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# **Supporting Information**

## For

# Visible Light-Mediated Atom Transfer Radical Addition of Styrene: Base Controlled Selective (Phenylsulfonyl)difluoromethylation

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## **General Information:**

NMR spectra were recorded on Bruker-400 (400 MHz for <sup>1</sup>H; 100 MHz for <sup>13</sup>C and 376 MHz for <sup>19</sup>F {<sup>1</sup>H decoupled}) instruments internally referenced to SiMe<sub>4</sub> signal. High resolution mass spectra were recorded on P-SIMS-Gly of Bruker Daltonics Inc. using ESI-TOF (electrospray ionization-time of flight) or Micromass GCT using EI (electron impact). Ru(bpy)<sub>3</sub>Cl<sub>2</sub><sup>1</sup> was were prepared according to the literature procedures and recrystallized from hot water. K<sub>2</sub>HPO<sub>4</sub> were purchased from Alfa and used as received. NaOAc were purchased from Sinopharm and used as received. DABCO were purchased from Aladdin and used as received. CHCl<sub>3</sub> was distilled from CaH before use. Irradiation of visible light was performed with two 26 w compact fluorescent light bulbs, showed as below.



# **Tables of the Optimization of Reaction Conditions:**

la	+ PhSO <sub>2</sub> C	$F_{2}I = \frac{K_{2}HPO_{4} (2.0 \text{ G})}{Solvent (0.5 \text{ mL})}$ $F_{2}I = \frac{K_{2}HPO_{4} (2.0 \text{ G})}{Solvent (0.5 \text{ mL})}$	0 (1 mol%) equiv) ), 40 °C bulb h	P Za	R + 3a
entry	solvent	yield (2a:3a) (%) <sup>b</sup>	entry	solvent	yield (2a:3a) (%) <sup>b</sup>
1	DMF	trace/-	6	DCM	35/1
2	DMSO	trace/-	7	CHCI <sub>3</sub>	42/0
3	Acetone	12/0	8	Cl <sub>2</sub> CHCH	Cl <sub>2</sub> 34/1
4	MeCN	trace/-	9	MeOH	0/-
5	EtOAc	12/0	10	THF	25/0

Table S1. Solvent Screening:<sup>a</sup>

 $^{a}$  **1a** (0.3 mmol, 1.5 equiv), PhSO<sub>2</sub>CF<sub>2</sub>I (0.2 mmol, 1.0 equiv), N<sub>2</sub>, 24 h.  $^{b}$  Yield determined by  $^{19}\text{F}$  NMR yield

Table S2. Base Screening: <sup>a</sup>

		F	Ru(bpy) <sub>3</sub> Cl <sub>2</sub> •6H <sub>2</sub> O( Base (2.0 equiv	1 mol%) /)	R	R
	la +	P1150 <sub>2</sub> 0F <sub>2</sub> 1	CHCl <sub>3</sub> (0.5 mL), 40 °C 26 W light bulb R=SO <sub>2</sub> Ph <b>2</b>		2a +	3a
	entry	base	yield( <b>2a:3a</b> ) (%)	entry	base	yield( <b>2a:3a</b> ) (%)
	1	Na <sub>2</sub> CO <sub>3</sub>	40/1	8	DABCO	0/92
	2	$Cs_2CO_3$	0/-	9	none	31/2
	3	NaOAc	52/0	10	K <sub>3</sub> PO <sub>4</sub>	33/3
	4	AgOAc	trace/-	11	TsNa	45/4
	5	KHCO <sub>3</sub>	35/1	12	CF <sub>3</sub> COONa	trace/-
	6	NaHCO <sub>3</sub>	43/3	13	NaOH	0/-
_	7	NEt <sub>3</sub>	0/27			

<sup>a</sup> **1a** (0.3 mmol, 1.5 equiv), PhSO<sub>2</sub>CF<sub>2</sub>I (0.2 mmol, 1.0 equiv), N<sub>2</sub>, 24 h. <sup>b</sup> Yield determined by <sup>19</sup>F NMR yield

 Table S3. Concentration Screening:<sup>a</sup>

la la	+ PhSO <sub>2</sub> CF <sub>2</sub>	$ \begin{array}{c} \text{Ru(bpy)}_{3}\text{Cl}_{2} \cdot 6\text{H}_{2}\text{O} \\ \text{NaOAc (2.0 ed)} \\ \hline \text{CHCl}_{3} (x \text{ mL}), 40 \\ 26 \text{ W light b} \\ \text{R=SO}_{2}\text{P} \end{array} $	(1 mol%) quiv) ) °C ulb h	R 2a	R 3a
entry	CHCl <sub>3</sub> (mL)	yield (2a:3a) (%) <sup>b</sup>	entry	CHCl <sub>3</sub> (mL)	yield (2a:3a) (%) <sup>b</sup>
1	0.2	31/1	3	2	46/0
2	1	48/0	4	4	86/1

 $\frac{4 \text{ v}/\text{U}}{^{a} \text{ 1a} (0.3 \text{ mmol}, 1.5 \text{ equiv}), \text{PhSO}_2 \text{CF}_2 \text{I} (0.2 \text{ mmol}, 1.0 \text{ equiv}), \text{N}_2, 24 \text{ h}. \text{ }^{b} \text{ Yield determined by }}{^{19} \text{F NMR yield}}$ 

## **Preparation of Substrates:**

All the styrene derivatives **1b-s** were prepared according to the literature.<sup>2</sup> **4a** was purchased from Energy Chemistry and used as received. **4b**, **4e** and **4f** was were prepared according to the literature.<sup>3</sup> **4c** was prepared according to the literature.<sup>4</sup> **4d** was prepared according to the literature.<sup>5</sup> PhSO<sub>2</sub>CF<sub>2</sub>I was prepared according to the literature.<sup>6</sup>

## **General Procedure of ATRA Reaction of Styrene:**

Ru(bpy)<sub>3</sub>Cl<sub>2</sub>·6H<sub>2</sub>O (1.5 mg, 0.002 mmol), NaOAc (33 mg, 0.4 mmol) and PhSO<sub>2</sub>CF<sub>2</sub>I (63.6 mg, 0.2 mmol) were combined in an oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). Styrene **1a** (31.2 mg, 0.3 mmol) and CHCl<sub>3</sub> (4.0 mL) were added *via* syringe. The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 40 °C with two 26 W compact fluorescent light bulbs (one on either side of the vial approximately 5 cm away) for 24 h. The mixture was then cooled to room temperature, filtered through a plug of silica and washed with EtOAc. The filtrate was concentrated under vacuum and purified by preparatory thin layer chromatography (PE:EA = 30:1) to give product **2a** as a white solid (70.0 mg, 83%). (Note: All of the reactions were very sensitive to the size of the catalyst and powdery c atalyst was needed to obtain reproducible results).



**2a:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.93 (d, J = 7.6 Hz, 2H), 7.74 (t, J = 7.2 Hz, 1H), 7.61-7.57 (m, 2H), 7.42 (d, J = 7.2 Hz, 2H), 7.31-7.22 (m, 3H), 5.47 (dd, J = 9.2, 5.6

Hz, 1H), 3.62-3.40 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  142.6, 135.7, 131.6, 130.9, 129.4, 128.9, 128.6, 126.9, 122.9 (t, *J* = 287.2 Hz), 40.6 (t, *J* = 18.6 Hz), 17.1 (t, *J* = 2.3 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.4 (d, *J* = 227.8 Hz), -104.4 (d, *J* = 227.8 Hz). HRMS ESI (*m*/*z*): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>13</sub>O<sub>2</sub>F<sub>2</sub>SINa: 444.9547, found: 444.9558.



