



## RSC Advances

### PAPER

# Hydrothermal synthesis, characterization and enhanced photocatalytic activity and toxicity studies of rhombohedral Fe<sub>2</sub>O<sub>3</sub> nanomaterial

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### SUPPLEMENTARY INFORMATION

#### RESEARCH HIGHLIGHTS

1. Facile Hydrothermal synthesis of Rhombohedral Fe<sub>2</sub>O<sub>3</sub> for multi-functional application with eco-friendly approaches to compare already reported papers.
2. The synthesized material has high structural uniqueness, high surface area and narrow band gap for the increased photocatalytic performance and recycling performance after photodegradation of Rhodamine-B (RB). The XRD patterns of Rhodamine-B (RB): before and after successful cycles recycling processes, it gives a clear picture on recyclability of the material.
3. First report on, the recycled material was used for toxicity assessments like, histological alterations of the heart tissue of *Albino wistar* rat, *Musmusculus* skin melanoma cells (B16-F10), Human embryonic kidney (HEK)-293, and *Escherichia coli*, (MTCC 7410) bacterial cell wall damage for clear understand of before and after mechanism of material, and this material is safest and the superlative alternative in terms of toxicity in biological application in future
4. Finally, the combination of materials was high photocatalytic stability, non-toxicity and efficient up-conversion properties in the synthesized material paves the way for its applications in photocatalysis, energy conversion and in the biological field as well as environmental degradation and these results are evident gives an encouraging result to compare to already reported materials.

**LIST OF TABLES****Table S1** The elemental composition of Rhombohedral Fe<sub>2</sub>O<sub>3</sub> photocatalyst determined from energy dispersive spectroscopy (EDS)

Element	Weight (%)	Atomic (%)	compound
O K	197.03	51.90	270.99
Fe L	293.61	22.15	0.00
C	73.96	25.95	0.00
<b>Total</b>		<b>564.60</b>	

**Table S2** Surface (BET) and pore volume parameters of Rhombohedral Fe<sub>2</sub>O<sub>3</sub>

Photocatalyst	S <sub>BET</sub> (m <sup>2</sup> g <sup>-1</sup> )	Pore volume (cm <sup>3</sup> g <sup>-1</sup> )
Fe <sub>2</sub> O <sub>3</sub>	5.676	1.304

**Table S3** Electro-negativity, band-gap, conduction band (CB) edge and valence band (VB) edge potential of the catalysts on normalized hydrogen scale of Rhombohedral Fe<sub>2</sub>O<sub>3</sub>

Semiconductor catalyst	$\chi$ (eV)	E <sub>g</sub> (eV)	E <sub>CB</sub> (eV)	E <sub>VB</sub> (eV)
Fe <sub>2</sub> O <sub>3</sub>	5.89	1.96	-0.22	2.354

**Table S4** The Total organic carbon (TOC) and Chemical oxygen demand (COD) results of Rhodamine - B against Rhombohedral Fe<sub>2</sub>O<sub>3</sub>

Time (min)	COD (ppm)	TOC (ppm)
00	149.12	242.05
20	117.39	170.27
40	89.74	97.64
60	34.97	56.37
80	20.41	11.22
100	3.54	6.31
120	1.39	2.97

**Table S5** The Anti-bacterial activity of Fe<sub>2</sub>O<sub>3</sub> nanoparticle against *E. coli*

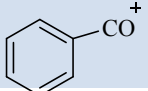
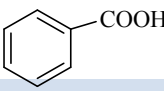
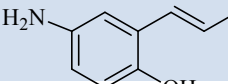
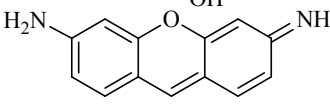
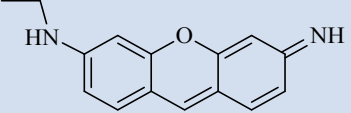
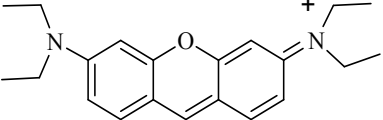
Test Sample	Zone of inhibition (in mm)		
	<i>E. coli</i> (0.3 mg)	<i>E. coli</i> (0.6 mg)	<i>E. coli</i> (1.0 mg)
Fe <sub>2</sub> O <sub>3</sub>	15.81 ± 0.15	18.35 ± 0.08	20.60 ± 0.24
Std. DW	00.00 ± 0.00	00.00 ± 0.00	00.00 ± 0.00

