

## Electronic Supplementary Information

### A turn-on fluorescent probe with a dansyl fluorophore for hydrogen sulfide sensing

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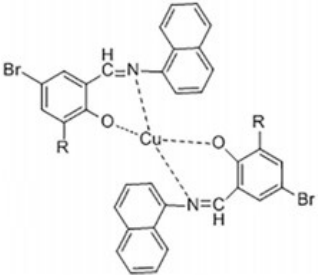
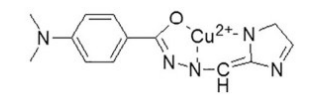
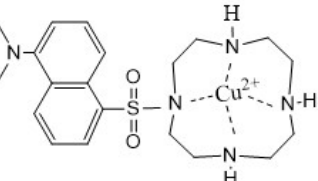
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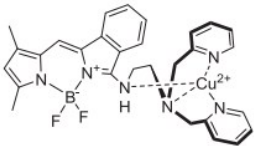
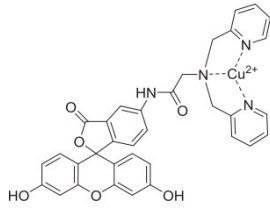
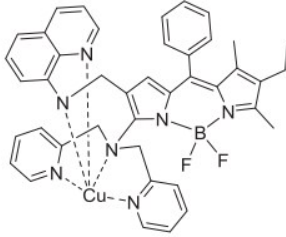
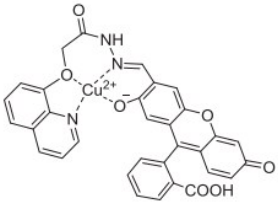
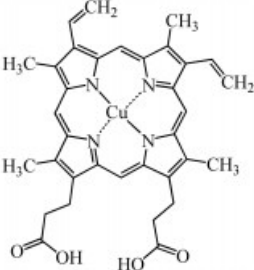
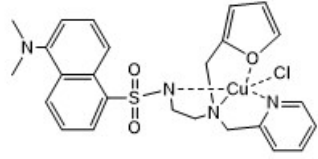
**Corresponding author:**

