SUPPORTING INFORMATION

Stereodivergent access to all four stereoisomers of chiral tetrahydrobenzo[*f*][1,4]oxazepines, through highly diastereoselective multicomponent Ugi-Joullié reaction.

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Experimental procedures

General methods

NMR spectra were taken at the indicated temperature in CDCl₃ or d6-DMSO at 300 MHz (¹H), and 75 MHz (¹³C), using, as internal standard, TMS (¹H NMR: 0.000 ppm) or the central peak of CDCl₃ (¹³C: 77.02 ppm), or the central peak of DMSO (¹H NMR in d₆-DMSO; 2.506 ppm), or the central peak of DMSO (¹³C in d₆-DMSO; 39.43 ppm). Chemical shifts are reported in ppm (δ scale). Peak assignments were made with the aid of gCOSY and gHSQC experiments. HRMS: samples were analysed with a Synapt G2 QToF mass spectrometer. MS signals were acquired from 50 to 1200 m/z in ESI positive ionization mode. TLC analyses were carried out on silica gel plates and viewed at UV (254 nm) and developed with Hanessian stain (dipping into a solution of (NH₄)₄MoO₄·4 H₂O (21 g) and Ce(SO₄)₂·4 H₂O (1 g) in H₂SO₄ (31 ml) and H₂O (469 ml) and warming) or with ninhydrin. R_f were measured after an elution of 7-9 cm. Column chromatographies were done with the "flash" methodology using 220-400 mesh silica. Petroleum ether (40-60 °C) is abbreviated as PE. In extractive work-up, aqueous solutions were always reextracted three times with the appropriate organic solvent. Organic extracts were always dried over Na₂SO₄ and filtered, before evaporation of the solvent under reduced pressure. All reactions using dry solvents were carried out under a nitrogen atmosphere.

General procedure for the synthesis of Boc-protected amino alcohols 7a-c from the corresponding Boc-α-amino acids.



Boc-amino acid was dissolved in dry THF (1 mL/mmol). This solution was cooled at -10 °C, and isobutyl chloroformate (1.2 eq.) and N,N-diisopropylethylamine (1.5 eq.) were added in this order. The mixture was stirred at – 10 °C for 1h. Then it was filtered and the filtrate was cooled at -10 °C and treated with NaBH₄ (1.5 eq.) and H₂O (3 mL/mmol of NaBH₄), in this sequence. After stirring at -10 °C for 30 min and at rt for 1h, the mixture was diluted with AcOEt and with saturated aqueous NH₄Cl and 5% aqueous (NH₄)H₂PO₄)NH₄ (resulting pH = 5). After phase separation, the organic phases were washed with brine, and evaporated to dryness. Chromatography afforded the pure products.

7a. Chromatography: PE/AcOEt/CH₂Cl₂ 2:1:2. Yield: 88%. Only (S)-compound was prepared and used. This compound is known.¹

7b. Chromatography: PE/AcOEt/CH₂Cl₂ 1:1:1. Yield: 87%. Only (S)-compound was prepared and used. This compound is known.²

7c. Chromatography: PE/AcOEt 3:1. Yield: 87%. Only (S)-compound was prepared and used. This compound is known.²

General procedure for the synthesis of Boc-protected amino alcohols 7d-e from the corresponding amino alcohol.



1.010 g (7.36 mmol) of R-(-)-2-phenyl glycinol was dissolved in 12.3 mL of dry CH_2Cl_2 . 1.13 mL (8.11 mmol, 1.1 eq.) of Et_3N and 1.771 g (8.11 mmol, 1.1 eq.) of (Boc)_2O were added at this solution. The mixture was stirred overnight at room temperature, then it was diluted with CH_2Cl_2 and with 1 M HCl. After phase separation, the organic phases were washed with saturated aqueous NaHCO₃, and evaporated to dryness. The crude product was purified by chromatography using PE:AcOEt:CH_2Cl_2 2:1:1. The product (1.500 g, 6.32 mmol) was obtained as a white solid with a yield of 86,0%. This product is known.³

Using the same procedure, both (*R*)- and (*S*)-7e were prepared in 93% yield from the corresponding amino alcohols. They are both known.²

Optimization of the Mitsunobu reaction to give 8a

The reaction was first optimized using stoichiometric quantities of the two reagents. Thus, all the following reactions were carried out in THF at 0°C for 3 h and then at rt overnight, using 1 eq. each of **6a** and **7a** and 1.5 eq. of PPh₃ and of azodicarboxylate. A main side-product was detected, corresponding to Boc-aziridine **9a**.



Then, also taking into account the fact that the Boc-amino alcohol is more precious than the salicyladehyde, we decided to use a slight excess (1.2 equiv.) of salicylaldehyde. Finally, we noticed that a slight better yield was achieved by adding PPh₃ and azodicarboxylate in three portions.

Typical procedure for the Mitsunobu reaction. (S)-tert-Butyl (1-(2-(dimethoxymethyl)phenoxy)-3-phenylpropan-2-yl)carbamate 8a.



A solution of salicylaldehyde (0.770 mL, 901 mg, 7.38 mmol) in dry MeOH (8.1 mL) was treated with trimethyl orthoformate (3.50 mL, 32.0 mmol) and Amberlyst 15 (100 mg). The resulting mixture was stirred at room temperature for 20 h, then diluted with CH_2Cl_2 and filtered. The filtrate was treated with solid NaHCO₃ (25 mg) and filtered again. The filtrate was evaporated, and the residue was taken up in CH_2Cl_2 /toluene, and again evaporated to azeothropically remove all methanol. The crude acetal **6a** was obtained as a yellow oil (1.380 g). This acetal (301.7 mg, theoretical 1.61 mmol, 1.2 equiv.) S3

was dissolved in dry THF (2.60 mL), cooled to 0°C, and treated sequentially with Boc-amino alcohol **7a** (330.5 mg, 1.31 mmol, 1 equiv.), PPh₃ (171.8 mg, 0.655 mmol, 0.5 equiv.) and 1,1'-(Azodicarbonyl)dipiperidine (ADDP) (165.5 mg, 0.656 mmol, 0.5 equiv.). After stirring at 0°C for 60 min, further PPh₃ (171.8 mg, 0.655 mmol) and ADDP (165,5 mg, 0.656 mmol) were added. Finally, after 60 minutes, an identical amount of both reagents was added. The mixture was further stirred overnight at rt and evaporated to dryness. The crude was taken up in Et₂O and washed with 1 M NaOH, to remove excess of **6a**. The organic phase was washed with aqueous saturated NH₄Cl, evaporated and purified by chromatography (PE / AcOEt 11:1) to give pure **8a** as an oil (411.4 mg, 78% from **7a**).

 R_f = 0.20 (PE / AcOEt 10:1). [α]_D = −15.1 (c 1.1, CHCl₃). ¹H NMR (300 MHz, CDCl₃, 20 °C): δ = 7.54 (1 H, dd, J = 7.6, 1.6 Hz), 7.34-7.14 (6 H, m), 6.98 (1 H, t, J = 7.5 Hz), 6.76 (1 H, d, J = 8.2 Hz), 5.73 (1 H, s, CH(OCH₃)₂), 5.63 (1 H, broad d, J = 8.1 Hz, NH), 4.21-4.05 (1 H, broad m, CHN), 3.98 (1 H, dd, J = 9.3, 2.6 Hz, CHHO), 3.84 (1 H, dd, J = 9.3, 3.8 Hz, CHHO), 3.47 (3 H, s, OCH₃), 3.29 (3 H, s, OCH₃), 3.10-2.94 (2 H, m, CH₂Ph), 1.43 (9 H, s, C(CH₃)₃). ¹³C NMR (75 MHz, CDCl₃, 20 °C): δ = 156.5, 155.5 (C=O and C-O), 138.1, 126.1 (quat.), 129.8, 129.5 (x2), 128.5 (x2), 127.6, 126.5, 120.7, 112.3 (aromatic CH), 99.4 (CH(OCH₃)₂), 79.3 (C(CH₃)₃), 68.6 (CH₂O), 54.4 (OCH₃), 51.7 (OCH₃ and CHN), 38.2 (CH₂Ph), 28.4 (C(CH₃)₃). I.R. (ATR): v_{max} 3346, 2978, 2930, 2897, 2827, 1700, 1606, 1590, 1522, 1494, 1455, 1442, 1388, 1365, 1337, 1284, 1244, 1203, 1164, 1125, 1096, 1047, 1033, 987, 973, 960, 921, 901, 881, 866, 850, 822, 780, 763, 742, 698, 670, 641, 623, 608 cm⁻¹. HRMS (ESI+): found 402.2287 [Calcd for C₂₃H₃₂NO₅⁺ (M + H)⁺ 402.2280].

(S)-tert-Butyl 2-benzylaziridine-1-carboxylate 9a.



9a

Oil. R_f = 0.49 (PE / AcOEt 10:1). ¹H NMR (300 MHz, CDCl₃, 20 °C): δ = 7.35-7.20 (5 H, m), 3.02-2.89 (1 H, m, C*H*N), 2.71-2.57 (2 H, m, C*H*₂N), 2.31 (1 H, d, J = 6.0 Hz, C*H*HPh), 2.03 (1 H, d, J = 3.6 Hz, C*H*HPh), 1.44 (9 H, s, C(CH₃)₃). This compound is known.⁴

(S)-tert-Butyl (1-(2-(dimethoxymethyl)phenoxy)-4-methylbutan-2-yl)carbamate 8b.



It was prepared from salicylaldehyde and Boc-amino alcohol **7b** following the same procedure described for **8a**, but using stoichiometric quantities of **6a** and Boc-amino alcohol **7b**. The isolated yield of **8b** was 65% from salicylaldehyde. Chromatography was carried out with PE: AcOEt 30:1 \rightarrow 10:1.