**2b**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.92 (d, *J* = 7.6 Hz, 2H), 7.75 (tt, *J* = 7.6, 1.2 Hz, 1H), 7.61-7.57 (m, 2H),

7.37-7.34 (m, 2H), 7.28-7.25 (m, 2H), 5.45-5.42 (m, 1H), 3.59-3.39 (m,2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  141.2, 135.7, 134.2, 131.5, 130.9, 129.5, 129.1, 128.3, 122.8 (t, 288.7 Hz), 40.7 (t, *J* = 18.8 Hz), 15.7 (t, *J* = 2.3 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -100.8 (d, *J* = 228.8 Hz), -104.4 (d, *J* = 228.8 Hz). HRMS ESI (*m*/*z*): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>12</sub>O<sub>2</sub>F<sub>2</sub>SIClNa: 478.9157, found: 478.9156.



2c: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.92 (d, J = 7.6 Hz, 2H), 7.74 (tt, J = 7.6, 1.2 Hz, 1H), 7.60-7.56 (m, 2H), 7.42-7.39 (m, 2H), 6.99-6.95 (m, 2H), 5.47-5.45 (m, 2H), 5.47-5

1H), 3.59-3.39 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  162.3 (d, J = 247.3 Hz), 138.6 (d, J = 3.4 Hz), 135.7, 131.5, 130.9, 129.5, 128.7 (d, J = 8.4 Hz), 122.9 (t, J = 288.8 Hz), 115.9 (d, J = 21.9 Hz), 40.9 (t, J = 18.7 Hz), 16.0 (t, J = 2.0 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.0 (d, J = 228.6 Hz), -104.4 (d, J = 228.6 Hz), -112.4. HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>12</sub>O<sub>2</sub>F<sub>3</sub>SINa: 462.9452, found: 462.9451.



**2d**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.92 (d, *J* = 7.7 Hz, 2H), 7.75 (t, *J* = 7.2 Hz, 1H), 7.62-7.56 (m, 2H), 7.43-7.41 (m, 2H), 7.33-7.27 (m, 2H), 5.44-5.40 (m,

1H), 3.58-3.38 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  141.7, 135.8, 132.1, 131.5, 131.0, 129.5, 128.6, 122.8 (t, *J* = 288.4 Hz), 122.4, 40.7 (t, *J* = 18.6 Hz), 15.7 (t, *J* = 2.2 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -100.7 (d, *J* = 228.8 Hz), -104.3 (d, *J* = 228.8 Hz). HRMS ESI (*m*/*z*): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>12</sub>O<sub>2</sub>F<sub>2</sub>SBrINa: 522.8652, found: 522.8661.



**2e**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.93 (d, J = 7.6 Hz, 2H), 7.75 (tt, J = 7.2, 1.2 Hz, 1H), 7.61-7.57 (m, 2H), 7.55 (t, J = 2.0 Hz, 1H), 7.38-7.34 (m, 2H), 7.17 (t, J = 8.0 Hz, 1H), 5.40-5.36 (m, 1H), 3.57-3.38 (m, 2H). <sup>13</sup>C

NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  144.8, 135.8, 131.7, 131.5, 131.0, 130.4, 130.0, 129.5, 125.7, 122.8 (t, *J* = 288.8 Hz), 122.6, 40.6 (t, *J* = 18.8 Hz), 15.0 (t, *J* = 2.3 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.0 (d, *J* = 228.7 Hz), -104.3 (d, *J* = 228.8 Hz). HRMS ESI (*m*/*z*): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>12</sub>O<sub>2</sub>F<sub>2</sub>SBrINa: 522.8652, found: 522.8653.



**2f:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.93 (d, *J* = 7.6 Hz, 2H), 7.75 (tt, *J* = 7.6, 1.2 Hz, 1H), 7.61-7.57 (m, 2H), 7.30-7.23 (m, 1H), 7.19 (dt, *J* = 7.6, 1.2 Hz, 1H), 7.12 (dt, *J* = 9.6, 2.0 Hz, 1H), 6.94 (tdd, *J* = 8.4, 2.4, 1.2 Hz, 1H),

5.44-5.40 (m, 1H), 3.58-3.37 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  162.6 (d, J = 245.9 Hz), 144.9 (d, J = 7.4 Hz), 135.8, 131.6, 131.0, 130.5 (d, J = 8.3 Hz), 129.5, 122.8 (t, J = 288.7 Hz ), 122.6 (d, J = 2.8 Hz ), 115.7 (d, J = 21.2 Hz), 114.1 (d, J = 22.5 Hz). 40.5 (t, J = 18.8 Hz), 15.3 (q, J = 2.3 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.1 (d, J = 228.2 Hz), -104.4 (d, J = 228.6 Hz), -111.8. HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>12</sub>O<sub>2</sub>F<sub>3</sub>ISNa: 462.9452, found: 462.9460.



**2g**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.86 (d, J = 7.6 Hz, 2H), 7.66 (t, J = 7.6 Hz, 1H), 7.52-7.48 (m, 3H), 7.21-7.17 (m, 2H), 7.12-7.08 (m, 1H), 5.85-5.82 (m, 1H), 3.64-3.33 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  139.6,

135.7, 131.7, 131.7, 131.5, 131.0, 130.2, 129.6, 129.5, 128.8, 127.7, 122.9 (t, J = 288.0 Hz), 39.2 (t, J = 18.8 Hz), 11.6. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -102.6 (d, J = 227.5 Hz), -104.0 (d, J = 228.9 Hz). HRMS ESI (*m*/*z*): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>12</sub>O<sub>2</sub>F<sub>2</sub>IClSNa: 478.9157, found: 478.9135.



2h: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.93 (d, J = 7.8 Hz,
2H), 7.74 (tt, J = 7.6, 1.2 Hz, 1H), 7.61-7.57 (m, 2H),
7.43 (td, J = 7.6, 1.6 Hz, 1H), 7.29-7.22 (m, 1H), 7.11 (td,
J = 7.6, 1.0 Hz, 1H), 7.01-6.96 (m, 1H), 5.67 (dd, J =

10.0, 5.3 Hz, 1H), 3.70-3.39 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  159.2 (d, *J* = 248.5 Hz), 135.7, 131.6, 131.0, 130.3 (d, *J* = 8.6 Hz,), 129.8 (d, *J* = 12.5 Hz), 129.5, 128.8 (d, *J* = 2.5 Hz), 124.7 (d, *J* = 3.6 Hz), 123.0 (t, *J* = 290.2 Hz), 116.3 (d, *J* = 21.7 Hz), 39.5 (td, *J* = 19.0, 2.2 Hz), 7.7 (q, *J* = 2.9 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -102.4 (d, *J* = 228.2 Hz), -104.7 (d, *J* = 228.2 Hz), -115.1. HRMS ESI (*m*/*z*): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>12</sub>O<sub>2</sub>F<sub>3</sub>SINa: 462.9452, found: 462.9449.



2i: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.94 (d, J = 7.6 Hz, 2H), 7.76 (tt, J = 7.6, 1.2 Hz, 1H), 7.62-7.58 (m, 2H), 7.53 (dd, J = 6.8, 2.4 Hz, 1H), 7.35 (ddd, J = 8.8, 4.4, 2.8 Hz, 1H), 6.89 (dd, J = 10.0, 8.8 Hz, 1H), 5.56 (dd, J = 10.0, 8.1 Hz, 1Hz, 1Hz), 5.56 (dd, J = 10.0, 8.1 Hz, 1Hz), 5.56 (dd, J = 10.0, 8.1 Hz), 5.56 (dd, J = 10.0, 8.1 Hz)

J = 10.0, 5.2 Hz, 1H), 3.64-3.36 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  158.4 (d, J = 249.2 Hz), 135.8, 133.2 (d, J = 8.6 Hz), 131.9 (d, J = 13.8 Hz), 131.6 (d, J = 2.8 Hz), 131.5,131.0, 129.5, 122.8 (t, J = 288.2 Hz ), 118.1 (d, J = 23.4 Hz), 116.9 (d, J = 3.5 Hz), 39.4 (td, J = 18.8, 2.2 Hz), 6.0 (q, J = 2.5 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -102.0 (d, J = 229.1 Hz), -104.7 (d, J = 229.0 Hz), -116.8. HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>11</sub>O<sub>2</sub>F<sub>3</sub>SBrINa: 540.8558, found: 540.8560.