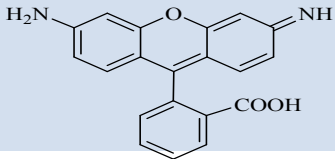
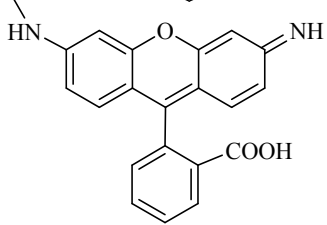
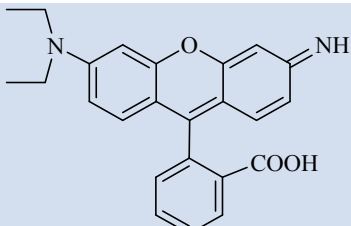
**Note:** Values are means of three independent replicates; ± indicate standard error.

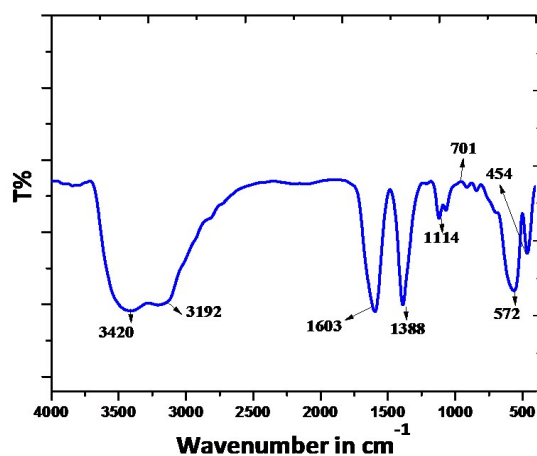
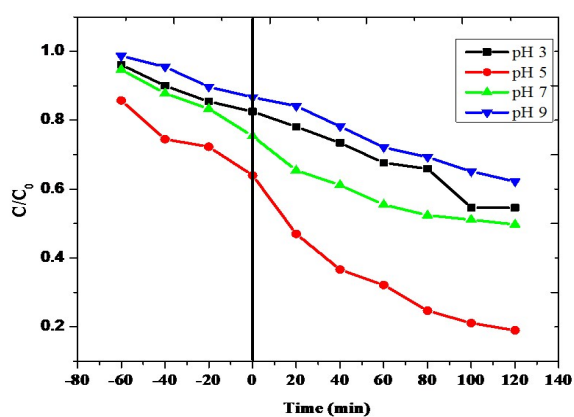
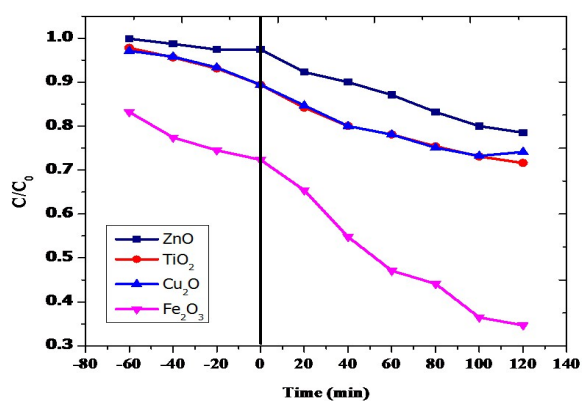
	IC <sub>50</sub> (µg/ml)
Rhombohedral Fe <sub>2</sub> O <sub>3</sub>	202.14 ± 0.14
STD (Cisplatin)	>500

The data represents in mean ± SD of three experiments

**Table S6** Inhibitory concentration values (IC<sub>50</sub>) of HEK-293 (human embryonic kidney – 293) cells against Rhombohedral Fe<sub>2</sub>O<sub>3</sub> nanoparticles.**Table S7** The Photocatalytic degradation intermediates of Rhodamine – B (RB-B) by LC-MS.

