

Supporting Information

Palladium-Catalyzed Cross-Coupling of (Hetero)Aryl or Alkenyl Sulfonates with Aryl Titanium as the Multi-Functional Reagent

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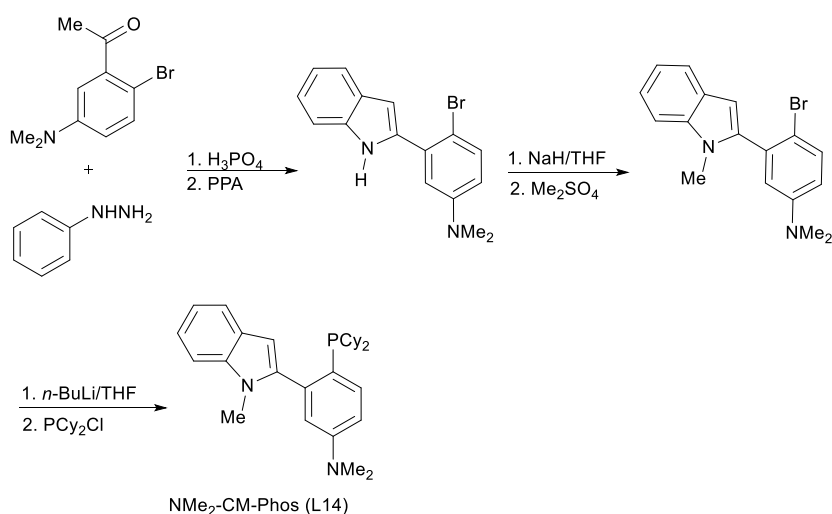
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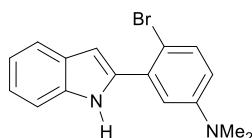
1. General considerations

Unless otherwise noted, all reagents were purchased from commercial suppliers and used without purification. All cross-coupling reactions were performed in resealable screw cap Schlenk tube (approx. 20 mL volume) in the presence of Teflon-coated magnetic stirrer bar (4.5 mm × 12 mm). Indolylphosphine ligand **L1**¹ and **L13**² were prepared according to the reported procedures. Ligands **L2-L12** were purchased from commercial suppliers. Alkenyl tosylates and mesylates were synthesized according to the reported procedures.³ Thin layer chromatography was performed on pre-coated silica gel 60 F₂₅₄ plates. Silica gel (Merck, 230-400 mesh) was used for column chromatography. Melting points were recorded on an uncorrected Büchi Melting Point B-545 instrument. NMR spectra were recorded on a Bruker spectrometer (400 MHz for ¹H, 100 MHz for ¹³C, 376 MHz for ¹⁹F and 162 MHz for ³¹P). Spectra were referenced internally to the residual proton resonance in CDCl₃ (δ 7.26 ppm) or TMS (δ 0.00 ppm) as the internal standard. Chemical shifts (δ) were reported as part per million (ppm) in δ scale downfield from TMS. ¹³C NMR spectra were referenced to CDCl₃ (δ 77.0 ppm, the middle peak). ¹⁹F NMR chemical shifts were determined relative to CFC₃ as the external standard and low field is positive. ³¹P NMR spectra were referenced to 85% H₃PO₄ externally. Coupling constants (*J*) were reported in Hertz (Hz). Mass spectra (EI-MS and ES-MS) were recorded on a HP 5989B Mass Spectrometer. High-resolution mass spectra (HRMS) were obtained on a Bruker APEX 47e FTICR mass spectrometer (ESI-MS), a Agilent 6540 Qtof MS and a Bruker UltrafleXtreme Maldi-Tof/Tof MS. GC-MS analysis was conducted on a HP 5977A GCD system using a HP5MS column (30 m × 0.25 mm). The products described in GC yield were accorded to the authentic samples/dodecane calibration standard from HP 7890B GC-FID system. All yields reported refer to isolated yield of compounds estimated to be greater than 95% purity as determined by capillary gas chromatography (GC) or ¹H NMR. Compounds described in the literature were characterized by comparison of their ¹H, ¹³C, ³¹P and/or ¹⁹F NMR spectra to the previously reported data. E factors were calculated according to the procedure reported by Lipshutz et.al.⁴ The procedures in this section are representative, and thus the yields may differ from those reported in tables.

2. Preparation of indolylphosphine ligand NMe₂-CM-Phos (L14)

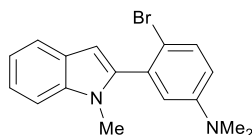


4-Bromo-3-(1H-indol-2-yl)-N,N-dimethylaniline



Fischer indole synthesis of 4-bromo-3-(1H-indol-2-yl)-N,N-dimethylaniline was conducted according to the previous reported procedure.^{1, 5} 1-(2-Bromo-5-(dimethylamino)phenyl)ethanone (5.7 mmol), phenylhydrazine (6.84 mmol), H₃PO₄ (2.0 mL) and PPA (20 g) were used to give the desired product as a light yellow powder (1.04 g, 58%). M.p. 113.0–114.9°C; ¹H NMR (400 MHz, CDCl₃) δ 3.03 (s, 6H), 6.67 (d, *J* = 8.0Hz, 1H), 6.90 (s, 1H), 6.98 (s, 1H), 7.24–7.35 (m, 2H), 7.48 (d, *J* = 8.0Hz, 1H), 7.56 (d, *J* = 8.0Hz, 1H), 7.78 (d, *J* = 8.0Hz, 1H), 8.68 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 40.4, 102.9, 107.4, 110.9, 113.6, 114.9, 119.9, 120.5, 122.2, 128.1, 133.4, 133.9, 135.9, 137.2, 149.6; MS (EI): *m/z* (relative intensity) 314 (M⁺, 100), 235 (43), 220 (43), 205 (8), 191 (39); HRMS: calcd. for C₁₆H₁₅N₂BrH⁺: 315.0497, found 315.0495.

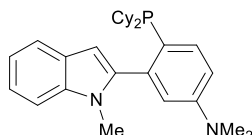
4-Bromo-N,N-dimethyl-3-(1-methyl-1H-indol-2-yl)aniline



A 100-mL three-necked round-bottomed flask equipped with dropping funnel is charged with sodium hydride (4.80 mmol, 60 % dispersion in mineral oil) which was washed free of mineral oil with hexane. The dropping funnel was charged with 4-Bromo-3-(1H-indol-2-yl) -

N,N-dimethylaniline (3.20 mmol, 1.00 g) and the setup is evacuated and backfilled with nitrogen three times. Freshly distilled THF (10 mL) was added to the dropping funnel and THF (5.0 mL) was added to the flask. The solution of 4-Bromo-3-(1*H*-indol-2-yl) *N,N*-dimethylaniline was added dropwise to the reaction flask and the reaction mixture was stirred for 15 min at room temperature. Me₂SO₄ (3.36 mmol, 0.318 mL) was then added to the mixture dropwise *via* syringe. After the mixture was stirred at room temperature for overnight, solvent was removed by high vacuum. Ethyl acetate and water was added to the mixture and the organic layer was separated. The organic layer was washed with water and brine for three times and concentrated. The residue was subjected to flash column chromatography on silica gel (230-400 mesh) to give the product as a white powder (0.84 g, 80%). M.p. 103–105°C; ¹H NMR (400 MHz, CDCl₃) δ 2.97 (s, 6H), 3.62 (s, 3H), 6.53 (s, 1H), 6.68 (d, *J* = 7.0Hz, 1H), 6.75 (s, 1H), 7.16 (t, *J* = 6.7Hz, 1H), 7.27 (t, *J* = 7.0Hz, 1H), 7.38 (d, *J* = 7.8Hz, 1H), 7.50 (d, *J* = 8.7Hz, 1H), 7.67 (d, *J* = 7.4Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 30.6, 40.4, 101.4, 109.4, 110.7, 114.0, 116.5, 119.6, 120.6, 121.5, 127.6, 132.7, 134.2, 137.1, 140.7, 143.4; MS (EI): *m/z* (relative intensity) 328 (M⁺, 100), 249 (42), 233 (31), 204 (44); HRMS: calcd. for C₁₇H₁₇N₂BrH⁺: 329.0653, found 329.0650.

4-(Dicyclohexylphosphino)-*N,N*-dimethyl-3-(1-methyl-1*H*-indol-2-yl)aniline (NMe₂-CM-Phos, L14)



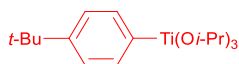
4-Bromo-*N,N*-dimethyl-3-(1-methyl-1*H*-indol-2-yl)aniline (1.42 mmol, 0.467 g) was dissolved in freshly distilled THF (10 mL) at room temperature under nitrogen atmosphere. The solution was cooled to –78 °C in dry ice/acetone bath. Titrated *n*-BuLi (1.56 mmol) was added dropwise by syringe. After the reaction mixture was stirred for 30 min at –78 °C, chlorodicyclohexylphosphine (1.56 mmol, 0.344 mL) in THF (3.0 mL) was added. The reaction was allowed to warm to room temperature and stirred overnight. Solvent was removed under high vacuum. The residue was successively washed with cold MeOH and then dried under high vacuum. 4-(Dicyclohexylphosphino)-*N,N*-dimethyl-3-(1-methyl-1*H*-indol-2-yl)aniline was obtained as a white solid (0.44 g, 69%). M.p. 150.6–151.9°C; ¹H NMR (400 MHz, CD₂Cl₂) δ 1.07–1.26 (m, 10H), 1.58–2.10 (m, 12 H), 3.02 (s, 6H), 3.53 (s, 3H), 6.38 (s, 1H), 6.74 (s, 1H), 6.71 (t, *J* = 2.8Hz, 1H), 6.86–6.89 (m, 1H), 7.13 (t, *J* = 7.2Hz, 1H), 7.23 (t, *J* = 7.3Hz, 1H), 7.37 (d, *J* = 8.1Hz, 1H), 7.49 (d, *J* = 8.6Hz, 1H), 7.62 (d, *J* = 7.8Hz, 1H); ¹³C

NMR (100 MHz, CD₂Cl₂) δ 26.5, 27.2, 30.60, 30.66, 39.8, 102.3, 102.4, 109.1, 112.0, 114.90, 114.97, 119.1, 119.8, 120.6, 121.5, 121.69, 127.7, 133.61, 133.65, 136.4, 141.5, 141.9, 142.6, 142.7, 150.0; ³¹P NMR (162 MHz, CD₂Cl₂) δ -12.97; MS (EI): *m/z* (relative intensity) 446 (M⁺, 47), 431 (9), 363 (100), 281 (96), 265 (18); HRMS: calcd. for C₂₉H₃₉N₂PH⁺: 447.2929, found 447.2913.

3. Preparation of aryl titanium reagents

All aryl titanium reagents were synthesized according or analogous to the reported procedures.⁶ A general procedure for the preparation of ArTi(O*i*-Pr)₃: A two-necked 250 mL round-bottomed flask equipped with a condenser, a magnetic stir bar, and an addition funnel was charged with magnesium turnings (2.40 g, 100 mmol), and the flask was evacuated under reduced pressure for 2 h. Under a nitrogen atmosphere, THF (100 mL) was added to the flask and aryl bromide (120 mmol) in THF (50 mL) was transferred into the addition funnel. The THF solution of aryl bromide was added slowly to the reaction flask, and the reaction mixture was controlled under gentle reflux using an ice bath if necessary. After the reaction was complete, the resulting Grignard reagent was cooled to 0 °C. In another two-necked 500 mL round bottomed flask under a nitrogen atmosphere, to a solution of Ti(O*i*-Pr)₄ (22.4 mL, 75.0 mmol) in 50 mL of THF at 0°C was added TiCl₄ (2.8 mL, 25.0 mmol). The resulting solution was warmed to room temperature and stirred for 30 min, giving a ClTi(O*i*-Pr)₃ solution (100 mmol). The ClTi(O*i*-Pr)₃ solution was cooled to 0 °C, and to this solution was transferred the ice cold Grignard solution via a cannula. The reaction mixture was warmed to room temperature and was allowed to react for 3 h. The volatile material was removed completely under reduced pressure, and under a nitrogen atmosphere, the residue was extracted with *n*-hexane (3 x 200 mL). The combined hexane solution was concentrated and was cooled to -20 °C, furnishing a crystalline product of the corresponding ArTi(O*i*-Pr)₃. The characterization data of new 4-*tert*-butylphenyltitanium triisopropoxide is showed below.

4-*tert*-Butylphenyltitanium triisopropoxide

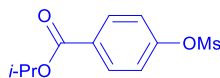


Obtained in 45% as pale yellow crystal; m.p. 74.2-77.0°C; ¹H NMR (400 MHz, CDCl₃) δ 1.36 (s, 9H), 1.37–1.38 (m, 18H), 4.73–4.77 (m, 3H), 7.23 (d, *J* = 6.8Hz, 2H), 7.61 (d, *J* = 7.1Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 26.3, 31.3, 34.5, 77.3, 123.0, 133.4, 149.7, 182.3; HRMS calcd for C₁₉H₃₄O₃Ti⁺: 358.1988, found 358.1972.

4. Preparation of aryl mesylates and tosylates substrates

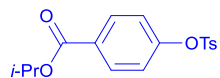
Aryl mesylates and tosylates were prepared according to the literature procedures.⁷ A general procedure for the preparation of aryl sulfonates: To a stirred solution of corresponding phenol (10 mmol) in anhydrous dichloromethane (20 mL) cooled to 0 °C was added distilled triethylamine (6.95 mL, 50 mmol). To this was added mesyl chloride (1.94 mL, 15 mmol) dropwise via syringe over 5 min or was added tosyl chloride (2.86 g, 15 mmol) as powder form. The reaction was stirred at room temperature. The reaction was quenched with water and the layer separated when the reaction was completed. The aqueous layer was extracted with dichloromethane and the combined organic solution was dried over Na₂SO₄ and concentrated. The residue was purified by column chromatography on silica gel using EA/hexane solvent mixtures as the eluent. The organic solution was concentrated and the product was dried to afford the corresponding aryl sulfonates. The characterization data of new isopropyl 4-(mesyloxy)benzoate and isopropyl 4-(tosyloxy)benzoate are showed below.

Isopropyl 4-(mesyloxy)benzoate



Obtained in 82% as white solid; m.p. 49.8-50.6°C; ¹H NMR (400 MHz, CDCl₃) δ 1.35 (d, *J* = 6.2Hz, 6H), 3.16 (s, 3H), 5.24 (sept, *J* = 6.2Hz, 1H), 7.33 (d, *J* = 8.1Hz, 2H), 8.09 (d, *J* = 8.4Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 21.8, 37.7, 68.8, 121.7, 129.9, 131.5, 152.2, 164.7; MS (EI): *m/z* (relative intensity) 258.3 (M⁺, 23), 216.2 (53), 199.2 (100), 138.2 (93), 121.2 (73), 92.2 (20), 59.2 (27); HRMS calcd for C₁₁H₁₅O₅S⁺: 259.0635, found 259.0632.

Isopropyl 4-(tosyloxy)benzoate



Obtained in 93% as white solid; m.p. 42.7-44.8°C; ¹H NMR (400 MHz, CDCl₃) δ 1.35 (d, *J* = 6.2Hz, 6H), 2.44 (s, 3H), 5.21 (sept, *J* = 6.2Hz, 1H), 7.04 (d, *J* = 8.7Hz, 2H), 7.30 (d, *J* = 8.1Hz, 2H), 7.69 (d, *J* = 8.2Hz, 2H), 7.96 (d, *J* = 8.7Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 21.6, 21.8, 68.7, 122.1, 128.4, 129.6, 129.8, 131.1, 132.0, 145.6, 152.7, 164.9; MS (EI): *m/z* (relative intensity) 334.4 (M⁺, 27), 292.3 (16), 275.3 (20), 207.3 (9), 155.2 (100), 120.2 (32), 91.2 (90), 65.2 (14); HRMS calcd for C₁₇H₁₉O₅S⁺: 335.0948, found 335.0955.

5. General procedures for palladium-catalyzed cross-coupling reaction of aryl mesylate with different nucleophiles

Pd(OAc)₂ (4.5 mg, 0.020 mmol) and L1 (32.2 mg, 0.080 mmol) were loaded into Schlenk tube with a Teflon-coated magnetic stirrer bar (4.5 mm x 12 mm). The tube was evacuated and backfilled with nitrogen (3 cycles). Precomplexation was applied by adding freshly distilled DCM (1.0 mL) and Et₃N (0.10 mL) into the tube. The solution was stirred and warmed using oil bath (~75 °C) for about 1 min until the solvent started boiling. The solvent in tube was removed under high vacuum. *For cross-coupling reaction using lithium, magnesium and zinc nucleophile*, PhLi (1.25 mL, 2.50 mmol, 2.0 M in dibutyl ether) or *p*-tolylMgBr (2.50 mL, 2.50 mmol, 1.0 M in THF) or *p*-tolylZnCl (2.50 mmol generated from 2.50 mmol *p*-tolylMgBr in 1.0 M THF and 2.50 mmol ZnCl₂ in 0.5 M THF) was added into the tube via syringe. The solvent in tube was removed under high vacuum. 4-*tert*-Butylphenyl mesylate (0.114 g, 0.50 mmol) was loaded into the Schlenk tube which was again evacuated and re-filled with nitrogen for three times. *For cross-coupling reaction using boron, silane, stannane and titanium nucleophile*, 4-*tert*-butylphenyl mesylate (0.114 g, 0.50 mmol) and *p*-tolylB(OH)₂ (0.339 g, 2.5 mmol) or *p*-tolylBpin (0.545 g, 2.5 mmol), *p*-tolylBF₃K (0.495 g, 2.5 mmol) or *p*-tolylSi(OMe)₃ (513 μL, 2.5 mmol) or *p*-tolylSnBu₃ (0.953 g, 2.5 mmol) or *p*-tolylTi(Oi-Pr)₃ (0.790 g, 2.5 mmol) were loaded into the Schlenk tube which was again evacuated and re-filled with nitrogen for three times. The Schlenk tube was resealed and magnetically stirred in a preheated 110 °C oil bath for 3 h. The reaction tube was allowed to reach room temperature. Water, ethyl acetate and dodecane (113 μL, internal standard) were added to the mixture. The organic solution was subjected to GC analysis. The GC yield was previously calibrated by authentic sample/dodecane calibration curve.

6. General procedure for initial ligand and reaction conditions screening

Pd(OAc)₂ (4.5 mg, 0.020 mmol) and ligand (Pd:L = 1:4) were loaded into Schlenk tube with a Teflon-coated magnetic stirrer bar (4.5 mm x 12 mm). The tube was evacuated and backfilled with nitrogen (3 cycles). Precomplexation was applied by adding freshly distilled DCM (1.0 mL) and Et₃N (0.10 mL) into the tube. The solution was stirred and warmed using oil bath (~75 °C) for about 1 min until the solvent started boiling. The solvent in tube was removed under high vacuum. 4-*tert*-Butylphenyl mesylate **1a** (0.114 g, 0.50 mmol) and *p*-tolylTi(*Oi*-Pr)₃ **2a** (the equivalence was indicated in Table 2) were loaded into the Schlenk tube which was again evacuated and re-filled with nitrogen for three times. The Schlenk tube was resealed and magnetically stirred in a preheated 110 °C oil bath for the time as indicated in Table 2. The reaction tube was allowed to reach room temperature. Water, ethyl acetate and dodecane (113 μL, internal standard) were added to the mixture. The organic solution was subjected to GC analysis. The GC yield was previously calibrated by authentic sample/dodecane calibration curve.

7. General procedures for palladium-catalyzed cross-coupling reaction of aryl/heteroaryl and alkenyl mesylates and tosylates with aryl titanium reagents

General procedure for cross-coupling reaction of aryl/heteroaryl mesylates and tosylates with aryl titanium reagents (Pd catalyst loading equal to or large than 2.0 mol%): Pd(OAc)₂ (Pd loading indicated in Table 3) and NMe₂-CM-Phos **L14** (Pd:L =1:4) were loaded in a Schlenk tube with a Teflon-coated magnetic stirrer bar (4.5 mm x 12 mm). The tube was evacuated and backfilled with nitrogen (3 cycles). Precomplexation was applied by adding freshly distilled DCM (1.0 mL) and Et₃N (0.10 mL) into the tube. The solution was stirred and warmed using oil bath (~75 °C) for about 1 min until the solvent started boiling. The solvent in the tube was removed under high vacuum. Aryl/heteroaryl mesylates or tosylates (0.50 mmol) and aryl titanium reagents (the equivalence was indicated in Table 3) were loaded into the tube which was again evacuated and flushed with nitrogen for three times. The tube was resealed and magnetically stirred in a preheated 110 °C oil bath for the time as indicated in Table 3. The tube was cooled and the reaction was quenched by addition of water and ethyl acetate. The organic layer was separated and the aqueous layer was washed with ethyl acetate. The combined organic solution was concentrated on a rotary evaporator. The residue was subjected to flash column chromatography on silica gel (230-400 mesh) to give the products.

General procedure for cross-coupling reaction of alkenyl mesylates and tosylates with aryl titanium reagents (Pd catalyst loading less than 0.50 mol%): Preparation of stock solution: Pd(OAc)₂ (5.6 mg, 0.025 mmol) with NMe₂-CM-Phos **L14** (44.6 mg, 0.010 mmol) were loaded into a tube with magnetic stirrer bar (4.5 mm x 12 mm). The tube was evacuated and backfilled with nitrogen (3 cycles). A freshly distilled DCM (10.0 mL) was then added to give the palladium stock solution (0.50 mol% Pd per 1 mL stock solution). A Schlenk tube with a Teflon-coated magnetic stirrer bar (4.5 mm x 12 mm) was evacuated and backfilled with nitrogen (3 cycles). The corresponding volume of the palladium stock solution was transferred to the tube. Precomplexation was applied by adding freshly distilled Et₃N (0.10 mL) into the tube. The solution was stirred and warmed using oil bath (~75 °C) for about 1 minute until the solvent started boiling. The solvent was removed under high vacuum. Alkenyl mesylates or tosylates (0.50 mmol) and aryl titanium reagents (the equivalence was indicated in Table 4) were loaded into the tube which was again evacuated and flushed with nitrogen for three times. The tube was resealed and magnetically stirred in a preheated 110

°C oil bath for the time as indicated in Table 4. The tube was cooled and the reaction was quenched by addition of water and ethyl acetate. The organic layer was separated and the aqueous layer was washed with ethyl acetate. The combined organic solution was concentrated on a rotary evaporator. The residue was subjected to flash column chromatography on silica gel (230-400 mesh) to give the products.

8. General procedure for a set of parallel experiments between Suzuki-type organoborn and organotitanium nucleophilles

A stock solution of Pd(OAc)₂ (2.3 mg, 0.010 mmol) with NMe₂-CM-Phos **L14** (17.8 mg, 0.04 mmol) in freshly distilled 10 mL DCM (0.2 mol% Pd per 1.0 mL stock solution) was initially prepared with continuously stirring at room temperature. An array of Schlenk tubes equipped with a Teflon-coated magnetic stir bar (4.5 mm x 12 mm) were evacuated and backfilled with nitrogen (3 cycles). The stock solutions (1.00 mL) and Et₃N (0.10 mL) were transferred accordingly to the each of the tubes *via* syringes. The solution was stirred and warmed using oil bath (~75 °C) for about 1 minute until the solvent started boiling. The solvent was then evaporated under high vacuum.

For the cross-coupling reaction using organotitanium nucleophile: 3,4-dihydronaphthalen-2-yl tosylate **4a** (0.150 g, 0.50 mmol) and *p*-tolylTi(Oi-Pr)₃ **2a** (0.316 g, 1.00 mmol) were loaded into the tubes that were further evacuated and flushed with nitrogen for three times.

For the cross-coupling reaction using organoboron nucleophile: 4-dihydronaphthalen-2-yl tosylate **4a** (0.150 g, 0.50 mmol), *p*-tolylB(OH)₂ (0.123 g, 1.0 mmol) and K₃PO₄•H₂O (0.345 g, 1.5 mmol) were loaded into the tubes which were further evacuated and flushed with nitrogen for three times. *t*-BuOH (1.5 mL) was then added via syringe. All the tubes were resealed and magnetically stirred in a preheated 110 °C oil bath for 10 min. The tubes were cooled and the reactions were quenched by addition of water and ethyl acetate. Dodecane (113 μL, internal standard) was then added into the tubes. The organic solutions were subjected to GC analysis. The GC yield was previously calibrated by authentic sample/dodecane calibration curve.

9. General procedures and data for palladium-catalyzed cross-coupling reaction of aryl mesylate and alkenyl tosylate using *in situ* generated aryl titanium reagent

The procedure for cross-coupling reaction of alkenyl tosylate with in situ preparation aryl titanium reagent:

Preparation of stock solution: Pd(OAc)₂ (2.3 mg, 0.010 mmol) with NMe₂-CM-Phos **L14** (17.8 mg, 0.040 mmol) were loaded into a tube with magnetic stirrer bar (4.5 mm x 12 mm). The tube was evacuated and backfilled with nitrogen (3 cycles). Precomplexation was applied by adding freshly distilled DCM (1.0 mL) and Et₃N (0.10 mL) into the tube. The solution was stirred and warmed using oil bath (~75 °C) for about 1 minute until the solvent started boiling. The solvent was removed under high vacuum. A freshly distilled THF (10.0 mL) was then added to give the palladium stock solution (0.2 mol% Pd per 1 mL stock solution).

A Schlenk tube equipped with a Teflon-coated magnetic stir bar (4.5 mm x 12 mm) was evacuated and backfilled with nitrogen (3 cycles). ClTi(Oi-Pr)₃ (0.24 mL, 1.00 mmol) and PhLi (0.50 mL, 1.00 mmol, 2.0 M in dibutyl ether) were added successively to the tube at 0 °C and the mixture was then allowed to stand at room temperature and stirred for 30 min. The solvent was then removed under high vacuum. The palladium stock solution (1.0 mL) was transferred to the tube and the solvent was then removed under high vacuum. 3,4-Dihydronaphthalen-2-yl tosylate **4a** (0.15 g, 0.50 mmol) was loaded into the tube which was resealed and magnetically stirred in a preheated 110 °C oil bath for 10 min. The tube was cooled and the reaction was quenched by addition of water and ethyl acetate. The organic layer was separated and the aqueous layer was washed with ethyl acetate. The combined organic solution was concentrated on a rotary evaporator. The residue was subjected to flash column chromatography on silica gel (230-400 mesh) to give the products.

The procedure for cross-coupling reaction of aryl mesylate with in situ preparation aryl titanium reagent:

Preparation of stock solution: Pd(OAc)₂ (5.6 mg, 0.025 mmol) with NMe₂-CM-Phos **L14** (44.6 mg, 0.10 mmol) were loaded into a tube with magnetic stirrer bar (4.5 mm x 12 mm). The tube was evacuated and backfilled with nitrogen (3 cycles). Precomplexation was applied by adding freshly distilled DCM (1.0 mL) and Et₃N (0.10 mL) into the tube. The palladium complex solution was stirred and warmed using oil bath (~75 °C) for about 1 minute

until the solvent started boiling. The solvent was then evaporated under high vacuum. A freshly distilled THF (1.0 mL) was then added to give the palladium stock solution (5.0 mol% Pd per 1 mL stock solution).

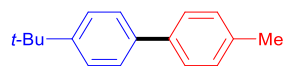
A Schlenk tube equipped with a Teflon-coated magnetic stir bar (4.5 mm x 12 mm) was evacuated and backfilled with nitrogen (3 cycles). $\text{ClTi}(\text{O}i\text{-Pr})_3$ (0.60 mL, 2.50 mmol) and PhLi (1.25 mL, 2.50 mmol, 2.0 M in dibutyl ether) were added successively to the tube at 0 °C and the mixture was then allowed to stand at room temperature and stirred for 30 min. The solvent was then removed under high vacuum. The palladium stock solution (0.80 mL) was transferred to the tube and the solvent was then removed under high vacuum. Sesamol mesylate **1g** (0.108 g, 0.50 mmol) was loaded into the tube which was resealed and magnetically stirred in a preheated 110 °C oil bath for 18 h. The tube was cooled and the reaction was quenched by addition of water and ethyl acetate. The organic layer was separated and the aqueous layer was washed with ethyl acetate. The combined organic solution was concentrated on a rotary evaporator. The residue was subjected to flash column chromatography on silica gel (230-400 mesh) to give the products.

10. General procedure for large-scale cross-coupling reaction of alkenyl tosylate with aryl titanium reagent

Pd(OAc)₂ (4.5 mg, 0.020 mmol) and NMe₂-CM-Phos **L14** (35.6 mg, 0.080 mmol) were loaded to a two necked 100 mL round bottom flask which equipped with an air condenser, magnetic stirrer bar (25 mm x 6 mm) and fitted with septum. The flask was carefully evacuated and backfilled with nitrogen (3 cycles). Precomplexation was accomplished by adding freshly distilled DCM (1.0 mL) and Et₃N (0.1 mL) into the flask and the solution was stirred and warmed using oil bath (~75 °C) until the solvent started boiling. The solvent was removed under high vacuum. 3,4-Dihydronaphthalen-2-yl tosylate **4a** (3.00 g, 10.0 mmol) and *m*-anisylTi(O*i*-Pr)₃ **2c** (6.64 g, 20.0 mmol) were charged to the flask and the septum was replaced with a glass stopper. The reaction flask was placed in a preheated 110 °C oil bath for 10 min. The flask was cooled to room temperature. Water was added to quench the reaction. Ethyl acetate was then added to the flask and the mixture was stirred for 5 min. The mixture was filtered through a short pad of sodium sulphate which was then eluted with ethyl acetate. The filtrate was then transferred to separating funnel and the organic layer was washed with water for two times. The organic solution was concentrated on a rotary evaporator. The residue was subjected to flash column chromatography on silica gel (230-400 mesh) to give the 3-(3-methoxyphenyl)-1,2-dihydronaphthalene (2.24 g, 95%).

11. Characterization data for coupling products

4-(*tert*-Butyl)-4'-methyl-1,1'-biphenyl (Table 3, **3aa**)⁸



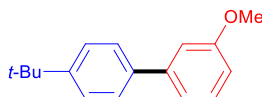
Eluents (Hexane, $R_f = 0.5$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 1.40 (s, 9H), 2.43 (s, 3H), 7.28 (d, $J = 6.6\text{Hz}$, 2H), 7.48-7.57 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 21.0, 31.3, 34.3, 125.6, 126.5, 126.8, 129.4, 136.6, 138.1, 138.2, 149.9; MS (EI): m/z (relative intensity) 224.4 (M^+ , 45), 209.4 (100), 181.3 (20), 165.3 (15), 90.6 (9).

4-(*tert*-Butyl)-4'-methoxy-1,1'-biphenyl (Table 3, **3ab**)⁹



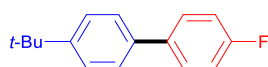
Eluents (Hexane : Dichloromethane = 20:1, $R_f = 0.3$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 1.40 (s, 9H), 3.87 (s, 3H), 7.00 (d, $J = 8.6\text{Hz}$, 2H), 7.48 (d, $J = 8.3\text{Hz}$, 2H), 7.55-7.57 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 31.3, 34.4, 55.2, 114.1, 125.6, 126.3, 127.9, 133.6, 137.9, 149.6, 158.9; MS (EI): m/z (relative intensity) 240.4 (M^+ , 47), 225.4 (100), 197.3 (14), 98.7 (14).

4-(*tert*-Butyl)-3'-methoxy-1,1'-biphenyl (Table 3, **3ac** and **3hf**)¹⁰



Eluents (Hexane : Dichloromethane = 20:1, $R_f = 0.30$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 1.39 (s, 9H), 3.88 (s, 3H), 6.90 (d, 8.4Hz, 1H), 7.15 (s, 1H), 7.21 (d, $J = 7.6\text{Hz}$, 1H), 7.36 (t, $J = 8.0\text{Hz}$, 1H), 7.48 (d, $J = 8.4\text{Hz}$, 2H), 7.56 (d, $J = 8.4\text{Hz}$, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 31.3, 34.5, 55.2, 112.4, 112.7, 119.5, 125.6, 126.8, 129.6, 138.2, 142.6, 150.4, 159.9; MS (EI): m/z (relative intensity) 240.4 (M^+ , 38), 225.4 (100), 197.3 (15), 98.7 (15).

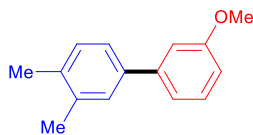
4-(*tert*-Butyl)-4'-fluoro-1,1'-biphenyl (Table 3, **3ad** and **3if**)¹¹



Eluents (Hexane, $R_f = 0.50$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 1.43 (s, 9H), 7.16 (t, $J = 8.8\text{Hz}$, 2H), 7.54 (d, 8.4Hz, 4H), 7.57-7.61 (m, 2H); ^{13}C

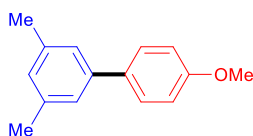
NMR (100 MHz, CDCl₃) δ 31.3, 34.4, 115.4 (d, $J = 21\text{Hz}$), 125.7, 126.6, 128.4 (d, $J = 8\text{Hz}$), 137.1 (d, $J = 3\text{Hz}$), 137.3, 150.2, 162.2 (d, $J = 244\text{Hz}$); ¹⁹F NMR (376 MHz, CDCl₃) δ -116.0; MS (EI): m/z (relative intensity) 228.4 (M⁺, 33), 213.3 (100), 195.3 (25), 92.7 (15).

3,4-Dimethyl-3'-methoxy-1,1'-biphenyl (Table 3, **3bc**)



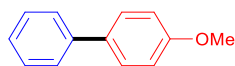
Eluents (Hexane : Dichloromethane = 9:1, $R_f = 0.50$) was used for flash column chromatography. Colourless liquid; ¹H NMR (400 MHz, CDCl₃) δ 2.38 (s, 3H), 2.40 (s, 3H), 3.92 (s, 3H), 6.94 (d, $J = 8.0\text{Hz}$, 1H), 7.20 (m, 1H), 7.26 (m, 2H), 7.40 (m, 2H), 7.45 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 19.3, 19.8, 55.1, 112.2, 112.6, 119.4, 124.4, 128.3, 129.5, 129.9, 135.8, 136.8, 138.6, 142.7, 159.8; MS (EI): m/z (relative intensity) 212.4 (M⁺, 100), 197.3 (32), 165.3 (13), 153.2 (13), 128.2 (6); HRMS calcd for C₁₅H₁₆O: 212.1196, found 212.1194.

4-Methoxy-3',5'-dimethyl-1,1'-biphenyl (Table 3, **3cb**)¹²

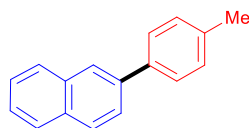


Eluents (Hexane : Dichloromethane = 20:1, $R_f = 0.50$) was used for flash column chromatography. ¹H NMR (400 MHz, CDCl₃) δ 2.38 (s, 6H), 3.85 (s, 3H), 6.97 (m, 3H), 7.18 (m, 2H), 7.52 (d, $J = 8.7\text{Hz}$, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 21.4, 55.2, 114.0, 124.6, 128.1, 128.2, 133.9, 138.1, 140.7, 158.9; MS (EI): m/z (relative intensity) 212.1 (M⁺, 100), 197.1 (45), 169.1 (18), 153.1 (9), 128.0 (5).

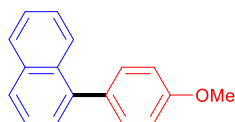
4-Methoxy-1,1'-biphenyl (Table 3, **3db**)¹⁰



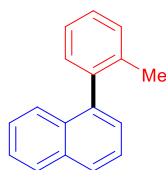
Eluents (Hexane : Dichloromethane = 20:1, $R_f = 0.40$) was used for flash column chromatography. ¹H NMR (400 MHz, CDCl₃) δ 3.88 (s, 3H), 7.02 (t, $J = 8.8\text{Hz}$, 2H), 7.34 (t, $J = 7.3\text{Hz}$, 1H), 7.46 (t, $J = 7.4\text{Hz}$, 2H), 7.59 (m, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 55.2, 114.1, 126.61, 126.69, 128.1, 128.6, 133.7, 140.7, 159.1; MS (EI): m/z (relative intensity) 184.1 (M⁺, 100), 169.1 (40), 141.1 (27), 115.1 (20), 76.1 (4).

2-(4-Methylphenyl)-naphthalene (Table 3, **3ea**)¹³

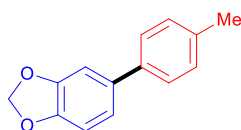
Eluents (Hexane, $R_f = 0.50$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 2.53 (s, 3H), 7.39 (d, $J = 7.9\text{Hz}$, 2H), 7.55–7.61 (m, 2H), 7.74 (d, $J = 8.1\text{Hz}$, 2H), 7.84 (d, $J = 8.5\text{Hz}$, 1H), 7.94–8.00 (m, 3H), 8.14 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 21.0, 125.3, 125.5, 125.7, 126.1, 127.2, 127.5, 128.1, 128.3, 129.5, 132.4, 133.7, 137.0, 138.1, 138.4; MS (EI): m/z (relative intensity) 218.4 (M^+ , 100), 202.3 (25), 107.9 (6), 73.1 (6).

1-(4-Methoxyphenyl)-naphthalene (Table 3, **3fb**)¹⁴

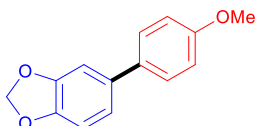
Eluents (Hexane : Dichloromethane = 20:1, $R_f = 0.30$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 3.94 (s, 3H), 7.11 (d, $J = 8.6\text{Hz}$, 2H), 7.48–7.50 (m, 4H), 7.54–7.60 (m, 2H), 7.91 (d, $J = 8.1\text{Hz}$, 1H), 7.97 (d, $J = 7.9\text{Hz}$, 1H), 8.03 (d, $J = 8.3\text{Hz}$, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 55.2, 113.6, 125.3, 125.6, 125.8, 126.0, 126.8, 127.2, 128.2, 131.0, 131.8, 133.0, 133.8, 139.8, 158.9; MS (EI): m/z (relative intensity) 234.3 (M^+ , 100), 219.3 (32), 189.3 (40), 94.7 (8).

1-(*o*-Tolyl)naphthalene (Table 3, **3fe**)¹⁵

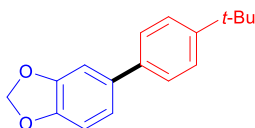
Eluents (Hexane, $R_f = 0.5$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 2.03 (s, 3H), 7.24–7.26 (m, 1H), 7.28–7.40 (m, 5H), 7.45–7.55 (m, 3H), 7.87 (d, $J = 8.3\text{Hz}$, 1H), 7.90 (d, $J = 8.1\text{Hz}$, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 20.0, 125.3, 125.5, 125.6, 125.9, 126.1, 126.6, 127.4, 127.5, 128.2, 129.8, 130.4, 132.0, 133.5, 136.8, 139.8, 140.2; MS (EI): m/z (relative intensity) 218.3 (M^+ , 100), 203.3 (53), 189.3 (8), 108.1 (12), 94.8 (7).

5-(4-Methylphenyl)benzo[d][1,3]dioxole (Table 3, **3ga**)¹⁶

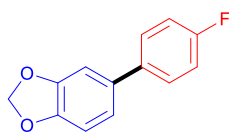
Eluents (Hexane : Dichloromethane = 20:1, $R_f = 0.30$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 2.44 (s, 3H), 6.02 (s, 2H), 6.93 (d, $J = 8.0\text{Hz}$, 1H), 7.09–7.13 (m, 2H), 7.27 (d, $J = 7.8\text{Hz}$, 2H), 7.48 (d, $J = 8.1\text{Hz}$, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 20.9, 100.9, 107.4, 108.4, 120.2, 126.6, 129.3, 135.5, 136.5, 138.0, 146.7, 148.0; MS (EI): m/z (relative intensity) 212.1 (M^+ , 100), 153.1 (19), 139.1 (13), 105.1 (6), 76.1 (5).

5-(4-Methoxyphenyl)benzo[d][1,3]dioxole (Table 3, **3gb**)¹⁷

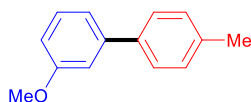
Eluents (Hexane : Dichloromethane = 7:3, $R_f = 0.50$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 3.84 (s, 3H), 5.98 (s, 2H), 6.86 (d, $J = 7.9\text{Hz}$, 1H), 6.95 (d, $J = 8.4\text{Hz}$, 2H), 7.00–7.03 (m, 2H), 7.44 (d, $J = 8.6\text{Hz}$, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 55.3, 101.0, 107.3, 108.4, 114.1, 120.0, 127.8, 133.5, 135.2, 146.5, 148.0, 158.8; MS (EI): m/z (relative intensity) 228.3 (M^+ , 100), 213.3 (63), 185.2 (15), 127.2 (13), 114.1 (6).

5-(4-*tert*-Butylphenyl)benzo[d][1,3]dioxole (Table 3, **3gf**)

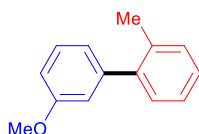
Eluents (Hexane : Dichloromethane = 9:1, $R_f = 0.50$) was used for flash column chromatography. Colourless oily paste; ^1H NMR (400 MHz, CDCl_3) δ 1.42 (s, 9H), 6.02 (s, 2H), 6.93 (d, $J = 8.0\text{Hz}$, 1H), 7.11–7.14 (m, 2H), 7.48–7.54 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 31.3, 34.4, 101.0, 107.5, 108.4, 120.3, 125.6, 126.4, 135.4, 138.0, 146.8, 148.0, 149.8; MS (EI): m/z (relative intensity) 254.4 (M^+ , 60), 239.4 (100), 211.3 (14), 181.3 (14), 165.2 (12), 105.3 (17); HRMS calcd. for $\text{C}_{17}\text{H}_{18}\text{O}_2$: 254.1307, found 254.1305.

5-(4-Fluorophenyl)benzo[d][1,3]dioxole (Table 3, **3gd**)¹⁰

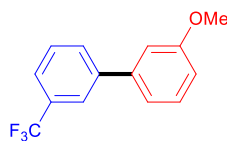
Eluents (Hexane : Dichloromethane = 9:1, $R_f = 0.50$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 6.00 (s, 2H), 6.88 (d, $J = 7.8\text{Hz}$, 1H), 7.01 (d, $J = 8.2\text{Hz}$, 2H), 7.11 (t, $J = 8.6\text{Hz}$, 2H), 7.45–7.47 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 101.1, 107.4, 108.4, 115.4 (d, $J = 21\text{Hz}$), 120.3, 128.2 (d, $J = 8\text{Hz}$), 134.5, 137.0 (d, $J = 3\text{Hz}$), 146.9, 148.0, 162.1 (d, $J = 244\text{Hz}$); ^{19}F NMR (376 MHz, CDCl_3) δ -116.1; MS (EI): m/z (relative intensity) 216.2 (M^+ , 100), 157.2 (27), 107.8 (11), 69.2 (5).

3-Methoxy-4'-methyl-1,1'-biphenyl (Table 3, **3ha**)¹⁶

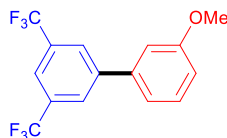
Eluents (Hexane : Dichloromethane = 20:1, $R_f = 0.40$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 2.56 (s, 3H), 4.01 (s, 3H), 7.05 (d, $J = 8.0\text{ Hz}$, 1H), 7.30 (s, 1H), 7.35 (d, $J = 7.6\text{Hz}$, 1H), 7.41 (d, $J = 8.0\text{Hz}$, 2H), 7.51 (t, $J = 7.6\text{Hz}$, 1H), 7.67. (d, $J = 7.6\text{Hz}$, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 21.0, 55.1, 112.3, 112.6, 119.4, 126.7, 126.9, 129.4, 129.6, 137.1, 138.1, 142.6; MS (EI): m/z (relative intensity) 198.3 (M^+ , 100), 167.3 (17), 155.3 (13), 115.2 (6).

3-Methoxy-2'-methyl-1,1'-biphenyl (Table 3, **3he**)¹⁸

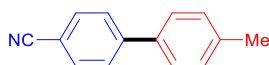
Eluents (Hexane : Dichloromethane = 9:1, $R_f = 0.50$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 2.37 (s, 3H), 3.89 (s, 3H), 6.96–7.00 (m, 3H), 7.32 (m, 4H), 7.40 (t, $J = 8.4\text{Hz}$, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 20.3, 55.1, 112.2, 114.7, 121.6, 125.6, 127.2, 128.9, 129.5, 130.2, 135.2, 141.7, 143.3, 159.2; MS (EI): m/z (relative intensity) 198.3 (M^+ , 100), 183.3 (18), 167.3 (64), 153.2 (25), 128.2 (14), 115.2 (13).

3-Methoxy-3'-(trifluoromethyl)-1,1'-biphenyl (Table 3, **3jc**)¹⁹

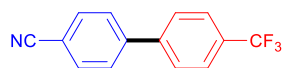
Eluents (Hexane : Dichloromethane = 9:1, $R_f = 0.30$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 3.89 (s, 3H), 6.97 (d, $J = 8.2\text{Hz}$, 1H), 7.15 (s, 1H), 7.20 (d, $J = 7.6\text{Hz}$, 1H), 7.43 (t, $J = 7.9\text{Hz}$, 1H), 7.88 (t, $J = 7.6\text{Hz}$, 1H), 7.63 (d, $J = 7.6\text{Hz}$, 1H), 7.77 (d, $J = 7.6\text{Hz}$, 1H), 7.86 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 55.4, 113.0, 113.3, 119.6, 123.8, 124.0 (m), 124.2 (q, $J = 270\text{Hz}$), 129.1, 130.0, 130.4, 131.1 (q, $J = 34\text{Hz}$), 141.2, 141.8, 160.1; ^{19}F NMR (376 MHz, CDCl_3) δ -62.5; MS (EI): m/z (relative intensity) 252.3 (M^+ , 100), 222.3 (21), 209.3 (21), 183.2 (13).

3-Methoxy-3',5'-di(trifluoromethyl)-1,1'-biphenyl (Table 3, **3kc**)²⁰

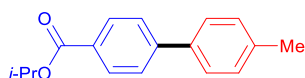
Eluents (Hexane : Dichloromethane = 9:1, $R_f = 0.40$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 3.90 (s, 3H), 7.01 (d, $J = 8.2\text{Hz}$, 1H), 7.13 (s, 1H), 7.19 (d, $J = 7.6\text{Hz}$, 1H), 7.43 (t, $J = 7.9\text{Hz}$, 1H), 7.88 (s, 1H), 8.02 (s, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 55.3, 113.1, 114.0, 119.6, 121.0 (m), 123.4 (q, $J = 270\text{Hz}$), 127.2, 130.3, 131.9 (q, $J = 38\text{Hz}$), 139.6, 143.2, 160.3; ^{19}F NMR (376 MHz, CDCl_3) δ -62.9; MS (EI): m/z (relative intensity) 320.4 (M^+ , 100), 301.4 (14), 290.3 (29), 277.3 (21), 251.3 (13), 188.2 (7).

4'-Methyl[1,1'-biphenyl]-4-carbonitrile (Table 3, **3la**)¹⁶

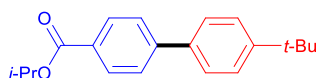
Eluents (Hexane : Ethyl acetate = 9:1, $R_f = 0.50$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 2.45 (s, 3H), 7.32 (d, $J = 7.9\text{Hz}$, 2H), 7.51 (d, $J = 9.2\text{Hz}$, 2H), 7.66–7.71 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 20.9, 110.2, 118.8, 126.8, 127.1, 129.6, 132.3, 135.9, 138.5, 145.3; MS (EI): m/z (relative intensity) 193.3 (M^+ , 100), 178.2 (10), 165.2 (20), 91.2 (7).

4'-(Trifluoromethyl)-[1,1'-biphenyl]-4-carbonitrile (Table 3, **3lg**)²¹

Eluents (Hexane : Ethyl acetate = 9:1, R_f = 0.50) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 7.68–7.77 (m, 8H); ^{13}C NMR (100 MHz, CDCl_3) δ 111.9, 118.5, 122.6, 125.3, 126.0 (q, J = 4.0Hz), 127.7 (d, J = 33Hz), 130.6 (d, J = 32Hz), 132.7, 142.6, 144.0; ^{19}F NMR (376 MHz, CDCl_3) δ -62.6; MS (EI): m/z (relative intensity) 247.3 (M^+ , 100), 228.3 (13), 197.2 (6), 177.2 (10), 151.2 (6), 98.7 (5).

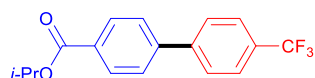
Isopropyl 4'-methyl-[1,1'-biphenyl]-4-carboxylate (Table 3, **3ma**)

Eluents (Hexane : Ethyl acetate = 9:1, R_f = 0.40) was used for flash column chromatography. White solid; m.p. 115.7–117.6°C; ^1H NMR (400 MHz, CDCl_3) δ 1.41 (d, J = 6.2Hz, 6H), 2.42 (s, 3H), 5.30 (sept, J = 6.2Hz, 1H), 7.28 (d, J = 7.8Hz, 2H), 7.54 (d, J = 8.0Hz, 2H), 7.65 (d, J = 8.2Hz, 2H), 8.11 (d, J = 8.2Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 21.1, 21.9, 68.2, 126.6, 127.0, 129.3, 129.6, 129.9, 137.1, 137.7, 145.3, 166.0; MS (EI): m/z (relative intensity) 254.4 (M^+ , 98), 212.3 (100), 195.3 (92), 165.3 (36), 152.2 (40), 97.3 (7); HRMS calcd. for $\text{C}_{17}\text{H}_{18}\text{NaO}_2^+$: 277.1199, found 277.1209.

Isopropyl 4'-(*tert*-butyl)-[1,1'-biphenyl]-4-carboxylate (Table 3, **3mf**)

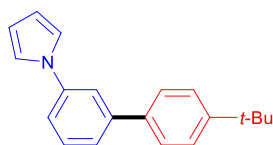
Eluents (Hexane : Ethyl acetate = 9:1, R_f = 0.40) was used for flash column chromatography. White solid; m.p. 53.8–56.7°C; ^1H NMR (400 MHz, CDCl_3) δ 1.40–1.43 (m, 15H), 5.31 (sept, J = 6.1Hz, 1H), 7.51 (d, J = 6.6Hz, 2H), 7.60 (d, J = 8.0Hz, 2H), 7.68 (d, J = 8.0Hz, 2H), 8.13 (d, J = 8.0Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 21.9, 31.3, 34.5, 68.2, 125.8, 126.7, 126.9, 129.3, 130.0, 137.1, 145.2, 151.2, 166.0; MS (EI): m/z (relative intensity) 296.5 (M^+ , 38), 281.4 (100), 239.3 (27), 97.2 (10); HRMS calcd. for $\text{C}_{20}\text{H}_{25}\text{O}_2^+$: 297.1849, found 297.1857.

Isopropyl 4'-(trifluoromethyl)-[1,1'-biphenyl]-4-carboxylate (Table 3, **3mg**)²¹



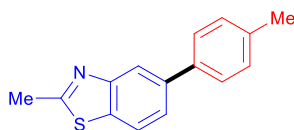
Eluents (Hexane : Ethyl acetate = 20:1, $R_f = 0.30$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 1.40 (d, $J = 6.2\text{Hz}$, 6H), 5.29 (sept, $J = 6.2\text{Hz}$, 1H), 7.64 (d, $J = 8.3\text{Hz}$, 2H), 7.70–7.71 (m, 4H), 8.13 (d, $J = 8.3\text{Hz}$, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 21.8, 68.5, 124.1 (q, $J = 270\text{Hz}$), 125.8 (q, $J = 4\text{Hz}$), 127.0, 127.5, 130.0 (q, $J = 32\text{Hz}$), 130.1, 130.6, 143.5, 143.7, 165.6; ^{19}F NMR (376 MHz, CDCl_3) δ -62.5; MS (EI): m/z (relative intensity) 308.4 (M^+ , 32), 266.3 (70), 249.3 (100), 201.3 (30), 152.2 (35), 59.2 (18).

4-(*tert*-Butylphenyl)-1*H*-benzopyrrole (Table 3, **3nf**)

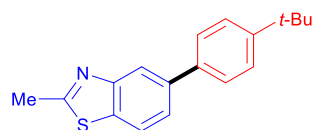


Eluents (Hexane : Ethyl acetate = 10:1, $R_f = 0.40$) was used for flash column chromatography. White solid; m.p. 123.8-125.2°C; ^1H NMR (400 MHz, CDCl_3) δ 1.49 (s, 9H), 6.48–6.49 (m, 2H), 7.25 (m, 2H), 7.42–7.44 (m, 1H), 7.53 (d, $J = 6.2\text{Hz}$, 2H), 7.60 (d, $J = 8.3\text{Hz}$, 2H), 7.66 (d, $J = 8.3\text{Hz}$, 2H), 7.72 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 31.3, 34.5, 110.4, 119.0, 119.1, 119.3, 124.2, 125.7, 126.7, 129.7, 137.4, 141.1, 142.6, 150.7; MS (EI): m/z (relative intensity) 275.4 (M^+ , 58), 260.4 (100), 232.3 (13), 116.2 (13); HRMS calcd. for $\text{C}_{20}\text{H}_{22}\text{N}^+$: 276.1747, found 276.1754.

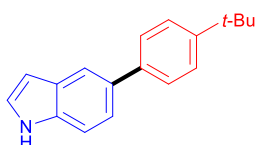
2-Methyl-5-(4-methylphenyl)-benzothiazole (Table 3, **3oa**)²²



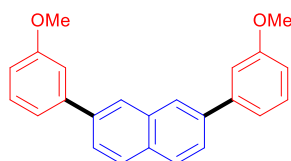
Eluents (Hexane : Ethyl acetate = 9:1, $R_f = 0.50$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 2.41 (s, 3H), 2.84 (s, 3H), 7.28 (d, $J = 7.6\text{Hz}$, 2H), 7.56–7.59 (m, 3H), 7.83 (d, $J = 8.4\text{Hz}$, 1H), 8.18–8.19 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 20.0, 21.0, 31.7, 120.3, 121.3, 124.0, 127.0, 129.5, 134.1, 137.7, 139.4, 153.9, 167.5; MS (EI): m/z (relative intensity) 239.3 (M^+ , 100), 197.2 (12), 165.2 (12), 152.2 (10), 119.6 (5).

2-Methyl-5-(4-*tert*-butylphenyl)-benzothiazole (Table 3, **3of**)²³

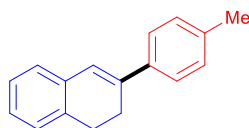
Eluents (Hexane : Ethyl acetate = 9:1, $R_f = 0.50$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 1.42 (s, 9H), 2.88 (s, 3H), 7.54 (d, $J = 6.5\text{Hz}$, 2H), 7.61–7.67 (m, 3H), 7.86 (d, $J = 8.3\text{Hz}$, 1H), 8.23–8.24 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 20.0, 34.49, 31.3, 120.4, 121.3, 124.04, 125.8, 126.9, 134.2, 137.7, 139.3, 150.4, 154.0, 167.5; MS (EI): m/z (relative intensity) 281.4 (M^+ , 46), 266.4 (100), 238.3 (13), 119.2 (13).

5-(4-*tert*-Butylphenyl)-1*H*-indole (Table 3, **3pf**)²⁴

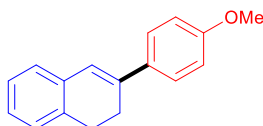
Eluents (Hexane : Ethyl acetate = 9:1, $R_f = 0.30$) was used for flash column chromatography. ^1H NMR (400 MHz, CD_2Cl_2) δ 1.47 (s, 9H), 6.67–6.68 (m, 1H), 7.27–7.29 (m, 1H), 7.49–7.51 (m, 2H), 7.57 (d, $J = 8.5\text{Hz}$, 2H), 7.70 (d, $J = 8.4\text{Hz}$, 2H), 7.95 (s, 1H), 8.31 (s, 1H); ^{13}C NMR (100 MHz, CD_2Cl_2) δ 31.2, 34.3, 102.6, 111.2, 118.7, 121.5, 125.0, 125.6, 126.7, 128.4, 132.9, 135.3, 139.4, 149.3; MS (EI): m/z (relative intensity) 249.4 (M^+ , 56), 234.4 (100), 204.3 (13), 103.2 (16), 89.2(5).

2,7-Di(3-methoxyphenyl)-naphthalene (Table 3, **3qc**)

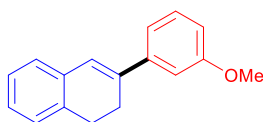
Eluents (Hexane : Dichloromethane = 10:1, $R_f = 0.30$) was used for flash column chromatography. White solid; m.p. 81.2–83.9°C; ^1H NMR (400 MHz, CDCl_3) δ 3.94 (s, 6H), 7.01 (d, $J = 7.3\text{Hz}$, 2H), 7.36–7.48 (m, 6H), 7.80 (d, $J = 8.2\text{ Hz}$, 2H), 7.96 (d, $J = 8.3\text{ Hz}$, 2H), 8.16 (s, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 55.2, 112.8, 113.0, 119.8, 125.6, 126.0, 128.0, 129.7, 131.8, 133.7, 138.7, 142.4, 159.9; MS (EI): m/z (relative intensity) 340.5 (M^+ , 100), 297.4 (13), 254.4 (6), 170.3 (9); HRMS calcd. for $\text{C}_{24}\text{H}_{21}\text{O}_2^+$: 341.1536, found 341.1540.

3-(4-Methylphenyl)-1,2-dihydronaphthalene (Table 4, **5aa**)²⁵

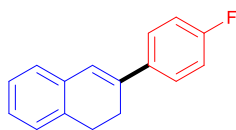
Eluents (Hexane, $R_f = 0.50$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 2.48 (s, 3H), 2.84 (t, $J = 8.2\text{Hz}$, 2H), 3.06 (t, $J = 7.6\text{Hz}$, 2H), 6.95 (s, 1H), 7.23–7.30 (m, 6H), 7.56 (d, $J = 8.2\text{Hz}$, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 21.1, 26.2, 28.1, 123.4, 124.9, 126.4, 126.5, 126.7, 127.2, 129.1, 134.6, 134.8, 137.0, 138.1, 138.5; MS (EI): m/z (relative intensity) 220.2 (M^+ , 100), 205.1 (35), 128.1 (17), 105.1 (30).

3-(4-Methoxyphenyl)-1,2-dihydronaphthalene (Table 4, **5ab**)²⁵

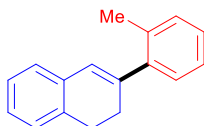
Eluents (Hexane : Dichloromethane = 9:1, $R_f = 0.25$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 2.77 (t, $J = 8.0\text{Hz}$, 2H), 3.00 (t, $J = 7.6\text{Hz}$, 2H), 3.87 (s, 3H), 6.85 (s, 1H), 6.97 (d, $J = 8.8\text{Hz}$, 2H), 7.17–7.24 (m, 4H), 7.55 (d, $J = 8.8\text{Hz}$, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 26.2, 28.1, 55.2, 113.7, 122.5, 126.24, 126.27, 126.51, 126.55, 127.0, 133.4, 134.4, 134.9, 138.1, 159.0; MS (EI): m/z (relative intensity) 236.2 (M^+ , 100), 221.1 (26), 178.1 (15), 121.1 (26).

3-(3-Methoxyphenyl)-1,2-dihydronaphthalene (Table 4 and Scheme 2, **5ac**)²⁶

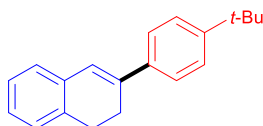
Eluents (Hexane : Dichloromethane = 9:1, $R_f = 0.25$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 2.83 (t, $J = 8.2\text{Hz}$, 2H), 3.04 (t, $J = 7.7\text{Hz}$, 2H), 3.92 (s, 3H), 6.94–6.96 (m, 2H), 7.18–7.26 (m, 6H), 7.38 (t, $J = 7.9\text{Hz}$, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 26.3, 28.0, 55.1, 110.9, 112.5, 117.6, 124.5, 126.53, 126.58, 126.9, 127.1, 129.3, 134.5, 134.7, 138.4, 142.5, 159.6; MS (EI): m/z (relative intensity) 236.4 (M^+ , 100), 221.3 (12), 205.3 (14), 189.3 (12), 165.2 (10), 121.2 (37).