**2j**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.92 (d, *J* = 8.0 Hz, 2H), 7.74 (t, *J* = 7.6 Hz, 1H), 7.60-7.56 (m, 2H), 7.31 (d, *J* = 8.1 Hz, 2H), 7.10 (d, *J* = 8.0 Hz, 2H), 5.47 (dd, *J* = 9.6, 5.6 Hz, 1H), 3.62-3.39 (m, 2H), 2.29 (s, 3H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 139.8, 138.6, 135.6, 131.6, 130.9, 129.6, 129.4 126.8, 123.0 (t, J = 289.0 Hz), 40.6 (t, J = 18.6 Hz), 21.3, 17.5 (t, J = 2.2 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -101.39 (d, J = 228.0 Hz), -104.61 (d, J = 228.0 Hz). RMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>15</sub>O<sub>2</sub>F<sub>2</sub>SINa: 458.9703, found: 458.9687.



2k: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.93 (d, J = 7.6 Hz, 2H), 7.74 (t, J = 7.6 Hz, 1H), 7.60-7.56 (m, 2H), 7.21 (t, J = 7.6 Hz, 1H), 7.00 (d, J = 7.6 Hz, 1H), 6.94 (s, 1H), 6.78 (dd, J = 8.0, 2.0 Hz, 1H), 5.42 (dd, J = 9.6, 5.6 Hz,

1H), 3.80 (s, 3H), 3.61-3.40 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  159.6, 144.1, 135.7, 131.6, 130.9, 129.9, 129.4, 122.9 (t, *J* = 288.9 Hz), 119.2, 114.0, 112.7, 55.4, 40.5 (t, *J* = 18.6 Hz), 16.9 (t, *J* = 2.1 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.6 (1F, *J* = 227.8 Hz), -104.5 (1F, *J* = 227.8 Hz). HRMS ESI (*m*/*z*): [M+Na]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>15</sub>O<sub>3</sub>F<sub>2</sub>SINa: 474.9652, found: 474.9654.



**2l**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) 
$$\delta$$
 7.92 (d,  $J$  = 7.8 Hz,  
2H), 7.75 (tt,  $J$  = 7.6, 1.2 Hz, 1H), 7.61-7.52 (m, 6H),

5.49-5.45 (m, 1H), 3.62-3.40 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  146.5, 135.8, 131.5, 131.0, 130.6 (q, *J* = 32.5 Hz), 129.5, 127.4, 125.9 (q, *J* = 3.7 Hz), 123.8 (q, *J* = 270.6 Hz), 122.8 (t, *J* = 288.6 Hz), 40.6 (t, *J* = 18.9 Hz), 14.9 (t, *J* = 2.2 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -62.7, -100.72 (d, *J* = 229.1 Hz), -104.33 (d, *J* = 229.0 Hz). HRMS ESI (*m*/*z*): [M+Na]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>12</sub>O<sub>2</sub>F<sub>5</sub>SINa: 512.9421, found: 512.9426.



**2m**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.92 (d, *J* = 7.6 Hz, 2H), 7.75 (t, *J* = 7.5 Hz, 1H), 7.64-7.56 (m, 4H), 7.50 (d, *J* = 7.6 Hz, 1H), 7.44 (t, *J* = 7.6 Hz, 1H), 5.48 (dd, *J* = 9.2, 6.0 Hz, 1H), 3.62-3.41 (m, 2H). <sup>13</sup>C NMR (101 MHz,

CDCl<sub>3</sub>)  $\delta$  143.7, 135.8, 131.5, 131.3 (q, J = 32.4 Hz ) 130.9, 130.4, 129.6, 129.5, 125.4 (q, J = 3.7 Hz), 123.7 (q, J = 270.9 Hz), 123.7 (q, J = 3.6 Hz), 122.8 (t, 288.4 Hz), 40.7 (t, J = 18.9 Hz), 14.9 (t, J = 2.3 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -62.73, -100.8 (d, J = 229.3 Hz), -104.2 (d, J = 229.2 Hz). HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>12</sub>O<sub>2</sub>F<sub>5</sub>SINa: 512.9421, found: 512.9418.



**2n**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.92 (d, *J* = 8.0 Hz, 2H), 7.85 (d, *J* = 8.0 Hz, 1H), 7.74 (t, *J* = 7.6 Hz, 1H), 7.60-7.56 (m, 3H), 7.53 (d, *J* = 8.0 Hz, 1H), 7.35 (t, *J* = 7.6 Hz, 1H), 5.78 (dd, *J* = 9.6, 5.2 Hz, 1H), 3.75-3.40 (m,

2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  141.5, 135.7, 132.8, 131.4, 131.0, 130.4, 129.5, 128.4, 125.9 (q, *J* = 5.8 Hz), 125.5 (q, *J* = 30.6 Hz), 124.0 (q, *J* = 272.5 Hz), 122.9 (t, *J* = 288.6 Hz), 40.4 (t, *J* = 18.7 Hz), 9.3. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -59.7, -103.7 (d, *J* = 227.4 Hz), 104.6 (dq, *J* = 227.4, 4.8 Hz). HRMS ESI (*m*/*z*): [M+Na]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>12</sub>O<sub>2</sub>F<sub>5</sub>SINa: 512.9421, found: 512.9427.



20: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.91 (d, J = 7.7 Hz, 2H), 7.76 (tt, J = 7.6, 1.2 Hz, 1H), 7.61-7.56 (m, 4H), 7.53-7.50 (m, 2H), 5.46-5.43 (m, 1H), 3.60-3.38 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 147.6, 135.9,

132.7, 131.4, 130.9, 129.5, 127.8, 122.7 (t, J = 288.3 Hz), 118.3, 112.3, 40.4 (t, J = 19.0 Hz), 14.4 (d, J = 2.5 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -100.43 (d, J = 229.5

Hz), -104.19 (d, J = 229.5 Hz). HRMS ESI (m/z):  $[M+Na]^+$  calcd. for C<sub>16</sub>H<sub>12</sub>NO<sub>2</sub>F<sub>2</sub>SINa: 469.9499, found: 469.9501.



**2p**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.96 (d, *J* = 8.4 Hz, 2H), 7.91 (d, *J* = 8.0 Hz, 2H), 7.74 (t, *J* = 7.6 Hz, 1H), 7.59-7.55 m, 2H), 7.48 (d, *J* = 8.4 Hz, 2H), 5.47 (dd, *J* = 9.6, 5.6 Hz, 1H), 3.89 (s,

3H), 3.62-3.40 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  166.4, 147.4, 135.7, 131.5, 130.9, 130.2, 129.5, 127.0, 122.8 (t, *J* = 288.6 Hz), 52.3, 40.4 (t, *J* = 18.8 Hz), 15.4 (t, *J* = 2.2 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -100.8 (d, *J* = 228.9 Hz), -104.3 (d, *J* = 228.9 Hz). HRMS ESI (*m*/*z*): [M+Na]<sup>+</sup> calcd. for C<sub>17</sub>H<sub>15</sub>O<sub>4</sub>F<sub>2</sub>SINa: 502.9601, found: 502.9604.



**2q**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.91 (d, J = 7.6 Hz, 2H), 7.89-7.86 (m, 2H), 7.76 (tt, J = 7.6, 1.2 Hz), 7.63-7.58 (m, 4H), 5.50-5.46 (m, 1H), 3.62-3.42 (m, 2H), 3.06 (s, 3H). <sup>13</sup>C NMR (101

MHz, CDCl<sub>3</sub>)  $\delta$  148.5, 140.4, 135.9, 131.3, 130.9, 129.5, 128.1, 128.0, 122.7 (t, *J* = 288.2 Hz), 44.5, 40.5 (t, *J* = 18.9 Hz), 14.2 (t, 2.5 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -100.3 (d, *J* = 228.9 Hz), -104.1 (d, *J* = 229.3 Hz). HRMS ESI (*m*/*z*): [M+Na]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>15</sub>O<sub>4</sub>F<sub>2</sub>S<sub>2</sub>INa: 522.9322, found: 522.9319.



