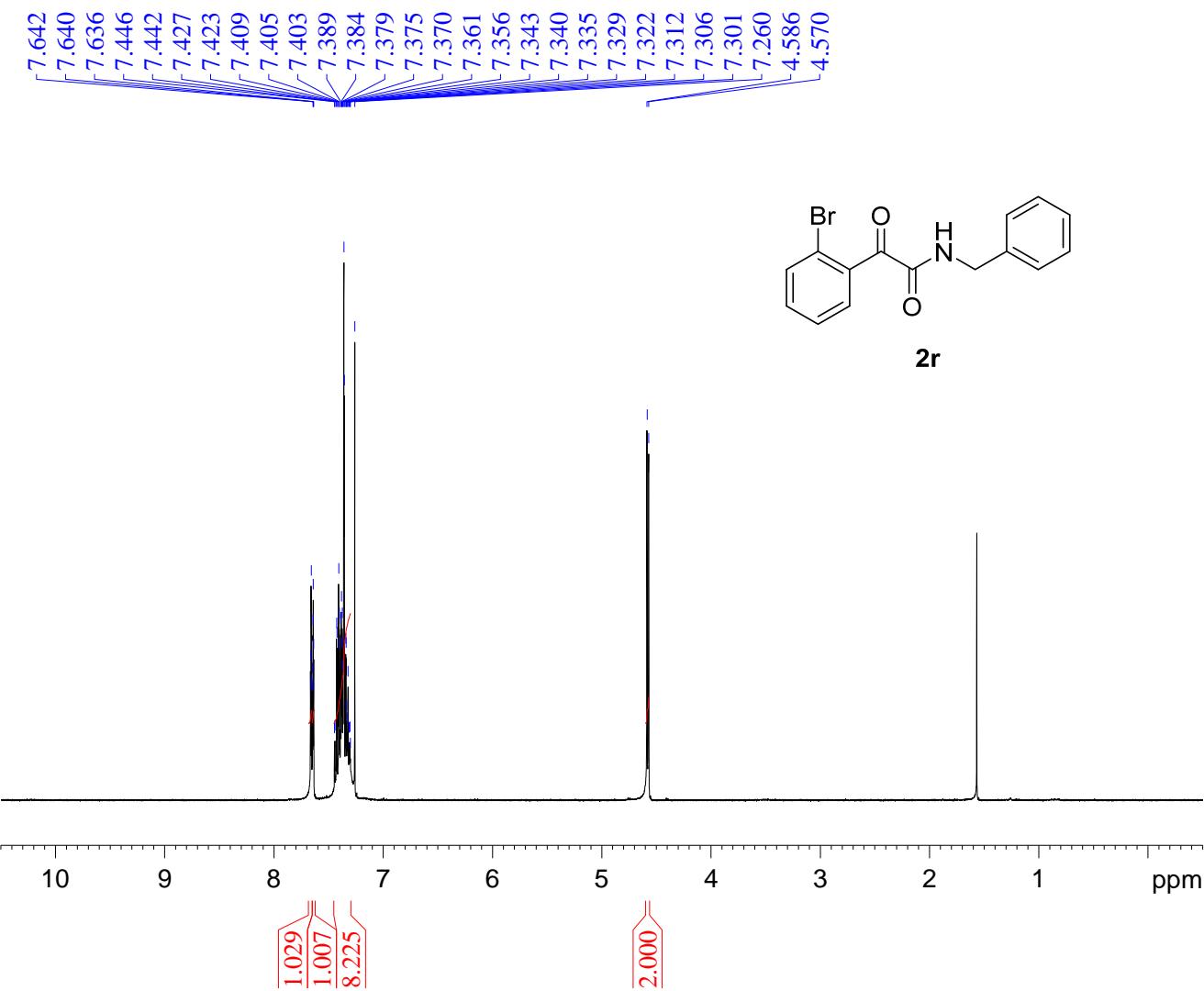




NAME 20160120
 EXPNO 1
 PROCNO 1
 Date_ 20160120
 Time 19.55
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 32768
 SOLVENT CDCl₃
 NS 48
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.195625 Hz
 AQ 2.5559540 sec
 RG 4
 DW 78.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 TD0 1

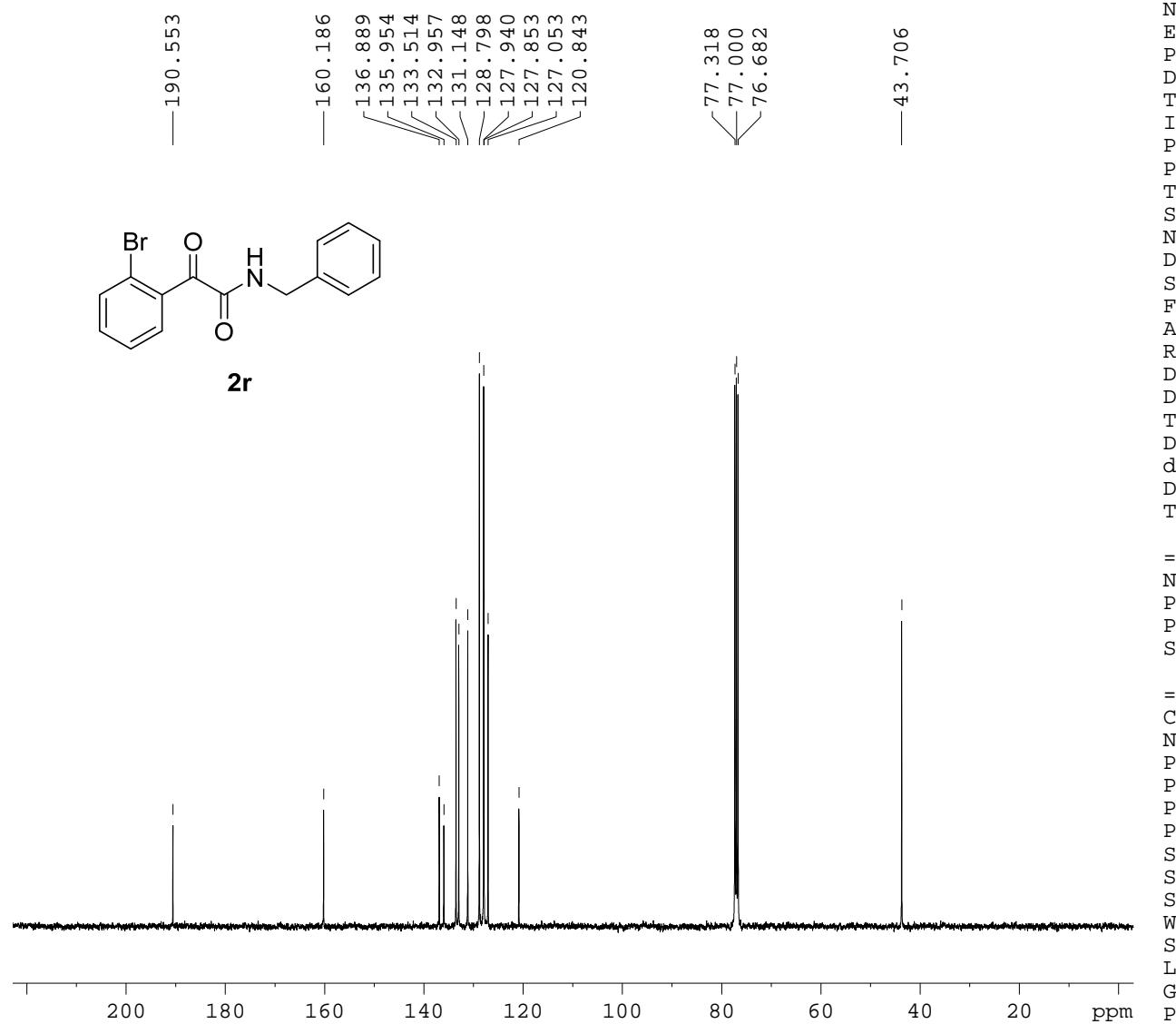
===== CHANNEL f1 ======
