

Supporting Information

Enantioselective Photoredox Dehalogenative Protonation

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1. General information

General procedures and methods

Experiments involving moisture and/or air sensitive components were performed under a positive pressure of argon in oven-dried glassware equipped with a rubber septum inlet. Dried solvents and liquid reagents were transferred by oven-dried syringes or hypodermic syringe cooled to ambient temperature in a desiccator. Reaction mixtures were stirred in 10 mL sample vial with Teflon-coated magnetic stirring bars unless otherwise stated. Moisture in non-volatile reagents/compounds was removed in high *vacuo* by means of an oil pump and subsequent purging with nitrogen. Solvents were removed *in vacuo* under ~30 mmHg and heated with a water bath at 30–35 °C using rotary evaporator with aspirator. The condenser was cooled with running water at 0 °C.

All experiments were monitored by analytical thin layer chromatography (TLC). TLC was performed on pre-coated plates, 60 F₂₅₄. After elution, plate was visualized under UV illumination at 254 nm and 365 nm for UV active material. Further visualization was achieved by staining phosphomolybdic acid and anisaldehyde solution. For those using the aqueous stains, the TLC plates were heated on a hot plate.

Columns for flash chromatography (FC) contained *silica gel* 200–300 mesh. Columns were packed as slurry of *silica gel* in petroleum ether and equilibrated solution using the appropriate solvent system. The elution was assisted by applying pressure of about 2 atm with an air pump.

Instrumentations

Proton nuclear magnetic resonance (¹H NMR), carbon NMR (¹³C NMR) and fluorine (¹⁹F NMR) spectra were recorded in CDCl₃ otherwise stated. Chemical shifts are reported in parts per million (ppm), using the residual solvent signal as an internal standard: CDCl₃ (¹H NMR: δ 7.26, singlet; ¹³C NMR: δ 77.0, triplet). Coupling constants (*J*) were recorded in Hertz (Hz). The number of proton atoms (*n*) for a given resonance was indicated by *n*H. The number of carbon atoms (*n*) for a given resonance was indicated by *n*C. HRMS (Analyzer: TOF) was reported in units of mass of charge ratio (*m/z*). Mass samples were dissolved in CH₃CN (HPLC Grade) unless otherwise stated. Optical rotations were recorded on a polarimeter with

a sodium lamp of wavelength 589 nm and reported as follows; $[\alpha]_{\lambda}^{T^{\circ}C}$ (c = g/100 mL, solvent). Melting points were determined on a melting point apparatus.

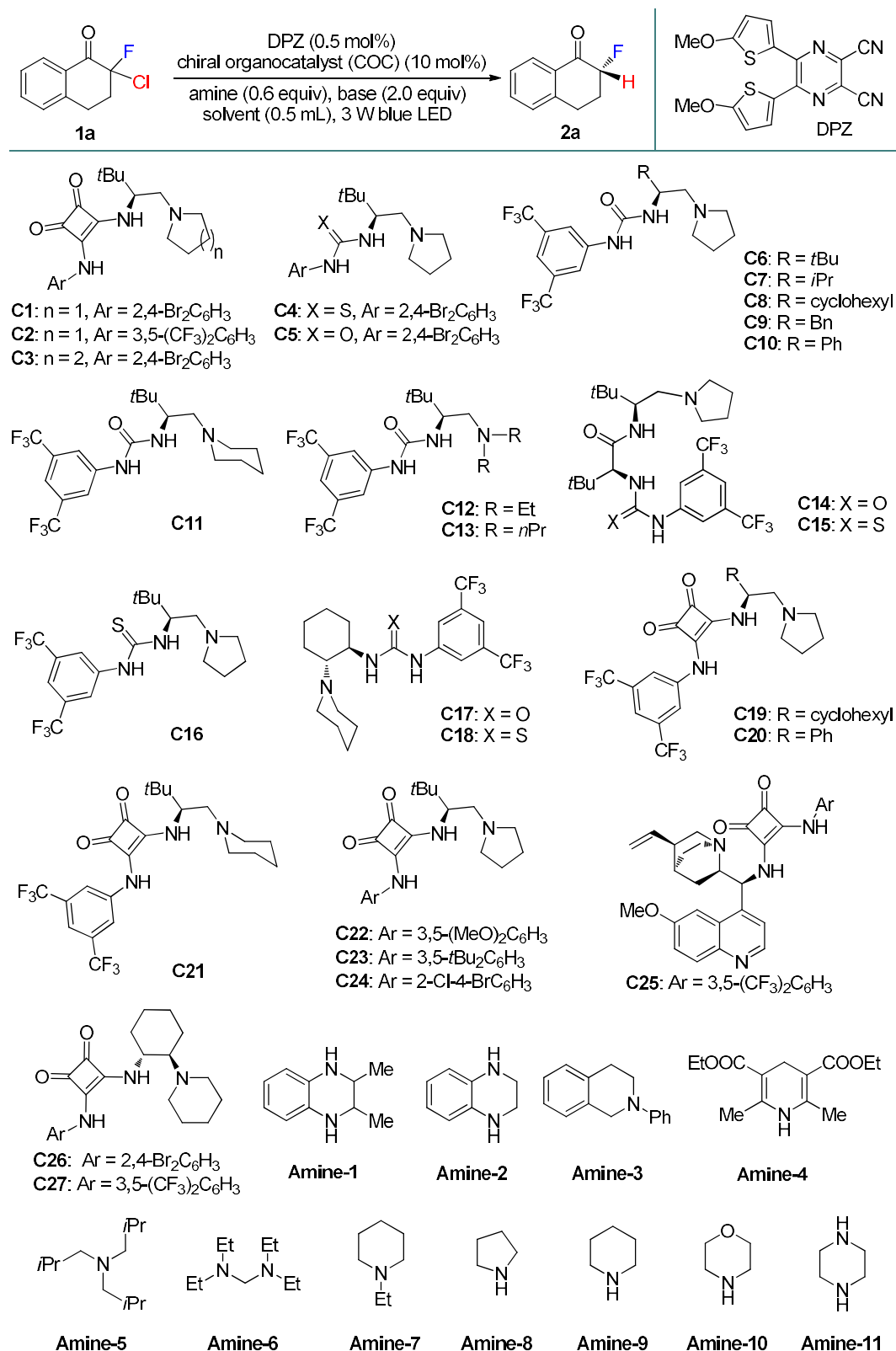
Enantiomeric excesses were determined by chiral High Performance Liquid Chromatography (HPLC) analysis. UV detection was monitored at 254 nm, 230 nm and 210 nm at the same time. HPLC samples were dissolved in HPLC grade isopropanol (IPA) unless otherwise stated.

Materials

All commercial reagents were purchased with the highest purity grade. They were used without further purification unless specified. All solvents used, mainly petroleum ether (PE) and ethyl acetate (EtOAc) were distilled. Anhydrous dichloromethane (DCM), 1,2-dichloroethane (DCE) and chloroform (CHCl_3) were freshly distilled from CaH_2 and stored under N_2 atmosphere. THF, Et_2O and toluene were freshly distilled from sodium/benzophenone before use. Other solvent were purchased with the highest purity grade and without further treatment. Substrates **1**, **3**, **4** and **7** were prepared according to the relevant literatures.¹⁻⁴ **D-Amine-1** was prepared according to the relevant literature.⁵ All compounds synthesized were stored in a $-20\text{ }^{\circ}\text{C}$ freezer and light-sensitive compounds were protected with aluminium foil.

2. Optimization of reaction conditions

Table S1. Optimization of Reaction Conditions for Enantioselectivity of Approaching Chiral Secondary α -Fluoroketones^a

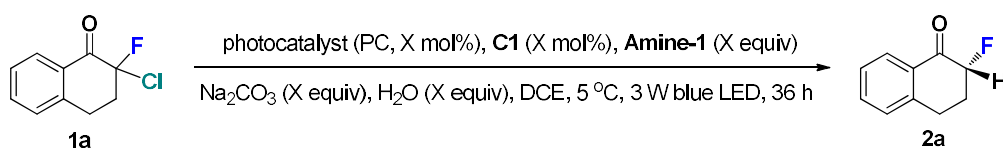


entry	COC	amine	base	solvent	<i>T</i> (°C)	ee (%) ^b
1	C6	Et ₃ N	Na ₂ CO ₃	PhBr	25	12
2	C6	DIPEA	Na ₂ CO ₃	PhBr	25	49
3	C6	Amine-3	Na ₂ CO ₃	PhBr	25	0
4	C6	Amine-4	Na ₂ CO ₃	PhBr	25	26
5	C6	Amine-5	Na ₂ CO ₃	PhBr	25	20
6	C6	Amine-6	Na ₂ CO ₃	PhBr	25	−4
7	C6	Amine-7	Na ₂ CO ₃	PhBr	25	27
8	C6	Amine-8	Na ₂ CO ₃	PhBr	25	0
9	C6	Amine-9	Na ₂ CO ₃	PhBr	25	2
10	C6	Amine-10	Na ₂ CO ₃	PhBr	25	31
11	C6	Amine-11	Na ₂ CO ₃	PhBr	25	45
12	C6	Amine-2	Na ₂ CO ₃	PhBr	25	73
13	C6	Amine-2	NaF	PhBr	25	18
14	C6	Amine-2	Na ₂ SO ₃	PhBr	25	22
15	C6	Amine-2	PhCOONa	PhBr	25	27
16	C6	Amine-2	NaHCO ₃	PhBr	25	22
17	C6	Amine-2	NaOAc	PhBr	25	38
18	C6	Amine-2	EtONa	PhBr	25	61
19	C6	Amine-2	Na ₂ HPO ₄	PhBr	25	43
20	C6	Amine-2	Na ₃ PO ₄	PhBr	25	68
21	C6	Amine-2	PhSO ₂ Na	PhBr	25	68
22	C6	Amine-2	KF	PhBr	25	69
23	C6	Amine-2	KOAc	PhBr	25	49
24	C6	Amine-2	K ₂ HPO ₄	PhBr	25	67
25	C6	Amine-2	K ₂ CO ₃	PhBr	25	62
26	C6	Amine-2	K ₃ PO ₄	PhBr	25	30
27	C6	Amine-2	Cs ₂ CO ₃	PhBr	25	15
28	C6	Amine-2	Li ₂ CO ₃	PhBr	25	32
29	C6	Amine-2	Li ₃ PO ₄	PhBr	25	13
30	C7	Amine-2	Na ₂ CO ₃	PhBr	25	60
31	C8	Amine-2	Na ₂ CO ₃	PhBr	25	51
32	C9	Amine-2	Na ₂ CO ₃	PhBr	25	35
33	C10	Amine-2	Na ₂ CO ₃	PhBr	25	34
34	C11	Amine-2	Na ₂ CO ₃	PhBr	25	62
35	C12	Amine-2	Na ₂ CO ₃	PhBr	25	11
36	C13	Amine-2	Na ₂ CO ₃	PhBr	25	43
37	C14	Amine-2	Na ₂ CO ₃	PhBr	25	52
38	C15	Amine-2	Na ₂ CO ₃	PhBr	25	60
39	C16	Amine-2	Na ₂ CO ₃	PhBr	25	78
40	C17	Amine-2	Na ₂ CO ₃	PhBr	25	−32
41	C18	Amine-2	Na ₂ CO ₃	PhBr	25	−54
42	C2	Amine-2	Na ₂ CO ₃	PhBr	25	81
43	C19	Amine-2	Na ₂ CO ₃	PhBr	25	76

44	C20	Amine-2	Na ₂ CO ₃	PhBr	25	50
45	C21	Amine-2	Na ₂ CO ₃	PhBr	25	8
46	C22	Amine-2	Na ₂ CO ₃	PhBr	25	16
47	C23	Amine-2	Na ₂ CO ₃	PhBr	25	75
48	C24	Amine-2	Na ₂ CO ₃	PhBr	25	82
49	C25	Amine-2	Na ₂ CO ₃	PhBr	25	46
50	C26	Amine-2	Na ₂ CO ₃	PhBr	25	48
51	C27	Amine-2	Na ₂ CO ₃	PhBr	25	44
52	C1	Amine-2	Na ₂ CO ₃	PhBr	25	84
53	C1	Amine-1	Na ₂ CO ₃	PhBr	25	85
54	C1	Amine-1	Na ₂ CO ₃	toluene	25	73
55	C1	Amine-1	Na ₂ CO ₃	PhCl	25	82
56	C1	Amine-1	Na ₂ CO ₃	PhCF ₃	25	82
57	C1	Amine-1	Na ₂ CO ₃	C ₆ F ₅ H	25	81
58	C1	Amine-1	Na ₂ CO ₃	<i>t</i> BuPh	25	68
59	C1	Amine-1	Na ₂ CO ₃	anisole	25	80
60	C1	Amine-1	Na ₂ CO ₃	2-bromoanisole	25	83
61	C1	Amine-1	Na ₂ CO ₃	3-bromoanisole	25	84
62	C1	Amine-1	Na ₂ CO ₃	4-bromoanisole	25	85
63	C1	Amine-1	Na ₂ CO ₃	2-fluoroanisole	25	84
64	C1	Amine-1	Na ₂ CO ₃	3-fluoroanisole	25	84
65	C1	Amine-1	Na ₂ CO ₃	4-fluoroanisole	25	85
66	C1	Amine-1	Na ₂ CO ₃	DCM	25	73
67	C1	Amine-1	Na ₂ CO ₃	DCE	25	90
68	C1	Amine-1	Na ₂ CO ₃	CHCl ₃	25	80
69	C1	Amine-1	Na ₂ CO ₃	THF	25	55
70	C1	Amine-1	Na ₂ CO ₃	Et ₂ O	25	64
71	C1	Amine-1	Na ₂ CO ₃	DCE	15	91
72	C1	Amine-1	Na ₂ CO ₃	DCE	10	93
73	C1	Amine-1	Na ₂ CO ₃	DCE	5	94 ^c
74	C1	Amine-1	Na ₂ CO ₃	DCE	0	94 ^d
75	C1	Amine-1	Na ₂ CO ₃	DCE	-5	93
76	C1	Amine-1	Na ₂ CO ₃	DCE	-10	93

^a0.05 mmol scale. For catalyst C3-C5, we attempted their viability under the standard reaction conditions. The results could be checked in Table 1 of the manuscript. ^bDetermined by HPLC analysis on a chiral stationary phase. 36 h, ^cYield = 72%. 36 h, ^dYield = 52%.

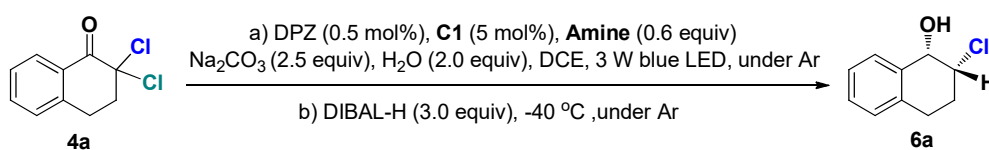
Table S2. Optimization of Reaction Conditions for Isolated Yield of Approaching Chiral Secondary α -Fluoroketones^a



entry	PC (X mol%)	mol% of C1	equiv of Amine-1	equiv of Na ₂ CO ₃	equiv of H ₂ O	yield (%) ^b	ee (%) ^c
1	DPZ (0.2)	10	0.6	2.0	--	68	94
2	DPZ (1.0)	10	0.6	2.0	--	73	92
3	DPZ (2.0)	10	0.6	2.0	--	72	92
4	DPZ (0.5)	10	0.6	1.5	--	71	91
5	DPZ (0.5)	10	0.6	2.5	--	74	94
6	DPZ (0.5)	10	0.6	3.0	--	72	91
7	DPZ (0.5)	10	0.4	2.5	--	67	93
8	DPZ (0.5)	10	1.0	2.5	--	74	95
9	DPZ (0.5)	10	2.0	2.5	--	74	93
10	DPZ (0.5)	15	0.6	2.5	--	73	92
11	DPZ (0.5)	5	0.6	2.5	--	74	95
12	DPZ (0.5)	5	0.6	2.5	1.0	92	96
13	DPZ (0.5)	5	0.6	2.5	2.0	91	96
14	DPZ (0.5)	5	0.6	2.5	5.0	89	96
15	DPZ (0.5)	5	0.6	2.5	10.0	72	96
16	Rose Bangel (0.5)	5	0.6	2.5	1.0	54	95
17	Eosin Y (0.5)	5	0.6	2.5	1.0	47	93
18	Ru(bpy) ₃ Cl ₂ ·6H ₂ O (0.5)	5	0.6	2.5	1.0	87	95
19	Ir(ppy) ₂ (dtbbpy)PF ₆ (0.5)	5	0.6	2.5	1.0	75	90
21	[Acr-Mes]ClO ₄ (0.5)	5	0.6	2.5	1.0	44	84

^a0.05 mmol scale. ^bYield of isolated product. ^cDetermined by HPLC analysis on a chiral stationary phase.

Table S3. Optimization of Reaction Conditions for Chiral α -Chlorohydrin^a



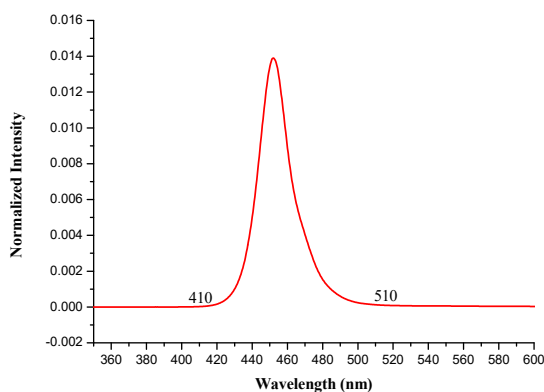
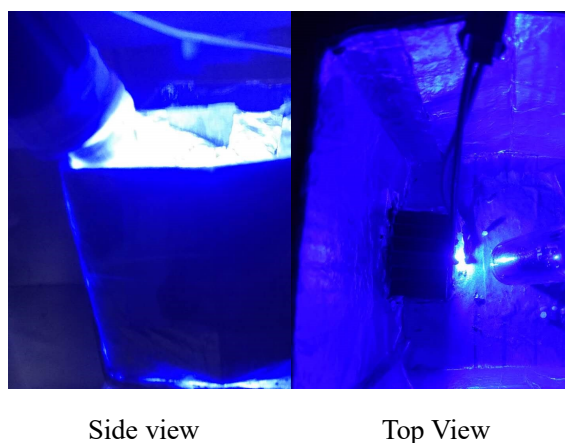
entry	amine	temperature	dr	yield (%) ^b	ee (%) ^c
1	Amine-1	0 °C	>20:1	62%	92%
2	Amine-2	0 °C	>20:1	73%	89%
3	Amine-2	-30 °C	>20:1	79%	92%

^a0.1 mmol scale. ^bYield of isolated product. ^cDetermined by HPLC analysis on a chiral stationary phase.

3. General experimental procedures

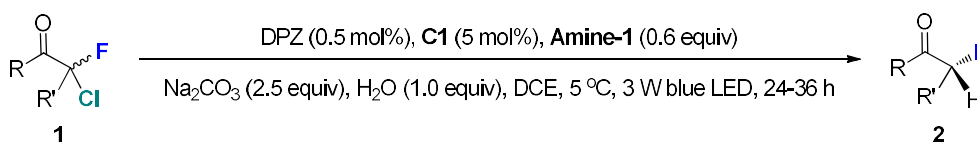
(1) Reaction setup

10 mL Schlenk tube is placed at the center of a stir plate. A 3 W blue LED lamp (HW-450-455LED-3W) is placed to one sidewall of reaction tube (at approximately 2 cm away from the light source). The transformations were conducted in a cryostat which allows a stable and certain temperature.



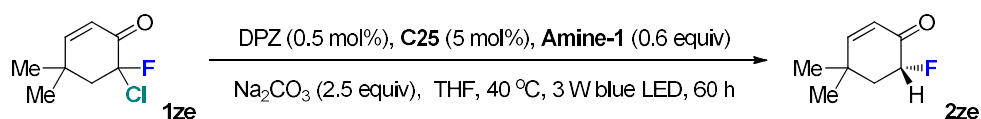
Emission spectrum of the 3 W LED light.

(2) General procedure for the synthesis of chiral secondary α -fluoketones



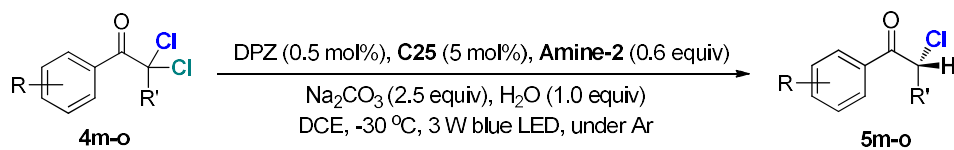
35 μL (0.0005 mmol, 0.005 equiv) of DPZ solution (1.0 mg of DPZ in 200 μL of anhydrous toluene) was added into a 10 mL Schlenk tube, and then solvent was removed in *vacuo*. Subsequently, **C1/C25/C26** (0.005 mmol, 0.05 equiv), Na_2CO_3 (0.25 mmol, 2.5 equiv),

Amine-1 (0.06 mmol, 0.6 equiv), **1** (0.1 mmol, 1.0 equiv), distilled H₂O (0.1 mmol, 1.0 equiv), stir bar and DCE (1.0 mL) were added sequentially and then degassed for three times by freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at 5 °C for 30 min without light, then irradiated by a 3 W blue LED ($\lambda = 450\text{--}455$ nm) from a 2.0 cm distance for another 24–36 hours. The reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with petroleum ether/dichloromethane (5/1–2/1 ratio). Removing the solvent in *vacuo*, afforded products **2a–zd**.



35 μ L (0.0005 mmol, 0.005 equiv) of DPZ solution (1.0 mg of DPZ in 200 μ L of anhydrous toluene) was added into a 10 mL Schlenk tube, and then solvent was removed in *vacuo*. Subsequently, **C25** (0.005 mmol, 0.05 equiv), Na₂CO₃ (0.25 mmol, 2.5 equiv), **Amine-2** (0.06 mmol, 0.6 equiv), **1ze** (0.1 mmol, 1.0 equiv), stir bar and THF (1.0 mL) were added sequentially and then degassed for three times by freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at 40 °C for 30 min without light, then irradiated by a 3 W blue LED ($\lambda = 450\text{--}455$ nm) from a 2.0 cm distance for another 60 hours. The reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with petroleum ether/ ethyl acetate (40/1–10/1 ratio). Removing the solvent in *vacuo*, afforded products **2ze**.

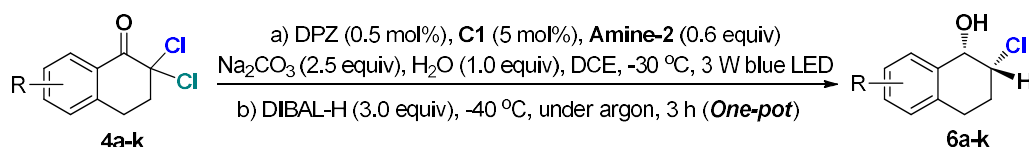
(3) General procedure for the synthesis of chiral secondary α -chloroketones



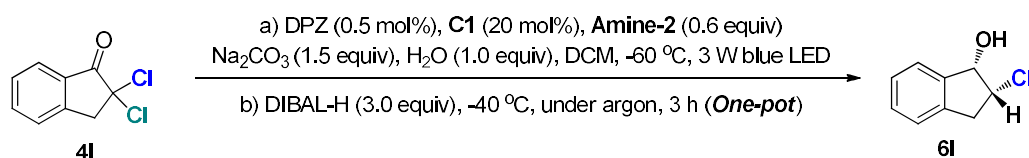
35 μ L (0.0005 mmol, 0.005 equiv) of DPZ solution (1.0 mg of DPZ in 200 μ L of anhydrous toluene) was added into a 10 mL Schlenk tube, and then solvent was removed in *vacuo*. Subsequently, **C25** (0.005 mmol, 0.05 equiv), Na₂CO₃ (0.25 mmol, 2.5 equiv), **Amine-2** (0.06 mmol, 0.6 equiv), **4** (0.1 mmol, 1.0 equiv), distilled H₂O (0.1 mmol, 1.0 equiv), stir bar and DCE (1.0 mL) were added sequentially and then degassed for three times by freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at

–30 °C for 30 min without light, then irradiated by a 3 W blue LED ($\lambda = 450\text{--}455$ nm) from a 2.0 cm distance for another 22–42 hours until **4** exhausted monitored by TLC. The reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with petroleum ether/dichloromethane (5/1–2/1 ratio). Removing the solvent in *vacuo*, afforded products **5m-o**.

(4) General procedure for the synthesis of chiral α -chlorohydrins



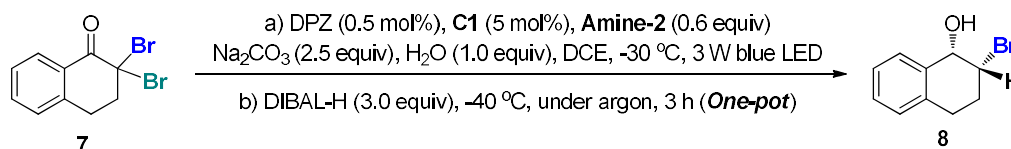
35 μL (0.0005 mmol, 0.005 equiv) of DPZ solution (1.0 mg of DPZ in 200 μL of anhydrous toluene) was added into a 10 mL Schlenk tube, and then solvent was removed in *vacuo*. Subsequently, **C1** (0.005 mmol, 0.05 equiv), Na_2CO_3 (0.25 mmol, 2.5 equiv), **Amine-2** (0.06 mmol, 0.6 equiv), **4a-k** (0.1 mmol, 1.0 equiv), distilled H_2O (0.1 mmol, 1.0 equiv), stir bar and DCE (1.0 mL, **4b** for 2 mL) were added sequentially and then degassed for three times by freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at –30 °C for 30 min without light, then irradiated by a 3 W blue LED ($\lambda = 450\text{--}455$ nm) from a 2.0 cm distance for another 5–18 hours until **4a-k** exhausted monitored by TLC. Then 3.0 equiv DIBAL-H (1.0 M in toluene) was added slowly at –40 °C under an argon atmosphere. After stirring for 3 hours, added two drops of H_2O and warmed to room temperature. The reaction mixture was filtered and washed with DCM (5.0 mL), concentrated in *vacuo*, followed by gradient elution with petroleum ether/dichloromethane (5/1–2/1 ratio). Removing the solvent in *vacuo*, afforded products **6a–6k**.



35 μL (0.0005 mmol, 0.005 equiv) of DPZ solution (1.0 mg of DPZ in 200 μL of anhydrous toluene) was added into a 10 mL Schlenk tube, and then solvent was removed in *vacuo*. Subsequently, **C1** (0.02 mmol, 0.2 equiv), Na_2CO_3 (0.15 mmol, 1.5 equiv), **Amine-2** (0.06 mmol, 0.6 equiv), **4l** (0.1 mmol, 1.0 equiv), distilled H_2O (0.1 mmol, 1.0 equiv), stir bar and DCM (1.0 mL) were added sequentially and then degassed for three times by

freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at $-60\text{ }^{\circ}\text{C}$ for 30 min without light, then irradiated by a 3 W blue LED ($\lambda = 450\text{--}455\text{ nm}$) from a 2.0 cm distance for another 48 hours until **4l** exhausted monitored by TLC under Ar atmosphere. Then 3.0 equiv DIBAL-H (1.0 M in toluene) was added slowly at $-40\text{ }^{\circ}\text{C}$ under an argon atmosphere. After stirring for 3 hours, added two drops of H_2O and warmed to room temperature. The reaction mixture was filtered and washed with DCM (5 mL), concentrated in *vacuo*, followed by gradient elution with petroleum ether/dichloromethane (5/1–2/1 ratio). Removing the solvent in *vacuo*, afforded products **6l**.

(5) Procedure for the synthesis of chiral secondary α -bromohydrin **8**



35 μL (0.0005 mmol, 0.005 equiv) of DPZ solution (1.0 mg of DPZ in 200 μL of anhydrous toluene) was added into a 10 mL Schlenk tube, and then solvent was removed in *vacuo*. Subsequently, **C1** (0.005 mmol, 0.05 equiv), Na_2CO_3 (0.25 mmol, 2.5 equiv), **Amine-2** (0.06 mmol, 0.6 equiv), **7** (0.1 mmol, 1.0 equiv), distilled H_2O (0.1 mmol, 1.0 equiv), stir bar and DCE (1.0 mL) were added sequentially and then degassed for three times by freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at $-30\text{ }^{\circ}\text{C}$ for 30 min without light, then irradiated by a 3 W blue LED ($\lambda = 450\text{--}455\text{ nm}$) from a 2.0 cm distance for another 2 hours until **7** exhausted monitored by TLC. Then 3.0 equiv DIBAL-H (1.0 M in toluene) was added slowly at $-40\text{ }^{\circ}\text{C}$ under an argon atmosphere. After stirring for 3 hours, added two drops of H_2O and warmed to room temperature. The reaction mixture was filtered and washed with DCM (5.0 mL), concentrated in *vacuo*. followed by gradient elution with petroleum ether/dichloromethane (5/1–2/1 ratio). Removing the solvent in *vacuo*, afforded products **8**.

Note: All racemic samples for determining HPLC conditions and ee values of chiral compounds were prepared through: 0.5 mol% DPZ, 0.6 equiv. of **Amine-1**, 2.5 equiv. of Na_2CO_3 in DCE at $25\text{ }^{\circ}\text{C}$ (for racemic products **2** and **5m-o**); 0.5 mol% DPZ, 0.6 equiv. of **Amine-1**, 2.5 equiv. of Na_2CO_3 in DCE at $25\text{ }^{\circ}\text{C}$. After 6 h, 3.0 equiv. of DIBAL-H was added and the transformations were conducted at $-40\text{ }^{\circ}\text{C}$ (for racemic products **6** and **8**)

4. Mechanism studies

Emission quenching experiments

Emission intensities were recorded on a spectrofluorometer. DPZ solution was excited at 448 nm and the emission intensity at 544 nm was observed. A solution of DPZ (2.5×10^{-4} M) in MeCN was added to the appropriate amount of quencher in 5.0 mL volumetric flask under N_2 . The solution was transferred to a 1.5 mL quartz cell and the emission spectrum of the sample was collected.

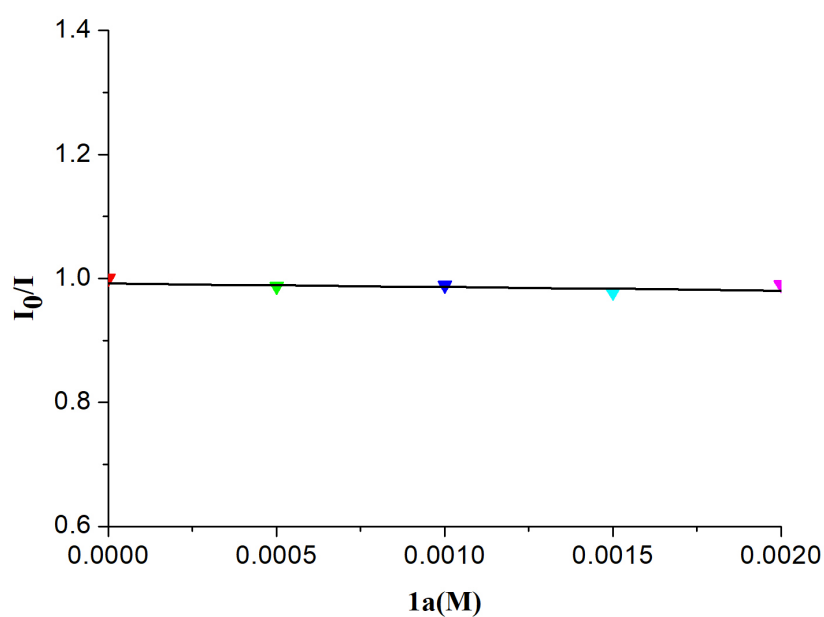


Fig. S1 Stern–Volmer quenching experiment of DPZ and **1a**.

(Result: no quenching observed)

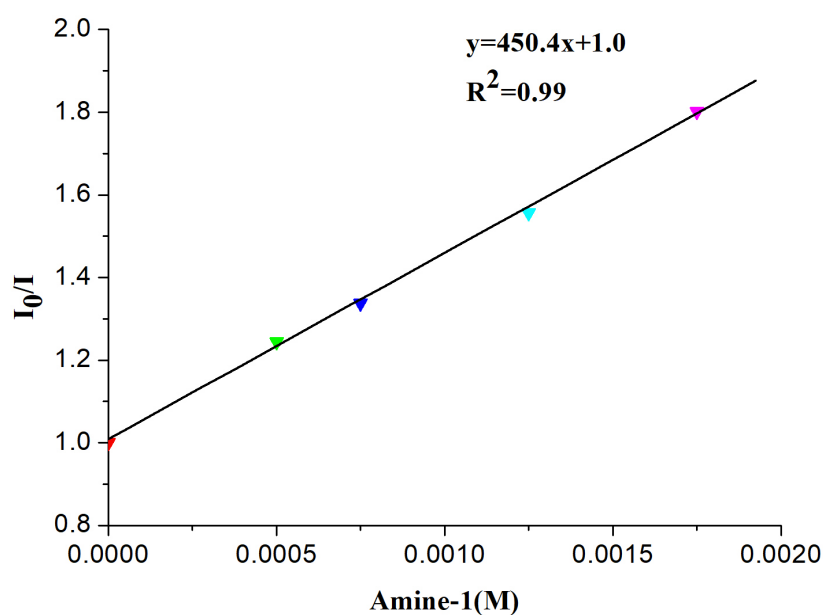


Fig. S2 Stern–Volmer quenching experiment of DPZ and **Amine-1**.

(Result: quenching observed)

Cyclic voltammetry measurement

Cyclic voltammetry experiments were performed on a CHI600E Workstation. Measurements were performed for anhydrous acetonitrile solutions ($[\text{sample}] = 1.0 \text{ mM}$, $[(\text{NBu}_4)\text{PF}_6] = 0.10 \text{ M}$) with a radium glassy carbon (working electrode) and platinum wire (counter electrode), and a Ag/AgCl(AgNO_3) reference electrode under N_2 at room temperature. The scan rate was 50 mV/s. Ferrocene (Cp_2Fe) was used as a reference. The obtained value was referenced to Ag/AgCl and converted to SCE by adding 0.03 V. The obtained value was referenced to Ag/AgNO₃ and converted to SCE by adding 0.337 V.

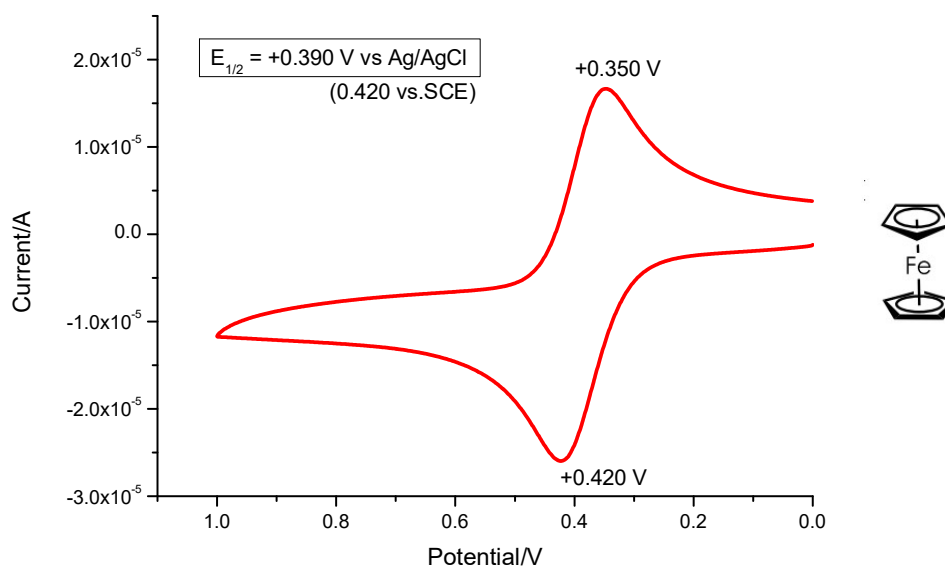


Fig. S3. Cyclic voltammogram of Ferrocene with a Ag/AgCl reference electrode.

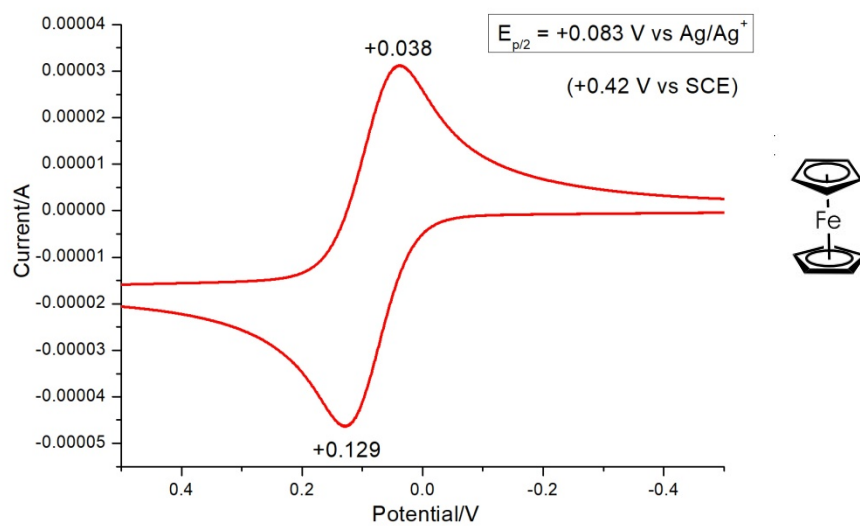


Fig. S4. Cyclic voltammogram of Ferrocene in MeCN with a Ag/AgNO₃ reference electrode.

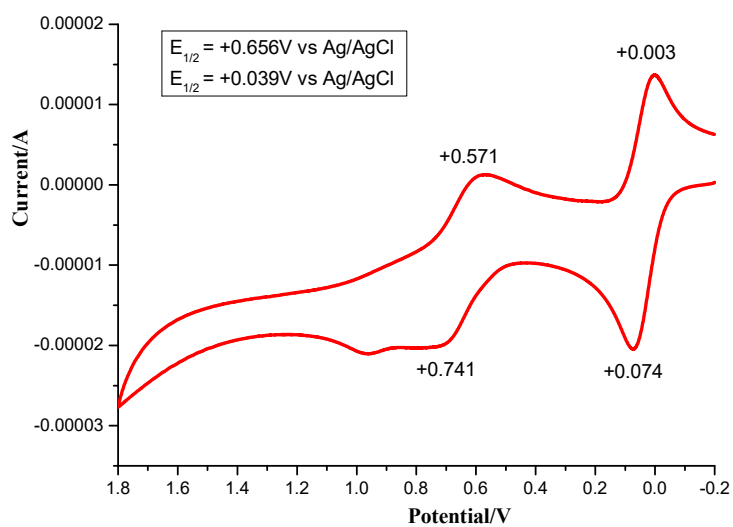


Fig. S5. Cyclic voltammogram of **Amine-1** in MeCN. $E_{1/2} = +0.686$ V versus SCE in CH_3CN , $E_{1/2} = +0.069$ V versus SCE in CH_3CN .

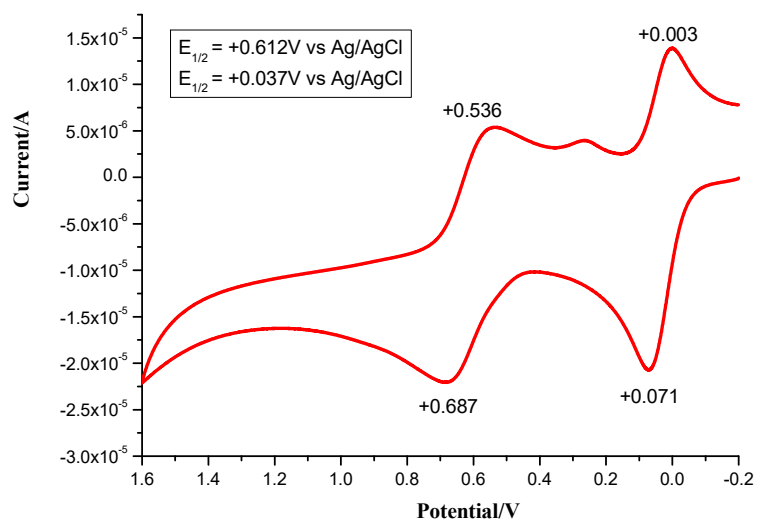


Fig. S6 Cyclic voltammogram of **Amine-2** in MeCN. $E_{1/2} = +0.642$ V versus SCE in CH_3CN , $E_{1/2} = +0.067$ V versus SCE in CH_3CN .

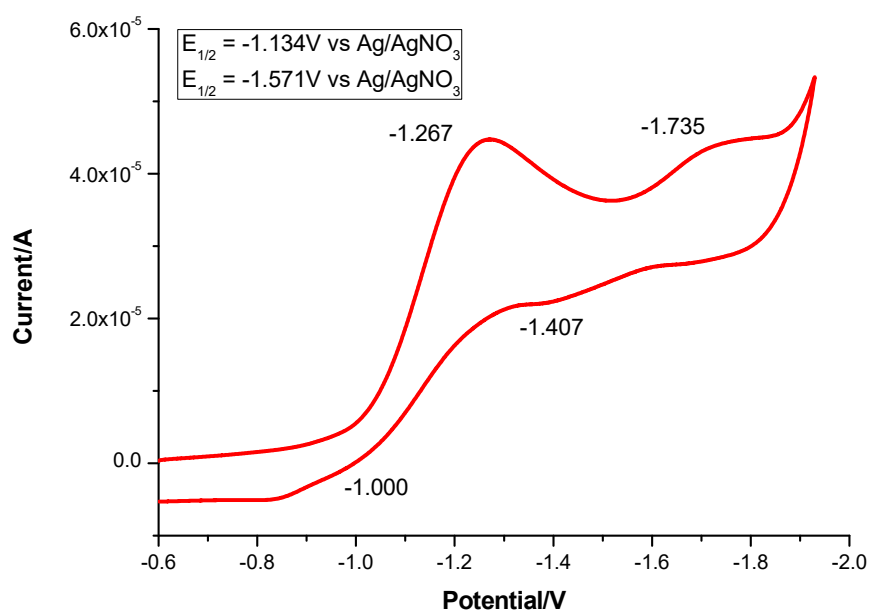


Fig. S7 Cyclic voltammogram of **1a** in MeCN. $E_{1/2} = -0.797\text{ V}$ versus SCE in CH_3CN , $E_{1/2} = -1.234\text{ V}$ versus SCE in CH_3CN .