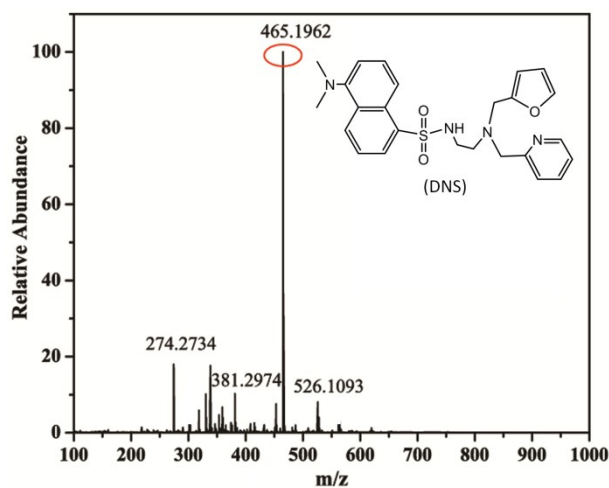
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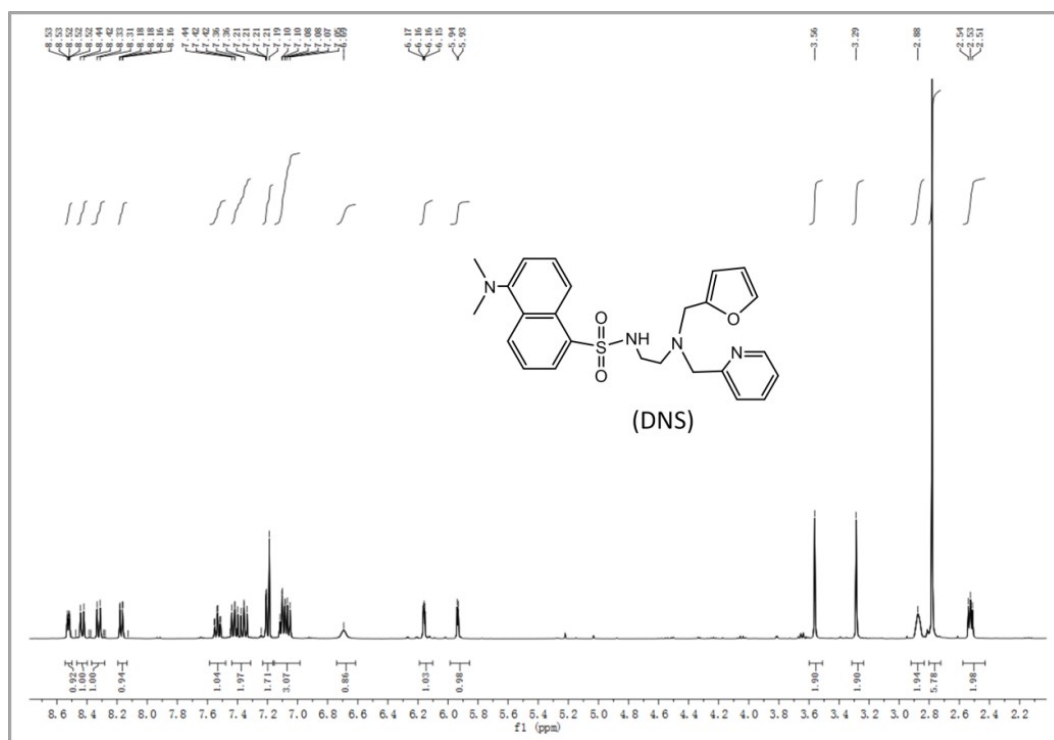
**Table S1.** The comparison of probe based copper metal complexes for H<sub>2</sub>S. In terms of their solvent, excitation wavelength (Ex) and emission wavelength (Em) or the maximum absorption wavelength, limit of detection (LOD), cell imaging, H<sub>2</sub>S gas test.

| NO. | Probe  | Solvent  | Ex/Em<br>or $\lambda_{max}$<br>(nm)           | LOD   | Cell<br>imaging | H <sub>2</sub> S<br>gas<br>test |
|-----|--|--|---|---|-----------------|---------------------------------|
| 1   | Ves-1.Cu-EY  | HEPES<br>or<br>PBS                                 | 515/540 nm<br>or 564 nm                       | 0.59 $\mu$ M<br>HEPES/<br>4.06 $\mu$ M<br>PBS | Yes             | No                              |
| 2   | naphthalimide-rhodamine B-<br>Cu derivative<br><i>colorimetric and fluorescent</i>   | CH <sub>3</sub> CN-H <sub>2</sub> O<br>(v/v = 7:3) | 325/ 528 and<br>610 nm<br>or 564 nm           | 0.23 $\mu$ M                                  | Yes             | No                              |
| 3   | <br><i>colorimetric and fluorescent</i> | DMSO-H <sub>2</sub> O<br>(v/v = 9:1)               | 337/430 nm<br>or<br>shift 390 nm to<br>530 nm | 0.13 $\mu$ M                                  | No              | No                              |
| 4   | <br><i>colorimetric and fluorescent</i> | CH <sub>3</sub> CN/H <sub>2</sub> O<br>(v/v = 3:2) | 350/470 nm<br>or<br>shift 520 to 569<br>nm    | 0.68 $\mu$ M                                  | No              | No                              |
| 5   |   | PBS<br>(10% DMSO)                                  | --/491 nm                                     | 4.07 $\mu$ M                                  | Yes             | No                              |

