

Exploiting the Trifluoroethyl Group as a Precatalyst Ligand in Nickel-Catalyzed Suzuki- type Alkylations

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I. General specifications

All reagents were used as received from commercial sources, unless specified otherwise, or prepared as described in the literature. All manipulations were conducted using standard Schlenk and high-vacuum techniques or in a nitrogen-filled glovebox. Solvents were distilled from Na/benzophenone or CaH_2 . ^1H NMR and ^{13}C NMR spectra were recorded on a 500 MHz or 600 MHz spectrometer (TMS as internal standard). ^{19}F NMR was also recorded on a 400 MHz or 500 MHz spectrometer (FCCl_3 as outside standard and low field is positive). Chemical shifts (δ) are reported in parts per million, and coupling constants (J) are in hertz. High resolution mass spectrometry (EI/TOF or ESI-TOF) was performed at the Mass Spectrometry Facility.

Thin layer chromatography monitoring (TLC) was performed using precoated silica gel plate (0.2 mm thickness, GF254). Subsequent to elution, plates were first visualized using UV radiation (254 nm). Further visualization was conducted by staining with basic solution of potassium permanganate or acidic solution of ceric molybdate, followed by heating on a hot plate. Flash chromatography was performed using silica gel (200-300 mesh) with HPLC solvents. Columns were typically packed as slurry and equilibrated with petroleum ether prior to use.

II. Synthesis of precatalyst [(bipy)Ni(CH₂CF₃)₂]

A 100 mL round-bottom flask was charged with 2,2'-bipyridine (0.500 g, 3.20 mmol), Ni(COD)₂ (0.880 g, 3.20 mmol) and THF (40 mL). After stirring at room temperature for 12 h, a THF solution of CF₃CH₂I (0.740 g, 3.52 mmol in 10 mL of THF) were added to the flask. The reaction mixture was stirred overnight, and then the volatiles were removed under reduced pressure. The residue was redissolved in benzene (80 mL), and the suspension solution was passed through a sand-core glass funnel to remove the insoluble [(bipy)NiI₂]. After the solvent of the filtrate were removed under reduced pressure, the residue solid was washed with pentane (5 mL × 2) and dried in *vacuo* to furnish [(bipy)Ni(CH₂CF₃)₂] as a dark red powder (0.252 g, Yield 41%). Suitable single crystals for X-ray analysis were obtained by recrystallization from THF/pentane solution at -25 °C. ¹H NMR (500 MHz, THF-d₈): δ 8.69 (d, *J* = 7.8 Hz, 2H), 8.23 (d, *J* = 7.8 Hz, 2H), 8.09 (dt, *J* = 7.8 Hz, 1.3 Hz, 2H), 7.59 (m, 2H), 1.06 (q, *J* = 16.2 Hz, 4H); ¹⁹F NMR (470 MHz, CDCl₃): δ -47.98 (t, *J* = 16.2 Hz, 6F); ¹³C NMR (125 MHz, CDCl₃): δ 156.22, 150.56, 139.39, 135.38 (q, *J* = 273.8 Hz), 127.16, 122.24, 6.11 (q, *J* = 23.6 Hz); Elemental analysis calcd (%) for C₁₄H₁₂F₆N₂Ni: C, 44.14; H, 3.18; found: C, 44.01; H, 3.27.

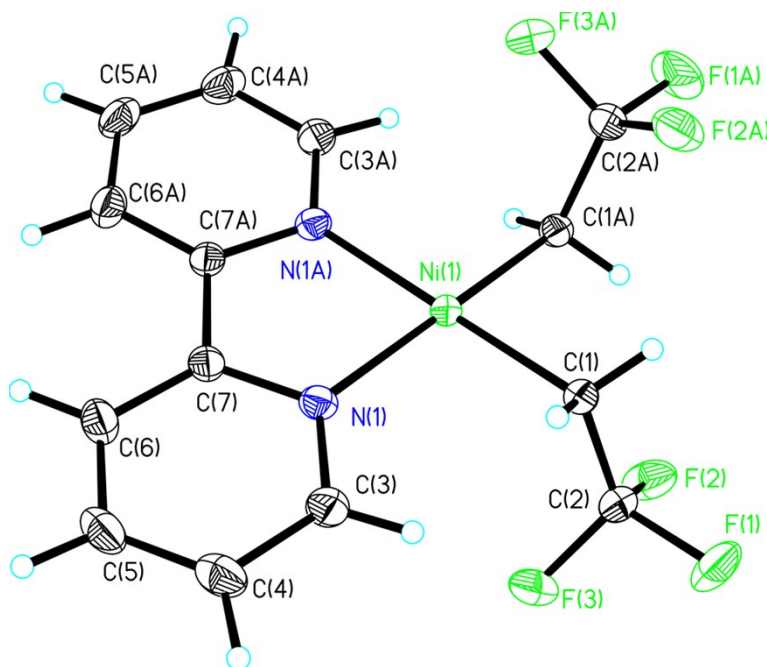
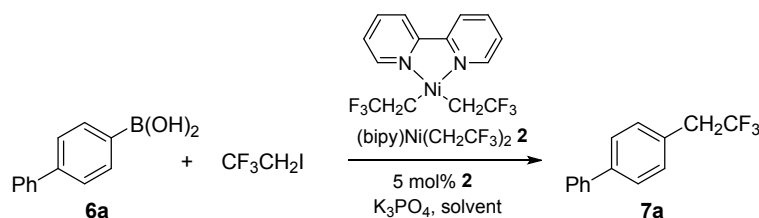


Figure S1. ORTEP diagram of [(bipy)Ni(CH₂CF₃)₂] (**2**). (CCDC 1436475)

III. Examination of the catalytic activities of [(bipy)Ni(CH₂CF₃)₂] and reaction condition optimization

4-biphenylboronic acid **2a** (1.5 equiv), base (2.0 equiv), followed by a solution of CF₃CH₂I (1.0 equiv) in the indicated solvent (1.0 mL) were loaded into a 25 mL of Schlenk tube which was subject to evacuating/flushing with nitrogen gas three times. The precatalyst **2** (5.0 mol%) in the indicated solvent (0.5 mL) was added dropwise into the reaction system subsequently. The Schlenk tube was screw capped and put into a preheated oil bath (80 °C). After stirring for 12-24 h, the reaction mixture was cooled to room temperature and poured into a saturated aqueous ammonium chloride solution (10.0 mL). The aqueous phase was extracted with ether three times (10.0 mL×3) and the combined organic phase was dried over sodium sulfate. After removing the solvent in *vacuo*, the residue was purified by flash chromatography on silica gel or preparative TLC to afford the corresponding ArCH₂CF₃ products.

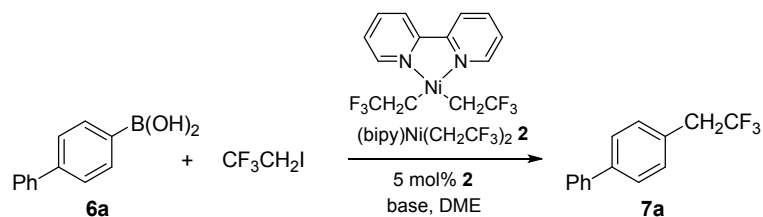
Table S1. Solvent effect on the precatalyst **2** catalyzed Suzuki-type trifluoroethylation



Entry	Base	Solvent	T/ °C	Isolated Yield
1	K ₃ PO ₄	DME	80	93%
2	K ₃ PO ₄	DMSO	80	91%
3	K ₃ PO ₄	DMF	80	47%
4	K ₃ PO ₄	CH ₃ CN	80	35%
5	K ₃ PO ₄	toluene	80	43%
6	K ₃ PO ₄	THF	80	54%

7 K₃PO₄ dioxane 80 78%

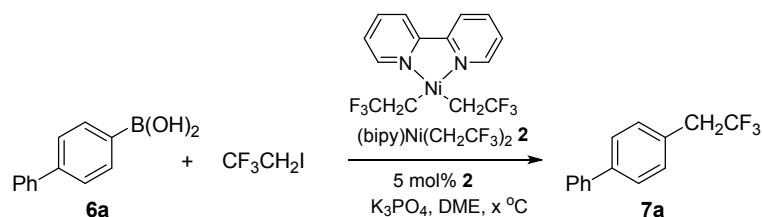
Table S2. Base effect on the precatalyst **2** catalyzed Suzuki-type trifluoroethylation



Entry	Base	Solvent	T/ °C	Isolated Yield
1	K ₃ PO ₄	DME	80	93%
2	Na ₂ CO ₃	DME	80	7% ^a
3	K ₂ CO ₃	DME	80	23% ^a
4	Cs ₂ CO ₃	DME	80	50% ^a
5	tBuOK	DME	80	Trace ^b
6	NaOAc	DME	80	ND ^c

^aPartial dehydrofluorination of coupling product. ^bComplete dehydrofluorination of coupling product. ^cNo conversion of CF₃CH₂I.

Table S3. Temperature effect of Suzuki-type trifluoroethylation^a



Entry	Base	Solvent	T/ °C	Isolated Yield
1	K ₃ PO ₄	DME	80	93%
2	K ₃ PO ₄	DME	50	77%
3	K ₃ PO ₄	DME	RT	<5%

^aThis experiment indicated the activation temperature of precatalyst [(bipy)Ni(CH₂CF₃)₂] is approximately 50

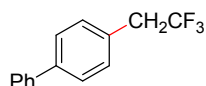
°C which is lower than the commercialized (TMEDA)Ni(*o*-tol)(Cl) (the lower bound of activation is 60 °C) (J. D. Shields, E. E. Gray and A. G. Doyle, *Org. Lett.*, 2015, **17**, 2166).

IV. Suzuki-type direct trifluoroethylation and alkylation of ArB(OH)₂ and characterization of Ar-R products

(a) Typical Procedure

Arylboronic acid (1.5 equiv), base (2.0 equiv), followed by a solution of CF₃CH₂I (0.4 mmol, 1.0 equiv) or the corresponding R-X (0.4 mmol, 1.0 equiv) in the DME solvent (1.0 mL) were loaded into a 25 mL of Schlenck tube which was subject to evacuating/flushing with nitrogen gas three times. The precatalyst **2** (5.0 mol%) in the DME solvent (0.5 mL) was added dropwise into the reaction system subsequently. The Schlenck tube was screw capped and put into a preheated oil bath (80 °C). After stirring for 12-24 h, the reaction mixture was cooled to room temperature and poured into a saturated aqueous ammonium chloride solution (10.0 mL). The aqueous phase was extracted with ether three times (10.0 mL × 3) and the combined organic phase was dried over sodium sulfate. After removing the solvent in *vacuo*, the residue was purified by flash chromatography on silica gel or preparative TLC to afford the corresponding Ar-CH₂CF₃ or Ar-R products.

(b) Characterization of Ar-CH₂CF₃ and Ar-R Products

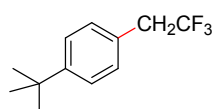


(7a). Colorless oil, TLC R_f (hexane) = 0.65, 91% yield (87 mg). A gram-scale synthesis was

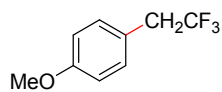
implemented as the following procedure and the isolated yield can be comparatively effective (83%). ¹H NMR (600 MHz, CDCl₃) δ 7.58 (d, *J* = 7.7 Hz, 4H), 7.44 (t, *J* = 7.6 Hz, 2H), 7.35 (t, *J* = 8.5 Hz, 3H), 3.40 (q, *J* = 10.8 Hz, 2H); ¹⁹F NMR (470 MHz, CDCl₃) δ -65.85 (t, *J* = 10.8 Hz, 3F); ¹³C NMR (151 MHz, CDCl₃) δ 141.12 (s), 140.53 (s), 130.60 (s), 129.14 (q, *J* = 2.9 Hz), 128.86 (s), 127.54 (s), 127.44 (s), 127.14 (s), 125.84 (q, *J* = 276.8 Hz), 39.92 (q, *J* = 29.8 Hz). MS (EI) *m/z* 236 (M⁺); HRMS (EI-TOF) *m/z* [M]⁺ Calcd for C₁₄H₁₁F₃ 236.0813, found 236.0808.

Gram-scale synthesis procedure: 4-biphenylboronic acid (2.38 g, 12.0 mmol, 1.5 equiv), K₃PO₄ (5.10 g, 24.0 mmol, 3.0 equiv), followed by a solution of CF₃CH₂I (0.80 mL, 8.0 mmol, 1.0 equiv) in the DME solvent (20.0 mL) were loaded into a 100 mL of Schlenck tube which was subject to evacuating/flushing with nitrogen gas three times. The precatalyst **2** (76.0 mg, 2.5 mol%) in the DME solvent (5.0 mL) was added dropwise into

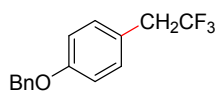
the reaction system subsequently. The Schlenck tube was screw capped and put into a preheated oil bath (80 °C). After stirring for 24 h, the reaction mixture was cooled to room temperature and poured into a saturated aqueous ammonium chloride solution (100.0 mL). The aqueous phase was extracted with ether three times (70.0 mL \times 3) and the combined organic phase was dried over sodium sulfate. After removing the solvent in *vacuo*, the residue was purified by flash chromatography on silica gel (eluent: petroleum ether) afford the corresponding 4-(2,2,2-trifluoroethyl)-1,1'-biphenyl **7a** (1.57 g, yield 83%).



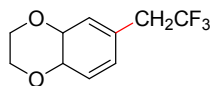
(7b). Colorless oil, TLC R_f (hexane) = 0.60, 79% yield (68 mg). ^1H NMR (500 MHz, CDCl_3): δ 7.38 (d, J = 8.4 Hz, 2H), 7.23 (d, J = 8.1 Hz, 2H), 3.33 (q, J = 10.9 Hz, 2H), 1.33 (s, 9H); ^{19}F NMR (470 MHz, CDCl_3): δ -65.97 (t, J = 10.9 Hz, 3F); ^{13}C NMR (125 MHz, CDCl_3): δ 151.27, 130.06, 127.35 (q, J = 2.7 Hz), 126.14 (q, J = 276.6 Hz), 125.83, 39.93 (q, J = 29.6 Hz), 34.75, 31.50. MS (EI) m/z 216 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{12}\text{H}_{15}\text{F}_3$ 216.1126, found 216.1120.



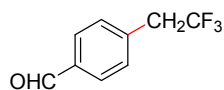
(7c). Colorless oil, TLC R_f (hexane) = 0.55, 86% yield (65 mg). ^1H NMR (500 MHz, CDCl_3): δ 7.21 (d, J = 8.4 Hz, 2H), 6.90-6.87 (m, 2H), 3.80 (s, 3H), 3.29 (q, J = 10.8 Hz, 2H); ^{19}F NMR (470 MHz, CDCl_3): δ -66.44 (t, J = 10.8 Hz, 3F); ^{13}C NMR (125 MHz, CDCl_3): δ 159.65, 131.46, 126.11 (q, J = 276.6 Hz), 122.35 (q, J = 2.8 Hz), 114.27, 55.42, 39.57 (q, J = 29.7 Hz); MS (EI) m/z 190 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_9\text{H}_9\text{F}_3\text{O}$ 190.0605, found 190.0608.



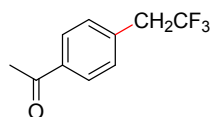
(7d). White solid, TLC R_f (hexane) = 0.55, 83% yield (88 mg). ^1H NMR (500 MHz, CDCl_3): δ 7.45 (d, J = 7.0 Hz, 2H), 7.41 (t, J = 7.0 Hz, 2H), 7.37-7.33 (m, 1H), 7.23 (d, J = 8.5 Hz, 2H), 7.00-6.97 (m, 2H), 5.07 (s, 2H), 3.31 (q, J = 10.9 Hz, 2H); ^{19}F NMR (470 MHz, CDCl_3): δ -66.31 (t, J = 10.9 Hz, 3F); ^{13}C NMR (125 MHz, CDCl_3): δ 158.87, 137.02, 131.49, 128.83, 128.24, 127.68, 126.08 (q, J = 276.7 Hz), 122.63 (q, J = 2.8 Hz), 115.19, 70.21, 39.57 (q, J = 29.7 Hz). MS (EI) m/z 266 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{15}\text{H}_{13}\text{F}_3\text{O}$ 266.0918, found 266.0921.



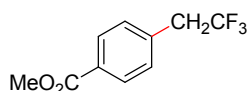
(7e). Light yellow oil, TLC R_f (hexane : EtOAc = 10 : 1) = 0.55, 80% yield (71 mg). ^1H NMR (500 MHz, CDCl_3): δ 6.83 (d, J = 8.3 Hz, 1H), 6.81 (d, J = 1.8 Hz, 1H), 6.75 (dd, J = 8.3 Hz, 1.8 Hz, 1H), 4.23 (s, 4H), 3.24 (q, J = 10.8 Hz, 2H); ^{19}F NMR (470 MHz, CDCl_3): δ -66.26 (t, J = 10.8 Hz, 3F); ^{13}C NMR (125 MHz, CDCl_3): δ 143.71, 143.69, 126.02 (q, J = 276.7 Hz), 123.35, 121.60, 119.20, 117.60, 64.50, 64.48, 39.64 (q, J = 29.8 Hz). MS (EI) m/z 220 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{10}\text{H}_{11}\text{F}_3\text{O}_2$ 220.0711, found 220.0715.



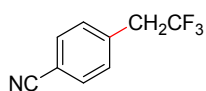
(7f). Light yellow oil, TLC R_f (hexane : EtOAc = 9 : 1) = 0.45, 70% yield (53 mg). ^1H NMR (500 MHz, CDCl_3): δ 10.01 (s, 1H), 7.86 (d, J = 8.3 Hz, 2H), 7.46 (d, J = 8.3 Hz, 2H), 3.44 (q, J = 10.6 Hz, 2H); ^{19}F NMR (470 MHz, CDCl_3): δ -65.47 (t, J = 10.6 Hz, 3F); ^{13}C NMR (125 MHz, CDCl_3): δ 191.89, 136.92 (q, J = 2.8 Hz), 136.33, 131.10, 130.18, 125.52 (q, J = 277.0 Hz), 40.52 (q, J = 30.0 Hz). MS (EI) m/z 188 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_9\text{H}_7\text{F}_3\text{O}$ 188.0449, found 188.0447.



(7g). Colorless oil, TLC R_f (hexane : EtOAc = 5 : 1) = 0.48, 74% yield (60 mg). ^1H NMR (500 MHz, CDCl_3): δ 7.93 (dt, J = 8.3 Hz, 2.0 Hz, 2H), 7.38 (d, J = 8.1 Hz, 2H), 3.41 (q, J = 10.7 Hz, 2H), 2.58 (s, 3H); ^{19}F NMR (470 MHz, CDCl_3): δ -65.57 (t, J = 10.6 Hz, 3F); ^{13}C NMR (125 MHz, CDCl_3): δ 197.74, 137.06, 135.45 (q, J = 2.7 Hz), 130.63, 128.84, 125.60 (q, J = 277.0 Hz), 40.33 (q, J = 30.0 Hz), 26.81. MS (EI) m/z 202 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{10}\text{H}_9\text{F}_3\text{O}$ 202.0605, found 202.0600.

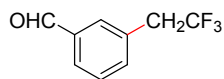


(7h). White solid, TLC R_f (hexane : EtOAc = 6 : 1) = 0.45, 80% yield (70 mg). ^1H NMR (500 MHz, CDCl_3): δ 8.01 (d, J = 8.3 Hz, 2H), 7.35 (d, J = 8.3 Hz, 2H), 3.90 (s, 3H), 3.40 (q, J = 10.7 Hz, 2H); ^{19}F NMR (470 MHz, CDCl_3): δ -65.62 (t, J = 10.7 Hz, 3F); ^{13}C NMR (125 MHz, CDCl_3): δ 166.84, 135.28 (q, J = 2.6 Hz), 130.42, 130.24, 130.10, 125.62 (q, J = 276.9 Hz), 52.40, 40.35 (q, J = 30.0 Hz). MS (EI) m/z 218 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{10}\text{H}_9\text{F}_3\text{O}_2$ 218.0555, found 218.0552.

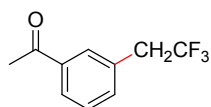


(7i). White solid, TLC R_f (hexane : EtOAc = 6 : 1) = 0.35, 72% yield (53 mg). ^1H NMR (500 MHz, CDCl_3): δ 7.64 (d, J = 8.1 Hz, 2H), 7.40 (d, J = 8.1 Hz, 2H), 3.42 (q, J = 10.5 Hz, 2H); ^{19}F NMR (470 MHz, CDCl_3): δ -65.55 (t, J = 10.6 Hz, 3F); ^{13}C NMR (125

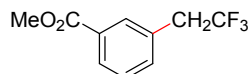
MHz, CDCl₃): δ 135.50 (q, J = 2.8 Hz), 132.65, 131.16, 125.33 (q, J = 277.0 Hz), 118.52, 112.56, 40.42 (q, J = 30.3 Hz). MS (EI) m/z 185 (M^+); HRMS (EI-TOF) m/z [M]⁺ Calcd for C₉H₆F₃N 185.0452, found 185.0452.



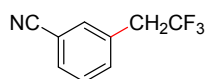
(7j). Yellow oil, TLC R_f (hexane : EtOAc = 9 : 1) = 0.45, 48% yield (36 mg). ¹H NMR (600 MHz, CDCl₃) δ 10.03 (s, 1H), 7.87 (d, J = 7.3 Hz, 1H), 7.83 (s, 1H), 7.57 (dt, J = 14.9, 7.6 Hz, 2H), 3.47 (q, J = 10.6 Hz, 2H); ¹⁹F NMR (470 MHz, CDCl₃) δ -65.84 (t, J = 10.8 Hz, 3F); ¹³C NMR (151 MHz, CDCl₃) δ 191.77 (s), 136.83 (s), 136.06 (s), 131.33 (q, J = 3.2 Hz), 131.13 (s), 129.49 (s), 128.19 (s), 125.44 (q, J = 276.8 Hz), 39.99 (q, J = 30.2 Hz). MS (EI) m/z 188 (M^+); HRMS (EI-TOF) m/z [M]⁺ Calcd for C₉H₇F₃O 188.0449, found 188.0447.



(7k). Yellow oil, TLC R_f (hexane : EtOAc = 5 : 1) = 0.50, 67% yield (54 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.94 (d, J = 7.6 Hz, 1H), 7.90 (s, 1H), 7.52 (d, J = 7.6 Hz, 1H), 7.48 (t, J = 7.6 Hz, 1H), 3.44 (q, J = 10.7 Hz, 2H), 2.62 (s, 3H); ¹⁹F NMR (470 MHz, CDCl₃) δ -65.87 (t, J = 10.6 Hz, 3F); ¹³C NMR (151 MHz, CDCl₃) δ 197.62 (s), 137.57 (s), 134.70 (s), 130.79 (q, J = 2.9 Hz), 129.95 (s), 129.03 (s), 128.21 (s), 125.53 (q, J = 276.9 Hz), 40.10 (q, J = 29.9 Hz), 26.64 (s). MS (EI) m/z 202 (M^+); HRMS (EI-TOF) m/z [M]⁺ Calcd for C₁₀H₉F₃O 202.0605, found 202.0600.

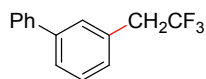


(7l). Colorless oil, TLC R_f (hexane : EtOAc = 6 : 1) = 0.45, 63% yield (55 mg). ¹H NMR (600 MHz, CDCl₃) δ 8.03 (d, J = 7.7 Hz, 1H), 7.99 (s, 1H), 7.50 (d, J = 7.6 Hz, 1H), 7.45 (t, J = 7.7 Hz, 1H), 3.93 (s, 3H), 3.43 (q, J = 10.7 Hz, 2H); ¹⁹F NMR (470 MHz, CDCl₃) δ -65.91 (t, J = 10.6 Hz, 3F); ¹³C NMR (151 MHz, CDCl₃) δ 166.62 (s), 134.57 (s), 131.33 (s), 130.72 (s), 130.54 (q, J = 2.9 Hz), 129.38 (s), 128.83 (s), 125.54 (q, J = 276.9 Hz), 52.26 (s), 40.04 (q, J = 30.0 Hz). MS (EI) m/z 218 (M^+); HRMS (EI-TOF) m/z [M]⁺ Calcd for C₁₀H₉F₃O₂ 218.0555, found 218.0552.



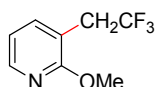
(7m). White solid, TLC R_f (hexane : EtOAc = 6 : 1) = 0.35, 44% yield (33 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.66 (d, J = 7.7 Hz, 1H), 7.61 (s, 1H), 7.56 (d, J = 7.8 Hz, 1H), 7.50 (t, J = 7.7 Hz, 1H), 3.42 (q, J = 10.5 Hz, 2H); ¹⁹F NMR (470 MHz, CDCl₃) δ -65.83 (t, J = 10.8 Hz, 3F); ¹³C NMR (151 MHz, CDCl₃) δ 134.59 (s), 133.64 (s), 131.94 (s), 131.65 (q, J = 2.9 Hz), 129.64 (s), 125.16 (q, J = 277.0 Hz), 118.24 (s), 113.11 (s), 39.83 (q, J = 30.4 Hz). MS (EI) m/z 185 (M^+); HRMS (EI-TOF) m/z [M]⁺ Calcd for C₉H₆F₃N 185.0452, found

185.0452.



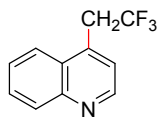
(7n). Colorless oil, TLC R_f (hexane) = 0.55, 74% yield (70 mg). ^1H NMR (600 MHz, CDCl_3)

δ 7.57 (t, J = 8.7 Hz, 3H), 7.51 (s, 1H), 7.47 – 7.40 (m, 3H), 7.36 (t, J = 7.4 Hz, 1H), 7.28 (d, J = 7.6 Hz, 1H), 3.43 (q, J = 10.8 Hz, 2H); ^{19}F NMR (470 MHz, CDCl_3) δ -65.77 (t, J = 10.6 Hz, 3F). ^{13}C NMR (151 MHz, CDCl_3) δ 141.81 (s), 140.66 (s), 130.67 (q, J = 5.6 Hz), 129.12 (s), 129.07 (s), 129.01 (s), 128.86 (s), 128.86 (s), 127.58 (s), 127.23 (s), 127.23 (s), 126.98 (s), 125.82 (q, J = 276.9 Hz), 40.33 (q, J = 29.7 Hz). MS (EI) m/z 236 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{14}\text{H}_{11}\text{F}_3$ 236.0813, found 236.0808.



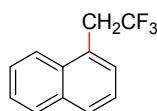
(7o). Colorless oil, TLC R_f (hexane: EtOAc = 5 : 1) = 0.40, 78% yield (60 mg). ^1H NMR (500

MHz, CDCl_3): δ 8.13 (dd, J = 5.0 Hz, 1.8 Hz, 1H), 7.52 (d, J = 7.3 Hz, 1H), 6.86 (dd, J = 7.3 Hz, 5.0 Hz, 1H), 3.95 (s, 3H), 3.40 (q, J = 10.8 Hz); ^{19}F NMR (470 MHz, CDCl_3): δ -65.50 (t, J = 10.8 Hz, 3F); ^{13}C NMR (125 MHz, CDCl_3): δ 162.62, 147.01, 140.09, 126.02 (q, J = 277.3 Hz), 116.93, 113.48 (q, J = 2.8 Hz), 53.86, 33.62 (q, J = 30.6 Hz). MS (EI) m/z 191 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_8\text{H}_8\text{F}_3\text{NO}$ 191.0558, found 191.0554.



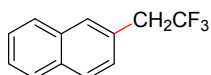
(7p). White solid, TLC R_f (hexane: EtOAc = 2 : 1) = 0.40, 82% yield (69 mg). ^1H NMR (500 MHz,

CDCl_3): δ 9.24 (s, 1H), 8.49 (s, 1H), 8.00 (d, J = 8.2 Hz, 1H), 7.96 (d, J = 8.5 Hz, 1H), 7.78 (ddd, J = 8.4 Hz, 6.9 Hz, 1.2 Hz, 1H), 7.64 (t, J = 7.5 Hz, 1H), 3.79 (q, J = 10.4 Hz, 2H); ^{19}F NMR (470 MHz, CDCl_3): δ -64.71 (t, J = 10.3 Hz, 3F); ^{13}C NMR (125 MHz, CDCl_3): δ 153.63, 145.63, 135.21, 131.25, 128.63, 127.63, 125.92 (q, J = 277.6 Hz), 122.93, 120.48, 34.60 (q, J = 30.9 Hz). MS (EI) m/z 211 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{11}\text{H}_8\text{F}_3\text{N}$ 211.0609, found 211.0601.



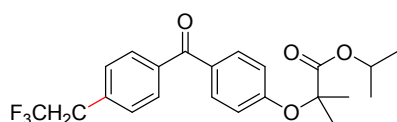
(7q). White solid, TLC R_f (hexane) = 0.55, 36% yield (30 mg). ^1H NMR (500 MHz, CDCl_3): δ

8.03 (d, J = 8.5 Hz, 1H), 7.90 (d, J = 7.9 Hz, 1H), 7.87 (dd, J = 7.4 Hz, 1.8 Hz, 1H), 7.59 (ddd, J = 8.5 Hz, 6.8 Hz, 1.4 Hz, 1H), 7.54 (ddd, J = 8.0 Hz, 6.9 Hz, 1.1 Hz, 1H), 7.50-7.46 (m, 2H), 3.87 (q, J = 10.6 Hz, 2H); ^{19}F NMR (470 MHz, CDCl_3): δ -64.66 (t, J = 10.6 Hz, 3F); ^{13}C NMR (125 MHz, CDCl_3): δ 134.09, 132.54, 129.66, 129.28, 129.05, 126.82, 126.61 (q, J = 2.3 Hz), 126.33 (q, J = 276.3 Hz), 126.10, 125.46, 123.79, 36.94 (q, J = 30.0 Hz). MS (EI) m/z 210 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{12}\text{H}_9\text{F}_3$ 210.0656, found 210.0662.



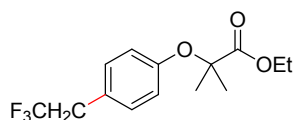
(7r). White solid, TLC R_f (hexane) = 0.57, 42% yield (35 mg). ^1H NMR (500 MHz, CDCl_3):

δ 7.88-7.84 (m, 3H), 7.79 (s, 1H), 7.55-7.51 (m, 2H), 7.43 (d, J = 8.3 Hz, 1H), 3.54 (q, J = 10.8 Hz, 2H); ^{19}F NMR (470 MHz, CDCl_3): δ -66.79 (t, J = 10.5 Hz, 3F); ^{13}C NMR (125 MHz, CDCl_3): δ 133.47, 133.08, 129.70, 128.59, 128.00, 127.90, 127.80, 126.61, 126.54, 126.13 (q, J = 277.0 Hz), 40.54 (q, J = 29.7 Hz). MS (EI) m/z 210 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{12}\text{H}_9\text{F}_3$ 210.0656, found 210.0662.



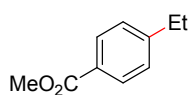
(7s). White solid, TLC R_f (hexane: EtOAc = 4 : 1) = 0.50, 61% yield (100

mg). ^1H NMR (600 MHz, CDCl_3) δ 7.76 (dd, J = 10.8, 8.5 Hz, 4H), 7.41 (d, J = 8.0 Hz, 2H), 6.87 (d, J = 8.8 Hz, 2H), 5.09 (dt, J = 12.5, 6.3 Hz, 1H), 3.46 (q, J = 10.7 Hz, 2H), 1.66 (s, 6H), 1.21 (s, 3H), 1.20 (s, 3H); ^{19}F NMR (470 MHz, CDCl_3) δ -65.53 (t, J = 10.6 Hz, 3F); ^{13}C NMR (151 MHz, CDCl_3) δ 194.92 (s), 173.14 (s), 159.72 (s), 137.97 (s), 134.07 (s), 132.05 (s), 130.35 (s), 130.05 (s), 126.41 (s), 124.58 (s), 117.22 (s), 79.42 (s), 69.35 (s), 40.19 (q, J = 30.0 Hz), 25.38 (s), 21.53 (s). MS (ESI) m/z 409 ($\text{M}+\text{H}^+$); HRMS (ESI-TOF) m/z [$\text{M}+\text{H}$] $^+$ Calcd for $\text{C}_{22}\text{H}_{24}\text{F}_3\text{O}_4$ 409.1627, found 409.1633.



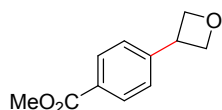
(7t). White solid, TLC R_f (hexane: EtOAc = 6 : 1) = 0.60, 44% yield (51 mg). ^1H

NMR (600 MHz, CDCl_3) δ 7.15 (d, J = 8.5 Hz, 2H), 6.81 (d, J = 8.6 Hz, 2H), 4.23 (q, J = 7.1 Hz, 2H), 3.29 (q, J = 10.9 Hz, 2H), 1.60 (s, 6H), 1.23 (t, J = 7.1 Hz, 3H); ^{19}F NMR (470 MHz, CDCl_3) δ -66.44 (t, J = 10.5 Hz, 3F); ^{13}C NMR (151 MHz, CDCl_3) δ 174.38, 155.60, 131.16, 126.01 (q, J = 277.0 Hz), 123.75 (q, J = 6.2 Hz), 119.20, 79.38, 61.69, 39.64 (q, J = 29.9 Hz), 25.59, 14.24. MS (EI) m/z 290 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{14}\text{H}_{17}\text{F}_3\text{O}_3$ 290.1130, found 290.1137.

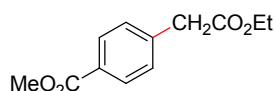


(13a). Colorless oil, TLC R_f (hexane: EtOAc = 6 : 1) = 0.60, 78% yield (51 mg). ^1H NMR (600

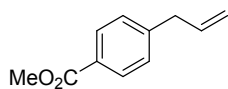
MHz, CDCl_3) δ 7.96 (d, J = 8.2 Hz, 2H), 7.26 (d, J = 8.1 Hz, 2H), 3.90 (s, 3H), 2.70 (q, J = 7.6 Hz, 2H), 1.25 (t, J = 7.6 Hz, 3H); ^{13}C NMR (151 MHz, CDCl_3) δ 167.22, 149.76, 129.71, 127.90, 127.65, 51.96, 28.96, 15.23.



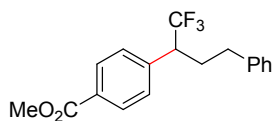
(13b). Colorless oil, TLC R_f (hexane: EtOAc = 6 : 1) = 0.55, 46% yield (35 mg). ^1H NMR (600 MHz, CDCl_3) δ 8.05 (d, J = 8.3 Hz, 2H), 7.47 (d, J = 8.2 Hz, 2H), 5.11 (dd, J = 8.3, 6.1 Hz, 2H), 4.77 (t, J = 6.4 Hz, 2H), 4.32 – 4.22 (m, 1H), 3.92 (s, 3H); ^{13}C NMR (151 MHz, CDCl_3) δ 166.85, 146.76, 130.12, 128.99, 126.83, 78.41, 52.13, 40.27. MS (EI) m/z 192 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{11}\text{H}_{12}\text{O}_3$ 192.0786, found 192.0777.



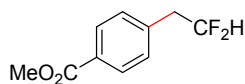
(13c). Colorless oil, TLC R_f (hexane: EtOAc = 6 : 1) = 0.55, 72% yield (64 mg). ^1H NMR (600 MHz, CDCl_3) δ 8.00 (d, J = 8.3 Hz, 2H), 7.36 (d, J = 8.2 Hz, 2H), 4.16 (q, J = 7.1 Hz, 2H), 3.91 (s, 3H), 3.67 (s, 2H), 1.25 (t, J = 7.1 Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 170.86, 166.8, 139.29, 129.85, 129.35, 129.01, 61.10, 52.10, 41.38, 14.15. MS (EI) m/z 222 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{12}\text{H}_{14}\text{O}_4$ 222.0892, found 222.0891.



(13d). Colorless oil, TLC R_f (hexane: EtOAc = 6 : 1) = 0.65, 71% yield (50 mg). ^1H NMR (600 MHz, CDCl_3) δ 7.97 (d, J = 8.2 Hz, 2H), 7.26 (d, J = 8.0 Hz, 2H), 5.95 (ddt, J = 16.9, 10.2, 6.7 Hz, 1H), 5.13 – 5.06 (m, 2H), 3.90 (s, 3H), 3.44 (d, J = 6.7 Hz, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 167.12, 145.51, 136.41, 129.78, 128.63, 128.10, 116.60, 52.01, 40.16. MS (EI) m/z 176 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{11}\text{H}_{12}\text{O}_2$ 176.0837, found 176.0833.

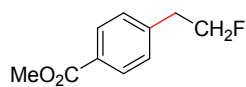


(13e). Light yellow solid, TLC R_f (hexane: EtOAc = 10 : 1) = 0.60, 75% yield (97 mg). ^1H NMR (600 MHz, CDCl_3) δ 8.07 (d, J = 8.3 Hz, 2H), 7.37 (d, J = 8.2 Hz, 2H), 7.27 (d, J = 7.7 Hz, 2H), 7.21 (s, 1H), 7.06 (d, J = 7.4 Hz, 2H), 3.94 (s, 3H), 3.28 (m, 1H), 2.57 (m, 1H), 2.38 (m, 2H), 2.22 (m, 1H). ^{19}F NMR (376 MHz, CDCl_3) δ -69.40 (s, 3F); ^{13}C NMR (151 MHz, CDCl_3) δ 166.68, 140.17, 139.54, 130.25, 130.03, 129.30, 128.60, 128.38, 126.89 (q, J = 191.8 Hz), 126.40, 52.25, 49.11 (q, J = 26.9 Hz), 32.38, 30.02. MS (EI) m/z 322 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{18}\text{H}_{17}\text{F}_3\text{O}_2$ 322.1181, found 322.1190.

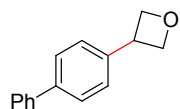


(13f). Light yellow oil, TLC R_f (hexane: EtOAc = 10 : 1) = 0.55, 41% yield (33 mg). ^1H

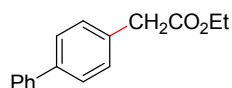
NMR (600 MHz, CDCl₃) δ 8.02 (d, J = 8.2 Hz, 2H), 7.34 (d, J = 8.0 Hz, 2H), 5.95 (tt, J = 56.3, 4.5 Hz, 1H), 3.92 (s, 2H), 3.20 (td, J = 17.3, 4.4 Hz, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 166.80 (s), 137.52 (s), 129.95 (s), 129.90 (s), 129.48 (s), 116.05 (t, J = 241.7 Hz), 52.18 (s), 40.82 (t, J = 22.2 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ -114.86 (s, 2F).



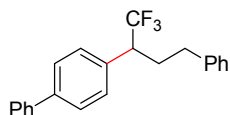
(13g). Light yellow oil, TLC R_f (hexane: EtOAc = 10 : 1) = 0.55, 58% yield (42 mg). ¹H NMR (400 MHz, CDCl₃) δ 7.99 (d, J = 8.3 Hz, 2H), 7.31 (d, J = 8.1 Hz, 2H), 4.65 (dt, J = 47.0, 6.3 Hz, 2H), 3.91 (s, 3H), 3.07 (dt, J = 24.3, 6.3 Hz, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 167.08 (s), 142.78 (s), 130.00 (s), 129.12 (s), 128.88 (s), 83.60 (d, J = 169.7 Hz), 52.16 (s), 37.04 (d, J = 20.6 Hz); ¹⁹F NMR (377 MHz, CDCl₃) δ -216.10 (s, 1F).



(14b). Colorless oil, TLC R_f (hexane: EtOAc = 20 : 1) = 0.60, 57% yield (48 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.59 (d, J = 7.0 Hz, 2H), 7.57 (d, J = 7.0 Hz, 2H), 7.50 – 7.41 (m, 4H), 7.35 (t, J = 7.4 Hz, 1H), 5.10 (dd, J = 8.4, 6.1 Hz, 2H), 4.81 (t, J = 6.4 Hz, 2H), 4.31 – 4.22 (m, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 140.76, 140.61, 140.07, 128.83, 127.50, 127.34, 127.28, 127.06, 78.92, 40.09. MS (EI) m/z 210 (M^+); HRMS (EI-TOF) m/z [M]⁺ Calcd for C₁₅H₁₄O 210.1045, found 210.1054.

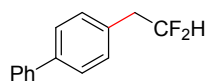


(14c). Colorless oil, TLC R_f (hexane: EtOAc = 10 : 1) = 0.50, 87% yield (84 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.56 (d, J = 7.7 Hz, 2H), 7.54 (d, J = 7.7 Hz, 2H), 7.42 (t, J = 7.7 Hz, 2H), 7.34 – 7.31 (m, 3H), 4.16 (q, J = 7.1 Hz, 2H), 3.64 (s, 2H), 1.26 (t, J = 7.1 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 171.65, 140.85, 140.06, 133.22, 129.71, 128.79, 127.34, 127.29, 127.10, 60.97, 41.08, 14.24. MS (EI) m/z 240 (M^+); HRMS (EI-TOF) m/z [M]⁺ Calcd for C₁₆H₁₆O₂ 240.1150, found 240.1144.

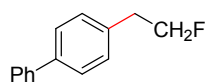


(14e). Light yellow solid, TLC R_f (hexane: EtOAc = 10 : 1) = 0.60, 90% yield (123 mg). ¹H NMR (600 MHz, CDCl₃) δ 7.61 (d, J = 8.2 Hz, 4H), 7.50 – 7.42 (m, 2H), 7.39 – 7.35 (m, 3H), 7.29 (t, J = 7.5 Hz, 2H), 7.21 (t, J = 7.3 Hz, 1H), 7.11 (d, J = 7.3 Hz, 2H), 3.26 (m, 1H), 2.63 (m, 1H), 2.46 (m, 1H), 2.41 – 2.31 (m, 1H), 2.31 – 2.17 (m, 1H); ¹⁹F NMR (376 MHz, CDCl₃) δ -69.18 (s, 3F); ¹³C NMR (151 MHz, CDCl₃) δ 141.36, 140.79, 140.69, 133.59, 129.80, 129.04, 128.74, 128.64,

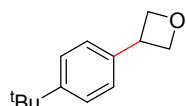
127.71, 127.67, 127.32, 127.15 (q, $J = 279.4$ Hz), 49.03 (q, $J = 26.5$ Hz), 32.72, 30.34. MS (EI) m/z 340 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $C_{22}H_{19}F_3$ 340.1439, found 340.1435.



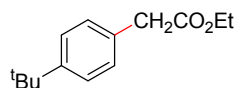
(14f). Light yellow solid, TLC R_f (hexane: EtOAc = 10 : 1) = 0.65, 75% yield (65 mg). 1H NMR (600 MHz, $CDCl_3$) δ 7.56 (t, $J = 8.6$ Hz, 4H), 7.42 (t, $J = 7.6$ Hz, 2H), 7.37 – 7.27 (m, 3H), 5.94 (tt, $J = 56.6$, 4.6 Hz, 1H), 3.16 (td, $J = 17.3$, 4.5 Hz, 2H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 140.80 (s), 140.64 (s), 131.58 (t, $J = 5.9$ Hz), 130.34 (s), 128.94 (s), 127.56 (s), 127.53 (s), 127.21 (s), 116.75 (t, $J = 241.5$ Hz), 40.69 (t, $J = 21.9$ Hz); ^{19}F NMR (377 MHz, $CDCl_3$) δ -114.70 (s, 2F).



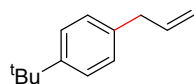
(14g). White solid, TLC R_f (hexane: EtOAc = 10 : 1) = 0.65, 78% yield (62 mg). 1H NMR (600 MHz, $CDCl_3$) δ 7.57 (d, $J = 7.5$ Hz, 2H), 7.54 (d, $J = 7.7$ Hz, 2H), 7.42 (t, $J = 7.4$ Hz, 2H), 7.34 (d, $J = 7.2$ Hz, 1H), 7.30 (d, $J = 7.7$ Hz, 2H), 4.66 (dt, $J = 47.1$, 6.5 Hz, 2H), 3.05 (dt, $J = 23.4$, 6.4 Hz, 2H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 141.04 (s), 139.85 (s), 136.33 (d, $J = 6.2$ Hz), 129.51 (s), 128.89 (s), 127.44 (s), 127.33 (s), 127.18 (s), 84.14 (d, $J = 169.2$ Hz), 36.71 (d, $J = 20.4$ Hz); ^{19}F NMR (377 MHz, $CDCl_3$) δ -215.25 (s, 1F).



(15b). Colorless oil, TLC R_f (hexane: EtOAc = 20 : 1) = 0.60, 81% yield (62 mg). 1H NMR (600 MHz, $CDCl_3$) δ 7.40 (d, $J = 7.9$ Hz, 2H), 7.34 (d, $J = 8.0$ Hz, 2H), 5.08 – 5.01 (m, 2H), 4.79 (t, $J = 6.4$ Hz, 2H), 4.21 (p, $J = 7.7$ Hz, 1H), 1.33 (s, 9H); ^{13}C NMR (151 MHz, $CDCl_3$) δ 150.03, 138.51, 126.58, 125.67, 79.08, 39.96, 34.52, 31.38. MS (EI) m/z 190 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $C_{13}H_{18}O$ 190.1358, found 190.1366.

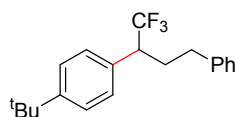


(15c). Colorless oil, TLC R_f (hexane: EtOAc = 10 : 1) = 0.55, 75% yield (66 mg). 1H NMR (600 MHz, $CDCl_3$) δ 7.34 (d, $J = 8.3$ Hz, 2H), 7.22 (d, $J = 8.3$ Hz, 2H), 4.15 (q, $J = 7.1$ Hz, 2H), 3.58 (s, 2H), 1.31 (s, 9H), 1.26 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (151 MHz, $CDCl_3$) δ 171.85, 149.88, 131.12, 128.90, 125.51, 60.80, 40.90, 34.46, 31.36, 14.22. MS (EI) m/z 220 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $C_{14}H_{20}O_2$ 220.1463, found 220.1460.



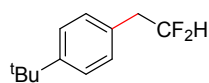
(15d). Colorless oil, TLC R_f (hexane) = 0.50, 69% yield (48 mg). ^1H NMR (600 MHz, CDCl_3)

δ 7.32 (d, J = 8.3 Hz, 2H), 7.13 (d, J = 8.3 Hz, 2H), 5.97 (ddt, J = 16.9, 10.0, 6.8 Hz, 1H), 5.07 (dddd, J = 10.0, 4.2, 3.2, 1.4 Hz, 2H), 3.36 (d, J = 6.8 Hz, 2H), 1.31 (s, 9H); ^{13}C NMR (151 MHz, CDCl_3) δ 148.91, 137.64, 137.06, 128.21, 125.34, 115.64, 39.76, 34.40, 31.43. MS (EI) m/z 174 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{13}\text{H}_{18}$ 174.1409, found 174.1411.



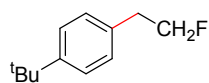
(15e). Light yellow oil, TLC R_f (hexane) = 0.50, 51% yield (65 mg). ^1H NMR (600 MHz, CDCl_3)

δ 7.39 (d, J = 8.3 Hz, 2H), 7.27 (dd, J = 14.7 Hz, 7.1 Hz, 2H), 7.22 (d, J = 13.6 Hz, 2H), 7.20 (t, J = 7.3 Hz, 1H), 7.10 (d, J = 7.3 Hz, 2H), 3.27–3.10 (m, 1H), 2.59 (m, 1H), 2.42 (m, 1H), 2.36–2.26 (m, 1H), 2.20 (m, 1H), 1.34 (s, 9H); ^{19}F NMR (376 MHz, CDCl_3) δ -69.70 (s, 3F); ^{13}C NMR (151 MHz, CDCl_3) δ 151.30, 141.01, 131.48, 128.97, 128.69, 128.64, 126.38, 125.64, 127.52 (q, J = 280.1 Hz), 48.90 (q, J = 26.5 Hz), 34.76, 32.80, 31.54, 30.38. MS (EI) m/z 320 (M^+); HRMS (EI-TOF) m/z [M] $^+$ Calcd for $\text{C}_{20}\text{H}_{23}\text{F}_3$ 320.1752, found 320.1760.



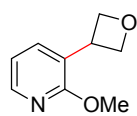
(15f). Light yellow oil, TLC R_f (hexane) = 0.50, 44% yield (35 mg). ^1H NMR (600 MHz, CDCl_3)

δ 7.40 (s, 2H), 7.22 (s, 2H), 5.95 (t, J = 56.6 Hz, 1H), 3.14 (t, J = 17.3 Hz, 2H), 1.35 (s, 9H); ^{13}C NMR (151 MHz, CDCl_3) δ 150.62 (s), 129.67 (s), 125.84 (s), 117.02 (t, J = 241.3 Hz), 40.63 (t, J = 21.8 Hz), 34.70 (s), 31.53 (s); ^{19}F NMR (376 MHz, CDCl_3) δ -114.66 (dt, J = 56.7, 17.4 Hz, 2F).



(15g). Colorless oil, TLC R_f (hexane) = 0.50, 56% yield (40 mg). ^1H NMR (600 MHz, CDCl_3)

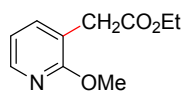
δ 7.34 (d, J = 8.3 Hz, 2H), 7.17 (d, J = 8.2 Hz, 2H), 4.62 (dt, J = 47.1, 6.7 Hz, 2H), 2.99 (dt, J = 22.7, 6.7 Hz, 2H), 1.31 (s, 9H); ^{13}C NMR (151 MHz, CDCl_3) δ 149.56 (s), 133.98 (d, J = 6.7 Hz), 128.66 (s), 125.49 (s), 84.20 (d, J = 168.7 Hz), 36.40 (d, J = 20.3 Hz), 34.44 (s), 31.38 (s); ^{19}F NMR (376 MHz, CDCl_3) δ -214.86 (s, 1F).



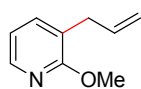
(16b). Yellow oil, TLC R_f (hexane: EtOAc = 4 : 1) = 0.40, 42% yield (28 mg). ^1H NMR (600 MHz, CDCl_3)

δ 8.07 (d, J = 5.0 Hz, 1H), 7.55 (d, J = 7.2 Hz, 1H), 6.92 (dd, J = 7.2, 5.1 Hz, 1H), 5.01 (dd, J = 8.5, 5.9 Hz, 2H), 4.81 (d, J = 13.3 Hz, 2H), 4.43 (p, J = 8.0 Hz, 1H), 3.93 (s, 3H); ^{13}C NMR (151 MHz, CDCl_3) δ 161.56, 145.06, 134.94, 123.58, 116.76, 65.86, 53.36, 34.55.

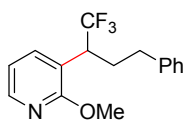
MS (EI) m/z 165 (M^+); HRMS (EI-TOF) m/z $[M]^+$ Calcd for $C_9H_{11}NO_2$ 165.0790, found 165.0788.



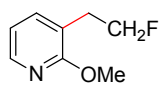
(16c). Colorless oil, TLC R_f (hexane: EtOAc = 4 : 1) = 0.40, 51% yield (40 mg). 1H NMR (600 MHz, $CDCl_3$) δ 8.09 (d, J = 4.9 Hz, 1H), 7.47 (d, J = 7.1 Hz, 1H), 6.89 – 6.81 (m, 1H), 4.17 (q, J = 7.1 Hz, 2H), 3.95 (s, 3H), 3.58 (s, 2H), 1.26 (t, J = 7.1 Hz, 3H); ^{13}C NMR (151 MHz, $CDCl_3$) δ 171.09, 162.12, 145.69, 139.02, 117.53, 116.71, 60.85, 53.46, 35.56, 14.20. MS (EI) m/z 195 (M^+); HRMS (EI-TOF) m/z $[M]^+$ Calcd for $C_{10}H_{13}NO_3$ 195.0895, found 195.0899.



(16d). Colorless oil, TLC R_f (hexane: EtOAc = 4 : 1) = 0.50, 67% yield (40 mg). 1H NMR (600 MHz, $CDCl_3$) δ 8.03 (dd, J = 5.0, 1.7 Hz, 1H), 7.39 (d, J = 6.4 Hz, 1H), 6.83 (dd, J = 7.1, 5.0 Hz, 1H), 5.97 (ddt, J = 16.9, 10.4, 6.7 Hz, 1H), 5.10 – 5.05 (m, 2H), 3.96 (s, 3H), 3.33 (d, J = 6.6 Hz, 2H); ^{13}C NMR (151 MHz, $CDCl_3$) δ 161.95, 144.48, 137.62, 135.67, 122.86, 116.73, 116.28, 53.32, 33.82. MS (EI) m/z 149 (M^+); HRMS (EI-TOF) m/z $[M]^+$ Calcd for $C_9H_{11}NO$ 149.0841, found 149.0839.



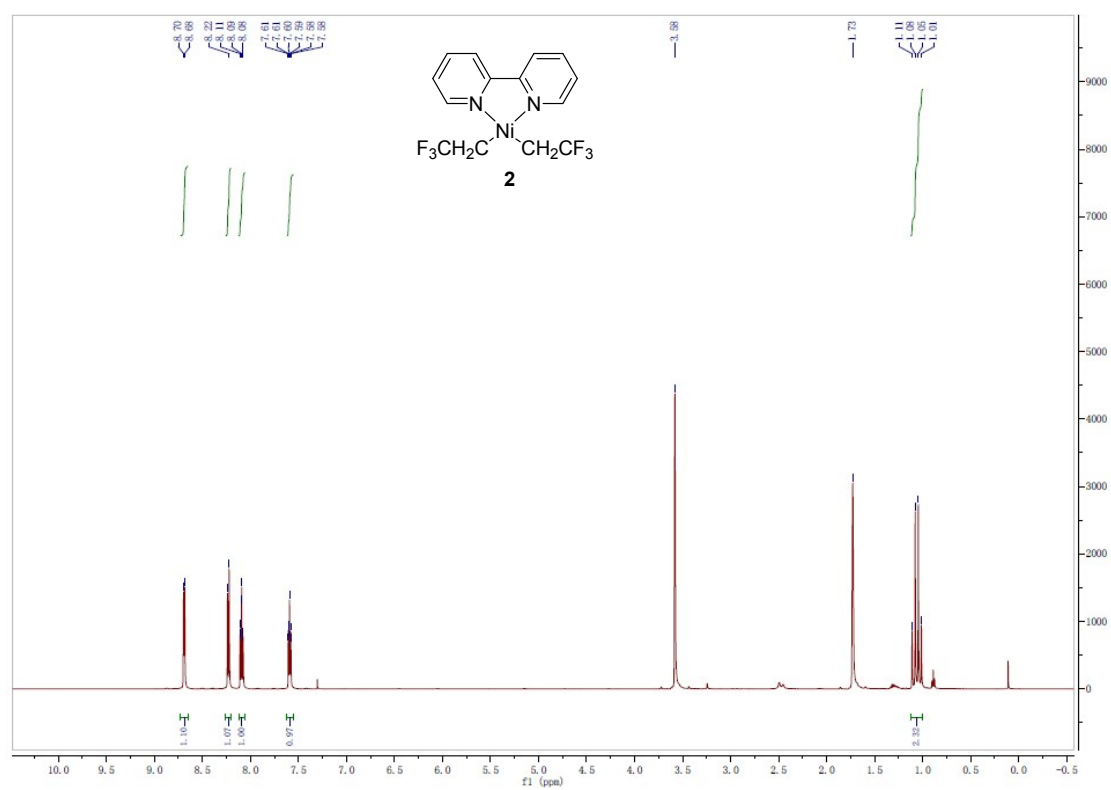
(16e). Colorless oil, TLC R_f (hexane: EtOAc = 5 : 1) = 0.40, 85% yield (100 mg). 1H NMR (600 MHz, $CDCl_3$) δ 8.16 (dd, J = 4.9, 1.8 Hz, 1H), 7.64 (d, J = 7.3 Hz, 1H), 7.29 – 7.25 (m, 2H), 7.19 (t, J = 7.4 Hz, 1H), 7.07 (d, J = 7.1 Hz, 2H), 6.94 (dd, J = 7.4, 5.0 Hz, 1H), 3.94 (s, 3H), 3.89 (m, 1H), 2.54 (m, 1H), 2.47 (m, 1H), 2.36 – 2.26 (m, 1H), 2.18 – 2.07 (m, 1H); ^{19}F NMR (376 MHz, $CDCl_3$) δ -69.54 (s, 3F); ^{13}C NMR (151 MHz, $CDCl_3$) δ 162.64, 146.68, 128.63, 128.54, 127.14 (q, J = 280.9 Hz), 53.85, 40.54 (q, J = 26.8 Hz), 32.76, 30.39. MS (EI) m/z 295 (M^+); HRMS (EI-TOF) m/z $[M]^+$ Calcd for $C_{16}H_{16}F_3NO$ 295.1184, found 295.1180.



(16g). Yellow oil, TLC R_f (hexane: EtOAc = 5 : 1) = 0.45, 62% yield (39 mg). 1H NMR (600 MHz, $CDCl_3$) δ 8.07 (d, J = 6.4 Hz, 1H), 7.46 (d, J = 7.1 Hz, 1H), 6.84 (dd, J = 7.1, 5.1 Hz, 1H), 4.64 (dt, J = 47.1, 6.3 Hz, 2H), 3.96 (s, 3H), 2.98 (dt, J = 23.8, 6.3 Hz, 2H); ^{13}C NMR (151 MHz, $CDCl_3$) δ 161.07 (s), 144.16 (s), 138.00 (s), 118.66 (d, J = 5.8 Hz), 115.75 (s), 81.27 (d, J = 167.4 Hz), 52.34 (s), 30.31 (d, J = 21.0 Hz); ^{19}F NMR (377 MHz, $CDCl_3$) δ -216.62 (s, 1F).