2r: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.57 (ddd, J = 4.8, 1.6, 0.8 Hz, 1H), 7.92 (d, J = 7.6 Hz, 2H), 7.73 (td, J = 7.6, 1.2 Hz, 1H), 7.63-7.55 (m, 3H), 7.31 (dt, J = 7.8, 0.8 Hz,

1H), 7.14 (ddd, J = 7.6, 4.8, 1.2 Hz, 1H), 5.54 (dd, J = 10.0, 4.4 Hz, 1H), 4.00-3.86 (m, 1H), 3.47-3.33 (m, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 160.0, 149.7, 136.9, 135.5, 131.8, 130.8, 129.3, 123.0, 123.0 (t, J = 289.7 Hz), 120.9, 38.7 (t, J = 19.0 Hz), 17.5 (t, J = 9.6 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -101.23 (d, J = 229.6 Hz), -103.79 (d, J = 229.6 Hz). HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>14</sub>H<sub>12</sub>NO<sub>2</sub>F<sub>2</sub>SINa: 445.9499, found: 445.9498.

## **General Procedure of ATRA Reaction of Aliphatic Alkene:**

 $Ru(bpy)_{3}Cl_{2}{\cdot}\,6H_{2}O$  (1.5mg, 0.002 mmol),  $K_{2}CO_{3}$  (55.2 mg, 0.4 mmol) and

PhSO<sub>2</sub>CF<sub>2</sub>I (63.6 mg, 0.2 mmol) were combined in an oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). 1-Octene **4a** (33.6 mg, 0.3 mmol) and CHCl<sub>3</sub> (0.5 mL) were added via syringe. The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 40 °C with two 26 W compact fluorescent light bulbs (one on either side of the vial approximately 5 cm away) for 24 h. The mixture was then cooled to room temperature, filtered through a plug of silica and washed with EtOAc. The filtrate was concentrated under vacuum and purified by preparatory thin layer chromatography (PE:EA = 30:1) to give product **5a** as a pale yellow oil (75.6 mg, 88%).



5a: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.98 (d, J = 7.6 Hz, 2H), 7.77 (td, J = 7.6, 1.2 Hz, 1H), 7.64-7.60 (m, 2H), 4.38-4.31 (m, 1H), 3.23-2.99 (m, 2H), 1.87-1.70 (m, 2H),

1.58-1.47 (m, 1H), 1.46-1.35 (m, 1H), 1.33-1.22 (m, 6H), 0.87 (t, J = 7.2 Hz). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  135.7, 131.8, 130.9, 129.5, 123.5 (t, J = 287.4 Hz), 40.3, 39.8 (t, J = 18.9 Hz,), 31.6, 29.4, 28.2, 22.6, 21.4 (t, J = 7.2 Hz), 14.1. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.6 (d, J = 227.4 Hz), -103.2 (d, J = 227.1 Hz). HRMS ESI (*m*/*z*): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>21</sub>O<sub>2</sub>F<sub>2</sub>SINa: 453.0173, found: 453.0171.



**5b**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.03-8.01 (m, 2H), 7.95 (d, *J* = 7.6 Hz, 2H), 7.76 (tt, *J* = 7.6, 0.8 Hz, 1H), 7.62-7.58 (m, 2H), 7.54 (tt, J = 7.2 Hz, 1.2 Hz, 1H), 7.43-7.39 (m, 2H),

4.47-4.40 (m, 1H), 4.35 (t, J = 6.0 Hz, 2H), 3.29-3.02 (m, 2H), 2.09-1.86 (m, 4H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  166.5, 135.7, 133.0, 131.7, 130.9, 130.1, 129.6, 129.5, 128.4, 123.3 (t, J = 289.2 Hz), 63.6, 39.9 (t, J = 19.1 Hz), 36.8, 28.9, 20.1. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.1 (d, J = 228.4 Hz), -103.1 (d, J = 228.4 Hz). HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>19</sub>H<sub>19</sub>O<sub>4</sub>F<sub>2</sub>SINa: 530.9914, found: 530.9909.



**5c:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.97 (d, *J* = 7.6 Hz, 2H), 7.76 (tt, *J* = 7.6, 1.2 Hz, 1H), 7.63-7.59 (m, 2H), 7.35-7.24 (m, 5H), 4.48 (s,

2H), 4.41-4.34 (m, 1H), 3.49 (t, J = 5.6 Hz, 2H), 3.25-3.00 (m, 2H), 1.97-1.82 (m, 3H), 1.71-1.66 (m, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  138.4, 135.7, 131.7, 130.9, 129.5, 128.4, 127.7, 127.6, 123.4 (t, J = 287.5 Hz), 73.0, 68.9, 39.8 (t, J = 19.0 Hz), 37.3, 29.8, 20.9 (t, J = 1.8 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.6 (d, J = 227.4 Hz), -103.2 (d, J = 227.4 Hz). HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>19</sub>H<sub>21</sub>O<sub>3</sub>F<sub>2</sub>SINa: 517.0122, found: 517.0125.



**5d**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.97 (d, *J* = 7.6 Hz, 2H), 7.79 (t, *J* = 7.2 Hz, 1H), 7.75 (d, *J* = 8.4 Hz, 2H), 7.66-7.62 (m, 2H), 7.30 (d, *J* = 8.0 Hz, 2H), 4.78 (t, *J* 

= 6.0 Hz, 1H), 4.30-4.23 (m, 1H), 3.20-2.91 (m, 4H), 2.42 (s, 3H), 1.78-1.65 (m, 2H), 1.53-1.33 (m, 4H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  143.5, 136.9, 135.8, 131.7, 130.9, 129.8, 129.5, 127.2, 123.3 (t, *J* = 287.3 Hz), 42.9, 39.8 (t, *J* = 19.0 Hz), 39.5, 28.5, 26.5, 21.6, 20.6. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.3 (d, *J* = 227.8 Hz), -103.0 (d, *J* = 227.8 Hz). HRMS ESI (*m*/*z*): [M+Na]<sup>+</sup> calcd. for C<sub>20</sub>H<sub>24</sub>NO<sub>4</sub>F<sub>2</sub>S<sub>2</sub>INa: 594.0057, found: 594.0050.



**5e**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.97 (d, J = 7.6 Hz, 2H), 7.79 (t, J = 7.6 Hz, 1H), 7.65-7.61 (m, 2H), 4.71-4.64 (m, 1H), 3.99 (dd, J = 14.0, 8.0 Hz, 1H), 3.81 (dd, J = 13.6, 7.6 Hz, 1H), 3.23-3.04 (m, 2H), 2.76 (s,

4H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  176.7, 135.8, 131.5, 130.9, 129.5, 122.9 (t, *J* = 287.4 Hz), 46.3, 38.0 (t, *J* = 20.2 Hz), 28.2, 12.3 (t, *J* = 2.3 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -100.5 (d, *J* = 229.5 Hz), -103.7 (d, *J* = 229.5 Hz). HRMS ESI (*m*/*z*): [M+H]<sup>+</sup> calcd. for C<sub>14</sub>H<sub>15</sub>NO<sub>4</sub>IF<sub>2</sub>S: 457.9735, found: 457.9741.



5f: <sup>1</sup>H NMR (400 MHz, CDCl3) δ 7.96 (d, J = 7.9 Hz, 2H), 7.81-7.76 (m, 2H), 7.64 (t, J = 7.2 Hz, 2H), 7.55 (t, J = 8.4 Hz, 1H), 7.32 (d, J = 8.1 Hz, 1H), 7.25 (d, J = 8.8 Hz, 1H), 5.67 (s, 1H), 4.49

(s, 1H), 4.18 (s, 2H), 3.33-3.05 (m, 2H), 2.27-2.02 (m, 4H). <sup>13</sup>C NMR (101 MHz, CDCl3)  $\delta$  165.5, 162.9, 153.4, 135.8, 132.5, 131.7, 130.9, 129.5, 124.0, 123.3 (t, *J* = 289.87 Hz), 123.1, 116.9, 115.7, 90.7, 68.2, 40.1 (t, *J* = 19.2 Hz), 36.7, 28.7, 19.8 (t, *J* 

= 2.2 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -100.72 (d, J = 228.9 Hz), -102.88 (d, J = 228.9 Hz). HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>21</sub>H<sub>19</sub>O<sub>5</sub>F<sub>2</sub>SINa: 570.9864, found: 570.9858.

