

## Supporting Information

for

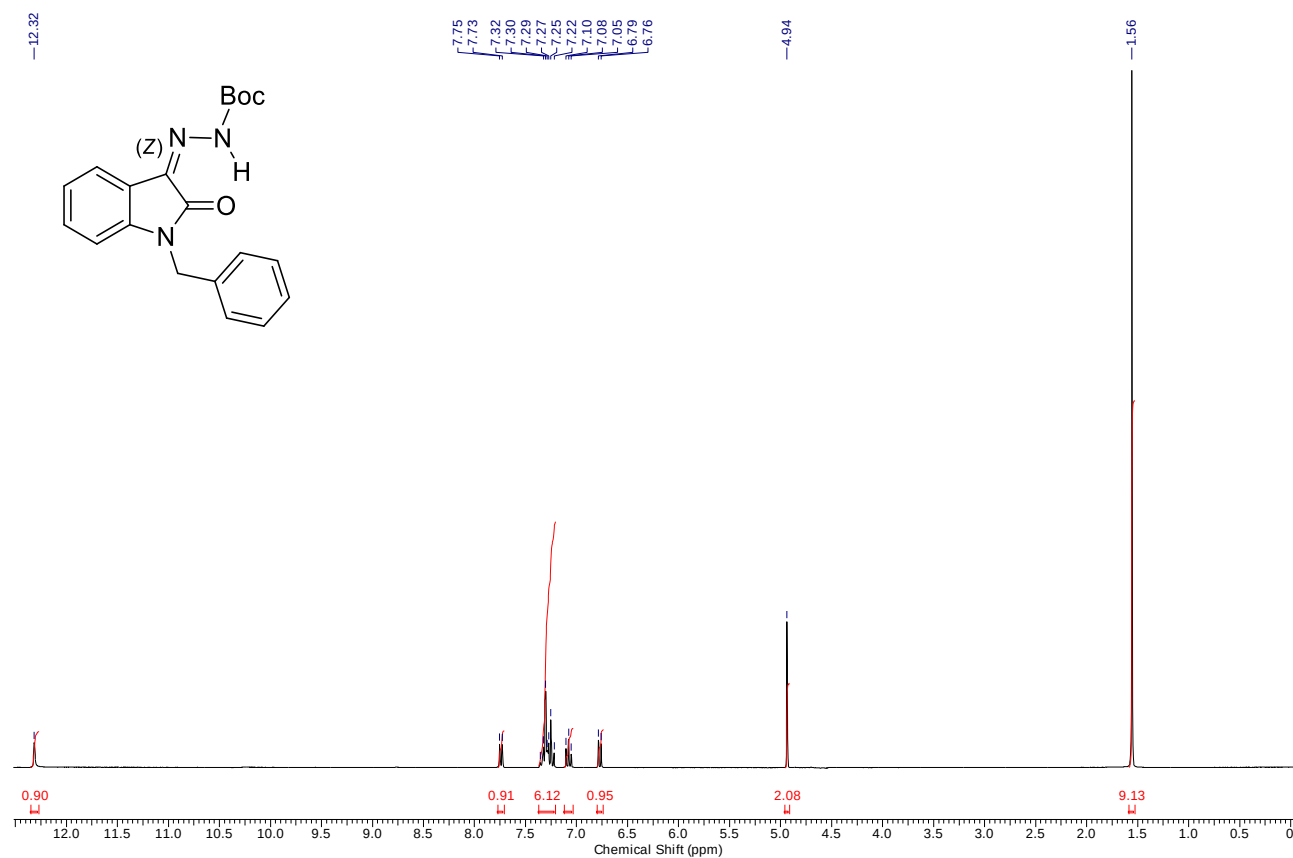
### **Allylation of isatin-derived N-Boc-hydrazones followed by Pd-catalyzed carboamination reaction: an entry to 3-spiro-pyrazolidyl-oxindoles**

Stefano Gazzotti, Marco Manenti, Leonardo Lo Presti and Alessandra Silvani\*

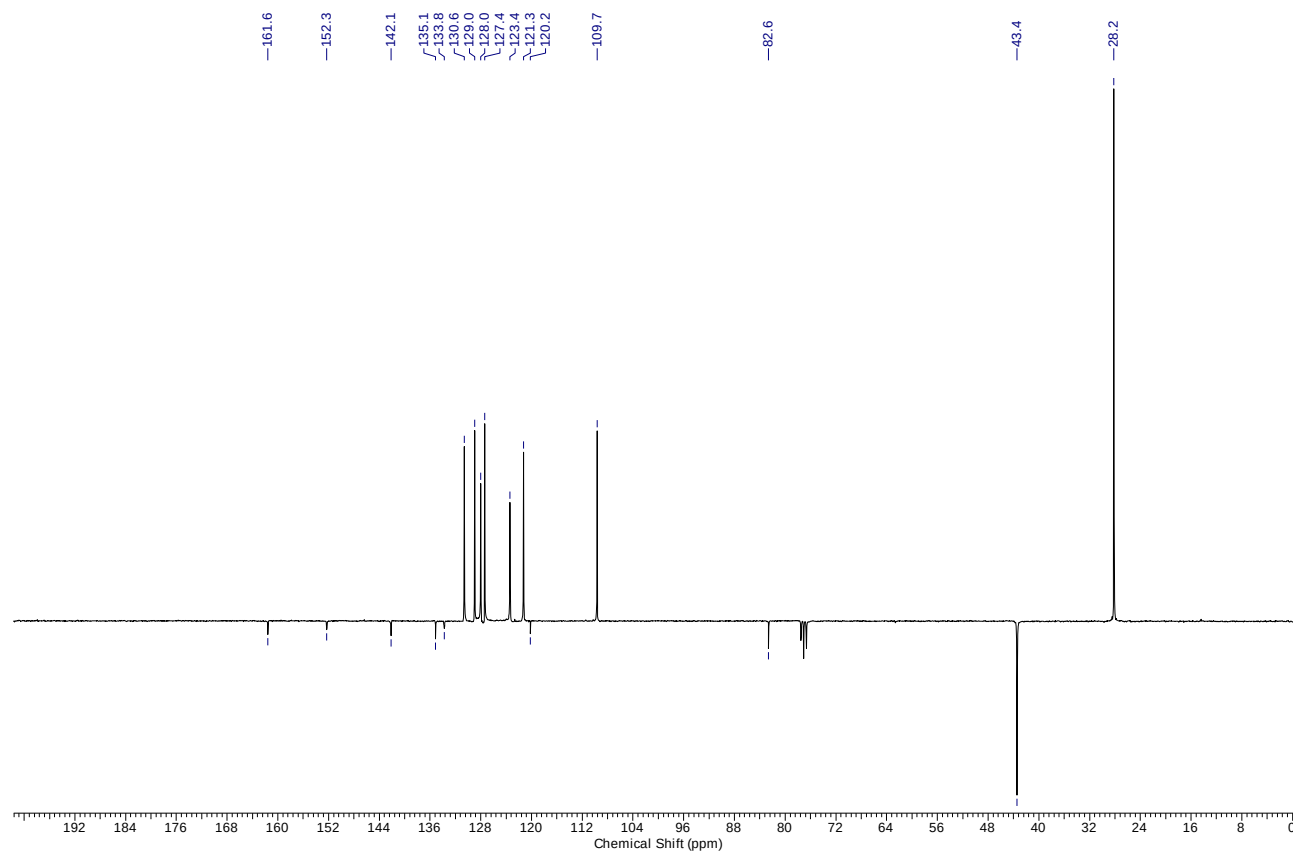
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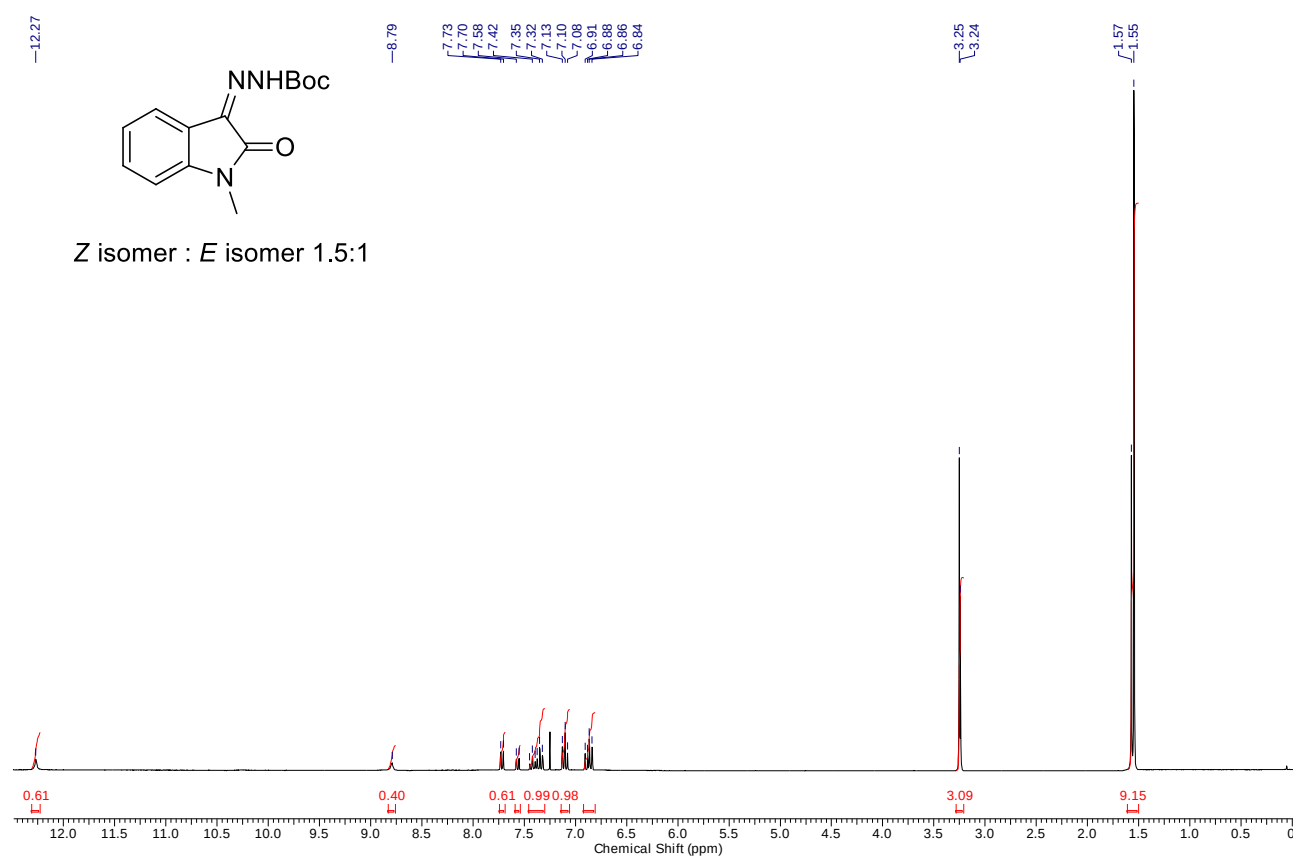
**4a**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



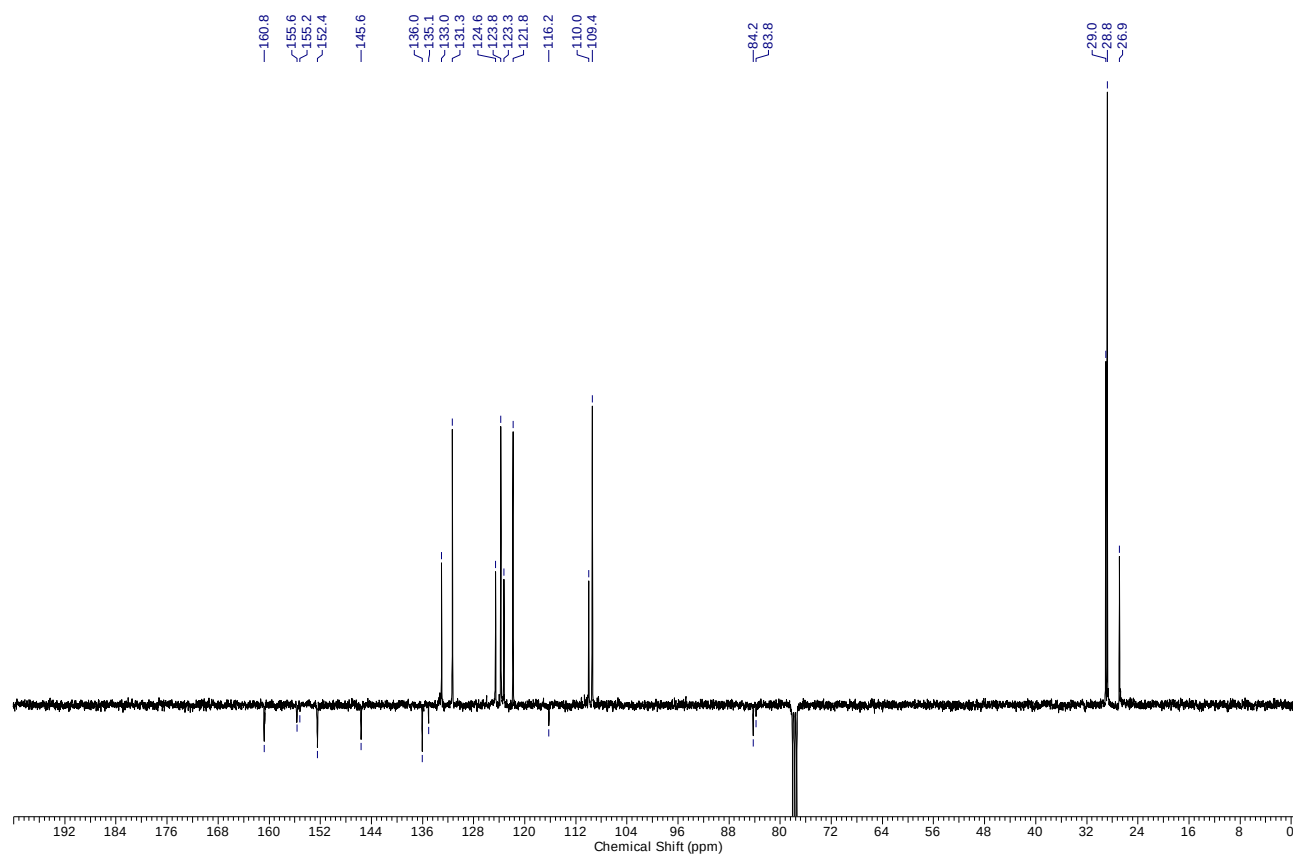
**4a**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



**4b**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



**4b**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



Chemical structure of compound 10: CC1=CC=C(C=C1)c2c(c3ccccc3n2C/C=C/c4ccc(Cl)c(Cl)c4)C(=O)NN1C(=O)OCC1(C)C

<sup>1</sup>H NMR spectrum (CDCl<sub>3</sub>) of compound 10. The x-axis represents the chemical shift in ppm, ranging from 0 to 12.0. The spectrum shows several peaks, with integration values provided below the baseline.

Peak list (Chemical Shift in ppm):

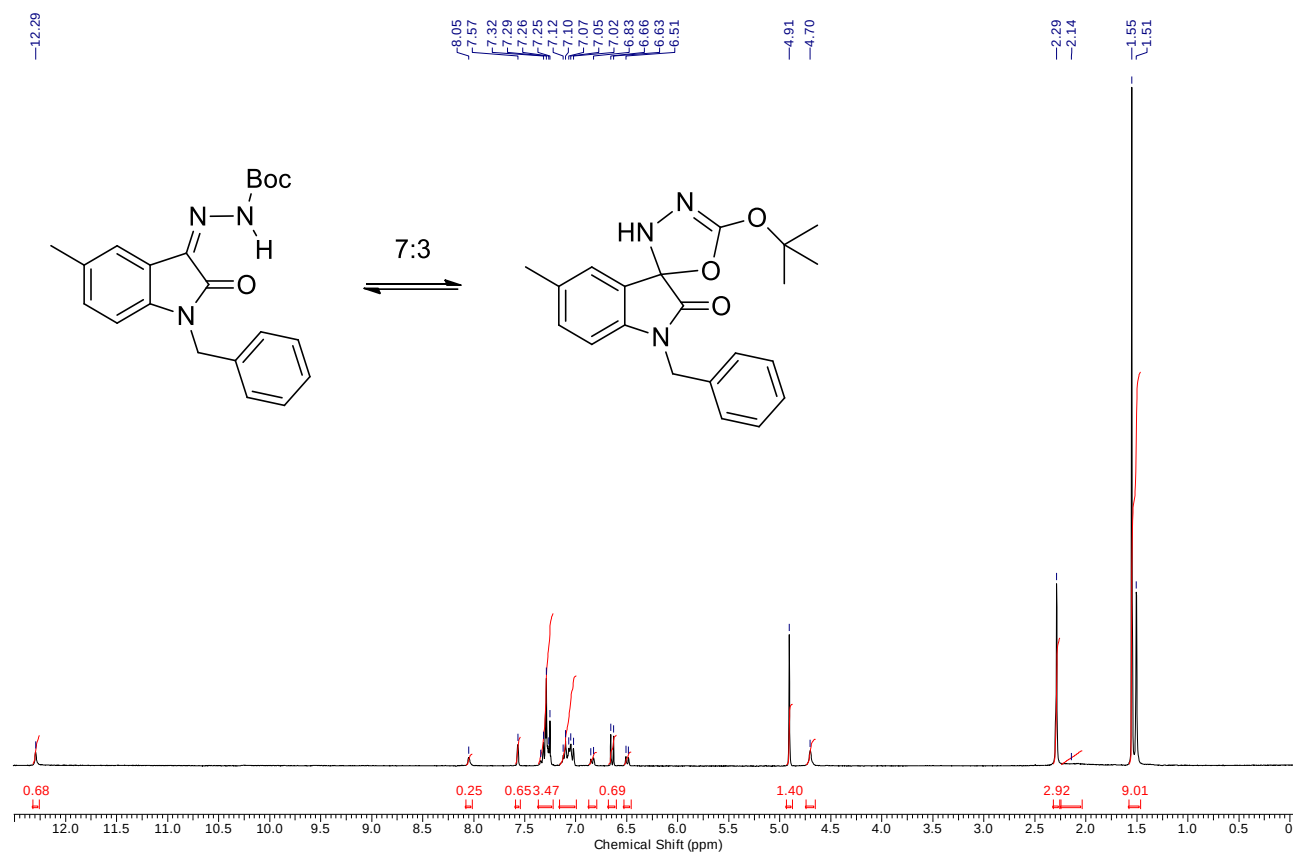
- 1.55 (t, 9.17H)
- 4.51 (d, 2.02H)
- 6.17 (m, 0.92H)
- 6.22 (m, 0.94H)
- 6.49 (m, 0.92H)
- 6.52 (m, 0.98H)
- 6.86 (m, 3.00H)
- 7.12 (m, 0.98H)
- 7.14 (m, 0.91H)
- 7.16 (m, 3.00H)
- 7.30 (m, 0.91H)
- 7.32 (m, 3.00H)
- 7.34 (m, 0.91H)
- 7.35 (m, 3.00H)
- 7.37 (m, 0.91H)
- 7.41 (m, 3.00H)
- 7.75 (m, 0.91H)
- 7.77 (m, 3.00H)

Integration values (from left to right): 0.90, 0.91, 3.00, 0.98, 0.92, 0.94, 2.02, 9.17.

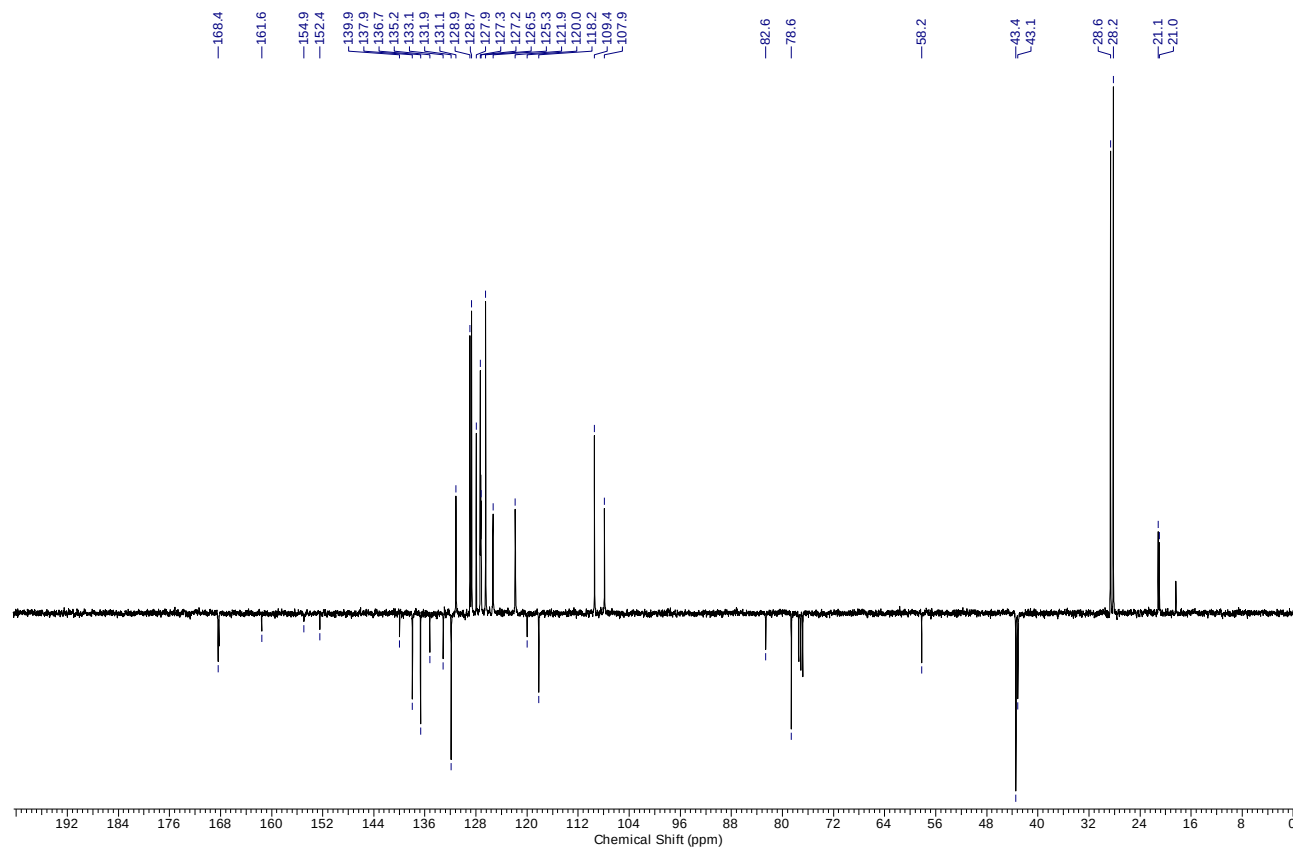
Chemical Shift (ppm)

161.2  
155.1  
152.2  
141.9  
136.0  
132.8  
132.3  
131.8  
131.0  
130.7  
128.3  
125.7  
124.2  
123.5  
121.4  
120.2  
109.3  
82.7  
41.3  
28.1

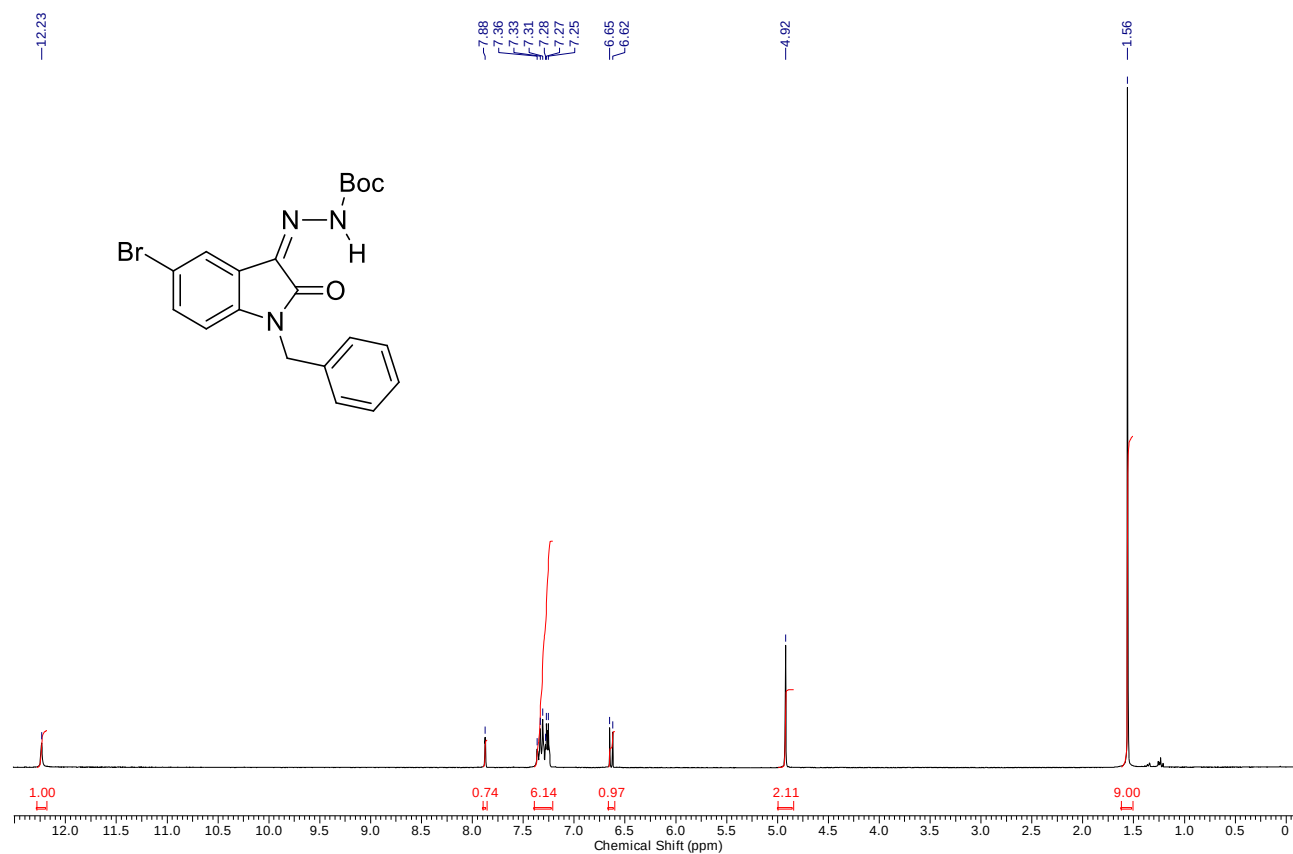
**4d**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



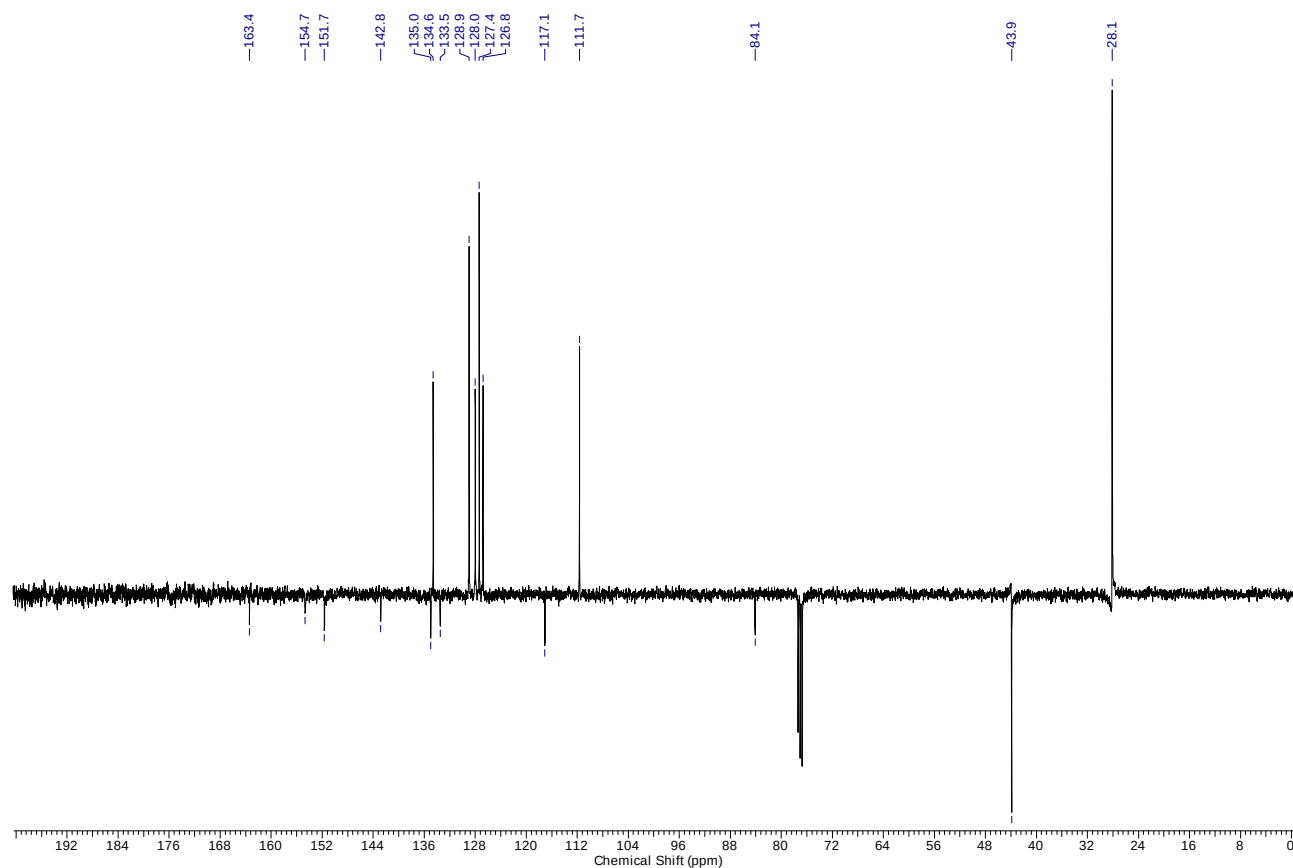
**4d**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



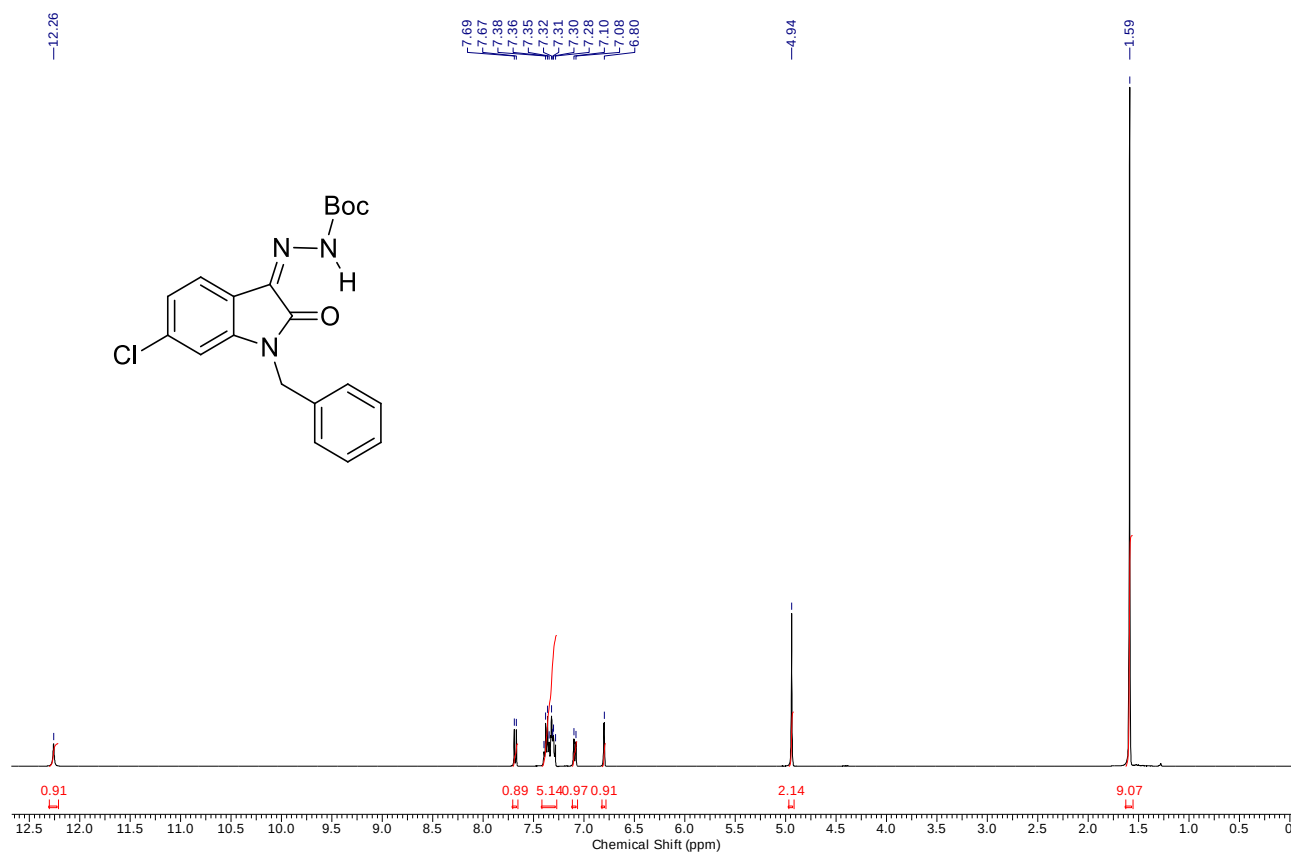
**4e**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



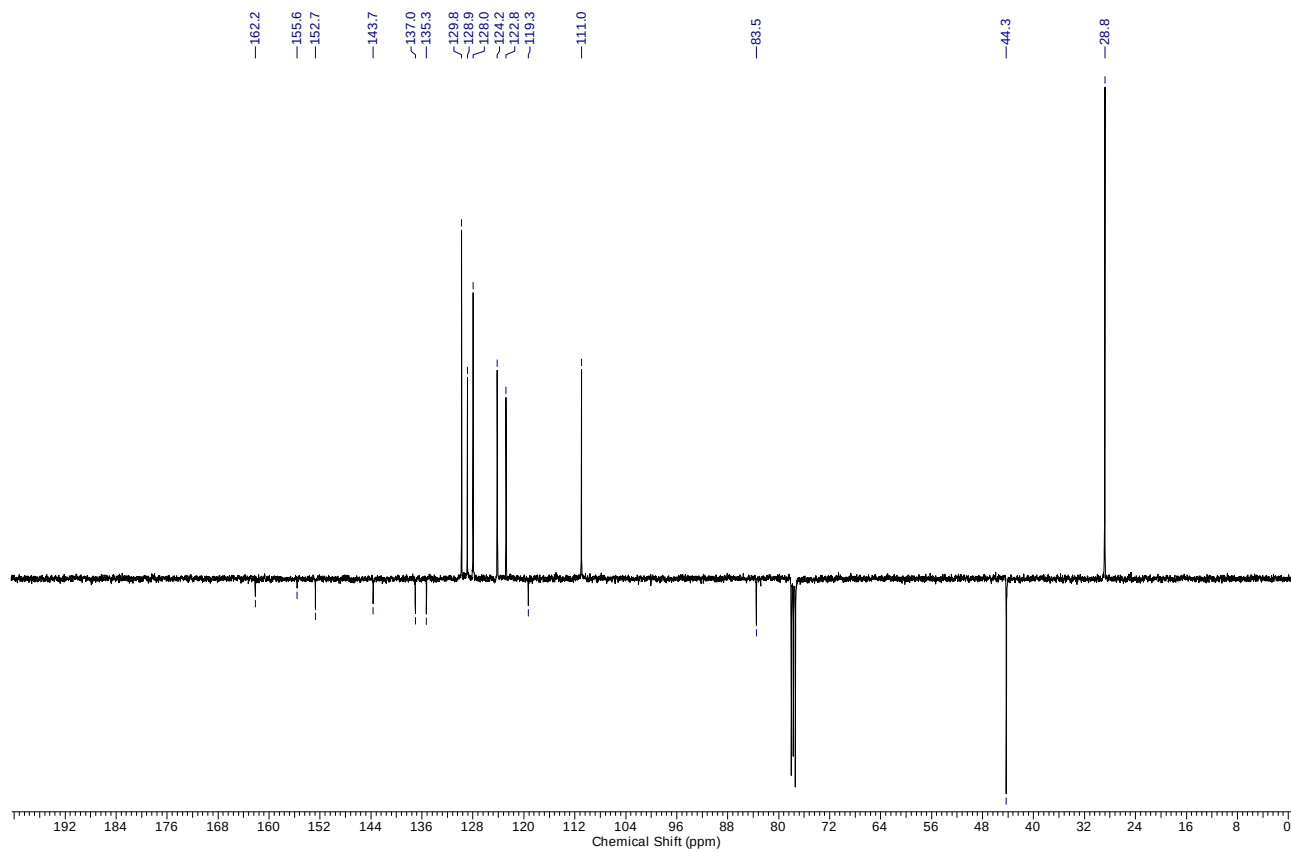
**4e**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):



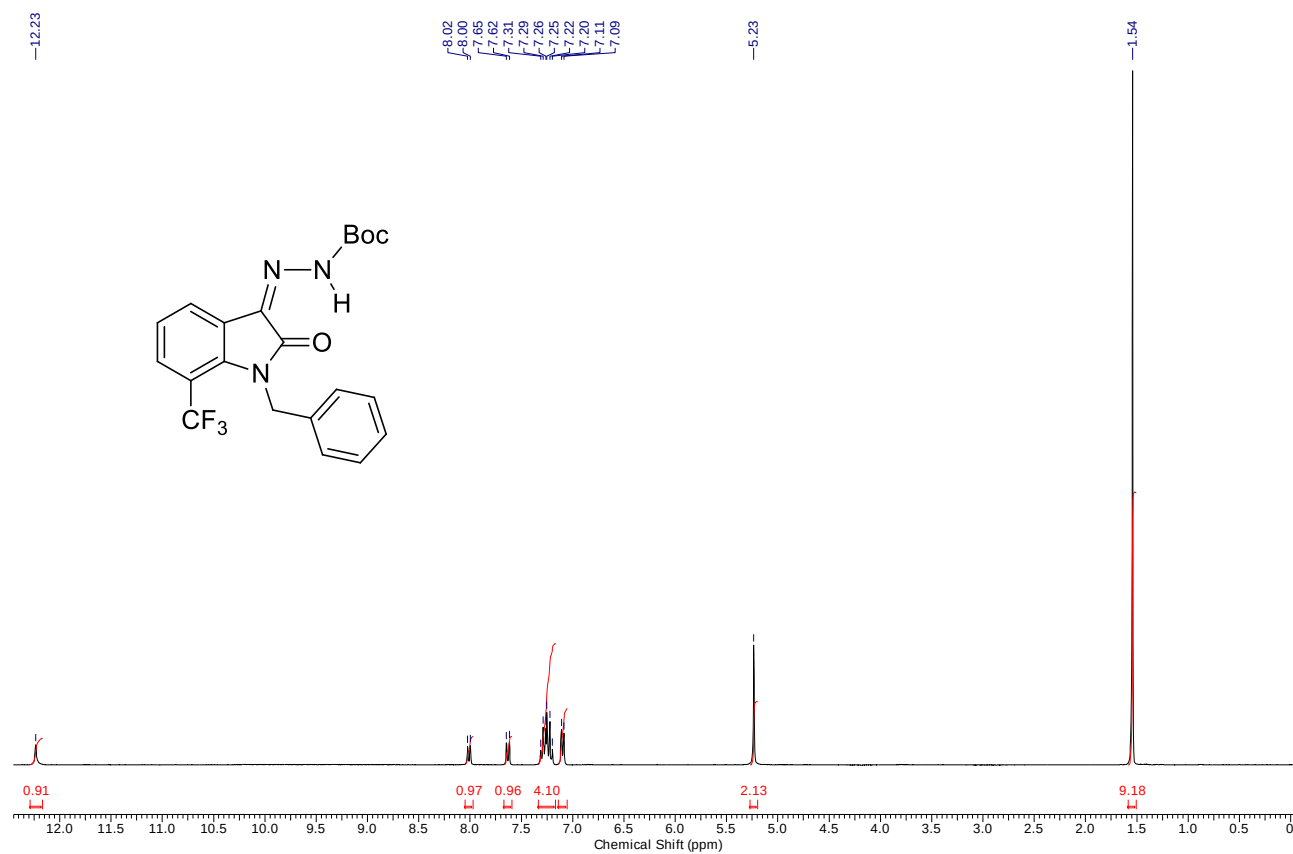
**4f**  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):



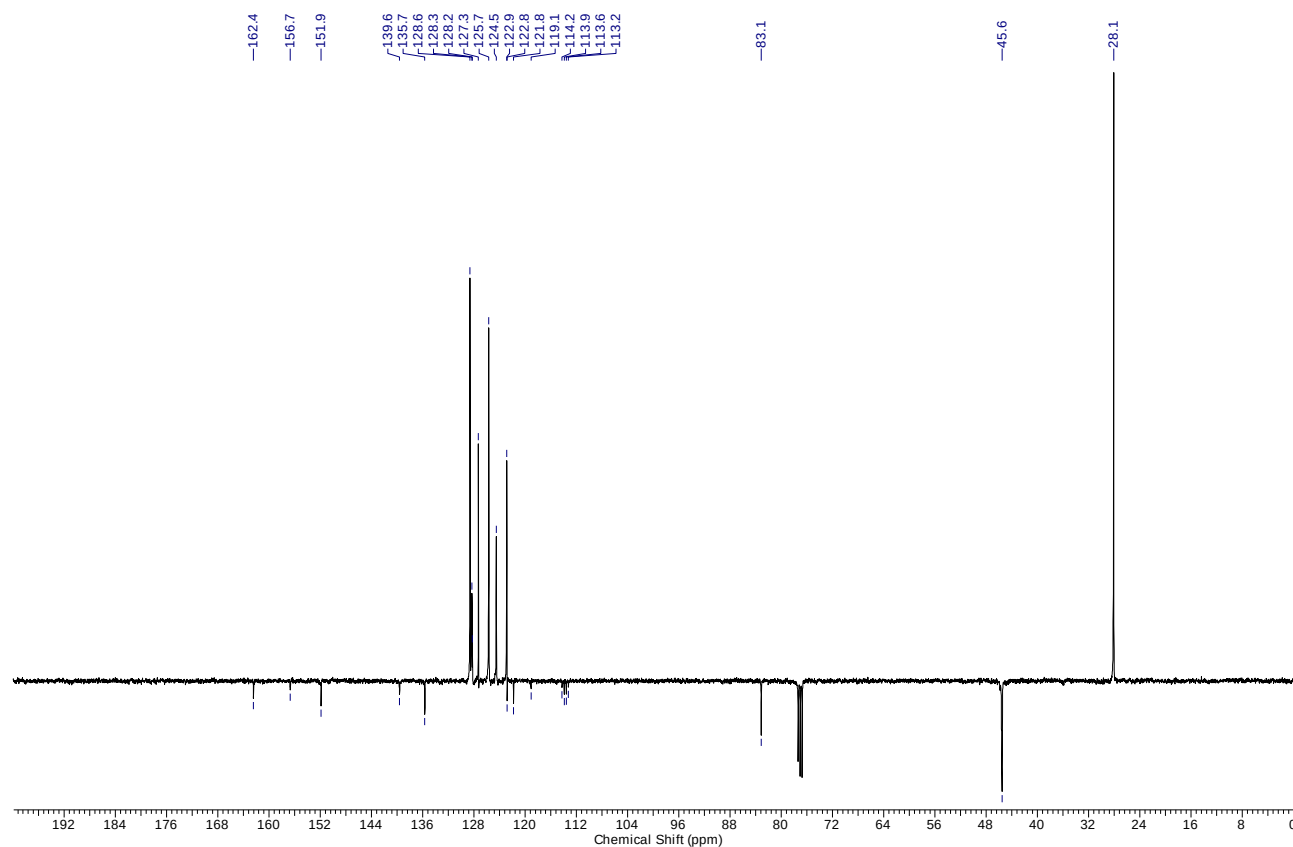
**4f**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):