Table S4. The reference literatures of reported compounds

Known Compound Number	Reference
7a, 7b, 7d, 7e, 7f, 7g, 7h, 7i, 7k, 7n, 7q, 7r	Zhao, Y.; Hu, J. <i>Angew. Chem. Int. Ed.</i> 2012 , <i>51</i> , 1033.
7c	Leng, F.; Wang, Y.; Li, H.; Li, J.; Zou, D.; Wu, Y.; Wu, Y. <i>Chem. Commun.</i> 2013 , <i>49</i> , 10697.
7j, 7l	Xu, S.; Chen, H.-H.; Dai, J.-J.; Xu, H.-J. <i>Org. Lett.</i> 2014 , <i>16</i> , 2306.
7m	Kautzky, J. A.; Wang, T.; Evans, R. W.; MacMillan, D. W. C. <i>J. Am. Chem. Soc.</i> 2018 , <i>140</i> , 6522.
7p	Akira, S.; Tetsuo, Y.; Hiroaki, I.; Masayuki, N.; Keiko, T.; Noriko, O. <i>Bull. Chem. Soc. Jpn.</i> 1986 , <i>59</i> , 3905.
13a	Rushworth, P. J.; Hulcoop, D. G.; Fox, D. J. <i>J. Org. Chem.</i> 2013 , <i>78</i> , 9517.
13b, 13c, 13d, 13f, 14f, 15f	Zhang, X.; Yang, C. <i>Adv. Synth. Catal.</i> 2015 , <i>357</i> , 2721.
14c	Xie, P.; Xie, Y.; Qian, B.; Zhou, H.; Xia, C.; Huang, H. <i>J. Am. Chem. Soc.</i> 2012 , <i>134</i> , 9902.
15c	Chen, Z.; Wen, Y.; Fu, Y.; Chen, H.; Ye, M.; Luo, G. <i>Synlett</i> 2017 , <i>28</i> , 981.
15d	Mayer, M.; Czaplik, W. M.; Jacobi von Wangelin, A. <i>Adv. Synth. Catal.</i> 2010 , <i>352</i> , 2147.
16d	Struk, Ł.; Sośnicki, J. G. <i>Synthesis</i> 2012 , <i>44</i> , 735-746.
13g, 14g, 15g, 16g	Yang, Y.; Cai, J.; Luo, G.; Jiang, Y.; Su, Y.; Su, Y.; Li, C.; Zheng, Y.; Zeng, J.; Liu, Y. <i>Org. Chem. Front.</i> 2019 , doi:

V. Copies of ^1H NMR, ^{19}F NMR and ^{13}C NMR spectra**Figure S2. ^1H NMR of **2** in d_8 -THF**

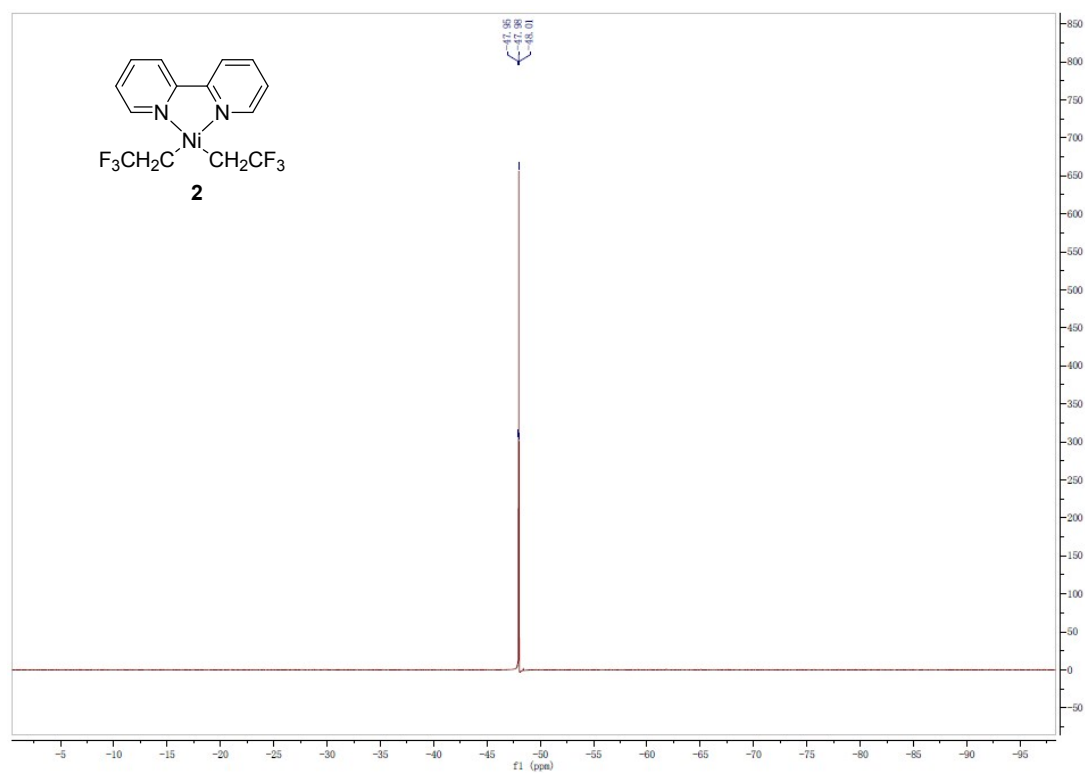


Figure S3. ¹⁹F NMR of **2** in *d*₈-THF

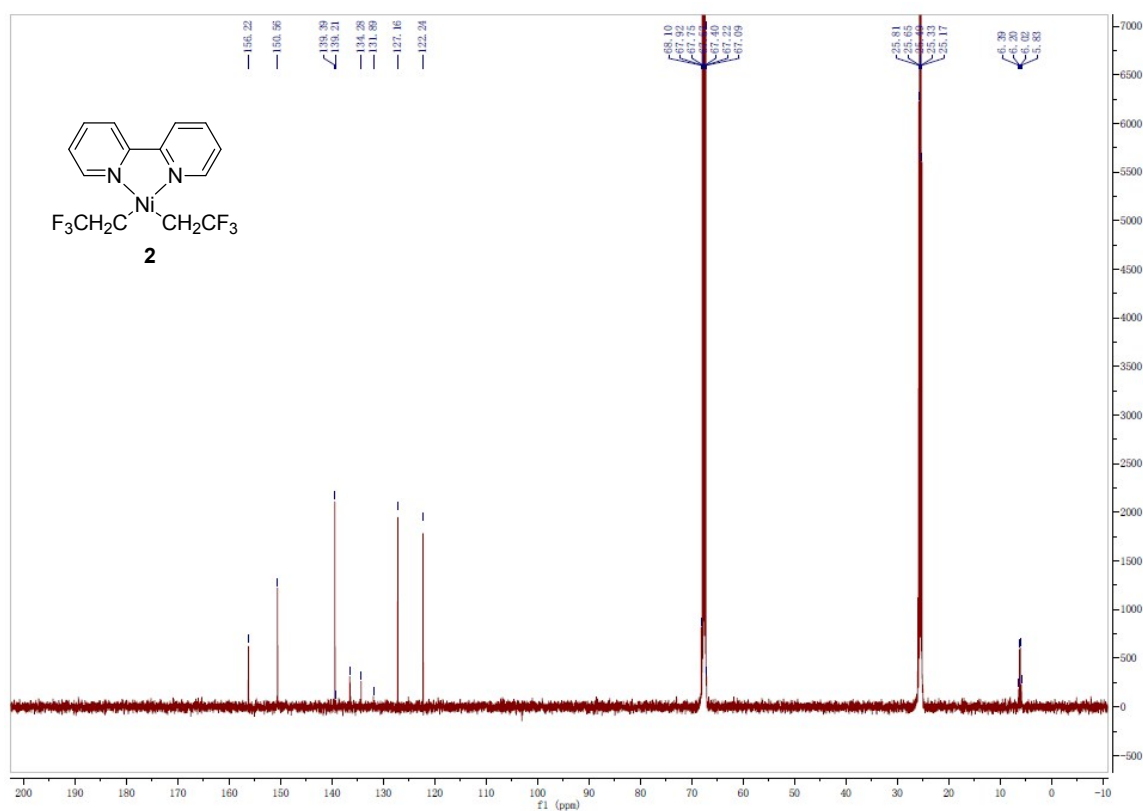


Figure S4. ¹³C NMR of **2** in *d*₈-THF

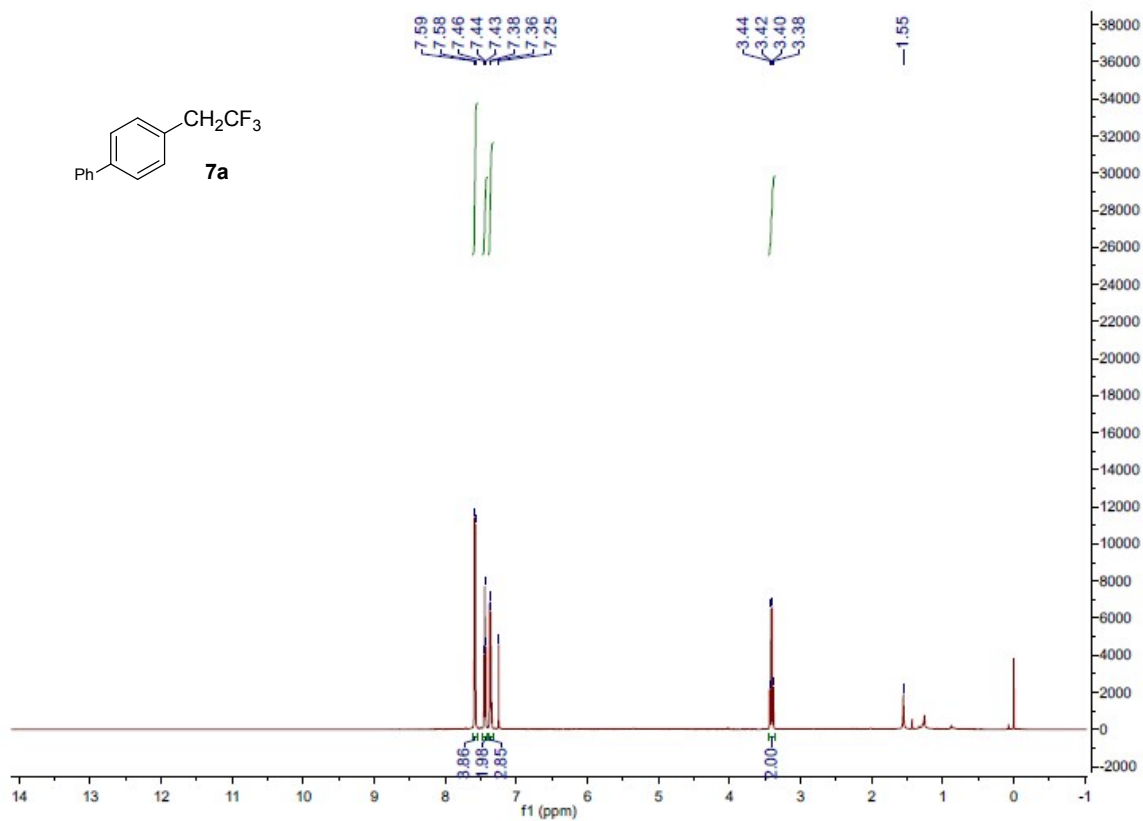


Figure S5. ¹H NMR of **7a** in CDCl₃

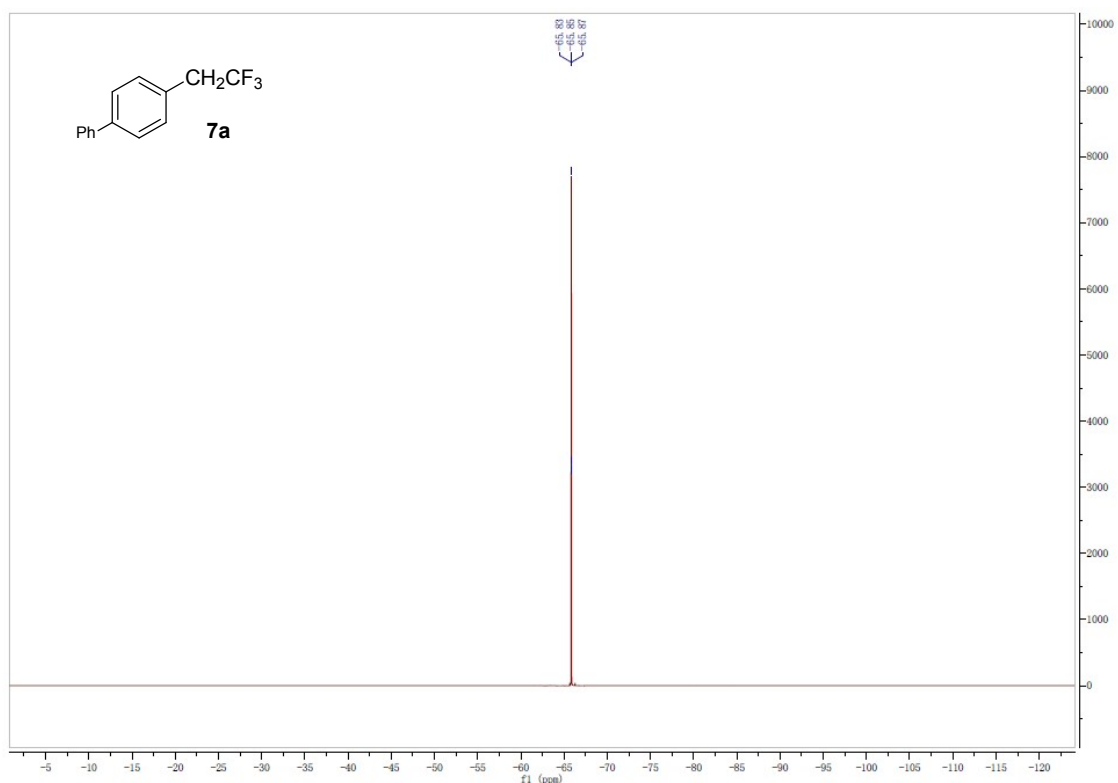


Figure S6. ¹⁹F NMR of **7a** in CDCl₃

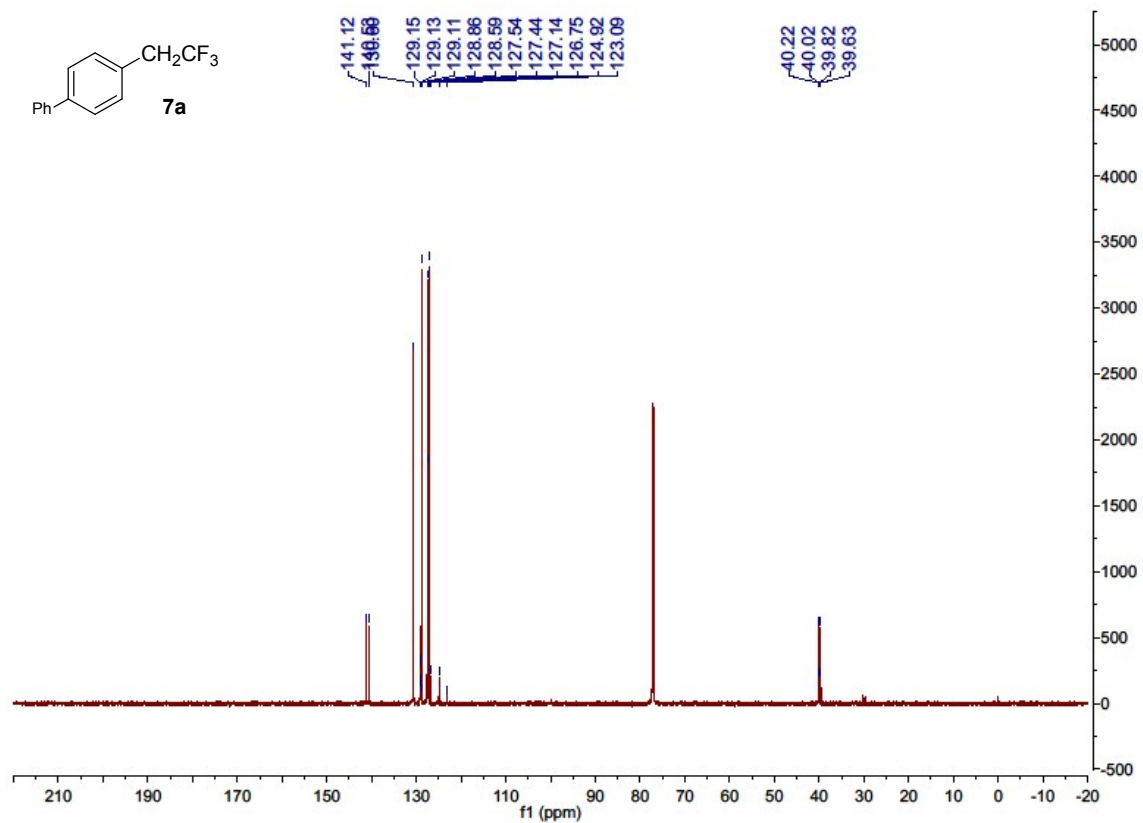


Figure S7. ¹³C NMR of **7a** in CDCl₃

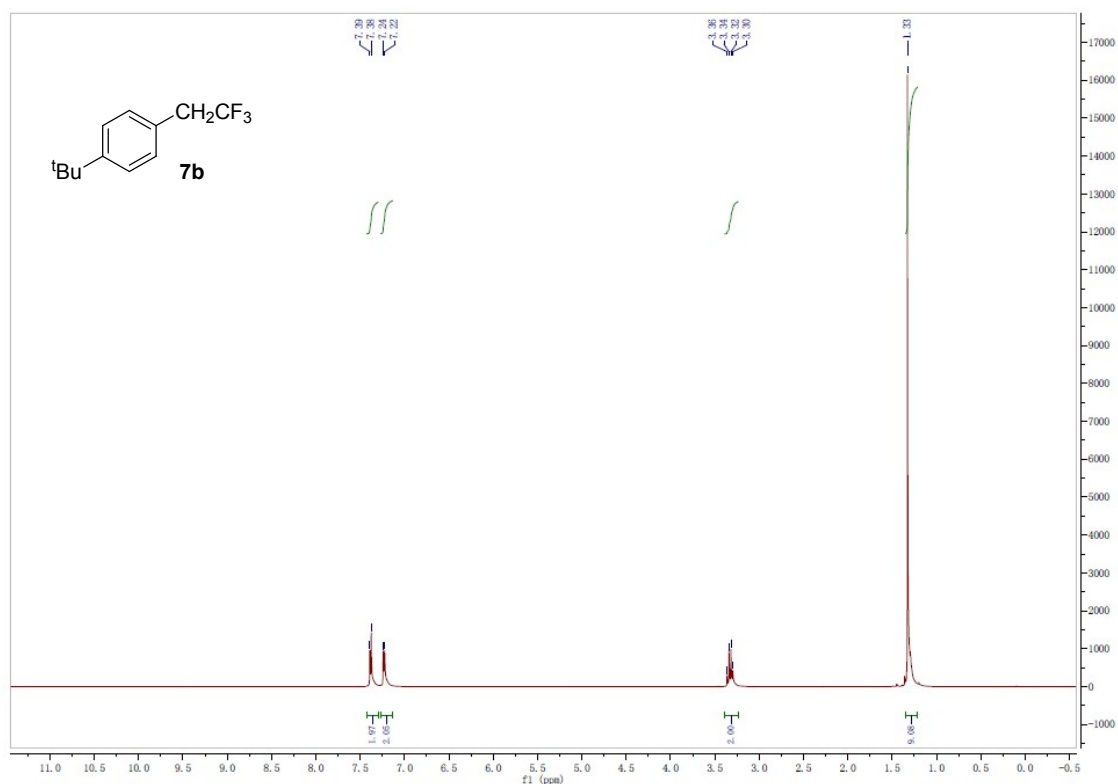


Figure S8. ¹H NMR of **7b** in CDCl₃

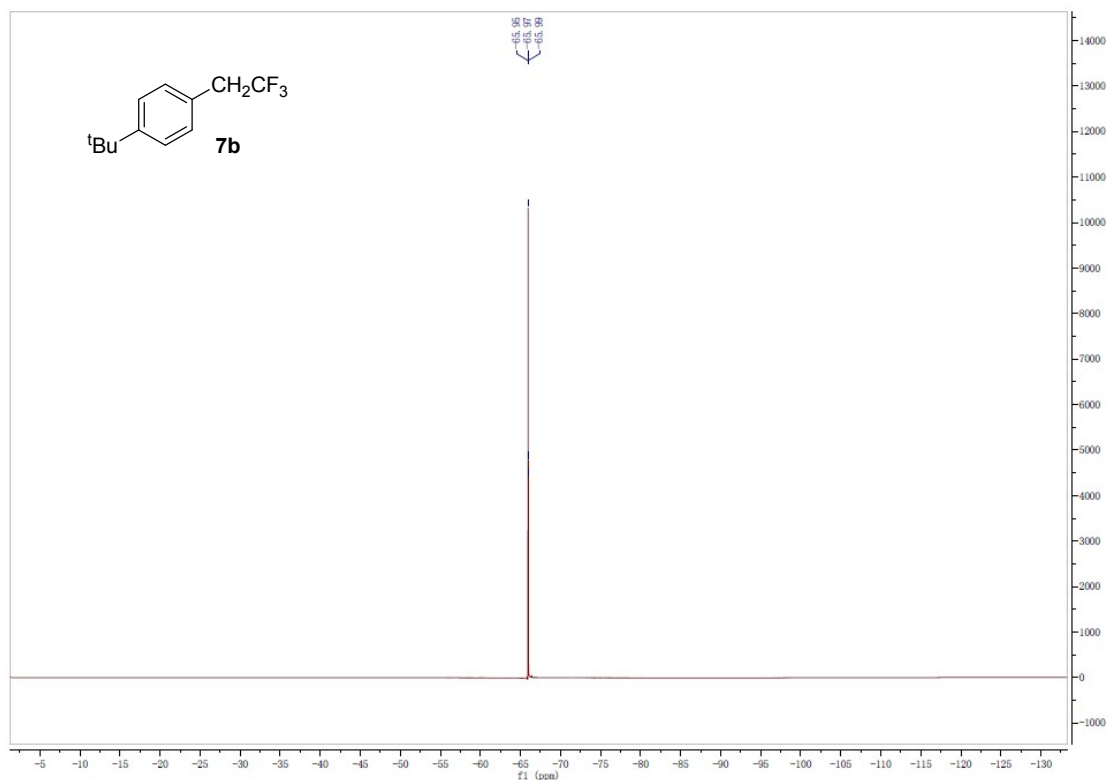


Figure S9. ¹⁹F NMR of **7b** in CDCl₃

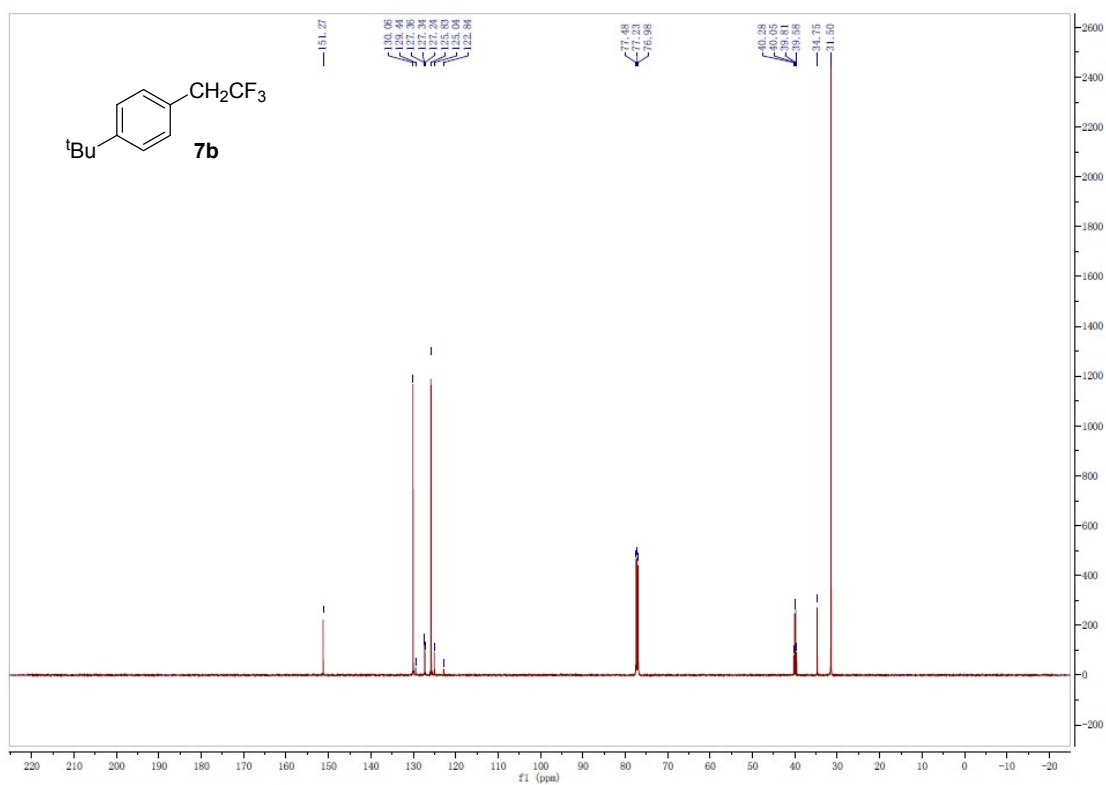


Figure S10. ¹³C NMR of **7b** in CDCl₃

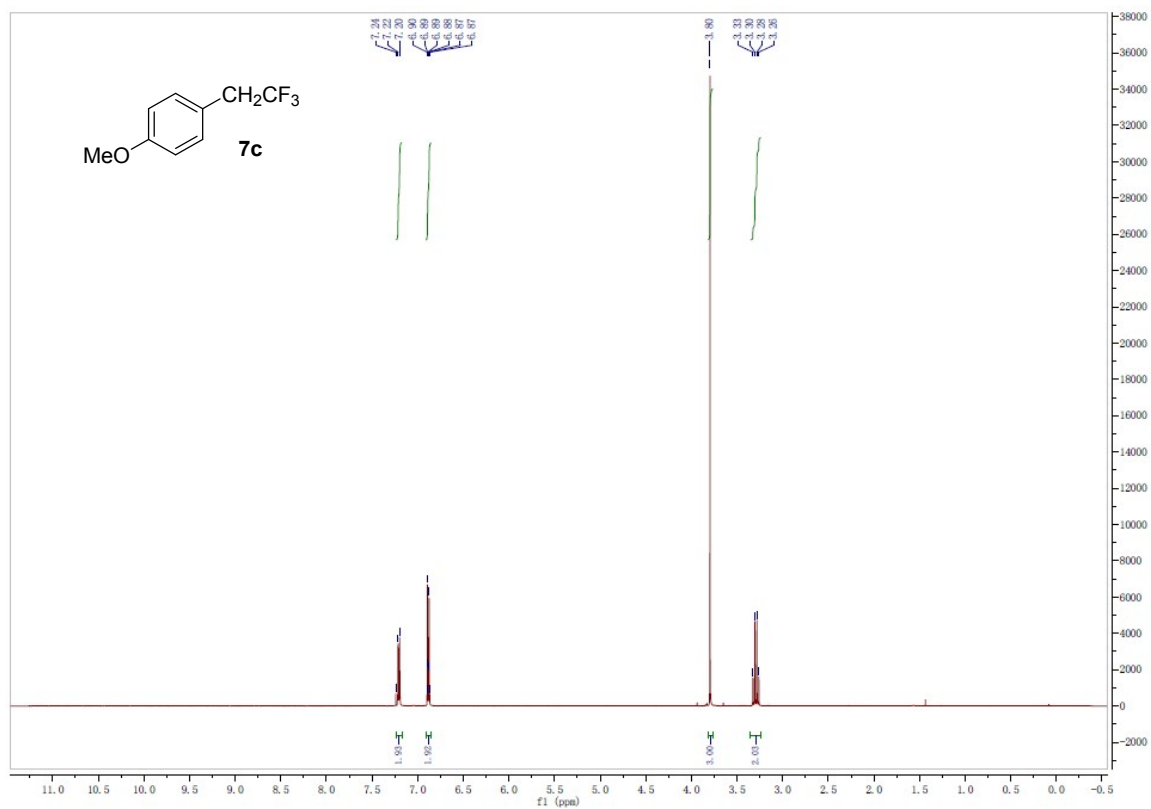


Figure S11. ¹H NMR of 7c in CDCl₃

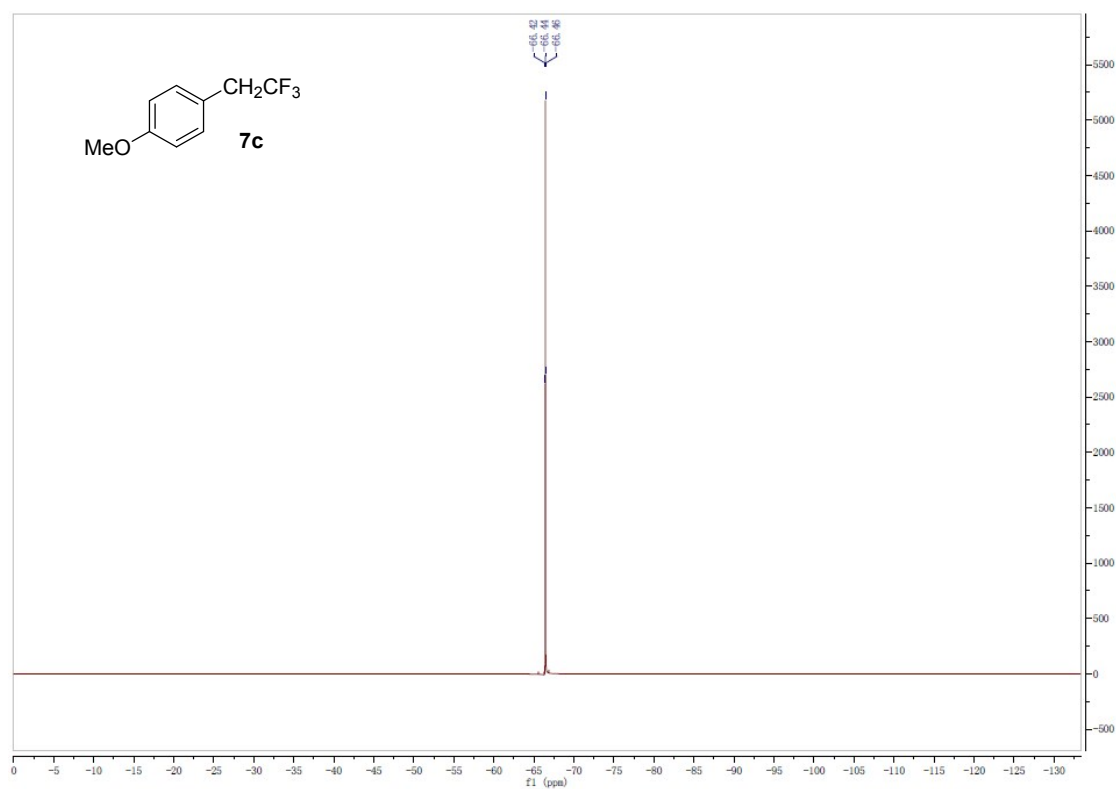


Figure S12. ¹⁹F NMR of 7c in CDCl₃

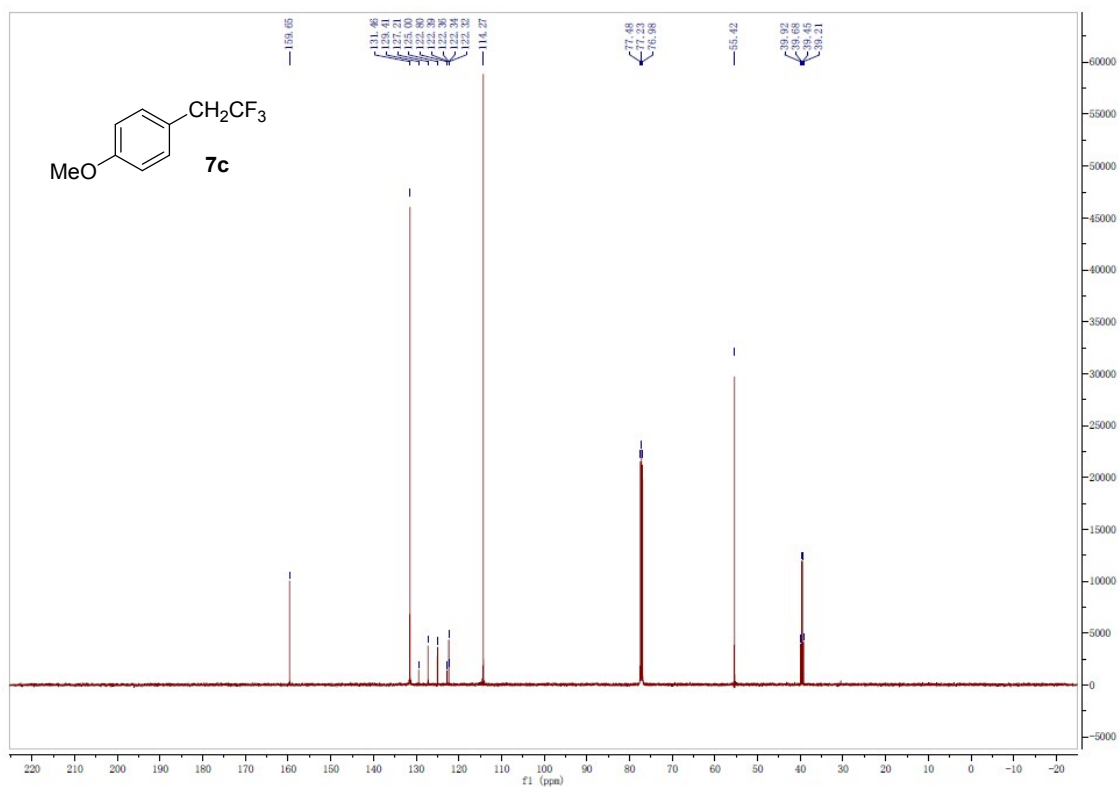
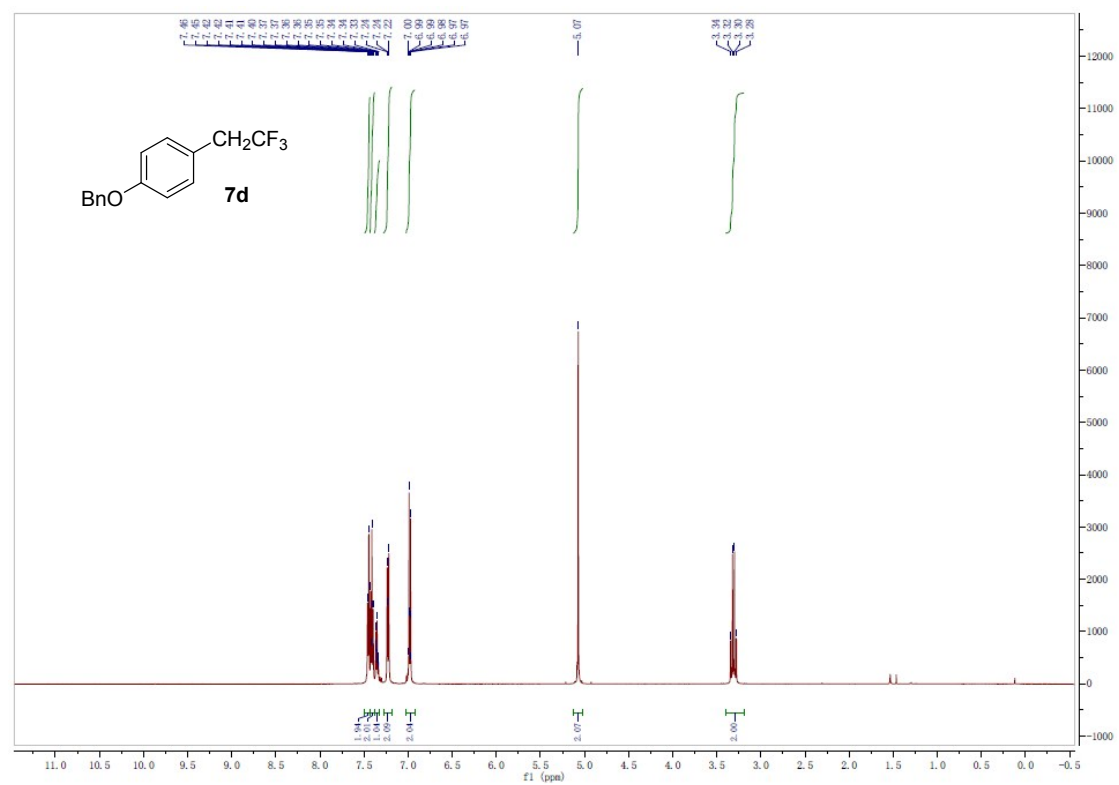


