Electronic Supplementary Information

Screening of metal ions and organocatalysts on solid supportcoupled DNA oligonucleotides guides design of DNA-encoded reactions

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General methods and materials

Unless otherwise noted, chemicals were purchased from *abcr, Acros Organics, Alfa Aesar, Fisher Scientific, Merck, Sigma Aldrich, TCI* and *VWR* and were used as provided without further purifications. Dry solvents (CH₂Cl₂, DMF, MeOH, ACN, EtOH, THF, toluene) were used as commercially available.

5'-Aminolinker-modified DNA oligonucleotides coupled to controlled pore glass solid support (CPG, 1000 Å porosity) were synthesized by *IBA* (Göttingen, Germany). Oligonucleotide-small molecule conjugates coupled to CPG were filtered and washed through synthesis columns using a vacuum manifold (*Vac-Man*[®]) from *Promega*.

Semi-preparative ion pair RP-HPLC. Compound purification was performed on a *Shimadzu Prominence* HPLC System equipped with a C₁₈ stationary phase (*Phenomenex*, Gemini, 5 μ m, C₁₈, 110 Å, 100 x 4.6 mm). A gradient from 100 mM aqueous triethylammonium acetate (pH = 8.0, eluent A) to MeOH (eluent B) was used at a flow rate of 5 mL/min. Fractions containing the desired product were pooled and concentrated.

Method: Step gradient of 20 % to 70 % B within 13 min, then 70 % to 100 % B within 1 min followed by 100 % B for 3 min using 100 mM aqueous triethylammonium acetate (pH = 8.0, eluent A) and MeOH (eluent B) at a flow rate of 5 mL/min.

Analytical RP-HPLC. HPLC analysis was performed on an *Agilent 1100 series* chromatograph equipped with 1100 Quaternary Pump (*G1311A*), a 1100 Multi-Wavelength Detector (*G1365B*) and an *Agilent Eclipse Plus* C_{18} (4.6 x 100 mm, 3.5 µm) column. The conversion and purity of DNA conjugates were determined by integration of peaks recorded at 254 nm wavelength.

Method-I: Step gradient of 10 % to 60 % B within 10 min, then 60 % to 100 % B within 2 min followed by 100 % B for 2 min using 10 mM aqueous triethylammonium acetate (pH = 8.0, eluent A) and MeOH (eluent B) at a flow rate of 0.6 mL/min.

Method-II: Step gradient of 10 % to 70 % B within 10 min, then 70 % to 100 % B within 2 min followed by 100 % B for 2 min using 10 mM aqueous triethylammonium acetate (pH = 8.0, eluent A) and MeOH (eluent B) at a flow rate of 0.6 mL/min.

Method-III: A linear gradient of 10 % to 100 % B within 10 min followed by 100 % B for 4 min using 10 mM aqueous triethylammoinum acetate (pH 8.0, eluent A) and MeOH (eluent B) at a flow rate of 0.6 mL/min.

MALDI-TOF. Mass analysis was performed on a MALDI TOF/TOF MS from *Bruker Daltonics* using 2',4',6'-trihydroxyacetophenone (THAP) matrix (*Dichrom*).

Representative procedures

DMT-deprotection and capping of solid support-coupled hexT (RP_01)

The DMT-protecting group of 5'-amino linker modified hexT-DNA strand coupled to 1000 Å controlled pore glass (CPG) solid support (1 µmol, ca. 36 mg) was cleaved by addition of 3% trichloroacetic acid in CH₂Cl₂ (3x 200 µL) for 1 min (yellow to orange color indicated successful removal of protecting group). The CPG was washed with DMF (3x 200 µL), MeOH (3x 200 µL), ACN (3x 200 µL) and CH₂Cl₂ (3x 200 µL). Deprotected CPG-coupled oligonucleotide was treated with 200 µL of capping solution (1:1 mixture of THF/methylimidazole (9:1, vol:vol) and THF/pyridine/acetic acid anhydride (8:1:1, vol/vol)) for 1 min. Finally, the CPG was washed three times with each 200 µL of DMF, MeOH, ACN and CH₂Cl₂ and dried *in vacuo* for 15 min. Capping was repeated three times.

To assess completion of deprotection and capping, a small portion of CPG (0.7 mg) was treated with 500 μ L of an AMA solution (AMA = aqueous ammonia (30%)/ aqueous methylamine (40%), 1:1, vol/vol) for 30 min at room temperature. Afterwards 20 μ L if 1 M Tris buffer (pH = 7.5) were added, the mixture was dried under reduced pressure (SpeedVac) and dissolved in 100 μ L of distilled water. The product was analyzed by Analytical RP-HPLC and MALDI-TOF-MS.

DMT-deprotection of solid support-coupled 10mer DNA strands (RP_02)

The DMT-protecting group of DNA strand coupled to 1000 Å controlled pore glass (CPG) solid support (1 μ mol, ~40 mg of 10mer DNA (TC-, ATC-, ATCG-Sequences)) was cleaved by addition of 3% trichloroacetic acid in CH₂Cl₂ (3x 200 μ L) for 1 min (yellow to orange color indicated successful removal of protecting group). The CPG was washed three times with each 200 μ L of DMF, MeOH, ACN and CH₂Cl₂ and dried *in vacuo* for 15 min.

Treatment of solid support-coupled DNA with metal salts (RP_03)

20 nmol of CPG-coupled oligonucleotide (ca. 0.7 mg solid phase) were treated with 200 equiv. of metal salt (4 μ mol) dissolved in 50 μ L dry solvent. The suspension was shaken at ambient temperature for 22 h. Afterwards the solvent was removed by filtration, and the CPG was washed three times with each 200 μ L of 0.1 M EDTA solution, 0.1 M MgCl₂ solution, water, DMF, MeOH, ACN and CH₂Cl₂ and dried *in vacuo*.

<u>Cleavage and analysis:</u> DNA was deprotected and cleaved from CPG by shaking with 500 μ L of an AMA solution (AMA = aqueous ammonia (30%)/ aqueous methylamine (40%), 1:1, vol/vol)

for 4 h (30 min for hexT-Ac) at room temperature. Afterwards, 20 μ L of 1 M Tris buffer (pH = 7.5) were added, the mixture was dried under reduced pressure (SpeedVac) and the DNA was dissolved in 200 μ L distilled water. The product was analyzed by Analytical RP-HPLC and MALDI-TOF-MS.

Treatment of solid support-coupled DNA with organocatalysts (RP_04)

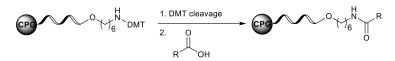
20 nmol of CPG-coupled oligonucleotide (ca. 0.7 mg of solid phase) were treated with 200 equiv. of organocatalyst (4 μ mol) dissolved in 50 μ L dry solvent. The suspension was shaken at ambient temperature for 22 h. Afterwards, the solvent was removed by filtration, the CPG was washed three times with each 0.1 M MgCl₂ solution, water, DMF, MeOH, ACN and CH₂Cl₂ and dried *in vacuo*.

<u>Cleavage and analysis:</u> DNA was deprotected and cleaved from CPG by shaking in 500 μ L of an AMA solution (AMA = aqueous ammonia (30%)/ aqueous methylamine (40%), 1:1, vol/vol) for 4 h (30 min for hexT-Ac) at room temperature. Afterwards 20 μ L of 1 M Tris buffer (pH = 7.5) were added, the mixture was dried under reduced pressure (SpeedVac) and DNA was dissolved in 200 μ L distilled water. The product was analyzed by Analytical RP-HPLC and MALDI-TOF-MS.

Treatment of solid support-coupled DNA with acids (RP_05)

20 nmol of CPG-coupled oligonucleotide (ca. 0.7 mg of solid phase) were treated with 50 μ L acid. The suspension was shaken at ambient temperature for 22 h. Afterwards solution was removed by filtration, and the CPG was washed three times with each 200 μ L of 0.1 M MgCl₂ solution, water, DMF, MeOH, ACN and CH₂Cl₂ and dried *in vacuo*.

<u>Cleavage and analysis:</u> DNA was deprotected and cleaved from CPG by shaking in 500 μ L of an AMA solution (AMA = aqueous ammonia (30%)/ aqueous methylamine (40%), 1:1, vol/vol) for 4 h (30 min for hexT-Ac) at room temperature. Afterwards 20 μ L if 1 M Tris buffer (pH = 7.5) were added, the mixture was dried under reduced pressure (SpeedVac) and dissolved in 200 μ L of distilled water. The product was analyzed by Analytical RP-HPLC and MALDI-TOF-MS.



<u>Step 1:</u> The DMT-protecting group of CPG-coupled oligonucleotide (250 nmol, ca. 10 mg of solid phase material) was removed by addition of 200 μ L 3 % trichloroacetic acid in CH₂Cl₂ for 1 min. Orange coloring of the solution indicated successful removal of protecting group. The deprotection was repeated 3-5 times until no further coloring of the solution was observed. The CPG-coupled deprotected DNA was washed three times with each 200 μ L of 1 % TEA in ACN, DMF, MeOH, ACN and CH₂Cl₂ and dried *in vacuo*.

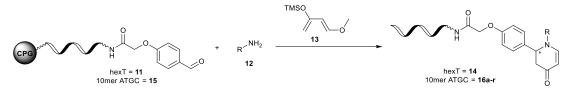
<u>Step 2:</u>

CPG-coupled oligonucleotide, carboxylic acid and HATU were dried *in vacuo* for 15 min. Stock solutions of all reactants in dry DMF were prepared before the reaction was started. To the solution of carboxylic acid (25 µmol, 100 equiv.) in 75 µL dry DMF, HATU (25 µmol, 100 equiv.) dissolved in 75 µL dry DMF and DIPEA (62.5 µmol, 250 equiv.) were added. The mixture was shaken for 5 min and added to CPG-coupled DNA suspended in 75 µL dry DMF (250 nmol, 1 equiv.). The amide coupling reaction was shaken at ambient temperature for 2 hours. Next, the CPG-coupled conjugate was filtered over a filter column, washed three times with each 200 µL of DMF, MeOH, ACN and CH₂Cl₂ and dried *in vacuo*. Amide coupling was repeated two times.

Completion of amide coupling was controlled by cleaving off a small portion of CPG-coupled oligonucleotide conjugate (0.7–0.9 mg, ~20 nmol) with 500 μ L AMA (AMA = aqueous ammonia (30 %)/ aqueous methylamine (40 %), 1:1, vol/vol) for 30 min (hexT) or 4 h (ATGC-sequences) at ambient temperature. Afterwards 20 μ L of 1 M Tris buffer (pH = 7.5) were added, the mixture was dried under reduced pressure (SpeedVac) and DNA was dissolved in 200 μ L distilled water. The crude reaction mixture was analyzed by analytical RP-HPLC and MALDI-MS. In case of uncompleted coupling (<90%) the reaction was repeated a third time.

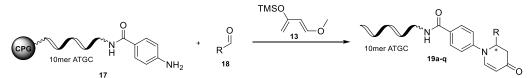
Unreacted amines were capped with acetic acid anhydride (three times 200 μ L, 30 s, 1:1 mixture of THF/methylimidazole, 9:1, vol/vol, and THF/pyridine/acetic acid anhydride 8:1:1, vol/vol). The capped CPG-coupled oligonucleotide conjugate was washed three times with each 200 μ L of DMF, MeOH, ACN and CH₂Cl₂ and dried *in vacuo*.

aza-Diels-Alder reaction with Danishefsky's diene on CPG-coupled oligonucleotidealdehyde conjugates (RP-07)



Prior to reaction, CPG-coupled oligonucleotide, solid amines and ZnCl₂ were dried *in vacuo* for 15 min. Amine **14** (10 µmol, 500 equiv.) was dissolved in 24 µL acetonitrile. The solution was added to CPG-coupled oligonucleotide-aldehyde conjugate **13** (20 nmol) suspended in 12 µL triethyl orthoformate. The suspension was shaken at ambient temperature for 4 h. Afterwards 30 µL of ZnCl₂ (2 µmol, 100 equiv.) in ACN followed by Danisheskys's diene **15** (20 µmol, 1000 equiv.) was added. The reaction mixture was shaken for 1 h at ambient temperature. Then the CPG-coupled oligonucleotide conjugate was filtered over a filter column, washed three times with each 200 µL of 0.1 M EDTA solution, 0.1 M MgCl₂ solution, water, DMF, MeOH, ACN and CH₂Cl₂ and dried *in vacuo*. CPG-coupled oligonucleotide conjugates **16-19** (**16** = hexT, **18** = 10mer ATGC) were cleaved from solid support and deprotected with 200 µL aqueous ammonia (30 %) at 50 °C for 6 h. Afterwards 20 µL of 1 M Tris buffer (pH = 7.5) were added, the mixture was dried under reduced pressure (SpeedVac) and DNA was dissolved in 200 µL distilled water. The crude reaction mixture was analyzed by analytical RP-HPLC (Method I) and MALDI-TOF-MS. The product was purified by preparative RP-HPLC.

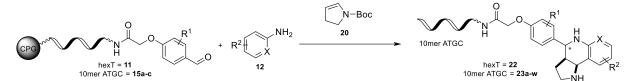
aza-Diels-Alder reaction with Danishefsky's diene on CPG-coupled oligonucleotide-aniline conjugates (RP-08)



Prior to reaction, CPG-coupled oligonucleotide, solid aldehydes and $ZnCl_2$ were dried *in vacuo* for 15 min. Aldehyde **14** (30 µmol, 1500 equiv.) was dissolved in 24 µL tetrahydrofuran. The solution was added to CPG-coupled oligonucleotide-aniline conjugate **13** (20 nmol) suspended in 12 µL triethyl orthoformate. The suspension was shaken at ambient temperature for 4 h. Afterwards 30 µL of $ZnCl_2$ (2 µmol, 100 equiv.) in tetrahydrofuran followed by Danishefsky's diene **15** (20 µmol, 1000 equiv.) were added. The reaction mixture was shaken at ambient temperature for 1 h. Then the CPG-coupled oligonucleotide conjugate was filtered over a filter column, washed three times with each 200 µL of 0.1 M EDTA solution, 0.1 M MgCl₂ solution,

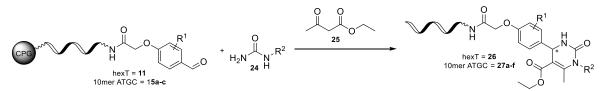
water, DMF, MeOH, ACN and CH_2CI_2 and dried *in vacuo*. CPG-coupled oligonucleotide conjugates **16** and **19** (**16** = hexT, **19** = 10mer ATGC) were cleaved from solid support and deprotected with 200 µL aqueous ammonia (30 %) at 50 °C for 6 h. Afterwards, 20 µL of 1 M Tris buffer (pH = 7.5) were added, the mixture was dried under reduced pressure (SpeedVac) and DNA was dissolved in 200 µL distilled water. The crude reaction mixture was analyzed by analytical RP-HPLC (Method I) and MALDI-TOF-MS. The product was purified by preparative RP-HPLC.

(R)-(-)-BNDHP-mediated Povarov reaction on CPG-coupled oligonucleotides (RP-09)



Prior to use, CPG-coupled oligonucleotide, solid anilines and (R)-(-)-BNDHP were dried in vacuo for 15 min. Aniline 12 (10 µmol, 500 equiv.) was dissolved in 24 µL ethanol. The solution was added to CPG-coupled oligonucleotide-aldehyde conjugate 15 (20 nmol) suspended in 12 µL triethyl orthoformate. The suspension was shaken at ambient temperature for 4 h. Afterwards 30 µL of (R)-(-)-BNDHP (2 µmol, 100 equiv.) in ethanol followed by N-Boc-2,3-dihydro-1H-pyrrole 20 (10 µmol, 500 equiv.) were added. The reaction mixture was shaken at 50 °C for 16 h. Then the CPG-coupled oligonucleotide conjugate was filtered over a filter column, washed three times with each DMF, MeOH, ACN and CH₂Cl₂ and dried in vacuo. Boc-protecting group was removed by addition of 200 µL 75 % trifluoroacetic acid in CH₂Cl₂ for 30 sec.. Afterwards CPG-coupled DNA was washed with excess of 1 % TEA and CH_2CI_2 . The deprotection was repeated 4 times. The solid phase was washed three times with each 200 µL of 1 % TEA in ACN, DMF, MeOH, ACN and CH₂Cl₂ and dried in vacuo. CPG-coupled oligonucleotide conjugates 22-23 were cleaved from solid support and deprotected with 500 µL AMA (AMA = aqueous ammonia (30 %)/ aqueous methylamine (40 %), 1:1, vol/vol) for 30 min (hexT 22) or 4 h (ATGC-sequences 23) at ambient temperature. Afterwards 20 μ L of 1 M Tris buffer (pH = 7.5) were added, the mixture was dried under reduced pressure (SpeedVac) and DNA was dissolved in 200 µL distilled water. The crude reaction mixture was analyzed by analytical RP-HPLC (Method I) and MALDI-TOF-MS. The product was purified by preparative RP-HPLC.

(R)-(-)-BNDHP-mediated Biginelli reaction on CPG-coupled oligonucleotides (RP-10)



Prior to reaction, CPG-coupled oligonucleotide, ureas and (*R*)-(-)-BNDHP were dried *in vacuo* for 15 min. Urea **24** (10 µmol, 500 equiv.) and (*R*)-(-)-BNDHP (1 µmol, 50 equiv.) were dissolved both in 30 µL ethanol. The solutions were added to CPG-coupled oligonucleotide-aldehyde conjugate **15** (20 nmol) followed by ethyl acetoacetate **25** (10 µmol, 500 equiv.). The reaction mixture was shaken at 50 °C for 20 h. Then the CPG-coupled oligonucleotide conjugate was filtered over a filter column, washed three times with each DMF, MeOH, ACN and CH₂Cl₂ and dried *in vacuo*. CPG-coupled oligonucleotide conjugates **26-27** were cleaved from solid support and deprotected with 500 µL AMA (AMA = aqueous ammonia (30 %)/ aqueous methylamine (40 %), 1:1, vol/vol) for 30 min (hexT **26**) or 4 h (ATGC-sequences **27**) at ambient temperature. Afterwards 20 µL of 1 M Tris buffer (pH = 7.5) were added, the mixture was dried under reduced pressure (SpeedVac) and DNA was dissolved in 200 µL distilled water. The crude reaction mixture was analyzed by analytical RP-HPLC (Method I) and MALDI-TOF-MS. The product was purified by preparative RP-HPLC.

Investigation in the stability of solid phase coupled oligonucleotides charged with metal ions and organocatalysts

Entry	Metal salt	Solvent	hexT-Ac	ТС	ATC	ATCG
1	2% TFA					
2	10% TFA					
3	3.7% HCI					
4	AgOAc	CH ₂ Cl ₂				
5	AgOTf	ACN				
6	AgSbF ₆	ACN				
7 ^b	BiBr₃	ACN				
8	Bi(OTf)₃	MeOH				
9	Ce(NH ₄) ₂ (NO ₃) ₆	MeOH				
10	Co(acac) ₃	ACN				
11 ^b	CuCl	MeOH				
12	Cu(MeCN) ₄ PF ₆	ACN				
13	Cu(OTf) ₂	ACN				
14	Fe(acac)₃	ACN				
15	FeCl ₂ • 4 H ₂ O	ACN				
16	InCl₃	ACN				
17 ^b	La(O <i>i</i> -Pr)₃	THF				
18	LiBr	ACN				
19	Mg(ClO ₄) ₂	MeOH				
20	Ni(acac) ₂	ACN				
21	Ni(PPh ₃) ₂ Cl ₂	MeOH				
22 ^b	Pd(dba)₃	MeOH				
23a	Pd(OAc) ₂	ACN				
23b ^c	Pd(OAc) ₂	ACN				
24 ^c	Pd(PPh ₃) ₄	MeOH				
25 ^b	[Rh(cod)Cl]2	MeOH				
26	RuCl₃	ACN				
27	[Ru(p-cymene)Cl ₂]2	CH ₂ Cl ₂				
28	Ru(Me-allyl) ₂ (COD)	CH ₂ Cl ₂				
29	Grubbs 1 st Gen.	CH ₂ Cl ₂				
30	SbCl₃	ACN				
31	Sc(OTf)₃	ACN				
32	SeO ₂	MeOH				
33	Ti(O <i>i</i> −Pr)₄	MeOH				
34	VO(acac) ₂	MeOH				
35	Yb(OTf)₃	MeOH				
36	ZnCl ₂	ACN				

Table S1 – Stability of DNA against metal salts^a

^a for each: 20 nmol DNA, 200 eq. transition metal salt, 50 μL solvent, r.t., 22 h. ^b poor solubility, added as suspension. ^c 5 equiv. of metal salt were used. ACN = acetonitrile, MeOH = methanol, THF = tetrahydrofuran.

				DNA degradation
0-20%	21-40%	41-60%	> 61%	DNA degradation

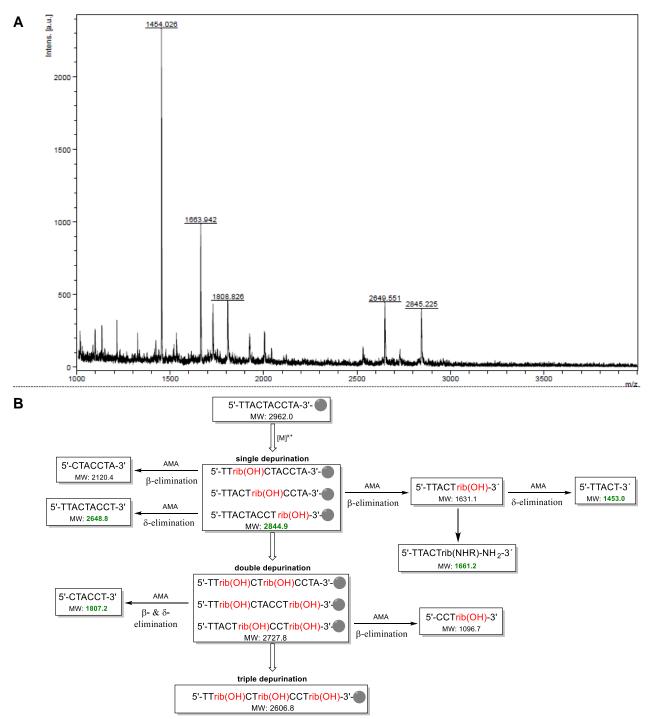
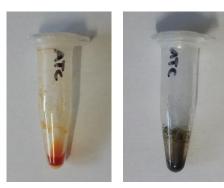


Figure S1 Representative mass spectrometric analysis of degraded DNA. A: MALDI-TOF spectrum for 10mer ATC-oligonucleotide **9** after treatment with aqueous 3.7% HCI. B: Possible depurination and fragmentation products of 10mer ATC-oligonucleotide **9**.



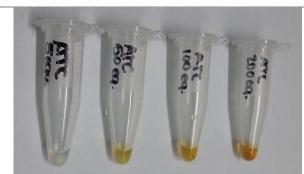
Colouring of the Pd(PPh₃)₄ solution indicated redox processes which are plausible as all experiments were performed without protective gas.

Pd(PPh₃)₄, 3 h

Α

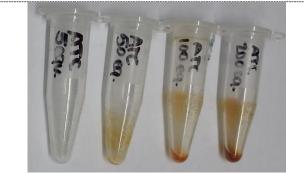
В

Pd(PPh₃)₄, 22 h



CPG-coupled DNA suspended in a solution of x equiv. $Pd(OAc)_2$ for 0 h at ambient temperature.

5 equiv. 50 equiv. 100 equiv. 200 equiv.



50 equiv.

CPG-coupled DNA suspended in a solution of x equiv. $Pd(OAc)_2$ for 22 h at ambient temperature.

5 equiv.

100 equiv. 200 equiv.

D

С



CPG-coupled DNA after incubation with a solution of x equiv. $Pd(TFA)_2$, for 22 h at ambient temperature and multiple washing steps. Slight coloring of CPG treated with higher amounts of $Pd(OAc)_2$ is observable.

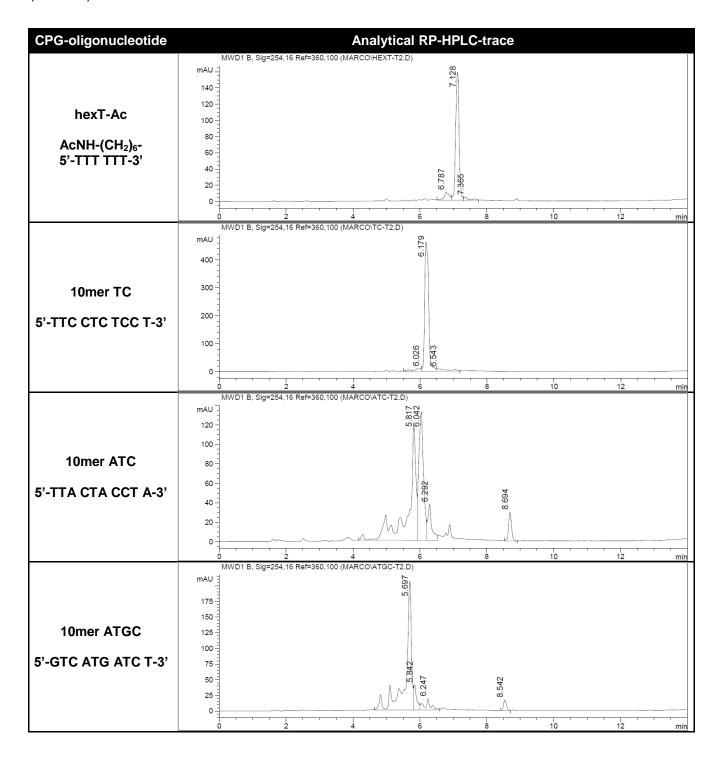
5 equiv. 50 equiv. 100 equiv. 200 equiv.

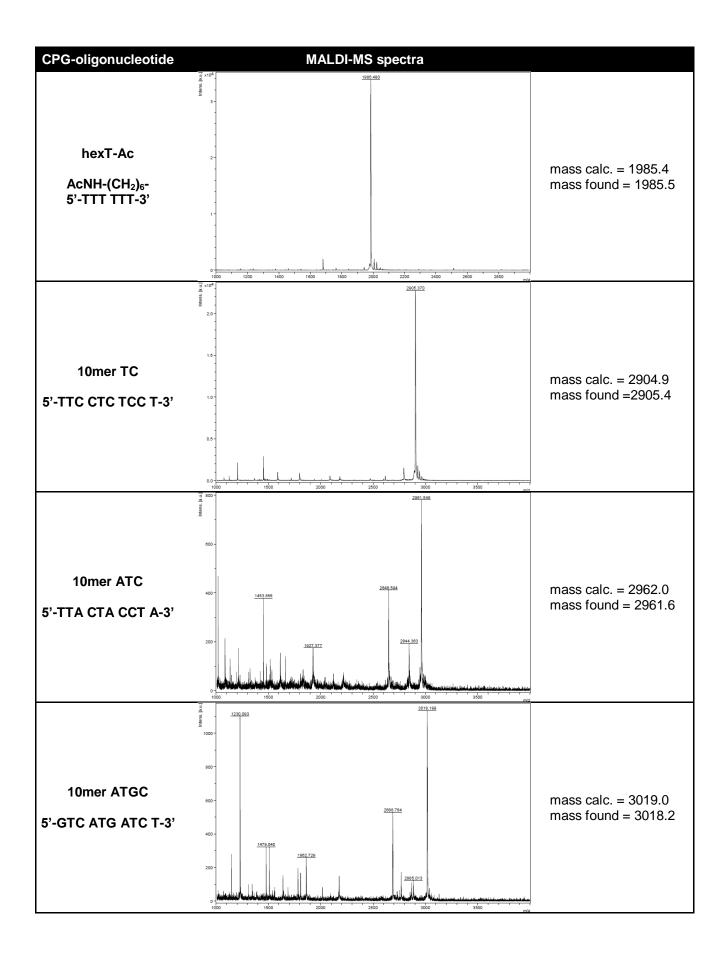
Figure S2 Photos of CPG-coupled DNA-oligonucleotides treated with Pd(O) and Pd(II).

HPLC traces and MALDI-MS spectra of metal ion screens

CPG-oligonucleotide + 2% TFA

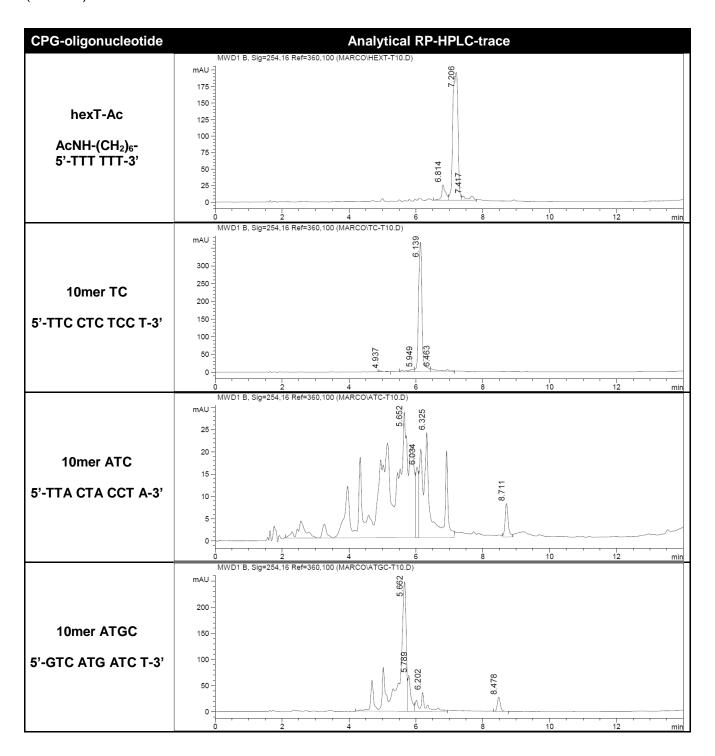
According to the representative procedure (RP-05) solid support coupled oligonucleotide (20 nmol) was treated with 2% TFA.

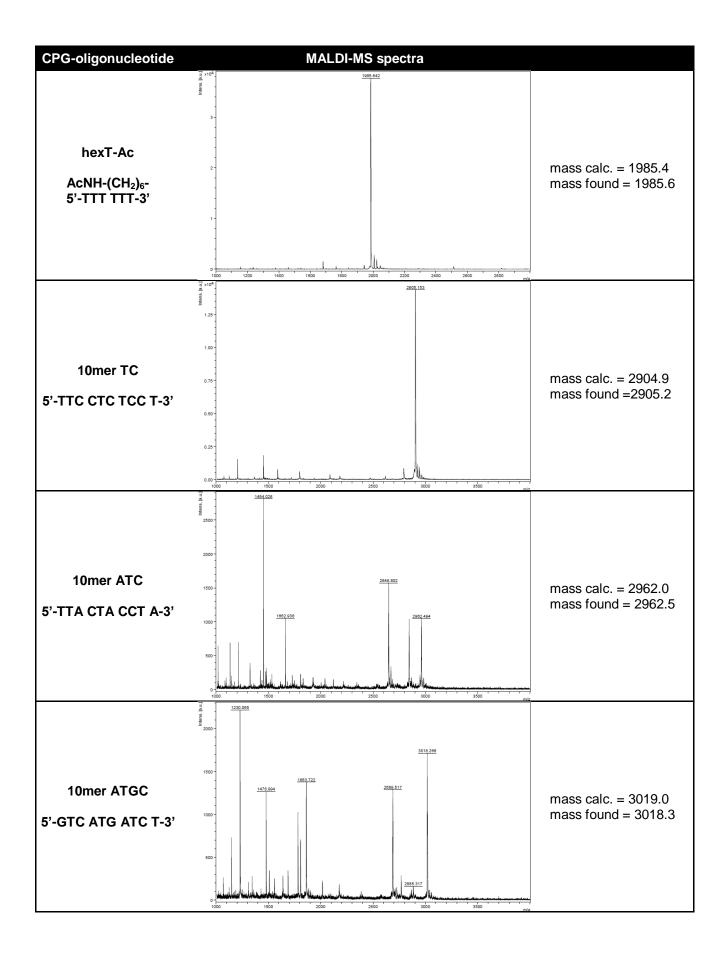




CPG-oligonucleotide + 10 % TFA

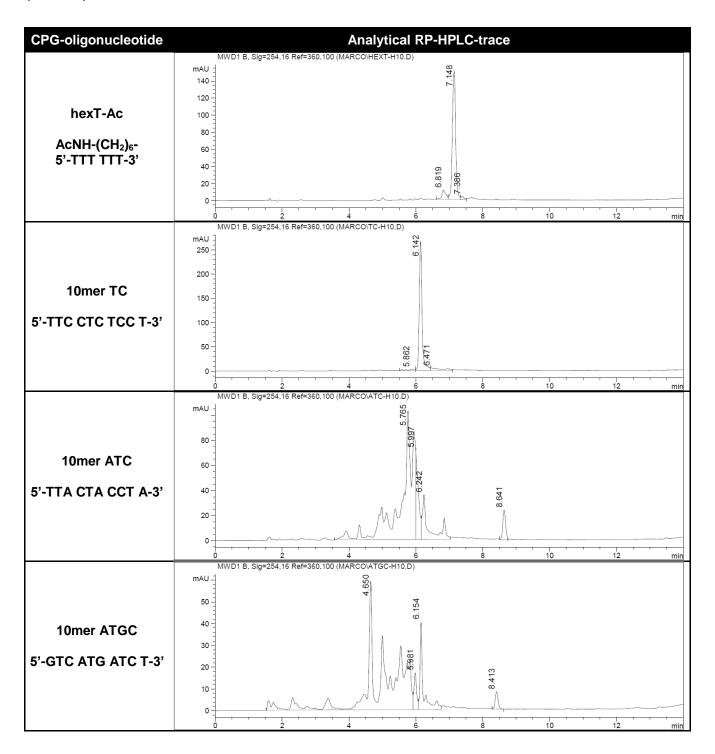
According to the representative procedure (RP-05) solid support coupled oligonucleotide (20 nmol) was treated with 10 % TFA.

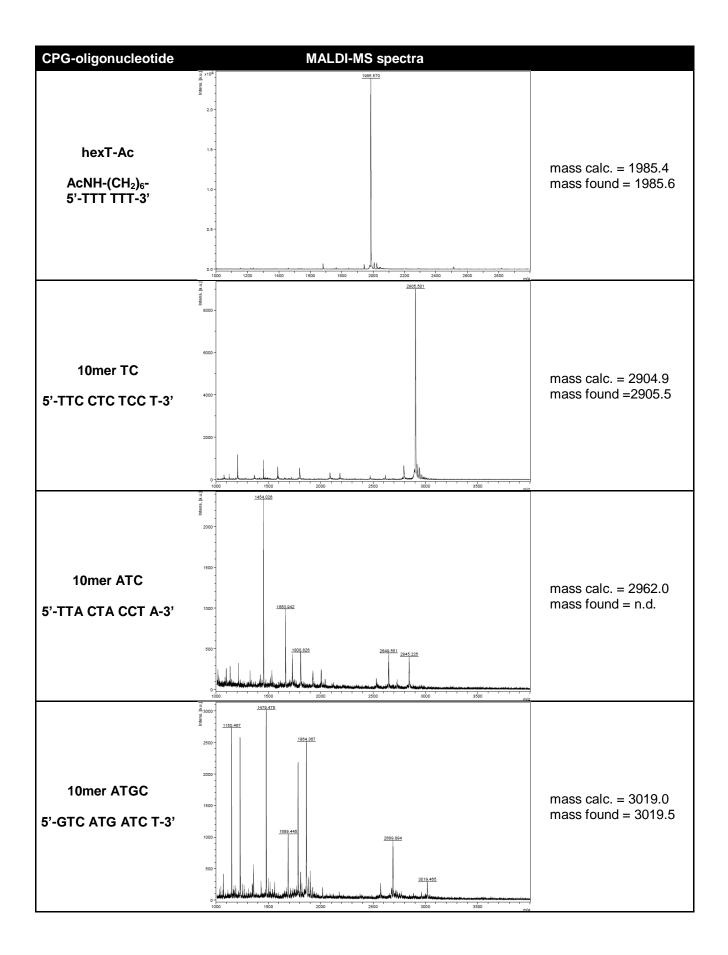




CPG-oligonucleotide + 3.7% HCl

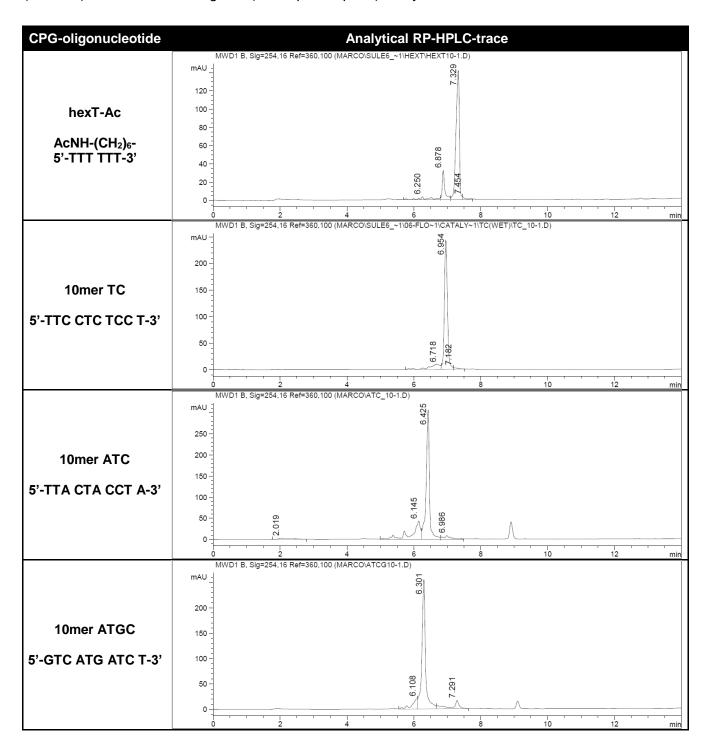
According to the representative procedure (RP-05) solid support coupled oligonucleotide (20 nmol) was treated with 3.7% HCl.

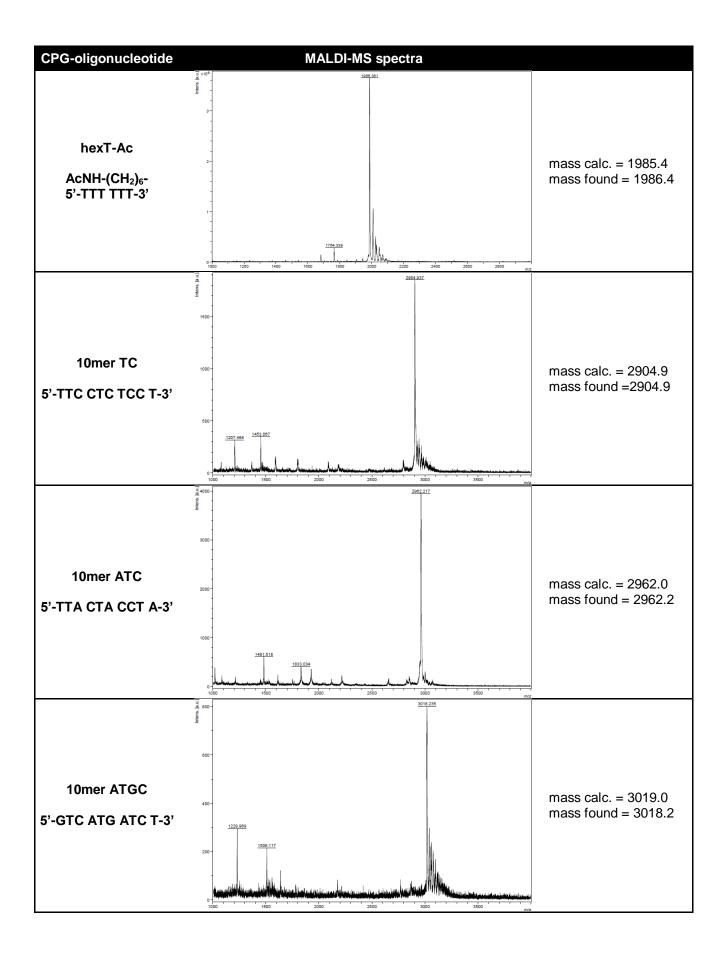




CPG-oligonucleotide + AgOAc

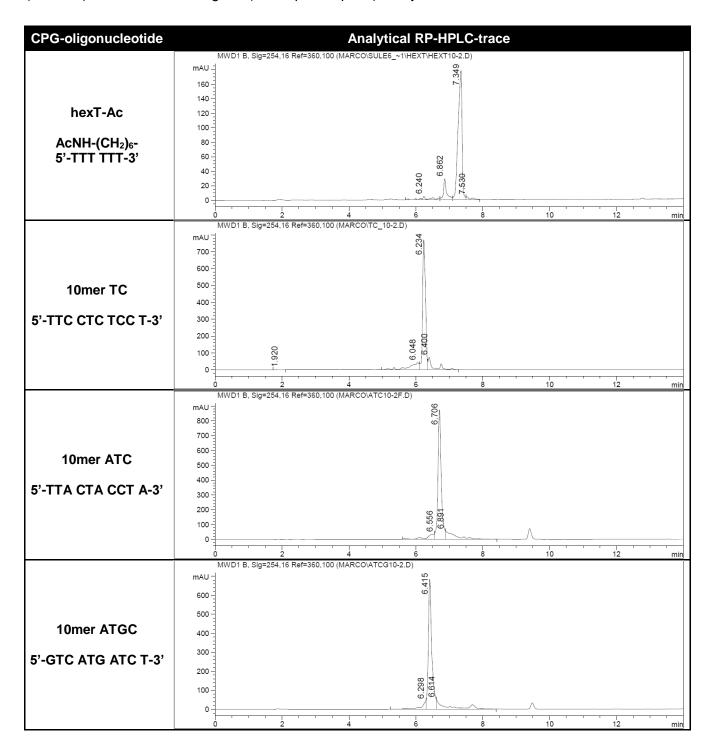
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with AgOAc (200 equiv., 4 µmol) in dry CH₂Cl₂.

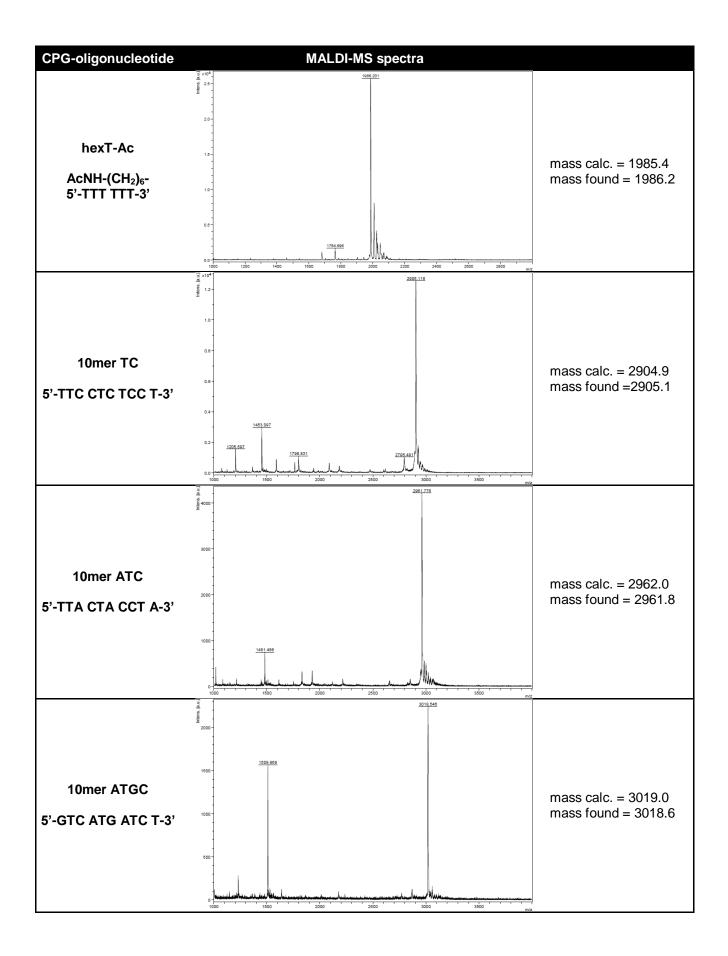




CPG-oligonucleotide + AgOTf

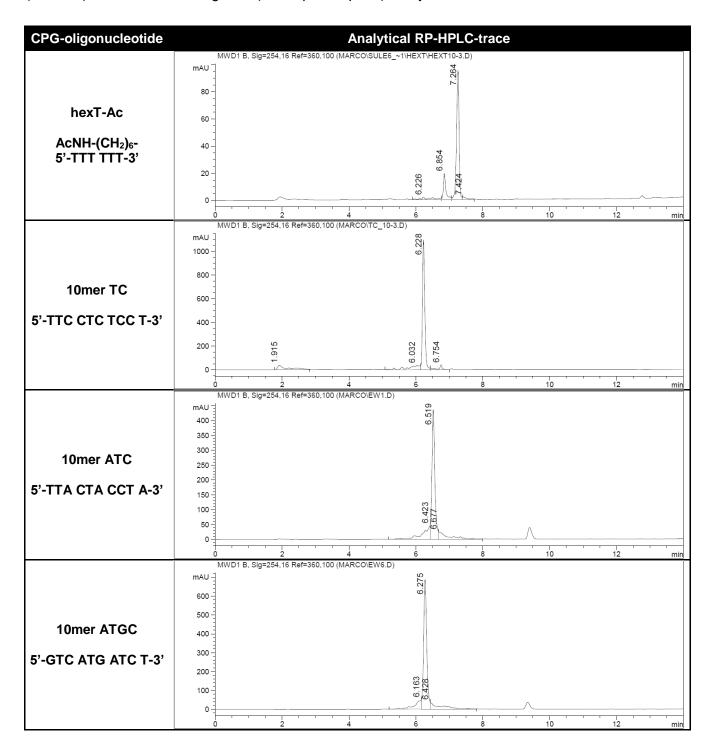
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with AgOTf (200 equiv., 4 µmol) in dry ACN.

