# **Supporting Information**

## Development of improved dual-diazonium reagents for faster crosslinking of tobacco mosaic virus as hydrogels

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#### 1. Synthesis.



Scheme S1. Synthesis of 5.

#### Synthesis of 5

4,4'-methylenediphenol (203 mg, 1.0 mmol) was dissolved in 140 mL CH<sub>3</sub>CN and 60 mL phosphate buffer (200 mM, pH 6.0). **DDA-2** (584 mg, 1.0 mmol) was dissolved in 50 mL CH<sub>3</sub>CN and the reaction mixture was stirred at room temperature for 1 h. After removing CH<sub>3</sub>CN under reduced pressure, the mixture was extracted with EtOAc. Next, the combined organic extracts were washed with brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. After the solution was removed, the product was purified by silica gel column chromatography (CH<sub>2</sub>Cl<sub>2</sub>: MeOH = 99 : 1) to give **5** (64 mg, 13.7%).<sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.56 (s, 2H), 8.69 (d, *J* = 11.2 Hz, 2H), 8.07 (d, *J* = 8.1 Hz, 4H), 8.00 (dd, *J* = 11.0, 7.6 Hz, 2H), 7.70 (td, *J* = 7.8, 2.4 Hz, 2H), 7.33 (dd, *J* = 8.4, 2.2 Hz, 2H), 6.91 (d, *J* = 8.3 Hz, 2H), 4.03 (d, *J* = 14.6 Hz, 1H), 3.80 (d, *J* = 14.6 Hz, 1H), 2.25 (d, *J* = 13.7 Hz, 3H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  152.2, 151.8, 151.7, 139.3, 137.0, 136.0, 134.0, 132.6, 132.4, 132.4, 130.0, 129.8, 127.4, 127.3, 124.1, 123.1, 117.6, 37.8, 17.9, 17.2; <sup>31</sup>P NMR (162 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  26.62 (s). HRMS (ESI): *m*/*z* [M+H]<sup>+</sup> calculated for C<sub>26</sub>H<sub>22</sub>N<sub>4</sub>O<sub>3</sub>P<sup>+</sup>: 469.1424; found: 469.1423.

### 2. Supplementary figures



**Fig. S1.** Time-dependent UV-Vis absorbance spectra change of **DDA-2** (20  $\mu$ M) at 400 nm, when treated with different concentrations of H-Tyr-OBzl·Tos in PBS buffer. (a. 200  $\mu$ M; b. 400  $\mu$ M; c. 600  $\mu$ M; d. 800  $\mu$ M; e. 1000  $\mu$ M). The second-order rate constant was obtained by plotting the pseudo first-order rates against the concentration of tyrosine used (f).



**Fig. S2.** Time-dependent UV-Vis absorbance spectra change of **DDA-3** (20  $\mu$ M) at 400 nm, when treated with different concentrations of H-Tyr-OBzl·Tos in PBS buffer. (a. 100  $\mu$ M; b. 150  $\mu$ M; c. 250  $\mu$ M; d. 300  $\mu$ M). The second-order rate constant was obtained by plotting the pseudo first-order rates against the concentration of tyrosine used (e).



**Fig. S3.** Plots of the Hammett parameters vs. rate constant for three azo coupling reactions. We obtain the corresponding Hammett constant by two kinds of similar structural substituents. a) The Hammett constants of the substituent COEt, PO(Me)<sub>2</sub>, SO<sub>2</sub>Et are 0.48, 0.43, 0.77, respectively; b) The Hammett constants of the substituent COC<sub>6</sub>H<sub>5</sub>, PO(C<sub>6</sub>H<sub>5</sub>)<sub>2</sub>, SO<sub>2</sub>C<sub>6</sub>H<sub>5</sub> are 0.43, 0.38, 0.68, respectively. The  $\sigma$ value is plotted against lg K,  $\rho$  is the slope of the Hammett line and is called the reaction constant, depending on the type and condition of the reaction; K is the reaction rate constant of the substituted homologue.



**Fig. S4.** The comparison of TMV gelation at 25 °C (a) and 37 °C (b). The reaction was performed by mixing 9  $\mu$ L TMV virus (1  $\mu$ g/ $\mu$ L) and 1  $\mu$ L **DDA-1** with various concentrations (100, 50, 25, 12.5, 6.25, 3.12, 1.56, mM). The reaction was incubated at 25 °C or 37 °C for 60 min. After reaction, 10  $\mu$ L sample for each treatment was directly used for 12% SDS-PAGE assay. (c) The crosslinking rate of TMV virus (%) was calculated based on the group without the addition of **DDA-1**.



**Fig. S5.** The comparison of TMV gelation at 25 °C (a) and 37 °C (b). The reaction was performed by mixing 9  $\mu$ L TMV virus (1  $\mu$ g/ $\mu$ L) and 1  $\mu$ L **DDA-2** with various concentrations (100, 50, 25, 12.5, 6.25, 3.12, 1.56, mM). The reaction was incubated at 25 °C or 37 °C for 60 min. After reaction, 10  $\mu$ L sample for each treatment was directly used for 12% SDS-PAGE assay. (c) The crosslinking rate of TMV virus (%) was calculated based on the group without the addition of **DDA-2**.



**Fig. S6.** The comparison of TMV gelation at 25 °C (a) and 37 °C (b). The reaction was performed by mixing 9  $\mu$ L TMV virus (1  $\mu$ g/ $\mu$ L) and 1  $\mu$ L **DDA-3** with various concentrations (100, 50, 25, 12.5, 6.25, 3.12, 1.56, mM). The reaction was incubated at 25 °C or 37 °C for 60 min. After reaction, 10  $\mu$ L sample for each treatment was directly used for 12% SDS-PAGE assay. (c) The crosslinking rate of TMV virus (%) was calculated based on the group without the addition of **DDA-3**.



