Supporting Information

Reductive Coupling of Imines with Redox-Active Esters by Visible Light Photoorganocatalysis

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1. General methods

Unless otherwise noted, all reactions of substrates preparation were conducted in flame-dried glassware under argon atmosphere using anhydrous degassed solvent. Commercially available reagents were used without further purification. Solvents for chromatography were technical grade and distilled prior to use. Dry dichloromethane and chloroform used in reactions were obtained by distilling over calcium hydride and were stored over activated molecular sieves (4 Å). Diethyl ether, tetrahydrofuran, dimethylformamide and toluene used in reactions were obtained by distilling over sodium-benzophenone ketyl. Analytical thin-layer chromatography (TLC) was performed on Macherey-Nagel silica gel 60 aluminium plates with F-254 indicator, visualized by UV irradiation. Column chromatography was performed using MN silica gel (particle size 0.040-0.063 mm). ¹H-NMR and ¹³C-NMR spectra were recorded on a vnmrs-400 or vnmrs-600 spectrometer in CDCl₃ with residual proton signal of the deuterated solvents as the internal reference ($\delta H = 7.26$ ppm and $\delta C = 77$ ppm for CDCl₃). Data are reported in the following order: chemical shift (δ) in ppm; multiplicities are indicated s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), dd (doublet of doublet), tt (triplet of triplet), dt (doublet of triplet), td (triplet of doublet); coupling constants (J) are in Hertz (Hz). IR spectra were recorded on a Jasco FT/IR-420 spectrometer and are reported in terms of frequency of absorption (cm⁻¹). Mass spectra were acquired on a Finnigan SSQ7000 (EI/CI) spectrometer and high resolution mass spectra on a Finnigan MAT 95 (EI/CI) or on a ThermoFisher Scientific LTQOrbitrap XL (ESI) spectrometer. Melting points were recorded on a Büchi 560 Melting Point Apparatus.

2. Substrate preparation

2.1 General procedure for the synthesis of imines^[1]

$$Ar^{1} H + Ar^{2}-NH_{2} \xrightarrow{\text{molecular sieves}} Ar^{1} N_{Ar^{2}}$$

In an oven-dried flask equipped with a magnetic stir bar and molecular sieves (4 Å, 2 g) was added a solution of the aldehyde (20 mmol) in dry DCM (50 mL). Then the amine (20 mmol) was added slowly and the stirring continued overnight. After filtration over Celite, the solvent was evaporated under reduced pressure and the product was purified by crystallization from a mixture of Et_2O /Pentane, or by distilling off the excess of aniline via Kugelrohr distillation.

2.2 General procedure for the synthesis of *N*-hydroxyphthalimide esters (NHP esters)^[2]



A round bottom flask containing a teflon-coated magnetic stir bar was charged with the corresponding acid (5.00 mmol, 1.00 equiv.) and dry EtOAc (25 mL). After sequential addition of N,N'-dicyclohexylcarbodiimide (7.50 mmol, 1.5 equiv.), DMAP (30.54 mg, 0.25 mmol, 0.05 equiv.), and N-hydroxyphthalimide (1.35 g, 8.30 mmol, 1.66 equiv.), the reaction mixture was allowed to stir overnight at room temperature. After this time, the heterogeneous mixture was filtered and the filtrate was concentrated under reduced pressure. Purification on silica gel (hexane: EA=6:1) afforded the N-hydroxyphthalimide esters as colorless solids.

3. Full optimization studies

3.1 Photoredox catalysts and light source screening



Entry	Photocatalyst	NMR Yield $(\%)^b$
1^c	[Ir(ppy) ₂ (dtbbpy)]PF ₆	69
2	Fluorescein	48
3	Rose Bengal (RB)	53
4	Eosin Y	50
5	Methylen Blue	NR
6	Alizarin Red S	NR
7	Rhodamine B	trace
8^d	Rose Bengal	42
9^e	Rose Bengal	57

Table S1 Catalyst screening^{*a*}

^{*a*}The reactions of **1a** (0.2 mmol) with **2a** (0.1 mmol) were carried out in the presence of photocatalyst (5 mol%), iPr_2NEt (5 equiv.) and HBF (5 equiv., 50% in water) in DMF (degased, 1 ml) under irradiation with blue LEDs (11 W, 450 nm) for 20 h at rt; ^{*b*}Yields were determined by ¹H-NMR analysis of the crude mixture relative to CH₂Br₂ as internal standard; ^{*c*}Photocatalyst (1 mol%); ^{*d*}High power blue LEDs irradiation; ^{*e*}Green LED irradiation.

3.2 Reducing agent screening



Entry	Reducing agent	NMR Yield $(\%)^b$
1	<i>i</i> Pr ₂ NEt	57
2	<i>n</i> Bu ₃ N	52
3	Et ₃ N	43
4	HE	32
5	iPr ₂ NEt (2.5 equiv) + HE (2.5 equiv)	40

Table S2 Reducing agent screening^a

^{*a*}The reactions of **1a** (0.2 mmol) with **2a** (0.1 mmol) were carried out in the presence of photocatalyst (5 mol%), Reducing agent (5 equiv.) and HBF₄ (5 equiv., 50% in water) in DMF (degassed, 1 ml) under irradiation with Green LED irradiation for 20 h at rt; ^{*b*}Yields were determined by ¹H-NMR analysis of the crude mixture relative to CH₂Br₂ as internal standard.