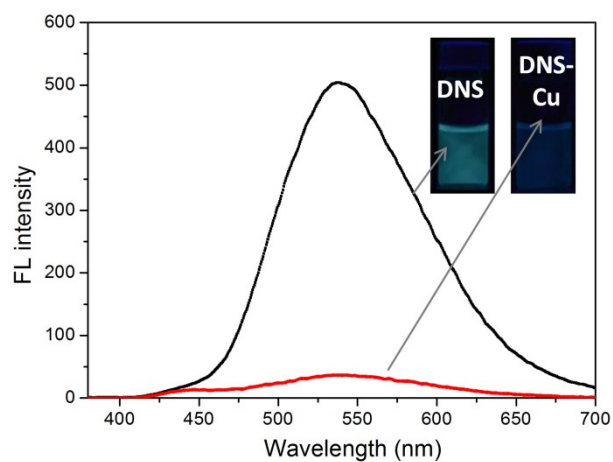
|    |   |  |                           |        |     |     |
|----|---|--|---------------------------|--------|-----|-----|
| 6  |                      | PBS<br>pH 7.4  | 480 /546 nm               | 250 nM | Yes | No  |
| 7  |                      | H <sub>2</sub> O   | 470/517 nm                | 420 nM | No  | No  |
| 8  | <br><br>colorimetric | HEPES<br>(10% DMSO)  | shift 520 nm to<br>569 nm | 167 nM | No  | No  |
| 9  |                    | PBS/CH <sub>3</sub> CN<br>(1:1, v/v, pH<br>7.2)                  | 495/534 nm                | ---    | Yes | No  |
| 10 |                    | H <sub>2</sub> O   | 386/623 and 682<br>nm     | 1 μM   | NO  | NO  |
| 11 | <br><br>This work  | C <sub>2</sub> H <sub>5</sub> OH/H <sub>2</sub> O<br>(v/v = 1:1) | 338/534 nm                | 11 nM  | No  | Yes |



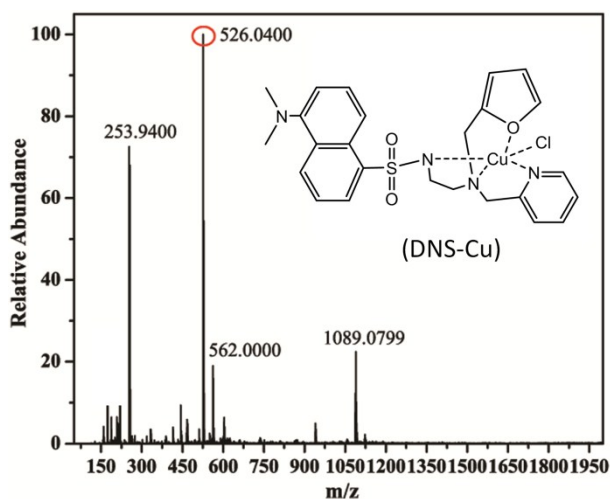
**Fig. S1.** ESI-Mass spectrum (positive mode) of DNS.



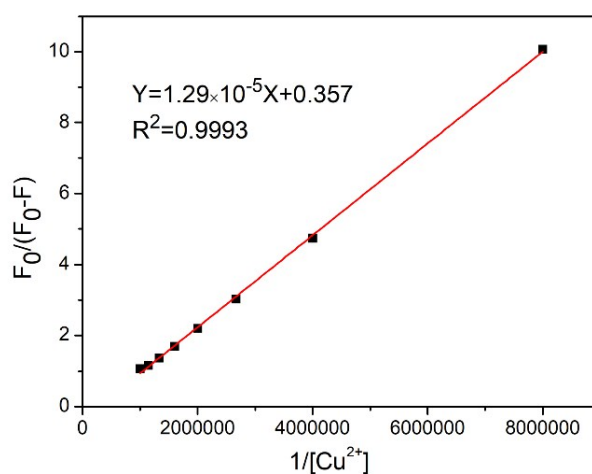
**Fig. S2.** <sup>1</sup>H-NMR spectrum of the DNS. <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>) δ 8.56-8.51 (m, 1H), 8.43 (d, *J* = 8.5 Hz, 1H), 8.32 (d, *J* = 8.7 Hz, 1H), 7.53 (td, *J* = 7.7, 1.8 Hz, 1H), 7.45-7.39 (m, 1H), 7.36 (dd, *J* = 8.6, 7.6 Hz, 1H), 7.21 (dd, *J* = 1.8, 0.8 Hz, 1H), 7.13-7.03 (m, 3H), 6.69 (s, 1H), 6.15 (dt, *J* = 9.9, 4.9 Hz, 1H), 5.94 (d, *J* = 2.9 Hz, 1H), 3.56 (s, 2H), 3.20 (s, 2H), 2.82-2.92 (m, 2H), 2.75 (s, 6H), 2.53 ppm (t, *J* = 4 Hz, 2H).