Figure S13. ¹³C NMR of **7c** in CDCl₃



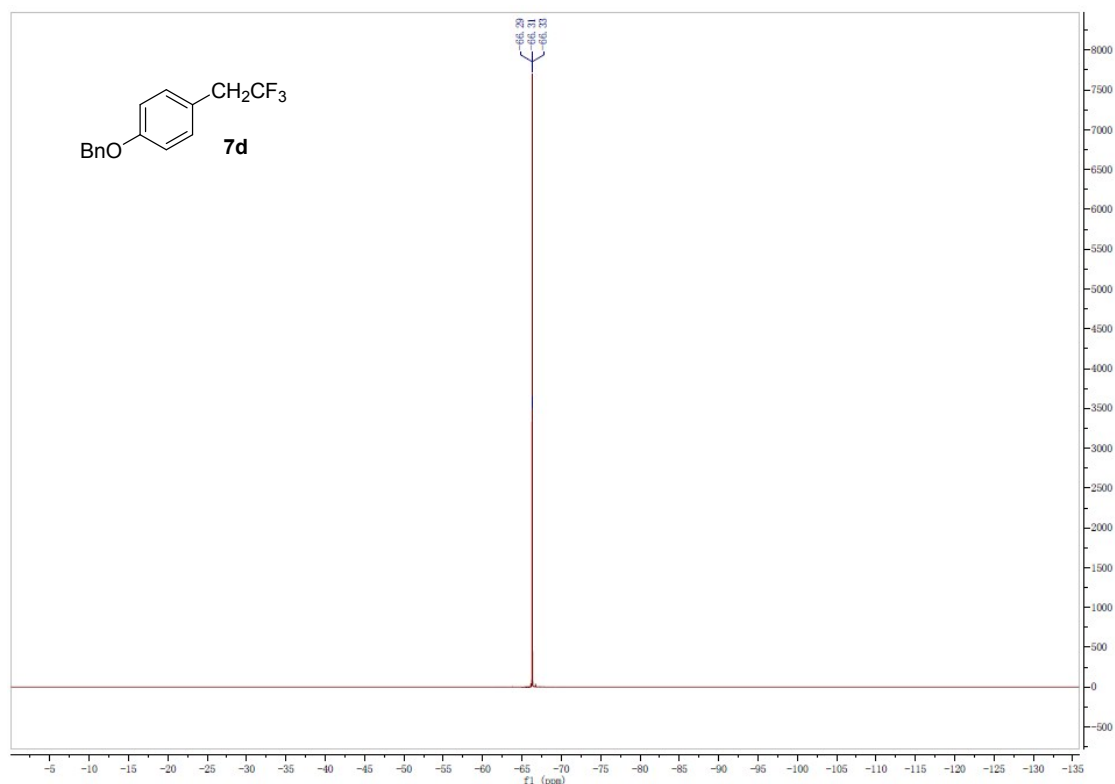


Figure S15. ¹⁹F NMR of **7d** in CDCl₃

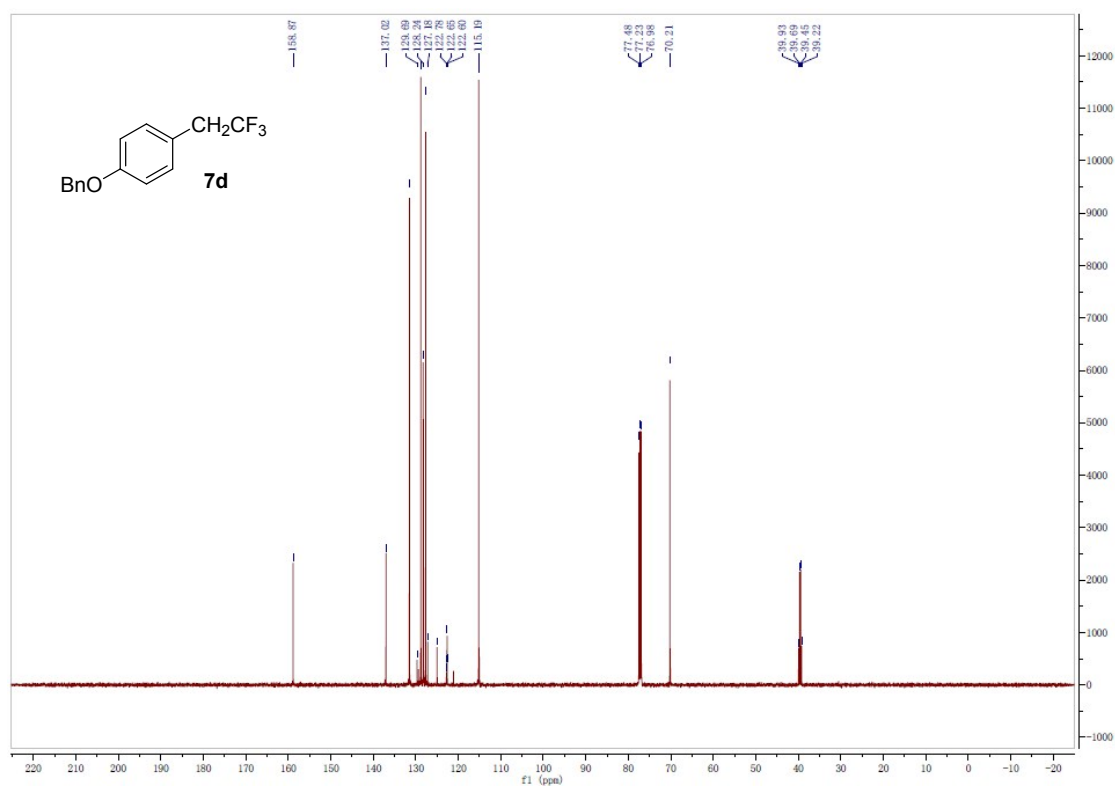


Figure S16. ¹³C NMR of **7d** in CDCl₃

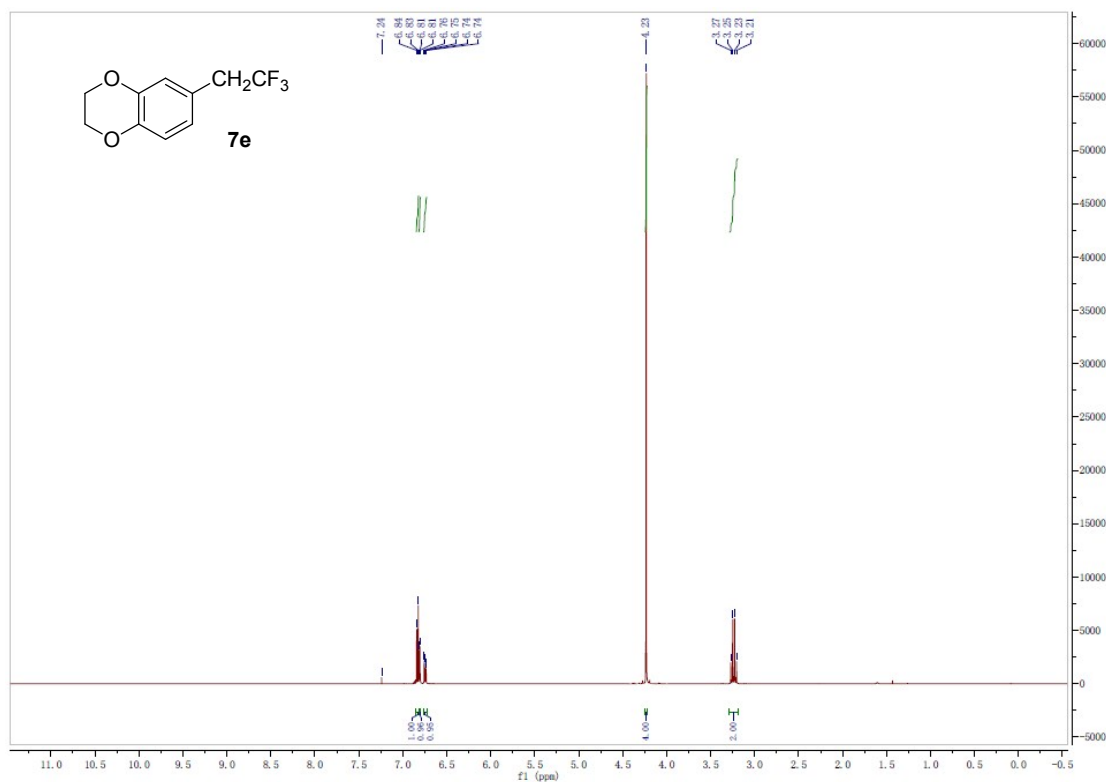


Figure S17. ¹H NMR of **7e** in CDCl₃

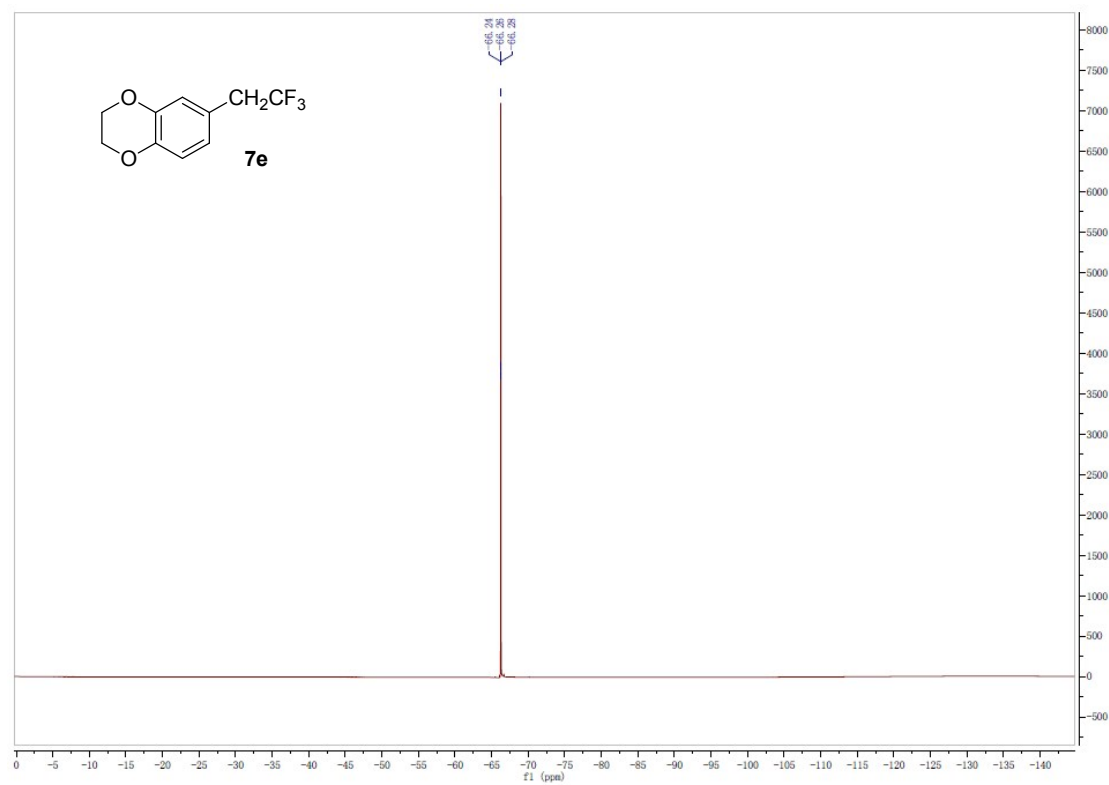


Figure S18. ¹⁹F NMR of **7e** in CDCl₃

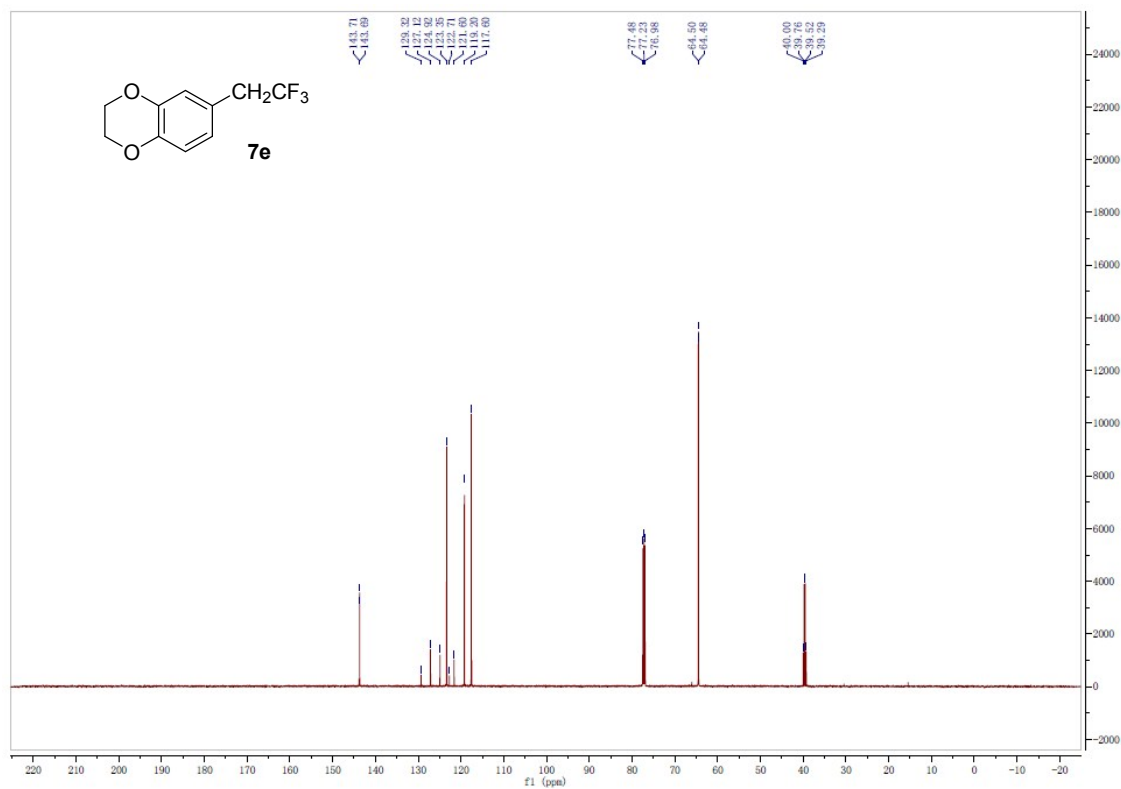


Figure S19. ¹³C NMR of **7e** in CDCl₃

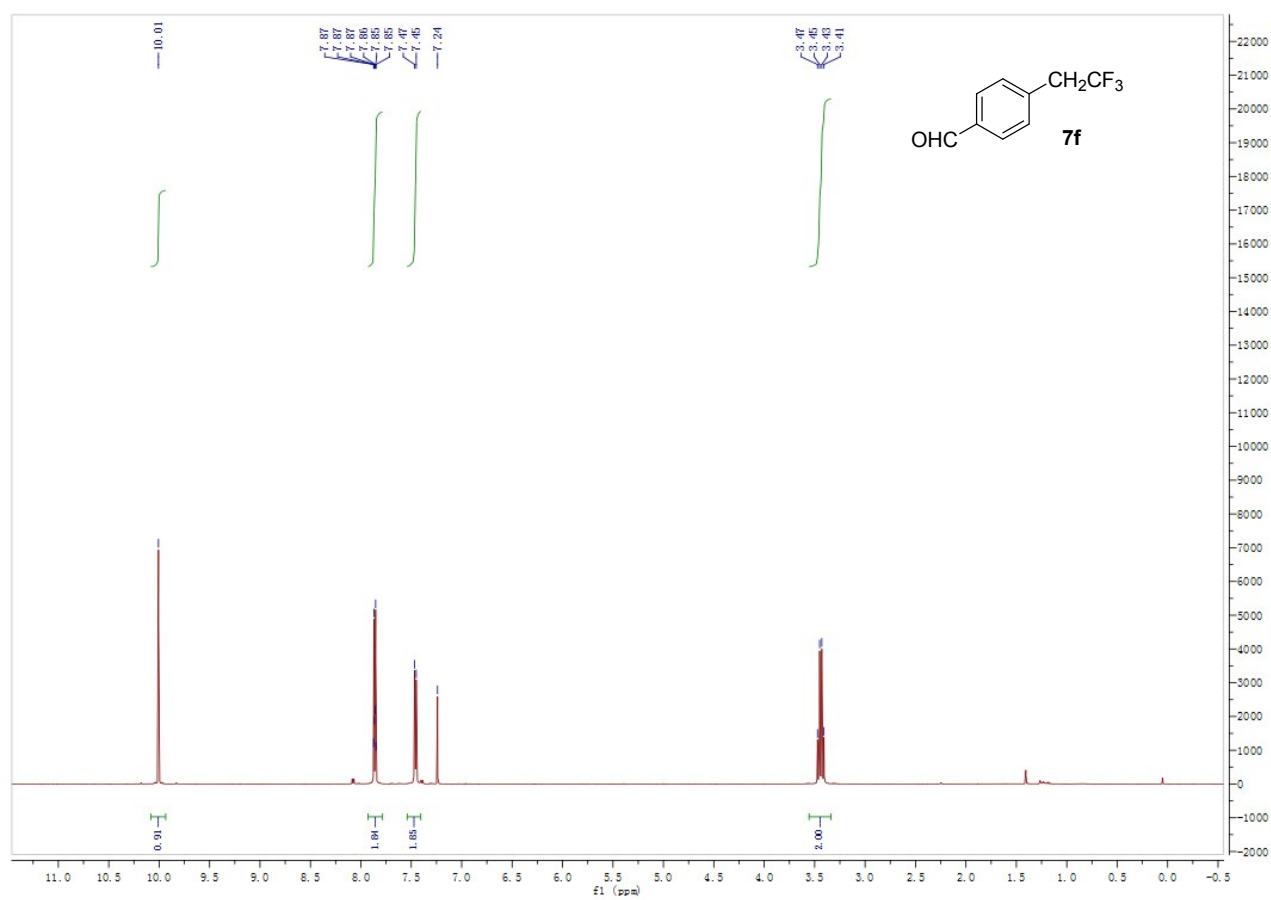


Figure S20. ¹H NMR of **7f** in CDCl₃



Figure S21. ¹⁹F NMR of **7f** in CDCl₃

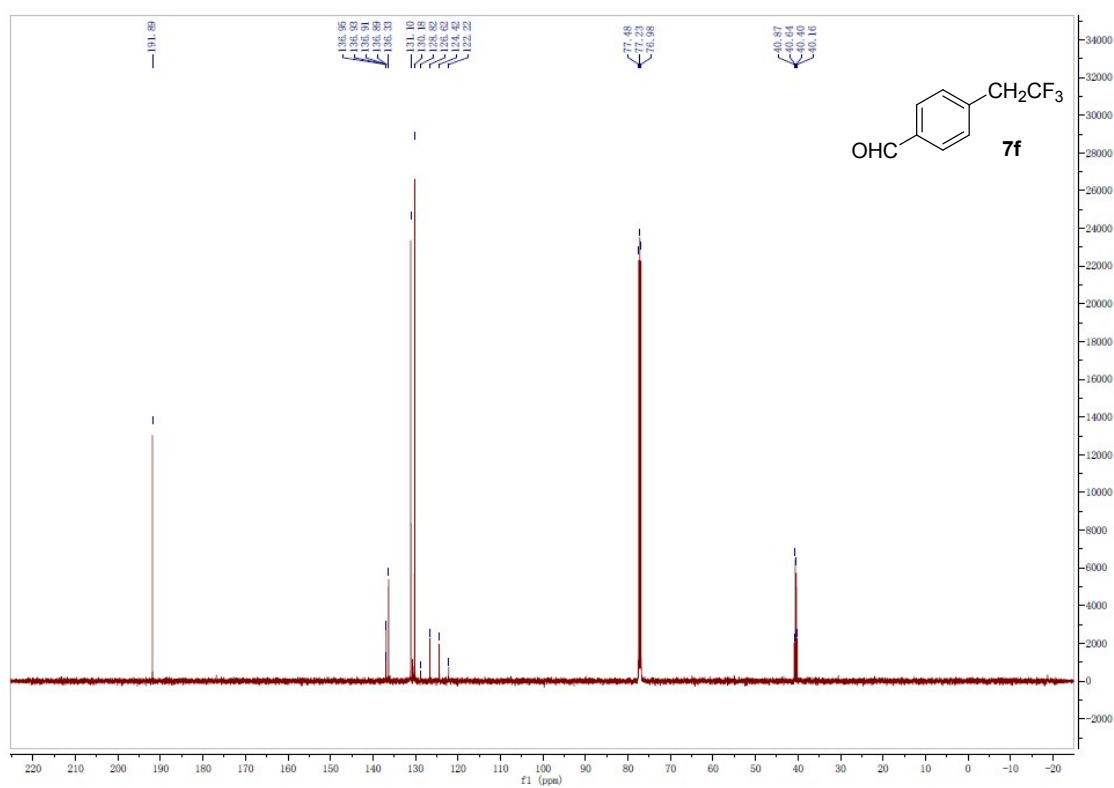


Figure S22. ¹³C NMR of **7f** in CDCl₃

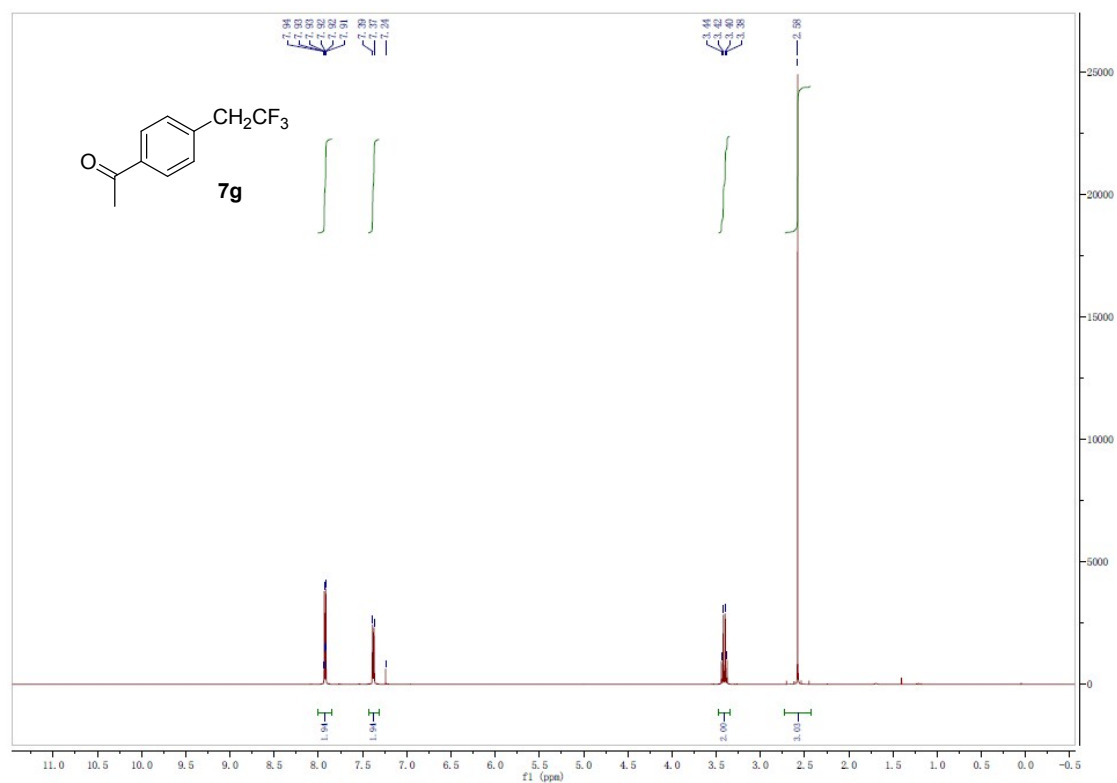


Figure S23. ¹H NMR of **7g** in CDCl₃

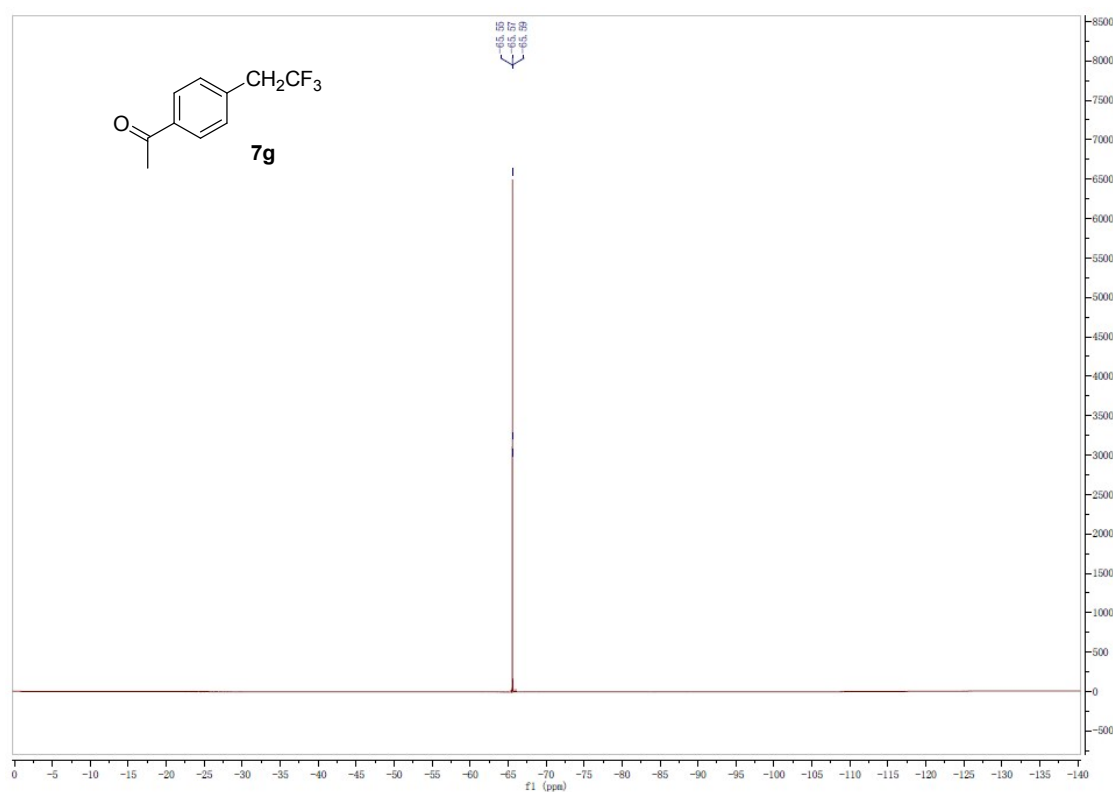


Figure S24. ¹⁹F NMR of **7g** in CDCl₃

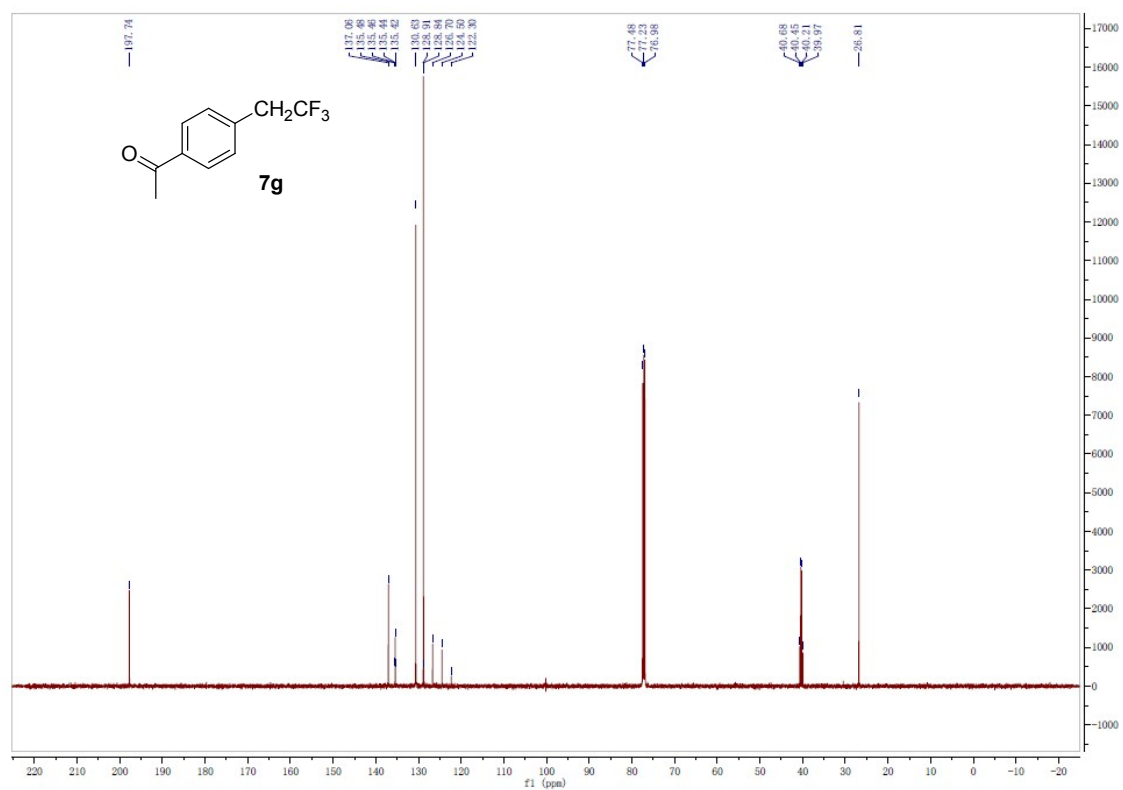


Figure S25. ¹³C NMR of **7g** in CDCl₃

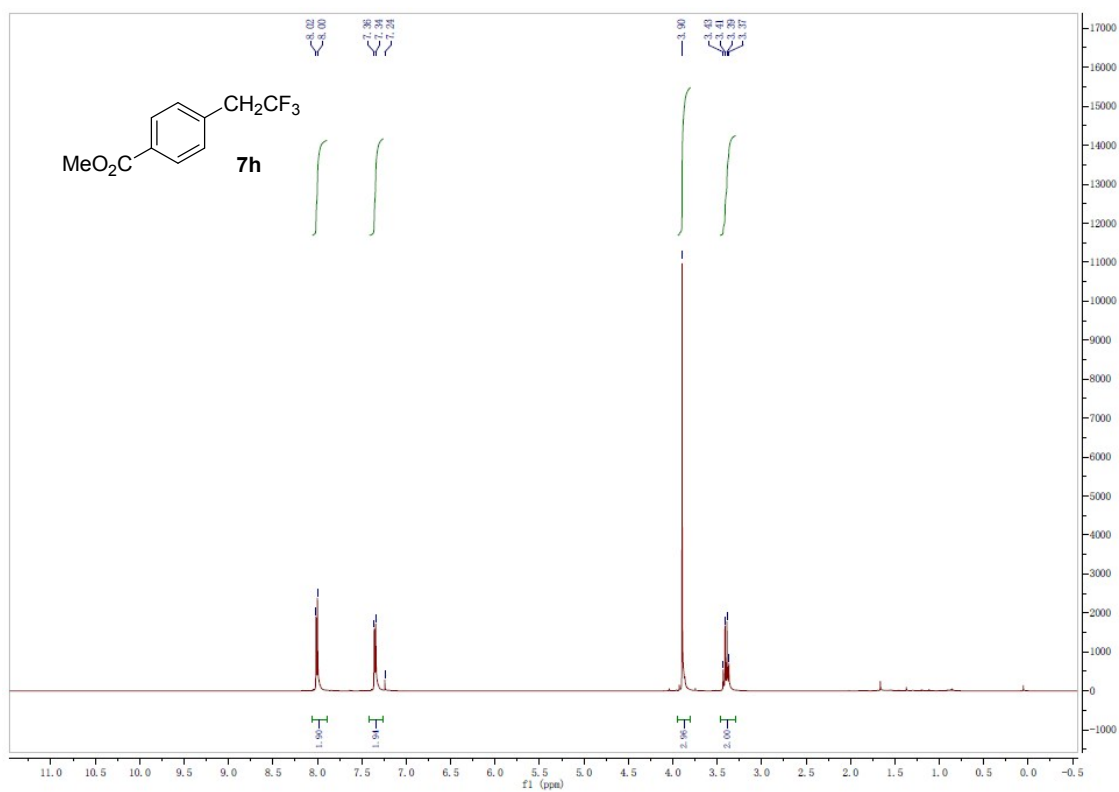


Figure S26. ¹H NMR of **7h** in CDCl₃

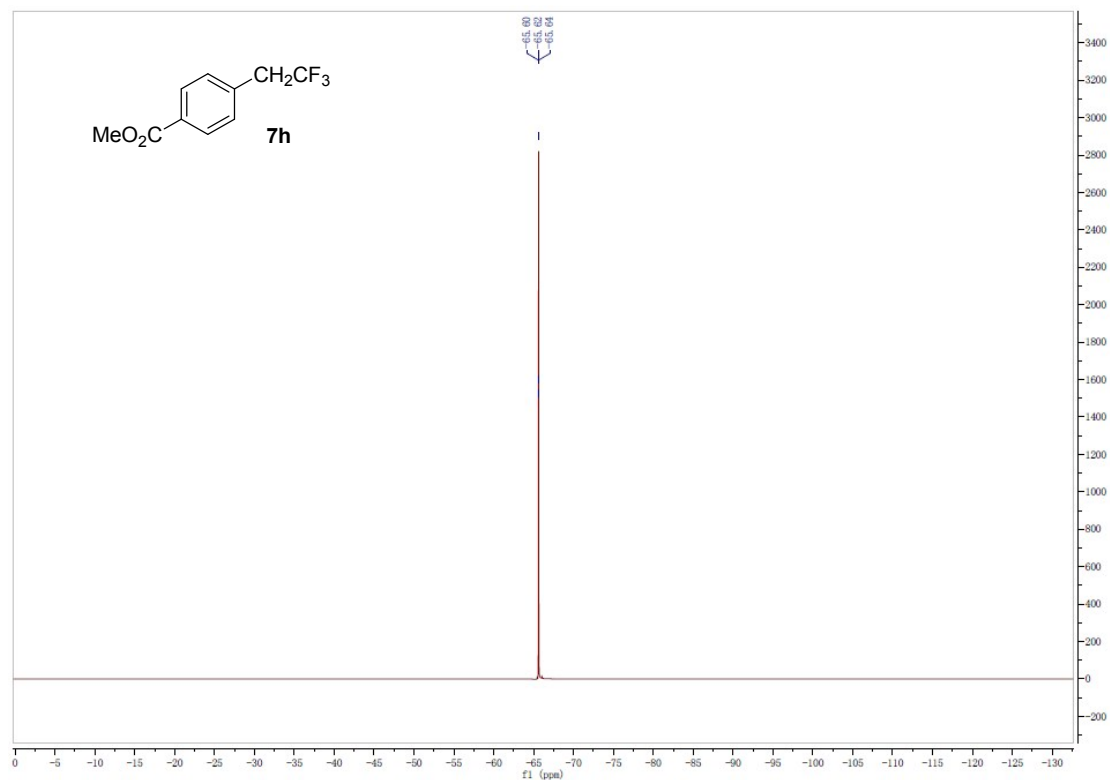


Figure S27. ¹⁹F NMR of **7h** in CDCl₃

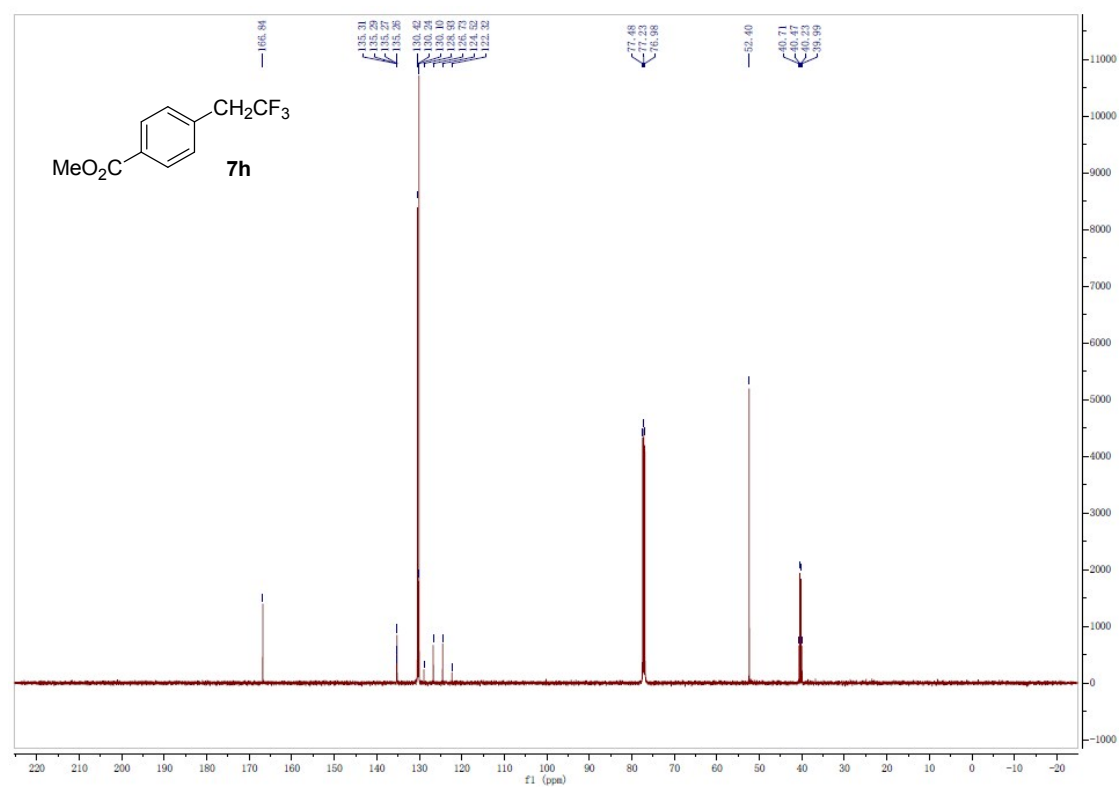
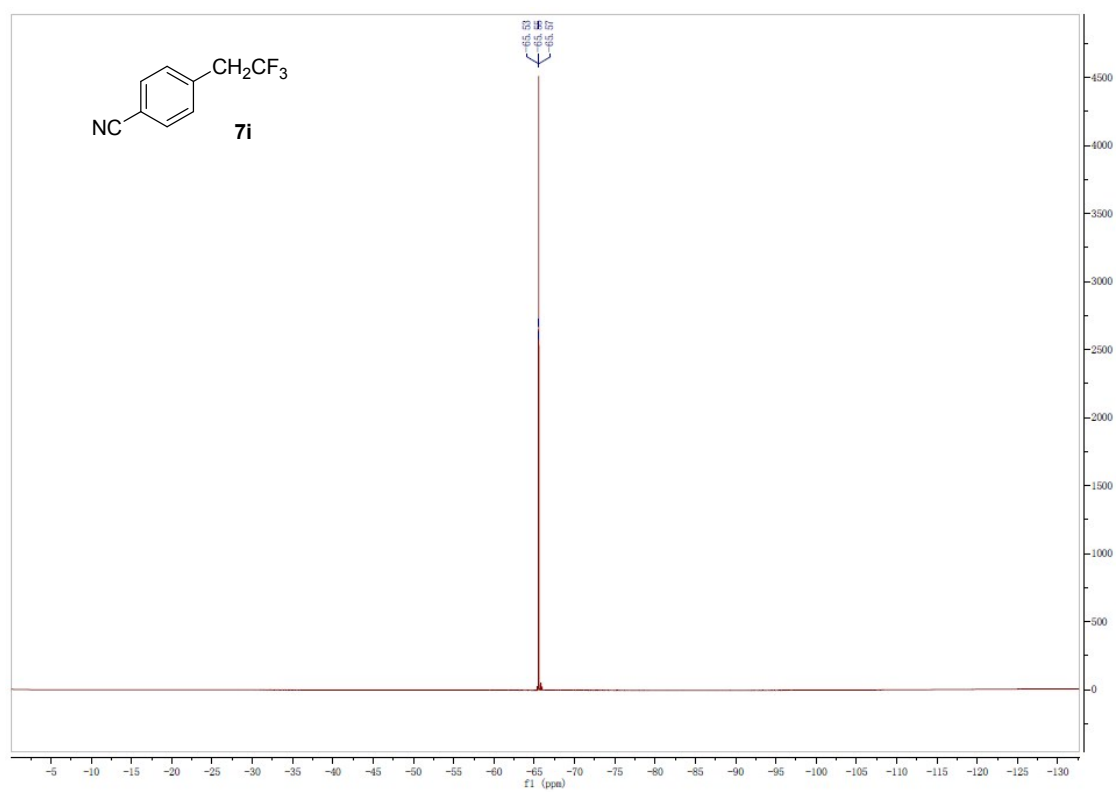
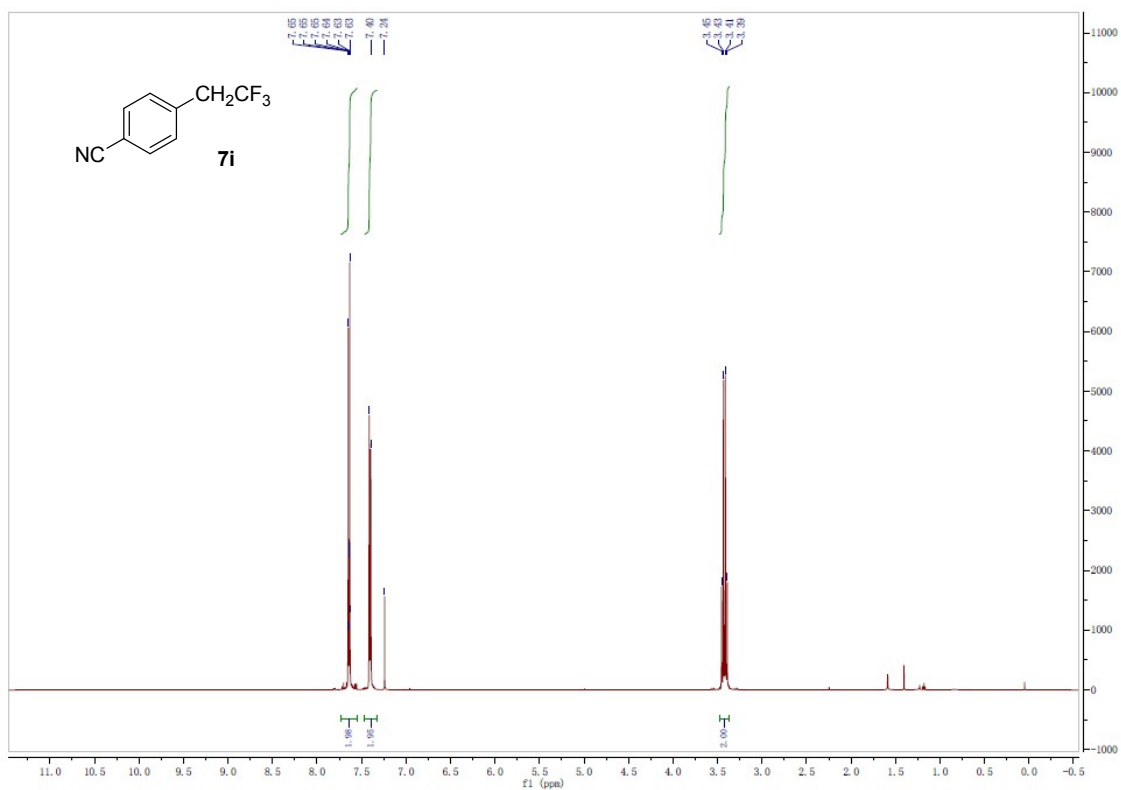


