

Supporting Information

Tuning catalysis of boronic acids in microgels by in situ reversibly structural variations

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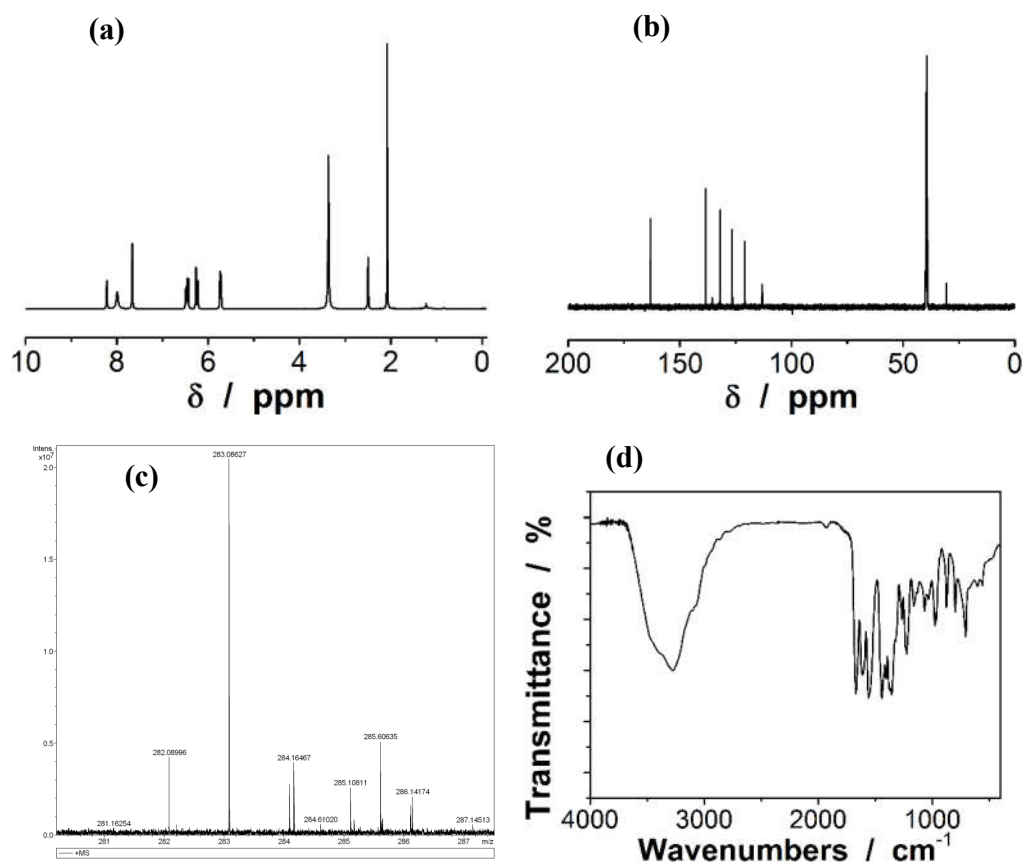


Fig. S1. (a) ¹H NMR (in DMSO-d₆), (b) ¹³C NMR (in DMSO-d₆), (c) FT-MS, and (d) IR spectra of **1**. ¹H NMR: δ = 10.10 (2H, NH), 8.22 (1H, ArH), 7.99 (2H, B(OH)₂), 7.67 (2H, ArH), 6.47

(2H, $CH=CH_2$), 6.25 (2H, $CH=CH_2$), 5.73 (2H, $CH=CH_2$); ^{13}C NMR: $\delta = 113.14, 120.90, 126.53, 132.05, 135.51, 138.42, 163.10$; FT-MS: m/z 283.09 [$M+Na^+$].

