SUPPORTING INFORMATION

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General Methods

Reactions were performed under a nitrogen atmosphere in flame dried glassware. NMR spectra were obtained on a Varian spectrometer operating at 400 or 500 MHz for ¹H, 125 MHz for ¹³C, and 202 MHz for ³¹P at 25 °C unless noted otherwise. ¹³C NMR spectra were referenced relative to CD_2Cl_2 (δ 53.8) or $CDCl_3$ (δ 77.2), ¹H NMR spectra were referenced relative to residual $CHCl_3$ (δ 7.26) or $CHDCl_2$ (δ 5.32). ³¹P NMR spectra was referenced to an external solution of triphenylphosphine oxide in CD_2Cl_2 (δ 26.9). Flash column chromatography was performed employing 200-400 mesh silica gel 60 (EM). Thin layer chromatography (TLC) was performed on silica gel 60 F254. CD_2Cl_2 was dried over CaH_2 and degassed prior to use. Ether, methylene chloride, and THF were purified by passage through columns of activated alumina under nitrogen. Reagents and other materials were obtained through major chemical suppliers and were used as received unless noted otherwise. Room temperature is 25 °C. Glassware and NMR tubes used for generation of allenylidenes was silanized before use.^{\$1} Propargylic ethers **7a**

and 7**b** were prepared as white solids employing published procedures.^{S2,S3} Acetylene-¹³C₂ was prepared from amorphous carbon-¹³C (99% ¹³C) employing a published procedure and condensed at -78 °C.^{S4} Elemental Analyses were performed by Atlantic Microlab, Inc.

Synthesis of Vinyl Boronic esters



Scheme S1. Propargylic alcohols and ethers and vinyl boronic esters and their isotopomers.

(*E*)-Vinyl boronic esters were prepared employing a modification of the procedure published by Frost.^{S5}

(*E*)-BpinC(H)=C(H)-C(OMe)(C₆H₄OMe)₂ [(*E*)-8b]. A yellow solution of 7b (1.10 g, 3.90 mmol), Cp₂ZrHCl (100 mg, 0.39 mmol), HBPin (0.60 mL, 4.1 mmol) and Et₃N (55 μ L, 0.39 mmol) in CH₂Cl₂ (5 mL) was stirred in a sealed tube at 60 °C for 2 days in the dark. The reaction mixture was diluted with hexanes, filtered through silica gel, and concentrated under vacuum. The resulting residue was chromatographed (SiO₂; hexanes–EtOAc = 85:15) to give a yellow oil, which was chromatographed a second time (SiO₂: hexanes–EtOAc = 9:1) to give yellow solid, which was then crystalized from ⁱPrOH at -20 °C to give (*E*)-S2 as white solid (0.35 g, 22%). ¹H NMR (400 MHz; CDCl₃): δ 7.22 (d, *J* = 8.8 Hz, 4 H), 7.08 (d, *J* = 18.2 Hz, 1H), 6.83 (d, *J* = 8.8 Hz, 4H), 5.63 (d, *J* = 18.1 Hz, 1H), 3.79 (s, 6H), 3.08 (s, 3H), 1.26 (s, 12H). ¹³C{¹H} NMR (125 MHz; CDCl₃): δ 158.74, 153.91, 135.82, 129.58, 113.37, 84.80, 83.42, 55.39, 52.07, 25.03. HRMS (ESI) calcd. (found) for C₂₃H₂₈BO4 [M-OMe]⁺: 379.2770 (379.2083).

(*E*)-BpinC(D)=C(H)-C(OMe)(C₆H₄OMe)₂ [(*E*)-8b- d_1]. A solution of 7b (0.80 g, 2.8 mmol) in methanol-O-d (5 mL) was added into a solution of NaOMe (46 mg, 0.85 mmol) in methanol-O-d (2 mL)

at room temperature and stirred for 2 h and the resulting mixture was concentrated under reduced pressure and diluted with fresh methanol-*O*-*d* (4 mL). This procedure was repeated three times and the resulting suspension was treated with D₂O (4 mL) and extracted with CH₂Cl₂. The combined organic extracts were dried (MgSO₄) and concentrated to give 4,4'-(1-methoxyprop-2-yne-1,1-diyl-3-*d*)bis(methoxybenzene) (**7b**-*d*₁) as white solid (0.61 g, 76%) with >95% deuterium incorporation at terminal alkynyl position. **7b**-*d*₁ was converted into (*E*)-**8b**-*d*₁ employing a procedure analogous to that used to synthesize (*E*)-**8b**. ¹H NMR analysis of (*E*)-**8b**-*d*₁ established >95% deuterium incorporation at C₁ position, with δ 7.08 (s, 1H) at C₂ position.

(*E*)-Bpin¹³C(H)=¹³C(H)-C(OMe)(C₆H₄OMe)₂ [(*E*)-8b-1,2-¹³C₂]. THF (10 mL) was added to condensed H¹³C=¹³CH [generated from reaction of carbon-¹³C (0.47 g) with Ca(0) (2.1 g)]^{S4} at –116 °C and the resulting solution was treated sequentially with *n*-BuLi (2.1 mL, 2.5 M in hexanes, 5.3 mmol) at –78 °C and a solution of 4,4'-bismethoxybenzophenone (1.3 g, 5.4 mmol) in THF (20 mL). The resulting solution was stirred at –78 °C for 20 min, during which time the initially pale yellow solution became colorless. The solution was warmed gradually to room temperature, stirred for 12 h, and then treated with water (25 mL). The layers were separated, the aqueous phase was extracted with diethyl ether, and the combined organic extracts were dried (MgSO₄), filtered, and concentrated under vacuum. The resulting colorless oil was chromatographed (SiO₂: hexanes–EtOAc = 9:1) to give 1,1-bis(4-methoxyphenyl)prop-2-yn-1-ol (S1-1,2-¹³C₂) (197 mg, 0.73 mmol). Bisanisyl ethynyl alcohol S1-1,2-¹³C₂ was methylated with NaH/MeI employing a procedure analogous to that used to synthesize (*E*)-8b gave (*E*)-8b-1,2-¹³C₂ as colorless oil (79%).

For S1-1,2-¹³C₂: ¹H NMR (400 MHz; CDCl₃; selected resonances): δ 2.85 (dd, ¹J_{HC} = 250.5, ²J_{HC} = 49.8 Hz, 1H). ¹³C{¹H} NMR (100 MHz; CDCl₃): δ 86.97 (d, ¹J_{CC} = 171.3 Hz), 75.18 (d, ¹J_{CC} = 171.3 Hz).

For (*E***)-8b-1,2-¹³C₂:** ¹H NMR (400 MHz; CDCl₃): δ 5.62 (ddd, ¹*J*_{HC} = 142.5, ³*J*_{HH} = 18.3, ²*J*_{HC} = 7.3 Hz, 1H). ⁻¹³C{¹H} NMR (100 MHz; CDCl₃): δ 153.86 (d, ¹*J*_{CC} = 62.3 Hz), 117.96 (m). ¹³C NMR (100 MHz; CDCl₃): δ 153.86 (ddd, ¹*J*_{HC} = 156.6, ¹*J*_{CC} = 62.4, ²*J*_{HC} = 7.1 Hz).