### **General Procedure of Heck-type Reaction of Styrene:**

Ru(bpy)<sub>3</sub>Cl<sub>2</sub>·6H<sub>2</sub>O (1.5mg, 0.002 mmol), DABCO (44.8 mg, 0.4 mmol) and PhSO<sub>2</sub>CF<sub>2</sub>I (63.6 mg, 0.2 mmol) were combined in an oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). Styrene **1a** (31.2 mg, 0.3 mmol) and CHCl<sub>3</sub> (0.5 mL) were added via syringe. The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 40 °C with two 26 W compact fluorescent light bulbs (one on either side of the vial approximately 5 cm away) for 24 h. The mixture was then cooled to room temperature, filtered through a plug of silica and washed with EtOAc. The filtrate was concentrated under vacuum and purified by preparatory thin layer chromatography (PE:EA = 30:1) to give product **3a** as a white solid (54.0 mg, 92%).



**3a**<sup>7</sup>: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.03 (d, *J* = 7.6 Hz, 2H), 7.76 (t, *J* = 7.6 Hz, 1H), 7.64-7.60 (m, 2H), 7.50-7.47 (m, 2H), 7.40-7.38 (m, 3H), 7.19 (dt, *J* = 16.4

Hz, 2.0 Hz, 1H), 6.39 (dt, J = 16.0 Hz, 12.0 Hz, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$ 142.4 (t, J = 8.8 Hz), 135.4, 133.7, 133.1, 130.9, 130.5, 129.4, 129.0, 128.0, 121.6 (t, J = 281.7 Hz), 112.5 (t, J = 22.2 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.2.



**3b**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.03 (d, *J* = 7.6 Hz, 2H), 7.76 (dt, *J* = 7.6, 1.2 Hz, 1H), 7.67 -7.60 (m, 2H), 7.50-7.45 (m, 2H), 7.16 (dt, *J* = 16.4, 2.0 Hz, 1H),

7.11-7.05 (m, 2H), 6.31 (dt, J = 16.2, 11.8 Hz, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$ 164.0 (d, J = 250.0 Hz), 141.1 (t, J = 8.9 Hz), 135.4, 133.0, 130.9, 129.9, 129.9, 129.4, 121.6 (t, J = 281.8 Hz), 116.2 (d, J = 22.0 Hz), 112.2 (td, J = 22.5, 2.3Hz ). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.1, -109.4. HRMS ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>12</sub>O<sub>2</sub>F<sub>3</sub>S: 313.0510, found: 313.0509.



**3c**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.02 (d, J = 7.6 Hz, 2H), 7.76 (tt, J = 7.2, 0.8 Hz, 1H), 7.65-7.61 (m, 2H), 7.43-7.41 (m, 2H), 7.37-7.35 (m, 2H), 7.15 (dt, *J* = 16.0,

2.0 Hz, 1H), 6.36 (dt, J = 16.4, 11.6 Hz, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  141.1 (t, *J* = 8.9 Hz), 136.5, 135.4, 133.0, 132.2, 130.9, 129.4, 129.3, 129.2, 121.5 (t, *J* = 281.9 Hz), 113.2 (t, J = 22.4 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -101.3. HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>11</sub>O<sub>2</sub>F<sub>2</sub>SClNa: 351.0034, found: 351.0028.



**3d**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.02 (d, J = 7.2 Hz, 2H), 7.77 (tt, J = 7.6, 1.2 Hz, 1H), 7.66-7.61 (m, 2H), 7.56-7.52 (m, 2H), 7.38-7.34 (m, 2H), 7.14 (dt, J = 16.0,

2.2 Hz, 1H), 6.39 (dt, J = 16.4, 11.6 Hz, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  141.2 (t, *J* = 8.9 Hz), 135.5, 133.0, 132.6, 132.3, 130.9, 129.4, 129.4, 124.8, 121.5 (t, *J* = 281.9 Hz), 113.3 (t, J = 22.4 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -101.3. HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>11</sub>O<sub>2</sub>F<sub>2</sub>SBrNa: 394.9529, found: 394.9527.



**3e**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.03 (d, J = 7.7 Hz, 2H), 7.78 (tt, J = 7.6 Hz, 1H), 7.65-7.61 (m, 2H), 7.40-7.34 (m, 1H), 7.27 (d, J = 8.0 Hz, 1H), 7.20-7.14 (m, 2H, 7.10 (td, J = 8.4, 2.0 Hz, 1H), 6.40 (dt, J = 16.0, 11.6 Hz, 1H). <sup>13</sup>C NMR (101) MHz, CDCl<sub>3</sub>)  $\delta$  163.1 (d, J = 245.7 Hz), 141.1 (td, J = 8.9, 2.6 Hz), 135.8 (d, J = 7.7 Hz), 135.5, 132.9, 130.9, 130.6 (d, J = 8.3 Hz), 129.4, 123.9 (d, J = 2.7 Hz), 121.4 (t, *J* = 281.9 Hz), 117.4 (d, *J* = 21.3 Hz), 114.4 (d, *J* = 22.0 Hz), 114.1 (t, *J* = 22.4 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.5, -112.2. HRMS ESI (*m/z*): [M+H]<sup>+</sup> calcd. for



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C<sub>15</sub>H<sub>12</sub>O<sub>2</sub>F<sub>3</sub>S: 313.0510, found: 313.0527.

**3f**:<sup>7</sup> <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.03 (d, J = 7.6 Hz, 2H), 7.78 (tt, J = 7.6, 0.8 Hz, 1H), 7.65-7.61 (m, 2H), 7.48 (s, 1H), 7.39-7.31 (m, 3H), 7.14 (dt, *J* = 16.2, 2.1 Hz,

 $\overline{1H}$ , 6.41 (dt, J = 16.2, 11.8 Hz, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  140.9 (t, J = 8.9Hz), 135.5, 135.4, 135.1, 132.9, 130.9, 130.4, 130.3, 129.4, 127.8, 126.1, 121.3 (t, J = 282.0 Hz), 114.2 (t, J = 22.5 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -101.5.



**3g**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.03 (d, J = 7.7 Hz, 2H), 7.77 (tt, J = 7.6, 0.8 Hz, 1H), 7.67 -7.60 (m, 3H), 7.52 (d, J = 8.0 Hz, 1H), 7.41 (d, J = 7.8 Hz, 1H), 7.27 (t, J = 8.0 Hz, 1H), 7.12 (tt, J = 16.0, 2.0 Hz, 1H), 6.40 (dt, J

= 16.0, 11.8 Hz, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  140.8 (t, *J* = 8.9 Hz), 135.7, 135.5, 133.3, 132.9, 130.9, 130.7, 130.5, 129.4, 126.6, 123.1, 119.9 (t, *J* = 281.9 Hz), 114.2 (t, *J* = 22.5 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.6. HRMS ESI (*m*/*z*): [M+H]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>12</sub>O<sub>2</sub>F<sub>2</sub>SBr: 372.9709, found: 372.9699.



**3h**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.03 (d, J = 7.6 Hz, 2H), 7.77 (t, J = 7.6 Hz, 1H), 7.65-7.61 (m, 2H), 7.51 (t, J = 8.4 Hz, 1H), 7.40-7.35 (m, 1H), 7.32 (dt, J = 16.4, 2.2

Hz, 1H), 7.18 (td, J = 7.6, 1.2 Hz, 1H), 7.13-7.08 (m, 1H), 6.52 (dt, J = 16.4, 11.8 Hz, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 161.2 (d, J = 252.3 Hz), 135.4, 135.2 (td, J = 9.3, 3.1 Hz), 133.0, 132.0 (d, J = 8.8 Hz), 130.9, 129.4, 129.1, 124.6 (d, J = 3.6 Hz), 121.7 (d, J = 11.5 Hz), 121.4 (d, J = 281.8 Hz), 116.3 (d, J = 21.8 Hz), 115.2 (td, J = 22.3, 7.1 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -101.7, -114.8. HRMS ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>12</sub>O<sub>2</sub>F<sub>3</sub>S: 313.0510, found: 313.0519.