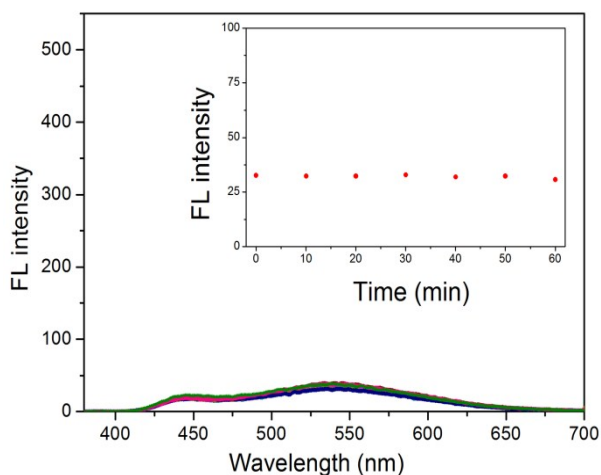
**Fig. S3.** The fluorescence spectra of DNS and the probe complex DNS-Cu. The insets are their corresponding photographs.



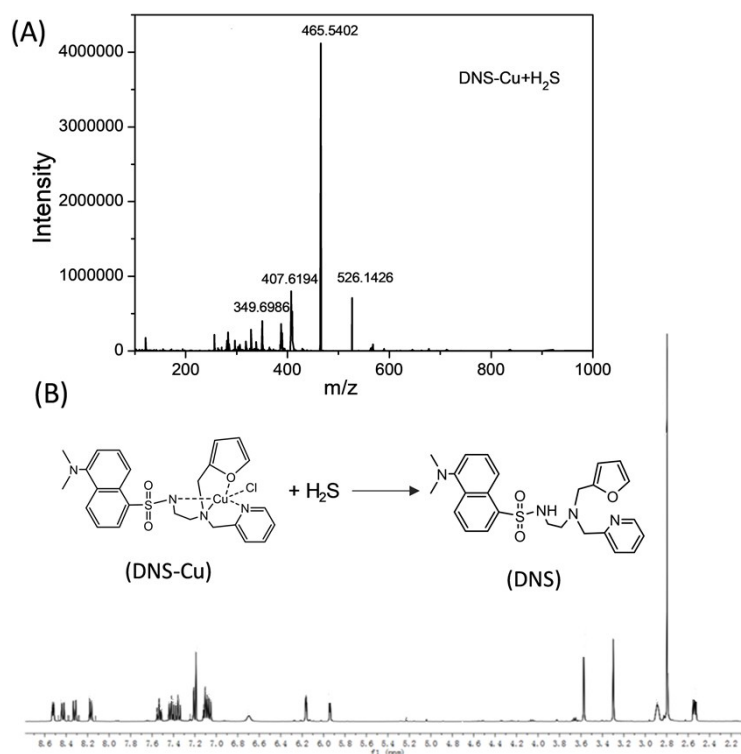
**Fig. S4.** The mass spectrum (positive mode) of DNS-Cu complex probe.