(*Z*)-BpinC(H)=C(H)-C(OMe)(C₆H₄OMe)₂ [(*Z*)-8b]. Compound (*Z*)-8b was prepared employing a modification of the procedure published by Miyaura.^{S6} A mixture of **7b** (195 mg, 0.69 mmol), HBpin (0.11 mL, 0.79mmol), Et₃N (0.1 mL, 0.69 mmol), [Rh(COD)CI]₂ (10.2 mg, 2.1×10^{-2} mmol), and PCy₃ (23.5 mg, 8.4×10^{-2} mmol) in cyclohexane (2.1 mL) was stirred at room temperature for 3.5 h. The resulting mixture was concentrated under vacuum and the resulting residue was chromatographed (SiO₂; hexanes–EtOAc = 90:10) to give (*Z*)-8b (110 mg, 39%) as a yellow oil. ¹H NMR (500 MHz; CDCl₃): δ 7.31 (d, *J* = 8.8 Hz, 4H), 6.85 (d, *J* = 8.8 Hz, 4H), 6.44 (d, *J* = 13.9 Hz, 1H), 5.40 (d, *J* = 13.9 Hz, 1H), 3.81 (s, 6H), 2.95 (s, 3H), 1.31 (s, 12H). ¹³C{¹H} NMR (125 MHz; CDCl₃): δ 158.80, 148.74, 134.57, 130.05, 113.42, 84.95, 83.16, 55.4, 52.26, 25.33. HRMS (ESI) calcd. (found) for C₂₄H₃₁BO₅ [M+H]⁺: 411.2337 (411.2327). [M+Na]⁺: 433.2157 (433.2152) [M+K]⁺: 499.1896 (499.1895).

(*E*)-BpinC(H)=C(H)C(OMe)Ph₂ [(*E*)-8a]. Zirconium-catalyzed hydroboration of 7a (1.0 g, 4.5 mmol) with HBpin employing a procedure similar to that used to synthesize (*E*)-8b led to isolation of (*E*)-8a (1.0 g, 64 %) as colorless oil, which solidified upon cooling at -20 °C. ¹H NMR (400 MHz; CDCl₃): δ 7.30 - 7.16 (m, 10H), 7.09 (d, *J* = 18.1 Hz, 1H), 5.62 (d, *J* = 18.1 Hz, 1H), 3.06 (s, 3H), 1.21 (s, 12H). ¹³C{¹H} NMR (125 MHz; CDCl₃): δ 153.24, 143.61, 128.32, 128.10, 127.29, 119.02 (br), 85.30, 83.47, 52.26, 25.03. HRMS (ESI) calcd. (found) for C₂₂H₂₇BO₃ [M-OMe]⁺: 319.1864 (319.1871).

Synthesis of Gold Vinyl Complexes





(*E*)-(IPr)Au[n^1 -C(H)=C(H)-C(OMe)(C₆H₅)₂ [(*E*)-6a]. A suspension of 7a (45 mg, 0.13 mmol), IPrAuCl (80 mg, 0.13 mmol) and Cs₂CO₃ (42 mg, 0.13 mmol) in iPrOH (1.3 mL) was stirred for one day at 60 °C. The reaction mixture was cooled to room temperature and concentrated under vacuum. The resulting white solid was dissolved in Et₂O and filtered through thin pad of basic alumina, concentrated, and the procedure was repeated with hexanes/EtOAc (v/v = 9/1). The resulting solid was recrystallized by layering a concentrated CH₂Cl₂ solution with pentane at -20 °C to give (*E*)-6a as colorless crystals (62 mg, 59%). ¹H NMR (400 MHz; CDCl₃): δ 7.47 (t, *J* = 7.8 Hz, 2H), 7.30 – 7.24 (m, 10H), 7.19 – 7.05 (m, 8H), 6.26 (d, *J* = 19.1 Hz, 1H), 5.76 (d, *J* = 19.2 Hz, 1H), 3.08 (s, 3H), 2.63 (h, *J* = 7.1 Hz, 4H), 1.35 (d, *J* = 6.9 Hz, 12H), 1.21 (d, *J* = 7.0 Hz, 12H). ¹³C{¹H} NMR (125 MHz; CDCl₃): δ 198.19, 161.01, 146.22, 145.93, 143.08, 134.88, 130.25, 128.22, 127.44, 126.00, 124.07, 122.87, 86.62, 51.22, 28.94, 24.56, 24.17. HRMS (ESI) calcd. (found) for C₄₃H₅₂AuN₂O (M)⁺: 809.3740 (809.3748).

(*E*)-(IPr)Au[η^1 -C(H)=C(H)-C(OMe)(4-C₆H₄OMe)₂ [(*E*)-6b]. A suspension of (*E*)-8b (93 mg, 0.23 mmol), IPrAuCl (141 mg, 0.23 mmol) and Cs₂CO₃ (74 mg, 0.23 mmol) in iPrOH (2.3 mL) was stirred for one day at 60 °C. The reaction mixture was cooled to room temperature and concentrated under vacuum. The resulting white solid was dissolved in Et₂O and filtered through thin pad of basic alumina, concentrated, and the procedure was repeated. The resulting solid was recrystallized by layering a concentrated CH₂Cl₂ solution with pentane at –20 °C to give (*E*)-6b as colorless crystals (117 mg, 59%). ¹H NMR (400 MHz; CDCl₃): δ 7.46 (t, *J* = 7.8 Hz, 2H), 7.27 (d, *J* = 8.3 Hz, 4H), 7.15 (d, *J* = 8.7 Hz, 4H),

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7.09 (s, 2H), 6.71 (d, J = 8.8 Hz, 4H), 6.22 (d, J = 19.1 Hz, 1H), 5.75 (d, J = 19.1 Hz, 1H), 3.73 (s, 6H), 3.05 (s, 3H), 2.63 (hept, J = 6.8 Hz, 4H), 1.35 (d, J = 6.8 Hz, 12H), 1.21 (d, J = 6.9 Hz, 12H). ¹³C{¹H} NMR (125 MHz; CDCl₃): δ 198.3, 160.2, 157.9, 145.9, 143.6, 138.6, 134.9, 130.2, 129.4, 124.1, 122.9, 112.8, 86.1, 55.3, 51.0, 24.6, 24.2. Anal. calcd. (found) for C₄₅H₅₆AuN₂O₃: C, 62.13 (62.20); H, 6.49 (6.25); N, 3.22 (3.21).

(*E*)-(IPr)Au[η^1 -C(D)=C(H)-C(OMe)(4-C₆H₄OMe)₂] [(*E*)-6b-*d*₁]. Gold vinyl complex (*E*)-6b-*d*₁ was isolated as a white solid in 100 % yield from reaction of (*E*)-8b-*d*₁ (31 mg, 7.3 × 10⁻² mmol) and (IPr)AuCl (45 mg, 7.3 × 10⁻² mmol) employing a procedure analogous to that used to synthesize (*E*)-6b. ¹H NMR analysis of (*E*)-6b-*d*₁ established >95% deuterium incorporation at C₁ position.