**4g**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):

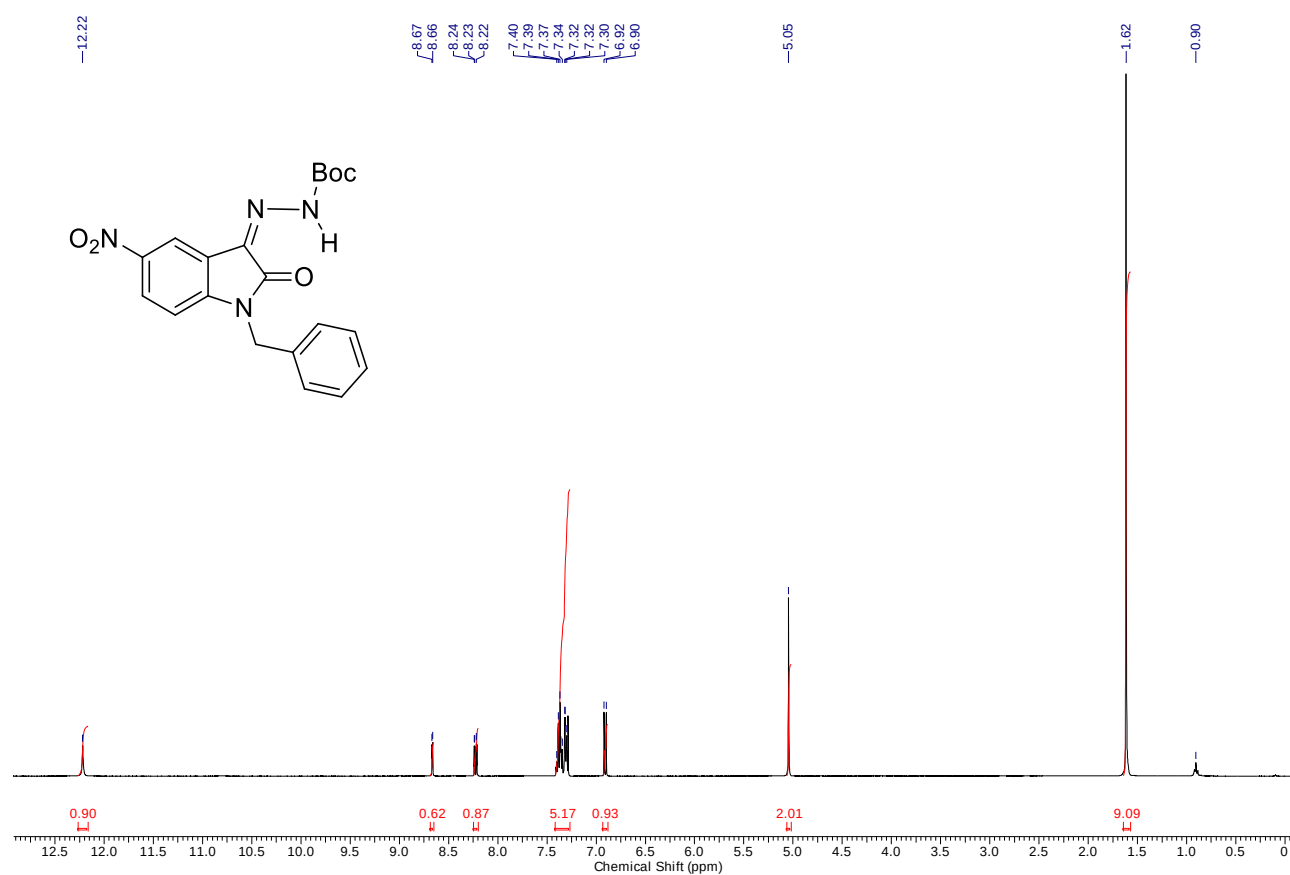


**4g**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):

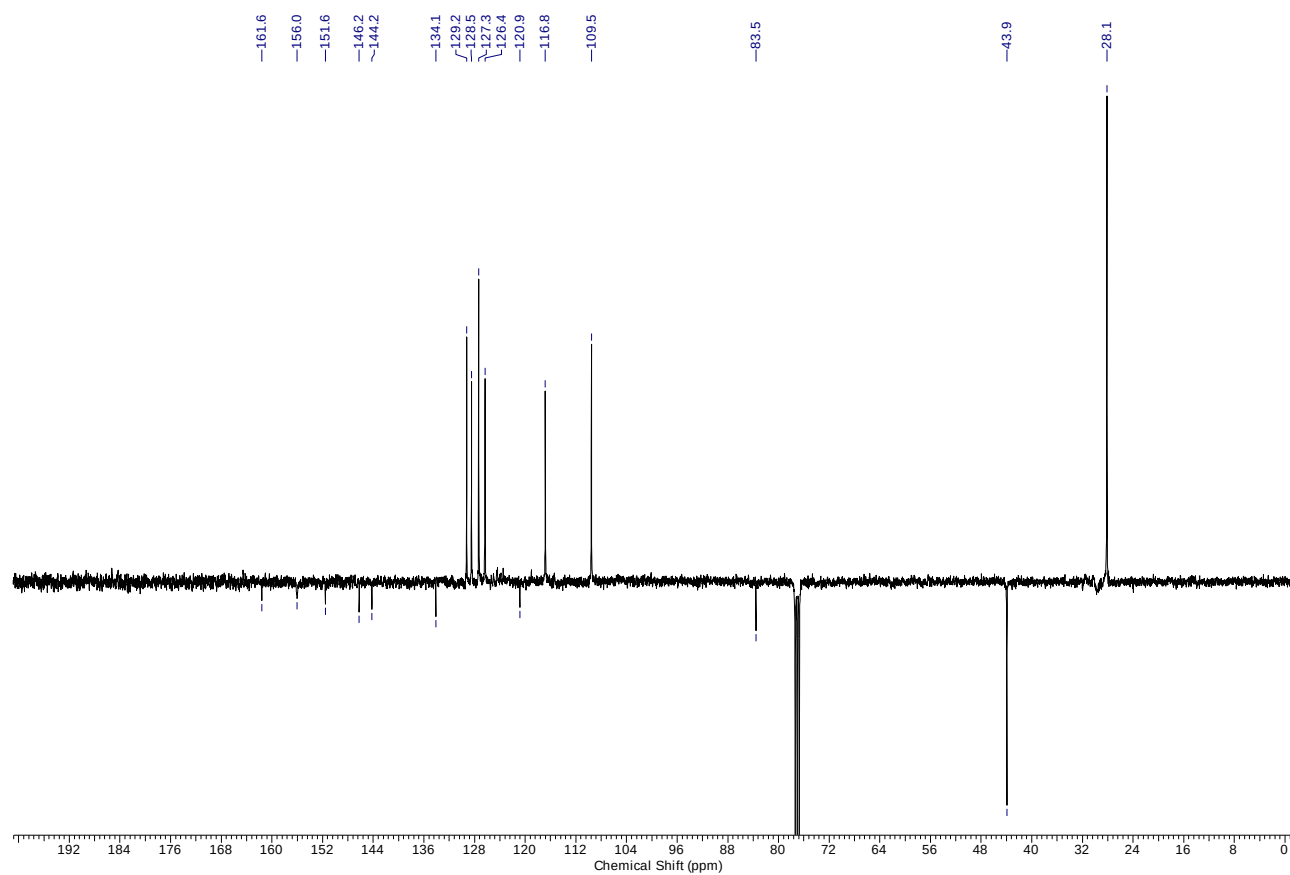




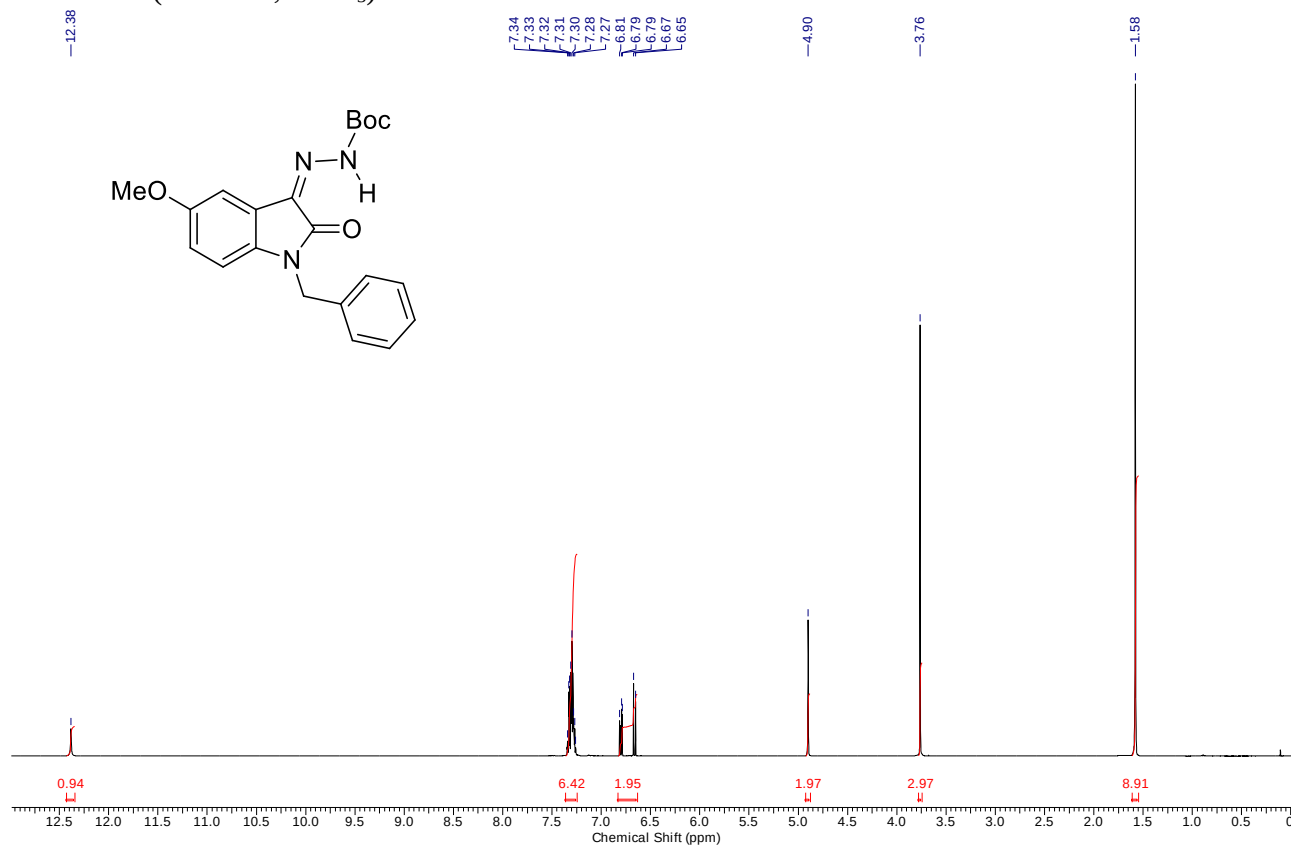
**4h**  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):



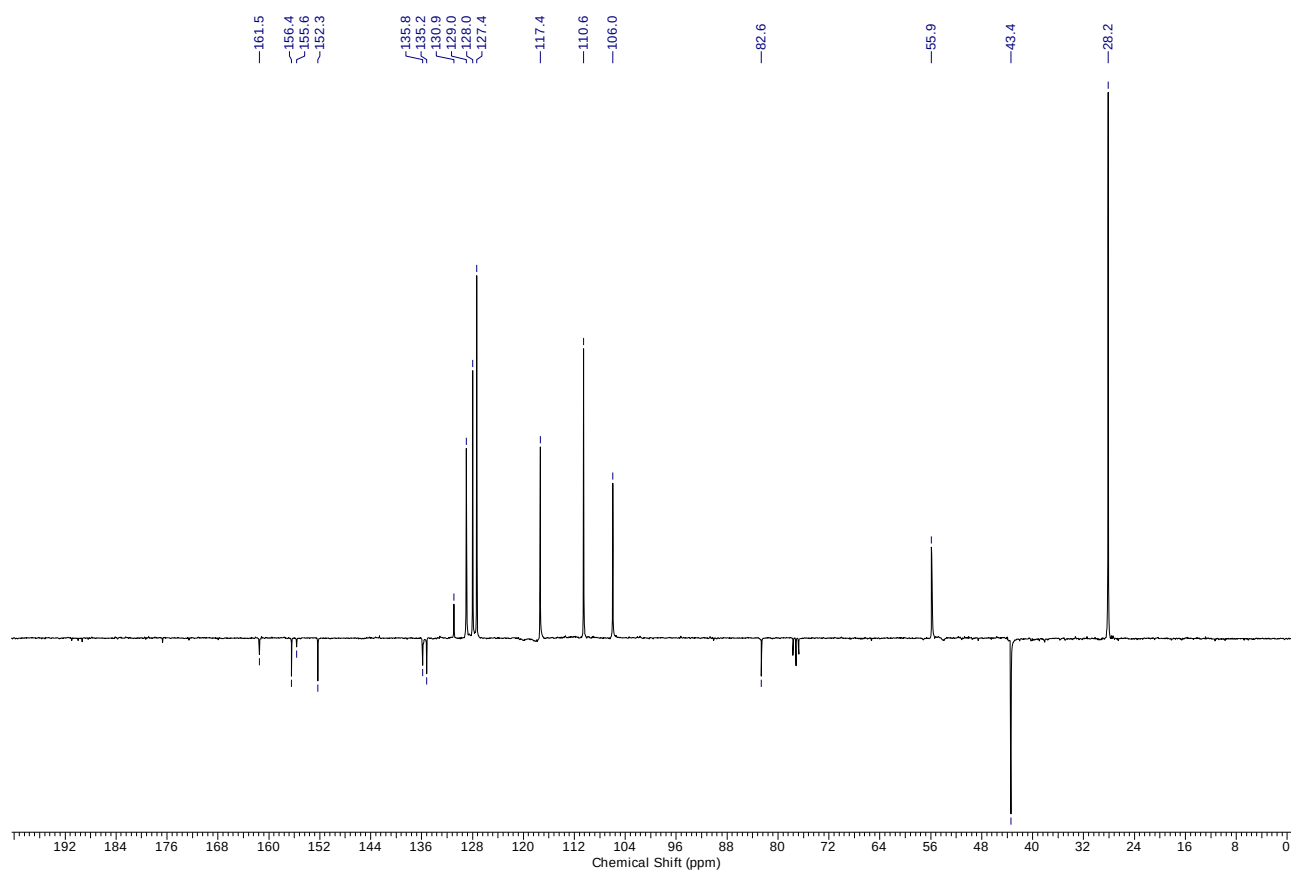
**4h**  $^{13}\text{C}$  NMR (101 MHz, APT,  $\text{CDCl}_3$ ):



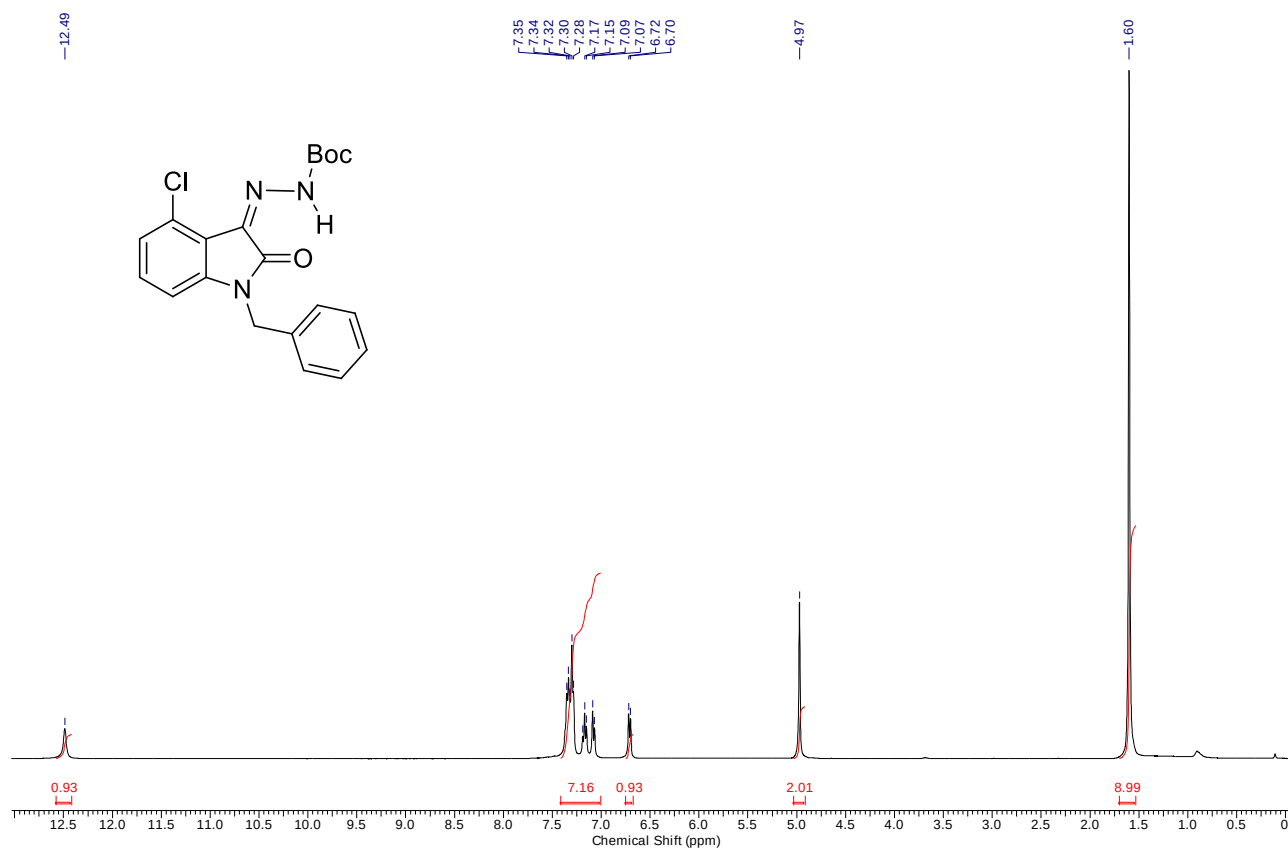
**4i**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



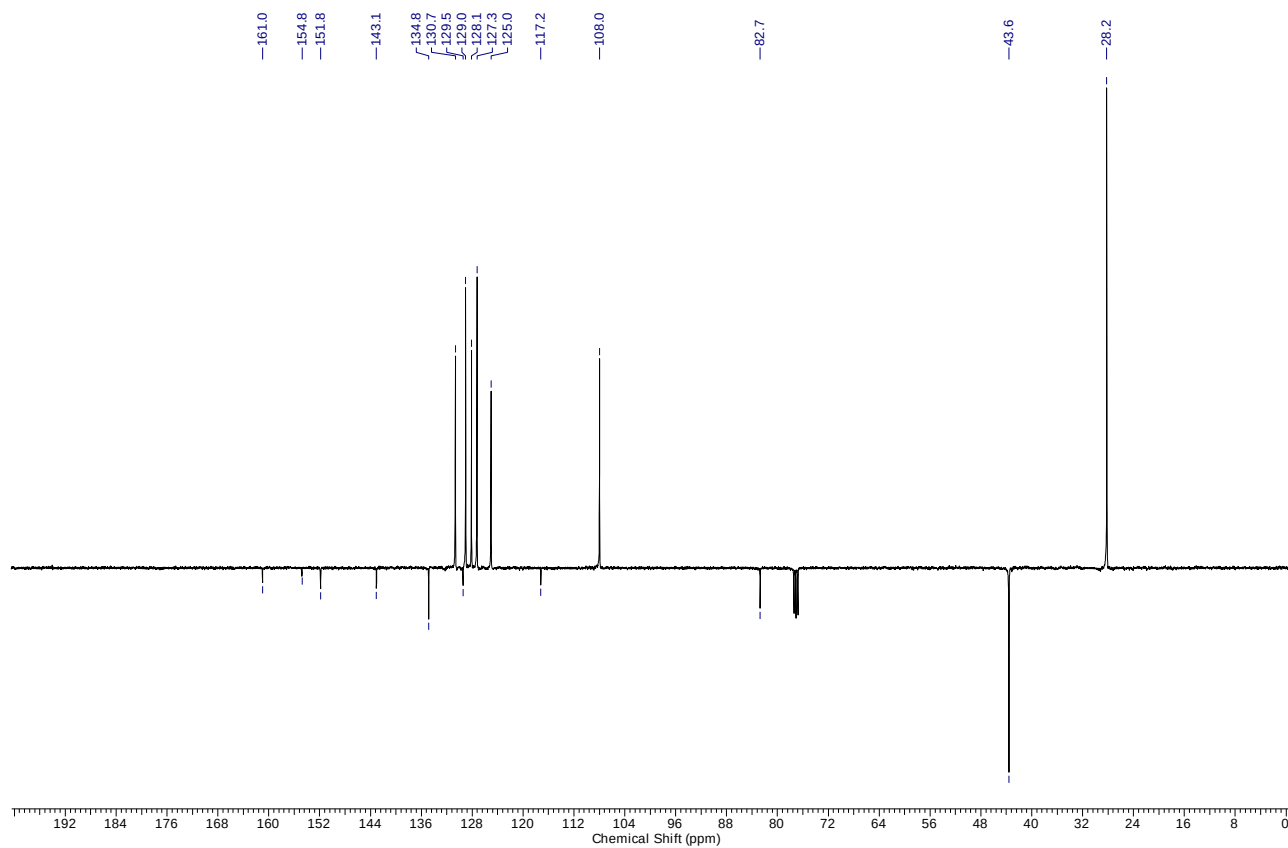
**4i**  $^{13}\text{C}$  NMR (75 MHz, APT,  $\text{CDCl}_3$ ):



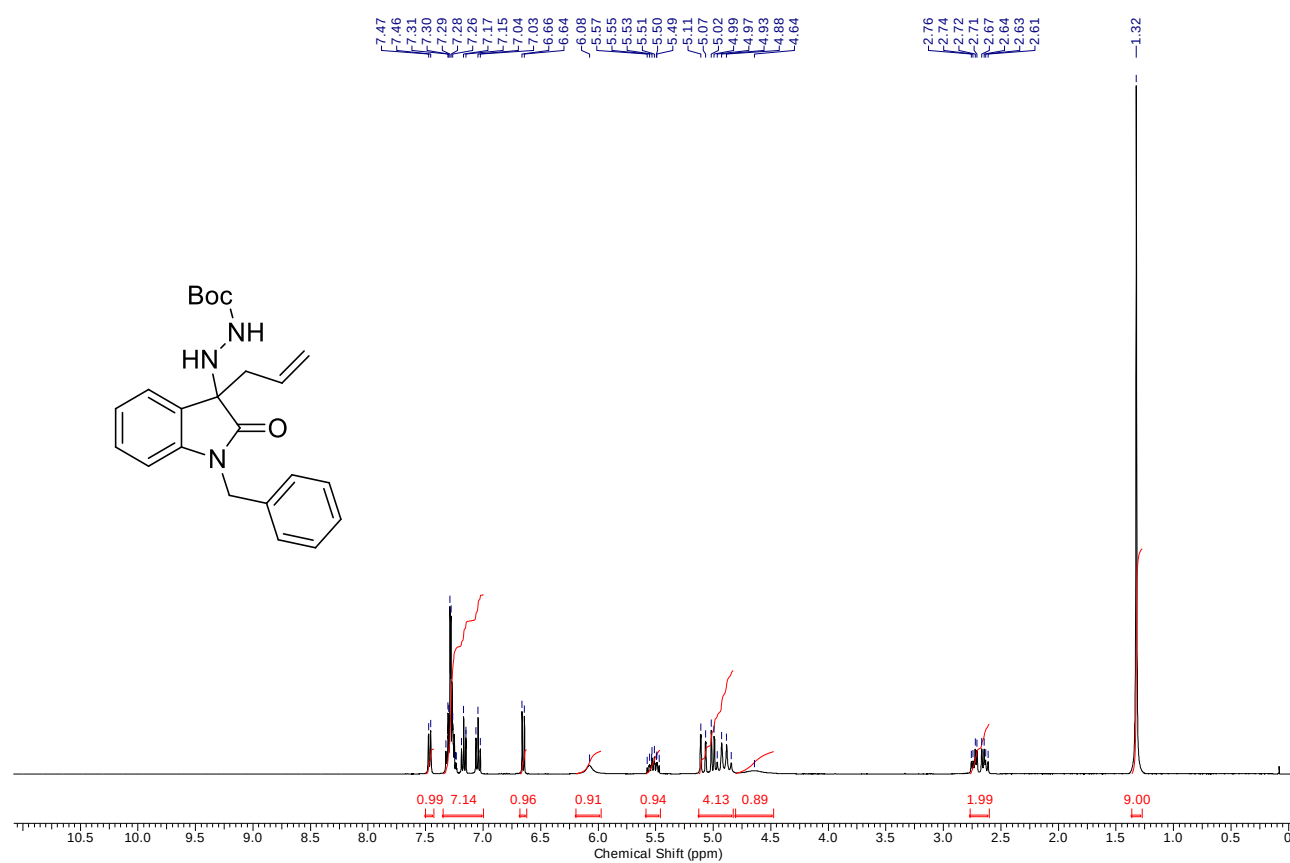
**4j**  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):



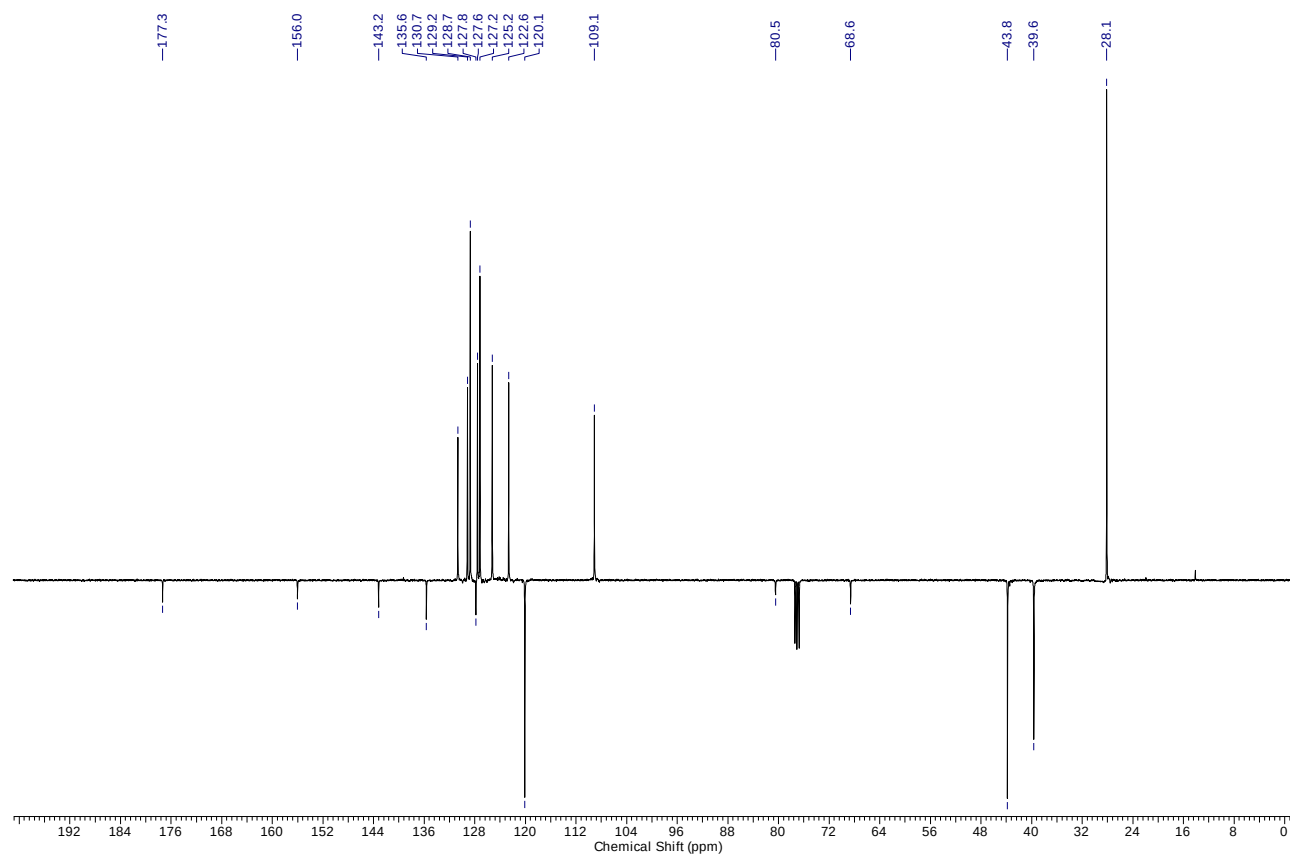
**4j**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):



**5a**  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):



**5a**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



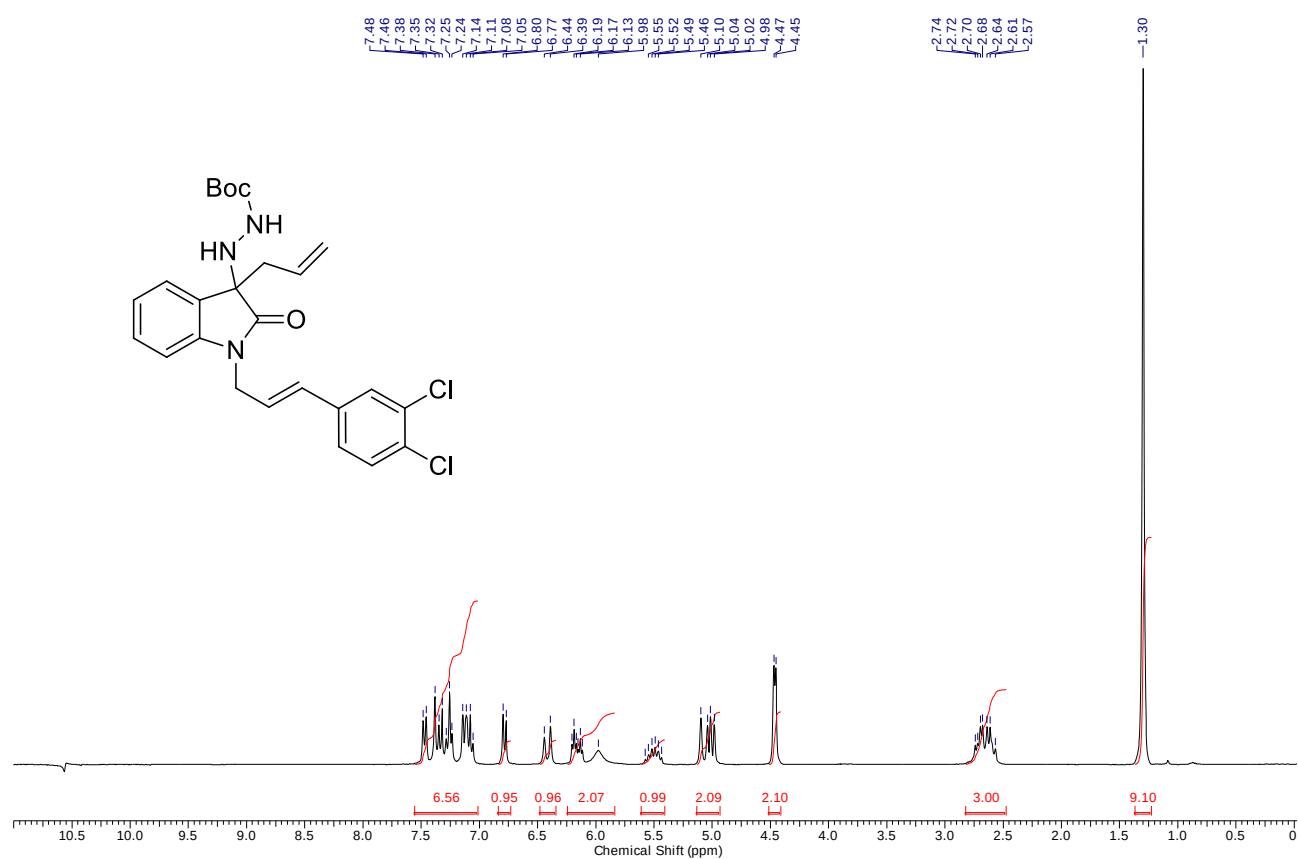
Chemical structure of the compound is shown above the spectrum. The spectrum displays peaks corresponding to the structure, with chemical shifts (ppm) labeled above the peaks and integration values below the peaks.

Chemical Shift (ppm): 7.46, 7.44, 7.38, 7.36, 7.29, 7.07, 7.04, 6.81, 6.78, 5.98, 5.56, 5.53, 5.51, 5.50, 5.47, 5.46, 5.45, 5.42, 5.06, 3.00, 4.96, 3.18, 2.94, 2.71, 2.69, 2.66, 2.63, 2.58, 2.55, 2.54, 2.51, 1.31.

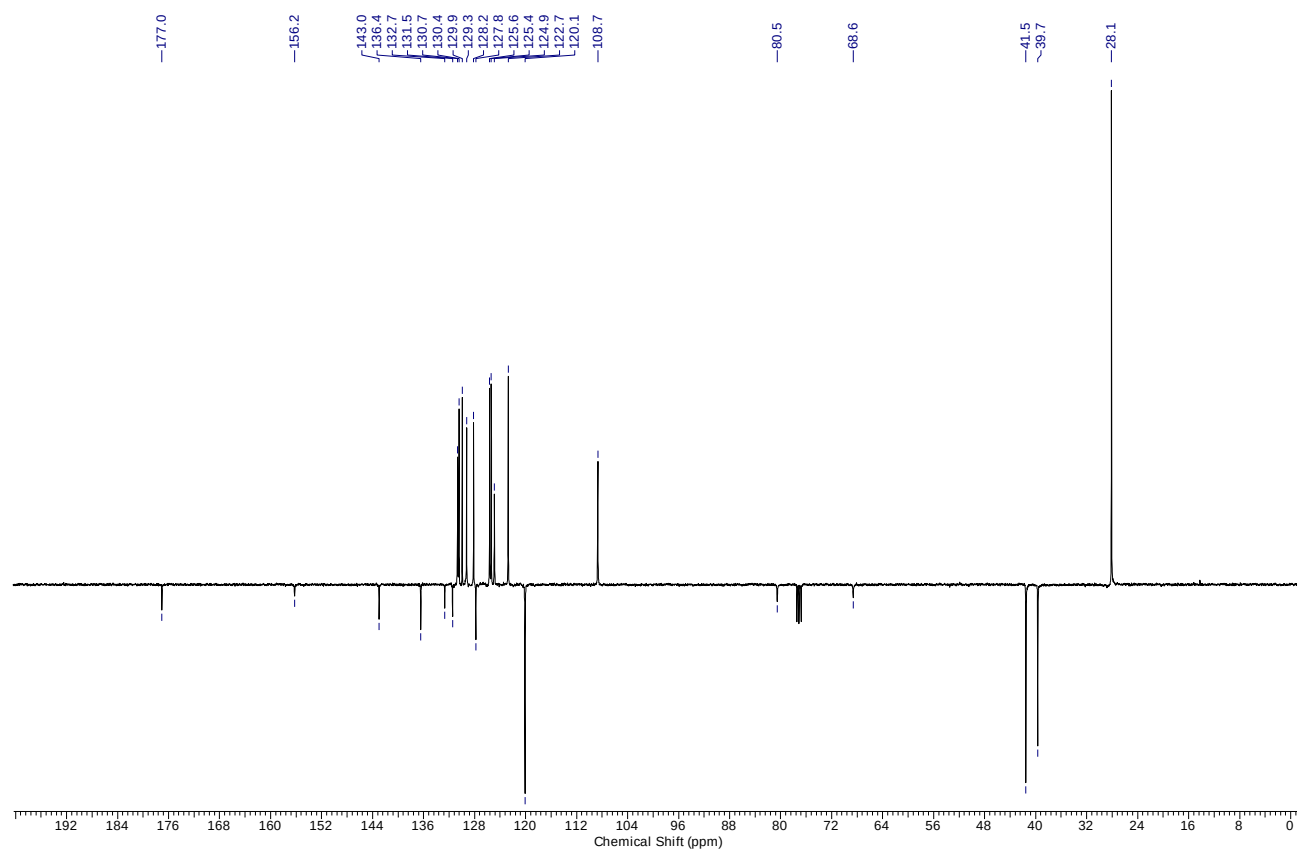
Integration values: 1.13, 0.92, 0.74, 0.88, 1.97, 6.04, 9.07.

13C NMR spectrum of 1,2,3,4,5-pentachlorobenzene. The spectrum shows peaks at 177.3, 156.1, 143.9, 130.6, 129.2, 127.7, 125.2, 122.5, 119.8, 107.9, 80.3, 68.6, 39.5, 28.1, and 26.1 ppm. The x-axis is labeled 'Chemical Shift (ppm)' and ranges from 192 to 0.

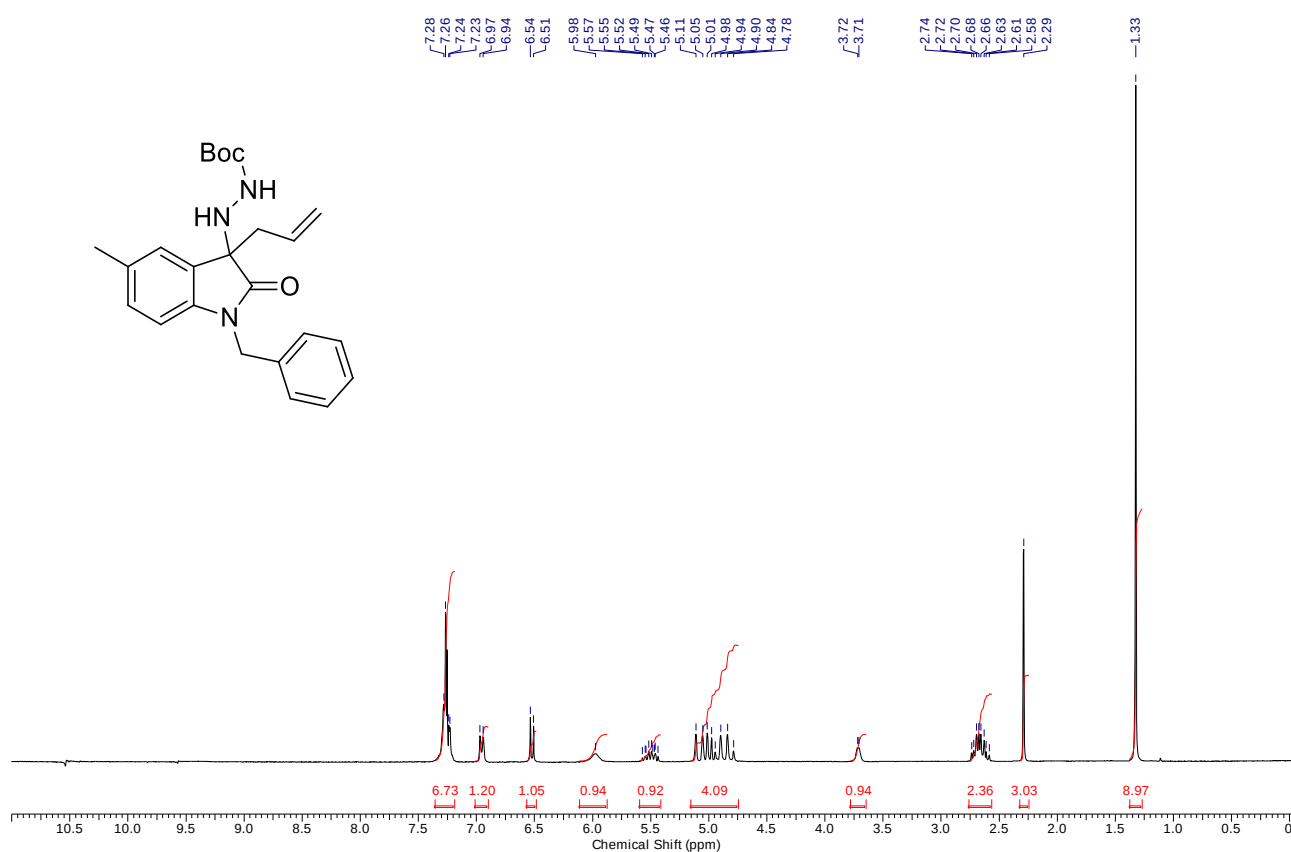
**5c**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



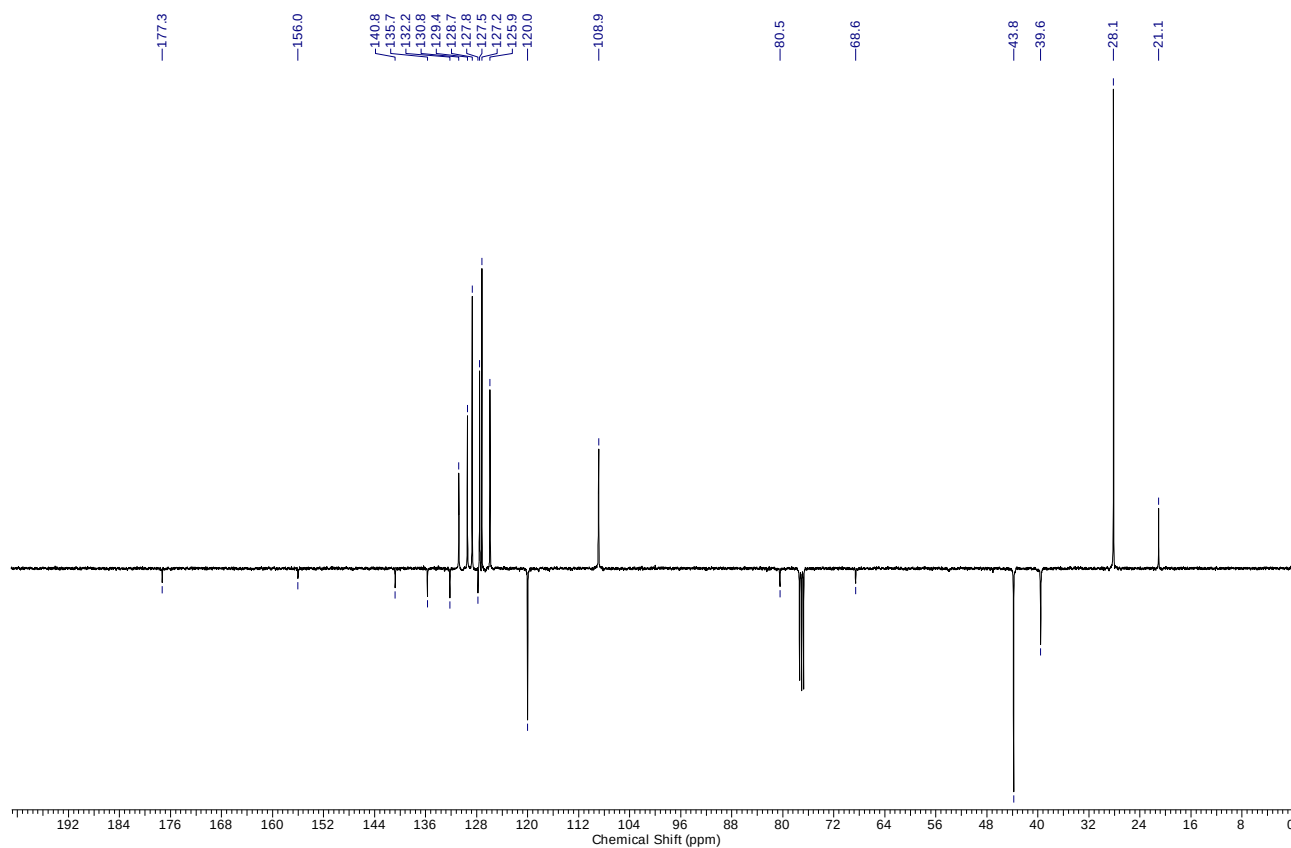
**5c**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



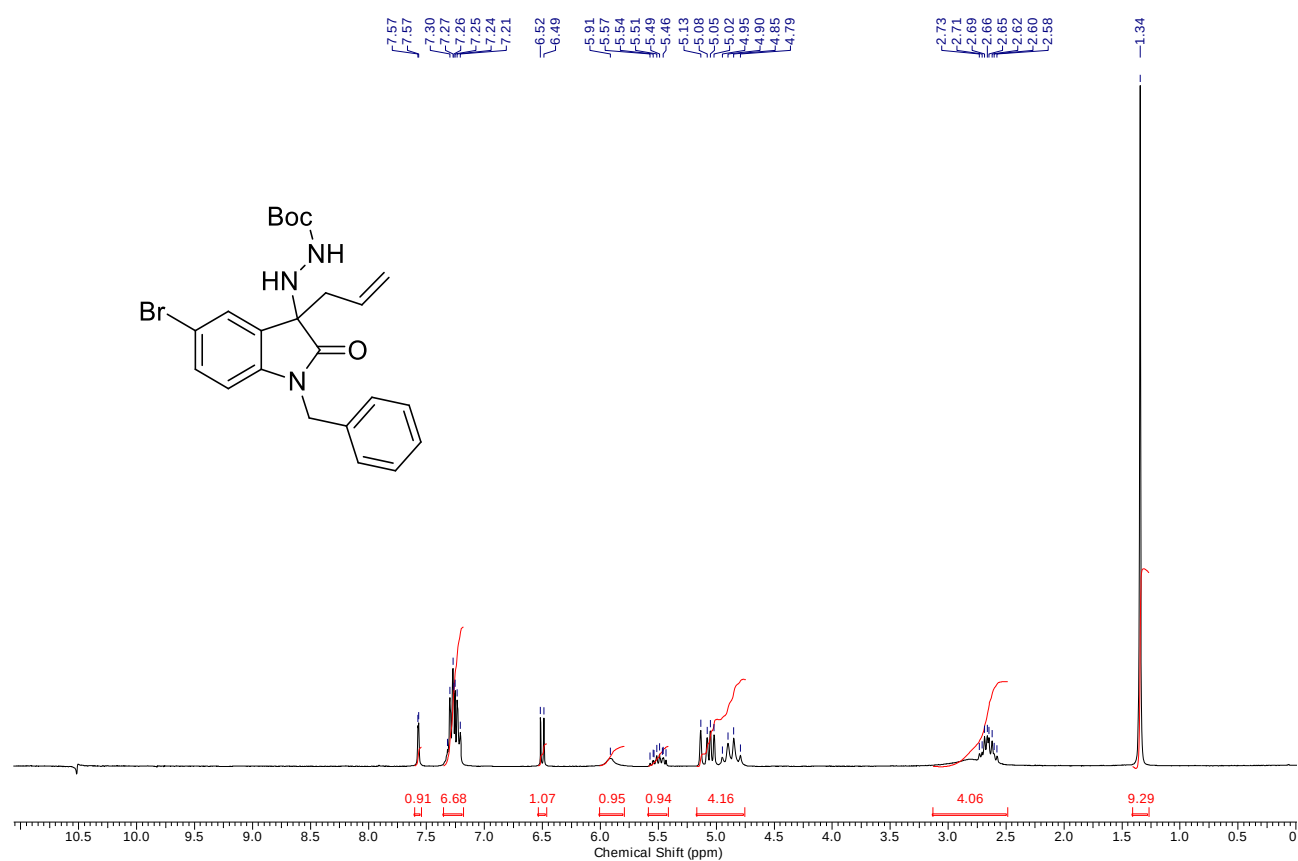
**5d**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



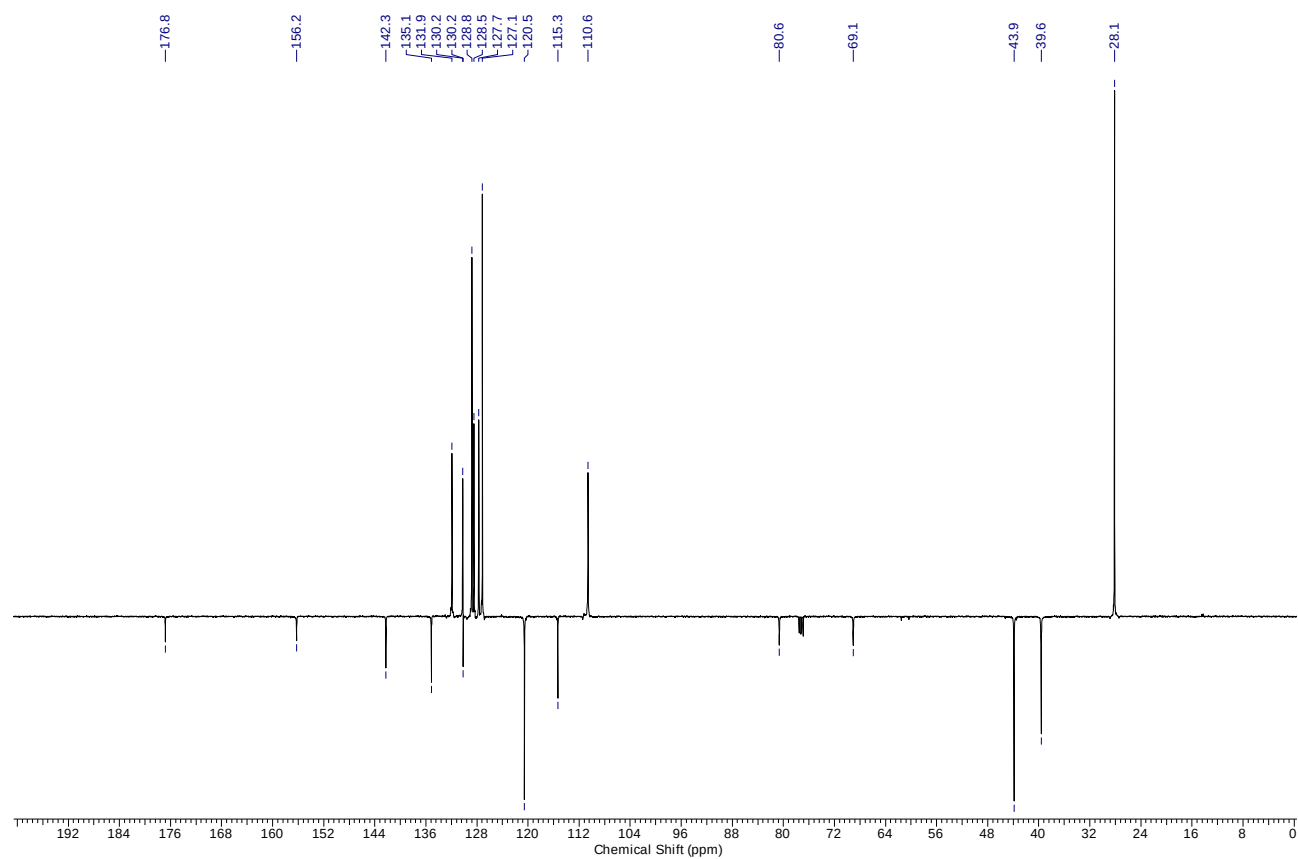
**5d**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



