# Synthesis, human topoisomerase Πα inhibitory properties and molecular modeling studies of anti-proliferative curcumin mimics

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**Fig. S85.** ORTEP view of compound **25** showing the atom-numbering scheme. Displacement ellipsoids are drawn at the 50 % probability level and H atoms are shown as small spheres of arbitrary radii.

**Fig. S86.** ORTEP view of compound **34** showing the atom-numbering scheme. Displacement ellipsoids are drawn at the 50 % probability level and H atoms are shown as small spheres of arbitrary radii.

**Fig. S87**. The crystal packing of compound **25**. The H atoms not engaged in the intermolecular interactions (dashed lines) have been skipped for clarity.

**Fig. S88**. The crystal packing of compound **34**. The H atoms not engaged in the intermolecular interactions (dashed lines) have been skipped for clarity.

**Fig. S89.** A projection of the optimized structure of compound **25** by DFT/B3LYP method with 3-21G\* basis set.

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Fig. S91. Overlay diagram of compound 25; red (X-ray structure), green (DFT).

Fig. S92. Overlay diagram of compound 34; red (X-ray structure), green (DFT).

**Fig. S93.** Dose-response curve for the tested compounds against HCT116 (colon cancer) cell line.

**Fig. S94.** Dose-response curve for the tested compounds against MCF7 (breast cancer) cell line.

**Fig. S95.** Dose-response curve for the tested compounds against A431 (squemous cancer) cell line.

**Fig. S96.** Dose-response curve for the tested compounds against RPE1 (retinal pigment epithelium) cell line.

**Fig. S97.** BMLR-QSAR model plot of correlations representing the observed *vs.* predicted  $1/IC_{50}$ ,  $\mu$ M values for the tested compounds against HCT116 (colon) carcinoma cell line.

**Fig. S98.** BMLR-QSAR model plot of correlations representing the observed *vs.* predicted IC<sub>50</sub>,  $\mu$ M values for the tested compounds against MCF7 (breast) carcinoma cell line.

**Fig. S99.** BMLR-QSAR model plot of correlations representing the observed *vs.* predicted log(IC<sub>50</sub>),  $\mu$ M values for the tested compounds against A431 (squamous) carcinoma cell line.

**Fig. S100.** Constraint distances "H-1 – H-2 = 10.672, H-1 – H-3 = 7.810, H-2 – H-3 = 11.639, H-1 – HBA = 4.686, H-2 – HBA = 6.093, H-3 – HBA = 8.206 Å" of the generated 3D-pharmacophore for the tested piperidinecarboxamides **24–47** against HCT116 (colon) carcinoma cell line which contains three hydrophobics (H-1, H-2, H-3; light blue) and one hydrogen bonding acceptor (HBA; green).

**Fig. S101.** Constraint angles "H-1 – H-2 – H-3 = 40.69, H-3 – H-1 – HBA = 77.68, H-3 – H-2 – HBA = 42.09 °" of the generated 3D-pharmacophore for the tested piperidinecarboxamides **24–47** against HCT116 (colon) carcinoma cell line which contains three hydrophobics (H-1, H-2, H-3; light blue) and one hydrogen bonding acceptor (HBA; green).

Fig. S102. 3D-pharmacophore model mapped on the tested piperidinecarboxamides 24–47 against HCT116 (colon) carcinoma cell line.

**Fig. S103.** Constraint distances "H-1 – H-2 = 9.356, H-1 – HBA = 8.527, H-2 – HBA = 6.917 Å" of the generated 3D-pharmacophore for the tested piperidinecarboxamides **24–47** against MCF7 (breast) carcinoma cell line which contains two hydrophobics (H-1, H-2; light blue) and one hydrogen bonding acceptor (HBA; green).

**Fig. S104.** Constraint angle "H-2 – H-1 – HBA = 45.21 °" of the generated 3D-pharmacophore for the tested piperidinecarboxamides **24–47** against MCF7 (breast) carcinoma cell line which contains two hydrophobics (H-1, H-2; light blue) and one hydrogen bonding acceptor (HBA; green).

**Fig. S105.** 3D-pharmacophore model mapped on the tested piperidinecarboxamides 24–47 against MCF7 (breast) carcinoma cell line.

**Fig. S106.** Constraint distances "H-1 – H-2 = 8.270, H-1 – HBA-1 = 6.866, H-1 – HBA-2 = 4.726, H-2 – HBA-1 = 8.788, H-2 – HBA-2 = 4.683, HBA-1 – HBA-2 = 6.077 Å" of the generated 3D-pharmacophore for the tested piperidinecarboxamides **24–47** against A431 (squamous) carcinoma cell line which contains two hydrophobics (H-1, H-2; light blue) and two hydrogen bonding acceptor (HBA-1, HBA-2; green).