3-(4-Fluorophenyl)-1,2-dihydronaphthalene (Table 4, **5ad**)²⁵

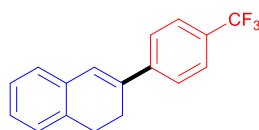
Eluents (Hexane, $R_f = 0.50$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 2.76 (t, $J = 8.0\text{Hz}$, 2H), 3.00 (t, $J = 7.7\text{Hz}$, 2H), 6.85 (s, 1H), 7.11 (t, $J = 8.6\text{Hz}$, 2H), 7.11–7.26 (m, 4H), 7.53–7.57 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 26.3, 28.0, 115.2 (d, $J = 21\text{Hz}$), 124.1, 126.5, 126.60, 126.68, 126.99, 127.1, 134.5, 137.1 (d, $J = 3\text{Hz}$), 137.5, 160.9, 162.1 (d, $J = 245\text{Hz}$); ^{19}F NMR (376 MHz, CDCl_3) δ -114.8; MS (EI): m/z (relative intensity) 224.3 (M^+ , 100), 202.3 (20), 128.2 (17), 109.2 (46).

3-(2-Methylphenyl)-1,2-dihydronaphthalene (Table 4, **5ae**)²⁵

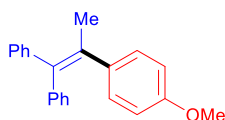
Eluents (Hexane, $R_f = 0.50$) was used for flash column chromatography. Colourless liquid. ^1H NMR (400 MHz, CDCl_3) δ 2.49 (s, 3H), 2.69 (t, $J = 7.8\text{Hz}$, 2H), 3.05 (t, $J = 7.7\text{Hz}$, 2H), 6.54 (s, 1H), 7.19 (d, $J = 6.7\text{Hz}$, 1H), 7.18–7.33 (m, 7H); ^{13}C NMR (100 MHz, CDCl_3) δ 20.4, 28.3, 28.8, 125.7, 126.1, 126.4, 126.5, 126.8, 127.0, 127.2, 128.0, 130.3, 134.5, 134.6, 134.9, 141.1, 142.8; MS (EI): m/z (relative intensity) 220.3 (M^+ , 100), 205.3 (33), 128.2 (25), 105.2 (60).

3-(4-*tert*-Butylphenyl)-1,2-dihydronaphthalene (Table 4, **5af**)²⁵

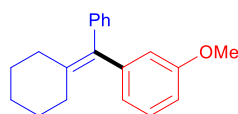
Eluents (Hexane, $R_f = 0.50$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 1.49 (s, 9H), 2.87 (t, $J = 7.6\text{Hz}$, 2H), 3.07 (t, $J = 7.6\text{Hz}$, 2H), 6.99 (s, 1H), 7.24–7.33 (m, 4H), 7.54 (d, $J = 8.5\text{Hz}$, 2H), 7.64 (d, $J = 8.6\text{Hz}$, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 26.2, 28.1, 31.2, 34.5, 123.6, 124.8, 125.3, 126.4, 126.5, 126.7, 127.1, 134.7, 134.8, 138.0, 138.3, 150.3; MS (EI): m/z (relative intensity) 262.4 (M^+ , 77), 247.4 (100), 202.3 (7), 131.2 (12), 109.6 (11), 91.2 (6).

3-(4-Trifluoromethylphenyl)-1,2-dihydronaphthalene (Table 4, **5ag**)²⁵

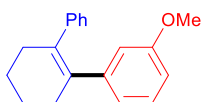
Eluents (Hexane, $R_f = 0.50$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 2.72–2.76 (m, 2H), 2.95–2.99 (m, 2H), 6.92 (s, 1H), 7.15–7.62 (m, 4H), 7.65–7.67 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 26.2, 28.0, 124.2 (q, $J = 270\text{Hz}$), 125.2, 125.3 (q, $J = 4\text{Hz}$), 126.3, 126.7, 127.0, 127.3, 127.6, 129.0 (q, $J = 32\text{Hz}$), 134.1, 134.8, 137.1, 144.6; ^{19}F NMR (376 MHz, CDCl_3) δ –62.3; MS (EI): m/z (relative intensity) 274.4 (M^+ , 100), 259.3 (13), 233.3 (19), 202.3 (16), 115.2 (40).

1,1-Diphenyl-2-(4-methoxyphenyl)propene (Table 4, **5bb**)²⁷

Eluents (Hexane : Dichloromethane = 20:1, $R_f = 0.30$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 2.17 (s, 3H), 3.77 (s, 3H), 6.74 (d, $J = 8.5\text{Hz}$, 2H), 6.96 (d, $J = 7.2\text{Hz}$, 2H), 7.04–7.13 (m, 5H), 7.29 (d, $J = 6.4\text{Hz}$, 3H), 7.38 (t, $J = 7.2\text{Hz}$, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 23.2, 55.0, 113.2, 125.6, 126.4, 127.4, 128.0, 129.8, 130.3, 130.8, 135.0, 136.1, 138.7, 143.3, 143.7, 157.8; MS (EI): m/z (relative intensity) 300.1 (M^+ , 100), 285.1 (12), 191.1 (8), 165.1 (6).

1-(Cyclohexylidenephénylmethyl)-3-methoxy-benzene (Table 4, **5cc**)²⁸

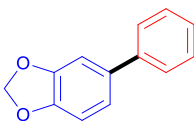
Eluents (Hexane : Dichloromethane = 20:1, $R_f = 0.30$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 1.57–1.60, (m, 6H), 2.21–2.24 (m, 4H), 6.67–6.68 (m, 1H), 6.72–6.73 (m, 2H), 7.11–7.17 (m, 4H), 7.22–7.24 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 26.7, 28.64, 28.66, 32.3, 32.4, 55.0, 111.2, 115.5, 122.3, 126.0, 127.8, 128.7, 129.6, 134.3, 139.2, 142.8, 144.5, 159.1; MS (EI): m/z (relative intensity) 264.4 (M^+ , 100), 235.4 (14), 221.3 (20), 115.2 (20), 91.2 (18).

1-Phenyl-2-(3-methoxyphenyl)-cyclohex-1-ene (Table 4, **5dc**)²⁹

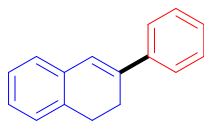
Eluents (Hexane : Dichloromethane = 20:1, $R_f = 0.30$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 1.83–1.86 (m, 4H), 2.45–2.49 (m, 4H), 3.56 (s, 3H), 6.52–6.53 (m, 1H), 6.59–6.63 (m, 2H), 7.00–7.07 (m, 4H), 7.09–7.13 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 23.1, 23.2, 31.7, 31.9, 54.8, 111.6, 114.5, 121.3, 125.6, 127.6, 128.4, 128.8, 134.7, 135.1, 143.9, 145.1, 158.8; MS (EI): m/z (relative intensity) 264.4 (M^+ , 100), 221.3 (17), 205.3 (13), 115.2 (7), 91.2 (13).

1-[4'-*tert*-Butylcyclohex-1-enyl]-4-methoxybenzene (Table 4, **5eb**)³⁰

Eluents (Hexane : Dichloromethane = 9:1, $R_f = 0.40$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 0.9 (s, 9H), 1.34–1.42 (m, 2H), 1.99–2.01 (m, 2H), 2.24–2.28 (m, 1H), 2.38–2.45 (m, 1H), 2.50–2.54 (m, 1H), 3.82 (s, 3H), 6.06–6.07 (m, 1H), 6.87 (d, $J = 8.5\text{Hz}$, 2H), 7.34 (d, $J = 8.5\text{Hz}$, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 24.4, 27.2, 27.3, 28.8, 32.1, 43.8, 55.1, 113.5, 123.1, 125.8, 134.8, 135.6, 158.3; MS (EI): m/z (relative intensity) 244.4 (M^+ , 88), 187.3 (35), 160.3 (100), 121.2 (42), 57.2 (20).

5-Phenyl-benzo[d][1,3]dioxole (Scheme 3, **3gh**)¹⁰

Eluents (Hexane : Dichloromethane = 20:1, $R_f = 0.30$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 6.02 (s, 2H), 6.93 (d, $J = 8.0\text{Hz}$, 1H), 7.10–7.13 (m, 2H), 7.36 (t, $J = 7.3\text{Hz}$, 1H), 7.46 (t, $J = 6.8\text{Hz}$, 2H), 7.56 (d, $J = 7.0\text{Hz}$, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 101.0, 107.6, 108.5, 120.5, 126.82, 126.86, 128.6, 135.5, 140.8, 147.0, 148.0; MS (EI): m/z (relative intensity) 198.2 (M^+ , 100), 139.2 (49), 98.8 (10).

3-Phenyl-1,2-dihydronaphthalene (Scheme 3, **5ah**)²⁵

Eluents (Hexane : Dichloromethane = 20:1, $R_f = 0.30$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 2.85 (t, $J = 8.0\text{Hz}$, 2H), 3.05 (t, $J = 7.6\text{Hz}$, 2H), 6.96 (s, 1H), 7.22–7.31 (m, 3H), 7.38 (t, $J = 7.3\text{Hz}$, 1H), 7.47 (t, $J = 7.8\text{Hz}$, 2H), 7.64 (d, 7.3Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 26.2, 28.1, 124.3, 125.0, 126.5, 126.9, 127.1, 127.2, 128.4, 134.6, 134.7, 138.6, 141.0; MS (EI): m/z (relative intensity) 206.2 (M^+ , 100), 191.2 (23), 128.2 (35), 91.2 (62).

12. X-ray crystallography data of NMe₂-CM-Phos L14

Table S1. Crystal data and structure refinement for p.

Identification code	BSYU001	
Empirical formula	C ₂₉ H ₃₉ N ₂ P	
Formula weight	446.59	
Temperature	296(2) K	
Wavelength	0.71073 Å	
Crystal system	Orthorhombic	
Space group	Pbca	
Unit cell dimensions	a = 15.0158(12) Å	= 90°.
	b = 15.6985(10) Å	= 90°.
	c = 21.9190(16) Å	= 90°.
Volume	5166.9(6) Å ³	
Z	8	
Density (calculated)	1.148 Mg/m ³	
Absorption coefficient	0.125 mm ⁻¹	
F(000)	1936	
Crystal size	0.500 x 0.400 x 0.300 mm ³	
Theta range for data collection	2.094 to 25.249°.	
Index ranges	-18<=h<=18, -18<=k<=18, -26<=l<=26	
Reflections collected	93019	
Independent reflections	4669 [R(int) = 0.0625]	
Completeness to theta = 25.242°	99.9 %	
Absorption correction	multi-scan	
Max. and min. transmission	0.7456 and 0.6423	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	4669 / 0 / 290	
Goodness-of-fit on F ²	1.027	
Final R indices [I>2sigma(I)]	R1 = 0.0398, wR2 = 0.1064	
R indices (all data)	R1 = 0.0598, wR2 = 0.1266	
Extinction coefficient	0.0065(7)	
Largest diff. peak and hole	0.232 and -0.174 e.Å ⁻³	

Table S2. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for p. $U(\text{eq})$ is defined as one third of the trace of the orthogonalized U^{ij} tensor.

	x	y	z	U(eq)
P(1)	8943(1)	1522(1)	3161(1)	42(1)
N(1)	8361(1)	-56(1)	2132(1)	47(1)
N(2)	10731(1)	-1791(1)	3825(1)	51(1)
C(1)	8306(1)	108(1)	1516(1)	48(1)
C(2)	7561(2)	133(1)	1137(1)	63(1)
C(3)	7707(2)	307(2)	529(1)	76(1)
C(4)	8547(2)	445(2)	306(1)	76(1)
C(5)	9282(2)	419(1)	674(1)	65(1)
C(6)	9168(2)	247(1)	1298(1)	50(1)
C(7)	9748(2)	158(1)	1808(1)	50(1)
C(8)	9245(1)	-14(1)	2306(1)	43(1)
C(9)	7614(2)	-263(2)	2509(1)	66(1)
C(10)	9519(1)	-147(1)	2951(1)	40(1)
C(11)	9965(1)	-891(1)	3096(1)	42(1)
C(12)	10255(1)	-1071(1)	3693(1)	41(1)
C(13)	10025(1)	-475(1)	4144(1)	46(1)
C(14)	9586(1)	262(1)	3995(1)	46(1)
C(15)	9339(1)	470(1)	3401(1)	40(1)
C(16)	10855(2)	-2444(1)	3368(1)	62(1)
C(17)	11022(2)	-1964(2)	4439(1)	62(1)
C(18)	9962(1)	2180(1)	3299(1)	45(1)
C(19)	10753(1)	1824(1)	2943(1)	57(1)
C(20)	11549(2)	2426(2)	2961(1)	70(1)
C(21)	11809(2)	2647(2)	3606(1)	74(1)
C(22)	11025(2)	2986(2)	3965(1)	71(1)
C(23)	10244(2)	2363(1)	3954(1)	59(1)
C(24)	8201(1)	1859(1)	3790(1)	46(1)
C(25)	7380(2)	1294(1)	3825(1)	65(1)
C(26)	6742(2)	1601(2)	4316(1)	85(1)
C(27)	6479(2)	2522(2)	4215(1)	92(1)
C(28)	7279(2)	3095(2)	4176(1)	77(1)
C(29)	7927(2)	2791(1)	3693(1)	61(1)

Table S3. Bond lengths [\AA] and angles [$^\circ$] for p.

P(1)-C(15)	1.8325(17)
P(1)-C(24)	1.8504(19)
P(1)-C(18)	1.8711(19)
N(1)-C(1)	1.379(2)
N(1)-C(8)	1.382(2)
N(1)-C(9)	1.430(3)
N(2)-C(12)	1.367(2)
N(2)-C(17)	1.442(2)
N(2)-C(16)	1.444(2)
C(1)-C(2)	1.394(3)
C(1)-C(6)	1.396(3)
C(2)-C(3)	1.378(3)
C(3)-C(4)	1.371(4)
C(4)-C(5)	1.368(3)
C(5)-C(6)	1.405(3)
C(6)-C(7)	1.422(3)
C(7)-C(8)	1.356(3)
C(8)-C(10)	1.487(2)
C(10)-C(11)	1.384(2)
C(10)-C(15)	1.408(2)
C(11)-C(12)	1.407(2)
C(12)-C(13)	1.404(2)
C(13)-C(14)	1.371(3)
C(14)-C(15)	1.394(2)
C(18)-C(23)	1.524(3)
C(18)-C(19)	1.527(3)
C(19)-C(20)	1.523(3)
C(20)-C(21)	1.507(3)
C(21)-C(22)	1.510(3)
C(22)-C(23)	1.526(3)
C(24)-C(25)	1.520(3)
C(24)-C(29)	1.534(3)
C(25)-C(26)	1.520(3)
C(26)-C(27)	1.514(4)
C(27)-C(28)	1.503(4)
C(28)-C(29)	1.516(3)

C(15)-P(1)-C(24)	103.89(8)
C(15)-P(1)-C(18)	100.73(8)
C(24)-P(1)-C(18)	102.37(8)
C(1)-N(1)-C(8)	108.55(16)
C(1)-N(1)-C(9)	124.21(16)
C(8)-N(1)-C(9)	127.20(16)
C(12)-N(2)-C(17)	120.75(16)
C(12)-N(2)-C(16)	120.52(15)
C(17)-N(2)-C(16)	118.28(16)
N(1)-C(1)-C(2)	129.5(2)
N(1)-C(1)-C(6)	107.98(16)
C(2)-C(1)-C(6)	122.52(18)
C(3)-C(2)-C(1)	116.9(2)
C(4)-C(3)-C(2)	121.5(2)
C(5)-C(4)-C(3)	121.8(2)
C(4)-C(5)-C(6)	118.9(2)
C(1)-C(6)-C(5)	118.34(19)
C(1)-C(6)-C(7)	106.52(17)
C(5)-C(6)-C(7)	135.1(2)
C(8)-C(7)-C(6)	108.11(19)
C(7)-C(8)-N(1)	108.82(16)
C(7)-C(8)-C(10)	129.76(18)
N(1)-C(8)-C(10)	121.41(16)
C(11)-C(10)-C(15)	120.82(15)
C(11)-C(10)-C(8)	118.09(15)
C(15)-C(10)-C(8)	121.09(15)
C(10)-C(11)-C(12)	122.23(15)
N(2)-C(12)-C(13)	122.08(16)
N(2)-C(12)-C(11)	121.59(15)
C(13)-C(12)-C(11)	116.33(16)
C(14)-C(13)-C(12)	120.97(16)
C(13)-C(14)-C(15)	123.14(16)
C(14)-C(15)-C(10)	116.31(16)
C(14)-C(15)-P(1)	124.38(13)
C(10)-C(15)-P(1)	118.79(12)
C(23)-C(18)-C(19)	109.54(17)
C(23)-C(18)-P(1)	118.94(13)

C(19)-C(18)-P(1)	110.57(13)
C(20)-C(19)-C(18)	111.70(17)
C(21)-C(20)-C(19)	111.75(19)
C(20)-C(21)-C(22)	111.54(19)
C(21)-C(22)-C(23)	111.45(19)
C(18)-C(23)-C(22)	110.36(17)
C(25)-C(24)-C(29)	110.28(17)
C(25)-C(24)-P(1)	110.99(13)
C(29)-C(24)-P(1)	109.28(13)
C(24)-C(25)-C(26)	111.23(18)
C(27)-C(26)-C(25)	111.3(2)
C(28)-C(27)-C(26)	111.8(2)
C(27)-C(28)-C(29)	111.3(2)
C(28)-C(29)-C(24)	111.98(18)

Symmetry transformations used to generate equivalent atoms:

Table S4. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for p. The anisotropic displacement factor exponent takes the form: $-2 \pi^2 [h^2 a^{*2} U^{11} + \dots + 2 h k a^* b^* U^{12}]$

	U ¹¹	U ²²	U ³³	U ²³	U ¹³	U ¹²
P(1)	50(1)	40(1)	37(1)	1(1)	-3(1)	2(1)
N(1)	52(1)	48(1)	40(1)	-3(1)	-6(1)	1(1)
N(2)	62(1)	48(1)	44(1)	-2(1)	-10(1)	10(1)
C(1)	65(1)	39(1)	41(1)	-4(1)	-13(1)	4(1)
C(2)	70(1)	61(1)	58(1)	-7(1)	-21(1)	10(1)
C(3)	101(2)	71(2)	55(1)	-2(1)	-33(1)	11(1)
C(4)	114(2)	72(2)	41(1)	4(1)	-17(1)	-1(2)
C(5)	89(2)	64(1)	42(1)	-2(1)	-2(1)	-8(1)
C(6)	69(1)	42(1)	39(1)	-6(1)	-7(1)	-1(1)
C(7)	56(1)	55(1)	40(1)	-6(1)	-3(1)	-1(1)
C(8)	51(1)	39(1)	40(1)	-6(1)	-8(1)	2(1)
C(9)	57(1)	81(2)	58(1)	-5(1)	-1(1)	-8(1)
C(10)	42(1)	42(1)	37(1)	-2(1)	-3(1)	-3(1)
C(11)	50(1)	41(1)	37(1)	-8(1)	-4(1)	2(1)
C(12)	44(1)	38(1)	42(1)	-1(1)	-3(1)	-1(1)
C(13)	62(1)	45(1)	32(1)	1(1)	-5(1)	0(1)
C(14)	62(1)	41(1)	35(1)	-3(1)	-1(1)	1(1)
C(15)	46(1)	38(1)	37(1)	0(1)	-1(1)	-1(1)
C(16)	69(1)	51(1)	66(1)	-10(1)	-12(1)	16(1)
C(17)	70(2)	67(1)	50(1)	9(1)	-10(1)	16(1)
C(18)	53(1)	39(1)	43(1)	6(1)	-2(1)	0(1)
C(19)	53(1)	62(1)	55(1)	-1(1)	3(1)	0(1)
C(20)	54(1)	77(2)	79(2)	7(1)	6(1)	-5(1)
C(21)	58(1)	68(2)	95(2)	10(1)	-16(1)	-14(1)
C(22)	79(2)	64(1)	69(1)	-6(1)	-14(1)	-22(1)
C(23)	70(1)	60(1)	47(1)	-2(1)	-6(1)	-13(1)
C(24)	49(1)	46(1)	44(1)	-2(1)	-3(1)	6(1)
C(25)	57(1)	65(1)	73(1)	-6(1)	7(1)	-4(1)
C(26)	59(2)	96(2)	100(2)	-7(2)	24(1)	-3(1)
C(27)	65(2)	111(2)	99(2)	-9(2)	9(2)	32(2)
C(28)	82(2)	70(2)	77(2)	-13(1)	-2(1)	31(1)
C(29)	69(2)	51(1)	64(1)	-2(1)	-2(1)	14(1)

Table S5. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^{-3}$) for p.

	x	y	z	U(eq)
H(2A)	6991	36	1288	76
H(3A)	7224	331	264	91
H(4A)	8620	560	-107	91
H(5A)	9847	514	514	78
H(7A)	10365	208	1800	61
H(9A)	7812	-350	2921	98
H(9B)	7192	197	2499	98
H(9C)	7337	-773	2360	98
H(11A)	10077	-1285	2789	51
H(13A)	10174	-581	4548	56
H(14A)	9446	642	4307	55
H(16A)	11202	-2900	3537	93
H(16B)	10285	-2658	3242	93
H(16C)	11160	-2207	3023	93
H(17A)	11345	-2492	4447	94
H(17B)	11403	-1511	4576	94
H(17C)	10514	-2004	4703	94
H(18A)	9836	2737	3118	54
H(19A)	10924	1279	3115	68
H(19B)	10578	1732	2522	68
H(20A)	12050	2160	2758	84
H(20B)	11403	2944	2742	84
H(21A)	12277	3073	3599	88
H(21B)	12041	2144	3807	88
H(22A)	10838	3526	3793	85
H(22B)	11206	3085	4384	85
H(23A)	10417	1836	4153	71
H(23B)	9747	2604	4178	71
H(24A)	8527	1815	4176	55
H(25A)	7561	714	3913	78
H(25B)	7080	1296	3433	78
H(26A)	6212	1247	4312	102

H(26B)	7021	1543	4712	102
H(27A)	6139	2566	3840	110
H(27B)	6101	2707	4548	110
H(28A)	7577	3108	4569	92
H(28B)	7087	3669	4080	92
H(29A)	8455	3147	3701	73
H(29B)	7654	2849	3294	73

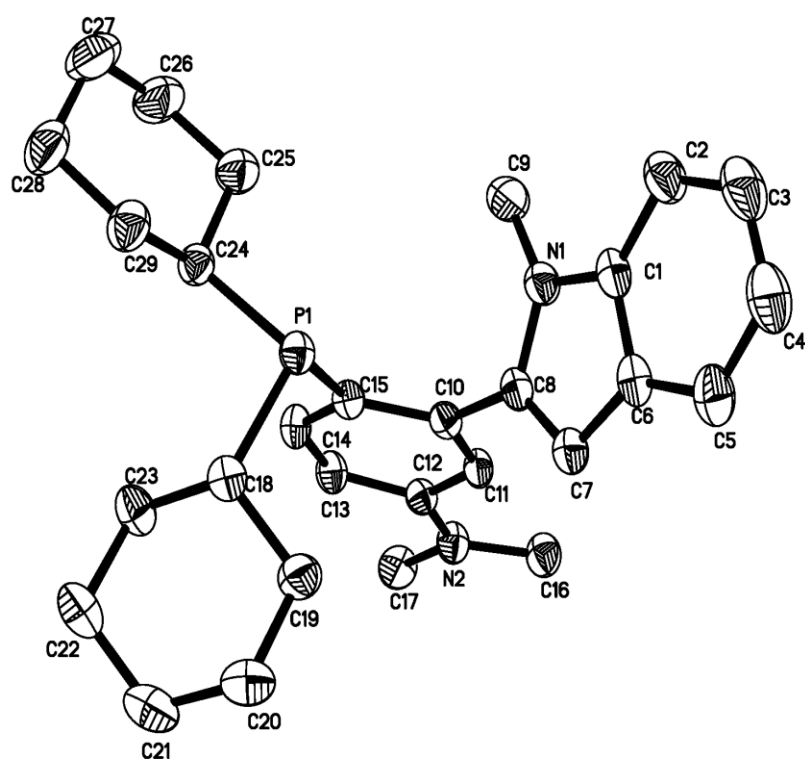
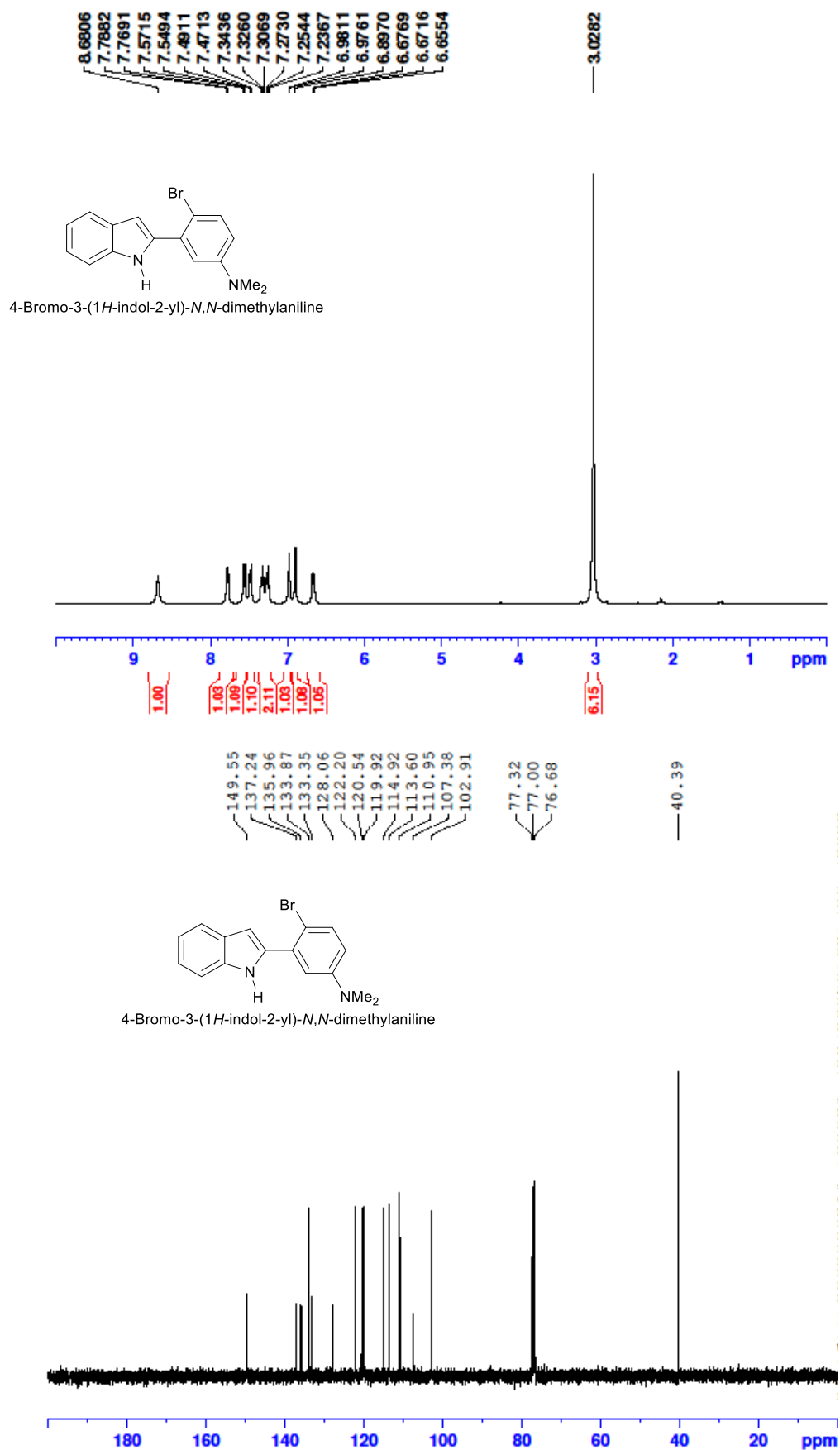
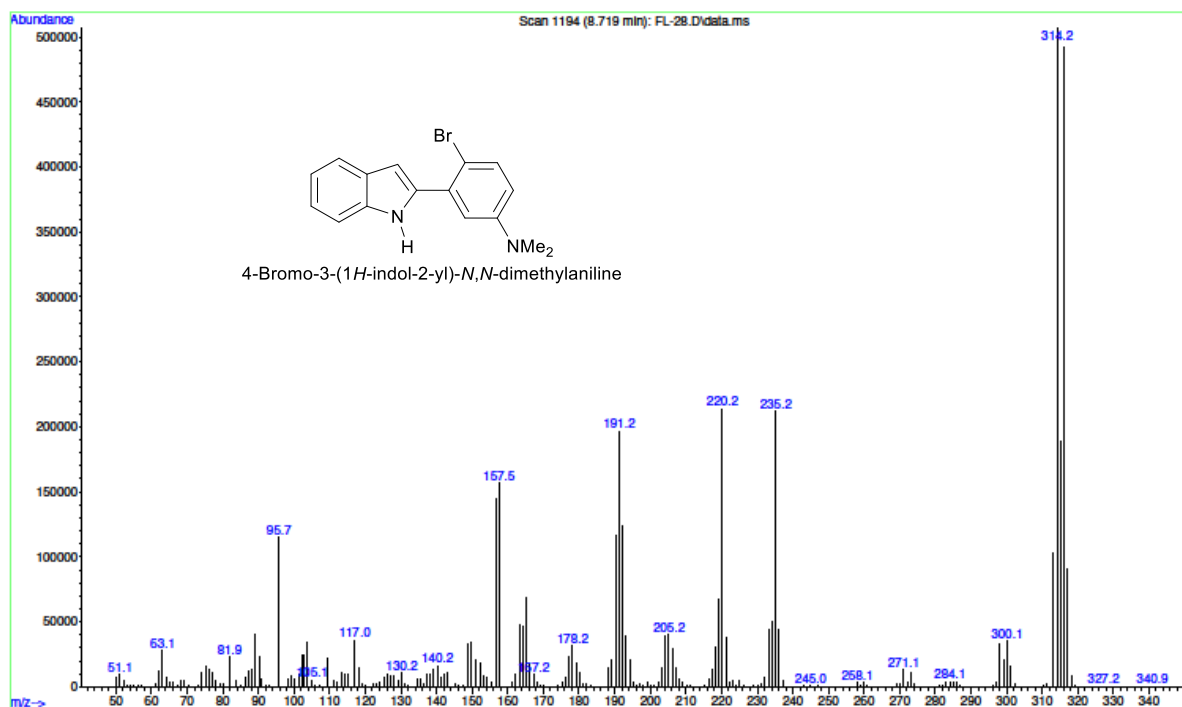
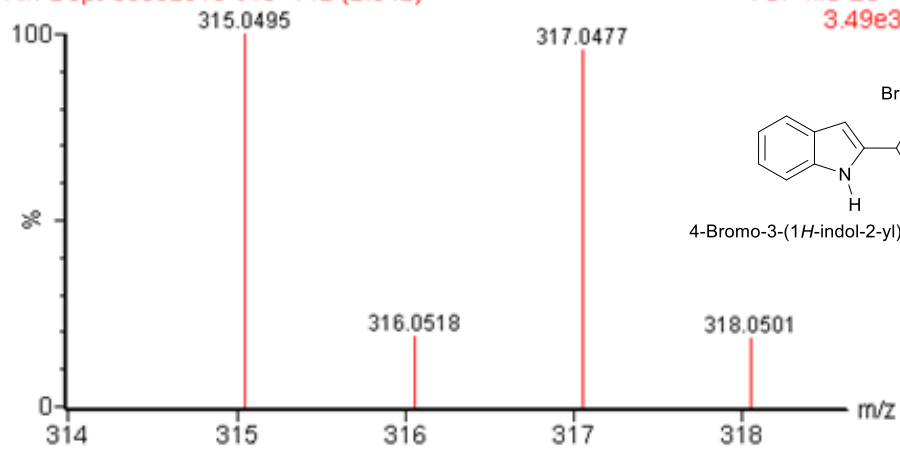


Figure S1. ORTEP drawing of NMe₂-CM-Phos (30% probability for the thermal ellipsoid)

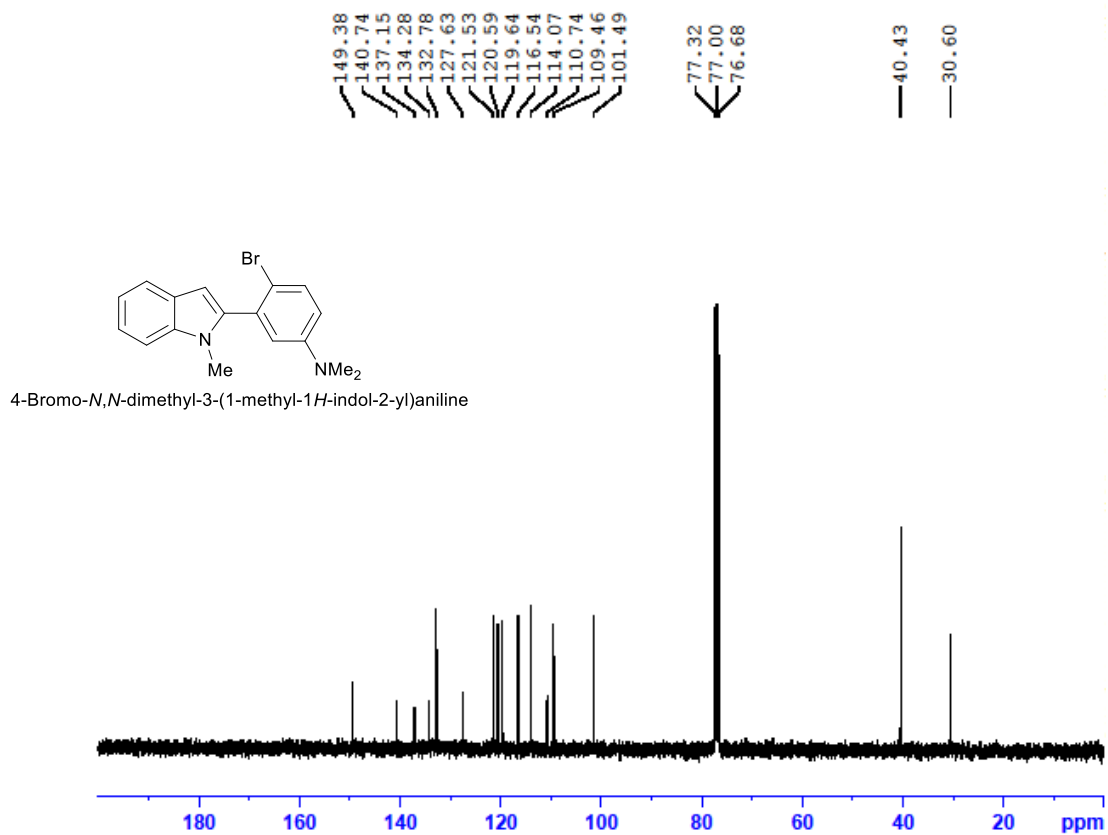
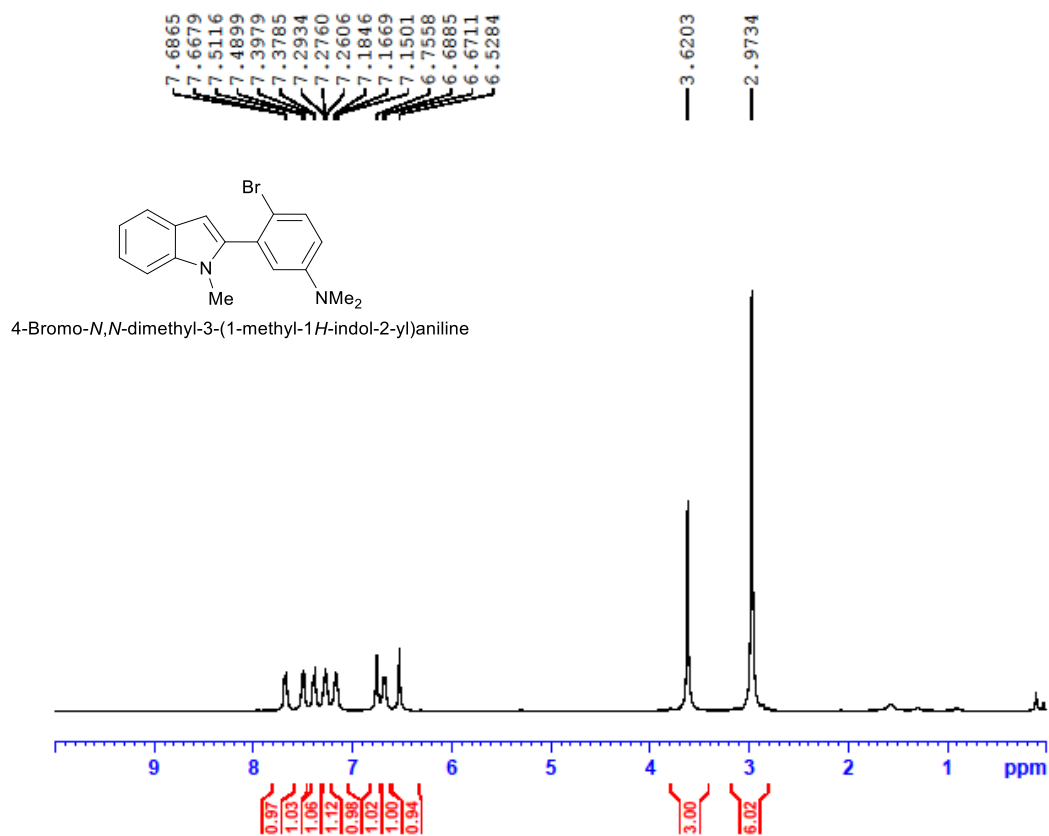
13. ^1H , ^{13}C , ^{19}F , ^{31}P NMR, MS and HRMS spectra

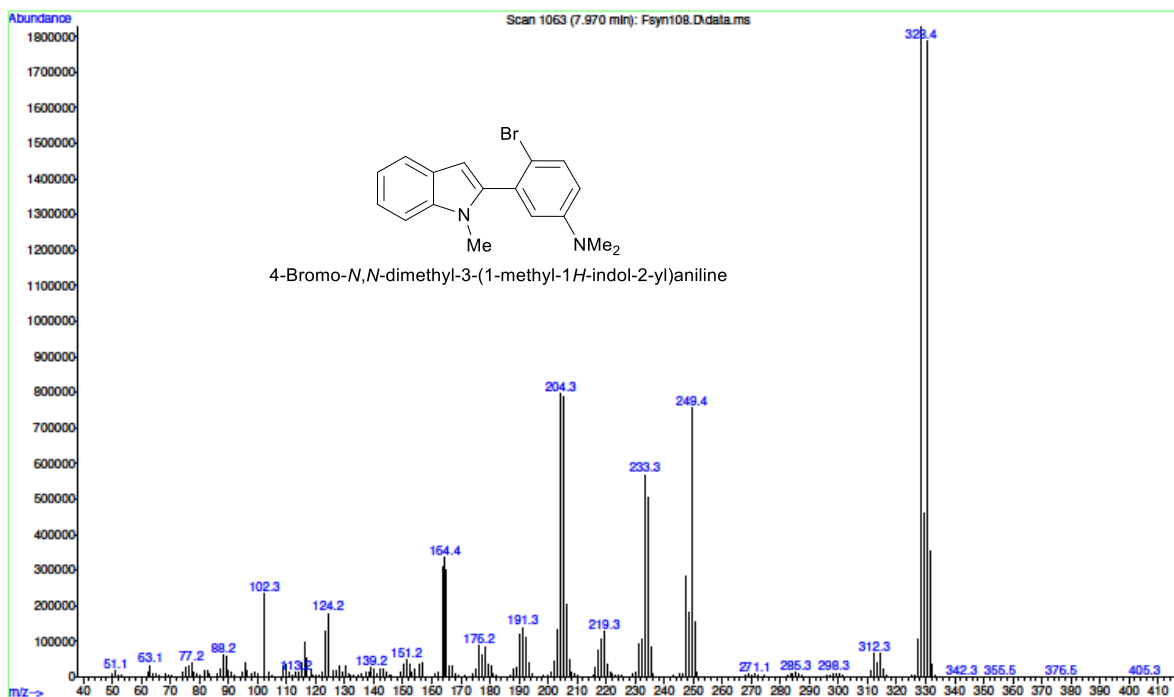


Kin-Dept-05062013-s18 142 (2.642)

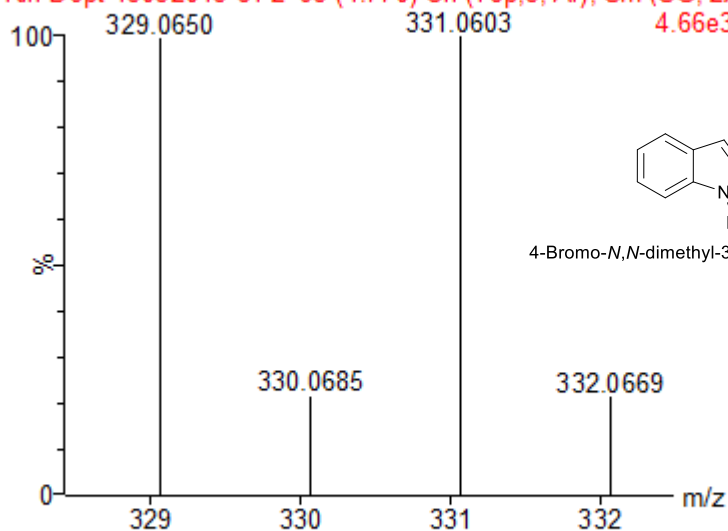


Mass	Calc. Mass	mDa	PPM	DBE	Score	Formula
315.0495	315.0497	-0.2	-0.6	9.5	1	C16 H16 N2 Br

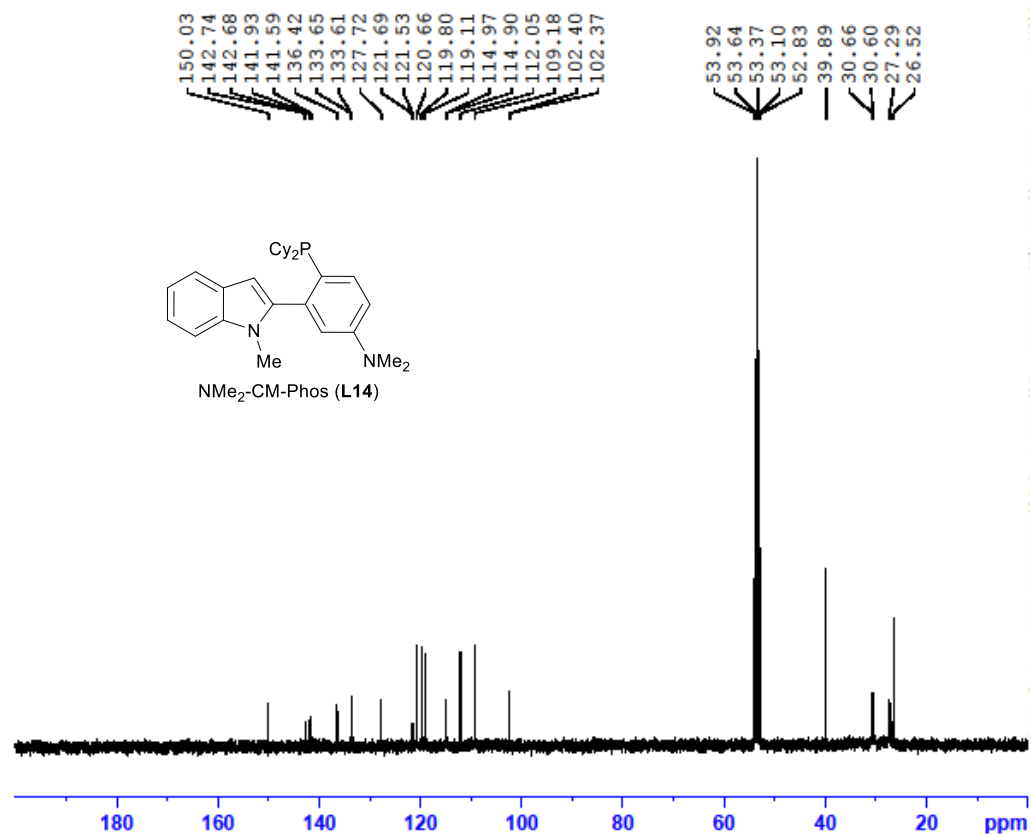
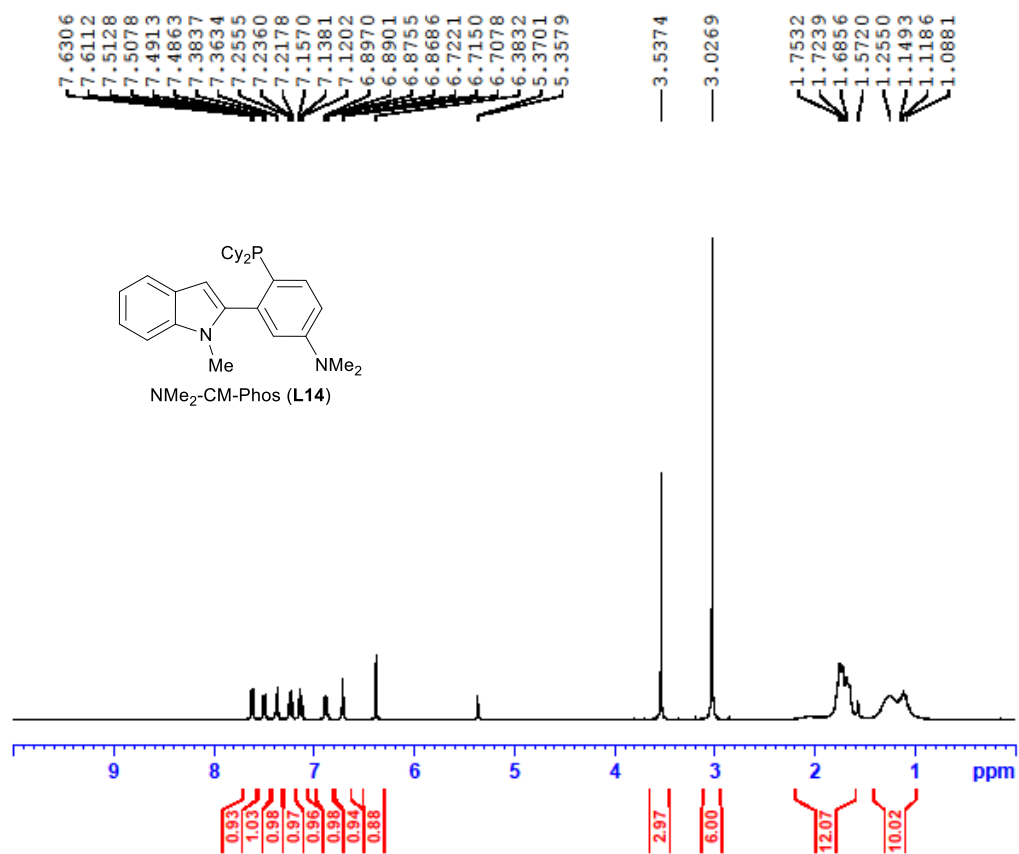


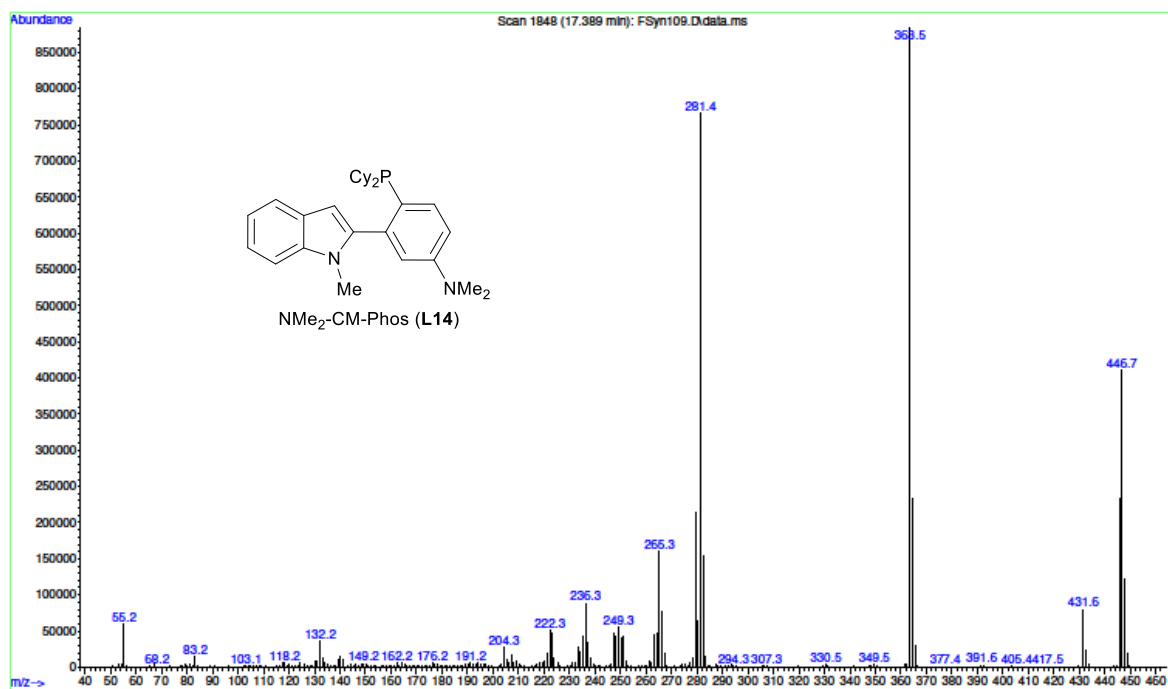
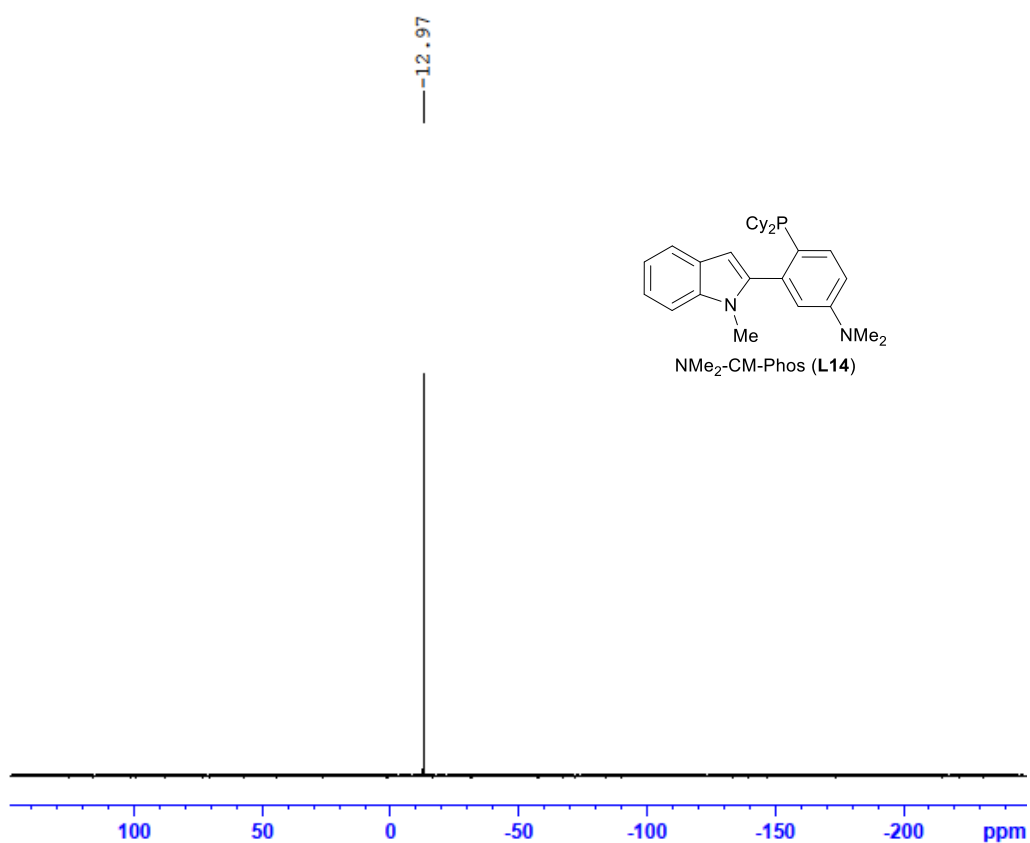


Kin-Dept-15052013-s4 2 95 (1.770) Cn (Top,5, Ar); Sm (SG, 2x
4.66e3

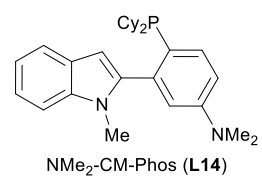
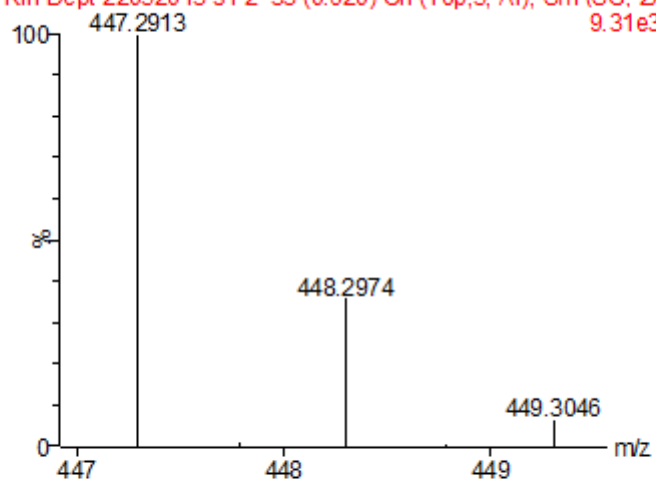


Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Formula
329.0650	329.0653	-0.3	-0.9	9.5	4.2	C17 H18 N2 Br

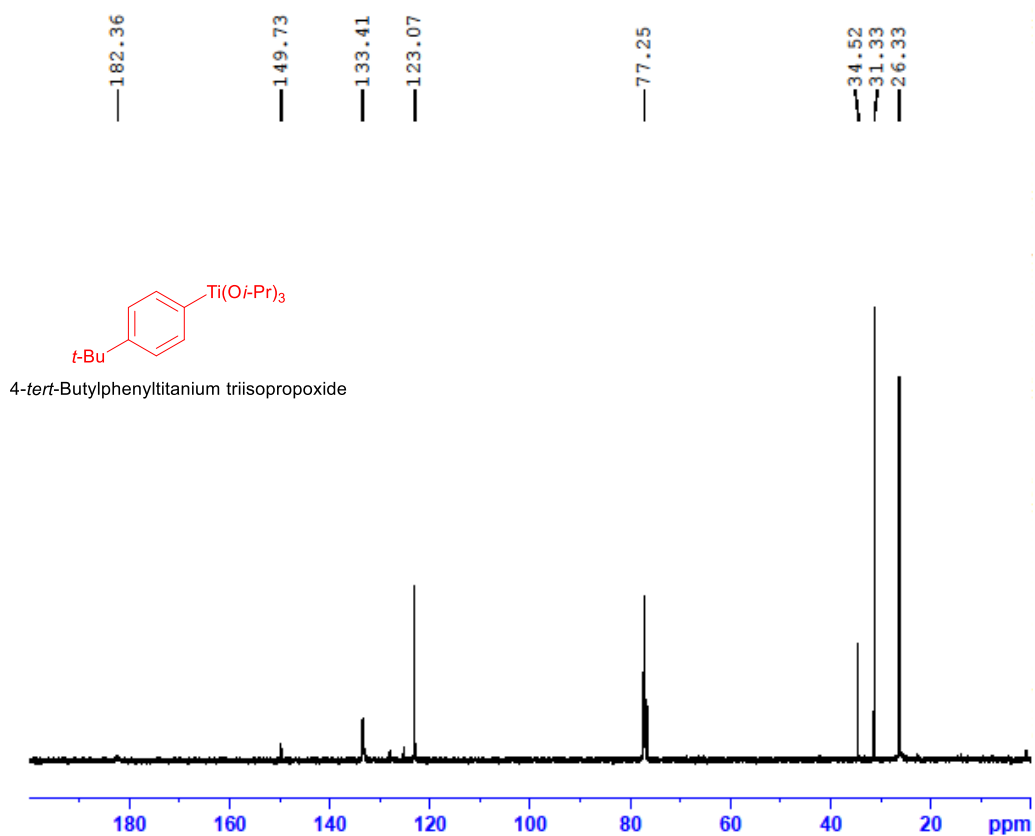
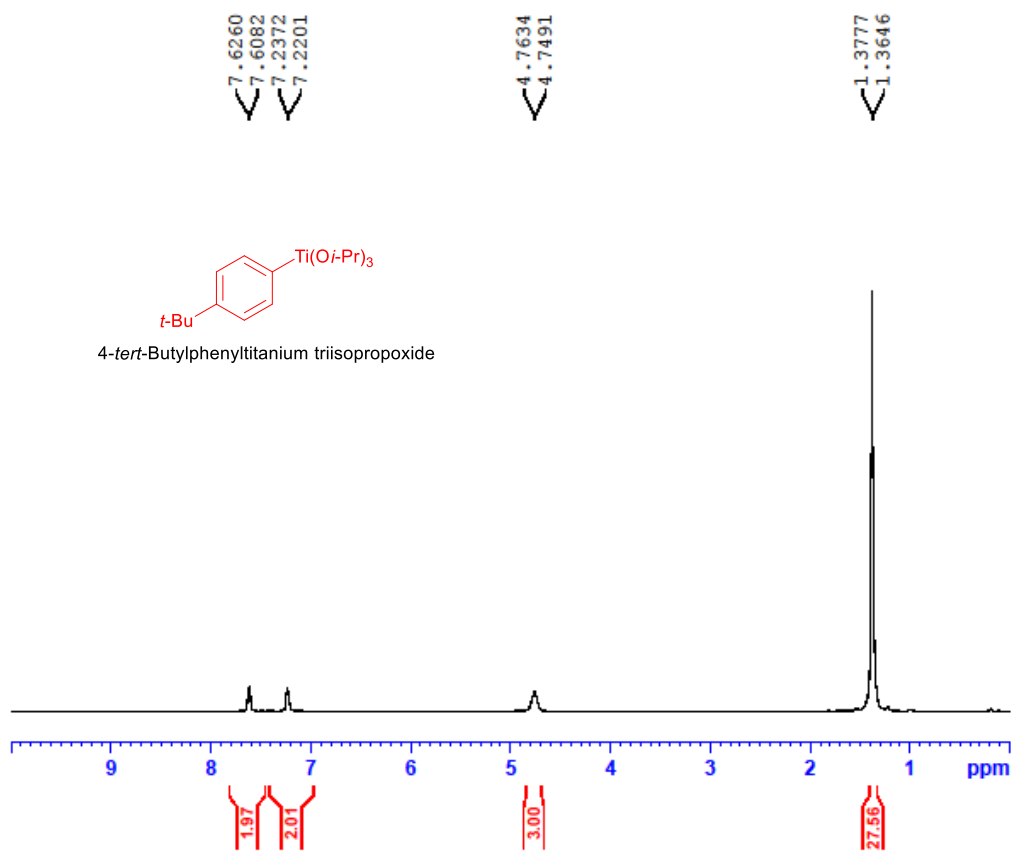