 NUC1 1H
 P1 10.00 usec
 PL1 -2.40 dB
 SFO1 400.1528010 MHz
 SI 16384
 SF 400.1500089 MHz
 WDW EM
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00

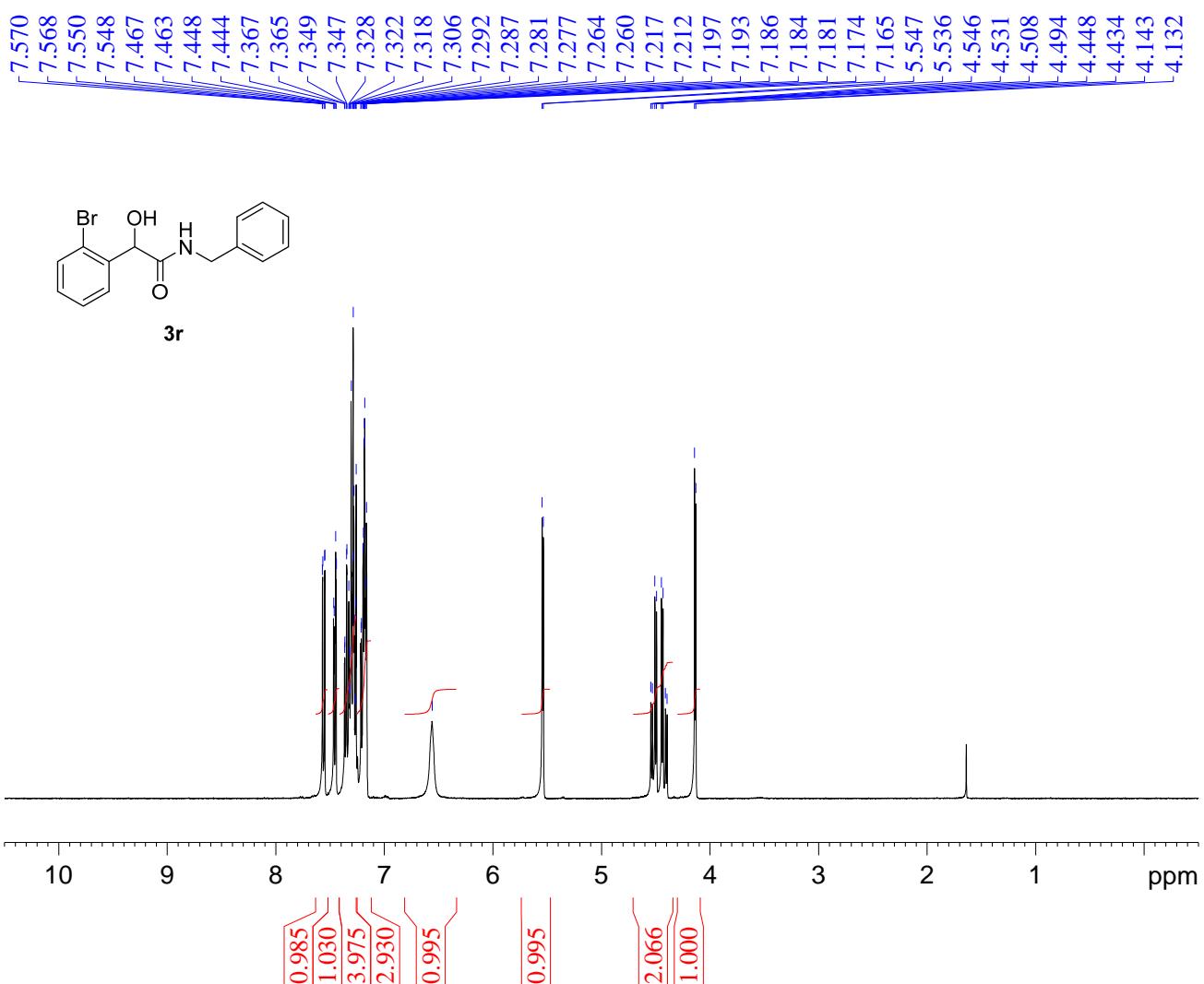




NAME 20151111
 EXPNO 1
 PROCNO 1
 Date_ 20151111
 Time 20.12
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 32768
 SOLVENT CDCl₃
 NS 16
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.195625 Hz