(*E*)-(IPr)Au[η^{1} -1³C(H)=1³C(H)-C(OMe)(4-C₆H₄OMe)₂] [(*E*)-6b-1,2-1³C₂]. Complex (*E*)-6b-1,2-1³C₂ was isolated as a white solid in 62% yield from reaction of (*E*)-8b-1,2-1³C₂ (67 mg, 1.6 × 10⁻¹ mmol) and (IPr)AuCl (101 mg, 1.6 × 10⁻¹ mmol) employing a procedure analogous to that used to synthesize (*E*)-6b. ¹H NMR (400 MHz; CDCl₃, selected resonances): δ 6.14 (ddd, $^{1}J_{HC}$ = 171.7, $^{3}J_{HH}$ = 19.1, $^{2}J_{HC}$ = 5.8 Hz, 1H), 5.78 (ddd, $^{1}J_{HC}$ = 155.1, $^{3}J_{HH}$ = 19.1, $^{2}J_{HC}$ = 4.7 Hz, 1H). $^{13}C{}^{1}H$ NMR (100 MHz; CDCl₃): δ 160.2 (d, *J* = 59.1 Hz), 143.5 (d, *J* = 58.7 Hz).

(*Z*)-(IPr)Au[η^1 -C(H)=C(H)-C(OMe)(4-C₆H₄OMe)₂] [(*Z*)-6b]. Complex (*Z*)-6b was isolated as a white solid in 25% yield from reaction of (*Z*)-8b with (IPr)AuCl employing a procedure analogous to that used to synthesize (*E*)-6b after three successive recrystallizations from diethyl ether. ¹H NMR (500 MHz; CDCl₃): δ 7.44 (t, *J* = 7.8 Hz, 2H), 7.26 (d, *J* = 7.8 Hz, 4H), 7.14 (d, *J* = 8.7 Hz, 4H), 7.10 (s, 2H), 6.78 (d, *J* = 14.1 Hz, 1H), 6.62 (d, *J* = 13.4 Hz, 1H), 6.62 (d, *J* = 8.9 Hz, 4H), 3.73 (s, 6H), 2.32 (s, 3H), 1.38 (d, *J* = 6.9 Hz, 12H), 1.21 (d, *J* = 6.8 Hz, 12H). ¹³C{¹H} NMR (125 MHz; CDCl₃): δ 198.01, 157.61, 156.12, 146.02, 143.54, 140.44, 135.19, 130.20, 129.41, 124.47, 124.08, 122.95, 112.72, 84.09, 55.25, 50.71, 28.96, 24.54, 24.17.

(*E*)-(P1)Au[η^1 -C(H)=C(H)-C(OMe)(4-C₆H₄OMe)₂] [(*E*)-6c]. Gold vinyl complex (*E*)-6c was isolated as a white solid in 71% yield from reaction of (*E*)-8b (67 mg, 1.6 × 10⁻¹ mmol) and (P1)AuCl (86 mg, 1.6 × 10⁻¹ mmol) employing a procedure analogous to that used to synthesize (*E*)-6b. ¹H NMR (400 MHz; CDCl₃): δ 7.86 (td, *J* = 7.0, 1.8 Hz, 1H), 7.45 - 7.40 (m, 2H), 7.30 (d, *J* = 8.8 Hz, 4H), 7.25 - 7.21 (m, 3H), 7.13 (d, *J* = 6.9 Hz, 2H), 7.05 (t, *J* = 7.5 Hz, 1H), 6.84 (d, *J* = 8.7 Hz, 4H), 5.93 (dd, *J* = 19.0, ³*J*_{PH} = 6.6 Hz, 1H), 5.83 (dd, *J* = 18.9, ⁴*J*_{PH} = 4.1 Hz, 1H), 3.81 (s, 6H), 3.18 (s, 3H), 1.39 (d, *J* = 14.3 Hz, 18H). ¹³C{¹H} NMR (125 MHz; CDCl₃): δ 158.09, 150.56 (d, *J* = 18.4 Hz), 143.96, 142.66, 138.55, 135.20, 132.99 (d, *J* = 7.6 Hz), 129.91, 129.75, 129.59, 128.26, 127.51, 126.50 (d, *J* = 5.2 Hz), 112.81, 86.46 (d, *J* = 8.6 Hz), 55.40, 51.35, 37.38 (d, *J* = 18.4 Hz), 31.22 (d, *J* = 7.2 Hz). ³¹P{¹H} (162 Hz; CDCl₃) δ 64.58. Anal. calcd. (found) for C₃₈H₄₆AuO₃P: C, 58.61 (58.76); H, 5.95 (5.88).

Ionization of gold vinyl complexes



Scheme S3. Gold vinyl carbene complexes and their isotopomers generated via ionization of gold vinyl complexes.

[(*E*)-(IPr)Au[η^1 -C(H)=C(H)-C(4-C₆H₄OMe)₂]⁺ OTf⁻ [(*E*)-5b]. A solution of (*E*)-6b (14.5 mg, 1.7 × 10⁻² mmol) in CD₂Cl₂ (0.35 mL) was added dropwise with constant agitation to an NMR tube containing a solution of TMSOTf (4.4 mg, 2.0 × 10⁻² mmol) and CH₂Br₂ (0.77 mg, 4.4 µmol; internal standard) in CD₂Cl₂ (0.35 mL) at –95 °C to give a bright orange solution of (*E*)-5b that was analyzed without isolation by NMR spectroscopy at or below –85 °C. A yield of 96% for the conversion of (*E*)-6b to (*E*)-5b was determined by integrating the H₁ resonance of (*E*)-5b at δ 9.52 relative to the resonance of CH₂Br₂ at δ

4.96 in the ¹H NMR spectrum. ¹H NMR (500 MHz; CD₂Cl₂; -95 °C): δ 9.51 (d, *J* = 17.8 Hz, 1H), 7.88 (dd, *J* = 9.2, 2.4 Hz, 1H), 7.71 (d, *J* = 17.8 Hz, 1H), 7.52 (t, *J* = 7.7 Hz, 2H), 7.33 (s, 2H), 7.31 (d, *J* = 3.0 Hz, 4H), 7.29 (d, *J* = 8.9 Hz, 4H), 7.13 (dd, *J* = 8.9, 2.8 Hz, 1H), 6.99 (d, *J* = 8.7 Hz, 4H), 3.96 (s, 3H), 3.89 (s, 3H), 2.44 (sept, *J* = 6.9 Hz, 4H), 1.28 (d, *J* = 6.8 Hz, 12H), 1.20 (d, *J* = 6.9 Hz, 12H). ¹³C{¹H} NMR (125 MHz; CD₂Cl₂; -95 °C): δ 243.7, 188.7, 185.7, 168.3, 167.6, 146.9, 145.0, 142.4, 139.8, 136.4, 132.9, 129.8, 129.6, 123.5, 123.3, 116.0, 114.4, 56.6, 56.3, 28.0, 24.5, 22.6.

Fluxional behavior of (*E***)-5b**. When a solution of (*E*)-5b was warmed above –95 °C, the aryl methoxy resonances at δ 3.96 and 3.89 broadened and coalesced at –42 °C (Figure S1). An energy barrier of $\Delta G^{\ddagger}_{231K}$ = 10.5 kcal/mol was determined for interconversion of the anisyl groups of (*E*)-5b from the slow-exchange peak separation (Δv = 32 Hz) and coalescence temperature (T_c = –42 °C) according to the relationship [$k = \pi(\Delta v)/\sqrt{2}$].



Figure S1. Temperature dependence of the methoxy ¹H NMR resonances of (*E*)-**5b** in CD₂Cl₂ from – 80 °C to -37 °C in CD₂Cl₂.