**5e**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):

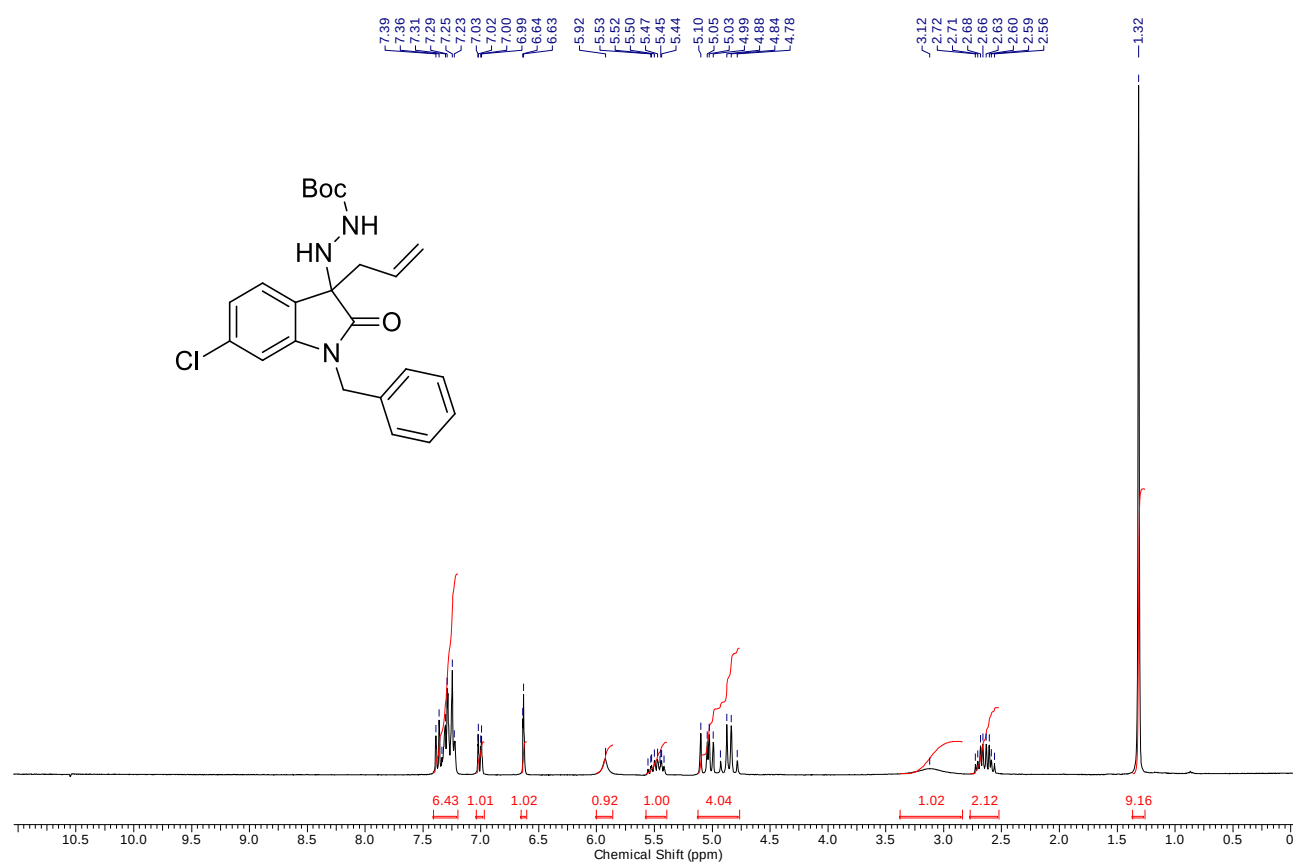


**5e**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):

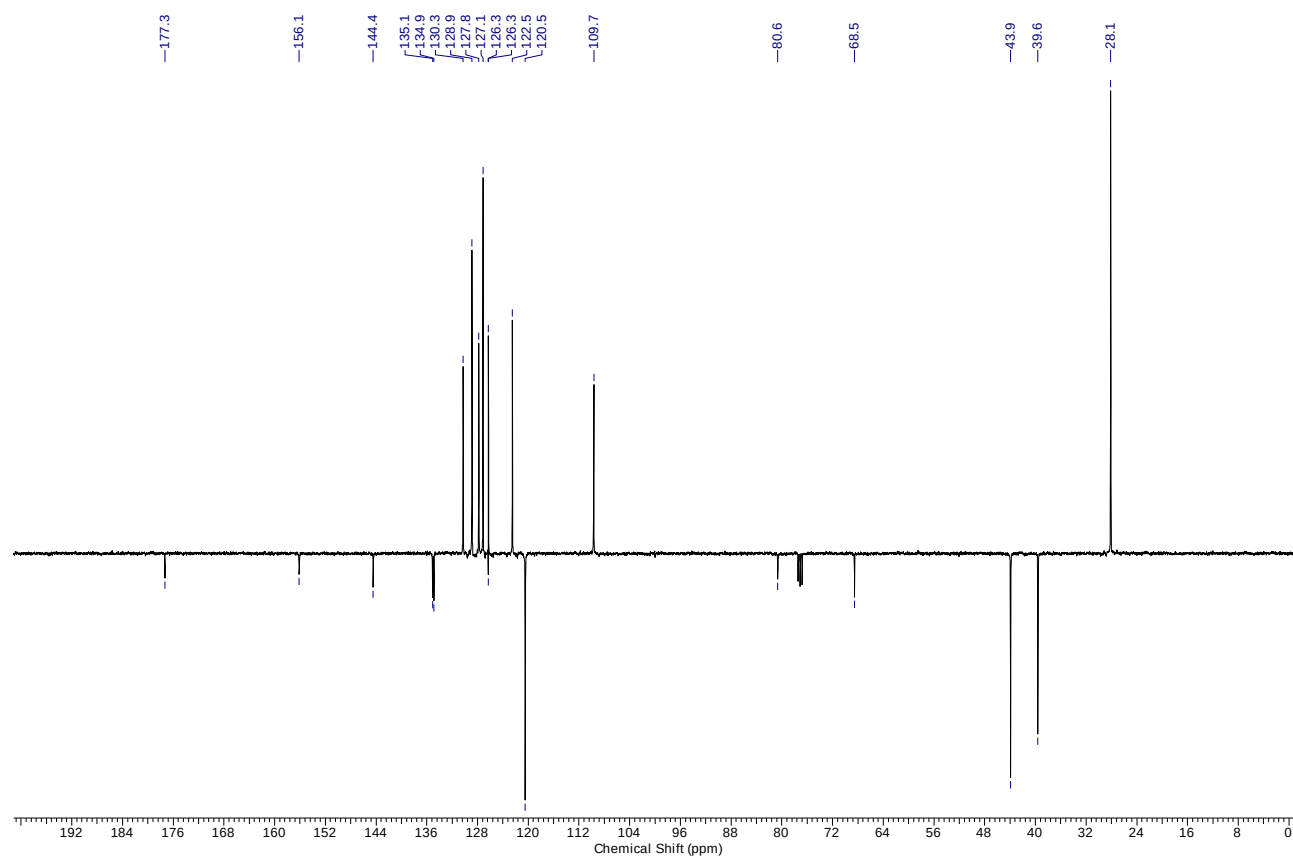




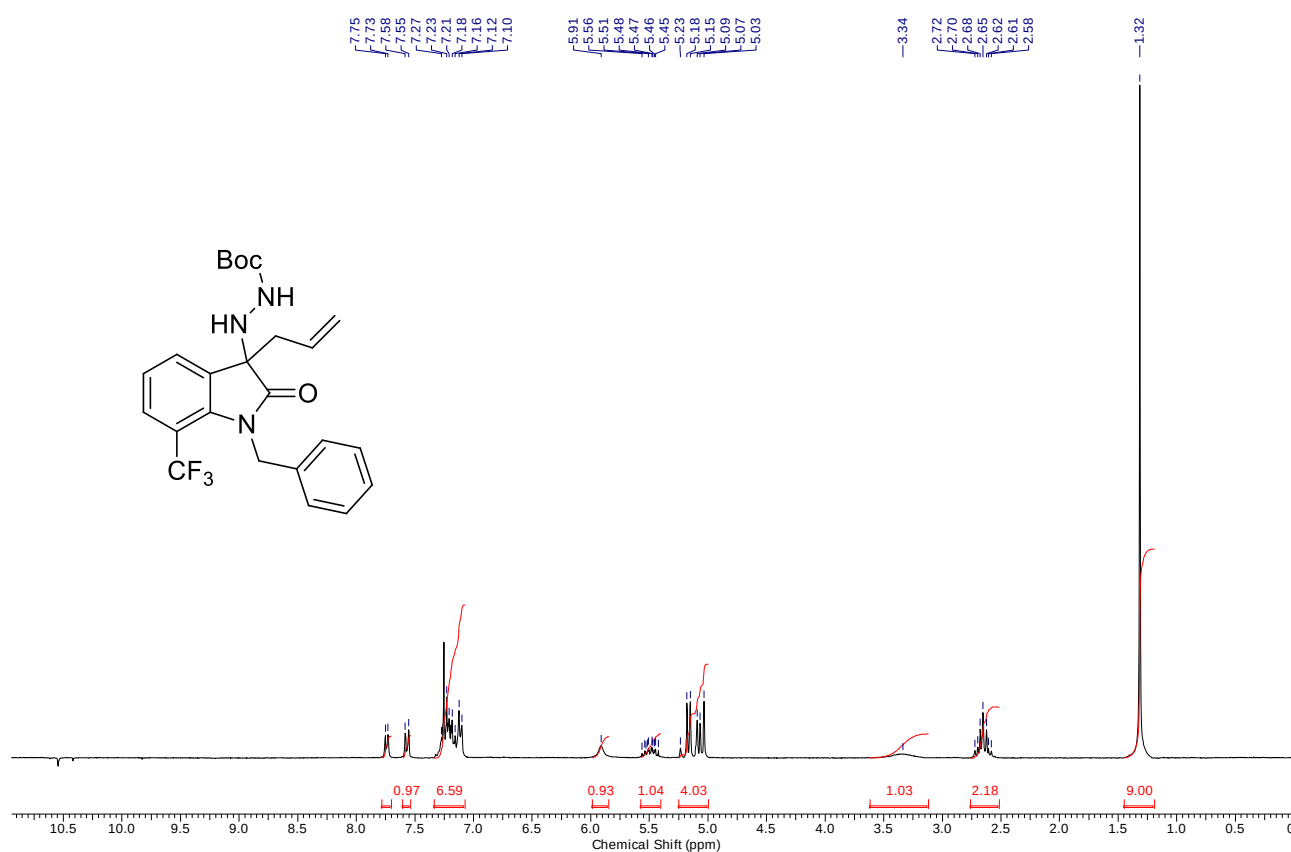
**5f**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



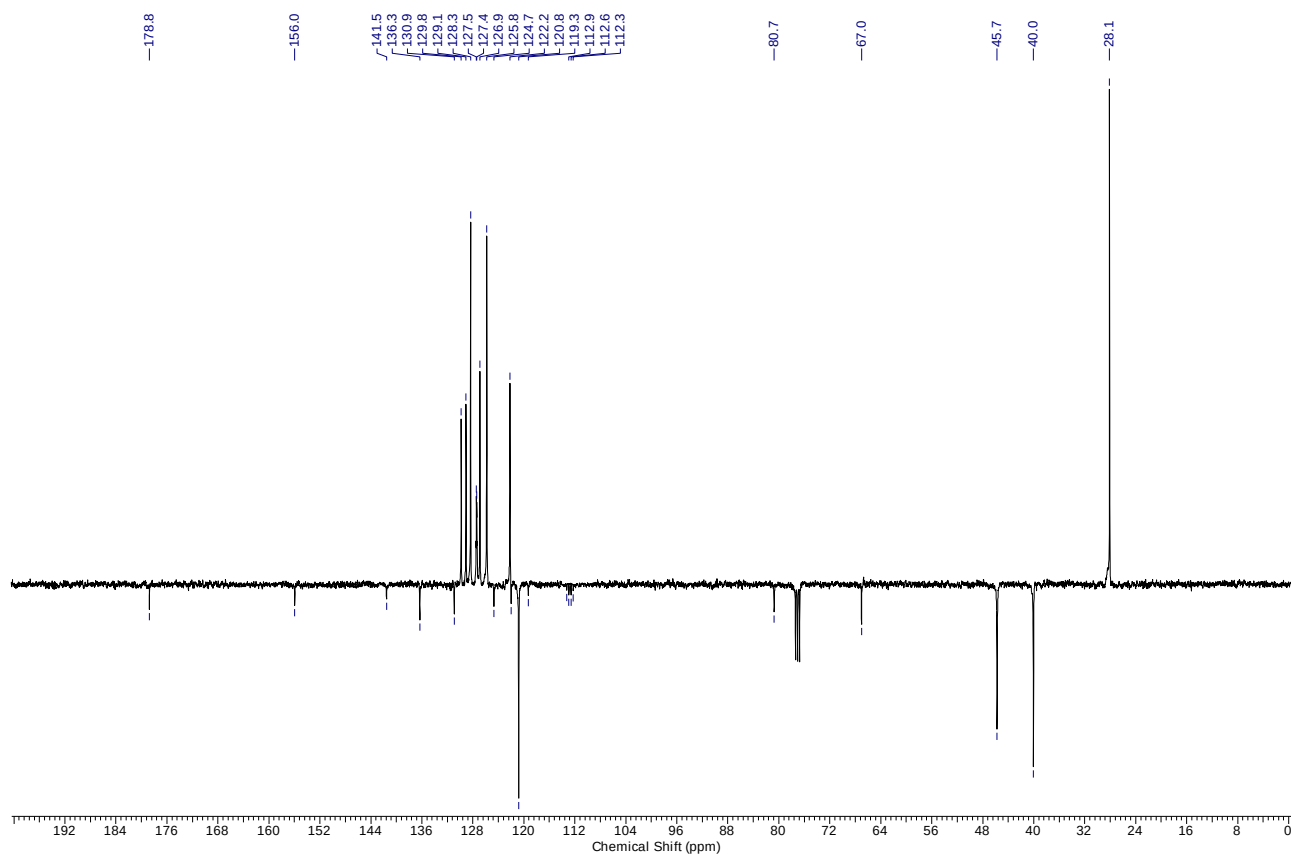
**5f**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):



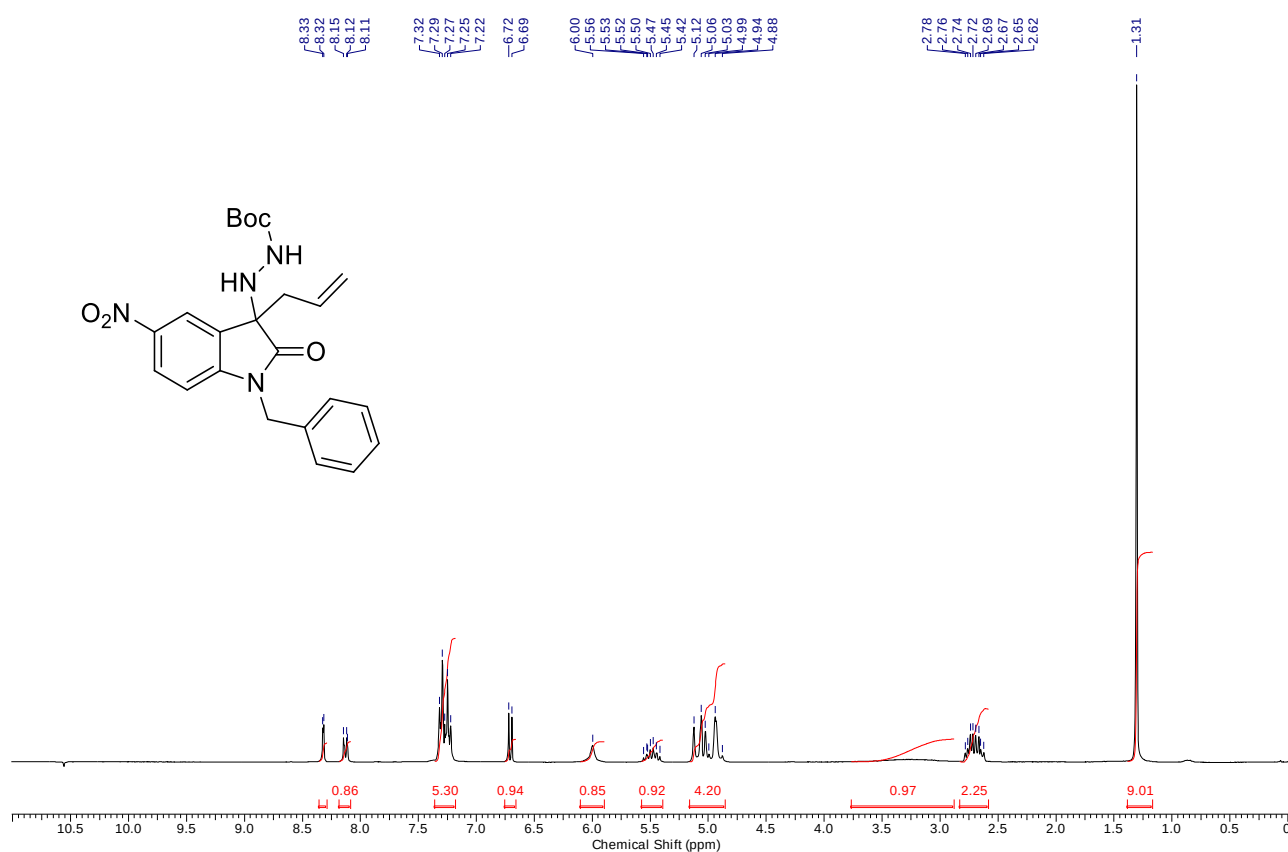
**5g**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



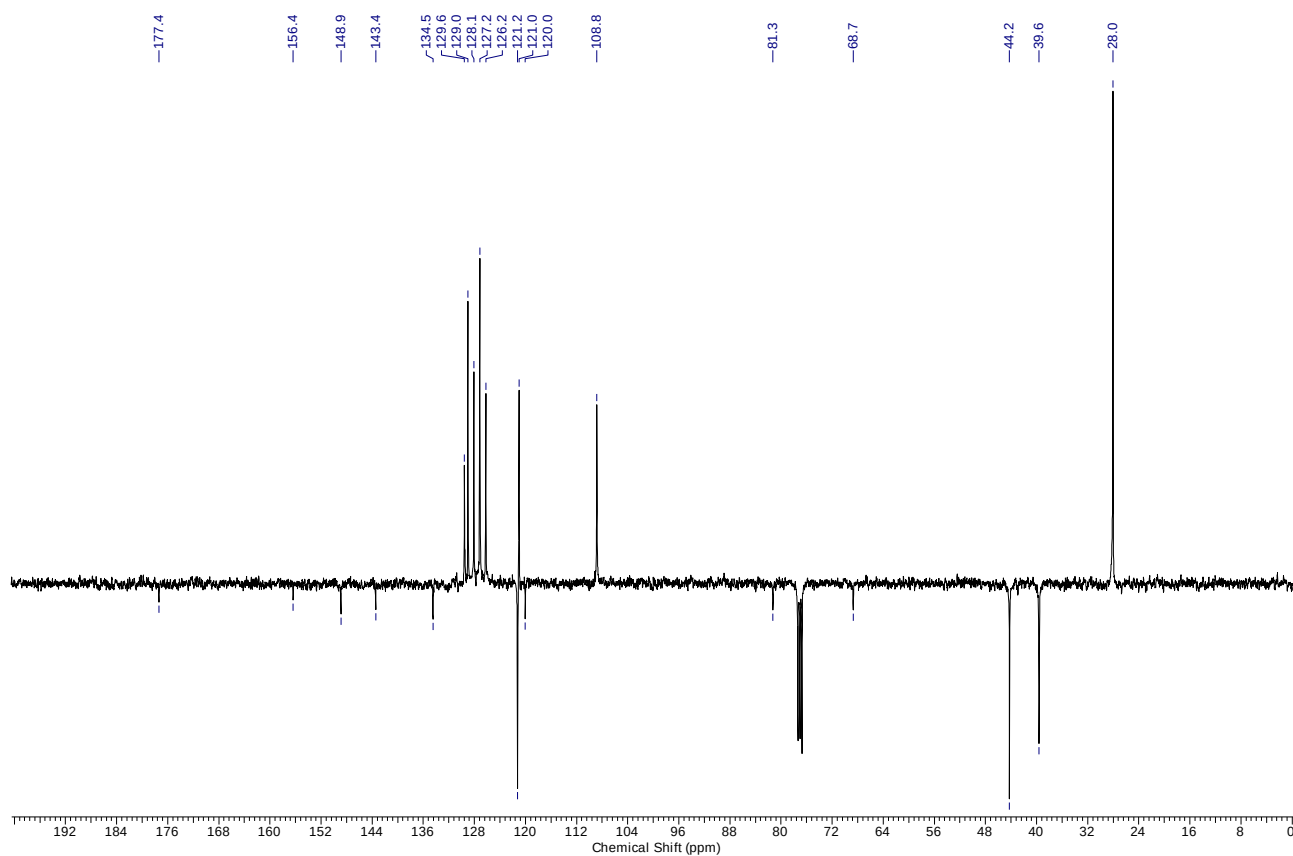
**5g**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



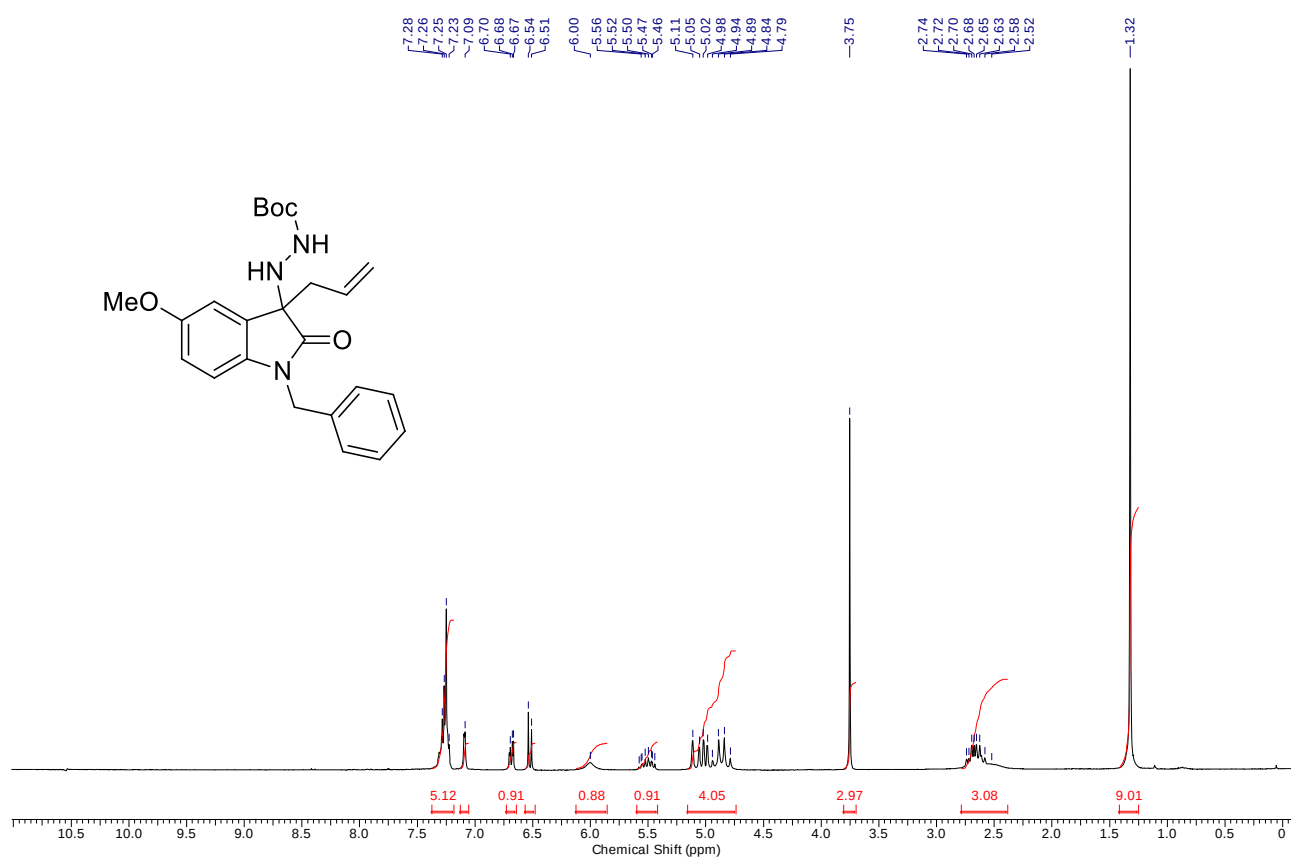
**5h**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



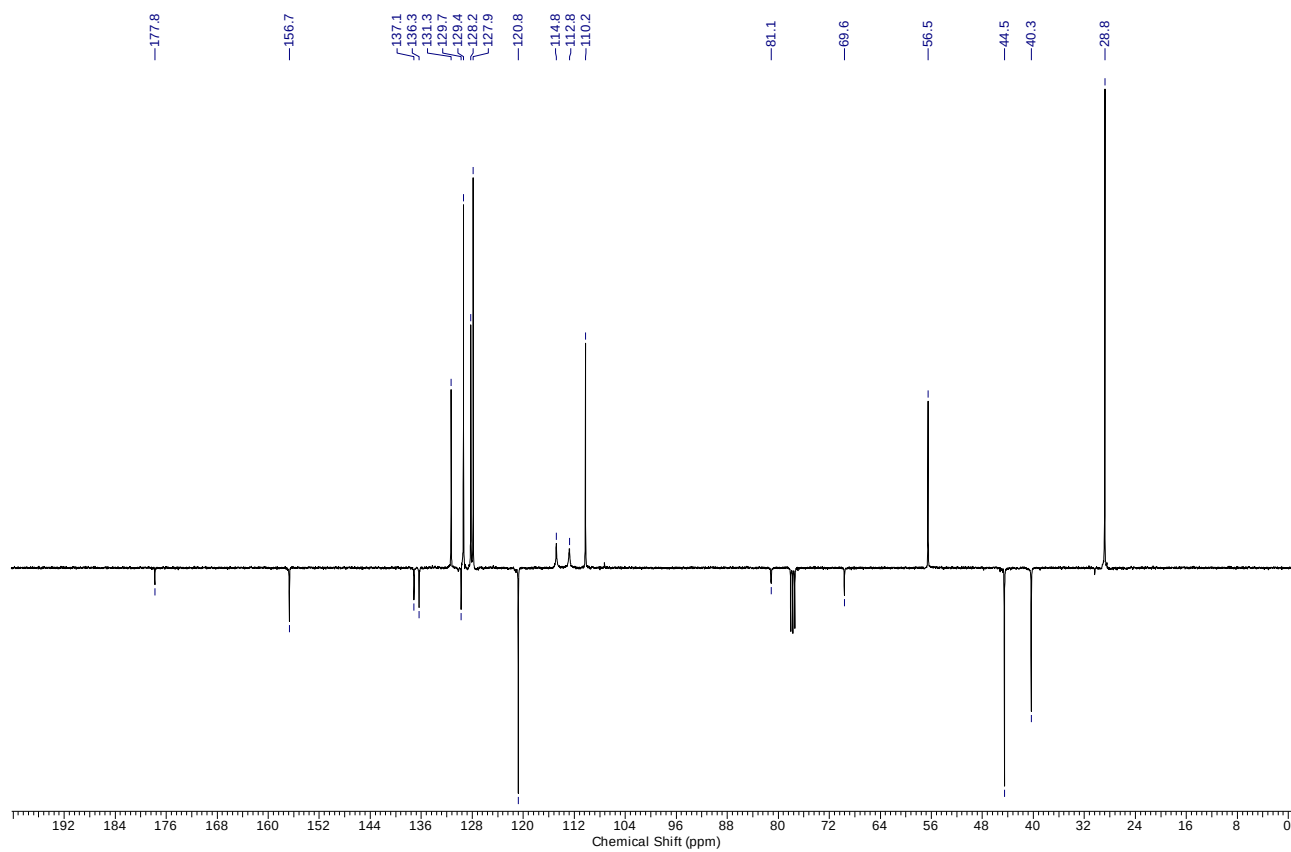
**5h**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



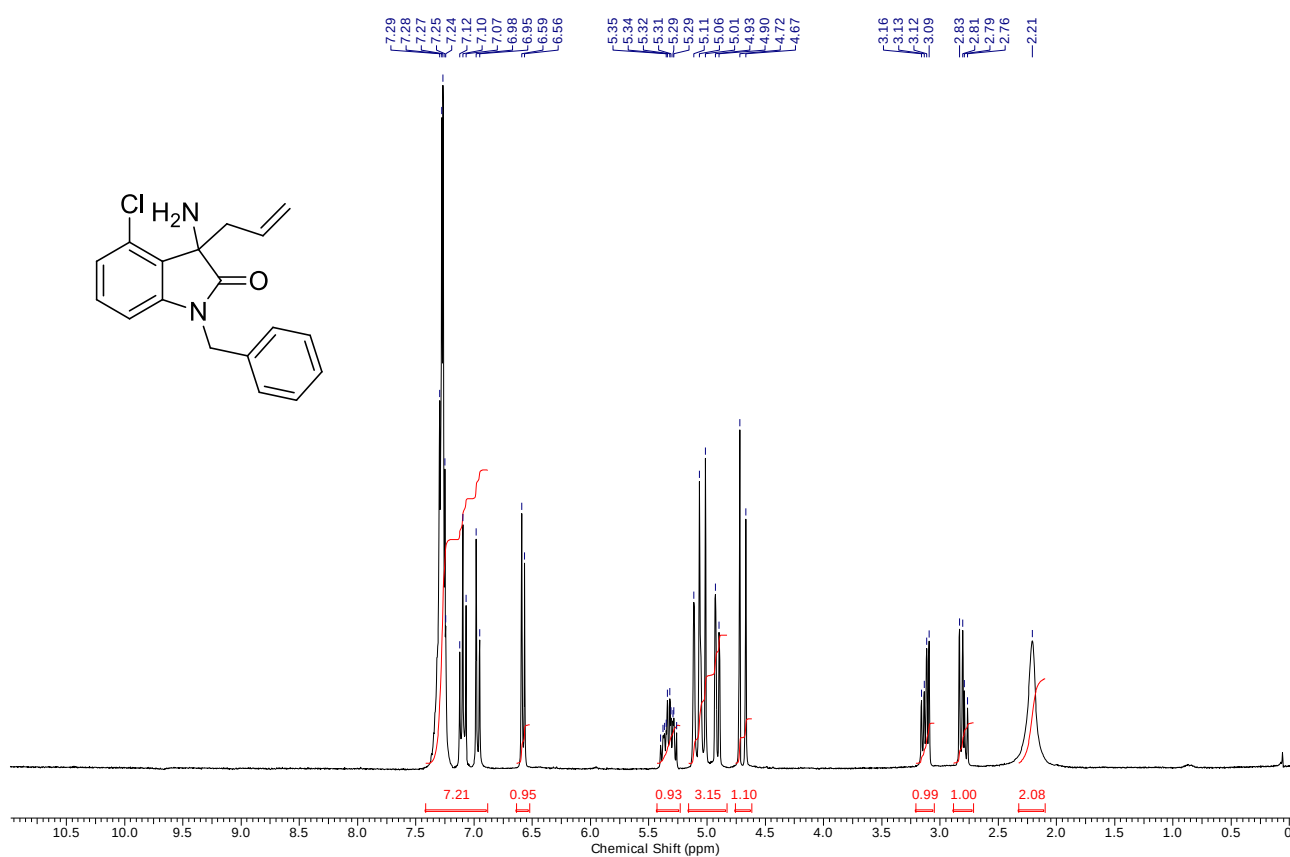
**5i**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



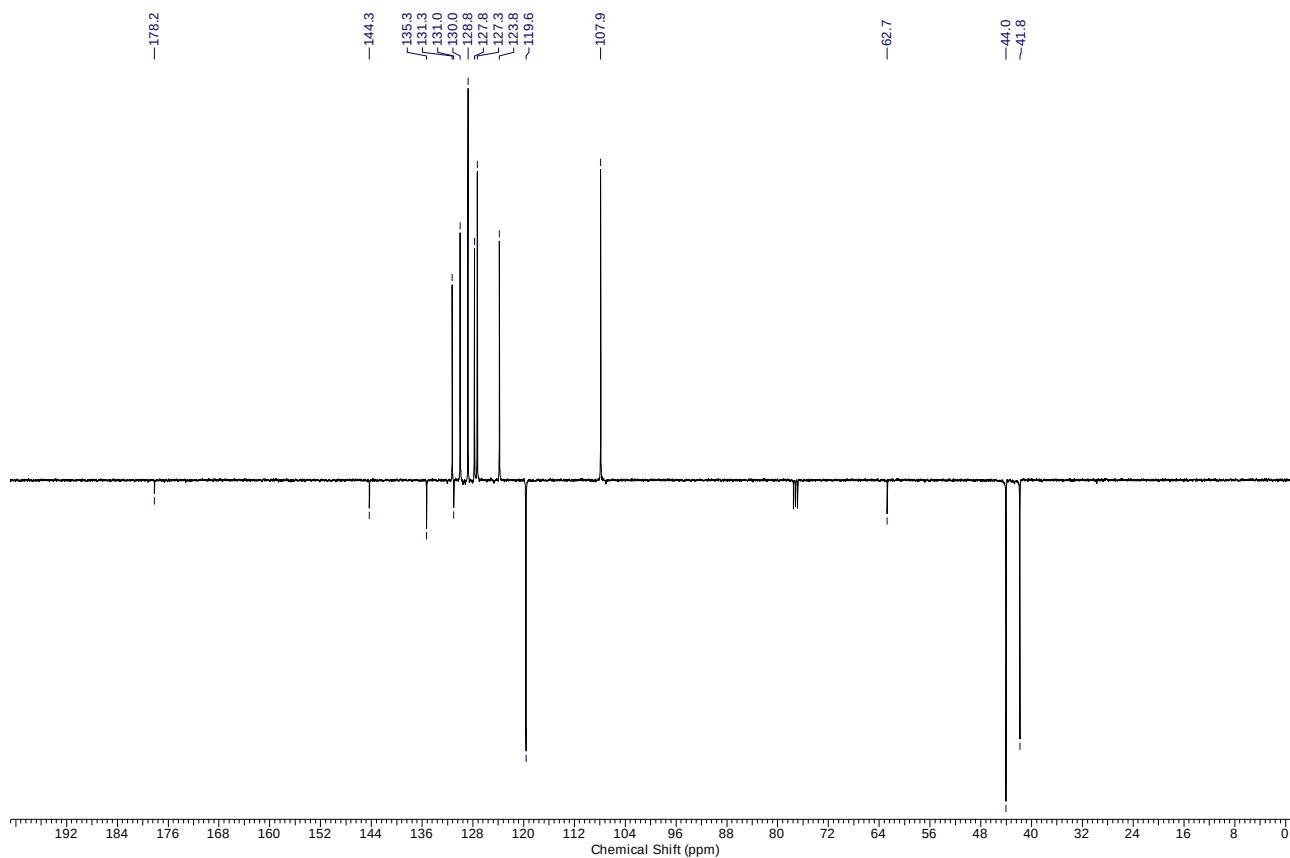
**5i**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):



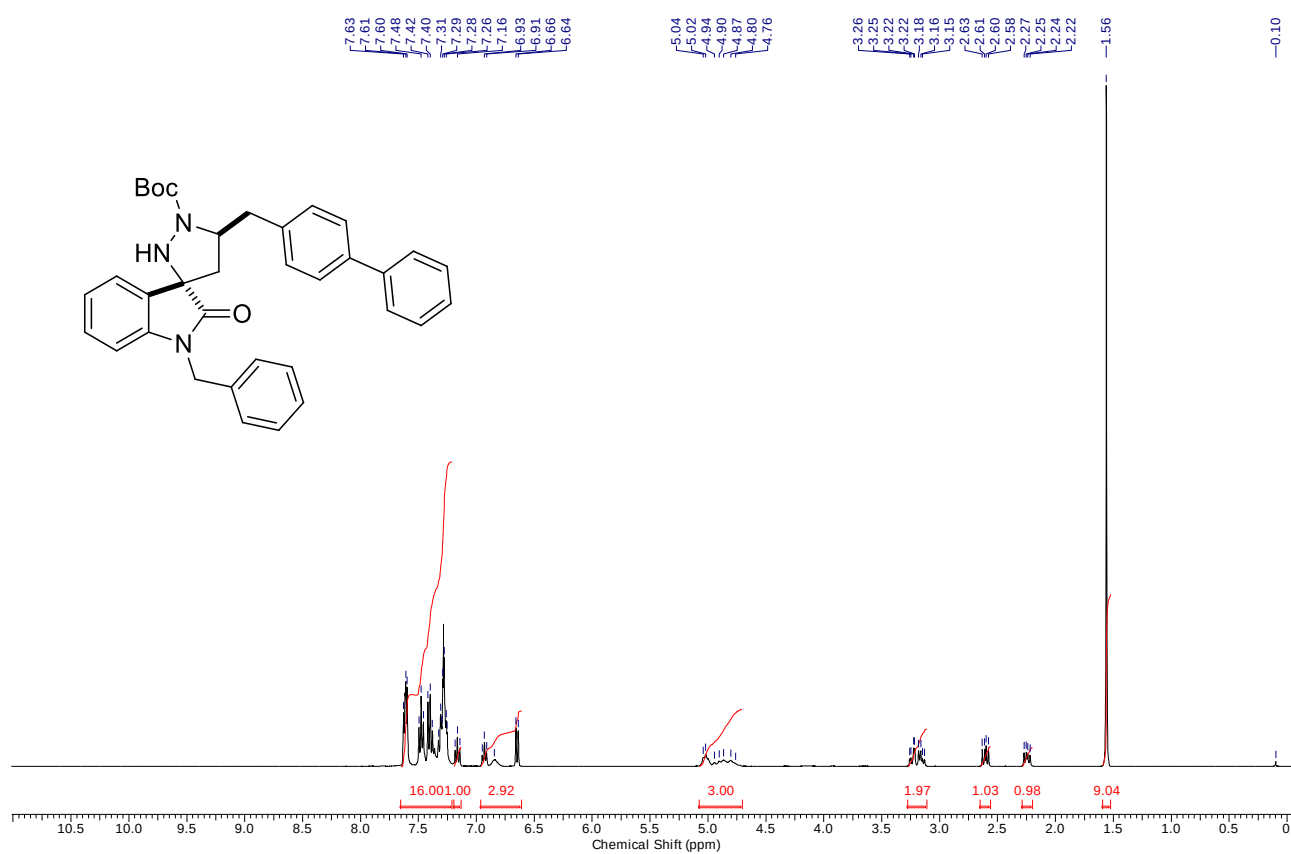
**5j**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



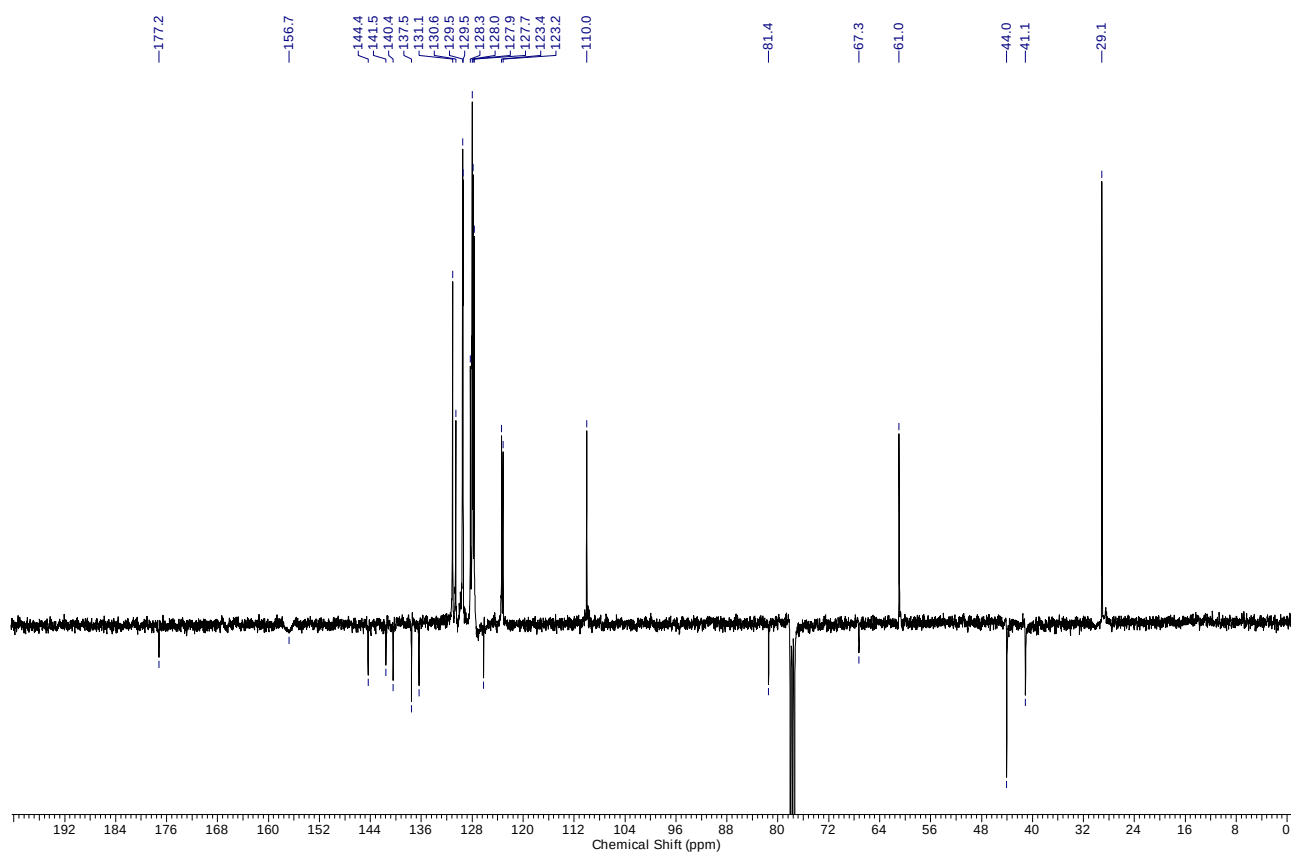
**5j**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):



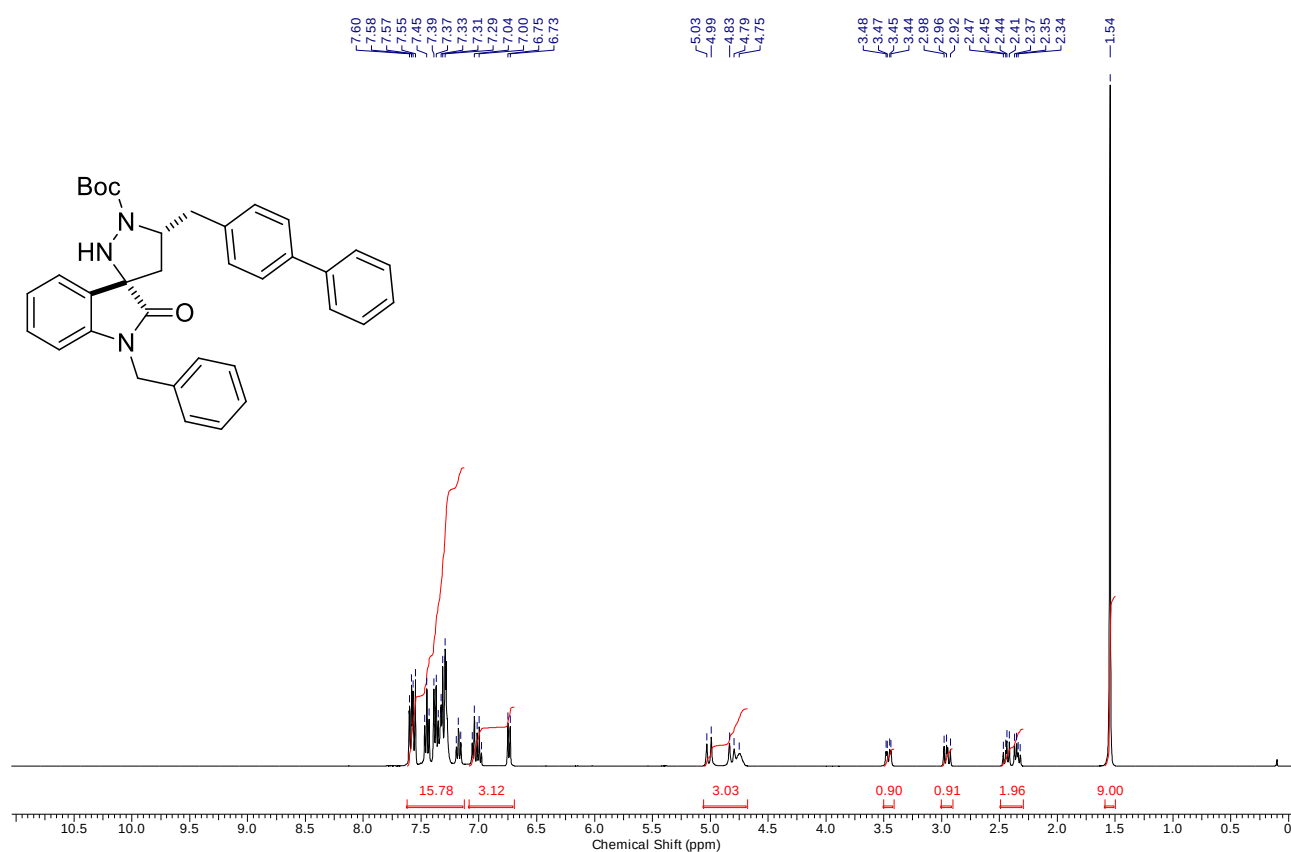
**6a**  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):



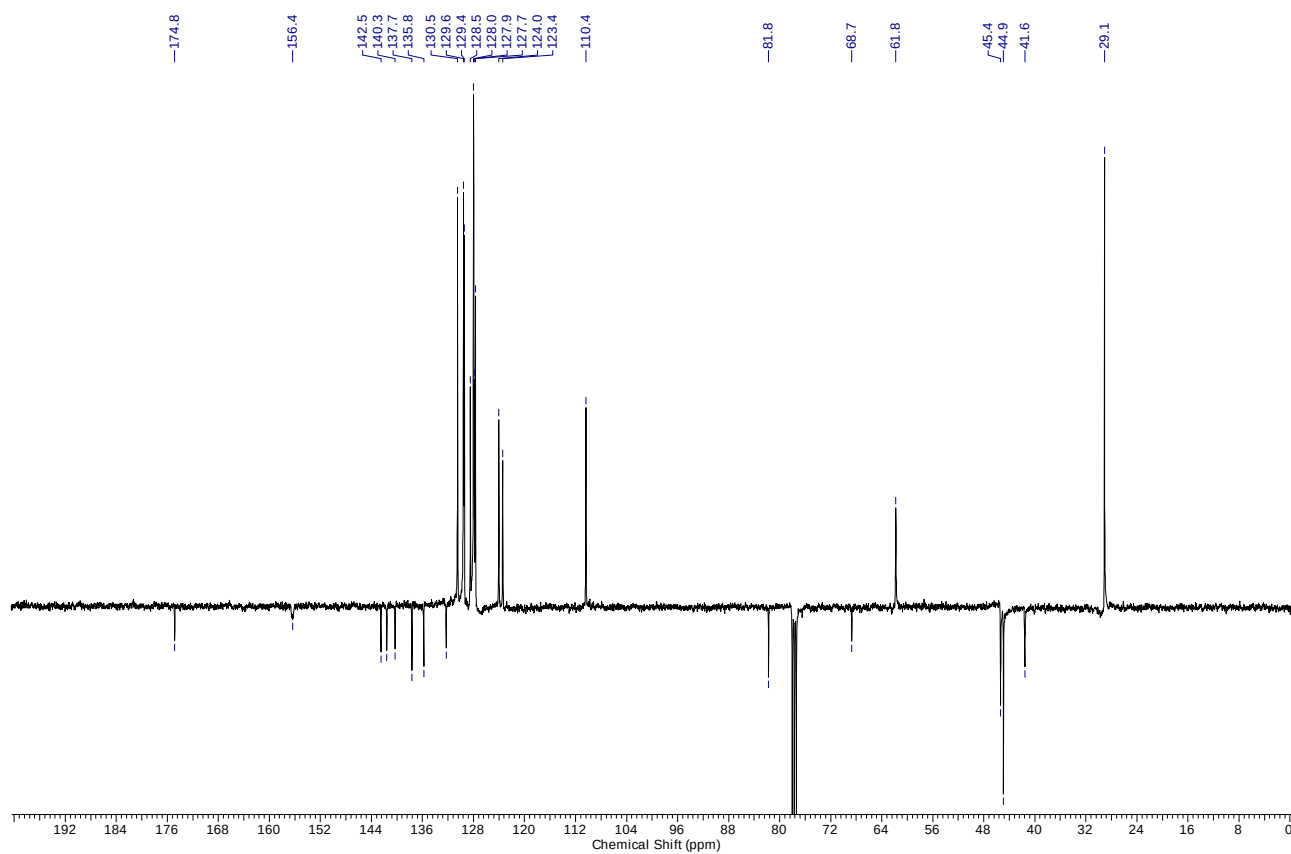
**6a**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):



