

Facile Triflic Acid-Catalyzed α -1,2-Cis-Thiol Glycosylations: Scope and Applications to the Synthesis of S-Linked Oligosaccharides, Glycolipids, Sublancin Glycopeptide and TN/TF Antigens

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1. Supporting Figure

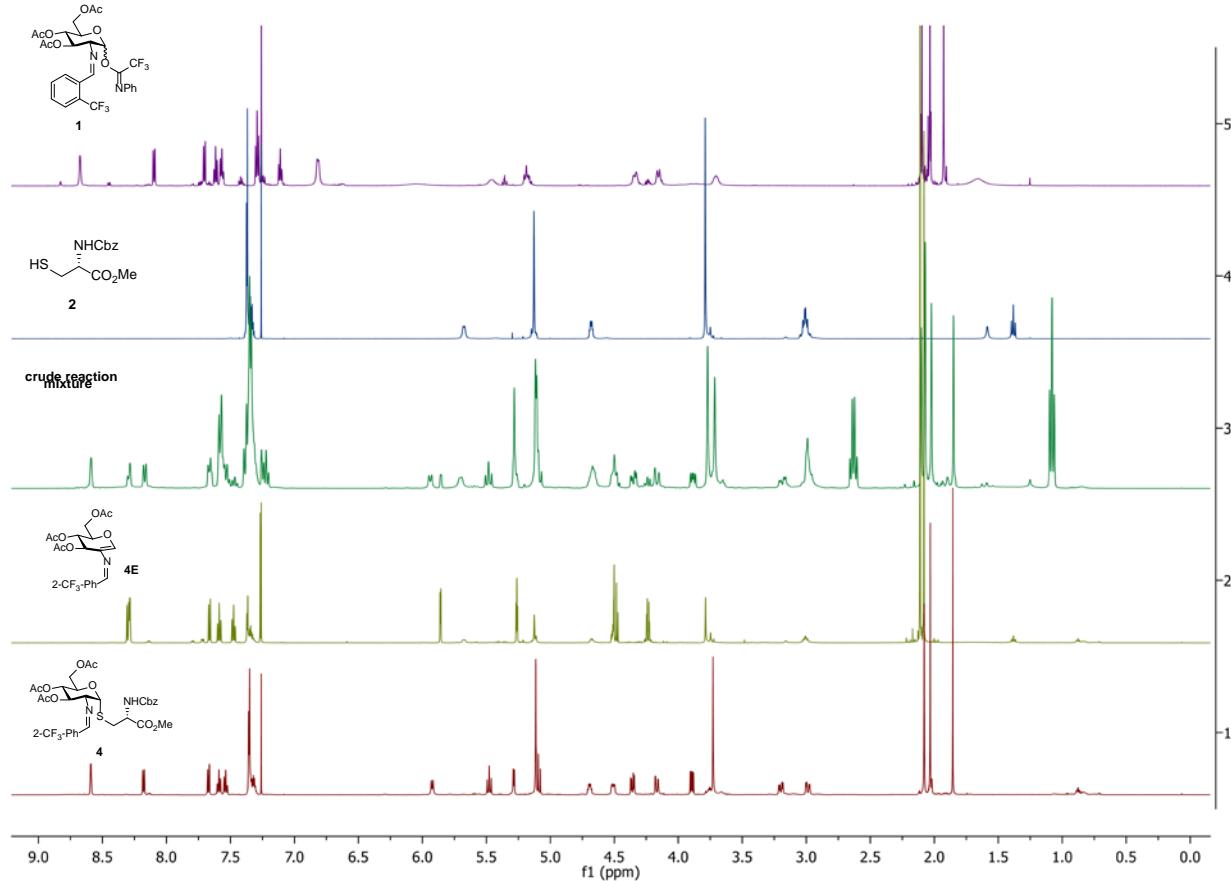


Figure S1. ¹H NMR Analysis of Glycosylation of Cysteine Amino acid Nucleophile 2 with *N*-Phenyl Trifluoroacetimidate Electrophile 1

The crude mixture resulted from the coupling of cysteine amino acid nucleophile **2** with *N*-phenyl trichloroacetimidate electrophile **1** was first analyzed by ¹H NMR spectroscopy. The crude mixture was purified by silica gel flash chromatography to separate the desired product **4** from the undesired elimination product **4E**. The result indicated the electrophile **1** was fully converted to the desired product **4** as exclusive α -isomer accompanied with the elimination product **4E**.

2. General Information

Methods and Reagents. All reactions were performed in oven-dried flasks fitted with septa under a positive pressure of nitrogen atmosphere. Organic solutions were concentrated using a Buchi rotary evaporator below 40 °C at 25 torr. Analytical thin-layer chromatography was routinely utilized to monitor the progress of the reactions and performed using pre-coated glass plates with 230-400 mesh silica gel impregnated with a fluorescent indicator (250 nm). Visualization was then achieved using UV light, iodine, or ceric ammonium molybdate. Flash column chromatography was performed using 40-63 µm silica gel (SiliaFlash F60 from Silicycle). Dry solvents were obtained from a SG Waters solvent system utilizing activated alumina columns under an argon pressure. All other commercial reagents were used as received from Sigma Aldrich, Alfa Aesar, Acros Organics, TCI, and Combi-Blocks, unless otherwise noted.

Instrumentation. All new compounds were characterized by Nuclear Magnetic Resonance (NMR) spectroscopy and High-Resolution Mass spectrometry (HRMS). All ¹H NMR spectra were recorded on either Agilent 400 or 600 MHz spectrometers. All ¹³C NMR spectra were recorded on either Agilent 100 or 150 MHz spectrometer. Chemical shifts are expressed in parts per million (δ scale) referenced to the residual proton in the NMR solvent (CDCl_3 : δ 7.26 ppm, δ 77.16 ppm). Data are presented as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, and bs = broad singlet), integration, and coupling constant in hertz (Hz).

High resolution mass spectra (HRMS) were recorded using a Micromass LCT Premier XE instrument (Waters) and were determined by electrospray ionization (ESI).

3. Optimization Studies

Table S1. Reaction Development^[a]

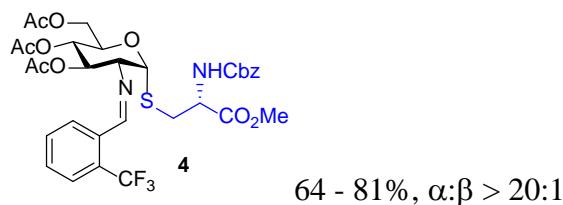
The reaction scheme illustrates the thiol glycosylation of donor **1** (N-phenyl trifluoroacetimidate glycosyl donor) with cysteine acceptors **2** or **3** (cysteine derivatives with R groups Cbz or Fmoc). The reaction conditions involve CH₂Cl₂ as the solvent and various catalysts (Ni(OTf)₂, TfOH) at different temperatures (35 °C or 25 °C) for varying times (1–20 min). The products are the α and β anomers of the thioether-linked glycosides, labeled **4** (R = Cbz) and **5** (R = Fmoc).

entry	1 (equiv.)	2 or 3 (equiv.)	catalyst	temp (°C)	time (min)	4 or 5 yield (α:β)
1	1	2 (1.5)	15 mol% Ni(OTf) ₂	35	16	4 : 66% (>20:1)
2	1	2 (1.5)	5 mol% TfOH	35	1	4 : 64% (>20:1)
3	1	2 (1.5)	5 mol% TfOH	25	2	4 : 68% (>20:1)
4	1	2 (1.5)	1 mol% TfOH	25	20	4 : 67% (>20:1)
5	1.5	2 (1.0)	3 mol% TfOH	25	3	4 : 76% (>20:1)
6	2	2 (1.0)	3 mol% TfOH	25	3	4 : 81% (>20:1)
7	2	2 (1.0)	5 mol% TfOH	25	1	4 : 80% (>20:1)
8	2	3 (1.0)	5 mol% TfOH	25	1	5 : 78% (>20:1)

[a] The reaction was conducted with 0.1 – 0.2 mmol of **1**. Yields of isolated product averaged two runs. The (α/β) ratios were determined by ¹H NMR analysis.

Procedure for Thiol Glycosylations Optimization:

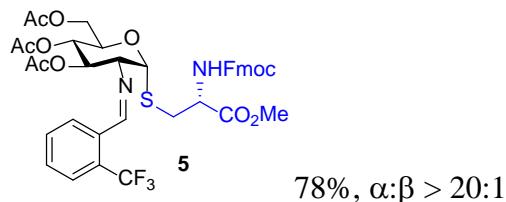
A 10 mL Schlenk flask was charged with *N*-phenyl trifluoroacetimidate glycosyl donor **1** (0.1 – 0.2 mmol, 1 – 2 equiv.), cysteine acceptor **2** or **3** (0.1 – 0.15 mmol, 1 – 1.5 equiv.) and dichloromethane (1 mL). The resulting solution was stirred at room temperature for 5 min under a nitrogen atmosphere before the catalyst (1 – 15 mol% with respect to donor **1**) was added. After the TLC shows completion of the reaction, it was quenched with 1 drop of Et₃N and concentrated. A crude ¹H NMR was taken to determine the (α/β) ratio. Further purification by silica gel column chromatography (ethyl acetate/hexane: 1/4 → 1/2) was performed to give the desired product.



¹H NMR (CDCl₃, 600 MHz): δ 8.59 (d, *J* = 2.1 Hz, 1H), 8.18 (d, *J* = 7.7 Hz, 1H), 7.67 (d, *J* = 7.7 Hz, 1H), 7.59 (t, *J* = 7.5 Hz, 1H), 7.54 (t, *J* = 7.6 Hz, 1H), 7.40 – 7.29 (m, 5H), 5.92 (d, *J* = 8.5 Hz, 1H), 5.48 (t, *J* = 9.7 Hz, 1H), 5.29 (d, *J* = 5.6 Hz, 1H), 5.10 (dd, *J* = 15.9, 6.0 Hz, 3H), 4.73 – 4.64 (m, 1H), 4.51 (dd, *J* = 10.1, 3.1 Hz, 1H), 4.36 (dd, *J* = 12.4, 4.9 Hz, 1H), 4.17 (d, *J* = 10.9 Hz, 1H), 3.89 (dd, *J* = 10.0, 5.6 Hz, 1H), 3.73 (s, 3H), 3.20 (dd, *J* = 14.2, 5.4 Hz, 1H), 2.99 (dd, *J* = 14.2, 4.2 Hz, 1H), 2.08 (s, 3H), 2.03 (s, 3H), 1.85 (s, 3H).

¹³C NMR (CDCl₃, 150 MHz): δ = 171.02, 170.59, 169.85, 169.59, 160.37, 155.74, 136.09, 133.03, 132.27, 130.90, 129.07, 128.50, 128.20, 128.11, 125.48, 125.45, 85.18, 71.97, 71.33, 68.56, 68.52, 67.11, 62.09, 53.93, 53.75, 52.63, 33.42, 20.66, 20.64, 20.34.

HRMS (ESI): calc. for C₃₂H₃₅F₃N₂O₁₁SNa (M+Na)⁺: 735.1806; found: 735.1818.



¹H NMR (CDCl₃, 600 MHz): δ 8.62 (d, *J* = 1.9 Hz, 1H), 8.19 (d, *J* = 7.7 Hz, 1H), 7.75 (t, *J* = 7.8 Hz, 2H), 7.67 (d, *J* = 7.6 Hz, 1H), 7.63 – 7.56 (m, 3H), 7.53 (t, *J* = 7.5 Hz, 1H), 7.43 – 7.36 (m, 2H), 7.34 – 7.27 (m, 2H), 6.00 (d, *J* = 8.6 Hz, 1H), 5.49 (t, *J* = 9.7 Hz, 1H), 5.27 (d, *J* = 5.6 Hz, 1H), 5.14 – 5.06 (m, 1H), 4.74 – 4.69 (m, 1H), 4.52 (dd, *J* = 10.1, 3.0 Hz, 1H), 4.42 (qd, *J* = 10.6, 7.3 Hz, 2H), 4.28 (dd, *J* = 12.3, 5.1 Hz, 1H), 4.24 – 4.19 (m, 2H), 3.92 (dd, *J* = 10.0, 5.6 Hz, 1H), 3.74 (s, 3H), 3.20 (dd, *J* = 14.3, 5.5 Hz, 1H), 2.99 (dd, *J* = 14.3, 4.1 Hz, 1H), 2.06 (s, 3H), 2.03 (s, 3H), 1.87 (s, 3H).

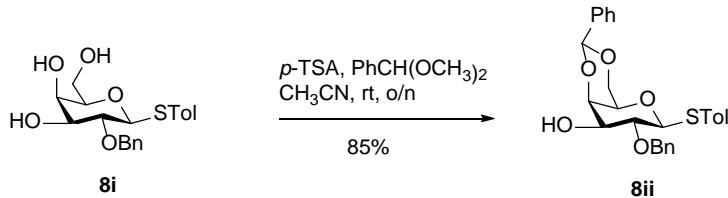
¹³C NMR (CDCl₃, 150 MHz): δ 170.96, 170.55, 169.85, 169.57, 160.36, 155.72, 143.64, 141.31, 141.28, 132.26, 130.90, 129.05, 127.71, 127.69, 127.06, 127.03, 125.49, 125.46, 125.03, 119.97, 119.94, 85.43, 72.03, 71.33, 68.64, 68.63, 67.03, 62.21, 54.05, 52.63, 47.11, 33.77, 20.65, 20.60, 20.33.

HRMS (ESI): calc. for C₃₉H₃₉F₃N₂O₁₁SNa (M+Na)⁺: 823.2119; found: 823.2116.

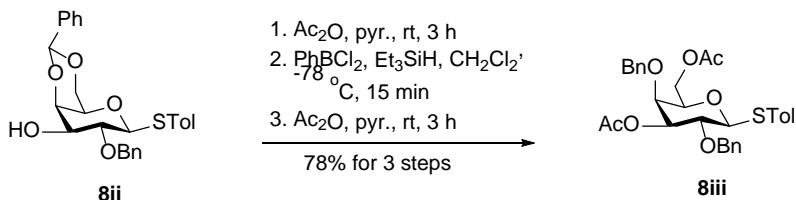
4. Preparation of Glycosyl Donors

Glycosyl donors **1**, **6** and **7** were synthesis according to our former literature.^[1]

4.1 Preparation of glycosyl donor **8**

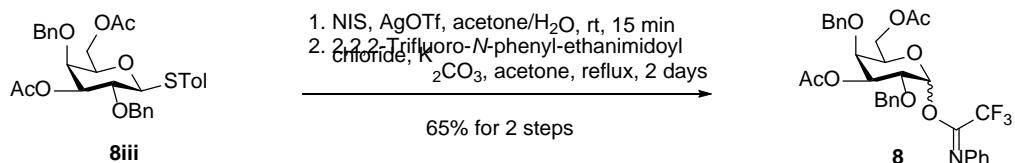


To a solution of **8i** (3.18 g, 8.46 mmol) in CH₃CN (80 mL) were added benzaldehyde dimethyl acetal (1.9 mL, 12.67 mmol) and *p*-TSA (161 mg, 0.846 mmol). After the reaction mixture was stirred at room temperature for overnight, it was quenched with Et₃N and concentrated. The residue was subjected to flush silica gel column chromatography (1:1.5, EtOAc–hexane) to give **8ii** (3.34 g, 85%) as a white foam. *R*_f 0.30 (1:1.5, EtOAc–hexane); ¹H NMR (600 MHz, CDCl₃) δ: 7.62 (d, *J* = 8.1 Hz, 2H), 7.54 – 7.48 (m, 2H), 7.46 – 7.39 (m, 5H), 7.36 (t, *J* = 7.4 Hz, 2H), 7.30 (d, *J* = 7.3 Hz, 1H), 7.07 (d, *J* = 7.9 Hz, 2H), 5.54 (s, 1H), 4.79 (d, *J* = 10.6 Hz, 1H), 4.70 (d, *J* = 10.6 Hz, 1H), 4.57 (d, *J* = 9.5 Hz, 1H), 4.38 (dd, *J* = 12.4, 1.3 Hz, 1H), 4.17 (d, *J* = 3.6 Hz, 1H), 4.00 (dd, *J* = 12.4, 1.4 Hz, 1H), 3.78 (dd, *J* = 9.0, 3.6 Hz, 1H), 3.62 (t, *J* = 9.3 Hz, 1H), 3.45 (s, 1H), 2.35 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ: 138.38, 137.71, 133.26, 129.65, 129.28, 128.88, 128.38, 128.20, 128.11, 127.78, 126.61, 101.37, 86.34, 77.10, 75.76, 75.23, 74.38, 69.75, 69.28, 21.19. HR ESI-TOF MS (m/z): calcd for C₂₇H₂₈O₅Na [M + Na]⁺, 487.1550; found, 487.1555.



A solution of **8ii** (2.0 g, 4.31 mmol) in 50 mL anhydrous pyridine was added acetic anhydride (814 μL, 8.62 mmol) and stirred at room temperature for 3 h. The reaction mixture was concentrated under reduced pressure, and the residue was subjected to flush silica gel column chromatography (1:3, EtOAc–hexane). The obtained intermediate was dissolved in 50 mL anhydrous CH₂Cl₂, PhBCl₂ (1.12 mL, 8.62 mmol) and Et₃SiH (1.37 mL, 8.62 mmol) were added at -78 °C under a N₂

atmosphere. 15 min later, the reaction was quenched by addition of CH₃OH and Et₃N. The mixture was concentrated and purified by silica gel column chromatography (1:2, EtOAc–hexane) to afford the intermediate product as syrup. It was dissolved in pyridine (50 mL) again, followed by addition of acetic anhydride (814 µL, 2.21 mmol). After the reaction mixture was stirred at room temperature for 3 h, it was concentrated and the residue was subjected to flush silica gel column chromatography (1:4, EtOAc–hexane) to give **8iii** (1.85 g, 78% for 3 steps) as a syrup. *R*_f 0.50 (1:3, EtOAc–hexane); ¹H NMR (600 MHz, CDCl₃) δ: 7.48 (d, *J* = 8.1 Hz, 2H), 7.39 – 7.28 (m, 10H), 7.06 (d, *J* = 7.9 Hz, 2H), 4.96 (dd, *J* = 9.7, 2.9 Hz, 1H), 4.87 (d, *J* = 11.0 Hz, 1H), 4.68 (d, *J* = 11.5 Hz, 1H), 4.65 – 4.52 (m, 3H), 4.30 (dd, *J* = 11.1, 6.7 Hz, 1H), 4.10 (dd, *J* = 11.2, 6.3 Hz, 1H), 3.98 – 3.89 (m, 2H), 3.71 (t, *J* = 6.5 Hz, 1H), 2.33 (s, 3H), 2.02 (s, 3H), 1.94 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ: 170.40, 170.29, 138.06, 137.69, 137.63, 132.46, 129.71, 129.60, 128.38, 128.34, 127.99, 127.96, 127.88, 127.77, 88.05, 77.00, 75.59, 75.38, 75.36, 74.85, 74.13, 62.60, 21.12, 20.90, 20.78. HR ESI-TOF MS (m/z): calcd for C₃₁H₃₄O₇SNa [M + Na]⁺, 573.1917; found, 573.1911.

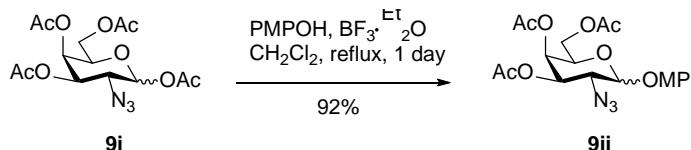


To a solution of **8iii** (1.5 g, 2.73 mmol) in acetone-H₂O (v/v, 50/1, 30 mL) were added NIS (1.23 g, 5.45 mmol) and AgOTf (140 mg, 0.545 mmol). After the reaction mixture was stirred at room temperature for 15 min, it was quenched with Et₃N, and concentrated. The residue was subjected to flush silica gel column chromatography (1:2, EtOAc–hexane) to give a hemiacetal. To a solution of this hemiacetal in anhydrous acetone (30 mL) was added 2,2,2-Trifluoro-N-phenyl-ethanimidoyl chloride (1.31 mL, 8.19 mmol) and K₂CO₃ (752 mg, 5.45 mmol) under N₂ protection. After 2 days of stirring under reflux, the mixture was filtered and concentrated in vacuum, and the residue was purified by Et₃N-neutralized silica gel column with EtOAc and hexanes (1:4) as the eluent to afford compound **8** (1.09 g, 65% for 2 steps) as a syrup.

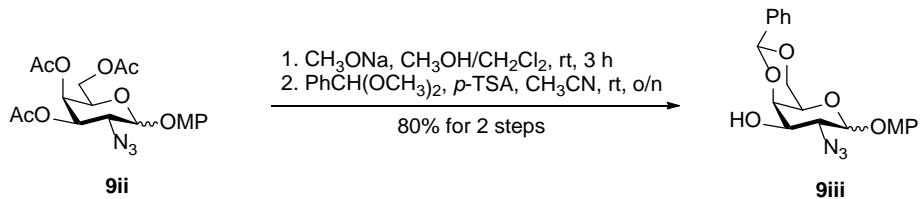
For α anomer: *R*_f 0.55 (1:3, EtOAc–hexane); ¹H NMR (600 MHz, CDCl₃) δ: 7.39 – 7.22 (m, 12H), 7.09 (t, *J* = 7.5 Hz, 1H), 6.74 (s, 2H), 6.54 (s, 1H), 5.30 (d, *J* = 10.0 Hz, 1H), 4.70 (dd, *J* = 26.1, 11.6 Hz, 3H), 4.53 (d, *J* = 11.3 Hz, 1H), 4.20 (d, *J* = 7.4 Hz, 3H), 4.10 (d, *J* = 8.1 Hz, 2H), 2.05 (s, 3H), 2.04 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ: 170.32, 170.30, 137.68, 137.31, 128.70, 128.51,

128.43, 128.11, 127.90, 127.60, 119.34, 75.29, 74.51, 73.27, 72.91, 72.11, 70.65, 62.38, 20.96, 20.72. HR ESI-TOF MS (m/z): calcd for $C_{31}H_{35}N_3O_7SNa$ [M + Na]⁺, 616.2093; found, 616.2088. For β anomer: R_f 0.50 (1:3, EtOAc–hexane); ¹H NMR (600 MHz, CDCl₃) δ : 7.42 – 7.27 (m, 12H), 7.11 (t, J = 7.5 Hz, 1H), 6.82 (d, J = 7.6 Hz, 2H), 5.70 (s, 1H), 4.97 (s, 1H), 4.84 (d, J = 11.5 Hz, 1H), 4.69 (d, J = 11.4 Hz, 2H), 4.55 (d, J = 11.5 Hz, 1H), 4.27 (dd, J = 11.0, 6.6 Hz, 1H), 4.16 – 4.02 (m, 2H), 3.94 (s, 1H), 3.75 (s, 1H), 1.99 (s, 3H), 1.98 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ : 170.25, 170.24, 143.33, 137.67, 137.31, 128.70, 128.50, 128.42, 128.25, 128.11, 128.02, 127.96, 124.30, 119.14, 97.03, 75.48, 75.04, 75.02, 74.61, 73.47, 72.89, 61.99, 20.83, 20.68. HR ESI-TOF MS (m/z): calcd for $C_{32}H_{32}F_3NO_8Na$ [M + Na]⁺, 638.1972; found, 638.1976.

4.2 Preparation of glycosyl donor **9**



To a solution of **9i** (4.5 g, 12.03 mmol) in CH₂Cl₂ (70 mL) was added 4-Methoxyphenol (4.475 g, 36.09 mmol) and boron trifluoride diethyl etherate (2.97 mL, 24.06 mmol) at room temperature. After the reaction mixture was stirred under reflux for 1 day, it was diluted with CH₂Cl₂ and washed with 1 N NaOH and brine, dried over anhydrous Na₂SO₄, and concentrated. The residue was subjected to flush silica gel column chromatography (1:3, EtOAc–hexane) to give **9ii** (4.85 g, 92%) as an inseparable α/β mixture. R_f 0.50 (1:2, EtOAc–hexane); ¹H NMR (600 MHz, CDCl₃) δ : 7.04 (dd, J = 9.0, 1.4 Hz, 5H), 6.92 – 6.76 (m, 5H), 5.57 (dd, J = 11.1, 3.2 Hz, 1.7H), 5.51 (dd, J = 4.6, 2.2 Hz, 3H), 5.37 (d, J = 3.3 Hz, 1H), 4.84 (dd, J = 10.9, 3.4 Hz, 1H), 4.79 (d, J = 8.1 Hz, 1H), 4.39 (t, J = 6.6 Hz, 1.7H), 4.21 (dd, J = 11.3, 6.9 Hz, 1H), 4.17 – 4.04 (m, 5.5H), 4.00 – 3.90 (m, 1.7H), 3.84 – 3.68 (m, 8.5H), 2.17 (s, 3H), 2.16 (s, 5H), 2.08 (s, 5H), 2.07 (s, 3H), 2.04 (s, 3H), 1.98 (s, 5H). ¹³C NMR (150 MHz, CDCl₃) δ : 170.26, 170.01, 169.96, 169.78, 169.73, 155.91, 155.63, 150.78, 150.19, 118.71, 118.21, 114.67, 114.58, 101.89, 97.94, 70.92, 68.13, 67.48, 67.37, 66.18, 61.49, 61.25, 60.62, 60.34, 57.35, 55.62, 20.63, 20.58, 20.57. HR ESI-TOF MS (m/z): calcd for $C_{19}H_{23}N_3O_9Na$ [M + Na]⁺, 460.1327; found, 460.1328.



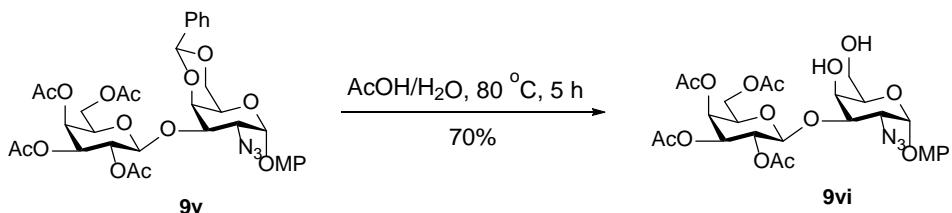
To a solution of **9ii** (4.2 g, 9.59 mmol) in $\text{CH}_3\text{OH}/\text{CH}_2\text{Cl}_2$ (v/v, 3/1, 80 mL) was added CH_3ONa (259 mg, 4.79 mmol). After the reaction mixture was stirred at room temperature for 3 h, it was quenched with Amberlyst 15 hydrogen resin, and concentrated. The obtained residue was dissolved in CH_3CN (80 mL), benzaldehyde dimethyl acetal (2.16 mL, 14.39 mmol) and *p*-TSA (182 mg, 0.959 mmol) were added. After the reaction mixture was stirred at room temperature for overnight, it was quenched with Et_3N and concentrated. The residue was subjected to flush silica gel column chromatography (1:2, EtOAc–hexane) to give **9iii** (3.07 g, 80% for 2 steps) as a white solid.

For α anomer: R_f 0.55 (1:2, EtOAc–hexane); ^1H NMR (600 MHz, CDCl_3) δ : 7.55 – 7.48 (m, 2H), 7.44 – 7.35 (m, 3H), 7.12 – 7.02 (m, 2H), 6.88 – 6.81 (m, 2H), 5.60 (s, 1H), 5.58 (d, J = 3.3 Hz, 1H), 4.39 (dd, J = 10.5, 3.7 Hz, 1H), 4.37 – 4.33 (m, 1H), 4.27 (dd, J = 12.7, 1.4 Hz, 1H), 4.06 (dd, J = 12.7, 1.7 Hz, 1H), 3.88 (d, J = 0.9 Hz, 1H), 3.78 (s, 3H), 3.70 (dd, J = 10.5, 3.3 Hz, 1H), 2.60 (s, 1H). ^{13}C NMR (150 MHz, CDCl_3) δ : 155.28, 150.52, 137.21, 129.40, 128.35, 126.20, 117.56, 114.72, 101.27, 98.14, 75.35, 69.14, 67.39, 63.36, 60.49, 55.65. HR ESI-TOF MS (m/z): calcd for $\text{C}_{31}\text{H}_{35}\text{N}_3\text{O}_7\text{SNa} [\text{M} + \text{Na}]^+$, 616.2093; found, 616.2088.

For β anomer: R_f 0.20 (1:2, EtOAc–hexane); ^1H NMR (600 MHz, CDCl_3) δ : 7.52 (dd, J = 6.6, 3.0 Hz, 2H), 7.42 – 7.34 (m, 3H), 7.11 – 7.04 (m, 2H), 6.87 – 6.79 (m, 2H), 5.57 (s, 1H), 4.74 (d, J = 8.1 Hz, 1H), 4.36 (dd, J = 12.5, 1.4 Hz, 1H), 4.22 – 4.17 (m, 1H), 4.08 (dd, J = 12.6, 1.7 Hz, 1H), 3.87 (dd, J = 10.2, 8.1 Hz, 1H), 3.78 (s, 3H), 3.61 (dd, J = 10.2, 3.8 Hz, 1H), 3.50 (d, J = 1.1 Hz, 1H), 2.66 (s, 1H). ^{13}C NMR (150 MHz, CDCl_3) δ : 155.68, 151.06, 137.24, 129.41, 128.31, 126.41, 118.90, 114.53, 101.79, 101.44, 74.33, 71.34, 68.92, 66.63, 63.74, 55.63. HR ESI-TOF MS (m/z): calcd for $\text{C}_{20}\text{H}_{21}\text{N}_3\text{O}_6\text{Na} [\text{M} + \text{Na}]^+$, 422.1323; found, 422.1313.

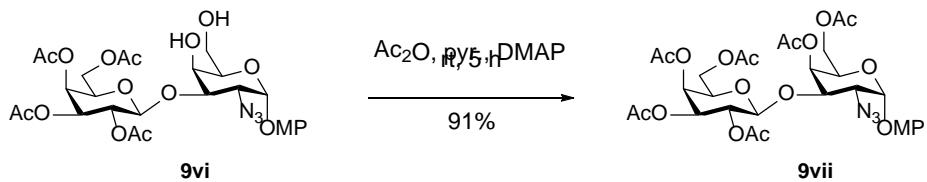


To a stirred mixture of donor **9iv** (4.06 g, 8.25 mmol), acceptor **9iii** (2.2 g, 5.5 mmol), and freshly activated MS 4Å (5 g), in anhydrous CH₂Cl₂ (100 mL) was added TMSOTf (99.4 µL, 0.55 mmol) under N₂ protection at 0 °C. After the reaction mixture was stirred for another 30 min, it was neutralized with Et₃N, filtered, and concentrated. The residue was subjected to silica gel column chromatography with EtOAc and hexanes (1:2) as the eluent to afford the **9v** (3.53 g, 88%) as a syrup. *R*_f 0.45 (1:1, EtOAc–hexane); ¹H NMR (600 MHz, CDCl₃) δ: 7.59 – 7.52 (m, 2H), 7.42 – 7.31 (m, 3H), 7.10 – 7.01 (m, 2H), 6.88 – 6.80 (m, 2H), 5.60 (d, *J* = 3.3 Hz, 1H), 5.58 (s, 1H), 5.44 – 5.40 (m, 1H), 5.32 (dd, *J* = 10.4, 7.9 Hz, 1H), 5.06 (dd, *J* = 10.4, 3.5 Hz, 1H), 4.86 (d, *J* = 7.9 Hz, 1H), 4.46 (d, *J* = 3.1 Hz, 1H), 4.32 (dd, *J* = 10.8, 3.3 Hz, 1H), 4.24 (ddd, *J* = 17.8, 11.9, 3.9 Hz, 2H), 4.14 (dd, *J* = 10.0, 5.3 Hz, 1H), 4.04 (dd, *J* = 12.5, 1.2 Hz, 1H), 3.97 (ddd, *J* = 14.4, 9.1, 5.0 Hz, 2H), 3.84 (s, 1H), 3.77 (s, 3H), 2.17 (s, 3H), 2.07 (s, 3H), 2.04 (s, 3H), 1.99 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ: 170.27, 170.24, 170.12, 169.41, 155.33, 150.46, 137.54, 128.89, 128.13, 126.09, 117.61, 114.72, 102.52, 100.60, 98.32, 75.79, 75.67, 71.03, 70.91, 69.07, 68.68, 66.97, 63.67, 61.40, 58.74, 55.63, 20.71, 20.70, 20.68, 20.54. HR ESI-TOF MS (m/z): calcd for C₃₄H₃₉N₃O₁₅Na [M + Na]⁺, 752.2273; found, 752.2278.

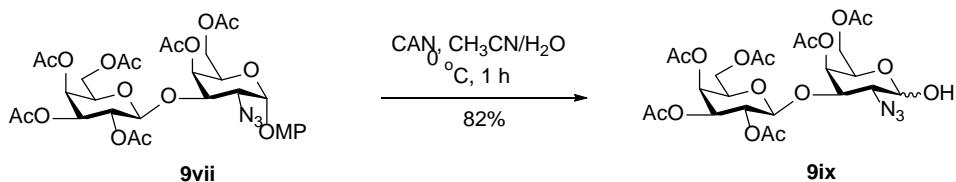


Compound **9v** (3.3 g, 4.52 mmol) was dissolved in mixed solvent of acetic acid/H₂O (4:1, 50 mL). After the reaction mixture was stirred at 80 °C for 5 h, it was concentrated and the residue was subjected to silica gel column chromatography with acetone and hexanes (1:2) as the eluent to afford the **9vi** (2.03 g, 70%) as a syrup. R_f 0.50 (1:1, acetone–hexane); ¹H NMR (600 MHz, CDCl₃) δ: 7.06 – 6.98 (m, 2H), 6.85 – 6.78 (m, 2H), 5.46 (d, J = 3.5 Hz, 1H), 5.40 (d, J = 2.7 Hz, 1H), 5.29 (dd, J = 10.5, 8.0 Hz, 1H), 5.05 (dd, J = 10.5, 3.4 Hz, 1H), 4.78 (d, J = 8.0 Hz, 1H), 4.27 (d, J = 1.7 Hz, 1H), 4.22 (dd, J = 10.5, 3.1 Hz, 1H), 4.17 (dd, J = 11.5, 7.3 Hz, 1H), 4.10 (dd, J = 11.5, 5.5 Hz, 1H), 4.03 (t, J = 5.6 Hz, 1H), 3.98 (t, J = 6.6 Hz, 1H), 3.88 (dd, J = 11.7, 5.6 Hz, 1H), 3.80 (dd, J = 11.7, 5.1 Hz, 1H), 3.77 – 3.73 (m, 4H), 2.15 (s, 3H), 2.09 (s, 3H), 2.04 (s, 3H), 1.98 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ: 170.44, 170.11, 170.04, 169.63, 155.50, 150.40, 118.28, 114.72, 101.98, 98.40, 78.16, 71.29, 70.66, 69.93, 69.23, 68.39, 66.99, 62.44, 61.55, 58.42, 55.61,

20.63, 20.58, 20.56, 20.50. HR ESI-TOF MS (m/z): calcd for $C_{27}H_{35}N_3O_{15}Na$ [M + Na]⁺, 664.1960; found, 664.1969.

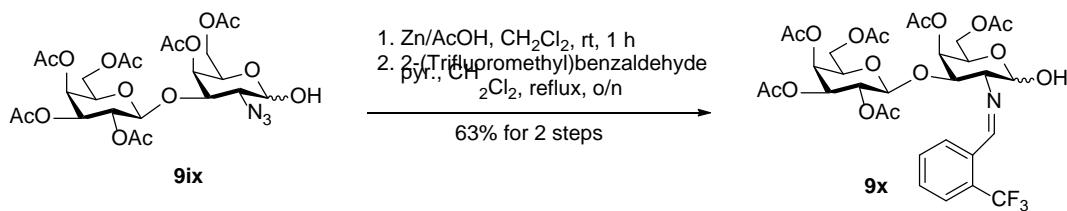


To a solution of **9vi** (1.8 g, 2.80 mmol) in pyridine (30 mL) was added acetic anhydride (1.06 mL, 11.21 mmol) and DMAP (34.2 mg, 0.28 mmol). After the reaction mixture was stirred at room temperature for 5 h, it was concentrated and the residue was subjected to flush silica gel column chromatography (1:2, EtOAc–hexane) to give **9vii** (1.85 g, 91%) as a serup. R_f 0.50 (1:1, EtOAc–hexane); ¹H NMR (600 MHz, CDCl₃) δ : 7.04 – 6.97 (m, 2H), 6.83 – 6.78 (m, 2H), 5.51 (d, J = 3.2 Hz, 1H), 5.44 (d, J = 3.5 Hz, 1H), 5.36 – 5.34 (m, 1H), 5.19 (dd, J = 10.5, 7.8 Hz, 1H), 5.01 (dd, J = 10.5, 3.4 Hz, 1H), 4.75 (d, J = 7.8 Hz, 1H), 4.30 – 4.26 (m, 2H), 4.15 (dt, J = 11.0, 5.2 Hz, 2H), 4.08 (dd, J = 7.0, 4.2 Hz, 1H), 3.99 – 3.90 (m, 2H), 3.78 – 3.72 (m, 4H), 2.13 (s, 3H), 2.12 (s, 3H), 2.07 (s, 3H), 2.03 (s, 3H), 1.96 (s, 3H), 1.93 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ : 170.36, 170.33, 170.19, 170.02, 169.64, 169.47, 155.62, 150.20, 149.05, 136.57, 118.43, 114.61, 101.56, 98.02, 74.73, 70.84, 70.77, 69.32, 68.77, 68.13, 66.78, 62.54, 61.06, 59.40, 55.59, 20.69, 20.62, 20.60, 20.59, 20.50. HR ESI-TOF MS (m/z): calcd for $C_{31}H_{39}N_3O_{17}Na$ [M + Na]⁺, 748.2172; found, 748.2162.

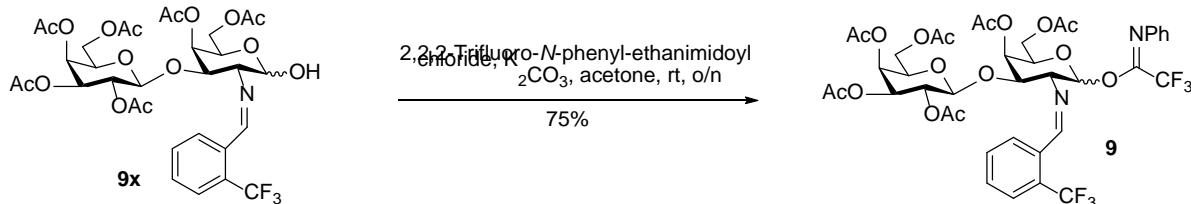


To a solution of **9vii** (2.5 g, 3.44 mmol) in acetonitrile/H₂O (1:1, 60 mL) was added cerium ammonium nitrate (5.66 g, 10.33 mmol) at 0 °C. After the reaction mixture was stirred at the same temperature for 1 h, it was diluted with ethyl acetate and washed with aqueous NaHCO₃ and brine, dried over anhydrous Na₂SO₄, and concentrated. The residue was subjected to flush silica gel column chromatography (1.5:1, EtOAc–hexane) to give **9ix** (1.75 g, 82%) as a α/β mixture. R_f 0.30 (1.5:1, EtOAc–hexane); ¹H NMR (600 MHz, CDCl₃) δ : 5.48 (d, J = 2.9 Hz, 1.1H), 5.40 (d, J = 3.4 Hz, 1.1H), 5.37 – 5.32 (m, 2.3H), 5.16 (ddd, J = 15.2, 10.5, 7.8 Hz, 1.8H), 5.00 (dd, J = 10.4, 3.4 Hz, 1.7H), 4.71 (dd, J = 12.0, 7.8 Hz, 1.8H), 4.60 (d, J = 6.9 Hz, 1H), 4.37 – 4.32 (m, 1.1H), 4.22 – 4.05 (m, 7.4H), 3.98 (ddd, J = 20.8, 11.7, 7.4 Hz, 2.1H), 3.89 (dt, J = 16.9, 6.5 Hz, 1.9H),

3.81 – 3.76 (m, 1.3H), 3.73 (dd, J = 10.6, 3.5 Hz, 1.2H), 3.55 (p, J = 10.3 Hz, 1.7H), 3.24 (s, 1.3H), 2.15 (d, J = 0.4 Hz, 4.6H), 2.13 (d, J = 3.2 Hz, 4.4H), 2.08 (s, 2.2H), 2.06 (d, J = 0.4 Hz, 6.7H), 2.05 (d, J = 1.3 Hz, 4.4H), 2.04 (s, 1.9H), 1.97 (d, J = 2.3 Hz, 4.5H). ^{13}C NMR (150 MHz, CDCl_3) δ : 101.45, 101.42, 96.35, 92.32, 77.35, 74.40, 71.70, 70.84, 70.81, 70.77, 70.63, 69.52, 68.81, 68.77, 68.12, 67.47, 66.80, 66.75, 64.49, 62.76, 62.58, 61.06, 60.97, 60.39, 60.16, 20.77, 20.73, 20.66, 20.62, 20.60, 20.52. HR ESI-TOF MS (m/z): calcd for $\text{C}_{24}\text{H}_{33}\text{N}_3\text{O}_{16}\text{Na} [\text{M} + \text{Na}]^+$, 642.1753; found, 642.1759.



To a solution of **9ix** (1.7 g, 2.74 mmol) in CH_2Cl_2 (30 mL) was added zinc powder (5.35 g, 82.25 mmol) and acetic acid (4.7 mL, 82.25 mmol). After the reaction mixture was stirred at room temperature for 1 h, it was filtered and concentrated. The obtained residue was dissolved in pyridine/ CH_2Cl_2 (1:10, 30 mL), 2-(Trifluoromethyl) benzaldehyde (433 μL , 3.29 mmol) was added. After the reaction mixture was stirred under reflux for overnight, it was concentrated. The residue was subjected to flush silica gel column chromatography (2:1, EtOAc–hexane) to give **9x** (1.3 g, 63% for 2 steps) as a α/β mixture. R_f 0.30 (2:1, EtOAc–hexane); ^1H NMR (600 MHz, CDCl_3) δ : 8.83 (d, J = 16.1 Hz, 2.2H), 8.32 (dd, J = 21.5, 7.4 Hz, 2.2H), 7.86 (d, J = 7.2 Hz, 2.3H), 7.77 (t, J = 7.3 Hz, 1H), 7.75 – 7.65 (m, 3.5H), 5.66 (s, 1H), 5.58 (s, 1.6H), 5.44 (s, 2.2H), 5.34 (s, 1H), 5.17 (dd, J = 12.3, 5.9 Hz, 2.2H), 5.05 – 4.96 (m, 3.6H), 4.73 (d, J = 7.4 Hz, 2.2H), 4.67 (s, 1H), 4.58 (d, J = 10.3 Hz, 1H), 4.44 – 4.37 (m, 2.5H), 4.34 – 4.13 (m, 11H), 4.09 (d, J = 3.6 Hz, 1.7H), 4.04 – 3.96 (m, 3H), 3.80 (s, 1H), 3.69 (t, J = 8.3 Hz, 1.6H), 2.28 (d, J = 13.7 Hz, 13H), 2.25 – 2.16 (m, 15H), 2.03 (d, J = 1.3 Hz, 5.4H), 1.62 (d, J = 11.0 Hz, 5.4H). ^{13}C NMR (150 MHz, CDCl_3) δ : 170.66, 170.39, 170.38, 170.28, 170.06, 170.02, 168.77, 161.72, 161.48, 133.49, 132.11, 131.99, 131.00, 130.75, 129.27, 128.46, 128.23, 125.84, 124.80, 122.98, 100.68, 95.41, 94.16, 77.43, 74.07, 73.72, 71.84, 70.83, 70.77, 70.63, 70.61, 70.25, 68.93, 68.85, 68.69, 67.95, 67.83, 66.72, 66.68, 63.03, 63.01, 60.98, 60.94, 60.37, 20.98, 20.81, 20.77, 20.76, 20.72, 20.64, 20.59, 20.43, 19.87. HR ESI-TOF MS (m/z): calcd for $\text{C}_{32}\text{H}_{38}\text{F}_3\text{NO}_{16}\text{Na} [\text{M} + \text{Na}]^+$, 772.2035; found, 772.2041.

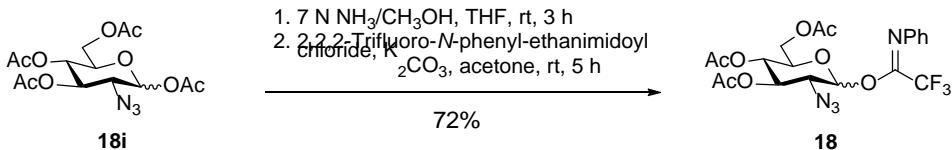


To a solution of **9x** (1.04 g, 1.39 mmol) in anhydrous acetone (12 mL) was added 2,2,2-Trifluoro-*N*-phenyl-ethanimidoyl chloride (443 μ L, 2.77 mmol) and K_2CO_3 (382 mg, 2.77 mmol) under N_2 protection. After overnight stirring at room temperature, the mixture was filtered and concentrated in vacuum, and the residue was purified by Et_3N -neutralized silica gel column with $EtOAc$ and hexanes (1:1) as the eluent to afford compound **9** (0.96 g, 75%) as a yellow solid.

For α anomer: R_f 0.65 (1.5:1, $EtOAc$ –hexane); 1H NMR (400 MHz, $CDCl_3$) δ : 8.69 (s, 1H), 8.26 (d, J = 7.7 Hz, 1H), 7.74 (d, J = 7.7 Hz, 1H), 7.67 (t, J = 7.5 Hz, 1H), 7.60 (t, J = 7.4 Hz, 1H), 7.14 (t, J = 7.7 Hz, 2H), 7.02 (t, J = 7.4 Hz, 1H), 6.59 (d, J = 6.9 Hz, 2H), 6.25 (s, 1H), 5.61 (d, J = 2.3 Hz, 1H), 5.32 (d, J = 3.3 Hz, 1H), 5.04 (dd, J = 10.4, 7.9 Hz, 1H), 4.87 (dd, J = 10.4, 3.4 Hz, 1H), 4.61 (d, J = 7.8 Hz, 1H), 4.52 – 4.40 (m, 2H), 4.28 (dd, J = 11.7, 4.7 Hz, 1H), 4.19 – 3.98 (m, 4H), 3.89 (t, J = 6.6 Hz, 1H), 2.16 (s, 3H), 2.13 (s, 3H), 2.10 (s, 3H), 2.06 (s, 3H), 1.89 (s, 3H), 1.52 (s, 3H). ^{13}C NMR (100 MHz, $CDCl_3$) δ : 170.47, 170.40, 170.23, 170.01, 169.80, 168.79, 161.70, 133.39, 132.07, 131.02, 128.96, 128.61, 128.35, 125.83, 124.23, 119.76, 119.27, 100.60, 70.88, 70.83, 70.37, 68.98, 68.51, 68.04, 66.80, 62.62, 61.14, 20.72, 20.59, 20.58, 20.45, 19.90. HR ESI-TOF MS (m/z): calcd for $C_{40}H_{42}F_6N_2O_{16}Na$ [M + Na] $^+$, 943.2331; found, 943.2338.

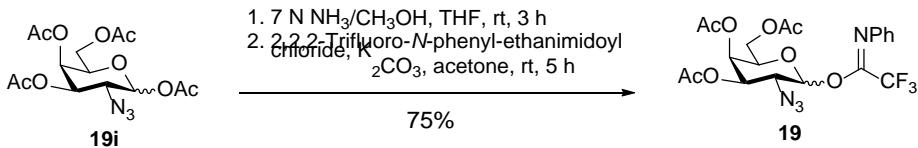
For β anomer: R_f 0.55 (1.5:1, $EtOAc$ –hexane); 1H NMR (400 MHz, $CDCl_3$) δ : 8.71 (d, J = 1.4 Hz, 1H), 8.17 (d, J = 7.7 Hz, 1H), 7.64 (ddd, J = 28.0, 21.1, 7.6 Hz, 3H), 7.23 (dd, J = 14.8, 7.0 Hz, 2H), 7.05 (t, J = 7.4 Hz, 1H), 6.70 (d, J = 7.7 Hz, 2H), 5.83 (s, 1H), 5.46 (d, J = 2.9 Hz, 1H), 5.30 (d, J = 3.2 Hz, 1H), 5.02 (dd, J = 10.4, 7.9 Hz, 1H), 4.83 (dd, J = 10.5, 3.4 Hz, 1H), 4.59 (d, J = 7.8 Hz, 1H), 4.28 – 3.93 (m, 6H), 3.85 (t, J = 6.7 Hz, 2H), 2.16 (s, 3H), 2.11 (s, 3H), 2.03 (s, 3H), 2.01 (s, 3H), 1.87 (s, 3H), 1.50 (s, 3H). ^{13}C NMR (100 MHz, $CDCl_3$) δ : 170.46, 170.32, 170.21, 169.99, 169.88, 168.73, 162.52, 143.25, 133.28, 132.06, 131.03, 129.73, 129.42, 128.65, 128.22, 125.97, 125.91, 125.19, 124.29, 122.46, 119.05, 100.78, 95.27, 77.16, 72.64, 71.48, 70.74, 70.70, 68.82, 67.65, 66.70, 62.26, 61.03, 20.67, 20.63, 20.62, 20.56, 20.40, 19.86. HR ESI-TOF MS (m/z): calcd for $C_{40}H_{42}F_6N_2O_{16}Na$ [M + Na] $^+$, 943.2331; found, 943.2336.

4.3 Preparation of glycosyl donor **18**



To a solution of **18i** (1.3 g, 3.48 mmol) in THF (20 mL) was added 7 N $\text{NH}_3/\text{CH}_3\text{OH}$ (3 mL). After the reaction mixture was stirred at room temperature for 1 h, it was diluted with ethyl acetate and washed with 1 N HCl and brine. The organic layer was dried over Na_2SO_4 , filtered and concentrated. The obtained residue was dissolved in anhydrous acetone (20 mL), 2,2,2-Trifluoro-N-phenyl-ethanimidoyl chloride (1.11 mL, 6.96 mmol) and K_2CO_3 (960 mg, 6.96 mmol) were added under N_2 protection. After 5 h stirring at room temperature, the mixture was filtered and concentrated in vacuum, and the residue was purified by Et_3N -neutralized silica gel column with EtOAc and hexanes (1:4) as the eluent to afford **18** (1.26 g, 72%) as a α/β mixture. R_f 0.50 (1:2, EtOAc–hexane); ^1H NMR (600 MHz, CDCl_3) δ : 7.32 (td, $J = 7.9, 4.2$ Hz, 2.5H), 7.14 (dt, $J = 11.4, 5.7$ Hz, 1.3H), 6.84 (t, $J = 8.2$ Hz, 2.6H), 6.48 (s, 1H), 5.62 (s, 1H), 5.50 (t, $J = 9.9$ Hz, 0.6H), 5.14 (t, $J = 9.7$ Hz, 1H), 5.06 (s, 1H), 4.36 – 4.22 (m, 1H), 4.20 – 4.03 (m, 1.5H), 3.87 – 3.65 (m, 2.3H), 2.11 (s, 3H), 2.10 (s, 2H), 2.09 (s, 3H), 2.07 (s, 2H), 2.06 (s, 3H), 2.02 (s, 2H). ^{13}C NMR (150 MHz, CDCl_3) δ : 170.44, 170.39, 169.77, 169.56, 169.48, 142.79, 128.83, 124.70, 119.19, 119.09, 95.23, 92.95, 72.70, 72.49, 70.59, 70.03, 67.74, 62.78, 61.37, 60.44, 20.61, 20.59, 20.52, 20.50. HR ESI-TOF MS (m/z): calcd for $\text{C}_{20}\text{H}_{21}\text{F}_3\text{N}_4\text{O}_8\text{Na}$ [M + Na]⁺, 525.1204; found, 525.1213.

4.4 Preparation of glycosyl donor **19**



Compound **19** (678 mg, 75%) was prepared from **19i** (672 mg, 1.8 mmol) by the same procedure as the synthesis of **18**. R_f 0.55 (1:2, EtOAc–hexane); ^1H NMR (400 MHz, CDCl_3) of α/β anomers δ : 7.36 – 7.27 (m, 4.2H), 7.13 (t, $J = 7.5$ Hz, 2H), 6.84 (d, $J = 7.8$ Hz, 4H), 6.47 (s, 0.5H), 5.60 (s, 0.5H), 5.53 (s, 1H), 5.41 – 5.30 (m, 2H), 4.87 (d, $J = 8.9$ Hz, 1H), 4.31 (d, $J = 6.0$ Hz, 0.8H), 4.19 – 4.05 (m, 5H), 3.96 (dd, $J = 24.4, 10.4$ Hz, 3H), 2.18 (s, 3.4H), 2.16 (s, 2.7H), 2.08 (s, 2.7H), 2.07 (s, 3.4H), 2.05 (s, 2.7H), 2.00 (s, 3.4H). ^{13}C NMR (100 MHz, CDCl_3) δ : 170.21, 169.91, 169.58, 142.87, 128.83, 124.66, 119.23, 119.07, 95.47, 93.32, 71.75, 71.13, 69.10, 68.61, 66.83, 66.03,

61.20, 60.90, 59.86, 56.90, 20.58, 20.54, 20.52. HR ESI-TOF MS (m/z): calcd for C₂₀H₂₁F₃N₄O₈Na [M + Na]⁺, 525.1204; found, 525.1217.

5. Preparation of Thiol Nucleophiles

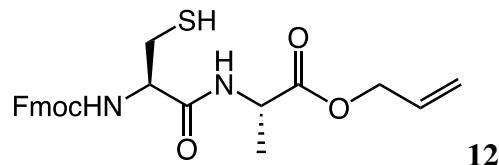
5.1 Preparation of cysteine-containing acceptors

General procedure for peptide coupling:

To a stirred solution of *N*-protected amino acid in dry DMF (0.5 M) was added HATU (1.1 eq), DIPEA (3.0 eq) followed by *C*-protected amino acid (1.05 eq) at room temperature under nitrogen atmosphere. The reaction mixture was stirred at same temperature for 3 to 6 h (TLC control). The reaction mixture was quenched with water and extracted with EtOAc (3×). The combined organic layer was dried over anhydrous Na₂SO₄ and evaporated to dryness. The obtained crude product was purified by flash chromatography.

General procedure for the removal of trityl protection:

The peptide was dissolved in dry dichloromethane (0.1 M) and purged with argon gas for few minutes, then was added TIPS (2.0 eq) followed TFA (10.0 eq) dropwise under argon atmosphere at room temperature. The reaction mixture was allowed to stir at same temperature for 1-2 hours (TLC control). The reaction mixture was evaporated, to the obtained crude gummy crude product was added diethyl ether (10 vol). The solids precipitated were filtered using sintered funnel and washed again with diethyl ether to get pure product (no column purification required).

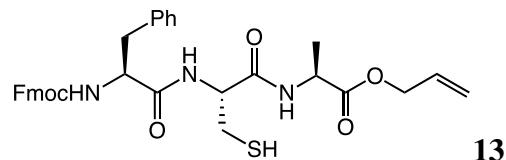


¹H NMR (CDCl₃, 400 MHz): δ 7.77 (d, *J* = 7.5 Hz, 2H), 7.59 (d, *J* = 7.3 Hz, 2H), 7.41 (t, *J* = 7.4 Hz, 2H), 7.32 (t, *J* = 7.4 Hz, 2H), 6.71 (d, *J* = 5.2 Hz, NH), 5.90 (ddt, *J* = 16.0, 10.6, 5.6 Hz, 1H), 5.72 (s, NH), 5.30 (dd, *J* = 26.5, 13.7 Hz, 2H), 4.70 – 4.53 (m, 3H), 4.53 – 4.32 (m, 3H), 4.23 (t, *J*

= 6.7 Hz, 1H), 3.12 – 2.98 (m, 1H), 2.74 (bs, 1H), 1.69 (t, J = 7.2 Hz, SH), 1.45 (d, J = 7.2 Hz, 3H).

^{13}C NMR (CDCl₃, 100 MHz): δ 172.1, 169.1, 143.6, 141.3, 141.30, 131.3, 127.8, 127.1, 125.0, 120.0, 120.0, 119.1, 67.2, 66.2, 56.0, 48.5, 47.1, 27.1, 18.2.

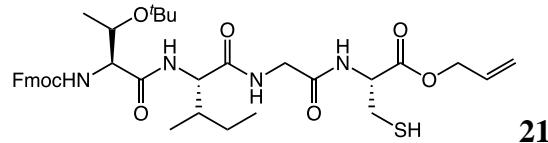
HRMS (ESI): calc. for C₂₄H₂₆N₂O₅SNa [M+Na]⁺: 477.1455; found: 477.1454.



^1H NMR (CDCl₃, 400 MHz): δ 7.74 (d, J = 7.5 Hz, 2H), 7.50 (t, J = 7.8 Hz, 2H), 7.38 (t, J = 7.4 Hz, 2H), 7.34 – 7.12 (m, 7H), 6.89 (d, J = 26.1 Hz, NH), 5.87 (ddt, J = 16.4, 10.9, 5.8 Hz, 1H), 5.42 – 5.19 (m, 2H, NH), 4.68 – 4.45 (m, 5H), 4.43 – 4.27 (m, 2H), 4.17 (t, J = 6.9 Hz, 1H), 3.21 – 2.95 (m, 3H), 2.73 – 2.61 (m, 1H), 1.63 (t, J = 7.2 Hz, SH), 1.38 (d, J = 6.6 Hz, 3H).

^{13}C NMR (CDCl₃, 100 MHz): δ 171.9, 170.9, 168.7, 143.6, 141.3, 135.9, 131.4, 129.2, 128.9, 128.8, 127.8, 127.3, 127.1, 124.9, 120.0, 118.8, 67.2, 66.1, 56.3, 54.2, 48.4, 47.0, 38.1, 26.6, 17.9.

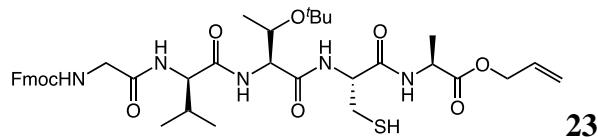
HRMS (ESI): calc. for C₃₃H₃₅N₃O₆SNa [M+Na]⁺: 624.2139; found: 624.2134.



^1H NMR (CDCl₃, 400 MHz): δ 7.76 (d, J = 7.5 Hz, 2H), 7.61 (d, J = 7.4 Hz, 2H), 7.54 (d, J = 6.7 Hz, NH), 7.39 (t, J = 7.5 Hz, 2H), 7.30 (t, J = 7.4 Hz, 2H), 6.92 (bd, NH), 6.12 (m, NH), 5.94 – 5.80 (m, 1H), 5.36 – 5.18 (m, 2H), 4.87 (td, J = 12.5, 4.9 Hz, 1H), 4.64 (d, J = 4.9 Hz, 2H), 4.44 – 4.34 (m, 2H), 4.33 – 4.06 (m, 5H), 3.88 (ddd, J = 34.4, 16.8, 5.3 Hz, 1H), 3.08 – 2.98 (m, 2H), 2.05 – 1.93 (m, 1H), 1.84 – 1.69 (m, 1H), 1.65 – 1.50 (m, 2H), 1.33 (s, 9H), 1.21 – 1.04 (m, 4H), 1.04 – 0.86 (m, 6H).

^{13}C NMR (CDCl₃, 100 MHz): δ 171.7, 170.4, 170.1, 169.5, 156.1, 143.8, 143.62, 141.3, 131.3, 131.3, 127.7, 127.0, 125.1, 120.1, 119.2, 76.0, 67.2, 66.9, 66.4, 59.3, 58.4, 54.1, 47.2, 43.2, 36.3, 28.2, 26.6, 25.04, 16.8, 15.8, 11.4.

HRMS (ESI): calc. for C₃₇H₅₁N₄O₈S [M+H]⁺: 711.3432; found: 711.3435.

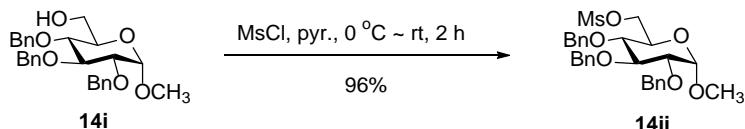


¹H NMR (CDCl₃, 400 MHz): δ 8.22 (s, NH), 7.94 (s, NH), 7.74 (d, *J* = 7.4 Hz, 2H), 7.57 (d, *J* = 7.2 Hz, 2H), 7.37 (t, *J* = 7.3 Hz, 2H), 7.27 (d, *J* = 9.6 Hz, 2H), 7.05 (s, NH), 6.12 (s, NH), 5.90 – 5.73 (m, 1H), 5.21 (dd, *J* = 26.3, 13.8 Hz, 2H), 5.06 (s, 1H), 4.83 – 4.73 (m, 1H), 4.65 – 4.51 (m, 3H), 4.45 – 4.15 (m, 3H), 4.08 – 3.87 (m, 3H), 3.09 – 2.98 (m, 1H), 2.87 – 2.72 (m, 1H), 2.02 (dd, *J* = 12.9, 6.6 Hz, 1H), 1.65 (t, *J* = 7.4 Hz, SH), 1.37 (d, *J* = 7.1 Hz, 3H), 1.25 (s, 9H), 1.05 (d, *J* = 5.5 Hz, 3H), 0.93 (d, *J* = 6.3 Hz, 6H).

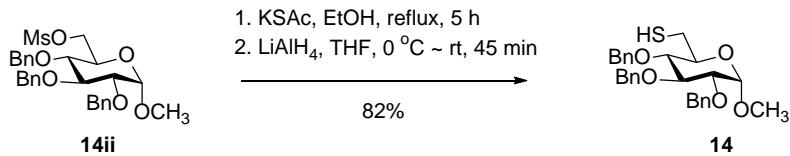
¹³C NMR (CDCl₃, 100 MHz): δ 172.2, 170.8, 169.6, 169.3, 168.9, 156.7, 143.8, 141.2, 131.5, 127.7, 127.0, 125.1, 119.9, 118.7, 75.4, 67.3, 66.1, 61.5, 58.1, 57.6, 53.7, 48.2, 47.0, 44.4, 32.1, 29.3, 28.3, 19.0, 18.5, 18.1.

HRMS (ESI): calc. for C₃₉H₅₄N₅O₉S [M+H]⁺: 768.5.

5.2 Preparation of glucosyl acceptor **14**

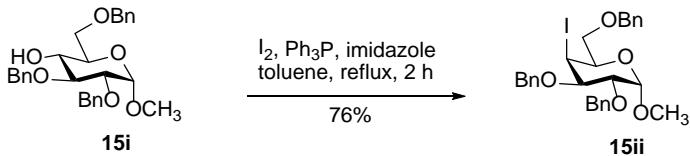


To a solution of **14i** (2.7 g, 5.82 mmol) in 30 mL of anhydrous pyridine was added MsCl (901 μL, 11.64 mmol) under N₂ protection at 0 °C. After 2 hours of stirring at the same temperature, the reaction mixture was quenched with methanol and concentrated. The residue was subjected to silica gel column chromatography with EtOAc and hexanes (1:3) as the eluent to afford compound **26** (3.03 g, 96%) as a syrup. *R*_f 0.45 (1:2, EtOAc–hexane); ¹H NMR (600 MHz, CDCl₃) δ: 7.43 – 7.21 (m, 1H), 5.01 (d, *J* = 10.9 Hz, 1H), 4.91 (d, *J* = 10.8 Hz, 1H), 4.81 (dd, *J* = 21.3, 11.5 Hz, 1H), 4.70 – 4.54 (m, 1H), 4.36 (qd, *J* = 11.0, 3.2 Hz, 1H), 4.02 (t, *J* = 9.2 Hz, 1H), 3.84 (ddd, *J* = 10.1, 4.1, 2.2 Hz, 1H), 3.56 – 3.45 (m, 1H), 3.38 (s, 1H), 2.97 (s, 1H). ¹³C NMR (150 MHz, CDCl₃) δ: 138.48, 137.92, 137.73, 128.51, 128.50, 128.43, 128.07, 128.06, 128.00, 127.97, 127.89, 127.69, 98.16, 81.79, 79.76, 76.93, 75.75, 75.11, 73.46, 68.62, 68.39, 55.45, 37.51. HR ESI-TOF MS (m/z): calcd for C₂₉H₃₄SO₈Na [M + Na]⁺, 565.1867; found, 565.1878.



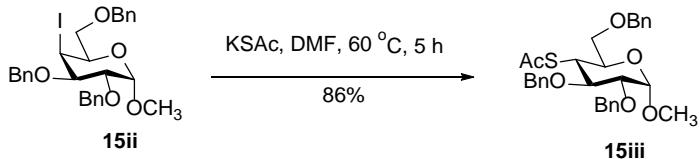
To a solution of **14ii** (3.3 g, 6.09 mmol) in ethanol (60 mL) was added KSAc (2.08 g, 18.27 mmol). After it was stirred under reflux for 5 h, it was diluted with ethyl acetate and washed with aqueous NaHCO₃ and brine. The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was subjected to flush silica gel column chromatography (1:6, EtOAc–hexane) to give an intermediate. To the obtained syrup in THF (60 mL) was added LiAlH₄ (1.85 g, 48.7 mmol) at 0 °C. After it was stirred at rt for 45 min, it was quenched with addition of ethyl acetate. The mixture was diluted with CH₂Cl₂ and washed with 1 N HCl and brine. The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was subjected to flush silica gel column chromatography (1:8, EtOAc–hexane) to give **14** (2.4 g, 82%) as a syrup. *R*_f 0.65 (1:4, EtOAc–hexane); ¹H NMR (600 MHz, CDCl₃) δ: 7.44 – 7.16 (m, 1H), 5.01 (d, *J* = 10.8 Hz, 1H), 4.92 (d, *J* = 11.1 Hz, 1H), 4.86 – 4.77 (m, 1H), 4.71 – 4.56 (m, 1H), 4.00 (t, *J* = 9.2 Hz, 1H), 3.73 (ddd, *J* = 9.7, 7.1, 2.6 Hz, 1H), 3.53 (dd, *J* = 9.6, 3.6 Hz, 1H), 3.45 (t, *J* = 9.3 Hz, 1H), 3.41 (s, 1H), 2.87 (ddd, *J* = 13.8, 8.7, 2.7 Hz, 1H), 2.65 – 2.57 (m, 1H), 1.52 (t, *J* = 8.3 Hz, 1H). ¹³C NMR (150 MHz, CDCl₃) δ: 138.64, 138.08, 128.47, 128.46, 128.40, 128.08, 127.96, 127.93, 127.86, 127.63, 97.93, 81.99, 80.13, 79.67, 75.72, 75.07, 73.37, 70.69, 55.21, 26.39. HR ESI-TOF MS (m/z): calcd for C₂₈H₃₂SO₅Na [M + Na]⁺, 503.1863; found, 503.1859.

5.3 Preparation of glucosyl acceptor **15**

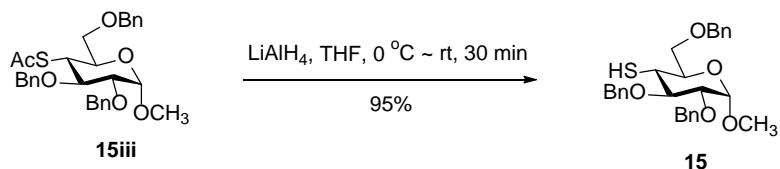


To a solution of **15i** (1.6 g, 3.45 mmol) in toluene (60 mL) were added I₂ (1.75 g, 6.9 mmol), imidazole (703 mg, 10.35 mmol) and Ph₃P (2.71 g, 10.35 mmol). After it was stirred under reflux for 2 h, it was filtered and concentrated. The residue was subjected to flush silica gel column chromatography (1:10, EtOAc–hexane) to give **15ii** (3.34 g, 76%) as a syrup. *R*_f 0.70 (1:3, EtOAc–hexane); ¹H NMR (400 MHz, CDCl₃) δ: 7.54 – 7.12 (m, 1H), 4.86 (d, *J* = 12.0 Hz, 1H), 4.75 (d, *J* = 11.6 Hz, 1H), 4.71 – 4.52 (m, 1H), 3.85 (dd, *J* = 9.5, 3.8 Hz, 1H), 3.64 (dd, *J* = 9.6, 6.0 Hz, 1H),

3.52 (dd, $J = 9.6, 6.4$ Hz, 1H), 3.39 (s, 1H), 3.31 (t, $J = 6.1$ Hz, 1H), 3.21 (dd, $J = 9.5, 4.0$ Hz, 1H). ^{13}C NMR (100 MHz, CDCl_3) δ : 138.34, 137.93, 137.80, 128.43, 128.39, 128.35, 128.12, 127.81, 127.78, 127.75, 127.68, 99.04, 77.84, 75.71, 74.18, 73.96, 73.74, 71.24, 67.08, 55.34, 41.23. HR ESI-TOF MS (m/z): calcd for $\text{C}_{28}\text{H}_{31}\text{IO}_5\text{Na} [\text{M} + \text{Na}]^+$, 597.1108; found, 597.1106.



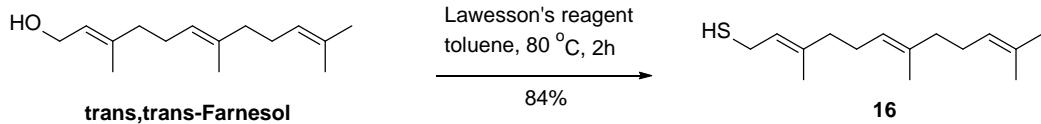
To a solution of **15ii** (1.5 g, 2.61 mmol) in DMF (30 mL) was added KSAc (1.19 g, 10.45 mmol). After it was stirred at 60 °C for 5 h, it was diluted with ethyl acetate and washed with H₂O and brine. The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was subjected to flush silica gel column chromatography (1:6, EtOAc–hexane) to give **15iii** (1.17 g, 86%) as a syrup. *R*_f 0.40 (1:4, EtOAc–hexane); ¹H NMR (600 MHz, CDCl₃) δ: 7.39 – 7.17 (m, 1H), 4.93 (d, *J* = 11.1 Hz, 1H), 4.79 (d, *J* = 12.0 Hz, 1H), 4.73 – 4.62 (m, 1H), 4.52 (s, 1H), 3.93 – 3.88 (m, 1H), 3.84 (dd, *J* = 10.6, 9.5 Hz, 1H), 3.68 (t, *J* = 11.1 Hz, 1H), 3.65 – 3.58 (m, 1H), 3.41 (s, 1H), 2.24 (s, 1H). ¹³C NMR (150 MHz, CDCl₃) δ: 193.42, 138.63, 138.07, 138.03, 128.44, 128.23, 128.19, 128.10, 127.91, 127.71, 127.63, 127.50, 127.42, 98.33, 97.44, 80.93, 78.34, 75.93, 73.45, 73.35, 69.87, 69.82, 55.39, 45.80, 30.61. HR ESI-TOF MS (m/z): calcd for C₃₀H₃₄SO₆Na [M + Na]⁺, 545.1968; found, 545.1963.



To a solution of **15iii** (590 mg, 1.13 mmol) in THF (20 mL) was added LiAlH₄ (344 mg, 9.04 mmol) at 0 °C. After it was stirred at rt for 30 min, it was quenched with addition of ethyl acetate. The mixture was diluted with CH₂Cl₂ and washed with 1 N HCl and brine. The organic layer was dried over Na₂SO₄, filtered and concentrated. The residue was subjected to flush silica gel column chromatography (1:7, EtOAc–hexane) to give **15** (515 mg, 95%) as a syrup. *R*_f 0.45 (1:4, EtOAc–hexane); ¹H NMR (600 MHz, CDCl₃) δ: 7.54 – 7.21 (m, 1H), 4.99 (d, *J* = 10.6 Hz, 1H), 4.82 (dd, *J* = 24.7, 11.3 Hz, 1H), 4.72 – 4.61 (m, 1H), 4.52 (d, *J* = 12.1 Hz, 1H), 3.80 (dd, *J* = 10.7, 3.9 Hz, 1H), 3.72 (ddd, *J* = 19.2, 12.0, 5.9 Hz, 1H), 3.55 (dd, *J* = 9.3, 3.5 Hz, 1H), 3.41 (s, 1H), 3.09 (td, *J* = 10.5, 6.7 Hz, 1H), 1.67 (d, *J* = 6.7 Hz, 1H). ¹³C NMR (150 MHz, CDCl₃) δ: 138.41, 138.05,

137.95, 128.46, 128.40, 128.34, 128.19, 128.10, 127.94, 127.76, 127.73, 127.65, 98.53, 81.81, 80.64, 76.29, 73.56, 73.20, 72.30, 69.49, 55.39, 41.89. HR ESI-TOF MS (m/z): calcd for C₂₈H₃₂SO₅Na [M + Na]⁺, 503.1863; found, 503.1857.

5.4 Preparation of thiol-Farnesol acceptor **16**

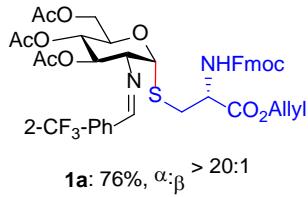


To a solution of **trans,trans-Farnesol** (0.4 mL, 1.58 mmol) in dry toluene (5 mL) was added Lawesson's reagent (0.42 g, 0.95 mmol). After it was stirred at 80 °C for 2 h, it was cooled down, filtered and concentrated. The residue was subjected to flush silica gel column chromatography (hexane) to give **16** (316 mg, 84%) as a colorless oil. *R*_f 0.25 (hexane); ¹H NMR (600 MHz, CDCl₃) δ: 5.34 (td, *J* = 7.8, 1.1 Hz, 1H), 5.12 – 5.06 (m, 2H), 3.16 (t, *J* = 7.4 Hz, 2H), 2.12 – 1.94 (m, 8H), 1.68 (s, 3H), 1.66 (s, 2H), 1.60 (s, 5H), 1.39 (t, *J* = 7.1 Hz, 1H). ¹³C NMR (150 MHz, CDCl₃) δ: 137.47, 135.27, 131.26, 124.31, 124.25, 124.09, 123.72, 123.61, 123.28, 39.67, 39.37, 26.70, 26.27, 25.67, 22.09, 17.66, 16.00, 15.75. HR ESI-TOF MS (m/z): calcd for C₁₅H₂₆SNa [M + Na]⁺, 261.1647; found, 261.1648.

6. Scope of Glycosylation

General Procedure for Triflic Acid-Catalyzed Thiol Glycosylation:

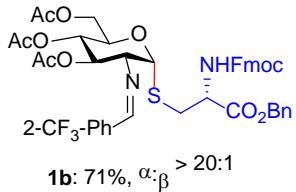
A 10 mL Schlenk flask was charged with *N*-phenyl trifluoroacetimidate glycosyl donor **1, 6 – 9, 18, 19** (0.2 mmol, 1 – 2 equiv.), thiol acceptor **10–17** (0.1 mmol, 1 equiv.) and dichloromethane (1 mL). The resulting solution was stirred at room temperature for 2 min under a nitrogen atmosphere before the TfOH (5 mol%) was added. After 1 h of stirring at room temperature, the reaction was quenched with 1 drop of Et₃N and concentrated. A crude ¹H NMR was taken to determine the (α/β) ratio. Further purification by silica gel column chromatography (ethyl acetate/hexane: 1/4 → 1/2) was performed to give the desired product.



¹H NMR (CDCl₃, 600 MHz): δ 8.61 (d, *J* = 2.0 Hz, 1H), 8.20 (d, *J* = 7.7 Hz, 1H), 7.76 (d, *J* = 7.6 Hz, 2H), 7.68 (d, *J* = 7.6 Hz, 1H), 7.64 – 7.51 (m, 4H), 7.40 (dd, *J* = 14.0, 7.0 Hz, 2H), 7.31 (dd, *J* = 15.2, 7.6 Hz, 2H), 6.01 (d, *J* = 8.6 Hz, 1H), 5.87 (ddd, *J* = 16.5, 11.2, 5.9 Hz, 1H), 5.49 (t, *J* = 9.7 Hz, 1H), 5.25 (ddd, *J* = 17.4, 11.5, 9.1 Hz, 3H), 5.10 (dd, *J* = 10.1, 9.6 Hz, 1H), 4.78 – 4.70 (m, 1H), 4.64 (d, *J* = 5.8 Hz, 2H), 4.52 (dd, *J* = 10.2, 3.2 Hz, 1H), 4.42 (p, *J* = 10.6 Hz, 2H), 4.25 (ddd, *J* = 12.5, 11.2, 6.2 Hz, 3H), 3.92 (dd, *J* = 10.0, 5.6 Hz, 1H), 3.23 (dd, *J* = 14.2, 5.4 Hz, 1H), 3.01 (dd, *J* = 14.3, 4.0 Hz, 1H), 2.05 (s, 3H), 2.03 (s, 3H), 1.87 (s, 3H).

¹³C NMR (CDCl₃, 150 MHz): δ 170.50, 169.87, 169.73, 155.69, 143.85, 143.62, 141.31, 137.07, 131.26, 128.51, 128.14, 127.79, 127.73, 127.71, 127.04, 125.07, 124.98, 119.99, 119.97, 119.34, 85.32, 76.11, 72.50, 71.86, 68.41, 67.07, 66.43, 62.07, 54.18, 47.05, 34.58, 20.73, 20.61, 20.58.

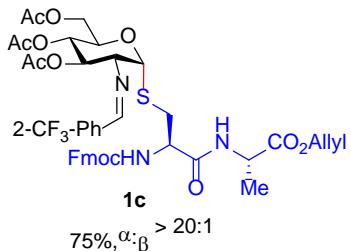
HRMS (ESI): calc. for C₄₁H₄₁F₃N₂O₁₁SNa [M + Na]⁺, 849.2275; found: 849.2278.



¹H NMR (CDCl₃, 600 MHz): δ 8.59 (d, *J* = 1.9 Hz, 1H), 8.18 (d, *J* = 7.7 Hz, 1H), 7.76 (d, *J* = 7.5 Hz, 2H), 7.68 (d, *J* = 7.6 Hz, 1H), 7.63 – 7.51 (m, 4H), 7.40 (dd, *J* = 13.5, 7.0 Hz, 2H), 7.36 – 7.27 (m, 8H), 6.05 (d, *J* = 8.6 Hz, 1H), 5.46 (t, *J* = 9.7 Hz, 1H), 5.16 (dd, *J* = 12.4, 3.4 Hz, 3H), 5.10 – 5.04 (m, 1H), 4.80 – 4.73 (m, 1H), 4.43 (ddt, *J* = 17.3, 13.7, 6.8 Hz, 3H), 4.28 – 4.16 (m, 3H), 3.85 (dd, *J* = 10.0, 5.6 Hz, 1H), 3.23 (dd, *J* = 14.3, 5.2 Hz, 1H), 2.99 (dd, *J* = 14.3, 4.0 Hz, 1H), 2.04 (s, 3H), 2.01 (s, 3H), 1.87 (s, 3H).

¹³C NMR (CDCl₃, 150 MHz): δ 170.55, 169.84, 169.56, 160.34, 155.73, 143.63, 141.27, 134.86, 132.28, 130.90, 129.09, 128.62, 128.59, 128.57, 127.71, 127.69, 127.06, 127.03, 125.49, 125.04, 119.97, 119.94, 85.35, 72.02, 71.29, 68.63, 67.65, 67.07, 62.18, 54.15, 53.77, 47.10, 33.77, 20.66, 20.59, 20.33.

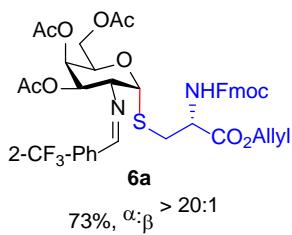
HRMS (ESI): calc. for C₄₅H₄₃F₃N₂O₁₁SNa [M + Na]⁺, 899.2432; found: 899.2433.



¹H NMR (CDCl₃, 600 MHz): δ 8.62 (d, *J* = 1.7 Hz, 1H), 8.18 (d, *J* = 7.6 Hz, 1H), 7.75 (d, *J* = 7.5 Hz, 2H), 7.66 (d, *J* = 7.6 Hz, 1H), 7.54 (ddd, *J* = 24.2, 15.3, 7.5 Hz, 4H), 7.38 (dd, *J* = 12.8, 7.2 Hz, 2H), 7.30 (ddd, *J* = 16.4, 7.6, 1.0 Hz, 2H), 6.84 (d, *J* = 5.7 Hz, 1H), 5.88 (ddd, *J* = 16.3, 11.0, 5.7 Hz, 1H), 5.71 (s, 1H), 5.52 (t, *J* = 9.7 Hz, 1H), 5.41 (s, 1H), 5.36 – 5.29 (m, 1H), 5.24 (dd, *J* = 10.5, 1.1 Hz, 1H), 5.11 (t, *J* = 9.8 Hz, 1H), 4.67 – 4.50 (m, 4H), 4.43 (dt, *J* = 17.0, 10.2 Hz, 3H), 4.25 (ddd, *J* = 24.6, 13.0, 5.8 Hz, 3H), 3.91 (dd, *J* = 9.9, 5.6 Hz, 1H), 3.14 – 3.06 (m, 1H), 2.93 – 2.85 (m, 1H), 2.06 (s, 3H), 2.02 (s, 3H), 1.86 (s, 3H), 1.38 (d, *J* = 7.2 Hz, 3H).

¹³C NMR (CDCl₃, 150 MHz): δ 171.96, 170.69, 169.87, 169.58, 160.45, 155.90, 143.69, 143.59, 141.29, 141.26, 133.05, 132.21, 131.44, 130.89, 129.09, 129.04, 127.75, 127.72, 127.10, 127.06, 125.50, 125.46, 125.00, 124.91, 119.98, 119.96, 118.81, 85.36, 71.99, 71.47, 68.82, 68.45, 67.19, 66.01, 62.16, 55.00, 48.39, 47.07, 33.38, 20.72, 20.65, 20.34, 18.00.

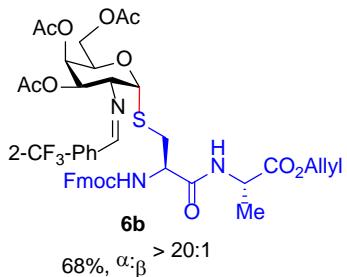
HRMS (ESI): calc. for $C_{44}H_{46}F_3N_3O_{12}SNa$ [M + Na]⁺, 920.2647; found: 920.2643.



¹H NMR (CDCl₃, 600 MHz): δ 8.63 (s, 1H), 8.19 (d, $J = 7.7$ Hz, 1H), 7.76 (d, $J = 7.6$ Hz, 2H), 7.68 (d, $J = 7.7$ Hz, 1H), 7.58 (ddd, $J = 33.7, 14.2, 7.5$ Hz, 4H), 7.40 (dd, $J = 13.5, 7.0$ Hz, 2H), 7.32 (dt, $J = 15.0, 7.6$ Hz, 2H), 5.96 (d, $J = 8.5$ Hz, 1H), 5.92 – 5.82 (m, 1H), 5.48 (s, 1H), 5.40 – 5.27 (m, 3H), 5.20 (d, $J = 10.5$ Hz, 1H), 4.72 – 4.60 (m, 3H), 4.40 (d, $J = 7.1$ Hz, 2H), 4.26 – 4.06 (m, 4H), 3.22 (dd, $J = 14.0, 5.4$ Hz, 1H), 3.00 (dd, $J = 14.0, 4.0$ Hz, 1H), 2.21 (s, 3H), 2.04 (s, 3H), 1.87 (s, 3H).

¹³C NMR (CDCl₃, 150 MHz): δ 170.46, 170.03, 169.62, 160.33, 155.67, 143.67, 141.29, 132.22, 131.30, 130.77, 129.03, 127.69, 127.04, 125.04, 119.96, 119.11, 85.43, 69.13, 67.62, 67.12, 66.79, 66.36, 62.18, 54.10, 47.07, 32.99, 29.67, 20.75, 20.61, 20.33.

HRMS (ESI): calc. for $C_{41}H_{41}F_3N_2O_{11}SNa$ [M + Na]⁺, 849.2275; found: 849.2273.

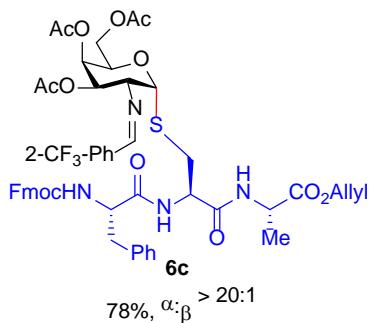


¹H NMR (CDCl₃, 600 MHz): δ 8.63 (d, $J = 1.9$ Hz, 1H), 8.18 (d, $J = 7.7$ Hz, 1H), 7.75 (d, $J = 7.5$ Hz, 2H), 7.66 (d, $J = 7.6$ Hz, 1H), 7.62 – 7.48 (m, 4H), 7.39 (dd, $J = 15.2, 7.6$ Hz, 2H), 7.36 – 7.28 (m, 2H), 6.97 (d, $J = 6.0$ Hz, 1H), 5.94 – 5.76 (m, 2H), 5.48 (d, $J = 2.4$ Hz, 2H), 5.31 (ddd, $J = 56.3, 24.8, 6.8$ Hz, 3H), 4.67 – 4.50 (m, 4H), 4.40 (dd, $J = 31.9, 5.9$ Hz, 3H), 4.21 (t, $J = 6.9$ Hz, 1H), 4.16 (d, $J = 6.0$ Hz, 2H), 4.10 (dd, $J = 10.4, 5.5$ Hz, 1H), 3.15 (dd, $J = 13.5, 5.7$ Hz, 1H), 2.93 – 2.85 (m, 1H), 2.19 (s, 3H), 2.00 (s, 3H), 1.87 (s, 3H), 1.38 (d, $J = 7.0$ Hz, 3H).

¹³C NMR (CDCl₃, 150 MHz): δ 172.00, 170.56, 170.08, 169.66, 169.60, 160.40, 155.90, 143.73, 143.61, 141.29, 141.27, 133.32, 132.18, 131.44, 130.74, 129.15, 129.04, 128.94, 127.74, 127.72, 127.09, 127.05, 125.48, 125.44, 125.01, 124.96, 123.15, 119.98, 119.97, 118.79, 85.31, 69.47,

69.24, 67.44, 67.11, 67.03, 66.84, 65.98, 62.22, 53.78, 48.35, 47.06, 32.54, 20.76, 20.63, 20.34, 18.08.

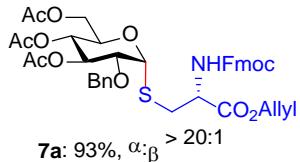
HRMS (ESI): calc. for $C_{44}H_{46}F_3N_3O_{12}SNa$ $[M + Na]^+$, 920.2647; found: 920.2641.



1H NMR (CDCl₃, 600 MHz): δ 8.63 (d, $J = 1.7$ Hz, 1H), 8.16 (d, $J = 7.6$ Hz, 1H), 7.74 (dd, $J = 17.5, 5.8$ Hz, 2H), 7.66 (d, $J = 7.5$ Hz, 1H), 7.55 – 7.46 (m, 4H), 7.39 (dd, $J = 13.4, 6.9$ Hz, 2H), 7.33 – 7.23 (m, 5H), 7.18 (s, 2H), 6.92 (s, 1H), 6.81 (s, 1H), 5.93 – 5.83 (m, 1H), 5.46 (dd, $J = 9.3, 3.8$ Hz, 2H), 5.32 (ddd, $J = 9.6, 9.0, 2.2$ Hz, 3H), 5.24 (dd, $J = 10.5, 1.1$ Hz, 1H), 4.66 – 4.41 (m, 7H), 4.31 – 4.12 (m, 4H), 4.05 (dd, $J = 10.5, 5.5$ Hz, 1H), 3.07 (d, $J = 36.9$ Hz, 3H), 2.87 (d, $J = 13.3$ Hz, 1H), 2.16 (s, 3H), 2.04 (s, 3H), 1.85 (s, 3H), 1.35 (d, $J = 7.0$ Hz, 3H).

^{13}C NMR (CDCl₃, 150 MHz): δ 171.90, 170.68, 170.09, 169.59, 169.07, 160.44, 143.72, 141.25, 133.56, 132.18, 131.49, 130.74, 129.30, 129.18, 129.03, 128.78, 127.72, 127.21, 127.05, 124.97, 119.97, 119.94, 118.76, 84.73, 69.23, 67.46, 67.15, 67.02, 66.72, 65.96, 62.11, 53.77, 48.38, 47.04, 31.47, 29.25, 20.73, 20.71, 20.32, 17.89.

HRMS (ESI): calc. for $C_{53}H_{55}F_3N_4O_{13}SNa$ $[M + Na]^+$, 1067.3331; found: 1067.3337.

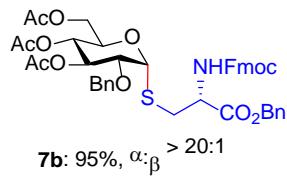


1H NMR (CDCl₃, 600 MHz): δ 7.76 (dd, $J = 7.5, 0.7$ Hz, 2H), 7.59 (dd, $J = 19.0, 7.5$ Hz, 2H), 7.40 (q, $J = 7.4$ Hz, 2H), 7.35 – 7.26 (m, 5H), 6.06 (d, $J = 8.7$ Hz, 1H), 5.91 (ddd, $J = 16.5, 11.2, 5.9$ Hz, 1H), 5.34 (d, $J = 17.2$ Hz, 1H), 5.24 (ddd, $J = 19.2, 9.9, 2.1$ Hz, 3H), 4.98 – 4.90 (m, 1H), 4.78 – 4.70 (m, 1H), 4.64 (dd, $J = 22.9, 9.0$ Hz, 3H), 4.55 (d, $J = 12.2$ Hz, 1H), 4.40 (dd, $J = 7.0,$

1.7 Hz, 2H), 4.35 – 4.26 (m, 1H), 4.24 – 4.12 (m, 3H), 3.80 (dd, J = 10.0, 5.5 Hz, 1H), 3.25 (dd, J = 14.5, 5.0 Hz, 1H), 2.90 (dd, J = 14.5, 3.8 Hz, 1H), 2.02 (s, 3H), 2.01 (s, 3H), 2.00 (s, 3H).

^{13}C NMR (CDCl₃, 150 MHz): δ 170.50, 169.94, 169.87, 169.73, 155.69, 143.85, 143.62, 141.31, 137.07, 131.26, 128.51, 128.14, 127.79, 127.73, 127.71, 127.04, 125.07, 124.98, 119.99, 119.97, 119.34, 85.32, 76.11, 72.50, 71.86, 68.41, 67.07, 66.43, 62.07, 54.18, 47.05, 34.58, 20.73, 20.61, 20.58.

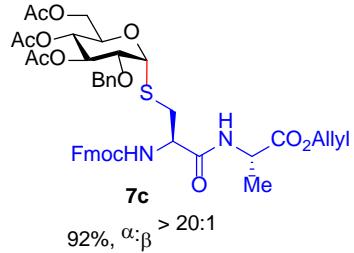
HRMS (ESI): calc. for C₄₀H₄₃NO₁₂SNa [M + Na]⁺, 784.2398; found: 784.2392.



^1H NMR (CDCl₃, 600 MHz): δ 7.76 (d, J = 7.5 Hz, 2H), 7.59 (dd, J = 18.7, 7.5 Hz, 2H), 7.46 – 7.23 (m, 14H), 6.12 (d, J = 8.8 Hz, 1H), 5.17 (dt, J = 23.4, 5.9 Hz, 4H), 4.92 (t, J = 9.8 Hz, 1H), 4.76 (dt, J = 8.6, 4.2 Hz, 1H), 4.59 (d, J = 12.3 Hz, 1H), 4.51 (d, J = 12.3 Hz, 1H), 4.40 (dd, J = 6.9, 4.2 Hz, 2H), 4.31 – 4.08 (m, 4H), 3.76 (dd, J = 10.0, 5.6 Hz, 1H), 3.26 (dd, J = 14.5, 4.9 Hz, 1H), 2.88 (dd, J = 14.5, 3.7 Hz, 1H), 2.03 (s, 3H), 2.01 (s, 3H), 1.97 (s, 3H).

^{13}C NMR (CDCl₃, 150 MHz): δ 170.50, 170.11, 169.86, 169.72, 155.71, 143.87, 143.62, 141.31, 141.27, 137.08, 134.84, 128.70, 128.65, 128.64, 128.51, 128.14, 127.80, 127.73, 127.71, 127.05, 125.08, 124.98, 119.99, 119.97, 85.30, 72.50, 71.82, 68.42, 68.39, 67.72, 67.08, 62.06, 54.20, 47.05, 34.68, 20.73, 20.63, 20.54.

HRMS (ESI): calc. for C₄₄H₄₅NO₁₂SNa [M + Na]⁺, 834.2555; found: 834.2565.

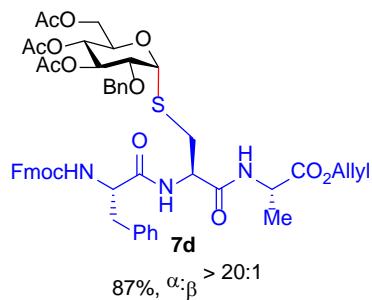


^1H NMR (CDCl₃, 600 MHz): δ 7.76 (dd, J = 7.5, 2.2 Hz, 2H), 7.59 (t, J = 8.0 Hz, 2H), 7.40 (dd, J = 13.1, 7.3 Hz, 2H), 7.30 (qdd, J = 20.3, 10.7, 6.6 Hz, 7H), 6.86 (d, J = 6.4 Hz, 1H), 5.96 – 5.79 (m, 2H), 5.47 (d, J = 5.3 Hz, 1H), 5.39 – 5.20 (m, 3H), 4.95 (t, J = 9.8 Hz, 1H), 4.73 – 4.48 (m, 5H), 4.47 – 4.34 (m, 3H), 4.34 – 4.26 (m, 1H), 4.26 – 4.17 (m, 2H), 4.12 (dd, J = 14.2, 7.1 Hz,

1H), 3.83 (dd, $J = 9.0, 5.2$ Hz, 1H), 3.11 (d, $J = 9.6$ Hz, 1H), 2.81 (dd, $J = 13.6, 4.9$ Hz, 1H), 2.02 (s, 3H), 2.01 (s, 3H), 2.00 (s, 3H), 1.37 (d, $J = 7.1$ Hz, 3H).

^{13}C NMR (CDCl₃, 150 MHz): δ 171.92, 170.58, 169.85, 169.73, 169.40, 155.86, 143.69, 143.58, 141.31, 141.28, 136.96, 131.49, 128.57, 128.51, 128.26, 128.17, 128.01, 127.91, 127.77, 127.75, 127.65, 127.59, 127.11, 127.08, 125.00, 120.01, 119.99, 118.81, 85.20, 76.21, 72.70, 72.14, 68.52, 68.43, 67.16, 66.01, 62.06, 54.94, 48.42, 47.06, 33.95, 20.75, 20.68, 20.61, 17.92.

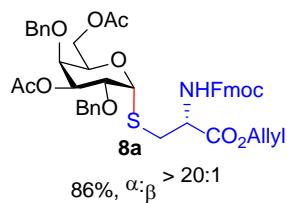
HRMS (ESI): calc. for C₄₃H₄₈N₂O₁₃SNa [M + Na]⁺, 855.2769; found: 855.2764.



^1H NMR (CDCl₃, 600 MHz): δ 7.75 (d, $J = 7.5$ Hz, 2H), 7.50 (dd, $J = 13.9, 7.5$ Hz, 2H), 7.42 – 7.36 (m, 2H), 7.36 – 7.22 (m, 10H), 7.18 (s, 2H), 6.85 (d, $J = 38.5$ Hz, 2H), 5.94 – 5.84 (m, 1H), 5.56 (d, $J = 4.3$ Hz, 1H), 5.27 (ddd, $J = 13.8, 12.1, 1.3$ Hz, 4H), 4.95 (t, $J = 9.7$ Hz, 1H), 4.71 – 4.38 (m, 8H), 4.36 – 4.11 (m, 5H), 3.81 (dd, $J = 9.9, 5.5$ Hz, 1H), 3.18 – 2.96 (m, 3H), 2.88 – 2.77 (m, 1H), 2.05 (s, 3H), 2.01 (s, 3H), 1.99 (s, 3H), 1.34 (d, $J = 6.7$ Hz, 3H).

^{13}C NMR (CDCl₃, 150 MHz): δ 171.79, 170.87, 170.68, 169.88, 169.73, 168.83, 143.68, 141.26, 136.97, 135.96, 131.53, 129.22, 128.82, 128.50, 128.14, 127.87, 127.72, 127.26, 127.08, 124.96, 119.97, 118.77, 84.81, 76.25, 72.66, 72.19, 68.61, 68.45, 67.11, 65.98, 61.92, 53.43, 48.45, 47.05, 38.05, 33.11, 20.74, 20.62, 17.70.

HRMS (ESI): calc. for C₃₈H₅₂N₃O₁₄SNa [M + Na]⁺, 1002.3453; found: 1002.3458.

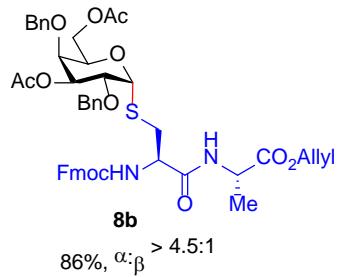


^1H NMR (CDCl₃, 600 MHz): δ 7.73 (d, $J = 7.5$ Hz, 2H), 7.60 (dd, $J = 17.0, 7.5$ Hz, 2H), 7.44 – 7.21 (m, 14H), 6.16 (d, $J = 8.8$ Hz, 1H), 5.97 – 5.83 (m, 1H), 5.40 – 5.20 (m, 3H), 5.07 (dd, $J = 10.5, 2.9$ Hz, 1H), 4.77 – 4.61 (m, 5H), 4.58 (d, $J = 12.0$ Hz, 1H), 4.52 (d, $J = 11.2$ Hz, 1H), 4.39

(td, $J = 17.9, 10.5$ Hz, 2H), 4.33 – 4.16 (m, 4H), 4.12 (dd, $J = 11.3, 4.9$ Hz, 1H), 3.98 (d, $J = 1.6$ Hz, 1H), 3.29 (dd, $J = 14.5, 5.1$ Hz, 1H), 2.88 (dd, $J = 14.5, 3.7$ Hz, 1H), 2.04 (s, 3H), 2.00 (s, 3H).

^{13}C NMR (CDCl₃, 150 MHz): δ 170.54, 170.20, 169.98, 155.75, 143.84, 143.71, 141.27, 141.24, 137.55, 137.41, 131.39, 128.52, 128.41, 128.18, 128.12, 127.93, 127.79, 127.65, 127.10, 127.05, 125.17, 125.08, 119.90, 119.11, 85.71, 75.29, 74.89, 73.35, 72.58, 72.49, 69.30, 67.06, 66.29, 62.88, 54.27, 47.06, 34.20, 20.95, 20.66.

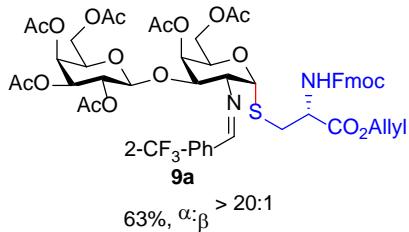
HRMS (ESI): calc. for C₃₈H₄₇NO₁₁SNa [M + Na]⁺, 832.2762; found: 832.2761.



^1H NMR (CDCl₃, 600 MHz): α/β mixture, inseparable, δ 7.79 – 7.70 (m, 2H), 7.64 – 7.54 (m, 2H), 7.44 – 7.25 (m, 14H), 7.17 (d, $J = 20.4$ Hz, 0.2H), 6.97 (d, $J = 6.8$ Hz, 1H), 6.09 (d, $J = 5.7$ Hz, 0.2H), 5.97 (d, $J = 6.7$ Hz, 1H), 5.93 – 5.82 (m, 1H), 5.45 (s, 1H), 5.31 (d, $J = 17.3$ Hz, 1H), 5.23 (d, $J = 10.4$ Hz, 1H), 5.16 – 5.05 (m, 1H), 4.97 (d, $J = 9.0$ Hz, 0.2H), 4.86 (d, $J = 10.8$ Hz, 0.2H), 4.74 – 4.48 (m, 7H), 4.44 (d, $J = 5.8$ Hz, 2H), 4.37 (d, $J = 5.3$ Hz, 1H), 4.21 (dd, $J = 18.3, 11.6$ Hz, 4H), 4.12 – 3.94 (m, 3H), 3.84 (d, $J = 10.1$ Hz, 1H), 3.19 (dd, $J = 32.9, 11.6$ Hz, 1.2H), 2.97 (d, $J = 8.1$ Hz, 0.2H), 2.80 (d, $J = 10.3$ Hz, 1H), 2.04 (s, 3H), 1.95 (s, 3H), 1.38 (d, $J = 6.9$ Hz, 3H).