[(*E*)-(IPr)Au[η¹-¹³C(H)=¹³C(H)-C(C₆H₄OMe)₂]⁺ OTf⁻ [(*E*)-5b-1,2-¹³C₂]. Complex (*E*)-5b-1,2-¹³C₂ was generated *in situ* from (*E*)-6b-1,2-¹³C₂ employing a procedure analogous to that used to synthesize (*E*)-5b. ¹H NMR (–95 °C, CD₂Cl₂, 500 MHz, selected resonances): δ 9.50 (dd, *J* = 135.8, 17.7 Hz, 1H; H₁), H₂ was obscured by aromatic protons. ¹³C{¹H} NMR (–80 °C, CD₂Cl₂, 125 MHz, selected resonances): δ 243.9 (d, *J* = 47.9 Hz, C1), 188.8 (d, *J* = 38.3 Hz, C_{lpr}), 185.8 (d, *J* = 48.0 Hz, C3), 146.9 (d, *J* = 47.9 Hz, C2). ¹³C NMR (–80 °C, CD₂Cl₂, 125 MHz, selected resonances): δ 244.4 (dd, *J*_{CH} = 135.4, *J*_{CC} = 48.5 Hz), 147.04 (dd, *J*_{CH} = 163.0, *J*_{CC} = 47.7 Hz).

[(*E*)-(P1)Au[η^1 -C(H)=C(H)-C(4-C₆H₄OMe)₂]⁺ OTf⁻ [(*E*)-5c]. Complex (*E*)-5c was synthesized in 95% yield from (*E*)-6c (¹H NMR) as a red solution employing a procedure analogous to that used to synthesize (*E*)-5b. ¹H NMR (500 MHz; CD₂Cl₂; -90 °C): δ 8.89 (d, *J* = 17.5 Hz, 1H), 8.16 (d, *J* = 9.1 Hz, 1H), 7.90 – 7.86 (m, 2H), 7.83 (dd, *J* = 17.5, 4.4 Hz, 1H), 7.62 – 7.48 (m, 3H), 7.37 (t, *J* = 7.6 Hz, 4H), 7.26 – 7.01 (m, 7H), 4.03 (s, 3H), 4.00 (s, 3H), 1.36 (d, *J* = 14.8 Hz, 18H). ¹³C{¹H} NMR (125 MHz; CD₂Cl₂; -90 °C): δ 252.33 (d, *J* = 93.7 Hz), 185.58, 168.29, 167.92, 148.15 (d, *J* = 16.1 Hz), 145.86, 142.70, 142.35, 140.03, 136.85 (br), 134.24, 131.71, 129.95 -124.95 (br), 115.97, 114.42, 56.53, 56.45, 36.32 (d, *J* = 21.8 Hz), 29.83 (br). ³¹P{¹H} (202 Hz; CD₂Cl₂; -95 °C): δ 61.8.

[(*E*)-(IPr)Au[η^1 -C(H)=C(H)-C(C₆H₅)₂]⁺ OTf⁻ [(*E*)-5a]. A solution of (*E*)-6a (11.2 mg, 1.4 × 10⁻² mmol) in CD₂Cl₂ (0.2 mL) was added via syringe to an NMR tube containing a solution of TMSOTf (3.7 mg, 1.7 × 10⁻² mmol) in ~1:1 mixture of CDFCl₂/CD₂Cl₂ at –116 °C to form a biphasic mixture consisting of a liquid phase of CDFCl₂/CD₂Cl₂ (bottom) and a solid phase of CD₂Cl₂ (top). As the top phase melted and dissolved in to the bottom layer, the bright orange color of (*E*)-5a was observed at the interface, ultimately forming a bright orange solution upon complete dissolution that was analyzed without isolation by NMR spectroscopy at –110 °C. ¹H NMR (500 MHz; CDFCl₂/CD₂Cl₂; –110 °C; selected resonances): δ 10.83 (d, *J* = 17.4 Hz, 1H), 8.18 (d, *J* = 17.5 Hz, 1H).

A solution of (*E*)-**6a** (11.8 mg, 1.5×10^{-2} mmol) in CD₂Cl₂ (0.3 mL) was added via syringe to a solution of TMSOTF (4.2 mg, 1.8×10^{-2} mmol) and CH₂Br₂ (0.75 mg, 4.3 µmol; internal standard) in CD₂Cl₂ (0.3 mL) at -95 °C to form a bright red solution that faded within 10 s. ¹H NMR analysis of the resulting solution revealed the formation of TMSOMe in 92 ± 5% yield, determined by integrating methoxy resonance of TMSOMe at δ 3.29 relative to the resonance of CH₂Br₂ at δ 4.96. ¹H NMR also revealed formation of corresponding to the known (*E*)-1,1,6,6-tetraphenylhexa-1,3,5-triene^{S7} at δ 6.74 (dd, *J* = 7.6, 3.1 Hz) and δ 6.51 (dd, *J* = 7.5, 3.1 Hz) and for (*Z*)-1,1,6,6-tetraphenylhexa-1,3,5-triene at δ 6.10 (br d, *J* = 8.8 Hz) [Lit. for (*E*)-1,1,6,6-tetraphenylhexa-1,3,5-triene (CDCl₃, 400 MHz): δ 6.72 (dd, *J* = 7.6, 3.2 Hz), 6.51 (dd, *J* = 7.6, 3.2 Hz). Lit. for (*Z*)-1,1,6,6-tetraphenylhexa-1,3,5-triene (CDCl₃, 400 MHz): δ 6.08 (dd, *J* = 8.4, 2.4 Hz)].^{S7} The yield of (*E*)-1,1,6,6-tetraphenylhexa-1,3,5-triene (64%) was determined by integrating vinyl proton at δ 6.51 relative to the resonance of CH₂Br₂ at δ 4.96 and the yield of (*Z*)-1,1,6,6-tetraphenylhexa-1,3,5-triene (64%) was determined by integrating vinyl proton at δ 6.50 relative to the resonance of CH₂Br₂ at δ 4.96 and the yield of (*Z*)-1,1,6,6-tetraphenylhexa-1,3,5-triene (64%) was determined by integrating vinyl proton at δ 6.50 relative to the resonance of CH₂Br₂ at δ 4.96 and the yield of (*Z*)-1,1,6,6-tetraphenylhexa-1,3,5-triene (64%) was determined by integrating vinyl proton at δ 6.10 relative to the resonance of CH₂Br₂ at δ 4.96.

To corroborate the NMR assignments of (*E*)- and (*Z*)-1,1,6,6-tetraphenylhexa-1,3,5-triene formed in the ionization of (*E*)-**6a**, a solution of (*E*)-**6a** (23.1 mg, 2.8×10^{-2} mmol) in CH₂Cl₂ (0.8 mL) was added via syringe to a solution of TMSOTf (7.6 mg, 3.4×10^{-2} mmol) in CH₂Cl₂ (1.2 mL) at –95 °C and warmed to room temperature. The resulting light green solution was concentrated under vacuum, extracted with hexanes/EtOAc (v/v = 9/1) and filtered through thin pad of silica gel to give yellow oil (5.2 mg). The resulting filtrate was dissolved in EtOAc and analyzed by GCMS on a standard non-polar column over temperature gradient 70 °C - 320 °C, with a hold at the high end. The GCMS trace displayed peaks at 16.49 min (10% mass yield) and 17.52 min (54% mass yield) with molecular weights (m/z = 384.8) consistent with a mixture of (*Z*)- and (*E*)-1,1,6,6-tetraphenylhexa-1,3,5-triene (Figure S2). No other resonances accounted for ≥5% of the reaction mixture. Mass yields were calculated from the total mass of isolated filtrate assuming that mass amount correlates to peak areas in the GC.