Oil. $R_f = 0.34$ (PE / AcOEt 30:1). $[\alpha]_D = +50.3$ (c 1.0, CHCl₃). ¹H NMR (300 MHz, CDCl₃, 20 °C): $\delta = 7.52$ (1 H, dd, J = 7.5, 1.5 Hz), 7.29 (1 H, td, J = 7.8, 1.5 Hz), 6.97 (1 H, td, J = 7.5, 1.2 Hz), 6.85 (1 H, dd, J = 8.1, 0.6 Hz), 5.65 (1 H, s, CH(OCH₃)₂), 5.29 (1 H, broad d, J = 8.1 Hz, NH), 4.10-3.91 (3 H, m, CHN, CHHO), 3.43 (3 H, s, OCH₃), 3.28 (3 H, s, OCH₃), 1.80-1.45 (2 H, m, CH₂CH(CH₃)₂), 1.43 (9 H, s, C(CH₃)₃), 0.97 (6 H, d, J = 6.6 Hz, (CH₃)₂CH). ¹³C NMR (75 MHz, CDCl₃, 20 °C): $\delta = 156.6$, 155.6 (C=O and C-O), 129.8, 127.5, 120.6, 112.2 (aromatic CH), 126.2 (quat.), 99.3 (CH(OCH₃)₂), 79.1 (C(CH₃)₃), 71.0 (CH₂O), 54.5 (OCH₃), 52.0 (OCH₃), 48.4 (CHN), 41.4 (CH₂/Pr), 28.4 (C(CH₃)₃), 24.9 (CH(CH₃)₂), 22.9, 22.5 (CH(CH₃)₃. I.R. (ATR): vmax 3349, 2956, 2933, 2871, 2830, 1697, 1604, 1516, 1491, 1455, 1391, 1366, 1329, 1285, 1241, 1163, 1118, 1091, 1047, 977, 910, 872, 847, 811, 754, 731, 672, 647, 610 cm⁻¹. HRMS (ESI+): found 368.2429 [Calcd for C₂₀H₃₄NO₅⁺ (M + H)⁺ 368.2437].

(S)-tert-Butyl (1-(2-(dimethoxymethyl)phenoxy)-3-methylpropan-2-yl)carbamate 8c.



It was prepared from salicylaldehyde and Boc-amino alcohol **7c** following the same procedure described for **8a**, but using 1.1 equiv. of **6a** relative to Bocamino alcohol **7c**. The isolated yield of **8c** was 55% from **7c**. Chromatography was carried out with PE: AcOEt $16:1 \rightarrow 10:1$.

Oil. $R_f = 0.25$ (PE / AcOEt 16:1). $[\alpha]_D = -56.2$ (c 1.0, CHCl₃). ¹H NMR (300 MHz, CDCl₃, 20 °C): $\delta = 7.52$ (1 H, dd, J = 7.6, 1.6 Hz), 7.29 (1 H, td, J = 7.5, 1.5 Hz), 6.98 (1 H, t, J = 7.5 Hz), 6.85 (1 H, d, J = 8.1 Hz), 5.64 (1 H, s, CH(OCH₃)₂), 5.43 (1 H, broad d, J = 9.0 Hz, NH), 4.21 (1 H, dd, J = 9.3, 2.8 Hz, CHO), 3.92 (1 H, dd, J = 9.3, 3.8 Hz, CHO), 3.73-3.60 (1 H, m, CHN), 3.43 (3 H, s, OCH₃), 3.26 (3 H, s, OCH₃), 2.02 (1 H, octuplet, J = 7.1 Hz, CH(CH₃)₂), 1.43 (9 H, s, C(CH₃)₃), 1.02 (3 H, d, J = 6.9 Hz, (CH₃)CH), 1.00 (3 H, d, J = 6.9 Hz, (CH₃)CH). ¹³C NMR (75 MHz, CDCl₃, 20 °C): $\delta = 156.7$, 155.9 (C=O and C-O), 129.8, 127.5, 120.5, 112.1 (aromatic CH), 126.2 (quat.), 99.3 (CH(OCH₃)₂), 79.0 (C(CH₃)₃), 69.1 (CH₂O), 55.6 (CHN), 54.5 (OCH₃), 51.7 (OCH₃), 30.2 (CH(CH₃)₂), 28.4 (C(CH₃)₃), 19.6, 19.3 (CH(CH₃)₃. I.R. (ATR): v_{max} 3348, 2962, 2934, 2829, 1711, 1604, 1591, 1516, 1491, 1456, 1390, 1365, 1286, 1240, 1170, 1121, 1092, 1073, 1045, 1028, 979, 946, 911, 883, 867, 807, 755, 737, 703, 675, 650, 620, 610 cm⁻¹. HRMS (ESI+): found 354.2284 [Calcd for C₁₉H₃₂NO₅⁺ (M + H)⁺ 354.2280].

(R)-tert-Butyl (2-(2-(dimethoxymethyl)phenoxy)-1-phenylethan-1-yl)carbamate 8d.



It was prepared from salicylaldehyde and Boc-amino alcohol **7d** following the same procedure described for **8a**, but using 1.1 equiv. of **6a** relative to Bocamino alcohol **7d**. The isolated yield of **8d** was 42% from **7c**. Chromatography was carried out with PE: AcOEt 8:1 \rightarrow 6:1.

Oil. $R_f = 0.20$ (PE / AcOEt 8:1). $[\alpha]_D = -33.5$ (c 1.2, CHCl₃). ¹H NMR (300 MHz, CDCl₃, 20 °C): $\delta = 7.49$ (1 H, dd, J = 7.6, 1.6 Hz), 7.46-7.22 (6 H, m), 6.97 (1 H, t, J = 7.5 Hz), 6.82 (1 H, d, J = 8.4 Hz), 6.12 (broad s, NH), 5.54 (1 H, s, CH(OCH₃)₂), 5.09 (1 H, broad s, CHNH), 4.32-4.15 (2 H, m, CH₂O), 3.40 (3 H, s, OCH₃), 3.28 (3 H, s, OCH₃), 1.43 (9 H, s, C(CH₃)₃). ¹³C NMR (75 MHz, CDCl₃, 20 °C): $\delta = 156.3$, 155.5 (C=O and C-O), 140.2, 126.5 (quat.), 129.8, 128.5 (x2), 127.6, 127.4, 126.8 (x2), 120.9, 112.4 (aromatic CH), 99.6 (CH(OCH₃)₂), 79.5 (C(CH₃)₃), 72.0 (CH₂O), 54.6 (OCH₃), 54.1 (CHN), 52.0 (OCH₃),

28.4 (C(CH₃)₃). I.R. (ATR): v_{max} 3335, 2977, 2933, 2830, 1706, 1604, 1591, 1515, 1491, 1454, 1391, 1366, 1284, 1239, 1164, 1121, 1092, 1070, 1047, 979, 947, 902, 867, 794, 751, 699, 666, 633 cm⁻¹. HRMS (ESI+): found 388.2119 [Calcd for C₂₂H₃₀NO_{5⁺} (M + H)⁺ 388.2124].

(S)-*tert*-Butyl (1-(2-(dimethoxymethyl)phenoxy)-propane-2-yl)carbamate 8e.



It was prepared from 4-bromosalicylaldehyde **1b** and Boc-amino alcohol (*S*) **7e** following the same procedure described for **8a**, but using 1.1 equiv. of **6b** relative to Boc-amino alcohol (*S*)-**7e**. The isolated yield of **8d** was 60% from **7e**. Chromatography was carried out with PE: AcOEt : CH₂Cl₂ 8:1:1 \rightarrow 6:1:1. Oil. R_f = 0.37 (PE / AcOEt / CH₂Cl₂ 8:1:1). [α]_D = -40.4 (c 1.0, CHCl₃). ¹H NMR (300 MHz, CDCl₃, 20 °C): δ = 7.64 (1 H, d, J = 2.4 Hz), 7.38 (1 H, dd, J = 8.7, 2.7 Hz), 6.74 (1 H, d, J = 8.7 Hz), 5.61 (1 H, s, CH(OCH₃)₂), 5.26 (broad s, NH), 4.05 (1 H, broad s, CHNH), 4.01 (1 H, dd, J = 8.9, 2.8 Hz, CHHO), 3.90 (1 H, dd, J = 8.9, 4.3 Hz, CHHO), 3.41 (3 H, s, OCH₃), 3.28 (3 H, s, OCH₃), 1.44 (9 H, s, C(CH₃)₃), 1.31 (3 H, d, J = 6.6 Hz, CH₃CH). ¹³C NMR (75 MHz, CDCl₃, 20 °C): δ = 155.6, 155.3 (C=O and C-O), 132.4, 130.6, 114.0 (aromatic CH), 128.4, 113.3 (quat.), 98.5 (CH(OCH₃)₂), 79.3 (C(CH₃)₃), 72.2 (CH₂O), 54.3 (OCH₃), 52.0 (OCH₃), 45.9 (CHN), 28.4 (C(CH₃)₃), 18.2 (CH₃CH). I.R. (ATR): vmax 3350, 2976, 2933, 2830, 1695, 1595, 1514, 1487, 1457, 1404, 1391, 1365, 1266, 1242, 1164, 1133, 1099, 1053, 980, 912, 875, 854, 806, 781, 752, 734, 677, 657, 645, 616 cm⁻¹. HRMS (ESI+): found 404.1075 [Calcd for C₁₇H₂₇BrNO₅⁺ (M + H)⁺ 404.1073].

(*R*)-*tert*-Butyl (1-(2-(dimethoxymethyl)phenoxy)-propane-2-yl)carbamate *ent*-8e.



It was prepared exactly as for **8e**, but starting from *ent*-**7e**. $[\alpha]_D = +39.9$ (c 1.0, CHCl₃). The other spectroscopical data were identical to those of **8e**.

Optimization of Ugi-Joulliè reaction to give 10a.