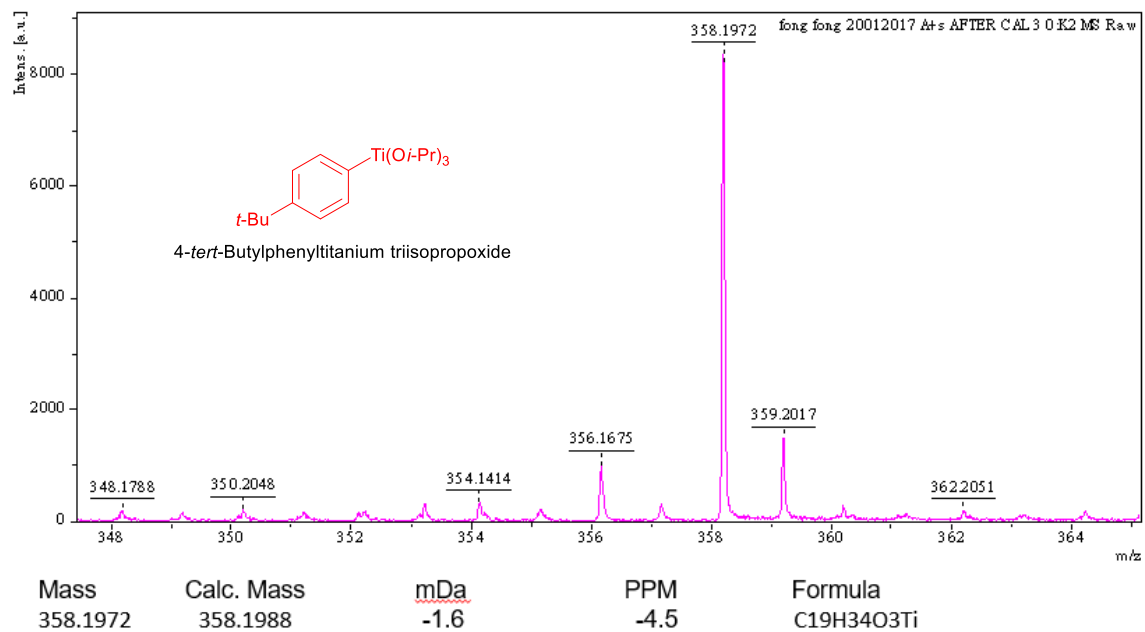


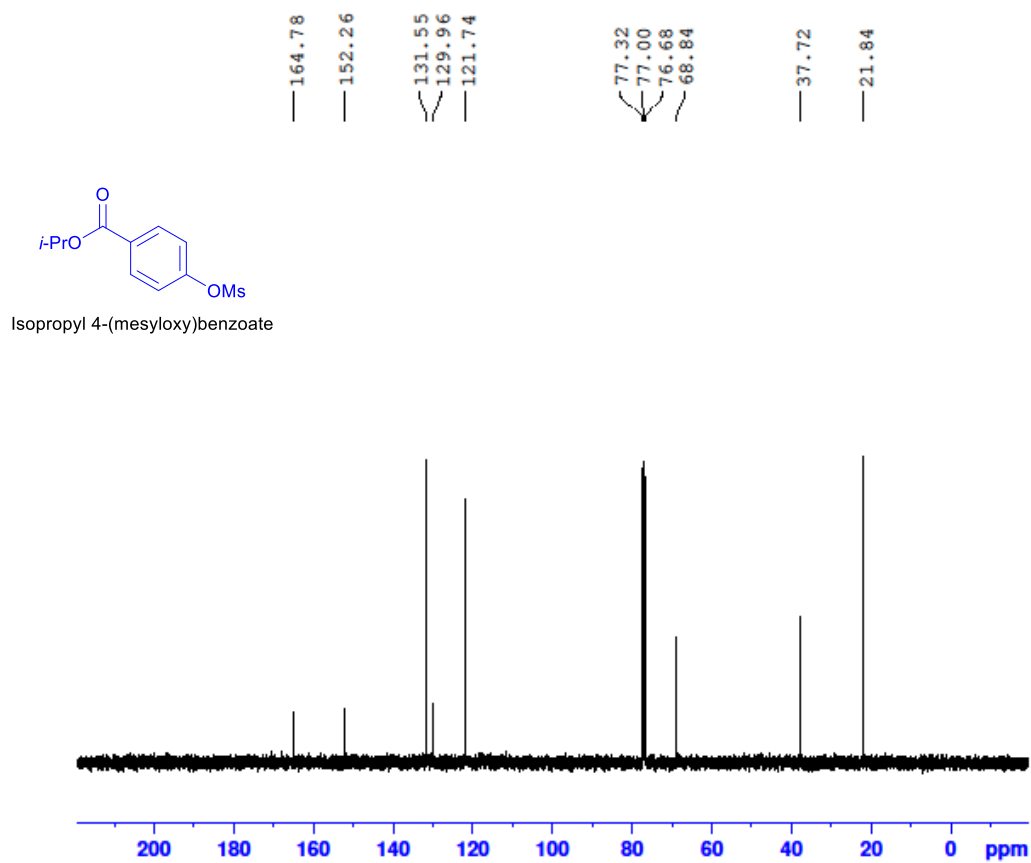
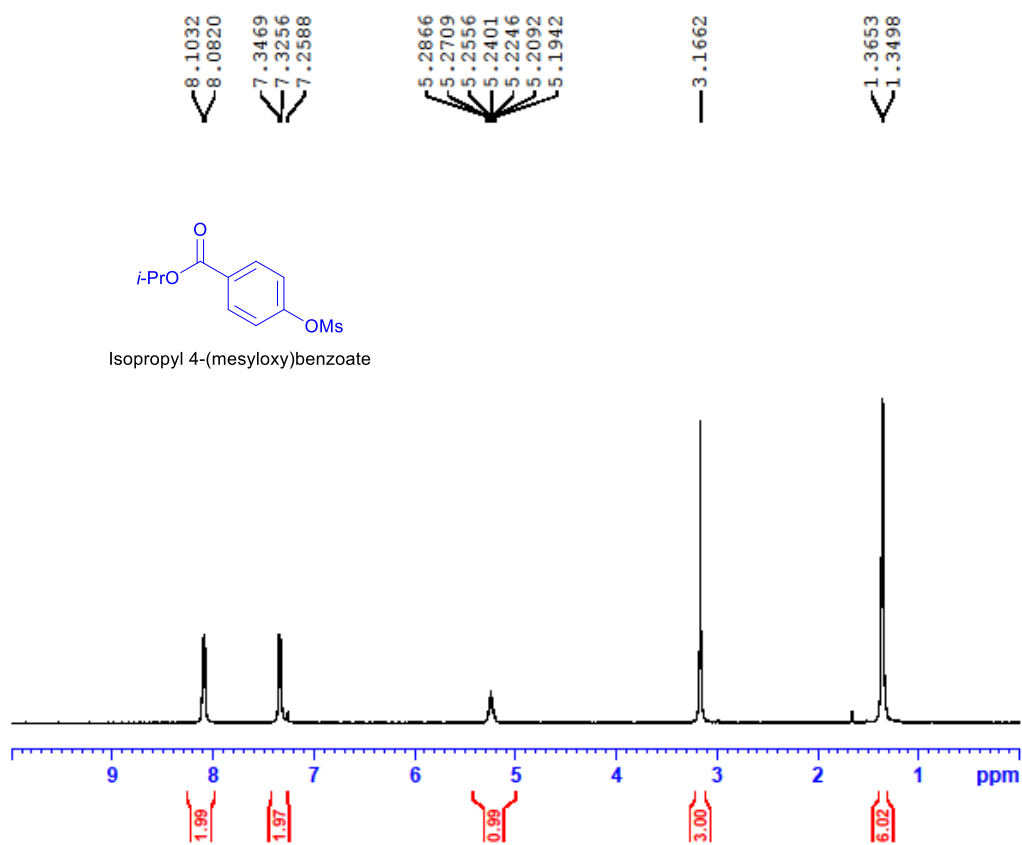
Kin-Dept-22052013-s1 2 33 (0.620) Cn (Top,5, Ar); Sm (SG, 2x
9.31e3

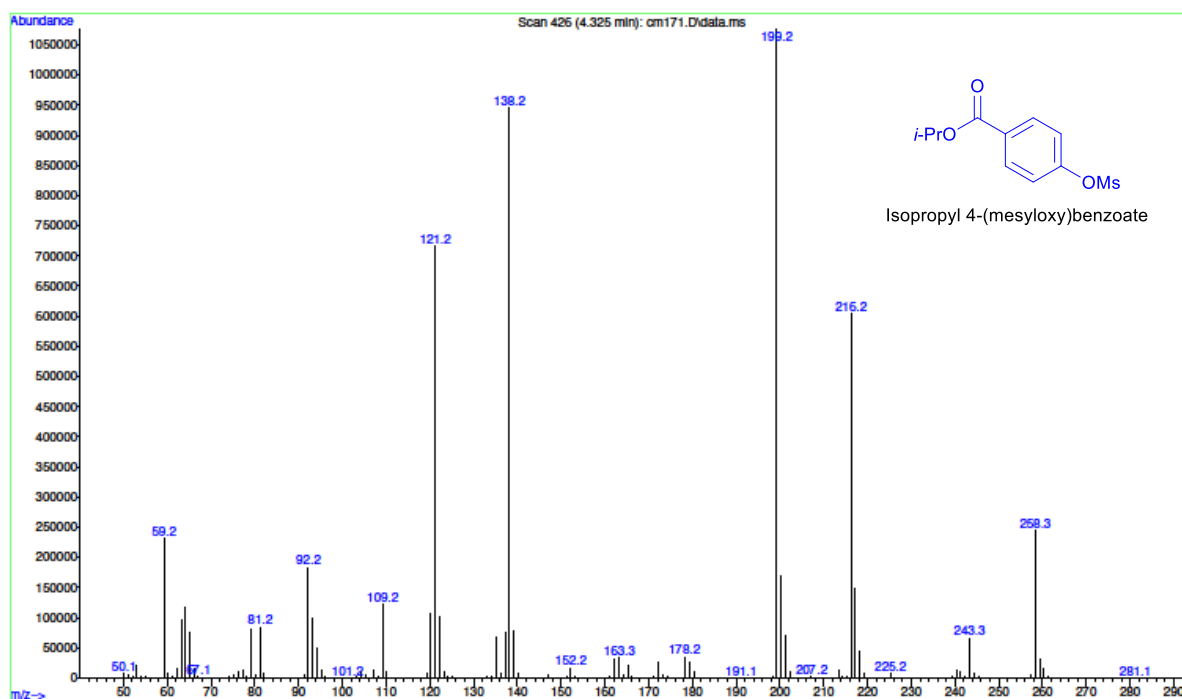


Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Formula
447.2913	447.2929	-1.6	-3.6	11.5	10.1	C ₂₉ H ₄₀ N ₂ P

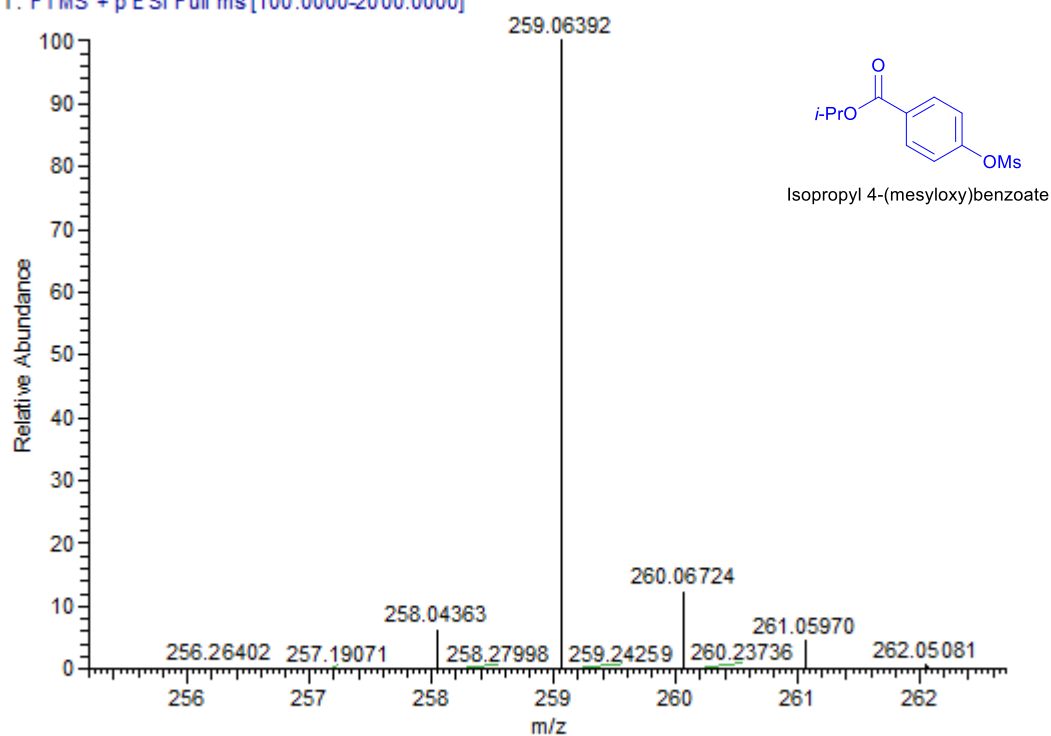




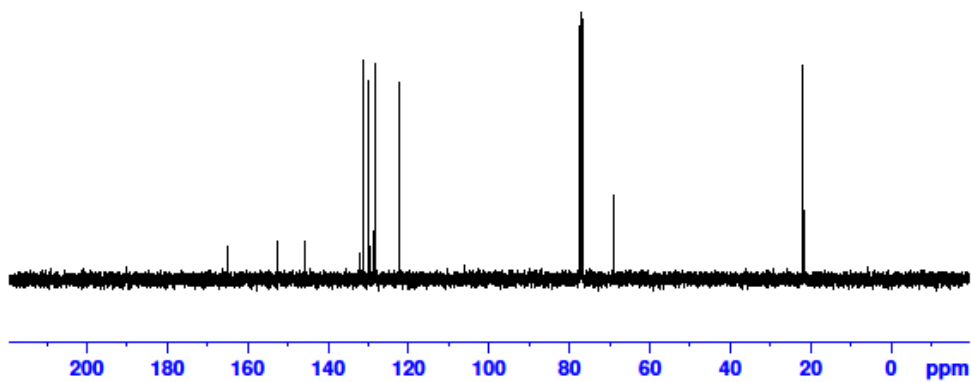
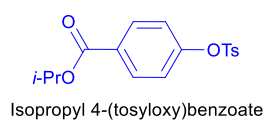
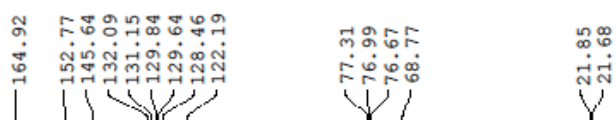
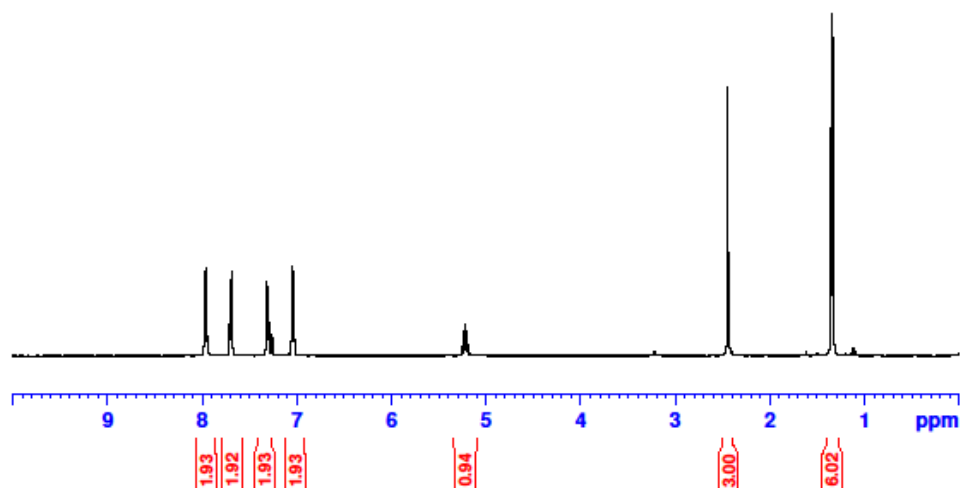
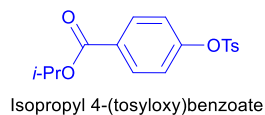


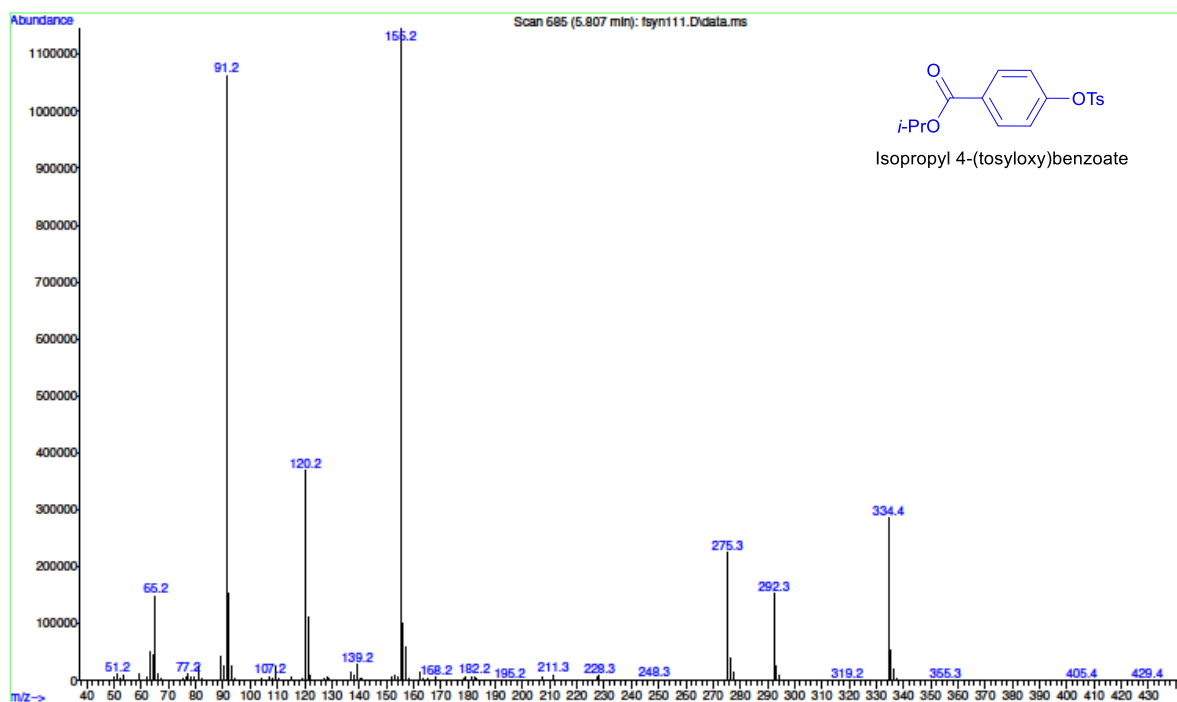


Kin-Dept 11012017 S6 #77-120 RT: 0.37-0.57 AV: 44 NL: 2.09E7
T: FTMS + p E SI Full ms [100.0000-2000.0000]

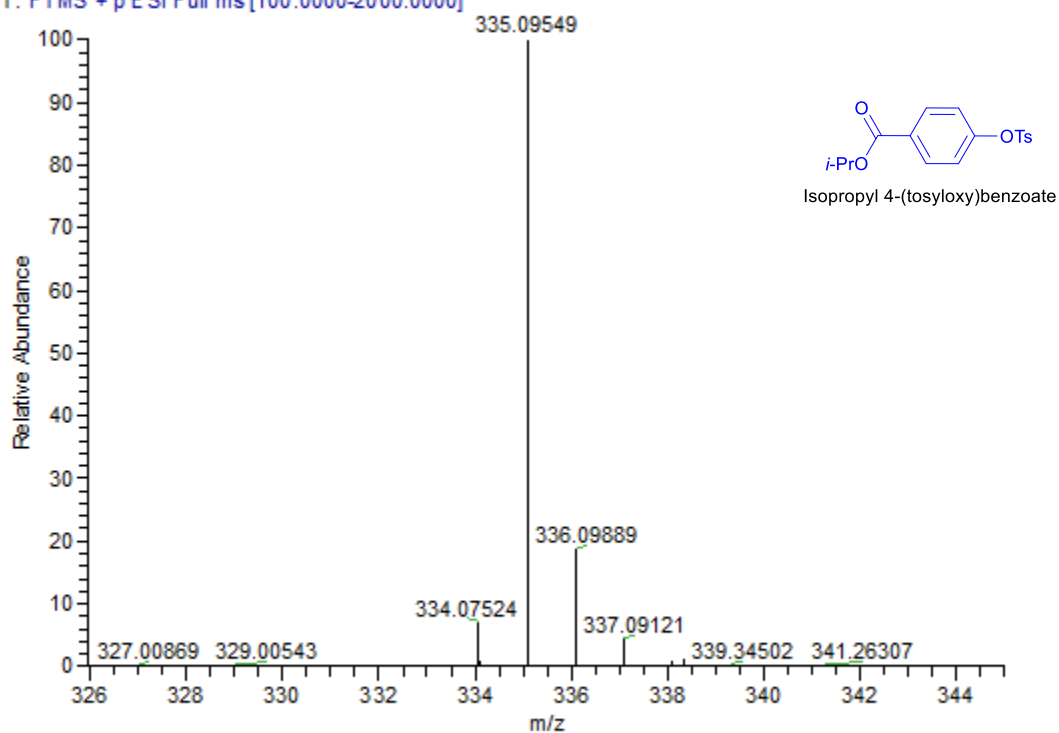


Mass	Calc. Mass	mDa	PPM	Formula
259.0639	259.0635	0.4	1.6	C ₁₁ H ₁₅ O ₅ S





Kin-Dept 11012017 S7 #77-131 RT: 0.37-0.63 AV: 55 NL: 4.45E8
T: FTMS + p ESI Full ms [100.0000-2000.0000]



Mass	Calc. Mass	mDa	PPM	Formula
335.0955	335.0948	0.7	2.2	C17 H19 O5 S

7.5771
7.5563
7.5422
7.5220
7.5050
7.5004
7.4842
7.2866
7.2837
7.2670

2.4319

1.4063

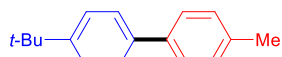
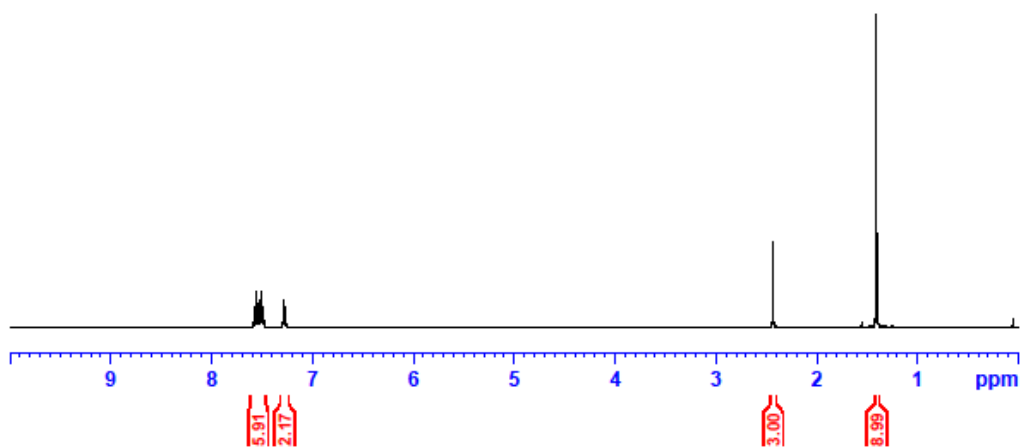


Table 3, 3aa



149.92
138.24
138.19
136.67
129.40
126.84
126.58
125.64

77.31
77.00
76.68

34.49
31.37
21.07

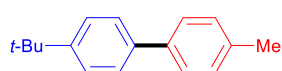
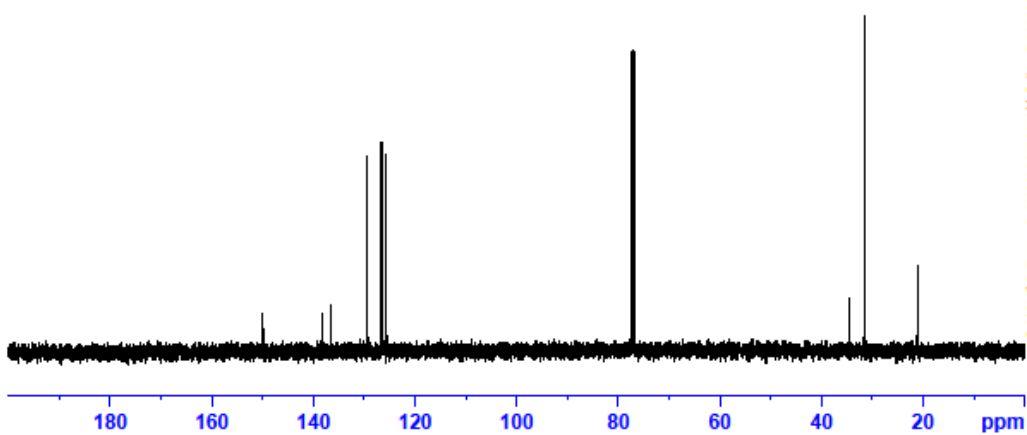
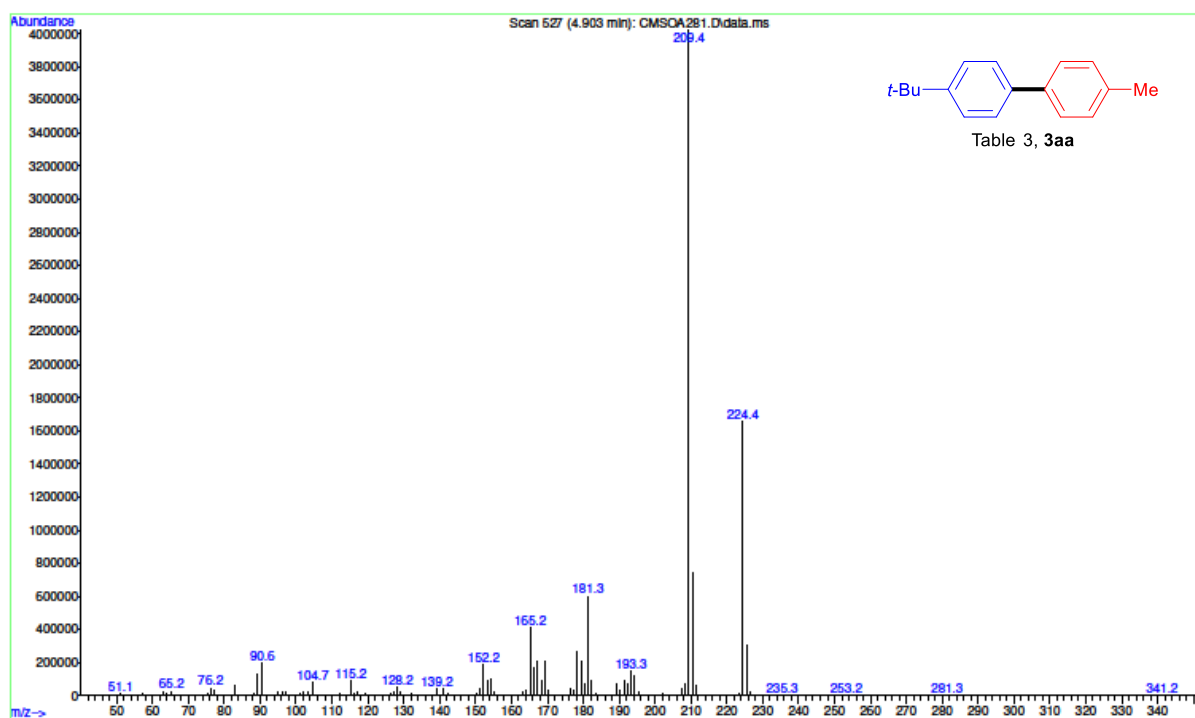
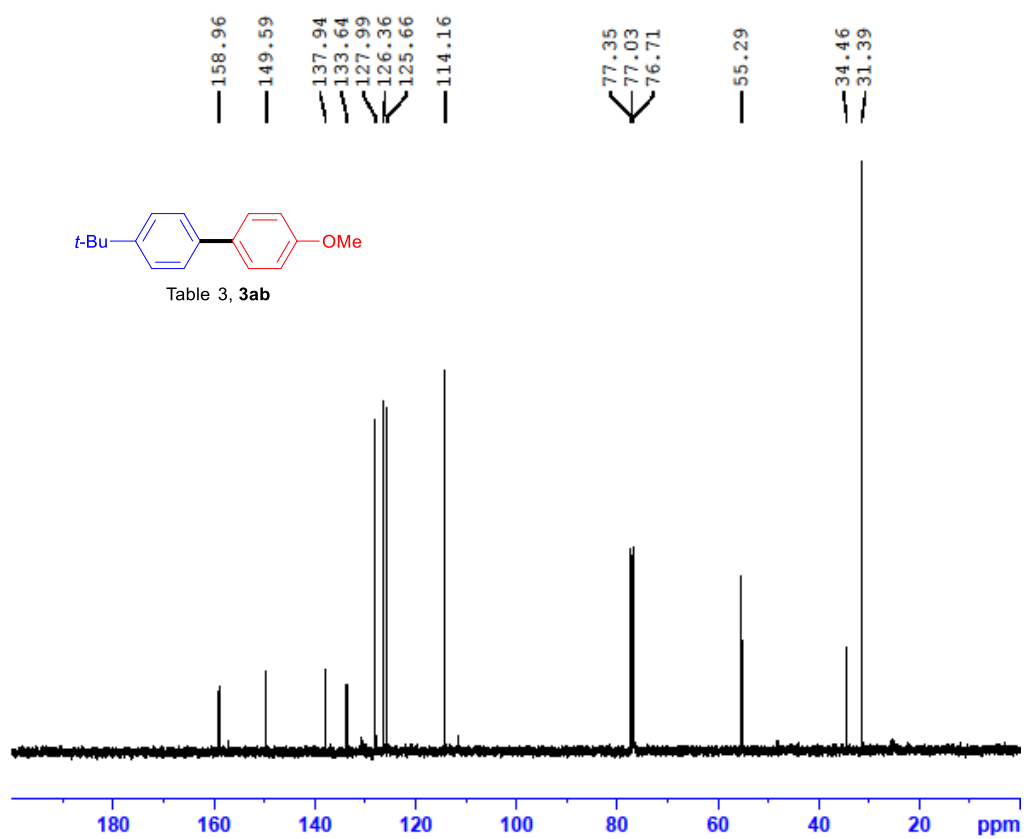
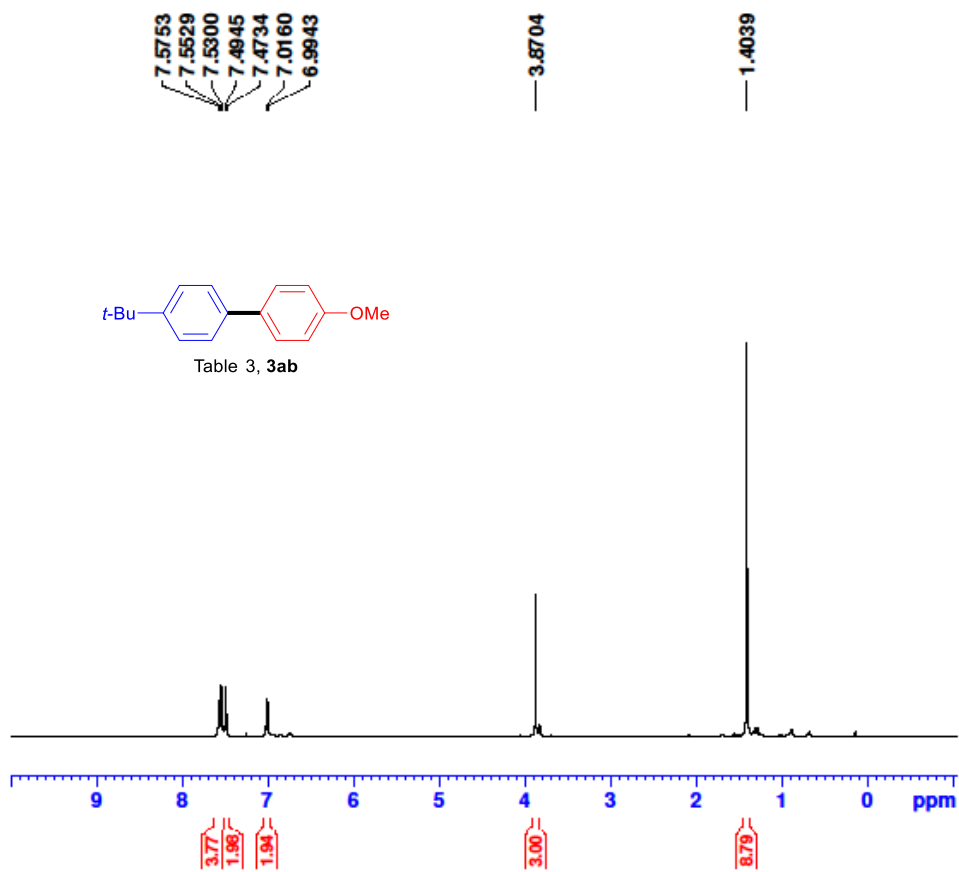
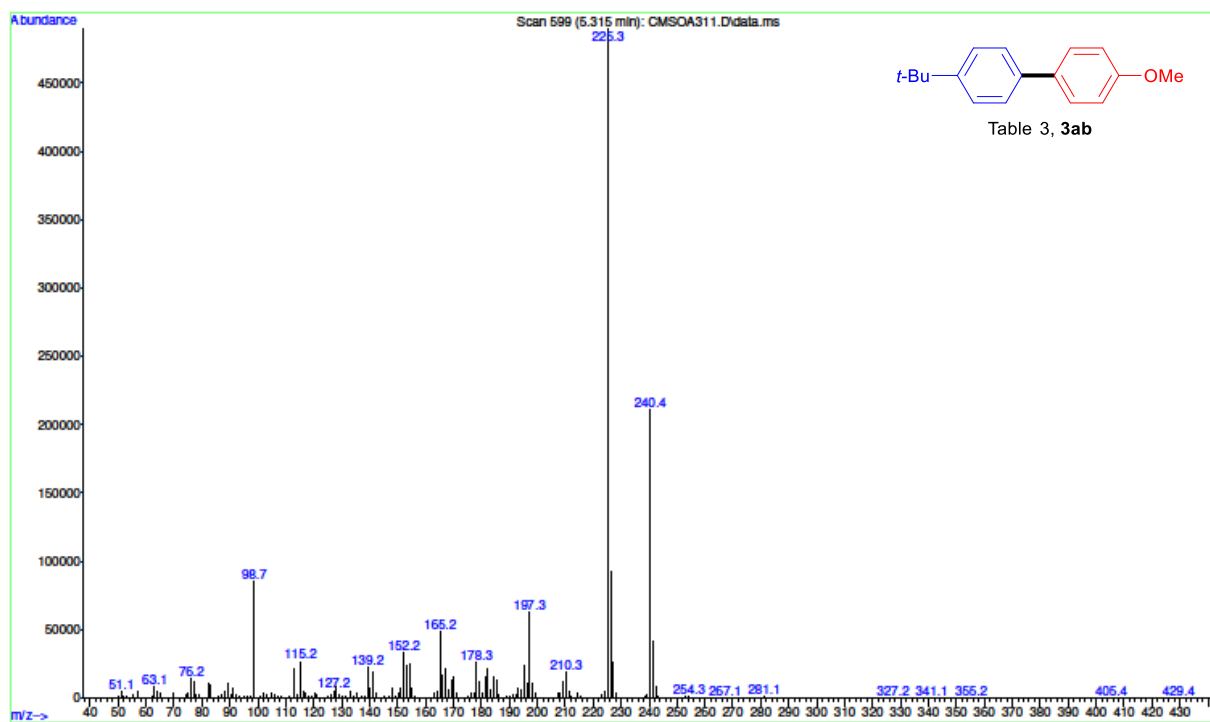


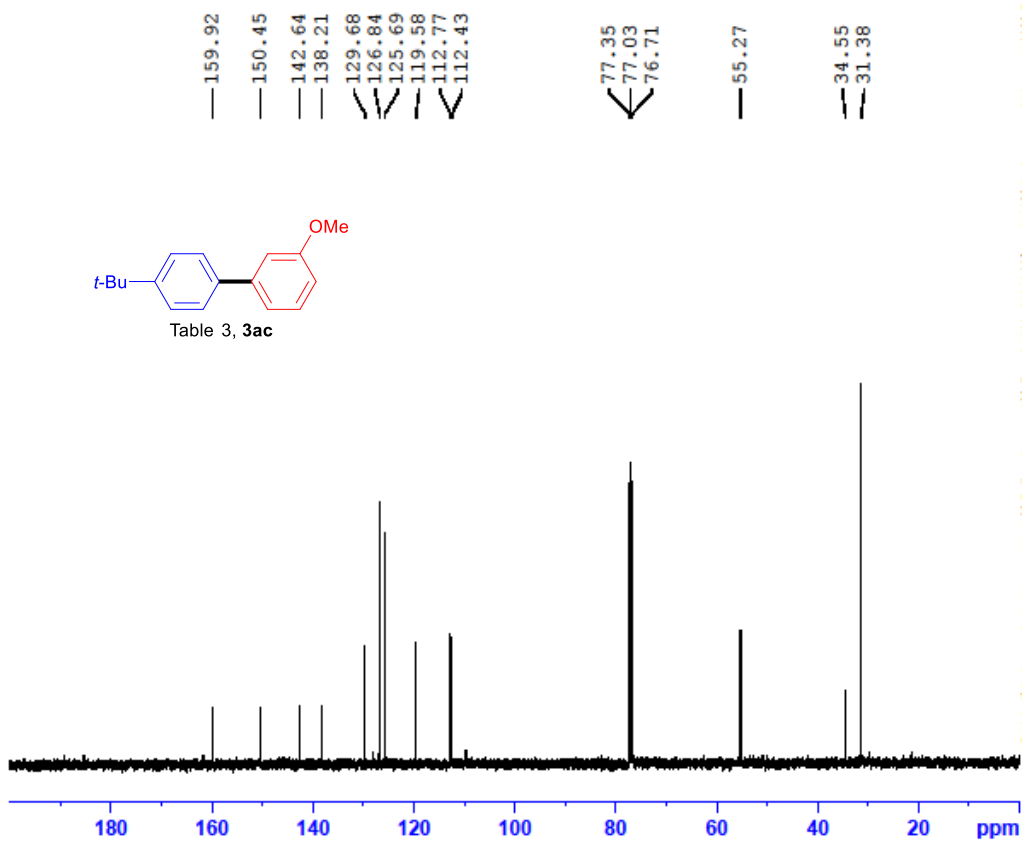
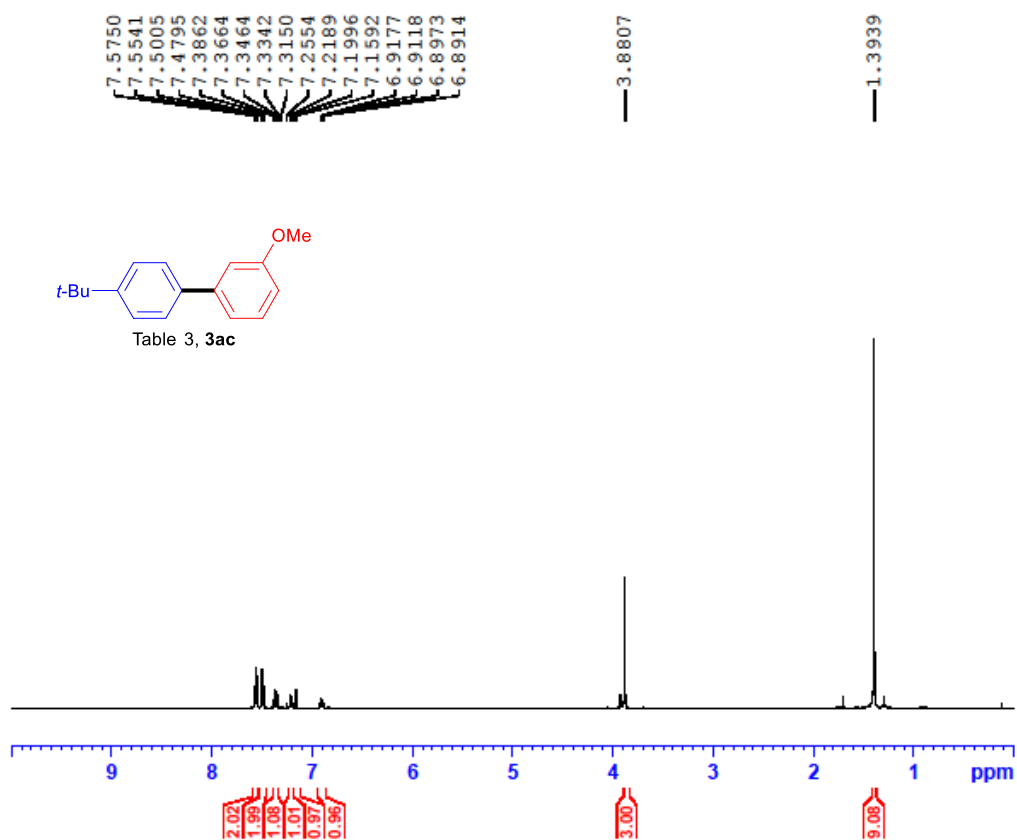
Table 3, 3aa

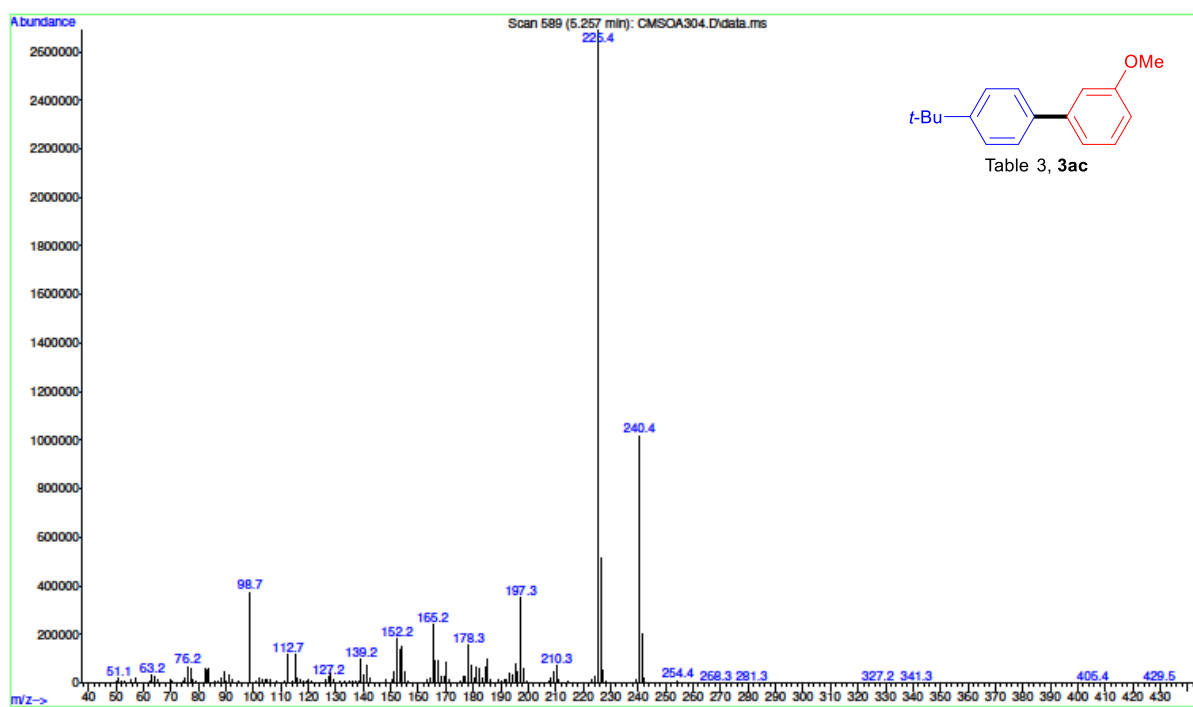


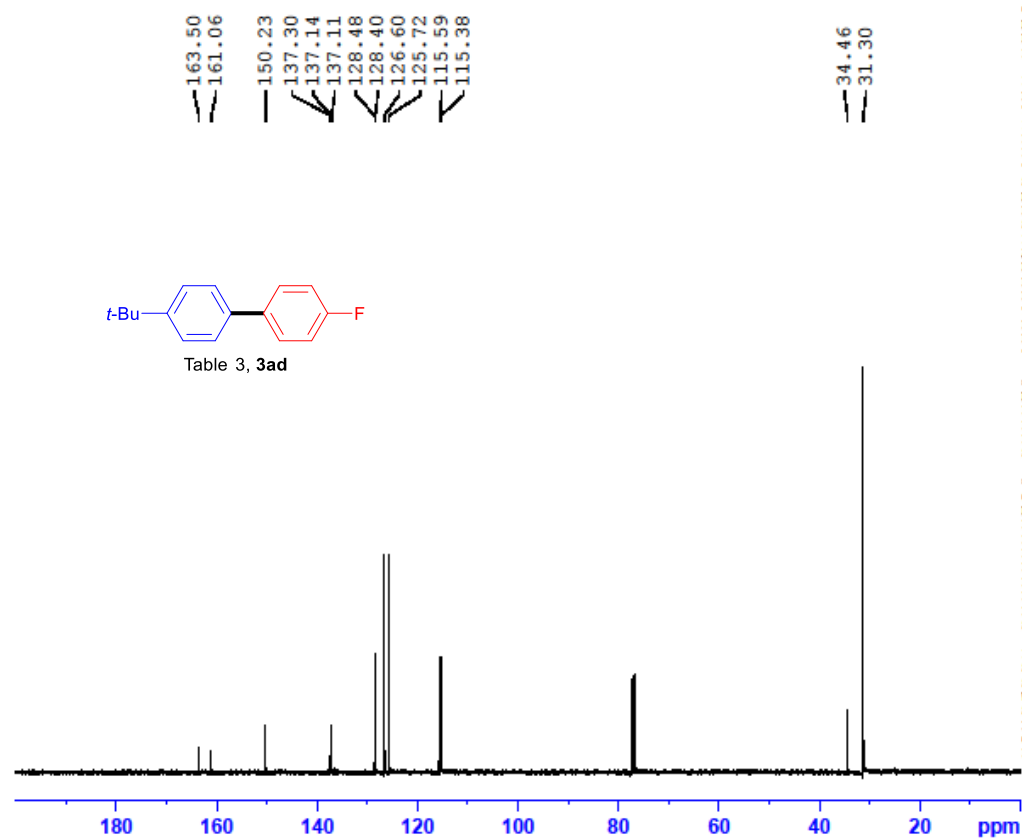
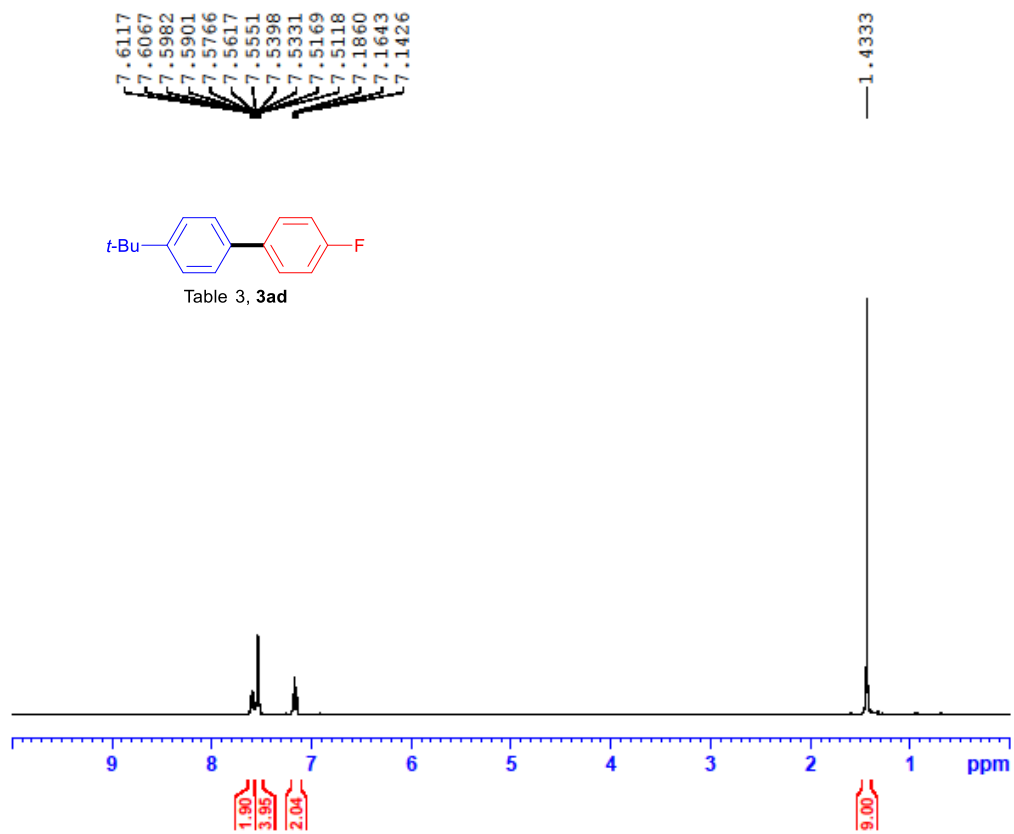












-116.06

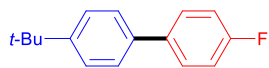
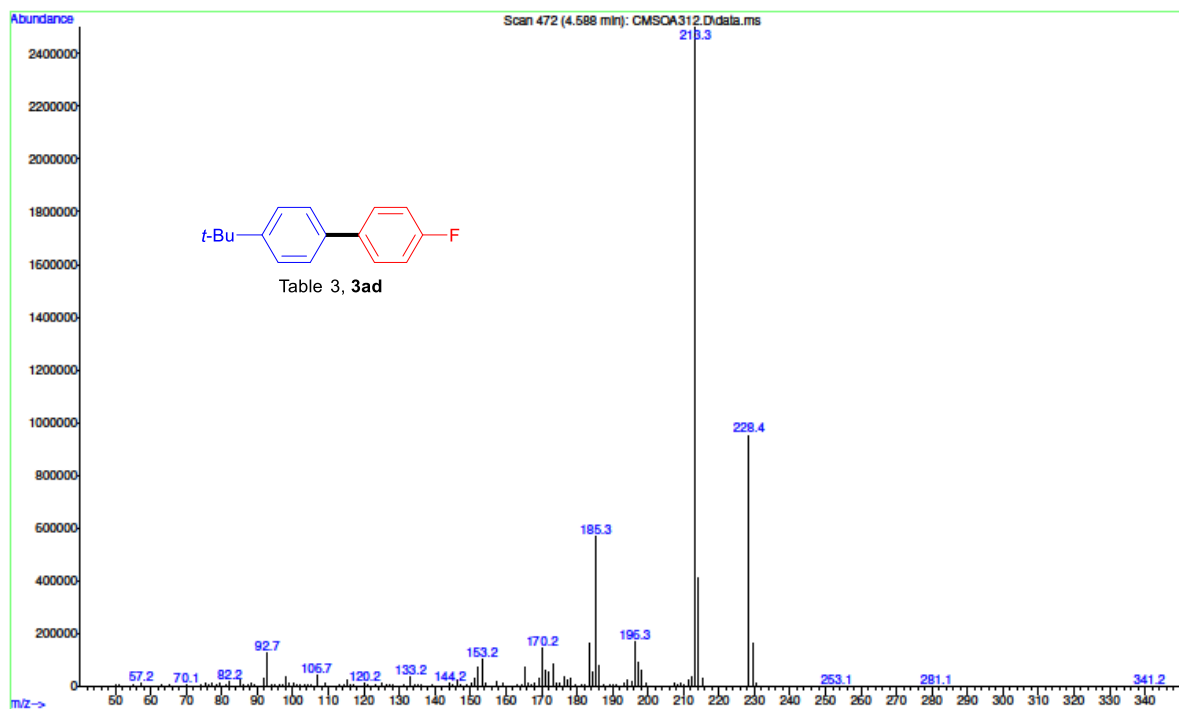
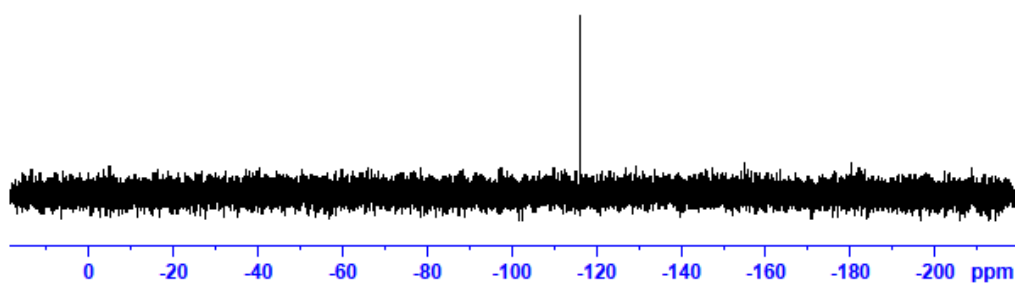
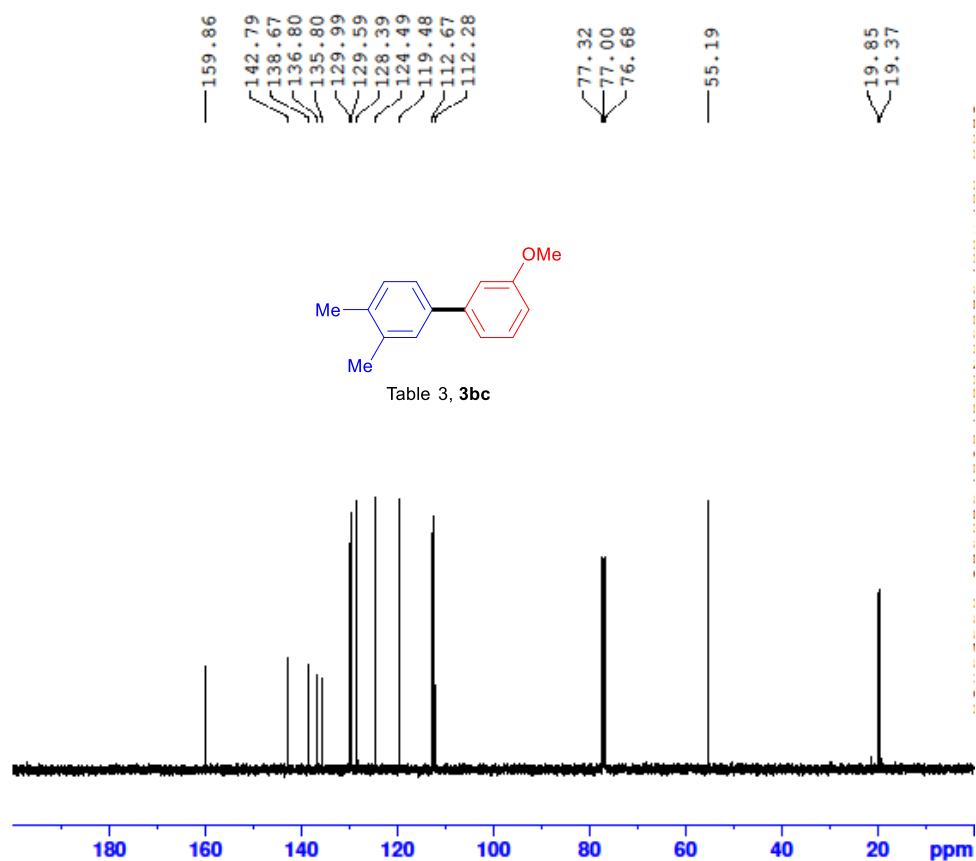
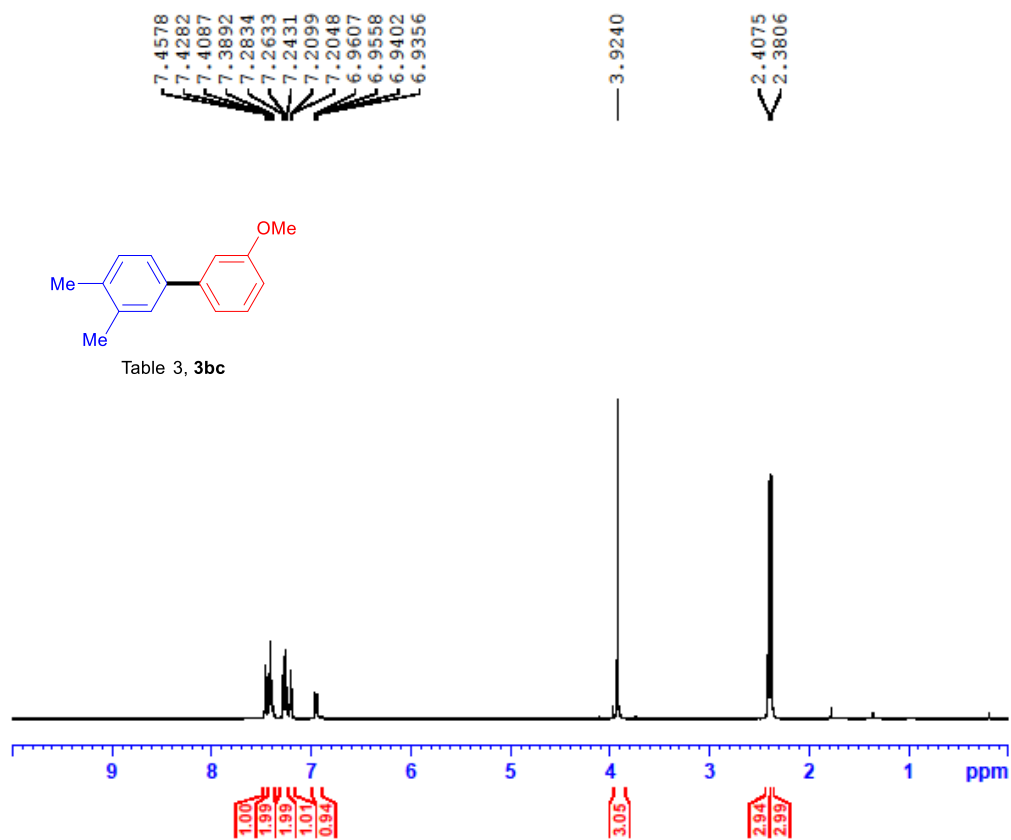
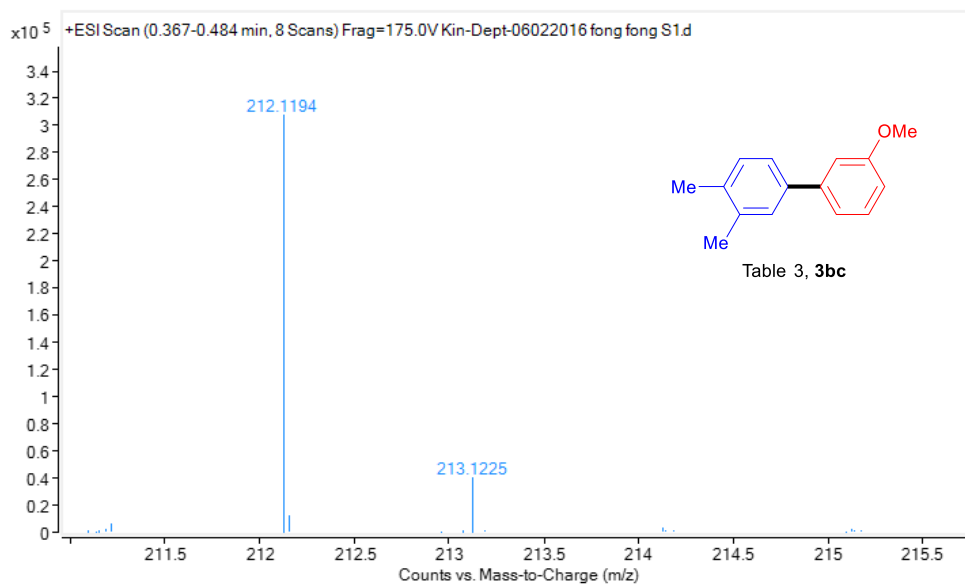
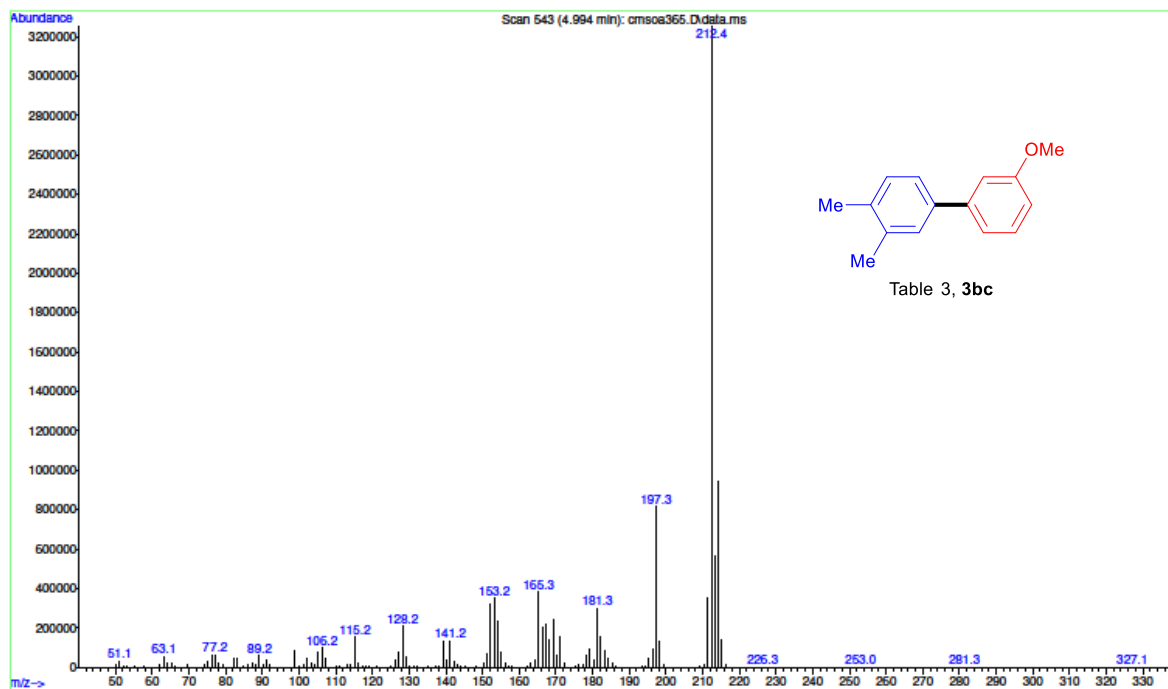