Figure S2. GCMS trace of the filtrated isolated from the reaction of (*E*)-**6a** with TMSOTf at -95 °C in CD₂Cl₂ with mass-selected (m/z = 384.8) GC trace (insert).

Ionization of (Z)-6b. A solution of (Z)-**6a** (10.0 mg, 1.2×10^{-2} mmol) in CD₂Cl₂ (0.3 mL) was added via syringe to a solution of TMSOTF (3.0 mg, 1.3×10^{-2} mmol) and CH₂Br₂ (0.62 mg, 3.5 µmol; internal standard) in CD₂Cl₂ (0.3 mL) at -95 °C to form an orange solution. ¹H NMR analysis of the

resulting solution displayed no resonances that could be attributed to the C1 or C2 vinyl protons of (*Z*)-**6b**, but displayed singlets at δ 4.63 and δ 4.59, as a small peak at δ 4.53 corresponding to the benzylic resonance of **9**. As the temperature was raised in 20 °C increments, the singlets at δ 4.63 and δ 4.59 disappeared and the benzylic resonance of **9** at δ 4.53 increased in intensity, ultimately accounting for a 58 ± 5% yield, as determined by integrating the δ 4.53 resonance relative to the resonance of CH₂Br₂ at δ 4.96. The crude reaction mixture was filtered through silica gel with hexanes–EtOAc = 9:1, concentrated, and the resulting residue was chromatographed (SiO₂; hexanes–EtOAc = 9:1) to give **9** (0.8 mg, 28%) as colorless oil, which solidified upon cooling at –20 °C.

For 9: ¹H NMR (500 MHz; CDCl₃): δ 7.11 (d, *J* = 8.1 Hz, 1H), 7.01 (d, *J* = 8.6 Hz, 1H), 6.95 (d, *J* = 2.4 Hz, 1H), 6.82 (dd, *J* = 5.3, 2.2 Hz, 1H), 6.80 (d, *J* = 8.8 Hz, 2H), 6.70 (dd, *J* = 8.2, 2.4 Hz, 1H), 6.58 (dd, *J* = 5.5, 1.9 Hz, 1H), 4.52 (t, *J* = 2.1 Hz, 1H), 3.83 (s, 3H), 3.77 (s, 3H). ¹³C{¹H} NMR (125 MHz; CDCl₃): 159.1, 158.5, 145.5, 141.5, 140.8, 131.4, 131.0, 128.7, 124.2, 114.1, 111.0, 106.8, 55.5, 55.3, 55.0. HRMS (ESI) calcd. (found) for C₁₇H₁₆O₂ [M+H]⁺: 253.1223 (253.1222).

Reactions of Nucleophiles with (E)-5b



Scheme S4. Products and isotopomers generated from reaction of (*E*)-**5b** with nucleophiles (Ar = $4-C_6H_4OMe$).

Reaction of (E)-5b with tetrahydrothiophene (THT). A solution of tetrahydrothiophene (1.9 mg, 2.21×10^{-2} mmol) in CD₂Cl₂ (0.10 mL) was added dropwise to a freshly prepared solution of (*E*)-**5b** (1.44 $\times 10^{-2}$ mmol) in CD₂Cl₂ (0.60 mL) at -95 °C and the resulting solution mixed thoroughly for < 1 min to give a yellow solution of (IPr)Au[η^{1} -C(H)($SCH_{2}CH_{2}CH_{2}CH_{2}$)=C(H)=C(4-C₆H₄OMe)₂]⁺ OTf⁻ (**10**) in 92 ± 5

yield (¹H NMR). The yield of **10** was determined by integrating the vinylic H₁ resonance of **10** at δ 5.61 relative to the resonance of CH₂Br₂ at δ 4.96 in the ¹H NMR spectrum. ¹H NMR (500 MHz; CD₂Cl₂; 0 °C): δ 7.56 (t, *J* = 7.9 Hz, 2H), 7.38 (d, *J* = 7.0 Hz, 2H), 7.37 (d, *J* = 7.8 Hz, 2H), 7.33 (s, 2H), 6.89 (d, *J* = 8.8 Hz, 2H), 6.77 (d, *J* = 8.8 Hz, 2H), 6.70 (d, *J* = 8.6 Hz, 2H), 6.56 (d, *J* = 8.6 Hz, 2H), 5.60 (d, *J* = 11.5 Hz, 1H), 3.79 (s, 3H), 3.77 (s, 3H), 3.22 (d, *J* = 11.5 Hz, 1H), 2.79 – 2.72 (m, 1H), 2.72 – 2.61 (m, 1H), 2.49 (h, *J* = 7.3 Hz, 4H), 2.08 - 1.95 (m, 1H), 1.81 - 1.72 (m, 1H), 1.35 - 1.20 (m, 24H). ¹³C{¹H} NMR (125 MHz; CD₂Cl₂; -80 °C): δ 185.8, 174.9, 157.7, 157.4, 145.3, 145.0, 137.3, 133.6, 133.0, 132.3, 130.6, 130.1, 128.7, 127.4, 123.9, 123.7, 123.5, 122.9, 120.6, 118.3, 112.9, 112.5, 54.9, 54.9, 49.7, 49.0, 43.9, 43.0, 31.3, 30.7, 28.6, 28.1, 24.7, 24.2, 23.1, 22.9, 22.8.

Reaction of (*E***)-5b with pyridine.** A solution of pyridine (3.3 mg; 4.21×10^{-2} mmol) in CD₂Cl₂ (0.10 mL) was added dropwise with agitation to a freshly prepared solution of (*E*)-**5b** (1.30 × 10⁻² mmol) in CD₂Cl₂ (0.60 mL) at -95 °C for <1 min to give a bright yellow solution of (IPr)Au[n¹-C(H)(NC₅H₅)=C(H)=C(4-C₆H₄OMe)₂]⁺ OTf⁻ (**11**) in 94 ± 5% NMR yield. The yield of **11** was determined by integrating the vinylic proton resonance of **11** at δ 5.80 relative to the resonance of CH₂Br₂ at δ 4.96 in the ¹H NMR spectrum. ¹H NMR (500 MHz; CD₂Cl₂; -20 °C): δ 8.23 (t, *J* = 7.8 Hz, 1H; py), 8.08 (d, *J* = 6.0 Hz, 2H; py), 7.68 - 7.53 (m, 4H), 7.46 - 7.38 (m, 2H), 7.38 - 7.30 (m, 4H), 6.87 (d, *J* = 8.8 Hz, 2H), 6.72 (d, *J* = 8.3 Hz, 2H), 6.63 (d, *J* = 5.7 Hz, 4H), 5.80 (d, *J* = 10.8 Hz, 1H), 4.90 (d, *J* = 10.7 Hz, 1H), 3.77 (s, 3H), 3.73 (s, 3H), 2.49 (h, *J* = 7.1 Hz, 4H), 1.36 - 1.15 (m, 24H). ¹³C{¹H} NMR (125 MHz; CD₂Cl₂; -80 °C): δ 187.5, 157.5, 157.3, 150.4, 145.5, 145.0, 143.1, 133.8, 133.2, 131.7, 130.5, 130.3, 130. 1, 128.9, 127.7, 126.7, 126.5, 123.9, 123.7, 123.5, 122.8, 121.1, 118.6, 112.7, 112.4, 81.41, 54.8, 28.2, 28.1, 24.7, 24.6, 24.0, 23.2, 22.9, 22.8, 22.8, 21.1.