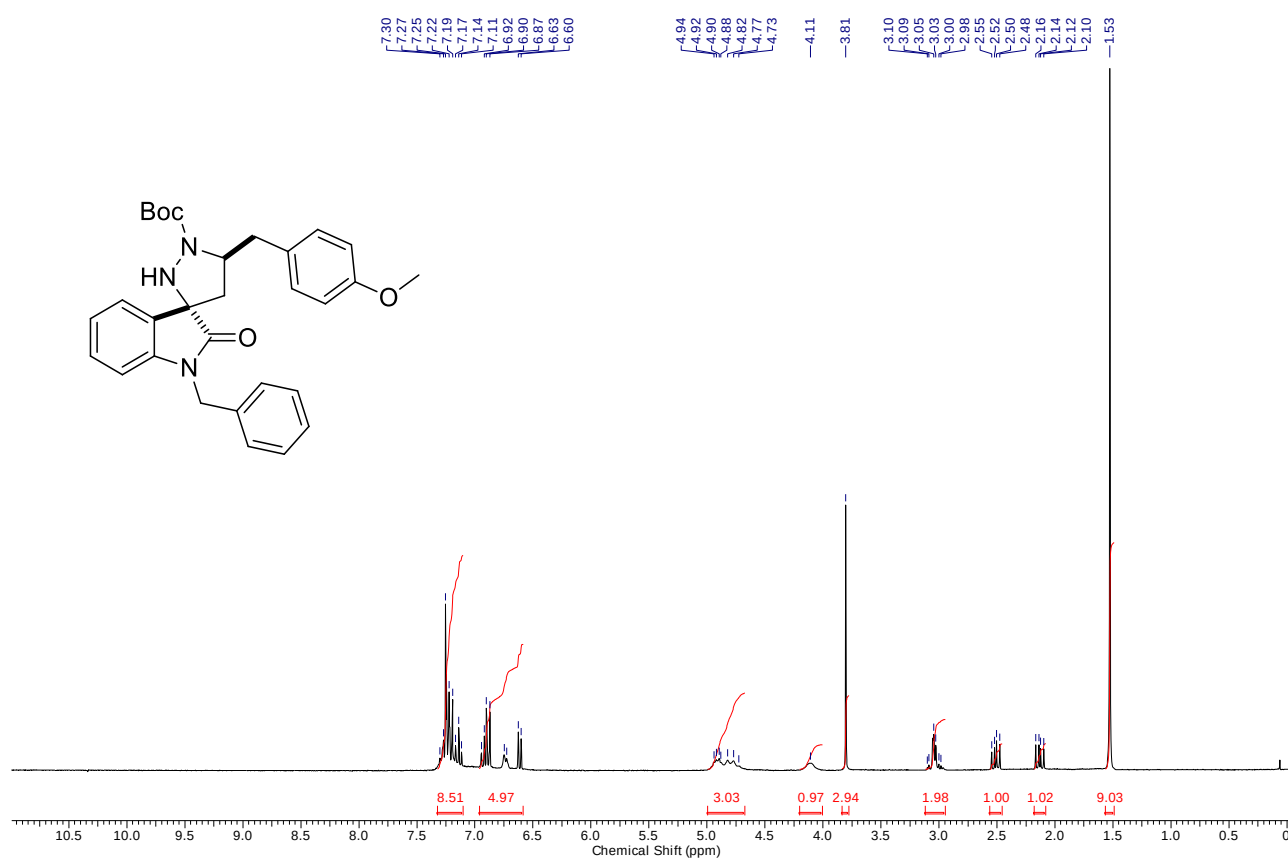
**6a'**  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):



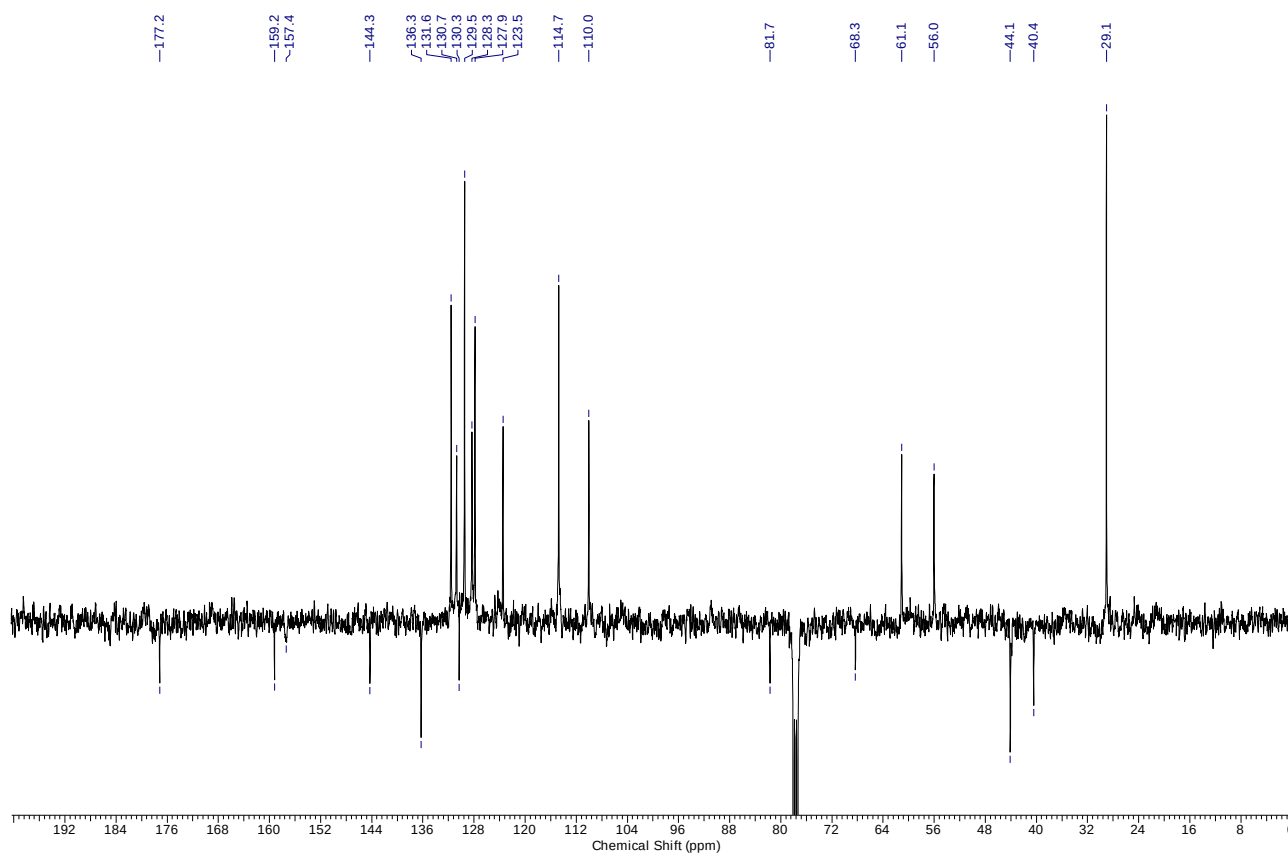
**6a'**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



**6b**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):

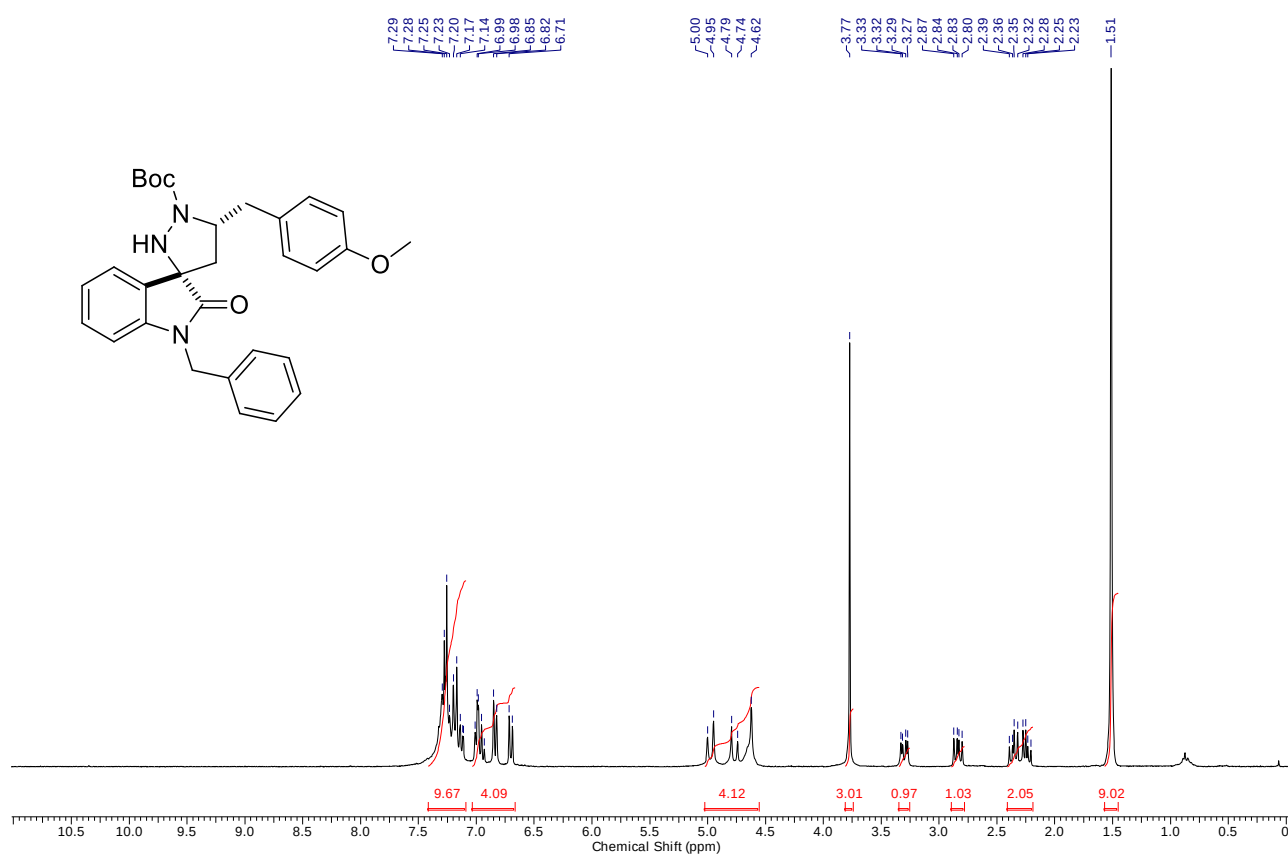


**6b**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):

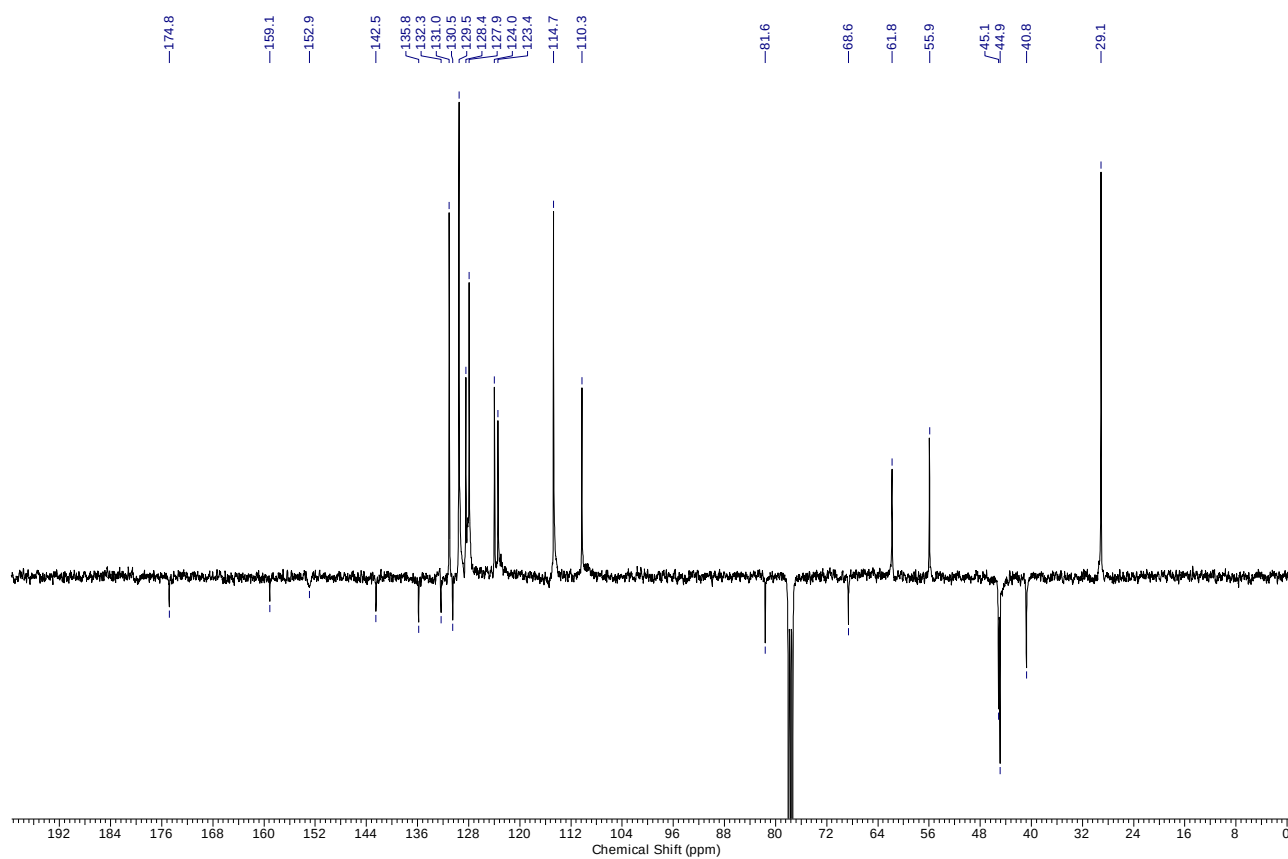




**6b'**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



**6b'**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



Chemical structure of compound 10: Cc1ccccc1N2C(=O)[C@H](Cc3ccc(cc3)N(C(=O)O)C(=O)O)C4=CC=CC=C4N2C(=O)O

<sup>1</sup>H NMR spectrum (CDCl<sub>3</sub>) of compound 10. The x-axis represents the chemical shift in ppm, ranging from 0 to 10.5. The spectrum shows several multiplets and singlets, with integration values indicated below the baseline. A list of chemical shifts (δ) is provided at the top of the spectrum.

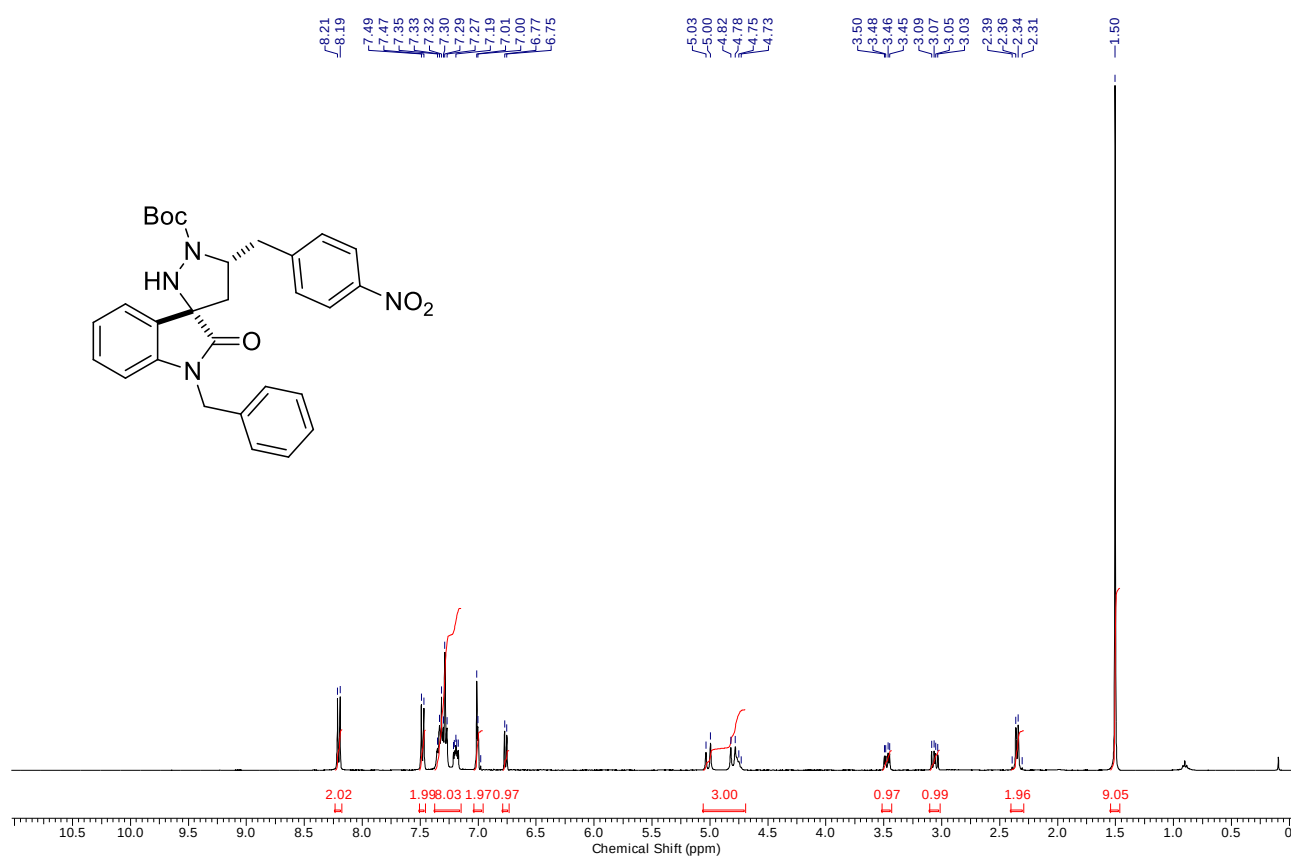
Chemical Shifts (ppm): 8.20, 8.17, 7.48, 7.45, 7.27, 7.25, 7.23, 7.20, 7.16, 7.14, 7.01, 7.00, 6.98, 6.67, 6.64, 4.98, 4.96, 4.93, 4.91, 4.89, 4.87, 4.81, 4.78, 4.73, 3.38, 3.36, 3.34, 3.32, 3.12, 3.10, 3.08, 3.05, 2.57, 2.55, 2.53, 2.51, 2.12, 2.09, 2.08, 2.05, 1.47.

Integration values (from left to right): 1.98, 2.08, 7.14, 2.06, 1.04, 2.99, 1.02, 1.05, 1.08, 1.06, 8.98.

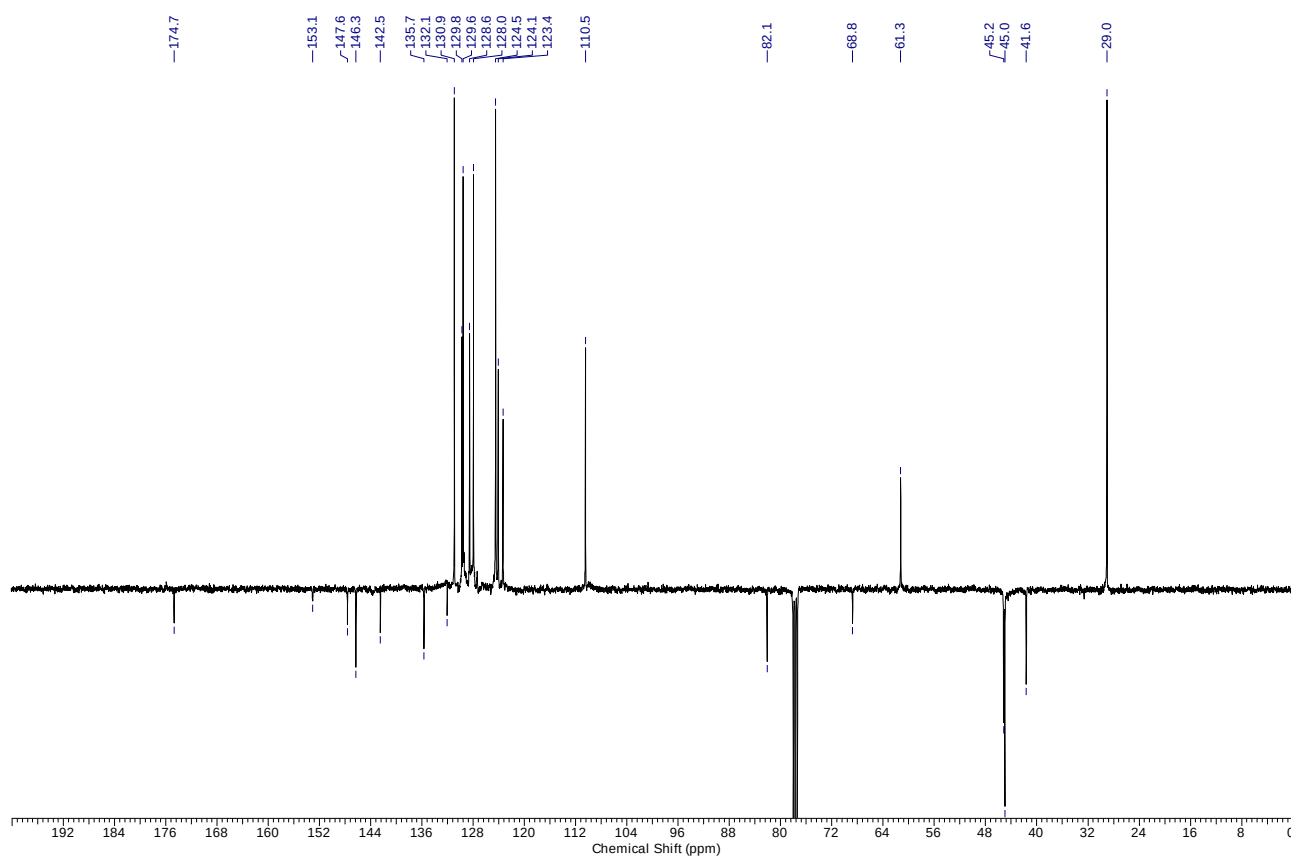
Chemical Shift (ppm)

176.3, 156.9, 146.9, 145.9, 143.8, 135.6, 130.5, 130.2, 130.0, 128.8, 127.7, 127.2, 123.7, 122.8, 122.4, 109.6, 81.1, 66.4, 60.2, 43.4, 42.3, 41.2, 28.3

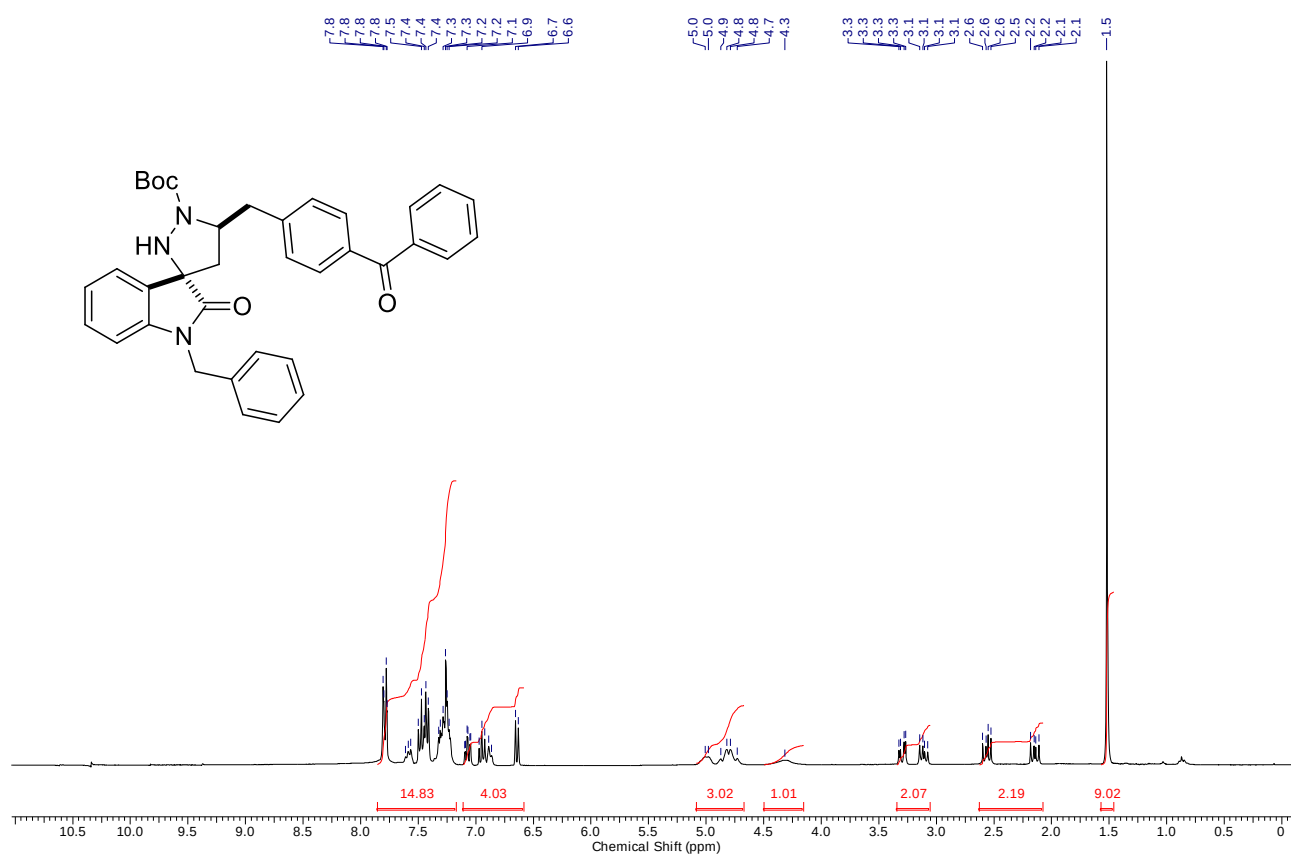
**6c'**  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):



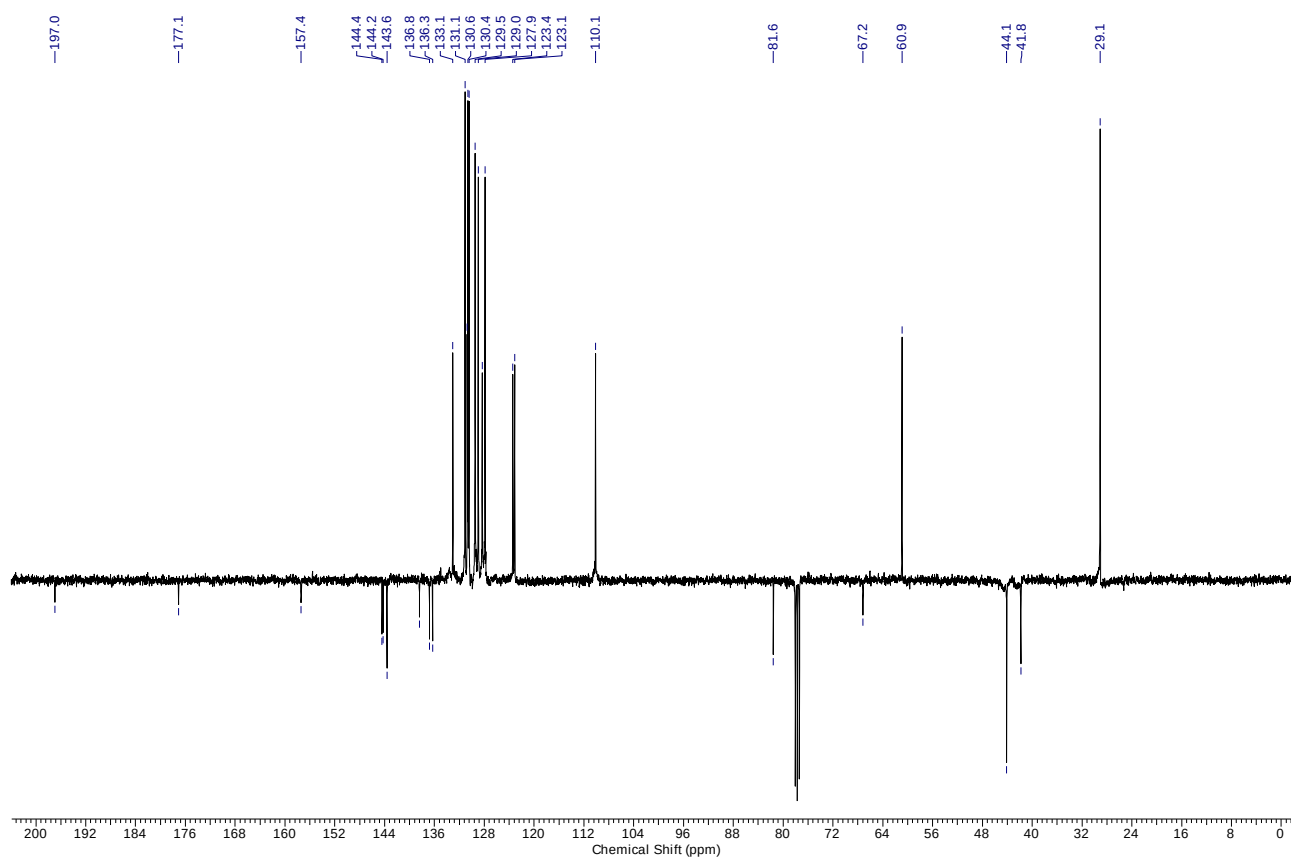
**6c'**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):



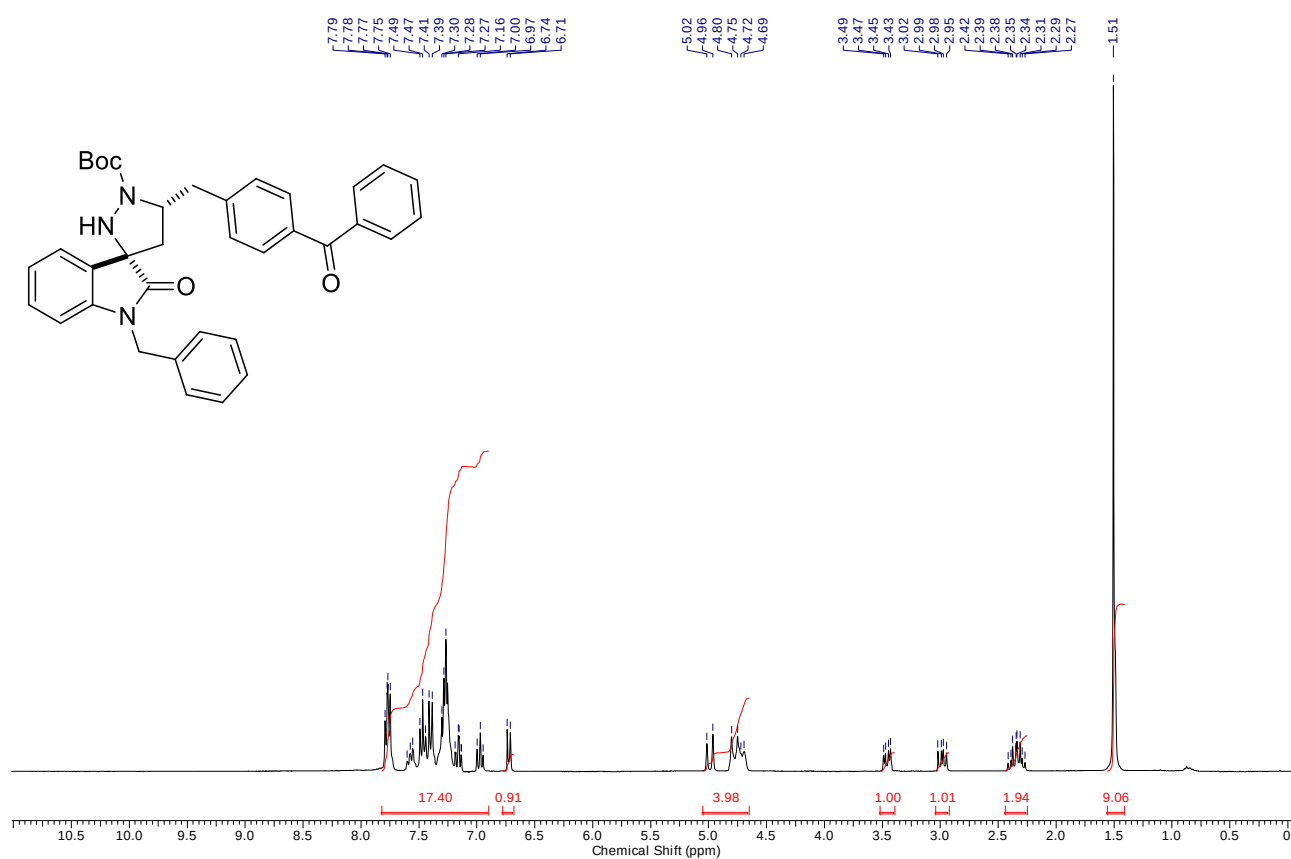
**6d**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



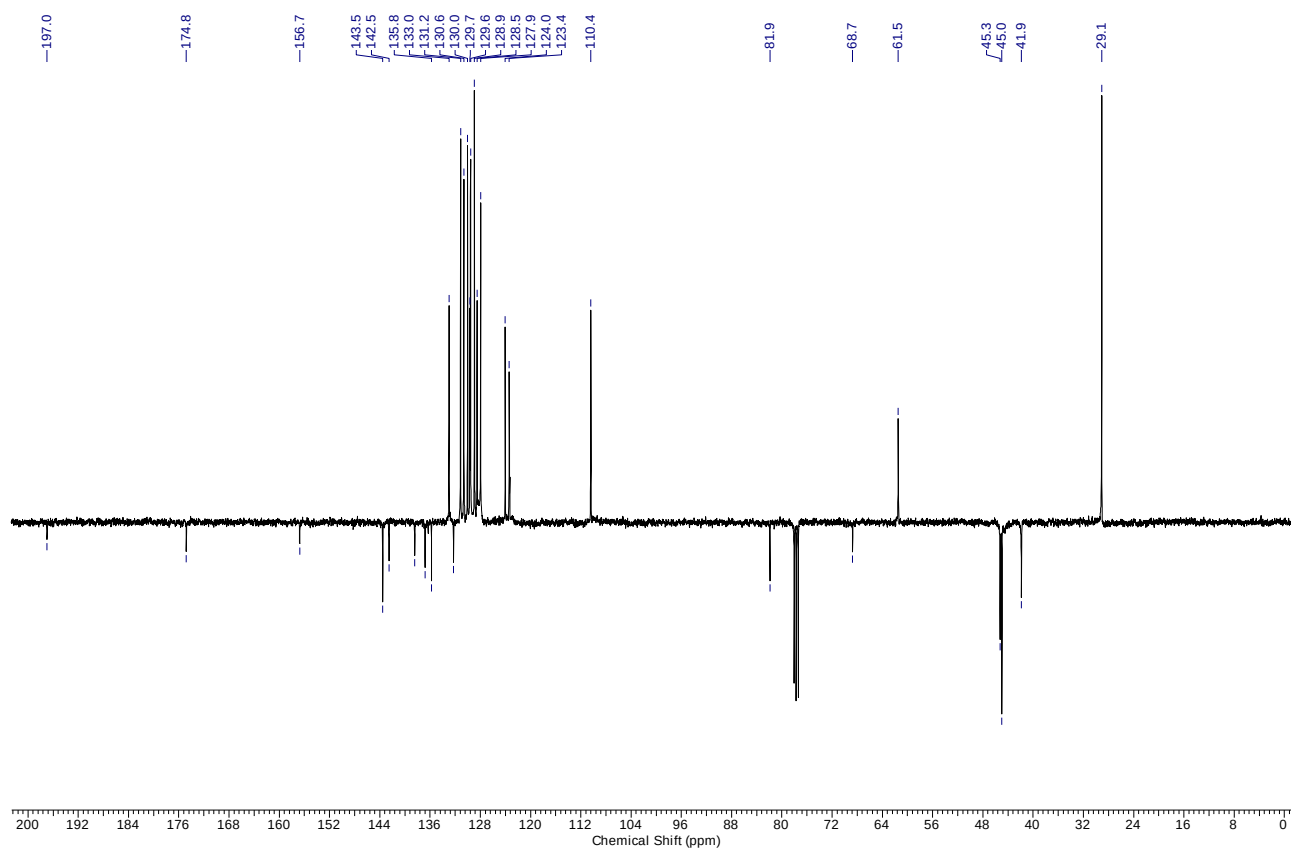
**6d**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



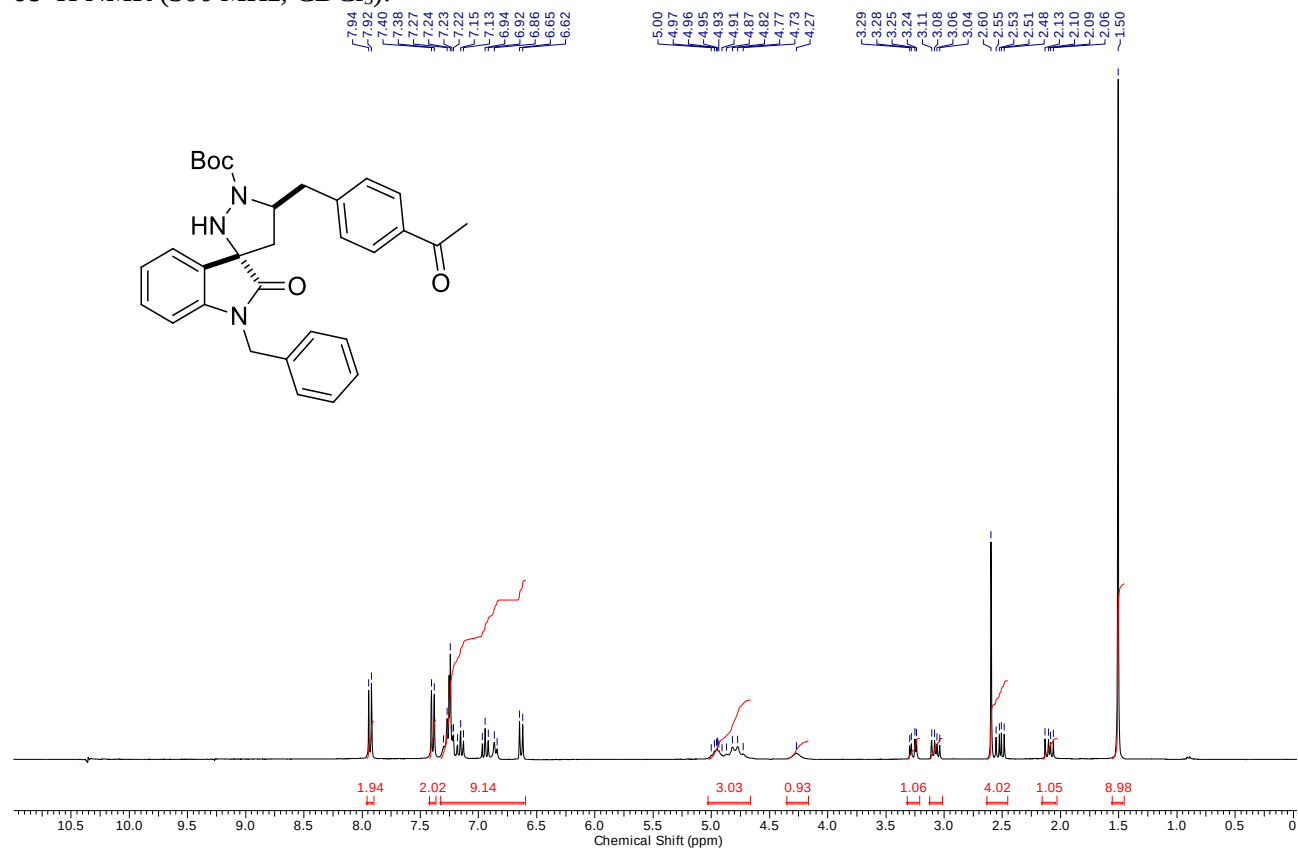
**6d'**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



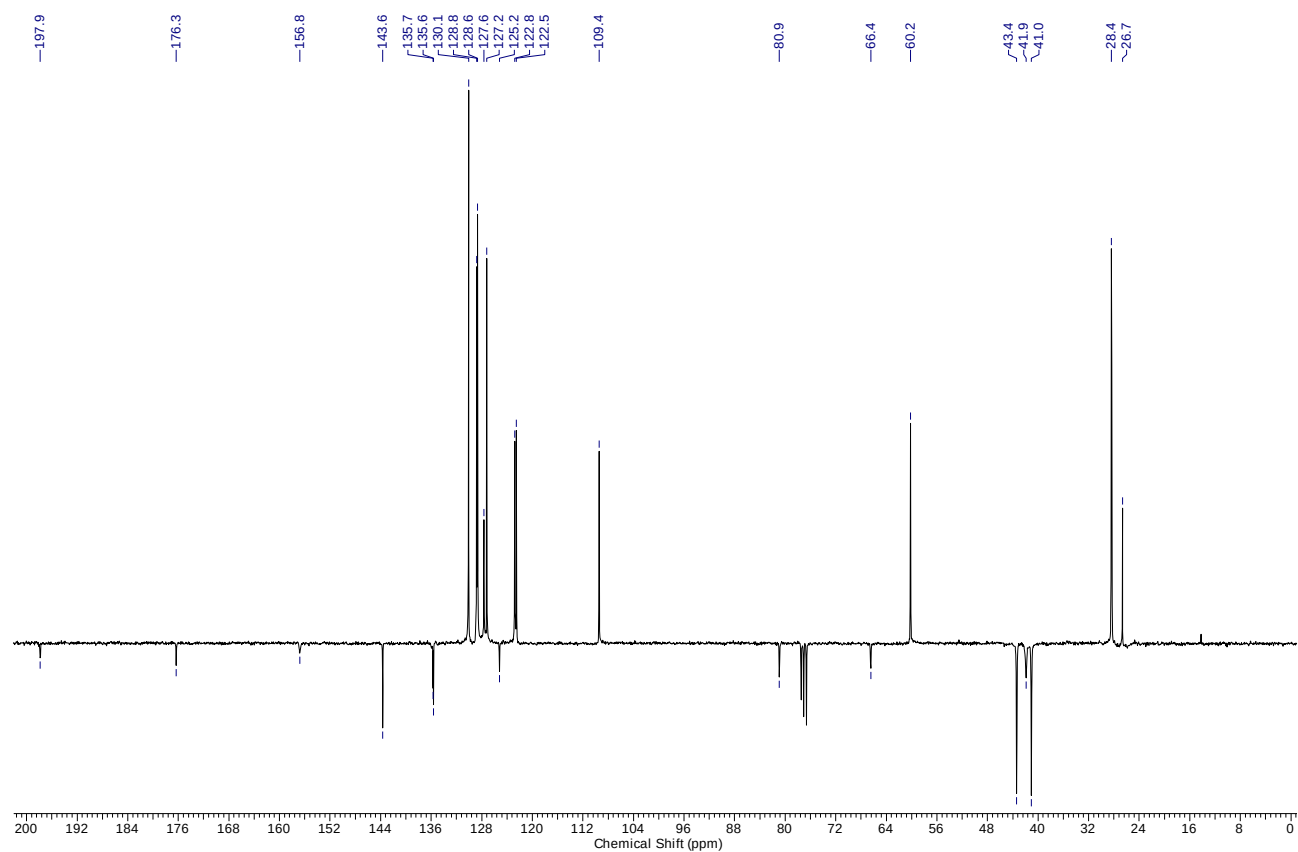
**6d'**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



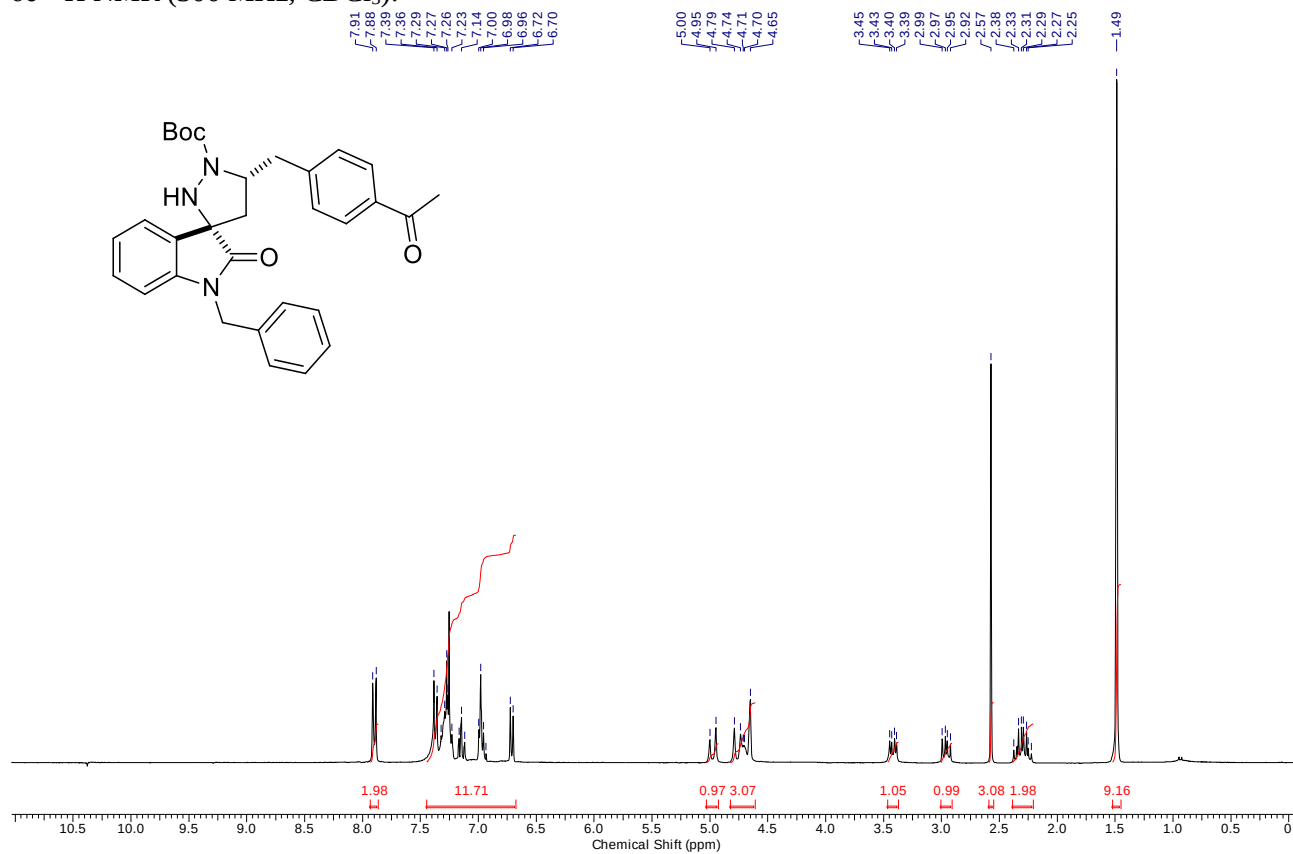
**6e**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