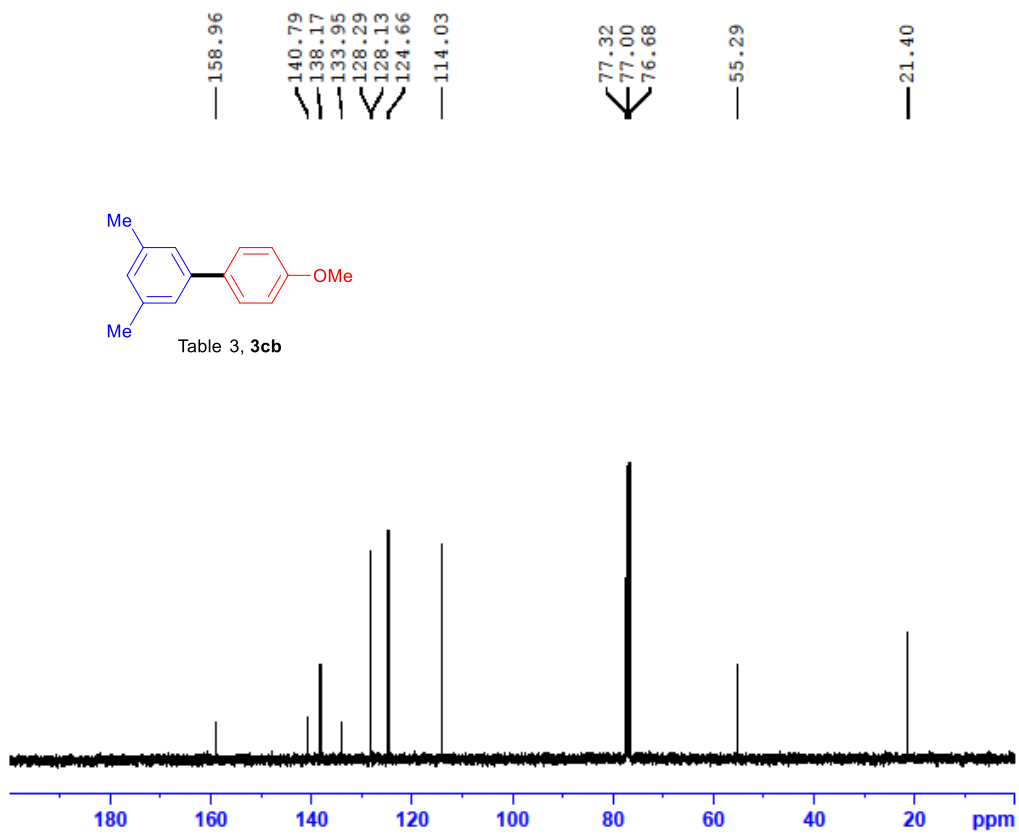
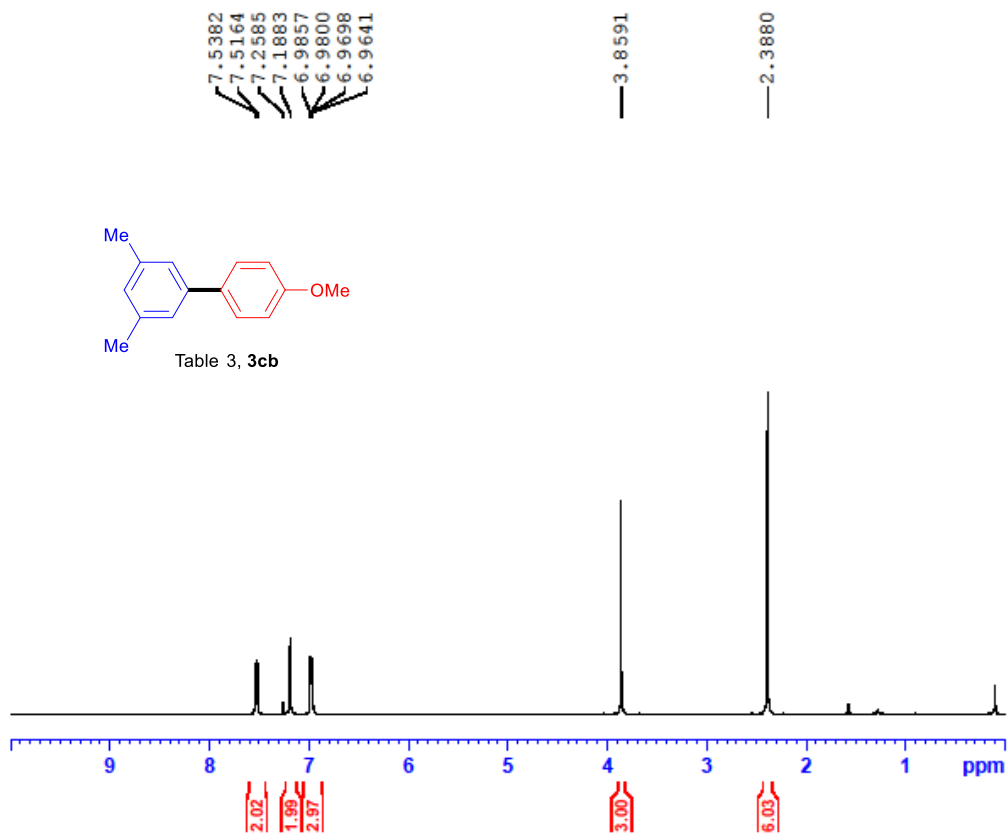
Table 3, 3ad

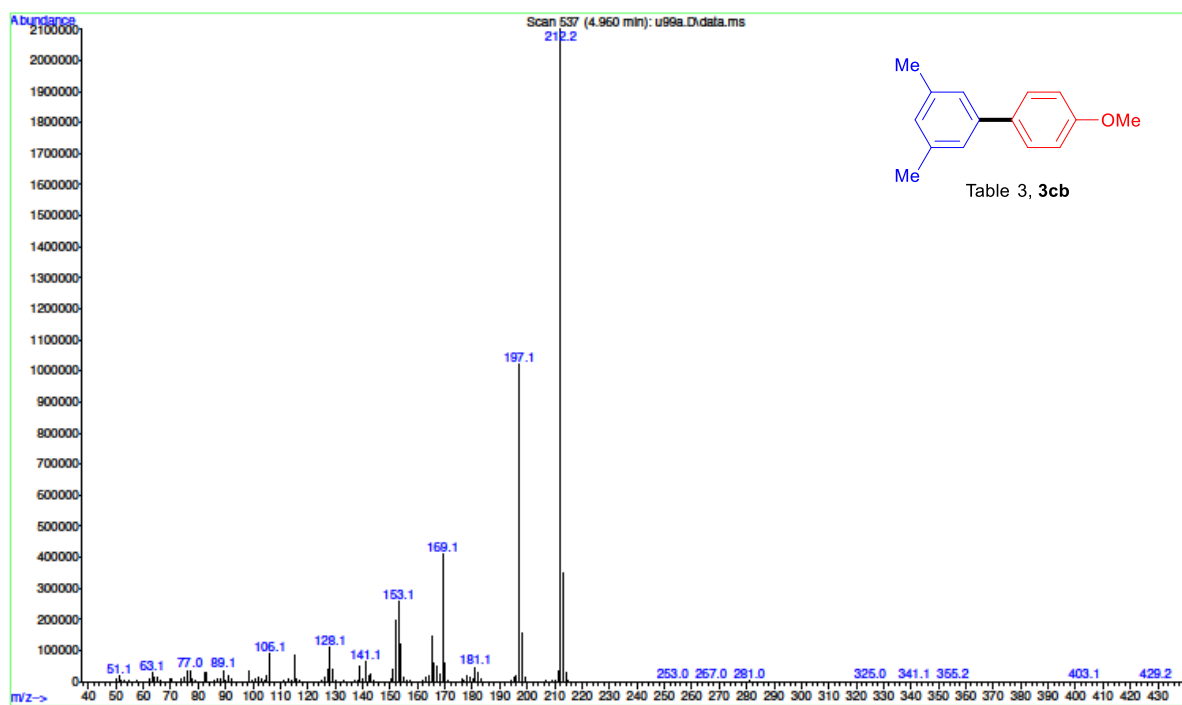


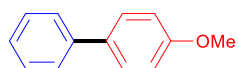
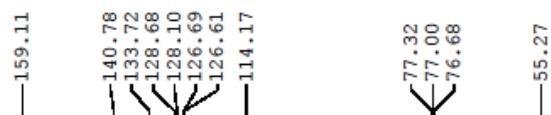
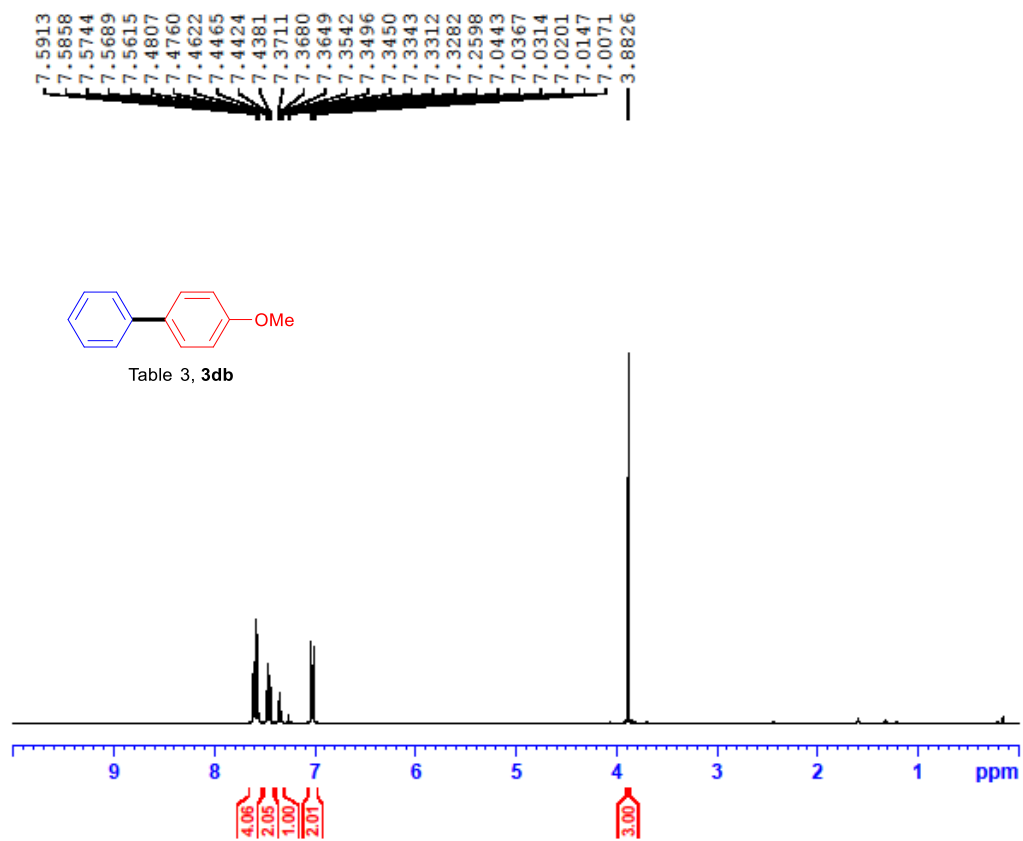
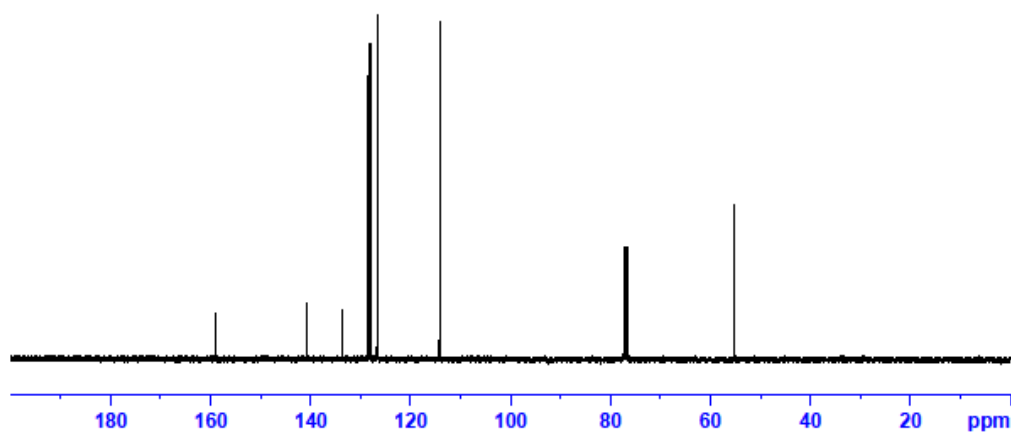


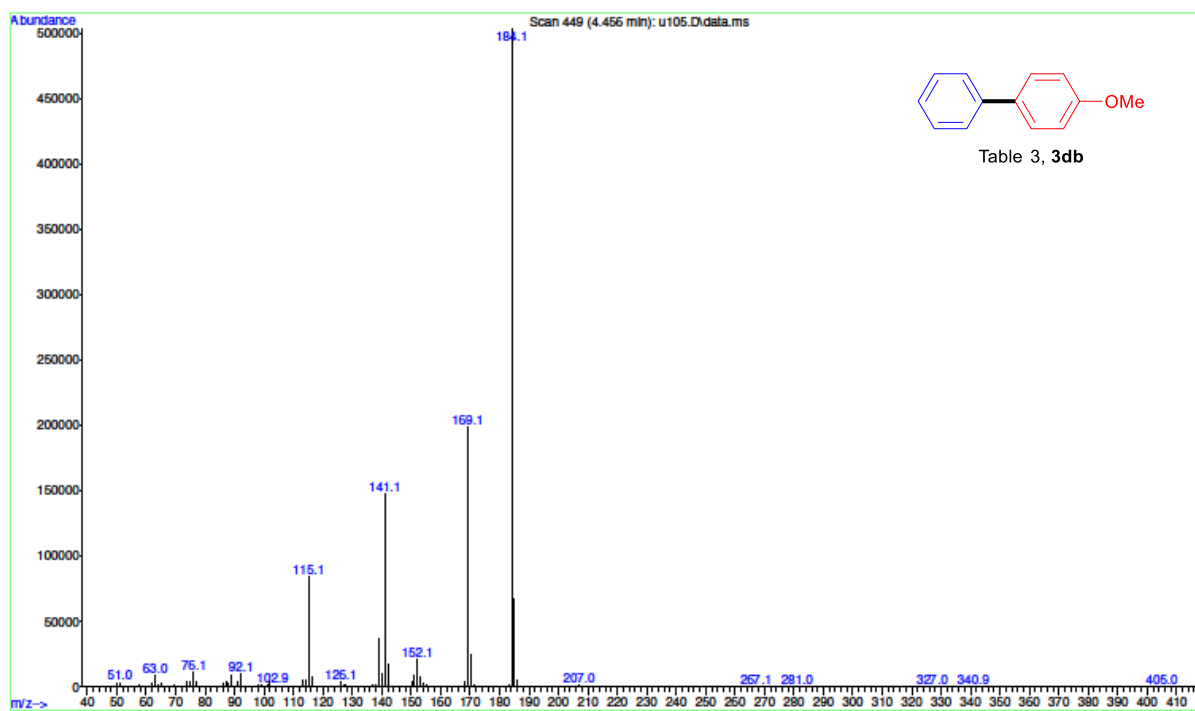


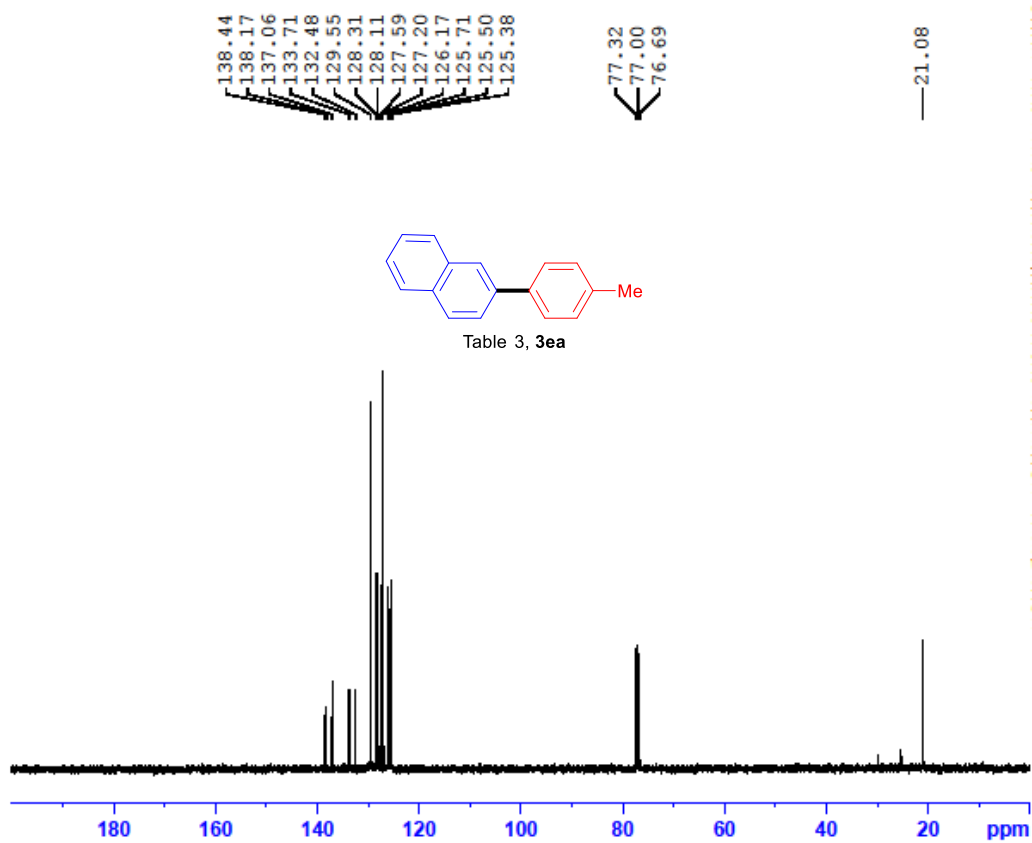
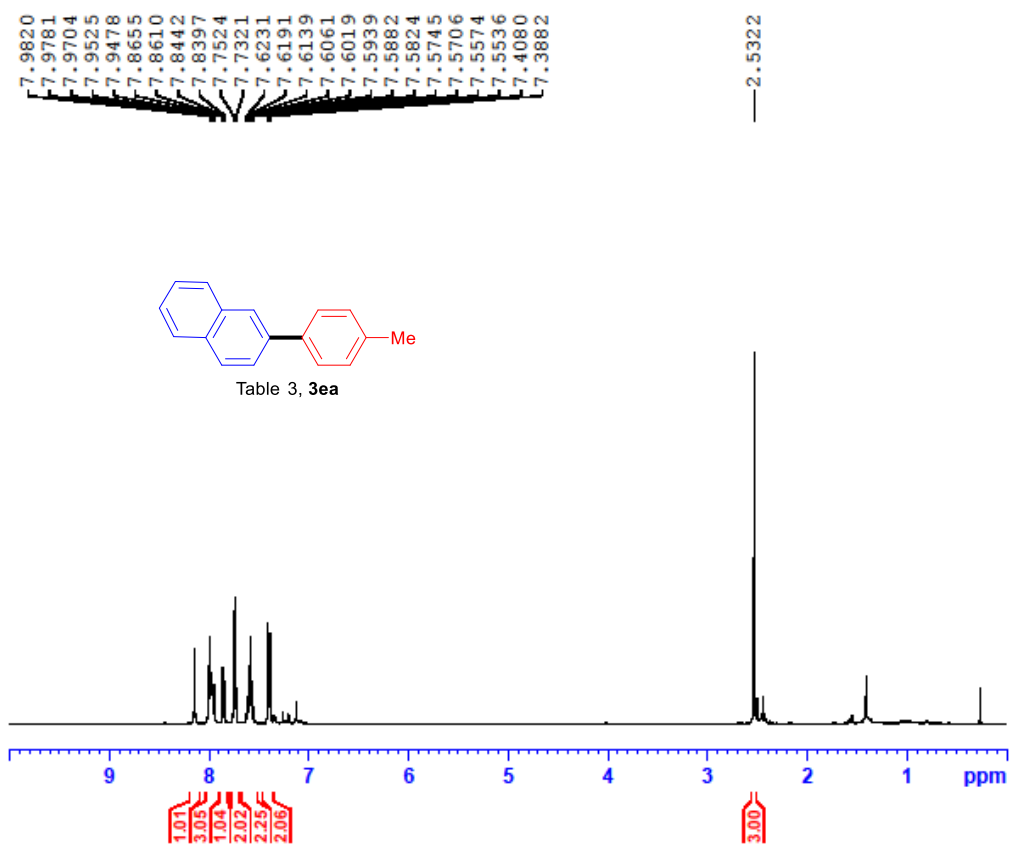
Mass	Calc. Mass	mDa	PPM	Formula
212.1194	212.1196	0.2	0.8	C ₁₅ H ₁₆ O

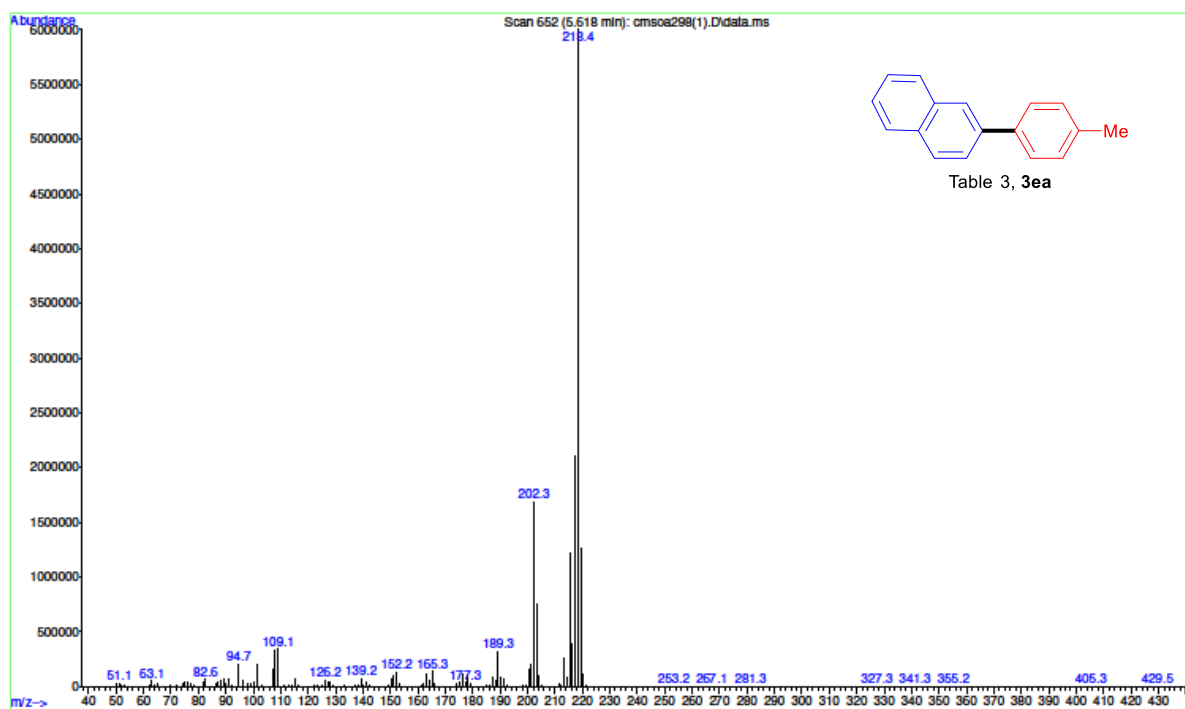


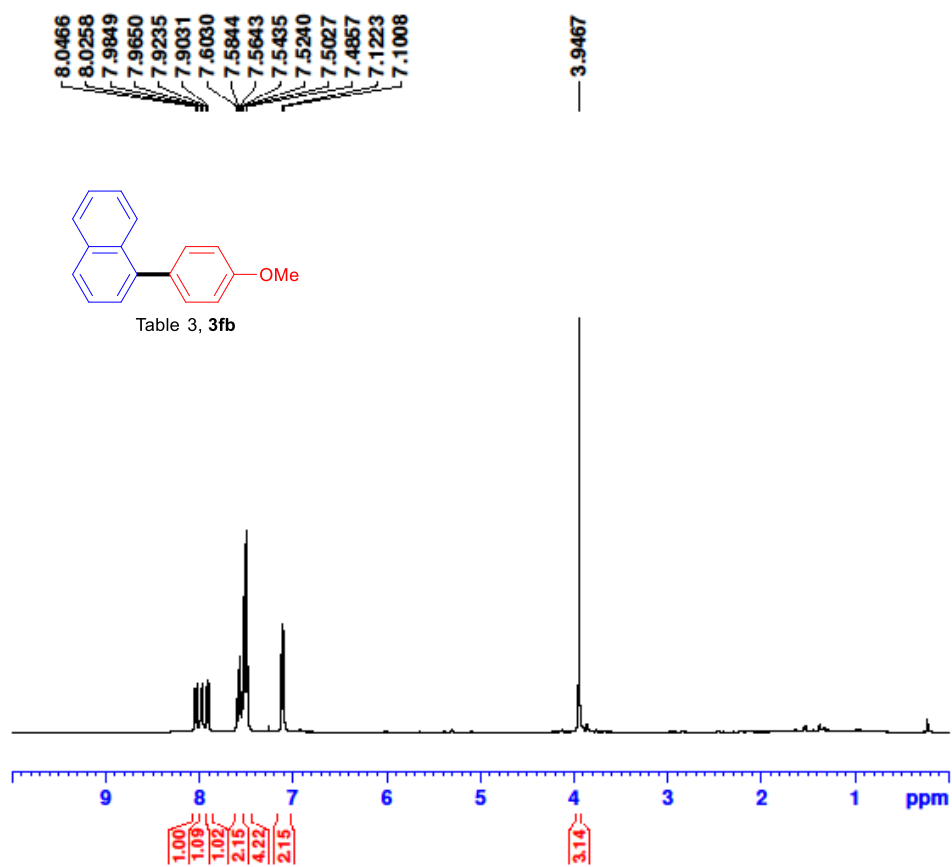


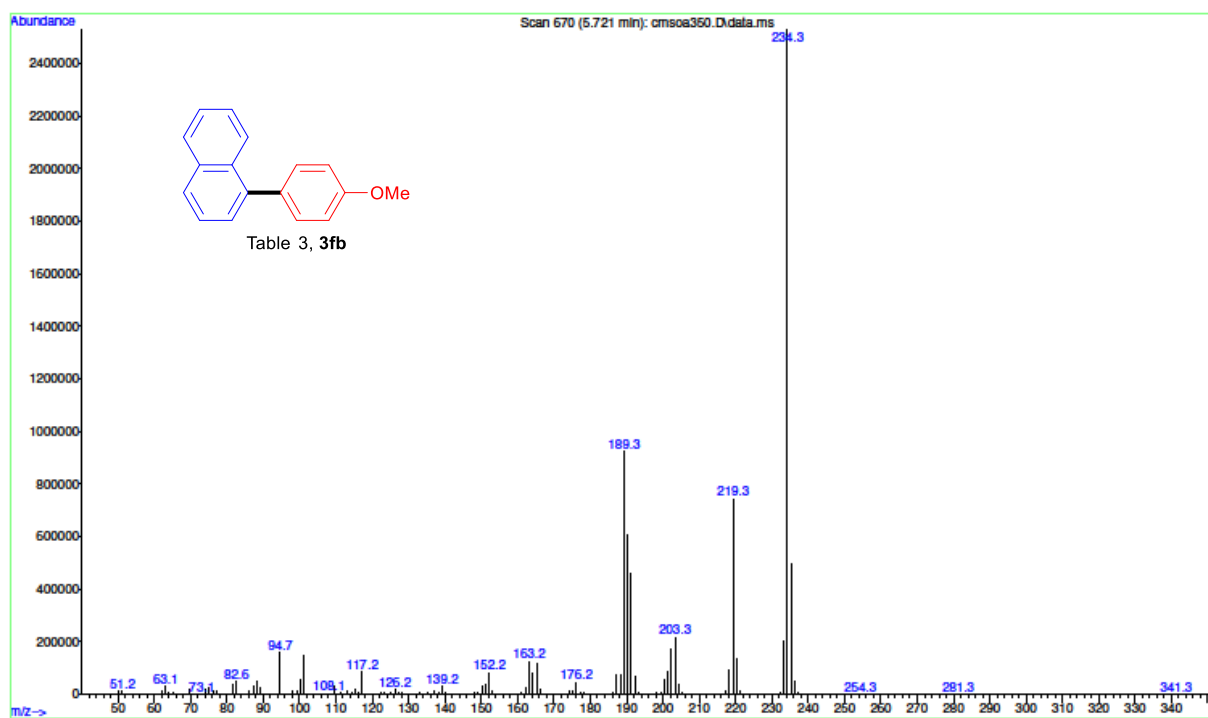
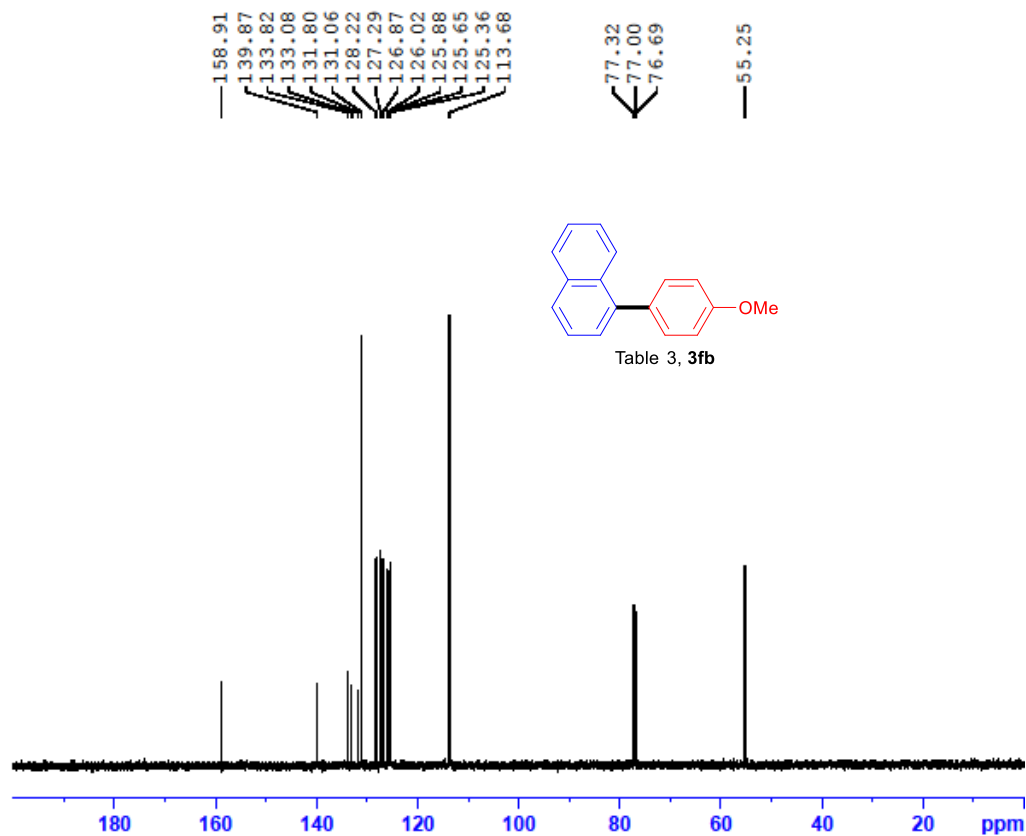
Table 3, **3db**

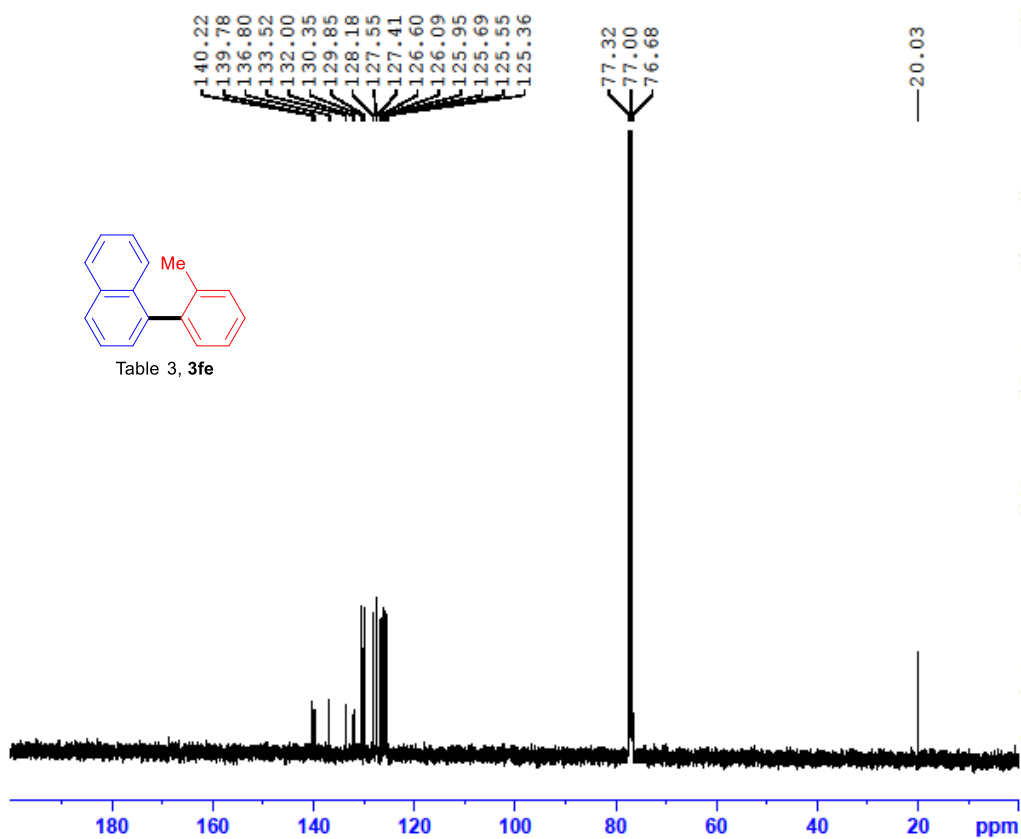
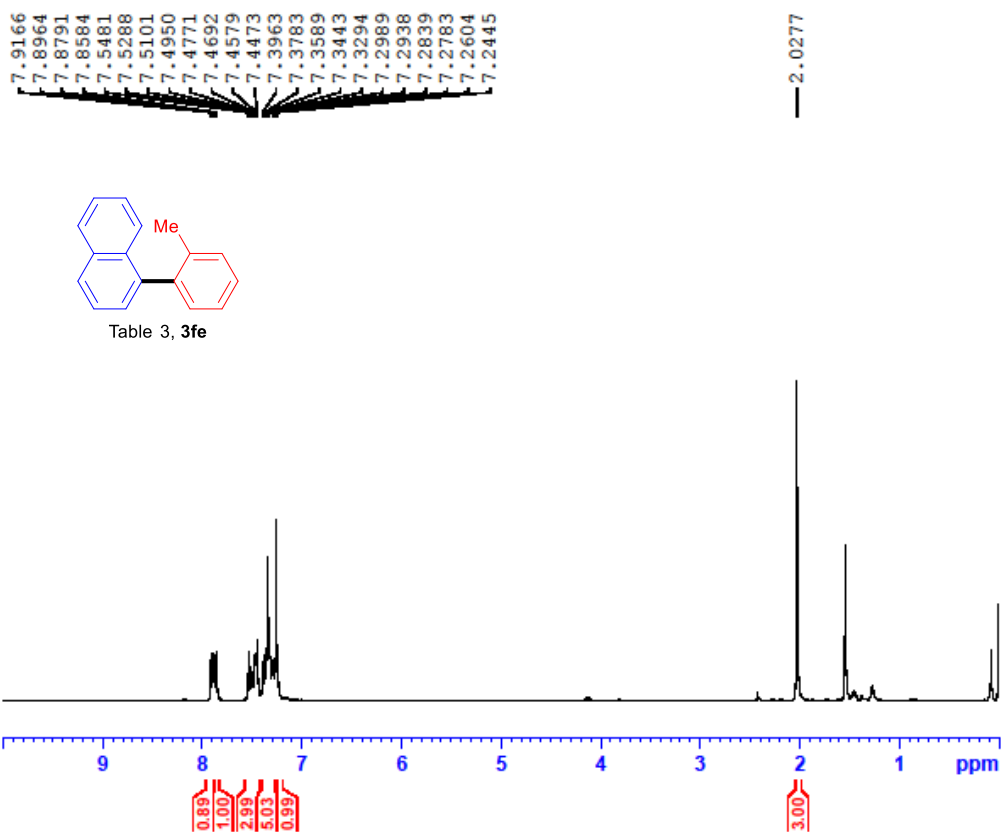


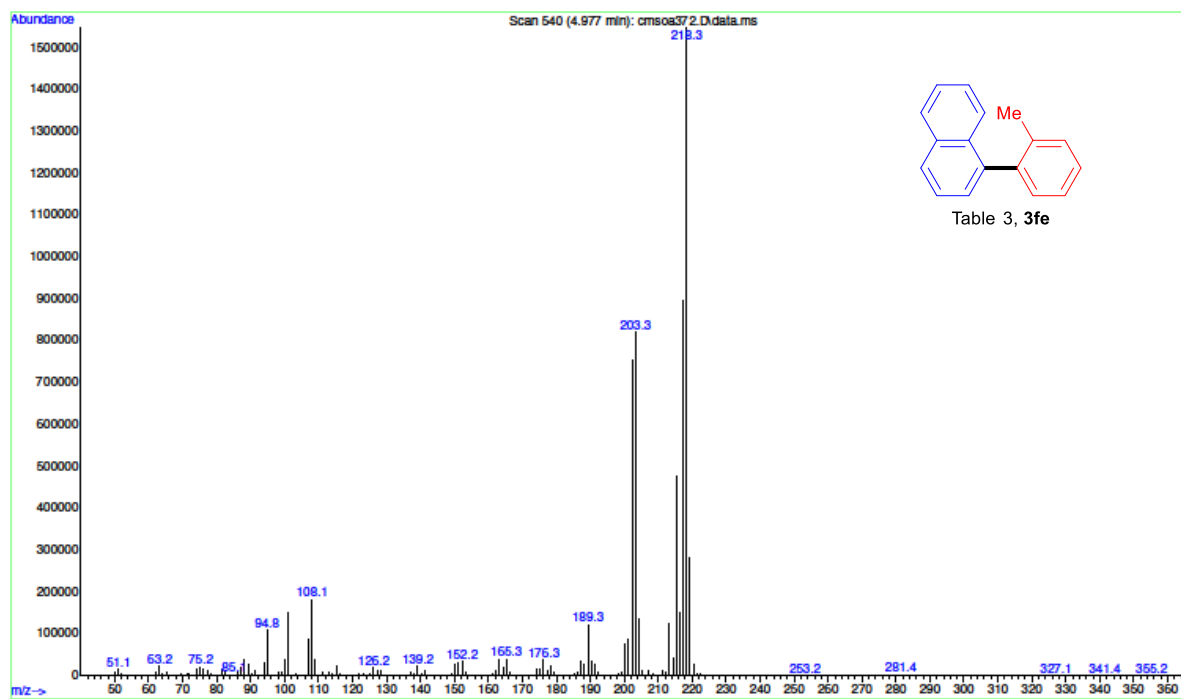












7.4916
7.4754
7.4712
7.2881
7.2684
7.2601
7.1359
7.1319
7.1217
7.1173
7.1017
7.0973
6.9406
6.9206
6.0229

— 2.4486

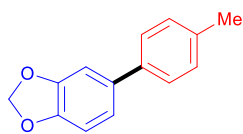
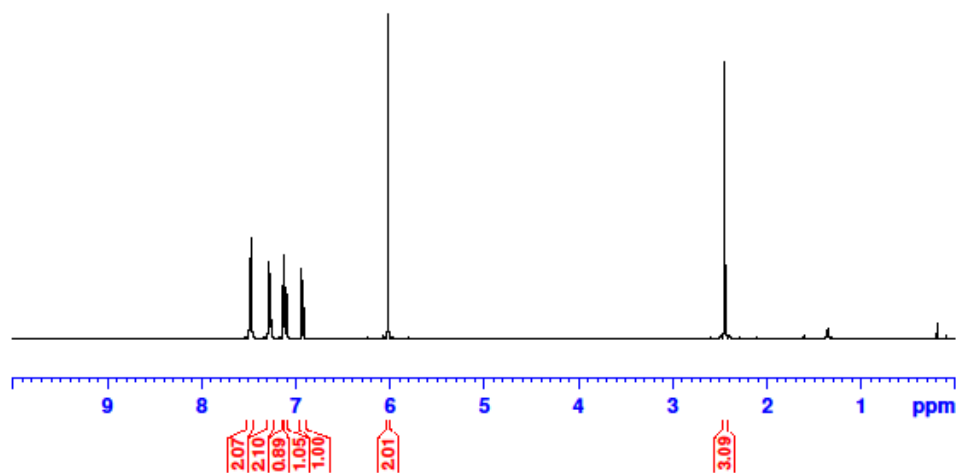


Table 3, 3ga



148.01
146.77
138.01
136.55
135.51
129.37
126.65
120.27

108.44
107.46
100.99

77.31
77.00
76.68

— 20.96

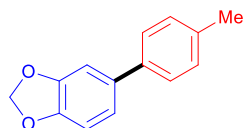
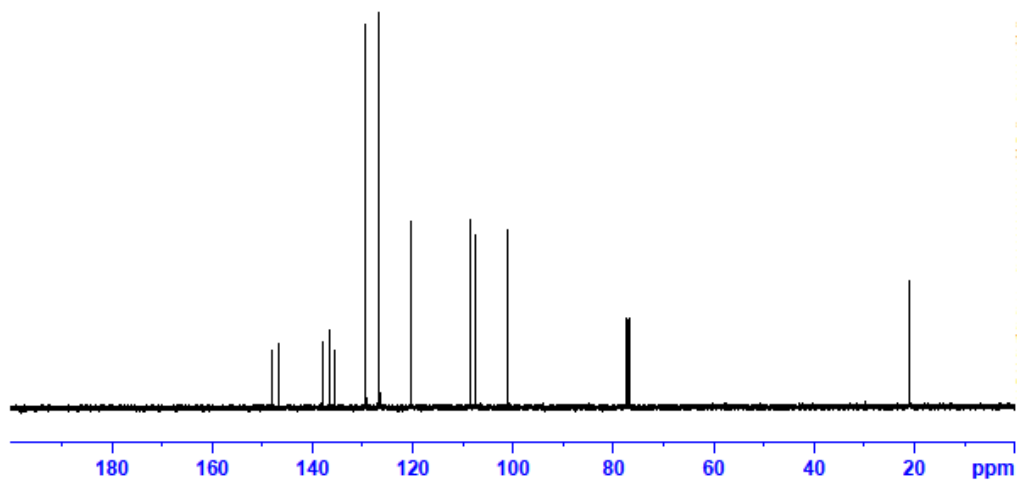
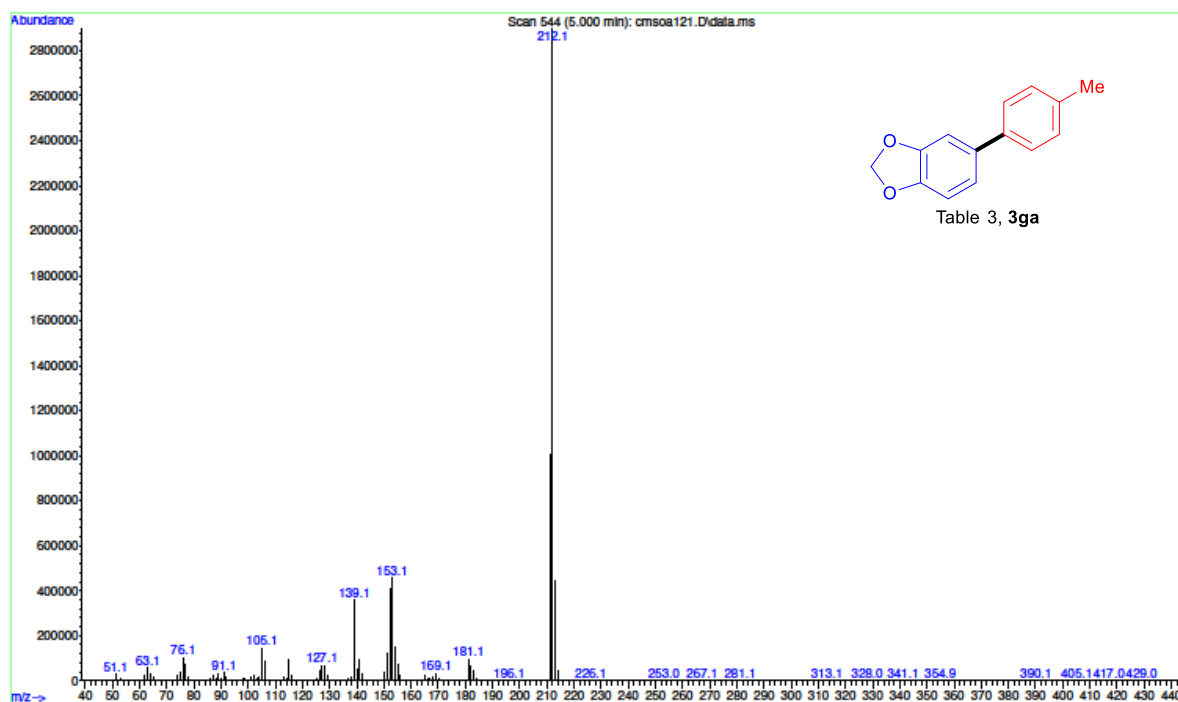
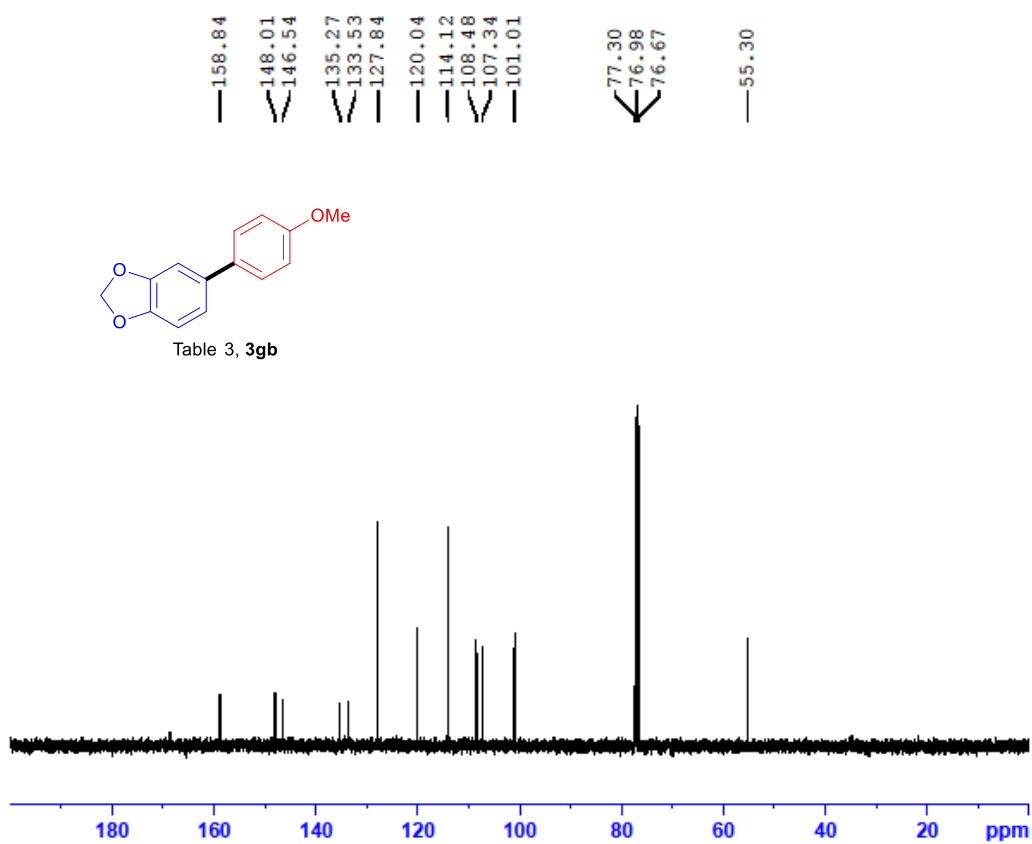
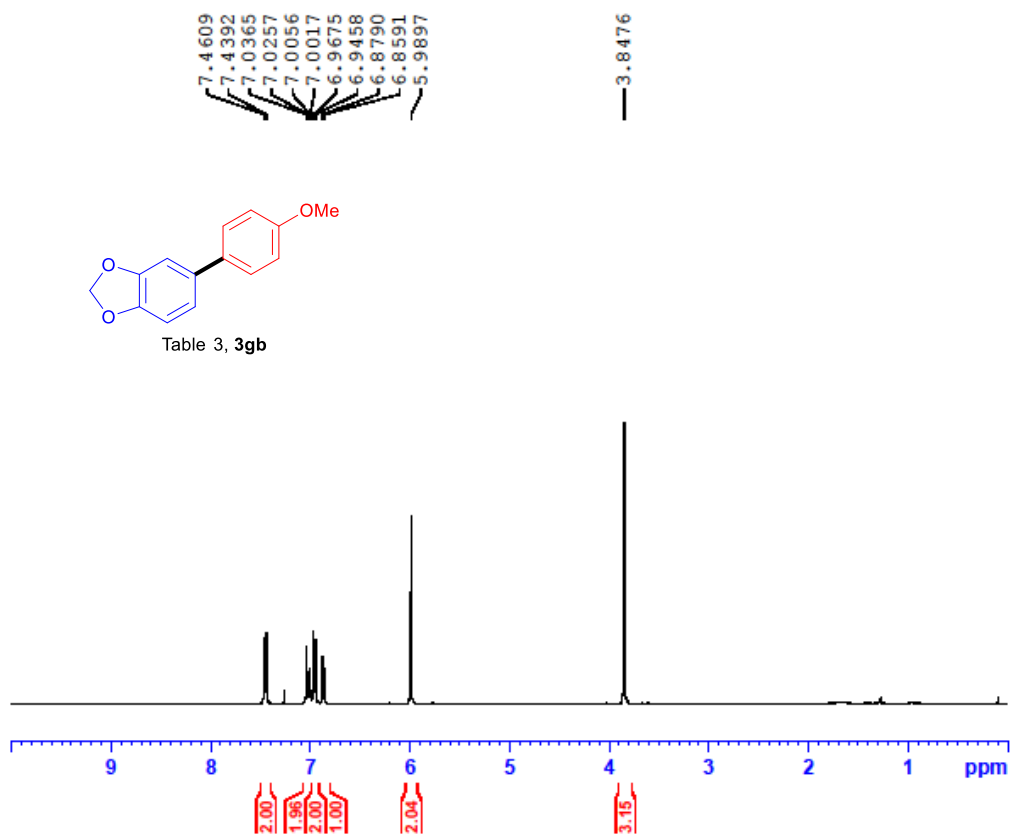
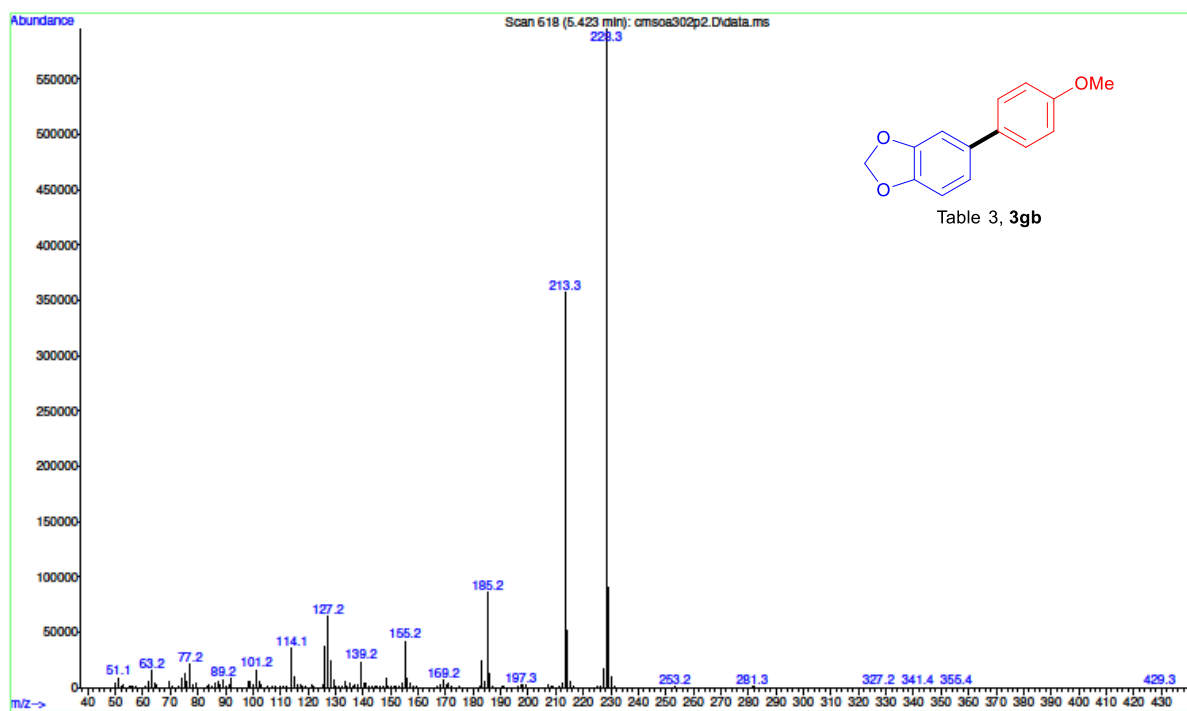


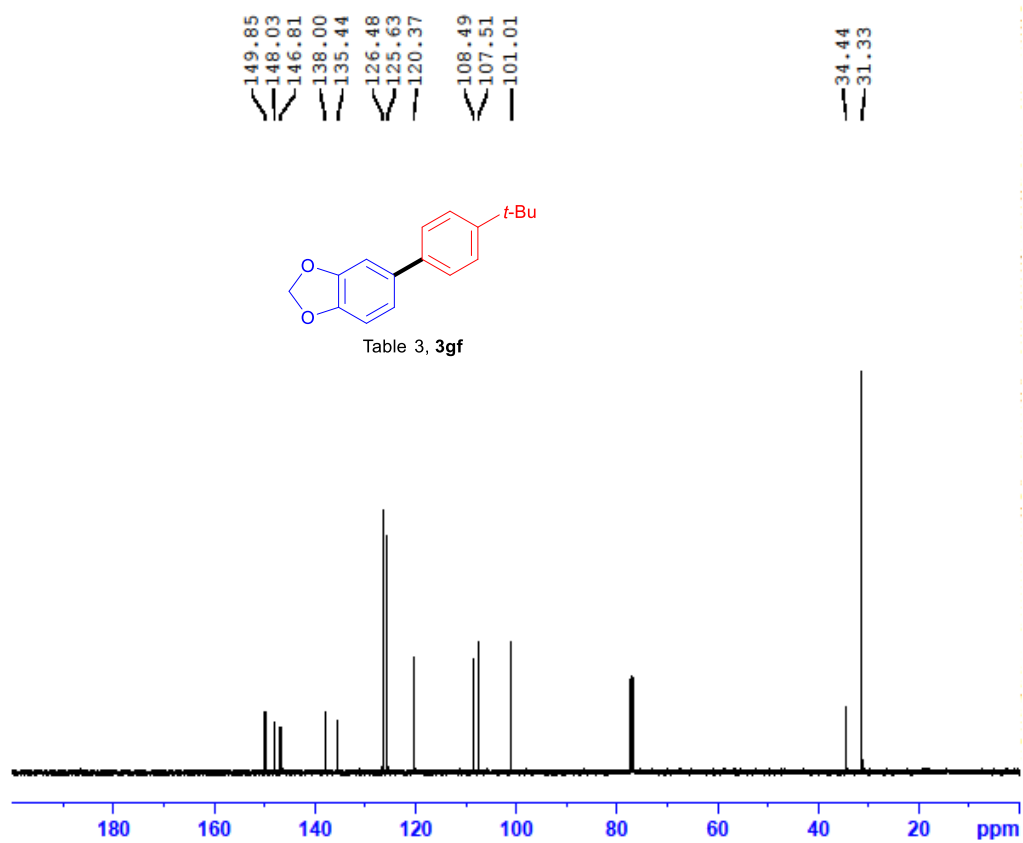
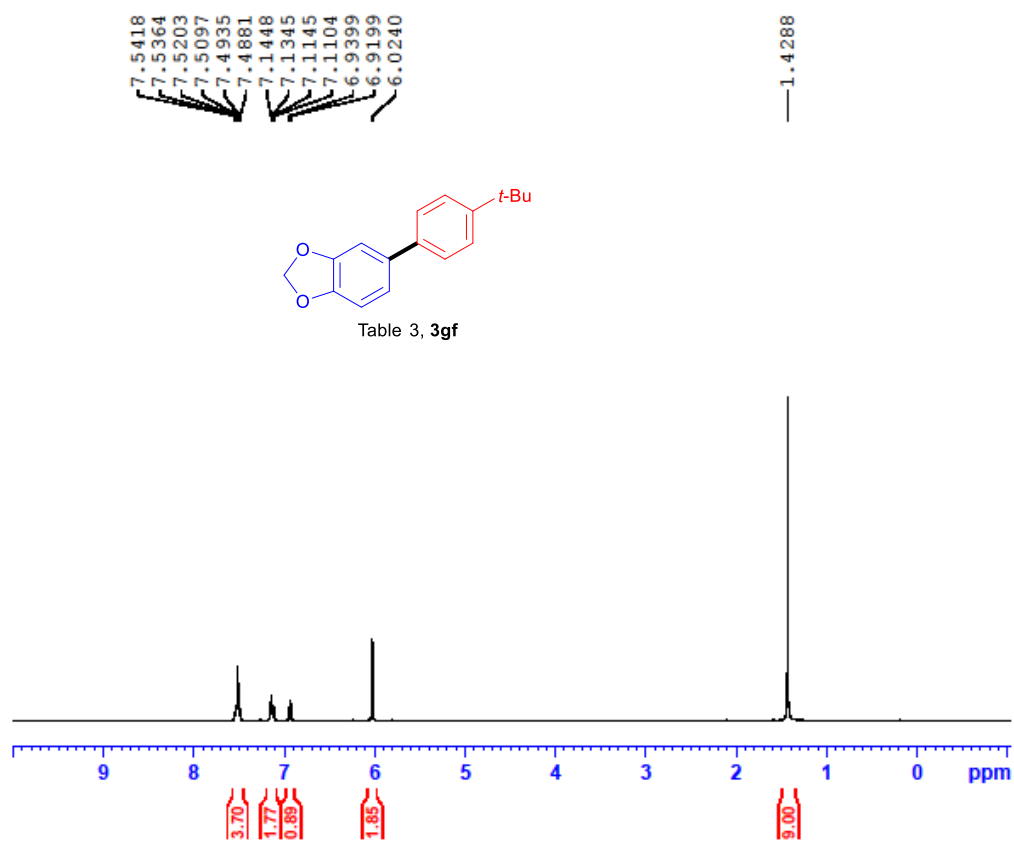
Table 3, 3ga