Figure S28. ¹³C NMR of **7h** in CDCl₃



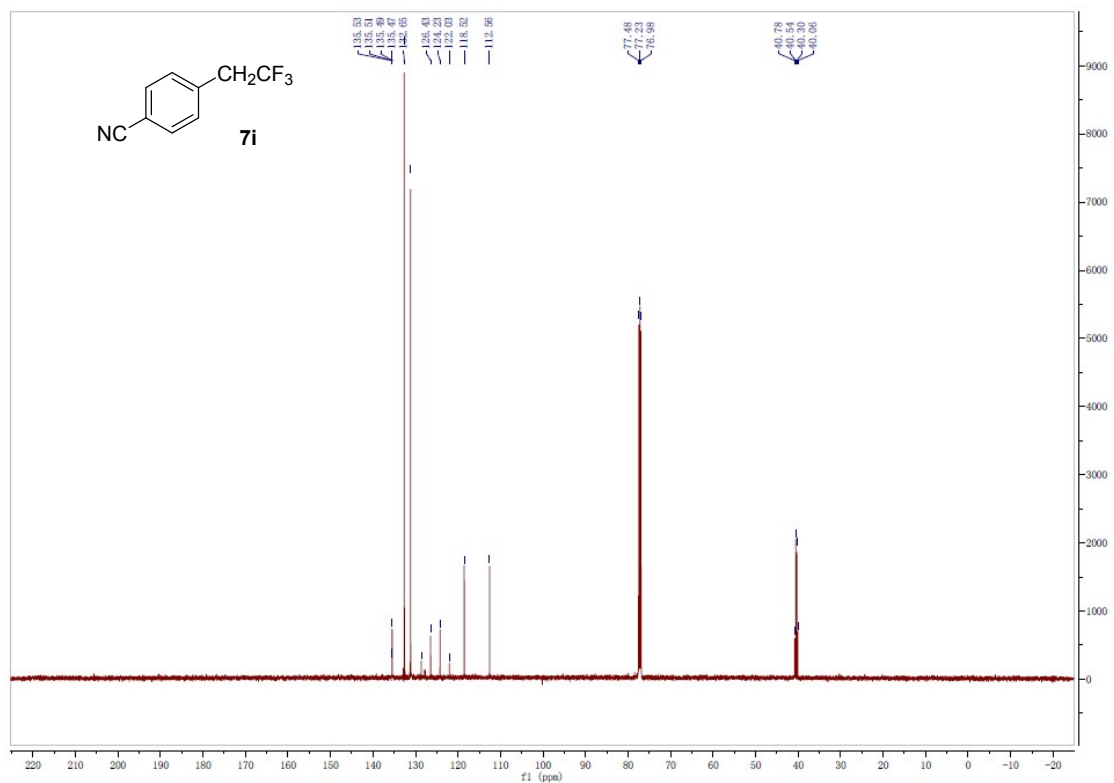


Figure S31. ¹³C NMR of **7i** in CDCl₃

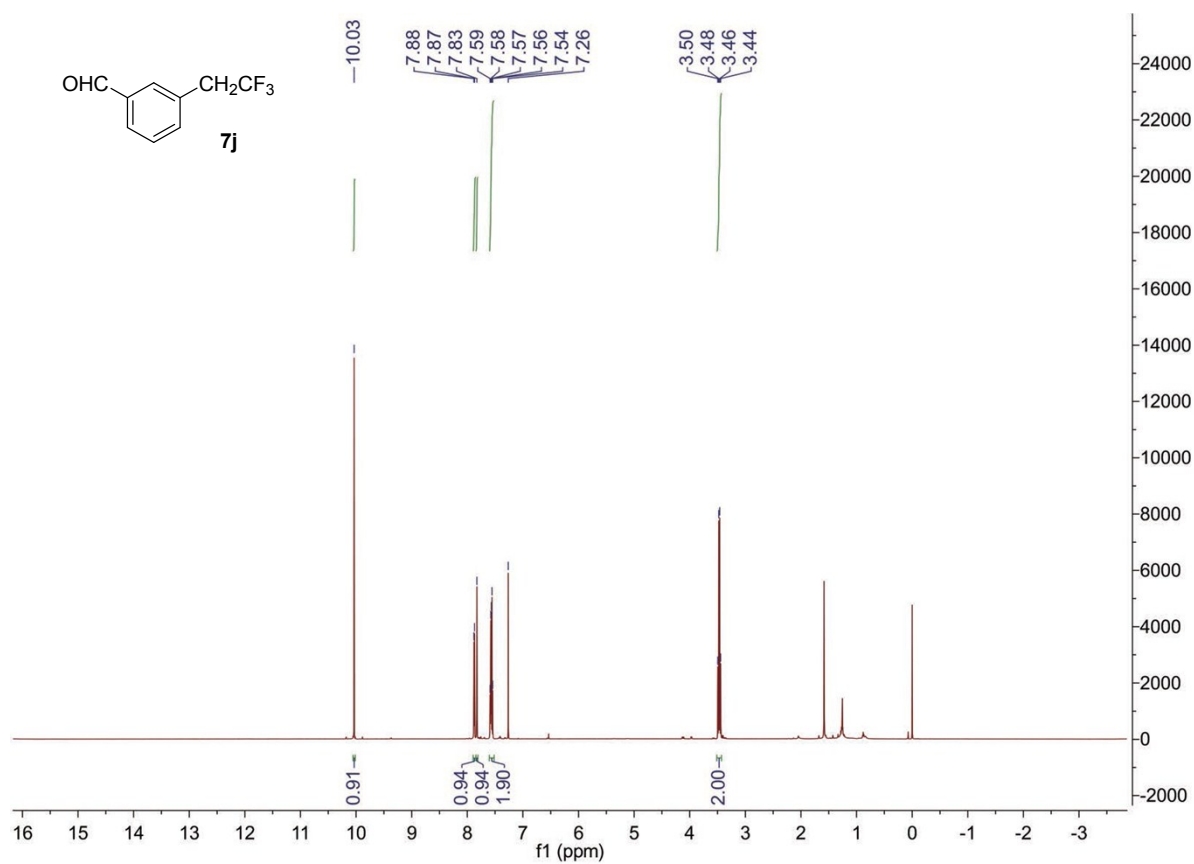


Figure S32. ¹H NMR of **7j** in CDCl₃

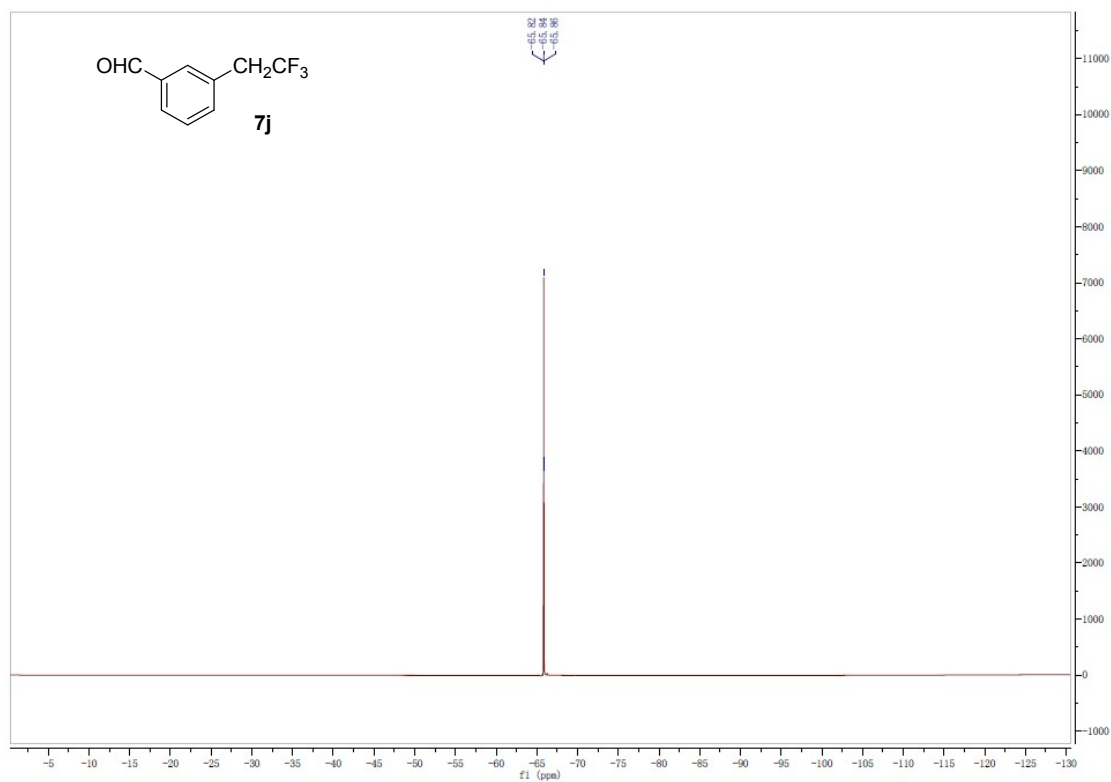


Figure S33. ¹⁹F NMR of **7j** in CDCl₃

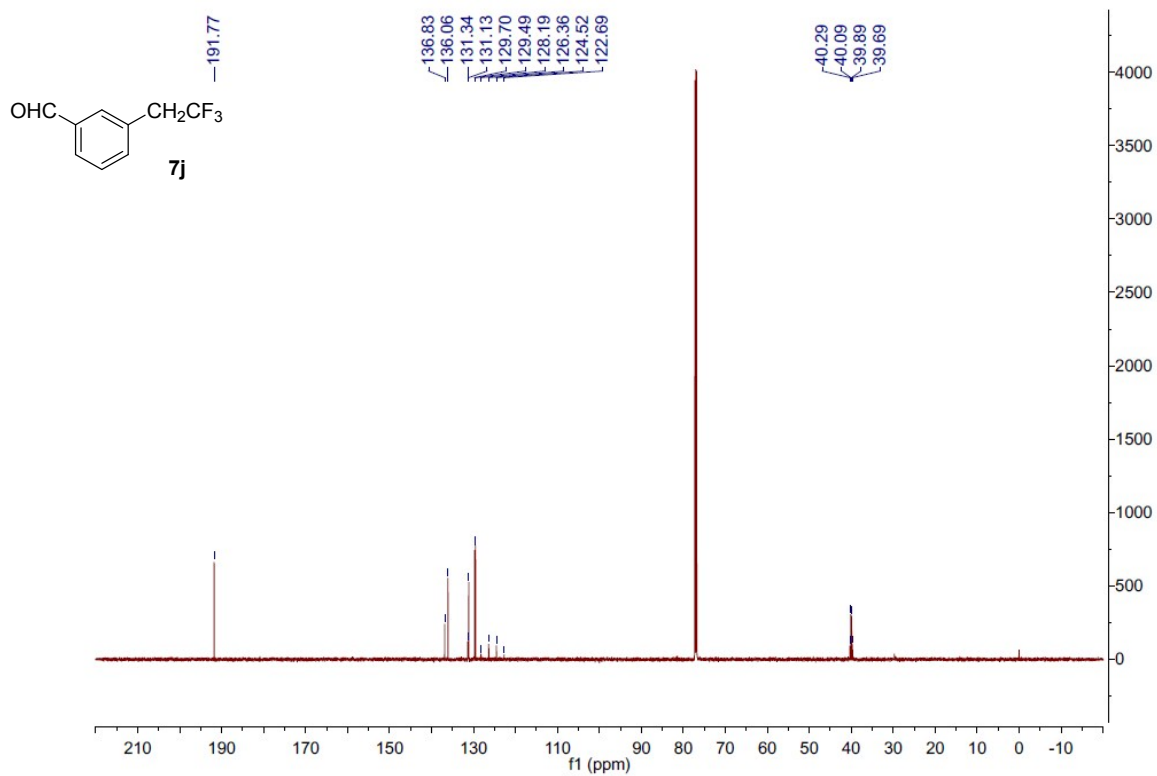


Figure S34. ¹³C NMR of **7j** in CDCl₃

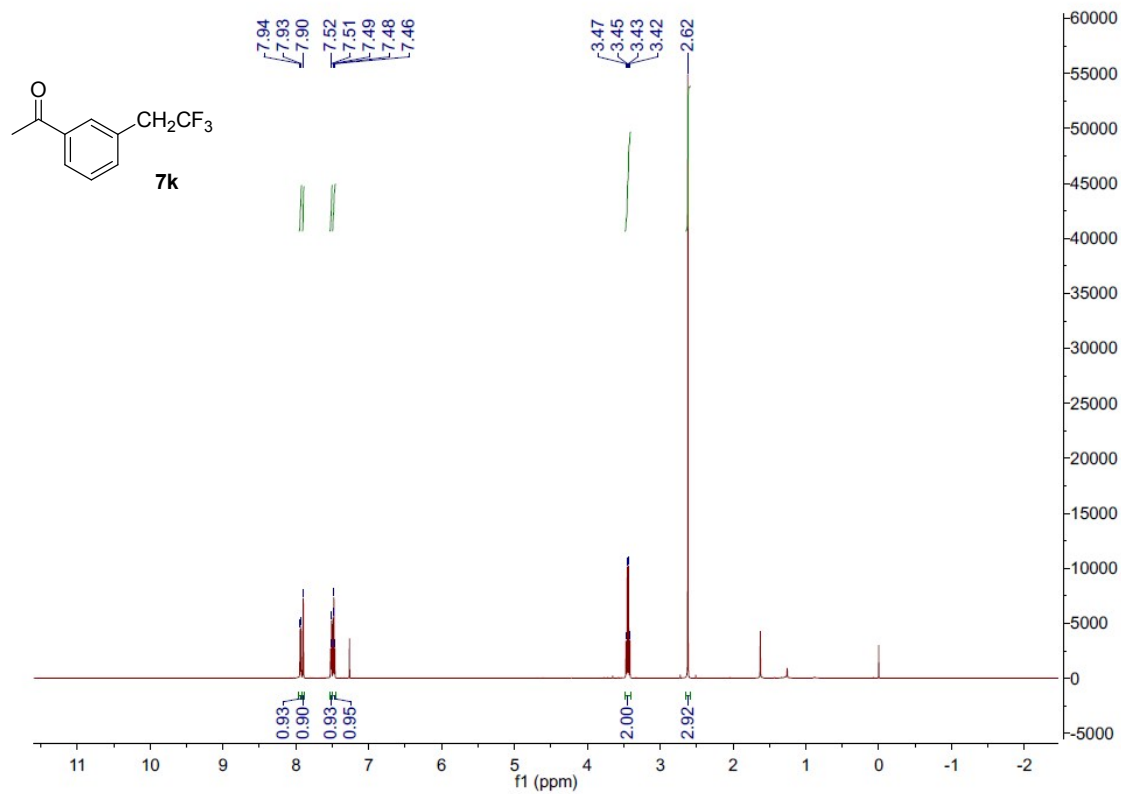


Figure S35. ¹H NMR of **7k** in CDCl₃

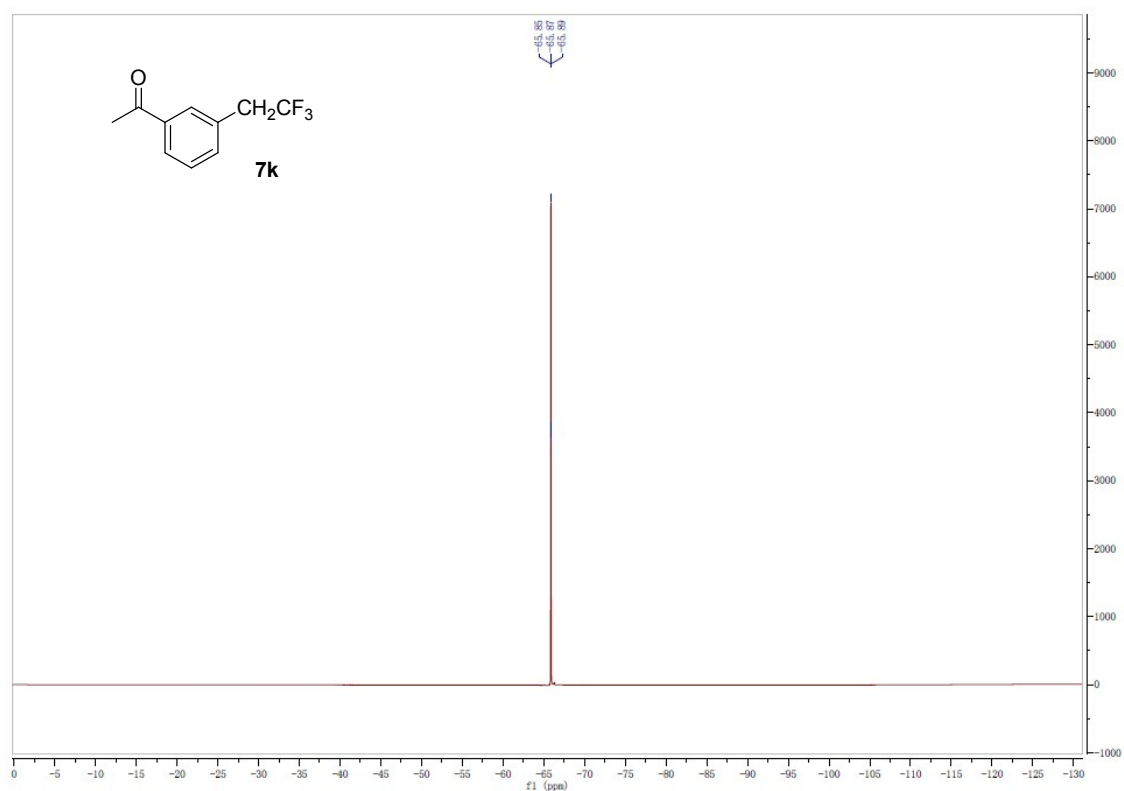


Figure S36. ¹⁹F NMR of **7k** in CDCl₃

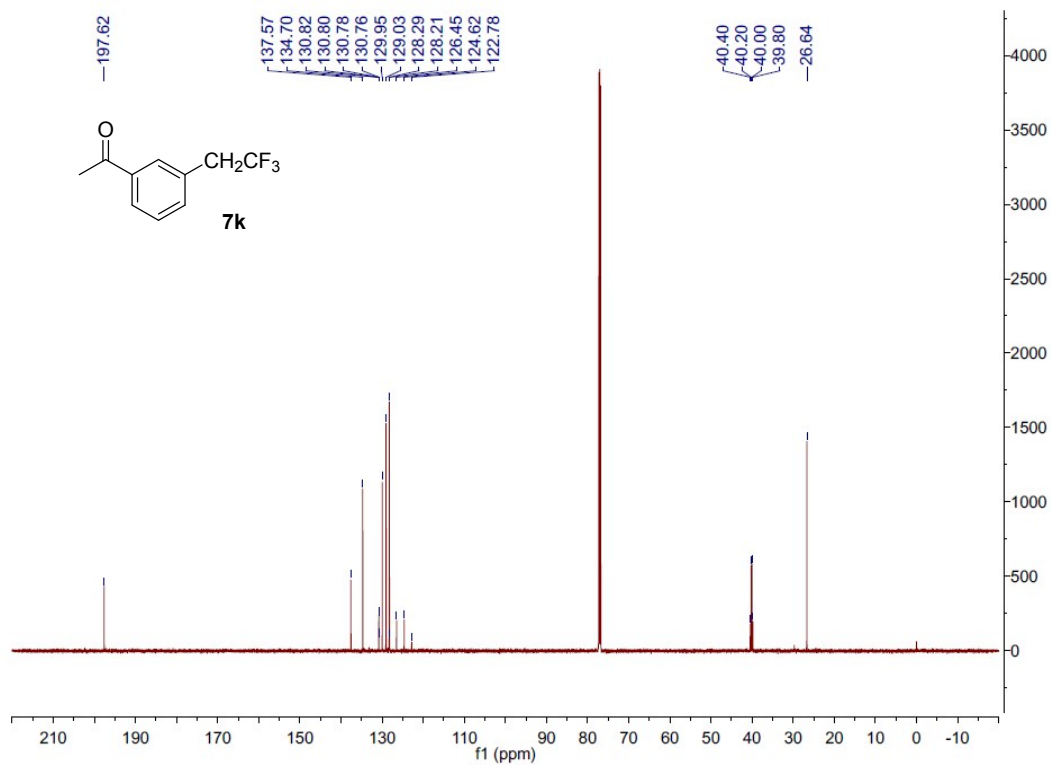


Figure S37. ¹³C NMR of **7k** in CDCl₃

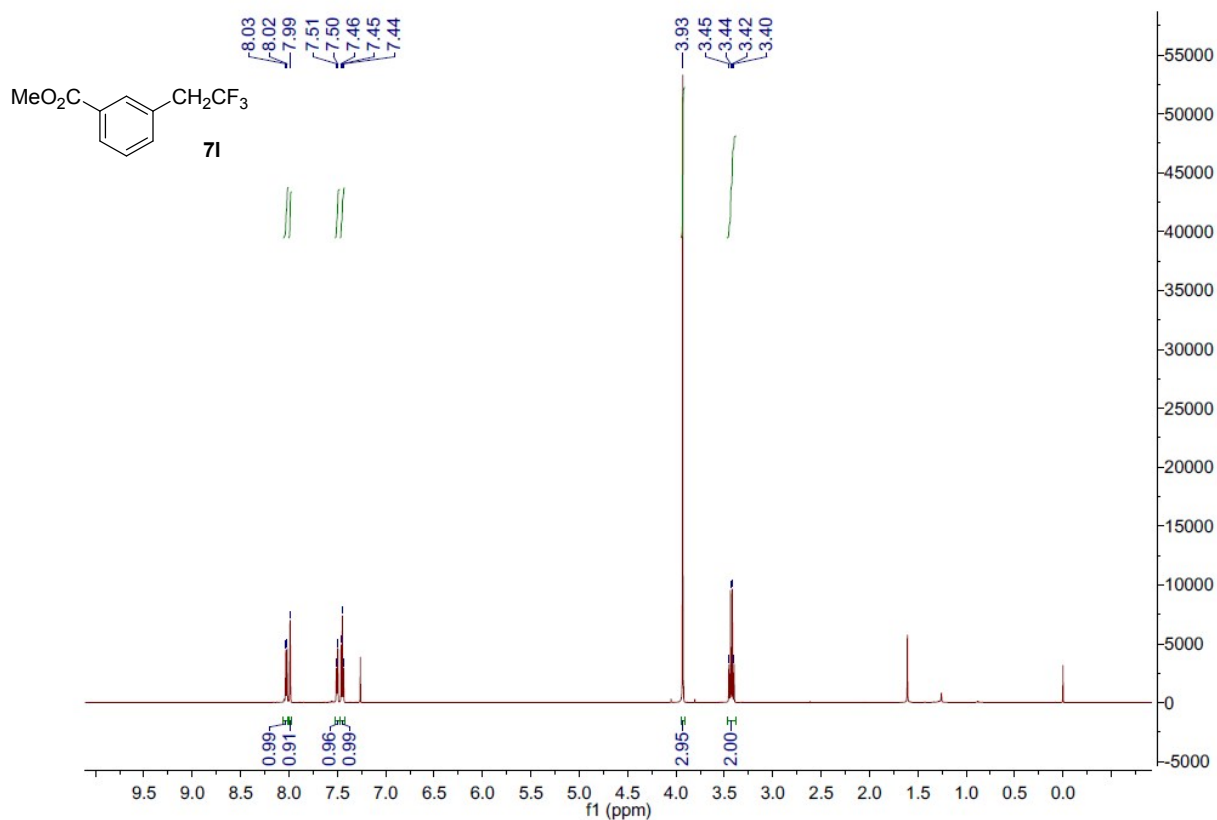


Figure S38. ¹H NMR of **7l** in CDCl₃

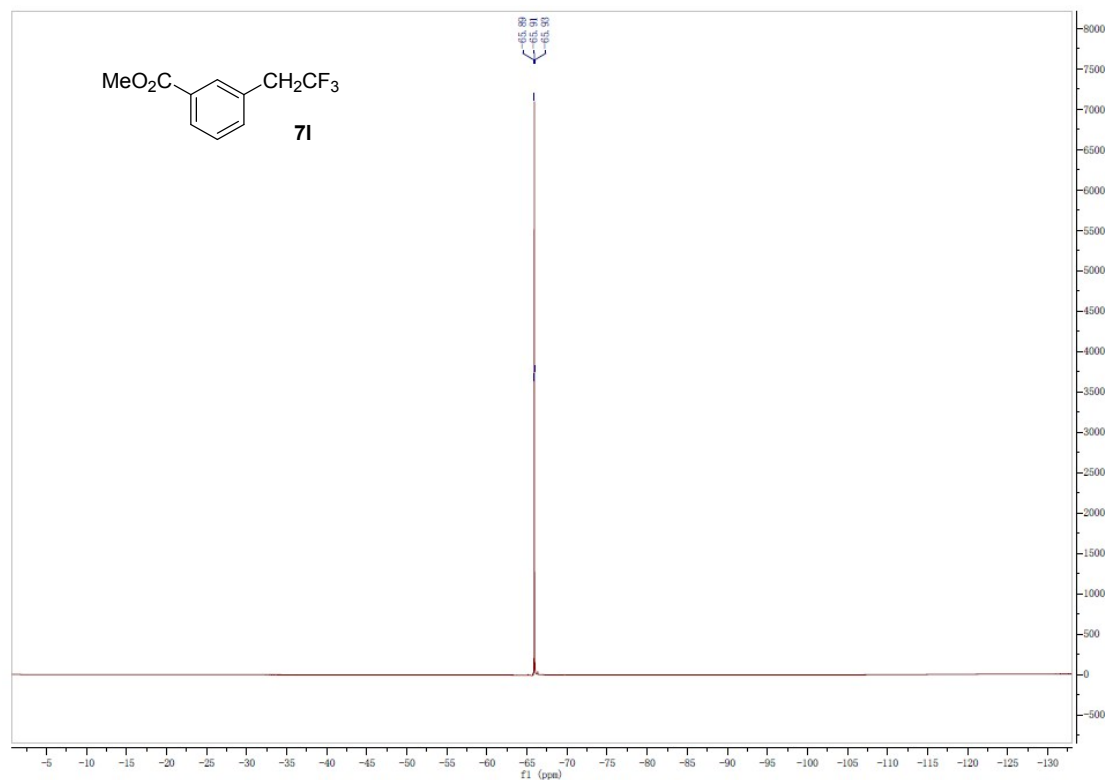


Figure S39. ¹⁹F NMR of **7l** in CDCl₃

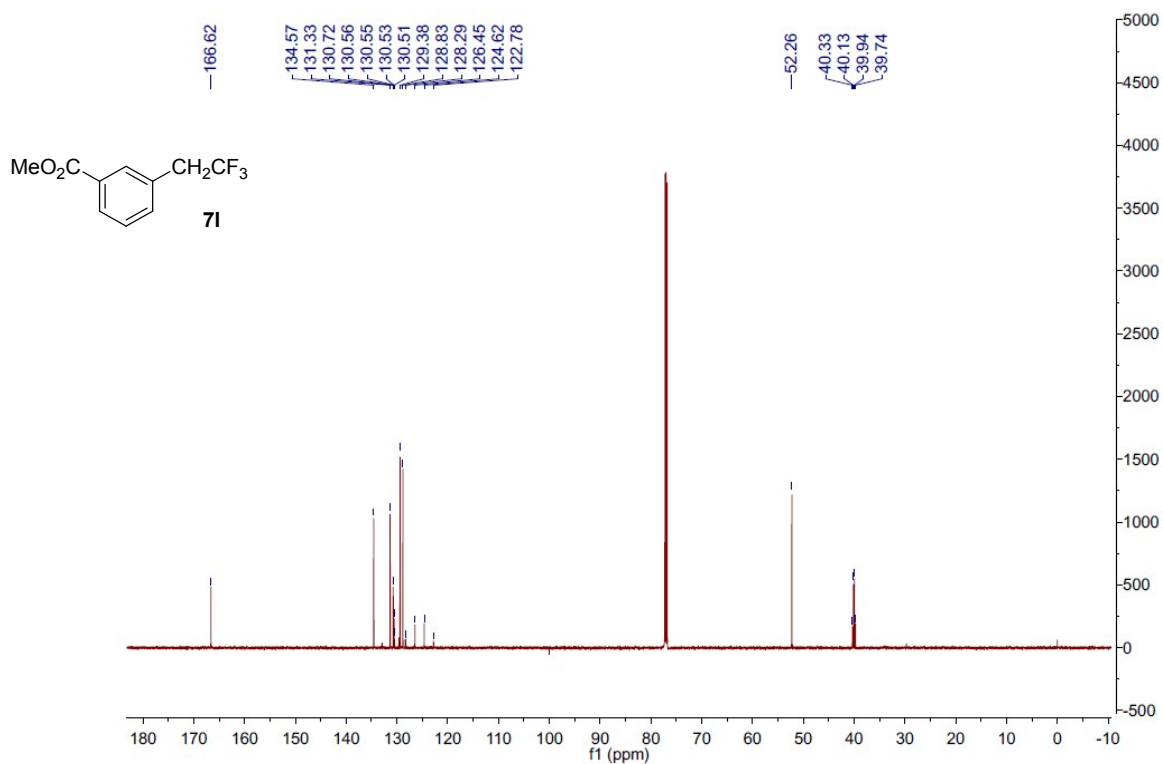


Figure S40. ¹³C NMR of **7l** in CDCl₃

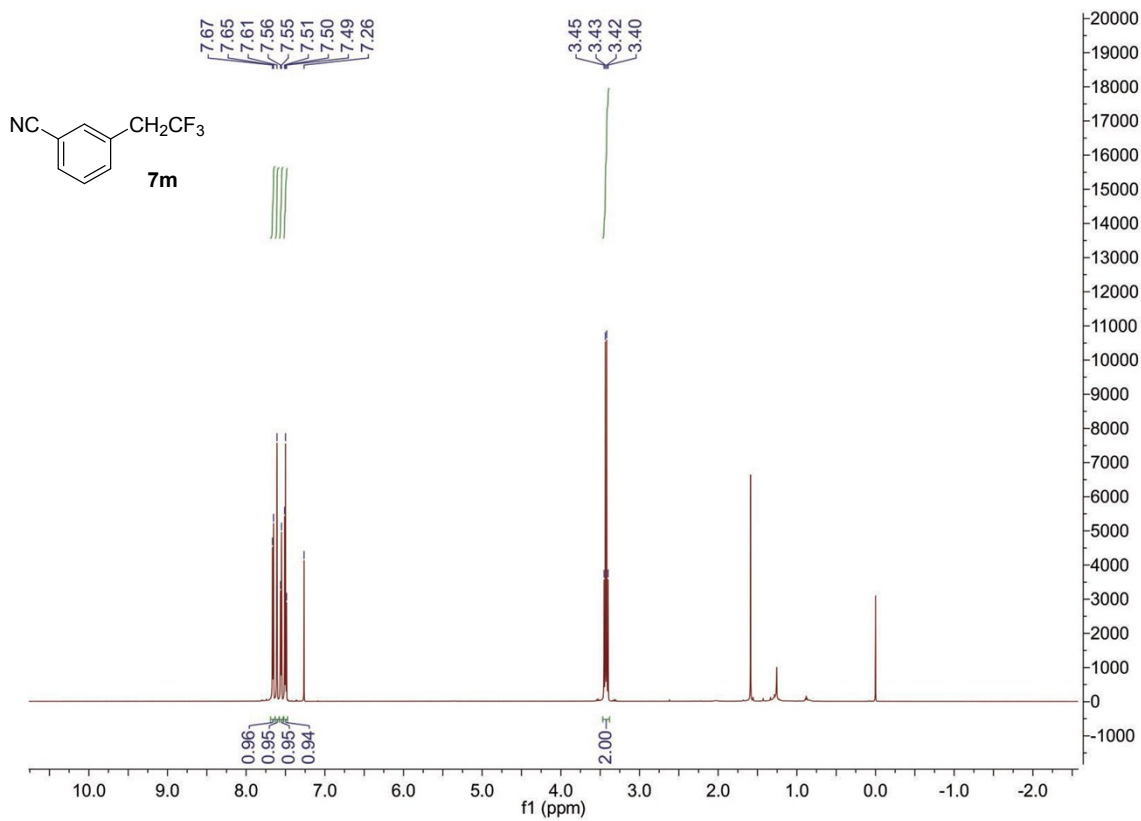


Figure S41. ¹H NMR of **7m** in CDCl₃

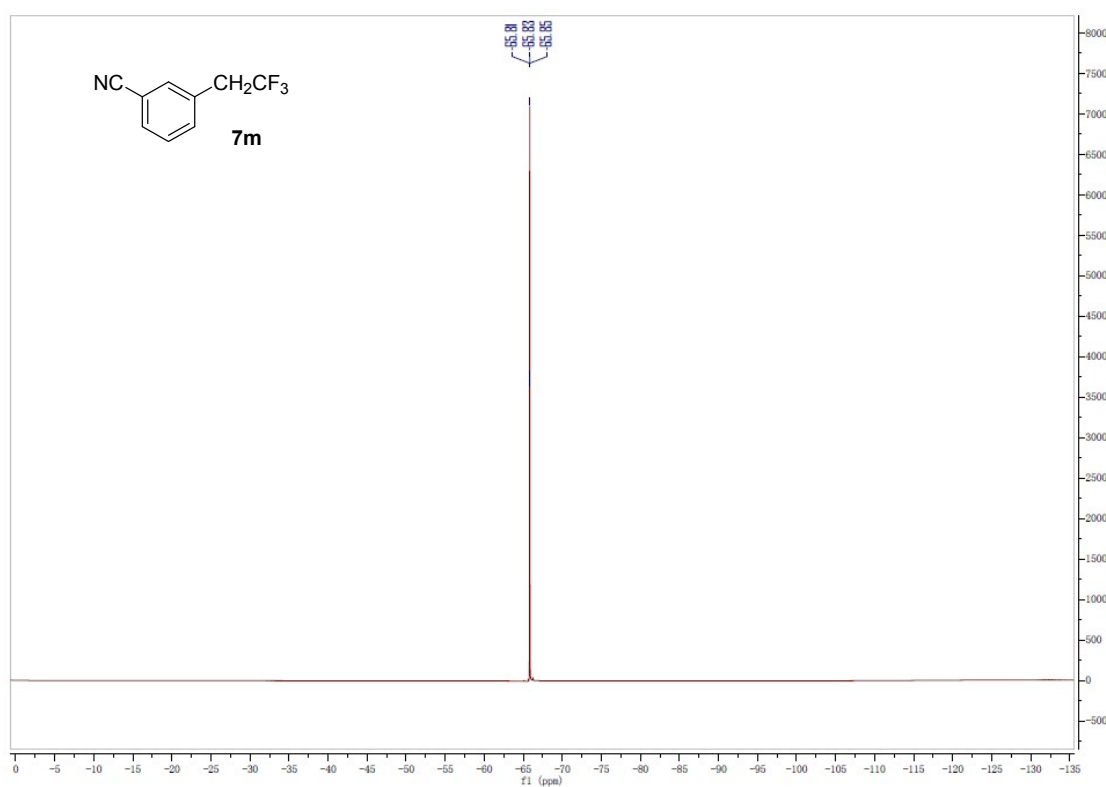


Figure S42. ¹⁹F NMR of **7m** in CDCl₃

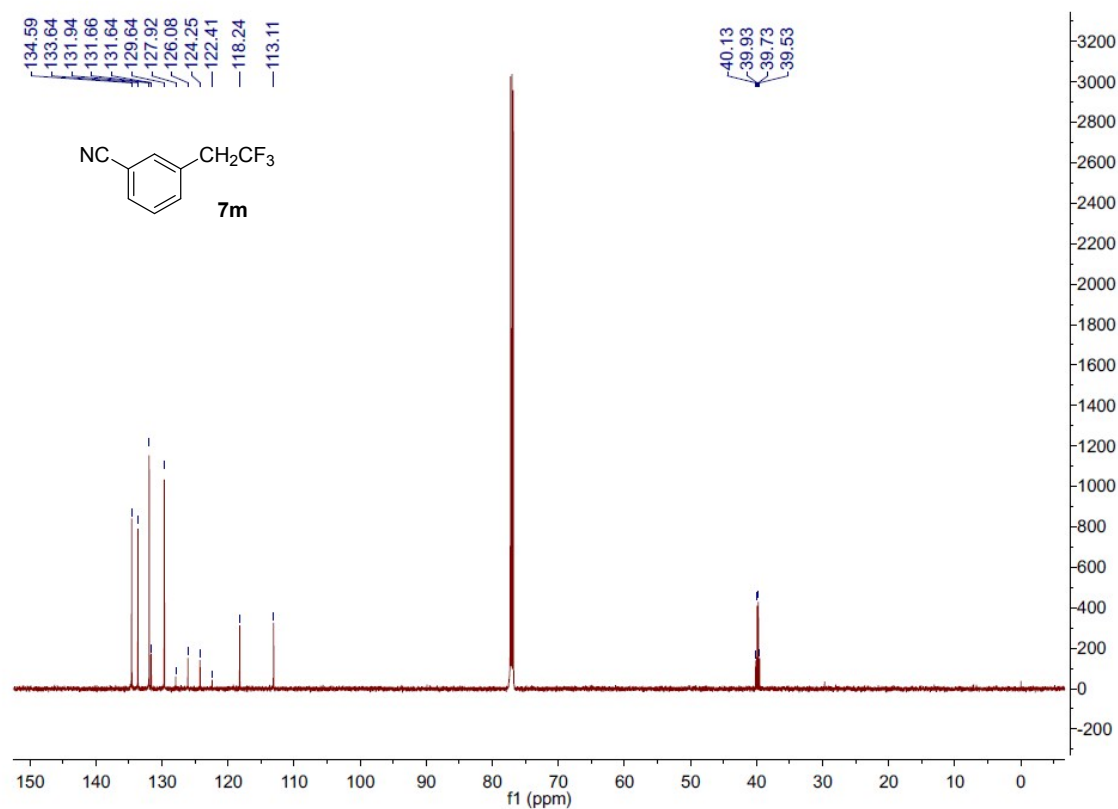


Figure S43. ¹³C NMR of **7m** in CDCl₃

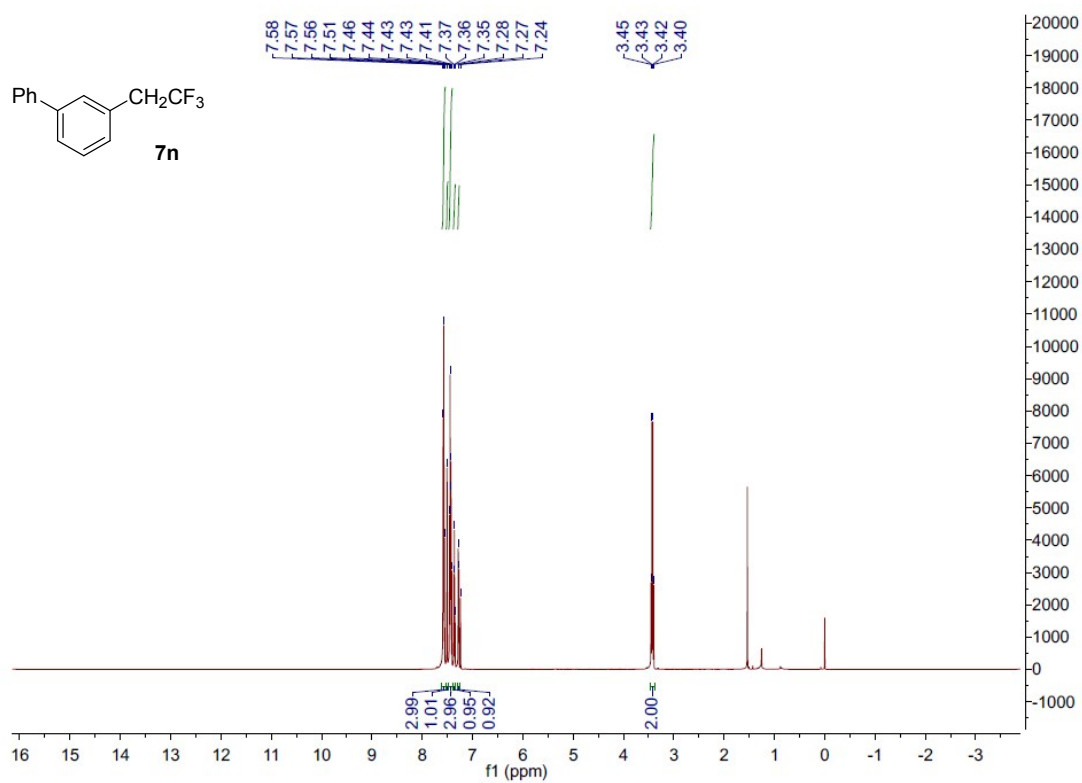


Figure S44. ¹H NMR of **7n** in CDCl₃

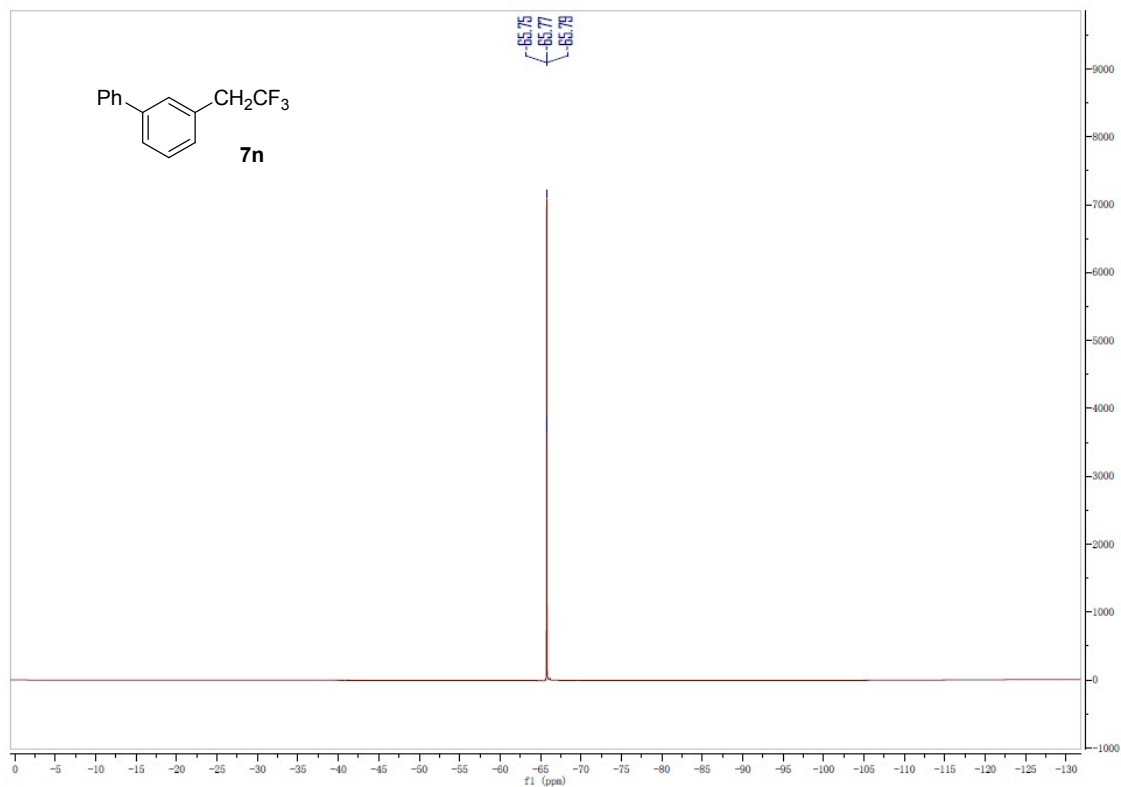


Figure S45. ¹⁹F NMR of **7n** in CDCl₃

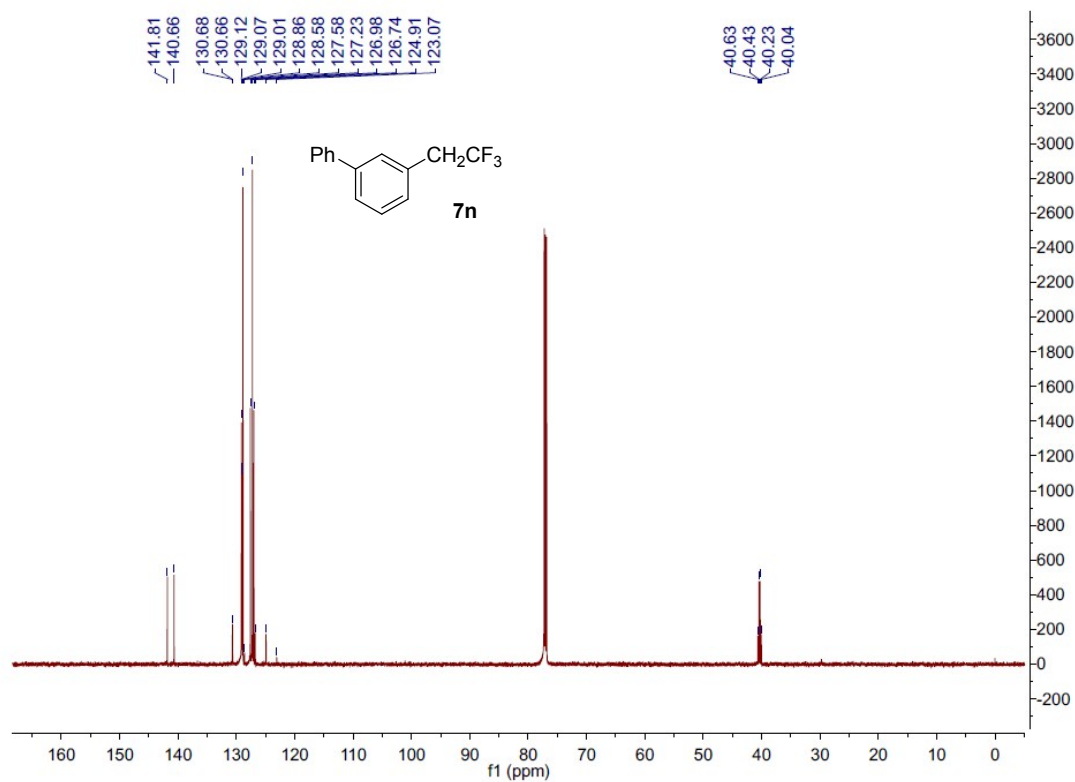


Figure S46. ¹³C NMR of **7n** in CDCl₃

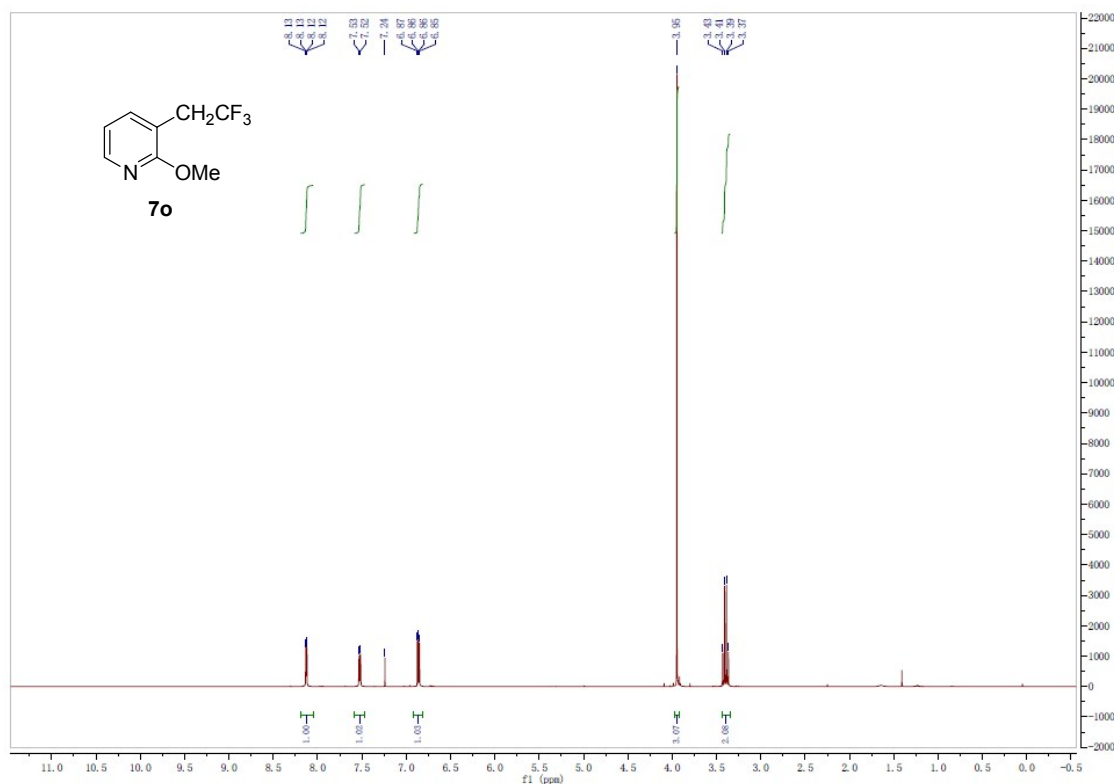


Figure S47. ¹H NMR of **7o** in CDCl₃

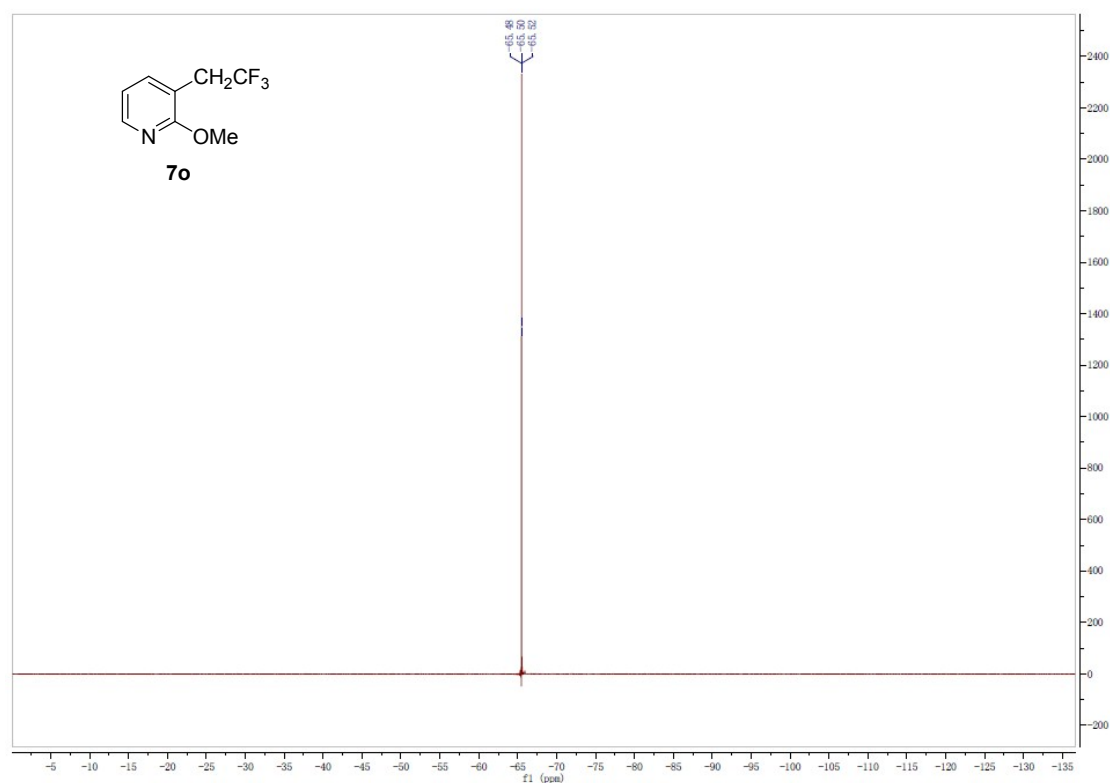
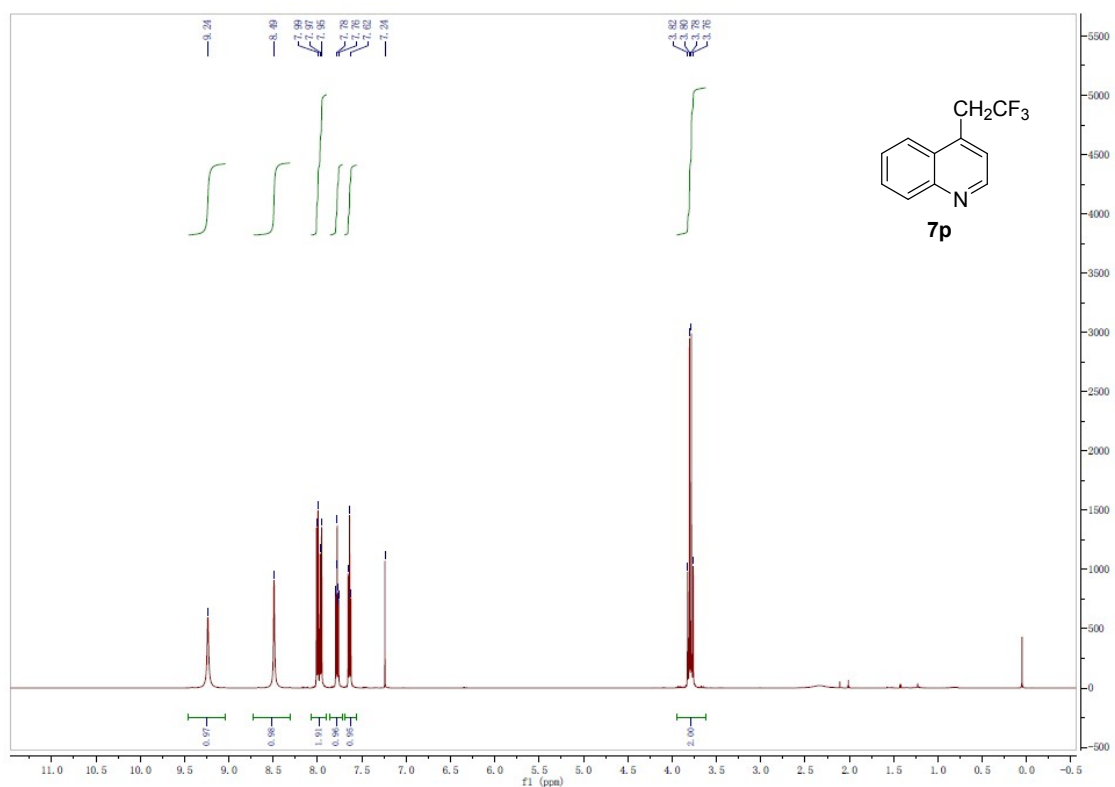
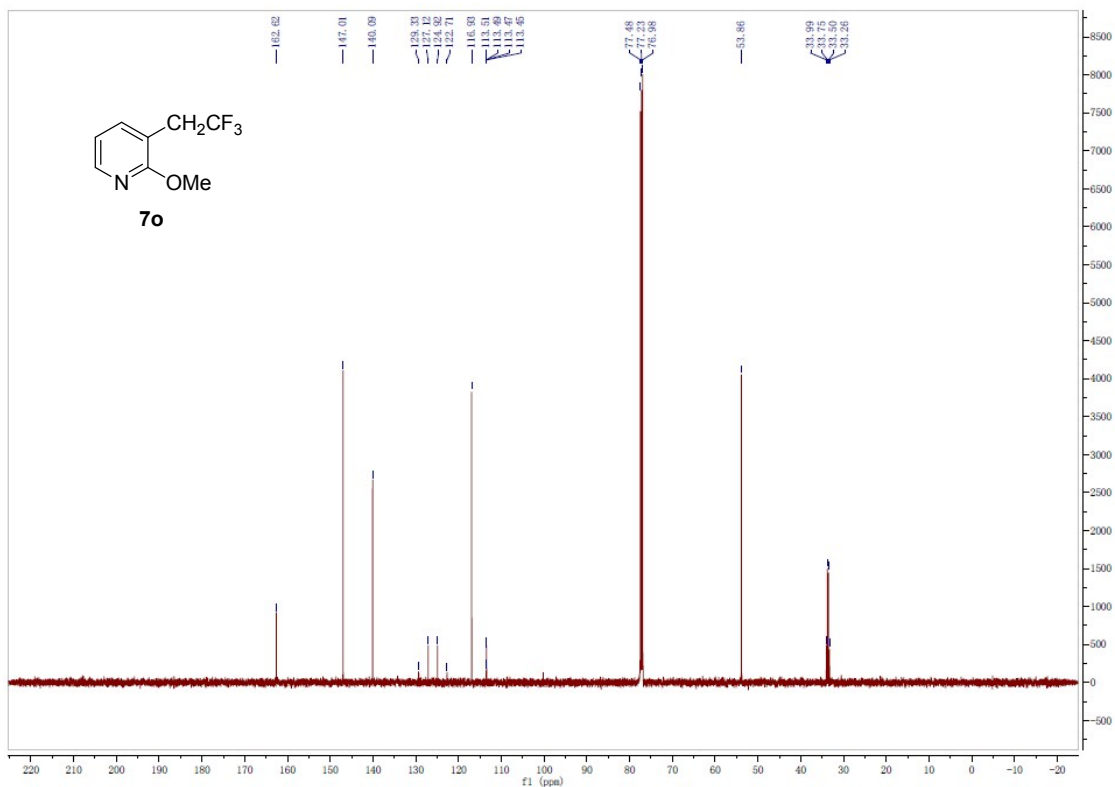


Figure S48. ¹⁹F NMR of **7o** in CDCl₃



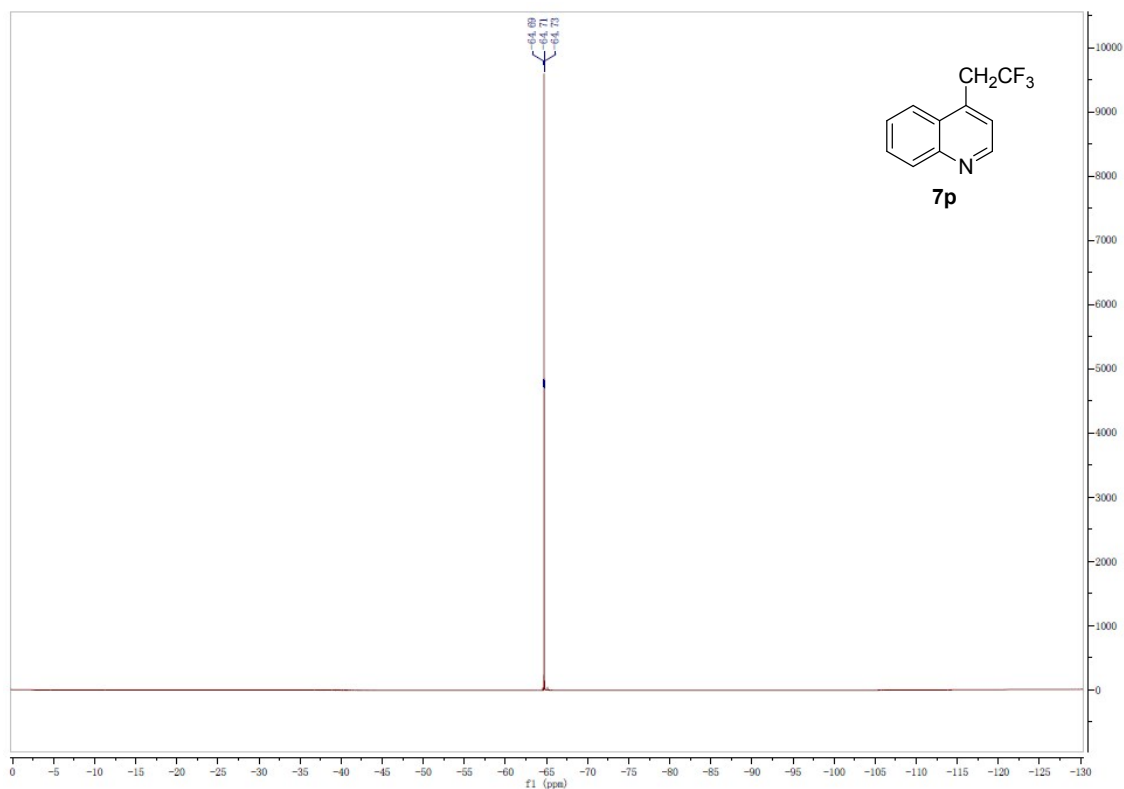


Figure S51. ¹⁹F NMR of 7p in CDCl₃

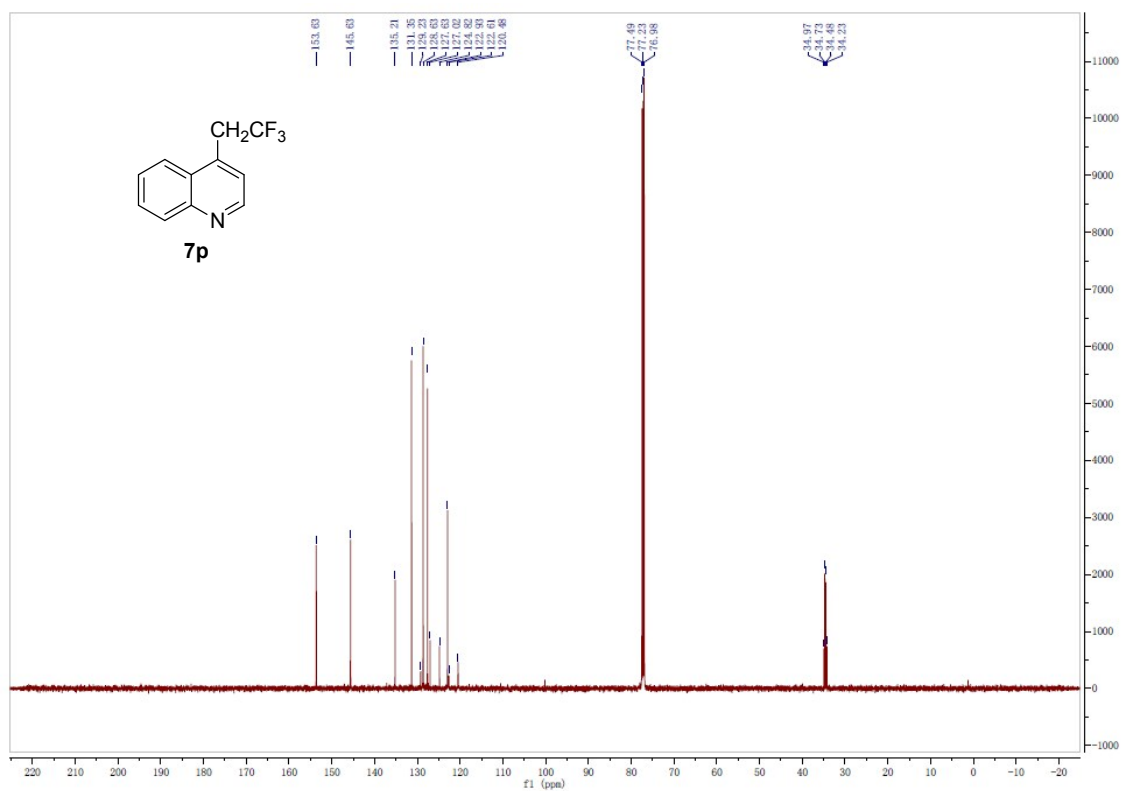


Figure S52. ¹³C NMR of 7p in CDCl₃

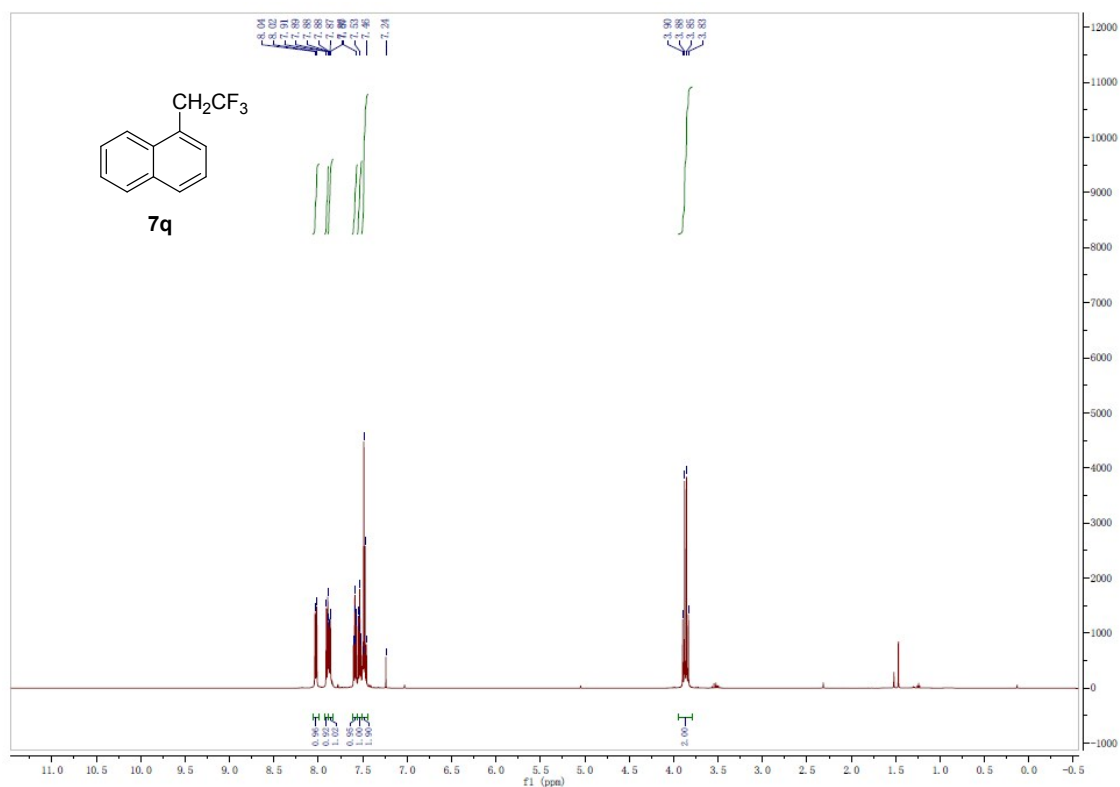
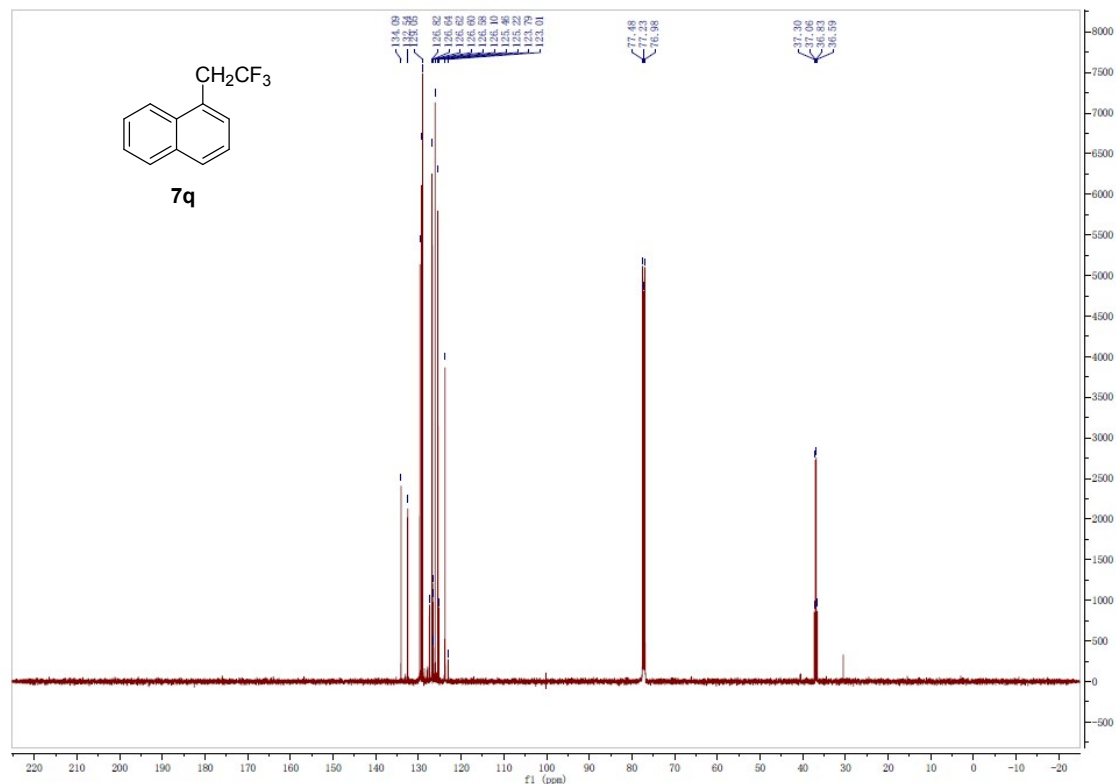