 AQ 2.5559540 sec
 RG 4
 DW 78.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 TD0 1

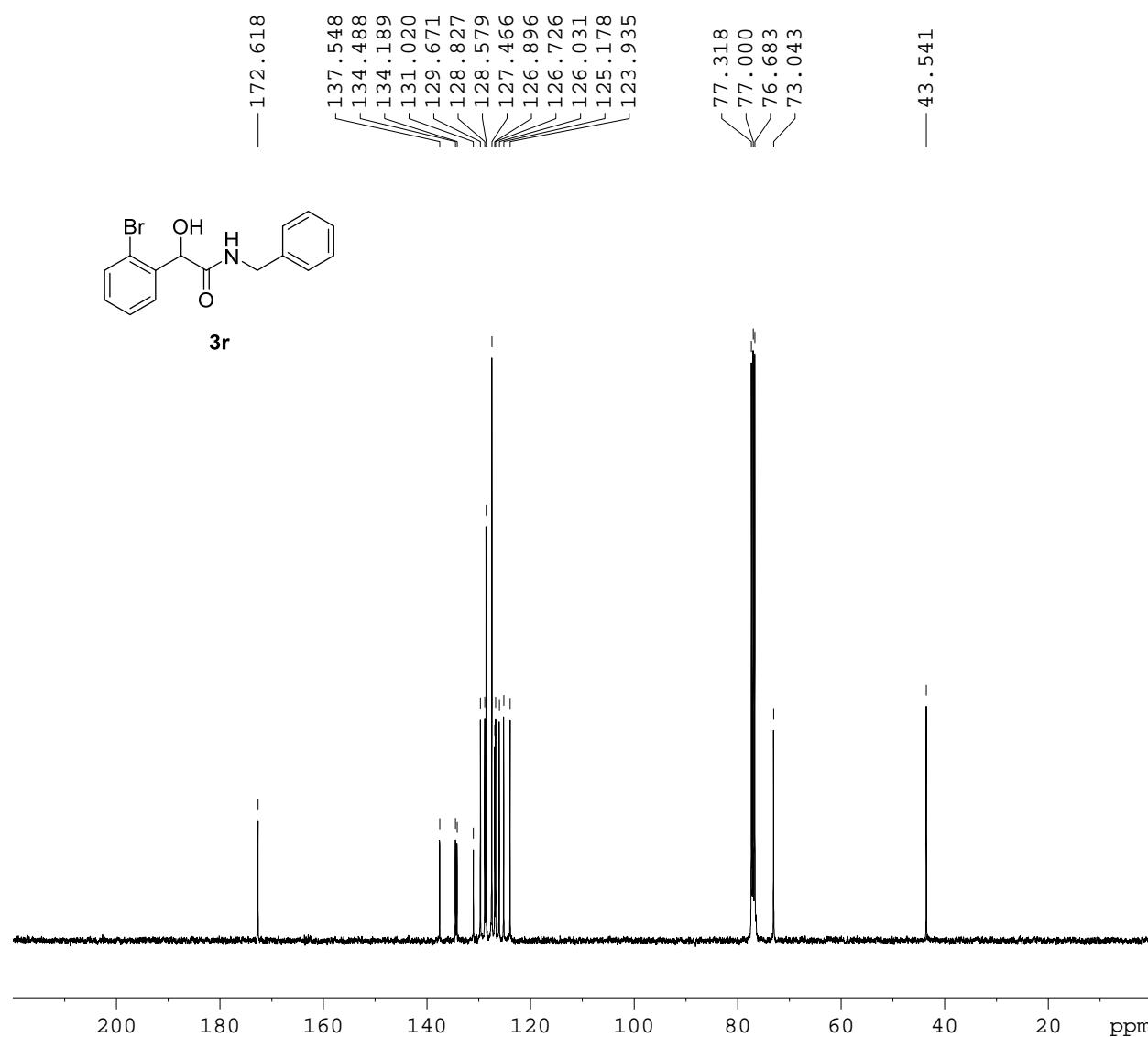
===== CHANNEL f1 ======
 NUC1 1H
 P1 10.00 usec
 PL1 -2.40 dB
 SFO1 400.1528010 MHz
 SI 16384
 SF 400.1500089 MHz
 WDW EM
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00



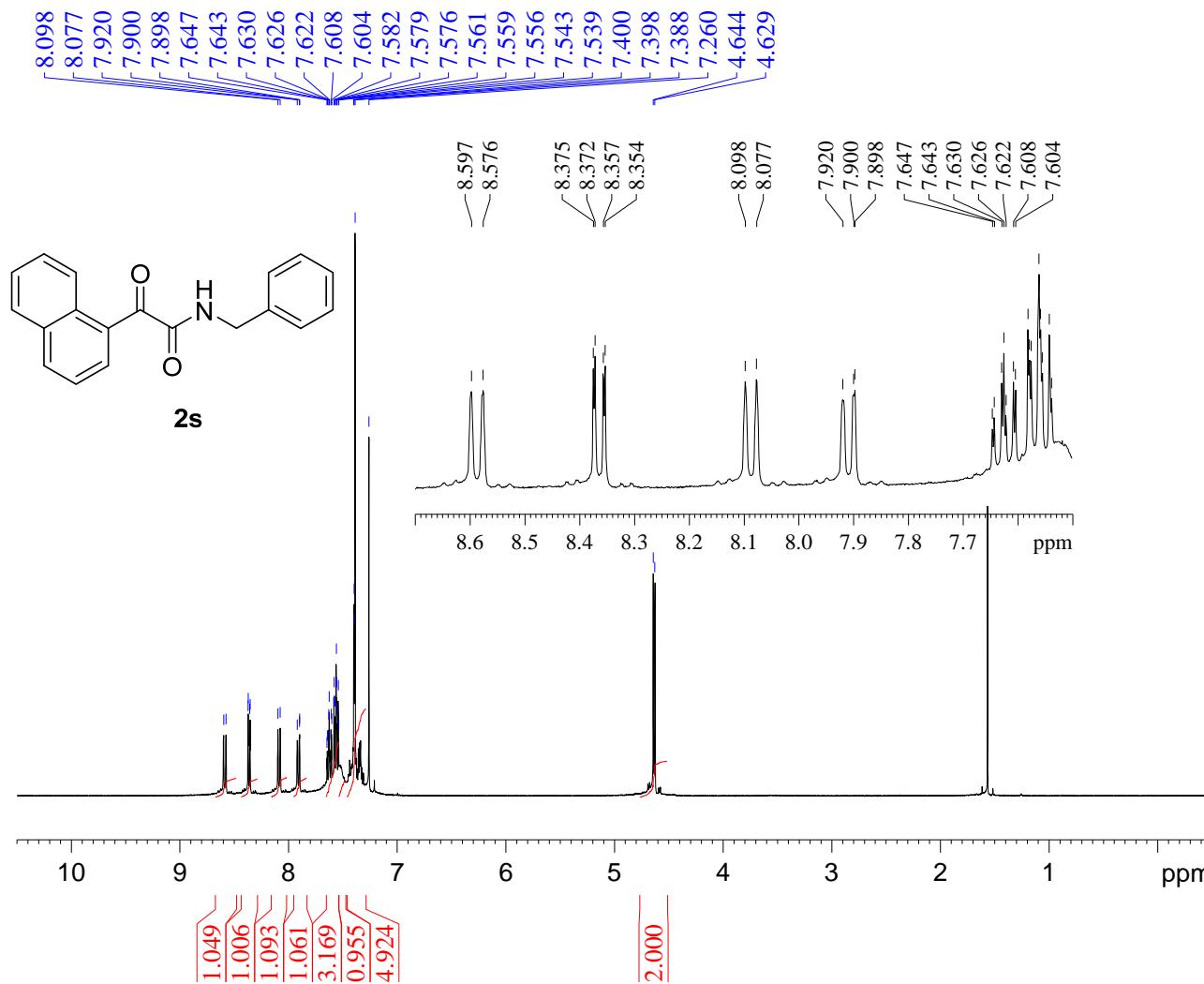


NAME Substrate
EXPNO 5
PROCNO 1
