

Supplementary Information for:

Improving the anticancer activity of platinum(IV) prodrugs using a dual-targeting strategy with dichloroacetate axial ligand

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1. NMR spectra of complexes 1a-2b

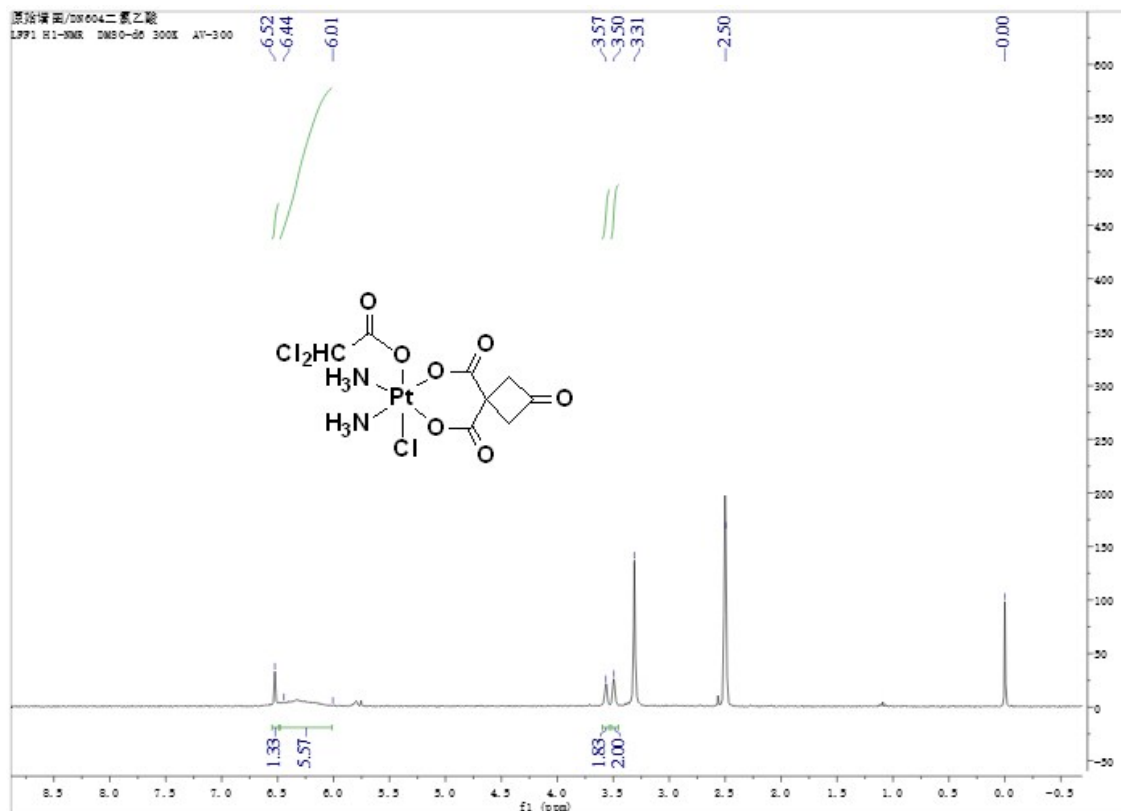


Fig. S1. ¹H NMR (DMSO) spectrum of complex 1a

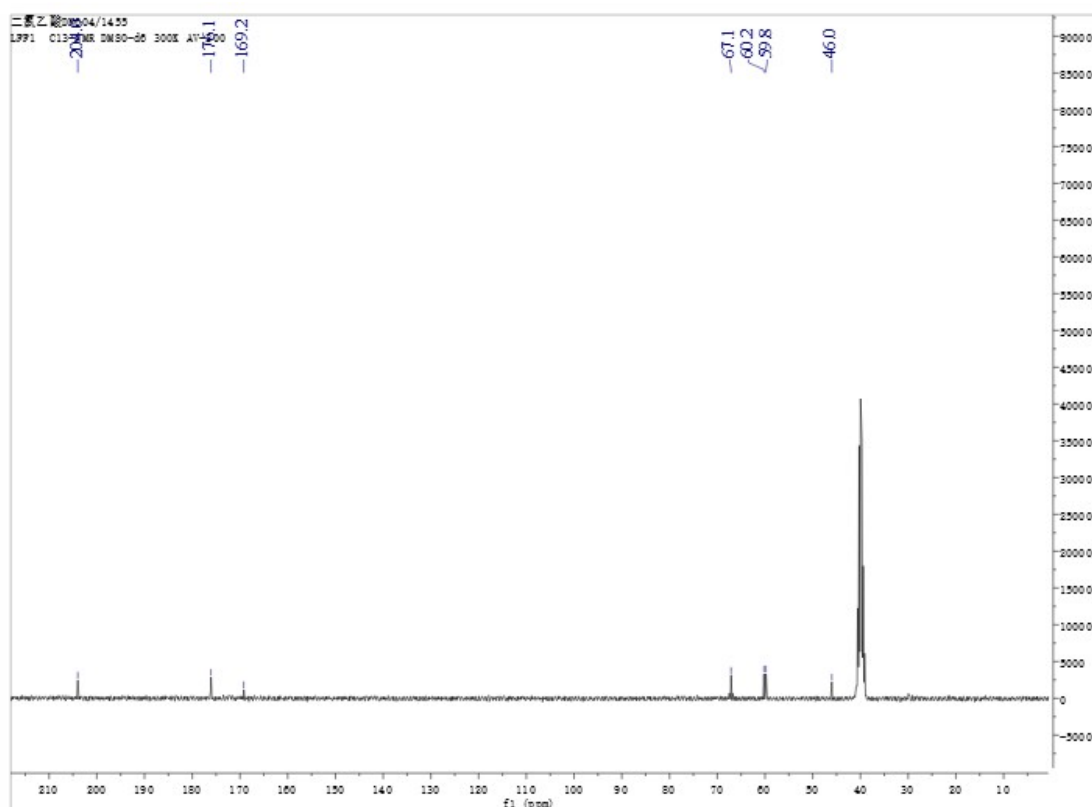
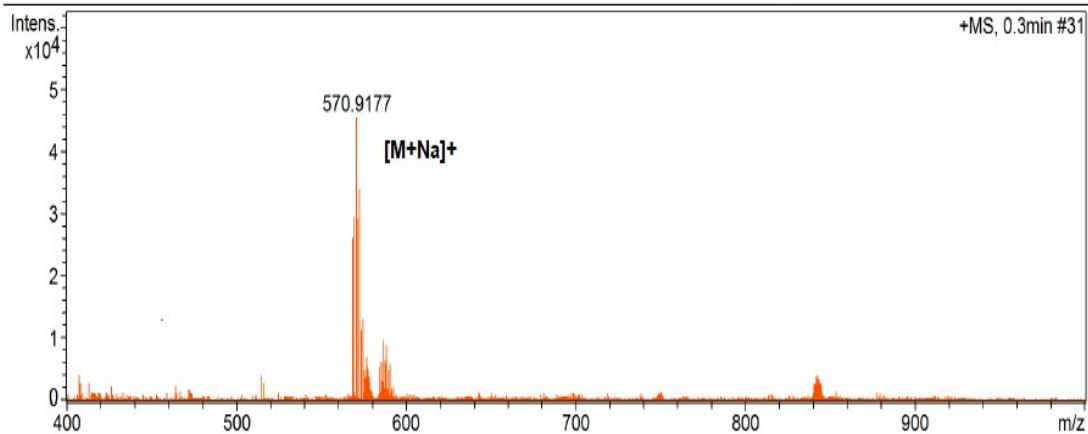


Fig. S2. ¹³C NMR (DMSO) spectrum of complex 1a

Acquisition Parameter

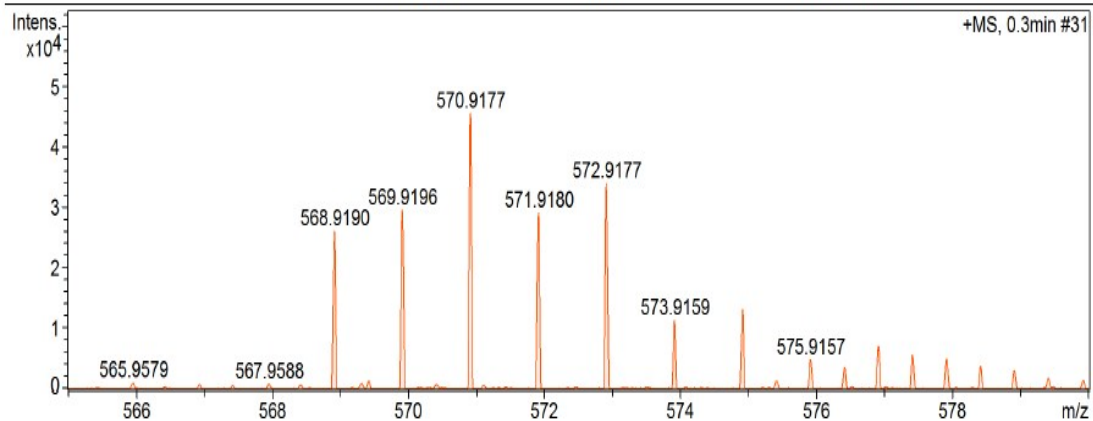
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Scan End	1200 m/z	Set Charging Voltage	2000 V	Set Divert Valve	Waste
		Set Corona	0 nA	Set APCI Heater	0 °C



Meas. m/z	Ion Formula	Sum Formula	m/z	err [ppm]
569.9196	C ₈ H ₁₁ Cl ₃ N ₂ NaO ₇ Pt	C ₈ H ₁₁ Cl ₃ N ₂ O ₇ Pt	569.9172	-4.0

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
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Scan End	1200 m/z	Set Charging Voltage	2000 V	Set Divert Valve	Waste
		Set Corona	0 nA	Set APCI Heater	0 °C



Meas. m/z	Ion Formula	Sum Formula	m/z	err [ppm]
569.9196	C ₈ H ₁₁ Cl ₃ N ₂ NaO ₇ Pt	C ₈ H ₁₁ Cl ₃ N ₂ O ₇ Pt	569.9172	-4.0