**3i**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.04 (d, J = 7.6 Hz, 2H), 7.77 (t, J = 7.6 Hz, 1H), 7.65-7.56 (m, 4H), 7.42-7.40 (m, 1H), 7.35-7.28 (m, 2H), 6.41 (dt, J = 16.2, 11.8 Hz, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  138.4 (t, J =

9.1 Hz), 135.5, 134.6, 132.9, 131.93, 131.3, 130.9, 130.2, 129.4, 127.8, 127.3, 121.3 (t, J = 283.8 Hz), 115.3 (t, J = 22.4 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.7. HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>11</sub>O<sub>2</sub>F<sub>2</sub>SClNa: 351.0034, found: 351.0036.



**3j**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.03 (d, *J* = 7.9 Hz, 2H), 7.78 (t, *J* = 7.2 Hz, 1H), 7.65-7.61 (m, 3H), 7.49-7.45 (m, 1H), 7.23 (d, *J* = 17.2 Hz), 7.01 (t, *J* =

9.2 Hz, 1H), 6.52 (dt, J = 16.3, 11.6 Hz, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  160.1 (d, J = 252.9 Hz), 135.5, 134.6 (d, J = 8.7 Hz), 133.8 (td, J = 9.4, 2.9 Hz,), 132.8, 131.6 (d, J = 2.7 Hz), 131.0, 129.4, 123.6 (d, J = 13.0 Hz), 121.1 (t, J = 282.4 Hz), 118.1 (d,

J = 23.5 Hz ), 117.2 (d, J = 3.4 Hz), 116.8 (td, J = 22.5, 6.9 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -102.0, -116.9. HRMS ESI (*m*/*z*): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>10</sub>O<sub>2</sub>F<sub>3</sub>SBrNa: 412.9435, found: 412.9430.



**3k**<sup>7</sup>: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.02 (d, *J* = 8.0 Hz, 2H), 7.76 (t, *J* = 7.6 Hz, 1H), 7.64-7.60 (m, 2H), 7.38 (d, *J* = 8.0 Hz), 7.20 (d, *J* = 8.0 Hz), 7.15 (d, *J* = 16.0 Hz, 1H), 6.32 (dt, *J* = 16.4, 11.6 Hz, 1H), 2.38 (s, 3H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 142.3 (t, J = 8.8 Hz), 140.9, 135.3, 133.2, 131.0, 130.9 129.7, 129.4, 128.0, 121.8 (t, J = 281.6 Hz), 111.2 (t, J = 22.3 Hz), 21.6. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -101.0.



**31:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.02 (d, *J* = 8.0 Hz, 2H), 7.76 (t, *J* = 7.6 Hz, 1H), 7.62 (t, *J* = 7.6 Hz, 2H), 7.44 (d, *J* = 8.8 Hz, 2H), 7.13 (d, *J* = 16.4 Hz, 1H),

 $\overline{6.91}$  (d, J = 8.4 Hz, 1H),  $\overline{6.22}$  (dt, J = 16.4, 12.0 Hz, 1H), 3.84 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  161.5, 141.9 (t, J = 8.9 Hz), 135.3, 133.2, 130.9, 129.6, 129.3, 126.5, 122.0 (t, J = 281.5 Hz), 114.4, 109.6, 22.2, 55.5. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -100.7. HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>14</sub>O<sub>3</sub>F<sub>2</sub>SNa: 347.0529, found: 347.0522.



**3m:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.94 (d, J = 7.6 Hz, 2H), 7.67 (t, J = 7.2 Hz, 1H), 7.55-7.51 (m, 2H), 7.23-6.84 (m, 5H), 6.30 (dt, J = 15.6, 12.0 Hz, 1H), 3.74 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  160.0, 142.3 (t, J

= 8.8 Hz), 135.4, 135.0, 133.0, 130.9, 130.0, 129.4, 121.6 (t, J = 281.8 Hz), 120.6, 116.4, 112.8, 112.7 (t, J = 22.2 Hz), 55.4. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.3. HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>14</sub>O<sub>3</sub>F<sub>2</sub>NaS: 347.0529, found: 347.0528.



**3n**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.03 (d, J = 7.7 Hz, 2H), 7.75 (t, J = 7.5 Hz, 1H), 7.62 (t, J = 7.8 Hz, 2H), 7.49-7.41 (m, 2H), 7.39-7.31 (m, 1H), 6.97 (t, J = 7.4 Hz,

1H), 6.92 (d, J = 8.3 Hz, 1H), 6.52 (dt, J = 16.3, 12.2 Hz, 1H), 3.88 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  158.2, 137.9 (t, J = 9.4 Hz), 135.2, 133.3, 131.7, 130.9, 129.3,

129.2, 122.6, 122.0 (t, J = 281.6 Hz), 120.8, 112.9 (t, J = 22.0 Hz), 111.2, 55.5. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.1. HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>14</sub>O<sub>3</sub>F<sub>2</sub>SNa: 347.0529, found: 347.0514.



**30**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.03 (d, *J* = 7.8 Hz, 2H), 7.79 (t, *J* = 7.5 Hz, 1H), 7.67-7.59 (m, 6H), 7.24 (d, *J* = 15.6 Hz, 2H), 6.49 (dt, *J* = 16.2, 11.7 Hz, 1H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 140.8 (t, J = 8.9 Hz), 137.0, 135.6, 132.8, 132.1 (q, J = 32.8 Hz), 130.9, 129.5, 128.2, 126.0 (q, J = 3.8 Hz),123.8 (q, J = 270.6Hz ) 121.2 (t, J = 282.6Hz), 115.4 (t, J = 22.5 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -62.8, -101.6. HRMS ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>12</sub>O<sub>2</sub>F<sub>5</sub>S: 363.0478, found: 363.0488.



**3p**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.04 (d, J = 7.7 Hz, 2H), 7.78 (t, J = 7.5 Hz, 1H), 7.73 (s, 1H), 7.69-7.62 (m, 4H), 7.54 (t, J = 8.0 Hz, 1H), 7.24 (dt, J = 16.4, 2.0 Hz, 1H), 7.22 (s, 0H), 6.48 (dt, J = 16.2, 11.7 Hz, 1H). <sup>13</sup>C

NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  140.8 (t, J = 8.9 Hz), 135.5, 134.4, 132.8, 131.6 (q, J = 32.7 Hz), 131.0, 129.6, 129.51, 126.9 (q, J = 3.7 Hz), 124.7 (q, J = 3.7 Hz), 123.8 (q, J = 270N Hz), 121.3 (t, J = 282.1 Hz), 114.8 (t, J = 22.5 Hz, 1H). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -62.9, -101.5 (d, J = 1.7 Hz), -101.5 (d, J = 1.7 Hz). HRMS ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>12</sub>O<sub>2</sub>F<sub>5</sub>S: 363.0478, found: 363.0487.



**3q**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.03 (d, *J* = 8.0 Hz, 2H), 7.77 (tt, *J* = 7.6, 1.2 Hz, 1H), 7.70 (d, *J* = 8.0 Hz, 2H), 7.64-7.54 (m, 4H), 7.49 (t, *J* = 7.6 Hz, 1H), 6.40 (dt,

J = 16.0, 11.6 Hz, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  138.2 (td, J = 9.0, 2.1 Hz), 135.6, 132.7, 132.5, 132.4, 130.9, 129.9, 129.4, 128.7 (q, J = 30.4 Hz), 128.2, 126.2 (q, J = 5.5 Hz), 123.9 (q, J = 272.4 Hz), 120.9 (t, J = 282.3 Hz), 117.2 (t, J = 22.2 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -59.0, -102.2. HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>11</sub>O<sub>2</sub>F<sub>5</sub>SNa: 385.0298, found: 385.0291.