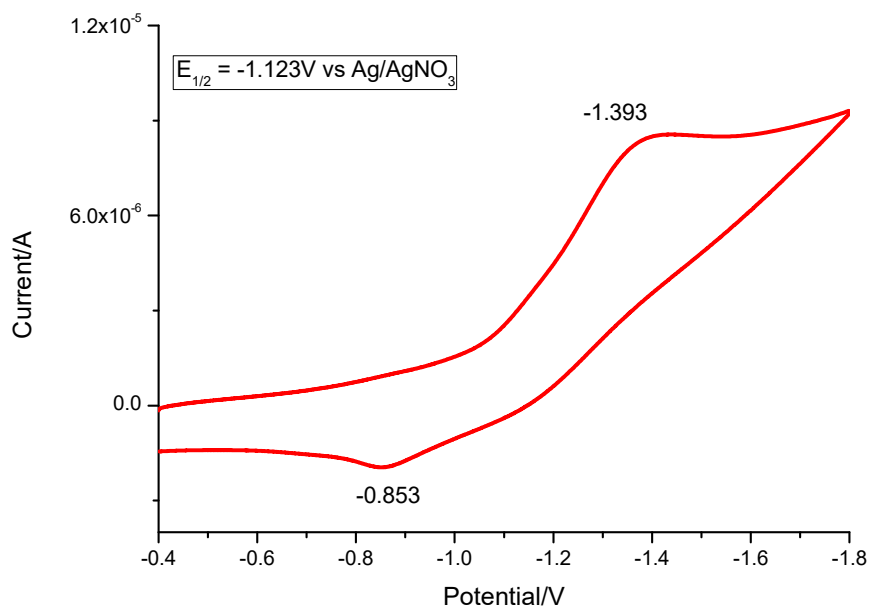


Fig. S8 Cyclic voltammogram of **2a** in MeCN. $E_{1/2} = -0.786\text{ V}$ versus SCE in CH_3CN .

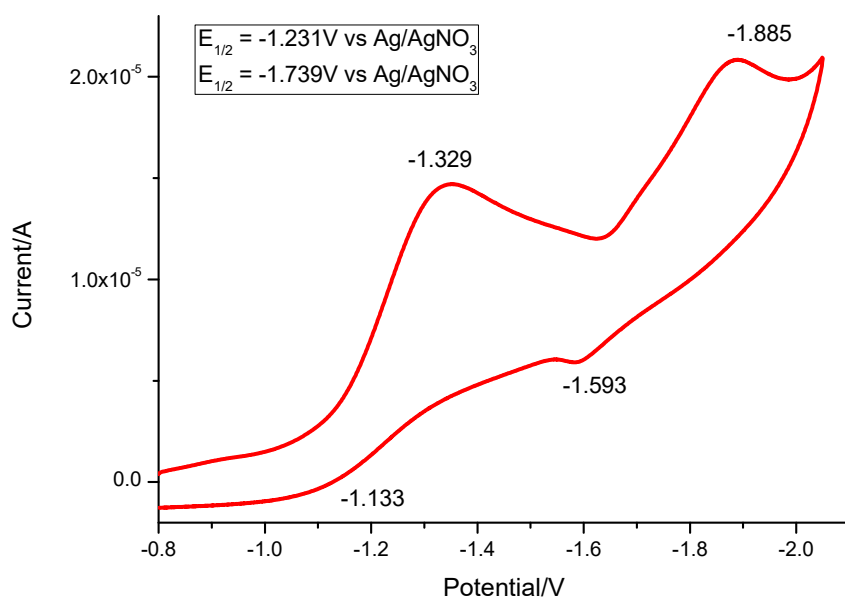


Fig. S9 Cyclic voltammogram of **1b** in MeCN. $E_{1/2} = -0.894$ V versus SCE in CH_3CN , $E_{1/2} = -1.402$ V versus SCE in CH_3CN .

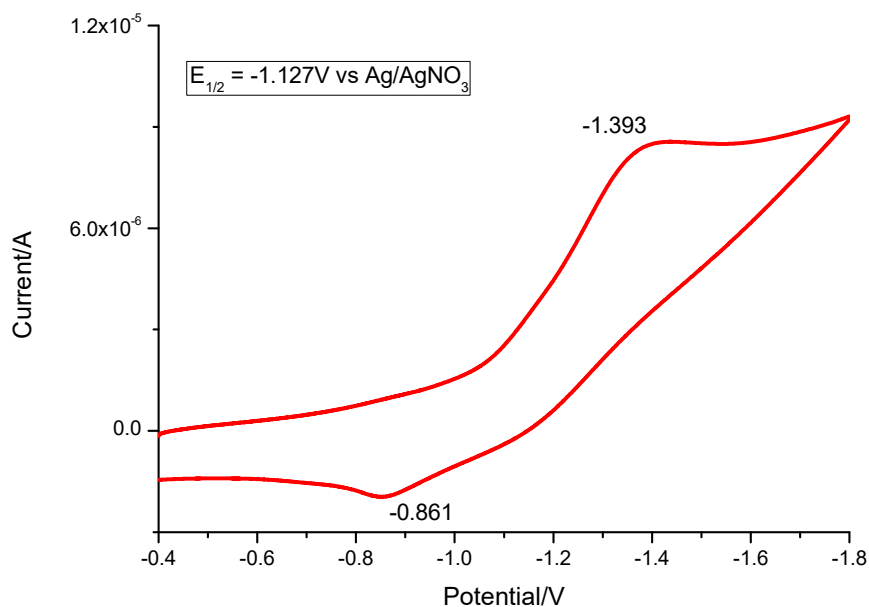


Fig. S10 Cyclic voltammogram of **2zb** in MeCN. $E_{1/2} = -0.790$ V versus SCE in CH_3CN .

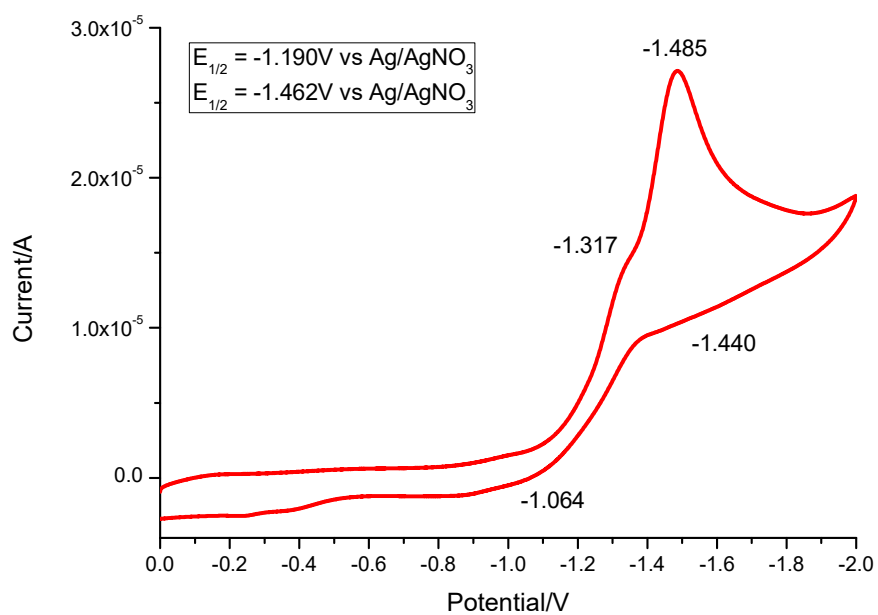


Fig. S11 Cyclic voltammogram of **4a** in MeCN. $E_{1/2} = -0.853\text{ V}$ versus SCE in CH_3CN , $E_{1/2} = -1.125\text{ V}$ versus SCE in CH_3CN .

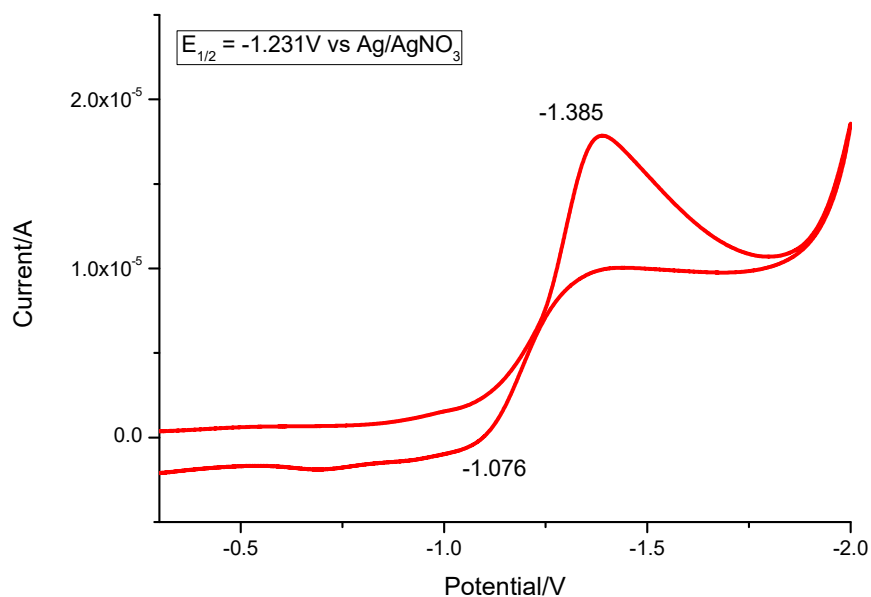


Fig. S12 Cyclic voltammogram of **5a** in MeCN. $E_{1/2} = -0.894\text{ V}$ versus SCE in CH_3CN .

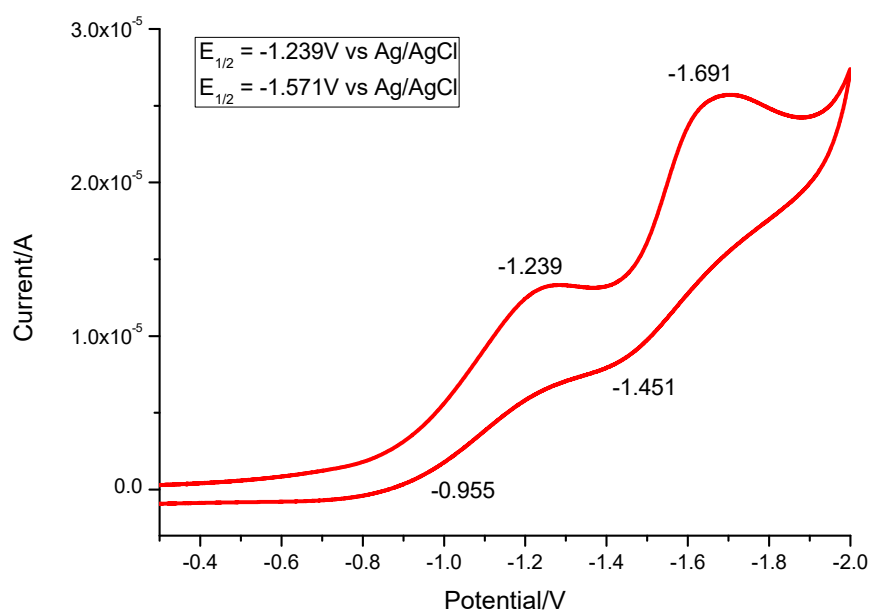


Fig. S13 Cyclic voltammogram of **4n** in MeCN. $E_{1/2} = -1.209$ V versus SCE in CH_3CN , $E_{1/2} = -1.541$ V versus SCE in CH_3CN .

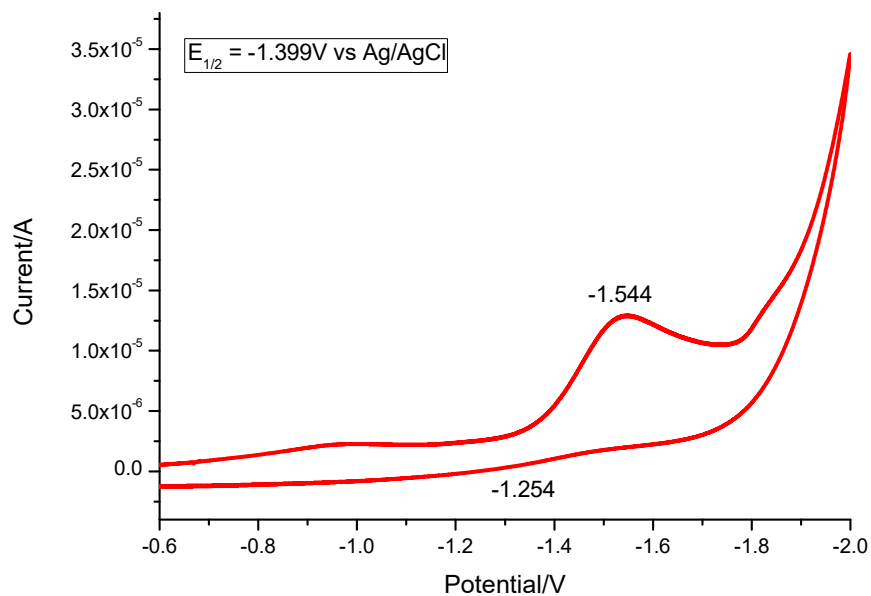


Fig. S14 Cyclic voltammogram of **5n** in MeCN. $E_{1/2} = -1.369$ V versus SCE in CH_3CN .

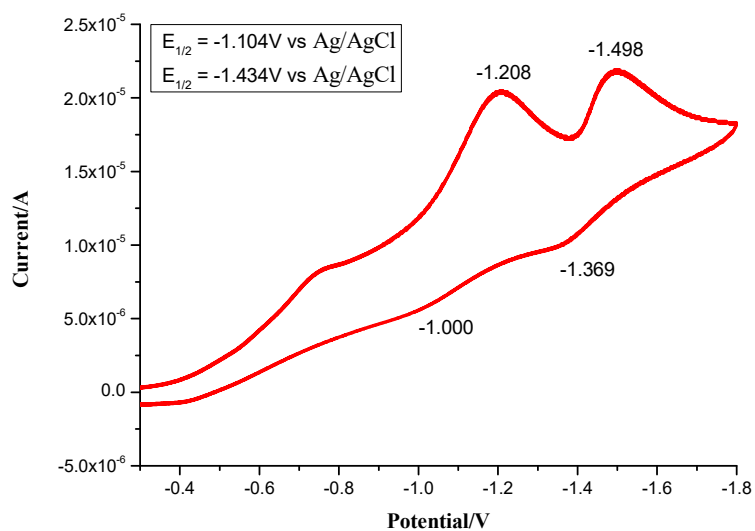


Fig. S15 Cyclic voltammogram of **7** in MeCN. $E_{1/2} = -1.074$ V versus SCE in CH_3CN , $E_{1/2} = -1.404$ V versus SCE in CH_3CN .

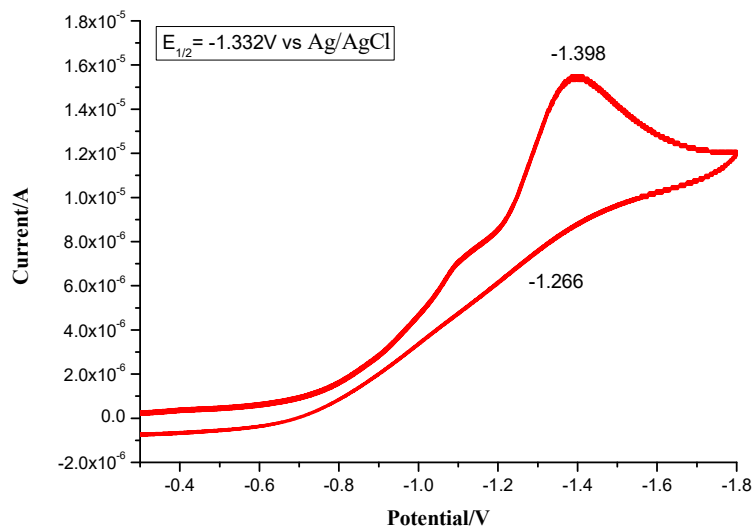


Fig. S16 Cyclic voltammogram of **8** in MeCN. $E_{1/2} = -1.302$ V versus SCE in CH_3CN .

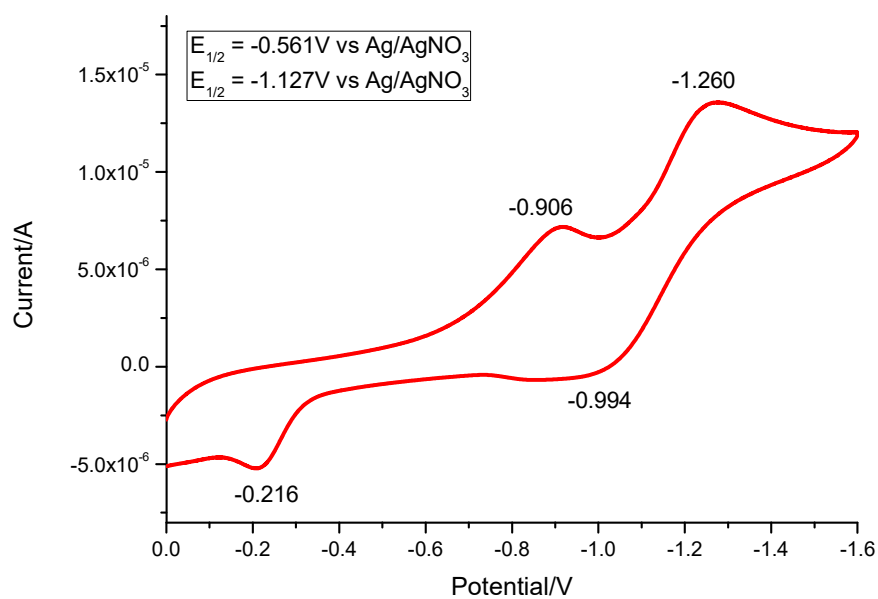


Fig. S17 Cyclic voltammogram of **C1** in MeCN. $E_{1/2} = -0.224$ V versus SCE in CH_3CN , $E_{1/2} = -0.790$ V versus SCE in CH_3CN .

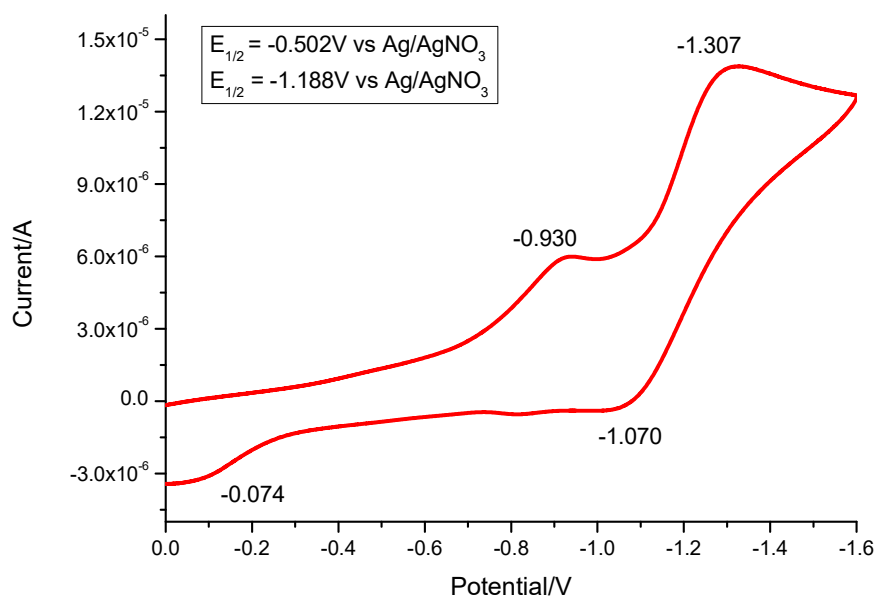


Fig. S18 Cyclic voltammogram of **C25** in MeCN. $E_{1/2} = -0.165$ V versus SCE in CH_3CN , $E_{1/2} = -0.851$ V versus SCE in CH_3CN .

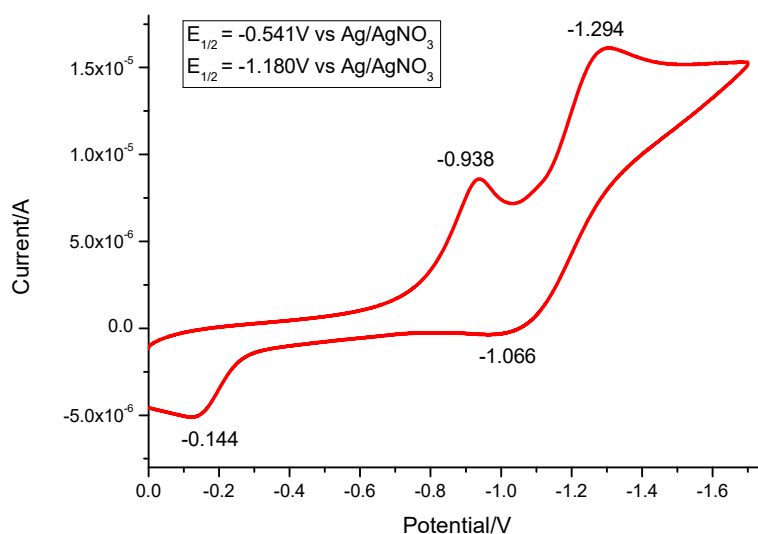


Fig. S19 Cyclic voltammogram of **C26** in MeCN. $E_{1/2} = -0.204$ V versus SCE in CH_3CN , $E_{1/2} = -0.843$ V versus SCE in CH_3CN .

UV-vis experiments.

Solutions of different complexes were introduced to a 3 cm path length quartz cuvette equipped with a Teflon[®] septum and analyzed using a TU-1901 spectrophotometer.

For the solutions of **1a** in DCE: **1a** (7.92 mg, 0.04 mmol) was dissolved in DCE (40 mL), then transformed to 3 cm path length quartz cuvettes, and sealed with a Teflon[®] septa.

For the solutions of **Amine-1** in DCE: **Amine-1** (3.89 mg, 0.024 mmol) was dissolved in DCE (40 mL), then transformed to 3 cm path length quartz cuvettes, and sealed with a Teflon[®] septa.

For the solutions of **1a** and Na_2CO_3 in DCE: **1a** (7.92 mg, 0.04 mmol) and Na_2CO_3 (10.6 mg, 0.1 mmol) were dissolved in DCE (40 mL), then transformed to 3 cm path length quartz cuvettes, and sealed with a Teflon[®] septa.

For the solutions of **1a** and **Amine-1** in DCE: **1a** (7.92 mg, 0.04 mmol) and **Amine-1** (3.89 mg, 0.024 mmol) were dissolved in DCE (40 mL), then transformed to 3 cm path length

quartz cuvettes, and sealed with a Teflon[®] septa.

For the solutions of **1a**, **Amine-1**, and Na₂CO₃ in DCE: **1a** (7.92 mg, 0.04 mmol), **Amine-1** (3.89 mg, 0.024 mmol), and Na₂CO₃ (10.6 mg, 0.1 mmol) were dissolved in DCE (40 mL), then transformed to 3 cm path length quartz cuvettes, and sealed with a Teflon[®] septa.

For the solutions of **1a**, **Amine-1**, Na₂CO₃, and **C1** in DCE: **1a** (7.92 mg, 0.04 mmol), **Amine-1** (3.89 mg, 0.024 mmol), Na₂CO₃ (10.6 mg, 0.1 mmol) and **C1** (0.99 mg, 0.002 mmol) were dissolved in DCE (40 mL), then transformed to 3 cm path length quartz cuvettes, and sealed with a Teflon[®] septa.

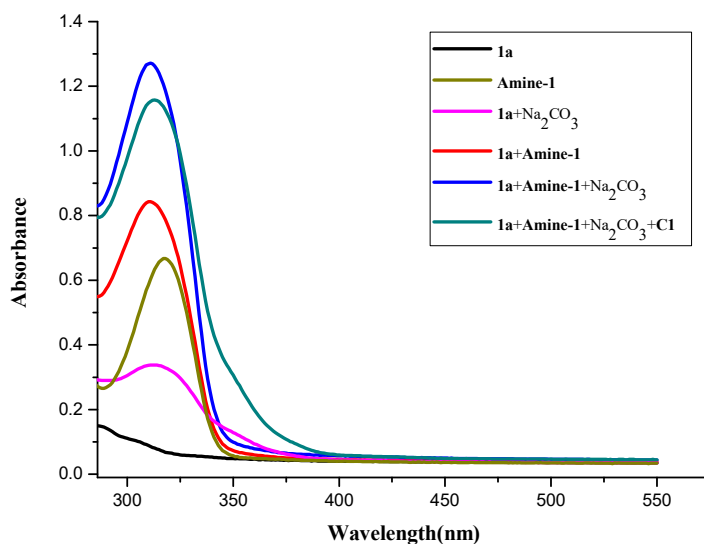


Fig. S20 Absorption spectra of various complexes.

For the solutions of **2a** in DCE: **2a** (6.56 mg, 0.04 mmol) was dissolved in DCE (40 mL), then transformed to 3 cm path length quartz cuvettes, and sealed with a Teflon[®] septa.

For the solutions of **Amine-1** in DCE: **Amine-1** (3.89 mg, 0.024 mmol) was dissolved in DCE (40 mL), then transformed to 3 cm path length quartz cuvettes, and sealed with a Teflon[®] septa.

For the solutions of **2a** and Na₂CO₃ in DCE: **2a** (6.56 mg, 0.04 mmol) and Na₂CO₃ (10.6 mg, 0.1 mmol) were dissolved in DCE (40 mL), then transformed to 3 cm path length quartz

cuvettes, and sealed with a Teflon[®] septa.

For the solutions of **2a** and **Amine-1** in DCE: **2a** (6.56 mg, 0.04 mmol) and **Amine-1** (3.89 mg, 0.024 mmol) were dissolved in DCE (40 mL), then transformed to 3 cm path length quartz cuvettes, and sealed with a Teflon[®] septa.

For the solutions of **2a**, **Amine-1**, and Na₂CO₃ in DCE: **2a** (6.56 mg, 0.04 mmol), **Amine-1** (3.89 mg, 0.024 mmol), and Na₂CO₃ (10.6 mg, 0.1 mmol) were dissolved in DCE (40 mL), then transformed to 3 cm path length quartz cuvettes, and sealed with a Teflon[®] septa.

For the solutions of **2a**, **Amine-1**, Na₂CO₃, and **C1** in DCE: **2a** (6.56 mg, 0.04 mmol), **Amine-1** (3.89 mg, 0.024 mmol), Na₂CO₃ (10.6 mg, 0.1 mmol) and **C1** (0.99 mg, 0.002 mmol) were dissolved in DCE (40 mL), then transformed to 3 cm path length quartz cuvettes, and sealed with a Teflon[®] septa.

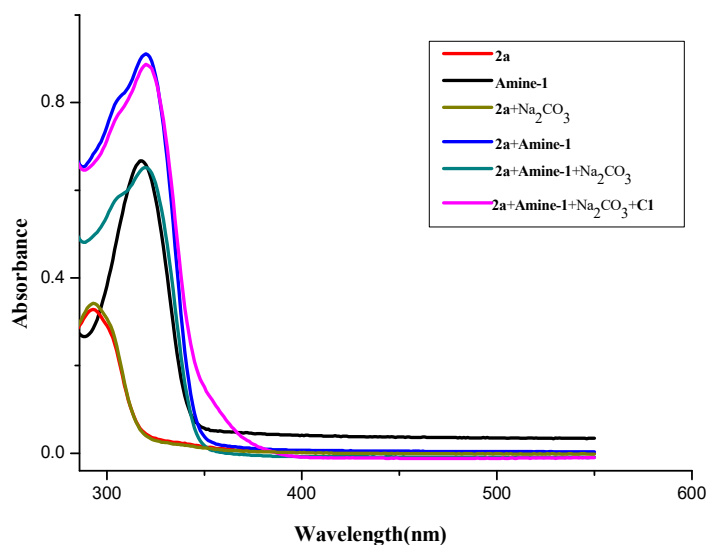
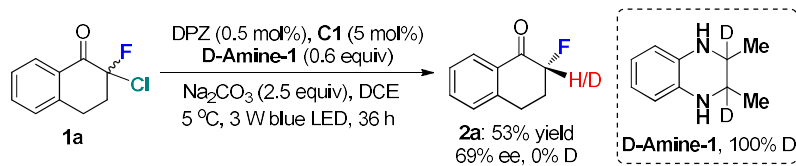
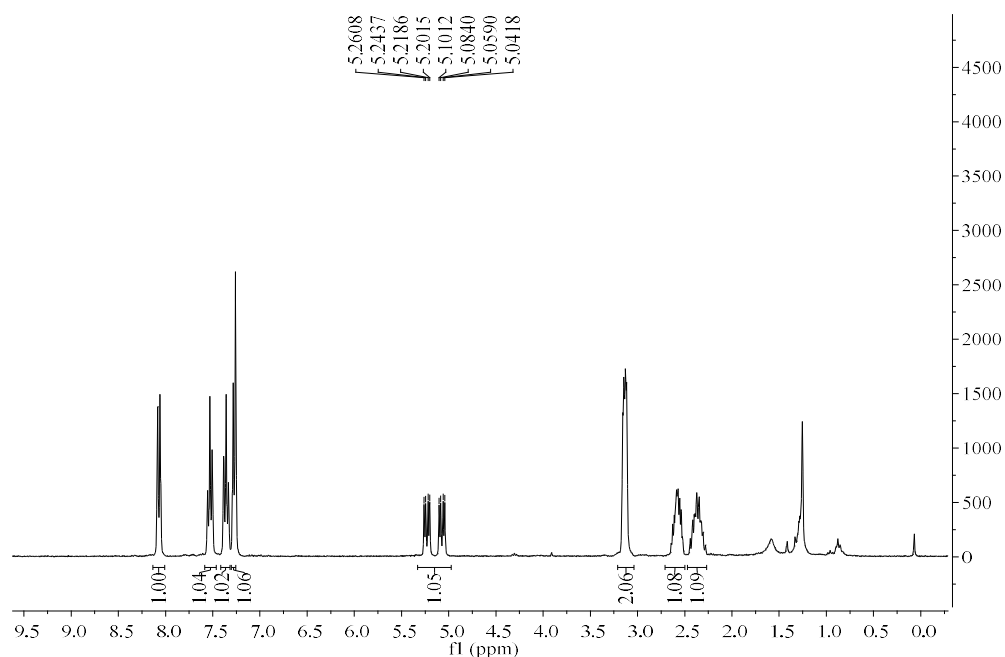


Fig. S21 Absorption spectra of various complexes.

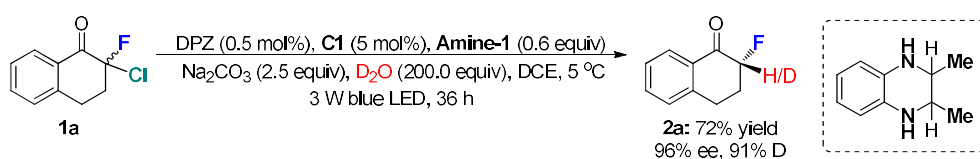
The study of the proton source



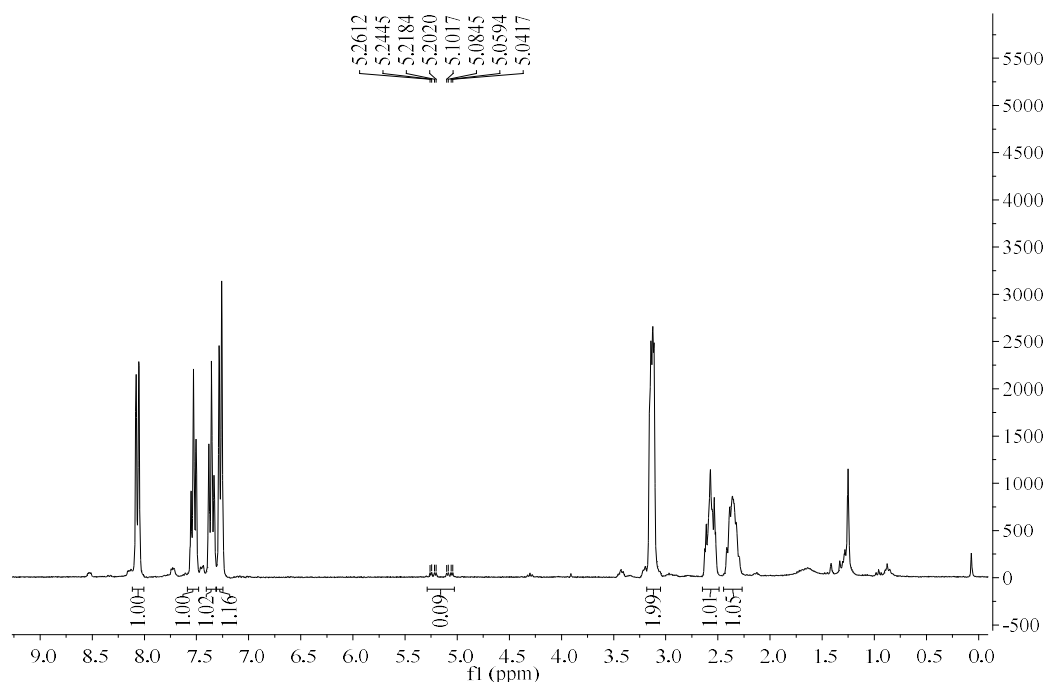
35 μL (0.0005 mmol, 0.005 equiv) of DPZ solution (1.0 mg of DPZ in 200 μL of anhydrous toluene) was added into a 10 mL Schlenk tube, and then solvent was removed in *vacuo*. Subsequently, **C1** (2.48 mg, 0.005 mmol, 0.05 equiv), Na_2CO_3 (26.5 mg, 0.25 mmol, 2.5 equiv), **D-Amine-1** (9.84 mg, 0.06 mmol, 0.6 equiv), **1a** (19.8 mg, 0.1 mmol, 1.0 equiv), and stir bar and DCE (1.0 mL) were added sequentially and then degassed for three times by freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at 5 °C for 30 min without light, then irradiated by a 3 W blue LED ($\lambda = 450\text{--}455\text{ nm}$) from a 2.0 cm distance for another 36 hours. The reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with petroleum ether/dichloromethane (5/1–2/1 ratio). Removing the solvent in *vacuo*, afforded products **2a** with 0% D incorporation.



^1H NMR of **2a** (D-Amine-1 as the sacrificial reductant)



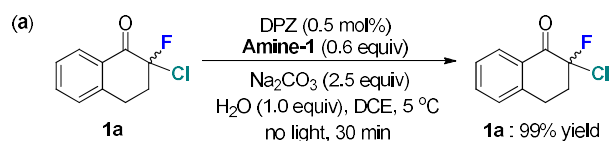
35 μL (0.0005 mmol, 0.005 equiv) of DPZ solution (1.0 mg of DPZ in 200 μL of anhydrous toluene) was added into a 10 mL Schlenk tube, and then solvent was removed in *vacuo*. Subsequently, **C1** (2.48 mg, 0.005 mmol, 0.05 equiv), Na_2CO_3 (26.5 mg, 0.25 mmol, 2.5 equiv), **Amine-1** (9.72 mg, 0.06 mmol, 0.6 equiv), **1a** (19.8 mg, 0.1 mmol, 1.0 equiv), D_2O (0.4 mL, 20 mmol, 200 equiv), stir bar and DCE (1.0 mL) were added sequentially and then degassed for three times by freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at 5 $^\circ\text{C}$ for 30 min without light, then irradiated by a 3 W blue LED ($\lambda = 450\text{--}455\text{ nm}$) from a 2.0 cm distance for another 36 hours. The reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with petroleum ether/dichloromethane (5/1–2/1 ratio). Removing the solvent in *vacuo*, afforded products **2a** in 72% yield with 96% ee and 91% D incorporation.



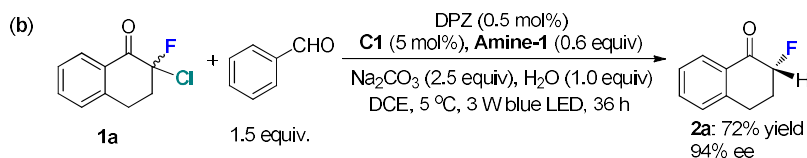
^1H NMR of **2a (D_2O as the proton source)**

These results indicated that the proton might be from free H^+ of the reaction system.

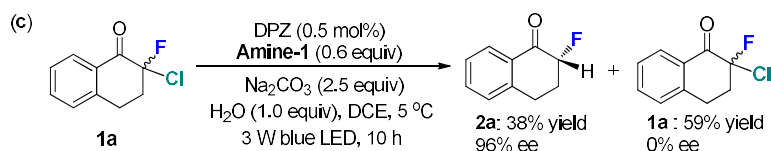
Control experiments



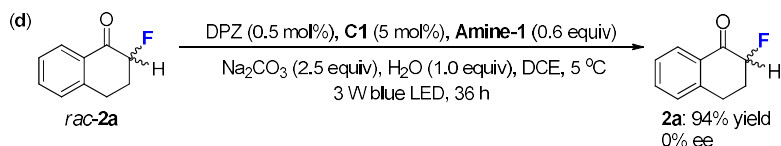
35 μ L (0.0005 mmol, 0.005 equiv) of DPZ solution (1.0 mg of DPZ in 200 μ L of anhydrous toluene) was added into a 10 mL Schlenk tube, and then solvent was removed in *vacuo*. Subsequently, **C1** (2.48 mg, 0.005 mmol, 0.05 equiv), Na₂CO₃ (26.5 mg, 0.25 mmol, 2.5 equiv), **Amine-1** (9.72 mg, 0.06 mmol, 0.6 equiv), **1a** (19.8 mg, 0.1 mmol, 1.0 equiv), H₂O (1.8 μ L, 0.1 mmol, 1.0 equiv), stir bar and DCE (1.0 mL) were added sequentially and then degassed for three times by freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at 5 °C for 30 min without light. The reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with petroleum ether/dichloromethane (5/1–3/1 ratio). Removing the solvent in *vacuo*, recovered **1a** in 99% yield.



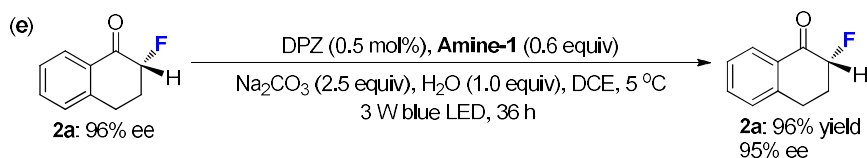
35 μ L (0.0005 mmol, 0.005 equiv) of DPZ solution (1.0 mg of DPZ in 200 μ L of anhydrous toluene) was added into a 10 mL Schlenk tube, and then solvent was removed in *vacuo*. Subsequently, **C1** (2.48 mg, 0.005 mmol, 0.05 equiv), Na₂CO₃ (26.5 mg, 0.25 mmol, 2.5 equiv), **Amine-1** (9.72 mg, 0.06 mmol, 0.6 equiv), **1a** (19.8 mg, 0.1 mmol, 1.0 equiv), benzaldehyde (15.3 μ L, 0.15 mmol, 1.5 equiv), H₂O (1.8 μ L, 0.1 mmol, 1.0 equiv), stir bar and DCE (1.0 mL) were added sequentially and then degassed for three times by freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at 5 °C for 30 min without light, then irradiated by a 3 W blue LED (λ = 450–455 nm) from a 2.0 cm distance for another 10 hours. The reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with petroleum ether/dichloromethane (5/1–2/1 ratio). Removing the solvent in *vacuo*, afforded products **2a** in 72% yield with 94% ee.



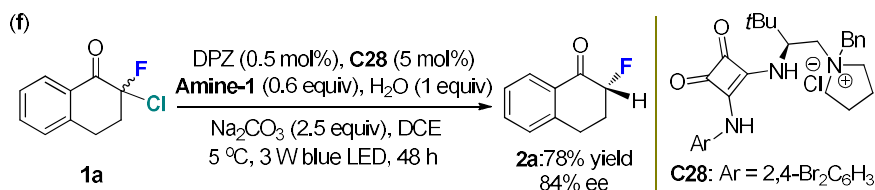
35 μL (0.0005 mmol, 0.005 equiv) of DPZ solution (1.0 mg of DPZ in 200 μL of anhydrous toluene) was added into a 10 mL Schlenk tube, and then solvent was removed in *vacuo*. Subsequently, **C1** (2.48 mg, 0.005 mmol, 0.05 equiv), Na_2CO_3 (26.5 mg, 0.25 mmol, 2.5 equiv), **Amine-1** (9.72 mg, 0.06 mmol, 0.6 equiv), **1a** (19.8 mg, 0.1 mmol, 1.0 equiv), H_2O (1.8 μL , 0.1 mmol, 1.0 equiv), stir bar and DCE (1.0 mL) were added sequentially and then degassed for three times by freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at 5 $^\circ\text{C}$ for 30 min without light, then irradiated by a 3 W blue LED ($\lambda = 450\text{--}455\text{ nm}$) from a 2.0 cm distance for another 10 hours. The reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with petroleum ether/dichloromethane (5/1–2/1 ratio). Removing the solvent in *vacuo*, afforded products **2a** in 38% yield with 96% ee, and recovered **1a** in 59% yield with 0 ee.



