

Copper-catalysed enantioselective intramolecular etherification of propargylic esters: Synthetic approach to chiral isochromans

Shiyao Liu,^[a] Kazunari Nakajima,^[b] and Yoshiaki Nishibayashi*^[a]

^[a] *Department of Systems Innovation, School of Engineering, The University of Tokyo, Hongo, Bunkyo-ku, Tokyo, 113-8656 (Japan)*

^[b] *Frontier Research Centre for Energy and Resources, School of Engineering, The University of Tokyo, Hongo, Bunkyo-ku, Tokyo, 113-8656 (Japan)*

List of Contents of Supporting Information

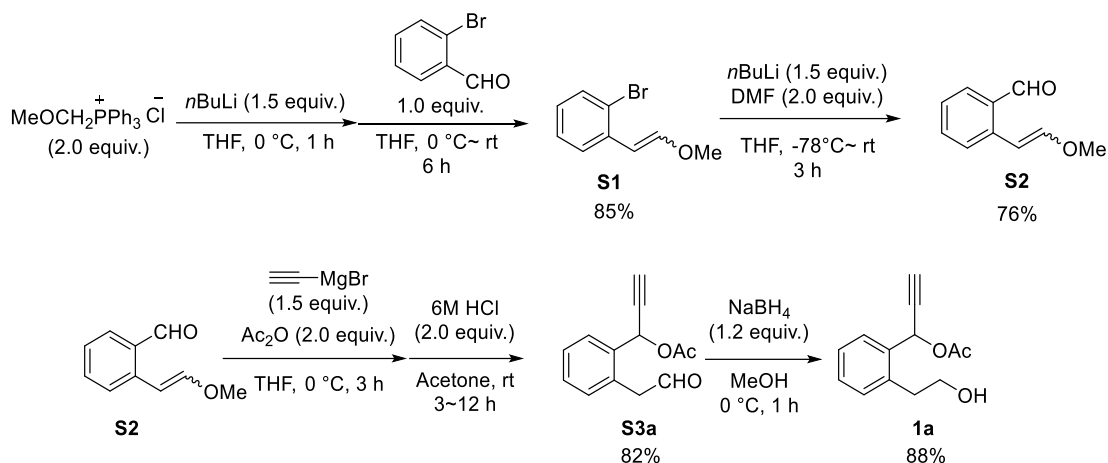
1. General Methods	Page S2
2. General Procedure for the Preparation of Propargylic Acetates	Page S3
3. Spectroscopic Data of Other Propargylic Acetates.....	Page S6
4. Enantioselective Intramolecular Propargylic etherification of Propargylic Acetates	Page S9
5. Spectroscopic Data and Isolated Yields of Other Products	Page S10
6. Preparation of (S)-1-benzyl-4-(isochroman-1-yl)-1H-1,2,3-triazole (3)	Page S14
7. Preparation of (R)-1-((4-chlorophenyl)ethynyl)isochromane (4)	Page S15
8. Preparation of (S)-1-benzyl-4-(7-chloroisochroman-1-yl)-1H-1,2,3-triazole (S16).....	Page S15
9. X-ray Diffraction Study of S16	Page S16
10. References	Page S18
11. ¹ H and ¹³ C NMR Spectra.....	Page S19
12. HPLC Charts.....	Page S52

General Methods.

^1H NMR (400 MHz) and ^{13}C NMR (100 MHz) spectra were measured on a JEOL ECS400 400 MHz spectrometer using CDCl_3 as solvent. The data are reported as (s = singlet, d = doublet, dd = double doublet, dt = double triplet, t = triplet, q = quartet, br = broad, m = multiplet or unresolved, coupling constant(s) in Hz, integration). HPLC analyses were performed on Hitachi L-7100 and GL-7480 apparatuses equipped with a UV detector using 25 cm x 4.6 mm DAICEL Chiralpak OJ, OZ columns. Mass spectra were measured on a JEOL JMS-700 mass spectrometer. Specific rotations were measured on a JASCO DIP-1000 polarimeter. All reactions were carried out under dry nitrogen atmosphere. The synthesis of racemic products were carried out by using racemic Ph-pybox ligand at 60 °C. Commercially obtained reagents were used without further purification. All reactions were monitored by TLC with silica gel-coated plates. Solvents were dried by the usual methods, then distilled under N_2 and degassed before use. Optically pure pybox ligands **L2**, **L5** are commercially available reagents. Optically pure pybox ligands **L1**^{S1}, **L3**^{S2}, **L4**^{S3} were prepared according to literature procedures. The absolute configuration of chiral derivative product **S16** were determined by unequivocally according to the X-ray diffraction analysis, and the absolute configuration chiral alkynyl isochromans **2** were deduced on the basis of this result.

General Procedure for the Preparation of Propargylic Acetates.

Scheme S1 Preparation of propargylic acetate **1a**



A typical experimental procedure for the preparation of 1-(2-(2-hydroxyethyl)phenyl)prop-2-yn-1-yl acetate **1a** is described below. In a 50 mL Schlenk flask were placed (methoxymethyl)triphenylphosphonium chloride (3.43 g, 10.0 mmol) and anhydrous THF (20 mL) under N₂ atmosphere and cooled down the solution to 0 °C. To the solution, *n*BuLi (1.57 M in hexane, 4.8 mL, 7.5 mmol) was added dropwise and the mixture was stirred at 0 °C for 1 h. 2-Bromobenzaldehyde was added to this solution by portion and the mixture was warmed to room temperature and stirred for another 6 h. After the reaction, mixture was quenched by saturated NH₄Cl aq., and the solution was extracted with CH₂Cl₂ (10 mL X 3). The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was passed through a short column chromatography with hexane/ethyl acetate (50:1) to give 1-bromo-2-(2-methoxyvinyl)benzene (**S1**) as an E/Z mixture (8.5 mmol, 85% isolated yield).

In a 50 mL Schlenk flask were placed **S1** (8.00 mmol) and anhydrous THF (20 mL) under N₂ atmosphere. The solution was cooled down to -78 °C. *n*BuLi (1.57 M in hexane, 7.60 mL, 12.0 mmol) was added dropwise and the mixture was stirred at 0 °C for 1 h. Anhydrous DMF (1.20 mL, 16.0 mmol) was added to the solution, and the mixture was warmed to room temperature and stirred for another 2 h. After the reaction, mixture was quenched by saturated NH₄Cl aq., and the solution was extracted with CH₂Cl₂ (10 mL X 3). The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by column chromatography with hexane/EtOAc (50:1-30:1) to give 2-(2-methoxyvinyl)benzaldehyde (**S2**) as an E/Z mixture (6.10 mmol, 76% isolated yield).

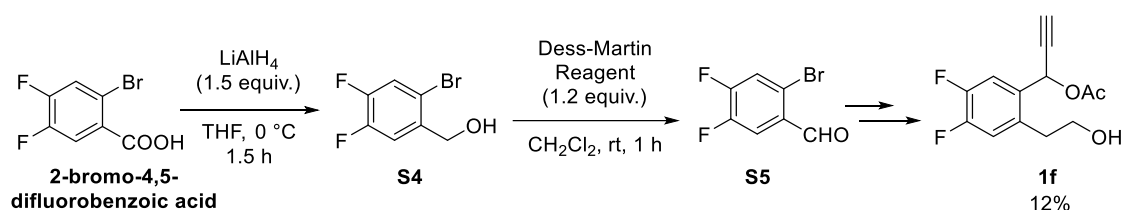
To a solution of **S2** (6.00 mmol) in anhydrous THF (15 mL) was added ethynylmagnesium bromide (0.5 M in THF, 18.0 mL, 9.00 mmol) at 0 °C and the mixture was stirred for 1 h. Acetic anhydride (1.10 mL, 12.0 mmol) was added to the solution and the mixture was allowed to warm to room temperature. After stirring for 2 h, the mixture was quenched by saturated NH₄Cl aq., and the solution was extracted with CH₂Cl₂ (10 mL X 3). The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was dissolved in 10 mL acetone, and 6M HCl (2.00 mL, 12.0 mmol) was added to the solution at room temperature. The reaction was monitored by TLC until the starting material vanished. After the reaction, the solution was diluted by H₂O and extracted with CH₂Cl₂ (10 mL X 3). The

combined organic layers were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by column chromatography with hexane/EtOAc (20:1-10:1) to give 1-(2-(2-oxoethyl)phenyl)prop-2-yn-1-yl acetate (**S3a**) as a yellow oil (4.90 mmol, 82% isolated yield).

S3a (865 mg, 4.00 mmol) was dissolved in 10 mL MeOH, and the solution was cooled down to 0 °C. NaBH₄ (182 mg, 4.80 mmol) was added to the solution and the mixture was stirred for 1 h at 0 °C. After the reaction, the solution was diluted by H₂O and extracted with CH₂Cl₂ (10 mL X 3). The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by column chromatography with hexane/EtOAc (10:1-4:1) to give 1-(2-(2-hydroxyethyl)phenyl)prop-2-yn-1-yl acetate (**1a**) as a pale yellow oil (3.5 mmol, 88% isolated yield). ¹H NMR δ 7.62 (d, J = 7.2 Hz, 1H), 7.27-7.20 (m, 3H), 6.61 (d, J = 2.4 Hz, 1H), 3.78 (t, J = 6.8 Hz, 2H), 3.01-2.90 (m, 3H), 2.65 (d, J = 2.4 Hz, 1H), 2.04 (s, 3H). ¹³C NMR δ 169.6, 136.5, 134.8, 130.3, 129.0, 128.1, 126.7, 80.2, 75.5, 62.80, 62.77, 35.3, 20.7. HRMS (FAB) Calcd. for C₁₃H₁₄NaO₃ [M+Na]: 241.0841. Found: 241.0840.

Substrates **1b**, **1c**, **1d**, **1e**, **1g**, **1h**, **1i**, **1k**, **1l**, **1m** were synthesized by using various substituted halogen benzaldehydes, as a similar method as the synthesis of **1a**.

Scheme S2 Preparation of propargylic acetate **1f**



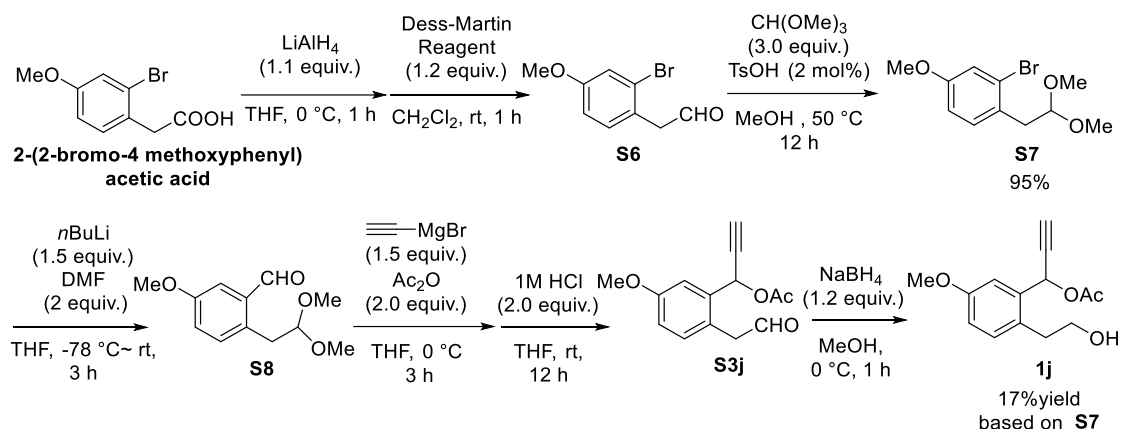
In a 50 mL Schlenk flask were placed 2-bromo-4,5-difluorobenzoic acid (1.19 g, 5.00 mmol) anhydrous THF (20 mL) under N₂ atmosphere. The solution was cooled down to 0 °C. LiAlH₄ (285 mg, 7.50 mmol) was added by portion and the mixture was stirred at 0 °C for 1.5 h. The mixture was quenched saturated NH₄Cl aq., and the suspension was filtered through a celite pad. Then the filtrate was extracted with EtOAc (10 mL X 3). The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. Crude (2-bromo-4,5-difluoro) benzylic alcohol (**S4**) was used to the next step without further purification.

To a solution of **S4** in 20 mL CH₂Cl₂, Dess-Martin periodinane (2.54 g, 6.00 mmol) was added. The mixture was stirred for 1 h at room temperature. After the reaction, the solution was diluted by H₂O and extracted with CH₂Cl₂ (10 mL X 3). The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by column chromatography with hexane/ethyl acetate (50:1-40:1) to give the corresponding 2-bromo-4,5-difluorobenzaldehyde (**S5**).

1f was synthesized from **S5**, as a similar method as the synthesis of **1a** (0.60 mmol, 12% total yield). A pale yellow oil. ¹H NMR δ 7.46 (dd, J = 11.2 Hz, 8.0 Hz, 1H), 7.07 (dd, J = 11.2 Hz, 8.0 Hz, 1H), 6.54 (d, J = 2.0 Hz, 1H), 3.84 (t, J = 6.4 Hz, 2H), 3.00-2.82 (m, 2H), 2.65 (d, J = 2.0 Hz, 1H), 2.10 (s, 3H), 1.79 (br, 1H). ¹³C NMR δ 169.5, 150.3 (dd, ¹J_{C-F} = 250 Hz and ²J_{C-F} = 12.4 Hz), 149.0 (dd, ¹J_{C-F} = 250 Hz and ²J_{C-F} = 12.4 Hz), 133.8 (dd, ³J_{C-F} = 5.7 Hz and ⁴J_{C-F} = 3.8 Hz), 132.0 (t, ³J_{C-F} = ⁴J_{C-F} = 4.3 Hz), 119.0 (d, ²J_{C-F} = 17.1 Hz), 117.3 (d, ²J_{C-F} = 18.1 Hz), 79.7, 75.9, 62.8, 61.9,

34.7, 20.8. HRMS (EI) Calcd. for $C_{13}H_{12}F_2O_3$ [M]: 254.0755. Found: 254.0760.

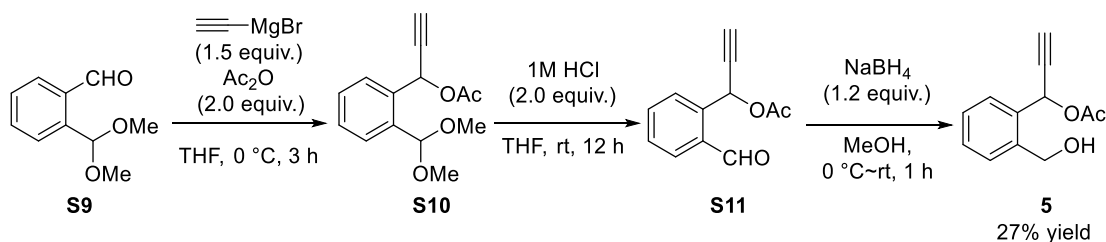
Scheme S3 Preparation of propargylic acetate **1j**



2-(2-Bromo-4-methoxyphenyl)acetaldehyde (**S6**) was synthesized from the 2-(2-bromo-4-methoxyphenyl)acetic acid as a similar method as the synthesis of **S5**. To the solution of crude **S6** (5.00 mmol) in 30 mL MeOH, trimethyl orthoformate (1.60 mL, 15.0 mmol) and TsOH.H₂O (19.0 mg, 0.10 mmol) were added. The mixture was stirred at 50 °C for 3 h. After the reaction, the resulting mixture was evaporated in vacuo and the residue was purified by silica gel column to give 2-bromo-1-(2,2-dimethoxyethyl)-4-methoxybenzene **S7** (4.75 mmol, 95% yield).

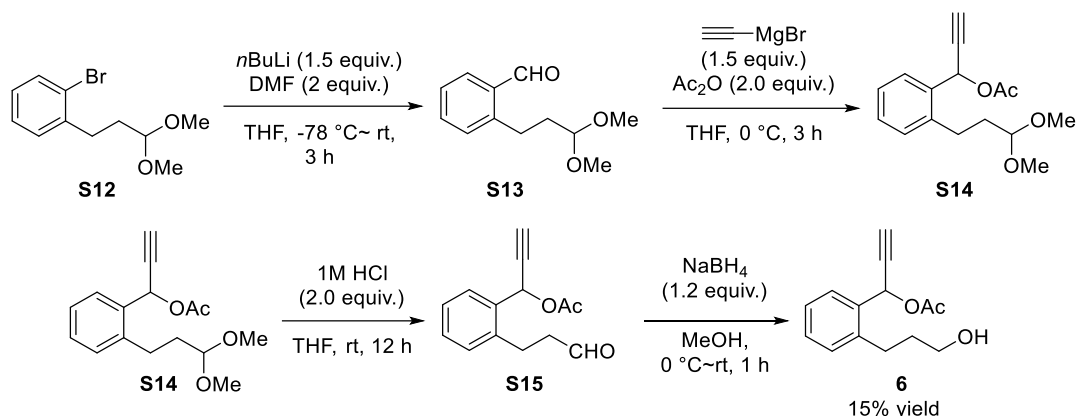
1-(2-(2-hydroxyethyl)-5-methoxyphenyl)prop-2-yn-1-yl acetate **1j** was synthesized from **S7** as a similar method as the synthesis of **1a** (0.80 mmol, 17% total yield from **S7**). A pale yellow oil. ¹H NMR δ 7.19 (d, J = 2.8 Hz, 1H), 7.15 (d, J = 9.0 Hz, 1H), 6.87 (dd, J = 9.0 Hz, 2.8 Hz, 1H), 6.59 (d, J = 2.4 Hz, 1H), 3.83-3.79 (m, 5H), 2.98-2.84 (m, 2H), 2.64 (d, J = 2.4 Hz, 1H), 2.10 (s, 3H), 2.00 (br, 1H). ¹³C NMR δ 169.6, 158.4, 136.2, 131.6, 128.4, 114.7, 113.6, 80.3, 75.5, 63.3, 62.9, 55.3, 34.8, 20.9. HRMS(EI) Calcd. for $C_{14}H_{16}O_4$ [M]: 248.1049. Found: 248.1052.

Scheme S4 Preparation of propargylic acetate **5**



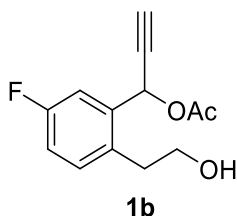
1-(2-(hydroxymethyl)phenyl)prop-2-yn-1-yl acetate **5** was synthesized from **S9**^{S4} (6.00 mmol) as a similar method as the synthesis of **1a** (1.60 mmol, 27% total yield). A pale yellow oil. ¹H NMR δ 7.73-7.70 (m, 1H), 7.44-7.31 (m, 3H), 5.69-5.68 (m, 1H), 5.35 (d, J = 12.8 Hz, 1H), 5.26 (d, J = 12.8 Hz, 1H), 3.31 (d, J = 5.4 Hz, 1H), 2.67 (d, J = 2.4 Hz, 1H), 2.08 (s, 3H). ¹³C NMR δ 171.0, 138.3, 133.4, 129.9, 128.8, 128.7, 127.4, 83.1, 75.0, 63.7, 61.8, 20.9. HRMS (EI) Calcd. for $C_{12}H_{12}O_3$ [M]: 204.0786. Found: 204.0784.

Scheme S5 Preparation of propargylic acetate **6**

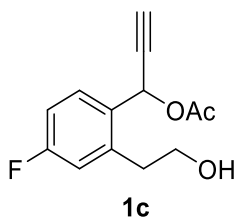


1-(2-(3-Hydroxypropyl)phenyl)prop-2-yn-1-yl acetate **6** was synthesized from **S12**⁵⁵ (6.00 mmol) according to a similar method as the synthesis of **1a** (0.90 mmol, 15% total yield). ¹H NMR δ 7.63 (dd, J = 7.2 Hz, 1.2 Hz, 1H), 7.32-7.19 (m, 3H), 6.63 (d, J = 2.2 Hz, 1H), 3.67 (t, J = 6.4 Hz, 2H), 2.85-2.77 (m, 2H), 2.62 (d, J = 2.2 Hz, 1H), 2.10 (s, 3H), 2.00 (br, 1H), 1.91-1.82 (m, 2H). ¹³C NMR δ 169.7, 139.8, 134.4, 129.8, 129.2, 128.2, 126.5, 80.5, 75.3, 62.7, 61.9, 34.0, 28.3, 21.0. HRMS (EI) Calcd. for C₁₄H₁₆O₃ [M]: 232.1099. Found: 232.1091.

Spectroscopic Data of Other Propargylic Acetates

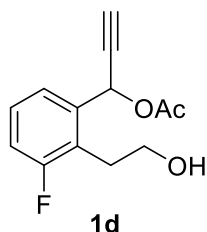


1-(5-fluoro-2-(2-hydroxyethyl)phenyl)prop-2-yn-1-yl acetate (1b) : a pale yellow oil. ¹H NMR δ 7.33 (dd, J = 9.6 Hz, 2.6 Hz, 1H), 7.18 (dd, J = 8.2 Hz, 6.0 Hz, 1H), 6.98 (td, J = 8.2 Hz, 2.6 Hz, 1H), 6.56 (d, J = 2.4 Hz, 1H), 3.78 (t, J = 6.8 Hz, 2H), 2.96-2.84 (m, 2H), 2.64 (d, J = 2.4 Hz, 1H), 2.42 (br, 1H), 2.08 (s, 3H). ¹³C NMR δ 169.6, 161.5 (d, ¹J_{C-F} = 244 Hz), 137.1 (d, ³J_{C-F} = 6.7 Hz), 132.1 (d, ⁴J_{C-F} = 2.9 Hz), 132.0 (d, ³J_{C-F} = 6.6 Hz), 116.0 (d, ²J_{C-F} = 21 Hz), 114.8 (d, ²J_{C-F} = 23 Hz), 79.8, 75.8, 62.9, 62.3, 34.8, 20.8. HRMS (EI) Calcd. for C₁₃H₁₃FO₃ [M]: 236.0849. Found: 236.0857.

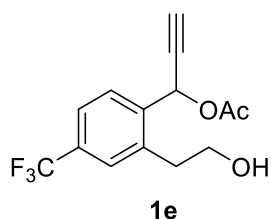


1-(4-fluoro-2-(2-hydroxyethyl)phenyl)prop-2-yn-1-yl acetate (1c) : a pale yellow oil. ¹H NMR δ 7.64 (dd, J = 9.6 Hz, 5.6 Hz, 1H), 7.00-6.95 (m, 2H), 6.60 (d, J = 2.4 Hz, 1H), 3.89 (t, J =

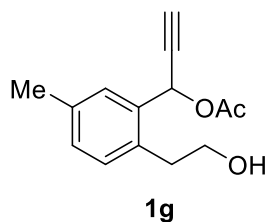
6.4 Hz, 2H), 3.06-2.93 (m, 2H), 2.65 (d, $J = 2.4$ Hz, 1H), 2.11 (s, 3H), 1.69 (br, 1H). ^{13}C NMR δ 169.7, 162.9 (d, $^1J_{\text{C-F}} = 247$ Hz), 139.5 (d, $^3J_{\text{C-F}} = 7.7$ Hz), 131.1 (d, $^4J_{\text{C-F}} = 1.9$ Hz), 130.5 (d, $^3J_{\text{C-F}} = 8.6$ Hz), 117.0 (d, $^2J_{\text{C-F}} = 21$ Hz), 113.9 (d, $^2J_{\text{C-F}} = 22$ Hz), 80.3, 75.6, 62.8, 62.4, 35.4, 20.9. HRMS (EI) Calcd. for $\text{C}_{13}\text{H}_{13}\text{FO}_3$ [M]: 236.0849. Found: 236.0844.



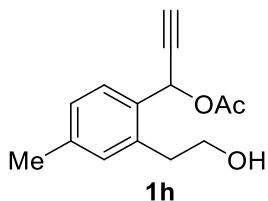
1-(3-fluoro-2-(2-hydroxyethyl)phenyl)prop-2-yn-1-yl acetate (1d): a pale yellow oil. ^1H NMR δ 7.44 (d, $J = 8.0$ Hz, 1H), 7.31-7.24 (m, 1H), 7.07 (t, $J = 9.0$ Hz, 1H), 6.66 (d, $J = 2.2$ Hz, 1H), 3.87 (t, $J = 6.8$ Hz, 2H), 3.11-3.04 (m, 2H), 2.66 (d, $J = 2.2$ Hz, 1H), 2.13 (s, 3H), 1.75 (br, 1H). ^{13}C NMR δ 169.6, 161.5 (d, $^1J_{\text{C-F}} = 244$ Hz), 137.8 (d, $^3J_{\text{C-F}} = 3.8$ Hz), 128.2 (d, $^3J_{\text{C-F}} = 9.5$ Hz), 124.4 (d, $^2J_{\text{C-F}} = 16.2$ Hz), 123.8 (d, $^4J_{\text{C-F}} = 2.9$ Hz), 116.0 (d, $^2J_{\text{C-F}} = 22.9$ Hz), 80.1, 75.7, 62.8 (d, $^4J_{\text{C-F}} = 3.8$ Hz), 62.3, 28.8, 21.0. HRMS (EI) Calcd. for $\text{C}_{13}\text{H}_{13}\text{FO}_3$ [M]: 236.0849. Found: 236.0842.



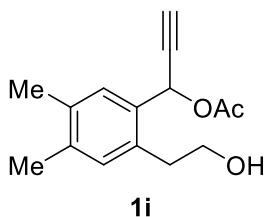
1-(2-(2-hydroxyethyl)-4-(trifluoromethyl)phenyl)prop-2-yn-1-yl acetate (1e): a pale yellow oil. ^1H NMR δ 7.75 (d, $J = 7.6$ Hz, 1H), 7.54-7.50 (m, 2H), 6.67 (s, 1H), 3.88 (t, $J = 6.8$ Hz, 2H), 3.13-3.00 (m, 2H), 2.67 (d, $J = 2.0$ Hz, 1H), 2.30 (br, 1H), 2.11 (s, 3H). ^{13}C NMR δ 169.6, 139.1, 137.7, 131.2 (q, $^2J_{\text{C-F}} = 32$ Hz), 128.5, 127.2 (q, $^3J_{\text{C-F}} = 3.8$ Hz), 123.8 (q, $^3J_{\text{C-F}} = 3.8$ Hz), 123.8 (q, $^1J_{\text{C-F}} = 271$ Hz), 79.7, 76.1, 62.7, 62.4, 35.4, 20.8. HRMS (FAB) Calcd. for $\text{C}_{14}\text{H}_{13}\text{F}_3\text{NaO}_3$ [M+Na]: 309.0714. Found: 309.0715.



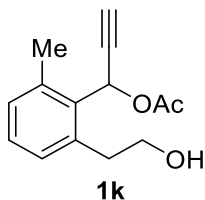
1-(2-(2-hydroxyethyl)-5-methylphenyl)prop-2-yn-1-yl acetate (1g): a pale yellow oil. ^1H NMR δ 7.44 (s, 1H), 7.12 (s, 2H), 6.60 (d, $J = 2.0$ Hz, 1H), 3.80 (t, $J = 6.4$ Hz, 2H), 3.03-2.87 (m, 2H), 2.64 (d, $J = 2.0$ Hz, 1H), 2.34 (s, 3H), 2.30 (br, 1H), 2.09 (s, 3H). ^{13}C NMR δ 169.6, 136.6, 134.7, 133.4, 130.4, 129.9, 128.8, 80.5, 75.4, 63.2, 62.9, 35.1, 20.9. HRMS (EI) Calcd. for $\text{C}_{14}\text{H}_{16}\text{O}_3$ [M]: 232.1099. Found: 232.1099.



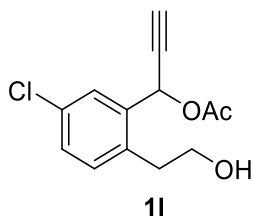
1-(2-(2-hydroxyethyl)-4-methylphenyl)prop-2-yn-1-yl acetate (1h): a pale yellow oil. ^1H NMR δ 7.54 (d, J = 7.6 Hz, 1H), 7.10 (d, J = 7.6 Hz, 1H), 7.07 (s, 1H), 6.61 (d, J = 2.2 Hz, 1H), 3.85 (t, J = 6.4 Hz, 2H), 3.06-2.88 (m, 2H), 2.63 (d, J = 2.2 Hz, 1H), 2.34 (s, 3H), 2.09 (br, 4H). ^{13}C NMR δ 169.7, 139.1, 136.4, 132.2, 131.1, 128.4, 127.8, 80.6, 75.3, 63.3, 62.9, 35.5, 21.0, 20.9. HRMS (FAB) Calcd. for $\text{C}_{14}\text{H}_{16}\text{NaO}_3$ [$\text{M}+\text{Na}$]: 255.0997. Found: 255.0998.



1-(2-(2-hydroxyethyl)-4,5-dimethylphenyl)prop-2-yn-1-yl acetate (1i): a pale yellow oil. ^1H NMR δ 7.42 (s, 1H), 7.03 (s, 1H), 6.60 (d, J = 2.2 Hz, 1H), 3.82 (t, J = 6.8 Hz, 2H), 3.04-2.86 (m, 2H), 2.66 (d, J = 2.2 Hz, 1H), 2.42 (br, 1H), 2.26 (s, 3H), 2.25 (s, 3H), 2.10 (s, 3H). ^{13}C NMR δ 169.6, 137.7, 135.2, 133.7, 132.2, 131.7, 129.5, 80.6, 75.2, 63.2, 62.8, 35.0, 20.8, 19.3, 19.1. HRMS (EI) Calcd. for $\text{C}_{15}\text{H}_{18}\text{O}_3$ [M]: 246.1256. Found: 246.1263.

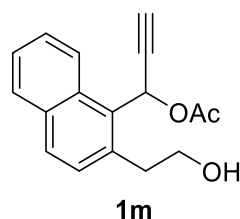


1-(2-(2-hydroxyethyl)-6-methylphenyl)prop-2-yn-1-yl acetate (1k): a pale yellow oil. ^1H NMR δ 7.20 (t, J = 7.2 Hz, 1H), 7.12-7.08 (m, 2H), 6.90 (d, J = 2.2 Hz, 1H), 3.93-3.83 (m, 2H), 3.30-3.24 (m, 1H), 3.10-3.03 (m, 1H), 2.61 (d, J = 2.2 Hz, 1H), 2.58 (s, 3H), 2.09 (s, 3H), 1.86 (br, 1H). ^{13}C NMR δ 169.6, 138.0, 137.5, 133.3, 130.0, 128.9, 128.8, 80.2, 75.0, 63.6, 61.2, 36.7, 20.9, 20.4. HRMS (FAB) Calcd. for $\text{C}_{14}\text{H}_{16}\text{NaO}_3$ [$\text{M}+\text{Na}$]: 255.0997. Found: 255.0992.



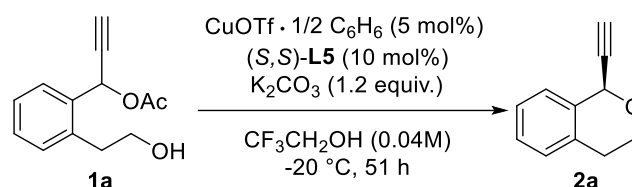
1-(5-chloro-2-(2-hydroxyethyl)phenyl)prop-2-yn-1-yl acetate (1l): a pale yellow oil. ^1H NMR δ 7.63 (d, J = 2.2 Hz, 1H), 7.28 (dd, J = 8.4 Hz, 2.2 Hz, 1H), 7.19 (d, J = 8.4 Hz, 1H), 6.59 (d, J = 2.2 Hz, 1H), 3.84 (t, J = 2.4 Hz, 2H), 3.02-2.90 (m, 2H), 2.67 (d, J = 2.2 Hz, 1H), 2.13 (s, 3H), 2.01 (br, 1H). ^{13}C NMR δ 169.6, 137.0, 135.0, 132.8, 131.8, 129.2, 128.1, 79.8, 75.9, 63.0, 62.3, 35.0,

20.9. HRMS (FAB) Calcd. for $C_{13}H_{13}ClO_3$ [M]: 252.0553. Found: 252.0557.



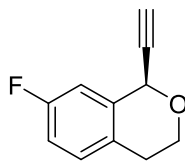
1-(2-(2-hydroxyethyl)naphthalen-1-yl)prop-2-yn-1-yl acetate (1m): a pale yellow oil. 1H NMR δ 8.66 (d, J = 8.8 Hz, 1H), 7.83 (d, J = 8.0 Hz, 1H), 7.79 (d, J = 8.8 Hz, 1H), 7.60-7.55 (m, 1H), 7.50-7.45 (m, 1H), 7.35-7.29 (m, 2H), 3.93 (t, J = 6.4 Hz, 2H), 3.44-3.39 (m, 1H), 3.20-3.10 (m, 1H), 2.67 (d, J = 2.4 Hz, 1H), 2.21 (br, 1H), 2.06 (s, 3H). ^{13}C NMR δ 169.8, 135.5, 133.2, 131.1, 130.0, 129.9, 128.52, 128.48, 126.2, 125.7, 125.4, 80.8, 75.8, 63.4, 61.2, 37.3, 20.8. HRMS (EI) Calcd. for $C_{17}H_{16}O_3$ [M]: 268.1099. Found: 268.1093.

Enantioselective Intramolecular Propargylic Etherification of Propargylic Acetates.