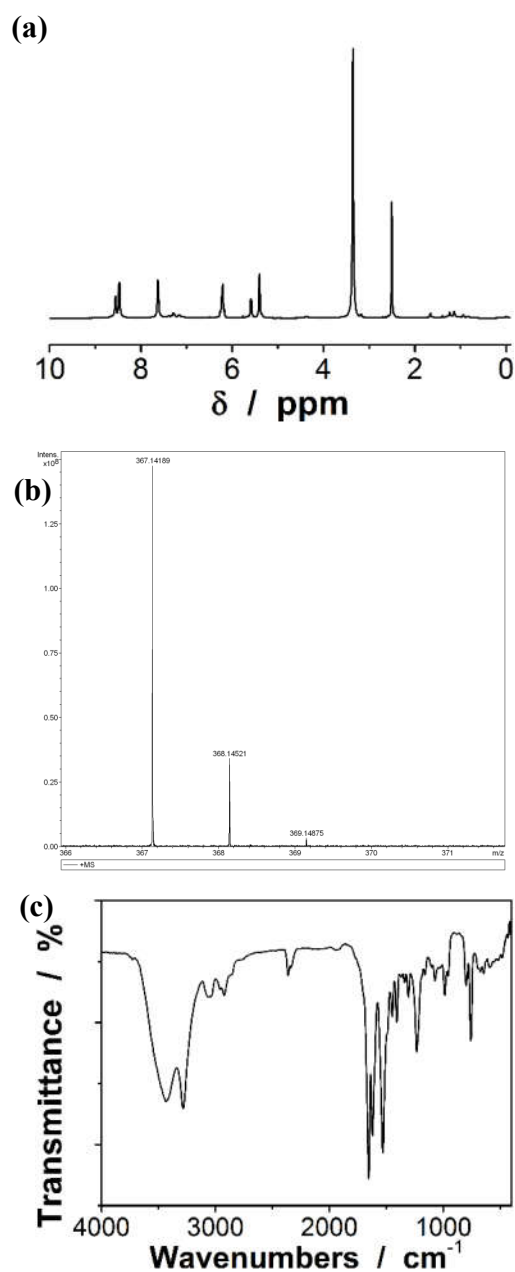


Fig. S2. (a) 1H NMR (in DMSO- d_6), (b) FT-MS, and (c) IR spectra of **2**. 1H NMR: $\delta = 8.56$ (2H, NH), 8.48 (4H, ArH), 7.63 (2H, ArH), 6.21 (4H, $ArCH_2$), 5.59 (2H, $CH=CH_2$), 5.40 (4H, $CH=CH_2$); FT-MS: m/z 367.14 [$M+Na^+$].

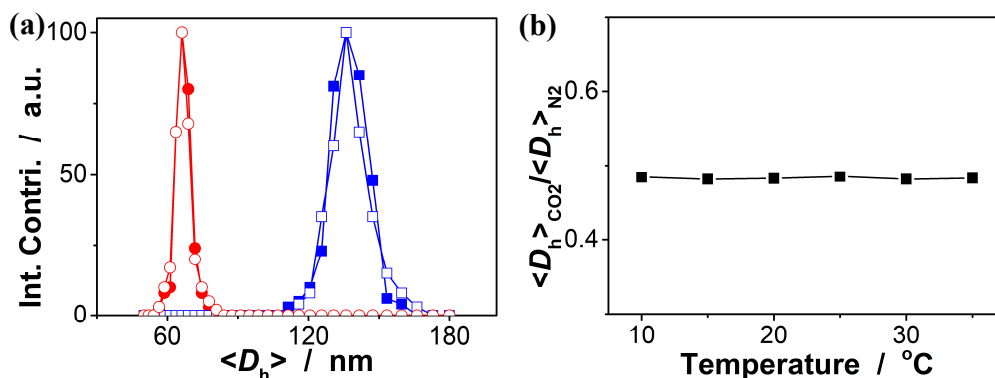


Fig. S3. (a) DLS size distribution of the PBA@PM in $\text{CH}_2\text{Cl}_2/\text{water}$ (9:1 in volume) upon heating (to 35.0 °C; ■,●) or cooling (to 10.0 °C; □,○) and bubbling with N_2 (■,□) or CO_2 (●,○) gas. (b) Influence of the solution temperature on the $\langle D_h \rangle_{\text{CO}_2} / \langle D_h \rangle_{\text{N}_2}$, where the $\langle D_h \rangle_{\text{CO}_2}$ and the $\langle D_h \rangle_{\text{N}_2}$ were measured on the same microgels at the same temperature upon bubbling with N_2 or CO_2 gas correspondingly.

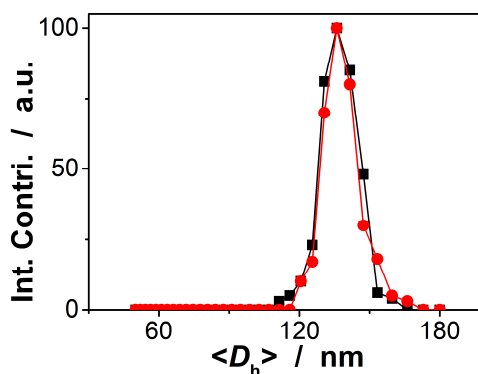


Fig. S4. DLS size distribution of the PBA@PM (■) in $\text{CH}_2\text{Cl}_2/\text{water}$ (9:1 in volume) before and (●) after 6 months' storage under the N_2 atmosphere at 10.0 °C.

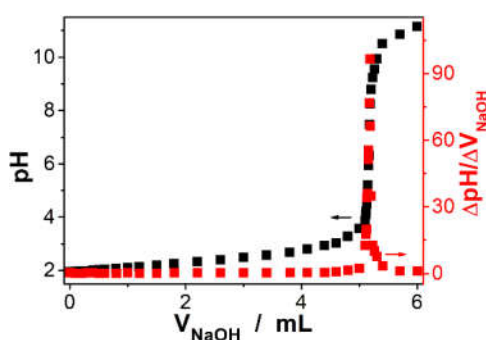


Fig. S5. Titration curve of the PBA@PM using 10.0 mM NaOH solution. All measurements were made at 10.0 °C.

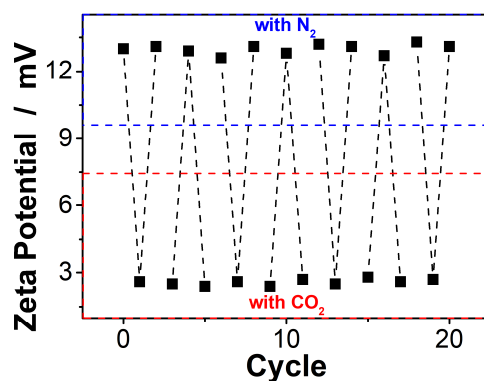


Fig. S6. The zeta potential of the PBA@PM dispersed in CH₂Cl₂/water (9:1 in volume) upon bubbling with N₂ or CO₂ gas.