^{13}C NMR (CDCl₃, 150 MHz): δ 171.99, 170.51, 170.17, 169.61, 155.98, 146.78, 143.73, 143.66, 141.31, 141.26, 137.48, 137.43, 135.83, 131.66, 131.53, 128.51, 128.45, 128.40, 128.38, 128.14, 128.09, 128.04, 127.98, 127.96, 127.90, 127.71, 127.70, 127.14, 127.08, 125.02, 124.96, 121.19, 119.95, 119.92, 118.71, 118.57, 85.41, 75.56, 75.12, 75.04, 74.56, 74.41, 73.50, 72.76, 72.52, 69.30, 66.98, 65.93, 65.84, 63.01, 62.72, 55.23, 54.62, 48.54, 48.35, 47.09, 33.68, 29.67, 20.97, 20.87, 20.66, 18.08, 17.93.

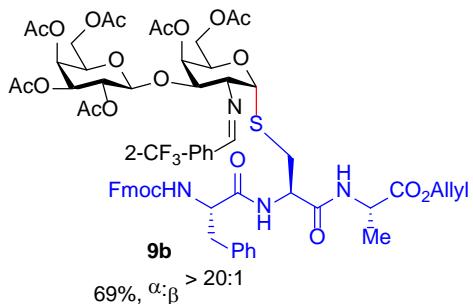
HRMS (ESI): calc. for C₃₈H₅₂N₂O₁₂SNa [M + Na]⁺, 903.3133; found: 903.3136.



¹H NMR (CDCl₃, 600 MHz): δ 8.61 (s, 1H), 8.24 (d, J = 7.7 Hz, 1H), 7.81 – 7.68 (m, 3H), 7.65 – 7.52 (m, 4H), 7.40 (dd, J = 13.1, 7.2 Hz, 2H), 7.31 (dd, J = 16.3, 7.9 Hz, 2H), 5.90 (d, J = 8.5 Hz, 1H), 5.87 – 5.77 (m, 1H), 5.51 – 5.48 (m, 1H), 5.23 (ddd, J = 21.7, 14.4, 6.8 Hz, 4H), 5.01 (dd, J = 10.4, 7.9 Hz, 1H), 4.83 (dd, J = 10.5, 3.5 Hz, 1H), 4.60 (ddd, J = 30.9, 23.7, 6.4 Hz, 4H), 4.42 – 4.19 (m, 5H), 4.17 – 4.01 (m, 5H), 3.84 (t, J = 6.6 Hz, 1H), 3.16 (dd, J = 14.1, 5.7 Hz, 1H), 2.95 (dd, J = 14.1, 4.0 Hz, 1H), 2.17 (s, 3H), 2.12 (s, 3H), 2.05 (d, J = 6.8 Hz, 6H), 1.89 (s, 3H), 1.48 (s, 3H).

¹³C NMR (CDCl₃, 150 MHz): δ 171.10, 170.53, 170.35, 170.25, 170.15, 170.04, 169.86, 168.76, 160.10, 155.66, 143.81, 143.64, 141.28, 133.56, 133.30, 132.28, 132.14, 131.23, 130.90, 129.04, 128.72, 127.70, 127.00, 125.73, 125.07, 119.99, 119.06, 100.65, 85.64, 75.21, 70.75, 70.70, 69.08, 68.84, 68.69, 68.47, 67.16, 67.11, 66.70, 66.31, 63.02, 60.98, 60.36, 54.06, 47.06, 32.95, 21.02, 20.76, 20.69, 20.67, 20.61, 20.46, 19.92.

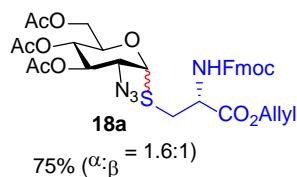
HRMS (ESI): calc. for C₅₃H₅₇F₃N₂O₁₉SNa [M + Na]⁺, 1137.3121; found: 1137.3125.



¹H NMR (CDCl₃, 600 MHz): δ 8.61 (s, 1H), 8.23 (d, J = 7.7 Hz, 1H), 7.80 – 7.66 (m, 3H), 7.52 (ddt, J = 30.9, 15.4, 7.6 Hz, 4H), 7.39 (dd, J = 14.6, 7.2 Hz, 2H), 7.34 – 7.20 (m, 6H), 7.15 (s, 2H), 6.93 (d, J = 6.7 Hz, 1H), 6.77 (s, 1H), 5.87 (ddd, J = 16.2, 11.0, 5.7 Hz, 1H), 5.46 (d, J = 3.0 Hz, 1H), 5.37 – 5.22 (m, 5H), 4.99 (dd, J = 10.4, 7.9 Hz, 2H), 4.80 (dd, J = 10.5, 3.3 Hz, 1H), 4.64 – 4.41 (m, 6H), 4.34 – 4.26 (m, 2H), 4.19 – 4.02 (m, 5H), 3.76 (d, J = 6.9 Hz, 1H), 3.07 (s, 2H), 2.96 (d, J = 6.8 Hz, 1H), 2.82 (d, J = 10.5 Hz, 1H), 2.13 (s, 3H), 2.10 (s, 3H), 2.04 (s, 3H), 2.01 (s, 3H), 1.88 (s, 3H), 1.43 (s, 3H), 1.33 (d, J = 7.1 Hz, 3H).

¹³C NMR (CDCl₃, 150 MHz): δ 171.99, 170.85, 170.82, 170.34, 170.24, 170.01, 169.96, 169.07, 168.74, 160.09, 143.65, 141.27, 141.23, 136.02, 133.34, 132.07, 131.46, 130.84, 129.26, 128.79, 128.72, 127.79, 127.75, 127.14, 127.08, 124.94, 120.02, 120.00, 118.71, 100.65, 84.97, 75.54, 70.75, 70.60, 68.92, 68.78, 68.49, 68.45, 67.11, 66.70, 65.92, 62.95, 60.93, 60.35, 55.94, 53.78, 48.33, 47.06, 30.89, 29.66, 29.24, 20.82, 20.77, 20.67, 20.63, 20.60, 20.45, 19.86, 17.86.

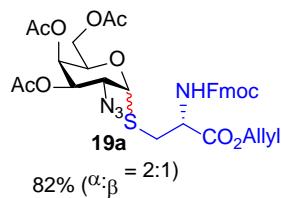
HRMS (ESI): calc. for C₆₅H₇₁F₃N₄O₂₁Na [M + Na]⁺, 1355.4176; found: 1355.4182.



¹H NMR (CDCl₃, 600 MHz): α/β mixture, inseparable, δ 7.76 (d, *J* = 7.3 Hz, 4.4H), 7.60 (d, *J* = 5.5 Hz, 5H), 7.33 (dd, *J* = 33.0, 25.9 Hz, 9H), 6.04 (d, *J* = 8.4 Hz, 1.3H), 5.91 (d, *J* = 5.9 Hz, 2.6H), 5.69 – 5.50 (m, 3.6H), 5.48 – 5.09 (m, 9.2H), 5.00 (t, *J* = 9.6 Hz, 1.6H), 4.94 (t, *J* = 9.7 Hz, 1H), 4.70 (d, *J* = 41.2 Hz, 7H), 4.55 – 4.00 (m, 17H), 3.98 – 3.92 (m, 1H), 3.51 (d, *J* = 8.8 Hz, 1.6H), 3.35 – 3.21 (m, 2.6H), 3.04 (dd, *J* = 20.4, 9.2 Hz, 2.6H), 2.08 (s, 3H), 2.05 (s, 4.8H), 2.03 (s, 4.8H), 1.99 (s, 4.8H), 1.65 (s, 3H), 1.56 (s, 3H).

¹³C NMR (CDCl₃, 150 MHz): δ 170.48, 170.30, 170.01, 169.69, 155.68, 143.84, 143.74, 141.26, 131.27, 131.18, 127.73, 127.69, 127.05, 125.14, 125.08, 124.91, 119.97, 119.47, 119.12, 92.28, 85.39, 71.64, 71.25, 68.80, 68.56, 67.97, 67.31, 66.75, 66.54, 66.44, 62.04, 61.95, 61.53, 61.14, 60.35, 54.22, 53.40, 47.17, 47.04, 35.49, 30.43, 29.78, 29.66, 29.24, 28.79, 20.67, 20.64, 20.60, 20.58, 20.55.

HRMS (ESI): calc. for C₃₃H₃₆N₄O₁₁Na [M + Na]⁺, 719.1993; found: 719.1998.

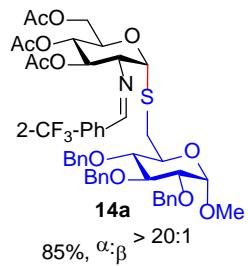


¹H NMR (CDCl₃, 600 MHz): α/β mixture, inseparable, δ 7.75 (t, *J* = 10.1 Hz, 5.2H), 7.61 (t, *J* = 8.8 Hz, 5.2H), 7.40 (dd, *J* = 12.1, 7.2 Hz, 5.5H), 7.36 – 7.29 (m, 5.4H), 6.07 (d, *J* = 8.5 Hz, 2H), 5.90 (ddd, *J* = 16.3, 10.5, 5.8 Hz, 2.8H), 5.66 (d, *J* = 8.3 Hz, 1H), 5.59 (d, *J* = 3.2 Hz, 1H), 5.46 –

5.23 (m, 11.2H), 5.03 (dd, $J = 11.1$, 2.8 Hz, 2H), 4.76 – 4.62 (m, 8H), 4.52 – 4.32 (m, 8H), 4.31 – 4.18 (m, 7H), 4.08 (dddd, $J = 22.7$, 18.2, 11.4, 6.4 Hz, 8H), 3.87 – 3.79 (m, 1H), 3.36 – 3.25 (m, 3H), 3.04 (td, $J = 14.0$, 5.3 Hz, 3H), 2.18 (s, 6H), 2.13 (s, 3H), 2.05 (d, $J = 9.0$ Hz, 6H), 1.99 (s, 6H), 1.64 (s, 3H), 1.56 (s, 3H).

^{13}C NMR (CDCl₃, 150 MHz): δ 170.34, 170.29, 170.04, 169.82, 169.70, 169.46, 155.66, 143.83, 143.76, 143.63, 141.35, 141.30, 141.26, 131.31, 131.23, 127.70, 127.68, 127.03, 127.00, 125.12, 125.03, 125.00, 119.95, 119.36, 119.07, 92.75, 85.63, 69.67, 69.17, 67.83, 67.50, 67.35, 66.97, 66.88, 66.49, 66.42, 61.76, 61.73, 60.35, 57.98, 57.75, 54.15, 47.12, 47.06, 34.90, 29.85, 21.01, 20.61, 20.55, 20.49.

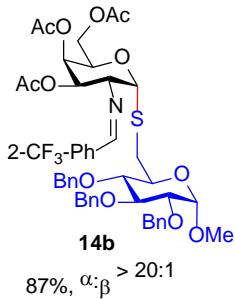
HRMS (ESI): calc. for C₃₃H₃₆N₄O₁₁SNa [M + Na]⁺, 719.1993; found: 719.1999.



^1H NMR (CDCl₃, 600 MHz): δ 8.60 (d, $J = 1.9$ Hz, 1H), 8.21 (d, $J = 7.6$ Hz, 1H), 7.66 (d, $J = 7.5$ Hz, 1H), 7.53 (dd, $J = 14.6$, 7.4 Hz, 2H), 7.36 – 7.22 (m, 15H), 5.57 (t, $J = 9.7$ Hz, 1H), 5.42 (d, $J = 5.6$ Hz, 1H), 5.13 (t, $J = 9.8$ Hz, 1H), 4.93 (dd, $J = 18.2$, 10.9 Hz, 2H), 4.78 (d, $J = 10.8$ Hz, 1H), 4.70 (d, $J = 12.0$ Hz, 1H), 4.63 (d, $J = 11.0$ Hz, 1H), 4.59 – 4.53 (m, 2H), 4.51 (d, $J = 3.5$ Hz, 1H), 4.33 (dd, $J = 12.3$, 4.3 Hz, 1H), 4.02 (dd, $J = 12.3$, 2.0 Hz, 1H), 3.97 – 3.88 (m, 2H), 3.85 – 3.79 (m, 1H), 3.46 – 3.40 (m, 2H), 3.31 (s, 3H), 2.99 (dd, $J = 13.7$, 2.3 Hz, 1H), 2.63 (dd, $J = 13.1$, 8.2 Hz, 1H), 2.06 (s, 3H), 2.03 (s, 3H), 1.86 (s, 3H).

^{13}C NMR (CDCl₃, 150 MHz): δ 170.56, 169.89, 169.68, 169.63, 159.89, 138.61, 138.05, 138.01, 132.23, 130.92, 130.78, 129.09, 128.72, 128.59, 128.41, 128.40, 128.37, 127.99, 127.96, 127.88, 127.79, 127.74, 127.62, 97.76, 84.21, 81.88, 80.42, 80.17, 75.69, 75.14, 73.33, 73.22, 72.10, 71.75, 69.70, 68.70, 67.90, 62.10, 55.07, 30.53, 20.70, 20.66, 20.37.

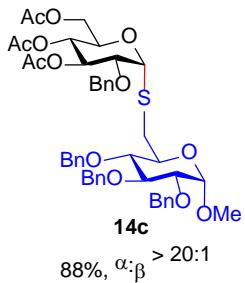
HRMS (ESI): calc. for C₄₈H₅₂F₃NO₁₂SNa [M + Na]⁺, 946.3055; found: 946.3063.



¹H NMR (CDCl₃, 600 MHz): δ 8.62 (d, $J = 2.0$ Hz, 1H), 8.21 (d, $J = 7.6$ Hz, 1H), 7.67 (d, $J = 7.6$ Hz, 1H), 7.54 (dt, $J = 24.8, 7.4$ Hz, 2H), 7.39 – 7.21 (m, 15H), 5.48 (dd, $J = 8.1, 4.0$ Hz, 2H), 5.42 (dd, $J = 10.5, 3.3$ Hz, 1H), 4.93 (dd, $J = 24.2, 10.9$ Hz, 2H), 4.81 – 4.56 (m, 5H), 4.52 (d, $J = 3.5$ Hz, 1H), 4.18 – 4.05 (m, 3H), 3.94 (t, $J = 9.2$ Hz, 1H), 3.86 – 3.78 (m, 1H), 3.46 – 3.40 (m, 2H), 3.29 (s, 3H), 3.00 (dd, $J = 13.8, 2.3$ Hz, 1H), 2.64 (dd, $J = 13.8, 7.6$ Hz, 1H), 2.18 (s, 3H), 1.98 (s, 3H), 1.86 (s, 3H).

¹³C NMR (CDCl₃, 150 MHz): δ 170.30, 170.11, 169.68, 159.89, 138.61, 138.06, 138.02, 132.18, 130.65, 129.07, 128.42, 128.40, 128.37, 127.99, 127.97, 127.88, 127.81, 127.76, 127.62, 125.44, 97.72, 84.47, 81.91, 80.50, 80.18, 75.70, 75.18, 73.32, 69.74, 69.54, 67.19, 66.85, 61.97, 54.99, 30.23, 20.75, 20.66, 20.37.

HRMS (ESI): calc. for C₄₈H₅₂F₃NO₁₂SNa [M + Na]⁺, 946.3055; found: 946.3061.

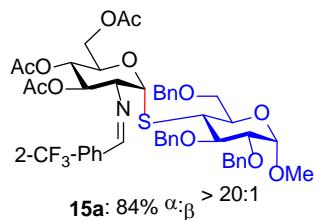


¹H NMR (CDCl₃, 600 MHz): δ 7.42 – 7.13 (m, 20H), 5.58 (d, $J = 5.6$ Hz, 1H), 5.30 (dd, $J = 15.4, 5.7$ Hz, 1H), 5.03 – 4.88 (m, 3H), 4.80 (dd, $J = 30.1, 11.5$ Hz, 2H), 4.68 – 4.58 (m, 3H), 4.53 (d, $J = 3.5$ Hz, 1H), 4.44 (d, $J = 12.2$ Hz, 1H), 4.39 (ddd, $J = 10.2, 4.0, 2.1$ Hz, 1H), 4.29 (dd, $J = 12.4, 4.2$ Hz, 1H), 4.00 (t, $J = 9.2$ Hz, 1H), 3.95 (dd, $J = 12.4, 2.0$ Hz, 1H), 3.90 – 3.85 (m, 1H), 3.78 (dd, $J = 9.9, 5.6$ Hz, 1H), 3.54 (t, $J = 9.3$ Hz, 1H), 3.48 (dd, $J = 9.6, 3.6$ Hz, 1H), 3.38 (s, 3H), 2.94 (dd, $J = 13.9, 2.5$ Hz, 1H), 2.67 (dd, $J = 13.9, 6.7$ Hz, 1H), 2.03 (s, 3H), 2.01 (s, 3H), 1.99 (s, 3H).

¹³C NMR (CDCl₃, 150 MHz): δ 170.49, 169.90, 169.76, 138.64, 138.03, 137.97, 137.12, 128.45, 128.39, 128.08, 128.03, 127.96, 127.86, 127.83, 127.81, 127.77, 127.60, 98.05, 83.62, 81.88, 80.10,

79.94, 75.66, 75.55, 75.26, 73.41, 72.09, 71.62, 70.20, 68.40, 67.56, 61.93, 55.34, 30.39, 20.75, 20.68, 20.62.

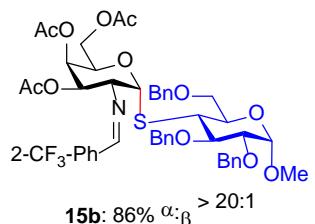
HRMS (ESI): calc. for $C_{47}H_{54}O_{13}SNa$ $[M + Na]^+$, 881.3177; found: 881.3178.



1H NMR (CDCl₃, 600 MHz): δ 8.32 (d, J = 1.8 Hz, 1H), 8.23 (d, J = 7.8 Hz, 1H), 7.66 (d, J = 7.7 Hz, 1H), 7.59 (t, J = 7.4 Hz, 1H), 7.54 (t, J = 7.6 Hz, 1H), 7.40 – 7.23 (m, 10H), 7.10 (t, J = 7.3 Hz, 1H), 7.04 (t, J = 7.5 Hz, 2H), 6.96 (d, J = 7.3 Hz, 2H), 5.82 (d, J = 5.8 Hz, 1H), 5.40 (t, J = 9.7 Hz, 1H), 5.06 (t, J = 9.8 Hz, 1H), 4.95 (d, J = 11.0 Hz, 1H), 4.74 (d, J = 12.1 Hz, 1H), 4.64 (ddd, J = 22.5, 16.1, 7.7 Hz, 5H), 4.40 – 4.33 (m, 1H), 4.21 (dd, J = 12.3, 3.9 Hz, 1H), 4.02 (t, J = 9.9 Hz, 1H), 3.90 – 3.76 (m, 4H), 3.71 (dd, J = 9.9, 5.7 Hz, 1H), 3.51 (dd, J = 9.5, 3.4 Hz, 1H), 3.41 (s, 3H), 3.02 (t, J = 10.5 Hz, 1H), 2.02 (d, J = 6.2 Hz, 6H), 1.82 (s, 3H).

^{13}C NMR (CDCl₃, 150 MHz): δ 170.49, 169.71, 169.68, 159.57, 138.21, 137.99, 137.96, 133.19, 132.16, 130.77, 128.92, 128.42, 128.40, 128.04, 127.92, 127.88, 127.69, 127.66, 127.23, 126.94, 125.43, 98.24, 84.83, 83.43, 80.72, 75.87, 73.65, 73.12, 72.01, 71.53, 70.05, 69.84, 68.41, 68.35, 61.82, 55.36, 43.99, 20.71, 20.66, 20.27.

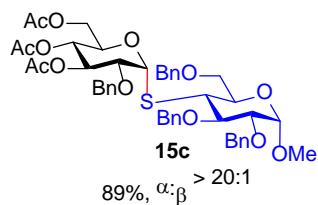
HRMS (ESI): calc. for $C_{48}H_{52}F_3NO_{12}SNa$ $[M + Na]^+$, 946.3055; found: 946.3052.



1H NMR (CDCl₃, 600 MHz): δ 8.36 (d, J = 1.9 Hz, 1H), 8.23 (d, J = 7.8 Hz, 1H), 7.67 (d, J = 7.7 Hz, 1H), 7.61 (t, J = 7.6 Hz, 1H), 7.55 (t, J = 7.6 Hz, 1H), 7.41 – 7.21 (m, 10H), 7.17 – 6.91 (m, 5H), 5.81 (d, J = 5.7 Hz, 1H), 5.36 (d, J = 2.4 Hz, 1H), 5.22 (dd, J = 10.6, 3.3 Hz, 1H), 4.95 (d, J = 11.1 Hz, 1H), 4.76 – 4.54 (m, 6H), 4.36 (t, J = 6.6 Hz, 1H), 4.06 – 3.73 (m, 7H), 3.51 (dd, J = 9.5, 3.4 Hz, 1H), 3.42 (s, 3H), 3.01 – 2.89 (m, 1H), 2.14 (s, 3H), 1.96 (s, 3H), 1.83 (s, 3H).

¹³C NMR (CDCl₃, 150 MHz): δ 170.15, 170.12, 169.66, 159.60, 138.34, 138.10, 138.00, 133.46, 132.17, 130.67, 128.98, 128.41, 128.38, 128.03, 127.98, 127.88, 127.72, 127.64, 127.19, 127.00, 125.41, 98.15, 85.13, 83.56, 80.77, 75.85, 73.33, 73.07, 70.34, 69.79, 69.12, 67.14, 66.52, 61.92, 55.32, 43.94, 29.67, 20.74, 20.69, 20.29.

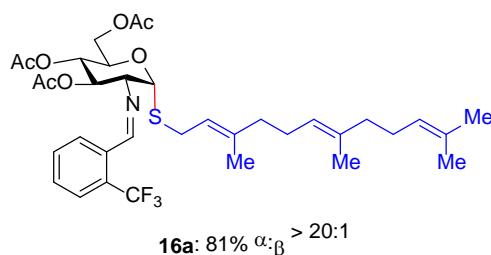
HRMS (ESI): calc. for C₄₈H₅₂F₃NO₁₂SNa [M + Na]⁺, 946.3055; found: 946.3058.



¹H NMR (CDCl₃, 600 MHz): δ 7.40 – 7.13 (m, 18H), 6.94 (dd, *J* = 6.4, 2.7 Hz, 2H), 6.02 (d, *J* = 5.6 Hz, 1H), 5.20 (dd, *J* = 20.3, 10.8 Hz, 2H), 4.89 (dd, *J* = 10.7, 8.6 Hz, 2H), 4.75 – 4.66 (m, 2H), 4.64 – 4.55 (m, 3H), 4.40 (d, *J* = 12.3 Hz, 1H), 4.18 (ddd, *J* = 16.4, 11.2, 2.9 Hz, 2zH), 4.02 (t, *J* = 9.9 Hz, 1H), 3.97 (d, *J* = 12.3 Hz, 1H), 3.81 (ddd, *J* = 21.8, 10.2, 5.8 Hz, 4H), 3.63 (dd, *J* = 9.8, 5.6 Hz, 1H), 3.57 (dd, *J* = 9.4, 3.4 Hz, 1H), 3.40 (s, 3H), 3.03 (t, *J* = 10.8 Hz, 1H), 2.00 (d, *J* = 5.7 Hz, 6H), 1.91 (s, 3H).

¹³C NMR (CDCl₃, 150 MHz): δ 170.44, 169.92, 169.59, 139.00, 137.96, 137.87, 137.07, 128.42, 128.39, 128.33, 128.17, 128.07, 127.91, 127.69, 127.67, 127.46, 127.11, 126.26, 98.10, 83.98, 82.41, 81.00, 75.51, 75.25, 73.65, 73.07, 71.89, 71.37, 70.04, 69.93, 68.19, 68.14, 61.75, 55.40, 44.21, 20.72, 20.71, 20.63.

HRMS (ESI): calc. for C₄₇H₅₄O₁₃SNa [M + Na]⁺, 881.3177; found: 881.3173.

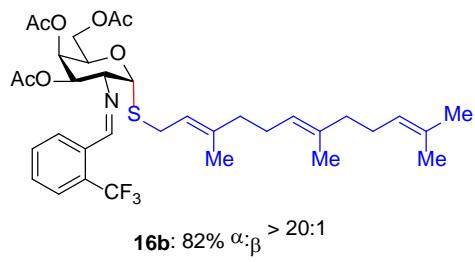


¹H NMR (CDCl₃, 600 MHz): δ 8.59 (d, *J* = 2.0 Hz, 1H), 8.21 (d, *J* = 7.8 Hz, 1H), 7.66 (d, *J* = 7.7 Hz, 1H), 7.59 (t, *J* = 7.6 Hz, 1H), 7.52 (t, *J* = 7.6 Hz, 1H), 5.58 (t, *J* = 9.6 Hz, 1H), 5.32 (d, *J* = 5.7 Hz, 1H), 5.23 (t, *J* = 7.7 Hz, 1H), 5.08 (ddd, *J* = 13.2, 8.3, 7.6 Hz, 3H), 4.63 (ddd, *J* = 10.2, 4.9, 2.0 Hz, 1H), 4.36 (dd, *J* = 12.2, 5.0 Hz, 1H), 4.14 (dd, *J* = 12.2, 2.0 Hz, 1H), 3.93 (dd, *J* = 9.9, 5.6

Hz, 1H), 3.29 (dd, $J = 13.2, 8.9$ Hz, 1H), 3.08 (dd, $J = 13.2, 6.6$ Hz, 1H), 2.13 – 1.99 (m, 12H), 1.97 – 1.91 (m, 2H), 1.86 (s, 3H), 1.67 (d, $J = 6.0$ Hz, 6H), 1.57 (d, $J = 10.6$ Hz, 6H).

^{13}C NMR (CDCl₃, 150 MHz): δ 170.62, 169.95, 169.62, 159.82, 139.93, 135.30, 133.26, 132.22, 131.27, 130.69, 129.15, 129.00, 125.39, 125.35, 124.24, 123.72, 119.12, 82.87, 72.15, 71.95, 69.00, 67.79, 62.47, 39.66, 39.60, 29.67, 26.78, 26.67, 26.38, 25.64, 20.74, 20.69, 20.37, 17.65, 15.95.

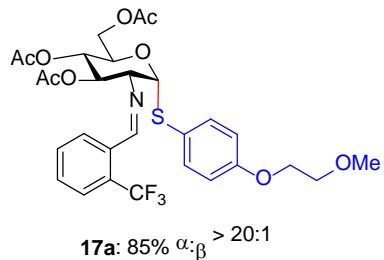
HRMS (ESI): calc. for C₃₅H₄₆F₃NO₇SNa [M + Na]⁺, 704.2839; found: 704.2833.



^1H NMR (CDCl₃, 600 MHz): δ 8.62 (d, $J = 2.0$ Hz, 1H), 8.20 (d, $J = 7.8$ Hz, 1H), 7.67 (d, $J = 7.7$ Hz, 1H), 7.59 (t, $J = 7.6$ Hz, 1H), 7.52 (t, $J = 7.6$ Hz, 1H), 5.50 (d, $J = 2.4$ Hz, 1H), 5.42 (dd, $J = 10.5, 3.3$ Hz, 1H), 5.36 (d, $J = 5.7$ Hz, 1H), 5.22 (t, $J = 7.7$ Hz, 1H), 5.06 (dt, $J = 7.1, 3.8$ Hz, 2H), 4.77 (t, $J = 6.4$ Hz, 1H), 4.18 (d, $J = 6.4$ Hz, 2H), 4.09 (dd, $J = 10.4, 5.7$ Hz, 1H), 3.31 (dd, $J = 13.2, 9.1$ Hz, 1H), 3.06 (dd, $J = 13.2, 6.5$ Hz, 1H), 2.19 (s, 3H), 2.12 – 1.99 (m, 9H), 1.98 – 1.91 (m, 2H), 1.86 (s, 3H), 1.67 (d, $J = 9.1$ Hz, 6H), 1.58 (t, $J = 7.6$ Hz, 6H).

^{13}C NMR (CDCl₃, 150 MHz): δ 170.43, 170.13, 169.66, 159.83, 139.81, 135.26, 133.52, 132.18, 131.25, 130.55, 129.13, 125.39, 125.35, 124.25, 123.76, 119.24, 83.07, 69.76, 67.22, 67.05, 66.80, 62.43, 39.65, 39.61, 29.67, 26.67, 26.54, 26.42, 25.64, 20.75, 20.72, 20.36, 17.64, 15.94, 15.91.

HRMS (ESI): calc. for C₃₅H₄₆F₃NO₇SNa [M + Na]⁺, 704.2839; found: 704.2836.

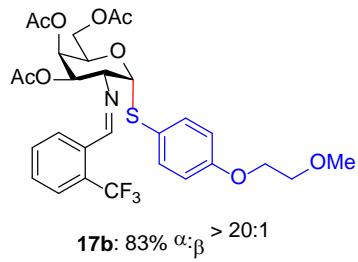


^1H NMR (CDCl₃, 600 MHz): δ 8.65 (d, $J = 1.7$ Hz, 1H), 8.28 (d, $J = 7.8$ Hz, 1H), 7.68 (d, $J = 7.8$ Hz, 1H), 7.62 (t, $J = 7.6$ Hz, 1H), 7.55 (d, $J = 7.6$ Hz, 1H), 7.40 (d, $J = 8.8$ Hz, 2H), 6.86 (d, $J = 8.8$ Hz, 2H), 5.64 (t, $J = 9.7$ Hz, 1H), 5.51 (d, $J = 5.6$ Hz, 1H), 5.13 (t, $J = 9.8$ Hz, 1H), 4.74 (ddd,

$J = 10.2, 4.8, 1.8$ Hz, 1H), 4.32 (dd, $J = 12.3, 5.0$ Hz, 1H), 4.14 – 4.06 (m, 2H), 4.02 – 3.93 (m, 2H), 3.79 – 3.68 (m, 2H), 3.43 (s, 3H), 2.05 (s, 3H), 2.03 (s, 3H), 1.88 (s, 3H).

^{13}C NMR (CDCl₃, 150 MHz): δ 170.57, 169.90, 169.65, 160.17, 158.83, 134.76, 133.23, 132.27, 130.81, 129.19, 129.05, 128.99, 128.18, 125.45, 125.41, 124.96, 123.46, 115.28, 87.38, 72.22, 71.71, 70.87, 68.87, 68.16, 67.32, 62.20, 59.18, 20.68, 20.36.

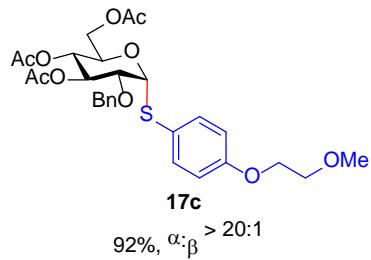
HRMS (ESI): calc. for C₂₉H₃₂F₃NO₉SNa [M + Na]⁺, 650.1642; found: 650.1637.



^1H NMR (CDCl₃, 600 MHz): δ ^1H NMR (600 MHz, CDCl₃) δ 8.67 (d, $J = 1.9$ Hz, 1H), 8.27 (d, $J = 7.8$ Hz, 1H), 7.69 (d, $J = 7.8$ Hz, 1H), 7.62 (t, $J = 7.6$ Hz, 1H), 7.55 (t, $J = 7.7$ Hz, 1H), 7.42 (d, $J = 8.8$ Hz, 2H), 6.86 (d, $J = 8.8$ Hz, 2H), 5.51 (ddd, $J = 13.8, 10.7, 4.5$ Hz, 3H), 4.87 (t, $J = 6.5$ Hz, 1H), 4.18 – 4.00 (m, 5H), 3.79 – 3.68 (m, 2H), 3.43 (s, 3H), 2.17 (s, 3H), 1.99 (s, 3H), 1.88 (s, 3H).

^{13}C NMR (CDCl₃, 150 MHz): δ 170.39, 170.10, 169.69, 160.17, 158.79, 135.00, 133.49, 132.24, 130.67, 129.17, 125.45, 125.41, 123.58, 115.18, 87.91, 70.88, 69.53, 67.37, 67.30, 67.16, 66.93, 62.02, 59.18, 20.72, 20.68, 20.36.

HRMS (ESI): calc. for C₂₉H₃₂F₃NO₉SNa [M + Na]⁺, 650.1642; found: 650.1635.

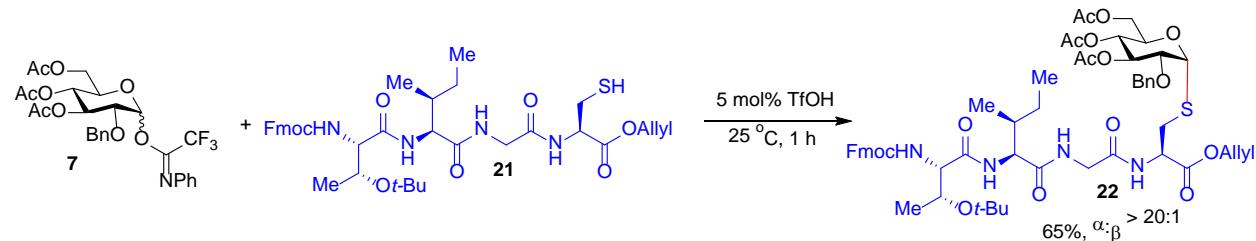


^1H NMR (CDCl₃, 600 MHz): δ 7.42 – 7.26 (m, 7H), 6.86 (d, $J = 8.7$ Hz, 2H), 5.45 (d, $J = 5.5$ Hz, 1H), 5.35 (t, $J = 9.6$ Hz, 1H), 4.97 (t, $J = 9.8$ Hz, 1H), 4.71 (d, $J = 12.2$ Hz, 1H), 4.63 – 4.55 (m, 2H), 4.27 (dd, $J = 12.3, 5.2$ Hz, 1H), 4.14 – 4.07 (m, 2H), 3.98 (dd, $J = 12.3, 2.0$ Hz, 1H), 3.85 (dd, $J = 9.9, 5.5$ Hz, 1H), 3.76 – 3.70 (m, 2H), 3.44 (s, 3H), 2.04 – 2.01 (m, 9H).

¹³C NMR (CDCl₃, 150 MHz): δ 170.50, 169.92, 169.79, 158.95, 137.20, 134.67, 128.47, 128.07, 127.88, 123.60, 115.33, 87.42, 76.29, 72.39, 72.14, 70.88, 68.69, 67.91, 67.36, 62.14, 59.20, 20.75, 20.67, 20.64.

HRMS (ESI): calc. for C₂₈H₃₄O₁₀SNa [M + Na]⁺, 585.1765; found: 585.1768.

7. Synthesis of Sublancin Glycopeptide Fragment 22

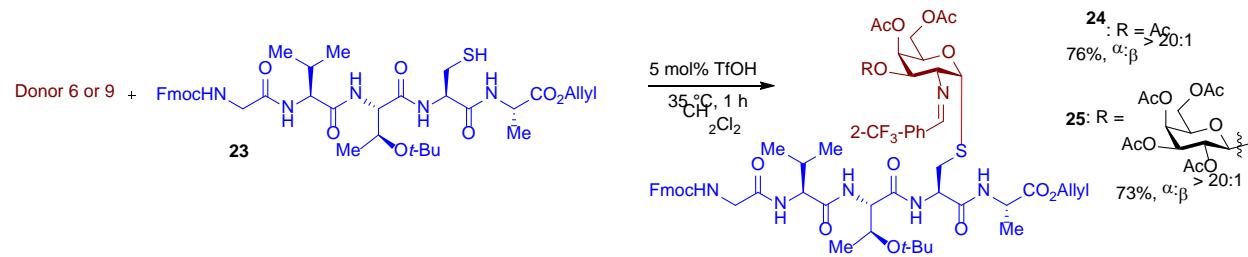


¹H NMR (CDCl₃, 600 MHz): δ 7.81 (d, *J* = 6.6 Hz, 1H), 7.76 (d, *J* = 7.5 Hz, 2H), 7.60 (d, *J* = 7.3 Hz, 2H), 7.52 (d, *J* = 8.7 Hz, 1H), 7.39 (t, *J* = 7.4 Hz, 2H), 7.36 – 7.26 (m, 5H), 6.46 (dd, *J* = 7.4, 4.8 Hz, 1H), 6.35 (d, *J* = 6.5 Hz, 1H), 5.81 (ddd, *J* = 25.2, 15.0, 5.5 Hz, 2H), 5.37 – 5.09 (m, 4H), 5.03 – 4.91 (m, 2H), 4.69 (d, *J* = 12.3 Hz, 1H), 4.61 (dd, *J* = 12.5, 6.6 Hz, 1H), 4.47 (d, *J* = 12.3 Hz, 1H), 4.40 – 4.16 (m, 7H), 4.13 – 4.00 (m, 2H), 3.91 (dd, *J* = 13.0, 6.3 Hz, 1H), 3.83 (dd, *J* = 9.8, 5.6 Hz, 1H), 3.63 (dd, *J* = 17.0, 4.4 Hz, 1H), 3.14 (dd, *J* = 14.0, 4.3 Hz, 1H), 2.93 (dd, *J* = 14.0, 8.8 Hz, 1H), 2.02 (s, 3H), 2.01 (s, 3H), 1.98 (s, 3H), 1.31 (s, 9H), 1.14 (d, *J* = 6.4 Hz, 3H), 0.99 – 0.92 (m, 5H), 0.90 – 0.84 (m, 4H).

¹³C NMR (CDCl₃, 150 MHz): δ 171.46, 170.81, 170.62, 170.49, 170.00, 169.76, 169.49, 168.97, 156.06, 143.84, 143.64, 141.24, 137.40, 131.08, 128.39, 128.37, 127.96, 127.88, 127.72, 127.56, 126.99, 125.16, 125.12, 120.01, 119.99, 119.27, 83.12, 81.74, 81.45, 75.55, 71.96, 71.36, 68.37, 67.91, 67.39, 67.01, 66.44, 61.89, 59.92, 58.13, 55.97, 53.72, 50.85, 47.09, 42.95, 36.59, 35.97, 31.91, 30.77, 29.69, 29.65, 29.35, 29.24, 28.12, 26.67, 25.25, 22.68, 20.77, 20.68, 20.65, 16.40, 15.58, 14.12, 11.12.

HRMS (ESI): calc. for C₅₆H₇₂N₄O₁₆SNa [M + Na]⁺, 1111.4556; found: 1111.4562.

8. Synthesis of S-linked TN and TF Glycopeptide Fragments 24 and 25



24:

¹H NMR (CDCl₃, 600 MHz): δ 8.60 (d, *J* = 1.7 Hz, 1H), 8.15 (d, *J* = 7.6 Hz, 1H), 7.75 (d, *J* = 7.5 Hz, 3H), 7.66 (d, *J* = 7.3 Hz, 1H), 7.62 – 7.49 (m, 3H), 7.39 (t, *J* = 7.4 Hz, 2H), 7.29 (t, *J* = 7.4 Hz, 2H), 6.69 (s, 1H), 5.84 (ddd, *J* = 22.8, 10.8, 5.7 Hz, 1H), 5.74 (s, 1H), 5.52 – 5.17 (m, 5H), 4.81 (s, 1H), 4.65 – 4.46 (m, 5H), 4.40 (dd, *J* = 19.9, 8.3 Hz, 3H), 4.26 – 3.87 (m, 8H), 3.11 (dd, *J* = 13.6, 6.6 Hz, 1H), 2.91 (dd, *J* = 13.6, 5.3 Hz, 1H), 2.17 (s, 3H), 2.10 – 2.00 (m, 4H), 1.84 (s, 3H), 1.31 (d, *J* = 7.2 Hz, 3H), 1.25 (s, 9H), 1.01 (d, *J* = 5.6 Hz, 3H), 0.92 (dd, *J* = 14.0, 6.8 Hz, 6H).

¹³C NMR (CDCl₃, 150 MHz): δ 171.97, 170.61, 170.10, 169.62, 169.45, 169.29, 160.13, 156.68, 143.73, 141.24, 133.57, 133.31, 132.06, 131.53, 130.72, 128.99, 127.71, 127.05, 125.44, 125.02, 123.12, 119.98, 118.60, 83.95, 75.64, 69.27, 67.27, 67.21, 67.18, 66.69, 66.22, 65.84, 61.99, 57.57, 55.95, 53.77, 52.97, 48.26, 47.04, 44.50, 29.67, 29.25, 28.07, 20.74, 20.68, 20.34, 19.14, 18.03, 18.00.

HRMS (ESI): calc. for C₅₉H₇₃F₃N₆O₁₆SNa [M + Na]⁺, 1233.4648; found: 1233.4656.

25:

¹H NMR (CDCl₃, 600 MHz): δ 8.58 (s, 1H), 8.23 (d, *J* = 7.7 Hz, 1H), 7.76 (d, *J* = 7.5 Hz, 2H), 7.69 (d, *J* = 7.8 Hz, 1H), 7.62 – 7.51 (m, 3H), 7.39 (t, *J* = 7.4 Hz, 2H), 7.30 (t, *J* = 7.3 Hz, 2H), 7.23 (s, 1H), 6.67 (s, 1H), 5.83 (ddd, *J* = 16.2, 10.9, 5.7 Hz, 1H), 5.76 (s, 1H), 5.47 (s, 1H), 5.35 – 5.18 (m, 4H), 5.11 (s, 1H), 5.00 (dd, *J* = 10.4, 7.9 Hz, 1H), 4.81 (dd, *J* = 10.5, 3.4 Hz, 1H), 4.73 (d, *J* = 4.9 Hz, 1H), 4.60 – 4.46 (m, 5H), 4.45 – 4.34 (m, 3H), 4.34 – 4.20 (m, 3H), 4.17 – 4.01 (m, 5H), 3.93 (d, *J* = 4.7 Hz, 2H), 3.82 (t, *J* = 6.8 Hz, 1H), 3.02 (dd, *J* = 13.6, 6.7 Hz, 1H), 2.87 (dd, *J* = 13.5, 5.2 Hz, 1H), 2.18 – 2.15 (m, 1H), 2.14 (s, 3H), 2.11 (s, 3H), 2.04 (d, *J* = 3.0 Hz, 6H), 1.88 (s, 3H), 1.45 (s, 3H), 1.27 (d, *J* = 7.2 Hz, 3H), 1.25 (s, 9H), 0.98 (s, 3H), 0.91 (d, *J* = 6.7 Hz, 3H), 0.89 (d, *J* = 6.7 Hz, 3H).

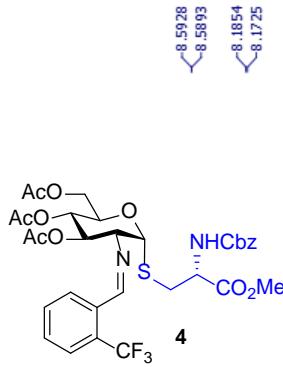
^{13}C NMR (CDCl₃, 150 MHz): δ 171.99, 170.72, 170.65, 170.37, 170.25, 170.04, 169.96, 169.36, 169.21, 168.73, 159.88, 143.73, 141.24, 133.33, 132.03, 131.48, 130.83, 128.71, 127.73, 127.07, 125.03, 120.00, 118.61, 100.65, 84.10, 75.59, 75.57, 70.77, 70.58, 69.14, 68.81, 68.39, 68.31, 67.28, 66.64, 66.22, 65.82, 62.87, 60.87, 58.27, 57.41, 55.94, 53.76, 52.95, 48.23, 47.04, 44.52, 29.66, 29.24, 27.96, 20.77, 20.67, 20.61, 20.46, 19.90, 19.15, 17.99, 17.91.

HRMS (ESI): calc. for C₇₁H₈₉F₃N₆O₂₄SNa [M + Na]⁺, 1521.5493; found: 1521.5498.

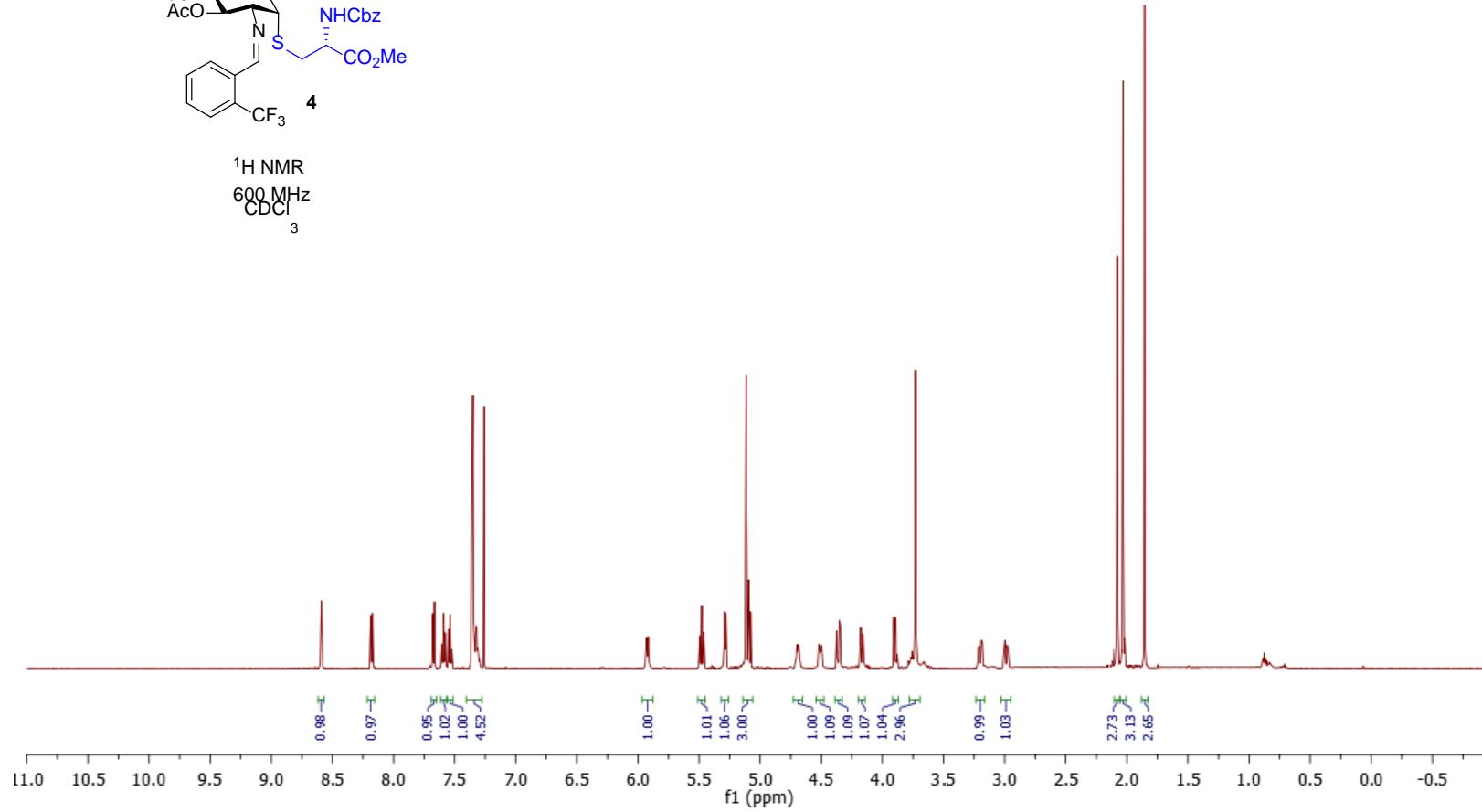
9. References

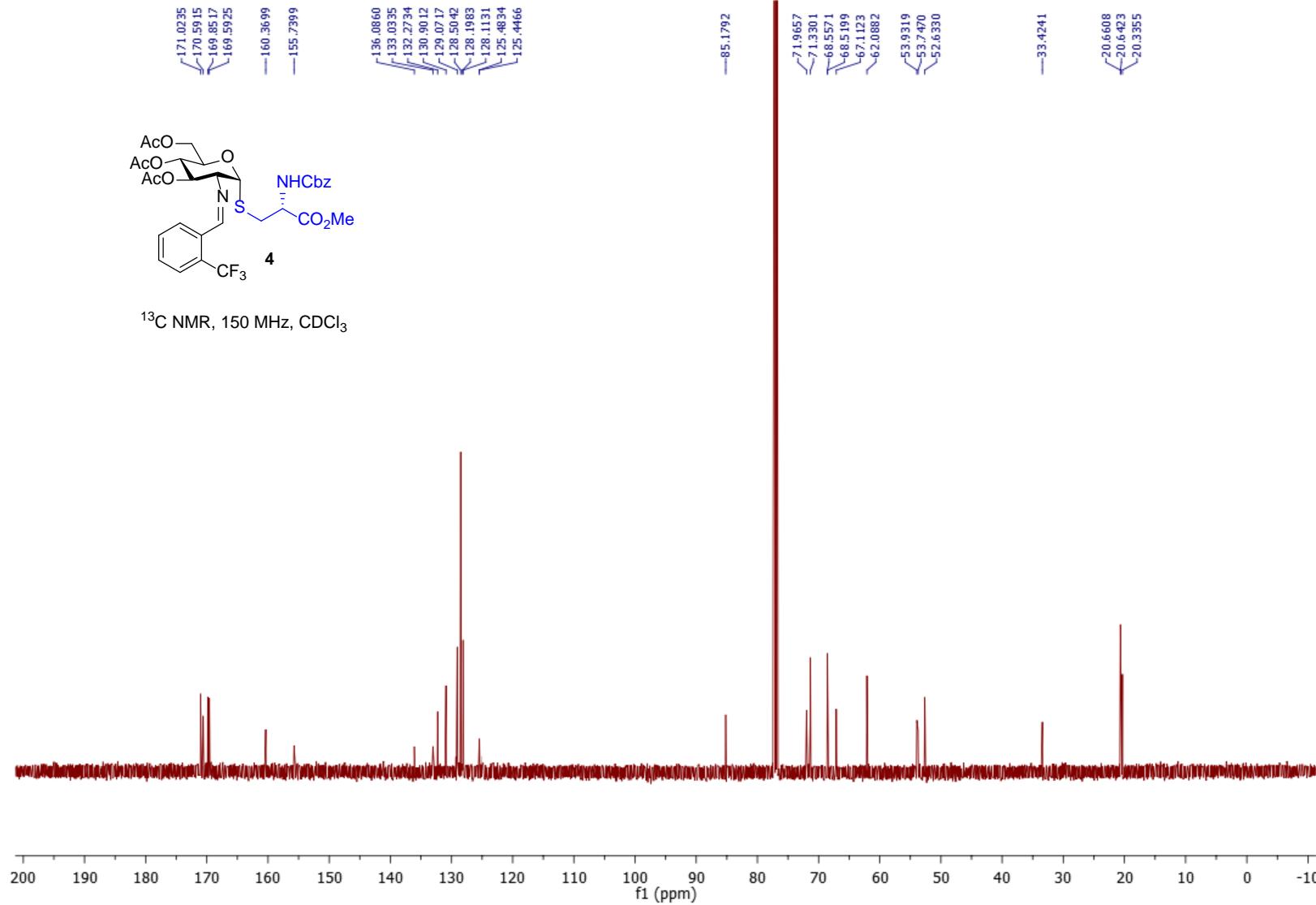
- [1] E. T. Sletten, Y.-J. Tu, H. B. Schlegel, H. M. Nguyen, *ACS Catal.* **2019**, *9*, 2110.

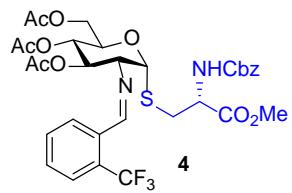
10. Spectral Data



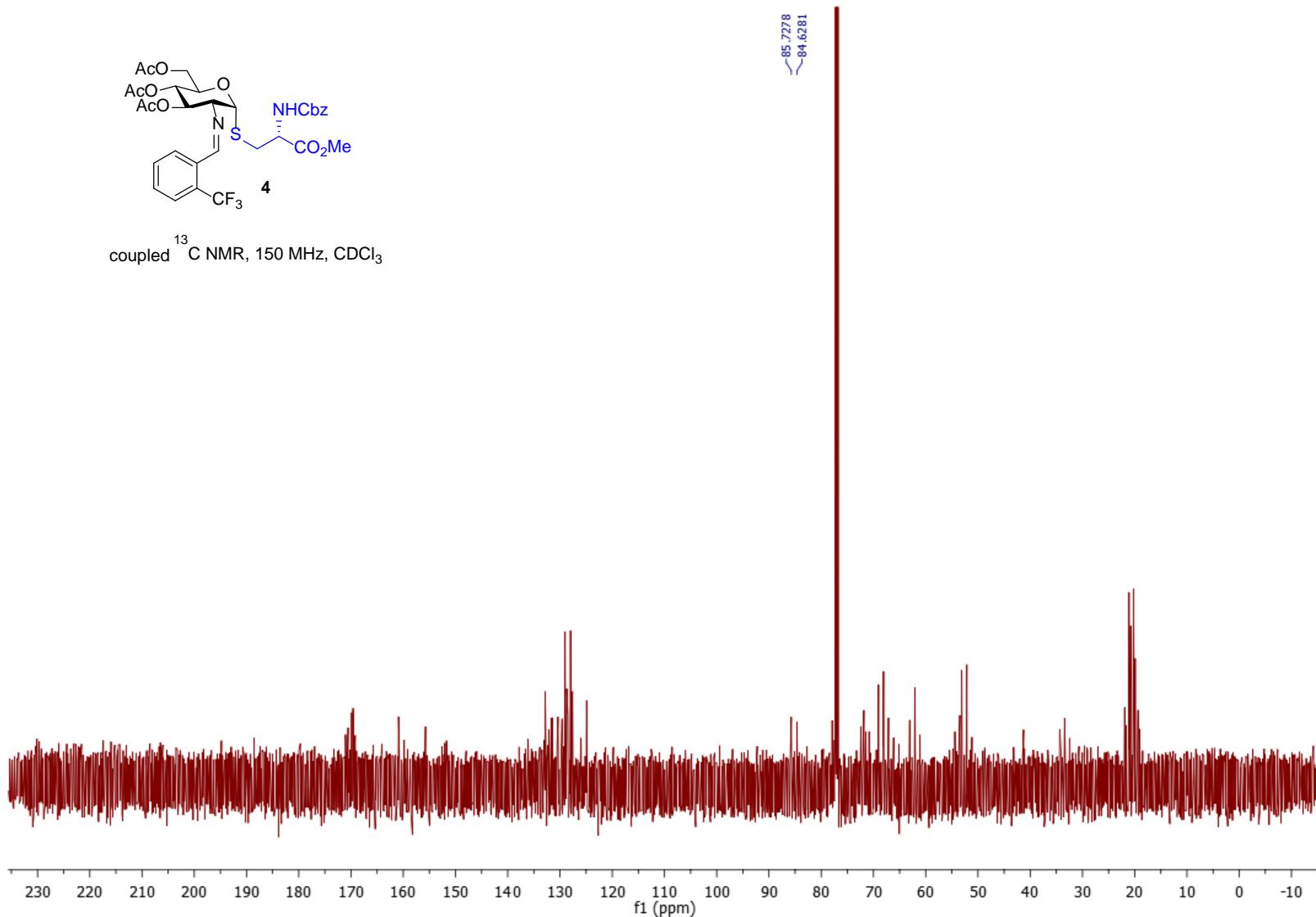
¹H NMR
600 MHz
CDCl₃

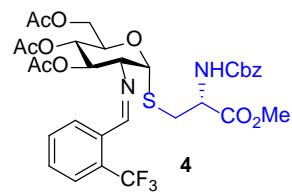




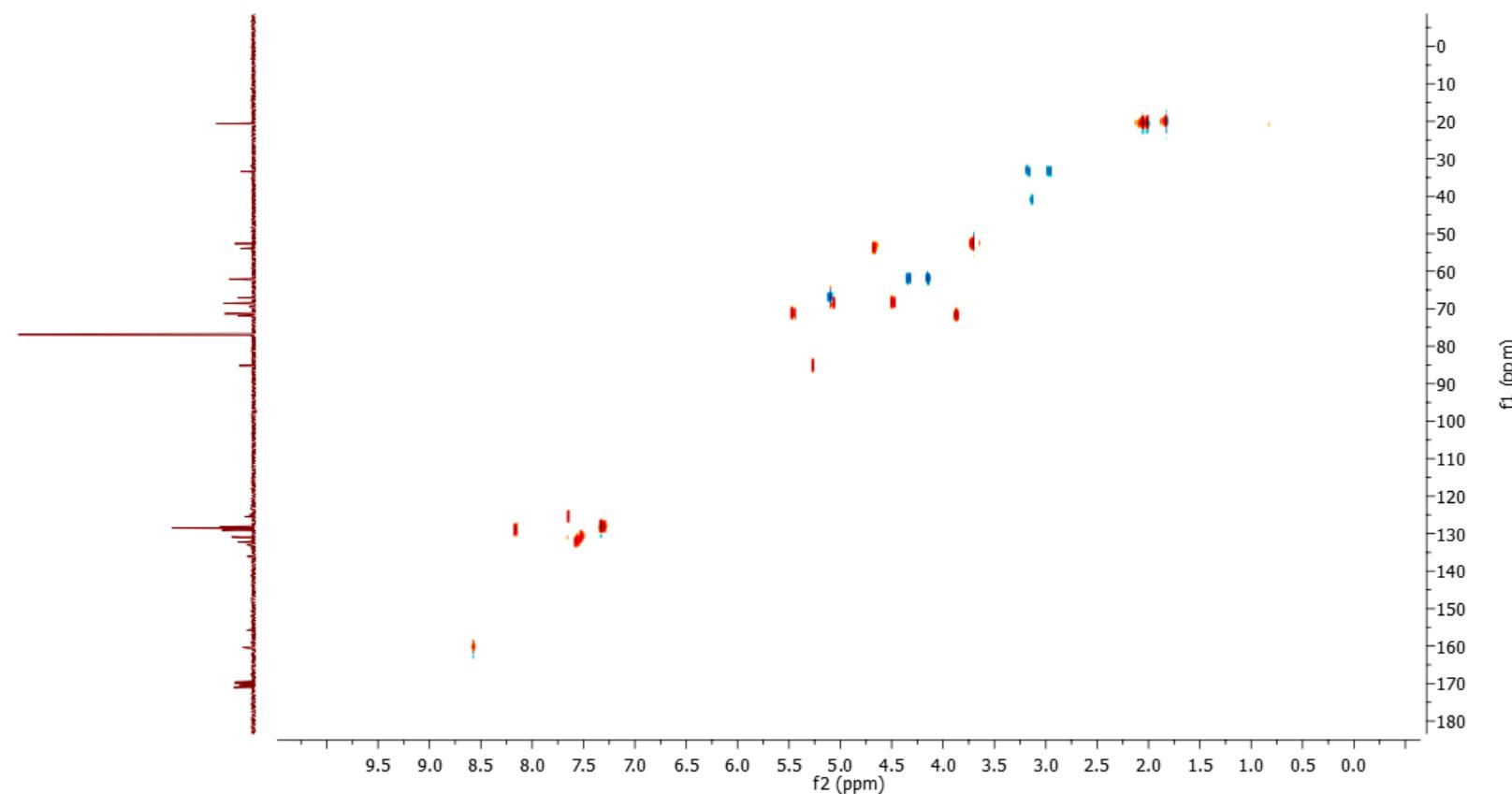


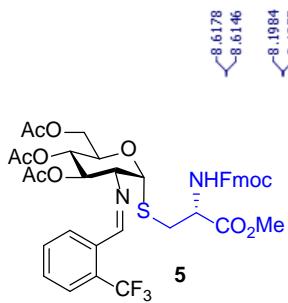
coupled ¹³C NMR, 150 MHz, CDCl₃





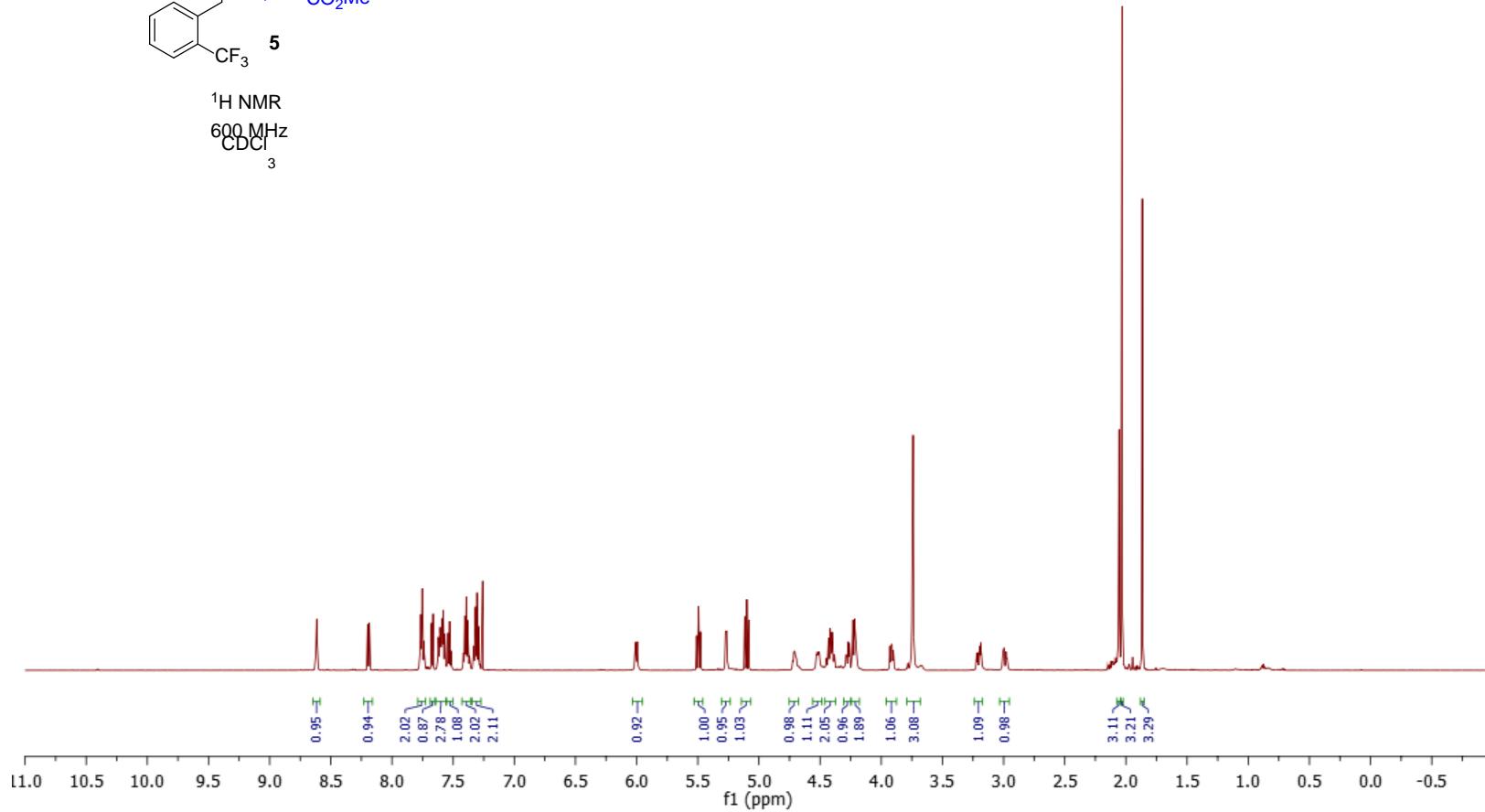
^1H - ^{13}C HSQC, 600/150MHz, CDCl_3

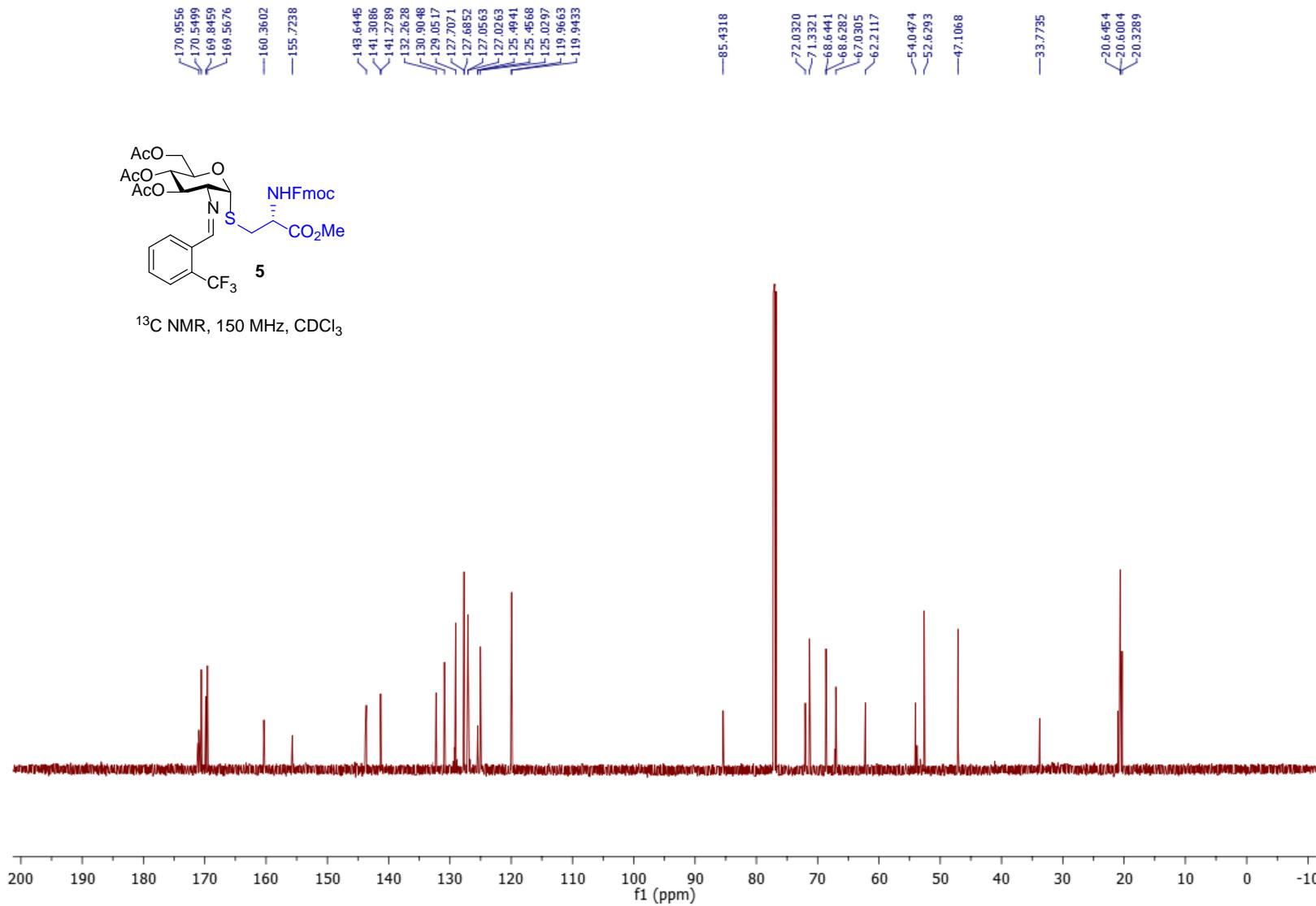


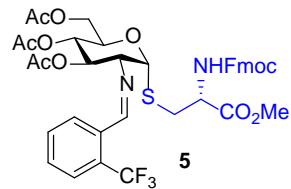


¹H NMR

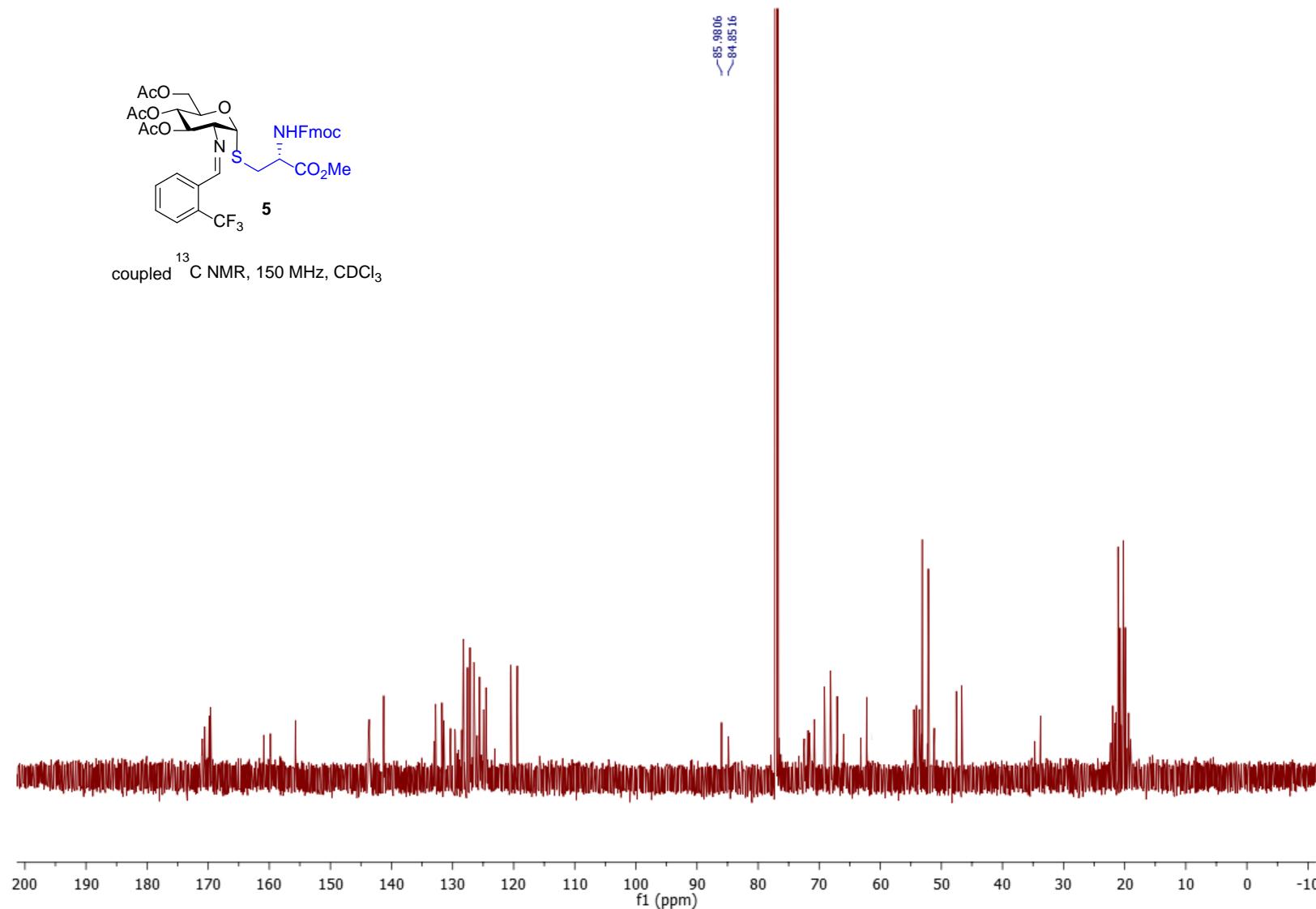
600 MHz

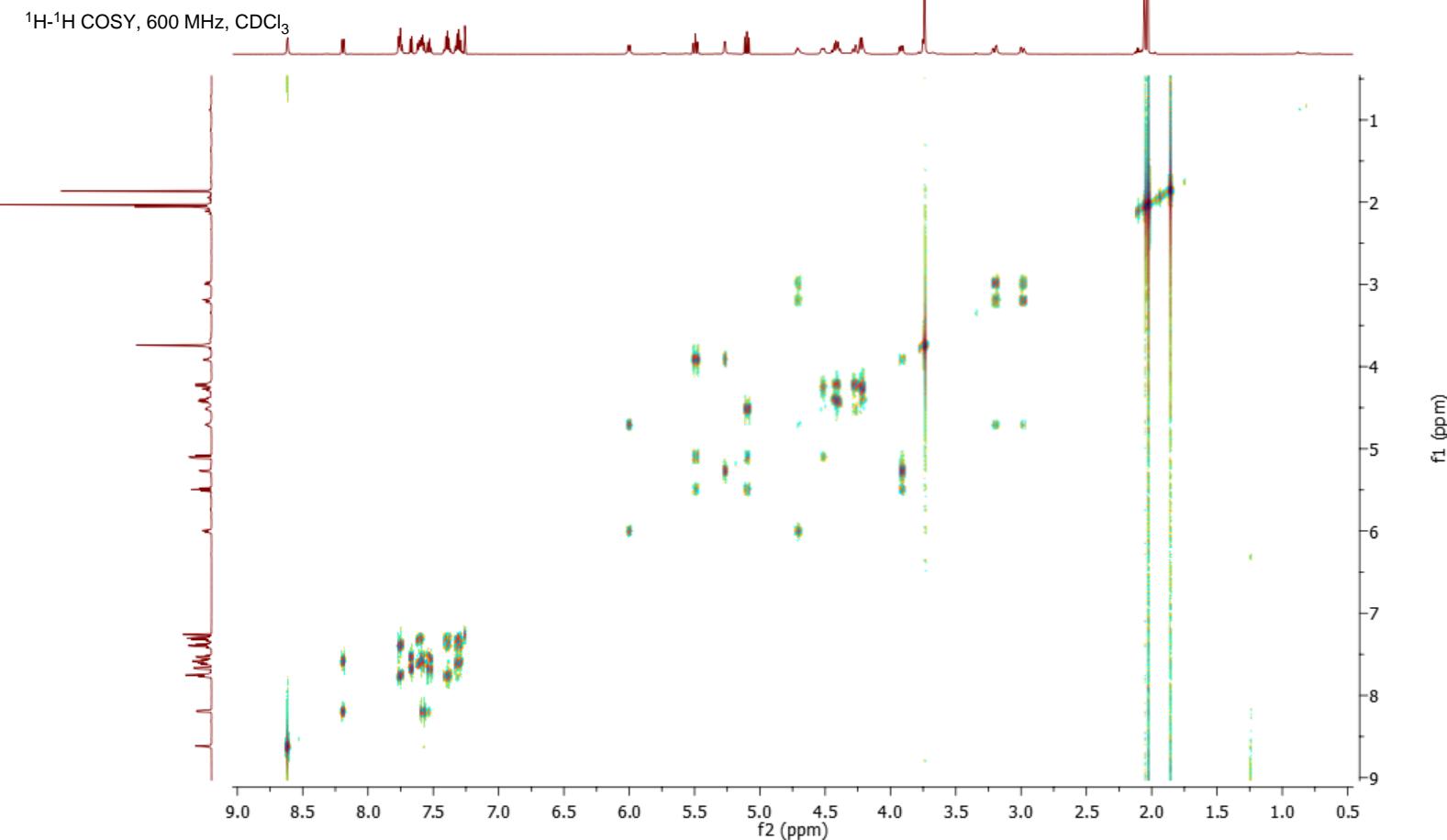
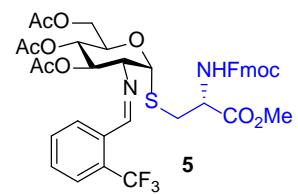