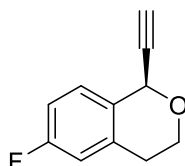
A typical experimental procedure for the reaction of 1-(2-(2-hydroxyethyl)phenyl)prop-2-yn-1-yl acetate (**1a**) is described below. In an oven dried 20 mL Schlenk flask were placed $CuOTf \cdot 1/2 C_6H_6$ (1.3 mg, 0.005 mmol) and (*S,S*)-Ph-pybox (**L5**) (3.7 mg, 0.01 mmol) under N_2 . Anhydrous methanol (1.0 mL) was added, and then the mixture was magnetically stirred at 60 °C for 1 h. After the solution was cooled to -20 °C, **1a** (21.8 mg, 0.10 mmol) in anhydrous methanol (1.5 mL) and K_2CO_3 (17 mg, 0.12 mmol) were added under N_2 , and the reaction mixture was kept at -20 °C for 51 h. The mixture was diluted by H_2O (10 mL) and extracted with CH_2Cl_2 (10 mL $\times$ 3). The combined organic layers were dried over anhydrous Na_2SO_4 and concentrated under reduced pressure *with caution*. The residue was purified by the column chromatography (SiO_2) with hexane/ Et_2O (50:1-40:1) to give 1-ethynylochromane (**2a**) as a pale yellow oil (14.1 mg, 0.089 mmol, 89% isolated yield). 1H NMR δ 7.27-7.18 (m, 3H), 7.15-7.10 (m, 1H), 5.55 (s, 1H), 4.26-4.20 (m, 1H), 4.01-3.95 (m, 1H), 2.94-2.80 (m, 2H), 2.56 (s, 1H). ^{13}C NMR δ 134.3, 132.7, 129.0, 127.3, 126.4, 125.8, 82.8, 73.8, 66.4, 62.4, 27.9. HRMS (EI) Calcd. for $C_{11}H_{10}O$ [M]: 158.0732. Found: 158.0731. $[\alpha]^{21}_D = +26.6$ (c = 0.50, $CHCl_3$). The enantiomeric excess of **2a** was determined by HPLC analysis; DAICEL Chiralpak OJ-H, hexane/*i*PrOH = 95/5, flow rate = 0.5 mL/min, λ = 220 nm, retention time: 18.6 min (major) and 26.0 min (minor), 93% ee.

Spectroscopic Data and Isolated Yields of Other Products.



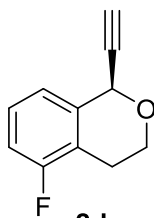
2b

1-ethynyl-7-fluoroisochromane (2b): Isolated yield 90%. A pale yellow oil. ^1H NMR δ 7.08 (dd J = 8.6 Hz, 6.0 Hz, 1H), 6.98 (dd J = 9.2 Hz, 2.4 Hz, 1H), 6.27 (td, J = 8.6 Hz, 2.4 Hz, 1H), 5.50 (s, 1H), 4.24-4.17 (m, 1H), 4.01-3.93 (m, 1H), 2.90-2.75 (m, 2H), 2.57 (d, J = 2.4 Hz, 1H). ^{13}C NMR δ 161.2 (d, $^1J_{\text{C-F}}$ = 246 Hz), 136.0 (d, $^3J_{\text{C-F}}$ = 6.7 Hz), 130.4 (d, $^3J_{\text{C-F}}$ = 6.6 Hz), 128.3 (d, $^4J_{\text{C-F}}$ = 2.9 Hz), 114.7 (d, $^2J_{\text{C-F}}$ = 21 Hz), 112.5 (d, $^2J_{\text{C-F}}$ = 23 Hz), 82.1, 74.2, 66.3, 62.6, 27.2. HRMS (EI) Calcd. for $\text{C}_{11}\text{H}_9\text{FO}$ [M]: 176.0637. Found: 176.0636. $[\alpha]^{22}_{\text{D}}$ = +49.6 (c = 0.32, CHCl_3). The enantiomeric excess of **2b** was determined by HPLC analysis; DAICEL Chiralpak OJ-H, hexane/*i*PrOH = 95/5, flow rate = 0.5 mL/min, λ = 220 nm, retention time: 17.3 min (major) and 18.5 min (minor), 87% ee.



2c

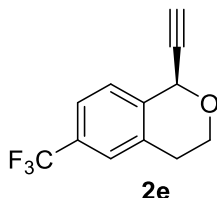
1-ethynyl-6-fluoroisochromane (2c): Isolated yield 75%. A pale yellow oil. ^1H NMR δ 7.22 (dd, J = 8.3 Hz, 5.2 Hz, 1H), 6.92 (td, J = 8.3 Hz, 2.6 Hz, 1H), 6.81 (dd, J = 9.6 Hz, 2.6 Hz, 1H), 5.50 (s, 1H), 4.24-4.17 (m, 1H), 4.00-3.93 (m, 1H), 2.95-2.77 (m, 2H), 2.57 (d, J = 2.0 Hz, 1H). ^{13}C NMR δ 161.8 (d, $^1J_{\text{C-F}}$ = 245 Hz), 135.0 (d, $^3J_{\text{C-F}}$ = 7.6 Hz), 130.0 (d, $^4J_{\text{C-F}}$ = 2.9 Hz), 127.5 (d, $^3J_{\text{C-F}}$ = 8.6 Hz), 115.3 (d, $^2J_{\text{C-F}}$ = 22 Hz), 113.7 (d, $^2J_{\text{C-F}}$ = 22 Hz), 82.6, 74.1, 66.1, 62.0, 28.0. HRMS (EI) Calcd. for $\text{C}_{11}\text{H}_9\text{FO}$ [M]: 176.0637. Found: 176.0630. $[\alpha]^{21}_{\text{D}}$ = +47.7 (c = 0.27, CHCl_3). The enantiomeric excess of **2c** was determined by HPLC analysis; DAICEL Chiralpak OZ-H, hexane/*i*PrOH = 95/5, flow rate = 0.5 mL/min, λ = 220 nm, retention time: 9.9 min (major) and 10.6 min (minor), 94% ee.



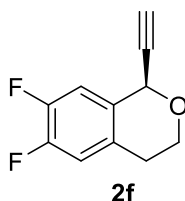
2d

1-ethynyl-5-fluoroisochromane (2d): Isolated yield 65%. A pale yellow oil. ^1H NMR δ 7.20 (dd J = 13.2 Hz, 8.1 Hz, 1H), 7.06 (d, J = 8.1 Hz, 1H), 6.95 (t, J = 8.1 Hz, 1H), 5.53 (s, 1H), 4.27-4.19 (m, 1H), 4.04-3.97 (m, 1H), 2.83 (t, J = 6.0, 2H), 2.58 (d, J = 2.8 Hz, 1H). ^{13}C NMR δ 160.3 (d, $^1J_{\text{C-F}}$ = 244 Hz), 136.4 (d, $^3J_{\text{C-F}}$ = 4.8 Hz), 127.2 (d, $^3J_{\text{C-F}}$ = 8.5 Hz), 121.2 (d, $^4J_{\text{C-F}}$ = 2.9 Hz), 120.7 (d, $^2J_{\text{C-F}}$ = 19 Hz), 113.6 (d, $^2J_{\text{C-F}}$ = 21 Hz), 82.2, 74.3, 65.9 (d, $^4J_{\text{C-F}}$ = 2.8 Hz), 61.6, 21.3 (d, $^3J_{\text{C-F}}$ =

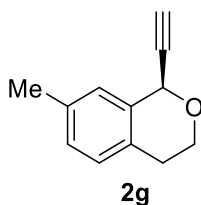
2.9 Hz). HRMS (EI) Calcd. for $C_{11}H_9FO$ [M]: 176.0637. Found: 176.0640. $[\alpha]^{22}_D = +34.1$ ($c = 0.57$, $CHCl_3$). The enantiomeric excess of **2d** was determined by HPLC analysis; DAICEL Chiralpak OJ-H, hexane/iPrOH = 95/5, flow rate = 0.5 mL/min, $\lambda = 220$ nm, retention time: 17.8 min (major) and 19.2 min (minor), 89% ee.



1-ethynyl-6-(trifluoromethyl)isochromane (2e): Isolated yield 87%. A pale yellow oil. 1H NMR δ 7.47 (d, $J = 7.6$ Hz, 1H), 7.40-7.36 (m, 2H), 5.57 (s, 1H), 4.28-4.21 (m, 1H), 4.05-3.98 (m, 1H), 3.02-2.83 (m, 2H), 2.59 (d, $J = 2.4$ Hz, 1H). ^{13}C NMR δ 138.2, 133.6, 129.7 (d, $^2J_{C-F} = 32$ Hz), 126.4, 126.0 (d, $^3J_{C-F} = 3.8$ Hz), 123.9 (d, $^1J_{C-F} = 272$ Hz), 123.2 (d, $^3J_{C-F} = 3.9$ Hz), 82.0, 74.6, 66.3, 62.0, 27.8. HRMS (EI) Calcd. for $C_{12}H_9F_3O$ [M]: 226.0605. Found: 226.0601. $[\alpha]^{22}_D = +27.5$ ($c = 0.50$, $CHCl_3$). The enantiomeric excess of **2e** was determined by HPLC analysis; DAICEL Chiralpak OZ-H, hexane/iPrOH = 95/5, flow rate = 0.5 mL/min, $\lambda = 220$ nm, retention time: 9.09 min (major) and 9.94 min (minor), 93% ee.

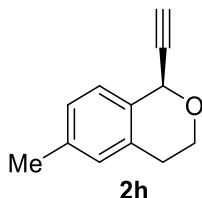


1-ethynyl-6,7-difluoroisochromane (2f): Isolated yield 51%. A pale yellow oil. 1H NMR δ 7.08 (dd, $J = 10.4$ Hz, 7.6 Hz, 1H), 6.93 (dd, $J = 10.4$ Hz, 7.6 Hz, 1H), 5.46 (s, 1H), 4.23-4.15 (m, 1H), 4.00-3.92 (m, 1H), 2.89-2.73 (m, 2H), 2.59 (d, $J = 2.4$ Hz, 1H). ^{13}C NMR δ 149.6 (dd, $^1J_{C-F} = 247$ Hz and $^2J_{C-F} = 12$ Hz), 148.9 (dd, $^1J_{C-F} = 247$ Hz and $^2J_{C-F} = 11$ Hz), 130.6 (t, $^3J_{C-F} = ^4J_{C-F} = 4.8$ Hz), 129.3 (t, $^3J_{C-F} = ^4J_{C-F} = 5.7$ Hz), 117.2 (d, $^2J = 17.1$ Hz), 114.6 (d, $^2J = 18.1$ Hz), 81.9, 74.5, 65.8, 62.1, 27.2. HRMS (EI) Calcd. for $C_{11}H_8F_2O$ [M]: 194.0543. Found: 194.0544. $[\alpha]^{20}_D = +34.5$ ($c = 0.35$, $CHCl_3$). The enantiomeric excess of **2f** was determined by HPLC analysis; DAICEL Chiralpak OJ-H, hexane/iPrOH = 95/5, flow rate = 0.5 mL/min, $\lambda = 220$ nm, retention time: 18.6 min (minor) and 19.6 min (major), 92% ee.

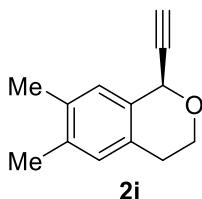


1-ethynyl-7-methylisochromane (2g): Isolated yield 75%. A pale yellow oil. 1H NMR δ 7.08-7.00 (m, 3H), 5.52 (d, $J = 2.0$ Hz, 1H), 4.24-4.19 (m, 1H), 4.01-3.94 (m, 1H), 2.89-2.75 (m, 2H), 2.56 (d, $J = 2.0$ Hz, 1H), 2.33 (s, 3H). ^{13}C NMR δ 136.0, 134.0, 129.6, 128.8, 128.2, 126.1, 83.0,

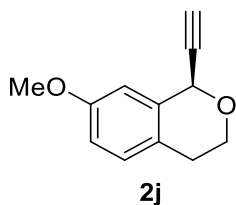
73.7, 66.4, 62.5, 27.5, 21.1. HRMS (EI) Calcd. for $C_{12}H_{12}O$ [M]: 172.0888. Found: 172.0894. $[\alpha]^{22}_D = +78.1$ ($c = 0.44$, $CHCl_3$). The enantiomeric excess of **2g** was determined by HPLC analysis; DAICEL Chiralpak OJ-H, hexane/iPrOH = 95/5, flow rate = 0.5 mL/min, $\lambda = 220$ nm, retention time: 14.6 min (major) and 17.9 min (minor), 88% ee.



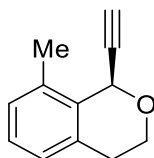
1-ethynyl-6-methylisochromane (2h): Isolated yield 90%. A pale yellow oil. 1H NMR δ 7.15 (d, $J = 7.5$ Hz, 1H), 7.03 (d, $J = 7.8$ Hz, 1H), 6.94 (s, 1H), 5.52 (s, 1H), 4.25-4.18 (m, 1H), 4.00-3.92 (m, 1H), 2.93-2.73 (m, 2H), 2.54 (d, $J = 2.4$ Hz, 1H), 2.32 (s, 3H). ^{13}C NMR δ 137.0, 132.5, 131.4, 129.4, 127.3, 125.7, 83.0, 73.7, 66.4, 62.4, 27.9, 21.1. HRMS (EI) Calcd. for $C_{12}H_{12}O$ [M]: 172.0888. Found: 172.0885. $[\alpha]^{21}_D = +42.0$ ($c = 0.37$, $CHCl_3$). The enantiomeric excess of **2h** was determined by HPLC analysis; DAICEL Chiralpak OZ-H, hexane/iPrOH = 95/5, flow rate = 0.5 mL/min, $\lambda = 220$ nm, retention time: 9.67 min (major) and 10.8 min (minor), 91% ee.



1-ethynyl-6,7-dimethylisochromane (2i): Isolated yield 66%. A pale yellow oil. 1H NMR δ 7.02 (s, 1H), 6.90 (s, 1H), 5.50 (s, 1H), 4.24-4.17 (m, 1H), 3.99-3.92 (m, 1H), 2.92-2.70 (m, 2H), 2.55 (d, $J = 2.4$ Hz, 1H), 2.24 (s, 3H), 2.23 (s, 3H). ^{13}C NMR δ 135.8, 134.8, 131.6, 130.0, 129.9, 126.6, 83.2, 73.6, 66.2, 62.5, 27.4, 19.42, 19.40. HRMS (EI) Calcd. for $C_{13}H_{14}O$ [M]: 186.1045. Found: 186.1051. $[\alpha]^{23}_D = +84.9$ ($c = 0.42$, $CHCl_3$). The enantiomeric excess of **2i** was determined by HPLC analysis; DAICEL Chiralpak OZ-H, hexane/iPrOH = 95/5, flow rate = 0.5 mL/min, $\lambda = 220$ nm, retention time: 9.41 min (major) and 10.6 min (minor), 87% ee.

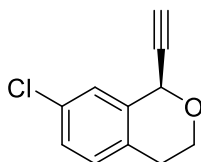


1-ethynyl-7-methoxyisochromane (2j): Isolated yield 72%. A pale yellow oil. 1H NMR δ 7.06-7.02 (m, 1H), 6.82-6.77 (m, 2H), 5.51 (s, 1H), 4.24-4.17 (m, 1H), 4.00-3.94 (m, 1H), 3.80 (s, 3H), 2.89-2.72 (m, 2H), 2.56 (d, $J = 2.0$ Hz, 1H). ^{13}C NMR δ 158.0, 135.2, 129.9, 124.7, 113.9, 110.4, 82.7, 73.8, 66.6, 62.7, 55.3, 27.1. HRMS (EI) Calcd. for $C_{12}H_{12}O_2$ [M]: 188.0837. Found: 188.0833. $[\alpha]^{22}_D = +45.0$ ($c = 0.47$, $CHCl_3$). The enantiomeric excess of **2j** was determined by HPLC analysis; DAICEL Chiralpak OZ-H, hexane/iPrOH = 95/5, flow rate = 0.5 mL/min, $\lambda = 220$ nm, retention time: 12.7 min (major) and 13.6 min (minor), 93% ee.



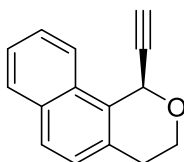
2k

1-ethynyl-8-methylisochromane (2k): Isolated yield 75%. A pale yellow oil. ^1H NMR δ 7.14 (t, $J = 7.3$ Hz, 1H), 7.02 (d, $J = 7.3$ Hz, 1H), 6.98 (d, $J = 7.3$ Hz, 1H), 5.50 (d, $J = 2.0$ Hz, 1H), 4.33-4.22 (m, 1H), 4.10-4.04 (m, 1H), 3.14-3.04 (m, 1H), 2.72-2.62 (m, 1H), 2.53 (d, $J = 2.0$ Hz, 1H), 2.33 (s, 3H). ^{13}C NMR δ 134.3, 132.7, 132.4, 128.3, 127.2, 126.9, 81.8, 74.1, 64.3, 60.7, 27.9, 18.4. HRMS (EI) Calcd. for $\text{C}_{12}\text{H}_{12}\text{O}$ [M]: 172.0888. Found: 172.0884. $[\alpha]^{20}_{\text{D}} = +18.6$ ($c = 0.60$, CHCl_3). The enantiomeric excess of **2k** was determined by HPLC analysis; DAICEL Chiralpak OJ-H, hexane/*i*PrOH = 95/5, flow rate = 0.5 mL/min, $\lambda = 220$ nm, retention time: 13.2 min (major) and 14.9 min (minor), 80% ee.



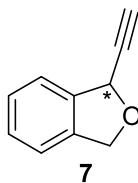
2l

7-chloro-1-ethynylisochromane (2l): Isolated yield 85%. A pale yellow oil. ^1H NMR δ 7.26 (s, 1H), 7.19 (dd, $J = 8.4$ Hz, 2.4 Hz, 1H), 7.06 (d, $J = 8.4$ Hz, 1H), 5.50 (s, 1H), 4.24-4.17 (m, 1H), 4.00-3.93 (m, 1H), 2.92-2.75 (m, 2H), 2.59 (d, $J = 2.4$ Hz, 1H). ^{13}C NMR δ 136.0, 131.9, 131.2, 130.3, 127.6, 125.8, 82.1, 74.4, 66.1, 62.3, 27.3. HRMS (EI) Calcd. for $\text{C}_{11}\text{H}_9\text{ClO}$ [M]: 192.0342. Found: 192.0346. $[\alpha]^{22}_{\text{D}} = +152.6$ ($c = 0.27$, CHCl_3). The enantiomeric excess of **2l** was determined by HPLC analysis; DAICEL Chiralpak OJ-H, hexane/*i*PrOH = 95/5, flow rate = 0.5 mL/min, $\lambda = 220$ nm, retention time: 17.7 min (major) and 20.4 min (minor), 80% ee.

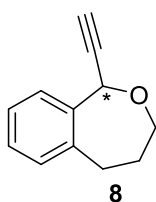


2m

1-ethynyl-3,4-dihydro-1H-benzo[h]isochromene (2m): Isolated yield 57%. A pale yellow oil. ^1H NMR δ 7.97 (d, $J = 8.4$, 1H), 7.82 (d, $J = 8.4$, 1H), 7.73 (d, $J = 8.6$, 1H), 7.58-7.52 (m, 1H), 7.47 (td, $J = 8.0$ Hz, 1.2 Hz, 1H), 7.23 (d, $J = 8.6$ Hz, 1H), 6.05 (s, 1H), 4.46-4.36 (m, 1H), 4.21-4.15 (m, 1H), 3.27-3.20 (m, 1H), 2.83-2.75 (m, 1H), 2.58 (d, $J = 2.0$ Hz, 1H). ^{13}C NMR δ 132.3, 130.4, 129.4, 128.9, 128.7, 127.9, 127.4, 126.5, 125.4, 122.4, 82.5, 74.7, 64.0, 60.7, 28.3. HRMS (EI) Calcd. for $\text{C}_{15}\text{H}_{12}\text{O}$ [M]: 208.0888. Found: 208.0882. $[\alpha]^{22}_{\text{D}} = +83.3$ ($c = 0.51$, CHCl_3). The enantiomeric excess of **2m** was determined by HPLC analysis; DAICEL Chiralpak OZ-H, hexane/*i*PrOH = 95/5, flow rate = 0.5 mL/min, $\lambda = 220$ nm, retention time: 9.96 min (major) and 11.8 min (minor), 83% ee.

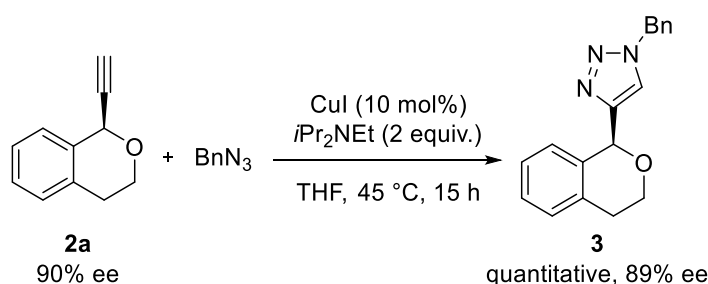


1-ethynyl-1,3-dihydroisobenzofuran (7): Isolated yield 36% (with **L2**). A pale yellow oil. ^1H NMR δ 7.39-7.31 (m, 3H), 7.27-7.22 (m, 1H), 5.88 (s, 1H), 5.26 (dd, $J = 12.2$ Hz, 2.0 Hz, 1H), 5.10 (d, $J = 12.2$, 1H), 2.60 (d, $J = 2.4$ Hz, 1H). ^{13}C NMR δ 138.8, 138.6, 128.3, 127.8, 121.7, 121.1, 82.0, 74.2, 73.1, 73.0. HRMS (EI) Calcd. for $\text{C}_{10}\text{H}_8\text{O}$ [M]: 144.0575. Found: 144.0582. $[\alpha]^{21}_{\text{D}} = +119.2$ ($c = 0.14$, CHCl_3). The enantiomeric excess of **7** was determined by HPLC analysis; DAICEL Chiralpak OJ-H, hexane/*i*PrOH = 95/5, flow rate = 0.5 mL/min, $\lambda = 220$ nm, retention time: 20.8 min (major) and 27.5 min (minor), 68% ee.



1-ethynyl-1,3,4,5-tetrahydrobenzo[c]oxepine (8): Isolated yield 40% (with **L5**). A pale yellow oil. ^1H NMR δ 7.57-7.49 (m, 1H), 7.24-7.15 (m, 3H), 5.50 (d, $J = 2.2$ Hz, 1H), 4.45-4.39 (m, 1H), 4.04-3.97 (m, 1H), 3.15-3.11 (m, 1H), 3.03-2.95 (m, 1H), 2.81 (d, $J = 2.2$ Hz, 1H), 1.89-1.79 (m, 2H). ^{13}C NMR δ 141.6, 138.7, 129.7, 128.4, 127.4, 126.3, 80.1, 77.5, 72.6, 72.4, 34.6, 29.8. HRMS (EI) Calcd. for $\text{C}_{12}\text{H}_{12}\text{O}$ [M]: 172.0888. Found: 172.0884. $[\alpha]^{22}_{\text{D}} = -2.40$ ($c = 0.15$, CHCl_3). The enantiomeric excess of **8** was determined by HPLC analysis; DAICEL Chiralpak OJ-H, hexane/*i*PrOH = 95/5, flow rate = 0.5 mL/min, $\lambda = 220$ nm, retention time: 43.1 min (minor) and 52.5 min (major), 29% ee.

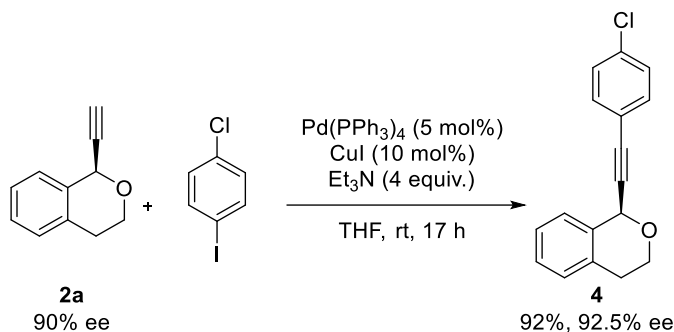
Preparation of (S)-1-benzyl-4-(isochroman-1-yl)-1H-1,2,3-triazole (**3**)



To a solution of **2a** (0.23 mmol) in anhydrous THF (4 mL) was added Benzyl azide (37.0 μL , 0.28 mmol), CuI (4.40 mg, 0.023 mmol), *i*Pr₂NEt (64.0 μL , 0.46 mmol). The mixture was stirred at 45 °C for 15 h. The mixture was concentrated under reduced pressure and purified by column chromatography with hexane/EtOAc (5:1-3:1) to give (S)-1-benzyl-4-(isochroman-1-yl)-1H-1,2,3-triazole **3** as a white solid (65.0 mg, 0.23 mmol, 98% isolated yield). ^1H NMR δ 7.36-7.00 (m, 10H), 6.05 (s, 1H), 5.51 (d, $J = 14.8$ Hz, 1H), 5.44 (d, $J = 14.8$ Hz, 1H), 4.16-4.10 (m, 1H), 3.96-3.88 (m, 1H), 3.05-2.96 (m, 1H), 2.84-2.76 (m, 1H). ^{13}C NMR δ 150.0, 135.7, 134.4, 133.4, 129.0, 128.8, 128.7, 128.1, 126.9, 126.3, 126.1, 122.0, 71.7, 63.6, 54.2, 28.5. HRMS (EI) Calcd.

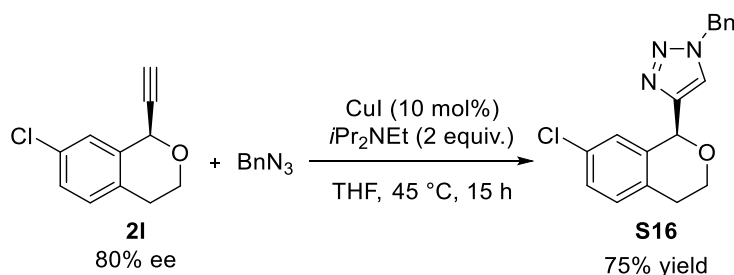
for C₁₈H₁₇N₃O [M]: 291.1372. Found: 291.1383. [α]_D²⁴ = +71.3 (c = 0.50, CHCl₃). The enantiomeric excess of **3** was determined by HPLC analysis; DAICEL Chiralpak OJ-H, hexane/iPrOH = 70/30, flow rate = 0.5 mL/min, λ = 220 nm, retention time: 28.6 min (major) and 52.7 min (minor), 89% ee.

Preparation of (*R*)-1-((4-chlorophenyl)ethynyl)isochromane (**4**)



To a solution of **2a** (0.17 mmol) in anhydrous THF (4 mL) was added 1-chloro-4-iodobenzene (80.0 mg, 0.34 mmol), Pd(PPh₃)₄ (9.80 mg, 0.0085 mmol), CuI (3.20 mg, 0.017 mmol), Et₃N (95.0 μ L, 0.68 mmol) at room temperature and the mixture was stirred for 17 h. The mixture was concentrated under reduced pressure and purified by column chromatography with hexane/EtOAc (100:0-25:1) to give (*R*)-1-((4-chlorophenyl)ethynyl)isochromane **4** as a yellow oil (42 mg, 0.156 mmol, 92% isolated yield). ¹H NMR δ 7.39-7.10 (m, 8H), 5.76 (s, 1H), 4.32-4.24 (m, 1H), 4.05-3.98 (m, 1H), 2.98-2.83 (m, 2H). ¹³C NMR δ 134.6, 134.5, 133.1, 132.8, 129.0, 128.6, 127.3, 126.4, 125.9, 121.0, 89.1, 84.5, 67.2, 62.7, 28.0. HRMS (EI) Calcd. for C₁₇H₁₃ClO [M]: 268.0655. Found: 268.0648. [α]_D²⁴ = +1.43 (c = 0.55, CHCl₃). The enantiomeric excess of **4** was determined by HPLC analysis; DAICEL Chiralpak OJ-H, hexane/iPrOH = 95/5, flow rate = 0.5 mL/min, λ = 220 nm, retention time: 24.1 min (minor) and 30.8 min (major), 92.5% ee.

Preparation of (*S*)-1-benzyl-4-(7-chloroisochroman-1-yl)-1H-1,2,3-triazole (**S16**)



(*S*)-1-benzyl-4-(7-chloroisochroman-1-yl)-1H-1,2,3-triazole **S16** was synthesized from **2l** (0.6 mmol) as a similar method as the synthesis of **3** from **2a**. Recrystallization from CH₂Cl₂ gave crystals of **S16** suitable for X-ray analysis (0.45 mmol, 75% yield). A white solid. ¹H NMR δ 7.39-7.32 (m, 4H), 7.29-7.23 (m, 2H), 7.14 (dd, J = 8.0 Hz, 1.6 Hz, 1H), 7.09-7.03 (m, 2H), 5.98 (s, 1H), 5.56 (d, J = 15.0 Hz, 1H), 5.47 (d, J = 15.0 Hz, 1H), 4.18-4.10 (m, 1H), 3.94-3.86 (m, 1H), 3.02-2.92 (m, 1H), 2.82-2.72 (m, 1H). ¹³C NMR δ 149.2, 137.4, 134.3, 131.8, 131.7, 130.2, 129.1, 128.8, 128.1, 127.2, 126.3, 122.0, 71.4, 63.5, 54.2, 27.9. HRMS (EI) Calcd. for C₁₈H₁₆N₃O [M]: 325.0982. Found: 325.0979.

X-ray diffraction studies of **S16**

Diffraction data for (S)-1-benzyl-4-(7-chloroisochroman-1-yl)-1H-1,2,3-triazole (**S16**, CCDC 1910939) were collected for the 2θ range of 4 to 55° at –100 °C on a Rigaku R-Axis RAPID imaging plate diffractometer with graphite-monochromated Mo-K α radiation (λ = 0.71075 Å) with VariMax optics. Intensity data were corrected for Lorentz and polarization effects and for empirical absorptions (ABSCOR),^{S6} whereas structure solutions and refinements were carried out by using *CrystalStructure* package.^{S7} Positions of non-hydrogen atoms were determined by direct methods (SHELXD Version 2013/2)^{S8} and subsequent Fourier syntheses SHELXL Version 2016/6,^{S9} and were refined on F_o^2 with all the unique reflections by full-matrix least squares with anisotropic thermal parameters. All the hydrogen atoms were placed at the calculated positions with fixed isotropic parameters. Anomalous dispersion effects were included in F_c ,^{S10} and mass attenuation coefficients, values for $\Delta f'$ and $\Delta f''$, and neutral atom scattering factors were taken from references.^{S11–S13} Refinement of the Flack parameter (0.09(3)) using 1202 Parsons' quotients demonstrates that the absolute configuration of **S16** is (S).^{S14} Details of the crystal and data collection parameters of **S16** are summarized in **Table S1**. ORTEP drawing of **S16** is shown in **Figure S1**.

Table S1. Crystallographic Data for **S16**.

compound	S16
CCDC number	1910939
chemical formula	C ₁₈ H ₁₆ ClN ₃ O
formula weight	325.80
crystal size (mm ³)	0.48 × 0.11 × 0.07
color, habit	colorless, needle
temperature (°C)	−100
crystal system	monoclinic
space group	<i>P</i> 2 ₁ (no. 4)
<i>a</i> (Å)	12.1916(6)
<i>b</i> (Å)	5.5082(3)
<i>c</i> (Å)	12.3900(6)
α (deg)	90
β (deg)	102.989(7)
γ (deg)	90
<i>V</i> (Å ³)	810.74(8)
<i>Z</i>	2
<i>d</i> _{calcd} (g cm ^{−3})	1.334
<i>F</i> (000)	340
μ (cm ^{−1})	2.430
transmission factors range	0.840 – 0.983
number of measured reflections	7587
number of unique reflections	3593
<i>R</i> _{int}	0.0258
number of refined parameters	208
<i>R</i> 1 [<i>I</i> > 2σ(<i>I</i>)] ^a	0.0485
<i>wR</i> 2 (all data) ^b	0.0965
GOF ^c	1.000
maximum and minimum residual peaks (e Å ^{−3})	+0.55 / −0.44 0.09(3)
Flack parameter	

^a $R1 = \sum ||F_o| - |F_c|| / \sum |F_o|$. ^b $wR2 = [\sum \{w(F_o^2 - F_c^2)^2\} / \sum w(F_o^2)^2]^{1/2}$, $w = 1/[\sigma^2(F_o^2) + rP]$, $P = (\text{Max}(F_o^2, 0) + 2 F_c^2)/3$ [$r = 0.5080$]. ^c $GOF = [\sum w(F_o^2 - F_c^2)^2 / (N_o - N_{\text{params}})]^{1/2}$.