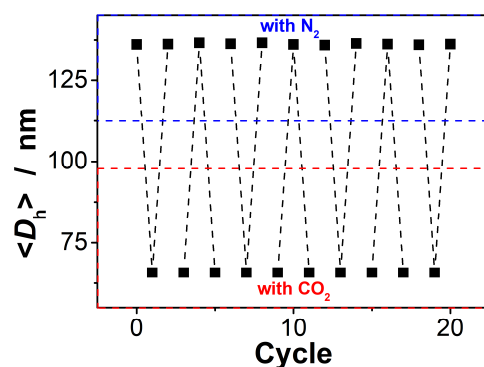


Fig. S7. The $\langle D_h \rangle$ of the PBA@PM dispersed in CH₂Cl₂/water (9:1 in volume) upon bubbling with N₂ or CO₂ gas.

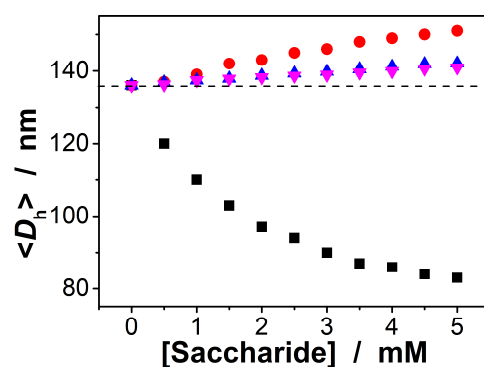


Fig. S8. [Saccharide]-dependent $\langle D_h \rangle$ of the PBA@PM in CH₂Cl₂/water (9:1 in volume) under the N₂ atmosphere at 10.0 °C, upon adding glucose (■), fructose (●), galactose (▲) or mannose (▼).

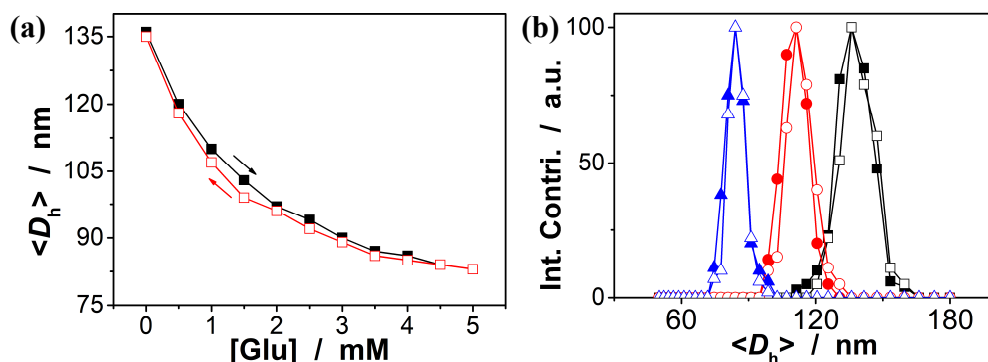


Fig. S9. (a) [Glu]-dependent $\langle D_h \rangle$ of the PBA@PM dispersed in $\text{CH}_2\text{Cl}_2/\text{water}$ (9:1 in volume) under the N_2 atmosphere at $10.0\text{ }^\circ\text{C}$. (b) DLS size distribution of the PBA@PM before (solid symbols) and after (open symbols) five cycles of adding ([Glu] = 1.0 mM: \bullet, \circ ; [Glu] = 5.0 mM: $\blacktriangle, \triangle$) and removing ([Glu] = 0.0 mM: \blacksquare, \square) glucose by dialysis.

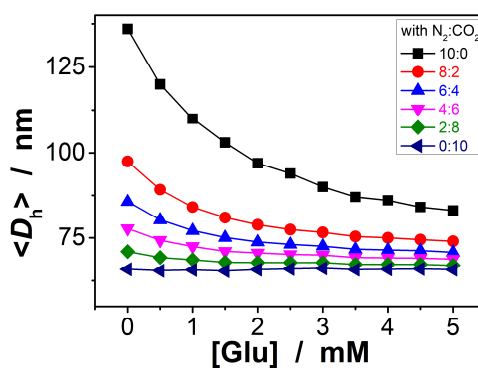


Fig. S10. Glucose-dependent $\langle D_h \rangle$ of the PBA@PM dispersed in $\text{CH}_2\text{Cl}_2/\text{water}$ (9:1 in volume) upon bubbling with gases containing appropriate amount (in volume) of CO_2 gas, measured at $10.0\text{ }^\circ\text{C}$ and at a scattering angle of 45° .