**3r**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.03 (d, *J* = 7.7 Hz, 2H), 7.79 (t, *J* = 7.5 Hz, 1H), 7.72-7.57 (m, 6H), 7.22

(dt, J = 16.4, 2.0 Hz, 1H), 6.51 (dt, J = 16.2, 11.6 Hz, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  140.3 (t, J = 8.9 Hz), 137.8, 135.6, 132.8, 132.6, 130.9, 129.5, 128.4, 121.0 (t, J = 282.4 Hz), 118.3, 116.5 (t, J = 22.6 Hz), 113.7. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -101.7. HRMS ESI (m/z): [M+H]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>12</sub>NO<sub>2</sub>F<sub>2</sub>S: 320.0557, found: 320.0550.

## **Mechanistic Investigation:**

**Competition Studies:** 



Ru(bpy)<sub>3</sub>Cl<sub>2</sub>·6H<sub>2</sub>O (1.5 mg, 0.002 mmol), NaOAc (33 mg, 0.4 mmol) and PhSO<sub>2</sub>CF<sub>2</sub>I (63.6 mg, 0.2 mmol) were combined in an oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). Styrene **1a** (31.2 mg, 0.3 mmol), **1k** (40.2 mg, 0.3 mmol) and CHCl<sub>3</sub> (4.0 mL) were added *via* syringe. The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 40 °C with two 26 W compact fluorescent light bulbs (one on either side of the vial approximately 5 cm away) for 24 h. The mixture was then cooled to room temperature, filtered through a plug of silica and washed with EtOAc. The filtrate was concentrated under vacuum and purified by preparatory thin layer chromatography (PE:EA = 30:1) to give product **2a** (20.2 mg, 24%) and **2k** (25.3 mg, 28%).



Ru(bpy)<sub>3</sub>Cl<sub>2</sub>·6H<sub>2</sub>O (1.5 mg, 0.002 mmol), NaOAc (33 mg, 0.4 mmol) and PhSO<sub>2</sub>CF<sub>2</sub>I (63.6 mg, 0.2 mmol) were combined in an oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). Styrene **1a** (31.2 mg, 0.3 mmol), **1o** (38.7 mg, 0.3 mmol) and CHCl<sub>3</sub> (4.0 mL) were added *via* syringe. The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 40 °C with two 26 W compact fluorescent light bulbs (one on either side of the vial approximately 5 cm away) for 24 h. The mixture was then cooled to room temperature, filtered through a plug of silica and washed with EtOAc. The filtrate was concentrated under vacuum and purified by preparatory thin layer chromatography (PE:EA = 30:1) to give product **2a** (30.4 mg, 36%) and **2o** (25.9 mg, 29%).

#### **Carbocation Trapping Experiment:**



Ru(bpy)<sub>3</sub>Cl<sub>2</sub>·6H<sub>2</sub>O (1.5 mg, 0.002 mmol), NaOAc (33 mg, 0.4 mmol), LiBr (43.4, 0.5 mmol) and PhSO<sub>2</sub>CF<sub>2</sub>I (63.6 mg, 0.2 mmol) were combined in an oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). Styrene **1a** (31.2 mg, 0.3 mmol) and CHCl<sub>3</sub> (4.0 mL) were added *via* syringe. The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 40 °C with two 26 W compact fluorescent light bulbs (one on either side of the vial approximately 5 cm away) for 24 h. The mixture was then cooled to room temperature, filtered through a plug of silica and washed with EtOAc. The filtrate was concentrated under vacuum and PhCF<sub>3</sub> (29.2 mg, 0.2 mmol) was added. None of **2a'** was detected by <sup>1</sup>H NMR and <sup>19</sup>F NMR of the crude material. **2a** was obtained with a low yield (10%, <sup>19</sup>F yield).



Ru(bpy)<sub>3</sub>Cl<sub>2</sub>·6H<sub>2</sub>O (1.5 mg, 0.002 mmol), NaOAc (33 mg, 0.4 mmol), KBr (59.5, 0.5 mmol) and PhSO<sub>2</sub>CF<sub>2</sub>I (63.6 mg, 0.2 mmol) were combined in an oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). Styrene **1a** (31.2 mg, 0.3 mmol) and CHCl<sub>3</sub> (4.0 mL) were added *via* syringe. The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 40 °C with two 26 W compact fluorescent light bulbs (one on either side of the vial approximately 5 cm away) for 24 h. The mixture was then cooled to room temperature, filtered through a plug of silica and washed with EtOAc. The filtrate was concentrated under vacuum and PhCF<sub>3</sub> (29.2 mg, 0.2 mmol) was added. None of **2a'** was detected by <sup>1</sup>H NMR and <sup>19</sup>F NMR of the crude material. **2a** was obtained with a moderate yield (42%, <sup>19</sup>F yield).

## **Propagation Experiments:**

In Stephenson's investigation,<sup>8</sup> ethyl bromoacetate which was inert to ATRA reaction with the optimized reaction conditions can undergo ATRA reaction when another atom transfer agent (ethyl bromomalonate or ethyl bromofluoroacetate) known to undergo ATRA reaction efficiently was added. It meaned that a propagation mechanism was operative. Here in we applied the same strategy to investigate the propagation mechanism. In our standard conditions PhSO<sub>2</sub>CF<sub>2</sub>I was a good substrate for the ATRA reaction while ethyl bromodifluoroacetate can not achieve the transformation at all. We mixed them in the same reaction to test if ethyl bromodifluoroacetate can be initiated to undergo ATRA reaction.



Ru(bpy)<sub>3</sub>Cl<sub>2</sub>·6H<sub>2</sub>O (1.5 mg, 0.002 mmol) and NaOAc (33 mg, 0.4 mmol) were combined in an oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). Styrene **1a** (31.2 mg, 0.3 mmol), EtOOCCF<sub>2</sub>Br (40.6 mg, 0.2 mmol) and CHCl<sub>3</sub> (4.0 mL) were added *via* syringe. The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 40 °C with two 26 W compact fluorescent light bulbs (one on either side of the vial approximately 5 cm away) for 24 h. None of the product **1a'** was detected by <sup>19</sup>F NMR.



Ru(bpy)<sub>3</sub>Cl<sub>2</sub>·6H<sub>2</sub>O (1.5 mg, 0.002 mmol), NaOAc (33 mg, 0.4 mmol) and PhSO<sub>2</sub>CF<sub>2</sub>I (63.6 mg, 0.2 mmol) were combined in an oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). Styrene **1a** (31.2 mg, 0.3 mmol), EtOOCCF<sub>2</sub>Br (81.2 mg, 0.4 mmol) and CHCl<sub>3</sub> (4.0 mL) were added *via* syringe. The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 40 °C with two 26 W compact fluorescent light bulbs (one on either side of the vial approximately 5 cm away) for 24 h. None of the product **1a'** was detected by <sup>19</sup>F NMR and **2a** was obtained with good yield (83%, isolated yield). *This result revealed that a propagation mechanism may not be operative*.

## General Procedure for Transformation of 2a to 3a:



Ru(bpy)<sub>3</sub>Cl<sub>2</sub>· $6H_2O$  (1.5 mg, 0.002 mmol), NaOAc (33 mg, 0.4 mmol) and **2a** (84.4 mg, 0.2 mmol) were combined in an oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 40 °C with two 26 W compact fluorescent light bulbs (one on either side of the vial

approximately 5 cm away) for 24 h. The mixture was then cooled to room temperature, filtered through a plug of silica and washed with EtOAc. The filtrate was concentrated under vacuum and purified by preparatory thin layer chromatography (PE:EA = 30:1) to give product **3a** as a white solid (54.7 mg, 93%).