In all cases (except entries 6,9) 1.2 equivalents of acetic acid and 1.2 equivalents of *tert*-butyl isocyanide were used.



Entry	Solvent	Additive	Time	Temp.	Yield % ^a	D.r. ^b
1	MeOH	none	45 h	rt	80	96:4
2	CH ₂ Cl ₂	none	38 h	rt	74	90.5 : 9.5
3	CH ₂ Cl ₂	ZnBr ₂ (0.5 equiv.)	45 h	rt	69	94 : 6
4	CH ₂ Cl ₂	ZnBr ₂ (1.0 equiv.)	48 h (incomplete)	rt	51	89:11
5	MeOH	ZnBr ₂ (0.5 equiv.)	48 h (incomplete)	rt	49	96:4
6	CH ₂ Cl ₂	Zn(OAc) ₂ (1.2 equiv.) ^c	48 h (largely incomplete)	rt	9	95 : 5
7	THF	none	48 h (incomplete)	rt	29	88 : 12
8	THF	ZnBr ₂ (0.5 equiv.)	48 h (incomplete)	rt	34	95 : 5
9	THF	Zn(OAc) ₂ (1.2 equiv.) ^c	48 h (no reaction)	rt	-	-
10	EtOH	none	48 h (incomplete)	rt	62	92:8
11	<i>i</i> BuOH	none	48 h (incomplete)	rt	46	87 : 13
^a Isolated yi	eld of major diaste	ereomer only. ^b Determined by HPL	C on the crude product. ^c Zn(OAc) ₂ was used inste	ad of Ac	ОН	

Typical procedure for the synthesis of Ugi-Joullié adducts: (3S,5R)-4-Acetyl-3-benzyl-N-(*tert*-butyl)-2,3,4,5-tetrahydrobenzo[f][1,4]oxazepine-5-carboxamide 10a.

Acetal **8a** (312.4 mg, 778 µmol) was dissolved in CH₂Cl₂ (3.0 mL) and treated with 37% aqueous HCl (640 µL, 7.73 mmol). The mixture was stirred at rt for 5 h. Then it was diluted with CH₂Cl₂, and cautiously treated with 5% aqueous Na₂CO₃ (25 mL). After checking that pH > 9, the two phases were separated, washed with brine, and evaporated to dryness. The resulting crude imine **3a** (oil) was taken up in dry methanol (3.90 mL), and treated with acetic acid (53 µL, 927 µmol), and *tert*-butyl isocyanide (106 µl, 937 µmol). The mixture was stirred at rt for 45 h and evaporated to dryness. It was taken up in AcOEt, washed with saturated aqueous NaHCO₃ (to remove excess carboxylic acid), evaporated, and chromatographed (PE / AcOEt 2:1) to give pure **10a** (237 mg, 80%). The ratio **10a** : **11a** was determined by HPLC on the crude product and resulted = 96:4. HPLC conditions: C6-Phenyl column 150 x 3 mm, 3 µm. Flow: 0.38 ml/min. Temp: 26°C. Eluent: H₂O : CH₃CN 55:45. Detection: UV 220 nm. R_t of **10a**: 13.62. R_t of **11a**: 14.41. Minor diastereomer **11a** was recognized using a MS (ESI) detector.

White solid. $R_f = 0.24$ (PE / AcOEt 2:1) (R_f of **11a** = 0.31). [α]_D = +10.0 (c 1.06, CHCl₃). M.p. = 119.7-122.0 °C. ¹H NMR (300 MHz, d6-DMSO, 90 °C): δ = 7.57 (1 H, dd, J = 7.7, 1.6 Hz), 7.36-7.16 (6 H, m), 7.00 (2 H, t, J = 8.2 Hz), 6.47 (1 H, broad s, NH), 5.66 (1 H, broad s, CHC=O), 4.77 (1 H, broad d, J = 11.1 Hz, CHHO), 4.55 (1 H, broad s, CHN), 4.03 (1 H, dd, J = 13.2, 4.8 Hz, CHHO), 2.79 (1 H, dd, J = 13.4, 5.9 Hz, CHHPh), 2.65-2.48 (1 H, m, CHHPh), 2.06 (3 H, s, CH₃C=O), 1.22 (9 H, s, C(CH₃)₃). ¹³C NMR (75 MHz, d6-DMSO, 90 °C): δ = 169.8, 168.8 (C=O), 155.6 (aromatic C-O), 137.6, 121.4 (quat.),

132.1, 129.6, 128.4 (x2), 127.8 (x2), 125.8, 120.9, 118.8 (aromatic CH), 66.6 (CH₂O), 62.8 (broad) (CHC=O), 57.5 (broad) (CHN), 50.2 ($C(CH_3)_3$), 36.4 (CH₂Ph), 27.8 ($C(CH_3)_3$), 21.2 (CH₃C=O). I.R. (ATR): v_{max} 3410, 3314, 3067, 3034, 2966, 2928, 2870, 1671, 1639, 1607, 1580, 1563, 1509, 1494, 1453, 1416, 1391, 1366, 1352, 1332, 1305, 1288, 1267, 1249, 1224, 1193, 1172, 1157, 1125, 1115, 1086, 1052, 1028, 1007, 973, 937, 908, 885, 871, 862, 827, 802, 752, 731, 719, 700, 665, 644, 631, 616, 603 cm⁻¹. HRMS (ESI+): found 381.2171 [Calcd for C₂₃H₂₉N₂O₃⁺ (M + H)⁺ 381.2178].



(3S,5R)-3-Benzyl-N-(cyclohexyl)-4-phenylacetyl-2,3,4,5-tetrahydrobenzo[f][1,4]oxazepine-5-carboxamide 10b.

It was prepared in 65% overall yield from (S)-8a, phenylacetic acid and cyclohexyl isocyanide, following the typical procedure described for 10a. Chromatography was carried out with PE / AcOEt / $CH_2Cl_2 5 : 1 \cdot 1$. Separation of the two diastereomers was not complete. We obtained fractions with pure 10b plus some fractions contaminated with little 11b. The overall yield was 65%. The calculated yield of 10b is therefore 62%. The d.r. was = 94 : 6 by HPLC of the crude product. Conditions: column Luna C8 150 x 4.6 mm, 5 μ . Flow: 0.8 ml/min. Temp: 25°C. Eluent: H₂O : MeOH 30:70. Detection: UV 220 nm. R_t of 10b: 21.25. R_t of 11b: 20.43. Minor diastereomer 11b was recognized using a MS (ESI) detector.

White solid. $R_f = 0.21$ (PE / AcOEt / CH₂Cl₂ 5 : 1 : 1) (R_f of **11b** = 0.24). [α]_D = +9.0 (c 1.0, CHCl₃). M.p. = 157.8-159.9 °C. ¹H NMR (300 MHz, d6-DMSO, 90 °C): δ = 7.42 (1 H, broad s), 7.36-7.15 (11 H, m), 7.05-6.95 (2 H, m), 6.80 (1 H, broad d, J = 6.6 Hz, N*H*), 5.72 (1 H, s, C*H*C=O), 4.75 (1 H, broad d, J = 12.3 Hz, C*H*HO), 4.61 (1 H, broad s, C*H*N), 4.02 (1 H, dd, J = 13.3, 4.6 Hz, C*H*HO), 3.84 (1 H, d, J = 15.6 Hz, C*H*HC=O), 3.67-3.50 (2 H, m, C*H*C=O, C*H*NH), 2.80 (1 H, dd, J = 13.2, 6.0 Hz, C*H*HPh), 2.65-2.48 (1 H, m, C*H*HPh), 1.80-1.45 (5 H, m, C*H*₂ cyclohexyl), 1.35-1.05 (5 H, m, C*H*₂ cyclohexyl). ¹³C NMR (75 MHz, d6-DMSO, 90 °C): δ = 170.2 168.4 (*C*=O), 155.6 (broad) (aromatic *C*-O), 137.5, 135.1, 121.3 (quat.), 132.1, 129.6 (x2), 128.5 (x2), 128.4 (x2), 127.8 (x2), 127.6, 125.83, 125.76, 121.0, 118.7 (aromatic CH), 66.7 (broad) (CH₂O), 65.9 (broad) (CHC=O), 57.3 (broad) (CHN), 47.7 (CHNH), 39.4 (CH₂C=O, hidden by DMSO signal, visible at qHSQC), 36.3 (broad) (CH₂Ph), 31.2 (x2), 24.6, 23.7, 23.6 (CH₂ cyclohexyl). I.R. (ATR): v_{max} 3271, 3023, 2977, 2940, 2920, 2855, 1656, 1633, 1604, 1582, 1525, 1492, 1451, 1416, 1360, 1343, 1319, 1306, 1289, 1270, 1250, 1222, 1193, 1150, 1115, 1094, 1076, 1060, 1029, 1012, 981, 968, 957, 935, 907, 893, 884, 875, 853, 826, 809, 781, 762, 749, 720, 696, 640, 619 cm⁻¹. HRMS (ESI+): found 483.2654 [Calcd for C₃₁H₃₅N₂O₃* (M + H)* 483.2648].





It was prepared in 80% overall yield from (*S*)-**8a**, *N*-benzyloxycarbonyl glycine and isopropyl isocyanide, following the typical procedure described for **10a**. Chromatography was carried out with PE / AcOEt 2 : 1. Separation of the two diastereomers was not complete. We obtained fractions with pure **10c** plus some fractions contaminated with little **11c**. The overall yield was 80%. The calculated yield of **10c** is therefore 74.4%. The d.r. was = 94 : 6 by HPLC of the crude product. Conditions: column Luna C8 150 x 4.6 mm, 5 μ . Flow: 1 ml/min. Temp: 26°C. Eluent: H₂O : CH₃CN 50:50. Detection: UV 220 nm. R_t of **10c**: 20.55. R_t of **11c**: 21.68. Minor diastereomer **11c** was recognized using a MS (ESI) detector.

