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Supporting Information

Fluorescence Lifetime Imaging of Upper Gastrointestinal pH $In\ Vivo$ with a Lanthanide Based Near-Infrared τ Probe

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General materials and methods

Unless otherwise stated, all reactions were performed under an inert atmosphere of nitrogen. UV-vis spectra were recorded on an Agilent 8453 UV-vis Spectrometer equipped with an Agilent 89090A Thermostat (±0.1 °C) at 25 °C. IR spectra were recorded on Bruker Tensor 27 FT-IR or PE Spectrum Spotlight 200 FT-IR Microscopy. Mass spectra were recorded on Bruker APEX IV FT-ICR Mass Spectrometer (ESI) or Solarix XR Fourier Transform Ion Cyclotron Resonance Mass Spectrometer. Simulated mass spectra were obtained at the website http://www.chemcalc.org. NMR spectra were recorded on Bruker ARX400 400MHz or AVANCE III 500MHz NMR Spectrophotometer. Transmission electron microscopy was obtained in JEOL JEM-2100F Field-emission High Resolution Transmission Electron Microscope. Dynamic light scattering was carried out in ALV/DLS/SLS-5022F Laser Light Scattering Spectrometer. Transient absorption spectra and triplet excited state decay dynamics were recorded on an Edinburgh LP920 spectrometer combined with OPO laser excitation pulse (10 Hz, 2 mJ pulse⁻¹). Phosphorescence spectra were recorded on frozen glasses of solutions of Gd³⁺ complexes (MeOH/EtOH 1:1, v/v) using a dewar cuvette filled with liquid N_2 (T = 77 K). HeLa cells were obtained from Peking University Health Science Center. All animal procedures were approved by the Institutional Animal Care and Use Committee of Fudan University. For the optical measurements in liquid solution, spectroscopic-grade dimethyl sulfoxide was used as purchased from Alfa-Aesar. Methyl *p*-formylbenzoate, anhydrous 1,2,4-trichlorobenzene (TCB), were purchased from J&K Scientific and used as received. Anhydrous CH₂Cl₂ was distilled from calcium hydride and H₂O were integral. obtained Milli-Q Sodium [(cyclopentadienyl)tris(di(methyl-d₃)phosphito) cobaltate] (NaL_{OCD3}) (D atom > 99%) 1,2 , **F-Yb-1**³ and **H-L**⁴ was synthesized according to literature methods.

Theoretical calculations

We performed single point energy calculation in the crystal structure with hybrid density functional, B3LYP,^{5,6} using the program package Gaussian 09 (Revision E.01)⁷. The 6-31G(d) basis set^{8,9} was used for all atoms except Yb³⁺ and Co²⁺ atoms, which was described by the Stuttgart relativistic pseudopotential and its accompanying basis set (ECP59MWB)^{10,11} and the effective core potential (ECP) Lanl2DZ basis set¹².

Synthesis of F-Ln (Ln = Yb, Gd)

Scheme S1. Synthetic procedure of F-Ln.

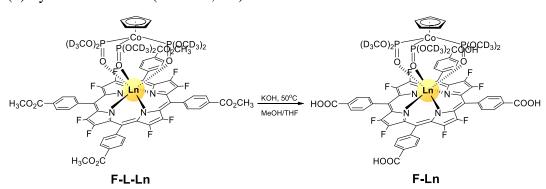
(1) Synthesis of 2,3,7,8,12,13,17,18-octafluoro-5,10,15,20-tetrakis[4-(methyl formyl)-phenyl]porphyrin (**F-L**).