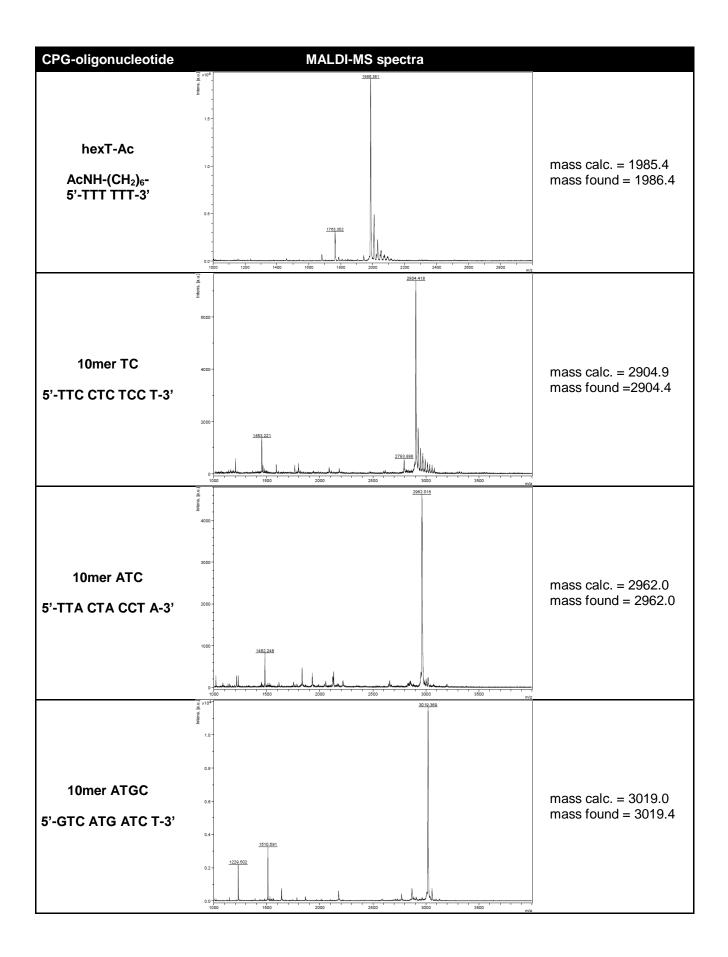




CPG-oligonucleotide + AgSbF₆

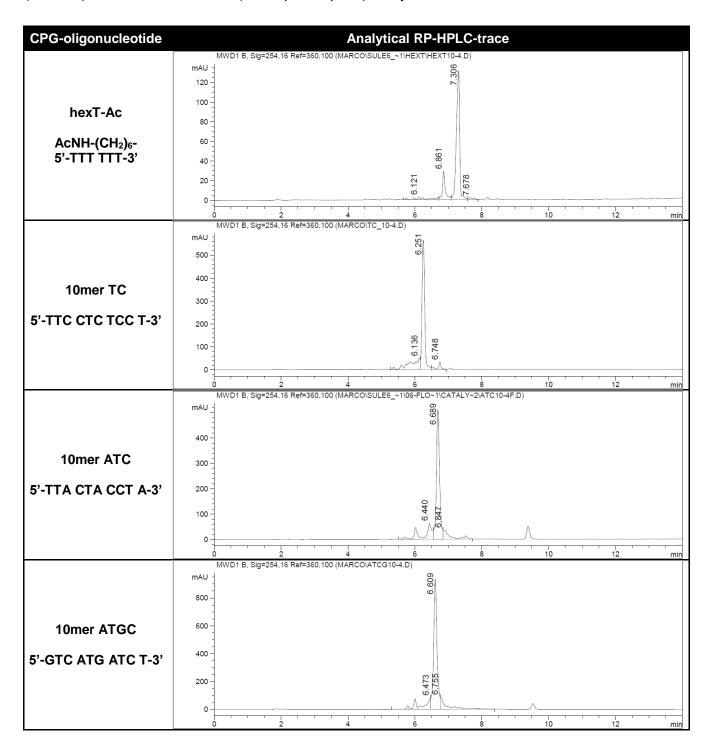
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with AgSbF₆ (200 equiv., 4 µmol) in dry ACN.

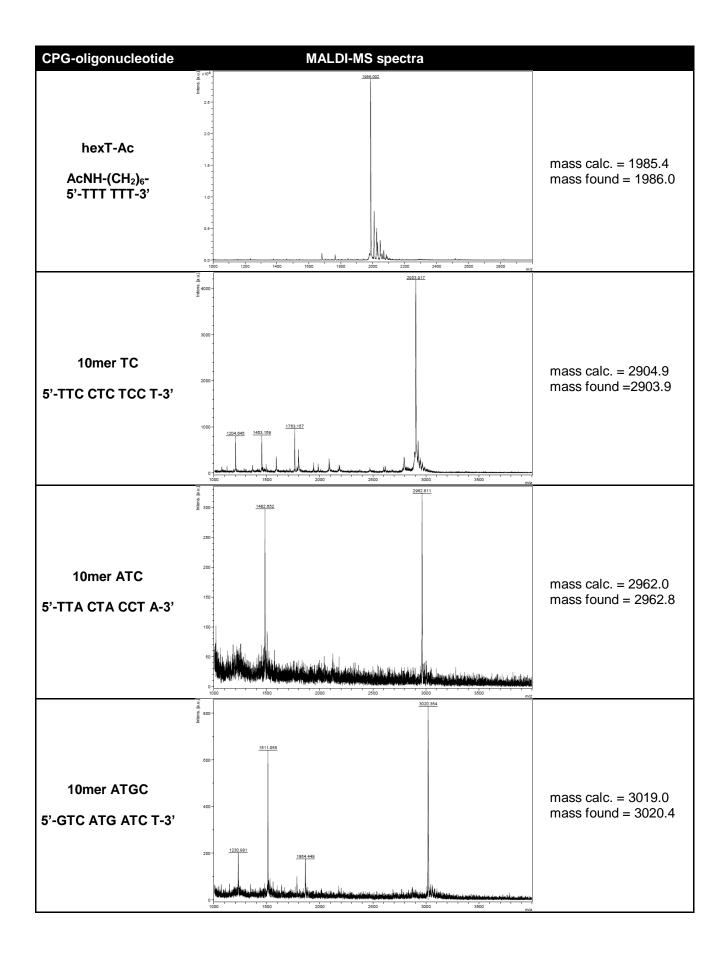




CPG-oligonucleotide + BiBr₃

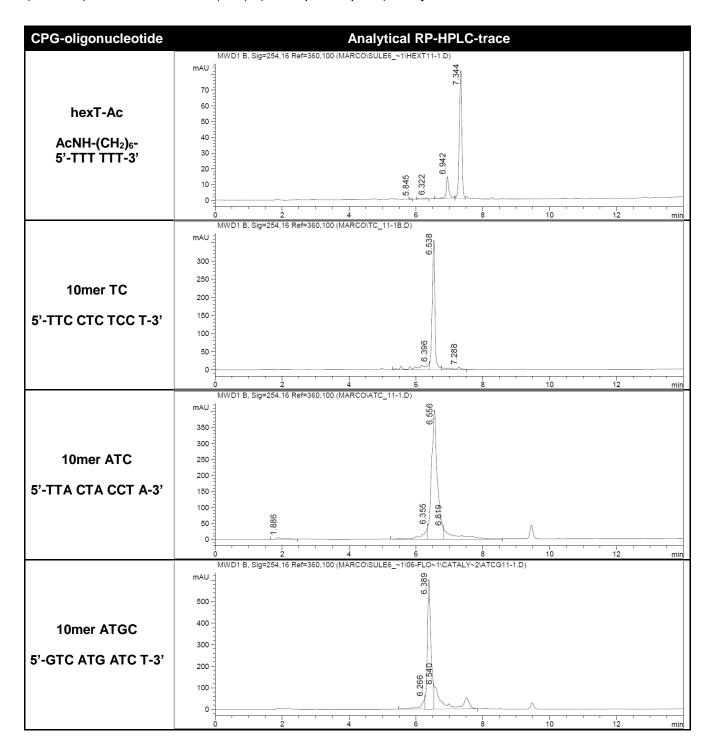
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with BiBr₃ (200 equiv., 4 µmol) in dry ACN.

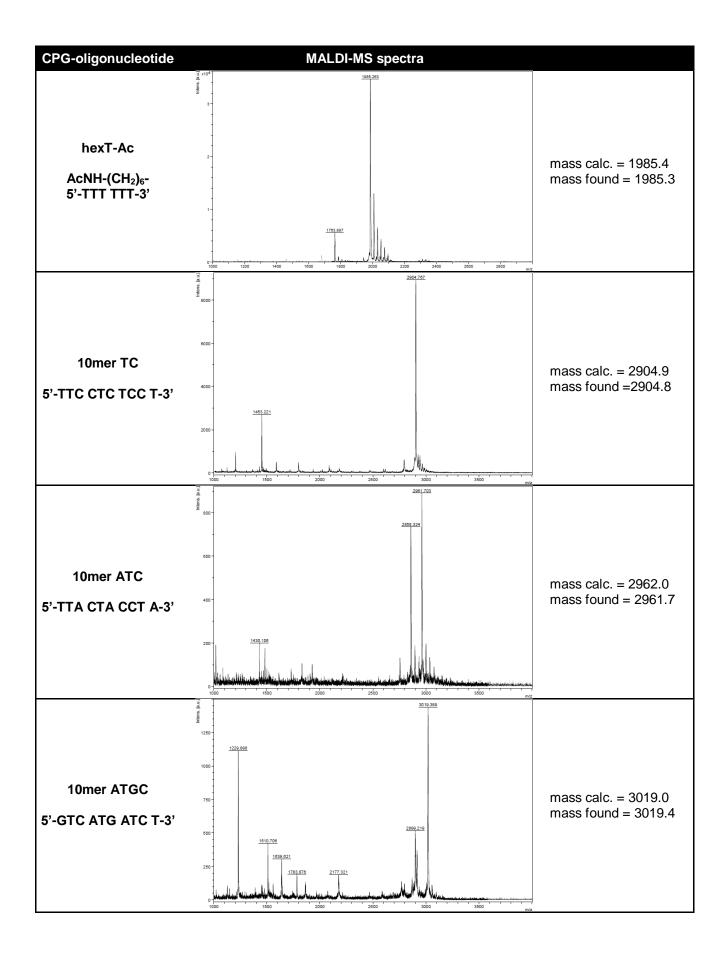




CPG-oligonucleotide + Bi(OTf)₃

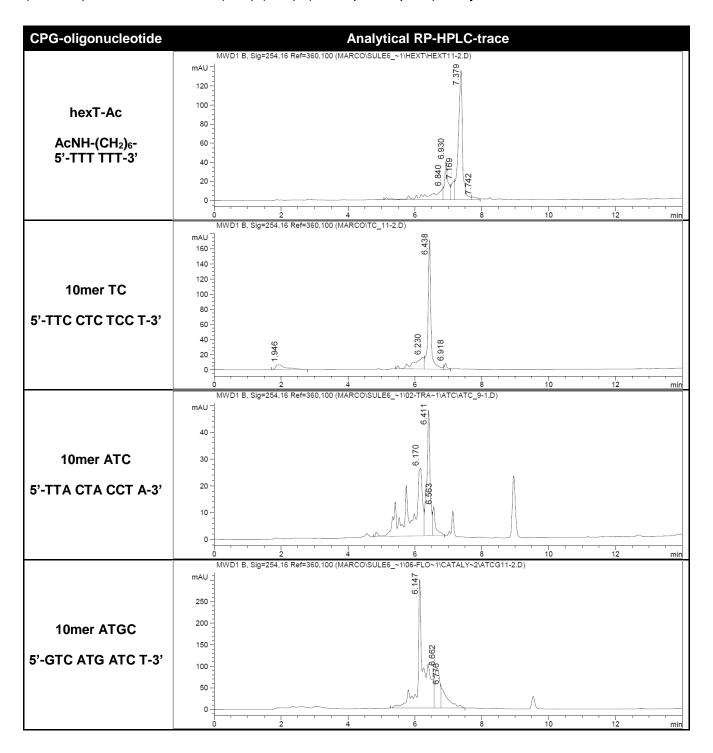
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with $Bi(OTf)_3$ (200 equiv., 4 µmol) in dry MeOH.

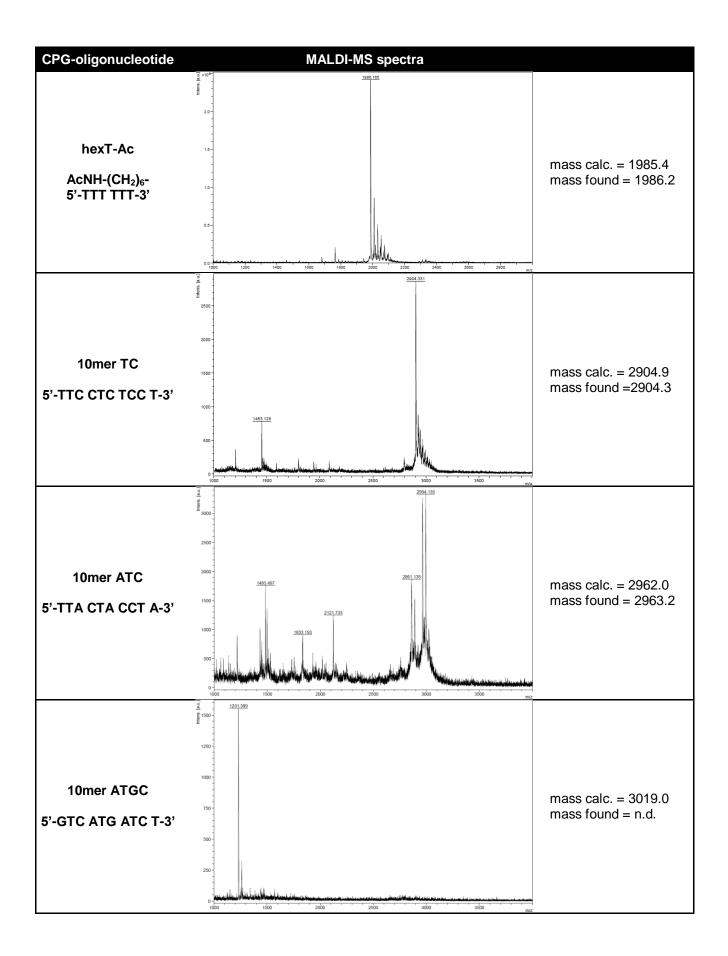




CPG-oligonucleotide + Ce(NH₄)₂(NO₃)₆

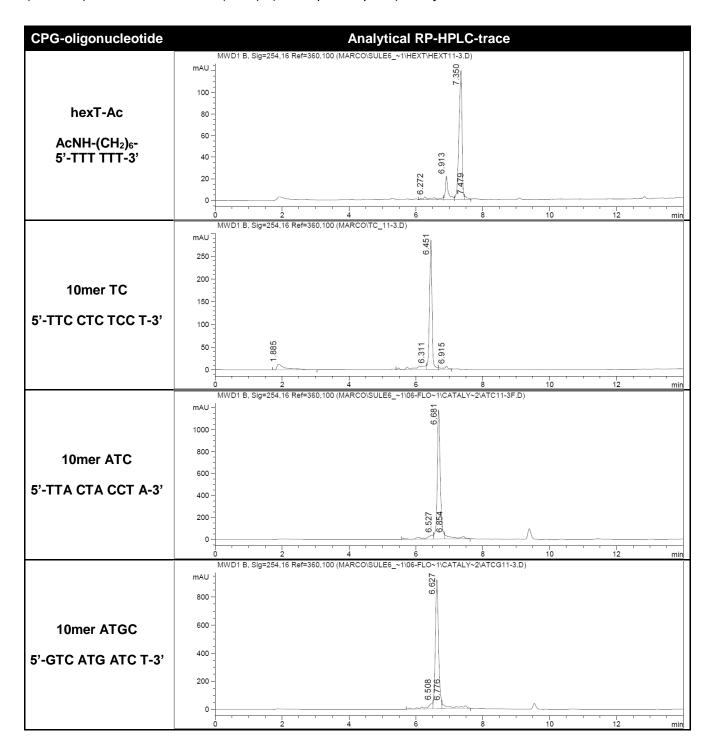
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with $Ce(NH_4)_2(NO_3)_6$ (200 equiv., 4 µmol) in dry MeOH.

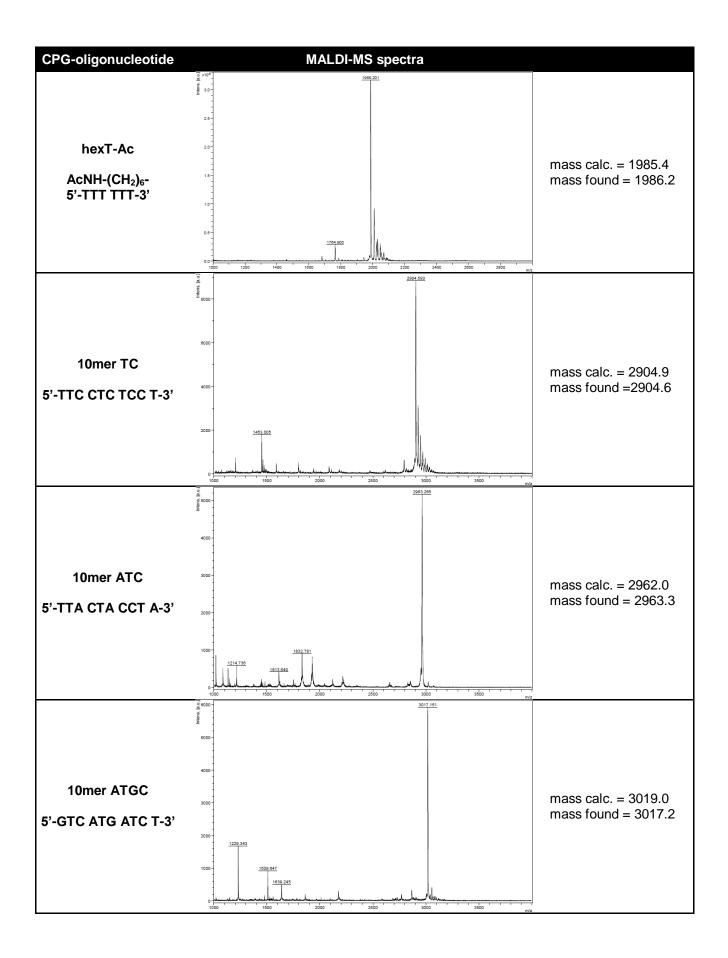




CPG-oligonucleotide + Co(acac)₃

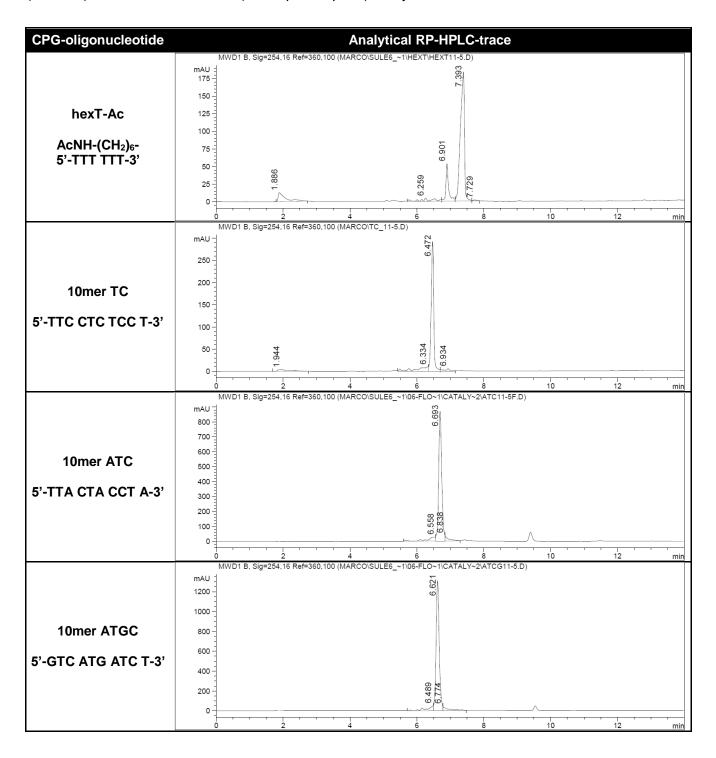
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with $Co(acac)_3$ (200 equiv., 4 µmol) in dry ACN.

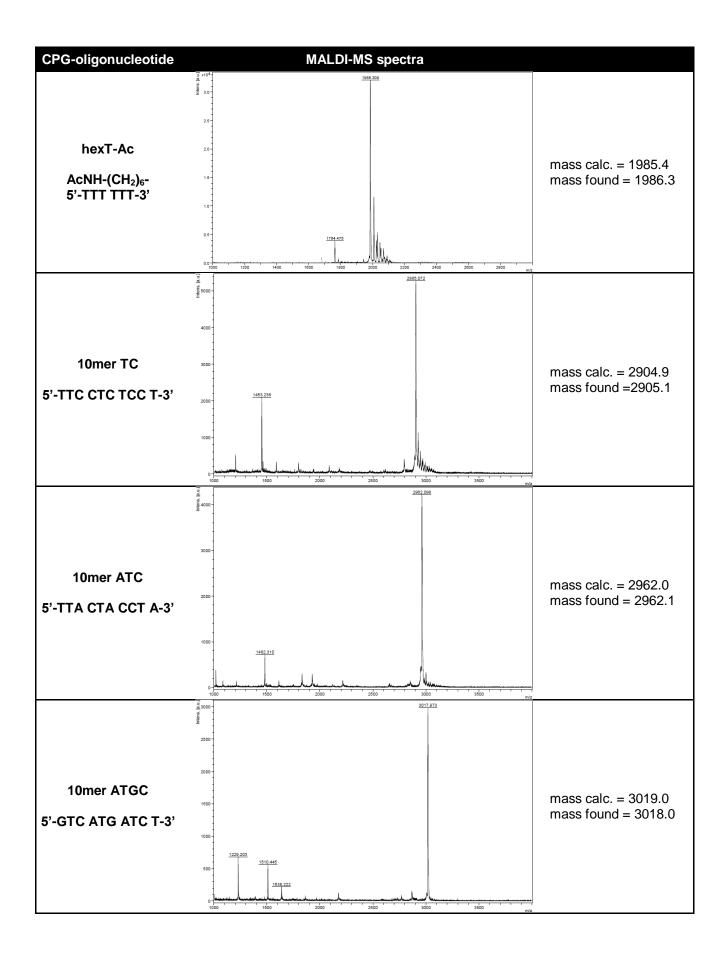




CPG-oligonucleotide + CuCl

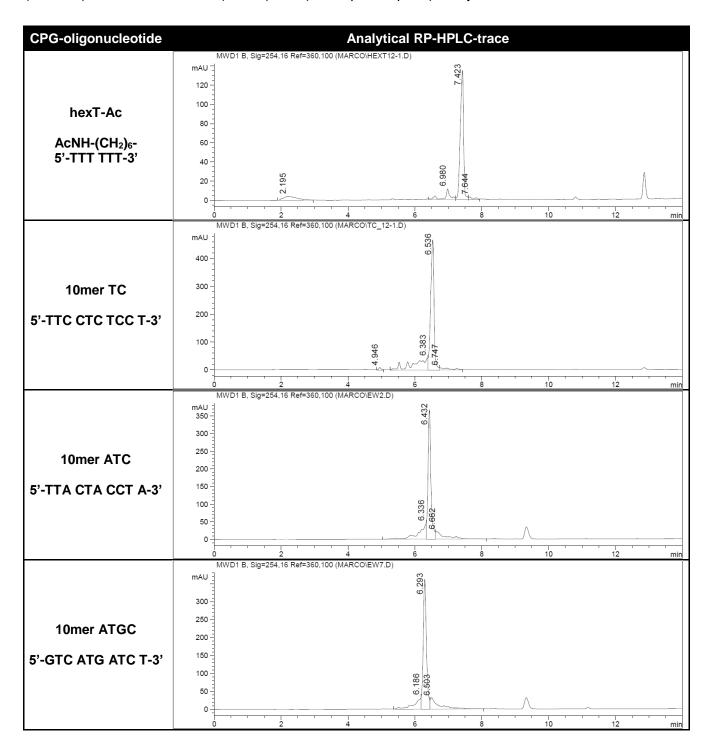
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with CuCl (200 equiv., 4 µmol) in dry MeOH.

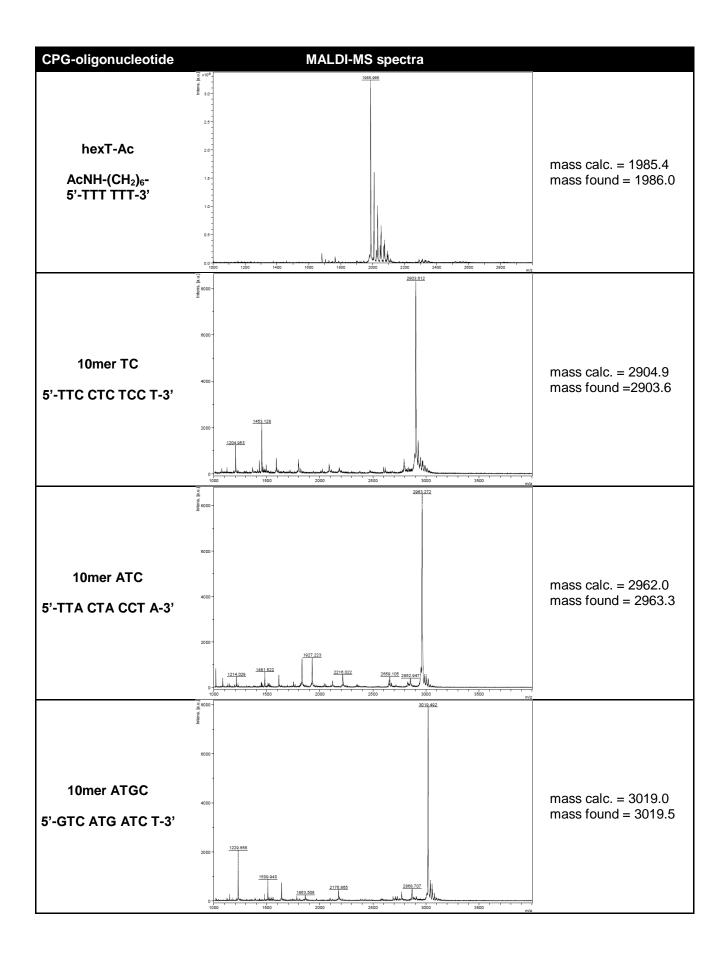




CPG-oligonucleotide + Cu(MeCN)₄PF₆

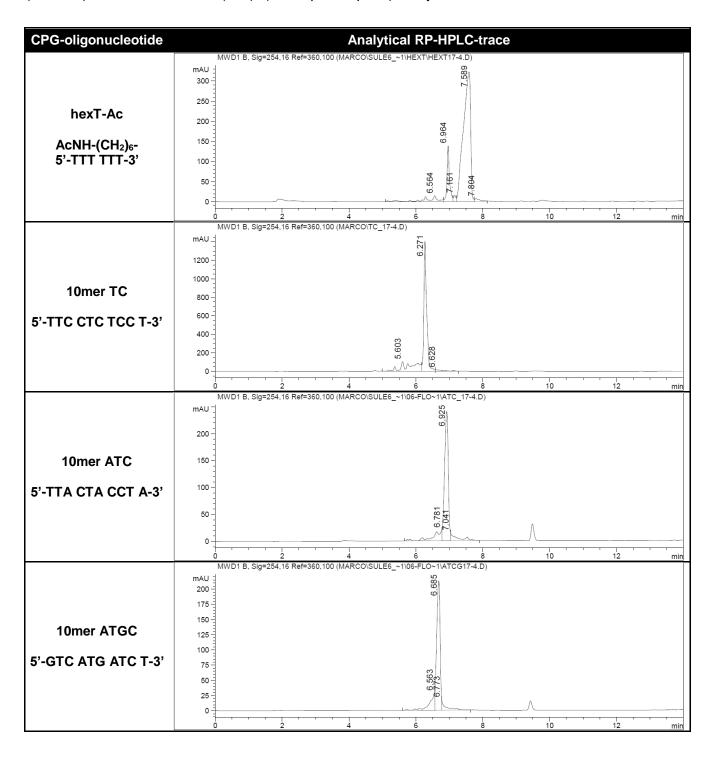
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Cu(MeCN)₄PF₆ (200 equiv., 4 µmol) in dry ACN.

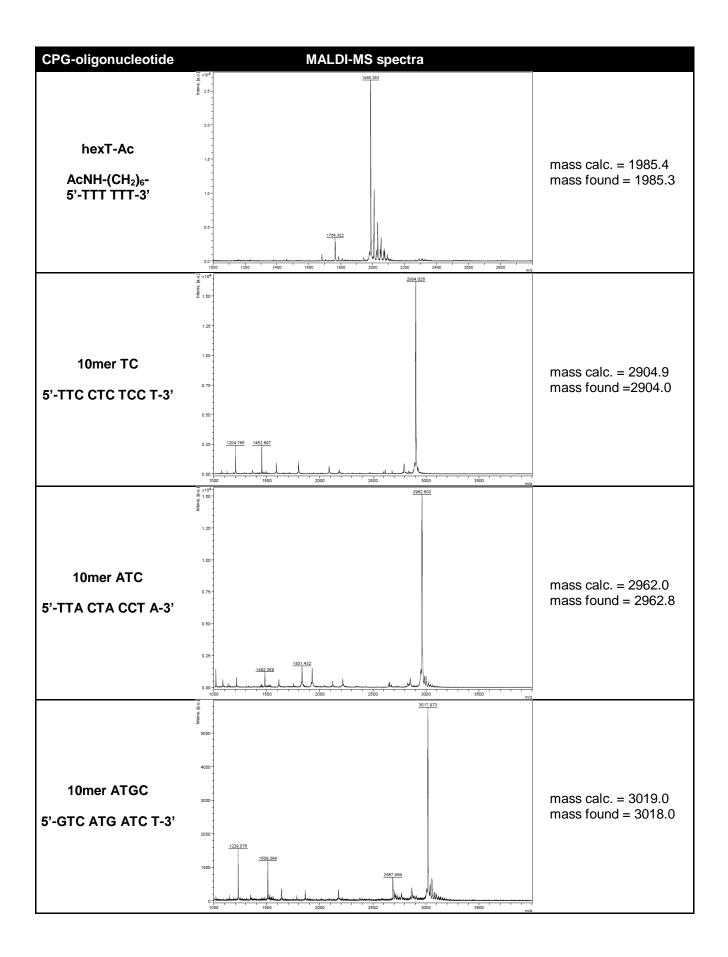




CPG-oligonucleotide + Cu(OTf)₂

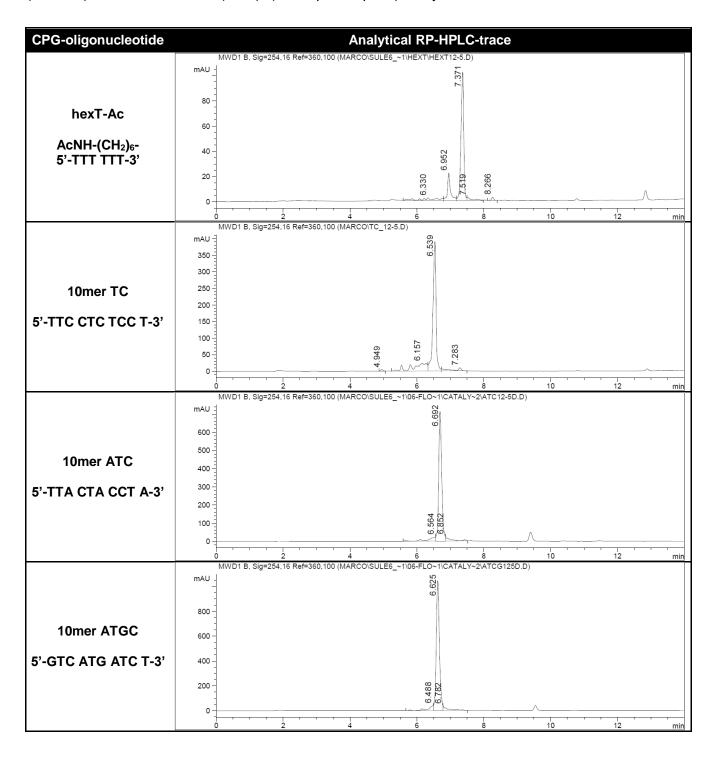
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with $Cu(OTf)_2$ (200 equiv., 4 µmol) in dry ACN.

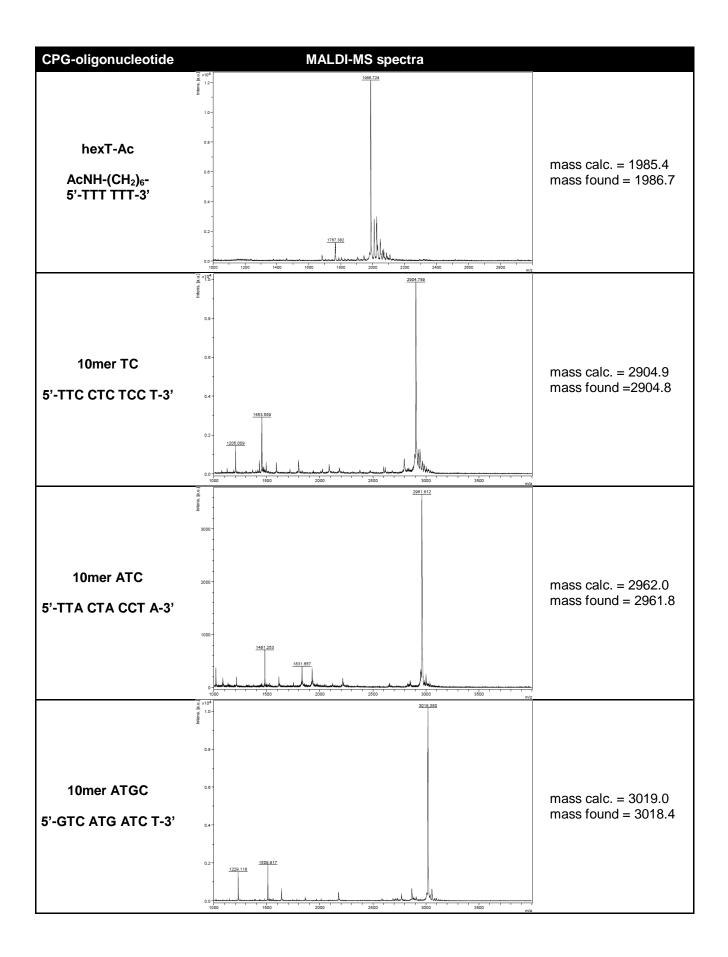




CPG-oligonucleotide + Fe(acac)₃

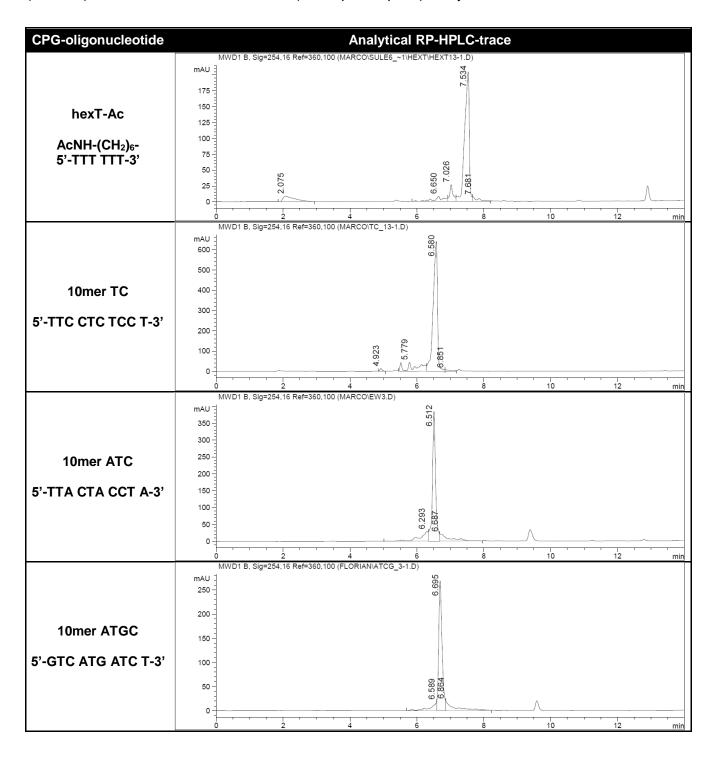
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Fe(acac)₃ (200 equiv., 4 µmol) in dry ACN.

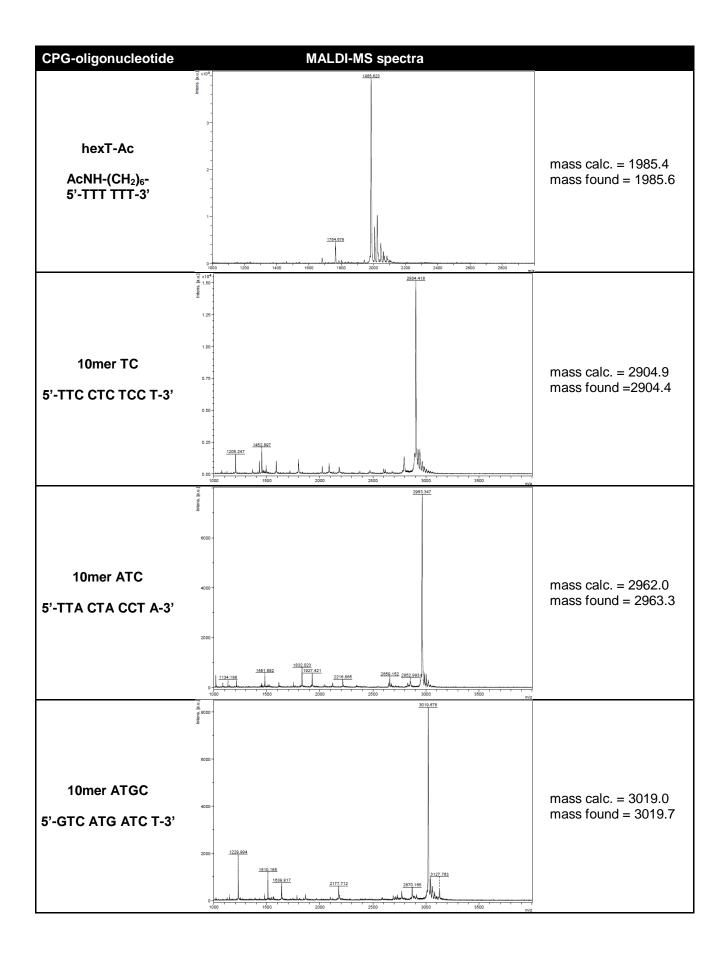




$CPG-oligonucleotide + FeCl_2 \cdot 4 H_2O$

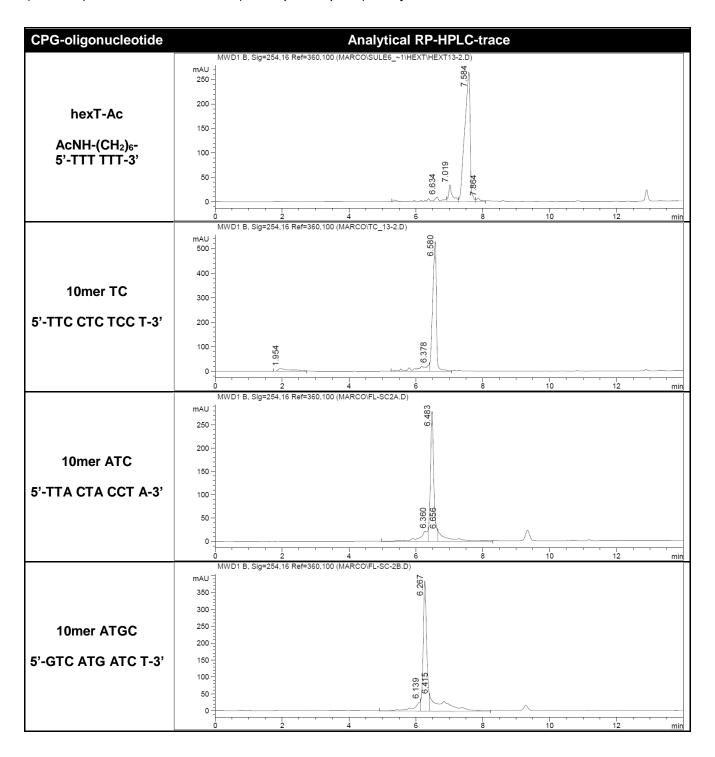
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with $FeCI_2 \cdot 4 H_2O$ (200 equiv., 4 µmol) in dry ACN.

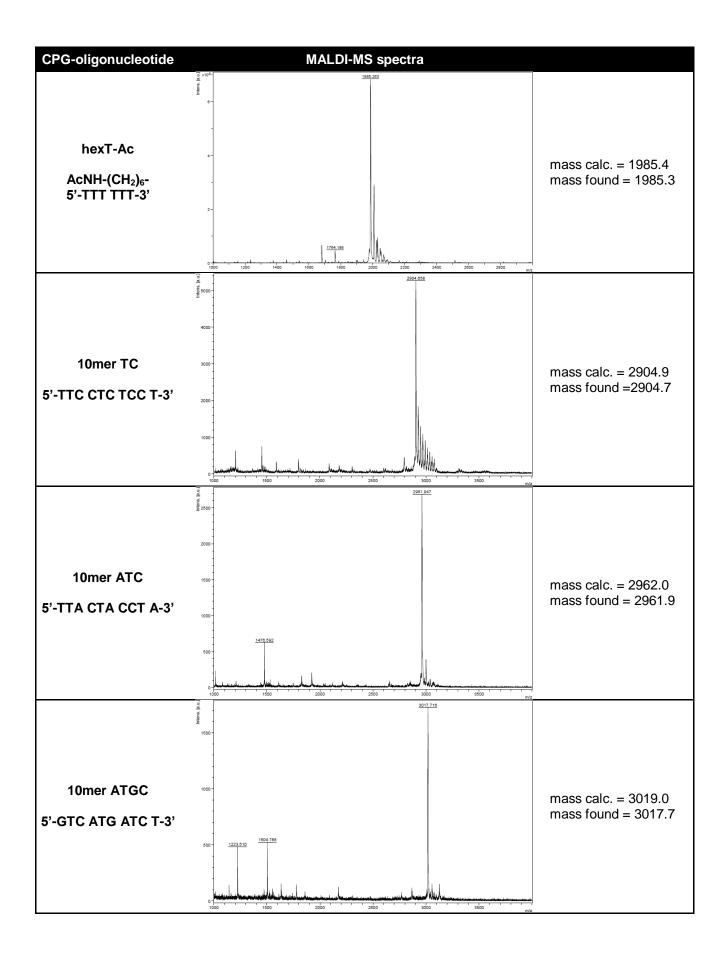




CPG-oligonucleotide + InCl₃

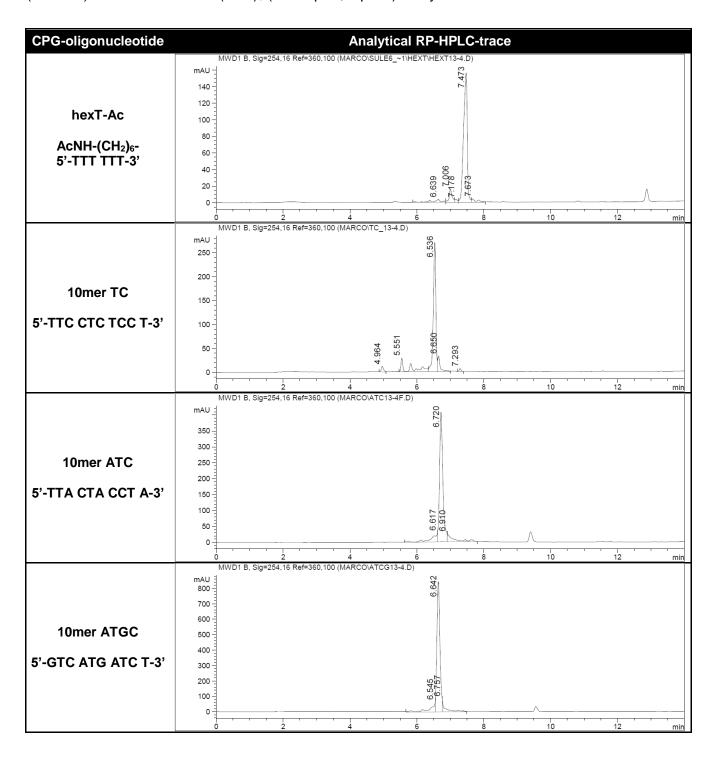
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with InCl₃ (200 equiv., 4 µmol) in dry ACN.

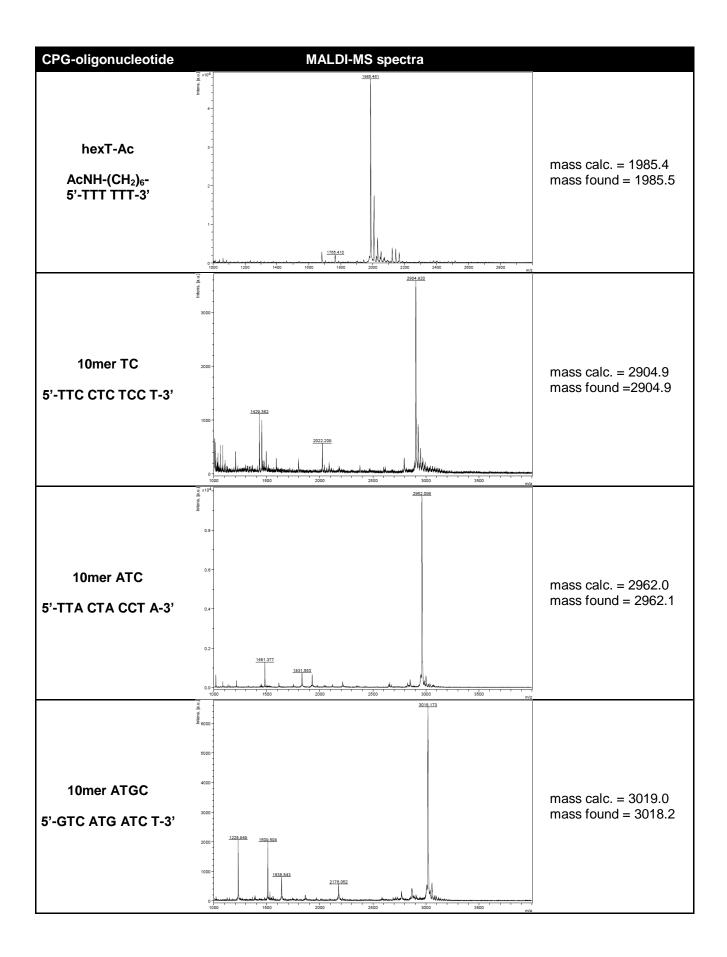




CPG-oligonucleotide + La(O*i*Pr)₃

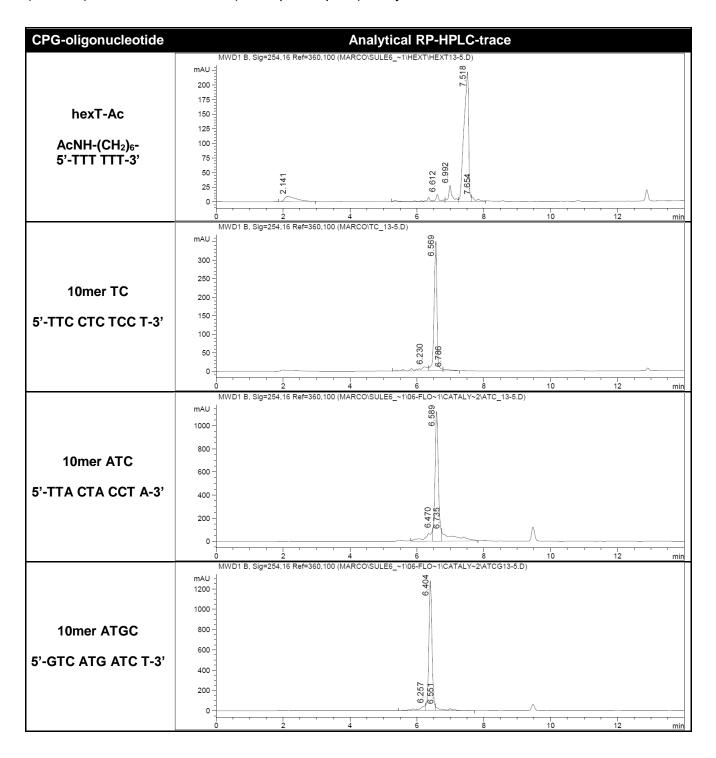
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with $La(OiPr)_3$ (200 equiv., 4 µmol) in dry THF.