Date_ 20150426
Time 17.39
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 32768
SOLVENT CDCl₃
NS 44
DS 0
SWH 6410.256 Hz
FIDRES 0.195625 Hz
AQ 2.5559540 sec
RG 4
DW 78.000 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 10.00 usec
PL1 -2.40 dB
SFO1 400.1528010 MHz
SI 16384
SF 400.1500092 MHz
WDW EM
SSB 0
LB 0.00 Hz
GB 0
PC 1.00

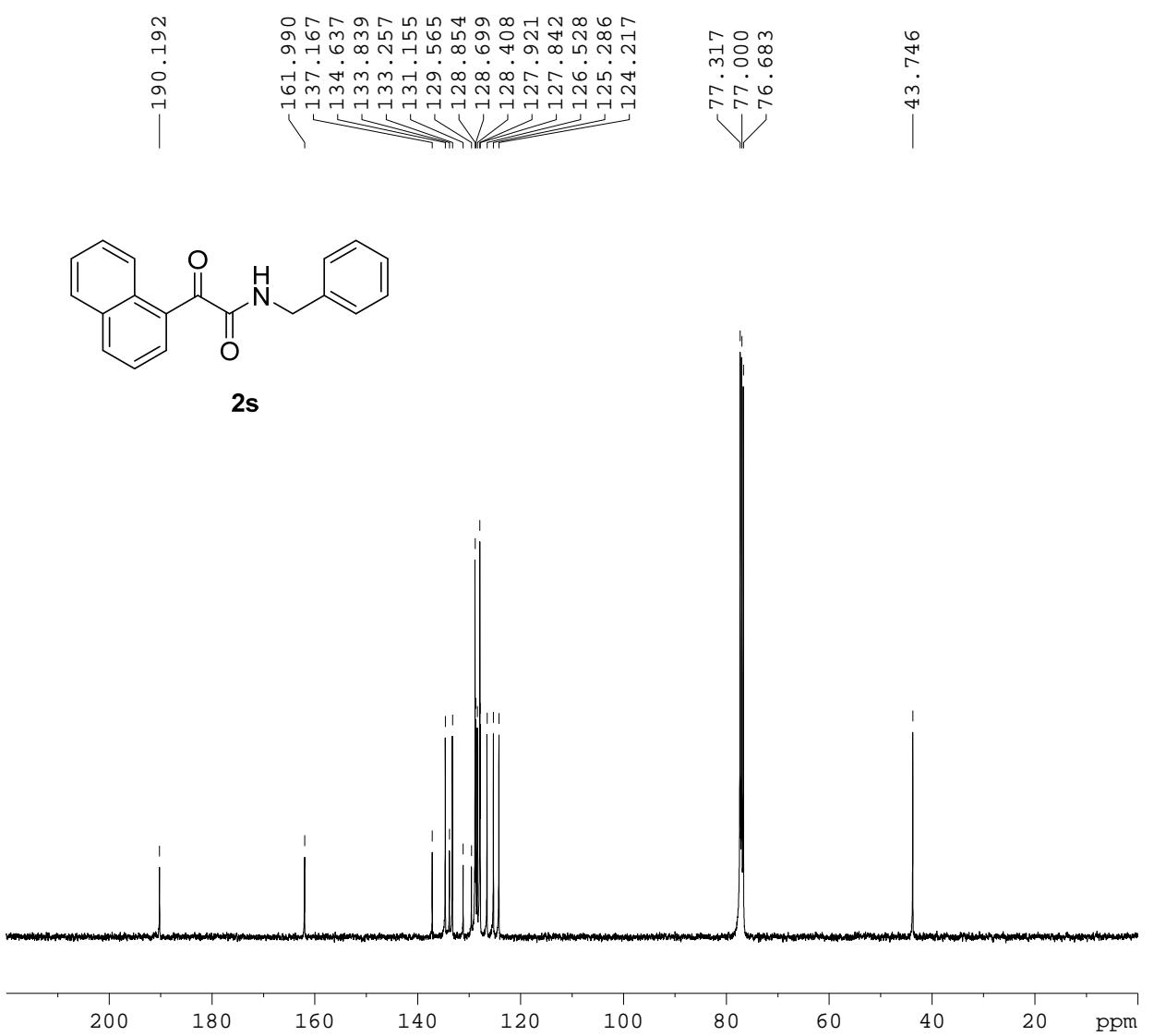