A concentrated solution (0.13 M) of **11** was layered with ten volumes of diethyl ether at room temperature and then maintained at –20 °C to give yellow crystals of **11** suitable for X-ray analysis. A yellow block-like crystal was used for the X-ray crystallographic analysis.

Conversion of 10 to 11. Pyridine $(1.1 \, \mu L, 1.2 \, \mu mol, 16 \, mM)$ was added to a freshly prepared solution of **10** (17 mM) in CD₂Cl₂ at -80 °C, mixed thoroughly, and placed in the probe of an NMR spectrometer cooled at -80 °C. The probe was warmed at 0 °C and the solution was monitored periodically by ¹H NMR spectroscopy. The concentrations of **10** and **11** were determined by integrating the C2 vinylic protons of **10** and **11** at δ 5.59 and 5.82, respectively, relative to the resonance for CH₂Br₂ at δ 4.96 (Figure S3). After 120 min the ratio of **10**:11 was 1.25, which remained unchanged after -20 °C for 24 h. From these data we determined an equilibrium constant for the conversion of **10** and pyridine to **11** and THT of $K_{eq} = [11]_{eq}[THT]_{eq}/[10]_{eq}[pyridine]_{eq} = 1.8 \pm 0.2$. The values for $[10]_{eq} + [11]_{eq}$ were determined from the ratio of **10**:11 at equilibrium and from the initial concentration of **10** assuming that $[10]_{eq} + [11]_{eq} = [10]_0 = 17 \, \text{mM}$, the value for $[THT]_{eq}$ was determined assuming that $[pyridine]_{eq} = [pyridine]_0 - [11]_{eq}$. A plot of $\ln([10]_{eq}) - [10]_{co}$) versus time was linear to 2.5 half-lives with an observed rate constant of $k_{obs} = k_r + k_r = 6.3 \pm 0.3 \times 10^{-4} \, \text{s}^{-1}$ (Figure S4). From this value and from the equilibrium constant for the conversion of **10** to **11**, we determined values for the forward and reverse first-order rate constants for the conversion of **10** to **11**, we determined values for the forward and reverse first-order rate constants for the conversion of **10** to **11** of $k_f = 4.1 \pm 0.9 \times 10^{-4} \, \text{s}^{-1}$ and $k_r = 2.3 \pm 0.5 \times 10^{-4} \, \text{s}^{-1}$.



Figure S3. Concentration versus time plot for the reaction of **10** (17 mM; O) with pyridine (16 mM) to form **11** (\Box) and THT in CD₂Cl₂ at 0 °C.



Figure S4. First-order plot of approach to equilibrium for the reaction of **10** (17 mM) with pyridine (16 mM) in CD₂Cl₂ at 0 °C.

Reaction of (*E*)-5b with 4-picoline *N*-oxide (4-PNO). A solution of 4-methylpyridine N-oxide (3.1 mg, 2.8×10^{-2} mmol) in CD₂Cl₂ (0.10 mL) was added dropwise with constant agitation to a freshly prepared solution of (*E*)-5b (1.5×10^{-2} mmol) in CD₂Cl₂ (0.60 mL) at –95 °C and mixed thoroughly to give transparent, pale green solution within 1 min. ¹H NMR analysis of the resulting solution revealed a mixture of the known 3,3-bis(4-methoxyphenyl)acrylaldehyde (**12**)^{S8} in 88 ± 5% yield and [(IPr)Au(4-pic)]⁺ OTf⁻ in quantitative yield, both of which were characterized in solution without isolation. The yield of **12** was determined by integrating the aldehyde resonance of **12** at δ 9.37 relative to the resonance of CH₂Br₂ at δ 4.96 in the ¹H NMR spectrum.

For [(IPr)Au(4-pic)]⁺ OTf⁻: ¹H NMR (500 MHz; CD₂Cl₂; 25 °C): δ 7.73 (d, *J* = 5.6 Hz, 2H), 7.59 (t, *J* = 7.8 Hz, 2H), 7.41 (s, 2H), 7.38 (d, *J* = 7.8 Hz, 4H), 7.30 (d, *J* = 5.8 Hz, 2H), 2.56 (h, *J* = 7.0 Hz, 4H), 2.39 (s, 3H), 1.34 (d, *J* = 6.8 Hz, 12H), 1.28 (d, *J* = 6.9 Hz, 12H). ¹³C{¹H} NMR (125 MHz; CD₂Cl₂; 25 °C): δ 168.1, 155.0, 150.2, 146.09, 133.7, 131.5, 127.9, 125.0, 124.9, 29.3, 24.8, 24.1 21.7.

Reaction of (*E***)-5b with triethylsilane**. A solution of triethylsilane (3.0 mg, 2.6×10^{-2} mmol) in CD₂Cl₂ (0.10 mL) was added to a freshly prepared solution of (*E*)-**5b** (1.4×10^{-2} mmol) and CH₂Br₂ (0.83 mg, 4.8 µmol; internal standard) in CD₂Cl₂ (0.60 mL) at –95 °C to give transparent, pale orange solution immediately upon mixing. ¹H NMR analysis of the resulting solution revealed formation of (IPr)Au(η^2 -Et₃SiCH₂C(H)=C(4-C₆H₄OMe)₂]⁺ OTf⁻ (**13**) as the exclusive product in 97% yield as determined by integrating vinyl resonance of **13** at δ 5.99 relative to the resonance of CH₂Br₂ at δ 4.96. Complex **13** was thermally unstable and was therefore characterized by NMR spectroscopy in solution without isolation and through isotopic labelling and independent synthesis (see below).

When the solution was warmed to -40 °C, conversion of **13** to (3,3-bis(4methoxyphenyl)allyl)triethylsilane (**14**) was observed and further warming the solution at 0 °C for 5 min. led to formation of **14** in 100% yield as determined by integrating the vinylic resonance of **14** at δ 6.00 relative to the resonance for CH₂Br₂ at δ 4.96 in the ¹H NMR spectrum. The resulting solution was concentrated and chromatographed (SiO₂; hexanes–EtOAc = 95: 5) to give **14** as colorless oil (3.5 mg, 67%).

Independent synthesis of 13. A solution of 14 (5.0 mg, 1.4×10^{-2} mmol) in CD₂Cl₂ (0.4 mL) was added to a suspension of IPrAuCl (8.4 mg, 1.4×10^{-2} mmol) and AgSbF₆ (5.0 mg, 1.5×10^{-2} mmol) in CD₂Cl₂ (0.3 mL) at -78 °C. ¹H NMR analysis of the resulting solution revealed formation of 13 in ~65% yield, determined by integrating the vinylic proton resonance of 13 at δ 5.98 relative to the resonance of CH₂Br₂ at δ 4.96 in the ¹H NMR spectrum, along with free 14. Complex 13 synthesized in this manner was indistinguishable from 13 generated from the reaction of (*E*)-5b with HSiEt₃. ¹H NMR spectrum of 13 displayed a doublet at δ 5.97 (*J* = 13 Hz) assigned to the vinylic proton and a triplet at δ 1.40 (*J* = 13 Hz) assigned to one of the diastereotopic allylic protons, with the second diastereotopic allylic proton presumably obscured by IPr resonances. This coupling pattern is consistent with strong and coincidentally equivalent (*J* = 13 Hz) coupling between the vinylic proton and the allylic proton at δ 1.40 and between the two allylic protons and weak ($J \le 2$ Hz) coupling between the vinylic proton and the obscured allylic resonance (Figure S5).