**Fig. S107.** Constraint angles "H-2 – H-1 – HBA-1 = 70.28, H-2 – HBA-2 – HBA-1 = 108.82 °" of the generated 3D-pharmacophore for the tested piperidinecarboxamides **24–47** against A431 (squamous) carcinoma cell line which contains two hydrophobics (H-1, H-2; light blue) and two hydrogen bonding acceptor (HBA-1, HBA-2; green). **Fig. S108.** 3D-pharmacophore model mapped on the tested piperidinecarboxamides

**24–47** against A431 (squamous) carcinoma cell line.

## X-ray crystallography

Single crystals of compounds 25 and 34 were grown using slow solvent evaporation method. For compound 25, good crystal has been selected, checked and mounted onto a thin glass fiber. The X-ray single crystal diffraction data were collected at room temperature (293 K) on an Enraf-Nonius 590 diffractometer with a Kappa CCD detector using graphite monochromated Mo- $K\alpha$  ( $\lambda = 0.71073$  Å) radiation, at National Research Center of Egypt.<sup>1,2</sup> Reflection data has been recorded in the rotation mode using the  $\phi$  and  $\omega$  scan technique with  $2\theta_{\text{max}} = 26.678$ °. Unit cell parameters were determined from least-squares refinement with  $\theta$  in the range  $2 \le \theta \le 26$ . Regarding compound 34, Data collections were performed at the X-ray diffraction beamline (XRD1) of the Elettra Synchrotron, Trieste, Italy.<sup>3</sup> Suitable crystal was dipped in NHV oil (Jena Bioscience GmbH) and mounted on the goniometer head with a nylon loop. Complete datasets were collected at room temperature through the rotating crystal method with  $2\theta_{\text{max}} = 30.961$  °. Data were acquired using a monochromatic wavelength of 0.700 Å on a Pilatus 2M hybrid-pixel area detector. The diffraction data were indexed with  $\theta$  in the range  $2 \le \theta \le$ 30 and integrated using XDS.<sup>4</sup> The structure was solved by direct methods using  $SIR-92^5$ and SUPERFLIP<sup>6</sup> implemented in CRYSTALS program suit.<sup>7</sup> The refinement was carried out by full-matrix least-squares method on the positional and anisotropic temperature parameters of all non-hydrogen atoms based on  $F^2$  using *CRYSTALS* package. All hydrogen atoms were positioned geometrically and were initially refined with soft restraints on the bond lengths and angles to regularize their geometry (C—H in the range 0.93–0.98 and N—H in the range 0.86–0.89) and  $U_{iso}(H)$  (in the range 1.2-1.5 times  $U_{eq}$ of the parent atom). Then, the positions were refined with riding constraints.<sup>8</sup> The general-purpose crystallographic tool *PLATON*<sup>9</sup> was used for the structure analysis and presentation of the results. The molecular graphics were carried out using ORTEP-3 for Windows<sup>10</sup> program. Details of the data collection conditions and the parameters of the refinement process are given in Table S11. Crystallographic data for the structures in this paper have been deposited with the Cambridge Crystallographic Data Center as supplementary publication numbers CCDC 1855508 and CCDC 1855509. Copies of the data can be obtained, free of charge, on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK [Fax: 144(0)1223 336033 or e-mail: deposit@ccdc.cam.ac.uk].

#### In-vitro antitumor screening

The targeted piperidinecarboxamides 24-51 were screened for their antitumor properties against HCT116 (colon), MCF7 (breast) and A431 (squamous skin) carcinoma cell lines by the standard mitochondrial dependent reduction of yellow MTT [3-(4,5dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide] to purple formazan technique.<sup>11</sup> Cells were suspended in McCoy's 5A medium for HCT116, DMEM for MCF-7 and A431 in addition to 1% antibiotic-antimycotic mixture (10000  $\mu g$  ml<sup>-1</sup> potassium penicillin, 10000  $\mu$ g ml<sup>-1</sup> streptomycin sulfate and 25  $\mu$ g ml<sup>-1</sup> amphotericin B), 10% fetal bovine serum and 1% L-glutamine at 37 °C, under 5% CO<sub>2</sub> and 95% humidity. Cells were seeded at concentration of 30000 cells per well in fresh complete growth medium in 96-well tissue culture microtiter plates for 24 h. Media was aspirated, fresh complete medium was added and cells were incubated with different concentrations of the tested compound to give a final concentration of [50, 25, 12.5 and 6.25  $\mu$ M (3, 1.5, 7.5 and 0.375  $\mu$ M for the high potent analogues)]. 0.5% DMSO was used as negative control and 5-fluorouracil was used as positive control (standard reference). Triplicate wells were prepared for each individual dose. After 72 h of incubation, medium was aspirated, 40  $\mu$ l MTT salt (2.5 mg ml<sup>-1</sup>) were added to each well and incubated for further 4 h at 37 °C. To stop the reaction and dissolve the formed crystals, 150  $\mu$ l of 10% sodium dodecyl sulfate (SDS) in deionized water were added to each well and incubated overnight at 37 °C. The absorbance was then measured at 570 nm and a reference wavelength of 595 nm.

Data were collected as mean values for experiments performed in triplicates for each individual dose which had been measured by MTT assay. Control experiments did not exhibit significant change compared to the DMSO vehicle. The percentage of cell survival was calculated according to equ. (1).

$$Surviving \ fraction = \frac{Optical \ density \ (O.D.)of \ treated \ cells}{O.D.of \ control \ cells} \qquad (1)$$

The synthesized compounds **24–51** were also tested against RPE1 (normal human immortalized retinal pigment epithelial cell line) cell to determine the toxicity/selectivity

towards normal cells relative to the carcinoma cell lines utilized. The  $IC_{50}$  (concentration required to produce 50% inhibition of cell growth compared to the control experiment) was determined using Graph-Pad PRISM version-5 software. Statistical calculations for determination of the mean and standard error values were determined by SPSS 16 software. The observed anti-proliferative properties are presented in Table 1 (Supplementary Figs. S93–S96).