Product Code	M/Z	Chemical Formula	Name	Structure
1 RB-B	105	C <sub>7</sub> H <sub>5</sub> O <sup>+</sup>	benzylidyeoxonium	
2 RB-B	122	C <sub>7</sub> H <sub>6</sub> O <sub>2</sub>	benzoic acid	
3 RB-B	149	C <sub>9</sub> H <sub>11</sub> NO	(E)-4-amino-2-(prop-1-en-1-yl)phenol	
4 RB-B	210	C <sub>13</sub> H <sub>10</sub> N <sub>2</sub> O	3-imino-3H-xanthen-6-amine	
5 RB-B	238	C <sub>15</sub> H <sub>14</sub> N <sub>2</sub> O	(Z)-3-(ethylimino)-3H-xanthen-6-amine	
6 RB-B	323	C <sub>22</sub> H <sub>29</sub> N <sub>2</sub> O <sup>+</sup>	(E)-N,N-diethyl-3-(diethylimmonium)-3H-xanthen-6-amine	

<b>7 RB-B</b>	330	$C_{20}H_{14}N_2O_3$	2-(6-amino-3-imino-3H-xanthen-9-yl)benzoic acid	
<b>8 RB-B</b>	358	$C_{22}H_{18}N_2O_3$	2-(6-(ethylamino)-3-imino-3H-xanthen-9-yl)benzoic acid	
<b>9 RB-B</b>	387	$C_{24}H_{22}N_2O_3$	2-(6-(diethylamino)-3-imino-3H-xanthen-9-yl)benzoic acid	

**LIST OF FIGURES****Fig. S1** FT-IR spectrum of the Fe<sub>2</sub>O<sub>3</sub> material.**Fig. S2** Role of pH photocatalytic degradation of Rhodamine-B (RB-B) by rhombohedral Fe<sub>2</sub>O<sub>3</sub>.**Fig. S3** Photocatalytic degradation of Rhodamine-B (RB-B) using various photocatalysts.

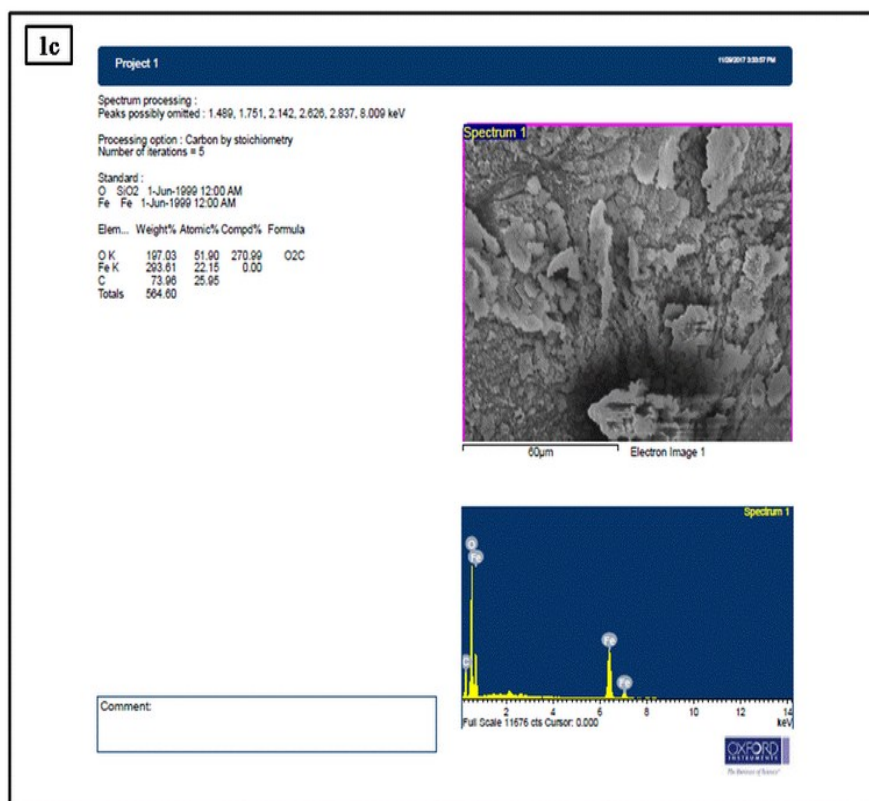


Fig. S4 The elemental composition and atomic percentage of  $\text{Fe}_2\text{O}_3$  in EDS (Sample code: 1c)