2,6,-Difluoropyrrole (103 mg, 1.0 mmol) and methyl *p*-formylbenzoate (164 mg, 1.0 mmol) were dissolved in 250 mL distilled methylene chloride. The mixture was stirred for 10 min after which a few drops of BF₃·Et₂O was added. The mixture was then stirred at room temperature for 1 h. 2,3-Dicyano-5,6-dichlorobenzoquinone (DDQ, 0.23 g, 1.0 mmol) was added. After reaction overnight at room temperature, silica gel (2g) was added to the dark solution and all solvent was evaporated. The absorbed products were placed on the top of a silica gel column. The product was obtained by using ethyl acetate as eluent (65% yield). ¹H NMR (400 MHz, CDCl₃) δ -4.21 (s, 2H), 4.08 (s, 12H), 8.12 (d, 4H), 8.39 (d, 4H). ¹⁹F NMR (377 MHz, CDCl₃) δ -139.81 (s, 4F), -145.28 (s, 4F). HR-MS (ESI⁺) m/z [M+H]⁺: Calcd for C₅₂H₃₁F₈N₄O₈ 991.2014; found: 991.1995. UV/Vis (CH₂Cl₂, 25 °C): λ max(nm) (log ϵ): 419 (5.69), 515 (4.45), 549 (4.43), 645(4.38).

(2) Synthesis of \mathbf{F} - \mathbf{L} - \mathbf{L} \mathbf{n} (Ln = Yb, Gd).

F-L (200 mg, 0.2 mmol), $Ln(acac)_3 \cdot 3H_2O$ (500 mg, 1,2,4-trichlorobenzene (TCB) were added to a Schlenk tube and refluxed overnight. After cooling to room temperature, the reaction mixtures were transferred to a silica column, TCB was first eluted by petroleum ether, and then the unreacted free base ligand was eluted by CH₂Cl₂, corresponding Yb³⁺ complex was obtained by using $CH_2Cl_2/MeOH$ (v/v = 5:1) as eluent and used directly to the next step. The obtained Ln complexes (acac as ancillary ligand) and 3 equiv. NaL_{OCD3} were dissolved in mixing solvent of CHCl₃/CH₃OH (v/v = 1:1, 5 mL). The mixture was refluxed for 8 h. After cooling to room temperature, the reaction mixtures were transferred to a silica column, F-L-Ln(III)-L_{OCD3} (F-L-Ln) was obtained by using ethyl acetate/petroleum ether (v/v = 1:2) as eluent. Complex **F-L-Yb**: Yield: 90%; ¹H NMR (400 MHz, CDCl₃) δ -5.21 (s, 5H), 4.89 (s, 12H), 9.00 (s, 4H), 9.67 (s, 4H), 11.43 (s, 4H), 17.24 (s, 4H). ¹⁹F NMR (377 MHz, CDCl₃) δ -131.53 (d, 8F). HR-MS (ESI⁺) m/z [M+H]⁺: Calcd for C₆₃H₃₄D₁₈CoF₈N₄O₁₇P₃Yb 1632.2263; found: 1632.2319. UV/Vis (CH₂Cl₂, 25 °C): $\lambda_{\text{max}}(\text{nm})$ (log ϵ): 409 (5.45), 550 (4.31), 591 (4.30). Complex **F-L-Gd**: Yield: 85%; HR-MS (ESI⁺) m/z [M+H]⁺: Calcd for C₆₃H₃₄D₁₈CoF₈GdN₄O₁₇P₃ 1616.2115; found: 1616.2160; elemental analysis calcd (%) for C₆₃H₃₃D₁₈CoF₈GdN₄O₁₇P₃: C, 46.84; H, 4.30; N, 3.47; found: C 46.55, H 4.61, N: 3.48; UV/Vis (CH₂Cl₂, 25 °C): $\lambda_{\text{max}}(\text{nm})$ (log ϵ): 409 (5.47), 550 (4.35), 591 (4.31).

(3) Synthesis of $\mathbf{F-Yb}$ (Ln = Yb, Gd).



F-L-Ln was dissolved in mixing solvent of tetrahydrofuran/ CH_3OH (v/v = 1:1, 10 mL), with the addition of 1 mL 10% KOH solution. The reaction mixtures were