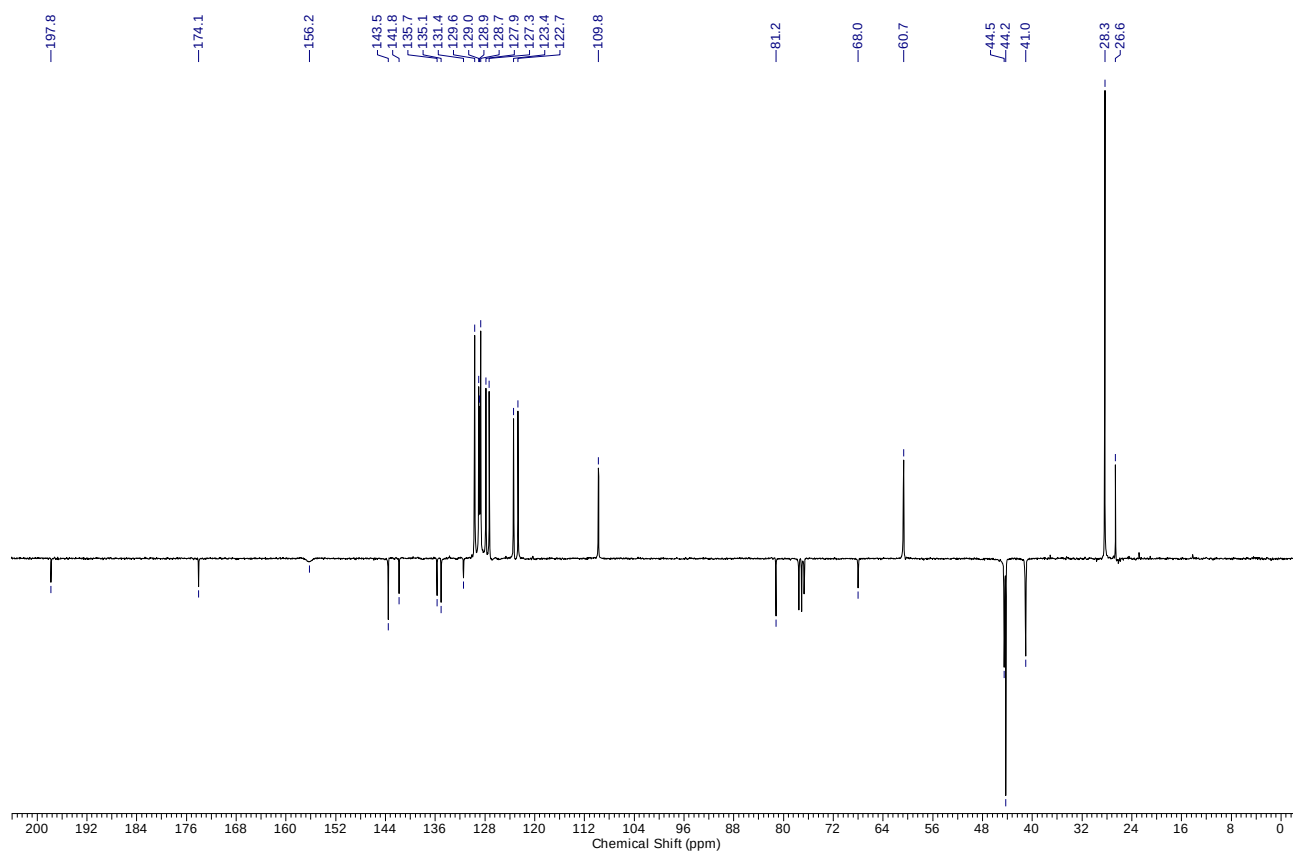
**6e**  $^{13}\text{C}$  NMR (75 MHz, ATP,  $\text{CDCl}_3$ ):



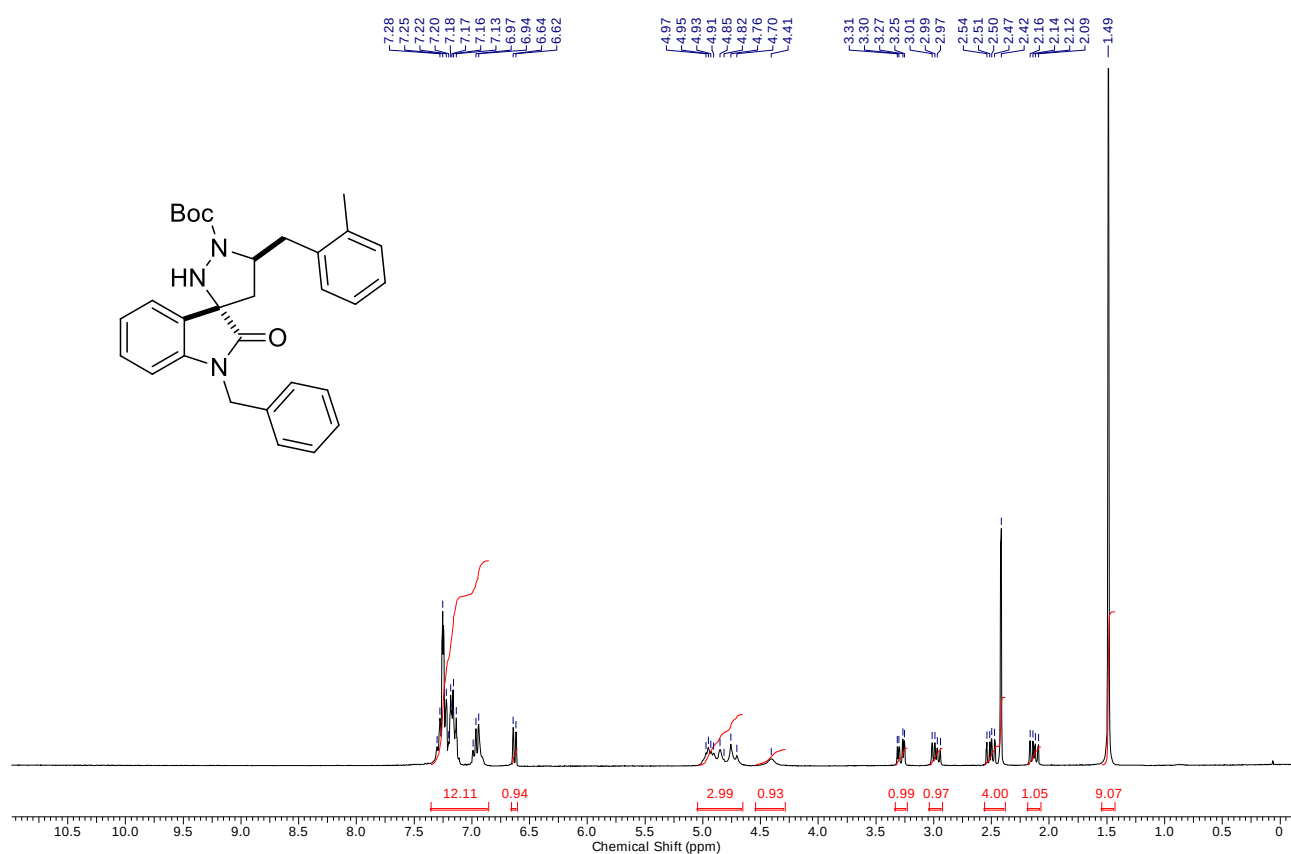
**6e'**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



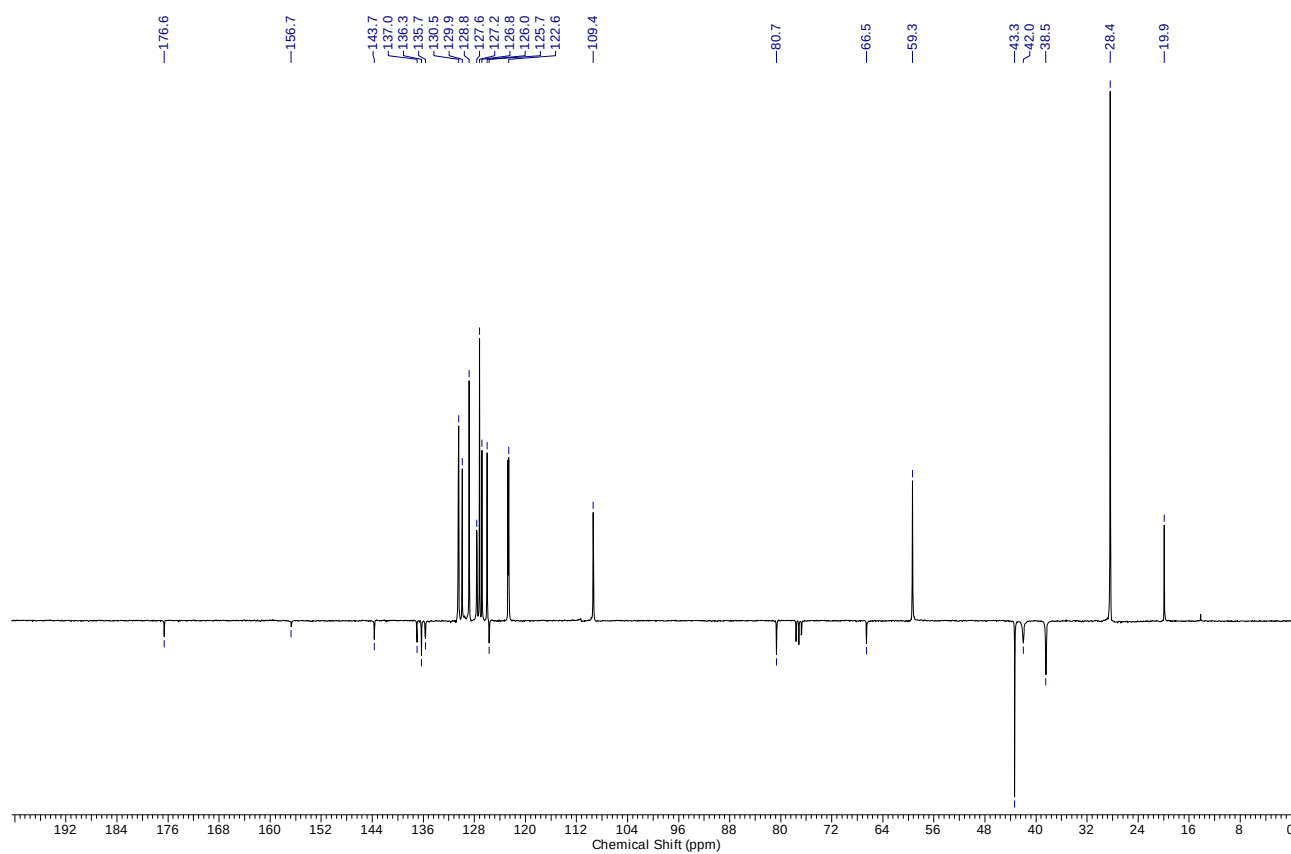
**6e'**  $^{13}\text{C}$  NMR (75 MHz, ATP,  $\text{CDCl}_3$ ):



**6f**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):

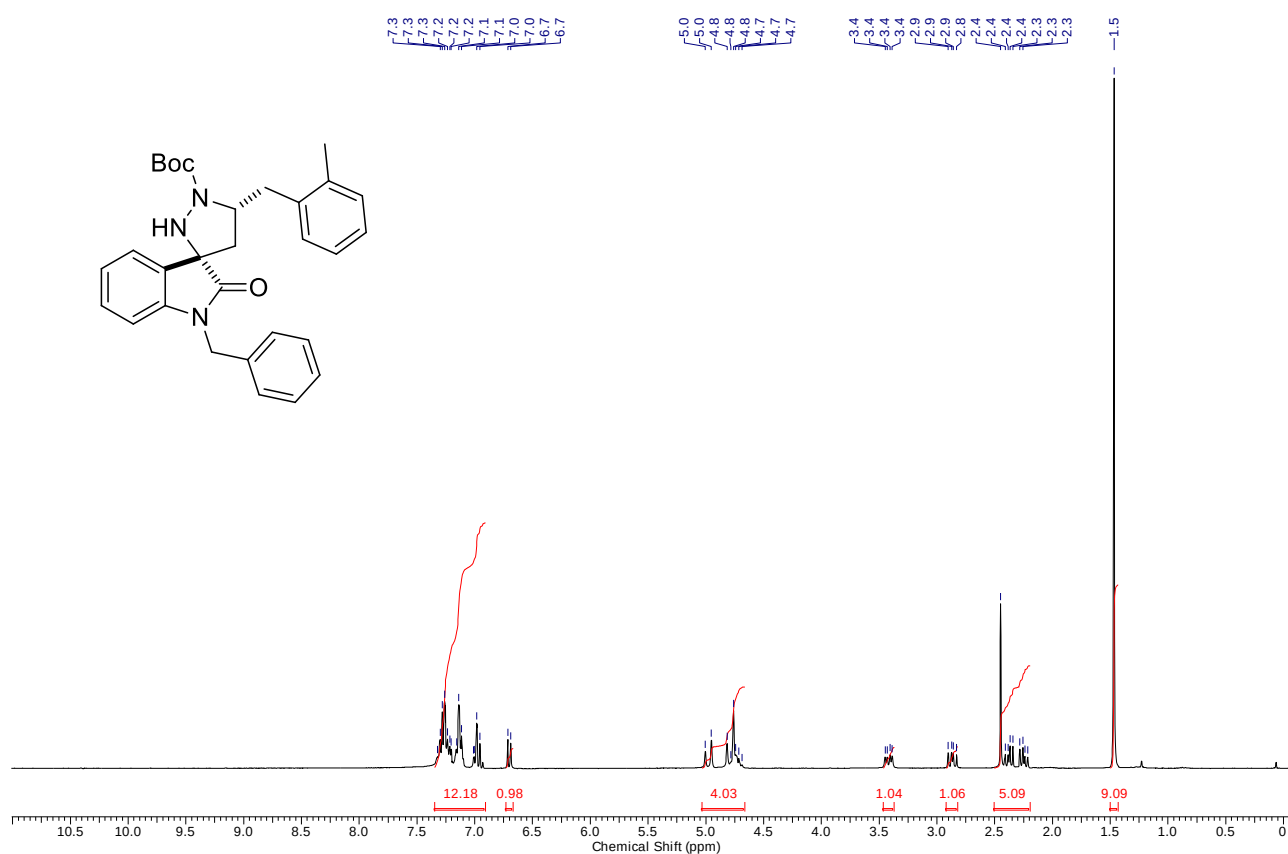


**6f**  $^{13}\text{C}$  NMR (75 MHz, ATP,  $\text{CDCl}_3$ ):

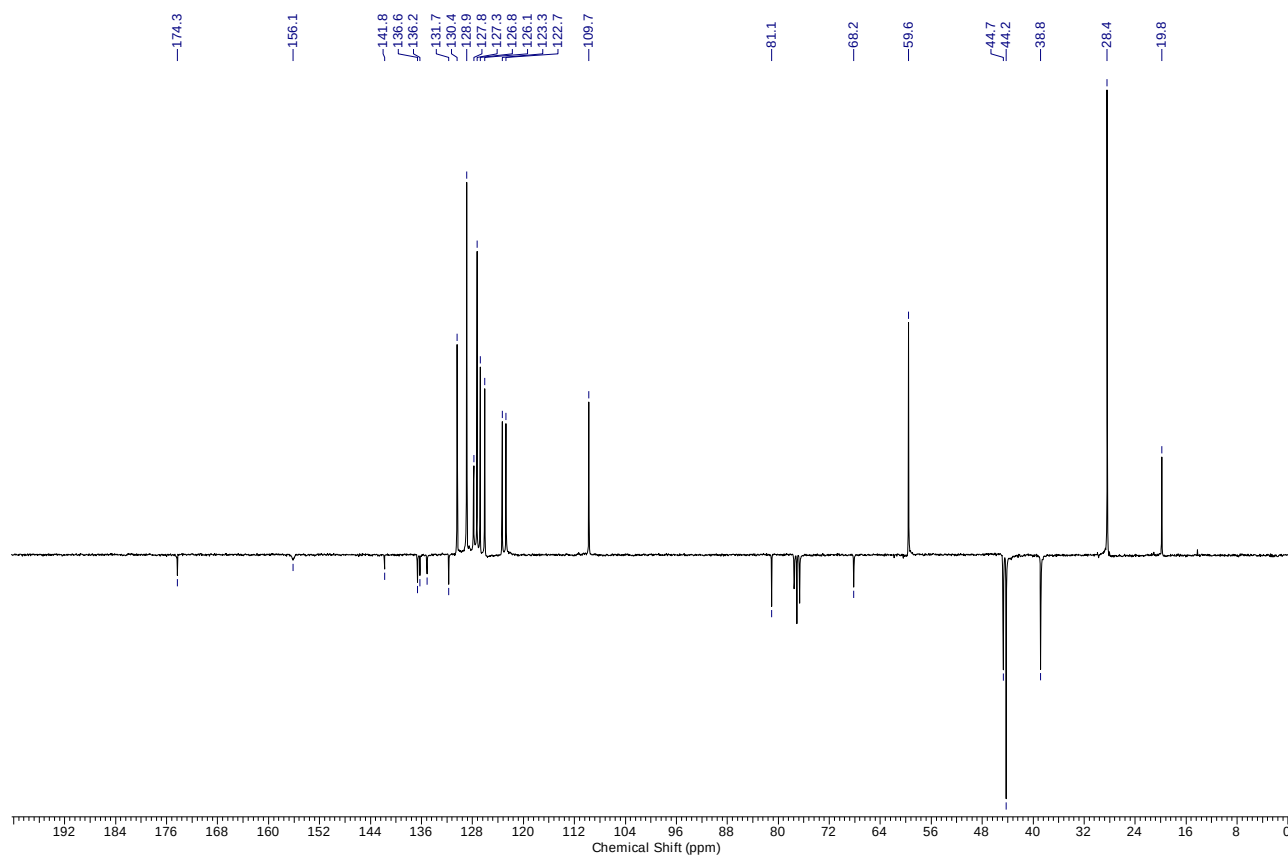




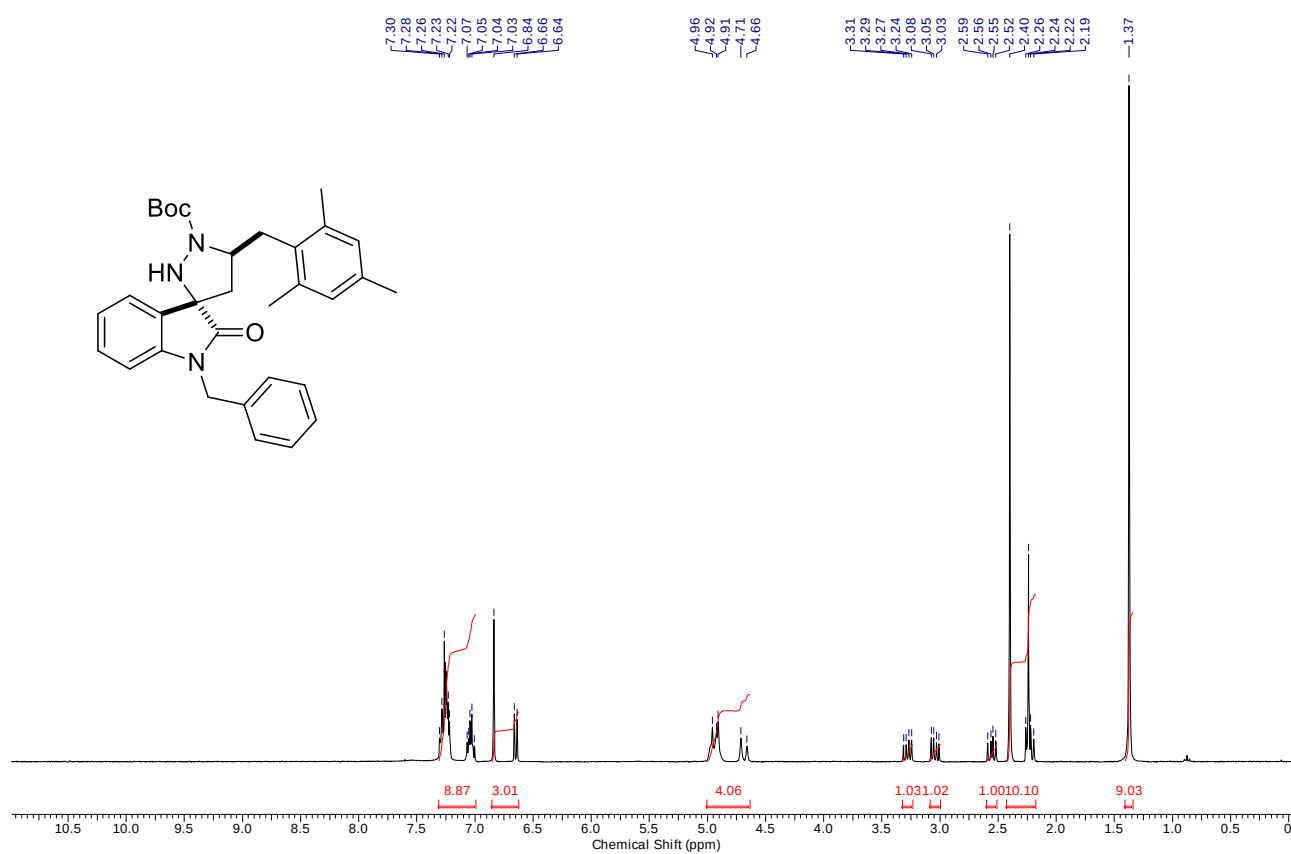
**6f'**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



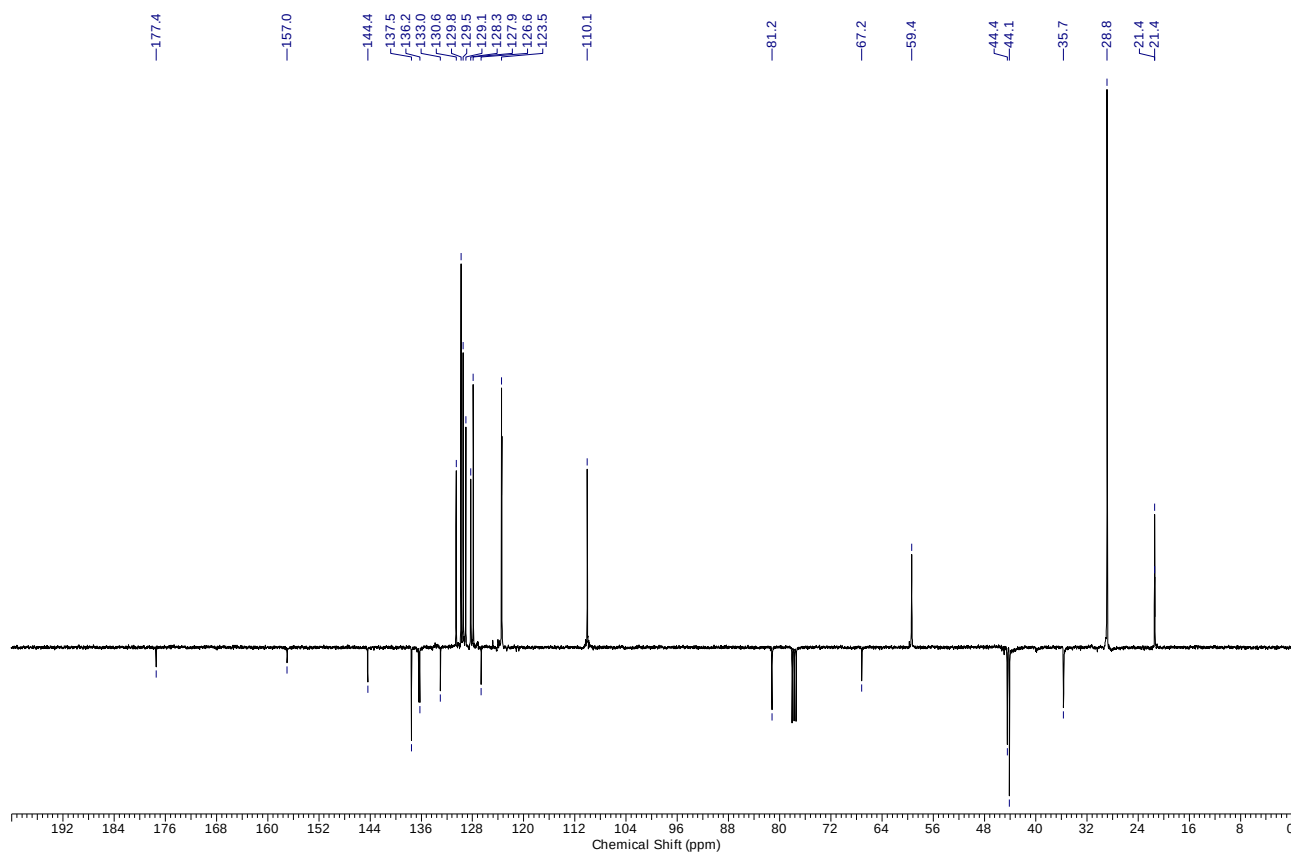
**6f'**  $^{13}\text{C}$  NMR (75 MHz, ATP,  $\text{CDCl}_3$ ):



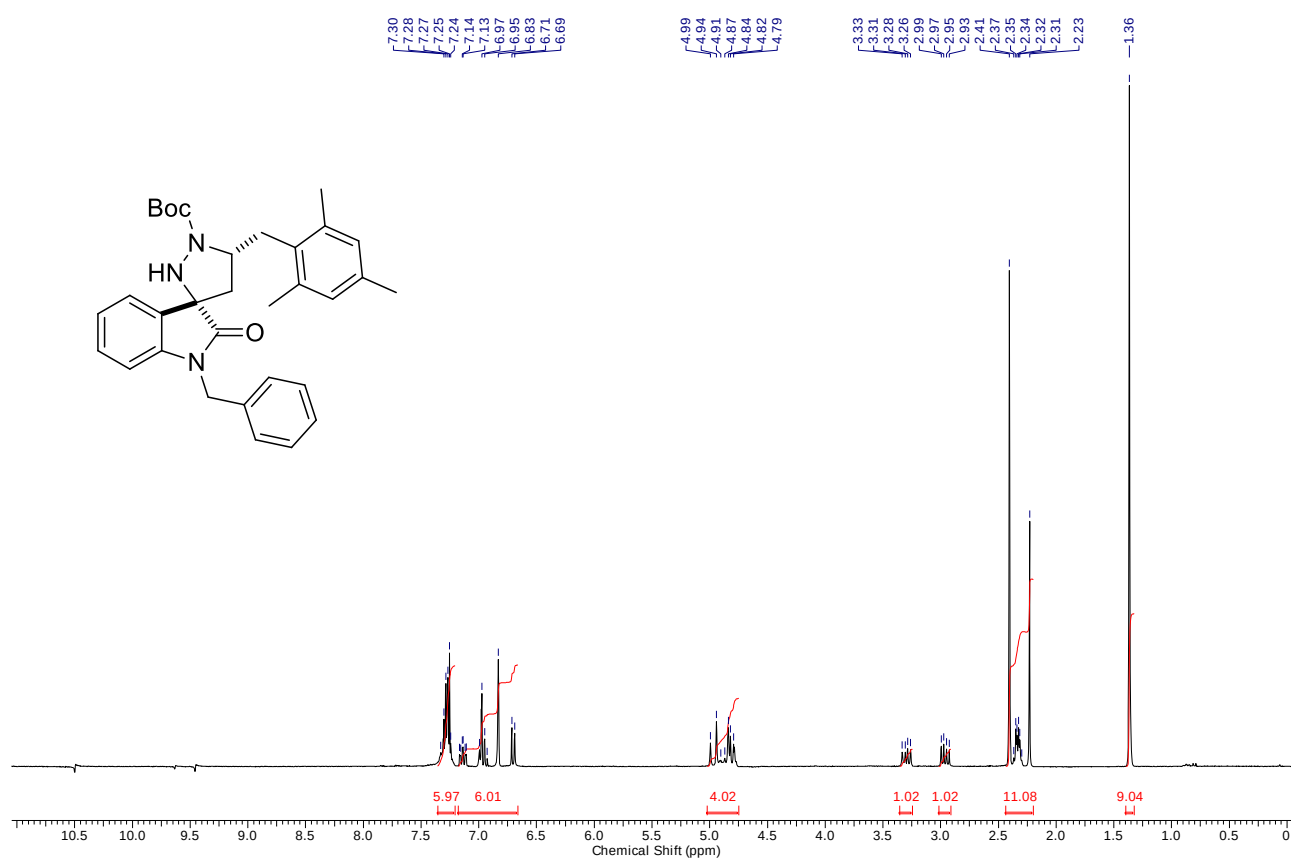
**6g**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



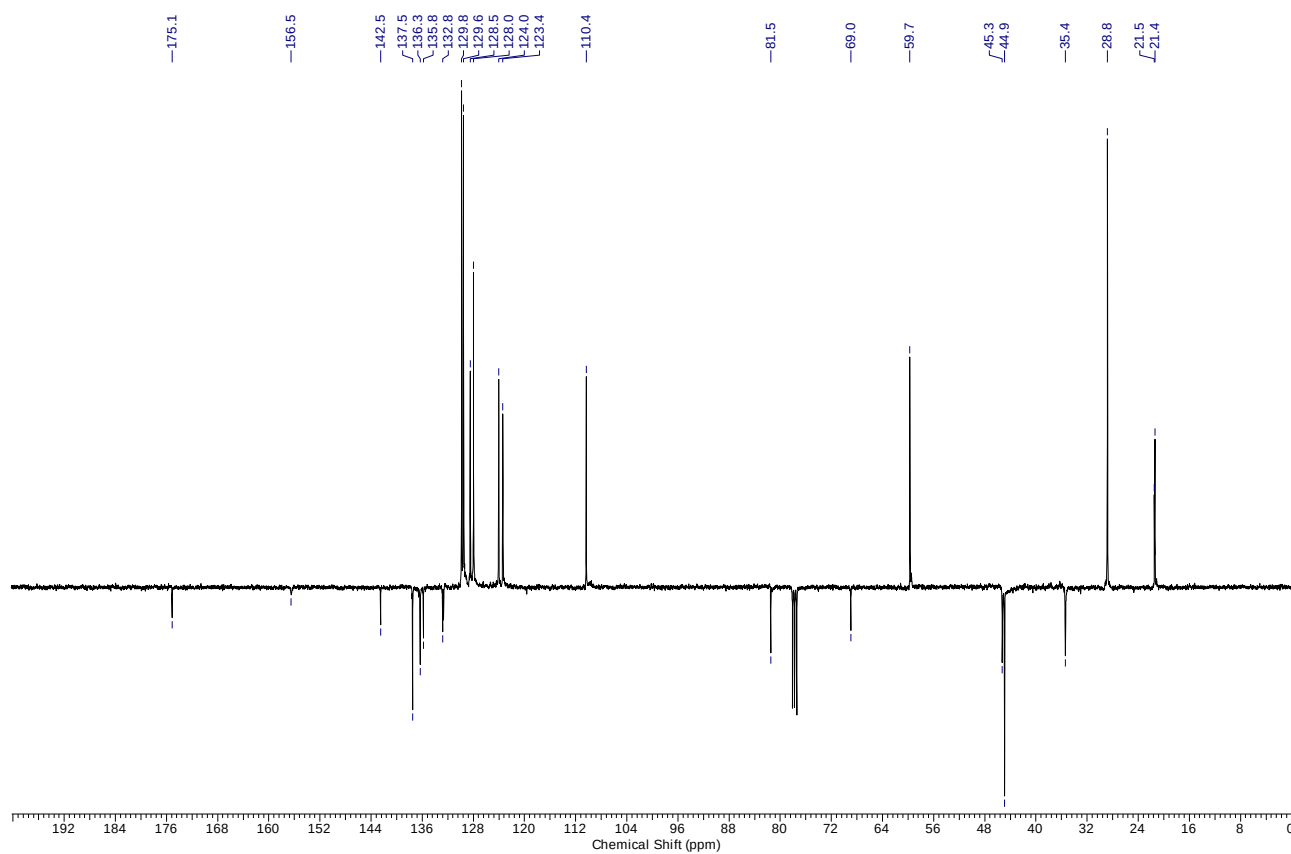
**6g**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



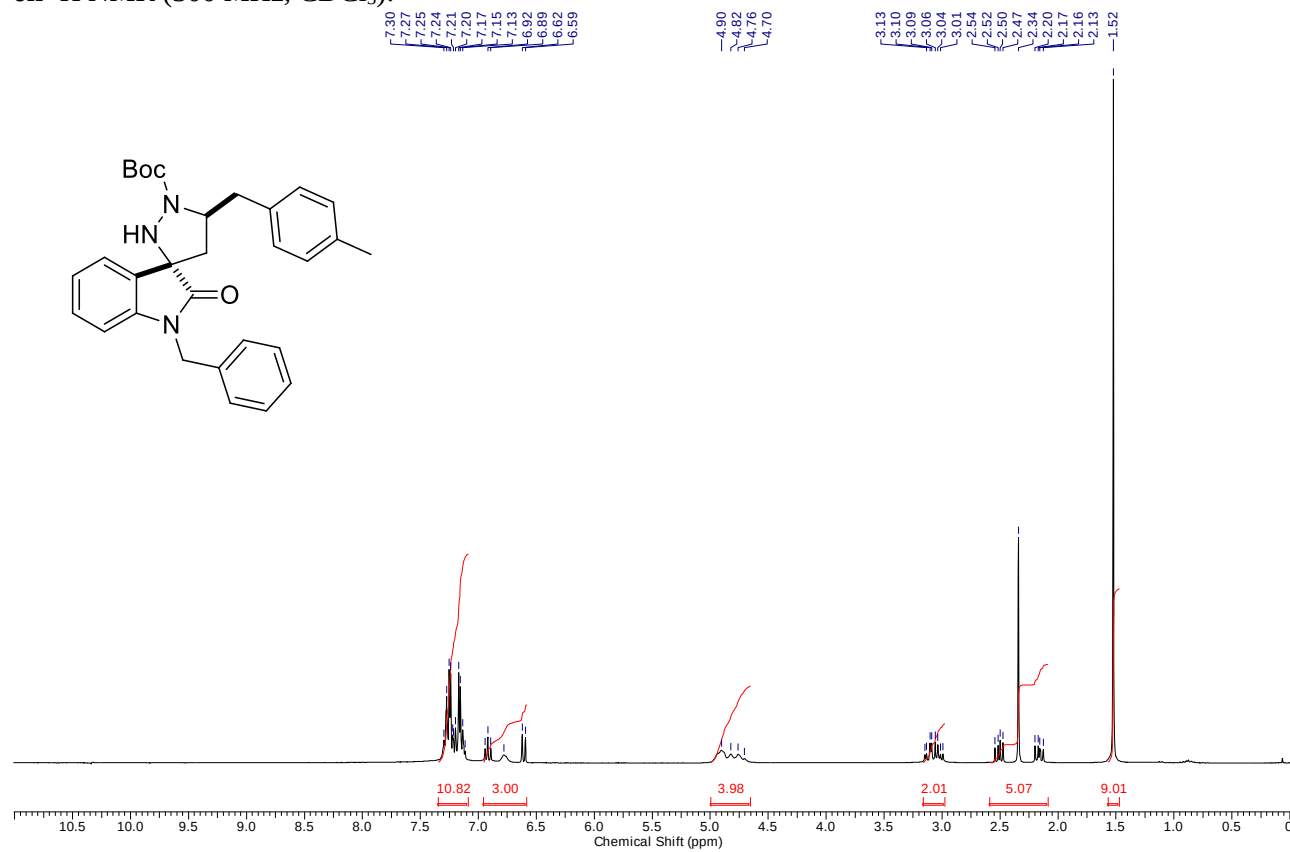
**6g'**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



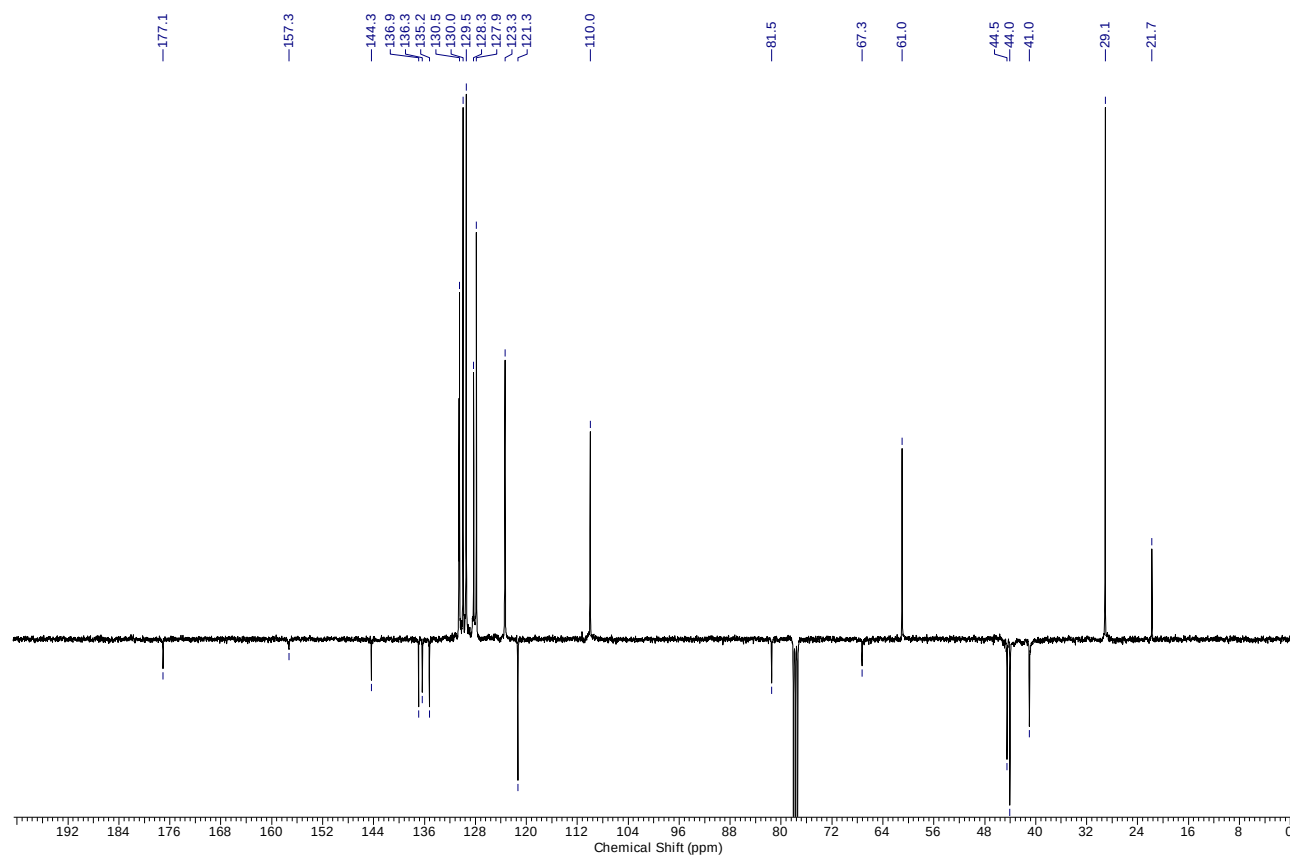
**6g'**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



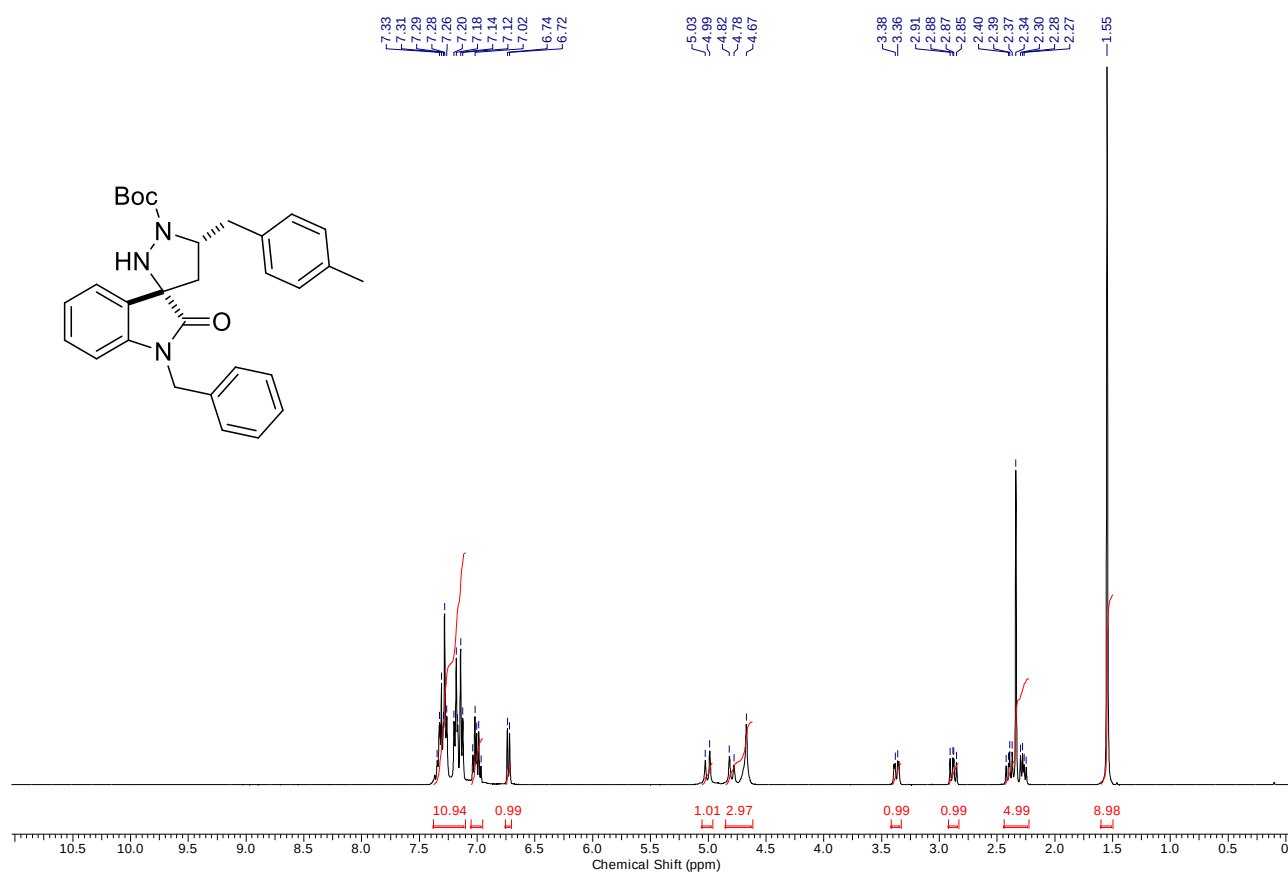
**6h**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



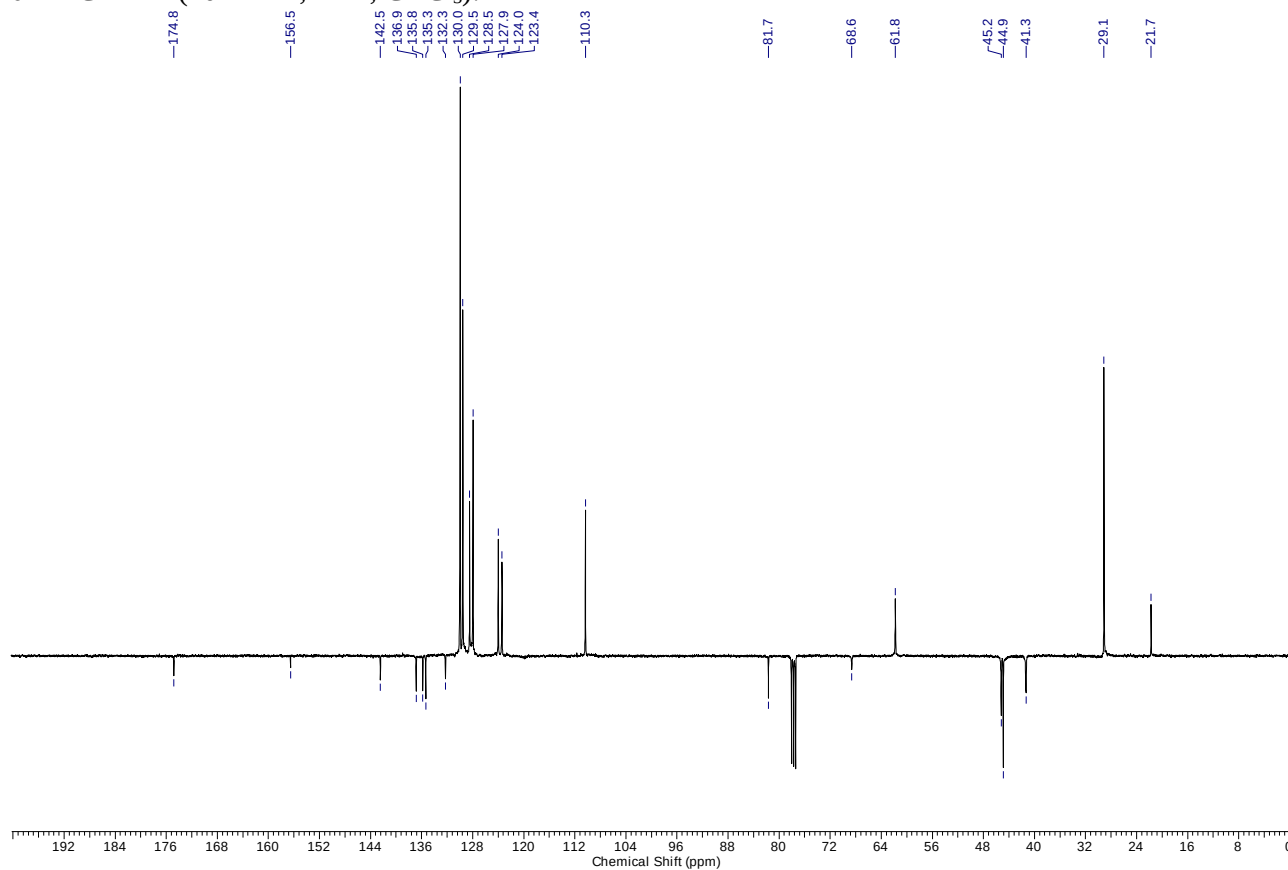
**6h**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):



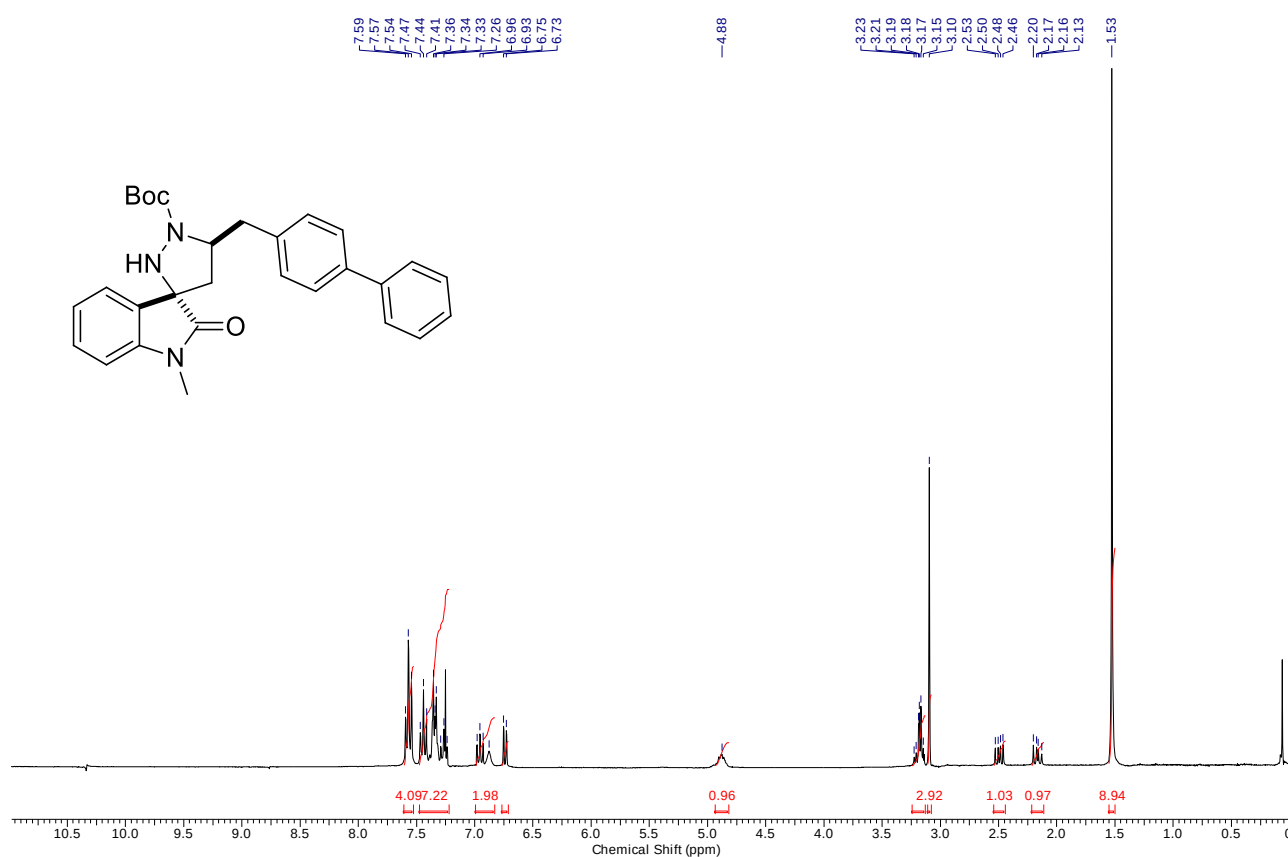
**6h'**  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):