Figure S53. ¹H NMR of 7q in CDCl₃



Figure S54. ¹⁹F NMR of 7q in CDCl₃



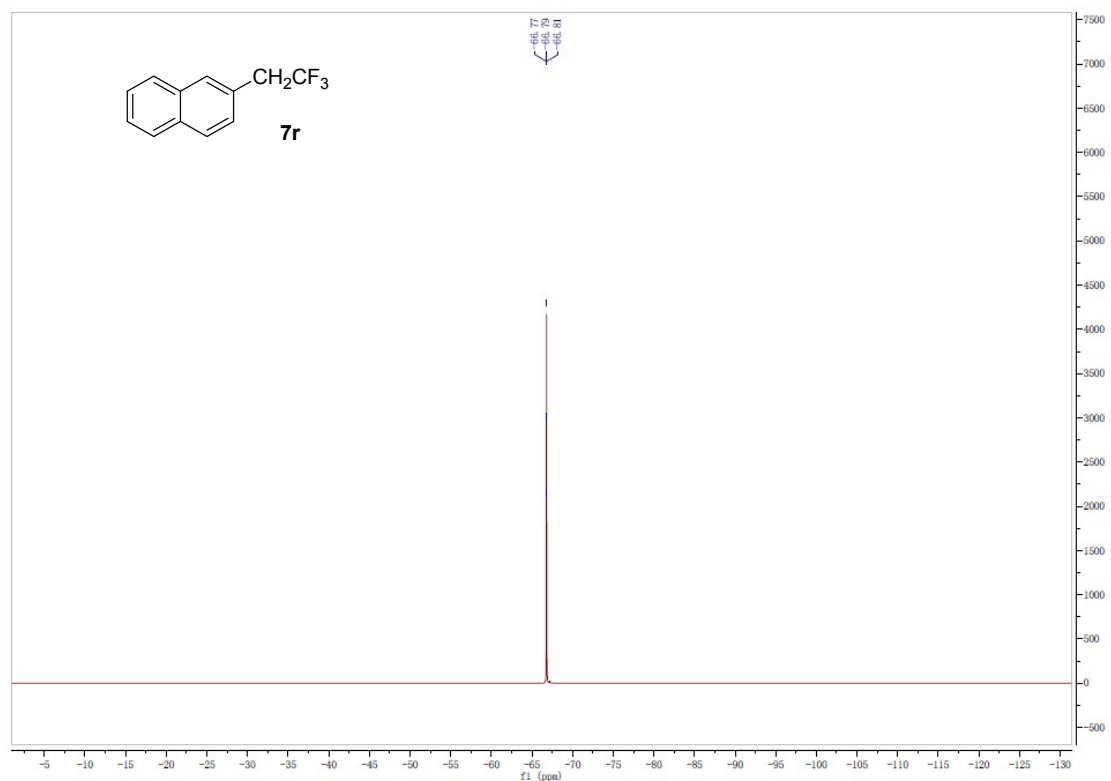


Figure S57. ¹⁹F NMR of **7r** in CDCl₃

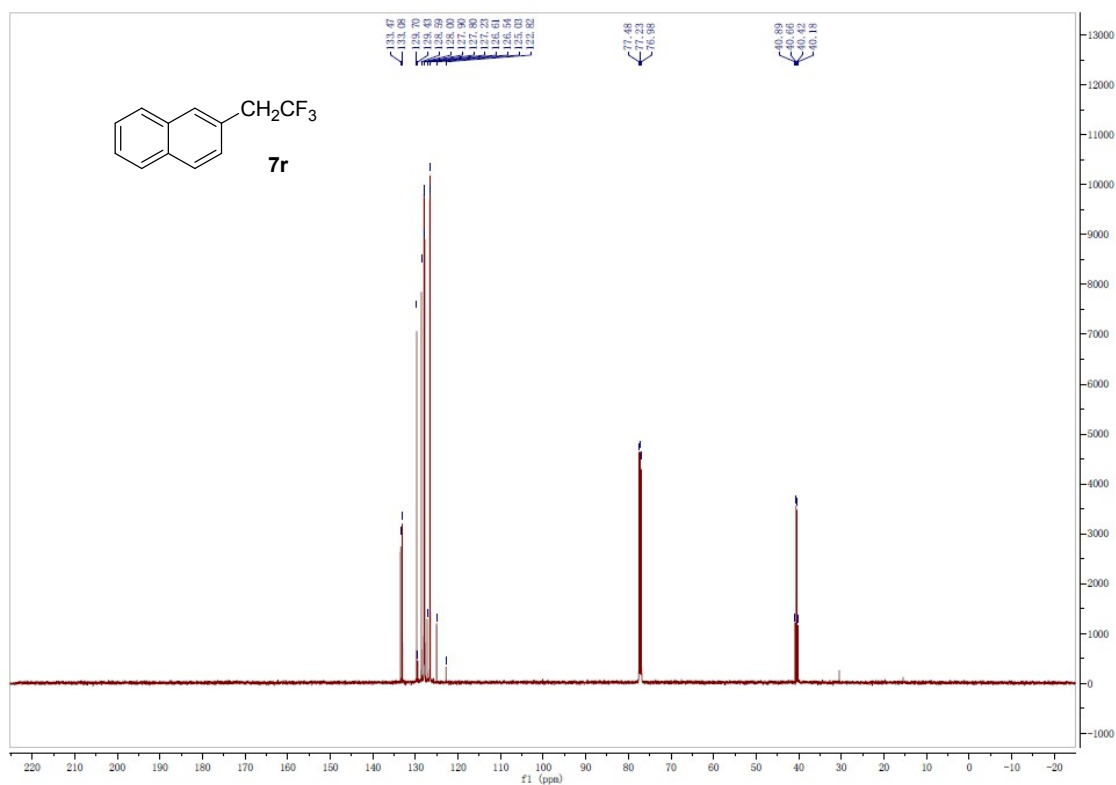


Figure S58. ¹³C NMR of **7r** in CDCl₃

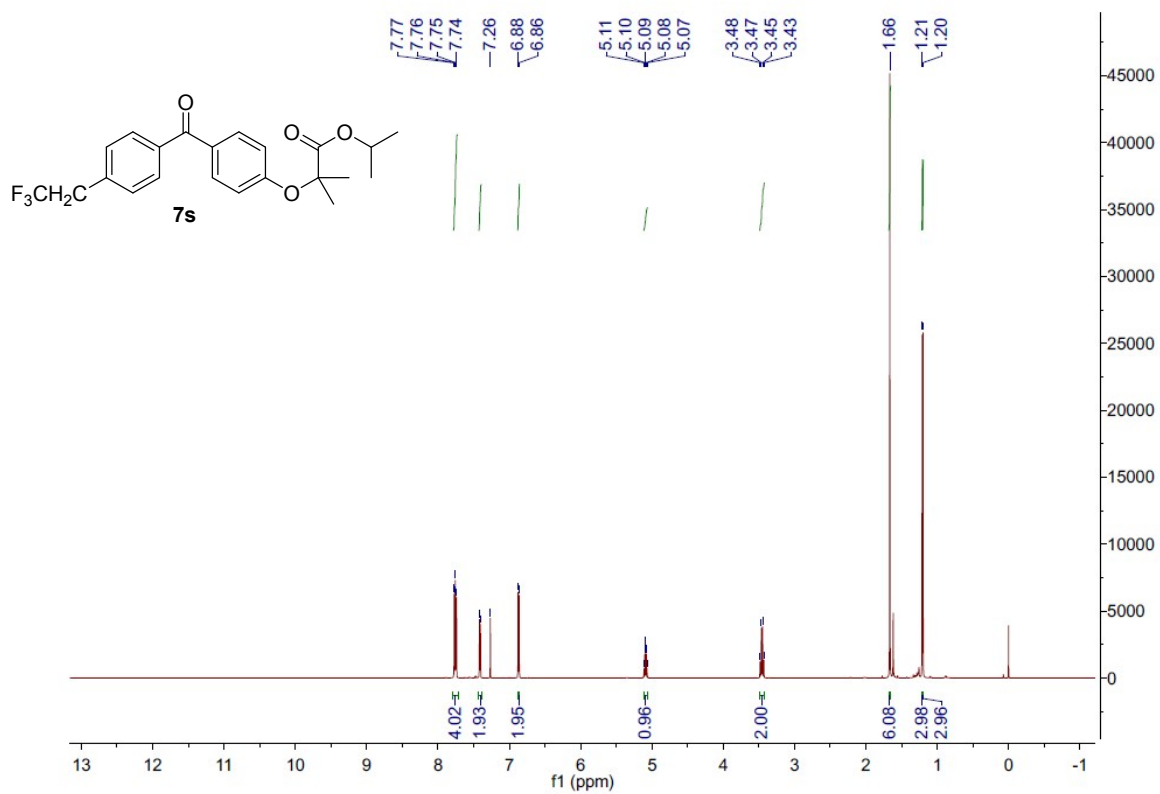


Figure S59. ¹H NMR of **7s** in CDCl₃

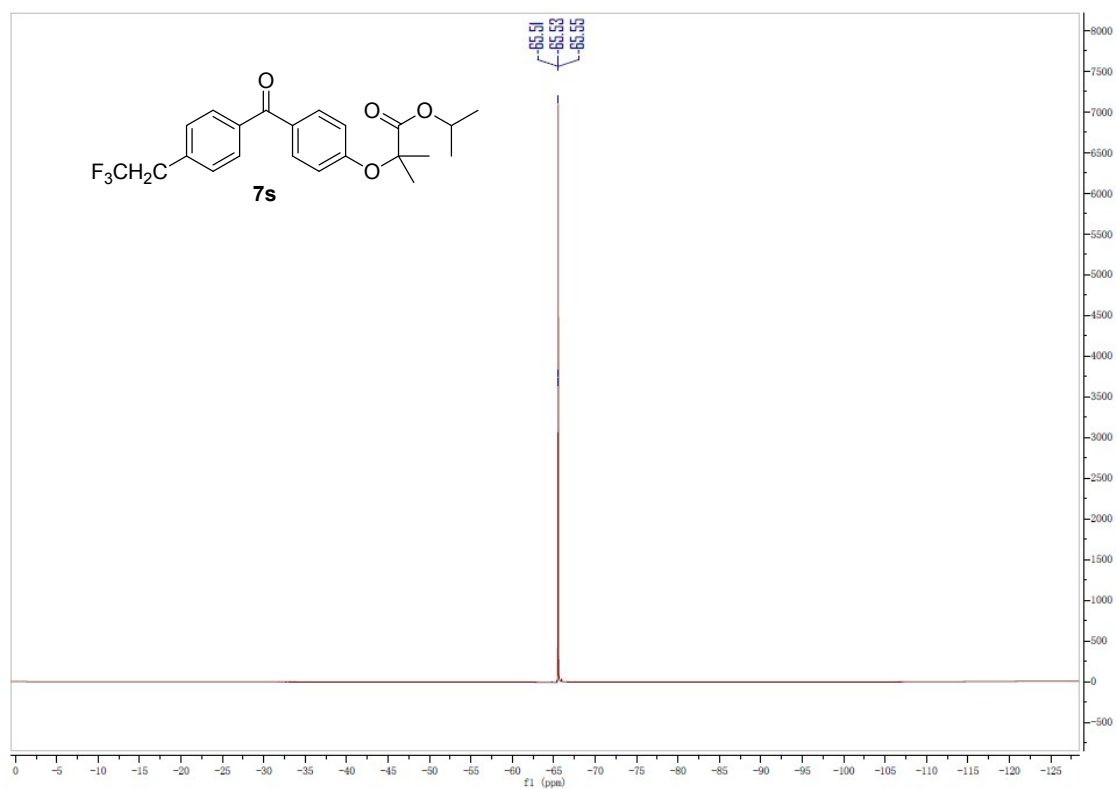


Figure S60. ¹⁹F NMR of **7s** in CDCl₃

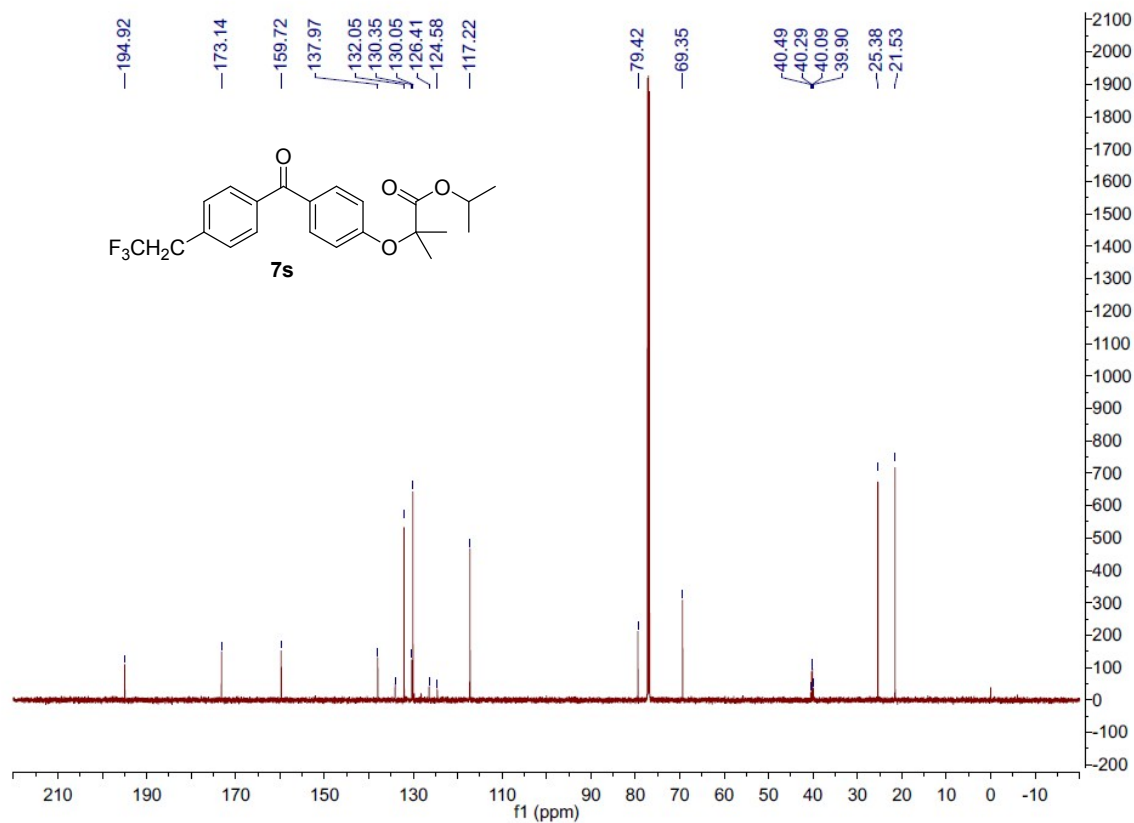


Figure S61. ¹³C NMR of **7s** in CDCl₃

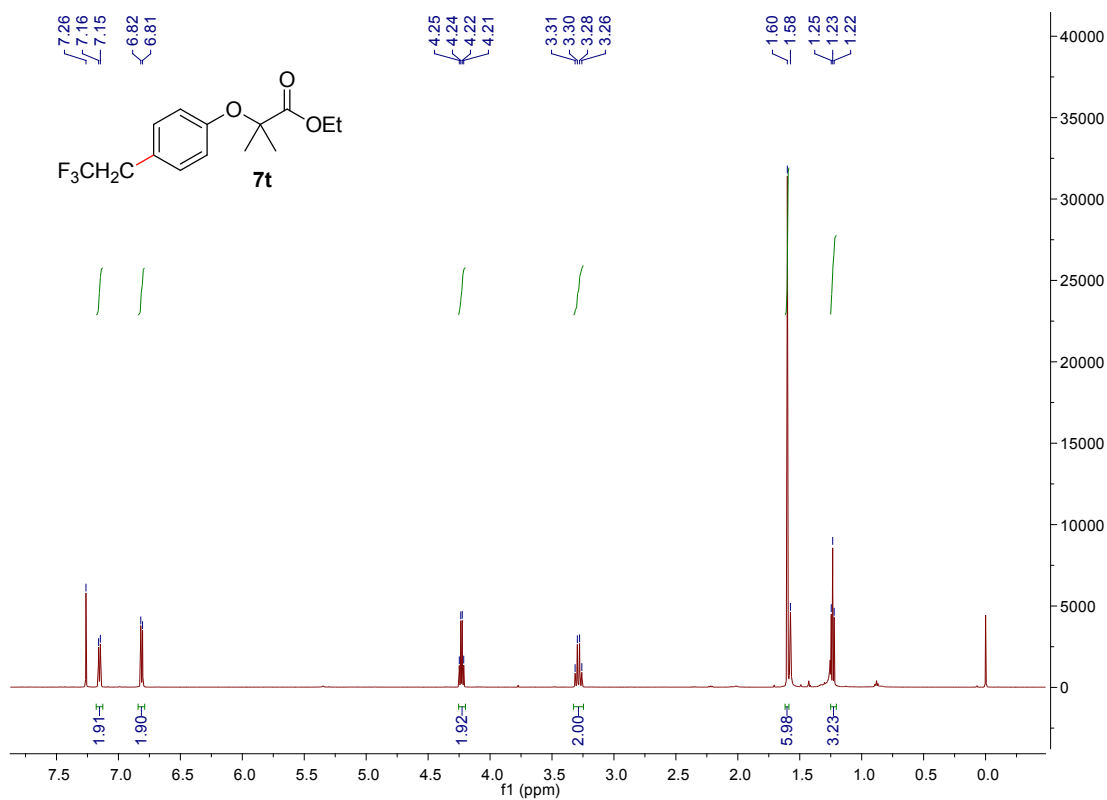


Figure S62. ¹H NMR of **7t** in CDCl₃

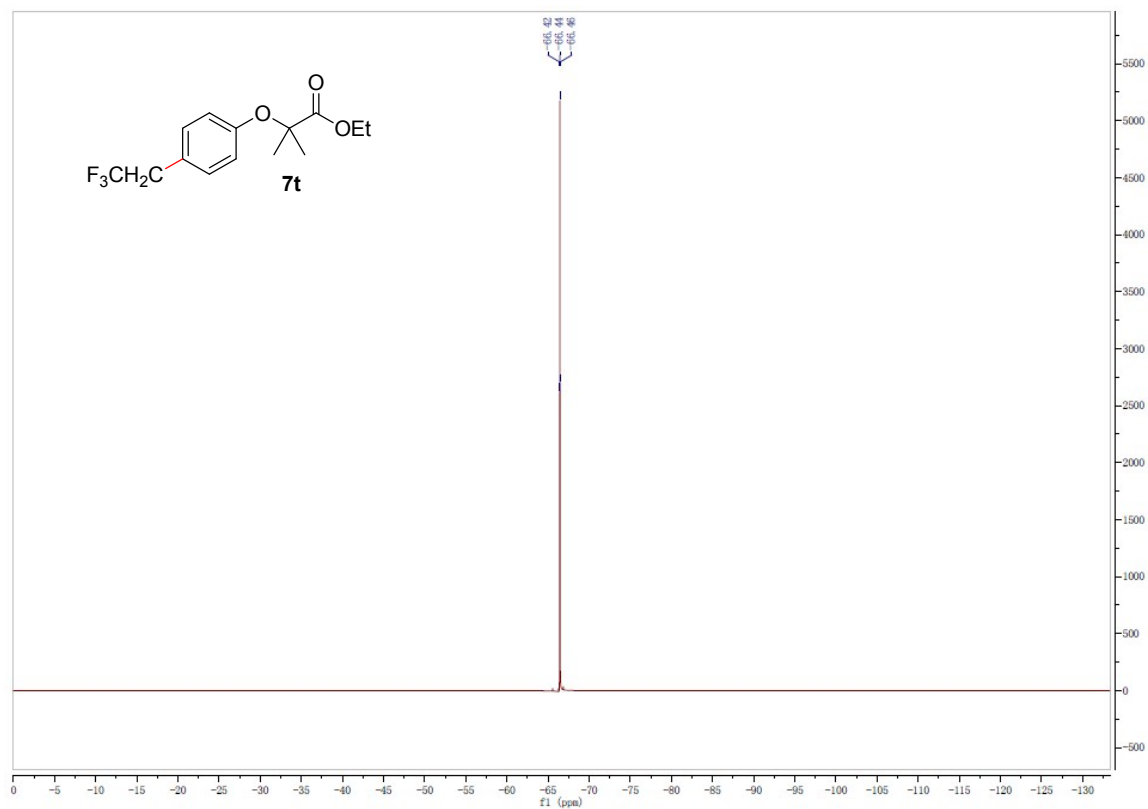


Figure S63. ¹⁹F NMR of 7t in CDCl₃

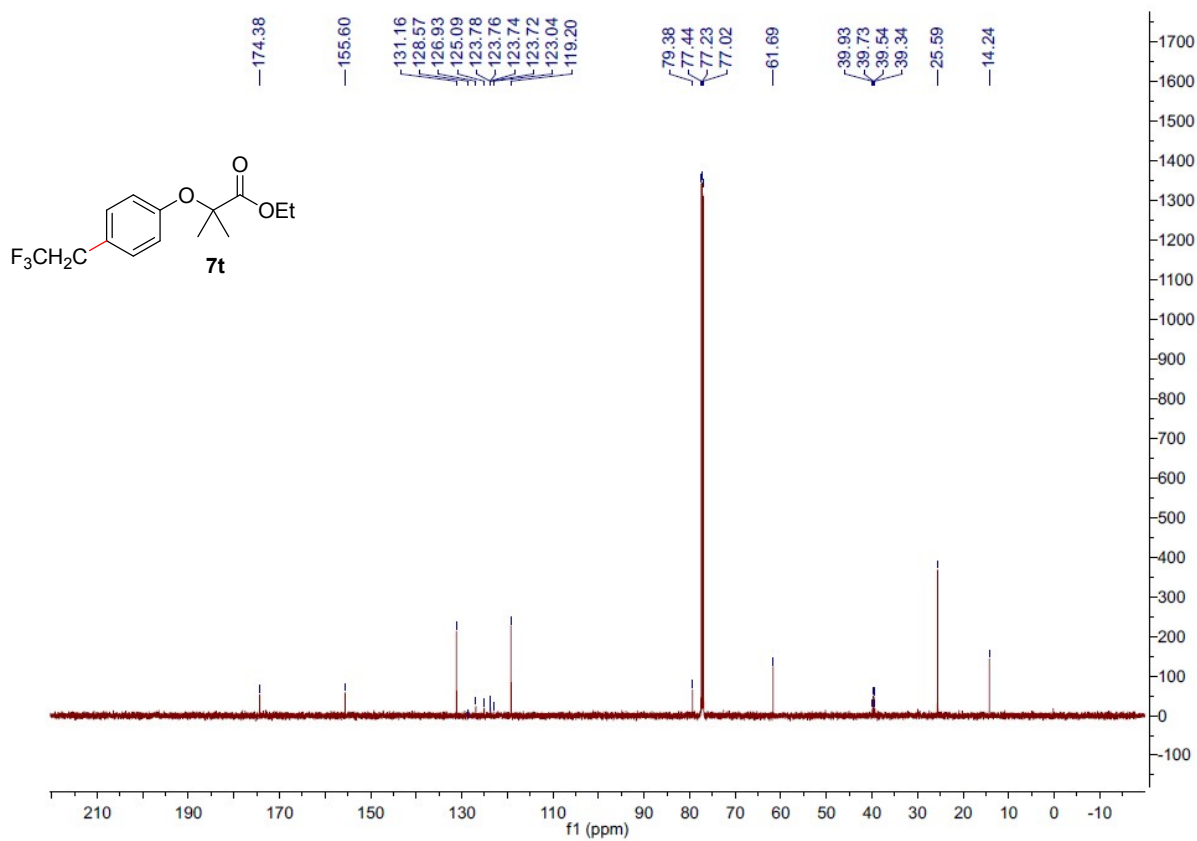


Figure S64. ¹³C NMR of 7t in CDCl₃

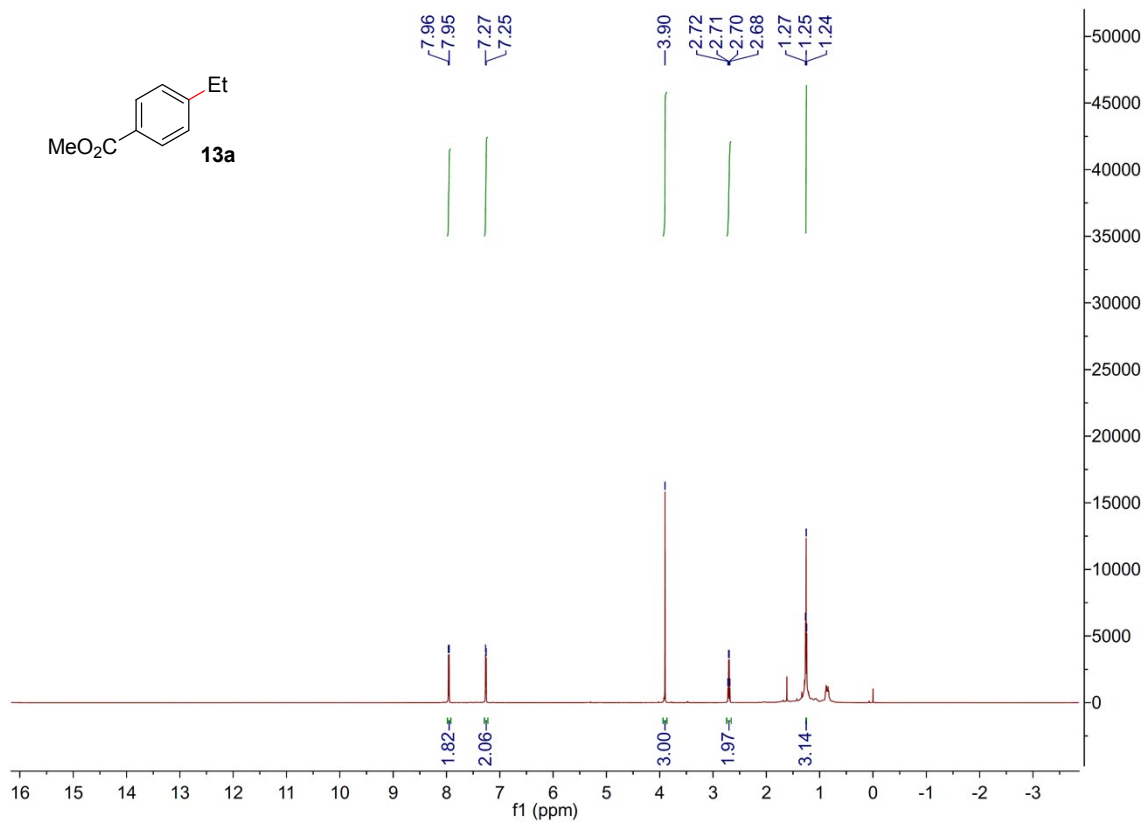


Figure S65. ¹H NMR of **13a** in CDCl₃

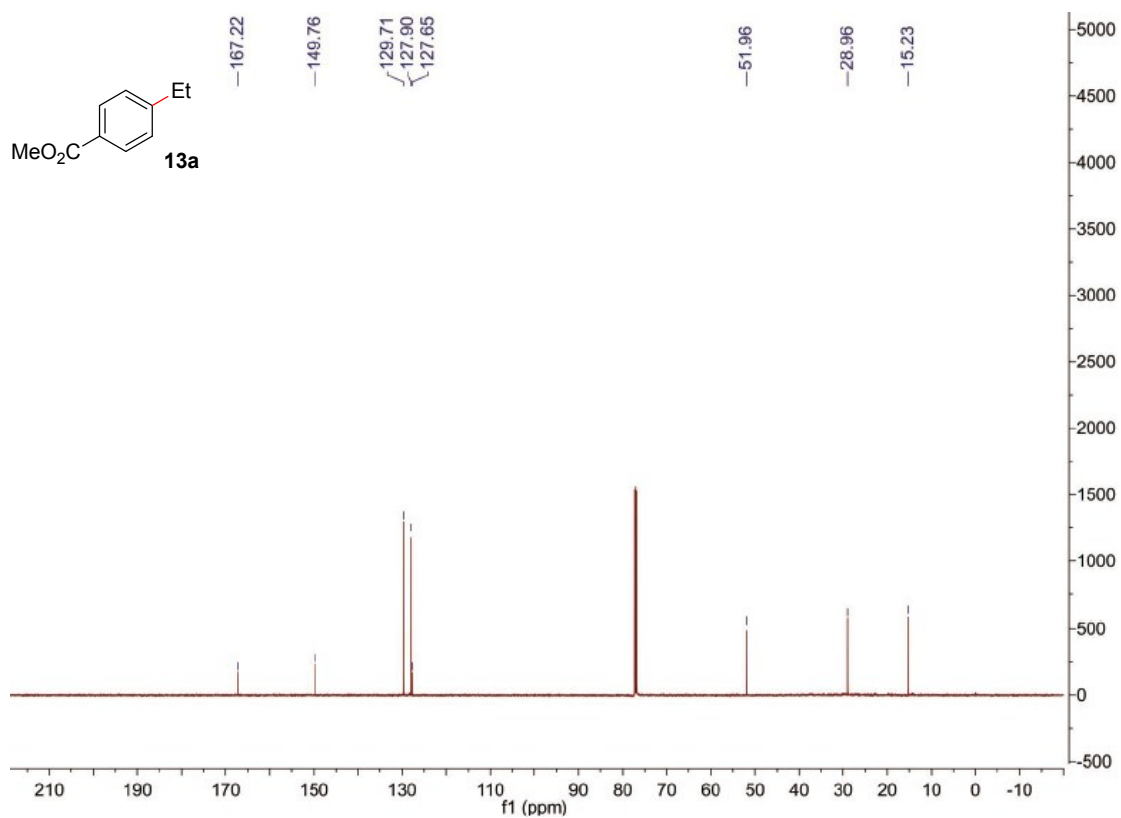


Figure S66. ¹³C NMR of **13a** in CDCl₃

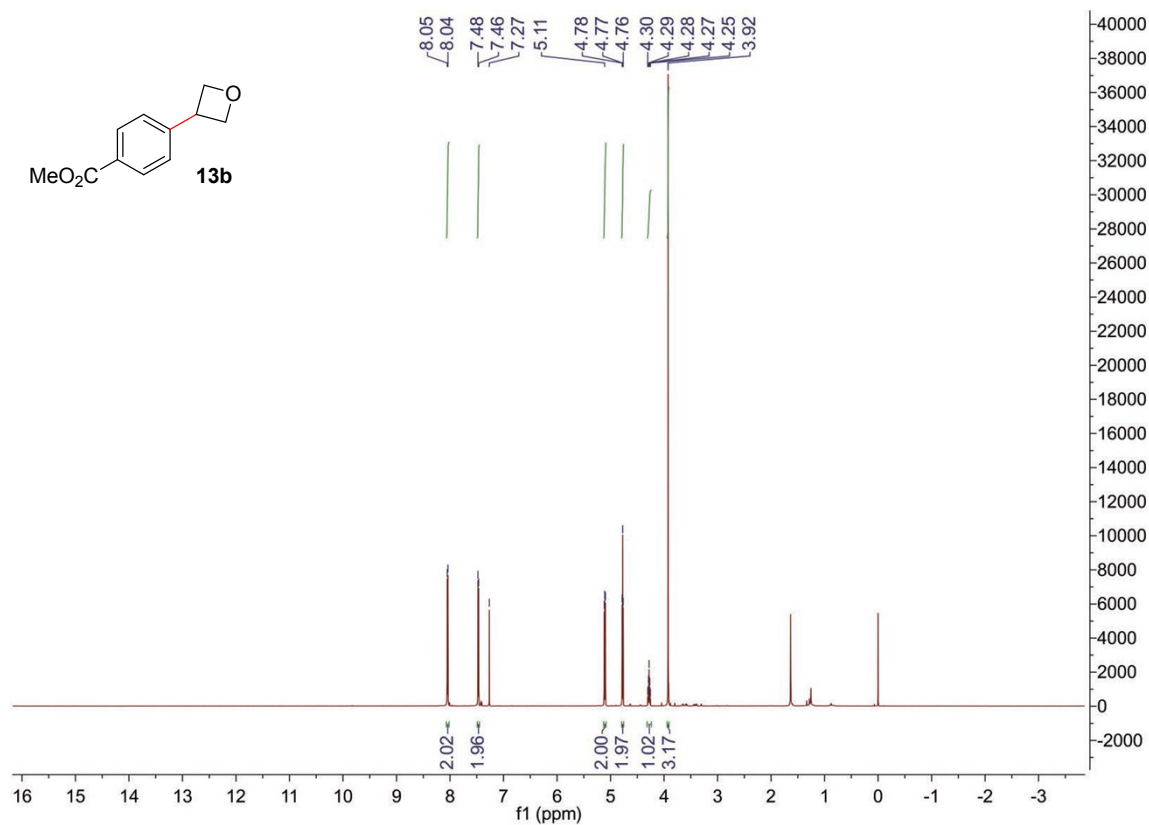


Figure S67. ¹H NMR of **13b** in CDCl₃

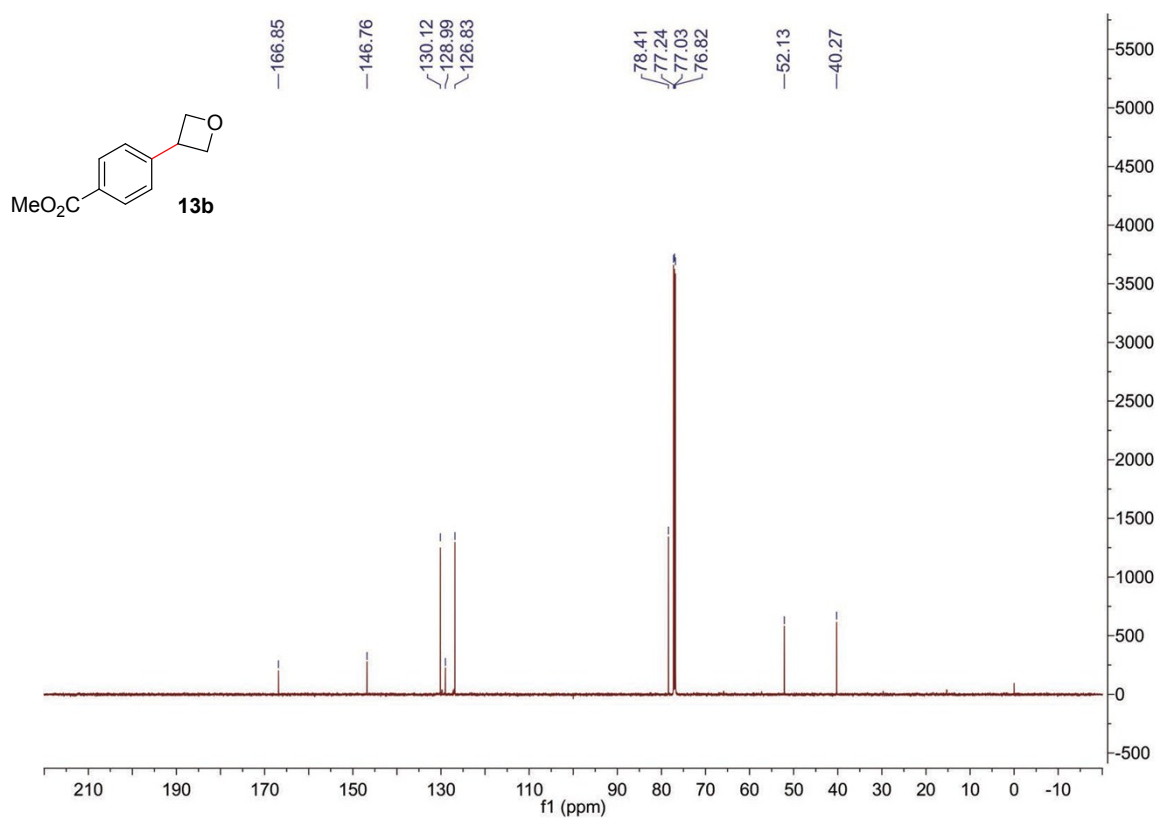


Figure S68. ¹³C NMR of **13b** in CDCl₃

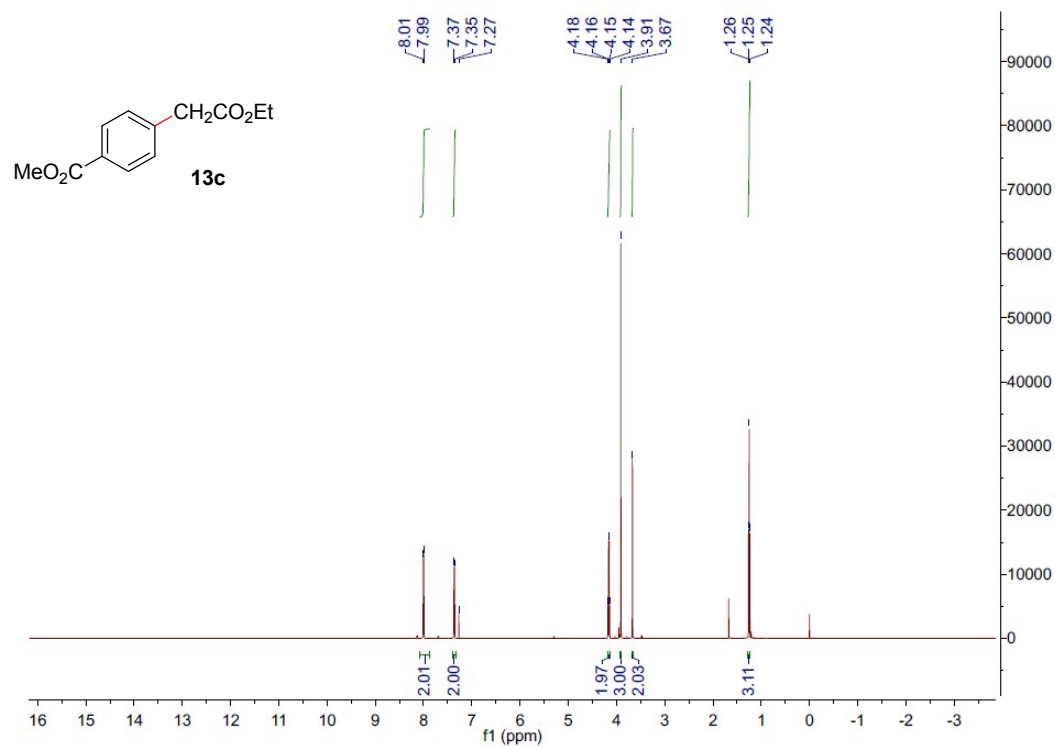


Figure S69. ¹H NMR of **13c** in CDCl₃

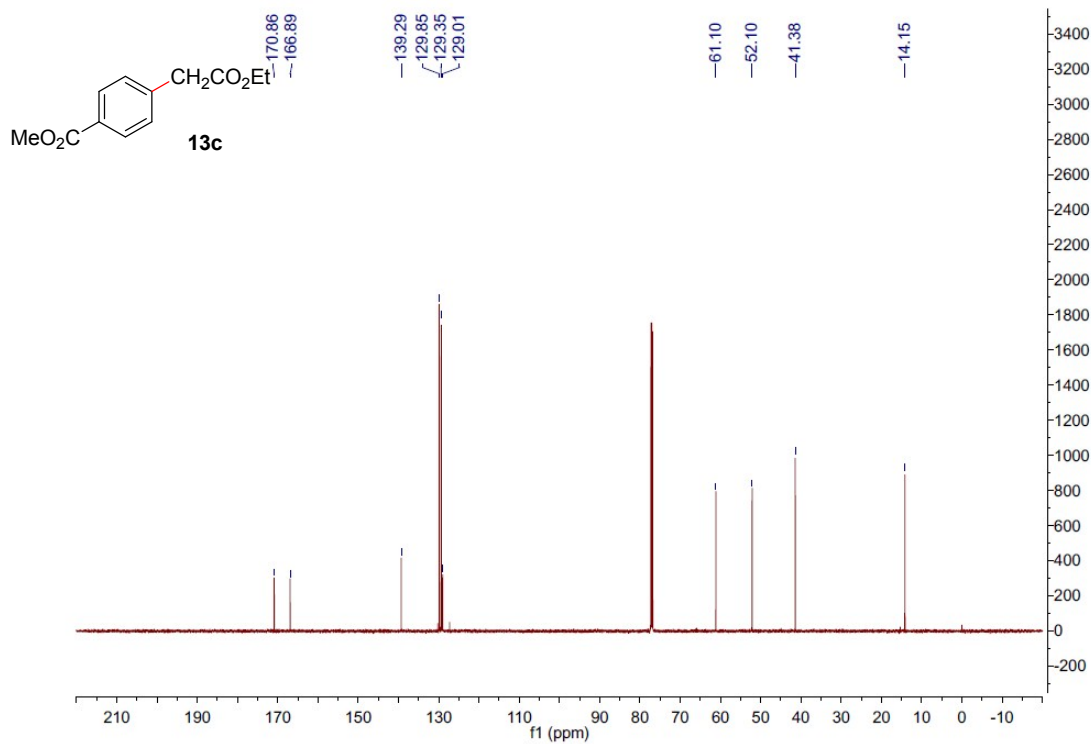


Figure S70. ¹³C NMR of **13c** in CDCl₃

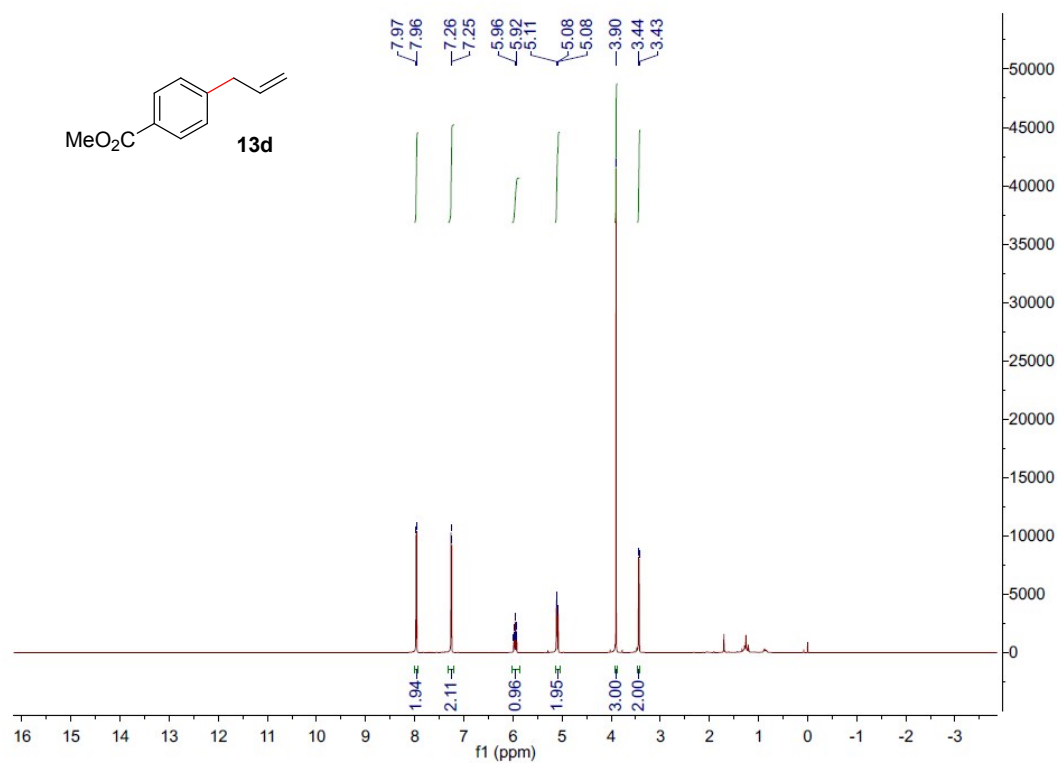


Figure S71. ¹H NMR of **13d** in CDCl₃

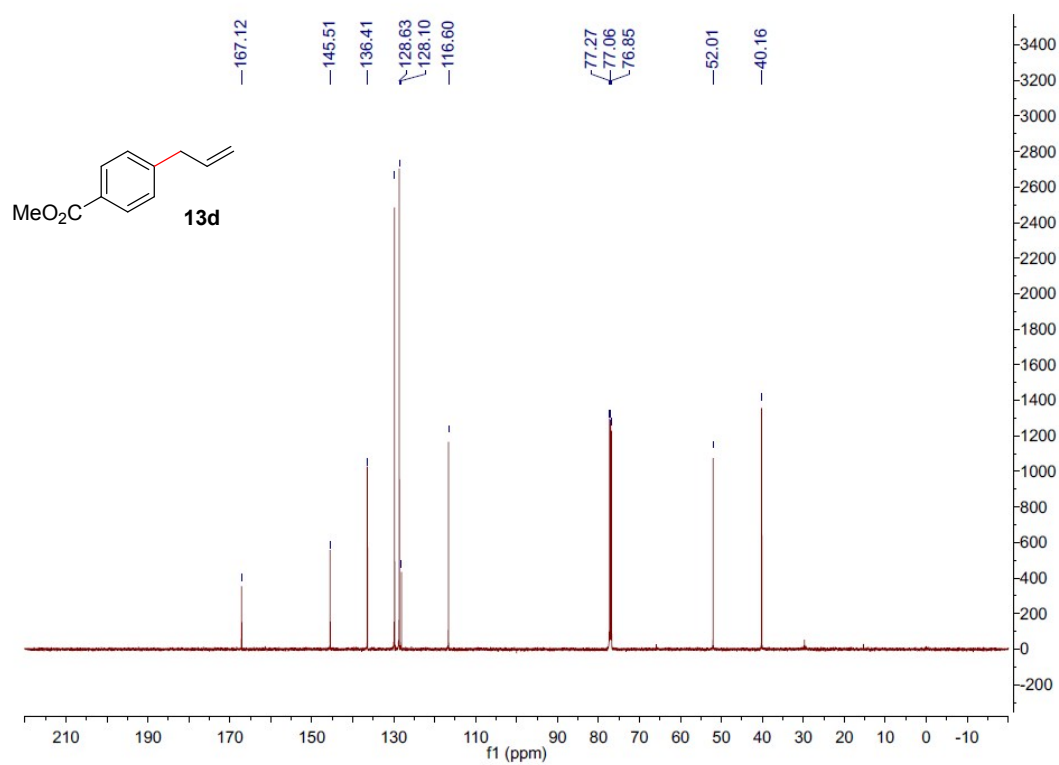


Figure S72. ¹³C NMR of **13d** in CDCl₃

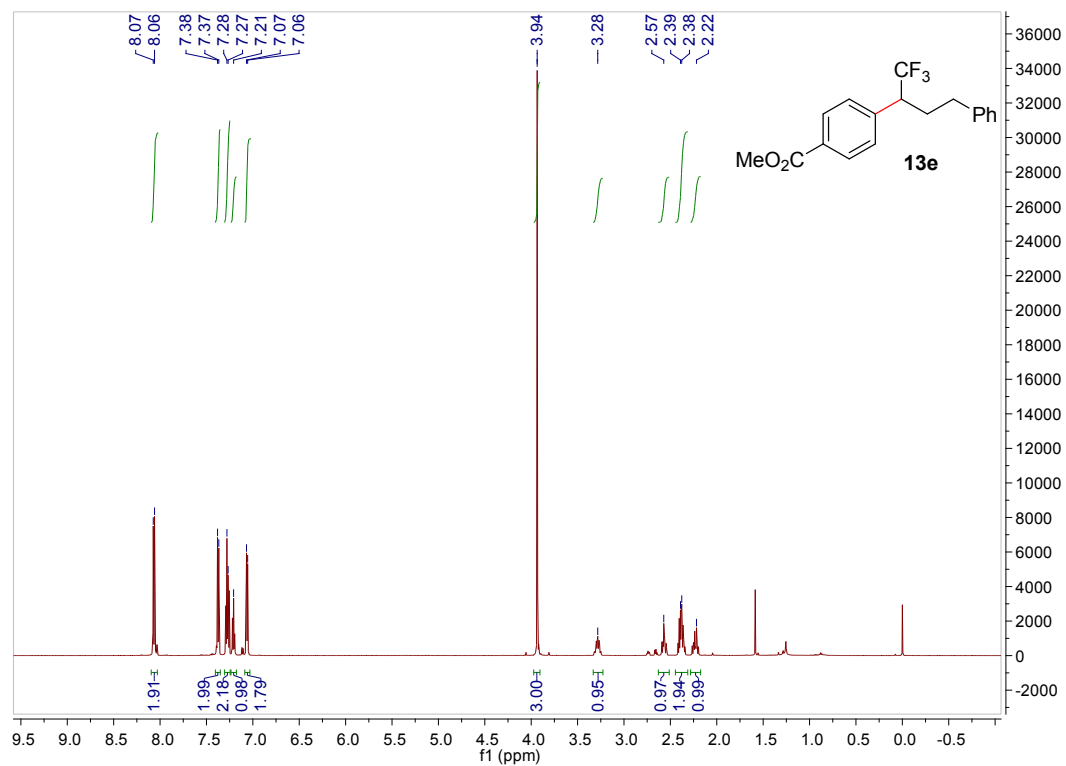


Figure S73. ^1H NMR of **13e** in CDCl_3

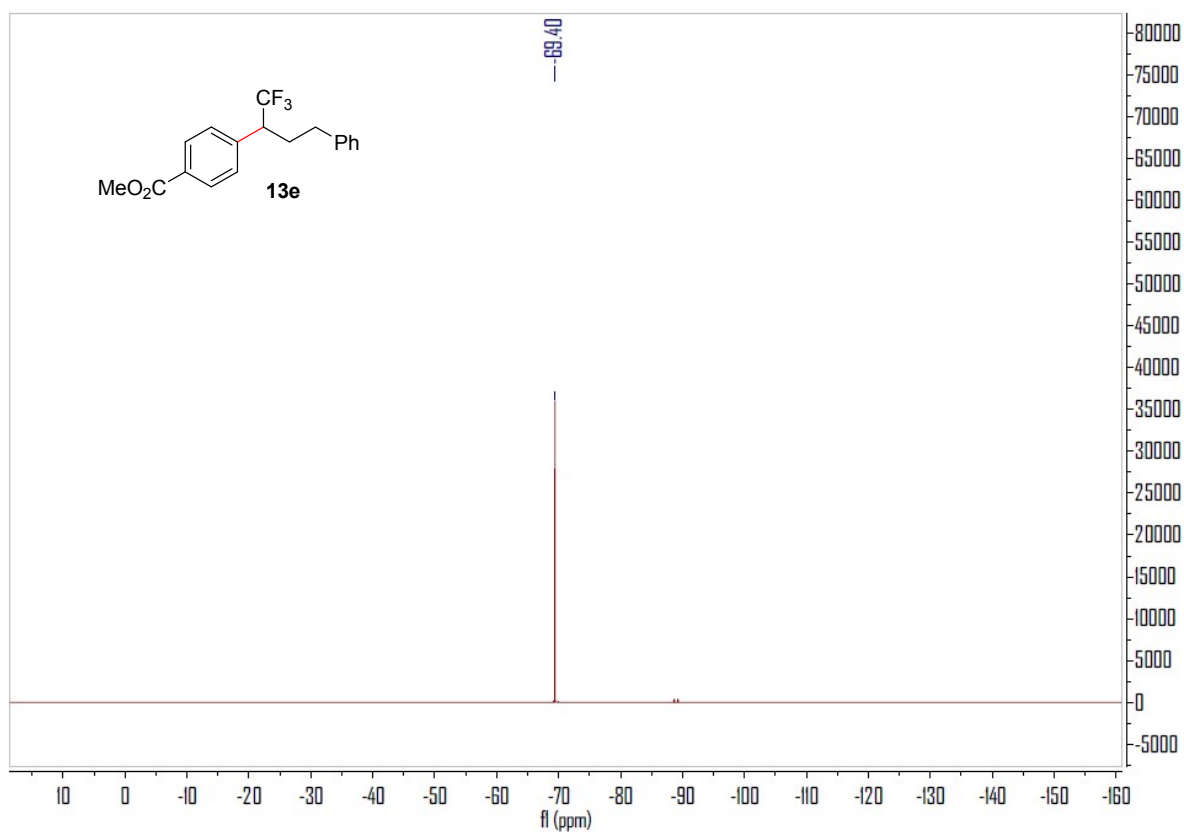


Figure S74. ^{19}F NMR of **13e** in CDCl_3

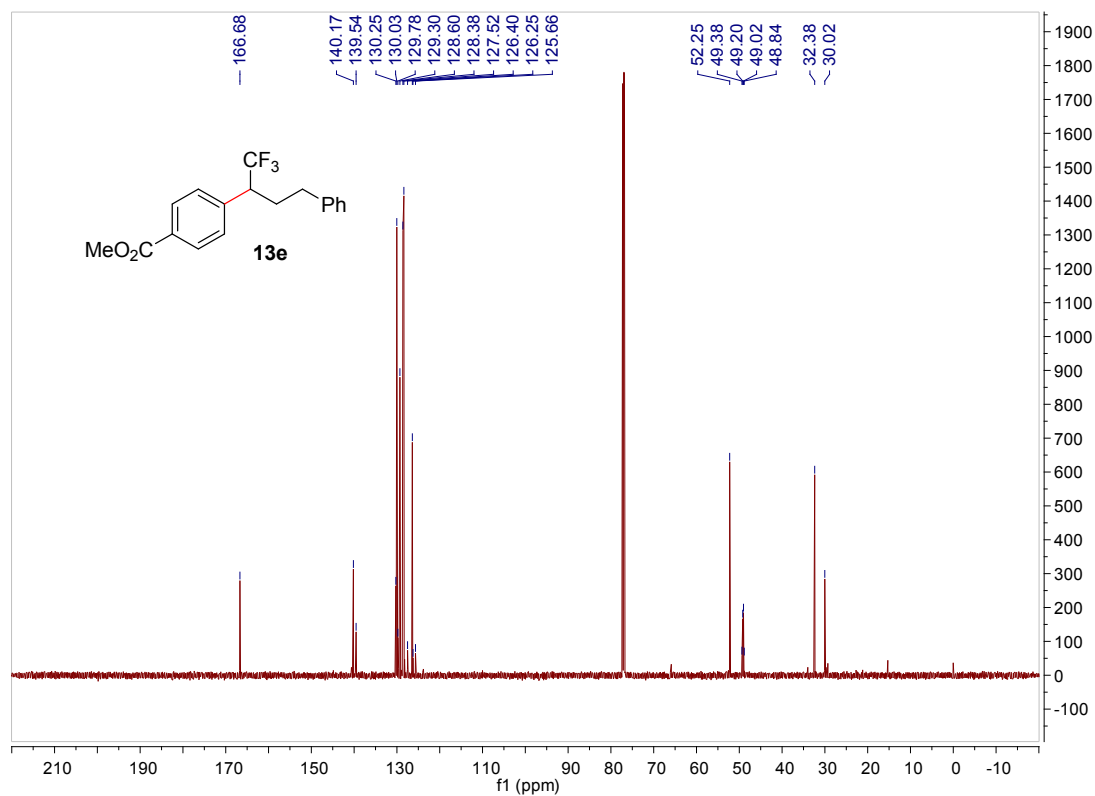


Figure S75. ¹³C NMR of **13e** in CDCl₃

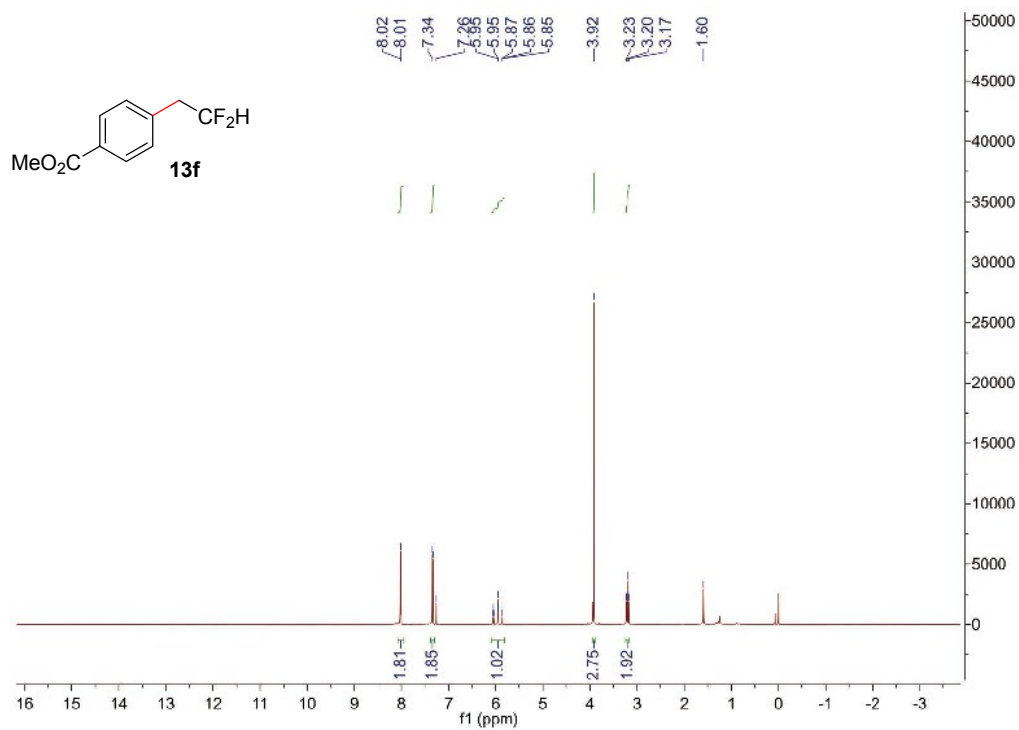


Figure S76. ¹H NMR of **13f** in CDCl₃

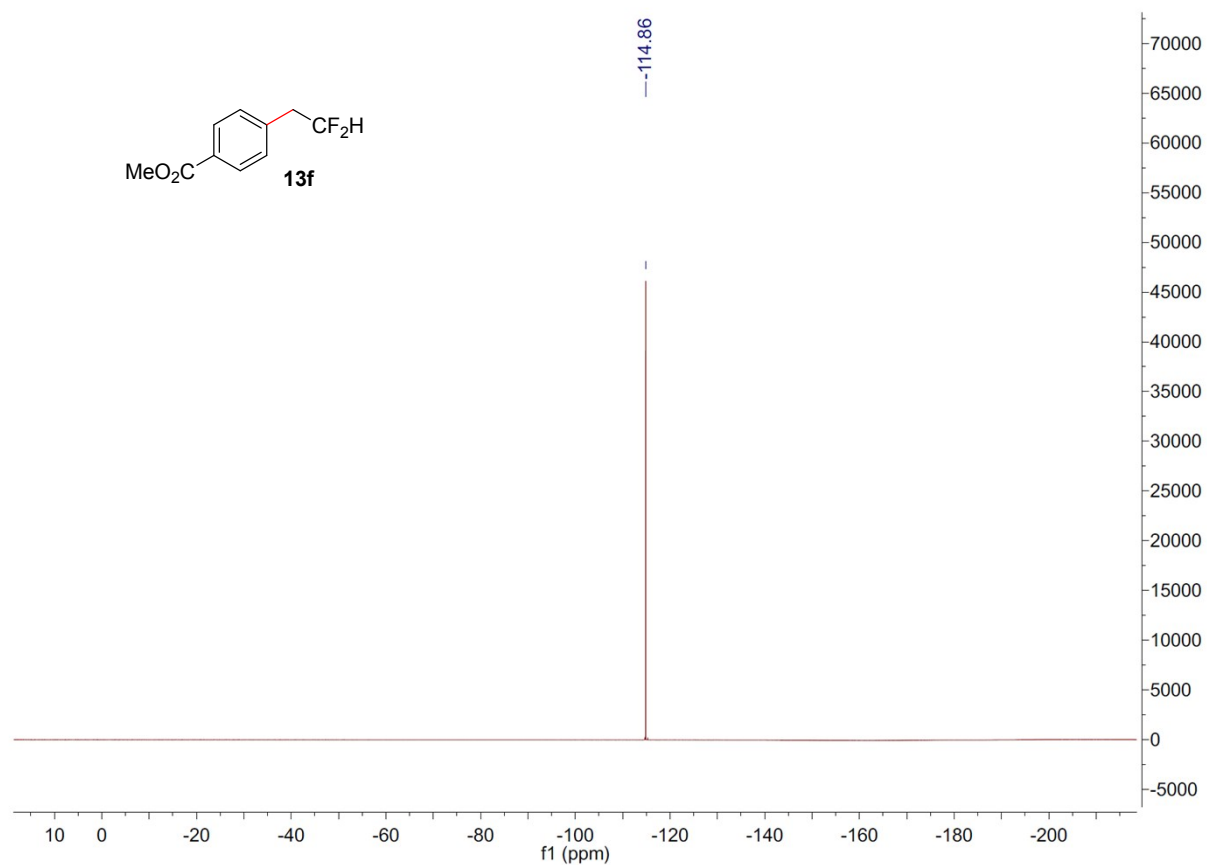


Figure S77. ¹⁹F NMR of **13f** in CDCl₃

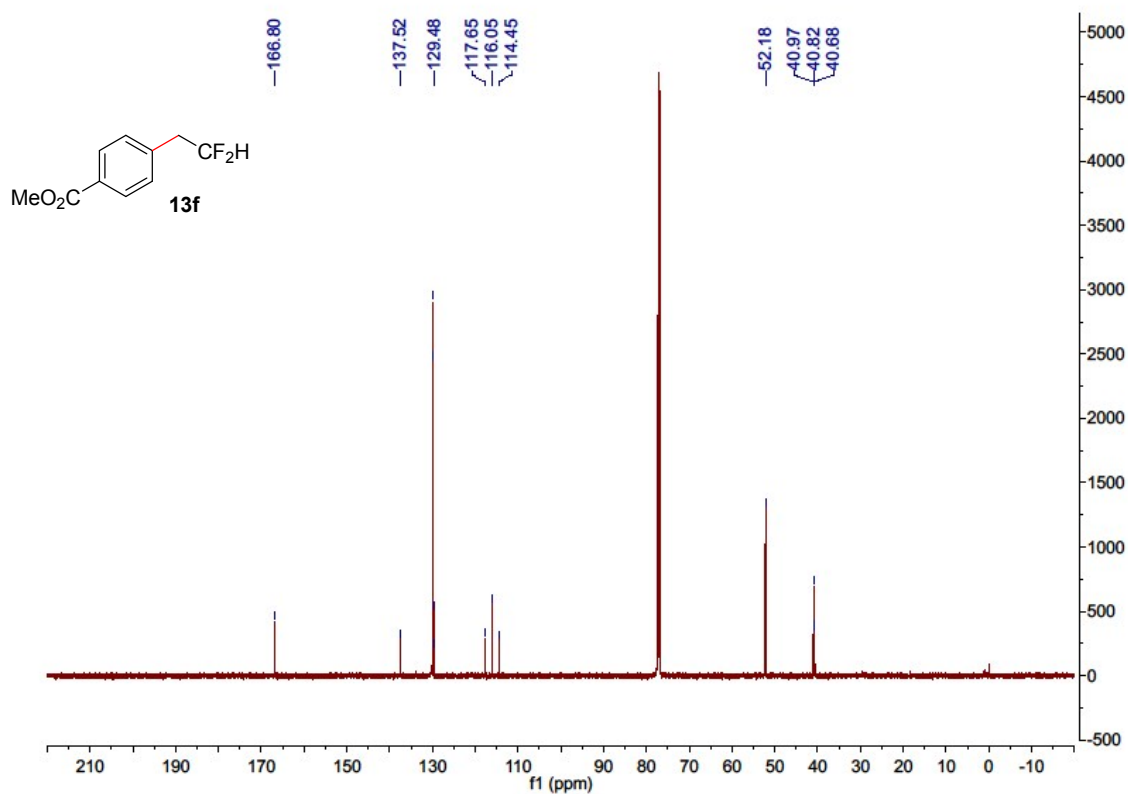


Figure S78. ¹³C NMR of **13f** in CDCl₃

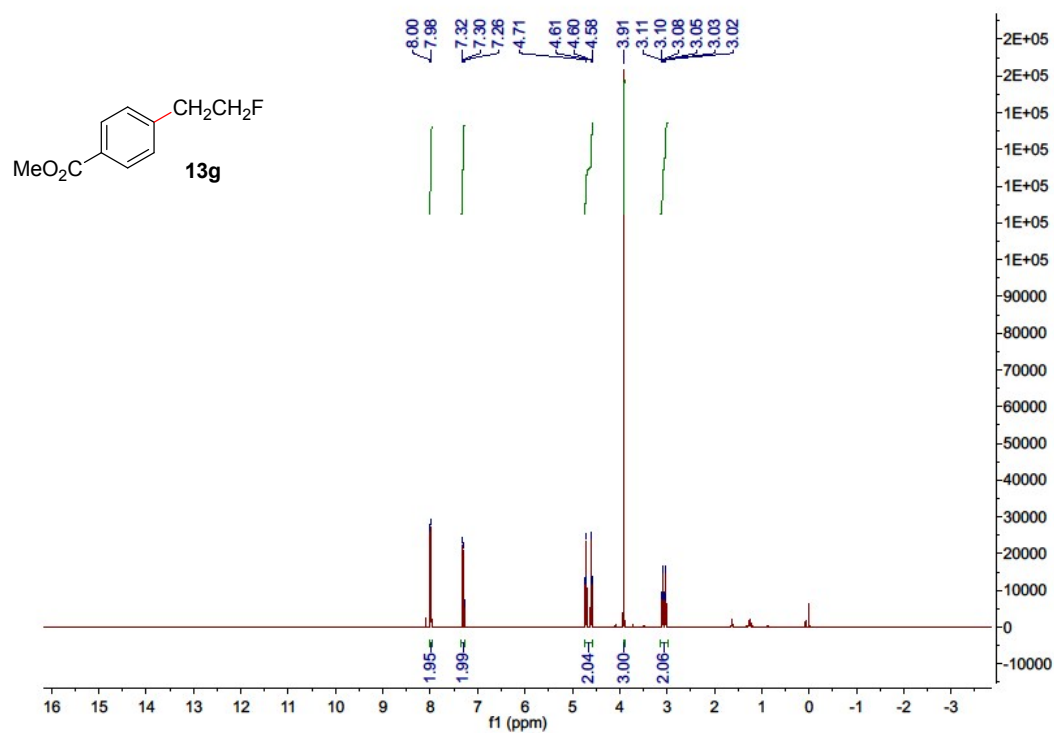


Figure S79. ¹H NMR spectra of **13g** in CDCl₃

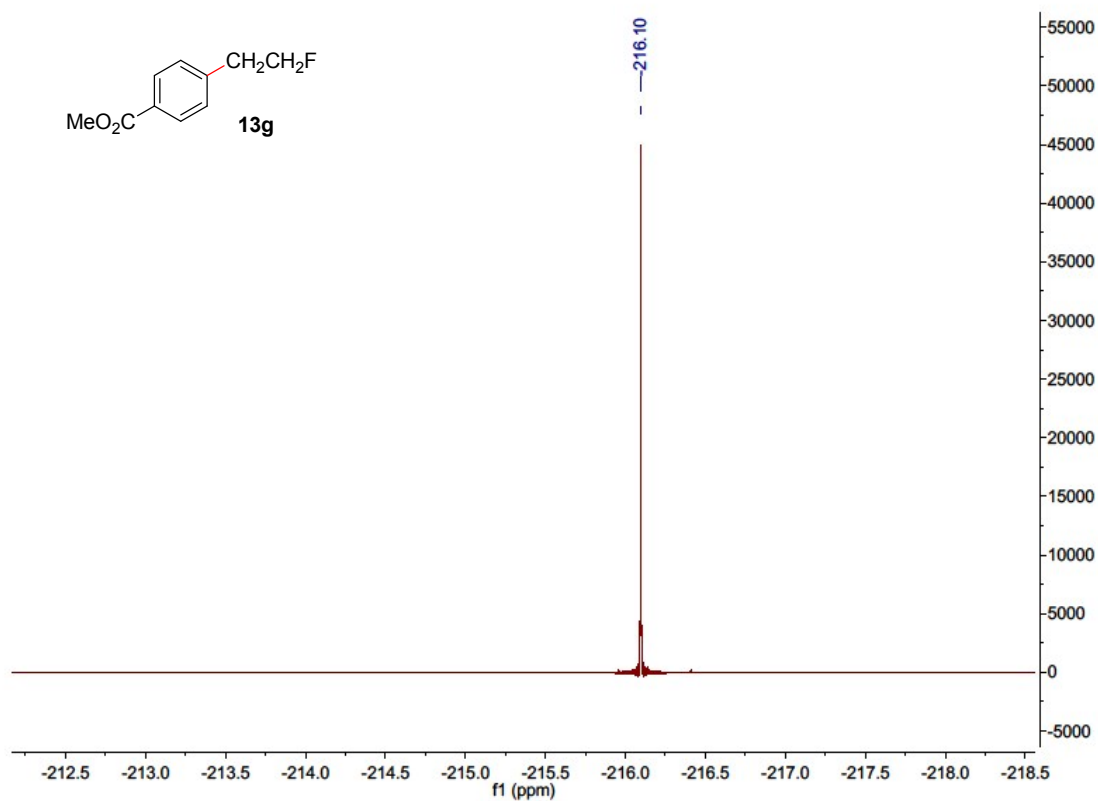


Figure S80. ¹⁹F NMR spectra of **13g** in CDCl₃

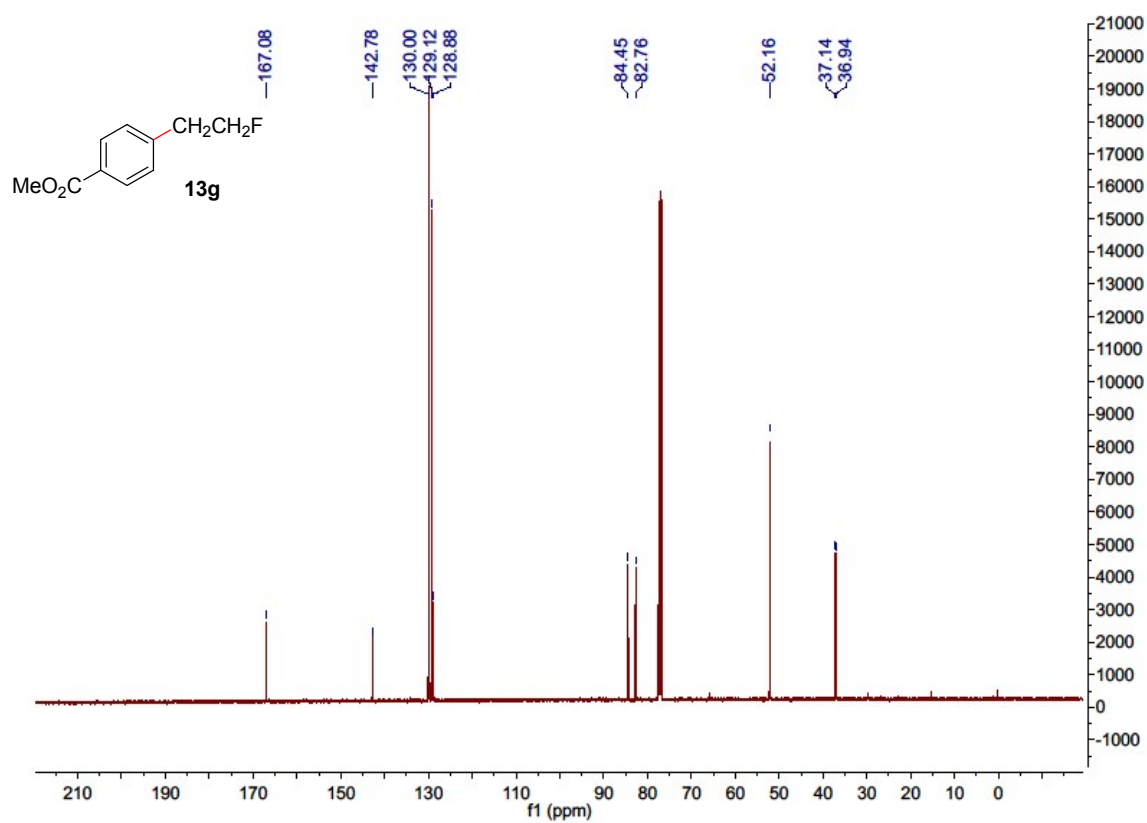


Figure S81. ¹³C NMR spectra of **3g** in CDCl₃

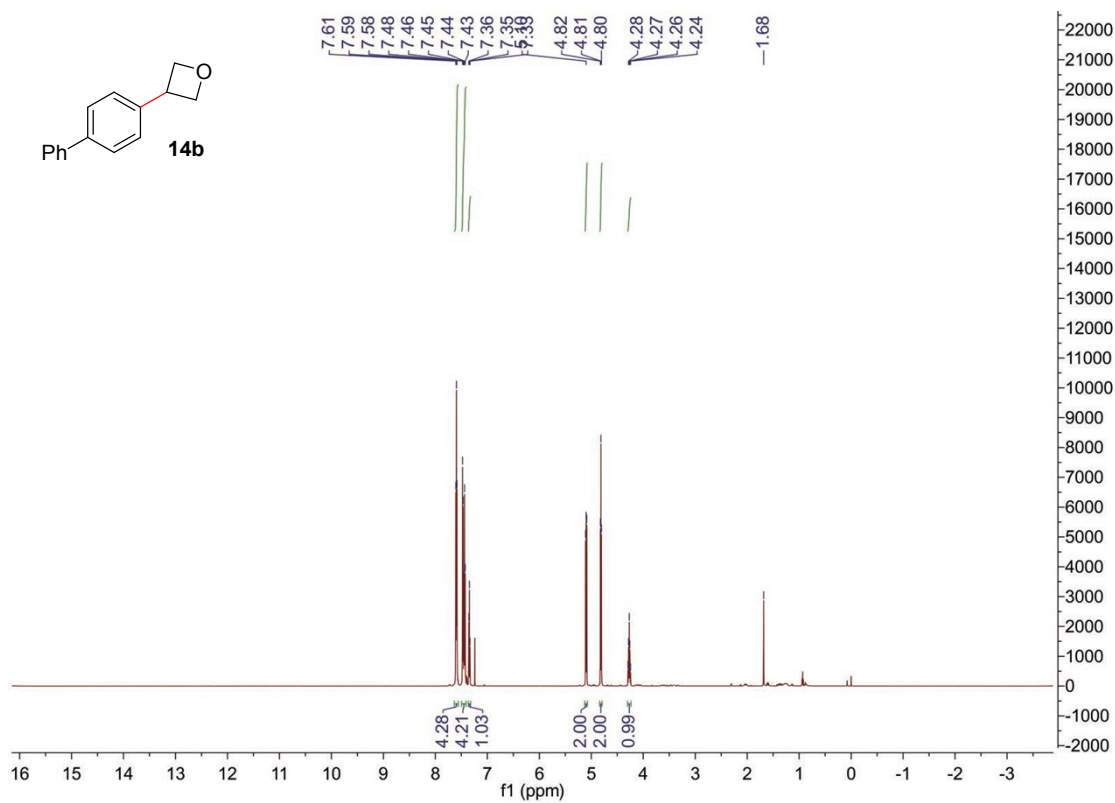


Figure S82. ¹H NMR of **14b** in CDCl₃

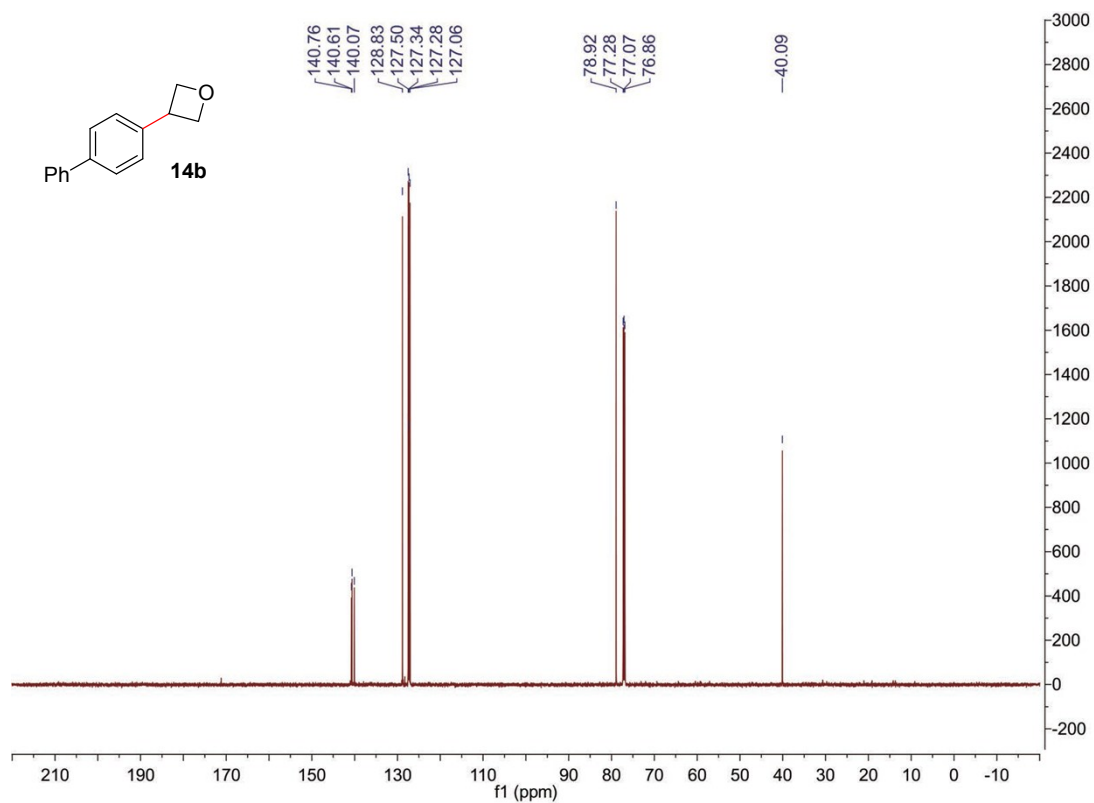


Figure S83. ¹³C NMR of **14b** in CDCl₃

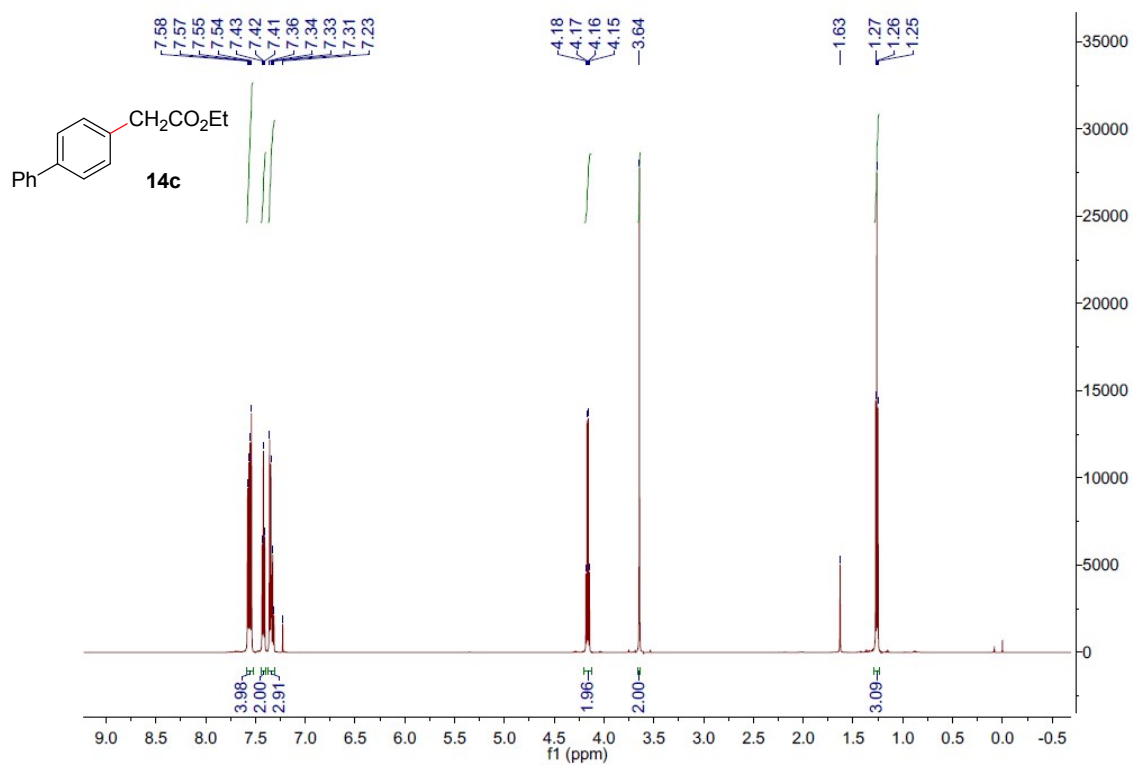


Figure S84. ¹H NMR of **14c** in CDCl₃

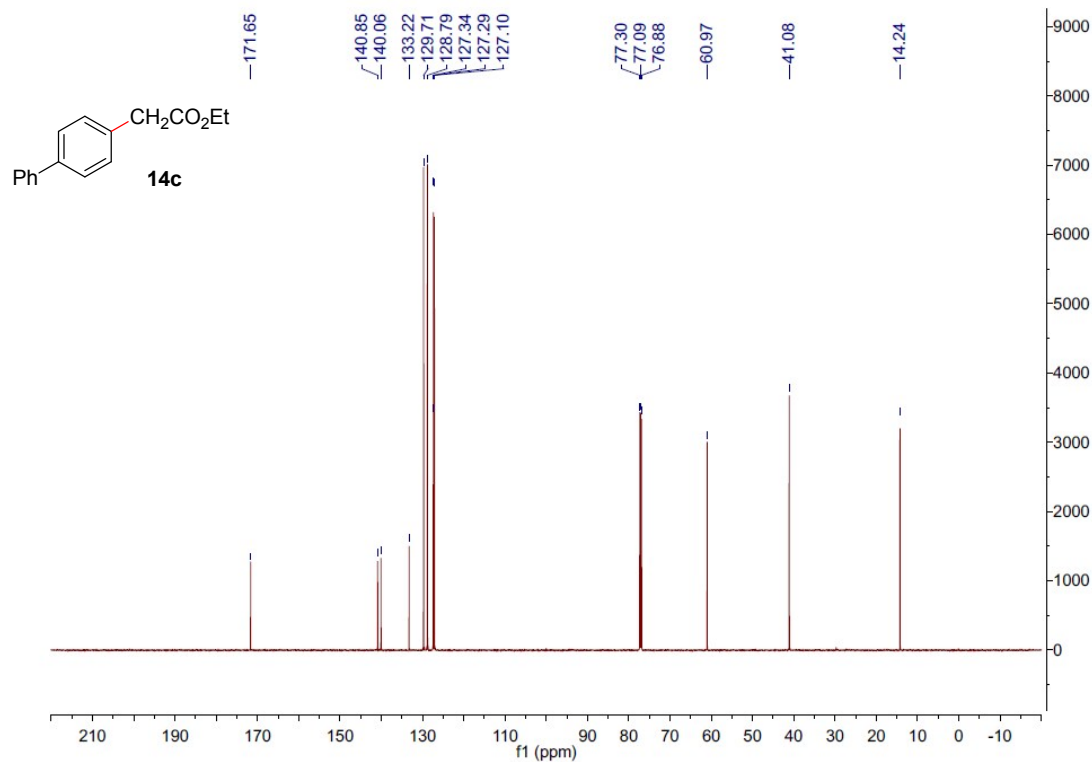


Figure S85. ¹³C NMR of **14c** in CDCl₃

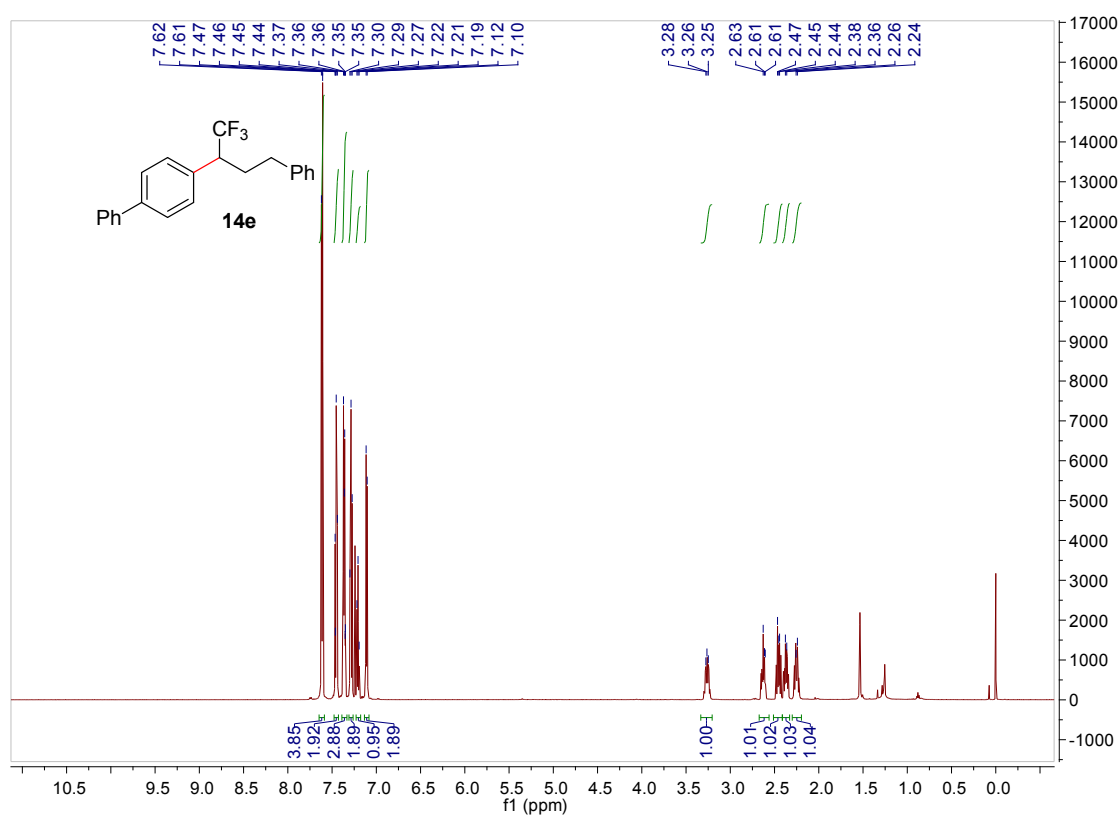


Figure S86. ¹H NMR of **14e** in CDCl₃

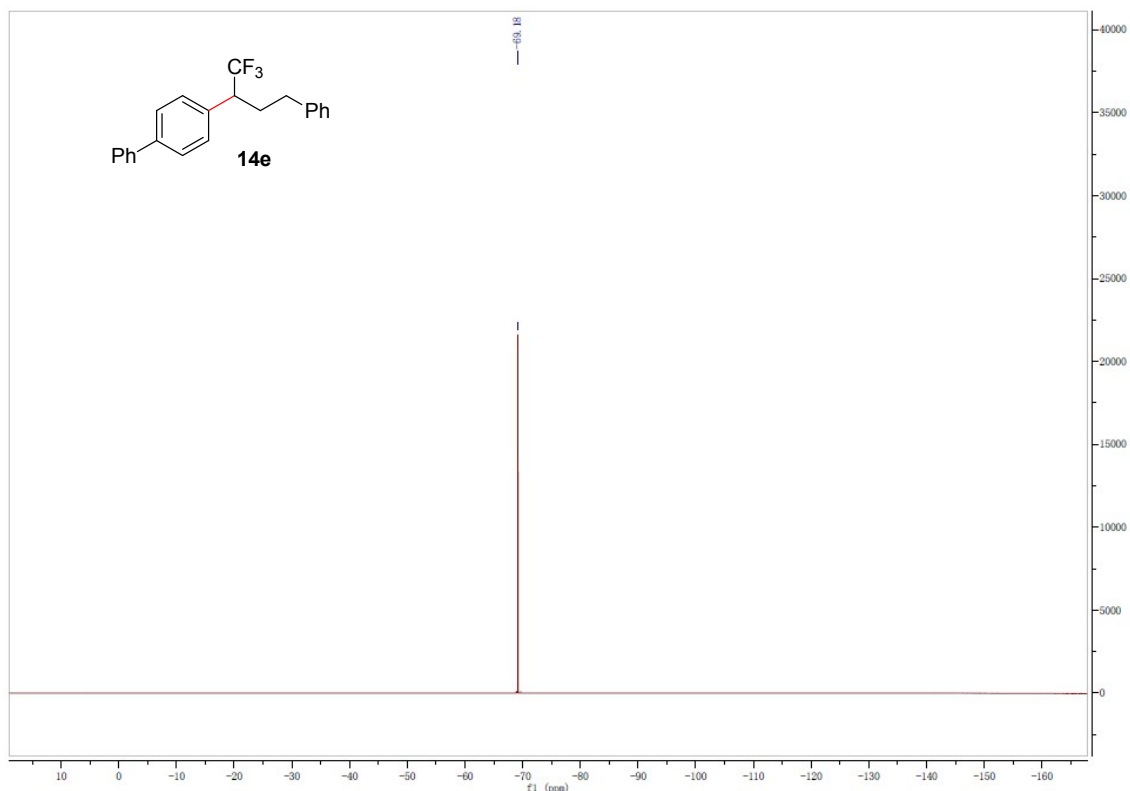


Figure S87. ¹⁹F NMR of **14e** in CDCl₃

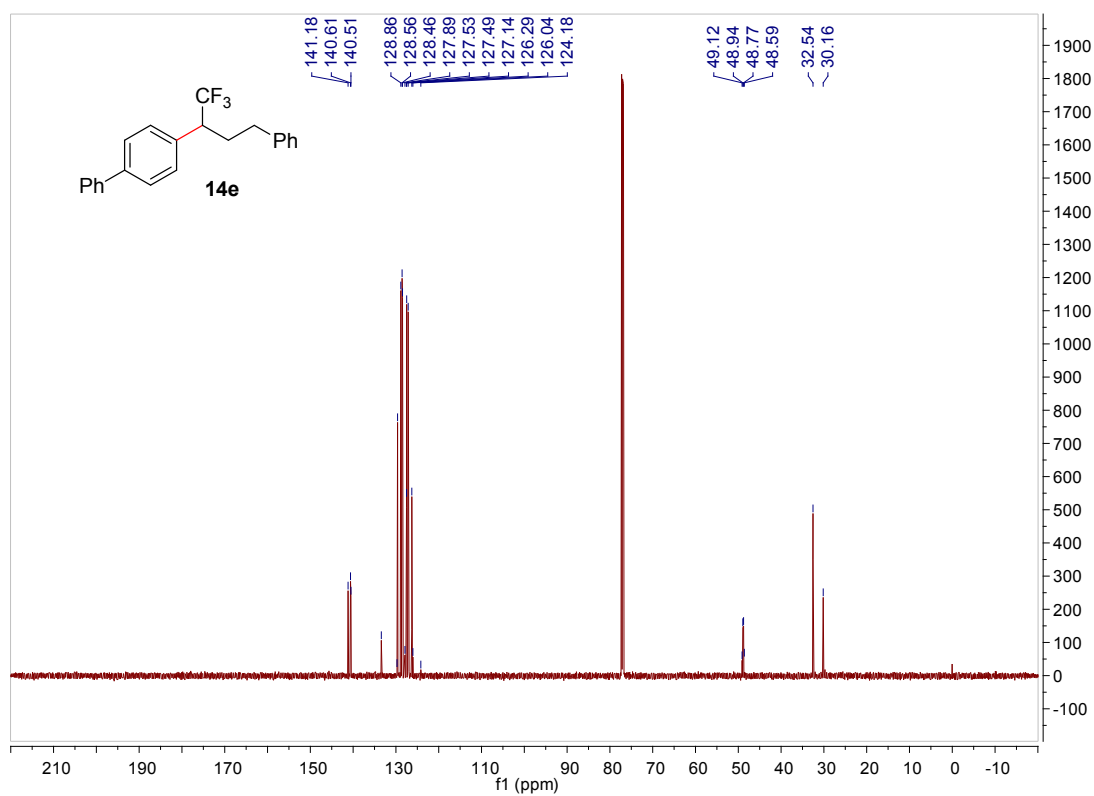


Figure S88. ¹³C NMR of **14e** in CDCl₃

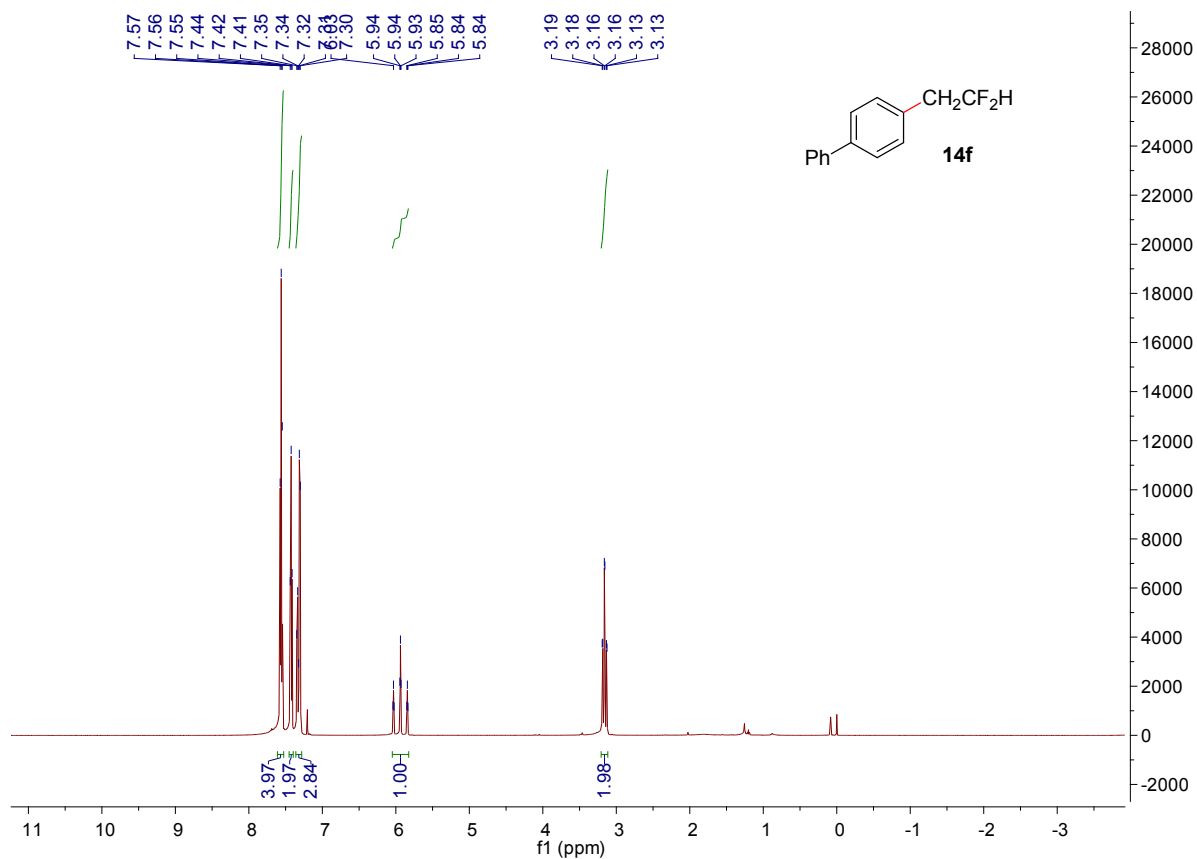


Figure S89. ¹H NMR of 14f in CDCl₃

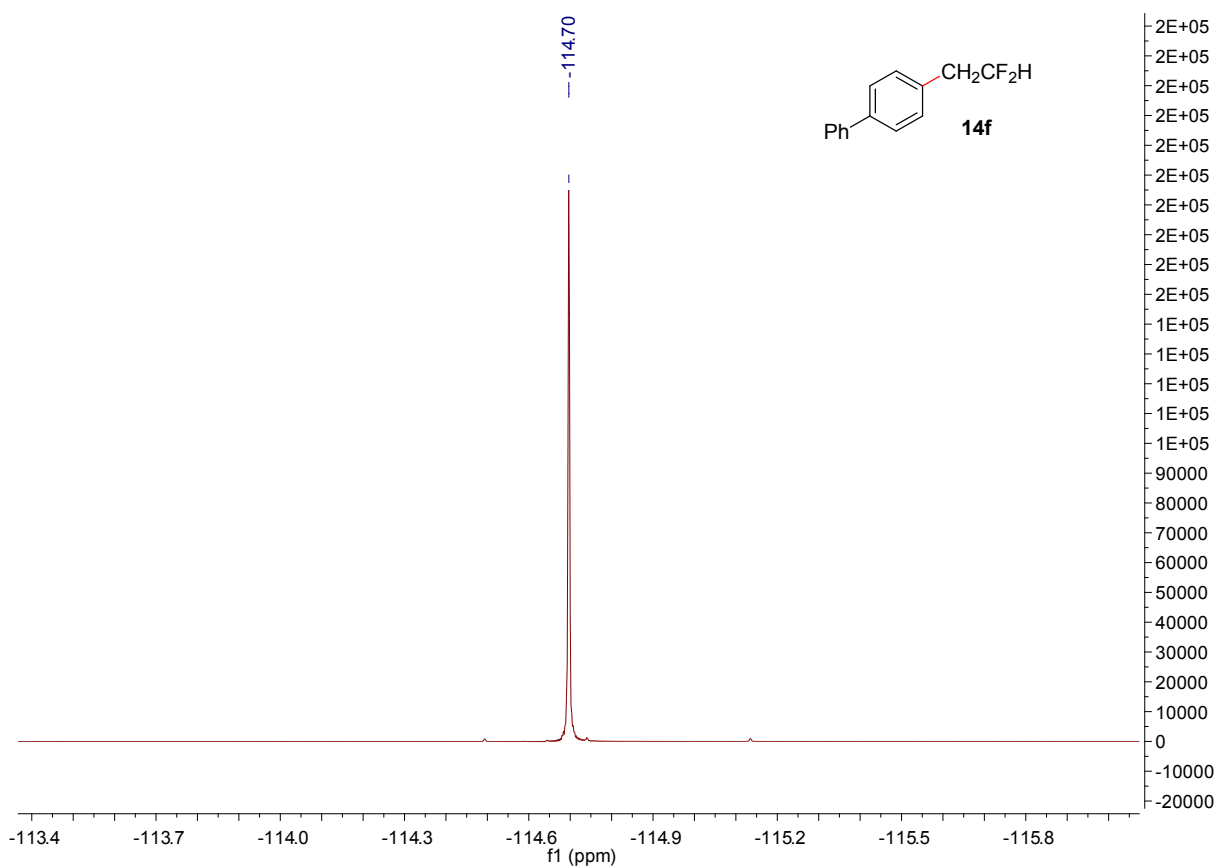


Figure S90. ¹⁹F NMR of 14f in CDCl₃

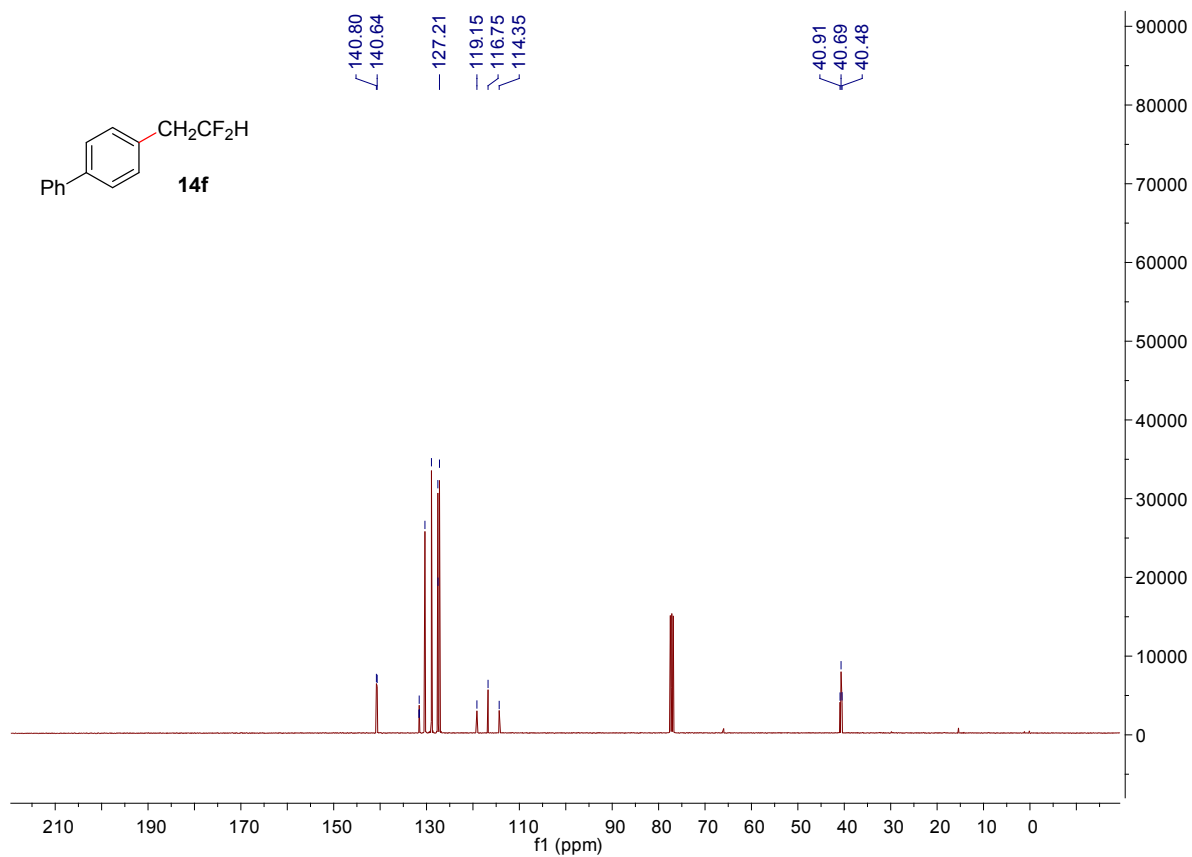


Figure S91. ¹³C NMR of **14f** in CDCl₃

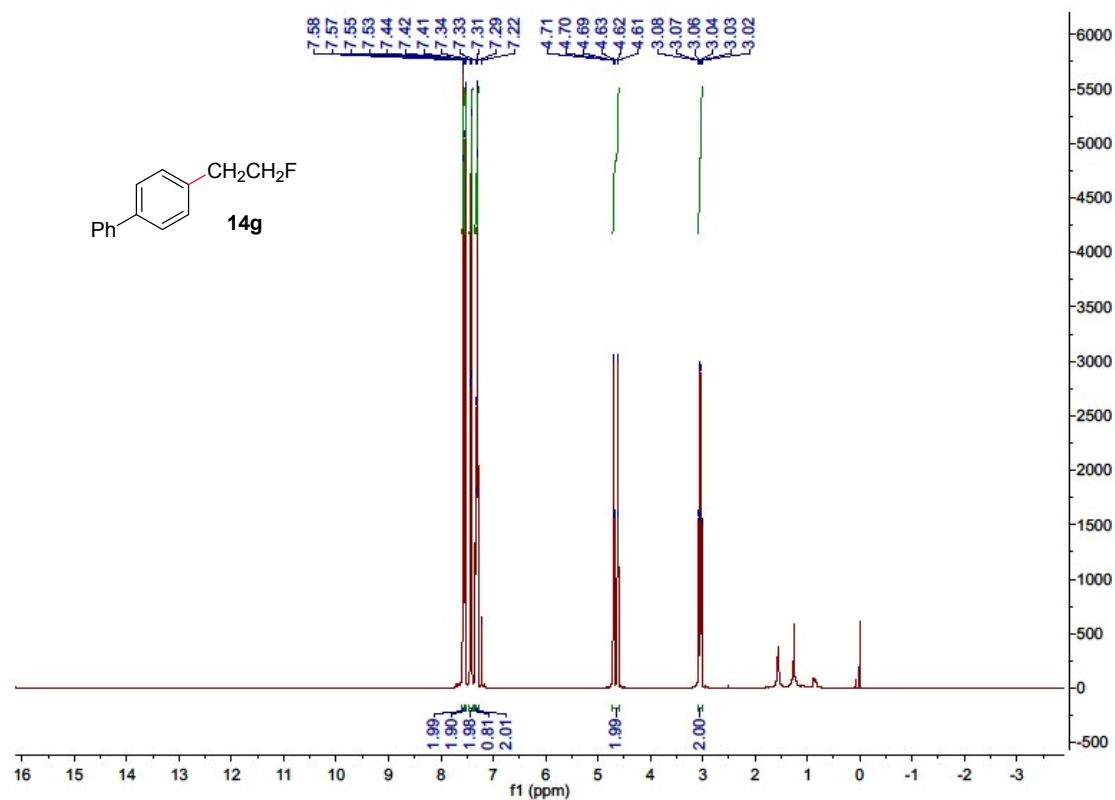


Figure S92. ¹H NMR of **14g** in CDCl₃

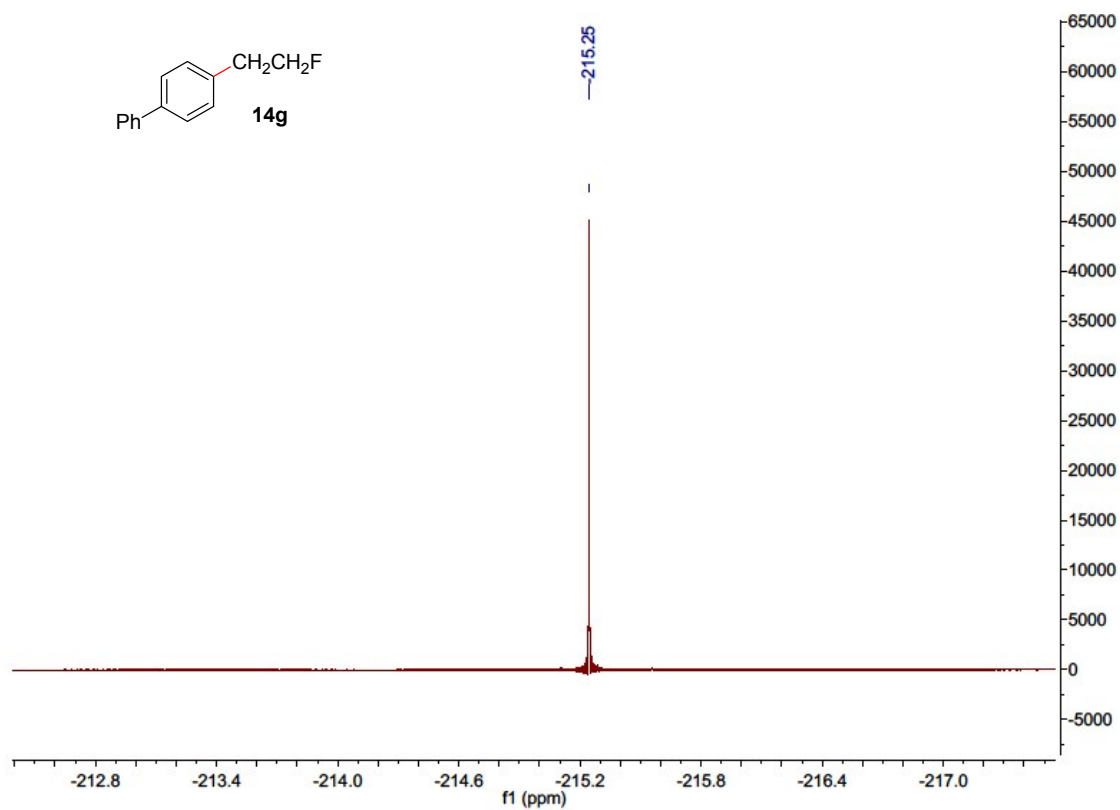


Figure S93. ¹⁹F NMR of **14g** in CDCl₃

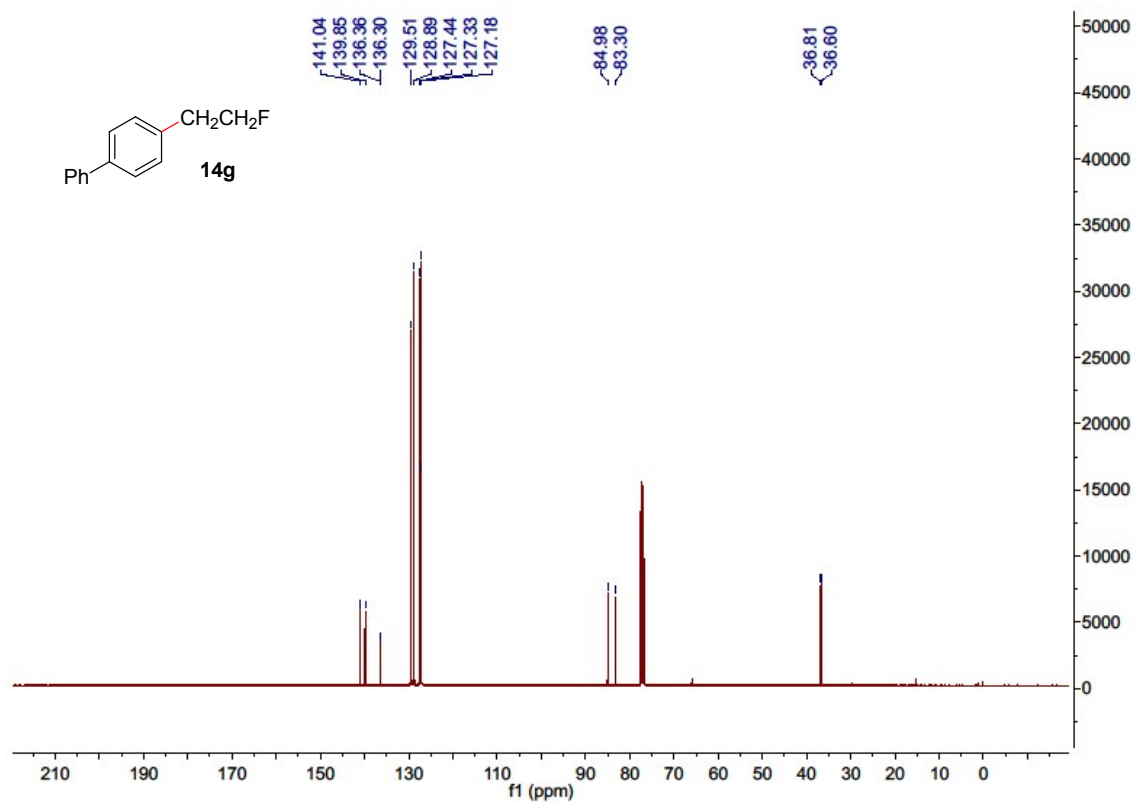


Figure S94. ¹³C NMR of **14g** in CDCl₃

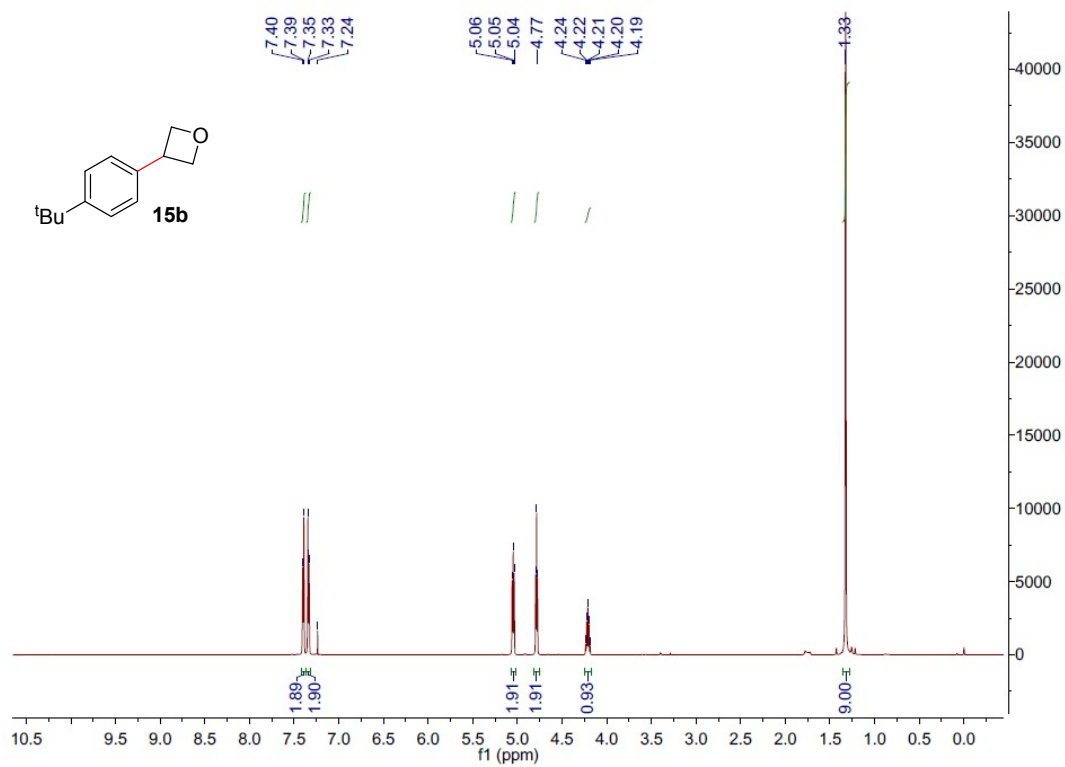


Figure S95. ¹H NMR of 15b in CDCl₃

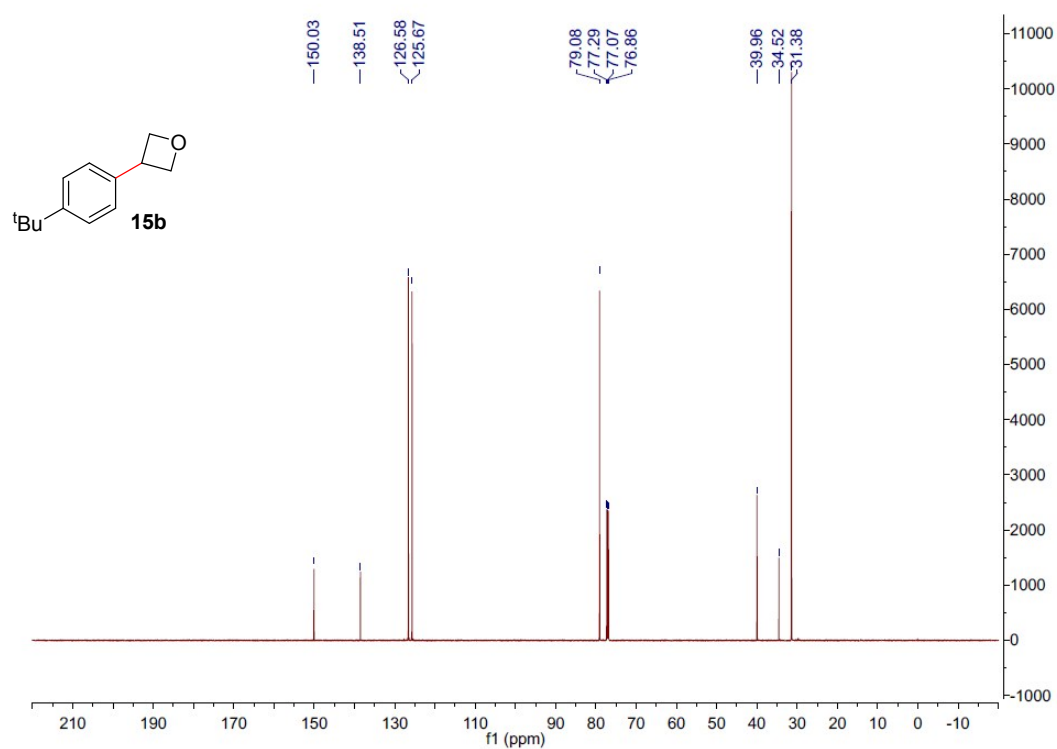


Figure S96. ¹³C NMR of 15b in CDCl₃

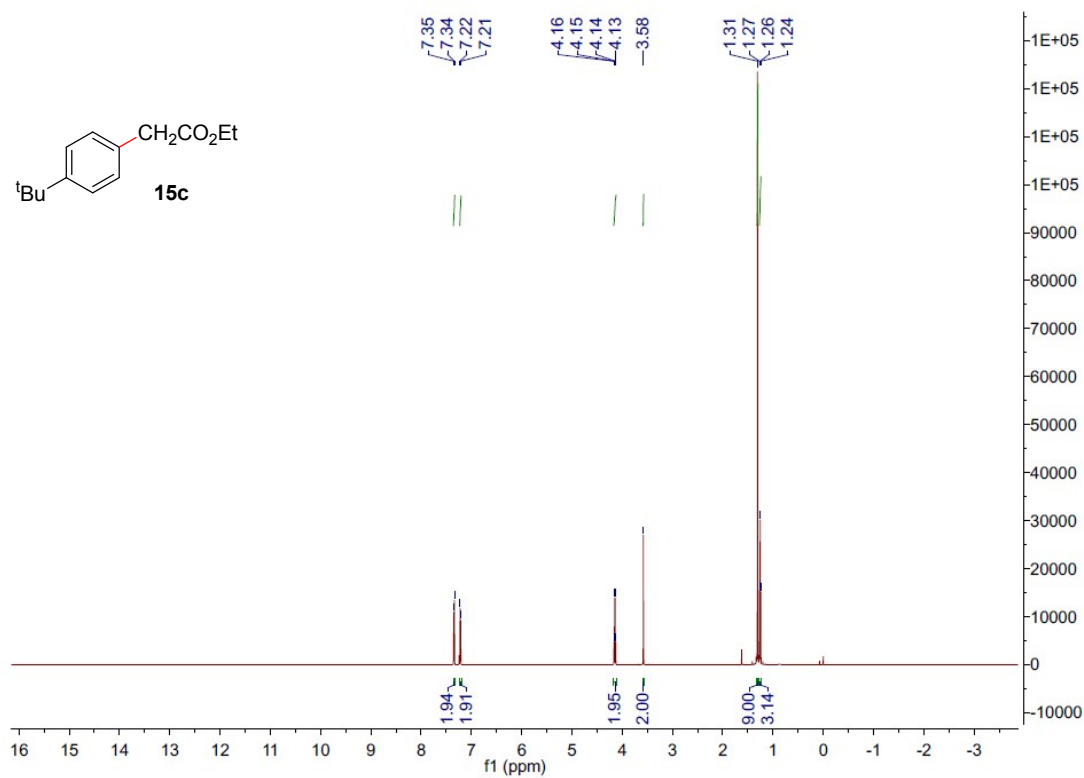


Figure S97. ^1H NMR of **15c** in CDCl_3

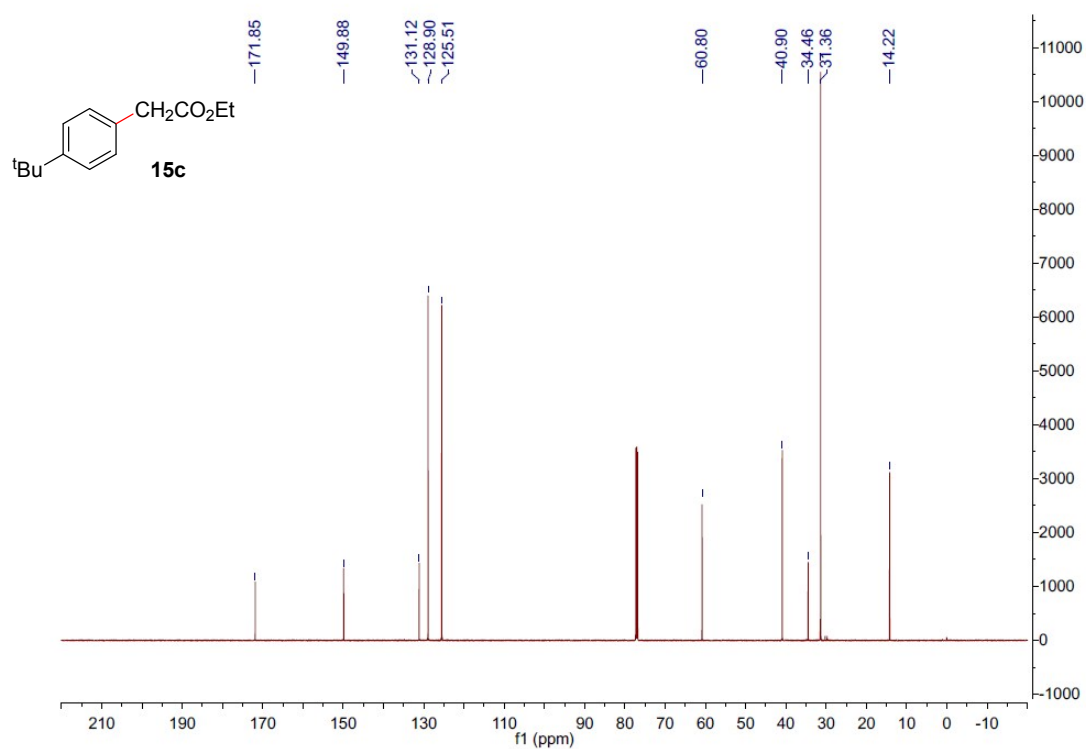


Figure S98. ^{13}C NMR of **15c** in CDCl_3

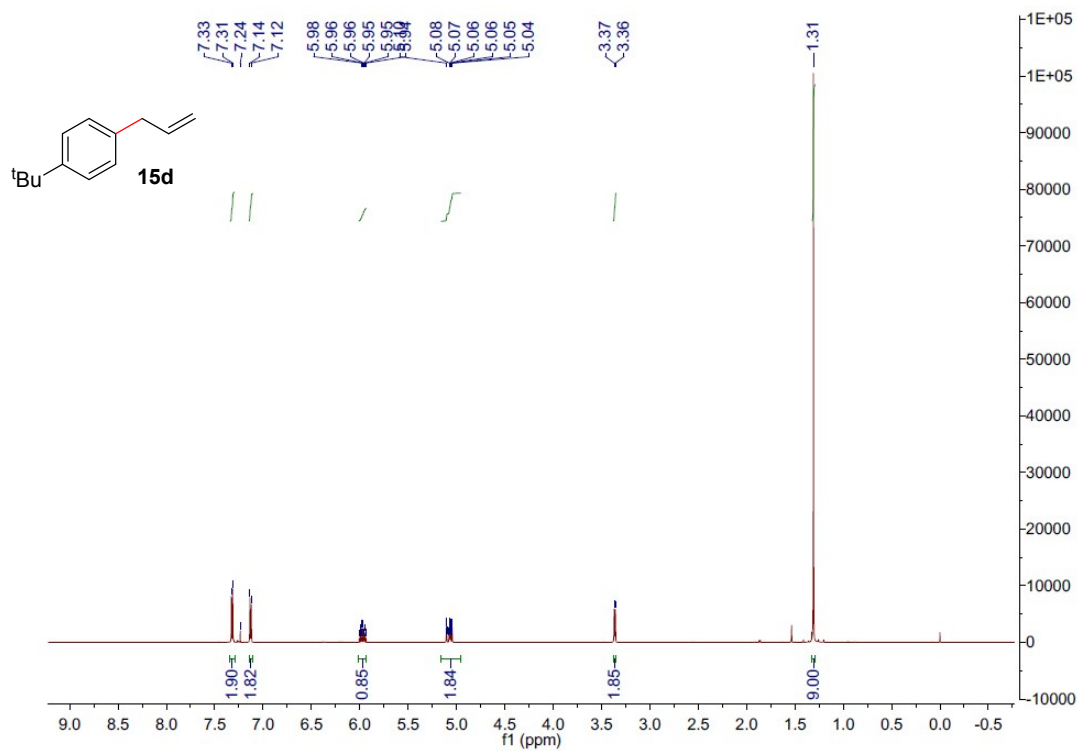


Figure S99. ¹H NMR of 15d in CDCl₃

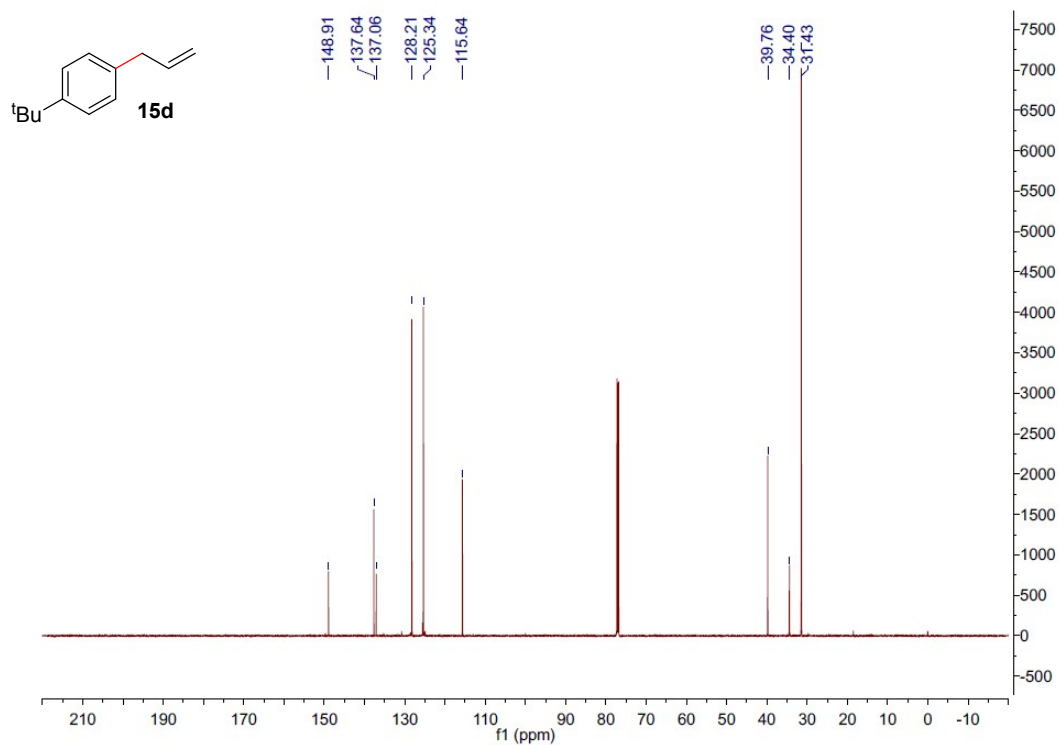


Figure S100. ¹³C NMR of 15d in CDCl₃

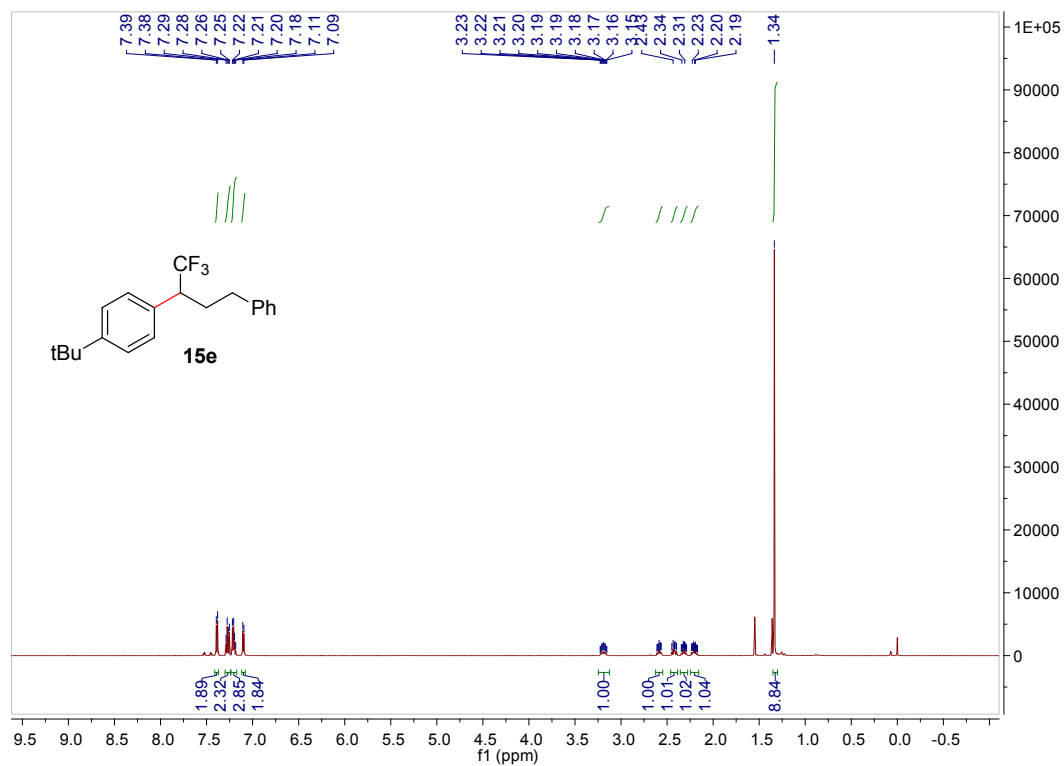


Figure S101. ¹H NMR of **15e** in CDCl₃

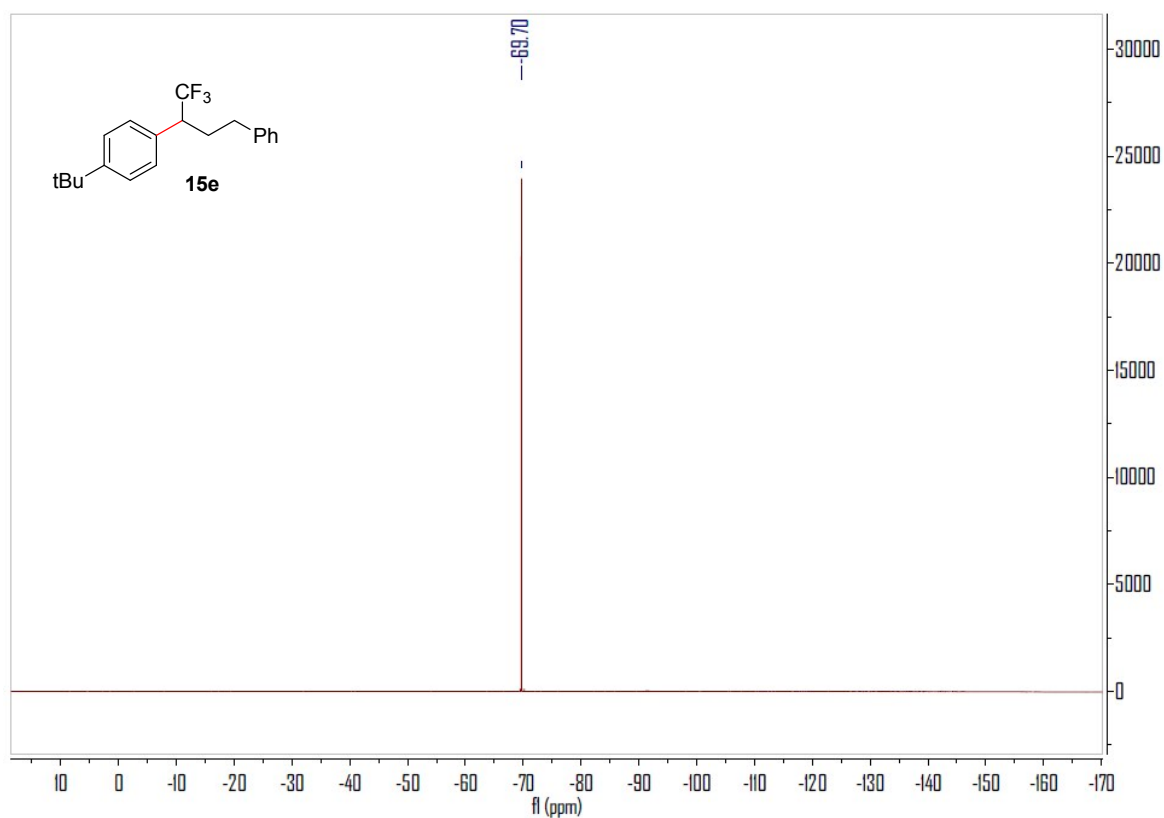


Figure S102. ¹⁹F NMR of **15e** in CDCl₃

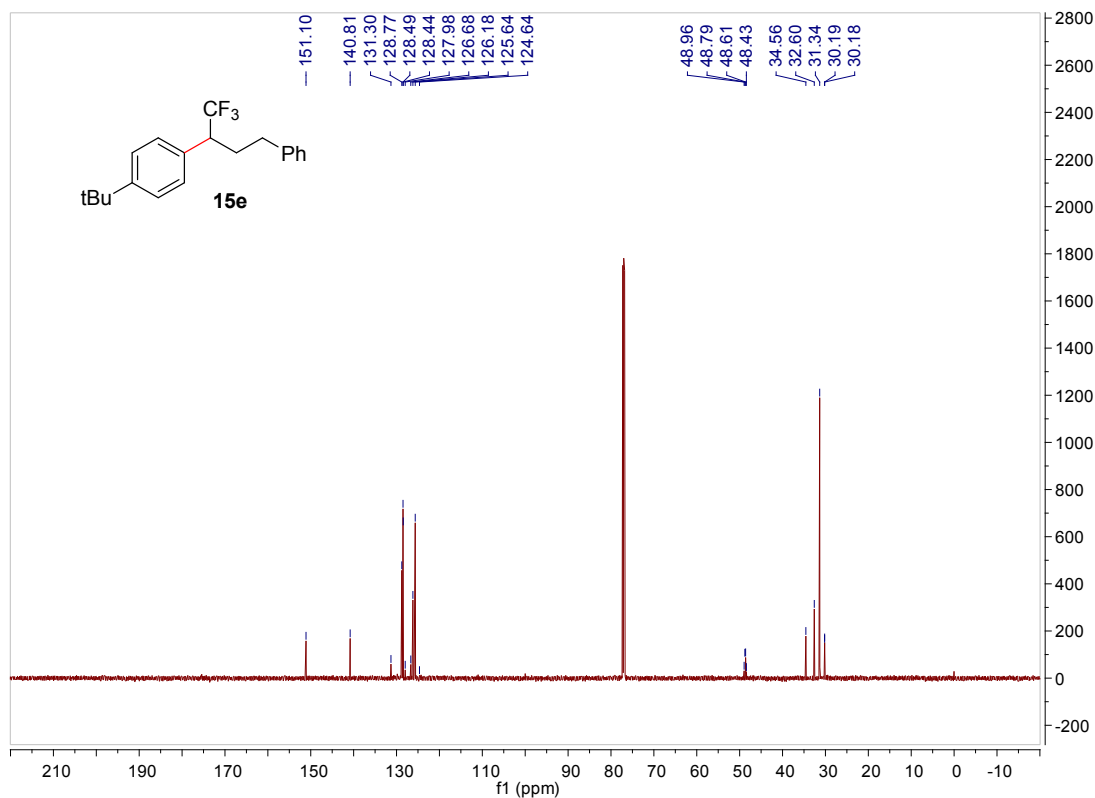


Figure S103. ¹³C NMR of **15e** in CDCl₃