Figure S5. Assignment of allylic and vinylic couplings in 13.

For 13: ¹H NMR (500 MHz; CD₂Cl₂; -80 °C): δ 7.59 (t, *J* = 7.7 Hz, 2H), 7.49 (t, *J* = 8.0 Hz, 0.5 H), 7.43 (s, 2H), 7.29 (d, *J* = 8.6 Hz, 4H), 7.27 (d, *J* = 8.3 Hz, 4H), 7.20 (d, *J* = 7.9 Hz, 0.5 H), 5.98 (d, *J* = 12.9 Hz, 1H), 3.73 (s, 3H), 3.73 (s, 3H), 2.23 (sept, *J* = 13.6 Hz, 4H), 1.40 (t, *J* = 13.0 Hz, 1H), 0.52 (t, *J* = 7.8 Hz, 9H), 0.33 – 0.12 (m, 6H). ¹³C{¹H} NMR (125 MHz; CD₂Cl₂; -80 °C): δ 174.0, 159.9, 158.8, 144.8, 144.7, 132.1, 131.2, 131.0, 130.9, 130.6, 128.8, 127.1, 124.0, 123.8, 113.4, 112.9, 103.4, 55.2, 55.1, 49.7, 28.1, 24.0, 23.4, 22.9.

For 14: ¹H NMR (500 MHz; CD₂Cl₂; 25 °C): δ 7.10 (dd, *J* = 8.7, 1.9 Hz, 4H), 6.90 (d, *J* = 8.6 Hz, 2H), 6.78 (d, *J* = 8.8 Hz, 2H), 6.00 (t, *J* = 8.7 Hz, 1H), 3.84 (s, 3H), 3.78 (s, 3H), 1.65 (d, *J* = 8.7 Hz, 2H), 0.87 (t, *J* = 7.9 Hz, 9H), 0.52 (q, *J* = 7.9 Hz, 6H). ¹³C{¹H} NMR (125 MHz; CD₂Cl₂; 25 °C): δ 158.43, 158.36, 138.50, 136.91, 133.20, 131.51, 128.14, 124.80, 113.75, 113.60, 55.46, 55.40, 15.92, 7.54, 3.59. HRMS (ESI) calcd. (found) for C₂₃H₃₂O₂Si (M)⁺: 369.2244 (369.2243).

Reactions of (*E***)-5b with DSiEt₃ and (***E***)-5b-***d***₁ with HSiEt₃. A solution of DSiEt₃ (3.4 mg, 2.9 × 10^{-2} mmol) in CD₂Cl₂ (0.10 mL) was added to a freshly prepared solution of (***E***)-5b (1.4 \times 10^{-2} mmol) in CD₂Cl₂ (0.60 mL) containing CH₂Br₂ (4.7 \times 10^{-3} mmol) at –95 °C to give transparent, pale orange mixture of (IPr)Au(\eta^2-Et₃SiCHDC(H)=C(4-C₆H₄OMe)₂]⁺ OTf⁻ (13**-*d*₁) in quantitative yield. In a separate experiment, a solution of HSiEt₃ (2.8 µL) in CD₂Cl₂ was added via syringe to a solution of (*E*)-5b-*d*₁ (11.9

mg, 1.4×10^{-2} mmol) in CD₂Cl₂ containing CH₂Br₂ (4.3×10^{-3} mmol) to form **13**-*d*₁ in 95% yield, which was indistinguishable from **13**-*d*₁ generated in the reaction of (*E*)-**5b** with DSiEt₃. The ¹H NMR spectrum of **13**-*d*₁ displayed a broad singlet at δ 5.97 assigned to the vinylic proton and a doublet at δ 1.40 (*J* = 13 Hz) assigned to one of the diastereotopic allylic protons, which integrated to 0.5 H relative to the vinylic resonance at δ 5.97. This observation is consistent with exclusive incorporation of deuterium into the allylic position of **13**-*d*₁ with the deuterium atom distributed equally between the diastereomeric positions.

Warming a solution of **13**- d_1 to 0 °C led to quantitative formation of **14**- d_1 with ≥95% deuterium incorporation at the allylic position as determined by integrating the allylic resonance at δ 1.63 relative to the vinylic resonance at δ 5.99.

For 13-*d*₁: ¹H NMR (CD₂Cl₂, –80 °C, selected resonances): δ 5.97 (br s, 1H), 1.38 (d, *J* = 13.0 Hz, 0.5H).

For 14-*d*₁: ¹H NMR (CD₂Cl₂, 25 °C, selected resonances): δ 5.99 (d, *J* = 8.7 Hz, 1H), 1.63 (d, *J* = 8.6 Hz, 1H).

Reaction of (E)-5b with 4-methoxystyrene. A solution of 4-methoxystyrene (6.5 mg, 4.8×10^{-2} mmol) in CD₂Cl₂ (0.10 mL) was added dropwise via syringe with constant agitation to a freshly prepared solution of (*E*)-**5b** (1.6×10^{-2} mmol) in CD₂Cl₂ (0.60 mL) at -95 °C to generate a transparent red solution. ¹H NMR analysis of the crude reaction mixture at -90 °C showed a distinct resonance at δ 4.99 (*J* = 10.2 Hz, 1H) assigned to the vinyl proton of **15**, which corresponded to a 75% yield from (*E*)-**5b** as determined by integrating vinylic resonance of **15** at δ 4.99 relative to the resonance of CH₂Br₂ at δ 4.96. Upon warming the solution above -40 °C, decomplexation of *cis*-**15** was observed with the concomitant appearance of **16**, as indicated by the appearance of a resonance at 5.44 (d, *J* = 9.6 Hz, 1H) assigned to the vinylic resonance of **16**. However, direct integration of this resonance was precluded by the presence of both excess 4-methoxystyrene and byproducts, presumably resulting from polymerization of 4-methoxystyrene. Therefore, the reaction mixture was filtered through a thin pad of silica gel, eluted with hexanes–EtOAc, and concentrated under vacuum. ¹H NMR analysis of the resulting residue established

the presence of **16** along with minor impurities. In a separate experiment, 4-methoxystyrene (9.2 mg, 6.8×10^{-2} mmol) was added dropwise with stirring to a freshly prepared solution of (*E*)-**5b** (1.6×10^{-2} mmol) and anthracene (2.3 mg, 1.3×10^{-2} mmol; internal standard) in CH₂Cl₂ (3.0 mL) at –95 °C and the resulting mixture was warmed slowly to room temperature. The crude reaction mixture was filtered through silica gel with hexanes–EtOAc and analyzed by ¹H NMR spectroscopy, which revealed formation of **16** in 67% yield as a ~3:1 mixture of cis/trans isomers as determined by integrating the vinylic proton resonance of *cis*-**16** at δ 5.44 and *trans*-**16** at δ 5.17 relative to the H9/H10 proton resonance of anthracene at δ 8.44. The solvent was evaporated and the resulting residue was chromatographed (SiO₂; hexanes–EtOAc = 9:1) to give **16** (8.4 mg, 51%, 3:1 cis/trans) as a colorless oil.