White solid. $R_f = 0.12$ (PE / AcOEt 2 : 1) (R_f of **11c** = 0.17). [α]_D = -19.7 (c 1.0, CHCl₃). M.p. = 70.0-72.2 °C. ¹H NMR (300 MHz, d6-DMSO, 90 °C): δ = 7.56 (1 H, d, J = 7.5 Hz), 7.40-7.17 (11 H, m), 7.01 (2 H, t, J = 7.5 Hz), 7.00-6.90 (1 H, m, NHCH₂), 6.89 (1 H, broad d, J = 7.2 Hz, NHCH), 5.66 (1 H, s, CHC=O), 5.08 (2 H, s, PhCH₂O), 4.76 (1 H, broad d, J = 12.9 Hz, CHHO), 4.54 (1 H, broad s, CHN), 4.16 (1 H, dd, J = 16.6, 5.6 Hz, CHHNH), 4.01 (1 H, dd, J = 13.2, 4.5 Hz, CHHO), 3.93-3.77 (2 H, m, CHNH, CHHNH), 2.82 (1 H, dd, J = 13.4, 5.1 Hz, CHHPh), 2.62-2.48 (1 H, m, CHHPh), 1.08 (3 H, d, J = 6.6 Hz, CH₃), 1.00 (3 H, d, J = 6.6 Hz, CH₃). ¹³C NMR (75 MHz, d6-DMSO, 90 °C): δ = 168.6 168.2 (C=O), 155.7, 155.5 (C=O urethane and aromatic C-O), 137.4, 136.6, 121.1 (quat.), 132.4, 129.6, 128.4 (x2), 127.8 (x2), 127.7 (x2), 127.1, 126.9 (x2), 125.9, 120.9, 118.7 (aromatic CH), 66.0 (broad) (CH₂O), 65.1 (PhCH₂O), 61.6 (broad) (CHC=O), 57.2 (CHN), 42.3 (CH₂NH), 40.8 (CHNH), 36.3 (broad) (CH₂Ph), 21.3 (CH₃). I.R. (ATR): v_{max} 3410, 3321, 3064, 3031, 2971, 2935, 1715, 1648, 1606, 1581, 1510, 1493, 1453, 1424, 1387, 1366, 1347, 1241, 1220, 1156, 1115, 1084, 1049, 1030, 1010, 929, 912, 892, 871, 826, 797, 749, 697, 658, 638 cm⁻¹. HRMS (ESI+): found 516.2490 [Calcd for C₃₀H₃₄N₃O₅* (M + H)* 516.2498].

(3S,5R)-N-(4-Allyloxyphenyl)-3-benzyl-4-(3-bromobenzoyl)-2,3,4,5-tetrahydrobenzo[f][1,4]oxazepine-5-carboxamide 10d.



A solution of N-(4-(allyloxy)phenyl)formamide (100 mg, 564 μmol)⁵ in dry CH₂Cl₂ (0,84 mL) was cooled at 0°C and treated with 161.2, mg (676 μmol, 1.2 eq.) of Burgess reagent (Methyl *N*-(triethylammoniosulfonyl)carbamate inner salt). The mixture was stirred at 0°C for 3h, then further 66.0 mg (276 μmol) S9

of Burgess Reagent were added to the mixture. After 10 min. the mixture was allowed to reach room temperature and stirred for 2h. This mixture was then added to a solution of imine **3a** (400 µmol) in MeOH (1.5 mL), prepared as described above for the synthesis of **10a**. Finally, 112 mg (557 µmol mmol) of 3-bromobenzoic acid were added at this mixture. The mixture was stirred at room temperature for 48h and then 30 °C overnight. The solvent was evaporated under reduced pressure and the crude product was purified by chromatography (PE: AcOEt 5:1) to give pure **10d** (128.8 mg, 54%). The d.r. was = 93 : 7 by HPLC of the crude product. HPLC conditions: C6-Phenyl column 150 x 3 mm, 3 µm. Flow: 0.34 ml/min. Temp: 25°C. Eluent: H₂O : CH₃CN 40:60 + 1% CF₃CO₂H. Detection: UV 220 nm. R_t of **10d**: 13.33. R_t of **11d**: 14.00. Minor diastereomer **11d** was recognized using a MS (ESI) detector. The overall yield, calculated from the d.r., was 58%.

White solid. $R_f = 0.13$ (PE / AcOEt 2 : 1). $[\alpha]_D = +57.0$ (c 1.0, CHCl₃). M.p. = 148.6-150.7 °C. ¹H NMR (300 MHz, d6-DMSO, 90 °C): $\delta = 9.17$ (1 H, s, N*H*), 7.64 (2 H, ddd, J = 8.1, 1.8, 1.0 Hz), 7.44-7.18 (9 H, m), 7.10 (1 H, td, J = 7.5, 1.2 Hz), 7.05 (1 H, dd, J = 8.1, 0.9 Hz), 7.01-6.92 (2 H, m), 6.88 (2 H, d, J = 9.0 Hz), 6.02 (1 H, ddt, J = 17.2, 10.5, 5.2 Hz, CH=CH₂), 5.81 (1 H, broad s, CHC=O), 5.37 (1 H, dq, J = 17.2, 1.6 Hz, CH=CHH), 5.23 (1 H, dq, J = 10.5, 1.5 Hz, CH=CHH), 4.95 (1 H, dd, J = 12.3, 1.8 Hz, CHHO), 4.53 (2 H, dt, J = 5.4, 1.5 Hz, CH₂CH=CH₂), 4.44 (1 H, broad s, CHN), 4.13 (1 H, dd, J = 13.0, 4.0 Hz, CHHO), 2.73 (1 H, dd, J = 13.2, 7.3 Hz, CHPh), 2.62 (1 H, dd, J = 13.2, 8.4 Hz, CHHPh). ¹³C NMR (75 MHz, d6-DMSO, 90 °C): $\delta = 169.1 167.4$ (C=O), 155.6, 154.4, 138.0, 137.2, 131.2, 121.7, 121.2 (quat.), 133.3 (CH=CH₂), 131.9, 131.7, 130.1, 130.0, 128.5 (x2), 128.3, 127.8 (x2), 125.9, 124.6, 121.8, 121.4 (x2), 119.6, 114.4 (x2) (aromatic CH), 116.5 (CH=CH₂), 68.2 (CH₂CH=CH₂), 68.1 (broad) (CH₂O), 63.4 (broad) (CHC=O), 58.4 (broad) (CHN), 37.0 (broad) (CH₂Ph). I.R. (ATR): vmax 3314, 3062, 2945, 2906, 2026, 1672, 1646, 1606, 1578, 1563, 1509, 1488, 1446, 1414, 1386, 1353, 1330, 1305, 1278, 1256, 1211, 1171, 1148, 1114, 1094, 1085, 1069, 1061, 1027, 1007, 967, 934, 914, 897, 887, 864, 824, 807, 752, 730, 709, 703, 682, 665, 649, 637, 605 cm⁻¹. HRMS (ESI+): found 597.1396 [Calcd for C₃₃H₃₀BrN₂O₄⁺ (M + H)⁺ 597.1389].

(3S,5R)-3-iso-Butyl-N-(tert-butyl)-4-(phenylacetyl)-2,3,4,5-tetrahydrobenzo[f][1,4]oxazepine-5-carboxamide 10e.



It was prepared in 57% overall yield from (S)-**8b**, phenylacetic acid and *tert*-butyl isocyanide, following the typical procedure described for **10a**. The reaction under these conditions was not complete. Chromatography was carried out with PE / AcOEt 5 : 1. Separation of the two diastereomers was not complete. We obtained fractions with pure **10e** plus some fractions contaminated with little **1e**. The overall yield was 57%. The calculated yield of **10e** is therefore 54%. The d.r. was 96 : 4 by HPLC of the crude product. Conditions: column Luna C8 150 x 4.6 mm, 5 μ . Flow: 0.8 ml/min. Temp: 25°C. Eluent: H₂O : MeOH 30:70 + 1% CF₃CO₂H. Detection: UV 220 nm. R_t of **10e**: 18.25. R_t of **11e**: 16.23. Minor diastereomer **11e** was recognized using a MS (ESI) detector. Oil. R_f = 0.22 (PE / AcOEt 5 : 1) (R_t of **11e** = 0.25). [α]_D = +12.2 (c 1.0, CHCl₃). ¹H NMR (300 MHz, d6-DMSO, 90 °C): δ = 7.26-7.07 (7 H, m), 6.32 (1 H, s, NH), 5.52 (1 H, s, CHC=O), 4.59 (1 H, d, J = 12.9 Hz, CHHO), 4.36 (1 H, broad s, CHN), 4.12 (1 H, dd, J = 13.2, 5.2 Hz, CHHO), 3.74 (1 H, d, J = 15.6 Hz, CHHPh), 3.57 (1 H, d, J = 15.6 Hz, CHHPh), 1.51 (1 H, nonuplet, J = 6.6 Hz, CH(CH₃)₂), 1.17 (2 H, t, J = 6.9 Hz, CH₂iPr), 1.11 (9 H, s, C(CH₃)₃), 0.74 (3 H, d, J = 6.6 Hz, CH₃CHCH₃), 0.67 (3 H, d, J = 6.6 Hz, CH₃CHCH₃). ¹³C NMR (75 MHz, d6-DMSO, 90 °C): δ = 170.3 168.7 (C=O), 155.7, 135.2, 121.6 (quat.), 131.6, 129.3, 128.5 (x2), 127.6 (x2), 125.8, 120.8, 118.6 (aromatic CH), 67.6 (CH₂O), 62.3 (broad) (CHC=O), 54.0 (broad) (CHN), 50.2 (CH₂*i*Pr),

39.6 (CH₂Ph), 27.7 (C(CH₃)₃), 24.3 (CH(CH₃)₂), 22.7, 21.2 (CH₃). I.R. (ATR): v_{max} 3413, 3331, 3063, 3030, 2959, 2930, 2870, 1683, 1644, 1604, 1580, 1508, 1492, 1452, 1411, 1393, 1365, 1348, 1309, 1264, 1248, 1219, 1168, 1144, 1115, 1096, 1076, 1057, 1030, 1004, 957, 869, 823, 804, 758, 721, 695, 634 cm⁻¹. HRMS (ESI+): found 423.2641 [Calcd for C₂₆H₃₆N₂O₃⁺ (M + H)⁺ 423.2648].