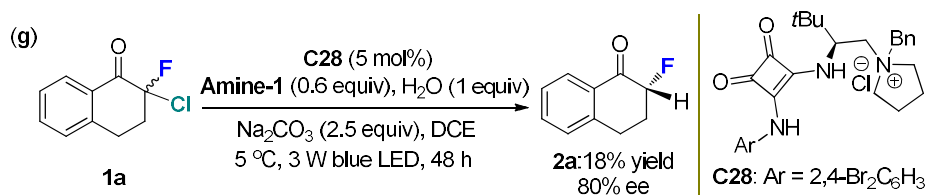
35 μL (0.0005 mmol, 0.005 equiv) of DPZ solution (1.0 mg of DPZ in 200 μL of anhydrous toluene) was added into a 10 mL Schlenk tube, and then solvent was removed in *vacuo*. Subsequently, **C1** (2.48 mg, 0.005 mmol, 0.05 equiv), Na_2CO_3 (26.5 mg, 0.25 mmol, 2.5 equiv), **Amine-1** (9.72 mg, 0.06 mmol, 0.6 equiv), *rac*-**2a** (16.4 mg, 0.1 mmol, 1.0 equiv), H_2O (1.8 μL , 0.1 mmol, 1.0 equiv), stir bar and DCE (1.0 mL) were added sequentially and then degassed for three times by freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at 5 $^\circ\text{C}$ for 30 min without light, then irradiated by a 3 W blue LED ($\lambda = 450\text{--}455\text{ nm}$) from a 2.0 cm distance for another 10 hours. The reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with petroleum ether/dichloromethane (5/1–2/1 ratio). Removing the solvent in *vacuo*, afforded products **2a** in 94% yield with 0 ee.



35 μL (0.0005 mmol, 0.005 equiv) of DPZ solution (1.0 mg of DPZ in 200 μL of anhydrous toluene) was added into a 10 mL Schlenk tube, and then solvent was removed in *vacuo*. Subsequently, Na_2CO_3 (26.5 mg, 0.25 mmol, 2.5 equiv), **Amine-1** (9.72 mg, 0.06 mmol, 0.6 equiv), **2a** (96% ee, 16.4 mg, 0.1 mmol, 1.0 equiv), H_2O (1.8 μL , 0.1 mmol, 1.0 equiv), stir bar and DCE (1.0 mL) were added sequentially and then degassed for three times by freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at 5 °C for 30 min without light, then irradiated by a 3 W blue LED ($\lambda = 450\text{--}455\text{ nm}$) from a 2.0 cm distance for another 10 hours. The reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with petroleum ether/dichloromethane (5/1–2/1 ratio). Removing the solvent in *vacuo*, afforded products **2a** in 96% yield with 95% ee.



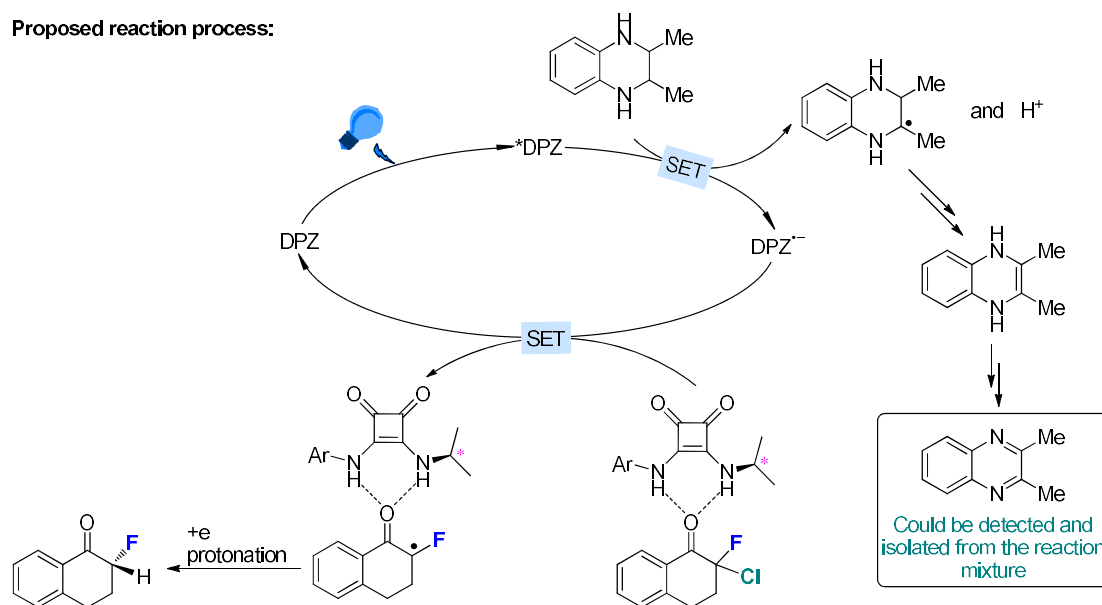
35 μL (0.0005 mmol, 0.005 equiv) of DPZ solution (1.0 mg of DPZ in 200 μL of anhydrous toluene) was added into a 10 mL Schlenk tube, and then solvent was removed in *vacuo*. Subsequently, **C28** (3.1 mg, 0.005 mmol, 0.05 equiv), Na_2CO_3 (26.5 mg, 0.25 mmol, 2.5 equiv), **Amine-1** (9.72 mg, 0.06 mmol, 0.6 equiv), **1a** (19.8 mg, 0.1 mmol, 1.0 equiv), H_2O (1.8 μL , 0.1 mmol, 1.0 equiv), stir bar and DCE (1.0 mL) were added sequentially and then degassed for three times by freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at 5 °C for 30 min without light, then irradiated by a 3 W blue LED ($\lambda = 450\text{--}455\text{ nm}$) from a 2.0 cm distance for another 10 hours. The reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with petroleum ether/dichloromethane (5/1–2/1 ratio). Removing the solvent in *vacuo*, afforded products **2a** in 78% yield with 84% ee.



Na₂CO₃ (26.5 mg, 0.25 mmol, 2.5 equiv), **Amine-1** (9.72 mg, 0.06 mmol, 0.6 equiv), **2a** (96% ee, 16.4 mg, 0.1 mmol, 1.0 equiv), H₂O (1.8 μ L, 0.1 mmol, 1.0 equiv), stir bar and DCE (1.0 mL) were added to a 10 mL Schlenk tube sequentially and then degassed for three times by freeze-pump-thaw method. The reaction mixture was stirred under an argon atmosphere at 5 °C for 30 min without light, then irradiated by a 3 W blue LED (λ = 450–455 nm) from a 2.0 cm distance for another 10 hours. The reaction mixture was directly loaded onto a short *silica gel* column, followed by gradient elution with petroleum ether/dichloromethane (5/1–2/1 ratio). Removing the solvent in *vacuo*, afforded products **2a** in 18% yield with 80% ee.

5. Proposed mechanism

Proposed reaction process:



Disfavoured and favoured transition states:

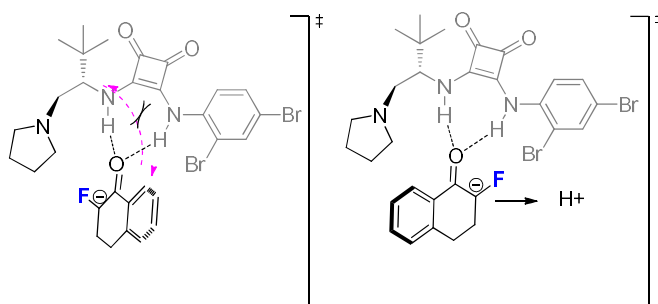


Fig. S22 Plausible reaction process and transition states.

6. Determination of the absolute configurations

Absolute configurations of **2** are determined by *X*-ray structure analysis of the product **2d**.

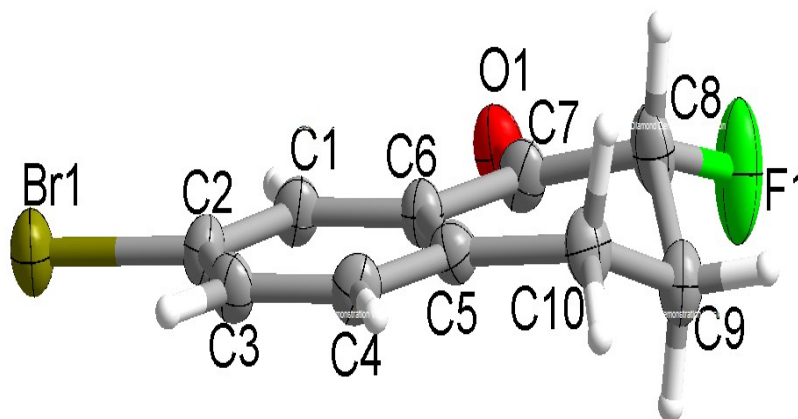


Fig. S21 Absolute configuration of **2d** (CCDC 1590512).

Displacement ellipsoids are drawn at the 30% probability level. (Solvent: dichloromethane)

Table S4 Crystal data and structure refinement.

Identification code	HMT70074
Empirical formula	C ₁₀ H ₈ BrFO
Formula weight	243.07
Temperature/K	293(2)
Crystal system	monoclinic
Space group	P2 ₁
a/Å	7.8209(6)
b/Å	6.9066(8)
c/Å	8.6288(6)
α/°	90
β/°	100.372(7)
γ/°	90
Volume/Å ³	458.47(7)
Z	2
ρ _{calc} /cm ³	1.761
μ/mm ⁻¹	5.892
F(000)	240.0
Crystal size/mm ³	0.23 × 0.13 × 0.08
Radiation	CuKα (λ = 1.54184)
2θ range for data collection/°	10.422 to 134.124
Index ranges	-9 ≤ h ≤ 8, -7 ≤ k ≤ 8, -10 ≤ l ≤ 10
Reflections collected	3410
Independent reflections	1530 [R _{int} = 0.0269, R _{sigma} = 0.0345]

Data/restraints/parameters	1530/1/118
Goodness-of-fit on F^2	1.064
Final R indexes [$I \geq 2\sigma(I)$]	$R_1 = 0.0342$, $wR_2 = 0.0855$
Final R indexes [all data]	$R_1 = 0.0410$, $wR_2 = 0.0922$
Largest diff. peak/hole / $e \text{ \AA}^{-3}$	0.43/-0.35
Flack parameter	-0.04(6)

Table S5 Fractional Atomic Coordinates ($\times 10^4$) and Equivalent Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$). U_{eq} is defined as 1/3 of of the trace of the orthogonalised U_{ij} tensor.

Atom	x	y	z	U(eq)
Br1	8986.6(7)	4963.8(2)	6006.1(7)	66.0(3)
C1	6489(7)	4910(30)	3177(6)	51.0(16)
C2	6706(6)	4950(30)	4785(6)	47.7(11)
C3	5288(7)	4980(30)	5553(6)	52.8(13)
C4	3654(7)	5010(30)	4661(6)	51.8(13)
C5	3375(6)	4990(30)	3020(6)	47.7(11)
C6	4830(6)	4940(30)	2284(5)	45.2(11)
C7	4619(7)	4950(30)	530(6)	55.1(15)
C8	2784(9)	4640(30)	-327(7)	67(5)
C9	1487(9)	5749(13)	420(8)	63(2)
C10	1564(6)	4940(30)	2080(6)	57.8(14)
F1	2697(6)	5260(30)	-1862(4)	121(4)
O1	5835(5)	5010(30)	-146(4)	68.9(13)

Table S6 Anisotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$). The Anisotropic displacement factor exponent takes the form: $-2\pi^2[h^2a^{*2}U_{11}+2hka^*b^*U_{12}+\dots]$.

Atom	U_{11}	U_{22}	U_{33}	U_{23}	U_{13}	U_{12}
Br1	49.8(3)	93.0(5)	51.7(3)	-1.9(9)	-0.6(2)	1.3(10)
C1	49(3)	65(4)	42(2)	-7(7)	14(2)	14(8)
C2	43(2)	58(3)	42(2)	6(8)	5(2)	4(9)
C3	56(3)	65(4)	37(2)	-8(8)	10(2)	6(10)
C4	51(3)	66(4)	44(2)	-1(8)	21(2)	9(8)
C5	47(2)	53(3)	45(2)	-6(8)	13(2)	1(9)
C6	46(2)	55(3)	36(2)	1(8)	11.0(19)	0(9)
C7	60(3)	71(4)	37(2)	-5(9)	16(2)	13(9)
C8	62(4)	100(14)	37(3)	-8(5)	4(3)	-2(5)
C9	51(4)	85(6)	50(4)	6(3)	2(3)	2(3)
C10	44(3)	77(4)	54(3)	0(11)	12(2)	-8(10)
F1	77(3)	242(11)	41.6(19)	30(6)	5.9(19)	0(7)
O1	63(2)	107(4)	41.0(18)	-10(8)	20.2(18)	-5(8)

Table S7 Bond Lengths.

Atom	Atom	Length/Å	Atom	Atom	Length/Å
Br1	C2	1.901(5)	C5	C10	1.501(7)
C1	C2	1.367(7)	C6	C7	1.492(6)
C1	C6	1.385(7)	C7	C8	1.506(11)
C2	C3	1.390(7)	C7	O1	1.203(7)
C3	C4	1.368(7)	C8	C9	1.504(15)
C4	C5	1.393(7)	C8	F1	1.380(12)
C5	C6	1.400(7)	C9	C10	1.530(12)

Table S8 Bond Angles.

Atom	Atom	Atom	Angle/°	Atom	Atom	Atom	Angle/°
C2	C1	C6	119.9(4)	C1	C6	C7	119.1(4)
C1	C2	Br1	119.7(4)	C5	C6	C7	120.6(5)
C1	C2	C3	121.3(4)	C6	C7	C8	114.8(6)
C3	C2	Br1	119.0(4)	O1	C7	C6	122.6(5)
C4	C3	C2	118.5(4)	O1	C7	C8	122.3(5)
C3	C4	C5	122.1(4)	C9	C8	C7	112.1(12)
C4	C5	C6	118.0(5)	F1	C8	C7	107.9(9)
C4	C5	C10	120.6(4)	F1	C8	C9	109.6(14)
C6	C5	C10	121.3(4)	C8	C9	C10	107.4(9)
C1	C6	C5	120.3(4)	C5	C10	C9	111.8(7)

Table S9 Torsion Angles.

A	B	C	D	Angle/°	A	B	C	D	Angle/°
Br1	C2	C3	C4	178.8(15)	C5	C6	C7	O1	176(2)
C1	C2	C3	C4	-1(3)	C6	C1	C2	Br1	-178.7(18)
C1	C6	C7	C8	170(2)	C6	C1	C2	C3	1(3)
C1	C6	C7	O1	-3(4)	C6	C5	C10	C9	-26(3)
C2	C1	C6	C5	-1(3)	C6	C7	C8	C9	41(3)
C2	C1	C6	C7	179(2)	C6	C7	C8	F1	162.1(19)
C2	C3	C4	C5	1(4)	C7	C8	C9	C10	-63.5(17)
C3	C4	C5	C6	0(3)	C8	C9	C10	C5	54.8(18)
C3	C4	C5	C10	177(2)	C10	C5	C6	C1	-177(2)
C4	C5	C6	C1	0(3)	C10	C5	C6	C7	3(3)
C4	C5	C6	C7	-179(3)	F1	C8	C9	C10	176.8(14)
C4	C5	C10	C9	156.1(19)	O1	C7	C8	C9	-144.9(18)
C5	C6	C7	C8	-10(3)	O1	C7	C8	F1	-24(3)

Table S10 Hydrogen Atom Coordinates (Å×10⁴) and Isotropic Displacement Parameters

(Å²×10³).

Atom	<i>x</i>	<i>y</i>	<i>z</i>	U(eq)
H1	7452	4868	2685	61
H3	5447	4983	6647	63
H4	2700	5048	5166	62
H8	2505	3261	-326	80
H9A	328	5594	-194	76
H9B	1772	7117	470	76
H10A	801	5684	2620	69
H10B	1150	3609	2007	69

Experimental

The crystal was kept at 293(2) K during data collection. Using Olex2, the structure was solved with the ShelXS structure solution program using Patterson Method and refined with the ShelXL [3] refinement package using Least Squares minimisation.

Crystal structure determination

Crystal Data for C₁₀H₈BrFO (*M* = 243.07 g/mol): monoclinic, space group P2₁ (no. 4), *a* = 7.8209(6) Å, *b* = 6.9066(8) Å, *c* = 8.6288(6) Å, *β* = 100.372(7)°, *V* = 458.47(7) Å³, *Z* = 2, *T* = 293(2) K, *μ*(CuKα) = 5.892 mm⁻¹, *D*_{calc} = 1.761 g/cm³, 3410 reflections measured (10.422° ≤ 2Θ ≤ 134.124°), 1530 unique (*R*_{int} = 0.0269, *R*_{sigma} = 0.0345) which were used in all calculations. The final *R*₁ was 0.0342 (*I* > 2σ(*I*)) and *wR*₂ was 0.0922 (all data).

Refinement model description

Number of restraints - 1, number of constraints - unknown.

Details:

1. Fixed Uiso

At 1.2 times of:

All C(H) groups, All C(H,H) groups

2.a Ternary CH refined with riding coordinates:

C8(H8)

2.b Secondary CH2 refined with riding coordinates:

C9(H9A,H9B), C10(H10A,H10B)

2.c Aromatic/amide H refined with riding coordinates:

C1(H1), C3(H3), C4(H4)

2) Absolute configurations of **6** and **8** are determined by *X*-ray structure analysis of the product **6f**

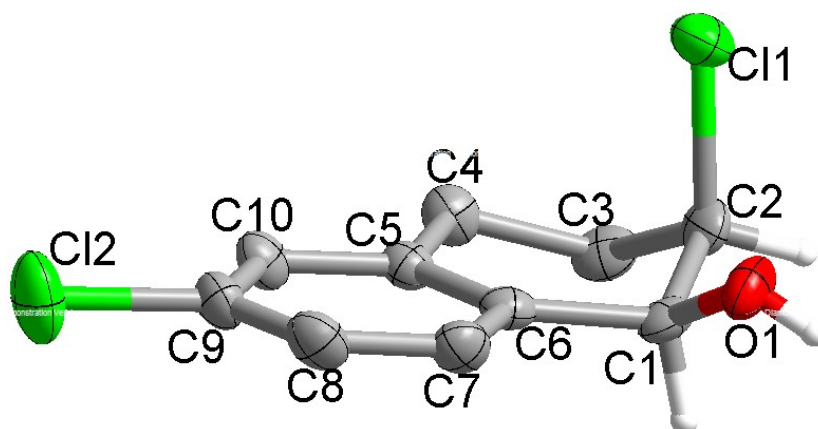


Fig. S22 Absolute configuration of **6f** (CCDC 1840318).

Table S11 Crystal data and structure refinement.

Identification code	HM23026
Empirical formula	C ₁₀ H ₁₀ Cl ₂ O
Formula weight	217.08
Temperature/K	293(2)
Crystal system	orthorhombic
Space group	P2 ₁ 2 ₁ 2 ₁
a/Å	5.04728(18)
b/Å	19.5159(6)
c/Å	20.2528(8)
α /°	90
β /°	90
γ /°	90
Volume/Å ³	1994.94(12)
Z	8
ρ_{calc} /cm ³	1.446
μ /mm ⁻¹	5.490
F(000)	896.0
Crystal size/mm ³	0.16 × 0.11 × 0.1
Radiation	CuK α (λ = 1.54184)
2 θ range for data collection/°	8.732 to 134.138
Index ranges	-6 ≤ h ≤ 4, -23 ≤ k ≤ 22, -24 ≤ l ≤ 23
Reflections collected	8713
Independent reflections	3534 [R_{int} = 0.0378, R_{sigma} = 0.0418]
Data/restraints/parameters	3534/0/243
Goodness-of-fit on F ²	1.032
Final R indexes [$I \geq 2\sigma(I)$]	R_1 = 0.0464, wR_2 = 0.1189
Final R indexes [all data]	R_1 = 0.0553, wR_2 = 0.1265
Largest diff. peak/hole / e Å ⁻³	0.38/-0.17

Flack parameter -0.014(19)

Table S12 Fractional Atomic Coordinates ($\times 10^4$) and Equivalent Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$). U_{eq} is defined as 1/3 of the trace of the orthogonalised U_{ij} tensor.

Atom	x	y	z	U(eq)
C1	4524(9)	6731(2)	7674(2)	47.7(10)
C2	4476(10)	6342(2)	7032(3)	53.2(11)
C3	4213(10)	6830(3)	6451(3)	55.9(12)
C4	6554(11)	7322(2)	6413(2)	51.8(12)
C5	7370(10)	7602.9(19)	7081(2)	42.4(9)
C6	6518(9)	7310(2)	7673(2)	43.0(10)
C7	7456(12)	7577(2)	8270(2)	52.5(11)
C8	9208(12)	8115(2)	8285(3)	58.1(13)
C9	9983(11)	8402(2)	7697(3)	54.0(12)
C10	9145(11)	8152(2)	7100(2)	50.7(12)
Cl1	7467(3)	5836.2(6)	6946.7(7)	65.1(3)
Cl2	12146(4)	9103.6(7)	7711.0(8)	88.9(6)
O1	5028(8)	6297(2)	8222(2)	59.8(10)
C1'	5328(10)	4323(3)	4308(3)	51.7(11)
C2'	5348(10)	3782(3)	4849(3)	57.0(13)
C3'	5594(11)	4117(3)	5515(3)	69.2(16)
C4'	3414(12)	4631(3)	5660(3)	65.0(15)
C5'	2453(12)	5030(2)	5072(2)	50.1(11)
C6'	3274(9)	4873(2)	4428(2)	47.4(11)
C7'	2238(12)	5243(2)	3900(2)	52.7(11)
C8'	414(12)	5757(2)	3999(2)	57.5(13)
C9'	-387(11)	5906(2)	4629(3)	56.0(12)
C10'	620(11)	5550(2)	5166(2)	54.8(13)
Cl1'	2346(3)	3271.8(6)	4820.3(7)	62.6(3)
Cl2'	-2718(4)	6557.0(7)	4756.7(8)	80.5(5)
O1'	4947(8)	4041(2)	3666.1(19)	58.4(9)

Table S13 Anisotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$). The Anisotropic displacement factor exponent takes the form: $-2\pi^2[h^2a^{*2}U_{11}+2hka^*b^*U_{12}+\dots]$.

Atom	U_{11}	U_{22}	U_{33}	U_{23}	U_{13}	U_{12}
C1	28(2)	61(2)	54(3)	5(2)	4(2)	5(2)
C2	32(2)	56(2)	71(3)	-3(2)	-4(2)	-6(2)
C3	38(3)	76(3)	54(3)	-5(2)	-12(2)	-2(2)
C4	60(3)	55(2)	40(2)	3.2(19)	-8(2)	-1(2)
C5	43(2)	41.4(19)	42.8(19)	-1.0(15)	-6(2)	5(2)

C6	39(2)	43(2)	47(2)	1.1(18)	0.2(19)	10.2(18)
C7	58(3)	57(2)	43(2)	-2.3(18)	9(3)	5(3)
C8	74(4)	53(3)	48(3)	-11(2)	-3(3)	2(3)
C9	60(3)	40(2)	62(3)	-4(2)	-4(3)	-6(2)
C10	65(3)	41(2)	46(2)	4.6(18)	-2(2)	0(2)
Cl1	63.0(8)	51.6(5)	80.7(8)	-7.7(5)	3.8(8)	10.5(7)
Cl2	120.9(15)	65.4(8)	80.3(9)	-5.5(6)	-9.9(11)	-41.3(10)
O1	40(2)	73(2)	67(2)	24.1(19)	6.0(18)	-4.7(18)
C1'	29(2)	71(3)	55(3)	-10(2)	0(2)	-14(2)
C2'	29(2)	71(3)	70(3)	2(3)	-7(2)	-2(2)
C3'	46(3)	99(4)	63(3)	11(3)	-16(3)	-14(3)
C4'	68(4)	82(3)	45(3)	1(2)	-7(3)	-14(3)
C5'	51(3)	56(2)	43(2)	-5.3(17)	-4(3)	-17(3)
C6'	41(3)	53(2)	48(2)	-4.4(19)	3(2)	-17.8(19)
C7'	59(3)	57(2)	42(2)	-4.3(18)	9(3)	-12(3)
C8'	65(3)	54(3)	54(3)	6(2)	4(3)	-6(3)
C9'	55(3)	53(2)	59(3)	-4(2)	9(2)	-9(2)
C10'	60(3)	61(3)	43(2)	-10(2)	10(2)	-18(3)
Cl1'	48.7(7)	63.0(6)	75.9(8)	6.5(5)	-5.8(7)	-11.1(6)
Cl2'	86.7(11)	70.6(8)	84.1(9)	-3.7(6)	22.0(10)	10.7(8)
O1'	40.4(19)	78(2)	57(2)	-20.1(18)	5.6(17)	-5.8(18)

Table S14 Bond Lengths.

Atom	Atom	Length/Å	Atom	Atom	Length/Å
C1	C2	1.506(7)	C1'	C2'	1.521(7)
C1	C6	1.513(6)	C1'	C6'	1.512(7)
C1	O1	1.420(6)	C1'	O1'	1.425(6)
C2	C3	1.519(7)	C2'	C3'	1.505(8)
C2	Cl1	1.812(5)	C2'	Cl1'	1.814(5)
C3	C4	1.524(7)	C3'	C4'	1.517(9)
C4	C5	1.517(6)	C4'	C5'	1.505(7)
C5	C6	1.395(6)	C5'	C6'	1.402(7)
C5	C10	1.397(7)	C5'	C10'	1.386(8)
C6	C7	1.400(6)	C6'	C7'	1.392(7)
C7	C8	1.373(7)	C7'	C8'	1.375(8)
C8	C9	1.372(7)	C8'	C9'	1.371(7)
C9	C10	1.371(7)	C9'	C10'	1.387(8)
C9	Cl2	1.751(5)	C9'	Cl2'	1.750(6)

Table S15 Bond Angles.

Atom	Atom	Atom	Angle/°	Atom	Atom	Atom	Angle/°
C2	C1	C6	112.7(4)	C6'	C1'	C2'	112.5(4)
O1	C1	C2	112.2(4)	O1'	C1'	C2'	112.9(4)
O1	C1	C6	109.2(4)	O1'	C1'	C6'	109.2(4)
C1	C2	C3	110.7(4)	C1'	C2'	C11'	110.6(3)
C1	C2	C11	110.1(4)	C3'	C2'	C1'	110.2(5)
C3	C2	C11	109.9(4)	C3'	C2'	C11'	109.6(4)
C2	C3	C4	111.5(4)	C2'	C3'	C4'	113.6(5)
C5	C4	C3	113.1(4)	C5'	C4'	C3'	115.0(5)
C6	C5	C4	122.3(4)	C6'	C5'	C4'	121.9(5)
C6	C5	C10	119.2(4)	C10'	C5'	C4'	119.0(4)
C10	C5	C4	118.4(4)	C10'	C5'	C6'	119.0(4)
C5	C6	C1	120.8(4)	C5'	C6'	C1'	120.5(5)
C5	C6	C7	119.1(4)	C7'	C6'	C1'	120.2(4)
C7	C6	C1	120.1(4)	C7'	C6'	C5'	119.3(5)
C8	C7	C6	121.4(4)	C8'	C7'	C6'	121.2(4)
C9	C8	C7	118.5(5)	C9'	C8'	C7'	119.2(5)
C8	C9	C12	118.9(4)	C8'	C9'	C10'	121.0(5)
C10	C9	C8	122.1(5)	C8'	C9'	C12'	119.4(4)
C10	C9	C12	119.0(4)	C10'	C9'	C12'	119.7(4)
C9	C10	C5	119.7(4)	C5'	C10'	C9'	120.3(4)

Table S16 Hydrogen Bonds.

D	H	A	d(D-H)/Å	d(H-A)/Å	d(D-A)/Å	D-H-A/°
O1	H1A	O1 ¹	0.71(7)	2.05(7)	2.747(6)	165(7)
O1'	H1'A	O1 ²	0.76(9)	2.03(9)	2.771(6)	163(9)

¹1/2-X,1-Y,1/2+Z; ²3/2-X,1-Y,-1/2+Z

Table S17 Torsion Angles.

A	B	C	D	Angle/°	A	B	C	D	Angle/°
C1	C2	C3	C4	61.4(6)	C1'	C2'	C3'	C4'	57.5(6)
C1	C6	C7	C8	-178.1(4)	C1'	C6'	C7'	C8'	-178.5(4)
C2	C1	C6	C5	23.3(6)	C2'	C1'	C6'	C5'	27.0(6)
C2	C1	C6	C7	-159.2(4)	C2'	C1'	C6'	C7'	-154.9(4)
C2	C3	C4	C5	-42.4(6)	C2'	C3'	C4'	C5'	-34.8(7)
C3	C4	C5	C6	15.2(7)	C3'	C4'	C5'	C6'	7.9(7)
C3	C4	C5	C10	-168.2(4)	C3'	C4'	C5'	C10'	-174.7(5)
C4	C5	C6	C1	-5.5(7)	C4'	C5'	C6'	C1'	-4.4(7)
C4	C5	C6	C7	176.9(4)	C4'	C5'	C6'	C7'	177.5(5)
C4	C5	C10	C9	-178.0(5)	C4'	C5'	C10'	C9'	-177.2(5)

C5 C6 C7 C8	-0.6(7)	C5' C6' C7' C8'	-0.3(8)
C6 C1 C2 C3	-50.6(5)	C6' C1' C2' C3'	-52.6(6)
C6 C1 C2 C11	71.2(4)	C6' C1' C2' C11'	68.8(5)
C6 C5 C10 C9	-1.2(7)	C6' C5' C10' C9'	0.2(8)
C6 C7 C8 C9	1.6(8)	C6' C7' C8' C9'	0.2(8)
C7 C8 C9 C10	-2.6(8)	C7' C8' C9' C10'	0.2(8)
C7 C8 C9 C12	178.3(4)	C7' C8' C9' C12'	-179.7(4)
C8 C9 C10 C5	2.4(8)	C8' C9' C10' C5'	-0.4(8)
C10 C5 C6 C1	177.9(4)	C10' C5' C6' C1'	178.2(4)
C10 C5 C6 C7	0.3(7)	C10' C5' C6' C7'	0.1(7)
C11 C2 C3 C4	-60.4(5)	C11' C2' C3' C4'	-64.5(6)
C12 C9 C10 C5	-178.5(4)	C12' C9' C10' C5'	179.5(4)
O1 C1 C2 C3	-174.2(4)	O1' C1' C2' C3'	-176.6(4)
O1 C1 C2 C11	-52.5(5)	O1' C1' C2' C11'	-55.2(5)
O1 C1 C6 C5	148.6(4)	O1' C1' C6' C5'	153.2(4)
O1 C1 C6 C7	-33.8(6)	O1' C1' C6' C7'	-28.7(6)

Table S18 Hydrogen Atom Coordinates ($\text{\AA} \times 10^4$) and Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$).

Atom	x	y	z	U(eq)
H1	2765	6934	7737	57
H2	2947	6033	7033	64
H3A	2586	7091	6495	67
H3B	4109	6569	6045	67
H4A	8051	7086	6217	62
H4B	6091	7702	6126	62
H7	6881	7386	8665	63
H8	9855	8281	8684	70
H10	9755	8346	6709	61
H1A	3870(130)	6190(30)	8400(30)	60(19)
H1'	7068	4546	4311	62
H2'	6875	3480	4781	68
H3'A	7292	4349	5541	83
H3'B	5570	3765	5853	83
H4'A	1921	4388	5851	78
H4'B	4053	4951	5990	78
H7'	2788	5141	3473	63
H8'	-268	5999	3642	69
H10'	64	5660	5590	66
H1'A	6180(180)	3880(40)	3510(40)	120(30)

Experimental

The crystal was kept at 293(2) K during data collection. Using Olex2, the structure was solved with the ShelXS structure solution program using Direct Methods and refined with the ShelXL refinement package using Least Squares minimisation.

Crystal structure determination

Crystal Data for $C_{10}H_{10}Cl_2O$ ($M = 217.08$ g/mol): orthorhombic, space group $P2_12_12_1$ (no. 19), $a = 5.04728(18)$ Å, $b = 19.5159(6)$ Å, $c = 20.2528(8)$ Å, $V = 1994.94(12)$ Å³, $Z = 8$, $T = 293(2)$ K, $\mu(\text{CuK}\alpha) = 5.490$ mm⁻¹, $D_{\text{calc}} = 1.446$ g/cm³, 8713 reflections measured ($8.732^\circ \leq 2\theta \leq 134.138^\circ$), 3534 unique ($R_{\text{int}} = 0.0378$, $R_{\text{sigma}} = 0.0418$) which were used in all calculations. The final R_1 was 0.0464 ($I > 2\sigma(I)$) and wR_2 was 0.1265 (all data).

Refinement model description

Number of restraints - 0, number of constraints - unknown.

Details:

1. Fixed Uiso

At 1.2 times of:

All C(H) groups, All C(H,H) groups

2.a Ternary CH refined with riding coordinates:

C1(H1), C2(H2), C1'(H1'), C2'(H2')

2.b Secondary CH2 refined with riding coordinates:

C3(H3A,H3B), C4(H4A,H4B), C3'(H3'A,H3'B), C4'(H4'A,H4'B)

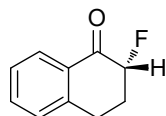
2.c Aromatic/amide H refined with riding coordinates:

C7(H7), C8(H8), C10(H10), C7'(H7'), C8'(H8'), C10'(H10')

7. References

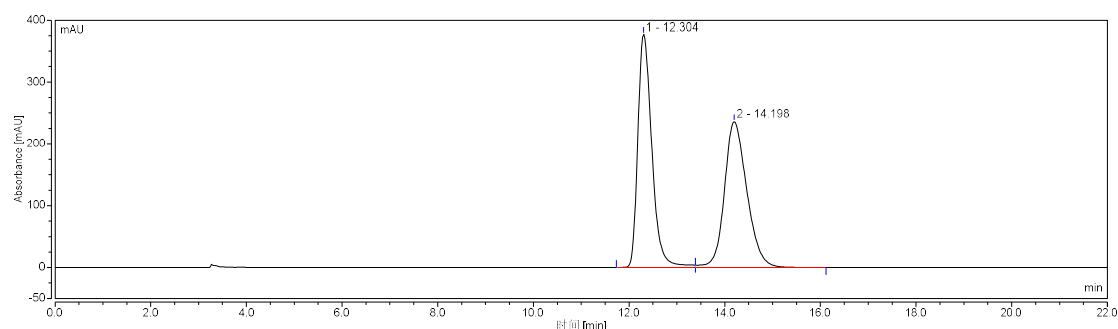
1. K. Shibatomi, H. Yamamoto, *Angew. Chem. Int. Ed.*, 2008, **47**, 5796.
2. J. D. Hamel, M. Cloutier, J. F. Paquin, *Org. Lett.*, 2016, **18**, 1852.
3. B. Šket, N. Zupančič, M. Zupan, *J. Fluorine Chem.*, 1989, **45**, 313.
4. T. Maji, A. Karmakar, O. Reiser, *J. Org. Chem.*, 2010, **76**, 736.
5. S. Liu, Y. Zhou, Y. Sui, H. Liu, H. Zhou, *Org. Chem. Front.*, 2017, **4**, 2175.

8. Characterization of products

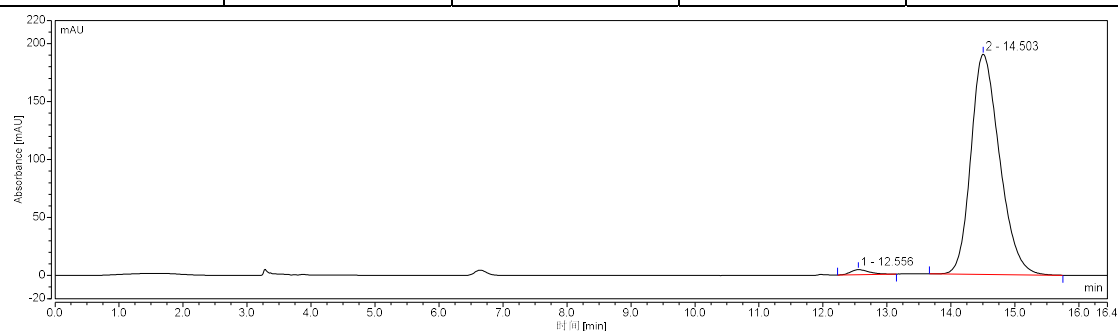


2a: white solid; Mp 44.6–46.0 °C; 15.1 mg, 92% yield; 96% ee (On a 1.0 mmol scale: 141.0 mg, 86% yield; 97% ee); $[\alpha]_D^{22} +157.1$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.99 (d, J = 7.8 Hz, 1H), 7.46 (t, J = 7.5 Hz, 1H), 7.30 (d, J = 7.6 Hz, 1H), 7.21 (d, J = 5.8 Hz, 1H), 5.08 (ddd, J = 47.9, 12.7, 5.2 Hz, 1H), 3.07 (dd, J = 9.1, 3.8 Hz, 2H), 2.57–2.45 (m, 1H), 2.36–2.20 (m, 1H); ^{13}C NMR (75 MHz, CDCl_3) δ 193.3 (d, J = 14.7 Hz), 143.0 (d, J = 1.4 Hz), 134.1, 131.2, 128.6, 127.8 (d, J = 2.2 Hz), 127.1, 91.2 (d, J = 188.0 Hz), 30.1 (d, J = 19.1 Hz), 27.0 (d, J = 11.5 Hz); ^{19}F NMR (376 MHz, CDCl_3) δ -190.3; HRMS (ESI) m/z 187.0541 ($\text{M}+\text{Na}^+$), calc. for $\text{C}_{10}\text{H}_9\text{OFNa}^+$ 187.0535.

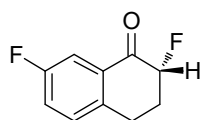
The ee was determined by HPLC analysis: LUX CELLULOSE-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 12.6 min (minor) and 14.5 min (major).



Entry	Retention Time	Area	Height	%Area
1	12.304	127.9981	376.82	49.98
2	14.198	128.1223	235.75	50.02

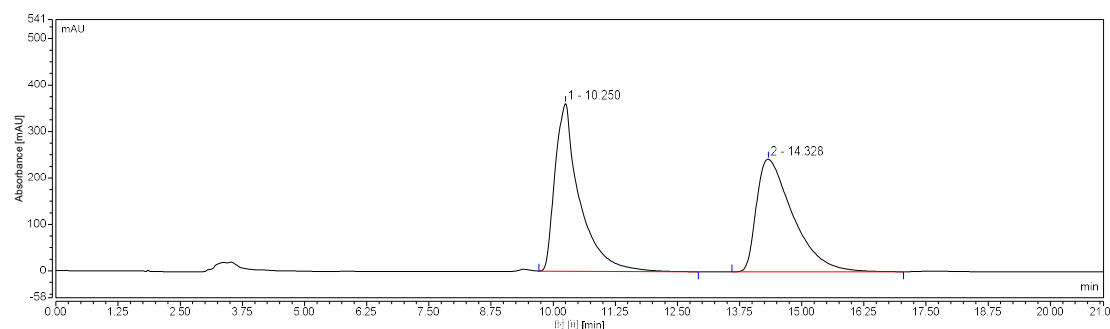


Entry	Retention Time	Area	Height	%Area
1	12.556	1.4671	4.39	1.47
2	14.503	98.3747	189.86	98.53

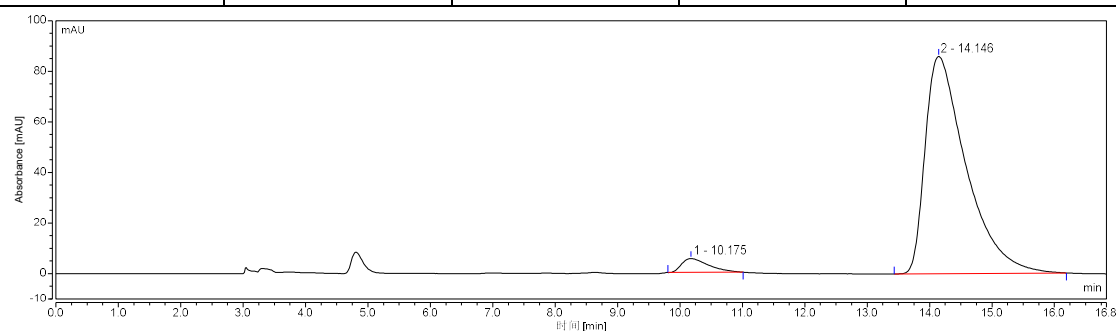


2b: yellow solid; Mp 119.8–121.0 °C; 17.5 mg, 96% yield; 92% ee; $[\alpha]_D^{22} +70.0$ (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.70 (dd, *J* = 8.8, 2.0 Hz, 1H), 7.29–7.19 (m, 2H), 5.14 (ddd, *J* = 47.7, 12.6, 5.1 Hz, 1H), 3.10 (dd, *J* = 8.1, 3.6 Hz, 2H), 2.57 (qd, *J* = 9.3, 4.3 Hz, 1H), 2.43–2.27 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 192.3 (dd, *J* = 15.0 Hz, 1.8 Hz), 161.7 (d, *J* = 247.2 Hz), 138.8 (dd, *J* = 3.0 Hz, 1.4 Hz), 132.8 (dd, *J* = 6.6 Hz, 1.2 Hz), 130.6 (d, *J* = 7.3 Hz), 121.6 (d, *J* = 22.4 Hz), 113.5 (dd, *J* = 22.2 Hz, 2.3 Hz), 90.9 (dd, *J* = 188.3 Hz, 1.1 Hz), 30.0 (d, *J* = 19.2 Hz), 26.3 (d, *J* = 11.5 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ -114.1, -191.0; HRMS (ESI) *m/z* 205.0438 (*M*+Na)⁺, calc. for C₁₀H₈OF₂Na⁺ 205.0441.

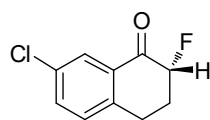
The ee was determined by HPLC analysis: CHIRALPAK AS-H (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 10.2 min (minor) and 14.1min (major).