stirred at 50 °C overnight and the solution was neutralized with 0.2 M HCl. After evaporating of the organic solvent, the product **F-Ln** was obtained by centrifugalization and further recrystallization from CH₃OH. Complex **F-Yb**: Yield: 99%; ¹H NMR (400 MHz, D₆-DMSO) δ -5.45 (s, 5H), 9.39 (s, 4H), 9.69 (s, 4H), 11.45 (s, 4H), 14.32 (s, 4H), 17.46 (s, 4H). ¹⁹F NMR (377 MHz, CDCl₃) δ -132.71 (s, 8F). HR-MS (ESI⁺) m/z [M+H]⁺: Calcd for C₅₉H₂₆D₁₈CoF₈N₄O₁₇P₃Yb 1576.1637; found: 1576.1631. UV/Vis (DMSO, 25 °C): λ_{max} (nm) (log ϵ): 409 (5.43), 550 (4.34), 591 (4.29). Complex **F-Gd**: Yield: 95%; HR-MS (ESI⁺) m/z [M]: Calcd for C₅₉H₂₅D₁₈CoF₈N₄O₁₇P₃Gd 1559.1411; found: 1559.1441; elemental analysis calcd (%) for C₅₉H₂₅D₁₈CoF₈N₄O₁₇P₃Gd: C, 45.45; H, 3.94; N, 3.59; found: C 45.30, H 4.12, N: 3.51; UV/Vis (H₂O, 25 °C): λ_{max} (nm) (log ϵ): 409 (5.51), 550 (4.33), 591 (4.30).

Synthesis of H-Yb

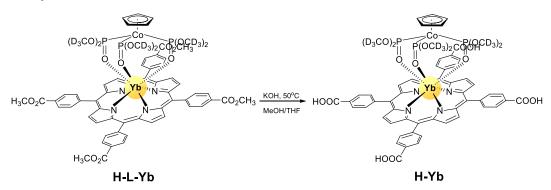
Scheme S2. Synthetic procedure of H-Yb.

(1) Synthesis of **H-L-Yb**

H-L (200 mg, 0.2 mmol), Yb(acac)₃•3H₂O (500 mg, 1 mmol) and 1,2,4-trichlorobenzene (TCB) were added to a Schlenk tube and refluxed overnight. After cooling to room temperature, the reaction mixtures were transferred to a silica column, TCB was first eluted by petroleum ether, and then the unreacted free base ligand was eluted by CH_2Cl_2 , corresponding Yb^{3+} complex was obtained by using $CH_2Cl_2/MeOH$ (v/v = 5:1) as eluent and used directly to the next step. The obtained Yb^{3+} complexes (acac as ancillary ligand) and 3 equiv. NaL_{OCD3} were dissolved in mixing solvent of $CHCl_3/CH_3OH$ (v/v = 1:1, 5 mL). The mixture was refluxed for 8 h. After cooling to room temperature, the reaction mixtures were transferred to a silica

column, **H-L-Yb(III)-L_{OCD3}** (**H-L-Yb)** was obtained by using ethyl acetate/petroleum ether (v/v = 1:2) as eluent. Complex **H-L-Yb**: Yield: 65%; ¹H NMR (400 MHz, CDCl₃) δ -4.64 (s, 5H), 4.84 (s, 12H), 8.71 (s, 4H), 9.47 (s, 4H), 11.29 (s, 4H), 15.28 (s, 8H), 17.05(s, 4H). HR-MS (ESI⁺) m/z [M+H]⁺: Calcd for C₆₃H₄₁D₁₈CoN₄O₁₇P₃Yb 1487.2938; found: 1487.2982. UV/Vis (CH₂Cl₂, 25 °C): λ_{max} (nm) (log ϵ): 425 (5.42), 555 (4.30), 597 (4.28).

(2) Synthesis of **H-Yb**



H-L-Yb was dissolved in mixing solvent of tetrahydrofuran/CH₃OH (v/v = 1:1, 10 mL), with the addition of 1 mL 10% KOH solution. The reaction mixtures were stirred at 50 °C overnight and the solution was neutralized with 0.2 M HCl. After evaporating of the organic solvent, the product **H-Yb** was obtained by centrifugalization and further recrystallization from CH₃OH. Complex **H-Yb**: Yield: 99%; ¹H NMR (400 MHz, CD₃OD) δ -5.53 (s, 5H), 8.56 (s, 4H), 8.89 (s, 4H), 9.57 (s, 4H), 11.29 (s, 4H), 15.45 (s, 8H), 16.91(s, 4H). HR-MS (ESI⁺) m/z [M+H]⁺: Calcd for C₅₉H₃₃D₁₈CoN₄O₁₇P₃Yb 1431.2312; found: [M-4H]⁴⁺ 356.8017. UV/Vis (H₂O, 25 °C): λ_{max} (nm) (log ε): 426 (5.39), 556 (4.25), 597 (4.21).