Fig. S3. HR-MS (ESI) spectrum of complex 1a

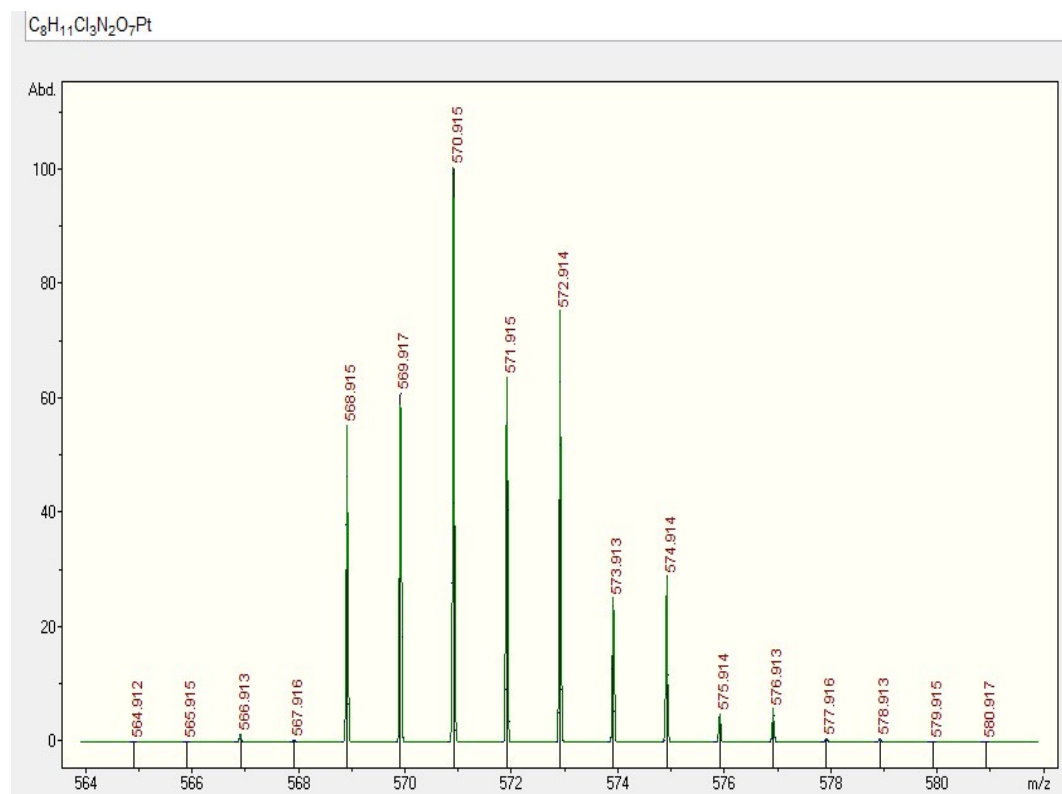


Fig. S4. The simulation of HR-MS (ESI) spectrum of complex 1a

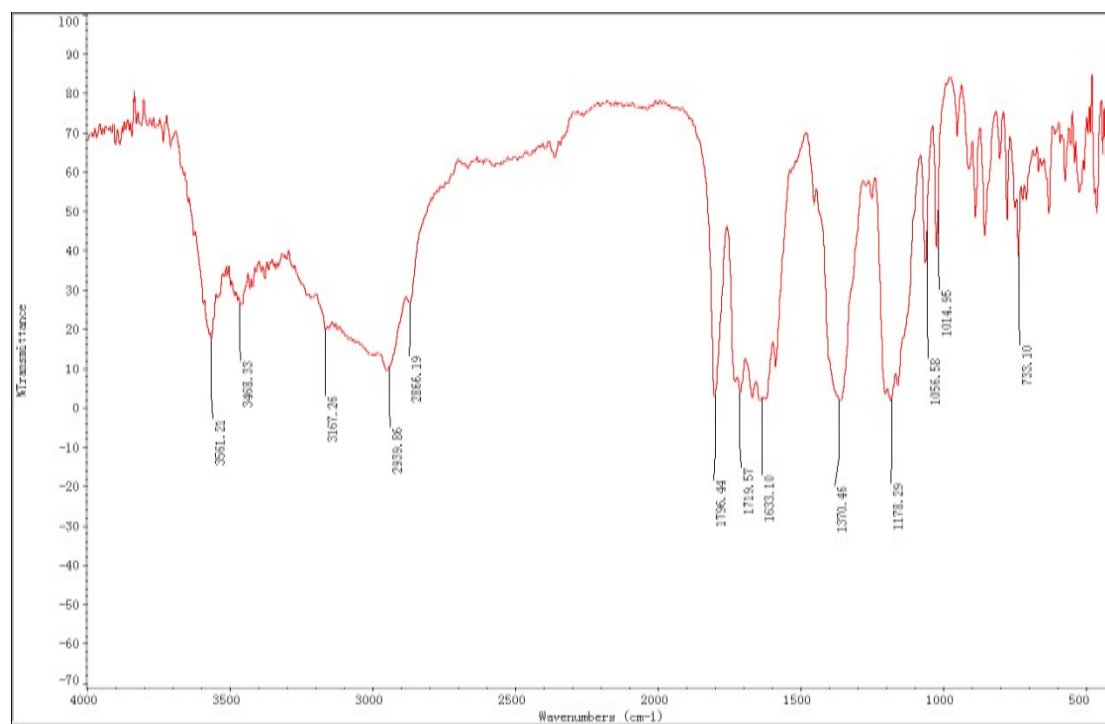


Fig. S5. IR spectrum of complex 1a

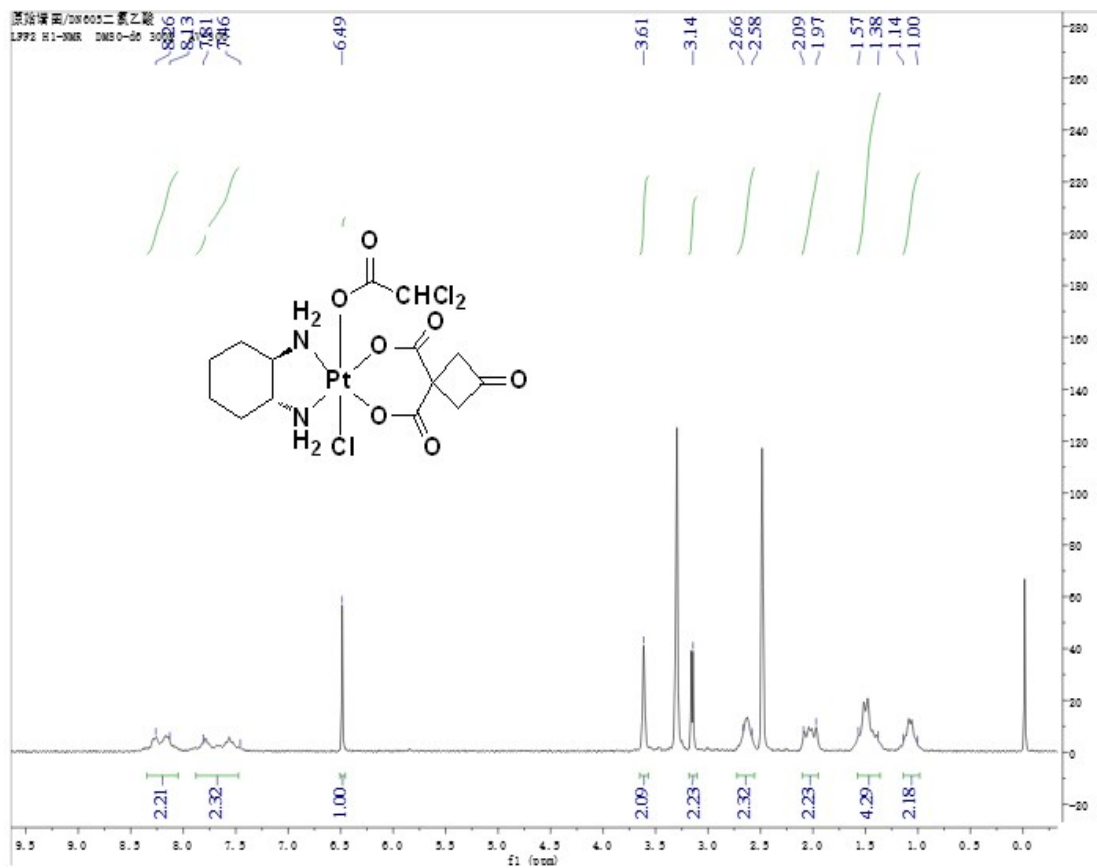


Fig. S6. ¹H NMR (DMSO) spectrum of complex 1b

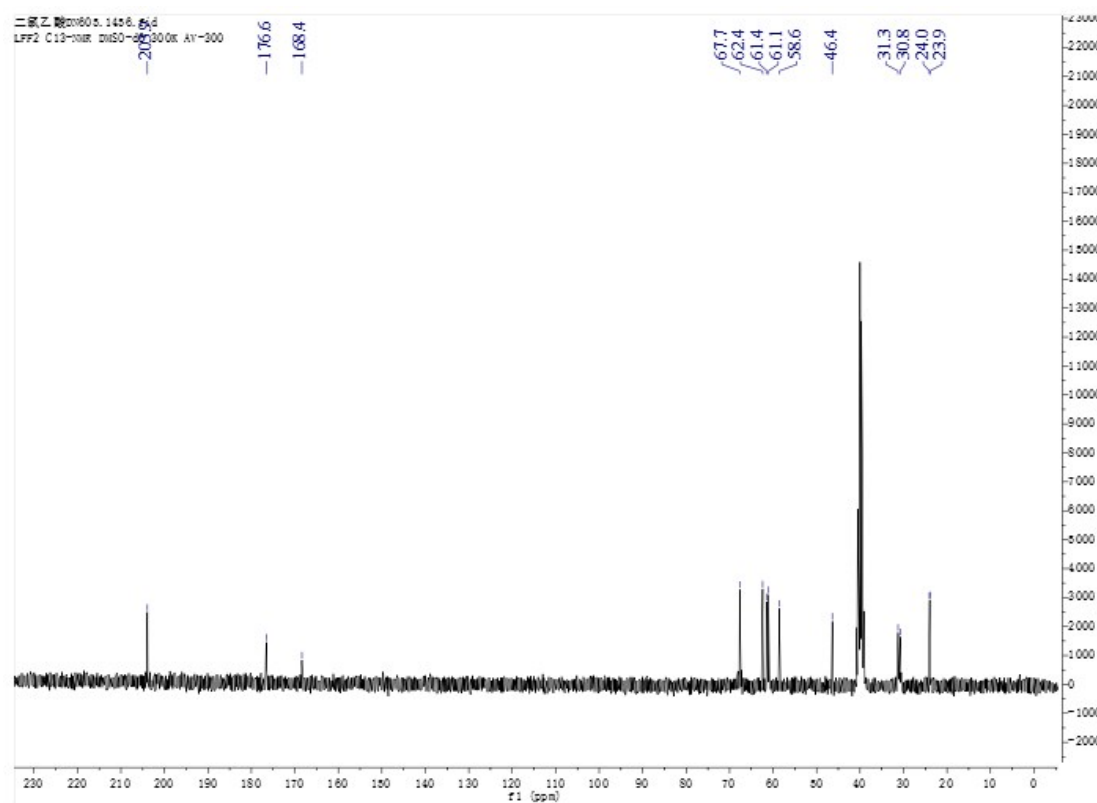
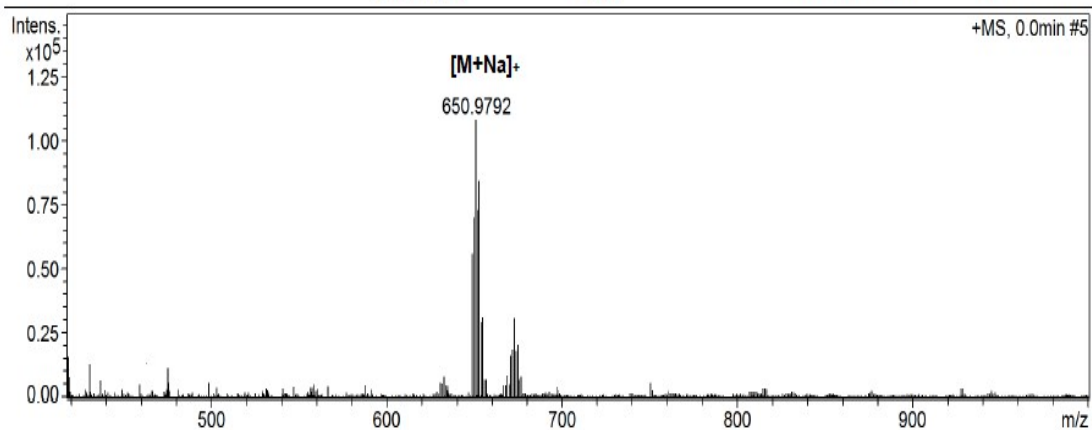


Fig. S7. ¹³C NMR (DMSO) spectrum of complex 1b

Acquisition Parameter

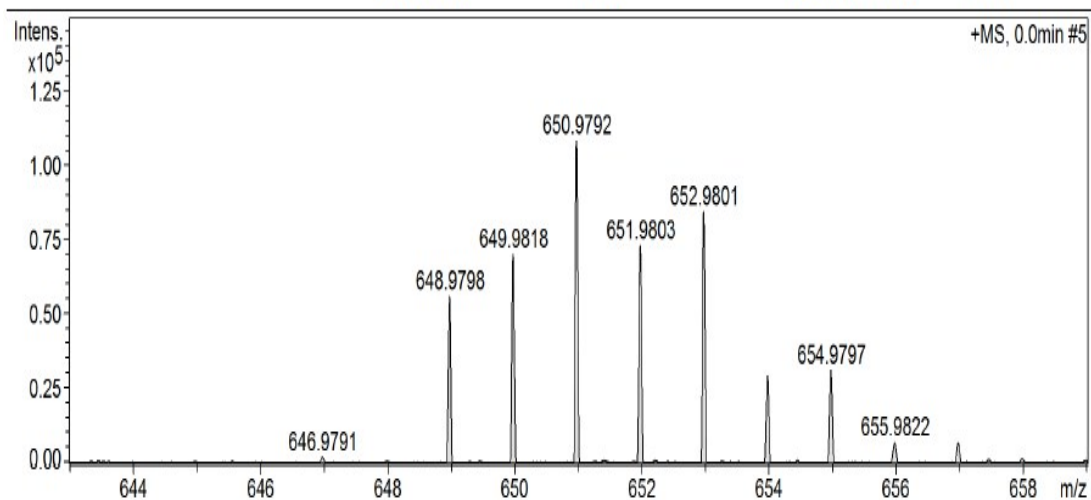
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Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	3.0 l/min
Scan End	1200 m/z	Set Charging Voltage	2000 V	Set Divert Valve	Waste
		Set Corona	0 nA	Set APCI Heater	0 °C



Meas. m/z	Ion Formula	Sum Formula	m/z	err [ppm]
649.9818	C ₁₄ H ₁₉ Cl ₃ N ₂ NaO ₇ Pt	C ₁₄ H ₁₉ Cl ₃ N ₂ O ₇ Pt	649.9798	-2.9

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
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Scan End	1200 m/z	Set Charging Voltage	2000 V	Set Divert Valve	Waste
		Set Corona	0 nA	Set APCI Heater	0 °C



Meas. m/z	Ion Formula	Sum Formula	m/z	err [ppm]
649.9818	C ₁₄ H ₁₉ Cl ₃ N ₂ NaO ₇ Pt	C ₁₄ H ₁₉ Cl ₃ N ₂ O ₇ Pt	649.9798	-2.9