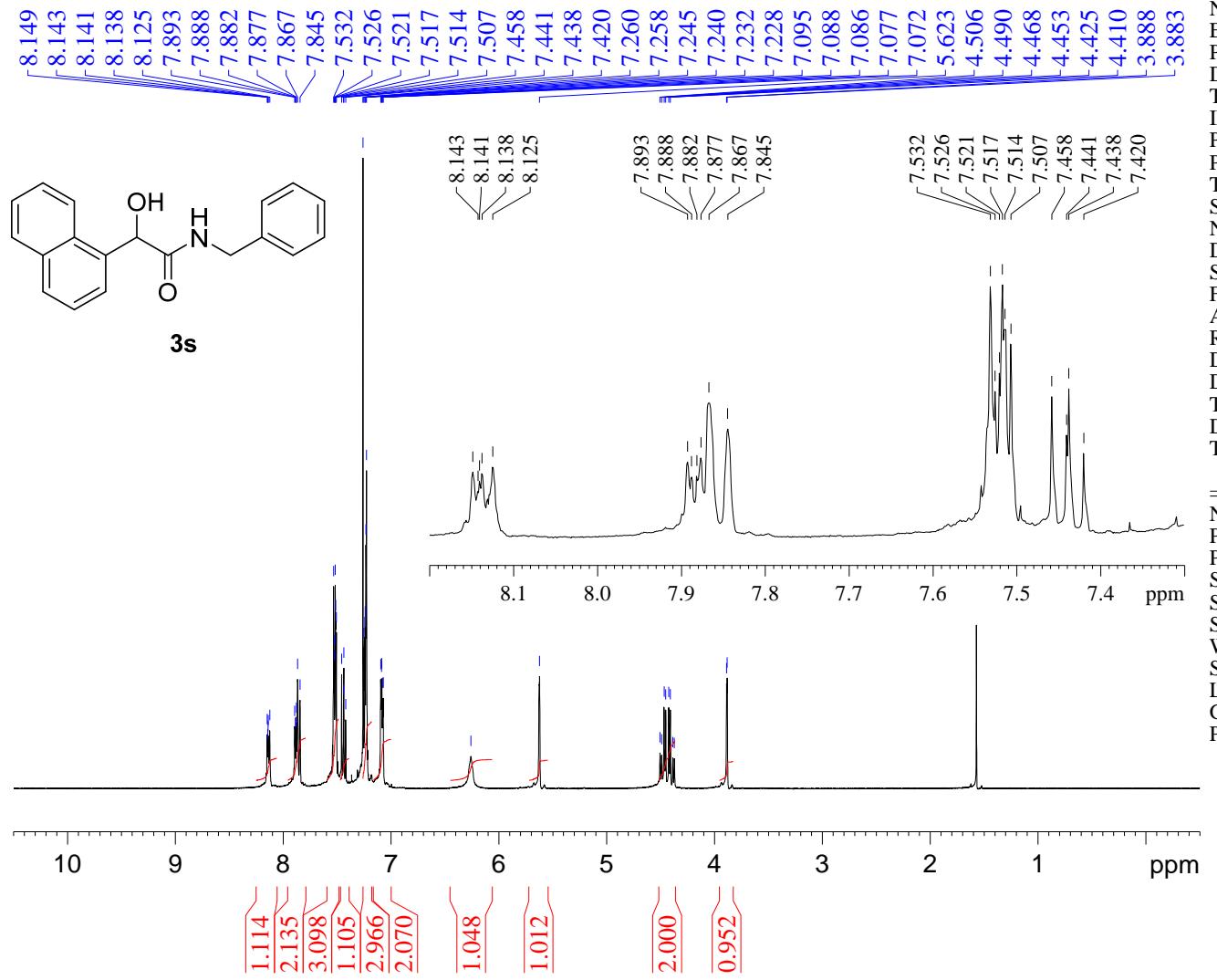
NAME	Substrate	¹³ C
EXPNO		7
PROCNO		1
Date_	20150712	
Time	15.35	
INSTRUM	spect	
PROBHD	5 mm DUL	¹³ C-1
PULPROG	zgpg30	
TD	65536	
SOLVENT	CDCl ₃	
NS	1029	
DS	0	
SWH	22727.273	Hz
FIDRES	0.346791	Hz
AQ	1.4418420	sec
RG	57	
DW	22.000	usec
DE	6.00	usec
TE	300.0	K
D1	2.00000000	sec
d11	0.03000000	sec
DELTA	1.89999998	sec
TDO	1	
 ===== CHANNEL f1 =====		
NUC1	¹³ C	
P1	9.70	usec