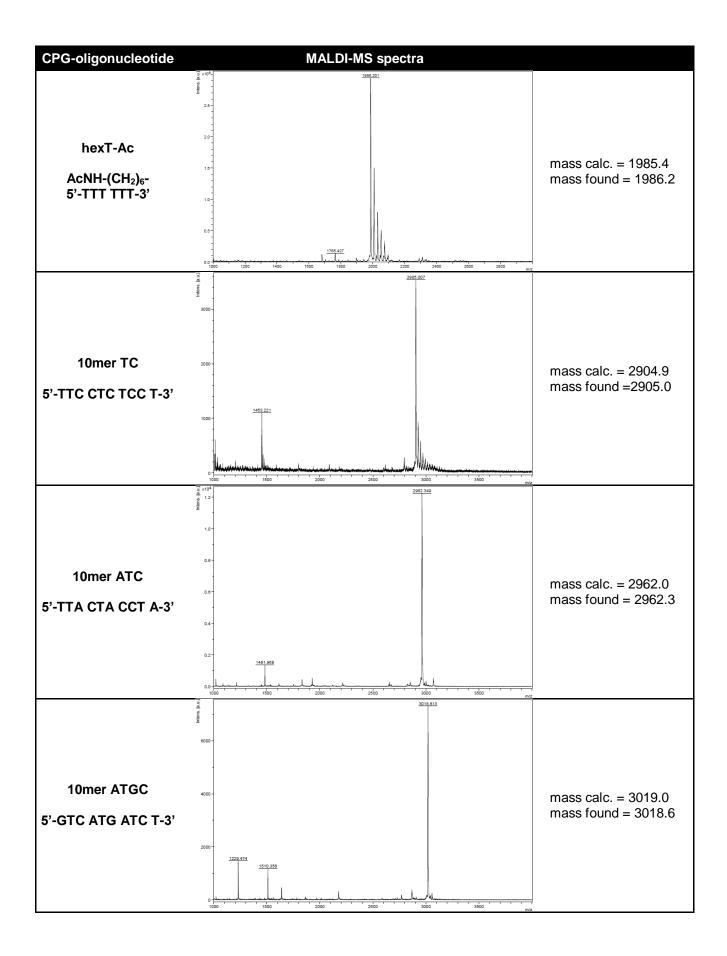




CPG-oligonucleotide + LiBr

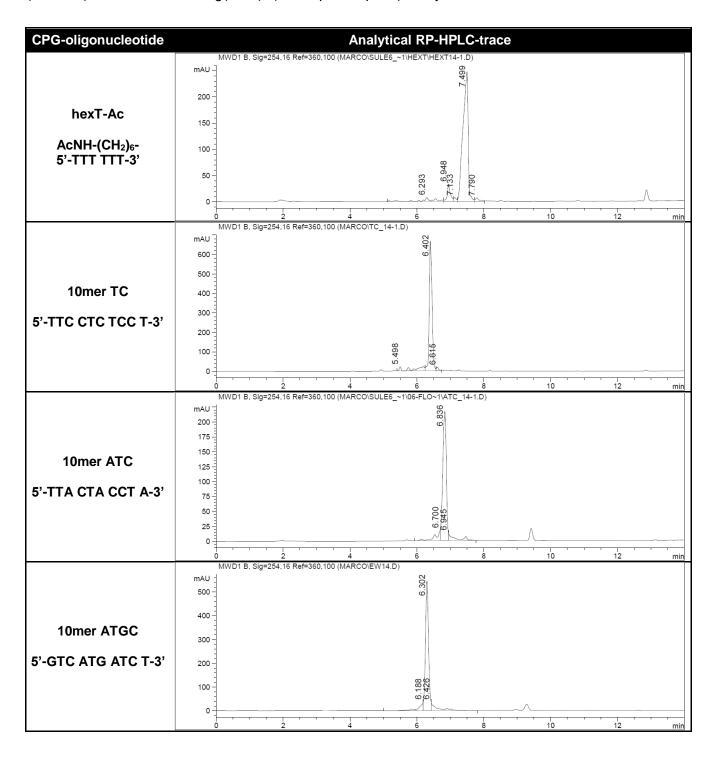
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with LiBr (200 equiv., 4 µmol) in dry ACN.

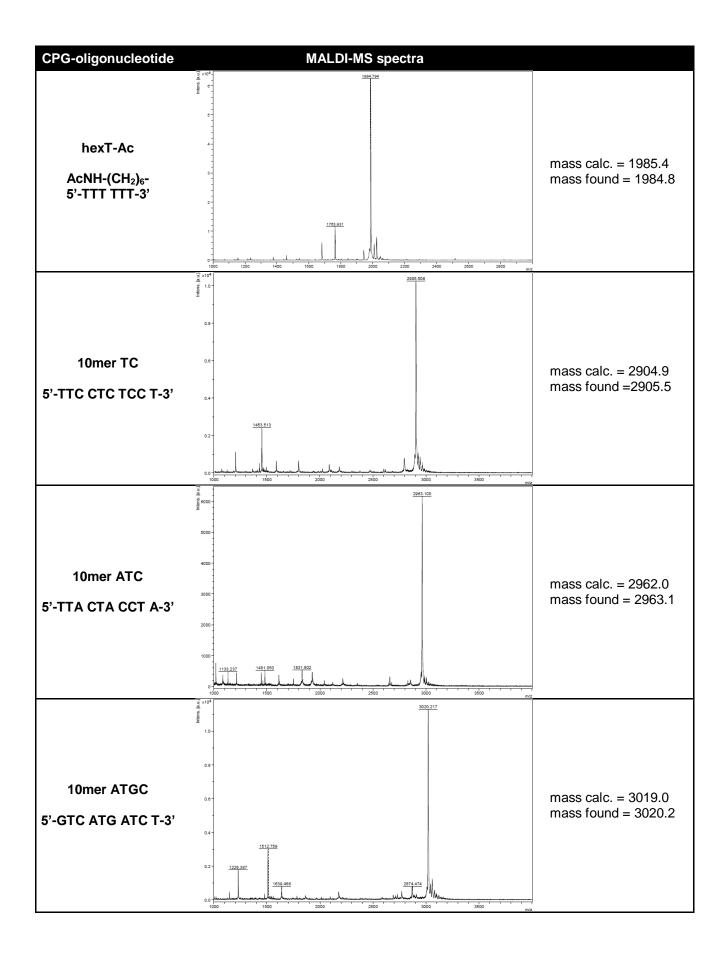




CPG-oligonucleotide + Mg(ClO₄)₂

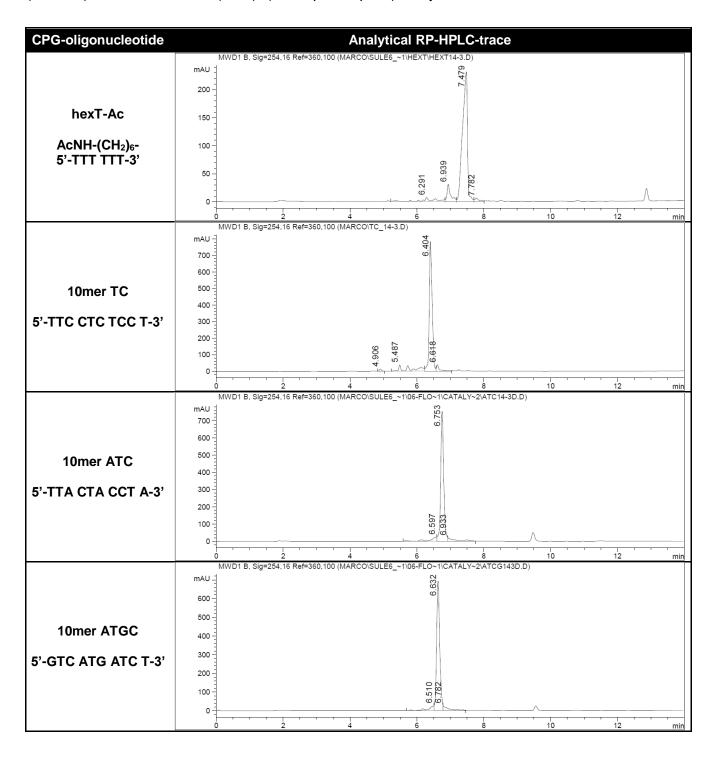
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Mg(ClO₄)₂ (200 equiv., 4 μ mol) in dry MeOH.

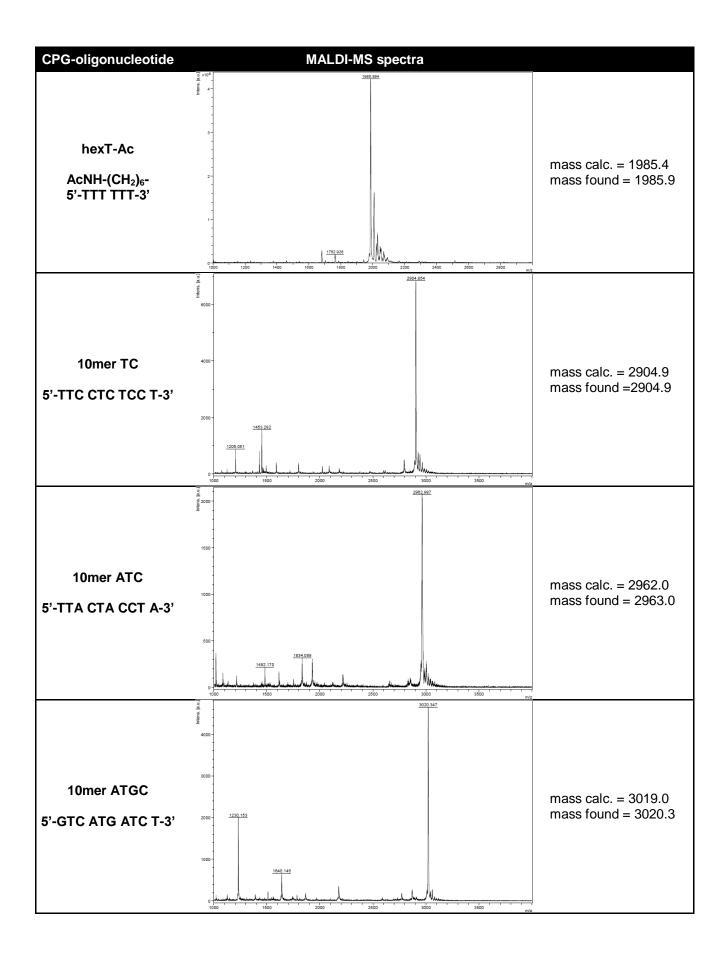




CPG-oligonucleotide + Ni(acac)₂

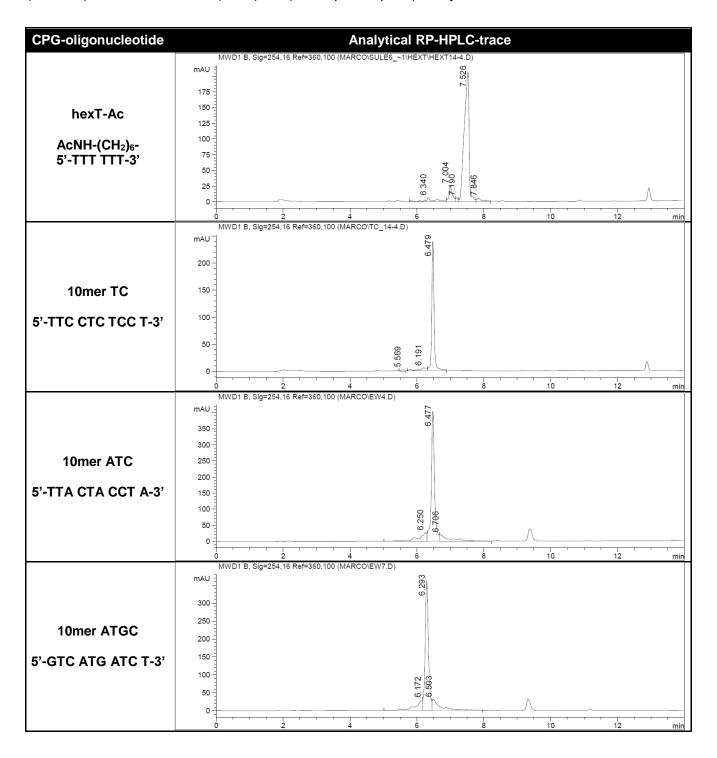
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Ni(acac)₂ (200 equiv., 4 µmol) in dry ACN.

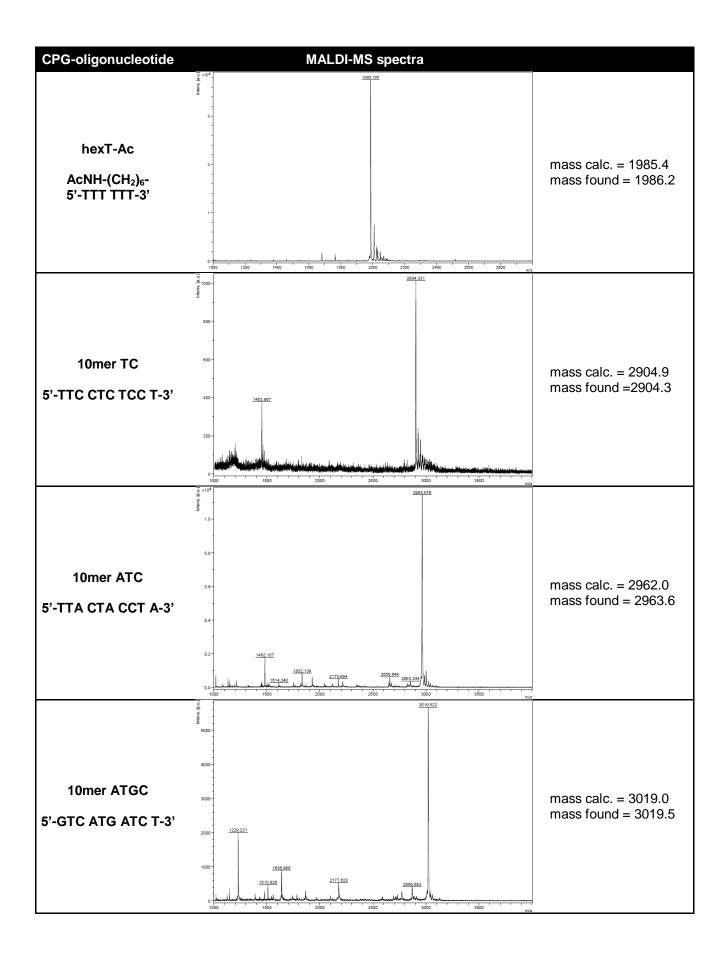




CPG-oligonucleotide + Ni(PPh₃)₂Cl₂

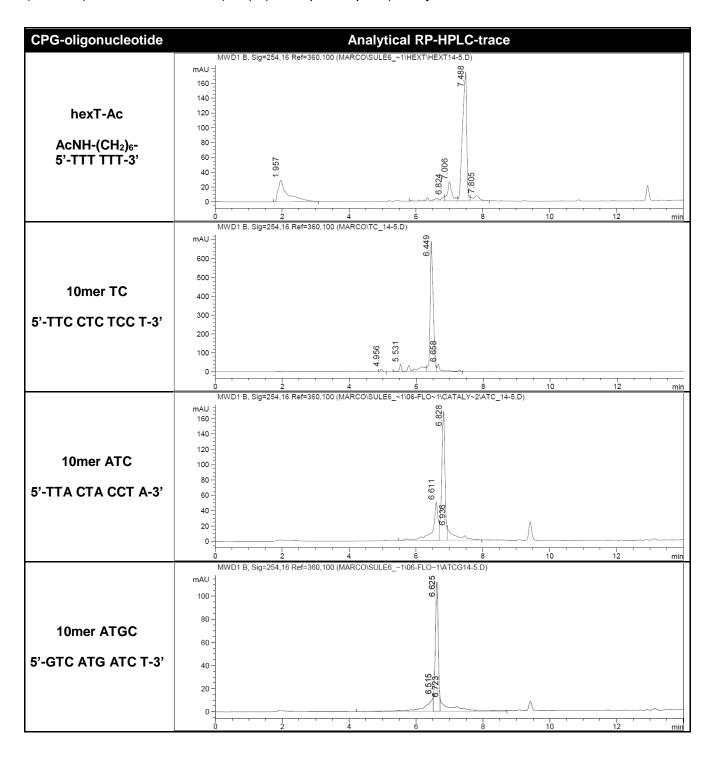
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Ni(PPh₃)₂Cl₂ (200 equiv., 4 µmol) in dry MeOH.

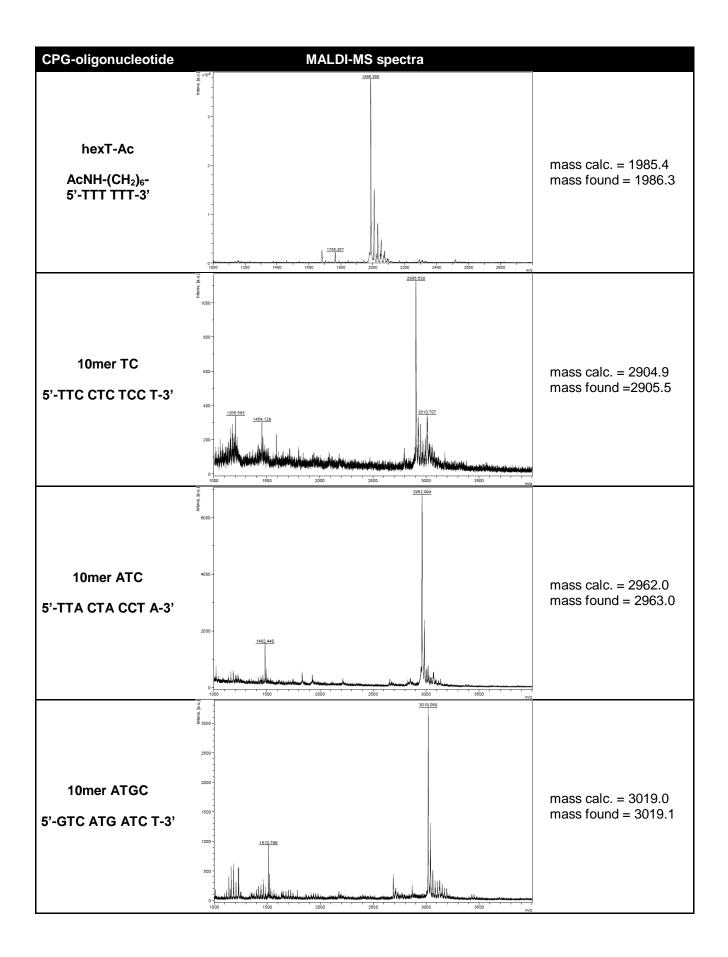




CPG-oligonucleotide + Pd(dba)₃

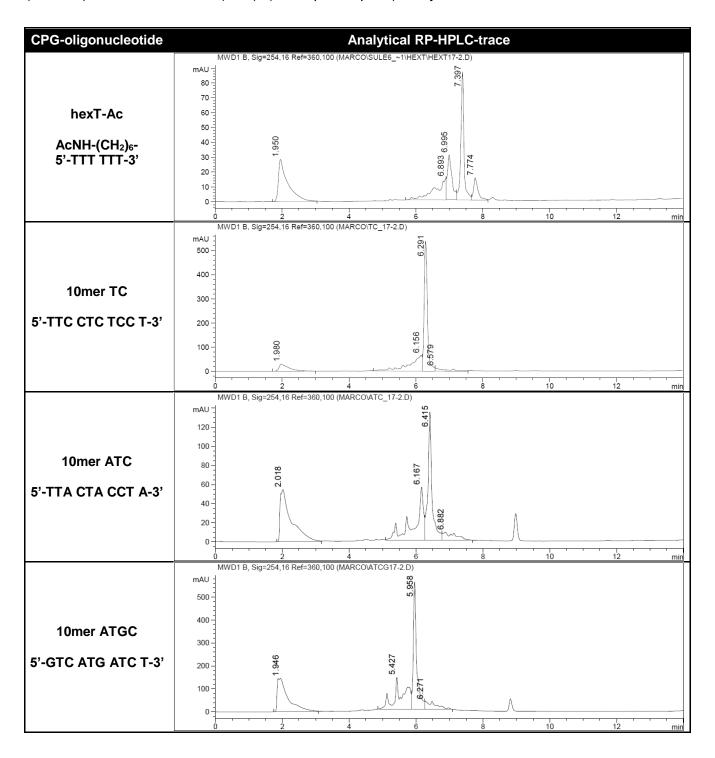
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Pd(dba)₃ (200 equiv., 4 µmol) in dry MeOH.

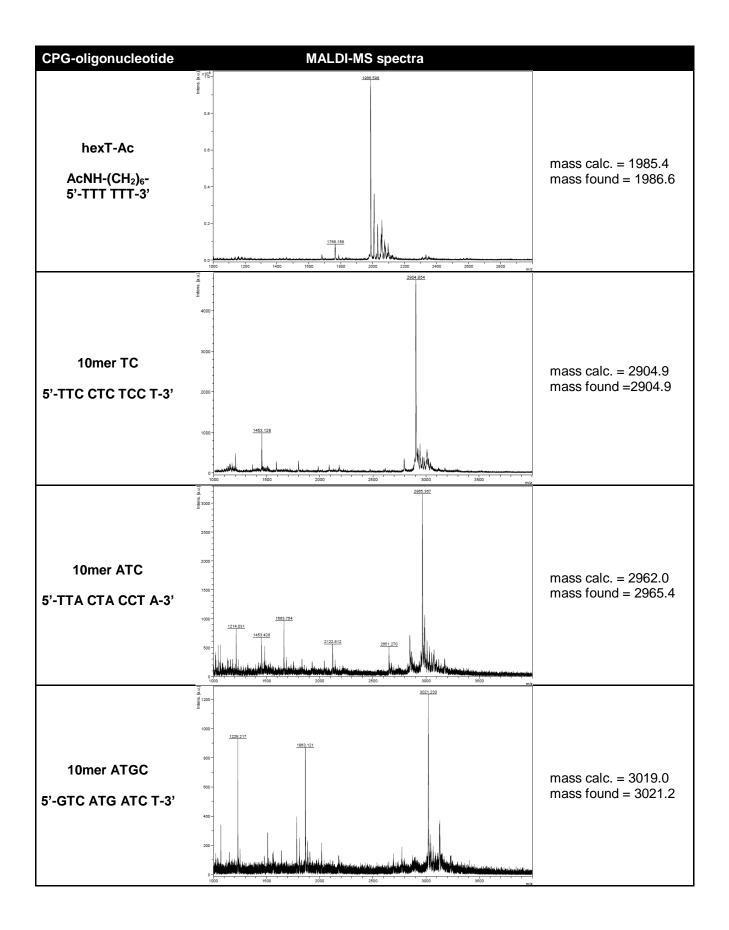




CPG-oligonucleotide + Pd(OAc)₂

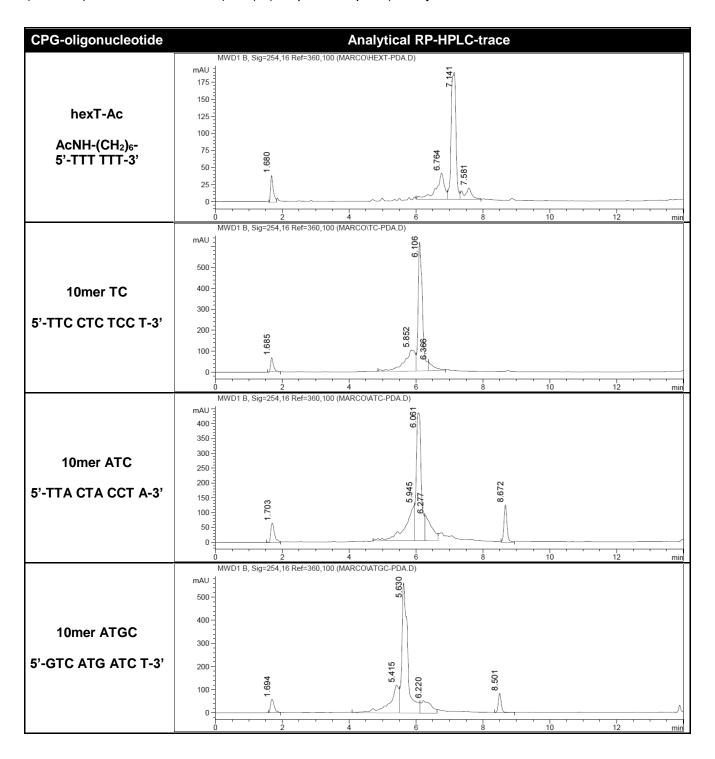
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Pd(OAc)₂ (200 equiv., 4 µmol) in dry ACN.

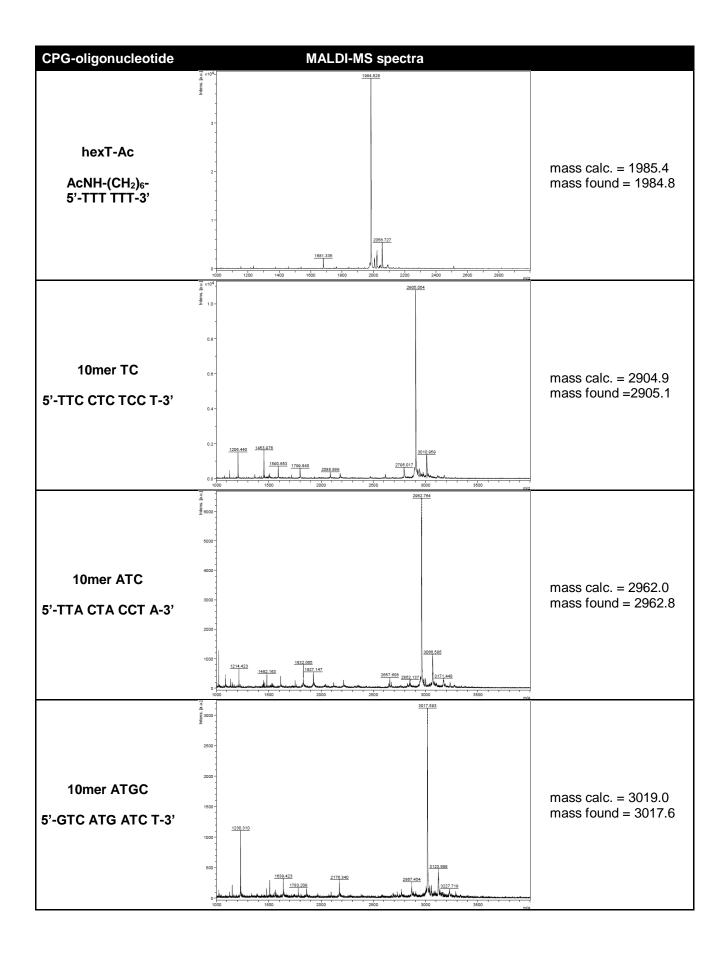




CPG-oligonucleotide + Pd(OAc)₂

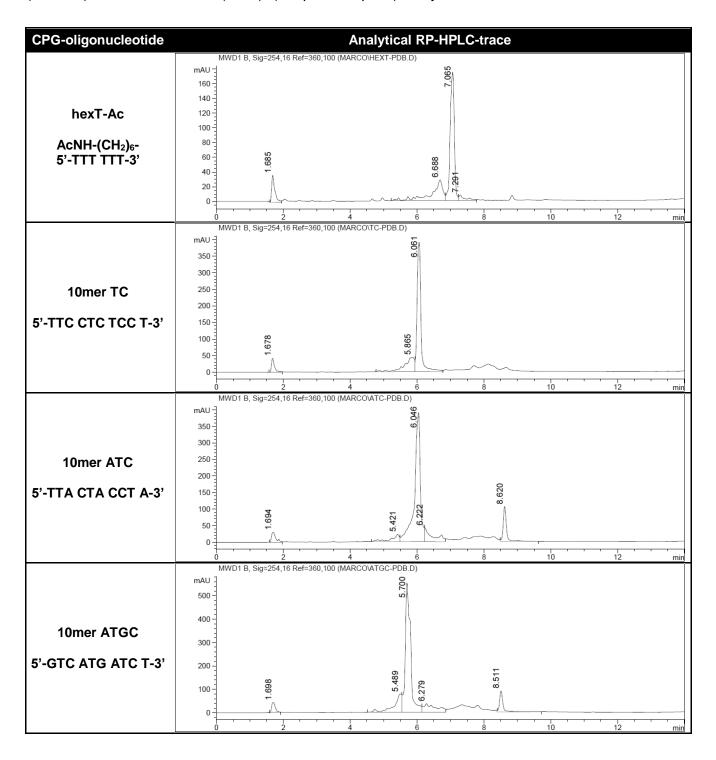
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Pd(OAc)₂ (5 equiv., 0.1 µmol) in dry ACN.

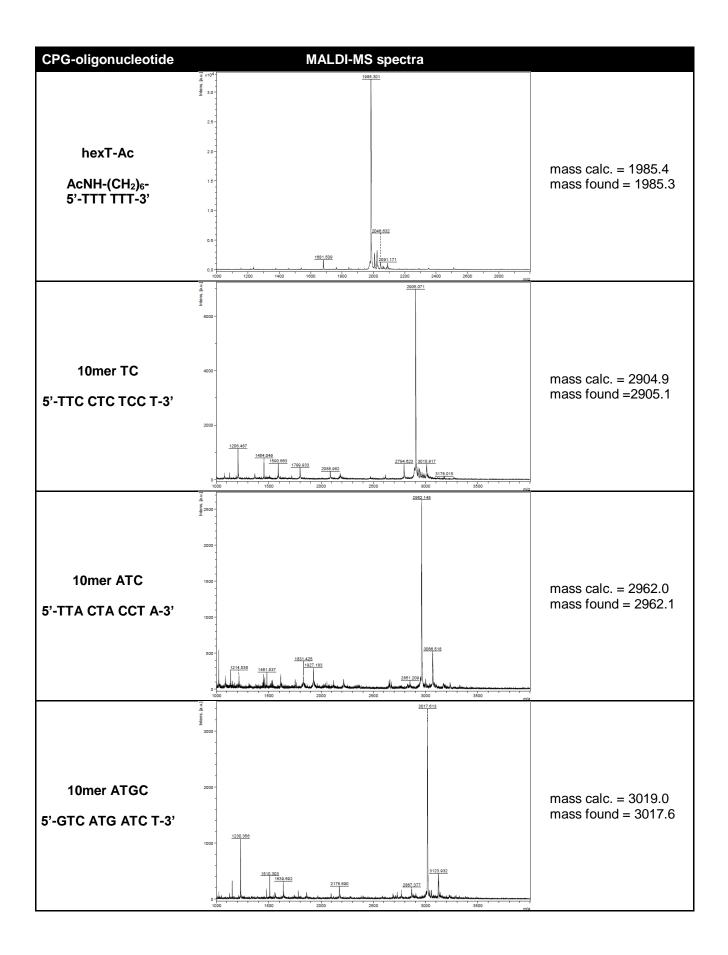




CPG-oligonucleotide + Pd(PPh₃)₄

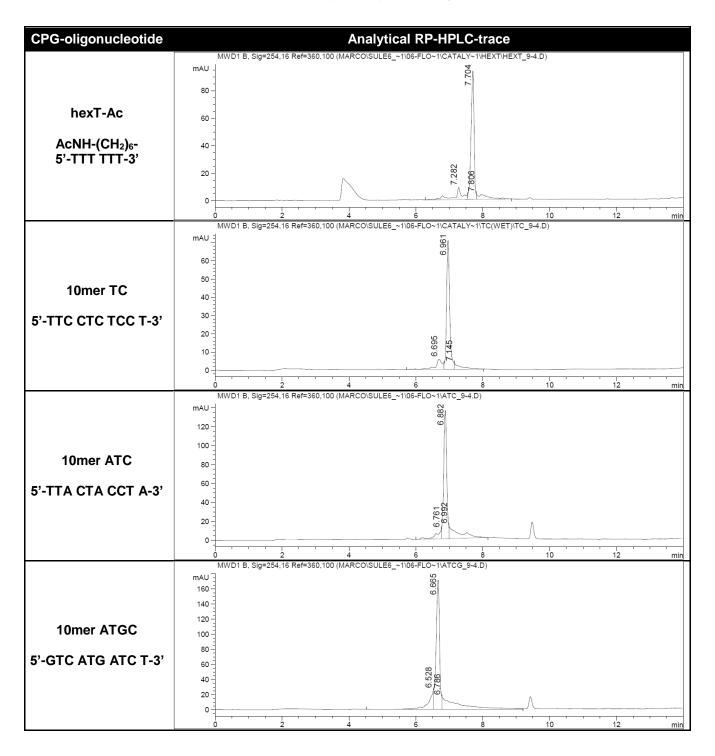
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with $Pd(PPh_3)_4$ (5 equiv., 0.1 µmol) in dry MeOH.

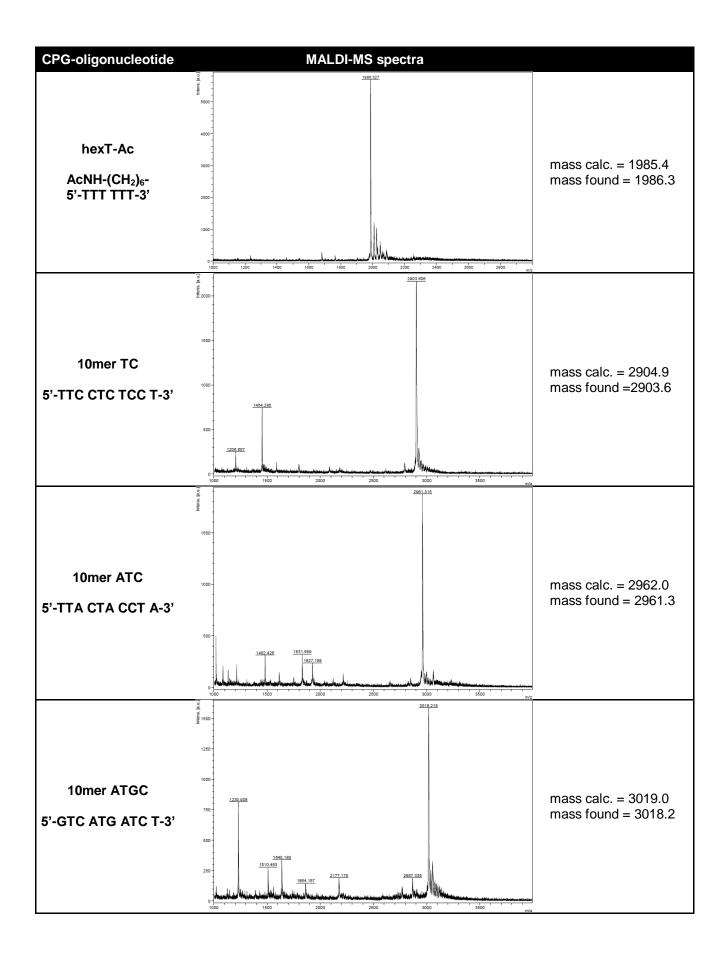




CPG-oligonucleotide + [Rh(cod)Cl]₂

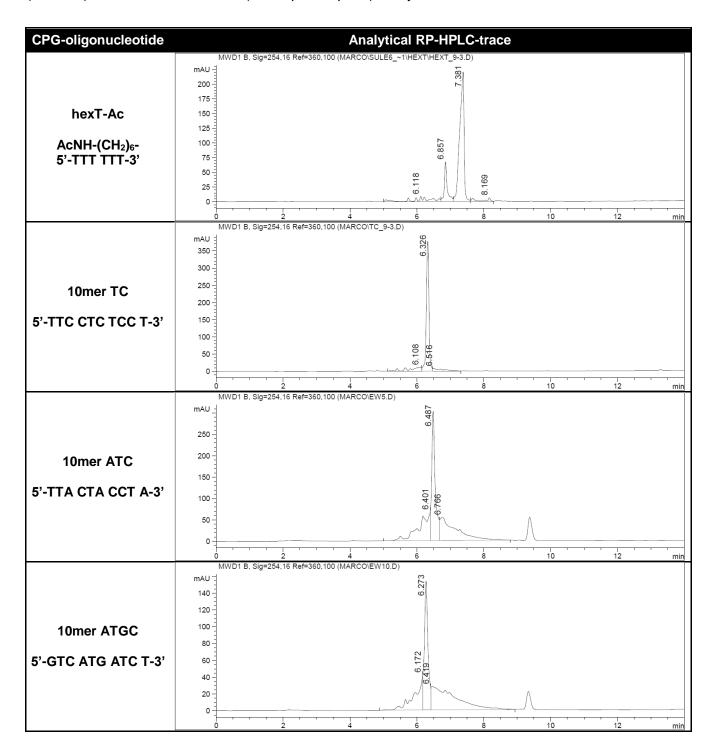
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with [Rh(cod)Cl]₂ (200 equiv., 4 µmol) in dry MeOH.

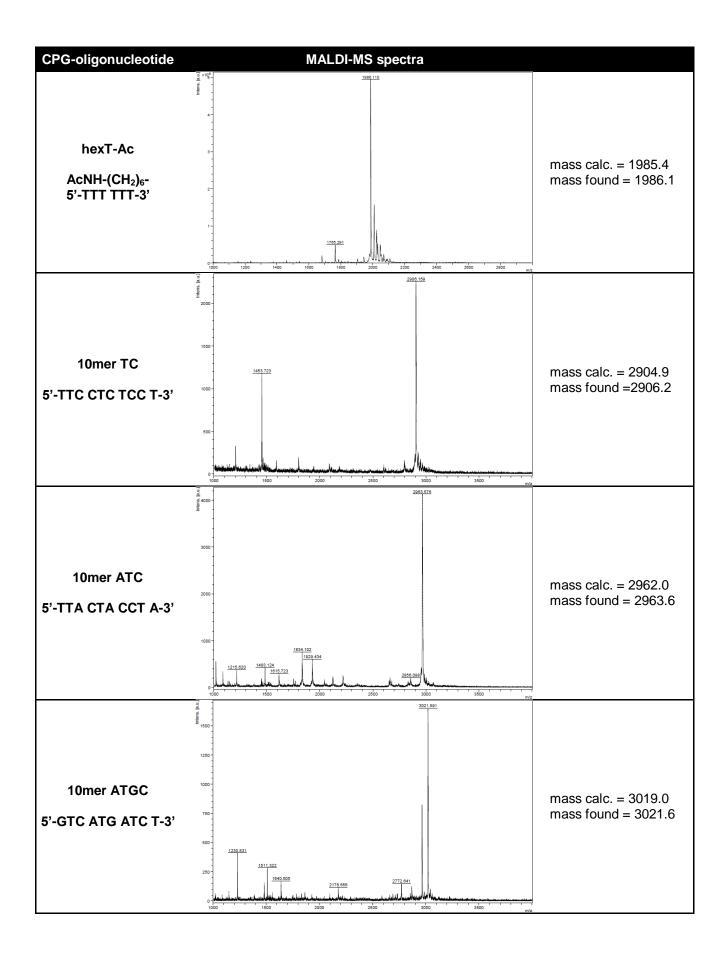




CPG-oligonucleotide + RuCl₃

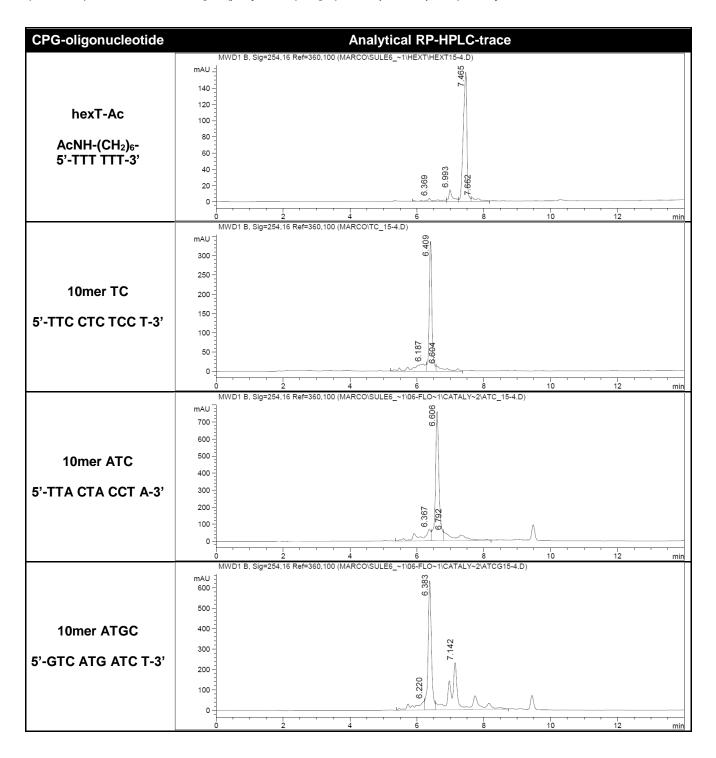
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with RuCl₃ (200 equiv., 4 µmol) in dry ACN.

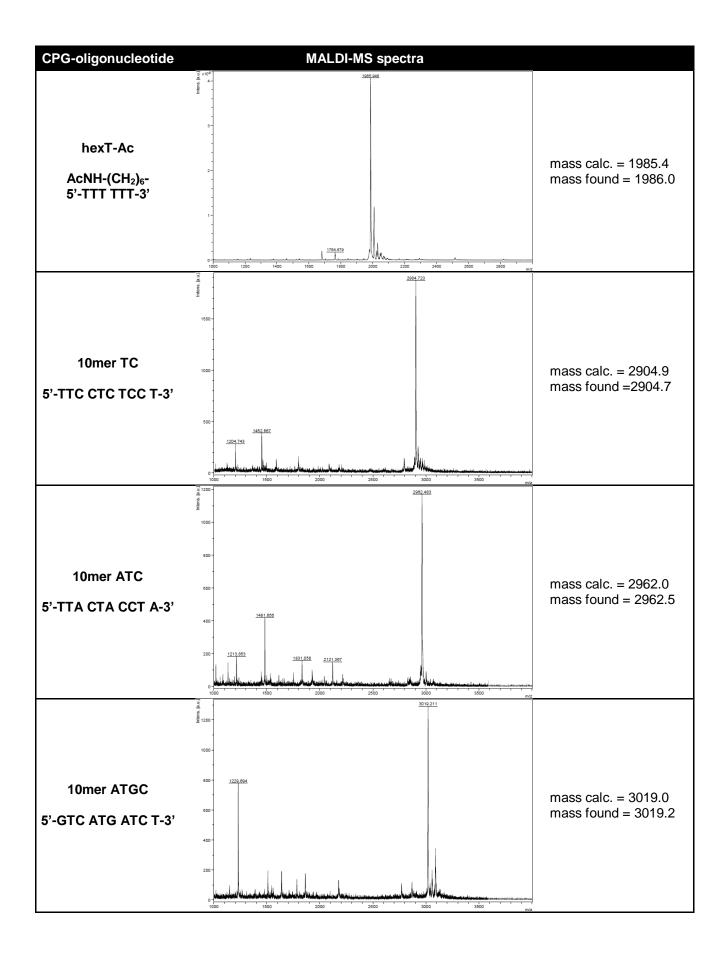




CPG-oligonucleotide + [Ru(p-cymene)Cl₂]₂

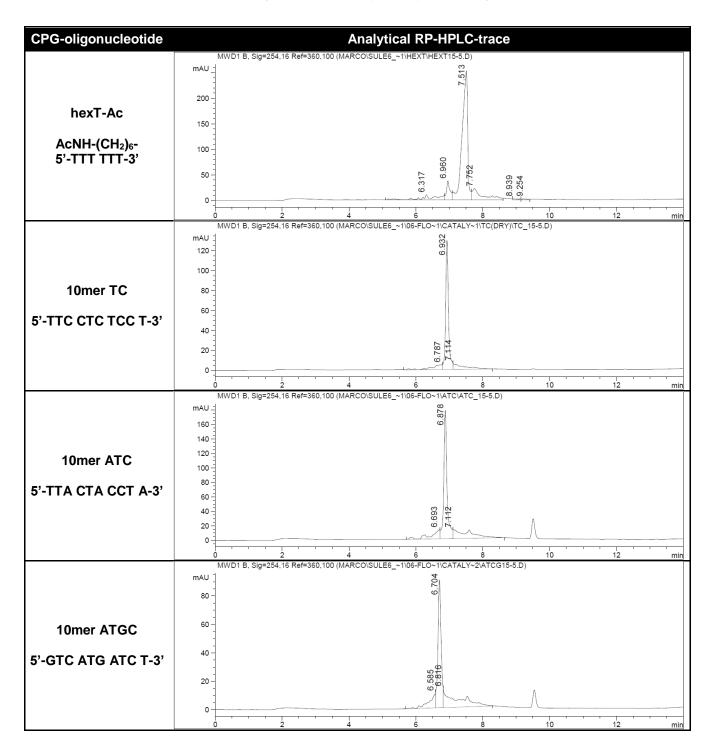
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with $[Ru(p-cymene)Cl_2]_2$ (200 equiv., 4 µmol) in dry CH_2Cl_2 .