#### **2D-QSAR studies**

3,5-bis(arylidene)-N-substituted-4-oxo-piperidine-1-The synthesized carboxamides 24-47 revealing variable anti-proliferative properties were utilized for developing the 2D-QSAR modeling by CODESSA-Pro (comprehensive descriptors for structural and statistical analysis) software. Geometry of the compounds was initially optimized by AM1 technique using hyperChem 8.0 then, uploaded to CODESSA-Pro for final geometrical structure optimization by MOPAC.<sup>12</sup> CODESSA-Pro calculated 673 (for the tested compounds against HCT116 and MCF7 cell lines) and 711 (for the tested compounds against A431 cell line) molecular descriptors (constitutional, topological, geometrical, charge-related, semi-empirical, molecular-type, atomic-type and bond-type descriptors in addition to, thermodynamic descriptors in case of A431 cell line) for the exported anti-proliferative active agents. Mathematical transformation of the experimental values (including IC<sub>50</sub>, 1/IC<sub>50</sub>, log(IC<sub>50</sub>) and 1/ log(IC<sub>50</sub>) µM) were used searching for the best QSAR model. The best multi-linear regression (BMLR) technique was utilized which is a stepwise search for the best *n*-parameter regression equations (where *n* stands for the number of descriptors used), based on the highest  $R^2$  (squared correlation coefficient),  $R^2$  cvOO (squared cross-validation "leave one-out, LOO" coefficient),  $R^2$  cvMO (squared cross-validation "leave many-out up to 20% of the training set, LMO" coefficient), F (Fisher statistical significance criteria) values, and  $s^2$ (standard deviation). The QSAR up to 3-descriptor model describing the biological activity of the anti-proliferative active agents were generated (obeying the thumb rule of 8:1 which is the ratio between the data points and the number of QSAR descriptor).

### Human DNA topoisomerase IIa inhibitory properties

Human DNA topoisomerase IIα activity assay was undertaken for compounds **29**, **30** and **34–38** by the Confirmatory Diagnostic Unit, Egyptian Company for Production of Vaccines, Sera and Drugs (VACSERA), Cairo, Egypt according to the Topoisomerase II Assay Kit, (plasmid based) manufacturer's instructions (TopoGEN, Inc., 108 Aces Alley, Port Orange, Florida 32128, USA) using ROBONIK EIA reader (450 nm), DMSO as solvent and Methotrexate as a standard reference.

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D—H···A	<i>D</i> —Н	Н…А	$D \cdots A$	D—H···A
$N1$ — $H262$ ···· $O2^{i}$	0.86(2)	2.13(3)	2.950(3)	161(2)
C26—H261…O1 <sup>ii</sup>	0.95	2.57	3.172(5)	122

**Table S1.** Hydrogen-bond geometry (Å,  $^{\circ}$ ) for compound **25**.

Symmetry codes: (i) 1/2-x,1/2-y,1-z; (ii) 1/2-x,1/2+y,1/2-z.

**Table S2.** Hydrogen-bond geometry (Å, °) for compound 34.

<i>D</i> —H···A	D—H	Н…А	D···A	D—H···A
$N1 - H2 \cdots O2^{i}$	0.85(3)	2.07(2)	2.908(3)	165(2)
C23—H231…O1 <sup>ii</sup>	0.95	2.49	3.398(3)	161

Symmetry codes: (i) 1-x,1-y,-z; (ii) -1/2+x,3/2-y,-1/2+z.

Coometrie nonometers	Compound 25		Compound <b>34</b>	
Geometric parameters	Exp. X-ray data	DFT	Exp. X-ray data	DFT
O1—C7	1.229	1.245	1.215	1.245
O2—C18	1.234	1.248	1.237	1.248
N1—C6	1.41	1.41	1.410	1.414
N1—C7	1.371	1.391	1.368	1.387
N2—C7	1.361	1.397	1.373	1.401
N2—C8	1.45	1.469	1.457	1.468
N2—C9	1.451	1.470	1.450	1.470
C1—C2	1.381	1.391	1.391	1.392
C1—C6	1.388	1.406	1.382	1.406
C2—C3	1.377	1.393	1.354	1.397
C3—C4	1.373	1.392	1.360	1.396
C4—C5	1.385	1.393	1.383	1.394
C5—C6	1.389	1.405	1.384	1.404
C8—C19	1.512	1.514	1.507	1.514
C9—C17	1.5	1.526	1.505	1.526
C10—C11	1.383	1.393	1.373	1.390
C10—C15	1.386	1.411	1.398	1.412
C11—C12	1.368	1.397	1.338	1.390
C12—C13	1.374	1.398	1.347	1.390
C13—C14	1.381	1.391	1.408	1.388
C14—C15	1.393	1.413	1.385	1.413
C15—C16	1.472	1.461	1.457	1.459
C16—C17	1.341	1.353	1.339	1.353
C17—C18	1.496	1.503	1.485	1.502
C18—C19	1.49	1.503	1.484	1.502
C19—C20	1.342	1.347	1.353	1.347
C20—C21	1.459	1.471	1.454	1.469

**Table S3.** Selected intramolecular experimental (X-ray) and computational optimizedgeometrical parameters (bond lengths, Å) of compounds 25 and 34.