Entry	Retention Time	Area	Height	%Area
1	10.250	205.5293	361.14	49.91
2	14.328	206.2378	242.43	50.09

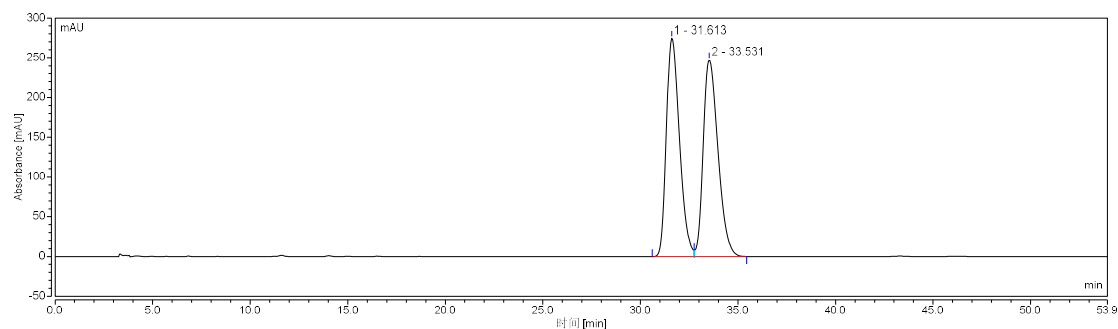


Entry	Retention Time	Area	Height	%Area
1	10.175	2.7524	5.49	4.04
2	14.146	65.2943	85.89	95.96

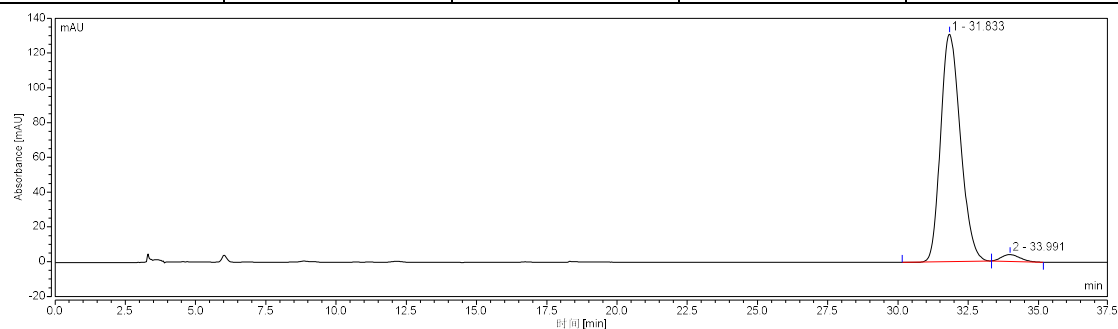


2c: white solid; Mp 119.8–121.5 °C; 16.7 mg, 84% yield; 95% ee; $[\alpha]_D^{22}$ +27.1 (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.07 (d, *J* = 2.1 Hz, 1H), 7.53 (dd, *J* = 8.2, 2.3 Hz, 1H), 7.30 (d, *J* = 5.7 Hz, 1H), 5.19 (ddd, *J* = 47.7, 12.6, 5.1 Hz, 1H), 3.15 (dd, *J* = 8.8, 4.2 Hz, 2H), 2.62 (qd, *J* = 9.1, 4.3 Hz, 1H), 2.48–2.32 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 192.1 (d, *J* = 15.0 Hz), 141.2 (d, *J* = 1.3 Hz), 134.1, 133.5, 132.5 (d, *J* = 1.2 Hz), 130.2, 127.4 (d, *J* = 2.2 Hz), 90.8 (d, *J* = 188.7 Hz), 29.8 (d, *J* = 19.3 Hz), 26.4 (d, *J* = 11.4 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ –190.9; HRMS (ESI) *m/z* 221.0145 (M+Na)⁺, calc. for C₁₀H₈OFNaCl⁺ 221.0145.

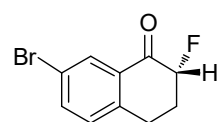
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 34.0min (minor) and 31.8 min (major).



Entry	Retention Time	Area	Height	%Area
1	31.613	214.5172	274.63	49.79
2	33.531	216.3284	247.32	50.21



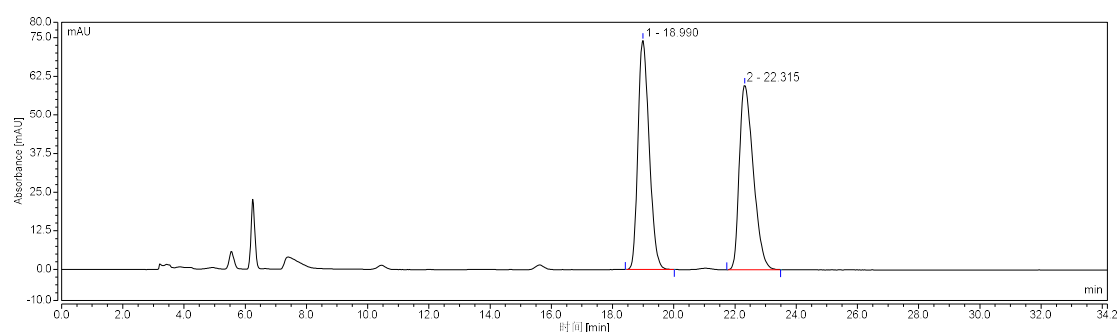
Entry	Retention Time	Area	Height	%Area
1	31.833	107.0252	130.86	97.28
2	33.991	2.9914	3.91	2.72



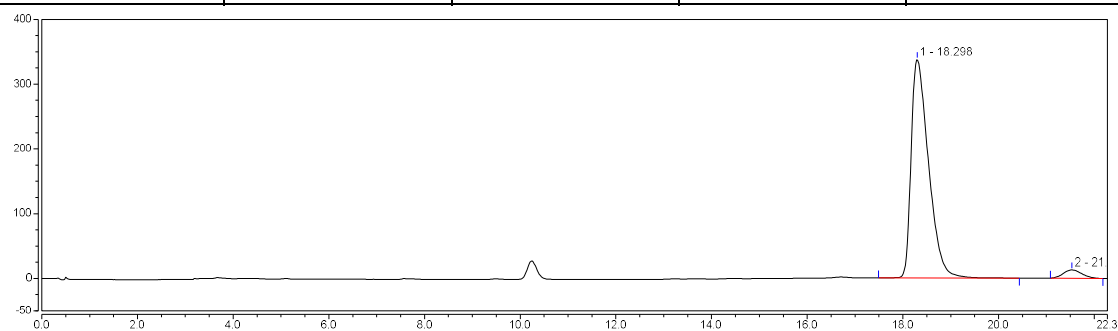
2d: white solid; Mp 122.1–123.8 °C; 22.6 mg, 93% yield; 92% ee; $[\alpha]_D^{22}$ +234.1 (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.16 (d, *J* = 1.7 Hz,

1H), 7.62 (dd, $J = 8.2, 1.9$ Hz, 1H), 7.16 (d, $J = 8.2$ Hz, 1H), 5.14 (ddd, $J = 47.7, 12.7, 5.1$ Hz, 1H), 3.07 (dd, $J = 8.7, 4.0$ Hz, 2H), 2.57 (qd, $J = 9.3, 4.3$ Hz, 1H), 2.41–2.25 (m, 1H); ^{13}C NMR (75 MHz, CDCl_3) δ 192.1 (d, $J = 14.9$ Hz), 141.6 (d, $J = 1.4$ Hz), 136.9, 132.7 (d, $J = 1.1$ Hz), 130.5, 121.2, 90.8 (d, $J = 188.9$ Hz), 29.8 (d, $J = 19.3$ Hz), 26.5 (d, $J = 11.5$ Hz); ^{19}F NMR (376 MHz, CDCl_3) δ -190.8; HRMS (ESI) m/z 264.9637 ($\text{M}+\text{Na}$) $^+$, calc. for $\text{C}_{10}\text{H}_8\text{OFNaBr}^+$ 264.9640.

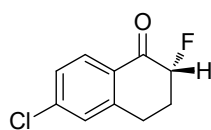
The ee was determined by HPLC analysis: CHIRALPAK ID (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 21.5 min (minor) and 18.3 min (major).



Entry	Retention Time	Area	Height	%Area
1	18.990	31.5624	74.07	50.07
2	22.315	31.4733	59.70	49.93



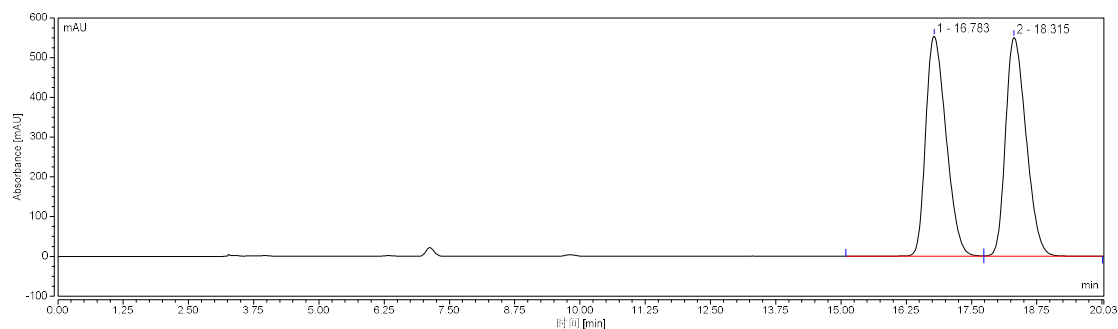
Entry	Retention Time	Area	Height	%Area
1	18.298	142.0097	337.21	96.09
2	21.531	5.7725	12.86	3.91



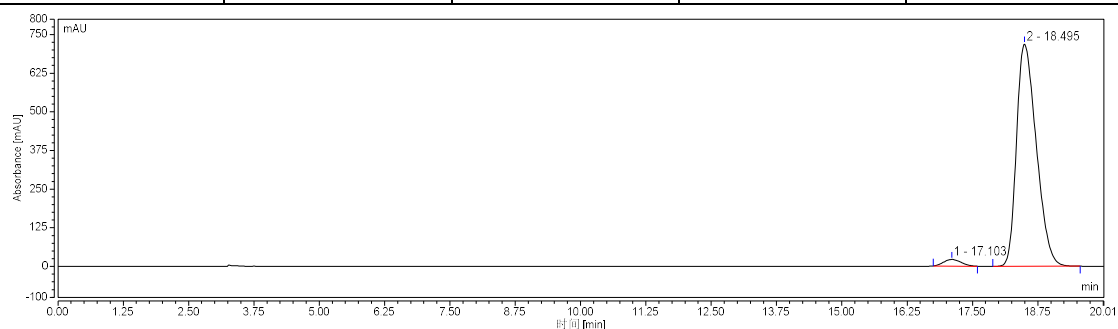
2e: white solid; Mp 106.9–108.8 °C; 15.9 mg, 80% yield; 95% ee; $[\alpha]_{\text{D}}^{22} +53.7$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.99 (d, $J = 8.4$ Hz, 1H), 7.32 (d, $J = 8.4$ Hz, 1H), 7.28 (s, 1H), 5.13 (ddd, $J = 47.7, 12.6, 5.1$ Hz, 1H), 3.10 (dd, $J = 9.0, 4.0$ Hz, 2H), 2.63–2.51 (m, 1H), 2.43–2.27 (m, 1H); ^{13}C NMR (75 MHz, CDCl_3) δ 192.2 (d, $J = 14.9$ Hz), 144.5 (d, $J = 1.4$ Hz), 140.6, 129.6 (d, $J = 0.9$ Hz),

129.4 (d, $J = 2.2$ Hz), 128.5, 127.7, 90.8 (d, $J = 188.3$ Hz), 29.8 (d, $J = 19.4$ Hz), 26.7 (d, $J = 11.5$ Hz); ^{19}F NMR (376 MHz, CDCl_3) δ -190.7; HRMS (ESI) m/z 199.0334 ($\text{M}+\text{H}$) $^+$, calc. for $\text{C}_{10}\text{H}_9\text{OCIF}^+$ 199.0326.

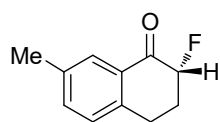
The ee was determined by HPLC analysis: CHIRALPAK ID (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 17.1 min (minor) and 18.5 min (major).



Entry	Retention Time	Area	Height	%Area
1	16.783	251.0569	553.69	49.95
2	18.315	251.5918	550.62	50.05

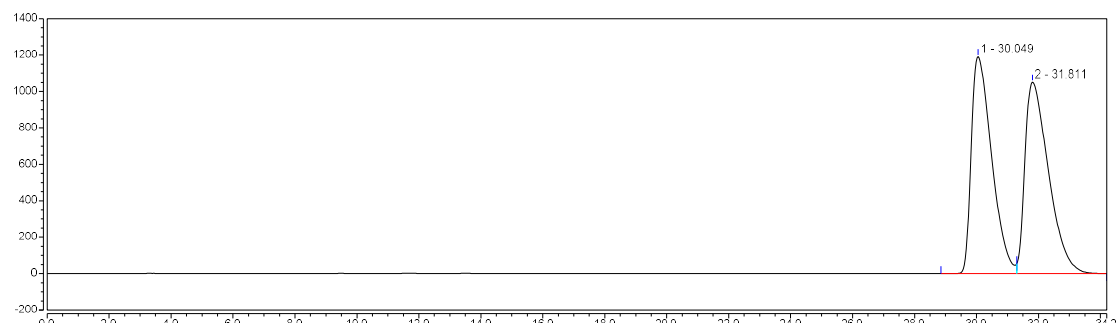


Entry	Retention Time	Area	Height	%Area
1	17.103	8.2298	21.37	2.54
2	18.495	315.2299	719.29	97.46

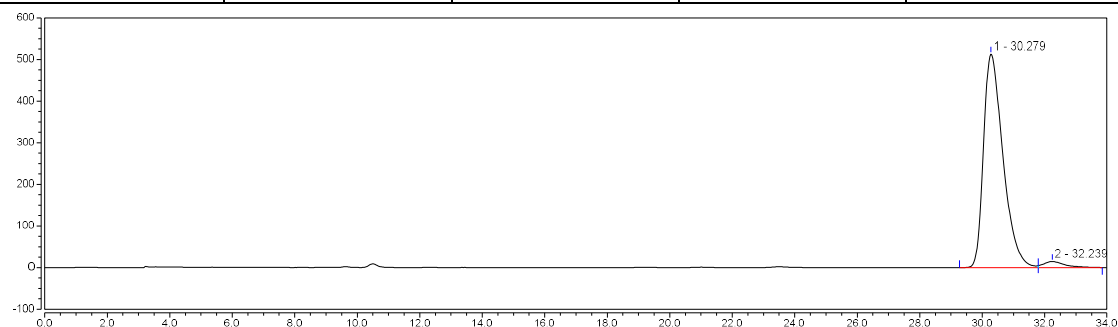


2f: white solid; Mp 51.7–53.4 °C; 13.5 mg, 76% yield; 94% ee; $[\alpha]_{\text{D}}^{22}$ +74.6 (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.85(s, 1H), 7.33 (d, $J = 7.8$ Hz, 1H), 7.15 (d, $J = 7.8$ Hz, 1H), 5.13 (ddd, $J = 47.9, 12.8, 5.2$ Hz, 1H), 3.08 (d, $J = 5.7$ Hz, 2H), 2.61–2.50 (m, 1H), 2.36 (s, 3H), 2.35–2.23 (m, 1H); ^{13}C NMR (75 MHz, CDCl_3) δ 193.6 (d, $J = 14.5$ Hz), 140.1 (d, $J = 1.4$ Hz), 136.9, 135.2, 130.9 (d, $J = 0.7$ Hz), 128.5, 127.7 (d, $J = 2.2$ Hz), 91.3 (d, $J = 187.9$ Hz), 30.2 (d, $J = 18.9$ Hz), 26.6 (d, $J = 11.7$ Hz), 20.8; ^{19}F NMR (376 MHz, CDCl_3) δ -190.3; HRMS (ESI) m/z 201.0695 ($\text{M}+\text{Na}$) $^+$, calc. for $\text{C}_{11}\text{H}_{11}\text{OFNa}^+$ 201.0692.

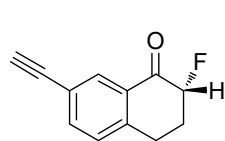
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 32.3 min (minor) and 30.3min (major).



Entry	Retention Time	Area	Height	%Area
1	30.049	909.1999	1191.13	49.56
2	31.811	925.2250	1051.03	50.44

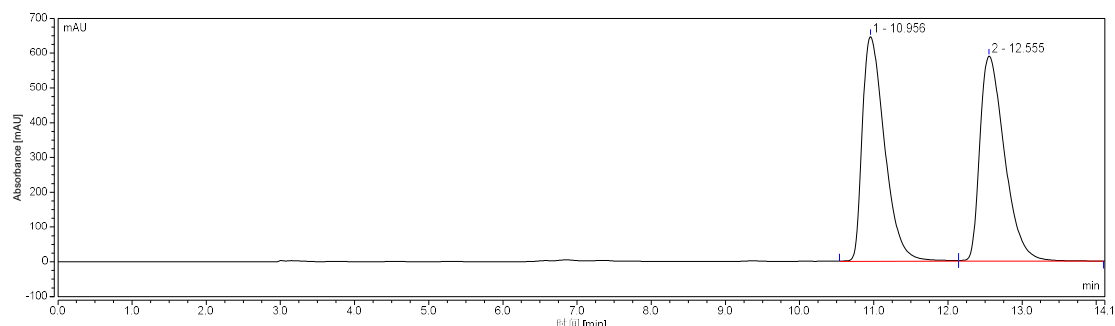


Entry	Retention Time	Area	Height	%Area
1	30.279	369.5150	512.59	97.05
2	32.239	11.2422	14.38	2.95

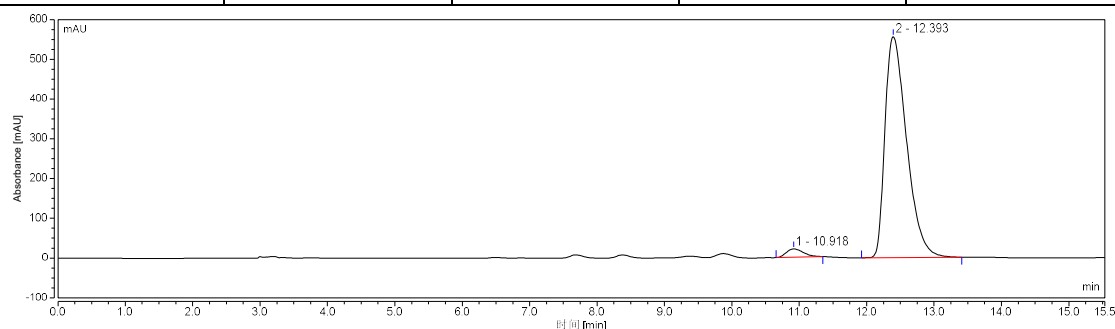


2g: yellow solid; Mp 162.8–163.4 °C; 15.2 mg, 81% yield; 94% ee; $[\alpha]_D^{22} +48.3$ (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.21 (s, 1H), 7.64 (d, *J* = 7.7 Hz, 1H), 7.29 (s, 1H), 5.18 (ddd, *J* = 47.7, 12.6, 5.0 Hz, 1H), 3.16 (d, *J* = 5.2 Hz, 2H), 3.14 (s, 1H), 2.66–2.57 (m, 1H), 2.47–2.31 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 192.4 (d, *J* = 14.9 Hz), 143.3 (d, *J* = 1.2 Hz), 137.0, 131.4 (d, *J* = 2.2 Hz), 131.2, 128.8, 121.3, 90.9 (d, *J* = 188.6 Hz), 82.0, 78.3, 29.8 (d, *J* = 19.2 Hz), 26.9 (d, *J* = 11.5 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ –190.6; HRMS (ESI) *m/z* 211.0539 (M+Na)⁺, calc. for C₁₂H₁₁OFNa⁺ 211.0535.

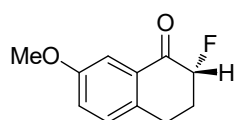
The ee was determined by HPLC analysis: CHIRALCEL OD-H (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 10.9 min (minor) and 12.4 min (major).



Entry	Retention Time	Area	Height	%Area
1	10.956	221.9637	644.15	49.83
2	12.555	223.4855	587.31	50.17

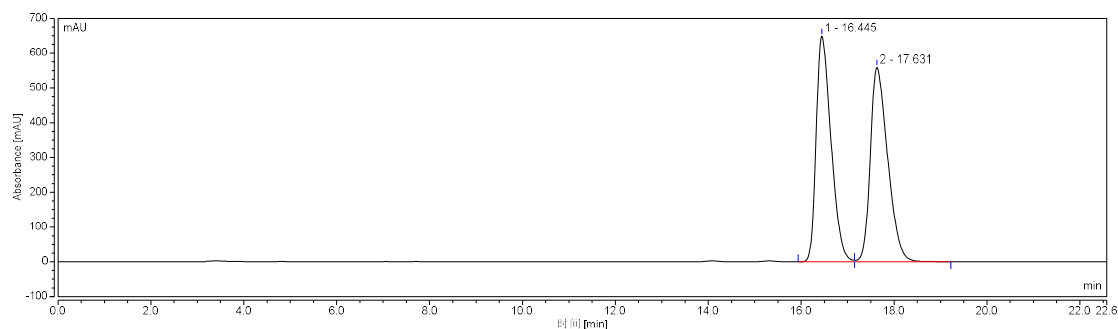


Entry	Retention Time	Area	Height	%Area
1	10.918	6.7516	21.82	3.17
2	12.393	206.3097	556.03	96.83

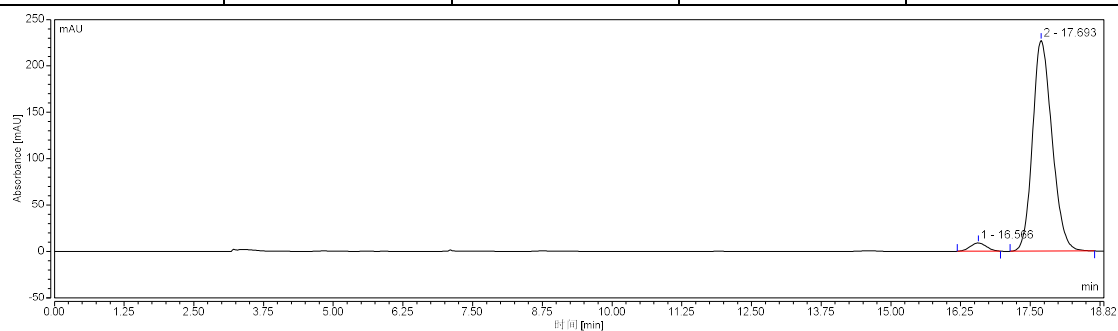


2h: white solid; Mp 87.2–88.7 °C; 15.2 mg, 86% yield; 94% ee; $[\alpha]_D^{22} +82.7$ (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.50 (d, *J* = 2.6 Hz, 1H), 7.17 (d, *J* = 8.5 Hz, 1H), 7.09 (dd, *J* = 8.5, 2.7 Hz, 1H), 5.13 (ddd, *J* = 47.9, 12.8, 5.2 Hz, 1H), 3.84 (s, 3H), 3.06 (dd, *J* = 9.3, 3.9 Hz, 2H), 2.61–2.50 (m, 1H), 2.40–2.24 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 193.3 (d, *J* = 14.7 Hz), 158.6, 135.6 (d, *J* = 1.4 Hz), 132.0 (d, *J* = 1.0 Hz), 129.9, 122.7, 109.3 (d, *J* = 2.3 Hz), 91.3 (d, *J* = 187.8 Hz), 55.5, 30.3 (d, *J* = 19.0 Hz), 26.2 (d, *J* = 11.7 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ –190.4; HRMS (ESI) *m/z* 217.0647 (M+Na)⁺, calc. for C₁₁H₁₁O₂FN⁺ 217.0641.

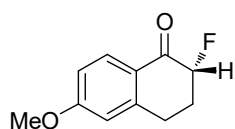
The ee was determined by HPLC analysis: CHIRALPAK ID (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 16.6 min (minor) and 17.7 min (major).



Entry	Retention Time	Area	Height	%Area
1	16.445	240.4402	649.34	49.77
2	17.631	242.6733	559.53	50.23



Entry	Retention Time	Area	Height	%Area
1	16.566	3.0162	8.94	3.17
2	17.693	92.2742	227.09	96.83



2i: white solid; Mp 113.4–115.0 °C; 13.2 mg, 68% yield; 97% ee; $[\alpha]_D^{22}$

+51.4 (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.03 (d, *J* = 8.8 Hz,

1H), 6.86 (dd, *J* = 8.8, 2.4 Hz, 1H), 6.69 (d, *J* = 2.1 Hz, 1H), 5.09 (ddd, *J*

= 47.9, 12.5, 5.1 Hz, 1H), 3.86 (s, 3H), 3.07 (dd, *J* = 9.1, 4.0 Hz, 2H), 2.59–2.47 (m, 1H),

2.40–2.24 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 192.0 (d, *J* = 14.7 Hz), 164.2, 145.5 (d, *J* =

1.4 Hz), 130.3 (d, *J* = 2.3 Hz), 124.6, 113.8, 112.4, 90.9 (d, *J* = 185.6 Hz), 55.5, 30.1 (d, *J* =

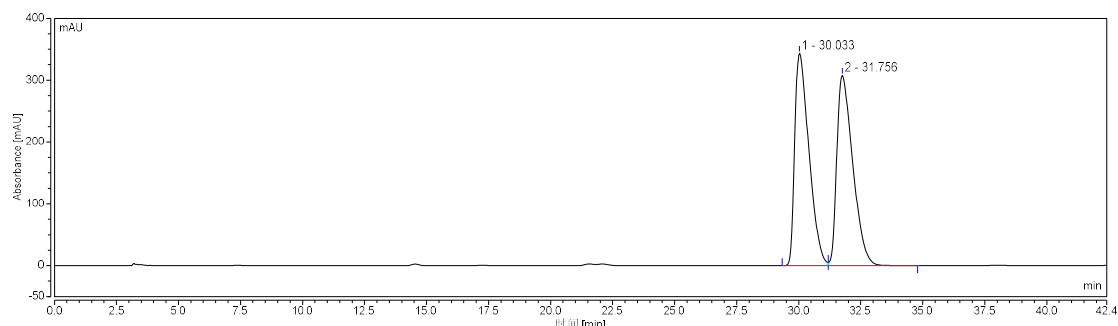
19.1 Hz), 27.2 (d, *J* = 11.5 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ –190.4; HRMS (ESI) *m/z*

217.0646 (M+Na)⁺, calc. for C₁₁H₁₁O₂FN⁺ 217.0641.

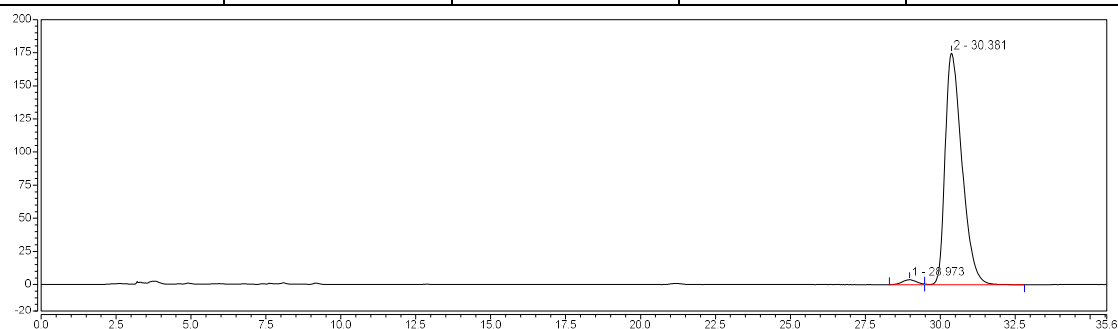
The ee was determined by HPLC analysis: CHIRALPAK ID (4.6 mm i.d. x 250 mm);

Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 29.0 min

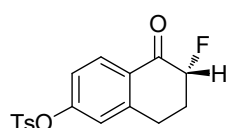
(minor) and 30.4 min (major).



Entry	Retention Time	Area	Height	%Area
1	30.033	228.3652	343.65	49.83
2	31.756	229.8979	307.77	50.17

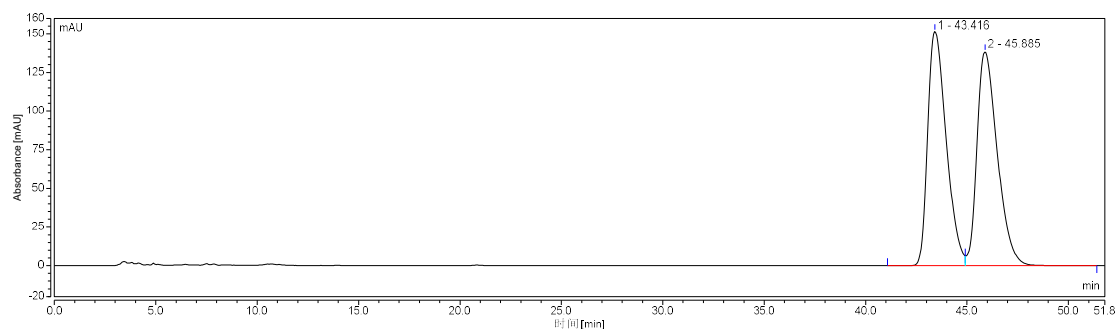


Entry	Retention Time	Area	Height	%Area
1	28.973	2.0324	3.70	1.45
2	30.381	114.0388	174.82	98.55

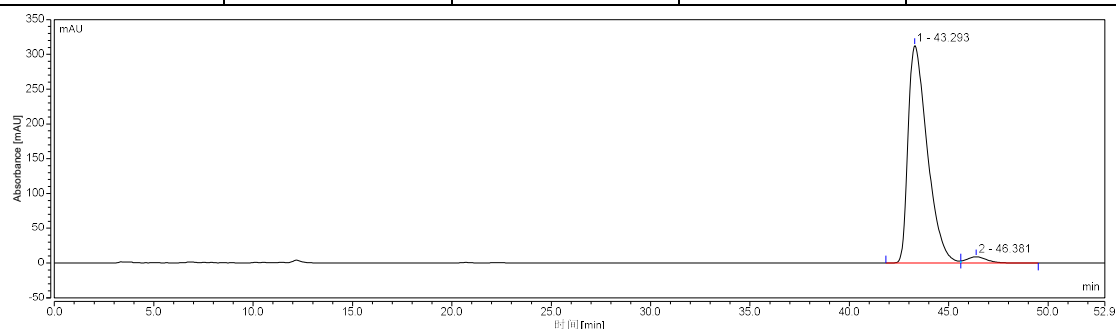


2j: yellow solid; Mp 87.2–88.7 °C; 26.1mg, 78% yield; 95% ee; $[\alpha]_D^{22}$ +29.3 (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.94 (d, *J* = 8.6 Hz, 1H), 7.70 (d, *J* = 8.2 Hz, 2H), 7.33 (d, *J* = 8.0 Hz, 2H), 7.04 (s, 1H), 6.82 (d, *J* = 8.5 Hz, 1H), 5.11 (ddd, *J* = 47.7, 12.6, 5.1 Hz, 1H), 3.08 (dd, *J* = 8.7, 3.7 Hz, 2H), 2.68–2.49 (m, 1H), 2.45 (s, 3H), 2.40–2.23 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 192.0 (d, *J* = 14.9 Hz), 153.4, 145.9, 145.1 (d, *J* = 1.4 Hz), 131.9, 129.9, 129.8, 129.8, 128.3, 122.3, 121.0, 90.7 (d, *J* = 187.3 Hz), 29.7 (d, *J* = 19.3 Hz), 26.8 (d, *J* = 11.5 Hz), 21.7; ¹⁹F NMR (376 MHz, CDCl₃) δ –190.8; HRMS (ESI) *m/z* 357.0578 (M+Na)⁺, calc. for C₁₇H₁₅O₄FNas⁺ 357.0573.

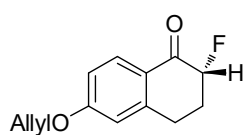
The ee was determined by HPLC analysis: CHIRALPAK IE (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 46.4 min (minor) and 43.3 min (major).



Entry	Retention Time	Area	Height	%Area
1	43.416	160.3057	151.45	49.95
2	45.885	160.6545	138.34	50.05

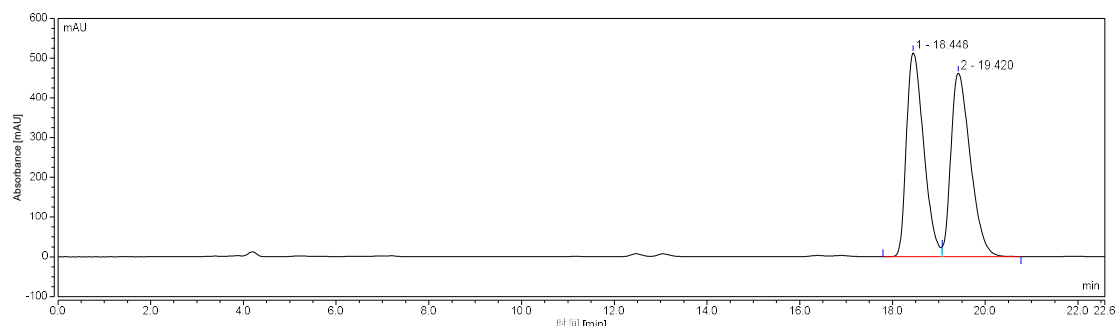


Entry	Retention Time	Area	Height	%Area
1	43.293	350.0930	312.40	97.34
2	46.381	10.3014	8.95	2.66

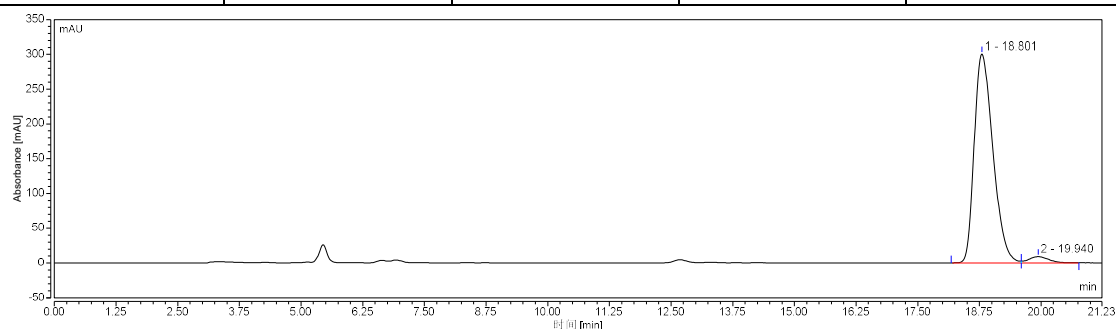


2k: white solid; Mp 100.9–102.2 °C; 15.2 mg, 69% yield; 94% ee; $[\alpha]_D^{22} +70.6$ (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.03 (d, *J* = 8.8 Hz, 1H), 6.87 (dd, *J* = 8.7, 2.1 Hz, 1H), 6.71 (s, 1H), 6.03 (ddd, *J* = 22.4, 10.5, 5.2 Hz, 1H), 5.37 (ddd, *J* = 13.9, 11.5, 1.1 Hz, 2H), 5.09 (ddd, *J* = 47.9, 12.4, 5.1 Hz, 1H), 4.60 (d, *J* = 5.2 Hz, 2H), 3.07 (dd, *J* = 8.8, 3.7 Hz, 2H), 2.54 (qd, *J* = 9.2, 4.2 Hz, 1H), 2.40–2.24 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 191.9 (d, *J* = 14.6 Hz), 163.2, 145.5 (d, *J* = 1.3 Hz), 132.2, 130.3 (d, *J* = 2.3 Hz), 124.7, 118.3, 114.3, 113.3, 90.9 (d, *J* = 185.6 Hz), 68.9, 30.1 (d, *J* = 19.2 Hz), 27.2 (d, *J* = 11.5 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ –190.4; HRMS (ESI) *m/z* 221.0979 (M+H)⁺, calc. for C₁₃H₁₄O₂F⁺ 221.0978.

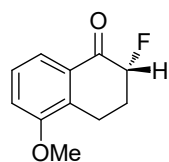
The ee was determined by HPLC analysis: CHIRALPAK IE (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 19.9 min (minor) and 18.8 min (major).



Entry	Retention Time	Area	Height	%Area
1	18.448	221.1624	513.09	49.73
2	19.420	223.5872	461.37	50.27

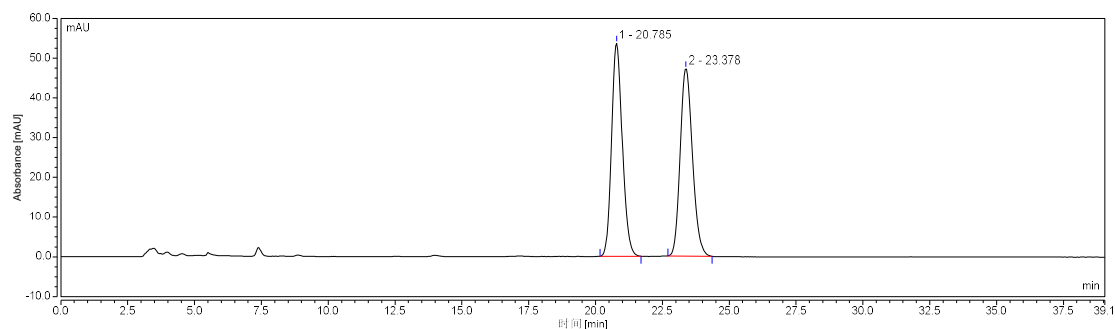


Entry	Retention Time	Area	Height	%Area
1	18.801	131.9842	300.36	96.96
2	19.940	4.1422	8.98	3.04

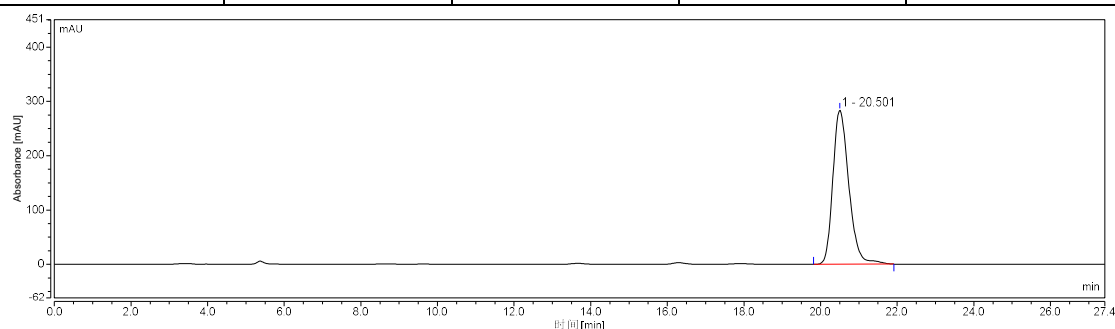


21: white solid; Mp 122.4–123.8 °C; 18.4 mg, 95% yield; >99% ee; $[\alpha]_D^{22} +26.9$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.50 (d, $J = 2.3$ Hz, 1H), 7.17 (d, $J = 8.5$ Hz, 1H), 7.09 (dd, $J = 8.5, 2.5$ Hz, 1H), 5.12 (ddd, $J = 47.9, 12.7, 5.1$ Hz, 1H), 3.83 (s, 3H), 3.05 (dd, $J = 8.9, 3.6$ Hz, 2H), 2.61–2.49 (m, 1H), 2.40–2.24 (m, 1H); ^{13}C NMR (75 MHz, CDCl_3) δ 193.3 (d, $J = 14.6$ Hz), 158.6, 135.6 (d, $J = 1.3$ Hz), 132.0 (d, $J = 0.7$ Hz), 129.9, 122.7, 109.3 (d, $J = 2.3$ Hz), 91.3 (d, $J = 186.6$ Hz), 55.5, 30.3 (d, $J = 18.8$ Hz), 26.2 (d, $J = 11.6$ Hz); ^{19}F NMR (376 MHz, CDCl_3) δ -190.4; HRMS (ESI) m/z 217.0647 ($\text{M}+\text{Na}$) $^+$, calc. for $\text{C}_{11}\text{H}_{11}\text{O}_2\text{FNa}^+$ 217.0641.

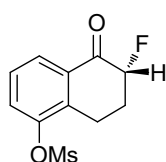
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 23.1 min (minor) and 20.5 min (major).



Entry	Retention Time	Area	Height	%Area
1	20.668	22.3796	47.30	50.21
2	23.215	22.1894	41.68	49.79

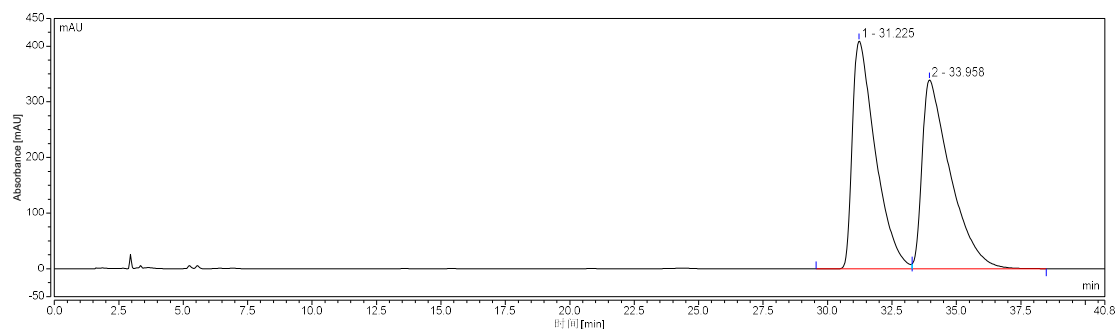


Entry	Retention Time	Area	Height	%Area
1	20.501	137.3159	283.76	100.00
2	23.1	0	0	0

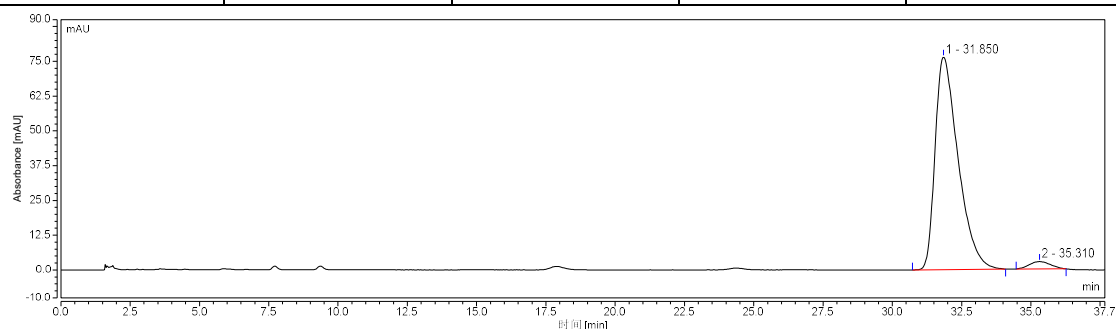


2m: white solid; Mp 104.3–105.9 °C; 17.6 mg, 68% yield; 94% ee; $[\alpha]_D^{22} +64.0$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 8.06 (d, $J = 7.8$ Hz, 1H), 7.54 (d, $J = 7.9$ Hz, 1H), 7.43 (t, $J = 7.9$ Hz, 1H), 5.18 (ddd, $J = 47.9, 12.8, 5.1$ Hz, 1H), 3.40–3.32 (m, 1H), 3.12–3.01 (m, 1H), 2.66–2.57 (m, 1H), 2.45–2.22 (m, 1H); ^{13}C NMR (75 MHz, CDCl_3) δ 192.1 (d, $J = 14.9$ Hz), 146.4, 136.5 (d, $J = 1.4$ Hz), 133.4 (d, $J = 0.7$ Hz), 128.2, 127.7, 126.8 (d, $J = 2.1$ Hz), 90.5 (d, $J = 188.9$ Hz), 38.5, 29.0 (d, $J = 19.5$ Hz), 21.7 (d, $J = 11.7$ Hz); ^{19}F NMR (376 MHz, CDCl_3) δ -191.1; HRMS (ESI) m/z 281.0259 ($\text{M}+\text{Na}^+$, calc. for $\text{C}_{11}\text{H}_{11}\text{O}_4\text{FNaS}^+$ 281.0260).