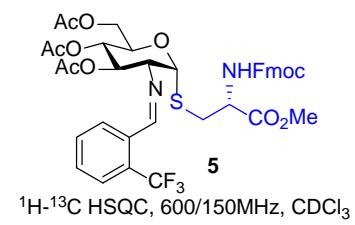




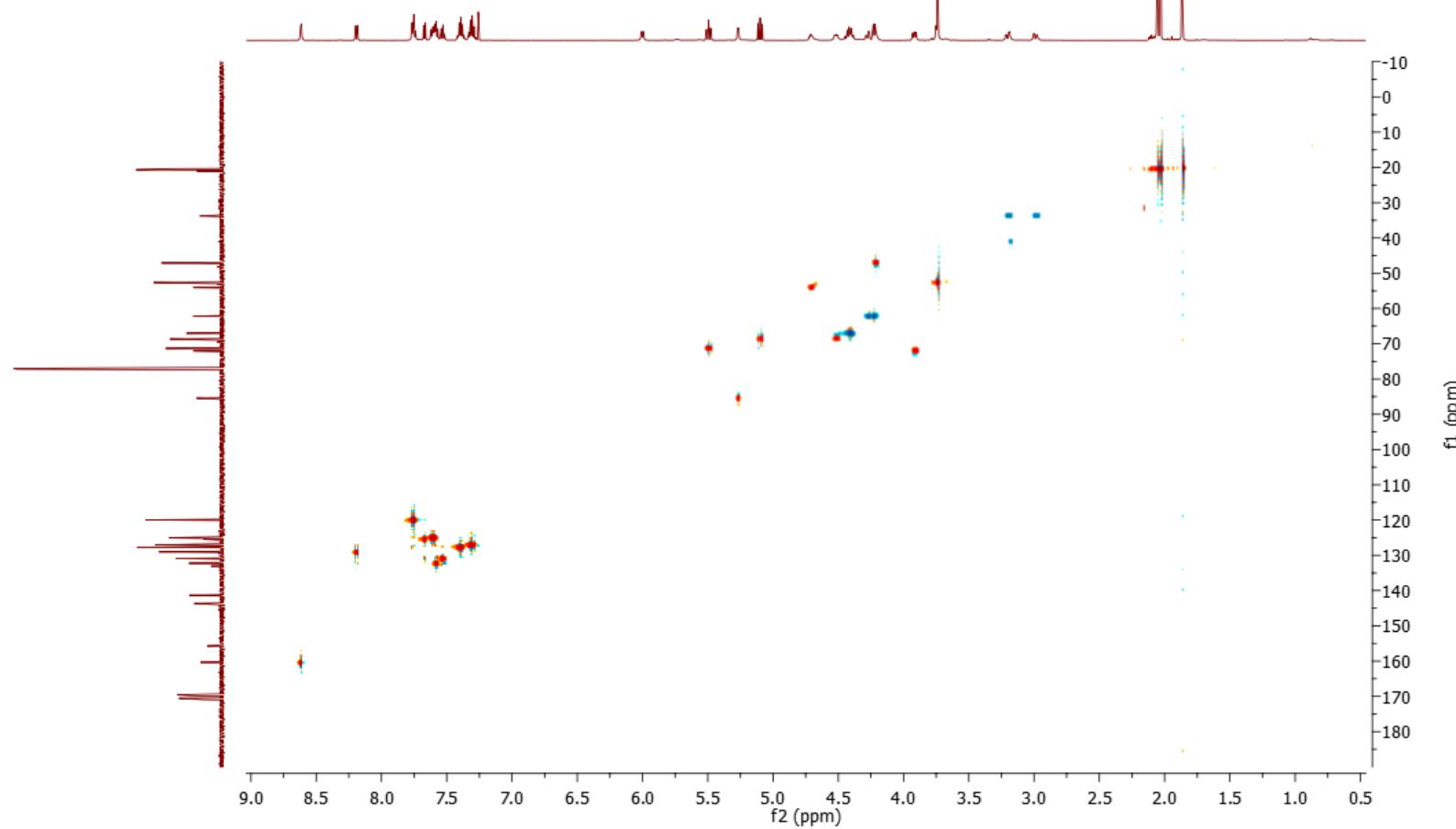
coupled ¹³C NMR, 150 MHz, CDCl₃

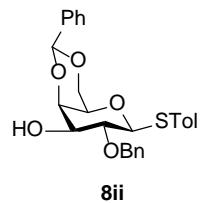




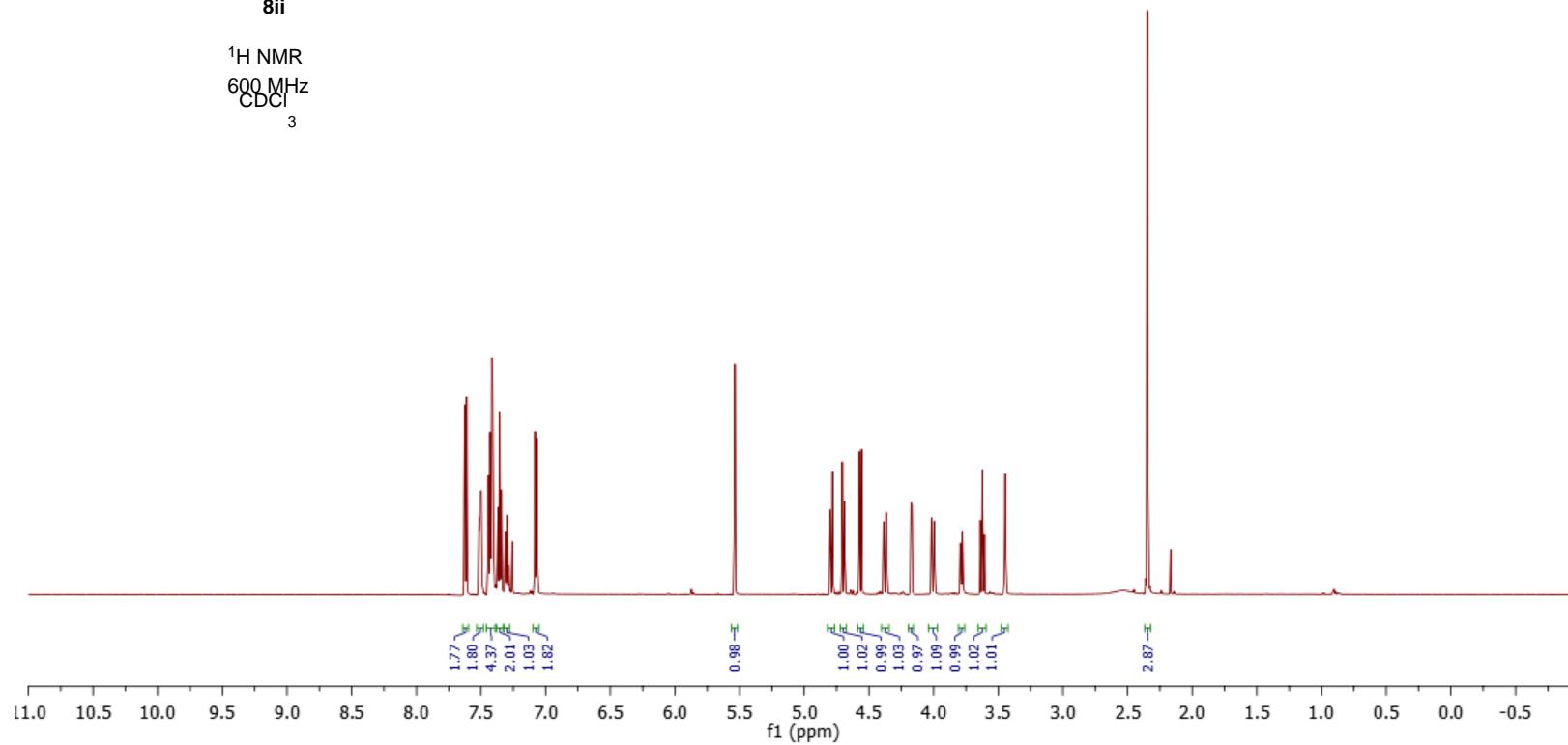


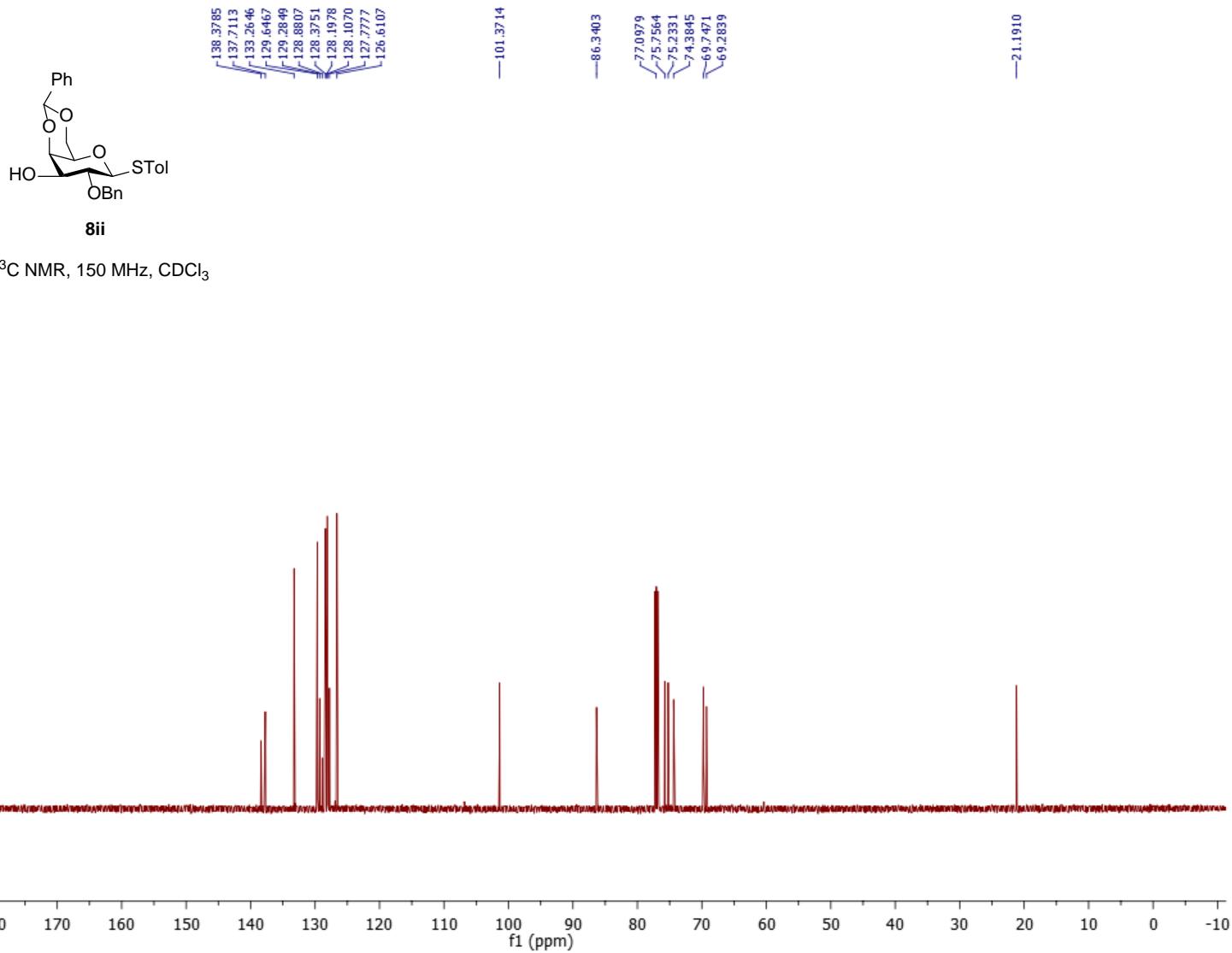
^1H - ^{13}C HSQC, 600/150MHz, CDCl_3

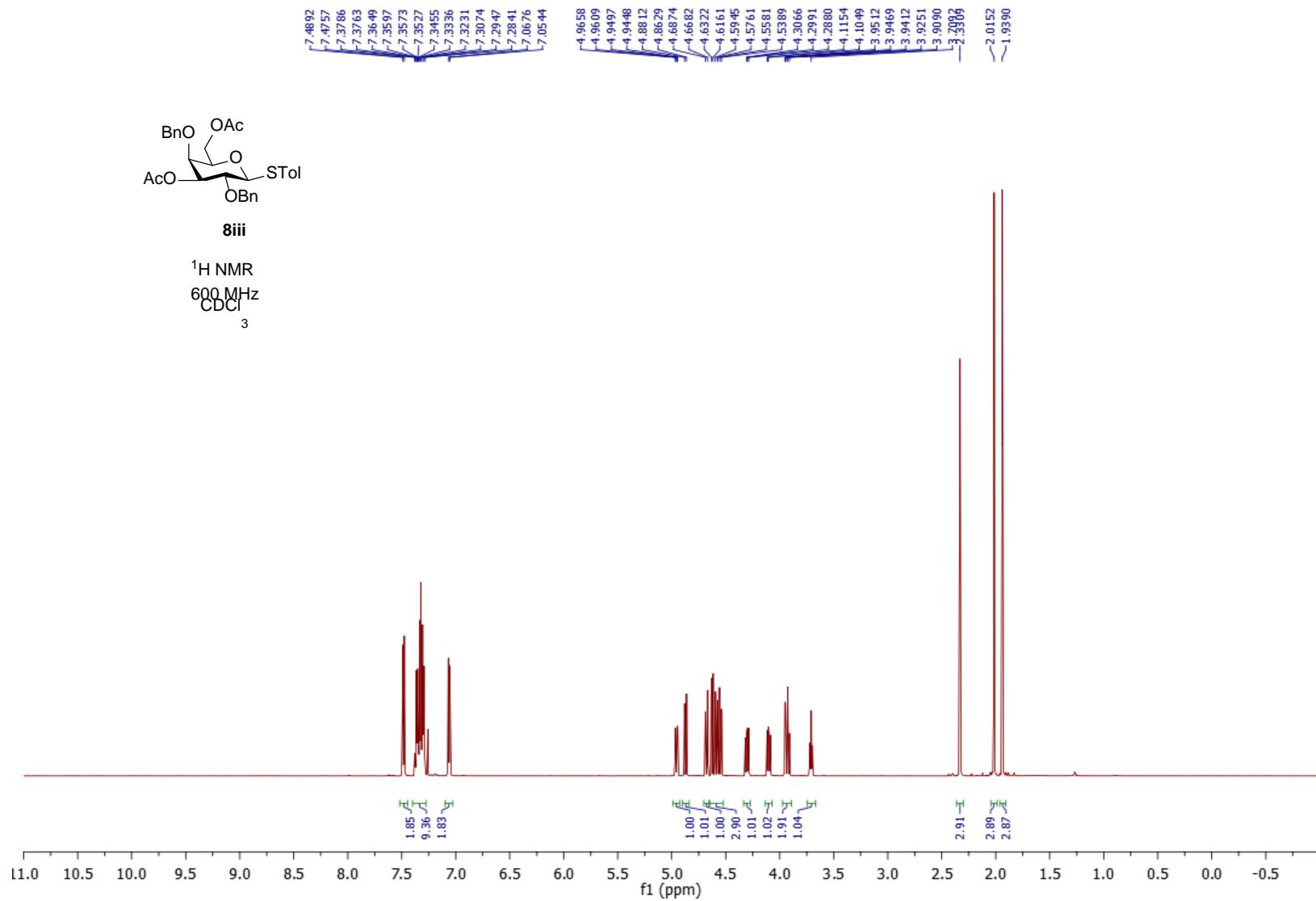


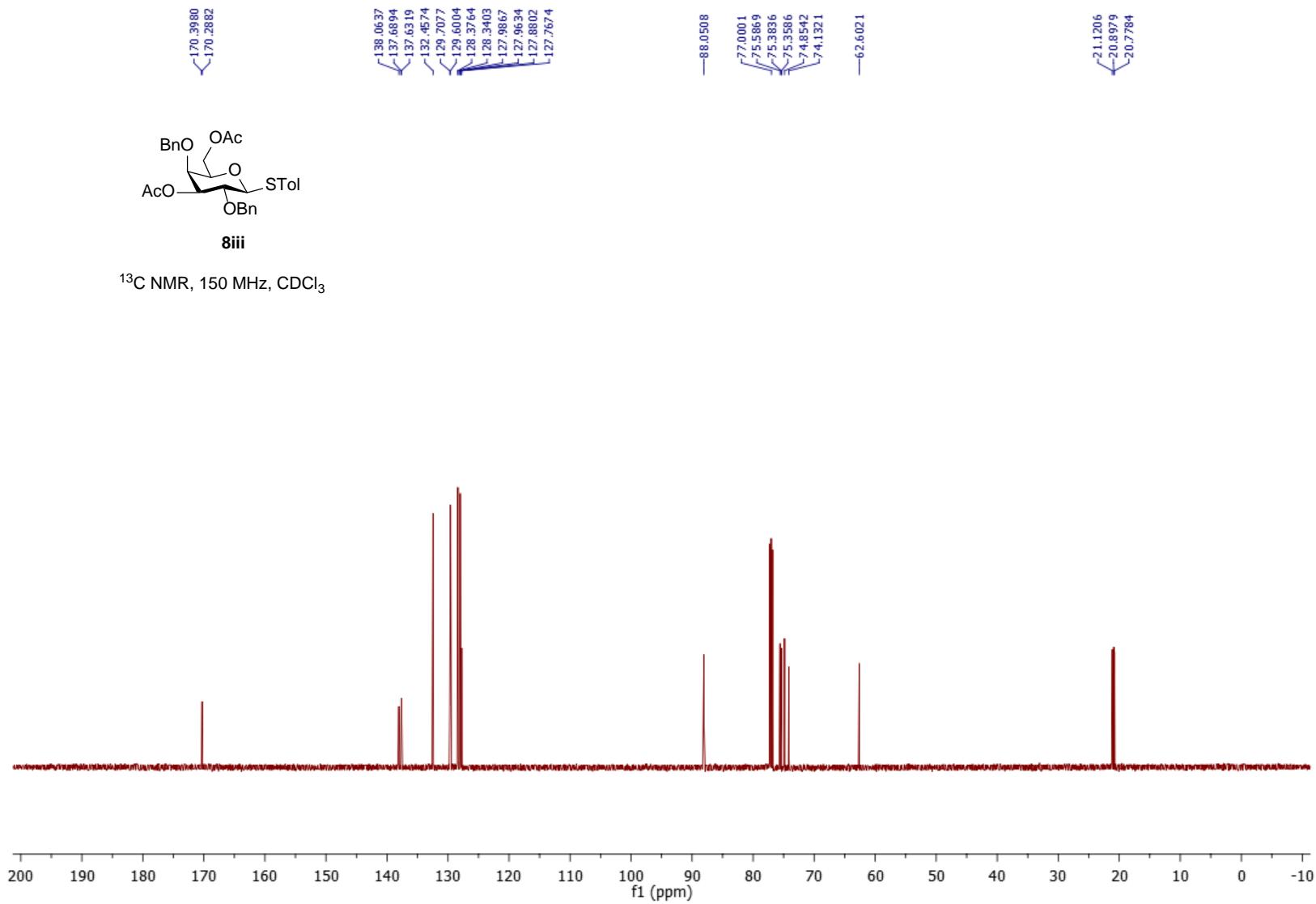


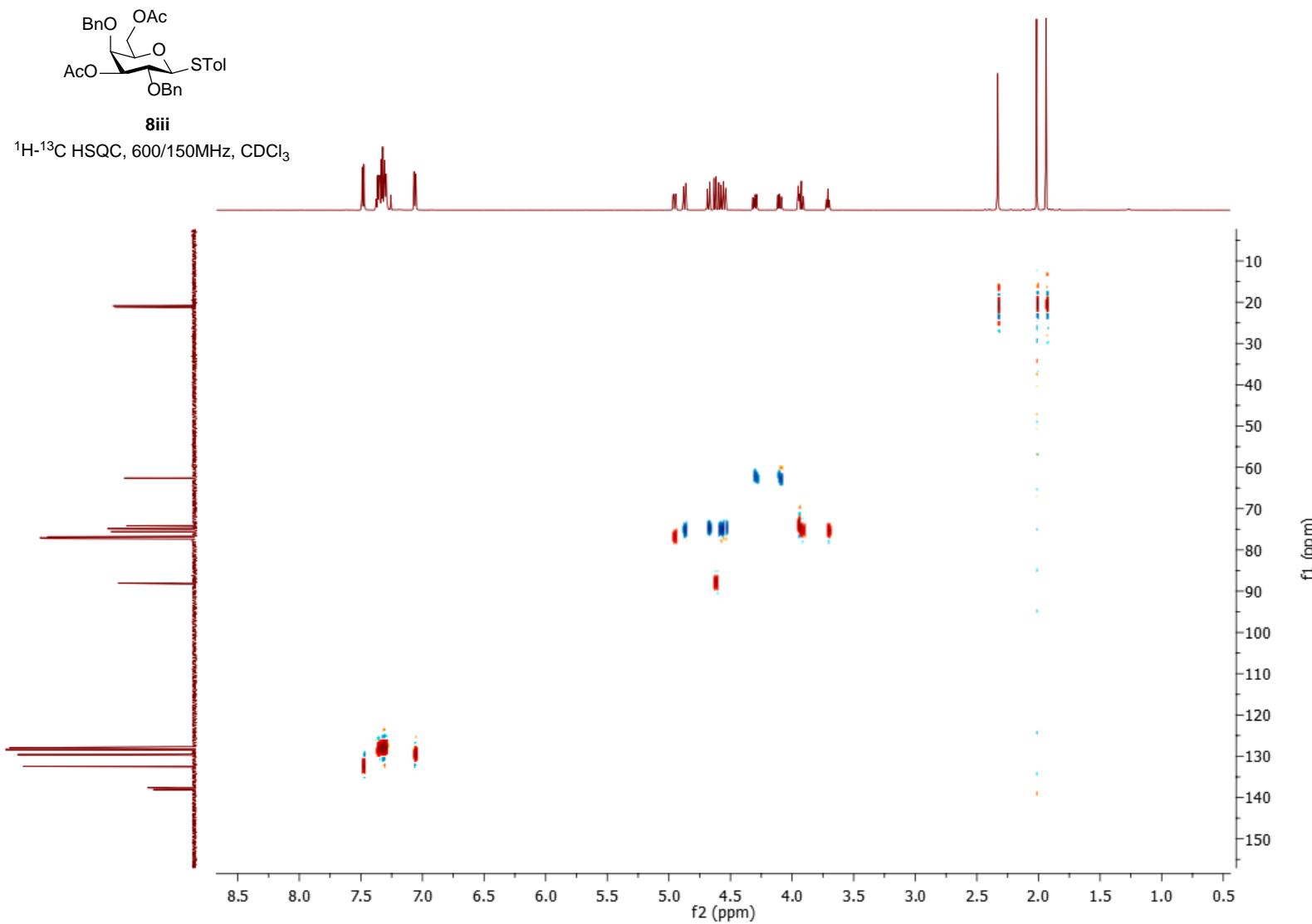
¹H NMR
600 MHz
CDCl₃

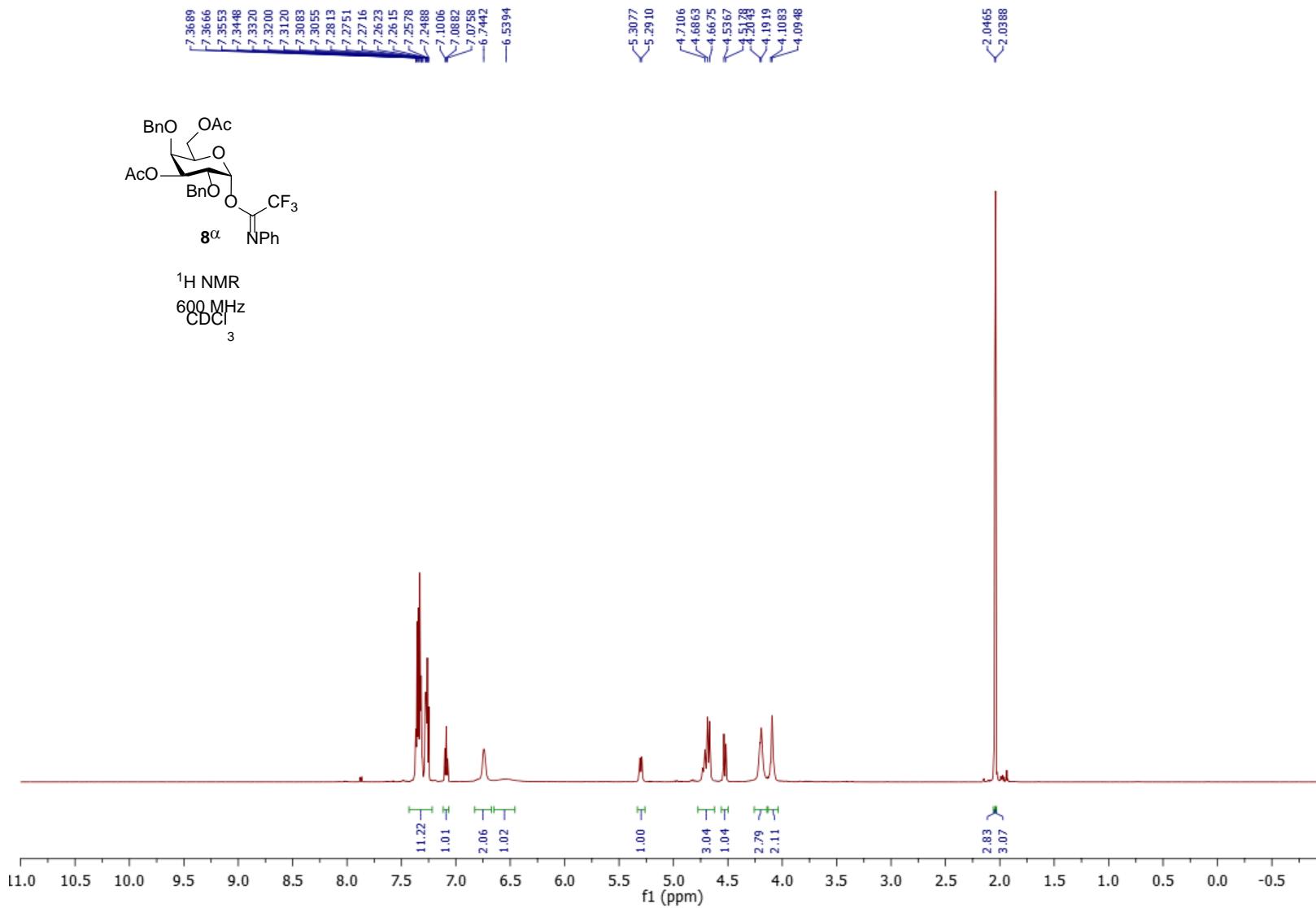


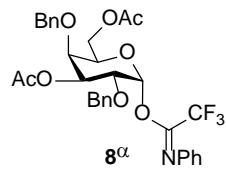




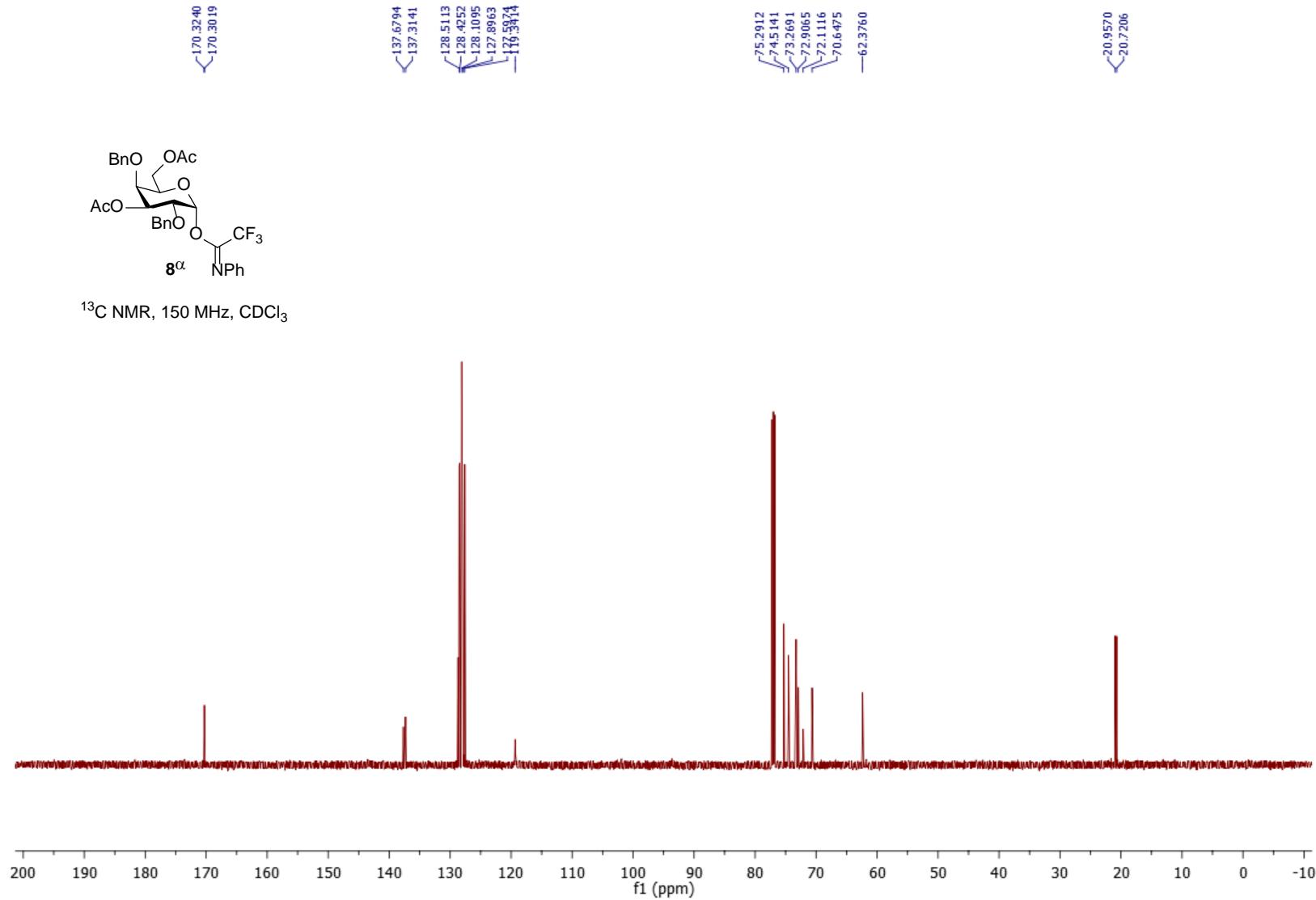


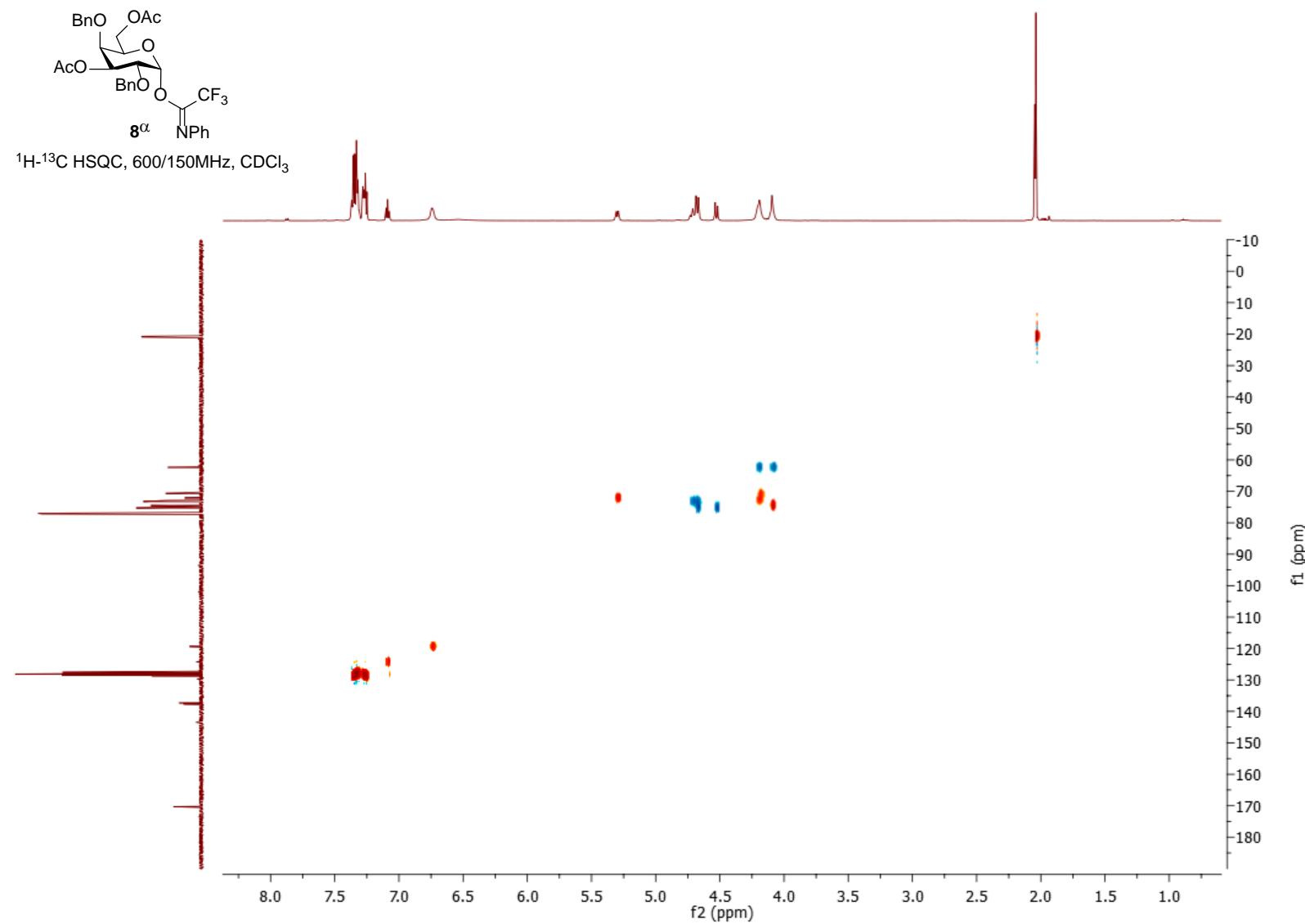


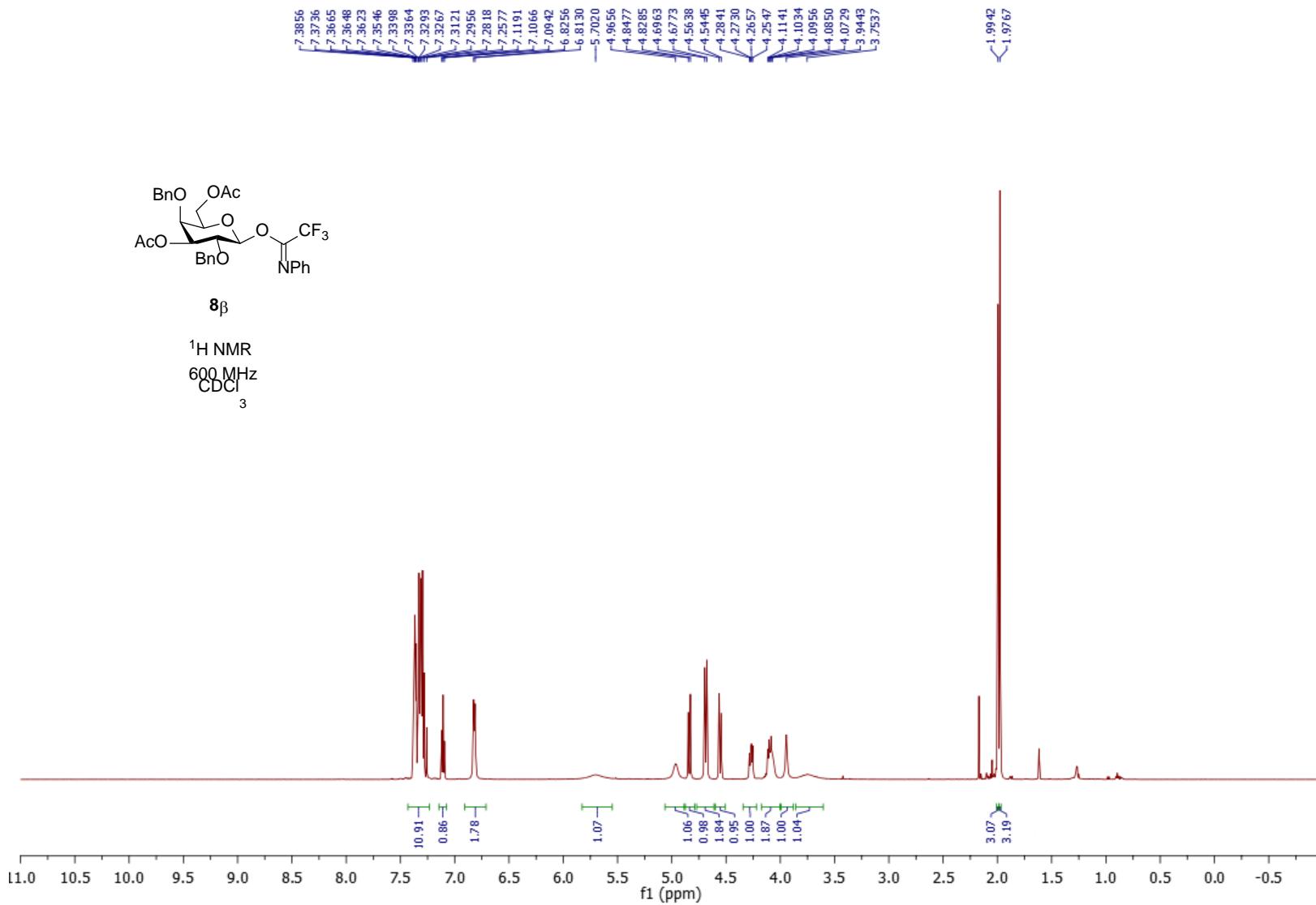


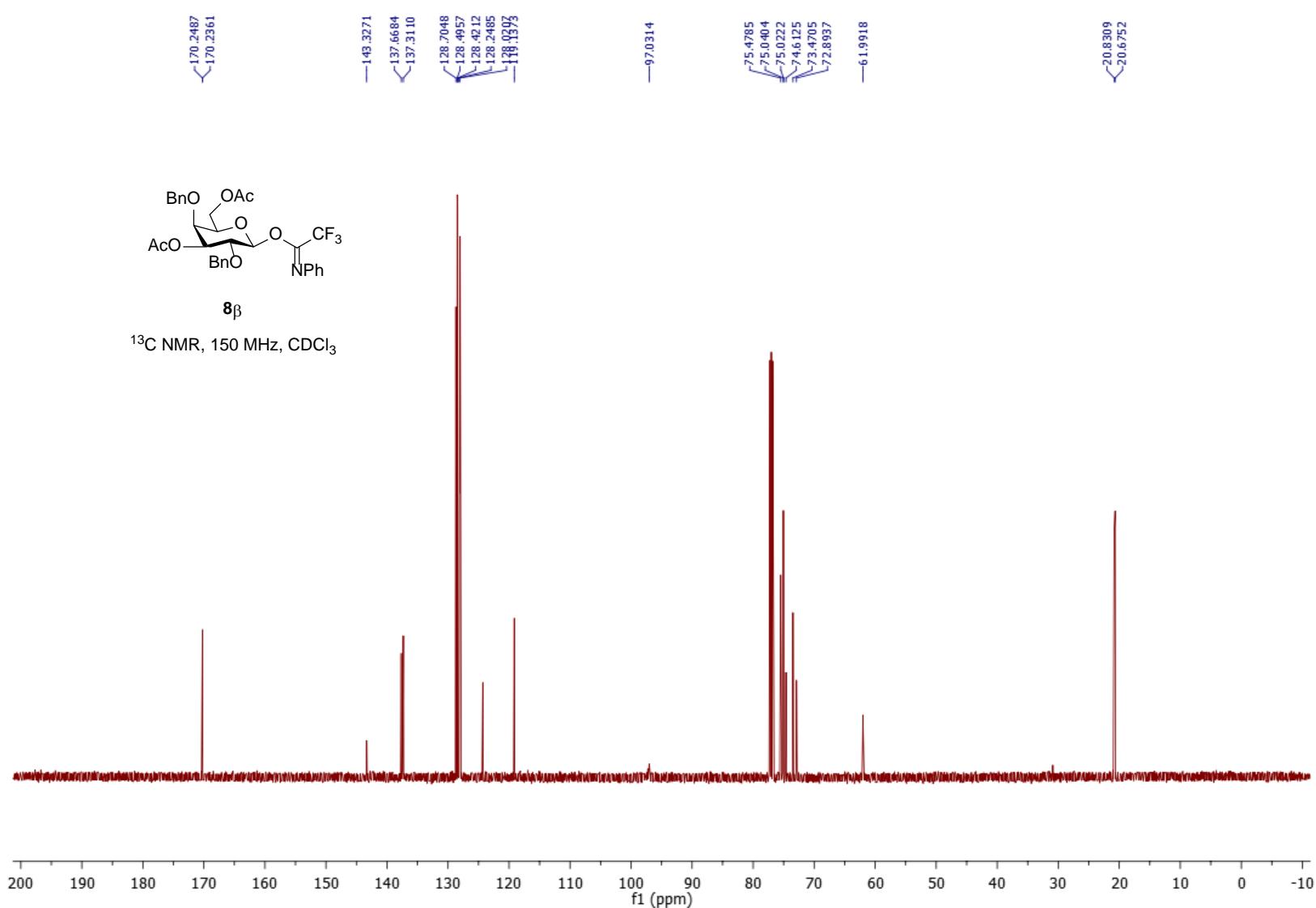


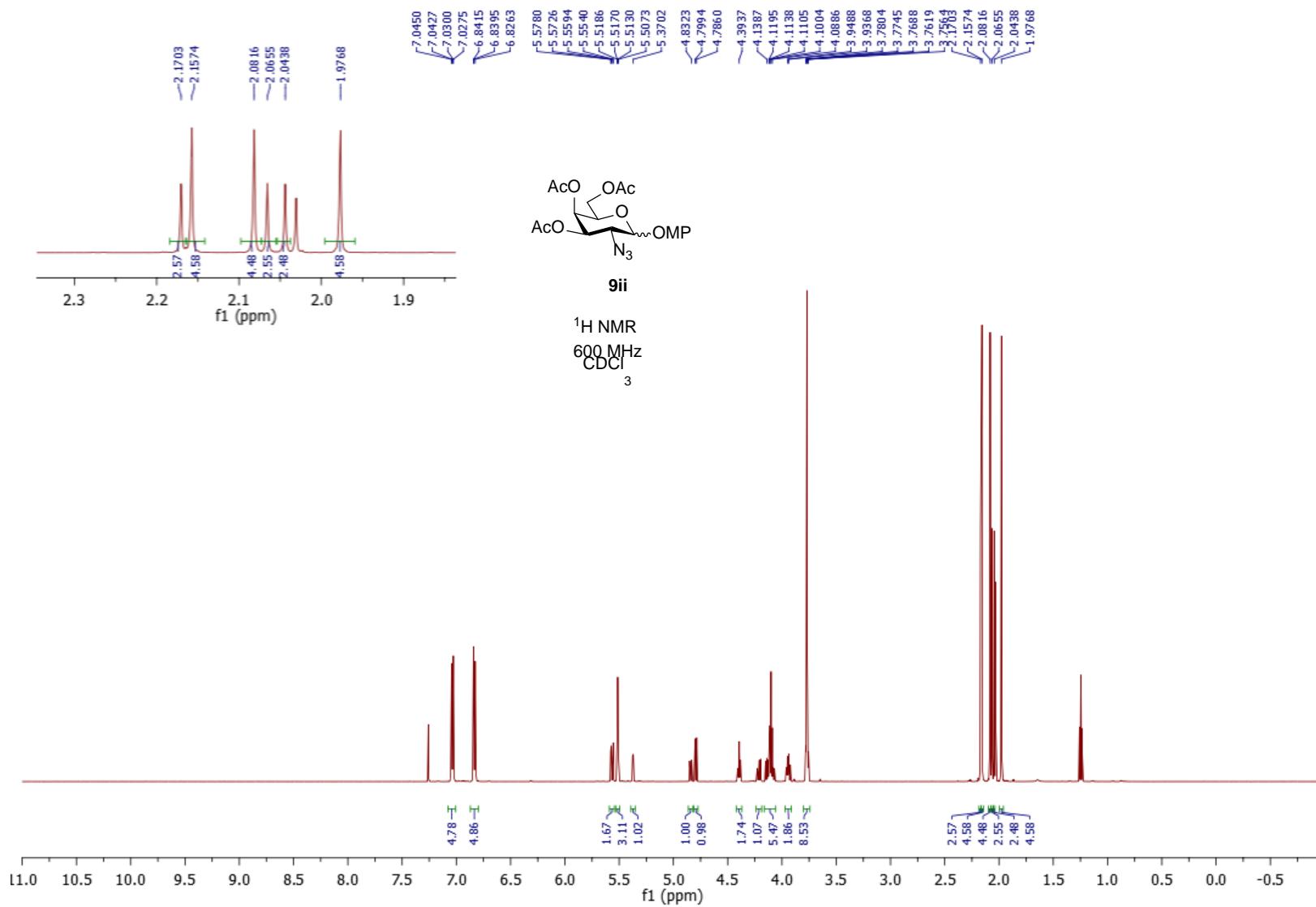
¹³C NMR, 150 MHz, CDCl₃









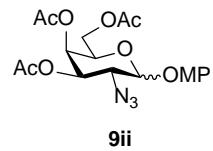


170.2606
170.0122
169.9595
169.7774
169.7266
155.9075
155.6280
150.7770
150.1889

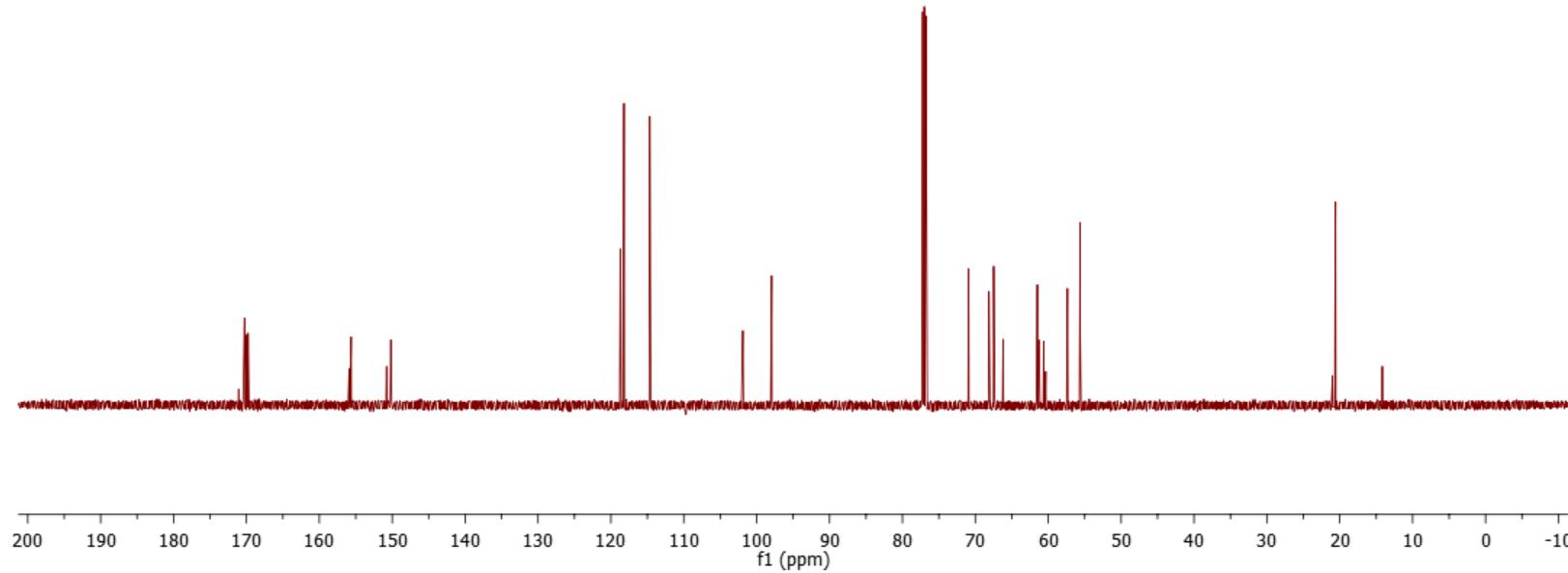
118.7053
118.2065
114.6729
114.5847

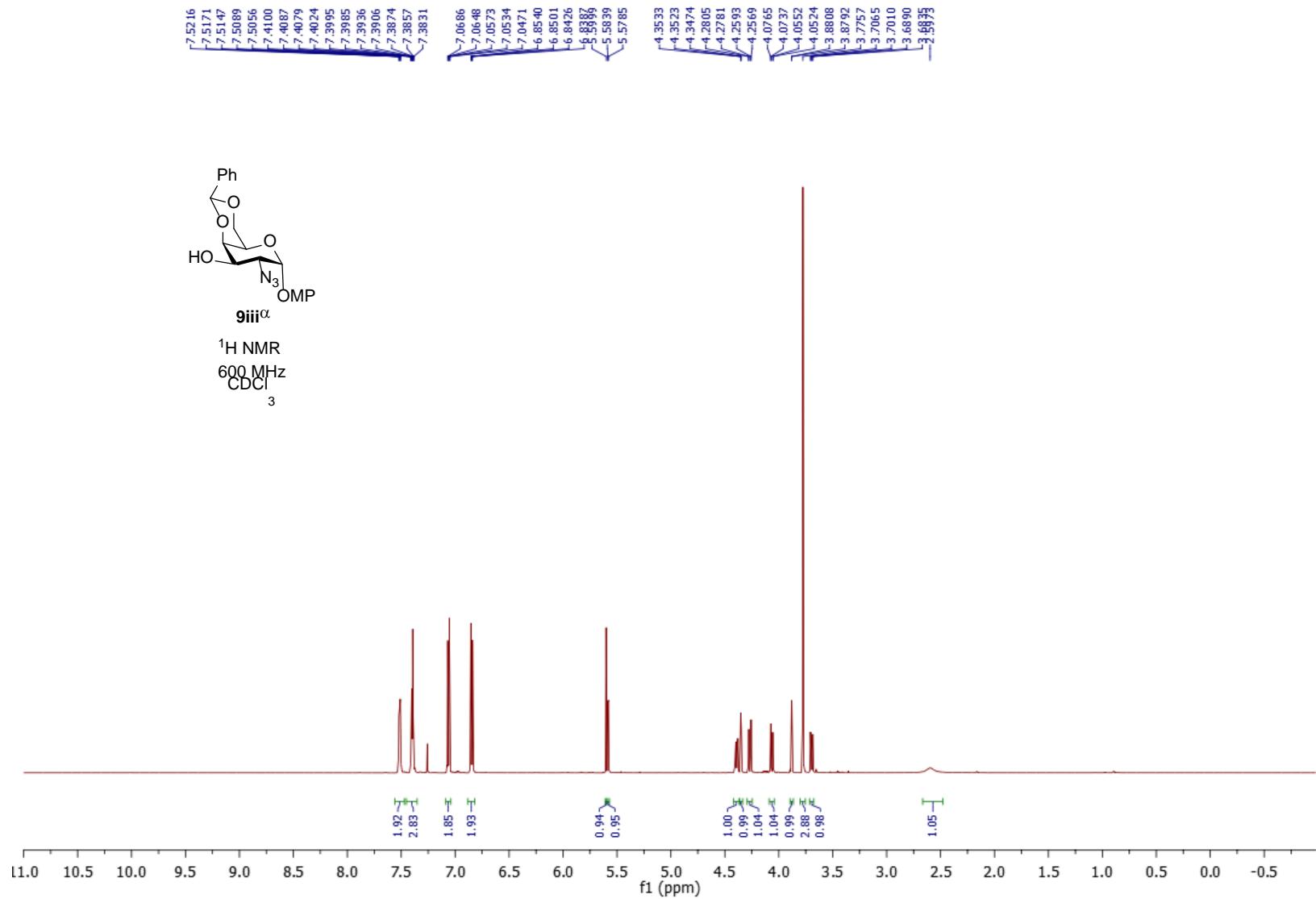
— 101.8931
— 97.9363

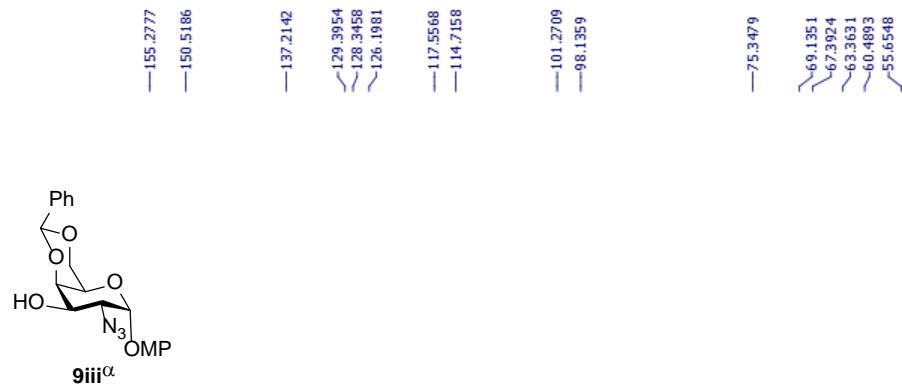
70.9167
68.1305
67.4788
67.3689
66.1817
61.4936
61.2548
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60.3391
57.3542
55.6235
20.6327
20.5810
20.5718



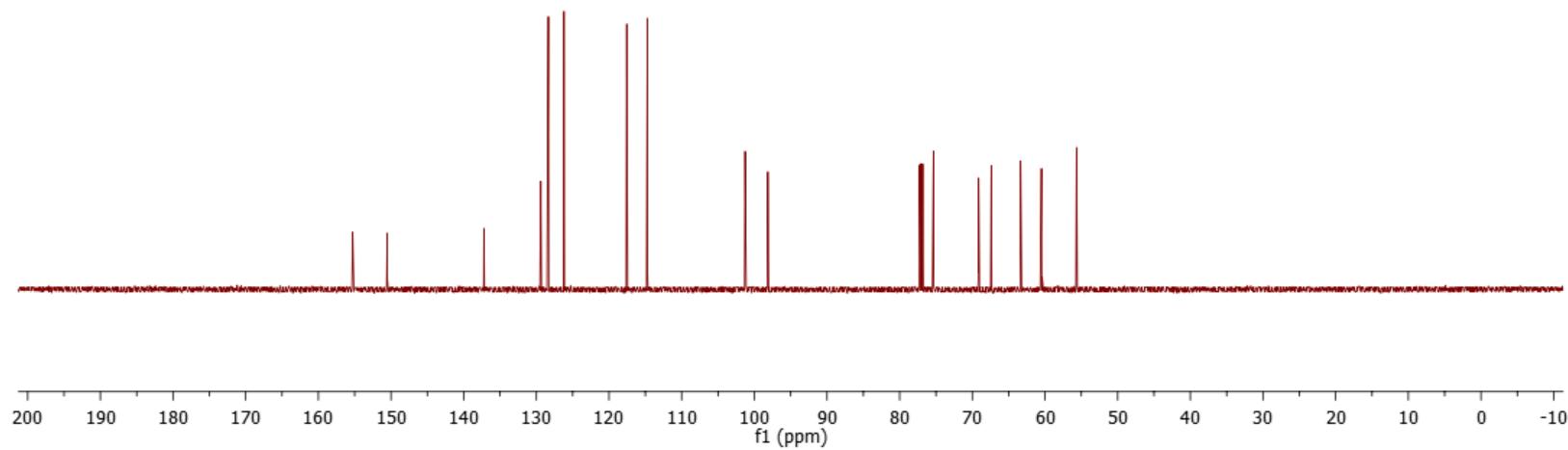
¹³C NMR, 150 MHz, CDCl₃

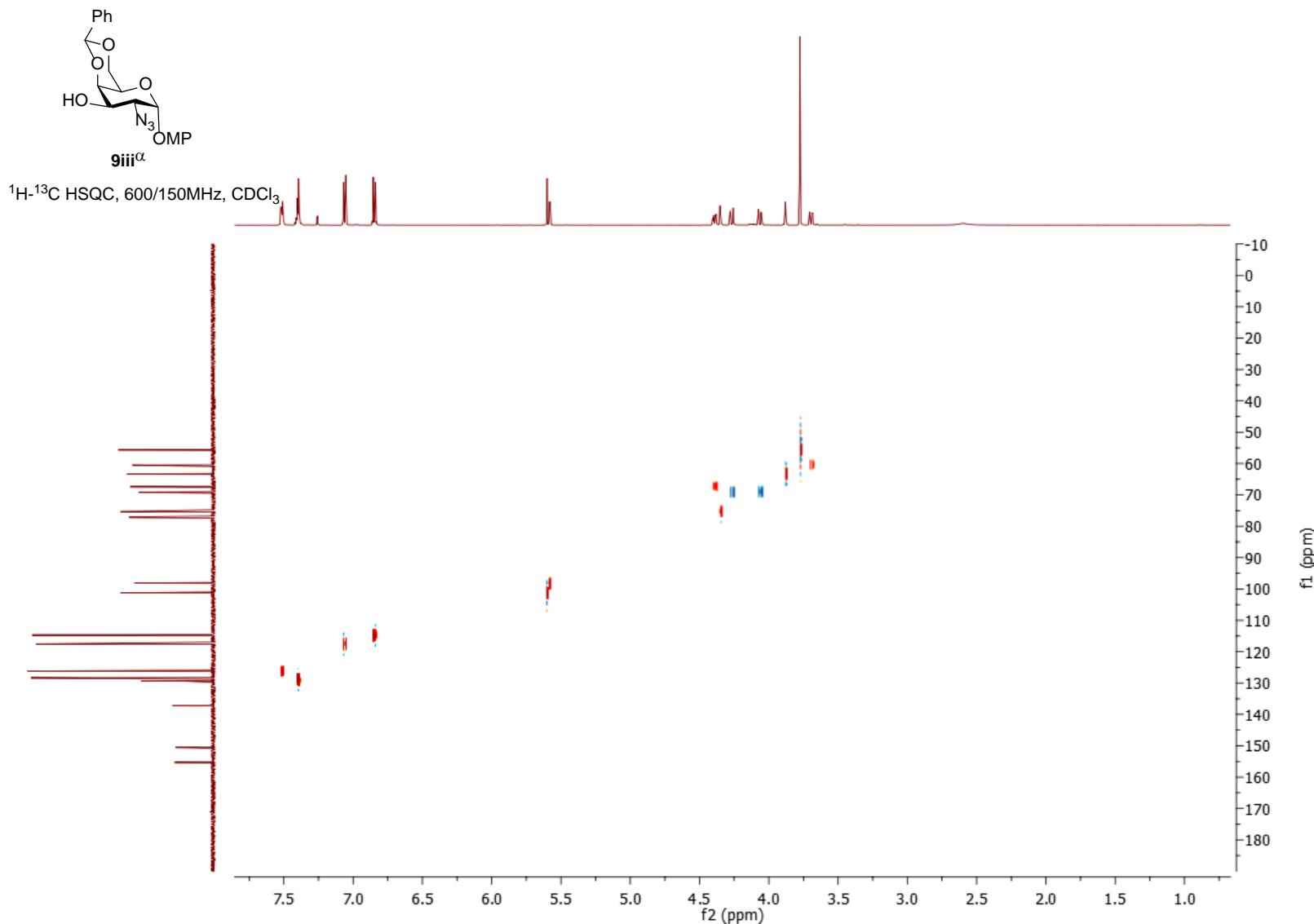


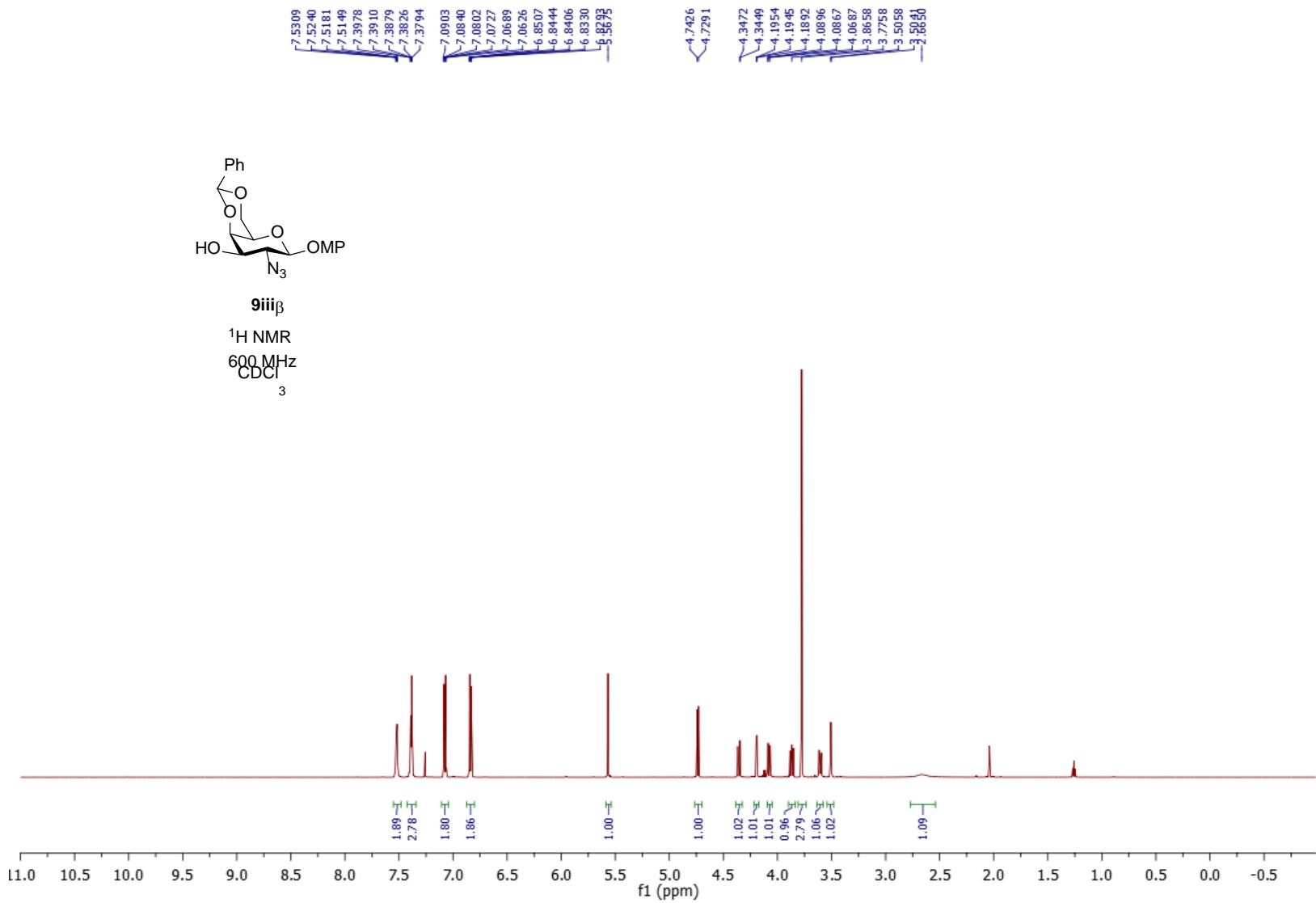




^{13}C NMR, 150 MHz, CDCl_3

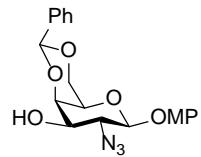






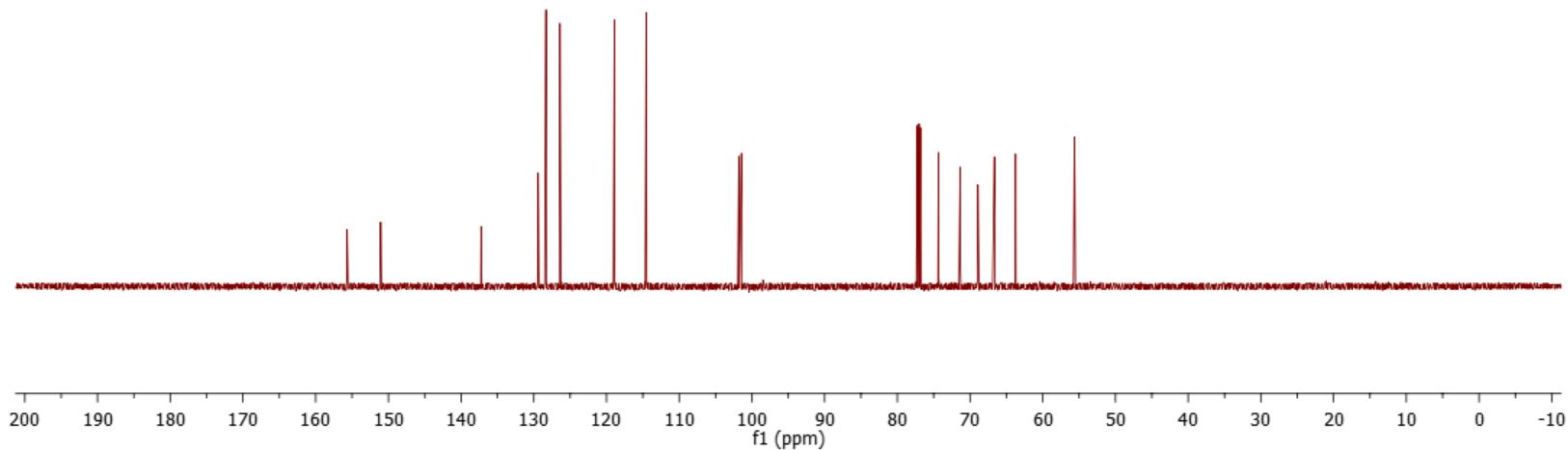
—155.6796
—151.0620
—137.2369
—129.4051
—>128.3103
—>126.4082
—118.9022
—114.5308

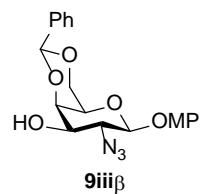
 $\swarrow^{101.7864}$
 $\searrow_{101.4411}$



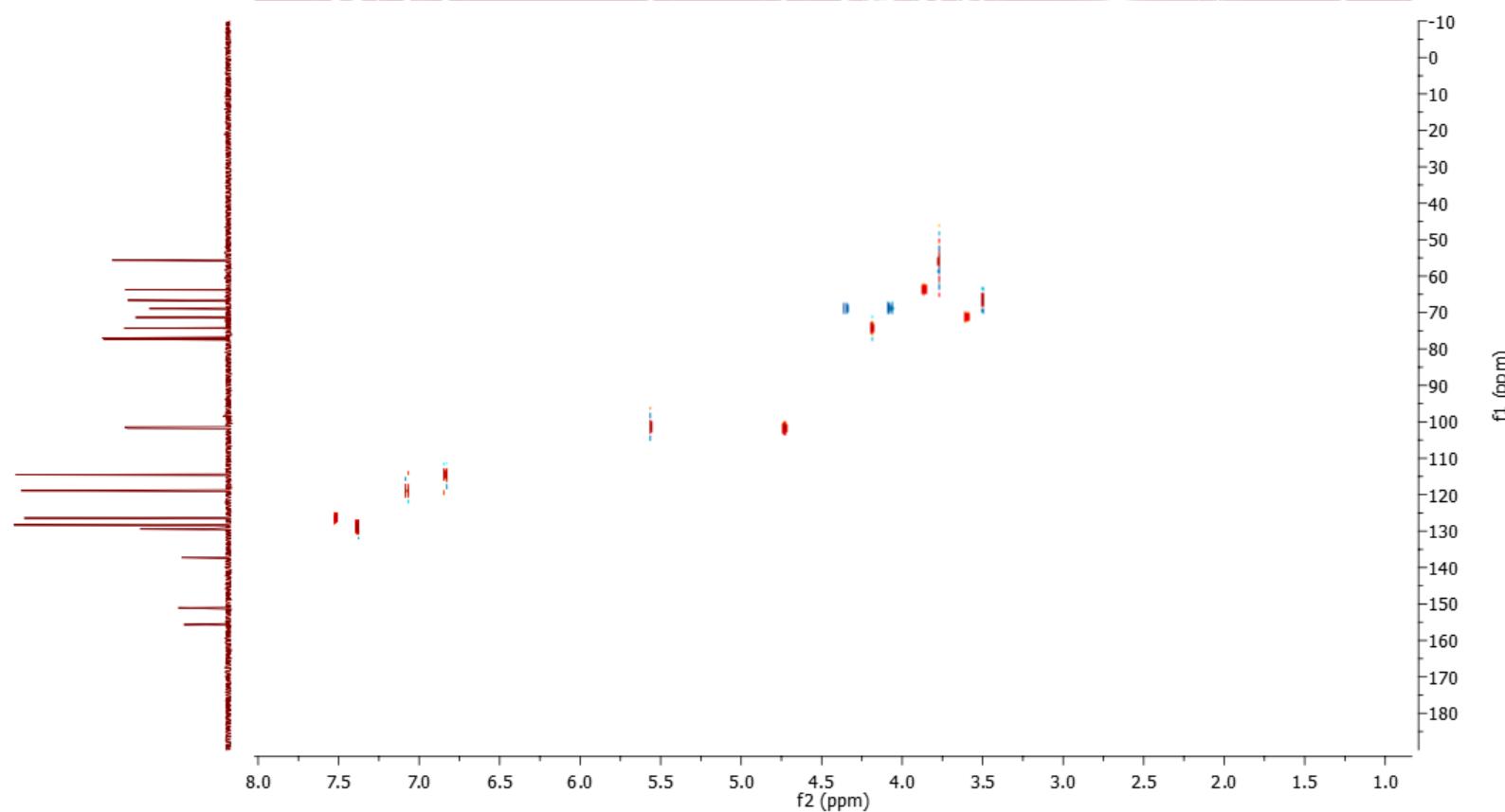
9iii β

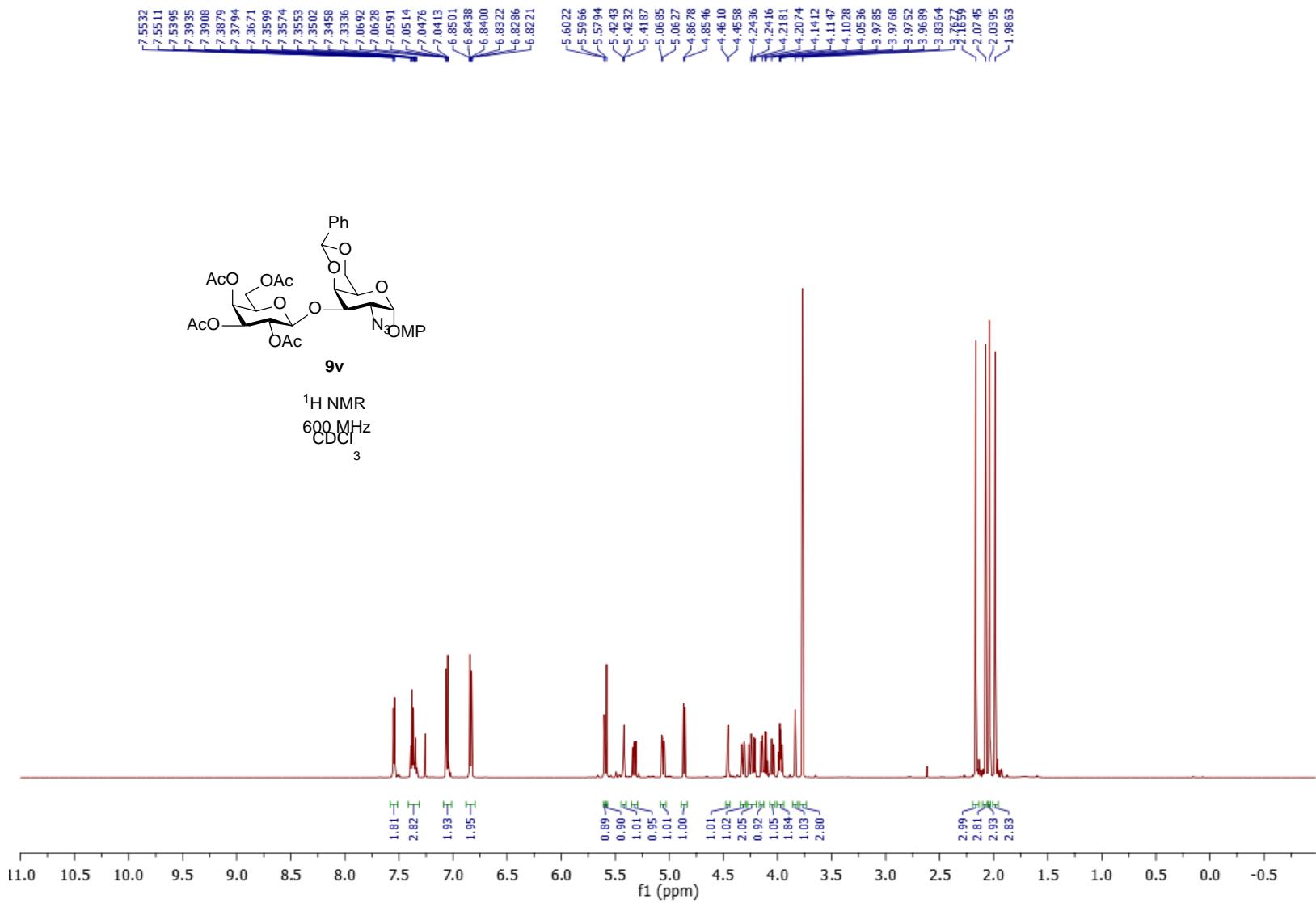
^{13}C NMR, 150 MHz, CDCl_3

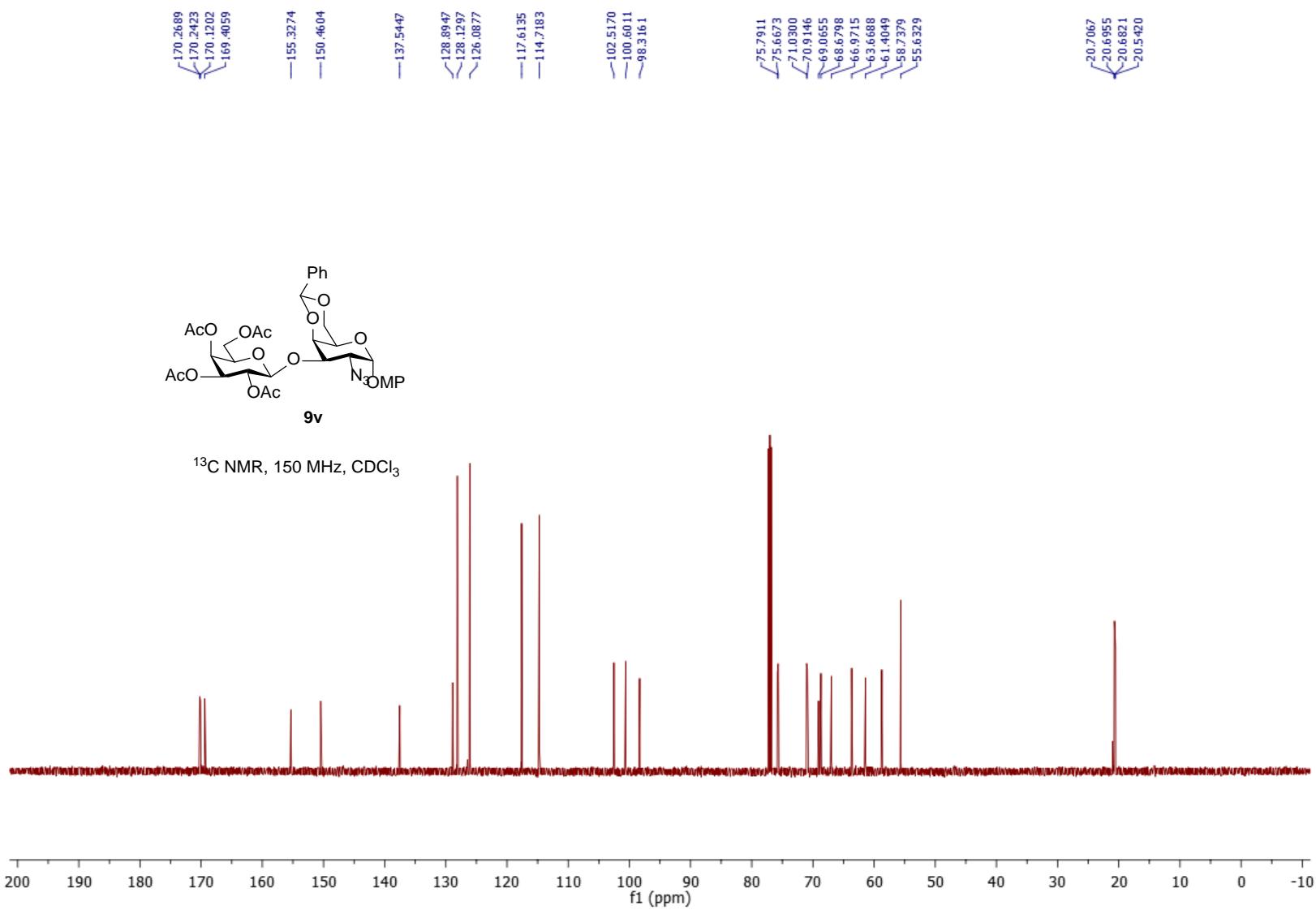


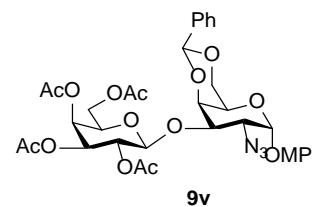


¹H-¹³C HSQC, 600/150MHz, CDCl₃

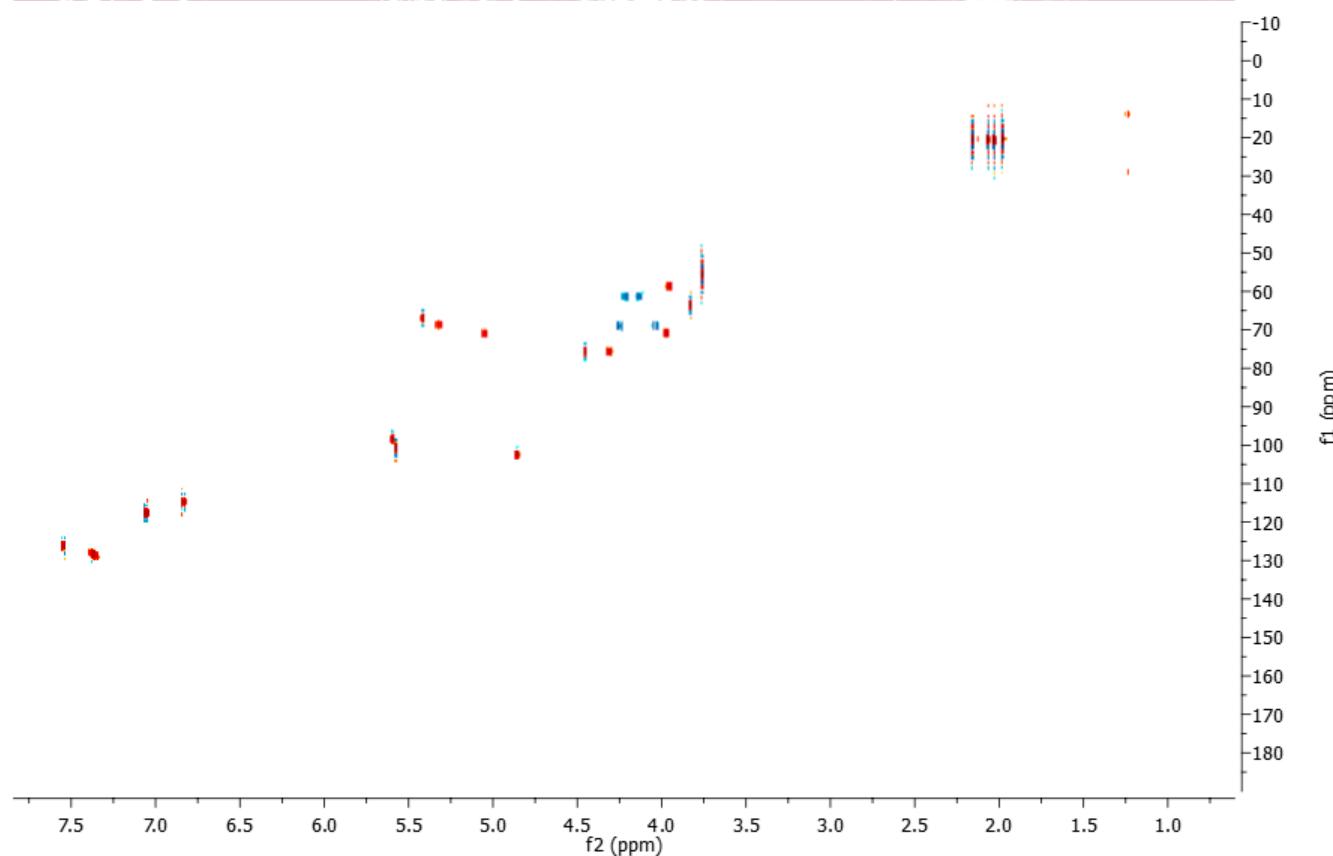


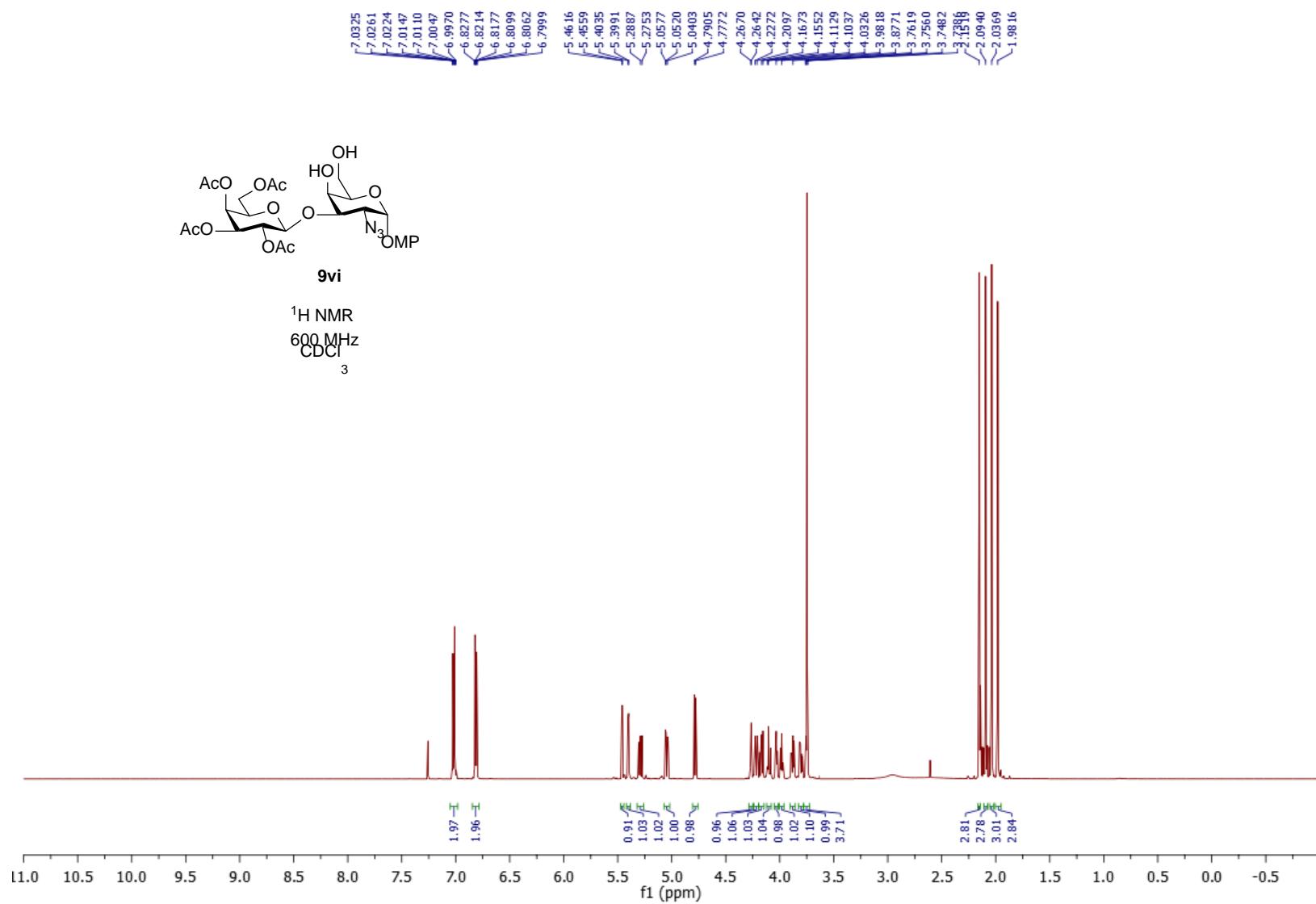


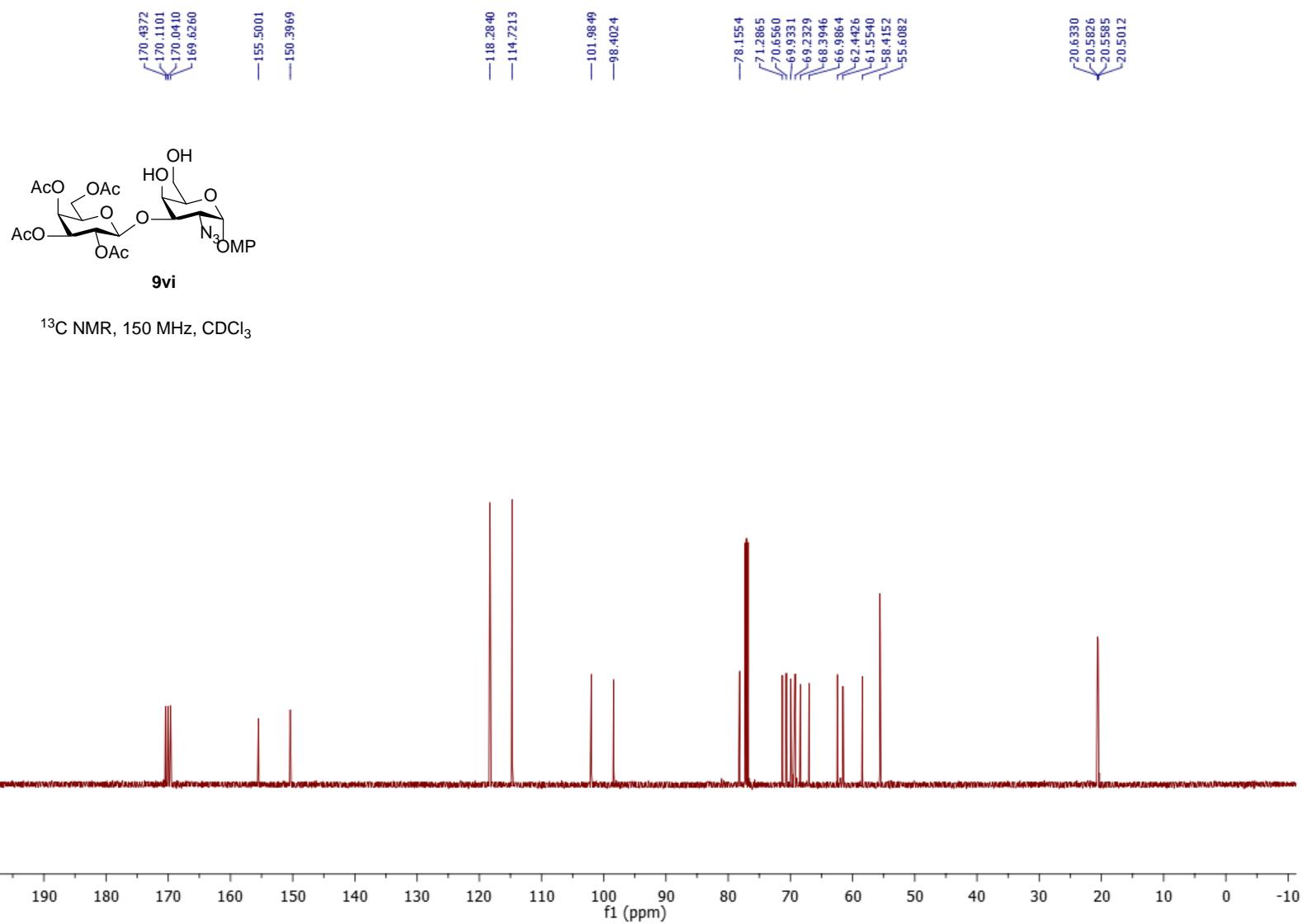


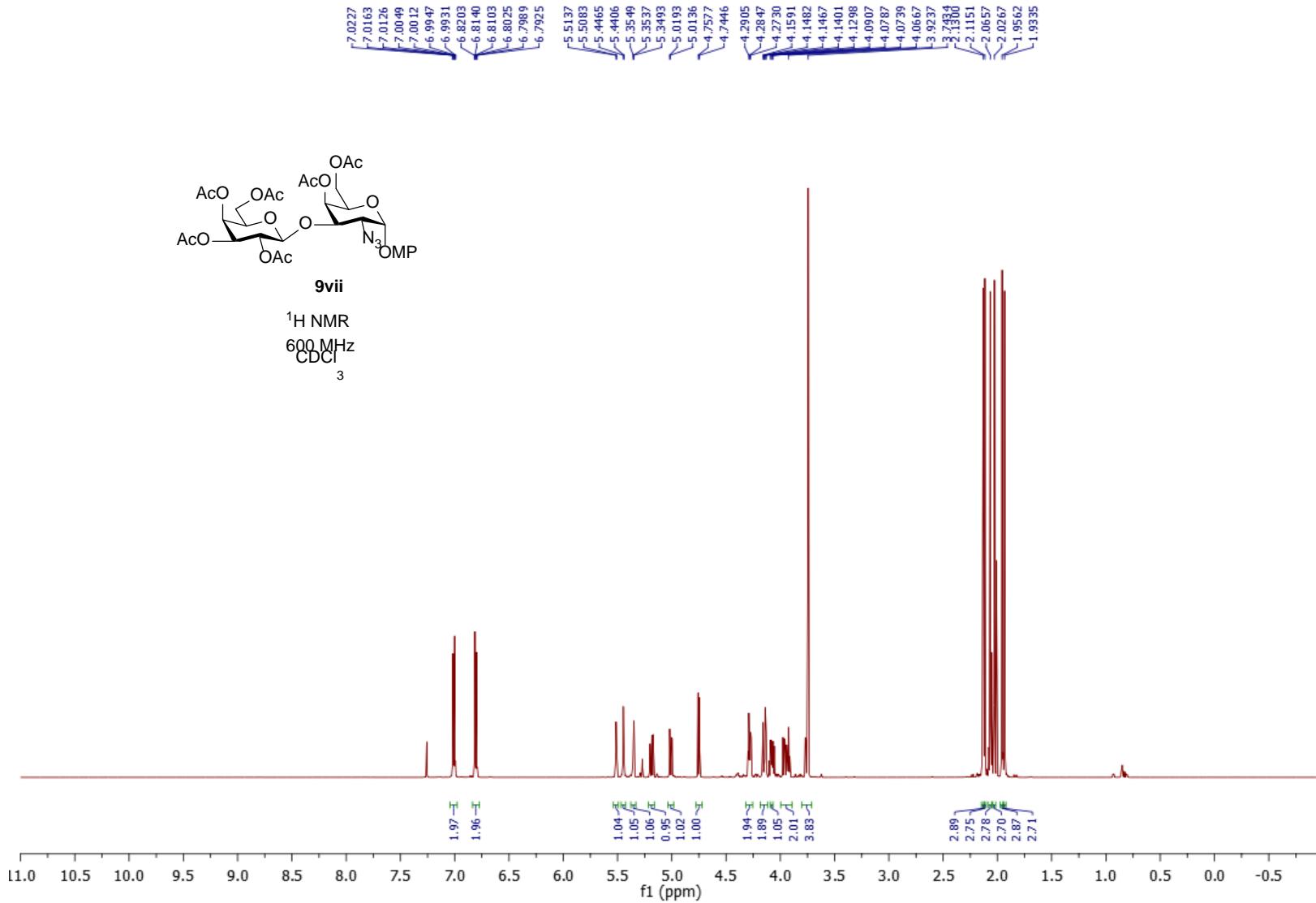


^1H - ^{13}C HSQC, 600/150MHz, CDCl_3









— 170.3563
— 170.3306
— 170.1876
— 170.0218
— 169.6365
— 169.4700

— 155.6250

— 150.2041

— 149.0507

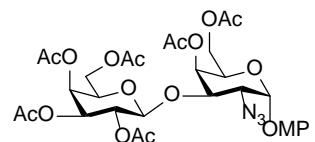
— 136.5736

— 118.4298
— 114.6033

— 101.5619
— 98.0235

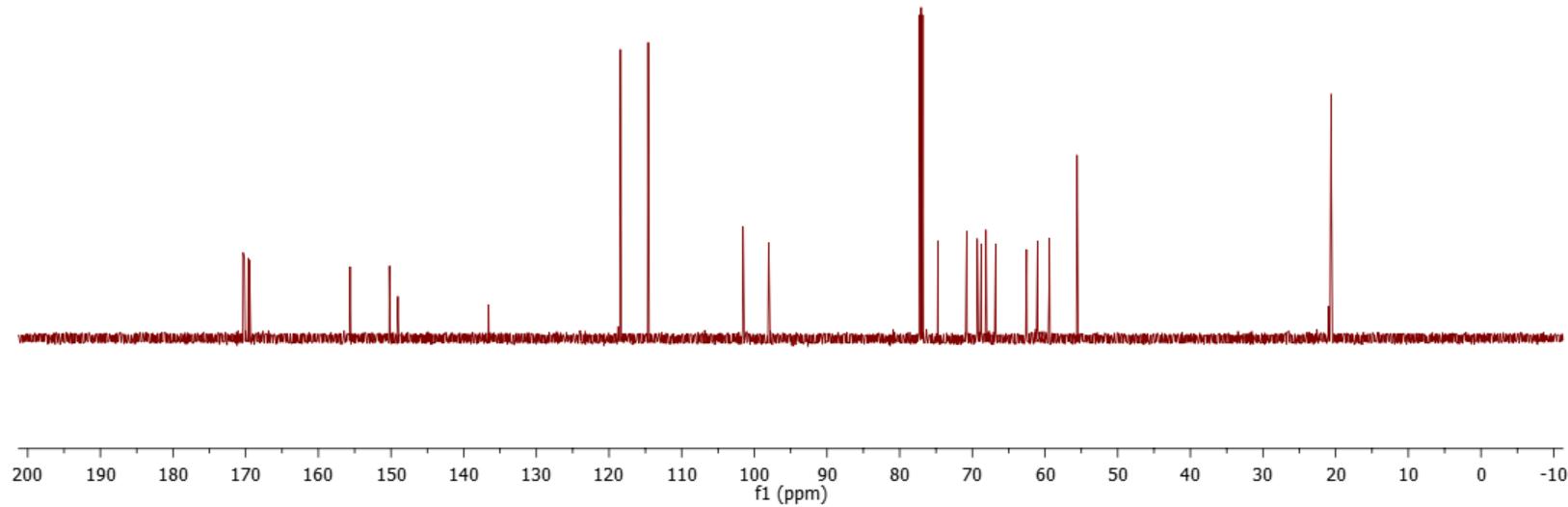
— 74.7339
— 70.8410
— 70.7675
— 69.3232
— 68.7655
— 68.1276
— 66.7825
— 62.5367
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— 59.3954
— 55.5912

— 20.6916
— 20.6235
— 20.5958
— 20.5865
— 20.4961

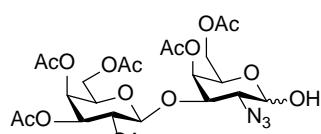


9vii

¹³C NMR, 150 MHz, CDCl₃



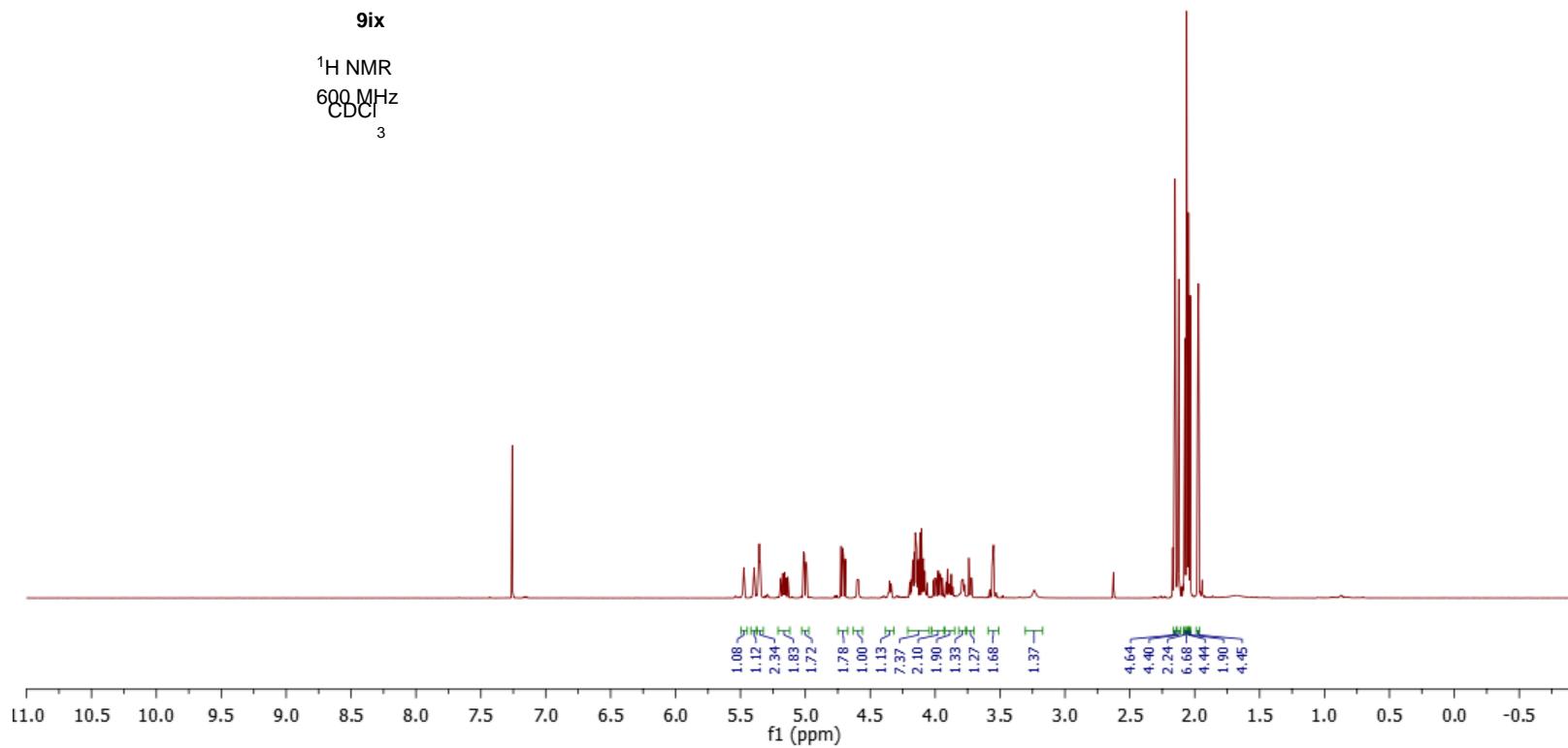
5.4795
5.4737
5.402
5.3945
5.3579
5.3549
5.3526
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5.1734
5.1655
5.1603
5.1524
5.1480
5.1349
5.0133
5.0076
4.9959
4.9901
4.7255
4.7124
4.7055
4.6924
4.6019
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4.3501
4.3393
4.1945
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4.1794
4.1748
4.1717
4.1669
4.1611
4.1530
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4.1302
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4.1015
4.0607
4.0931
4.0903
4.0823
4.0796
4.0713
4.0607
4.0115
3.9992
3.9919
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3.9157
3.9043
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3.7956
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3.5466
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1.9713

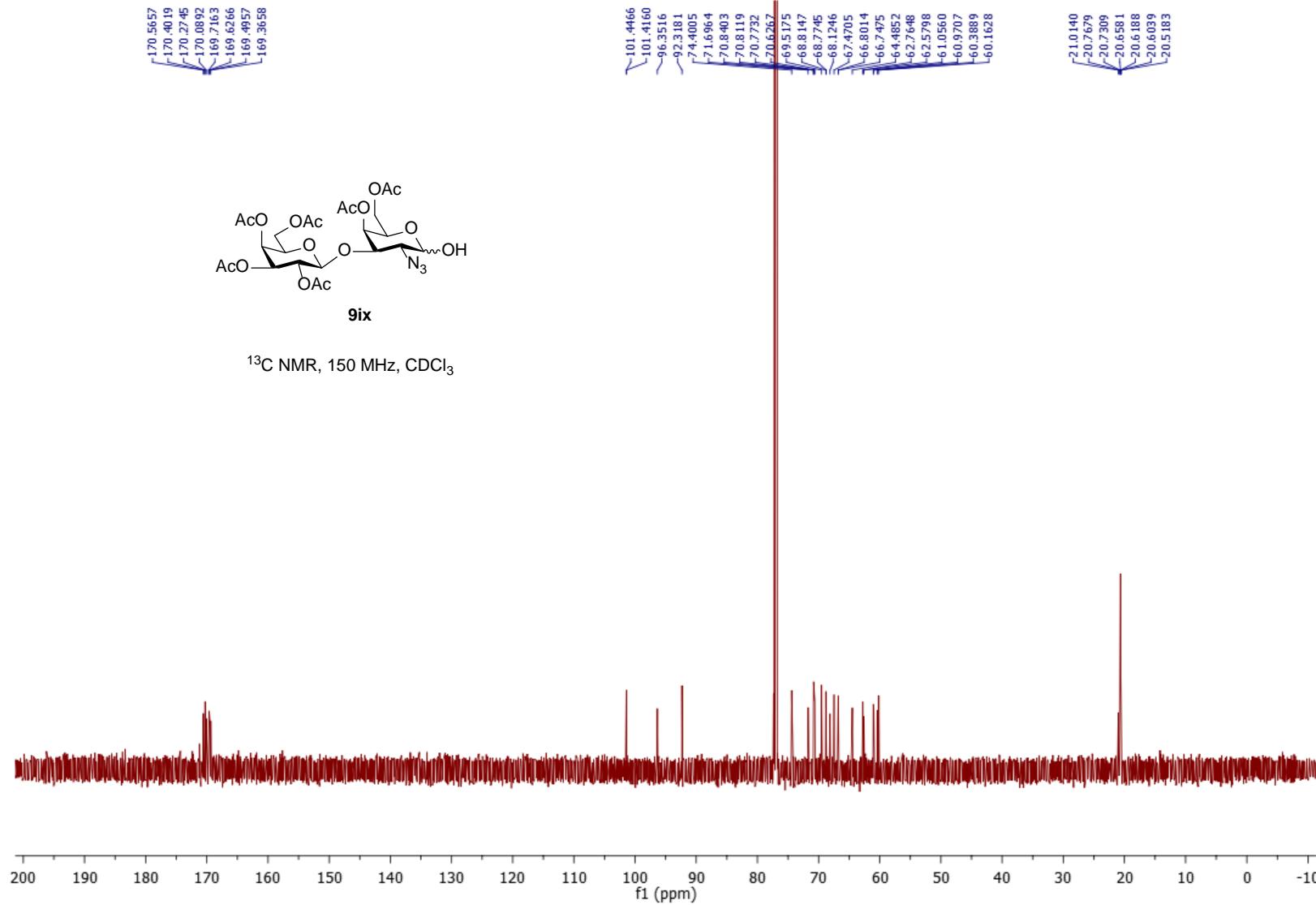


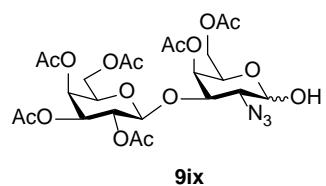
9ix

¹H NMR
600 MHz
CDCl₃

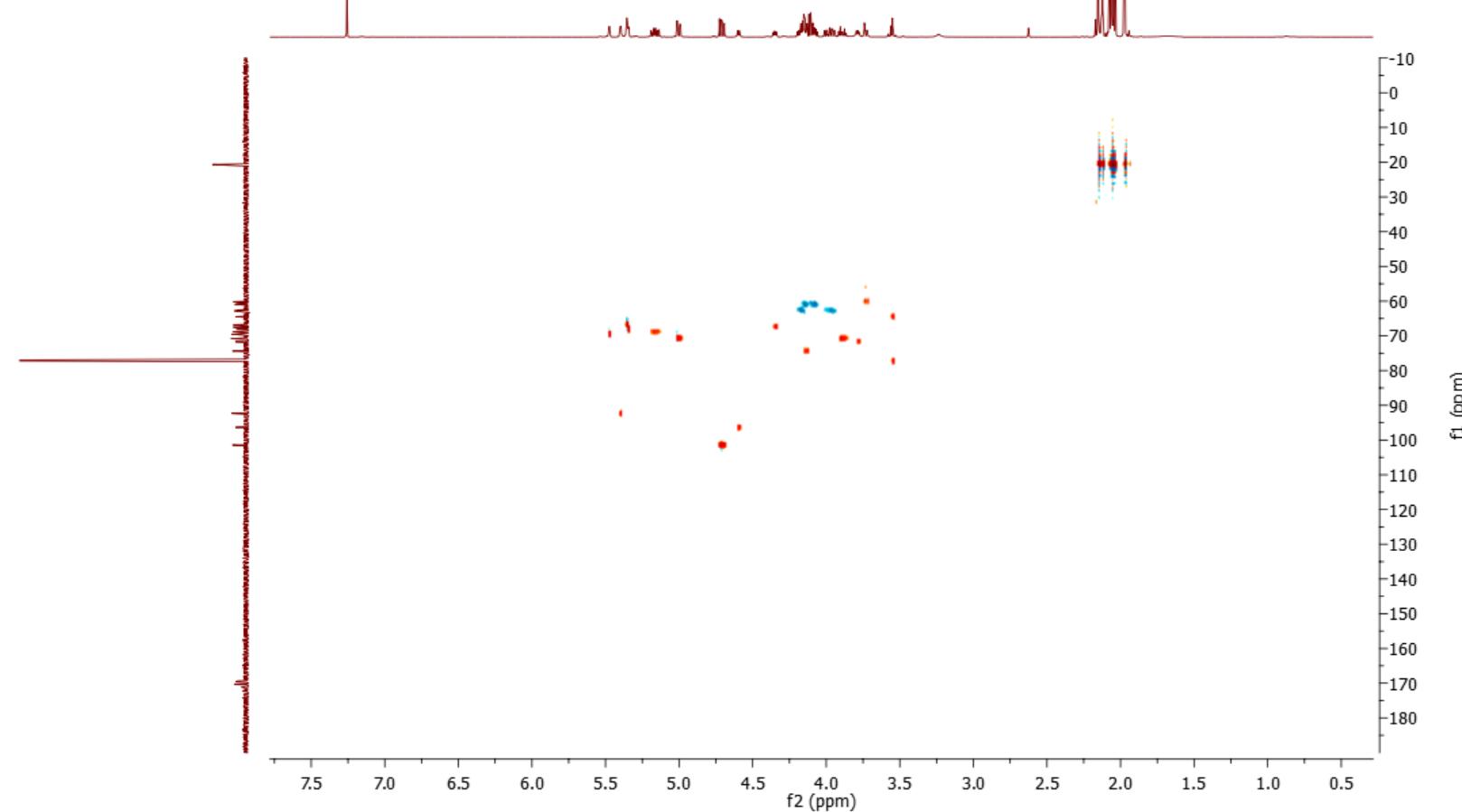
3

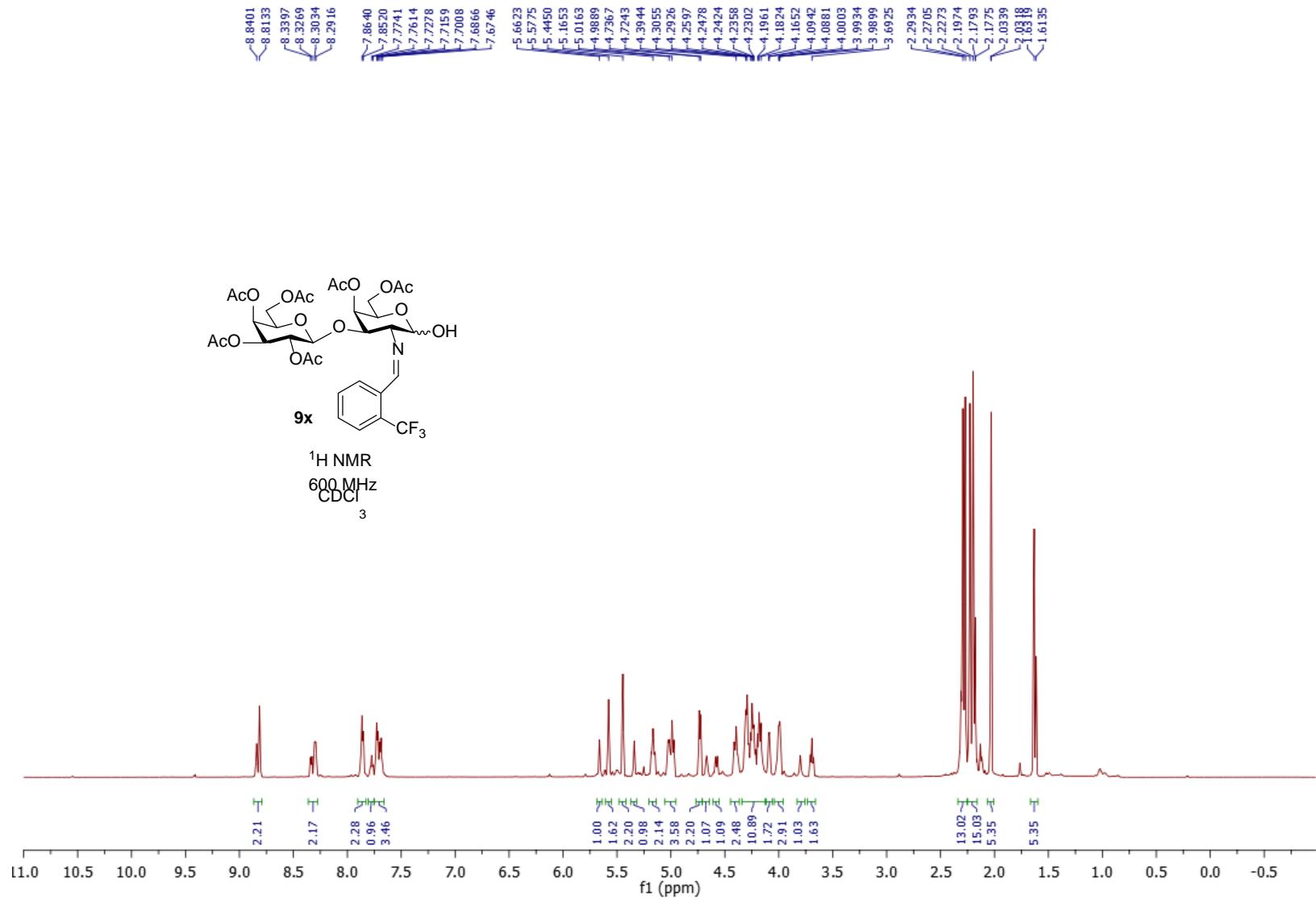


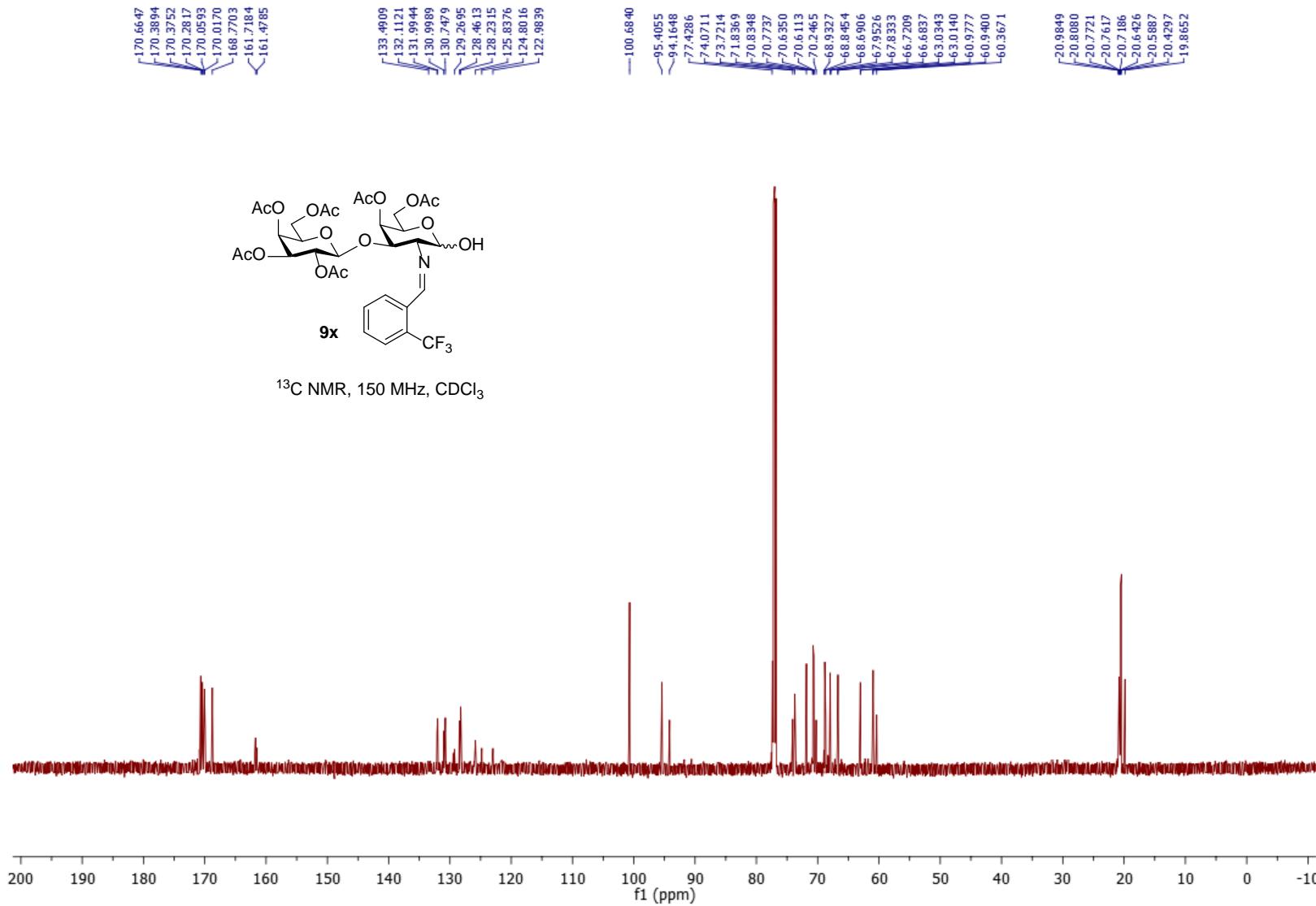


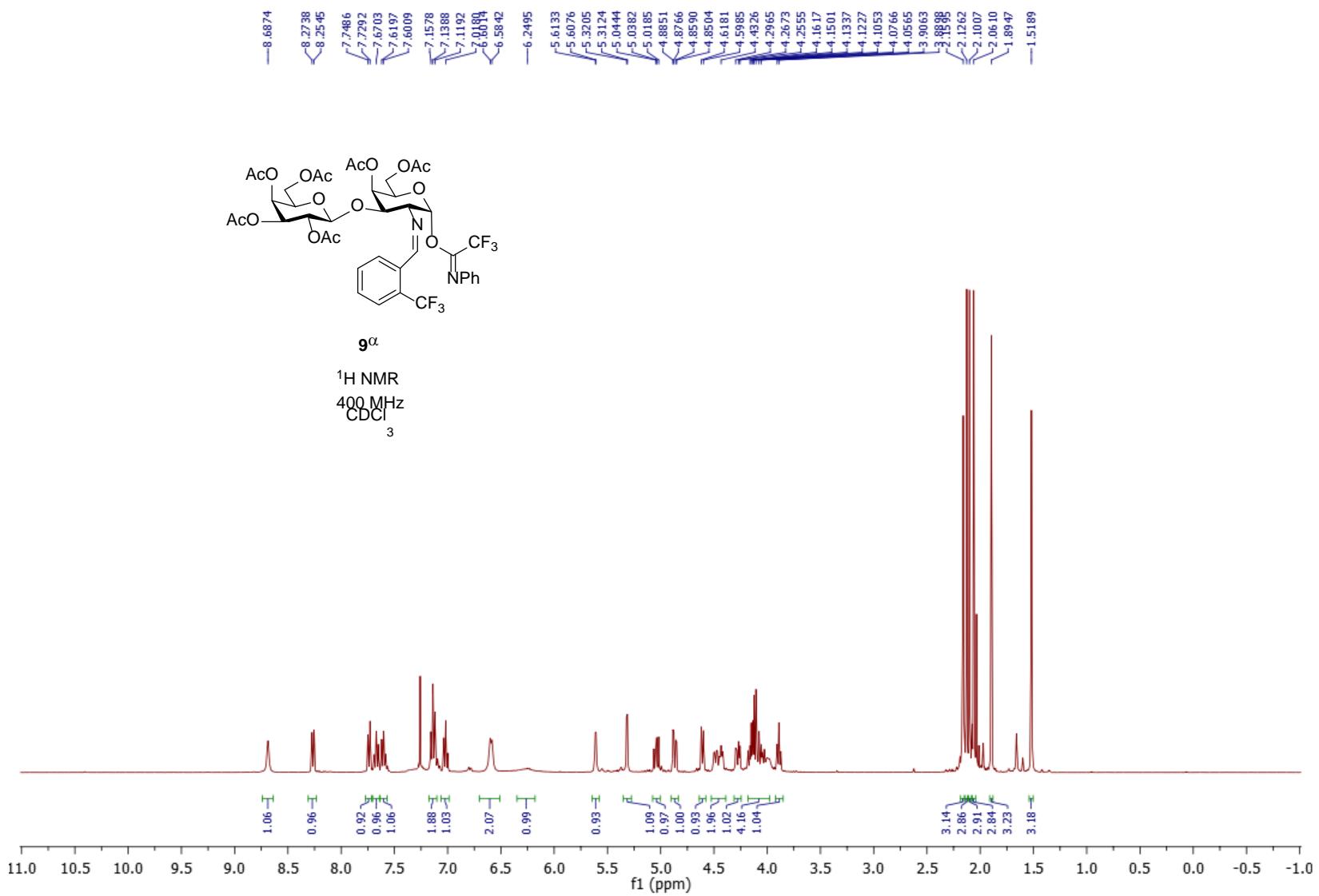


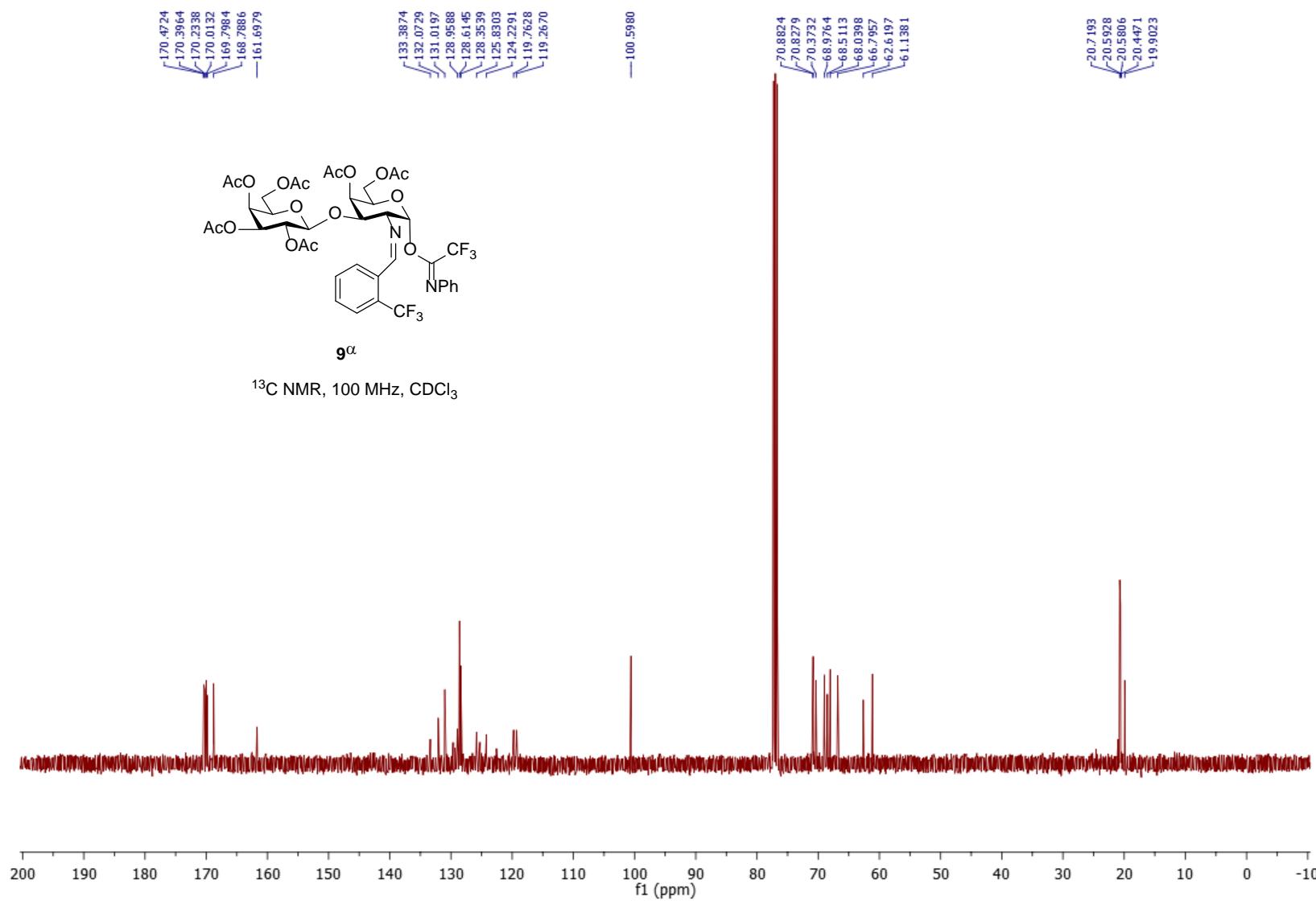
^1H - ^{13}C HSQC, 600/150MHz, CDCl_3

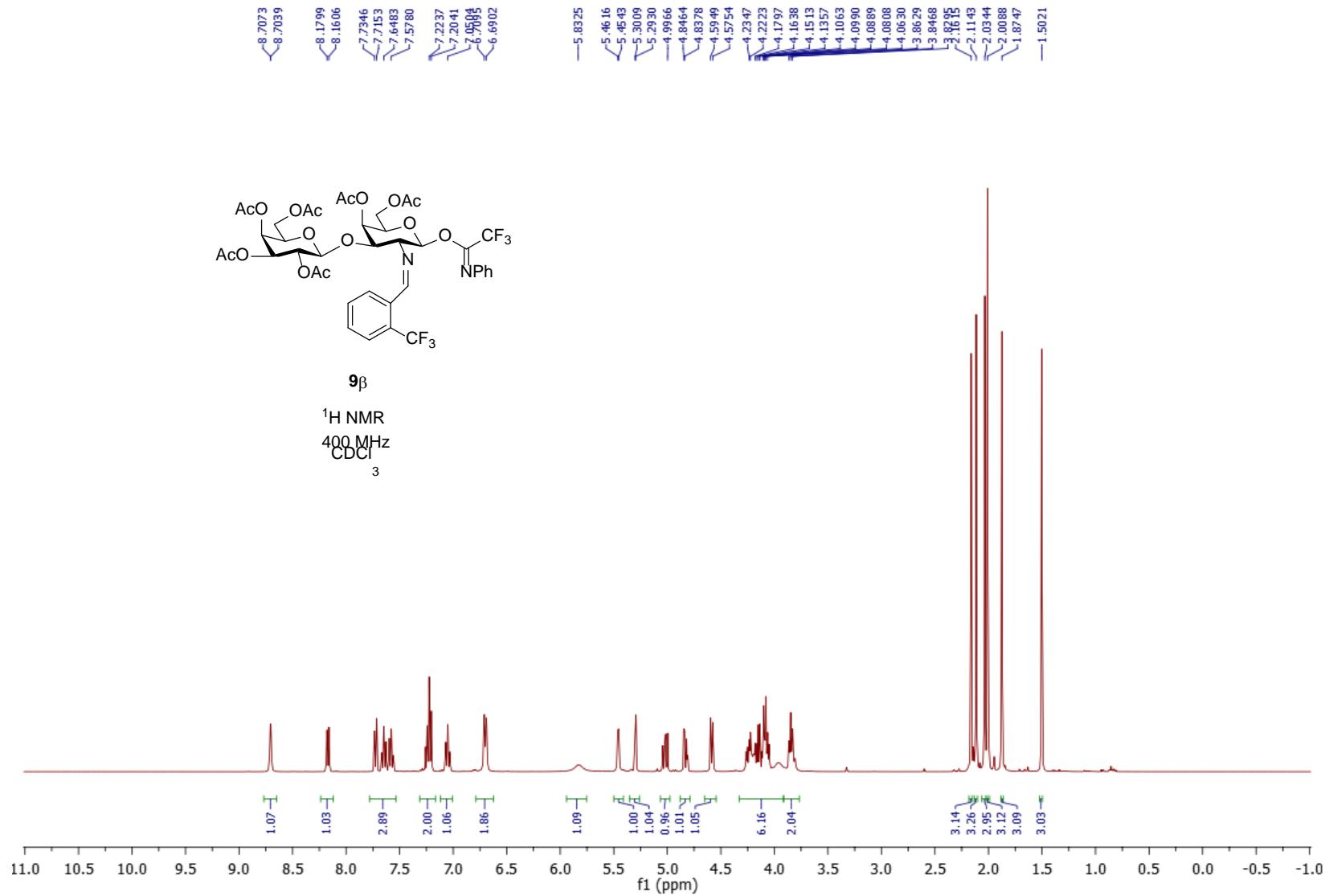












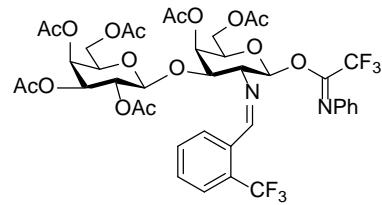
— 170.4595
— 170.3151
— 170.2134
— 169.9938
— 169.8789
— 168.7288
— 162.5167