Fig. S8. HR-MS (ESI) spectrum of complex 1b

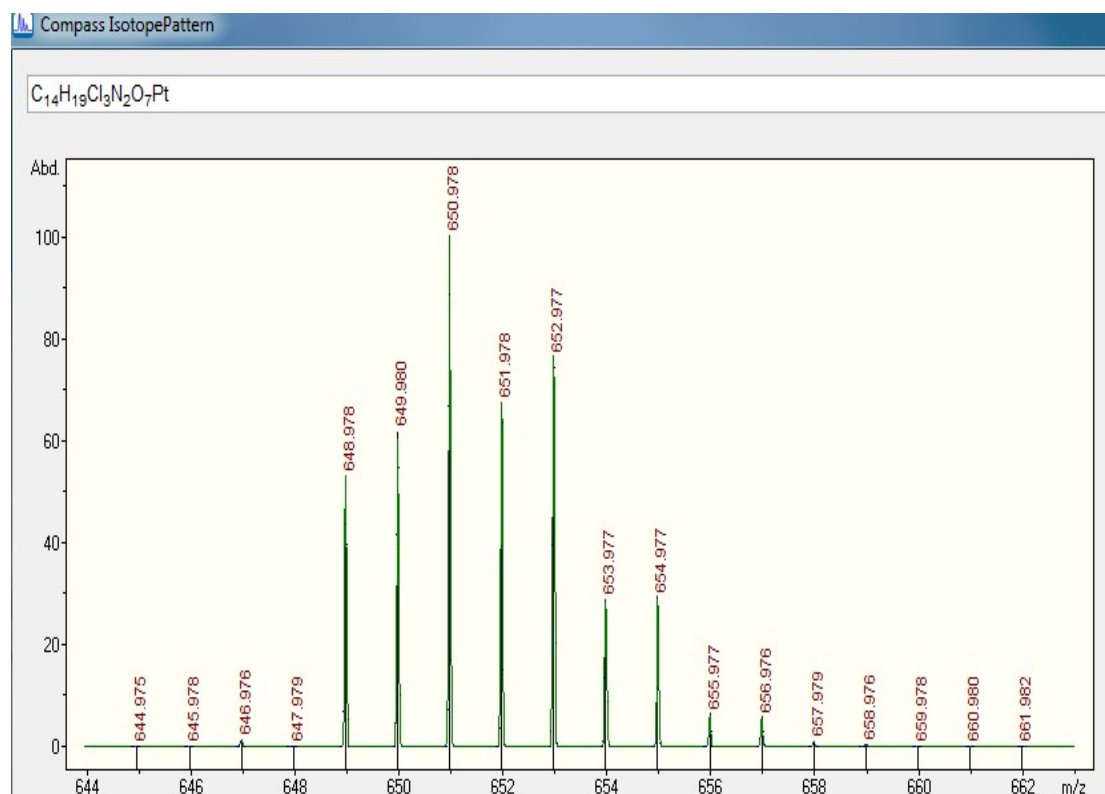


Fig. S9. The simulation of HR-MS (ESI) spectrum of complex 1b

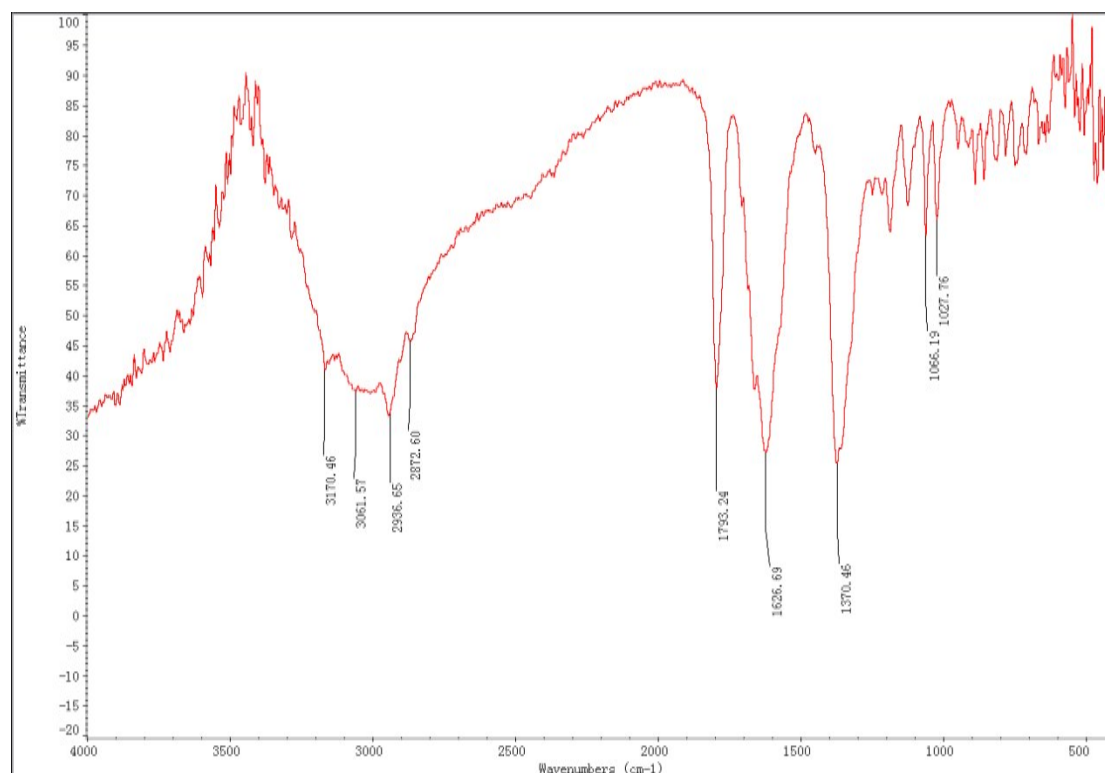


Fig. S10. IR spectrum of complex 1b

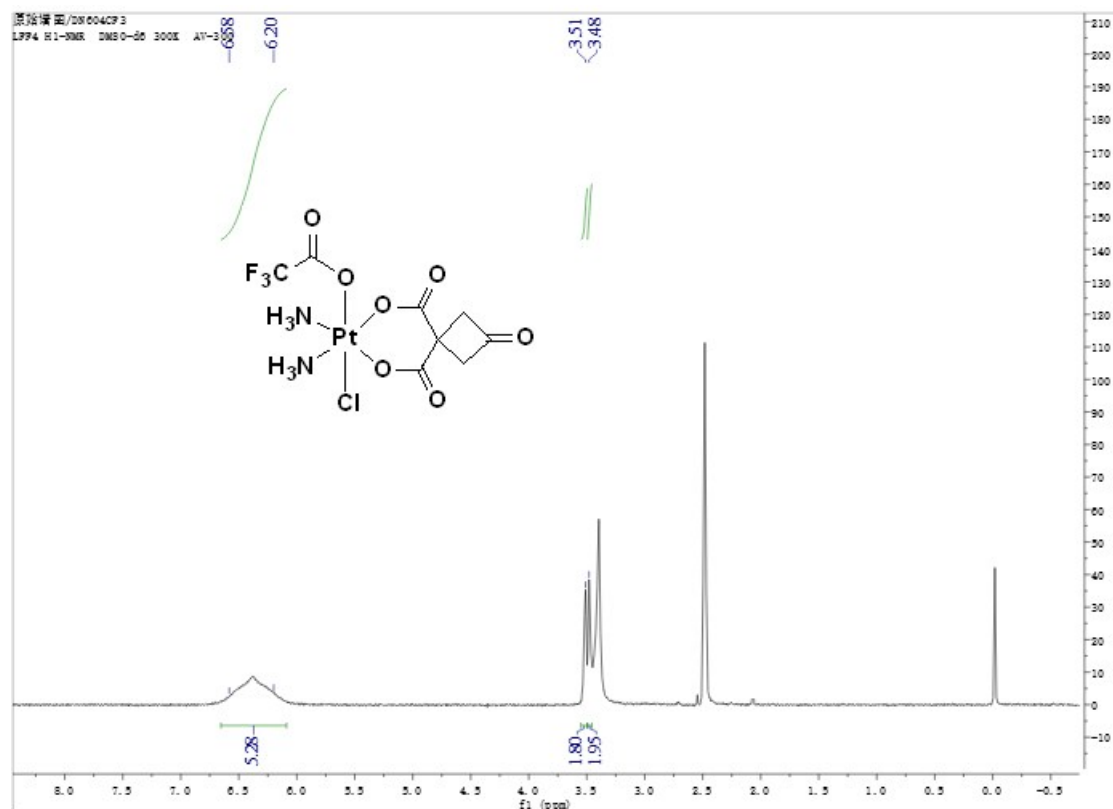


Fig. S11. ^1H NMR (DMSO) spectrum of complex 2a

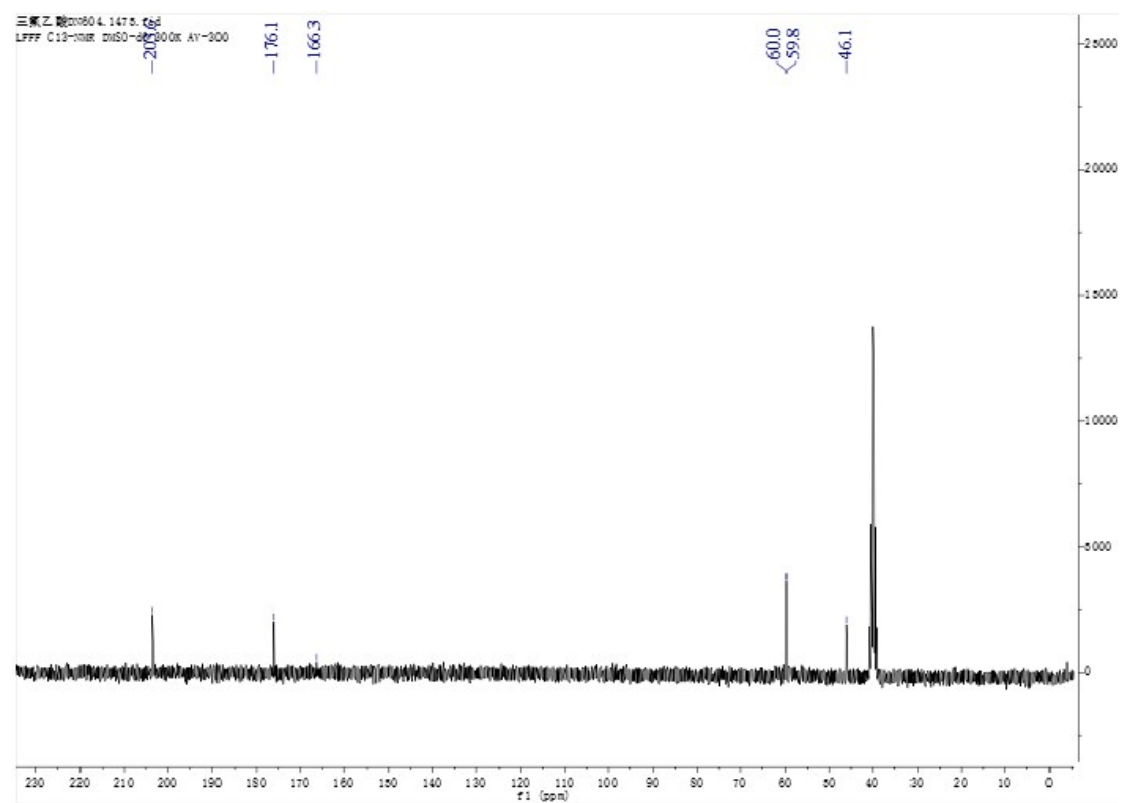
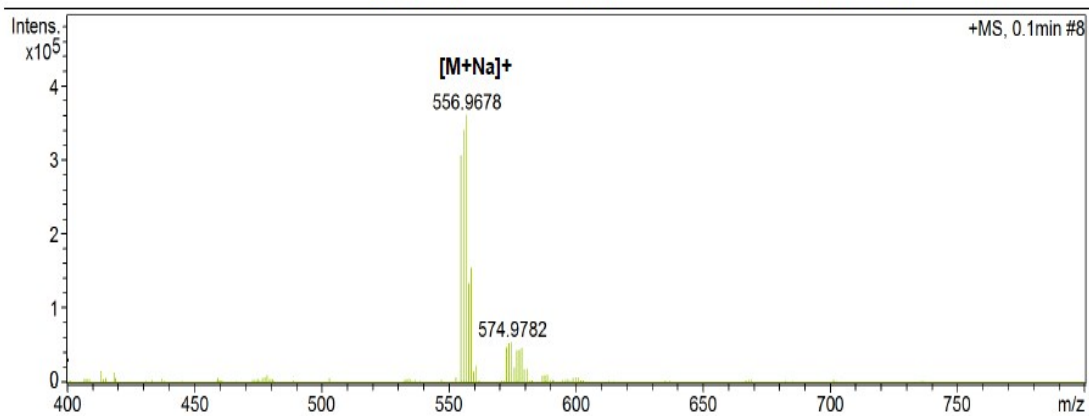


Fig. S12. ^{13}C NMR (DMSO) spectrum of complex 2a

Acquisition Parameter

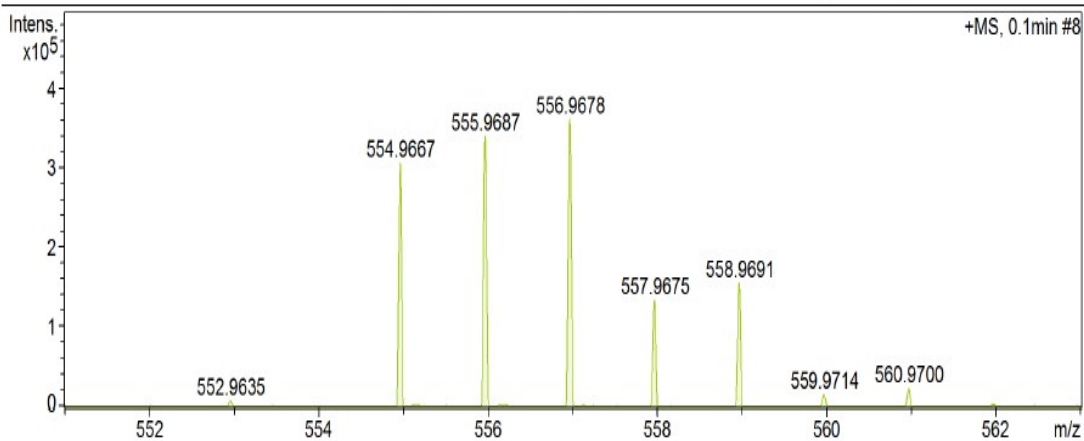
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Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	3.0 l/min
Scan End	1200 m/z	Set Charging Voltage	2000 V	Set Divert Valve	Waste
		Set Corona	0 nA	Set APCI Heater	0 °C



Meas. m/z	Ion Formula	Sum Formula	m/z	err [ppm]
555.9687	C ₈ H ₁₀ ClF ₃ N ₂ NaO ₇ Pt	C ₈ H ₁₀ ClF ₃ N ₂ O ₇ Pt	555.9669	-3.2

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	3.0 l/min
Scan End	1200 m/z	Set Charging Voltage	2000 V	Set Divert Valve	Waste
		Set Corona	0 nA	Set APCI Heater	0 °C



Meas. m/z	Ion Formula	Sum Formula	m/z	err [ppm]
555.9687	C ₈ H ₁₀ ClF ₃ N ₂ NaO ₇ Pt	C ₈ H ₁₀ ClF ₃ N ₂ O ₇ Pt	555.9669	-3.2