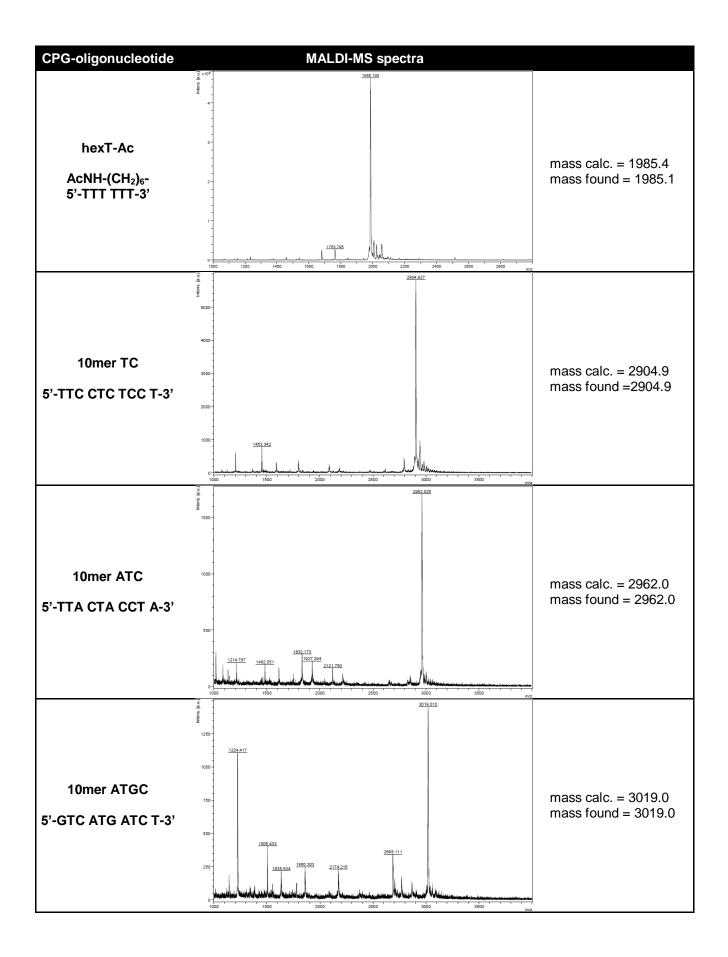




CPG-oligonucleotide + Ru(Me-allyl)₂(cod)

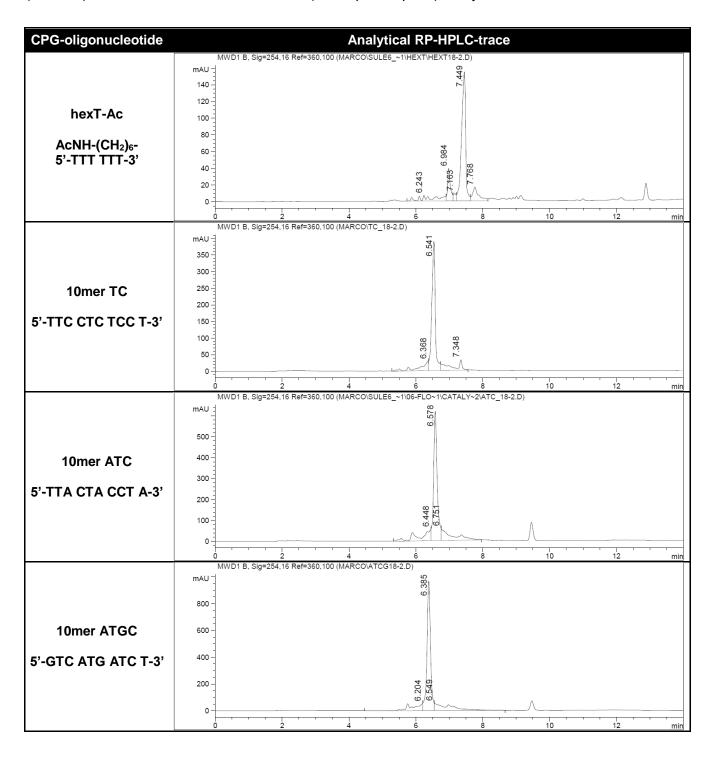
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Ru(Me-allyl)₂(cod) (200 equiv., 4 µmol) in dry CH₂Cl₂.

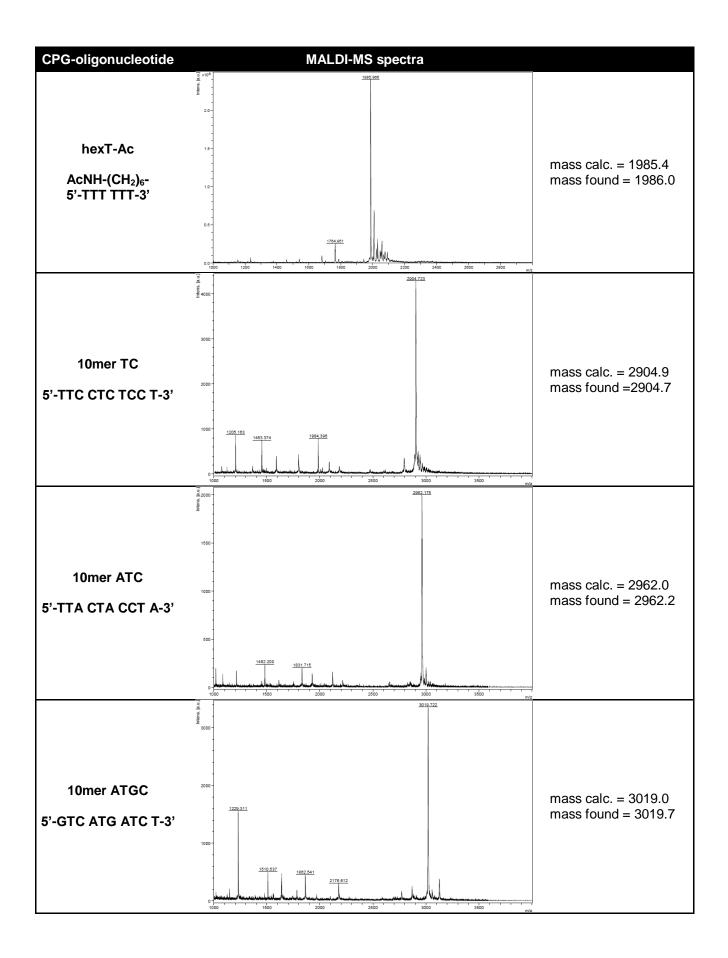




CPG-oligonucleotide + Grubbs 1st Gen.

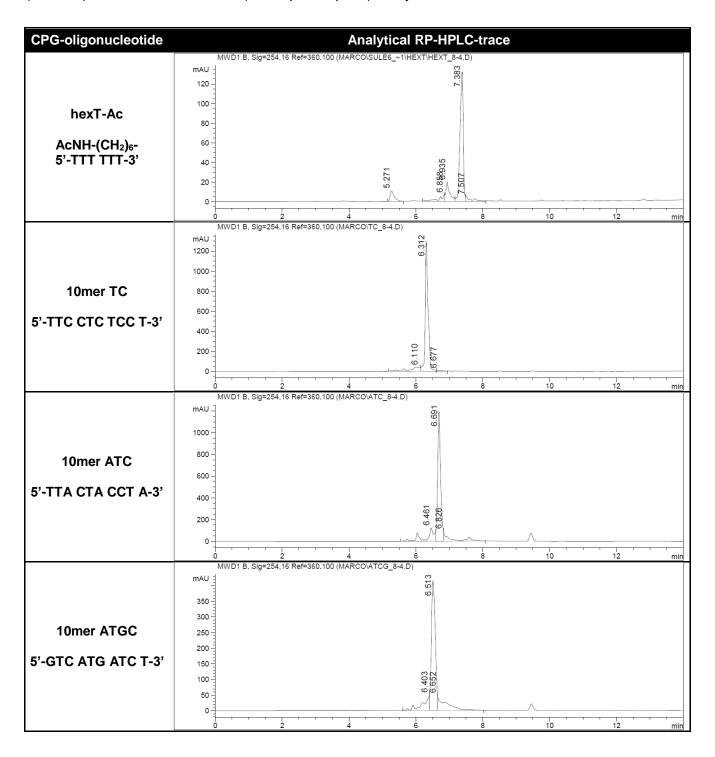
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Grubbs 1st Gen. (200 equiv., 4 µmol) in dry CH₂Cl₂.

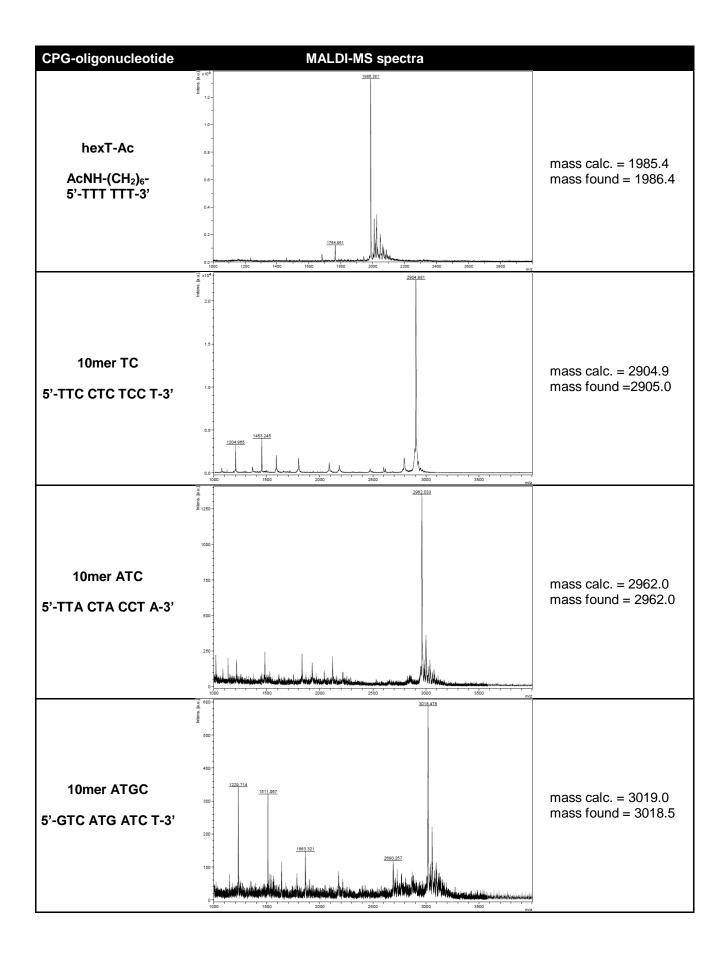




CPG-oligonucleotide + SbCl₃

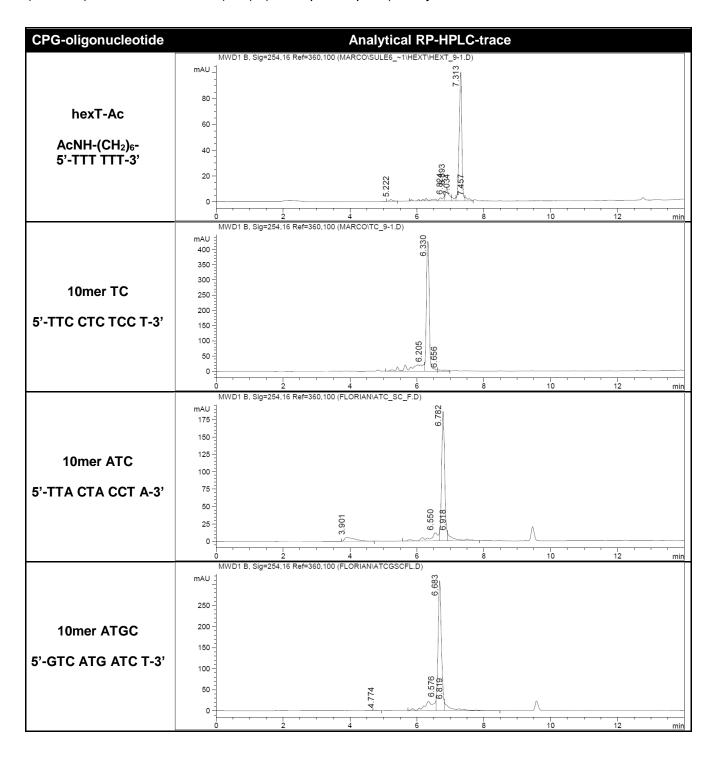
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with SbCl₃ (200 equiv., 4 µmol) in dry ACN.

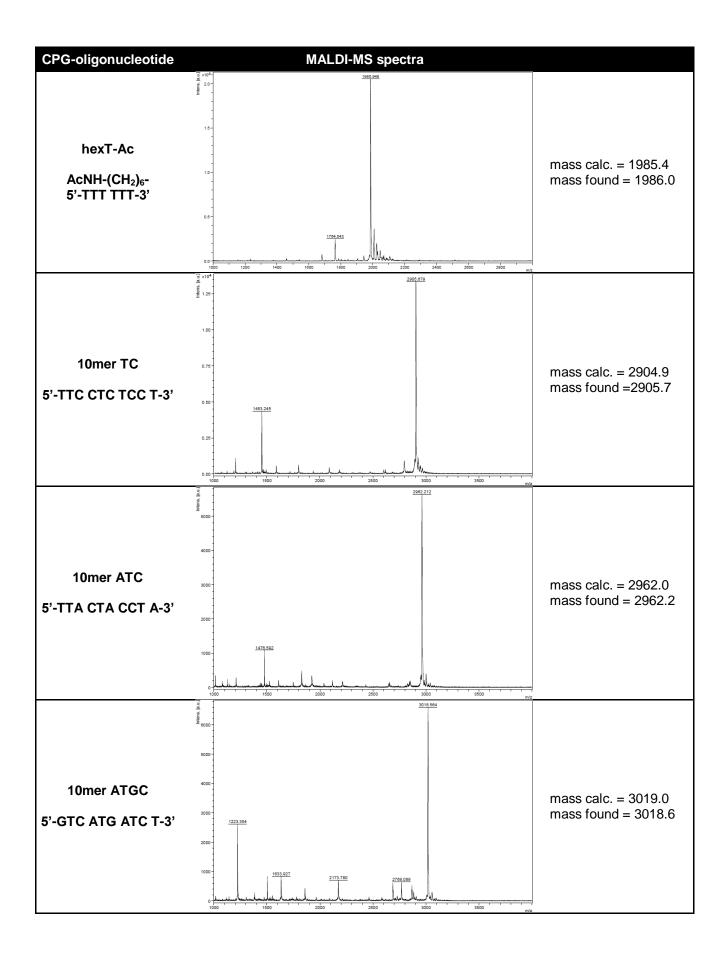




CPG-oligonucleotide + Sc(OTf)₃

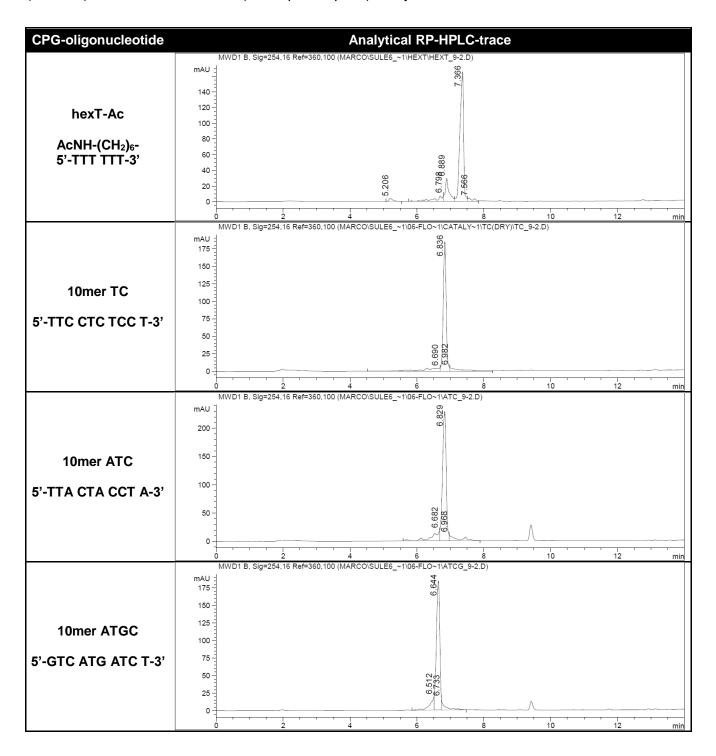
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with $Sc(OTf)_3$ (200 equiv., 4 µmol) in dry ACN.

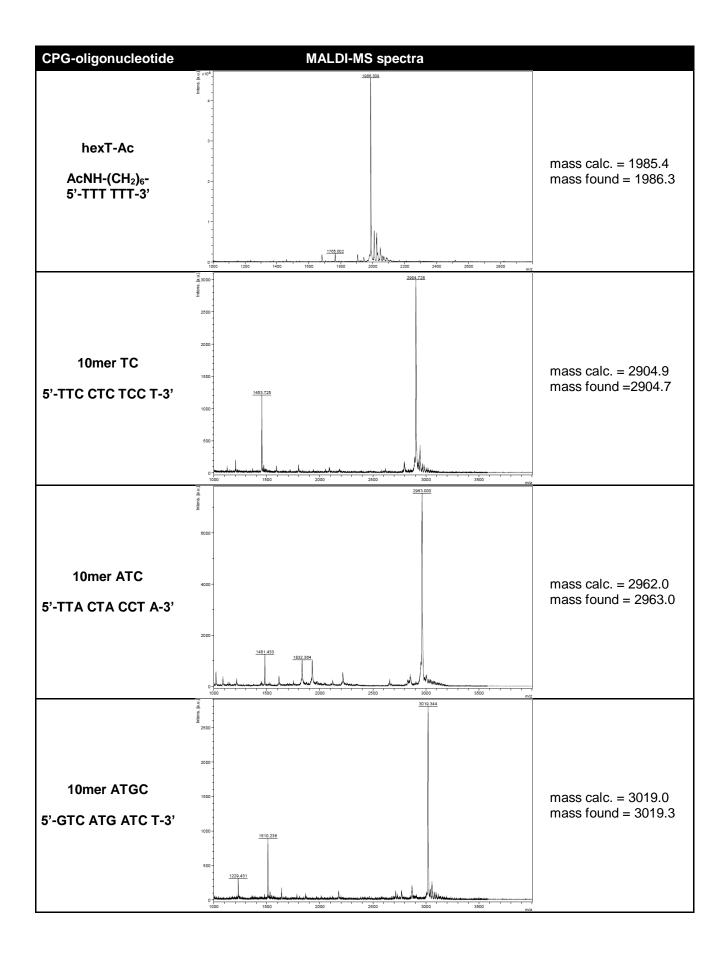




CPG-oligonucleotide + SeO₂

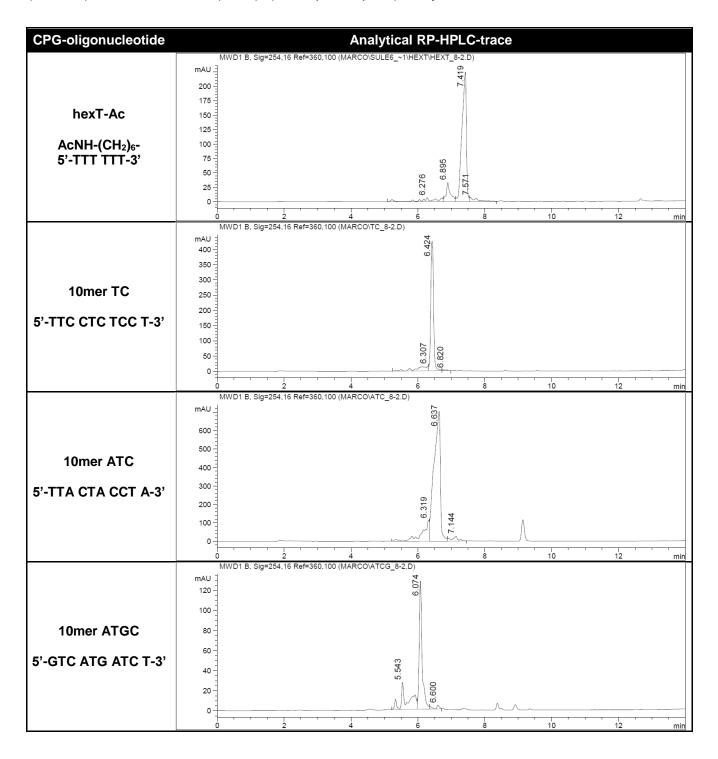
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with SeO₂ (200 equiv., 4 µmol) in dry MeOH.

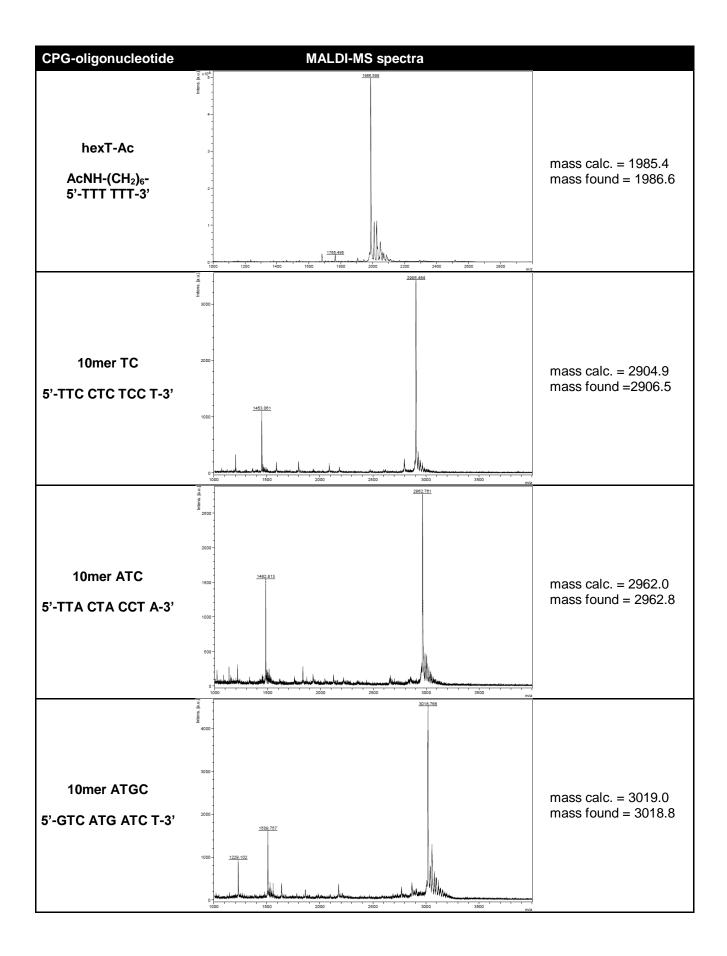




CPG-oligonucleotide + Ti(O*i*-Pr)₄

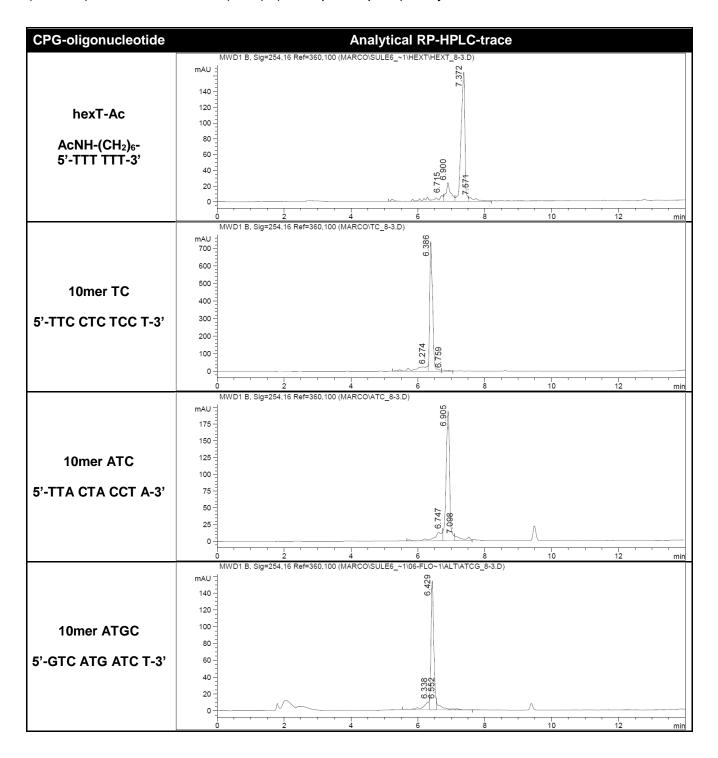
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Ti(O*i*-Pr)₄ (200 equiv., 4 µmol) in dry MeOH.

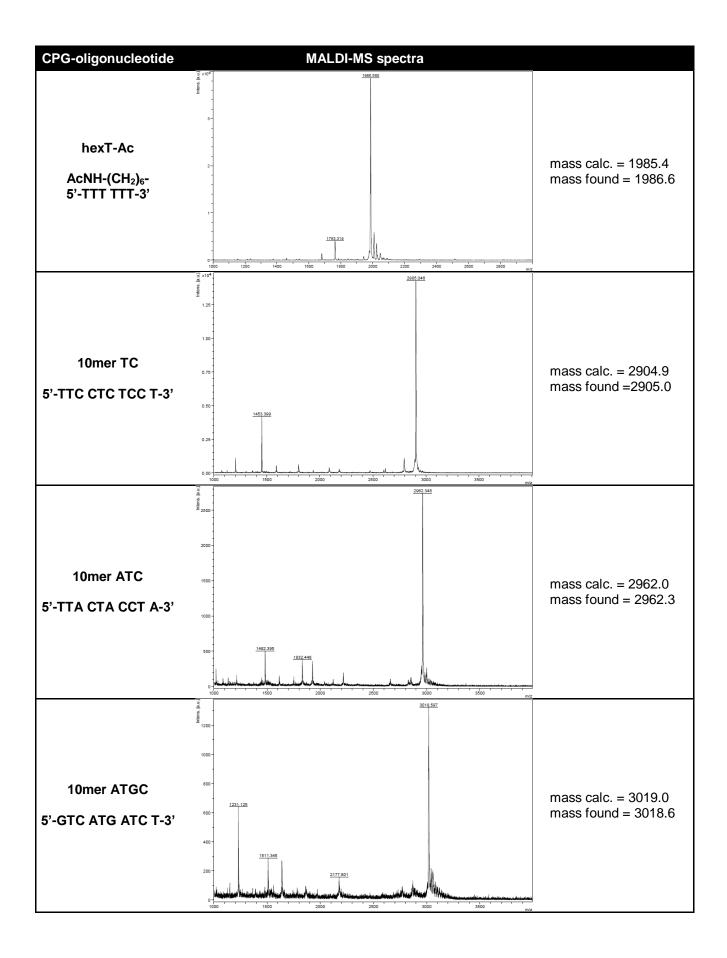




CPG-oligonucleotide + VO(acac)₂

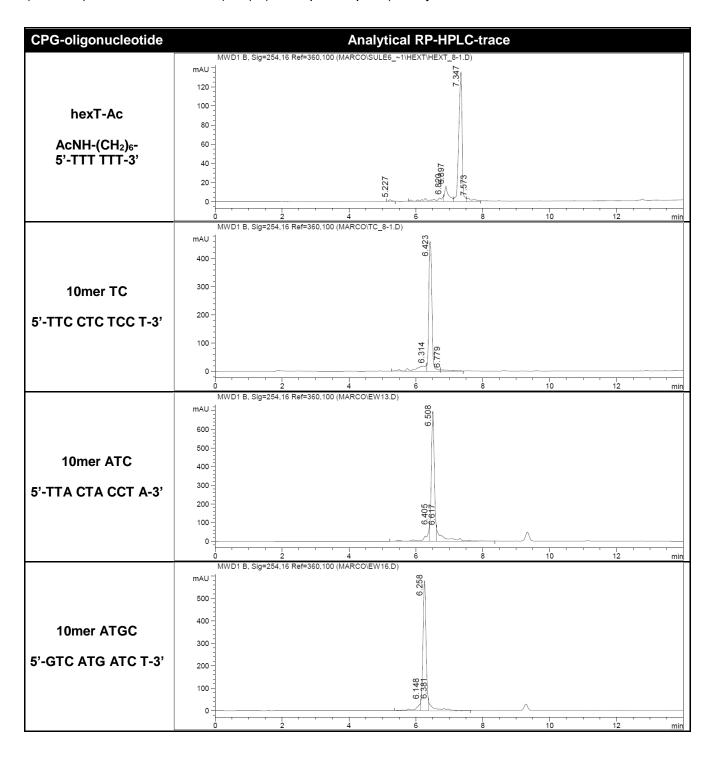
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with VO(acac)₂ (200 equiv., 4 µmol) in dry MeOH.

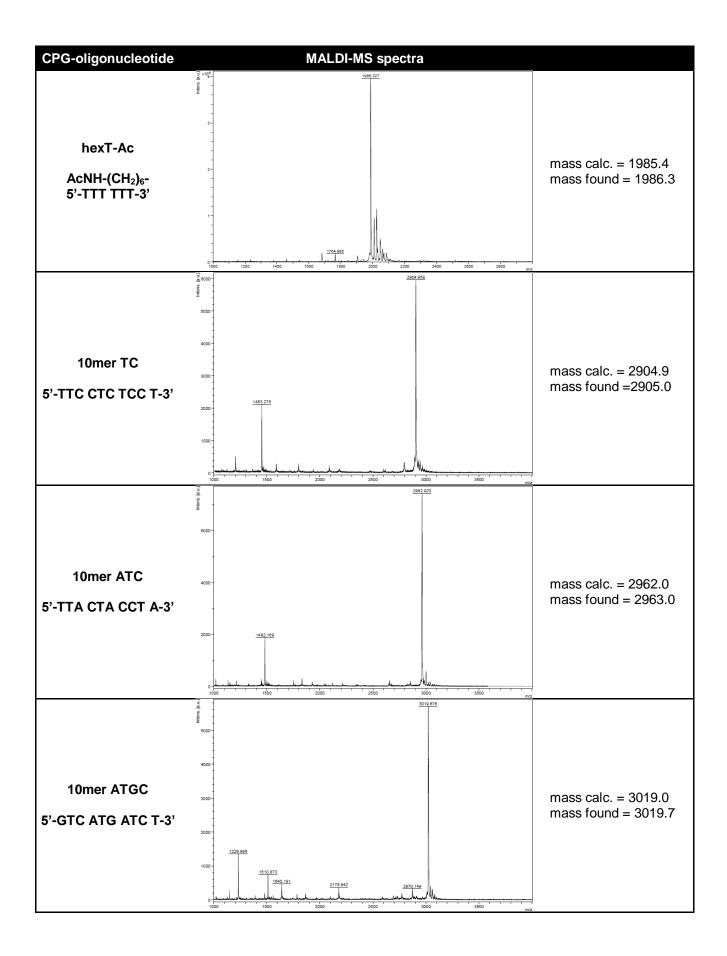




CPG-oligonucleotide + Yb(OTf)₃

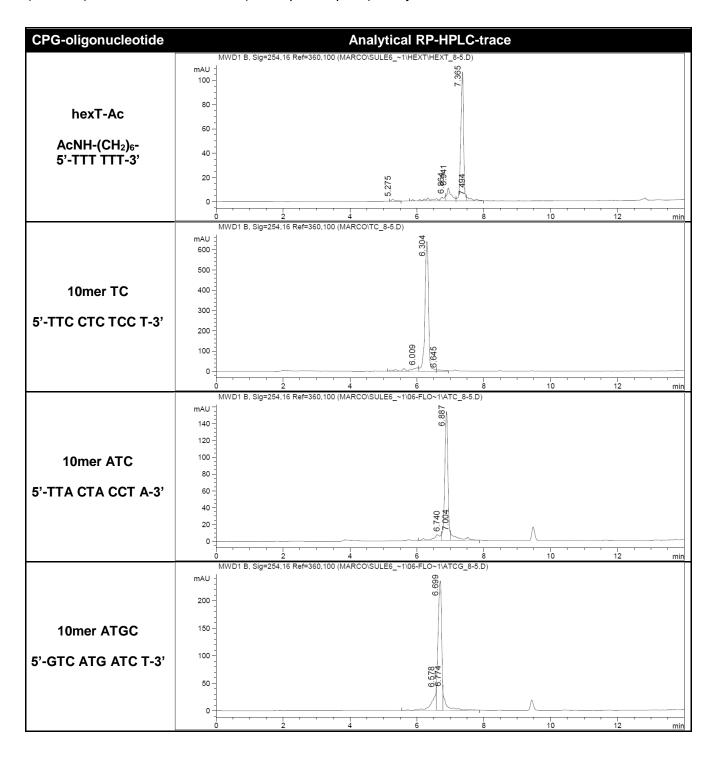
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Yb(OTf)₃ (200 equiv., 4 µmol) in dry MeOH.

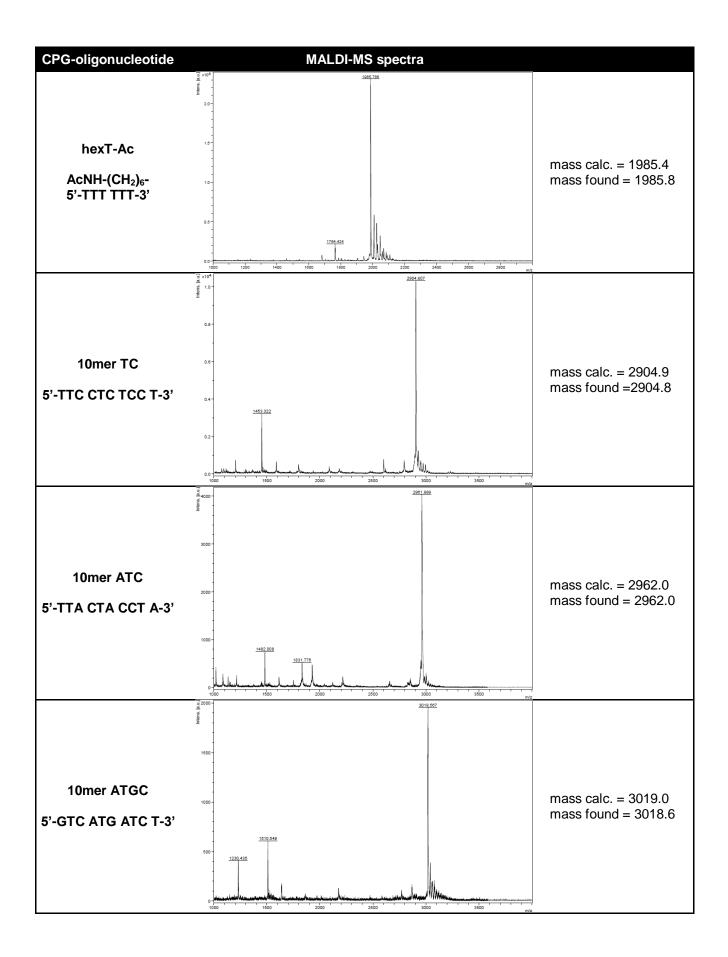




CPG-oligonucleotide + ZnCl₂

According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with ZnCl₂ (200 equiv., 4 µmol) in dry ACN.





TableS2 – Stability of DNA against metal salts at 40 °C^a

Entry	Metal salt	Solvent	ATC	ATCG
1	Cu(OTf) ₂	ACN		
2	InCl₃	ACN		
3	SeO ₂	MeOH		
4	Yb(OTf)₃	MeOH		
5	Sc(OTf) ₃	ACN		

^a for each: 20 nmol DNA, 200 eq. transition metal salt, 50 μL solvent (HPLC-grade), 40 °C., 22 h. ACN = acetonitrile, MeOH = methanol.

Entry	Metal salt	Solvent	ATC	ATCG
1	AgOTf	ACN		
2	BiBr₃	ACN		
3	InCl₃	ACN		
4	LiBr	ACN		
5	Ni(acac) ₂	ACN		
6	Yb(OTf) ₃	MeOH		

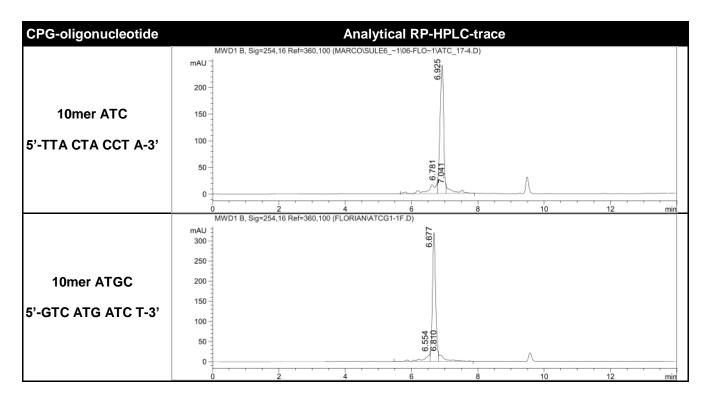
^a for each: 20 nmol DNA, 200 eq. transition metal salt, 50 µL solvent (HPLC-grade), r.t., 22 h. ACN = acetonitrile, MeOH = methanol.

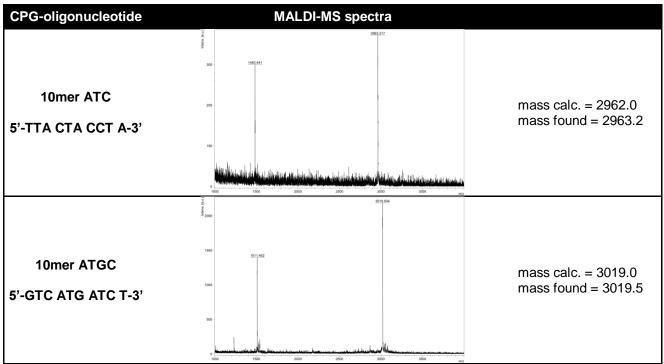
				DNA degradation
0-20%	21-40%	41-60%	> 61%	DNA degradation

HPLC traces and MALDI-MS spectra of metal ion screens

CPG-oligonucleotide + Cu(OTf)₂

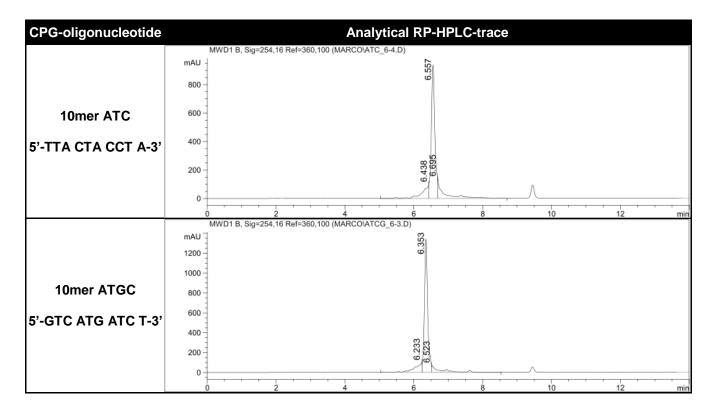
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Yb(OTf)₃ (200 equiv., 4 µmol) in dry ACN at 40 °C.

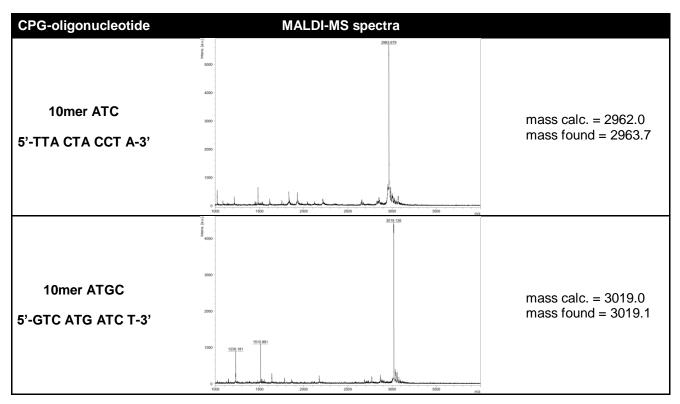




CPG-oligonucleotide + InCl₃

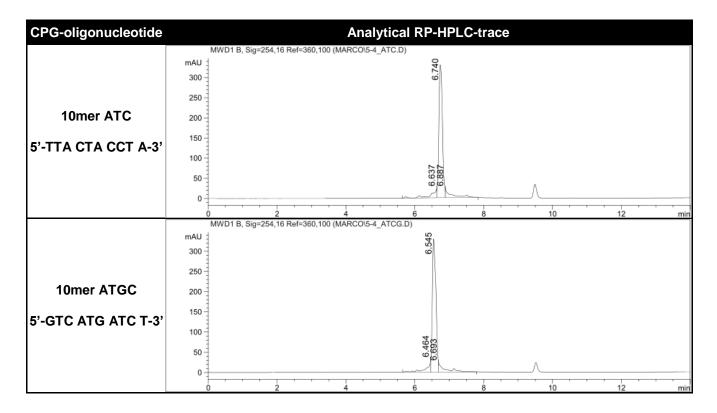
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with InCl₃ (200 equiv., 4 µmol) in dry ACN at 40 °C.

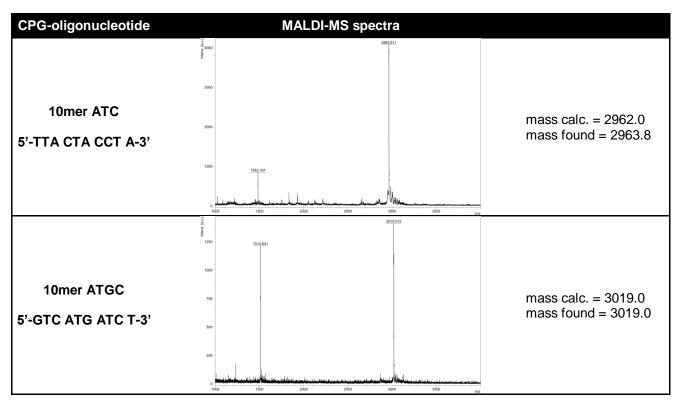




CPG-oligonucleotide + SeO₂

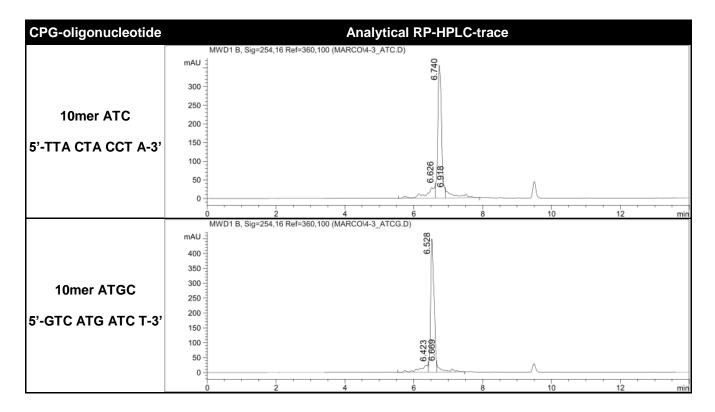
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with SeO₂ (200 equiv., 4 µmol) in dry MeOH at 40 °C.

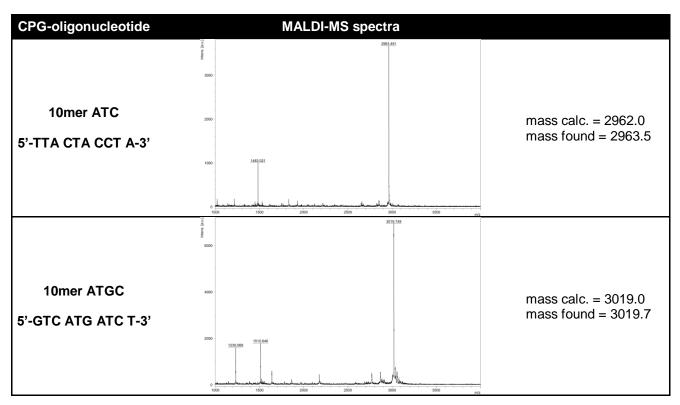




CPG-oligonucleotide + Yb(OTf)₃

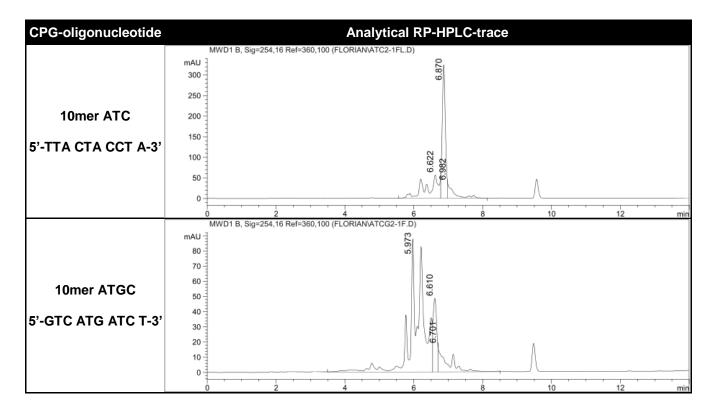
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Yb(OTf)₃ (200 equiv., 4 µmol) in dry MeOH at 40 °C.

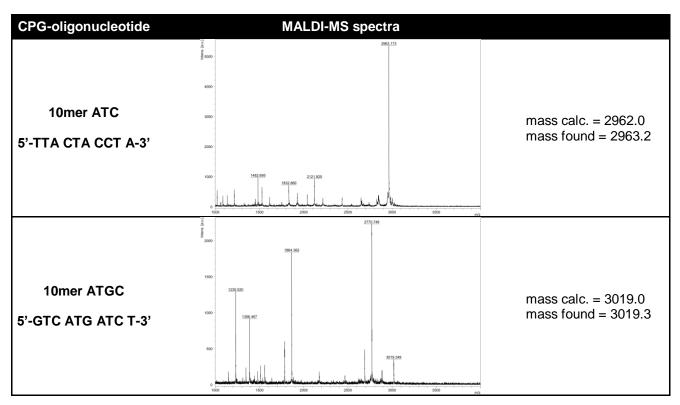




CPG-oligonucleotide + Sc(OTf)₃

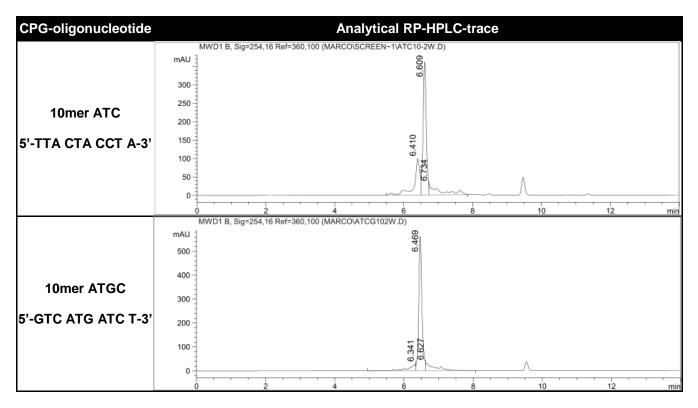
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with $Sc(OTf)_3$ (200 equiv., 4 µmol) in dry ACN at 40 °C.