3.3 Hydrogen source and additive screening



Entry	H-source	Additive	NMR Yield $(\%)^b$
1	HBF ₄ (5equiv., 50 % in ether)	-	trace
2	HCOOH (5 equiv.)	-	trace
3	CH ₃ COOH (5 equiv.)	-	trace
4	H ₂ O (10 equiv.)	-	40
5	H ₂ O (10 equiv.)	LiF (20 mol%)	48
6	H ₂ O (10 equiv.)	LiOAc (20 mol%)	54
7	H ₂ O (10 equiv.)	Li ₂ CO ₃ (20 mol%)	58

Table S2 Hydrogen source and additive screening^{*a*}

^{*a*}The reactions of **1a** (0.2 mmol) with **2a** (0.1 mmol) were carried out in the presence of rose bengal (RB, 5 mol%), iPr_2NEt (5 equiv.), H-source (5 or 10 equiv.) and additive (20 mol%) in DMF (degassed, 1 ml) by green LED irradiation (11 W, 450 nm) for 20 h at rt; ^{*b*}Yields were determined by ¹H-NMR analysis of the crude mixture relative to CH₂Br₂ as internal standard.

3.4 Solvent screening



Entry	Solvent	NMR Yield (%) ^b
1	DMF	57
2	THF	41
3	MeOH	trace
4	DMSO	24
5	MeCN	68
6	Tol	62
7	DCM	50
8	EA	72
9	CHCl ₃	53
10	Aceton	70
11	Et ₂ O	47

Table S3 Solvent screening^a

^{*a*}The reactions of **1a** (0.2 mmol) with **2a** (0.1 mmol) were carried out in the presence of rose bengal (5 mol%), *i*Pr₂NEt (5 equiv.), H₂O (10 equiv.) and Li₂CO₃ (20 mol%) in different solvents (degassed, 1 ml) under irradiation with green LEDs (11 W, 450 nm) for 20 h at rt; ^{*b*}Yields were determined by ¹H-NMR analysis of the crude mixture relative to CH₂Br₂ as internal standard.

4. Preparation and characterization of amines



A Schlenk-tube was charged with substrates **1a** (0.4 mmol, 2 equiv.), **2a** (0.2 mmol 1 equiv.), rose bengal (5 mol%), and Li₂CO₃ (20 mol%). It was capped with a rubber septum, evacuated and backfilled with argon. Then, degassed EtOAc (2 mL) was added via syringe, at the same time the *i*Pr₂NEt (5 equiv.) and H₂O (10 equiv.) were also added. The vial was placed in a 100 mL beaker wrapped with green LEDs inside and the reaction mixture was stirred at rt for 20 h. After checking the course of the reaction by TLC, the solvent was evaporated under reduced pressure and the product was purified by column chromatography using Pentane/EA (100:1-6:1) as eluent.

N-(1,4-diphenylbutyl)aniline (3a)^[3]

^{HN, Ph} Yellow oil was obtained after column chromatography (100% pentane to Ph 100:1 pentane/EtOAc). Yield = 62% (37.2 mg, 0.12 mmol). ¹H NMR (600 MHz, CDCl₃) δ 7.36 – 7.27 (m, 6H), 7.25 – 7.18 (m, 2H), 7.16 (d, *J* = 7.2 Hz, 2H), 7.09 (dd, *J* = 8.4, 7.4 Hz, 2H), 6.64 (t, *J* = 7.3 Hz, 1H), 6.51 (d, *J* = 7.8 Hz, 2H), 4.33 (t, *J* = 6.7 Hz, 1H), 4.05 (bs, 1H), 2.71 – 2.58 (m, 2H), 1.91 – 1.74 (m, 3H), 1.72 – 1.64 (m, 1H). ¹³C NMR (151 MHz, CDCl₃) δ 147.3, 143.9, 141.9, 129.0, 128.5, 128.3, 128.3, 126.8, 126.3, 125.8, 117.1, 113.1, 58.0, 38.2, 35.6, 27.9. MS (EI): *m/z* (%) = 302.2 ([M+H]⁺, 16), 301.2 ([M]⁺, 38), 182.1 (100).

IR (ATR): *v* =3383, 2938, 2285, 1704, 1477, 1464, 1350, 1240, 1139, 727.

N-(1,3-diphenylpropyl)aniline (3b)^[4]

HN, Ph Yellow oil was obtained after column chromatography (100% pentane to 100:1 Ph Ph pentane/EtOAc). Yield = 69% (39.6 mg, 0.14 mmol). ¹H NMR (400 MHz, CDCl₃) δ 7.38 – 7.27 (m, 6H), 7.27 – 7.16 (m, 4H), 7.08 (dd, *J* = 8.6, 7.3 Hz, 2H), 6.65 (t, *J* = 7.3 Hz, 1H), 6.50 (d, *J* = 8.6 Hz, 2H), 4.36 (t, *J* = 6.7 Hz, 1H), 4.08 (bs, 1H), 2.83 – 2.64 (m, 2H), 2.23 – 2.08 (m, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 147.2, 143.7, 141.4, 129.0, 128.5, 128.4, 128.4, 127.0, 126.4, 125.9, 117.2, 113.2, 57.6, 40.1, 32.6. MS (EI): *m/z* (%) = 288.2 ([M+H]⁺, 15), 287.2 ([M]⁺, 32), 182.1 (100).

