Supplementary Information

Extracellular electron transfer mediated by a cytocompatible redox polymer to study the crosstalk among the mammalian circadian clock, cellular metabolism, and cellular redox states

Masahito Ishikawa,^{1,4}* Kazuki Kawai¹, Masahiro Kaneko,² Kenya Tanaka,³ Shuji Nakanishi,^{3,4} and Katsutoshi Hori¹*

¹Department of Biomolecular Engineering, Graduate School of Engineering, Nagoya University, Furo-cho, Chikusa-ku, Nagoya 464-8603, Japan
²Department of Materials Engineering, School of Engineering, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8656, Japan
³Graduate School of Engineering Science, Osaka University, 1-3 Machikaneyama, Toyonaka, Osaka 560-8631, Japan
⁴Research Center for Solar Energy Chemistry, Osaka University, 1-3 Machikaneyama, Toyonaka, Osaka 560-8631, Japan

Supplementary figures

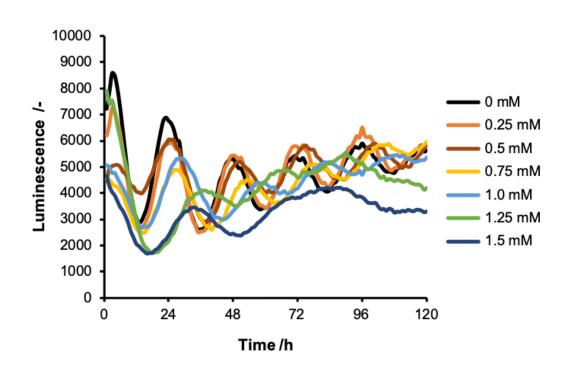


Figure S1. Representative curves of raw bioluminescence rhythm in U2OS cells in the presence of oxidized pMFc. Each concentration of pMFc corresponds to that of the vinyl ferrocene unit.

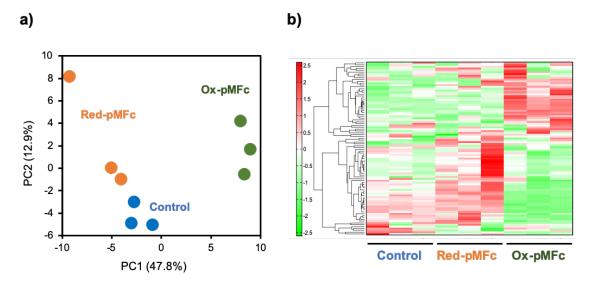
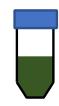
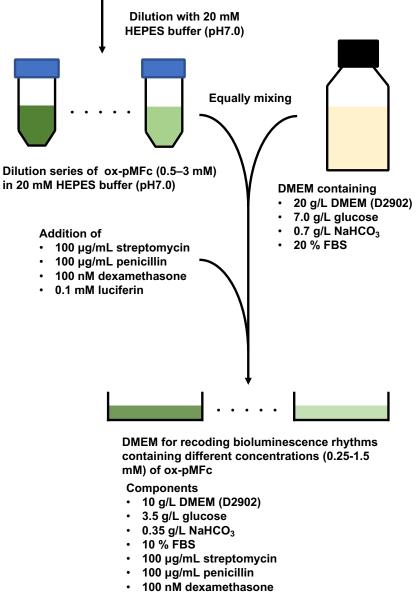


Figure S2. a) Principal component analysis and b) hierarchical cluster analysis of 116 metabolites extracted from U2OS cells treated with 1 mM of the oxidized form of pMFc (ox-pMFc) or 1 mM of the reduced form of pMFc (red-pMFc) and from untreated U2OS cells (control). The concentration of pMFc corresponds to that of vinyl ferrocene unit.



4 mM ox-pMFc in 20 mM HEPES buffer (pH7.0) sterilized by filtration.



0.1 mM luciferin

Figure S3. Schematic of the preparation of DMEM for recoding bioluminescence

rhythm containing the different concentrations of ox-pMFc.

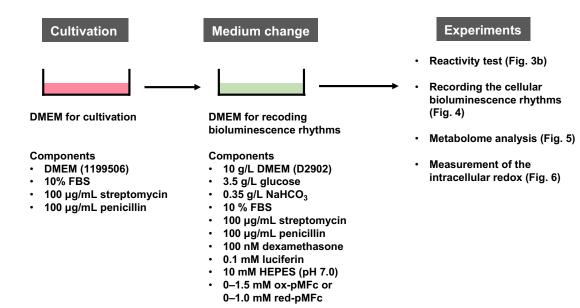


Figure S4. Schematic of the procedure for experiments using U2OS cells and pMFcs.