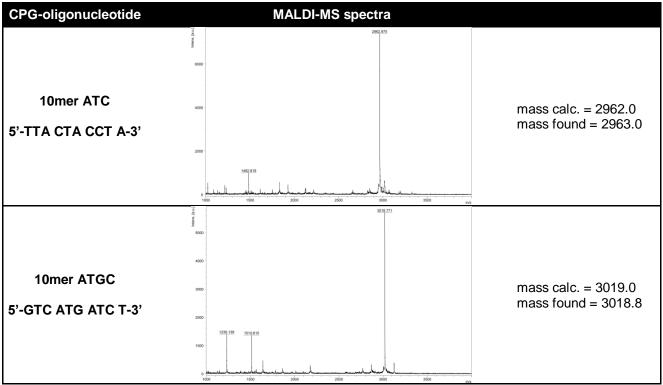




CPG-oligonucleotide + AgOTf

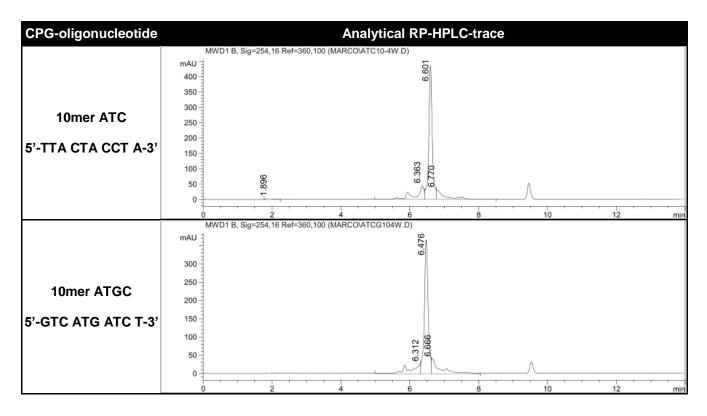
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with AgOTf (200 equiv., 4 µmol) in HPLC-grade ACN.

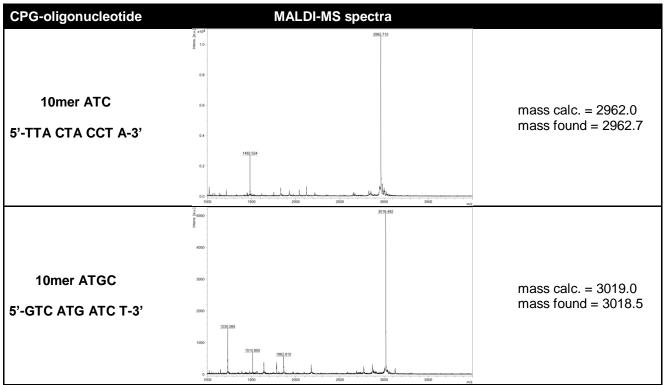




CPG-oligonucleotide + BiBr₃

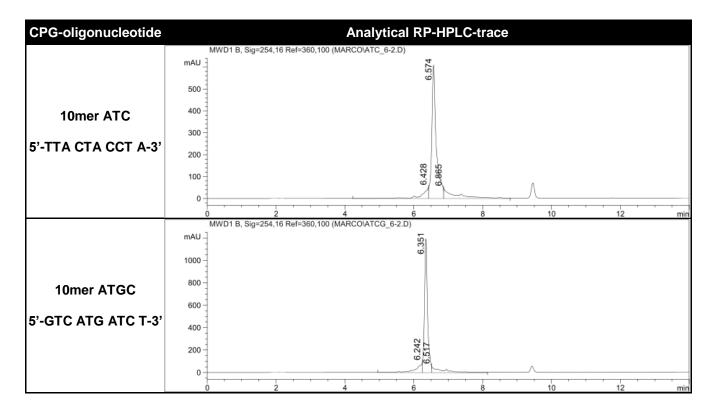
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with BiBr₃ (200 equiv., 4 µmol) in HPLC-grade ACN.

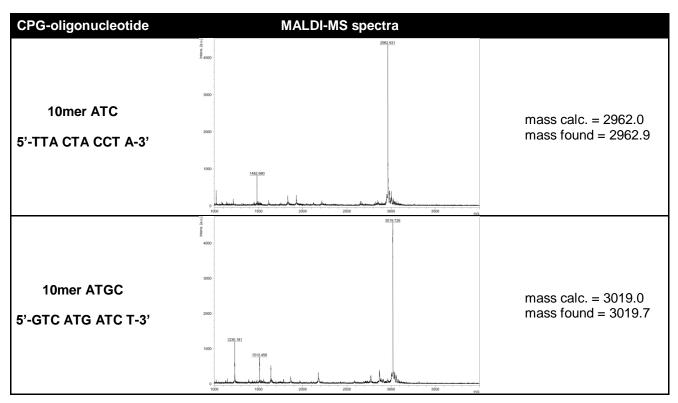




CPG-oligonucleotide + InCl₃

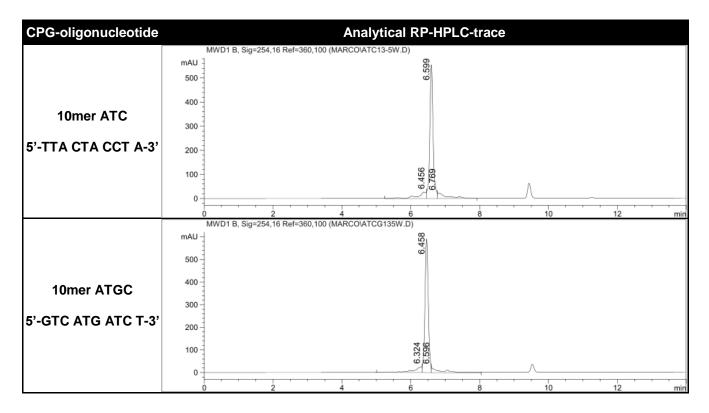
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with InCl₃ (200 equiv., 4 µmol) in HPLC-grade ACN.

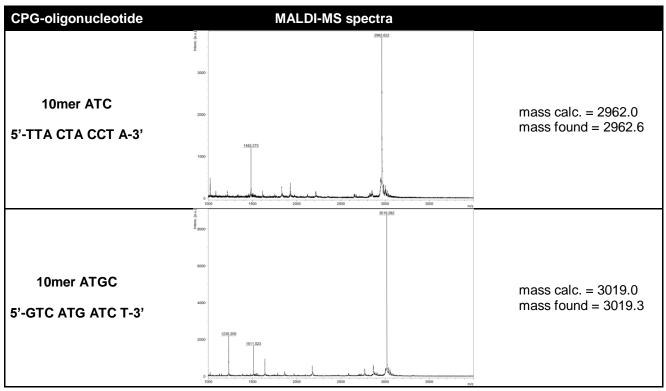




CPG-oligonucleotide + LiBr

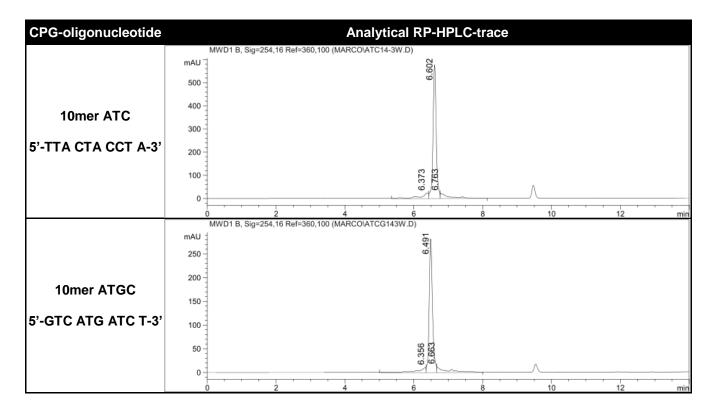
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with LiBr (200 equiv., 4 µmol) in HPLC-grade ACN.

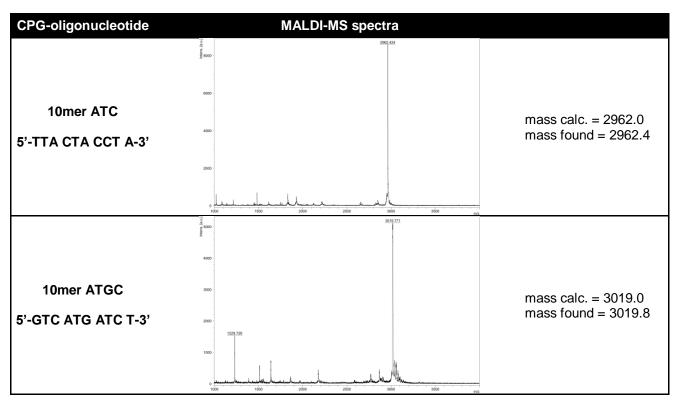




CPG-oligonucleotide + Ni(acac)₂

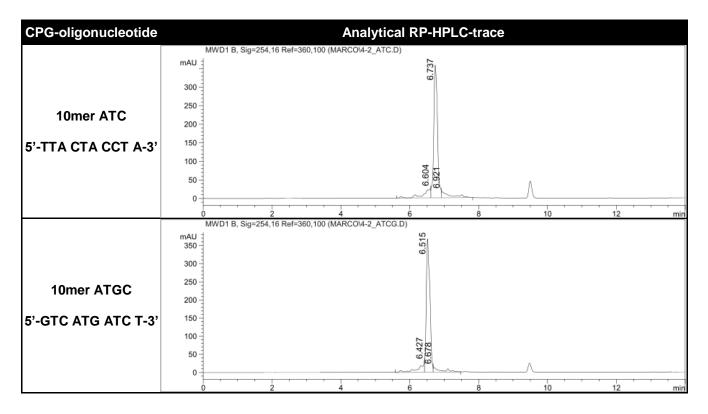
According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Ni(acac)₂ (200 equiv., 4 µmol) in HPLC-grade ACN.

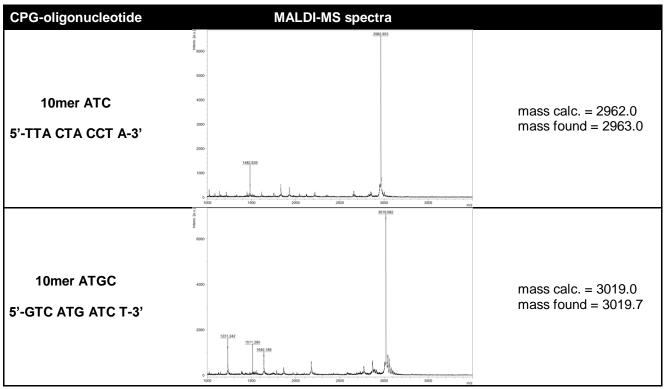




CPG-oligonucleotide + Yb(OTf)₃

According to the representative procedure (RP-03) solid support coupled oligonucleotide (20 nmol) was treated with Yb(OTf)₃ (200 equiv., 4 µmol) in HPLC-grade MeOH.





Entry ^[a]	Organo Catalyst	No.	Solvent	hexT	тс	ATC	ATCG
1	$\overset{\mathbb{H}_2}{\underset{()}{\overset{\mathbb{H}_2}}{\overset{\mathbb{H}_2}{\overset{\mathbb{H}_2}{\overset{\mathbb{H}_2}{\overset{\mathbb{H}_2}{\overset{\mathbb{H}_2}}{\overset{\mathbb{H}_2}{\overset{\mathbb{H}_2}{\overset{\mathbb{H}_2}}{\overset{\mathbb{H}_2}{\overset{\mathbb{H}_2}}{\overset{\mathbb{H}_2}}{\overset{\mathbb{H}_2}}{\overset{\mathbb{H}_2}}{\overset{\mathbb{H}_2}{\overset{\mathbb{H}_2}{\overset{\mathbb{H}_2}}{\overset{\mathbb{H}_2}{\overset{\mathbb{H}_2}}{\overset{\mathbb{H}_2}}{\overset{\mathbb{H}_2}}}{\overset{\mathbb{H}_2}}{\overset{\mathbb{H}_2}}}{\overset{\mathbb{H}_2}}}}}}}}}}}}}}}}}}}}}}}}}}}}}}}}}}}$	I	MeOH				
2	ð ð	J	ACN				
3		A	MeOH				
4	PPh ₂ PPh ₂	К	MeOH				
5	Ph Ph OH OH Ph Ph	В	MeOH				
6	OH C C C C C C C C C C C C C C C C C C C	С	MeOH				
7	H N N N N N N N N N N N N N N N N N N N	L	MeOH				

 Table S4 – Stability of DNA against organocatalysts and organic reagents^a

8		М	MeOH		
9	O NH OH	D	ACN		
10	∑N Ph N H H H	N	ACN		
11	NH OTMS	E	MeOH		
12		0	MeOH		
13		Ρ	MeOH		
14	C C C C C C C C C C C C C C C C C C C	Q	ACN		
15	OH NH ₂ NH ₂ OH	R	MeOH		

16	Ph- N N N N Ph	F	ACN		
17	F ₃ C F ₃ C	S	MeOH		
18		т	EtOH		
19	O. N	G	ACN		
20	i o i	н	ACN		

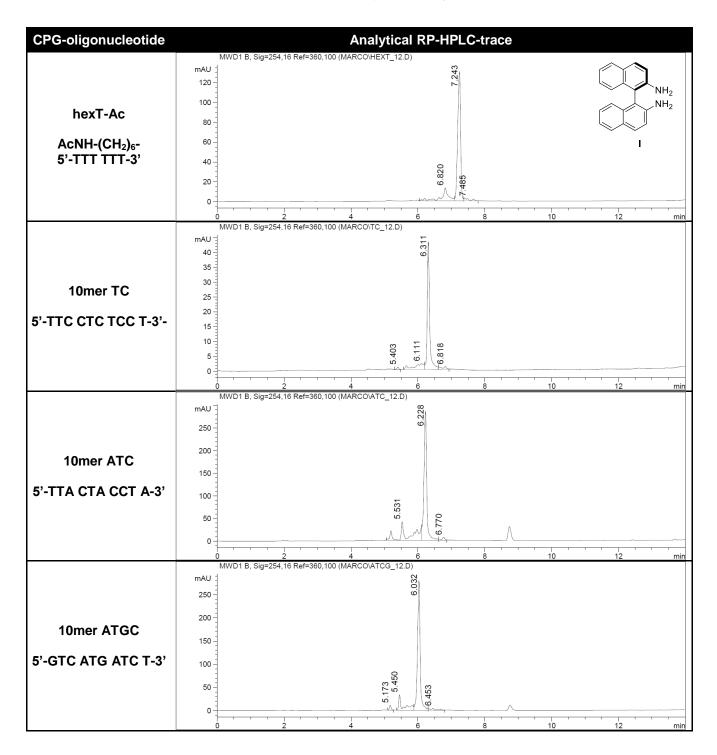
^a for each: 20 nmol DNA, 200 eq. organo catalyst, 50 µL solvent, r.t., 22 h; EtOH = ethanol, ACN = acetonitrile, MeOH = methanol.

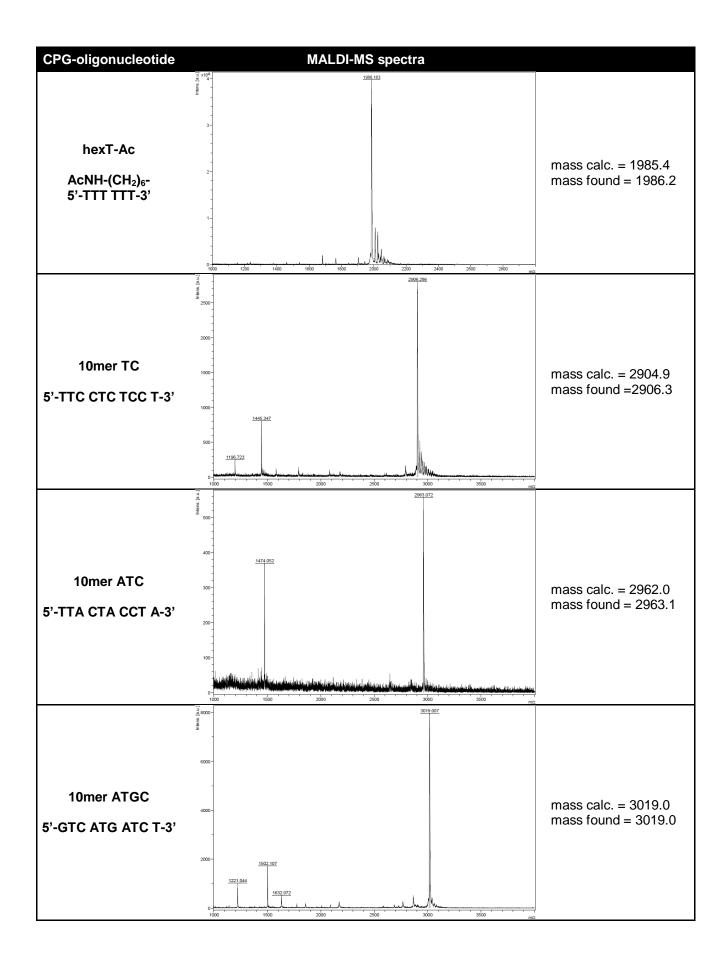
				DNA degradation
0-20%	21-40%	41-60%	> 61%	DNA degradation

HPLC traces and MALDI-MS spectra of organocatalyst and organic reagents screens

CPG-oligonucleotide + (R)-(+)-DABN I

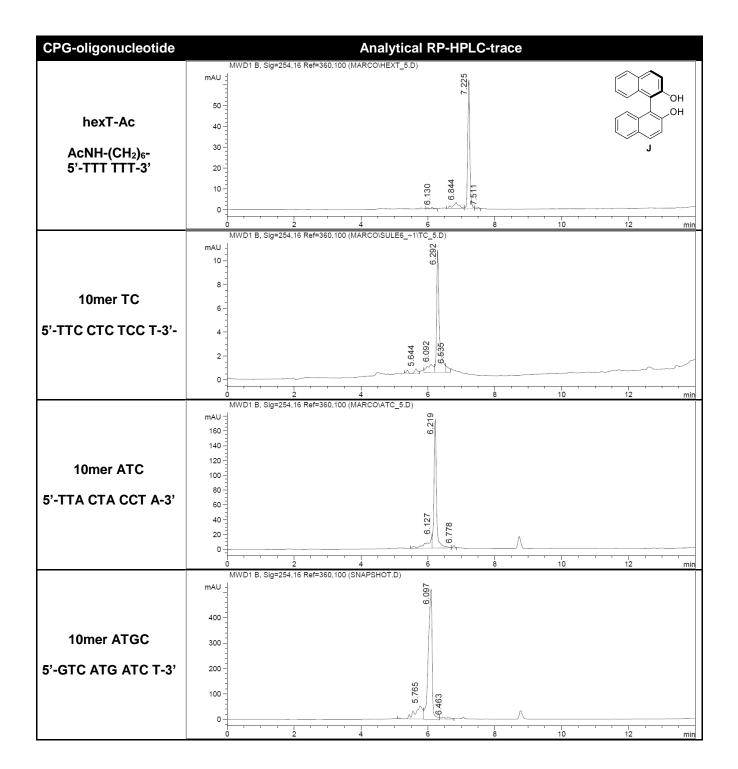
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with (R)-(+)-DABN I (200 equiv., 4 µmol) in dry MeOH.

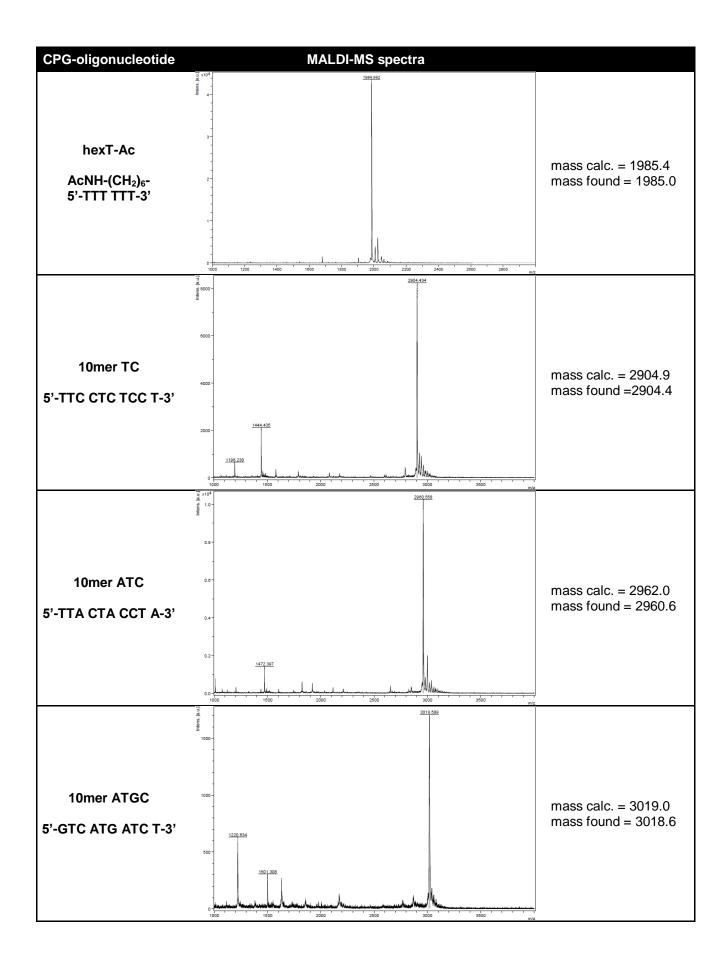




CPG-oligonucleotide + (*R*)-BINOL J

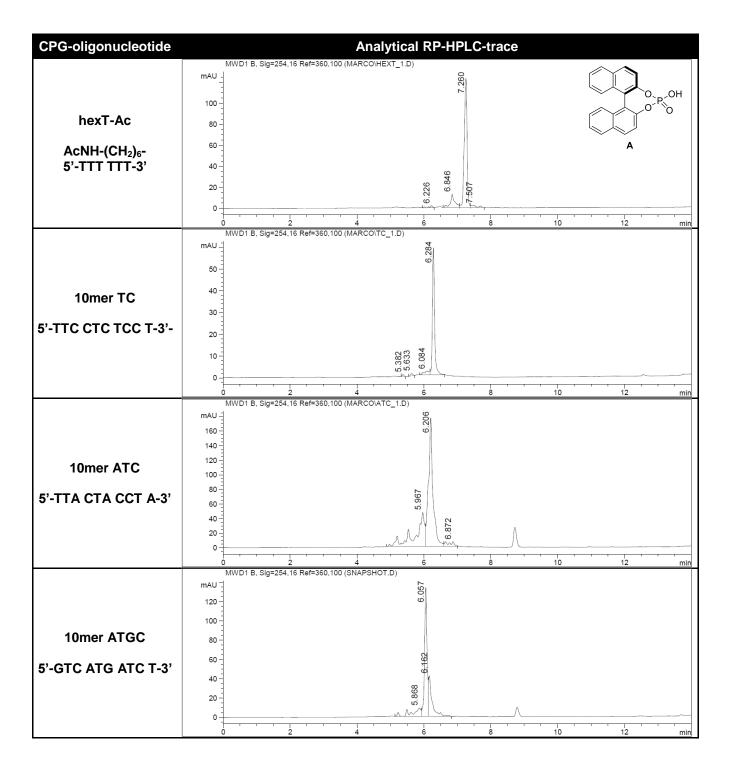
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with ((R)-BINOL J (200 equiv., 4 µmol) in dry ACN.

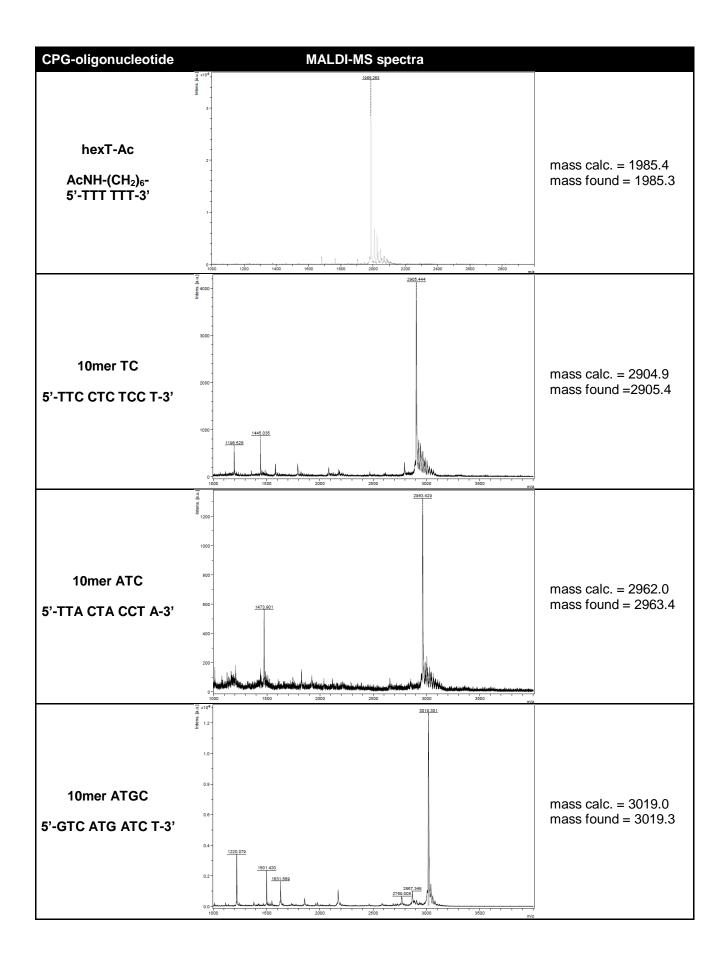




CPG-oligonucleotide + (R)-1,1-Binaphthyl-2,2-diyl hydrogenphosphate A

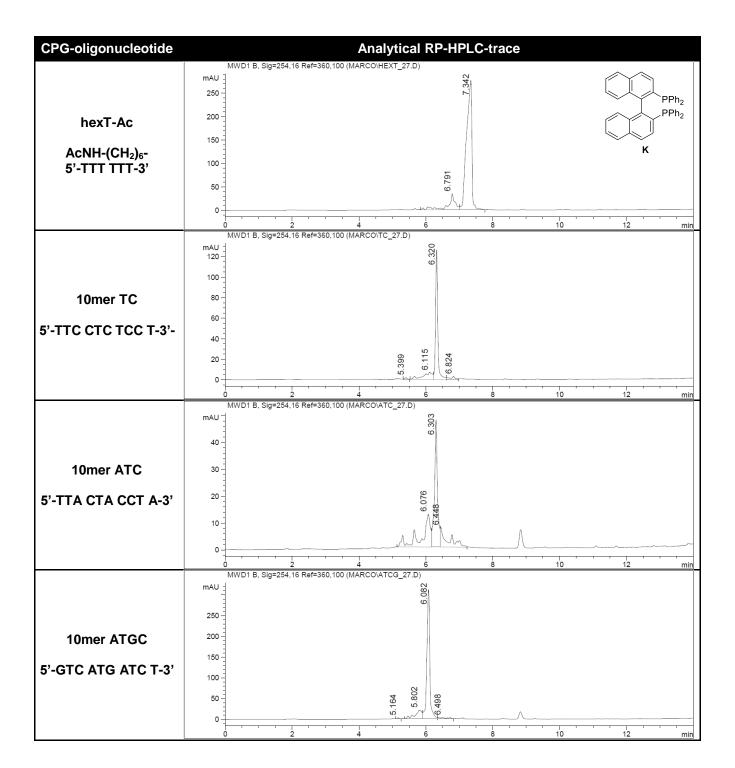
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with (*R*)-1,1-Binaphthyl-2,2-diyl hydrogenphosphate **A** (200 equiv., 4 μ mol) in dry MeOH.

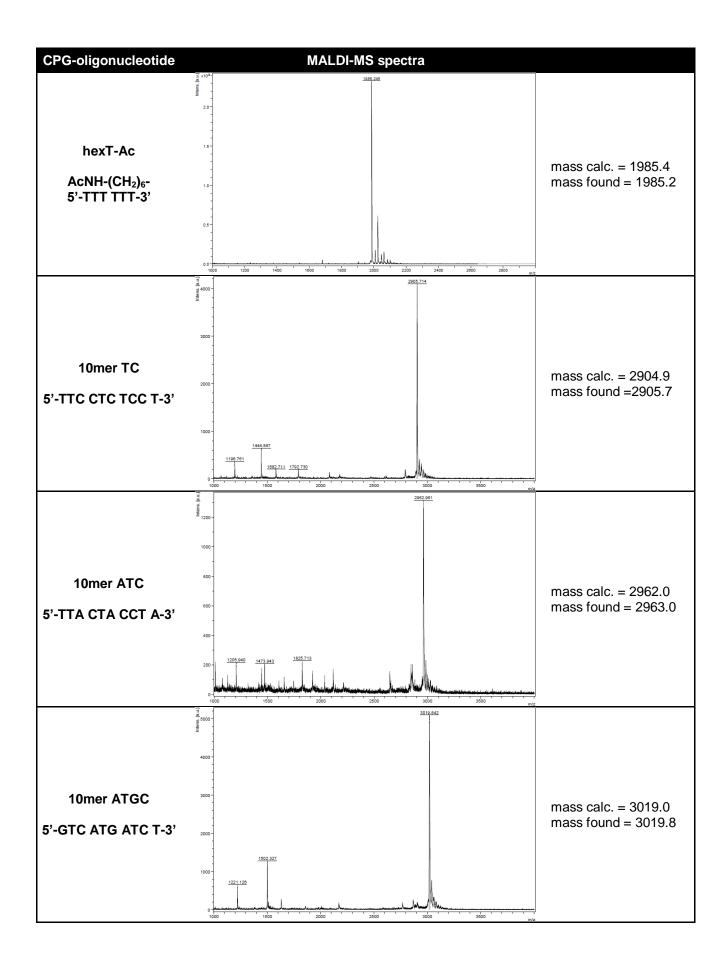




CPG-oligonucleotide + (±)-BINAP K

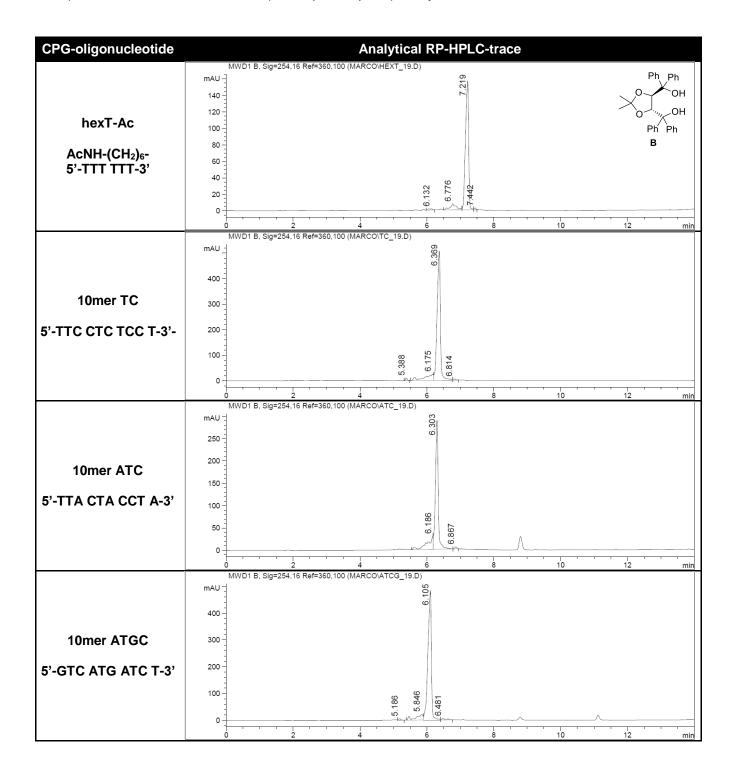
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with (\pm)-BINAP **K** (200 equiv., 4 µmol) in dry MeOH.

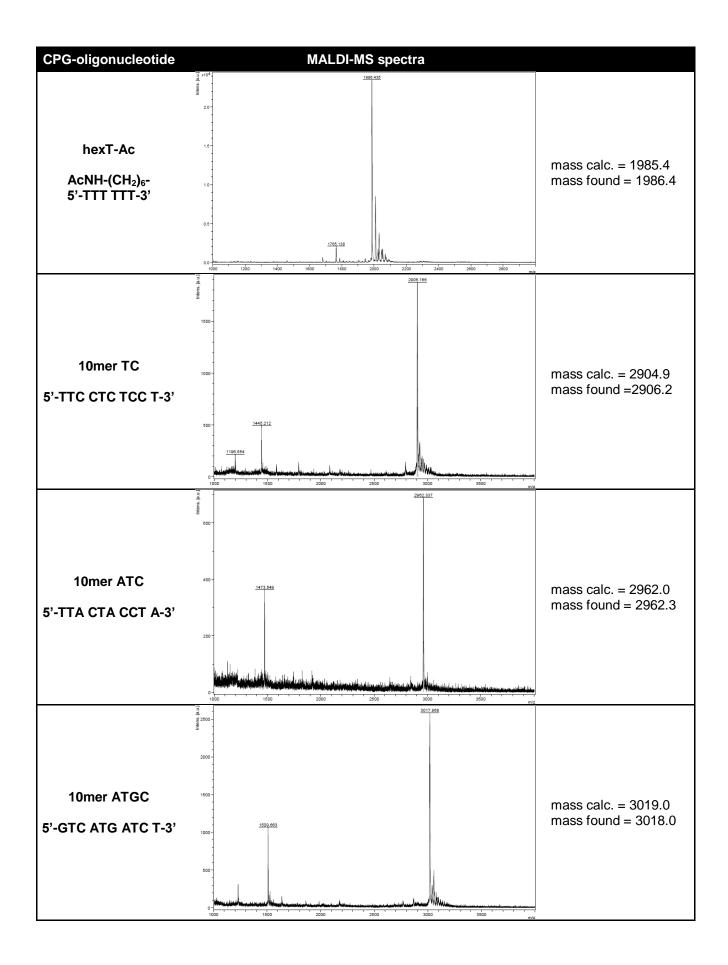




CPG-oligonucleotide + TADDOL B

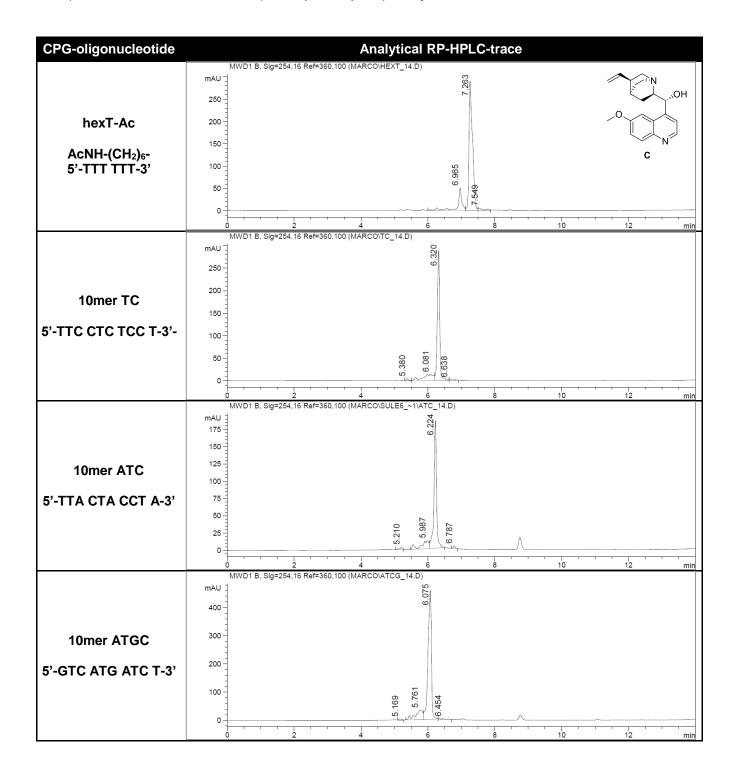
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with TADDOL **B** (200 equiv., 4 µmol) in dry MeOH.

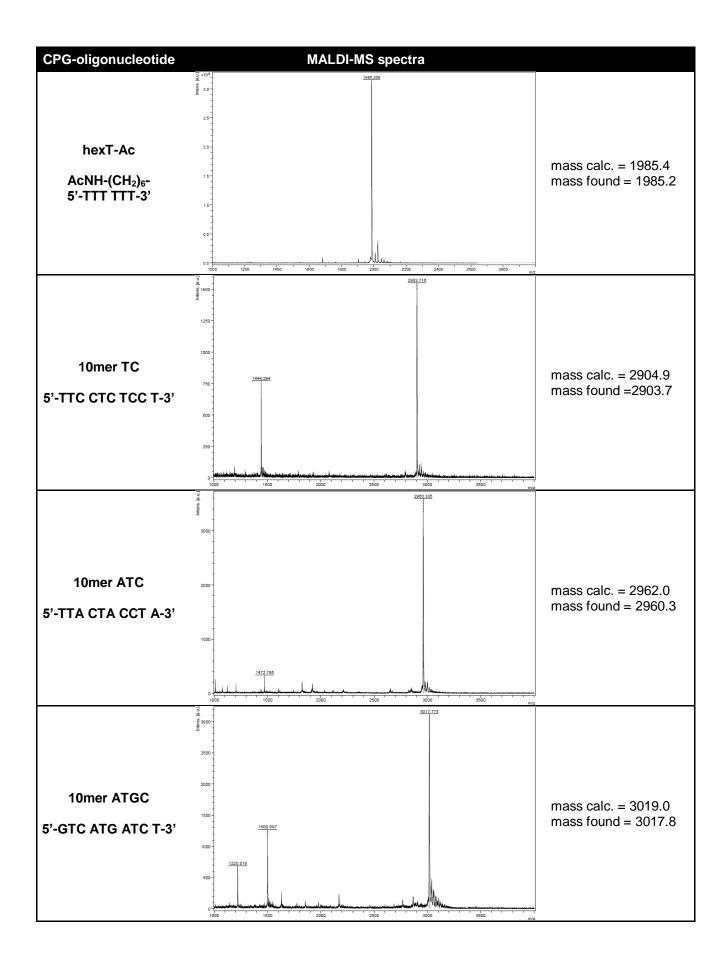




CPG-oligonucleotide + Quinine C

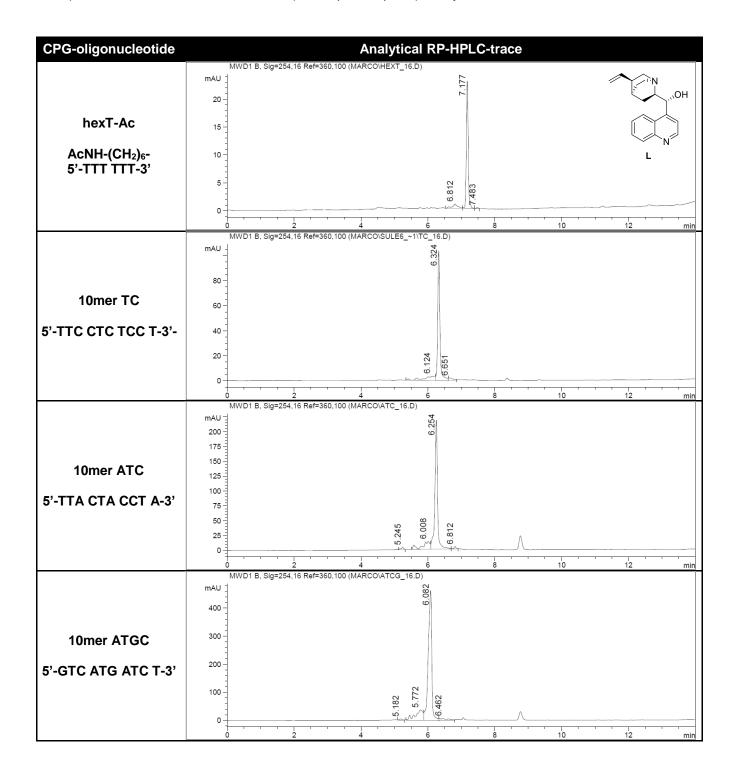
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with Quinine **C** (200 equiv., 4μ mol) in dry MeOH.

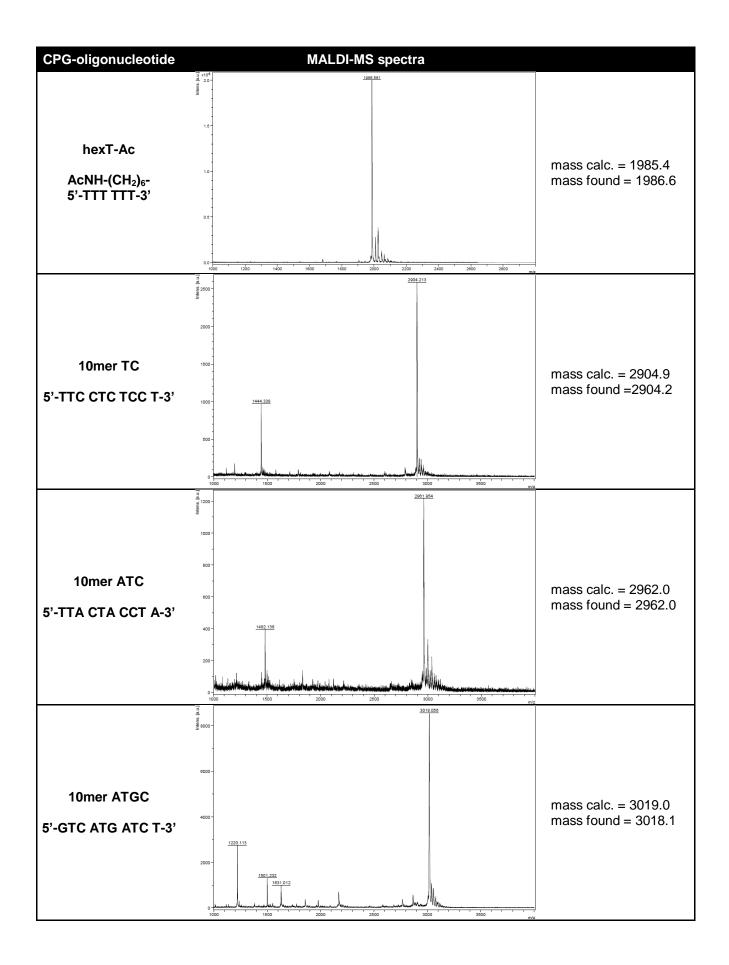




CPG-oligonucleotide + Cinchonidine L

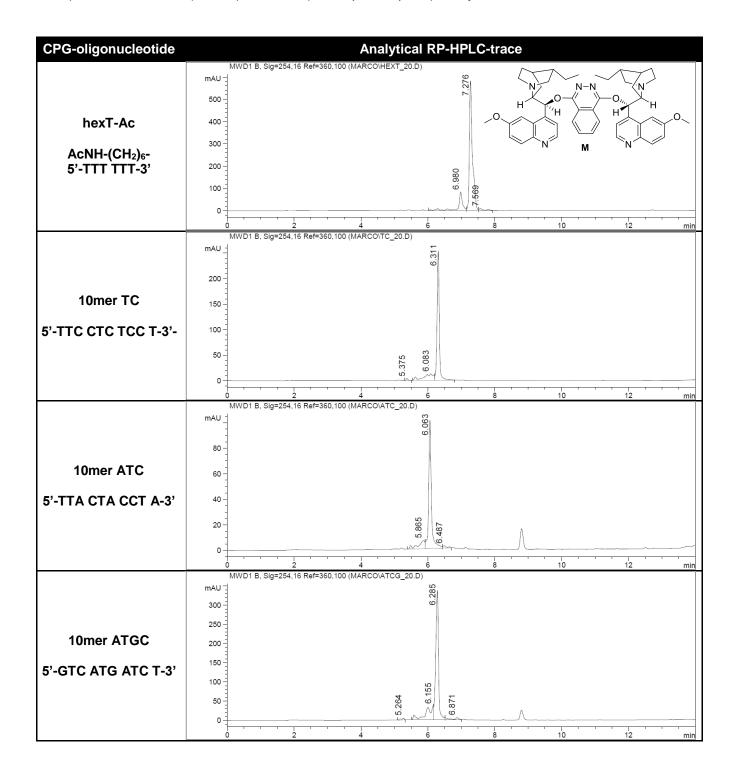
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with Cinchonidine L (200 equiv., 4 µmol) in dry MeOH.