— 143.2533
— 133.2816
— 132.0601
— 131.0298
— 129.7349
— 129.4230
— 128.6494
— 128.2228
— 125.9679
— 125.9116
— 125.1893
— 124.2870
— 122.4632
— 119.0458

— 100.7755
— 95.24667

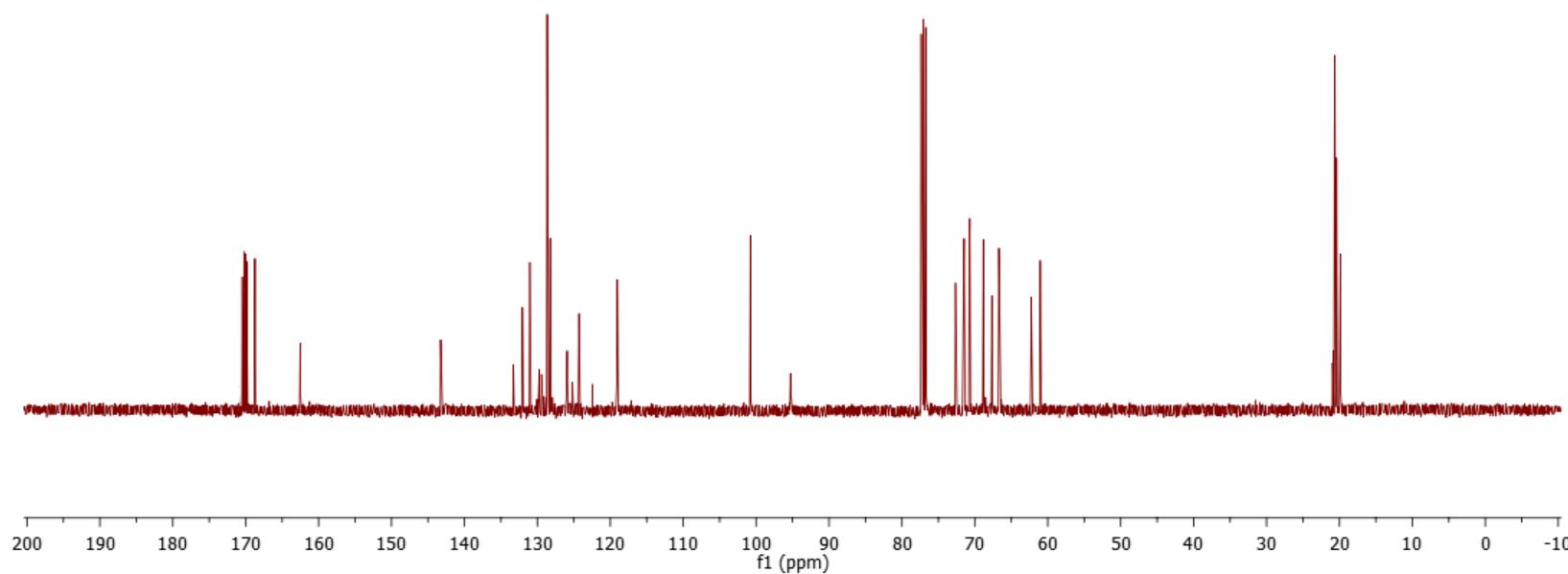
— 77.1649
— 72.6390
— 71.4766
— 70.7448
— 70.6892
— 68.8225
— 67.0502
— 66.7012
— 62.2607
— 61.0305

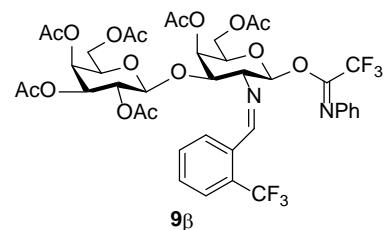
— 20.6723
— 20.6365
— 20.6212
— 20.5609
— 20.4044
— 19.8633