Supporting figures

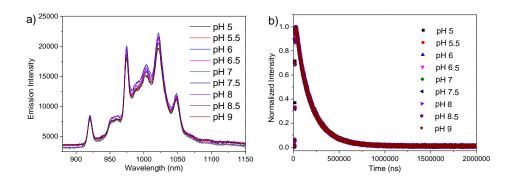


Fig. S1 a) Emission and b) decay curves monitored at 980 nm of **F-L-Yb** measured at different pH from pH 9 to pH 5 in the PBS buffer ($\lambda_{ex} = 420$ nm, $A_{420 \text{ nm}} = 0.1$).

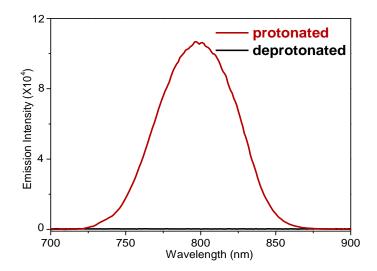


Fig. S2 Phosphorescence spectra of protonated and deprotonated **F-Gd**. The solvent is MeOH/EtOH 1:1, v/v at 77 K ($\lambda_{ex} = 410$ nm, $OD_{410 \text{ nm}} = 0.1$). For the protonated one, **F-Gd** was directly dissolved in MeOH/EtOH. For the deprotonated one, **F-Gd** was first dissolved in H₂O, with the addition of 0.1 mM NaOH to reach a final pH value of ca. 11. Then the solution was freeze-dried and the complex was re-dissolved in MeOH/EtOH.

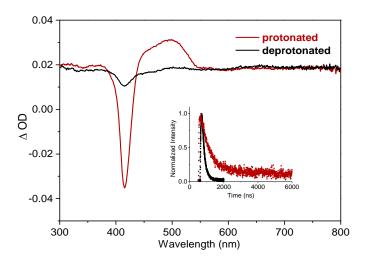


Fig. S3 Transient absorption spectra of protonated and deprotonated **F-Gd**. The spectra was measured in degassed MeOH ($OD_{410 \text{ nm}} = 0.05$) using the 410 nm OPO laser excitation pulse (10 Hz, 2 mJ pulse⁻¹). Inset: triplet decay lifetime monitored at 500 nm.

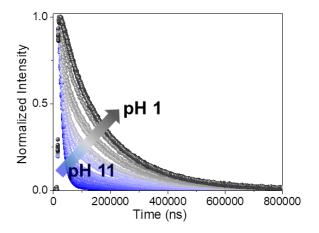


Fig. S4 Decay curves of **F-Yb** at different pH monitored at 980 nm from pH 11 to pH 1 in the Britton-Robison buffer.

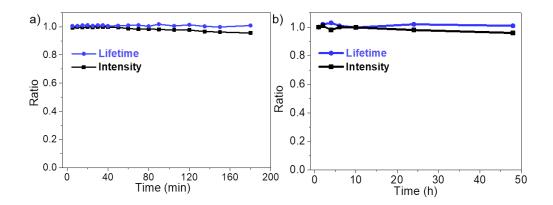


Fig. S5 Spectroscopic and lifetime evaluation of **F-Yb** at pH 1.5 KCl/HCl buffer a) with or b) without the irradiation of a 405 nm laser (0.2 W/cm²). Emission intensity and lifetime values were referenced to the emission intensity or lifetime of Yb³⁺ (980 nm) at 0 h.

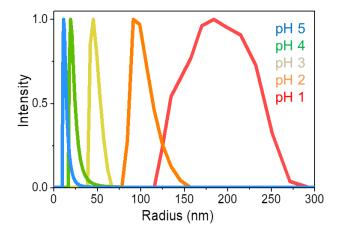


Fig. S6 Dynamic light scattering of F-Yb (1 μ M) at different pH Britton-Robison buffer.