C21—C22	1.403	1.410	1.409	1.411
C21—C26	1.38	1.409	1.391	1.409
C22—C23	1.376	1.394	1.376	1.390
C23—C24	1.378	1.399	1.366	1.391
C24—C25	1.366	1.397	1.377	1.389
C25—C26	1.387	1.395	1.388	1.392
C3—Cl1	1.746	1.766		
C24—F1			1.360	1.367
C12—F2			1.356	1.367
RMSE		0.018		0.022
Maximum difference		0.036		0.052

Coometrie response	Compound 25	Compound 25		Compound 34	
Geometric parameters	Exp. X-ray data	DFT	Exp. X-ray data	DFT	
C6—N1—C7	125.8	126.3	124.7	126.5	
C7—N2—C8	128.6	124.7	124.8	124.8	
C7—N2—C9	119.4	117.7	117.7	117.4	
C8—N2—C9	111.9	113.1	113.1	113.2	
C2—C1—C6	121.1	120.9	119.4	120.6	
C1—C2—C3	119.6	119.4	120.8	120.3	
C2—C3—C4	119.9	120.5	119.3	119.1	
C3—C4—C5	120.9	120.2	122.1	121.2	
C4—C5—C6	119.7	120.2	118.3	119.7	
N1—C6—C5	123.2	123.4	122.4	123.4	
N1—C6—C1	118	117.8	117.5	117.5	
C5—C6—C1	118.8	118.8	120.1	119.0	
N1—C7—N2	116.8	114.4	114.5	114.4	
N2—C7—O1	120.7	121.3	121.4	121.0	
N1—C7—O1	122.5	124.3	123.9	124.6	
N2—C8—C19	111.3	107.6	110.2	107.8	
N2—C9—C17	108.8	110.5	109.0	110.5	
C11—C10—C15	121.2	120.7	121.2	120.9	
C10-C11-C12	120	120.5	119.3	119.8	
C11—C12—C13	120.6	119.7	123.1	120.8	
C12—C13—C14	119	120.0	118.2	119.3	
C13—C14—C15	122	121.2	120.7	121.5	
C14—C15—C10	117.2	117.9	117.3	117.6	
C14—C15—C16	118.3	117.2	118.7	117.3	
C10-C15-C16	124.4	124.9	124.0	125.1	
C15—C16—C17	128.2	131.2	129.0	131.4	
C9—C17—C16	124.9	125.2	124.5	125.3	

**Table S4.** Selected intramolecular experimental (X-ray) and computational optimizedgeometrical parameters (bond angles, °) of compounds 25 and 34.

С9—С17—С18	116.5	119.5	117.3	119.4
C16—C17—C18	118.6	115.2	118.1	115.2
C17—C18—O2	120.3	121.6	120.4	121.5
C17—C18—C19	118.1	118.3	118.4	118.4
O2-C18-C19	121.5	120.1	121.2	120.1
C8—C19—C18	117.7	116.0	118.4	116.2
C8—C19—C20	123.5	126.1	123.8	126.1
C18—C19—C20	118.7	117.6	117.8	117.4
C19—C20—C21	130.8	127.1	131.4	127.2
C20—C21—C22	118.8	118.8	117.2	119.0
C20—C21—C26	124.3	122.7	125.3	122.8
C22—C21—C26	116.9	118.5	117.5	118.1
C21—C22—C23	121.2	120.7	122.0	121.0
C22—C23—C24	120.3	120.1	117.9	119.5
C23—C24—C25	119.8	119.8	123.0	120.9
C24—C25—C26	119.8	120.3	118.3	119.6
C25—C26—C21	122.1	120.6	121.3	120.9
C2—C3—Cl1	120.2	119.6		
C4—C3—Cl1	119.9	119.9		
C23—C24—F1			119.4	119.5
C25—C24—F1			117.6	119.6
C11—C12—F2			119.3	119.6
C13—C12—F2			117.6	119.6
RMSE		1.602		1.493
Maximum difference		3.9		4.2

Entry	ID	Coefficient	S	t	Descriptor
1	0	-0.833447	0.542	-1.536	Intercept
2	$D_1$	0.102299	0.007	15.070	Tot. molecular 1-center E-E
					repulsion / # of atoms
3	$D_2$	-88.6353	19.707	-4.497	Min. 1-electron react. index for
					atom O
4	$D_3$	-2.287	0.419	-5.456	Average information content
					(order 0)
$N = 24, n = 3, R^2 = 0.934, R^2 \text{cvOO} = 0.905, R^2 \text{cvMO} = 0.909, F = 93.751, s^2 = 0.019$					
$1/IC_{50} (\mu M) = -0.833447 + (0.102299 \text{ x } D_1) - (88.6353 \text{ x } D_2) - (2.287 \text{ x } D_3)$					

**Table S5.** Descriptors of the BMLR-QSAR model for the synthesizedpiperidinecarboxamides 24–47 against HCT116 (colon) carcinoma cell line.

**Table S6.** Descriptors of the BMLR-QSAR model for the synthesizedpiperidinecarboxamides 24–47 against MCF7 (breast) carcinoma cell line.