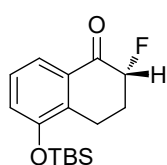
The ee was determined by HPLC analysis: CHIRALPAK ID (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 2.0 mL/min; 25 °C; 254 nm; retention time: 35.3 min (minor) and 31.8 min (major).



Entry	Retention Time	Area	Height	%Area
1	31.225	425.4863	409.32	49.70
2	33.958	430.6803	339.17	50.30

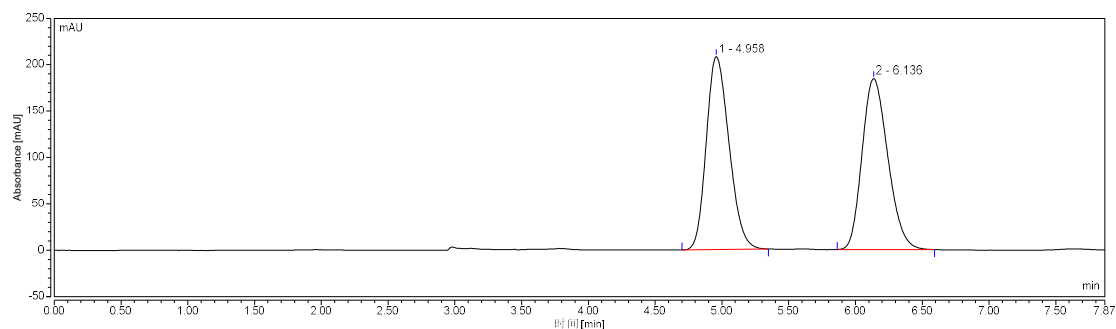


Entry	Retention Time	Area	Height	%Area
1	31.850	73.6254	76.38	96.92
2	35.310	2.3414	2.67	3.08

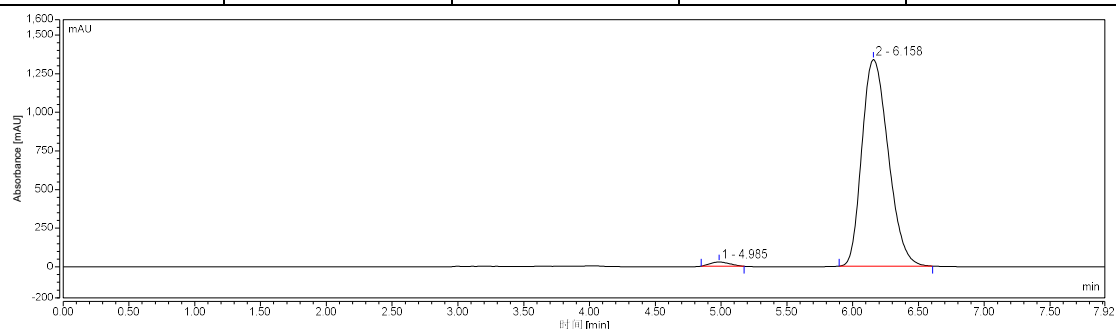


2n: yellow oil; 20.6 mg, 70% yield; 97% ee; $[\alpha]_D^{22} +50.5$ (*c* 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.99 (d, $J = 8.6$ Hz, 1H), 6.79 (dd, $J = 8.6, 2.1$ Hz, 1H), 6.66 (s, 1H), 5.10 (ddd, $J = 47.9, 12.5, 5.1$ Hz, 1H), 3.05 (dd, $J = 8.8, 4.0$ Hz, 2H), 2.54 (qd, $J = 9.2, 4.3$ Hz, 1H), 2.40–2.25 (m, 1H), 0.98 (s, 9H), 0.24 (s, 6H); ^{13}C NMR (75 MHz, CDCl_3) δ 192.2 (d, $J = 15.8$ Hz), 161.1, 145.4 (d, $J = 1.3$ Hz), 130.3 (d, $J = 2.3$ Hz), 125.1, 119.4, 119.0, 91.0 (d, $J = 185.7$ Hz), 30.1 (d, $J = 19.1$ Hz), 27.0 (d, $J = 11.5$ Hz), 25.5, 18.2, $-4.3, -4.3$; ^{19}F NMR (376 MHz, CDCl_3) δ -190.9 ; HRMS (ESI) m/z 295.1529 ($\text{M}+\text{H}$) $^+$, calc. for $\text{C}_{16}\text{H}_{24}\text{O}_2\text{FNSi}^+$ 295.1530.

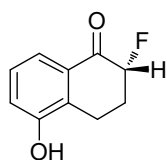
The ee was determined by HPLC analysis: CHIRALCEL OD-H (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 5.0 min (minor) and 6.2 min (major).



Entry	Retention Time	Area	Height	%Area
1	4.958	41.2355	208.11	49.99
2	6.136	41.2497	184.39	50.01

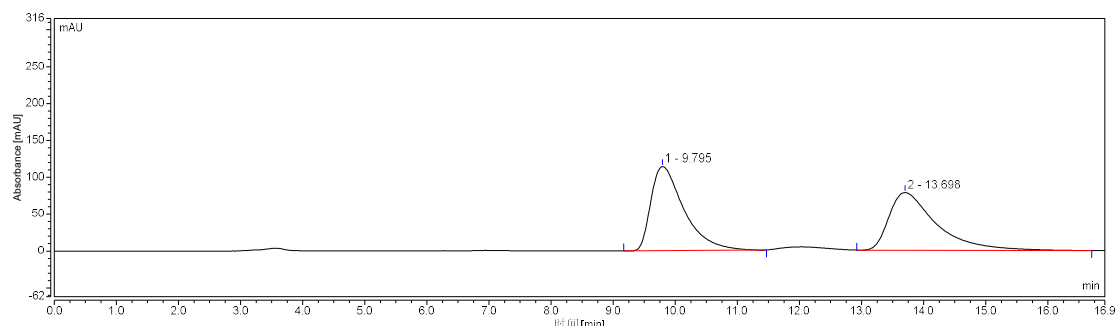


Entry	Retention Time	Area	Height	%Area
1	4.985	4.5785	26.63	1.45
2	6.158	310.1722	1336.86	98.55

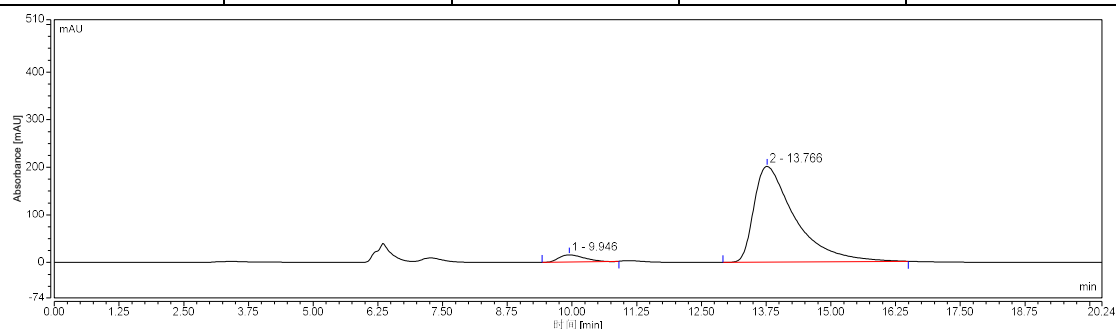


2o: white solid; Mp 174.9–196.4 °C; 20.6 mg, 72% yield; 92% ee; $[\alpha]_D^{22} +46.8$ (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CD₃OD) δ 7.46 (d, *J* = 7.7 Hz, 1H), 7.19 (t, *J* = 7.9 Hz, 1H), 7.02 (d, *J* = 7.9 Hz, 1H), 5.24 (ddd, *J* = 48.1, 13.1, 5.2 Hz, 1H), 3.23 (ddd, *J* = 17.9, 8.1, 4.7 Hz, 1H), 2.96–2.73 (m, 1H), 2.55 (tt, *J* = 14.6, 5.0 Hz, 1H), 2.41–2.08 (m, 1H); ¹³C NMR (75 MHz, CD₃OD) δ 196.3 (d, *J* = 14.2 Hz), 156.2, 133.3, 131.9 (d, *J* = 1.5 Hz), 128.4, 120.7, 118.8 (d, *J* = 2.2 Hz), 92.3 (d, *J* = 185.2 Hz), 30.4 (d, *J* = 18.7 Hz), 21.8 (d, *J* = 12.0 Hz); ¹⁹F NMR (376 MHz, CD₃OD) δ –192.6; HRMS (ESI) *m/z* 181.0671 (M+H)⁺, calc. for C₁₀H₁₀O₂F⁺ 181.0665.

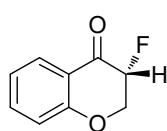
The ee was determined by HPLC analysis: CHIRALPAK AS-H (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 9.9 min (minor) and 13.8 min (major).



Entry	Retention Time	Area	Height	%Area
1	9.795	70.2708	113.95	50.47
2	13.698	68.9706	78.14	49.53



Entry	Retention Time	Area	Height	%Area
1	9.946	8.2645	14.92	4.18
2	13.766	184.7350	201.24	95.82



2p: white solid; Mp 71.8–73.2 °C; 14.8 mg, 89% yield; 93% ee; $[\alpha]_D^{22} +166.8$

(*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.92 (d, *J* = 7.9 Hz, 1H), 7.53

(dd, *J* = 11.3, 4.2 Hz, 1H), 7.09 (t, *J* = 7.5 Hz, 1H), 7.01 (d, *J* = 8.4 Hz, 1H),

5.17 (ddd, *J* = 47.0, 9.2, 4.8 Hz, 1H), 4.69–4.49 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 187.1

(d, *J* = 15.7 Hz), 161.2, 136.8, 127.6 (d, *J* = 1.3 Hz), 122.3, 119.4, 117.9, 85.5 (d, *J* = 187.0

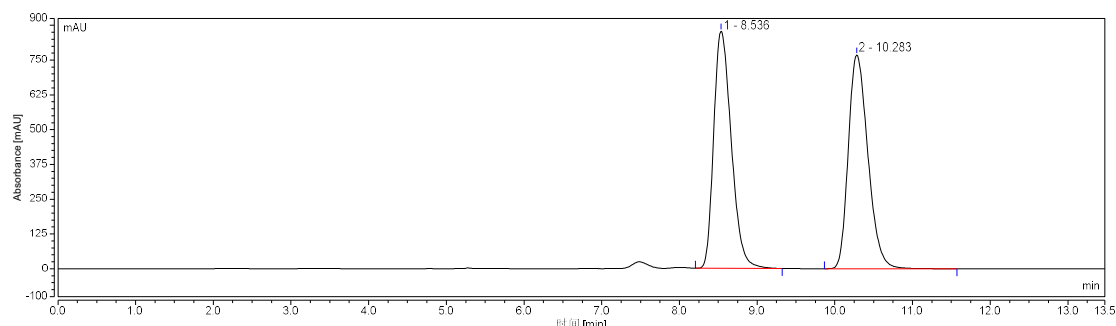
Hz), 68.6 (d, *J* = 25.8 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ –204.0; HRMS (ESI) *m/z*

189.0330 (M+Na)⁺, calc. for C₉H₇O₂FNa⁺ 189.0328.

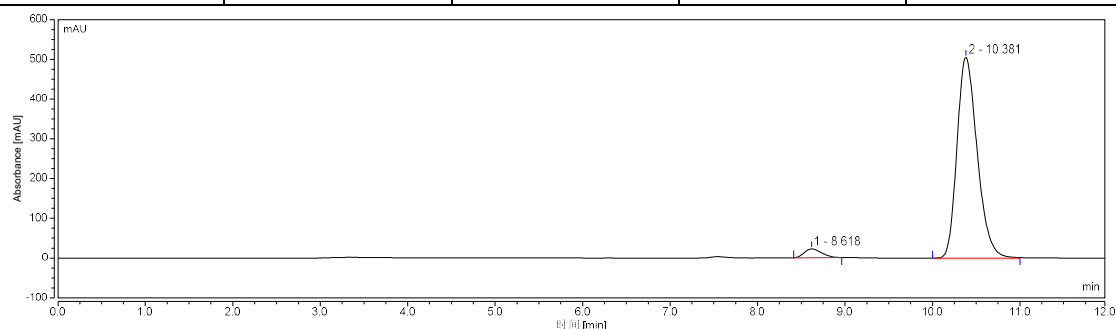
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm);

Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 8.6 min

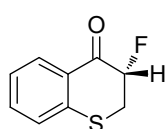
(minor) and 10.4 min (major).



Entry	Retention Time	Area	Height	%Area
1	8.536	222.9058	852.26	49.86
2	10.283	224.1198	768.28	50.14

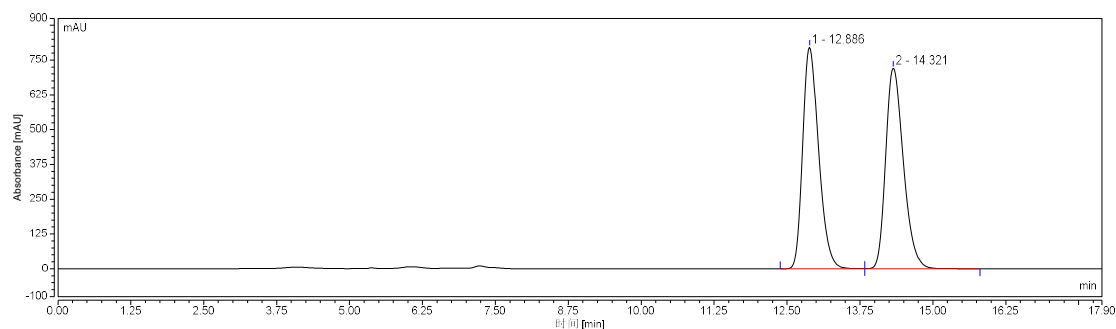


Entry	Retention Time	Area	Height	%Area
1	8.618	5.0461	22.03	3.60
2	10.381	135.1093	505.15	96.40

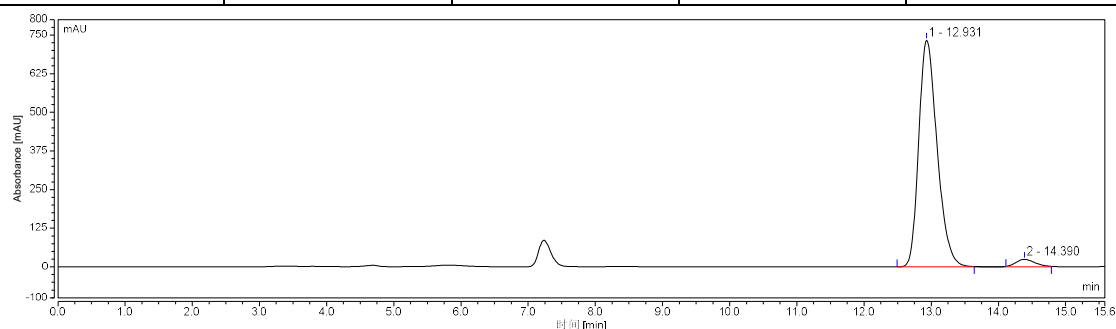


2q: white solid; Mp 87.3–88.9 °C; 16.4 mg, 90% yield; 93% ee; $[\alpha]_D^{22} +4.5$ (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.11 (*J* = 8.0 Hz, 1H), 7.46–7.41 (m, 1H), 7.28 (s, 1H), 7.26–7.21 (m, 1H), 5.43 (ddd, *J* = 47.5, 13.4, 4.9 Hz, 1H), 3.61 (td, *J* = 13.0, 2.9 Hz, 1H), 3.33–3.24 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 189.8 (d, *J* = 15.4 Hz), 140.4, 134.0, 130.2, 129.9 (d, *J* = 2.1 Hz), 127.1, 125.5, 89.3 (d, *J* = 195.0 Hz), 30.8 (d, *J* = 23.3 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ –184.0; HRMS (ESI) *m/z* 205.0099 (*M*+Na)⁺, calc. for C₉H₇OFSNa⁺ 205.0099.

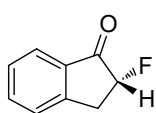
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 14.4 min (minor) and 12.9 min (major).



Entry	Retention Time	Area	Height	%Area
1	12.886	252.6142	796.43	49.94
2	14.321	253.1922	720.53	50.06

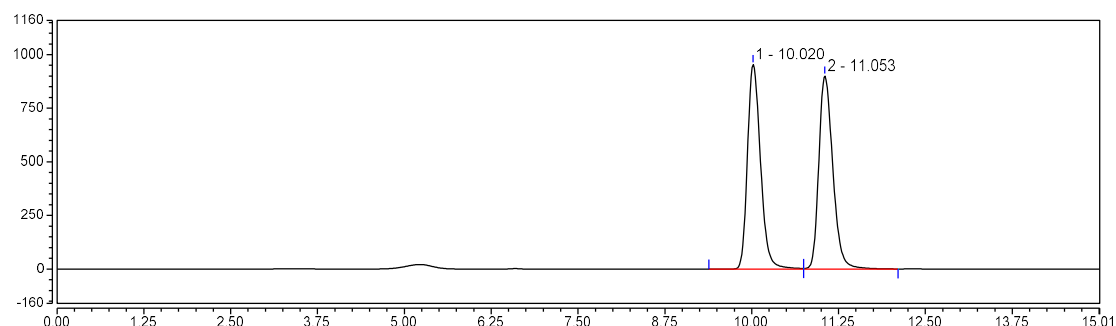


Entry	Retention Time	Area	Height	%Area
1	12.931	232.5183	733.59	96.61
2	14.390	8.1545	24.30	3.39

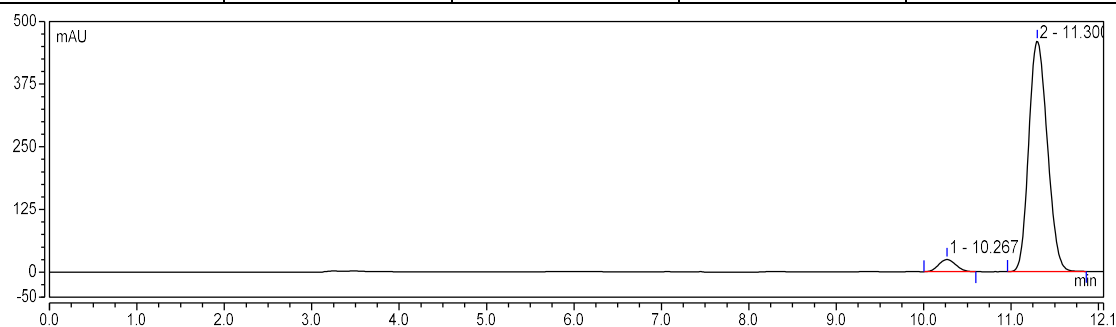


2r: white solid; Mp 54.6–56.0 °C; 11.3 mg, 75% yield; 91% ee; $[\alpha]_D^{22} +7.0$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.78 (d, $J = 7.7$ Hz, 1H), 7.67 (t, $J = 7.5$ Hz, 1H), 7.44 (dd, $J = 14.1, 7.3$ Hz, 2H), 5.27 (ddd, $J = 51.0, 7.8, 4.4$ Hz, 1H), 3.68–3.57 (m, 1H), 3.39–3.15 (m, 1H); ^{13}C NMR (75 MHz, CDCl_3) δ 200.0 (d, $J = 14.7$ Hz), 149.6 (d, $J = 5.7$ Hz), 133.8 (d, $J = 1.1$ Hz), 133.8, 128.4, 126.8 (d, $J = 1.6$ Hz), 124.7 (d, $J = 1.3$ Hz), 90.5 (d, $J = 189.2$ Hz), 33.4 (d, $J = 21.3$ Hz); ^{19}F NMR (376 MHz, CDCl_3) δ -194.0; HRMS (ESI) m/z 173.0376 ($\text{M}+\text{Na}^+$), calc. for $\text{C}_9\text{H}_7\text{OFNa}^+$ 173.0379.

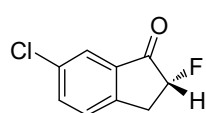
The ee was determined by HPLC analysis: CHIRALPAK ID (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 10.3 min (minor) and 11.3 min (major).



Entry	Retention Time	Area	Height	%Area
1	10.020	208.8269	953.60	49.71
2	11.053	211.2550	900.59	50.29

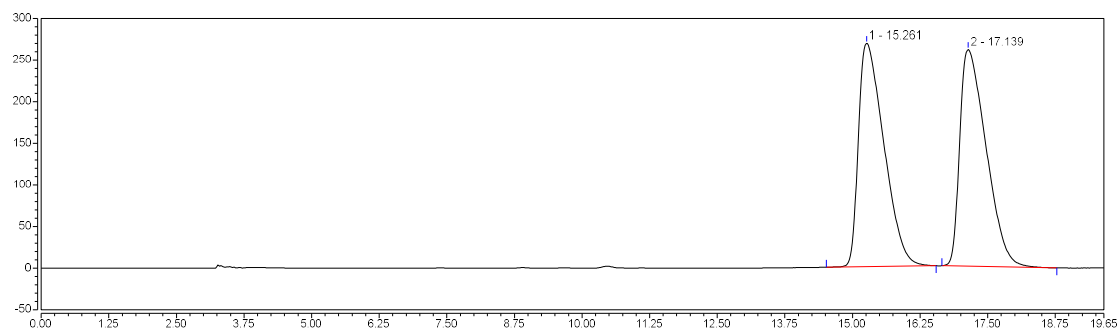


Entry	Retention Time	Area	Height	%Area
1	10.267	5.5552	24.36	4.68
2	11.300	113.1396	459.35	95.32

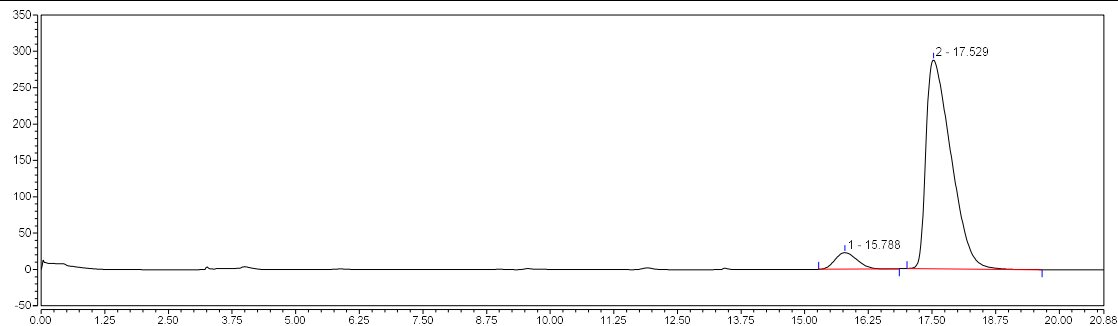


2s: white solid; Mp 89.1–91.3 °C; 13.5 mg, 73% yield; 87% ee; $[\alpha]_D^{22}$ –15.6 (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.72 (d, *J* = 8.2 Hz, 1H), 7.46–7.39 (m, 2H), 5.26 (ddd, *J* = 50.9, 7.7, 4.3 Hz, 1H), 3.66–3.55 (m, 1H), 3.28–3.13 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 198.3 (d, *J* = 14.9 Hz), 151.1 (d, *J* = 5.5 Hz), 142.9, 132.3 (d, *J* = 1.2 Hz), 129.3, 127.0 (d, *J* = 1.6 Hz), 125.9 (d, *J* = 1.3 Hz), 90.1 (d, *J* = 189.7 Hz), 33.2 (d, *J* = 22.0 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ –193.3; HRMS (ESI) *m/z* 206.9991 (M+Na)⁺, calc. for C₉H₆OFNaCl⁺ 206.9989.

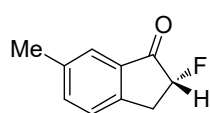
The ee was determined by HPLC analysis: CHIRALPAK ID (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 15.8 min (minor) and 17.5 min (major).



Entry	Retention Time	Area	Height	%Area
1	15.261	146.4709	268.32	49.95
2	17.139	146.7617	260.18	50.05

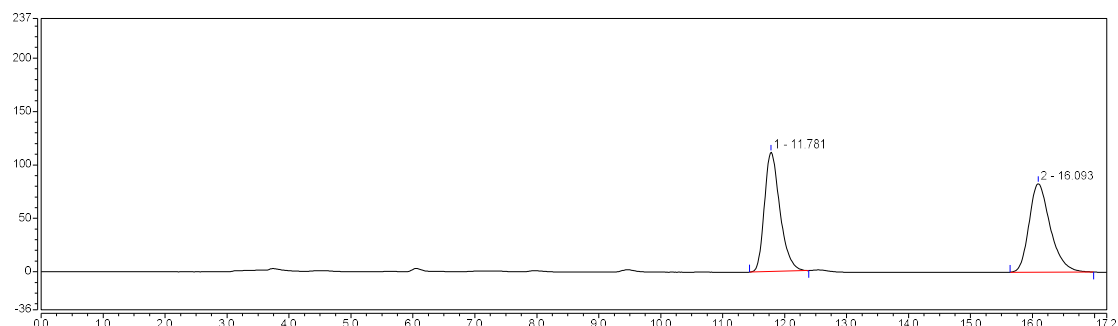


Entry	Retention Time	Area	Height	%Area
1	15.788	11.2989	22.61	6.48
2	17.529	163.0464	286.77	93.52

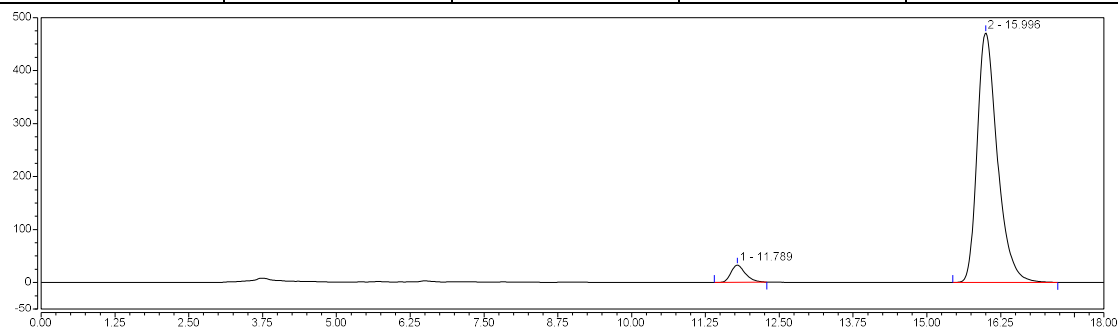


2t: white solid; Mp 67.9–69.4 °C; 13.6 mg, 83% yield; 91% ee; $[\alpha]_D^{22}$ –14.7 (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.58 (s, 1H), 7.48 (d, *J* = 7.9 Hz, 1H), 7.34 (d, *J* = 7.8 Hz, 1H), 5.25 (ddd, *J* = 51.0, 7.7, 4.3 Hz, 1H), 3.62–3.52 (m, 1H), 3.23–3.09 (m, 1H), 2.40 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 200.0 (d, *J* = 14.8 Hz), 147.0 (d, *J* = 5.8 Hz), 138.5, 137.6, 134.0 (d, *J* = 1.3 Hz), 126.4 (d, *J* = 1.6 Hz), 124.6 (d, *J* = 1.2 Hz), 90.8 (d, *J* = 189.1 Hz), 33.1 (d, *J* = 21.3 Hz), 21.1; ¹⁹F NMR (376 MHz, CDCl₃) δ –193.6; HRMS (ESI) *m/z* 187.0536 (M+Na)⁺, calc. for C₁₀H₉OFNa⁺ 187.0535.

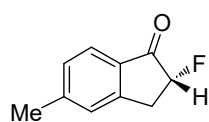
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 11.8 min (minor) and 16.0 min (major).



Entry	Retention Time	Area	Height	%Area
1	11.781	32.3695	111.76	49.60
2	16.093	32.8877	82.83	50.40

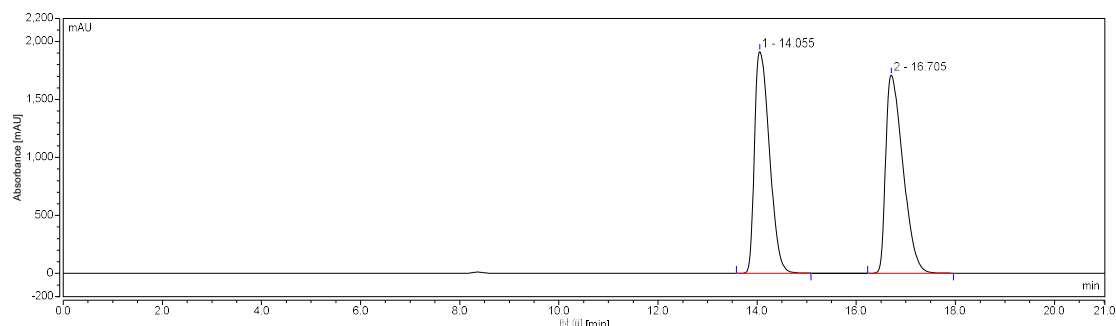


Entry	Retention Time	Area	Height	%Area
1	11.789	9.1073	32.51	4.65
2	15.996	186.7234	470.98	95.35

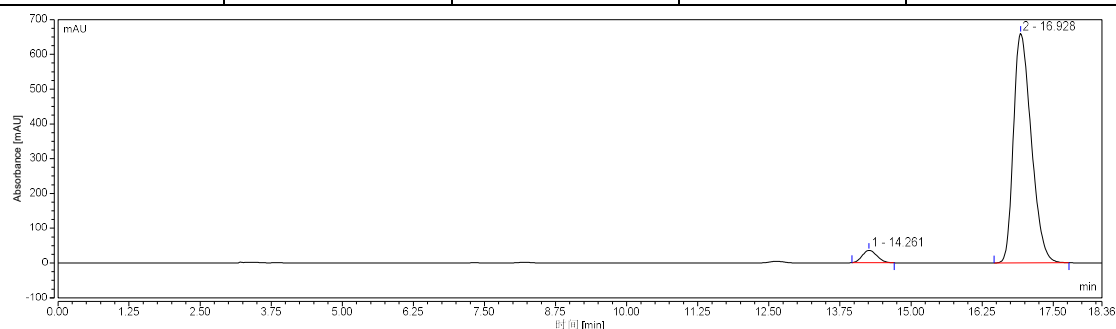


2u: white solid; Mp 81.1–82.5 °C; 13.6 mg, 83% yield; 92% ee; $[\alpha]_D^{22}$ -8.7 (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.69 (d, $J = 7.8$ Hz, 1H), 7.25 (t, $J = 5.5$ Hz, 2H), 5.25 (ddd, $J = 51.1, 7.7, 4.3$ Hz, 1H), 3.63–3.52 (m, 1H), 3.26–3.10 (m, 1H), 2.46 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 199.4 (d, $J = 14.8$ Hz), 150.1 (d, $J = 5.6$ Hz), 147.9, 131.6 (d, $J = 1.4$ Hz), 129.7, 127.1 (d, $J = 1.5$ Hz), 124.6 (d, $J = 1.4$ Hz), 90.6 (d, $J = 188.8$ Hz), 33.3 (d, $J = 21.4$ Hz), 22.3; ^{19}F NMR (376 MHz, CDCl_3) δ -193.4 ; HRMS (ESI) m/z 187.0540 ($\text{M}+\text{Na}^+$), calc. for $\text{C}_{10}\text{H}_9\text{OFNa}^+$ 187.0535.

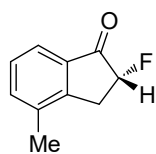
The ee was determined by HPLC analysis: CHIRALPAK ID (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 14.3 min (minor) and 16.9 min (major).



Entry	Retention Time	Area	Height	%Area
1	14.055	670.0501	1914.53	49.21
2	16.705	691.5219	1711.24	50.79

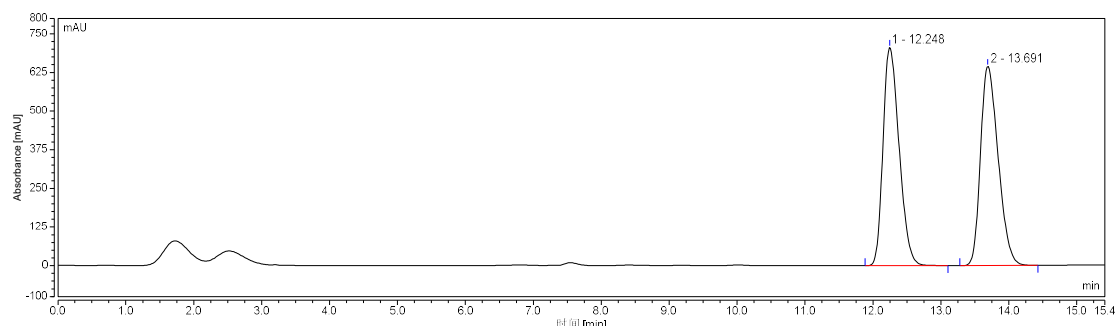


Entry	Retention Time	Area	Height	%Area
1	14.261	10.6185	35.28	4.25
2	16.928	239.3537	660.20	95.75

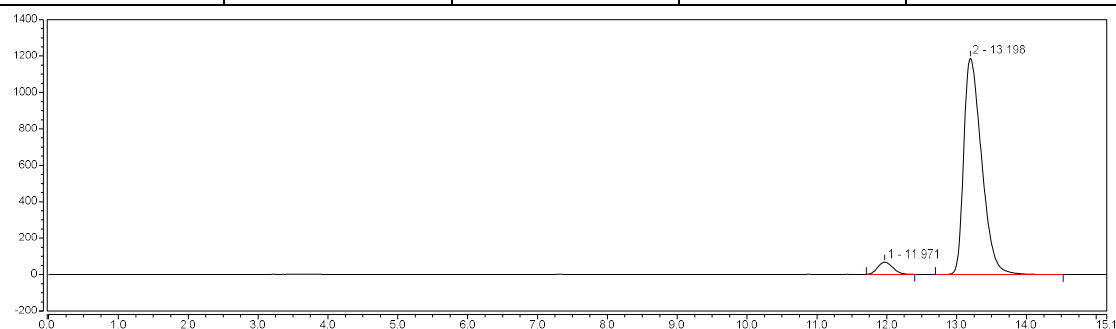


2v: white solid; Mp 64.8–66.3 °C; 14.8 mg, 90% yield; 91% ee; $[\alpha]_D^{22}$ –9.0 (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.62 (d, *J* = 7.6 Hz, 1H), 7.47 (d, *J* = 7.4 Hz, 1H), 7.34 (t, *J* = 7.5 Hz, 1H), 5.26 (ddd, *J* = 50.9, 7.7, 4.1 Hz, 1H), 3.59–3.49 (m, 1H), 3.14–2.99 (m, 1H), 2.35 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 200.3 (d, *J* = 14.6 Hz), 148.7 (d, *J* = 5.1 Hz), 136.8, 136.1 (d, *J* = 1.4 Hz), 133.7 (d, *J* = 1.2 Hz), 128.5, 122.0 (d, *J* = 1.2 Hz), 90.5 (d, *J* = 188.7 Hz), 32.1 (d, *J* = 21.3 Hz), 17.7; ¹⁹F NMR (376 MHz, CDCl₃) δ –193.9; HRMS (ESI) *m/z* 187.0537 (M+Na)⁺, calc. for C₁₀H₉OFNa⁺ 187.0535.

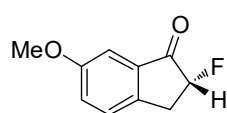
The ee was determined by HPLC analysis: CHIRALPAK ID (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 12.0 min (minor) and 13.2 min (major).



Entry	Retention Time	Area	Height	%Area
1	12.248	194.1255	706.12	50.00
2	13.691	194.1314	644.35	50.00

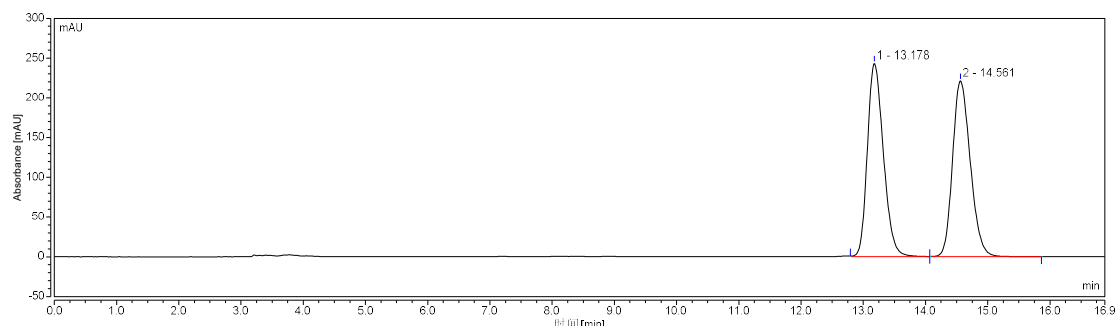


Entry	Retention Time	Area	Height	%Area
1	11.971	16.8186	67.60	4.56
2	13.198	352.1670	1188.46	95.44

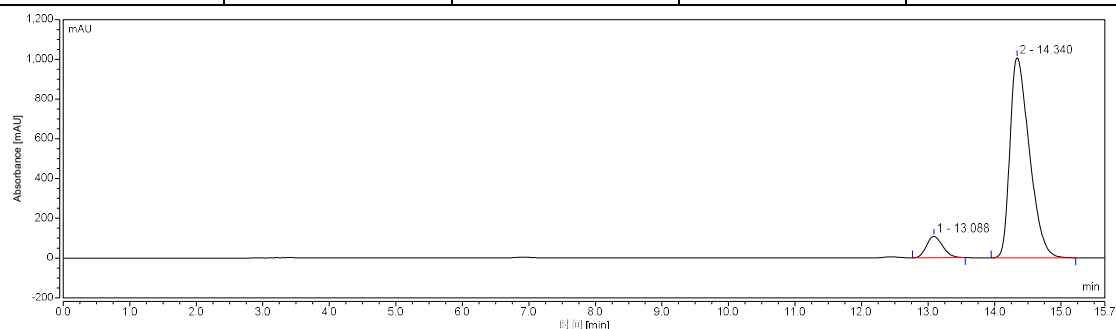


2w: white solid; Mp 67.8–68.8 °C; 14.9 mg, 83% yield; 84% ee; $[\alpha]_D^{22}$ –18.2 (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.42 (d, *J* = 8.4 Hz, 1H), 7.34–7.30 (m, 1H), 7.26 (d, *J* = 2.2 Hz, 1H), 5.33 (ddd, *J* = 51.0, 7.6, 4.2 Hz, 1H), 3.90 (s, 3H), 3.67–3.57 (m, 1H), 3.32–3.12 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 199.9 (d, *J* = 15.1 Hz), 159.9, 142.4 (d, *J* = 5.9 Hz), 134.9 (d, *J* = 1.1 Hz), 127.5 (d, *J* = 1.6 Hz), 125.7, 105.7 (d, *J* = 1.1 Hz), 91.0 (d, *J* = 189.3 Hz), 55.6, 32.7 (d, *J* = 21.2 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ –193.2; HRMS (ESI) *m/z* 203.0492 (M+Na)⁺, calc. for C₁₀H₉O₂FN⁺ 203.0484.

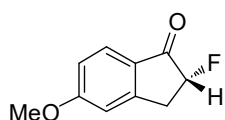
The ee was determined by HPLC analysis: CHIRALPAK ID (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 13.1 min (minor) and 14.3 min (major).