IR (ATR): $\tilde{v} = 3410, 3030, 2915, 2328, 1737, 1599, 1490, 1304, 718.$

N-(1-phenylheptyl)aniline (3c)

Yellow oil was obtained after column chromatography (100% pentane to Ph 100:1 pentane/EtOAc). Yield = 61% (32.5 mg, 0.12 mmol). ¹H NMR (400 MHz, CDCl₃) δ 7.42 – 7.27 (m, 4H), 7.26 – 7.19 (m, 1H), 7.07 (dd, *J* = 8.6, 7.3 Hz, 2H), 6.64 (t, *J* = 7.3 Hz, 1H), 6.52 (d, *J* = 8.1 Hz, 2H), 4.30 (t, *J* = 6.8 Hz, 1H), 4.07 (bs, 1H), 1.88 – 1.72 (m, 2H), 1.48 – 1.39 (m, 1H), 1.34 – 1.24 (m, 6H), 0.88 (t, *J* = 6.8 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 147.4, 144.3, 129.0, 128.4, 126.7, 126.3, 117.0, 113.2, 58.25, 39.0, 31.7, 29.1, 26.3, 22.5, 14.0. MS (EI): *m/z* (%) = 268.2 ([M+H]⁺, 13), 267.2 ([M]⁺, 33), 182.1 (100). IR (ATR): \tilde{v} =3411, 2925, 2339, 2095, 1733, 1599, 1494, 1308, 723.

HRMS (EI+, m/z): calculated for $C_{19}H_{26}N$ [M+H]+: 268.20598; found: 268.20590.

N-(1-phenylpropyl)aniline (3d)^[5]

^{HN}, ^{Ph} Yellow oil was obtained after column chromatography (100% pentane to 100:1 ^{Ph} pentane/EtOAc). Yield = 73% (30.8 mg, 0.15 mmol). ¹H NMR (600 MHz, CDCl₃) δ 7.38 – 7.29 (m, 4H), 7.24 (t, *J* = 7.0 Hz, 1H), 7.09 (dd, *J* = 8.6, 7.4 Hz, 2H), 6.65 (t, *J* = 7.4, 1H), 6.54 (d, *J* = 8.1, 2H), 4.25 (t, *J* = 6.7 Hz, 1H), 4.09 (bs, 1H), 1.92 – 1.80 (m, 2H), 0.98 (t, *J* = 7.4 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 147.4, 143.8, 129.0, 128.4, 126.8, 126.4, 117.0, 113.1, 59.68, 31.6, 10.8.

MS (EI): m/z (%) = 212.2 ([M+H]⁺, 16), 211.2 ([M]⁺, 37), 182.1 (100).

IR (ATR): \tilde{v} =3416, 2947, 2327, 2108, 1733, 1598, 1490, 1305, 707.

N-(4-methyl-1-phenyloctyl)aniline (3e)



N-(1-phenyltetradecyl)aniline (3f)



(37.9 mg, 0.10 mmol.

¹H NMR (400 MHz, CDCl₃) δ 7.37 – 7.28 (m, 4H), 7.25 – 7.19 (m, 1H), 7.09 (dd, J = 8.4, 7.2 Hz, 2H), 6.63 (t, J = 7.3 Hz, 1H), 6.52 (d, J = 8.1, 2H), 4.30 (t, J = 6.8 Hz, 1H), 4.06 (bs, 1H), 1.86 – 1.72 (m, 2H), 1.48 – 1.37 (m, 1H), 1.34 – 1.22 (m, 21H), 0.90 (t, J = 6.8 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 147.4, 144.3, 129.0, 128.4, 126.7, 126.3, 117.0, 113.1, 58.2, 39.0, 31.9, 29.6, 29.6, 29.6, 29.5, 29.5, 29.4, 29.3, 26.3, 22.6, 14.1.

MS (EI): m/z (%) = 366.3 ([M+H]⁺, 31), 365.3 ([M]⁺, 46), 182.1 (100).

IR (ATR): *v* = 3415, 2916, 2323, 1738, 1599, 1483, 1299, 719.

HRMS (EI+, m/z): calculated for C₂₆H₄₀N [M+H]+: 366.31553; found: 366.31577.

N-(cyclohexyl(phenyl)methyl)aniline (3g)^[6]

HN^{Ph} Yellow oil was obtained after column chromatography (100% pentane to 100:1 pentane/EtOAc). Yield = 57% (30.2 mg, 0.11 mmol).

¹H NMR (400 MHz, CDCl₃) δ 7.34 – 7.27 (m, 4H), 7.25 – 7.19 (m, 1H), 7.07 (dd, J = 8.6, 7.3 Hz, 2H), 6.61 (t, J = 7.2 Hz, 1H), 6.50 (d, J = 7.6 Hz, 2H), 4.15 (bs, 1H), 4.13 (d, J = 6.3 Hz, 1H), 1.94 – 1.86 (m, 1H), 1.81 – 1.62 (m, 5H), 1.58 – 1.51 (m, 1H), 1.24 – 1.03 (m, 5H).

¹³C NMR (101 MHz, CDCl₃) δ 147.7, 142.6, 129.1, 128.1, 127.1, 126.6, 116.8, 113.1, 63.3, 44.8, 30.2, 29.4, 26.4, 26.3, 26.3.

MS (EI): m/z (%) = 266.2 ([M+H]⁺, 21), 265.2 ([M]⁺, 20), 182.1 (100).