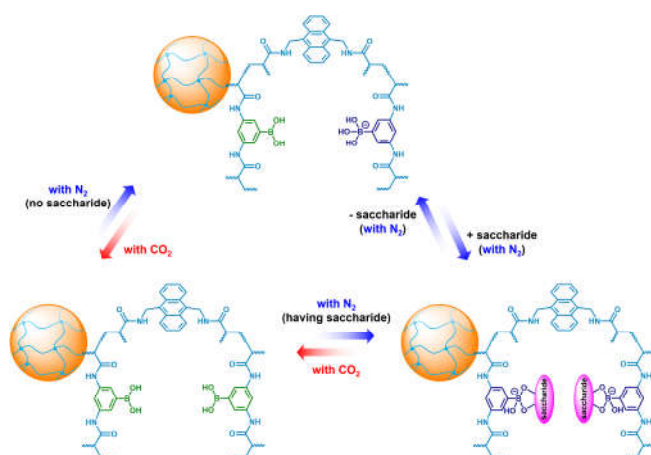


Fig. S11. A possible mechanism on structural modifications in the PBA@PM upon bubbling with N_2/CO_2 gases, in the presence or absence of fructose, galactose or mannose.

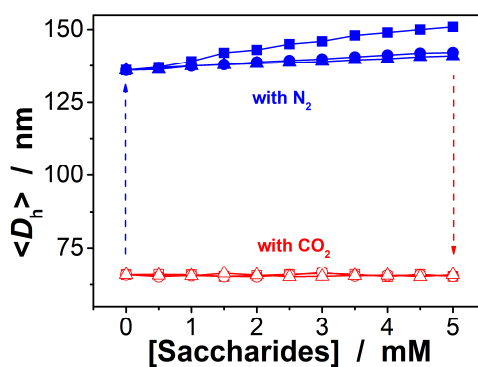


Fig. S12. [Saccharides]-dependent $\langle D_h \rangle$ of the PBA@PM in CH₂Cl₂/water (9:1 in volume) at 10.0 °C, upon adding fructose (■,□), galactose (●,○) or mannose (▲,△) and bubbling with N₂ or CO₂ gas.

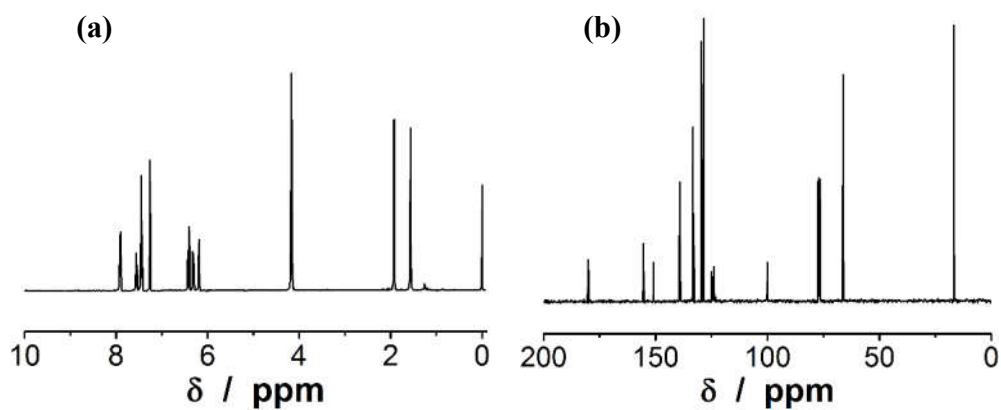


Fig. S13. (a) ¹H NMR (in CDCl₃) and (b) ¹³C NMR (in CDCl₃) spectra of **3**. ¹H NMR: δ = 7.91 (2H), 7.56 (1H), 7.44 (2H), 6.42 (1H), 6.31 (1H), 6.19 (1H), 4.17 (4H), 1.92 (3H); ¹³C NMR: δ = 180.3, 155.6, 150.9, 139.4, 133.4, 133.1, 129.6, 128.7, 125.0, 124.1, 100.2, 66.4, 16.9.

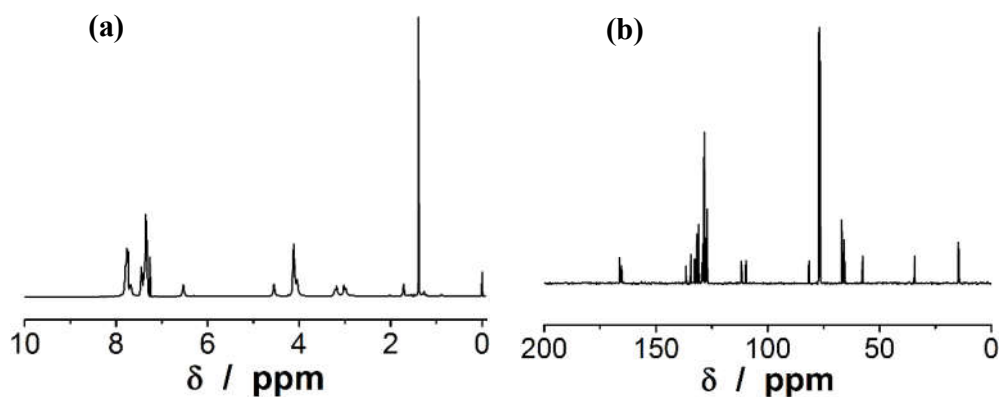


Fig. S14. (a) ^1H NMR (in CDCl_3) and (b) ^{13}C NMR (in CDCl_3) spectra of product **6**. ^1H NMR: δ = 7.80-7.74 (4H), 7.66 (1H), 7.48-7.30 (6H), 6.52 (1H), 4.55 (1H), 4.13-4.03 (4H), 3.20 (1H), 3.01 (1H), 1.38 (3H); ^{13}C NMR: δ = 166.2, 165.6, 136.5, 134.6, 132.8, 131.8, 130.8, 129.2, 128.3, 127.7, 127.1, 112.1, 109.7, 81.6, 66.8, 65.7, 57.7, 34.5, 14.6.