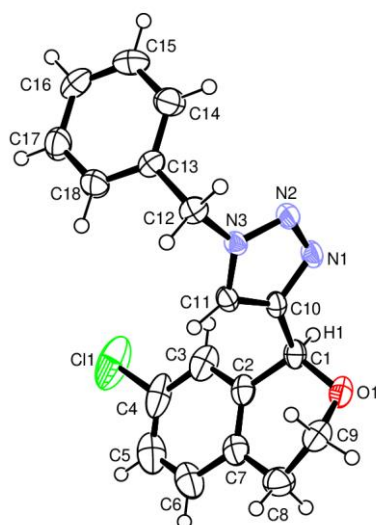
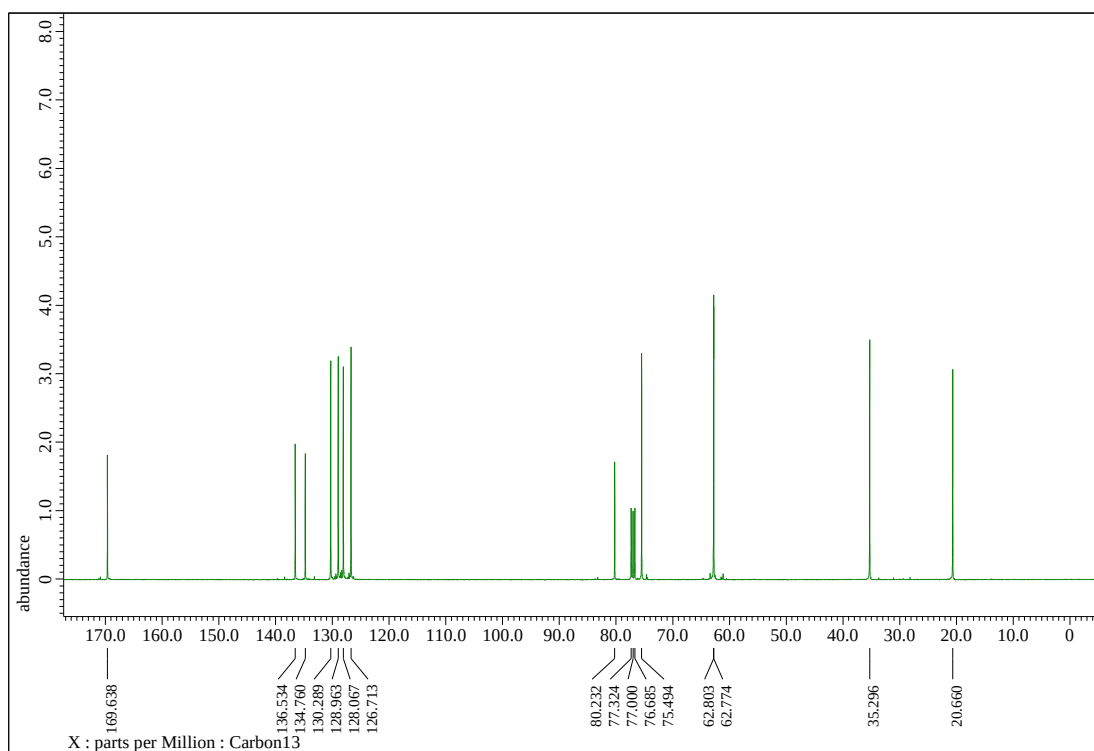
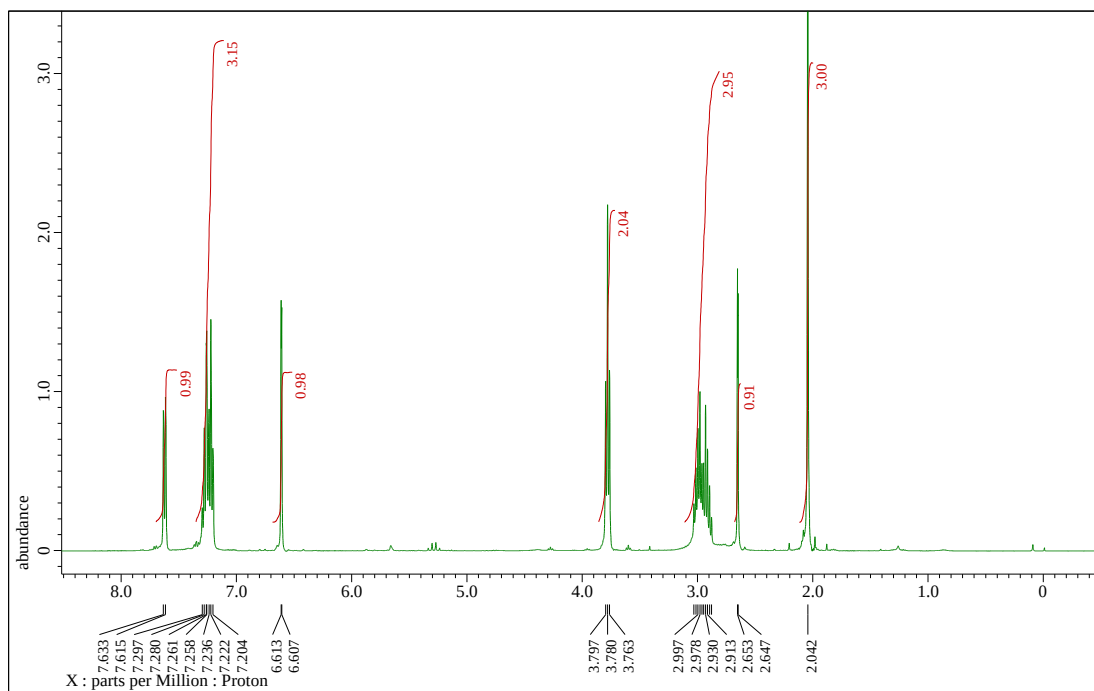
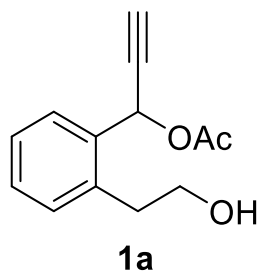


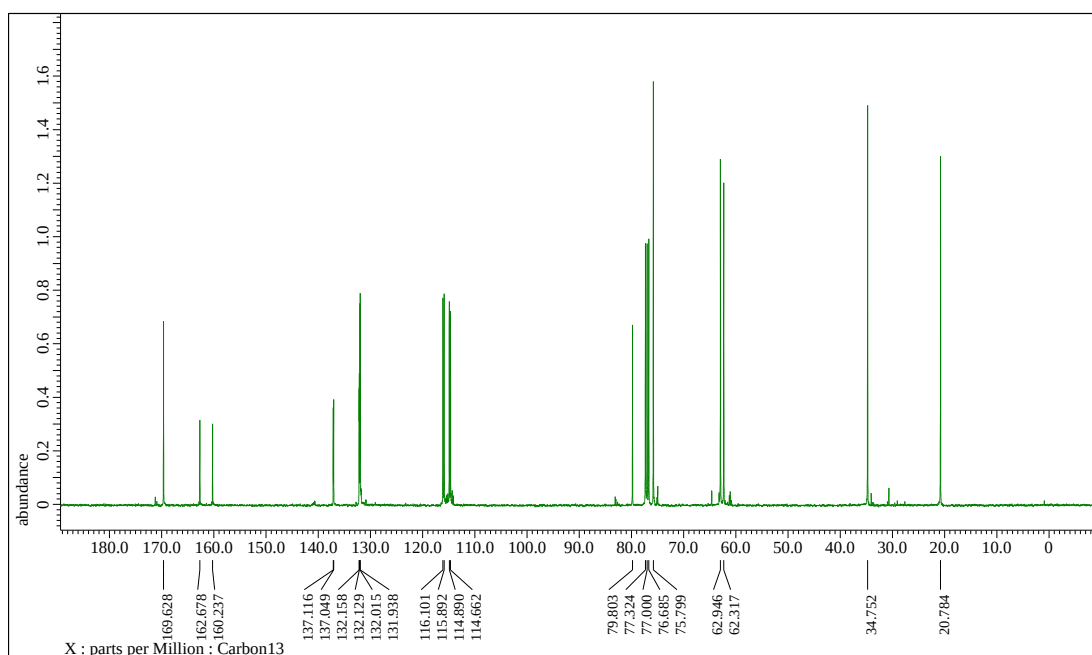
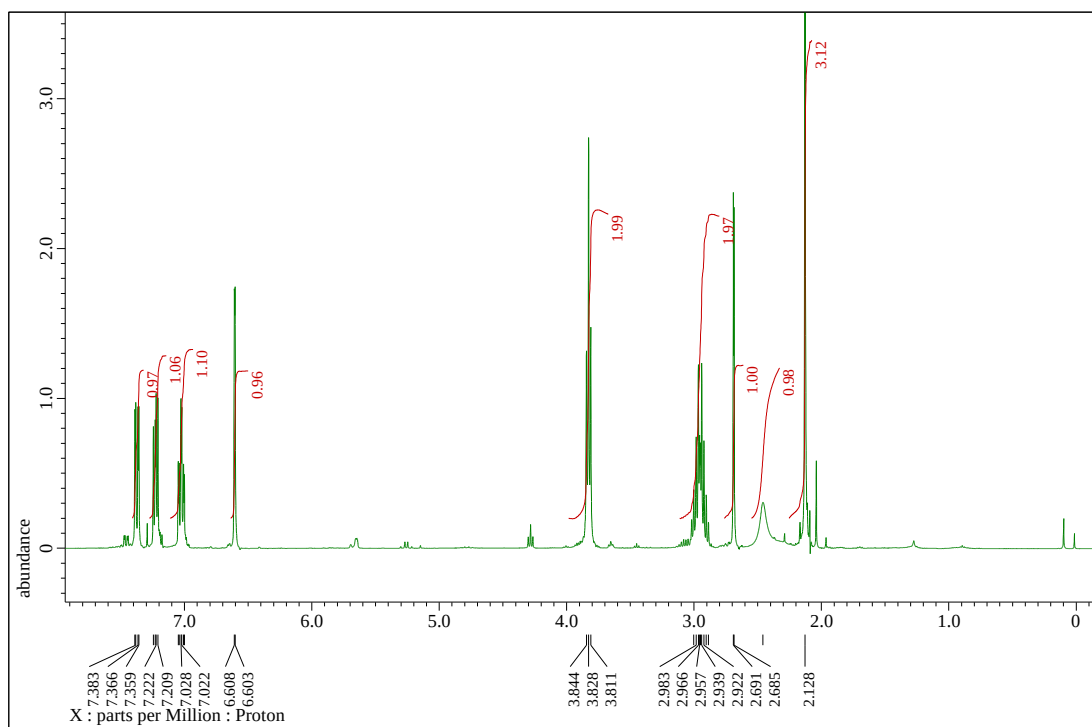
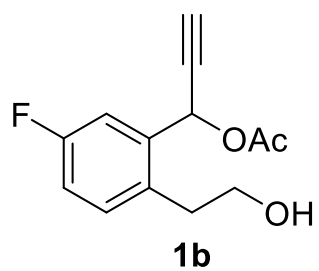
Figure S1. ORTEP drawing of **S16**. Thermal ellipsoids are given at the 50% probability level. Hydrogen atom labels except for H1 are omitted for clarity.

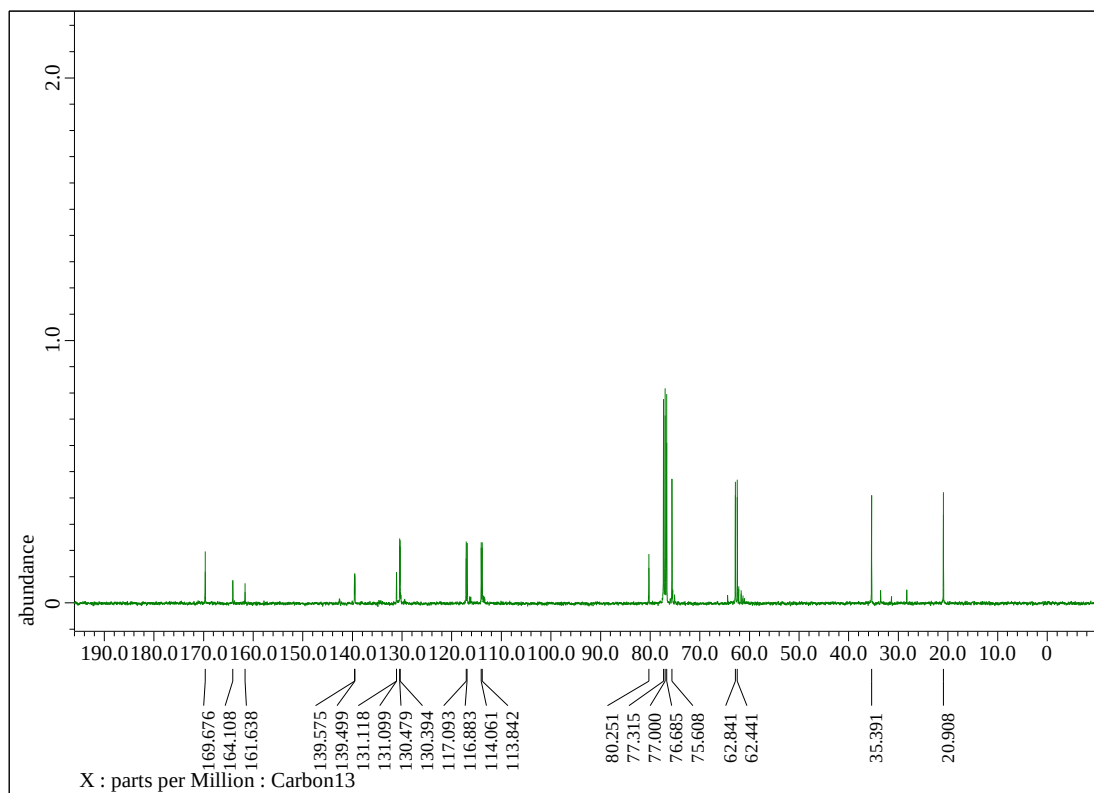
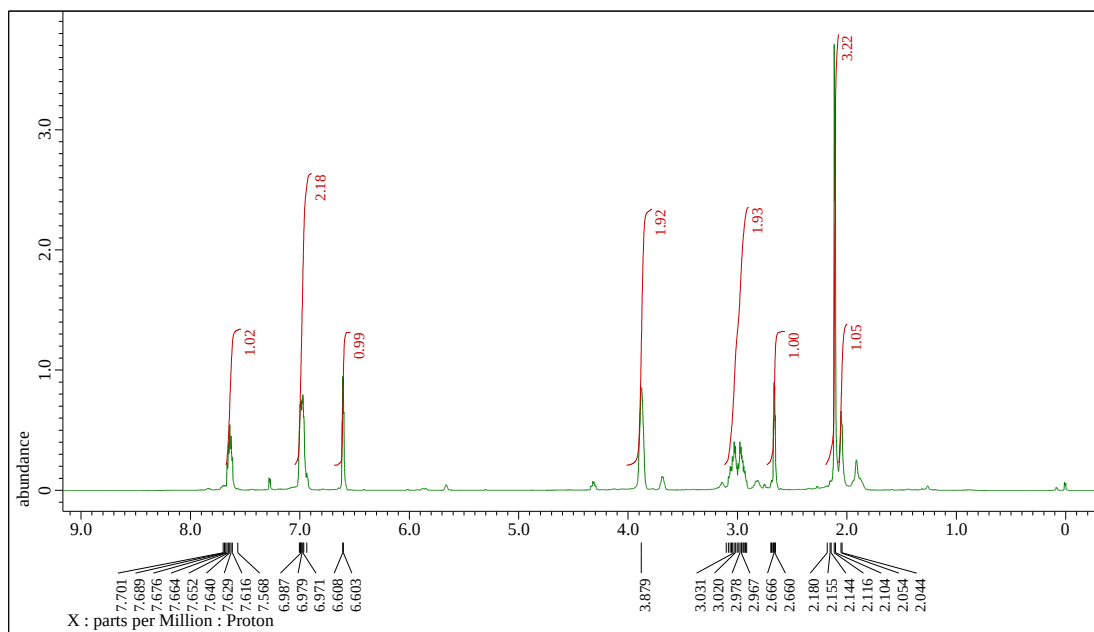
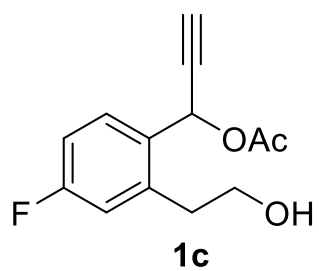
References

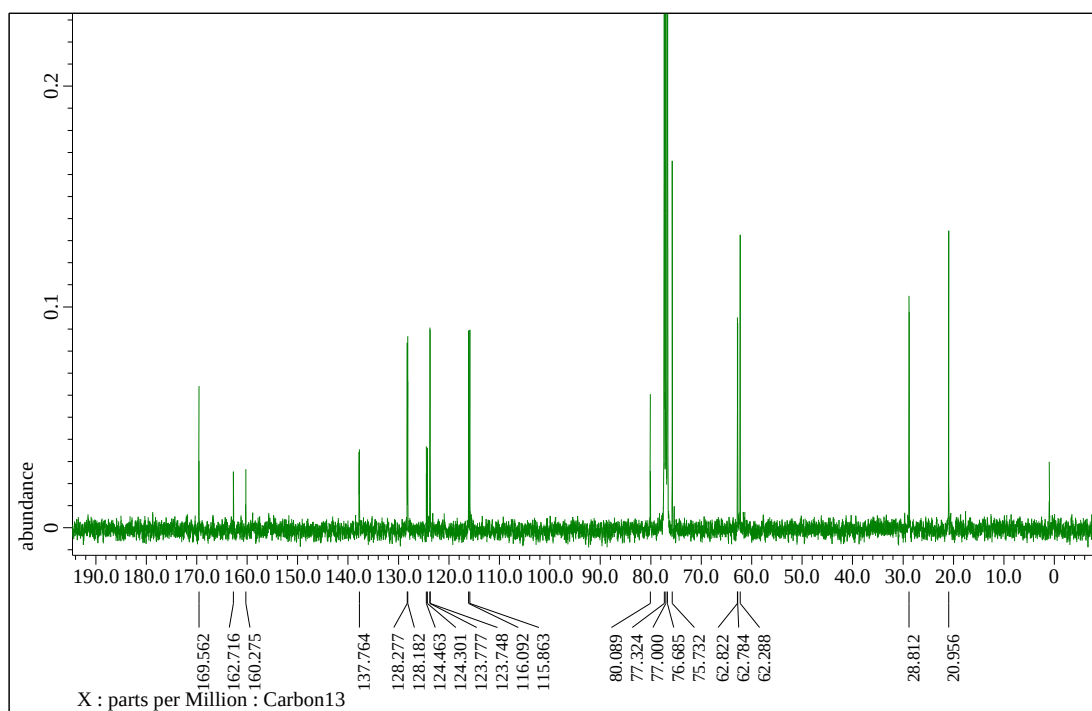
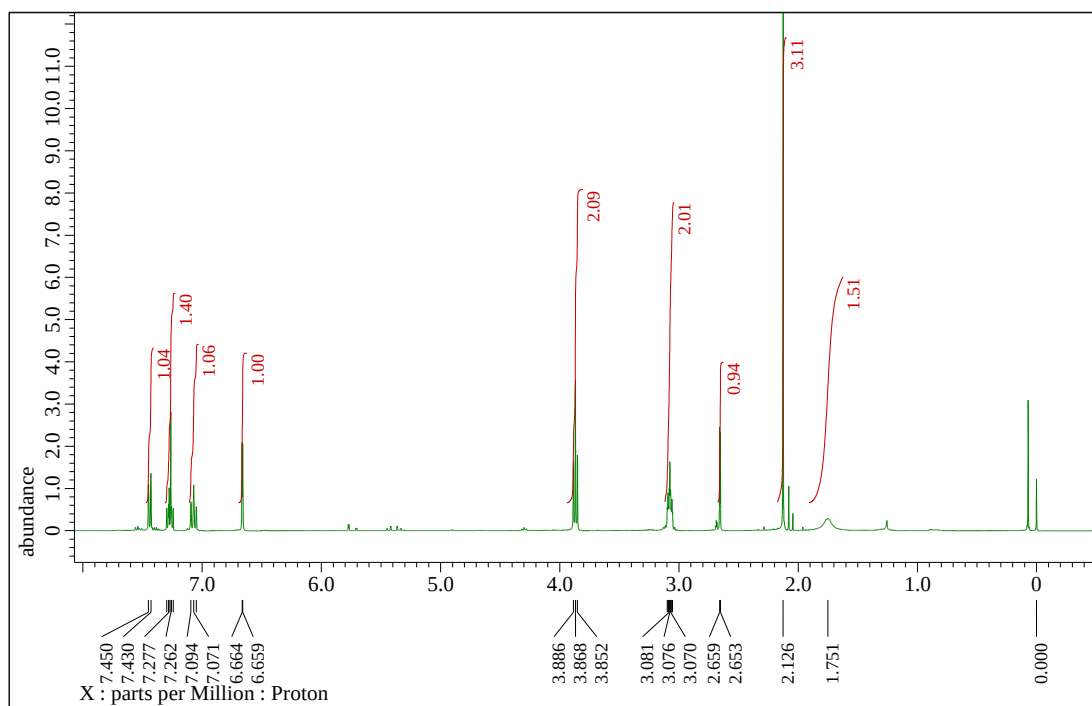
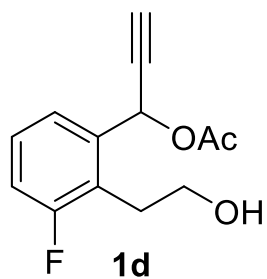
- S1) M. K. Tse; S. Bhor; M. Klawonn; G. Anilkumar; H. Jiao; C. Döbler; A. Spannenberg; W. Mägerlein; H. Hugel and M. Beller, *Chem. Eur. J.*, 2006, **12**, 1855-1874.
- S2) R. L. Espelt; I. S. McPherson; E. M. Wiensch and T. P. Yoon, *J. Am. Chem. Soc.*, 2015, **137**, 2452-2455.
- S3) Y. -Y. Zhu; C. Cui; N. Li; B. -W Wang; Z. -M. Wang and S. Gao, *Eur. J. Inorg. Chem.*, 2013, **17**, 3101-3111.
- S4) Y. Zhang; D. Guo; S. Ye; Z. Liu and G. Zhu, *Org. Lett.*, 2017, **19**, 1302-1305.
- S5) R. Dorel; P. R. McGonigal; A. M. Echavarren, *Angew. Chem., Int. Ed.*, 2016, **55**, 11120-11123.
- S6) T. Higashi, ABSCOR, Rigaku Corporation, Tokyo, Japan, 1995.
- S7) CrystalStructure 4.3: Crystal Structure Analysis Package, Rigaku Corporation, Tokyo, Japan, 2001–2018.
- S8) G. M. Sheldrick, *Acta Crystallogr., Sect. A: Found. Crystallogr.*, 2008, **A64**, 112–122.
- S9) G. M. Sheldrick, *Acta Crystallogr., Sect. C: Struct. Chem.*, 2015, **C71**, 3–8.
- S10) J. A. Ibers and W. C. Hamilton, *Acta Crystallogr.*, 1964, **17**, 781–782.
- S11) W. C. Creagh and J. H. Hubbell, in *International Tables for Crystallography*, ed. A. J. C. Wilson, Kluwer Academic Publishers, Dordrecht, 1992, vol. C, Table 4.2.4.3., pp. 200–206.
- S12) W. C. Creagh and W. J. McAuley, in *International Tables for Crystallography*, ed. A. J. C. Wilson, Kluwer Academic Publishers, Dordrecht, 1992, vol. C, Table 4.2.6.8., pp. 219–222.
- S13) E. N. Maslen, A. G. Fox and M. A. O’Keefe, in *International Tables for Crystallography*, ed. A. J. C. Wilson, Kluwer Academic Publishers, Dordrecht, 1992, vol. C, Table 6.1.1.4., pp. 500–503.
- S14) S. Parsons, H. D. Flack, T. Wagner, *Acta Crystallogr., Sect. B: Struct. Sci., Cryst. Eng. Mater.*, 2013, **B69**, 249–259.

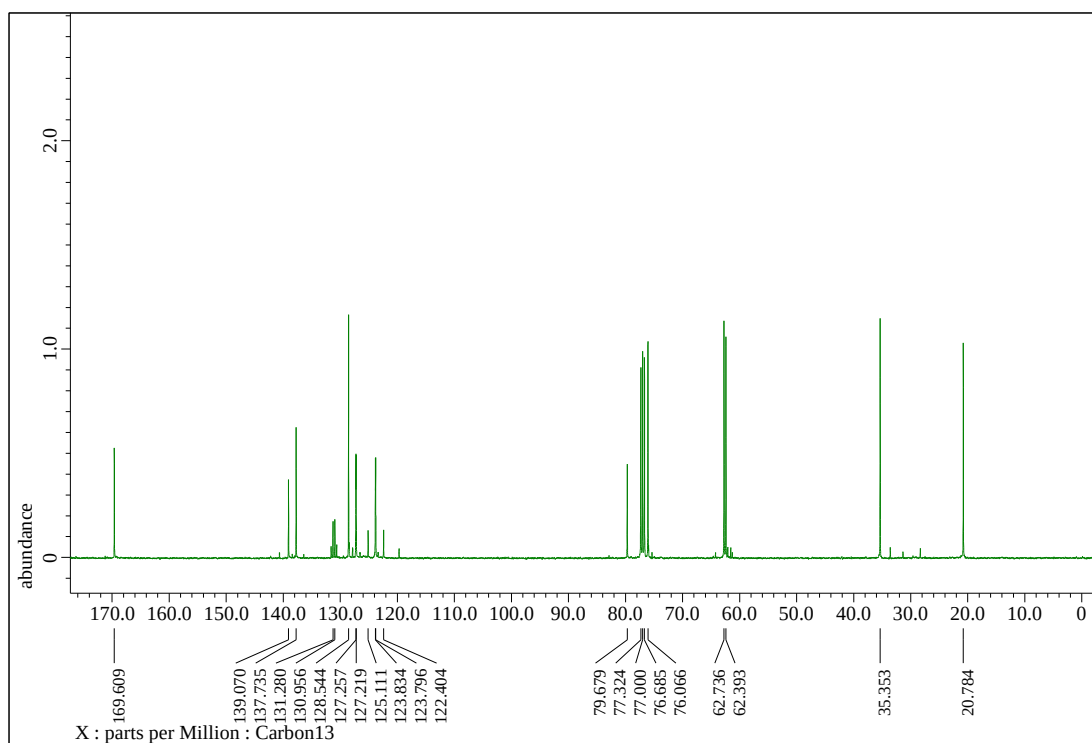
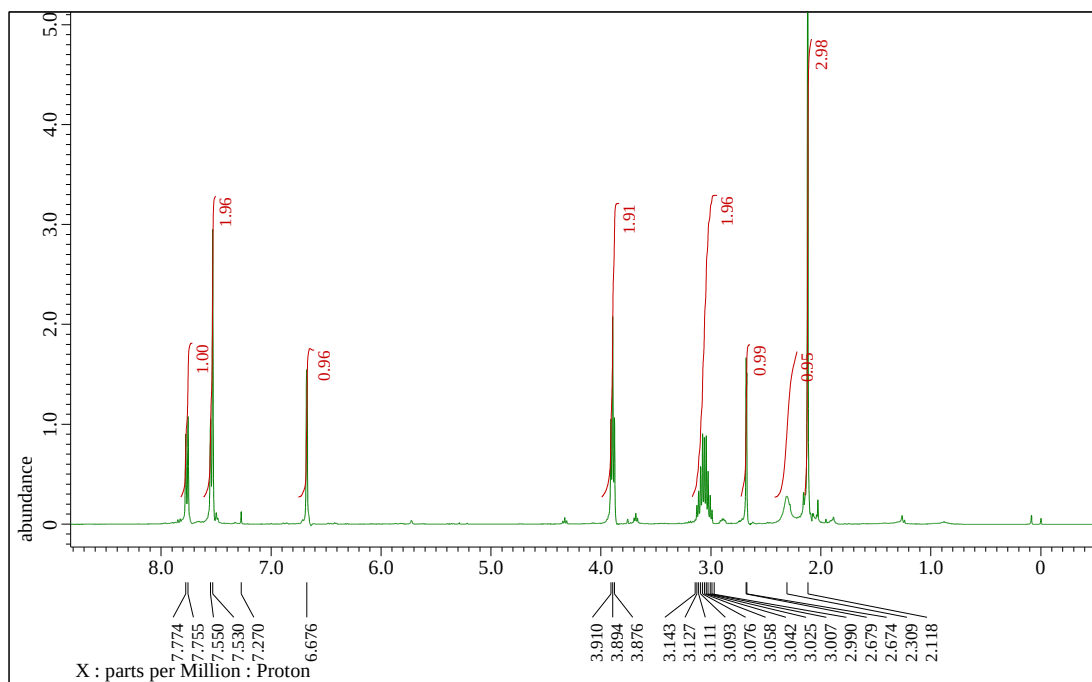
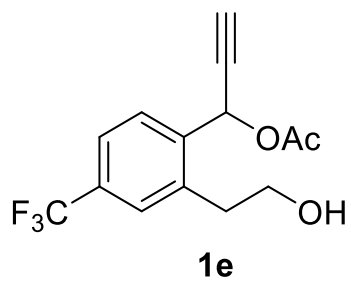
^1H and ^{13}C NMR Spectra

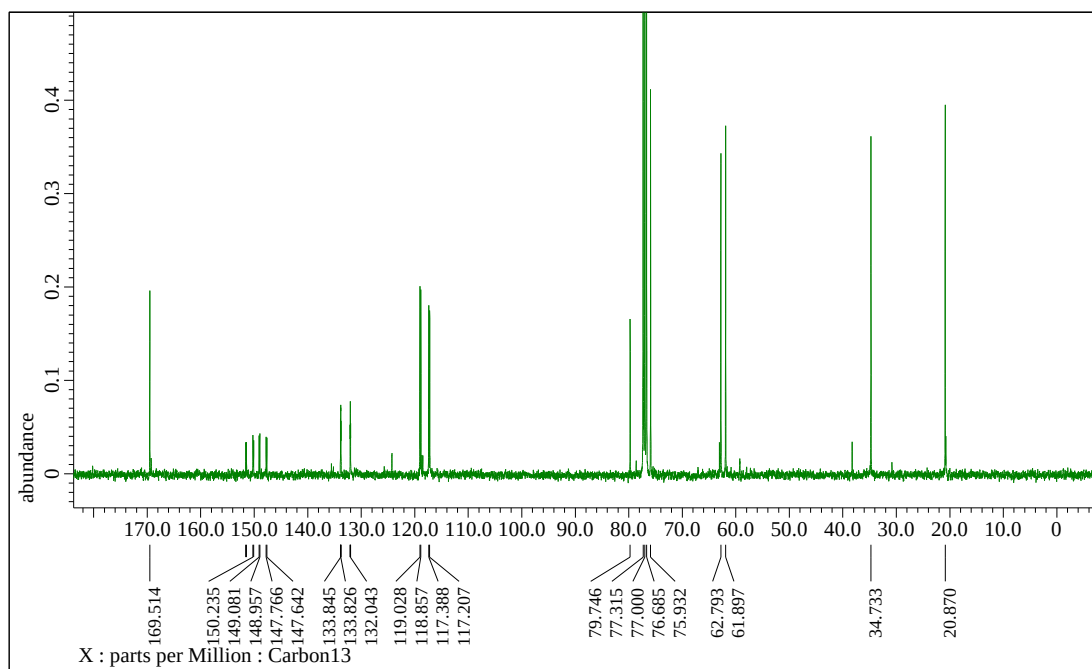
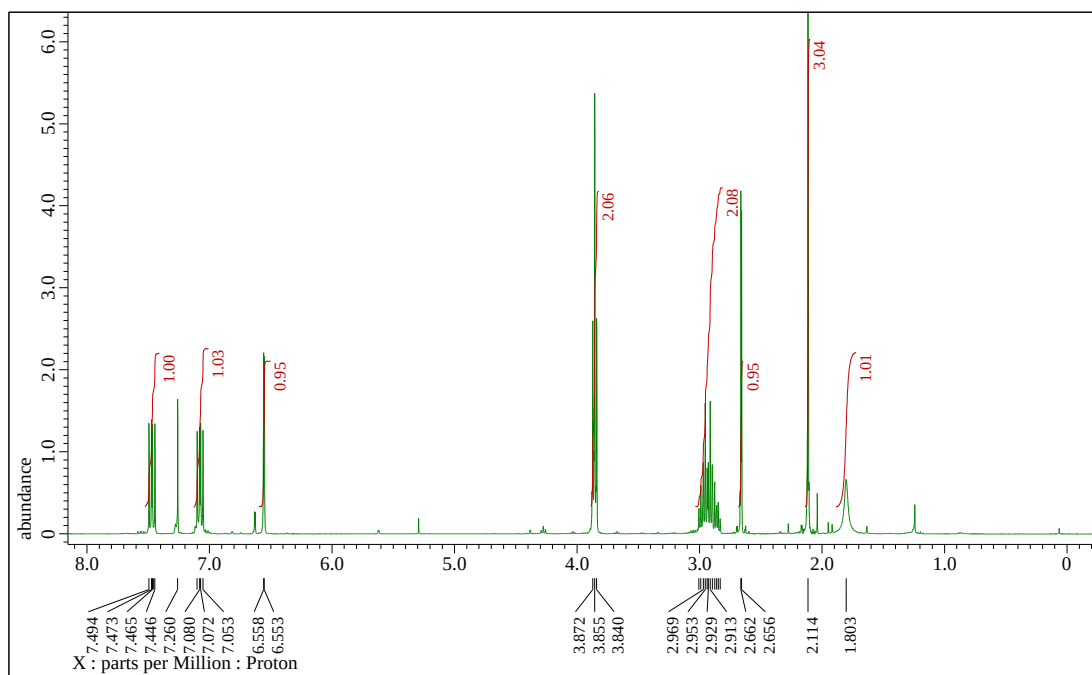
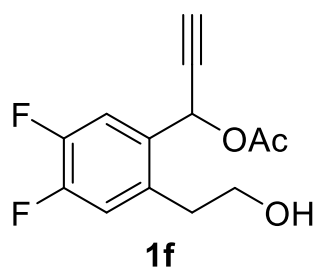


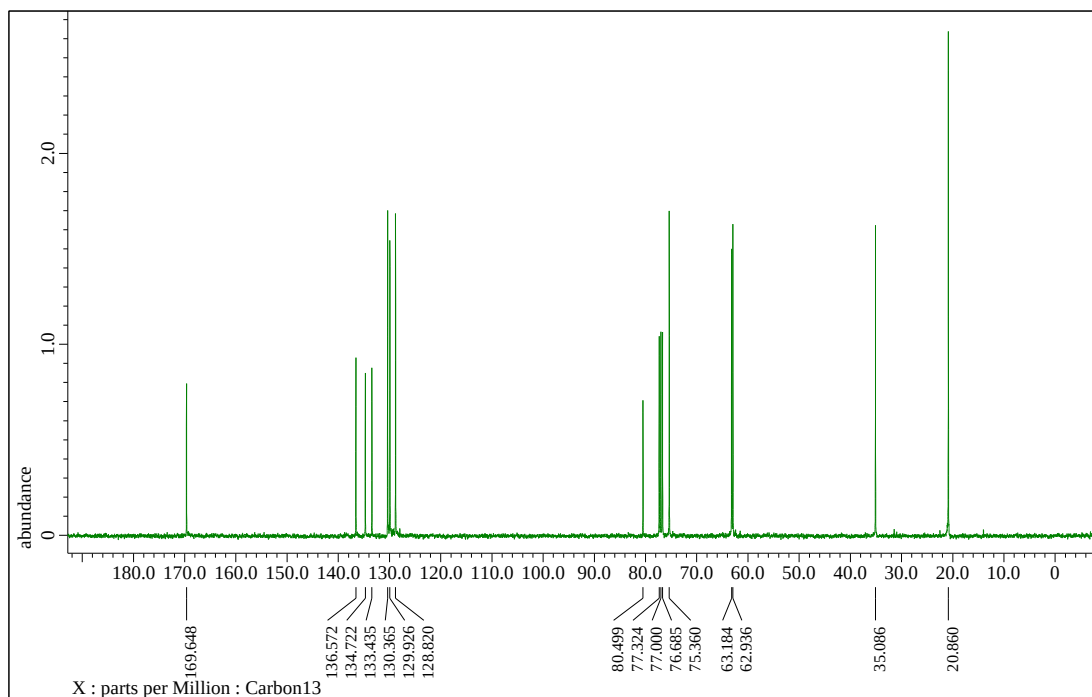
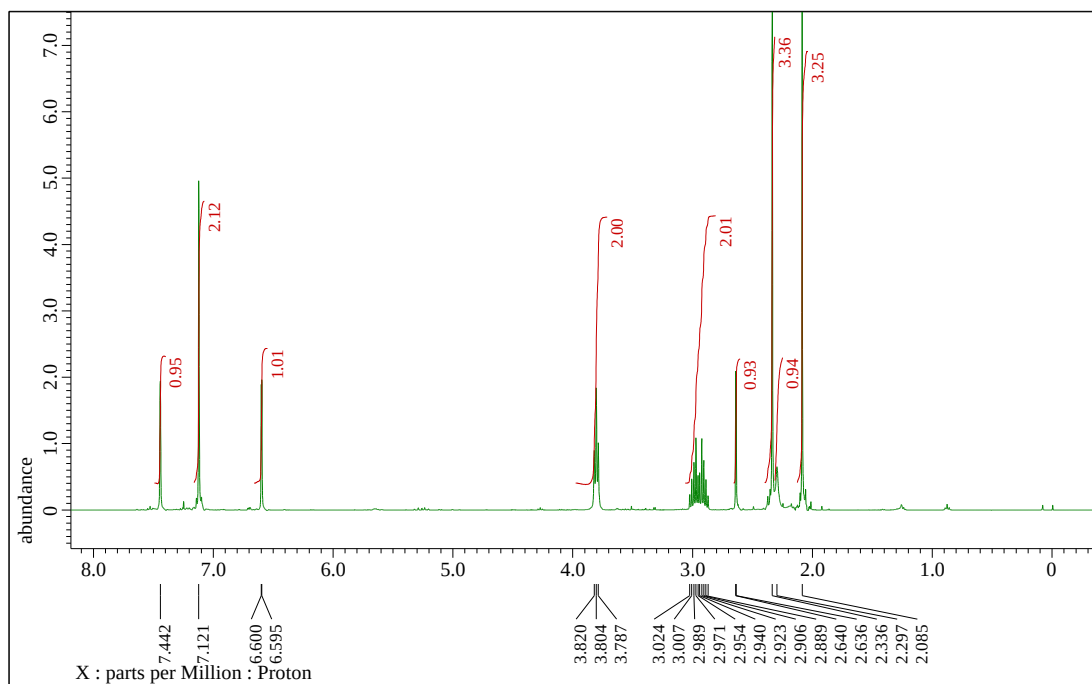
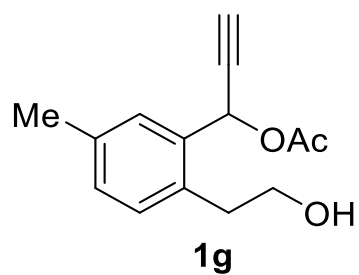