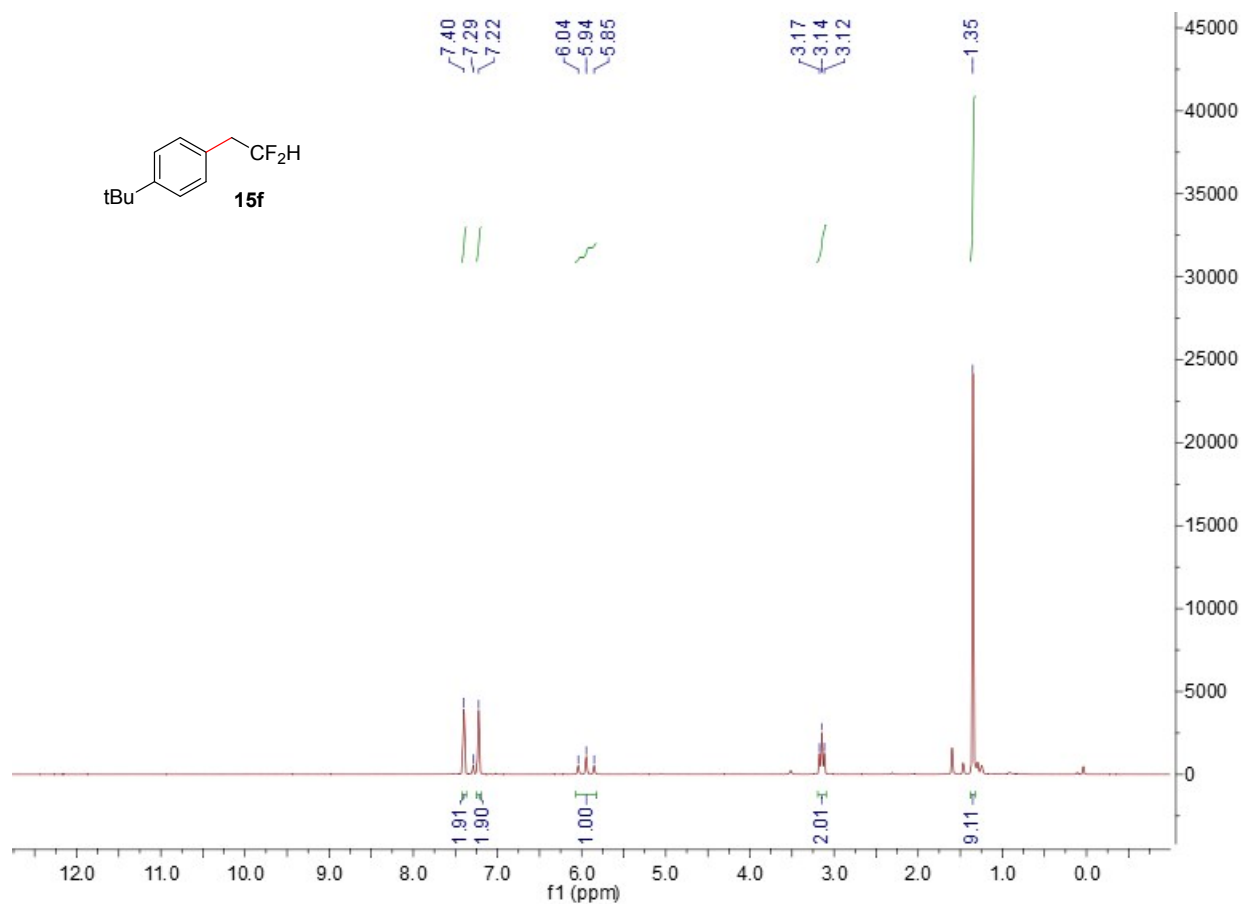


Figure S104. ¹H NMR of **15f** in CDCl₃

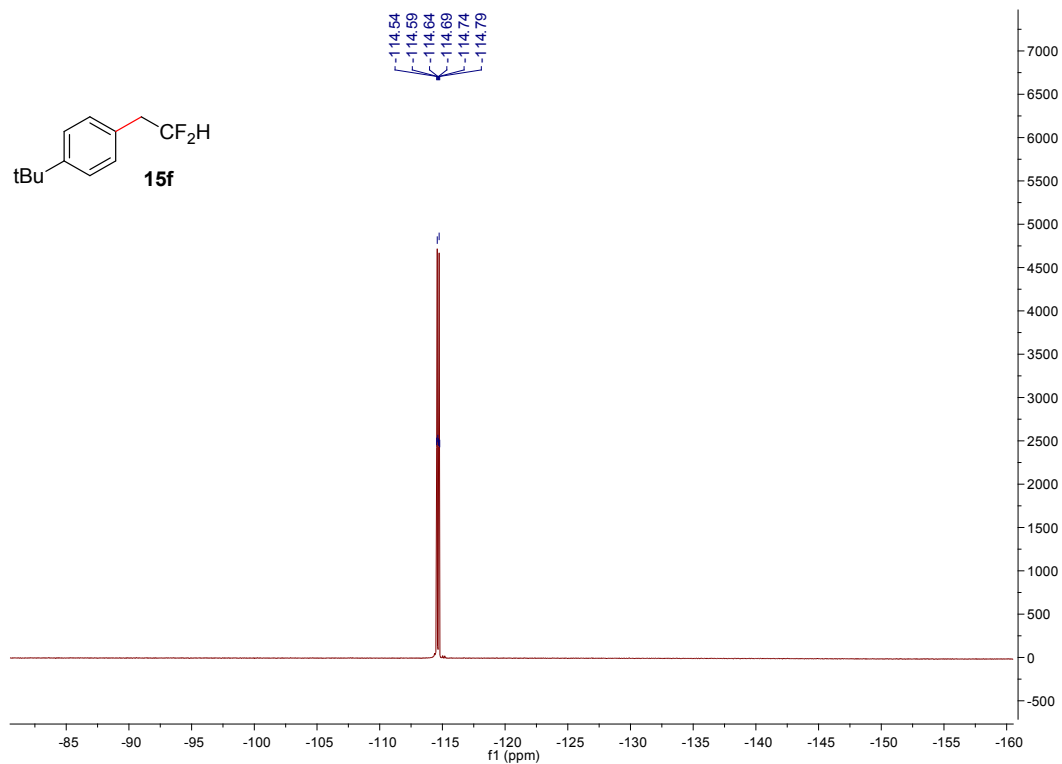


Figure S105. ^{19}F NMR of **15f** in CDCl_3

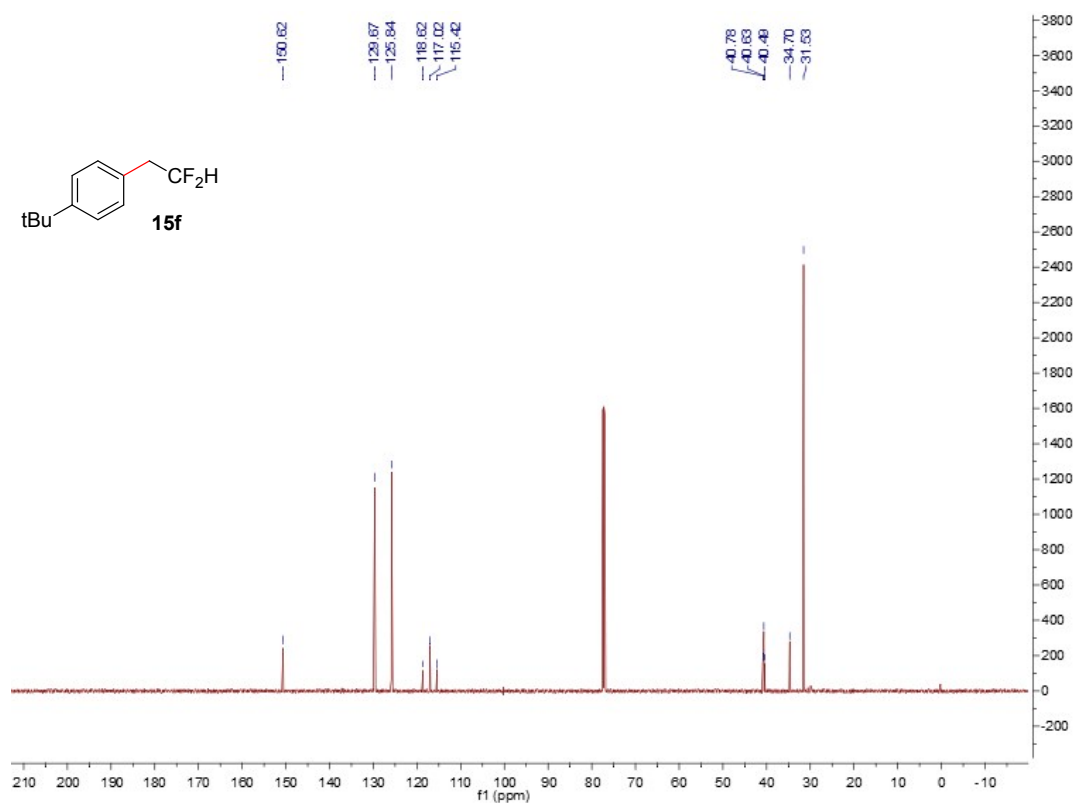


Figure S106. ^{13}C NMR of **15f** in CDCl_3

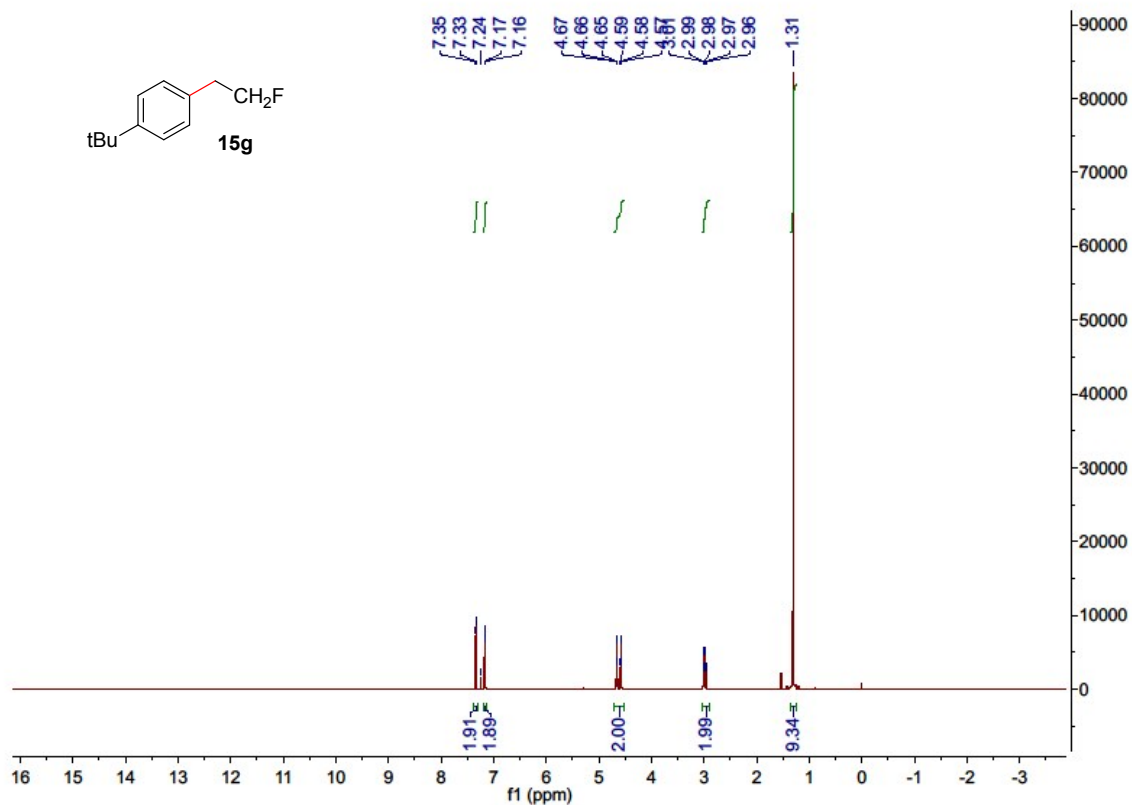


Figure S107. ^1H NMR of **15g** in CDCl_3

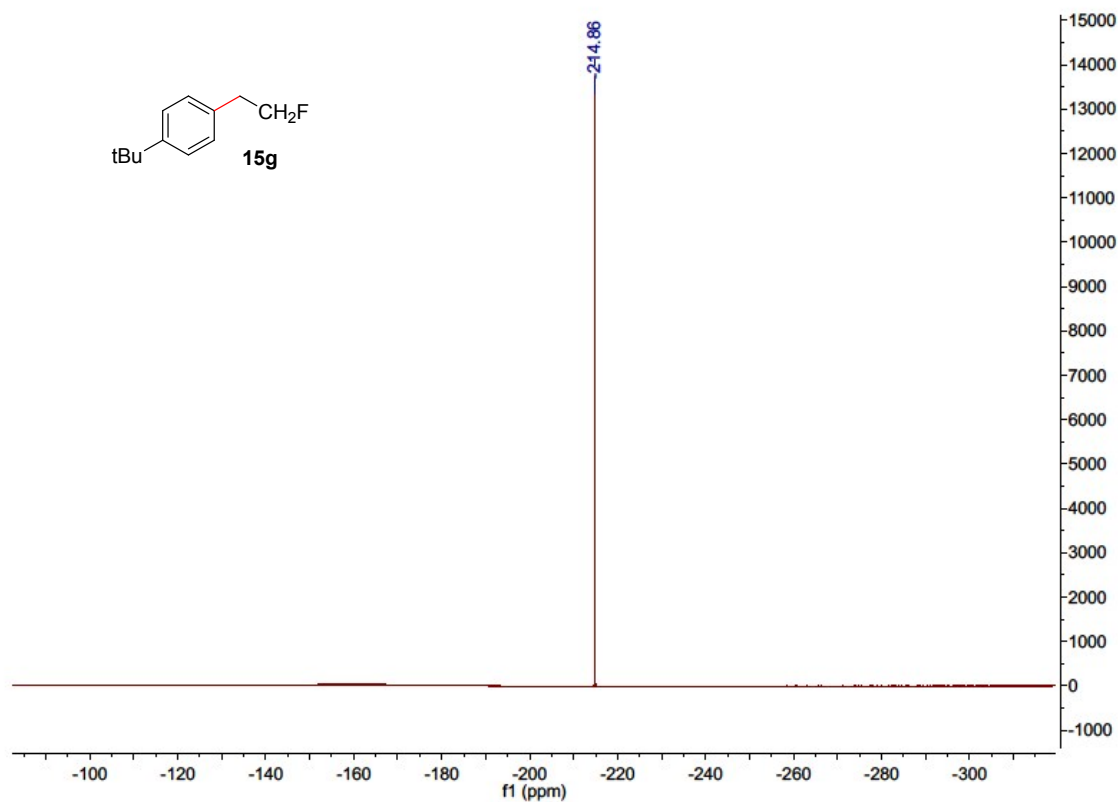


Figure S108. ^{19}F NMR of **15g** in CDCl_3

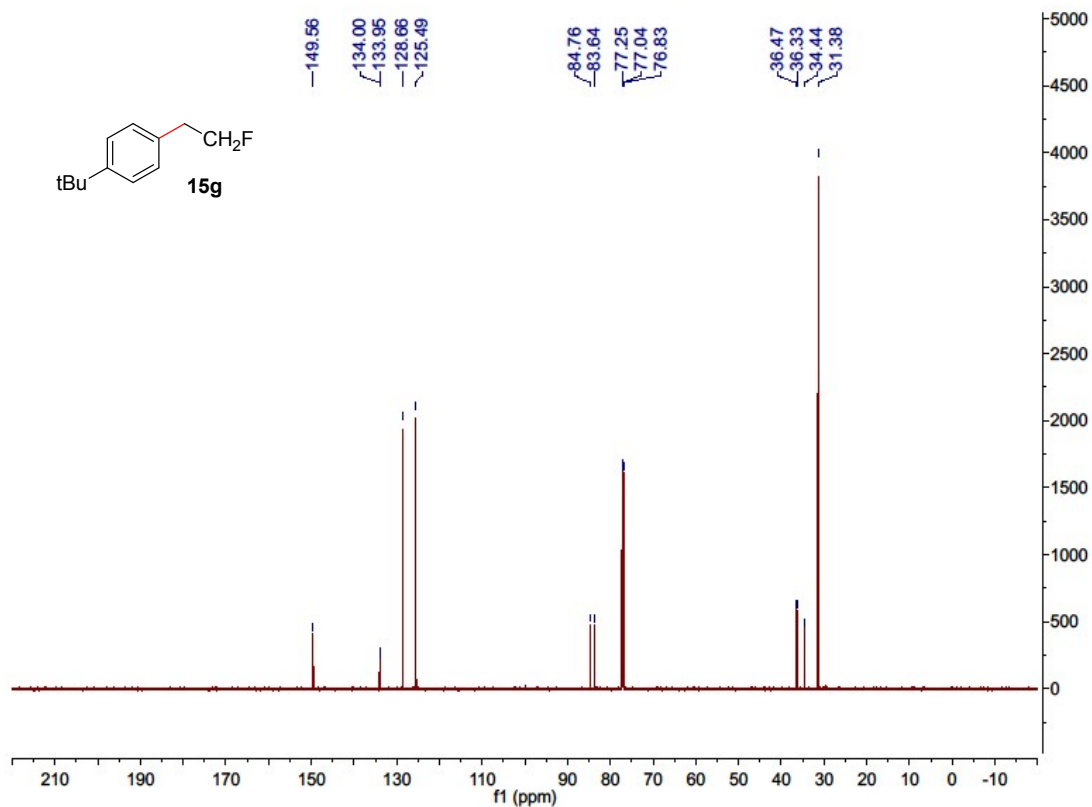


Figure S109. ¹³C NMR of **15g** in CDCl₃

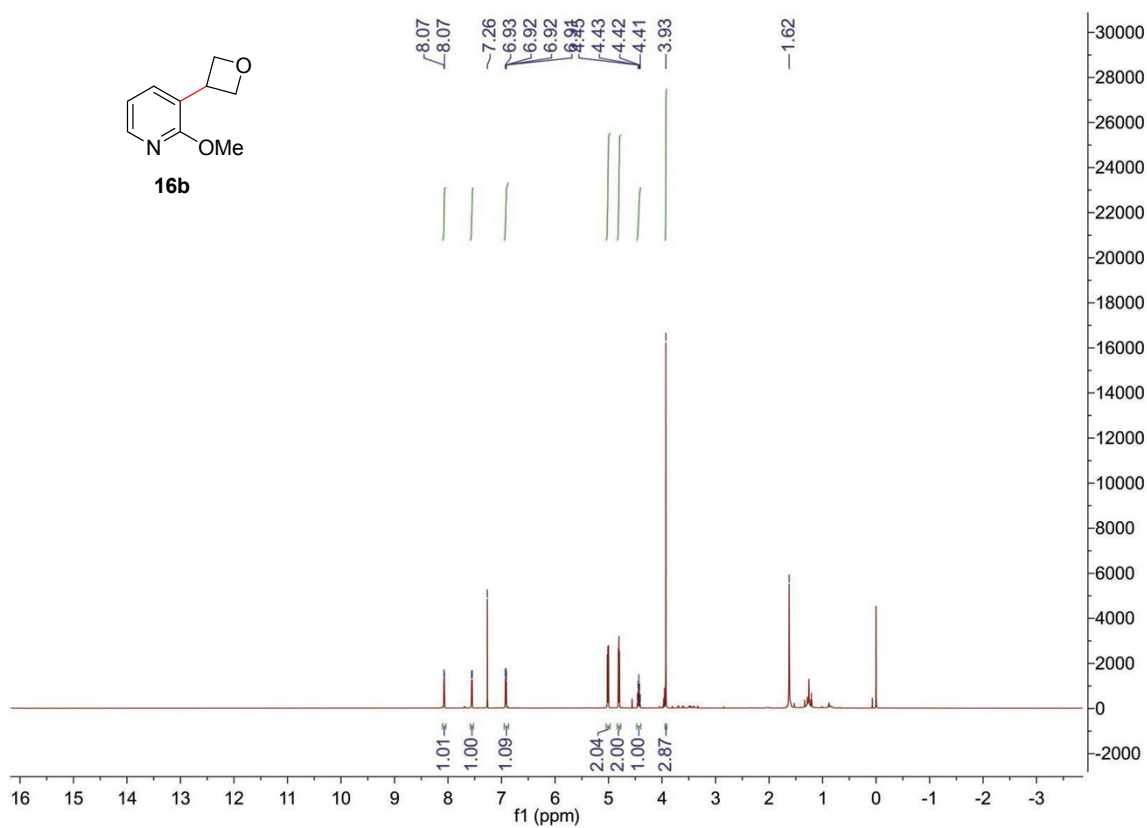


Figure S110. ¹H NMR of **16b** in CDCl₃

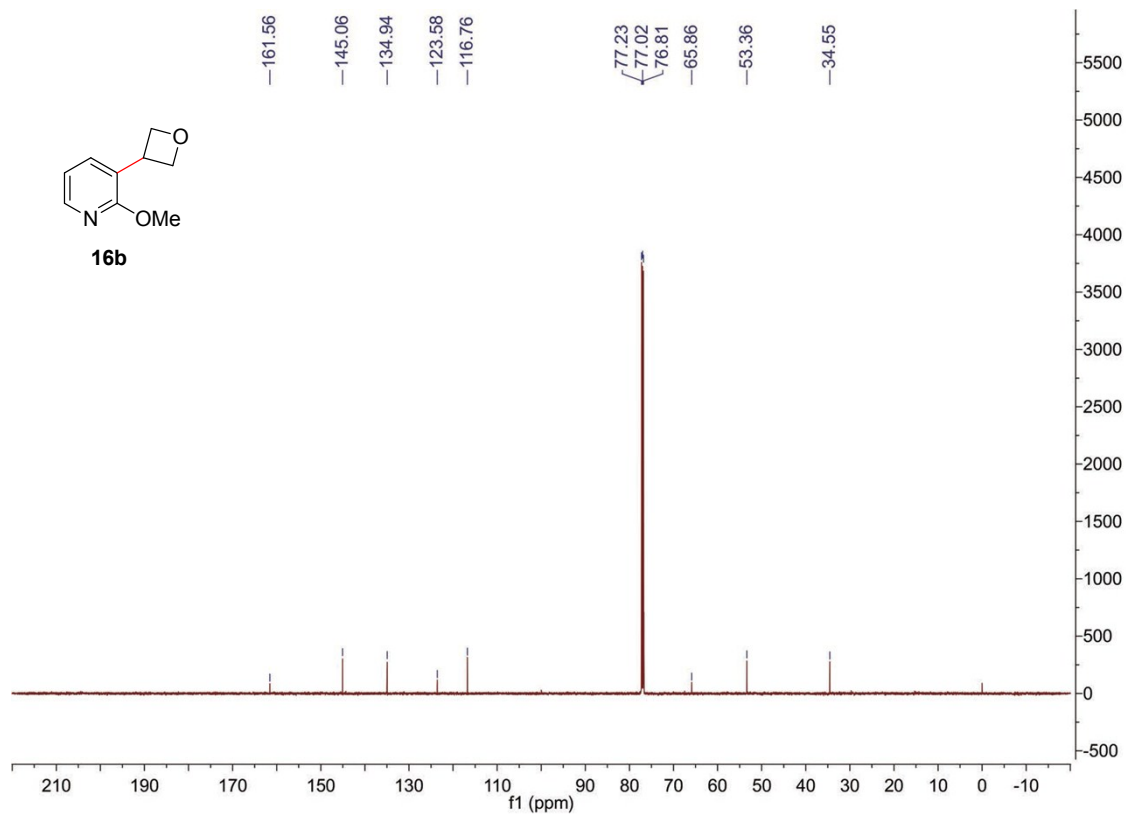


Figure S111. ¹³C NMR of **16b** in CDCl₃

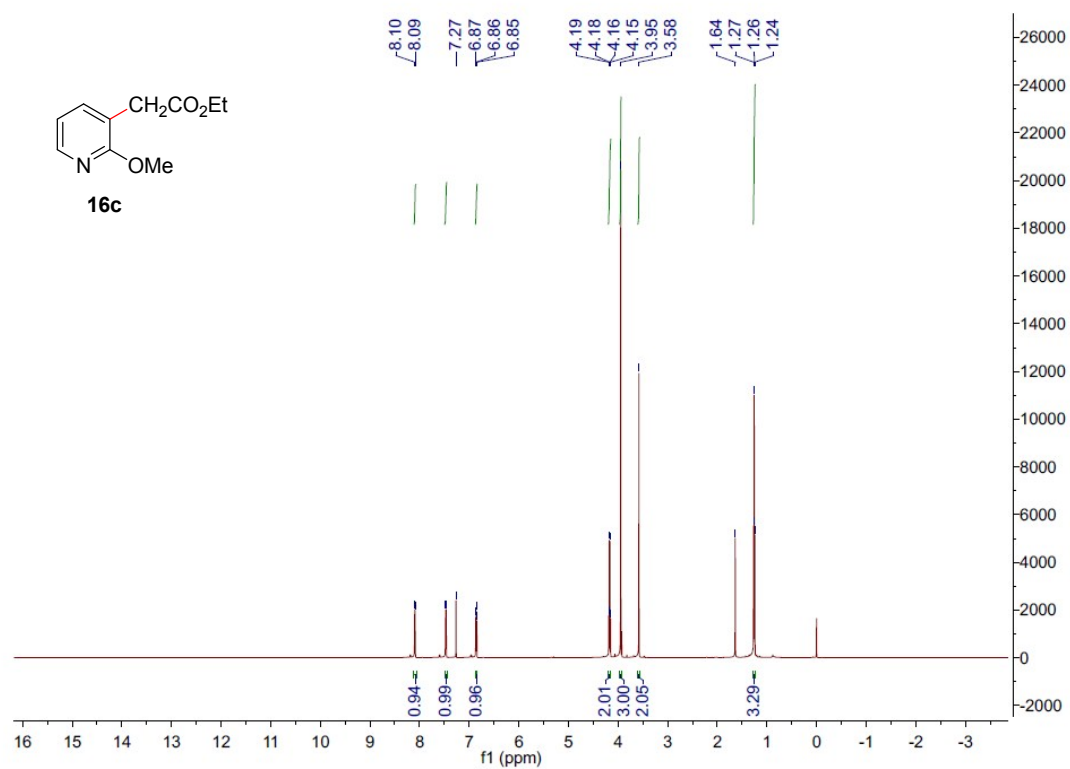


Figure S112. ¹H NMR of **16c** in CDCl₃

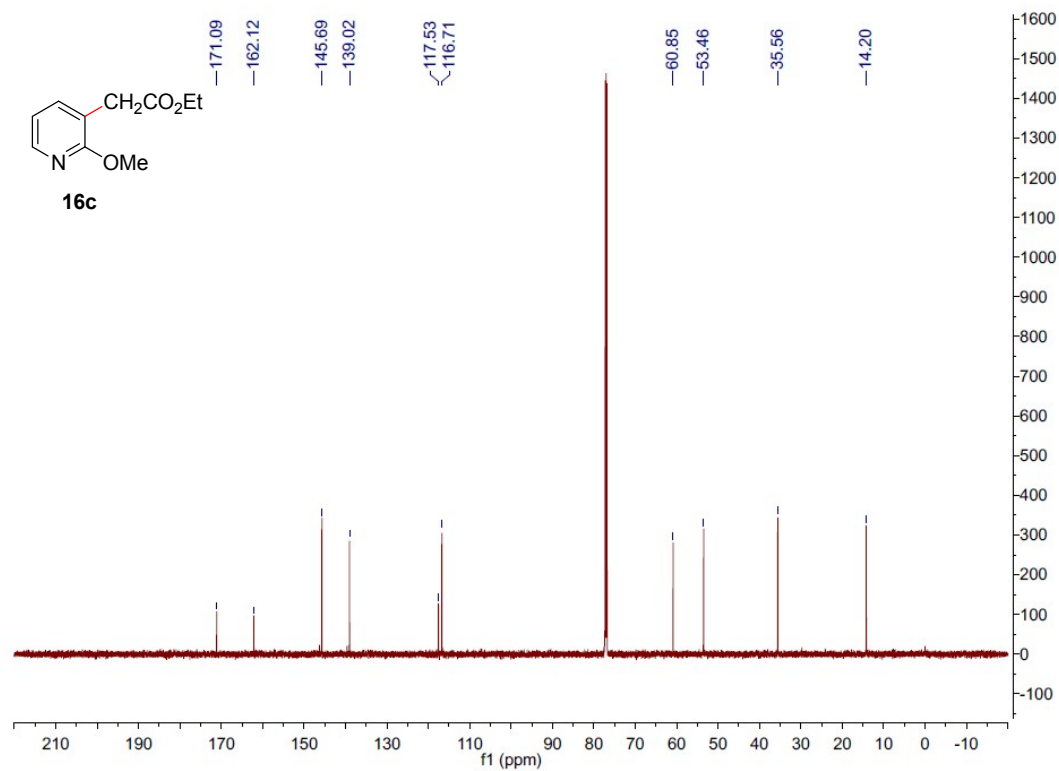


Figure S113. ¹³C NMR of **16c** in CDCl₃

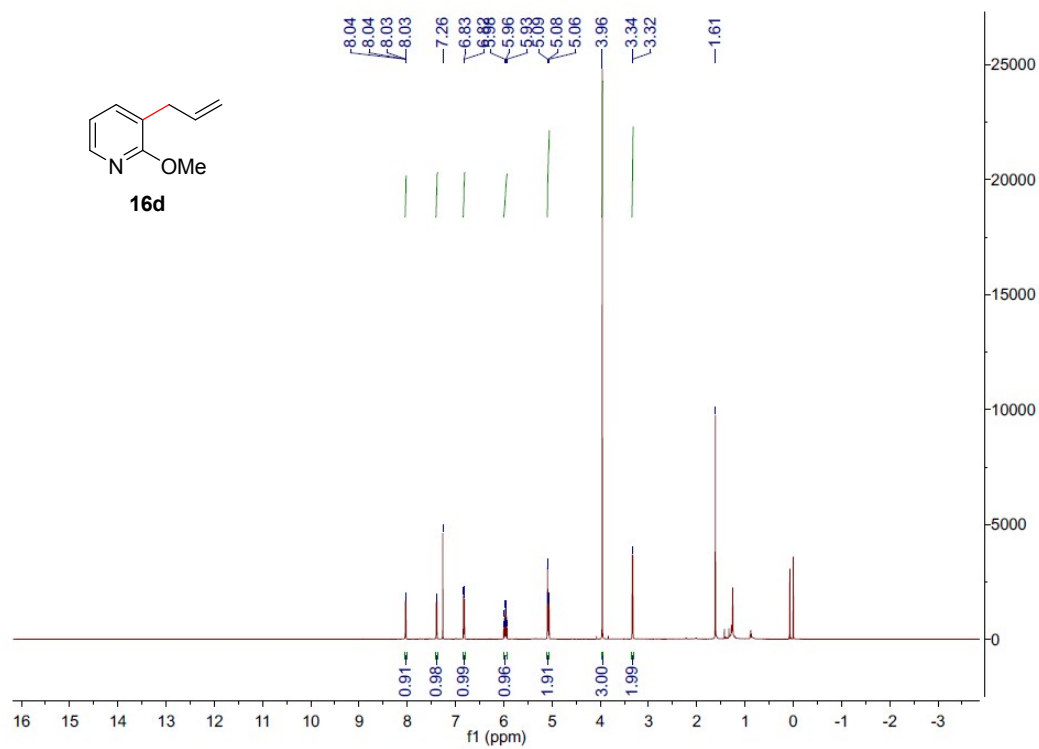


Figure S114. ¹H NMR of **16d** in CDCl₃

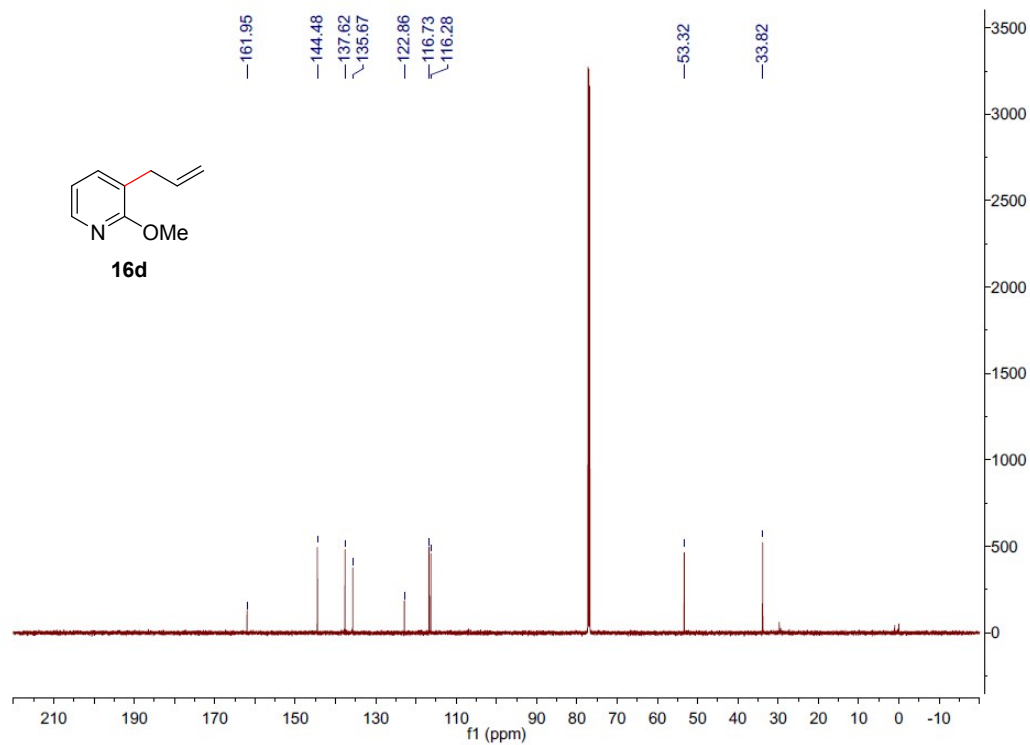


Figure S115. ¹³C NMR of 16d in CDCl₃

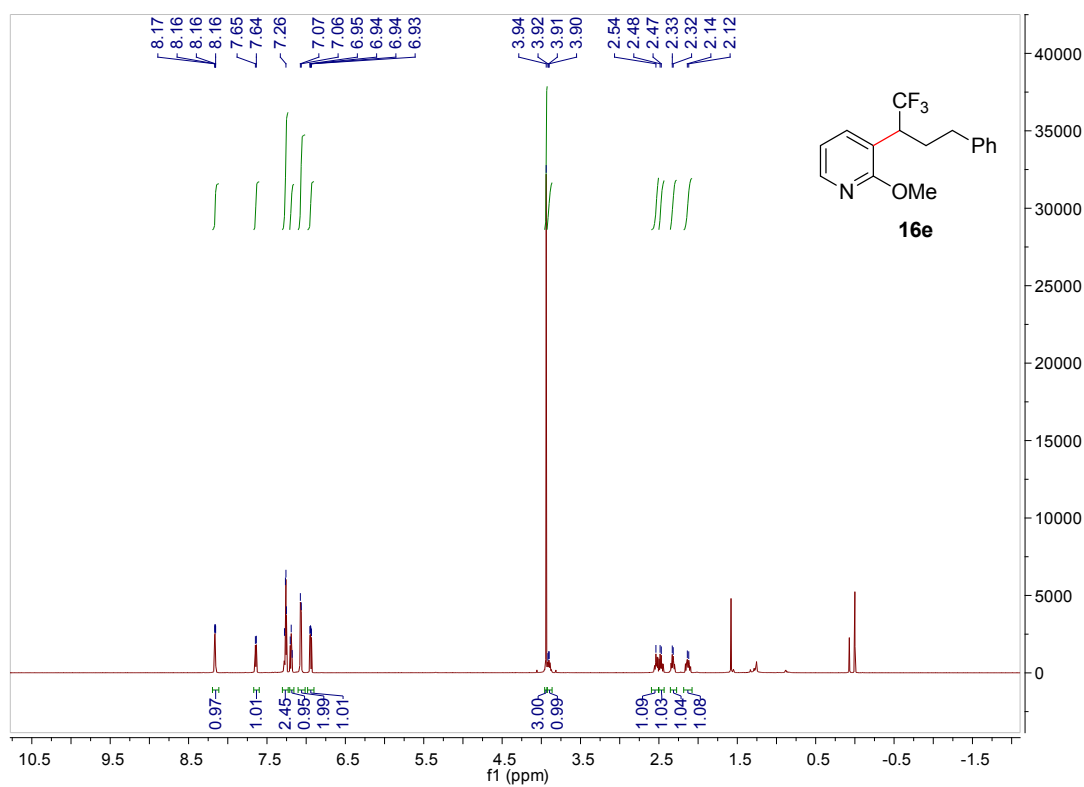


Figure S116. ¹H NMR of 16e in CDCl₃

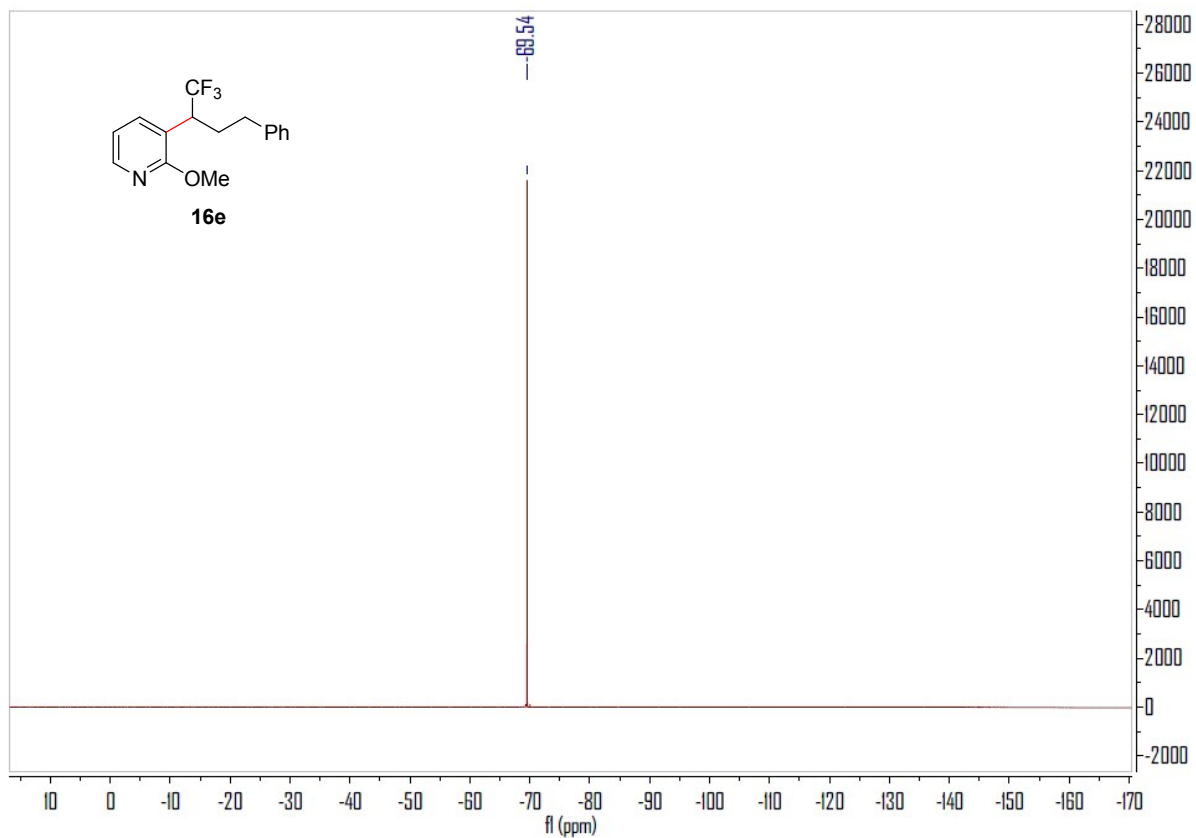


Figure S117. ¹⁹F NMR of **16e** in CDCl₃

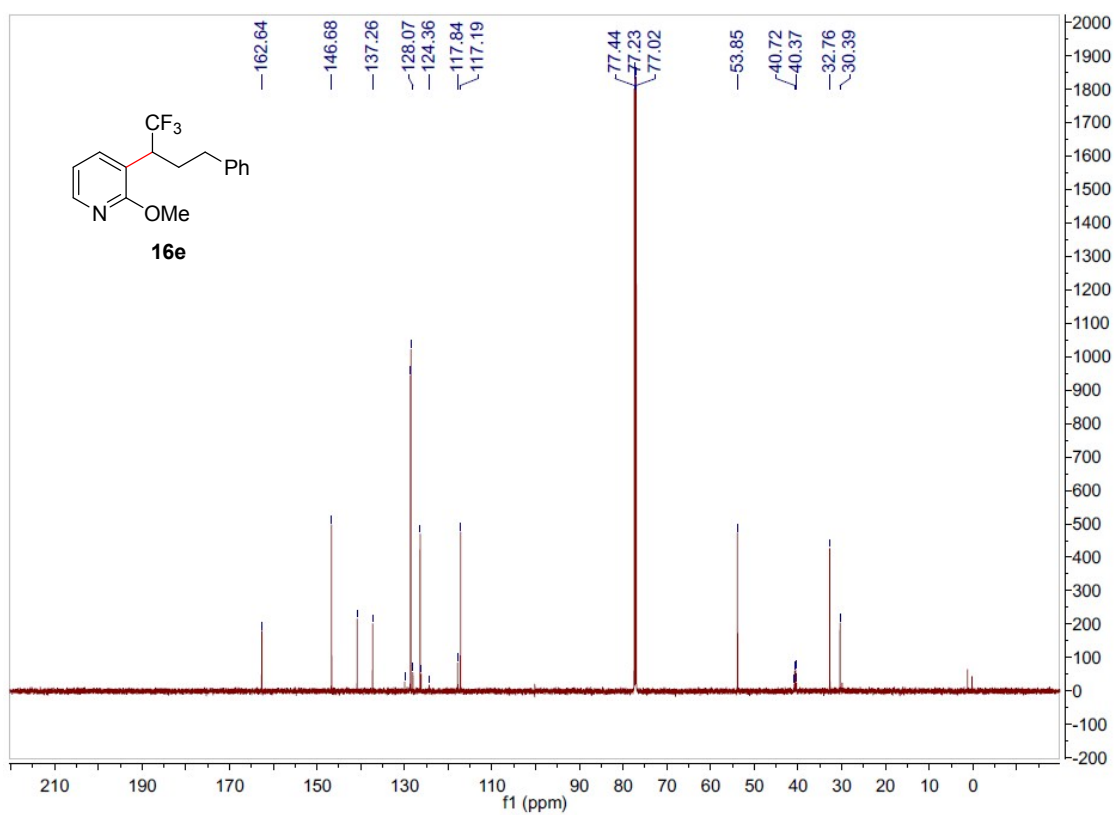


Figure S118. ¹³C NMR of **16e** in CDCl₃

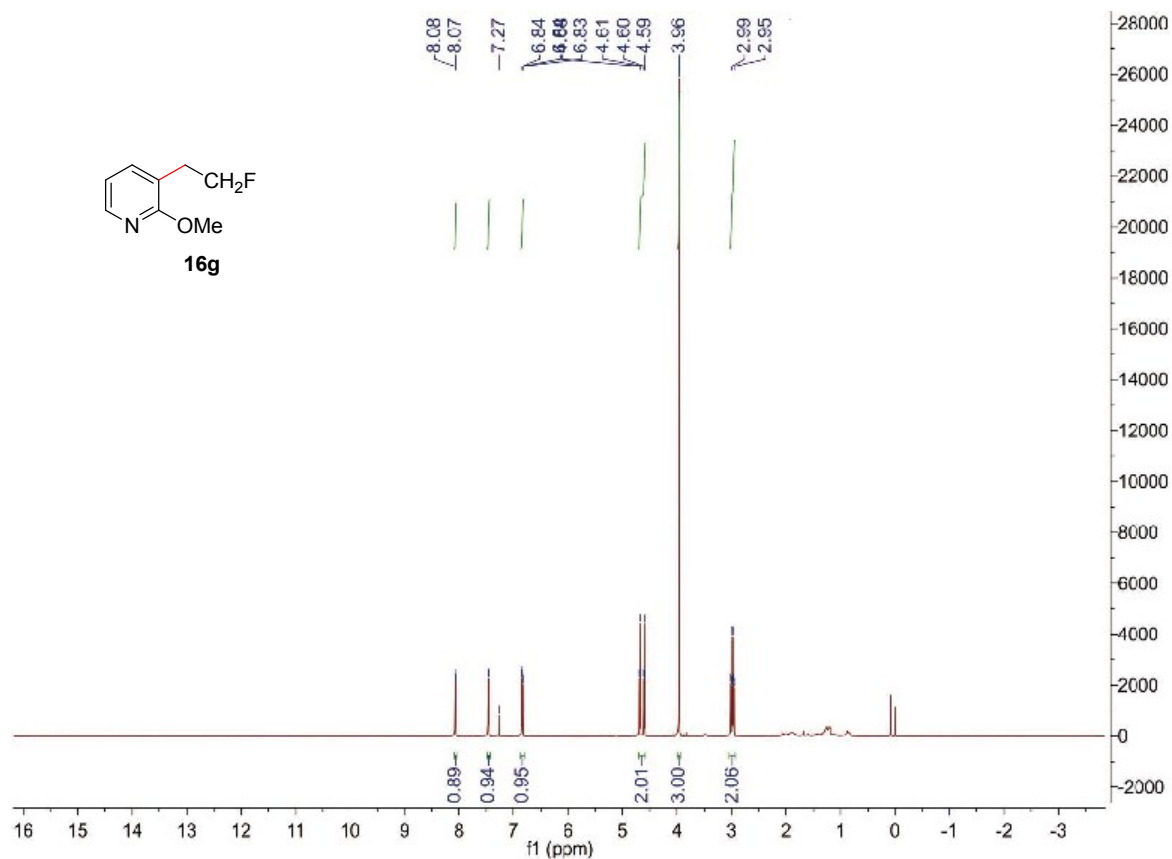


Figure S119. ^1H NMR of **16g** in CDCl_3

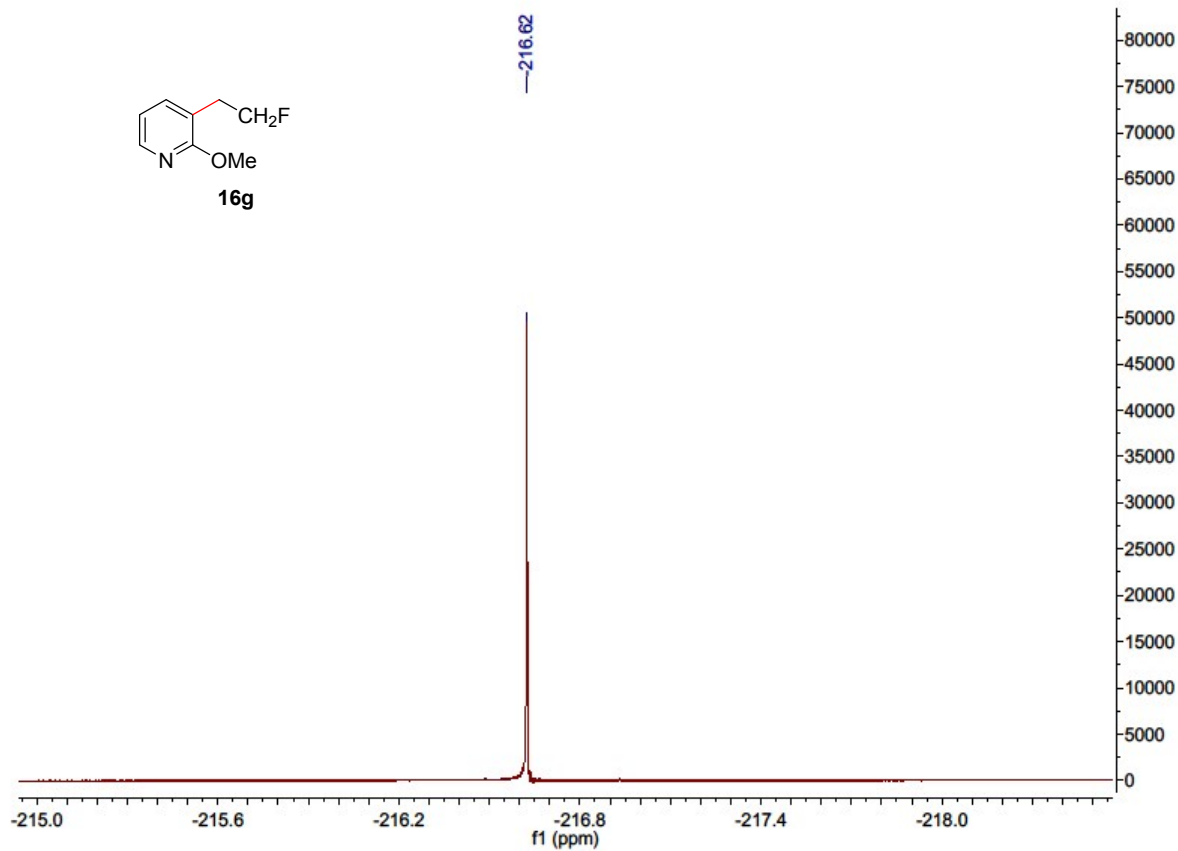


Figure S120. ^{19}F NMR of **16g** in CDCl_3

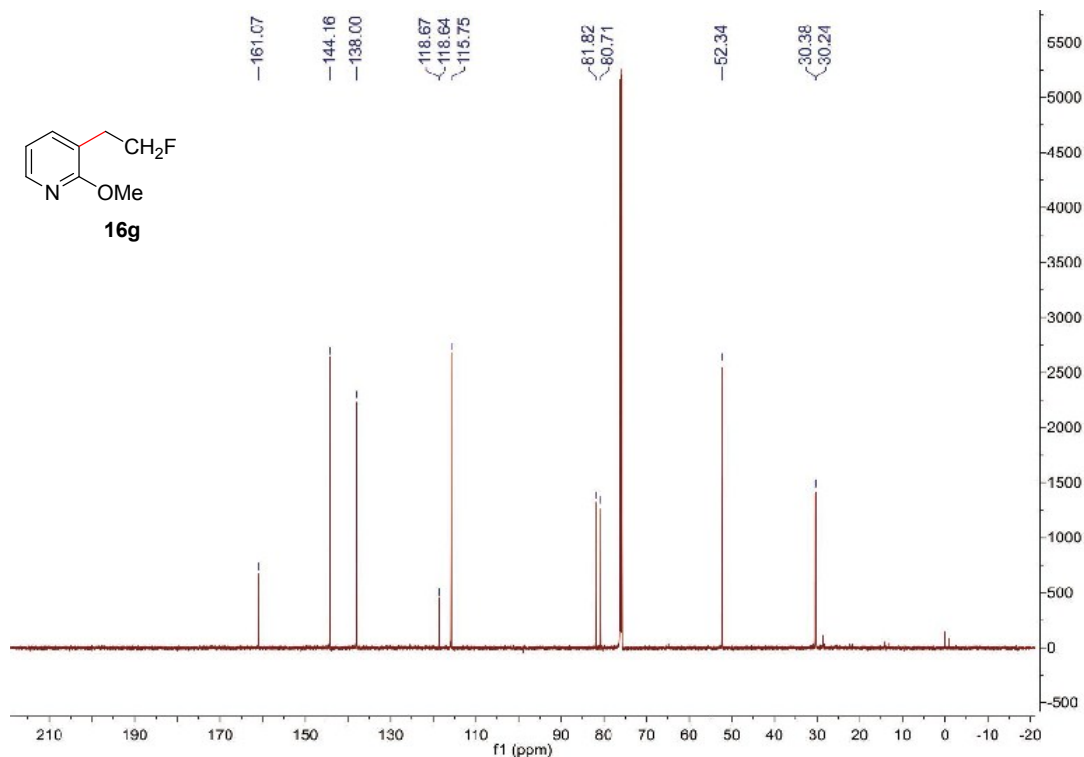
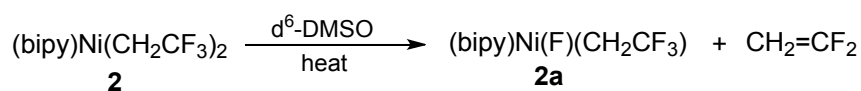


Figure S121. ^{13}C NMR of **16g** in CDCl_3

VI. Control experiments for mechanistic studies

(a) Control experiments for identifying the roles of trifluoroethyl ligands in precatalyst **2**

i) Continuous NMR monitoring of $(\text{bipy})\text{Ni}(\text{CH}_2\text{CF}_3)_2$ upon heating



The precatalyst $(\text{bipy})\text{Ni}(\text{CH}_2\text{CF}_3)_2$ **2** (4.6 mg) was dissolved in $\text{d}^6\text{-DMSO}$ (0.5 mL) with addition of PhCF_3 (3.0 μL) as internal standard which was loaded into a J. Young NMR tube. The solution was heated at the indicated temperature for fixed time and then recorded by a 400M NMR instrument. It was found that the decomposition of **2** started at a slight heating (approximately 40-50 $^\circ\text{C}$) for an evolution of $\text{CH}_2=\text{CF}_2$. When the temperature was elevated further to 60-80 $^\circ\text{C}$, fast extrusion of $\text{CH}_2=\text{CF}_2$ from **2** was observed. Attempts to fingerprint the transient $[(\text{bipy})\text{Ni}(\text{F})(\text{CH}_2\text{CF}_3)]$ **2a** via NMR were unsuccessful which might be attributed to the mentioned redistribution reaction to afford $[(\text{bipy})\text{Ni}(\text{CH}_2\text{CF}_3)_2]$.

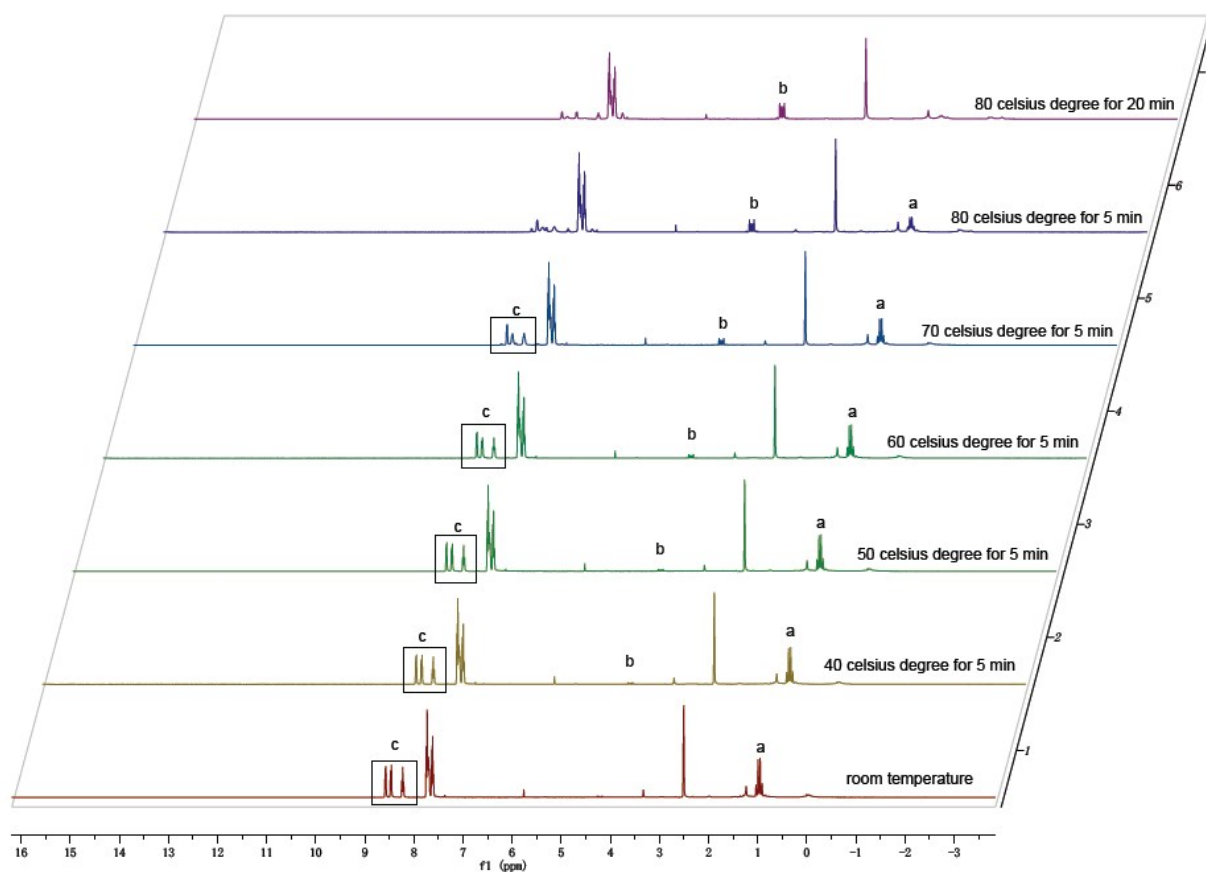


Figure S122. ^1H NMR spectrums of precatalyst **2 in a variable temperature experiment**

^amethylene peaks of precatalyst **2**; ^b $\text{CH}_2=\text{CF}_2$ gas peaks; ^cbipy peaks of precatalyst **2**.

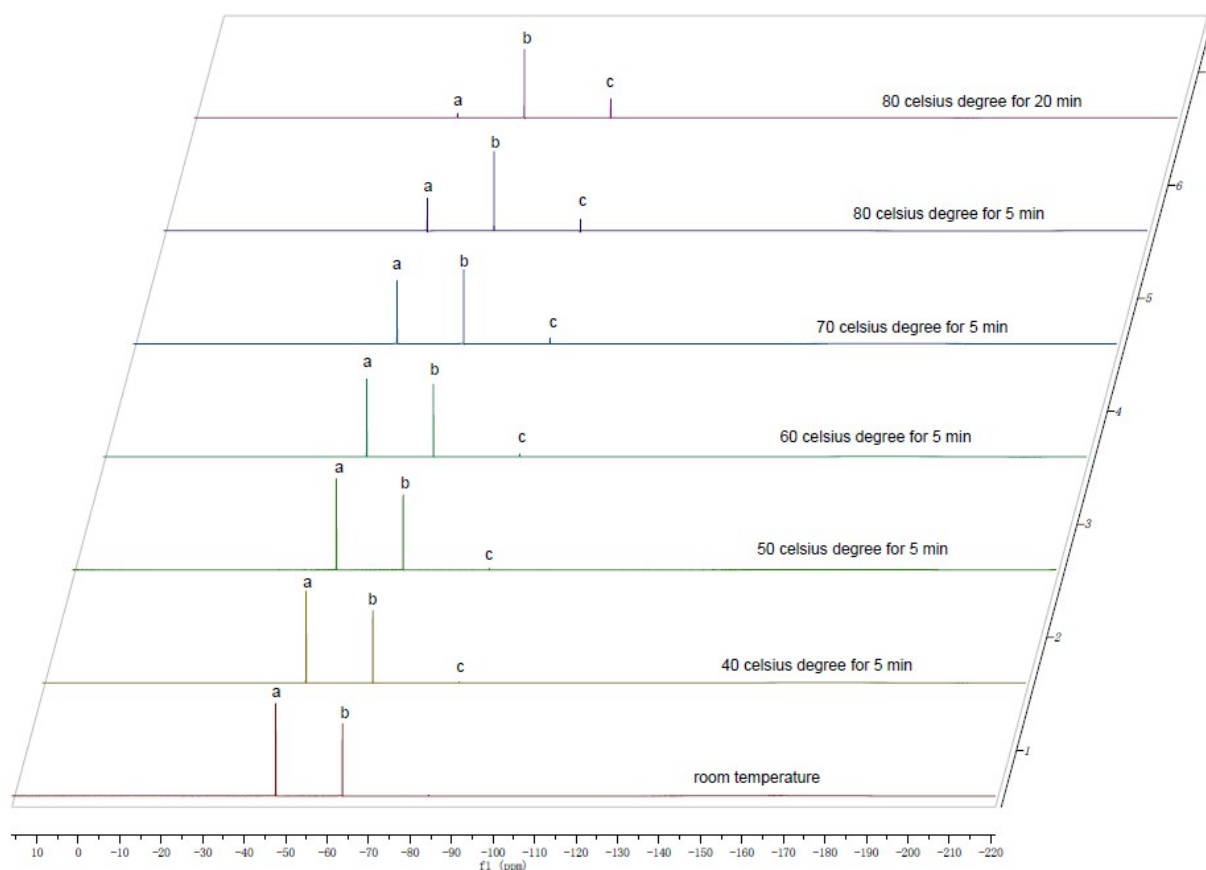
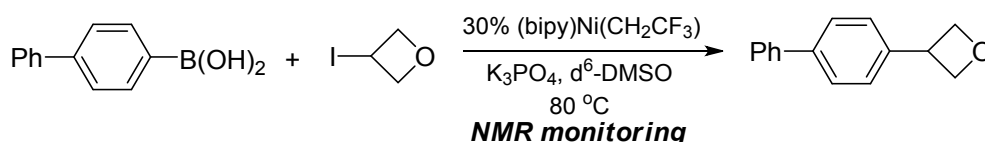


Figure S123. ^{19}F NMR spectrums of precatalyst **2 in a variable temperature experiment**

^atrifluoromethyl peak of precatalyst **2**; ^btrifluorotoluene (internal standard); ^c $\text{CH}_2=\text{CF}_2$ gas peaks.

ii) NMR experiments for identifying the role of trifluoroethyl groups bound to nickel



4-biphenylboronic acid (0.075 mmol, 1.5 equiv), K_3PO_4 (0.10 mmol, 3.0 equiv), followed by a solution of 3-iodooxetane (0.05 mmol, 1.0 equiv) and PhCF_3 (0.05 mmol, internal standard for ^{19}F NMR) in the $\text{d}^6\text{-DMSO}$ solvent (0.5 mL) were loaded into a 25 mL of Schlenck tube which was subject to evacuating/flushing with nitrogen gas three times. The precatalyst **2** (30.0 mol%) in the $\text{d}^6\text{-DMSO}$ solvent (0.5 mL) was added dropwise into the reaction system subsequently (*increasing the catalyst loading for clear identification of the reaction initiation*). The Schlenck tube was screw capped and put into a preheated oil bath (80 $^\circ\text{C}$). After stirring for the indicated time, the reaction mixture was cooled to room temperature and recorded by ^1H and ^{19}F NMR. The result indicated the gradual consumption of 3-iodooxetane, the formation of product **9b** as well as the extrusion of $\text{CH}_2=\text{CF}_2$ from precatalyst **2**.