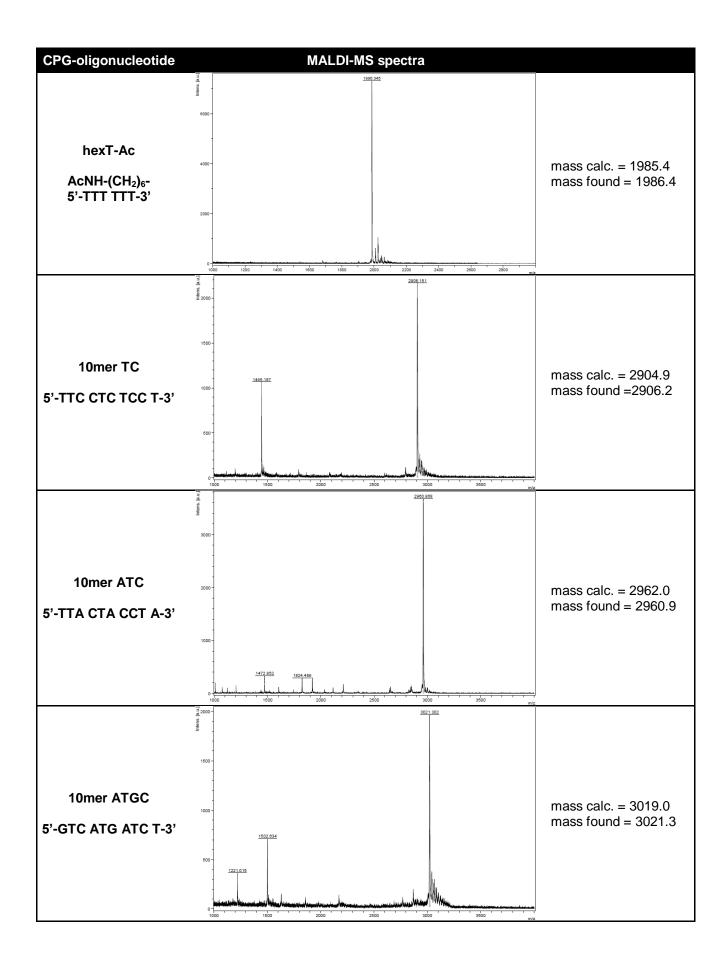




CPG-oligonucleotide + (DHQD)₂PHAL M

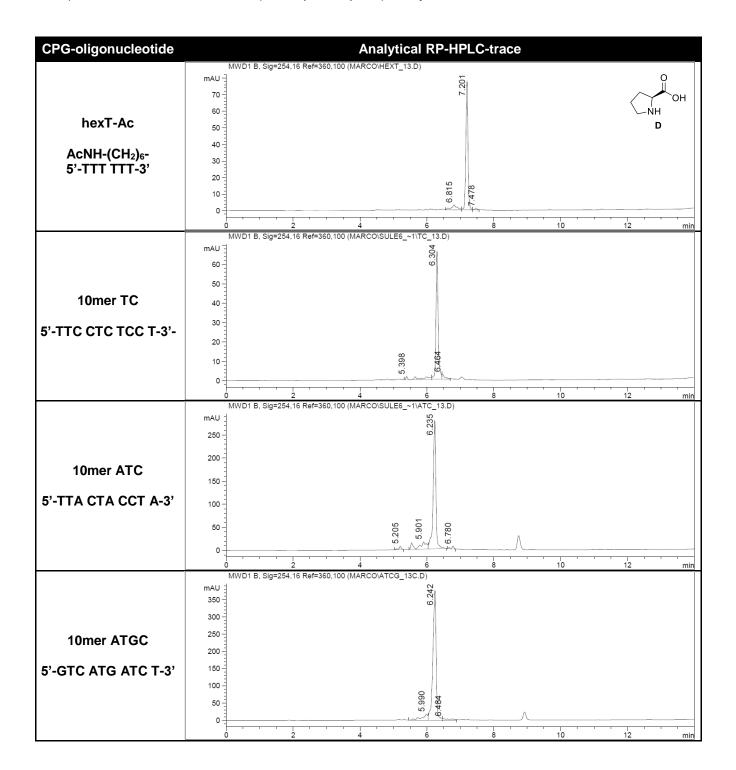
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with $(DHQD)_2PHAL \mathbf{M}$ (200 equiv., 4 µmol) in dry MeOH.

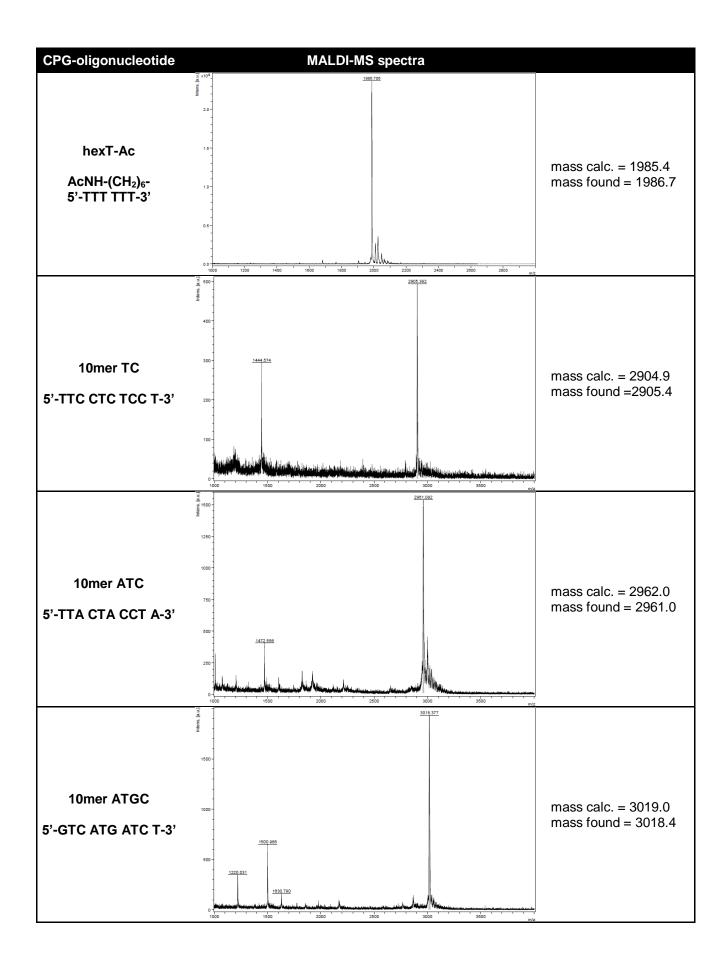




CPG-oligonucleotide + L-Prolin D

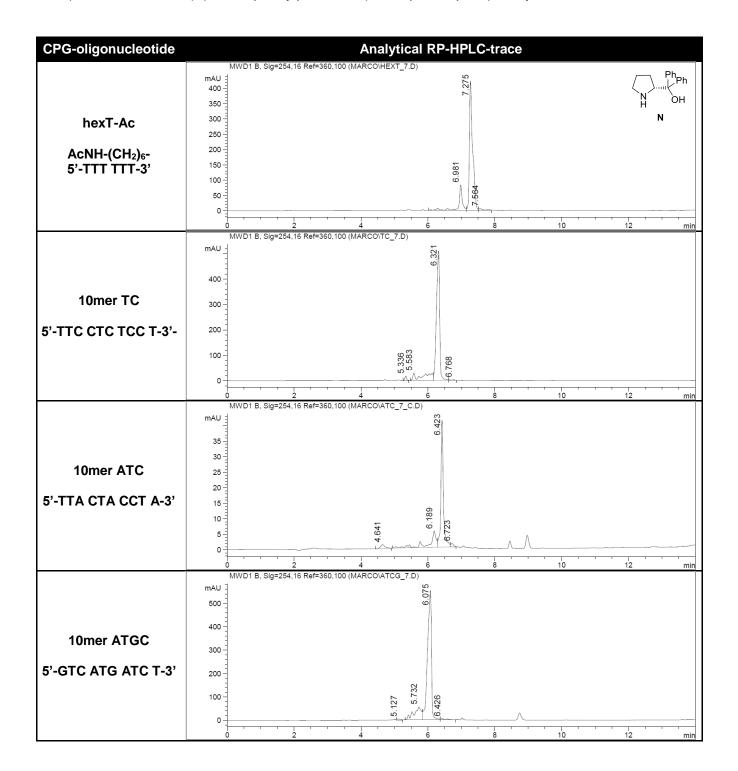
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with L-Prolin **D** (200 equiv., 4 µmol) in dry ACN.

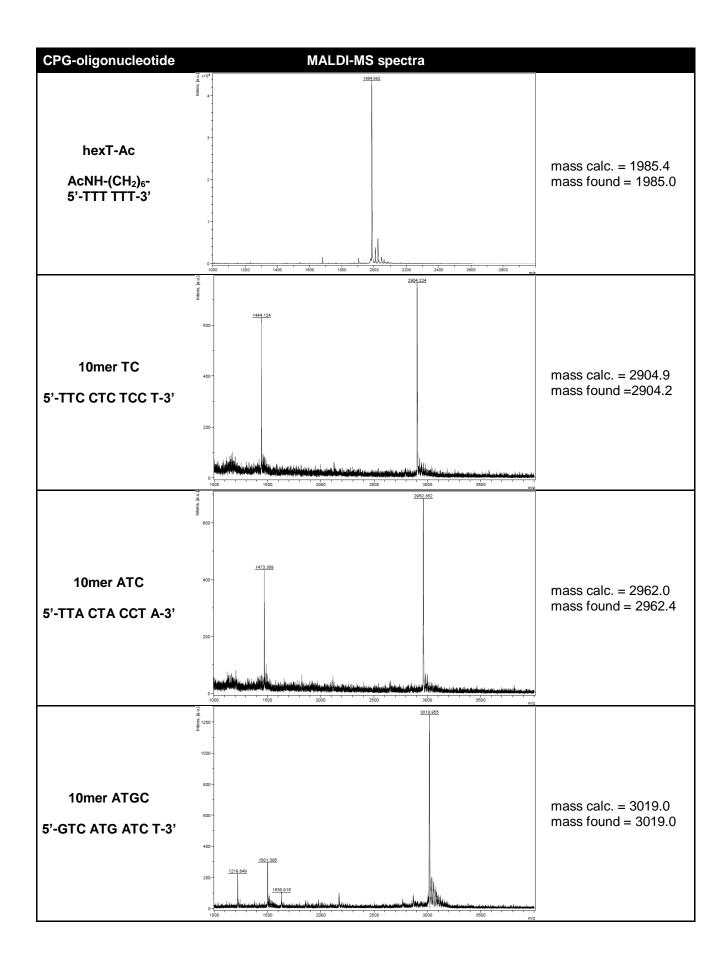




CPG-oligonucleotide + (*R*)-α,α-Diphenylprolinol N

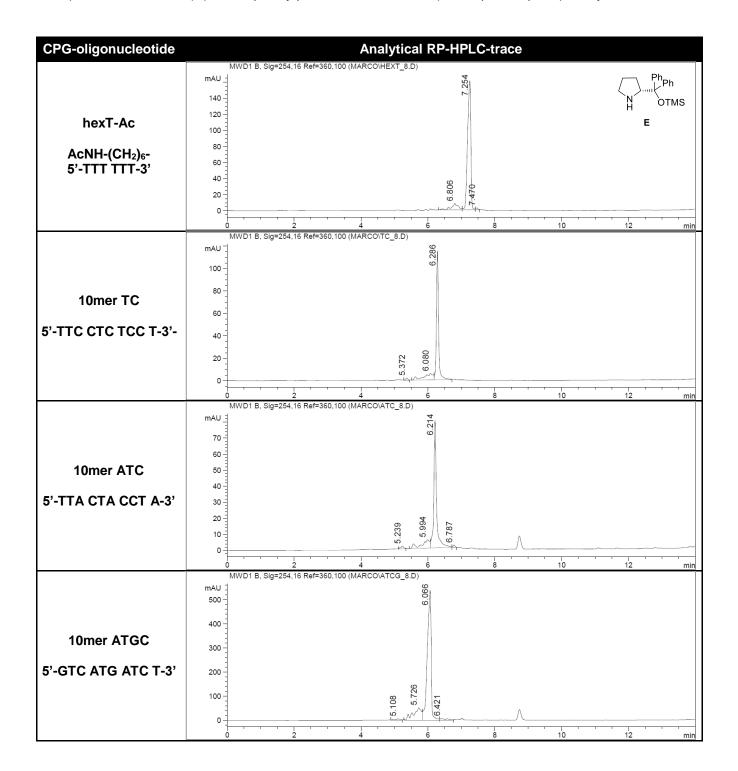
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with (*R*)- α , α -Diphenylprolinol **N** (200 equiv., 4 µmol) in dry ACN.

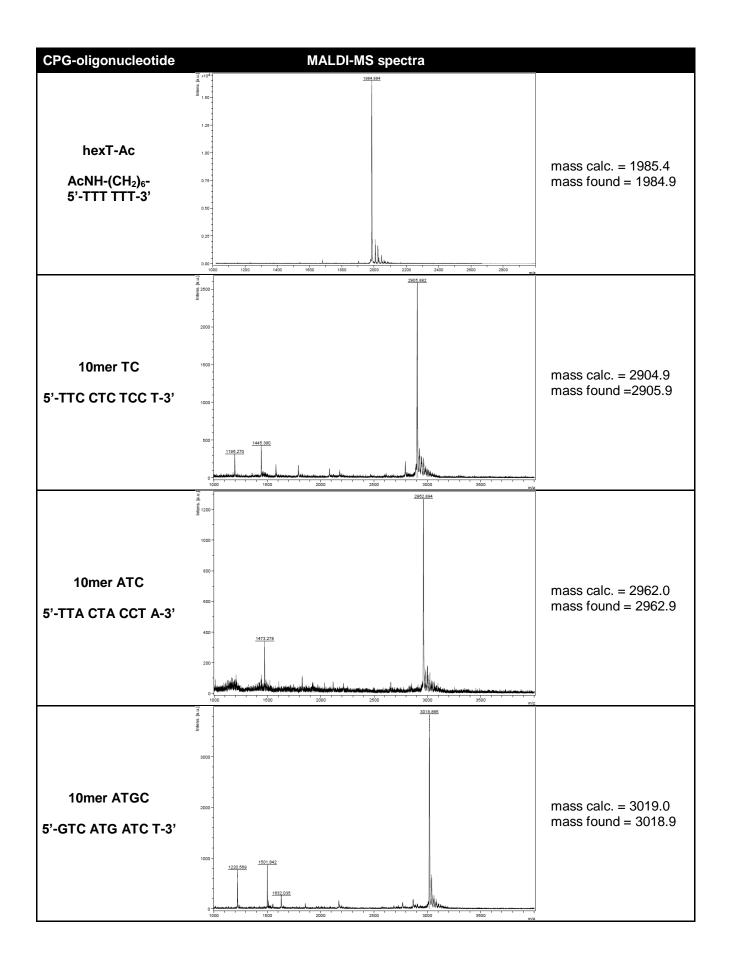




CPG-oligonucleotide + (*R*)- α , α -Diphenylprolinol TMS ether E

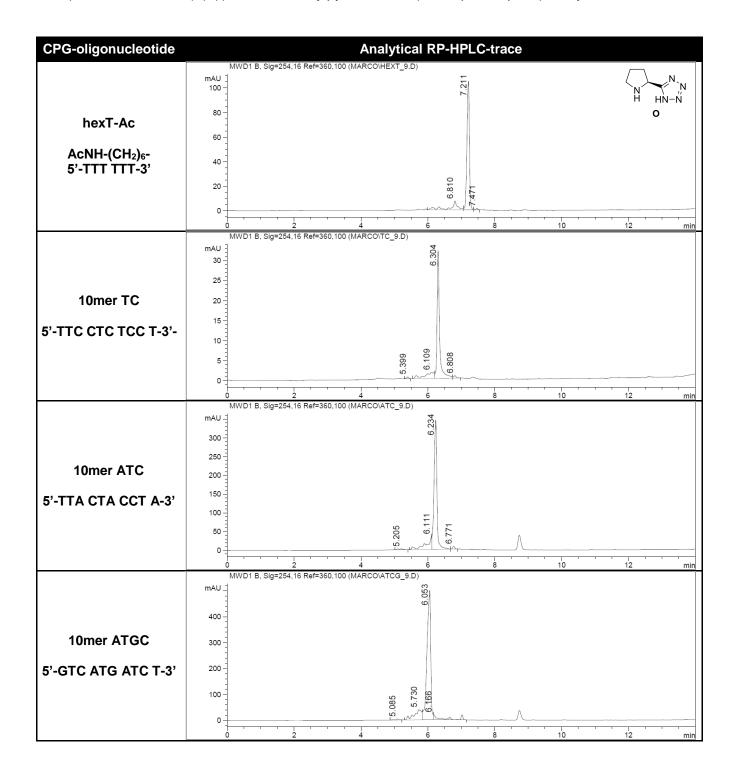
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with (*R*)- α , α -Diphenylprolinol TMS ether **E** (200 equiv., 4 µmol) in dry ACN.

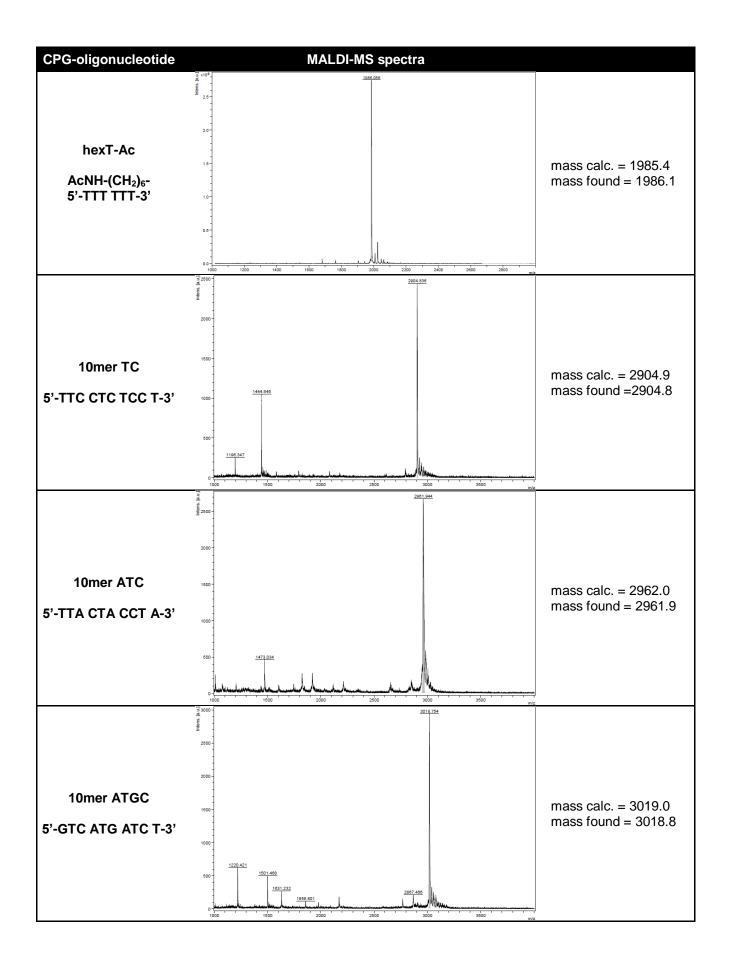




CPG-oligonucleotide + (S)-(-)-2-Tetrazol-5-ylpyrrolidine O

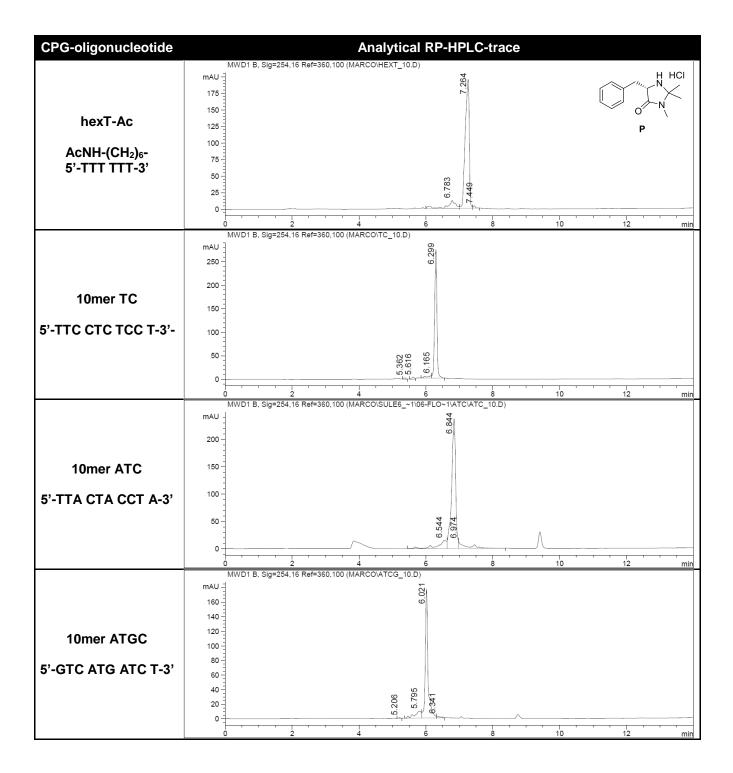
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with (*S*)-(-)-2-Tetrazol-5-ylpyrrolidine **O** (200 equiv., 4 μ mol) in dry MeOH.

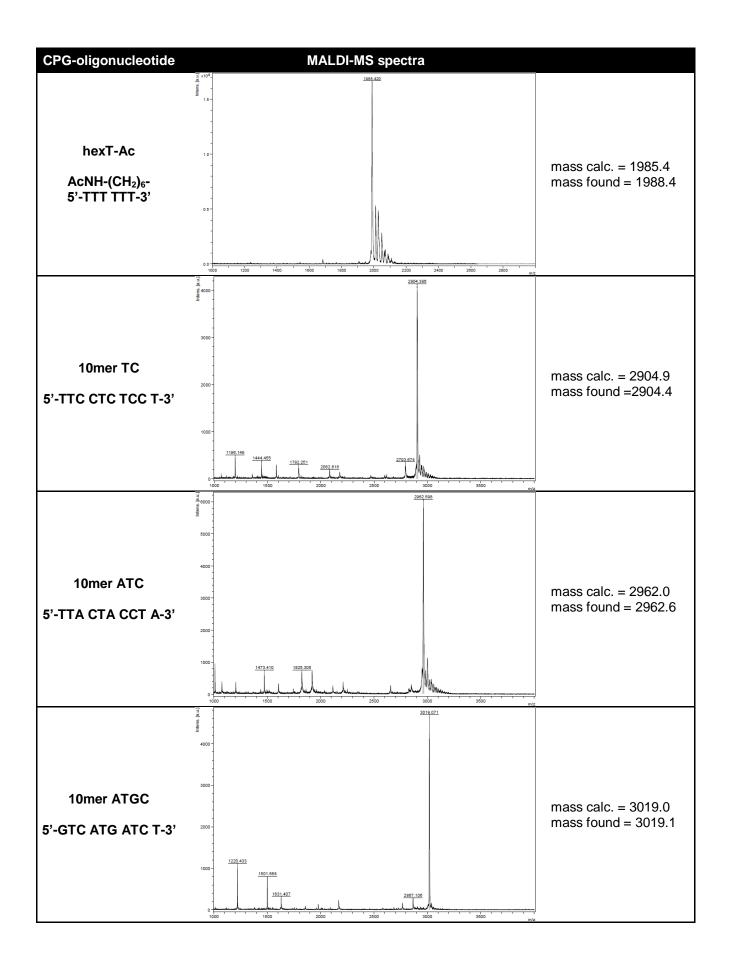




CPG-oligonucleotide + (5S)-(-)-2,2,3-Trimethyl-5-benzyl-4-imidazolidinone monohydrochloride P

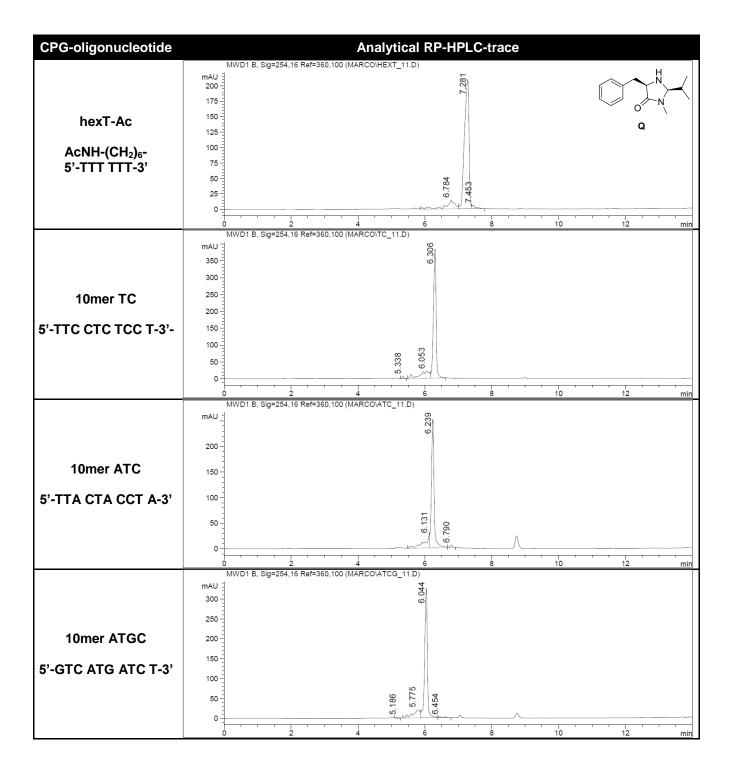
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with (5S)-(-)-2,2,3-Trimethyl-5-benzyl-4-imidazolidinone monohydrochloride **P** (200 equiv., 4 µmol) in dry MeOH.

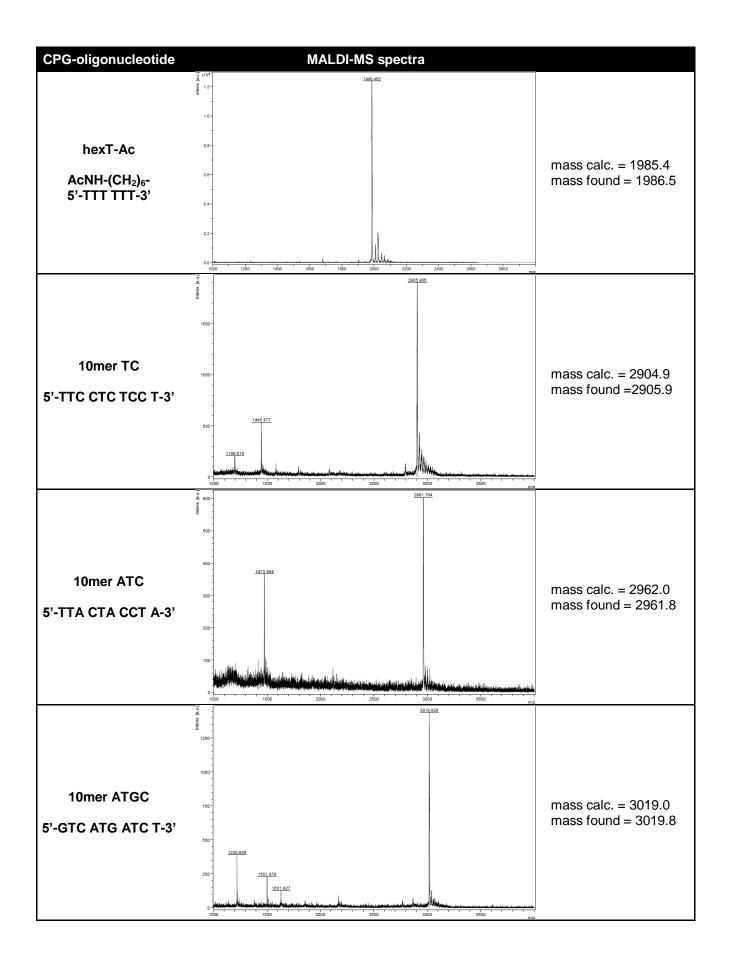




CPG-oligonucleotide + (2R,5R)-(+)-2-tert-Butyl-3-methyl-5-benzyl-4-imidazolidinone Q

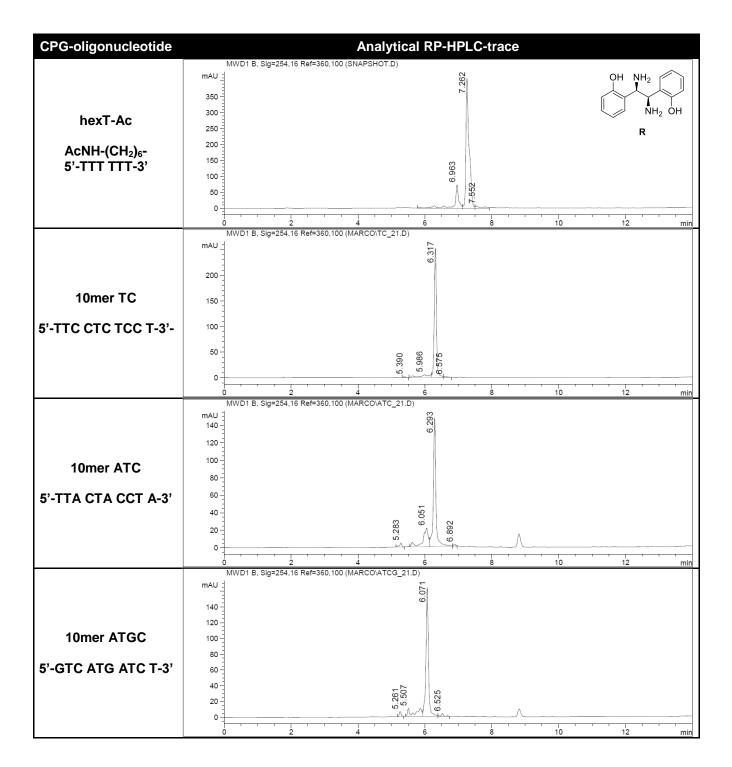
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with (2R,5R)-(+)-2-*tert*-Butyl-3-methyl-5-benzyl-4-imidazolidinone **Q** (200 equiv., 4 µmol) in dry ACN.

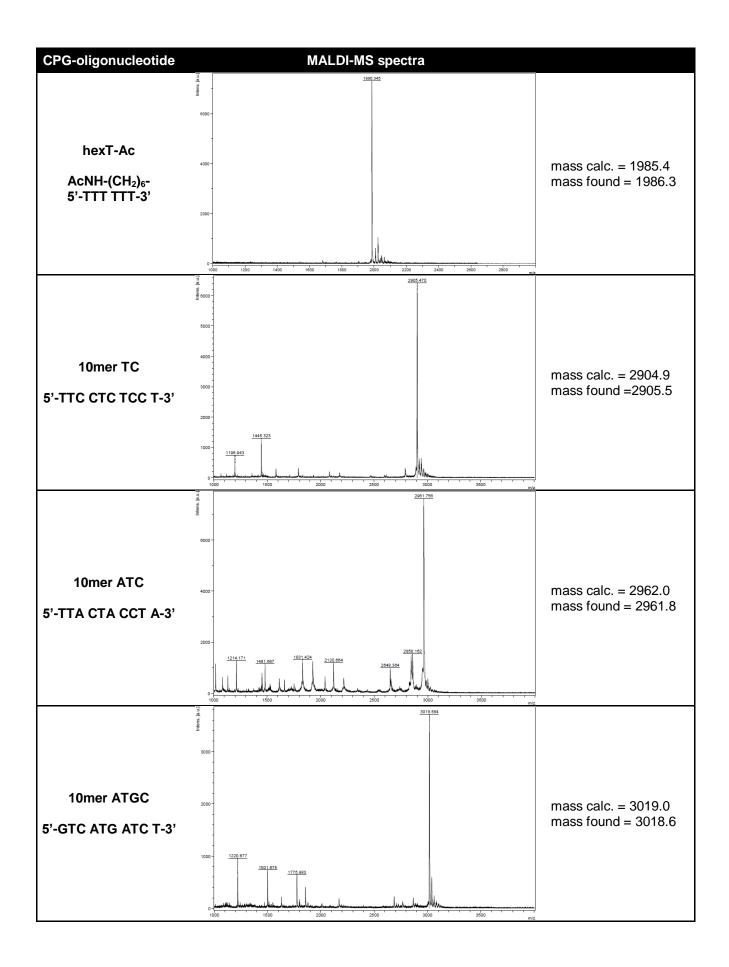




CPG-oligonucleotide + (1R,2R)-1,2-Bis(2-hydroxyphenyl)ethylenediamine R

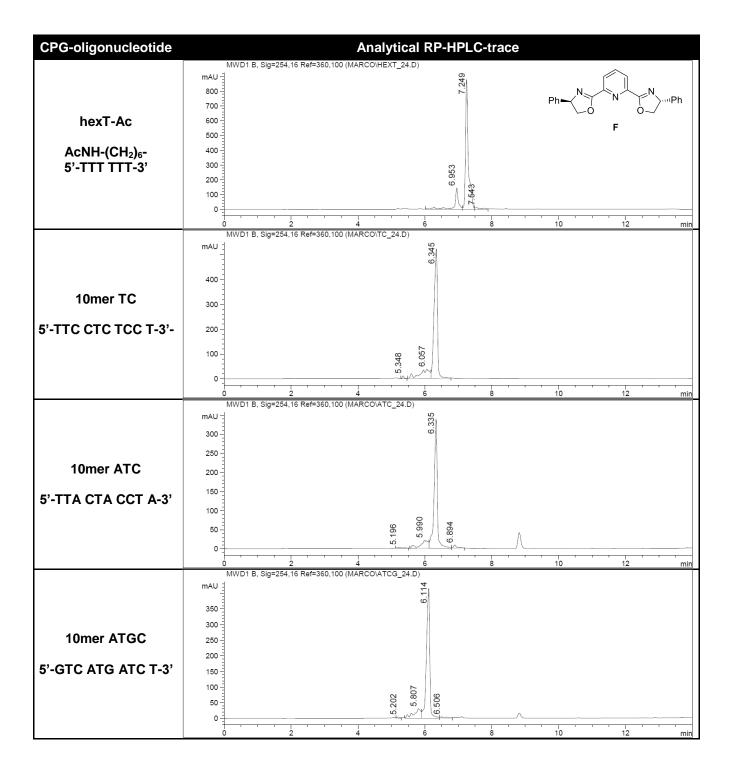
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with (1R,2R)-1,2-Bis(2-hydroxyphenyl)ethylenediamine **R** (200 equiv., 4 µmol) in dry MeOH.

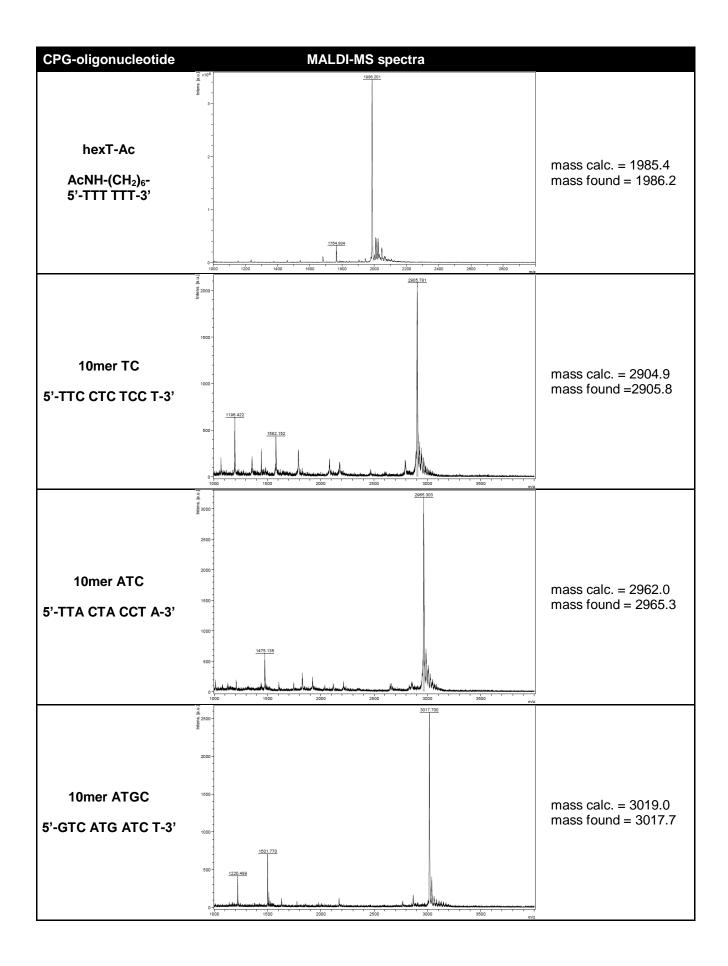




CPG-oligonucleotide + (R,R)-2,6-Bis(4-phenyl-2-oxazolin-2-yl)pyridine F

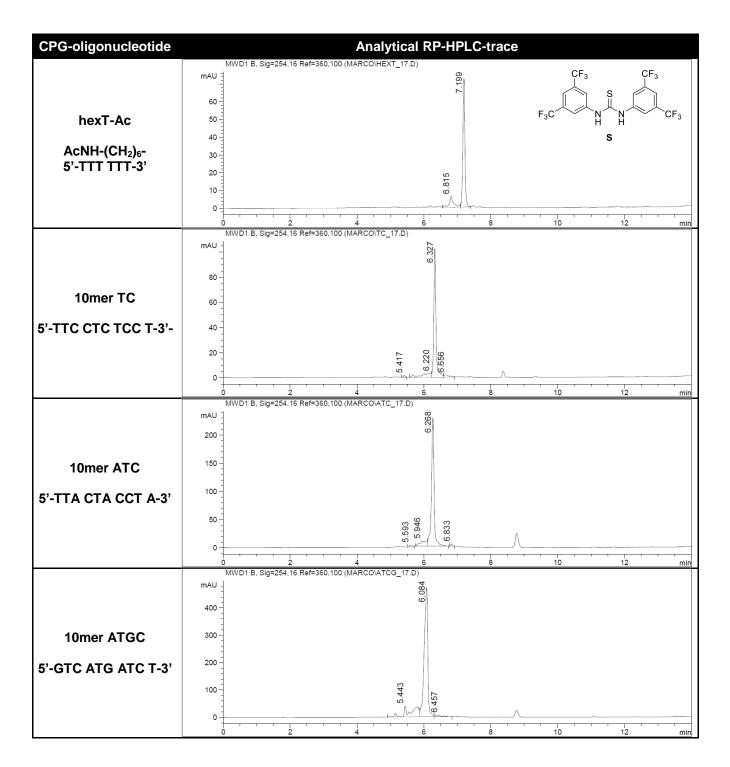
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with (R,R)-2,6-Bis(4-phenyl-2-oxazolin-2-yl)pyridine **F** (200 equiv., 4 µmol) in dry ACN.

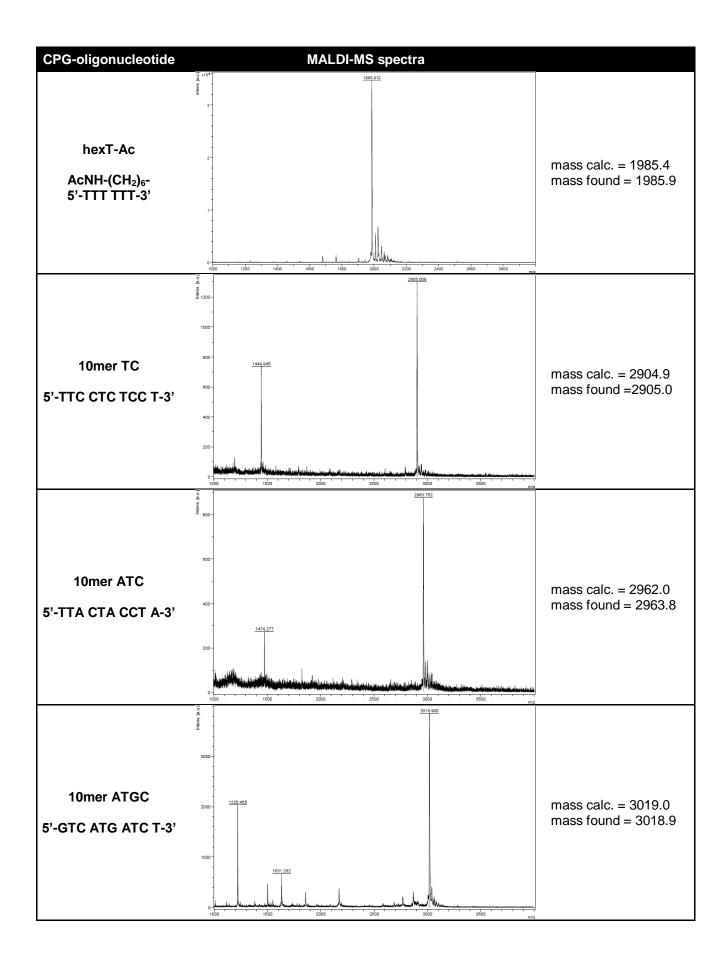




CPG-oligonucleotide + N,N-bis[3,5-bis(trifluoromethyl)phenyl]-thiourea S

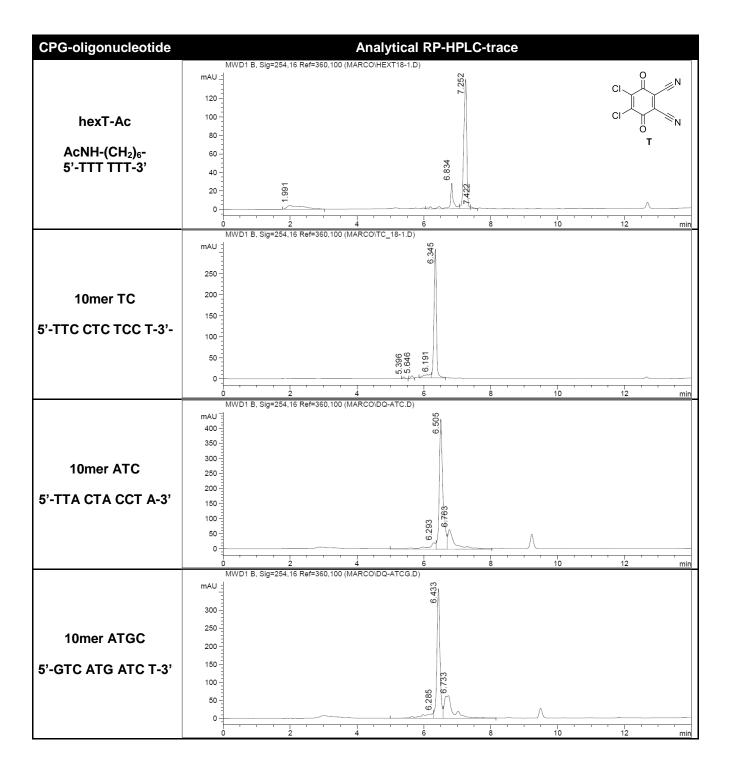
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with N,N-bis[3,5-bis(trifluoromethyl)phenyl]-thiourea **S** (200 equiv., 4 µmol) in dry MeOH.

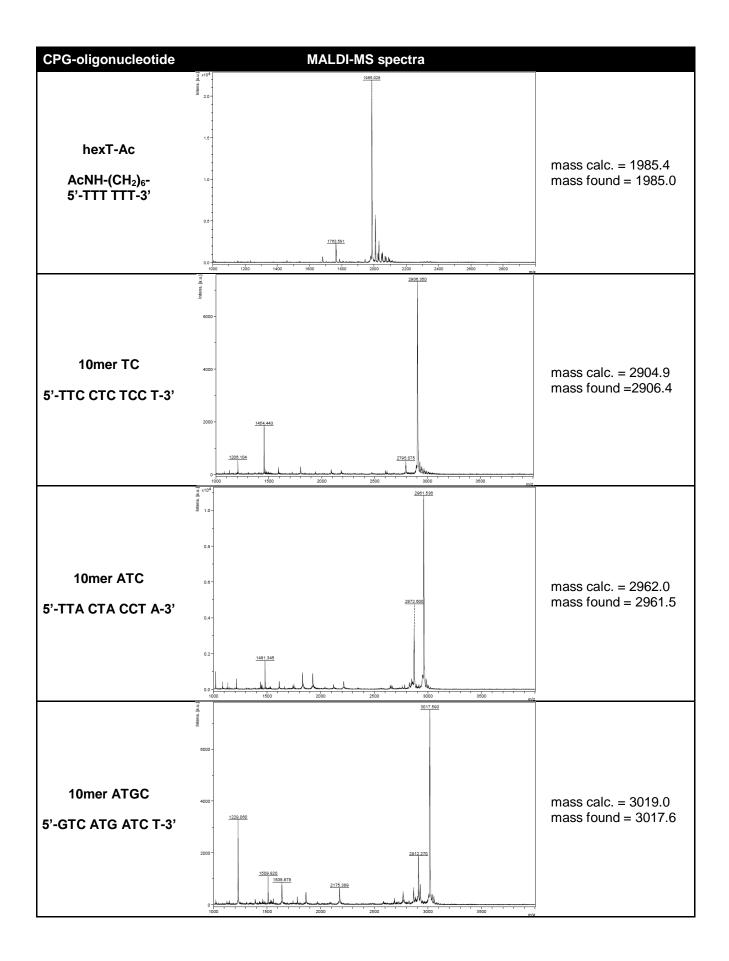




CPG-oligonucleotide + 2,3-Dichloro-5,6-dicyano-1,4-benzoquinone T

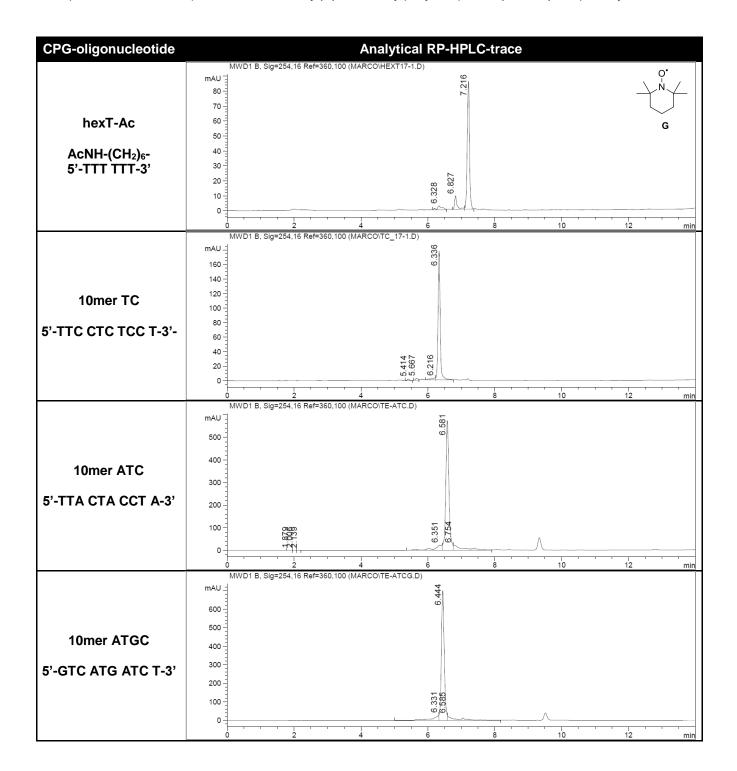
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with 2,3-Dichloro-5,6-dicyano-1,4-benzoquinone **T** (200 equiv., 4 μ mol) in dry EtOH.