IR (ATR): *v* = 3415, 2902, 2344, 2090, 1748, 1589, 1486, 1295, 719.

N-(2,2-dimethyl-1-phenylpropyl)aniline (3h)^[7]

^{HN} Ph Yellow oil was obtained after column chromatography (100% pentane to 100:1 pentane/EtOAc). Yield = 61% (29.1 mg, 0.12 mmol).

¹H NMR (600 MHz, CDCl₃) δ 7.30 (dt, *J* = 15.2, 4.6 Hz, 4H), 7.24 – 7.19 (m, 1H), 7.05 (dd, *J* = 8.5, 7.2 Hz, 2H), 6.59 (t, *J* = 7.3 Hz, 1H), 6.49 (d, *J* = 7.7 Hz, 2H), 4.27 (bs, 1H), 4.05 (s, 1H), 1.00 (s, 9H).

¹³C NMR (151 MHz, CDCl₃) δ 147.7, 141.1, 128.9, 128.4, 127.6, 126.7, 116.8, 113.1, 67.1, 34.8, 27.0.

MS (EI): m/z (%) = 240.2 ([M+H]⁺, 5), 239.2 ([M]⁺, 11), 182.1 (100).

IR (ATR): \tilde{v} =3425, 2942, 2323, 2098, 1731, 1599, 1491, 1302, 709.

Methyl 9-phenyl-9-(phenylamino)nonanoate (3i)

HN^{-Ph} O Ph O Vellow oil was obtained after column chromatography (100:1 to 20:1 pentane/EtOAc). Yield = 63% (42.7 mg, 0.13 mmol).

¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.28 (m, 4H), 7.25 – 7.19 (m, 1H), 7.08 (dd, J = 8.6, 7.3 Hz, 2H), 6.63 (t, J = 7.3 Hz, 1H), 6.52 (d, J = 7.6 Hz, 2H), 4.29 (t, J = 6.8 Hz, 1H), 4.07 (bs, 1H), 3.67 (s, 3H), 2.30 (t, J = 7.5 Hz, 2H), 1.85 – 1.73 (m, 2H), 1.72 – 1.49 (m, 2H), 1.46 – 1.38 (m, 1H), 1.34 – 1.28 (m, 7H).

¹³C NMR (101 MHz, CDCl₃) δ 174.2, 147.4, 144.2, 129.0, 128.4, 126.8, 126.3, 117.0, 113.17,

58.1, 51.4, 38.9, 34.0, 29.2, 29.0, 28.9, 26.2, 24.8.

MS (EI): m/z (%) = 340.2 ([M+H]⁺, 38), 339.2 ([M]⁺, 31), 182.1 (100).

IR (ATR): *v* = 3413, 2922, 2320, 2019, 1733, 1600, 1491, 1305, 1162, 705.

HRMS (EI+, m/z): calculated for C₂₂H₃₀NO₂ [M+H]+: 340.22711; found: 340.22723.

Methyl 4-(9-methoxy-9-oxo-1-(phenylamino)nonyl)benzoate (3j)



Yellow oil was obtained after column chromatography (20:1 to 9:1 pentane/EtOAc). Yield = 58% (46.1 mg, 0.12 mmol).

¹H NMR (400 MHz, CDCl₃) δ 7.98 (d, J = 8.4 Hz, 2H), 7.39 (d, J = 8.3 Hz, 2H), 7.07 (dd, J = 8.6, 7.3 Hz, 2H), 6.62 (t, J = 7.3 Hz, 1H), 6.45 (d, J = 7.8 Hz, 2H), 4.32 (t, J = 6.7 Hz, 1H), 4.05 (bs, 1H), 3.87 (s, 3H), 3.64 (s, 3H), 2.28 (t, J = 7.5 Hz, 2H), 1.84 – 1.70 (m, 2H), 1.63 – 1.52 (m, 3H), 1.26 (s, 7H).

¹³C NMR (101 MHz, CDCl₃) δ 174.20 166.94 149.8, 146.9, 129.9, 129.0, 128.8, 126.3, 117.4, 113.2, 58.1, 51.9, 51.3, 38.7, 33.9, 29.2, 29.0, 28.9, 26.1, 24.8.

MS (EI): m/z (%) = 398.3 ([M+H]⁺, 3), 397.2 ([M]⁺, 7), 240.2 (100).

IR (ATR): *v* = 3406, 2929, 2328, 2086, 1705, 1604, 1488, 1350, 1089, 710.

HRMS (EI+, m/z): calculated for C₂₄H₃₁NO₄K [M+K]+: 436.18847; found: 436.18842.

Methyl 9-(4-fluorophenyl)-9-(phenylamino)nonanoate (3k)



¹H NMR (600 MHz, CDCl₃) δ 7.29 (dt, *J* = 14.4, 7.2 Hz, 2H), 7.09 (t, *J* = 7.9 Hz, 2H), 7.00 (t, *J* = 8.6 Hz, 2H), 6.65 (t, *J* = 7.3 Hz, 1H), 6.49 (d, *J* = 7.9 Hz, 2H), 4.27 (t, *J* = 6.8 Hz, 1H), 4.05 (bs, 1H), 3.67 (s, 3H), 2.31 (t, *J* = 7.5 Hz, 2H), 1.81 – 1.69 (m, 2H), 1.65 – 1.58 (m, 2H), 1.43 – 1.36 (m, 1H), 1.29 (s, 7H).