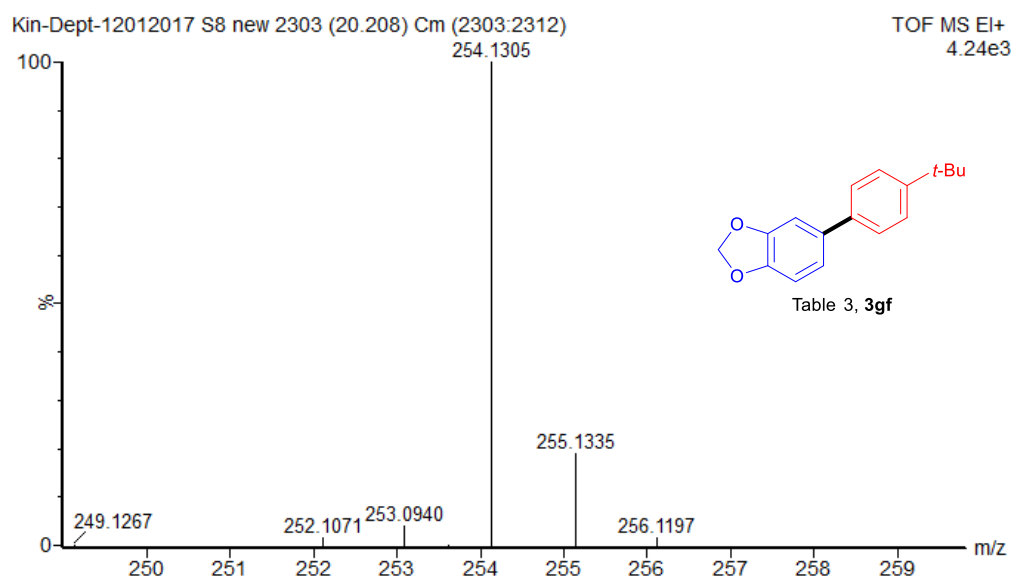
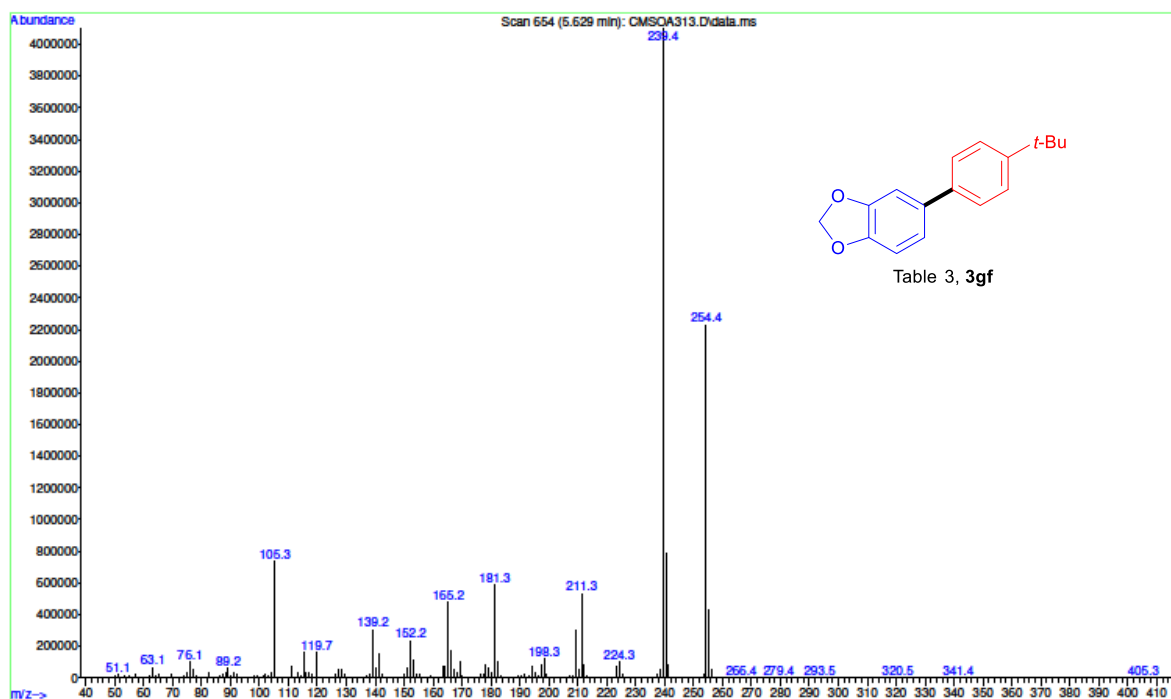




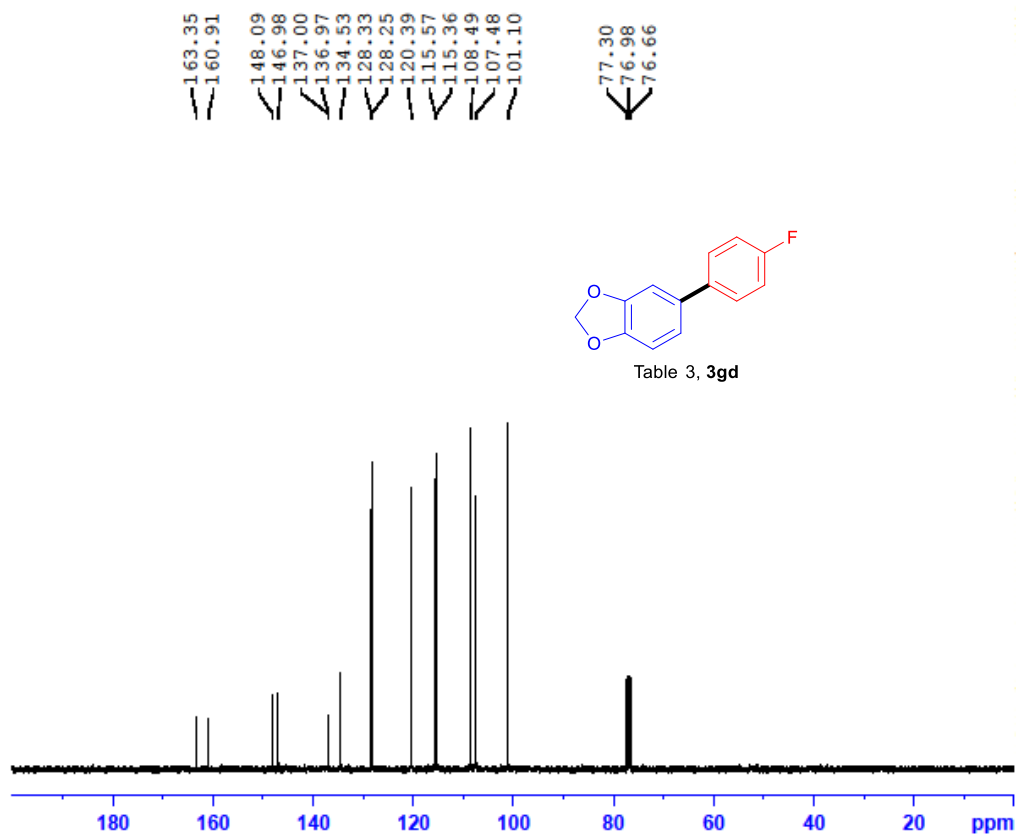
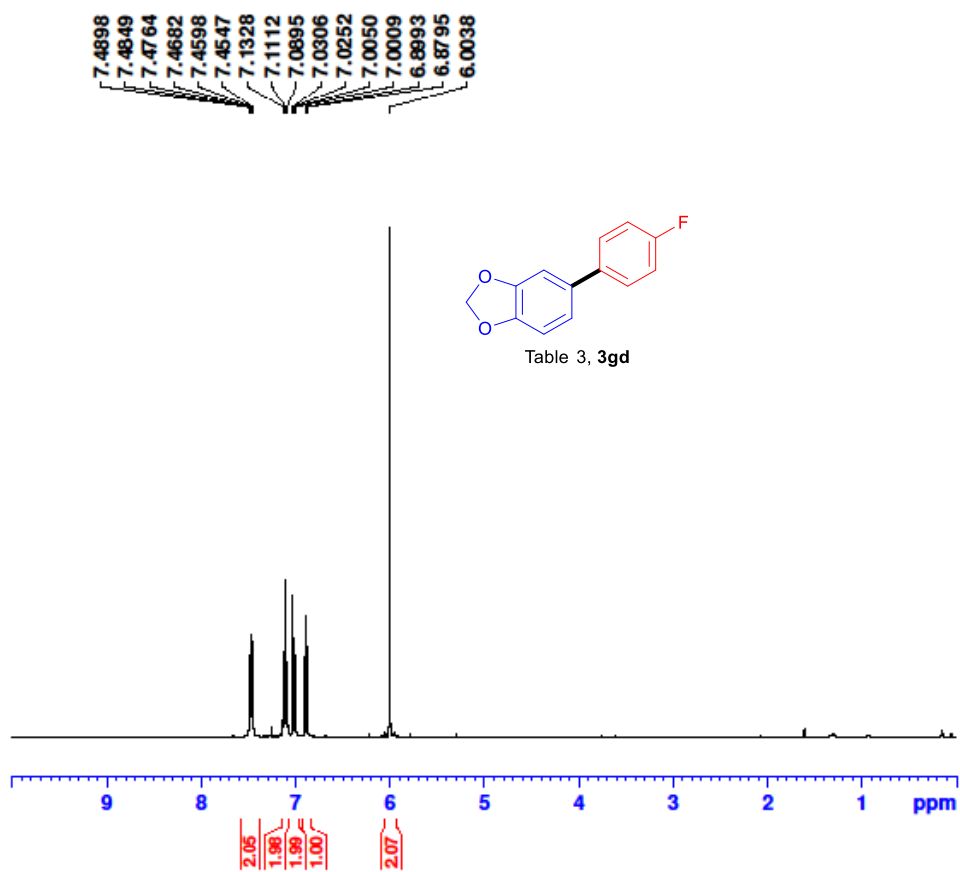


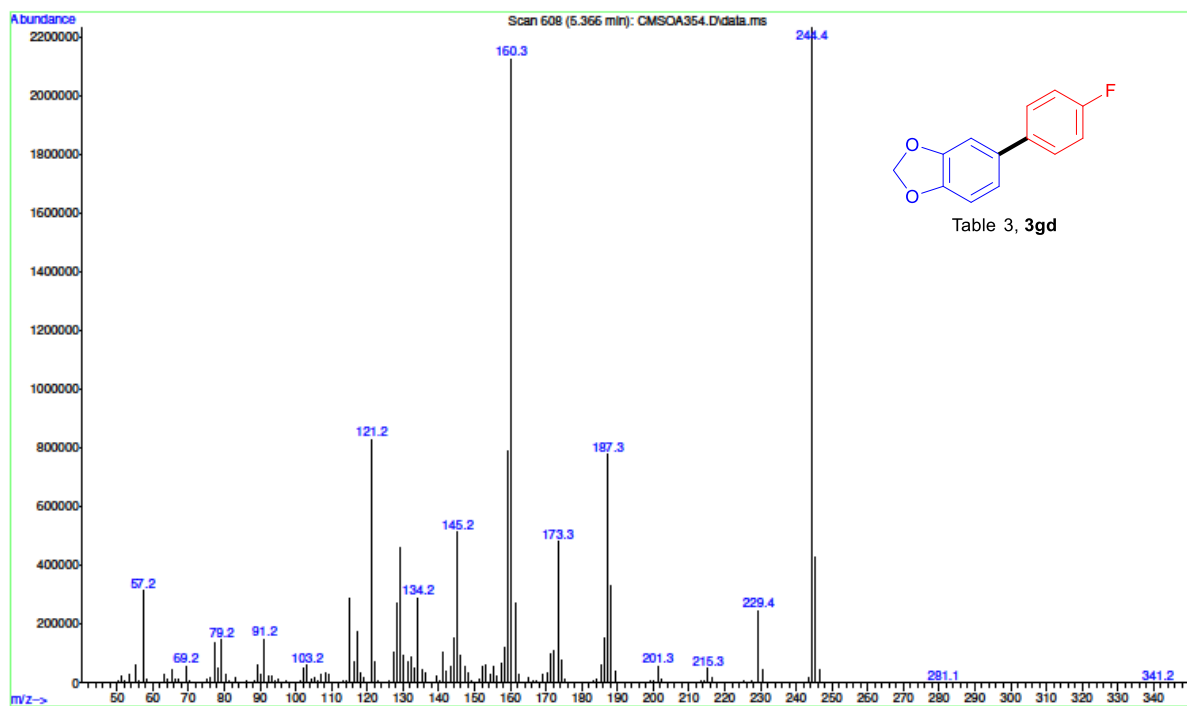
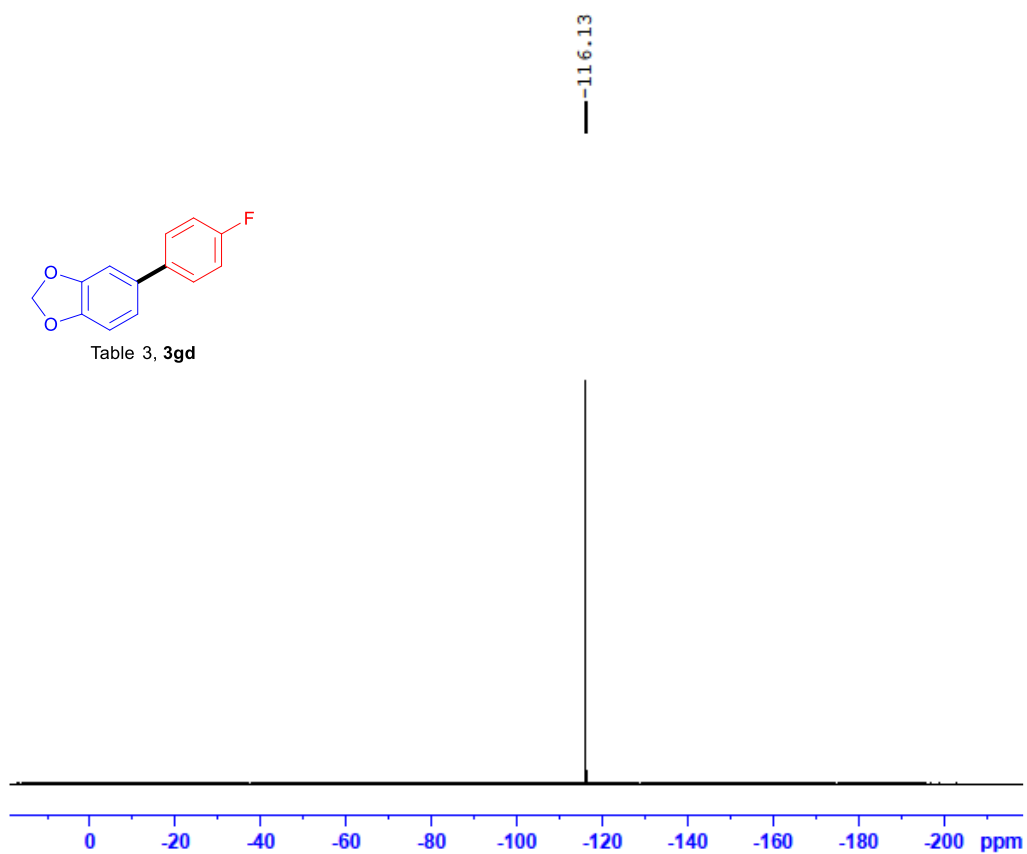


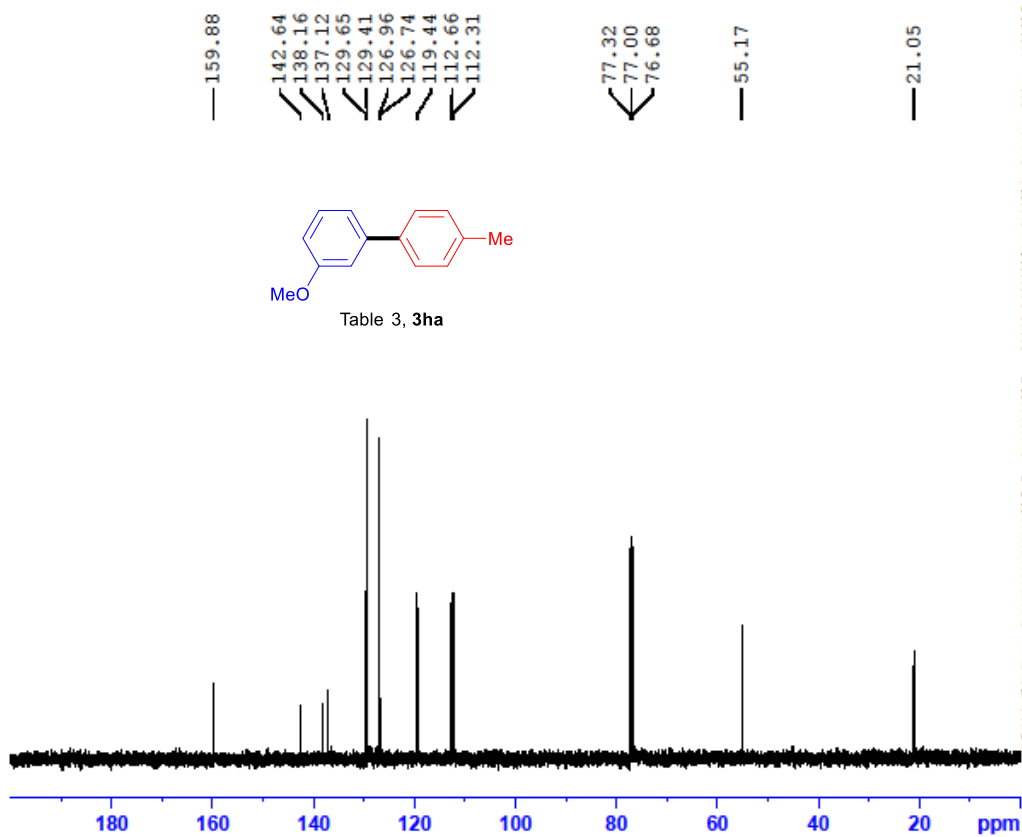
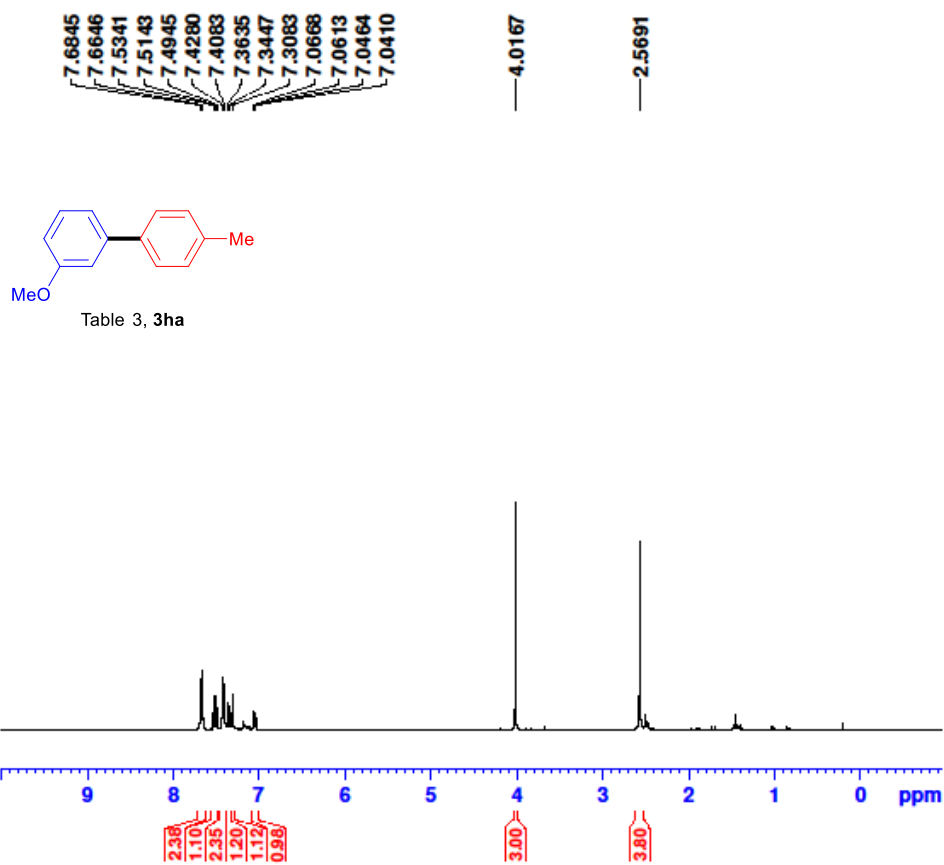


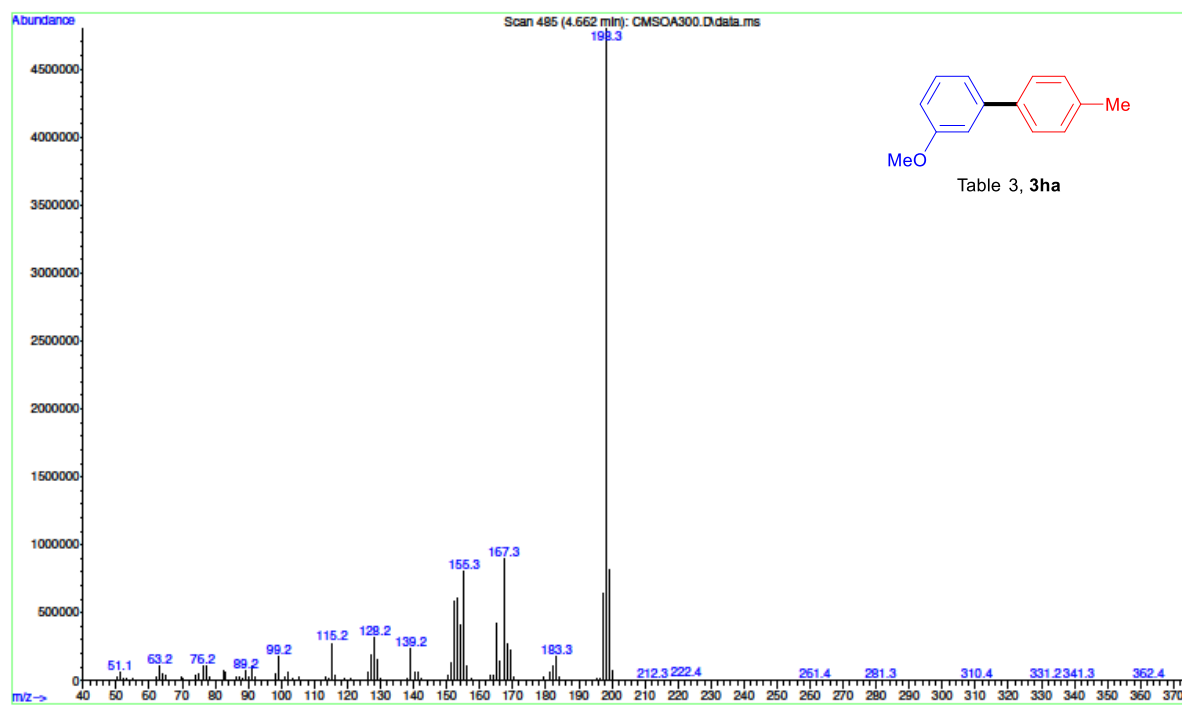


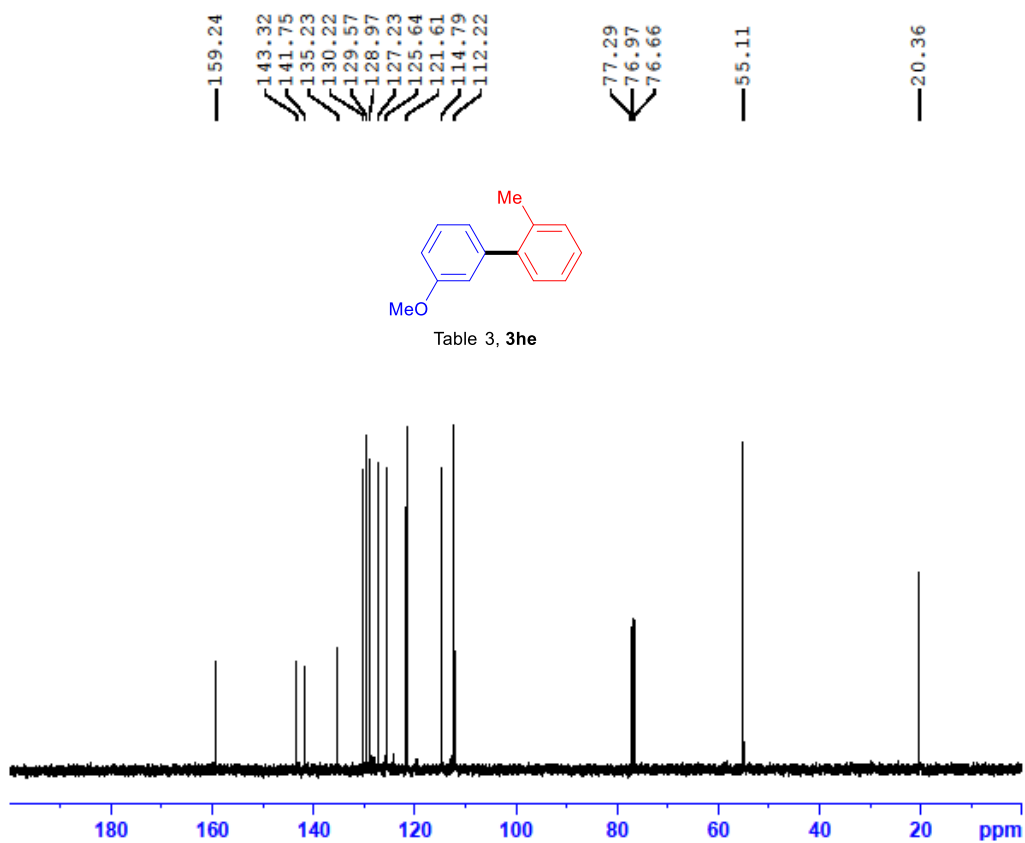
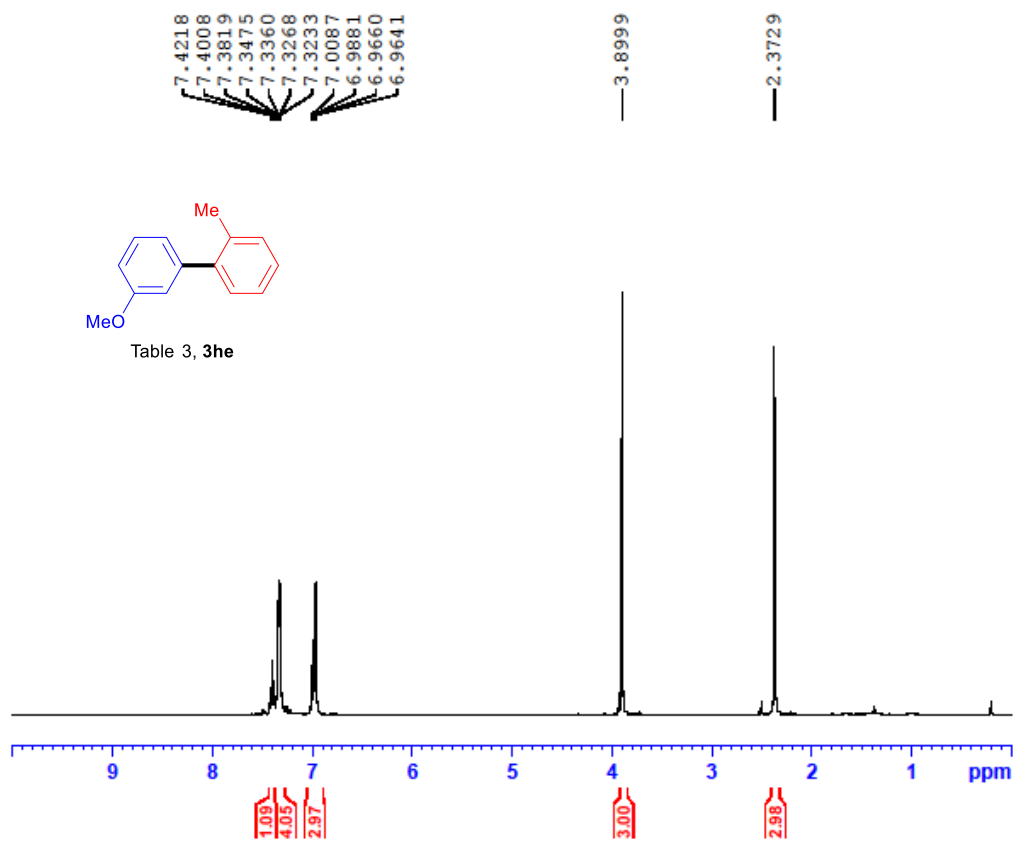
Mass	Calc. Mass	mDa	PPM	Formula
254.1305	254.1307	-0.2	-0.8	C ₁₇ H ₁₈ O ₂

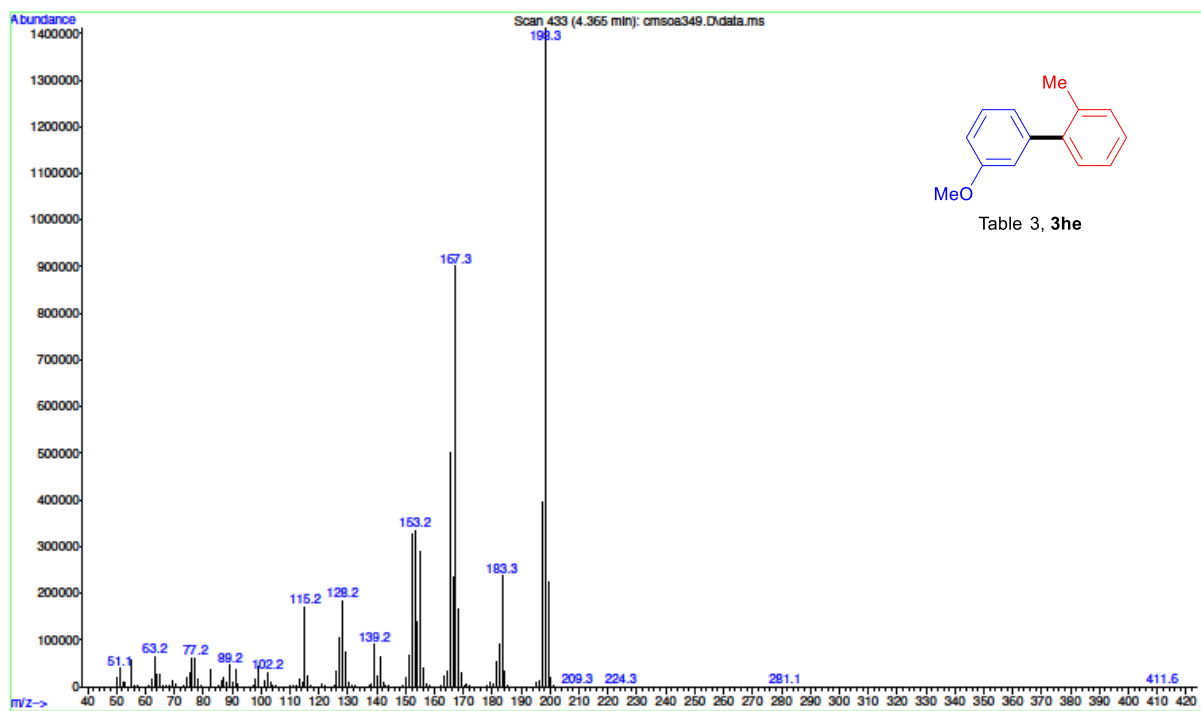




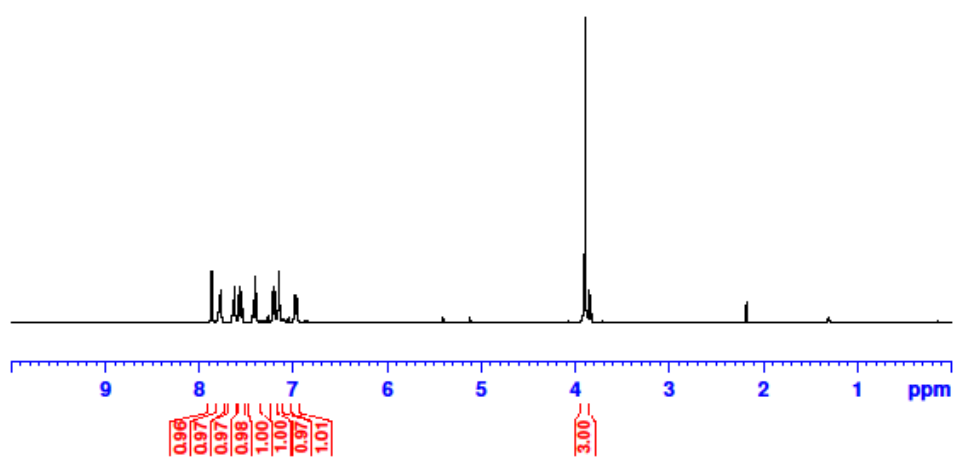
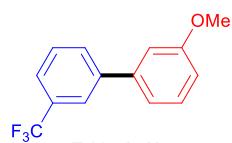




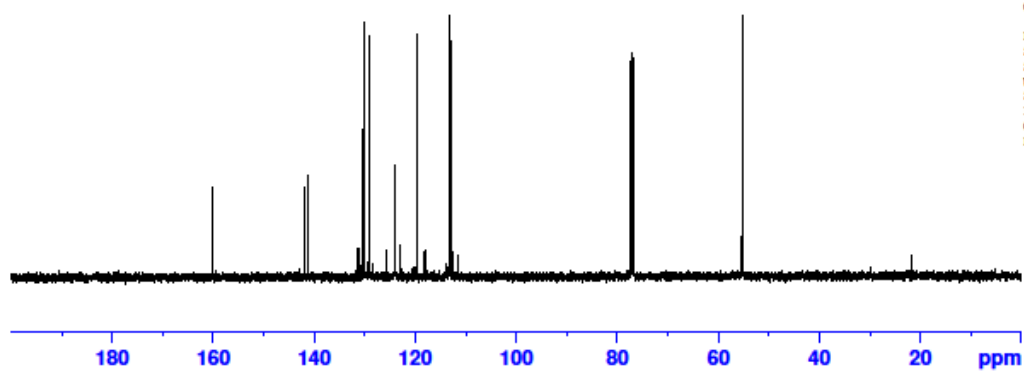
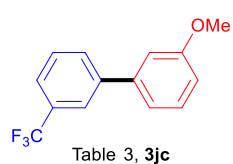


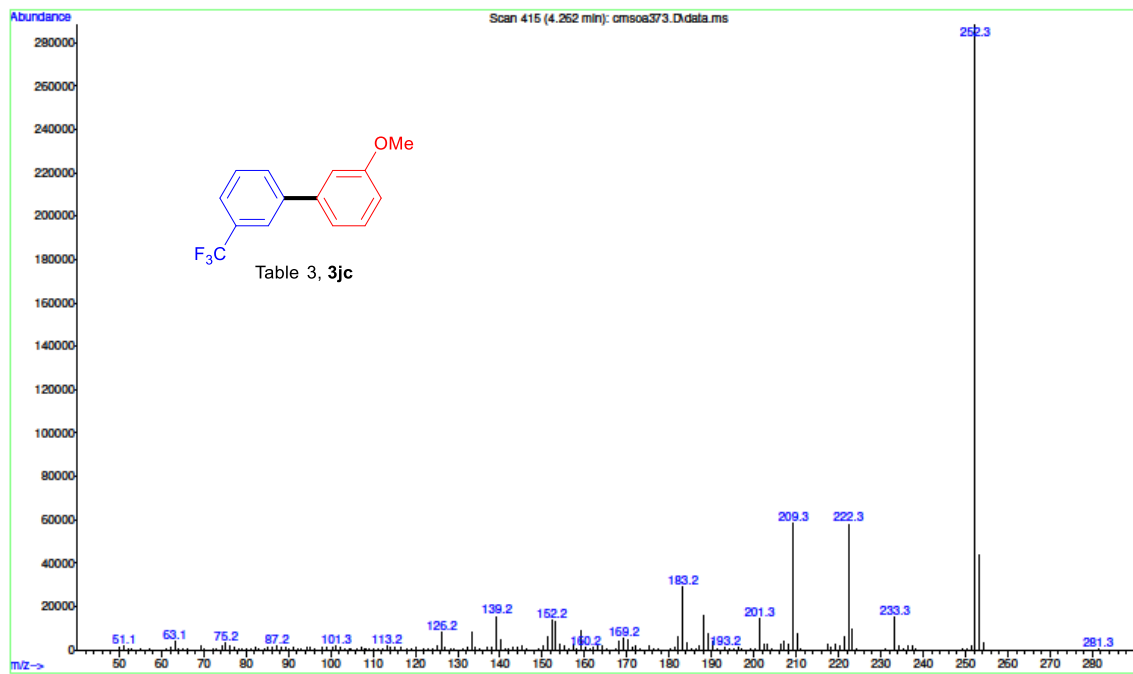
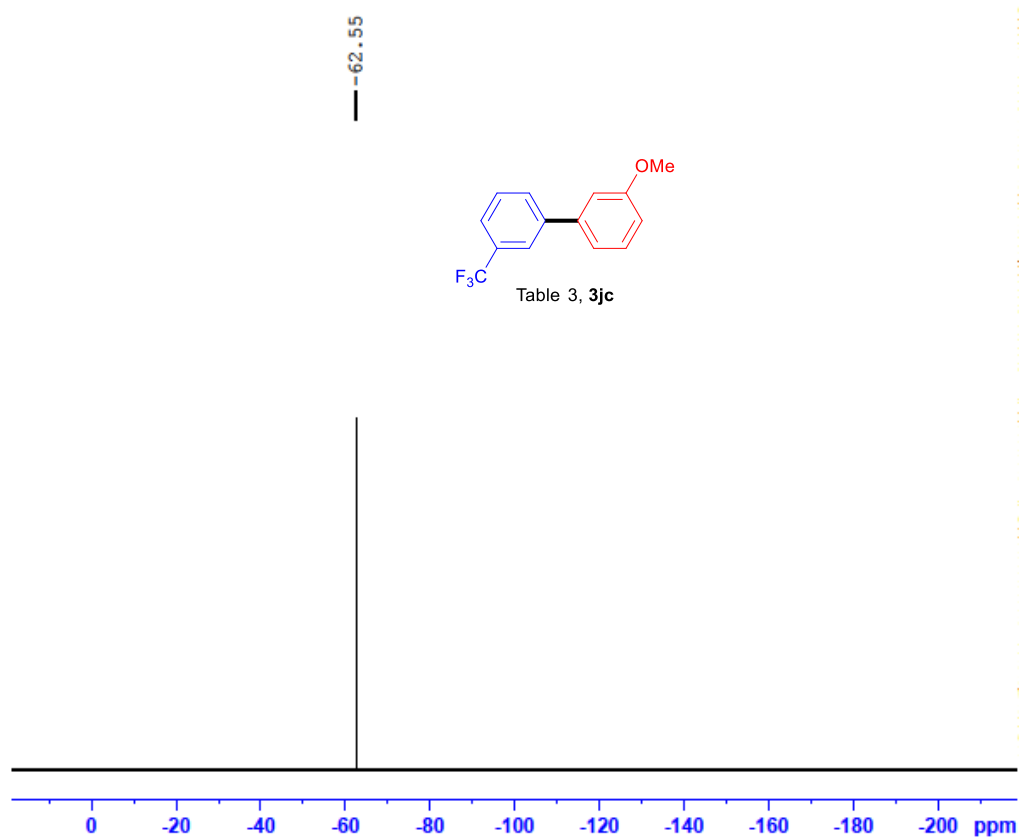


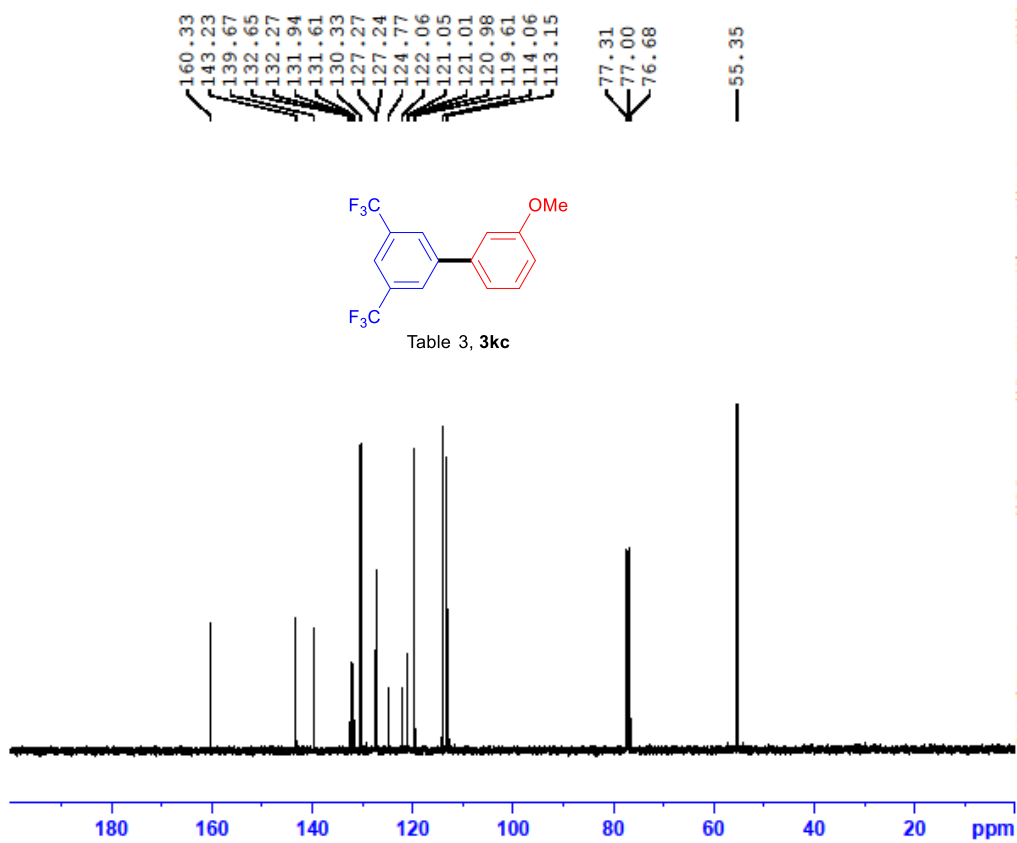
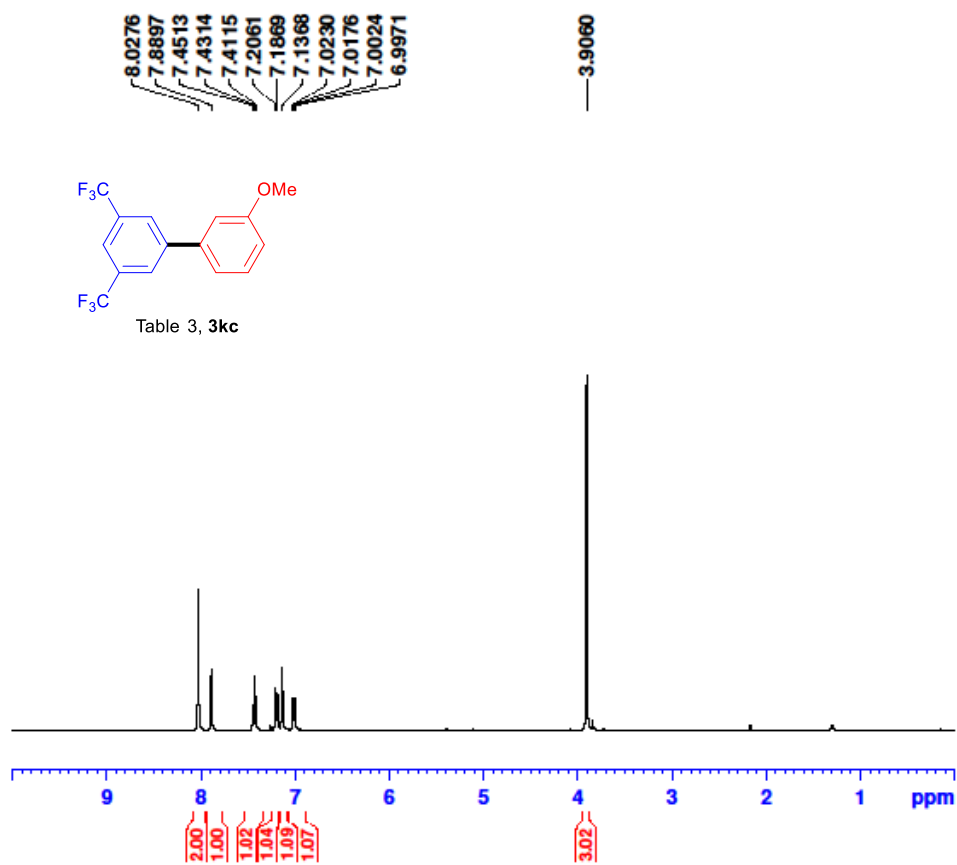
7.8683
7.7893
7.7703
7.6438
7.6246
7.5821
7.5629
7.5436
7.4298
7.4100
7.3901
7.2588
7.2142
7.1951
7.1519
6.9869
6.9816
6.9664
6.9611
3.8970

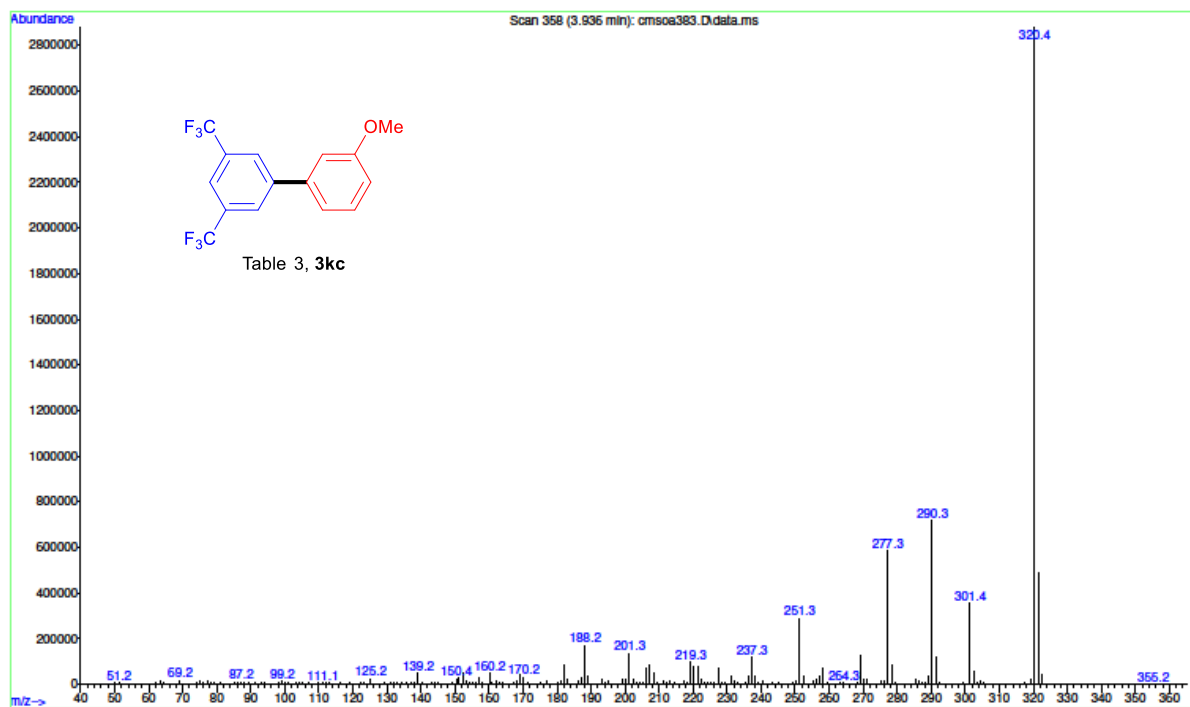
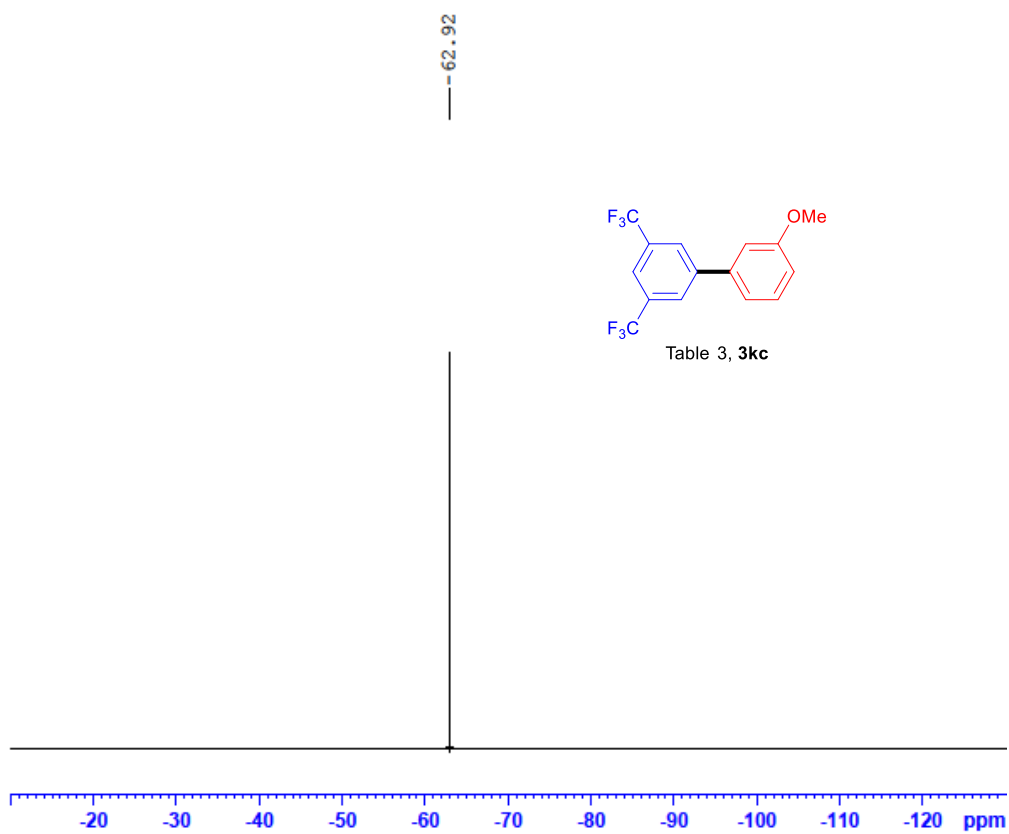


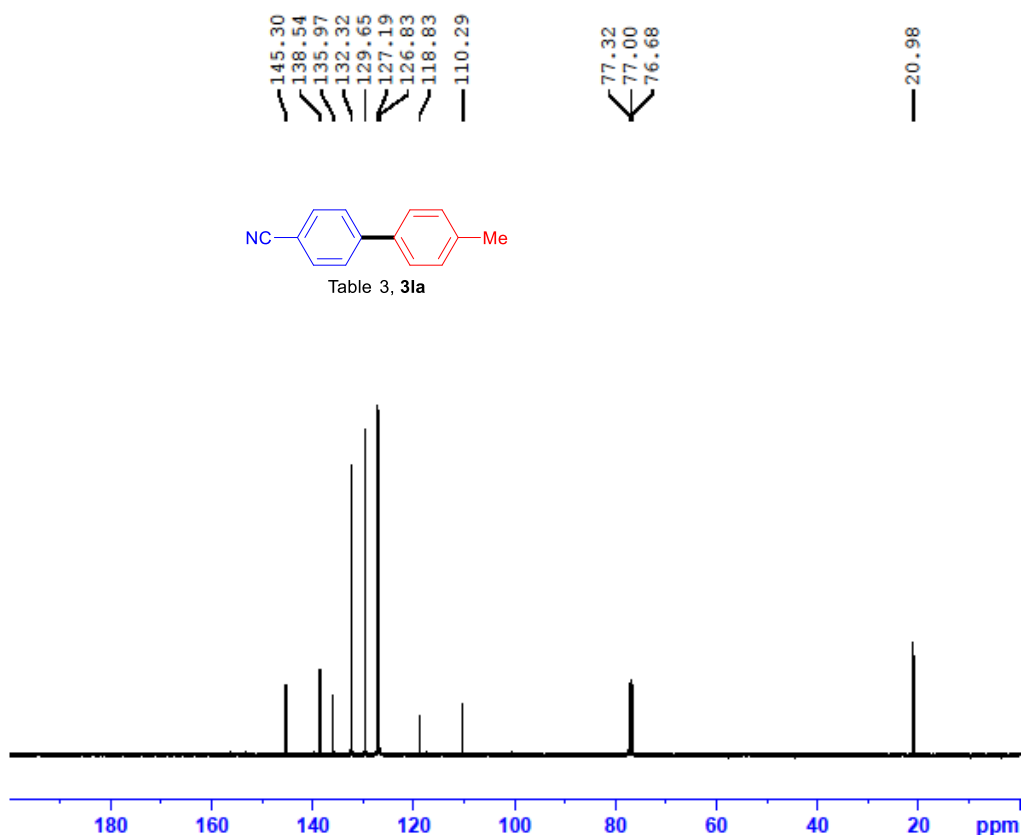
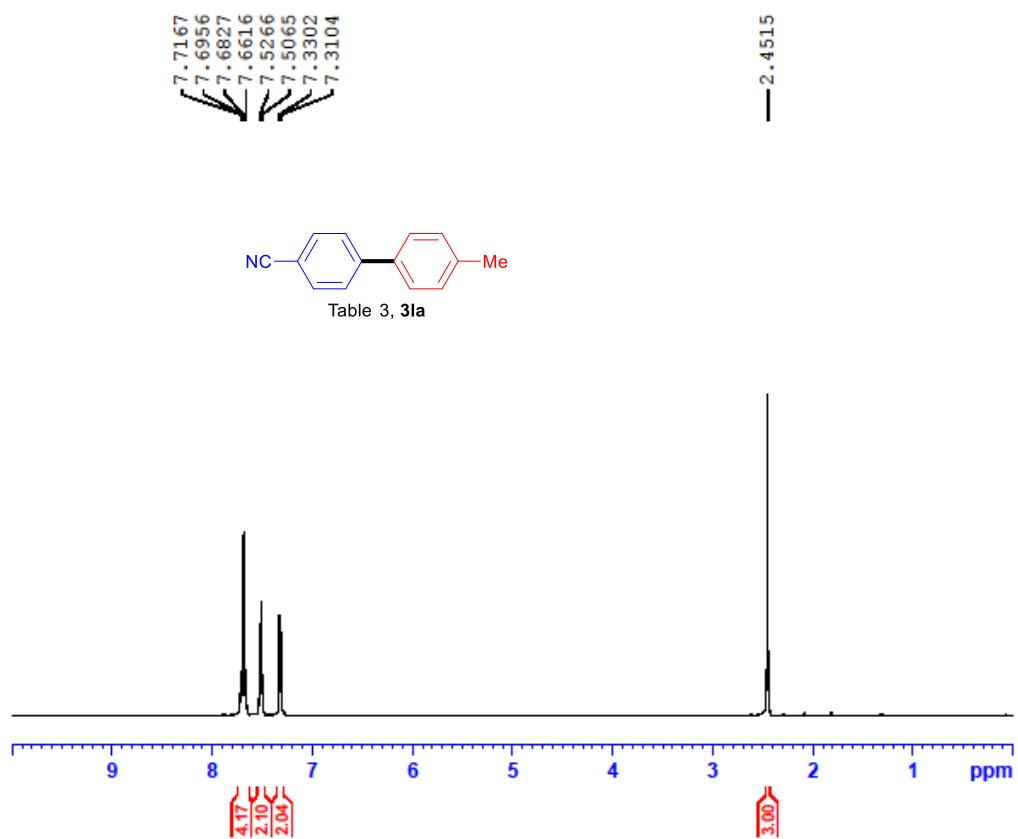
160.11
141.88
141.22
131.59
131.29
130.95
130.64
130.44
130.01
129.18
128.23
125.61
124.05
124.01
123.96
123.92
123.88
122.87
120.12
119.62
113.32
113.01
77.32
77.00
76.68
55.42