Fig. S13. HR-MS (ESI) spectrum of complex 2a

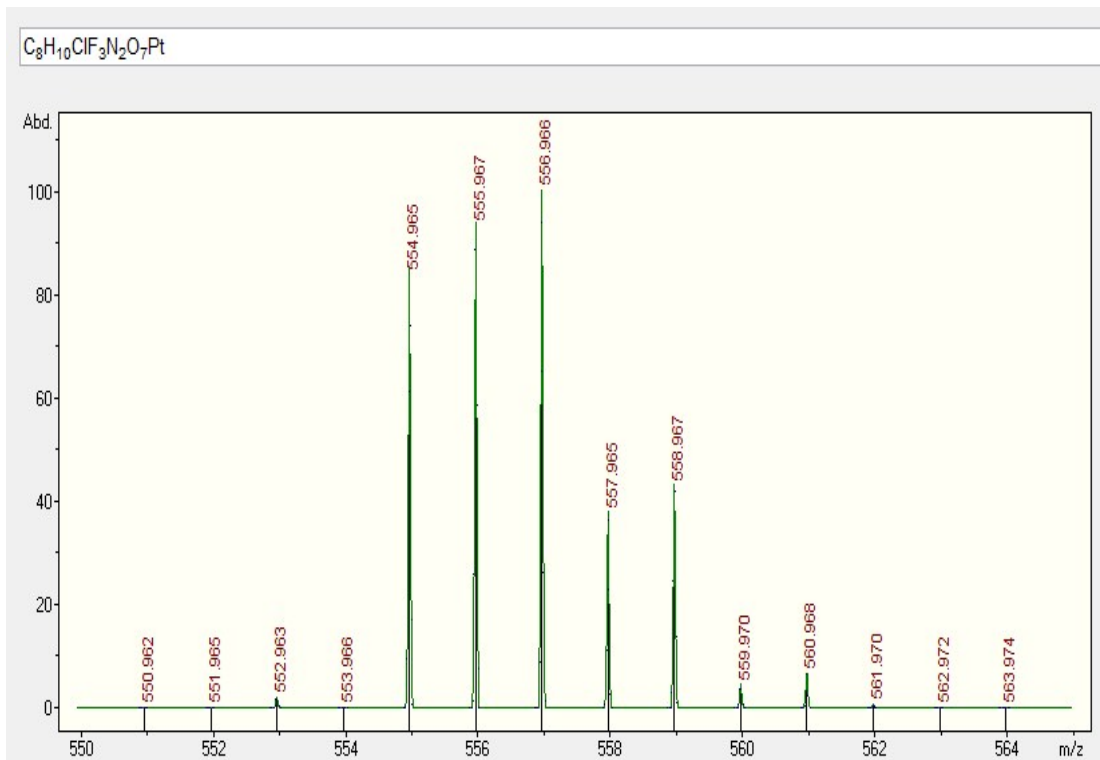


Fig. S14. The simulation of HR-MS (ESI) spectrum of complex 2a

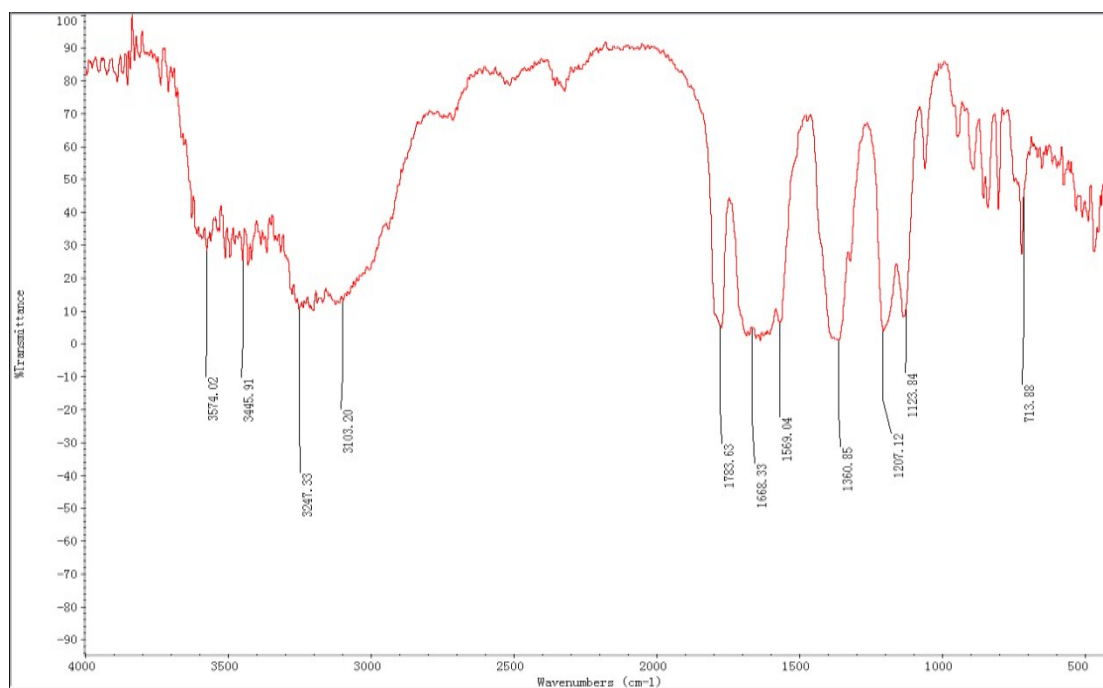


Fig. S15. IR spectrum of complex 2a

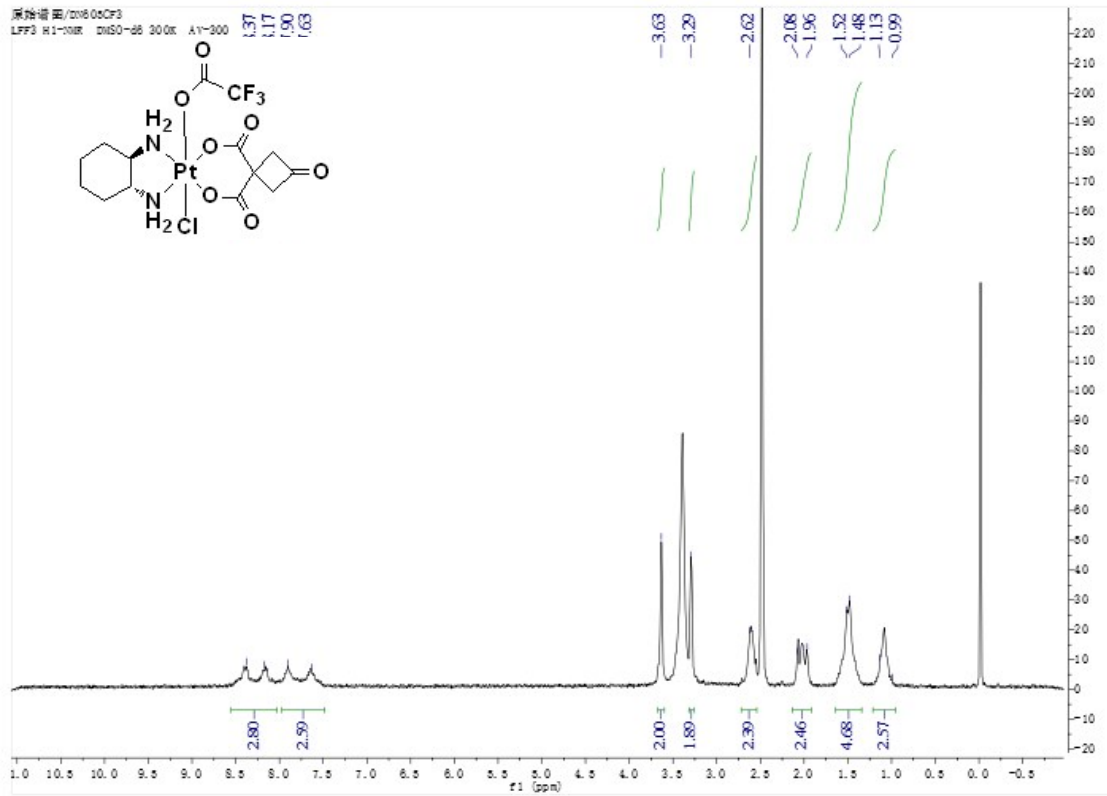


Fig. S16. ^1H NMR (DMSO) spectrum of complex 2b

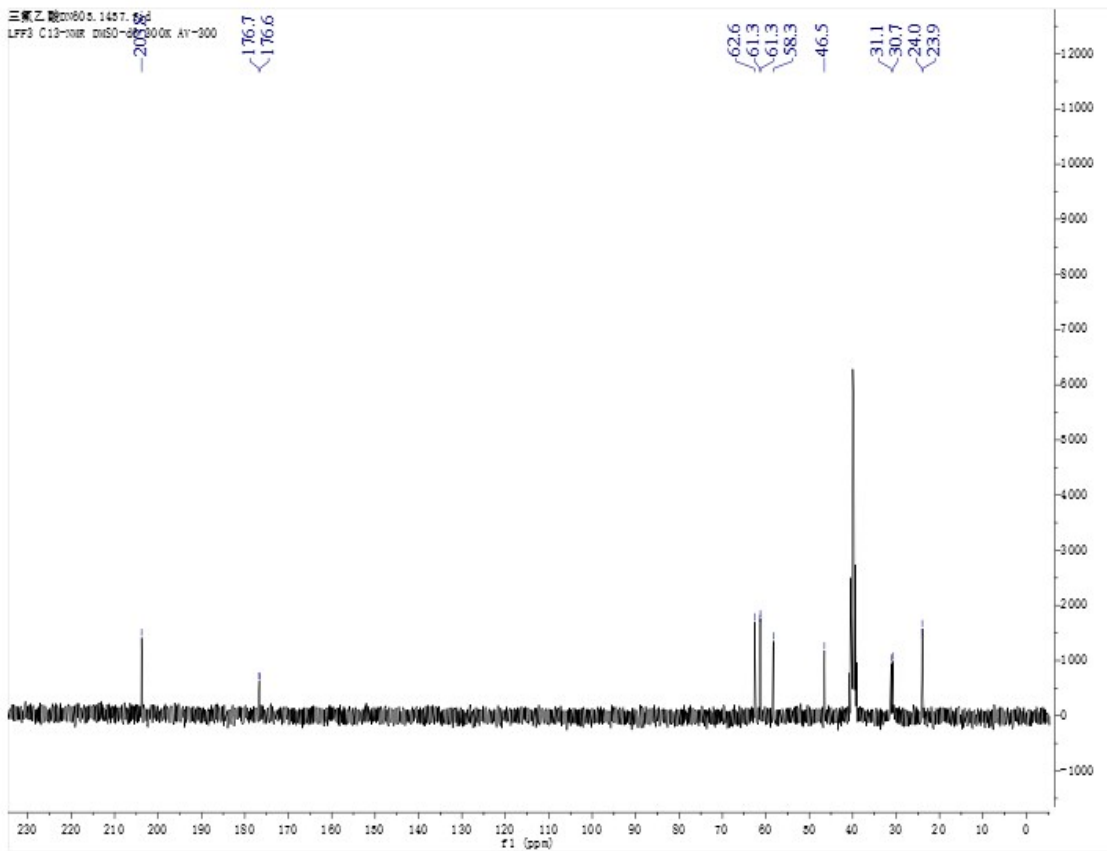
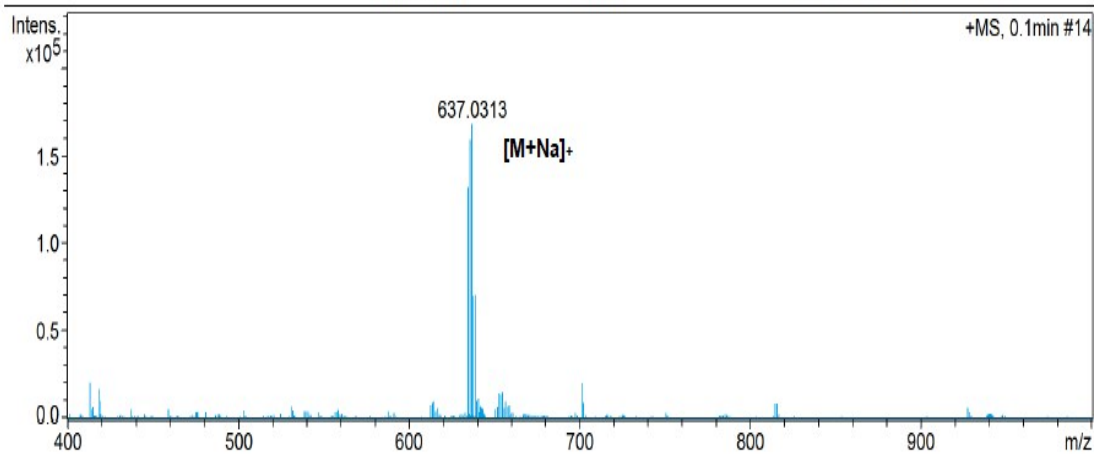


Fig. S17. ^{13}C NMR (DMSO) spectrum of complex 2b

Acquisition Parameter

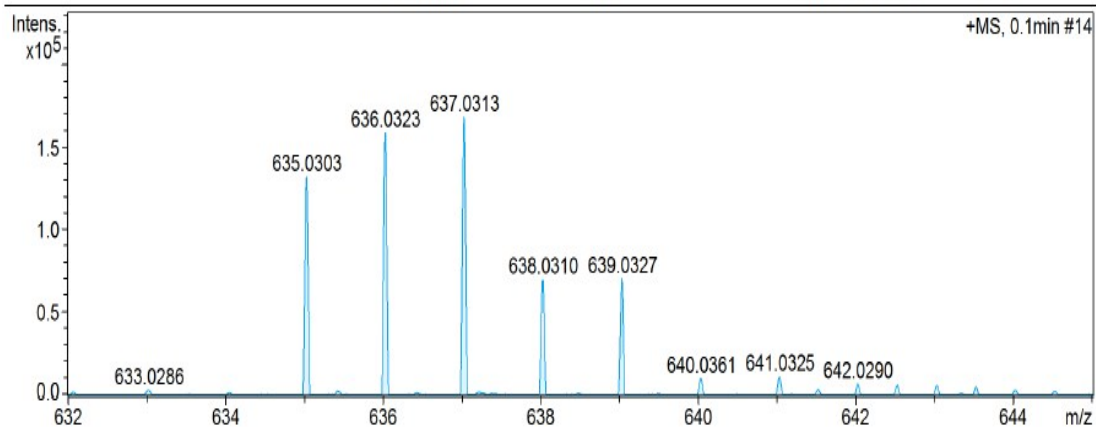
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Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	3.0 l/min
Scan End	1200 m/z	Set Charging Voltage	2000 V	Set Divert Valve	Waste
		Set Corona	0 nA	Set APCI Heater	0 °C



Meas. m/z	Ion Formula	Sum Formula	m/z	err [ppm]
636.0323	C ₁₄ H ₁₈ ClF ₃ N ₂ NaO ₇ Pt	C ₁₄ H ₁₈ ClF ₃ N ₂ O ₇ Pt	636.0295	-4.2

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	3.0 l/min
Scan End	1200 m/z	Set Charging Voltage	2000 V	Set Divert Valve	Waste
		Set Corona	0 nA	Set APCI Heater	0 °C



Meas. m/z	Ion Formula	Sum Formula	m/z	err [ppm]
636.0323	C ₁₄ H ₁₈ ClF ₃ N ₂ NaO ₇ Pt	C ₁₄ H ₁₈ ClF ₃ N ₂ O ₇ Pt	636.0295	-4.2