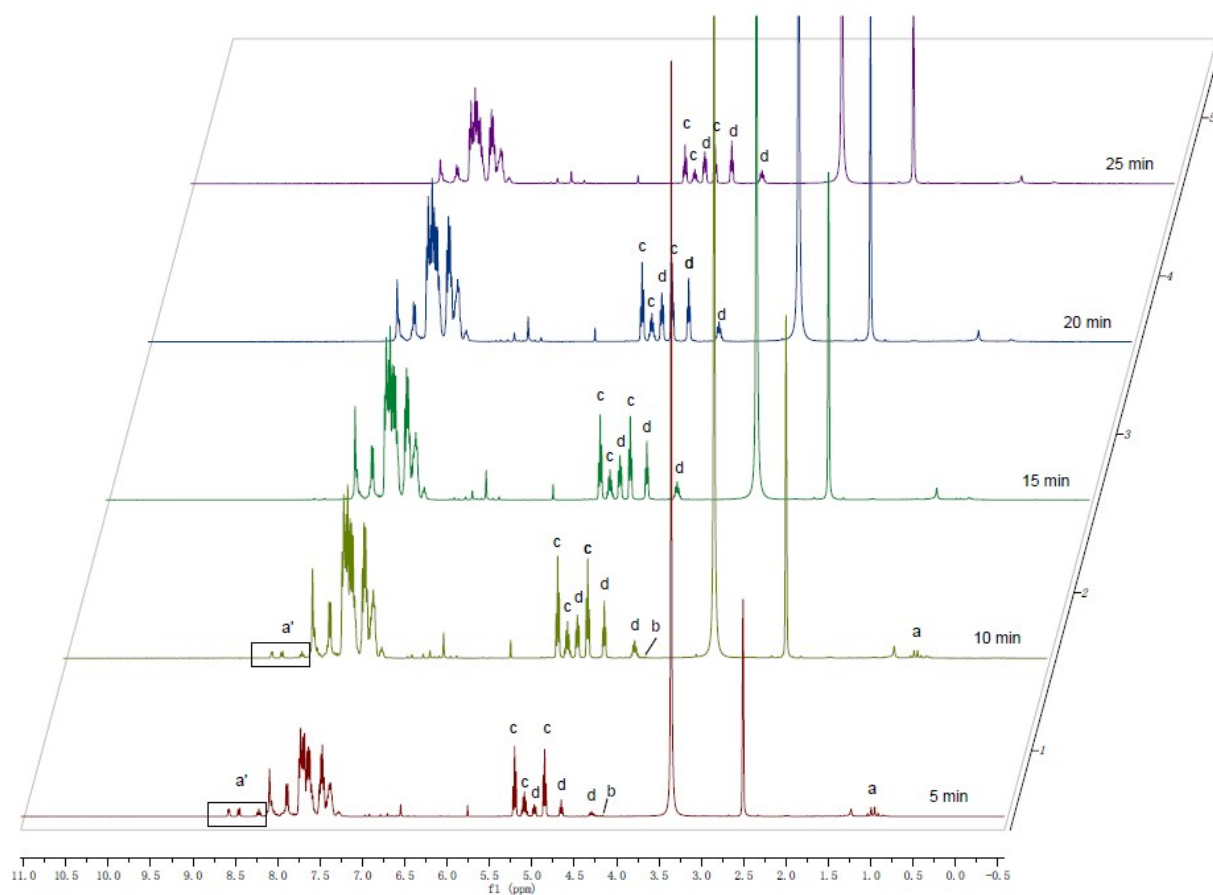


Figure S124. ^1H NMR monitoring of the synthetic reaction of product 14b

^amethylene peaks of precatalyst **2**; ^{a'}bipy peaks of precatalyst **2**; ^b $\text{CH}_2=\text{CF}_2$ gas peaks; ^c3-iodooxetane; ^dproduct **14b**

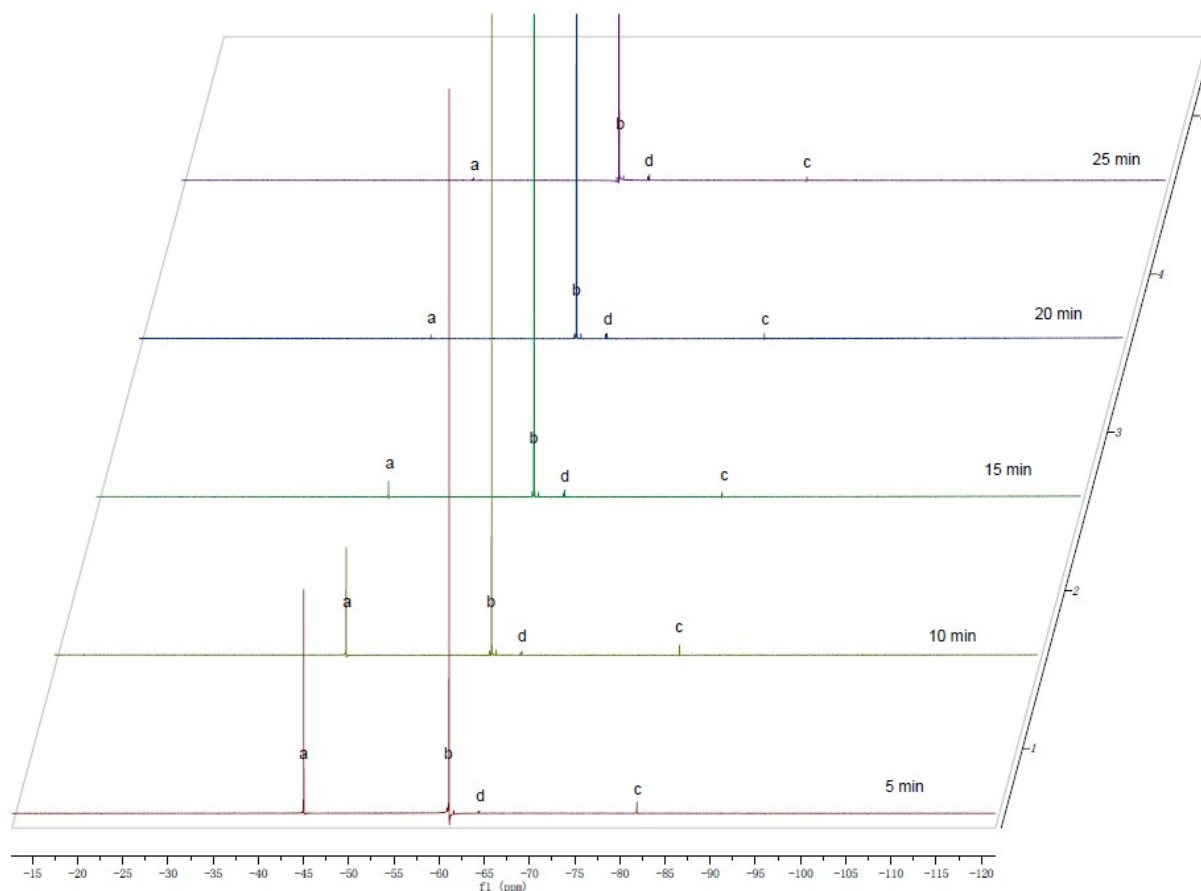
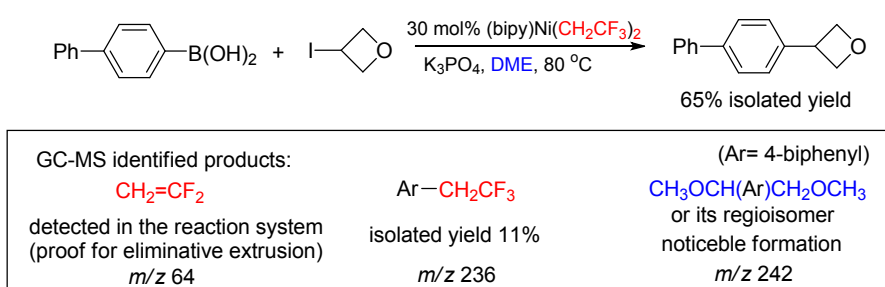


Figure S125. ^{19}F NMR monitoring of the synthetic reaction of product 14b

^atrifluoromethyl peak of precatalyst **2**; ^btrifluorotoluene (internal standard); ^c $\text{CH}_2=\text{CF}_2$ gas peaks; ^d $\text{Ar}-\text{CH}_2\text{CF}_3$ **7a** (produced from the retained CF_3CH_2 group during the initiation step of catalytic cycle).

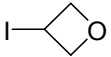
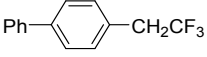
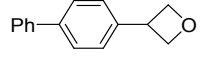
(iii) GC-MS analysis for identifying the role of trifluoroethyl groups bound to nickel



4-biphenylboronic acid (0.3 mmol, 1.5 equiv), K_3PO_4 (0.6 mmol, 3.0 equiv), followed by a solution of 3-iodooxetane (0.2 mmol, 1.0 equiv) in the DME solvent (0.5 mL) were loaded into a 25 mL of Schlenk tube which was subject to evacuating/flushing with nitrogen gas three times. The precatalyst **2** (30.0 mol%) in the DME solvent (0.5 mL) was added dropwise into the reaction system subsequently (increasing the catalyst loading for clear identification of the reaction initiation). The Schlenk tube was screw capped and put into a preheated oil bath (80 °C). After stirring for 24 h, the reaction mixture was cooled to room

temperature and poured into a saturated aqueous ammonium chloride solution (10.0 mL). The aqueous phase was extracted with ether three times (10.0 mL \times 3). The combined organic phase was analyzed by GC-MS with the *p*-xylene internal standard. The organic phase was condensed in *vacuo* to remove solvent, and the residue was purified by flash chromatography on preparative TLC to fingerprint the ArCH₂CF₃ mark **7a** (11% yield) and obtain the desired product **14b** (65% yield).

Table S5. GC-MS analysis for probing roles of trifluoroethyl groups bound to nickel

Retention time/minute	Detected species	Area%
1.62	F ₂ C=CH ₂	Trace ^a
8.56		3.30
12.98	Biphenyl	13.50
13.44	Bipyridine	5.77
13.80		2.38
17.37	CH ₃ OCH(Ar)CH ₂ OCH ₃	1.16
18.18	 (desired product)	12.92

^aF₂C=CH₂ was detected in trace amount in GC-MS due to the volatility of its gaseous properties. It could be observed clearly in ¹⁹F NMR (Figure S104).

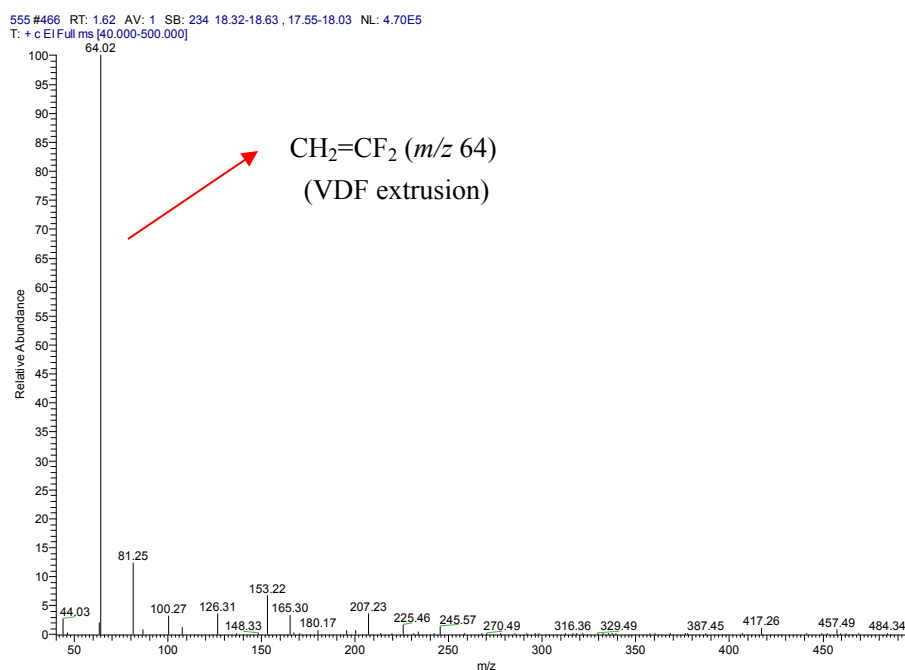


Figure S126. Detection of CH₂=CF₂ by GC-MS

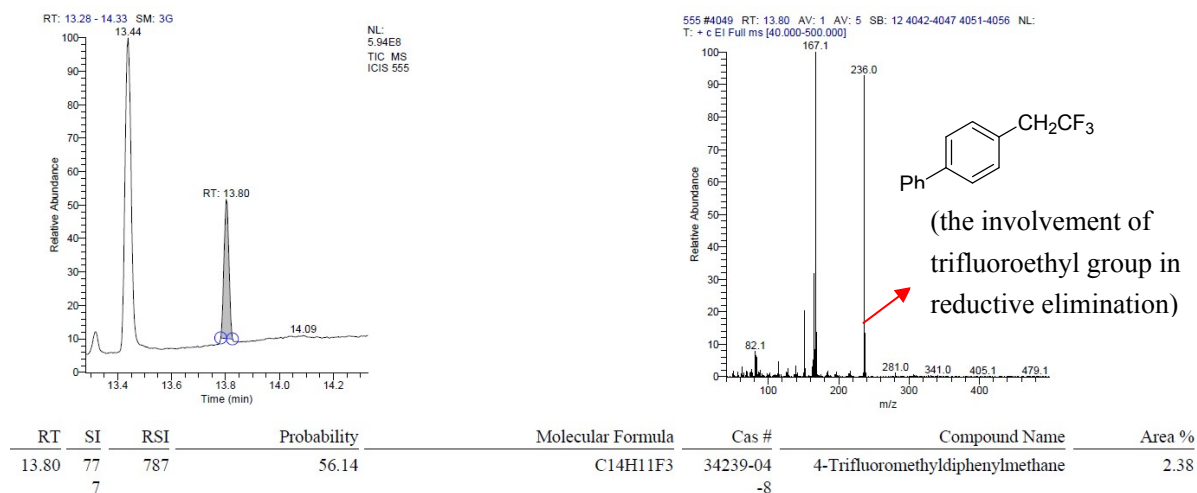


Figure S127. Detection of the Ar-CH₂CF₃ mark by GC-MS

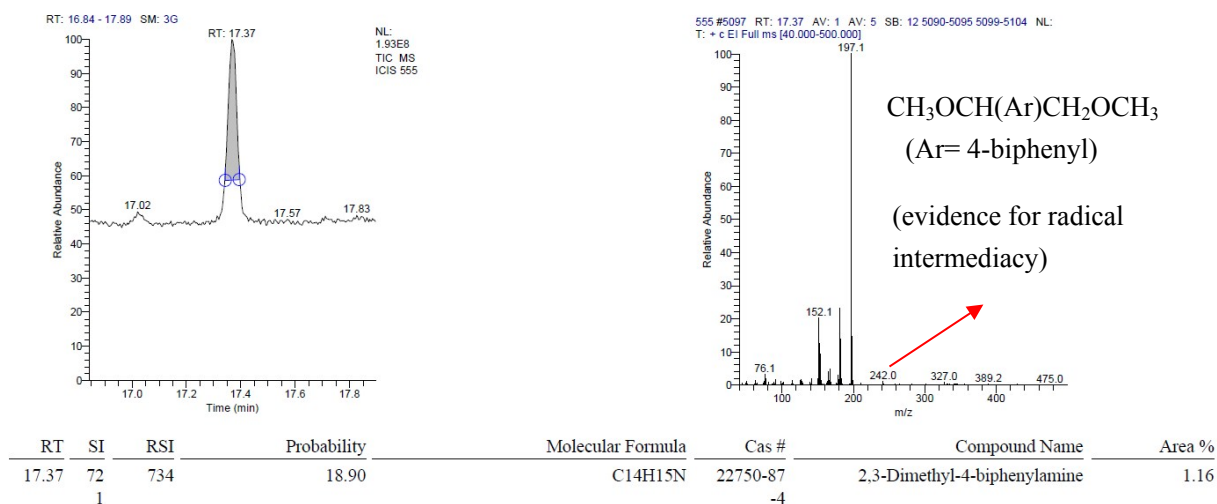


Figure S128. Detection of CH₃OCH(Ar)CH₂OCH₃ by GC-MS

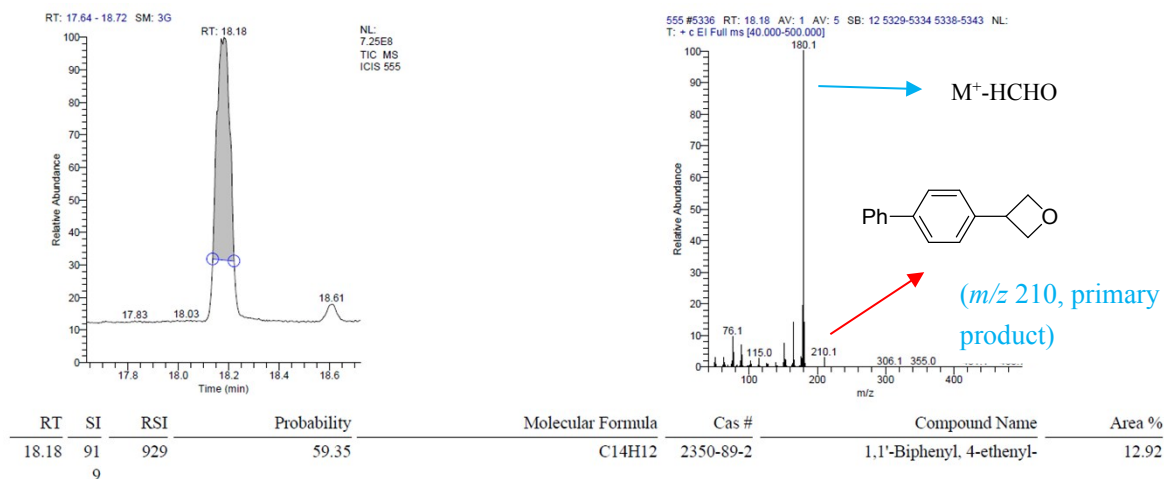
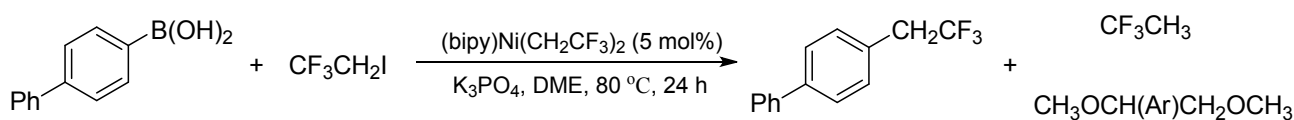


Figure S129. Detection of the product 14b by GC-MS

(b) Evidences for solvent-caged radical reactions

i) Detection of CF_3CH_3 (CF_3CH_2 radical abstracts ethereal α -hydrogen) and $\text{CH}_3\text{OCH}(\text{Ar})\text{CH}_2\text{OCH}_3$ to support the DME solvent-caged reactions

Table S6. GC-MS analysis of the coupling between $\text{CF}_3\text{CH}_2\text{I}$ and 4-biphenylboronic acid



Retention time/minute	Detected species	Area%
1.64	CF_3CH_3	Trace ^a
12.98	Ph-Ph	8.42
13.44	Bipy	5.02
13.81	<chem>c1ccc(cc1)CC(F)(F)F</chem>	17.38
14.26	<chem>c1ccc(cc1)C(F)(F)F</chem>	0.50
16.33	<chem>c1ccc(cc1)I</chem>	6.94
17.37	$\text{CH}_3\text{OCH}(\text{Ar})\text{CH}_2\text{OCH}_3$	9.79

^a CF_3CH_3 was detected in trace amount in GC-MS due to the volatility of its gaseous properties.