(3S,5R)-N-(Benzyl)-3-iso-propyl-4-(methoxyacetyl)-2,3,4,5-tetrahydrobenzo[f][1,4]oxazepine-5-carboxamide 10f.



It was prepared in 53% yield from (*S*)-8*c*, methoxyacetic acid and benzyl isocyanide, following the typical procedure described for **10a**. However, in this case, the Ugi-Joullié step reaction was carred out at 40°C. Chromatography was carried out with PE / AcOEt 1 : 1, affording a complete separation of the two diastereomers. The overall yield was 56%. The d.r. was = 94 : 6 by HPLC of the crude product. Conditions: column Luna C8 150 x 4.6 mm, 5 μ . Flow: 0.8 ml/min. Temp: 25°C. Eluent: H₂O : MeOH 30:70 + 1% TFA. Detection: UV 220 nm. R_t of **10f**: 6.02. R_t of **11f**: 5.49. Minor diastereomer **11f** was recognized using a MS (ESI) detector.

Oil. $R_f = 0.35$ (PE / AcOEt 1 : 1) (R_f of **11f** = 0.46). [α]_D = -42.4 (c 1.0, CHCl₃). ¹H NMR (300 MHz, d6-DMSO, 90 °C): δ = 7.80 (1 H, s, N*H*), 7.48 (1 H, d, J = 6.6 Hz), 7.31-7.14 (6 H, m), 6.99 (1 H, td, J = 7.5, 1.2 Hz), 6.91 (1 H, dd, J = 8.1, 1.1 Hz), 5.62 (1 H, s, CHC=O), 4.76 (1 H, broad d, J = 12.3 Hz, CHHO), 4.37 (1 H, dd, J = 13.2, 4.8 Hz, CHHO), 4.31 (2 H, broad t, J = 5.4 Hz, CH₂NH), 4.29 (1 H, J = 14.2 Hz, CHHOMe), 4.11 (1 H, broad s, CHN), 4.01 (1 H, J = 14.2 Hz, CHHOMe), 3.31 (3 H, s, CH₃O), 1.59 (d of heptuplets, J = 9.9, 6.6 Hz, CH(CH₃)₃), 0.88 (3 H, d, J = 6.6 Hz, CH₃CHCH₃), 0.75 (3 H, d, J = 6.8 Hz, CH₃CHCH₃). ¹³C NMR (75 MHz, d6-DMSO, 90 °C): δ = 169.3 (x2) (C=O), 156.0, 138.7, 122.3 (quat.), 131.6, 129.5, 127.5 (x2), 126.4 (x2), 126.0, 121.3, 119.1 (aromatic CH), 70.6 (CH₂OMe), 68.1 (broad) (CH₂O), 62.0 (CHC=O), 59.8 (broad) (CHN), 57.8 (OCH₃), 42.3 (CH₂NH), 28.5 (CH(CH₃)₂), 19.3, 18.8 (CH₃). I.R. (ATR): v_{max} 3314, 3062, 3031, 2964, 2931, 2873, 2825, 1653, 1605, 1580, 1516, 1493, 1453, 1424, 1390, 1367, 1320, 1248, 1218, 1198, 1156, 1124, 1107, 1080, 1028, 965, 948, 860, 838, 750, 731, 698, 640, 605 cm⁻¹. HRMS (ESI+): found 397.2135 [Calcd for C₂₃H₂₉N₂O₄⁺ (M + H)⁺ 397.2127].

(3S,5R)-N-(Cyclohexyl)-3-iso-propyl-4-(phenylacetyl)-2,3,4,5-tetrahydrobenzo[f][1,4]oxazepine-5-carboxamide 10g.



It was prepared in 79% yield from (*S*)-8c, phenylacetic acid and cyclohexyl isocyanide, following the typical procedure described for **10a**. However, in this case, the Ugi-Joullié step reaction was carried out at rt for 64 h. Chromatography was carried out with PE / CH_2CI_2 / AcOEt 3 : 1 . 1, affording a complete separation of the two diastereomers. The overall yield was 82%. The d.r. was = 96 : 4 by HPLC of the crude product. Conditions: column Luna C8 150 x 4.6 mm, 5 μ . Flow: 0.8 ml/min. Temp: 25°C. Eluent: H₂O : MeOH 30:70. Detection: UV 220 nm. R_t of **10g**: 15.14. R_t of **11g**: 13.89. Minor diastereomer **11g** was recognized using a MS (ESI) detector.

White solid. $R_f = 0.42$ (PE / CH₂Cl₂ / AcOEt 3 : 1 . 1). [α]_D = -72.8 (c 1.0, CHCl₃). M.p.: 65.0-67.2 °C. ¹H NMR (300 MHz, d6-DMSO, 120 °C): δ = 7.39 (1 H, d, J = 7.8 Hz), 7.33-7.19 (6 H, m), 6.98 (1 H, t, J = 7.5 Hz), 6.89 (1 H, d, J = 8.1), 6.78 (1 H, broad s, NH), 5.61 (1 H, s, CHC=O), 4.63 (1 H, dd, J = 12.5, 3.0 Hz, CHHO), 4.35 (1 H, dd, J = 12.5, 5.5 Hz, CHHO), 4.20 (1 H, broad s, CHN), 3.86 (1 H, d, J = 15.6 Hz, CHHPh), 3.71 (1 H, d, J = 15.6 Hz, CHHPh), 3.63 (1 H, broad m, CHNH), 1.83-1.48 (6 H, m, CH(CH₃)₂ and CH₂ cyclohexyl), 1.38-1.10 (5 H, m, CH₂ cyclohexyl), 0.89 (3 H, d, J = 6.6 Hz, CH₃CHCH₃), 0.75 (3 H, d, J = 6.8 Hz, CH₃CHCH₃). ¹³C NMR (75 MHz, d6-DMSO, 90 °C): δ = 171.3, 168.1 (C=O), 156.0, 135.4, 123.3 (quat.), 130.8, 129.3, 128.7 (x2), 127.6 (x2), 125.8, 121.4, 119.3 (aromatic CH), 68.6 (CH₂O), 62.5 (broad) (CHC=O), 60.4 (very broad) (CHN), 47.6 (CHNH), 39.9 (CH₂Ph), 31.43, 31.37, 24.6, 23.7, 23.6 (CH₂ cyclohexyl), 28.5 (CH(CH₃)₂), 19.3, 18.8 (CH₃). I.R. (ATR): v_{max} 3418, 3320, 3064, 3031, 2930, 2854, 1647, 1604, 1581, 1510, 1493, 1450, 1408, 1387, 1367, 1348, 1319, 1248, 1217, 1153, 1113, 1080, 1031, 949, 891, 855, 810, 795, 757, 747, 719, 696, 641 cm⁻¹. HRMS (ESI+): found 435.2639 [Calcd for C₂₇H₃₅N₂O₃+ (M + H)+ 435.2648].

(3S,5R)-4-(Acetyl)-N-(tert-butyl)-3-iso-propyl-2,3,4,5-tetrahydrobenzo[f][1,4]oxazepine-5-carboxamide 10h.



It was prepared in 52% yield from (S)-8c, acetic acid and *tert*-butyl isocyanide, following the typical procedure described for **10a**. The reaction was in this case incomplete and also 6% of starting material was recovered. Chromatography was carried out with CH_2Cl_2 / AcOEt 4 : 1, affording a complete separation of the two diastereomers. The minor diastereomer **11h** was not detected by HPLC-MS or NMR. The d.r. was thus supposed to be > 97 : 3. HPLC conditions: C6-Phenyl column 150 x 3 mm, 3 µm. Flow: 0.38 ml/min. Temp: 26°C. Eluent: H_2O : CH_3CN 60:40. Detection: UV 220 nm. R_t of **10h**: 11.66.

White solid. $R_f = 0.28 (CH_2CI_2 / AcOEt 4 : 1)$. $[\alpha]_D = -9.4 (c 0.9, CHCI_3)$. M.p.: 104.5-107.8 °C. ¹H NMR (300 MHz, d6-DMSO, 120 °C): $\delta = 7.46 (1 H, dd, J = 7.6, 1.4 Hz)$, 7.25 (1 H, td, J = 8.0, 1.6), 6.98 (1 H, td, J = 7.5, 1.1 Hz), 6.90 (1 H, dd, J = 8.1, 1.0), 6.62 (1 H, broad s, NH), 5.49 (1 H, s, CHC=O), 4.68 (1 H, dd, J = 12.5, 3.7 Hz, CHHO), 4.35 (1 H, dd, J = 12.5, 5.8 Hz, CHHO), 4.12 (1 H, broad s, CHN), 2.10 (3 H, s, CH_3C=O), 1.71 (1 H, d of heptuplets, J = 9.9, 6.6 Hz, CH(CH_3)_3), 1.26 (9 H, s, C(CH_3)_3), 0.90 (3 H, d, J = 6.6 Hz, CH_3CHCH_3), 0.79 (3 H, d, J = 6.8 Hz, CH_3CHCH_3). ¹³C NMR (75 MHz, d6-DMSO, 90 °C): $\delta = 170.3$, 168.4 (C=O), 155.9, 123.3 (quat.), 130.7, 129.1, 121.2, 119.2 (aromatic CH), 68.4 (CH₂O), 63.3 (broad) (CHC=O), 59.9 (very broad, barely visible) (CHN), 50.1 (CNH), 28.4 (CH(CH_3)_2), 27.8 (C(CH_3)_3), 21.7 (CH_3C=O), 19.3, 18.9 (CH_3). I.R. (ATR): vmax 3332, 2965, 2933, 2874, 1980, 1668, 1640, 1607, 1583, 1529, 1496, 1451, 1413, 1392, 1365, 1346, 1331, 1303, 1289, 1266, 1250, 1218, 1116, 1088, 1041, 979, 945, 900, 890, 858, 841, 812, 755, 746, 689, 654, 631, 621 cm⁻¹. HRMS (ESI+): found 333.2176 [Calcd for C₁₉H₂₉N₂O₃⁺ (M + H)⁺ 333.2178].