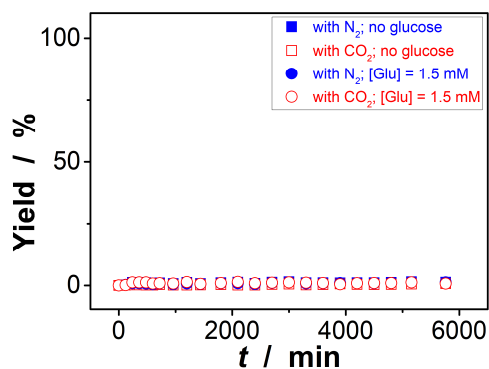


Fig. S15. Time trace of the yield of **6** of the reaction at 10.0 $^\circ\text{C}$, without any organoboron acid catalysts.

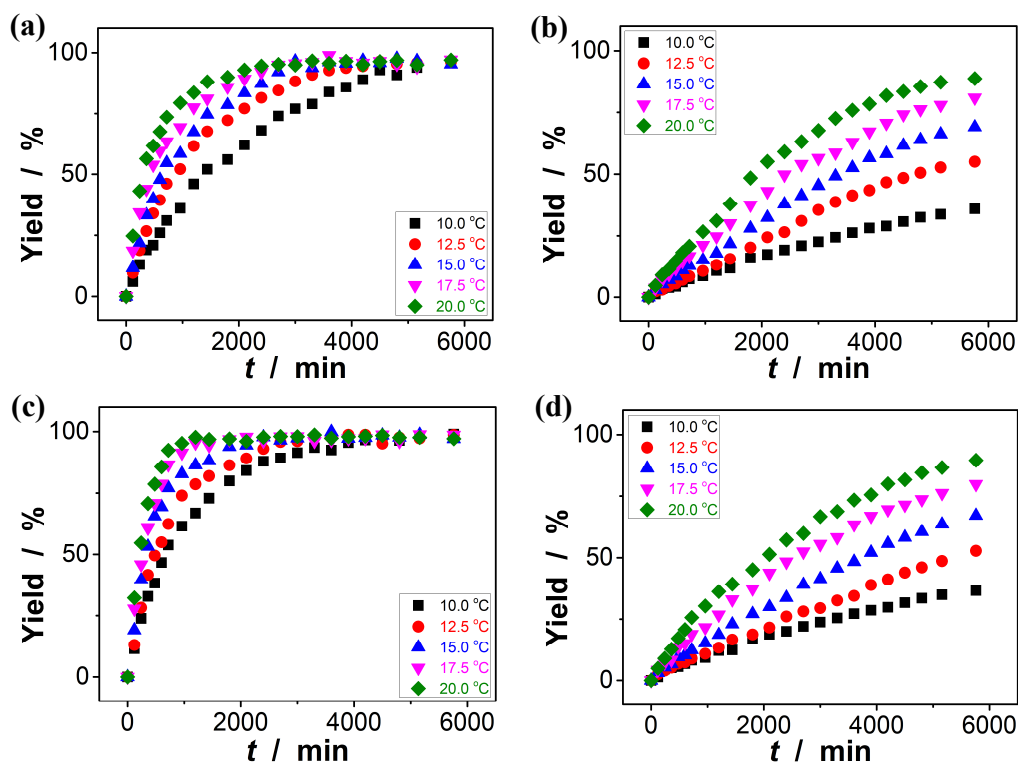


Fig. S16. Time trace of the yield of **6** of the reaction catalyzed by the PBA@PM in the absence (a,b) or presence (c,d) of glucose (1.5 mM), upon bubbling with N₂ (a,c) or CO₂ (b,d) gas.

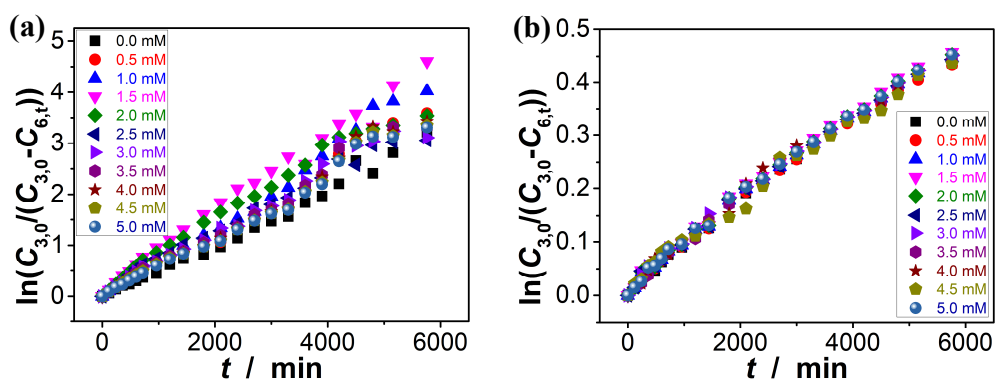


Fig. S17. Influence of the [Glu] on the k , measured in the reaction mixture upon bubbling with (a) N₂ or (b) CO₂ gas at 10.0 °C.