My Qual Report

Data File: 549_180328113743 Original Data Path: D:\GCMS\DATA\2018\03\0328
Scans: 7075 Low Mass(m/z): 40
High Mass(m/z): 500

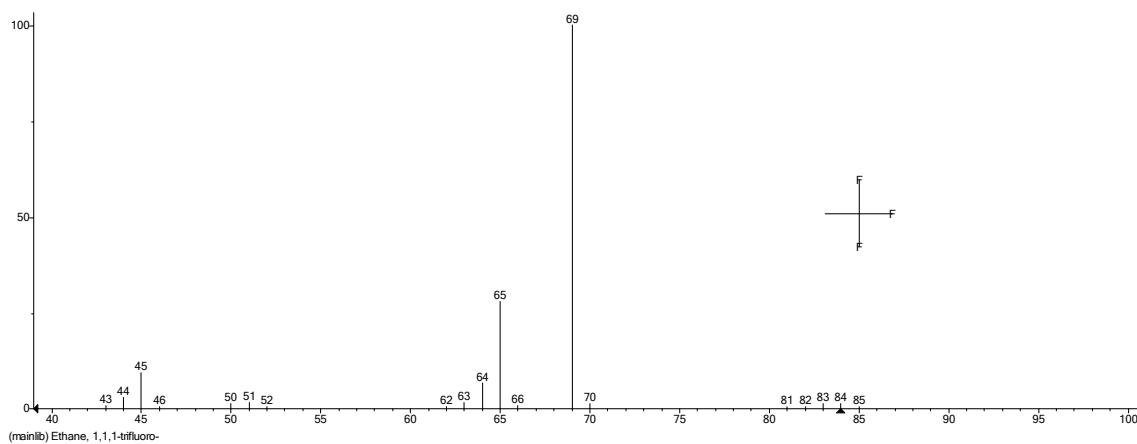
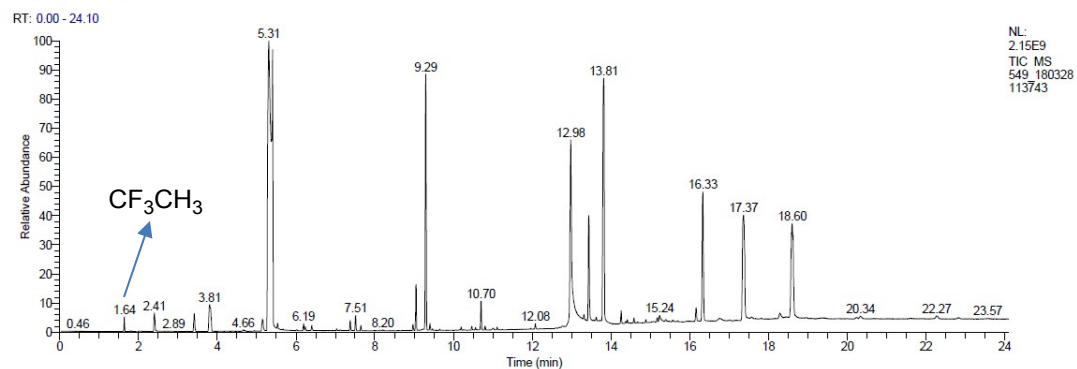


Figure S130. Detection CF₃CH₃ by GC-MS (CF₃CH₂ radical abstracts ethereal α -hydrogen)

ii) Detection of EtOAc (EtOOCCH₂ radical abstracts ethereal α -hydrogen) to support the DME solvent-caged reactions

Table S7. GC-MS analysis of the coupling between BrCH₂CO₂Et and 4-biphenylboronic acid

Retention time/minute	Detected species	Area%
4.65	CH ₃ COOEt	1.76
10.55		6.36
13.22		0.41
14.83		11.78

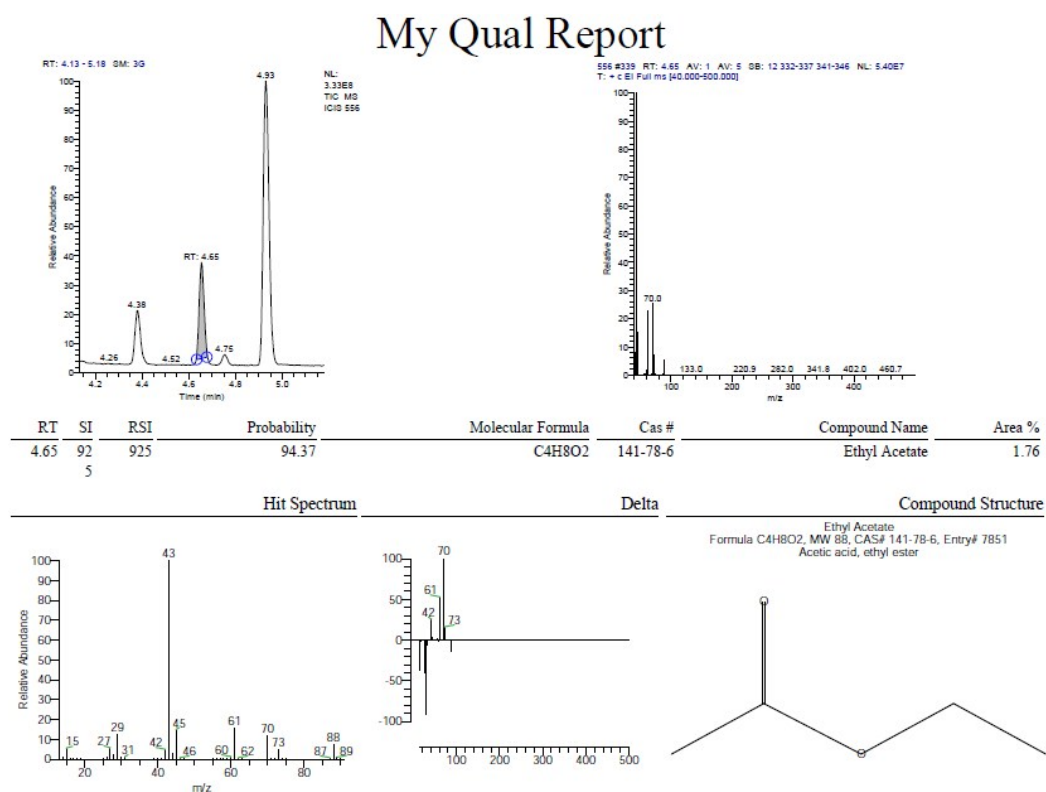


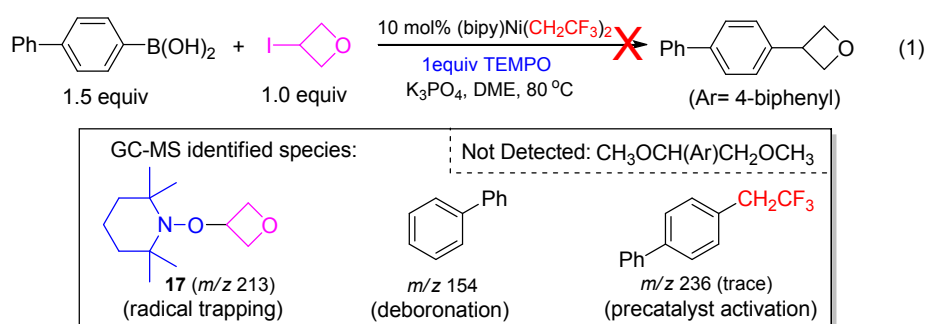
Figure S131. Detection of EtOAc by GC-MS (EtOOCCH₂ radical abstracts ethereal α -hydrogen)

(c) TEMPO radical trapping experiments

Example 1: 4-biphenylboronic acid (0.3 mmol, 1.5 equiv), K_3PO_4 (0.4 mmol, 2.0 equiv), TEMPO (0.2 mmol, 1.0 equiv), followed by a solution of 3-iodooxetane (0.2 mmol, 1.0 equiv) in the DME solvent (0.5 mL) were loaded into a 25 mL of Schlenck tube which was subject to evacuating/flushing with nitrogen gas three times. The precatalyst **2** (10.0 mol%) in the DME solvent (0.5 mL) was added dropwise into the reaction system subsequently.

Table S8. GC-MS analysis of TEMPO radical trapping for the coupling reaction of 4-biphenylboronic acid and 3-iodooxetane

a Radical inhibition test



Retention time/minute	Detected species	Area%
8.29	<i>p</i> -xylene(internal standard)	14.84
8.55	3-iodooxetane	7.20
9.00		0.42
10.28		trace ^a
10.96	TEMPO	12.10
12.96	Biphenyl	16.67
13.79		0.07

^aThis species was detected in trace amount possibly due to its instability.

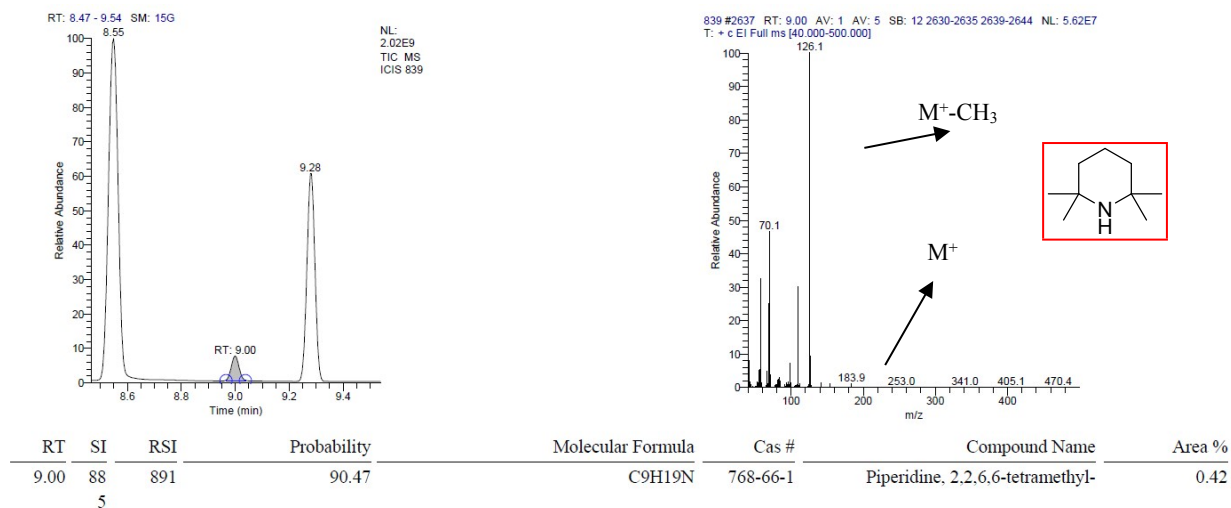


Figure S132. Detection of 2,2,6,6-tetramethylpiperidine by GC-MS

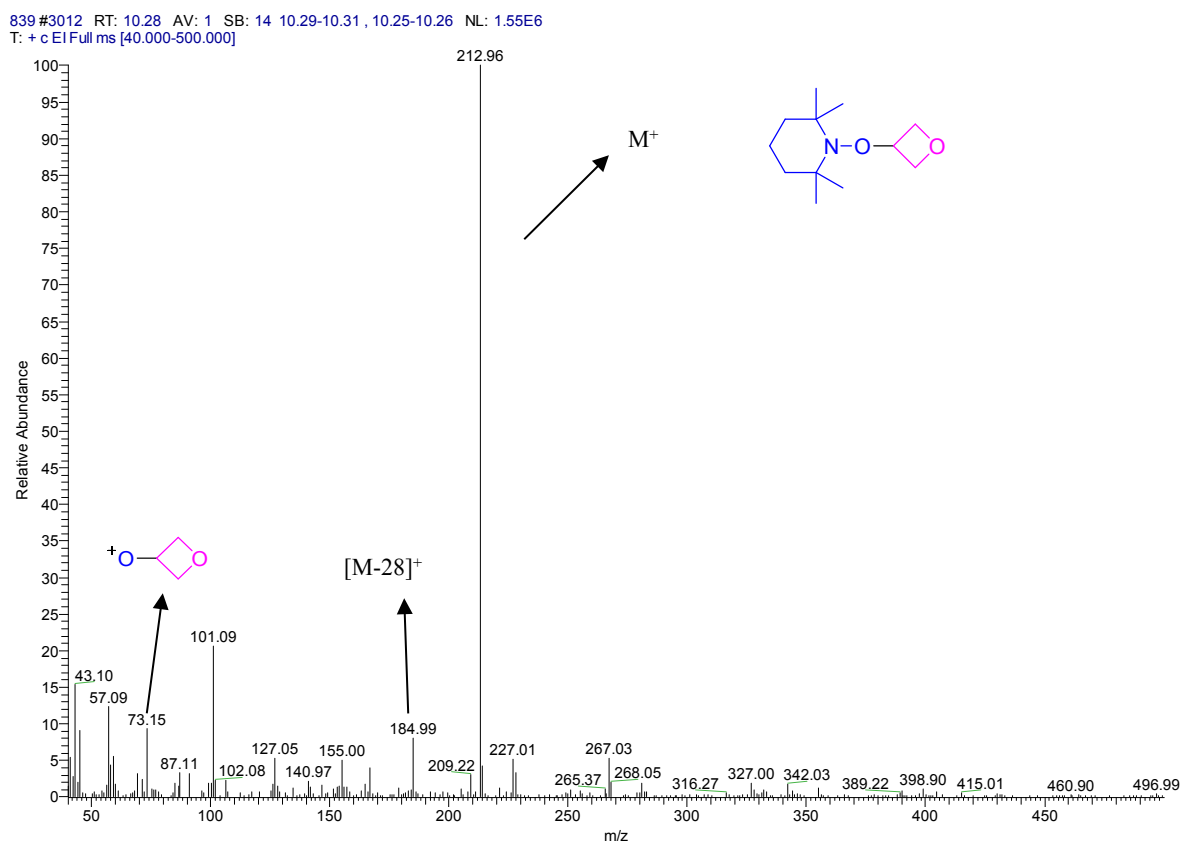


Figure S133. Detection of 2,2,6,6-tetramethyl-1-(oxetan-3-yloxy)piperidine by GC-MS

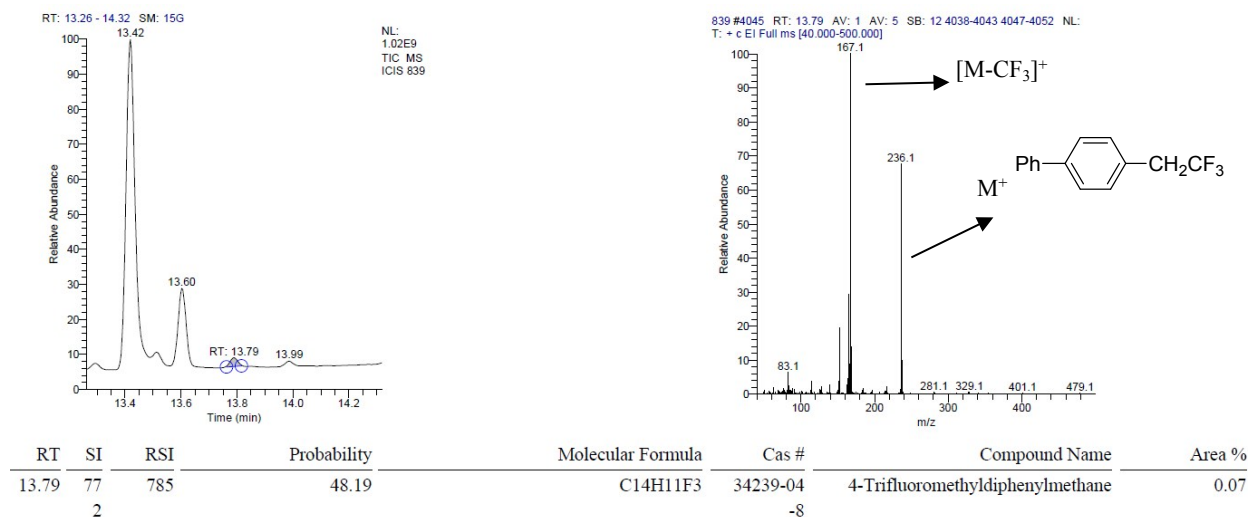
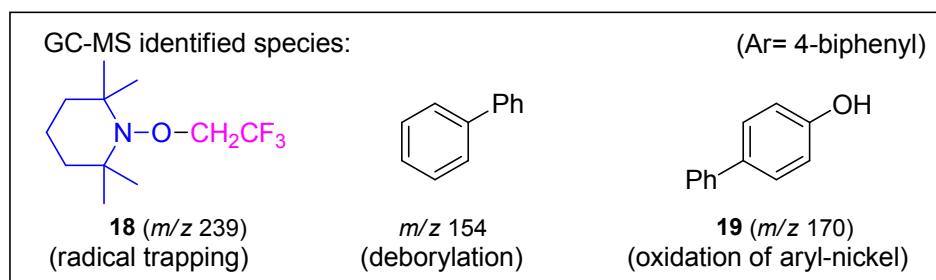
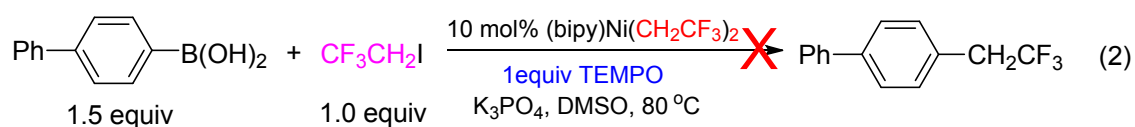


Figure S134. Detection of Ar-CH₂CF₃ by GC-MS (Ar= 4-biphenyl)

Example 2: 4-biphenylboronic acid (0.3 mmol, 1.5 equiv), K₃PO₄ (0.6 mmol, 3.0 equiv), TEMPO (0.2 mmol, 1.0 equiv), followed by a solution of CF₃CH₂I (0.2 mmol, 1.0 equiv) in the DMSO solvent (0.5 mL) were loaded into a 25 mL of Schlenk tube which was subject to evacuating/flushing with nitrogen gas three times. The precatalyst **2** (10.0 mol%) in the DMSO solvent (0.5 mL) was added dropwise into the reaction system subsequently. The Schlenk tube was screw capped and put into a preheated oil bath (80 °C). After stirring for 24 h, the reaction mixture was cooled to room temperature and poured into a saturated aqueous ammonium chloride solution (10.0 mL). The aqueous phase was extracted with ether three times (10.0 mL × 3). The combined organic phase was analyzed by GC-MS with the *p*-xylene internal standard.

Table S9. GC-MS analysis of TEMPO radical trapping for the coupling reaction of 4-biphenylboronic acid and CF₃CH₂I



Retention time/minute	Detected species	Area%
3.79	CF ₃ CH ₂ I	10.92
8.31	<i>p</i> -xylene (internal standard)	20.27
10.41		3.82
10.98	TEMPO	21.93
12.98	Biphenyl	15.14
13.45	Bipyridine	0.90
15.24		1.51

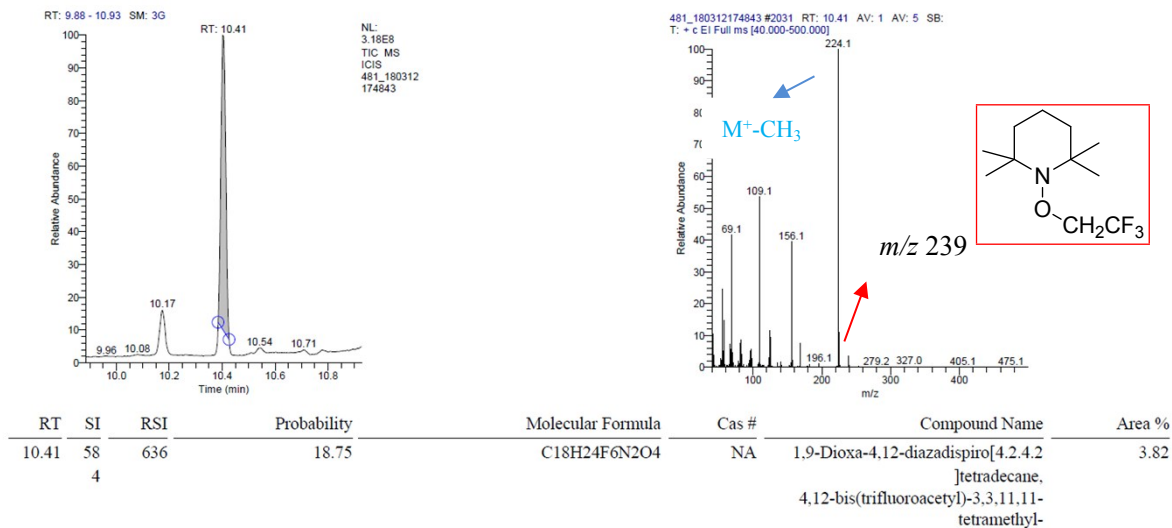


Figure S135. Detection of TEMPO-CH₂CF₃ by GC-MS

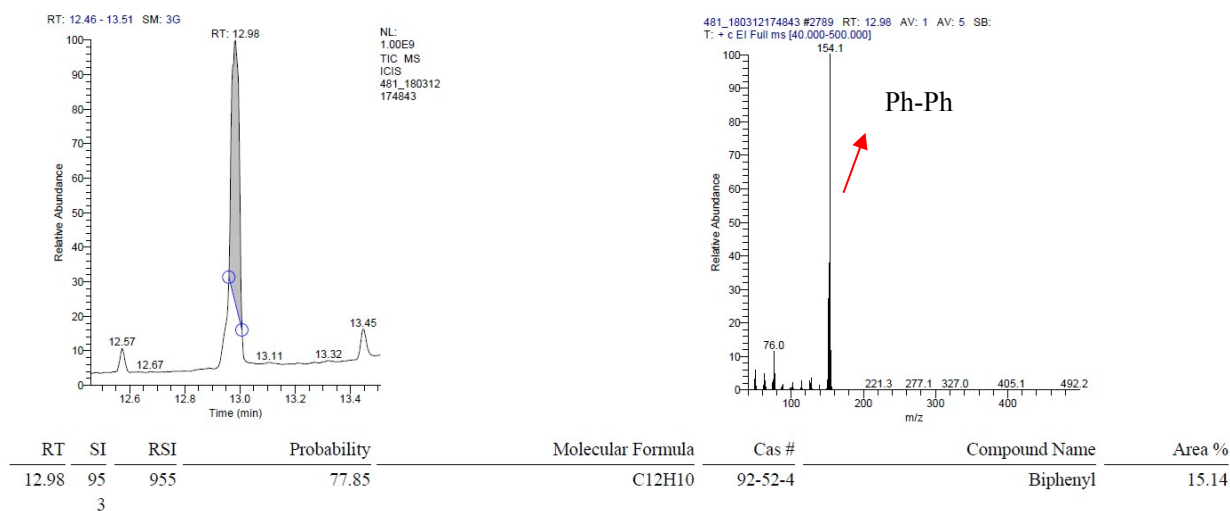


Figure S136. Detection of biphenyl by GC-MS

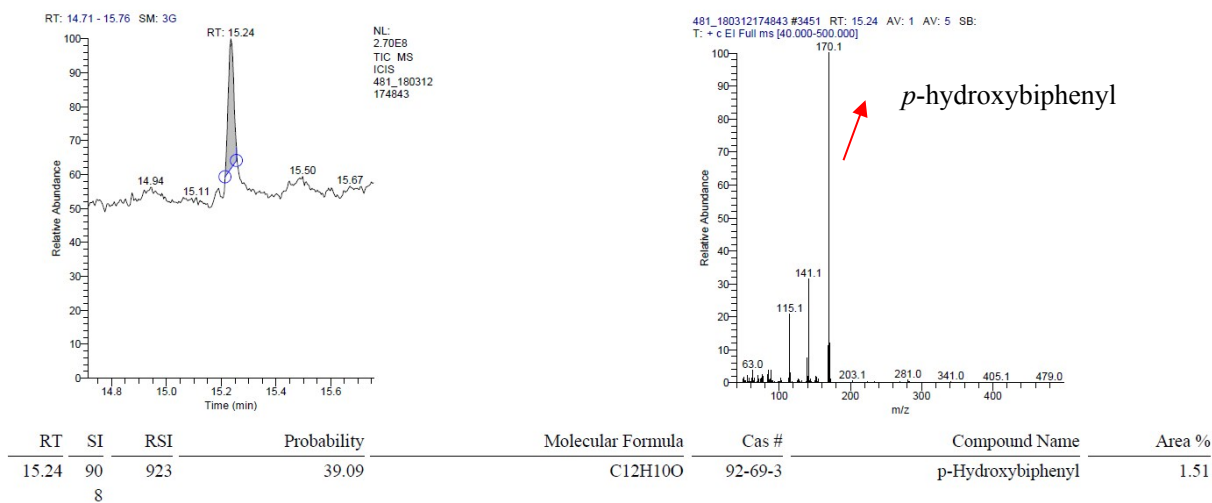
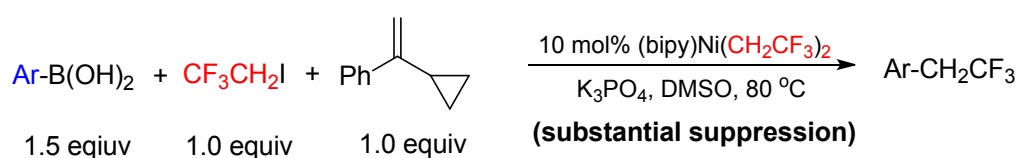


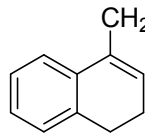
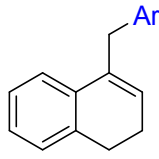
Figure S137. Detection of p-hydroxybiphenyl by GC-MS

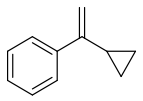
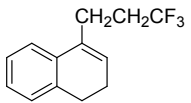
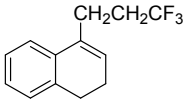
(c) Radical clock experiment

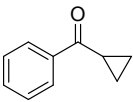
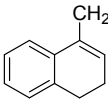
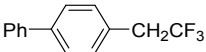
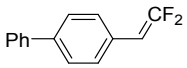

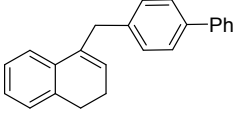
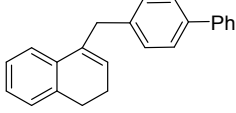
4-biphenylboronic acid (0.3 mmol, 1.5 equiv), K₃PO₄ (0.6 mmol, 3.0 equiv), followed by a solution of CF₃CH₂I (0.2 mmol, 1.0 equiv) and (1-cyclopropylvinyl)benzene (0.2 mmol, 1.0 equiv) in the DMSO solvent (0.5 mL) were loaded into a 25 mL of Schlenk tube which was subject to evacuating/flushing with nitrogen gas three times. The precatalyst **2** (10.0 mol%) in the DMSO solvent (0.5 mL) was added dropwise into the reaction system subsequently. The Schlenk tube was screw capped and put into a preheated oil bath (80 °C). After stirring for 24 h, the reaction mixture was cooled to room temperature and poured into a saturated aqueous ammonium chloride solution (10.0 mL). The aqueous phase was extracted with ether three times (10.0 mL × 3). The combined organic phase was analyzed by GC-MS with the *p*-xylene internal standard.

Table S10. GC-MS result of radical clock experiment



GC-MS identified species:		(Ar= 4-biphenyl)
		biphenyl (<i>m/z</i> 154) (deborylation)
20 (<i>m/z</i> 226) (trifluoroethyl trapping)	21 (<i>m/z</i> 296) (insertion of aryl-nickel)	4-hydroxybiphenyl 19 <i>m/z</i> 170 (oxidation of aryl-nickel)

Retention time/minute	Detected species	Area%
3.80	CF ₃ CH ₂ I	7.71
8.31	<i>p</i> -xylene (internal standard)	16.15
11.51		12.59
11.68		0.49
12.10	 (its isomer)	0.80

12.35		1.69
12.89	 (its isomer)	2.17
12.98	Biphenyl	11.35
13.80		1.71
14.26		0.44
15.23		1.46
15.57		0.49
16.19	 (its isomer)	0.85

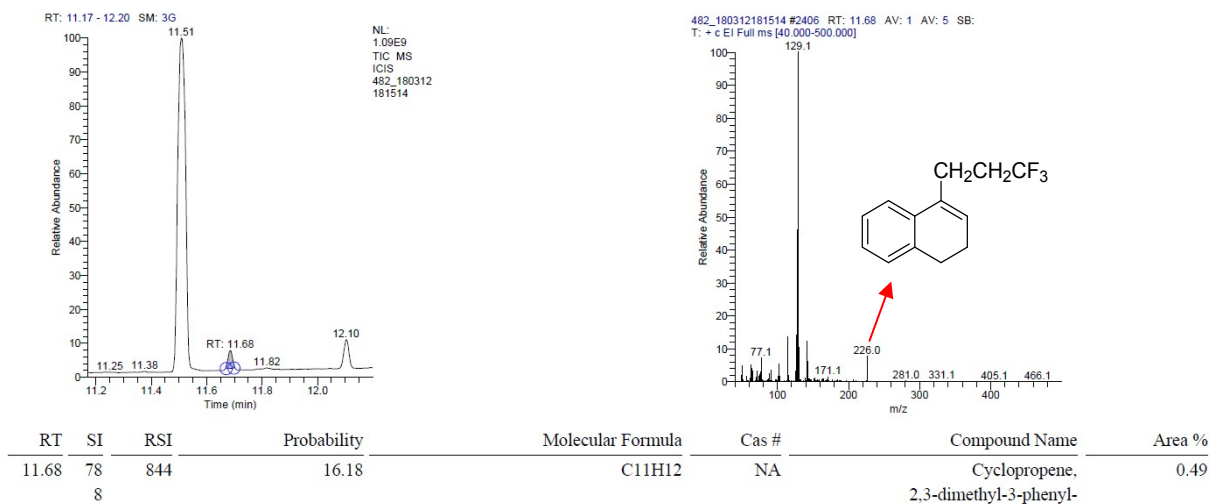


Figure S138. Detection of trapping of CF_3CH_2 radical by radical-clock

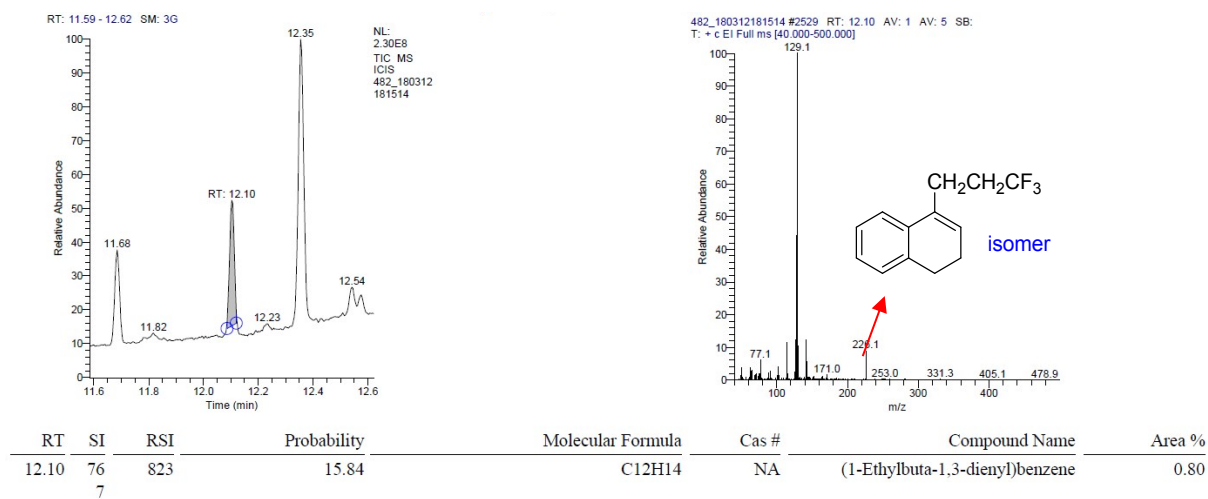


Figure S139. Detection of trapping of CF_3CH_2 radical by radical-clock (an isomeric product)

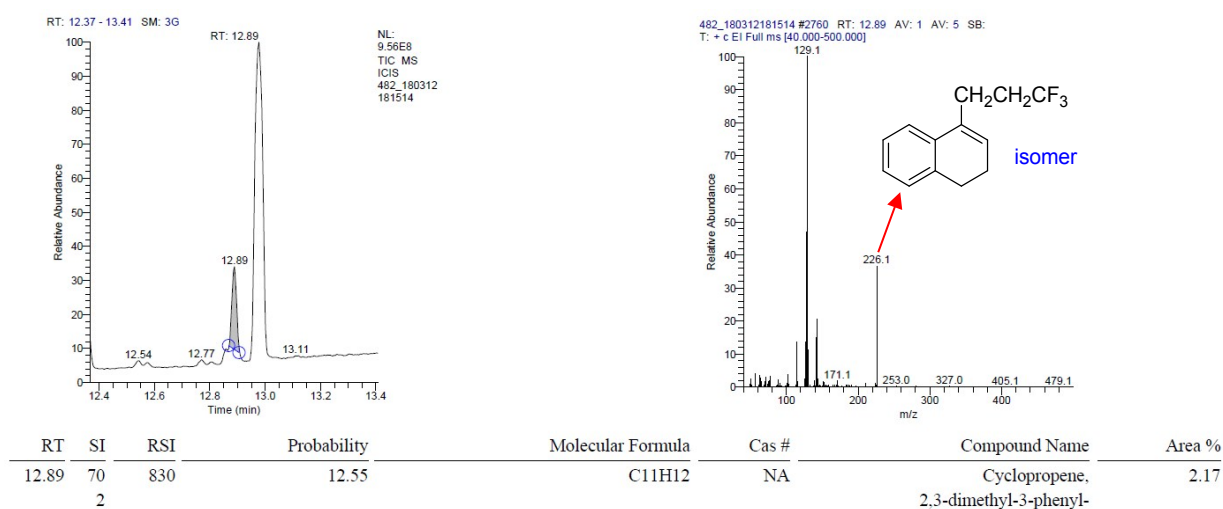


Figure S140. Detection of trapping of CF_3CH_2 radical by radical-clock (another isomer)

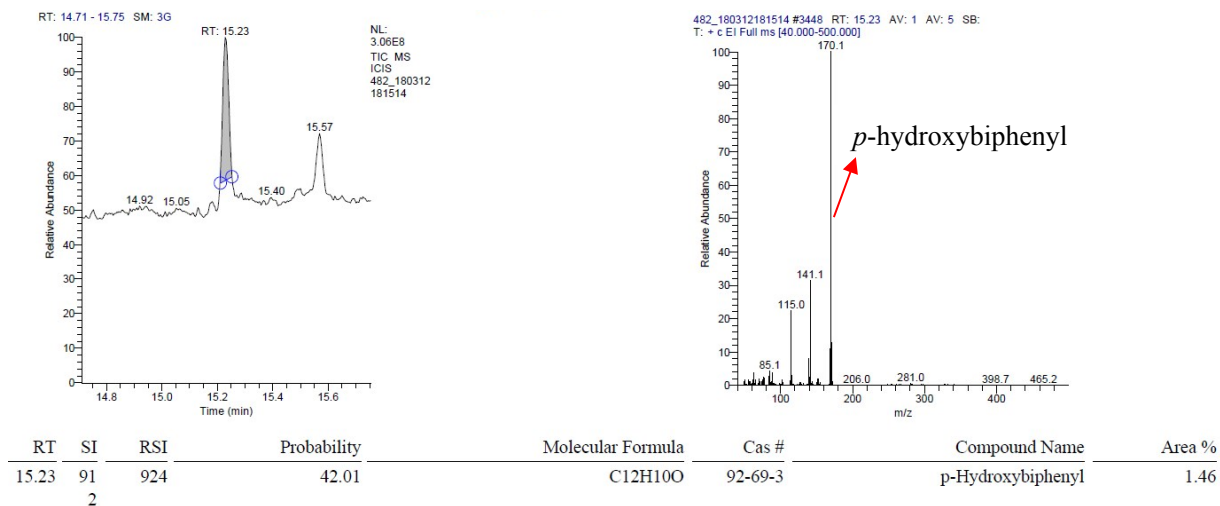


Figure S141. Detection of *p*-hydroxybiphenyl in radical-clock experiment

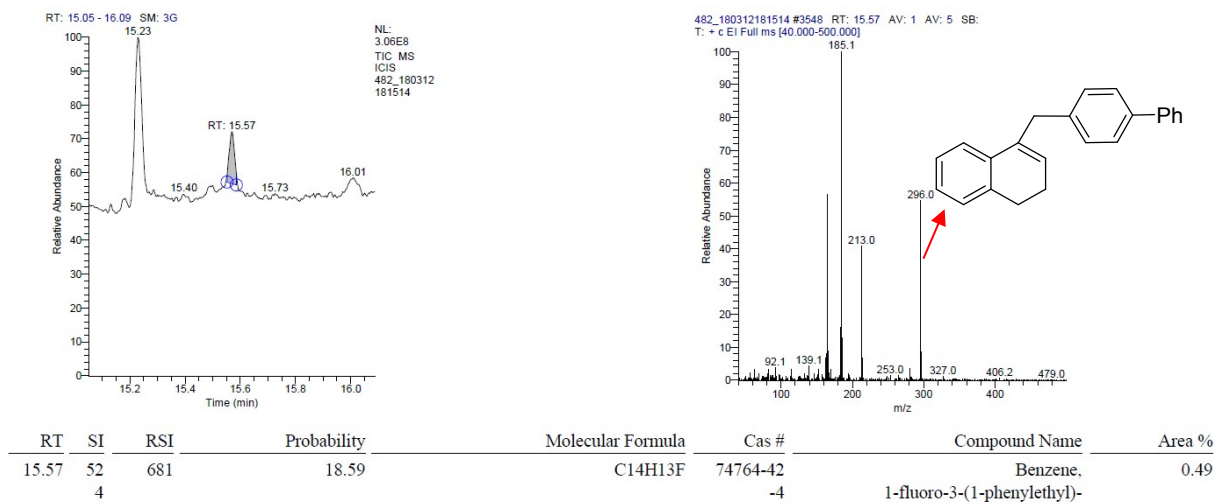


Figure S142. Detection of trapping of Aryl moiety by radical-clock

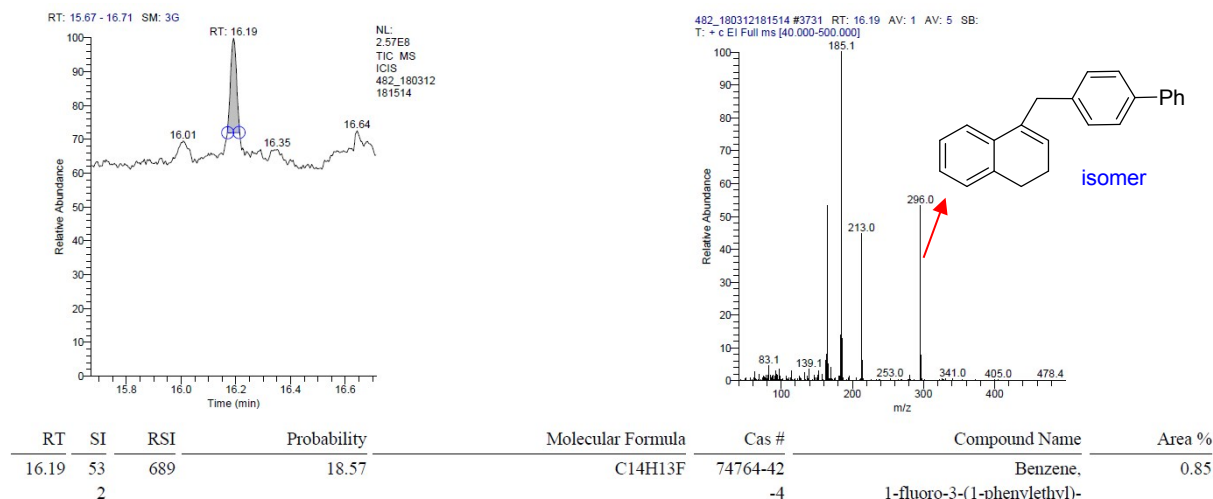
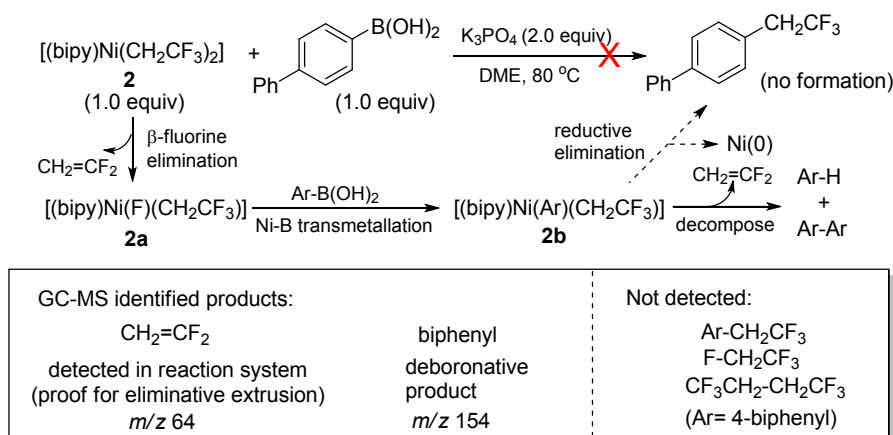


Figure S143. Detection of trapping of Aryl moiety by radical-clock (an isomeric product)

(d) Questioning over Ni(0)/Ni(II) or Ni(I)/Ni(III) catalytic cycle

i) Stoichiometric reaction of 4-biphenylboronic acid and 2 for verifying Ni(0)/Ni(II) cycle



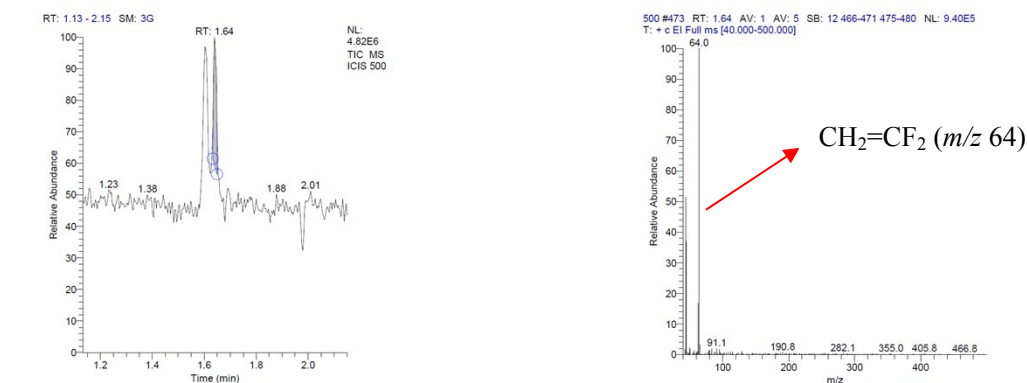
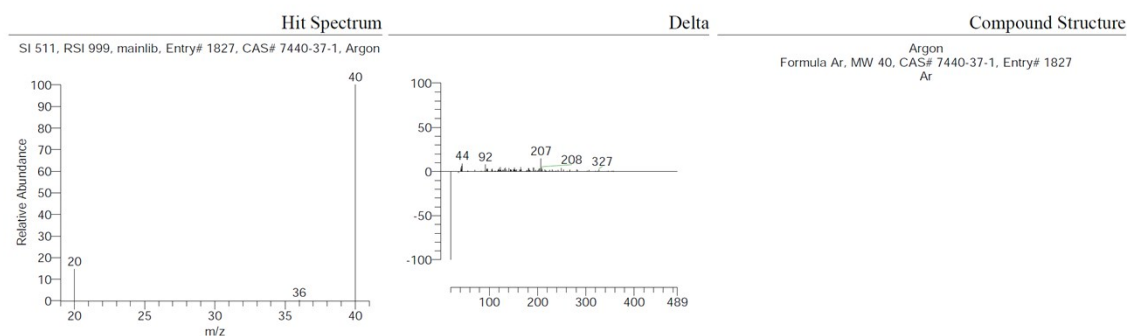
4-biphenylboronic acid (0.2 mmol, 1.0 equiv), K₃PO₄ (0.4 mmol, 2.0 equiv), followed by a solution of precatalyst **2** (0.2 mmol, 1.0 equiv) in the DME solvent (1 mL) were loaded into a 25 mL of Schlenk tube which was subject to evacuating/flushing with nitrogen gas three times. The Schlenk tube was screw capped and put into a preheated oil bath (80 °C). After stirring for 24 h, the reaction mixture was filtrated through a PTFE filter and analyzed by GC-MS.

Table S11. GC-MS analysis for probing the catalytic cycle of Ni(0)/Ni(II)

Retention Time/minute	Detected species	Area%
1.64	CF ₂ =CF ₂ (<i>m/z</i> 64)	0.03% ^a
12.91	Biphenyl (<i>m/z</i> 154)	37.39%

^aF₂C=CH₂ was detected in trace amount in GC-MS due to the volatility of its gaseous properties.

RT	SI	RSI	Probability	Molecular Formula	Cas #	Compound Name	Area %
1.60	51	999	50.16	Ar	7440-37-1	Argon	0.04
	1						



RT	SI	RSI	Probability	Molecular Formula	Cas #	Compound Name	Area %
1.64	76	899	90.09	C2H2F2	75-38-7	Ethene, 1,1-difluoro-	0.03
	2						

Figure S144. Detection of CH₂=CF₂ by GC-MS

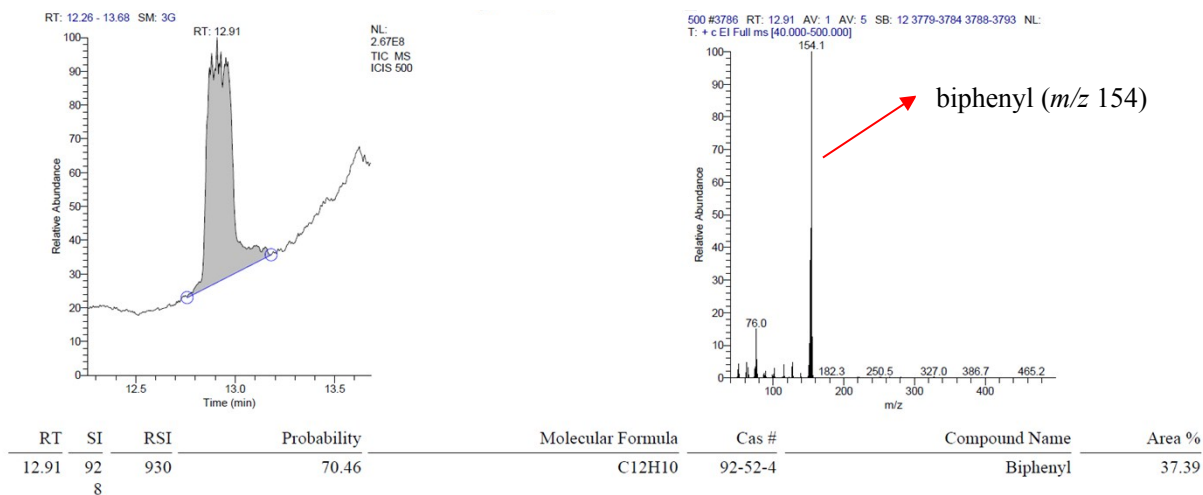
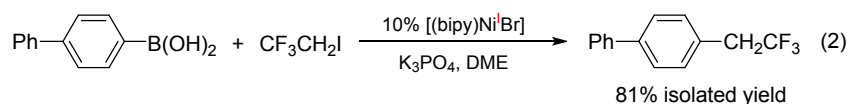


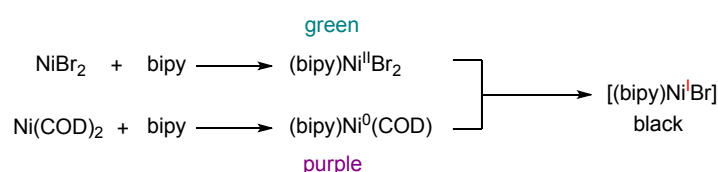
Figure S145. Detection of debornative product biphenyl by GC-MS

ii) Coupling reaction under the catalysis of Ni(I) precatalyst for verifying Ni(I)/Ni(III) cycle



4-biphenylboronic acid (0.3 mmol, 1.5 equiv), K₃PO₄ (0.6 mmol, 3.0 equiv), followed by a solution of CF₃CH₂I (0.2 mmol, 1.0 equiv) in the DME solvent (0.5 mL) were loaded into a 25 mL of Schlenck tube which was subject to evacuating/flushing with nitrogen gas three times. The presumed catalyst (bipy)Ni^IBr (0.02 mmol, 1.0 equiv) was added into the reaction system subsequently. The Schlenck tube was screw capped and put into a preheated oil bath (80 °C). After stirring for 24 h, the reaction mixture was cooled to room temperature and poured into a saturated aqueous ammonium chloride solution (10.0 mL). The aqueous phase was extracted with ether three times (10.0 mL×3). After removing the solvent in vacuo, the residue was purified by flash chromatography on silica gel to afford the corresponding ArCH₂CF₃ product (38 mg, isolated yield 81%).

iii) Procedure for the preparation of [(bipy)Ni^IBr]



Analogous to the preparation method of (dppf)Ni^(I)Cl of Hazari group (Guard, L. M.; Mohadjer Beromi, M.; Brudvig, G. W.; Hazari, N.; Vinyard, D. J. *Angew. Chem. Int. Ed.* **2015**, *54*, 13352.), the presumed catalyst [(bipy)Ni^IBr] was prepared according to the following procedure. To a THF solution (5.0 mL) of bipyridine (0.5 mmol) was added nickel(II) bromide (0.5 mmol) and the reaction mixture was stirred at room temperature for 24 h. The reaction mixture exhibited a suspended solution of green color. Meanwhile, a reaction mixture of Ni(COD)₂ (0.5 mmol) and bipyridine (0.5 mmol) in the solution of THF was also stirred at room temperature for 24 h to accomplish the coordination step (a purple solution was formed). The two reaction solutions were combined together and stirred at room temperature for an extra 24 h. A dark black precipitate was produced in the combined reaction solution and was filtrated. The black solid was washed with ether (10.0 mL) and pentane (10.0 mL), and dried in *vacuo* to furnish the presumed [(bipy)Ni^IBr] as a dark black powder (271 mg, Estimated Yield 92%). The insolubility of presumed catalyst [(bipy)Ni^IBr] in organic solvent made NMR characterization failed. Thus, XPS (X-ray Photoelectron Spectroscopy) and IR were used to characterize the presumable [(bipy)Ni^IBr]. The XPS of [(bipy)Ni^IBr] illustrated

the peaks corresponding to Ni(I) at 855.486 eV which was negatively shifted by 0.214 eV in comparison with a known $K_2[Ni^{(I)}(CN)_3]$ ($Ni^{(I)}$ at 855.70 eV) (Figure S123). The IR spectra of $[(bipy)Ni^I Br]$ (KBr pellets method) showed strong absorption bands in the ranges of 1599-1441 cm^{-1} for the stretching mode of pyridine ring, which were blue shifted toward higher frequency as compared to the free bipyridine.

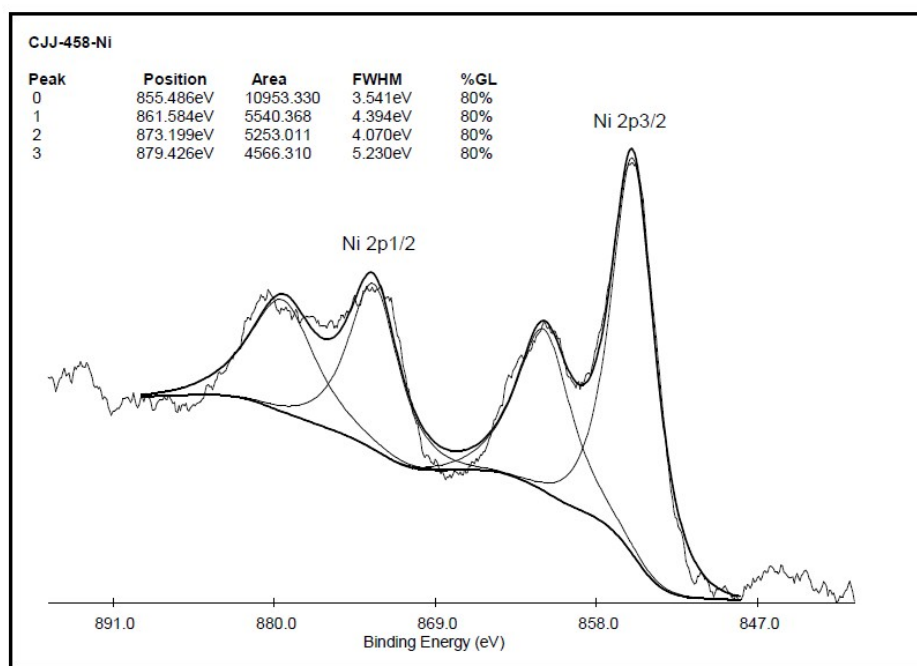


Figure S146. XPS (X-ray Photoelectron Spectroscopy) analysis of $[(bipy)Ni^I Br]$

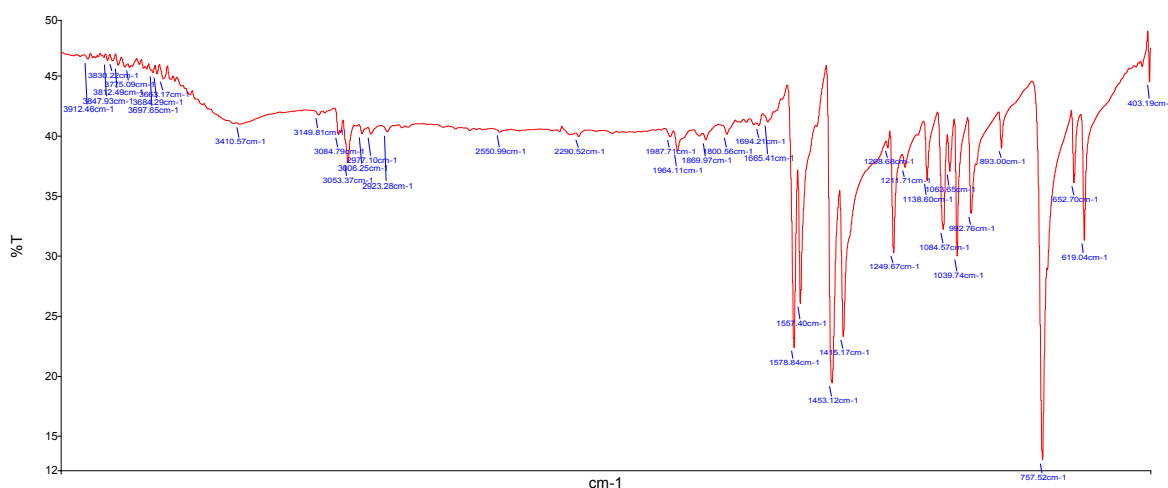


Figure S147. IR spectra of 2,2'-bipyridine

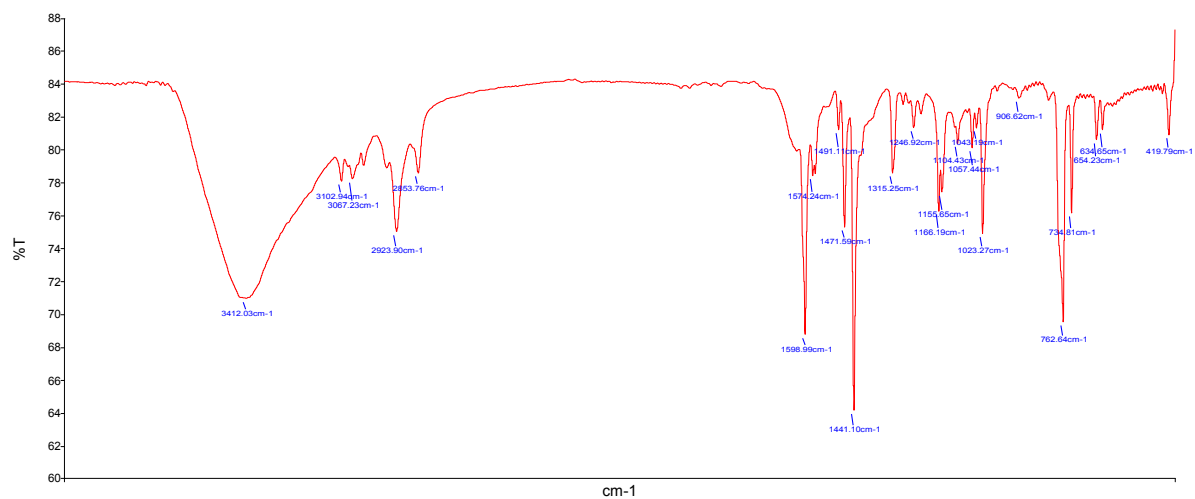


Figure S148. IR spectra of $[(bipy)Ni]Br$