**Fig. S7.** Time-dependent crosslinking of TMV with **DDA-3**. (a) The reaction was performed by mixing 9  $\mu$ L TMV virus (1  $\mu$ g/ $\mu$ L) and 1  $\mu$ L **DDA-3** (100 mM). The reaction was incubated at 25 °C for various time (0, 1, 5, 10, 15, 30, 45, 60, min). At the indicated time, the reaction was stopped with 5×SDS loading buffer and then used for 12% SDS-PAGE assay. (b) The crosslinking rate of TMV virus (%) was calculated based on the group at 0 min.



**Fig. S8.** Time-dependent crosslinking of TMV with **DDA-1**. (a) The reaction was performed by mixing 9  $\mu$ L TMV virus (1  $\mu$ g/ $\mu$ L) and 1  $\mu$ L **DDA-1** (100 mM). The reaction was incubated at 25 °C for various time (0, 1, 5, 10, 15, 30, 45, 60, min). At the indicated time, the reaction was stopped with 5×SDS loading buffer and then used for 12% SDS-PAGE assay. (b) The crosslinking rate of TMV virus (%) was calculated based on the group at 0 min.



**Fig. S9.** The minimum gelation time of three TMV hydrogels at room temperature. The concentration of **DDA-2** or **DDA-3** was 10 mM and the concentration of **DDA-4** was 2 mM; The concentration of TMV matrix was 2.5 mg/mL.

5 min							10 min							
DDA-3	DDA-3 (mM)						DDA-3 (mM)							
10	5	2.5	1.25	0.625	0.312	0	10	5	2.5	1.25	0.625	0.312	0	
	405	Nee -				(154		43.						
			<i>k</i> :		24	4			Aria Contraction	A82.8		121		
20 min														
	( <b>N</b> )	2	0 min							30 1	min			
DDA-3	(mM)	25	20 min	0.625	0.212		DDA-3 (1	mM)	25	301		0.212		
DDA-3 10	(mM) 5	2.5	20 min 1.25	0.625	0.312	0	DDA-3 (1) 10	mM) 5	2.5	30 I 1.25	min 0.625	0.312	0	

**Fig. S10.** The effect of the concentration of **DDA-3** on TMV gelation. The final concentration of **DDA-3** was varying from 0~10 mM and the concentration of TMV matrix was 2.5 mg/mL. The gelation time was kept within 0-30 min at room temperature.

		:	5 min							10 1	min		
TMV (r			TMV (mg/mL)										
10	5	2.5	1.25	0.625	0.312	0	10	5	2.5	1.25	0.625	0.312	0
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	1600	1150	-1.5 -1	1547	15			Ki	188	Ach		10	
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		2	0							20.			
TMV (r	ng/mL	2	0 min				TMV (m	g/mI)		30	min		
TMV (r	ng/mL)	2	0 min	0.625	0.212		TMV (m	ng/mL)		30	min	0.212	
TMV (r 10	ng/mL) 5	20	0 min 1.25	0.625	0.312	0	TMV (m 10	ng/mL) 5	2.5	<b>30</b> 1 1.25	0.625	0.312	0
TMV (r 10	ng/mL) 5	2.5	0 min 1.25	0.625	0.312	0	TMV (m 10	ng/mL) 5	2.5	<b>30</b> 1 1.25	0.625	0.312	0
TMV (r 10	ng/mL) 5	2.5	0 min 1.25	0.625	0.312	0	TMV (m 10	ng/mL) 5	2.5	<b>30</b> 1 1.25	min 0.625	0.312	0
10	ng/mL) 5	2.5	0 min 1.25	0.625	0.312	0	TMV (m	ng/mL) 5	2.5	30	0.625	0.312	0
10	ng/mL) 5	2.5	0 min 1.25	0.625	0.312	0	TMV (m	ng/mL) 5	2.5	30	min 0.625	0.312	0
TMV (r 10	ng/mL) 5	21 2.5	0 min 1.25	0.625	0.312	0	TMV (m	ng/mL) 5	2.5	30	0.625	0.312	0
10	ng/mL) 5	2.5	0 min 1.25	0.625	0.312	0	TMV (m	ng/mL) 5	2.5	30	0.625	0.312	0
TMV (r 10	ng/mL) 5	2.5	0 min 1.25	0.625	0.312	0	TMV (m 10	ng/mL) 5	2.5	30	min 0.625	0.312	0
10	ng/mL) 5	2.5	0 min 1.25	0.625	0.312	0	TMV (m	ng/mL) 5	2.5	30	0.625	0.312	
10	ng/mL) 5		0 min 1.25	0.625	0.312		TMV (m 10	ng/mL) 5	2.5	30	min 0.625	0.312	

**Fig. S11.** The effect of the concentration of TMV matrix on TMV gelation. The final concentration of TMV matrix was varying from 0~10 mg/mL and the concentration of **DDA-3** was 10 mM. The gelation time was kept within 0-30 min at room temperature.



**Fig. S12.** Degradation of virus-based hydrogel by DTT. Both gels were prepared by 2 mM **DDA-4** and 2.5 mg/mL TMV. 20 mL DTT (deionized water, 500 mM) was added to the petri dish on the right as the experimental group and 20 mL of deionized water was added to the petri dish on the left as a control. The gel degradation was imaged at different time (0 min, 30 min, 1 h, 2h, 3 h, 5h, 8 h, 10h, 12 h).