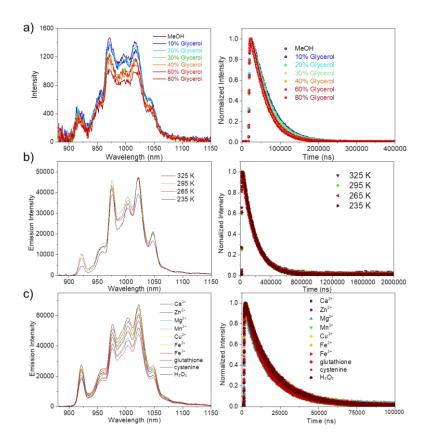


Fig. S7 Influence of the a) viscosity (measured in MeOH/Glycerol mixture), b) temperature (2-CH₃-THF solution) and c) metal ions (40 μ M, pH 7.4 PBS buffer) on the luminescence (left column) and decay curve (right column,) of **F-Yb** (1 μ M). The decay curves were monitored at 980 nm under the excitation of 410 nm.

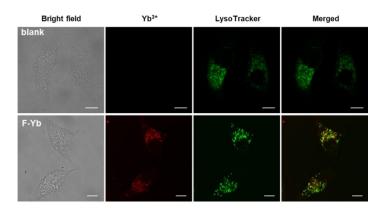


Fig. S8 Colocalization of **F-Yb** with LysoTracker Green. The NIR confocal images were performed on HeLa cells incubated with 10 μM of **F-Yb** for 4 h followed by 30 min incubation with 75 nM LysoTracker Green. a) Bright field; b) NIR signal arising from Yb³⁺ in channel 1 (λ_{ex} , 408 nm; λ_{em} , 935/170 nm bandpass); c) visible signal arising from LysoTracker Green in channel 2 (λ_{ex} , 470 nm; λ_{em} , 530/43 nm bandpass); d) merged b and c (P = 0.93). Scale bar: 10 μm.

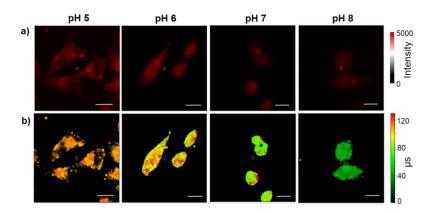


Fig. S9 The **F-Yb** images intracellular pH values with incubation of nigericin. a) NIR confocal fluorescence images and b) time-resolved fluorescence lifetime images of **F-Yb** (10 μM, 4 h) clamped at pH 5.0, 6.0, 7.0, 8.0 in HeLa cells in the presence of 10 μM nigericin (λ_{ex} , 408 nm; λ_{em} , 935/170 nm bandpass; dwell time, 1 ms). Scale bar: 10 μm.

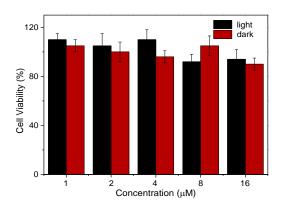
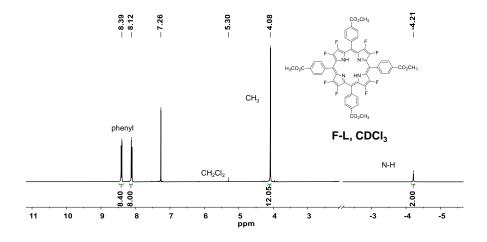
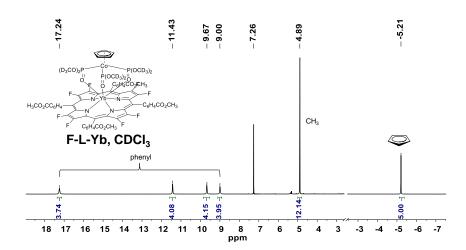
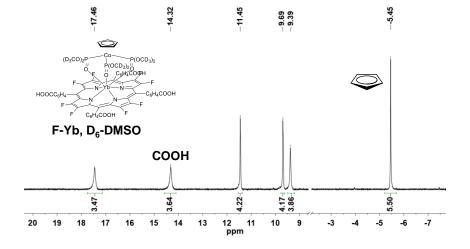