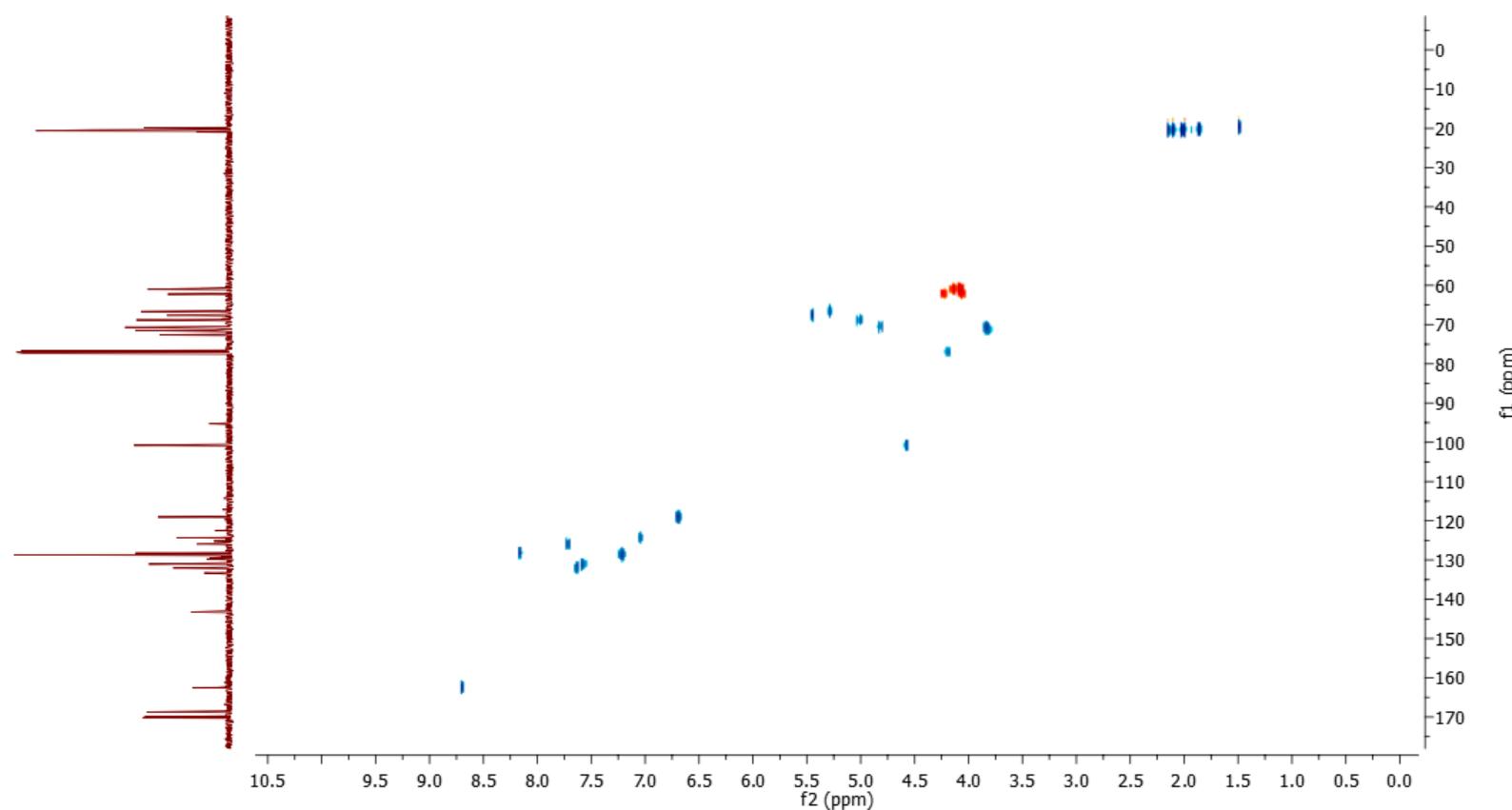
9 β

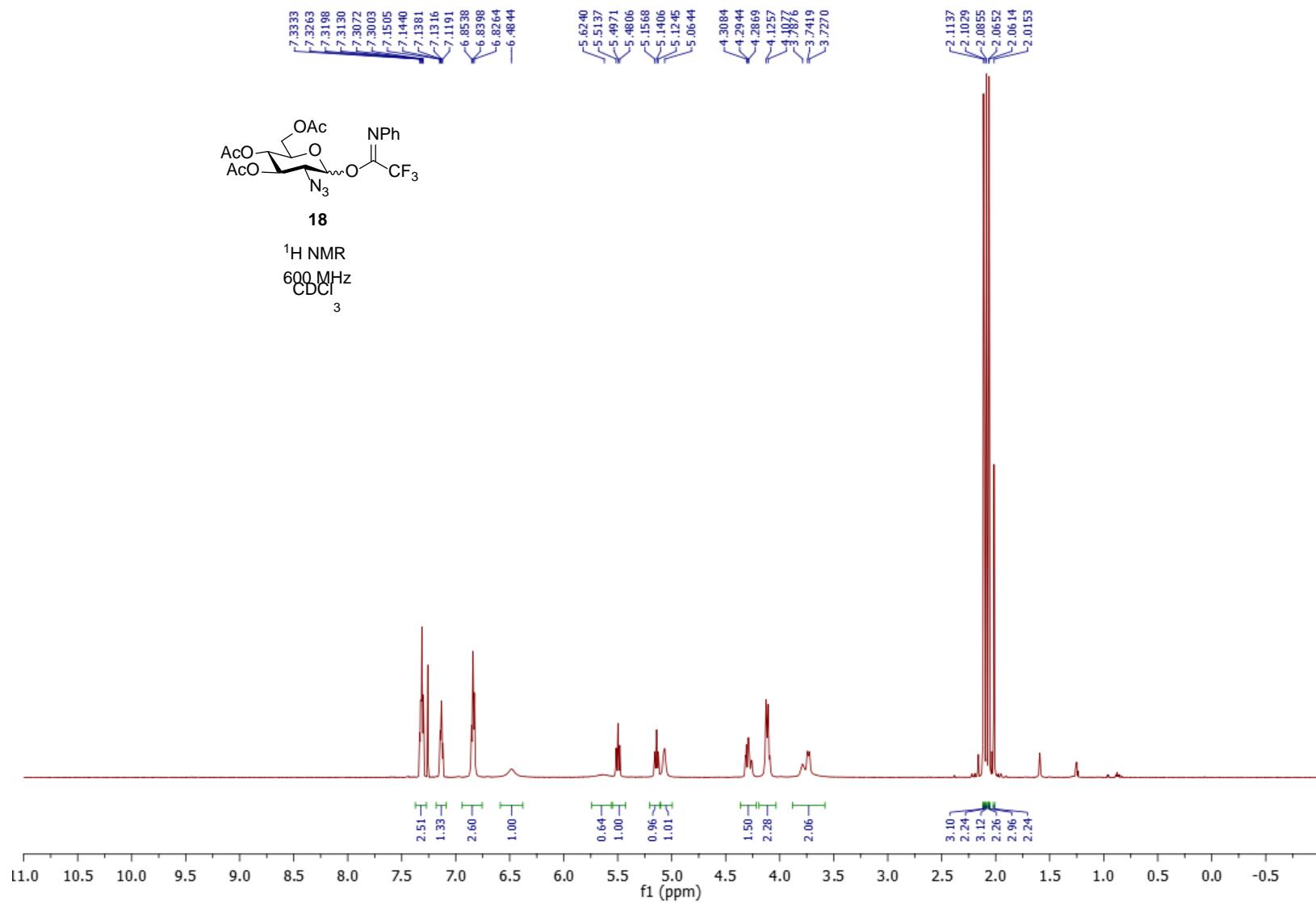
^{13}C NMR, 100 MHz, CDCl_3

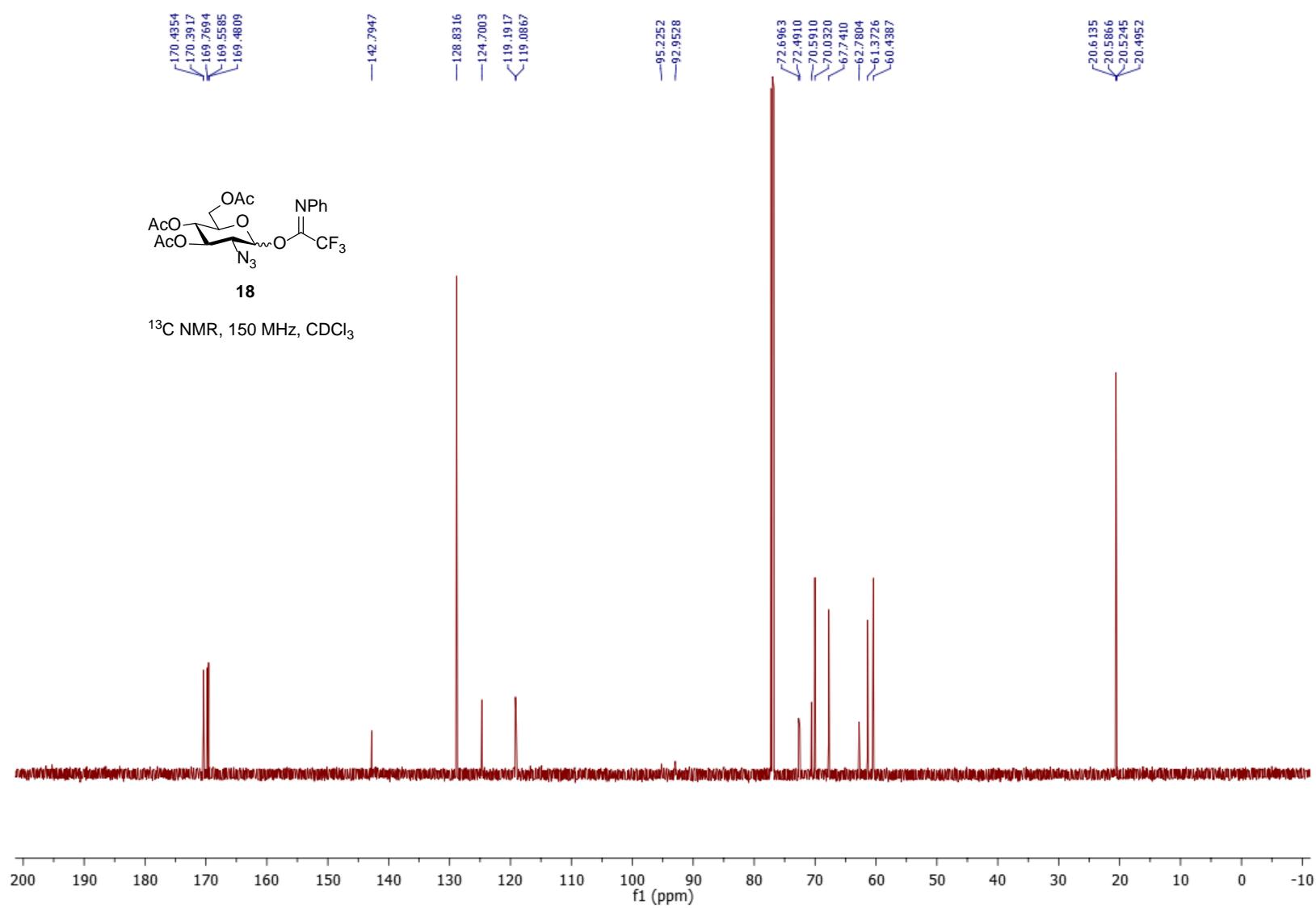


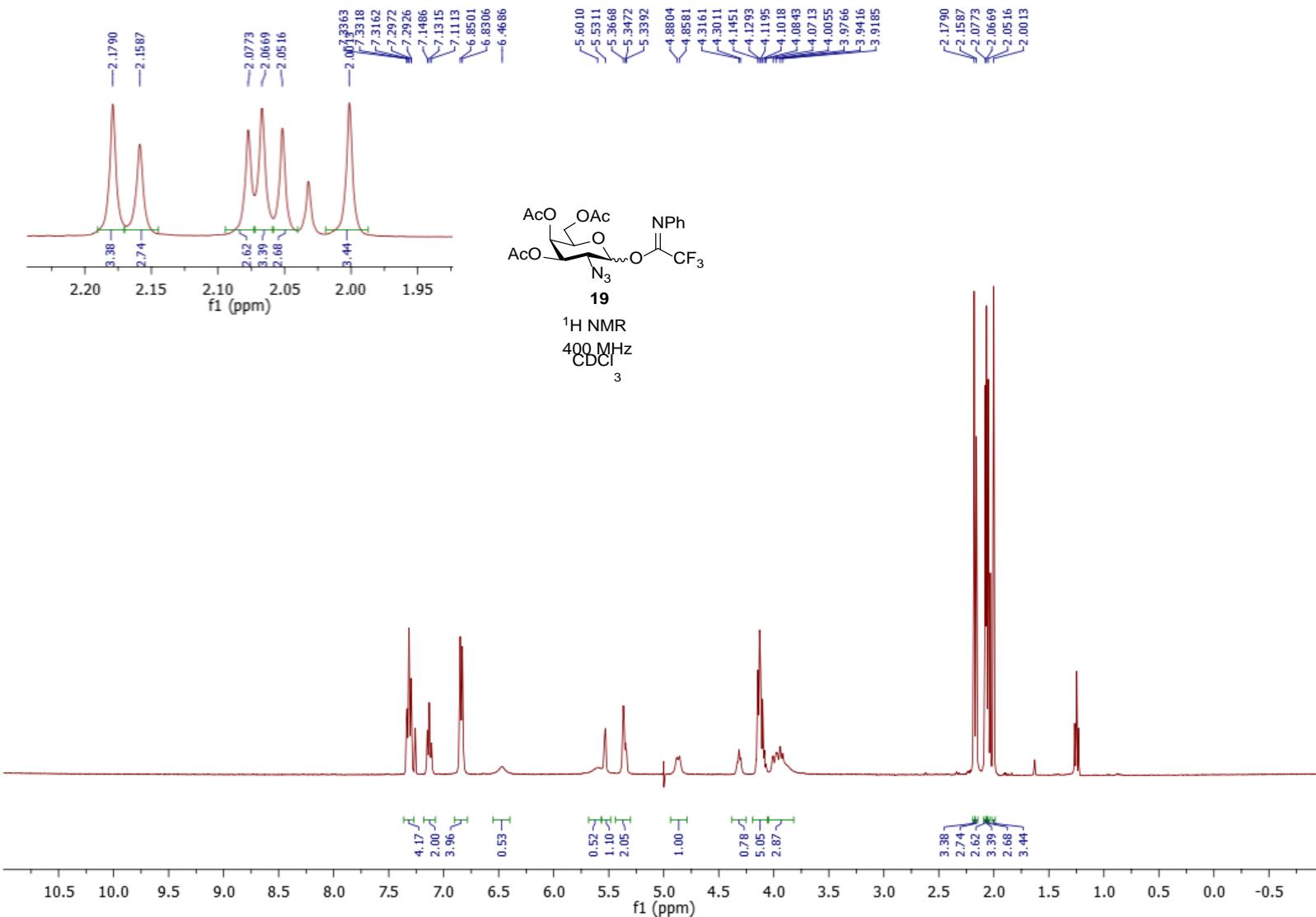


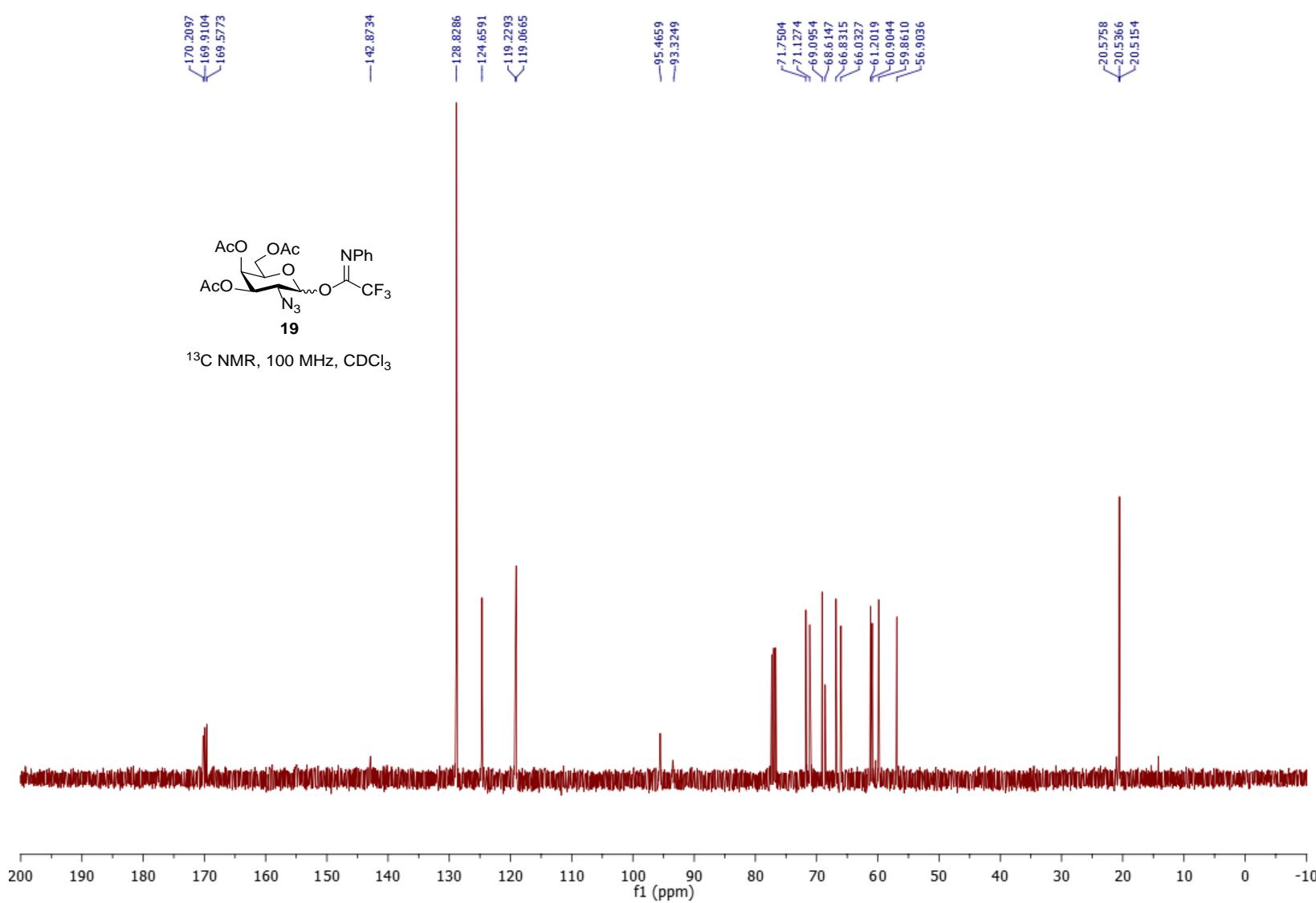
^1H - ^{13}C HSQC, 400/100MHz, CDCl_3







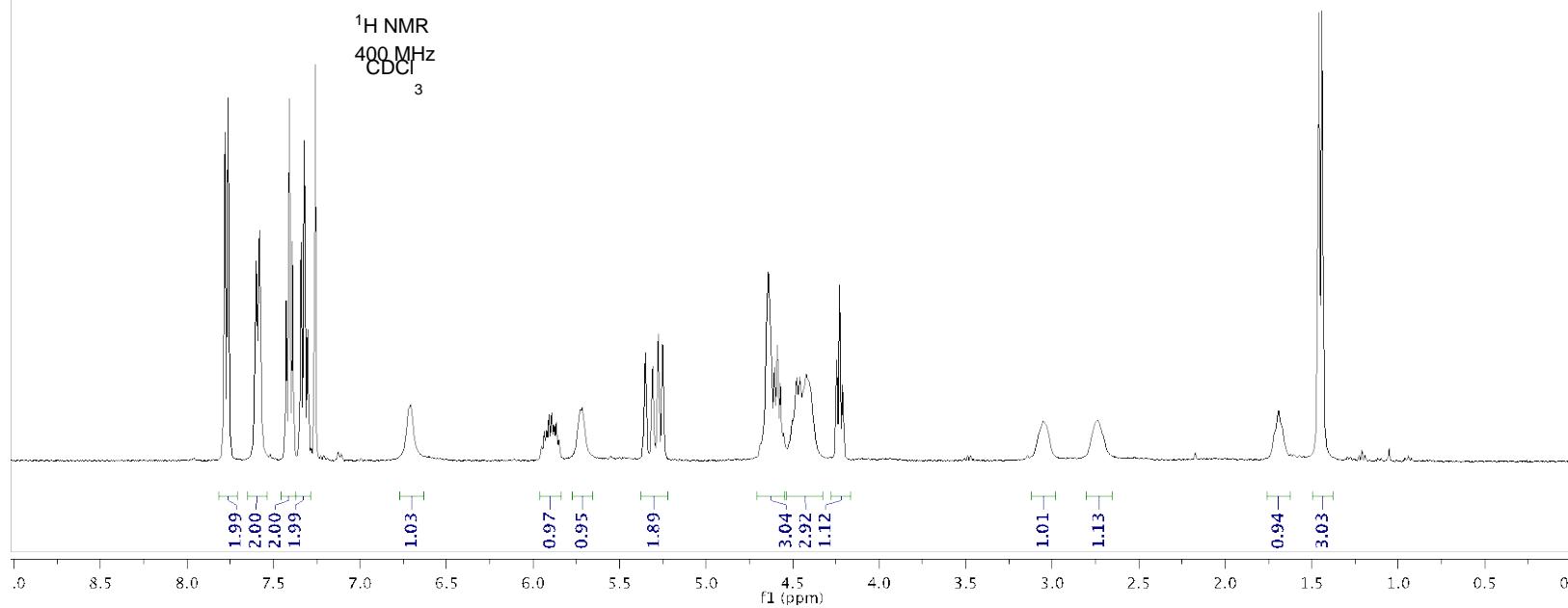


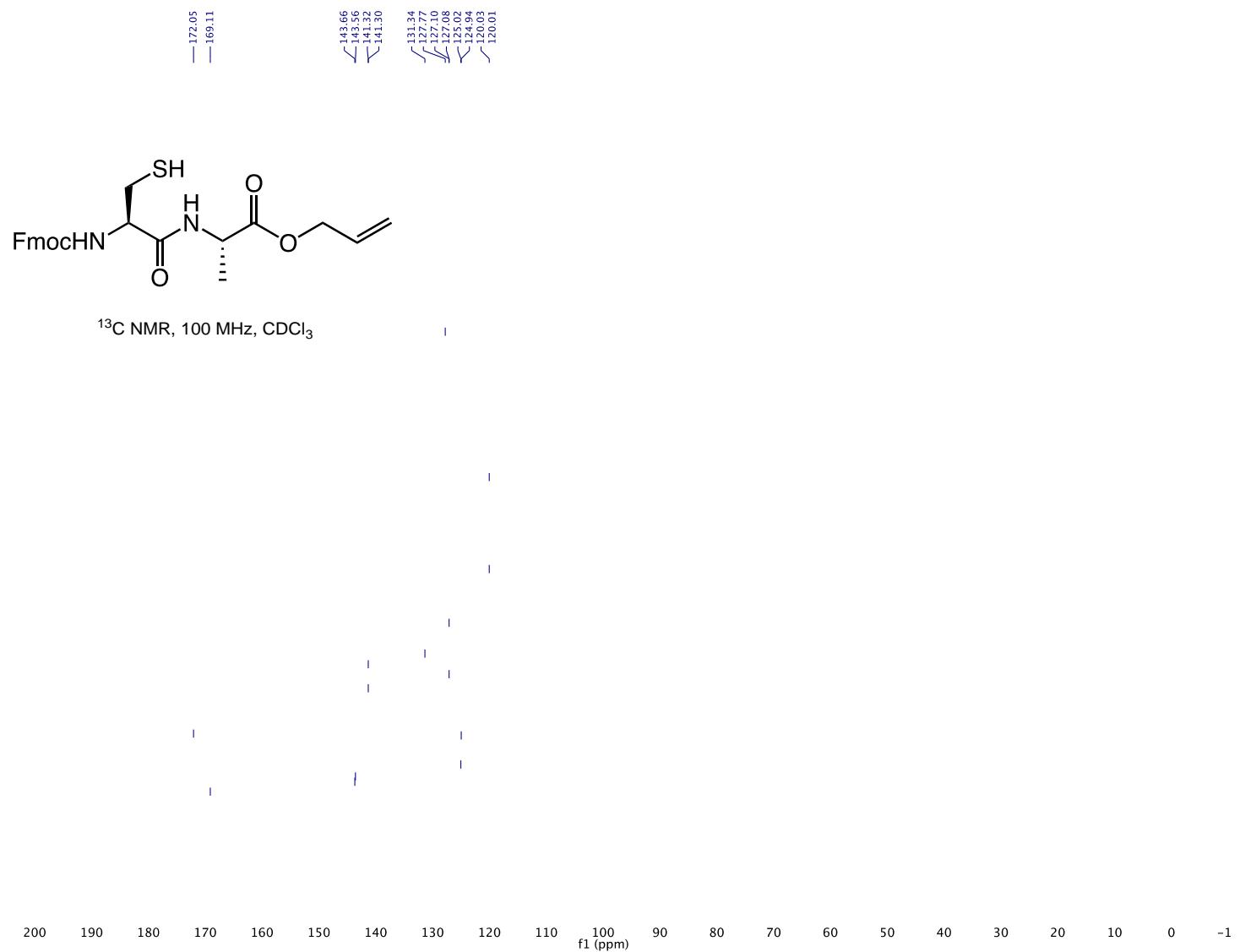


GS-IV-057-1H
GS-IV-054-1H

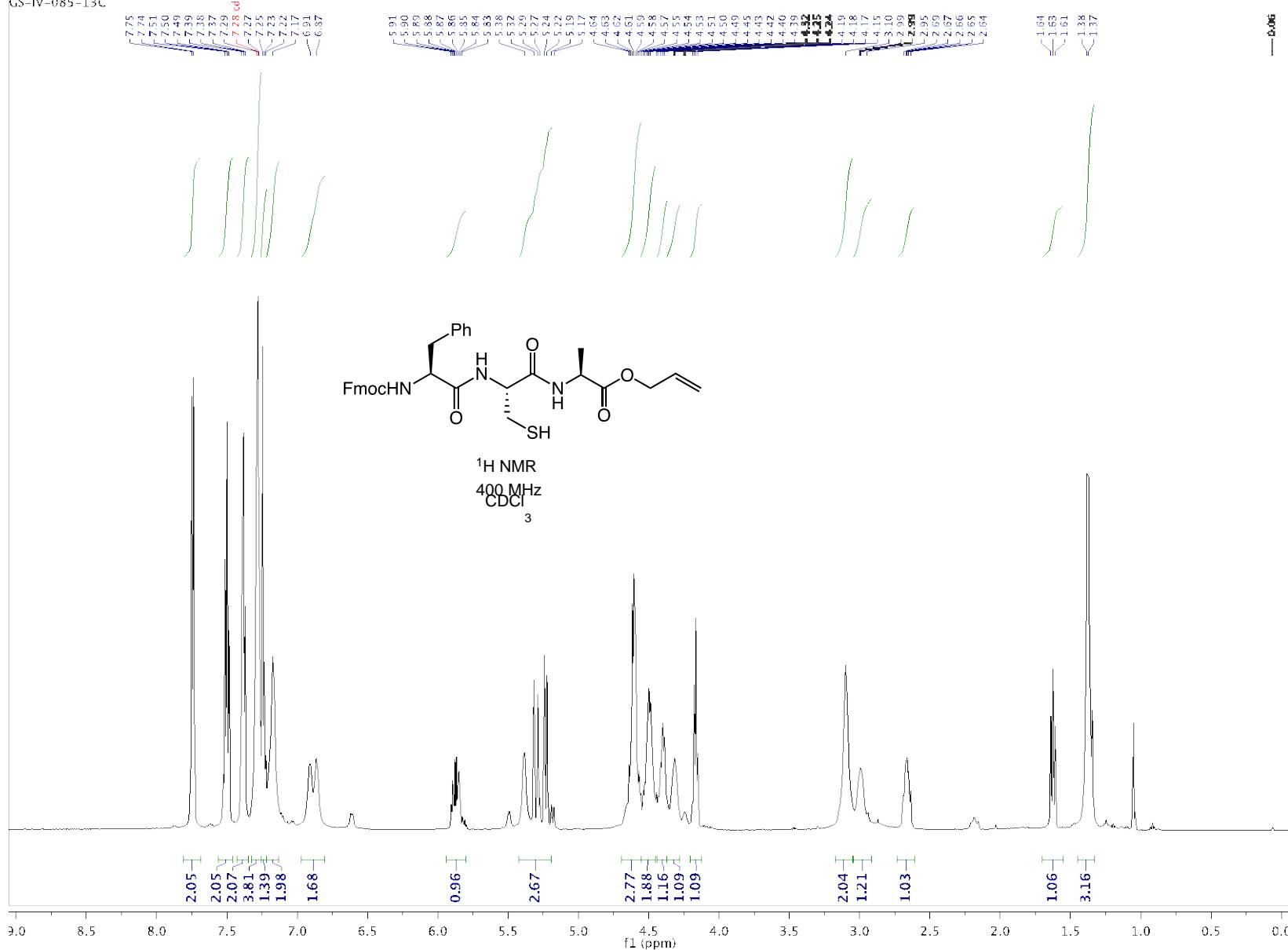


¹H NMR
400 MHz
CDCl₃



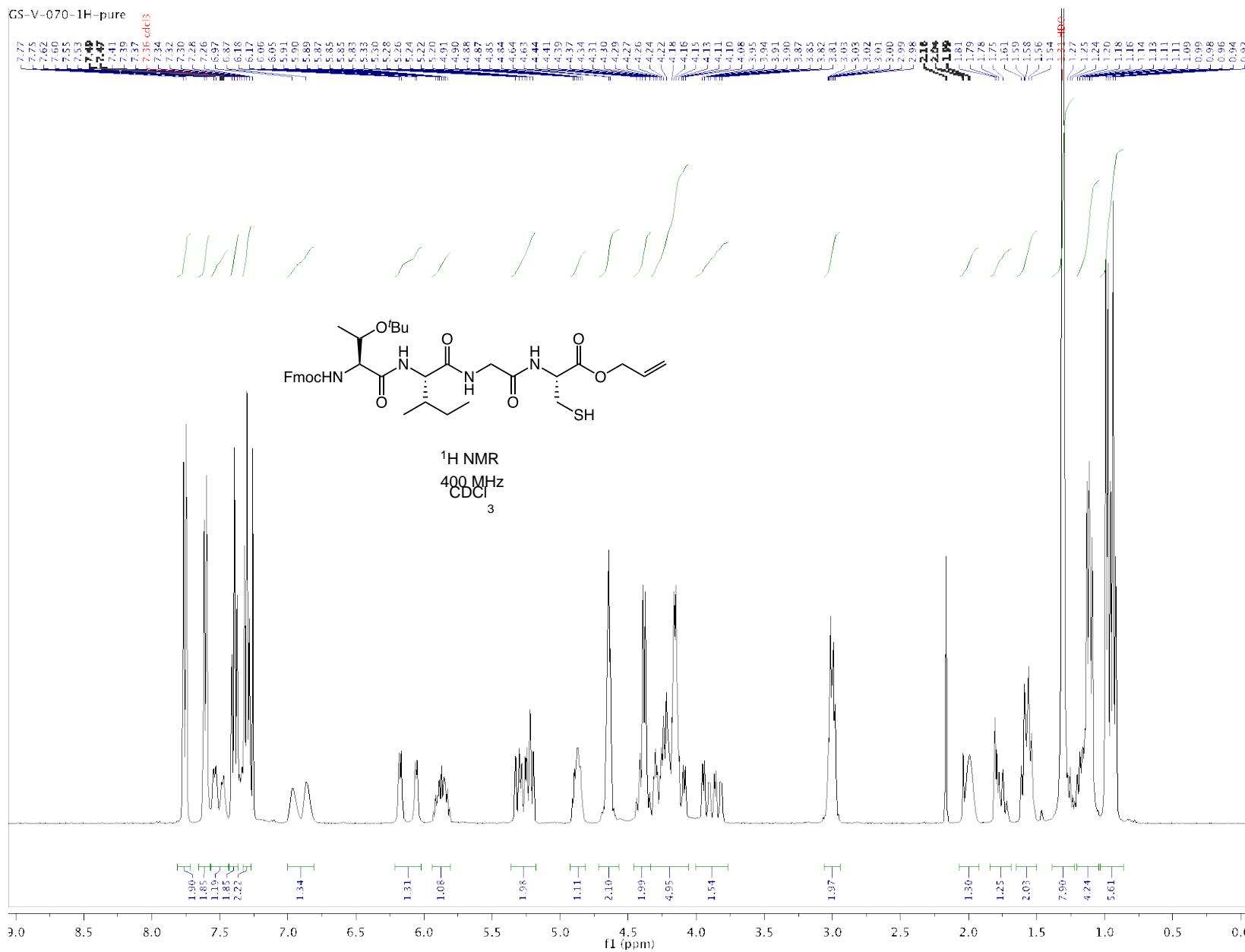


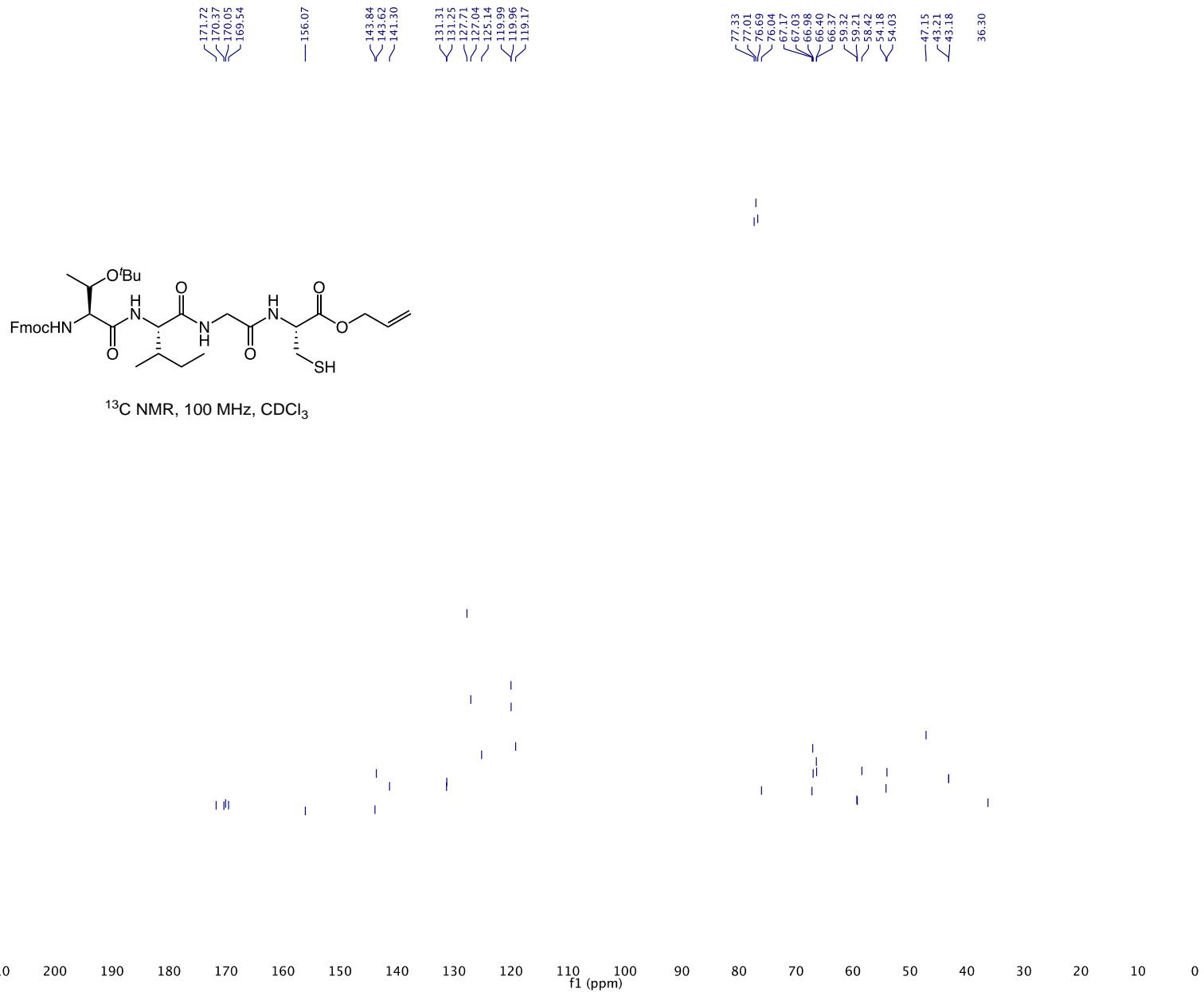
GS-IV-089-1H-pure
GS-IV-085-13C



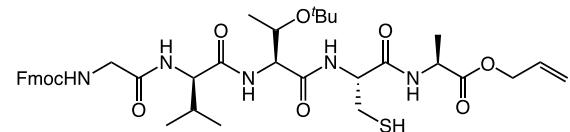


GS-V-070-1H-pure





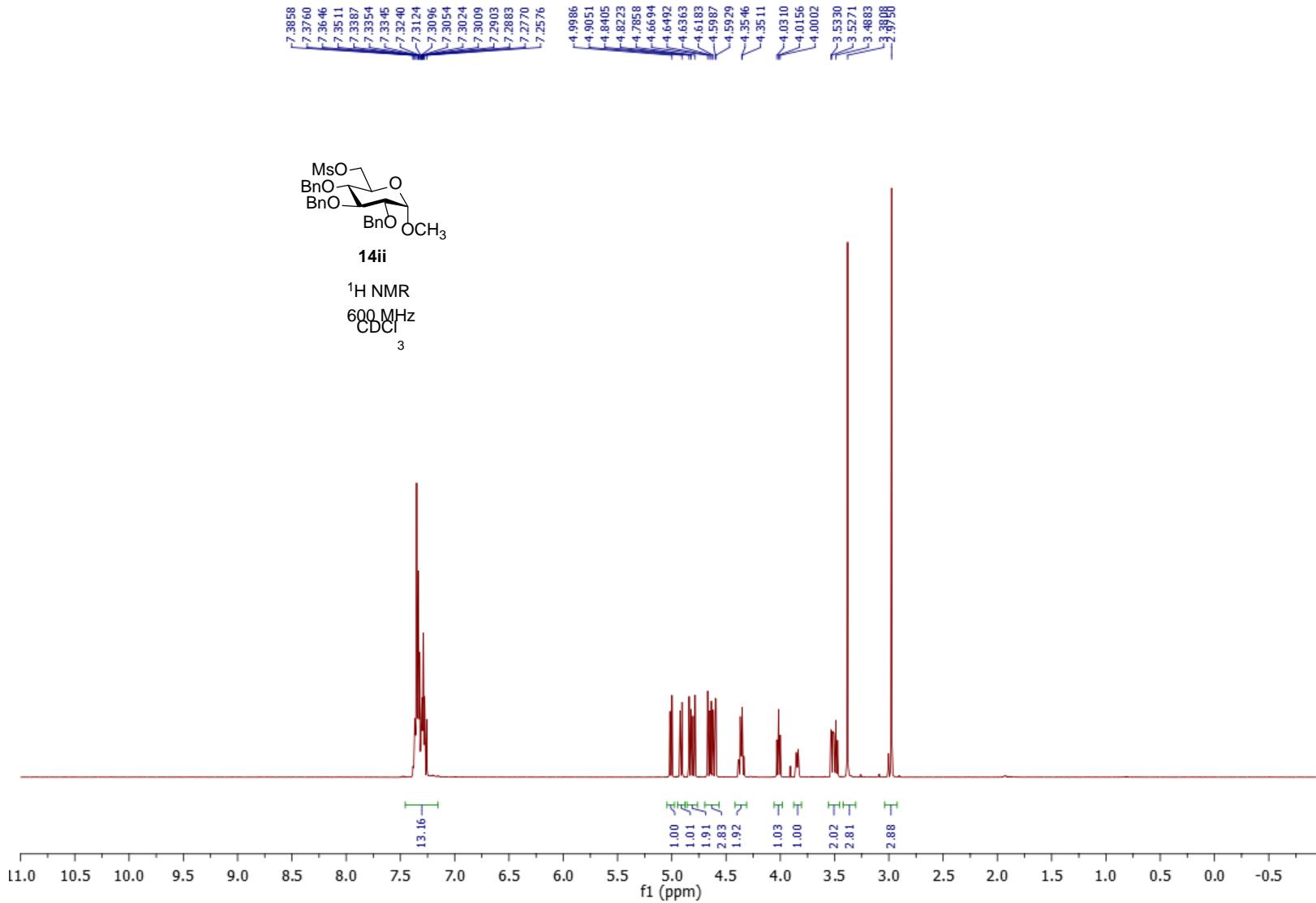
8.22

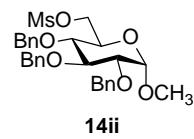


¹H NMR
400 MHz
³CDCl

9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0

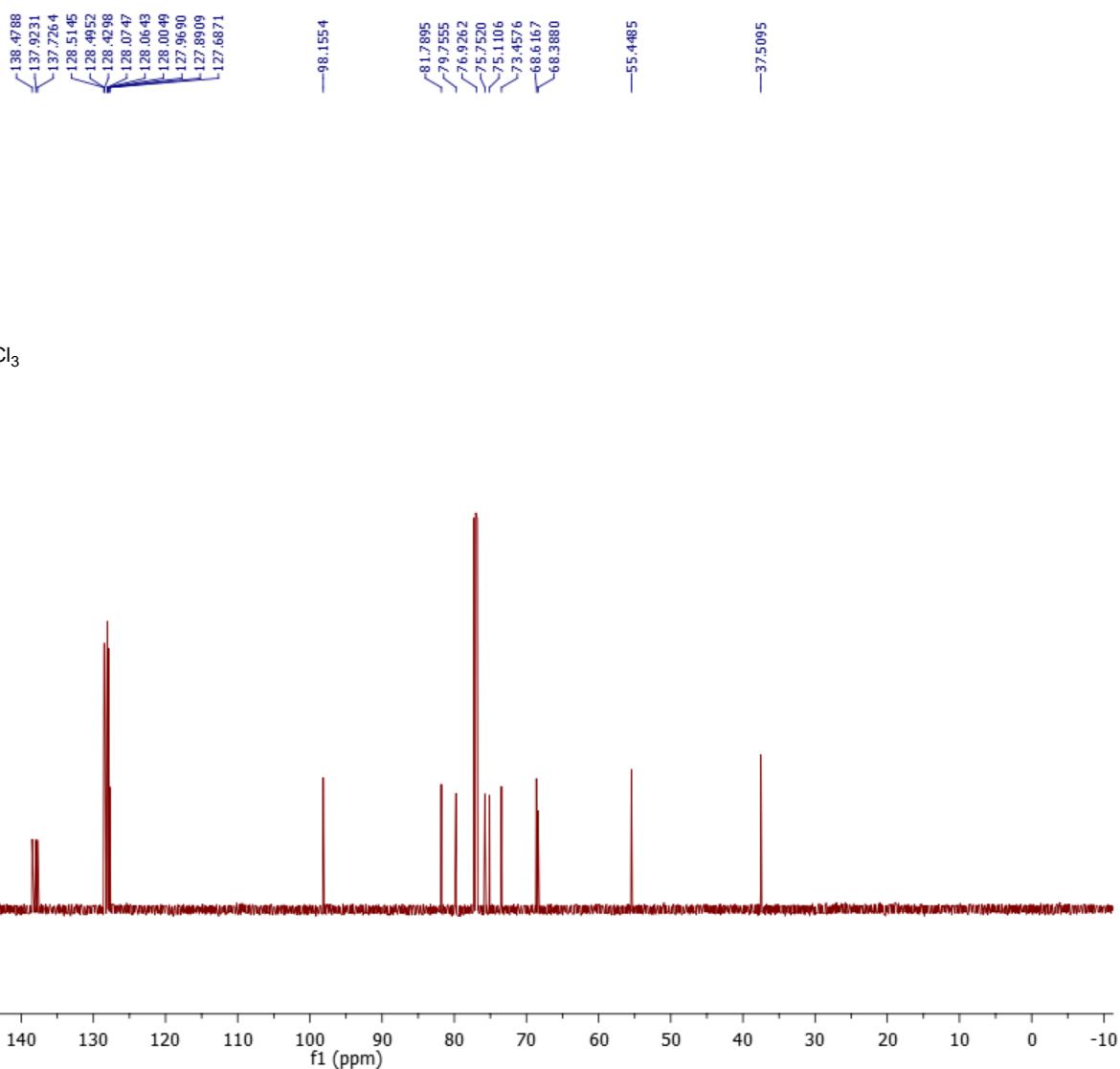


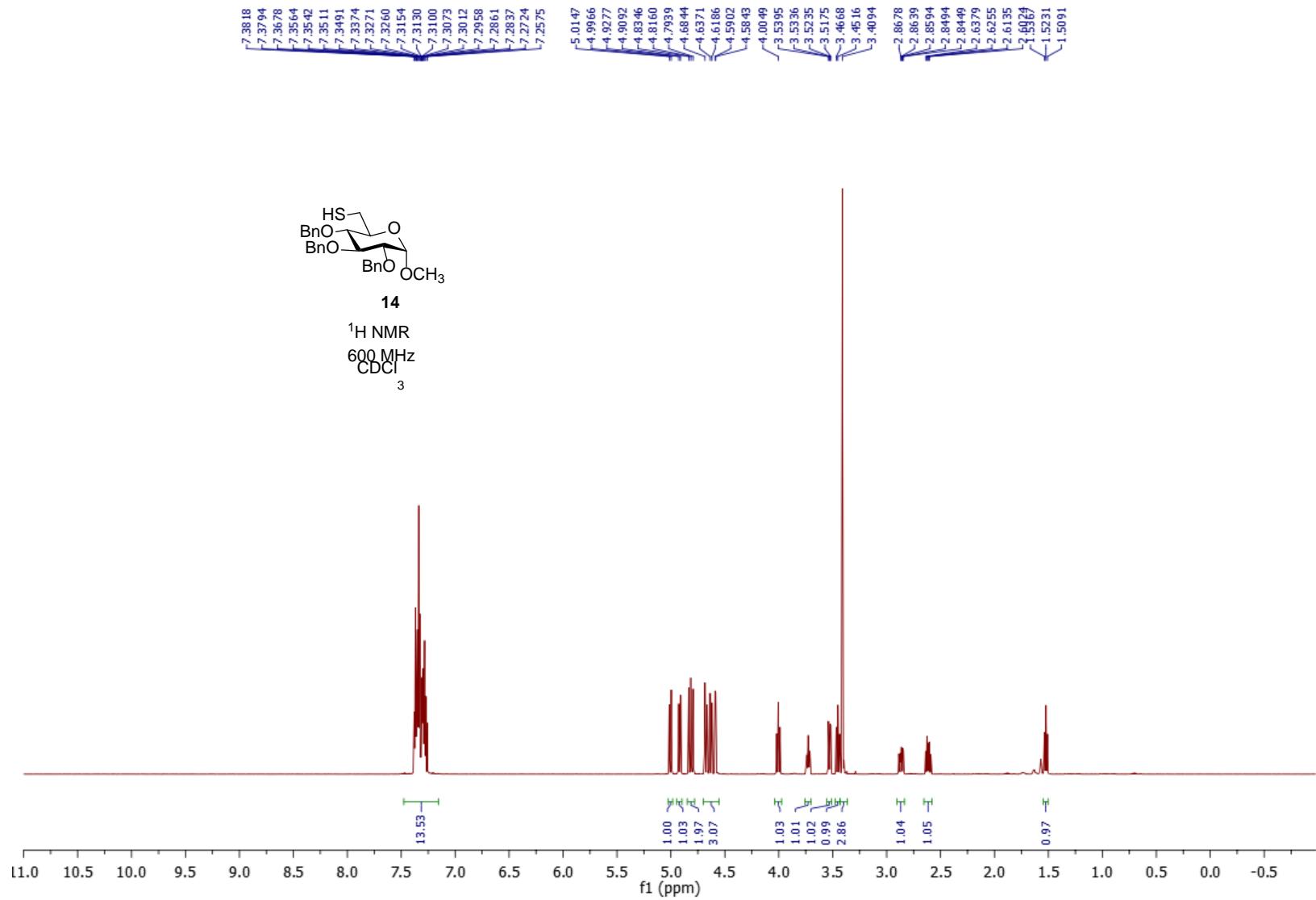


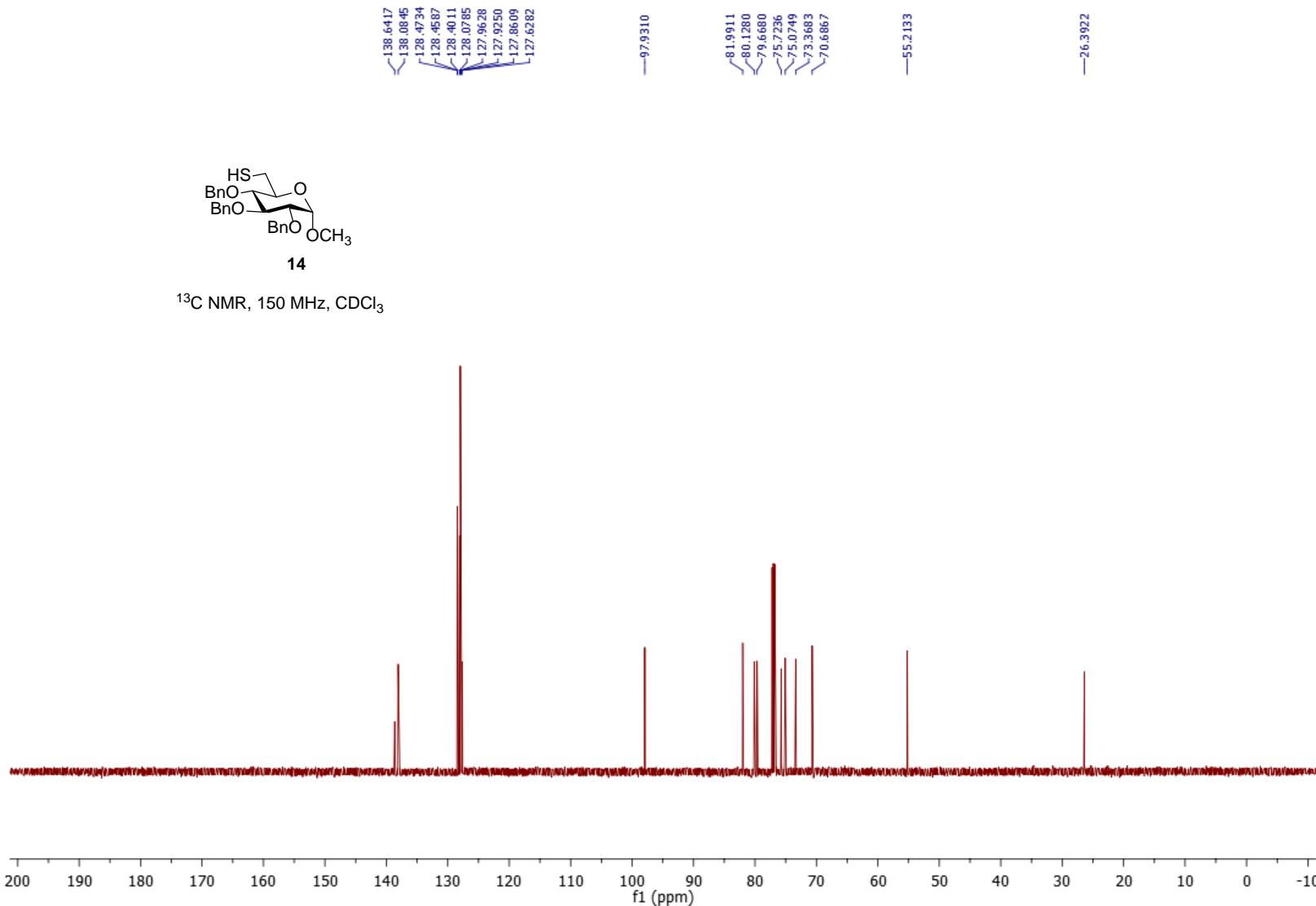


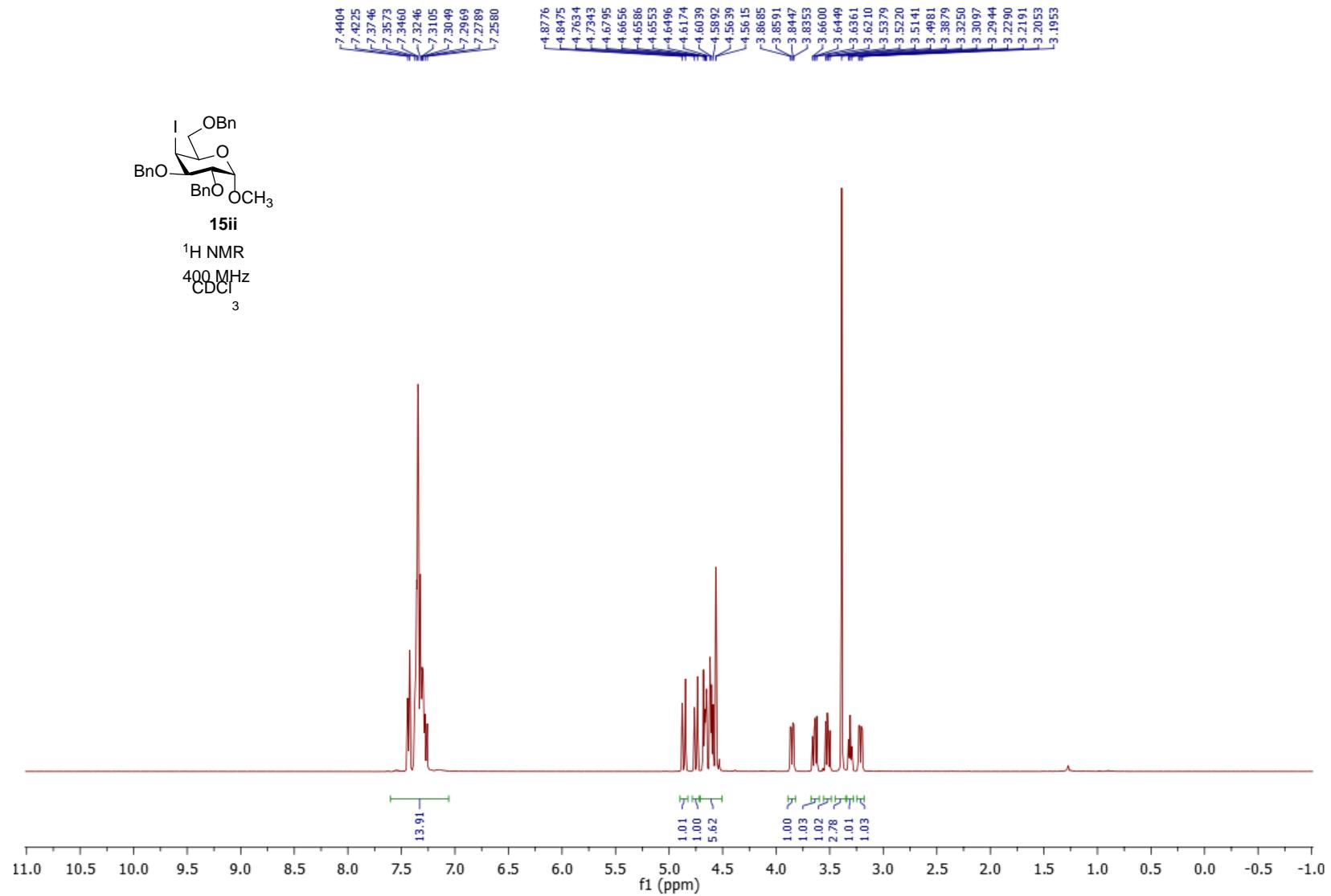
14ii

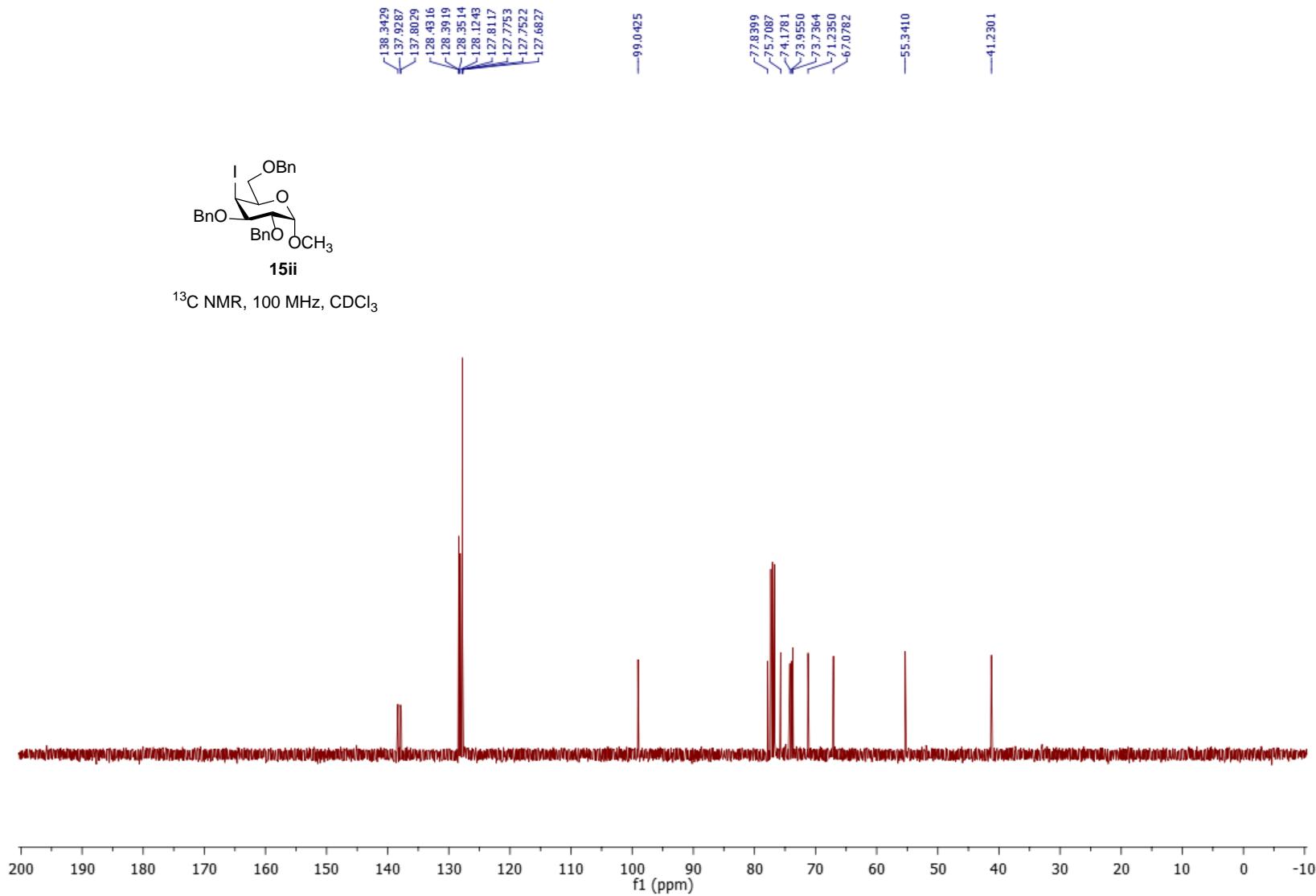
^{13}C NMR, 150 MHz, CDCl_3

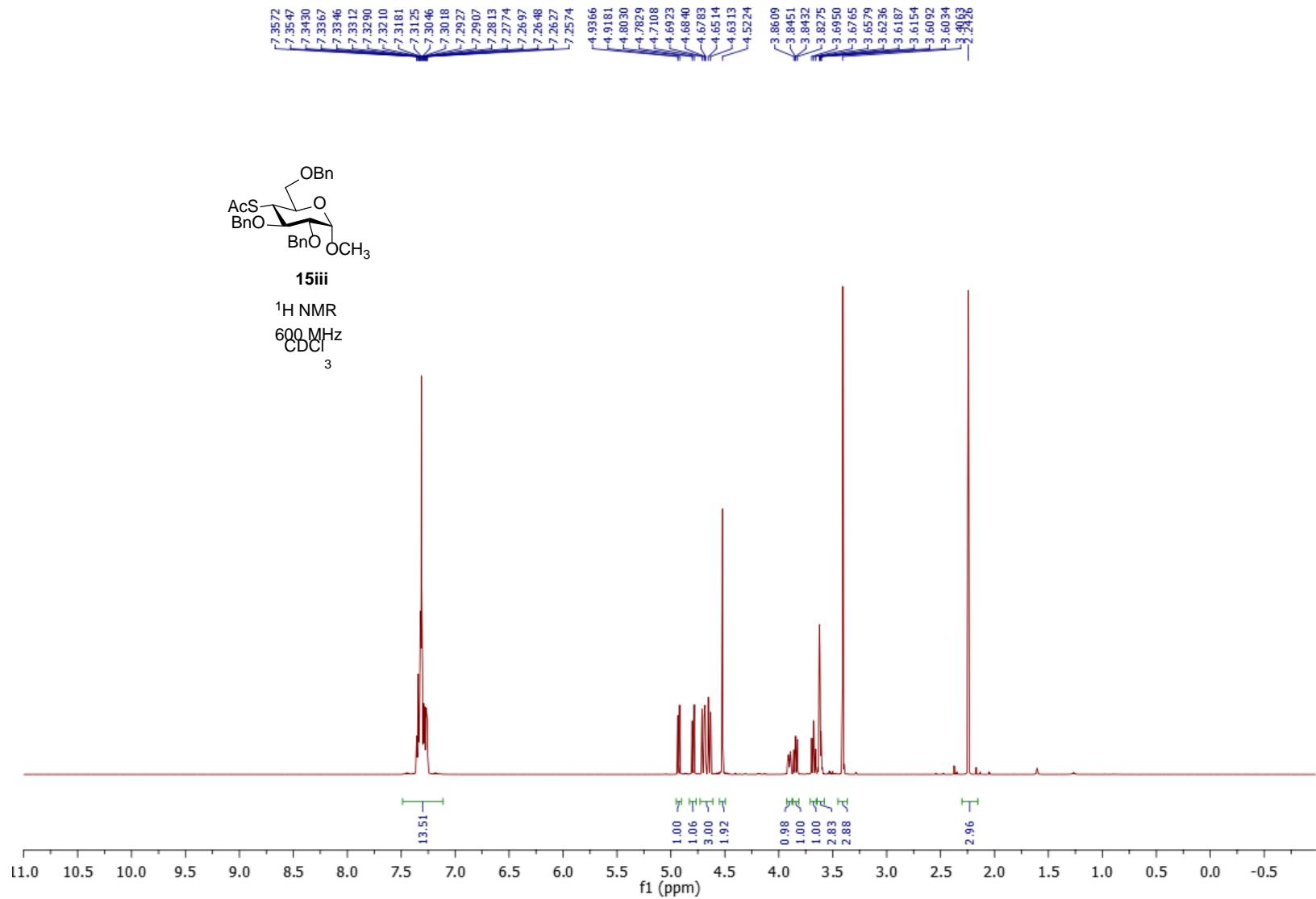




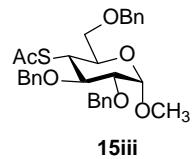






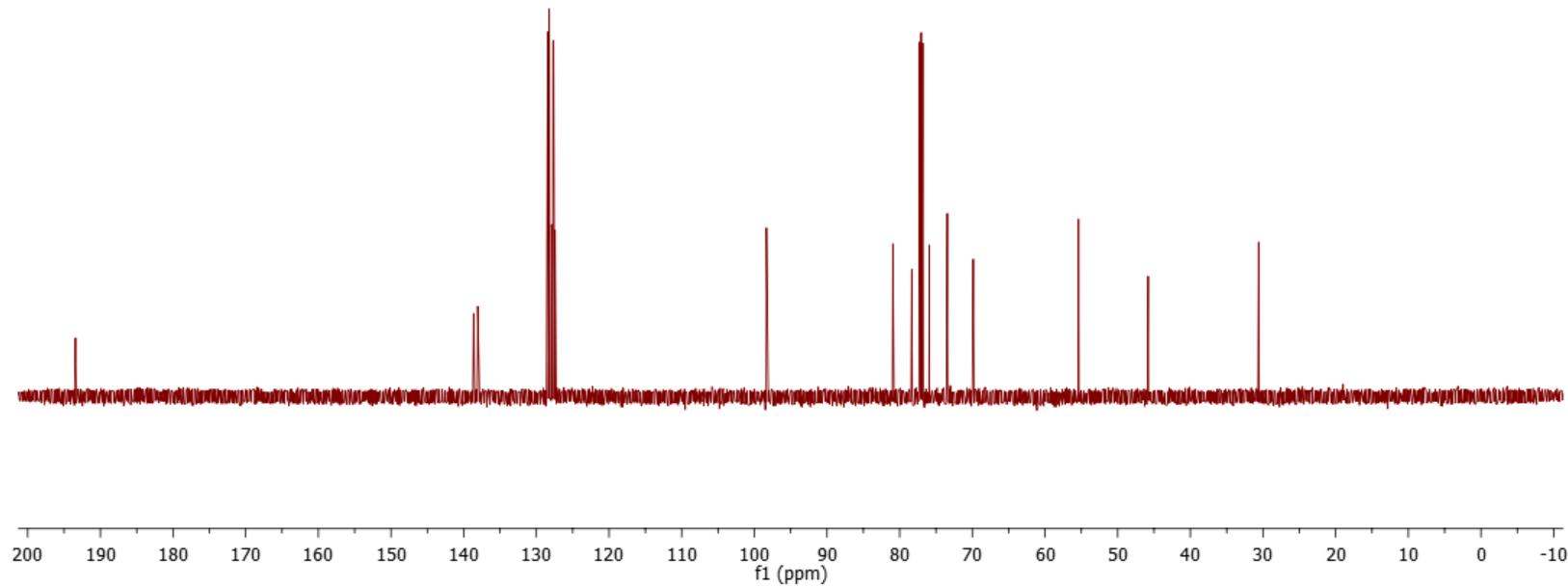


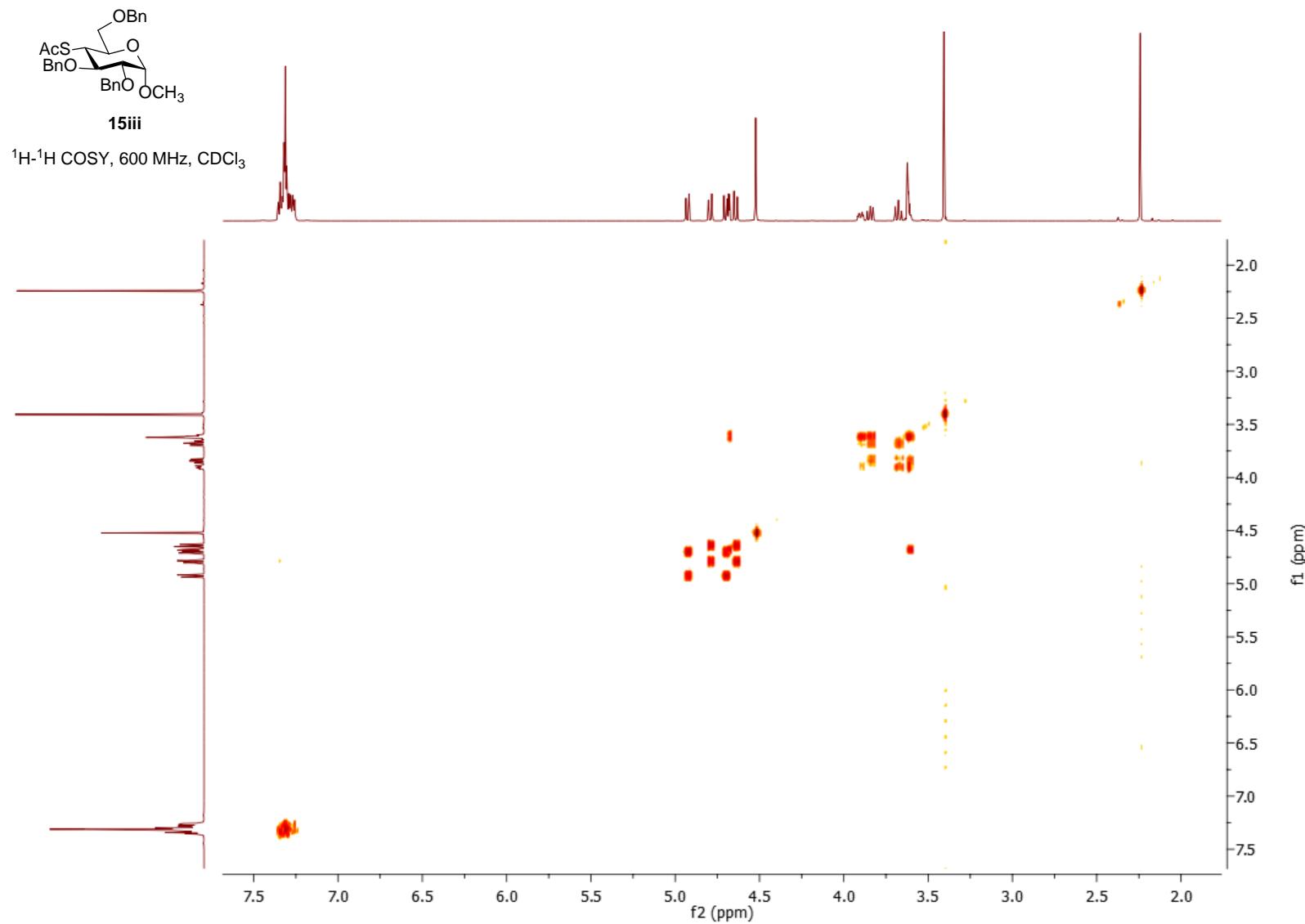
—193.4160

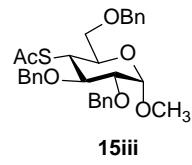


15iii

¹³C NMR, 150 MHz, CDCl₃

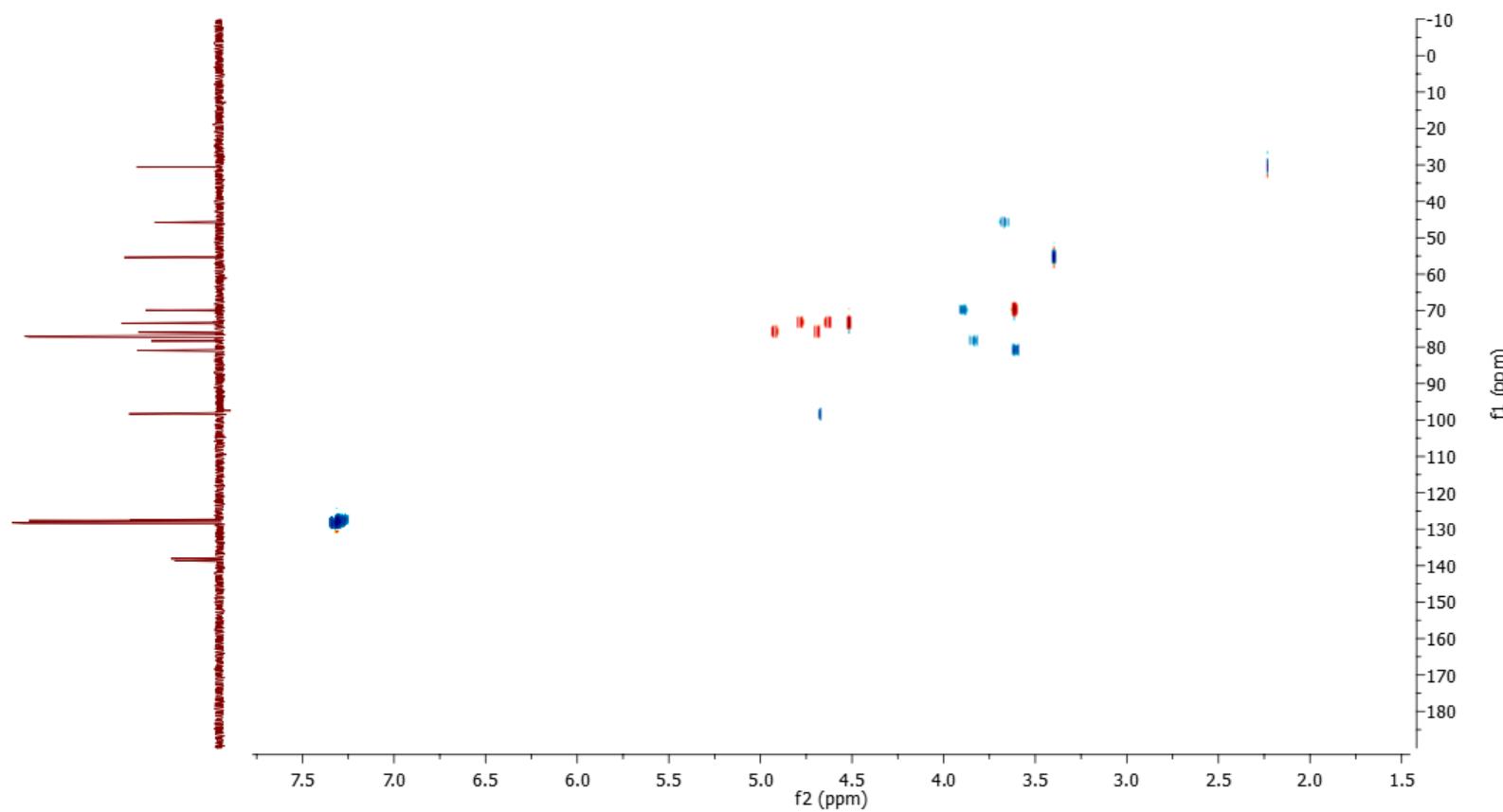


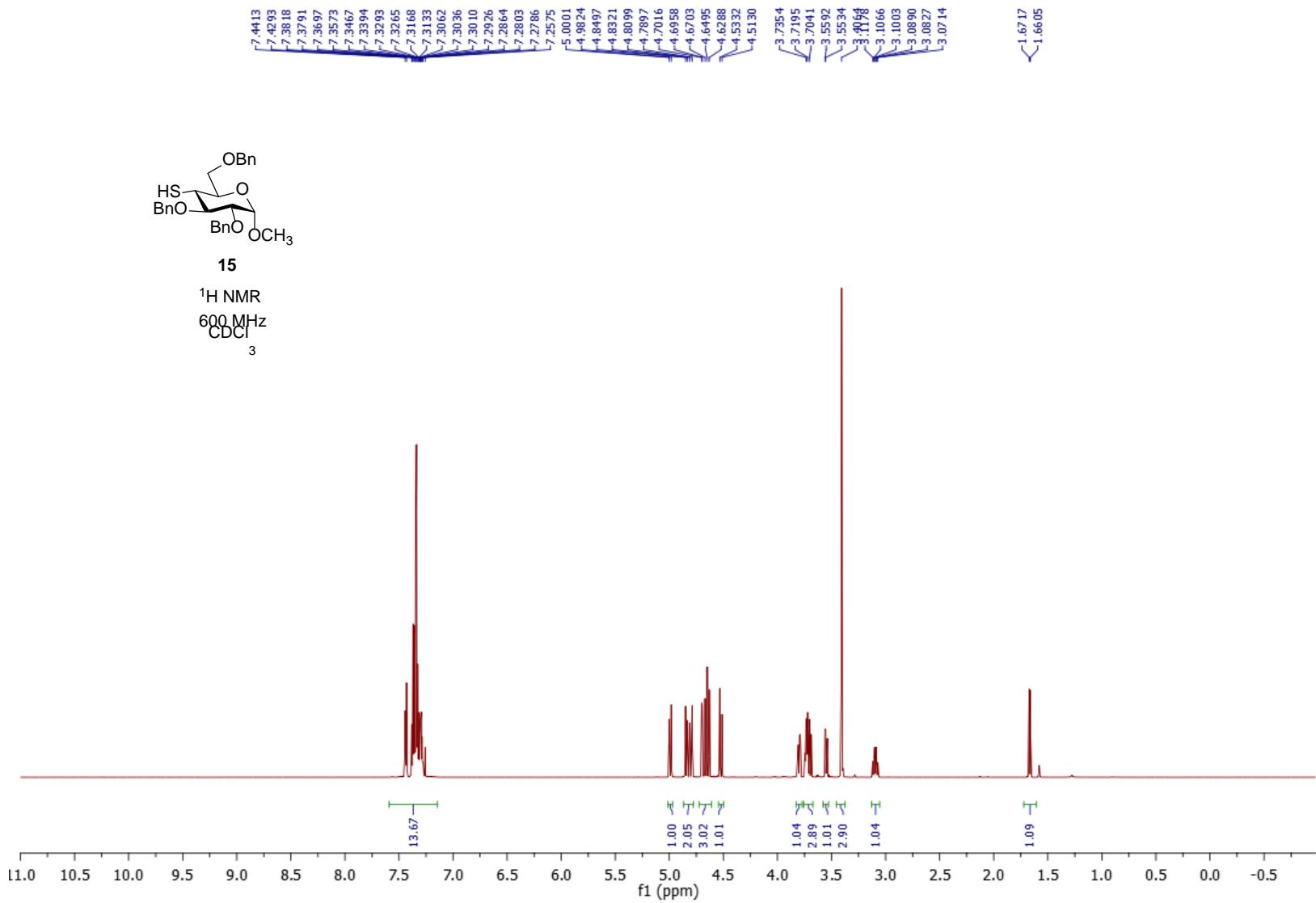


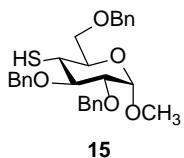


15iii

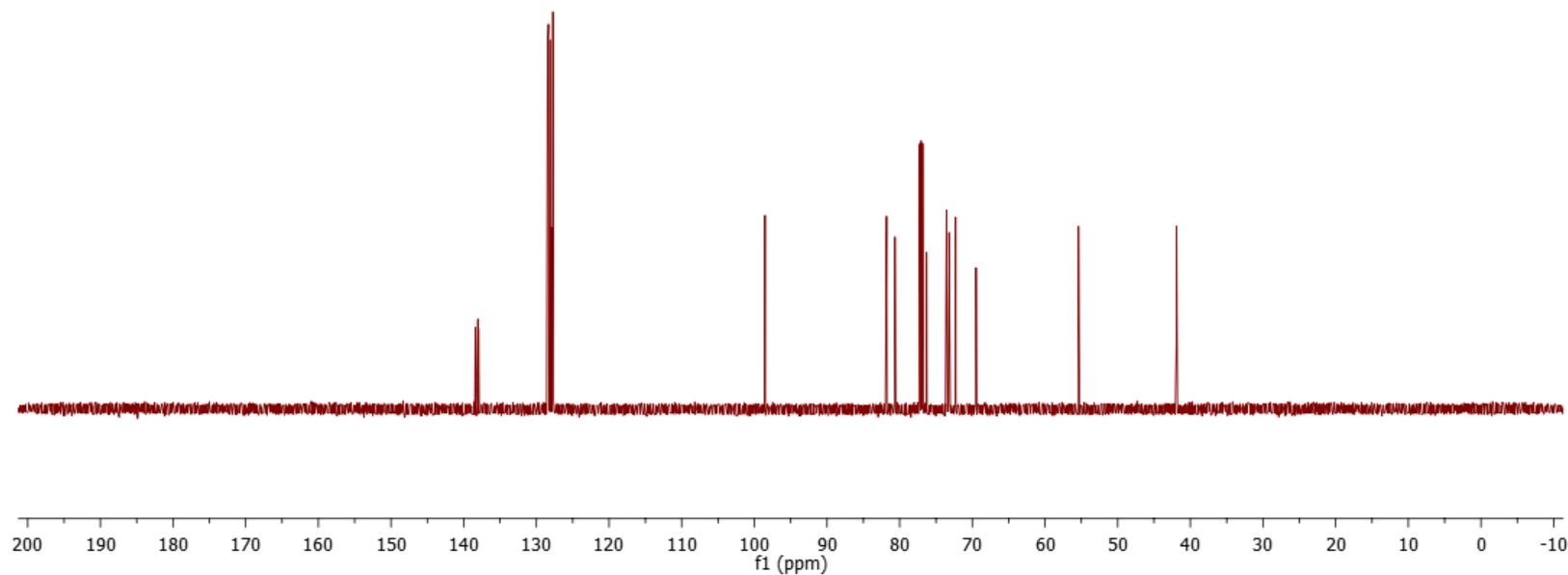
¹H-¹³C HSQC, 600/150MHz, CDCl₃

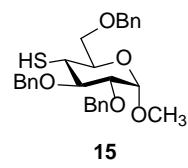




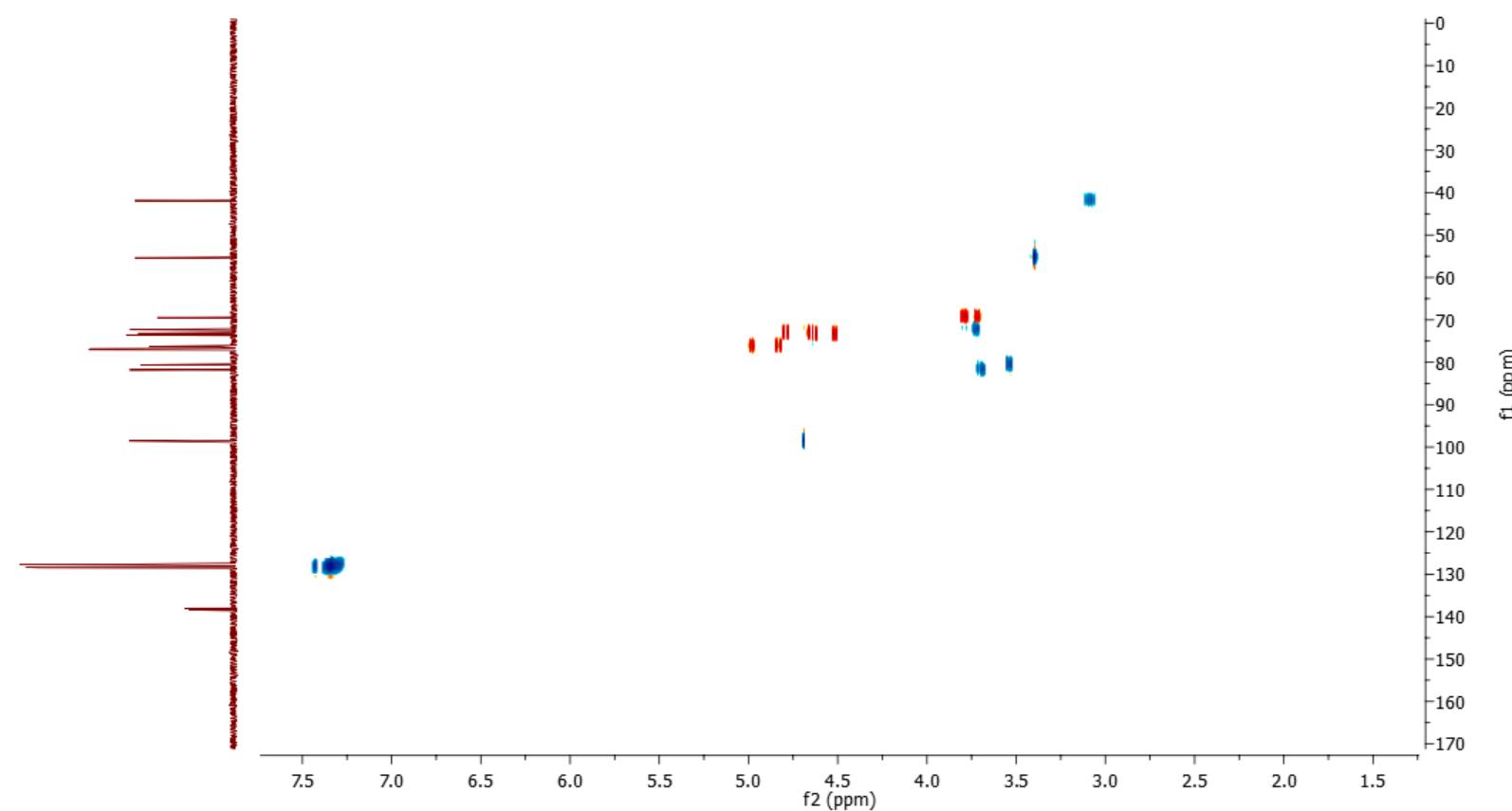


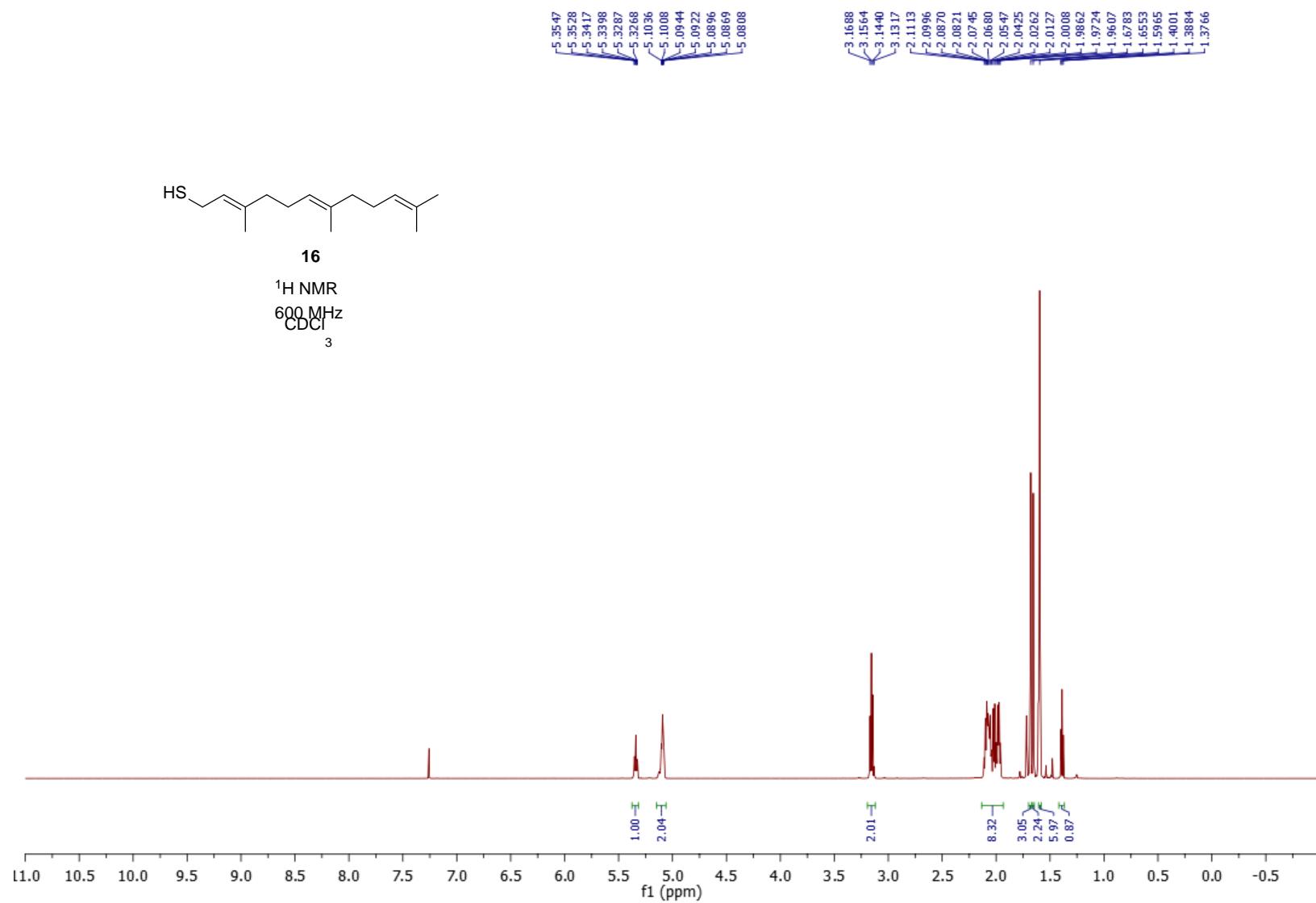
¹³C NMR, 150 MHz, CDCl₃

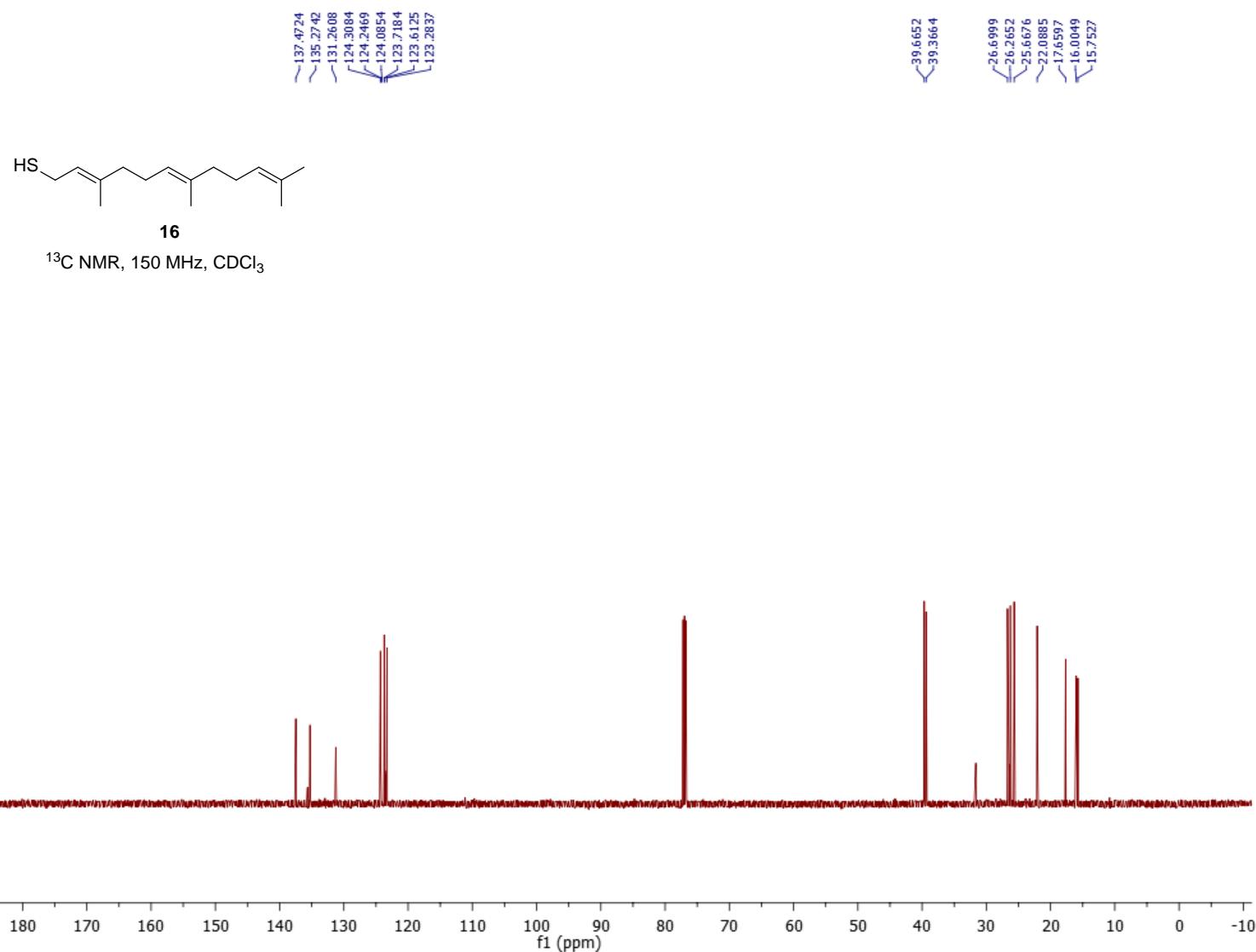


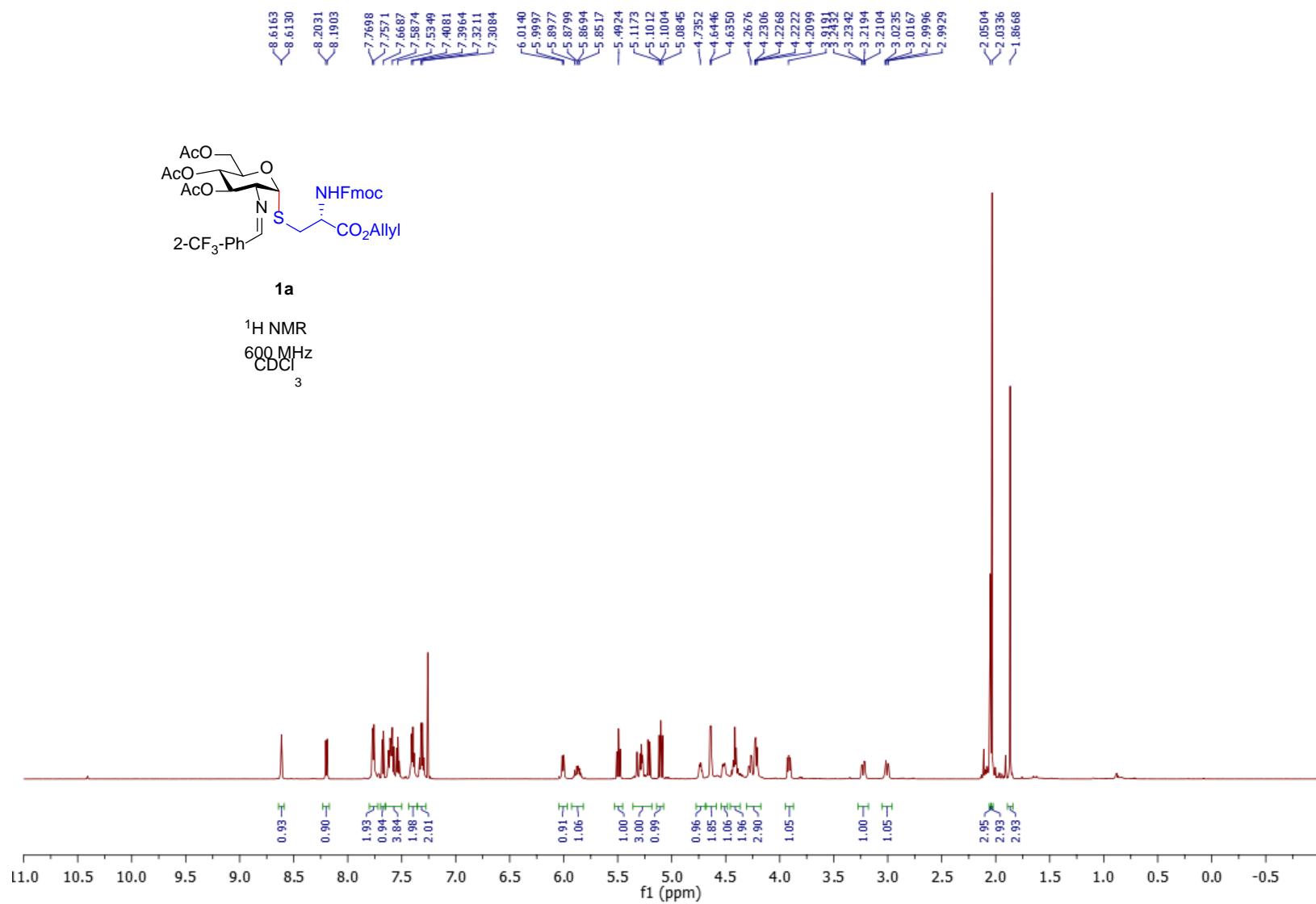


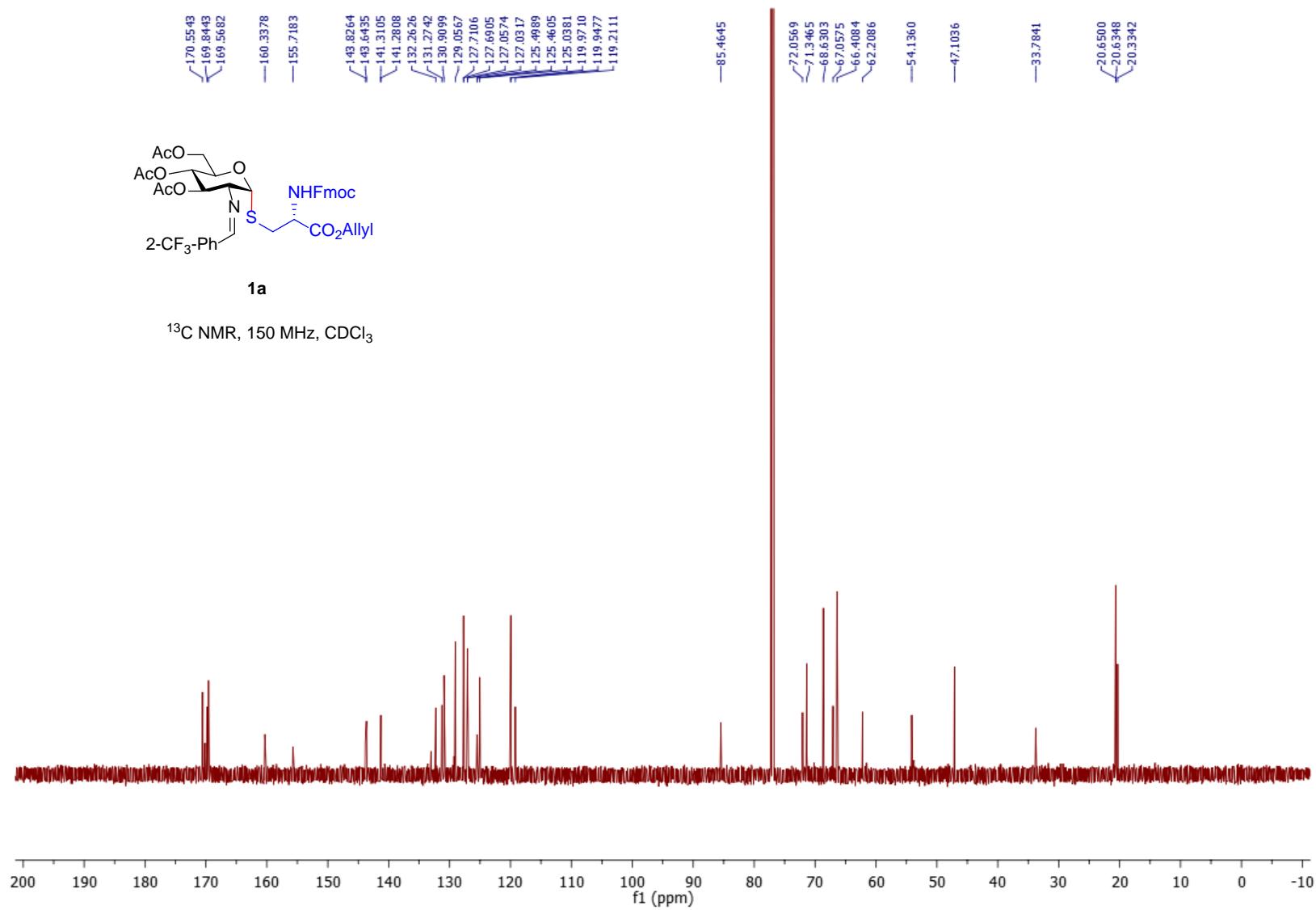
¹H-¹³C HSQC, 600/150MHz, CDCl₃

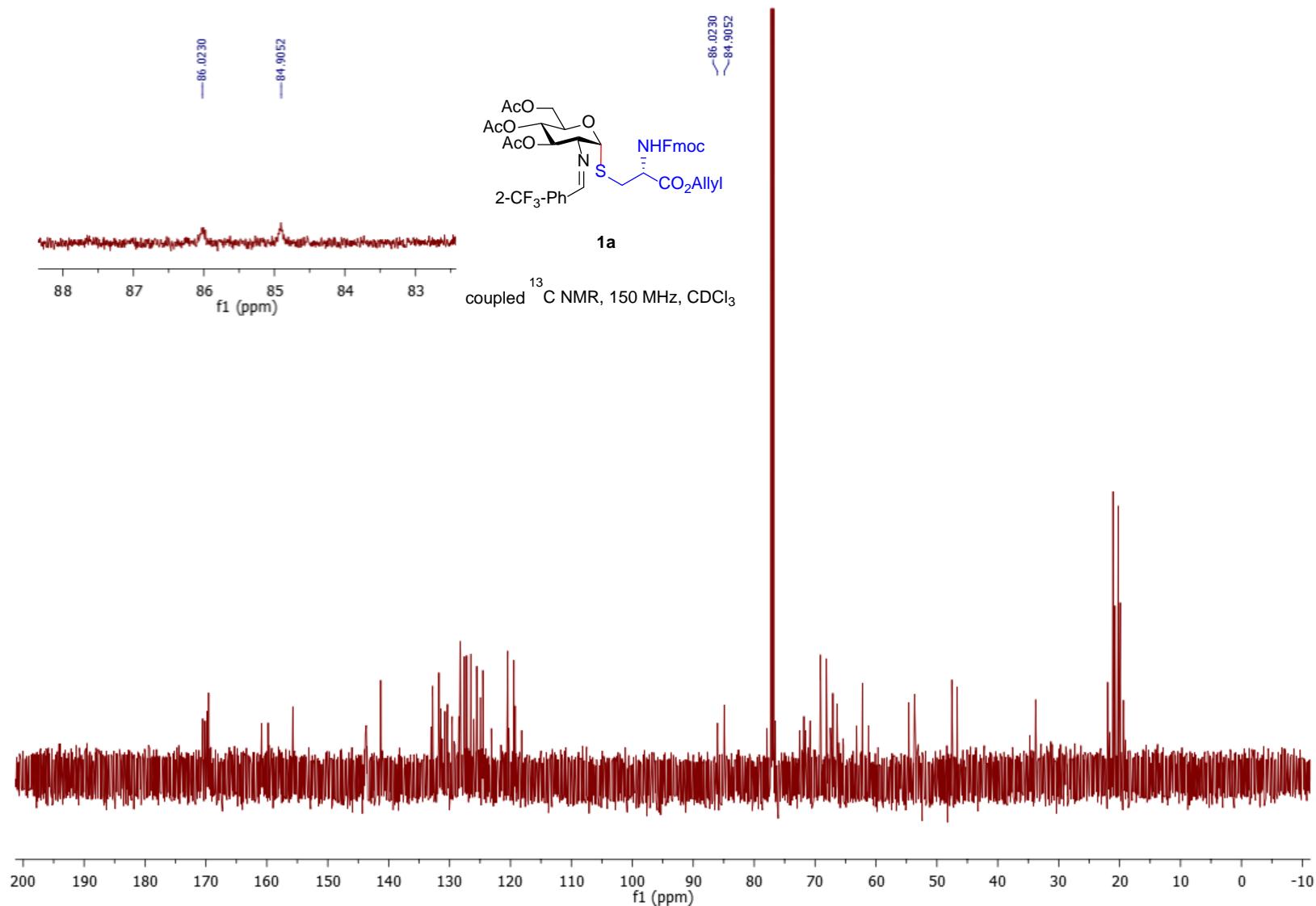


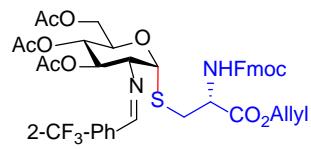






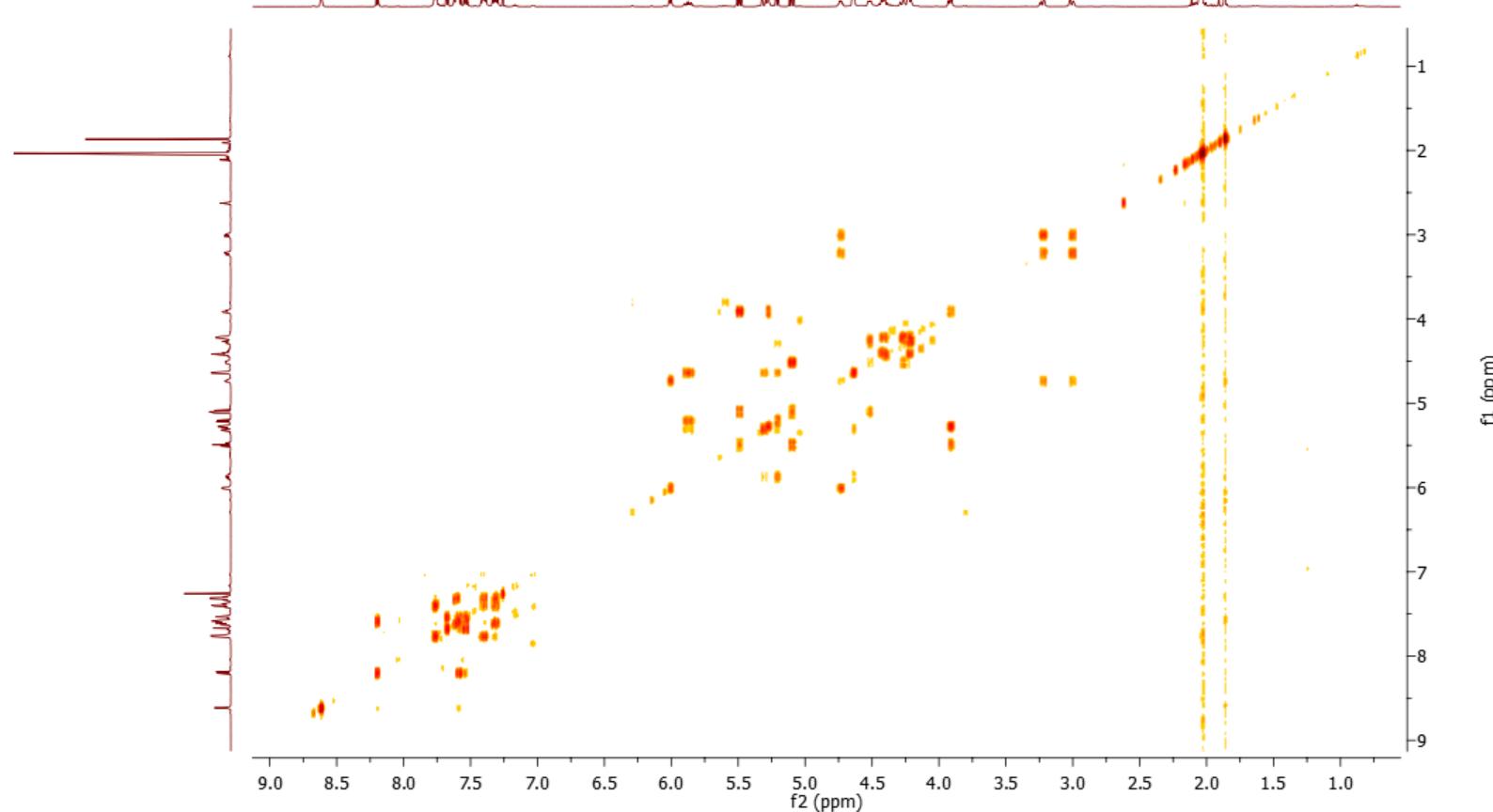


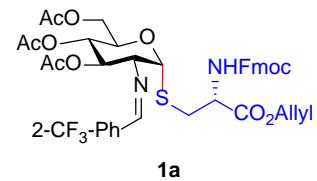




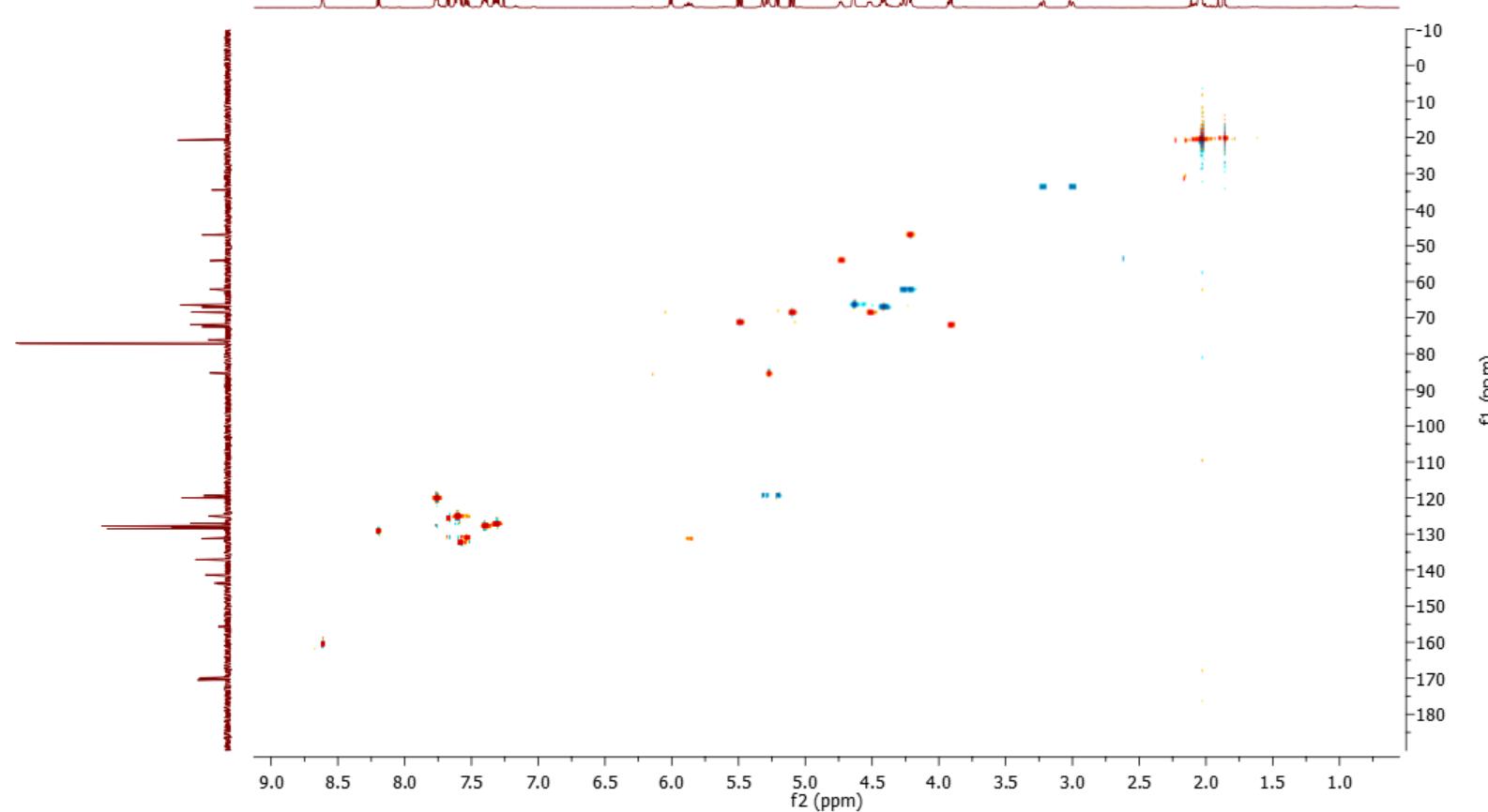
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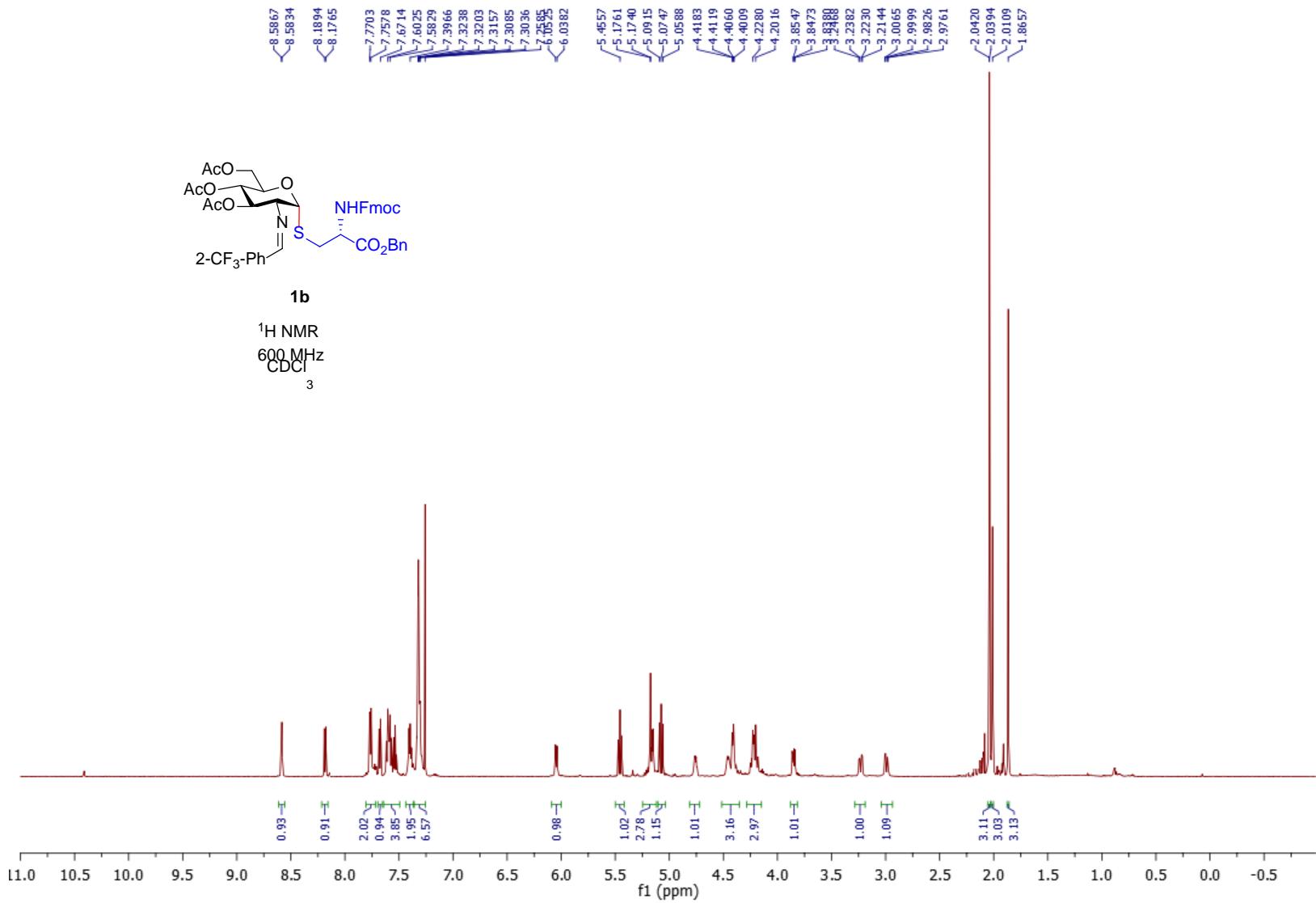
^1H - ^1H COSY, 600 MHz, CDCl_3

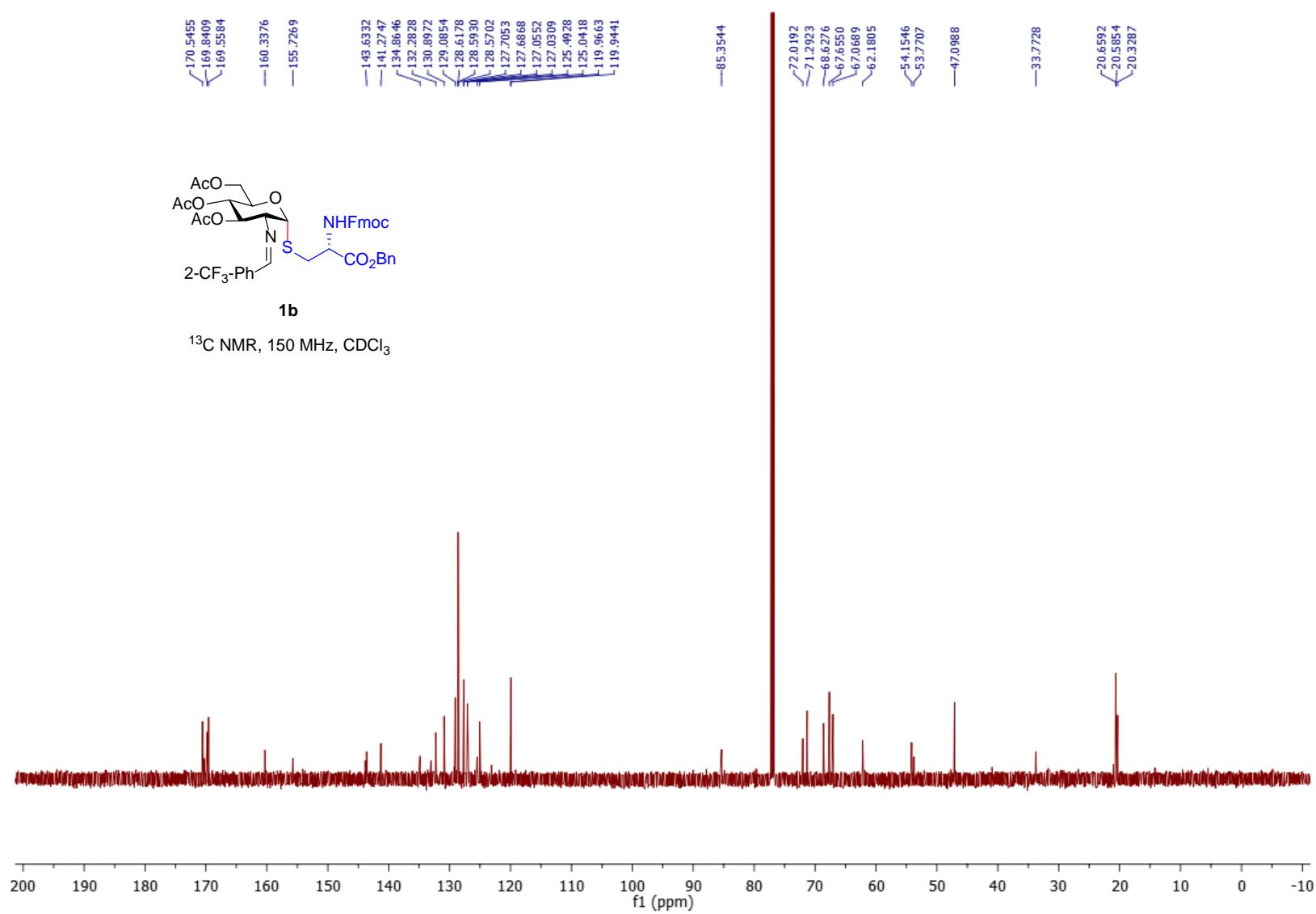


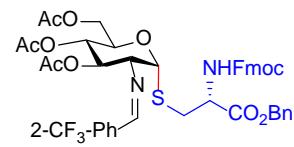


^1H - ^{13}C HSQC, 600/150MHz, CDCl_3

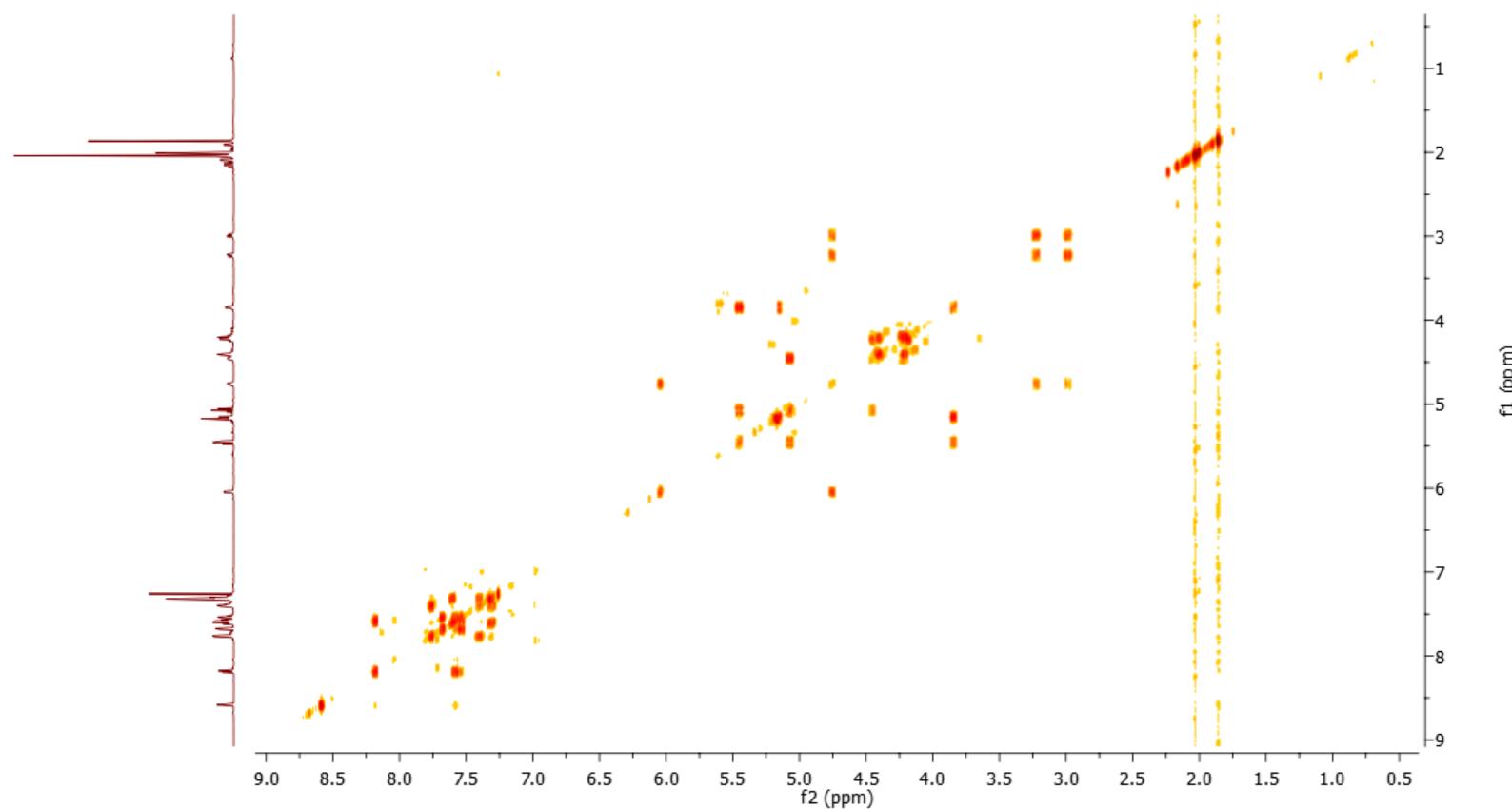


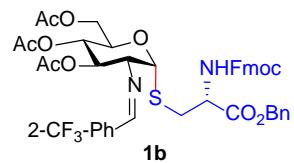




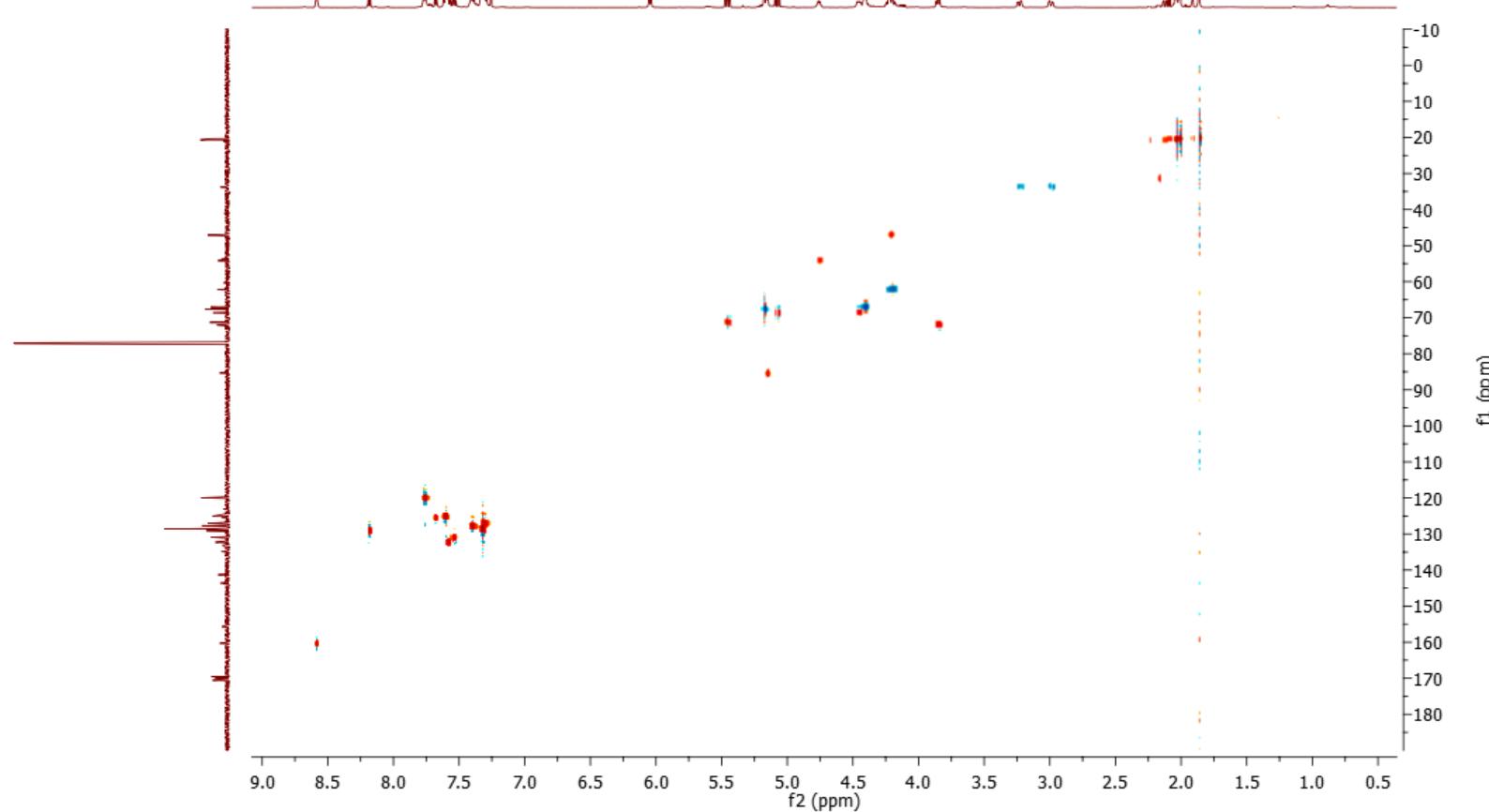


^1H - ^1H COSY, 600 MHz, CDCl_3

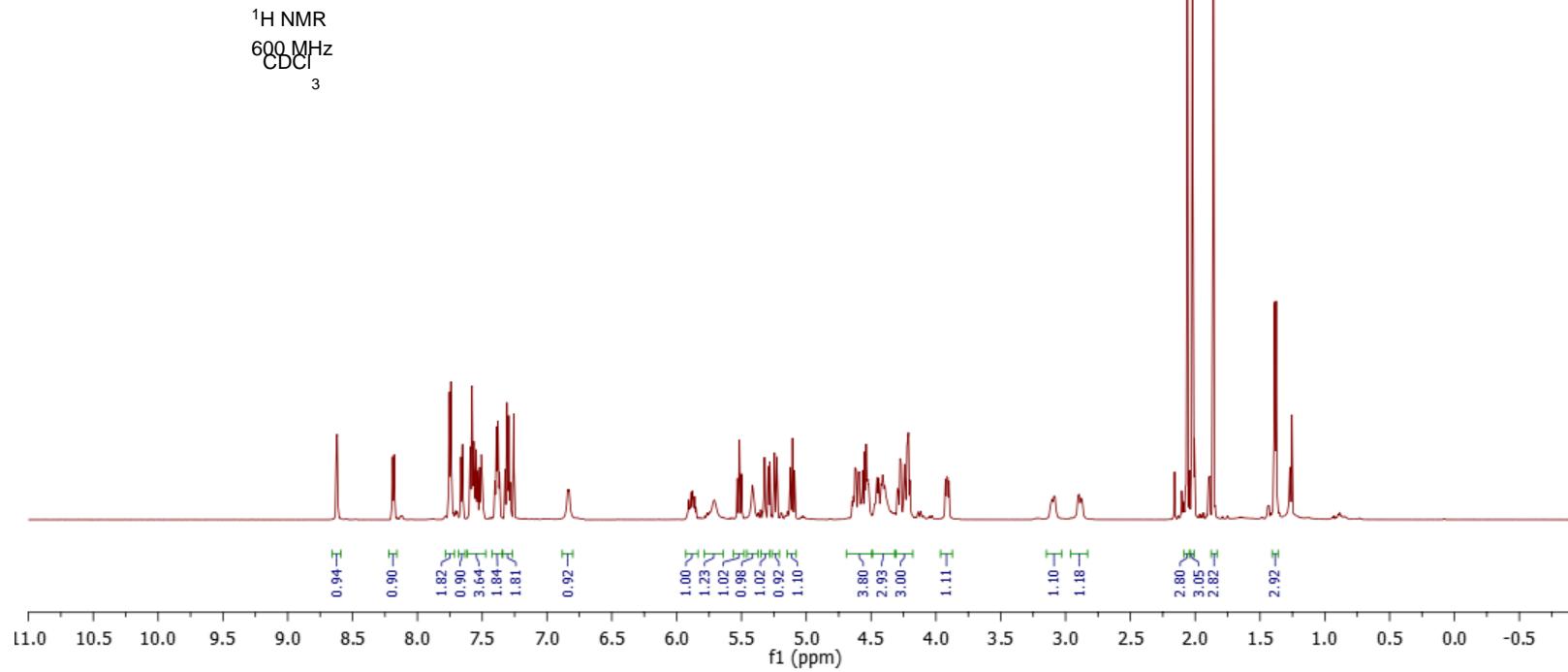
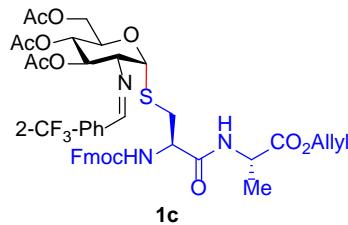


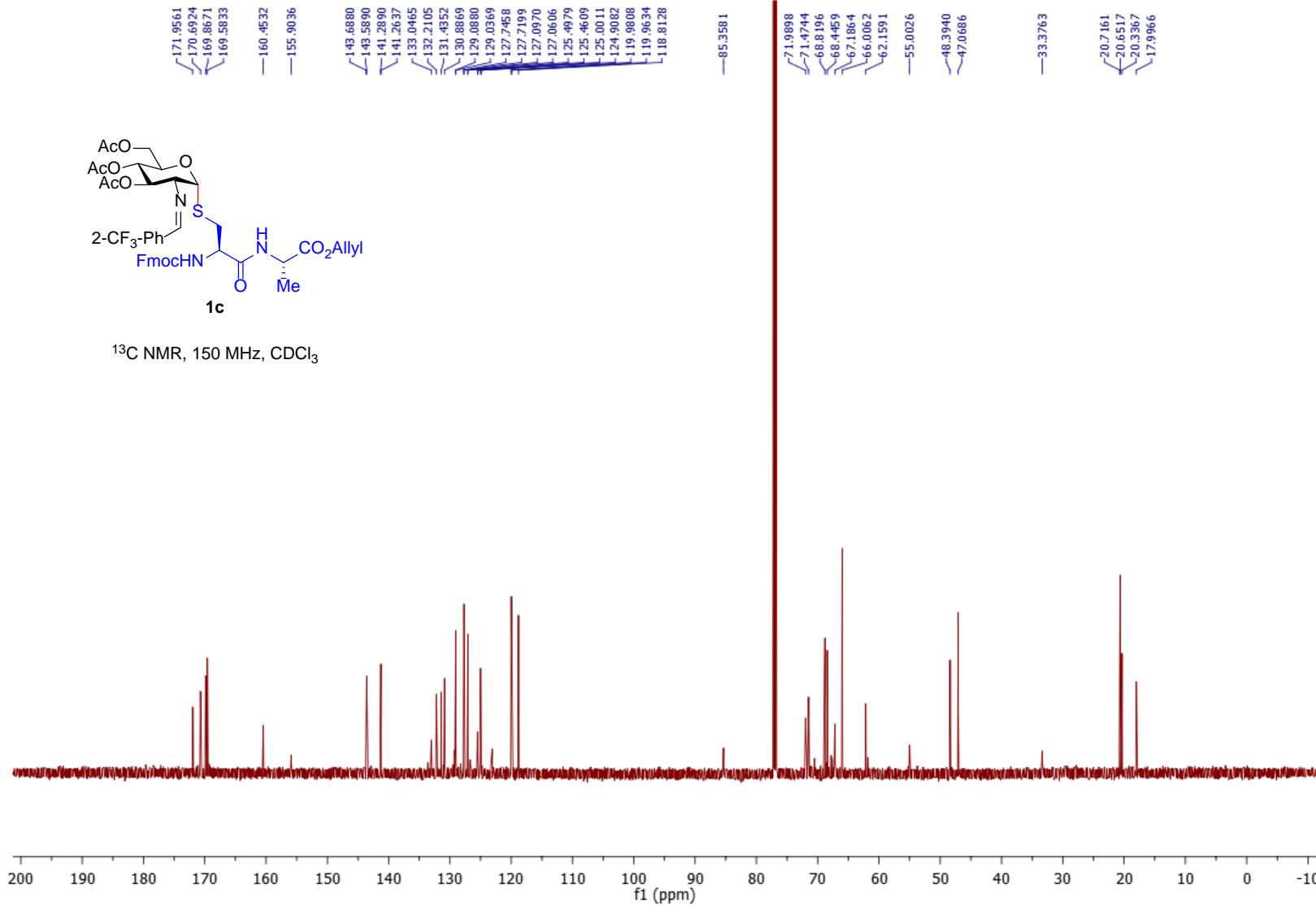


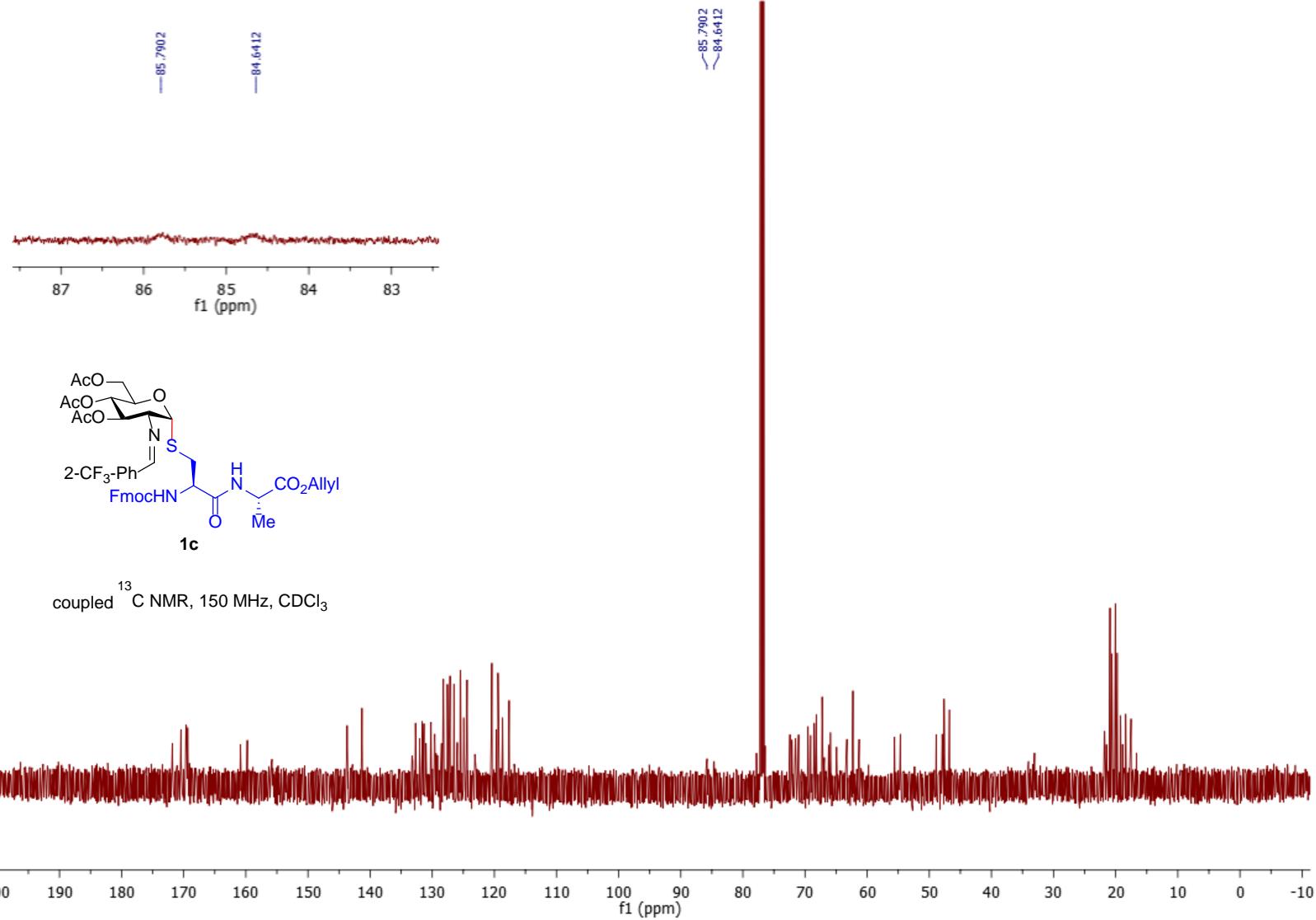
^1H - ^{13}C HSQC, 600/150MHz, CDCl_3

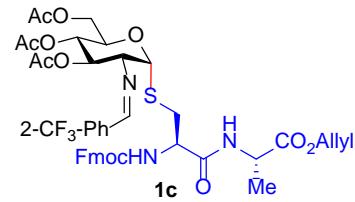


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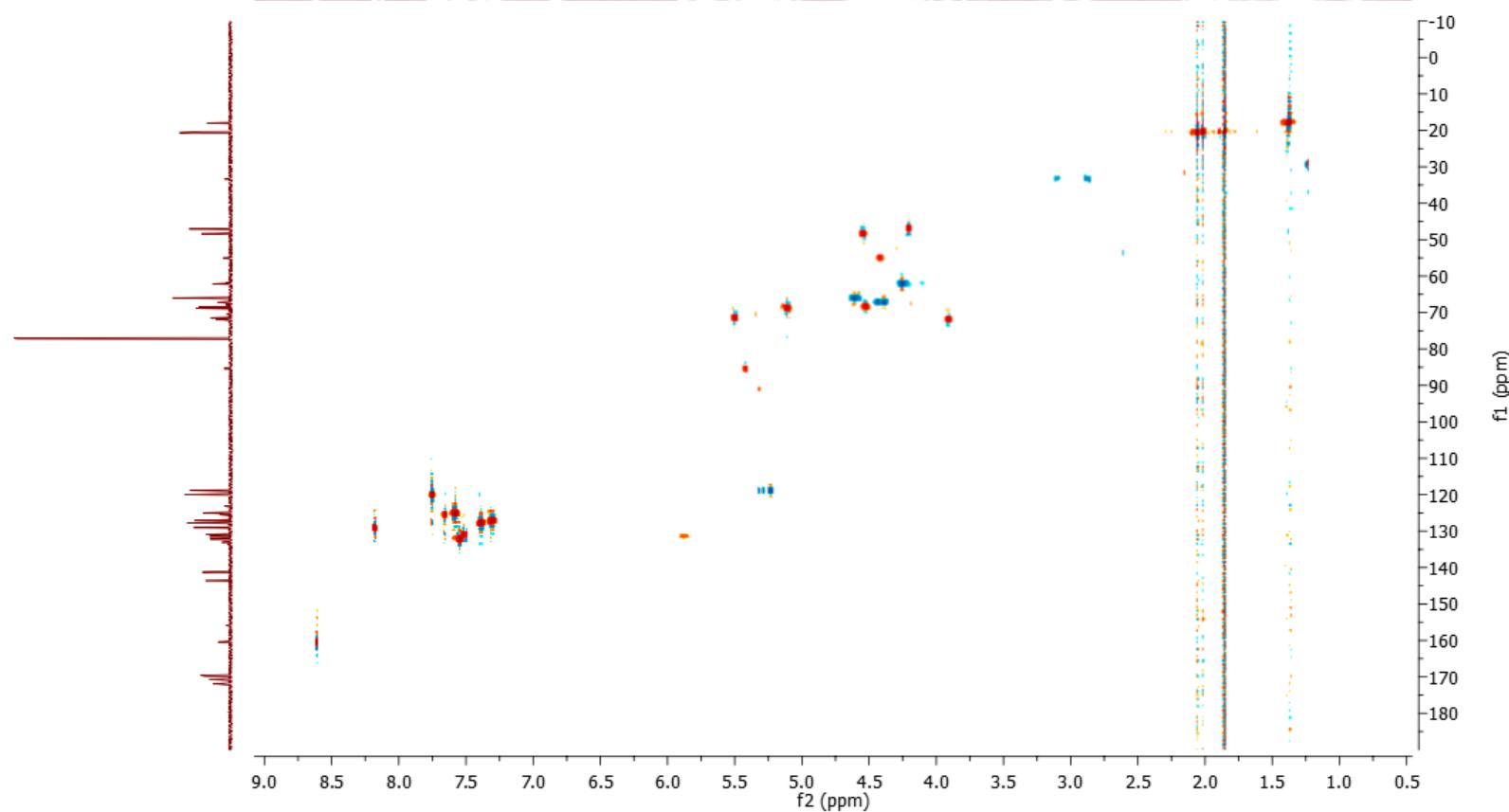


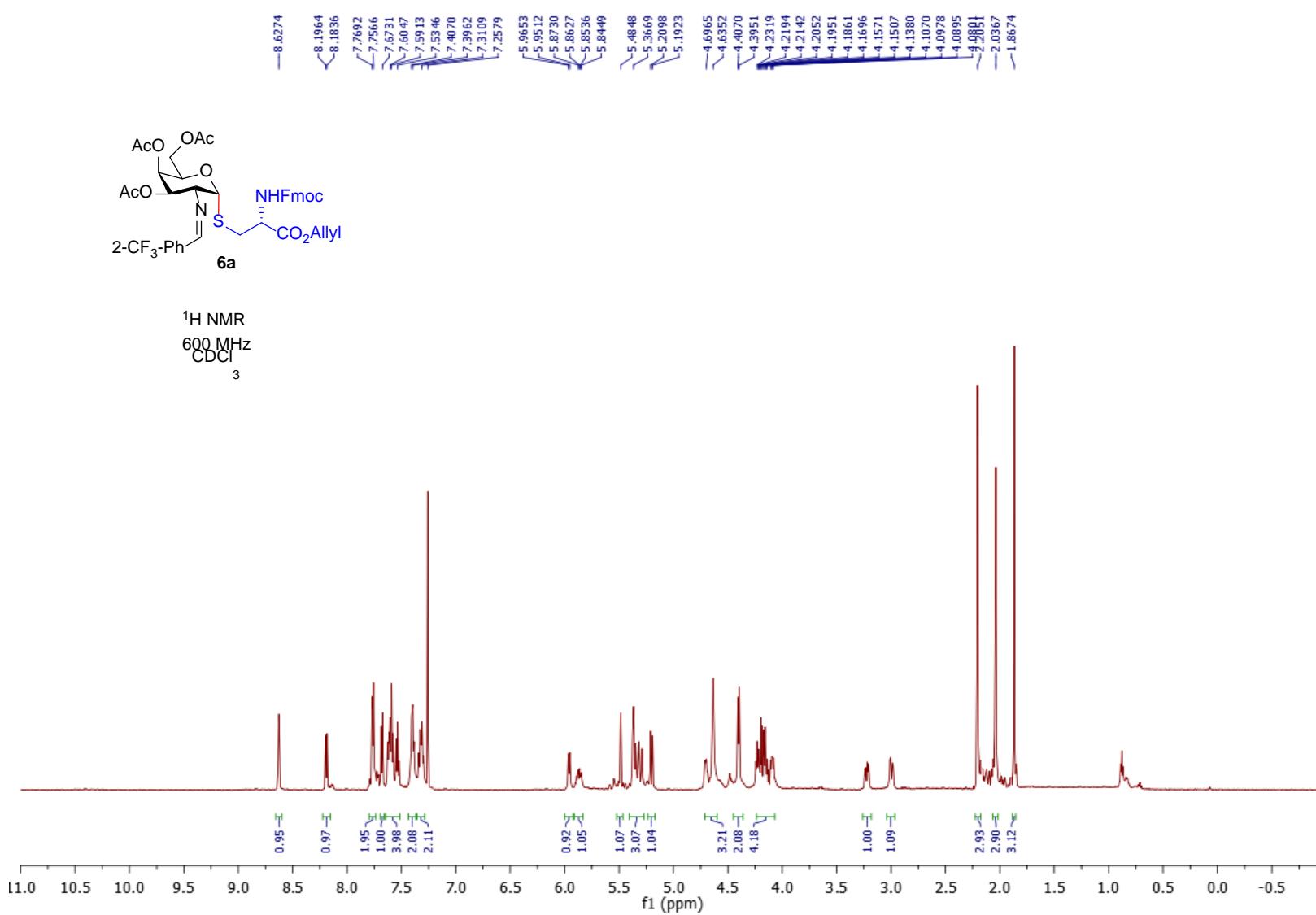


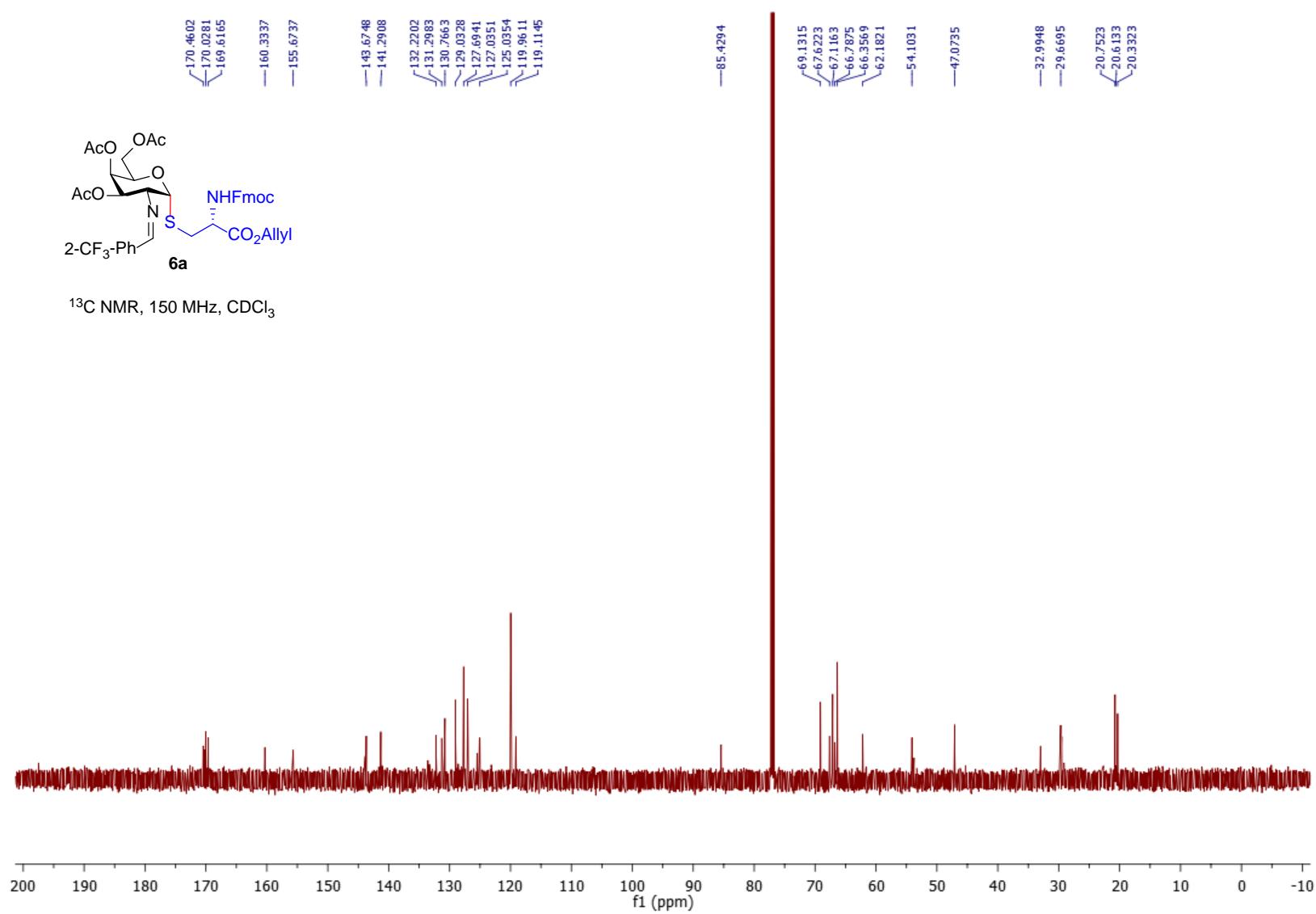


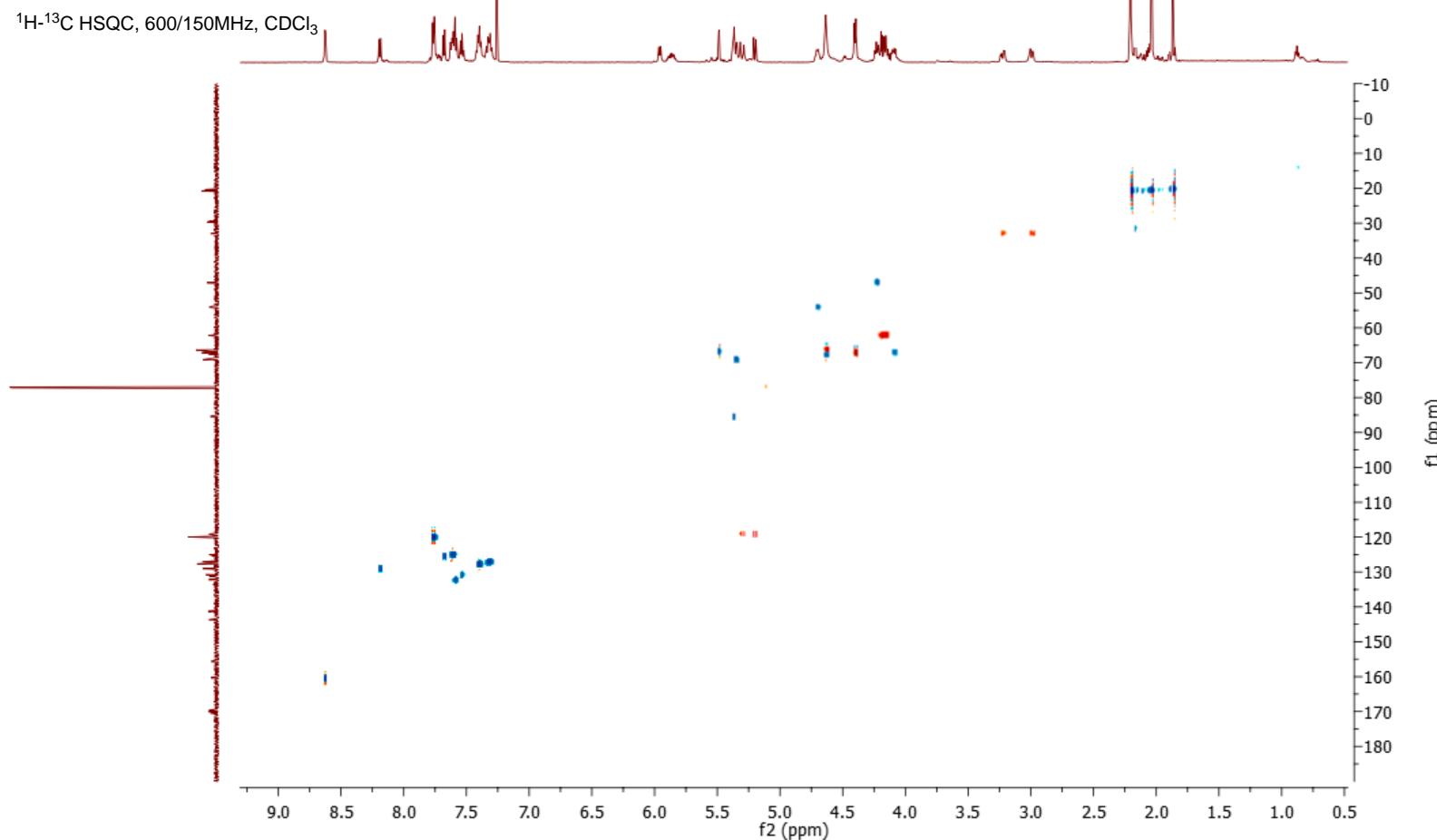
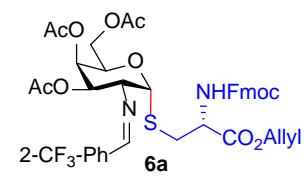


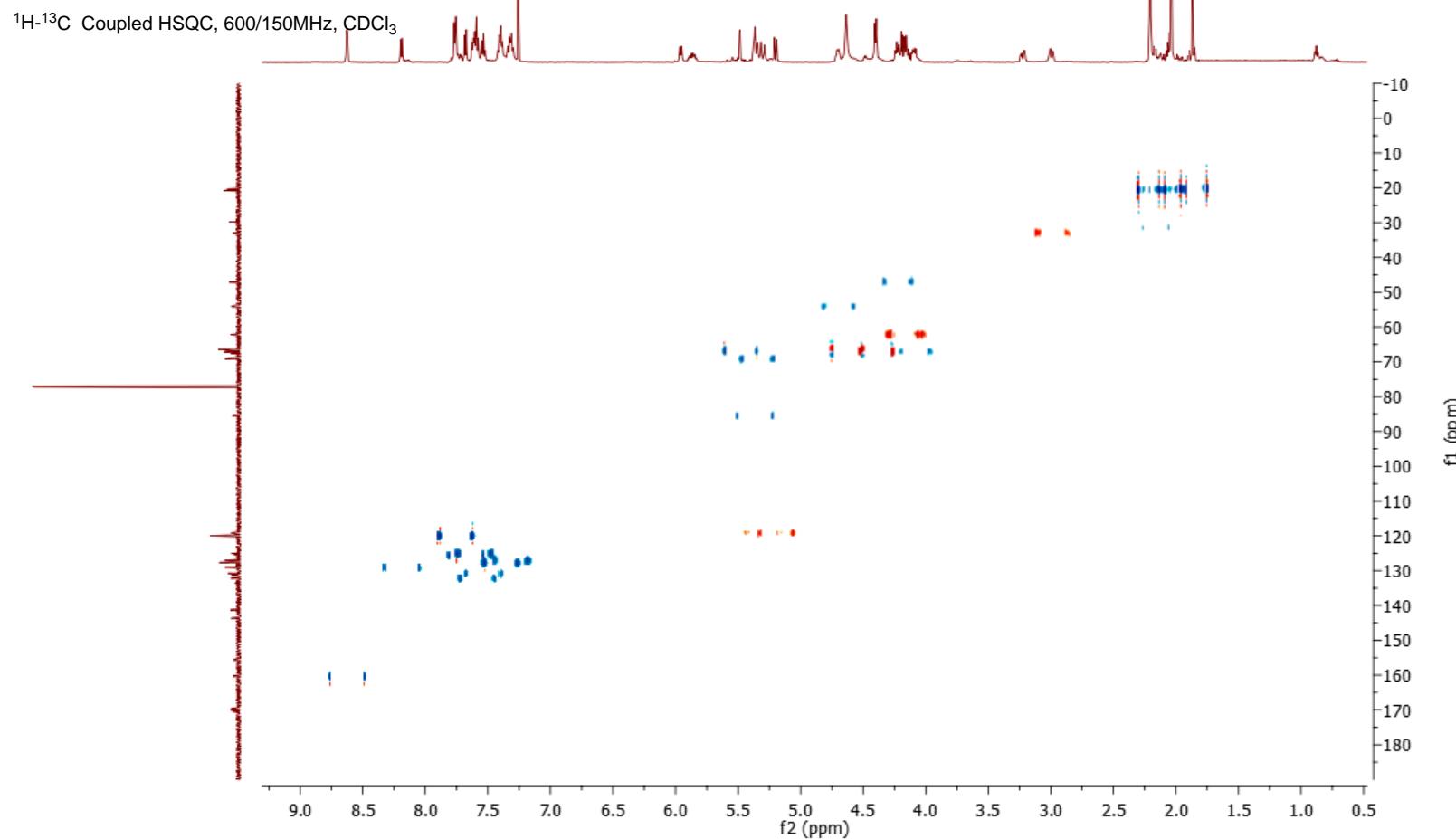
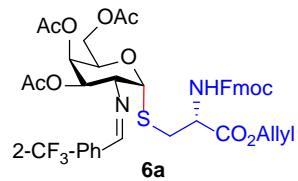
^1H - ^{13}C HSQC, 600/150MHz, CDCl_3

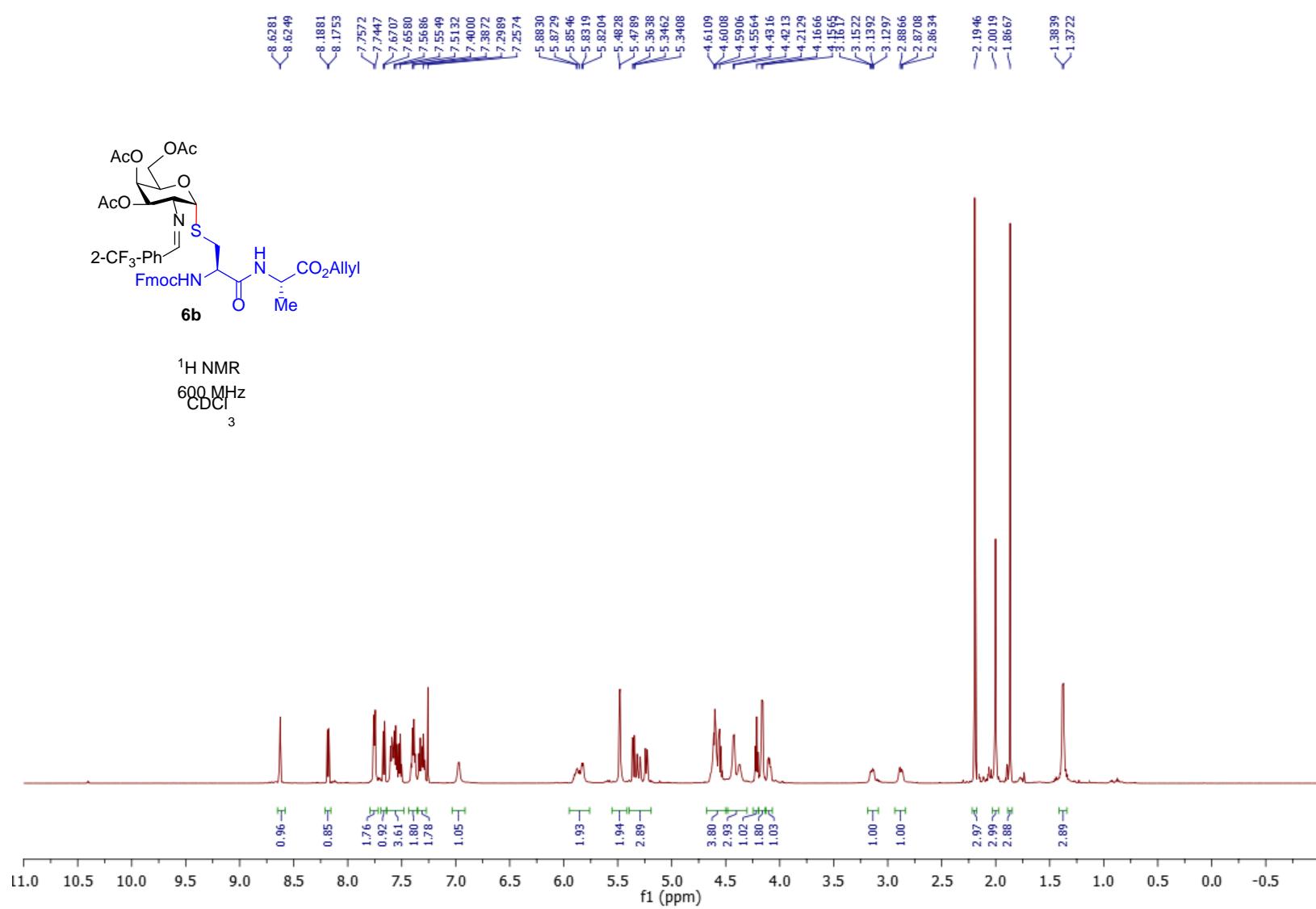


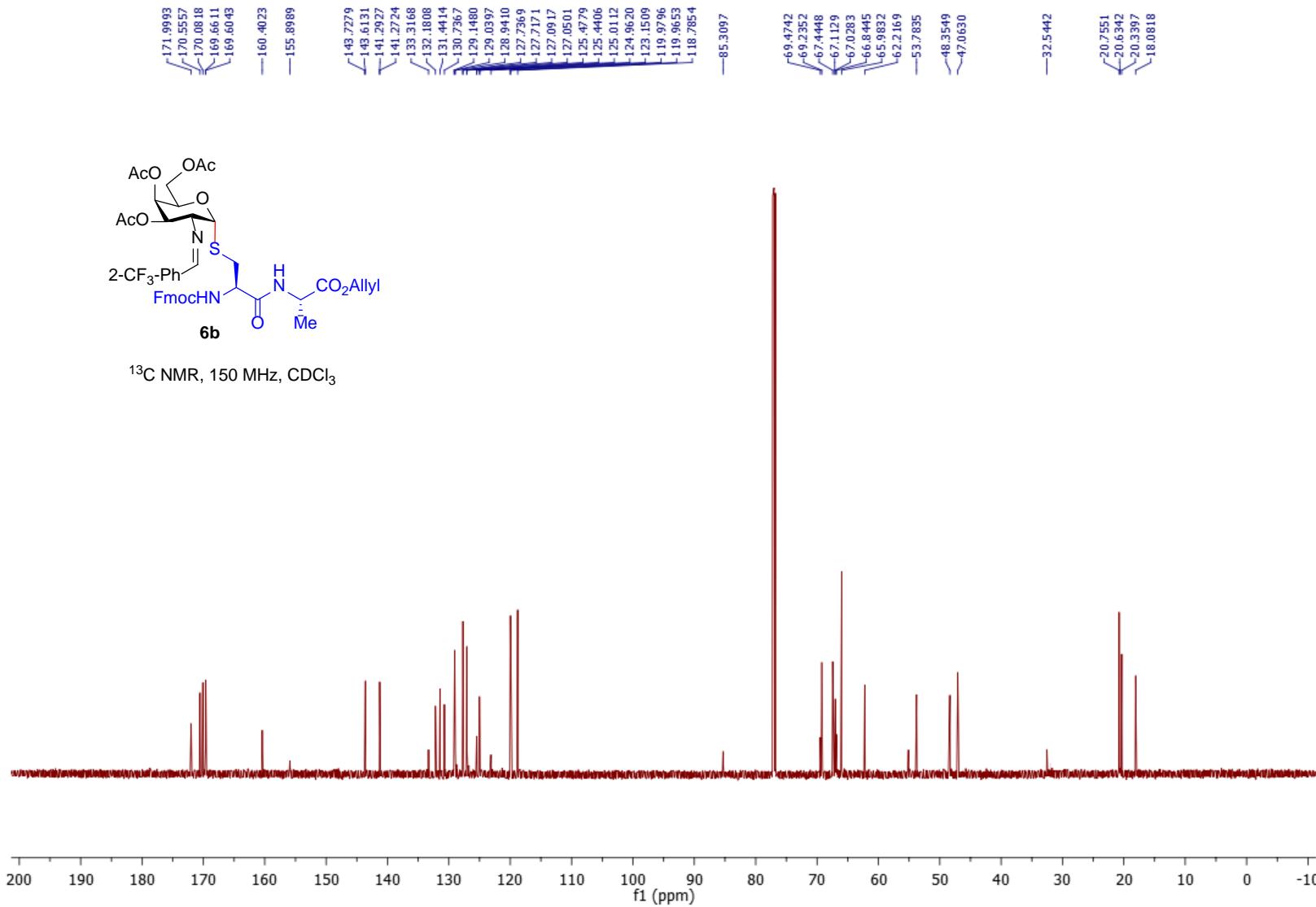


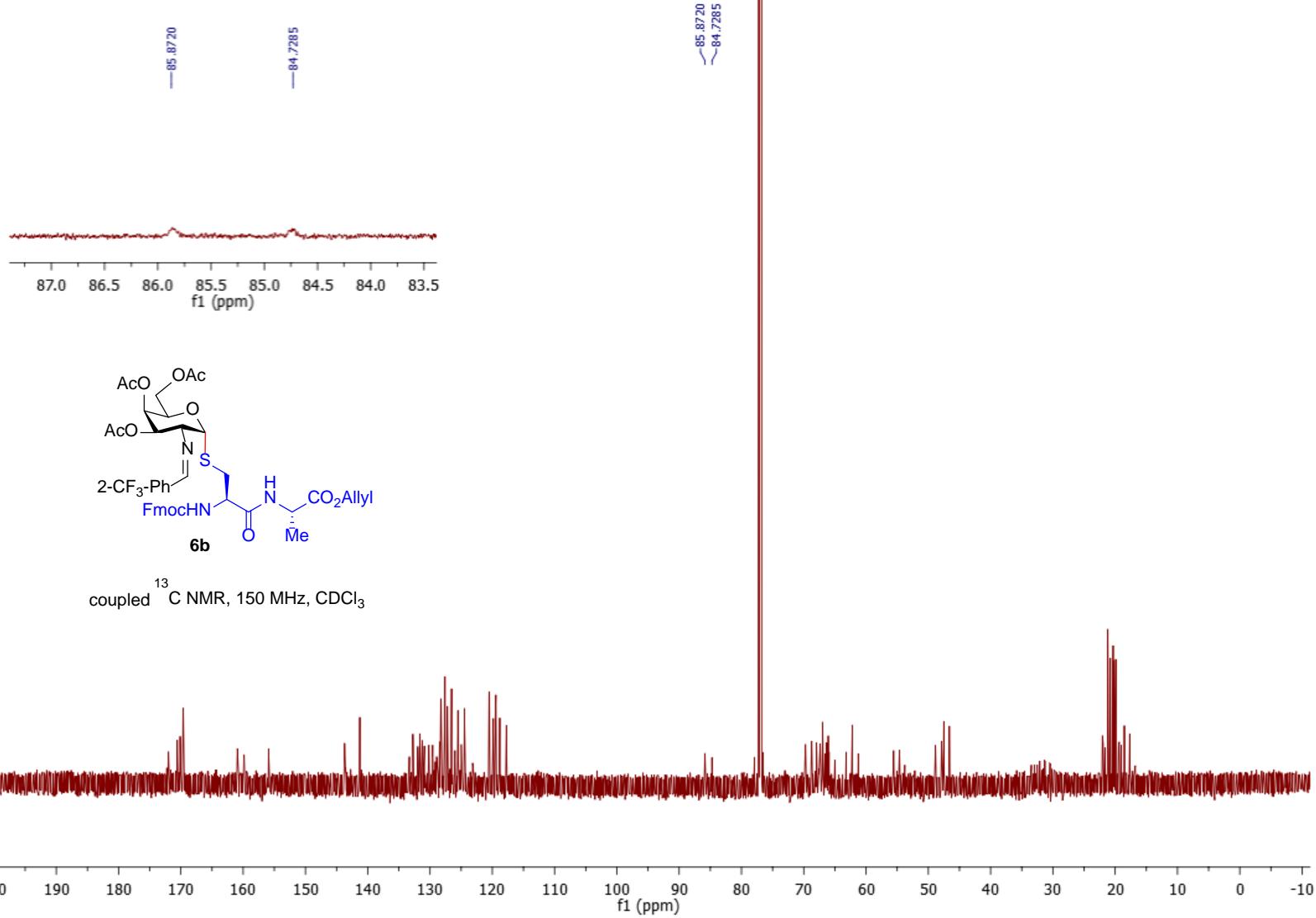


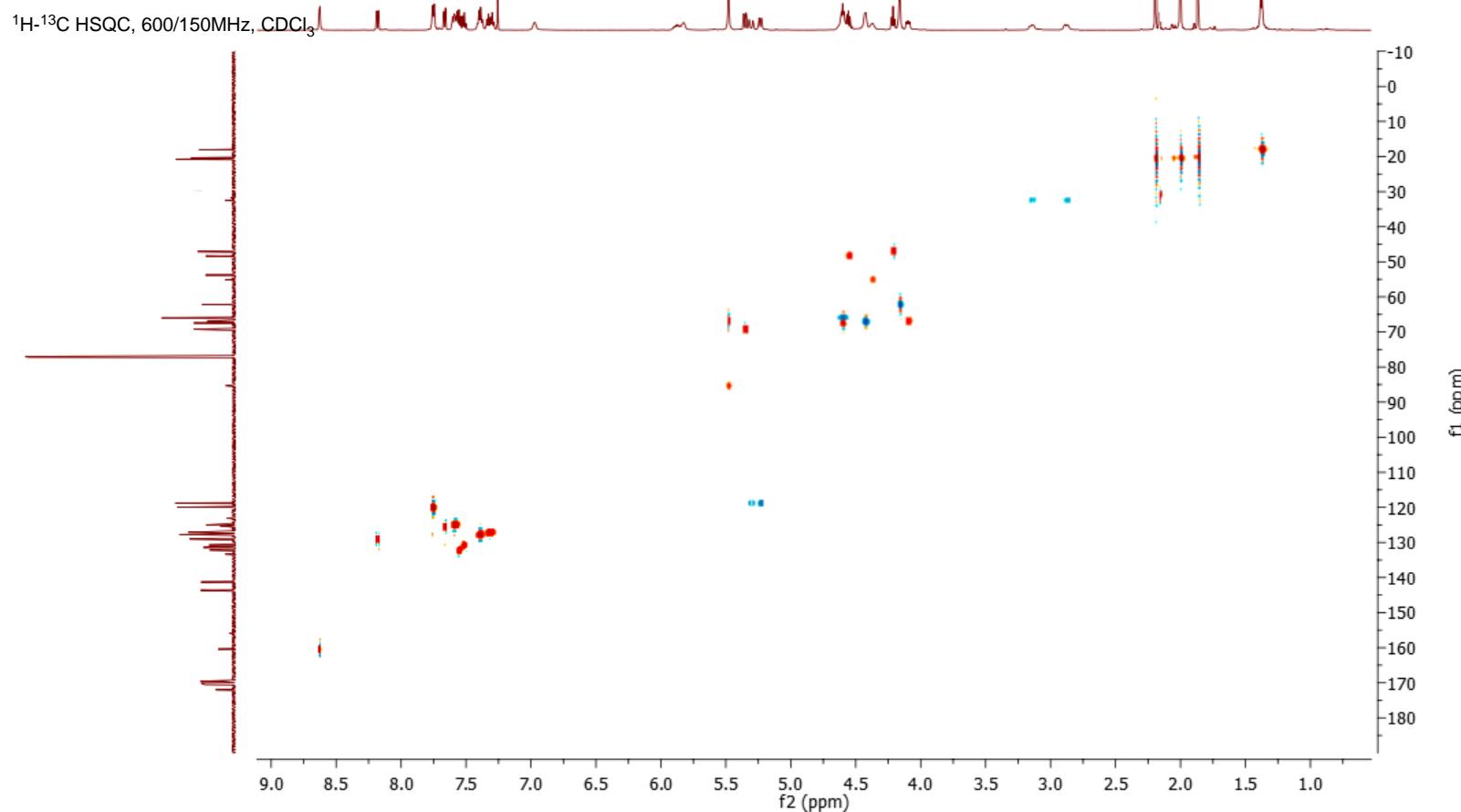
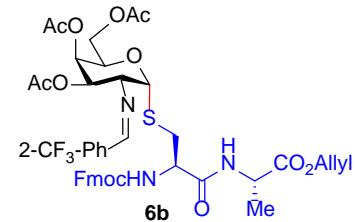