General Procedure for Transformation of 2s to 3s:



Ru(bpy)<sub>3</sub>Cl<sub>2</sub>· $6H_2O$  (1.5 mg, 0.002 mmol), NaOAc (33 mg, 0.4 mmol) and PhSO<sub>2</sub>CF<sub>2</sub>I (63.6 mg, 0.2 mmol) were combined in an oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). 2-Methoxystyrene **1s** (40.2 mg, 0.3 mmol) and CHCl<sub>3</sub> (4.0 mL) were added *via* syringe. The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 40 °C with two 26 W compact fluorescent light bulbs (one on either side of

the vial approximately 5 cm away) for 24 h. The mixture was then cooled to room temperature, filtered through a plug of silica and washed with EtOAc. The filtrate was concentrated under vacuum and purified by preparatory thin layer chromatography (PE:EA = 30:1) to give product **2s** (54.2 mg, 60%) and **3s** (14.9 mg, 23%).

Ru(bpy)<sub>3</sub>Cl<sub>2</sub>·6H<sub>2</sub>O (1.5 mg, 0.002 mmol), NaOAc (33 mg, 0.4 mmol) and PhSO<sub>2</sub>CF<sub>2</sub>I (63.6 mg, 0.2 mmol) were combined in an oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). 2-Methoxystyrene **1s** (40.2 mg, 0.3 mmol) and CHCl<sub>3</sub> (4.0 mL) were added *via* syringe. The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 40 °C with two 26 W compact fluorescent light bulbs (one on either side of the vial approximately 5 cm away) for 48 h. The mixture was then cooled to room temperature, filtered through a plug of silica and washed with EtOAc. The filtrate was concentrated under vacuum and purified by preparatory thin layer chromatography (PE:EA = 30:1) to give product **2s** (40.7 mg, 45%) and **3s** (27.8 mg, 43%). *This revealed that the heck-type product can be obtained from the ATRA product in the reaction*.



**2s**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.93 (d, J = 7.6 Hz, 2H), 7.73 (dt, J = 7.6, 1.2 Hz, 1H), 7.60-7.56 (m, 2H), 7.36 (dd, J = 7.6, 1.2 Hz, 1H), 7.27-7.23 (m, 1H), 6.90 (td, J = 7.6, 0.8 Hz, 1H), 6.83 (d, J = 8.4 Hz, 1H), 5.81

(dd, J = 9.6, 5.2 Hz, 1H), 3.90 (s, 3H), 3.78-3.36 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  156.2, 135.6, 131.8, 130.9, 130.4, 129.9, 129.4, 128.0, 123.3 (t, J = 288.3 Hz), 120.8, 115.4, 55.7, 38.8 (t, J = 18.6 Hz), 11.8. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -102.9 (d, J = 227.1 Hz, 1F), -104.6 (d, J = 227.1 Hz, 1F). HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>16</sub>H<sub>15</sub>O<sub>3</sub>F<sub>2</sub>SINa: 474.9652, found: 474.9659.

#### **General Procedure for Derivatization of Product:**

**Gram-scale reaction:** 



Ru(bpy)<sub>3</sub>Cl<sub>2</sub>·6H<sub>2</sub>O (22.5 mg, 0.030 mmol), NaOAc (495 mg, 6 mmol) and PhSO<sub>2</sub>CF<sub>2</sub>I (954 mg, 3 mmol) were combined in an oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). Styrene **1a** (468 mg, 4.5 mmol) and CHCl<sub>3</sub> (60 mL) were added *via* syringe. The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 40 °C with two 26 W compact fluorescent light bulbs (one on either side of the vial approximately 5 cm away) for 24 h. The mixture was then cooled to room temperature, filtered through a plug of silica and washed with EtOAc. The filtrate was concentrated under vacuum and purified by preparatory thin layer chromatography (PE:EA = 30:1) to give product **2a** as a white solid (1.47 g, 78%).

**Reaction of 2a with NaN3:** 



**2a** (21 mg, 0.05 mmol), NaN<sub>3</sub> (10 mg, 0.15 mmol) and AlCl<sub>3</sub> (3.5 mg, 0.025 mmol) were combined in a 5 mL oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). MeCN (0.5 mL) were added *via* syringe. The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 70 °C for 16 h. The mixture was then cooled to room temperature, filtered through a plug of silica and washed with EtOAc. The filtrate was concentrated under vacuum and purified by preparatory thin layer chromatography (PE:EA = 30:1) to give product **6** as a colorless oil (12.5 mg, 74%).



**6**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.98 (d, *J* = 7.6 Hz, 2H), 7.77 (tt, *J* = 7.2, 1.2 Hz, 1H), 7.64-7.60 (m, 2H), 7.44-7.34 (m, 5H), 4.97-4.94 (m, 1H), 2.94-2.73 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 138.1, 135.7, 131.8, 131.0, 129.5, 129.3, 129.1, 126.8, 123.0 (t, J = 286.6 Hz), 59.6 (t, J = 2.4 Hz), 35.9 (t, J = 19.0 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -101.4 (d, J = 229.7 Hz), -103.3 (d, J = 229.7 Hz). HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>13</sub>N<sub>3</sub>O<sub>2</sub>F<sub>2</sub>SNa: 360.0594, found: 360.0590.

## **Reductive De-iodination of 2a:**



**2a** (21 mg, 0.05 mmol) and AIBN (24.6 mg, 0.15 mmol) were combined in a 5 mL oven-dried sealing tube. The vessel was evacuated and backfilled with N<sub>2</sub> (repeated for 3 times). Toluene (0.5 mL) and <sup>t</sup>Bu<sub>3</sub>SnH (43.6 mg, 0.15 mmol) were added *via* syringe. The tube was sealed with a Teflon lined cap and the reaction mixture was placed into a preheated oil bath at 90 °C for 3 h. The mixture was then cooled to room temperature, concentrated under vacuum and purified by preparatory thin layer chromatography (PE:EA = 30:1) to give product **7** as a colorless oil (quant yield was obtained).



7: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.00 (d, J = 7.6 Hz, 2H), 7.76 (t, J = 7.6 Hz, 1H), 7.64-7.60 (m, 2H), 7.31 (t, J = 7.6 Hz, 2H), 7.24-7.20 (m, 3H), 2.97-2.93 (m, 2H),

2.72-2.58 (m, 2H). 139.2, 135.4, 132.5, 130.9, 129.4, 128.8, 128.4, 126.8, 124.2 (t, J = 284.5 Hz), 31.3 (t, J = 20.0 Hz), 27.2 (t, J = 3.7 Hz). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -103.7. HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>15</sub>H<sub>14</sub>O<sub>2</sub>F<sub>2</sub>SNa: 319.0580, found: 319.0568.

### **Reductive Desulfonylation:**



Into a 10 mL flask containing compound **3l** (32.4 mg, 0.1 mmol) in DMF (1 mL) at room temperature was added HOAc/NaOAc (1:1) buffer solution (8 mol/L, 1 mL).

Magnesium turnings (36 mg, 1.5 mmol) were added in portions. The reaction was stirred at 50 °C until the material was consumed as determined by TLC and quenched with water. The mixture was extracted with  $Et_2O$ , and the combined organic layer was washed with water and brine, dried over  $Na_2SO_4$ , filtrated and concentrated under vacuum. The residue was purified by flash column chromatography on silica gel (PE:EA = 10:1) to give product **8** as a white solid (16.6 mg, 90%).



8: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.38 (d, J = 8.8 Hz, 2H), 6.91-6.87 (m, 2H), 6.84-6.78 (m, 1H), 6.22 (td, J =56.0, 6.0 Hz, 1H), 6.17-6.07 (m, 1H), 3.83 (s, 3H). <sup>13</sup>C

NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  160.73, 136.8 (t, J = 12.2 Hz), 128.8, 127.2, 118.8 (t, J = 23.8 Hz), 115.9 (t, J = 231.7 Hz), 114.3, 55.4. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>). HRMS ESI (m/z): [M+Na]<sup>+</sup> calcd. for C<sub>10</sub>H<sub>10</sub>OF<sub>2</sub>Na: 207.0597, found: 207.0597.

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# Copies of <sup>1</sup>H, <sup>13</sup>C and <sup>19</sup>F NMR Spectra







0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200











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190 180 **1**40 130 



S35



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