Fig. S18. HR-MS (ESI) spectrum of complex 2b

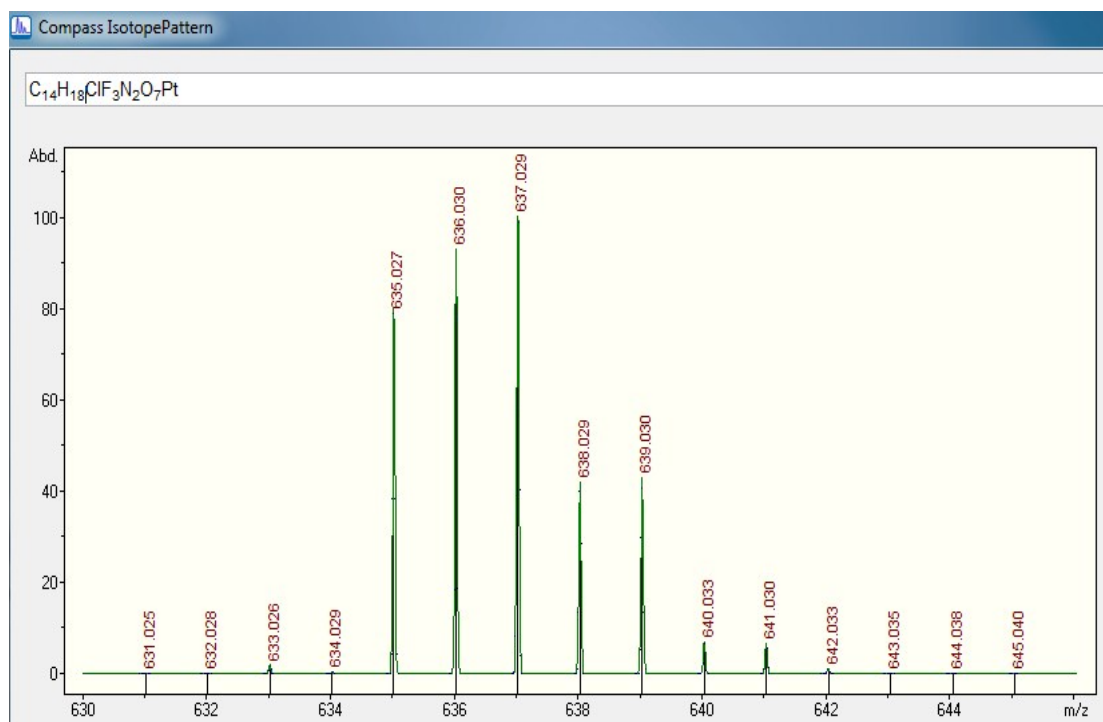


Fig. S19. The simulation of HR-MS (ESI) spectrum of complex **2b**

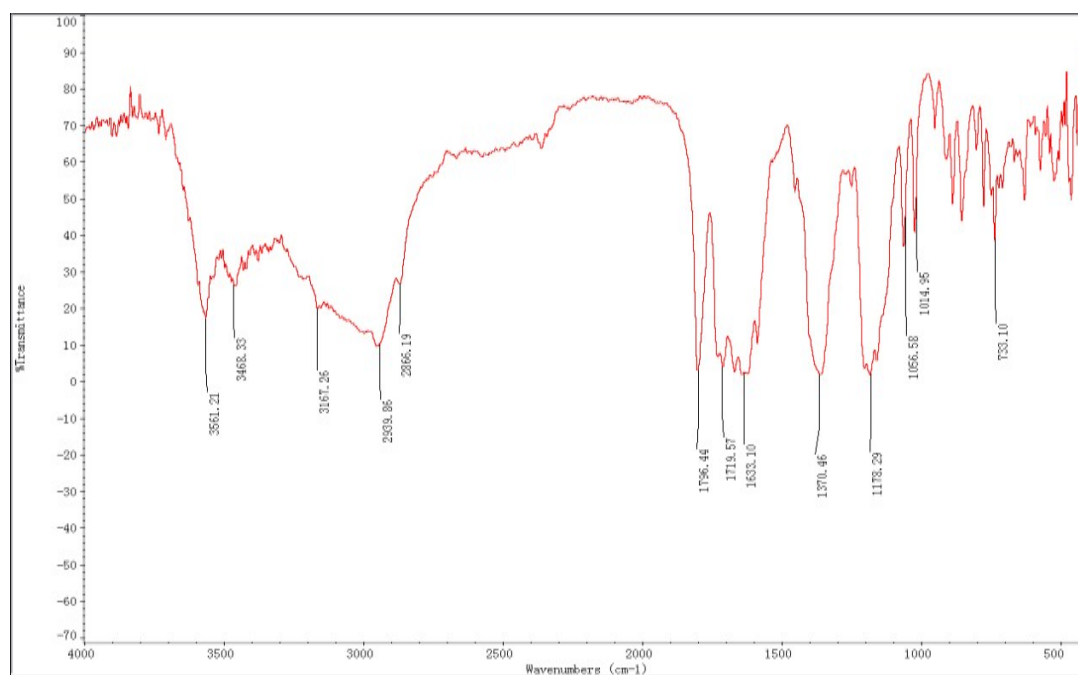


Fig. S20. IR spectrum of complex **2b**

2. The stability of complexes **1a-2b** in MeOH/PBS buffer

(a) ¹H-NMR (400 MHz) spectra of complex **1a** in deuterated aqueous, phosphate buffered solution

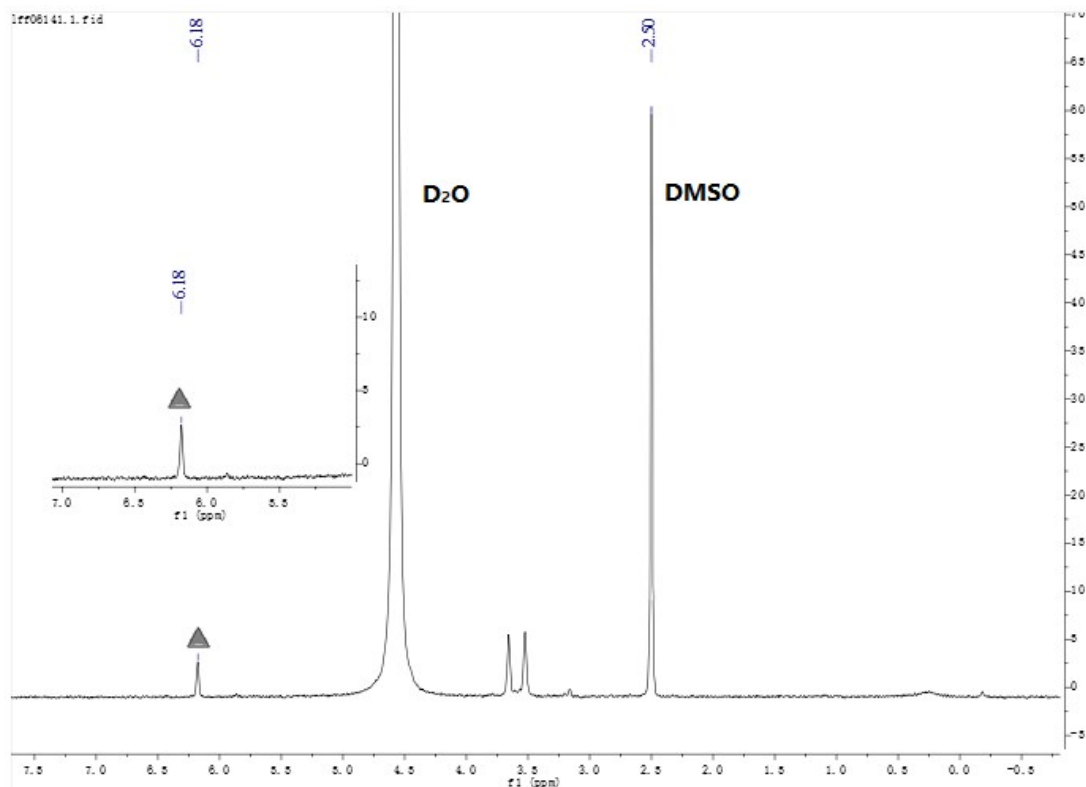


Fig.21. ^1H NMR spectra of complex **1a** treated with PBS (7.87 mM, pH 7.4; at 37°C) at 0 min (DMSO- d_6 /D $_2$ O, 10/90, v/v). ▲ indicates the peak of coordinated DCA in complex **1a**.

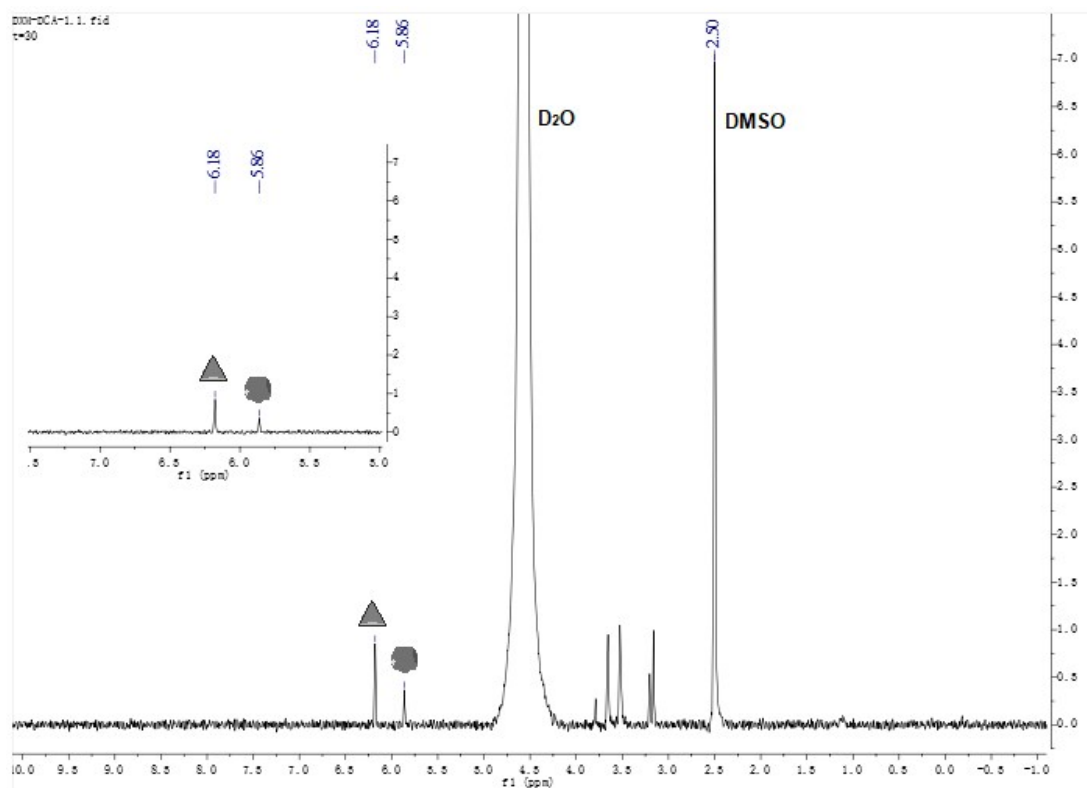


Fig.22. ^1H NMR spectra of complex **1a** treated with PBS (7.87 mM, pH 7.4; at 37°C) at 50 mins (DMSO- d_6 /D $_2$ O, 10/90, v/v). ▲ indicates the peak of coordinated DCA in complex **1a**. ● indicates the peak of free DCA.

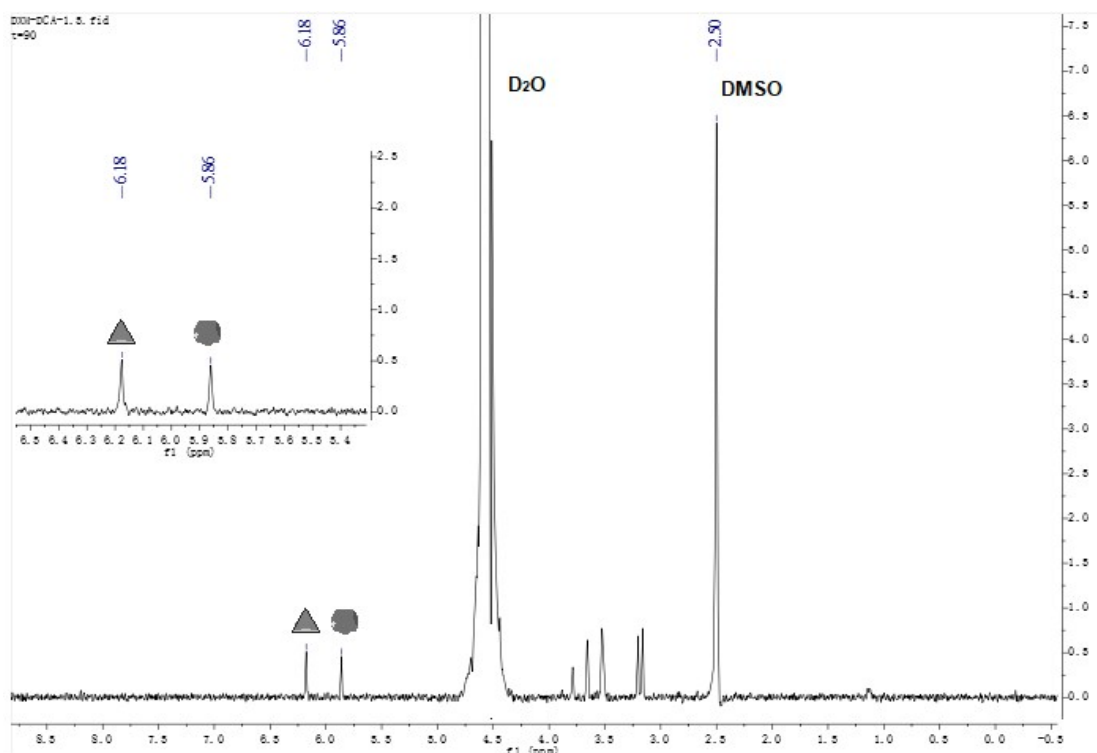


Fig.23. ¹H NMR spectra of complex **1a** treated with PBS (7.87 mM, pH 7.4; at 37°C) at 100 mins (DMSO-*d*₆/D₂O, 10/90, v/v). ▲ indicates the peak of coordinated DCA in complex **1a**. ● indicates the peak of free DCA.

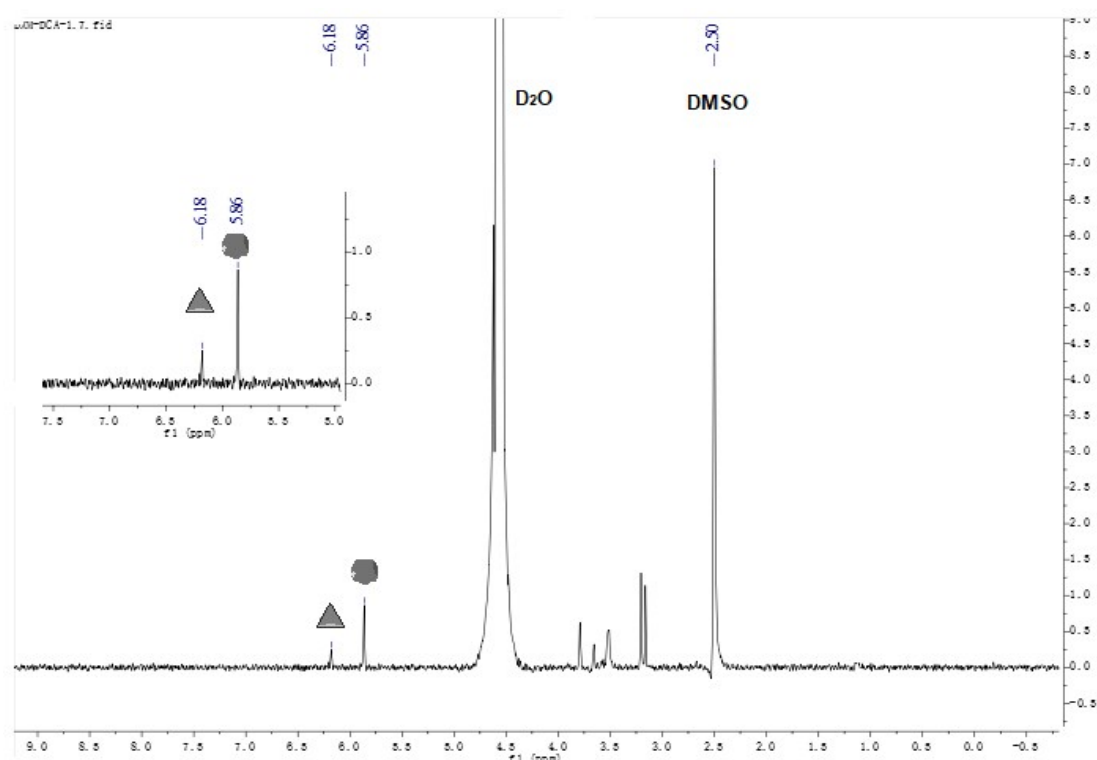


Fig.24. ¹H NMR spectra of complex **1a** treated with PBS (7.87 mM, pH 7.4; at 37°C) at 150 mins (DMSO-*d*₆/D₂O, 10/90, v/v). ▲ indicates the peak of coordinated DCA in complex **1a**. ● indicates the peak of free DCA.