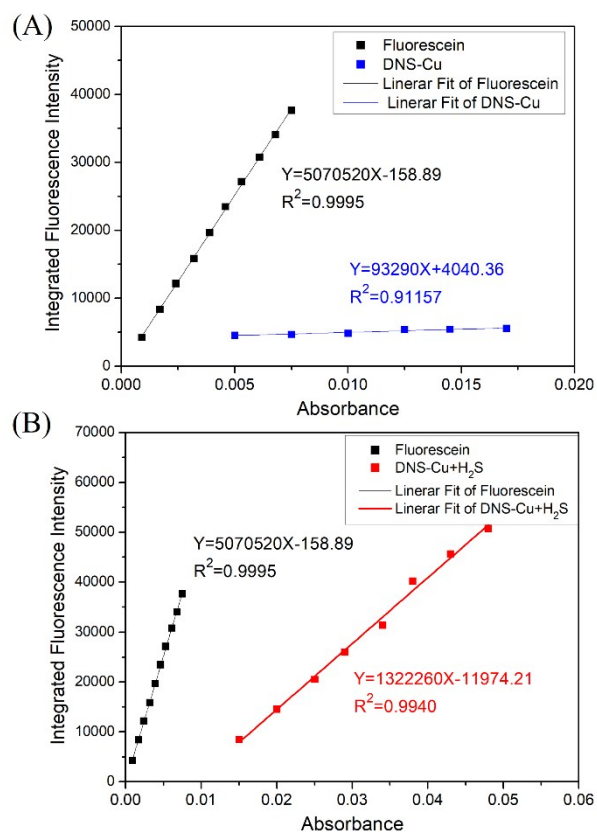
**Fig. S5.** Benesi-Hildebrand plot for determining the binding constant K of DNS-Cu complex.



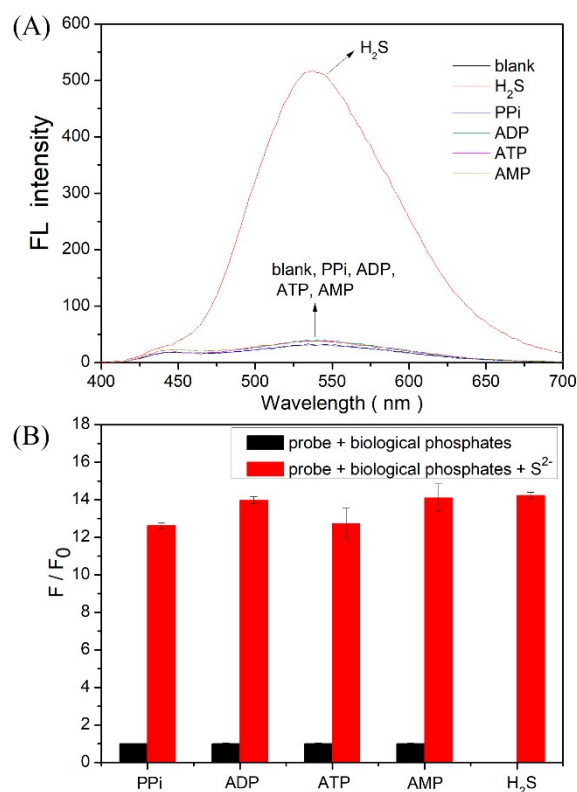
**Fig. S6.** The fluorescence spectra of probe recorded each 10 min for 1 h under ultraviolet irradiation at 338 nm. The fluorescence intensity has no significant change, implying probe exhibit good stability against photo-bleaching.



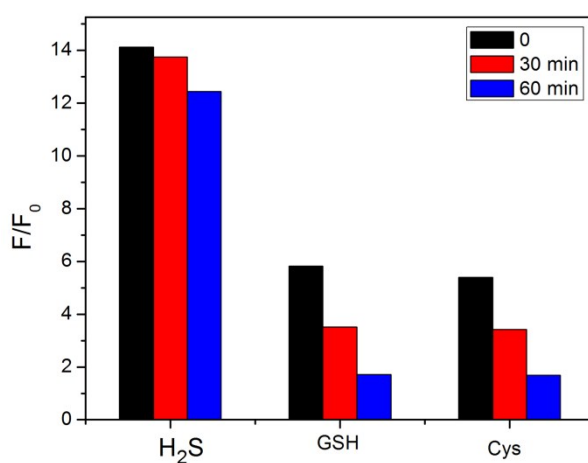
**Fig. S7.** (A) The mass spectrum (positive mode) and (B) <sup>1</sup>H-NMR spectrum (400 MHz, CDCl<sub>3</sub>) of the DNS-Cu complex after reaction with H<sub>2</sub>S. Clearly, the characteristic <sup>1</sup>H-NMR spectrum of DNS was restored.