PL1	-0.50	dB
SFO1	100.6288660	MHz
 ===== CHANNEL f2 =====		
CPDPRG2	waltz16	
NUC2	¹ H	
PCPD2	90.00	usec
PL2	-2.40	dB
PL12	15.10	dB
PL13	18.10	dB
SFO2	400.1516010	MHz
SI	32768	
SF	100.6178041	MHz
WDW	EM	
SSB	0	
LB	3.00	Hz
GB	0	
PC	1.00	



NAME	Substrate	1H
EXPNO	6	
PROCNO	1	
Date_	20150712	
Time	15.19	
INSTRUM	spect	
PROBHDI	5 mm DUL	13C-1
PULPROG	zg30	
TD	32768	
SOLVENT	CDCl ₃	
NS	52	
DS	0	
SWH	6410.256 Hz	
FIDRES	0.195625 Hz	
AQ	2.5559540 sec	
RG	4	
DW	78.0000 usec	
DE	6.00 usec	
TE	300.0 K	
D1	2.0000000 sec	
TD0	1	

```
===== CHANNEL f1 =====
NUC1          1H
P1           10.00 usec
PL1          -2.40 dB
SFO1        400.1528010 MHz
SI           16384
SF        400.1500088 MHz
WDW          EM
SSB            0
LB           0.00 Hz
GB            0
PC           1.00
```





NAME	Substrate	1H
EXPNO	7	
PROCNO	1	
Date_	20150712	
Time	15.27	
INSTRUM	spect	
PROBHD	5 mm DUL	13C-
PULPROG	zg30	
TD	32768	
SOLVENT	CDCl3	
NS	65	
DS	0	
SWH	6410.256 Hz	
FIDRES	0.195625 Hz	
AQ	2.5559540 sec	
RG	4	
DW	78.000 usec	
DE	6.00 usec	
TE	300.0 K	
D1	2.0000000 sec	
TD0	1	

===== CHANNEL f1 =====
NUC1 1H
P1 10.00 usec
PL1 -2.40 dB
SFO1 400.1528010 MHz
SI 16384
SF 400.1500092 MHz
WDW EM
SSB 0
LB 0.00 Hz
GB 0
PC 1.00