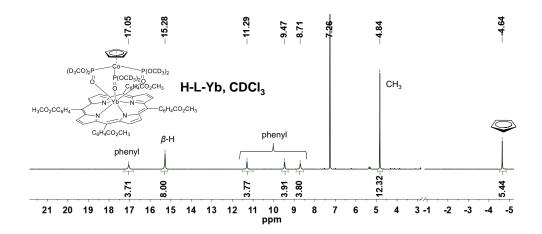
Fig. S10 Cytotoxicity of **F-Yb**. Cells were incubated with varying concentrations from 0 to 16 μM of the complex for 24 h in the dark while wells containing no cells were set as the controls. After washing three times with PBS, the cells were irradiated for 30 min in 100 mL PBS under the light irradiation (400–700 nm, 6.5 mW/cm²) for 30 min (this step was for the photocytotoxicity). Then, PBS was replaced by 200 mL of fresh culture media. After culture for 24 h, the cells were washed three times with PBS. Then, 10 μL Cell Counting Kit-8 (CCK-8) solution and 90 μL PBS were added per well. After 2 h, the absorbance at 450 nm was read by a 96-well plate reader. The viability of HeLa cells was calculated by $CV = (A_s - A_b)/(A_c - A_b)*100\%$, where CV stands for the viability of cells, A_s , A_c , and A_b stand for the absorbance of cells containing the studied complexes, cell control (no treated cells), and blank control (wells containing neither cells nor the studied complexes), respectively.