**Fig. S8** Determination of the fluorescence quantum yield of (A) the DNS-Cu (QY, 1.8%) (B) after reaction with H<sub>2</sub>S (QY, 25.5%), the fluorescein ( $\Phi_s=0.95$  in 0.1 M NaOH) as reference standard.



**Fig. S9.** (A) Fluorescence spectra of the DNS–Cu probe in presence of S<sup>2-</sup> ions (1.0 μM) and other biological phosphates P<sub>Pi</sub>, ATP, ADP and AMP (1.0 μM). (B) The effect of the biological phosphates on the fluorescence intensity of DNS–Cu complex. The black bars represent the probe in the presence of P<sub>Pi</sub>, ATP, ADP and AMP, the red bars represent the subsequent addition of S<sup>2-</sup> into the mixture solution. The concentration of S<sup>2-</sup> was 1.0 μM, and the concentration of other species was 10 μM.





**Fig. S10.** Fluorescence responses of the probe (1.0  $\mu\text{M}$ ) towards 1.0  $\mu\text{M}$  of  $\text{H}_2\text{S}$ , GSH, and Cys, respectively. The black bars represent the addition of analyte without any pretreatment. The red bars and blue bars represent the pretreatment with DMSO at 60°C for 30 min and 60 min, respectively. Clearly, the pretreatment of GSH and Cys with DMSO greatly decrease their interference on the detection of hydrogen sulfide.

## References

1. R. Kaushik, R. Sakla, A. Ghosh, G.T. Selvan, P. M. Selvakumar and D. A. Jose, *ACS Sens.* 2018, **3**, 1142-1148.
2. S. Wang, H.C. Ding, Y.S. Wang, C.B. Fan, Y.Y. Tu, G. Liu and S.Z. Pu, *RSC Adv.*, 2018, **8**, 33121–33128.
3. X. F. Shanga, X. J. Yueb, Y.M. Chena, C. S. Lia, H. L. Chenc and T.Y. Wang, *Inorg. Chem. Commun.*, 2019, **99**, 1–10.
4. H. Fang, P. C. Huang and F. Y. Wu, *Spectrochim. Acta, Part A*, 2018, **204**, 568–57.
5. J. Y. Kim, S. Sarkar, K. N. Bobba, P. T. Huynh, A. Bhise and J. Yoo, *Org. Biomol. Chem.* 2019, DOI: 10.1039/c9ob00948e.
6. X. Li, Y. Gong, K. Wu, S.H. Liang, J. Cao, B. Yang, Y. Hu and Y. Han, *RSC Adv.* 2014, **4**, 36106–36109.
7. M.G. Choi, S. Cha, H. Lee, H.L. Jeon and S. K. Chang, *Chem. Commun.* 2009, **47**, 7390–7392.
8. X. Gu, C. Liu, Y.C. Zhu and Y. Z. Zhu, *Tetrahedron Lett.* 2011, **52**, 5000–5003.
9. F. Hou, L. Huang, P. Xi, J. Cheng, X. Zhao, G. Xie, Y. Shi, F. Cheng, X. Yao, D. Bai, Z. Zeng, *Inorg. Chem.*, 2012, **51**, 2454–2460.
10. S. Mirra, S. Milione, M. Strianese and C. Pellicchia, *Eur. J. Inorg. Chem.* 2015, 2272–2276.