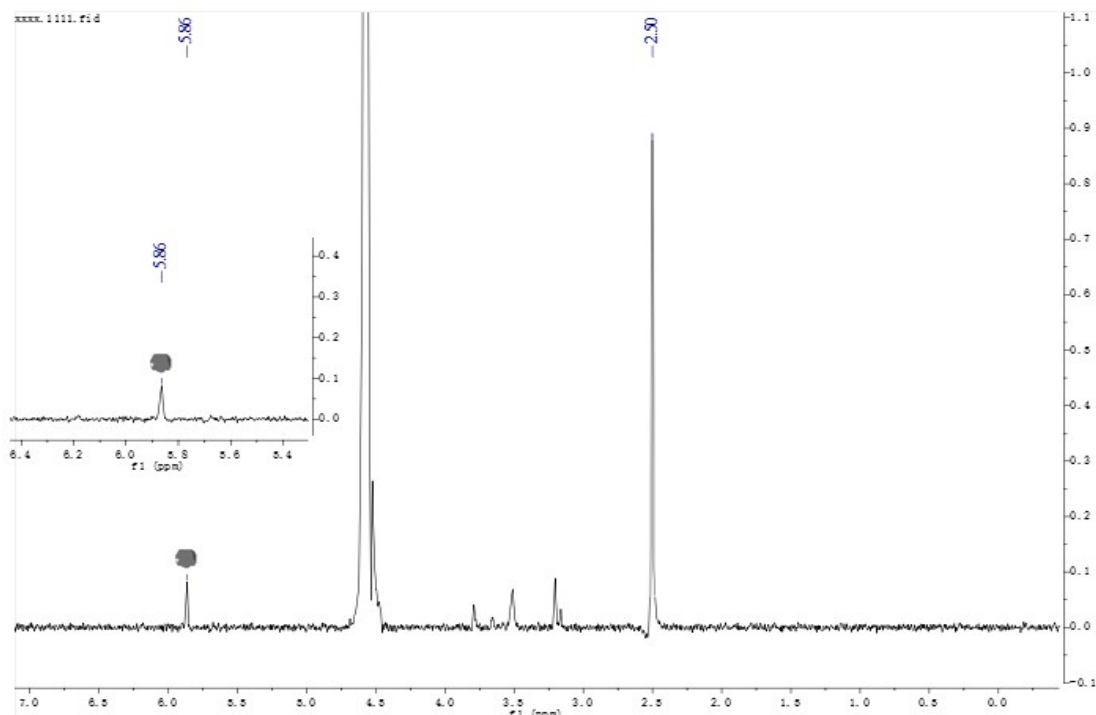


Fig.25. ^1H NMR spectra of complex **1a** treated with PBS (7.87 mM, pH 7.4; at 37°C) at 200 mins (DMSO- d_6 / D_2O , 10/90, v/v). \blacktriangle indicates the peak of coordinated DCA in complex **1a**. \bullet indicates the peak of free DCA.

(b) ^1H -NMR(400 MHz, ^1H) spectra of complex **1b** in deuterated aqueous, phosphate buffered solution

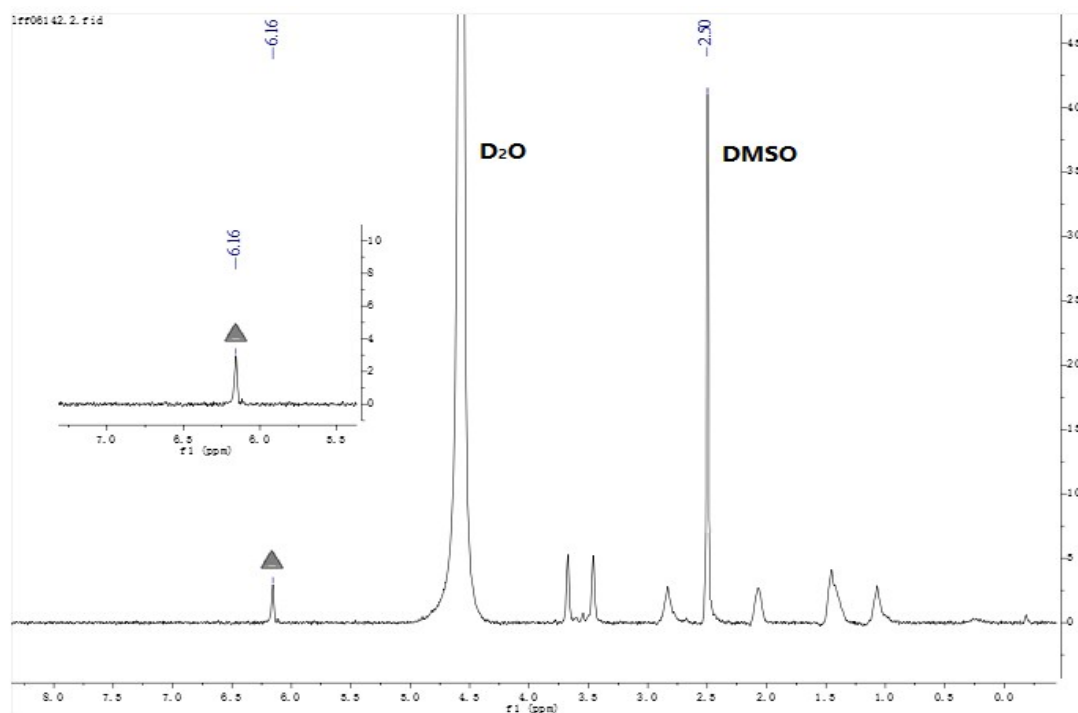


Fig.26. ^1H NMR spectra of complex **1b** treated with PBS (7.87 mM, pH 7.4; at 37°C) at 0 min (DMSO- d_6 / D_2O , 10/90, v/v). \blacktriangle indicates the peak of coordinated DCA in complex **1b**.

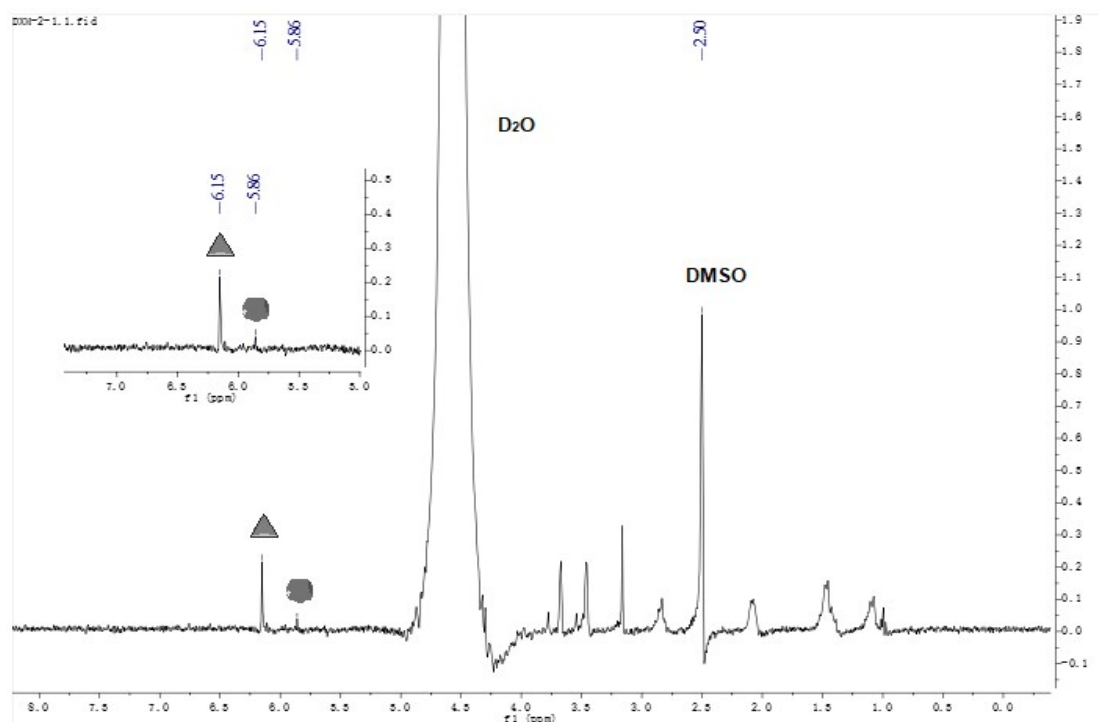


Fig.27. ^1H NMR spectra of complex **1b** treated with PBS (7.87 mM, pH 7.4; at 37°C) at 30 mins (DMSO- d_6 /D $_2$ O, 10/90, v/v). \blacktriangle indicates the peak of coordinated DCA in complex **1b**. \bullet indicates the peak of free DCA.

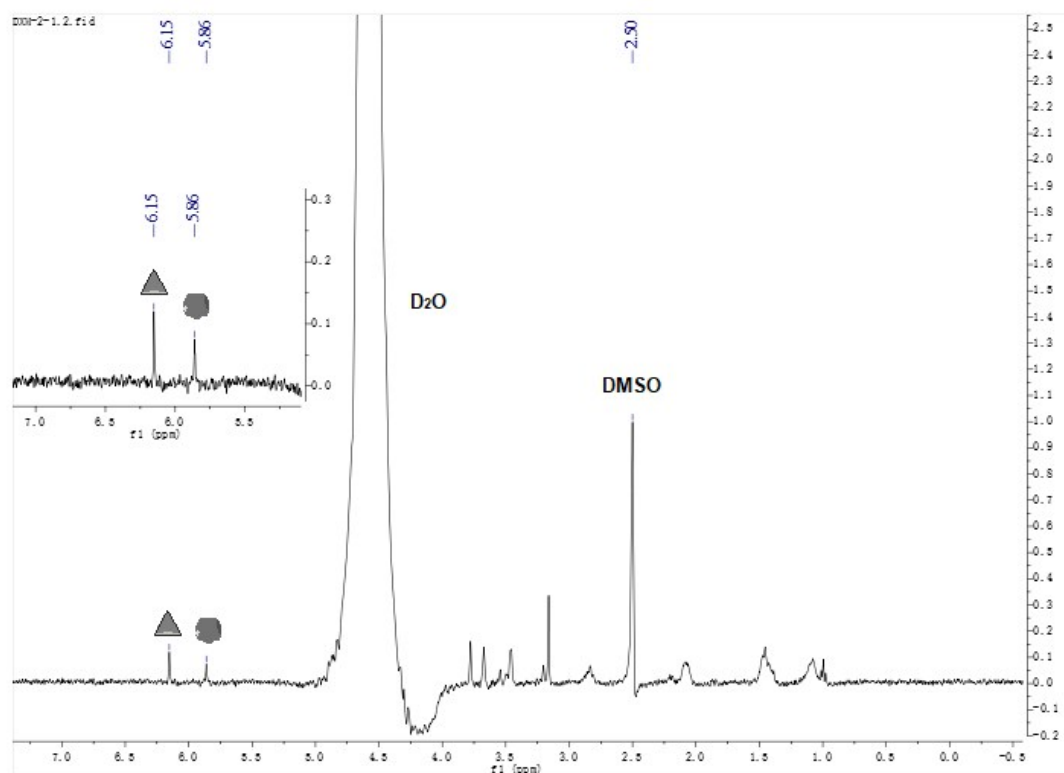


Fig.28. ^1H NMR spectra of complex **1b** treated with PBS (7.87 mM, pH 7.4; at 37°C) at 60 mins (DMSO- d_6 /D $_2$ O, 10/90, v/v). \blacktriangle indicates the peak of coordinated DCA in complex **1b**. \bullet indicates the peak of free DCA.

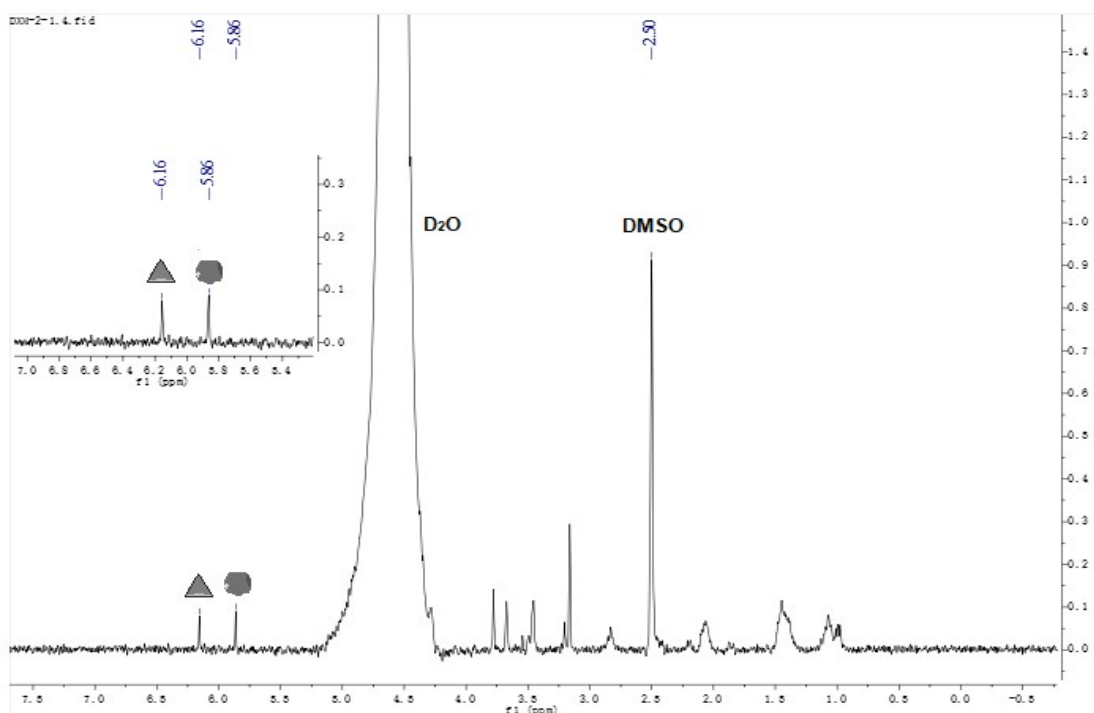


Fig.29. ^1H NMR spectra of complex **1b** treated with PBS (7.87 mM, pH 7.4; at 37°C) at 120 mins (DMSO- d_6 /D $_2$ O, 10/90, v/v). \blacktriangle indicates the peak of coordinated DCA in complex **1b**. \bullet indicates the peak of free DCA.