Entry	ID	Coefficient	S	t	Descriptor	
1	0	-1.26351	0.520	-2.431	Intercept	
2	$D_1$	4.27333	0.260	16.432	FPSA-2 Fractional PPSA	
					(PPSA-2/TMSA) (MOPAC PC)	
3	$D_2$	-0.0318232	0.006	-5.289	HA dependent HDSA-1 (Zefirov	
					PC)	
4	$D_3$	-866.471	104.142	-8.320	Partial charged surface area	
					(MOPAC PC) for atom C	
$N = 24, n = 3, R^2 = 0.951, R^2 \text{cvOO} = 0.931, R^2 \text{cvMO} = 0.933, F = 130.382, s^2 = 0.125$						
IC <sub>50</sub> (µ	$IC_{50} (\mu M) = -1.26351 + (4.27333 \text{ x } D_1) - (0.0318232 \text{ x } D_2) - (866.471 \text{ x } D_3)$					

Entry	ID	Coefficient	S	t	Descriptor		
1	0	166.573	19.779	8.422	Intercept		
2	$D_1$	0.068221	0.006	10.621	Count of H-donors sites		
					(MOPAC PC)		
3	$D_2$	-5.34709	1.134	-4.713	Min. resonance energy for bond		
					H-C		
4	$D_3$	-0.343666	0.056	-6.187	Min. e-n attraction for atom N		
$N = 24, n = 3, R^2 = 0.901, R^2 \text{cvOO} = 0.859, R^2 \text{cvMO} = 0.861, F = 60.784, s^2 = 0.012$							
$\log(\text{IC}_{50}, \mu\text{M}) = 166.573 + (0.068221 \text{ x } D_1) - (5.34709 \text{ x } D_2) - (0.343666 \text{ x } D_3)$							

**Table S7.** Descriptors of the BMLR-QSAR model for the synthesizedpiperidinecarboxamides 24–47 against A431 (squamous) carcinoma cell line.

Entry	Compd	Descriptor	Descriptors <sup><i>a</i></sup>		
Lifti y	Compa.	D <sub>1</sub>	$D_2$	D <sub>3</sub>	
1	24	51.78765	-0.00011	1.58224	
2	25	56.32968	0.00004	1.6951	
3	26	53.2964	-0.00043	1.72105	
4	27	50.09242	-0.00489	1.72733	
5	28	50.50078	-0.00001	1.62076	
6	29	60.69776	-0.00073	1.76818	
7	30	65.15003	0	1.82535	
8	31	61.53756	-0.00023	1.8984	
9	32	59.69894	-0.00263	1.92877	
10	33	58.90671	0.00002	1.80133	
11	34	64.11513	-0.00015	1.76818	
12	35	68.54209	-0.00024	1.87833	
13	36	64.7134	-0.00005	1.8984	
14	37	63.41592	-0.00398	1.92877	
15	38	62.16622	-0.00418	1.80133	
16	39	49.26383	-0.00335	1.64676	
17	40	50.76083	-0.00007	1.75654	
18	41	47.42904	-0.00261	1.74045	
19	42	48.2028	0	1.66427	
20	43	54.56279	-0.00167	1.80272	
21	44	58.44992	-0.00308	1.90463	
22	45	55.67702	-0.00037	1.85923	
23	46	53.28805	-0.00354	1.90057	
24	47	53.25566	-0.00133	1.81451	

**Table S8.** Molecular descriptor values of the BMLR-QSAR model for the synthesizedpiperidinecarboxamides 24–47 against HCT116 (colon) carcinoma cell line.

<sup>*a*</sup>  $D_1$  = Tot. molecular 1-center E-E repulsion / # of atoms,  $D_2$  = Min. 1-electron react. index for atom O,  $D_3$  = Average information content (order 0).

Entry	Compd	Descriptor	cs <sup>a</sup>		
Liiti y	Compu.	<b>D</b> <sub>1</sub>	$D_2$	D <sub>3</sub>	
1	24	2.24689	40.07887	0.00655	
2	25	2.12439	42.94164	0.00456	
3	26	2.50963	81.11199	0.00505	
4	27	2.28674	41.51026	0.00528	
5	28	2.30862	22.90221	0.00619	
6	29	1.81189	58.68691	0.0035	
7	30	1.3958	33.39906	0.00317	
8	31	1.8106	74.43218	0.00296	
9	32	1.72692	62.02682	0.00234	
10	33	1.84054	45.80442	0.00363	
11	34	2.05609	35.30757	0.00499	
12	35	1.71978	34.35332	0.00408	
13	36	2.2252	83.02051	0.00395	
14	37	2.0041	41.51026	0.004	
15	38	2.23988	52.0071	0.00476	
16	39	2.73898	61.07256	0.00466	
17	40	2.86792	79.68061	0.00455	
18	41	2.65675	60.1183	0.00382	
19	42	2.80097	38.17035	0.00506	
20	43	2.71423	93.51736	0.00344	
21	44	2.4645	93.51736	0.00272	
22	45	2.85685	91.13171	0.00256	
23	46	2.70558	85.88329	0.00282	
24	47	2.73771	80.15774	0.00363	

**Table S9.** Molecular descriptor values of the BMLR-QSAR model for the synthesizedpiperidinecarboxamides 24–47 against MCF7 (breast) carcinoma cell line.

<sup>*a*</sup>  $D_1$  = FPSA-2 Fractional PPSA (PPSA-2/TMSA) (MOPAC PC),  $D_2$  = HA dependent HDSA-1 (Zefirov PC),  $D_3$  = Partial charged surface area (MOPAC PC) for atom C.