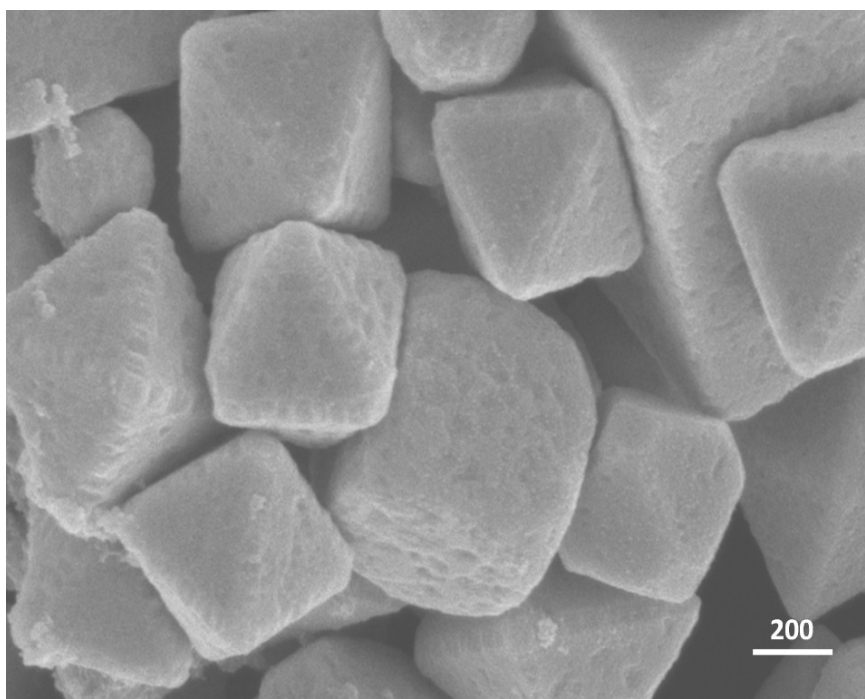
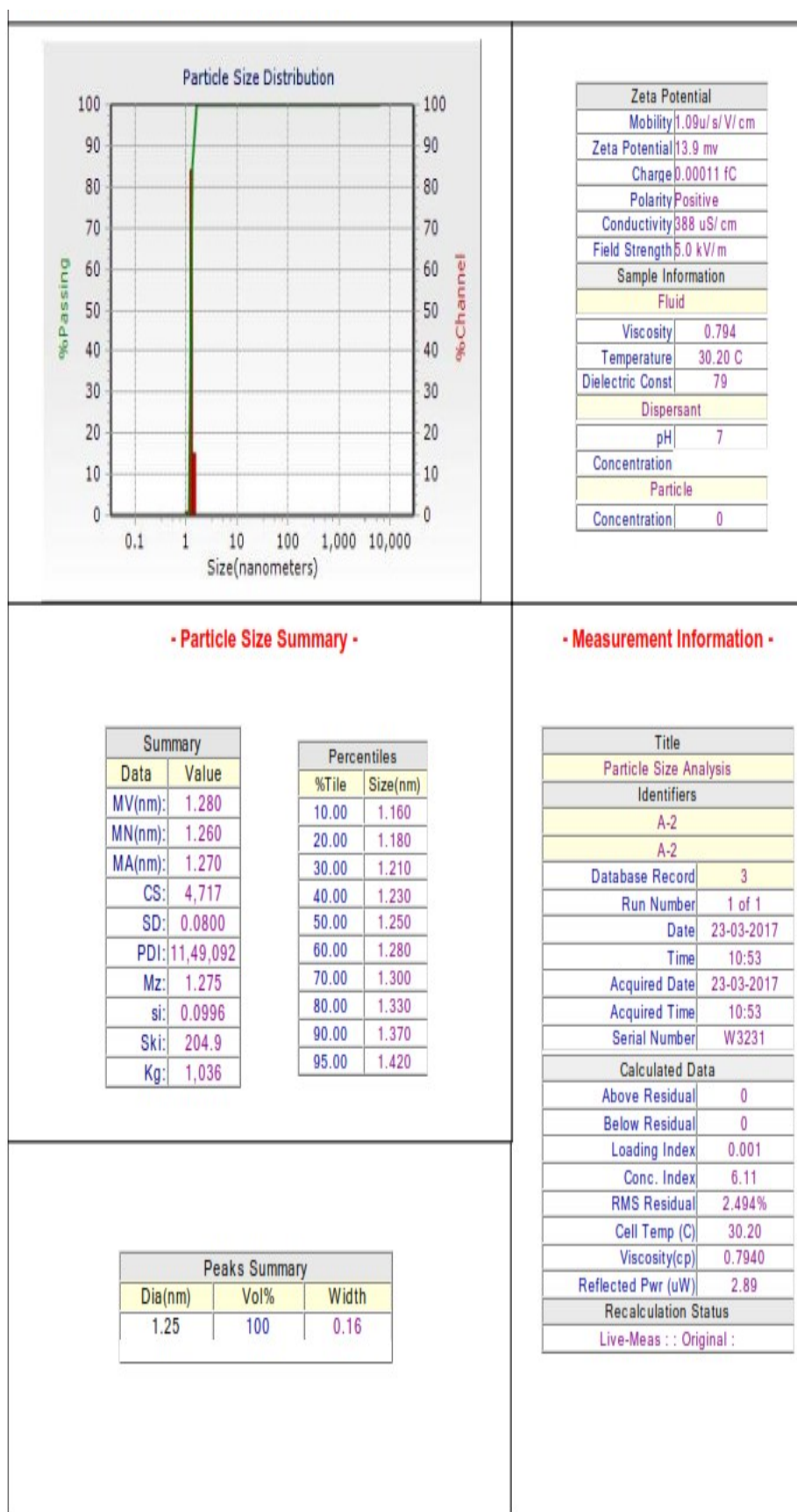