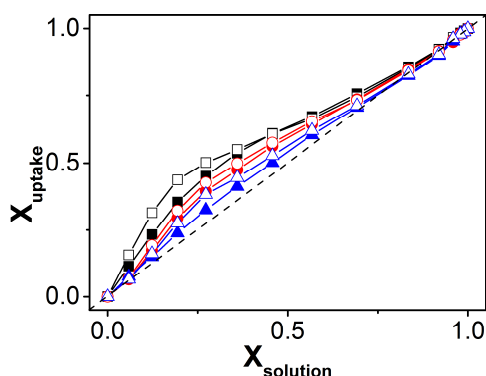


Fig. S18. A comparison of the uptaken mol fraction of **3** (■, [Glu] = 0.0 mM; ●, [Glu] = 1.5 mM; ▲, [Glu] = 5.0 mM), or **4** (□, [Glu] = 0.0 mM; ○, [Glu] = 1.5 mM; △, [Glu] = 5.0 mM), in the equilibrated PBA@PM at 10.0 °C, and corresponding mol fraction in initial solutions.

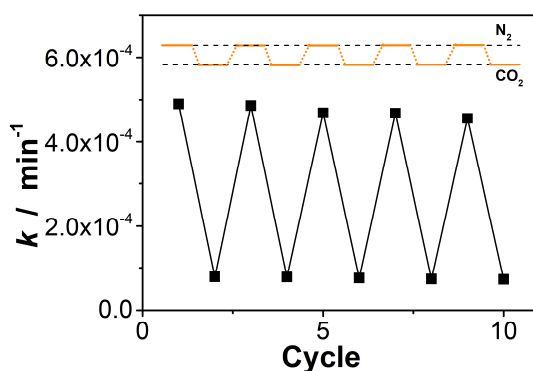


Fig. S19. A comparison of the catalysis activity of the PBA@PM upon repeated bubbling with N₂ or CO₂ gas, measured at 10.0 °C without glucose.

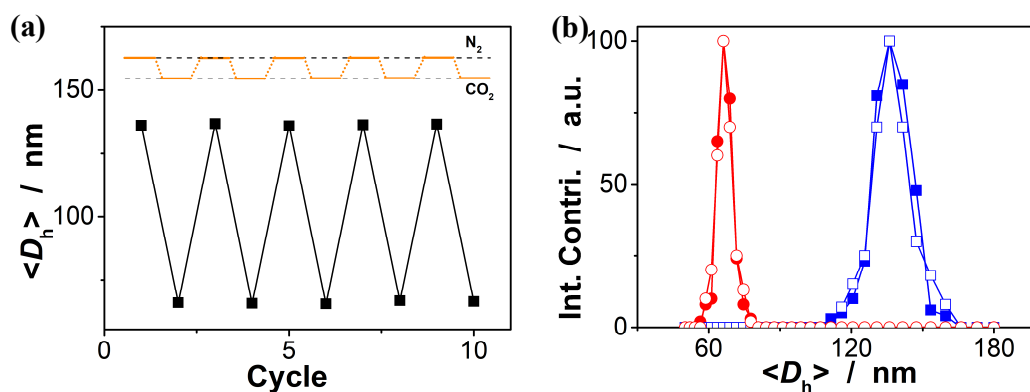


Fig. S20. (a) A comparison of the $\langle D_h \rangle$ of the PBA@PM upon repeated use in the reaction bubbling with N₂ or CO₂ gas. (b) DLS size distribution of the PBA@PM before (■, ●) and after (□, ○) ten cycles of use as catalyst and bubbling with N₂ (■, □) or CO₂ (●, ○) gas. The reused PBA@PM was collected after each reaction and purified for DLS measurements at 10.0 °C.

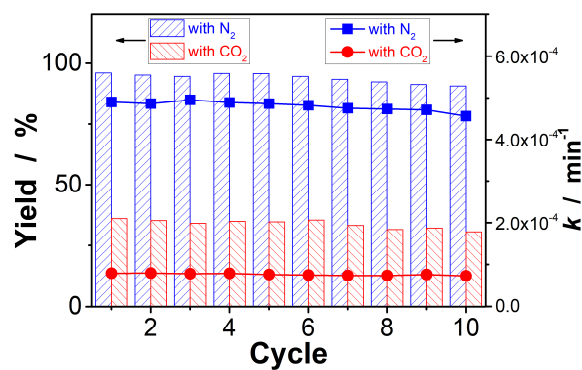


Fig. S21. A comparison of the catalysis activity of the PBA@PM upon bubbling with N₂ or CO₂ gas during the recycling experiments, measured at 10.0 °C without glucose. The PBA@PM was simply separated by centrifugation, and reused directly for the next cycle. The yield of **6** were harvested after 96 hours' reaction.