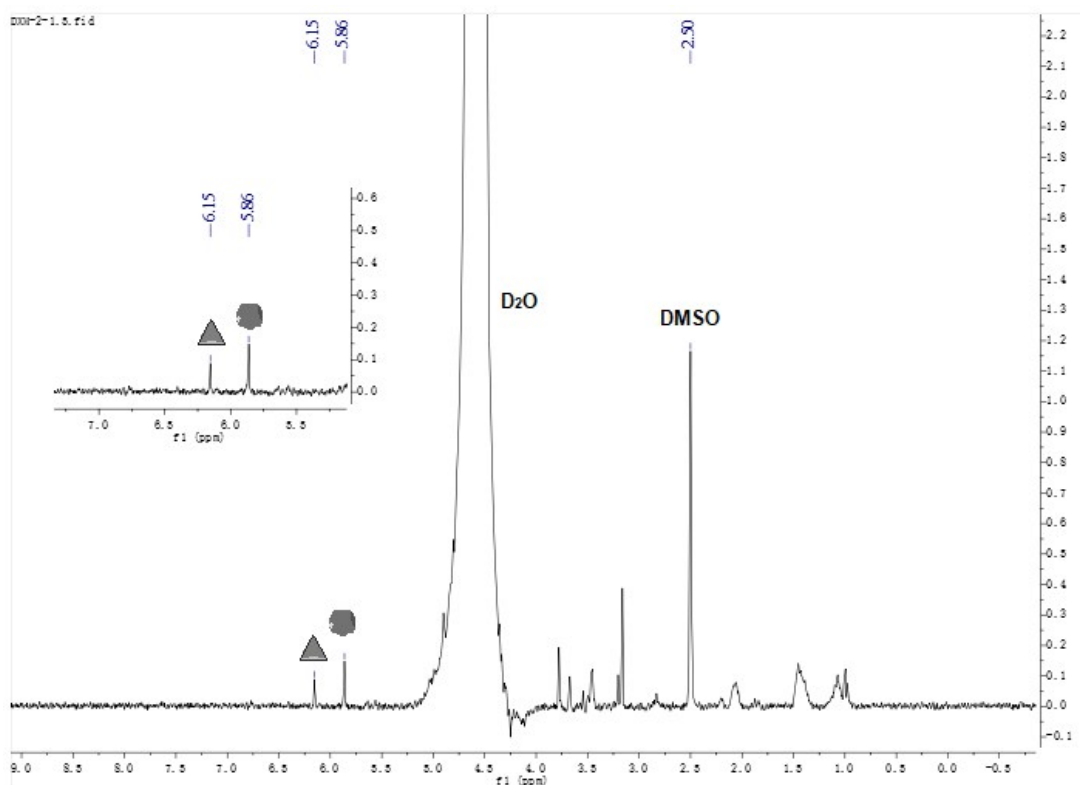


Fig.30. ^1H NMR spectra of complex **1b** treated with PBS (7.87 mM, pH 7.4; at 37°C) at 180 mins (DMSO- d_6 /D $_2$ O, 10/90, v/v). \blacktriangle indicates the peak of coordinated DCA in complex **1b**. \bullet indicates the peak of free DCA.

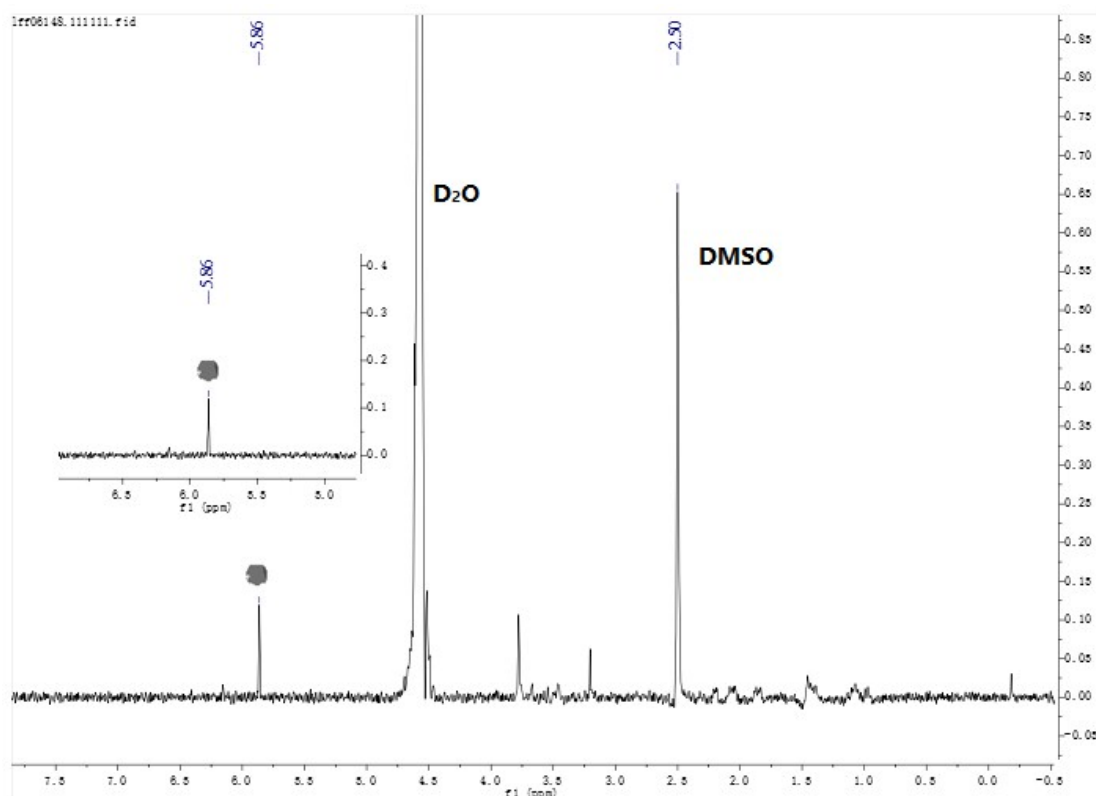


Fig.31. ^1H NMR spectra of complex **1b** treated with PBS (7.87 mM, pH 7.4; at 37°C) at 250 mins (DMSO- d_6 /D $_2$ O, 10/90, v/v). ● indicates the peak of free DCA.

(c) HPLC spectra of complex **1a** and **1b** in phosphate buffered solution

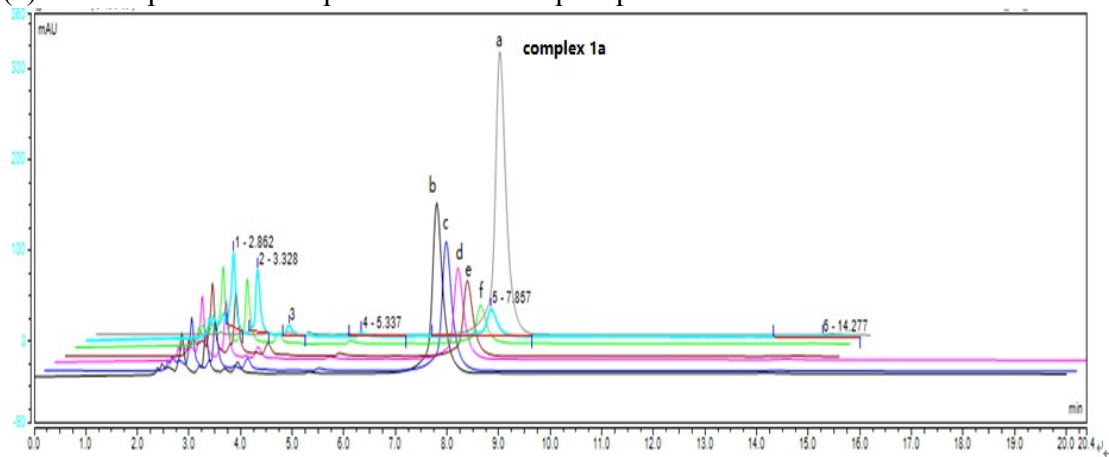


Fig. S32. Complex **1a** in PBS (PBS 7.87 mM, pH 7.4; at 37°C): (a) 0 min; (b) 50 min; (c) 80 min; (d) 100 min; (e) 150min; (f) 180min (250 \times 4.6 mm ODS; 226 nm; MeCN : H $_2$ O = 25:75 (v/v); 1.0 mL/min)

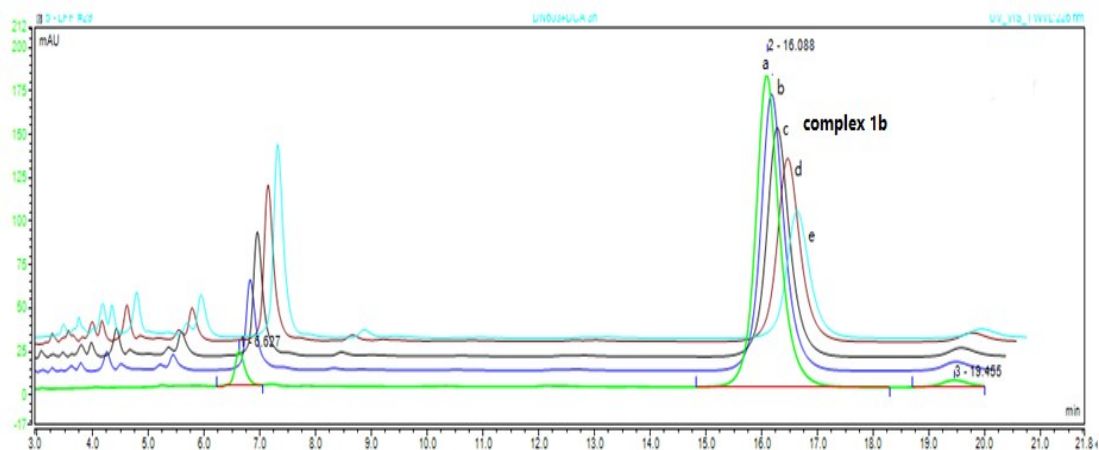


Fig. S33. Complex **1b** in PBS (PBS 7.87 mM, pH 7.4; at 37°C): (a) 0 min; (b) 15 min; (c) 30 min; (d) 60 min; (e) 150min (250×4.6 mm ODS; 226 nm; MeCN : H₂O = 25:75 (v/v); 1.0 mL/min)

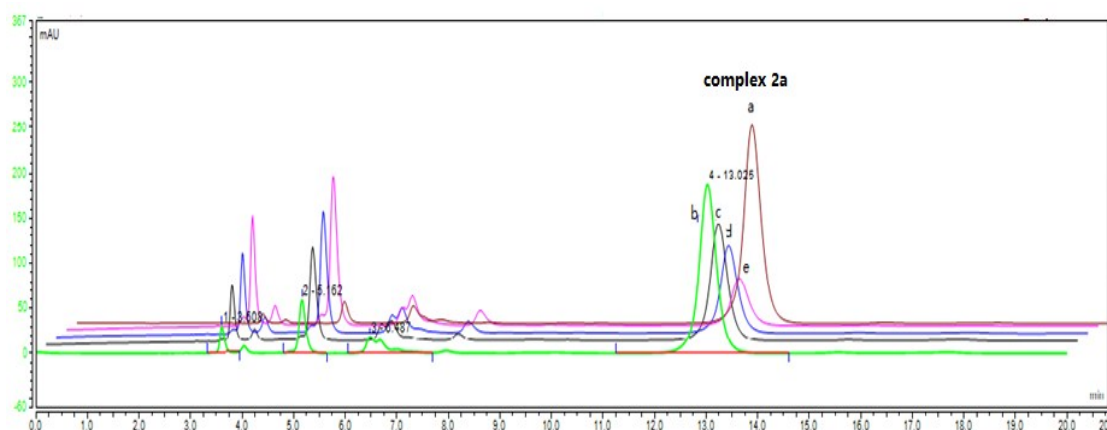


Fig. S34. Complex **2a** in PBS (PBS 7.87 mM, pH 7.4; at 37°C): (a) 0 min; (b)30min; (c)60 min; (d)90 min; (e)150min (250×4.6 mm ODS; 226 nm; MeCN : H₂O = 25:75 (v/v); 1.0 mL/min)

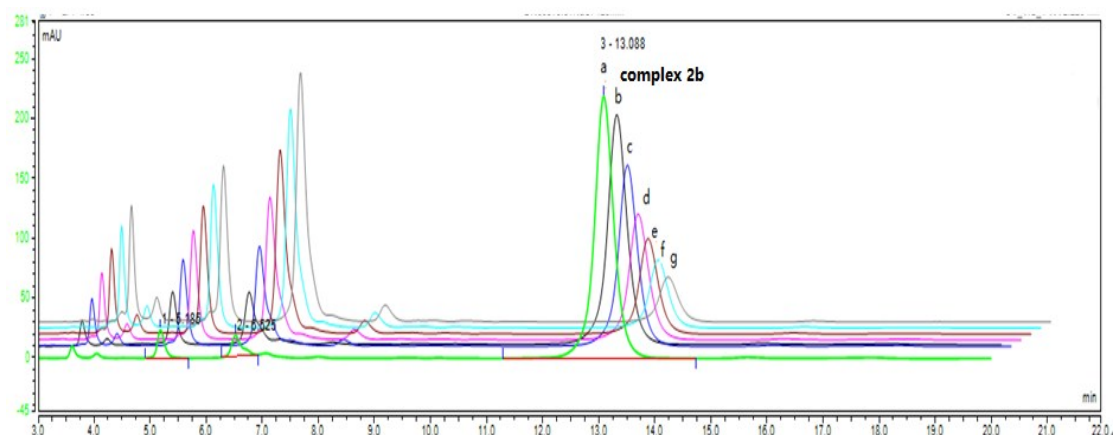


Fig. S35. Complex **2b** in PBS(PBS 7.87 mM, pH 7.4; at 37°C): (a) 0 min; (b)30min; (c)45 min; (d)60 min; (e)85min; (f)100min; (g)130min (250×4.6 mm ODS; 226 nm; MeCN : H₂O = 25:75 (v/v); 1.0 mL/min)