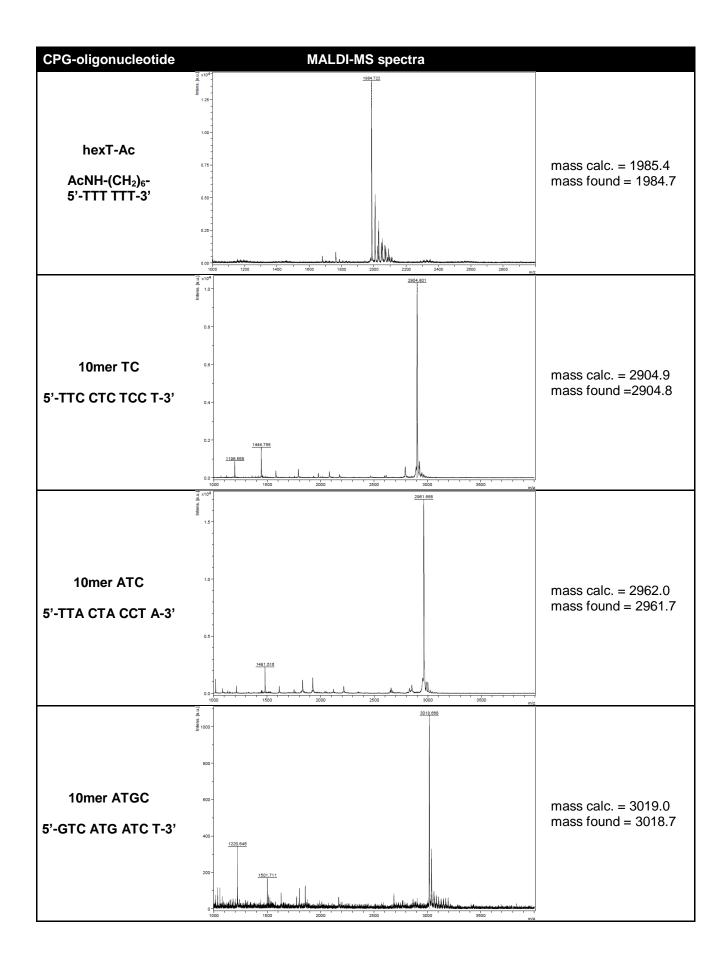




CPG-oligonucleotide + (2,2,6,6-Tetramethylpiperidin-1-yl)oxyl G

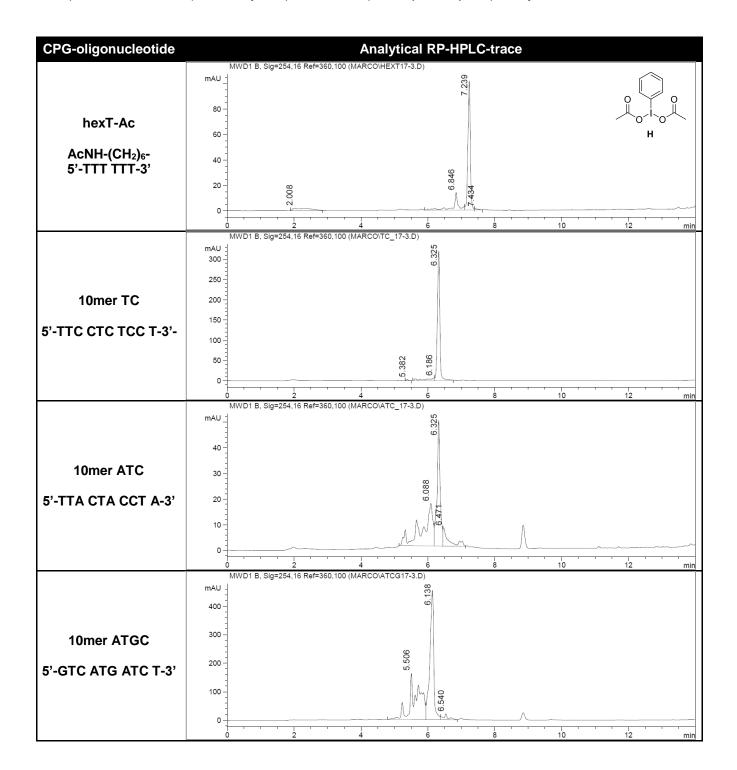
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with (2,2,6,6-Tetramethylpiperidin-1-yl)oxyl **G** (200 equiv., 4 µmol) in dry ACN.

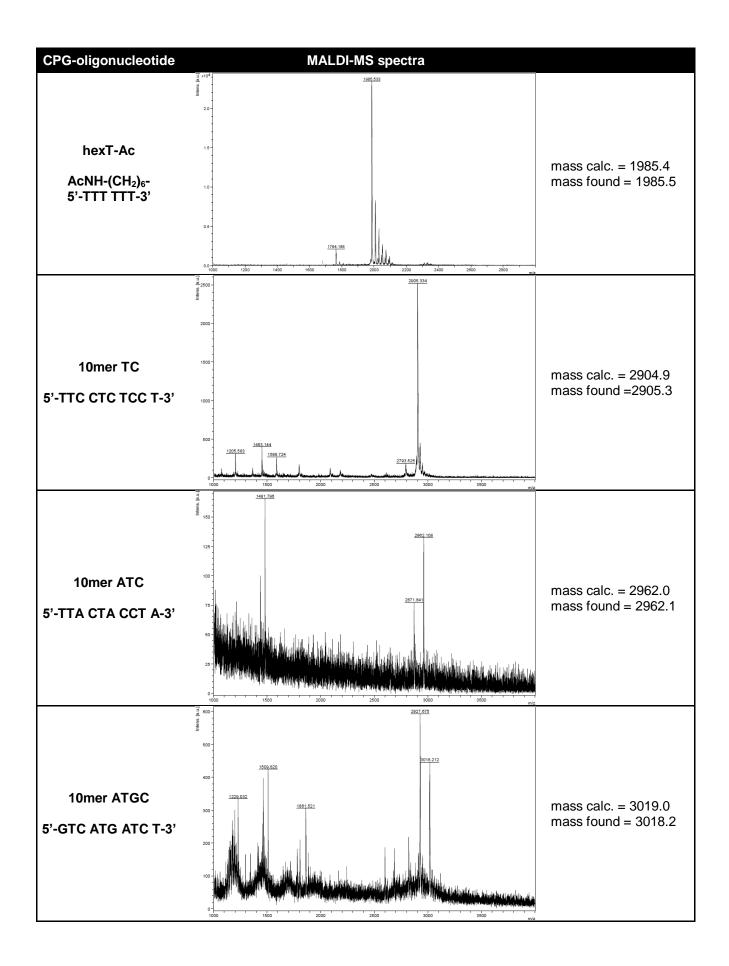




CPG-oligonucleotide + (Diacetoxyiodo)benzene H

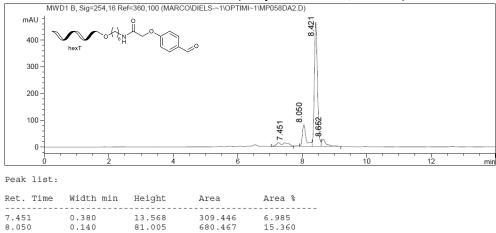
According to the representative procedure (RP-04) solid support coupled oligonucleotide (20 nmol) was treated with (Diacetoxyiodo)benzene **H** (200 equiv., 4 µmol) in dry ACN.





Synthesis of CPG-coupled oligonucleotide-aldehyde and -aniline conjugates

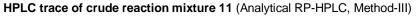
DNA conjugate 11: Following DMT removal, CPG-coupled hexT-C6-NH2 was reacted with 2-(4-formylphenoxy)acetic acid according to RP-06.



15.360

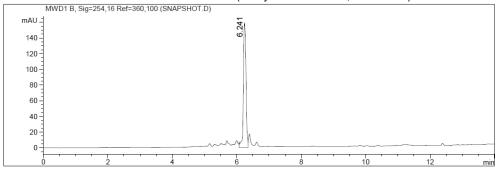
6.371

HPLC trace of crude reaction mixture 11 (Analytical RP-HPLC, Method-I)



680.467 3157.925

282.228



Peak list:

8.421

8.652

Ret. Time Width min Height Area % Area 6.241 0.089 159.216 887.480 100.000

81.005 465.707

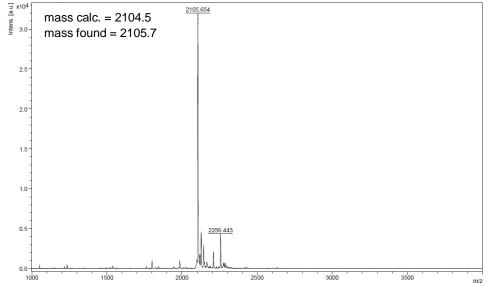
27.277

0.140

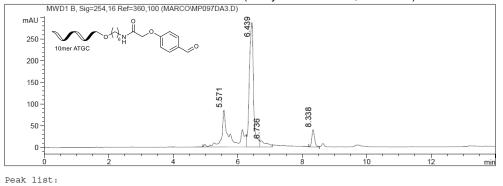
0.113

0.172





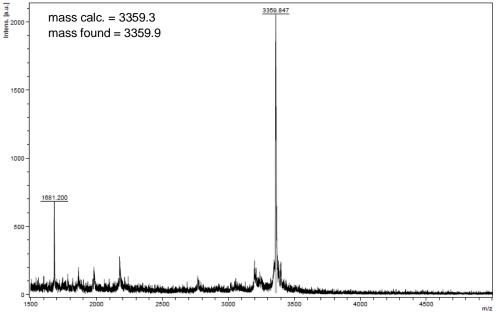
DNA conjugate 15a: Following DMT removal, CPG-coupled 10mer ATGC-C₆-NH₂ was reacted with 2-(4-formylphenoxy)acetic acid according to RP-06.



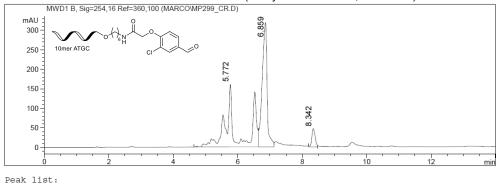
HPLC trace of crude reaction mixture 15a (Analytical RP-HPLC, Method-II)

Ret. Time	Width min	Height	Area	Area %
5.571	0.289	85.209	1478.684	32.859
6.439	0.151	287.006	2591.899	57.597
6.736	0.244	13.652	199.884	4.442
8.338	0.096	39.738	229.610	5.102





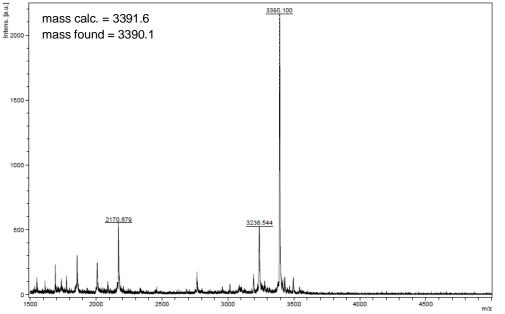
DNA conjugate 15b: Following DMT removal, CPG-coupled 10mer ATGC-C₆-NH₂ was reacted with 2-(2-chloro-4-formylphenoxy)acetic acid according to RP-06.



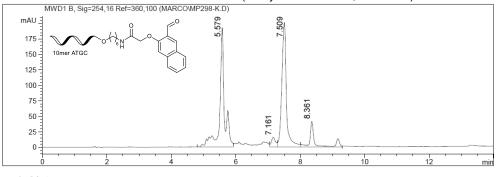
HPLC trace of crude reaction mixture 15b (Analytical RP-HPLC, Method-II)

Ret. Time	Width min	Height	Area	Area %
5.772	0.363	160.782	3497.436	48.597
6.859	0.176	318.828	3368.118	46.800
8.342	0.119	46.536	331.252	4.603





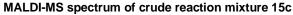
DNA conjugate 15c: Following DMT removal, CPG-coupled 10mer ATGC-C₆-NH₂ was reacted with 2-((3-formylnaphthalen-2-yl)oxy)acetic acid according to RP-06.

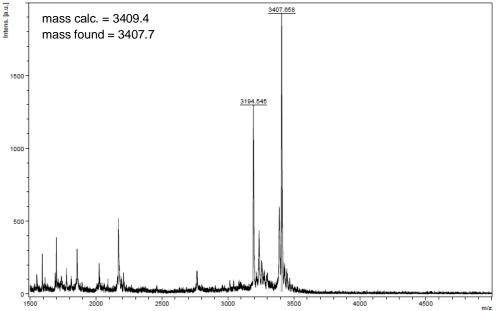


HPLC trace of crude reaction mixture 15c (Analytical RP-HPLC, Method-II)

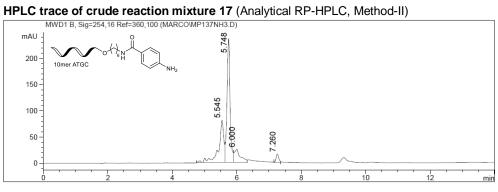
Peak list:

Ret. Time	Width min	Height	Area	Area %
5.579	0.169	192.695	1958.967	43.751
7.161	0.150	15.139	136.541	3.049
7.509	0.163	200.876	1960.796	43.792
8.361	0.171	41.176	421.236	9.408





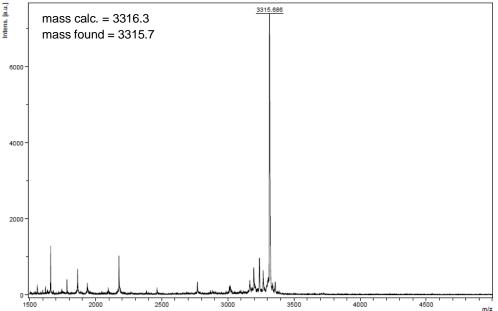
DNA conjugate 17: Following DMT removal, CPG-coupled 10mer ATGC-C6-NH2 was reacted with Fmoc-4aminobenzoic acid according to RP-06.



Peak list:

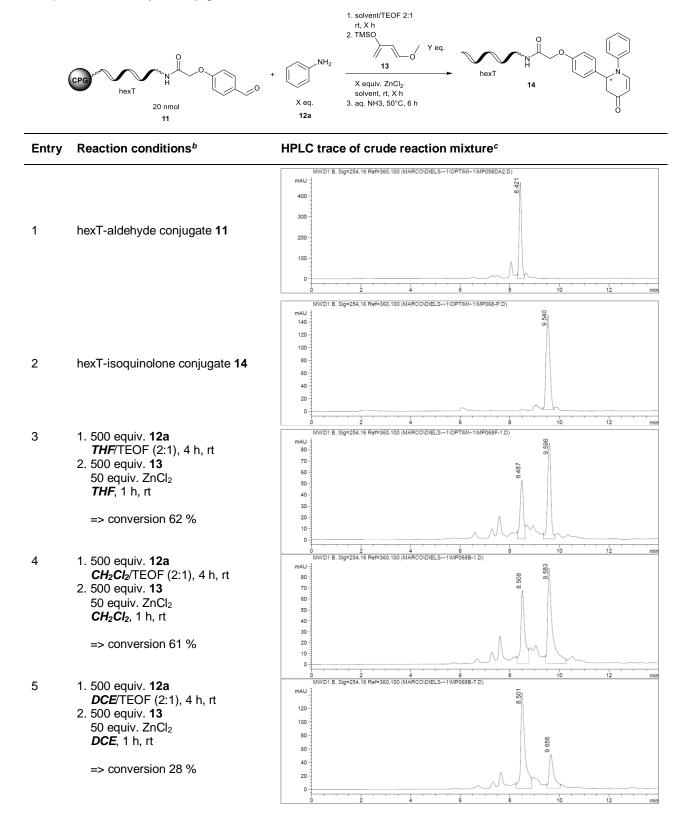
Ret. Time	Width min	Height	Area	Area %
5.545	0.185	80.235	889.637	31.150
5.748	0.108	236.329	1537.165	53.823
6.000	0.227	24.931	338.957	11.868
7.260	0.094	15.985	90.193	3.158

MALDI-MS spectrum of crude reaction mixture 17

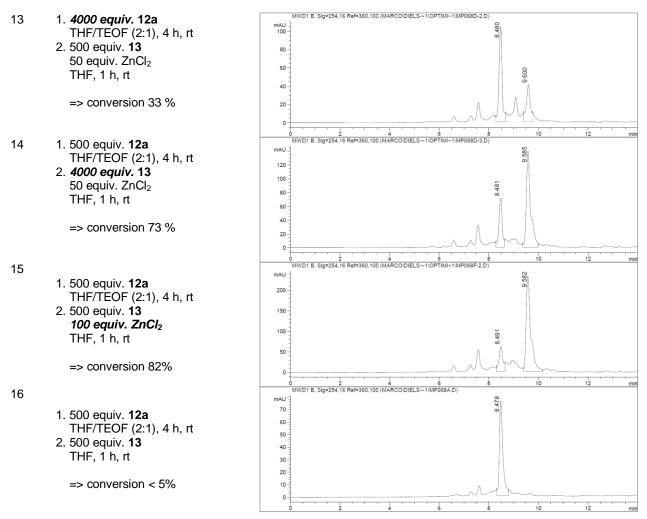


ZnCl2-mediated aza-Diels-Alder reaction on CPG-coupled oligonucleotides

Table S5 Optimization of ZnCl₂-mediated aza-Diels-Alder reaction with Danishefsky's diene on CPGcoupled hexT-aldehyde conjugate.^a



6	1. 500 equiv. 12a	mAU -	MWD1 B, Sig=254,16 Ref=360,100 (MARCOIDIELS~11MP068B-3.D)
	ACN /TEOF (2:1), 4 h, rt 2. 500 equiv. 13	100	43 0
	50 equiv. ZnCl ₂	80	
	ACN , 1 h, rt	60	ŵ
		40	99 90 80
	=> conversion 82 %	20	A And b.
		0-	
7	1. 500 equiv. 12a	mAU	MWD1 B, Sig=254,16 Ref=360,100 (MARCO\DIELS-~1\MP068B-4.D)
	MeOH /TEOF (2:1), 4 h, rt	120	9
	2. 500 equiv. 13 50 equiv. ZnCl ₂	100 -	
	<i>MeOH</i> , 1 h, rt	60	-
		40	
	=> conversion 80 %	20	
		0	
8	1. 500 equiv. 12a	mAU]	MWD1 B, Sig=254,16 Ref=360,100 (MARCO\DIELS-~1\MP068B-6.D)
	<i>EtOH</i> /TEOF (2:1), 4 h, rt	100	9 51 6
	2. 500 equiv. 13 50 equiv. ZnCl ₂	80	
	<i>EtOH</i> , 1 h, rt	60	。
		40	099 80 80
	=> conversion 80 %	20	
		0	
9	1. 500 equiv. 12a	mAU _	MWD1 B, Sig=254,16 Ref=360,100 (MARCO/DIELS-~1/MP068B-5.D)
	Toluene /TEOF (2:1), 4 h, rt	30	ග්
	2. 500 equiv. 13 50 equiv. ZnCl ₂	25	216
	<i>Toluene</i> , 1 h, rt	20	c,
		15 10	
	=> conversion 65 %	5	
		0	
10	1. <i>1000 equiv.</i> 12a	mAU _	MWD1 B, Sig=254,16 Ref=360,100 (MARCO\DIELS-~1\MP068C-2.D)
	THF/TEOF (2:1), 4 h, rt 2. <i>1000 equiv.</i> 13	80	ယ် ယ် ထဲ တို
	50 equiv. ZnCl ₂	70 60	
	THF, 1 h, rt	50 40	
	40.0/	30	
	=> conversion 49 %	20 10	
		0	
11	1. 2000 equiv. 12a	mAU -	MWD1 B, Sig=254.16 Ref=360.100 (MARCOIDIELS11MP068C-3.D)
	THF/TEOF (2:1), 4 h, rt 2. 2000 equiv. 13	70	225
	50 equiv. ZnCl ₂	60 - 50 -	o
	THF, 1 h, rt	40	
	=> conversion 43 %	30 20	A
		10	
		0-	2 4 6 8 10 12 min
12	1. 4000 equiv. 12a	mAU	MWD1 B, Sig=254.15 Ref=360.100 (MARCOIDIELS1/MP068C-4.D)
	THF/TEOF (2:1), 4 h, rt 2. <i>4000 equiv.</i> 13	80 -	ŵ
	50 equiv. $ZnCl_2$	60 -	N
	THF, 1 h, rt	40-	9 662
	=> conversion 34 %	20 -	
		-	
		0-	2 4 6 8 10 12 min



^a Condensation of CPG-coupled hexT aldehyde conjugate **11** (20 nmol) with aniline **12a** (X equiv.) in 36 μ L of indicated solvent/triethyl orthoformate (2:1) at ambient temperature for 4 h, then ZnCl₂ (X equiv.) suspended in 30 μ L of indicated solvent and Danishefsky's diene **13** (X equiv.) were added, the reaction mixture was shaken at ambient temperature for 1 h. ^{*b*} Changed parameters are in bold and italic. ^{*c*} Analytical RP-HPLC, Method-I. TEOF = triethyl orthoformate.

 Table S6 – Scope of ZnCl2-mediated aza-Diels-Alder reaction with Danishefsky's diene on CPG-coupled

 10mer ATGC oligonucleotide-aldehyde conjugate using different amines.^a

 1. ACN/TEOF 2:1

 rt, 4 h

CPG prof 10mer A	ATGC 0	12 ACN, rt	13 10mer ATGC 10mer ATGC	
Entry	Product	Amine	Conversion [%] ^b	Mass _{calc.} Mass _{found} ^c
1	16a	н ₂ м 12а	80	3502.5 3503.0
2	16b	H ₂ N	83	3530.6 3530.6
3	16c	H ₂ N	78	3530.6 3530.7
4	16d	H ₂ N	21	3530.6 3530.5
5	16e	H ₂ N-F 12e	82	3520.5 3522.4
6	16f	H ₂ N	75	3520.5 3518.2
7	16g	H ₂ N-F F 12g	63	3538.5 3540.5
8	16h	H ₂ N-Br 12h	75	3581.4 3582.0
9	16i	H ₂ N- Br 12i	< 5	3581.4 n.d.
10	16j	H ₂ N-()-(80	3544.6 3545.1
11	16k	H ₂ N	< 5	3558.6 n.d.
12	161		71	3562.6 3564.0
13	16m	H ₂ N	27	3562.6 3563.8

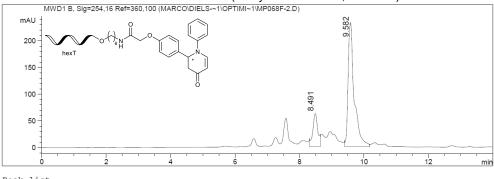
14	16n	H ₂ N- 12n	64	3560.5 n.d. cleaved ester 3546.5 3545.4
15	160	H ₂ N	70	3526.5 3527.2
16 ^{<i>d</i>}	16p	H ₂ N- N N N N H 12p	63	3570.6 3570.6
17 ^d	16q	H ₂ N-()N 12q	< 5	3503.5 n.d.
18 ^{<i>e,f,g</i>}	16r	H ₂ N 12r	56	3468.5 3469.1

^a Condensation of CPG-coupled oligonucleotide conjugate **13** (20 nmol) with amine **14** (500 equiv., 10 µmol) in 36 µL acetonitrile/triethyl orthoformate (2:1) at ambient temperature for 4 h, then addition of $ZnCl_2$ (100 equiv., 2 µmol) dissolved in 30 µL acetonitrile and Danishefsky's diene **15** (1000 equiv., 20 µmol) at ambient temperature for 1 h. DNA cleavage with 30 % aqueous ammonia at 50 °C for 6 h. ^b Determined by analytical RP-HPLC analysis. ^c Measured by MALDI-MS. ^d Dimethyl sulfoxid was used instead of acetonitrile. ^e 1000 equiv. of amine were used. ^d Yb(OTf)₃ was used instead of ZnCl₂. ^g 2nd step of the reaction was performed overnight at 35 °C . 10mer ATGC = 5'-GTC ATG ATC T-3'. n.d. = not detected.

Products of aza-Diels-Alder reaction with Danishefsky's diene on CPG-coupled 10mer ATGC oligonucleotide-aldehyde conjugate

DNA conjugate 14: CPG-coupled hexT-aldehyde conjugate **11** was reacted with aniline **12a** and Danishefsky's diene **13** according to RP-07.

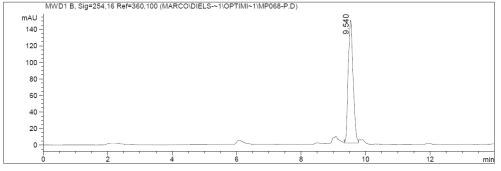




Peak list:

Ret. Time	Width min	Height	Area	Area %
8.491 9.582	0.183	61.121 231.477	672.861 3012.743	18.256
9.562		231.477		01./44

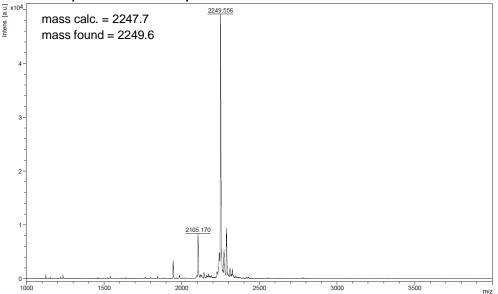
HPLC trace of isolated product 14 (Analytical RP-HPLC, Method-I) MWD1 B, Sig=254,16 Ref=360,100 (MARCO\DIELS-~1\OPTIMI~1\MP068-P.D)



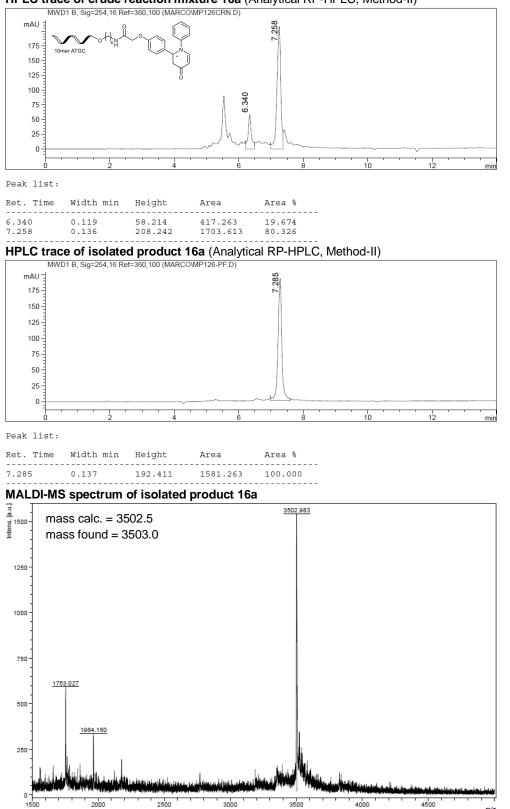
Peak list:

Ret. Time	Width min	Height	Area	Area %
9.540	0.174	148.892	1551.445	100.000

MALDI-MS spectrum of isolated product 14

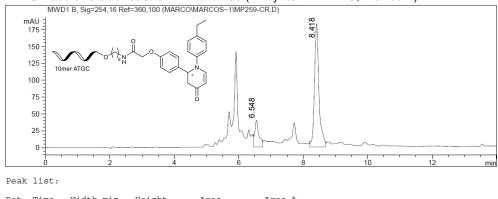


DNA conjugate 16a: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with aniline **12a** and Danishefsky's diene **13** according to RP-07.



HPLC trace of crude reaction mixture 16a (Analytical RP-HPLC, Method-II)

DNA conjugate 16b: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 4-ethylaniline **12b** and Danishefsky's diene **13** according to RP-07.



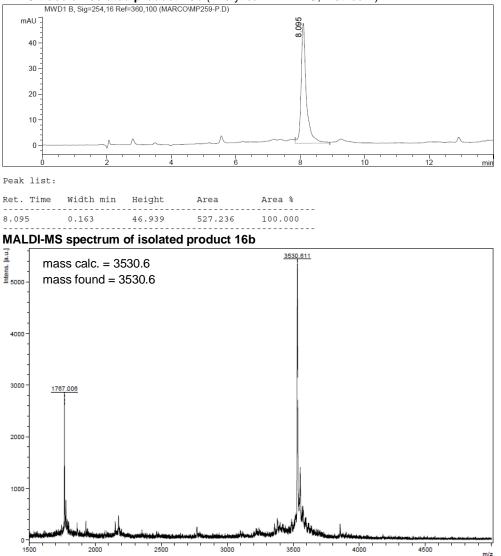
HPLC trace of crude reaction mixture 16b (Analytical RP-HPLC, Method-II)

 Ret. Time
 Width min
 Height
 Area
 Area %

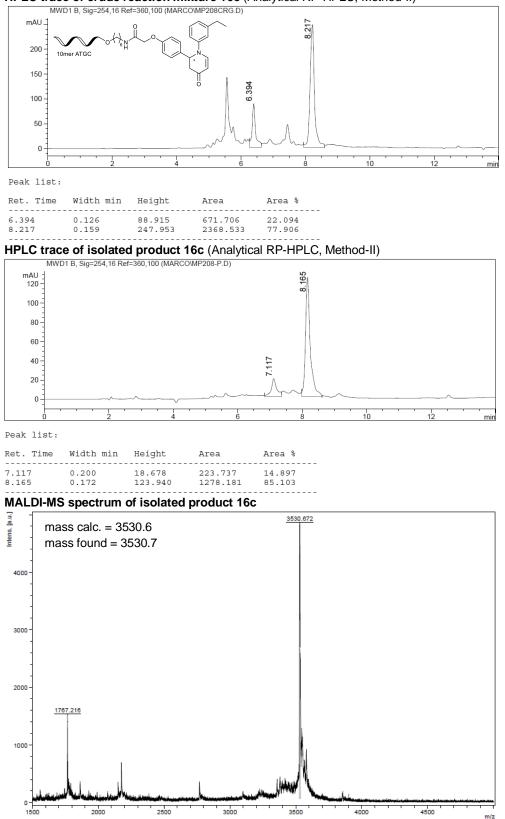
 6.548
 0.152
 40.113
 366.884
 17.142

 8.418
 0.161
 183.156
 1773.384
 82.858



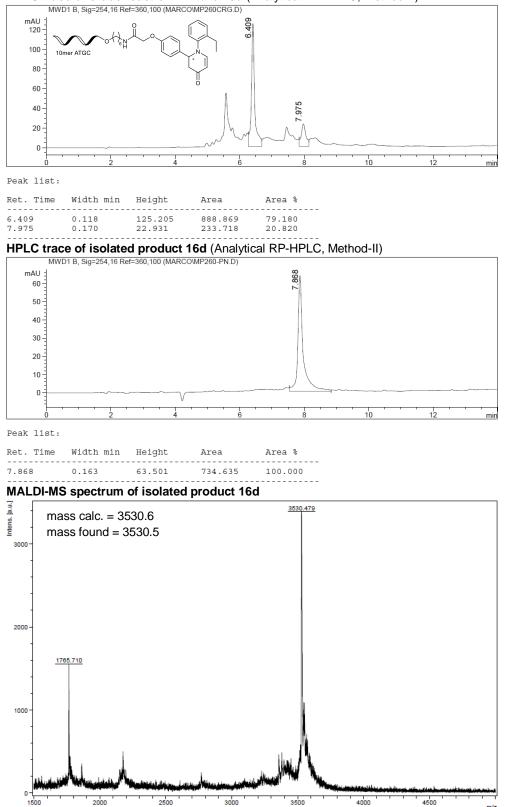


DNA conjugate 16c: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 3-ethylaniline **12c** and Danishefsky's diene **13** according to RP-07.



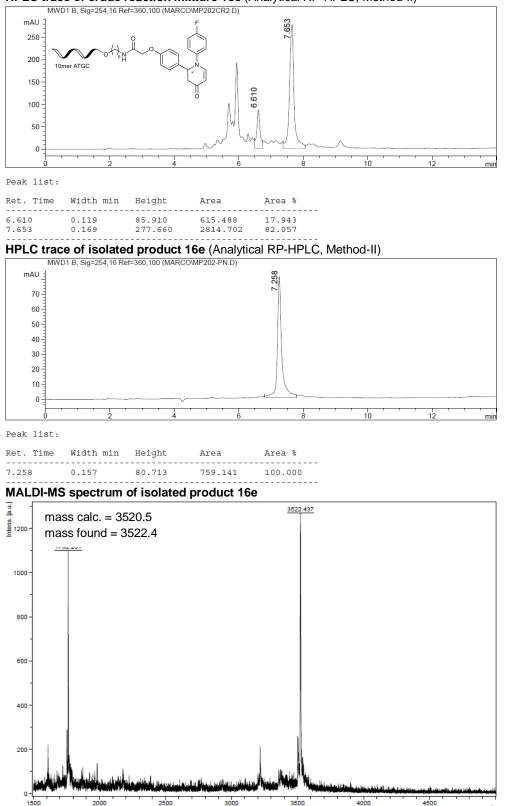
HPLC trace of crude reaction mixture 16c (Analytical RP-HPLC, Method-II)

DNA conjugate 16d: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 2-ethylaniline **12d** and Danishefsky's diene **13** according to RP-07.



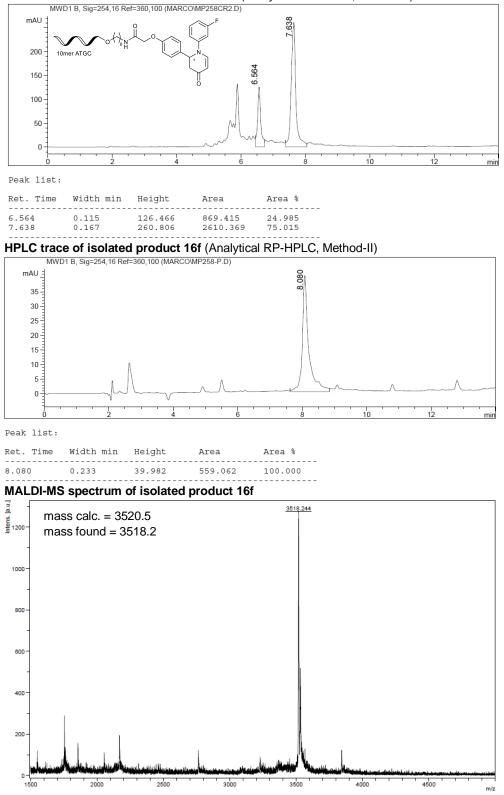
HPLC trace of crude reaction mixture 16d (Analytical RP-HPLC, Method-II)

DNA conjugate 16e: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 4-fluoroaniline **12e** and Danishefsky's diene **13** according to RP-07.



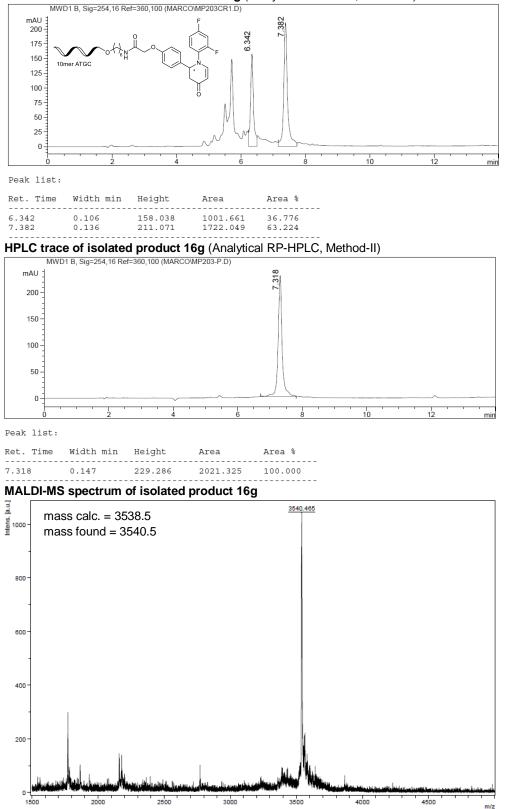
HPLC trace of crude reaction mixture 16e (Analytical RP-HPLC, Method-II)

DNA conjugate 16f: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 3-fluoroaniline **12f** and Danishefsky's diene **13** according to RP-07.



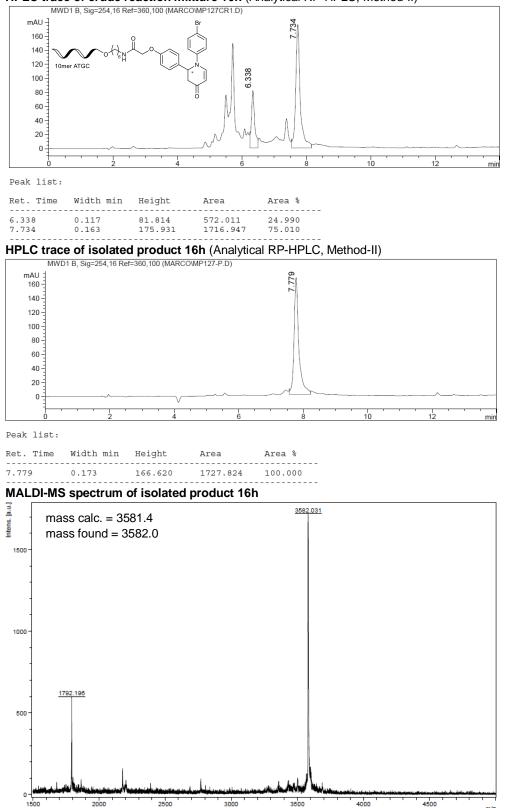
HPLC trace of crude reaction mixture 16f (Analytical RP-HPLC, Method-II)

DNA conjugate 16g: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 2,4-difluoroaniline **12g** and Danishefsky's diene **13** according to RP-07.



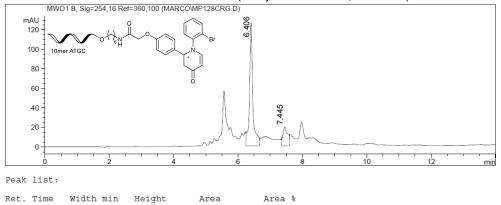
HPLC trace of crude reaction mixture 16g (Analytical RP-HPLC, Method-II)

DNA conjugate 16h: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 4-bromoaniline **12h** and Danishefsky's diene **13** according to RP-07.



HPLC trace of crude reaction mixture 16h (Analytical RP-HPLC, Method-II)

DNA conjugate 16i: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 2-bromoaniline **12i** and Danishefsky's diene **13** according to RP-07.



82.131

17.869

892.818

194.252

HPLC trace of crude reaction mixture 16i (Analytical RP-HPLC, Method-II)



126.845

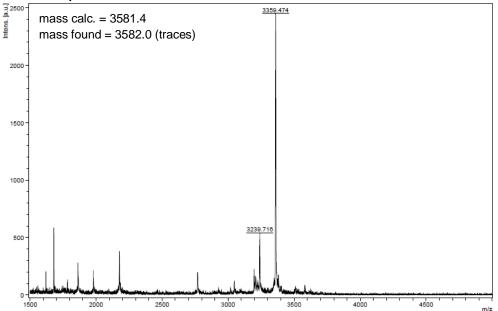
19.606

6.406

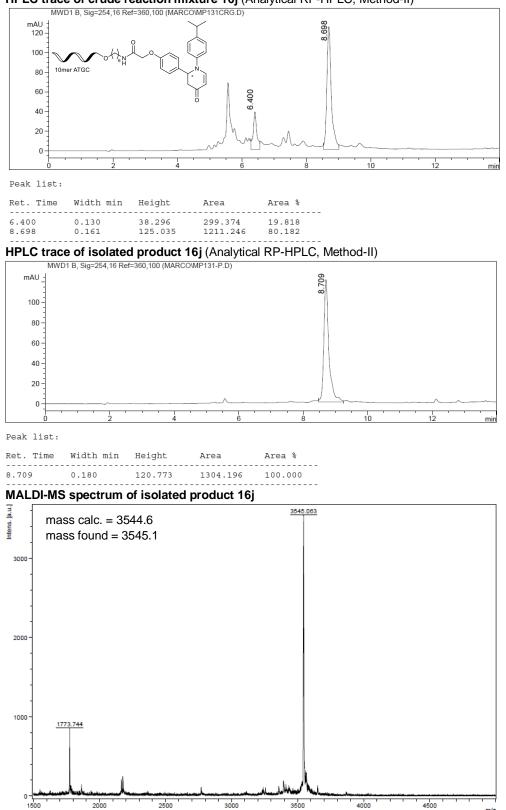
7.445

0.117

0.165

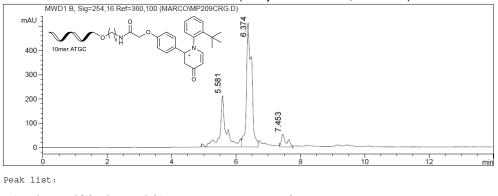


DNA conjugate 16j: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 4-isopropylaniline **12j** and Danishefsky's diene **13** according to RP-07.



HPLC trace of crude reaction mixture 16j (Analytical RP-HPLC, Method-II)

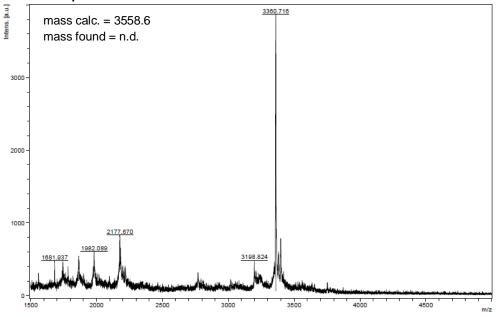
DNA conjugate 16k: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 2-*tert*-butylaniline **12k** and Danishefsky's diene **13** according to RP-07.



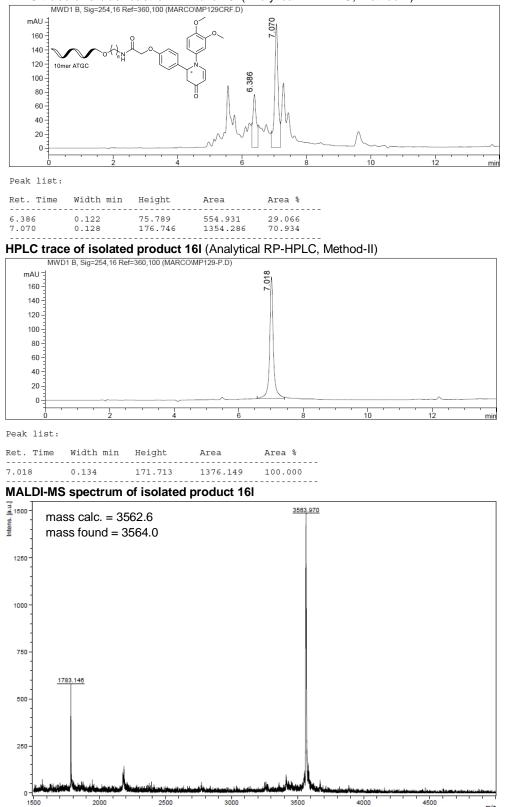
HPLC trace of crude reaction mixture 16k (Analytical RP-HPLC, Method-II)

Ret. Time	Width min	Height	Area	Area %
5.581	0.228	211.424	2894.779	31.714
6.374	0.184	513.326	5666.833	62.083
7.453	0.180	52.493	566.200	6.203

MALDI-MS spectrum of crude reaction mixture 16k

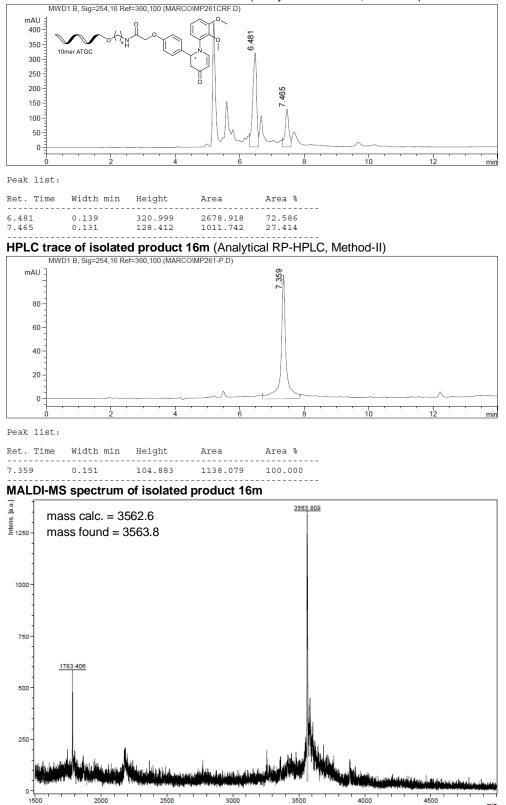


DNA conjugate 16I: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 3,4-dimethoxyaniline **12I** and Danishefsky's diene **13** according to RP-07.



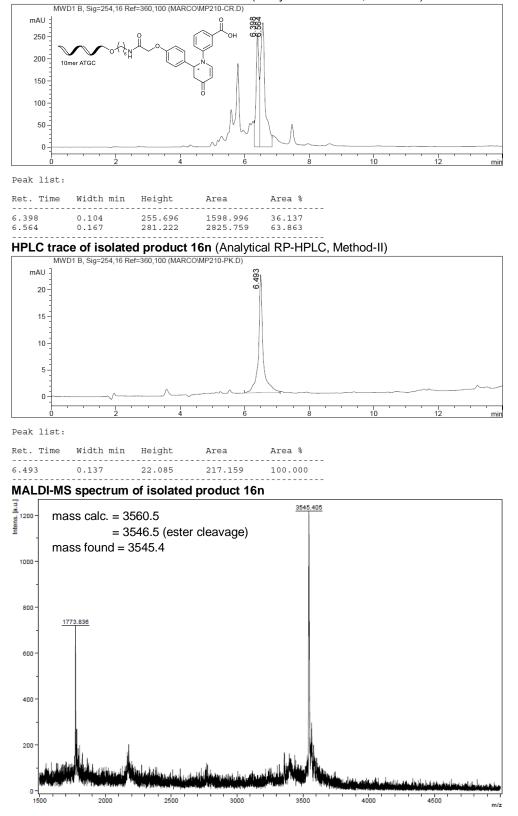
HPLC trace of crude reaction mixture 16I (Analytical RP-HPLC, Method-II)

DNA conjugate 16m: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 2,3-dimethoxyaniline **12m** and Danishefsky's diene **13** according to RP-07.



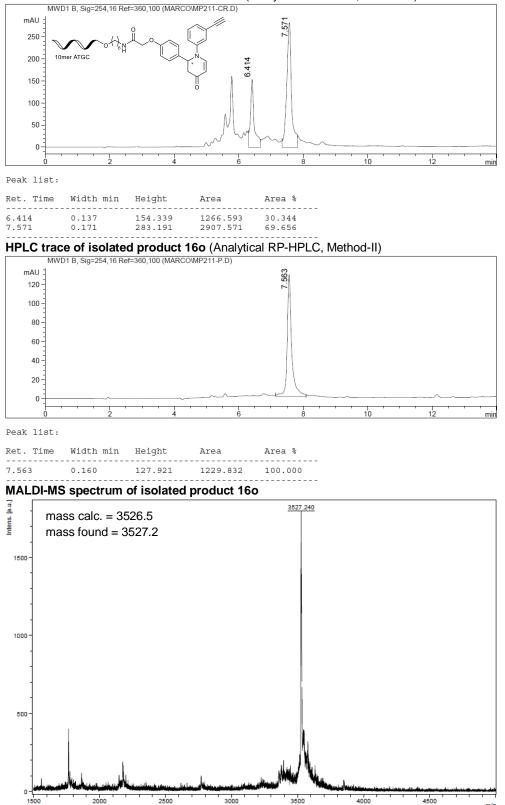
HPLC trace of crude reaction mixture 16m (Analytical RP-HPLC, Method-II)

DNA conjugate 16n: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with methyl 3-aminobenzoate **12n** and Danishefsky's diene **13** according to RP-07.