(3R,5S)-4-(3-Bromobenzoyl)-N-(cyclohexyl)-3-phenyl-2,3,4,5-tetrahydrobenzo[f][1,4]oxazepine-5-carboxamide 10i.



It was prepared in 86% yield from (*R*)-8d, 3-bromoacetic acid and cyclohexyl isocyanide, following the typical procedure described for **10a**. However, in this case, 1.3 equivalents of isocyanide were used. Chromatography was carried out with PE/AcOEt 3:1 \rightarrow AcOEt/MeOH 95:5, affording a complete separation of the two diastereomers. The minor diastereomer **11** was not isolated. An overall yield 89% was calculated on the basis of d.r. The d.r. was = 97 : 3 by HPLC of the crude product. Conditions: column Luna C8 150 x 4.6 mm, 5 μ . Flow: 0.8 ml/min. Temp: 25°C. Eluent: H₂O : MeOH 30:70 + 1% TFA. Detection: UV 220 nm. R_t of **10i**: 10.93. R_t of **11i**: 11.72. Minor diastereomer **11i** was recognized using a MS (ESI) detector.

White solid. $R_f = 0.36$ (PE/AcOEt 3 : 1). $[\alpha]_D = -98.5$ (c 1.0, CHCl₃). M.p.: 229.0-231.5 °C. ¹H NMR (300 MHz, d6-DMSO, 90 °C): $\delta = 7.61$ (1 H, d, J = 7.8 Hz), 7.41-7.22 (4 H, m), 7.21-7.05 (6 H, m), 6.95 (1 H, td, J = 7.4, 0.9 Hz), 6.71 (1 H, d, J = 8.1), 6.65 (1 H, broad d, J = 6.6 Hz, NH), 5.78 (1 H, broad s, CHC=O), 5.44 (1 H, broad s, CHPh), 5.03 (1 H, dd, J = 13.6, 1.4 Hz, CHHO), 4.53 (1 H, dd, J = 13.6, 4.0 Hz, CHHO), 3.70-3.55 (1 H, broad m, CHNH), 1.82-1.47 (5 H, m, CH₂ cyclohexyl), 1.40-1.05 (5 H, m, CH₂ cyclohexyl). ¹³C NMR (75 MHz, d6-DMSO, 90 °C): $\delta = 169.7$, 168.5 (C=O), 155.3, 138.8, 138.1, 121.0, 120.3 (quat.), 132.1, 131.8, 130.0, 129.4, 128.4, 127.2 (x2), 125.9, 125.6 (x2), 124.6, 120.9, 118.9 (aromatic CH), 68.5 (CH₂O), 64.1 (broad) (CHC=O), 60.5 (broad) (CHN), 48.1 (CHNH), 31.1 (x2), 24.5, 23.8, 23.7 (CH₂ cyclohexyl). I.R. (ATR): v_{max} 3320, 3063, 3028, 2924, 2850, 1663, 1650, 1610, 1581, 1563, 1512, 1490, 1464, 1449, 1414, 1389, 1352, 1332, 1311, 1275, 1249, 1219, 1178, 1148, 1115, 1080, 1046, 1027, 998, 977, 956, 917, 903, 894, 879, 866, 854, 823, 798, 777, 753, 741, 731, 691, 647 cm⁻¹. HRMS (ESI+): found 533.1429 [Calcd for C₂₉H₃₀BrN₂O₃+ (M + H)⁺ 533.1440].

(3S,5R)-4-(3-Acetyl)-7-bromo-N-(tert-butyl)-3-methyl-2,3,4,5-tetrahydrobenzo[f][1,4]oxazepine-5-carboxamide 10j.



It was prepared in 68% yield from (S)-8e, acetic acid and *tert*-butyl isocyanide, following the typical procedure described for **10a**. However, in this case, during Ugi-Joullié reaction, after 24 h, additional 0.2 equivalents of isocyanide were added. Chromatography was carried out with PE/AcOEt 2:1 \rightarrow 1:1, affording a complete separation of the two diastereomers. The minor diastereomer **11j** was not isolated. An overall yield 71% was calculated on the basis of d.r. The d.r. was = 95 : 5 by HPLC of the crude product. Conditions: column Luna C8 150 x 4.6 mm, 5 μ . Flow: 0.8 ml/min. Temp: 25°C. Eluent: H₂O : MeOH 50:50 + 1% CF₃CO₂H. Detection: UV 220 nm. R_t of **10j**: 13.59. R_t of **11j**: 12.51. Minor diastereomer **11j** was recognized using a MS (ESI) detector.

White solid. $R_f = 0.30$ (PE/AcOEt 2 : 1). R_f of **11** = 0.42. [α]_D = -46.2 (c 1.0, CHCl₃). M.p.: 180.3-181.8 °C. ¹H NMR (300 MHz, d6-DMSO, 90 °C): δ = 7.74 (1 H, d, J = 2.1 Hz), 7.39 (1 H, dd, J = 8.7, 2.5 Hz), 6.85 (1 H, d, J = 8.7 Hz), 6.62 (1 H, broad s, NH), 5.62 (1 H, broad s, CHC=O), 4.85 (1 H, d, J = 13.0 Hz, CHHO), 4.45 (1 H, broad quintuplet, J = 5.9 Hz, CHCH₃), 4.18 (1 H, dd, J = 13.0, 4.9 Hz, CHHO), 2.12 (3 H, s, CH₃C=O), 1.25 (9 H, s, C(CH₃)₃), 1.10 (3 H, d, J = 6.9 Hz, CH₃CH). ¹³C NMR (75 MHz, d6-DMSO, 90 °C): δ = 169.4, 168.6 (C=O), 154.9, 123.3, 111.6 (quat.), 134.3, 131.9, 120.7 (aromatic CH), 69.5 (CH₂O), 61.6 (broad) (CHC=O), 51.4 (broad) (CHN), 50.3 (CNH), 27.8 (C(CH₃)₃, 21.1 (CH₃=O), 16.2 (CH₃CH). I.R. (ATR): v_{max} 3413, 3336, 2969, 2930, 1685, 1630, 1570, 1514, 1487, 1449, 1418, 1389, 1369, 1335, 1319, 1299, 1281, 1267, 1254, 1221, 1179, 1146, 1125, 1106, 1090, 1066, 1042, 1000, 962, 929, 903, 877, 823, 766, 740, 709, 689, 659, 624 cm⁻¹.HRMS (ESI+): found 383.0974 [Calcd for C₁₇H₂₄BrN₂O₃⁺ (M + H)⁺ 383.0970].

(3R,5S)-4-(3-Acetyl)-7-bromo-N-(tert-butyl)-3-methyl-2,3,4,5-tetrahydrobenzo[f][1,4]oxazepine-5-carboxamide ent-10j.

It was prepared exactly as for the enantiomer **10***j*. $[\alpha]_D = +44.9$ (c 1.0, CHCl₃). All the other data were identical to those of the enantiomer.





It was prepared in 60% yield from (S)-8e, 5-chloro-2-thienoic acid and isopropyl isocyanide, following the typical procedure described for **10a**. However, in this case, for the Ugi-Joullié reaction, 1.4 equivalents of isocyanide were used. Chromatography was carried out with PE/AcOEt 4:1 \rightarrow 3:1. The minor diastereomer **11k** was not isolated. The overall yield was calculated to be 63% on the basis of d.r. The d.r. was = 95 : 5 by HPLC of the crude product. Conditions: column Luna C8 150 x 4.6 mm, 5 μ . Flow: 0.8 ml/min. Temp: 25°C. Eluent: H₂O : MeOH 50:50. Detection: UV 220 nm. R_t of **10k**: 16.27. R_t of **11k**: 15.68. Minor diastereomer **11k** was recognized using a MS (ESI) detector.

White solid. $R_f = 0.33$ (PE/AcOEt 4 : 1). [α]_D = -71.3 (c 1.0, CHCl₃). M.p.: 166.8-169.0 °C. ¹H NMR (300 MHz, CDCl₃, 20 °C): δ = 7.41 (1 H, dd, J = 8.7, 2.4 Hz), 7.22 (1 H, d, J = 2.3 Hz), 7.17 (1 H, d, J = 3.9 Hz), 6.90-6.85 (2 H, m), 5.63 (1 H, broad s, CHC=O), 5.49 (1 H, broad s, NH), 4.63 (1 H, d, J = 13.5 Hz, CHHO), 4.70-4.58 (1 H, m, CHCH₃), 1.31 (3 H, d, J = 6.8 Hz, CH₃CH), 1.12 (3 H, d, J = 6.6 Hz, CH₃CHCH₃), 1.10 (3 H, d, J = 6.6 Hz, CH₃CHCH₃). ¹³C NMR (75 MHz, CDCl₃, 20 °C): δ = 168.8, 164.6 (C=O), 155.5, 135.7, 134.6, 122.1, 113.7 (quat.), 134.1, 133.8, 128.6, 126.2, 121.9 (aromatic CH), 70.2 (CH₂O), 64.4 (CHC=O), 53.9 (CHN), 42.5 (CHNH), 22.4, 22.3 (CH₃)₂CH, 16.8 (CH₃CH). I.R. (ATR): v_{max} 3283, 3094, 2977, 2933, 2876, 1657, 1623, 1514, 1486, 1434, 1386, 1344, 1316, 1277, 1251, 1220, 1186, 1145, 1129, 1107, 1059, 998, 951, 933, 874, 859, 840, 771, 743, 688 cm⁻¹.HRMS (ESI+): found 471.0156 [Calcd for C₁₉H₂₁BrClN₂O₃S⁺ (M + H)⁺ 471.0145].