¹³C NMR (151 MHz, CDCl₃) δ 174.2, 161.7 (d, *J* = 244.3 Hz), 147.2, 139.9 (d, *J* = 3.0 Hz), 129.0, 127.7 (d, *J* = 7.9 Hz), 117.2, 115.3 (d, *J* = 21.2 Hz), 113.2, 57.5, 51.47, 39.0, 34.0, 29.2, 29.0, 29.0, 26.1, 24.8.

¹⁹F NMR (376 MHz, CDCl₃) δ -116.32.

MS (EI): m/z (%) = 358.2 ([M+H]⁺, 15), 357.2 ([M]⁺, 16), 200.1 (100).

IR (ATR): \tilde{v} = 3401, 2928, 2335, 2092, 1731, 1601, 1499, 1214, 744.

HRMS (EI+, m/z): calculated for C₂₂H₂₈NO₂FNa [M+Na]+: 380.19963; found: 380.20059.

Methyl 9-(4-chlorophenyl)-9-(phenylamino)nonanoate (3l)

¹H NMR (400 MHz, CDCl₃) δ 7.26 (s, 4H), 7.08 (dd, J = 8.6, 7.3 Hz, 2H), 6.63 (t, J = 7.3 Hz, 1H), 6.46 (d, J = 7.2 Hz, 2H), 4.25 (t, J = 6.8 Hz, 1H), 4.02 (bs, 1H), 3.65 (s, 3H), 2.28 (t, J = 7.5 Hz, 2H), 1.78 – 1.66 (m, 2H), 1.65 – 1.56 (m, 2H), 1.41 – 1.34 (m, 1H), 1.28 (s, 7H).

¹³C NMR (101 MHz, CDCl₃) δ 174.2, 147.1, 142.8, 132.3, 129.0, 128.6, 127.7, 117.3, 113.2, 57.6, 51.4, 38.8, 34.0, 29.2, 29.0, 28.9, 26.1, 24.8.

MS (EI): m/z (%) = 374.2 ([M+H]⁺, 40), 373.2 ([M]⁺, 24), 216.1 (100).

IR (ATR): *v* = 3403, 2928, 2322, 2080, 1730, 1600, 1491, 1183, 1012, 734.

HRMS (EI+, m/z): calculated for C₂₂H₂₉NO₂Cl [M+H]+: 374.18813; found: 374.18817.

Methyl 9-(4-bromophenyl)-9-(phenylamino)nonanoate (3m)

¹H NMR (400 MHz, CDCl₃) δ 7.43 (d, *J* = 8.4 Hz, 2H), 7.22 (d, *J* = 8.4 Hz, 2H), 7.08 (dd, *J* = 8.7, 7.3 Hz 2H), 6.63 (t, *J* = 7.3 Hz, 1H), 6.46 (d, *J* = 7.4 Hz, 2H), 4.23 (t, *J* = 6.8 Hz, 1H), 4.03 (bs, 1H), 3.65 (s, 3H), 2.28 (t, *J* = 7.5 Hz, 2H), 1.79 – 1.66 (m, 2H), 1.65 – 1.54 (m, 2H), 1.44 – 1.35 (m, 1H), 1.28 (s, 7H).

¹³C NMR (101 MHz, CDCl₃) δ 174.2, 147.1, 143.4, 131.6, 129.0, 128.1, 120.4, 117.3, 113.2,

57.7, 51.4, 38.8, 34.0, 29.2, 29.0, 28.9, 26.1, 24.8.

MS (EI): m/z (%) = 418.3 ([M+H]⁺, 16), 417.2 ([M]⁺, 24), 260.0 (100).

IR (ATR): *v* = 3401, 2926, 2338, 2091, 1731, 1600, 1490, 1195, 745.

HRMS (EI+, m/z): calculated for C₂₂H₂₉NO₂Br [M+H]+: 418.13762; found: 418.13763.

Methyl 9-(2-fluorophenyl)-9-(phenylamino)nonanoate (3n)

HN^{-Ph} O F Vellow oil was obtained after column chromatography (100:1 to 20:1 pentane/EtOAc). Yield = 65% (46.4 mg, 0.13 mmol).

¹H NMR (400 MHz, CDCl₃) δ 7.29 – 7.25 (m, 1H), 7.17 – 7.02 (m, 4H), 6.90 (t, *J* = 8.3 Hz, 1H), 6.65 (t, *J* = 7.3 Hz, 1H), 6.49 (d, *J* = 7.7 Hz, 2H), 4.28 (t, *J* = 6.7 Hz, 1H), 4.04 (bs, 1H), 3.66 (s, 3H), 2.29 (t, *J* = 7.5 Hz, 2H), 1.83 – 1.71 (m, 2H), 1.64 – 1.56 (m, 2H), 1.45 – 1.38 (m, 1H), 1.29 (s, 7H).

¹³C NMR (101 MHz, CDCl₃) δ 174.2, 163.14 (d, J = 245.8 Hz), 147.26 (d, J = 5.9 Hz), 147.1, 130.0, 129.96 (d, J = 8.1 Hz), 121.99 (d, J = 2.5 Hz), 117.3, 113.74 (d, J = 21.3 Hz)., 113.2, 113.10 (d, J = 21.7 Hz), 57.8, 51.4, 38.8, 34.01, 29.2, 29.0, 28.9, 26.1, 24.8.

¹⁹F NMR (376 MHz, CDCl₃) δ -113.12.