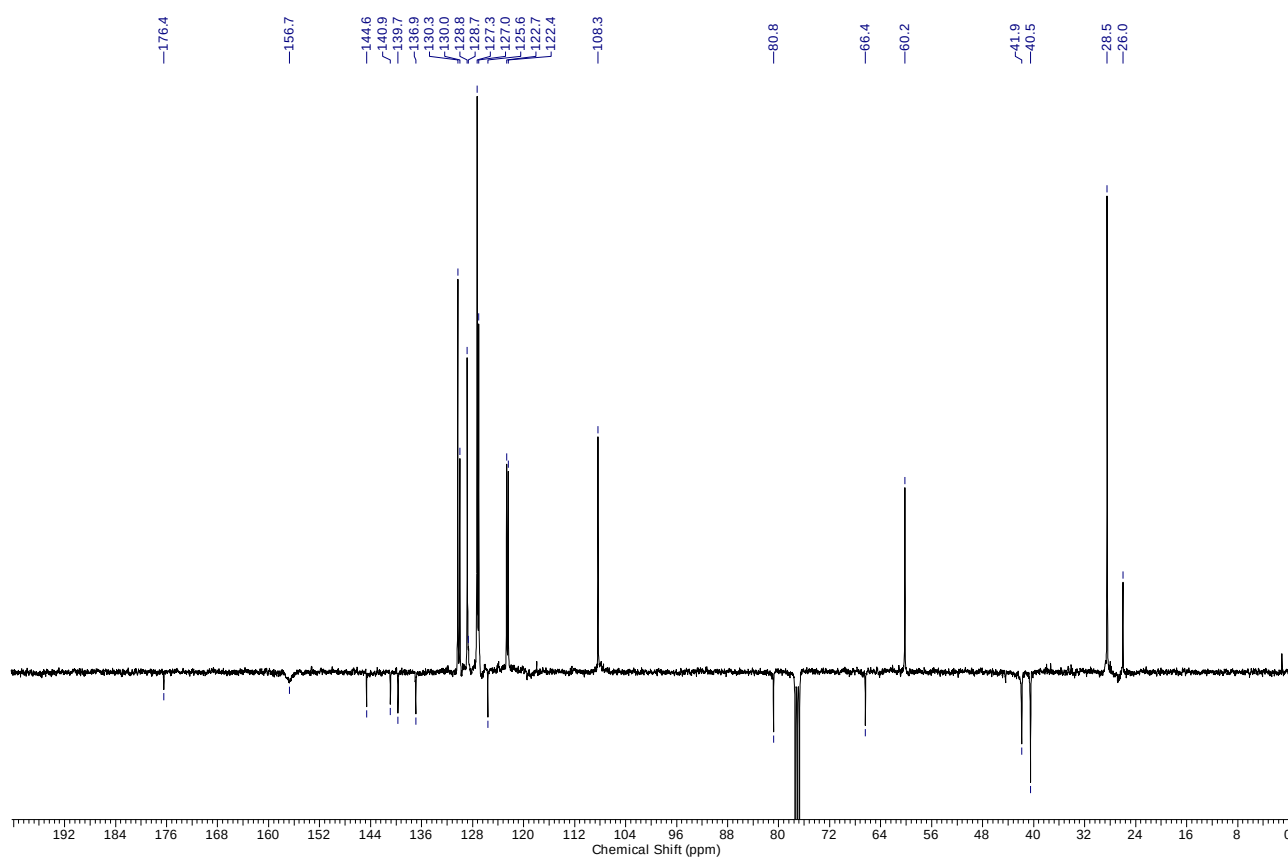
**6h'**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



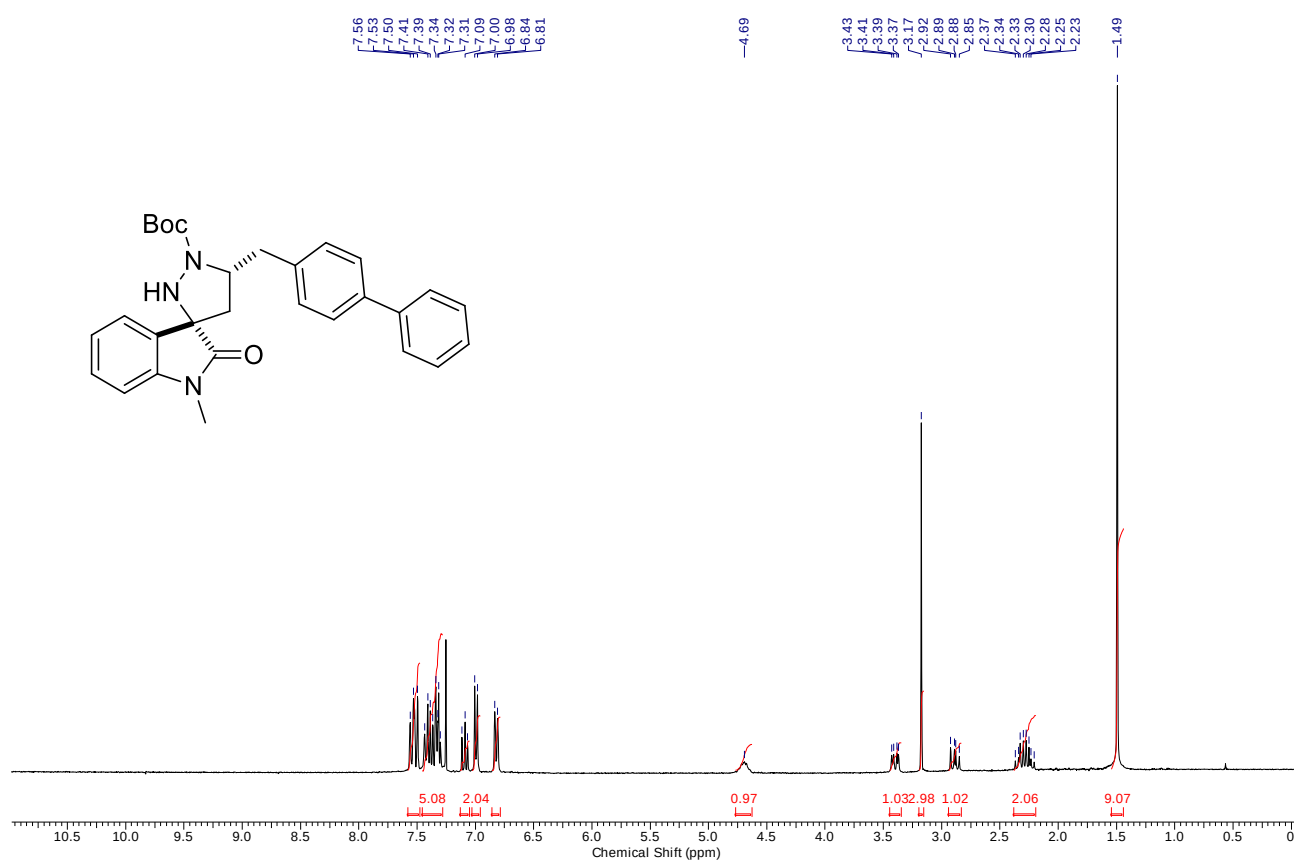
**6i**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



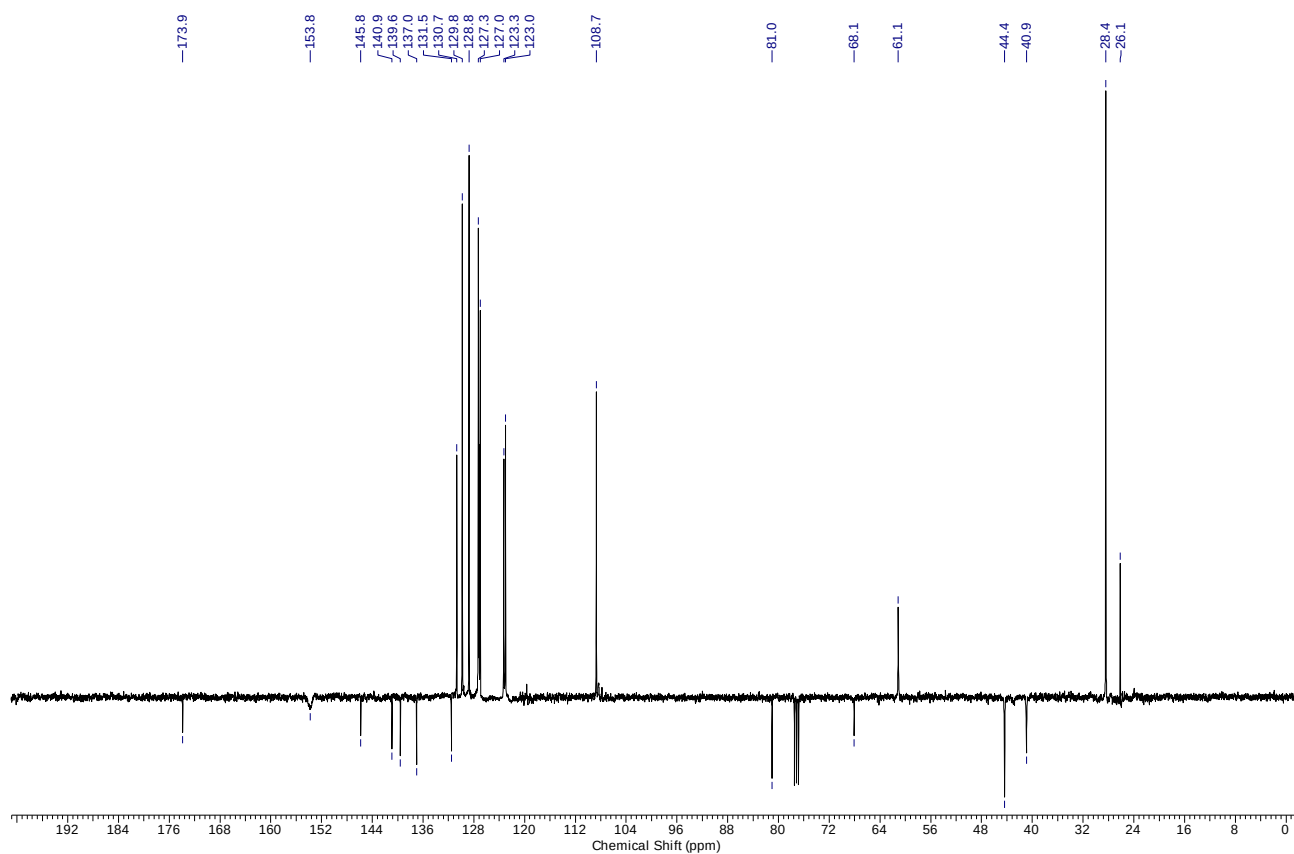
**6i**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):



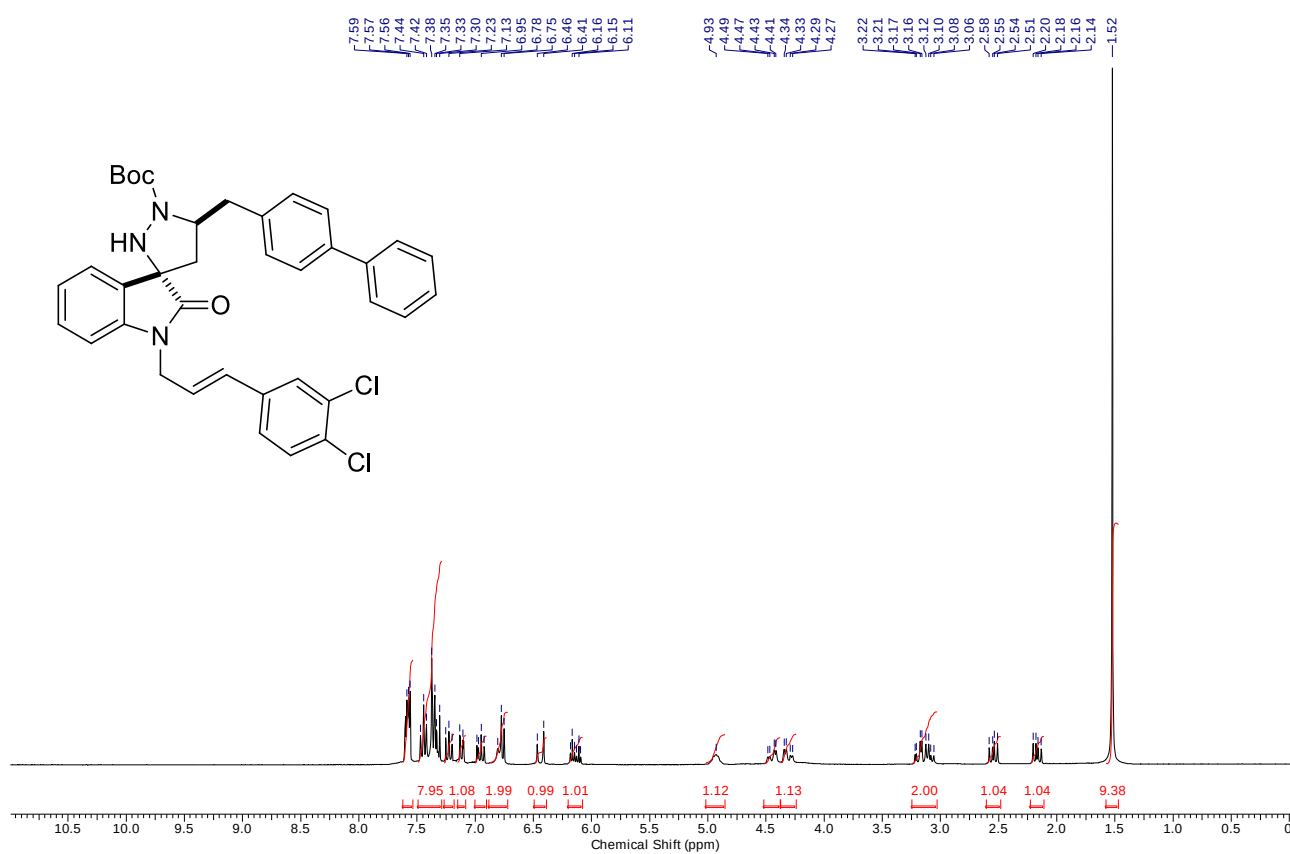
**6i'**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



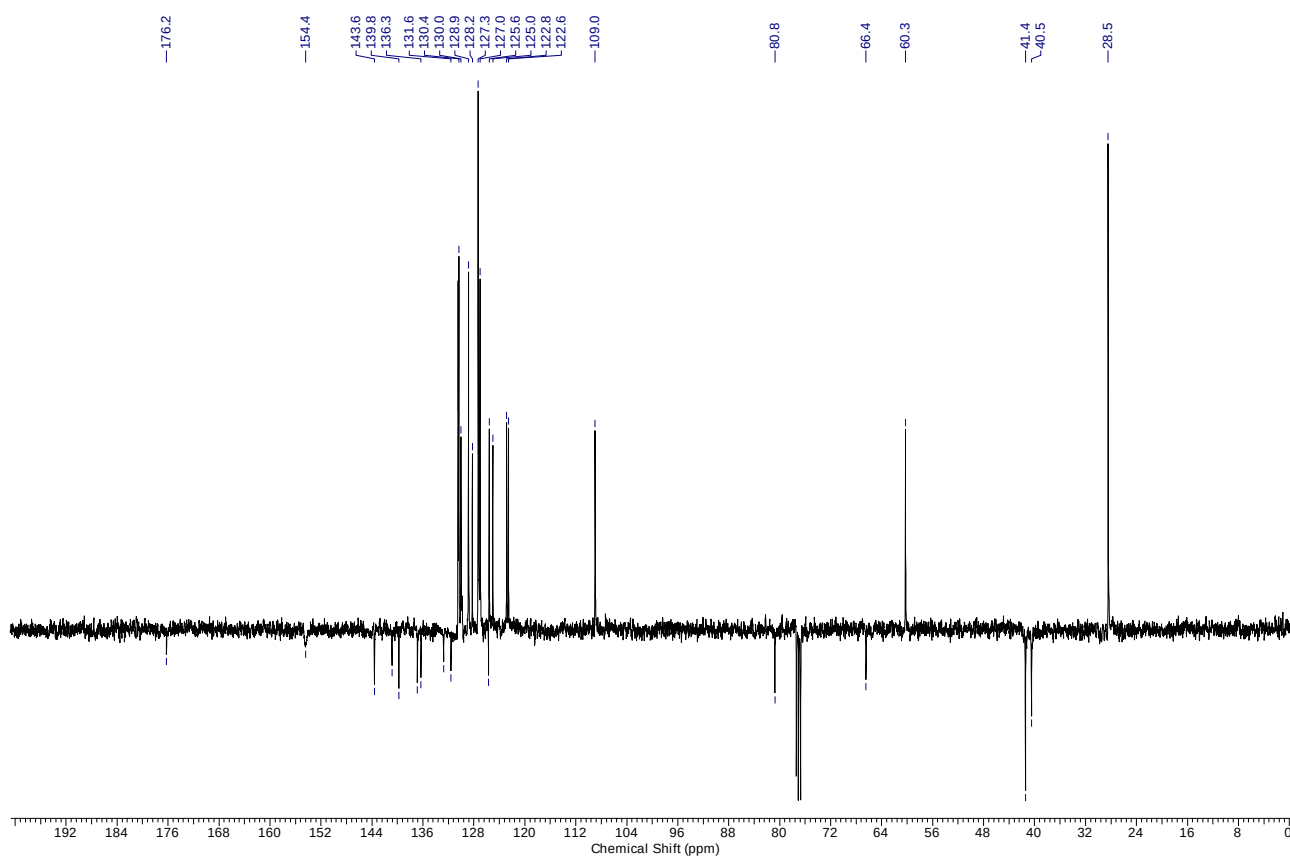
**6i'**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



**6j**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):

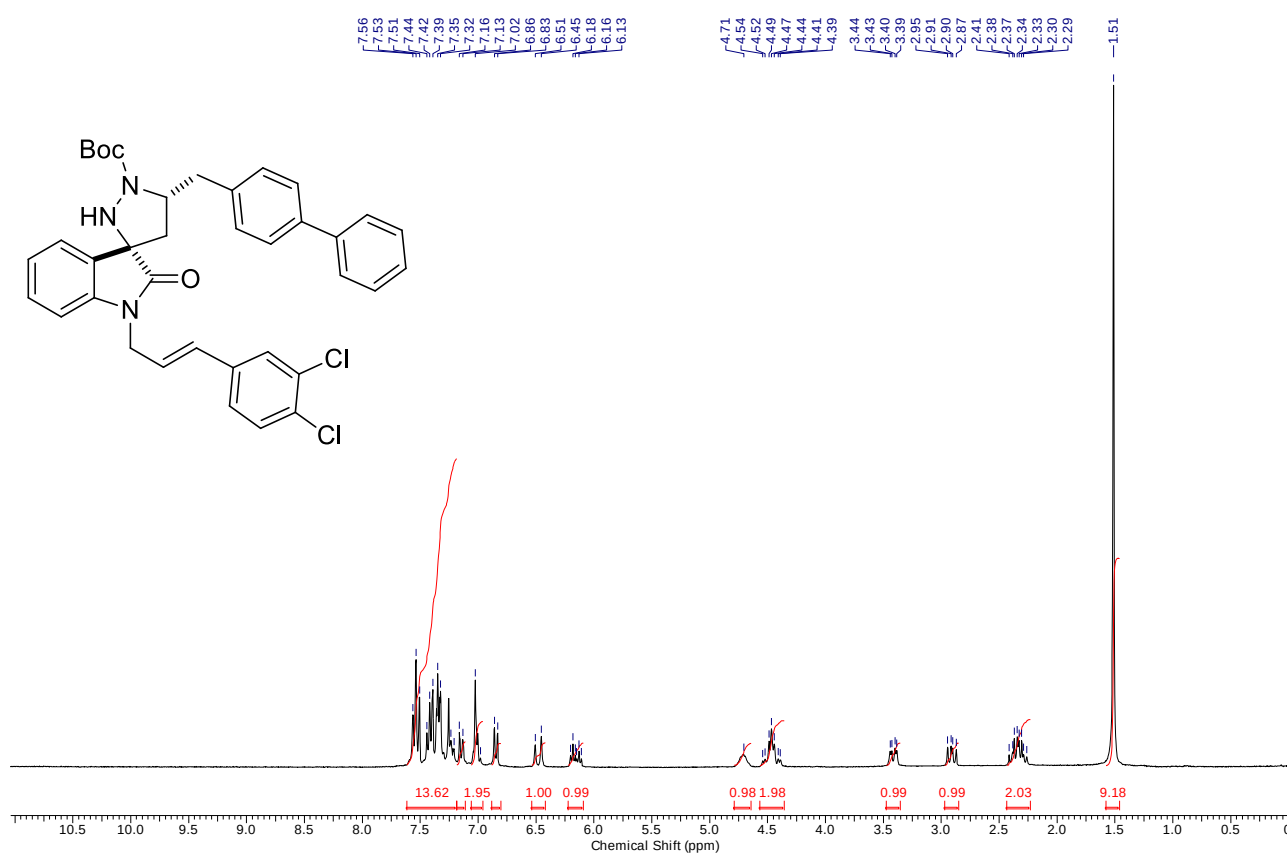


**6j**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):

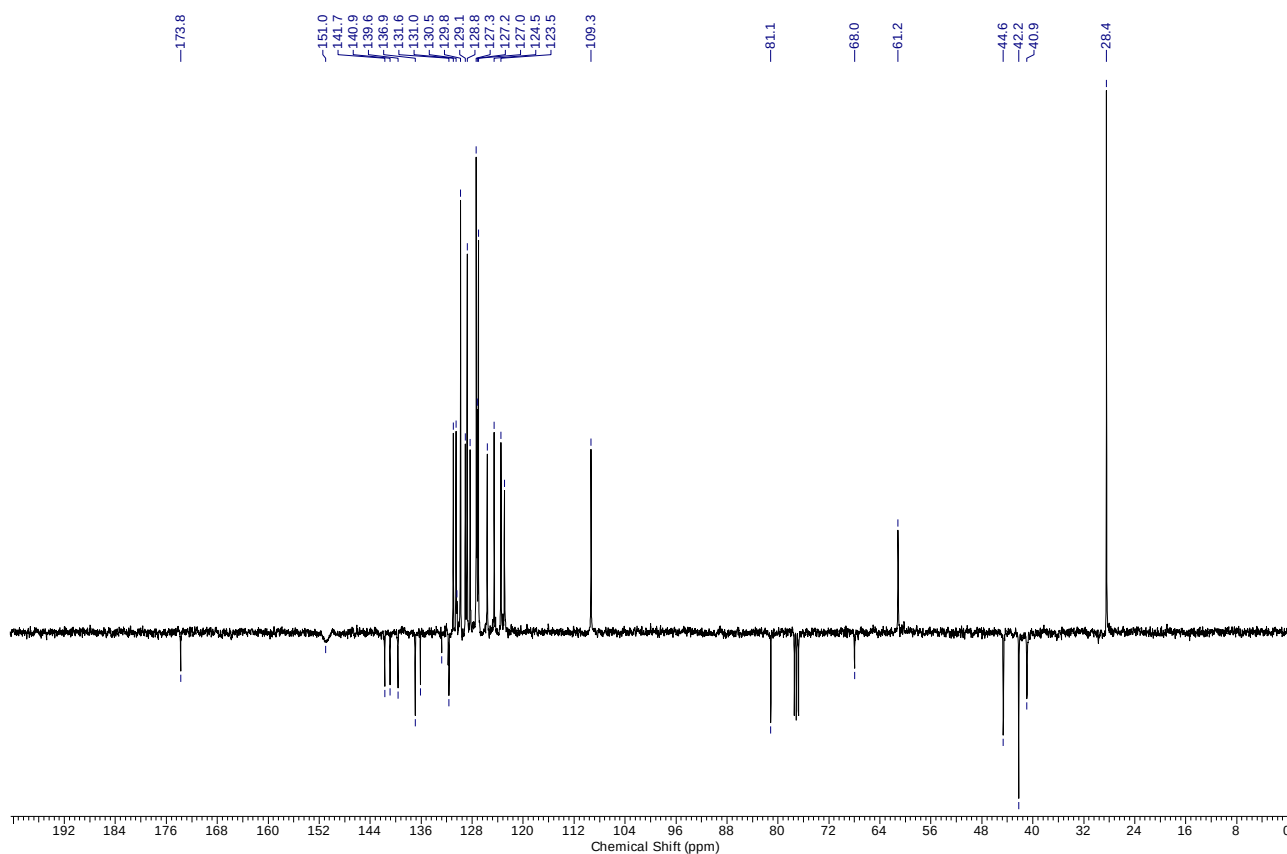




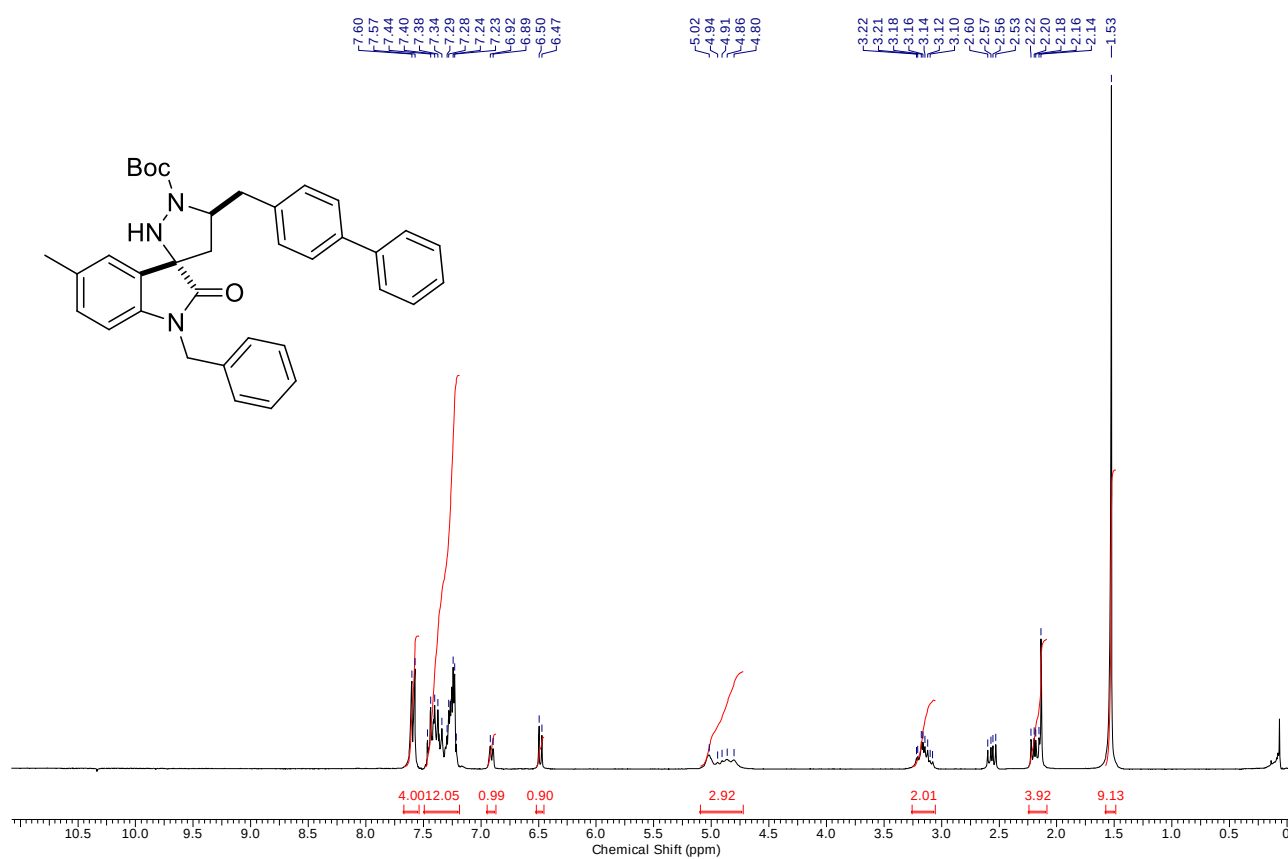
**6j'**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



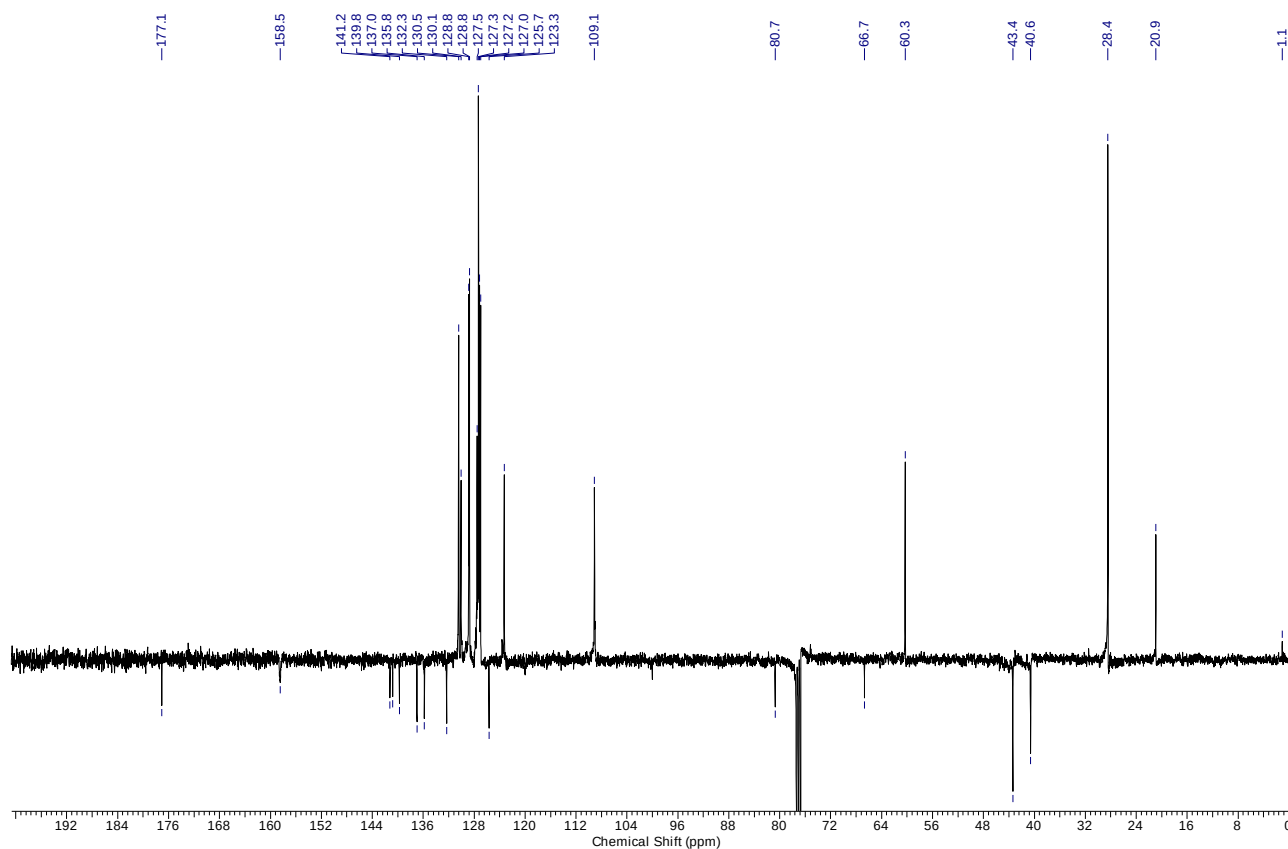
**6j'**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



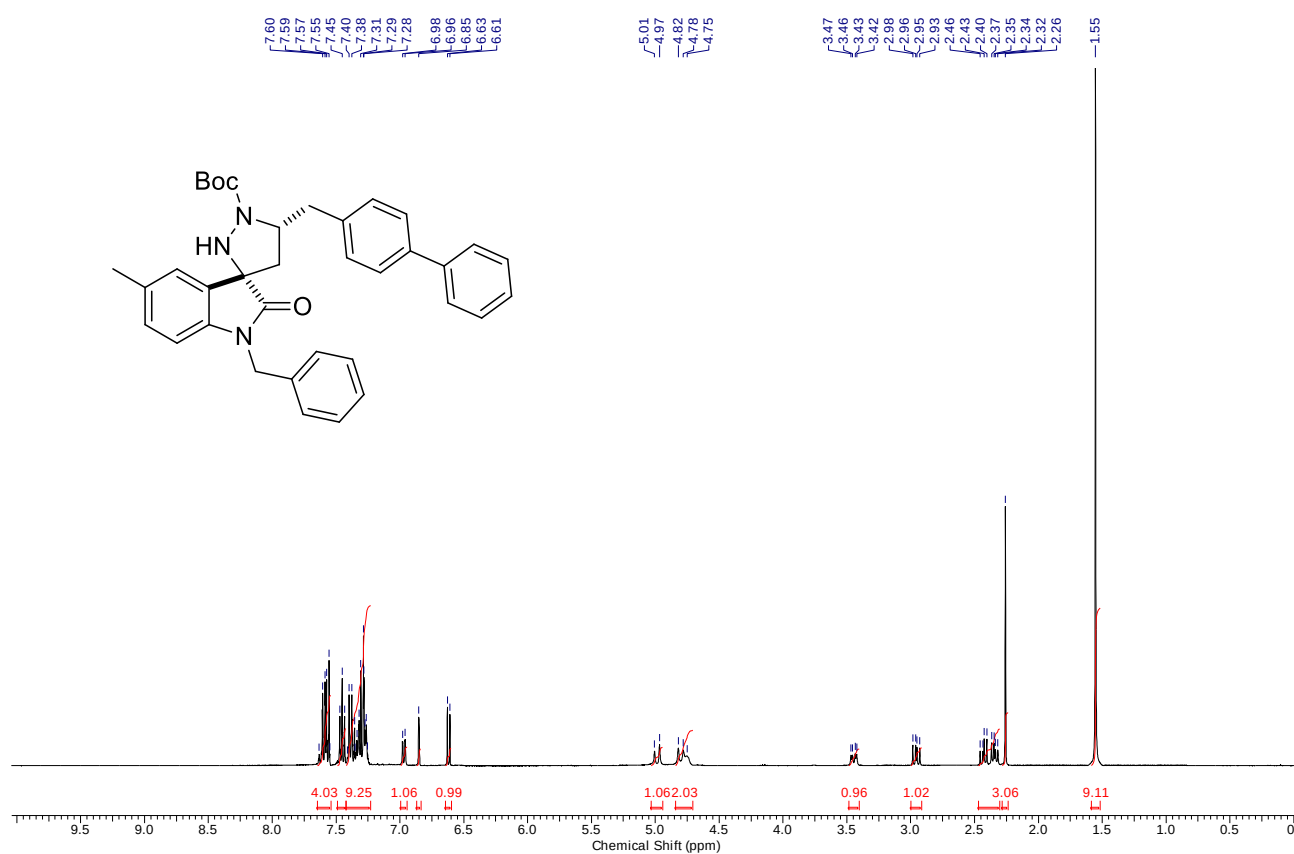
**6k**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



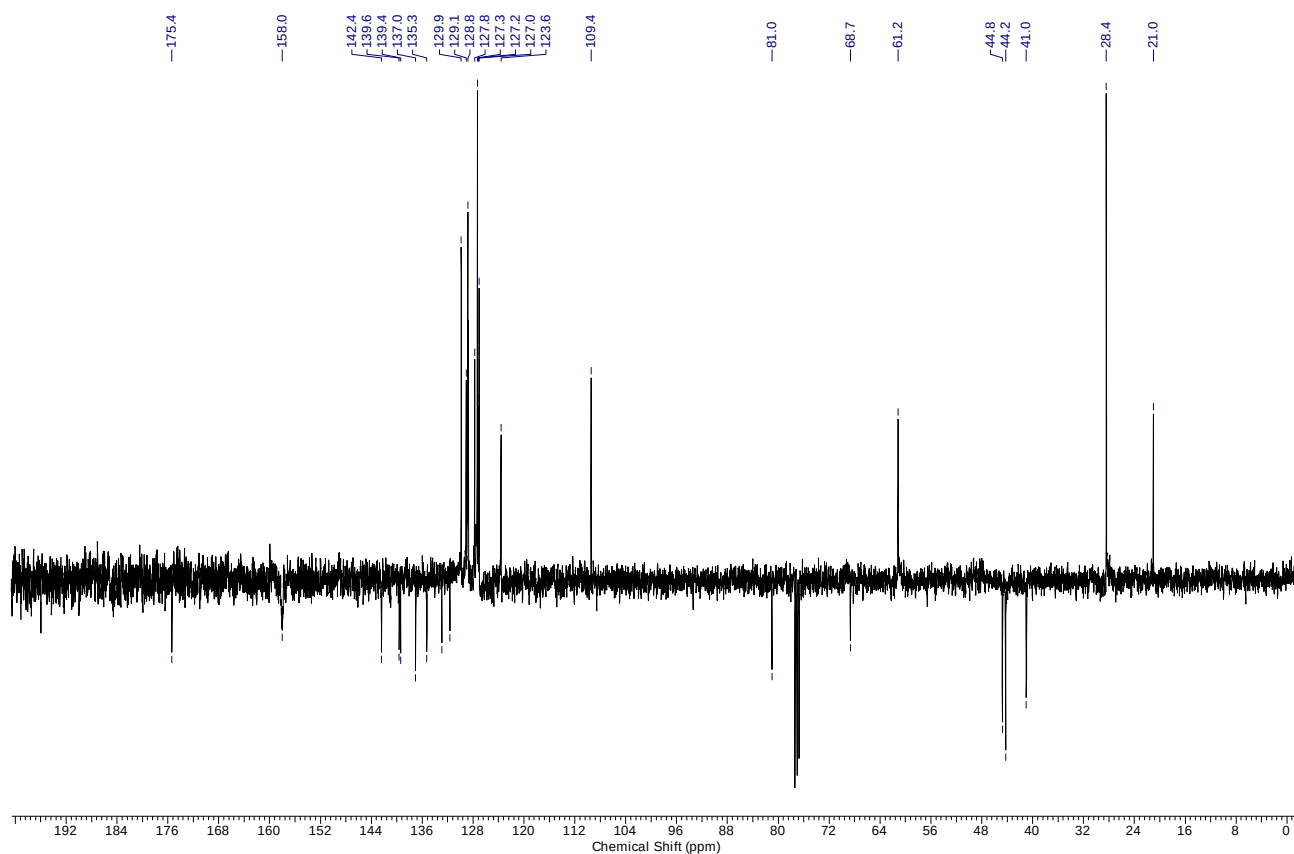
**6k**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):



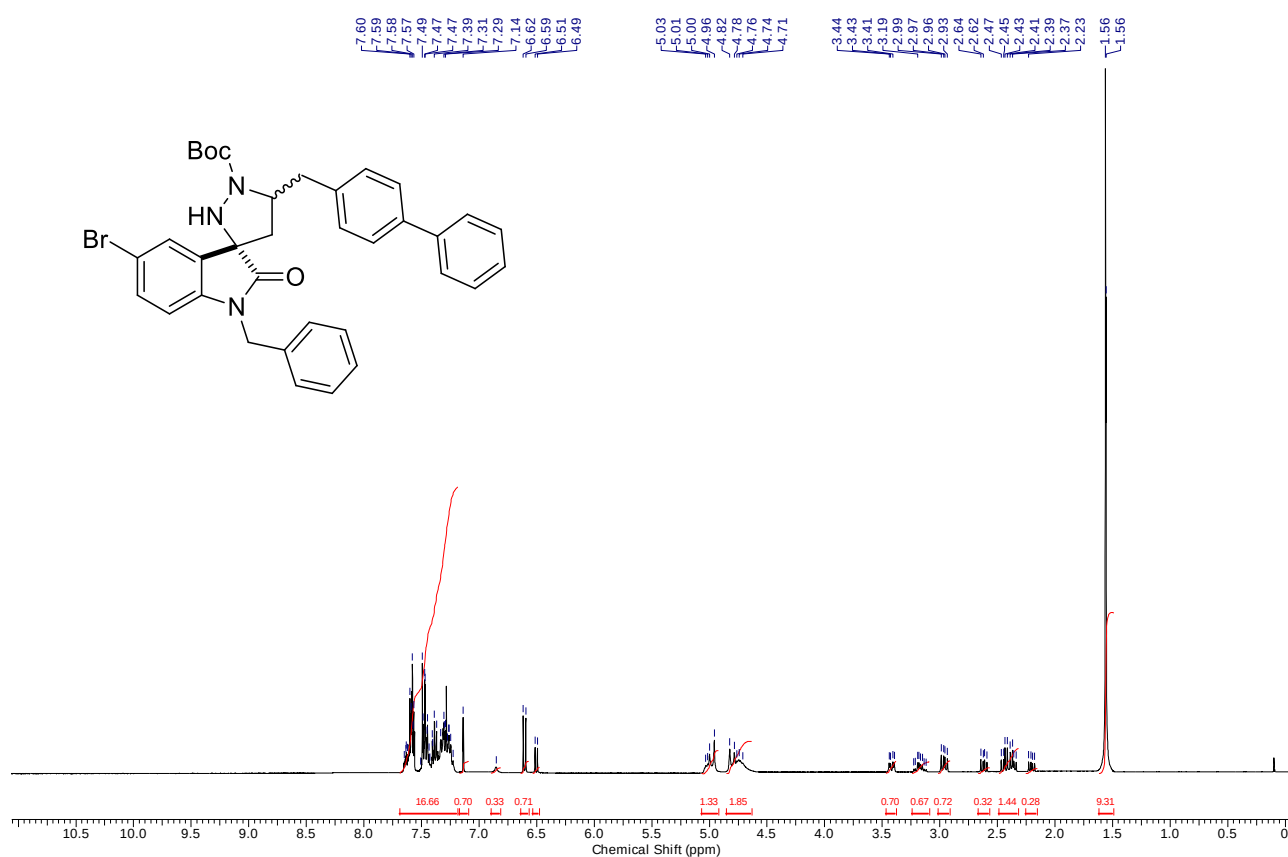
**6k'**  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):



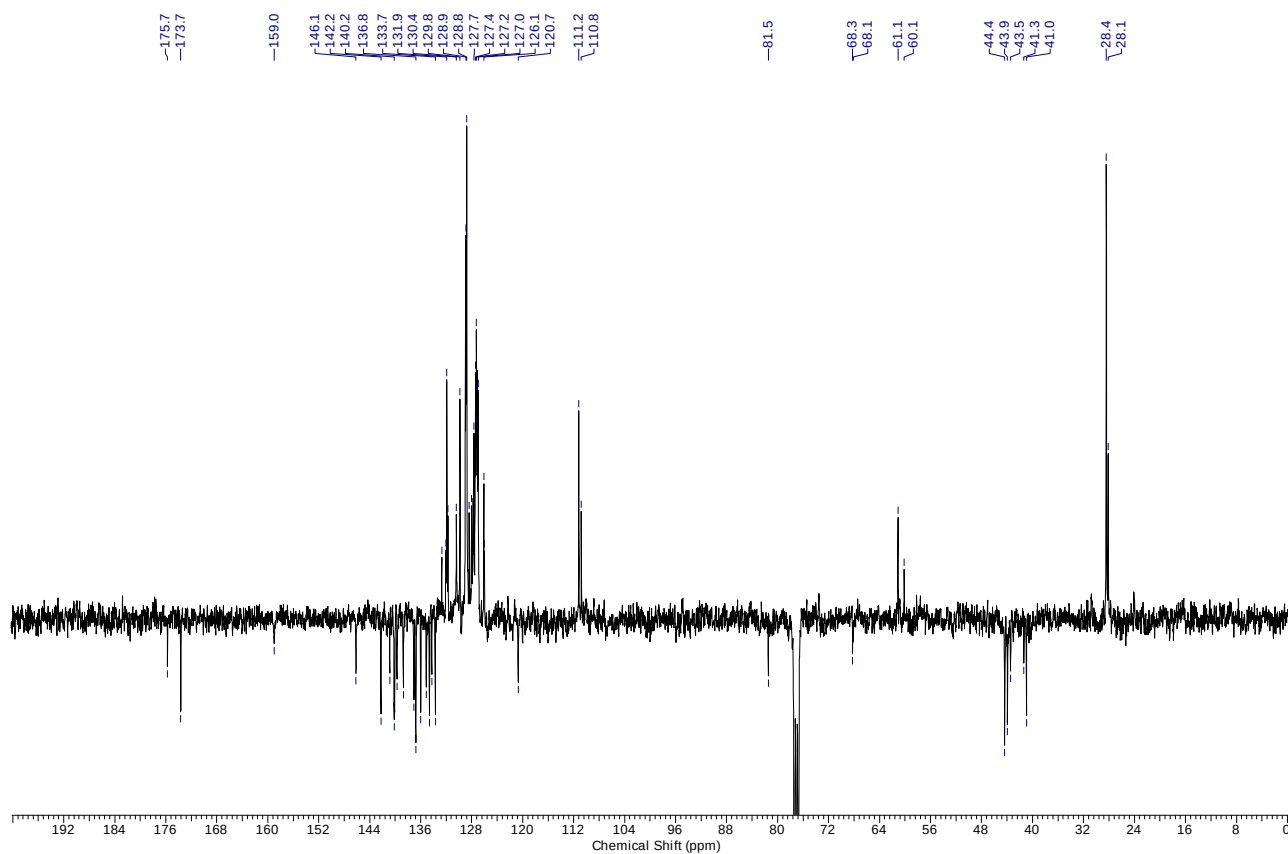
**6k'**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):