Fig. S5 The FE-SEM image of Rhombohedral  $\text{Fe}_2\text{O}_3$

Fig. S6 The Rhombohedral Fe<sub>2</sub>O<sub>3</sub> Particle size distribution and summary

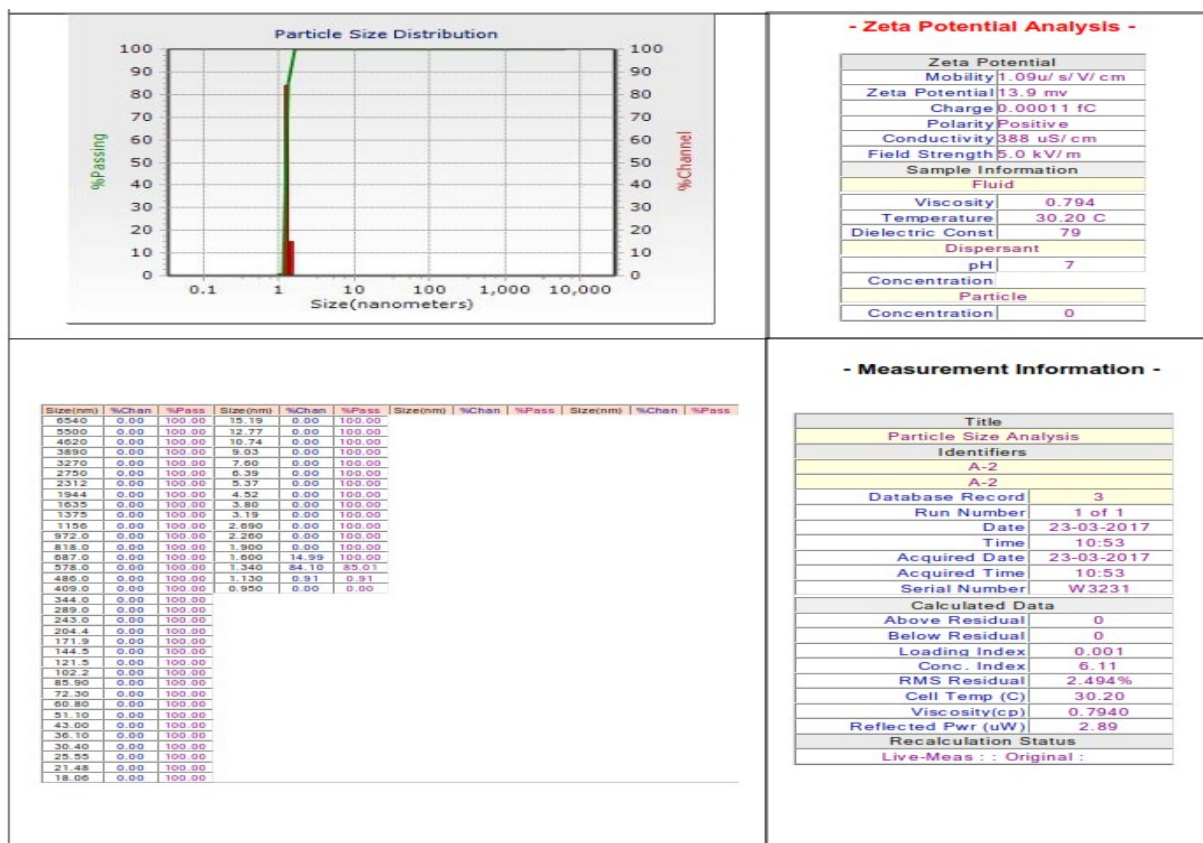


Fig. S7 The Rhombohedral Fe<sub>2</sub>O<sub>3</sub> zeta potential analysis results