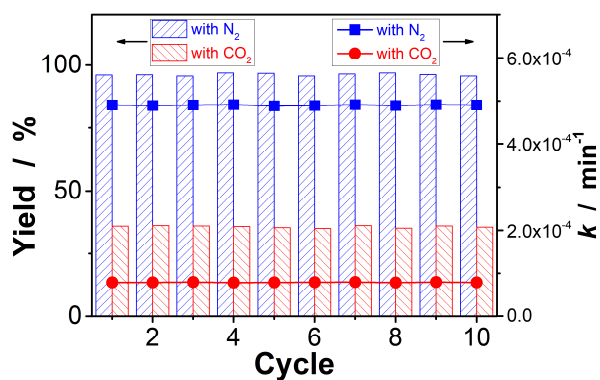


Fig. S22. A comparison of the catalysis activity of the PBA@PM upon bubbling with N₂ or CO₂ gas during the recycling experiments, measured at 10.0 °C without glucose. The recycled PBA@PM was purified before reuse. The yield of **6** were harvested after 96 hours' reaction.

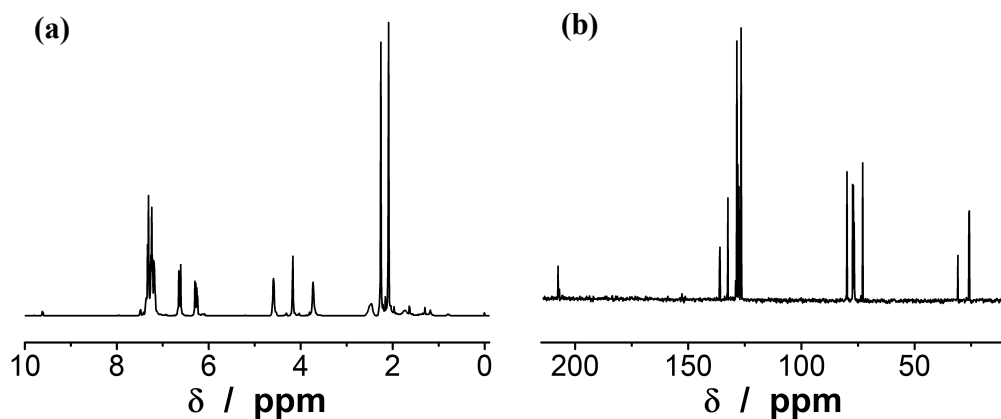


Fig. S23. (a) ^1H NMR (in CDCl_3) and (b) ^{13}C NMR (in CDCl_3) spectra of aldol adduct **7**. ^1H NMR: $\delta = 7.33\text{-}7.15$ (5H), 6.62 (1H), 6.26 (1H), 4.61 (1H), 4.18 (1H), 3.74 (1H), 2.47 (1H), 2.25 (3H); ^{13}C NMR: $\delta = 207.5, 136.2, 132.5, 128.5, 128.1, 127.6, 126.9, 80.2, 72.8, 26.2$.

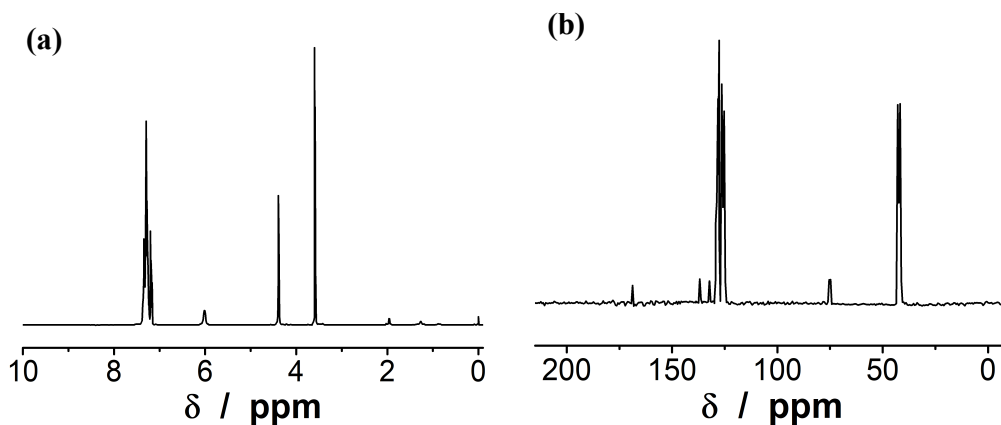


Fig. S24. (a) ^1H NMR (in CDCl_3) and (b) ^{13}C NMR (in CDCl_3) spectra of amide product **8**. ^1H NMR: $\delta = 7.33\text{-}7.15$ (10H), 6.02 (1H), 4.37 (2H), 3.59 (2H); ^{13}C NMR: $\delta = 169.8, 137.2, 134.6, 129.3, 128.9, 128.5, 127.4, 127.3, 127.2, 43.6, 43.4$.