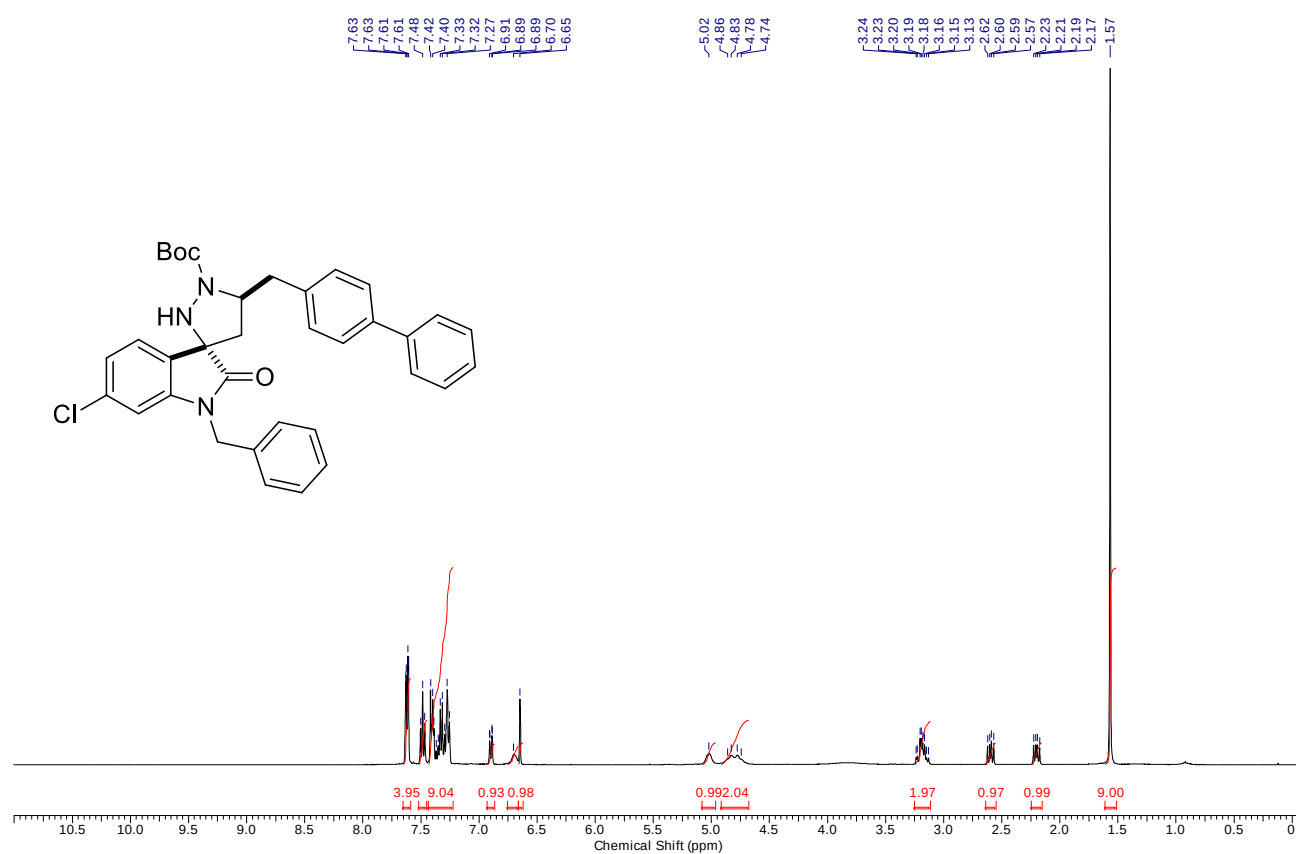
**6l,6l'** (7:3 mixture of diastereoisomers)  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):



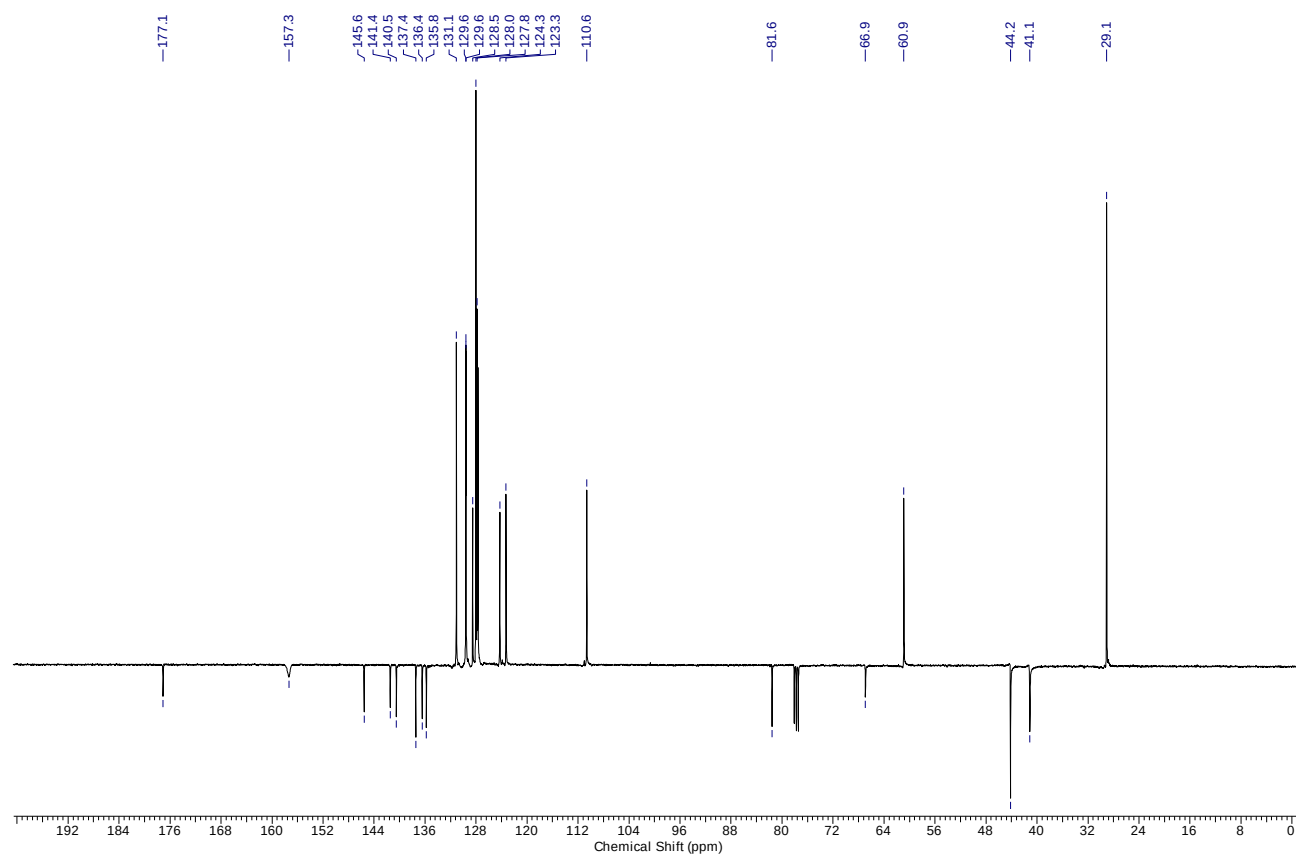
**6l,6l'** (7:3 mixture of diastereoisomers)  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):



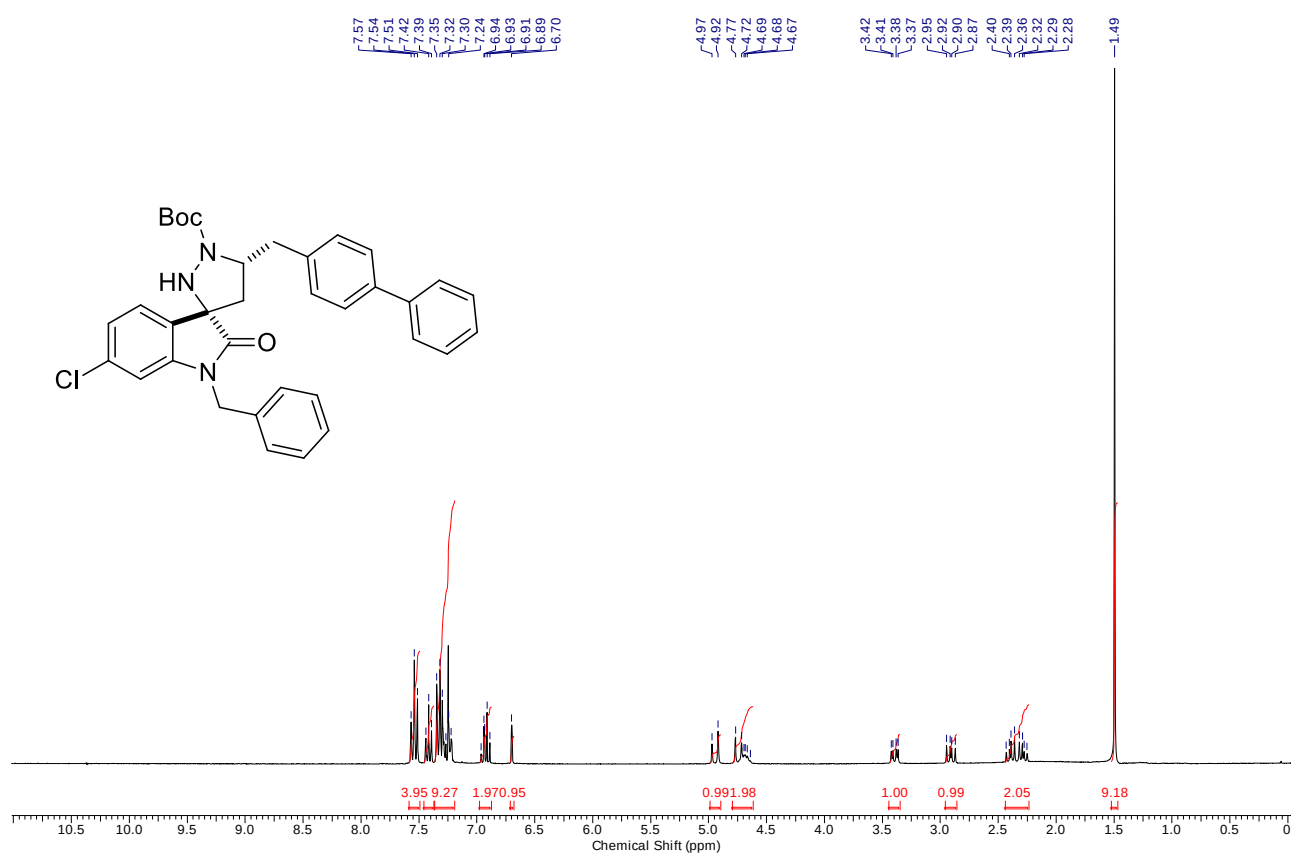
**6m**  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):



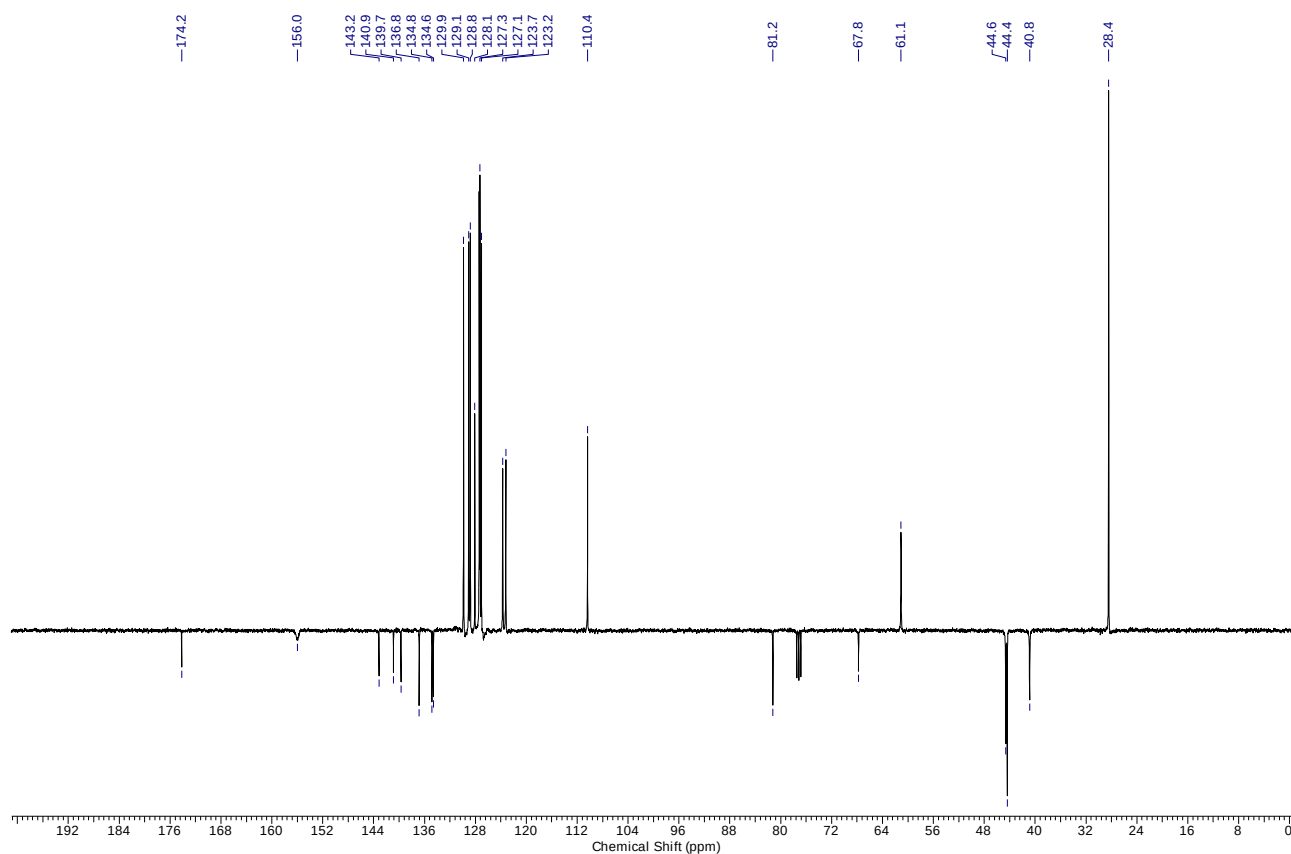
**6m**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



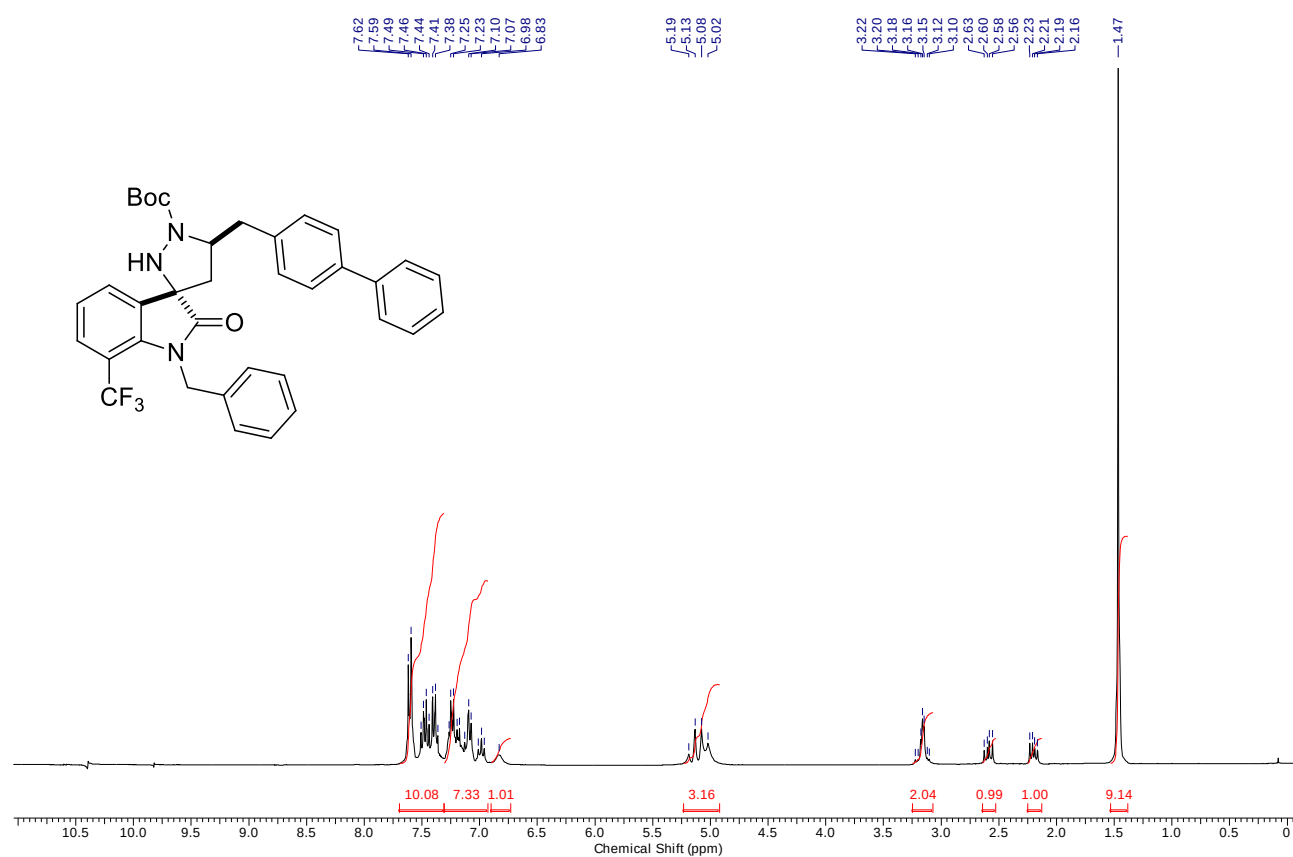
**6m'**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



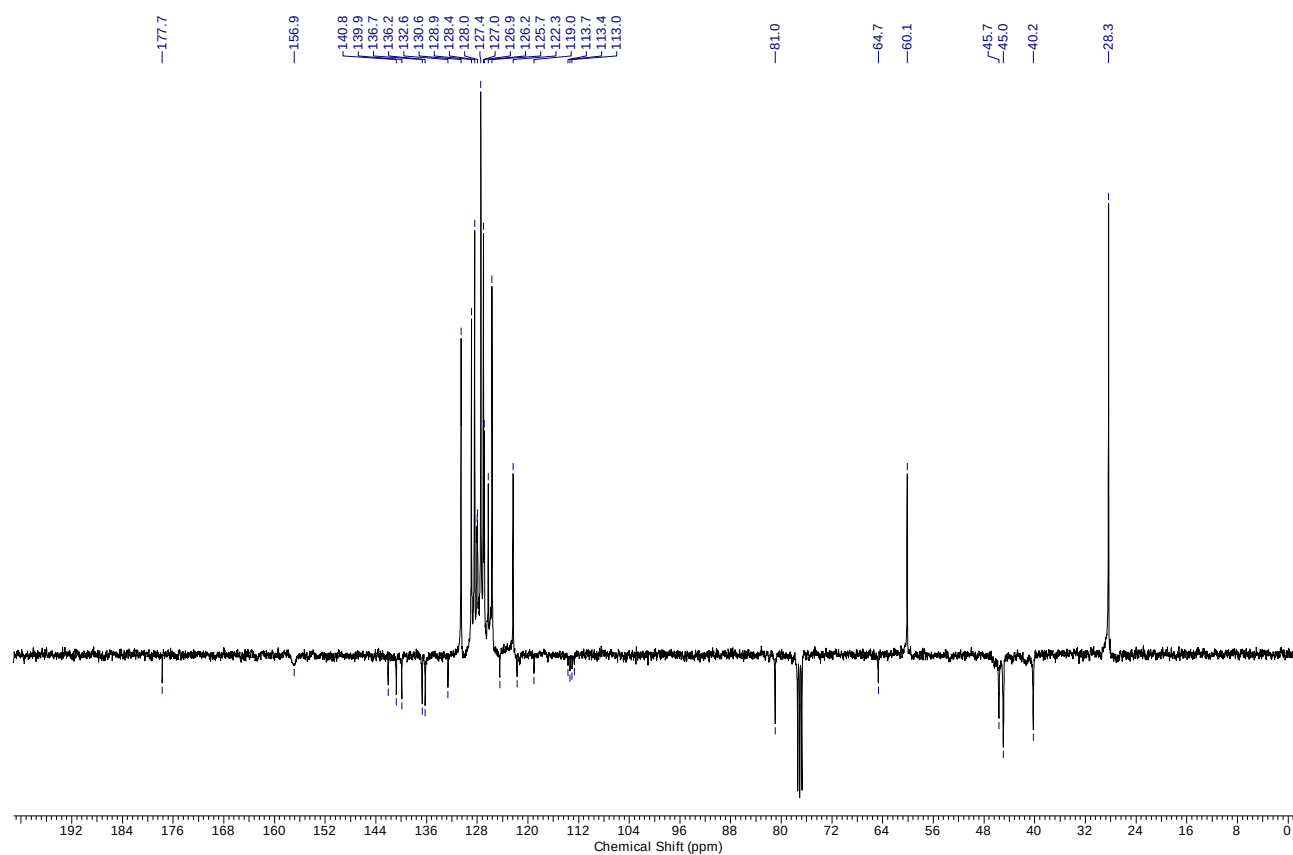
**6m'**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):



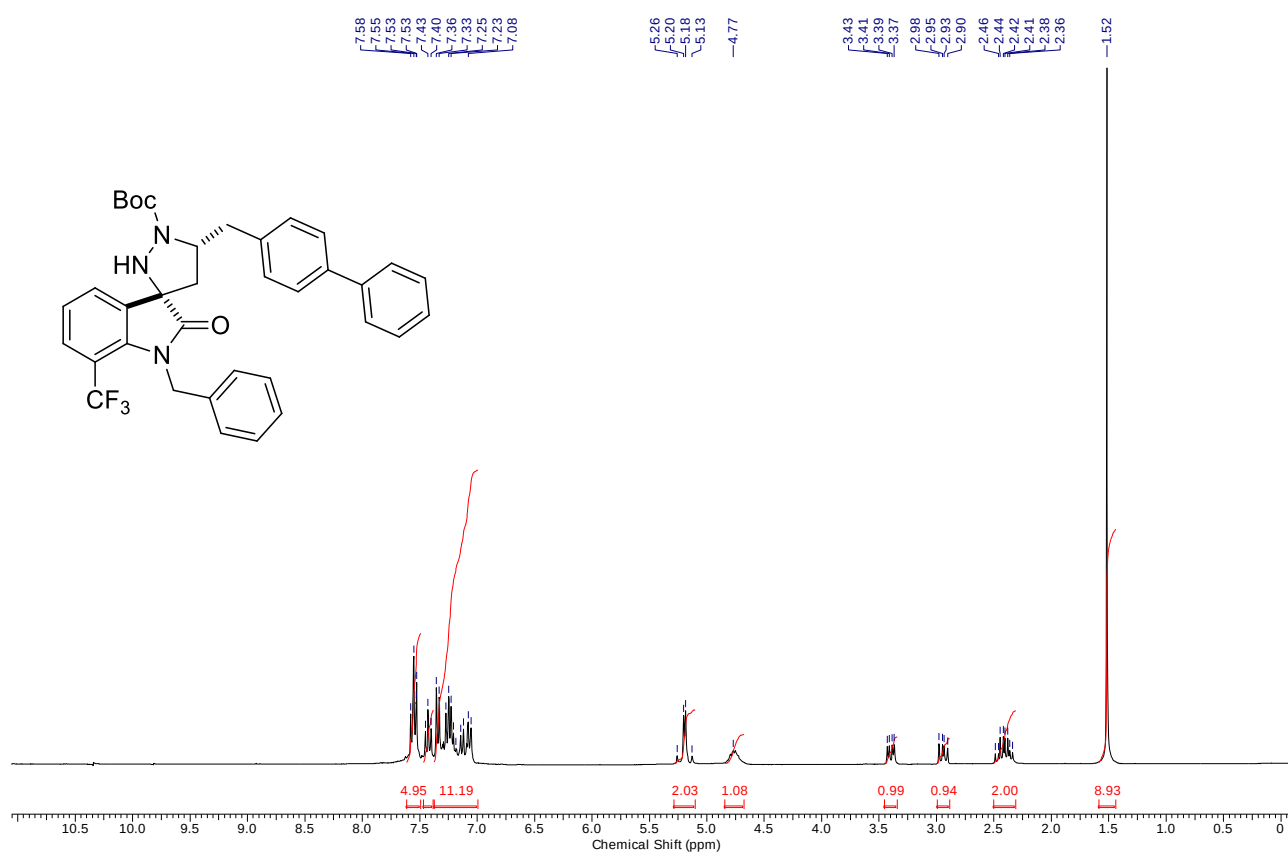
**6n**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



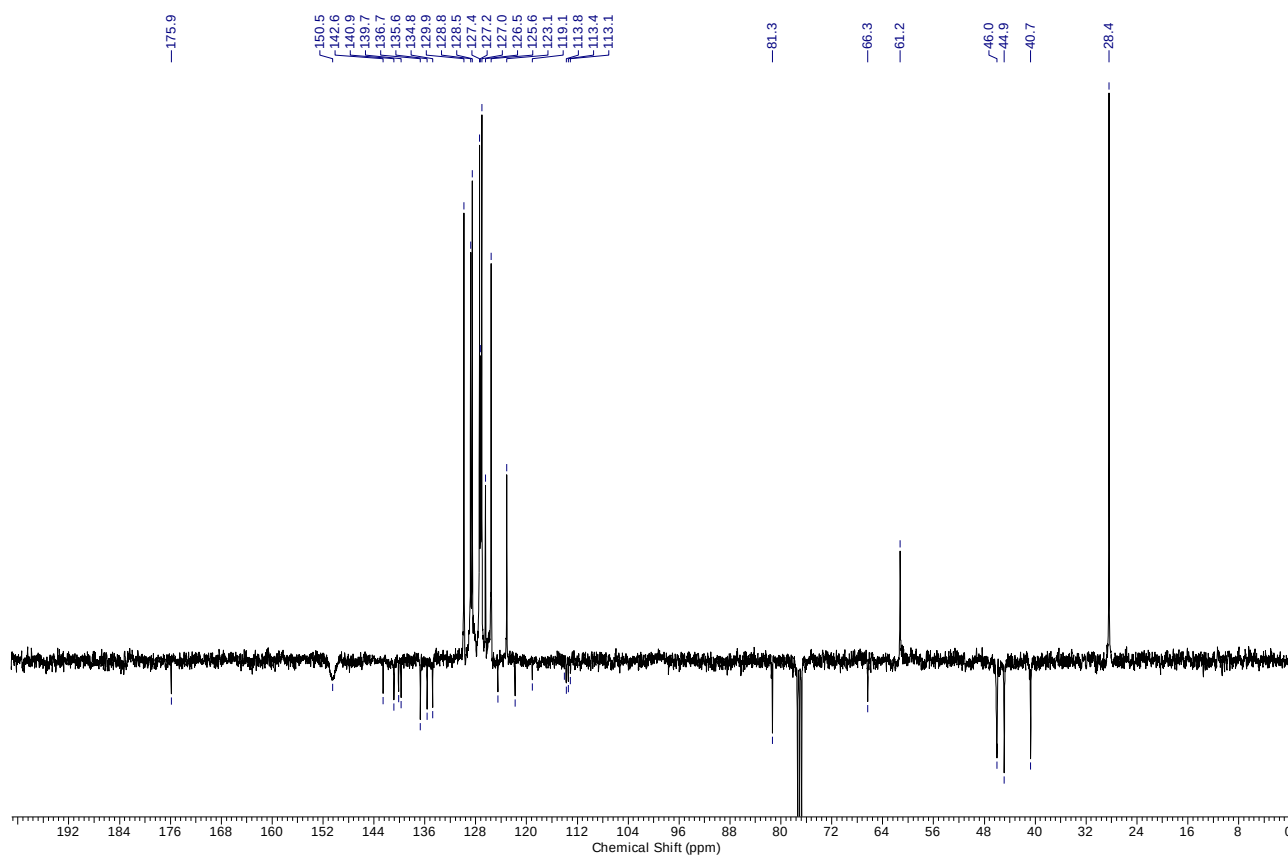
**6n**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



**6n'**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):

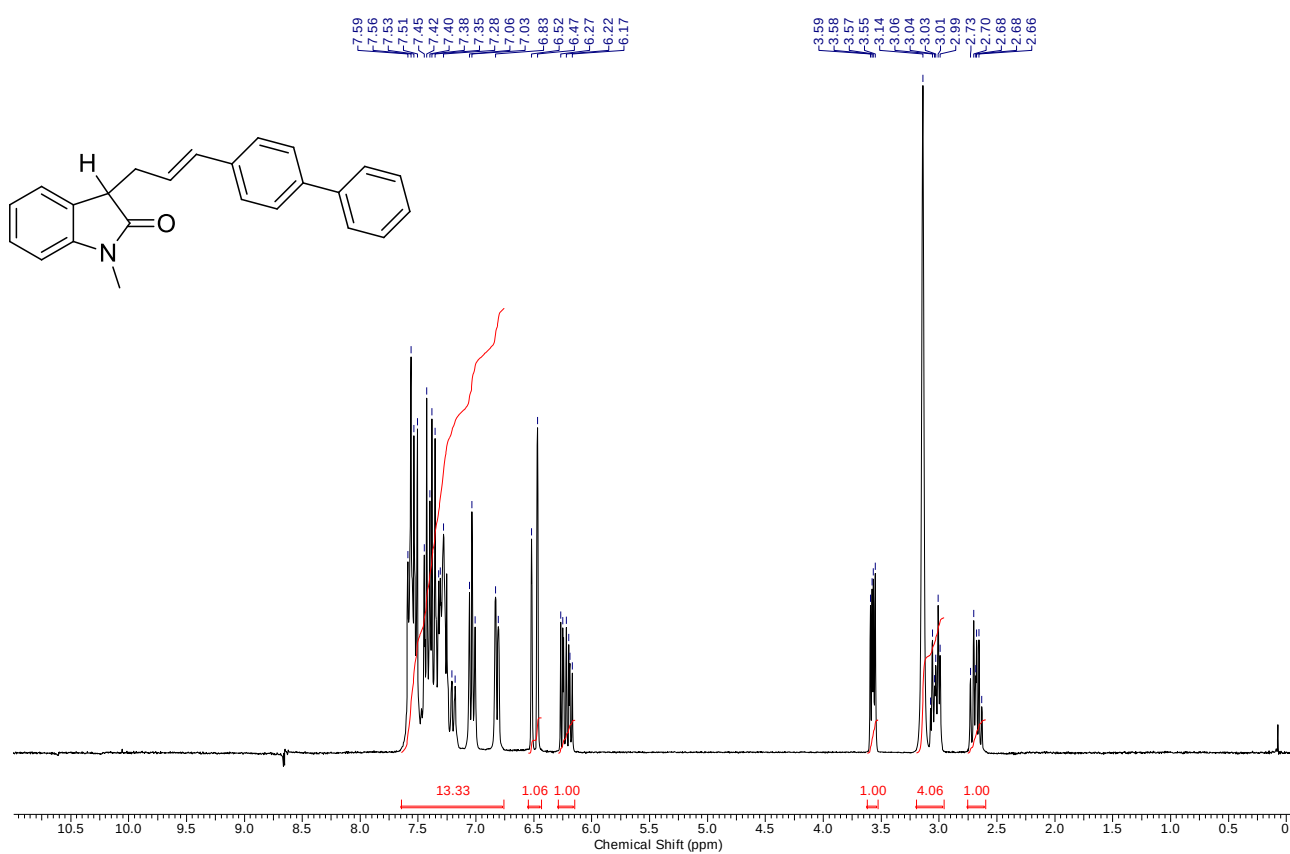


**6n'**  $^{13}\text{C}$  NMR (101 MHz, ATP,  $\text{CDCl}_3$ ):

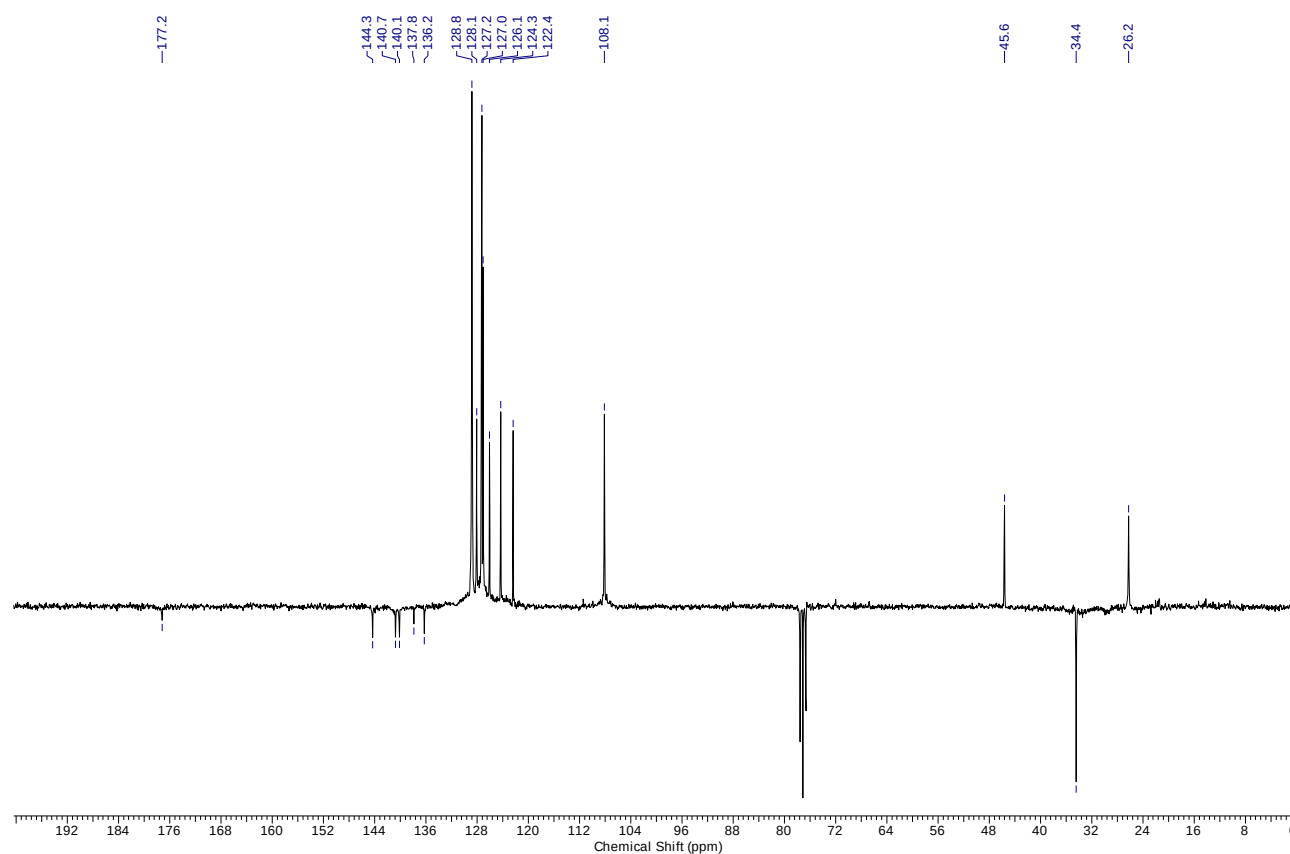




**3b**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



**3b**  $^{13}\text{C}$  NMR (75 MHz, ATP,  $\text{CDCl}_3$ ):



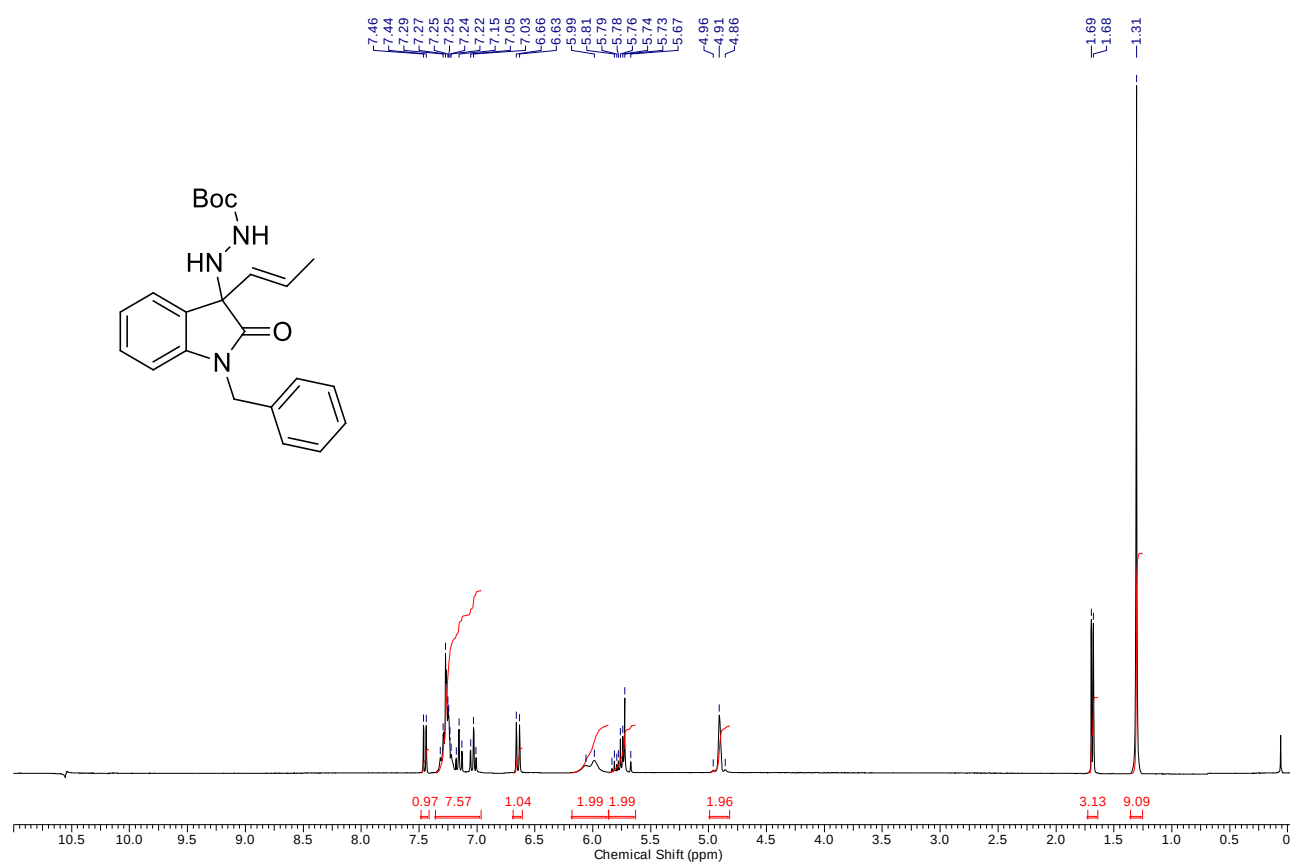
Chemical structure of compound 10 is shown above the spectrum. The spectrum displays peaks corresponding to the structure, with integration values provided below the baseline. The chemical shifts (ppm) are listed at the top of the spectrum.

Chemical Shifts (ppm): 7.57, 7.50, 7.49, 7.47, 7.46, 7.44, 7.31, 7.31, 7.30, 7.29, 7.27, 7.18, 7.03, 6.67, 6.65, 6.48, 6.44, 5.94, 5.92, 5.90, 5.86, 5.12, 5.08, 4.72, 4.68, 2.96, 2.93, 2.92, 2.89, 2.87, 2.85, 2.83, -1.36.

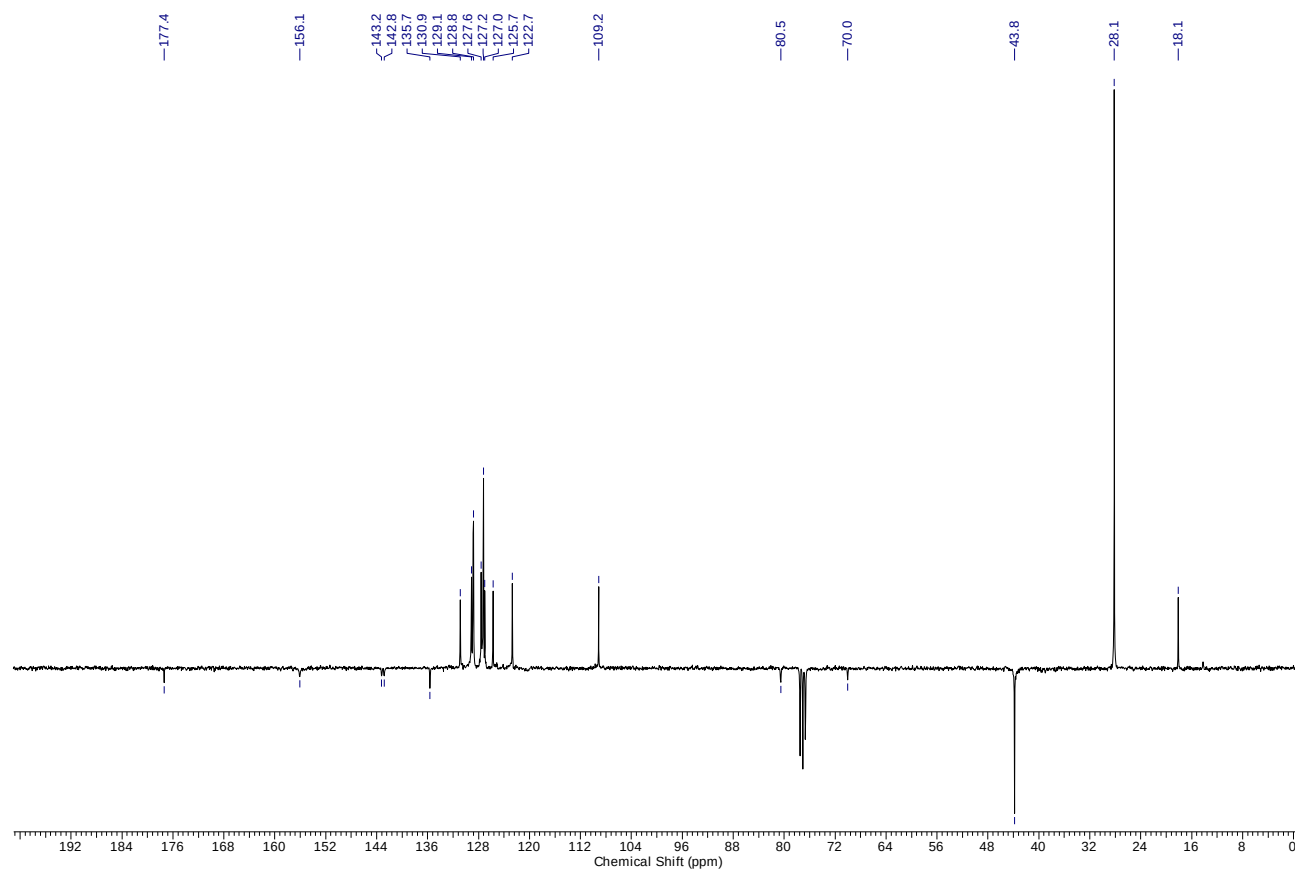
Integration values (from left to right): 17.19, 1.06, 3.00, 0.94, 0.93, 2.07, 9.01.

<sup>13</sup>C NMR spectrum of compound 10a in CDCl<sub>3</sub>. The spectrum shows peaks from 0 to 150 ppm. Key peaks are labeled: 177.4, 156.0, 144.2, 143.1, 141.9, 136.8, 134.5, 128.7, 127.6, 127.5, 127.2, 127.0, 126.8, 125.2, 122.8, 122.1, 109.3, 83.8, 68.5, 43.9, 38.9, and 28.2 ppm. The x-axis is labeled 'Chemical Shift (ppm)' and ranges from 0 to 192.