Fntry	Compd	Descriptors <sup>a</sup>		
Linu y	Compu.	D <sub>1</sub>	D <sub>2</sub>	D <sub>3</sub>
1	24	7	10.3613	324.0338
2	25	7	10.3565	323.9441
3	26	10	10.3648	324.111
4	27	12	10.3565	324.1108
5	28	9	10.3599	324.0577
6	29	7	10.4147	323.9467
7	30	7	10.3771	324.7383
8	31	10	10.3843	324.3601
9	32	12	10.4167	324.5874
10	33	9	10.3675	324.459
11	34	7	10.3789	324.594
12	35	7	10.3786	324.5205
13	36	10	10.4128	324.693
14	37	12	10.3689	325.709
15	38	9	10.3669	324.7718
16	39	13	10.3766	324.5288
17	40	16	10.3811	324.0791
18	41	18	10.4128	324.7377
19	42	15	10.3611	324.0454
20	43	13	10.4129	323.8358
21	44	13	10.3893	324.1668
22	45	16	10.3517	323.823
23	46	18	10.3888	324.4444
24	47	15	10.3635	324.0558

**Table S10.** Molecular descriptor values of the BMLR-QSAR model for the synthesizedpiperidinecarboxamides 24–47 against A431 (squamous) carcinoma cell line.

<sup>*a*</sup>  $D_1$  = Count of H-donors sites (MOPAC PC),  $D_2$  = Min. resonance energy for bond H-C,  $D_3$  = Min. e-n attraction for atom N.

Crystal data	Compound 25	Compound 34
Chemical formula	$C_{26}H_{21}ClN_2O_2$	$C_{26}H_{20}F_2N_2O_2$
$M_{ m r}$	428.90	430.45
Crystal system, space group	Monoclinic, C2/c	Monoclinic, P21/n
Temperature (K)	293	293
<i>a</i> , <i>b</i> , <i>c</i> (Å)	26.930 (3), 12.9179 (18),	14.374 (3), 8.0590 (16),
	12.9500 (14)	18.554 (4)
β (°)	107.491 (3)	96.61 (3)
$V(\text{\AA}^3)$	4296.7 (5)	2135.0 (8)
Ζ	8	4
Radiation type	Μο Κα	Μο Κα
$\mu (\mathrm{mm}^{-1})$	0.20	0.10
Crystal size (mm)	$0.28 \times 0.18 \times 0.14$	$0.05\times0.07\times0.07$
Data collection		
No. of measured,	11327, 2861, 1790	29314, 6693, 5048
independent and observed [I		
$> 2.0\sigma(I)$ ] reflections		
$R_{\rm int}$	0.036	0.056
$(\sin \theta / \lambda)_{max} (\text{\AA}^{-1})$	0.632	0.735
Refinement		
$R[F^2 > 2\sigma(F^2)], wR(F^2), S$	0.039, 0.077, 0.95	0.064, 0.177, 1.00
No. of reflections	1790	5048
No. of parameters	173	294
$\Delta \rho_{\text{max}}, \Delta \rho_{\text{min}} \ (e \ \text{\AA}^{-3})$	0.13, -0.16	0.49, -0.41

 Table S11. Crystal data and structure refinement parameters for compounds 25 and 34.



Fig. S1. IR spectrum of compound 24 (KBr pellet).



**Fig. S2.** <sup>1</sup>H-NMR spectrum of compound **24** in DMSO- $d_6$ .



**Fig. S3.** <sup>13</sup>C-NMR spectrum of compound **24** in DMSO- $d_6$ .



Fig. S4. IR spectrum of compound 25 (KBr pellet).



**Fig. S5.** <sup>1</sup>H-NMR spectrum of compound **25** in DMSO- $d_6$ .



**Fig. S6.** <sup>13</sup>C-NMR spectrum of compound **25** in DMSO- $d_6$ .



Fig. S7. IR spectrum of compound 26 (KBr pellet).



**Fig. S8.** <sup>1</sup>H-NMR spectrum of compound **26** in DMSO- $d_6$ .



**Fig. S9.** <sup>13</sup>C-NMR spectrum of compound **26** in DMSO- $d_6$ .



Fig. S10. IR spectrum of compound 27 (KBr pellet).



**Fig. S11.** <sup>1</sup>H-NMR spectrum of compound **27** in DMSO- $d_6$ .



Fig. S12. <sup>13</sup>C-NMR spectrum of compound 27 in DMSO- $d_6$ .



Fig. S13. IR spectrum of compound 28 (KBr pellet).



**Fig. S14.** <sup>1</sup>H-NMR spectrum of compound **28** in DMSO- $d_6$ .


Fig. S15. <sup>13</sup>C-NMR spectrum of compound 28 in DMSO- $d_6$ .



Fig. S16. IR spectrum of compound 29 (KBr pellet).



**Fig. S17.** <sup>1</sup>H-NMR spectrum of compound **29** in DMSO- $d_6$ .



**Fig. S18.** <sup>13</sup>C-NMR spectrum of compound **29** in DMSO- $d_6$ .



Fig. S19. IR spectrum of compound 30 (KBr pellet).



**Fig. S20.** <sup>1</sup>H-NMR spectrum of compound **30** in DMSO- $d_6$ .



Fig. S21. <sup>13</sup>C-NMR spectrum of compound 30 in DMSO- $d_6$ .



Fig. S22. IR spectrum of compound 31 (KBr pellet).



**Fig. S23.** <sup>1</sup>H-NMR spectrum of compound **31** in DMSO- $d_6$ .



**Fig. S24.** <sup>13</sup>C-NMR spectrum of compound **31** in DMSO- $d_6$ .



Fig. S25. IR spectrum of compound 32 (KBr pellet).



**Fig. S26.** <sup>1</sup>H-NMR spectrum of compound **32** in DMSO- $d_6$ .



Fig. S27. <sup>13</sup>C-NMR spectrum of compound 32 in DMSO- $d_6$ .



Fig. S28. IR spectrum of compound 33 (KBr pellet).