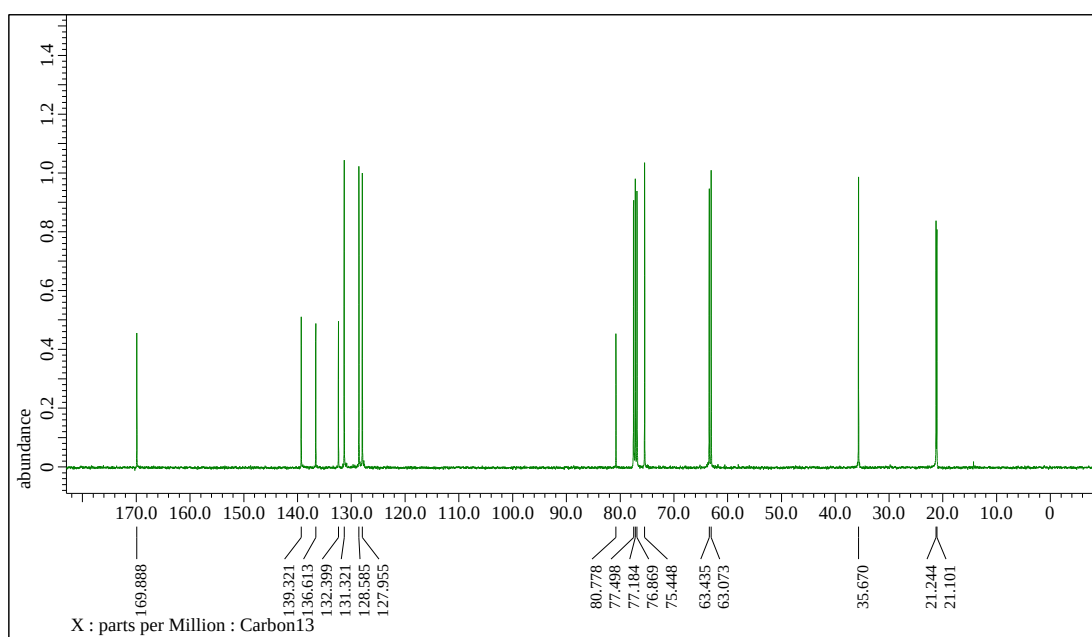
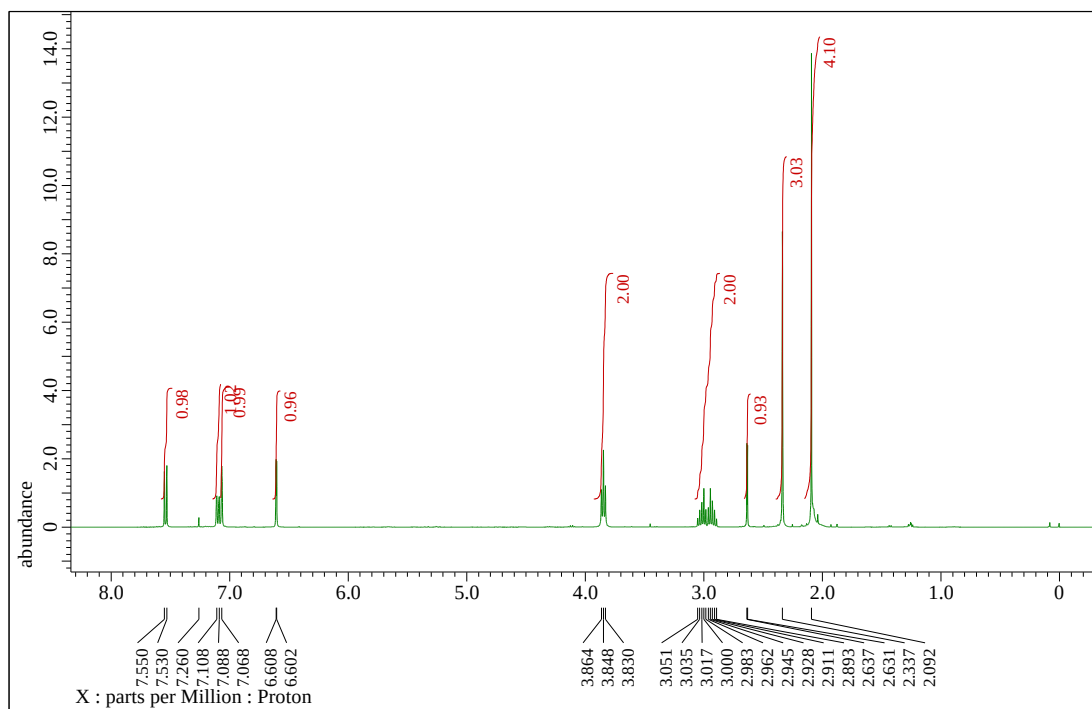
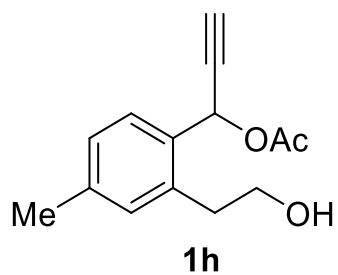


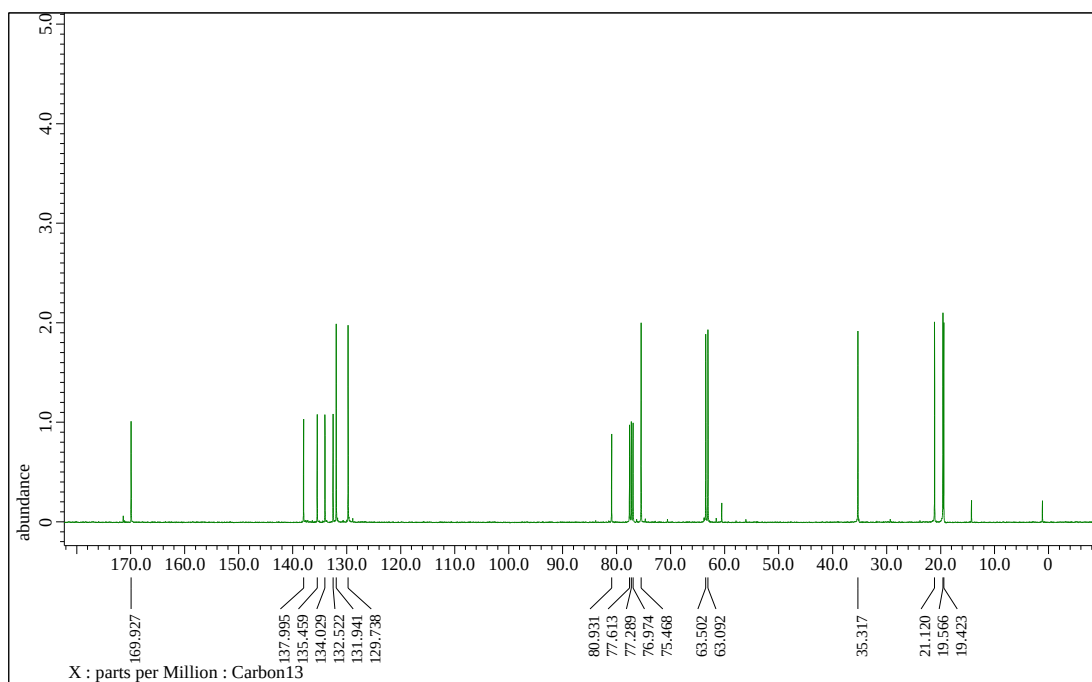
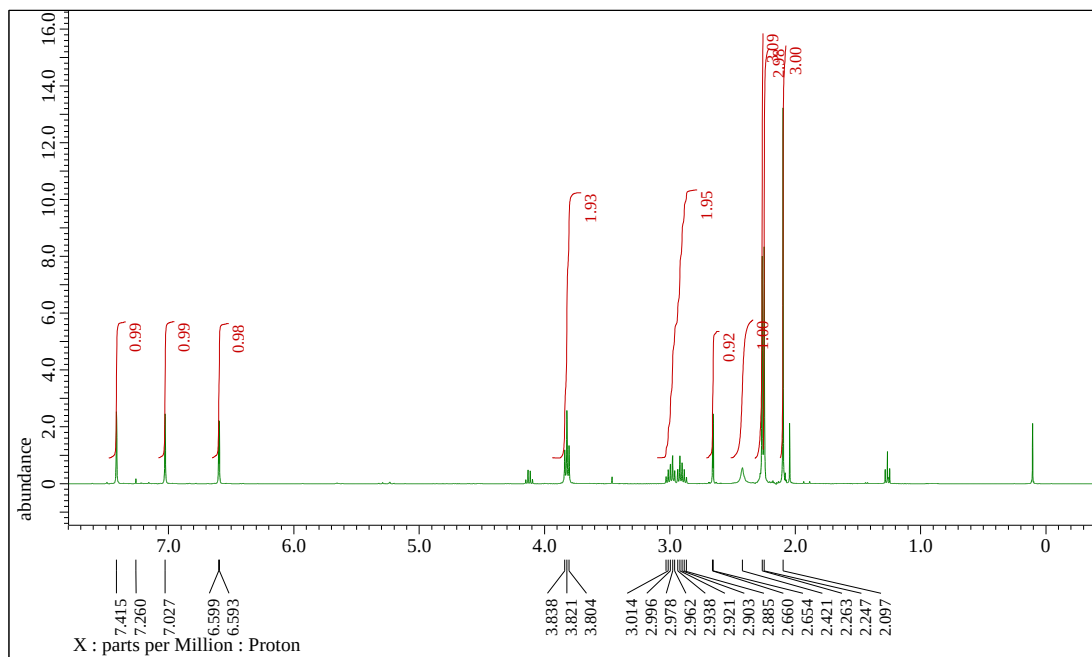
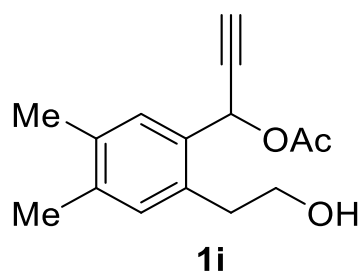


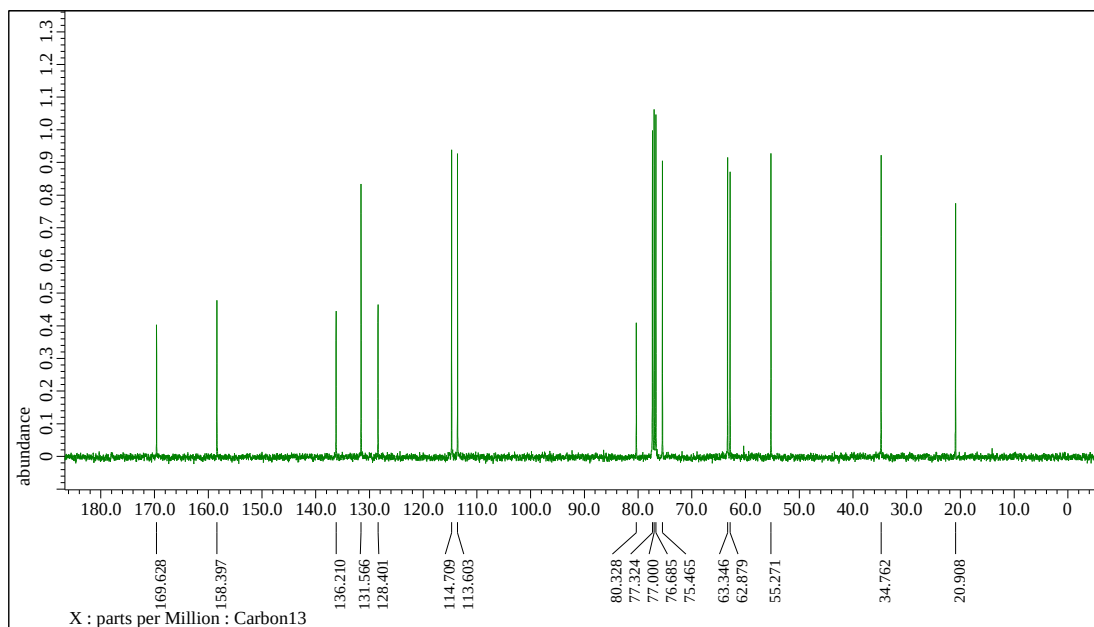
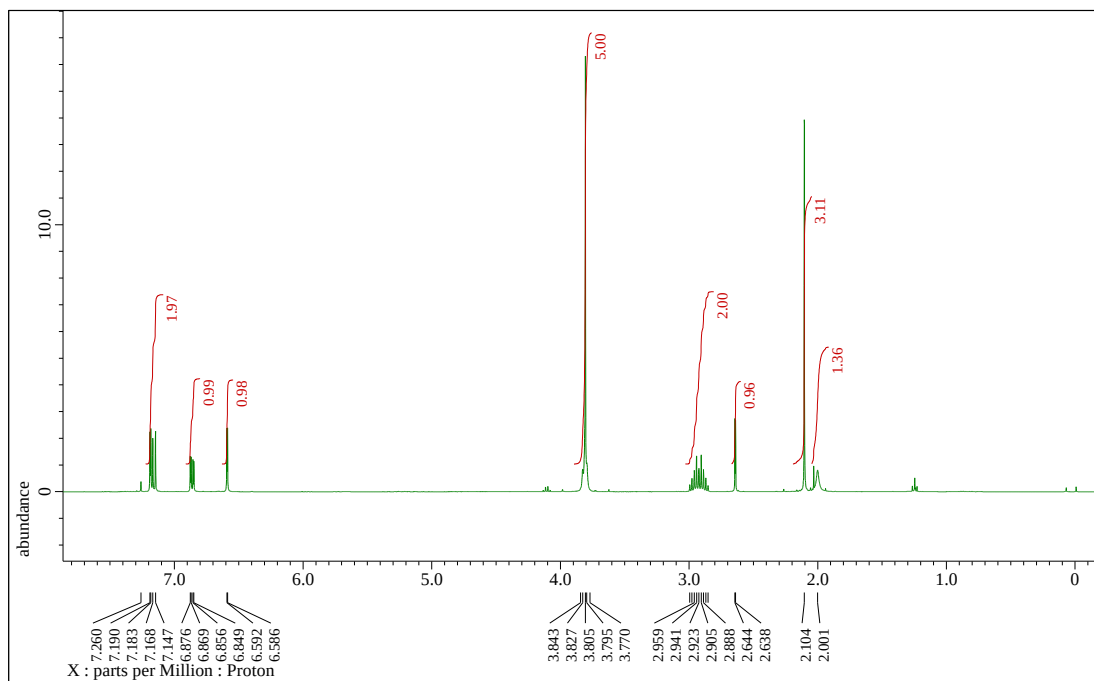
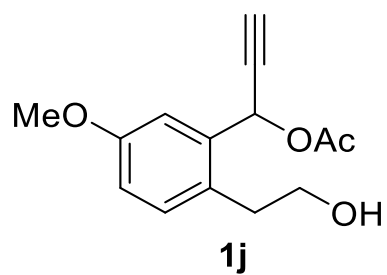


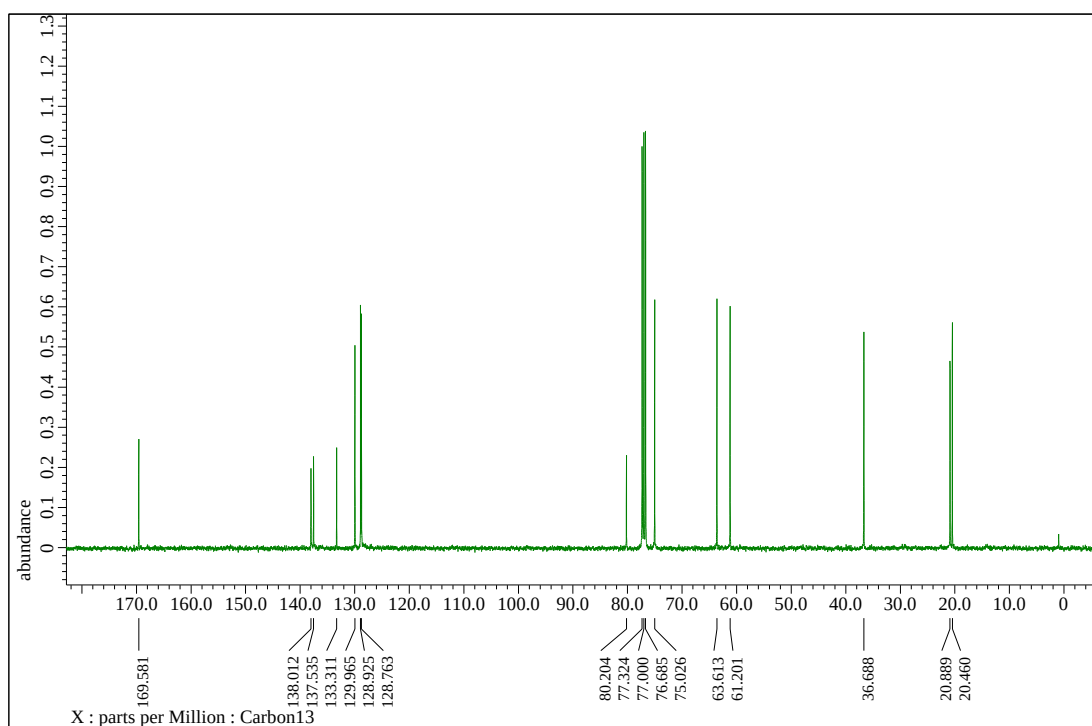
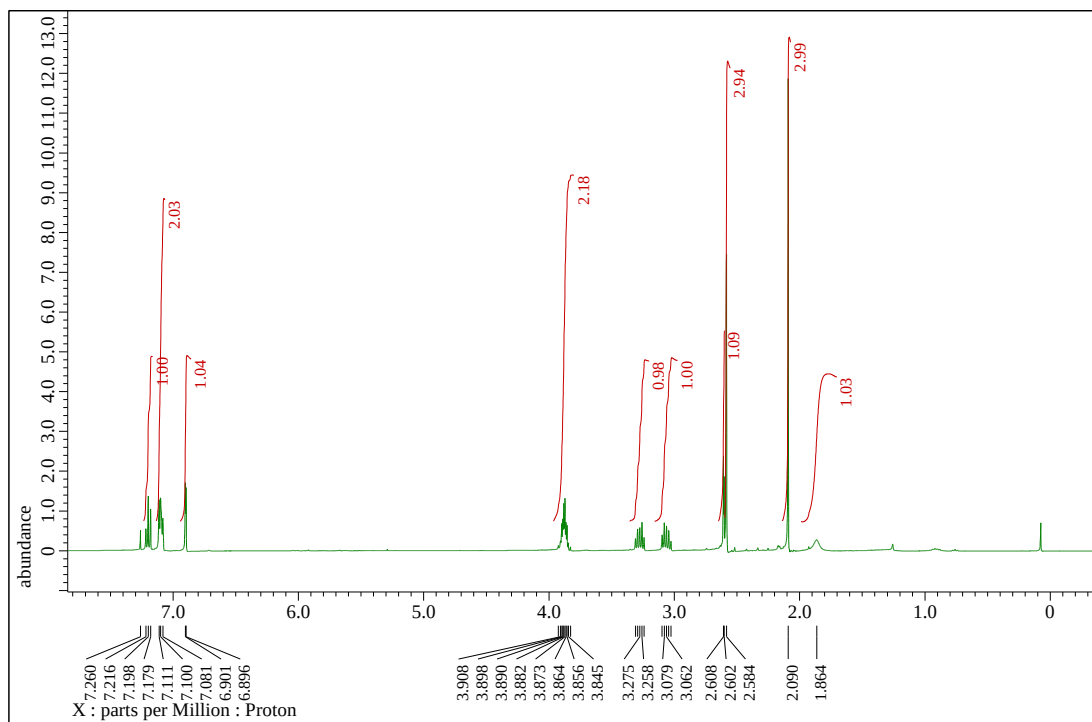
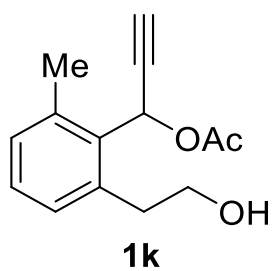


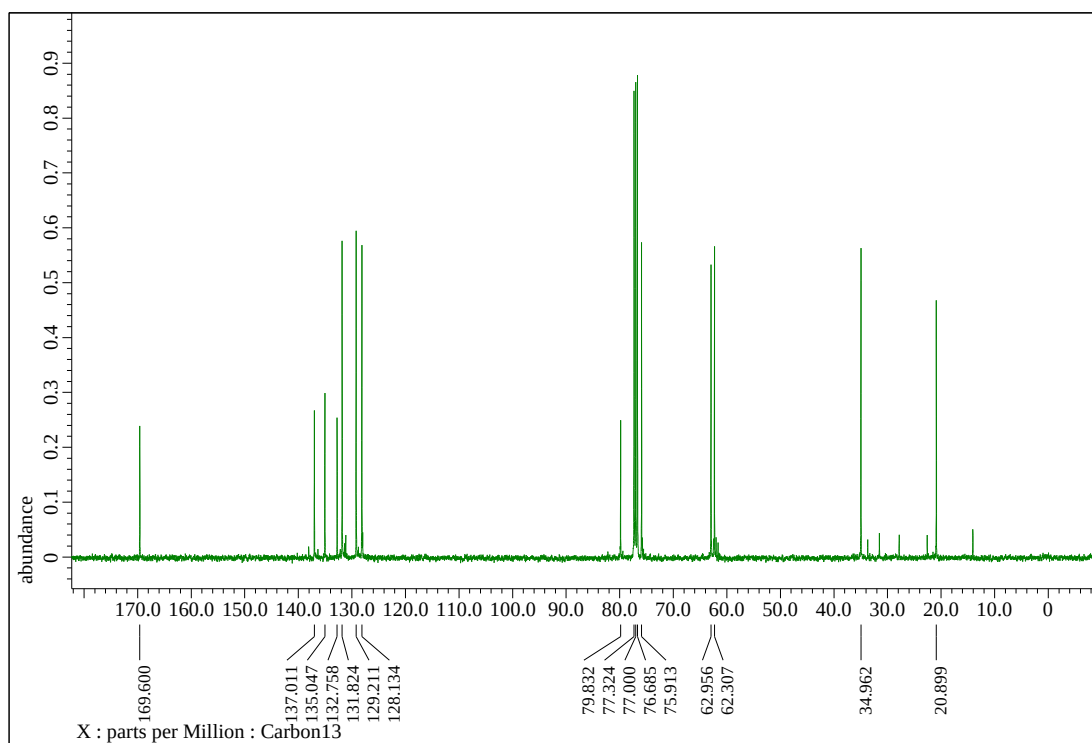
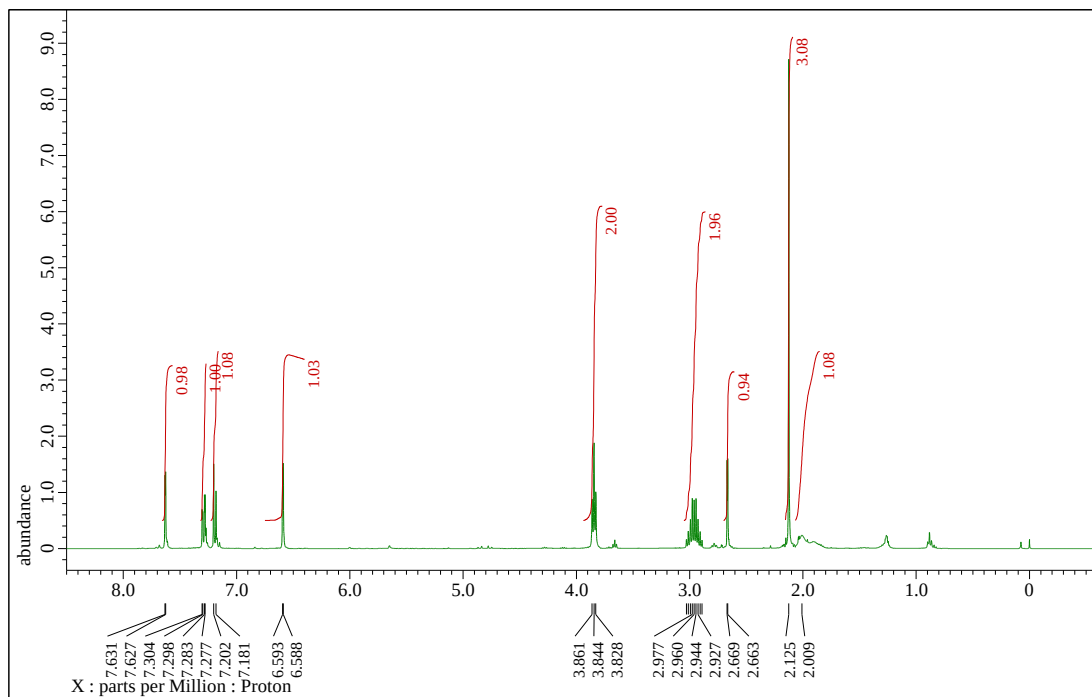
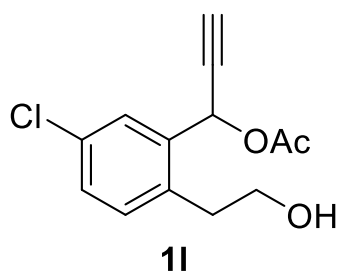


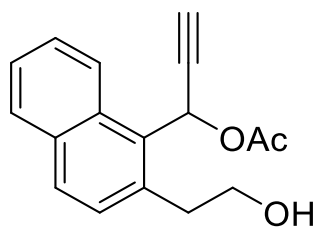




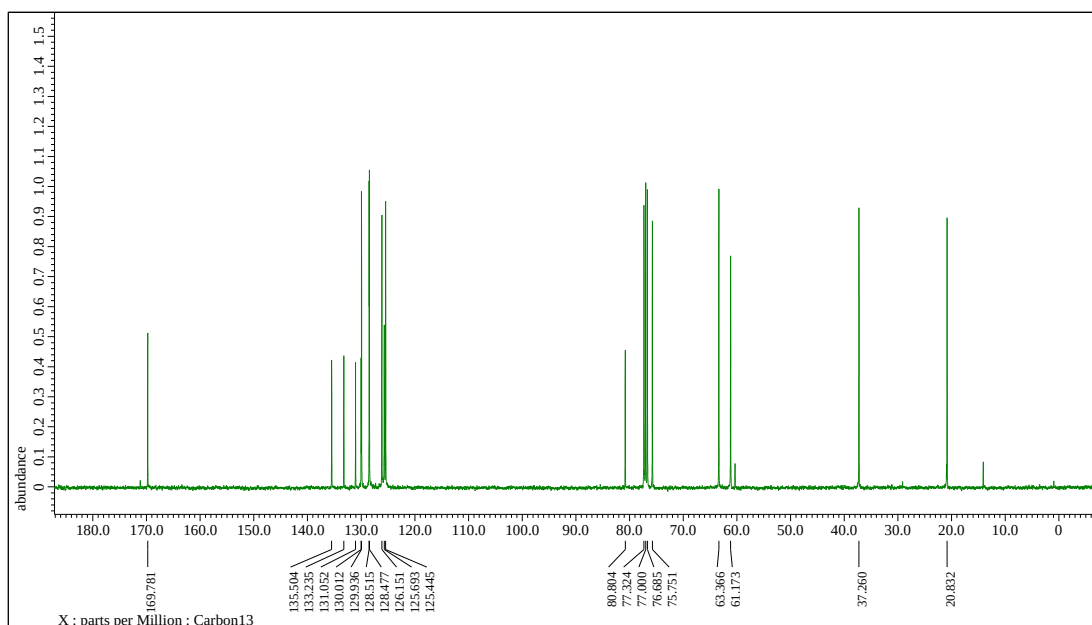
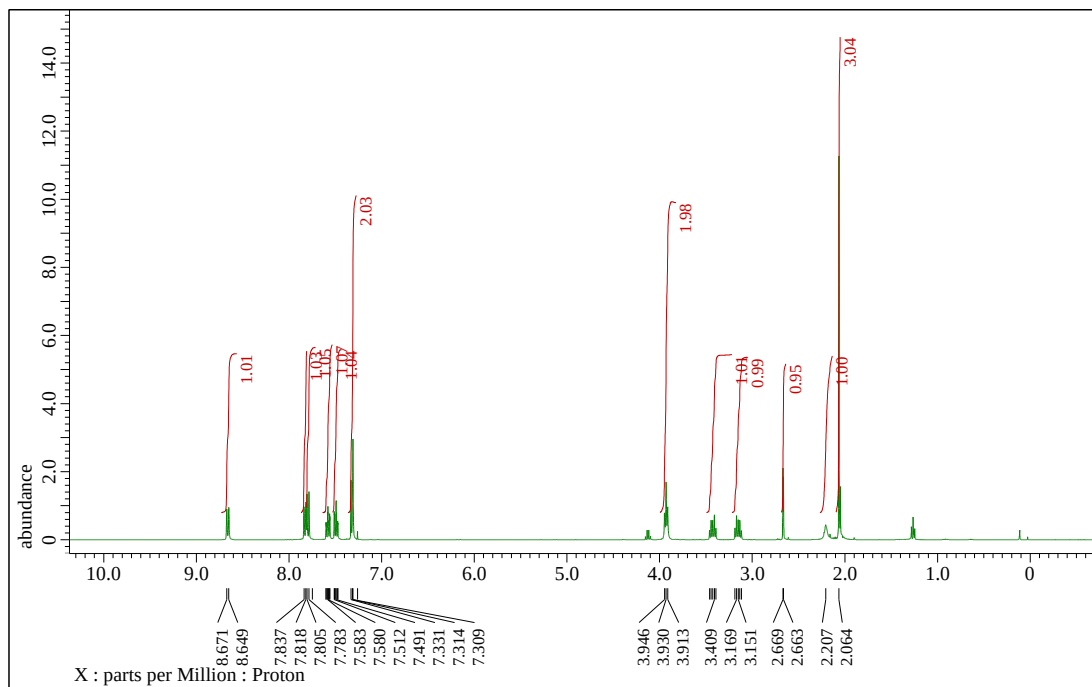