Entry	Retention Time	Area	Height	%Area
1	13.178	72.7721	243.42	49.96
2	14.561	72.8893	221.28	50.04

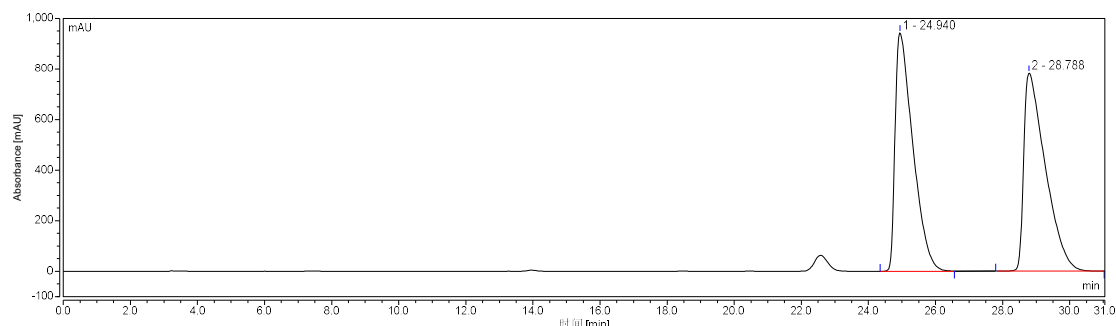


Entry	Retention Time	Area	Height	%Area
1	13.088	30.2802	107.65	8.16
2	14.340	340.8339	1006.39	91.84

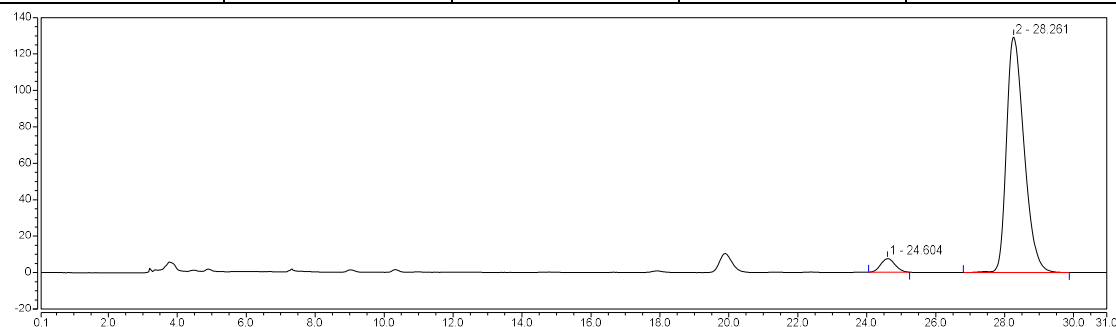


2x: white solid; Mp 122.9–124.1 °C; 9.9 mg, 55% yield; 91% ee; $[\alpha]_D^{22} +11.2$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.73 (d, $J = 8.6$ Hz, 1H), 6.87 (s, 1H), 5.22 (ddd, $J = 51.3, 7.6, 4.1$ Hz, 1H), 3.90 (s, 3H), 3.68–3.43 (m, 1H), 3.15 (ddd, $J = 25.3, 14.6, 6.0$ Hz, 1H); ^{13}C NMR (75 MHz, CDCl_3) δ 197.8 (d, $J = 15.0$ Hz), 166.5, 152.8 (d, $J = 5.6$ Hz), 127.1, 126.6 (d, $J = 1.2$ Hz), 116.4, 109.9 (d, $J = 1.6$ Hz), 90.5 (d, $J = 187.9$ Hz), 55.8, 33.6 (d, $J = 21.7$ Hz); ^{19}F NMR (376 MHz, CDCl_3) δ -192.2; HRMS (ESI) m/z 203.0487 ($\text{M}+\text{Na}^+$), calc. for $\text{C}_{10}\text{H}_9\text{O}_2\text{FNa}^+$ 203.0484.

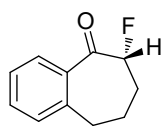
The ee was determined by HPLC analysis: CHIRALPAK ID (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 24.6 min (minor) and 28.3 min (major).



Entry	Retention Time	Area	Height	%Area
1	24.940	577.0831	943.36	49.72
2	28.788	583.5189	783.87	50.28

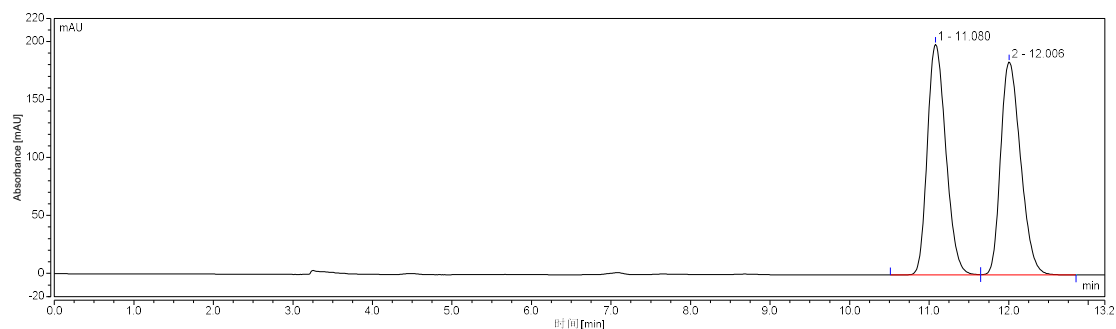


Entry	Retention Time	Area	Height	%Area
1	24.604	3.5844	7.47	4.47
2	28.261	76.5542	129.42	95.53

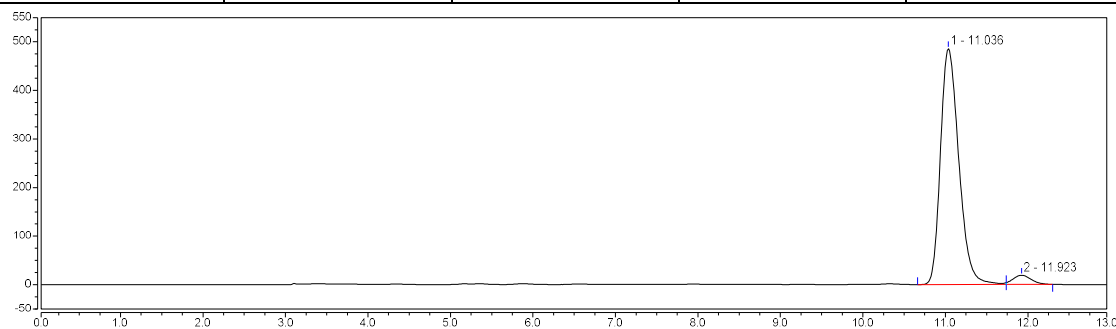


2y: yellow oil; 15.2 mg, 85% yield; 93% ee; $[\alpha]_D^{22} +95.9$ (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.76 (d, *J* = 6.0 Hz, 1H), 7.43 (t, *J* = 15.0, 9.0, 6.0 Hz, 1H), 7.32 (t, *J* = 15.0, 9.0, 6.0 Hz, 1H), 7.22 (d, *J* = 9.0 Hz, 1H), 5.36–5.15 (m, 1H), 3.08–2.90 (m, 2H), 2.40–2.27 (m, 1H), 2.19–1.81 (m, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 200.2 (d, *J* = 19.5 Hz), 141.8, 135.8, 132.3, 130.1, 129.2 (d, *J* = 7.5 Hz), 126.7, 94.7 (d, *J* = 183.0 Hz), 34.2, 30.5 (d, *J* = 21.0 Hz), 22.9 (d, *J* = 8.2 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ -182.6; HRMS (ESI) *m/z* 201.1920 (M+Na)⁺, calc. for C₁₀H₉O₂FNa⁺ 201.1921.

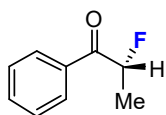
The ee was determined by HPLC analysis: CHIRALPAK IE (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 11.9 min (minor) and 11.0 min (major).



Entry	Retention Time	Area	Height	%Area
1	11.080	53.7328	198.94	50.13
2	12.006	53.4511	183.79	49.87



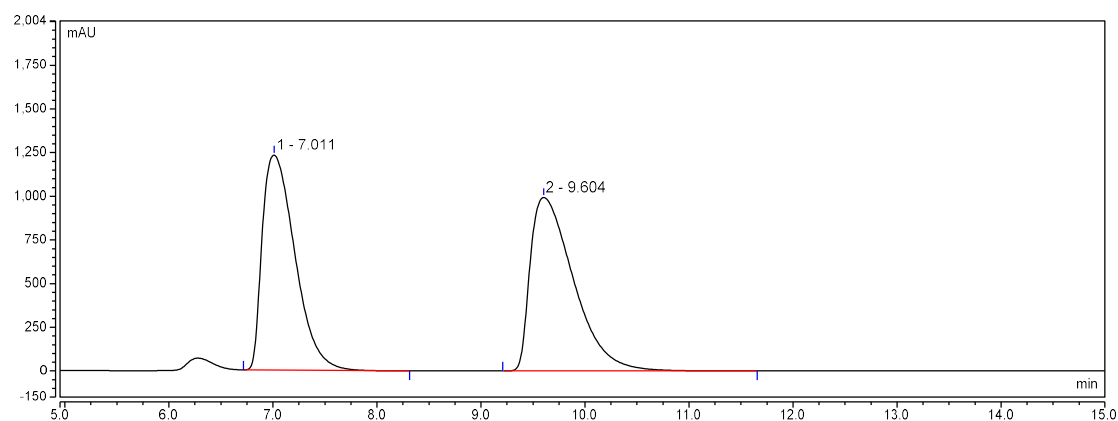
Entry	Retention Time	Area	Height	%Area
1	11.036	125.1085	485.78	96.28
2	11.923	4.8349	18.91	3.72



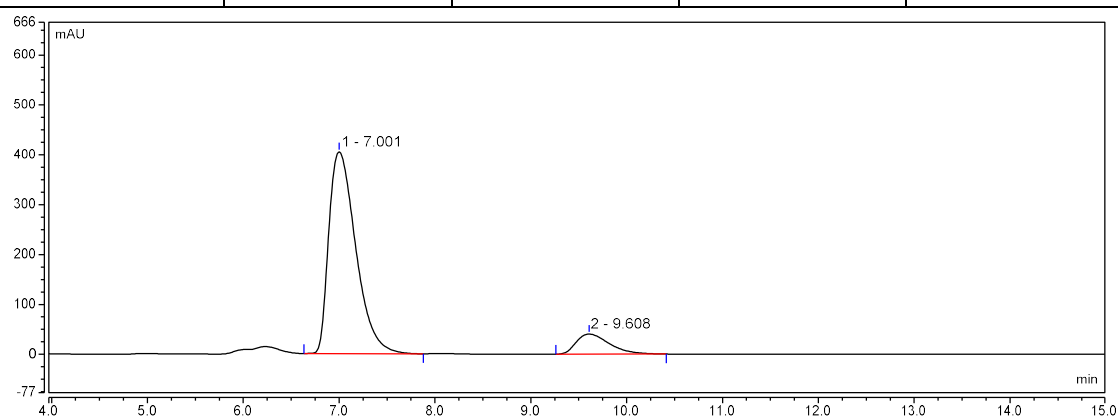
2a: colorless oil; 14.0 mg, 92% yield; 82% ee; $[\alpha]_D^{22}$ -55.1 (c 1.0, CHCl_3);

^1H NMR (300 MHz, CDCl_3) δ 7.95 (d, $J = 7.8$ Hz, 2H), 7.57 (t, $J = 7.4$ Hz, 1H), 7.45 (t, $J = 7.6$ Hz, 2H), 5.70 (dq, $J = 48.6, 6.8$ Hz, 1H), 1.63 (dd, $J = 24.0, 6.8$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 196.8 (d, $J = 19.4$ Hz), 133.9 (d, $J = 0.8$ Hz), 133.7, 128.8 (d, $J = 3.6$ Hz), 128.6, 90.0 (d, $J = 179.9$ Hz), 18.2 (d, $J = 22.8$ Hz); ^{19}F NMR (376 MHz, CDCl_3) δ -191.7 ; HRMS (ESI) m/z 175.0533 ($\text{M}+\text{Na}^+$), calc. for $\text{C}_9\text{H}_9\text{OFNa}^+$ 175.0529.

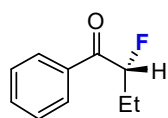
The ee was determined by HPLC analysis: CHIRALPAK AS-H (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 98/2; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 7.0 min (major) and 9.6 min (minor).



Entry	Retention Time	Area	Height	%Area
1	7.011	479.1758	1230.89	51.68
2	9.604	448.0692	992.85	48.32

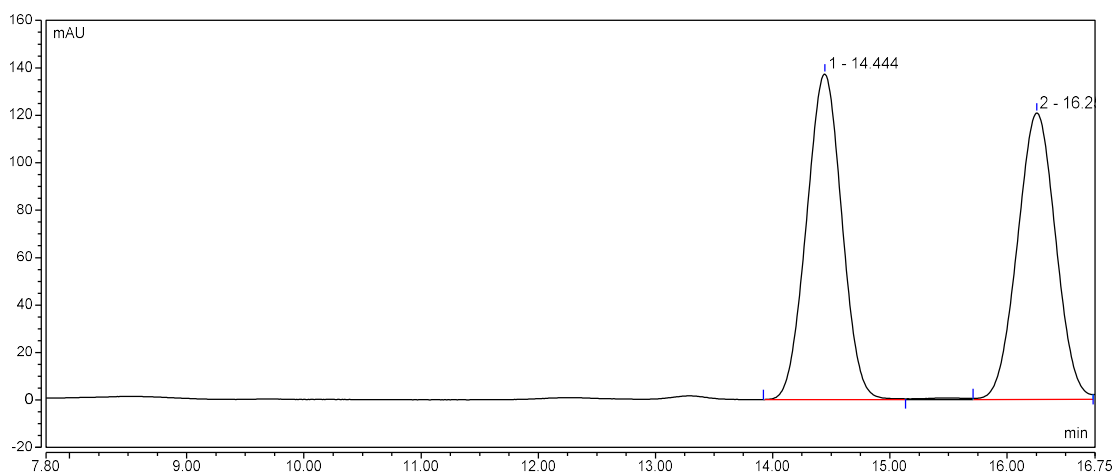


Entry	Retention Time	Area	Height	%Area
1	7.001	132.1674	404.57	90.81
2	9.608	13.3826	37.24	9.19

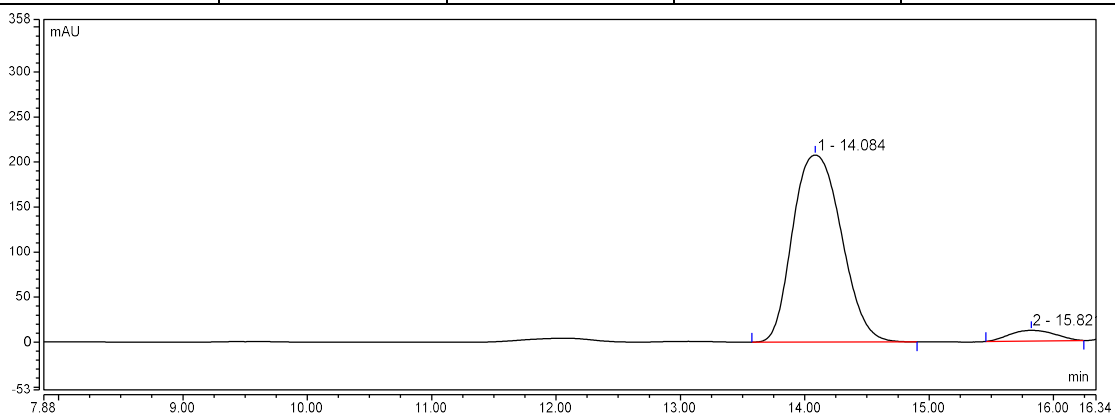


2b: colorless oil; 13.4 mg, 81% yield; 90% ee; $[\alpha]_D^{22} -37.1$ (*c* 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.94 (d, $J = 7.6$ Hz, 2H), 7.58 (t, $J = 7.4$ Hz, 1H), 7.46 (t, $J = 7.6$ Hz, 2H), 5.50 (ddd, $J = 49.3, 7.5, 4.7$ Hz, 1H), 2.17 – 1.85 (m, 2H), 1.06 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 196.7 (d, $J = 19.4$ Hz), 134.3 (d, $J = 0.7$ Hz), 133.62, 128.7 (d, $J = 3.8$ Hz), 128.6, 94.6 (d, $J = 183.4$ Hz), 26.0 (d, $J = 21.7$ Hz), 8.9 (d, $J = 4.5$ Hz); ^{19}F NMR (376 MHz, CDCl_3) δ -191.0; HRMS (ESI) m/z 189.0688 ($\text{M}+\text{Na}^+$), calc. for $\text{C}_{10}\text{H}_{11}\text{OFNa}^+$ 189.0686.

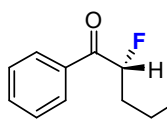
The ee was determined by HPLC analysis: CHIRALPAK IE-CHIRALPAK IE (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 98/2; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 14.1 min (major) and 15.8 min (minor).



Entry	Retention Time	Area	Height	%Area
1	14.444	47.8291	137.24	51.16
2	16.254	45.6551	120.76	48.84

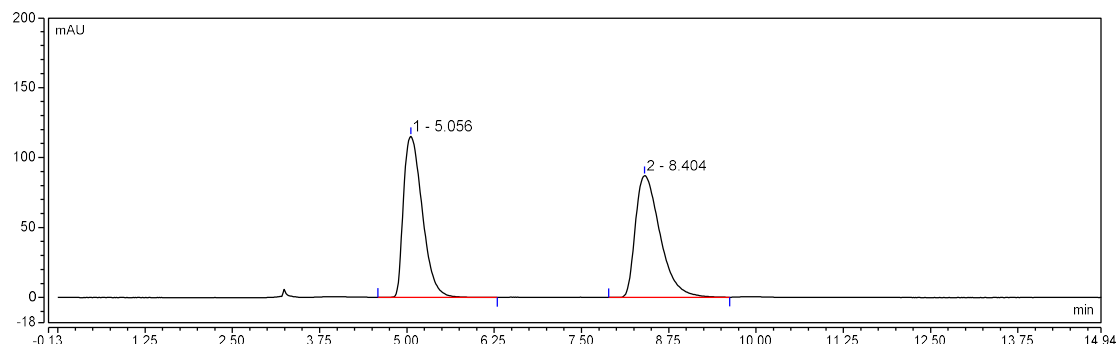


Entry	Retention Time	Area	Height	%Area
1	14.084	94.3707	207.68	95.00
2	15.821	4.9624	11.74	5.00

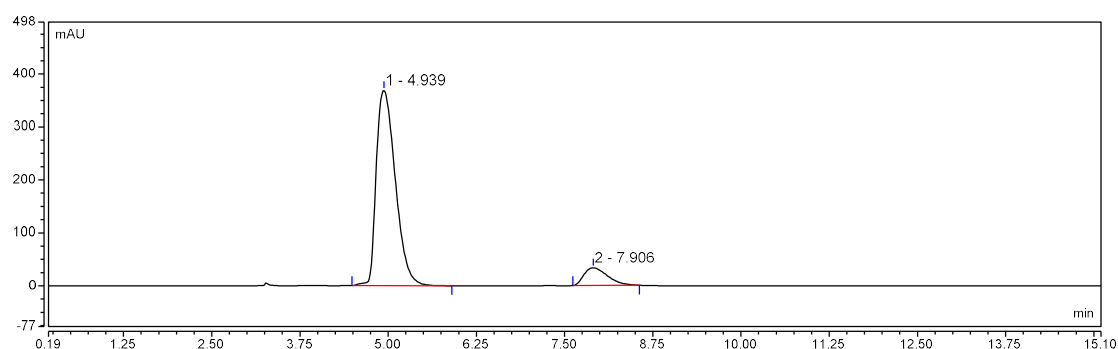


2zc: colorless oil; 18.3 mg, 88% yield; 83% ee; $[\alpha]_D^{22} -61.5$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.96 (d, $J = 7.6$ Hz, 2H), 7.60 (t, $J = 7.4$ Hz, 1H), 7.48 (t, $J = 7.6$ Hz, 2H), 5.56 (dt, $J = 12.4, 5.9$ Hz, 1H), 2.06 – 1.83 (m, 2H), 1.53 (d, $J = 7.1$ Hz, 2H), 1.38 – 1.27 (m, 4H), 0.89 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 197.0 (d, $J = 19.6$ Hz), 134.4 (d, $J = 0.8$ Hz), 133.7, 128.8 (d, $J = 3.8$ Hz), 128.7, 93.9 (d, $J = 183.1$ Hz), 32.7 (d, $J = 21.3$ Hz), 31.3, 24.4 (d, $J = 3.1$ Hz), 22.4, 13.9; ^{19}F NMR (376 MHz, CDCl_3) δ -189.5; HRMS (ESI) m/z 231.1152 ($\text{M}+\text{Na}^+$), calc. for $\text{C}_{13}\text{H}_{17}\text{OFNa}^+$ 231.1155.

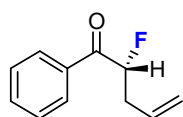
The ee was determined by HPLC analysis: CHIRALPAK AS-H (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 98/2; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 4.9 min (major) and 7.9 min (minor).



Entry	Retention Time	Area	Height	%Area
1	5.506	35.8924	114.95	50.26
2	8.404	35.5269	86.99	49.74



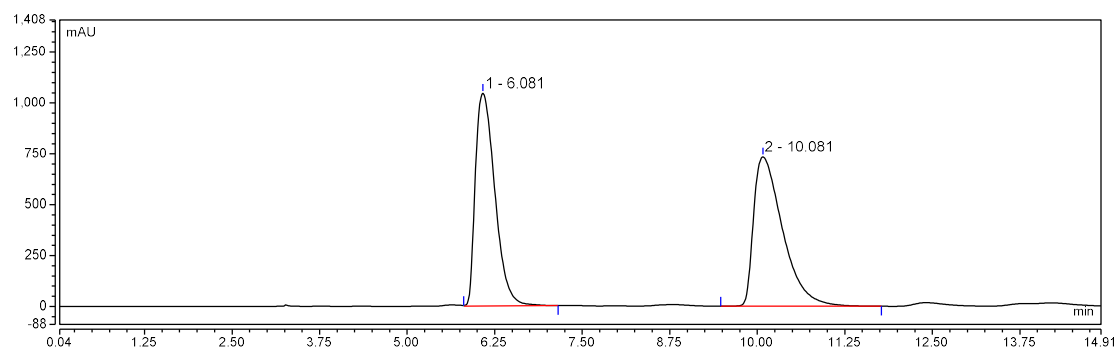
Entry	Retention Time	Area	Height	%Area
1	4.939	115.3989	369.10	91.69
2	7.906	10.4575	30.33	8.31



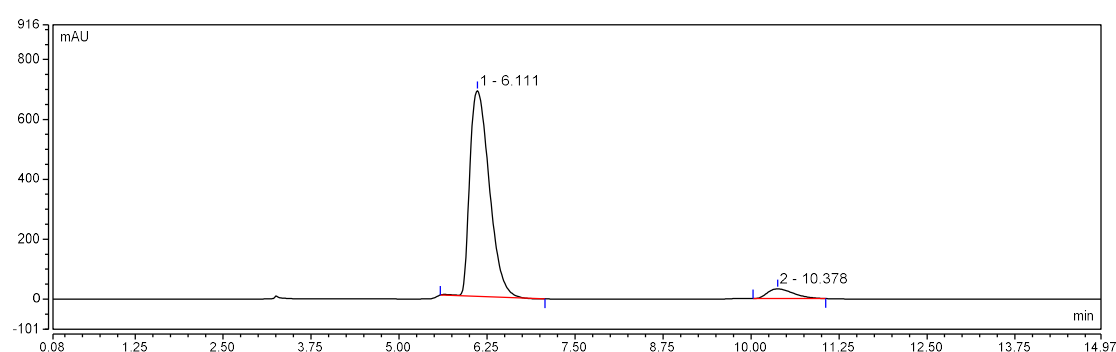
2zd: colorless oil; 15.8 mg, 89% yield; 90% ee; $[\alpha]_{\text{D}}^{22} -46.2$ (*c* 1.0, CHCl₃);

¹H NMR (300 MHz, CDCl₃) δ 7.96 (d, *J* = 7.7 Hz, 2H), 7.61 (t, *J* = 7.4 Hz, 1H), 7.48 (t, *J* = 7.6 Hz, 2H), 5.88 (ddt, *J* = 17.3, 10.4, 6.9 Hz, 1H), 5.61 (ddd, *J* = 48.9, 7.3, 4.8 Hz, 1H), 5.20 (d, *J* = 6.4 Hz, 1H), 5.15 (s, 1H), 2.88 – 2.58 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 196.1 (d, *J* = 19.5 Hz), 134.3, 133.8, 131.4 (d, *J* = 3.8 Hz), 128.9 (d, *J* = 3.9 Hz), 128.7, 92.8 (d, *J* = 185.1 Hz), 36.8 (d, *J* = 21.5 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ -181.5; HRMS (ESI) *m/z* 201.0683 (M+Na)⁺, calc. for C₁₁H₁₁OFNa⁺ 201.0686.

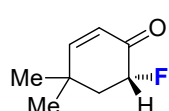
The ee was determined by HPLC analysis: CHIRALPAK AS-H (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 98/2; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 6.1 min (major) and 10.4 min (minor).



Entry	Retention Time	Area	Height	%Area
1	6.081	338.0516	1045.45	48.93
2	10.081	352.8729	734.21	51.07

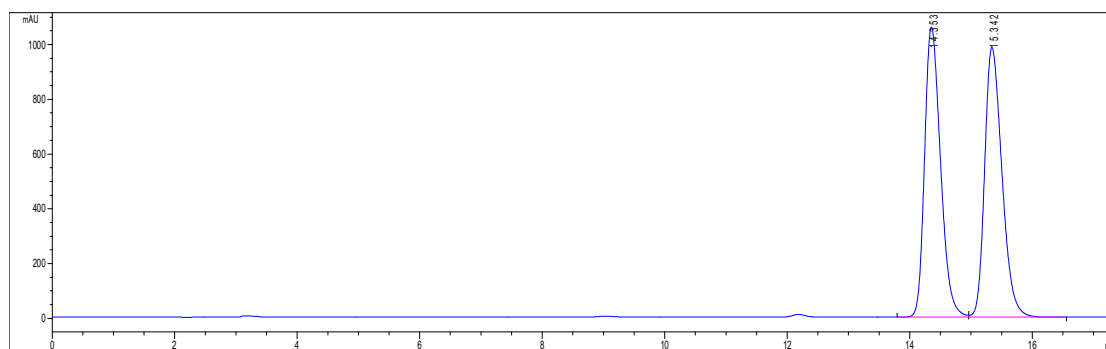


Entry	Retention Time	Area	Height	%Area
1	6.111	232.0810	695.35	95.15
2	10.387	11.8350	30.14	4.85

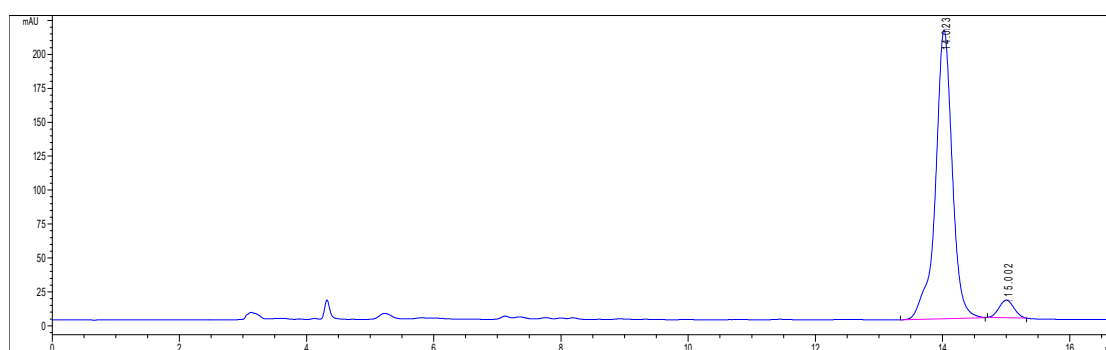


2ze: white solid; Mp 46.3–47.9 °C; 10.6 mg, 75% yield; 90% ee; $[\alpha]_D^{22}$ –30.2 (c 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 6.65 (d, *J* = 10.0 Hz, 1H), 5.84 (dd, *J* = 10.0, 4.4 Hz, 1H), 5.06 (ddd, *J* = 47.9, 13.2, 5.7 Hz, 1H), 2.24 (dt, *J* = 11.9, 5.9 Hz, 1H), 2.04 (dd, *J* = 22.4, 12.7 Hz, 1H), 1.22 (d, *J* = 13.3 Hz, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 194.5 (d, *J* = 14.7 Hz), 159.6 (d, *J* = 1.6 Hz), 124.4, 88.0 (d, *J* = 186.4 Hz), 42.2 (d, *J* = 16.8 Hz), 35.5 (d, *J* = 11.0 Hz), 30.5, 26.1; ¹⁹F NMR (376 MHz, CDCl₃) δ –195.6 (six peaks), –195.7 (three peaks), –195.8 (three peaks); HRMS (ESI) *m/z* 165.0684 (M+Na)⁺, calc. for C₈H₁₁OFNa⁺ 165.0686.

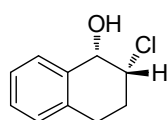
The ee was determined by HPLC analysis: CHIRALPAK IG (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 230 nm; retention time: 14.0 min (major) and 15.0 min (minor).



Entry	Retention Time	Area	Height	%Area
1	14.353	19164.6	1059.8	49.756
2	15.342	19352.4	985.4	50.244



Entry	Retention Time	Area	Height	%Area
1	14.023	3890.7	213	94.860
2	15.002	210.8	13	5.140



6a: white solid; Mp 80.4–81.8 °C; 14.4 mg, 79% yield; 92% ee; > 20:1 dr;

$[\alpha]_D^{22}$ -6.0 (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.52–7.49 (m, 1H),

7.27–7.25 (m, 2H), 7.15 (d, *J* = 3.3 Hz, 1H), 4.86 (s, 1H), 4.56 (dd, *J* = 5.5,

2.9 Hz, 1H), 3.13 (dt, *J* = 16.7, 6.3 Hz, 1H), 2.90–2.80 (m, 1H), 2.44 (td, *J* = 14.2, 6.8 Hz,

2H), 2.23–2.19 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 135.8, 134.8, 128.8, 128.5, 128.1,

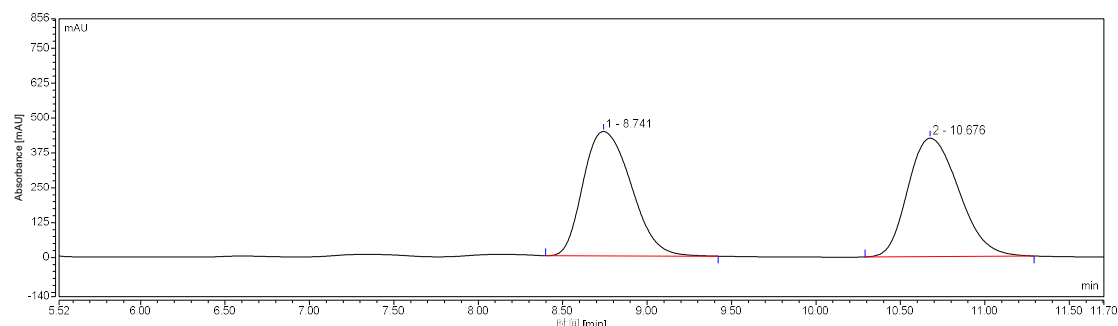
126.5, 70.1, 63.2, 27.5, 26.6; HRMS (ESI) *m/z* 183.0575 (M+H)⁺, calc. for C₁₀H₁₂OCl⁺

183.0577.

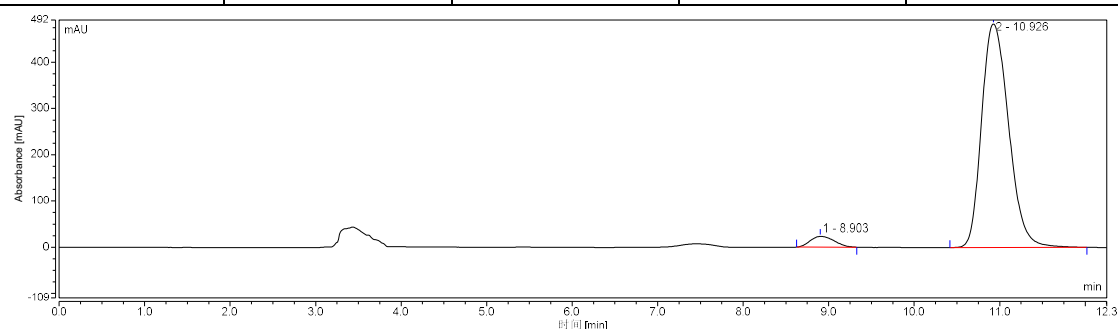
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm);

Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 210 nm; retention time: 8.9 min

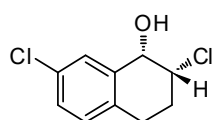
(minor) and 10.9 min (major).



Entry	Retention Time	Area	Height	%Area
1	8.741	146.6750	445.57	49.82
2	10.676	147.7391	424.21	50.18



Entry	Retention Time	Area	Height	%Area
1	8.903	8.903	8.903	4.08
2	10.926	10.926	10.926	95.92



6b: white solid; Mp 84.9–86.5 °C; 13.0 mg, 60% yield; 90% ee; > 20:1 dr;

$[\alpha]_D^{22}$ -56.1 (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.51 (s, 1H),

7.20 (d, *J* = 8.2 Hz, 1H), 7.05 (d, *J* = 8.2 Hz, 1H), 4.80 (s, 1H), 4.56 (dd, *J*

= 4.4, 3.0 Hz, 1H), 3.13–3.02 (m, 1H), 2.78 (dt, *J* = 17.3, 5.9 Hz, 1H), 2.47–2.34 (m, 2H),

2.24–2.15 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 137.7, 133.2, 132.1, 129.9, 128.3, 128.2,

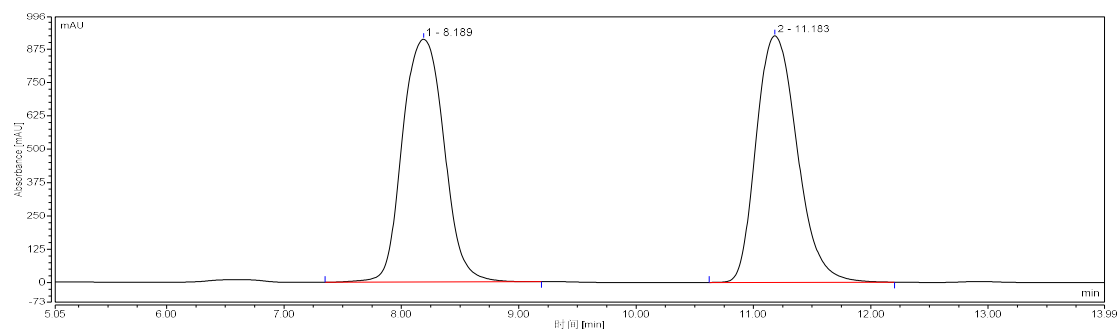
69.7, 62.7, 27.6, 25.4; HRMS (ESI) *m/z* 239.0003 (M+Na)⁺, calc. for C₁₀H₁₀ONaCl⁺

239.0006.

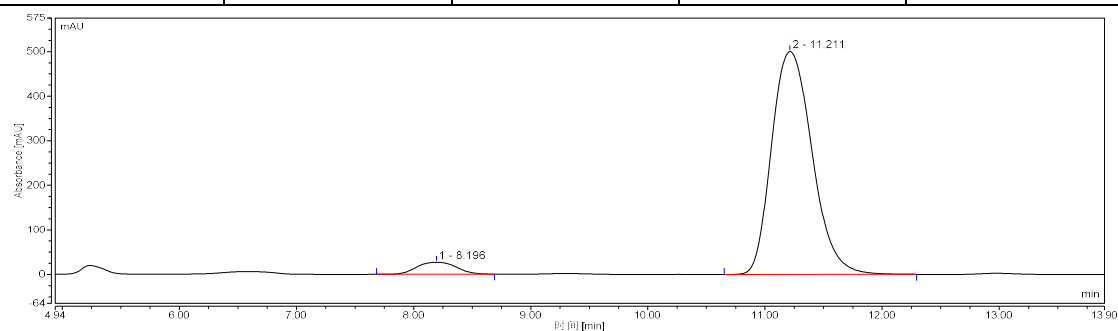
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm);

Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 210 nm; retention time: 8.2 min

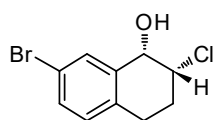
(minor) and 11.2 min (major).



Entry	Retention Time	Area	Height	%Area
1	8.189	372.8273	909.62	49.96
2	11.183	373.3544	923.61	50.04



Entry	Retention Time	Area	Height	%Area
1	8.196	10.4827	26.91	4.96
2	11.211	200.6826	500.11	95.04



6c: white solid; Mp 90.2–92.1 °C; 16.0 mg, 61% yield; 84% ee; > 20:1 dr;

$[\alpha]_D^{22}$ -46.6 (c 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.67 (d, *J* = 1.4

Hz, 1H), 7.35 (dd, *J* = 8.2, 1.9 Hz, 1H), 6.99 (d, *J* = 8.2 Hz, 1H), 4.81 (d,

J = 3.0 Hz, 1H), 4.65–4.46 (m, 1H), 3.11–3.01 (m, 1H), 2.76 (dt, *J* = 17.4, 6.0 Hz, 1H), 2.40

(td, *J* = 13.6, 5.9 Hz, 1H), 2.25–2.15 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 138.1, 133.8,

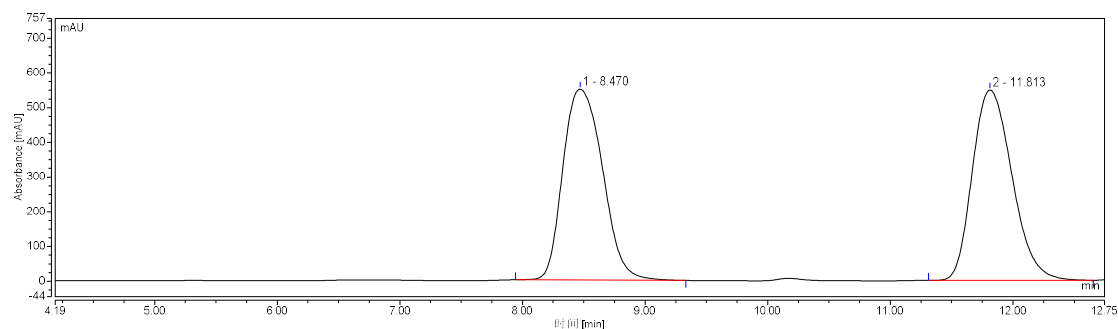
131.3, 131.1, 130.2, 120.1, 69.7, 62.7, 27.6, 25.6; HRMS (ESI) *m/z* 282.9504 (M+Na)⁺, calc.

for C₁₀H₁₀ONaClBr⁺ 282.9501.

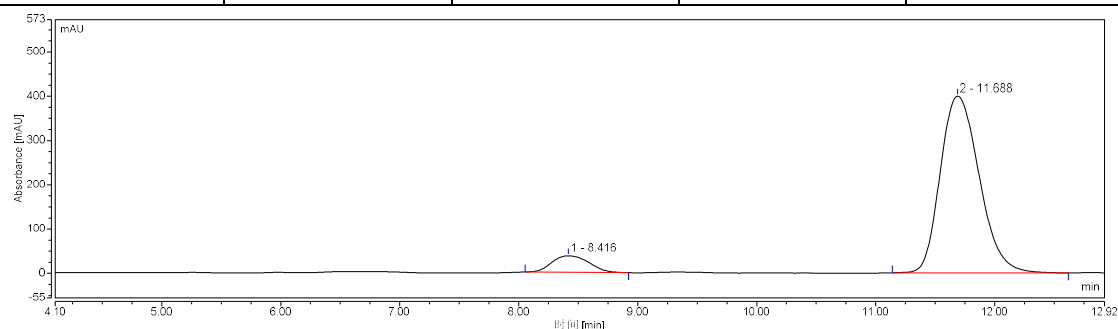
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm);

Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 210 nm; retention time: 8.4 min

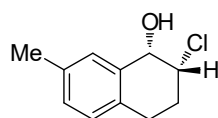
(minor) and 11.7 min (major).



Entry	Retention Time	Area	Height	%Area
1	8.470	205.9142	549.95	50.06
2	11.813	205.4494	548.22	49.94

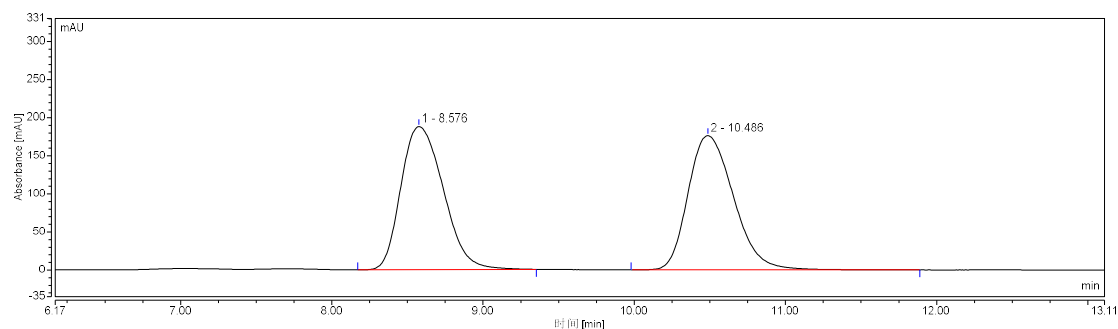


Entry	Retention Time	Area	Height	%Area
1	8.416	13.3986	37.15	8.13
2	11.688	151.3247	399.00	91.87

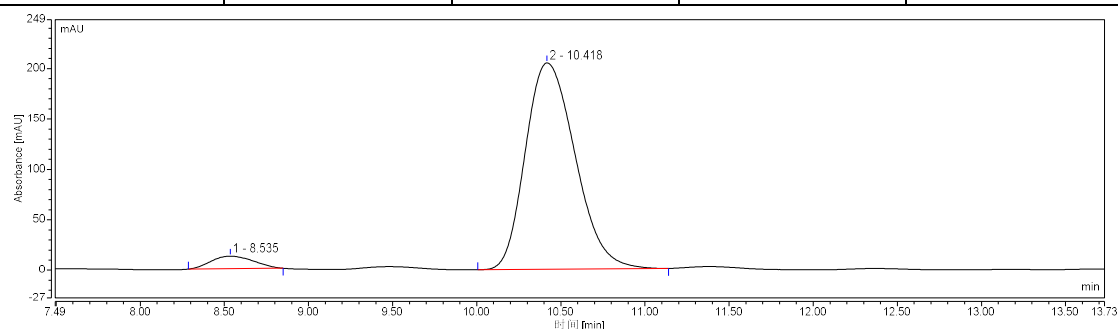


6d: white solid; Mp 103.9–105.1 °C; 13.1 mg, 67% yield; 90% ee; > 20:1 dr; $[\alpha]_D^{22}$ -9.5 (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.30 (s, 1H), 7.08–7.00 (m, 1H), 4.81 (d, J = 3.1 Hz, 1H), 4.53 (dt, J = 8.7, 3.0 Hz, 1H), 3.06 (dt, J = 16.8, 6.3 Hz, 1H), 2.85–2.75 (m, 1H), 2.47–2.35 (m, 1H), 2.33 (s, 2H), 2.18 (dtd, J = 9.3, 6.6, 2.7 Hz, 1H); ^{13}C NMR (75 MHz, CDCl_3) δ 136.1, 135.5, 131.8, 129.3, 129.1, 128.5, 70.1, 63.4, 27.7, 26.4, 21.0; HRMS (ESI) m/z 219.0554 ($\text{M}+\text{Na}^+$), calc. for $\text{C}_{11}\text{H}_{13}\text{OCINa}^+$ 219.0553.