**Fig. S29.** <sup>1</sup>H-NMR spectrum of compound **33** in DMSO- $d_6$ .



**Fig. S30.** <sup>13</sup>C-NMR spectrum of compound **33** in DMSO- $d_6$ .



Fig. S31. IR spectrum of compound 34 (KBr pellet).



**Fig. S32.** <sup>1</sup>H-NMR spectrum of compound **34** in DMSO- $d_6$ .



Fig. S33. <sup>13</sup>C-NMR spectrum of compound 34 in DMSO- $d_6$ .



Fig. S34. IR spectrum of compound 35 (KBr pellet).



**Fig. S35.** <sup>1</sup>H-NMR spectrum of compound **35** in DMSO- $d_6$ .



**Fig. S36.** <sup>13</sup>C-NMR spectrum of compound **35** in DMSO- $d_6$ .



Fig. S37. IR spectrum of compound 36 (KBr pellet).



**Fig. S38.** <sup>1</sup>H-NMR spectrum of compound **36** in DMSO- $d_6$ .



**Fig. S39.** <sup>13</sup>C-NMR spectrum of compound **36** in DMSO- $d_6$ .



Fig. S40. IR spectrum of compound 37 (KBr pellet).



**Fig. S41.** <sup>1</sup>H-NMR spectrum of compound **37** in DMSO- $d_6$ .



Fig. S42. <sup>13</sup>C-NMR spectrum of compound 37 in DMSO- $d_6$ .



Fig. S43. IR spectrum of compound 38 (KBr pellet).



**Fig. S44.** <sup>1</sup>H-NMR spectrum of compound **38** in DMSO- $d_6$ .



Fig. S45. <sup>13</sup>C-NMR spectrum of compound 38 in DMSO- $d_6$ .



Fig. S46. IR spectrum of compound 39 (KBr pellet).



**Fig. S47.** <sup>1</sup>H-NMR spectrum of compound **39** in DMSO- $d_6$ .



Fig. S48. <sup>13</sup>C-NMR spectrum of compound 39 in DMSO- $d_6$ .



Fig. S49. IR spectrum of compound 40 (KBr pellet).



**Fig. S50.** <sup>1</sup>H-NMR spectrum of compound **40** in DMSO- $d_6$ .


Fig. S51. <sup>13</sup>C-NMR spectrum of compound 40 in DMSO- $d_6$ .



Fig. S52. IR spectrum of compound 41 (KBr pellet).



Fig. S53. <sup>1</sup>H-NMR spectrum of compound 41 in DMSO- $d_6$ .



**Fig. S54.** <sup>13</sup>C-NMR spectrum of compound **41** in DMSO- $d_6$ .



Fig. S55. IR spectrum of compound 42 (KBr pellet).



**Fig. S56.** <sup>1</sup>H-NMR spectrum of compound **42** in DMSO- $d_6$ .



Fig. S57. <sup>13</sup>C-NMR spectrum of compound 42 in DMSO- $d_6$ .



Fig. S58. IR spectrum of compound 43 (KBr pellet).



**Fig. S59.** <sup>1</sup>H-NMR spectrum of compound **43** in DMSO- $d_6$ .



**Fig. S60.** <sup>13</sup>C-NMR spectrum of compound **43** in DMSO- $d_6$ .



Fig. S61. IR spectrum of compound 44 (KBr pellet).



**Fig. S62.** <sup>1</sup>H-NMR spectrum of compound 44 in DMSO- $d_6$ .



Fig. S63. <sup>13</sup>C-NMR spectrum of compound 44 in DMSO- $d_6$ .



Fig. S64. IR spectrum of compound 45 (KBr pellet).



**Fig. S65.** <sup>1</sup>H-NMR spectrum of compound **45** in DMSO- $d_6$ .



**Fig. S66.** <sup>13</sup>C-NMR spectrum of compound **45** in DMSO- $d_6$ .



Fig. S67. IR spectrum of compound 46 (KBr pellet).



**Fig. S68.** <sup>1</sup>H-NMR spectrum of compound **46** in DMSO- $d_6$ .



**Fig. S69.** <sup>13</sup>C-NMR spectrum of compound **46** in DMSO- $d_6$ .



Fig. S70. IR spectrum of compound 47 (KBr pellet).



**Fig. S71.** <sup>1</sup>H-NMR spectrum of compound 47 in DMSO- $d_6$ .



Fig. S72. <sup>13</sup>C-NMR spectrum of compound 47 in DMSO- $d_6$ .



Fig. S73. IR spectrum of compound 48 (KBr pellet).



**Fig. S74.** <sup>1</sup>H-NMR spectrum of compound **48** in DMSO- $d_6$ .



Fig. S75. <sup>13</sup>C-NMR spectrum of compound 48 in DMSO- $d_6$ .



Fig. S76. IR spectrum of compound 49 (KBr pellet).



**Fig. S77.** <sup>1</sup>H-NMR spectrum of compound **49** in DMSO- $d_6$ .



Fig. S78. <sup>13</sup>C-NMR spectrum of compound 49 in DMSO- $d_6$ .



Fig. S79. IR spectrum of compound 50 (KBr pellet).



**Fig. S80.** <sup>1</sup>H-NMR spectrum of compound **50** in DMSO- $d_6$ .



Fig. S81. <sup>13</sup>C-NMR spectrum of compound 50 in DMSO- $d_6$ .



Fig. S82. IR spectrum of compound 51 (KBr pellet).