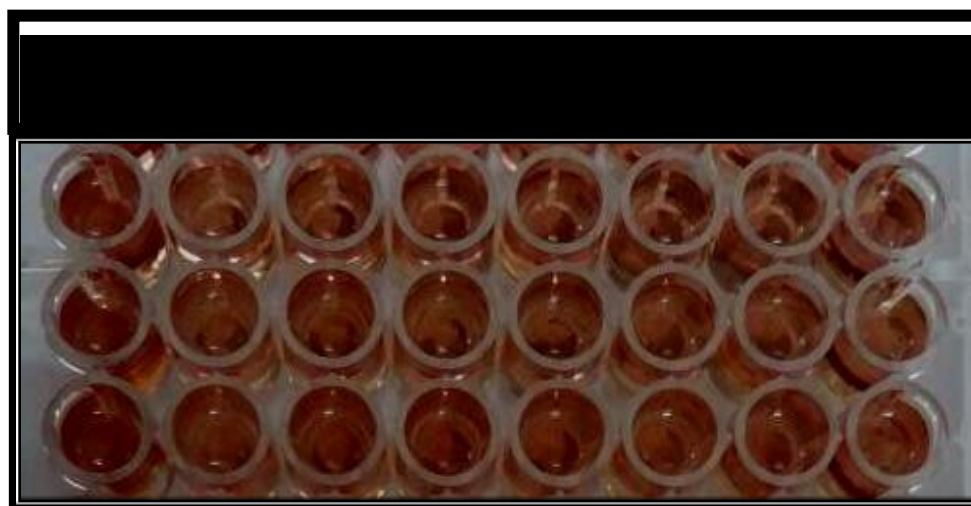


Fig. S8 Cell line activity of Fe<sub>2</sub>O<sub>3</sub> by using Human Embryonic Kidney cells – 293 (HEK - 293) at 48 Hours. Cells were grown in 96-well plates in a final volume of 100 μL of culture medium per well. Each well contained 1 × 10<sup>5</sup> cells/mL (In triplicate to maintain the accuracy as well as to get the better results) and was incubated for 24 h in a 5% CO<sub>2</sub> incubator at 37°C.



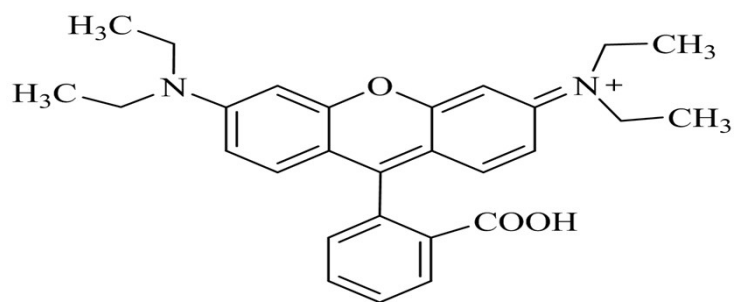


Fig. S9 The Rhombohedral structure

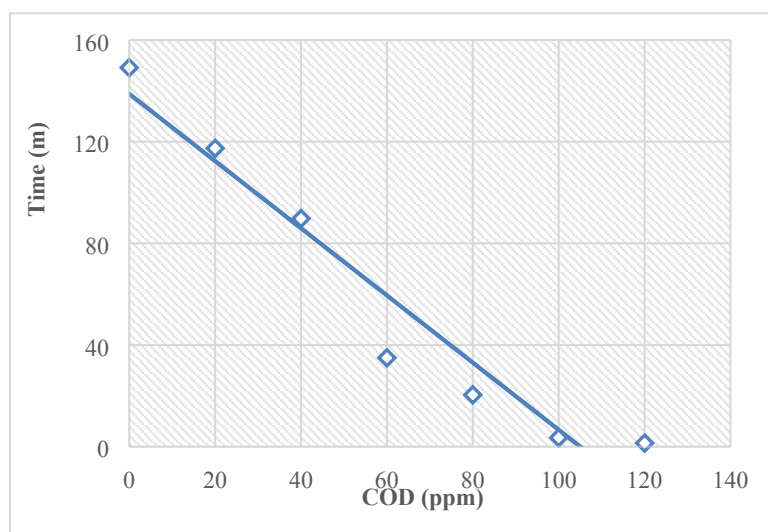


Fig. S10 The negative correlation result of COD vs. time, (Decreased level of COD with increasing time).