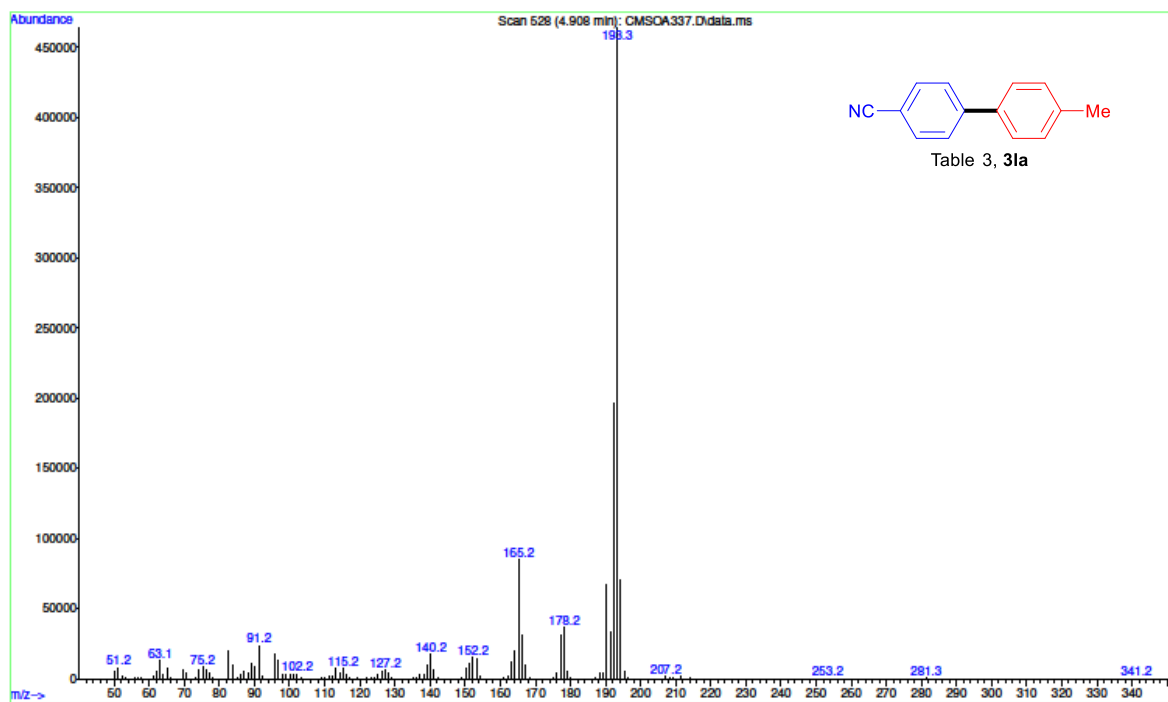












7.7770
7.7559
7.7338
7.7091
7.6885

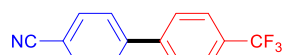
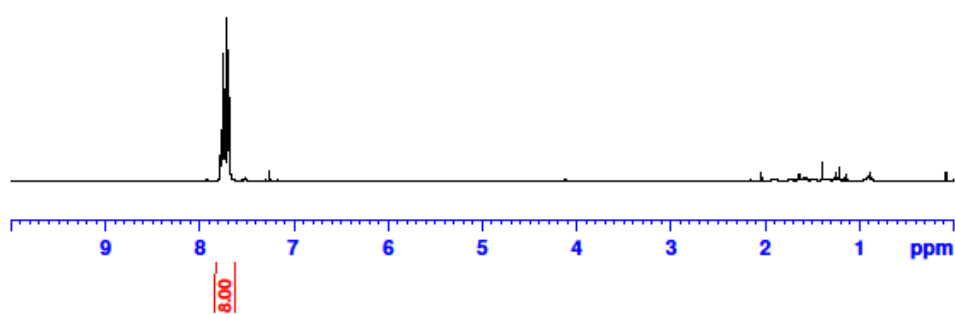


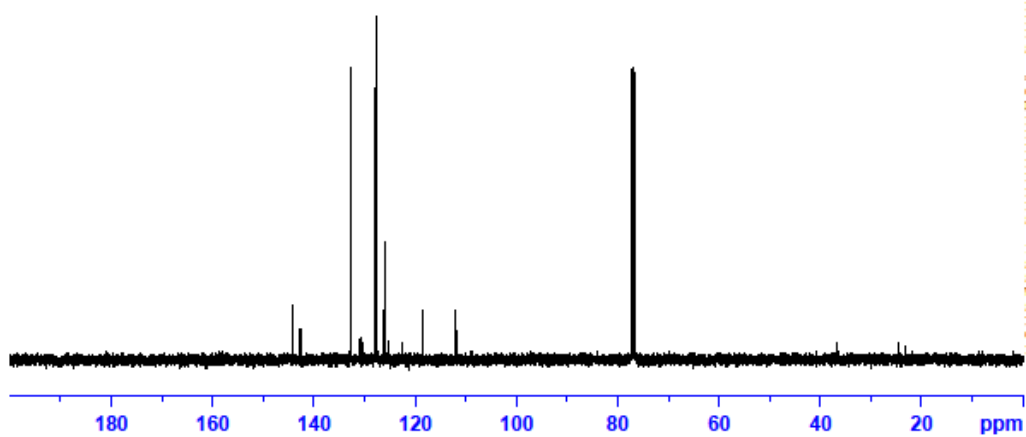
Table 3, 3Ig

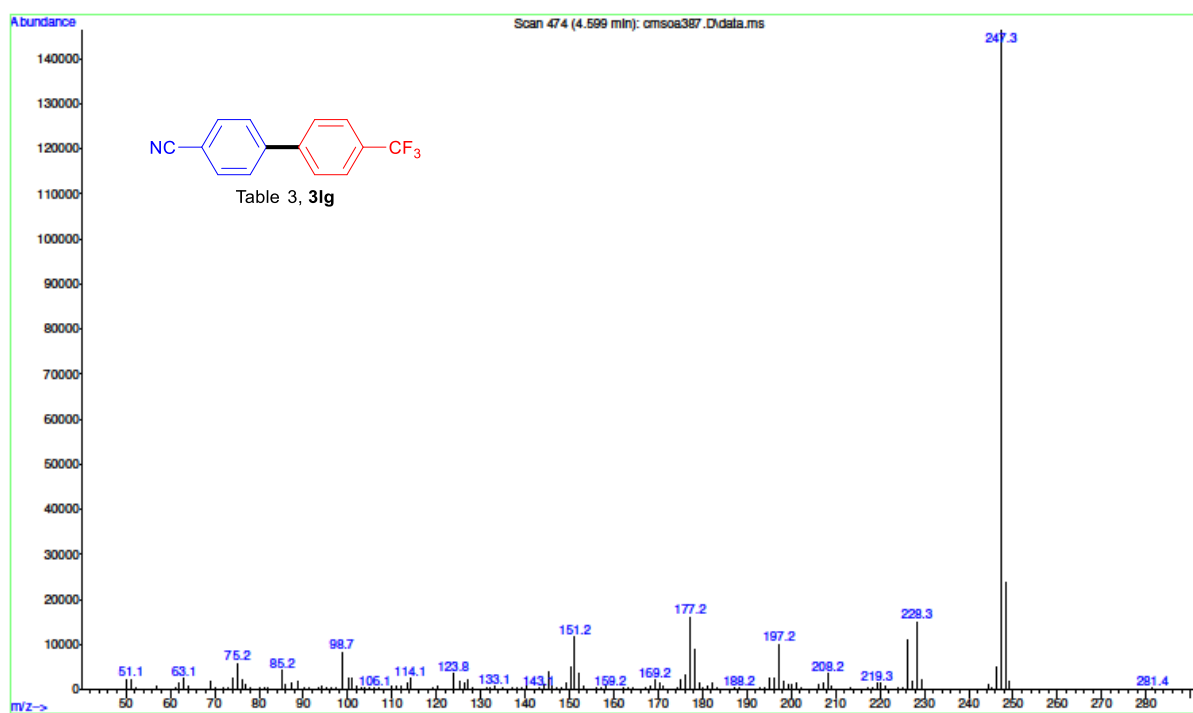
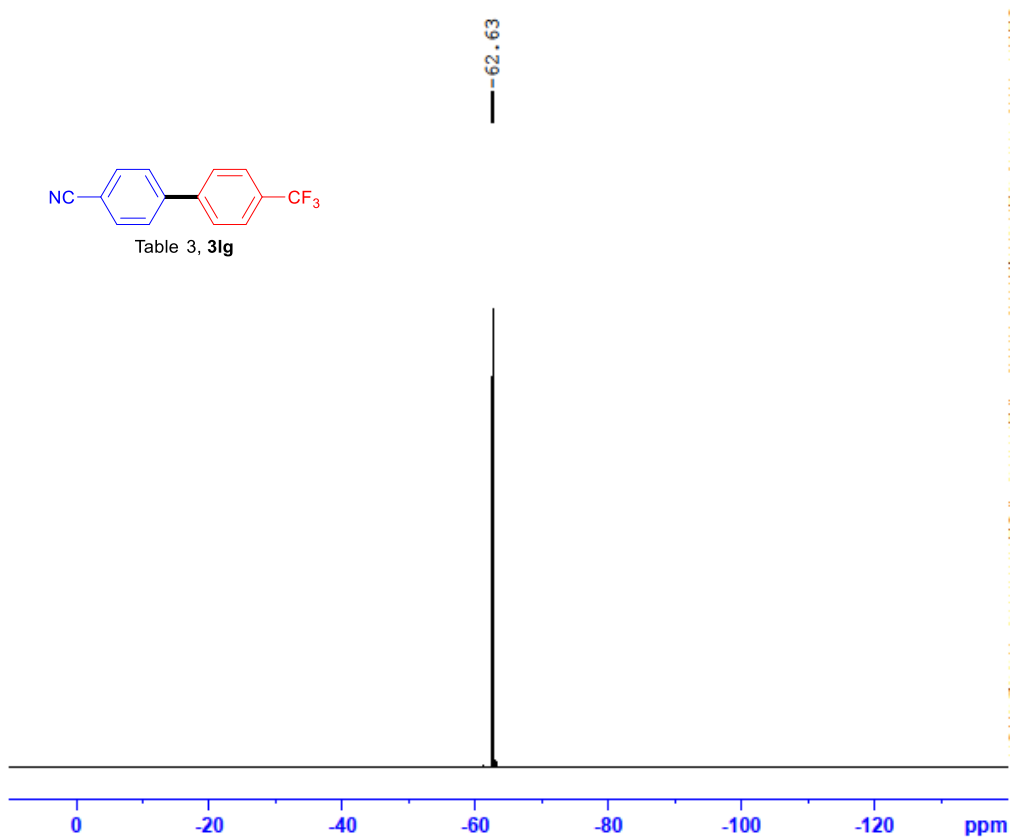


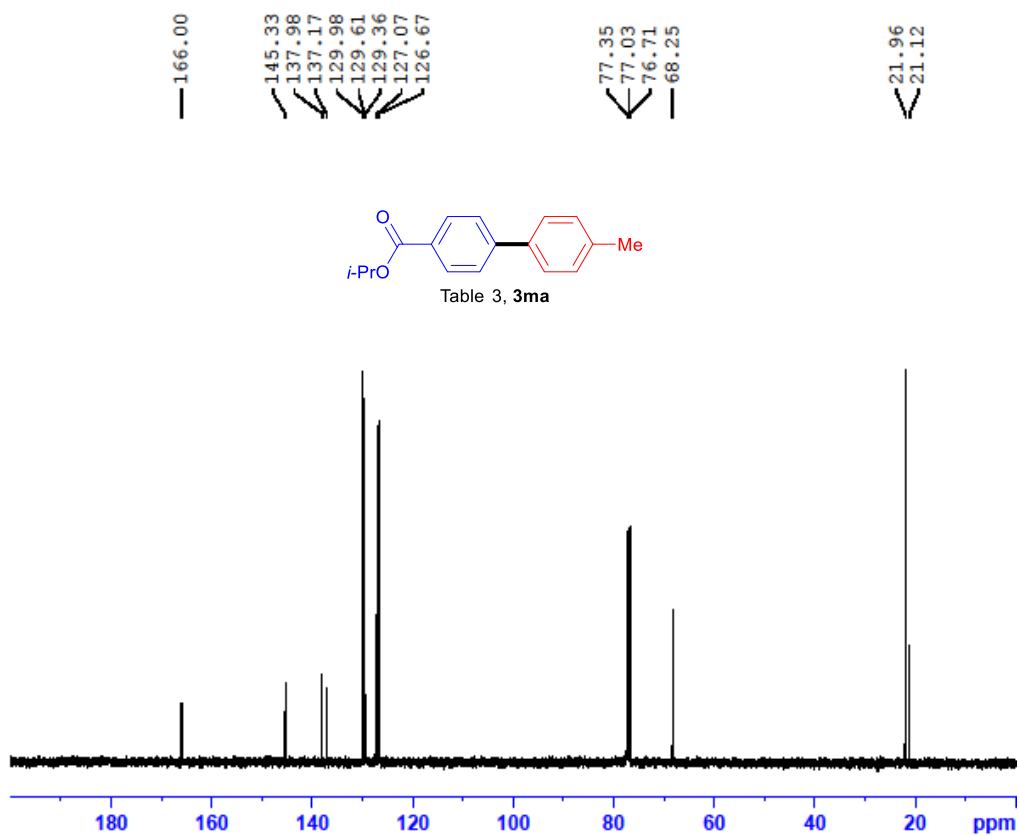
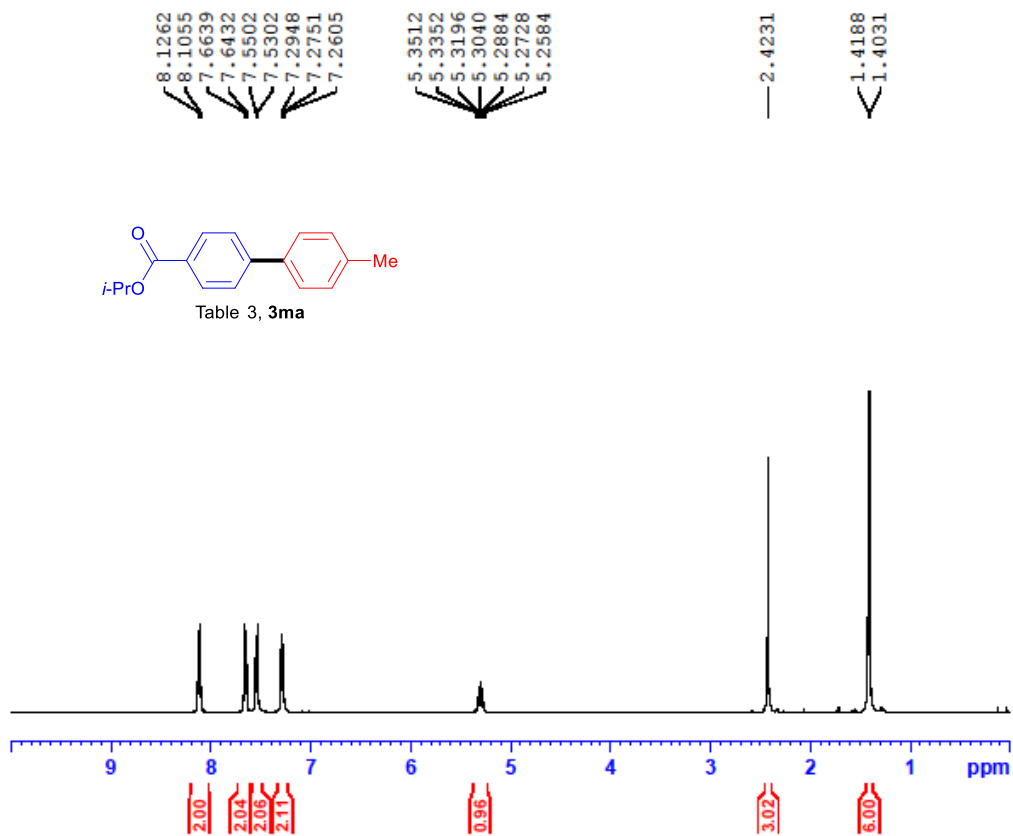
144.09
142.63
132.74
130.80
130.48
127.92
127.59
126.08
126.05
126.01
125.97
125.32
122.61
118.54
111.94
77.32
77.00
76.68

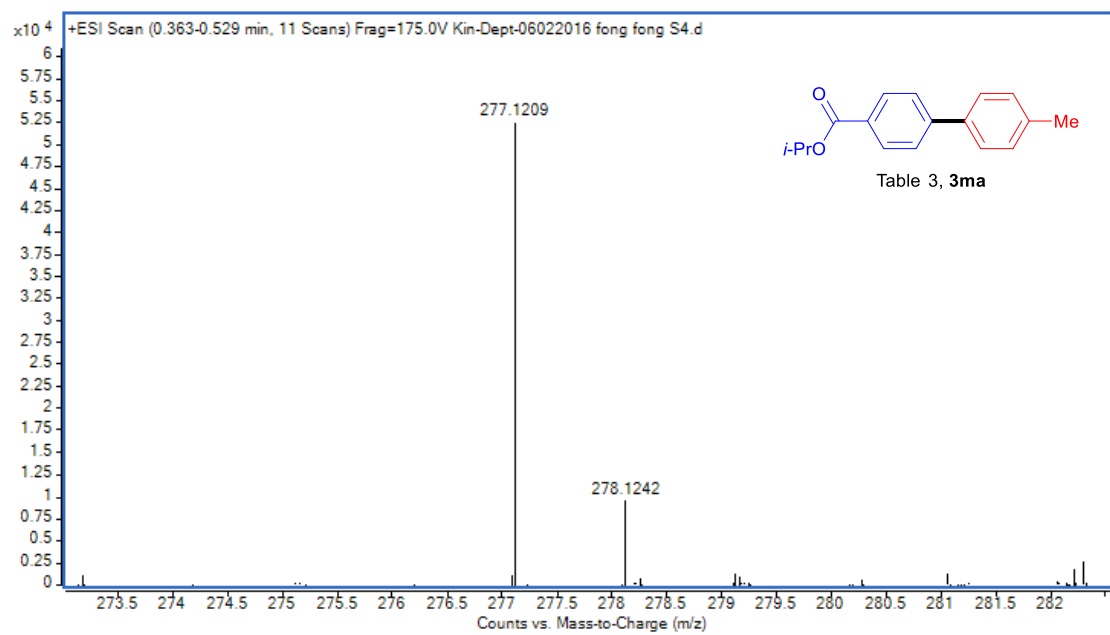
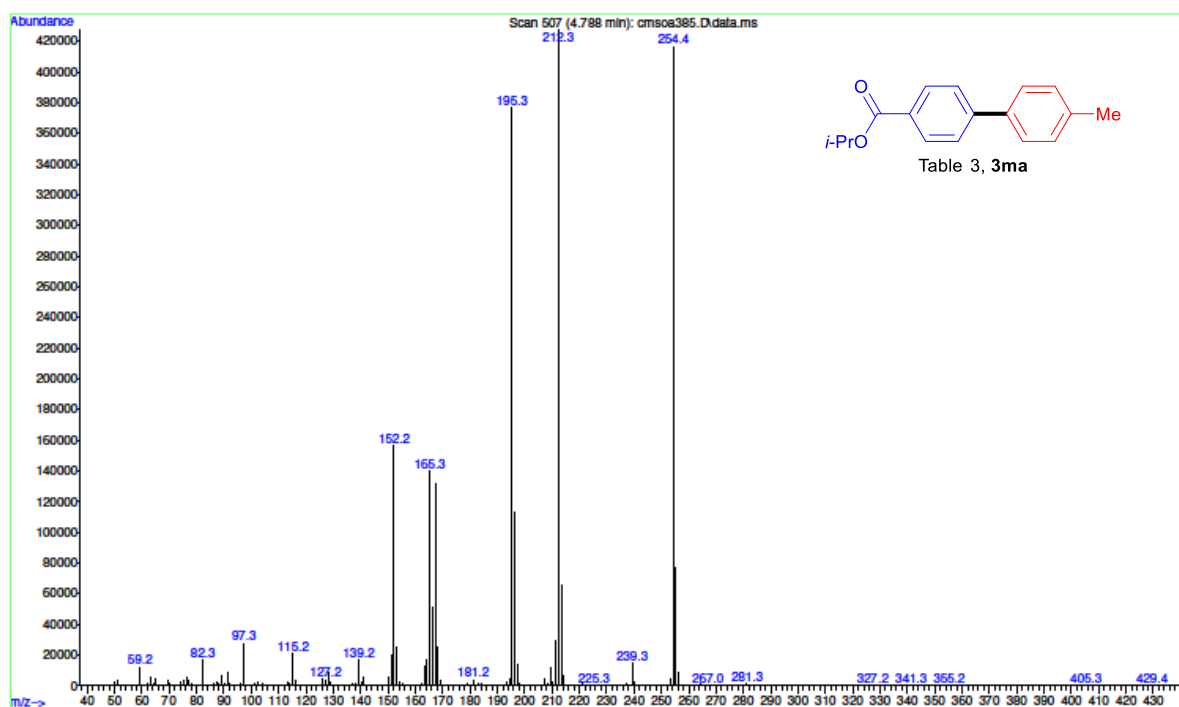


Table 3, 3Ig

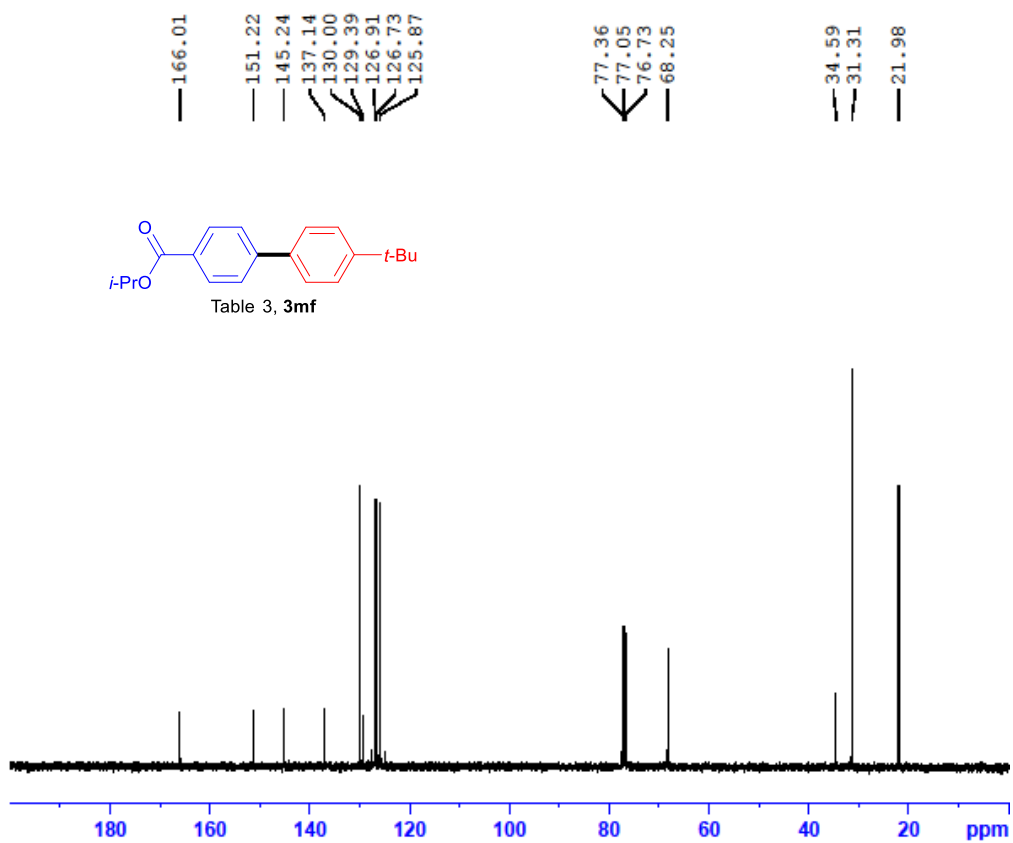
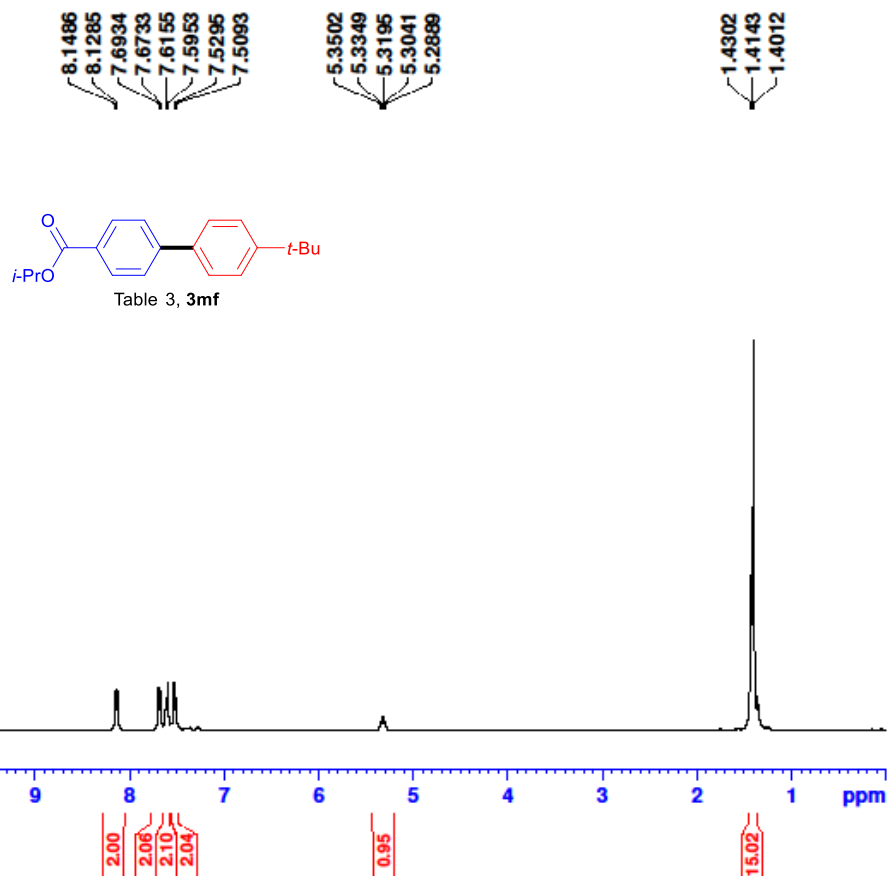


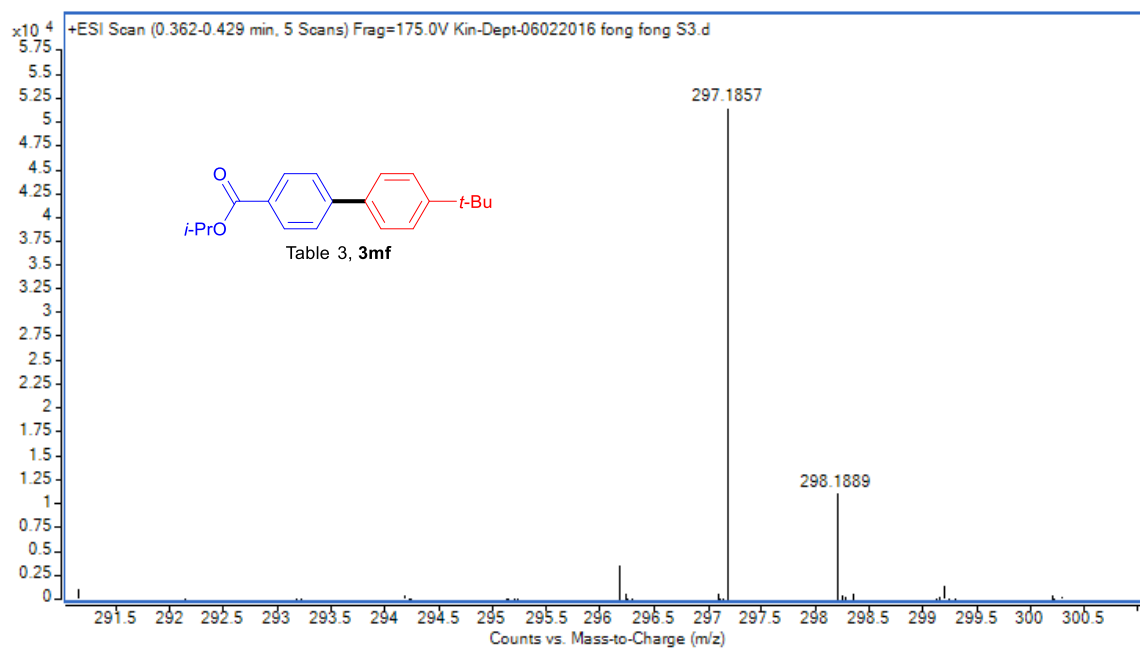
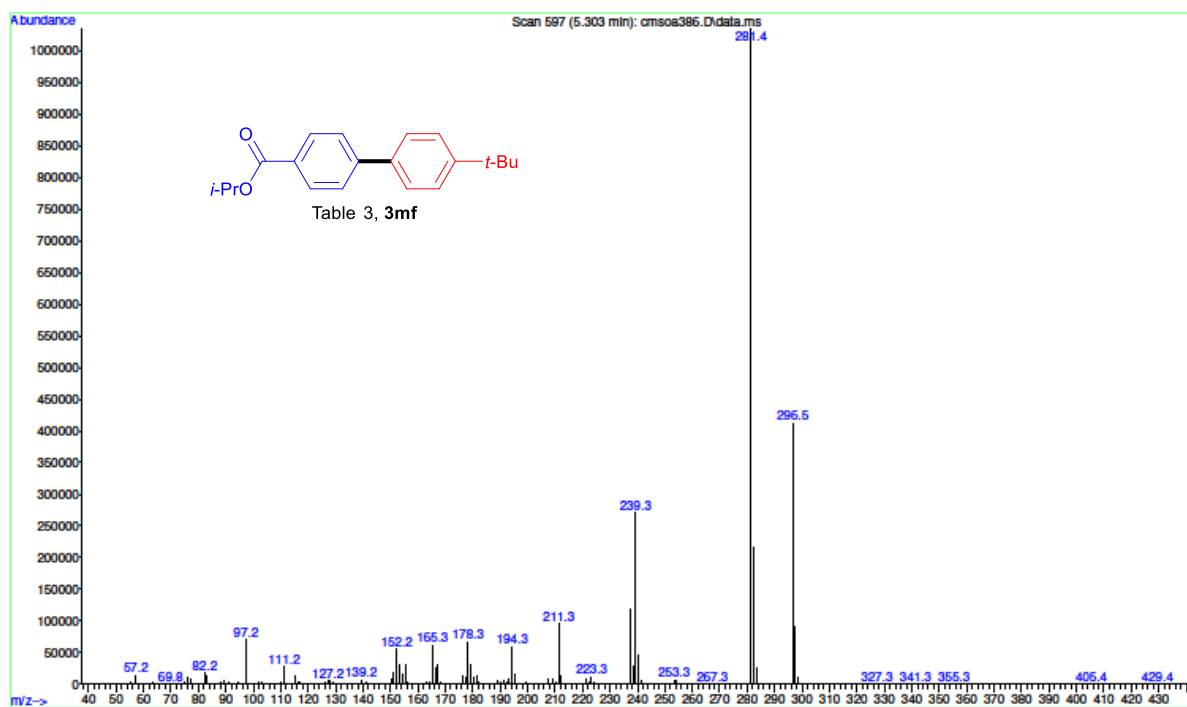




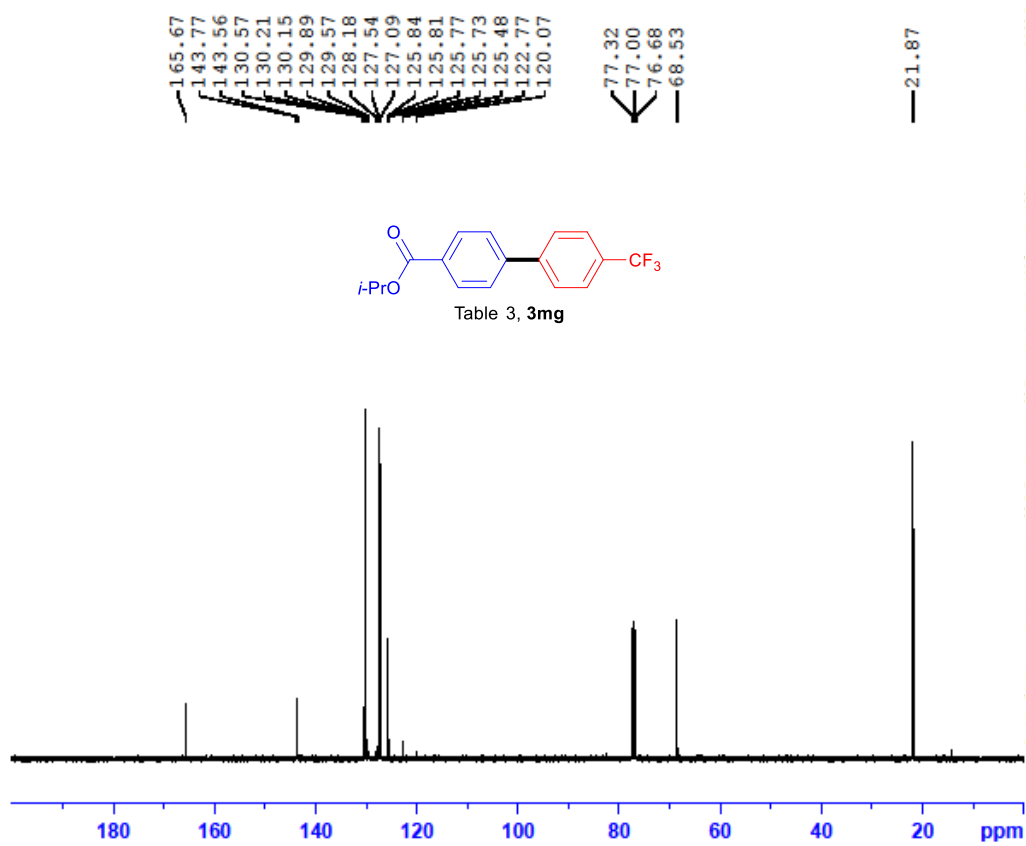
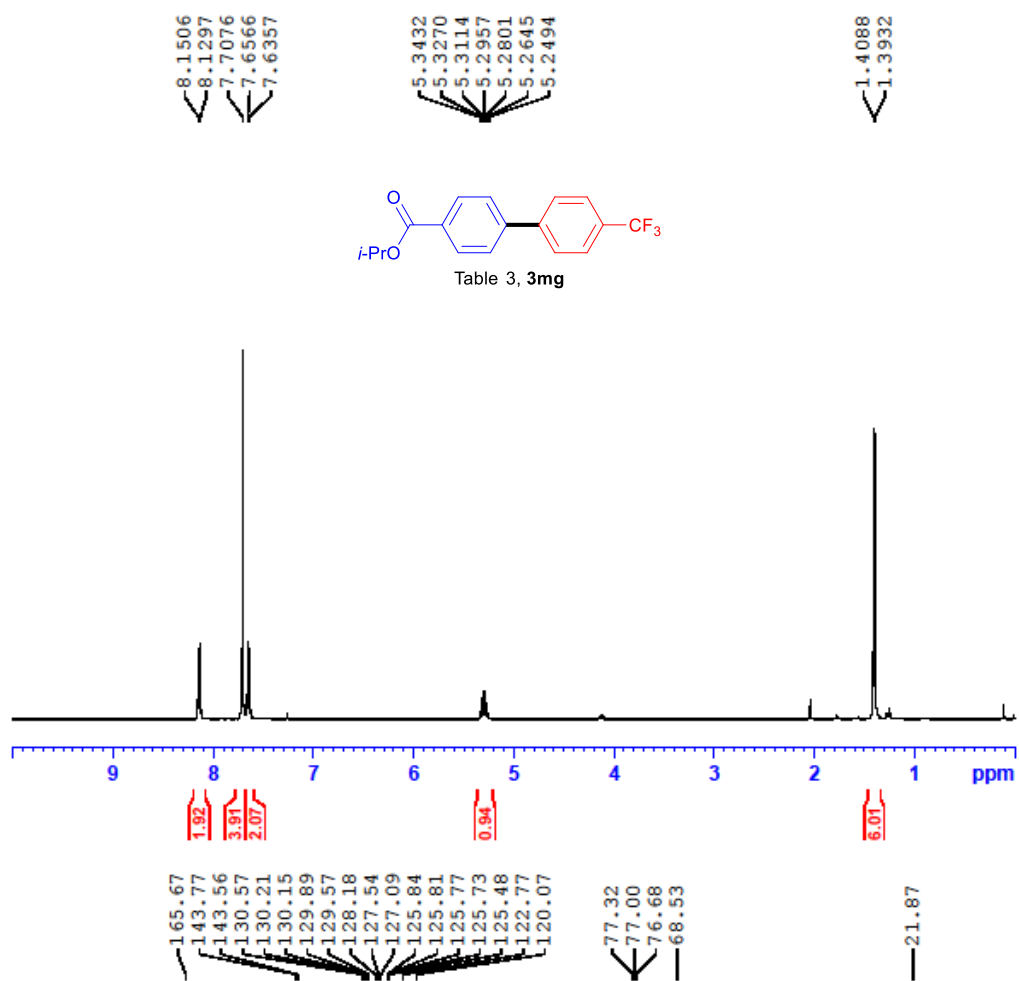


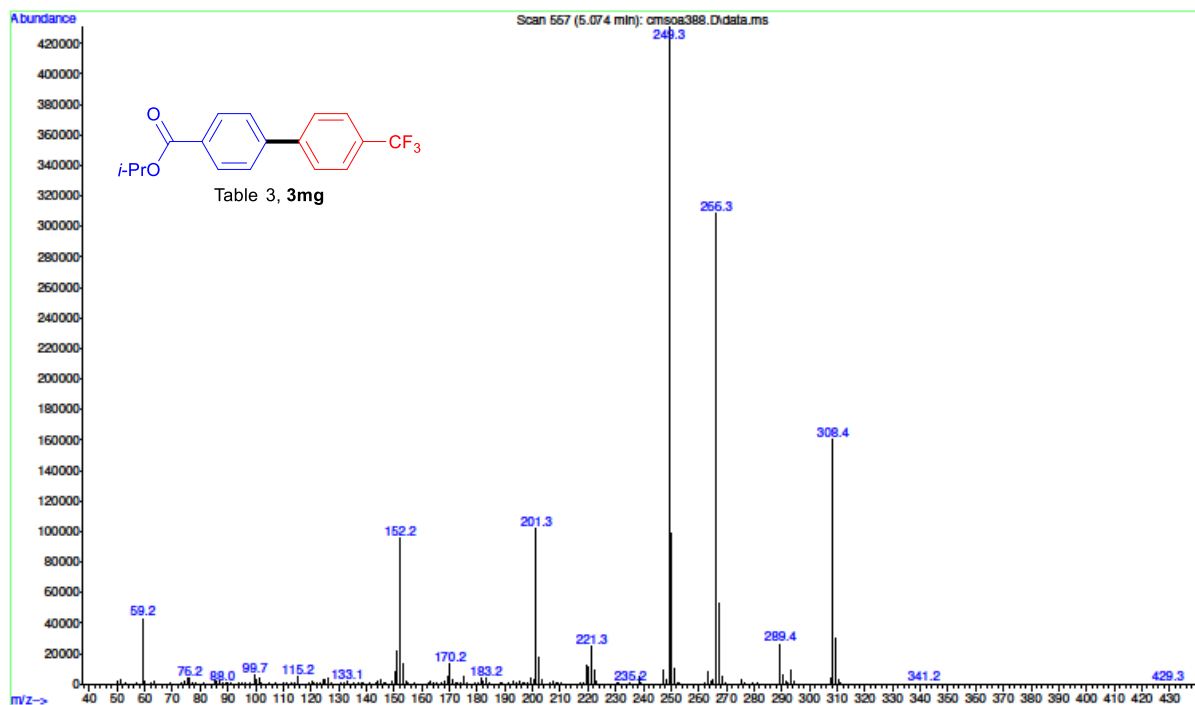
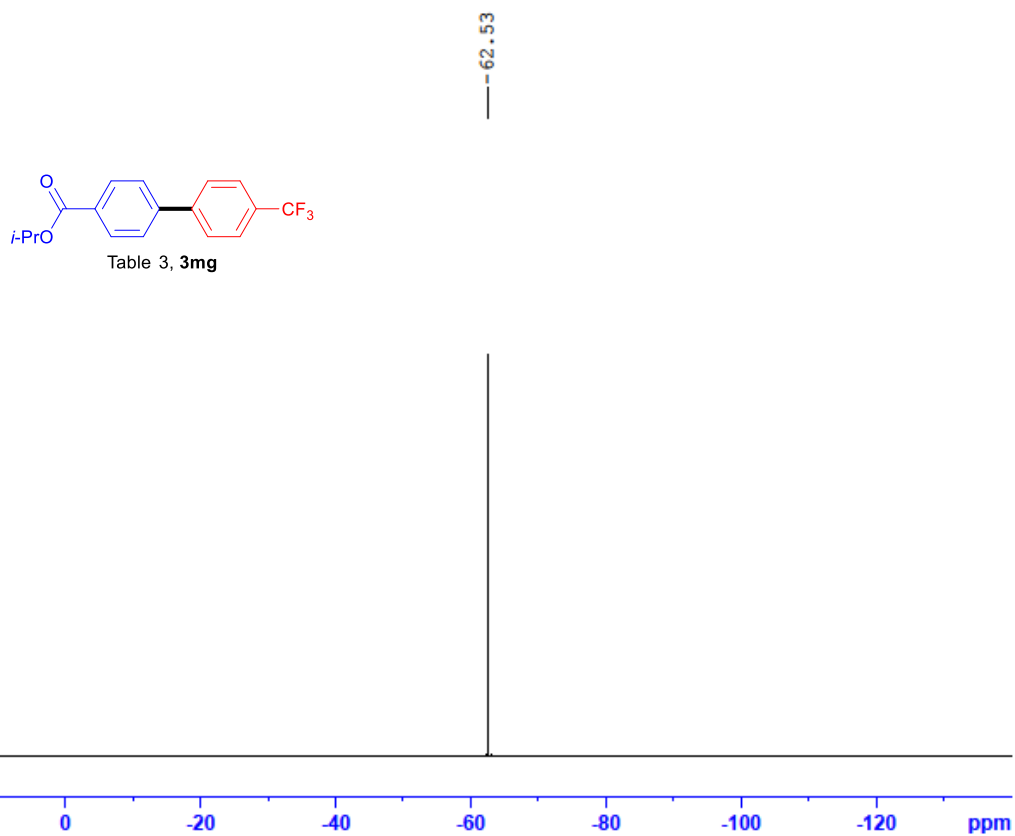
Mass	Calc. Mass	mDa	PPM	Formula
277.1209	277.1199	1.0	3.6	C17 H18 Na O2

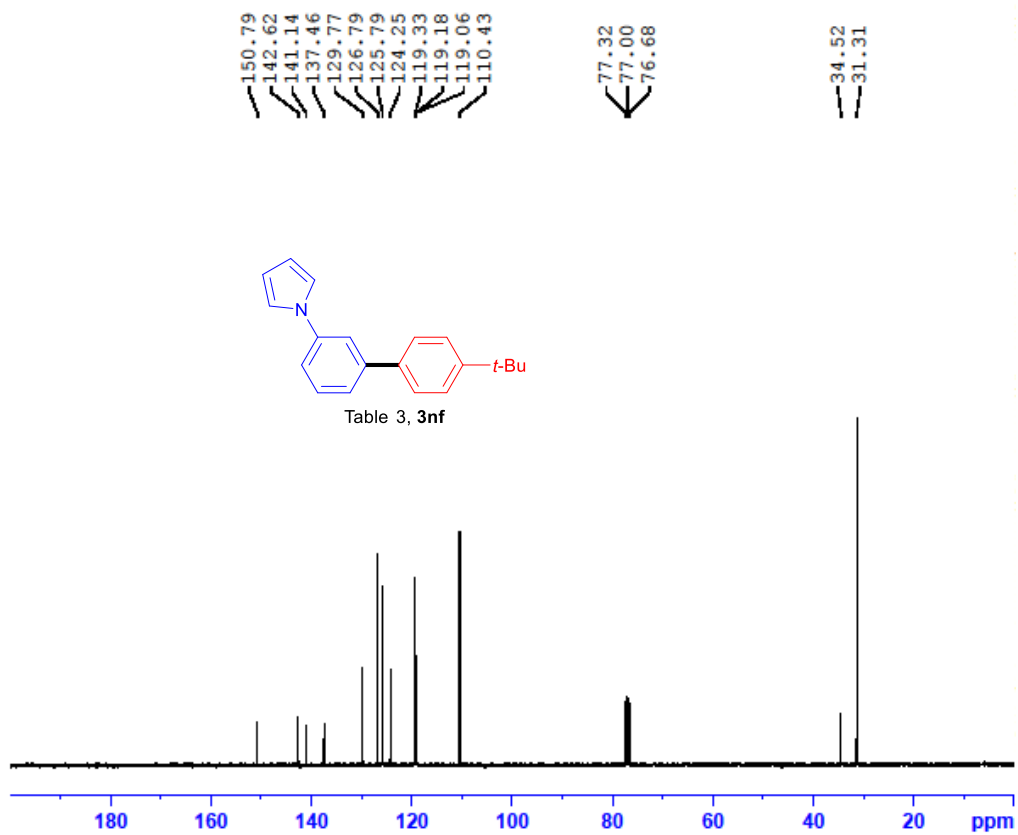
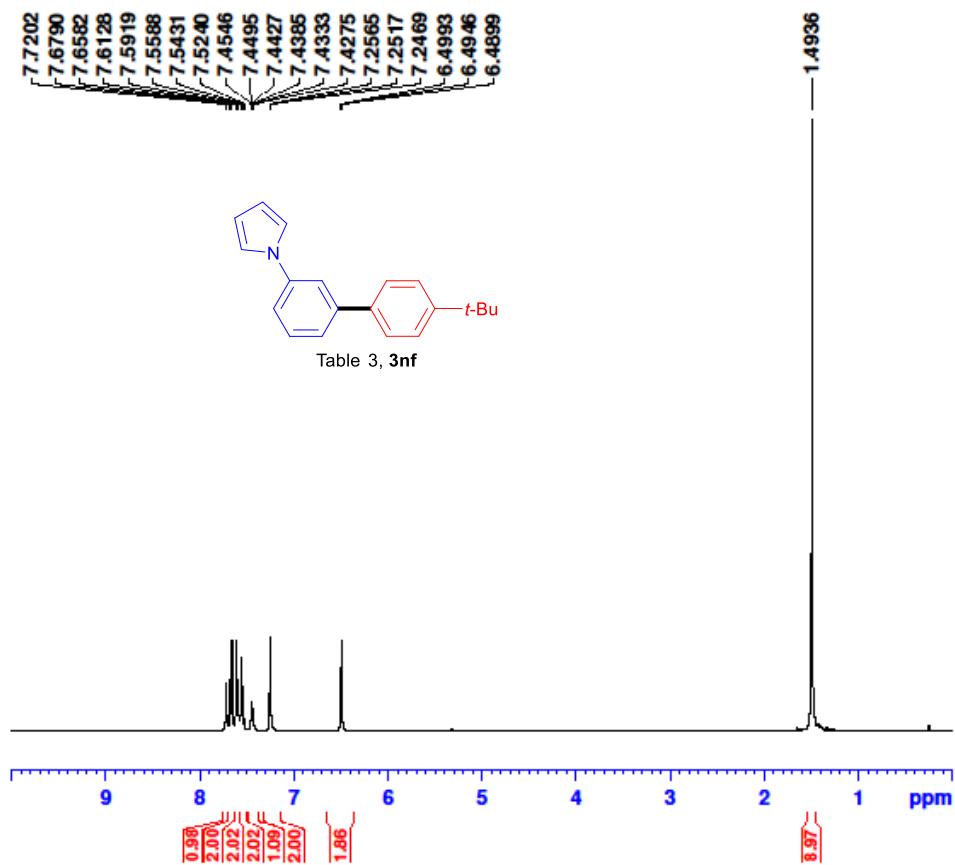


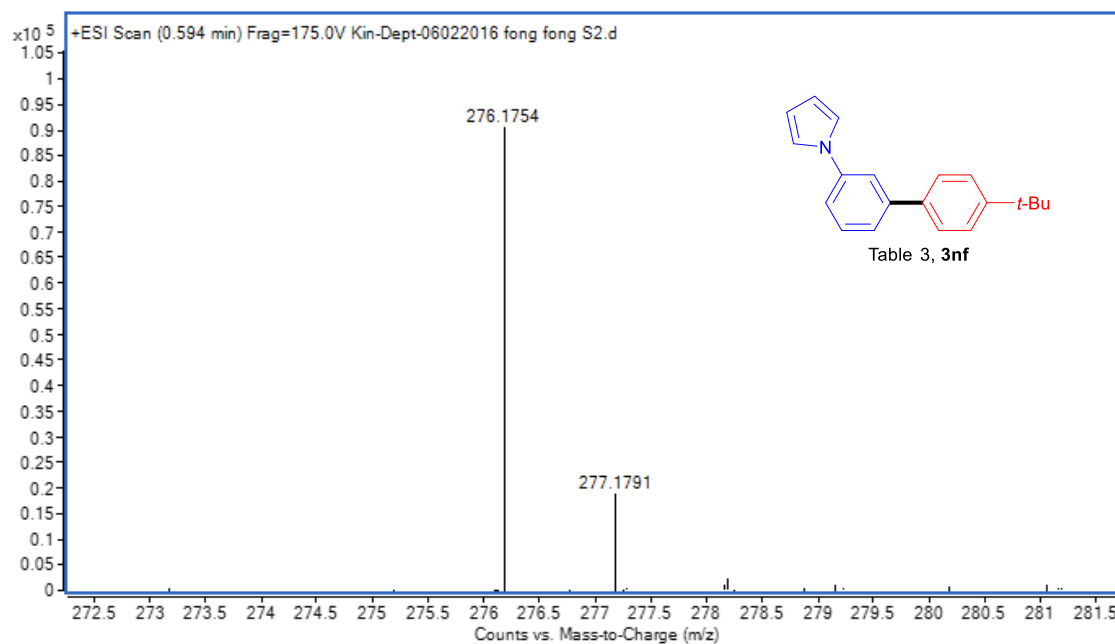
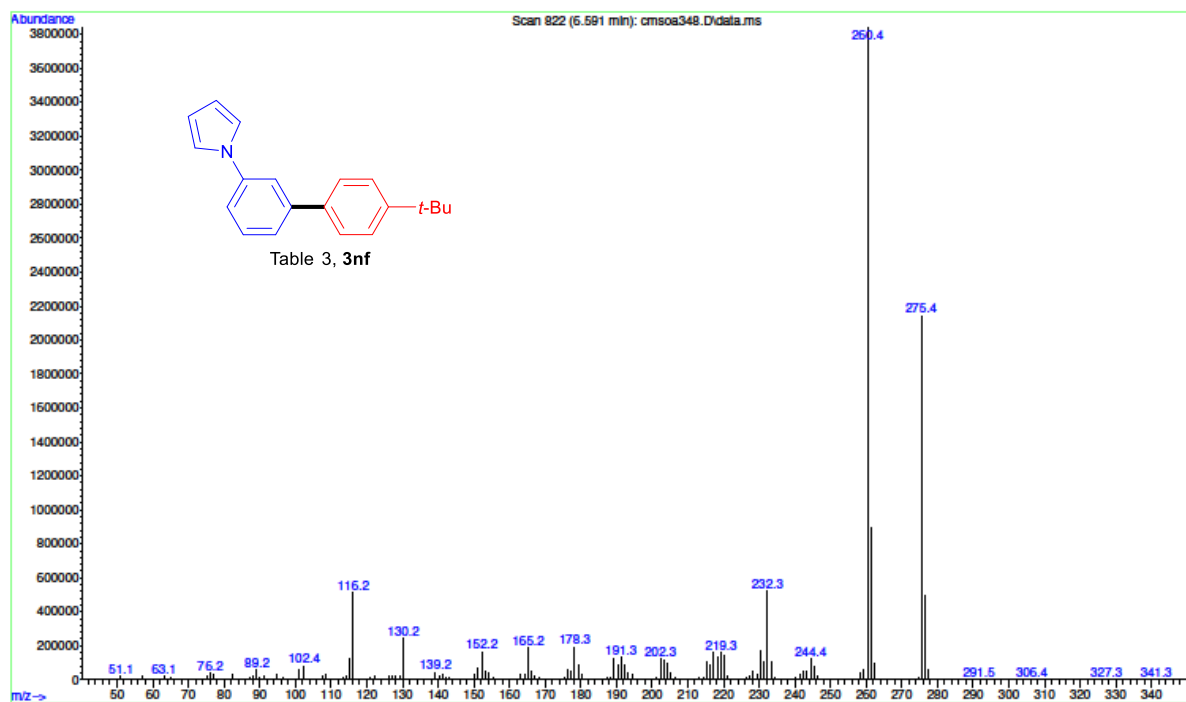


Mass	Calc. Mass	mDa	PPM	Formula
297.1857	297.1849	0.8	2.7	C20 H25 O2









Mass	Calc. Mass	mDa	PPM	Formula
276.1754	276.1747	0.7	2.6	C ₂₀ H ₂₂ N

8.1852
8.1820
7.8411
7.8204
7.5938
7.5897
7.5821
7.5734
7.5677
7.5620
7.2955
7.2757

2.8495
2.4187

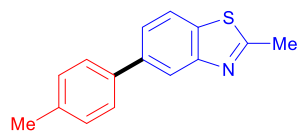
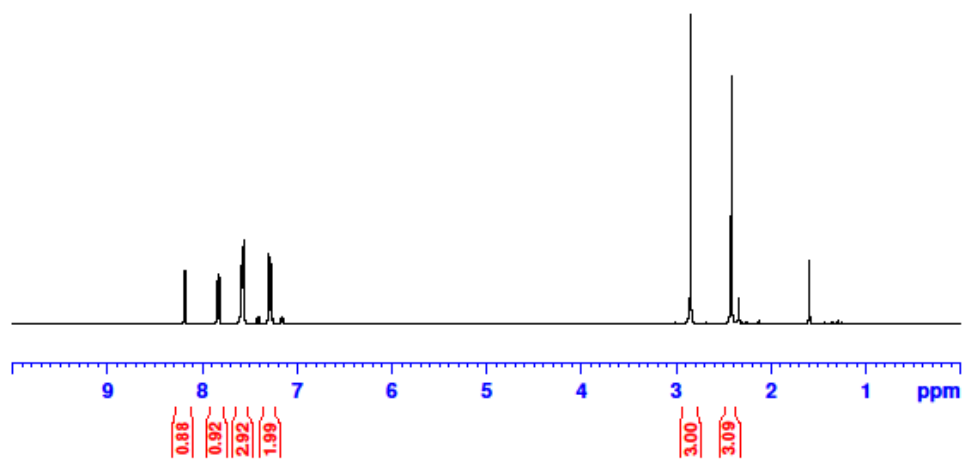


Table 3, 3oa



167.55
153.90
139.44
137.79
137.14
134.16
129.56
127.09
124.01
121.39
120.35

31.73
21.02
20.05

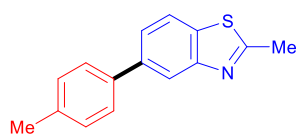
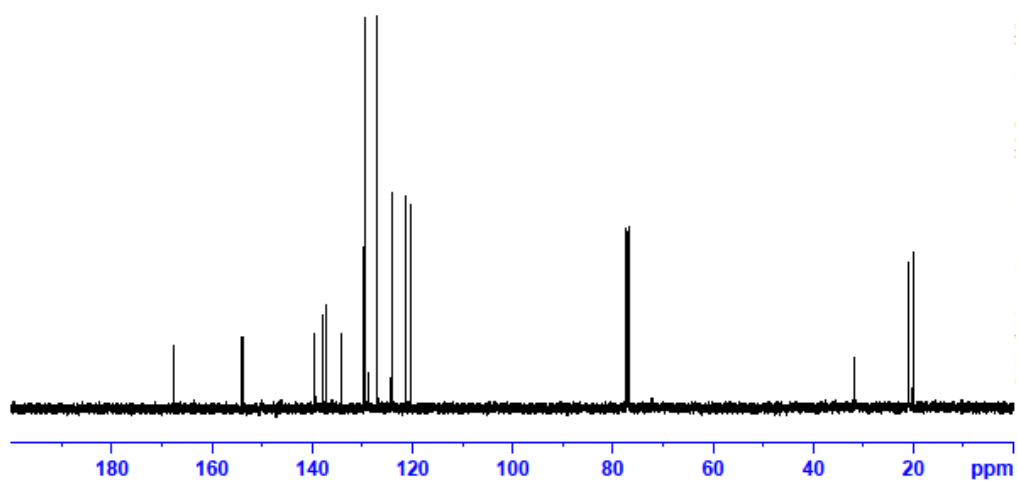
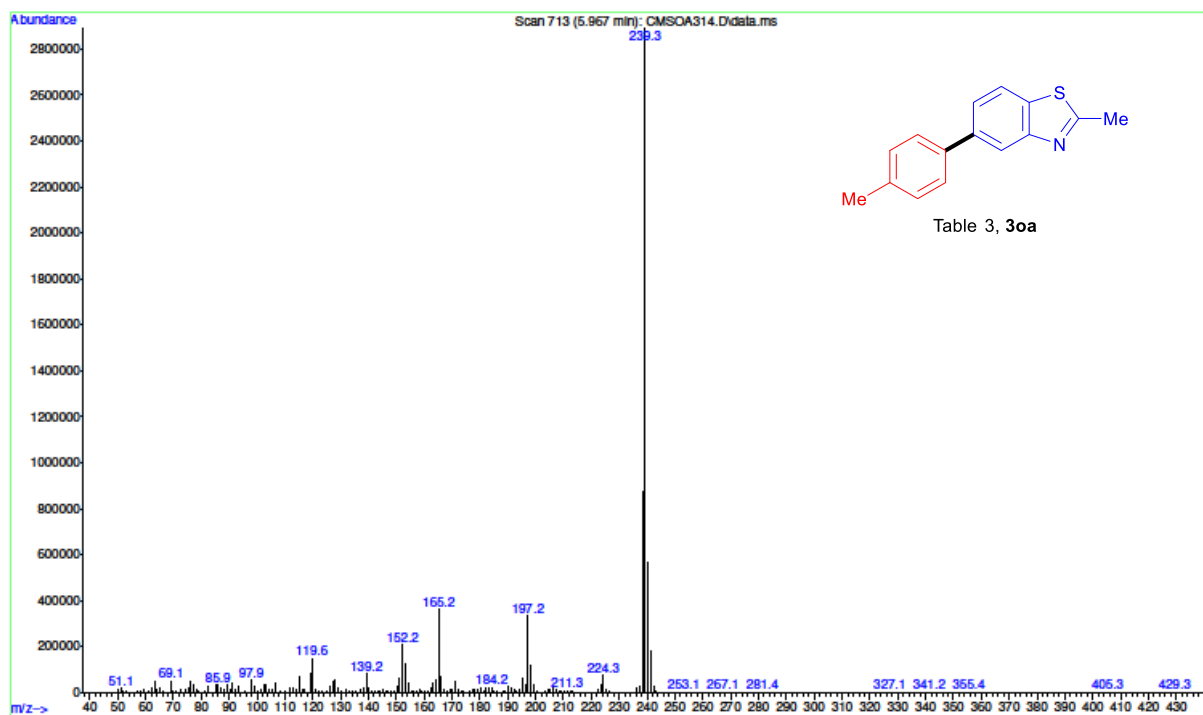
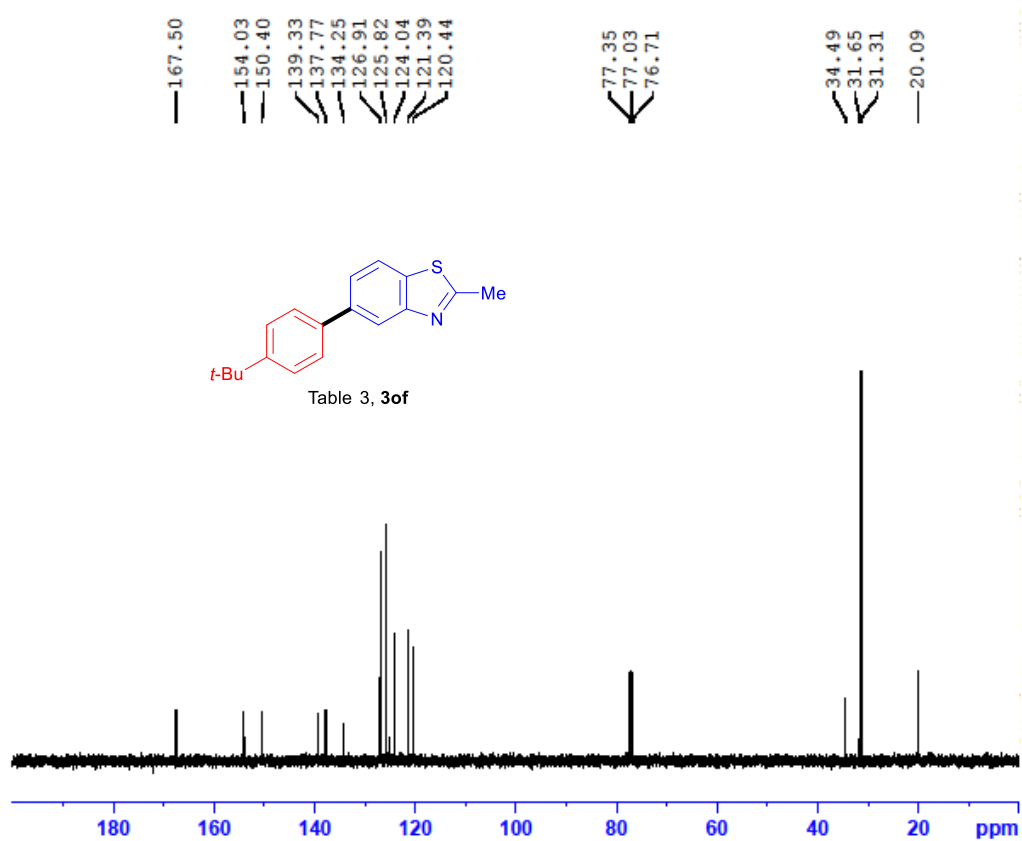
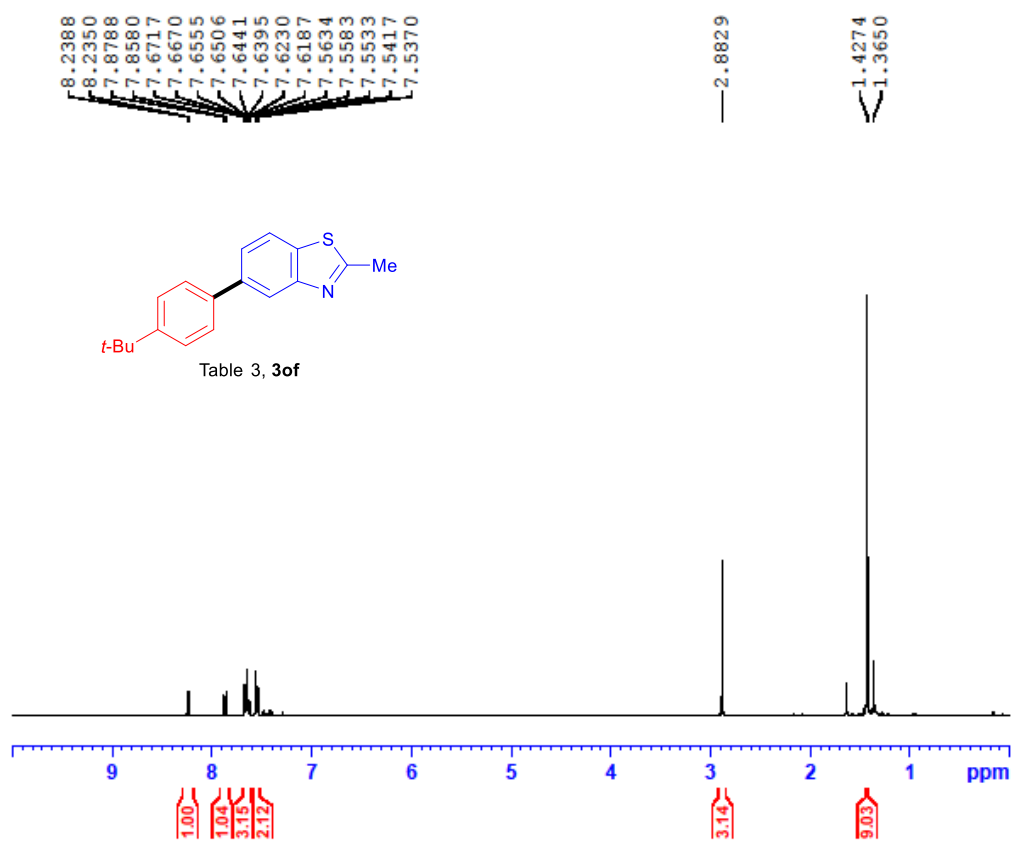
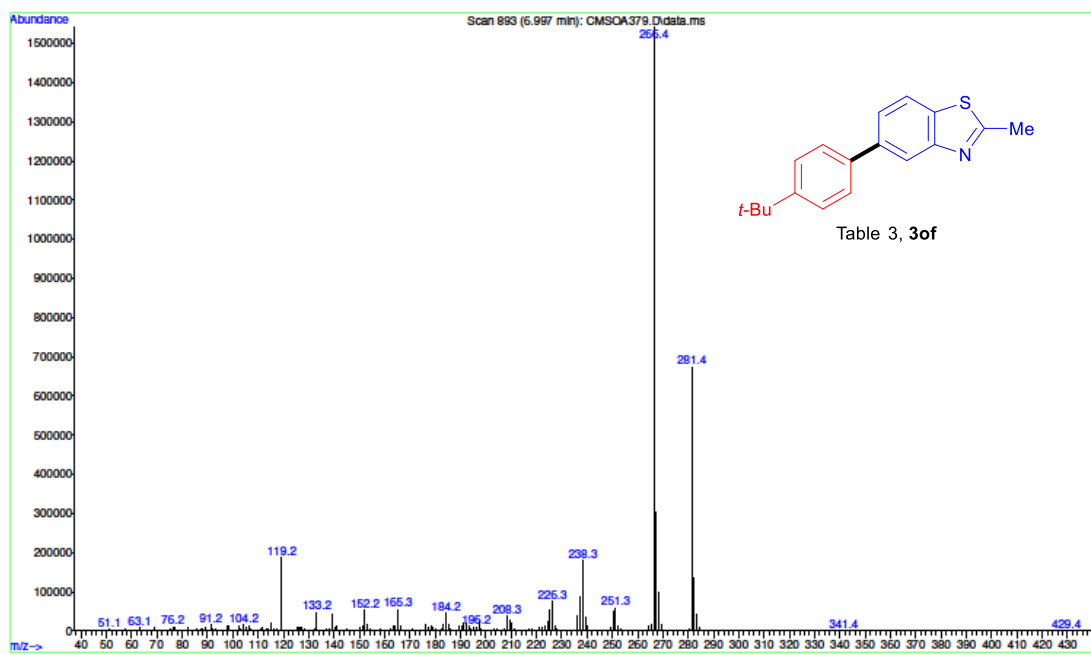