**Fig. S83.** <sup>1</sup>H-NMR spectrum of compound **51** in DMSO- $d_6$ .



**Fig. S84.** <sup>13</sup>C-NMR spectrum of compound **51** in DMSO- $d_6$ .



**Fig. S85.** ORTEP view of compound **25** showing the atom-numbering scheme. Displacement ellipsoids are drawn at the 50 % probability level and H atoms are shown as small spheres of arbitrary radii.



Fig. S86. ORTEP view of compound 34 showing the atom-numbering scheme.
Displacement ellipsoids are drawn at the 50 % probability level and H atoms are shown as small spheres of arbitrary radii.


**Fig. S87**. The crystal packing of compound **25**. The H atoms not engaged in the intermolecular interactions (dashed lines) have been skipped for clarity.



**Fig. S88**. The crystal packing of compound **34**. The H atoms not engaged in the intermolecular interactions (dashed lines) have been skipped for clarity.



Fig. S89. A projection of the optimized structure of compound 25 by DFT/B3LYP method with 3-21G\* basis set.



Fig. S90. A projection of the optimized structure of compound 34 by DFT/B3LYP method with 3-21G\* basis set.



Fig. S91. Overlay diagram of compound 25; red (X-ray structure), green (DFT).



Fig. S92. Overlay diagram of compound 34; red (X-ray structure), green (DFT).



























































Fig. S93. Dose-response curve for the tested compounds against HCT116 (colon cancer) cell line.



























































Fig. S94. Dose-response curve for the tested compounds against MCF7 (breast cancer) cell line.
























































Fig. S95. Dose-response curve for the tested compounds against A431 (squemous cancer) cell line.

























































Fig. S96. Dose-response curve for the tested compounds against RPE1 (retinal pigment epithelium) cell line.



Fig. S97. BMLR-QSAR model plot of correlations representing the observed vs. predicted  $1/IC_{50}$ ,  $\mu$ M values for the tested compounds against HCT116 (colon) carcinoma cell line.



Fig. S98. BMLR-QSAR model plot of correlations representing the observed vs. predicted IC<sub>50</sub>,  $\mu$ M values for the tested compounds against MCF7 (breast) carcinoma cell line.



**Fig. S99.** BMLR-QSAR model plot of correlations representing the observed *vs.* predicted log(IC<sub>50</sub>),  $\mu$ M values for the tested compounds against A431 (squamous) carcinoma cell line.



Fig. S100. Constraint distances "H-1 – H-2 = 10.672, H-1 – H-3 = 7.810, H-2 – H-3 = 11.639, H-1 – HBA = 4.686, H-2 – HBA = 6.093, H-3 – HBA = 8.206 Å" of the generated 3D-pharmacophore for the tested piperidinecarboxamides 24–47 against HCT116 (colon) carcinoma cell line which contains three hydrophobics (H-1, H-2, H-3; light blue) and one hydrogen bonding acceptor (HBA; green).



Fig. S101. Constraint angles "H-1 – H-2 – H-3 = 40.69, H-3 – H-1 – HBA = 77.68, H-3 – H-2 – HBA = 42.09 °" of the generated 3D-pharmacophore for the tested piperidinecarboxamides 24–47 against HCT116 (colon) carcinoma cell line which contains three hydrophobics (H-1, H-2, H-3; light blue) and one hydrogen bonding acceptor (HBA; green).
























Fig. S102. 3D-pharmacophore model mapped on the tested piperidinecarboxamides 24–47 against HCT116 (colon) carcinoma cell line.



Fig. S103. Constraint distances "H-1 – H-2 = 9.356, H-1 – HBA = 8.527, H-2 – HBA = 6.917 Å" of the generated 3D-pharmacophore for the tested piperidinecarboxamides
24–47 against MCF7 (breast) carcinoma cell line which contains two hydrophobics (H-1, H-2; light blue) and one hydrogen bonding acceptor (HBA; green).



**Fig. S104.** Constraint angle "H-2 – H-1 – HBA = 45.21 °" of the generated 3Dpharmacophore for the tested piperidinecarboxamides **24–47** against MCF7 (breast) carcinoma cell line which contains two hydrophobics (H-1, H-2; light blue) and one hydrogen bonding acceptor (HBA; green).

























Fig. S105. 3D-pharmacophore model mapped on the tested piperidinecarboxamides 24–47 against MCF7 (breast) carcinoma cell line.



Fig. S106. Constraint distances "H-1 – H-2 = 8.270, H-1 – HBA-1 = 6.866, H-1 – HBA-2 = 4.726, H-2 – HBA-1 = 8.788, H-2 – HBA-2 = 4.683, HBA-1 – HBA-2 = 6.077 Å" of the generated 3D-pharmacophore for the tested piperidinecarboxamides 24–47 against A431 (squamous) carcinoma cell line which contains two hydrophobics (H-1, H-2; light blue) and two hydrogen bonding acceptor (HBA-1, HBA-2; green).



**Fig. S107.** Constraint angles "H-2 – H-1 – HBA-1 = 70.28, H-2 – HBA-2 – HBA-1 = 108.82 °" of the generated 3D-pharmacophore for the tested piperidinecarboxamides **24–47** against A431 (squamous) carcinoma cell line which contains two hydrophobics (H-1, H-2; light blue) and two hydrogen bonding acceptor (HBA-1, HBA-2; green).
























Fig. S108. 3D-pharmacophore model mapped on the tested piperidinecarboxamides 24–47 against A431 (squamous) carcinoma cell line.