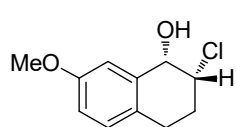
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 210 nm; retention time: 8.9 min (minor) and 10.9 min (major).



Entry	Retention Time	Area	Height	%Area
1	8.576	61.1470	187.91	49.87
2	10.486	61.4718	176.27	50.13

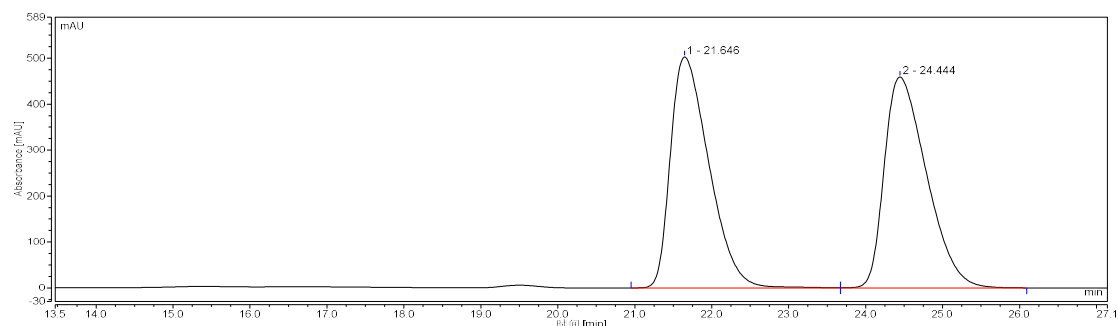


Entry	Retention Time	Area	Height	%Area
1	8.903	3.6232	12.39	4.91
2	10.926	70.2000	204.76	95.09

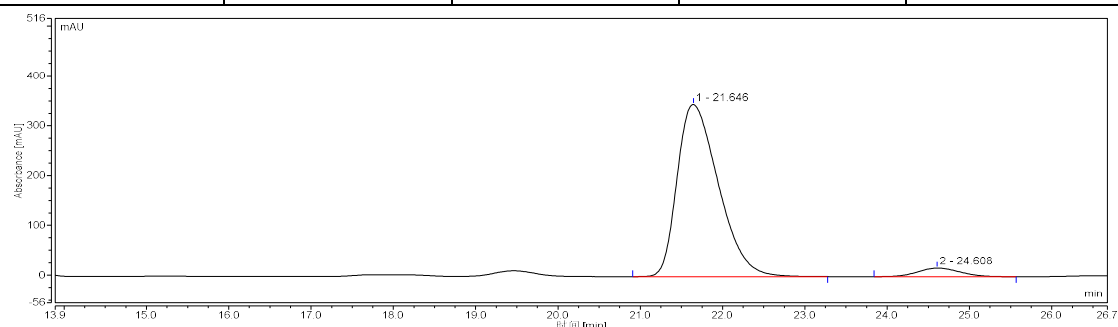


6e: white solid; Mp 94.9–96.5 °C; 16.0 mg, 75% yield; 90% ee; > 20:1 dr; $[\alpha]_{\text{D}}^{22} -5.0$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.03 (d, $J = 8.4$ Hz, 2H), 6.82 (dd, $J = 8.4, 2.4$ Hz, 1H), 4.81 (s, 1H), 4.56 (dd, $J = 4.8, 3.2$ Hz, 1H), 3.80 (s, 3H), 3.09–3.00 (m, 1H), 2.76 (dt, $J = 16.9, 6.2$ Hz, 1H), 2.39 (dt, $J = 13.9, 6.2$ Hz, 2H), 2.19 (ddd, $J = 13.7, 9.7, 4.0$ Hz, 1H); ^{13}C NMR (75 MHz, CDCl_3) δ 158.2, 136.9, 129.6, 126.7, 115.0, 112.6, 70.2, 63.3, 55.3, 27.8, 25.4; HRMS (ESI) m/z 235.0503 ($\text{M}+\text{Na}$) $^+$, calc. for $\text{C}_{11}\text{H}_{13}\text{O}_2\text{NaCl}^+$ 235.0502.

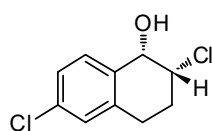
The ee was determined by HPLC analysis: CHIRALPAK IE (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 210 nm; retention time: 21.6 min (major) and 24.6 min (minor).



Entry	Retention Time	Area	Height	%Area
1	21.646	291.1048	502.78	49.98
2	24.444	291.2826	458.95	50.02



Entry	Retention Time	Area	Height	%Area
1	21.646	200.3435	345.93	95.12
2	24.608	10.2791	17.30	4.88

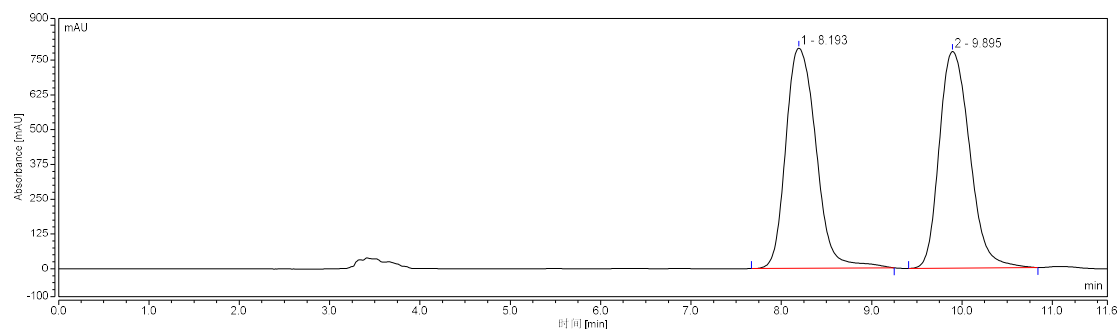


6f: white solid; Mp 133.9–136.8 °C; 14.1 mg, 65% yield; 90% ee; > 20:1

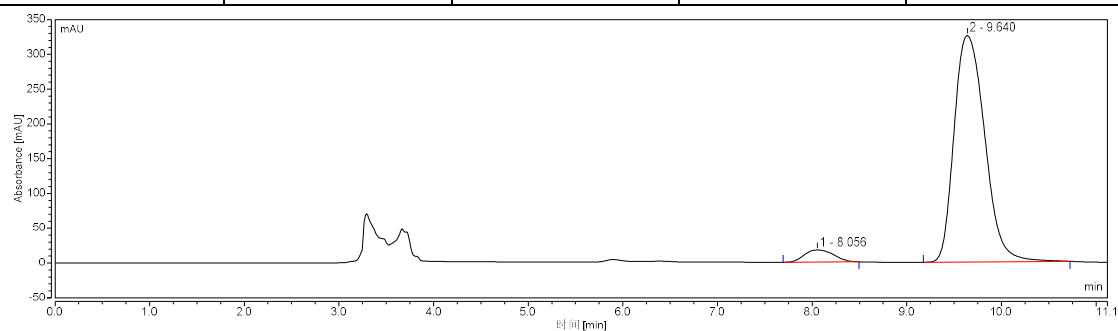
dr; $[\alpha]_D^{22} -7.3$ (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.44 (d, *J* = 8.3 Hz, 1H), 7.20 (d, *J* = 8.3 Hz, 1H), 7.12 (s, 1H), 4.81 (d, *J* = 3.2 Hz,

1H), 4.54 (dt, *J* = 7.9, 3.0 Hz, 1H), 3.09 (dt, *J* = 17.3, 6.8 Hz, 1H), 2.80 (dt, *J* = 17.5, 6.2 Hz, 1H), 2.40 (dt, *J* = 14.1, 6.2 Hz, 2H), 2.19 (ddd, *J* = 13.9, 8.6, 2.6 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 136.8, 134.4, 133.8, 130.1, 128.3, 126.8, 69.6, 62.8, 27.4, 26.2; HRMS (ESI) *m/z* 239.0011 (M+Na)⁺, calc. for C₁₀H₁₀OC₂Na⁺ 239.0006.

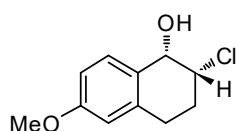
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 210 nm; retention time: 8.0 min (minor) and 9.6 min (major).



Entry	Retention Time	Area	Height	%Area
1	8.193	307.8653	791.58	50.31
2	9.895	304.1234	779.17	49.69

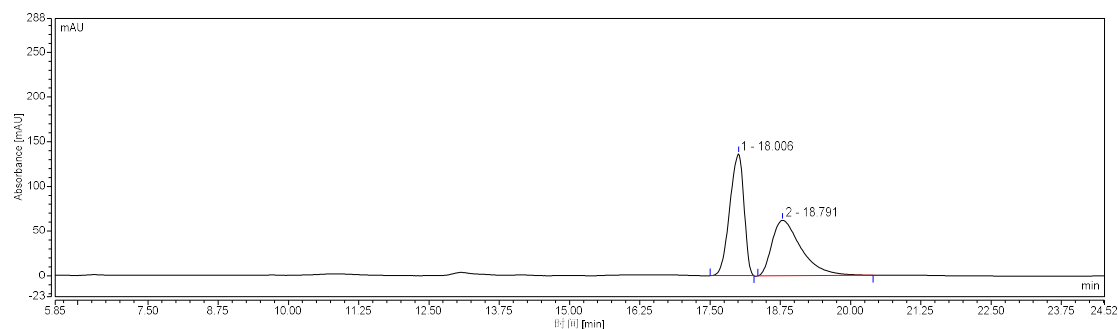


Entry	Retention Time	Area	Height	%Area
1	8.056	6.2293	17.42	4.85
2	9.640	122.1139	325.71	95.15

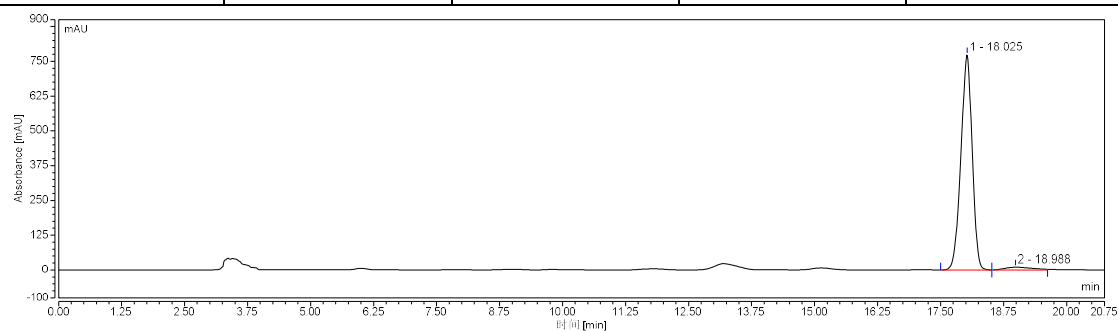


6g: white solid; Mp 115.7–117.1 °C; 15.3 mg, 72% yield; 94% ee; > 20:1 dr; $[\alpha]_{\text{D}}^{22} +8.1$ (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.38 (d, *J* = 8.6 Hz, 1H), 6.80 (dd, *J* = 8.5, 2.3 Hz, 1H), 6.64 (s, 1H), 4.80 (d, *J* = 2.9 Hz, 1H), 4.50 (dt, *J* = 9.0, 2.9 Hz, 1H), 3.79 (s, 3H), 3.07 (dt, *J* = 17.3, 6.2 Hz, 1H), 2.87–2.76 (m, 1H), 2.41 (dt, *J* = 13.8, 7.4 Hz, 1H), 2.17 (dtd, *J* = 8.9, 6.3, 2.7 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 159.4, 136.4, 130.3, 128.1, 113.0, 112.8, 69.7, 63.4, 55.2, 27.4, 27.2; HRMS (ESI) *m/z* 235.0500 (M+Na)⁺, calc. for C₁₁H₁₃O₂NaCl⁺ 235.0502.

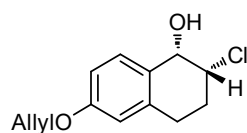
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 210 nm; retention time: 18.0 min (major) and 19.0 min (minor).



Entry	Retention Time	Area	Height	%Area
1	18.006	39.7601	136.18	51.53
2	18.791	37.3934	62.07	48.47



Entry	Retention Time	Area	Height	%Area
1	18.025	200.7196	773.56	97.01
2	18.988	6.1952	9.81	2.99

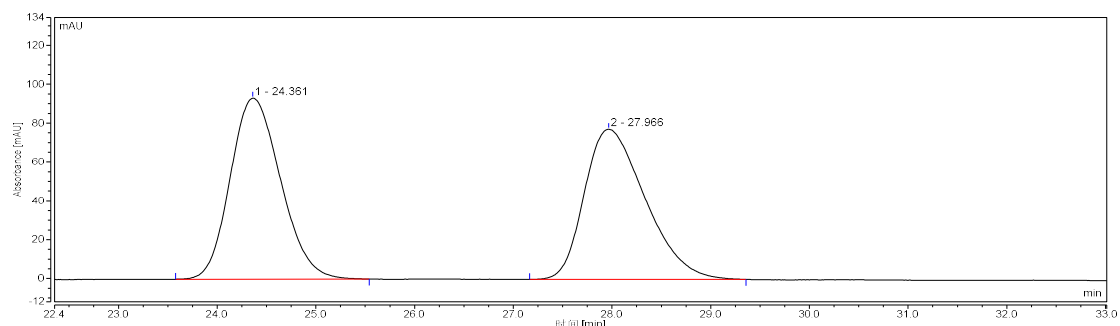


6h: white solid; Mp 92.8–93.9 °C; 16.5 mg, 69% yield; 94% ee; > 20:1

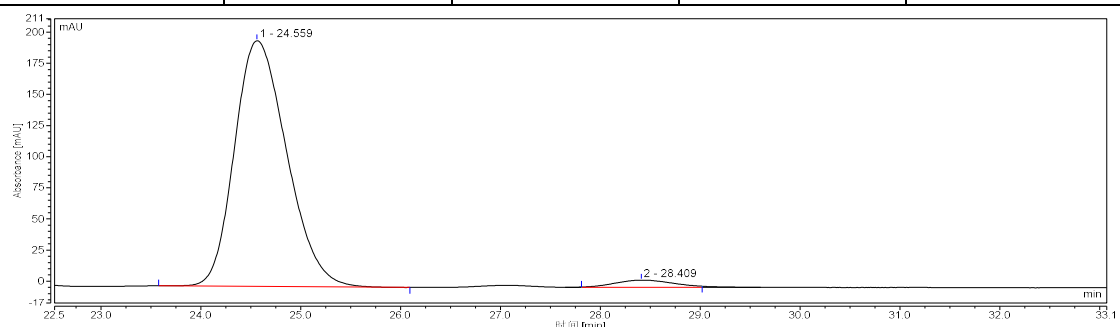
dr; $[\alpha]_{\text{D}}^{22} +3.8$ (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.37 (d, *J* = 8.5 Hz, 1H), 6.81 (dd, *J* = 8.5, 2.4 Hz, 1H), 6.65 (d, *J* = 2.1 Hz, 1H),

6.04 (ddd, *J* = 22.4, 10.5, 5.3 Hz, 1H), 5.40 (dd, *J* = 17.3, 1.4 Hz, 1H), 5.28 (dd, *J* = 10.5, 1.2 Hz, 1H), 4.80 (s, 1H), 4.52–4.51 (m, 2H), 4.48 (t, *J* = 3.1 Hz, 1H), 3.06 (dt, *J* = 17.3, 6.2 Hz, 1H), 2.81 (dt, *J* = 17.3, 6.9 Hz, 1H), 2.46–2.35 (m, 2H), 2.16 (dtd, *J* = 8.9, 6.3, 2.7 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 158.4, 136.4, 133.1, 130.3, 128.3, 117.7, 113.9, 113.5, 69.7, 68.7, 63.3, 27.4, 27.2; HRMS (ESI) *m/z* 261.0653 (M+Na)⁺, calc. for C₁₃H₁₅O₂NaCl⁺ 261.0651.

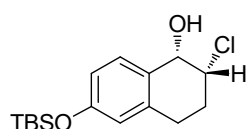
The ee was determined by HPLC analysis: CHIRALPAK IE (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 210 nm; retention time: 24.6 min (major) and 28.4 min (minor).



Entry	Retention Time	Area	Height	%Area
1	25.199	53.0888	86.83	49.84
2	29.038	53.4388	72.23	50.16

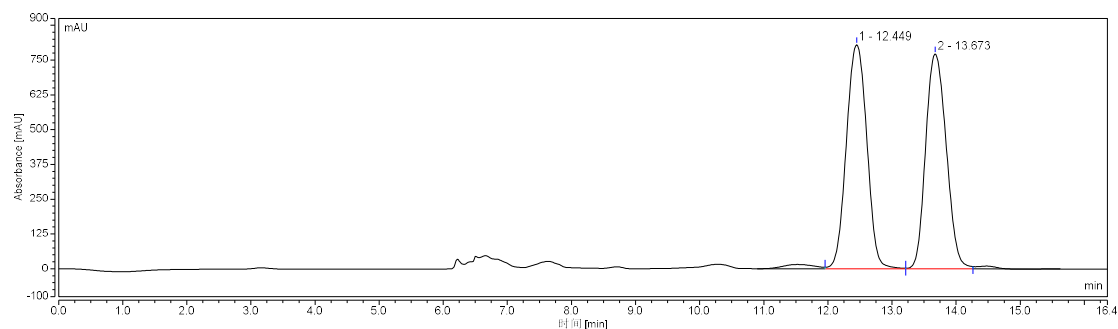


Entry	Retention Time	Area	Height	%Area
1	24.559	119.6923	197.34	96.90
2	28.409	3.8348	5.77	3.10

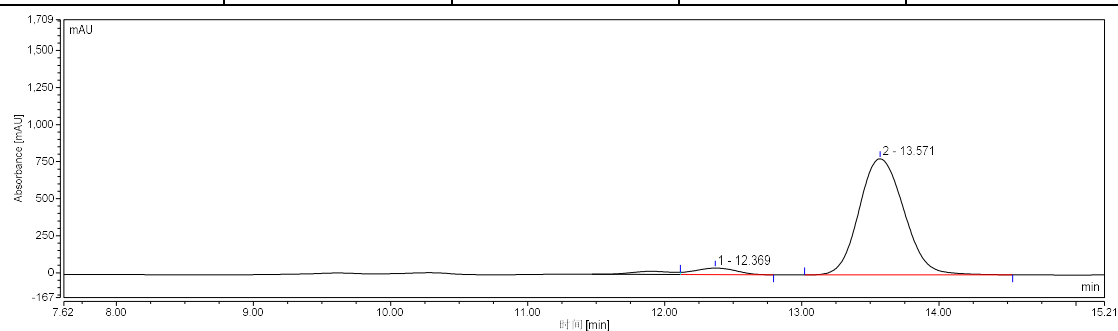


6i: colourless oil; 20.3 mg, 65% yield; 90% ee; > 20:1 dr; $[\alpha]_D^{22} +8.0$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.30 (d, $J = 8.4$ Hz, 1H), 6.71 (dd, $J = 8.4, 2.4$ Hz, 1H), 6.58 (d, $J = 2.2$ Hz, 1H), 4.78 (d, $J = 2.3$ Hz, 1H), 4.47 (dt, $J = 9.3, 3.1$ Hz, 1H), 3.02 (dt, $J = 17.3, 6.1$ Hz, 1H), 2.78 (dt, $J = 17.3, 7.0$ Hz, 1H), 2.47–2.34 (m, 2H), 2.14 (dtd, $J = 9.0, 6.2, 2.8$ Hz, 1H), 0.99 (s, 9H), 0.20 (s, 6H); ^{13}C NMR (75 MHz, CDCl_3) δ 155.5, 136.3, 130.3, 128.6, 119.4, 118.5, 69.8, 63.3, 27.3, 27.2, 25.6, 18.1, -4.4 ; HRMS (ESI) m/z 335.1205 ($\text{M}+\text{Na}$) $^+$, calc. for $\text{C}_{16}\text{H}_{25}\text{O}_2\text{NaCl}^+$ 335.1203.

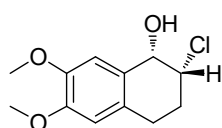
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 210 nm; retention time: 12.4 min (minor) and 13.6 min (major).



Entry	Retention Time	Area	Height	%Area
1	12.449	298.9512	805.95	50.93
2	13.673	288.0571	772.92	49.07

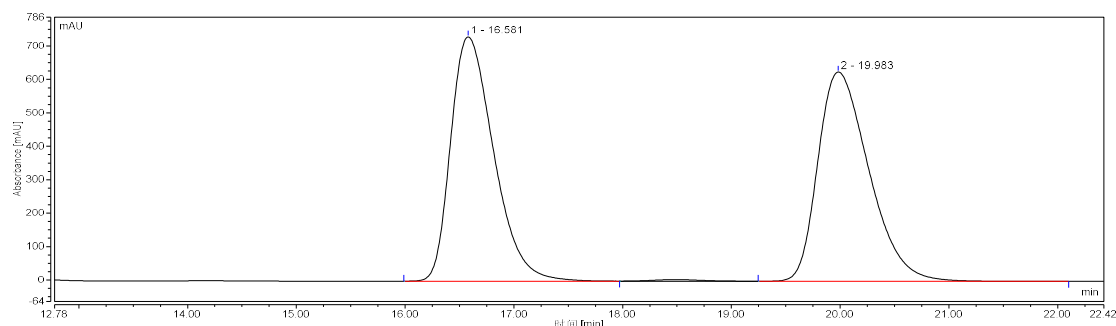


Entry	Retention Time	Area	Height	%Area
1	12.369	15.3647	43.85	4.93
2	13.571	296.4483	783.38	95.07

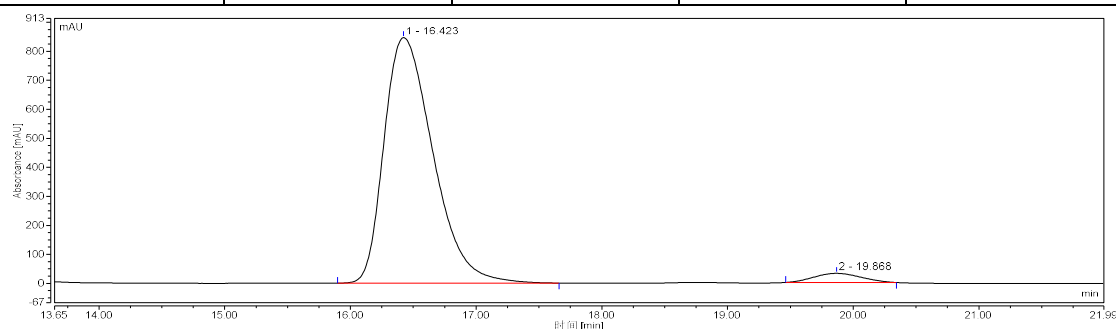


6j: colourless oil; 17.7 mg, 73% yield; 93% ee; > 20:1 dr; $[\alpha]_{\text{D}}^{22} -4.7$ (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 6.96 (s, 1H), 6.57 (s, 1H), 4.78 (d, *J* = 2.6 Hz, 1H), 4.51 (dt, *J* = 8.5, 2.8 Hz, 1H), 3.86 (s, 3H), 3.84 (s, 3H), 3.01 (dt, *J* = 16.8, 6.4 Hz, 1H), 2.75 (dt, *J* = 16.9, 6.5 Hz, 1H), 2.38 (dt, *J* = 14.5, 6.6 Hz, 2H), 2.15 (dtd, *J* = 9.0, 6.6, 2.4 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 148.9, 147.7, 127.6, 127.1, 111.1, 110.7, 69.8, 63.3, 55.8, 55.8, 27.6, 26.3; HRMS (ESI) *m/z* 265.0602 (*M*+Na)⁺, calc. for C₁₀H₉O₂FN⁺ 265.0599.

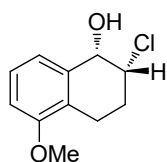
The ee was determined by HPLC analysis: CHIRALPAK IE (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 210 nm; retention time: 16.4 min (major) and 19.9 min (minor).



Entry	Retention Time	Area	Height	%Area
1	16.581	335.1269	730.99	49.63
2	19.983	340.0737	626.04	50.37



Entry	Retention Time	Area	Height	%Area
1	16.423	381.9103	846.15	96.55
2	19.868	13.6577	31.54	3.45



6k: white solid; Mp 119.8–121.1 °C; 17.4 mg, 82% yield; 91% ee; > 20:1 dr;

$[\alpha]_{\text{D}}^{22} -13.0$ (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.22 (d, *J* = 7.9 Hz,

1H), 7.11 (d, *J* = 7.7 Hz, 1H), 6.79 (d, *J* = 8.1 Hz, 1H), 4.83 (d, *J* = 3.1 Hz,

1H), 4.51 (dt, *J* = 8.7, 3.0 Hz, 1H), 3.83 (s, 3H), 2.96 (dt, *J* = 18.1, 6.6 Hz, 1H), 2.70 (dt, *J* =

18.1, 6.7 Hz, 1H), 2.47–2.35 (m, 1H), 2.18 (dtd, *J* = 9.1, 6.5, 2.5 Hz, 2H); ¹³C NMR (75 MHz,

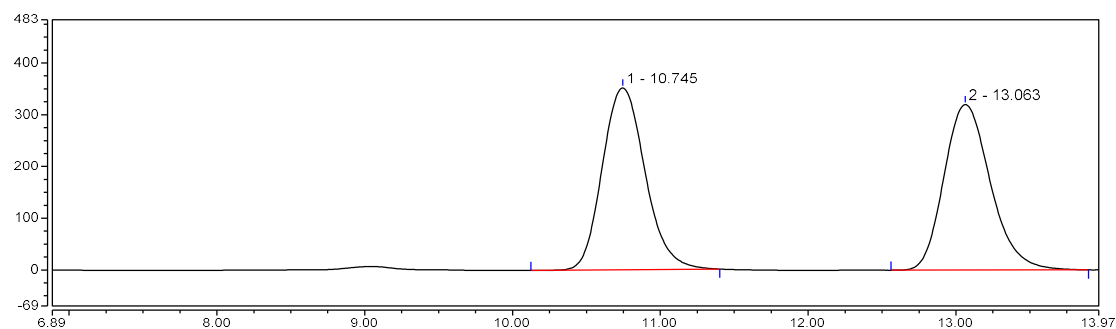
CDCl₃) δ 156.8, 137.0, 127.1, 124.0, 120.6, 109.2, 70.1, 63.0, 55.3, 27.0, 21.2; HRMS (ESI)

m/z 235.0499 (M+Na)⁺, calc. for C₁₁H₁₃O₂NaCl⁺ 235.0502.

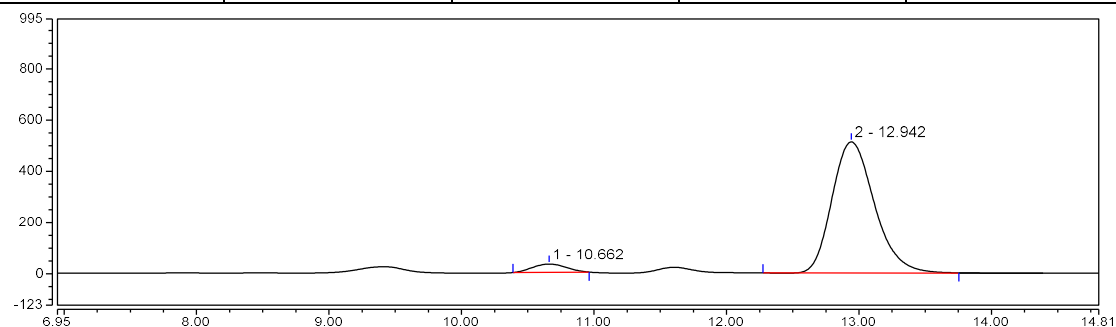
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm);

Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 210 nm; retention time: 10.7 min

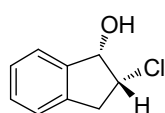
(minor) and 12.9 min (major).



Entry	Retention Time	Area	Height	%Area
1	10.745	118.4533	351.74	50.41
2	13.063	116.5267	319.96	49.59



Entry	Retention Time	Area	Height	%Area
1	10.662	9.4449	32.01	4.73
2	12.942	190.1631	512.34	95.27



6l: white solid; Mp 111.4–112.6 °C; 10.9 mg, 65% yield; 87% ee; >20:1 dr;

$[\alpha]_{\text{D}}^{22} -7.4$ (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.49–7.46 (m, 1H),

7.34–7.28 (m, 3H), 5.18 (d, *J* = 4.5 Hz, 1H), 4.83 (dd, *J* = 8.4, 5.0 Hz, 1H),

3.33 (qd, *J* = 16.6, 4.2 Hz, 2H), 2.53 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 141.4, 138.9,

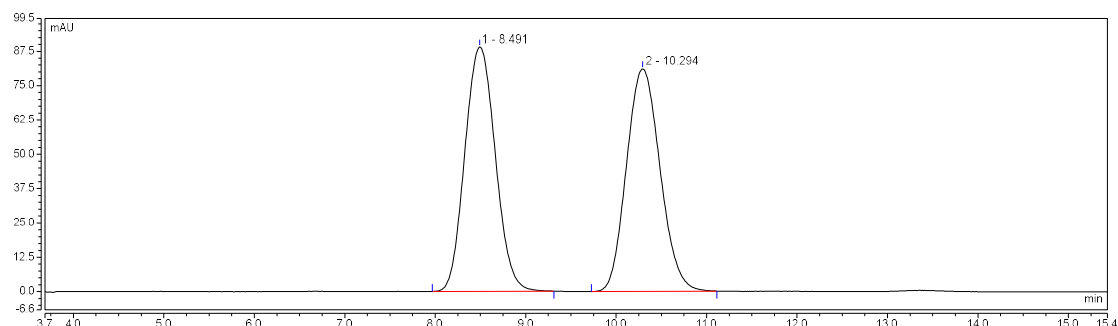
128.8, 127.5, 125.0, 124.7, 76.4, 65.9, 39.7; HRMS (ESI) *m/z* 191.0242 (M+Na)⁺, calc. for

C₁₀H₉O₂FNa⁺ 191.0240.

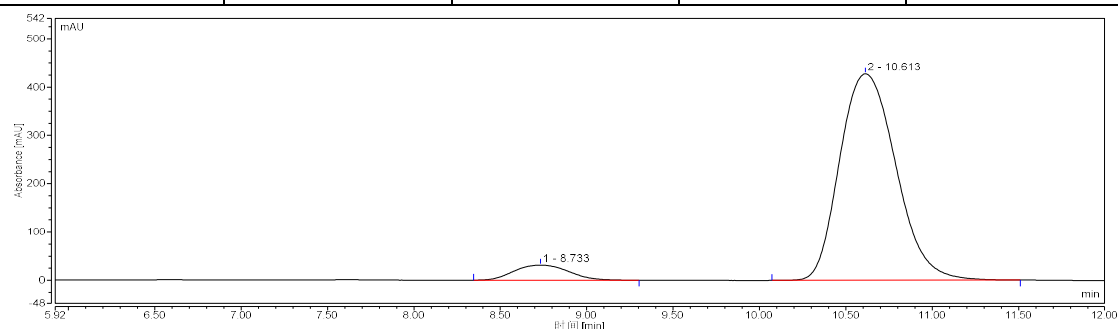
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm);

Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 210 nm; retention time: 8.5 min

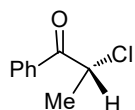
(minor) and 10.3 min (major).



Entry	Retention Time	Area	Height	%Area
1	8.491	34.78	34.7752	49.93
2	10.294	34.87	34.8705	50.07

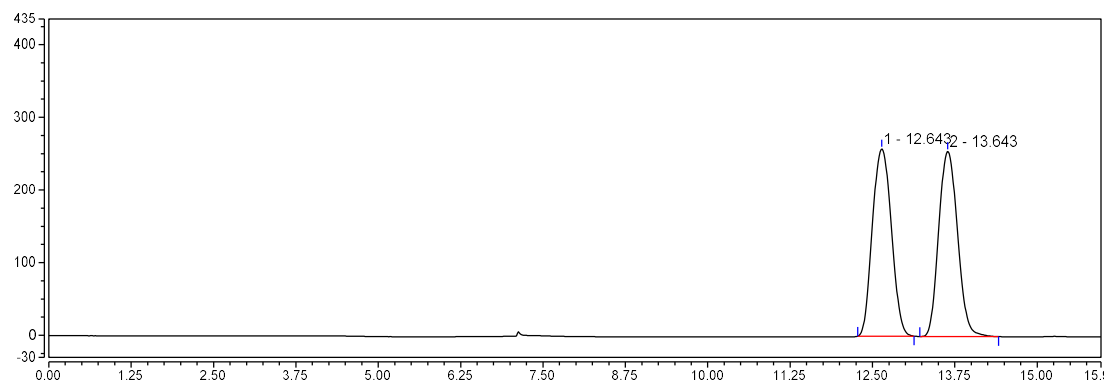


Entry	Retention Time	Area	Height	%Area
1	8.478	11.4190	31.25	6.63
2	10.294	160.9188	427.47	93.37

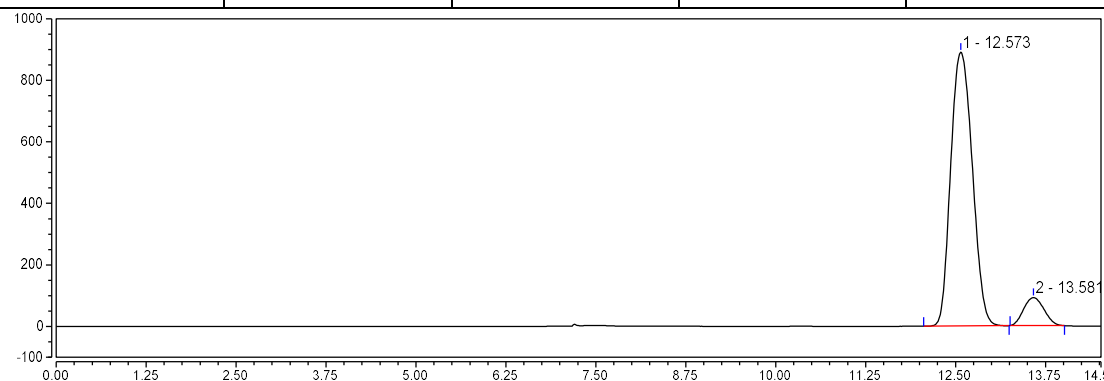


5m: colorless oil; 12.3 mg, 73% yield; 82% ee; $[\alpha]_{\text{D}}^{22} -28.2$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 8.06 – 7.97 (m, 2H), 7.59 (t, $J = 7.4$ Hz, 1H), 7.48 (t, $J = 7.5$ Hz, 2H), 5.25 (q, $J = 6.7$ Hz, 1H), 1.74 (d, $J = 6.7$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 193.6, 134.1, 133.7, 128.9, 128.7, 52.8, 19.9; HRMS (ESI) m/z 191.0236 ($\text{M}+\text{Na}^+$), calc. for $\text{C}_9\text{H}_9\text{OClNa}^+$ 191.0234.

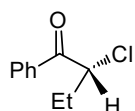
The ee was determined by HPLC analysis: CHIRALPAK IE–CHIRALPAK IE (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 98/2; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 12.6 min (major) and 13.6 min (minor).



Entry	Retention Time	Area	Height	%Area
1	12.643	86.1726	257.62	50.46
2	13.643	84.5988	254.76	49.54

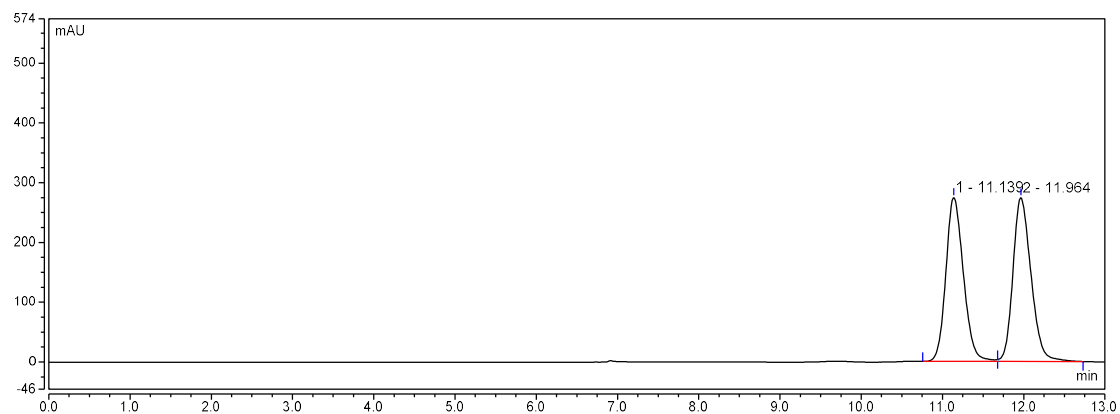


Entry	Retention Time	Area	Height	%Area
1	12.573	310.2536	889.72	91.25
2	13.581	29.7635	90.48	8.75

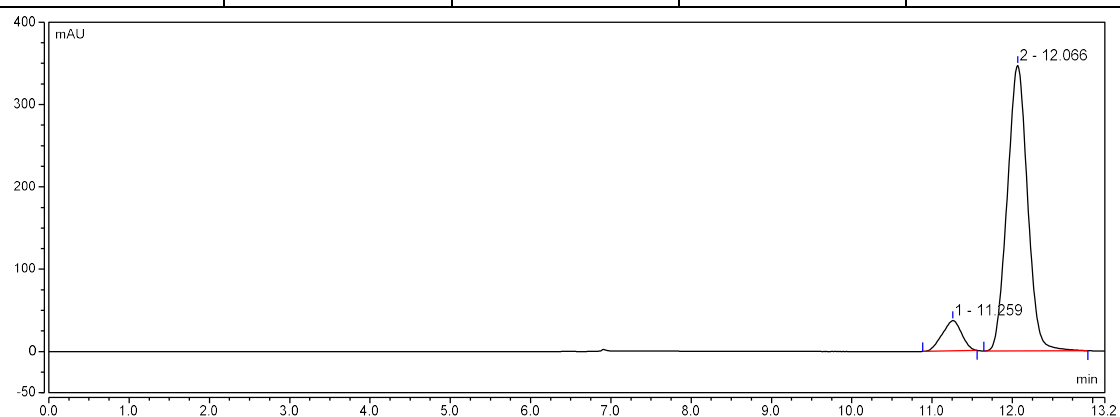


5n: colorless oil; 14.2 mg, 78% yield; 82% ee; $[\alpha]_{\text{D}}^{22} -41.3$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 8.00 (d, $J = 7.4$ Hz, 2H), 7.60 (t, $J = 7.3$ Hz, 1H), 7.49 (t, $J = 7.6$ Hz, 2H), 5.06 (dd, $J = 7.7, 5.9$ Hz, 1H), 2.27 – 1.91 (m, 2H), 1.08 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (75 MHz, DMSO) δ 193.5, 134.5, 133.6, 128.8, 128.7, 59.3, 27.0, 10.8; HRMS (ESI) m/z 205.0393 ($\text{M}+\text{Na}^+$), calc. for $\text{C}_{10}\text{H}_{11}\text{OCINa}^+$ 205.0390.

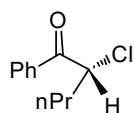
The ee was determined by HPLC analysis: CHIRALPAK ID–CHIRALPAK ID (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 98/2; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 11.3 min (minor) and 12.1 min (major).