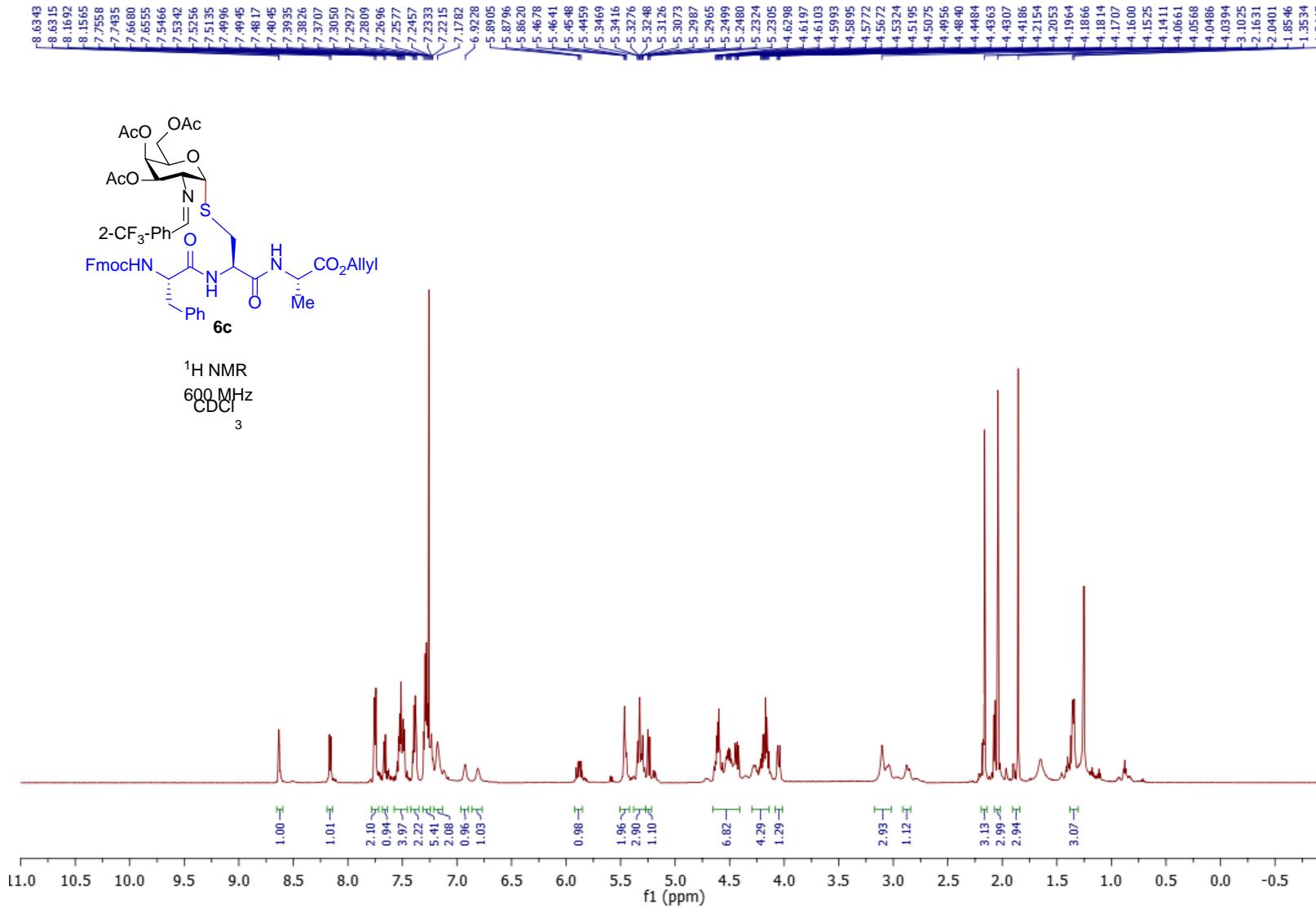


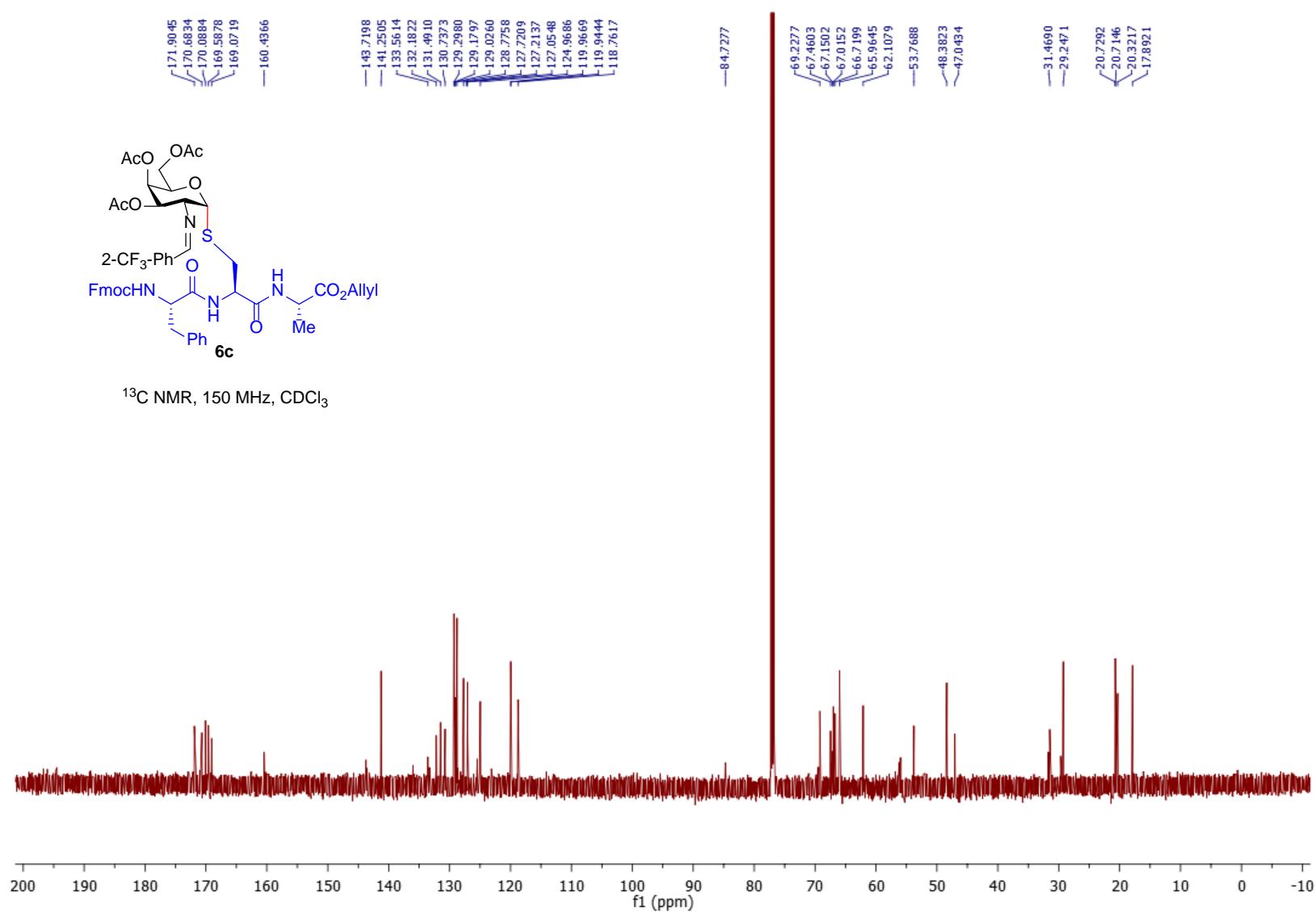


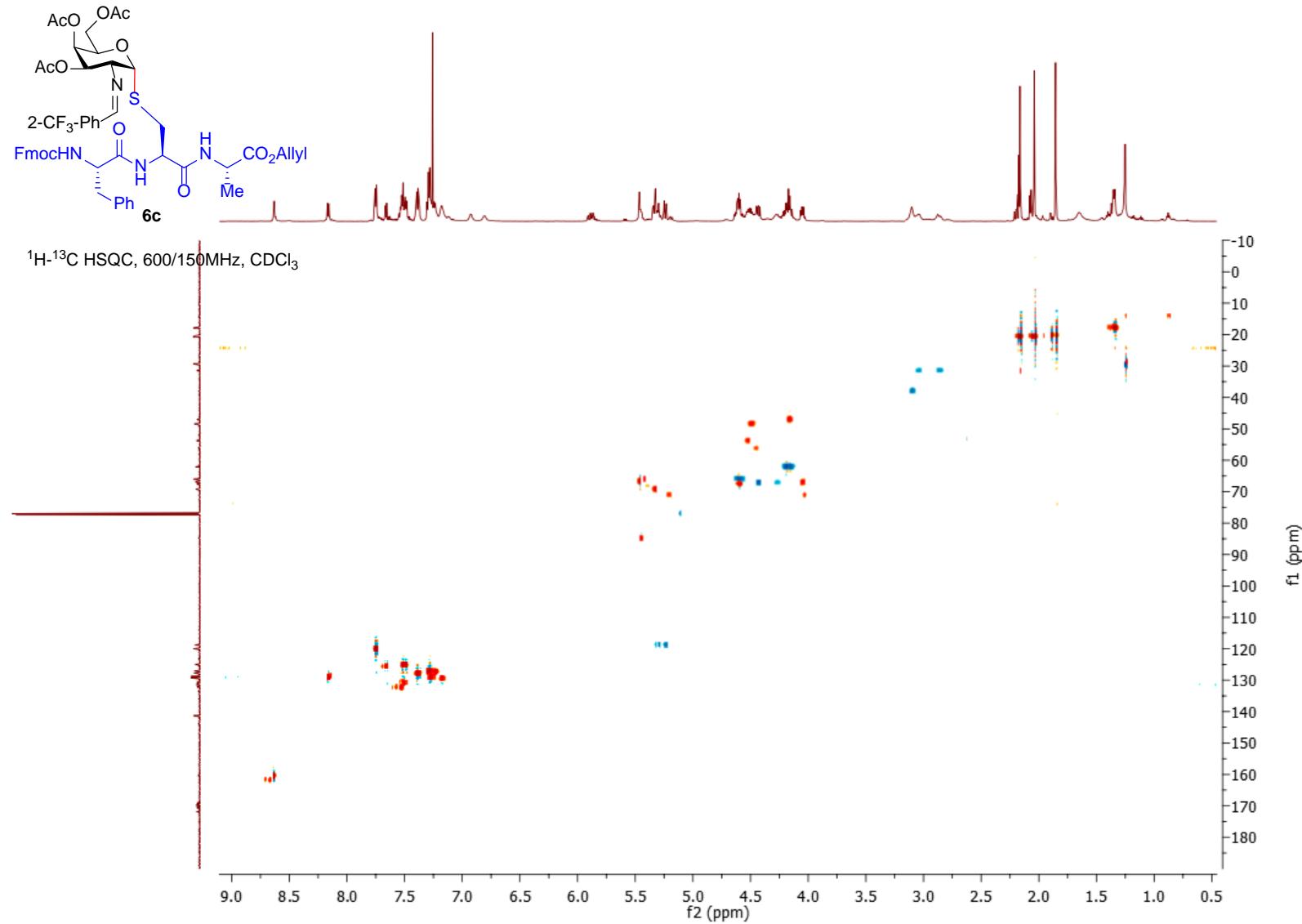


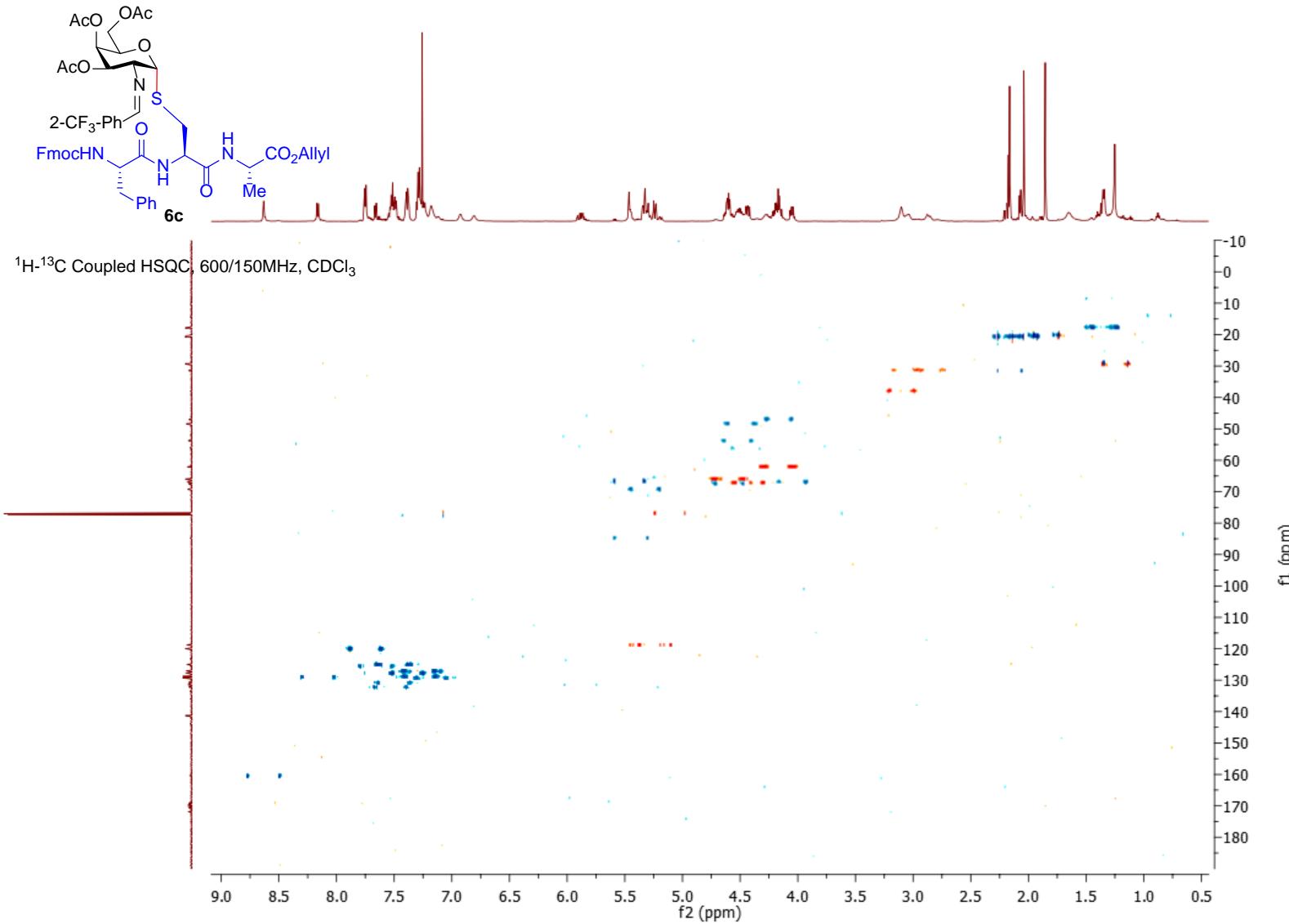


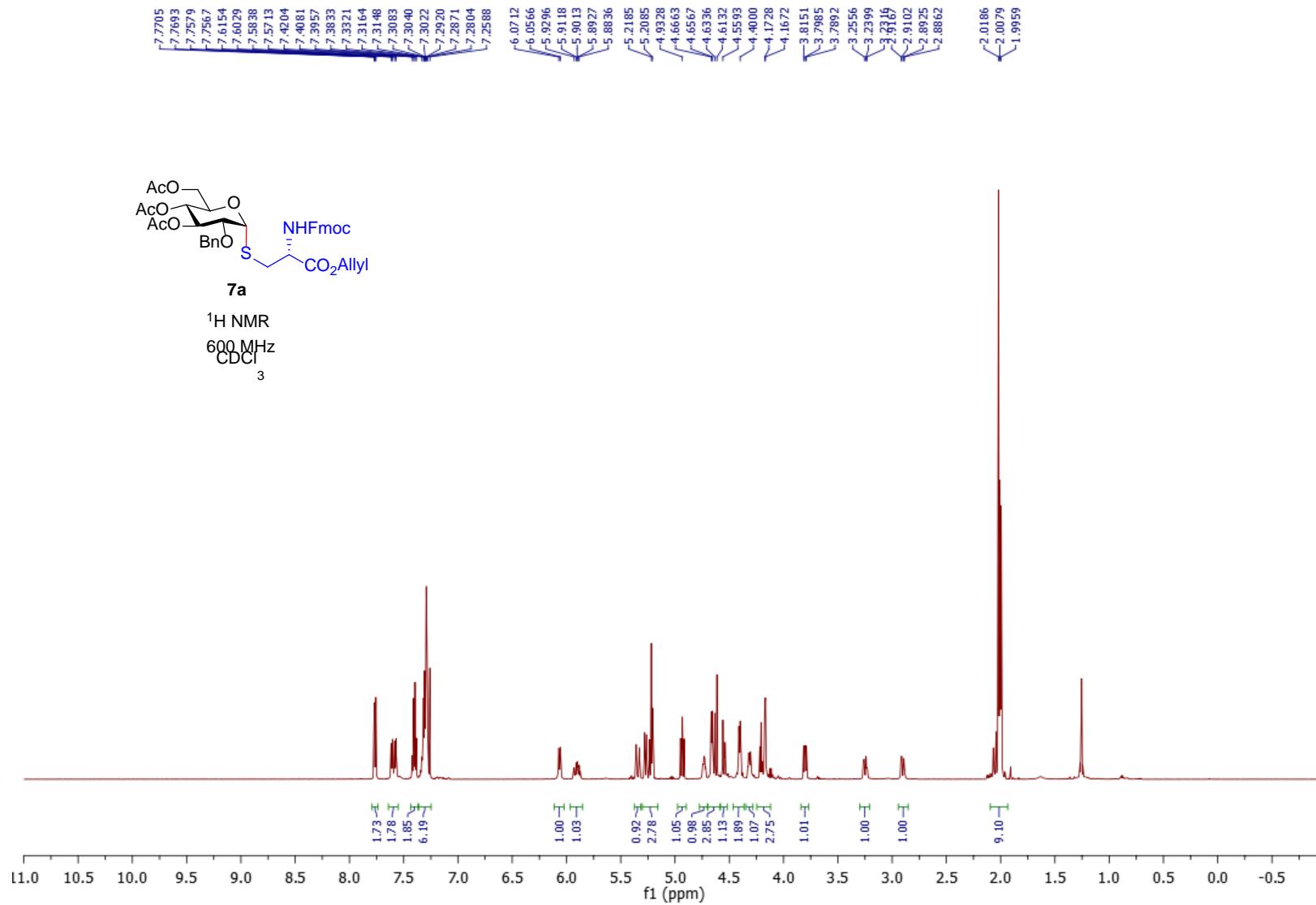


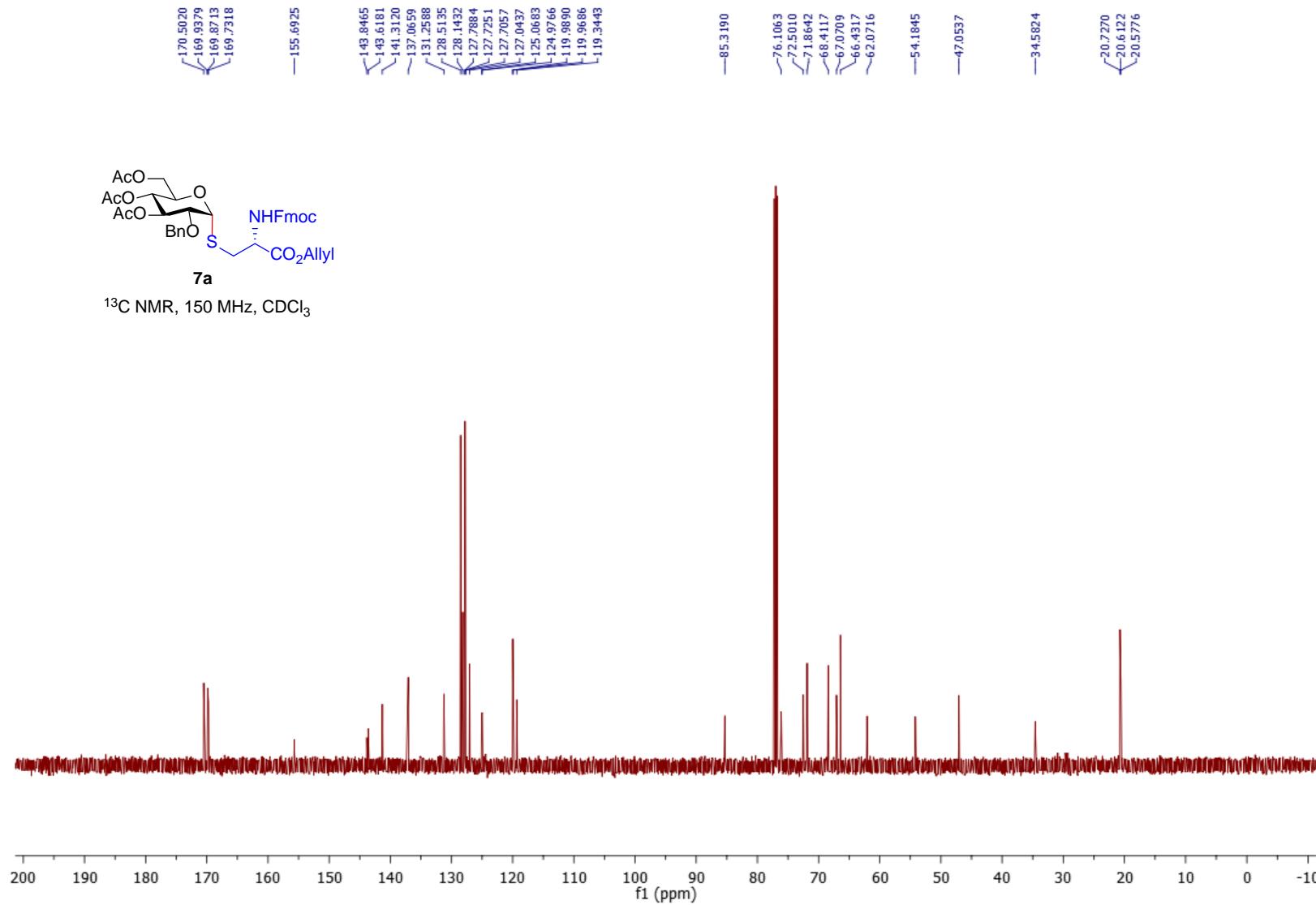


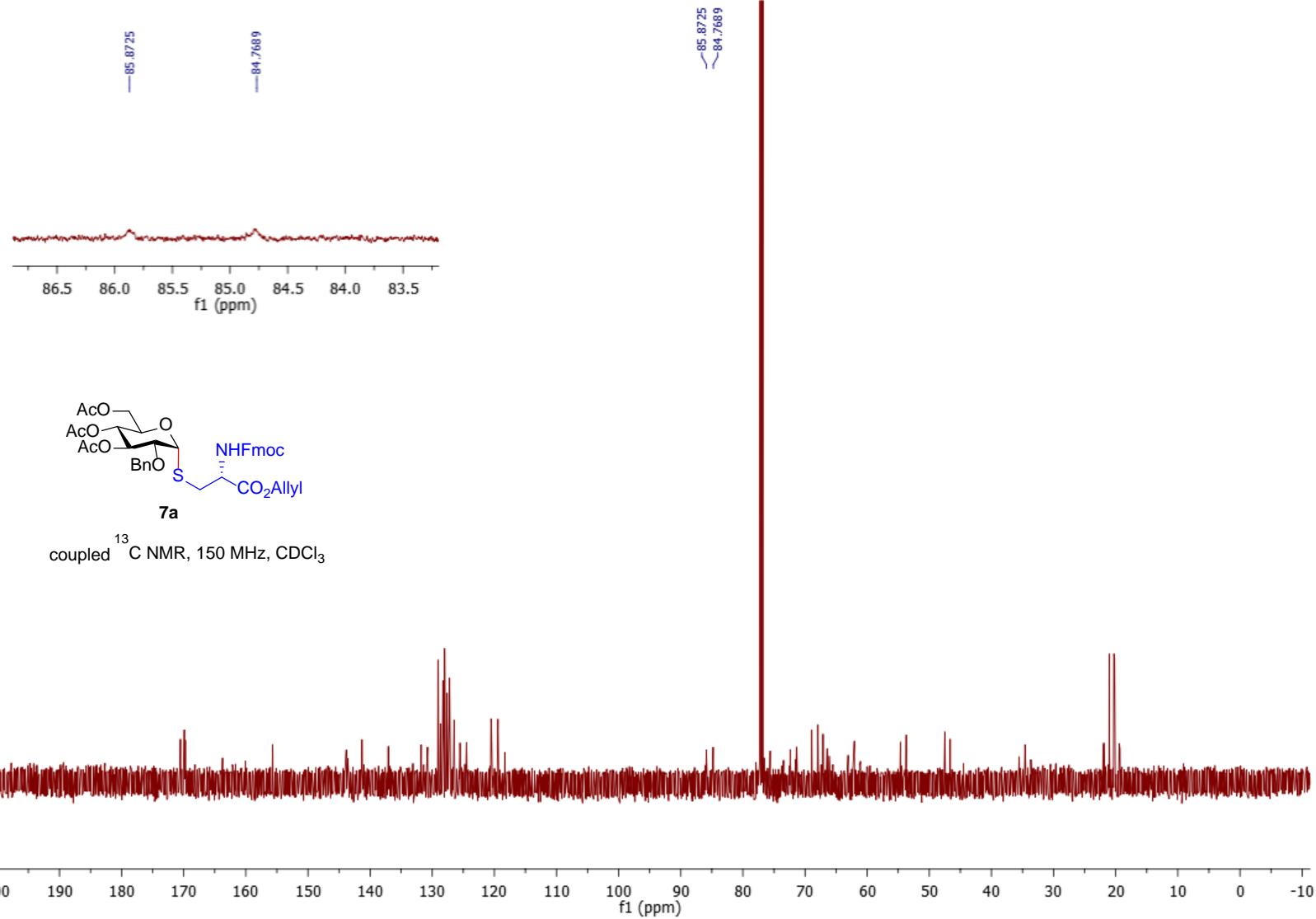


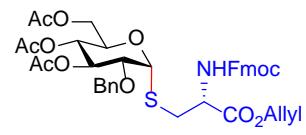




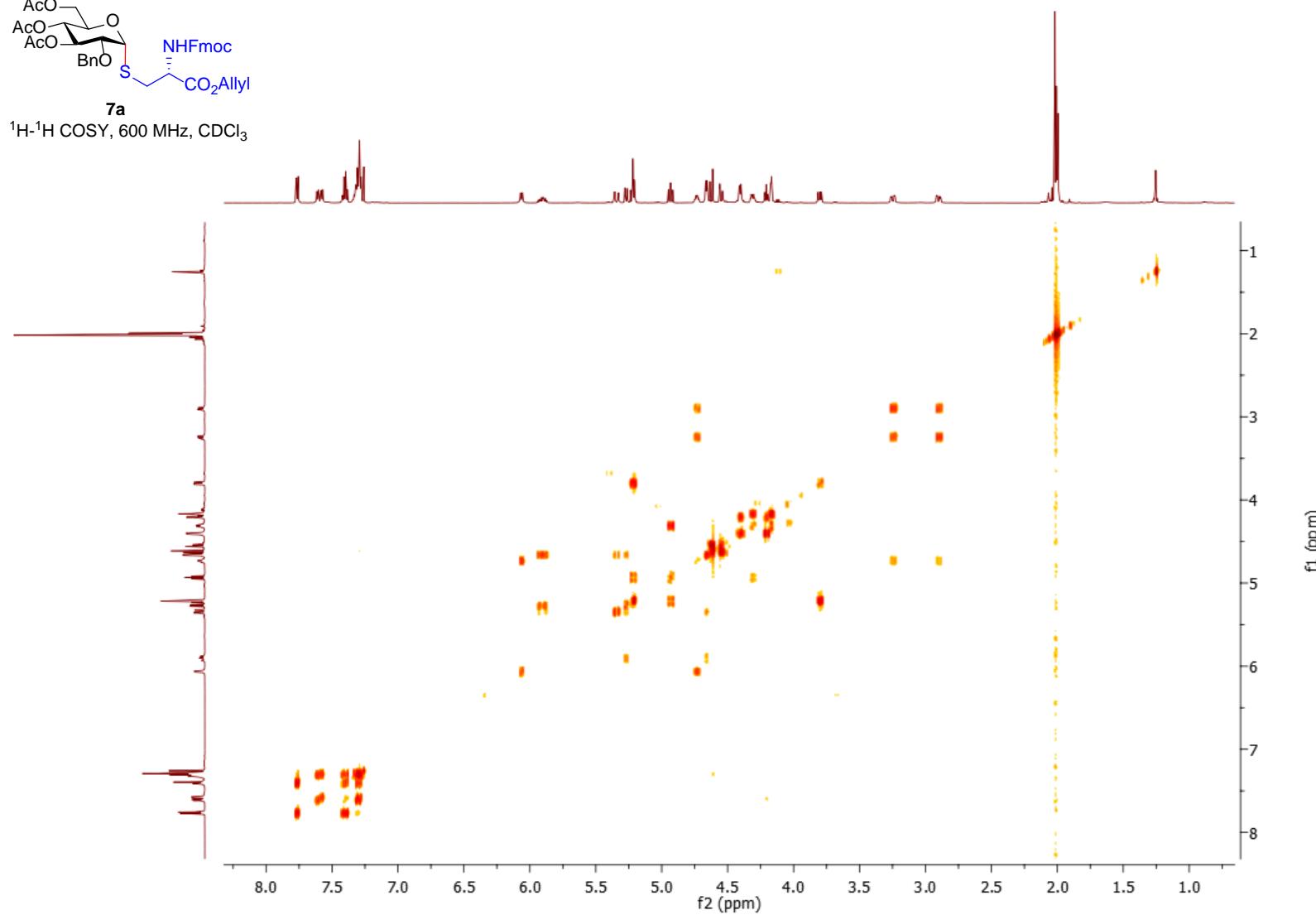


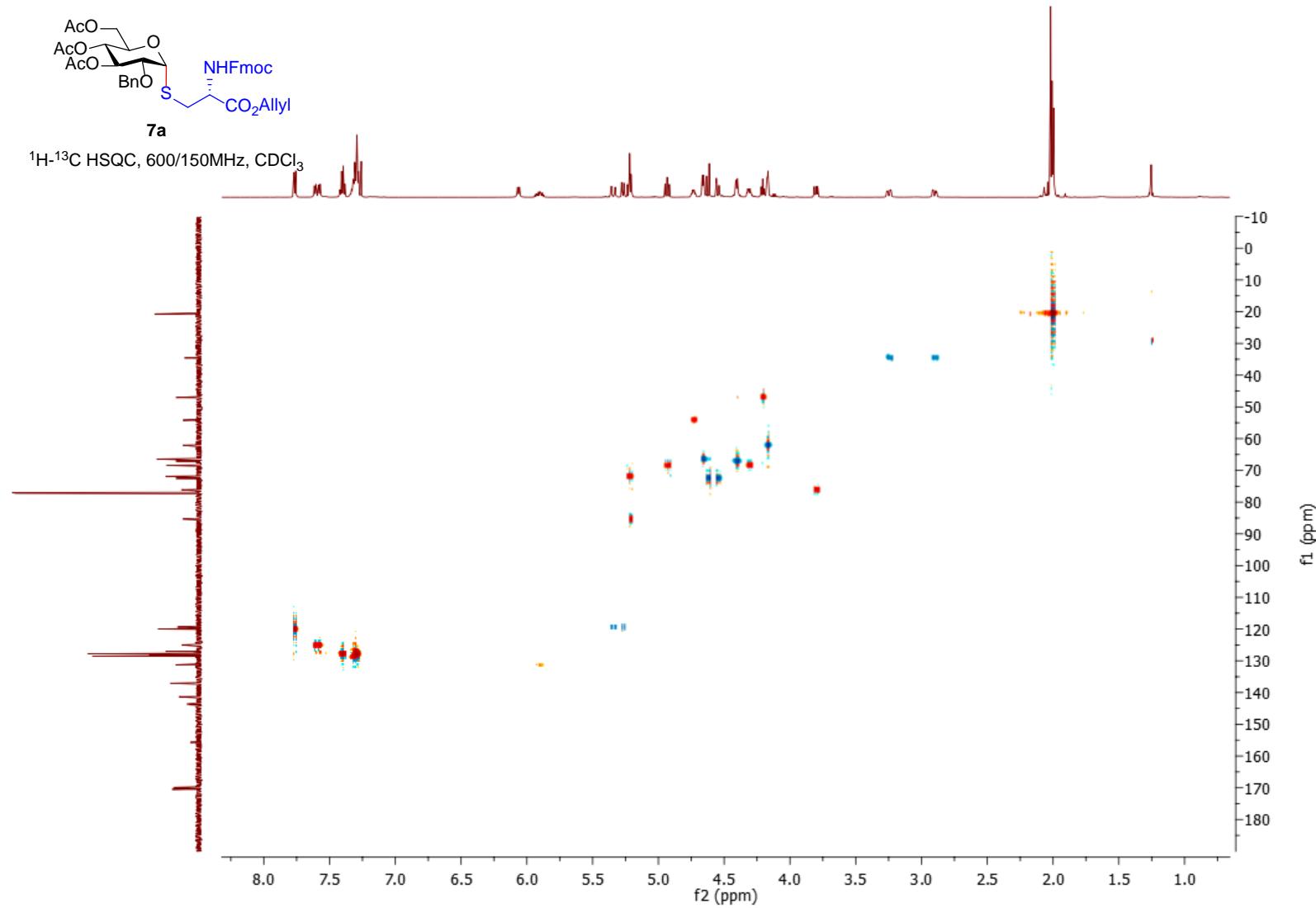


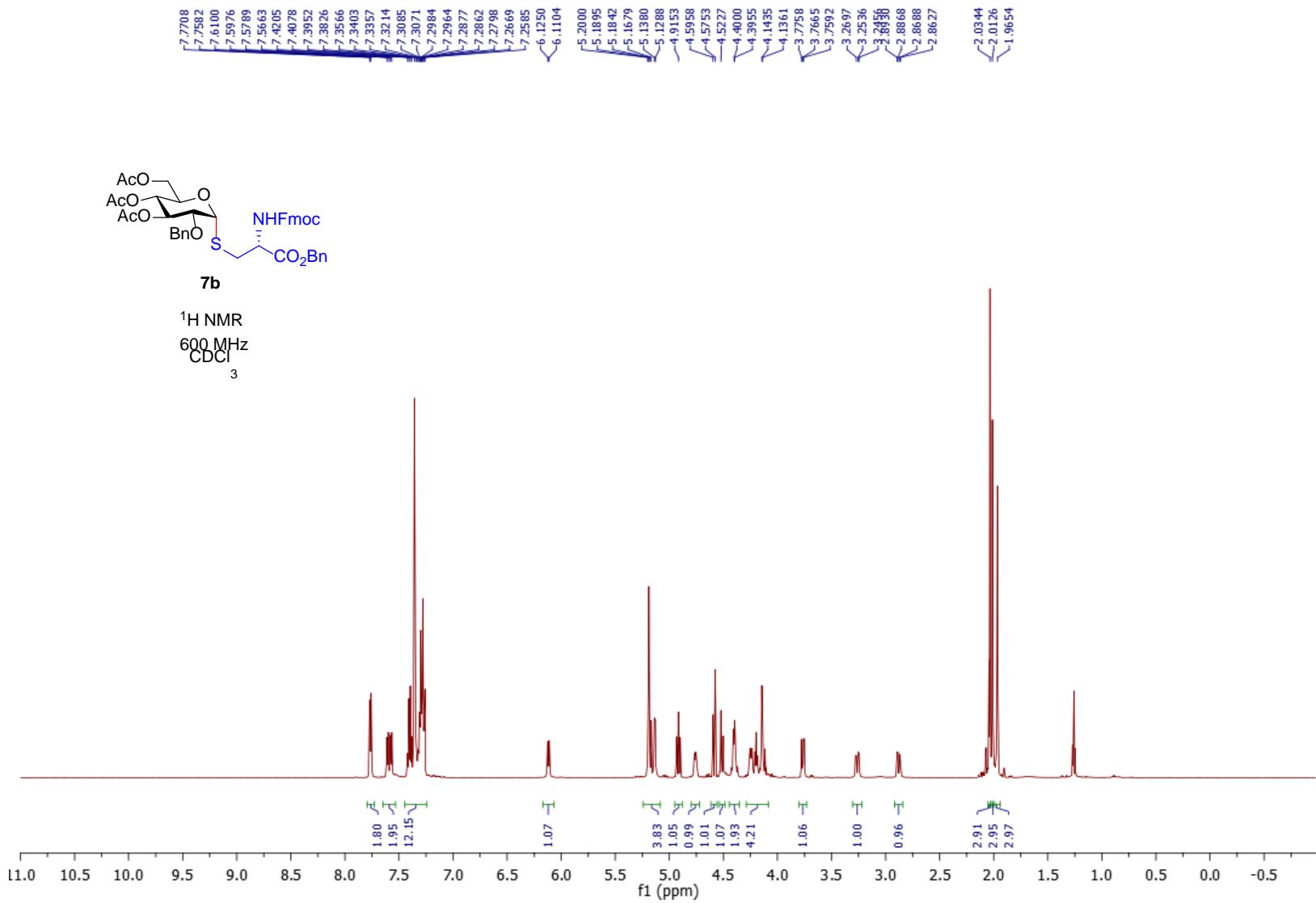


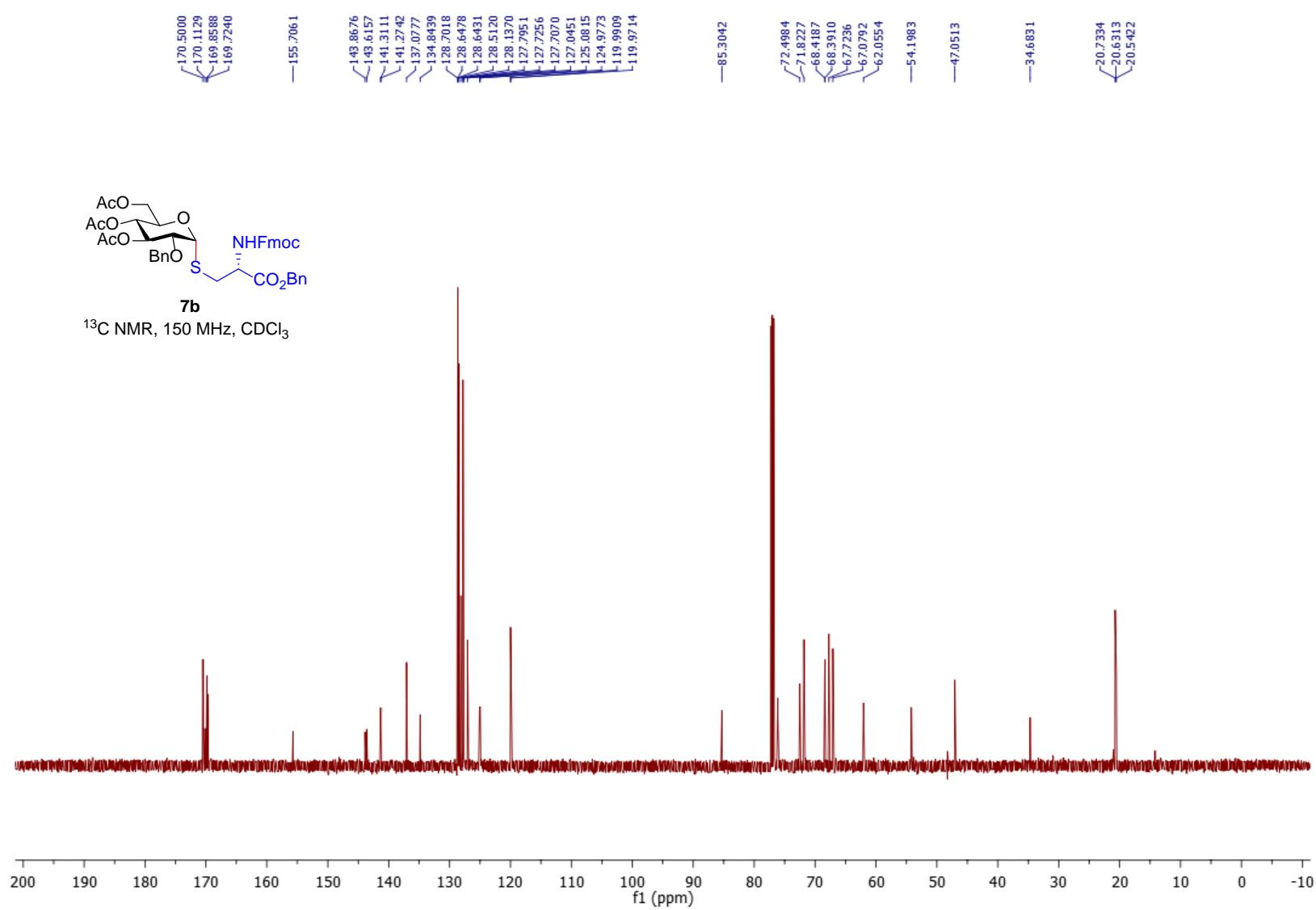


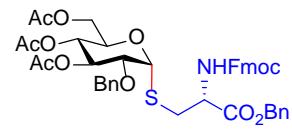
7a
¹H-¹H COSY, 600 MHz, CDCl₃



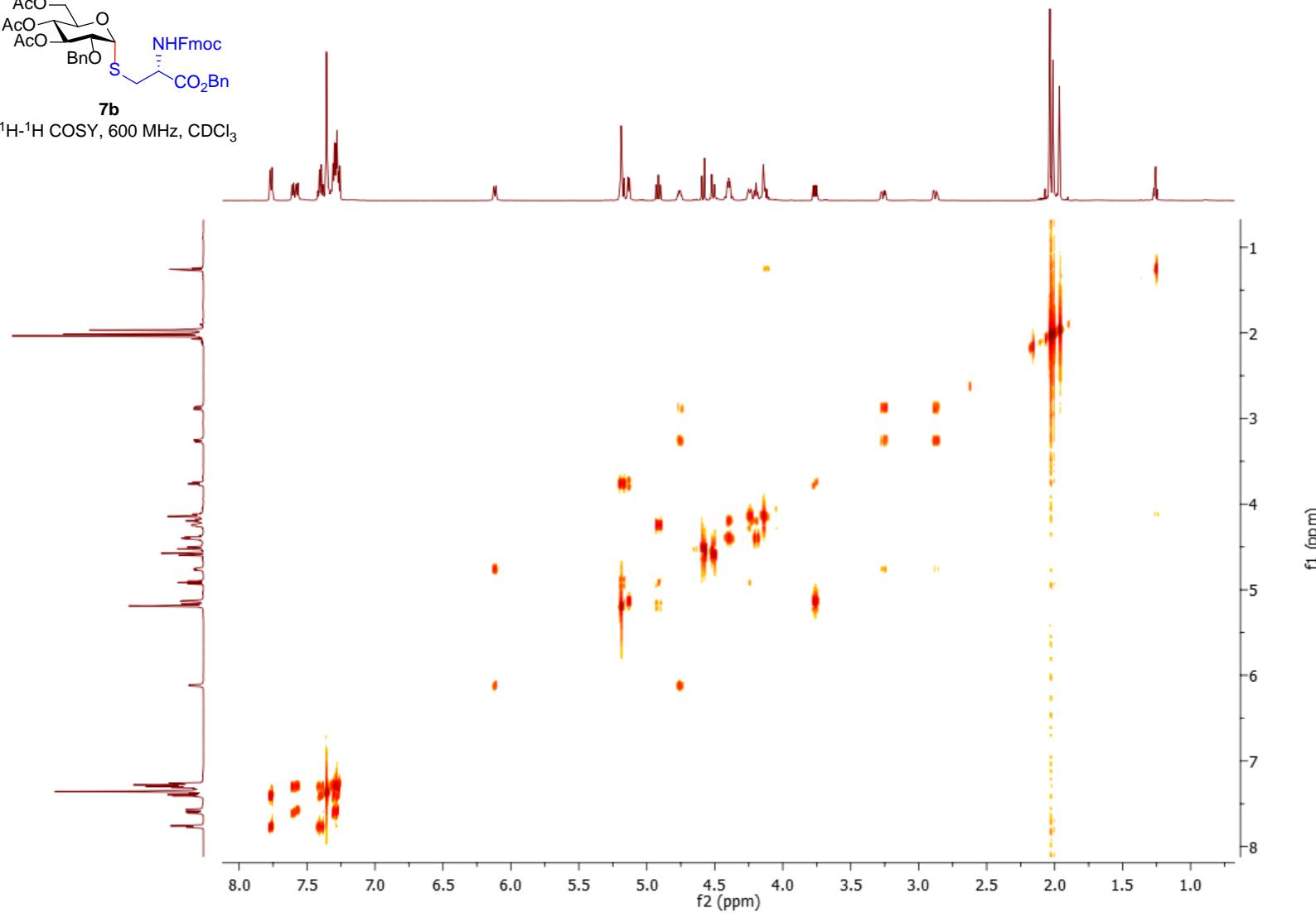


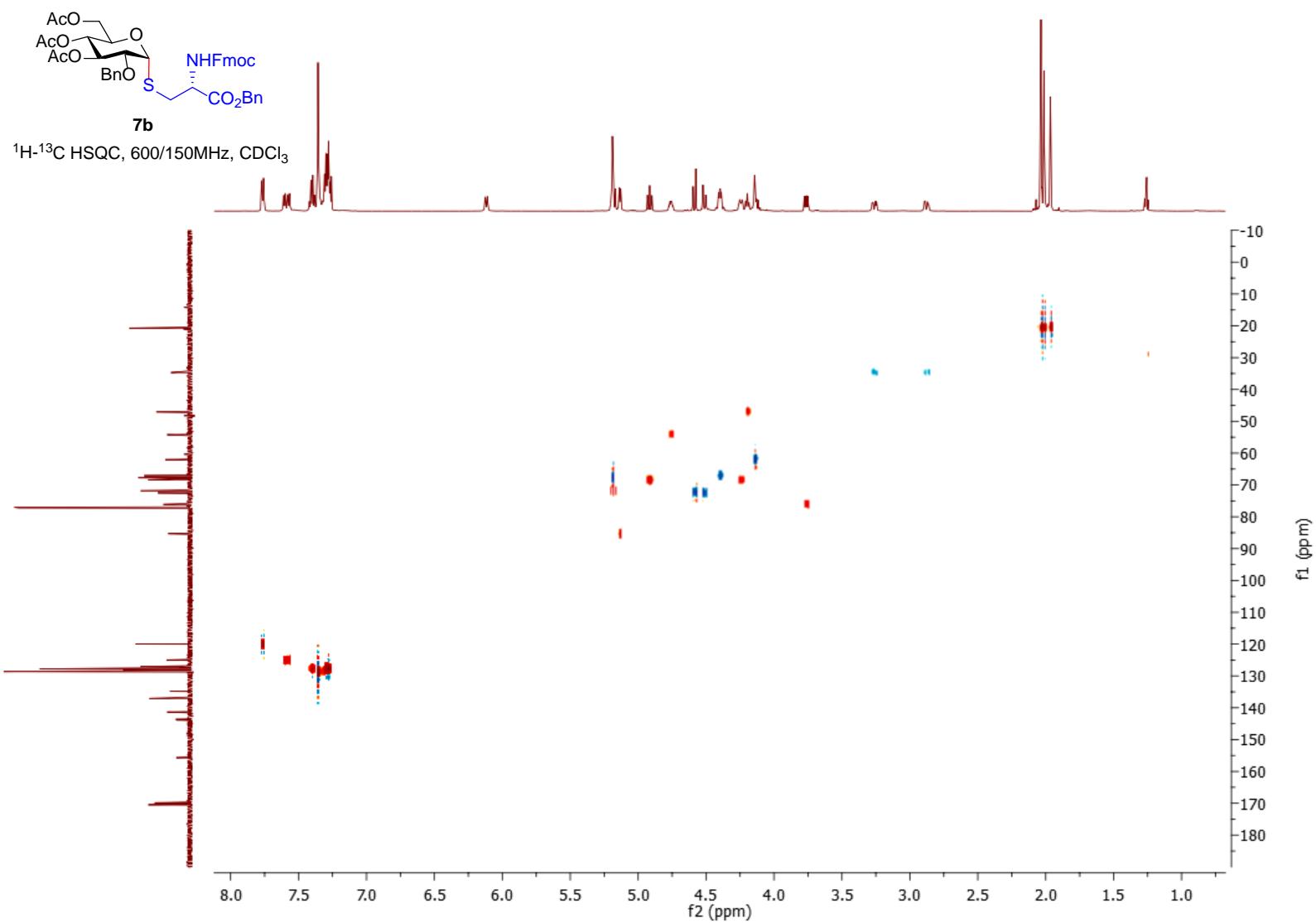


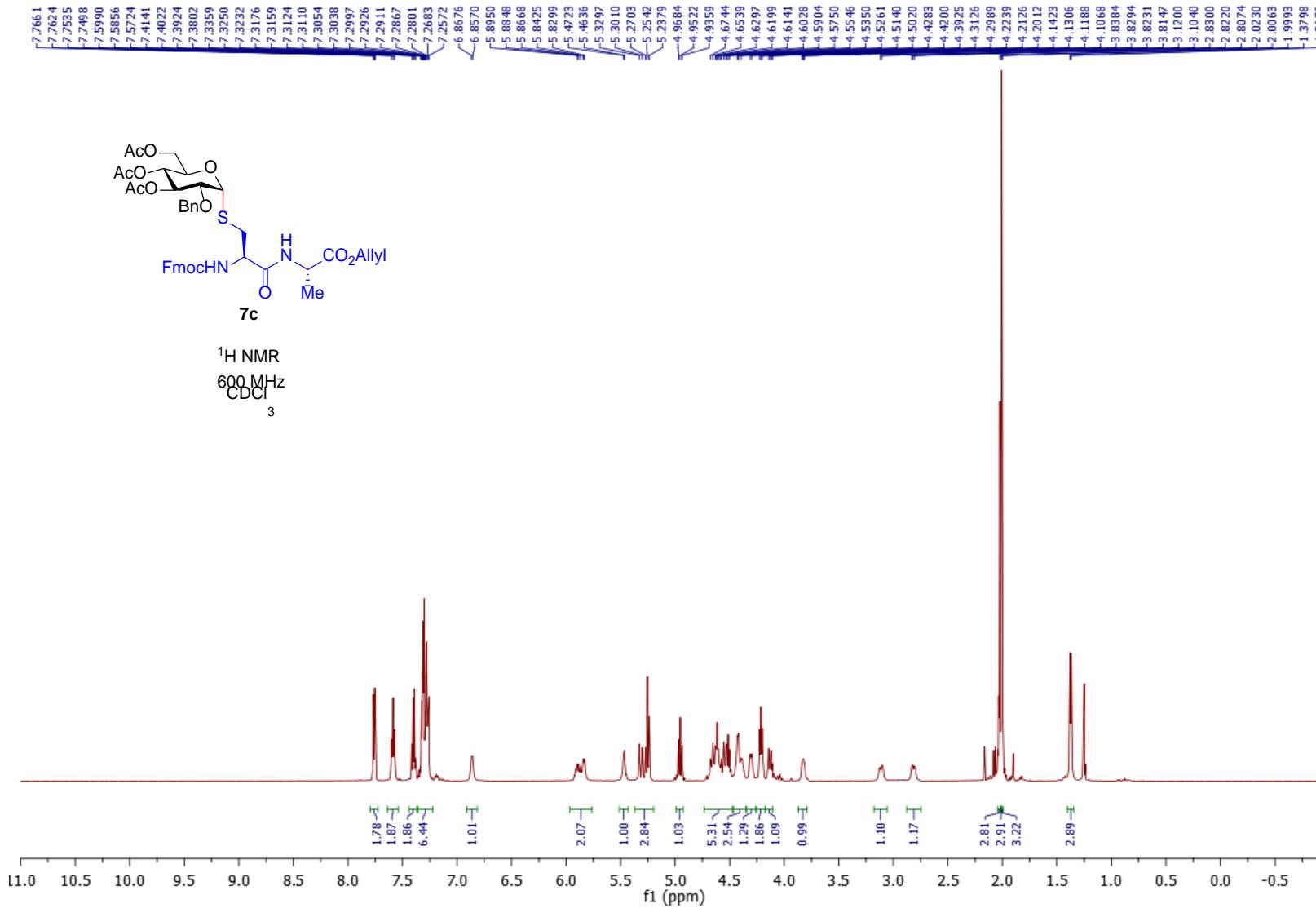


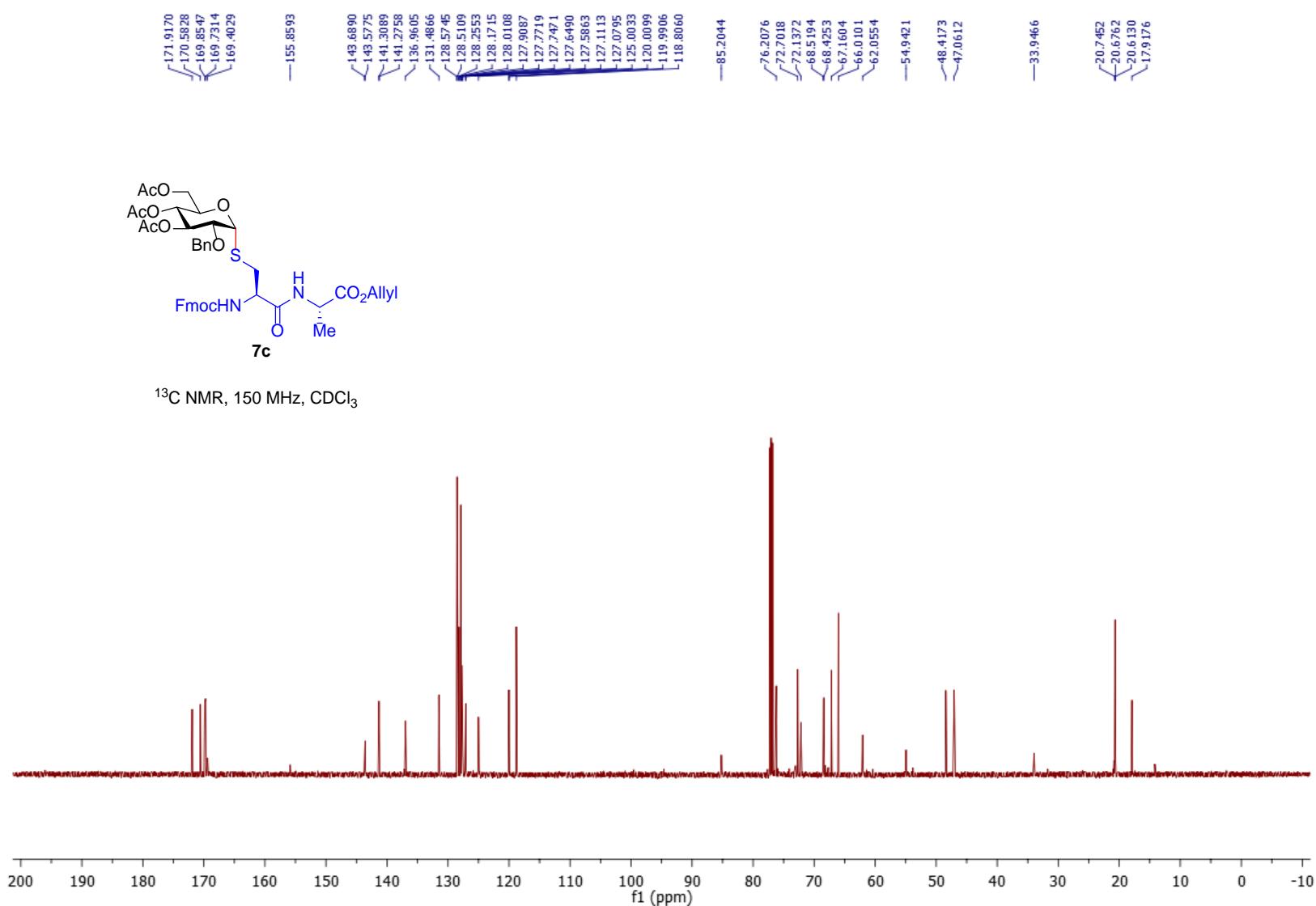


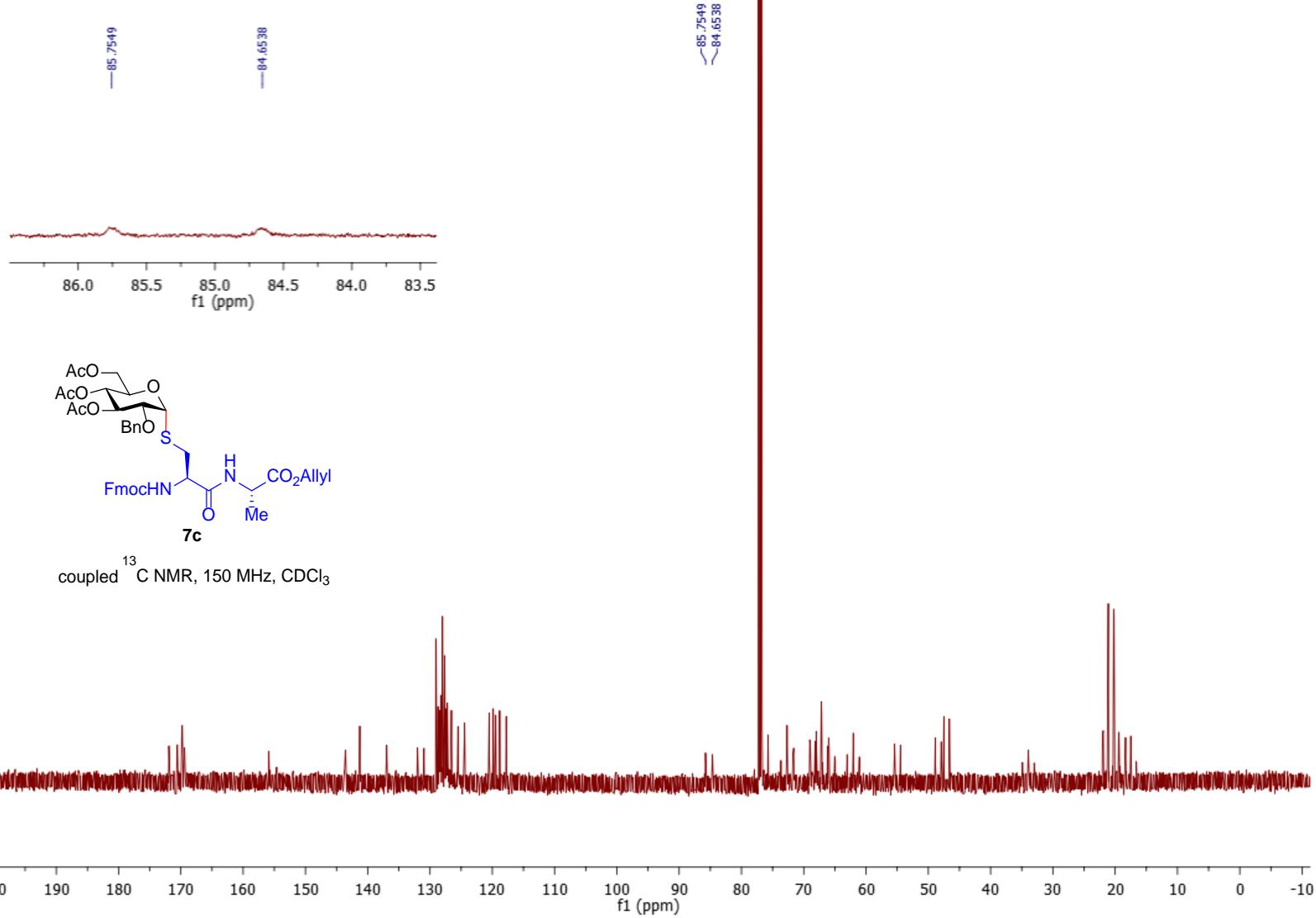
7b
¹H-¹H COSY, 600 MHz, CDCl₃

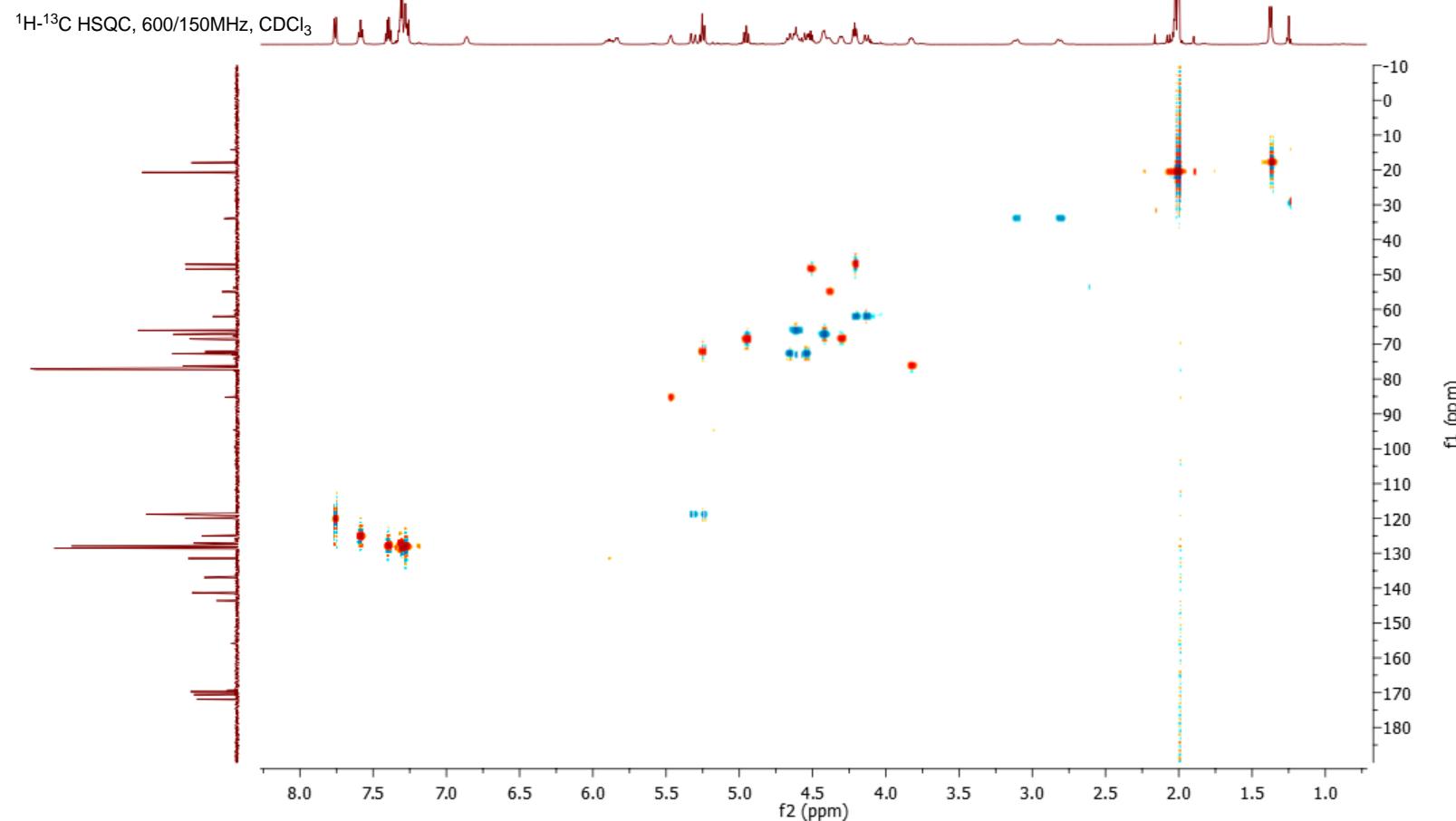
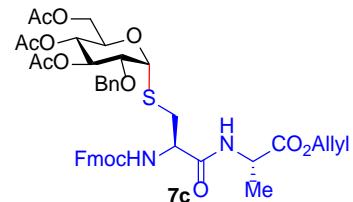




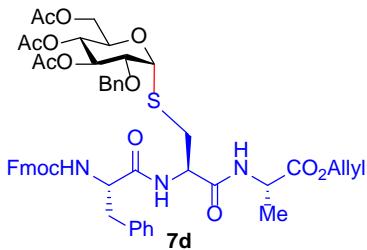






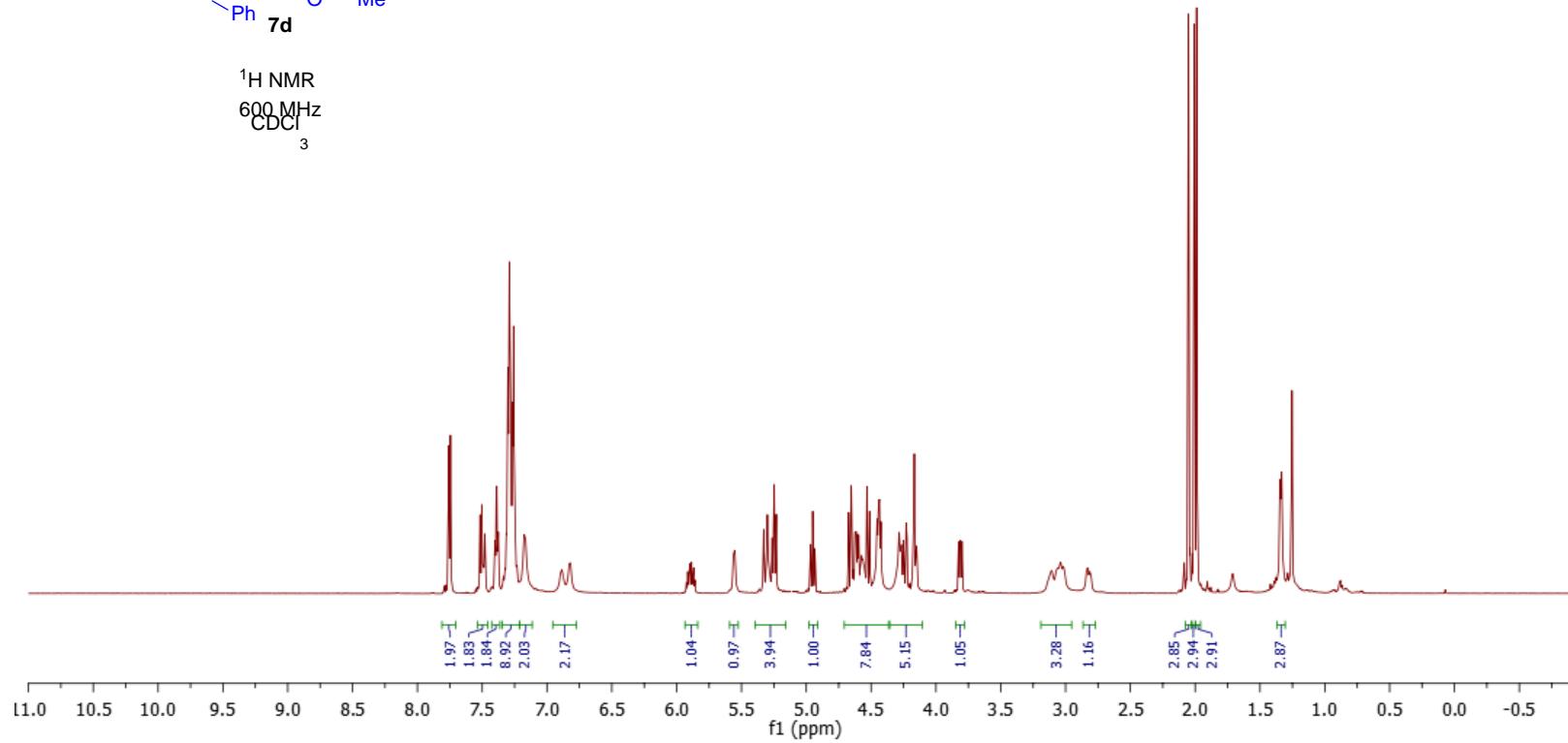


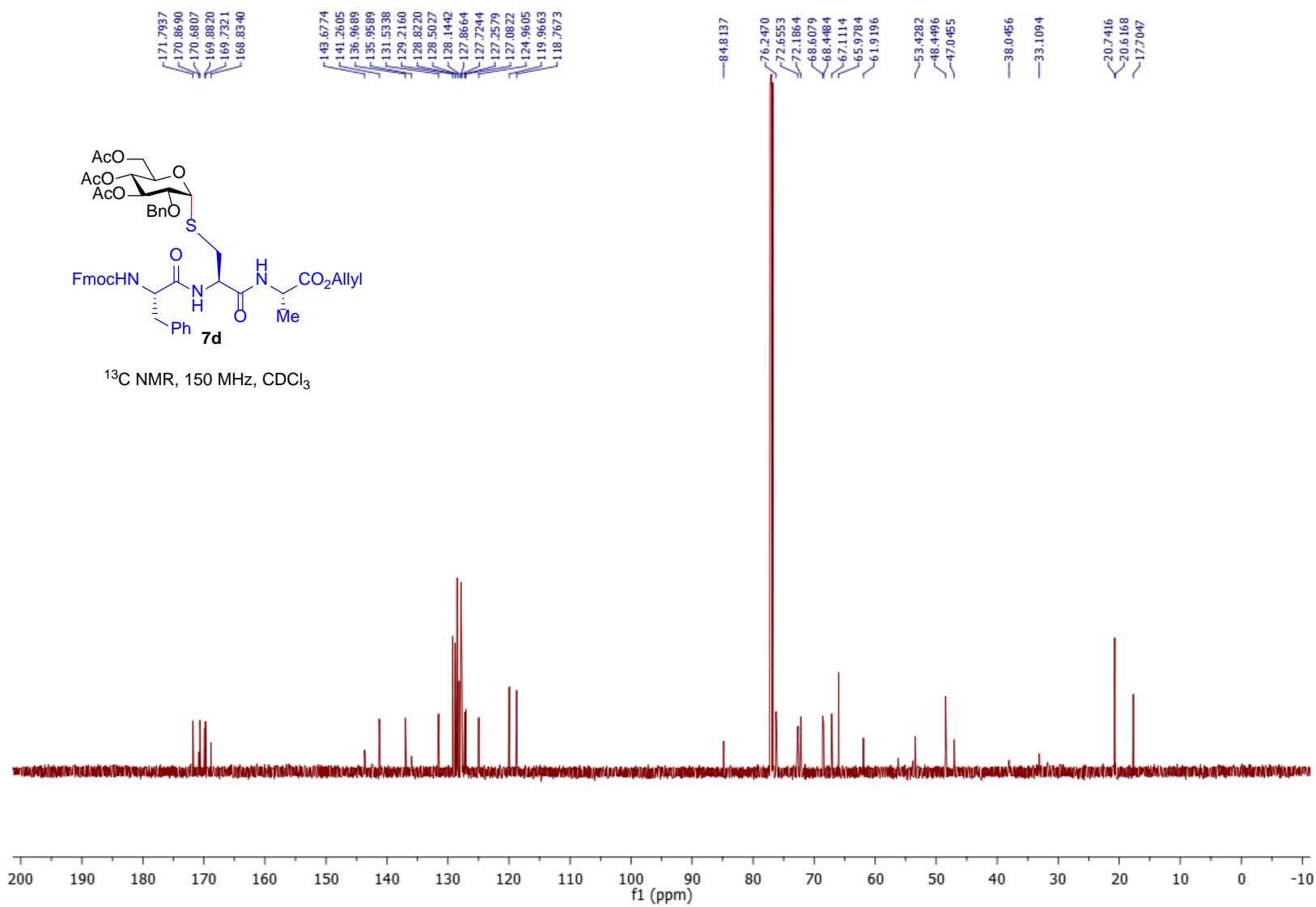
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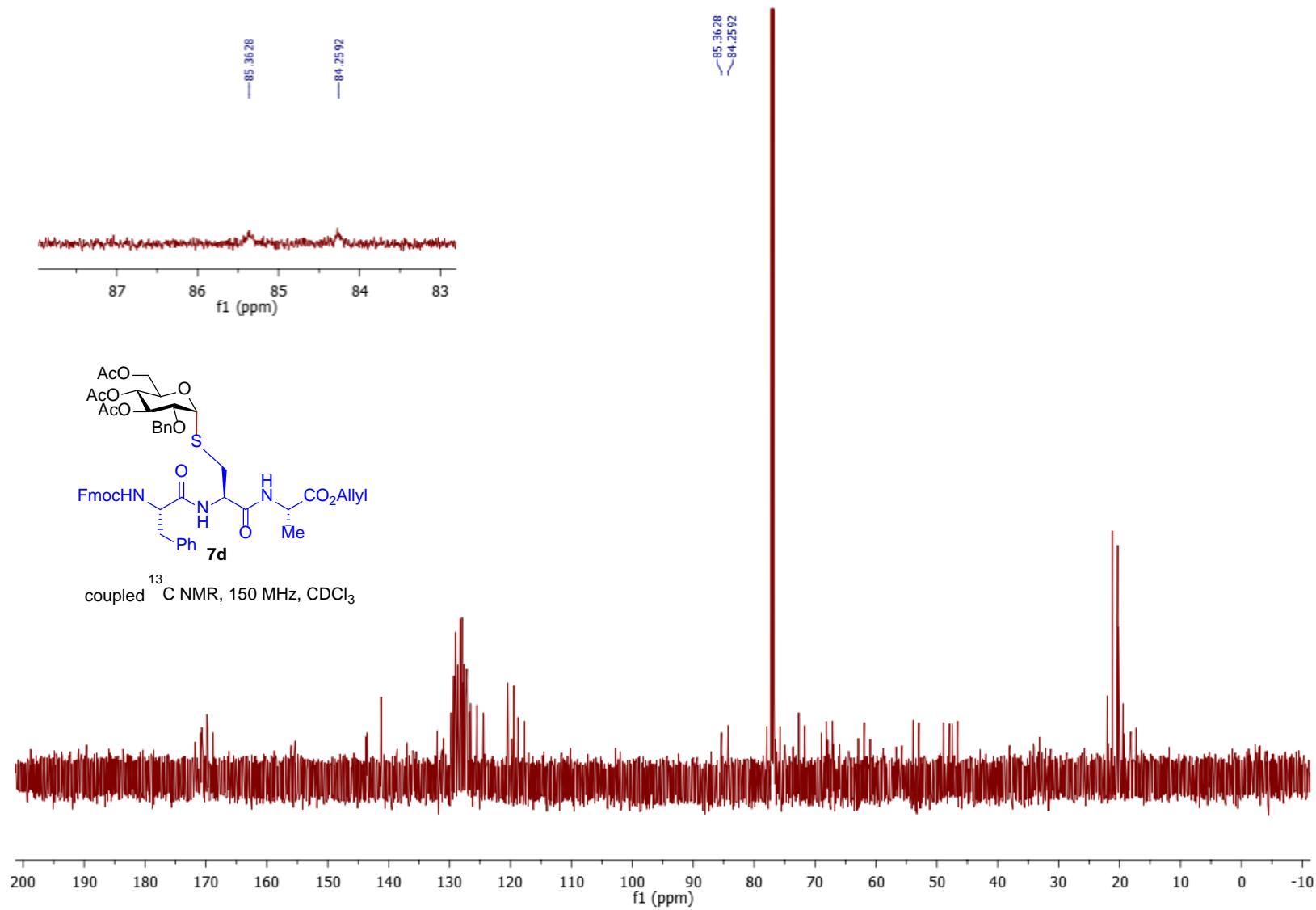


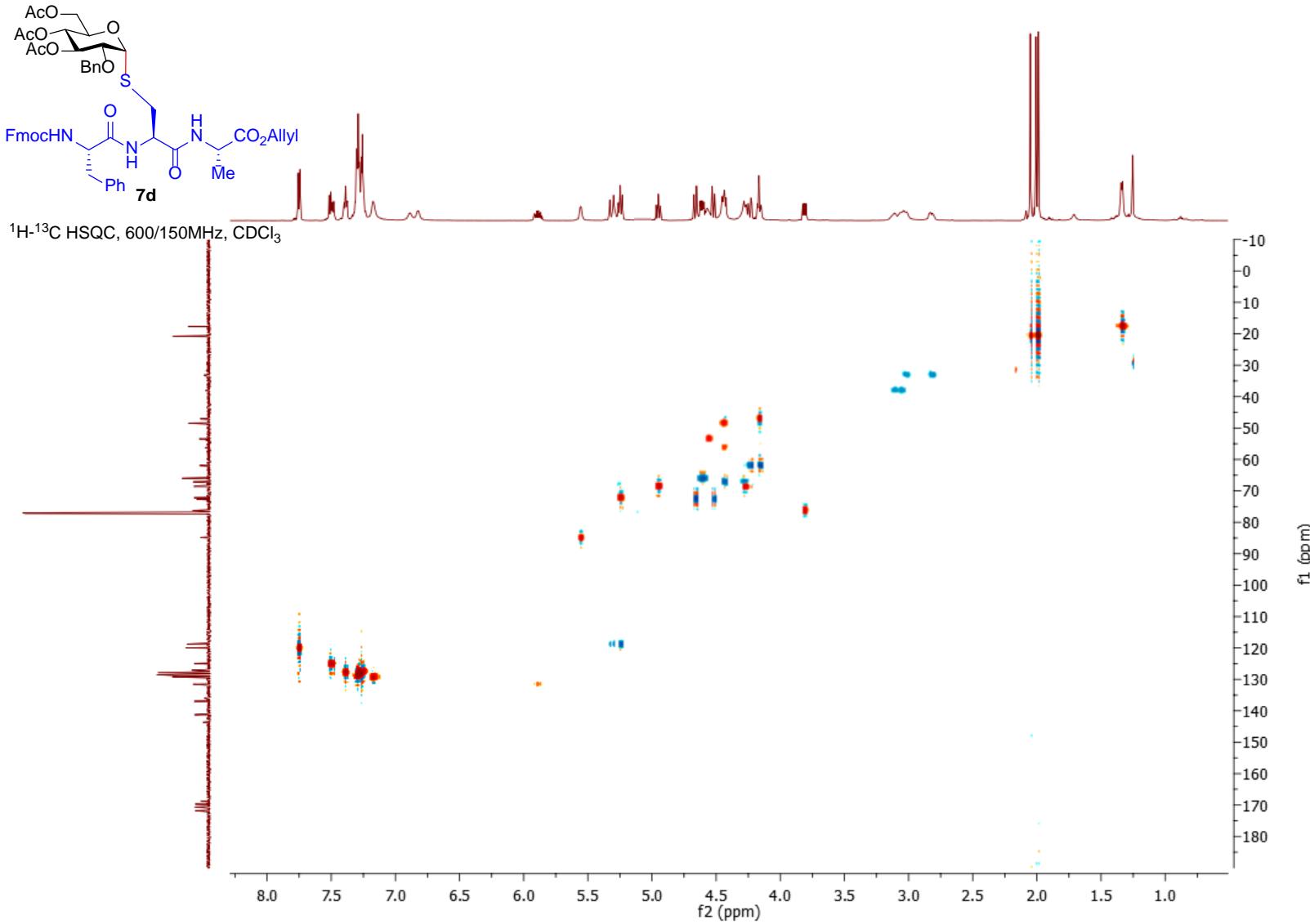
¹H NMR
 600 MHz
 CDCl₃

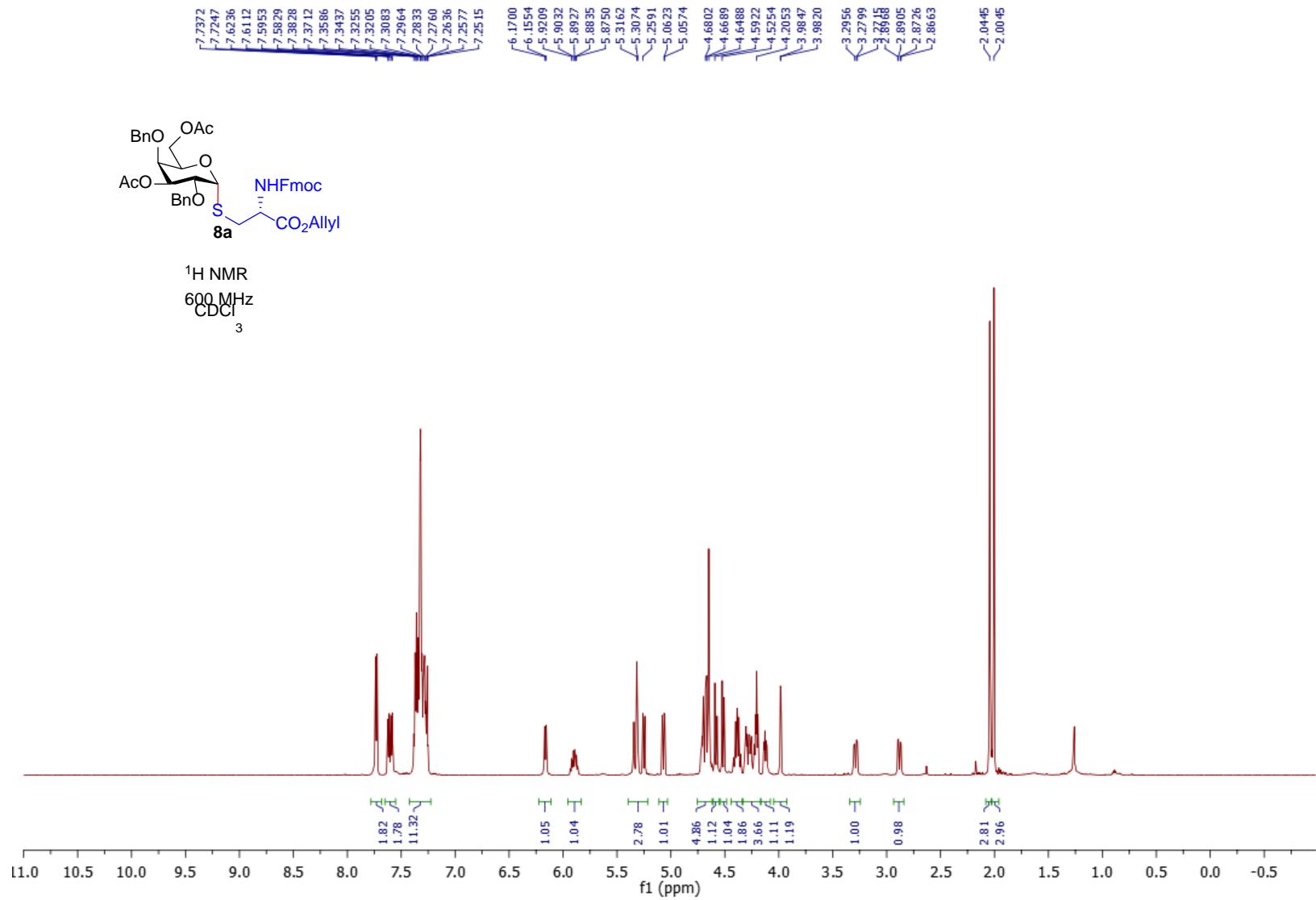
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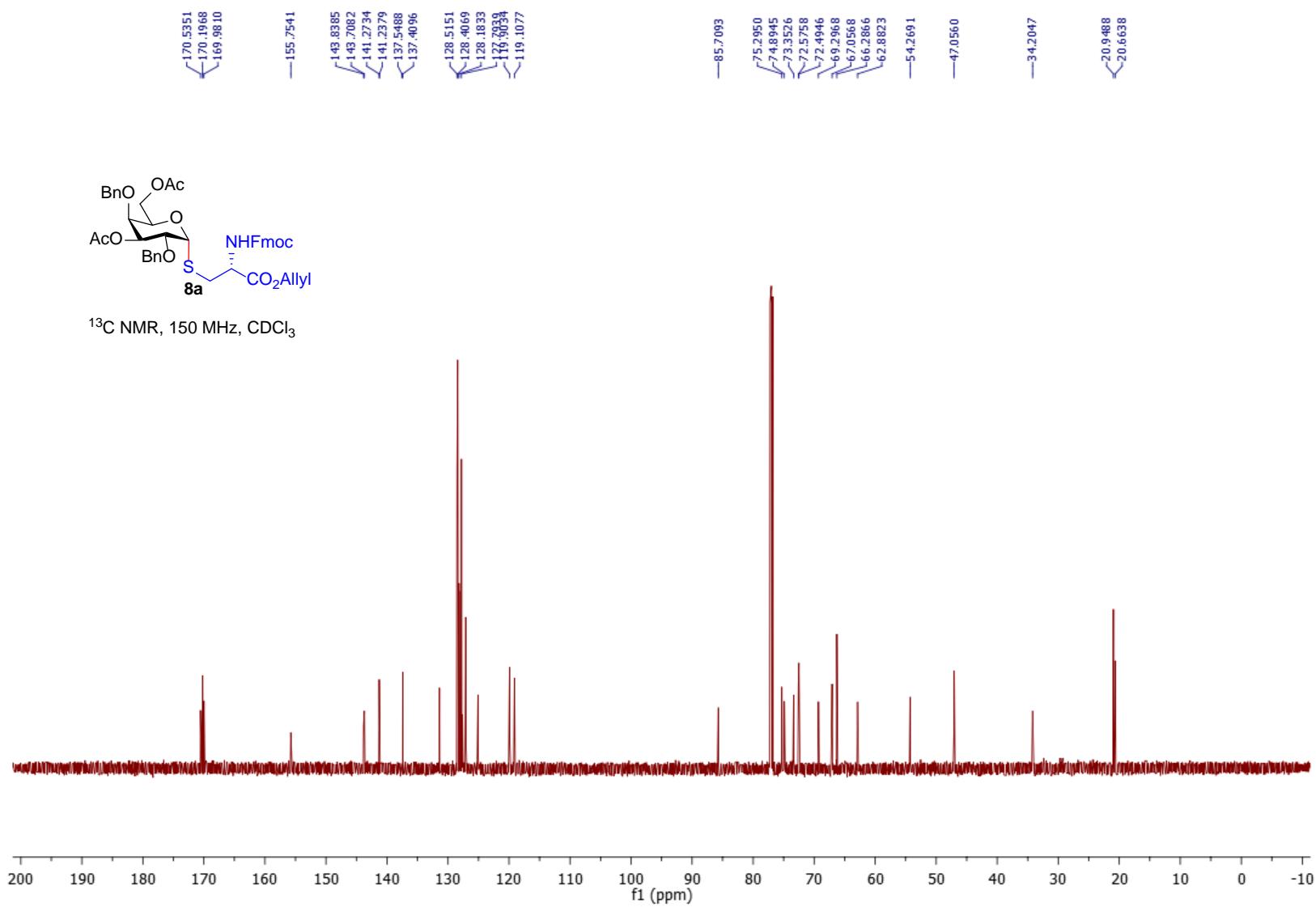


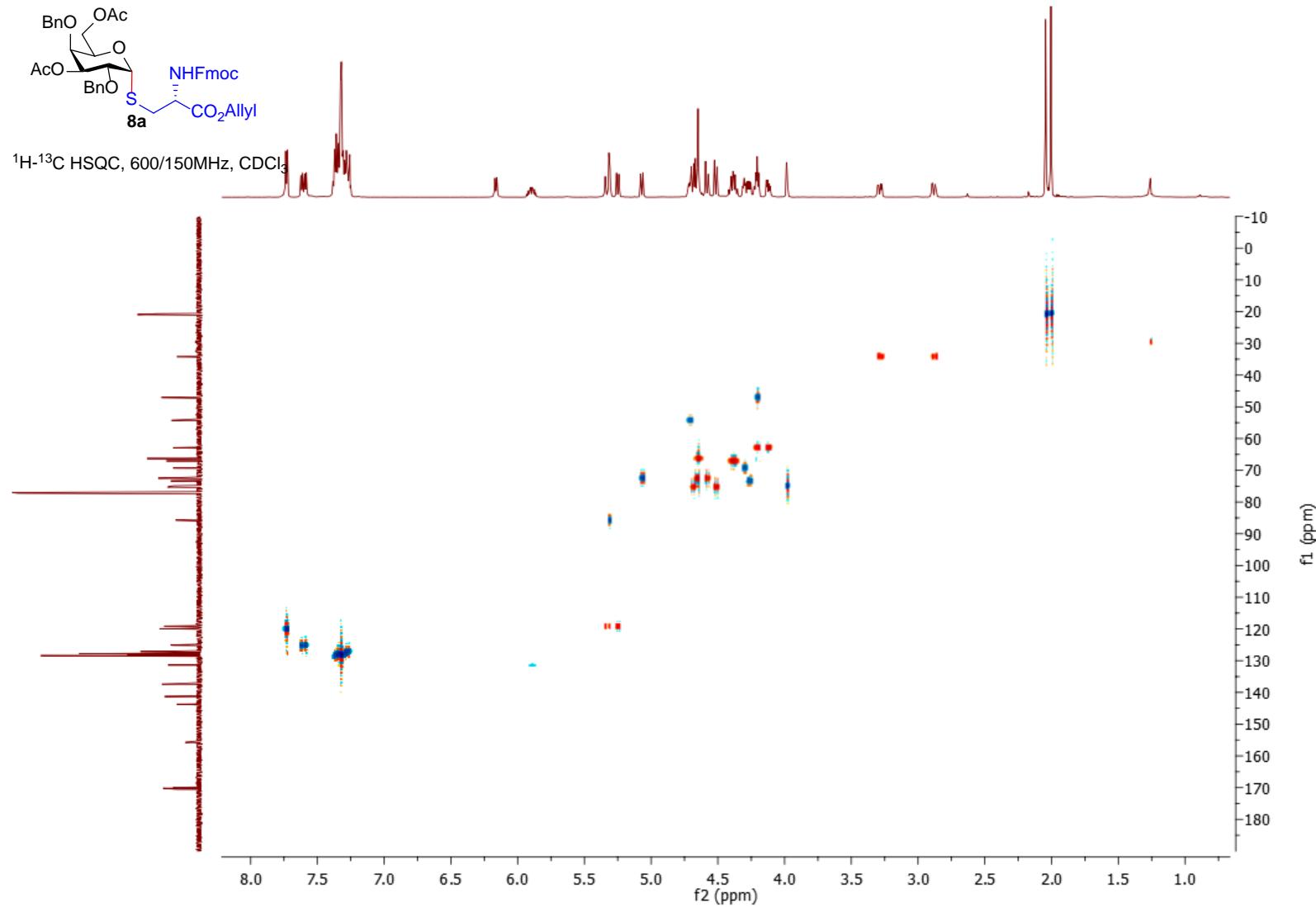


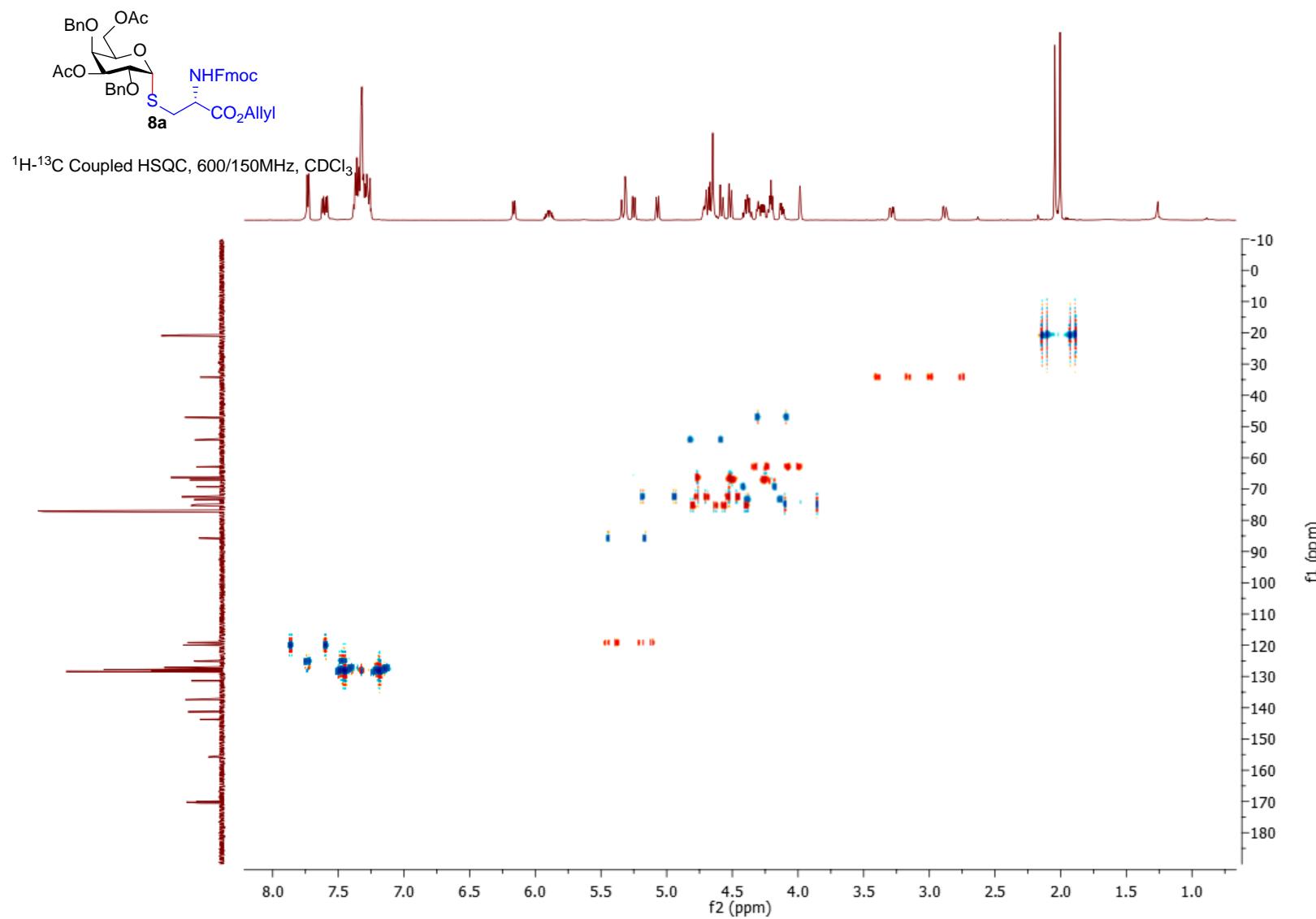


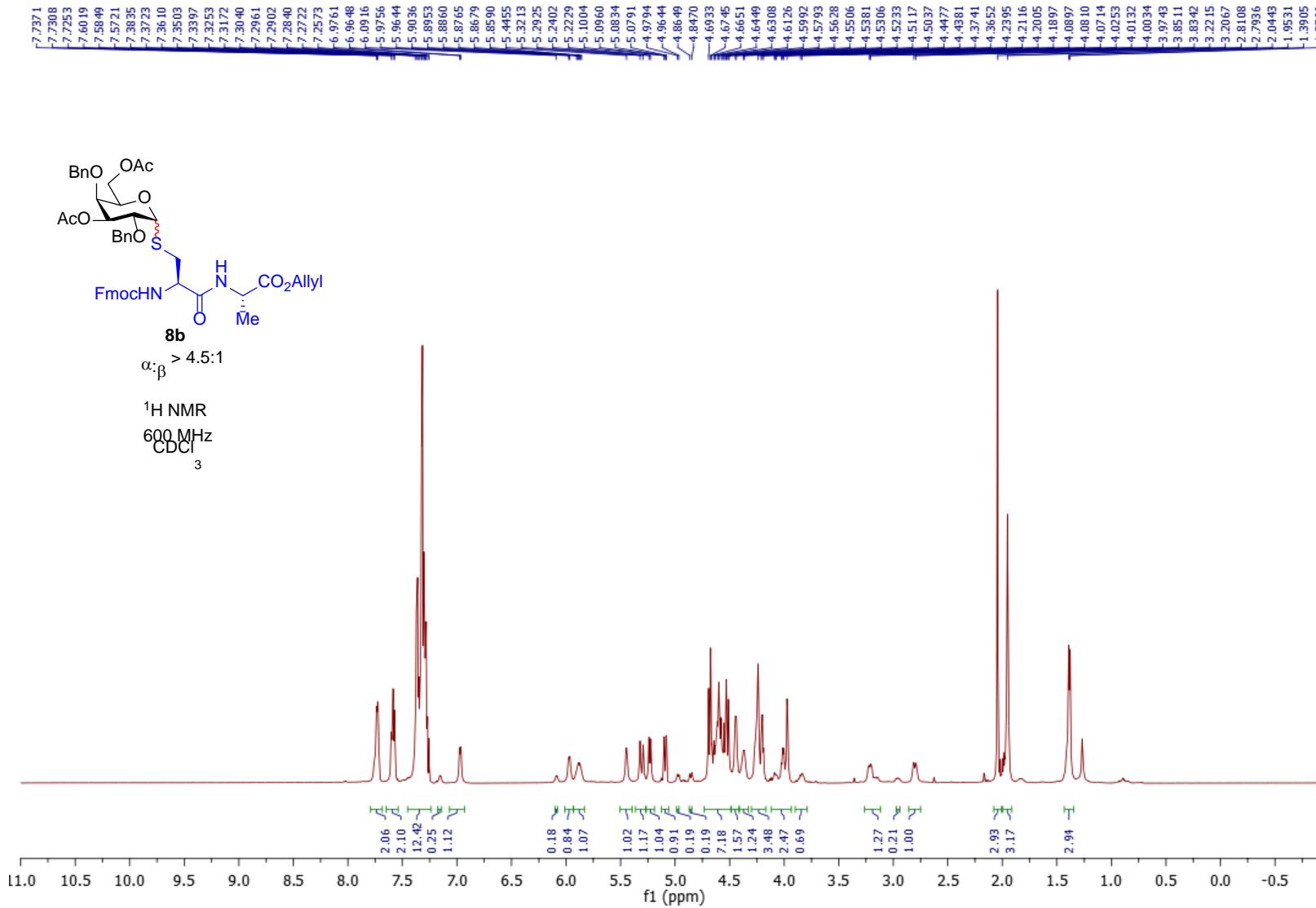


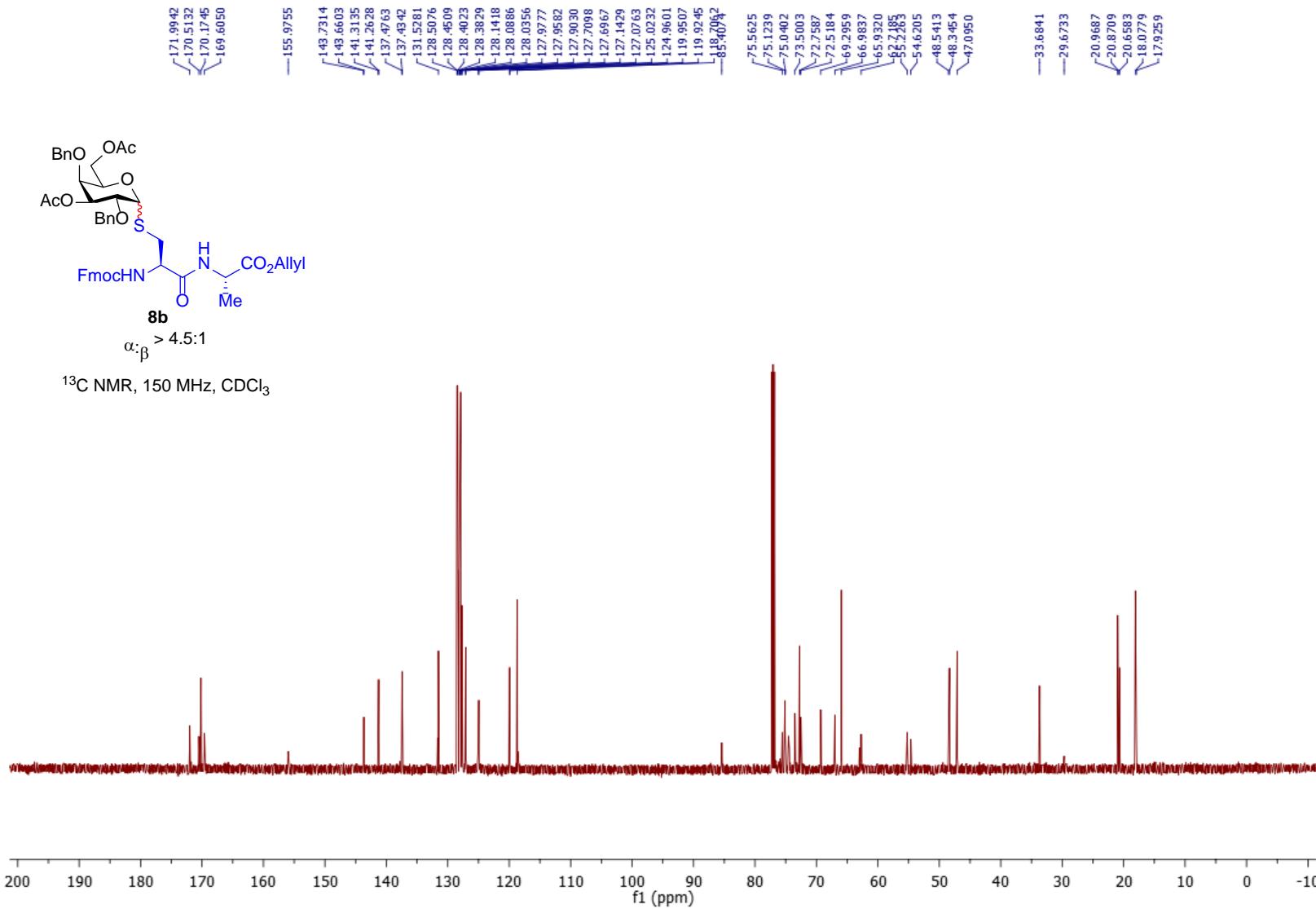


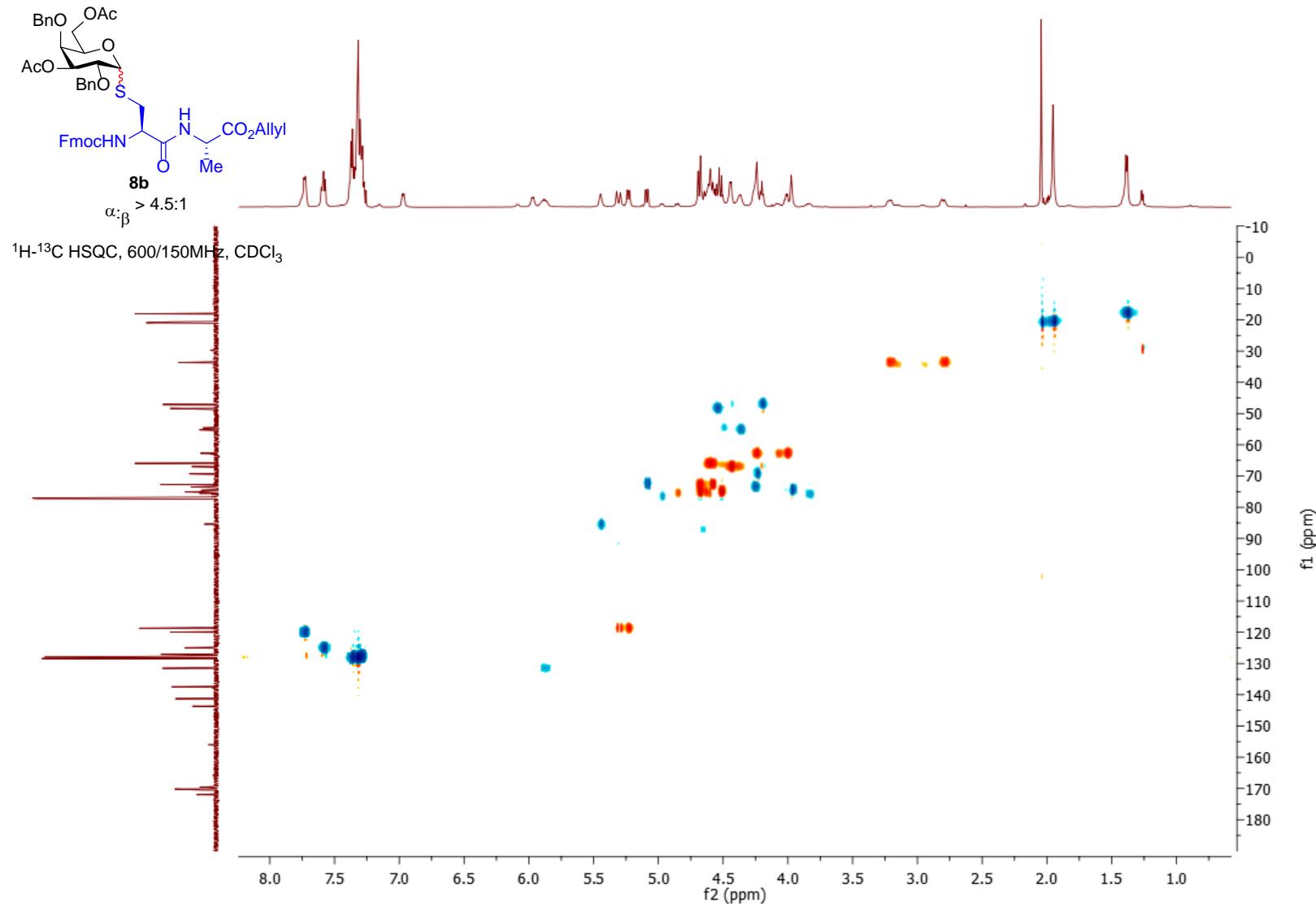


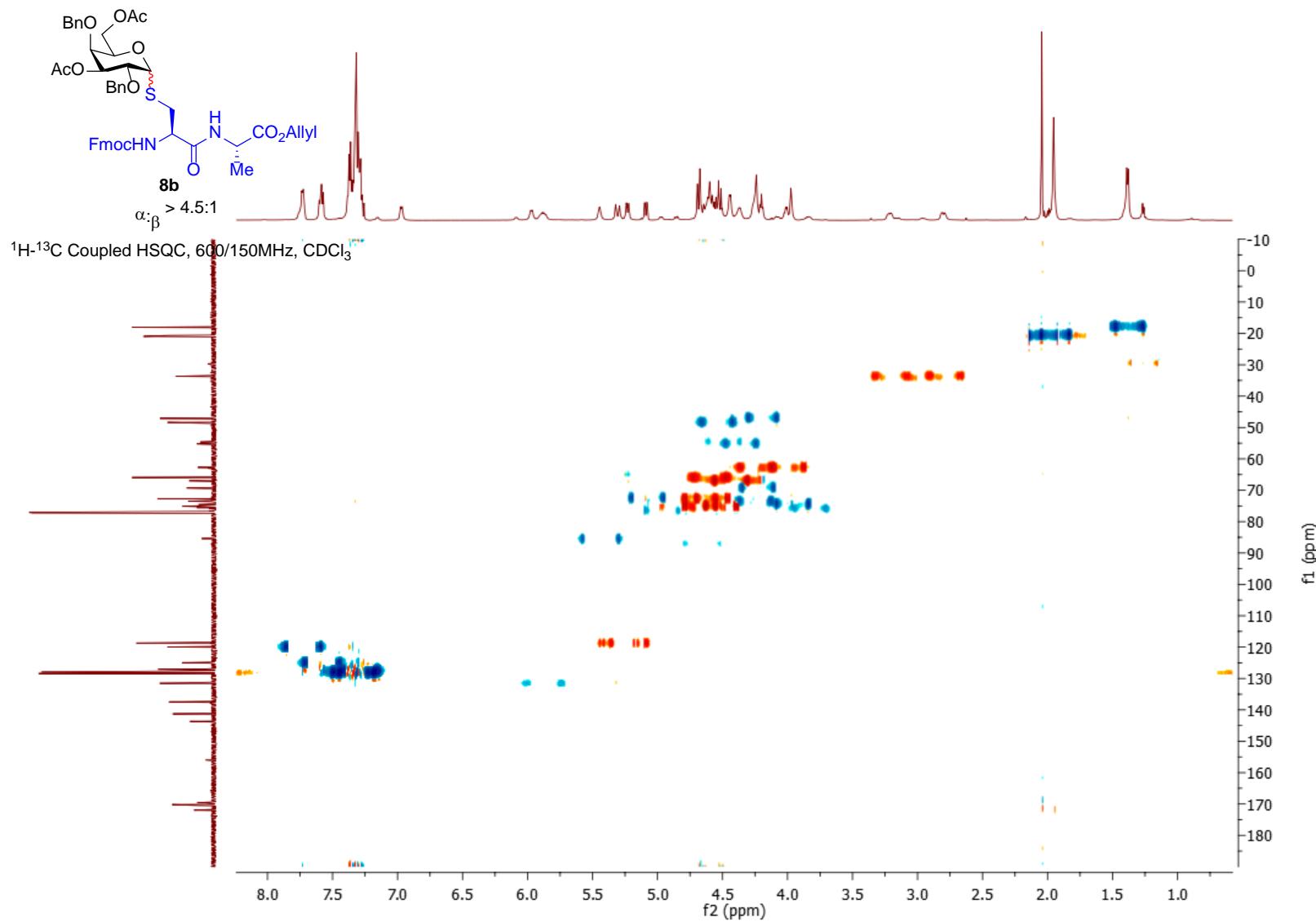


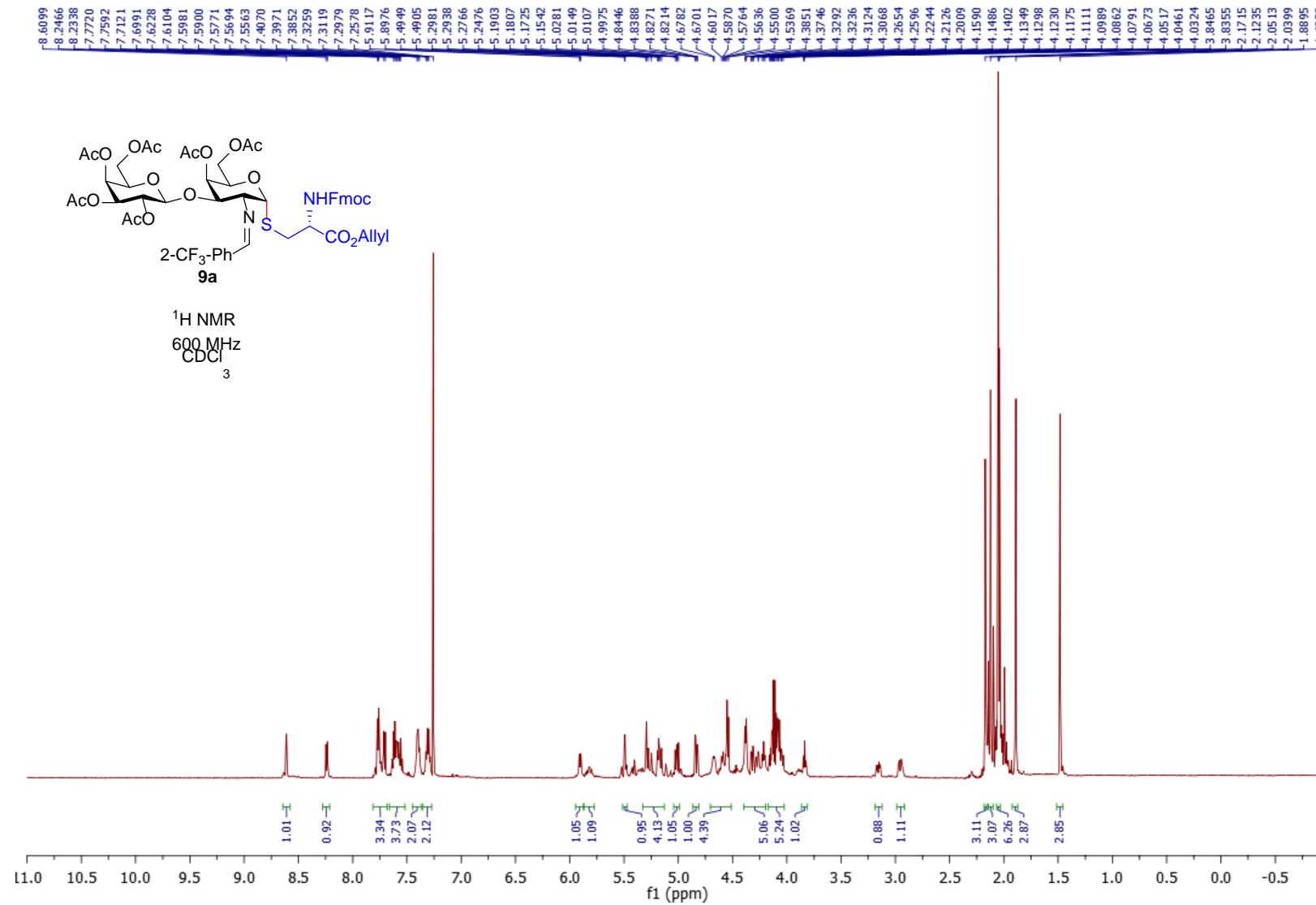


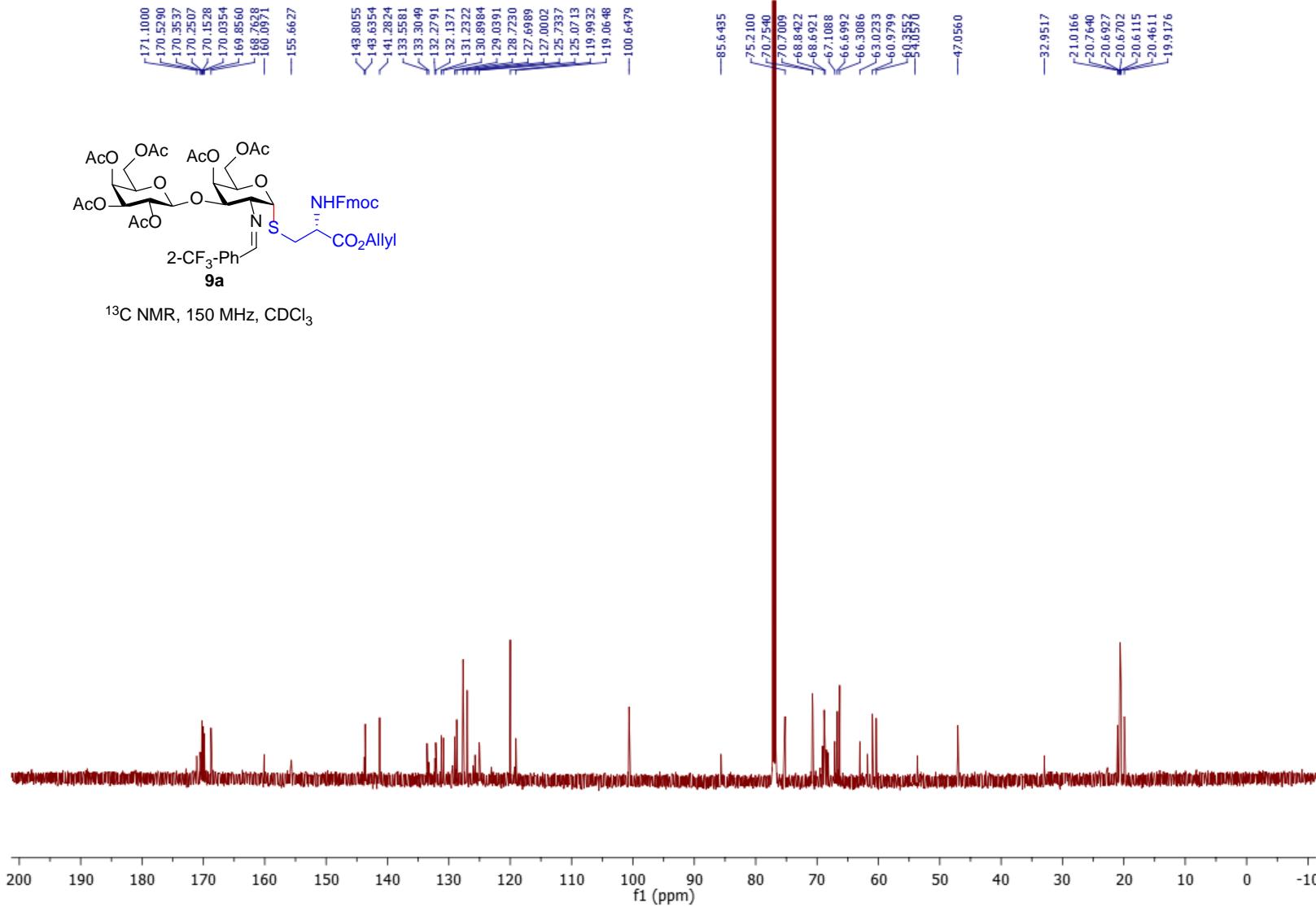


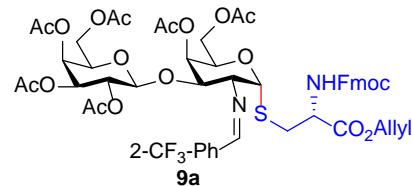




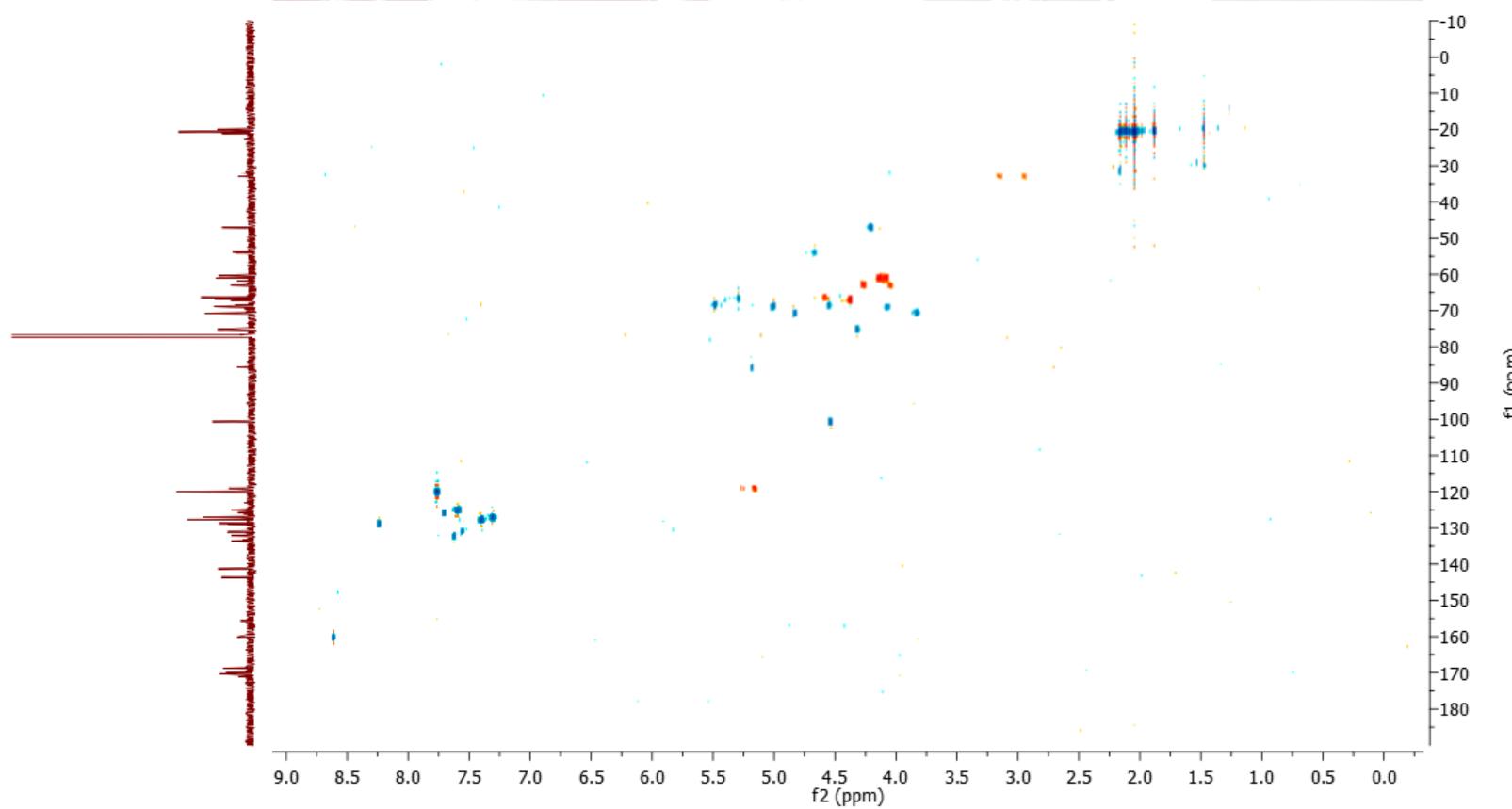


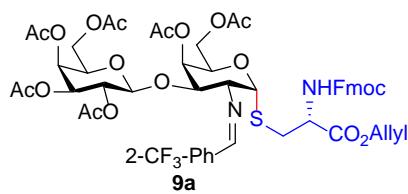




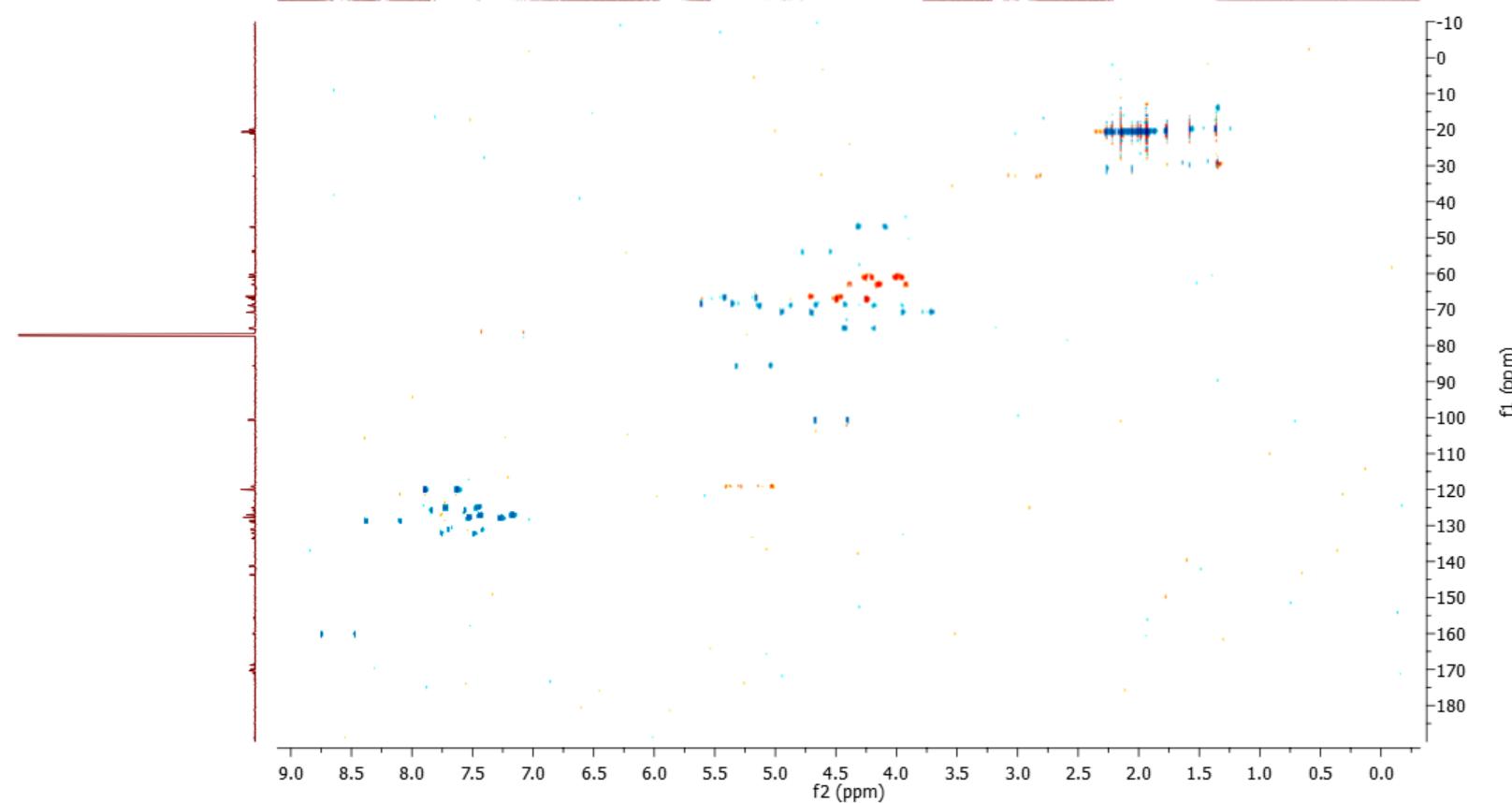


¹H-¹³C HSQC, 600/150MHz, CDCl₃

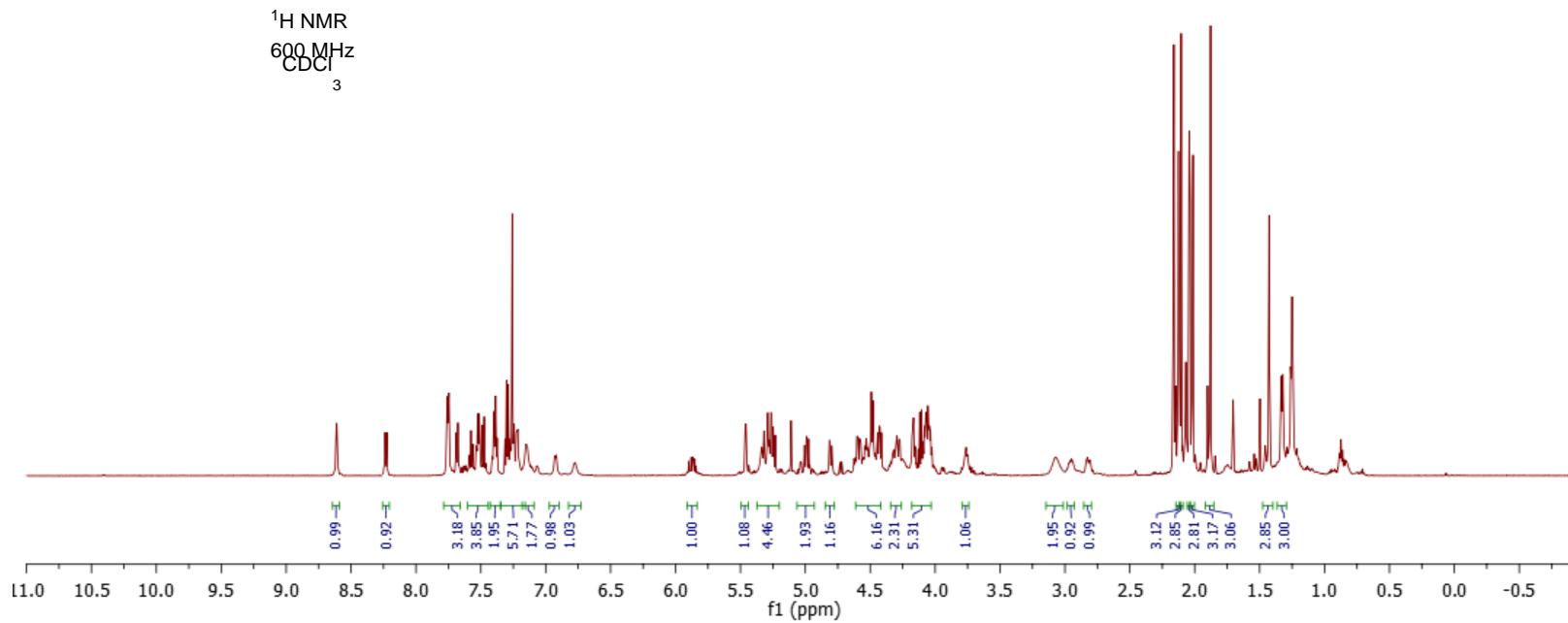
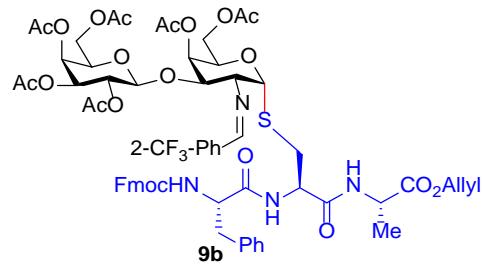


