The Role of Hollow Magnetic Nanoparticles in Drug Delivery

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Figure S1. The structure of IBU-PEG-coated Fe_2O_3 (a) and IBU-PEG-coated Fe_3O_4 (b) ¹



Figure S2. Scheme of the synthetic procedure for the preparation of hollow magnetic mesoporous spheres ²







Figure S4. Fabrication of the hollow magnetic-MOF composite through the interfacial growth approach induced by Fe₃O₄

stabilized Pickering emulsion ³



Figure S5. Drug release of ibuprofen from the porous MHSNs showing slow release ⁴





Fe₃O₄-SiO₂-NHFA ⁵



Figure S7. Ibuprofen released from Fe₃O₄–SiO₂–NH₂ and Fe₃O₄–SiO₂–NHFA microspheres curves ⁵



Figure S8. The schematic preparation process of magnetic mesoporous silica microspheres MZHM-MSS-NH₂ and MZHM-

MSS-NHFA⁶



Figure S9. Cumulative ibuprofen release from MZHM-MSS-NH₂ and MZHM-MSS-NHFA in PBS solution at room temperature

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Figure S11. In vitro cumulative drug release of cefradine from the Fe₃O₄/PE₅/CdTe/PE₁ in release media of different pH

values 7



Figure S12. The cumulative vancomycin release ratio from the control HAp nanoparticles (S0) and HAp hollow microspheres

(S1), and the fabricated magnetic HAp hollow microspheres (S2–S4) in PBS at pH = 7.40^{8}



Figure S13. The schematic formation mechanism of the CM-mediated, microwave-assisted HMSPs ⁹



Figure S 14. Drug release profiles of DOX from HMSPs as a function of time at pH 4.0, 5.0 and 7.4 at 37 °C ⁹



Figure S 15. The Schematic illustration of the mechanism of formation of the poly(MAA/EGDMA)/Fe₃O₄ composite

microcapsules with hollow structure ¹⁰



Figure S16. Doxorubicin cumulative release from the hollow poly(MAA/EGDMA)/Fe₃O₄ composite microcapsules versus incubation time. Release profiles at different pH values (a) pH = 2, (b) pH = 4 and (c) pH = 7. 10



Figure S17. Illustration of the synthetic procedure of FeOOH/HMSS(DOX)-PEG ¹¹



Figure S18. Schematic procedure for preparation and folate conjugation of Fe₃O₄@SiO₂ hollow mesoporous spheres ¹²



Figure S19. DOX release from Fe₃O₄@SiO₂ and Fe₃O₄@SiO₂-FA spheres in PBS at 37 °C (inset, DOX loading capacity in

 $Fe_3O_4@SiO_2$ and $Fe_3O_4@SiO_2$ -FA spheres) ¹²



Figure S20. Schematic illustration for the fabrication of the PNIPAM/Fe₃O₄–ZnS hybrid hollow spheres ¹³



Figure S21. (a) The release behaviour of DOX-loaded PNIPAM/Fe₃O₄–ZnS hollow spheres in PBS at different temperatures

and (b) Schematic diagram for the DOX releasing process ¹³



Figure S22. The procedure for the synthesis of the 1D magnetic Fe₃O₄/P(MBA-co-MAA)/Ag and y-Fe₂O₃@mSiO₂ nanochains

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Figure S23. Drug released kinetic curve of γ -Fe₂O₃@mSiO₂ nanochains ¹⁴



Figure S24. The effects of initial DXR concentrations on the drug loading capacity and encapsulation efficiency of magnetite

and tumour dual-targeting hollow P(MBAAm-coMAA) microspheres. ¹⁵



Figure S25. CDDP release from (a) HMS, (b) HMS-CMCS spheres in PBS at 37°C ¹⁶



Figure S26. The synthesis procedure of hollow Fe₃O₄/SiO₂@PEG–PLA $^{\rm 17}$



Figure S27. CDDP release from (a) HMS; (b) HMS@PEG–PLA spheres in PBS at 37 °C ¹⁷



Figure S28. Schematic illustration of simultaneous surfactant exchange and cisplatin loading into a PHNP and

functionalization of this PHNP with Herceptin. ¹⁸



Figure S29. Synthesis of fluorescent magnetic nanoparticles conjugated with CPT and folic acid ¹⁹



Figure S30. (a) Cumulative CPT releases from Fe₃O₄@m-SiO₂–CD–FA–CPT, (b) in vitrocell viability of HeLa with asprepared Fe₃O₄@m-SiO₂–CD, Fe₃O₄@m-SiO₂–CD–CPT and Fe₃O₄@m-SiO₂–CD–FA–CPT nanoparticles. ¹⁹



Figure S31. Schematic illustration of the drug loading and release of the hollow dual-responsive microspheres ²⁰



Figure S32. The release profile of the TSCHMSs at 35 and 50°C²⁰

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