Entry	Retention Time	Area	Height	%Area
1	11.139	68.7268	274.42	49.31
2	11.964	70.6429	274.22	50.69

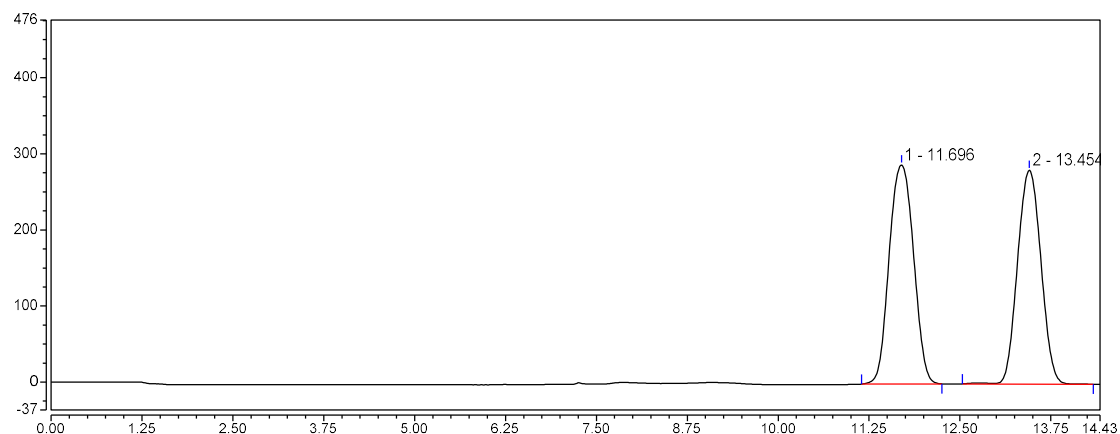


Entry	Retention Time	Area	Height	%Area
1	11.259	10.3188	36.76	9.24
2	12.066	101.4056	346.86	90.76

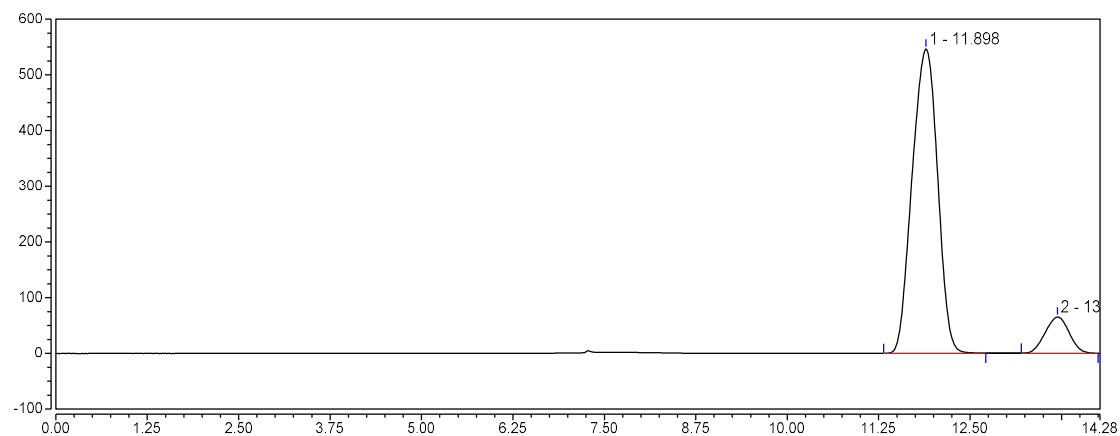


5o: colorless oil; 16.3 mg, 83% yield; 80% ee; $[\alpha]_D^{22} -67.9$ (c 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.05 – 7.95 (m, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.48 (t, *J* = 7.5 Hz, 2H), 5.13 (dd, *J* = 8.1, 5.8 Hz, 1H), 2.17 – 1.90 (m, 2H), 1.69 – 1.35 (m, 2H), 0.97 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 193.6, 134.5, 133.6, 128.8, 128.7, 57.5, 35.5, 19.5, 13.5; HRMS (ESI) *m/z* 219.0544 (M+Na)⁺, calc. for C₁₁H₁₃OCINa⁺ 219.0547.

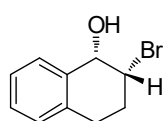
The ee was determined by HPLC analysis: CHIRALPAK IE–CHIRALPAK IE (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 98/2; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 11.9 min (major) and 13.7 min (minor).



Entry	Retention Time	Area	Height	%Area
1	11.696	112.0732	287.85	51.66
2	13.454	104.8790	280.93	48.34

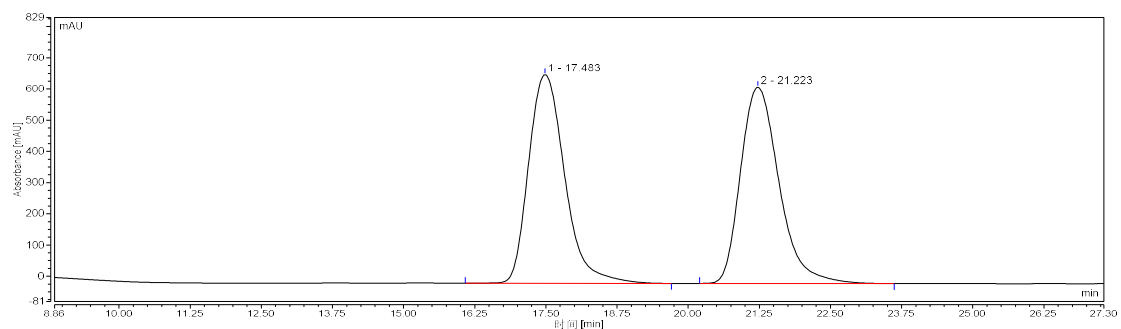


Entry	Retention Time	Area	Height	%Area
1	11.898	546.49	220.4470	89.89
2	13.696	65.04	24.8064	10.11

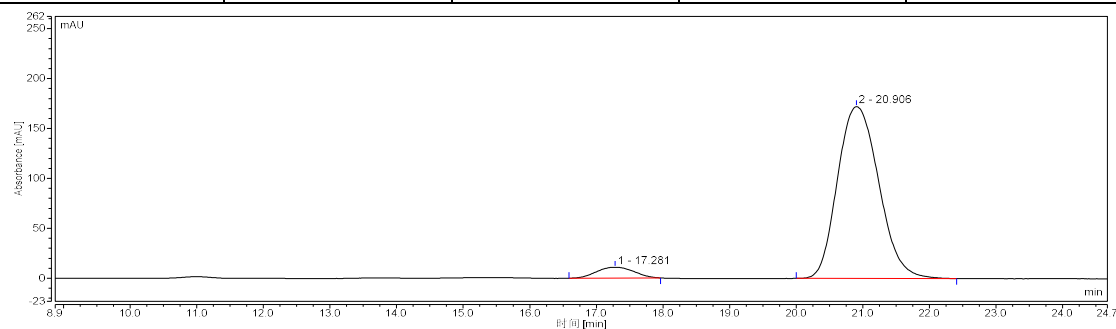


8: white solid; Mp 62.7–64.0 °C; 13.8 mg, 61% yield; 90% ee; > 20:1 dr; $[\alpha]_D^{22}$ –5.4 (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.50–7.47 (m, 1H), 7.26–7.23 (m, 2H), 7.13–7.10 (m, 1H), 4.80 (d, *J* = 3.0 Hz, 1H), 4.71 (dt, *J* = 8.3, 3.0 Hz, 1H), 3.11 (dt, *J* = 17.2, 6.5 Hz, 1H), 2.87 (dt, *J* = 17.2, 6.3 Hz, 1H), 2.52 (td, *J* = 14.3, 6.4 Hz, 1H), 2.30 (dtd, *J* = 9.4, 6.9, 2.7 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 136.0, 134.6, 128.6, 128.5, 128.2, 126.5, 70.2, 58.4, 28.3, 27.5; HRMS (ESI) *m/z* 248.9886 (M+Na)⁺, calc. for C₁₀H₁₁ONaBr⁺ 248.9891.

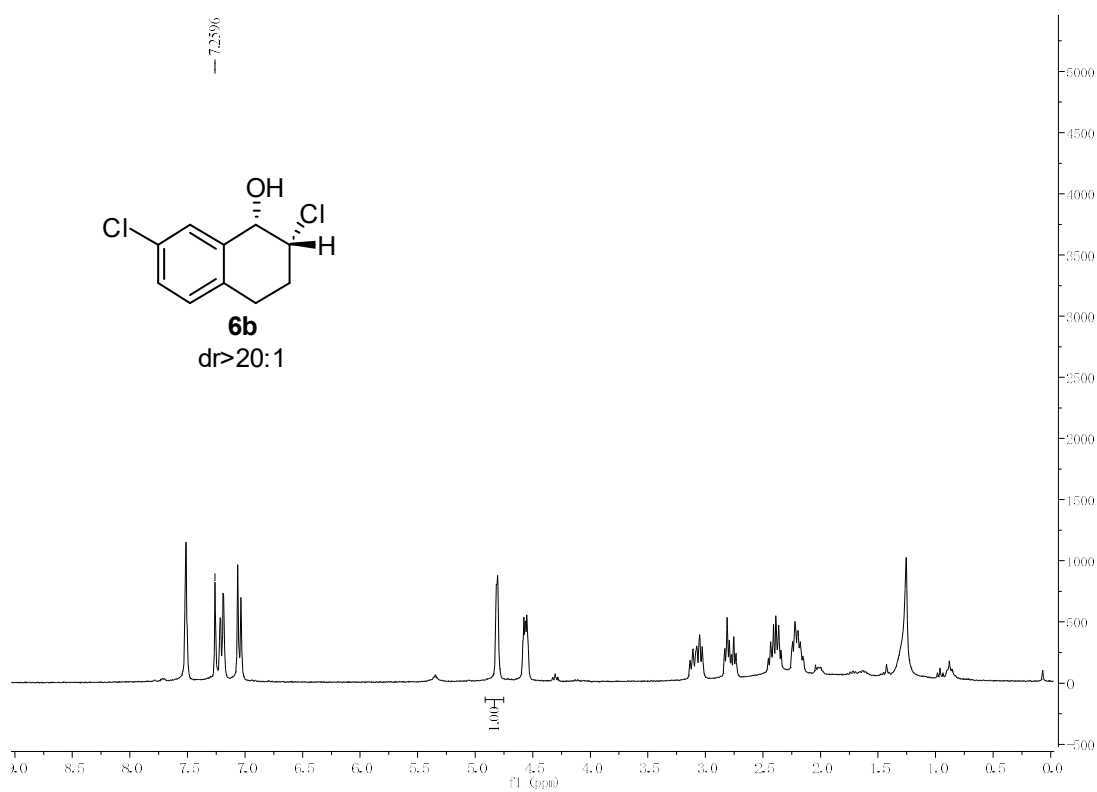
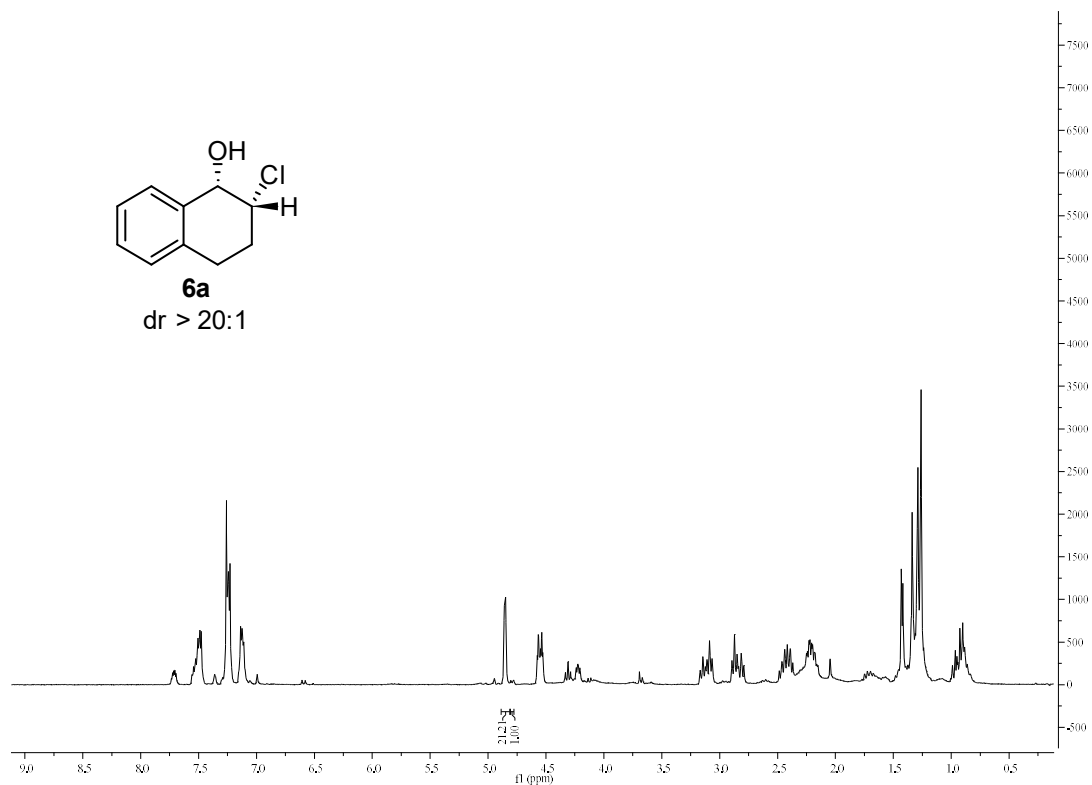
The ee was determined by HPLC analysis: CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 0.5 mL/min; 25 °C; 210 nm; retention time: 17.3 min (minor) and 20.9 min (major).

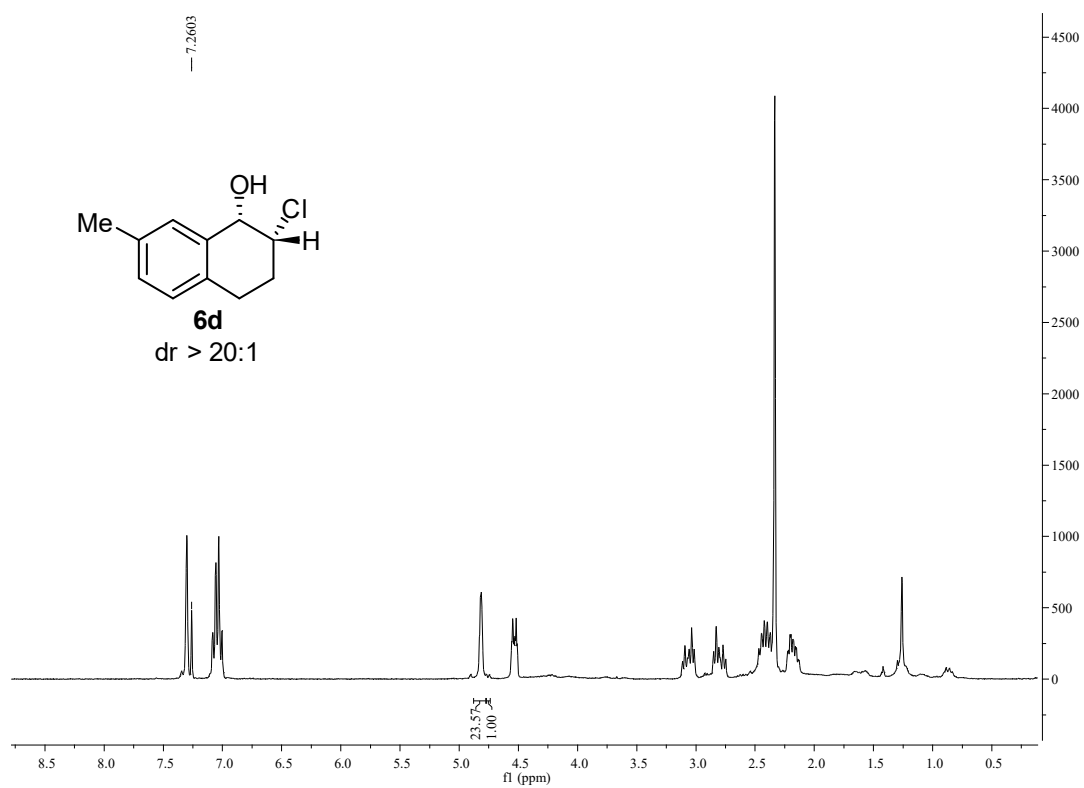
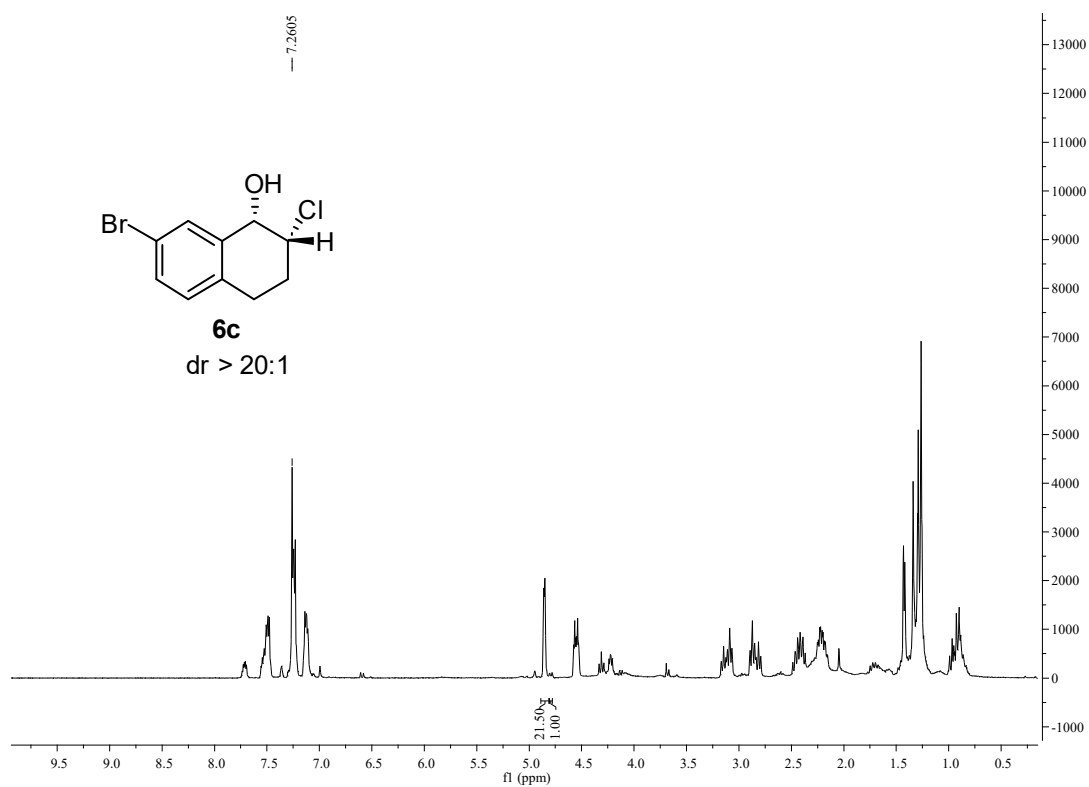


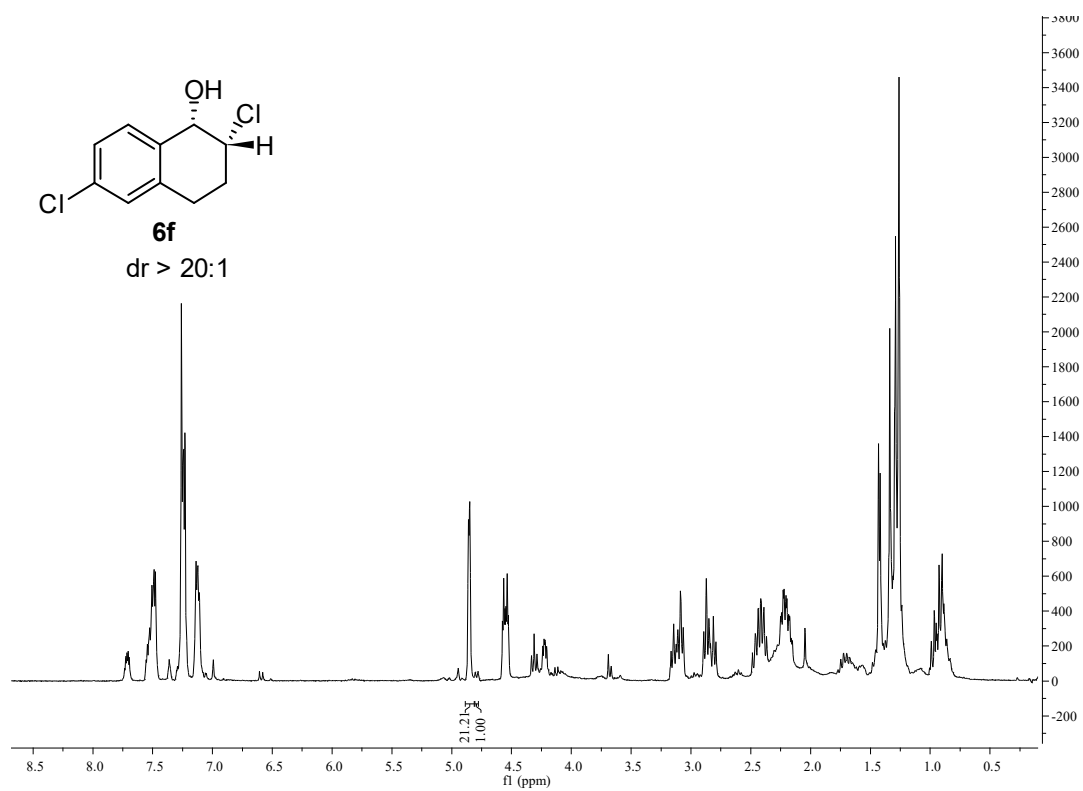
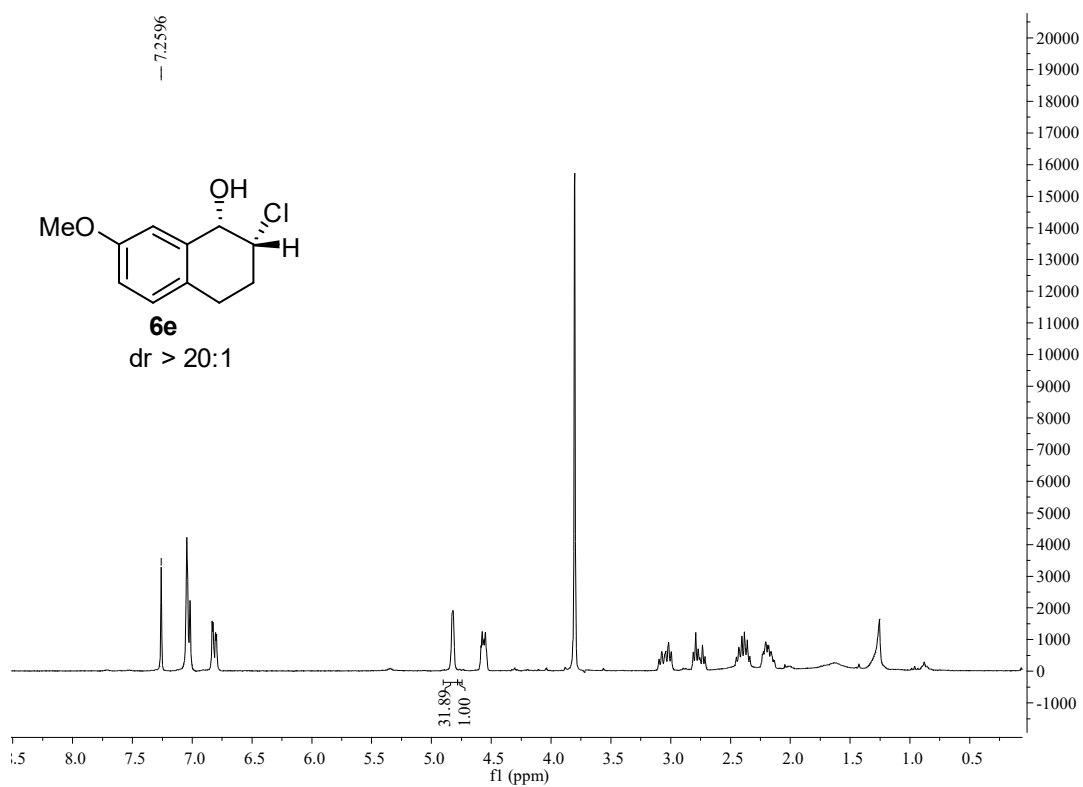
Entry	Retention Time	Area	Height	%Area
1	17.483	492.8670	668.75	50.02
2	21.223	492.5678	628.54	49.98

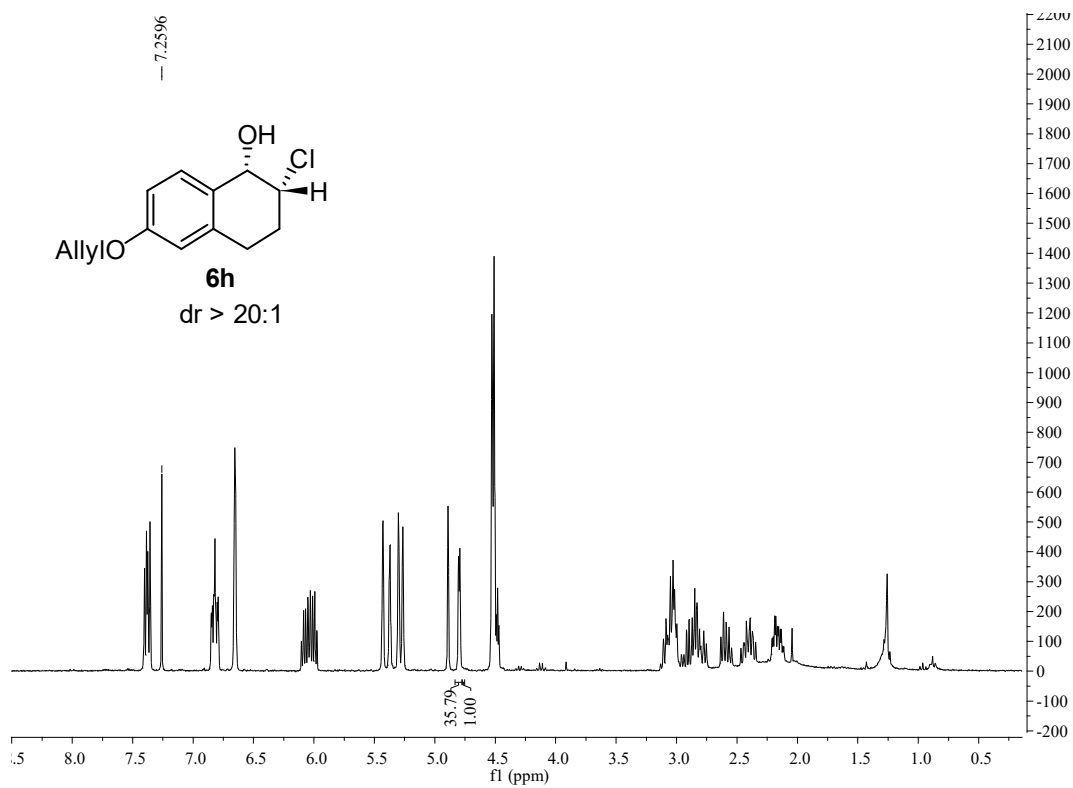
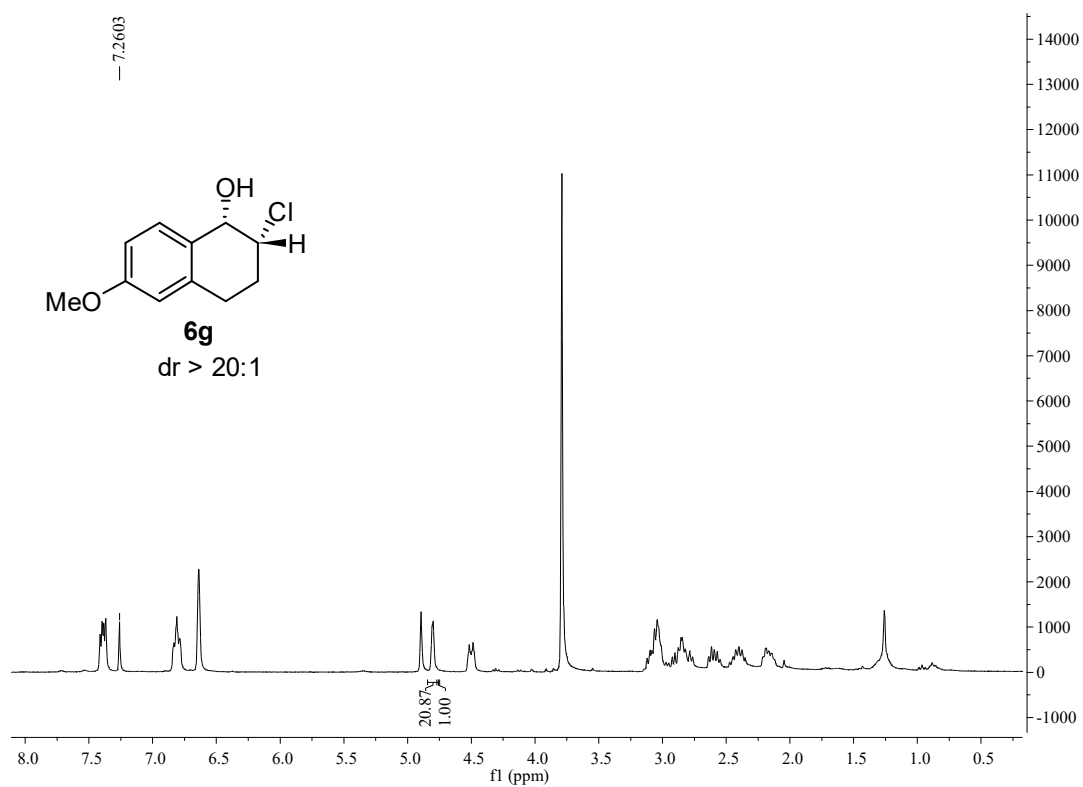


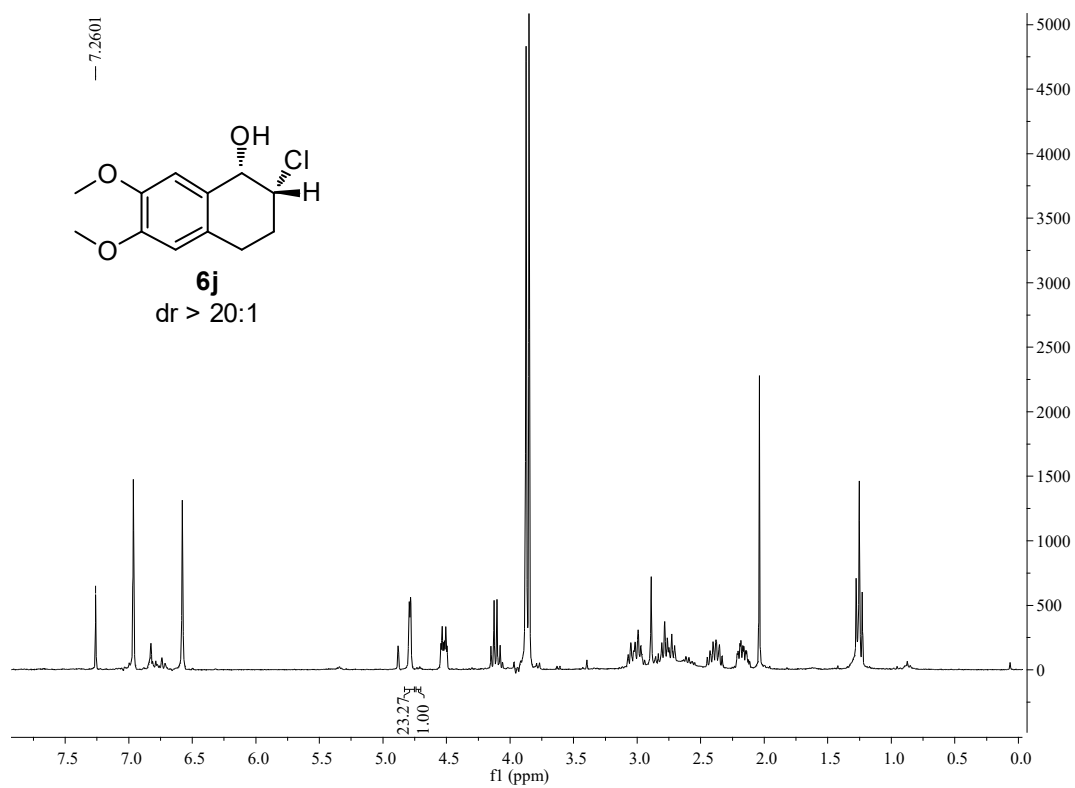
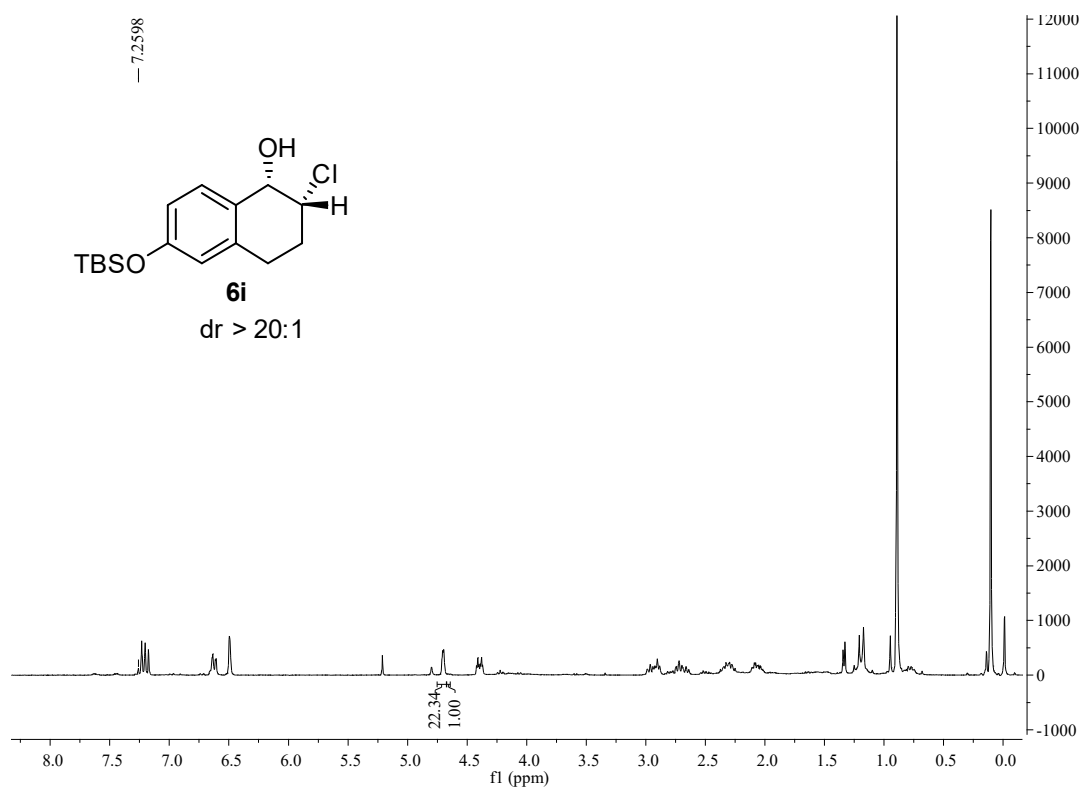
Entry	Retention Time	Area	Height	%Area
1	17.281	6.5102	10.44	5.00
2	20.906	123.7149	172.28	95.00

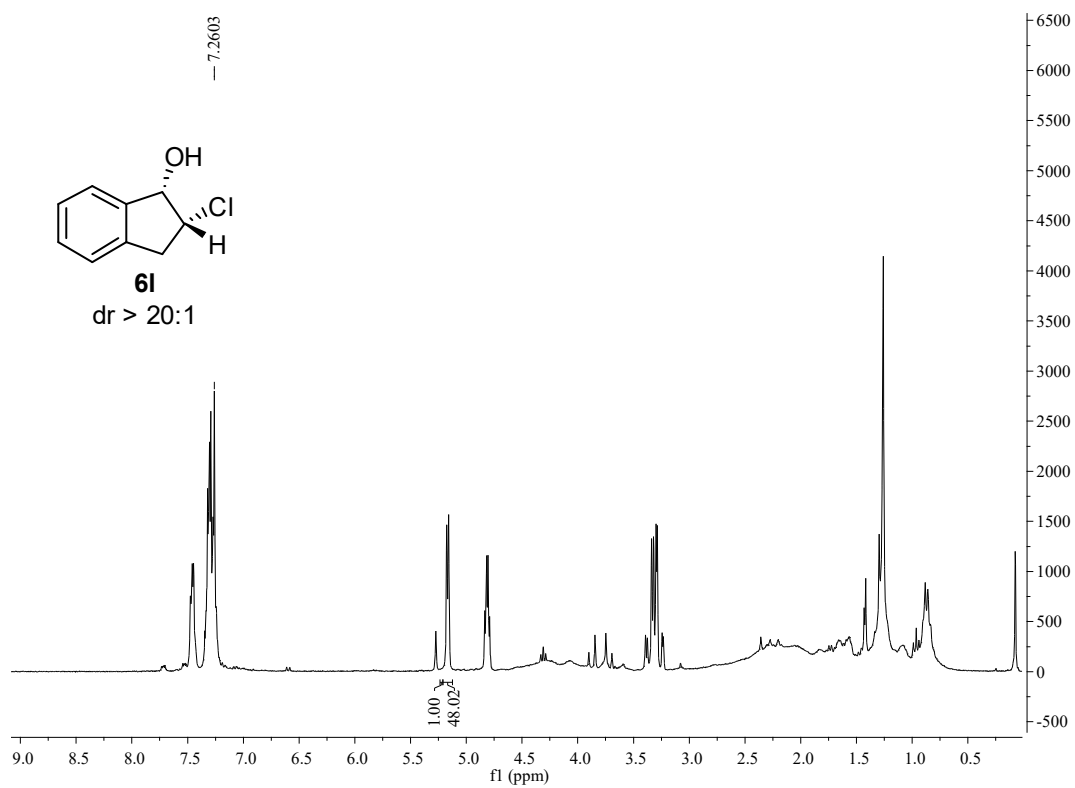
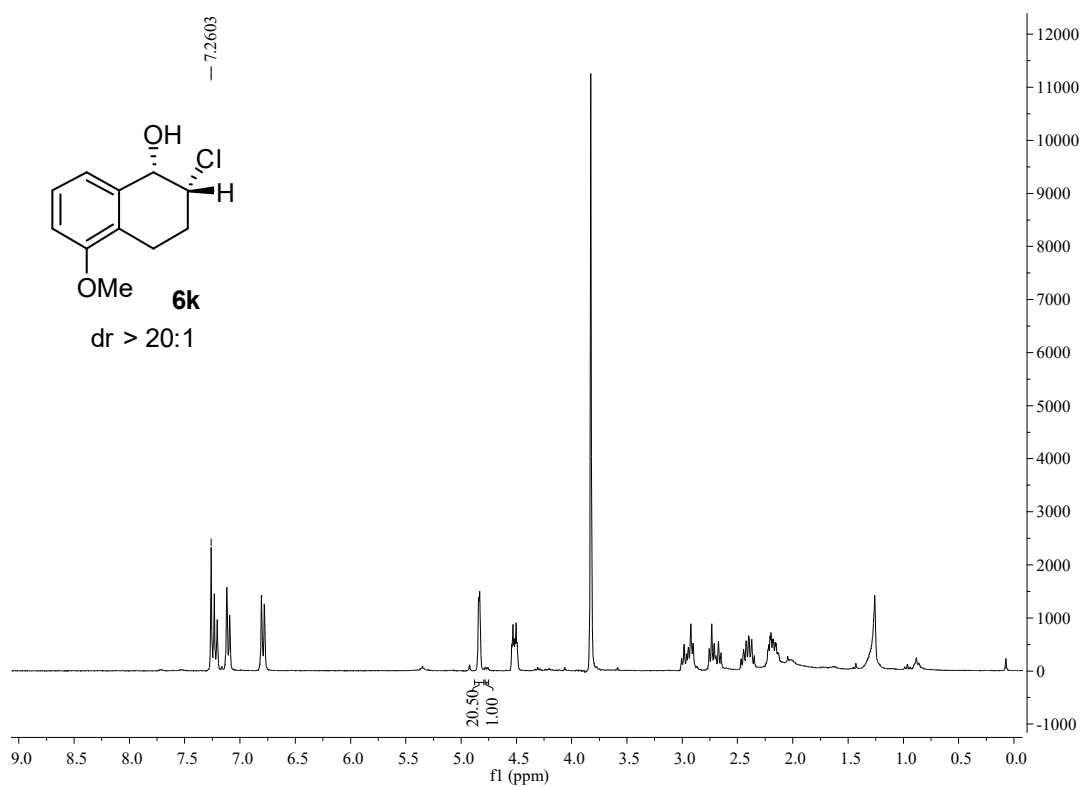
9. Crude ^1H NMR spectra to determine dr

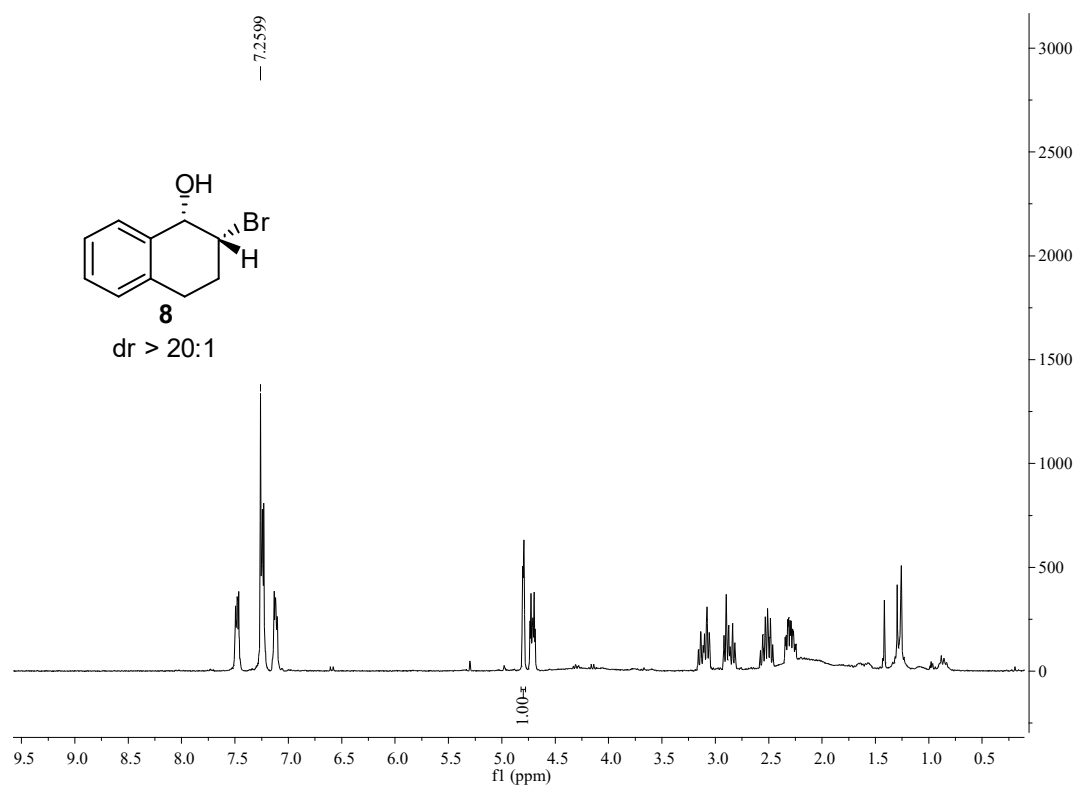












10. Copies of NMR spectra