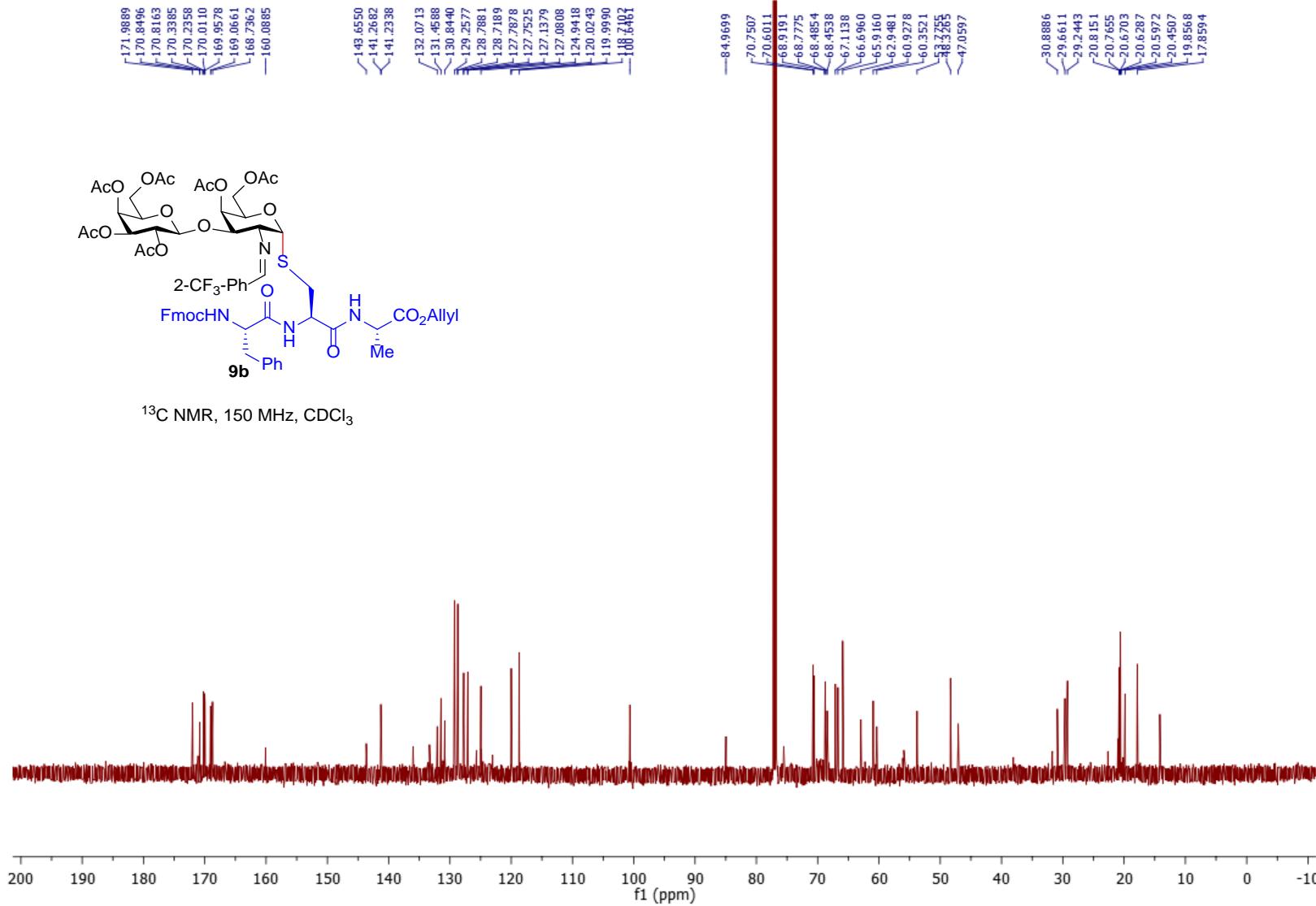


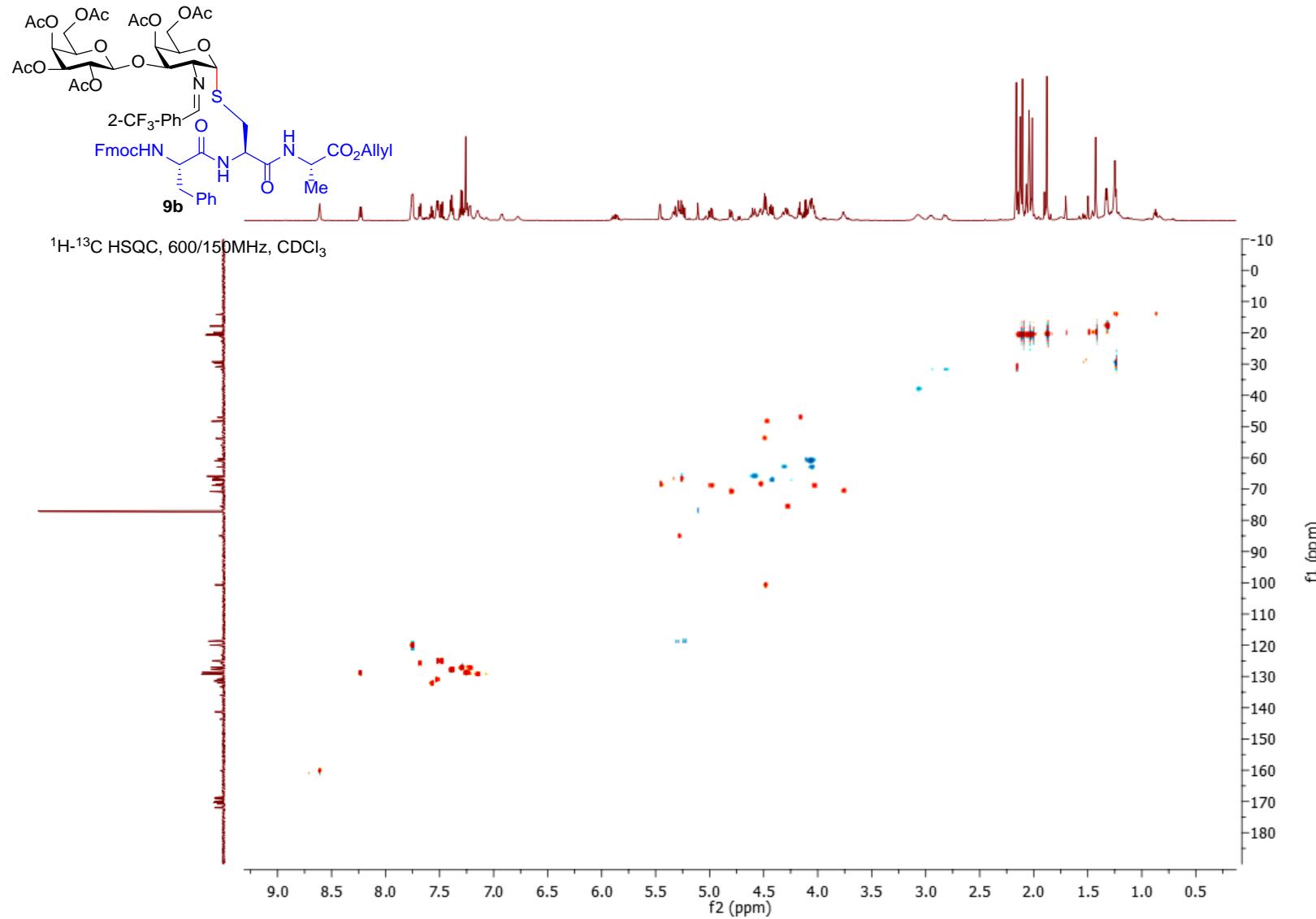
^1H - ^{13}C Coupled HSQC, 600/150MHz, CDCl_3

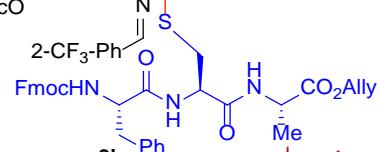
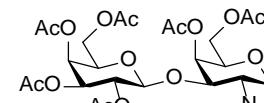


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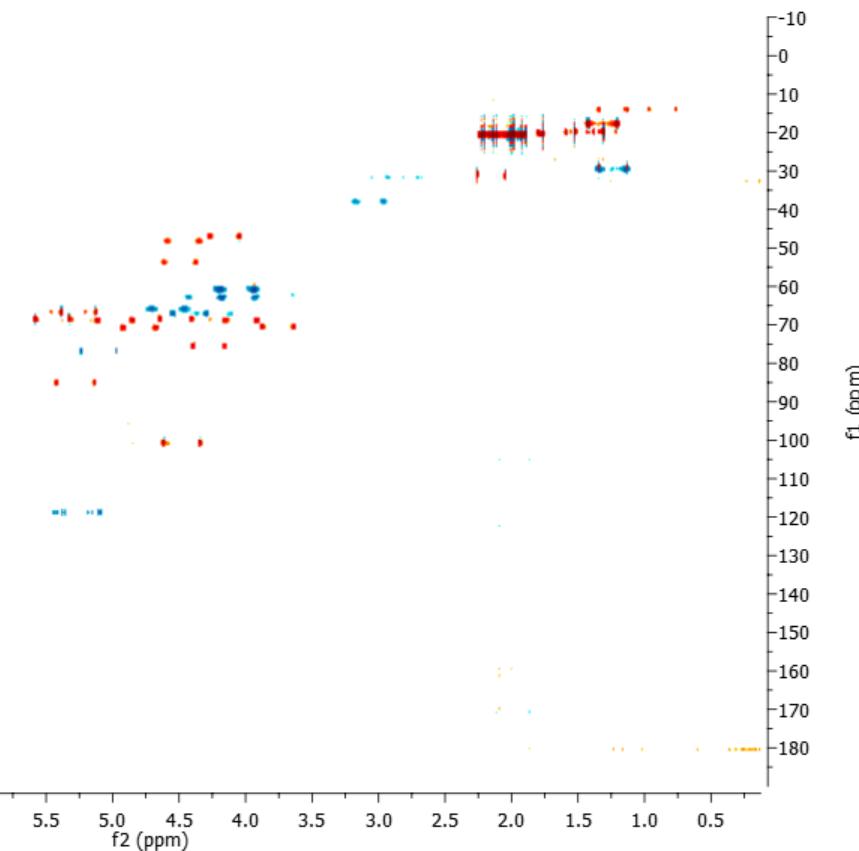


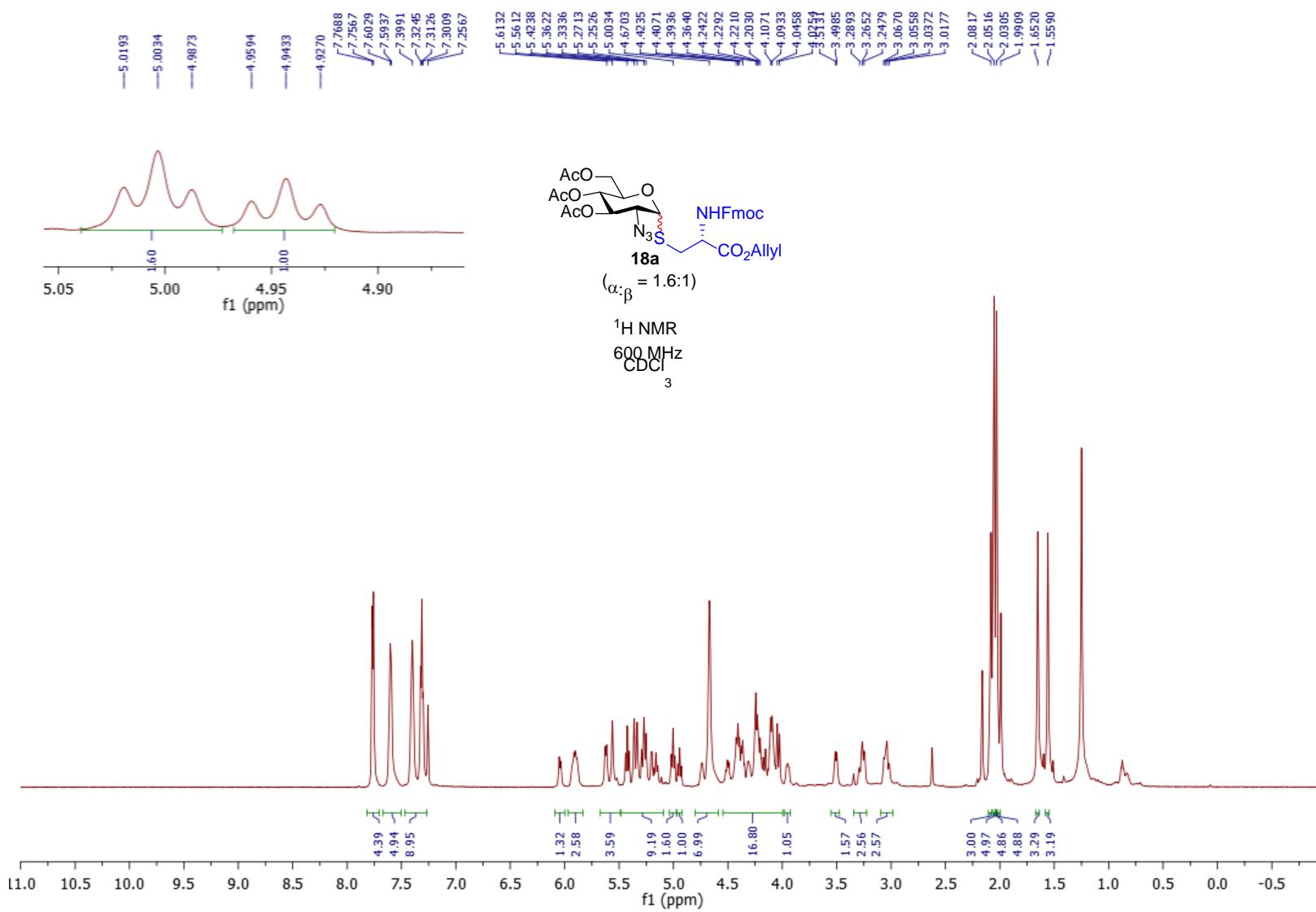


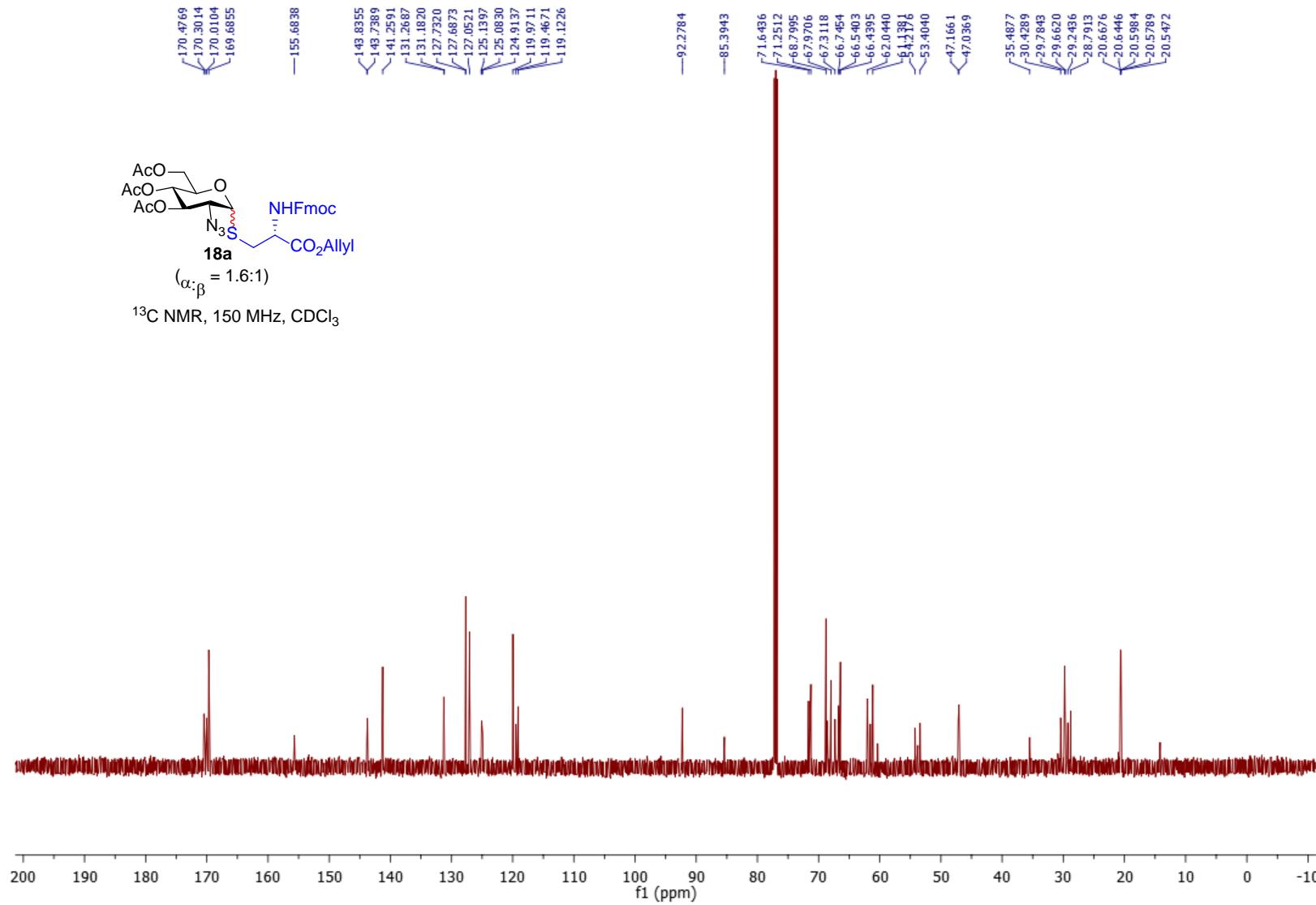


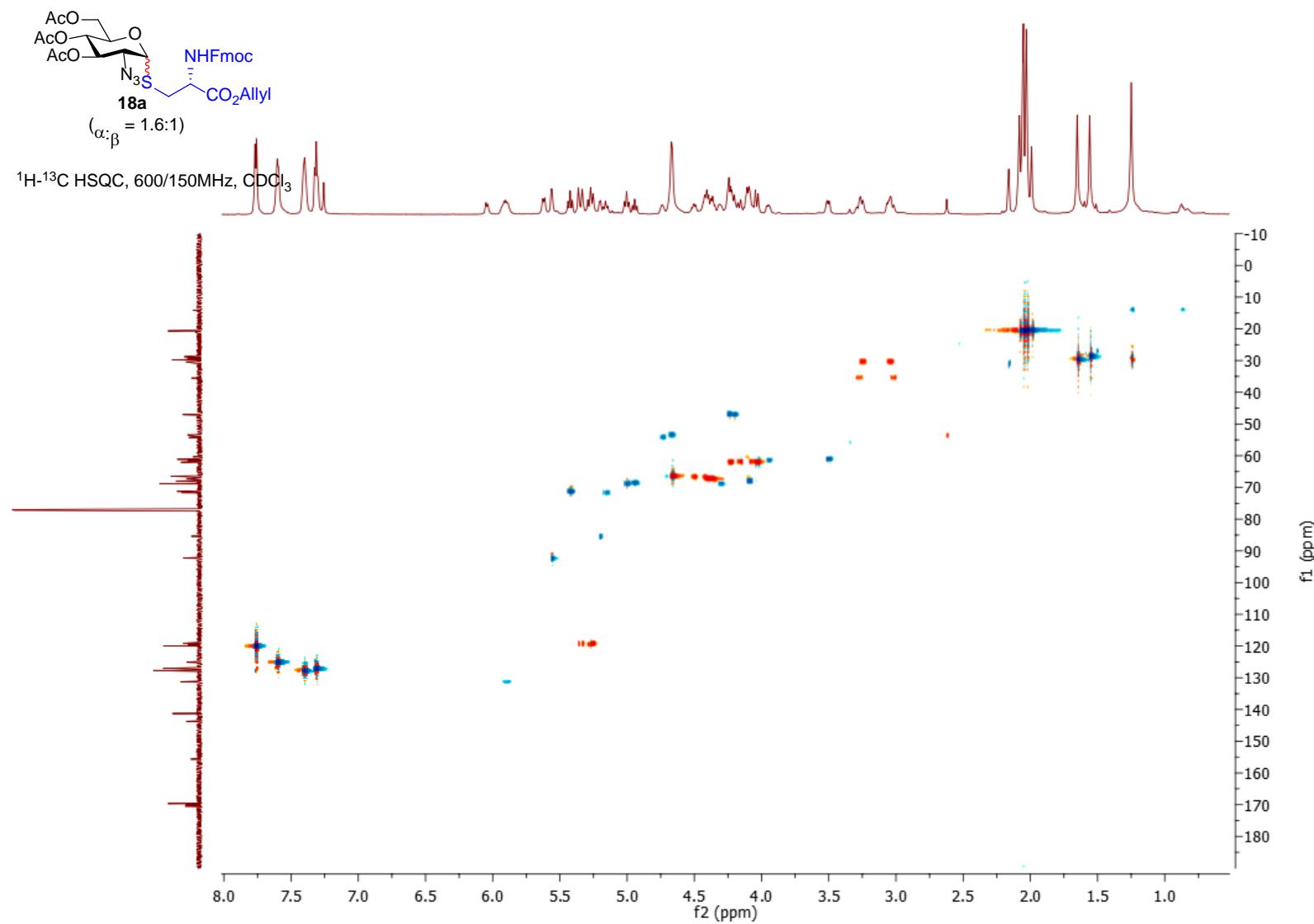


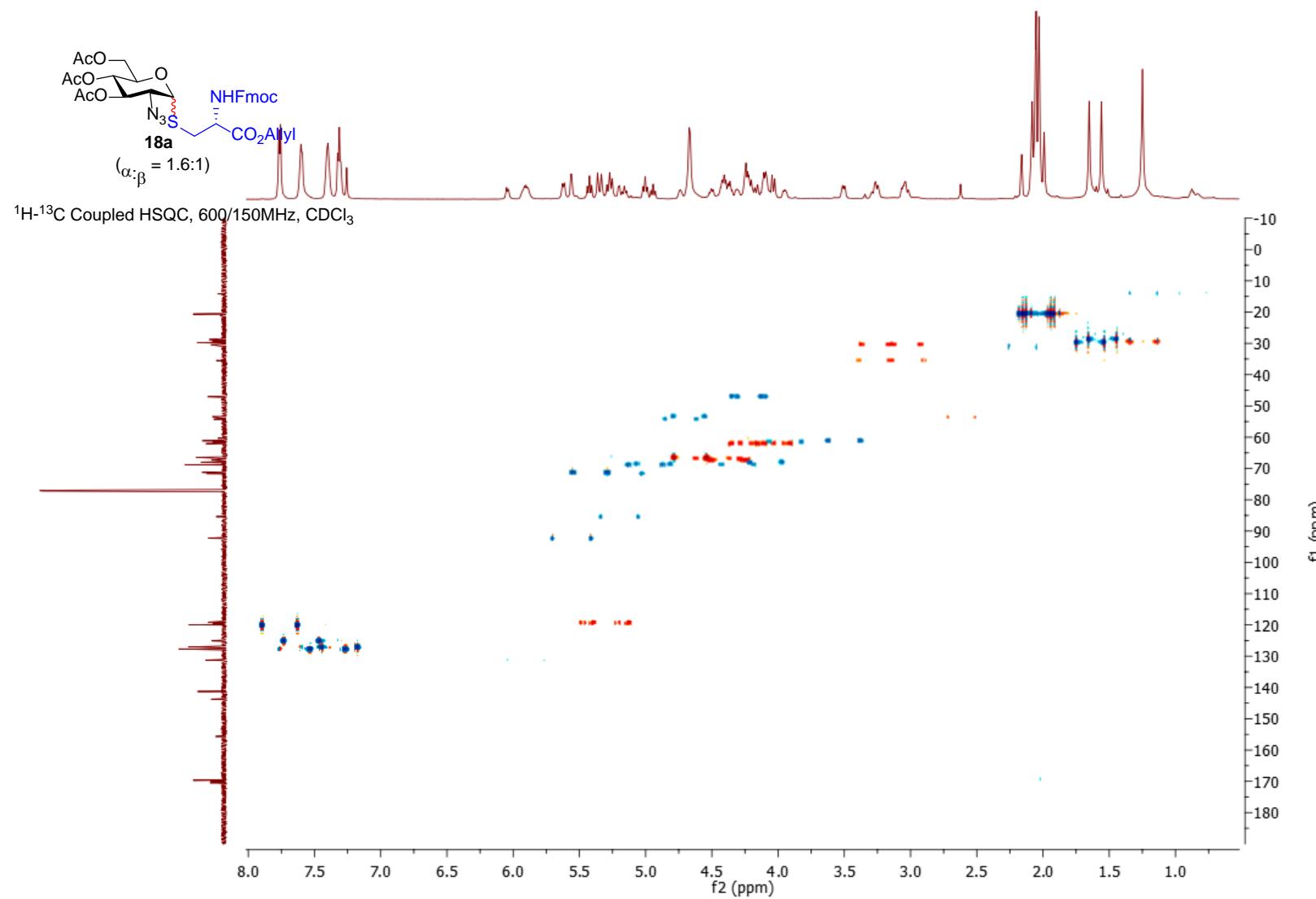
^1H - ^{13}C Coupled HSQC, 600/150MHz, CDCl_3

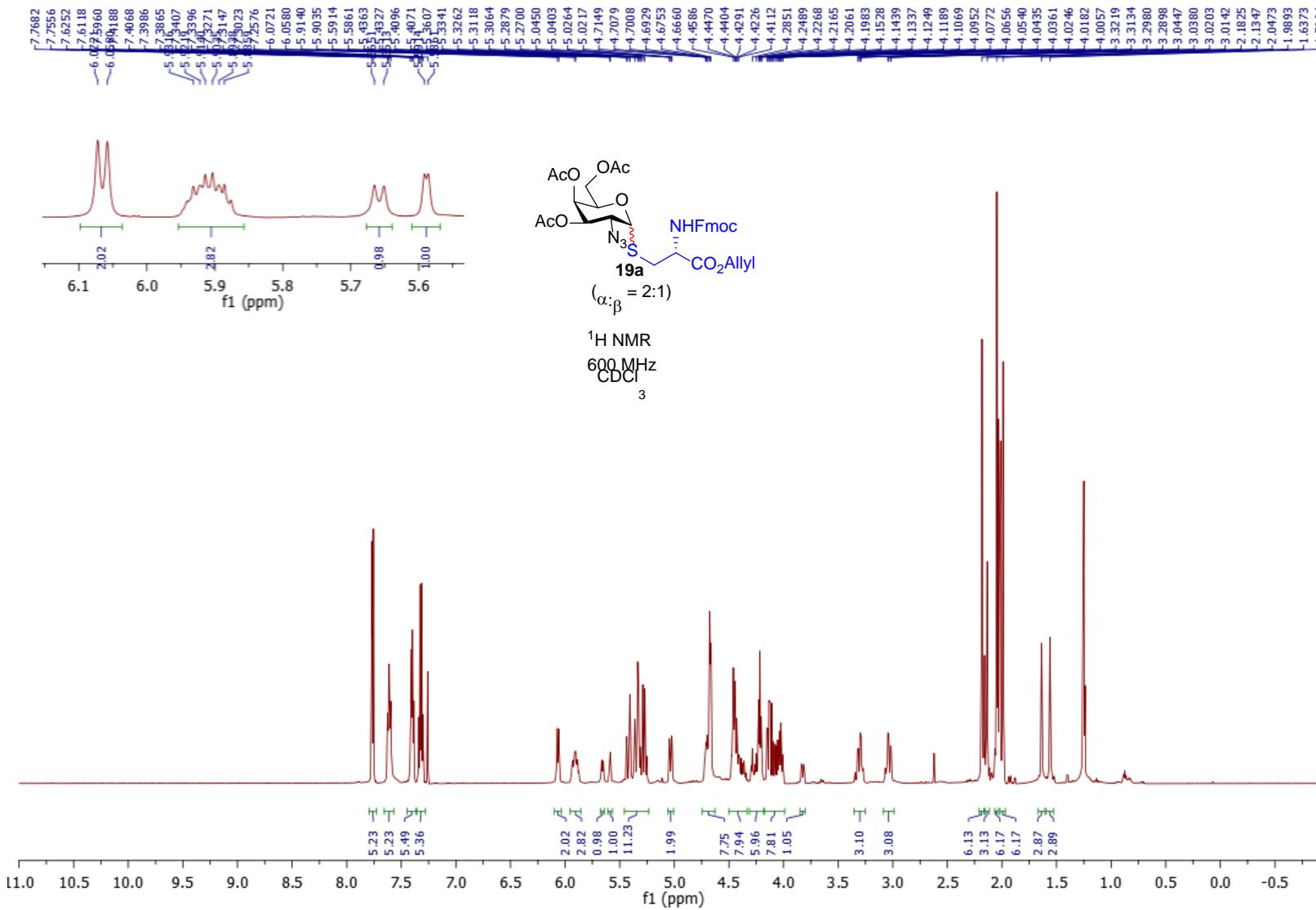


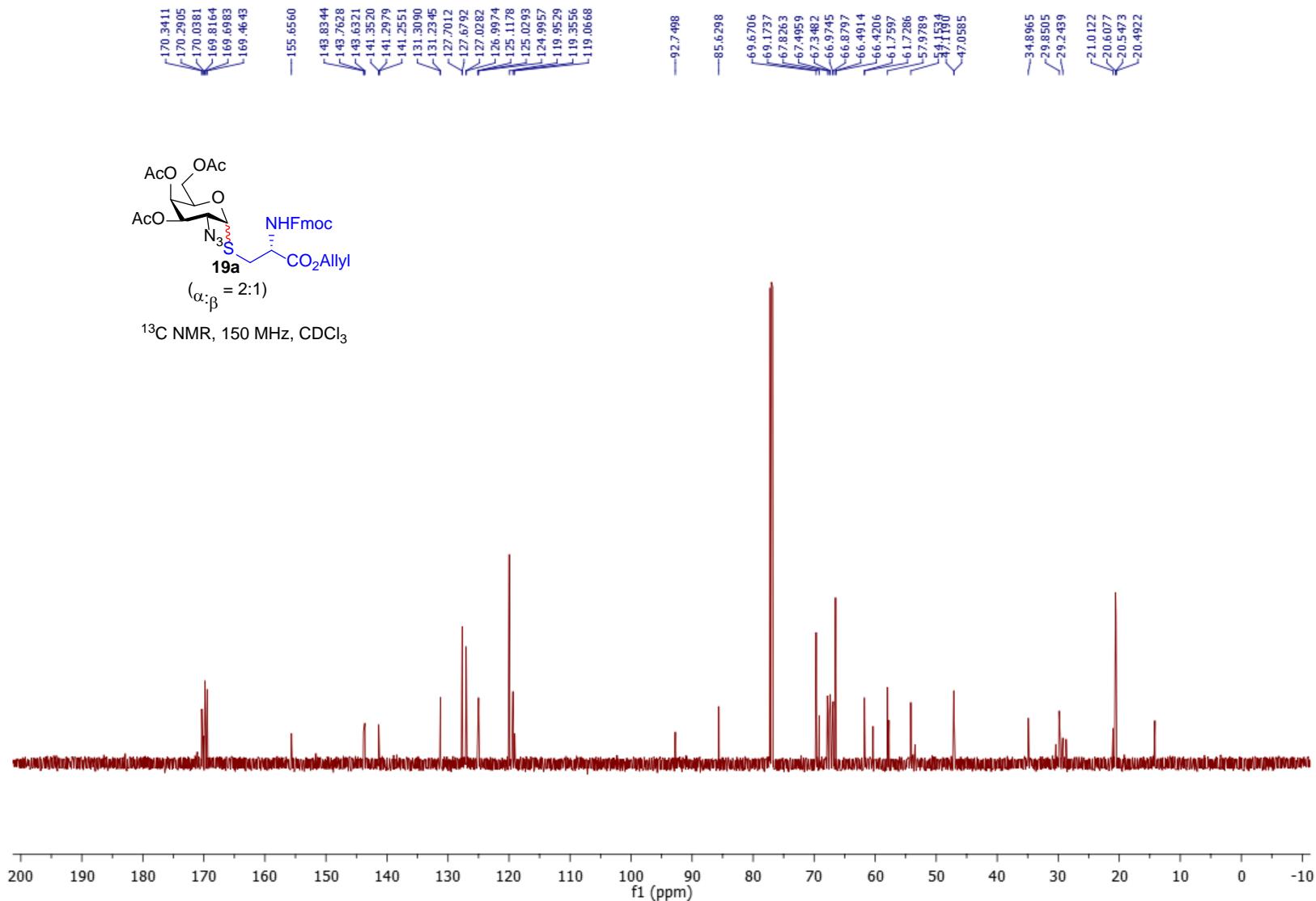


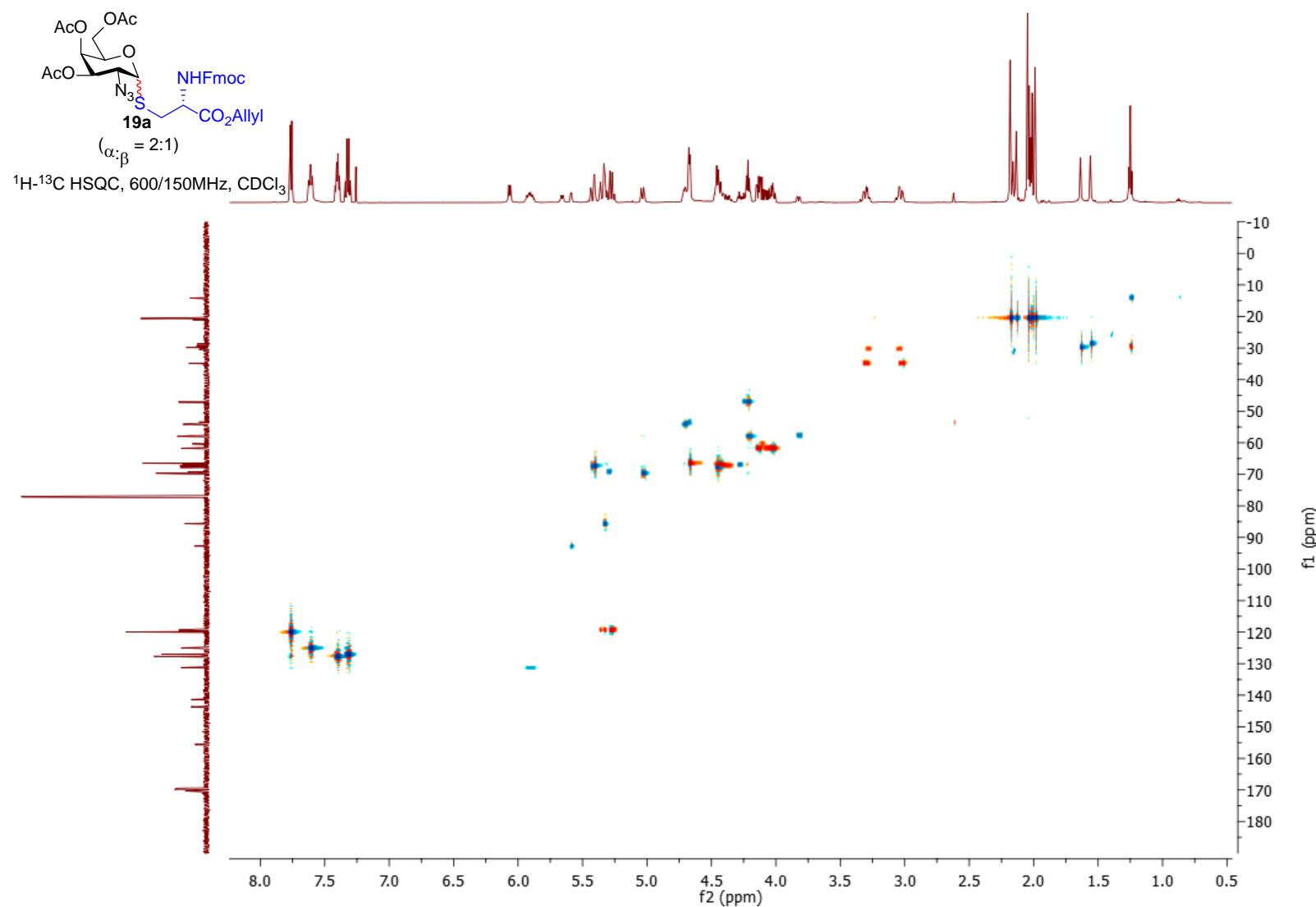


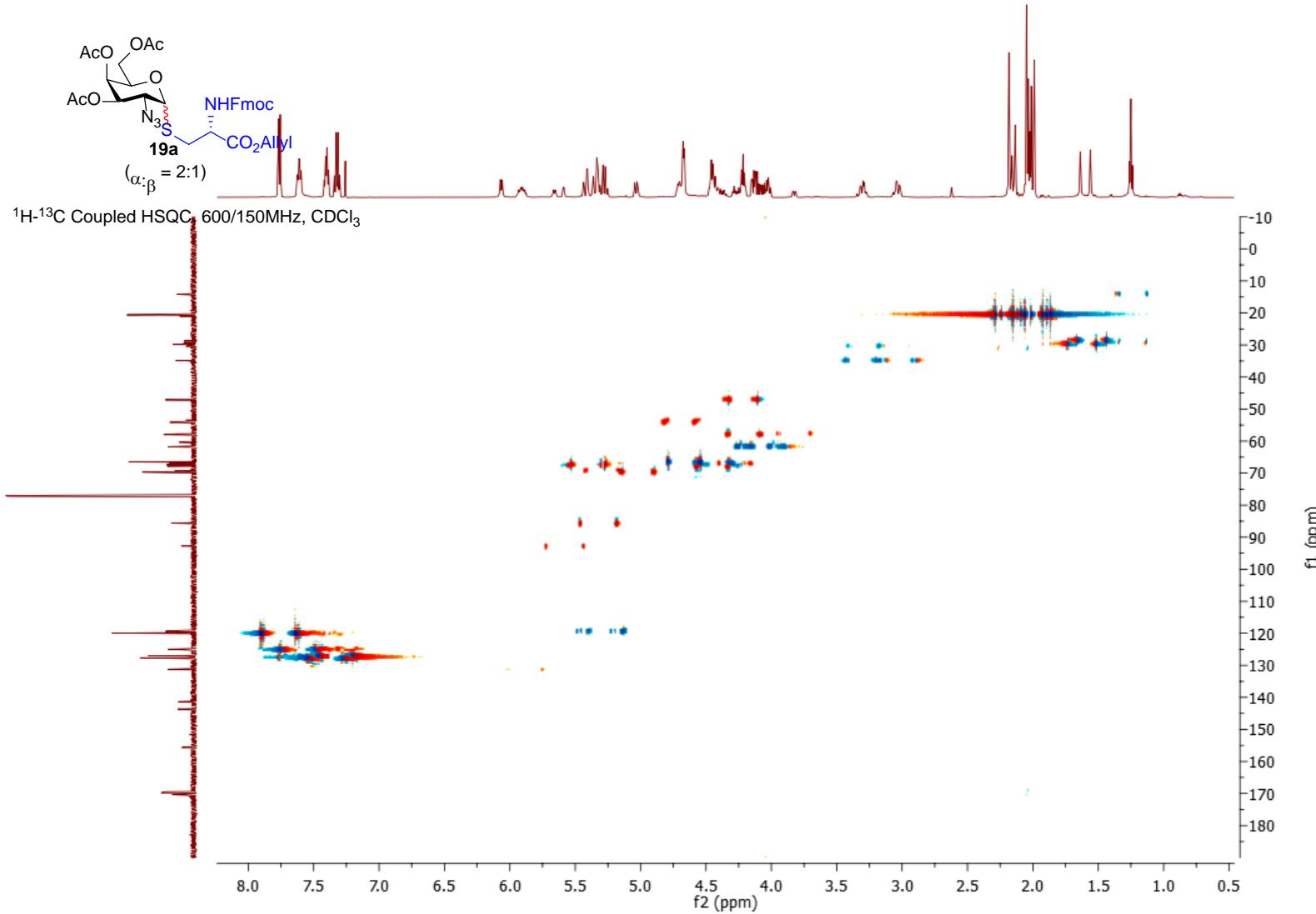


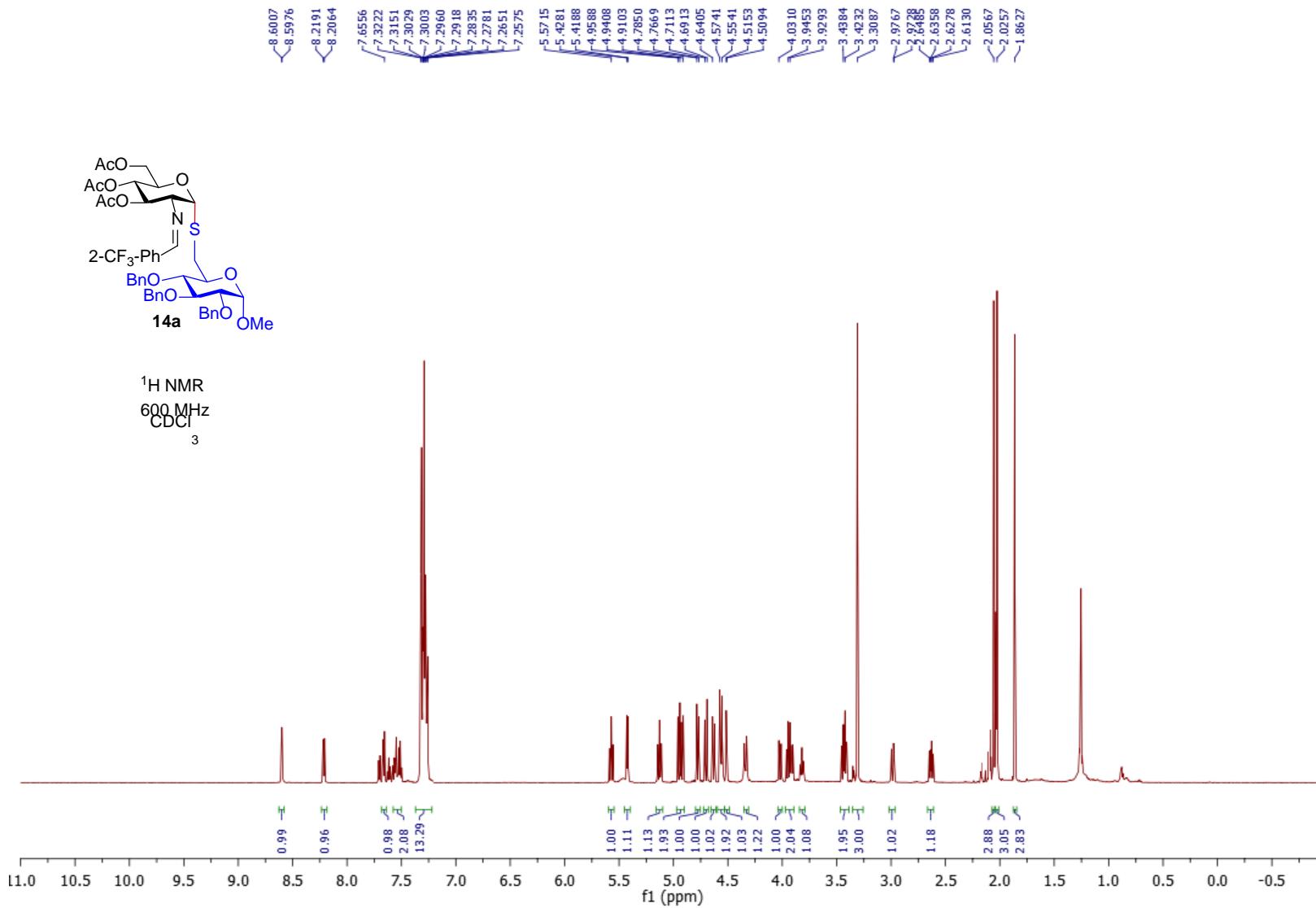


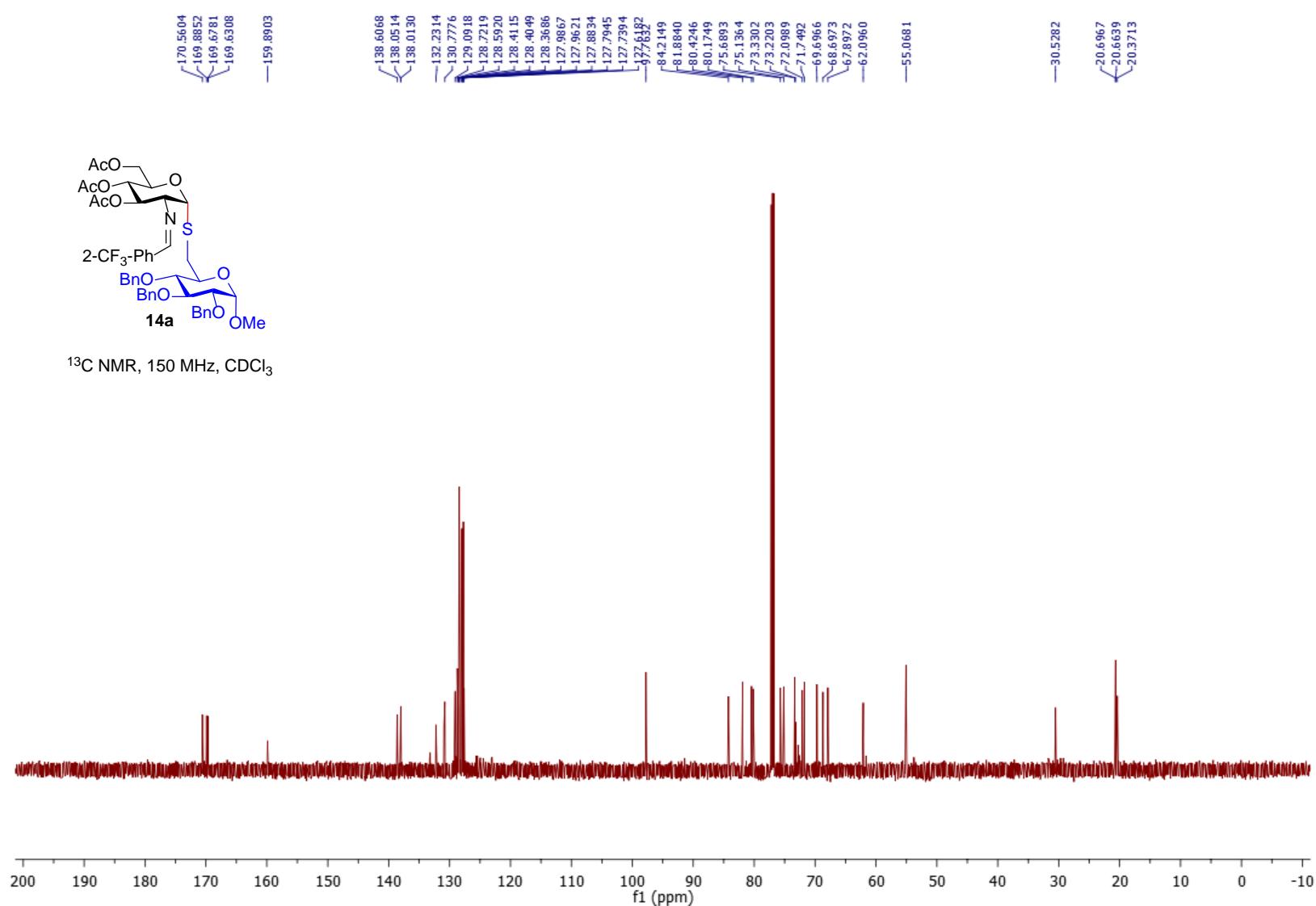


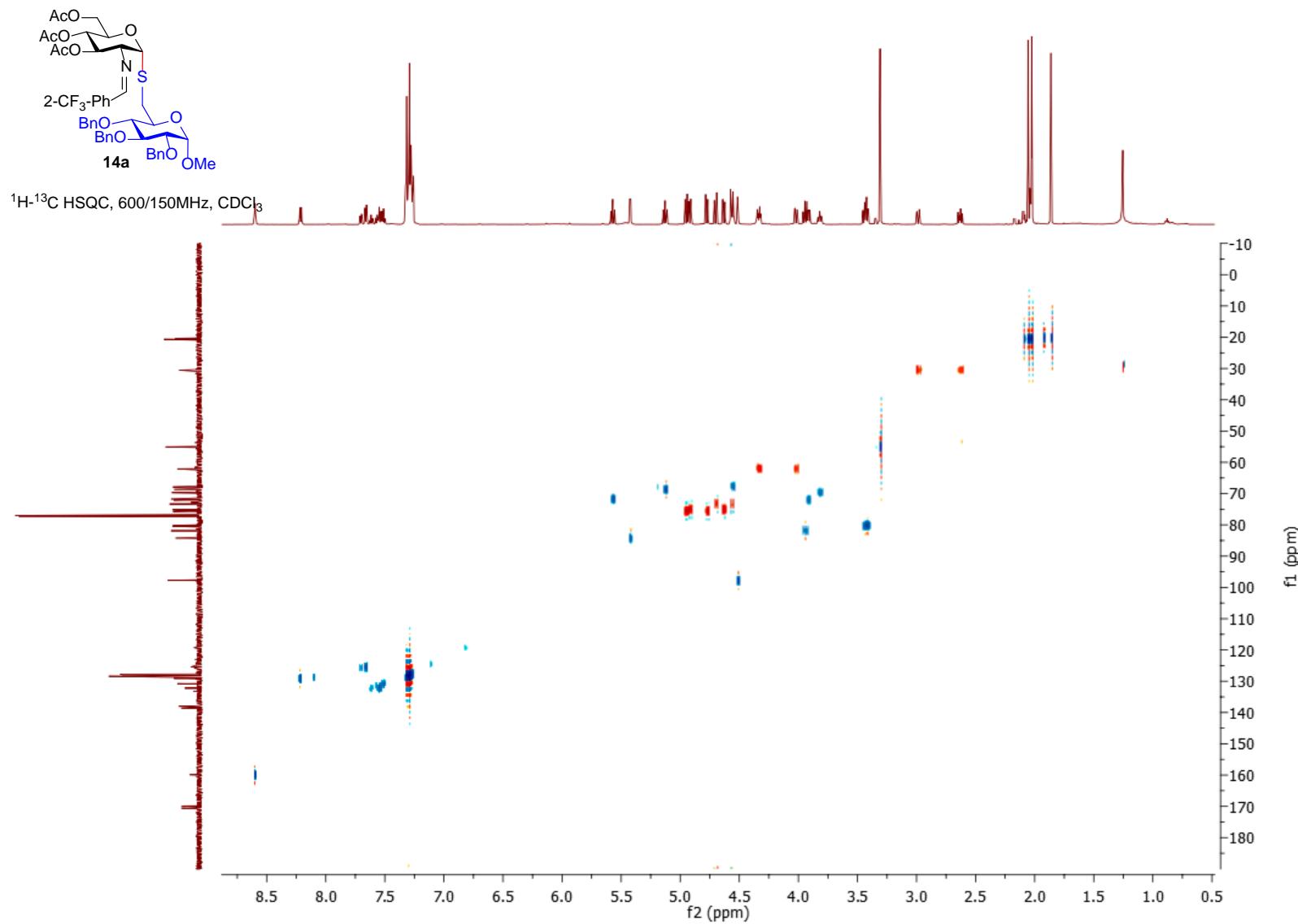


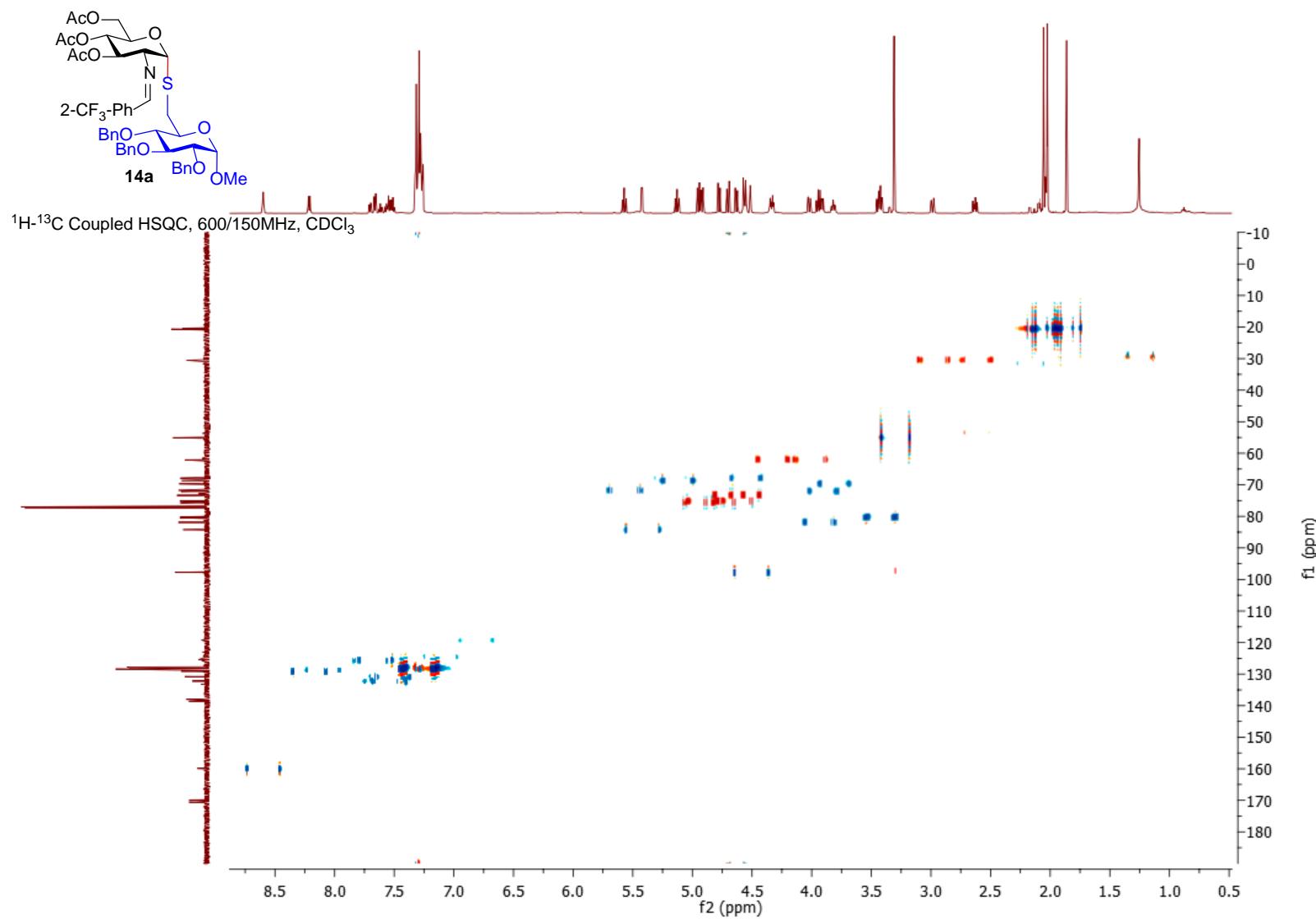


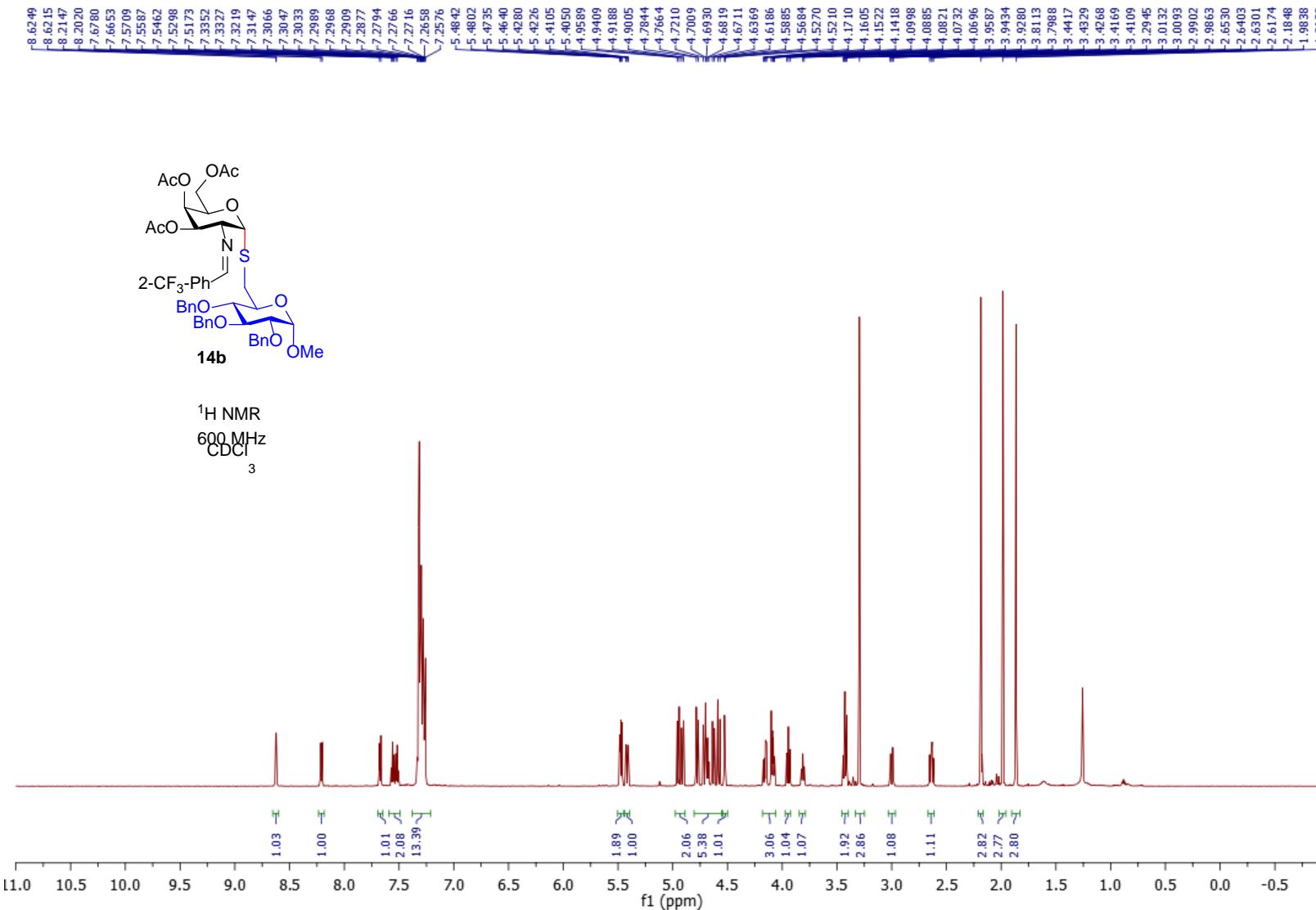


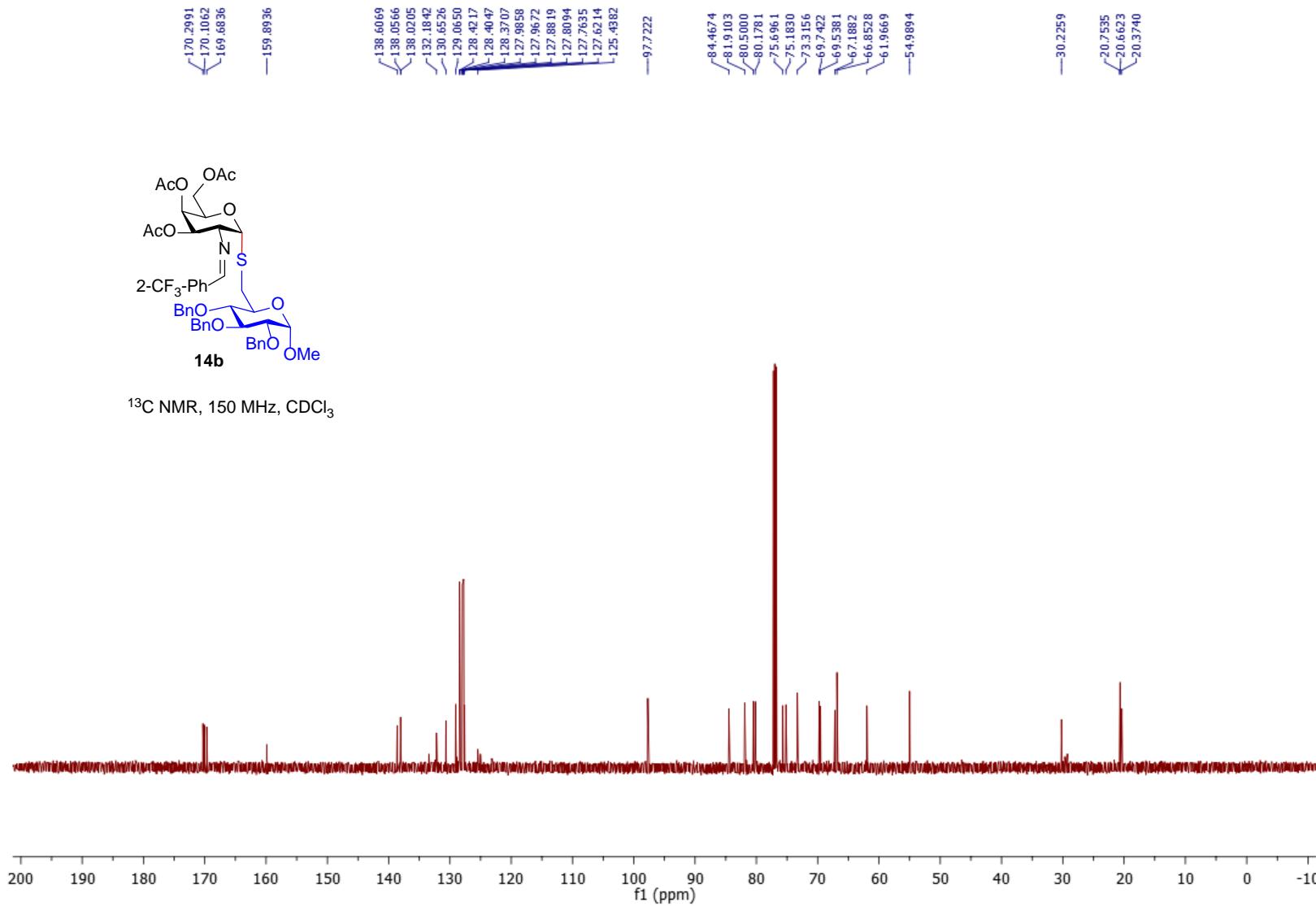


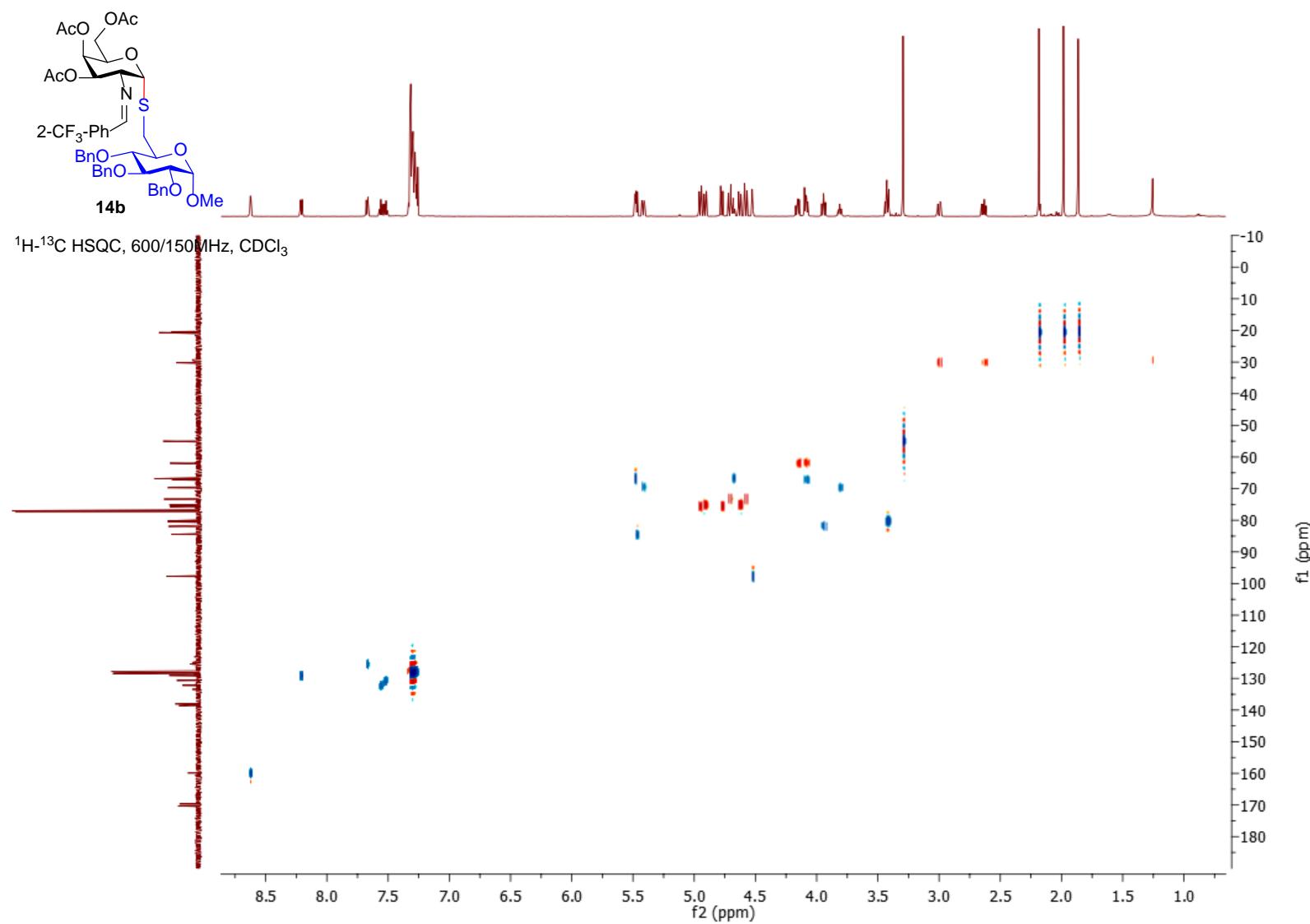


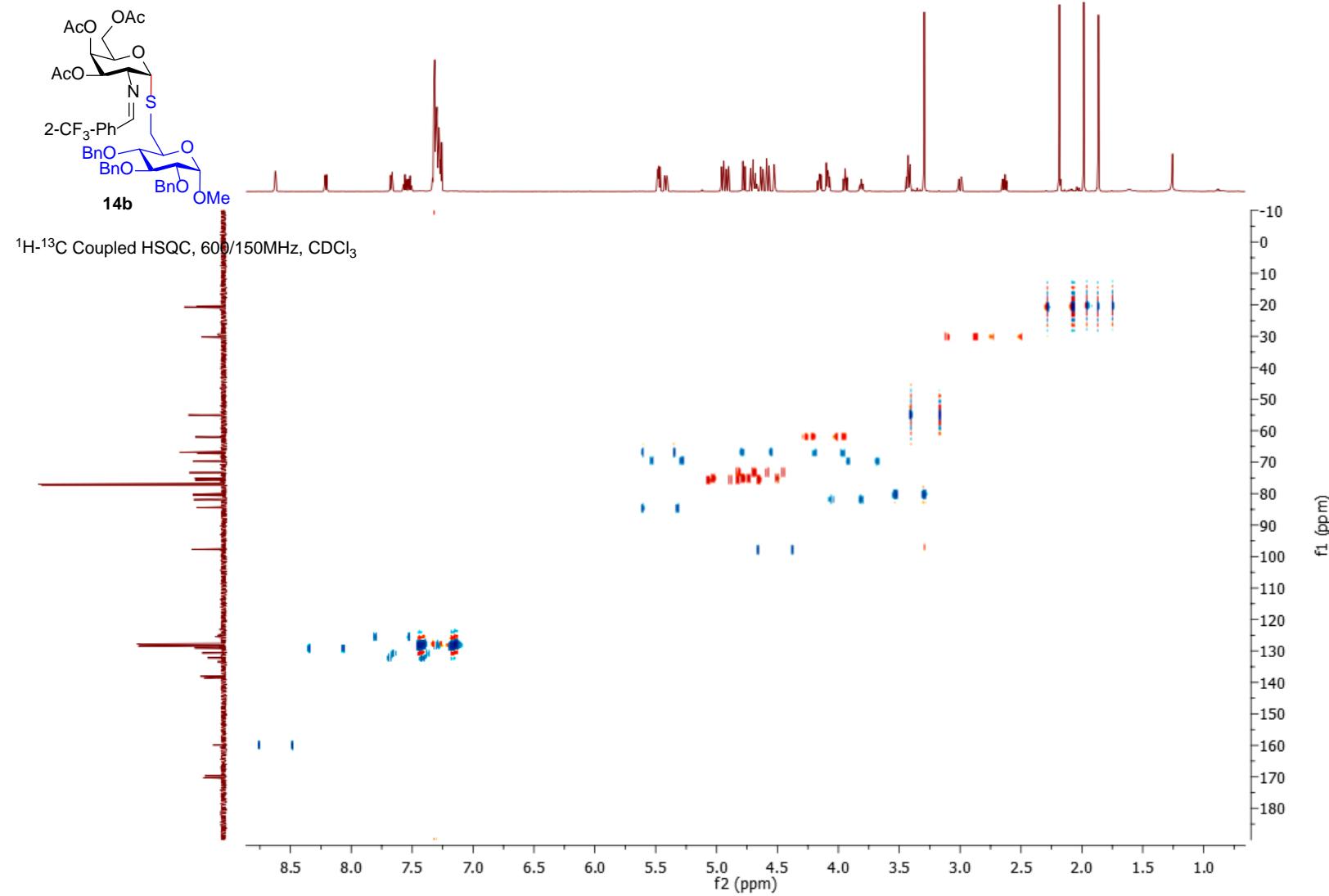




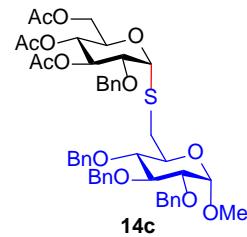




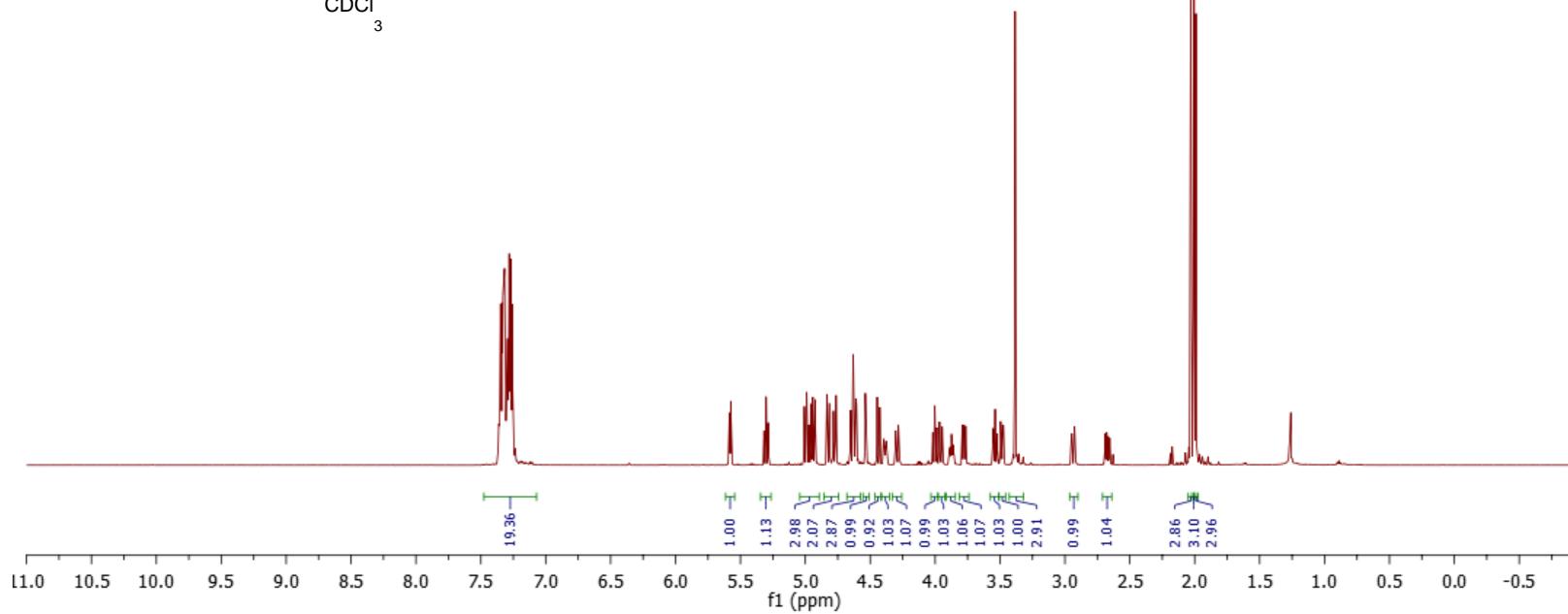


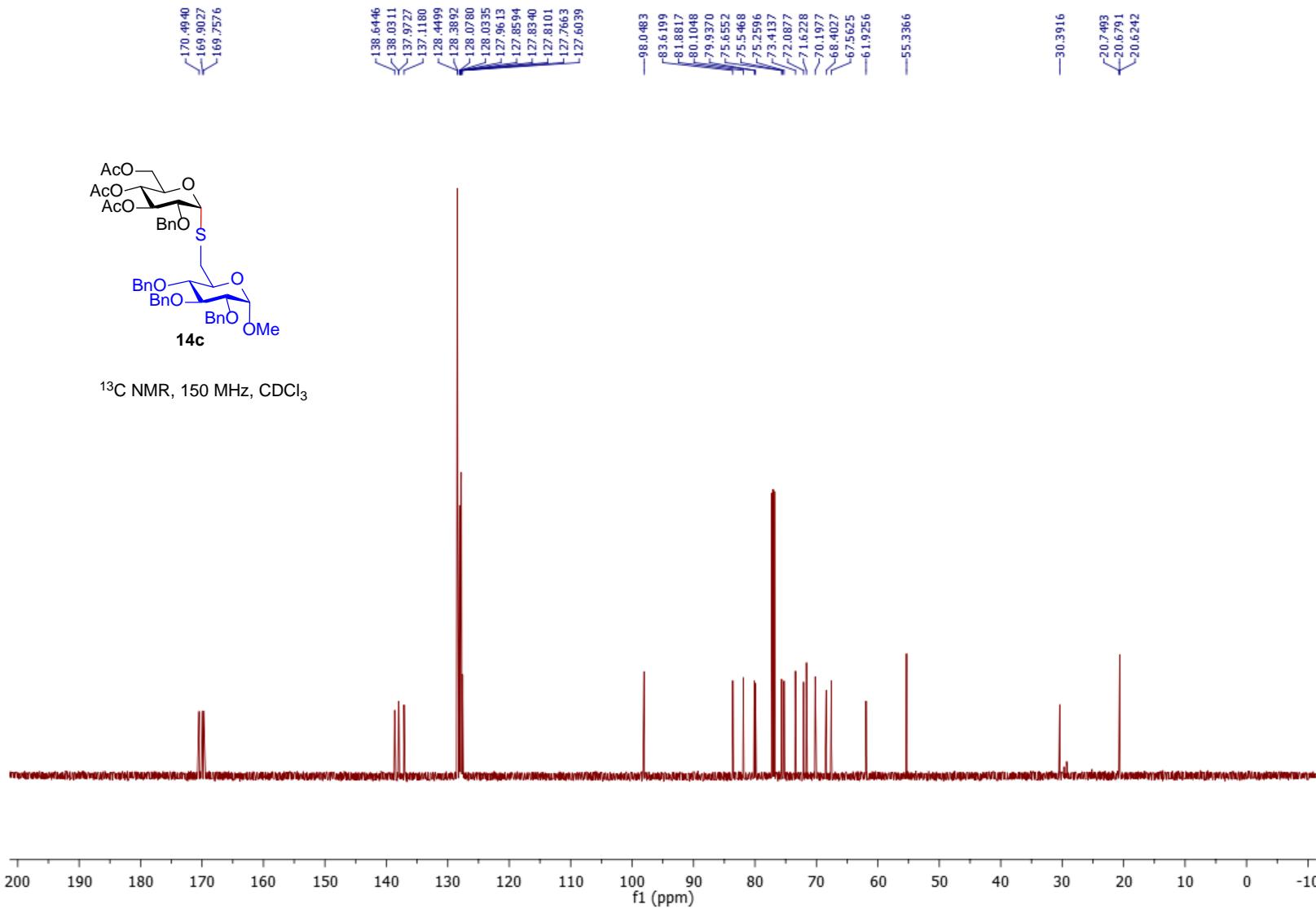


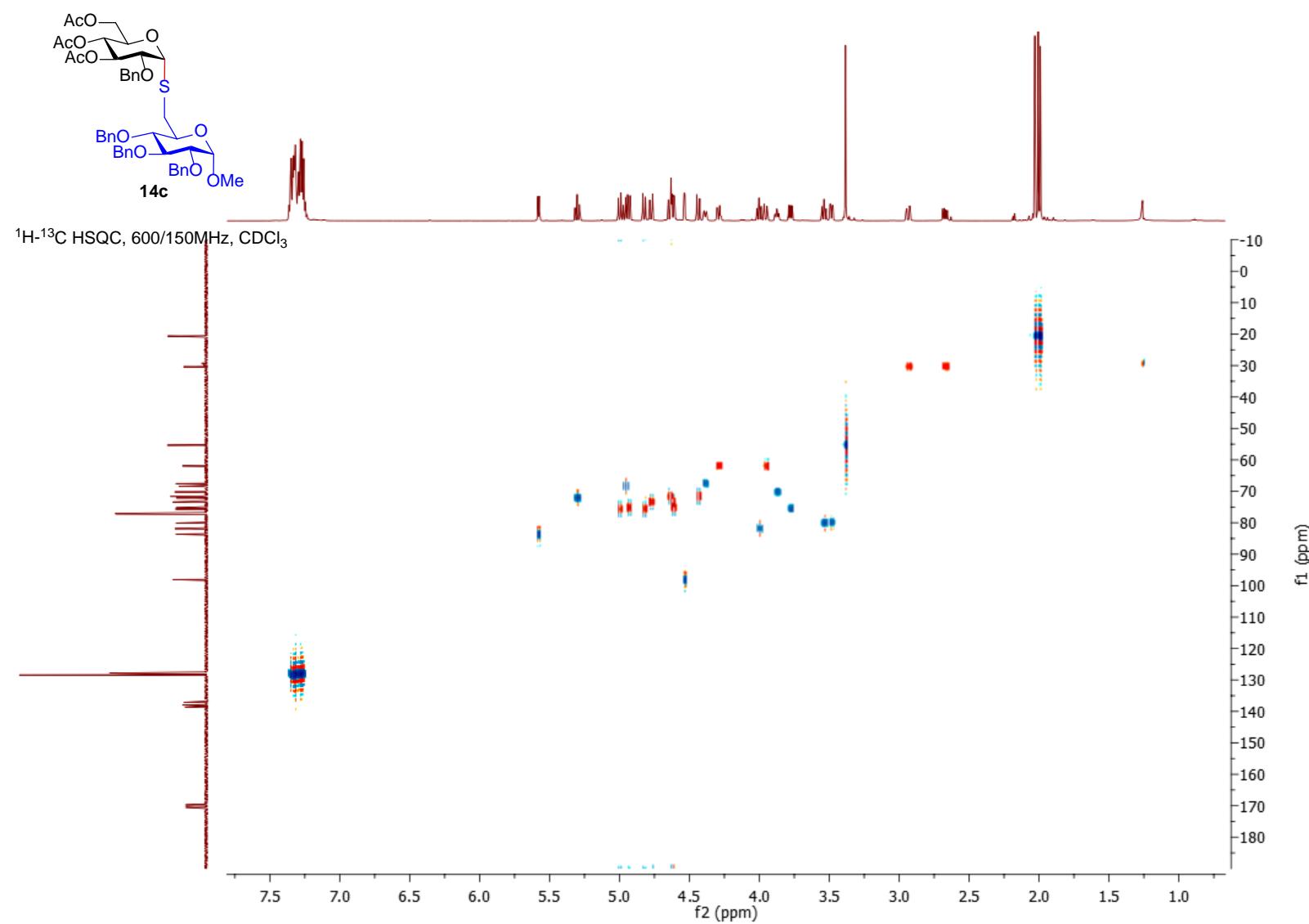
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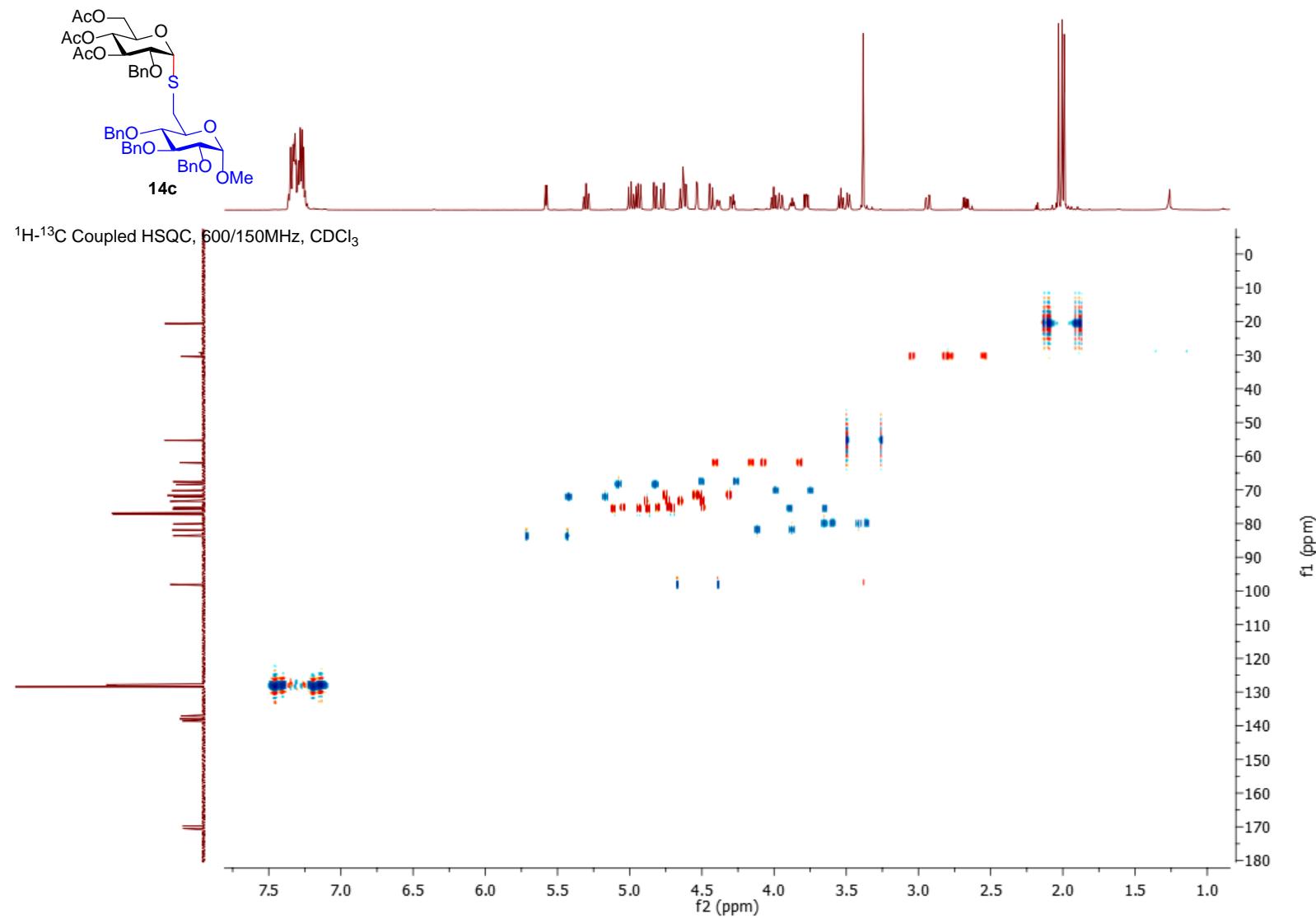


¹H NMR
 600 MHz
 CDCl₃

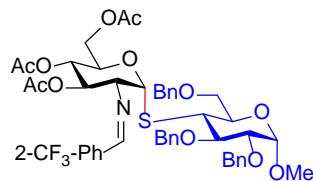




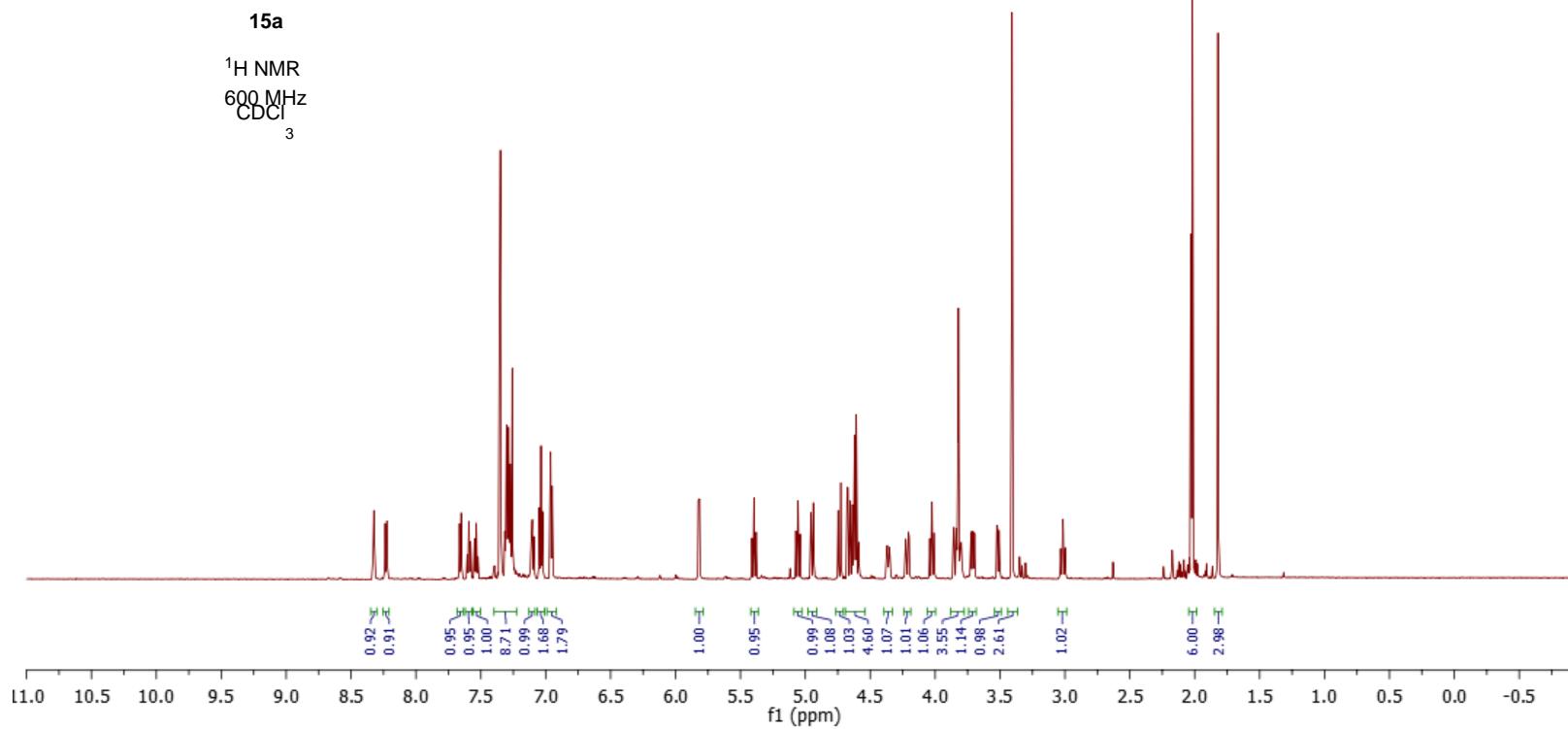


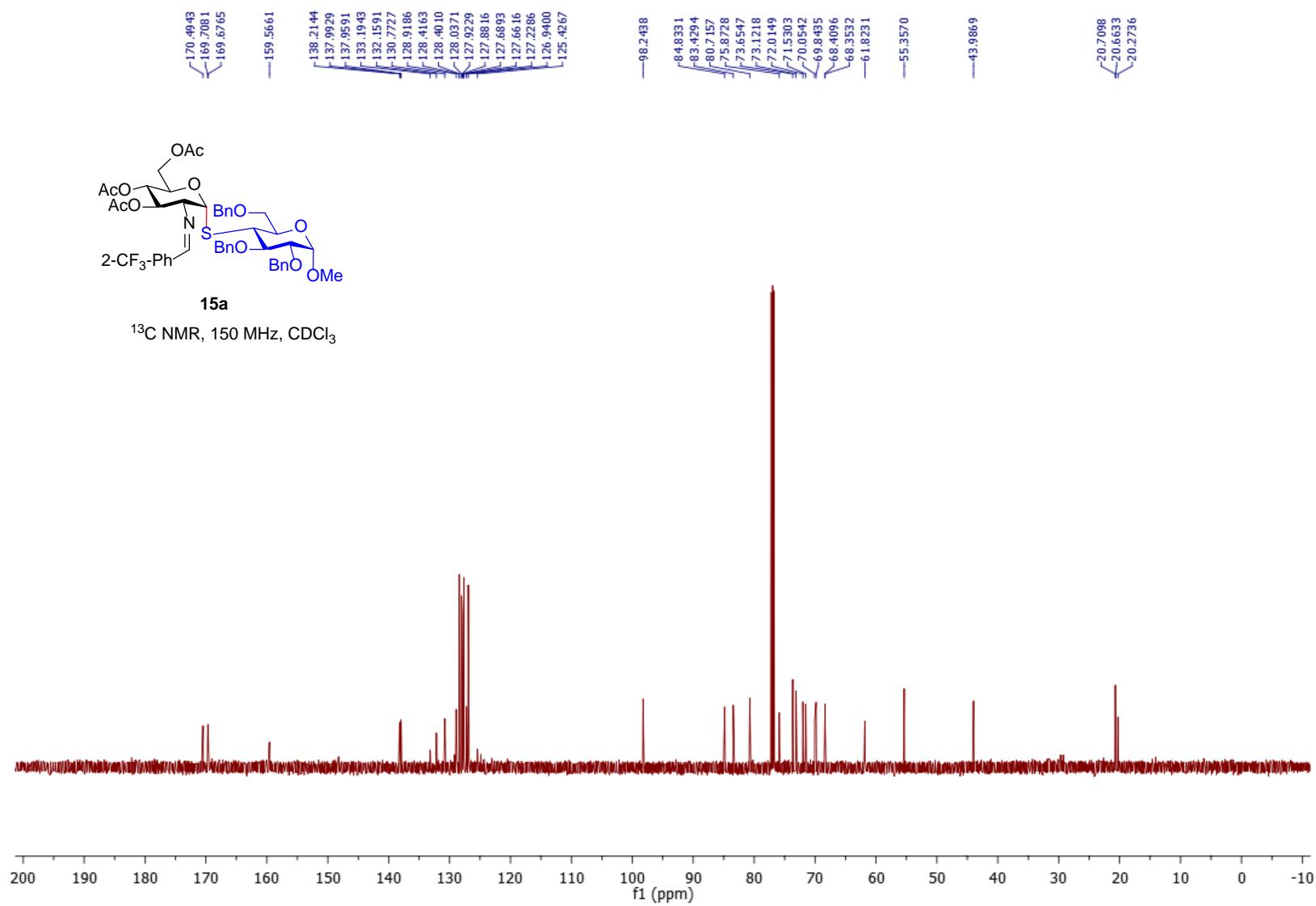


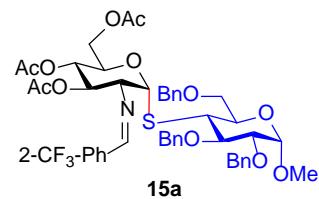
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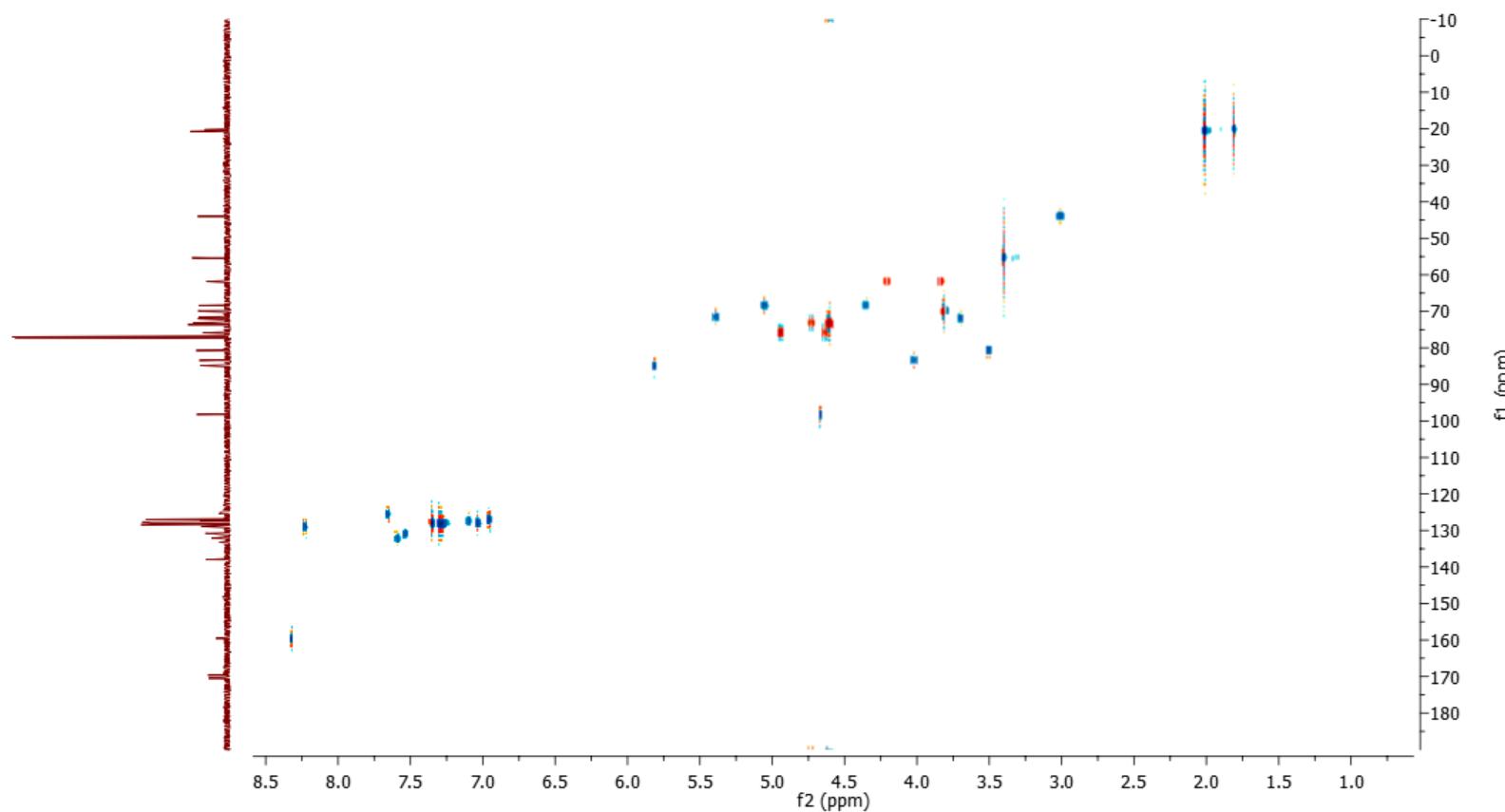
¹H NMR
600 MHz
CDCl₃