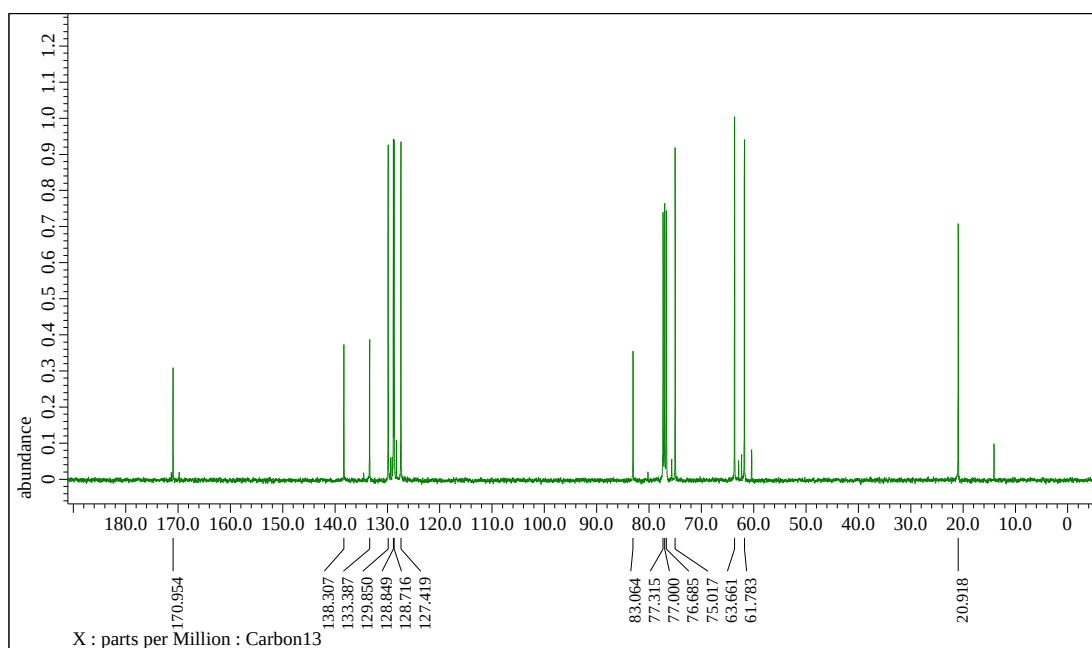
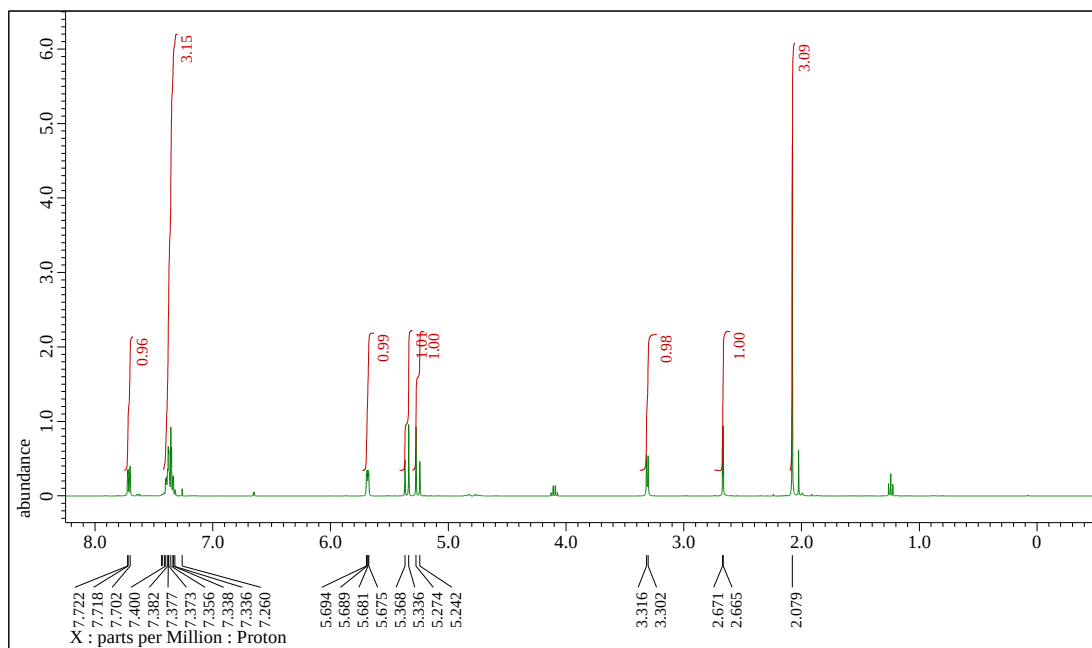
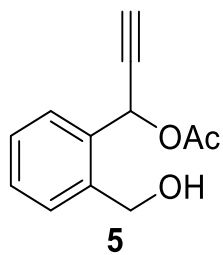


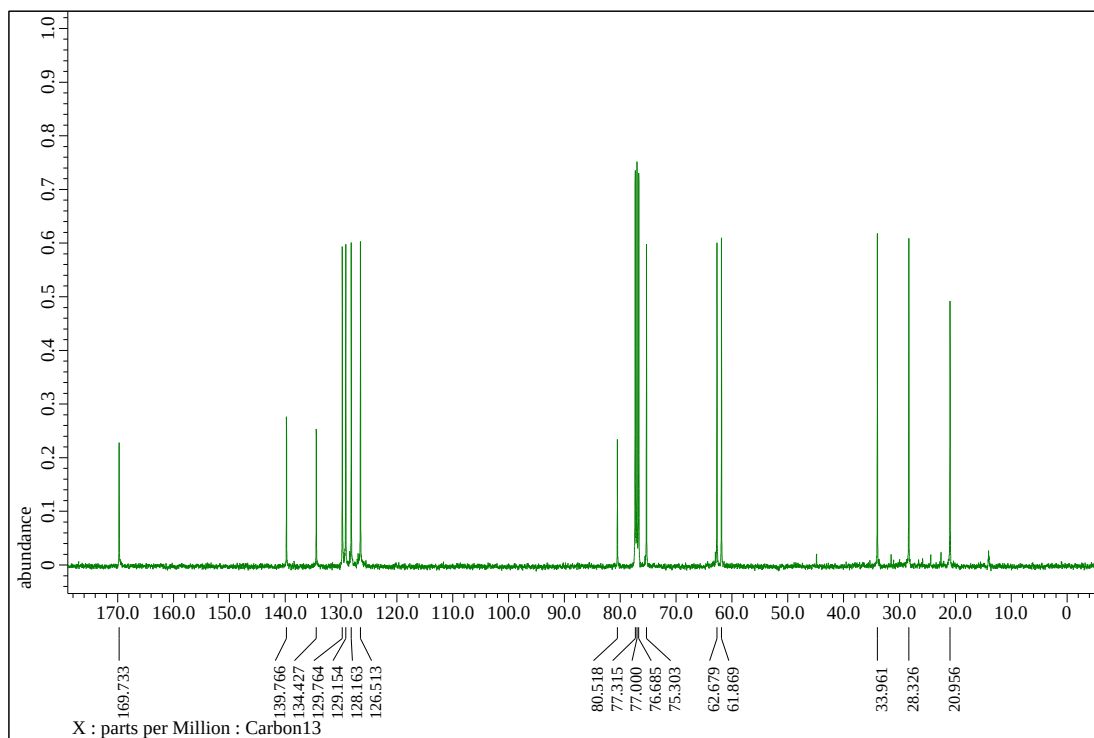
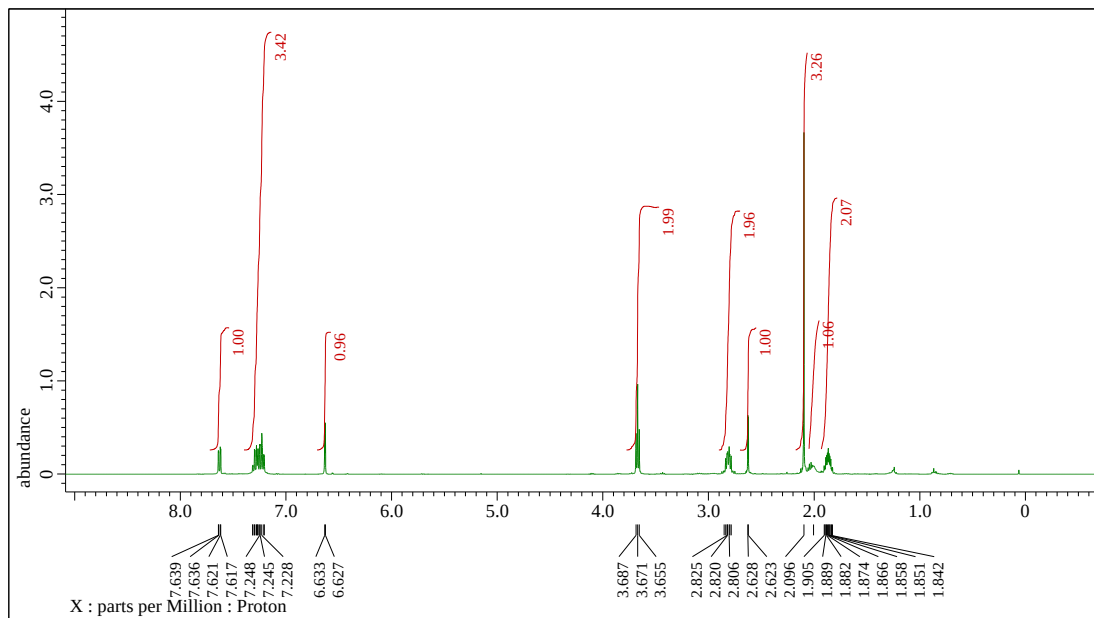
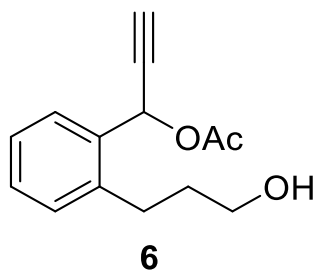


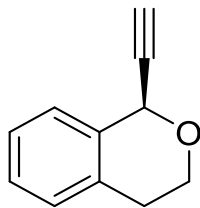


1m

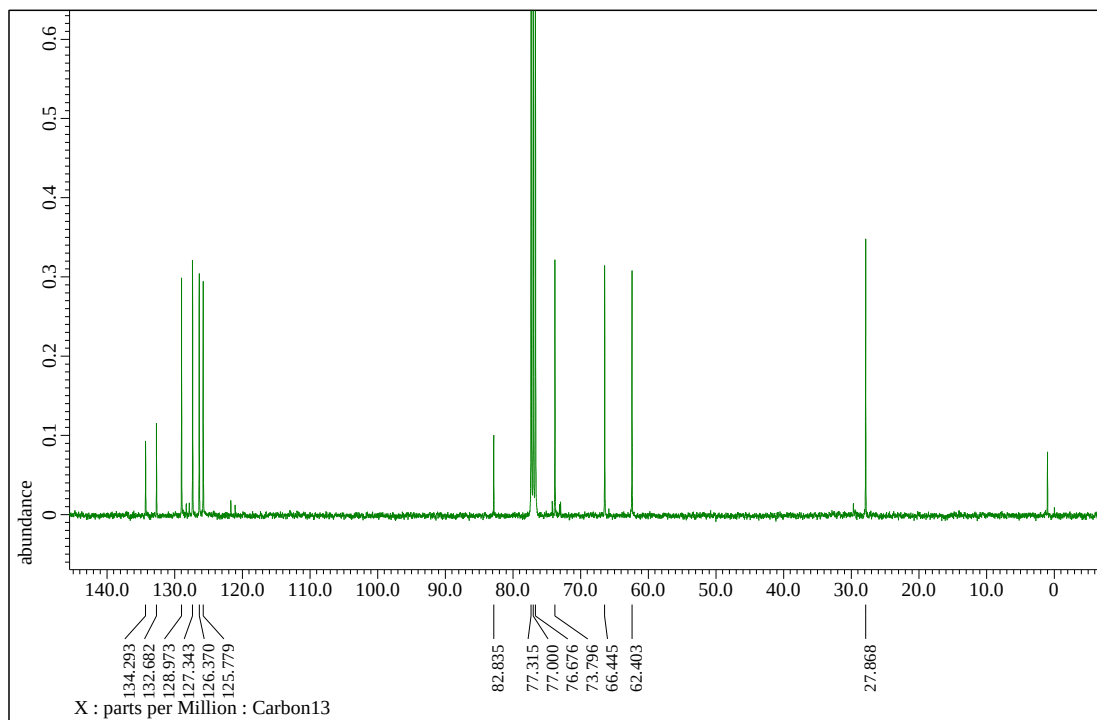
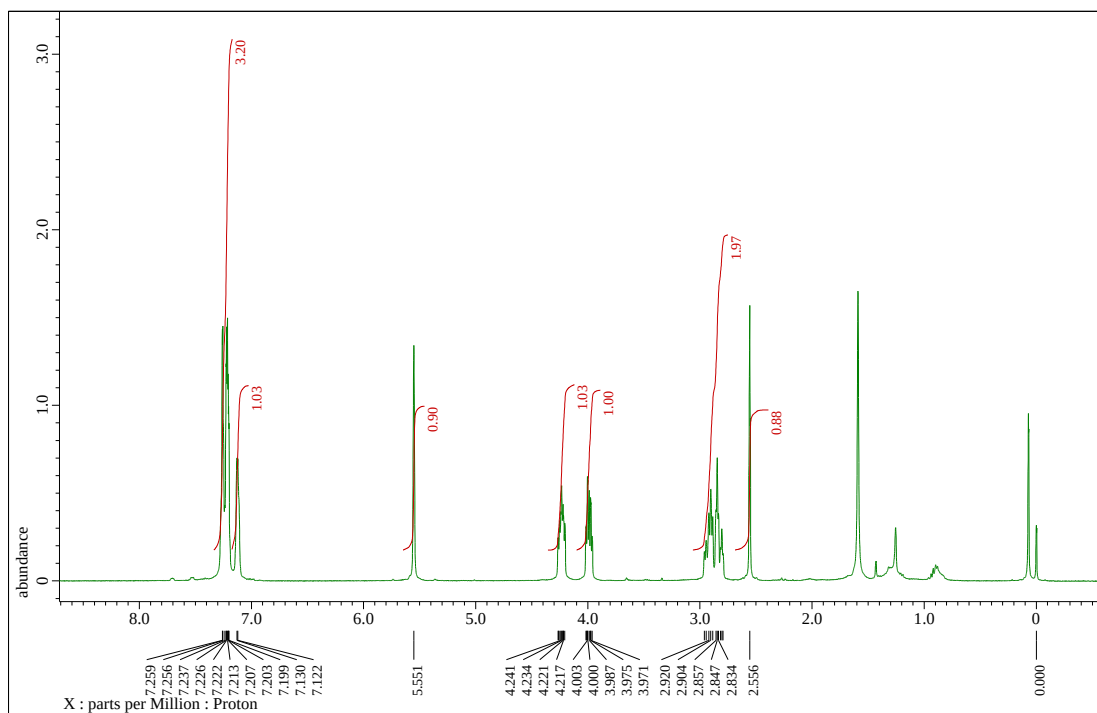


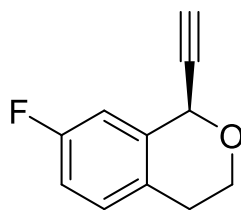




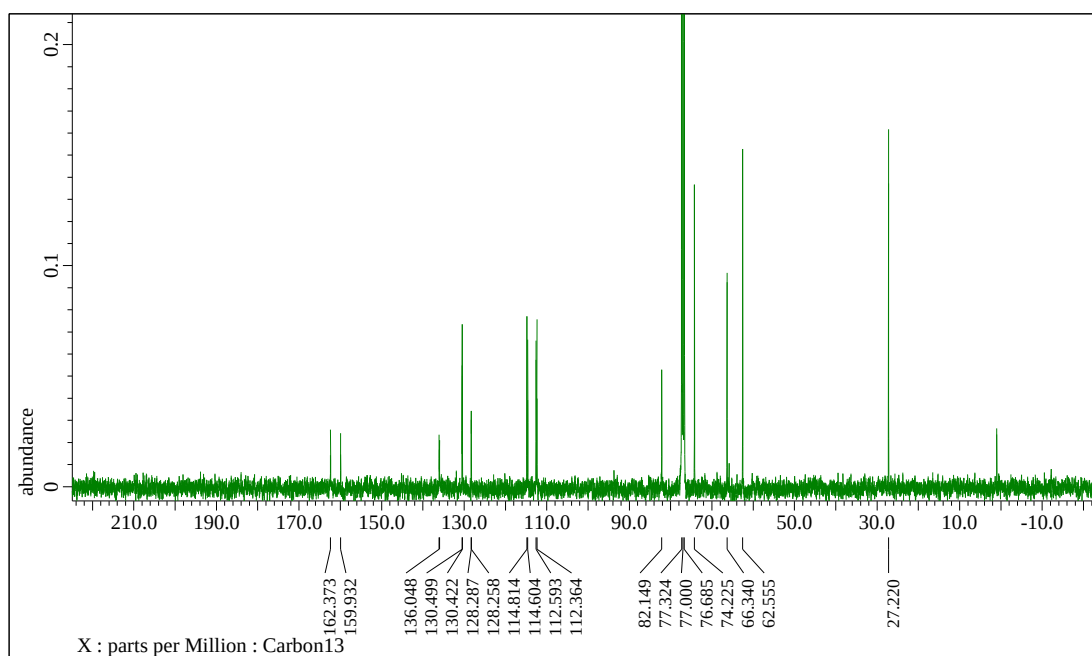
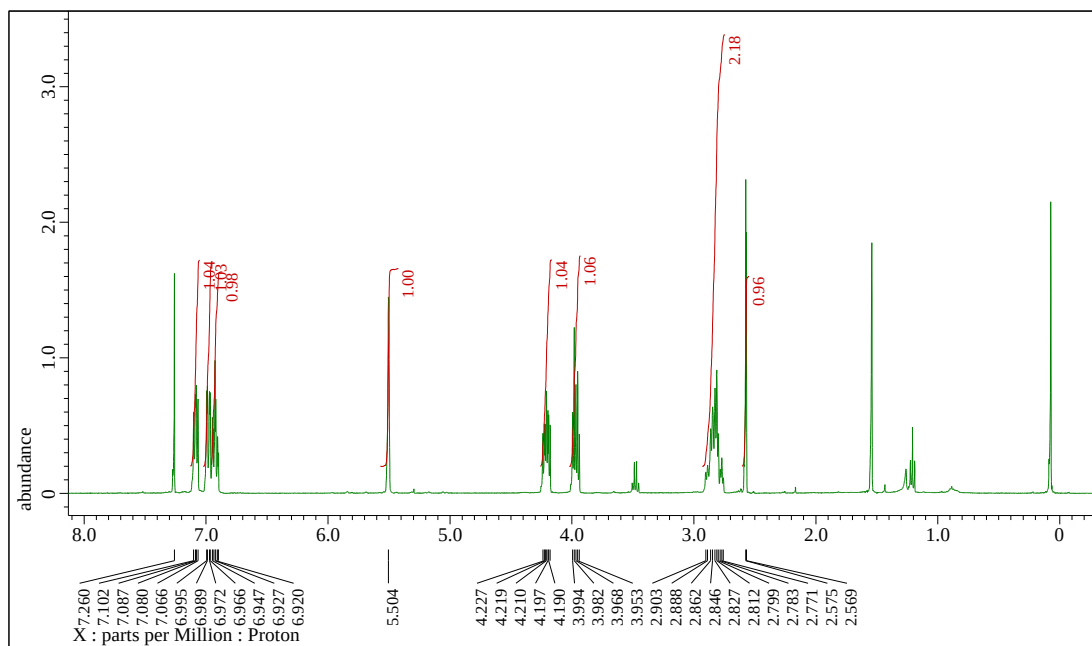


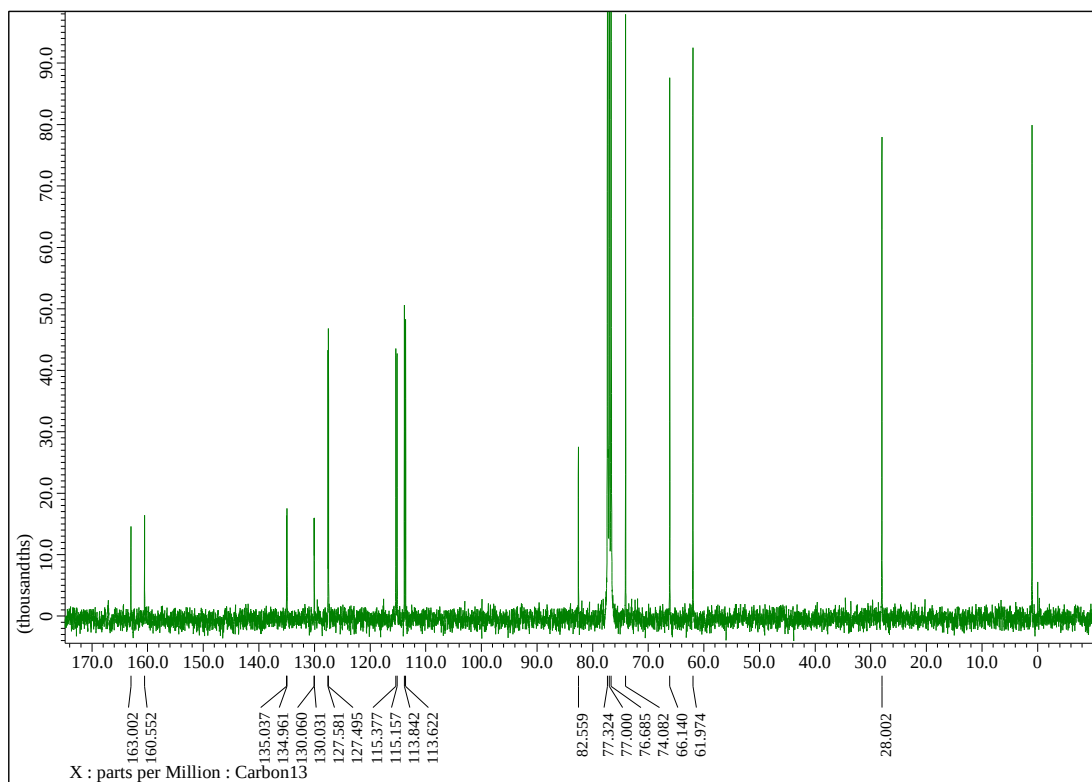
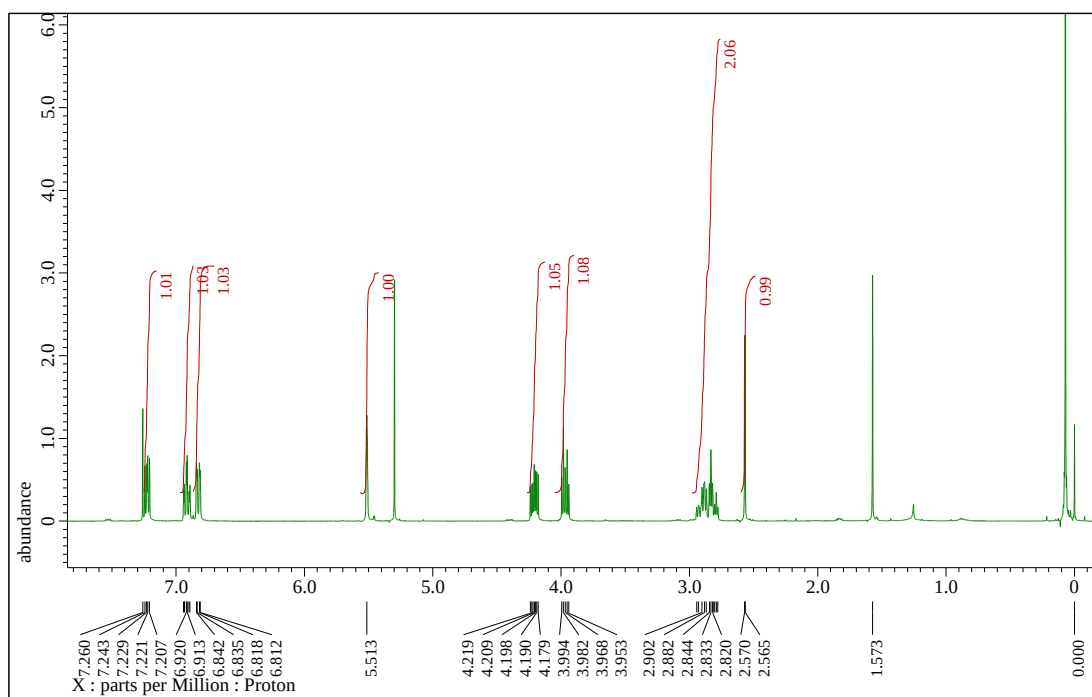
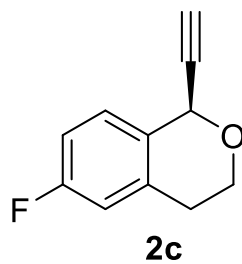
2a

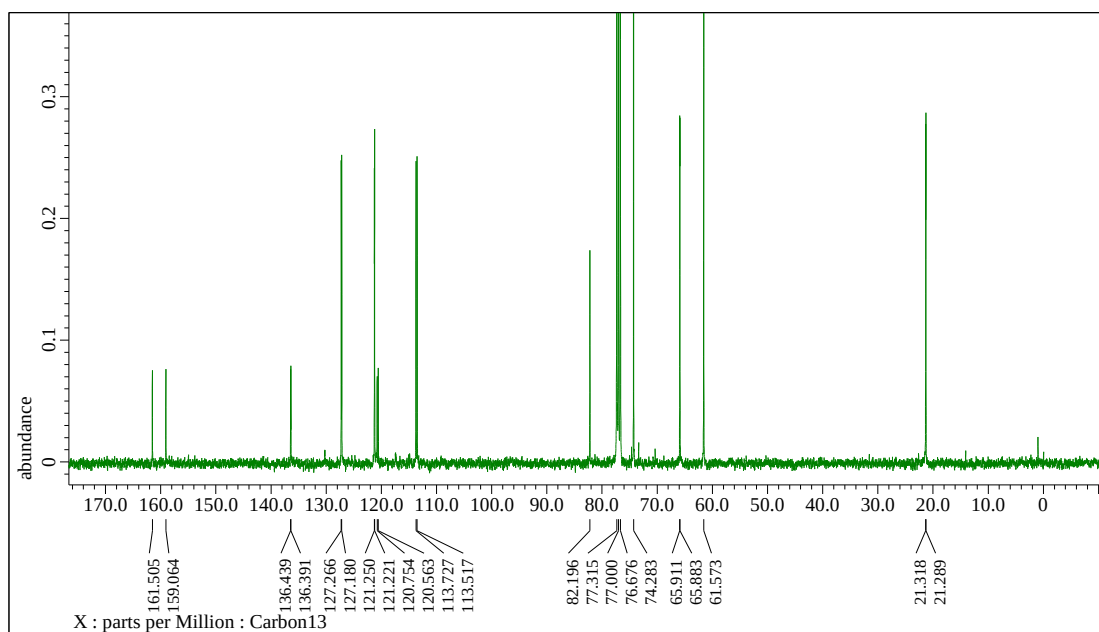
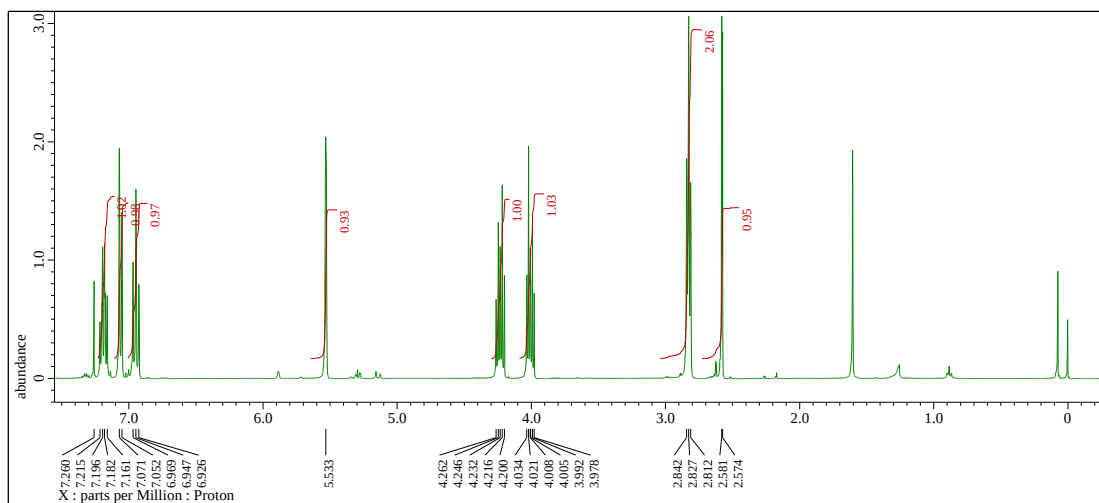
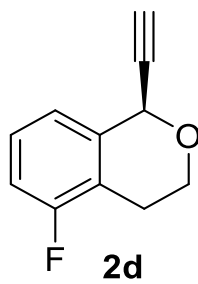


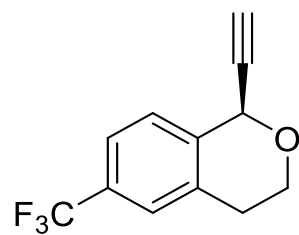


2b

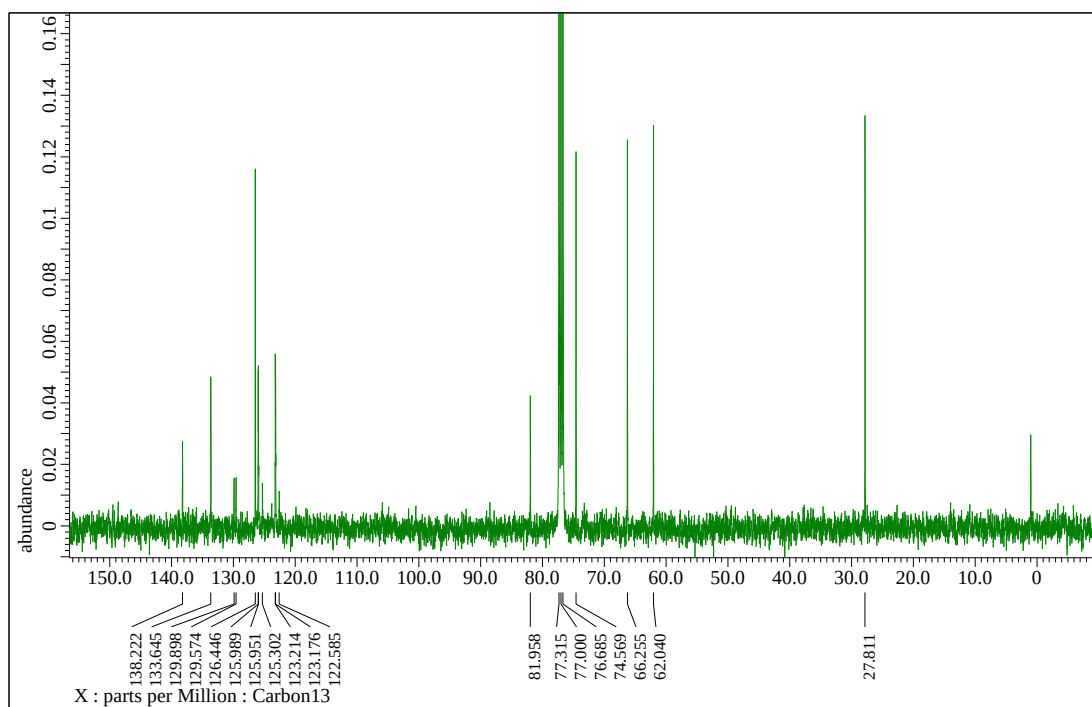
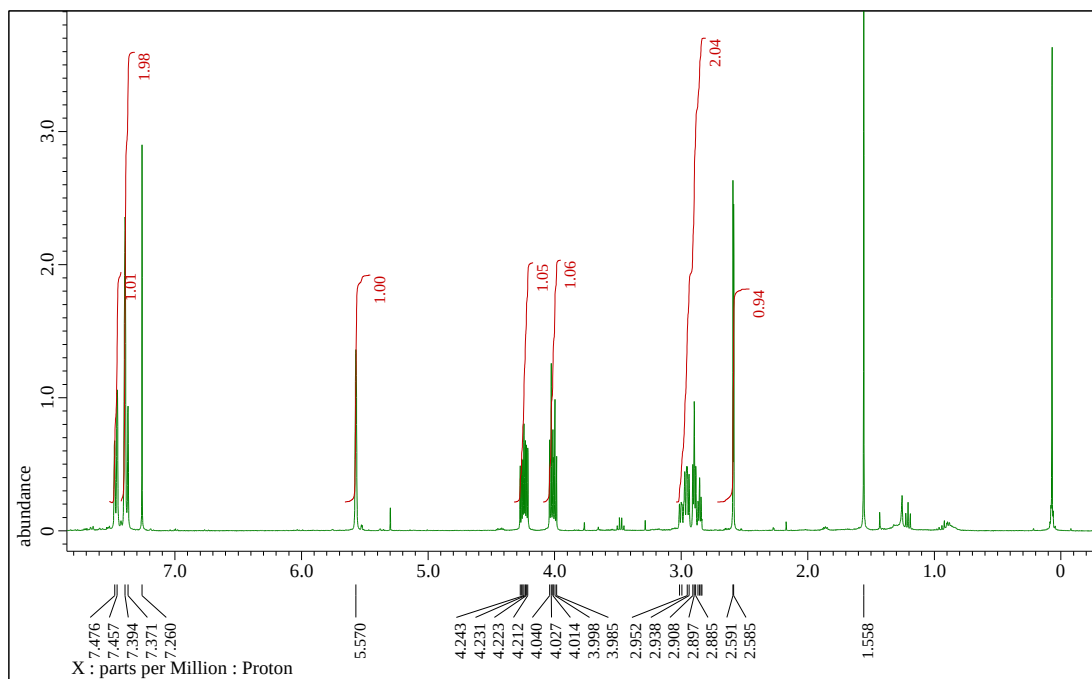