4. The stability of complexes 1a-2b in MeOH

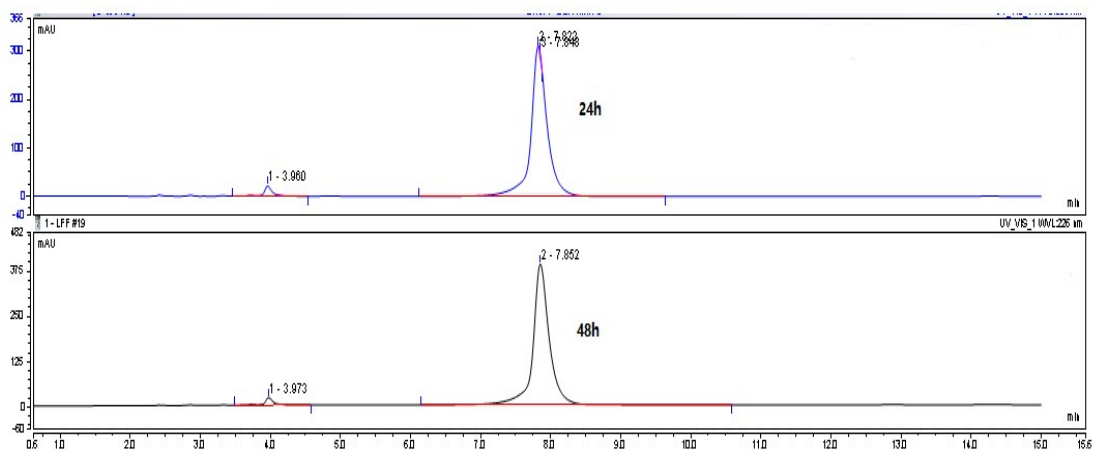


Fig.36. The stability of complexes **1a** in MeOH (0-48h); (250×4.6 mm ODS; 226 nm; MeCN : H₂O = 25:75 (v/v); 1.0 mL/min)

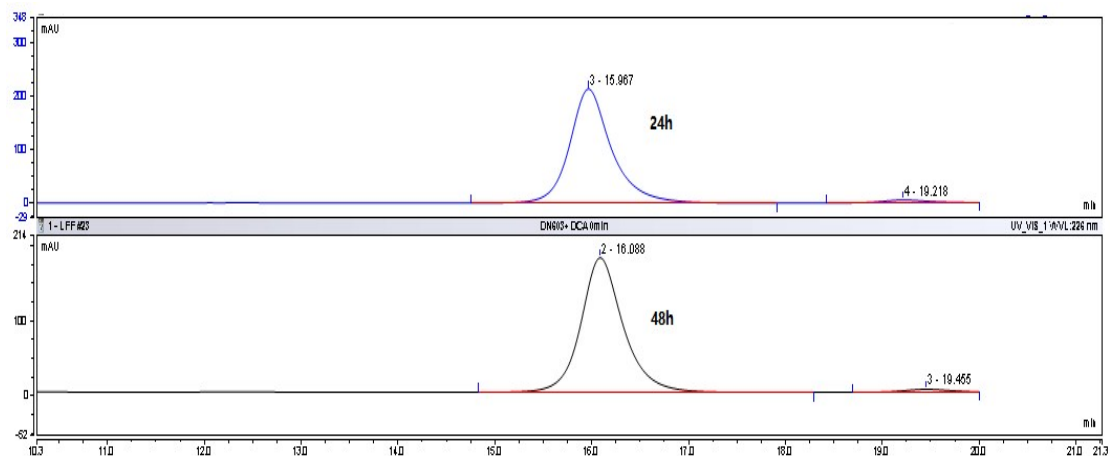


Fig.37. The stability of complexes **1b** in MeOH (0-48h); (250×4.6 mm ODS; 226 nm; MeCN : H₂O = 25:75 (v/v); 1.0 mL/min)

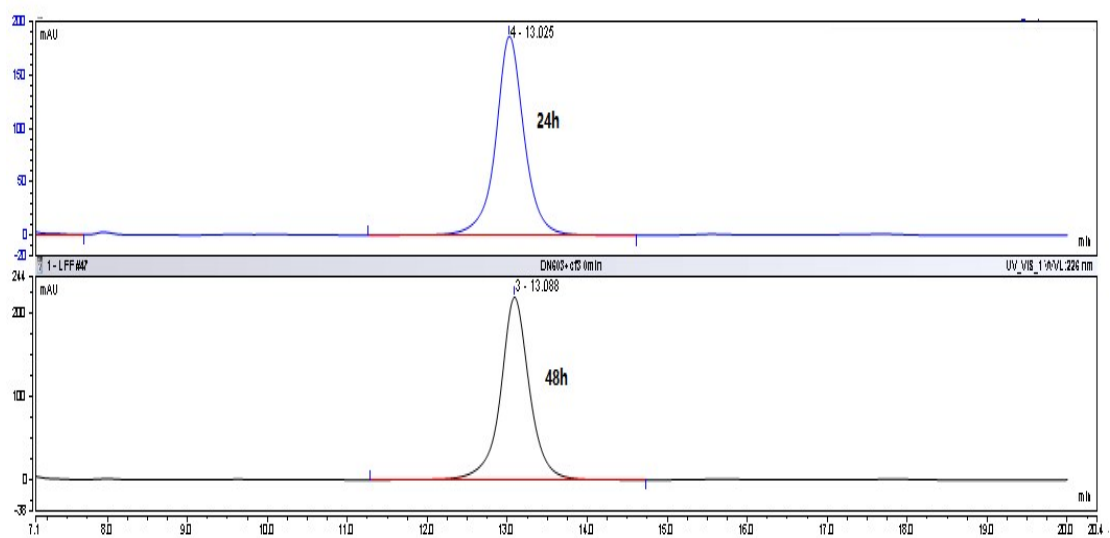


Fig.38. The stability of complexes **2a** in MeOH (0-48h); (250×4.6 mm ODS; 226 nm; MeCN : H₂O = 25:75 (v/v); 1.0 mL/min)

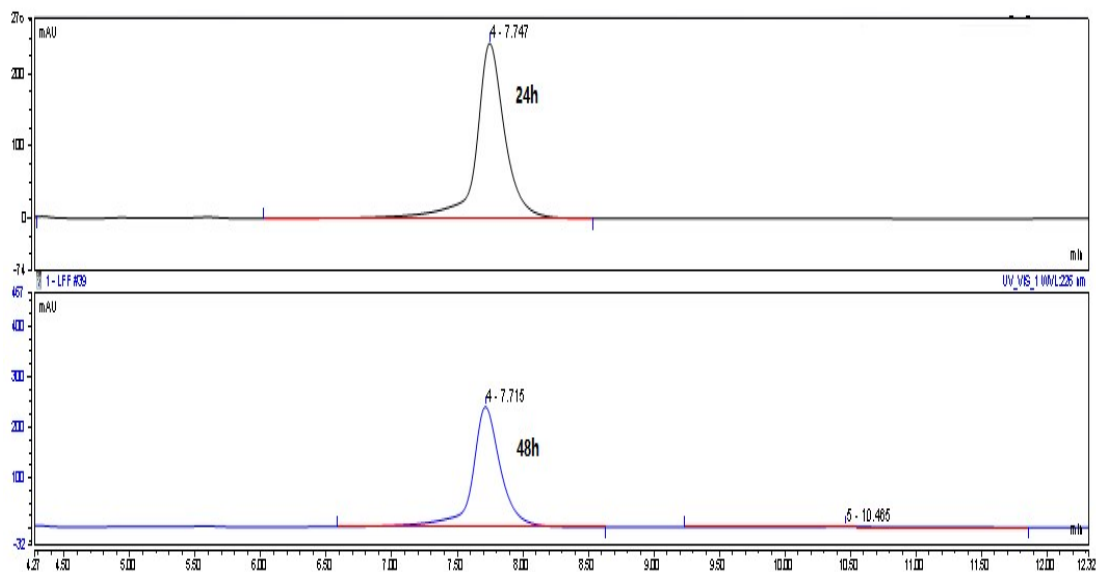


Fig.39. The stability of complexes **2b** in MeOH (0-48h); (250×4.6 mm ODS; 226 nm; MeCN : H₂O = 25:75 (v/v); 1.0 mL/min)

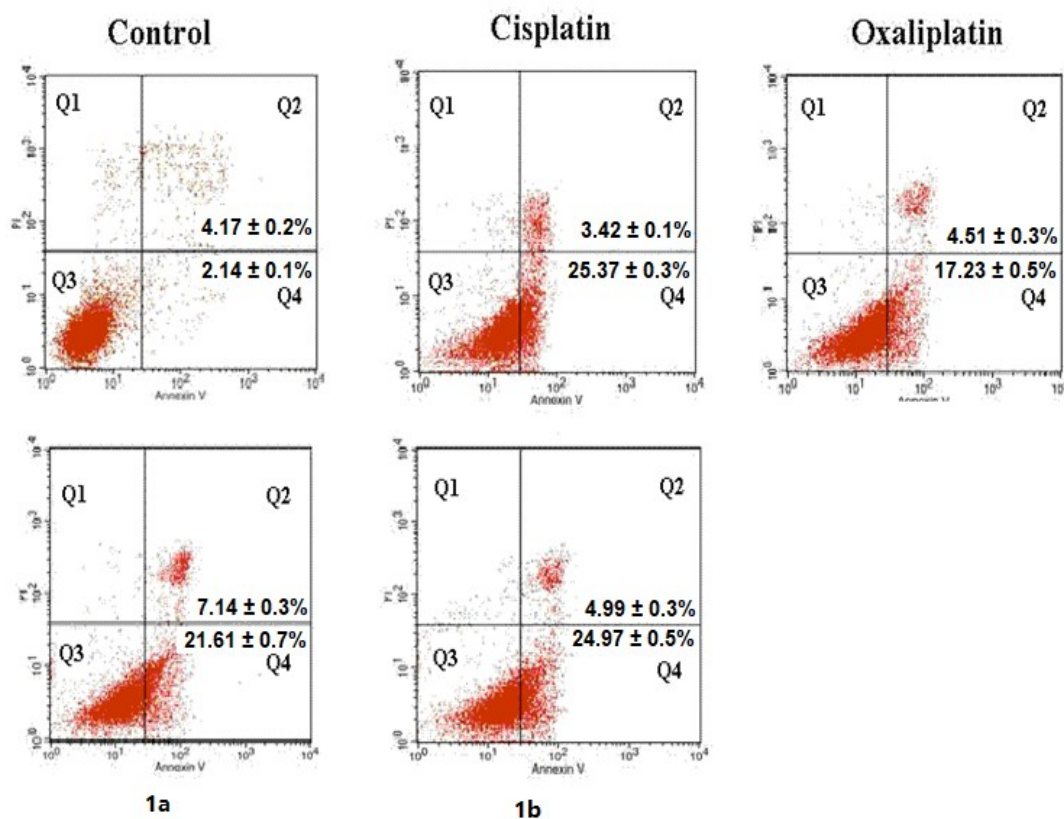


Fig.40. Flow cytometric analysis of the distribution of HepG-2 cells treated with 50 μ M of cisplatin, oxaliplatin, complex **1a** and **1b** for 24 hours (DMF final concentration < 0.4%). Cells were stained with 5 mL of Annexin V-FITC and incubated in the dark at 25 °C for 10 mins. The fluorescence was measured by using a flow cytometer. The results were expressed as the percentage of normal and apoptotic cells at various stages by FCSExpress software.

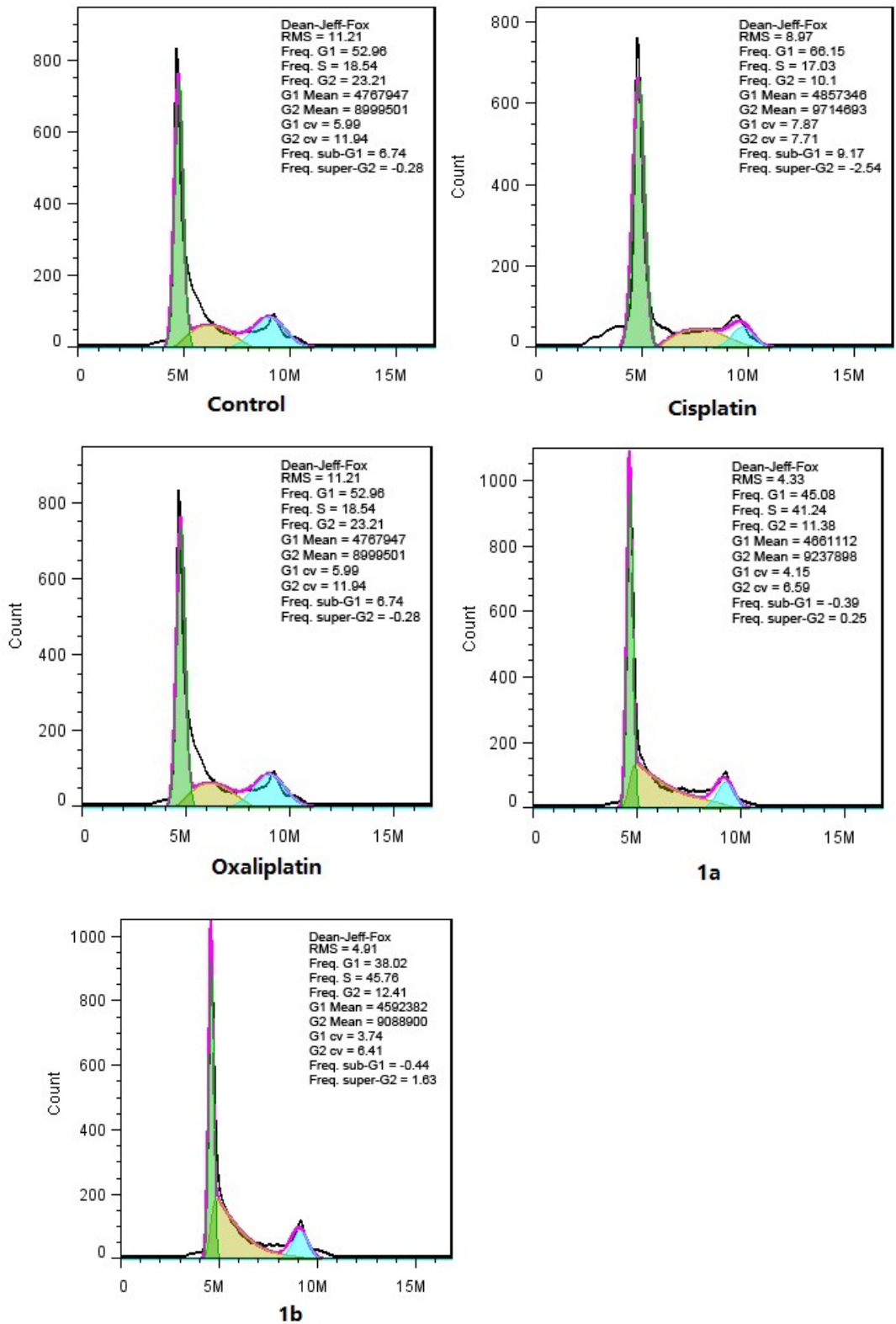


Fig.41. Cell cycle distribution of HepG-2 cells cultured in the presence of 30 μ M of cisplatin, oxaliplatin, complex **1a** and **1b** (DMF final concentration < 0.4%). The cells were seeded in 6-well plates for 12 hours, and then the cells were treated with cisplatin, oxaliplatin, complex **1a** and **1b**. After 12 hours of treatment, cells were harvested with trypsin and washed twice with PBS.