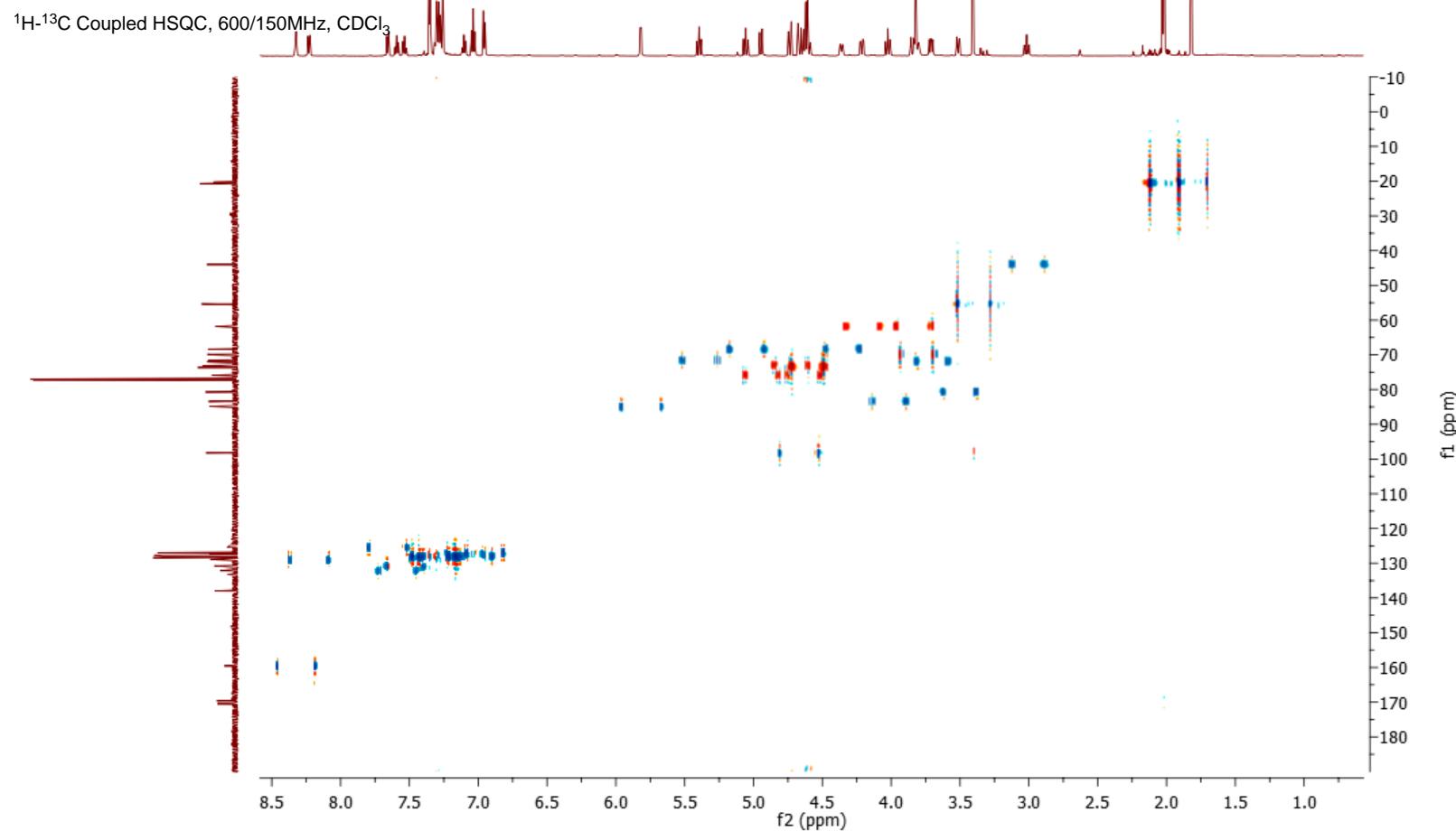
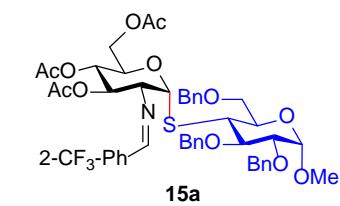


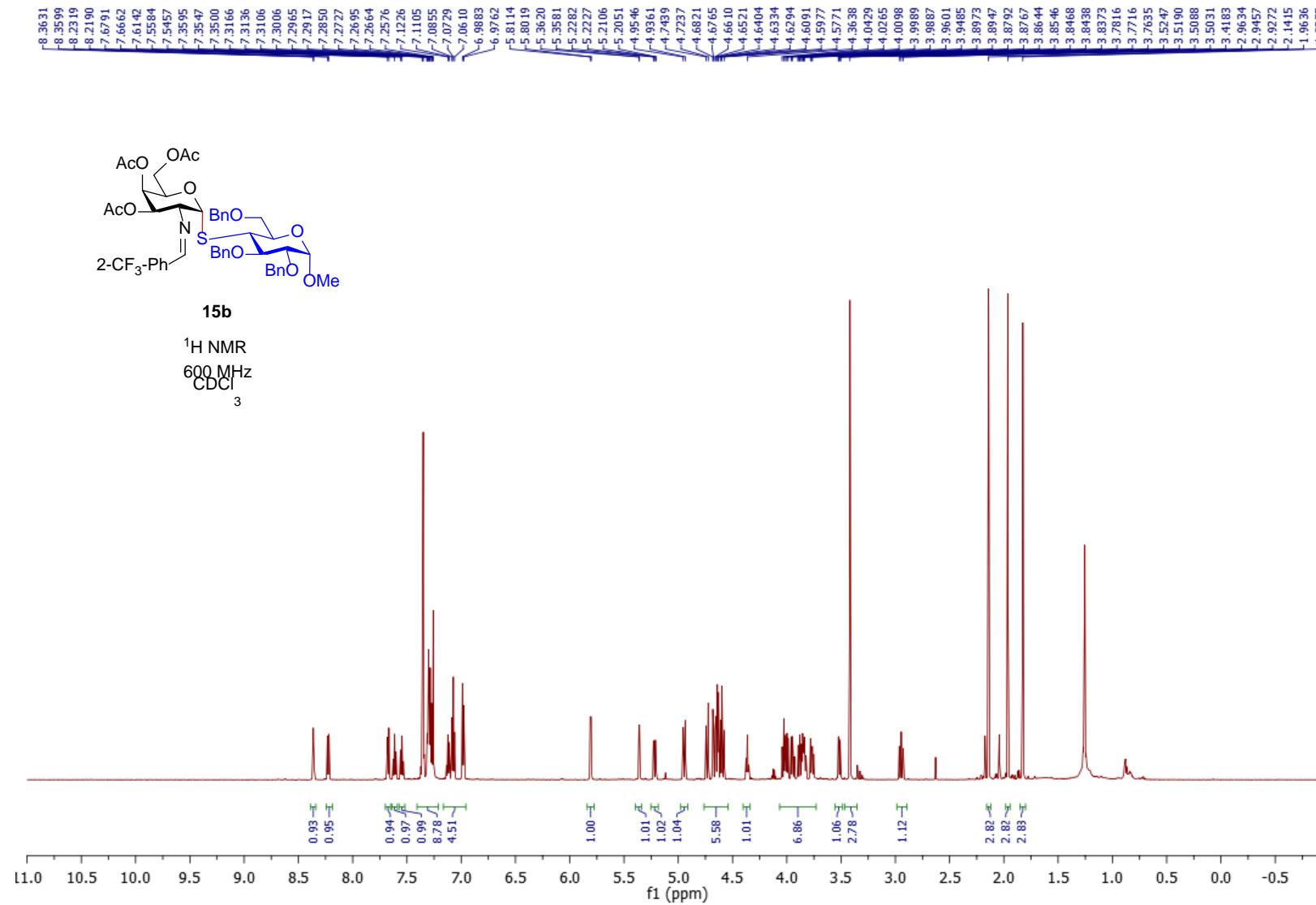


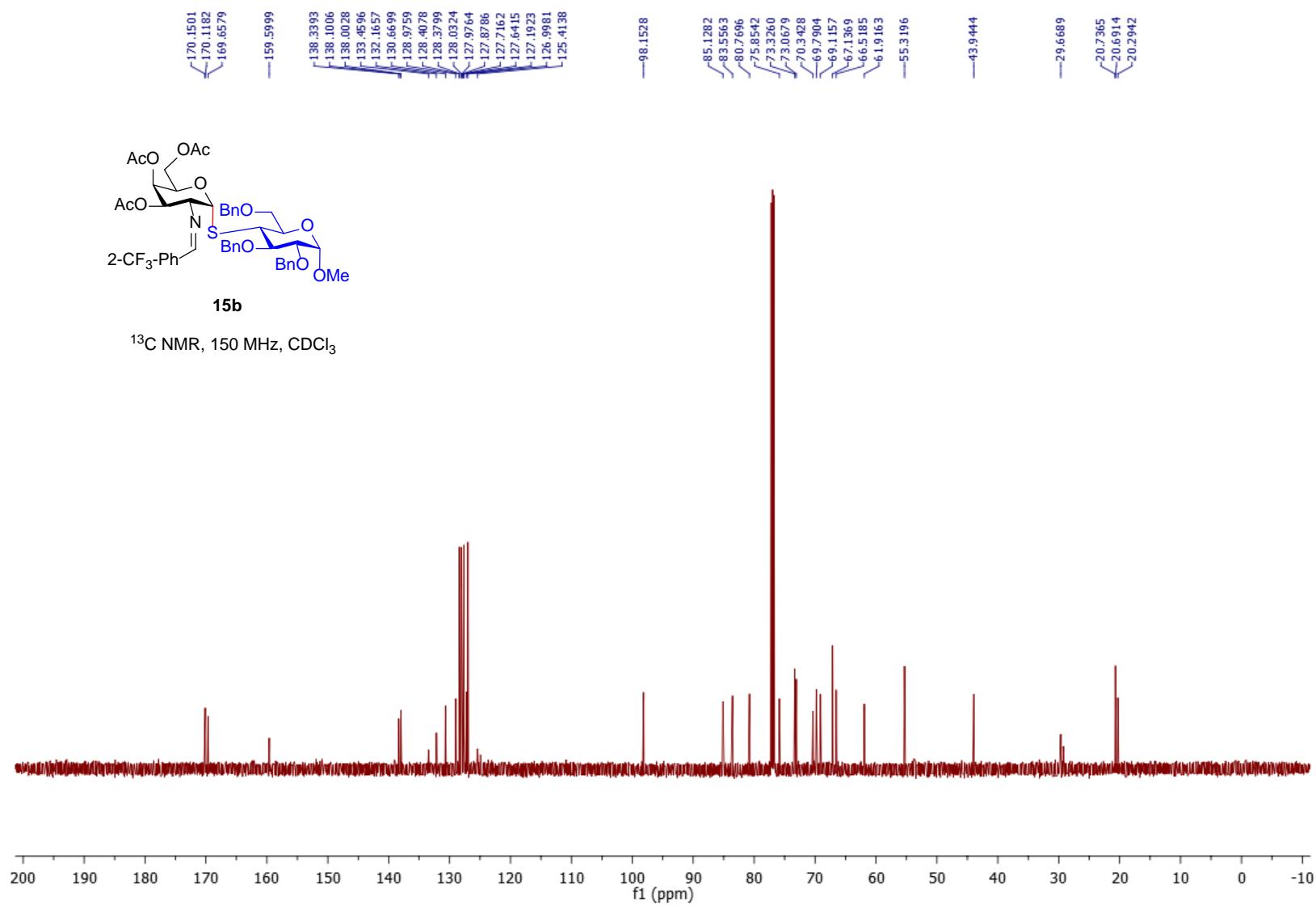


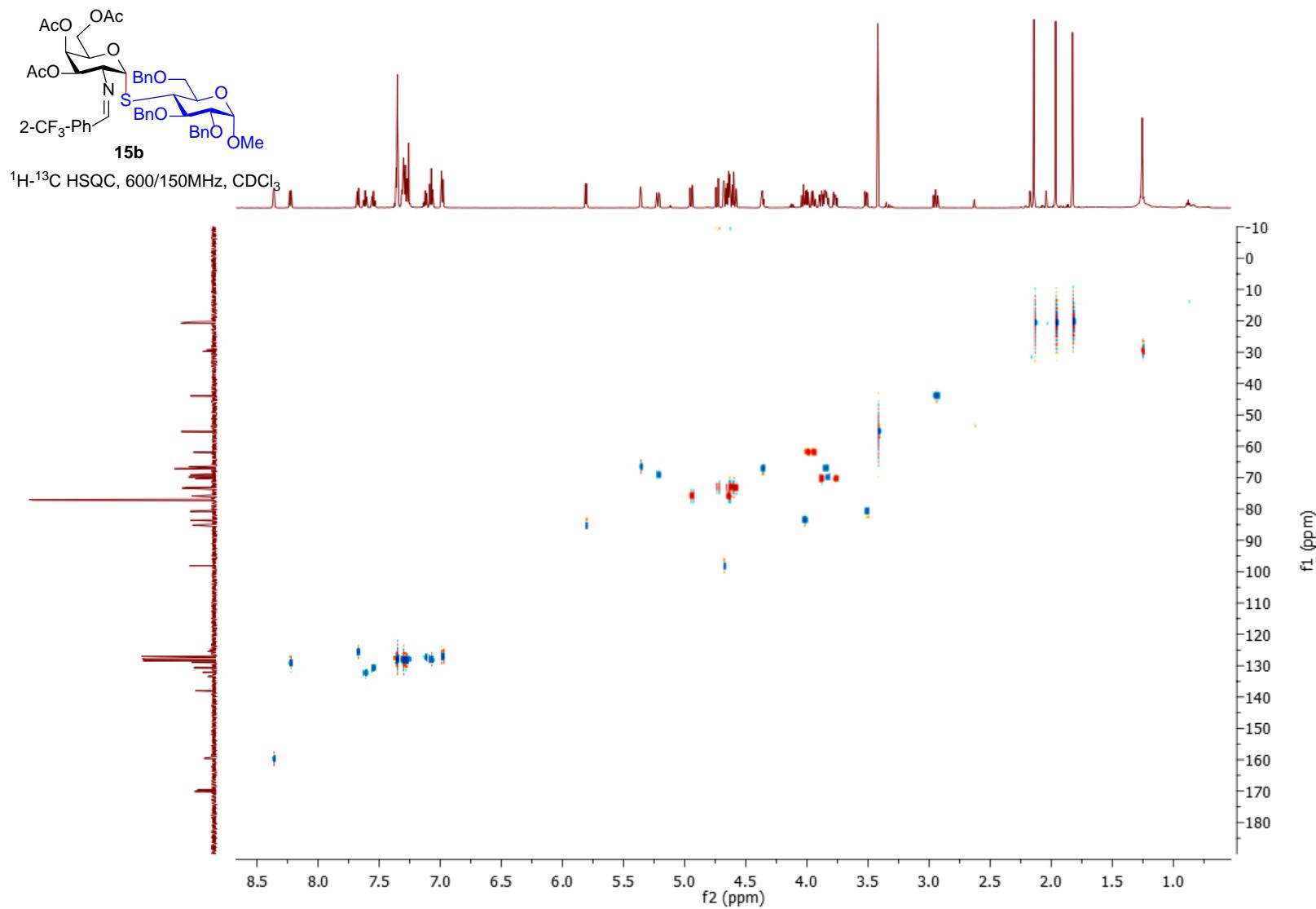
^1H - ^{13}C HSQC, 600/150MHz, CDCl_3

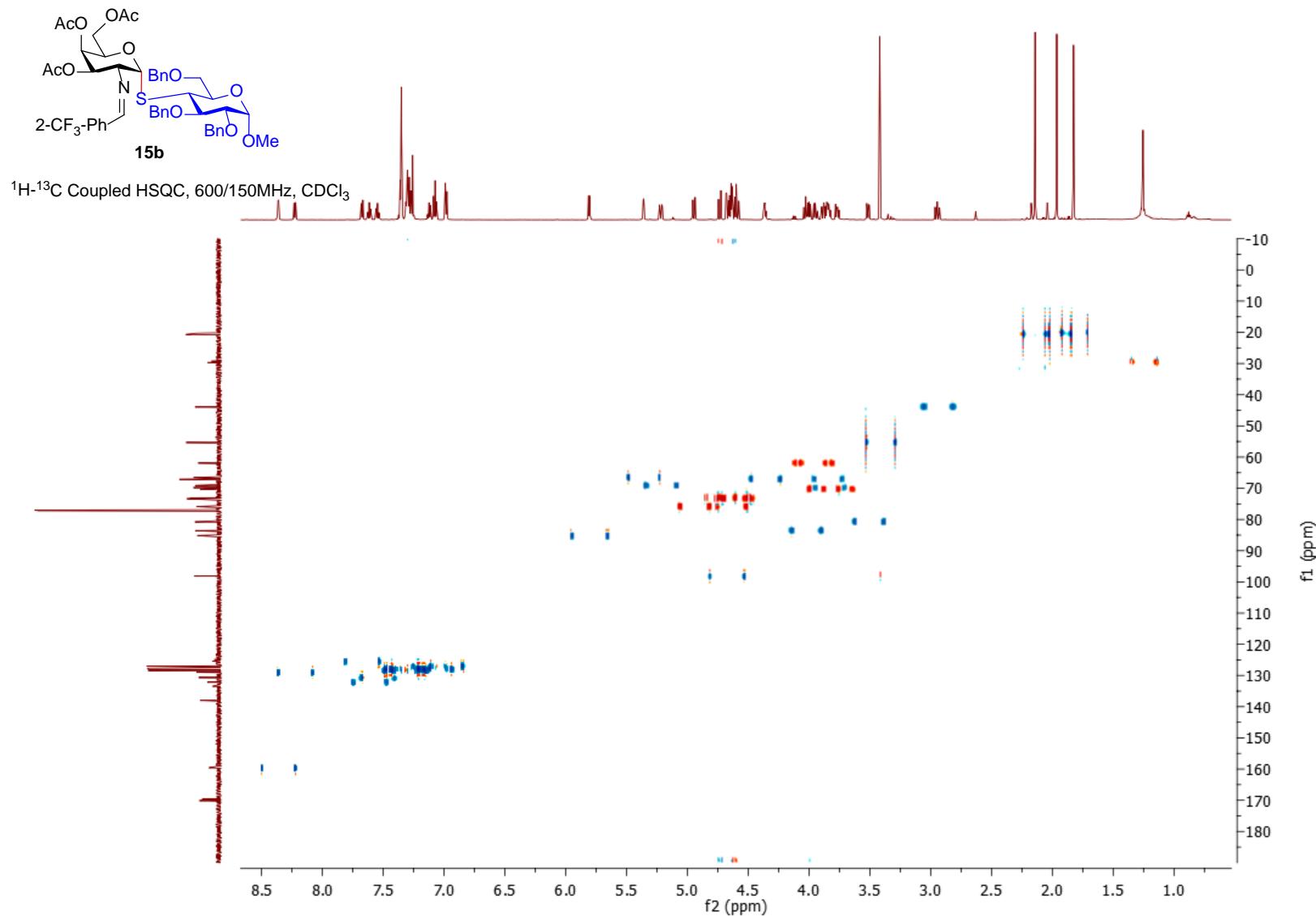


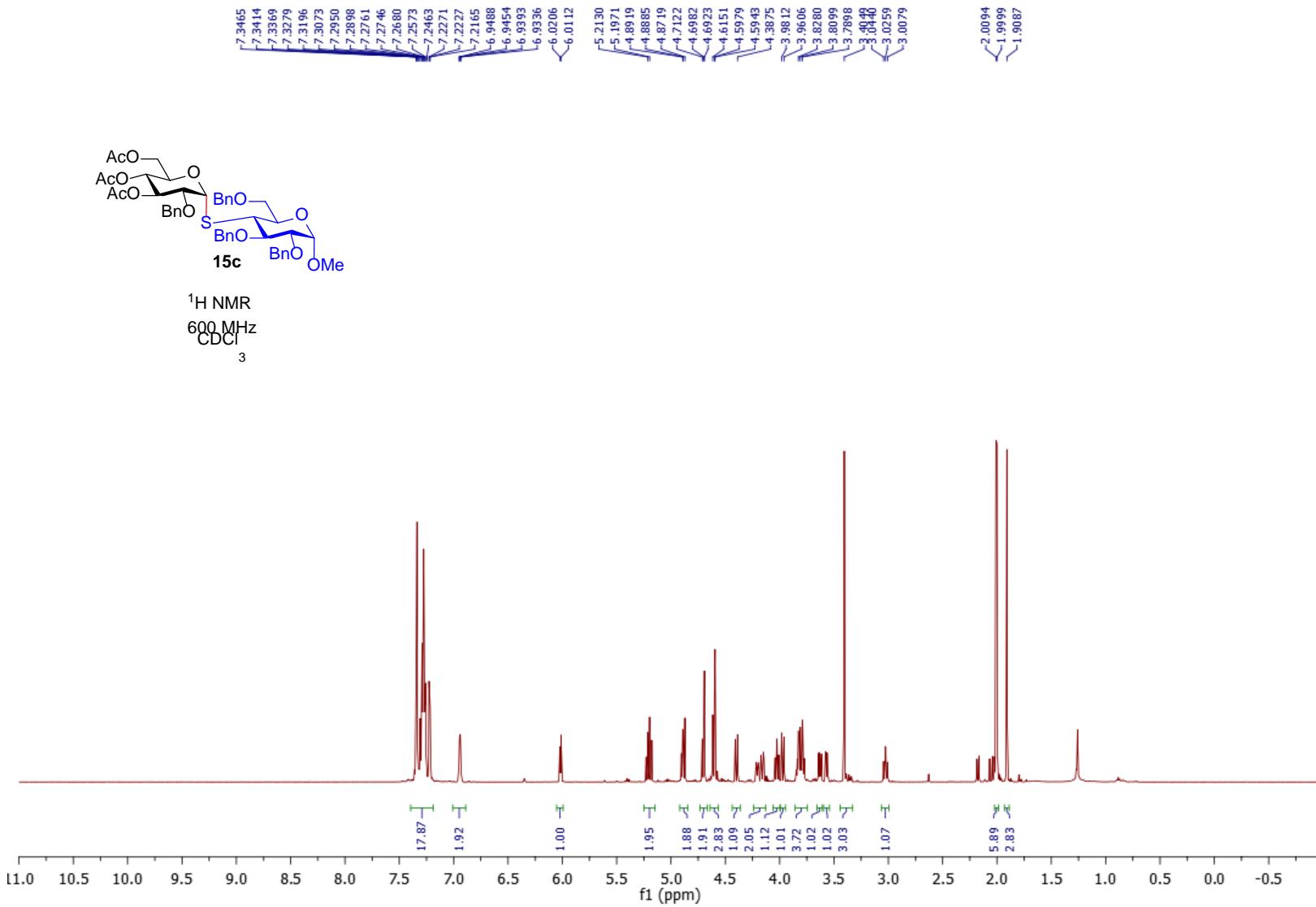


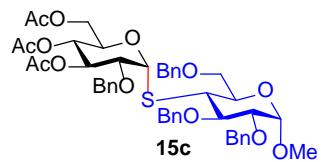




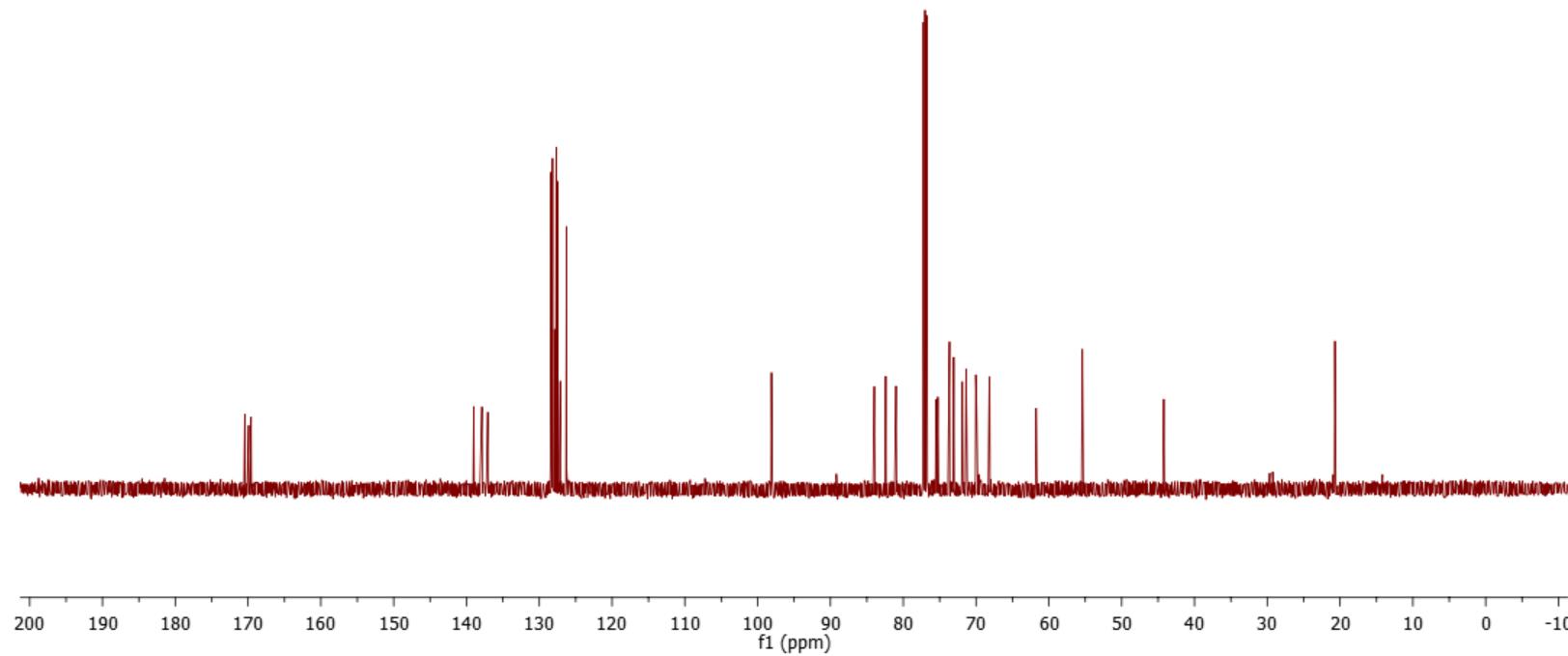


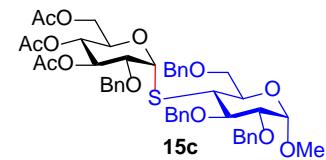




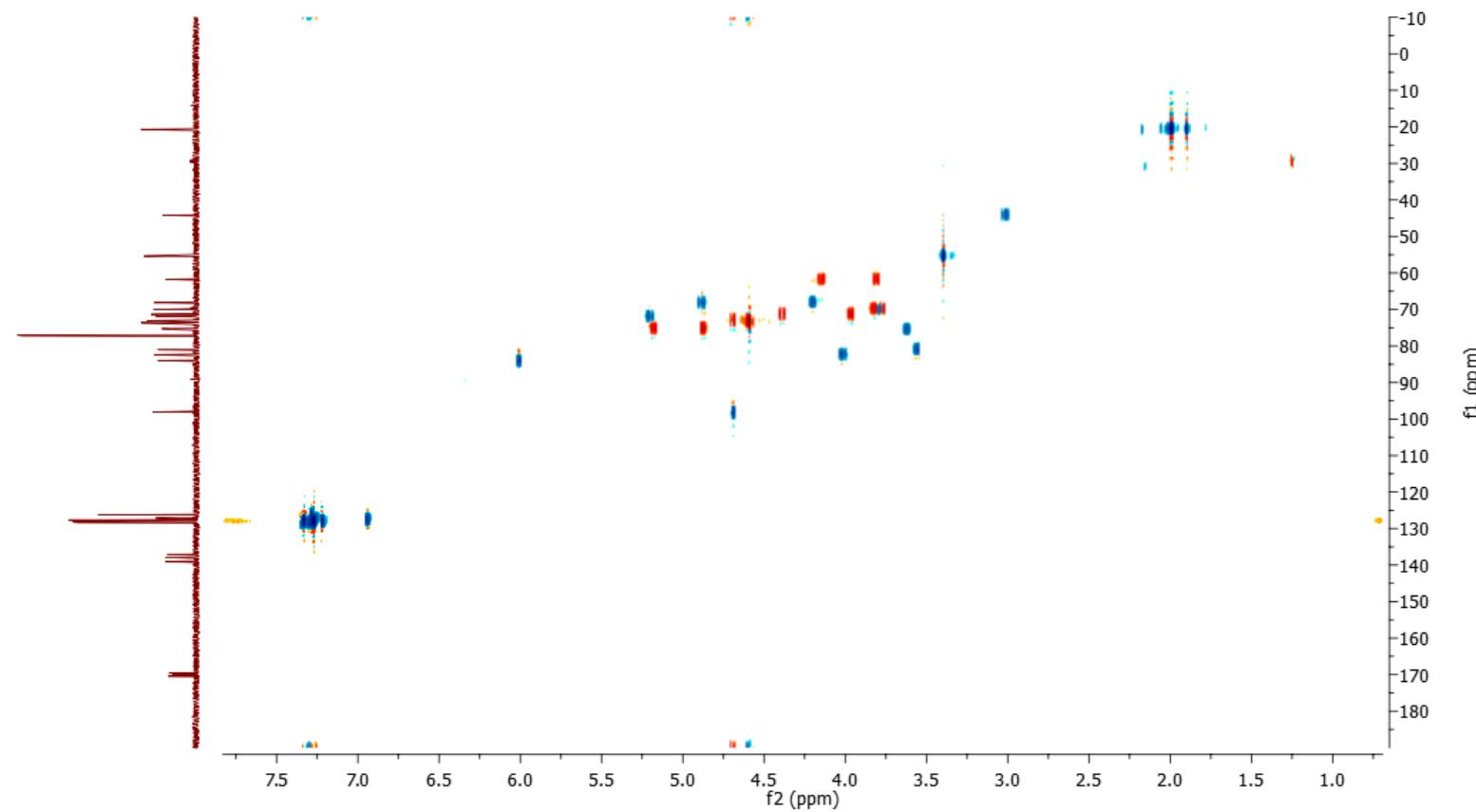


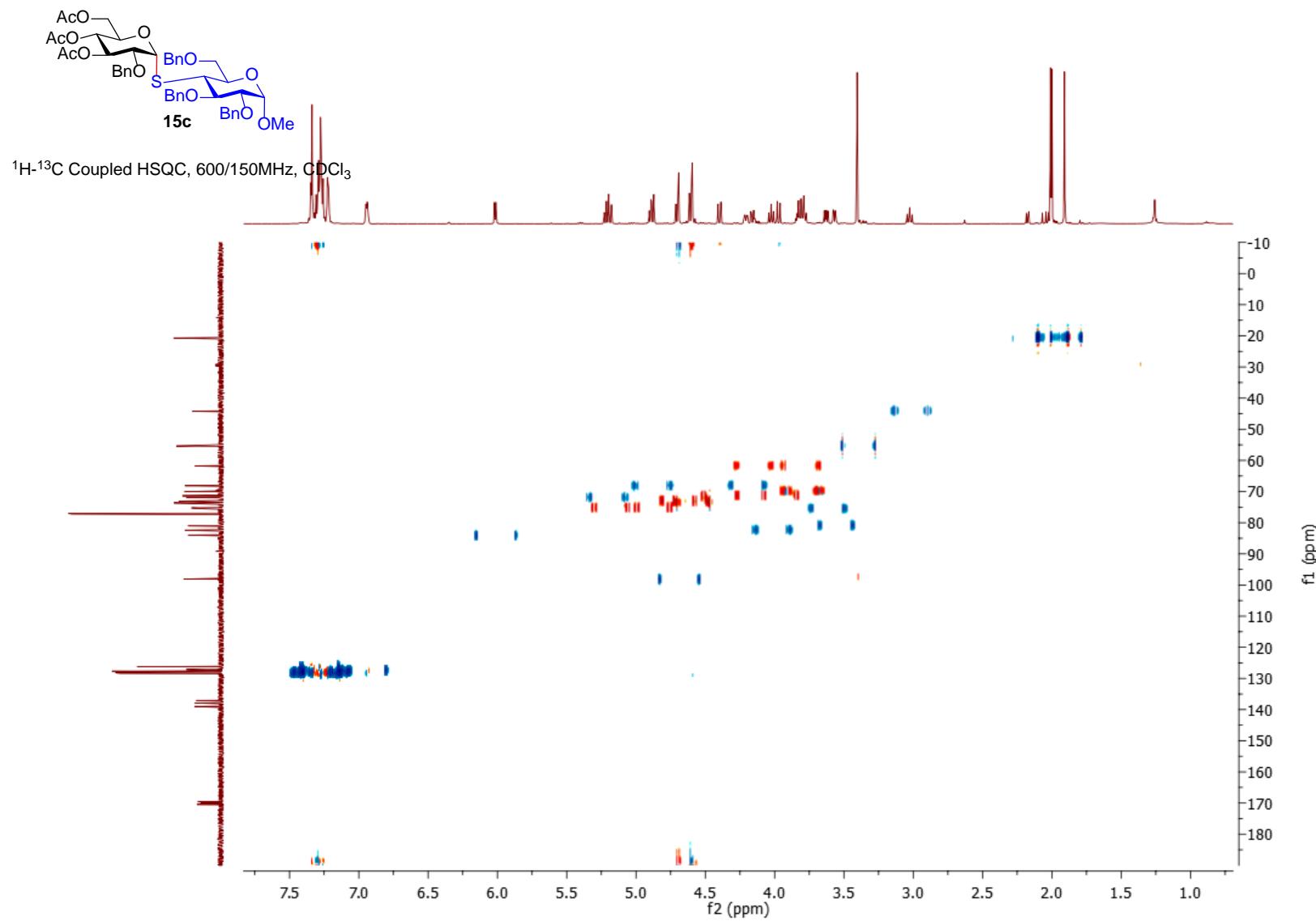
^{13}C NMR, 150 MHz, CDCl_3

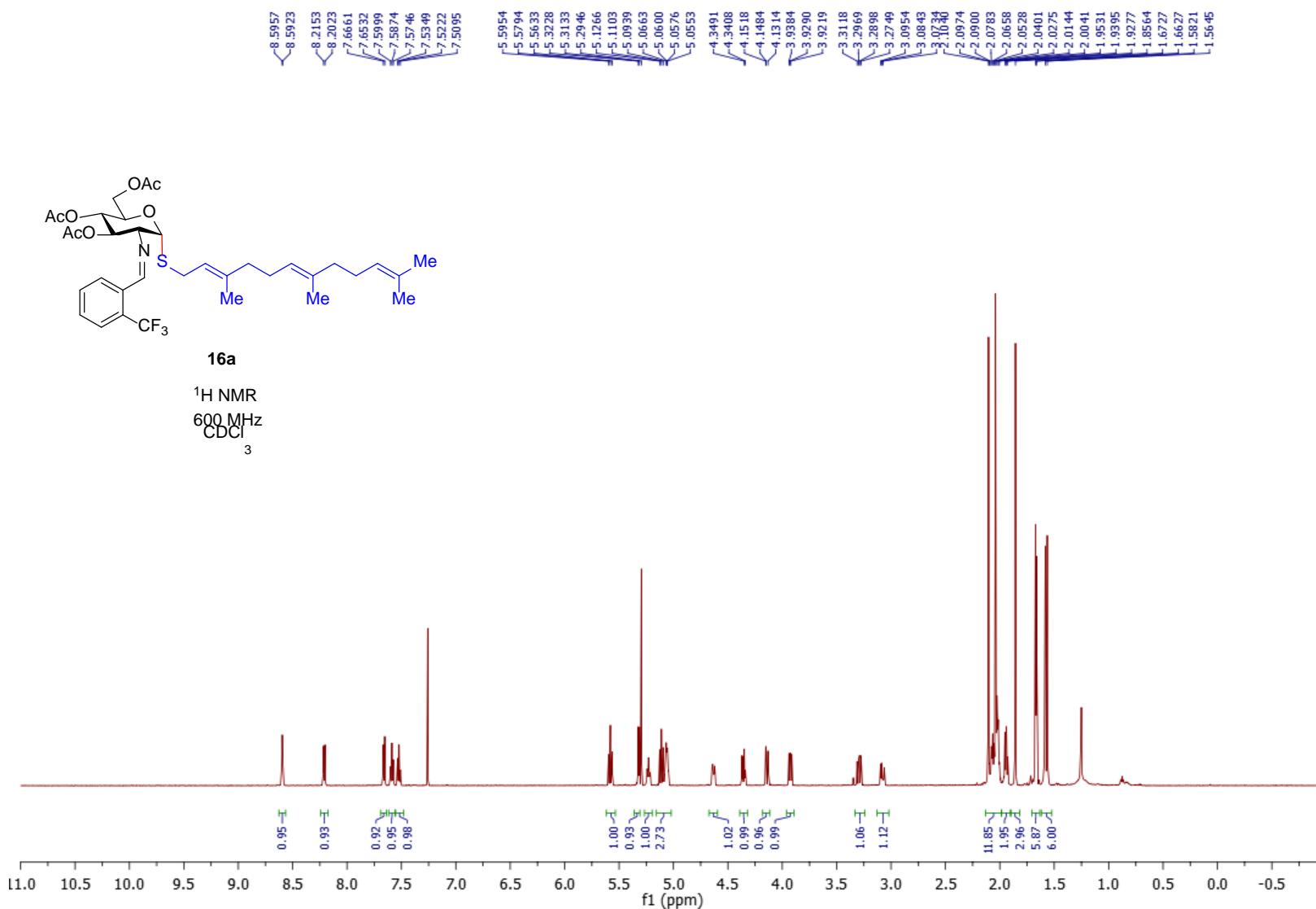


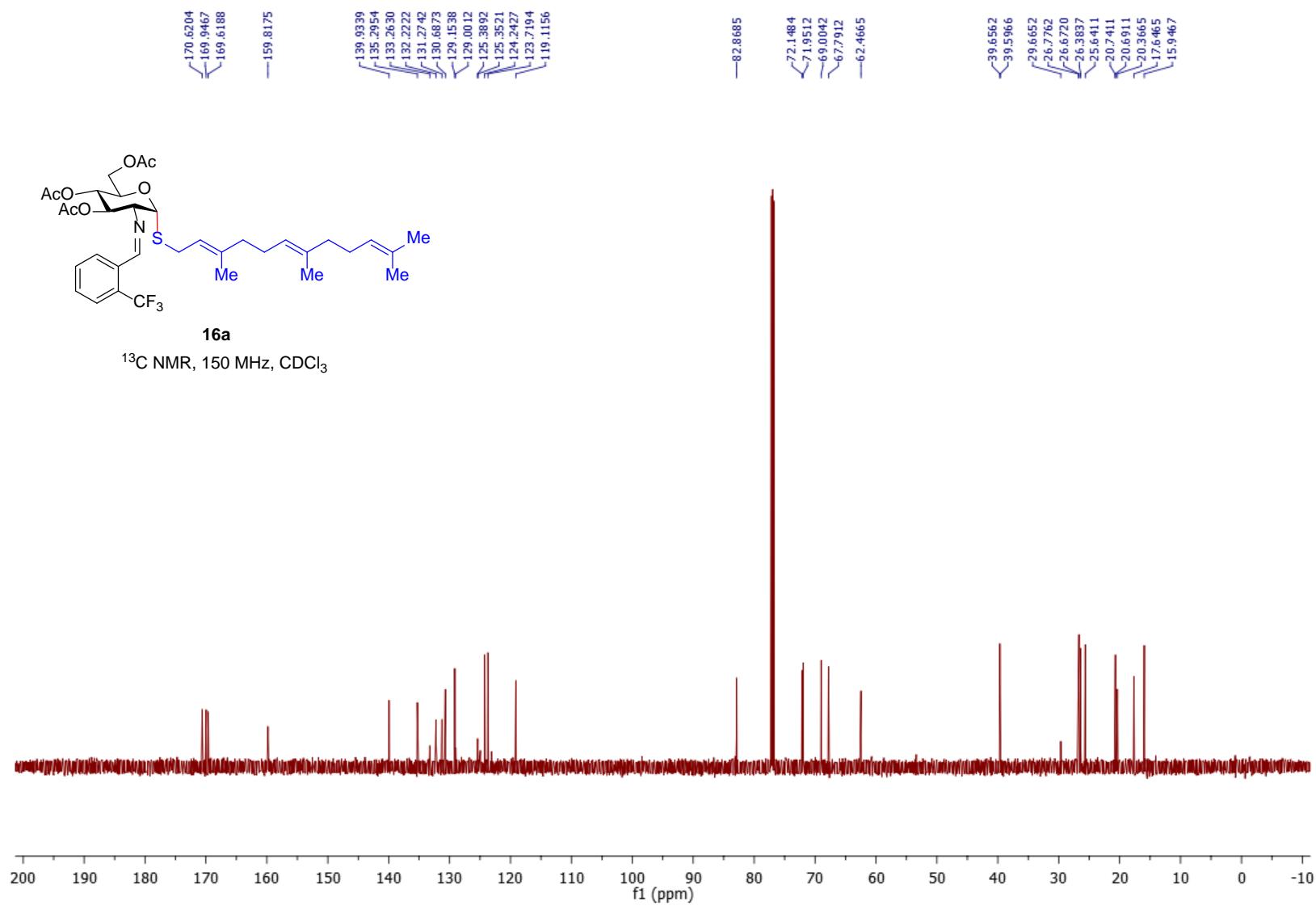


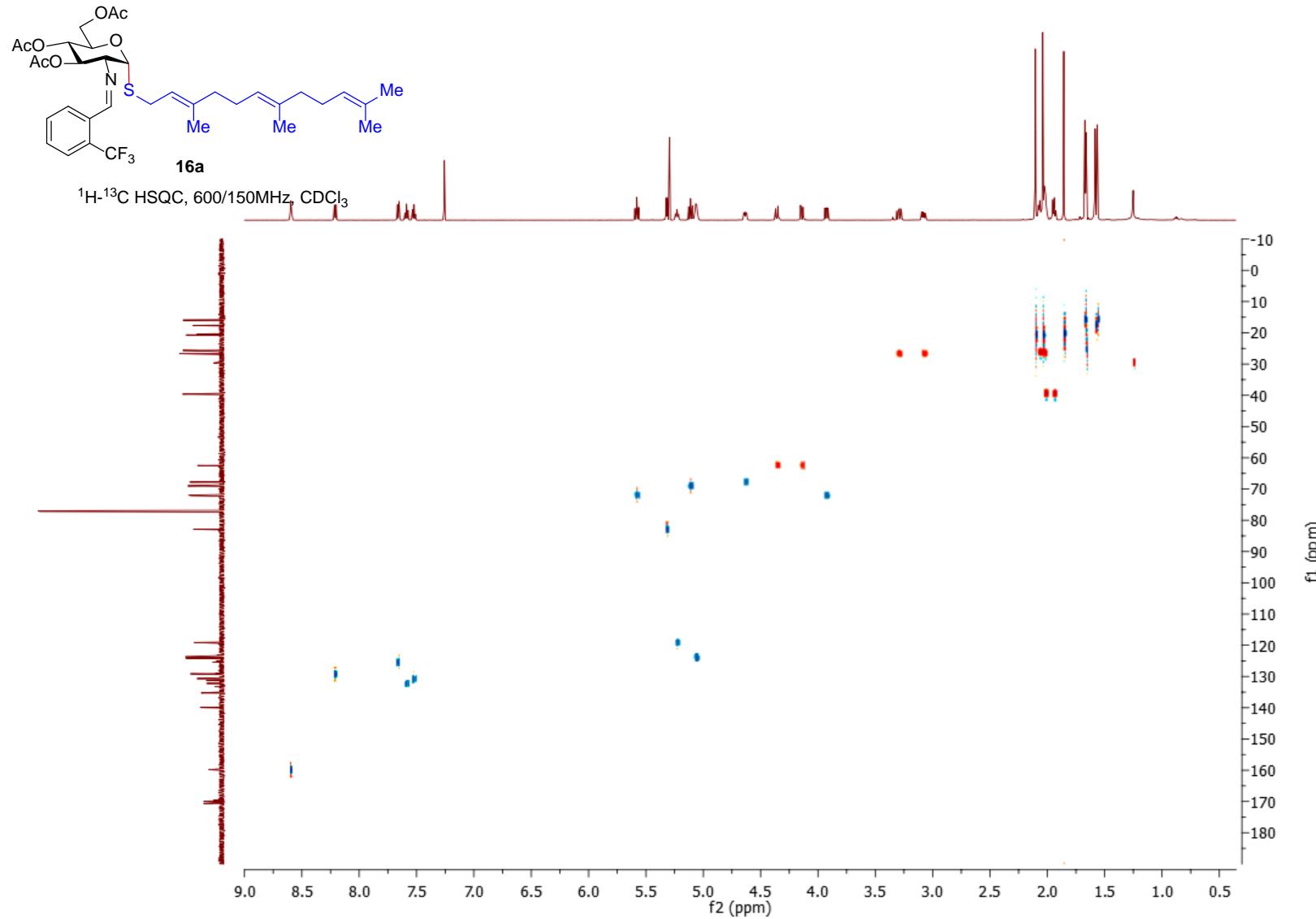
^1H - ^{13}C HSQC, 600/150MHz, CDCl_3

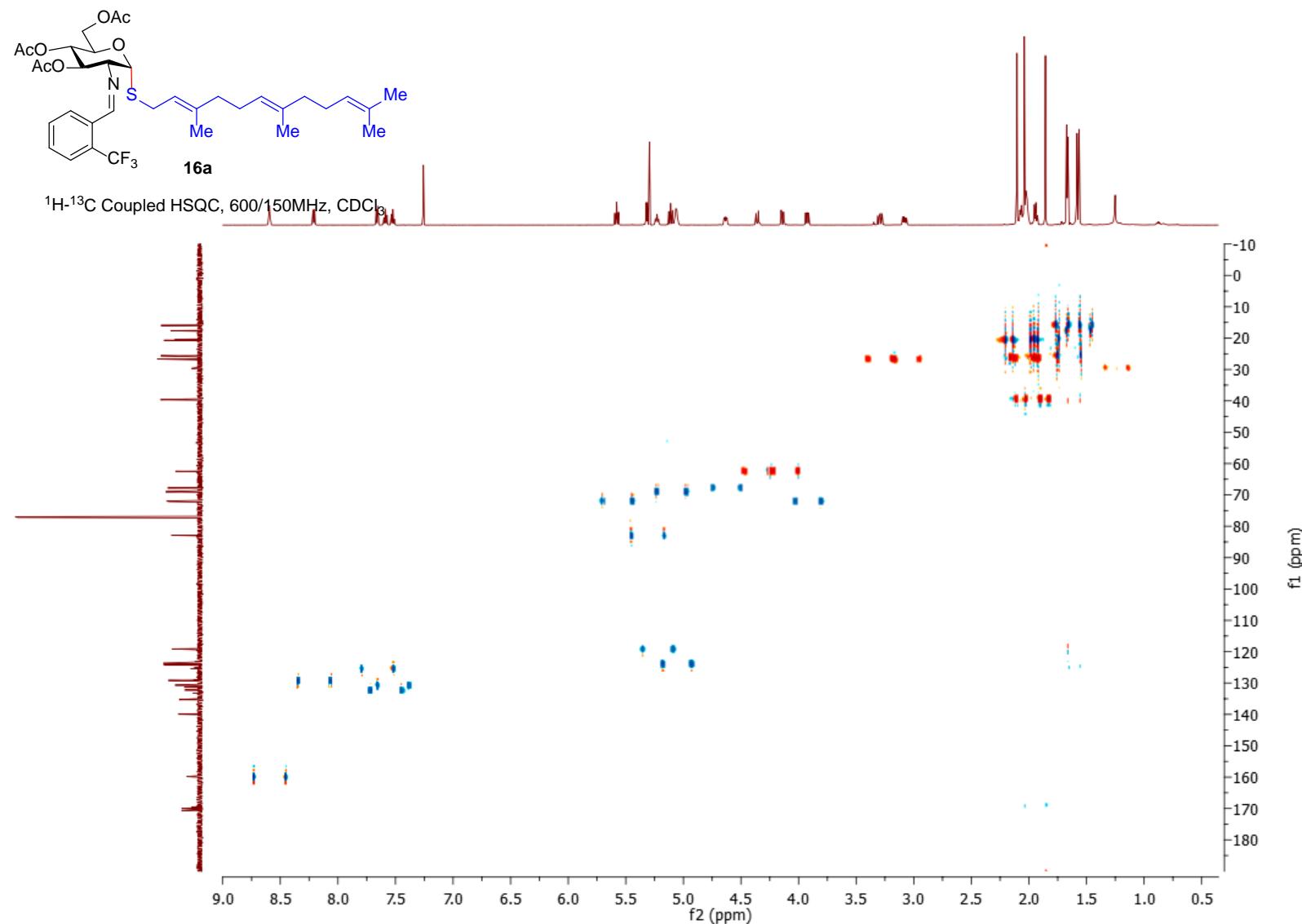


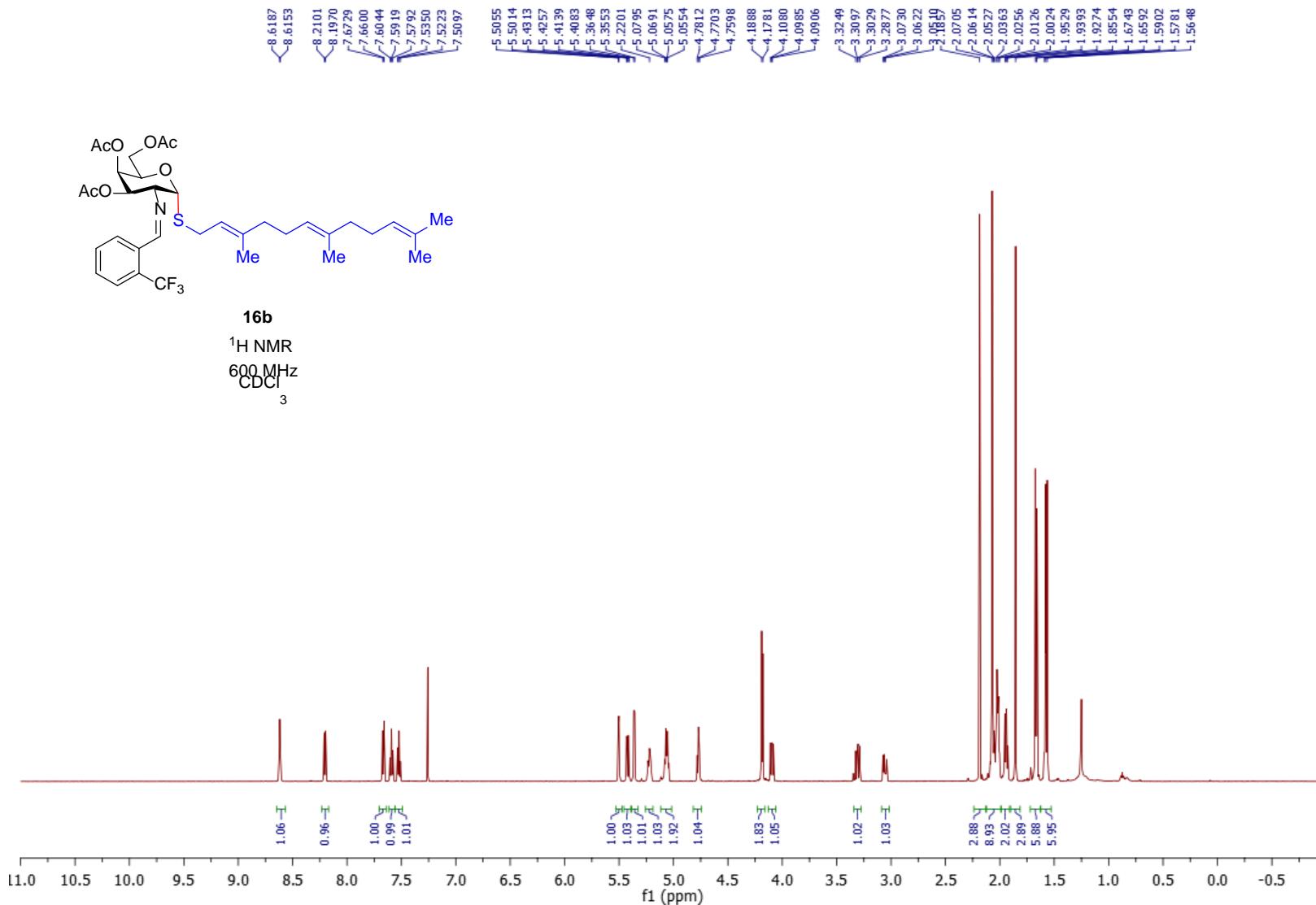


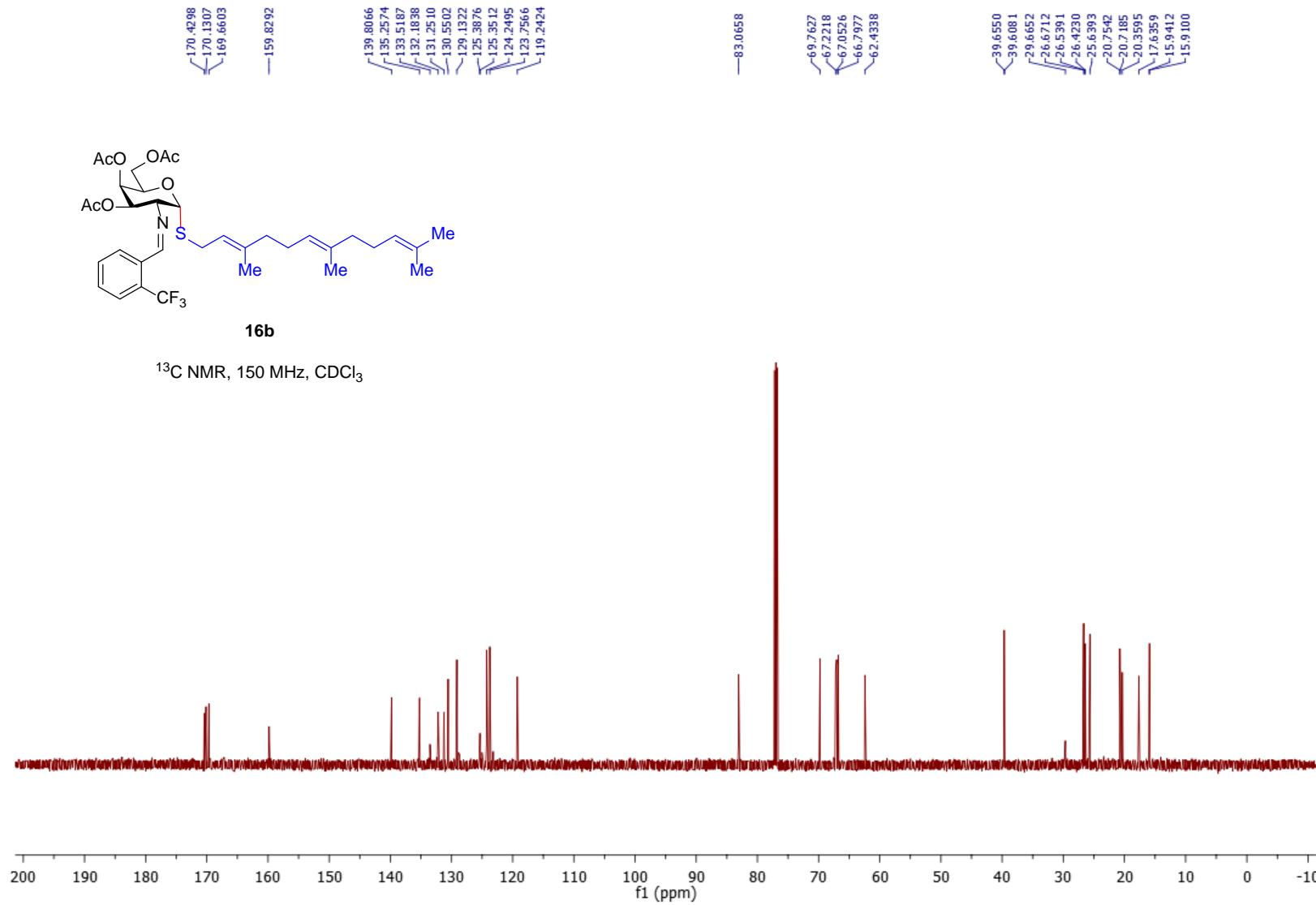


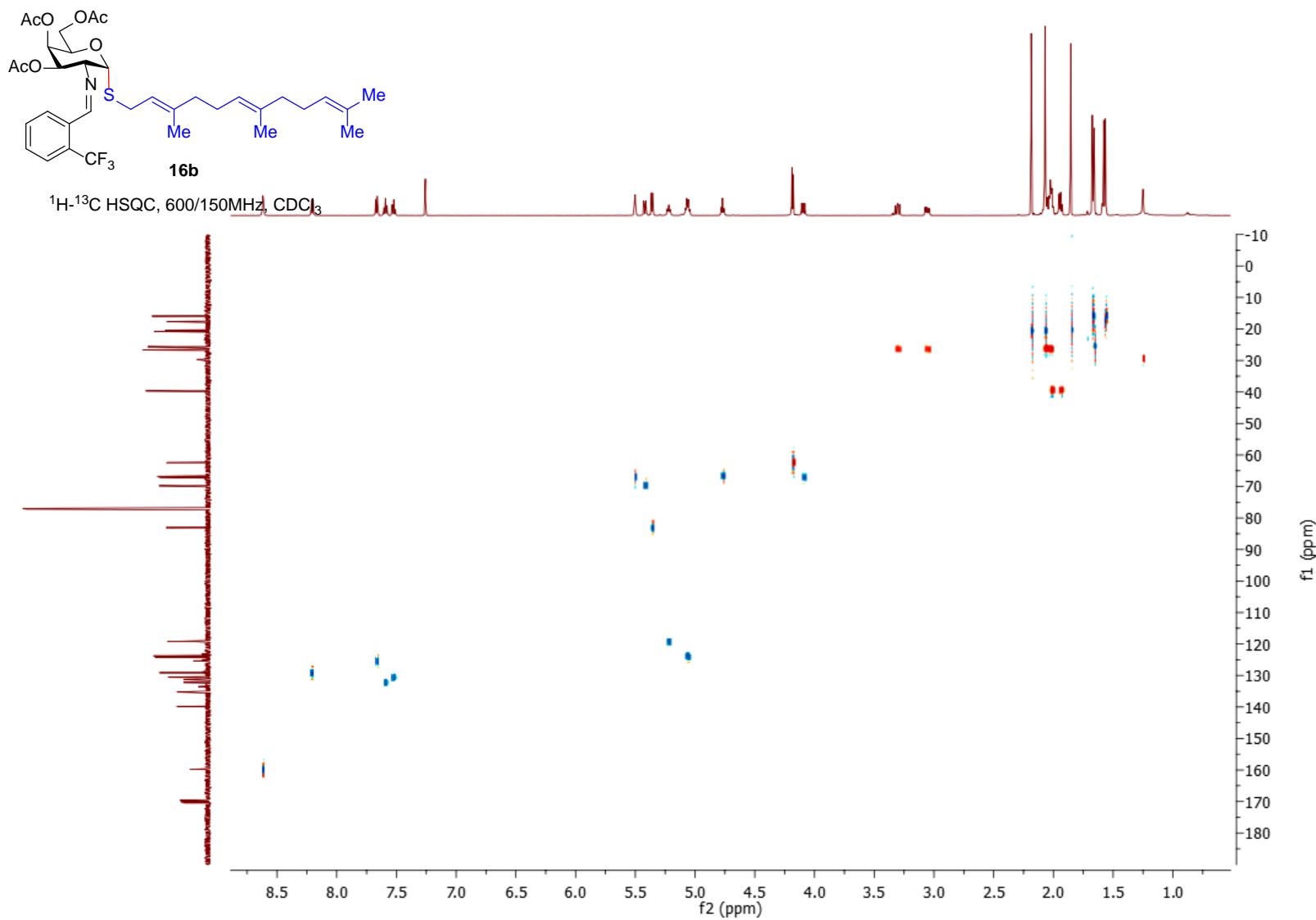


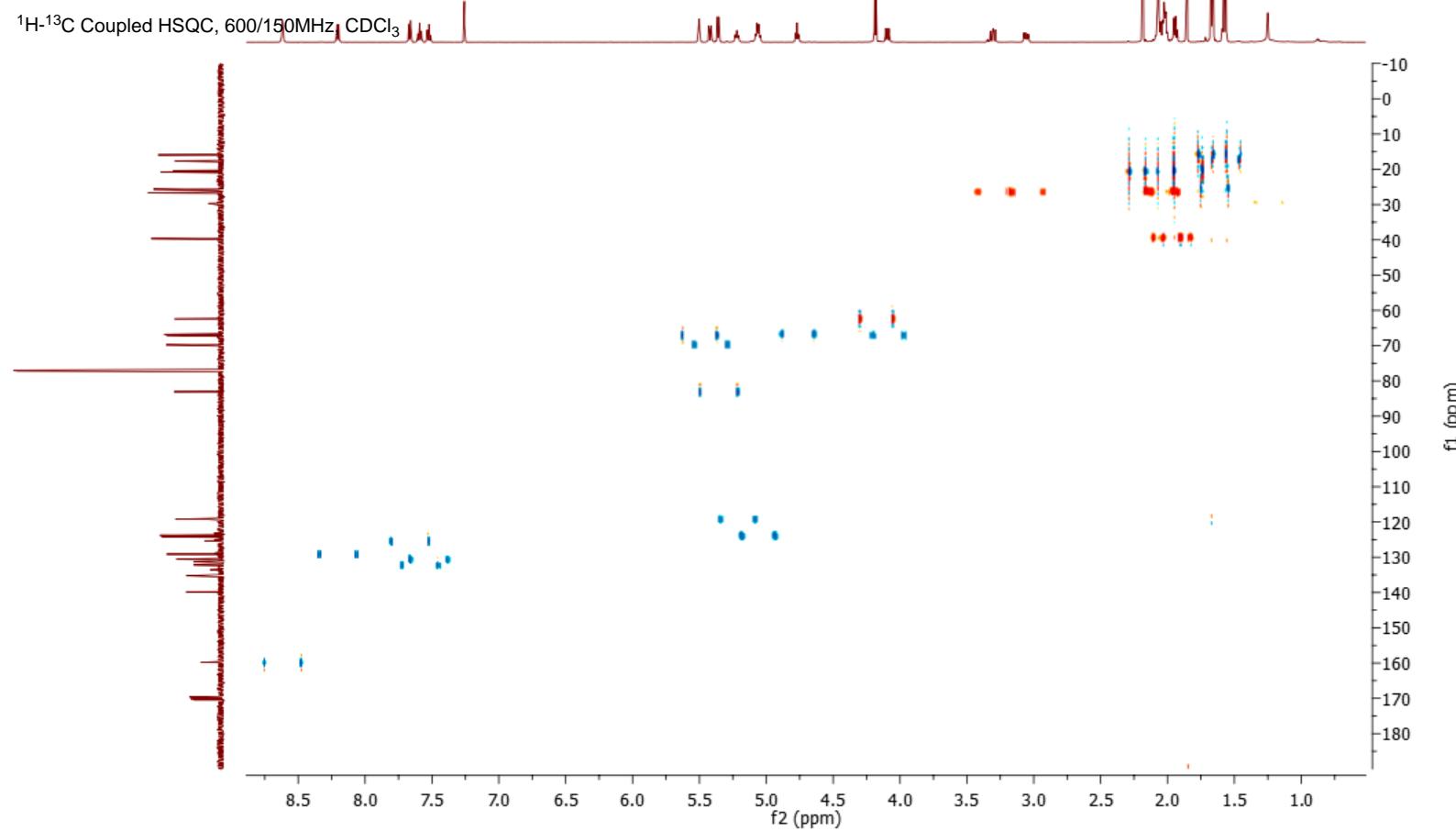
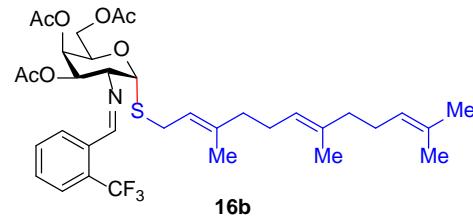


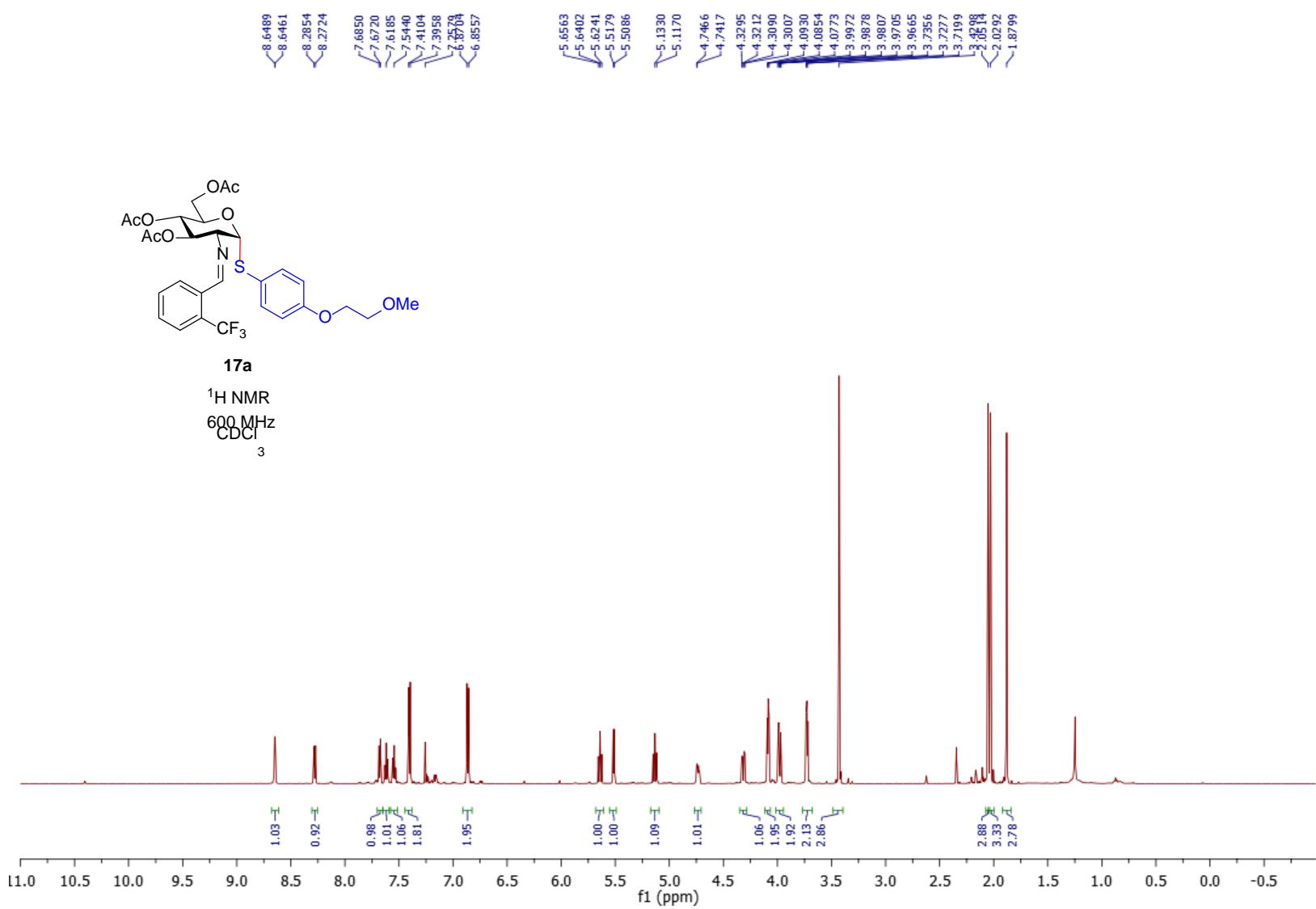


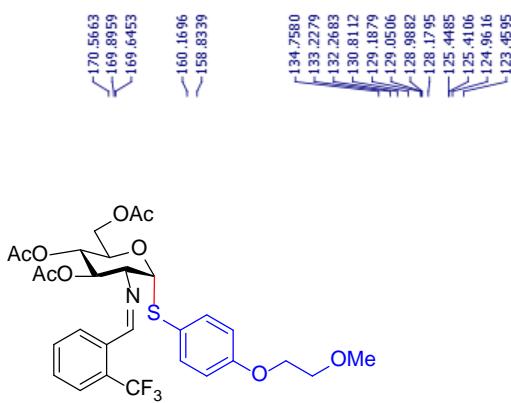




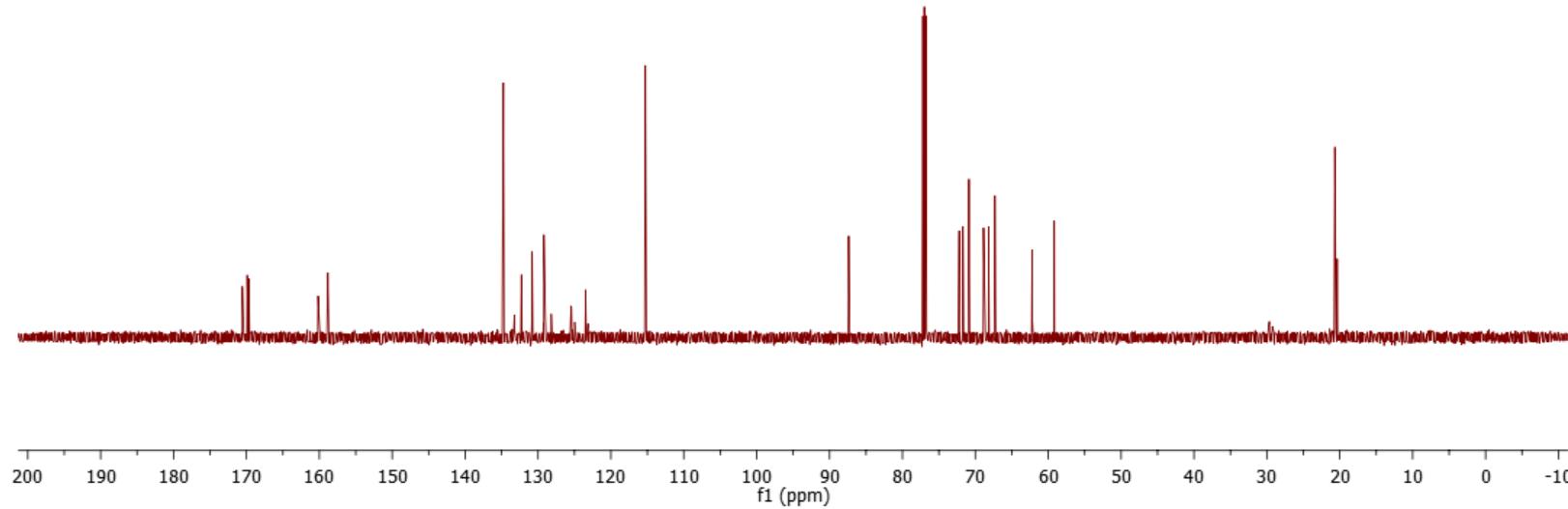


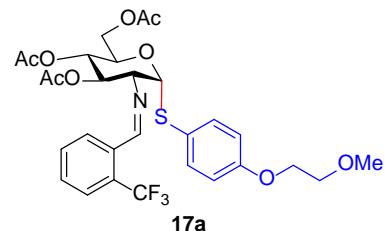




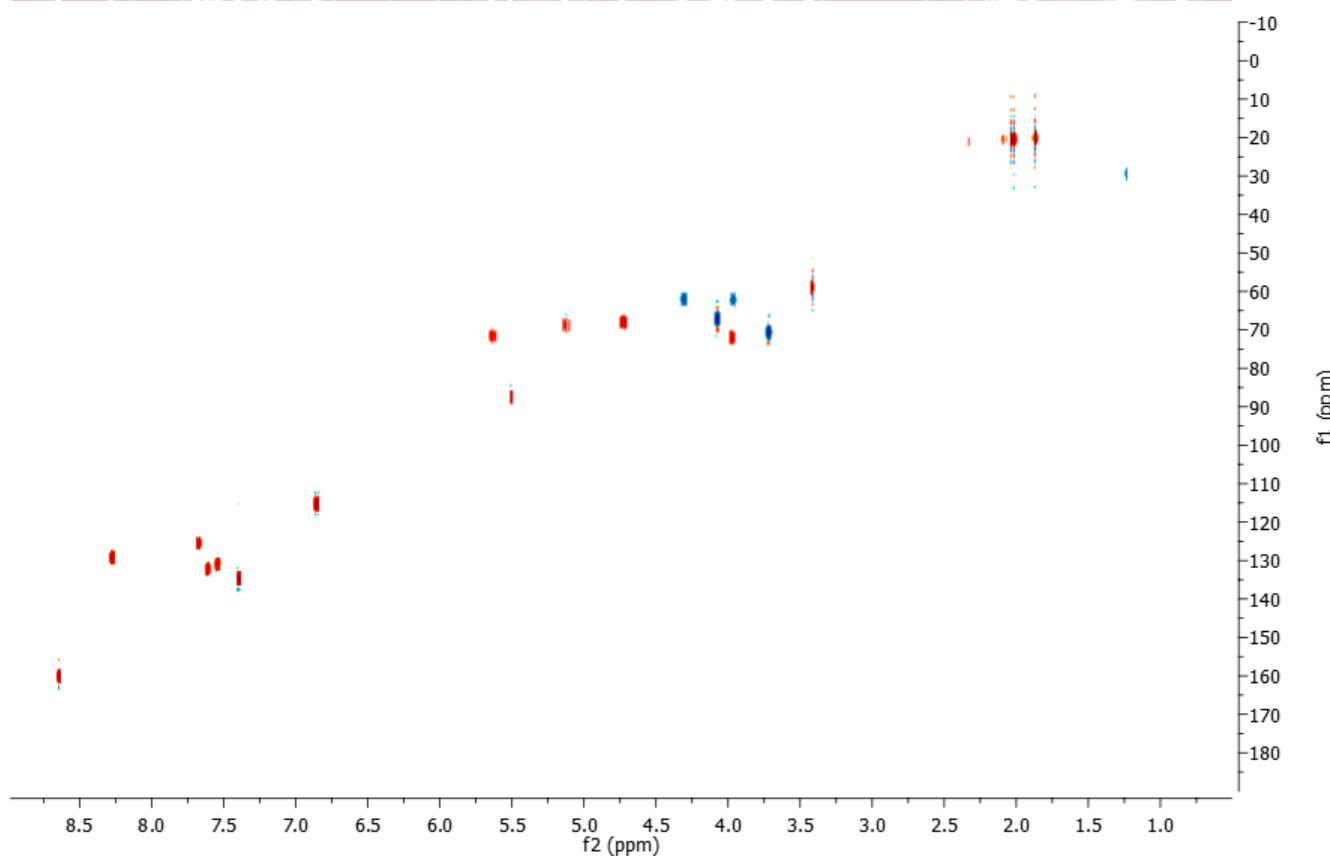


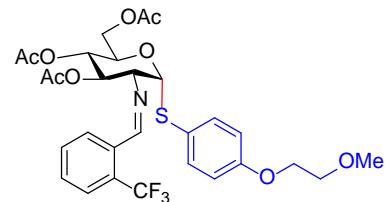
¹³C NMR, 150 MHz, CDCl₃



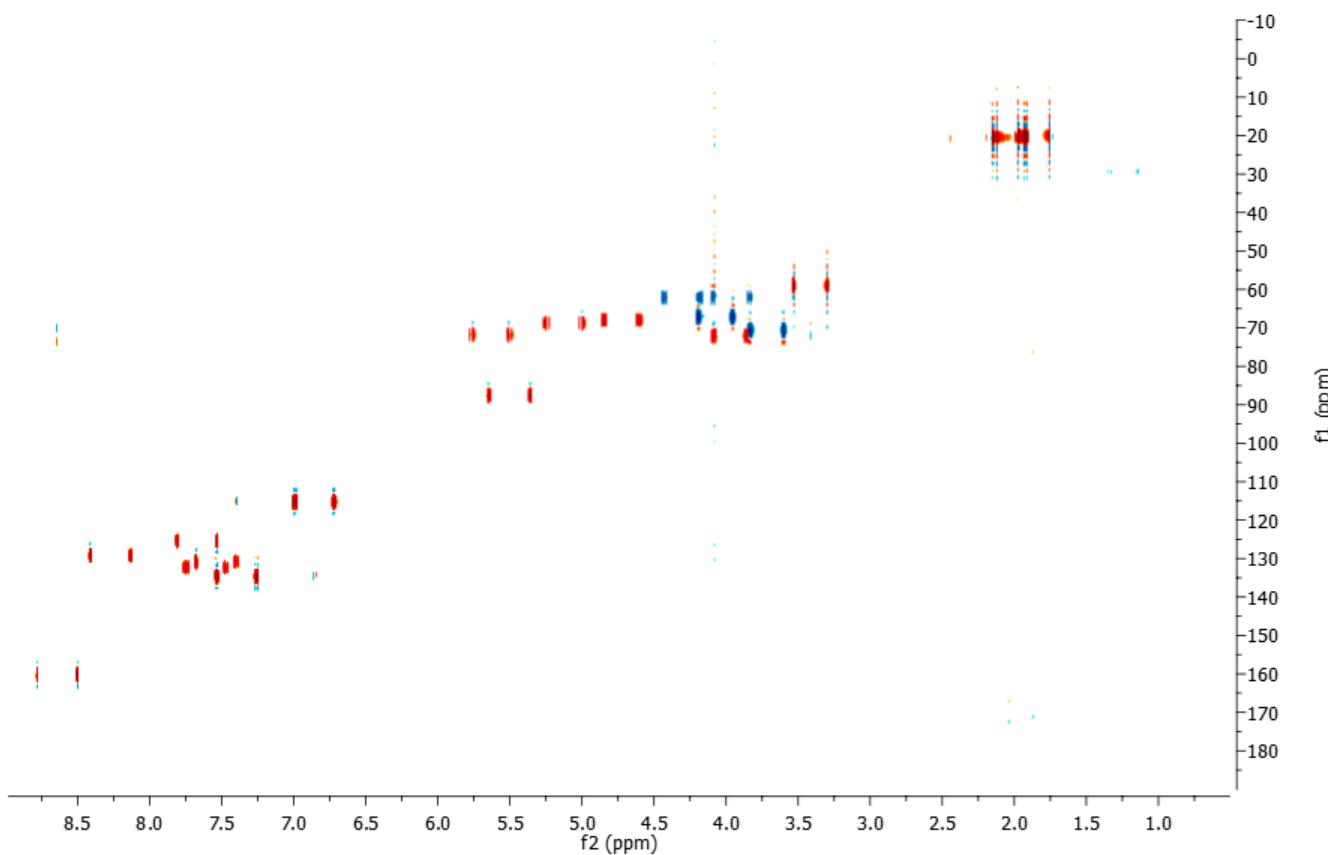


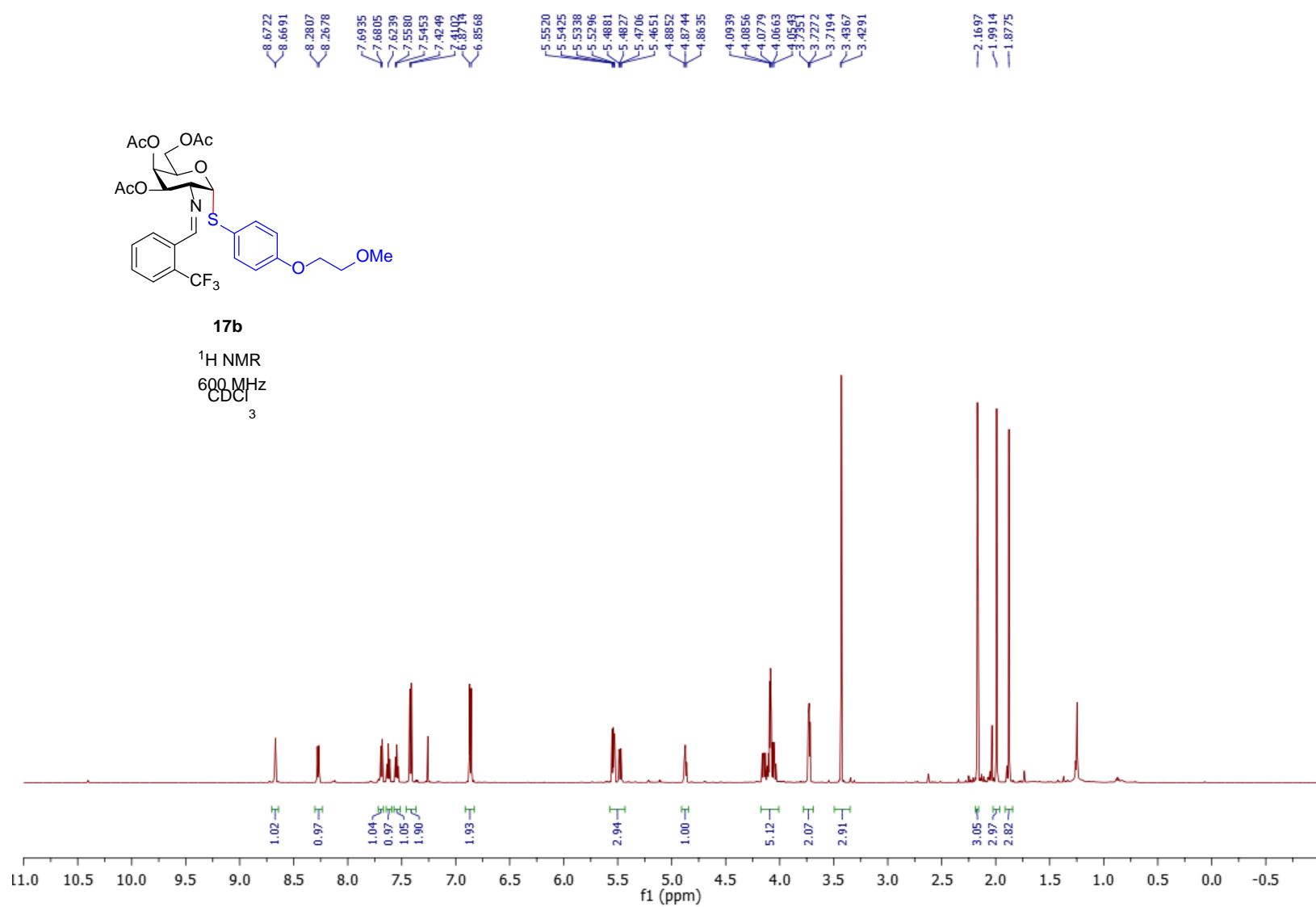
^1H - ^{13}C HSQC, 600/150MHz, CDCl_3

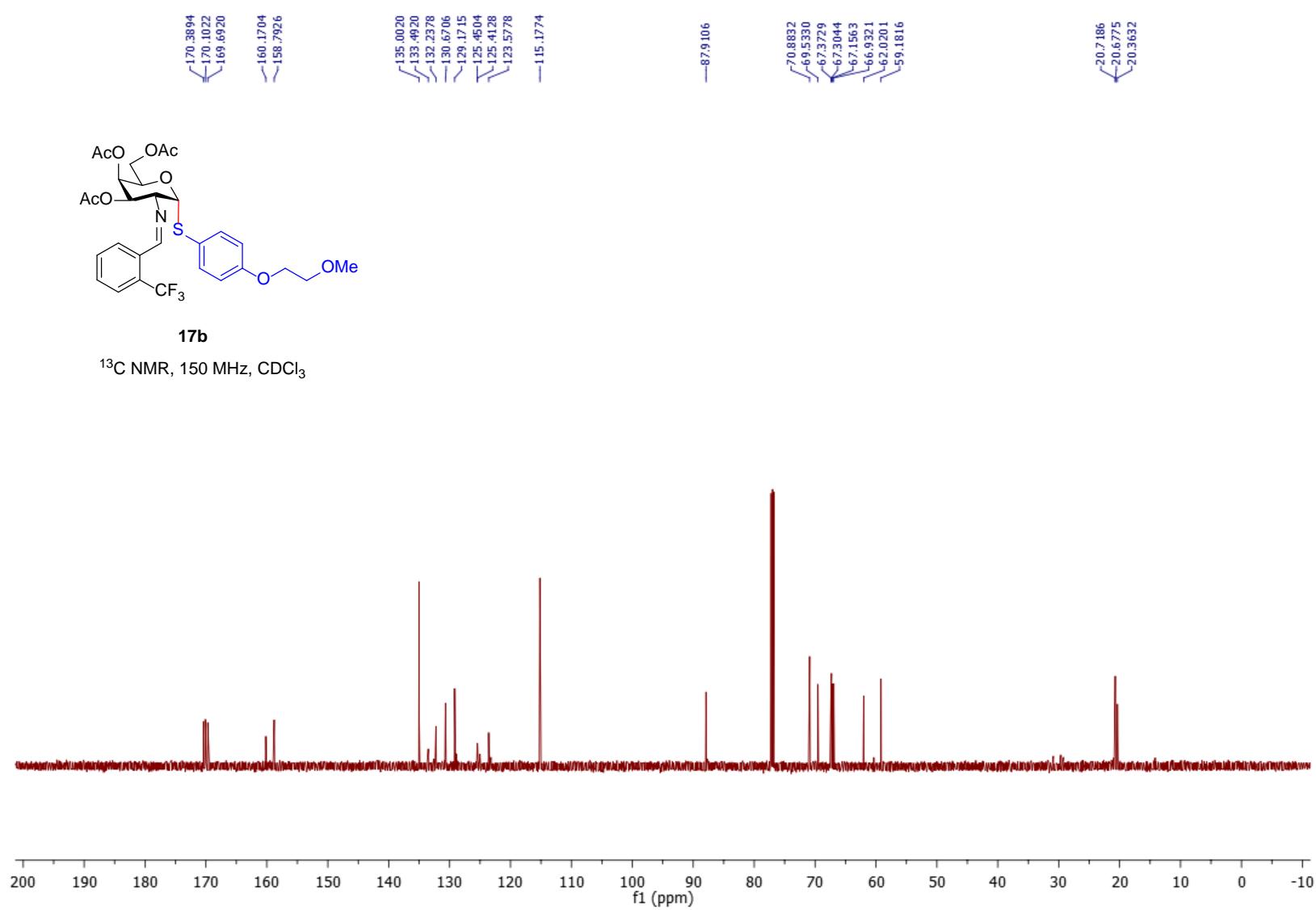


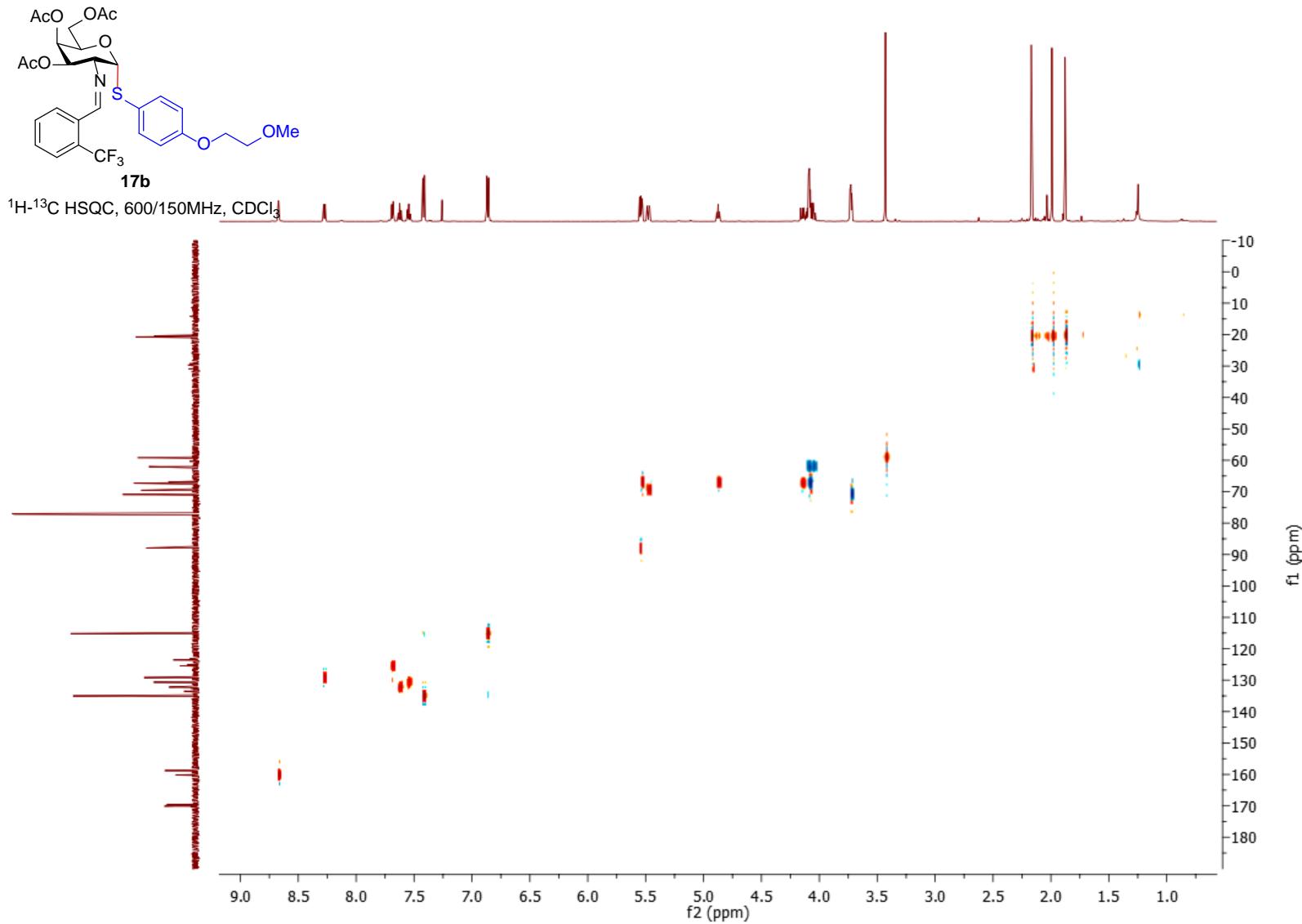


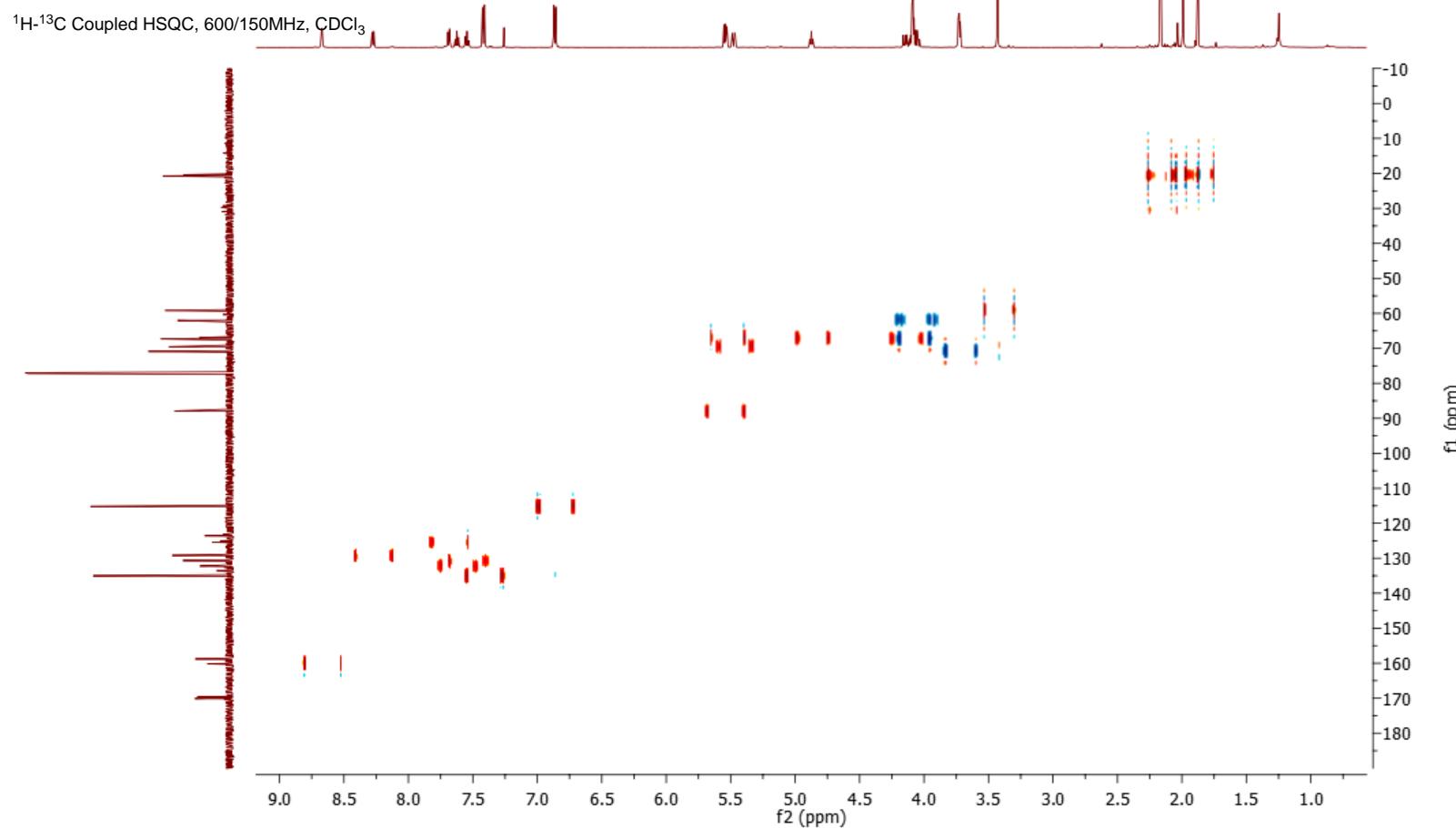
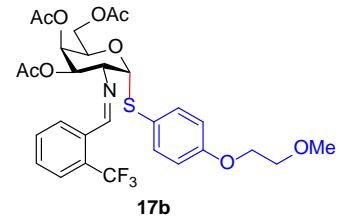
^1H - ^{13}C Coupled HSQC, 600/150MHz, CDCl_3

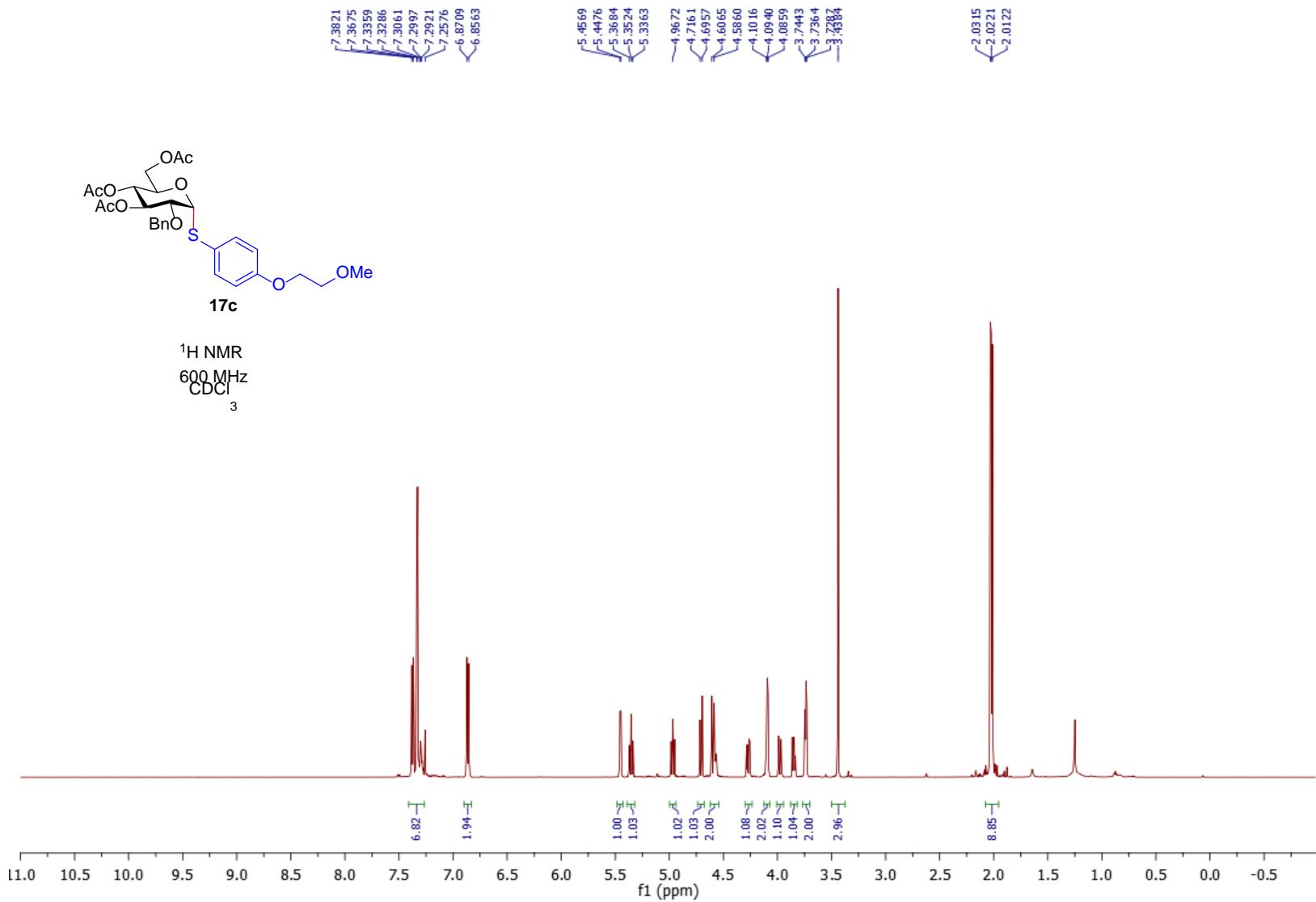


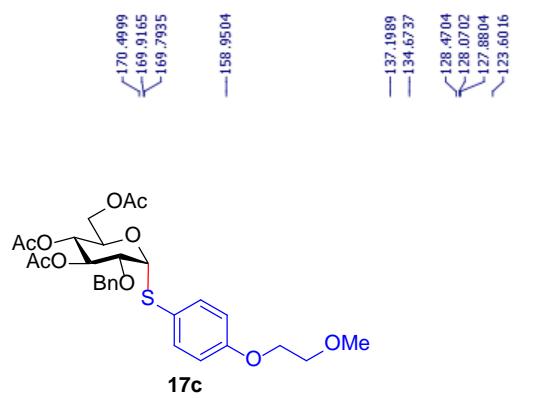




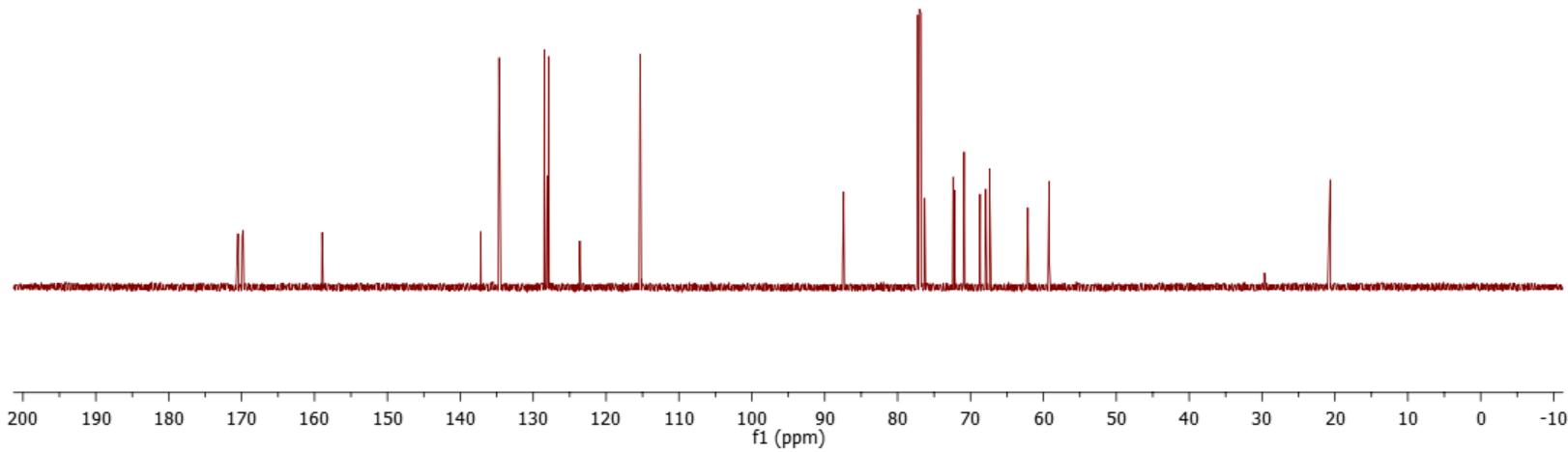


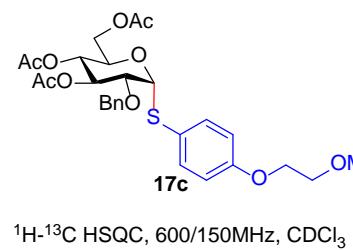






^{13}C NMR, 150 MHz, CDCl_3





^1H - ^{13}C HSQC, 600/150MHz, CDCl_3