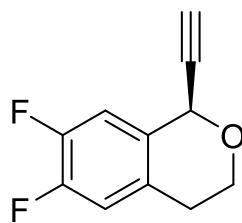




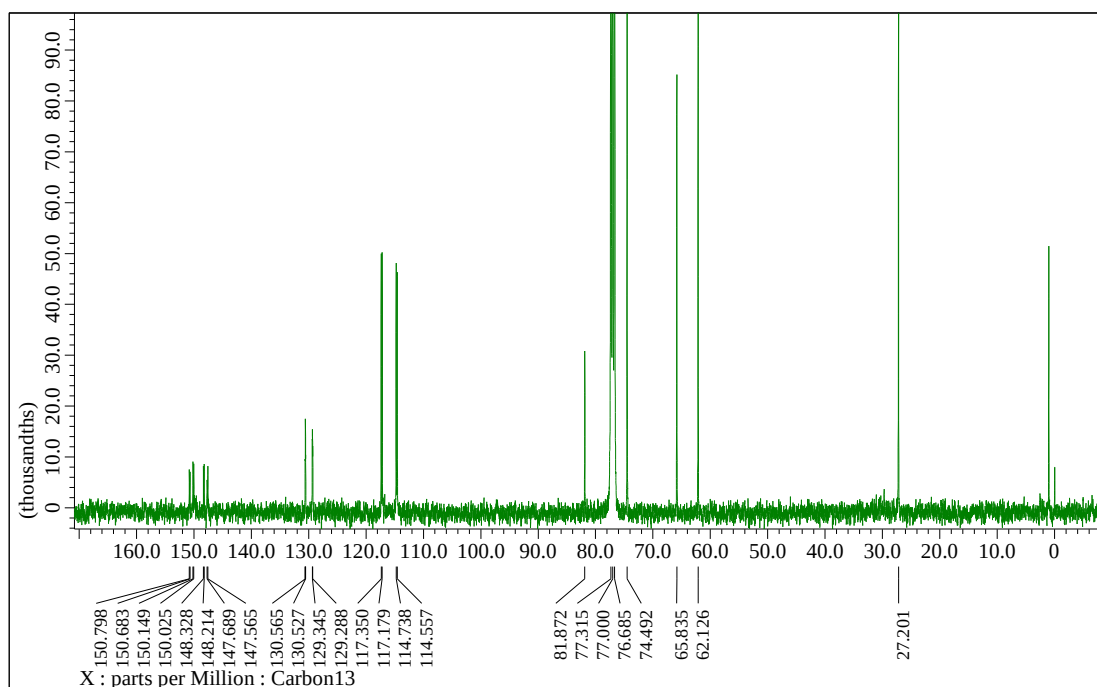
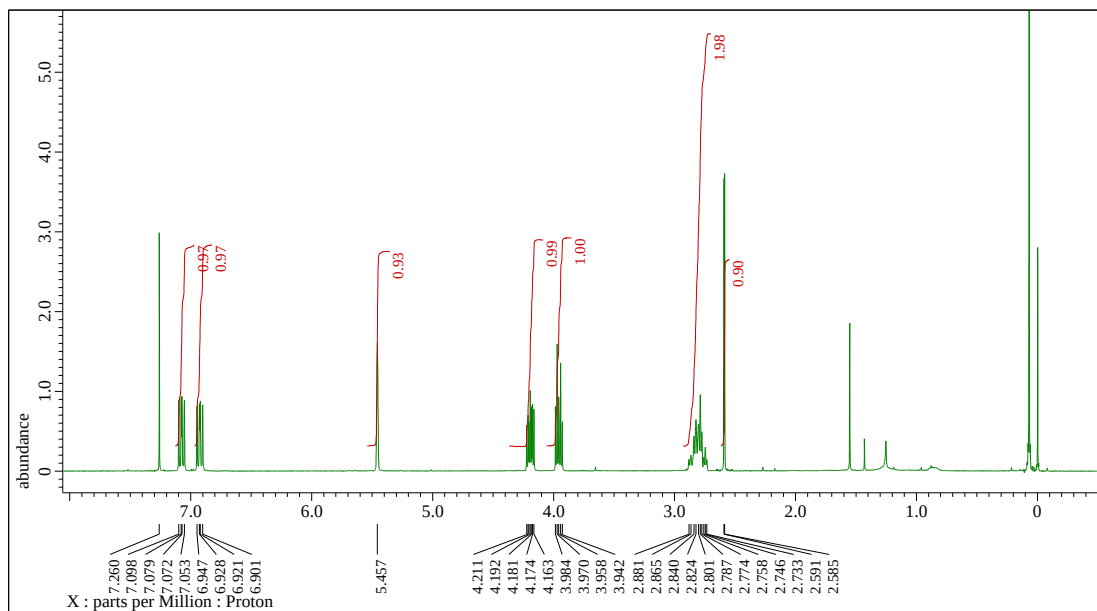


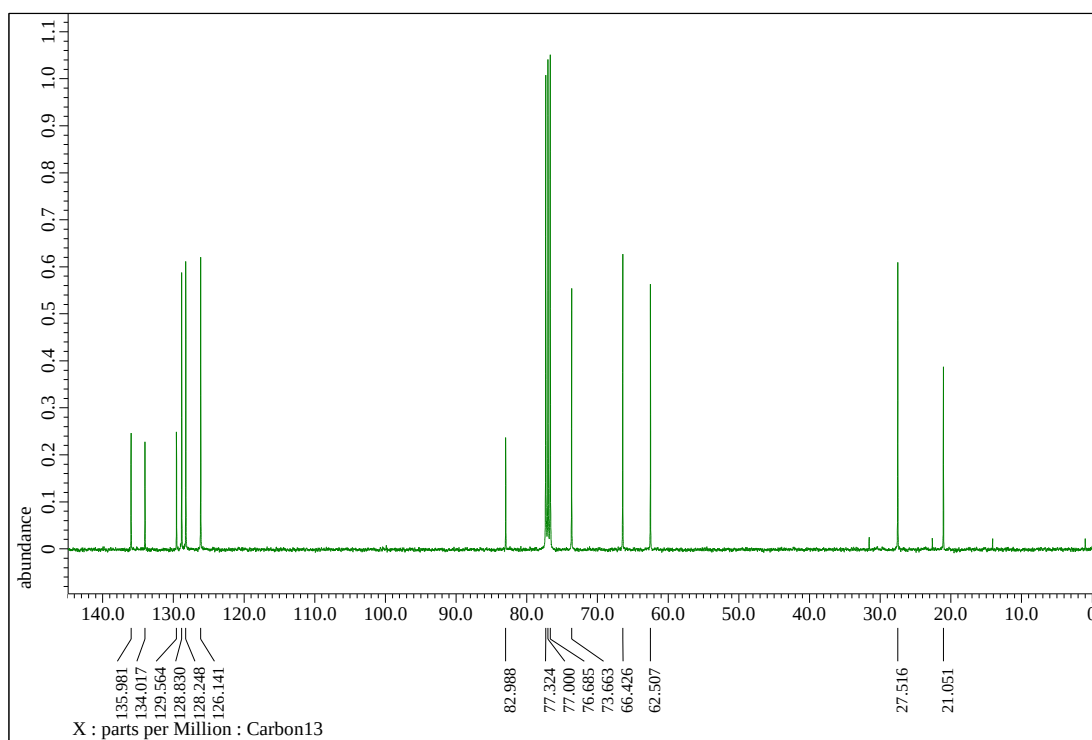
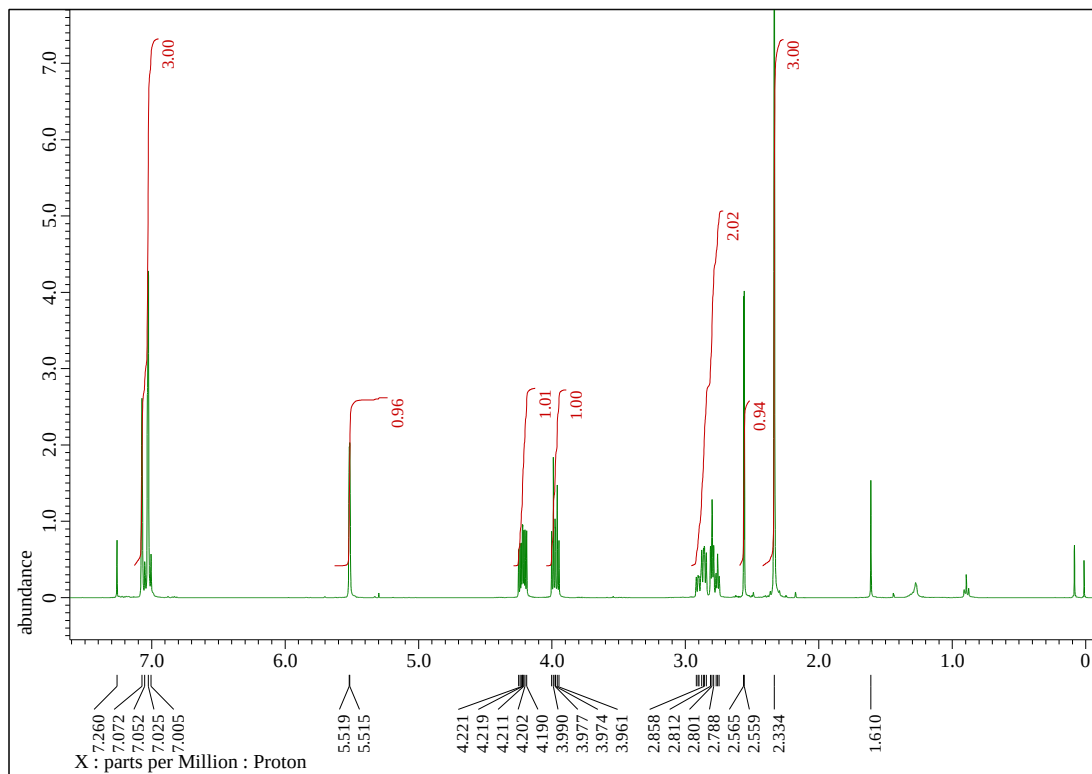
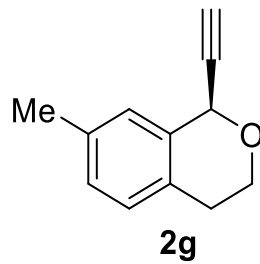
2e

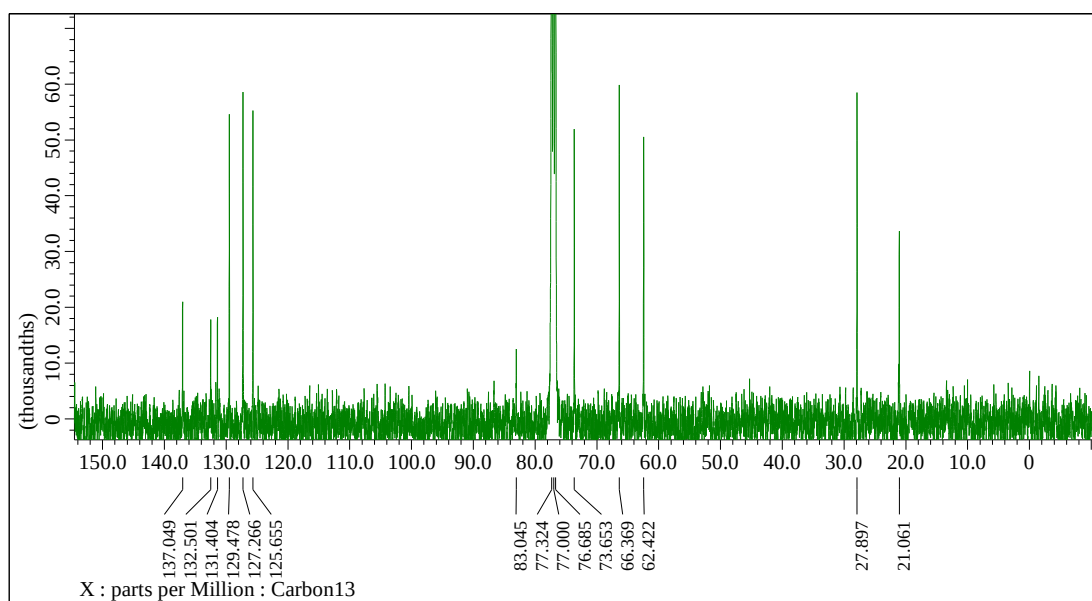
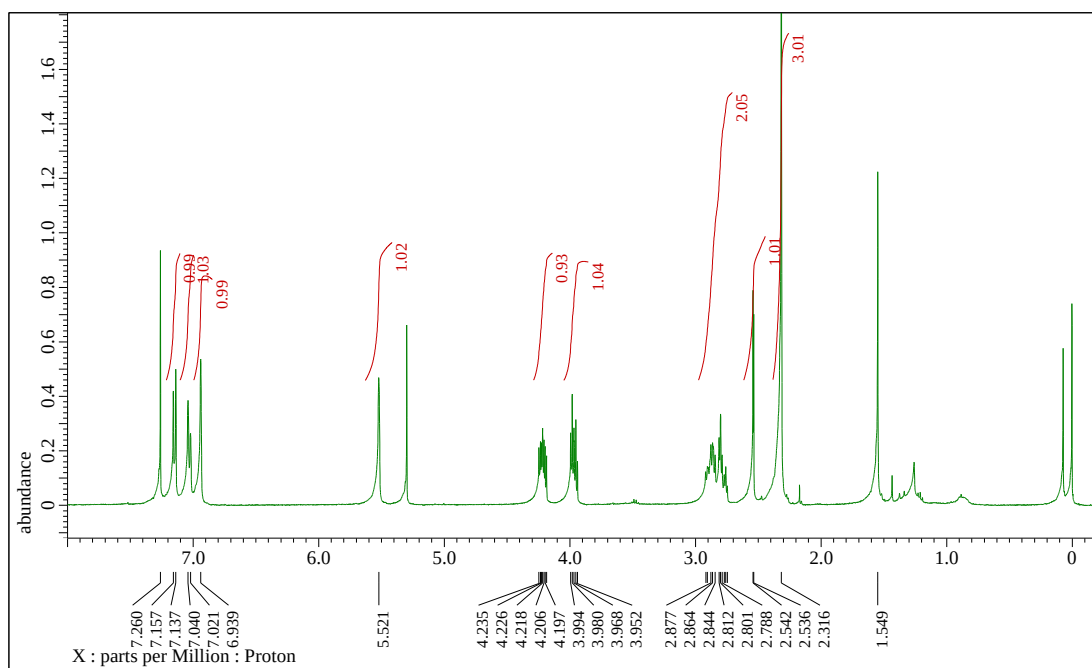
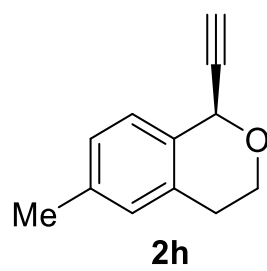


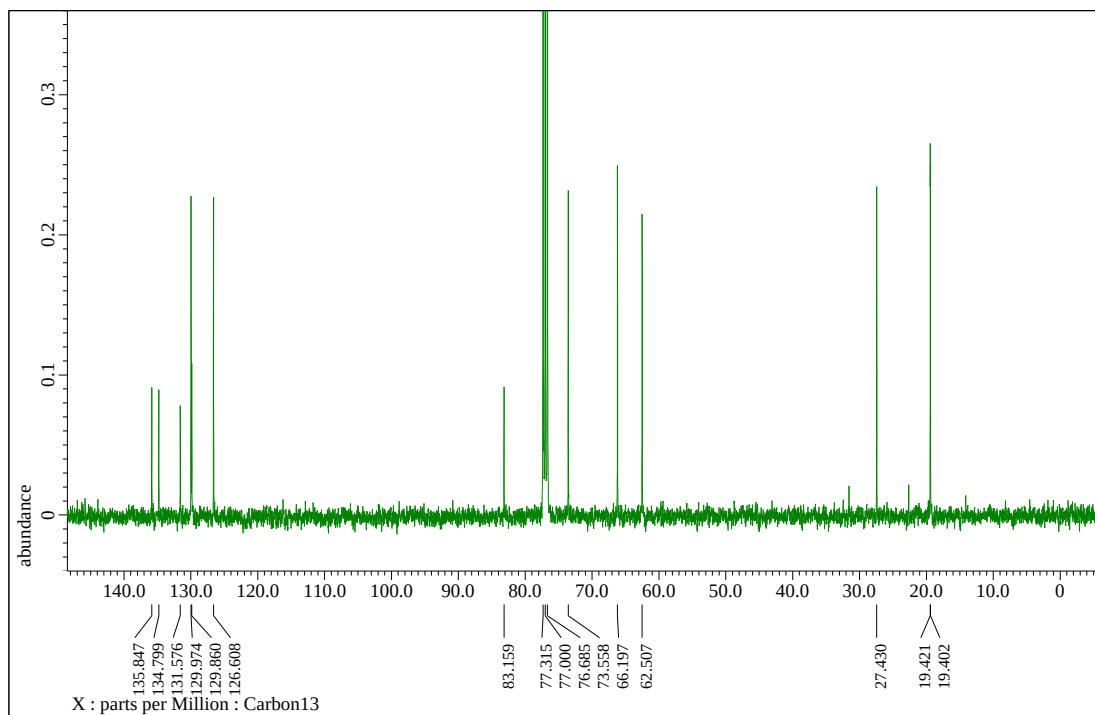
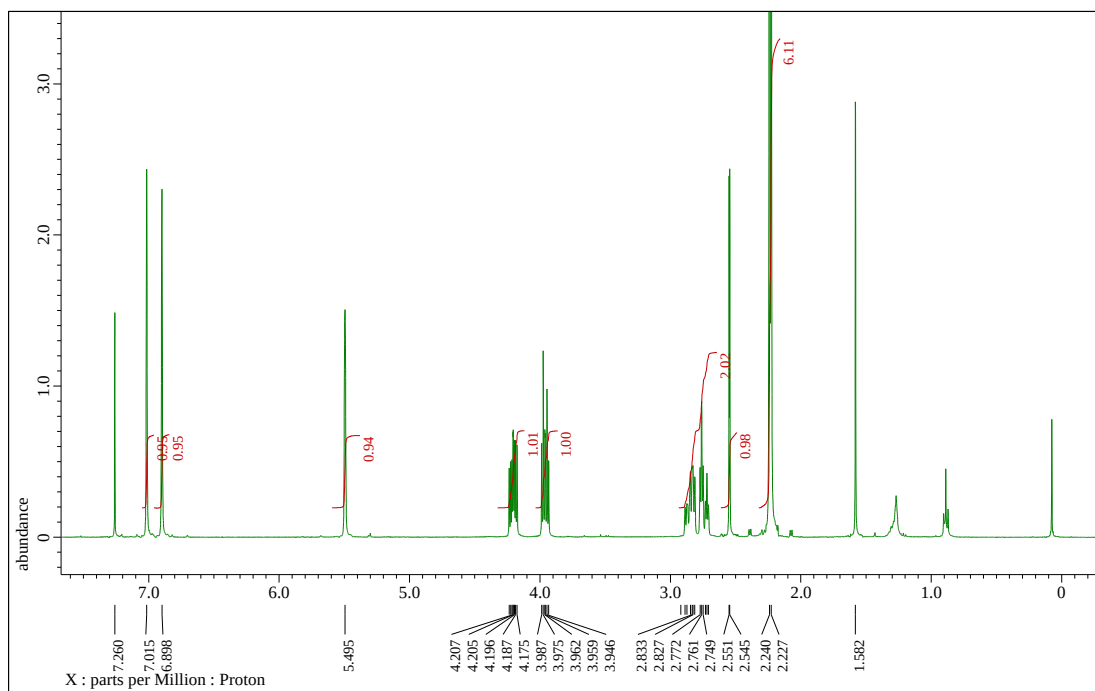
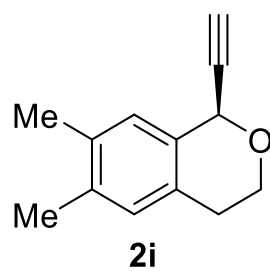


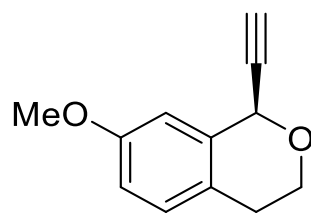
2f



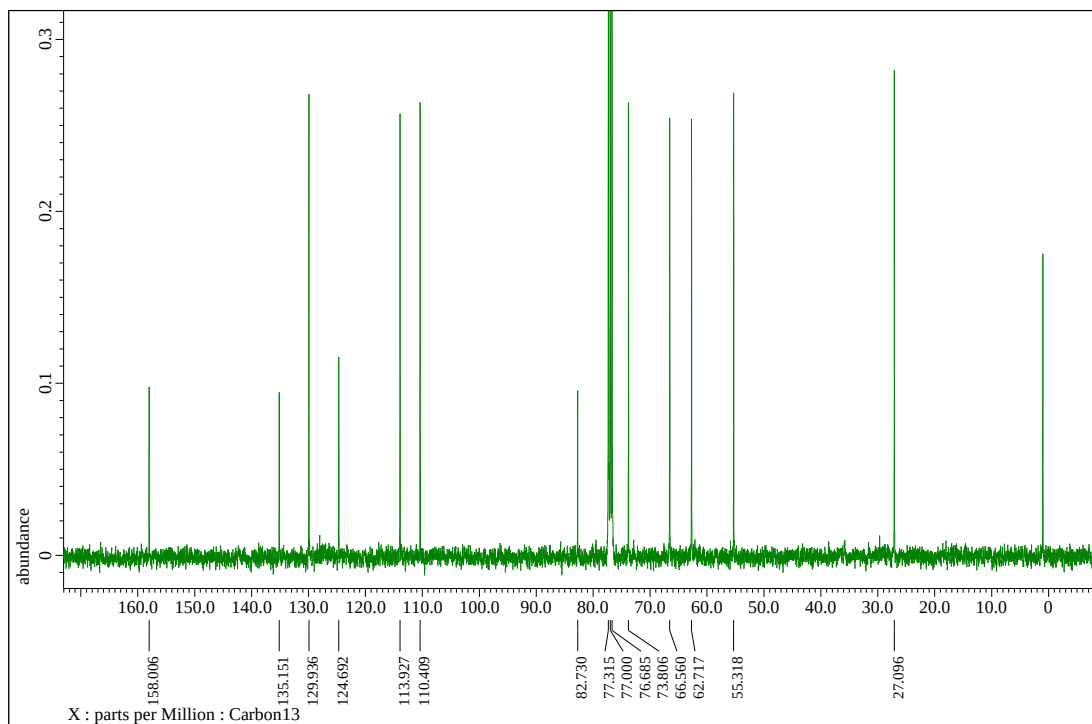
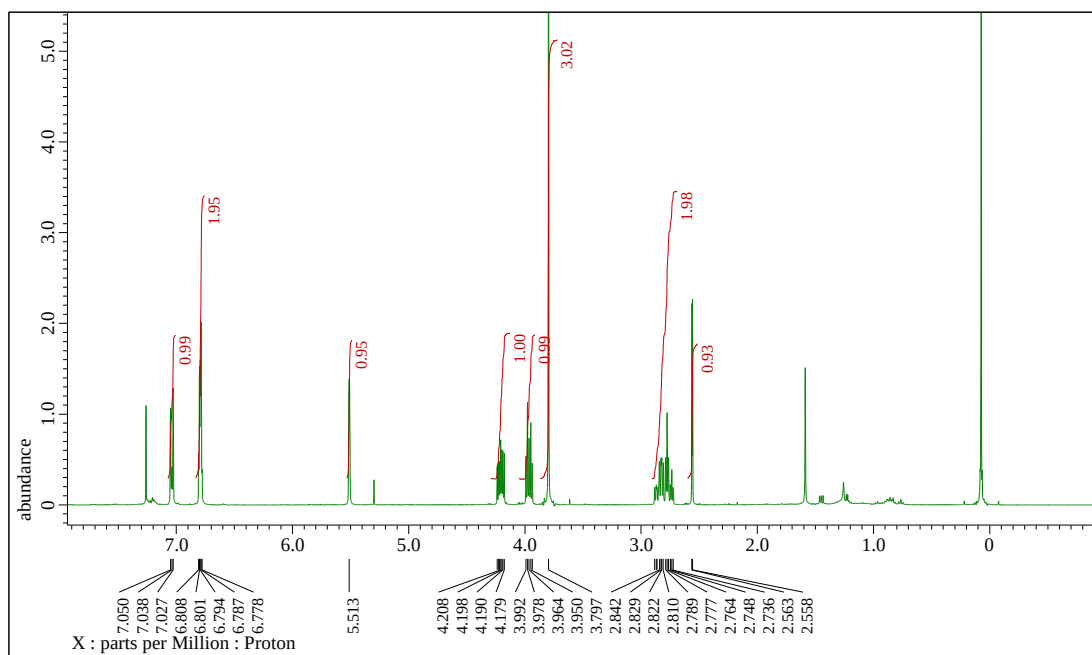


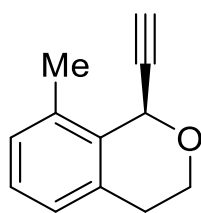




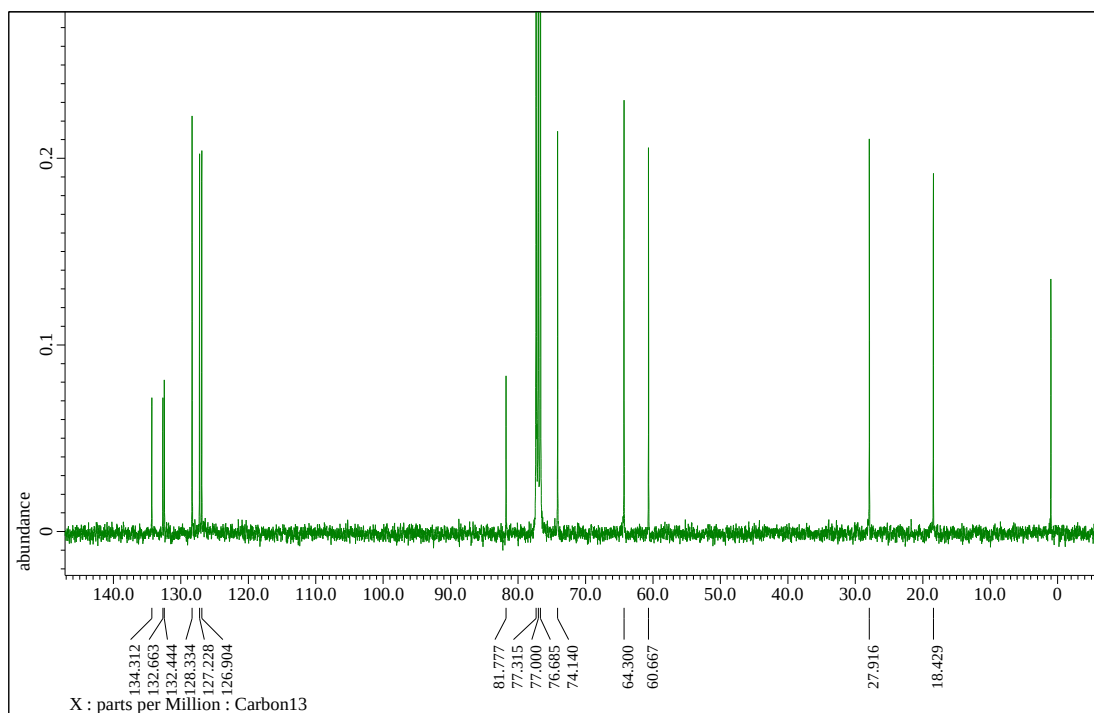
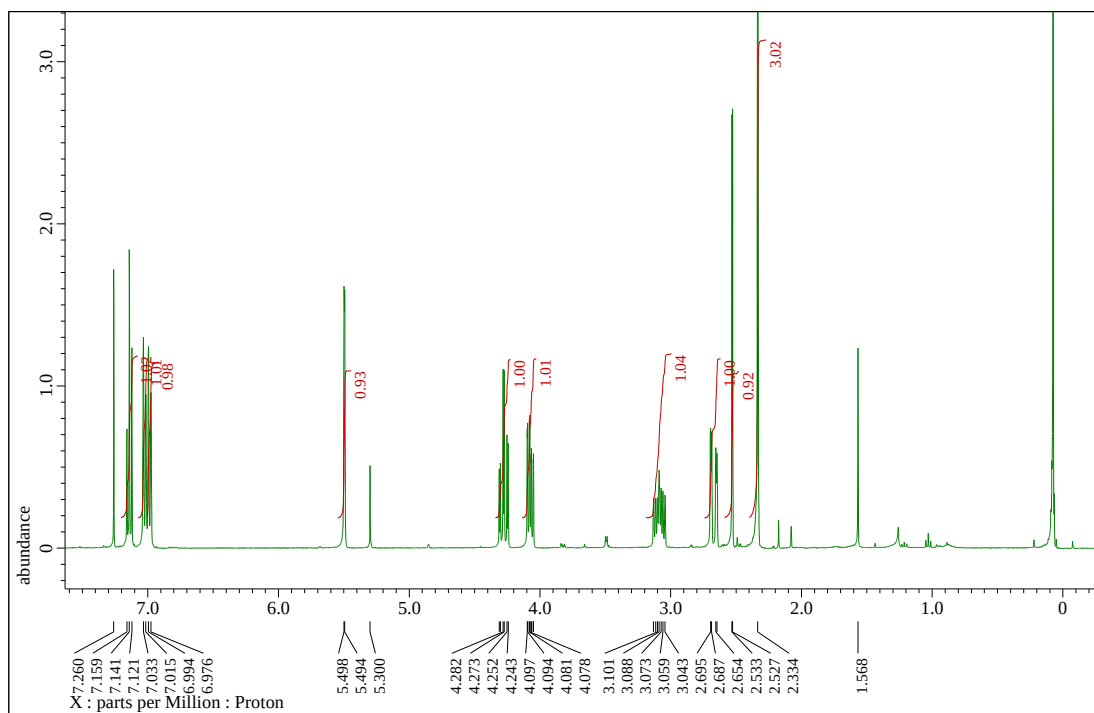


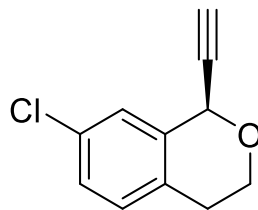
2j



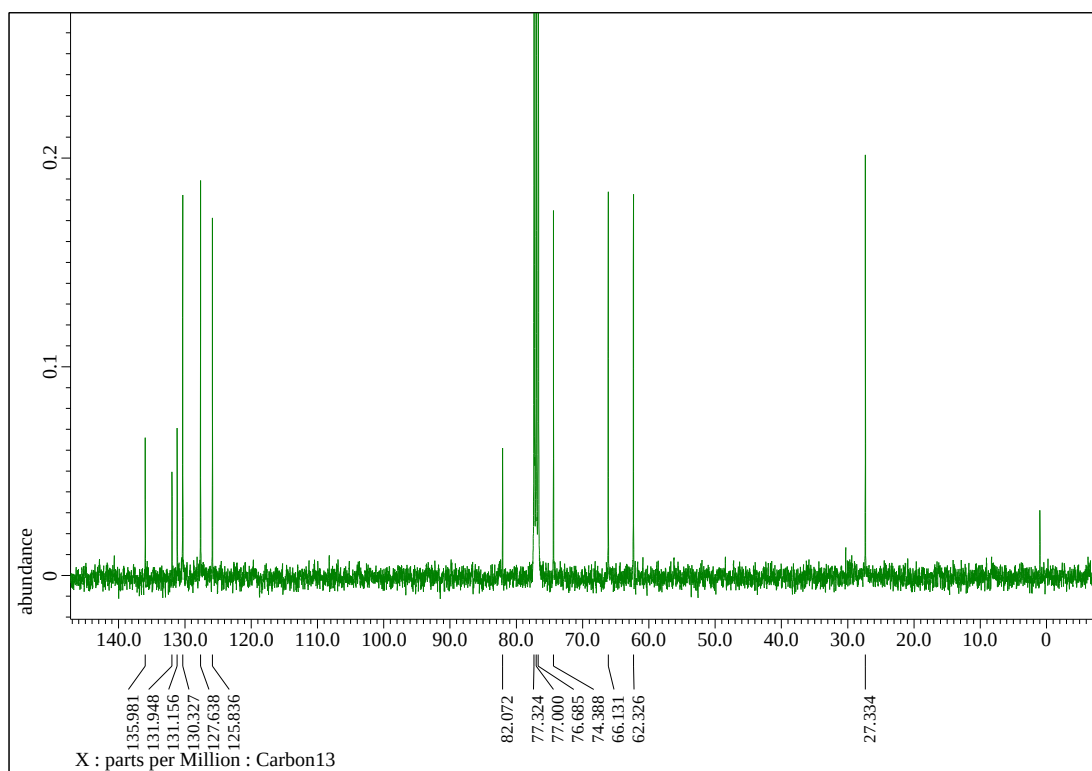
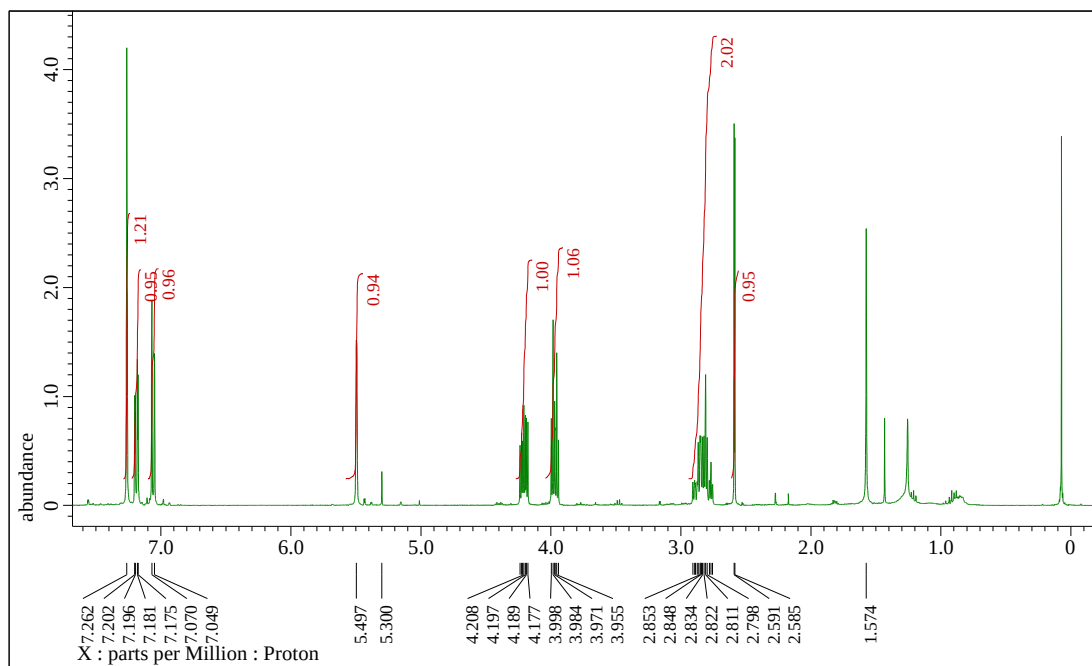