MS (EI): m/z (%) = 358.2 ([M+H]⁺, 13), 357.2 ([M]⁺, 12), 200.1 (100).

IR (ATR): *v* =3396, 2915, 2348, 2091, 1736, 1597, 1475, 1215, 736.

HRMS (EI+, m/z): calculated for C₂₂H₂₈NO₂FNa [M+Na]+: 380.19963; found: 380.20053.

Methyl 9-(phenylamino)-9-(p-tolyl)nonanoate (30)



Yellow oil was obtained after column chromatography (100:1 to 20:1 pentane/EtOAc). Yield = 58% (40.9 mg, 0.12 mmol).

¹H NMR (400 MHz, CDCl₃) δ 7.24 – 7.10 (m, 4H), 7.10 – 7.05 (m, 2H), 6.62 (t, *J* = 7.3 Hz, 1H), 6.51 (d, *J* = 8.6 Hz, 2H), 4.26 (t, *J* = 6.8 Hz, 1H), 4.03 (bs, 1H), 3.67 (s, 3H), 2.33 (s, 3H), 2.29 (t, *J* = 7.5 Hz, 2H), 1.84 – 1.71 (m, 2H), 1.64 – 1.56 (m, 2H), 1.45 – 1.38 (m, 1H), 1.31 (s, 7H).

¹³C NMR (101 MHz, CDCl₃) δ 174.2, 147.5, 141.1, 136.3, 129.1, 129.0, 126.2, 116.9, 113.1, 57.8, 51.4, 38.9, 34.0, 29.2, 29.0, 29.0, 26.2, 24.8, 21.0.
MS (EI): *m/z* (%) = 354.3 ([M+H]⁺, 9), 353.3 ([M]⁺, 12), 196.1 (100).
IR (ATR): *ṽ* = 3402, 2925, 2337, 2094, 1732, 1600, 1497, 1201, 739.
HRMS (EI+, m/z): calculated for C₂₃H₃₁NO₂Na [M+Na]+: 376.22470; found: 376.22495.

Methyl 9-(4-methoxyphenyl)-9-(phenylamino)nonanoate (3p)

HN^{-Ph} (20:1 to 9:1 pentane/EtOAc). Yield = 45% (33.2 mg, 0.09 mmol).

¹H NMR (400 MHz, CDCl₃) δ 7.24 (d, *J* = 5.7 Hz, 2H), 7.08 (t, *J* = 7.8 Hz, 2H), 6.85 (d, *J* = 8.6 Hz, 2H), 6.62 (t, *J* = 7.3 Hz, 1H), 6.54 – 6.48 (d, 2H), 4.24 (t, *J* = 6.8 Hz, 1H), 4.02 (bs, 1H), 3.79 (s, 3H), 3.66 (s, 3H), 2.29 (t, *J* = 7.5 Hz, 2H), 1.83 – 1.69 (m, 2H), 1.68 – 1.48 (m, 2H), 1.44 – 1.35 (m, 1H), 1.28 (s, 7H).

¹³C NMR (101 MHz, CDCl₃) δ 174.2, 158.4, 147.4, 136.2, 129.0, 127.3, 117.0, 113.8, 113.2, 57.5, 55.1, 51.4, 38.9, 34.0, 29.2, 29.0, 29.0, 26.2, 24.8.

MS (EI): m/z (%) = 370.3 ([M+H]⁺, 5), 369.3 ([M]⁺, 12), 212.1 (100).

IR (ATR): *v* = 3403, 2928, 2327, 2081, 1713, 1603, 1490, 1241, 1806, 713.

HRMS (EI+, m/z): calculated for C₂₃H₃₁NO₃Na [M+Na]+: 392.21962; found: 392.21994.

4-chloro-N-(9-methoxy-1-phenyldec-9-en-1-yl)aniline (3q)



Yellow oil was obtained after column chromatography (100:1 to 20:1 pentane/EtOAc). Yield = 64% (47.7 mg, 0.13 mmol).

Ph $^{-1}$ H NMR (400 MHz, CDCl₃) δ 7.33 – 7.26 (m, 4H), 7.23 – 7.18 (m, 1H), 7.01 (d, *J* = 8.8 Hz, 2H), 6.42 (d, *J* = 8.8 Hz, 2H), 4.22 (t, *J* = 6.8 Hz, 1H), 4.07 (bs, 1H), 3.64 (s, 3H), 2.27 (t, *J* = 7.5 Hz, 2H), 1.79 – 1.70 (m, 2H), 1.63 – 1.54 (m, 2H), 1.43 – 1.34 (m, 1H), 1.27 (s, 7H).

¹³C NMR (101 MHz, CDCl₃) δ 174.2, 145.9, 143.6, 128.8, 128.5, 126.9, 126.2, 121.6, 114.2, 58.3, 51.4, 38.8, 34.0, 29.2, 29.0, 28.9, 26.1, 24.8.

MS (EI): m/z (%) = 374.2 ([M+H]⁺, 10), 373.2 ([M]⁺, 16), 216.1 (100).

IR (ATR): \tilde{v} =3401, 2927, 2330, 2083, 1729, 1600, 1492, 1187, 812, 701.

HRMS (EI+, m/z): calculated for C₂₂H₂₉NO₂Cl [M+H]+: 374.18813; found: 374.18851.