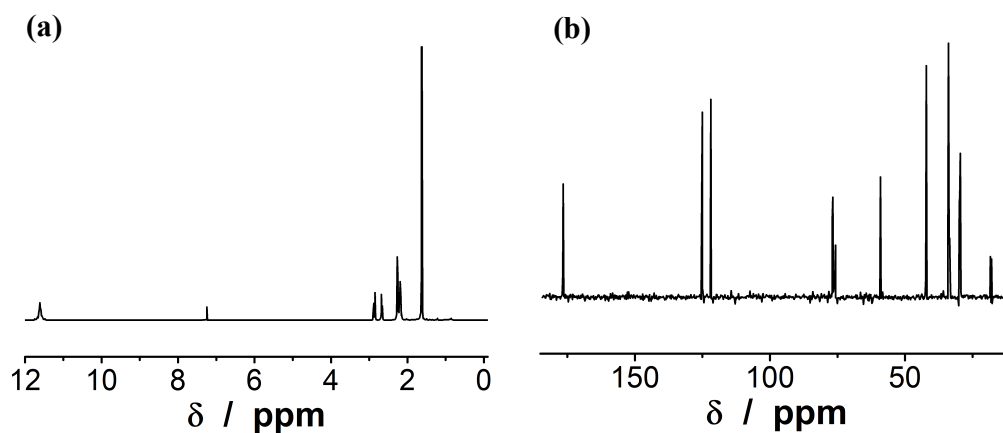


Fig. S25. (a) ^1H NMR (in CDCl_3) and (b) ^{13}C NMR (in CDCl_3) spectra of cycloadduct **9**. ^1H NMR: $\delta = 11.62$ (1H), 2.85 (1H), 2.68 (1H), 2.27 (2H), 2.18 (2H), 1.63 (6H); ^{13}C NMR: $\delta = 177.3, 125.0, 122.7, 59.2, 43.0, 34.1, 30.3, 19.0, 18.6$.

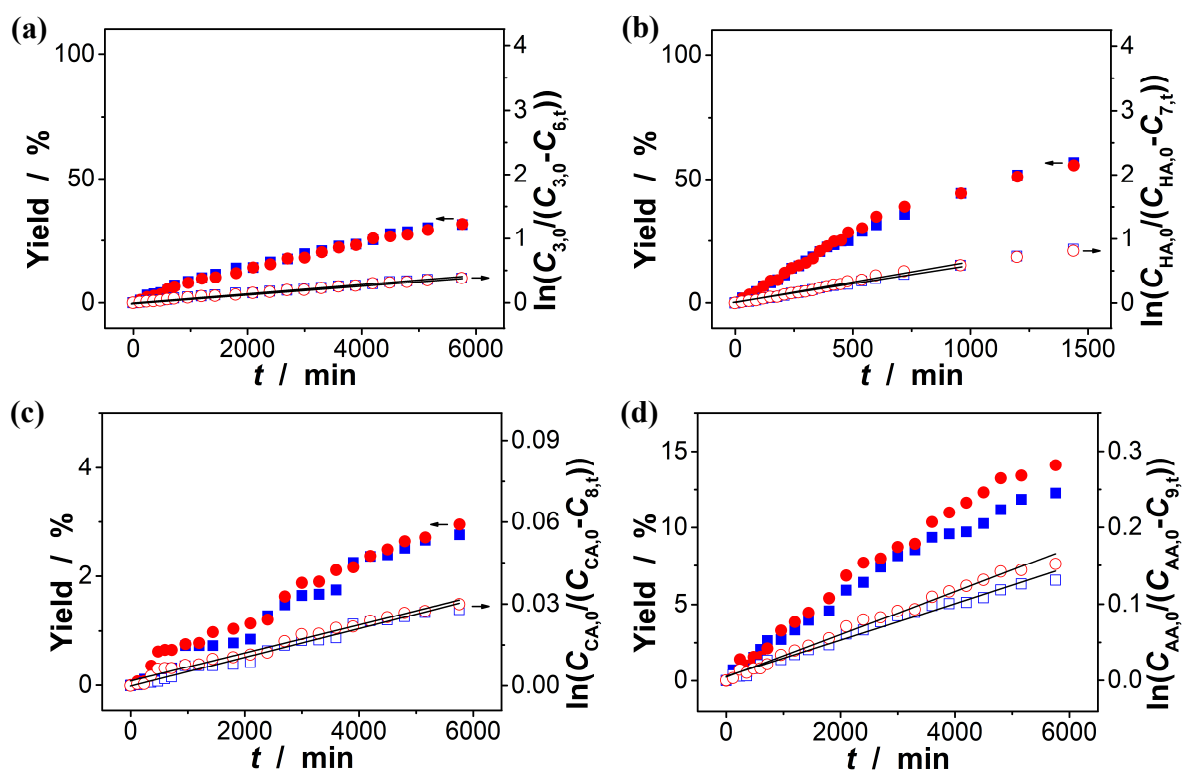


Fig. S26. Control experiments on the model (a) aza-Michael addition, (b) aldol, (c) amidation, and (d) [4+2] cycloaddition reactions catalyzed by **1**, upon bubbling with N_2 (\blacksquare, \square) or CO_2 (\bullet, \circ) gas, and at $10.0\text{ }^\circ\text{C}$, where lines are 1st-order kinetic fits.