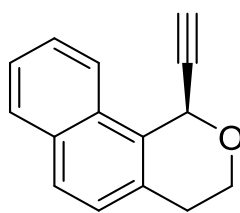
2k



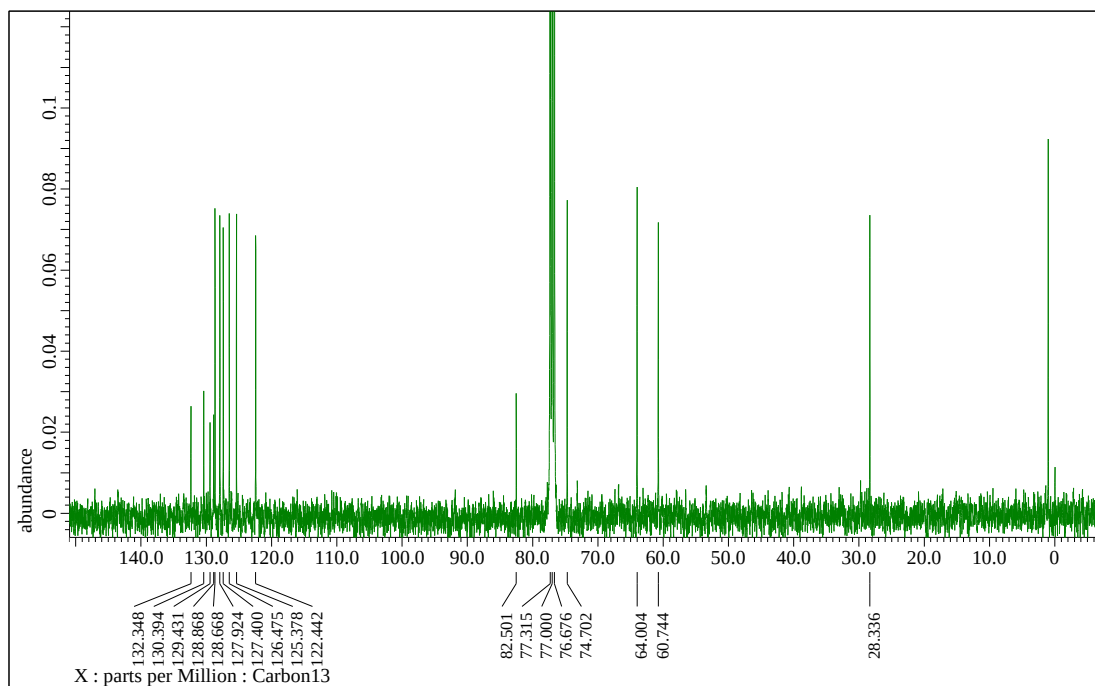
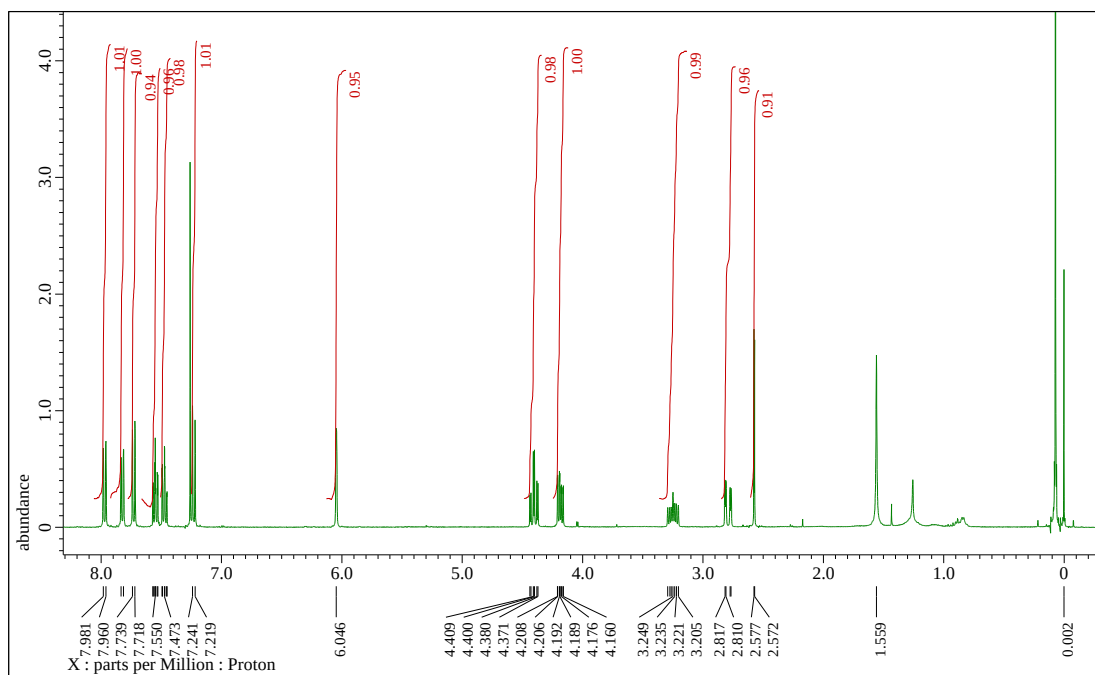


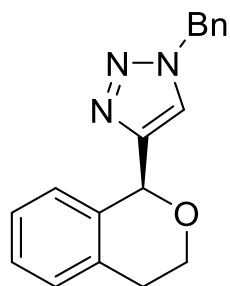
2I



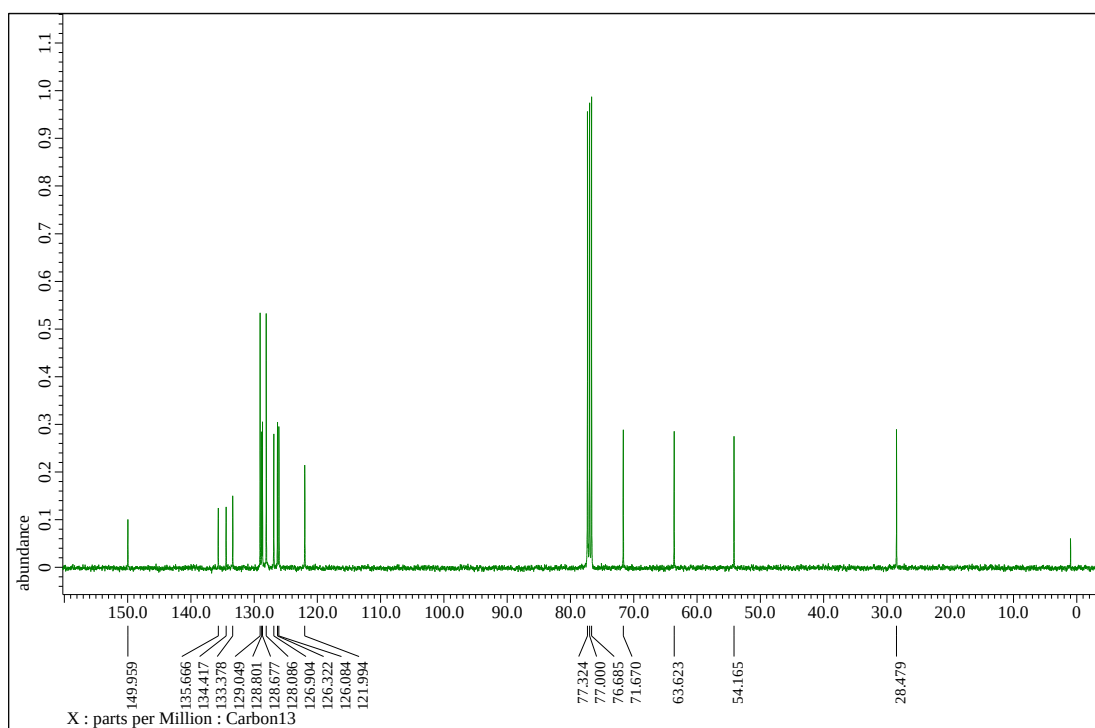
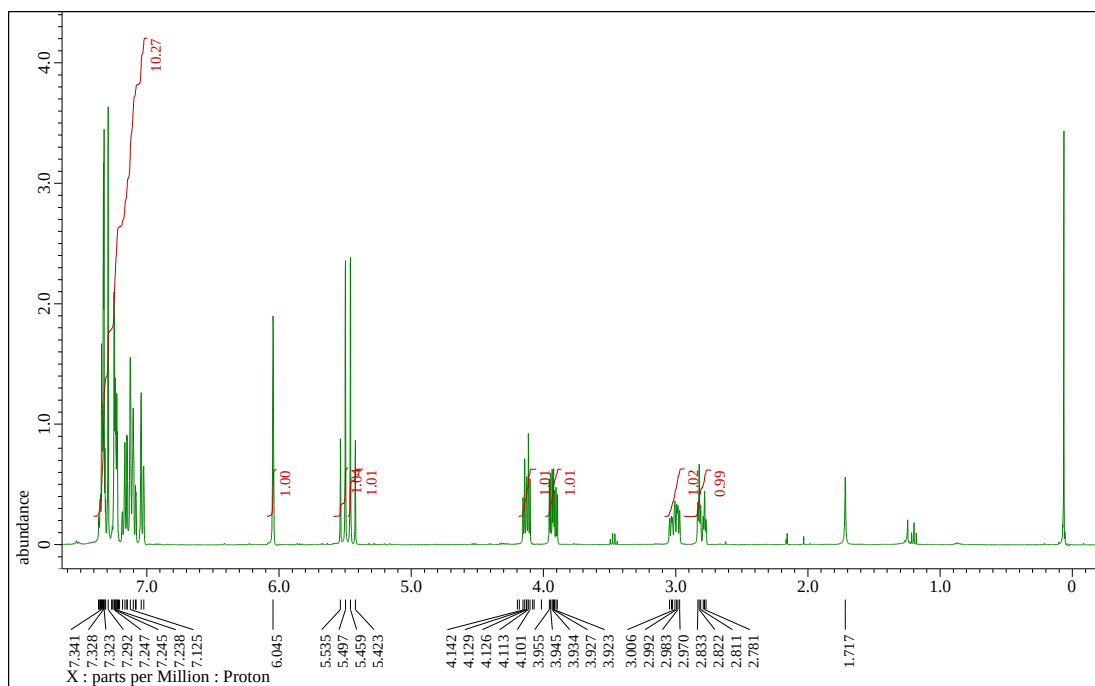


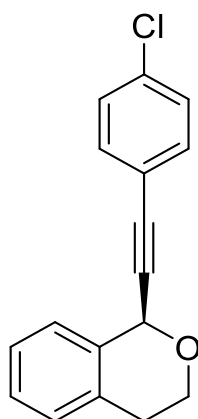
2m



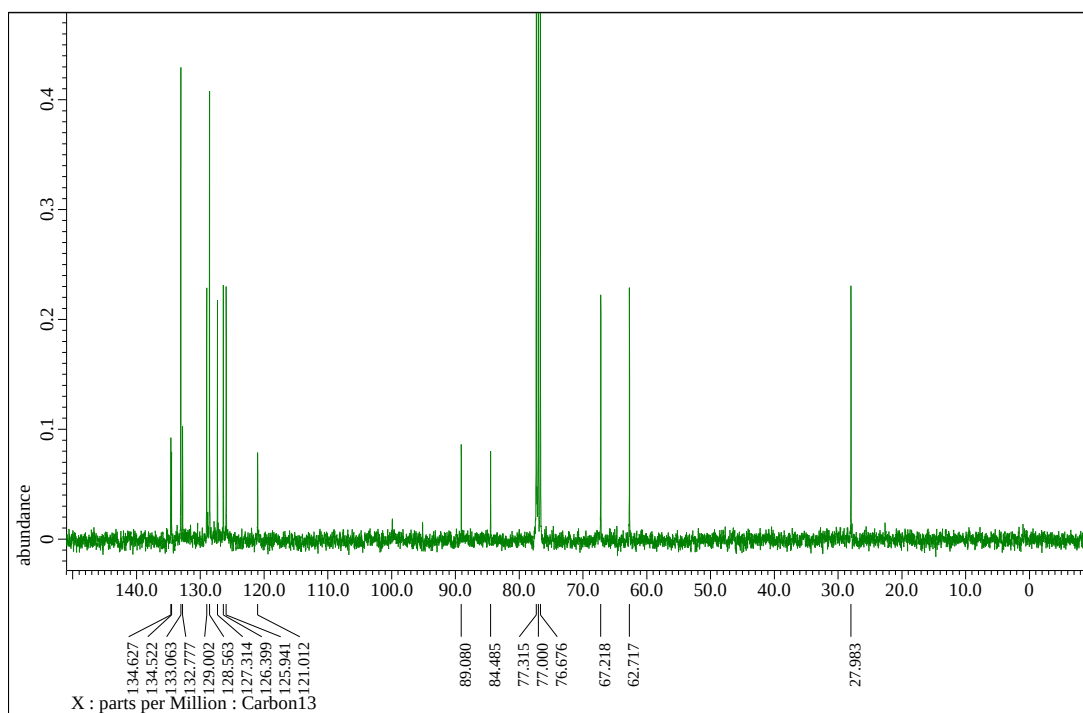
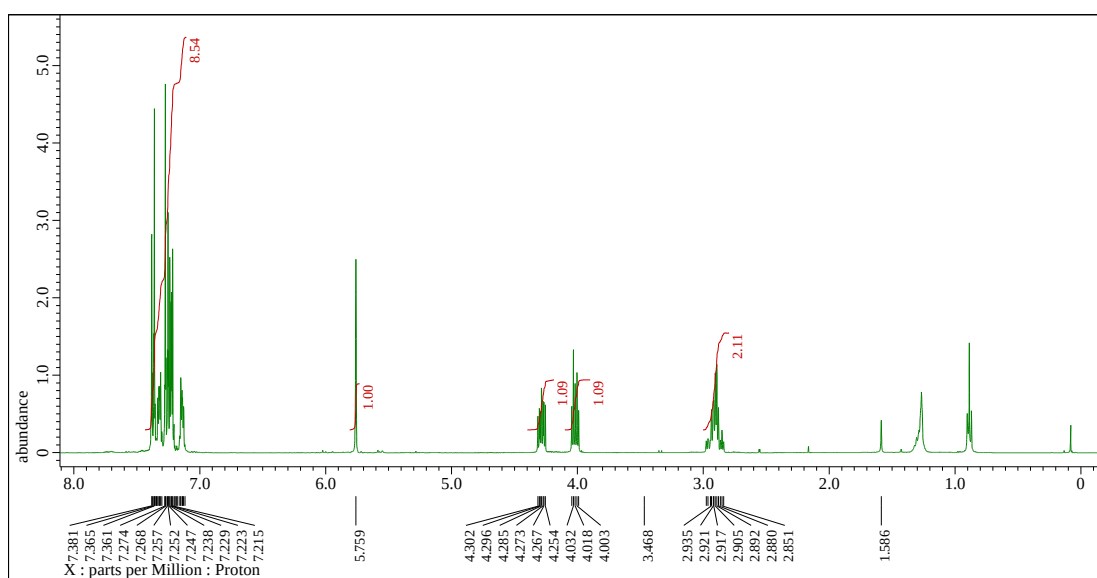


3

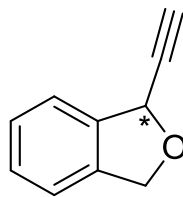




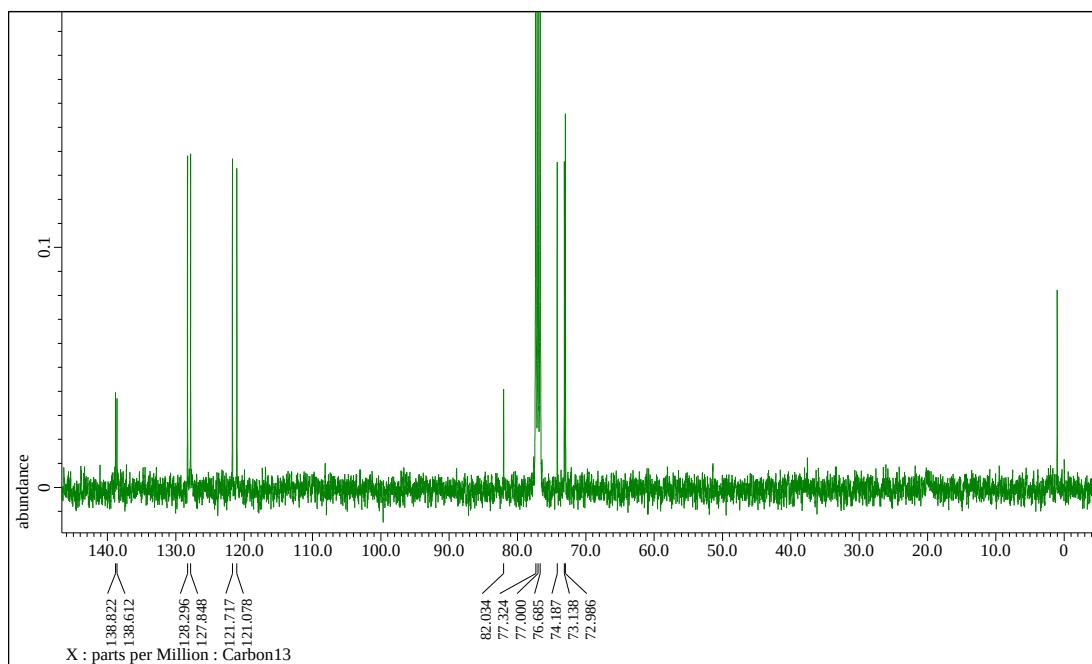
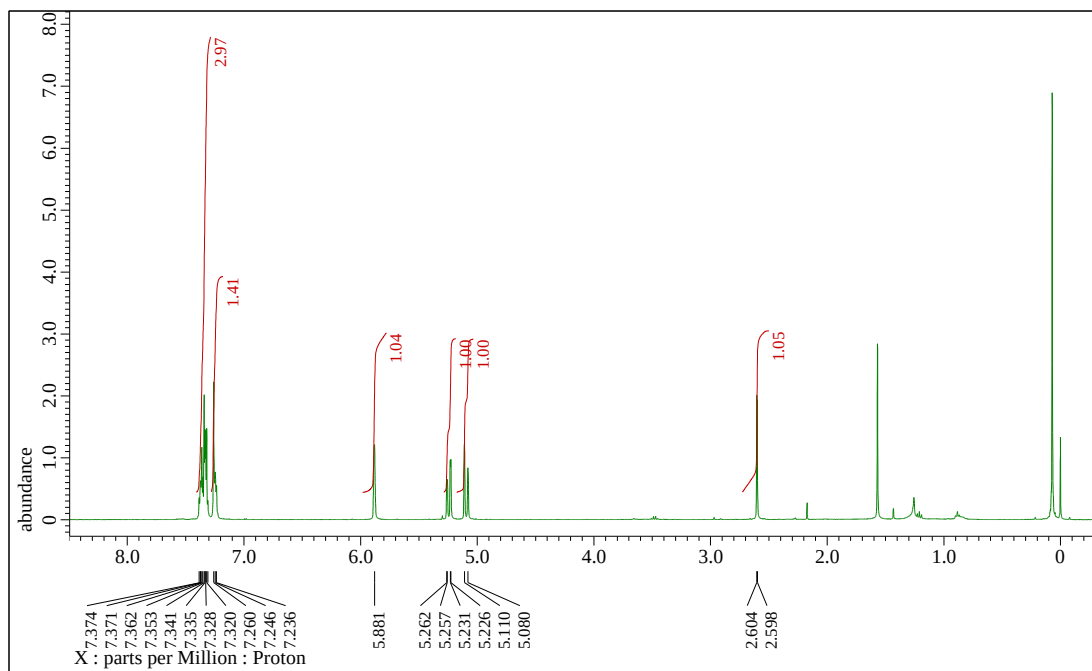
4

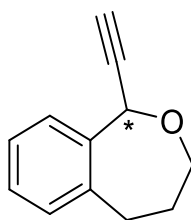


S48

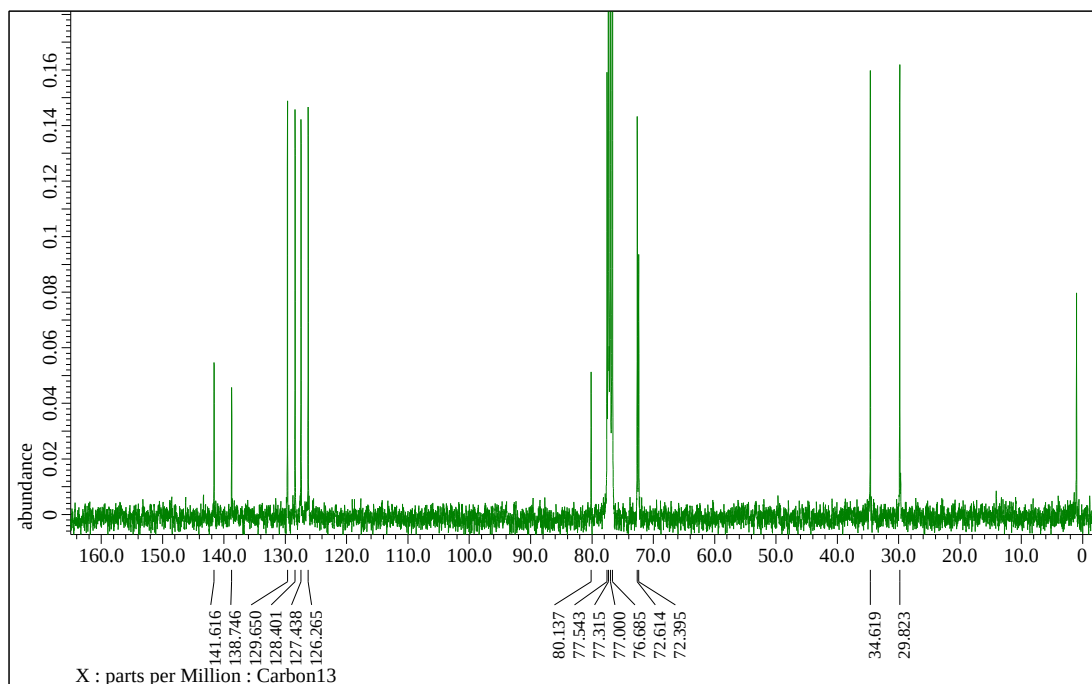
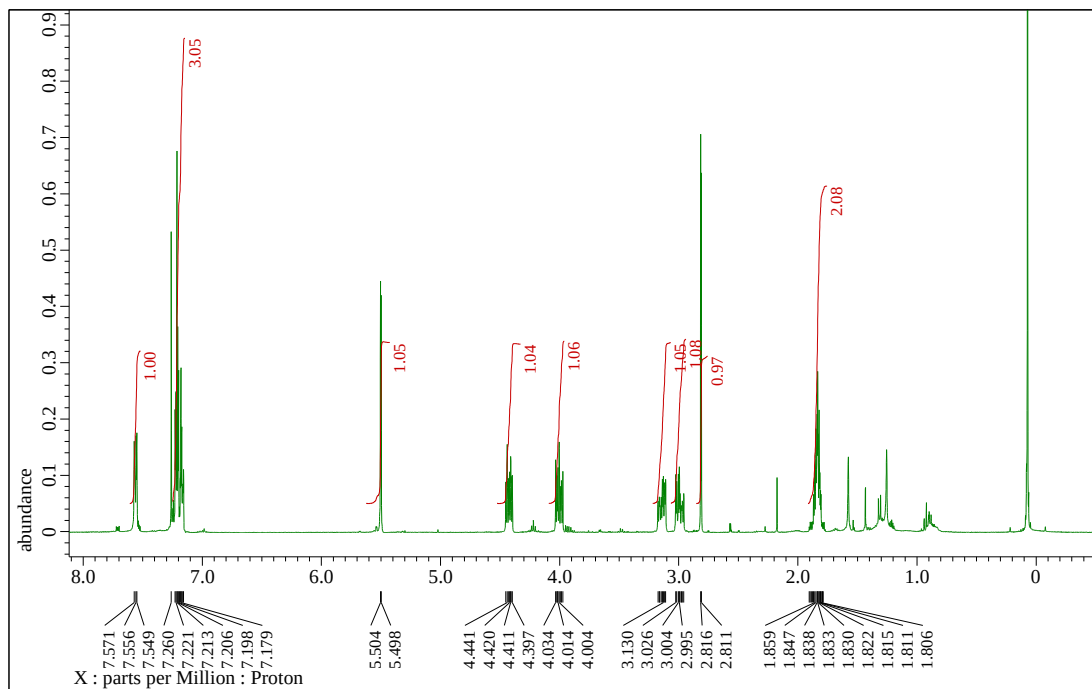


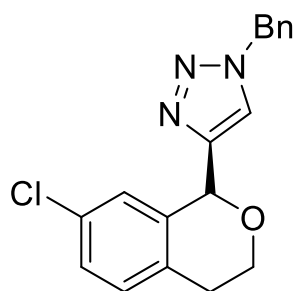
7



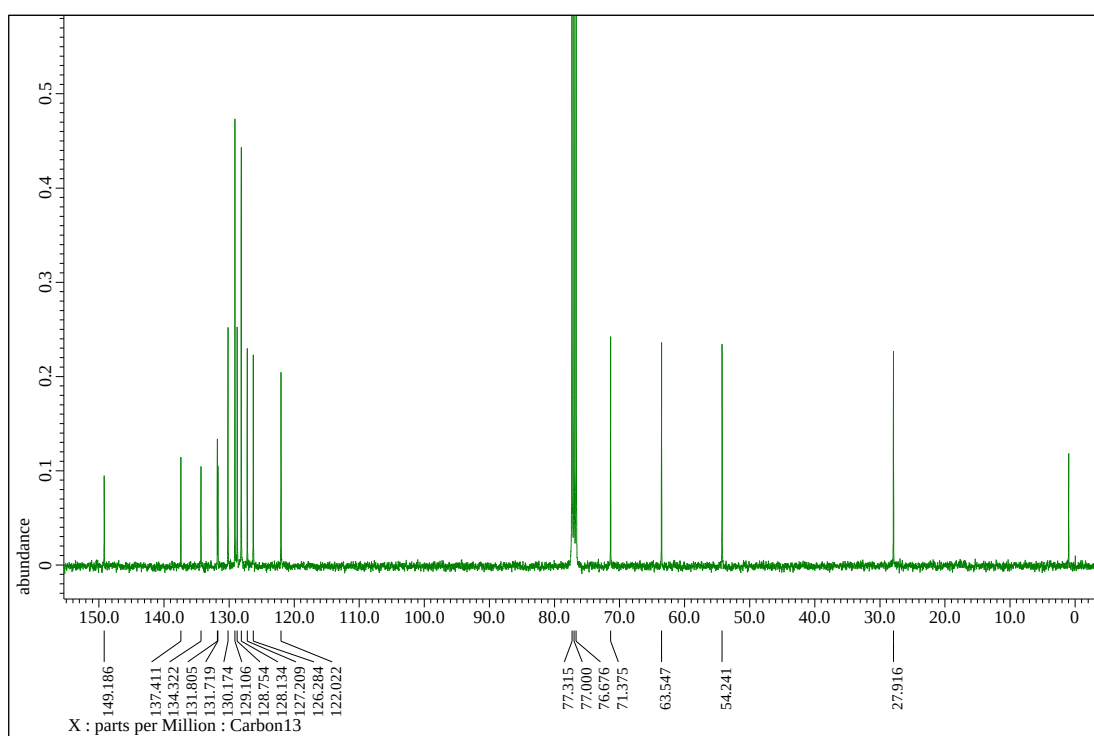
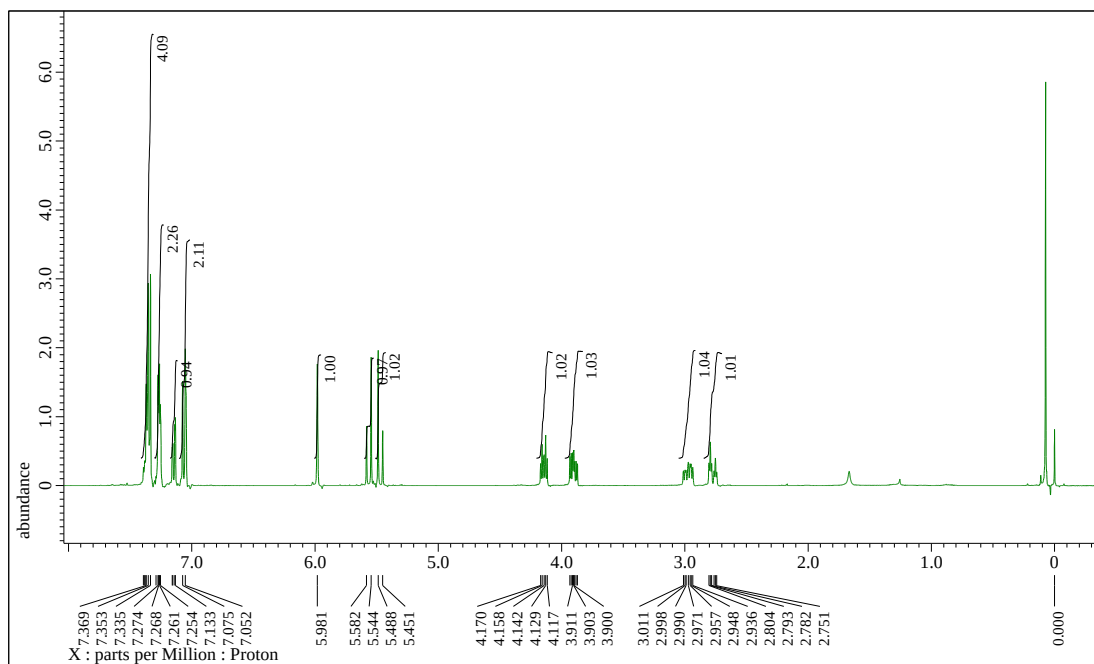


8





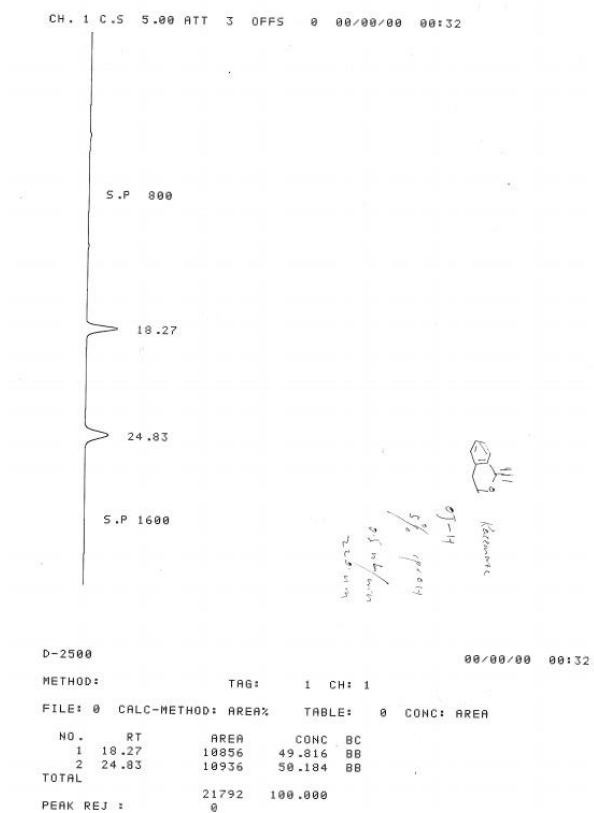
S16



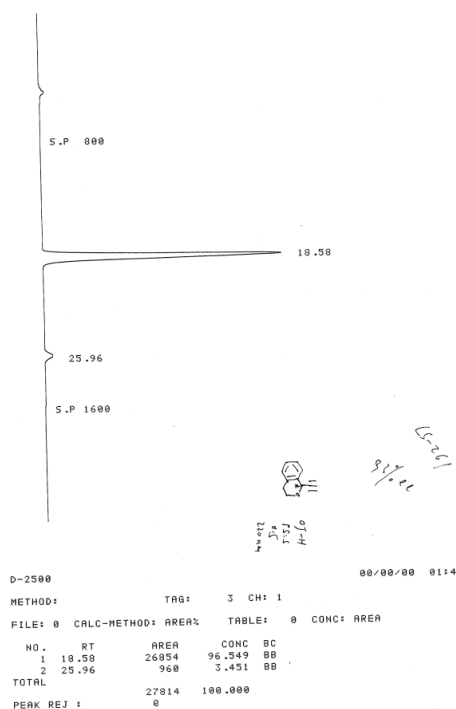
S51

HPLC Chart

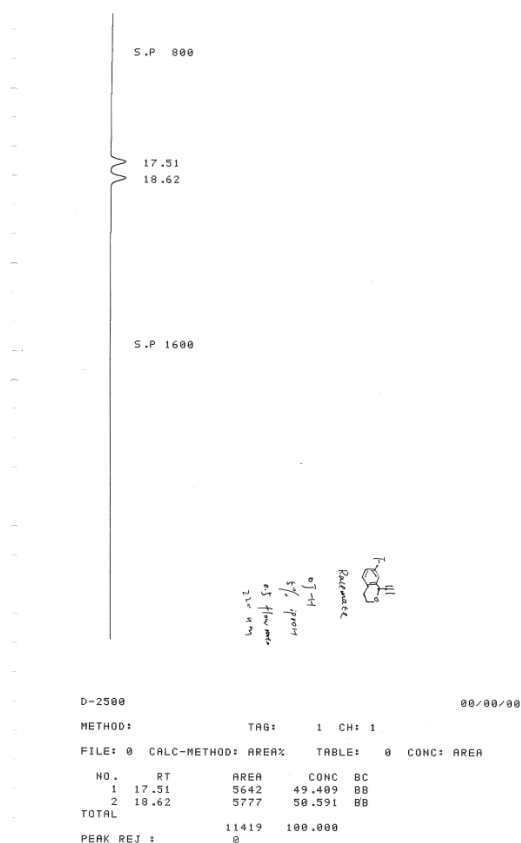
2a (rac)



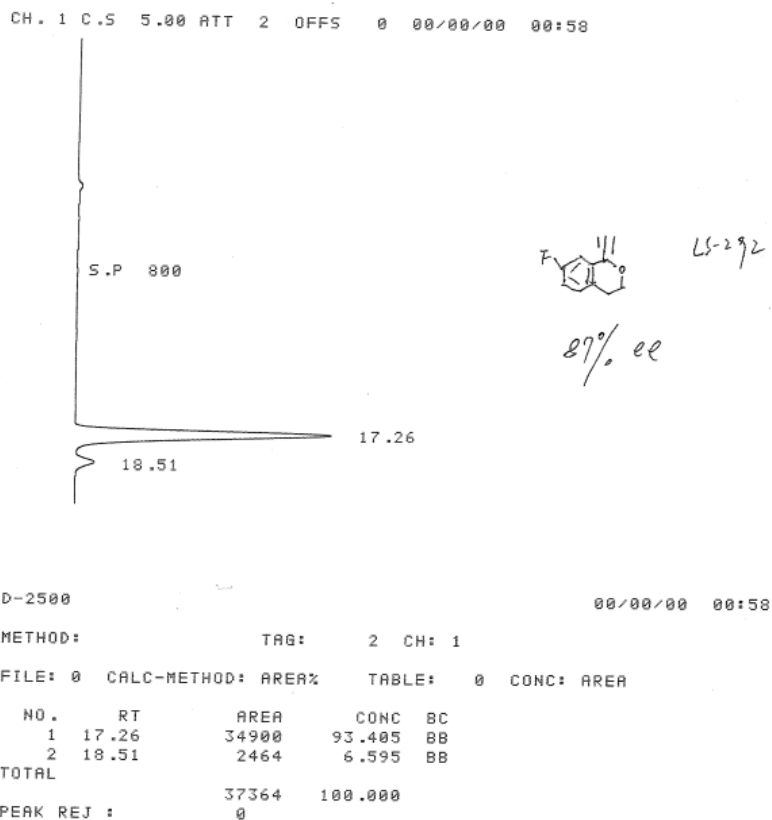
2a (chiral)