Methyl 9-((4-methoxyphenyl)amino)-9-phenylnonanoate (3r)

Yellow oil was obtained after column chromatography (20:1 to HN 9:1 pentane/EtOAc). Yield = 41% (30.2 mg, 0.08 mmol). ¹H NMR (400 MHz, CDCl₃) δ 7.36 – 7.27 (m, 4H), 7.24 – 7.17 (m, 1H), 6.67 (d, *J* = 8.9 Hz, 2H), 6.46 (d, *J* = 8.9 Hz, 2H), 4.21 (t, *J* = 6.8 Hz, 1H), 3.84 (bs, 1H), 3.68 (s, 3H), 3.66 (s, 3H), 2.28 (t, *J* = 7.5 Hz, 2H), 1.84 – 1.70 (m, 2H), 1.68 – 1.50 (m, 2H), 1.42 – 1.36 (m, 1H), 1.28 (s, 7H). ¹³C NMR (101 MHz, CDCl₃) δ 174.2, 151.8, 144.4, 141.7, 128.4, 126.7, 126.4, 114.7, 114.4, 59.0, 55.7, 51.4, 38.9, 34.0, 29.3, 29.0, 29.0, 26.2, 24.8.

MS (EI): m/z (%) = 370.3 ([M+H]⁺, 16), 369.2 ([M]⁺, 28), 212.1 (100).

IR (ATR): *v* =3398, 2927, 2331, 2079, 1731, 1505, 1228, 1035, 813, 723.

HRMS (EI+, m/z): calculated for C₂₃H₃₁NO₃Na [M+Na]+: 392.21962; found: 392.22009.

Methyl 9-phenyl-9-((4-(trifluoromethyl)phenyl)amino)nonanoate (3s)



Yellow oil was obtained after column chromatography (100:1 to 20:1 pentane/EtOAc). Yield = 51% (41.5 mg, 0.10 mmol). ¹H NMR (600 MHz, CDCl₃) δ 7.33 – 7.30 (m, 4H), 7.25 –

7.21 (m, 1H), 7.14 (t, *J* = 7.9 Hz, 1H), 6.85 (d, *J* = 7.6 Hz, 1H), 6.76 (s, 1H), 6.60 (d, *J* = 8.2 Hz, 1H), 4.30 (t, *J* = 6.7 Hz, 2H), 3.66 (s, 3H), 2.30 (t, *J* = 7.5 Hz, 2H), 1.85 – 1.74 (m, 2H), 1.62 – 1.58 (m 2H), 1.45 – 1.38 (m, 1H), 1.29 (s, 7H).

¹³C NMR (151 MHz, CDCl₃) δ 174.2, 147.4, 143.3, 131.2 (q, *J* = 31.7 Hz), 128.6, 127.1, 126.2, 124.2 (q, *J* = 270.2 Hz), 115.7, 113.4, 109.7, 58.1, 51.4, 38.7, 33.9, 29.1, 29.0, 28.9, 26.1, 24.8.

¹⁹F NMR (282 MHz, CDCl₃) δ -62.96.

MS (EI): m/z (%) = 408.3 ([M+H]⁺, 25), 407.3 ([M]⁺, 17), 250.2 (100).

IR (ATR): \tilde{v} =3395, 2930, 2331, 2103, 1727, 1613, 1492, 1440, 1342, 1161, 782, 698.

HRMS (EI+, m/z): calculated for C₂₃H₂₈NO₂F₃Na [M+Na]+: 430.19643; found: 430.19659.

Methyl 9-(phenylamino)-9-(thiophen-2-yl)nonanoate (3t)

¹H NMR (400 MHz, CDCl₃) δ 7.18 – 7.09 (m, 3H), 6.97 – 6.91 (m, 2H), 6.68 (t, *J* = 7.3 Hz, 1H), 6.60 (d, *J* =7.6 Hz, 2H), 4.61 (t, *J* = 6.8 Hz, 1H), 3.99 (bs, 1H), 3.67 (s, 3H), 2.29 (t, *J* = 7.5 Hz, 2H), 1.92 – 1.82 (m, 2H), 1.64 – 1.57 (m, 2H), 1.47 – 1.39 (m, 1H), 1.35 – 1.27 (m, 7H).

¹³C NMR (101 MHz, CDCl₃) δ 174.2, 149.3, 147.1, 129.1, 126.6, 123.5, 123.4, 117.6, 113.3, 54.1, 51.4, 38.8, 34.0, 29.1, 29.0, 29.0, 26.0, 24.8.

MS (EI): m/z (%) = 346.3 ([M+H]⁺, 16), 345.3 ([M]⁺, 31), 188.1 (100).

IR (ATR): \tilde{v} =3394, 3053, 2928, 2855, 2329, 2113, 1732, 1600, 1432, 1171, 749.

HRMS (EI+, m/z): calculated for C₂₀H₂₇NO₂SNa [M+Na]+: 368.16547; found: 368.16519.

Methyl 9-(phenylamino)-9-(pyridin-3-yl)nonanoate (3u)



Yellow oil was obtained after column chromatography (9:1 to 6:1 pentane/EtOAc). Yield = 49% (33.3 mg, 0.10 mmol). ¹H NMR (400 MHz, CDCl₃) δ 8.61 (s, 1H), 8.48 (d, *J* = 3.9 Hz, 1H), 7.65 (dt, *J* = 7.9, 1.8 Hz, 1H), 7.23 (dd, *J* = 7.8,

4.8 Hz, 1H), 7.12 – 7.05 (m, 2H), 6.65 (t, *J* = 7.3 Hz, 1H), 6.50 – 6.45 (m, 2H), 4.34 (t, *J* = 6.8 Hz, 1H), 3.66 (s, 3H), 2.29 (t, *J* = 7.5 Hz, 2H), 1.86 – 1.72 (m, 2H), 1.67 – 1.55 (m, 3H), 1.40 – 1.38 (m, 1H), 1.31 (s, 7H).