(3S,5R)-7-Bromo-4-(5-chlorothiophene-2-carbonyl)-N-isopropyl-3-methyl-2,3,4,5-tetrahydrobenzo[f][1,4]oxazepine-5-carboxamide 101.

Acetal 8e (395.4 mg, 976 µmol) was dissolved in CH₂Cl₂ (3.7 mL) and treated with 37% agueous HCI (808 µL, 9.67 mmol). The mixture was stirred at rt for 5 h. Then it was diluted with CH₂Cl₂, and cautiously treated with a solution of Na₂CO₃ (765 mg, 7.21 mmol) in water (16.0 mL). After checking that pH > 9, the two phases were separated, washed with brine, and evaporated to dryness. The resulting crude imine 3e (yellow oil)(197.5 mg) was taken up in dry methanol (4.75 mL), and treated with tert-butyl isocyanide (131 µl, 1.15 mmol) and with 3-((tert-butyldimethylsilyl)oxy)propanoic acid (252 mg, 1.15 mmol). The mixture was stirred at rt for 24 h. Then further tert-butyl isocyanide (21 µL, 0.25 mmol) were added. After strring for 24 h more, the mixture was evaporated to dryness and chromatographed (PE / AcOEt 5:1) to give pure 10I (284 mg, 54%). The minor diastereome 11I was not isolated. The ratio 101 : 111 was determined by HPLC on the crude product and resulted = 95:5. HPLC conditions: column Luna C8 150 x 4.6 mm, 5 µ. Flow: 0.8 ml/min. Temp: 25°C. Eluent: H₂O : MeOH 50:50 + 1% CF₃CO₂H up to 20 minutes. Then 100% MeOH + 1% CF₃CO₂H. Detection: UV 220 nm. R_f of **10I**: 20.82. R_f of 11I: 20.45. Minor diastereomer 11I was recognized using a MS (ESI) detector. The overall yield was calculated to be 57% on the basis of d.r. Oil. $R_f = 0.33$ (PE/AcOEt 5 : 1). [a]_D = -23.0 (c 1.0, CHCl₃). ¹H NMR (300 MHz, d6-DMSO, 90 °C): $\delta = 7.72$ (1 H, d, J = 2.5 Hz), 7.39 (1 H, dd, J = 8.7, 2.5) Hz), 6.85 (1 H, d, J = 8.7 Hz), 6.57 (1 H, broad s, NH), 5.64 (1 H, broad s, CHC=O), 4.82 (1 H, d, J = 13.1 Hz, CHHO), 4.47 (1 H, broad guintuplet, J = 6.1 Hz, CHCH₃), 4.18 (1 H, dd, J = 13.1, 4.9 Hz, CHHO), 3.64 (2 H, t, J = 6.4 Hz, CH₂OSi), 2.55 (1 H, dt, J = 15.6, 7.2 Hz, CHHC=O), 2.37 (1 H, dt, J = 15.6 7.2 Hz, CHHC=O), 1.77 (2 H, quintuplet, J = 6.9 Hz, CH₂CH₂OSi), 1.25 (9 H, s, C(CH₃)₃), 1.11 (3 H, d, J = 6.8 Hz, CH₃CH), 0.89 (9 H, s, SiC(CH₃)₃), 0.05 (6 H, s, (CH₃)₂Si). ¹³C NMR (75 MHz, d6-DMSO, 90 °C): δ = 171.7, 168.6 (C=O), 155.0, 123.4, 111.6 (quat.), 134.2, 131.9, 120.6 (aromatic CH), 69.4 (CH₂O), 61.4 (CH₂OSi), 61.2 (broad) (CHC=O), 51.1 (CHN), 50.2 (CNH), 28.4 (CH₂C=O), 27.8 (C(CH₃)₃, CH₂CH₂OSi), 25.3 (SiC(CH₃)₃), 17.3 (SiC(CH₃)₃)), 16.2 (CH₃CH), -5.9 ((CH₃)₂Si). I.R. (ATR): v_{max} 3418, 3344, 2956, 2929, 2857, 1686, 1625, 1538, 1507, 1486, 1472, 1462, 1454, 1412, 1393, 1365, 1323, 1273, 1250, 1235, 1181, 1124, 1100, 1060, 1005, 968, 939, 834, 775, 752, 679, 664, 626, 612 cm⁻¹.HRMS (ESI+): found 541.2103 [Calcd for

 $C_{25}H_{42}BrN_2O_4Si^+$ (M + H)⁺ 541.2097].

HPLC conditions for analysis of the diastereomeric ratio of 10m and 11m.



Column Luna C8 150 x 4.6 mm, 5 μ. Flow: 0.8 ml/min. Temp: 25°C. Eluent: H₂O : MeOH 90:10 + 1% CF₃CO₂H up to 20 minutes. Then 100% MeOH + 1% CF₃CO₂H. Detection: UV 220 nm. R_t of **10m**: 17.93. R_t of **11m**: 18.58. The ratio was 90 : 10 (**11m** . **10m**).

HPLC conditions for analysis of the enantiomeric composition of 10j, 11j, ent-10j, and ent-11j.

Column: Chiral Pak AD 250 x 4, 6 μ m. Flow: 0.8 ml/min. Temp: 25°C. Eluent: hexane : isopropanol 97:3. Detection: DAD (chromatograms are reported at 220 nm). R_t of **10***j*: 28.24. R_t of **11***j*: 24.31. R_t of *ent-10j*: 32.27 R_t of *ent-11j*: 29.43.

4-((tert-Butyldimethylsilyl)oxy)butanoic acid 12

 γ -Butyrolactone (0.763 mL, 10.0 mmol) was dissolved in a solution of NaOH (451 mg, 11.3 mmol) in MeOH (10.0 mL). The mixture was stirred at room temperature for 2 h. It was then evaporated and the crude product (1.2043 g) was taken up in dry DMF (20.0 mL). The mixture was cooled to 0°C and then *tert*-butyldimethylchlorosilane (3.299 g, 21.9 mmol) and imidazole (2.257 g, 33.1 mmol) were added. The mixture was stirred overnight at room temperature. It was diluted with PE / Et₂O 1:1 and washed with water. The organic phases were dried over Na₂SO₄, filtered and evaporated under reduced pressure at low temperature (the rotavapor bath was kept near 0°C). The resulting colourless oil (4.206 g) was taken up with a mixture of water (30.0 mL), MeOH (10.0 mL), THF (10.0 mL), and treated with K₂CO₃ (2.767g, 20.0 mmol). The resulting mixture was stirred at room temperature overnight. It was then concentrated to remove most MeOH and THF, diluted with Et₂O and extracted with water. The aqueous phase was acidified to pH = 4 with HCl 1M and extracted with Et₂O. The organic phases were dries over Na₂SO₄, filtered and evaporated product (1.4773 g, 68%) was obtained as a colourless oil and used as such without any purification.

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HPLC and NMR determination of enantiomeric excesses and diastereomeric ratios

HPLC of crude 10a + 11a after Ugi-Joullié reaction



HPLC of crude 10a + 11a after epimerization promoted by KOH



Sorted By		:	Signal	
Multiplier			1.0000	
Dilution		:	1.0000	
Use Multiplier	&	Dilution	Factor with	ISTDs

Peak #	RetTime [min]	Туре	Width [min]	Aı mAU	ea *s	Heig [mAU	ght]	Area %
1	13.704	MF	0.2605	4760.	31543	304.5	53952	32.6104
2	14.485	FM	0.2677	9837.	21387	612.3	36823	67.3896



HPLC of crude 10b + 11b



Area Percent Report

Sorted By		:	Signal	
Multiplier		:	1.0000	
Dilution		:	1.0000	
Use Multiplier	&	Dilution	Factor with	ISTDs

Signal 1: VWD1 A, Wavelength=220 nm

Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	20.427	MF	0.3592	333.90381	15.49456	5.5446
2	21.249	FM	0.3614	5688.26855	262.29486	94.4554



HPLC of crude 10c + 11c



Area Percent H	Report

Sorted By		:	Sign	nal	
Multiplier		:	1.00	000	
Dilution		:	1.00	000	
Use Multiplier	&	Dilution	Factor	with	ISTDs

Peak	RetTime	Туре	Width	A	rea	Hei	ght	Area
#	[min]		[min]	mAU	*s	[mA0		*
1 2	20.550 21.681	BV VB	0.4486 0.4790	3527 212	.66724 .34320	122. 6.	85065 94468	94.3224 5.6776
Total	s:			3740	.01044	129.	79533	



HPLC of crude 10d + 11d



Area Percent Report

Sorted By		:	Signal	
Multiplier		:	1.0000	
Dilution		:	1.0000	
Use Multiplier	&	Dilution	Factor with	ISTDs

Signal 1: VWD1 A, Wavelength=220 nm

Peak #	RetTime [min]	Туре	Width [min]	Area mAU *s	Height [mAU]	Area %
1	13.334	MF	0.2392	4715.04932	328.57242	92.8950
2	14.003	FM	0.2420	360.62830	24.84063	7.1050



HPLC of crude 10e + 11e



Sorted By		:	Signal	
Multiplier		:	1.0000	
Dilution		:	1.0000	
Use Multiplier	&	Dilution	Factor with	ISTDs

Peak	RetTime	Type	Width	Area	Height	Area
#	[min]		[min]	mAU *s	[mAU]	olo
1	16.232	MM	0.2911	172.34360	9.86690	4.0155
2	18.247	MM	0.3363	4119.61475	204.16066	95.9845