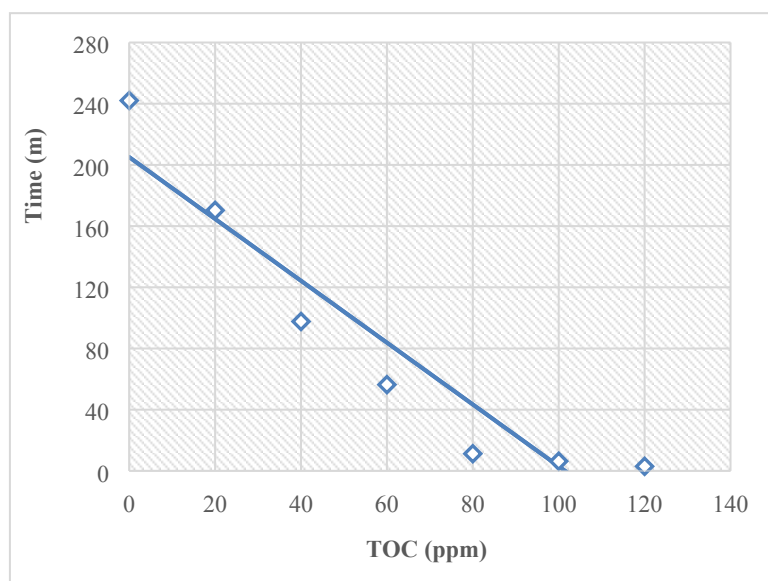


Fig. S11 The negative correlation result of TOC vs. time, (Decreased level of TOC with increasing time).

## **LIST OF INFORMATION**

### **Info. S1 B16-F10 Cell culture and treatment**

The (B16-F10) seeded in tissue culture flasks and full-grown in Dulbecco's modified Eagle's medium (DMEM; Thermo Fisher, USA Gibco), balance with 5% fetal bovine serum (FBS, Thermo Fisher; Gibco) and 1% penicillin/streptomycin blend (Santa Cruz Biotechnology, USA). A dose-dependent response curve with different concentrations (0.01, 0.05, 0.1, 0.5, 1.0, and 2.5 µg/l) was plotted using propagation and MTT [(2-(3, 5-diphenyltetrazol-2-ium-2-yl)-4,5-dimethyl-1,3-thiazole bromide)] assay, after 72 hours of exposure. For this, 105 cells were seeded in 24-well plates and cultured with fresh culture medium for 72 hours prior to treatment. Fe<sub>2</sub>O<sub>3</sub> be diluted in di-methyl sulfoxide (DMSO; maximum 1% final concentration) and, for this basis, 1% of DMSO was added in the culture medium of the control group. The concentrations of Fe<sub>2</sub>O<sub>3</sub> used in the other trails were 5, 50, 100, 200, 300, 400 and 500 µg/l, and they were monitored for duration of 72 hour. All the experiments were performed in triplicate and three independent repetitions.

### **Info. S2 Normal Cell line toxicity testing using HEK-293**

HEK-293 (human embryonic kidney), cell lines were gotten from the American Type Culture Collection (ACC), USA. The growing media: RPMI-1640, Mediated essential medium (MEM), and the phosphate buffer solution (PBS) tablets were obtained from Sigma Chemical Co., St Louis, USA. High quality fetal serum of calf penicillin–streptomycin, trypsin and XTT kit were taken from Sigma labs. Towards perform a cell proliferation assay (XTT) with one 96 wells microplate (1 × 10<sup>5</sup> cells/mL/well) were incubated for 24 hours in a 5% CO<sub>2</sub> incubator at 37°C. Same methodology involved in HEK-293 cell line IC<sub>50</sub> assessment, parallely the test Fe<sub>2</sub>O<sub>3</sub> nanomaterial inhibits the 50% of the proliferating the normal cells as per above environmental conditions for 48 hours. The control and effected cell growth were captured using (Olympus, USA).