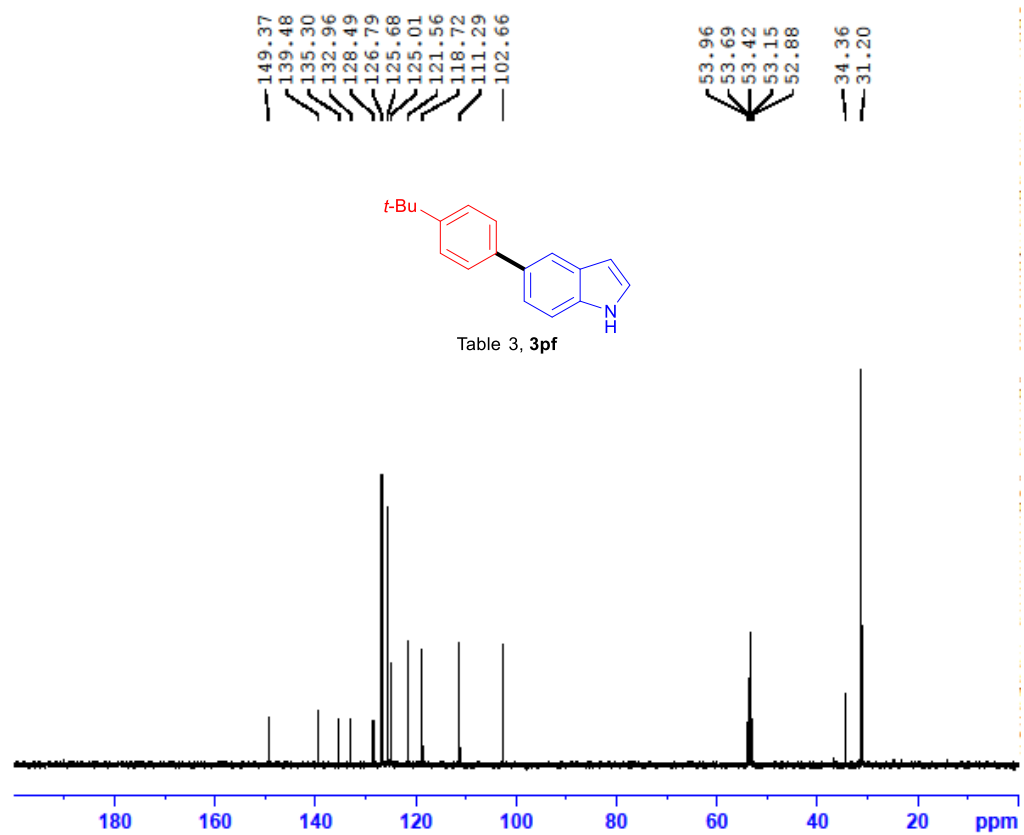
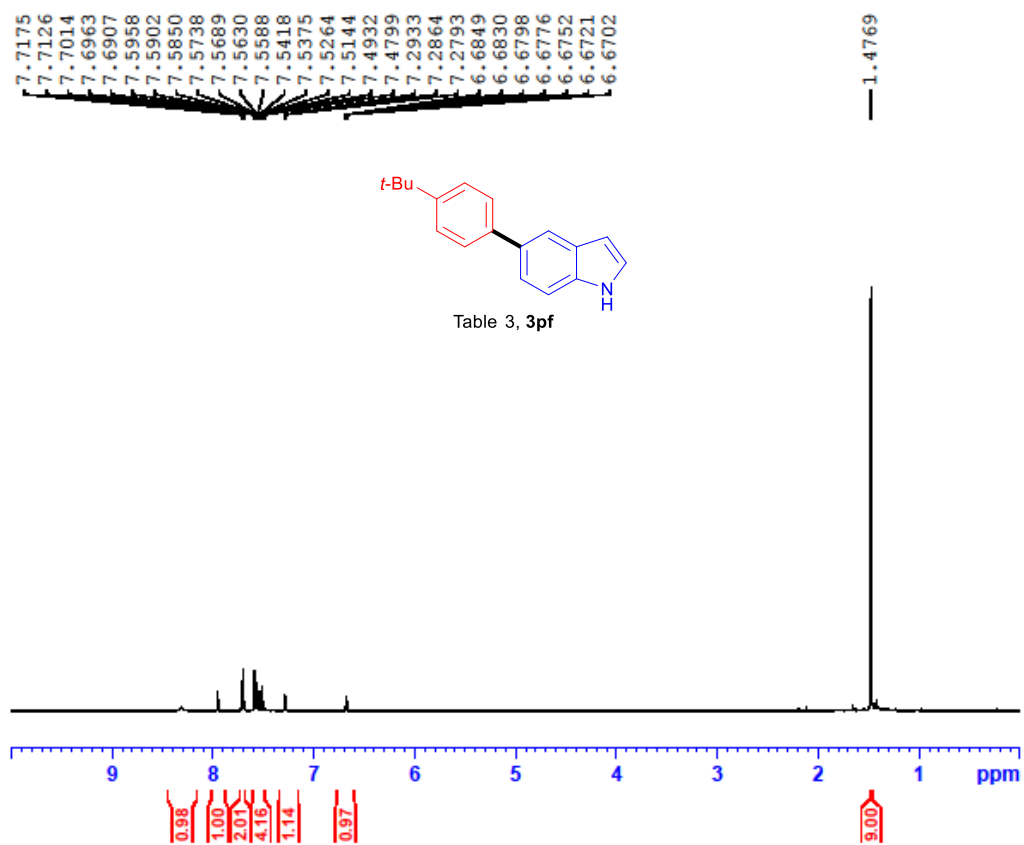
Table 3, 3oa

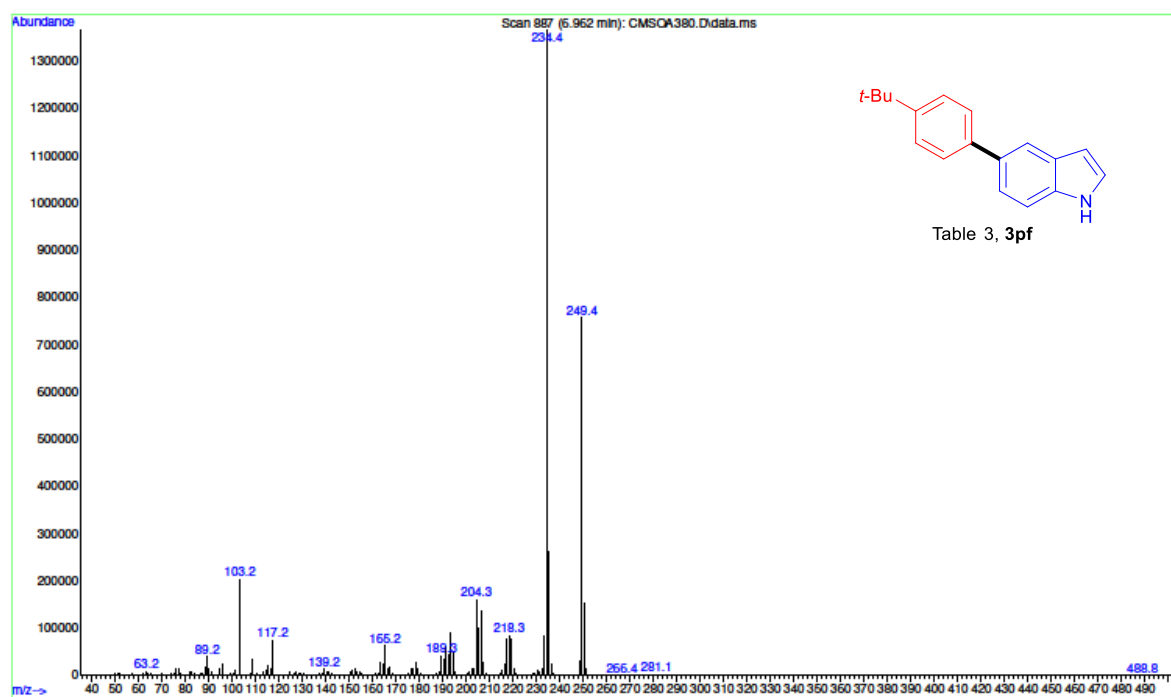












8.1640
7.9795
7.9586
7.8157
7.7951
7.4891
7.4699
7.4504
7.4058
7.3870
7.3601
7.0184
7.0002

3.9463

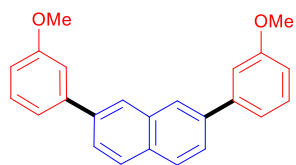
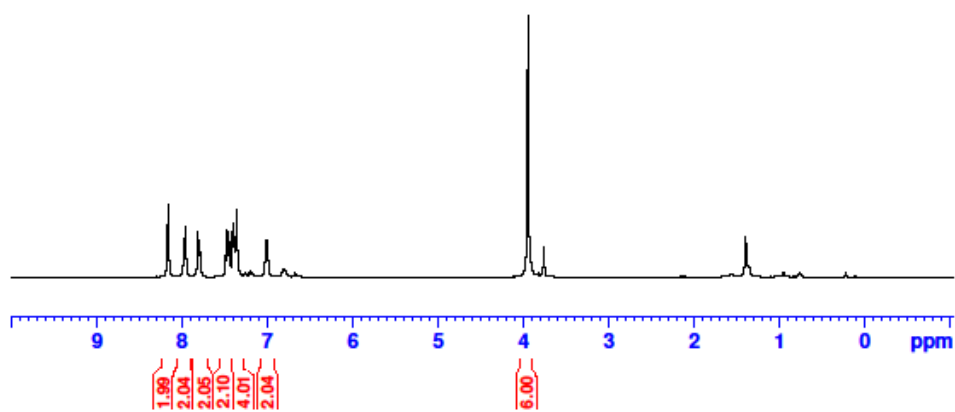


Table 3, 3qc



160.11
142.60
138.91
133.91
132.02
129.92
128.20
126.19
125.77
119.97
113.20
112.87

77.44
77.12
76.80

55.35

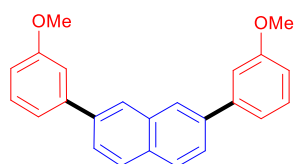
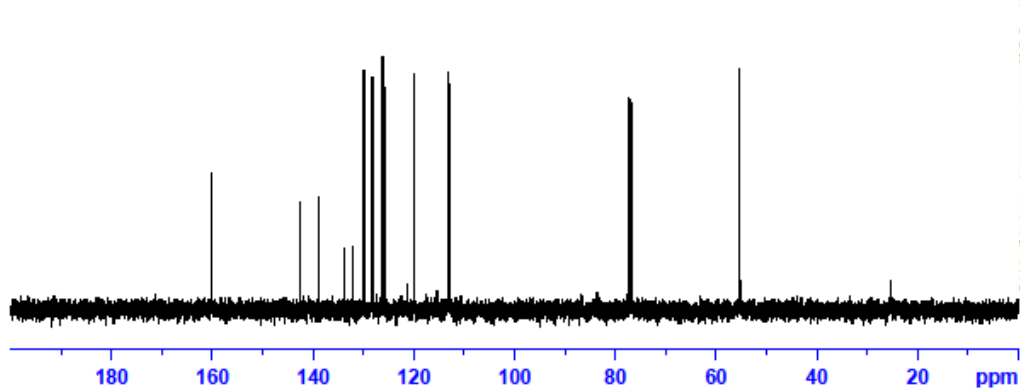
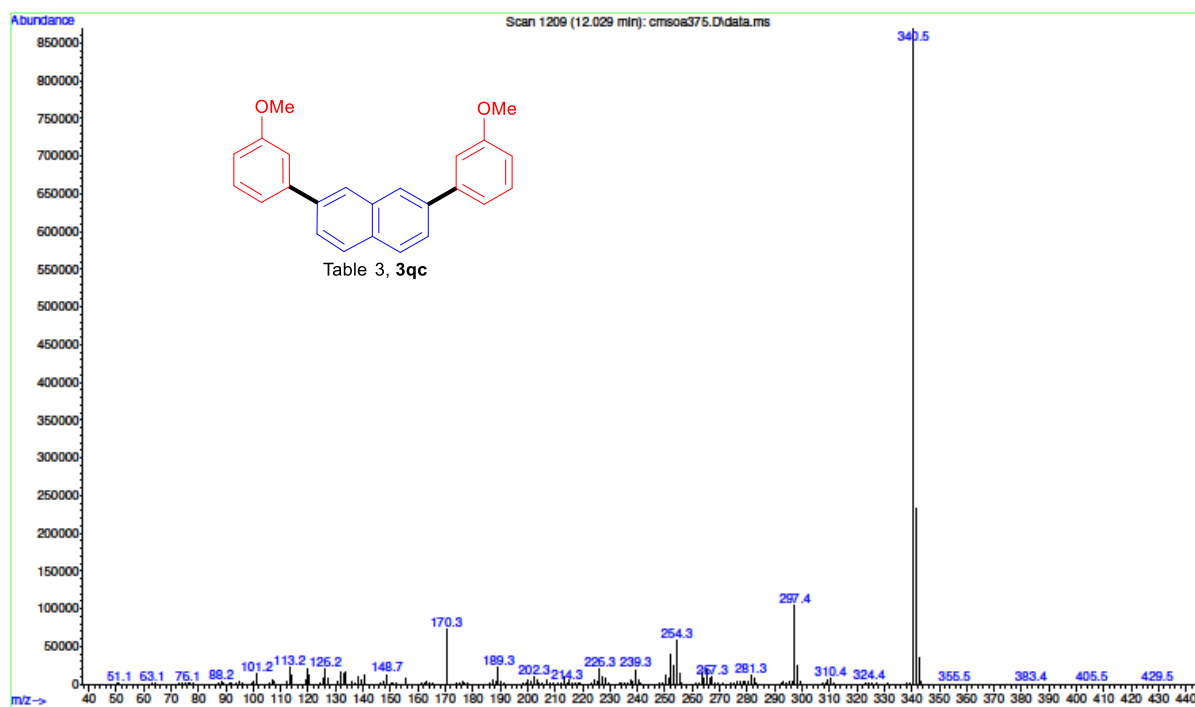
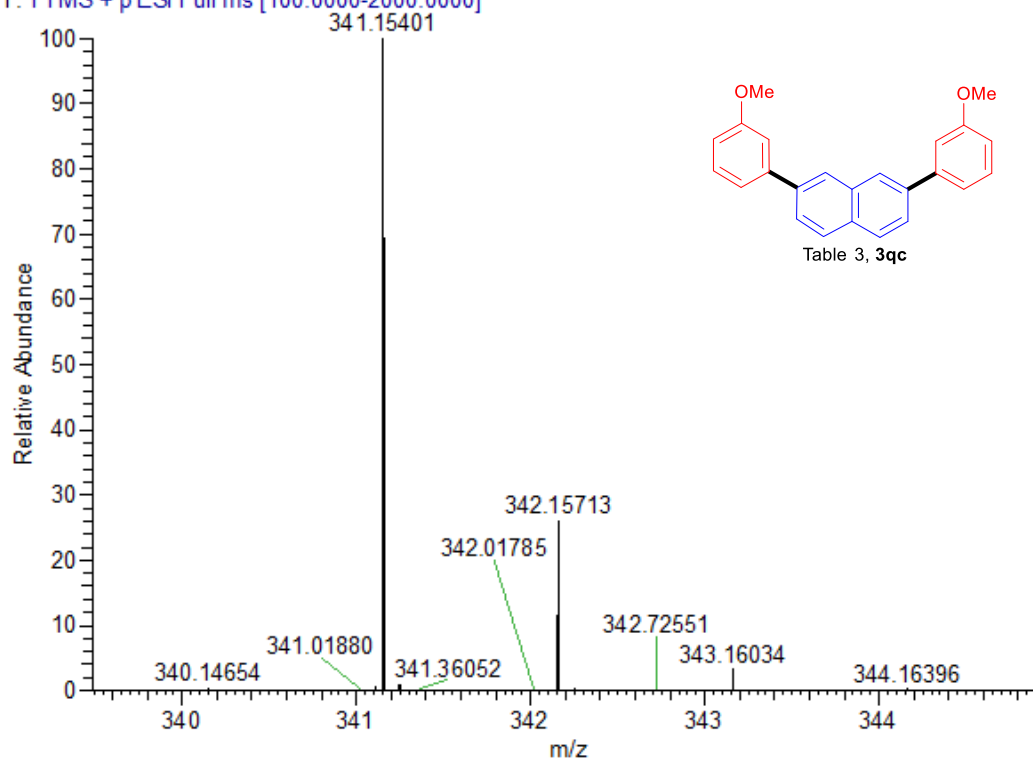


Table 3, 3qc





Kin-Dept 11012017 S5 #78-107 RT: 0.38-0.52 AV: 30 NL: 2.11E8
T: FTMS + p ESI Full ms [100.0000-2000.0000]



Mass	Calc. Mass	mDa	PPM	Formula
341.1540	341.1536	0.4	1.1	C ₂₄ H ₂₁ O ₂

7.5792
7.5588
7.3082
7.2880
7.2638
7.2606
7.2485
7.2453
7.2303
6.9521

3.0795
3.0603
3.0391
2.8686
2.8482
2.8280
2.4879

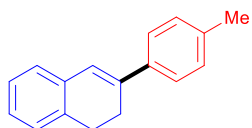
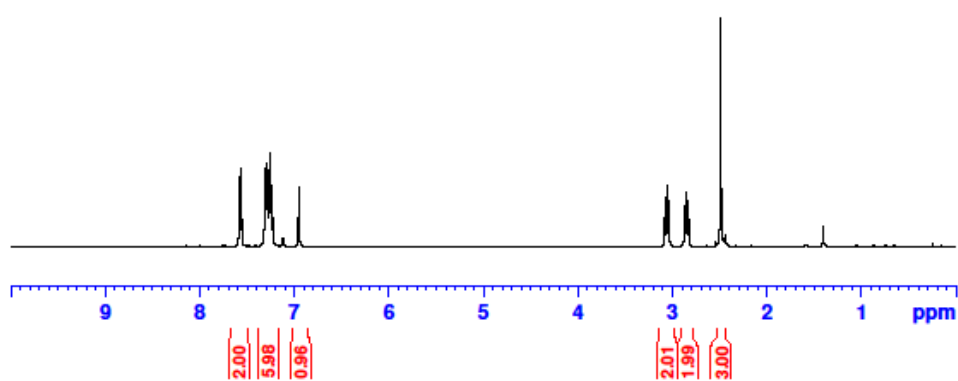


Table 4, 5a



138.51
138.12
137.07
134.82
134.67
129.10
127.12
126.72
126.52
126.42
124.98
123.45

77.32
77.00
76.68

28.15
26.25
21.10

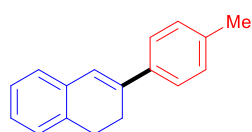
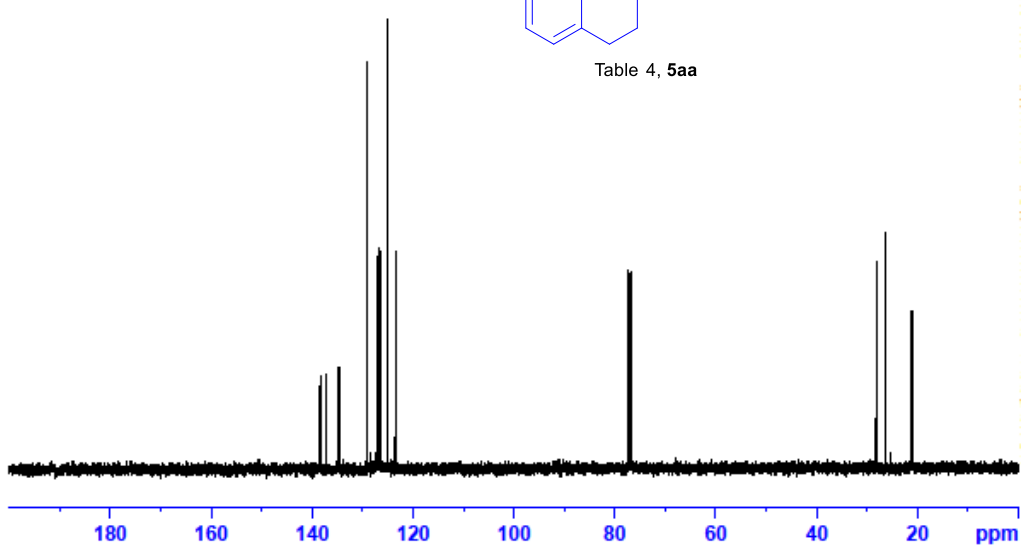
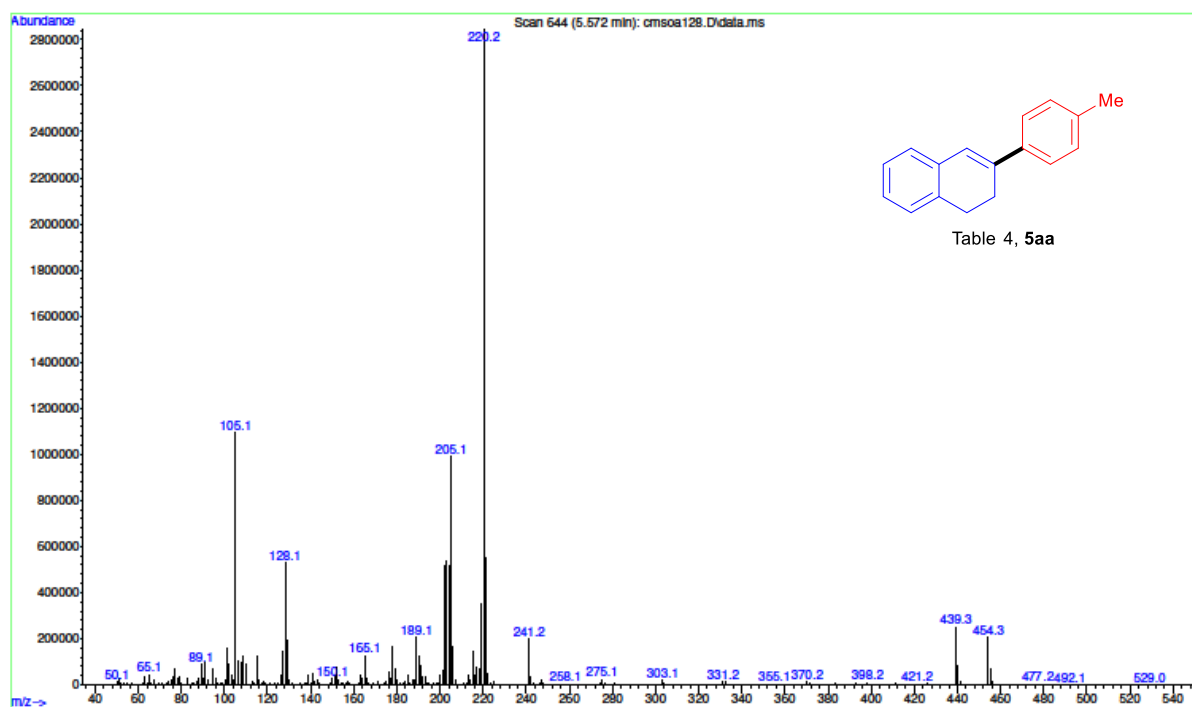


Table 4, 5a





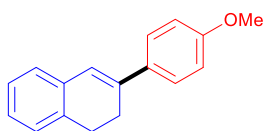
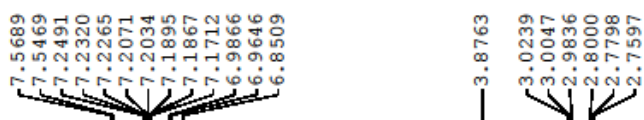


Table 4, 5ab

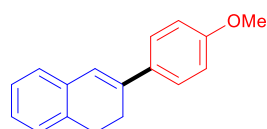
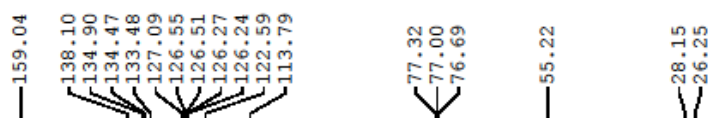
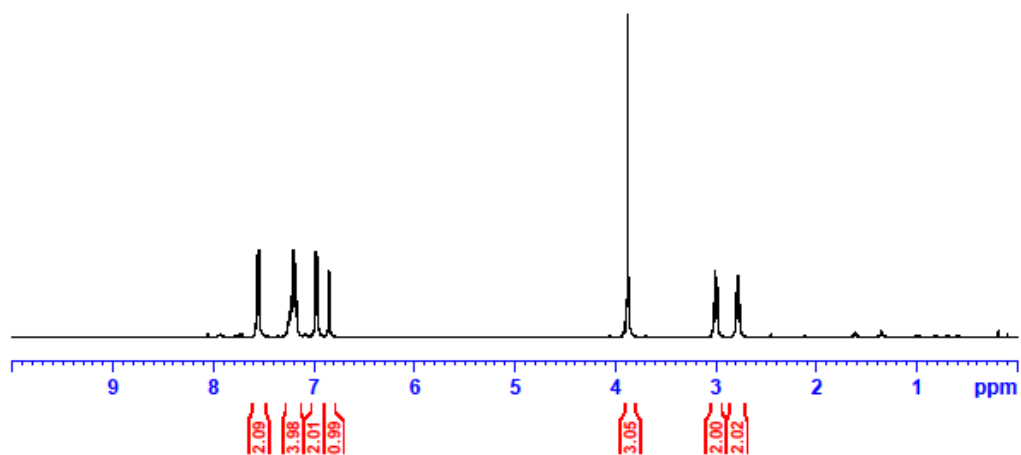
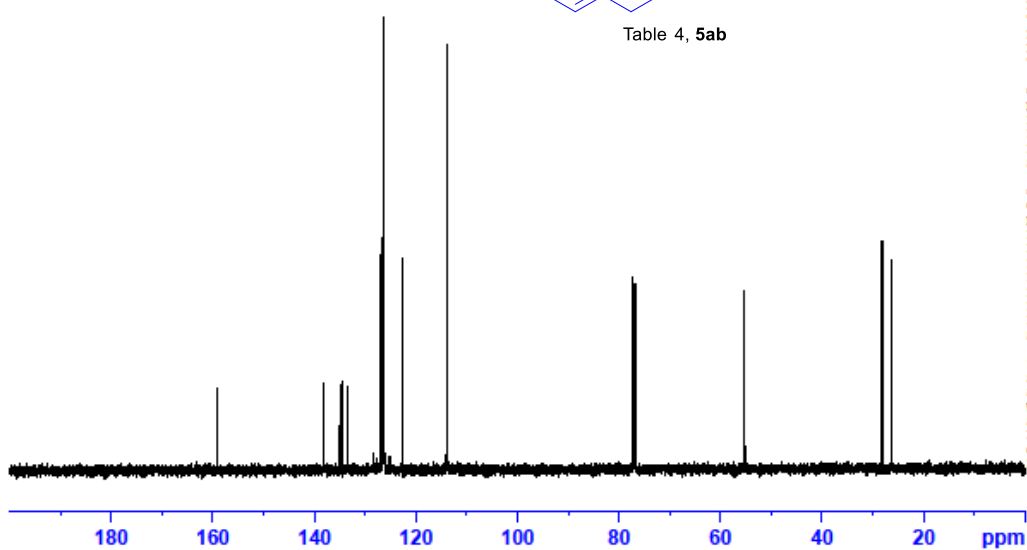
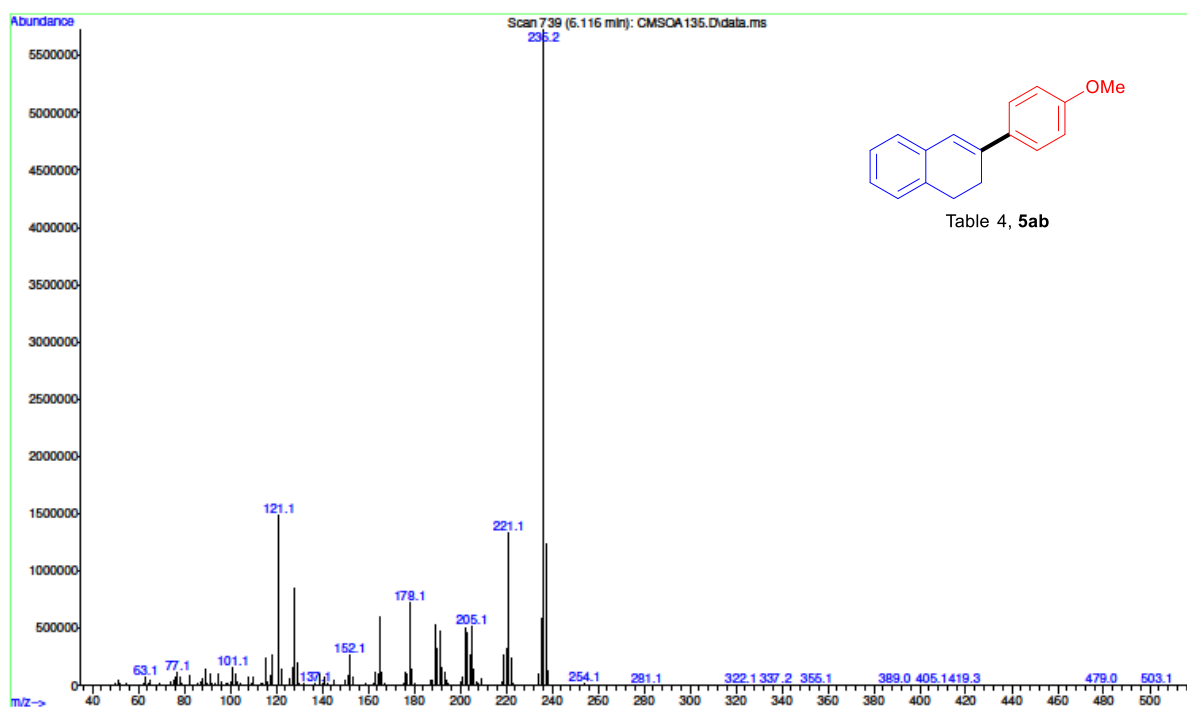
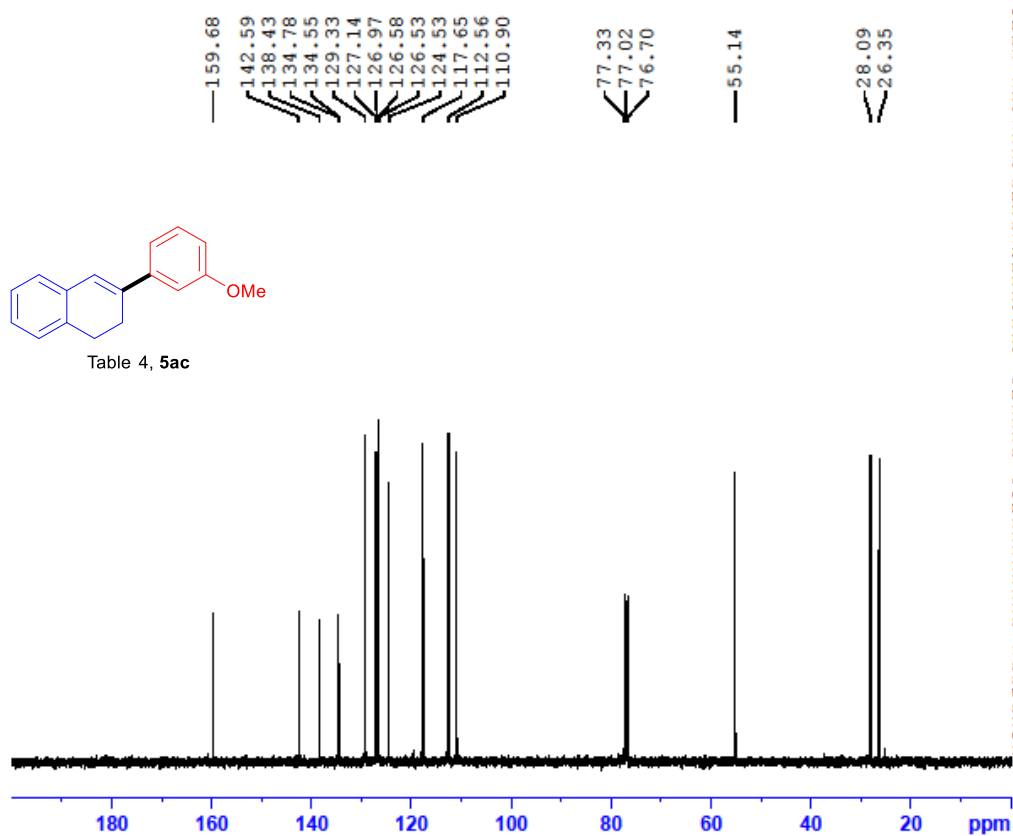
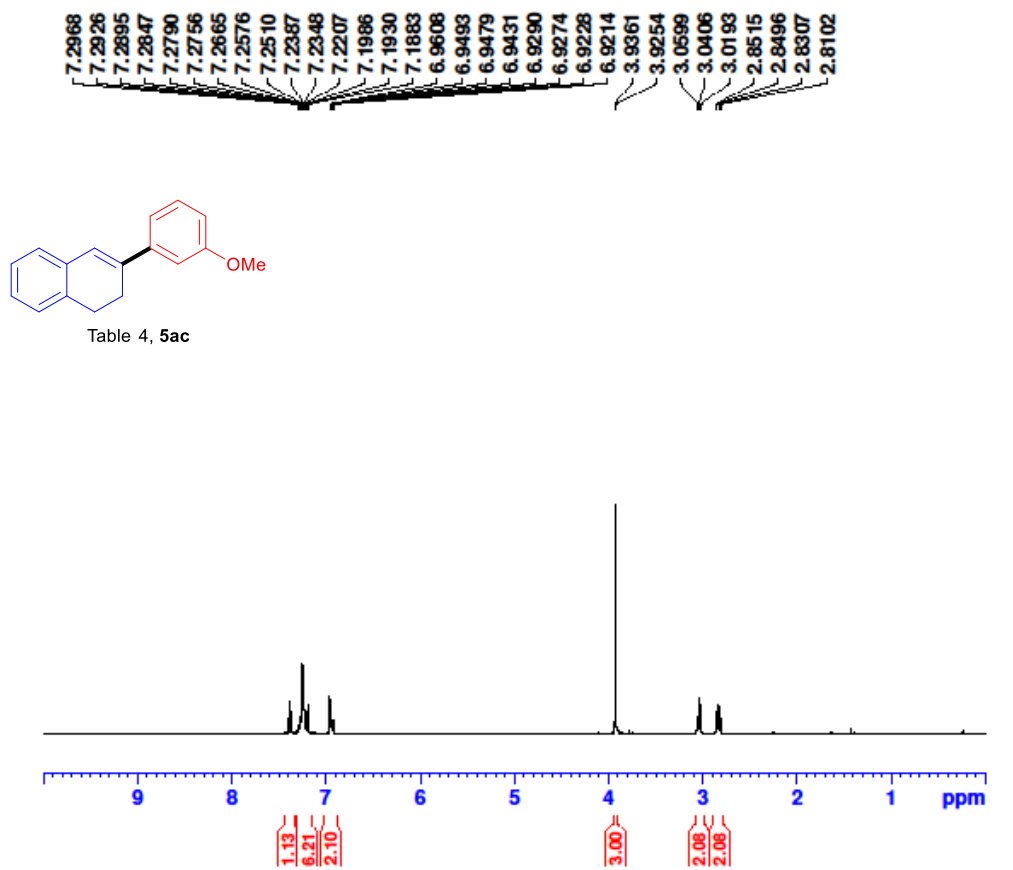
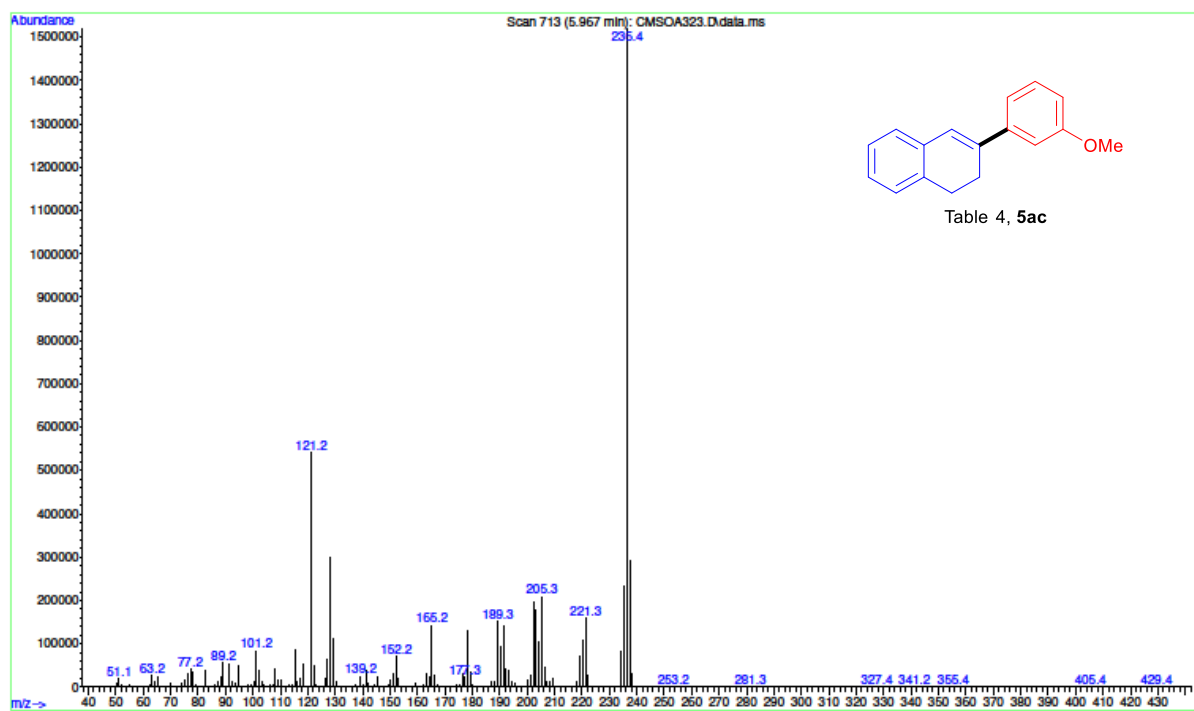


Table 4, 5ab









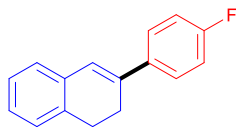
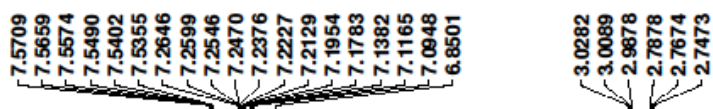


Table 4, 5ad

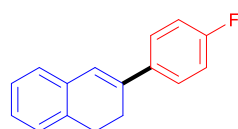
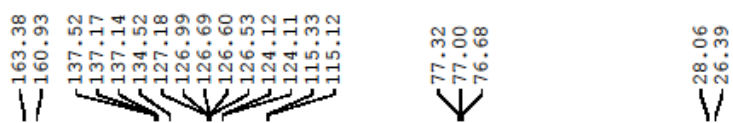
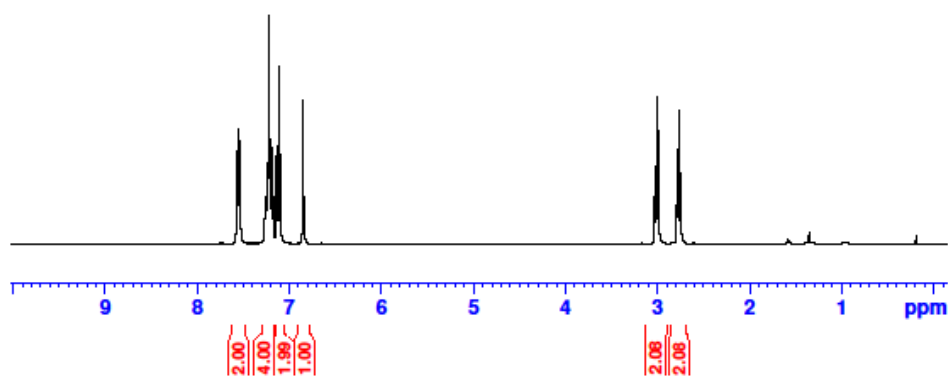
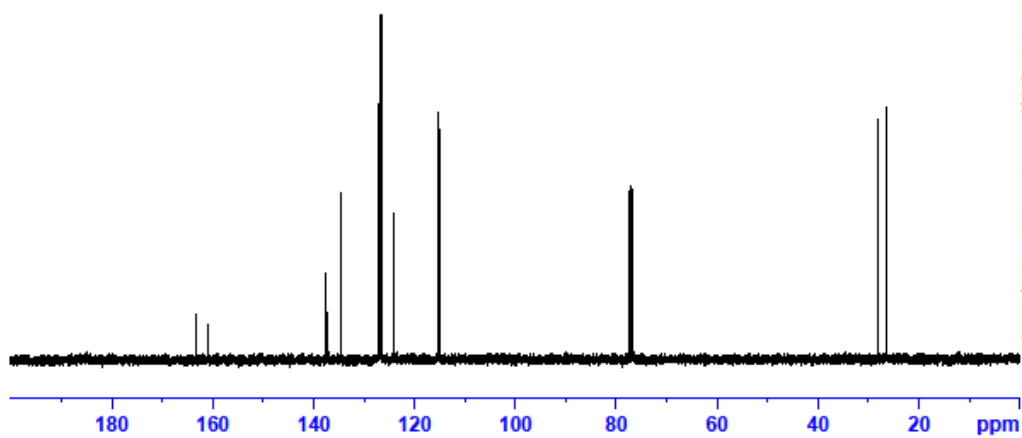
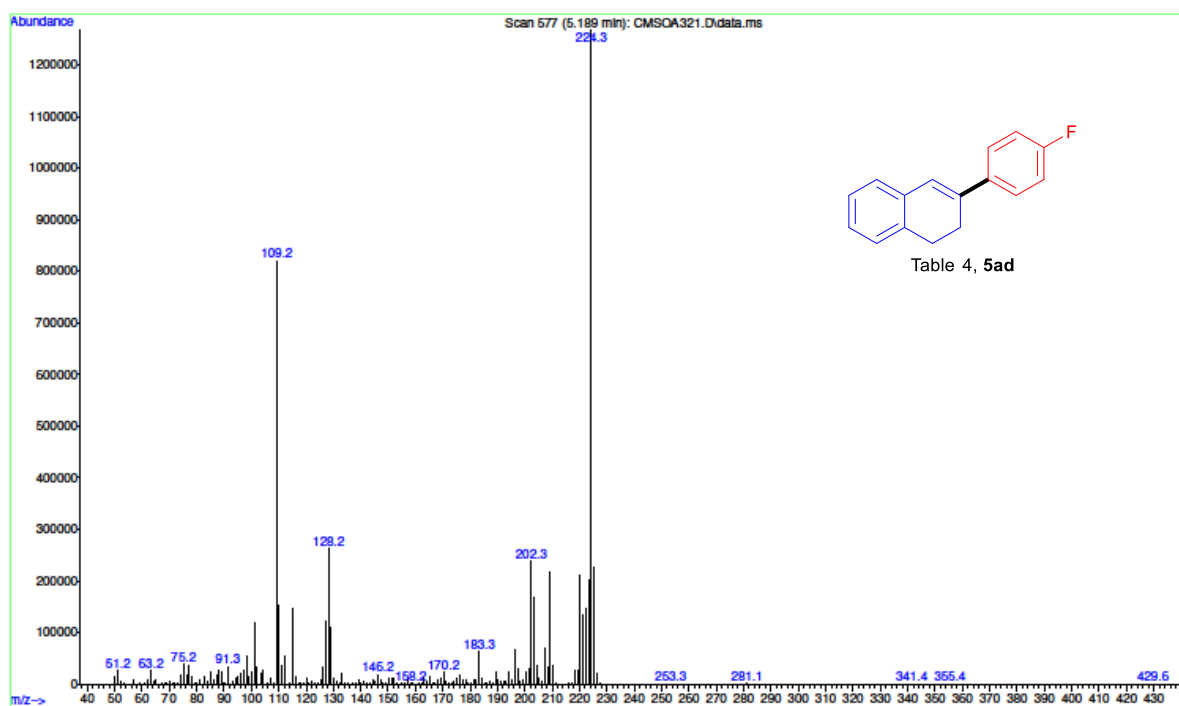
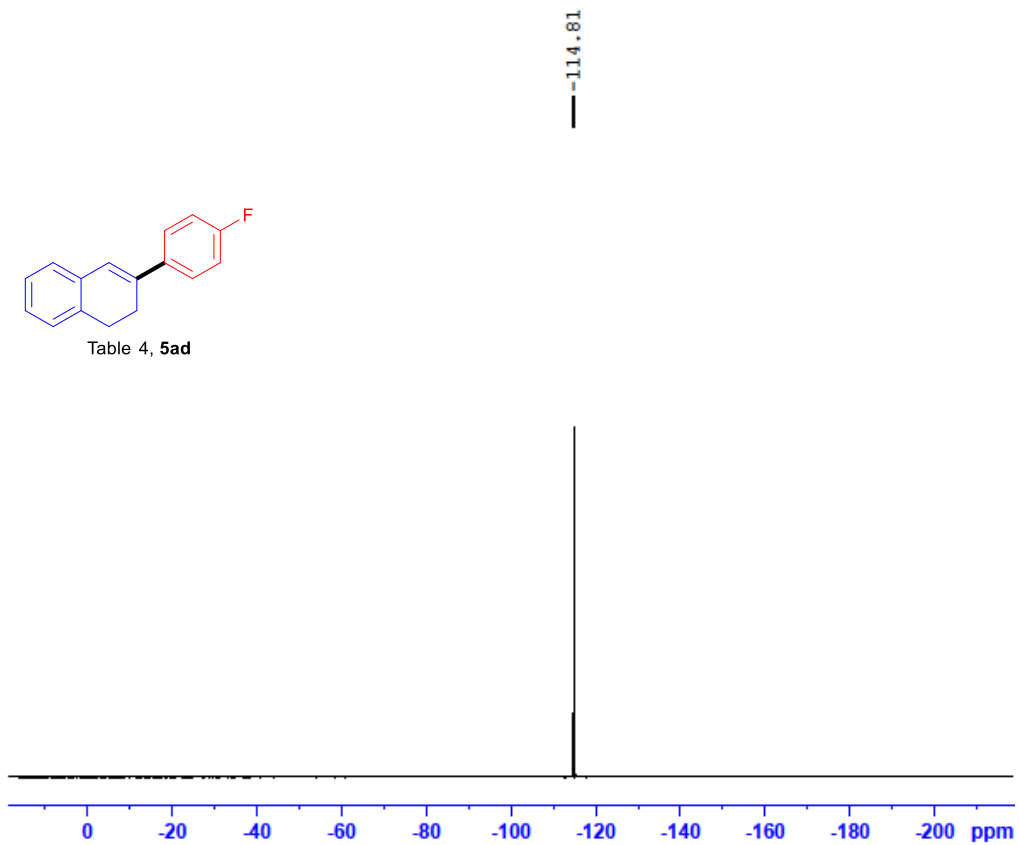


Table 4, 5ad





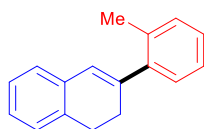


Table 4, 5ae

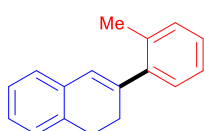
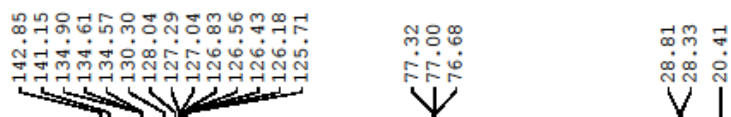
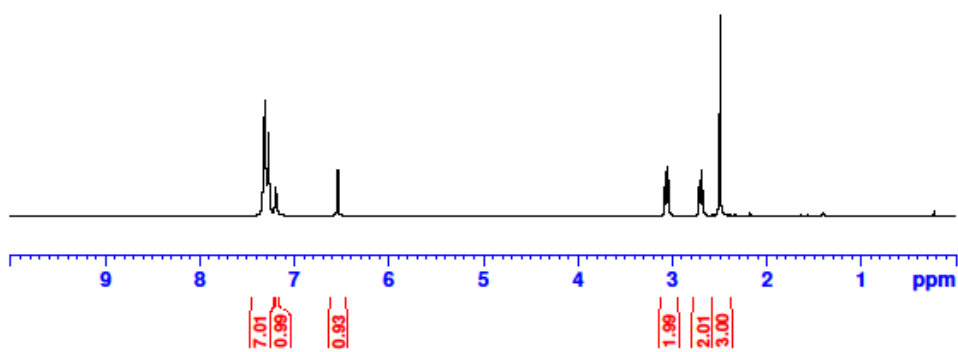
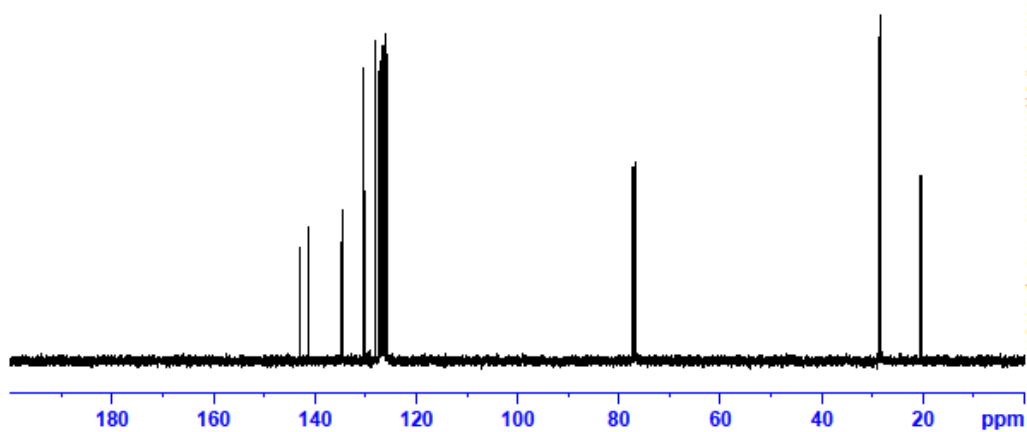
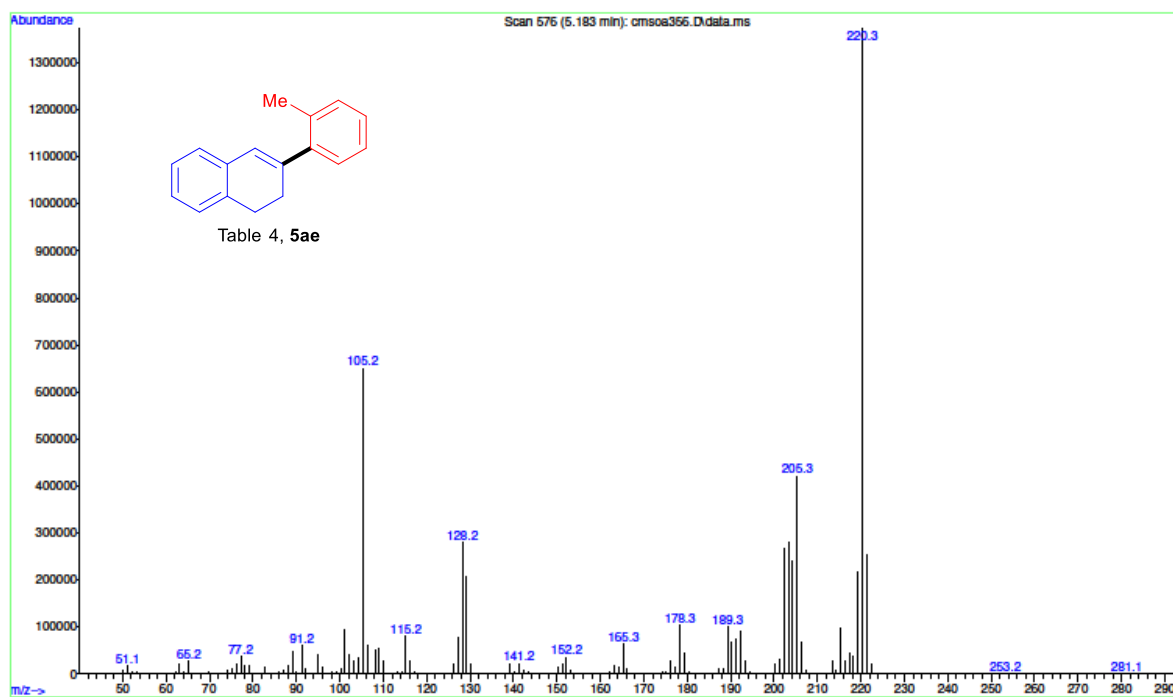
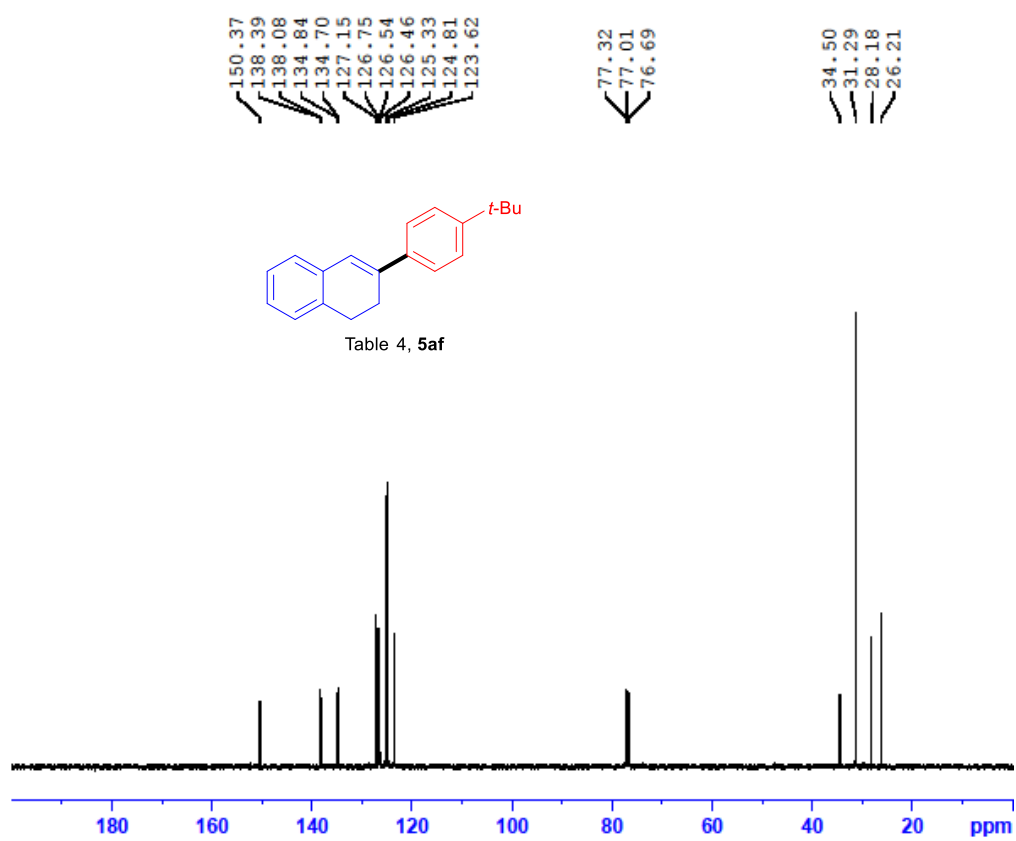
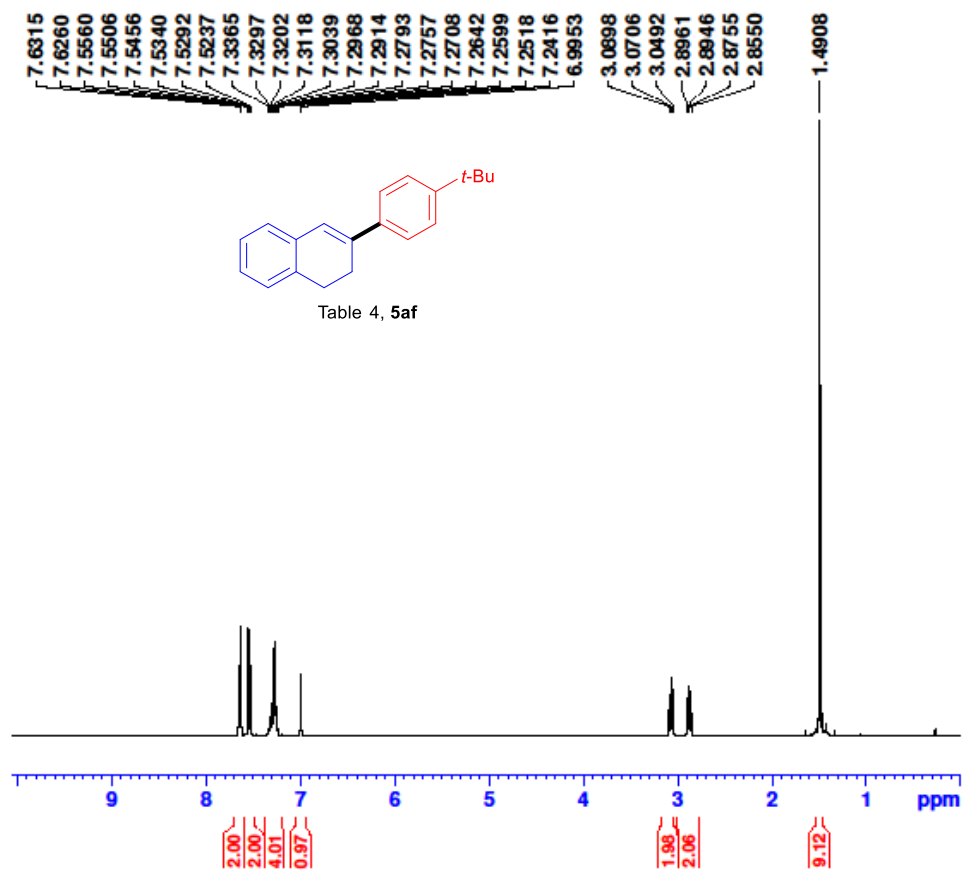
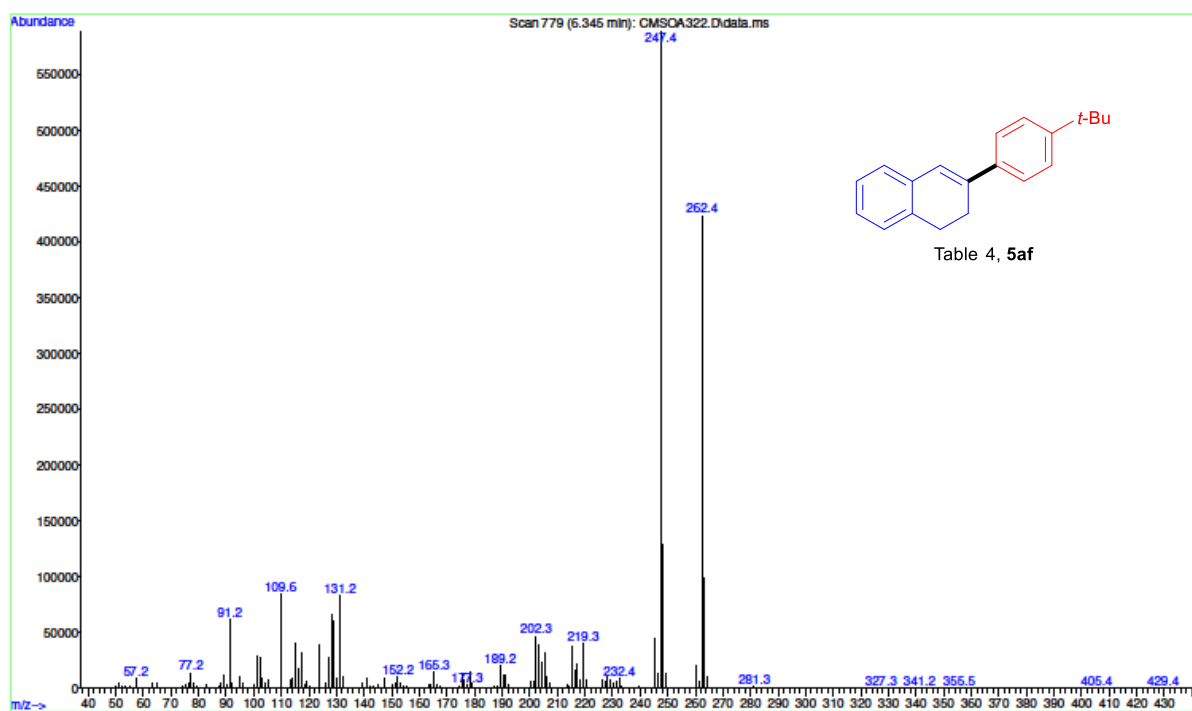