The cis and trans isomers of **16** (*cis*-**16** and *trans*-**16**, respectively) were unambiguously assigned by ¹H-¹H NOESY analysis. In particular, *cis*-**16** displayed a strong cross peak correlating the benzylic proton resonance at δ 2.03 (ddd, *J* = 8.7, 5.8, 4.3 Hz, 1H) with the allylic proton resonance at δ 1.75 (dddd, *J* = 9.8, 8.4, 5.6, 4.3 Hz, 1H), which was absent for *trans*-**16** (Figure S6).

Spectroscopic analysis was performed on a 3:1 mixture of *cis*-**16** and *trans*-**16** without further purification.

For *cis*-16: ¹H NMR (500 MHz; CDCl₃; 25 °C): δ 7.19 (d, *J* = 8.6 Hz, 2H), 7.15 (d, *J* = 8.8 Hz, 2H), 6.96 (d, *J* = 8.6 Hz, 2H), 6.84 (d, *J* = 8.7 Hz, 2H), 6.81 (d, *J* = 7.5 Hz, 2H), 6.79 (d, *J* = 7.4 Hz, 2H), 5.44 (d, *J* = 9.6 Hz, 1H), 3.81 (s, 3H), 3.80 (s, 3H), 3.78 (s, 3H), 2.03 (ddd, *J* = 8.7, 5.8, 4.3 Hz, 1H), 1.75 (dddd, *J* = 9.8, 8.4, 5.6, 4.3 Hz, 1H), 1.19 - 1.10 (m, 2H). ¹³C{¹H} NMR (125 MHz; CDCl₃; 25 °C): δ 158.82 (*p*), 158.64 (*p*), 157.88 (*p*), 139.73 (*ipso*), 136.04 (*ipso*), 134.14 (*ipso*), 132.80 (vinyl), 131.64 (*o*), 128.59 (*o*), 128.46 (vinyl), 127.10 (*o*), 113.96 (*m*), 113.65 (*m*), 113.61 (*m*), 55.44 (CH₃O–), 25.72 (benzylic), 24.85 (allylic), 18.00.

For trans-16: δ 7.24 (d, J = 8.7 Hz, 2H), 6.92 (dd, J = 8.6, 6.2 Hz, 2H), 6.70 (d, J = 8.8 Hz, 2H), 5.17 (d, J = 9.9 Hz, 1H), 3.81 (s, 11H), 3.85 (s, 3H), 3.74 (s, 3H), 2.28 (td, J = 8.5, 6.3 Hz, 1H), 1.89 (dtd, J = 10.0, 8.5, 5.5 Hz, 1H), 1.31 – 1.24 (m, 2H). Remaining aromatic resonances were obscured by those of *cis*-**16**. ¹³C{¹H} NMR (125 MHz; CDCl₃; 25 °C, distinct resonances only): δ 158.07, 140.52, 136.15, 133.24, 131.17, 130.93, 130.39, 128.46, 127.98, 23.90, 20.33, 13.86.

For *cis*-16/*trans*-16: HRMS (ESI) calcd. (found) for C₂₆H₂₆O₃(M)⁺: 387.1955 (387.1952).



Figure S6. Partial ¹H-¹H NOESY spectrum of a 3:1 mixture of *cis*-12 and *trans*-12.

Independent synthesis of 15. A solution of 16 (3.0 mg, 7.8×10^{-3} mmol; *cis:trans* = 3:1) in CD₂Cl₂ (0.35 mL) was added to a suspension of IPrAuCl (4.8 mg, 7.8×10^{-3} mmol) and AgSbF₆ (3.0 mg, 8.7×10^{-3} mmol) in CD₂Cl₂ (0.3 mL) at -80 °C to give dark purple solution. Although the ¹H NMR spectrum of the resulting was broadened, distinct resonances were observed for the vinylic protons of 15 and *cis*-16 at δ 5.54 (*J* = 9.6 Hz) and δ 4.99 (*J* = 10.2 Hz), respectively, in ~4:1 ratio. Upon gradual warming of

solution above –40 °C to room temperature over 5 h, the vinylic resonances for both **15** and *cis*-**16** were significantly reduced in intensity and eventually disappeared to give deep blue solution.

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Figure S7. ¹H NMR spectrum of (*E*)-**8b** (CDCl₃, 25 °C).



Figure S8. ¹³C{¹H} NMR spectrum of (E)-8b (CDCl₃, 25 °C).



Figure S9. ¹H NMR spectrum of (*E*)-**8a** (CDCl₃, 25 °C).



















Figure S14. ¹³C{¹H} NMR spectrum of (*E*)-**6a** (CDCl₃, 25 °C).



Figure S15. ¹H NMR spectrum of (*E*)-6b (CDCl₃, 25 °C).



Figure S16. ¹³C{¹H} NMR spectrum of (E)-6b (CDCl₃, 25 °C).



Figure S17. ¹H NMR spectrum of (*E*)-**6c** (CDCl₃, 25 °C).



Figure S18. ¹³C{¹H} NMR spectrum of (*E*)-6c (CDCl₃, 25 °C).







Figure S20. ¹³C{¹H} NMR spectrum of (*Z*)-6b (CDCl₃, 25 °C).





Figure S22. ¹³C{¹H} NMR spectrum of (*E*)-**5b** (CD₂Cl₂, –95 °C). Additional peaks observed at δ 21.3 (CH₂Br₂; internal standard), δ 49.6 (TMSOMe), –2.1 (TMSOMe), δ –0.5 (TMSOTf).









Figure S24. ¹³C{¹H} NMR spectrum of (*E*)-**5c** (CD₂Cl₂, -90 °C).



Figure S25. ¹H NMR spectrum of (*E*)-**5a** (CD₂Cl₂, -110 °C).







Figure S27. $^{13}C{}^{1}H$ NMR spectrum of 9 (CDCl₃, 25 °C).

Figure S28. ¹H NMR spectrum of **10** (CD₂Cl₂, 0 °C). Additional peaks observed at δ 4.96 (s, CH₂Br₂; internal standard), δ 3.30 (s, TMSOMe), 0.02 (s, TMSOMe).





Figure S29. ¹³C{¹H} NMR spectrum of **10** (CD₂Cl₂, -80 °C).

Figure S30. ¹H NMR spectrum of **11** (CD₂Cl₂, -20 °C). Additional peaks observed at δ 4.96 (s, CH₂Br₂; internal standard), δ 3.30 (s, TMSOMe), 0.02 (s, TMSOMe).





Figure S31. ¹³C{¹H} NMR spectrum of **11** (CD₂Cl₂, -80 °C).









Figure S34. ¹H NMR spectrum of **13** (CD₂Cl₂, –80 °C). Additional peaks observed at δ 4.96 (s, CH₂Br₂; internal standard), δ 3.30 (s, TMSOMe), δ 0.43 (s, TMSOTf), 0.02 (s, TMSOMe).



Figure S35. ¹³C{¹H} NMR spectrum of **13** (CD₂Cl₂, -80 °C). Additional peaks observed at δ 21.3 (CH₂Br₂; internal standard), δ 49.6 (TMSOMe), -2.1 (TMSOMe), δ -0.5 (TMSOTf).





Figure S36. ¹H NMR spectrum of **14** (CDCl₃, 25 °C).



Figure S37. $^{13}C{^1H}$ NMR spectrum of 14 (CDCl₃, 25 °C).



Figure S38. ¹H NMR spectrum of a 3:1 mixture of *cis*-16 and *trans*-16 (CDCl₃, 25 °C).



Figure S39. ¹³C{¹H} NMR spectrum of **16** (CDCl₃, 25 °C).