Electronic supplementary information

Rapid preparation of polydopamine coating as a multifunctional hair dye

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Experimental Section

Materials: Dopamine was purchased from Aladdin Chemical Co. Ltd, China. E. coli and S. aureus were purchased from Guangdong Microbial Culture Collection Center. Phosphate buffer solution (PBS) was purchased from Sangon Biotechnology Co. Ltd, China. Blonde hair was purchased from Amazon (Emosa, #60 platinum blonde). Commercial hair dye from Joybuy (Swarovski, 2.0 Pearl black). Shampoo from Joybuy (Qingyang, multi-effect water moisturizing maintenance). Other reagents, such as Hydrogen peroxide (30%, H_2O_2), copper sulfate pentahydrate (CuSO₄. 5H₂O), were purchased from Sinopharm Chemical Reagent Co., Ltd and used without further purification. Water used in all experiments was deionized and ultrafiltrated to 18.2 MΩ.

Hair dyeing with polydopamine: The hair sample were first washed with water thoroughly to eliminate impurities on the surface. Then, add the fresh prepared mixture contained with dopamine (5 mg/mL), Cu^{2+} (10 mM), and H_2O_2 (15 mM). The entire dyeing process can be completed about 5 min. Then, the dyed hair was washed and dried in air.

Durability test of PDA hair dye: The hair dyed with PDA was immersed in 5 vol% of shampoo in 40 mL of water in a 50 mL centrifugation tube. Then, the tube was vigorously shaken for 5 min on a vortex mixer. After washing, the hair was cleaned with water and dried in air. The PDA coating can endure over 30 washes without obvious decoloration.

Antibacterial test of PDA hair dye: First, the crude or PDA-dyed hair was placed into a 24-well culture plate. A 200 μ L of E. coli or S. aureus suspension in PBS (1×10⁷ cfu/mL) was used to cover the substrate. Add 1800 μ L of PBS to dilute the bacterial solution after 2.5 h at 37 °C for agar plate incubation. Then, the bacterial suspension and each sample were transferred to a new tube for 5 min ultrasonic treatment to detach the adhered bacteria. Finally, the bacterial solution was diluted to 1000 times with PBS solution and 100 μ L of the bacterial solution was taken to measure the viability of bacterial by using agar plates.

Characterization: Morphology of the samples was observed by scanning electron microscope (SEM, QUANTA250, USA). X-ray photoelectron spectra were collected by a spectrometer (XPS, Perkin Elmer, USA) with Al Kα excitation radiation (1486.6 eV). UV-vis absorption was tested with an ultraviolet spectro-photometer (UV 2450, Shimadzu, Japan). MTT assays were performed on a BioTek Epoch2 microplate reader. Inductively coupled plasma mass spectroscopy (ICP-MS) were conducted on an ICPOES730 instrument (Agilent). Gel permeation chromatography (GPC) was carried out with a Shimadzu LC-10AD (column, TSKgel GMPWXL; solvent, water; flow rate 0.6 mL/min; 35 °C). The digital images were taken on a shadow-less plate by a camera (5D Mark II, Canon) in a professional studio.



Fig. S1 GPC analysis of DA, DA+ Cu^{2+} , DA+ H_2O_2 , DA+ Cu^{2+} + H_2O_2 , respectively.



Fig. S2 Reaction kinetics of PDA monitored by UV/Vis spectrophotometer at 465 nm.



Fig. S3 (a) SEM micrographs of uncoated hair and PDA-coated hair. (b) SEM-EDS mapping of PDA-coated hair.



Fig. S4 (a) UV/Vis spectra and (b) absorbances (at 465 nm) of solutions containing 10 mM CuSO4, 15 mM H2O2 and various DA concentrations.



Fig. S5 (a) UV/Vis spectra and (b) absorbances (at 465 nm) of solutions containing 1 mg/mL DA, 15 mM H_2O_2 and various CuSO₄ concentrations.



Fig. S6 (a) UV/Vis spectra and (b) absorbances (at 465 nm) of solutions containing 1

mg/mL DA, 10 mM CuSO₄ and various $\mathrm{H_2O_2}$ concentrations.



Fig. S7 The diagram of changes of hair color responds to dyeing times.