HPLC of crude 10f + 11f



Peak	RetTime	Type	Width	Ar	rea	Heig	ght	Area
#	[min]		[min]	mAU	*s	[mAU	1	망
1	5.486	FM	0.1689	185.	27025	18.2	28515	5.7318
2	6.020	MF	0.1627	3047.	03931	312.0	5853	94.2682



HPLC of crude 10g + 11g



Area Percent Report

Sorted By		:	Sigr	nal	
Multiplier			1.00	000	
Dilution		:	1.00	000	
Use Multiplier	&	Dilution	Factor	with	ISTDs

Peak #	RetTime [min]	Туре	Width [min]	Area mAU *s	Height [mAU]	Area %
1	13.886	PP	0.2689	159.05804	9.19113	3.5708
2	15.137	VB	0.2858	4295.31494	235.29900	96.4292



HPLC of crude 10h



Area Percent Report

Sorted By	:	Signal	
Multiplier	:	1.0000	
Dilution	:	1.0000	
Use Multiplier	& Dilution	Factor with	ISTDs

Signal 1: VWD1 A, Wavelength=220 nm

Peak	RetTime	Type	Width	Aı	rea	Heig	ght	Area
#	[min]		[min]	mAU	*s	[mAU]	8
1	11.661	VV	0.2277	7715.	28467	505.1	L1163	100.0000



HPLC of crude 10i + 11i





HPLC of crude 10j + 11j



Area	Percent	Report	

Sorted By	:	Signal	
Multiplier	:	1.0000	
Dilution	:	1.0000	
Use Multiplier	& Dilution	Factor with	ISTDs

Signal 1: VWD1 A, Wavelength=220 nm

Peak #	RetTime [min]	Туре	Width [min]	Area mAU *s	Height [mAU]	Area %
1	12.507	MM	0.1441	237.96765	27.53019	4.8268
2	13.586	MM	0.1313	4692.12158	595.68738	95.1732



HPLC of crude 10k + 11k



Area Percent Report

Sorted By	:	Signal	
Multiplier	:	1.0000	
Dilution	:	1.0000	
Jse Multiplier	& Dilution	Factor with	ISTDs

Peak	RetTime	Type	Width	Ar	ea	Heig	ght	Area
#	[min]		[min]	mAU	*s	[mAU]	olo
1	15.676	MM	0.1331	201.	78844	25.2	26794	5.2093
2	16.267	MM	0.1256	3671.	84375	487.4	12706	94.7907



HPLC of crude 10I + 11I





HPLC of crude 11m + 10m (after reaction of 10l with TBAF)



Area Percent Report

Sorted By	:	Signal	
Aultiplier		1.0000	
Dilution	:	1.0000	
lse Multiplier &	Dilution	Factor with	ISTD

Peak	RetTime	Type	Width	Area	Height	Area
#	[min]		[min]	mAU *s	[mAU]	R
1	17.929	MM	0.1190	3121.02319	437.27203	90.2711
2	18.577	MM	0.1171	336.36569	47.86787	9.7289



HPLC of chiral stationary phase of crude $\mathbf{11j} + \mathbf{10j}$ after UgiJoullié reaction



Area Percent Report

Sorted By		:	Sigr	nal	
Multiplier		:	1.00	000	
Dilution		:	1.00	000	
Use Multiplier	&	Dilution	Factor	with	ISTDs

Signal 1: DAD1 A, Sig=220,8 Ref=450,50

Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	8
1	24.459	MM	0.8556	210.09351	4.09243	5.3066
2	28.248	MM	0.9804	3749.00391	63.73133	94.6934



HPLC of chiral stationary phase of **11j** + **10j** after epimerization (partially enriched in **11j** by chromatography).



Area Percent Report

Sorted By	:	Signal	
Multiplier	:	1.0000	
Dilution	:	1.0000	
Use Multiplier &	Dilution	Factor with ISTDs	\$

Signal 1: DAD1 A, Sig=220,8 Ref=450,50

Peak	RetTime	Type	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	90
1	24.314	MM	0.8527	5750.31738	112.39552	85.2836
2	28.235	MM	0.9750	992.26495	16.96245	14.7164





HPLC of chiral stationary phase of pure *ent-10j* isolated by chromatography after Ugi-Joullié reaction.

HPLC of chiral stationary phase of *ent***-11j** isolated by prep TLC after Ugi-Joullié reaction (impure of some starting imine **3e**).



Conformational analysis of compounds 13 and 14



This study was carried out Using the commercial software ChemBio3D Ultra and the MOPAC (PM3) interface. First of all we have searched for all the local minima of the parent compound

This parent compound gives rise to three conformations with similar energy. One of them resembles a chair, while the second one resembles a twist and the third one a boat. Thus, here we conventionally adopt these three terms. Obviously, each of these three conformations exist in two possible enantiomeric forms, which are in equilibrium.

A notable difference between them is that in the chair and twist conformations the axial hydrogen at C-5 is located on the same side of the median plane than C-2 and C-3. On the contrary, in the boat, they are opposite.



When passing to the real compound, the two conformations of each type become diastereomeric. Thus, we must consider 6 different conformations for each diastereomer. We have searched for the absolute minimum for each of them. According to the heat of formation found, the ones evidenced in yellow are more likely

	Conformations of cis compound (14)								
Number	Туре	Heat of formation (Kcal/mol)	Position of H-5	Position of H-3	Distance between H-5 and H-3 (Å)	Dihedral angles between H-2 and H-3			
1	Chair Chair	<mark>-60.49</mark>	<mark>axial</mark>	<mark>axial</mark>	<mark>2.31</mark>	<mark>178, 61</mark>			
2	Twist	<mark>-59.48</mark>	axial	axial	<mark>2.45</mark>	<mark>91, 29</mark>			

<mark>3</mark>	Boat	<mark>-59.39</mark>	equatorial	<mark>axial</mark>	<mark>2.50</mark>	<mark>153, 32</mark>
4	Chair	-57.76	equatorial	equatorial	4.09	68, 52
5	Twist	-57.15	equatorial	equatorial	4.11	175, 55
6	Boat	is converted to other conformations	axial	equatorial	-	-

	Conformations of trans compound (13)									
Number	Туре	Heat of formation (Kcal/mol)	Position of H-5	Position of H-3	Distance between H-5 and H-3 (Å)	Dihedral angles between H-2 and H-3				
<mark>1</mark>	Chair Chair	<mark>-60.06</mark>	equatorial	<mark>axial</mark>	<mark>3.61</mark>	<mark>67, 53</mark>				
2	Chair Chair	<mark>-59.79</mark>	axial	equatorial	<mark>3.68</mark>	<mark>174, 65</mark>				
<mark>3</mark>	Twist	<mark>-59.55</mark>	<mark>axial</mark>	equatorial	<mark>3.61</mark>	<mark>149, 28</mark>				
4	Twist	-58.73	equatorial	axial	3.73	97, 24				
5	Boat	is converted to other conformations	axial	axial	-	-				
6	Boat	is converted to other conformations	equatorial	equatorial	-	-				

From these results, it is clear that in general, as expected, conformations that place H-3 and/or H-5 in axial position are favoured. Obviously this meas that the substituents at C-5 and/or at C-3 are equatorial. Therefore, the lower energy is obtained for the chair conformation of the *cis* compound, where both hydrogens are axial.

In any case, the differences in heat of formation among the best three conformations of both isomers is too scarce to allow us to draw conclusion on the preferred one.

However, from the data in the tables, it is evident that no conformation of *trans* isomer may be able to give a nOe between H-3 and H-5. On the contrary, the best three conformations of *cis* compound can. NOESY experiments have shown that only the amine derived from the minor (kinetically unfavoured) diastereomer of the Ugi-Joullié reaction gives this nOe. We thus think that this is a proof of the onfigurational assignment.

Unfortunately, we are not able to measure precisely both J between H-3 and H-2, because one of the diastereotopic H-2 falls very near to H-3 in both isomers. These coupling constants could have given an useful information reagrding the stereochemical equilibria, because, as can be seen fro the tables, the dihedral angles are quite different among them.

Conformational analysis of imine 3e

Imine **3e** exists in just two half-chair conformations depicted below, The one on the left has the methyl substituent in axial position. The conformation on the right is more stable by about 0.80 Kcol/mol, according to MOPAC - PM3. In both conformations the C=N bond is coplanar to the benzene ring, and the face *trans* to the substituent is less encoumbered.



Conformational analysis of amides 10j-11j

This study was carried out Using the commercial software ChemBio3D Ultra and the MOPAC (PM3) interface.

As for the amines **13** and **14**, there are three types of conformations. One of them resembles a chair, while the second one resembles a twist and the third one a boat. Thus, here we conventionally adopt these three terms. Obviously, each of these three conformations exist in two possible diastereomeric forms, which are in equilibrium.

For the *cis* compound **11j**, two conformations clearly emerge as the most stable ones:

- Conformation A is a twist where the CONHtBu group is axial and the methyl equatorial. Heat of formation = -99.50 Kcal/mol
- Conformation **B** is a half-chair with the two substituents in axial position. Heat of formation = -98.86 Kcal/mol



CONFORMATIONS OF CIS COMPOUND 11j

For the *trans* compound **10j**, two conformations clearly emerge as the most stable ones:

- Conformation **A** is a twist where the CONH*t*Bu group is axial and the methyl axial. Heat of formation = -98.45 Kcal/mol
- Conformation **B** is a half-chair where the CONH*t*Bu group is axial and the methyl equatorial. Heat of formation = –98.29 Kcal/mol



CONFORMATIONS OF TRANS COMPOUND 10j

From these calculations, it is clear that the CONH*t*Bu group prefers an axial position. The *cis* compound seem to be more stable, as demonstrated experimentally by the equilibration under base catalysis.

¹H and ¹³C NMR spectra of all new compounds





























