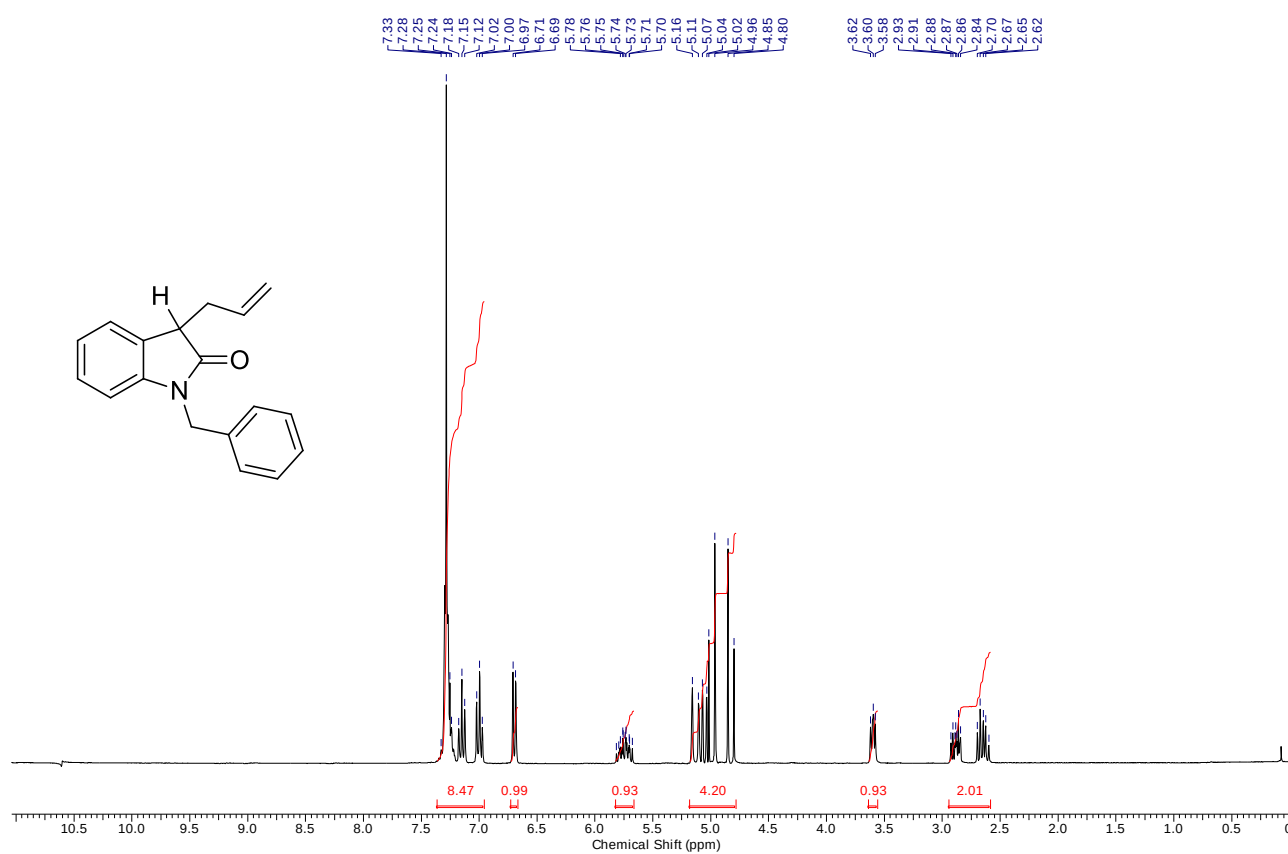
**8**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



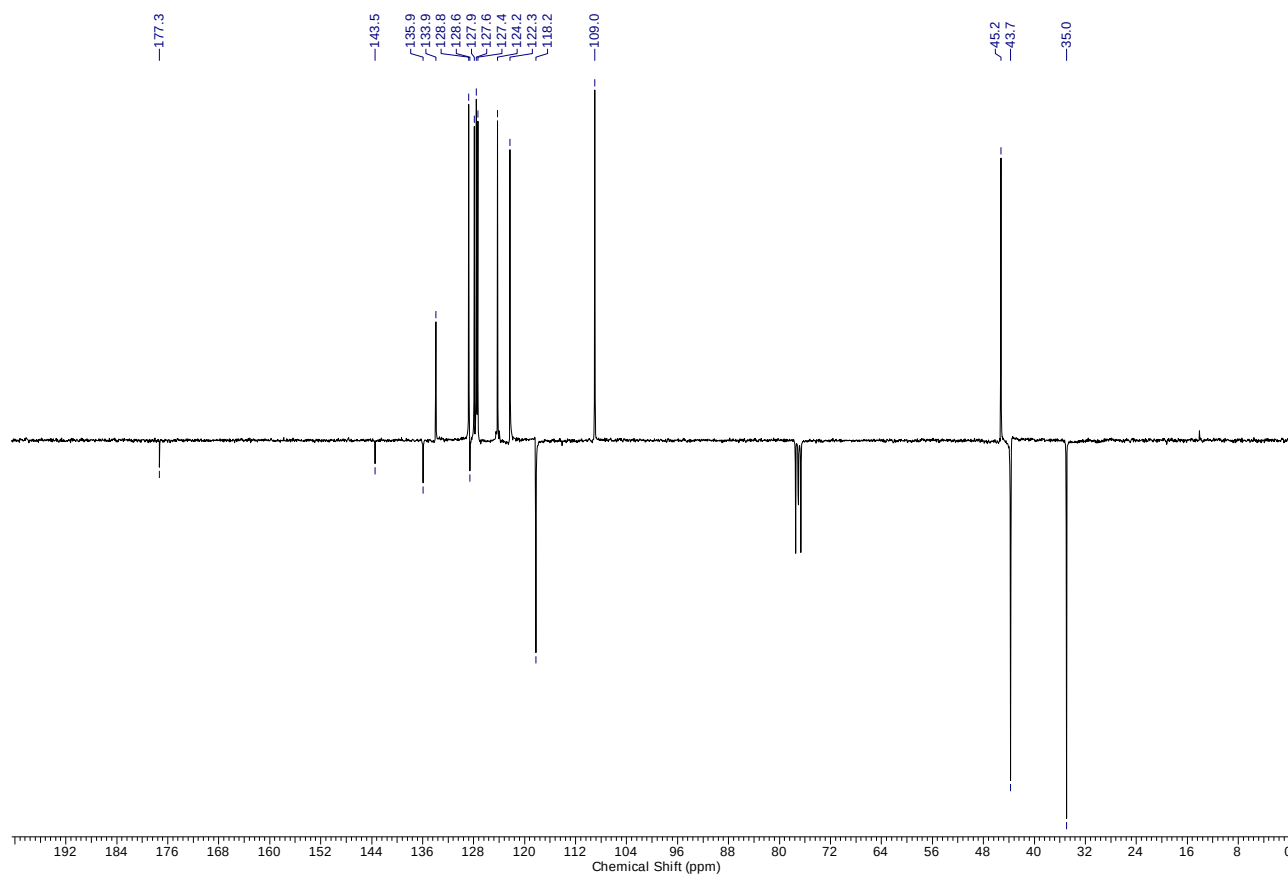
**8**  $^{13}\text{C}$  NMR (75 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



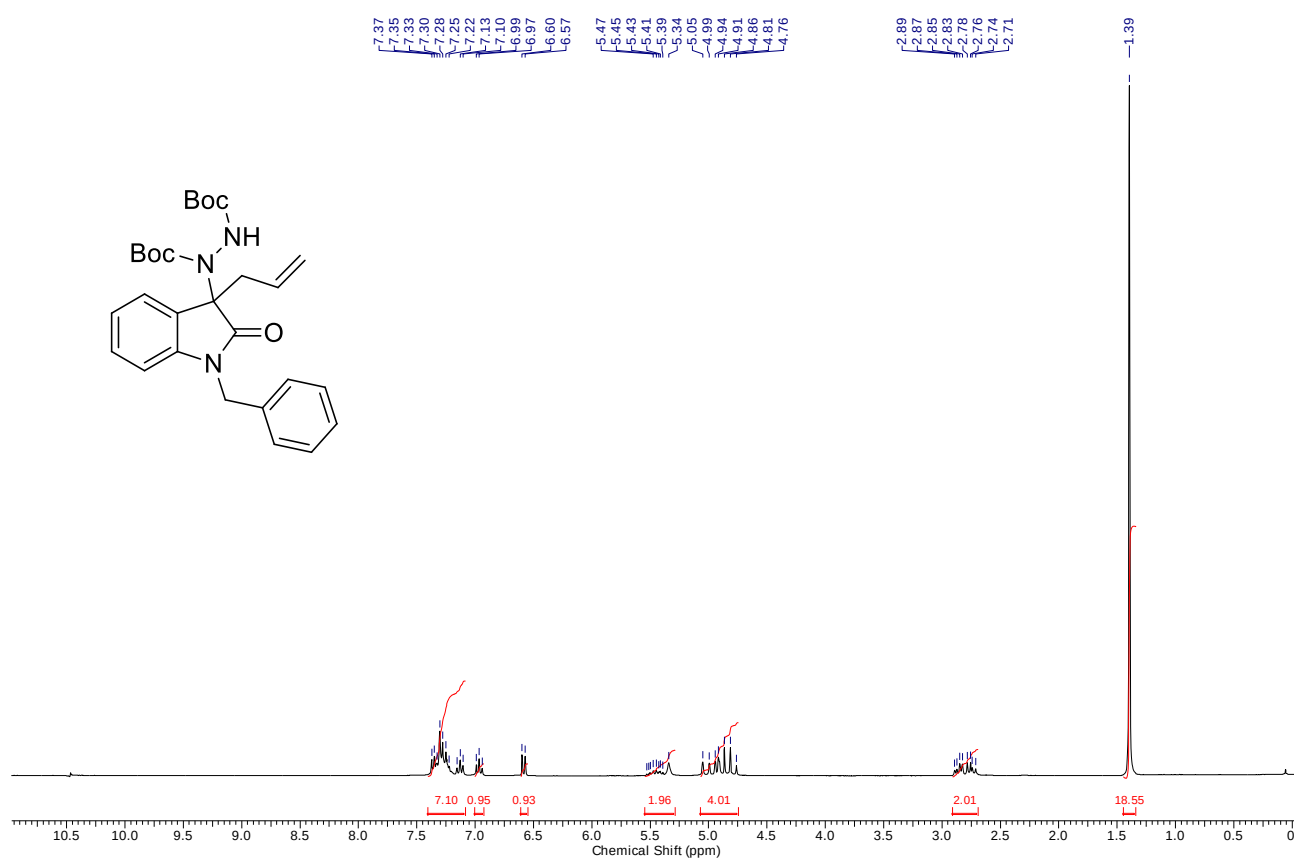
**9**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



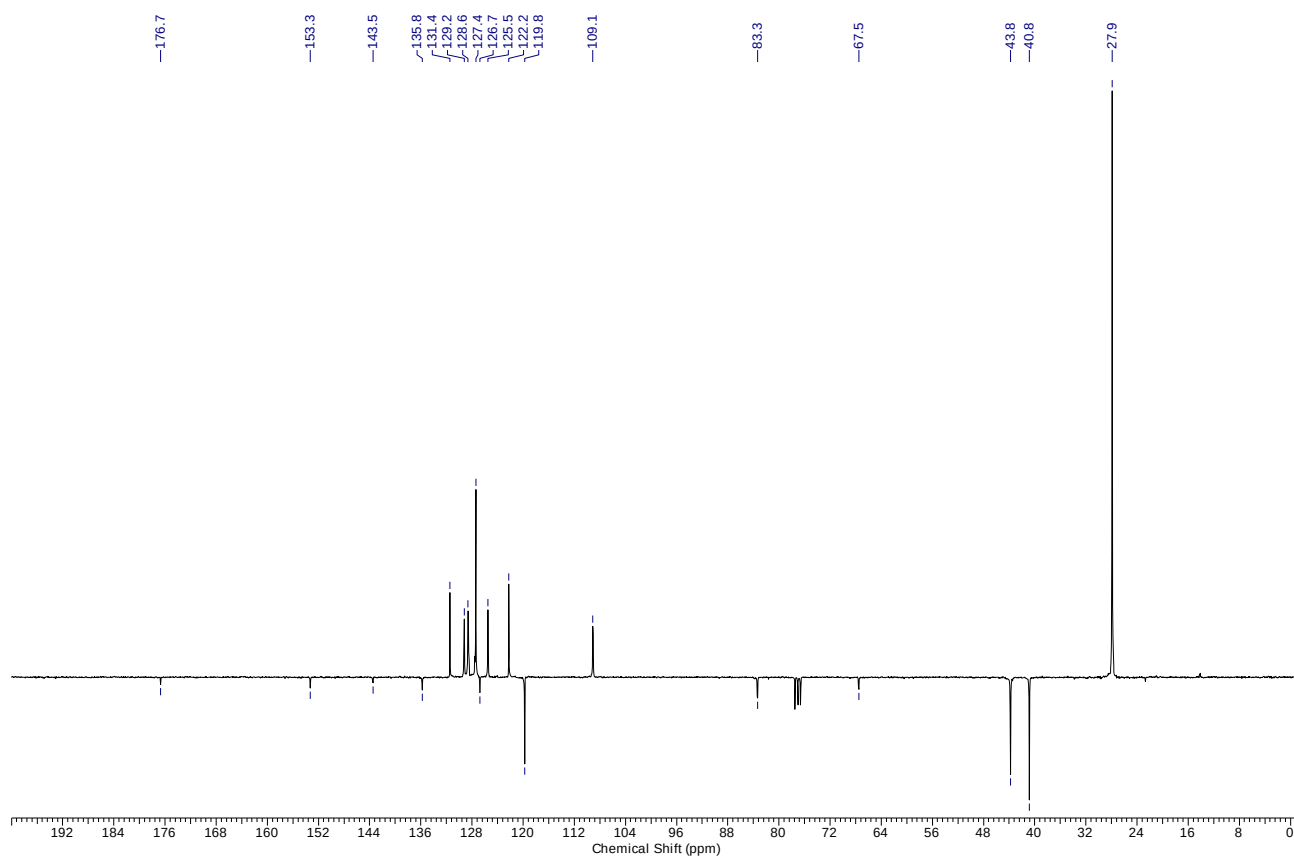
**9**  $^{13}\text{C}$  NMR (75 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



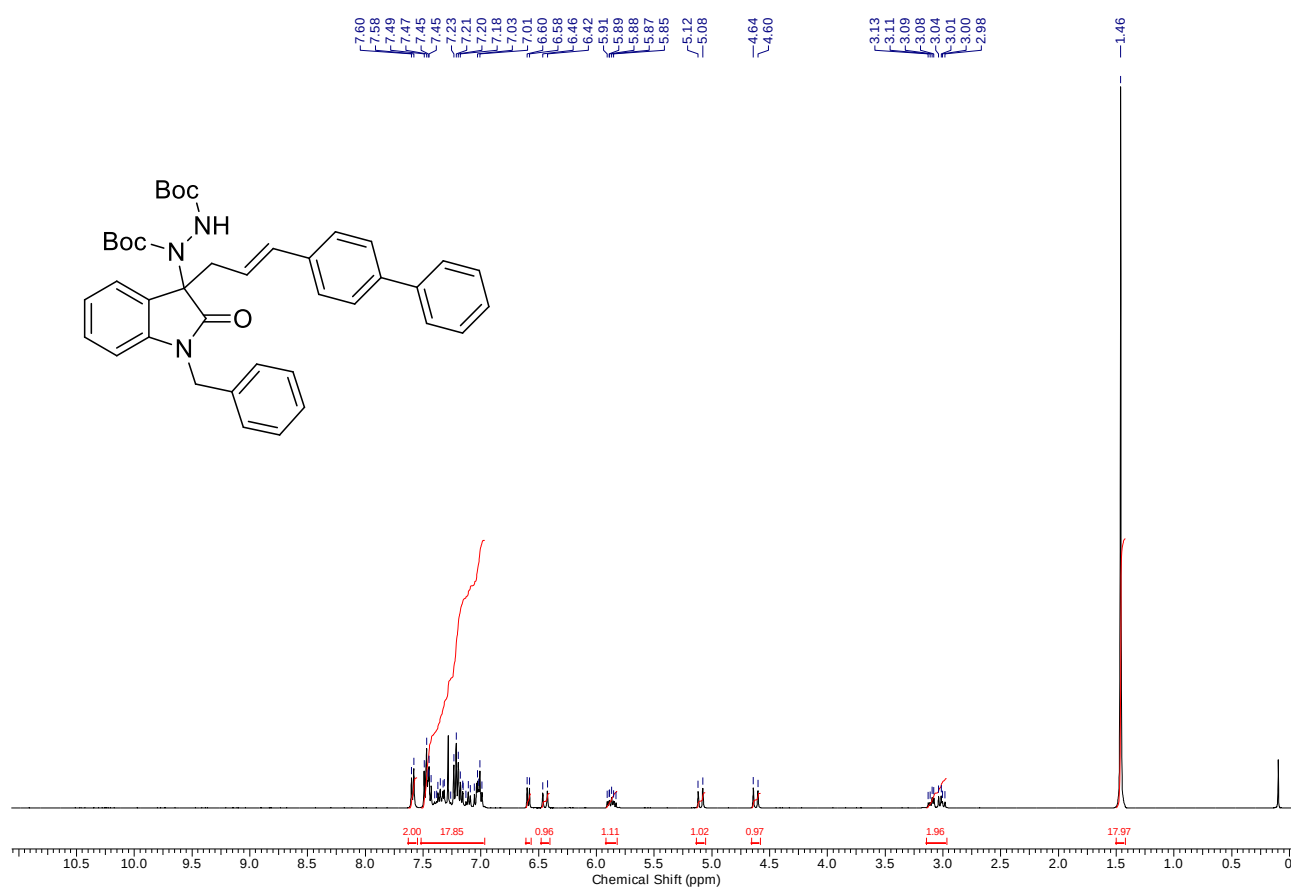
**10**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):



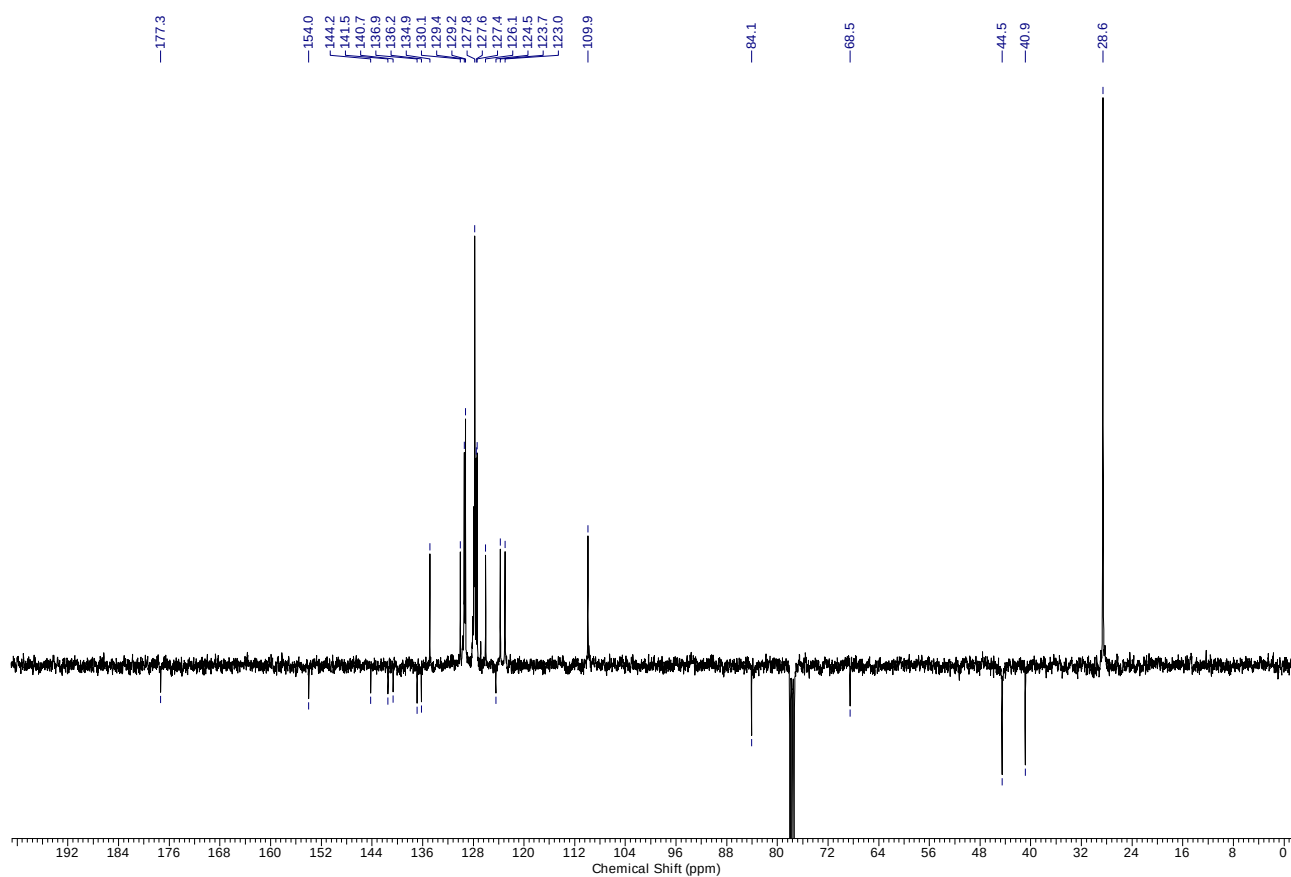
**10**  $^{13}\text{C}$  NMR (75 MHz, ATP,  $\text{CDCl}_3$ ):



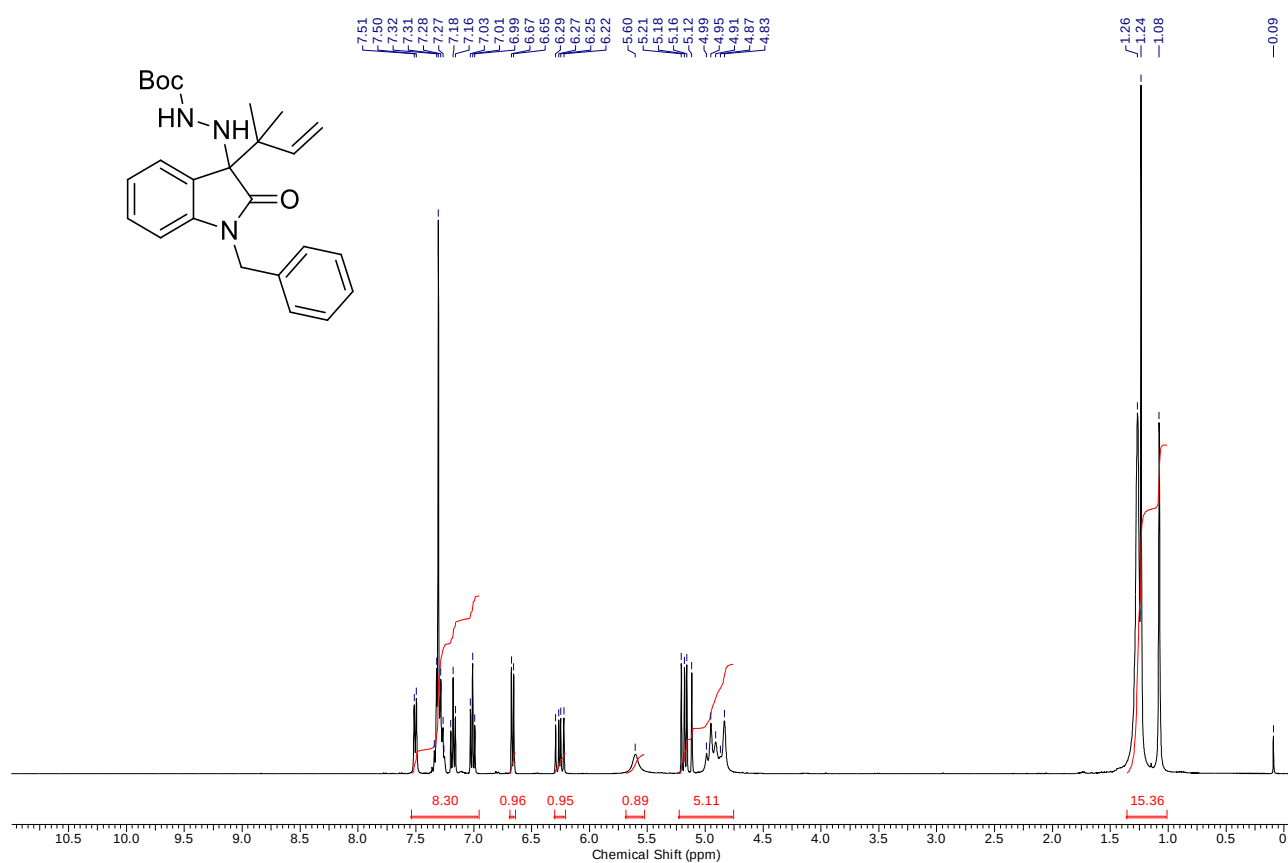
**11**  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):



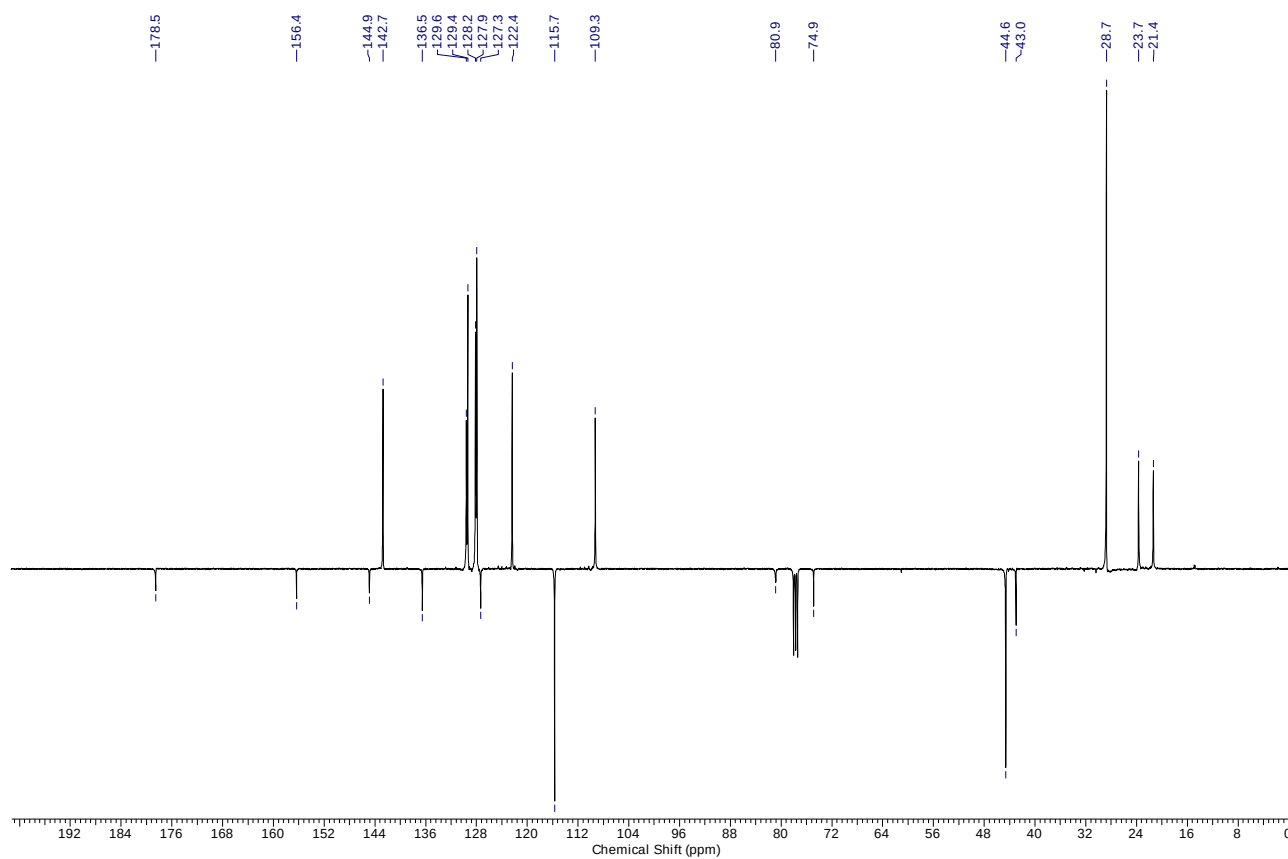
**11**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



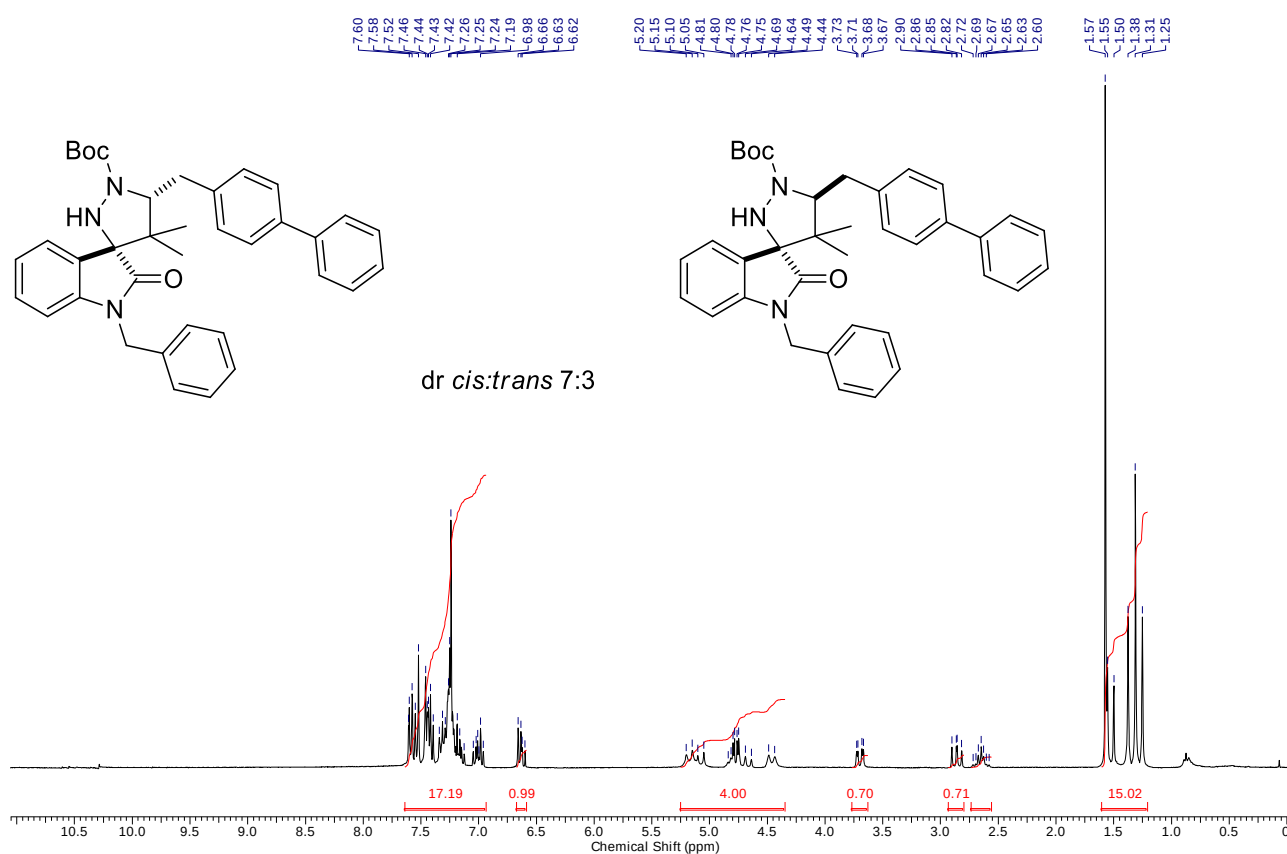
**12**  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):



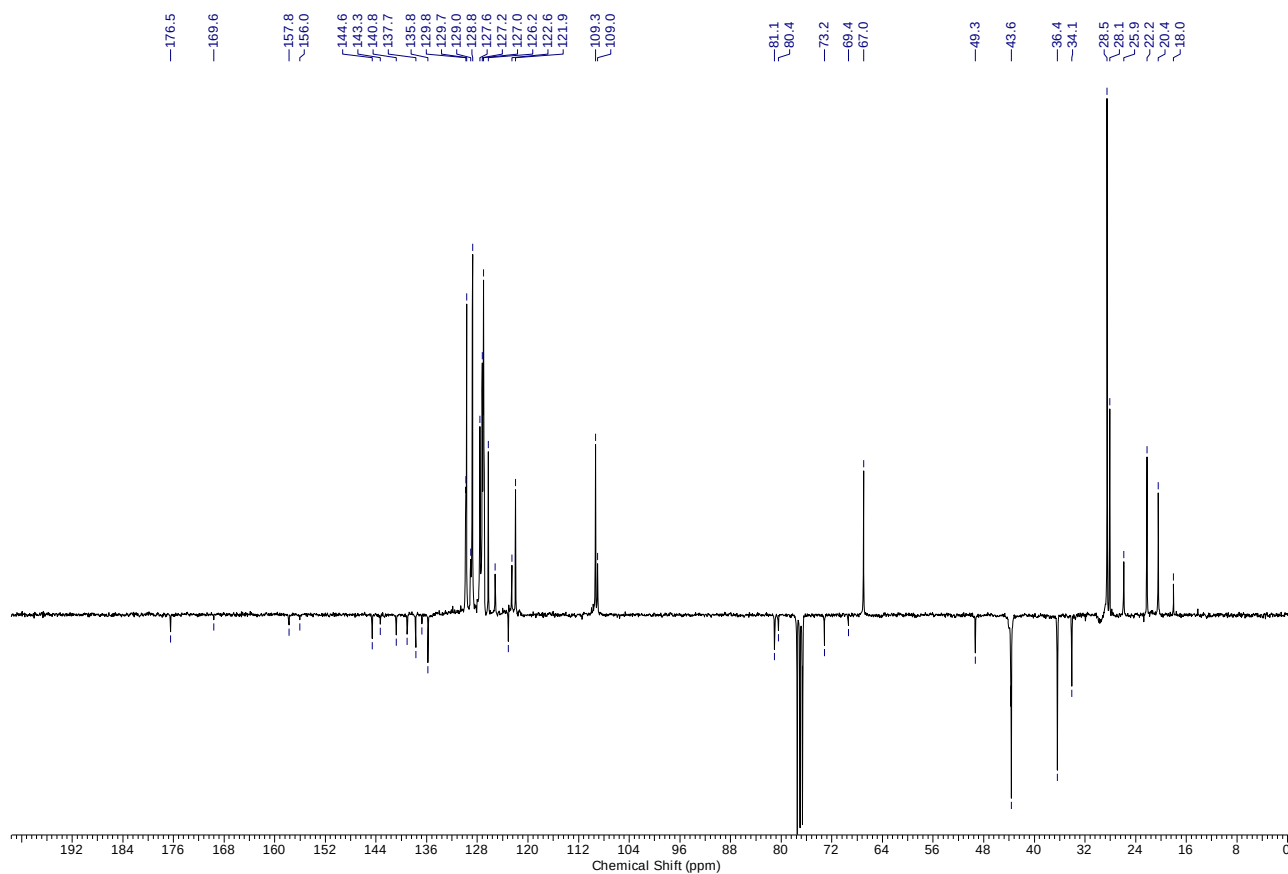
**12**  $^{13}\text{C}$  NMR (101 MHz,  $\text{ATP}$ ,  $\text{CDCl}_3$ ):



**13**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):

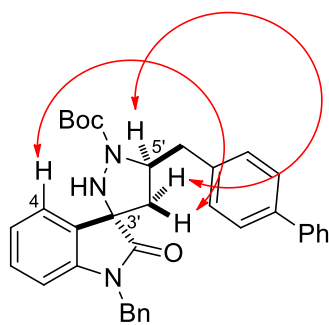
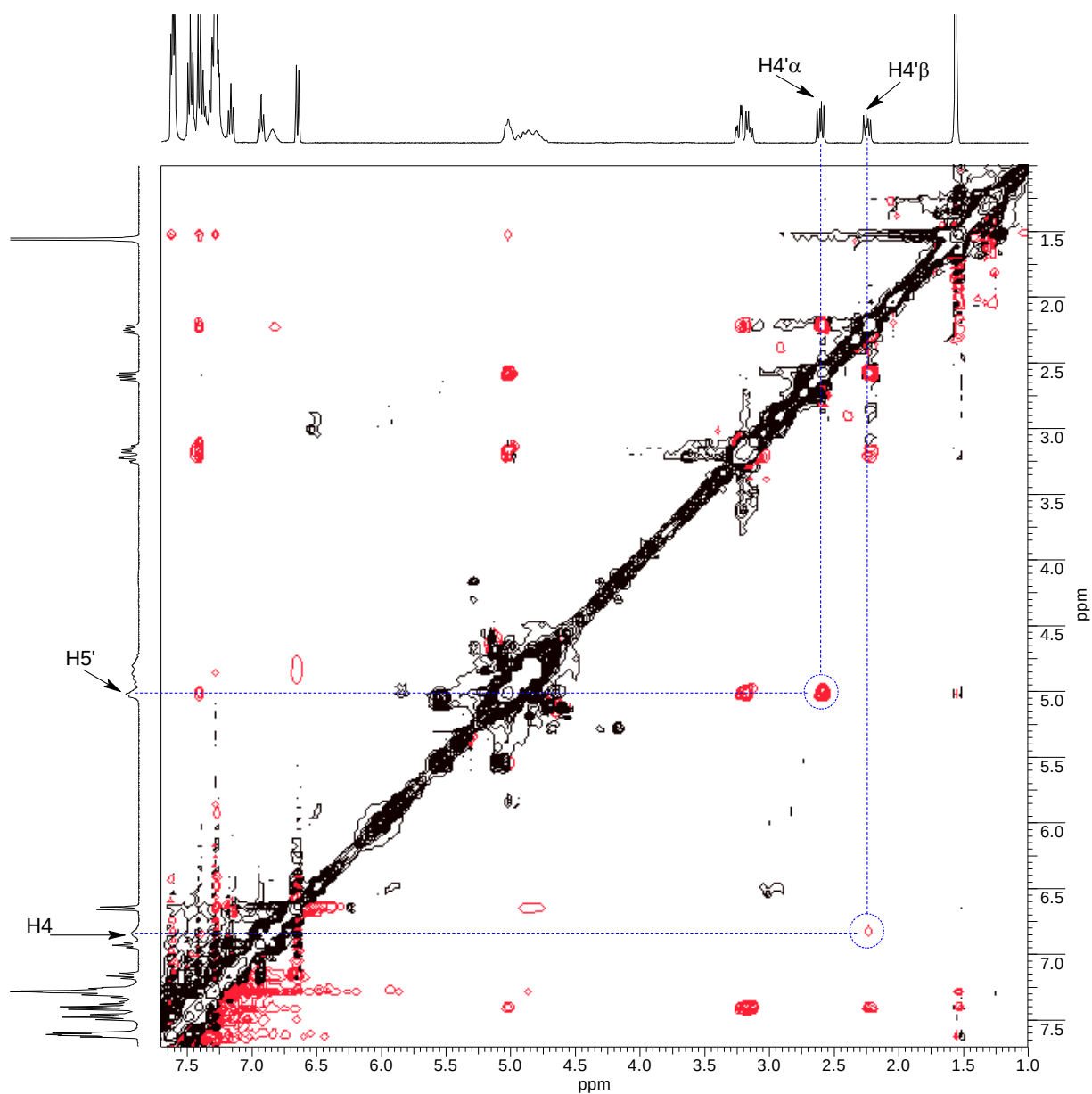


**13**  $^{13}\text{C}$  NMR (75 MHz, ATP,  $\text{CDCl}_3$ ):



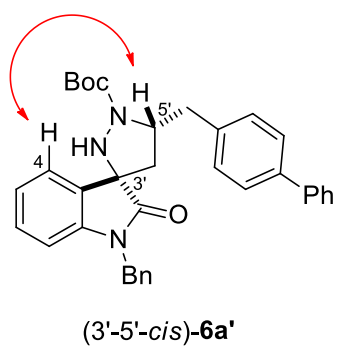
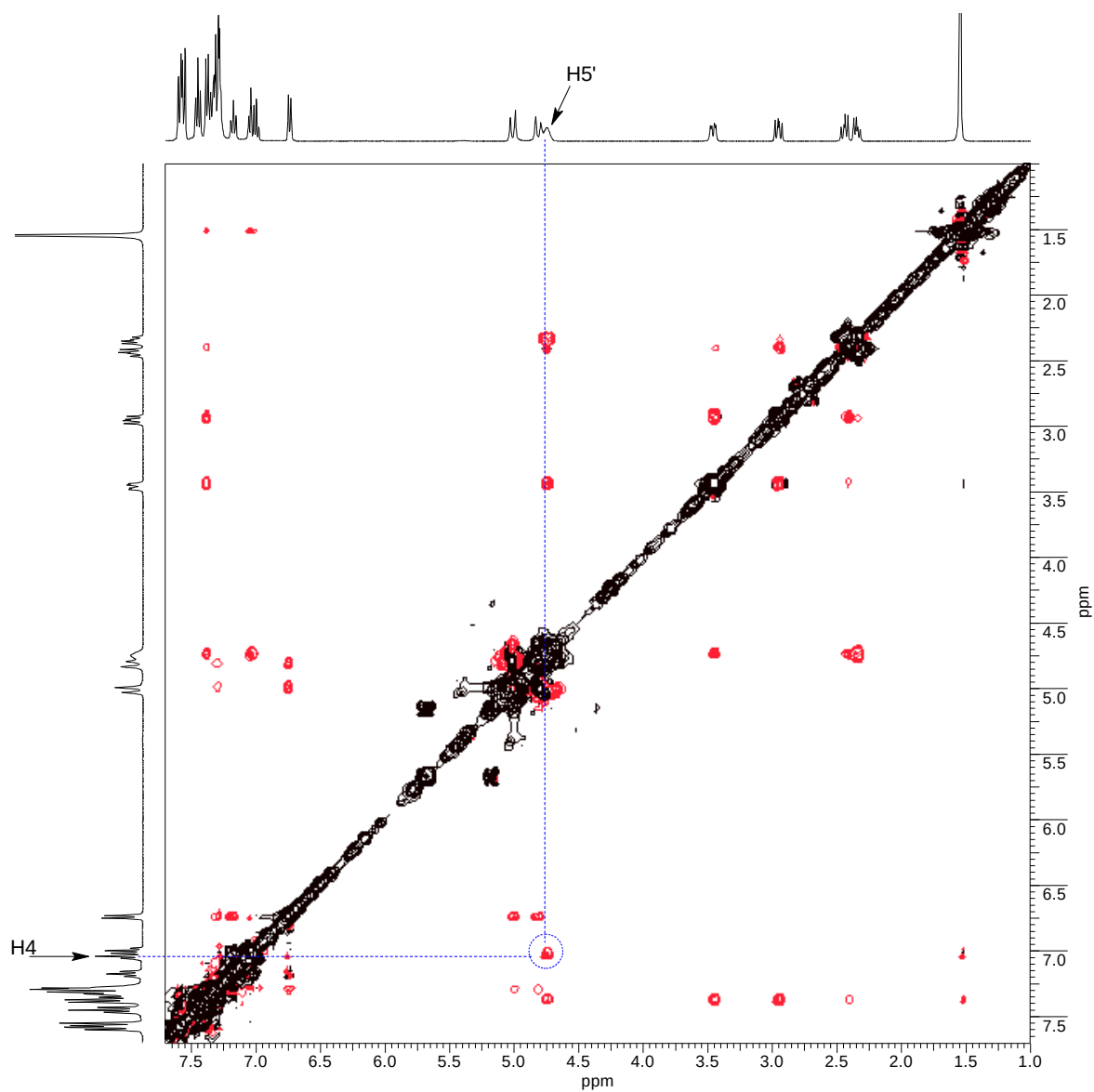


**6a**  $^1\text{H}$ - $^1\text{H}$  ROESY NMR (400 MHz,  $\text{CDCl}_3$ ):



**(3'-5'-trans)-6a**

**6a'**  $^1\text{H}$ - $^1\text{H}$  ROESY NMR (400 MHz,  $\text{CDCl}_3$ ):

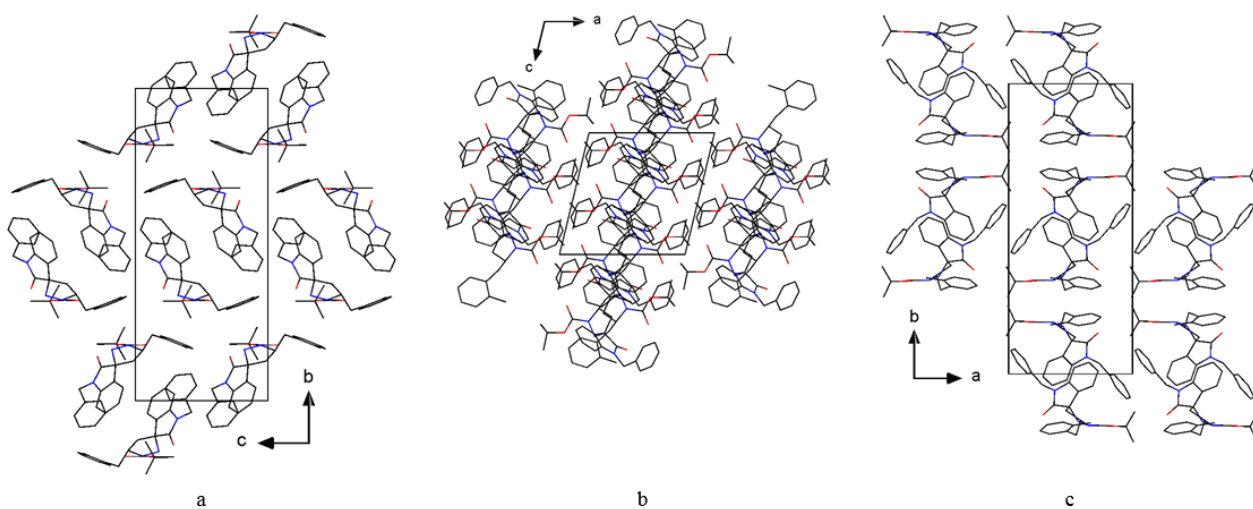


## Single-Crystal X-ray diffraction analysis of the compound **6f**

The specimen selected for the current diffraction experiment was a colourless and transparent prism, with dimensions 0.325 x 0.150 x 0.100 mm, cut from a larger aggregate with a stainless microblade and carefully polished into a drop of perfluorinated oil. The crystal was grown from MeOH by slow evaporation at room temperature (~24 h). The room temperature data collection was performed on a three-circle Bruker AXS Smart diffractometer at a nominal 50 kV x 30 mA power of the X-ray source, using a graphite-monochromatized Mo K $\alpha$  probe ( $\lambda = 0.71073$  Å). A 99.9 % complete full sphere of reflections was collected up to a maximum resolution of  $0.72$  Å $^{-1}$  in  $\sin\theta/\lambda$ , obtaining 39704 unmerged reflections and 6051 independent data (4090 with  $I > 2\sigma(I)$ ). The unit cell was determined by 4872 intense reflections among 5.16 and 40.18 deg of  $2\theta$ , obtaining the following estimates for the lattice parameters:  $a = 10.6314(2)$  Å,  $b = 24.1537(4)$  Å,  $c = 10.5798(2)$  Å,  $\beta = 103.6630(10)$  deg,  $V = 2639.9(1)$  Å $^3$ ,  $\rho = 1.217$  g/cm $^3$ .

Data collection and reduction were carried out with the software SAINT-plus (Bruker (2012). *Saint-plus*. Bruker AXS Inc., Madison, Wisconsin, USA), and raw data were subsequently corrected for beam anisotropy with SADABS (Bruker (2001). *SADABS*. Bruker AXS Inc., Madison, Wisconsin, USA). The structure was solved with direct methods and refined with the Shelx suite of programs (Sheldrick, G. M. (2015). *Acta Cryst.* **C71**, 3-8). The molecular model was developed through an unrestrained non-linear least-squares fit of the measured structure factor amplitudes.

Large librational motion affects rotatable terminal phenyl rings, which lie far from the main molecular inertial axes. As a consequence, the thermal motion of some atoms (C1, C2, C6, C20–C22, see Figure 3 in the main text for the atom numbering) is determined with low accuracy. The quality of the final least-squares estimates for the thermal motion of these atoms can be improved by performing low- $T$  X-ray diffraction experiments. However, these problems did not hamper us from obtaining a fully satisfactory structural determination, securing in particular the relative configuration of the molecular stereocentres (see below).



**Figure S1.** Wires-stick representation of the crystal packing of **6f**, as seen (a) along the  $a$  cell axis; (b) the  $b$  cell axis; (c) the  $c$  cell axis. Hydrogen atoms are omitted for clarity.

The compound **6f** is a racemate and crystallizes in the monoclinic centric space group  $P2_1/c$  ( $n^\circ 14$ ) with one molecule per asymmetric unit. Figure 3 in the main text shows the absolute configuration of

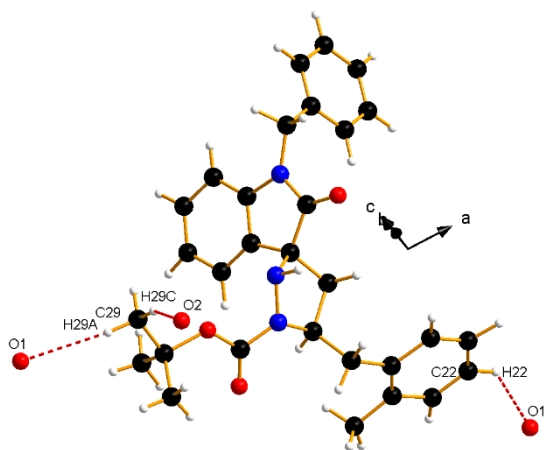
the 2 chiral centres, which were unequivocally determined as 3'S\*,5'R\* [*S*(C9), *R*(C17), Figure 3 in the main text]. Note, however, that also the *R*(C9), *S*(C17) enantiomer is present in an exact 1:1 ratio.

The relatively uncommon N–N covalent bond in the N2–N3–C17–C16–C9 5-membered pyrazolidine ring (Figure 3, main text) has a  $d_{\text{N2–N3}}$  length of 1.424(2) Å. This ring adopts an almost perfect envelope conformation, with puckering amplitude  $Q_2 = 0.360(2)$  Å and phase  $\phi_2 = 152.7(3)$  deg (see Cremer & Pople, J. Am. Chem. Soc. 1975, 97, 1354–1358).

Figure S1 above shows the main packing motifs of **6f**. Overall, the substance adopts a folding that maximizes close packing. However, no strong hydrogen bond donors are present, so only a small number of weak CH $\cdots$ O contacts are set up (Table S1 and Figure S2). Moreover, no direct hydrogen bond interactions are found that involve the secondary amine at N2. This is not unexpected, as NH donors without close possible acceptors are common in organic crystals, especially for molecules with complex foldings. Inversion-related molecules are arranged in a head-to-tail fashion (Figure S1a), allowing the stacking of facing indole-like rings along roughly the direction  $[1/3\ 1/3\ -1]$ , with a  $\sim 5.1$  Å large spacing.

**Table S1.** Geometric parameters for the hydrogen-bonded contacts in **6f** at room temperature. Only contacts with  $d_{\text{H}\cdots\text{A}} \leq 3.0$  and DHA angles in the 120–180° angular range were deemed as significant. Values in Å / deg, with standard deviations reported in parentheses. See Figure 4 for the atom labels.

| C–H $\cdots$ O       | D–H     | H $\cdots$ A | D $\cdots$ A | D–H $\cdots$ A | Symmetry operation                |
|----------------------|---------|--------------|--------------|----------------|-----------------------------------|
| C29–H29A $\cdots$ O1 | 1.00(2) | 2.77(2)      | 3.699(4)     | 155(1)         | $-1+x, y, z$                      |
| C29–H29C $\cdots$ O2 | 1.00(1) | 2.78(2)      | 3.738(4)     | 160(1)         | $x, \frac{1}{2}-y, \frac{1}{2}+z$ |
| C22–H22 $\cdots$ O1  | 1.09(2) | 2.69(2)      | 3.581(4)     | 138(1)         | $x, y, -1+z$                      |



**Figure S2.** Interaction geometry of the **6f** asymmetric unit. See Table S1 for the geometrical parameters of hydrogen-bonded contacts.

CCDC 1938731 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via [www.ccdc.cam.ac.uk/structures](http://www.ccdc.cam.ac.uk/structures).