Table 4, 5ae









7.6622
7.2688
7.2603
7.2530
7.2365
7.2264
7.2151
7.2096
7.1952
6.9630

3.0344
3.0152
2.9939
2.8102
2.7897
2.7696

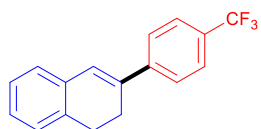
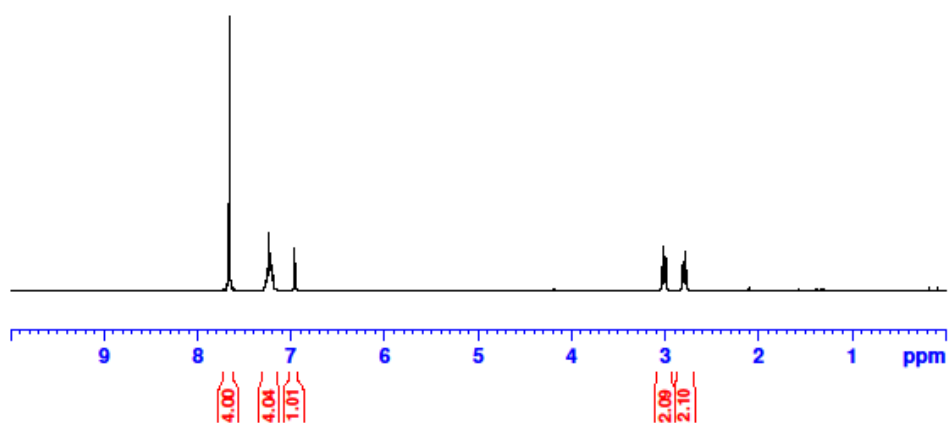


Table 4, 5ag



144.64
137.15
134.89
134.16
129.55
129.23
128.91
128.58
128.37
127.64
127.34
127.01
126.75
126.33
125.67
125.45
125.41
125.37
125.34
125.26
122.97
77.35
77.04
76.72

27.99
26.22

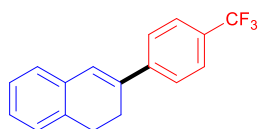
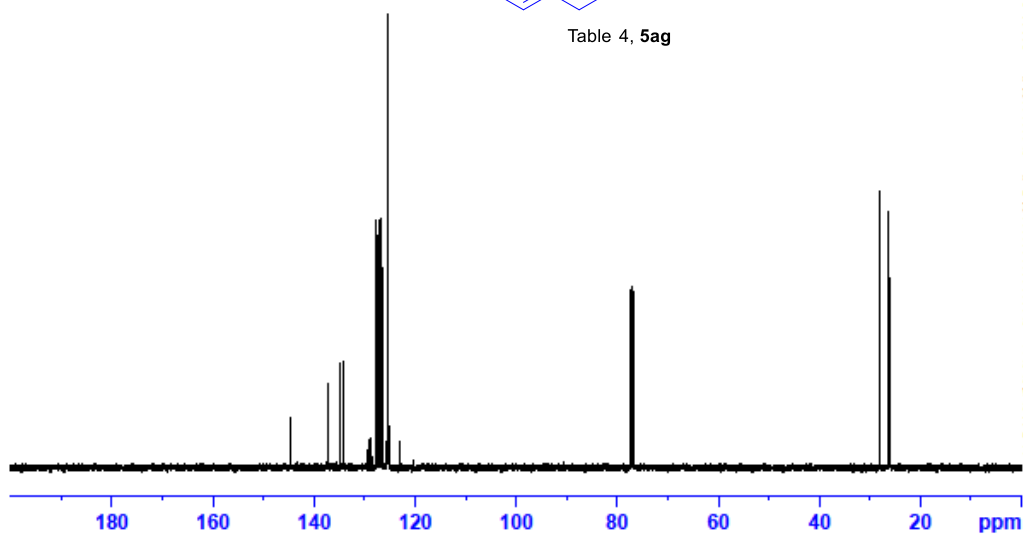
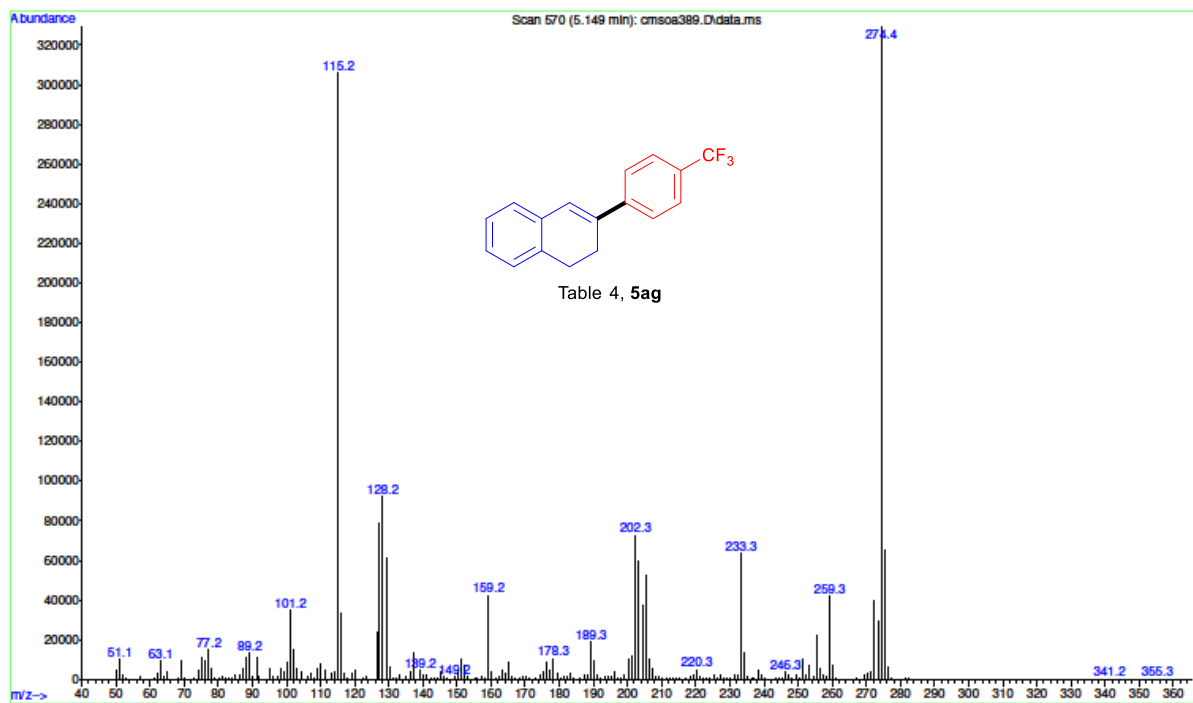
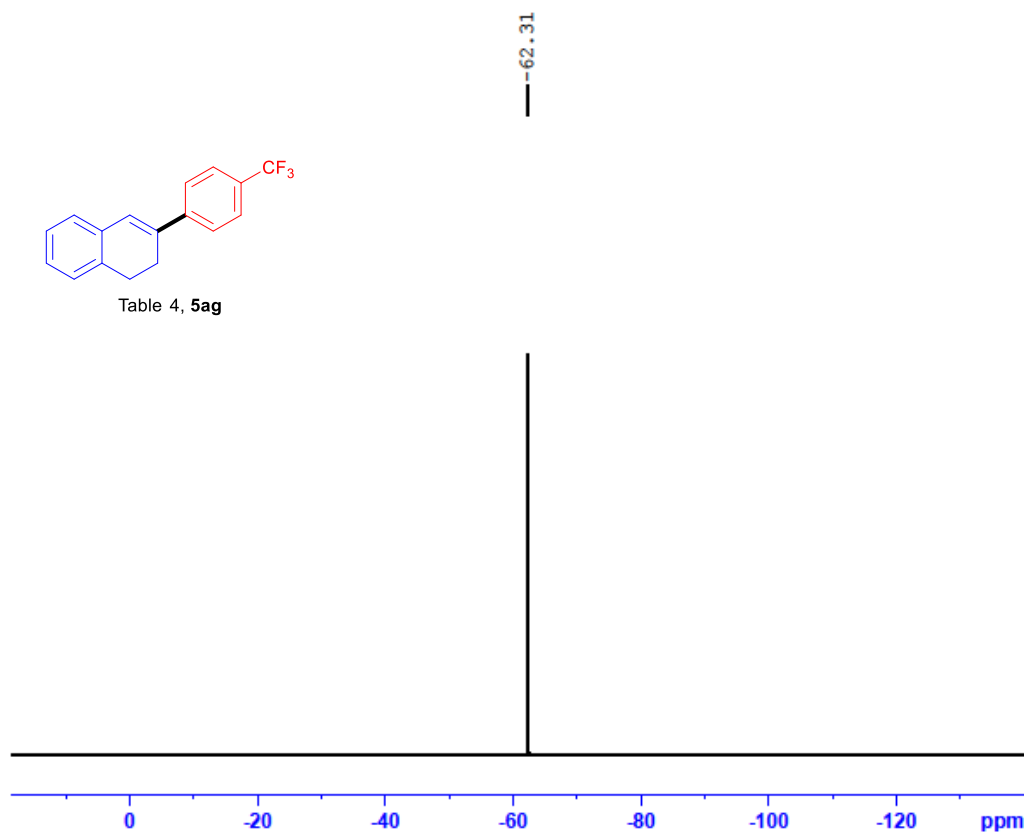


Table 4, 5ag





7.4033
7.3843
7.3664
7.3062
7.2902
7.2605
7.1348
7.1133
7.0901
7.0716
7.0492
6.9769
6.9589
6.7602
6.7389

3.7785

2.1708

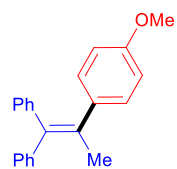
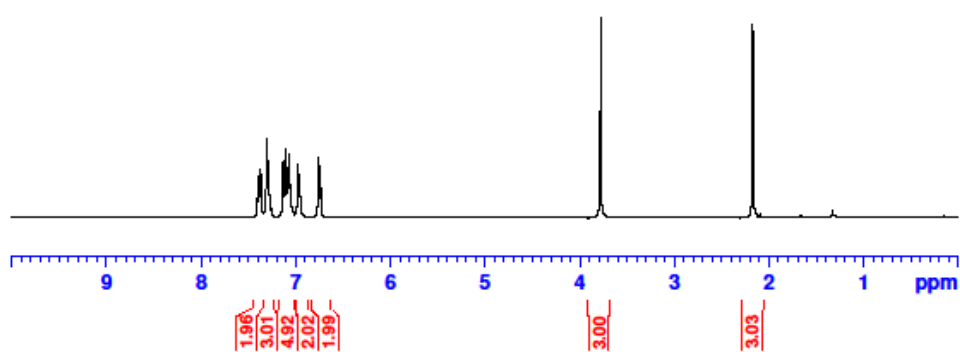


Table 4, 5bb



157.88
143.77
143.32
138.73
136.16
135.04
130.83
130.36
129.98
128.06
127.43
126.41
125.63
113.21

77.32
77.00
76.68

55.05

23.26

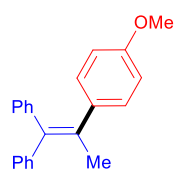
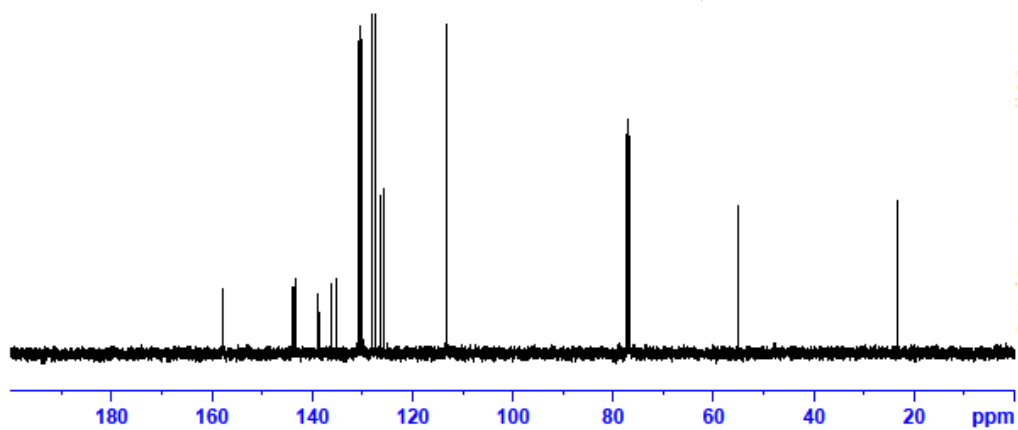
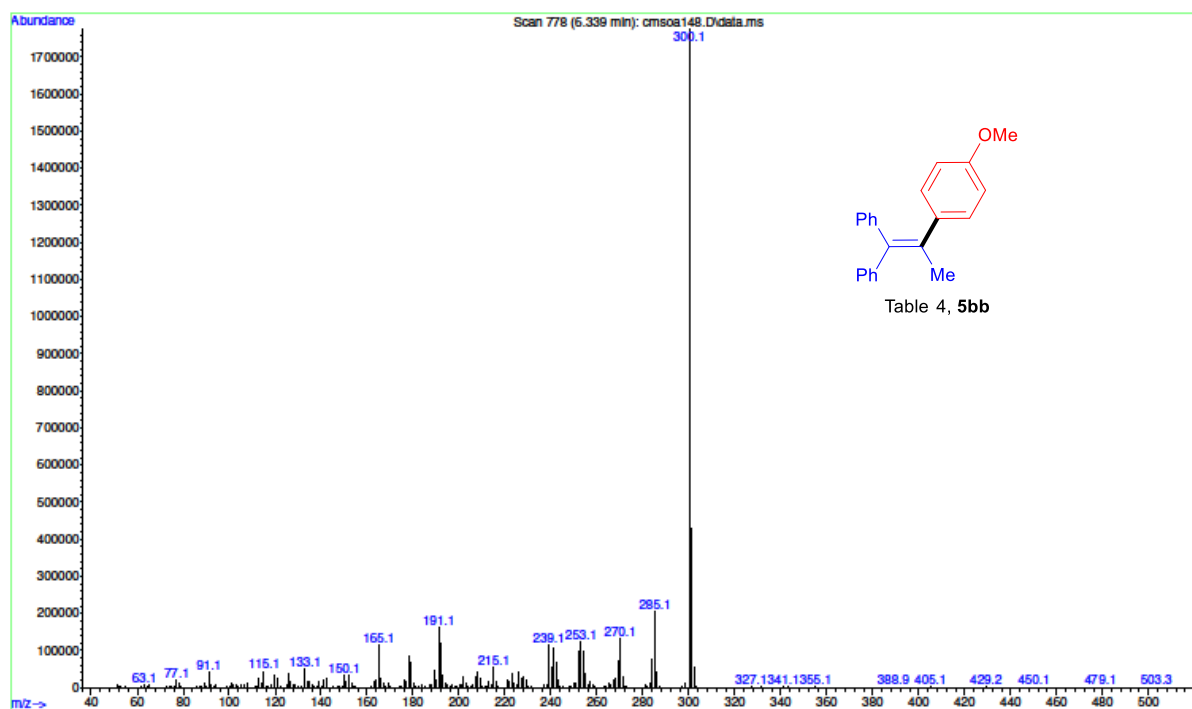
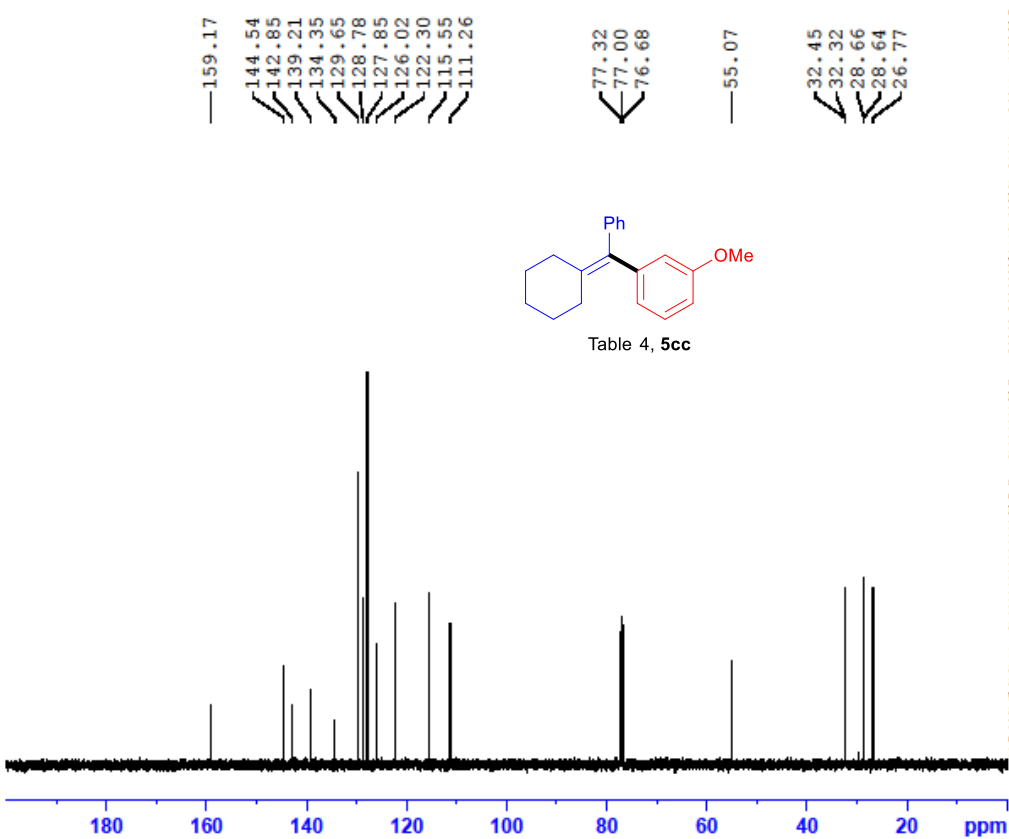
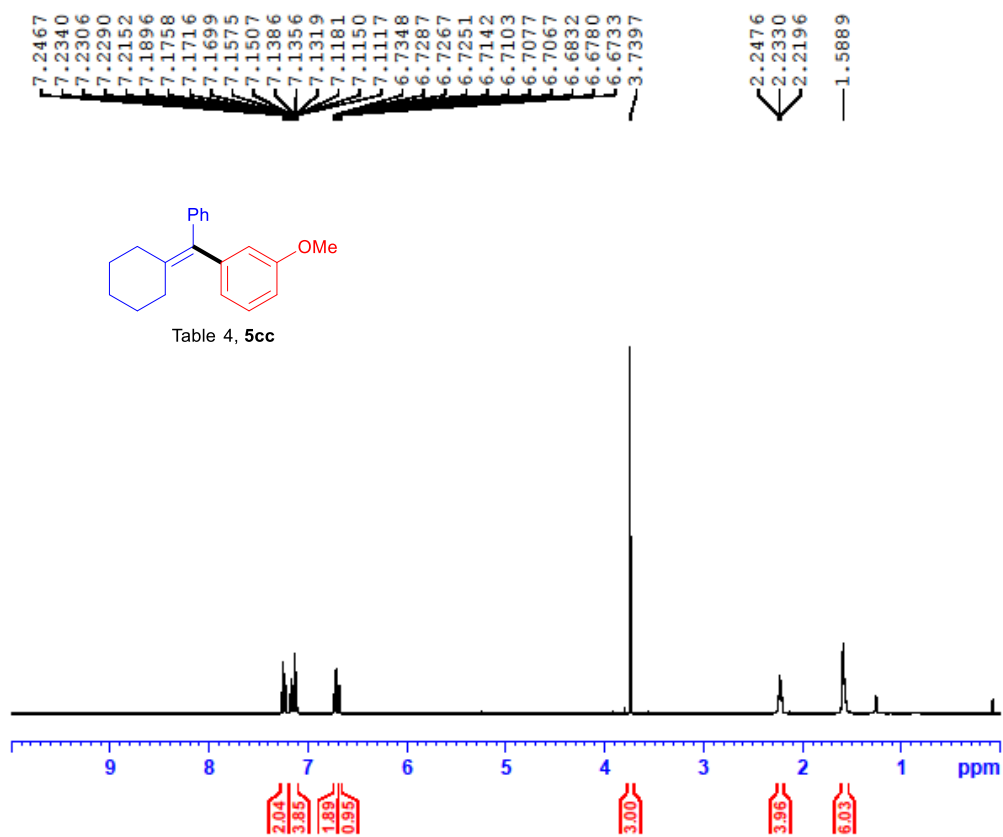
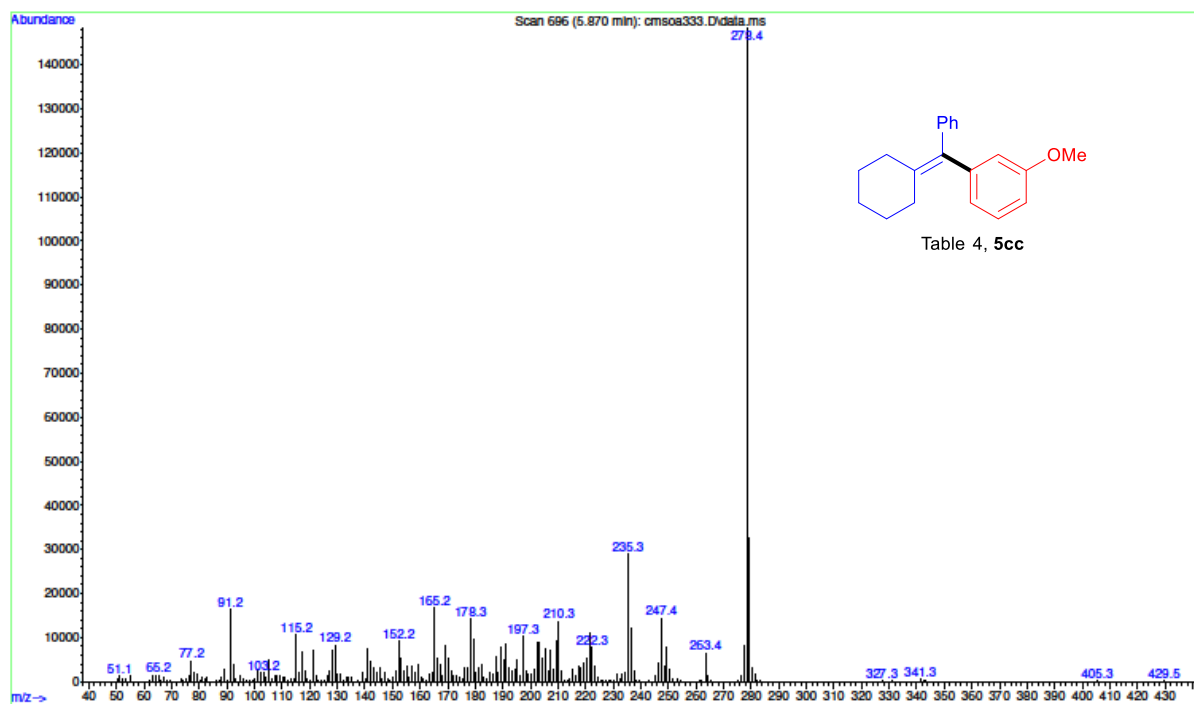


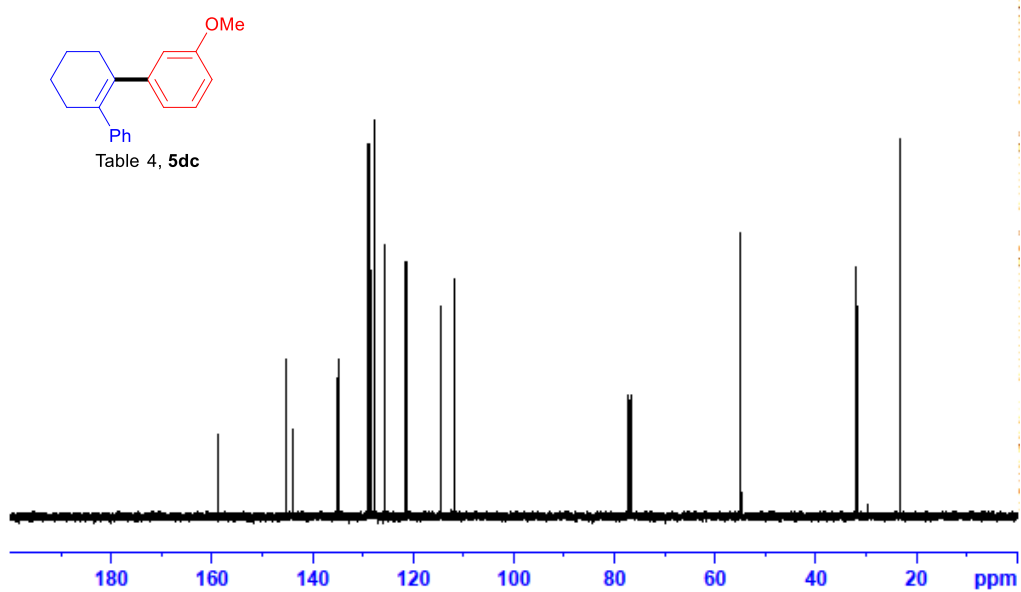
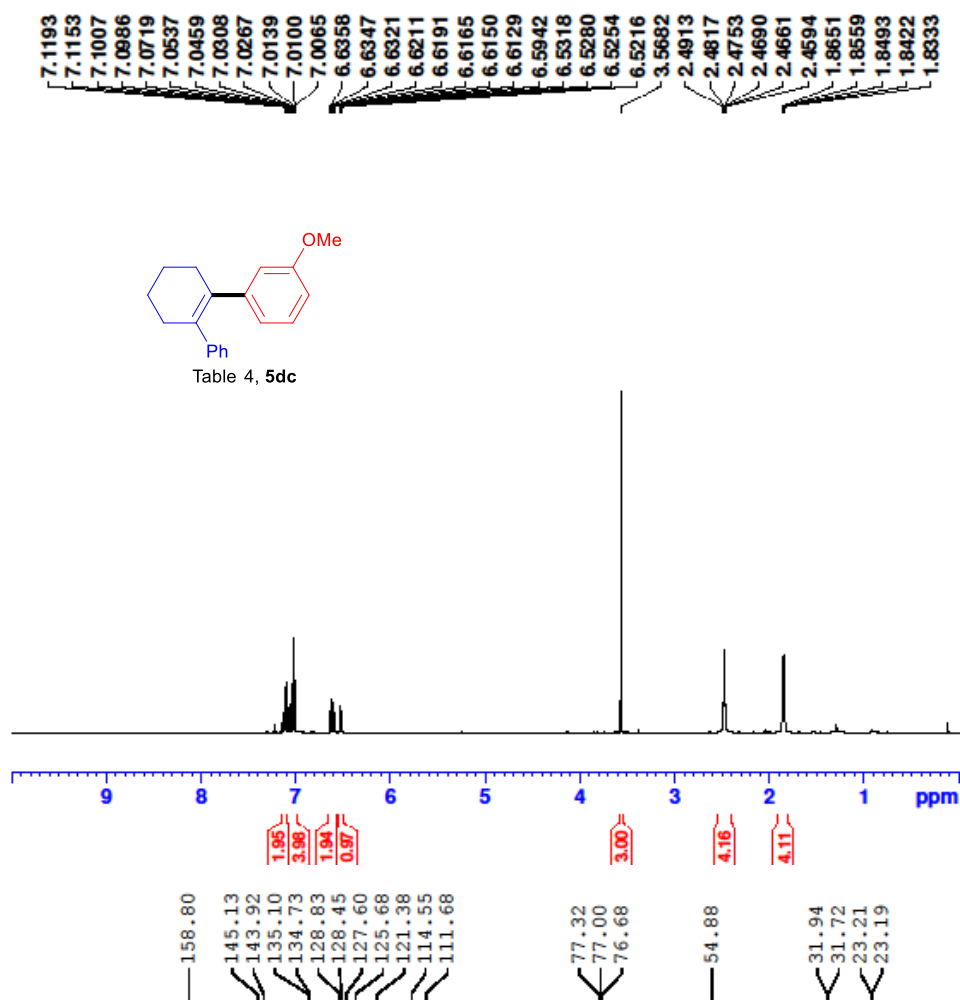
Table 4, 5bb

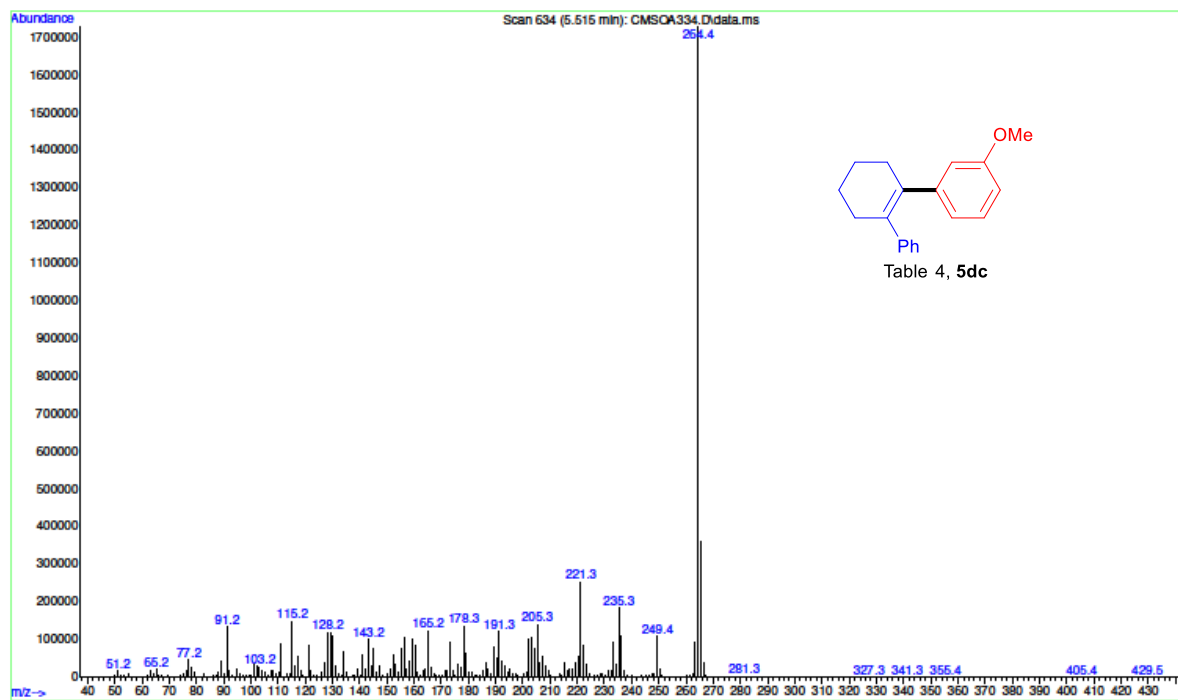


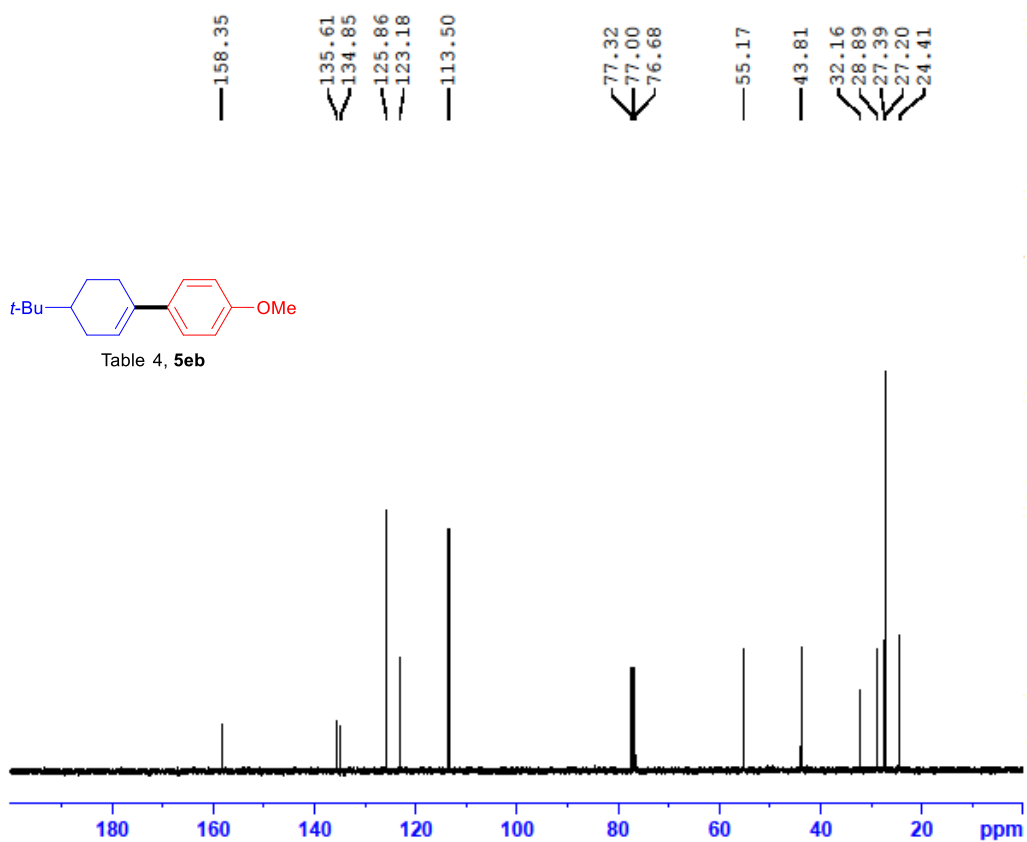
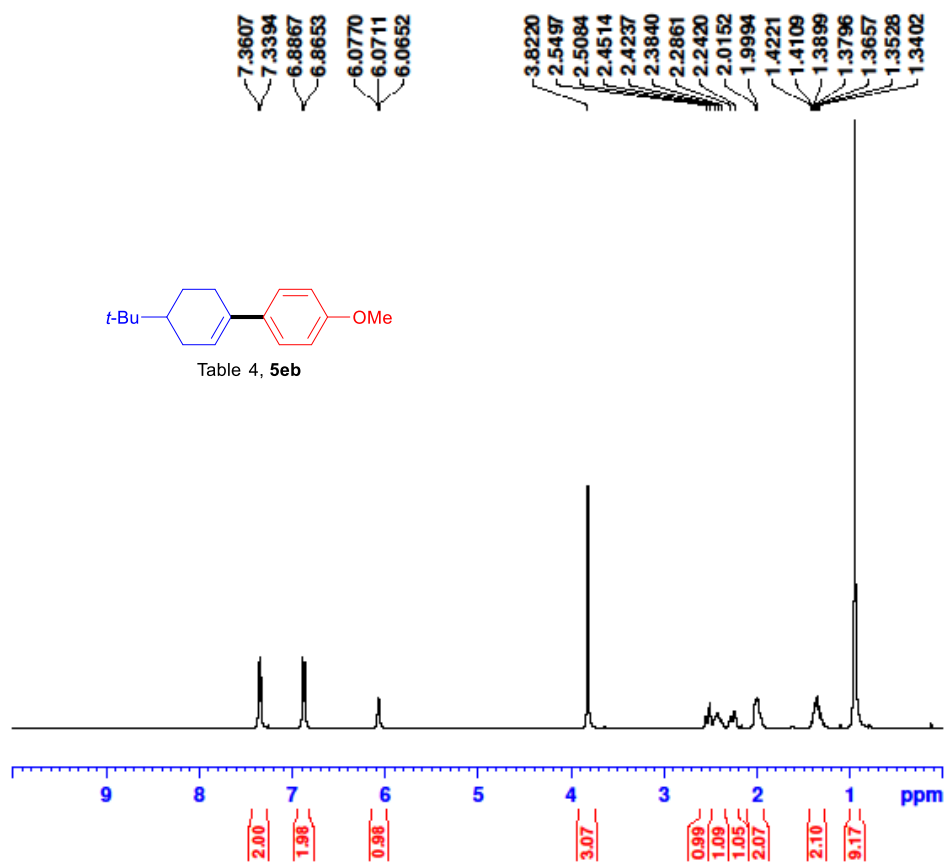


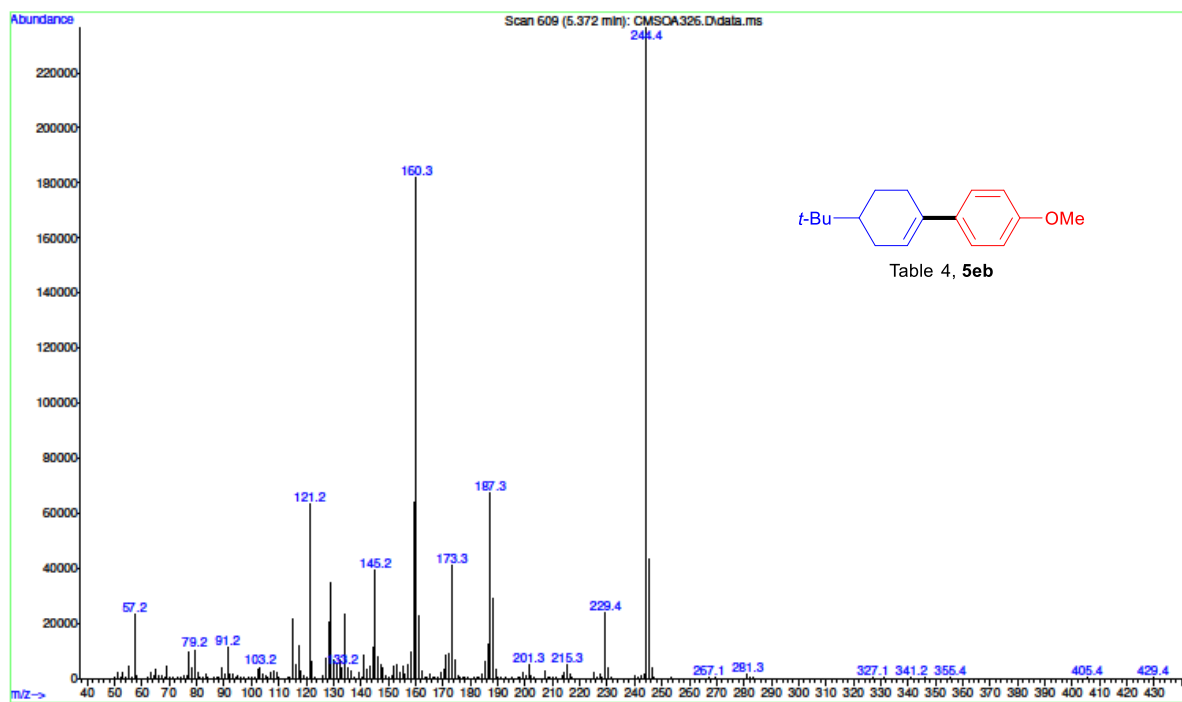




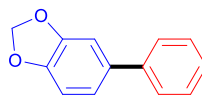




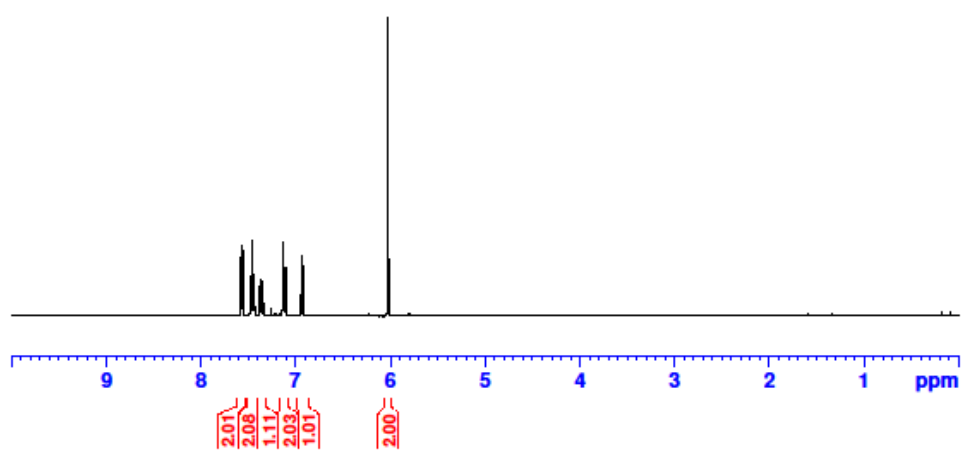




7.4710
7.4573
7.4540
7.4416
7.4377
7.4332
7.3822
7.3791
7.3759
7.3656
7.3607
7.3556
7.3455
7.3424
7.3393
7.2599
7.1372
7.1333
7.1263
7.1218
7.1064
7.1019
6.9413
6.9212
6.0215

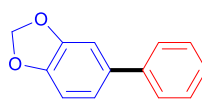


Scheme 3, 3gh

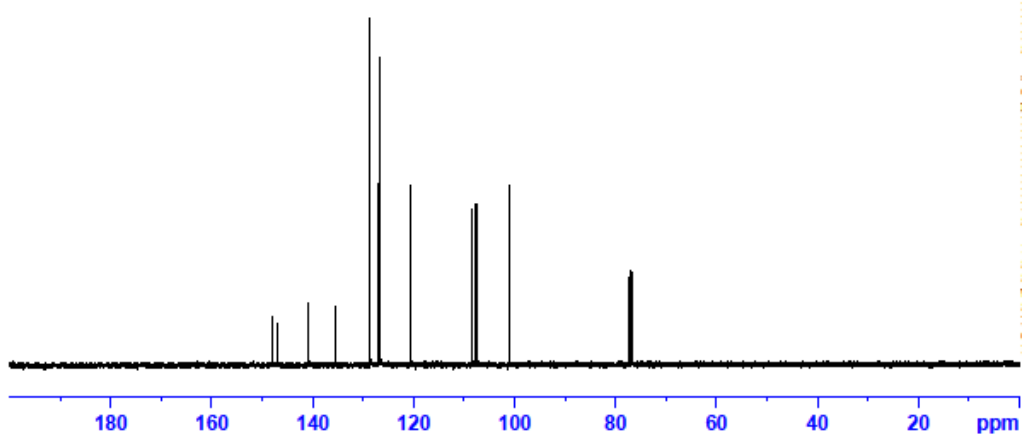


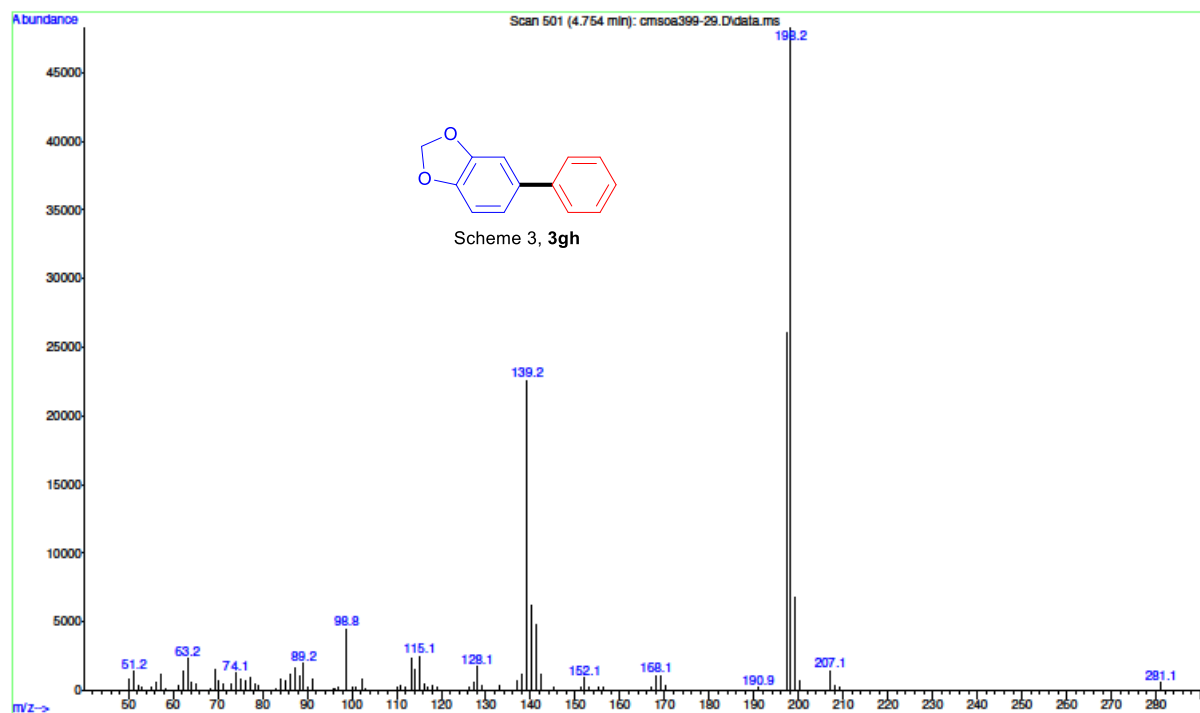
148.06
147.01
140.87
135.55
128.67
126.86
126.82
120.56
108.50
107.62
101.06

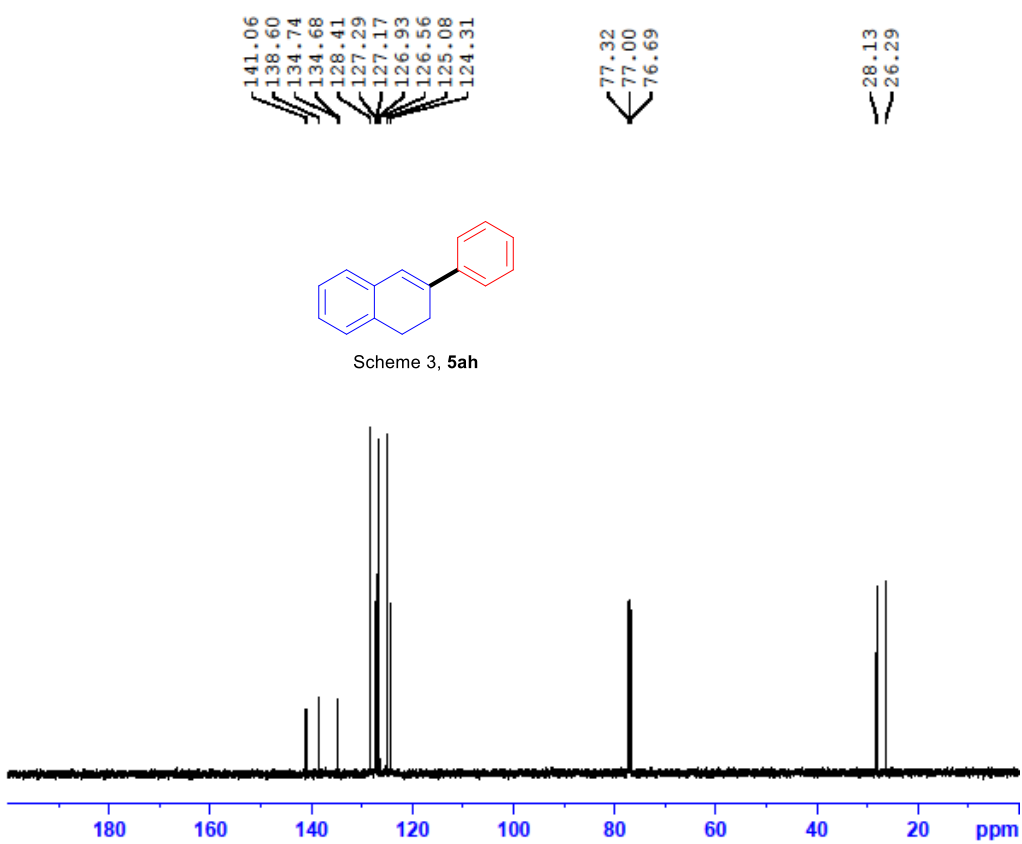
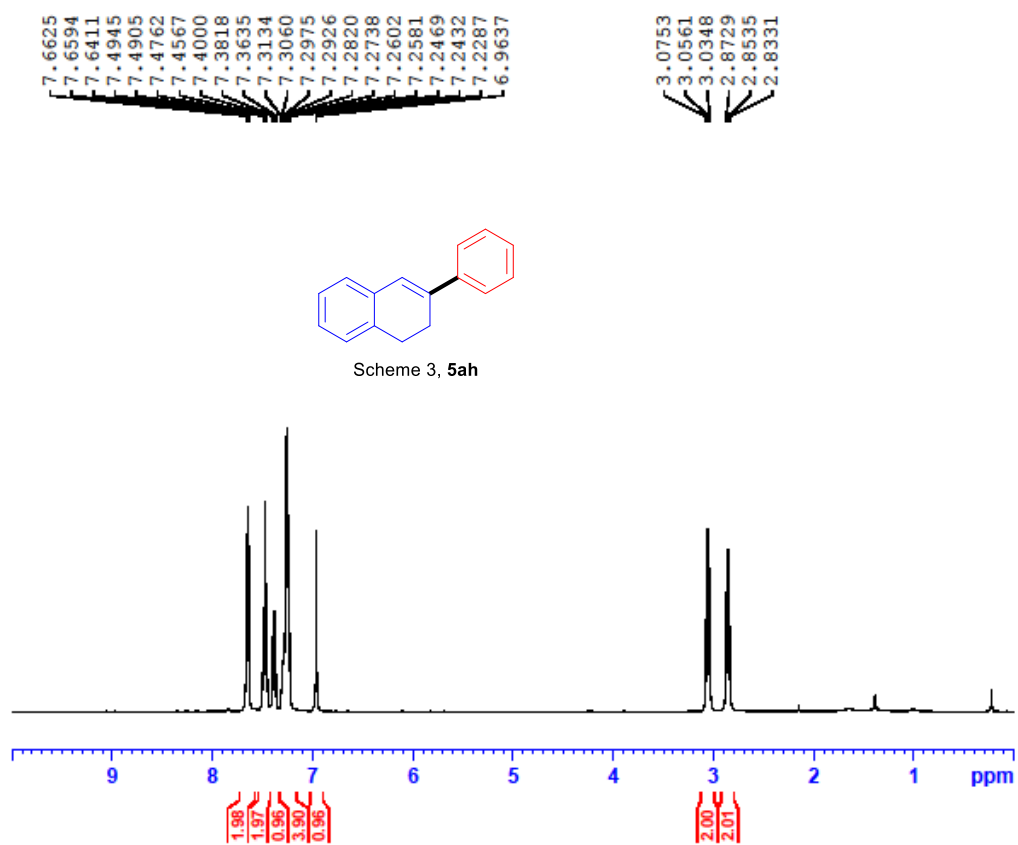
77.32
77.00
76.68

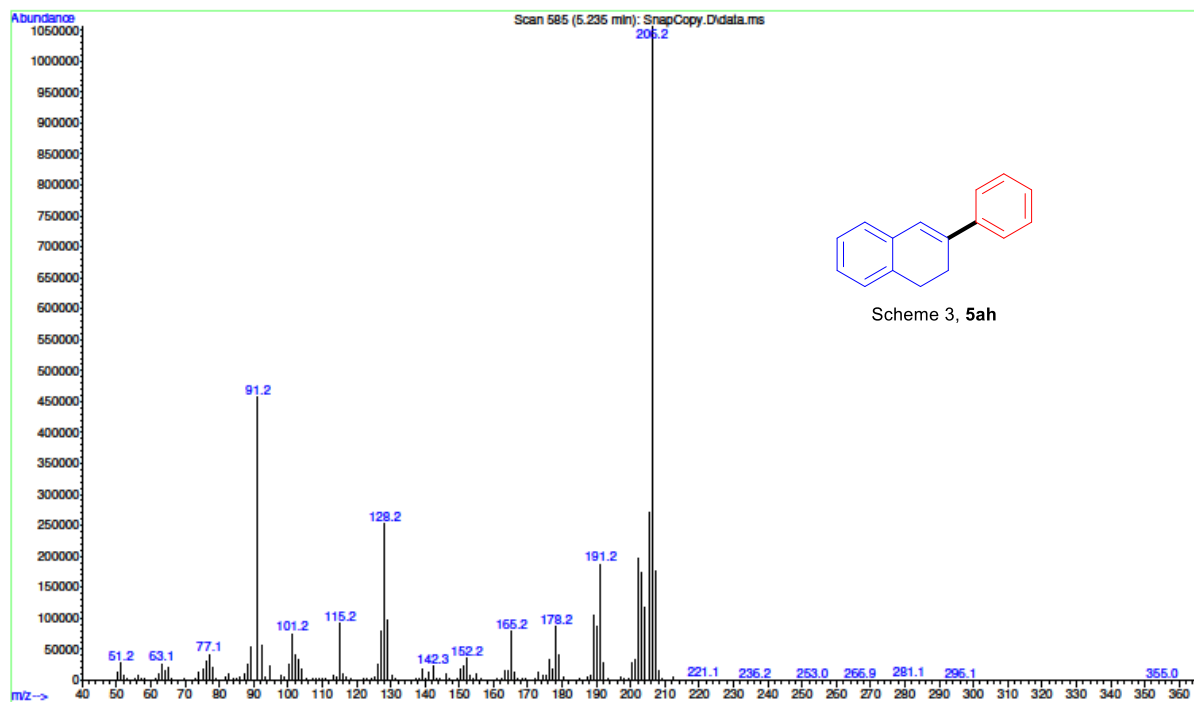


Scheme 3, 3gh









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