¹³C NMR (101 MHz, CDCl₃) δ 174.2, 148.5, 148.3, 146.8, 139.6, 133.9, 129.1, 123.6, 117.6, 113.2, 55.9, 51.4, 38.7, 33.9, 29.1, 29.0, 28.9, 26.0, 24.8.

MS (EI): m/z (%) = 341.3 ([M+H]⁺, 27), 340.3 ([M]⁺, 25), 183.1 (100).

IR (ATR): \tilde{v} =3368, 3253, 2928, 2856, 2315, 2100, 1738, 1599, 1496, 1429, 1318, 1166, 708. HRMS (EI+, m/z): calculated for C₂₁H₂₉N₂O₂ [M+H]+: 341.22235; found: 341.22244.

(3R,5S,7R,8R,9S,10S,12S,13R,14S,17R)-10,13-dimethyl-17-((2R)-5-phenyl-5-(phenylami no)pentan-2-yl)hexadecahydro-1H-cyclopenta[a]phenanthrene-3,7,12-triol (5)



After 48h yellow solid was obtained after column chromatography (2:1 pentane/EtOAc to 100% EtOAc). Yield = 54% (58.8 mg, 0.11 mmol).

¹H NMR (400 MHz, CDCl₃) δ 7.36 – 7.25 (m, 4H), 7.21 – 7.17 (m, 1H), 7.08 (t, *J* = 7.7 Hz, 2H), 6.64 (t, *J* = 7.3

Hz, 1H), 6.54 (d, J = 5.6 Hz, 2H), 4.28 – 4.18 (m, 1H),

3.99 (bs, 1H), 3.96 – 3.92 (m, 1H), 3.83 (s, 1H), 3.46 – 3.39 (m, 1H), 2.40 (s, 3H), 2.24 – 2.16 (m, 2H), 1.96 – 1.86 (m, 3H), 1.78 – 1.71 (m, 4H), 1.69 – 1.60 (m, 3H), 1.50 – 1.35 (m, 7H), 1.27 – 1.20 (m, 2H), 1.12 – 1.03 (m, 2H), 0.99 – 0.95 (m, 3H), 0.89 – 0.85 (m, 3H), 0.69 – 0.64 (m, 4H).

¹³C NMR (101 MHz, CDCl₃) δ 129.03, 128.47, 126.94, 126.89, 126.54, 126.38, 113.39, 73.03, 71.92, 68.44, 46.37, 41.66, 41.43, 39.59, 39.49, 36.84, 35.34, 35.23, 34.71, 32.33, 32.24, 30.47, 28.14, 27.45, 26.40, 23.19, 22.44, 17.73, 17.13, 12.46, 10.46.

LRMS (ESI): $m/z = 546.39215 ([M+H]^+)$.

IR (ATR): *v* =3386, 2929, 1901, 1738, 1601, 1366, 1217, 1052, 726.

m.p.: 117- 122 °C.

HRMS (ESI+, m/z): calculated for C₃₆H₅₂NO₃ [M+H]+: 546.39417; found: 546.39233.

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1. NMR spectra

N-(1,4-diphenylbutyl)aniline (3a)





N-(1,3-diphenylpropyl)aniline (3b)



N-(1-phenylheptyl)aniline (3c)



N-(1-phenylpropyl)aniline (3d)



N-(4-methyl-1-phenyloctyl)aniline (3e)



N-(1-phenyltetradecyl)aniline (3f)



N-(cyclohexyl(phenyl)methyl)aniline (3g)



N-(2,2-dimethyl-1-phenylpropyl)aniline (3h)



Methyl 9-phenyl-9-(phenylamino)nonanoate (3i)





Methyl 9-(4-fluorophenyl)-9-(phenylamino)nonanoate (3k)



Methyl 9-(4-fluorophenyl)-9-(phenylamino)nonanoate (3k)



Methyl 9-(4-chlorophenyl)-9-(phenylamino)nonanoate (3l)





Methyl 9-(4-bromophenyl)-9-(phenylamino)nonanoate (3m)





Methyl 9-(2-fluorophenyl)-9-(phenylamino)nonanoate (3n)





Methyl 9-(2-fluorophenyl)-9-(phenylamino)nonanoate (3n)



20 10 -60 -70 -80 fl (ppm) -100 -110 -120 0 -10 -20 -30 -40 -50 -90 -130 -140 -150 -160 -170





Methyl 9-(4-methoxyphenyl)-9-(phenylamino)nonanoate (3p)















Methyl 9-phenyl-9-((3-(trifluoromethyl)phenyl)amino)nonanoate (3s)



Methyl 9-(phenylamino)-9-(thiophen-2-yl)nonanoate (3t)





Methyl 9-(phenylamino)-9-(pyridin-3-yl)nonanoate (3u)





(3R, 5S, 7R, 8R, 9S, 10S, 12S, 13R, 14S, 17R) - 10, 13 - dimethyl - 17 - ((2R) - 5 - phenyl - 5 - (phenyl - 10, 10, 10)) - (10, 10



10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 fl (ppm)