2b (rac)

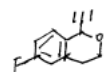
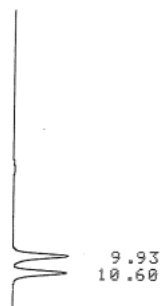


2b (chiral)



2c (rac)

CH. 1 C.S 5.00 ATT 3 OFFS 0 00/00/00 01:42



Racemate

D-2500

00/00/00 01:42

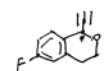
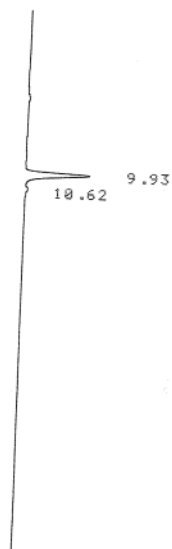
METHOD: TAG: 2 CH: 1

FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
1	9.93	7991	50.701	BB
2	10.60	7770	49.299	BB

TOTAL 15761 100.000
PEAK REJ : 0

2c (chiral)



02-14
5% iprocl
220 nm
0.3 flow rate

94% ee

D-2500

00/00/00 00:50

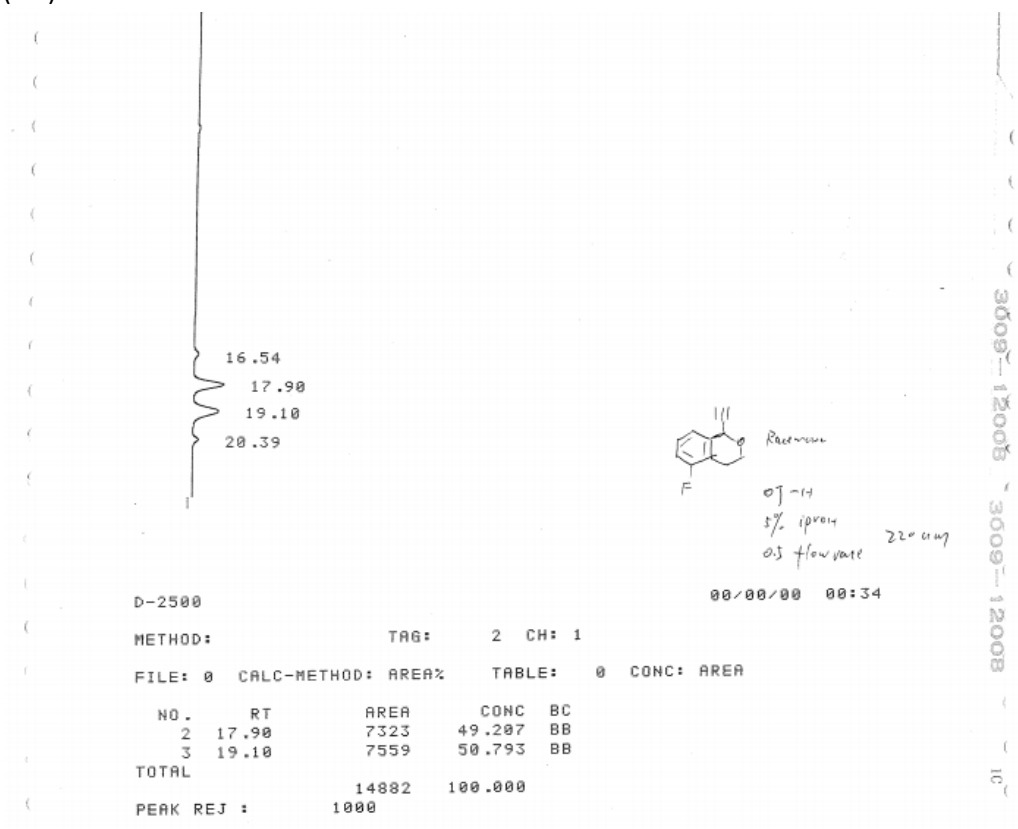
METHOD: TAG: 1 CH: 1

FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

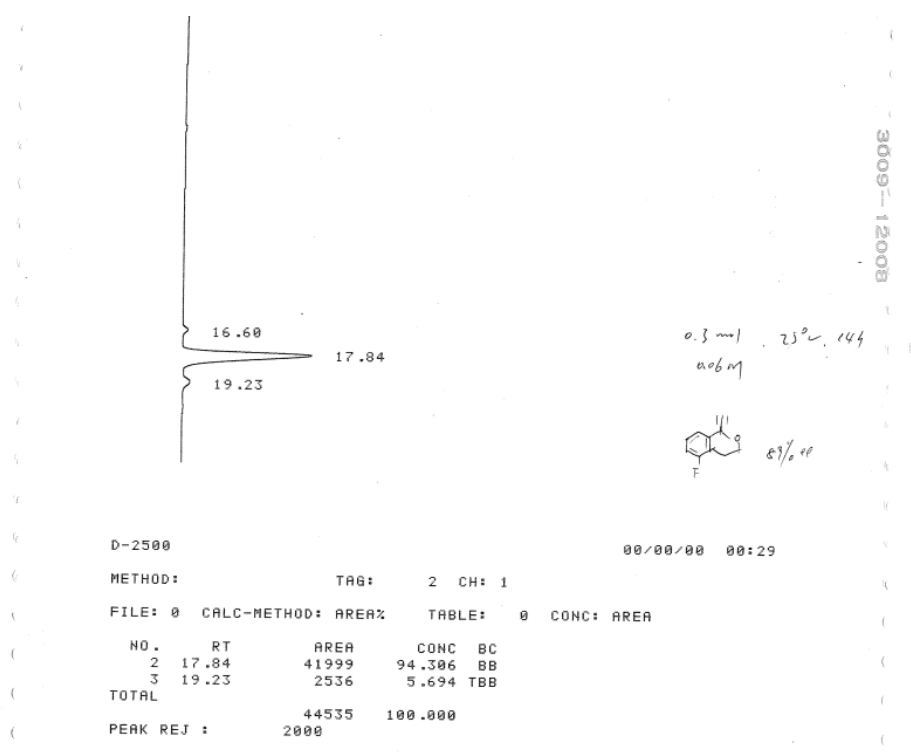
NO.	RT	AREA	CONC	BC
1	9.93	10312	96.936	BB
2	10.62	326	3.064	TBB

TOTAL 10638 100.000
PEAK REJ : 0

2d (rac)

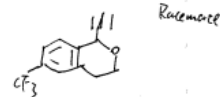
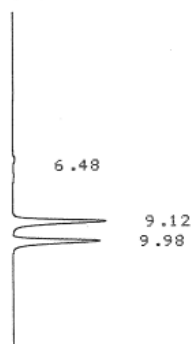


2d (chiral)



2e (rac)

CH. 1 C.S 5.00 ATT 3 OFFS 0 00/00/00 04:02



02-H
5% i-propyl
220 nm
0.5 flow rate

D-2500

00/00/00 04:02

METHOD: TAG: 4 CH: 1

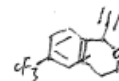
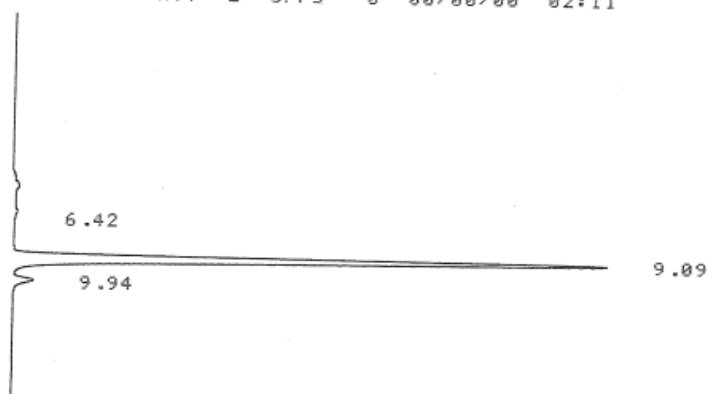
FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
2	9.12	12734	50.278	BB
3	9.98	12593	49.722	BB
TOTAL		25327	100.000	

PEAK REJ : 5000

2e (chiral)

CH. 1 C.S 5.00 ATT 2 OFFS 0 00/00/00 02:11



93% ee

D-2500

00/00/00 02:11

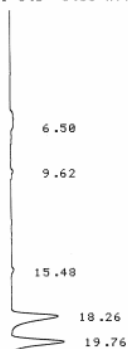
METHOD: TAG: 4 CH: 1

FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
2	9.89	35509	96.618	BB
3	9.94	1243	3.382	BB
TOTAL		36752	100.000	

PEAK REJ : 500

2f (rac)



07-19
5% protein
0.5 mL/min
220 nm

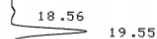
00/00/00 00:36

1 CH: 1

TABLE:

NO.	RT	AREA	CONC	BC
4	18.26	19276	49.272	BB
5	19.76	19846	50.728	BB
TOTAL				
		39122	100.000	
PEAK REJ :		5000		

2f (chiral)



92% ee

00/00/00 02:16

TAG:

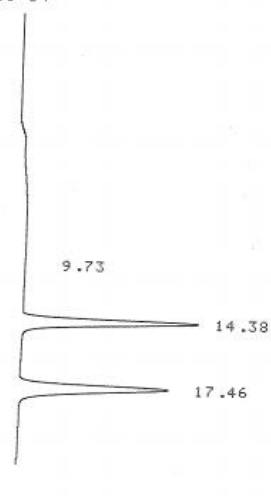
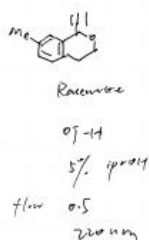
2 CH: 1

TABLE:	0	CONC:	AREA
--------	---	-------	------

NO.	RT	AREA	CONC	BC
1	18.56	948	3.880	BB
2	19.55	23488	96.120	BB
TOTAL		24436	100.000	

2g (rac)

CH. 1 C.S 5.00 ATT 4 OFFS 0 00/00/00 00:04



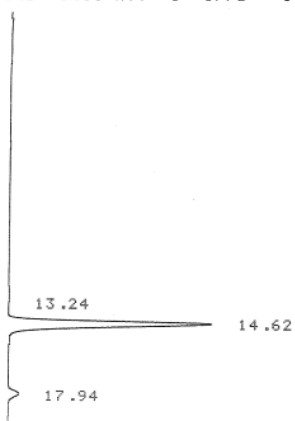
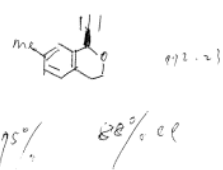
D-2500

00/00/00 00:04

NO.	RT	AREA	CONC	BC
2	14.38	78488	49.033	BB
3	17.46	81583	50.967	BB
TOTAL		160071	100.000	
PEAK REJ :		20000		

2g (chiral)

CH. 1 C.S 5.00 ATT 3 OFFS 0 00/00/00 00:36

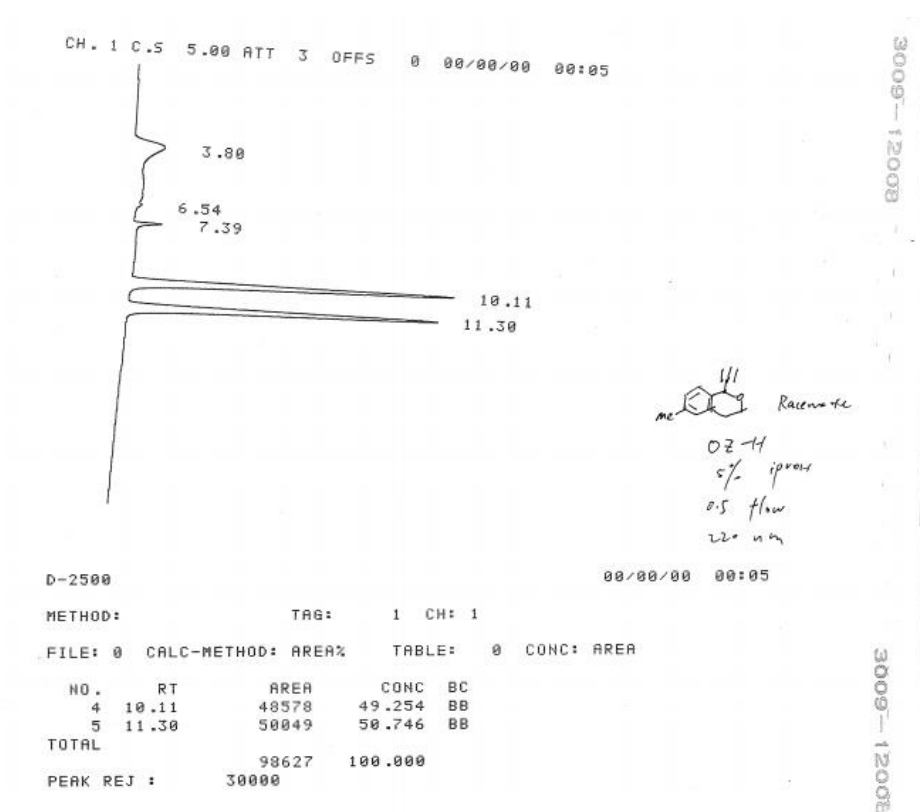


D-2500

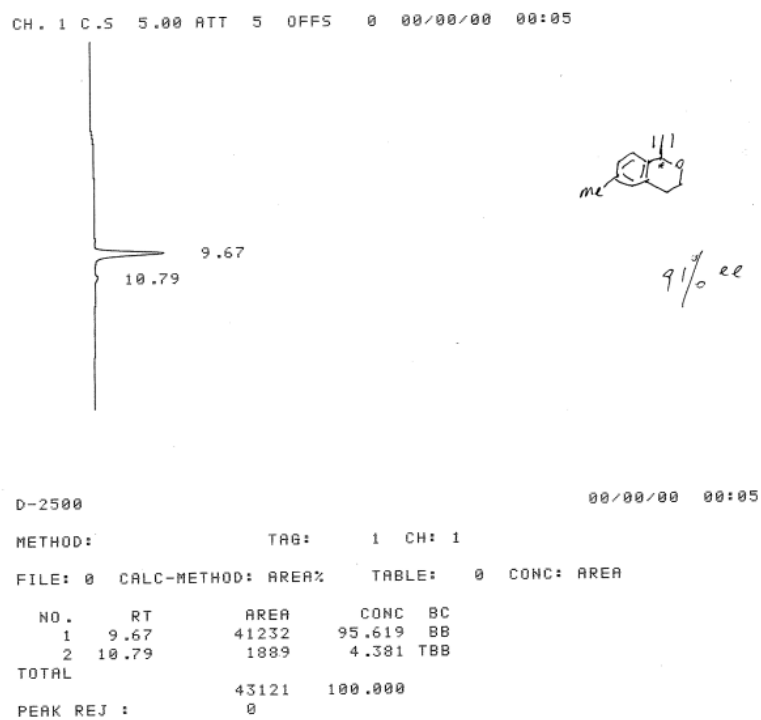
00/00/00 00:36

NO.	RT	AREA	CONC	BC
2	14.62	44736	93.833	BB
3	17.94	2940	6.167	BB
TOTAL		47676	100.000	
PEAK REJ :		2000		

2h (rac)

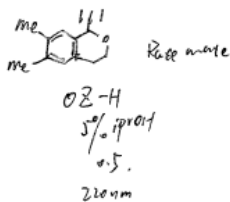
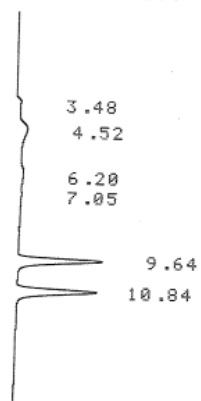


2h (chiral)



2i (rac)

CH. 1 C.S 5.00 ATT 5 OFFS 0 00/00/00 00:04



D-2500

00/00/00 00:04

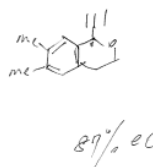
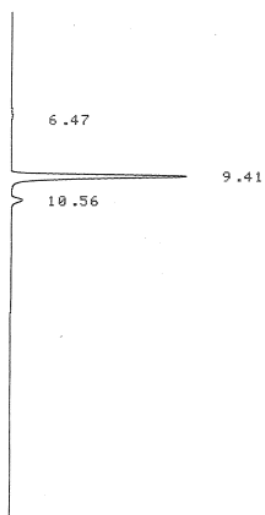
METHOD:

TAG: 1 CH: 1

FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
5	9.64	41159	49.541	BB
6	10.84	41922	50.459	BB
TOTAL		83081	100.000	
PEAK REJ :		10000		

2i (chiral)



D-2500

00/00/00 01:13

METHOD:

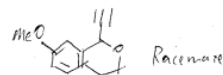
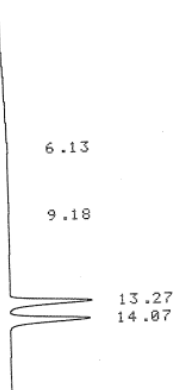
TAG: 1 CH: 1

FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
2	9.41	55516	93.617	BB
3	10.56	3785	6.383	BB
TOTAL		59301	100.000	
PEAK REJ :		500		

2j (rac)

CH. 1 C.S 5.00 ATT 5 OFFS 0 00/00/00 00:04



02-H
5% iprOH
220 nm
0.5 flow rate

3009-12008

D-2500

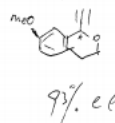
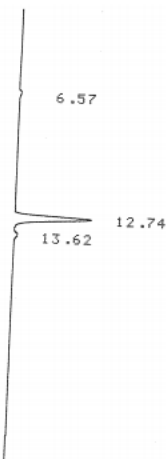
00/00/00 00:04

METHOD: TAG: 1 CH: 1
FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
3	13.27	55966	49.690	BU
4	14.07	56664	50.310	UB
TOTAL		112630	100.000	

PEAK REJ : 10000

2j (chiral)



3009-12008

D-2500

00/00/00 00:58

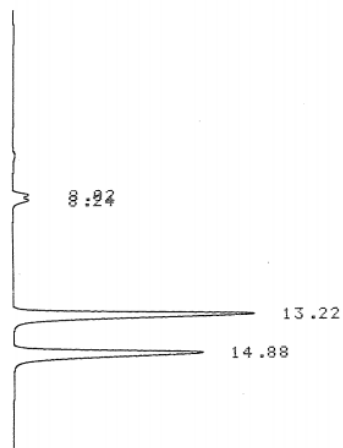
METHOD: TAG: 1 CH: 1
FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
2	12.74	15176	96.270	BB
3	13.62	588	3.730	TBB
TOTAL		15764	100.000	

PEAK REJ : 400

2k (rac)

CH. 1 C.S 5.00 ATT 3 OFFS 0 00/00/00 00:30



Racemate

0.5-1.4

5% isoproh

0.5 mL/min

220 nm

D-2500

00/00/00 00:30

METHOD: TAG: 1 CH: 1

FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

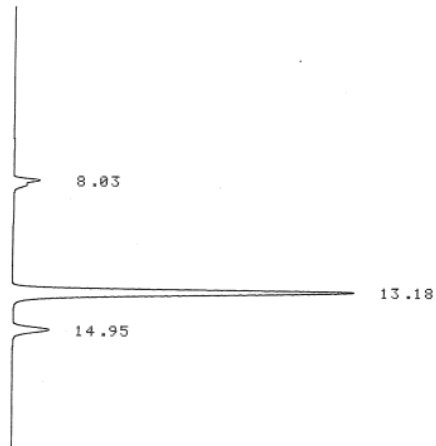
NO.	RT	AREA	CONC	BC
3	13.22	45208	53.318	BB
4	14.88	39582	46.682	BB

TOTAL 84790 100.000

PEAK REJ : 5000

2k (chiral)

CH. 1 C.S 5.00 ATT 3 OFFS 0 00/00/00 00:38



60% ee

D-2500

00/00/00 00:38

METHOD: TAG: 1 CH: 1

FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
2	13.18	71827	90.100	BB
3	14.95	7892	9.900	BB

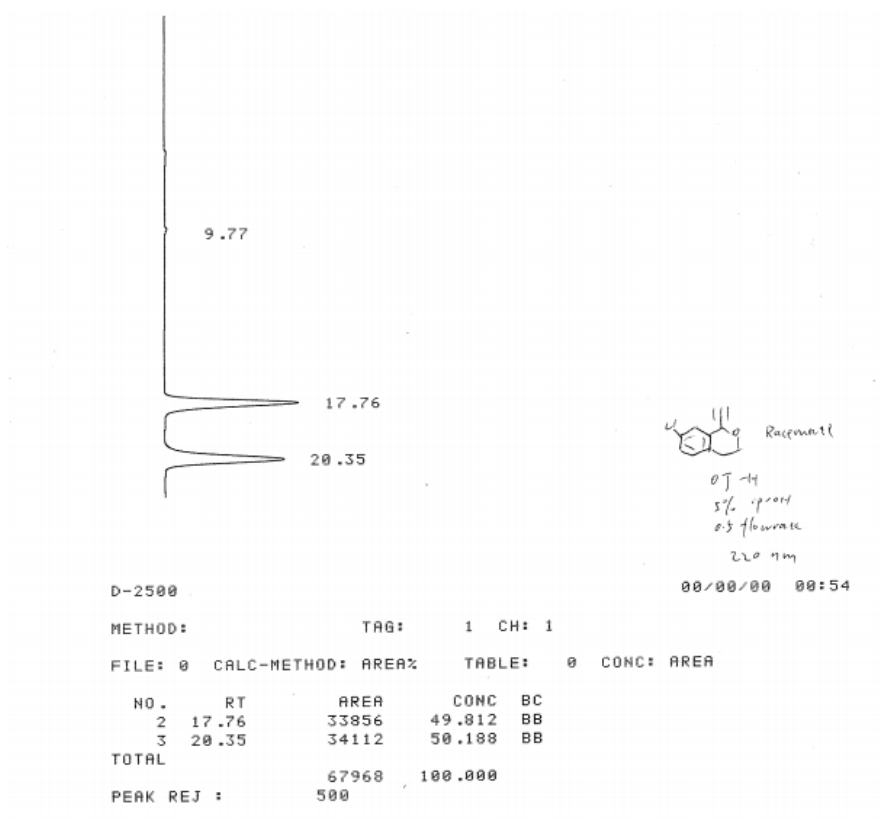
TOTAL 79719 100.000

PEAK REJ : 6000

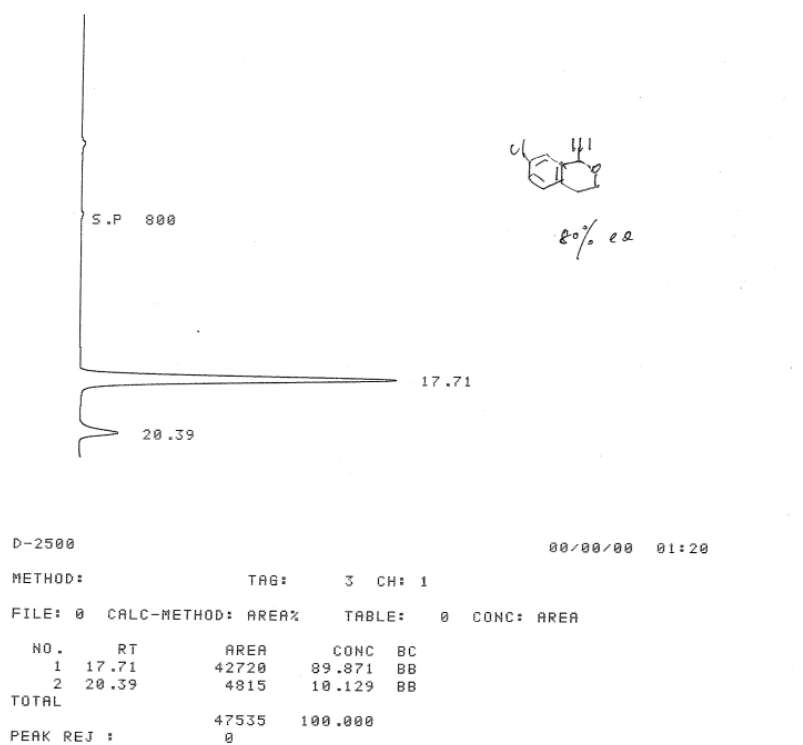
3009-12008

IC

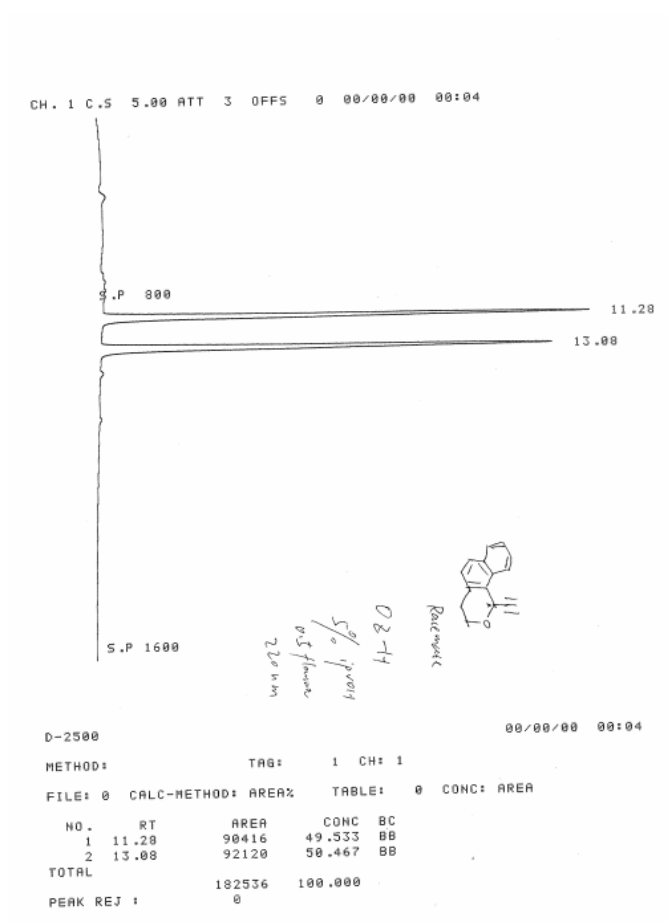
2I (rac)



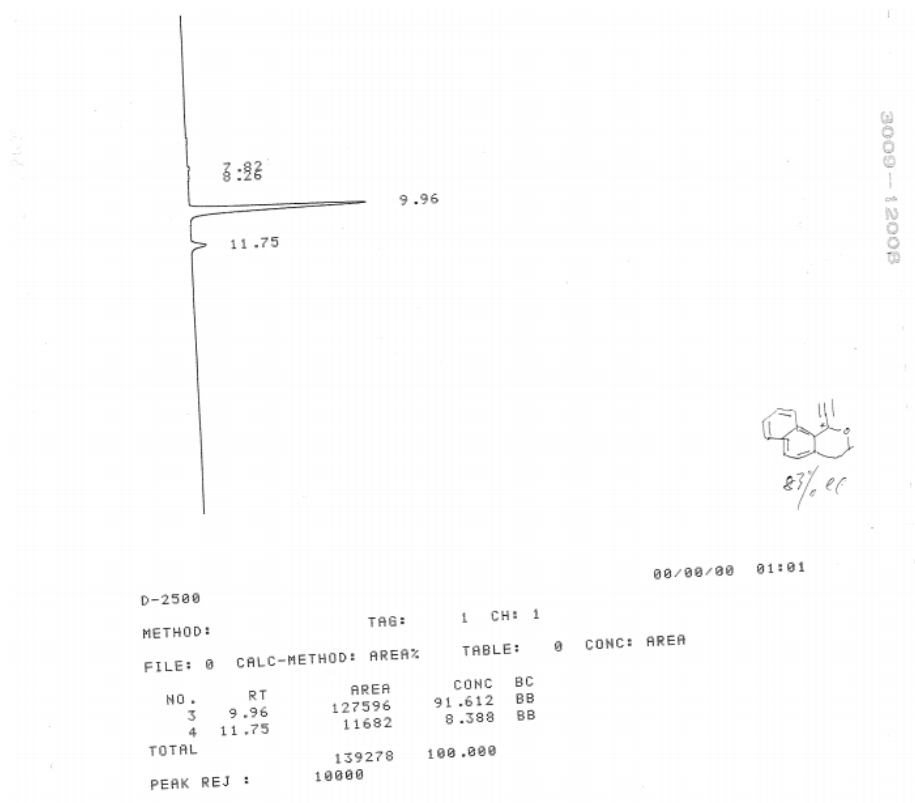
2I (chiral)



2m (rac)

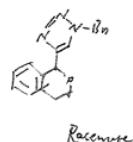
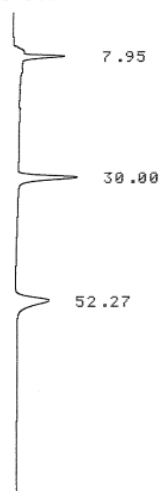


2m (chiral)



3 (rac)

CH. 1 C.S 1.25 ATT 3 OFFS 0 00/00/00 01:09



0J-H
3% iPrOH
0.3 mL/min
220 nm

D-2500

00/00/00 01:09

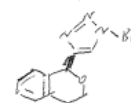
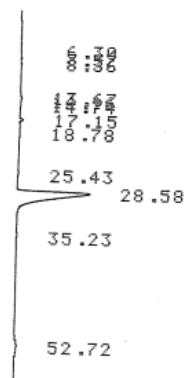
METHOD: TAG: 2 CH: 1

FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
2	30.00	35600	49.267	BB
3	52.27	36660	50.733	BB
TOTAL		72260	100.000	
PEAK REJ :		20000		

3 (chiral)

CH. 1 C.S 1.25 ATT 6 OFFS 0 00/00/00 00:04



83% ee

D-2500

00/00/00 00:04

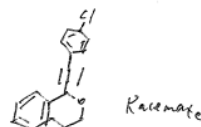
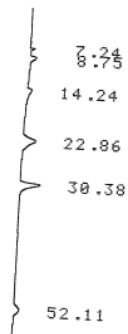
METHOD: TAG: 1 CH: 1

FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
10	28.58	411885	94.699	BB
12	52.72	23056	5.301	BB
TOTAL		434941	100.000	
PEAK REJ :		20000		

4 (rac)

CH. 1 C.S 1.25 ATT 5 OFFS 0 00/00/00 00:06



0J-H
0.5 ml/min
5% iproh
220 nm

D-2500

METHOD:

TAG: 1 CH: 1

FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

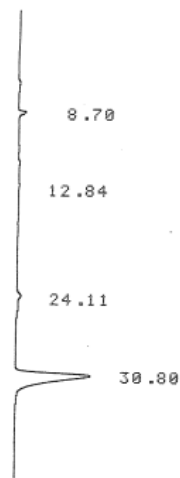
NO.	RT	AREA	CONC	BC
4	22.86	43763	49.874	BB
5	30.38	43984	50.126	BB

TOTAL 87747 100.000
PEAK REJ : 20000

00/00/00 00:06

4 (chiral)

CH. 1 C.S 2.50 ATT 3 OFFS 0 00/00/00 01:16



92.5% ee

D-2500

00/00/00 01:16

METHOD:

TAG: 2 CH: 1

FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
3	24.11	1496	3.706	BB
4	30.80	38873	96.294	BB

TOTAL 40369 100.000
PEAK REJ : 1400

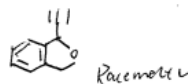
7 (rac)

CH. 1 C.S 2.50 ATT 2 OFFS 0 00/00/00 00:54

S.P 800

20.88

27.40



0J-H

5% iproh

220 nm
0.5 ml/min

00/00/00 00:54

D-2500

METHOD: TAG: 2 CH: 1

FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
1	20.88	1685	48.940	BB
2	27.40	1758	51.060	BB
TOTAL		3443	100.000	
PEAK REJ :		0		

7 (chiral)

CH. 1 C.S 2.50 ATT 2 OFFS 0 00/00/00 00:37

S.P 800

20.79

27.46



D-2500

00/00/00 00:37

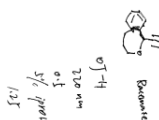
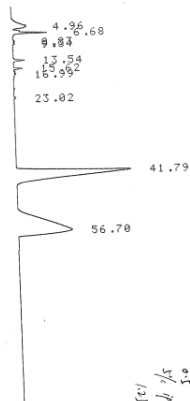
METHOD: TAG: 1 CH: 1

FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
1	20.79	18616	83.973	BB
2	27.46	3553	16.027	BB
TOTAL		22169	100.000	
PEAK REJ :		0		

8 (rac)

CH. 1 C.S 1.25 ATT 3 OFFS 0 00/00/00 01:48



D-2500 00/00/00
 METHOD: TAG: 2 CH: 1
 FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
9	41.79	190720	50.759	BB
10	56.70	185017	49.241	BB
TOTAL		375737	100.000	

 PEAK REJ : 100000

8 (chiral)

CH. 1 C.S 1.25 ATT 3 OFFS 0 00/00/00 01:36

S.P 800

S.P 1600

43.12

52.54

S.P 3200



Handwritten notes: 15-12-07, 28, 20.02

D-2500 00/00/00 01:
 METHOD: TAG: 2 CH: 1
 FILE: 0 CALC-METHOD: AREA% TABLE: 0 CONC: AREA

NO.	RT	AREA	CONC	BC
1	43.12	10350	35.322	BB
2	52.54	18952	64.678	BB
TOTAL		29302	100.000	

 PEAK REJ : 1000