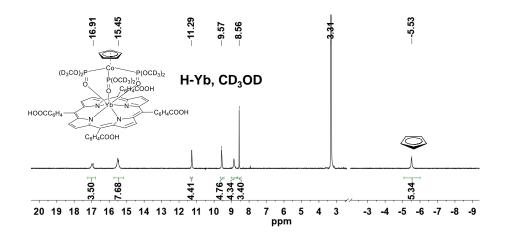
¹H NMR spectra



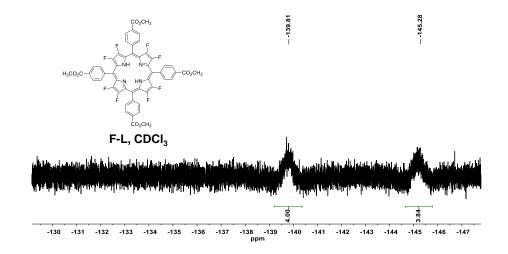


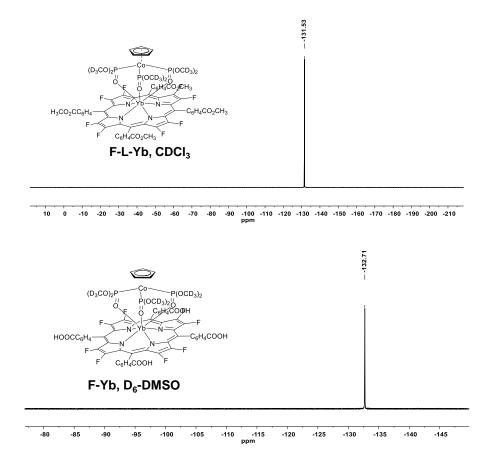




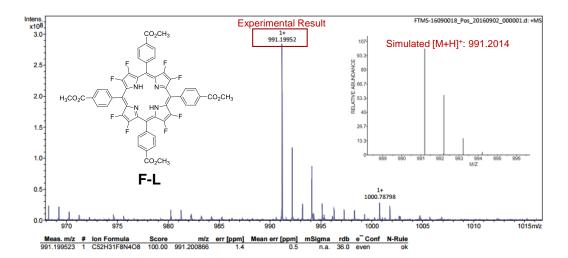


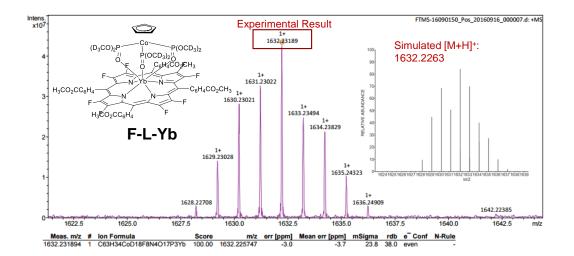
¹⁹F NMR spectra

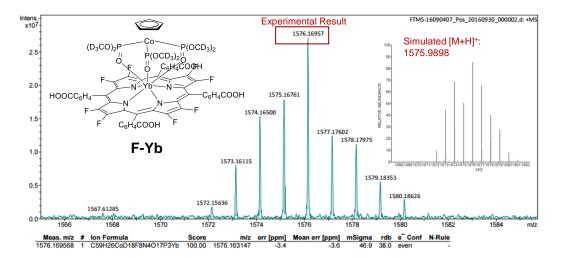


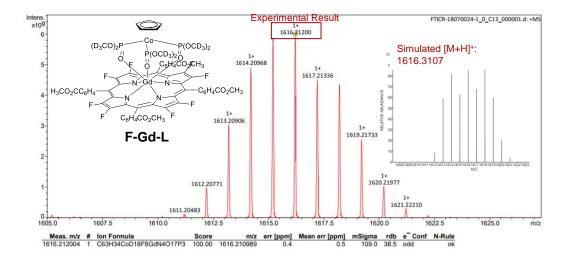


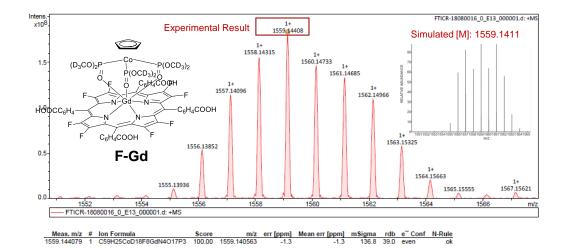
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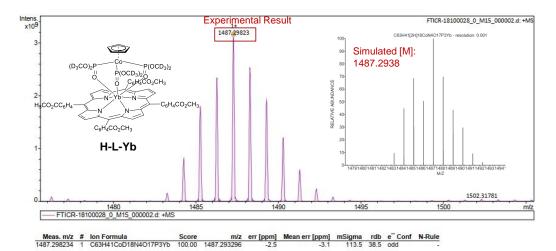


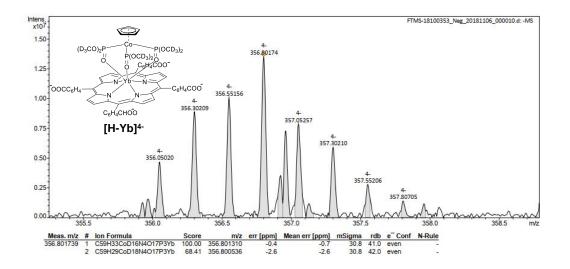




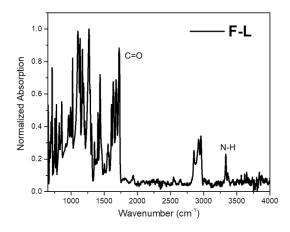


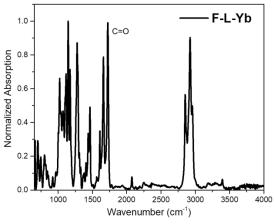


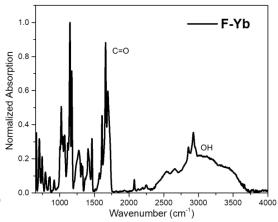


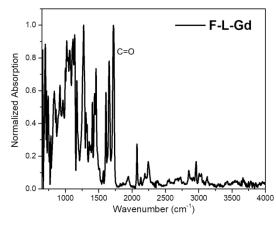


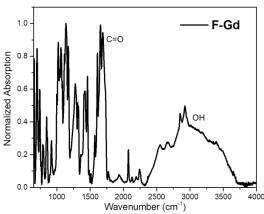
IR spectra

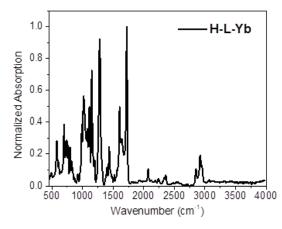


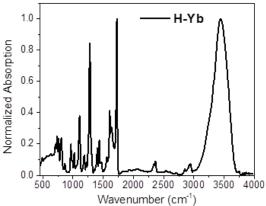












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