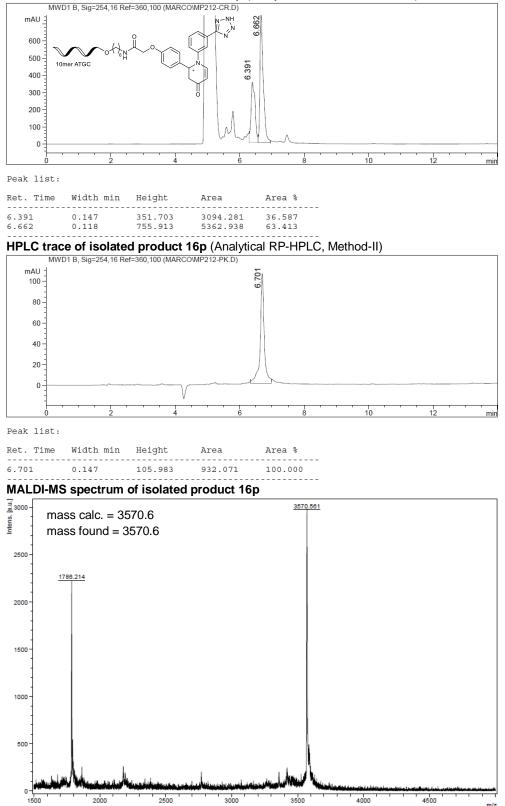
HPLC trace of crude reaction mixture 16n (Analytical RP-HPLC, Method-II)

DNA conjugate 16o: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 3-ethynylaniline **12o** and Danishefsky's diene **13** according to RP-07.



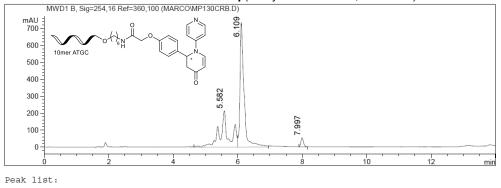
HPLC trace of crude reaction mixture 16o (Analytical RP-HPLC, Method-II)

DNA conjugate 16p: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 3-(2H-tetrazol-5-yl)aniline **12p** and Danishefsky's diene **13** according to RP-07 (Dimethyl sulfoxide was used instead of acetonitrile).



HPLC trace of crude reaction mixture 16p (Analytical RP-HPLC, Method-II)

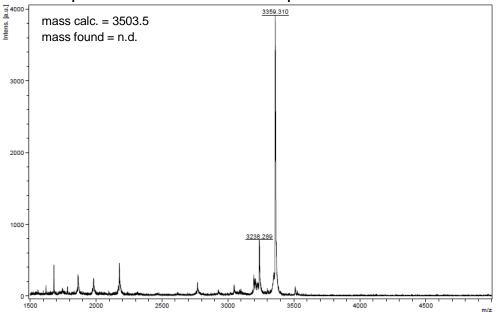
DNA conjugate 16q: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with pyridin-4-amine **12q** and Danishefsky's diene **13** according to RP-07 (Dimethyl sulfoxide was used instead of acetonitrile).



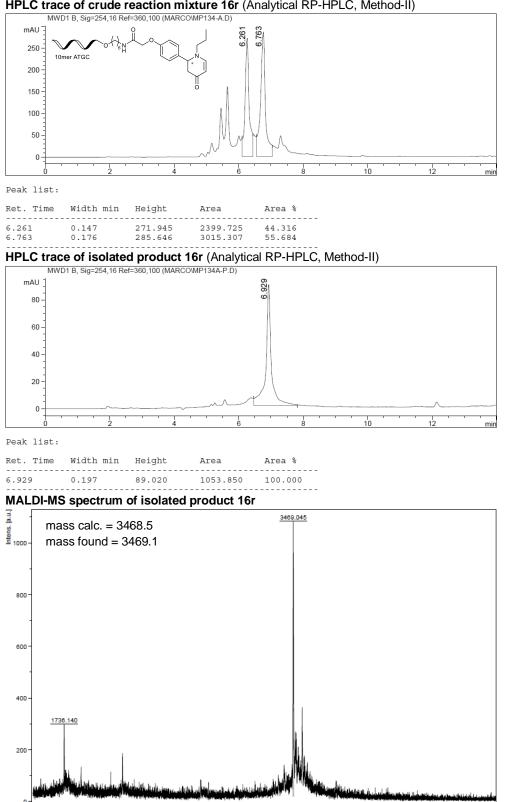
HPLC trace of crude reaction mixture 16q (Analytical RP-HPLC, Method-II)

Ret. Time	Width min	Height	Area	Area %
5.582	0.307	215.698	3966.932	37.123
6.109	0.145	737.833	6401.872	59.910
7.997	0.096	54.891	317.055	2.967





DNA conjugate 16r: CPG-coupled 10mer ATGC-aldehyde conjugate 15a was reacted with propylamine 12r and Danishefsky's diene 13 according to RP-07 (1000 equiv. of amine were used, 100 equiv. of Yb(OTf)₃ were used instead of 50 equiv. of ZnCl₂).



2500

2000

3000

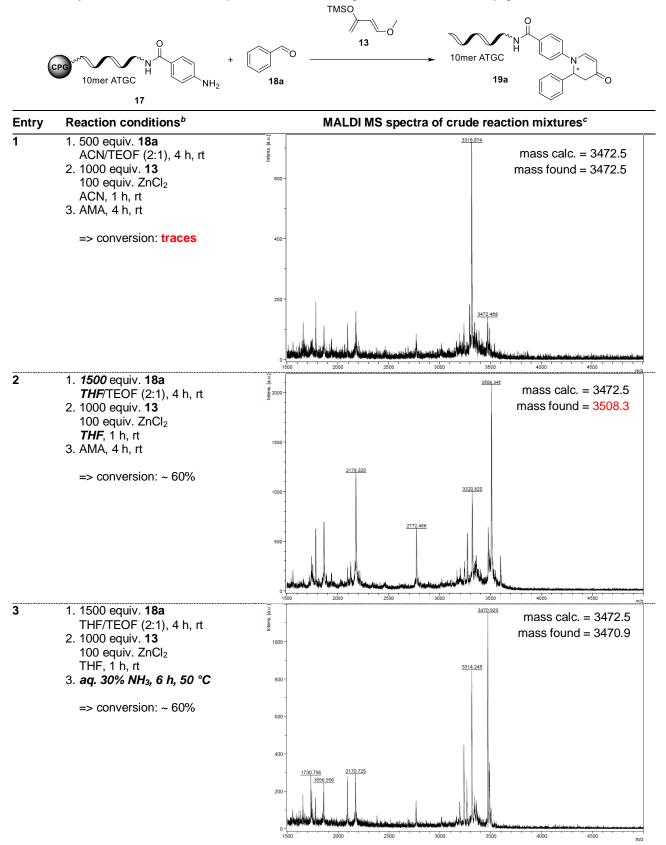
HPLC trace of crude reaction mixture 16r (Analytical RP-HPLC, Method-II)

3500

4000

4500

Table S7Optimization of ZnCl2-mediated aza-Diels-Alder reaction with benzaldehyde18a andDanishefsky's diene 13 on CPG-coupled 10mer ATGC oligonucleotide-aniline conjugate 17.

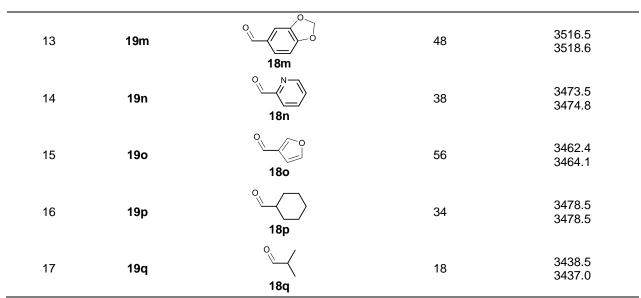


^a Condensation of CPG-coupled 10mer ATGC-aniline conjugate **17** (20 nmol) with benzaldehyde **18a** (X equiv.) in 36 µL of indicated solvent/triethyl orthoformate (2:1) at ambient temperature for 4 h, then ZnCl₂ (100 equiv.) suspended in 30 µL of indicated solvent and Danishefsky's diene **13** (1000 equiv.) were added, the reaction mixture was shaken at ambient temperature for 1 h. ^{*b*} Changed parameters are in bold and italic. ^c Measured by MALDI-MS. 10mer ATGC = 5'-GTC ATG ATC T-3'. n.d. = not detected. TEOF = triethyl orthoformate.

 Table S8 – Scope of ZnCl₂-mediated aza-Diels-Alder reaction with Danishefsky's diene on CPG-coupled

 10mer ATGC oligonucleotide-aniline conjugate using different aldehydes.^a

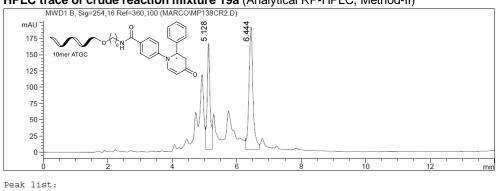
		1. THF/TEOF 2:1 rt, 4 h 2. TMSO		
	0	13		0
CPG	N ^N N ^N	+ P () .	10mer ATGC	
	mer ATGC	¹⁰⁰ equiv. ZnCl ₂ ¹¹⁷ NH ₂ 18 THF, rt, 1 h	19a-q	R R O
	17	3. aq. NH ₃ , 50°C, 6 h		
Entry	Product	Aldehyde	Conversion [%] ^b	Mass _{calc.} Mass _{found} ^c
1	19a	0 18a	62	3472.5 3473.3
2	19b	O Br 18b	60	3551.4 3552.4
3	19c	^o 18c	64	3551.4 3552.5
4	19d	Br 18d	56	3551.4 3551.3
5	19e		58	3502.5 3502.8
6	19 f	°	71	3502.5 3503.2
7	19g		64	3502.5 3503.2
8	19h	18g °F 18h	58	3490.5 3491.7
9	19i	° 18i	57	3486.5 3488.4
10	19j	° 18j	50	3528.6 3529.0
11	19k	° 18k	65	3548.6 3549.0
12	191		60	3522.5 3524.1



^{*a*} Condensation of CPG-coupled oligonucleotide conjugate **17** (20 nmol) with aldehyde **18** (1500 equiv., 30 µmol) in 36 µL tetrahydrofuran/triethyl orthoformate (2:1) at ambient temperature for 4 h, then addition of $ZnCl_2$ (100 equiv., 2 µmol) dissolved in 30 µL tetrahydrofuran and Danishefsky's diene **15** (1000 equiv., 20 µmol) at ambient temperature for 1 h. DNA cleavage with 30 % aqueous ammonia at 50 °C for 6 h. ^{*b*} Determined by analytical RP-HPLC analysis. ^{*c*} Measured by MALDI-MS. 10mer ATGC = 5'-GTC ATG ATC T-3'. n.d. = not detected.

Products of aza-Diels-Alder reaction with Danishefsky's diene on CPG-coupled 10mer ATGC oligonucleotide-aniline conjugate

DNA conjugate 19a: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with benzaldehyde **18a** and Danishefsky's diene **13** according to RP-08.

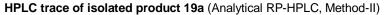


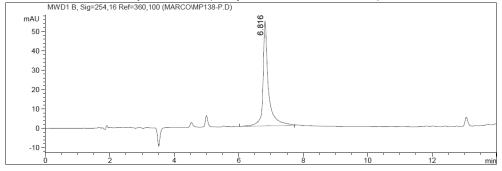
HPLC trace of crude reaction mixture 19a (Analytical RP-HPLC, Method-II)

 Ret. Time
 Width min
 Height
 Area
 Area %

 5.128
 0.101
 163.317
 993.368
 37.589

 6.444
 0.146
 188.069
 1649.351
 62.411

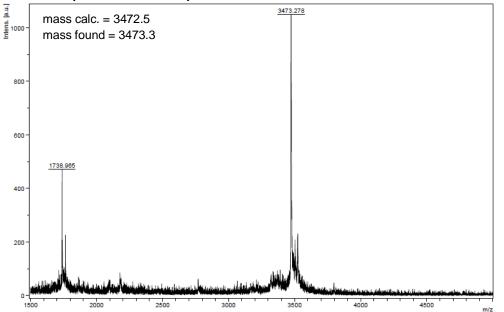




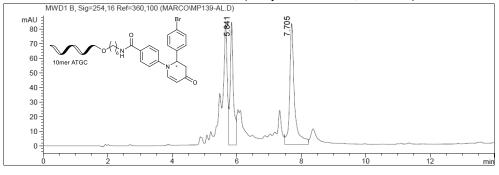
Peak list:

Ret. Time	Width min	Height	Area	Area %
6.816	0.164	54.110	632.311	100.000

MALDI-MS spectrum of isolated product 19a



DNA conjugate 19b: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with 4-bromobenzaldehyde **18b** and Danishefsky's diene **13** according to RP-08.

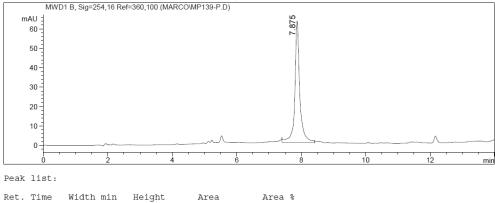


HPLC trace of crude reaction mixture 19b (Analytical RP-HPLC, Method-II)

Peak list:

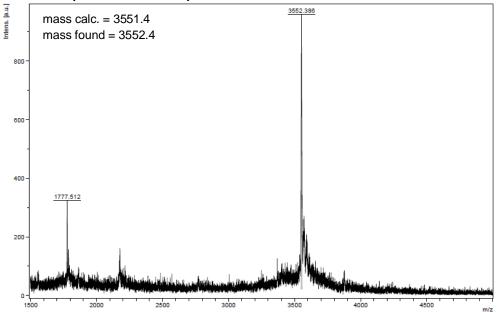
Ret. Time	Width min	Height	Area	Area %
5.841	0.113	84.160	571.159	39.796
7.705	0.173	83.182	864.044	60.204

HPLC trace of isolated product 19b (Analytical RP-HPLC, Method-II)

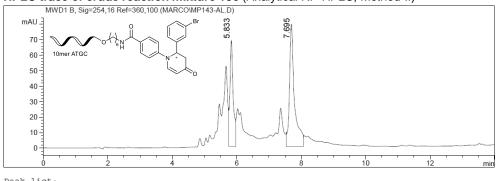


7.875 0.177 62.652 664.216 100.000

MALDI-MS spectrum of isolated product 19b



DNA conjugate 19c: CPG-coupled 10mer ATGC-aniline conjugate 17 was reacted with 3-bromobenzaldehyde 18c and Danishefsky's diene 13 according to RP-08.

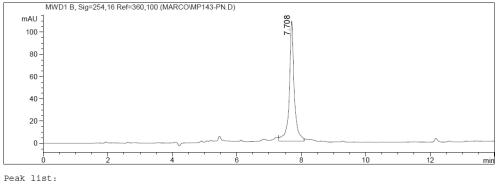


HPLC trace of crude reaction mixture 19c (Analytical RP-HPLC, Method-II)

Peak list:

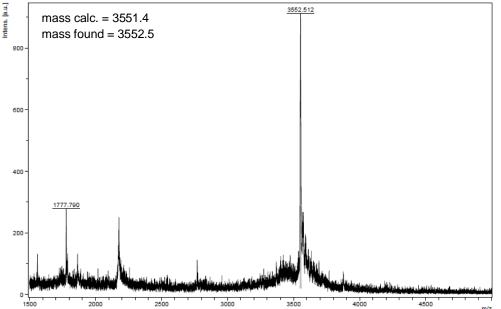
Ret. Time	Width min	Height	Area	Area %
5.833	0.117	68.815	484.264	35.791
7.695	0.183	79.125	868.751	64.209

HPLC trace of isolated product 19c (Analytical RP-HPLC, Method-II)

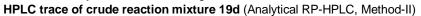


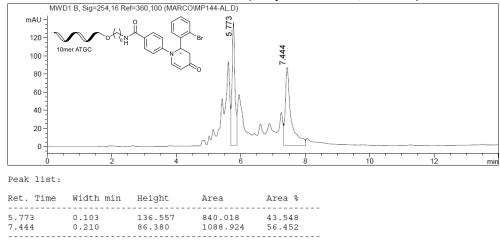
Ret. Time	Width min	Height	Area	Area %
7.708	0.177	108.018	1146.097	100.000

MALDI-MS spectrum of isolated product 19c

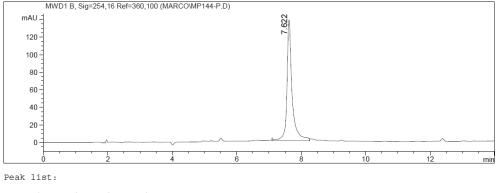


DNA conjugate 19d: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with 2-bromobenzaldehyde **18d** and Danishefsky's diene **13** according to RP-08.

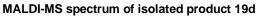


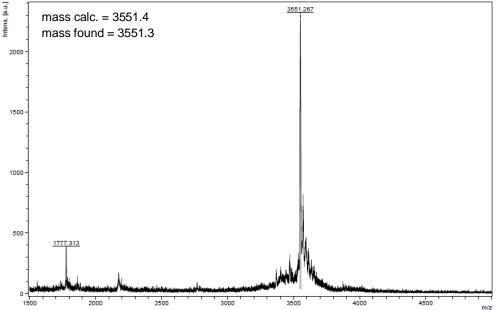




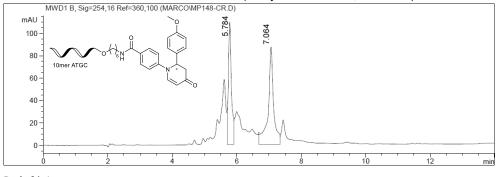


Ret. Time	Width min	Height	Area	Area %
7.622	0.185	137.808	1527.506	100.000





DNA conjugate 19e: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with 4-methoxybenzaldehyde **18e** and Danishefsky's diene **13** according to RP-08.

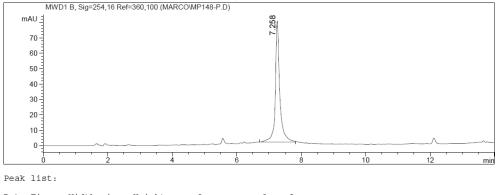


HPLC trace of crude reaction mixture 19e (Analytical RP-HPLC, Method-II)

Peak list:

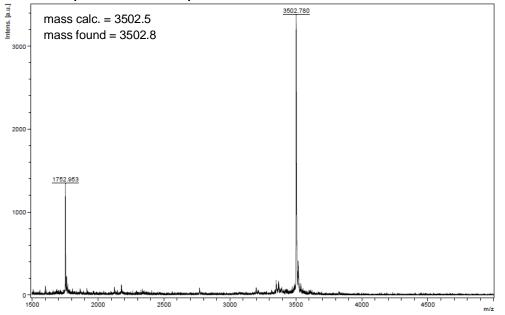
Ret. Time	Width min	Height	Area	Area %
5.784	0.111	109.344	725.500	41.666
7.064	0.194	87.057	1015.736	58.334

HPLC trace of isolated product 19e (Analytical RP-HPLC, Method-II)

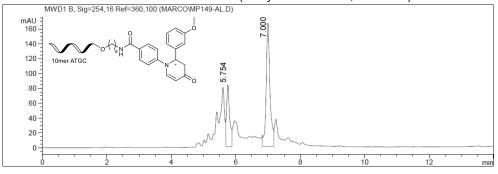


Ret. Time	Width min	Height	Area	Area %
7.258	0.162	78.696	763.349	100.000

MALDI-MS spectrum of isolated product 19e



DNA conjugate 19f: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with 3-methoxybenzaldehyde **18f** and Danishefsky's diene **13** according to RP-08.

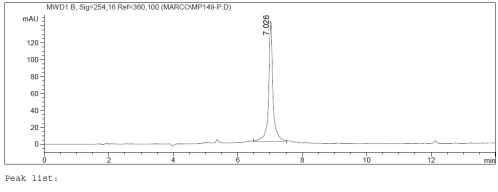


HPLC trace of crude reaction mixture 19f (Analytical RP-HPLC, Method-II)

Peak list:

Ret. Time	Width min	Height	Area	Area %
5.754	0.111	83.312	555.542	28.989
7.000	0.136	166.735	1360.813	71.011

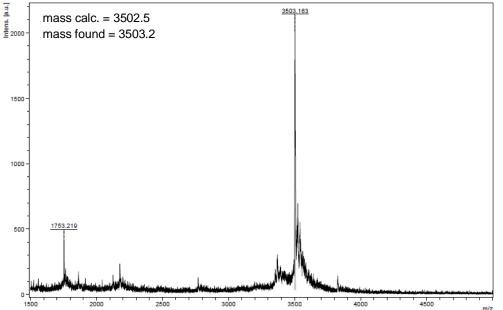
HPLC trace of isolated product 19f (Analytical RP-HPLC, Method-II)



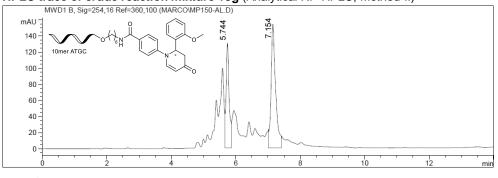
 Ret. Time
 Width min
 Height
 Area
 Area %

 7.026
 0.155
 142.647
 1325.466
 100.000

MALDI-MS spectrum of isolated product 19f



DNA conjugate 19g: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with 2-methoxybenzaldehyde **18g** and Danishefsky's diene **13** according to RP-08.

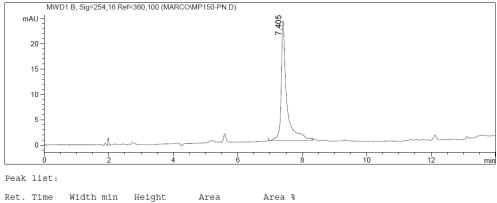


HPLC trace of crude reaction mixture 19g (Analytical RP-HPLC, Method-II)

Peak list:

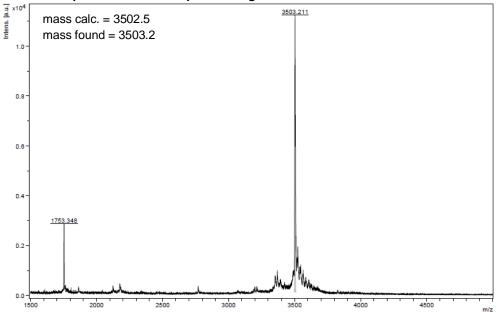
Ret. Time	Width min	Height	Area	Area %
5.744	0.108	130.706	845.267	35.788
7.154	0.164	154.065	1516.604	64.212

HPLC trace of isolated product 19g (Analytical RP-HPLC, Method-II)

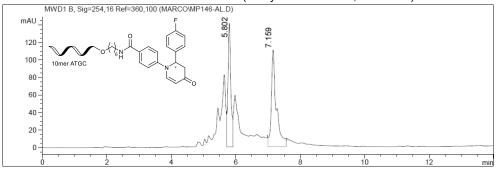


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 Ride
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MALDI-MS spectrum of isolated product 19g



DNA conjugate 19h: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with 4-fluorobenzaldehyde **18h** and Danishefsky's diene **13** according to RP-08.

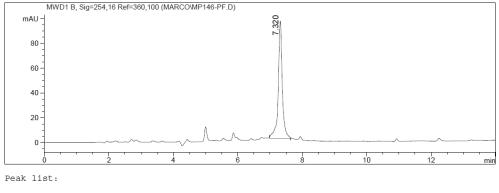


HPLC trace of crude reaction mixture 19h (Analytical RP-HPLC, Method-II)

Peak list:

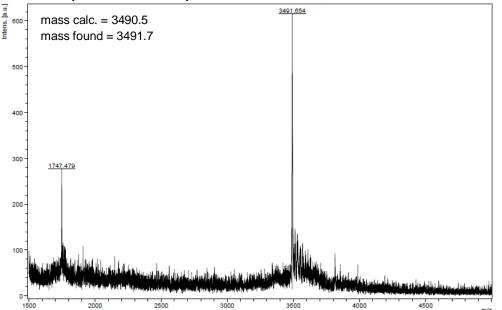
Ret. Time	Width min	Height	Area	Area %
5.802	0.100	141.245	847.449	41.605
7.159	0.180	110.390	1189.461	58.395

HPLC trace of isolated product 19h (Analytical RP-HPLC, Method-II)

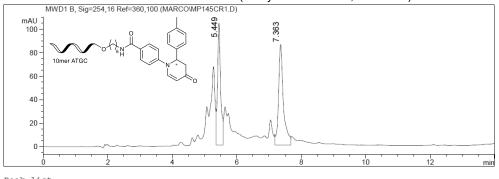


Ret. Time	Width min	Height	Area	Area %
7.320	0.145	94.654	824.639	100.000

MALDI-MS spectrum of isolated product 19h



DNA conjugate 19i: CPG-coupled 10mer ATGC-aniline conjugate 17 was reacted with 4-methylbenzaldehyde 18i and Danishefsky's diene 13 according to RP-08.

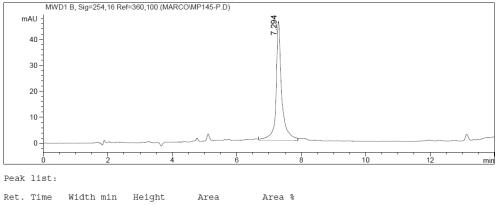


HPLC trace of crude reaction mixture 19i (Analytical RP-HPLC, Method-II)

Peak list:

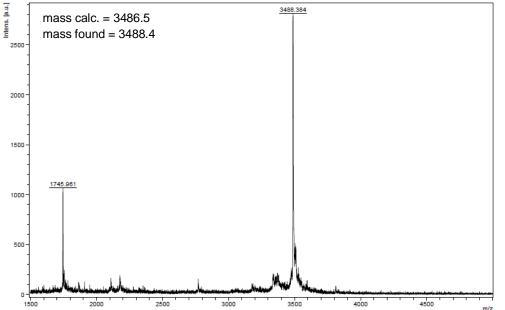
Ret. Time	Width min	Height	Area	Area %
5.449	0.110	104.130	688.043	43.079
7.363	0.177	85.811	909.125	56.921

HPLC trace of isolated product 19i (Analytical RP-HPLC, Method-II)

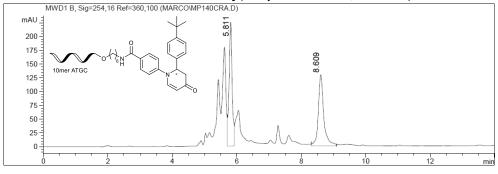


7.294 0.173 45.899 564.514 100.000

MALDI-MS spectrum of isolated product 19i



DNA conjugate 19j: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with 4-*tert*-butylbenzaldehyde **18j** and Danishefsky's diene **13** according to RP-08.

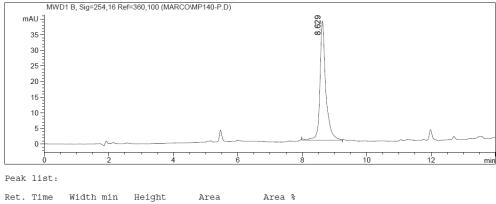


HPLC trace of crude reaction mixture 19j (Analytical RP-HPLC, Method-II)

Peak list:

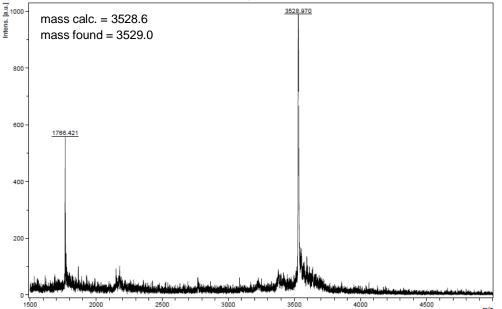
Ret. Time	Width min	Height	Area	Area %
5.811	0.111	225.544	1505.354	49.837
8.609	0.196	128.977	1515.207	50.163

HPLC trace of isolated product 19j (Analytical RP-HPLC, Method-II)

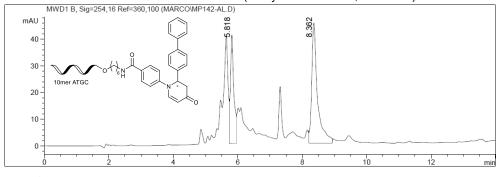


8.629 0.210 38.134 480.948 100.000

MALDI-MS spectrum of isolated product 19j



DNA conjugate 19k: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with 4-phenylbenzaldehyde **18k** and Danishefsky's diene **13** according to RP-08.

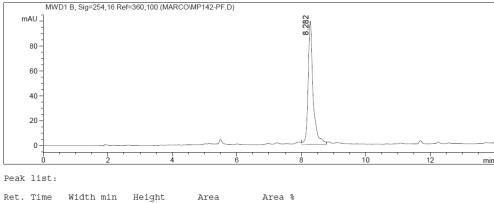


HPLC trace of crude reaction mixture 19k (Analytical RP-HPLC, Method-II)

Peak list:

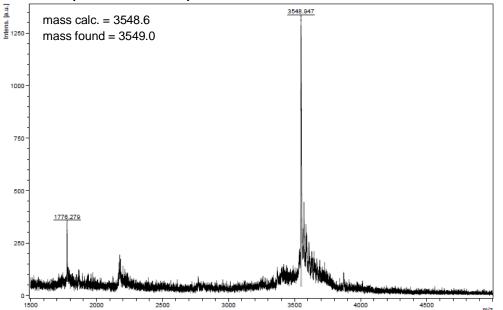
Ret. Time	Width min	Height	Area	Area %
5.818	0.125	40.785	305.138	35.237
8.362	0.208	44.986	560.832	64.763

HPLC trace of isolated product 19k (Analytical RP-HPLC, Method-II)

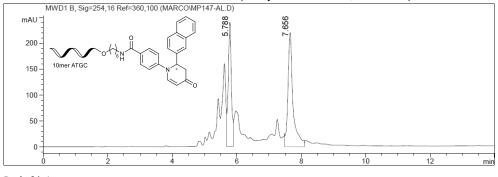


 Ret:
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MALDI-MS spectrum of isolated product 19k



DNA conjugate 19I: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with 2-Naphthaldehyde **18I** and Danishefsky's diene **13** according to RP-08.

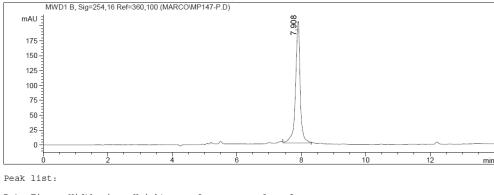


HPLC trace of crude reaction mixture 19I (Analytical RP-HPLC, Method-II)

Peak list:

Ret. Time	Width min	Height	Area	Area %
5.788	0.113	240.849	1630.942	40.280
7.656	0.183	220.402	2418.100	59.720

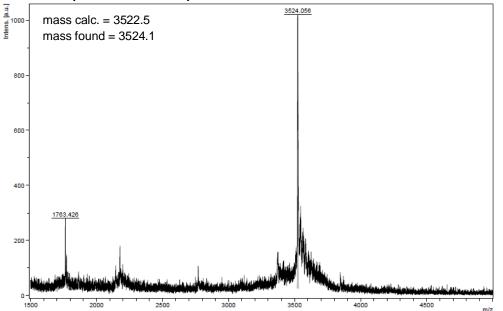
HPLC trace of isolated product 19I (Analytical RP-HPLC, Method-II)



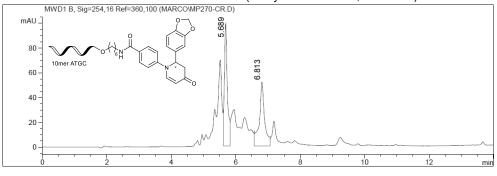
 Ret. Time
 Width min
 Height
 Area
 Area %

 7.908
 0.167
 204.996
 2055.109
 100.000

MALDI-MS spectrum of isolated product 19I



DNA conjugate 19m: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with piperonyl aldehyde **18m** and Danishefsky's diene **13** according to RP-08.

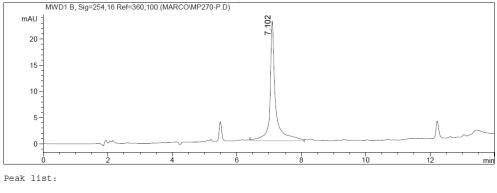


HPLC trace of crude reaction mixture 19m (Analytical RP-HPLC, Method-II)

Peak list:

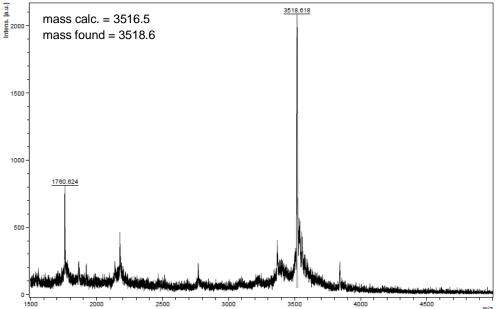
Ret. Time	Width min	Height	Area	Area %
5.689	0.107	99.124	638.691	52.502
6.813	0.187	51.512	577.815	47.498

HPLC trace of isolated product 19m (Analytical RP-HPLC, Method-II)

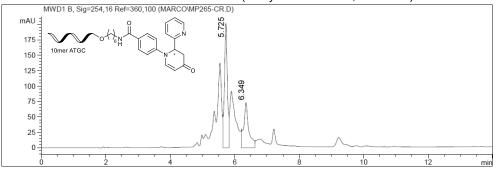


Ret. Time	Width min	Height	Area	Area %
7.102	0.163	22.801	272.139	100.000

MALDI-MS spectrum of isolated product 19m



DNA conjugate 19n: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with 2-pyridinecarbox-aldehyde **18n** and Danishefsky's diene **13** according to RP-08.

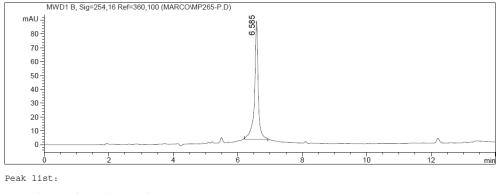


HPLC trace of crude reaction mixture 19n (Analytical RP-HPLC, Method-II)

Peak list:

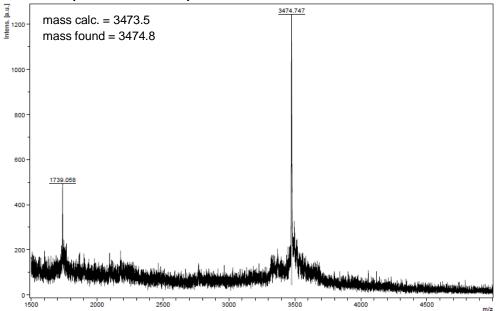
Ret. Time	Width min	Height	Area	Area %
5.725	0.106	202.771	1295.430	61.531
6.349	0.183	73.610	809.890	38.469

HPLC trace of isolated product 19n (Analytical RP-HPLC, Method-II)

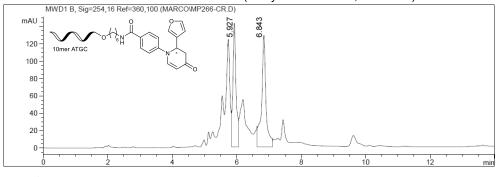


Ret. Time	Width min	Height	Area	Area %
6.585	0.133	85.834	684.034	100.000

MALDI-MS spectrum of isolated product 19n



DNA conjugate 19o: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with 3-furancarboxaldehyde **18o** and Danishefsky's diene **13** according to RP-08.

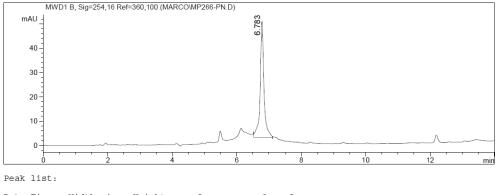


HPLC trace of crude reaction mixture 19o (Analytical RP-HPLC, Method-II)

Peak list:

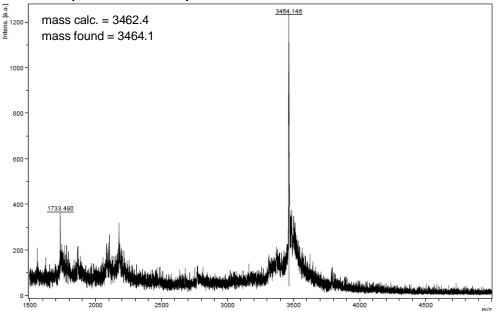
Ret. Time	Width min	Height	Area	Area %
5.927	0.113	143.131	972.186	44.381
6.843	0.157	129.107	1218.370	55.619

HPLC trace of isolated product 19o (Analytical RP-HPLC, Method-II)

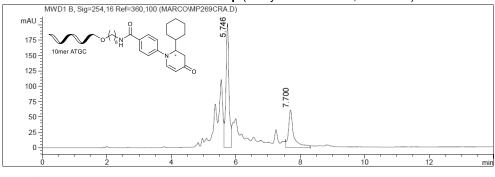


6.783	3	0.137		47.909	392.414	100.000
Ret.	Time	Width 1	min	Height	Area	Area %

MALDI-MS spectrum of isolated product 190



DNA conjugate 19p: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with cyclohexanecarbox-aldehyde **18p** and Danishefsky's diene **13** according to RP-08.

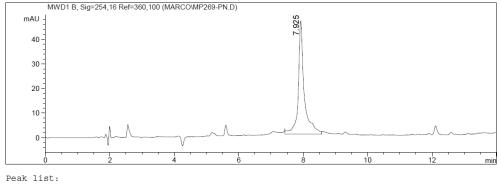


HPLC trace of crude reaction mixture 19p (Analytical RP-HPLC, Method-II)

Peak list:

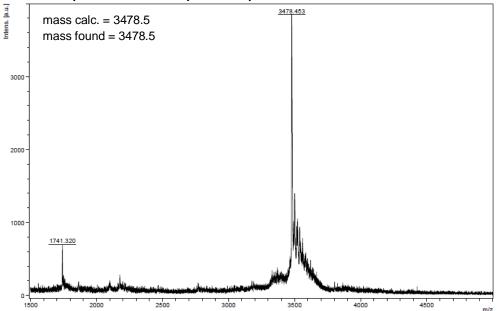
Ret. Time	Width min	Height	Area	Area %
5.746	0.109	203.173	1326.133	65.765
7.700	0.189	60.841	690.349	34.235

HPLC trace of isolated product 19p (Analytical RP-HPLC, Method-II)

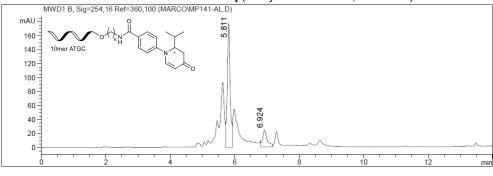


Ret. Time	Width min	Height	Area	Area %
7.925	0.213	46.069	590.128	100.000

MALDI-MS spectrum of isolated product 19p



DNA conjugate 19q: CPG-coupled 10mer ATGC-aniline conjugate **17** was reacted with isobutyraldehyde **18q** and Danishefsky's diene **13** according to RP-08.



HPLC trace of crude reaction mixture 19q (Analytical RP-HPLC, Method-II)

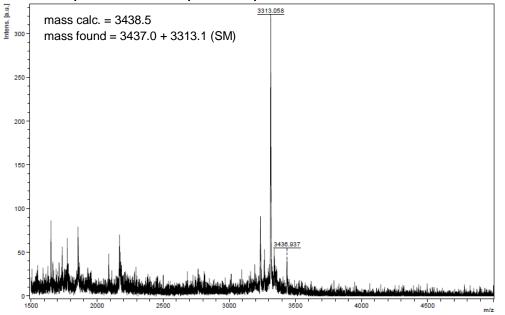
Peak list:

 Ret. Time
 Width min
 Height
 Area
 Area %

 5.811
 0.102
 178.303
 1095.891
 81.668

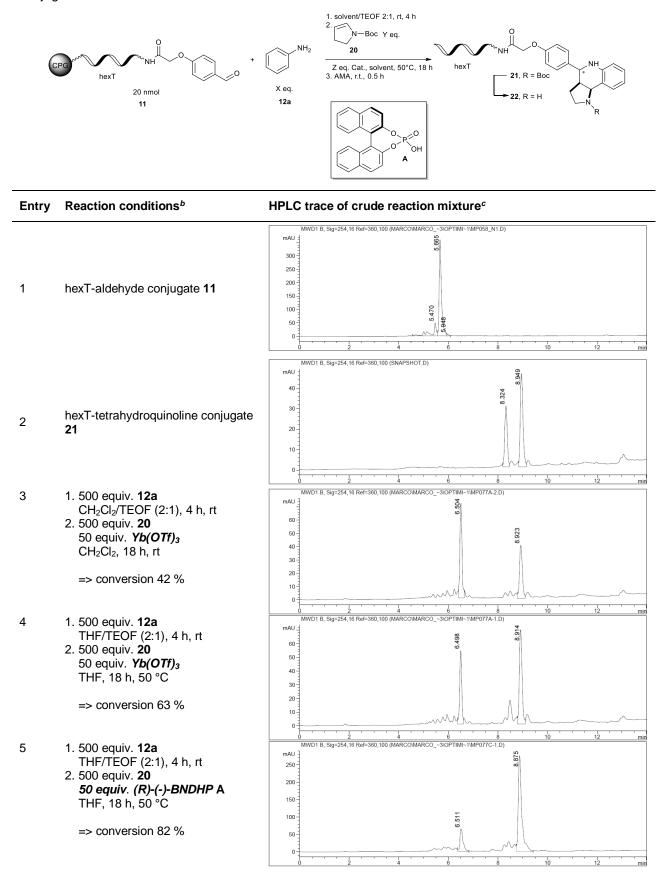
 6.924
 0.169
 24.237
 245.991
 18.332

MALDI-MS spectrum of isolated product 19q



(R)-(-)-BNDHP-mediated Povarov reaction on CPG-coupled oligonucleotides

Table S9 Optimization of (R)-(-)-BNDHP-mediated Povarov reaction CPG-coupled hexT-aldehyde conjugate.^{*a*}



MWD1 B, Sig=254,16 Ref=360,100 (MARCO\MP077B-2.D) 6 1. 500 equiv. 12a mAU -140 -THF/TEOF (2:1), 4 h, rt 2. 500 equiv. 20 120 100 -100 equiv. (R)-(-)-BNDHP A 80 THF, 18 h, 50 °C 60 · 40-=> conversion 94 % 20-0 2 4 MWD1 B, Sig=254,16 Ref=360,100 (SNAPSHOT.D 7 1. 500 equiv. 12a mAU _ THF/TEOF (2:1), 4 h, rt 140 -2. 500 equiv. 20 120 -THF, 18 h, 50 °C 100 -80 -60 -=> conversion 34 % 40 -20 -0 2 4 6 8 MWD1 B, Sig=254,16 Ref=360,100 (MARCO\MARCO_~3\OPTIMI~1\MP077C-2.D) 8 1. 1000 equiv. 12a mAU 160 -THF/TEOF (2:1), 4 h, rt 140 2. 500 equiv. 20 120 -50 equiv. (R)-(-)-BNDHP A 100 -THF, 18 h, 50 °C 80 -60 40 => conversion 55 % 20 · 9 1. 2000 equiv. 12a mAU THF/TEOF (2:1), 4 h, rt 300 2. 500 equiv. 20 250 50 equiv. (R)-(-)-BNDHP A 200 THF, 18 h, 50 °C 150 -100 -=> conversion 11 % 50· 0. 10 1. 4000 equiv. 12a mAU 🛓 THF/TEOF (2:1), 4 h, rt 2. 500 equiv. 20 400 -50 equiv. (R)-(-)-BNDHP A 300 -THF, 18 h, 50 °C 200 => conversion 7 % 100 -0. 11 1. 500 equiv. 12a mAU 200 THF/TEOF (2:1), 4 h, rt 175 2. 1000 equiv. 20 150 50 equiv. (R)-(-)-BNDHP A 125 -THF, 18 h, 50 °C 100 · 75 50 · => conversion 63 % 25 0 1. 500 equiv. **12a** THF/TEOF (2:1), 4 h, rt 12 mAU -140 2. 2000 equiv. 20 120 -50 equiv. (R)-(-)-BNDHP A 100 -80-THF, 18 h, 50 °C

PTIMI~1\MP077C-4.D) WD1 B, Sig=254,16 Ref=360,100 (MARCO\MARCO_~3\0 3.516 3.897 2 4 6 8 MWD1 B, Sig=254,16 Ref=360,100 (MARCO\MARCO ~3\OPTIMI~1\MP077C-3.D) 504 3.869 2 4 6 8 WD1 B, Sig=254,16 Ref=360,100 (MARCO\MARCO_~3\OPTIMI~1\MP077D-2.D) 834 6.515 2 4 6 8 MWD1 B, Sig=254,16 Ref=360,100 (MARCO\MARCO_~3\OPTIMI~1\MP077D-3.D 88 5.511 60 -40· 20

908

905

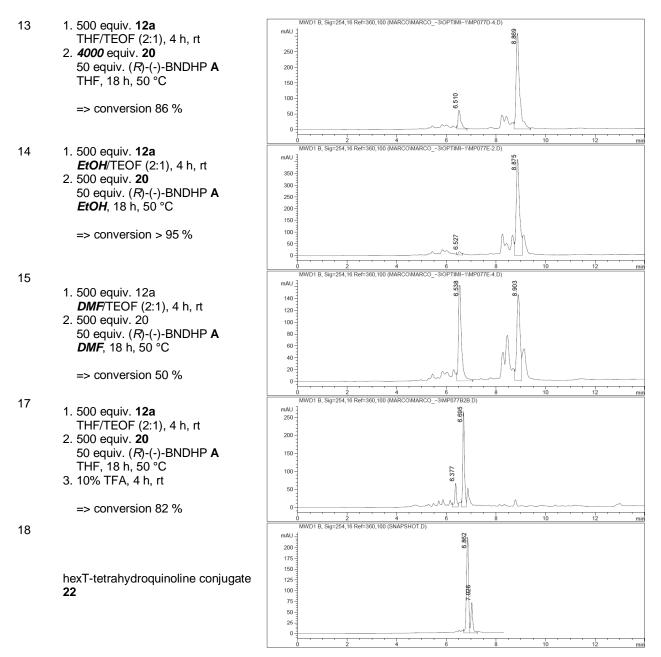
.855

6.484

5.464

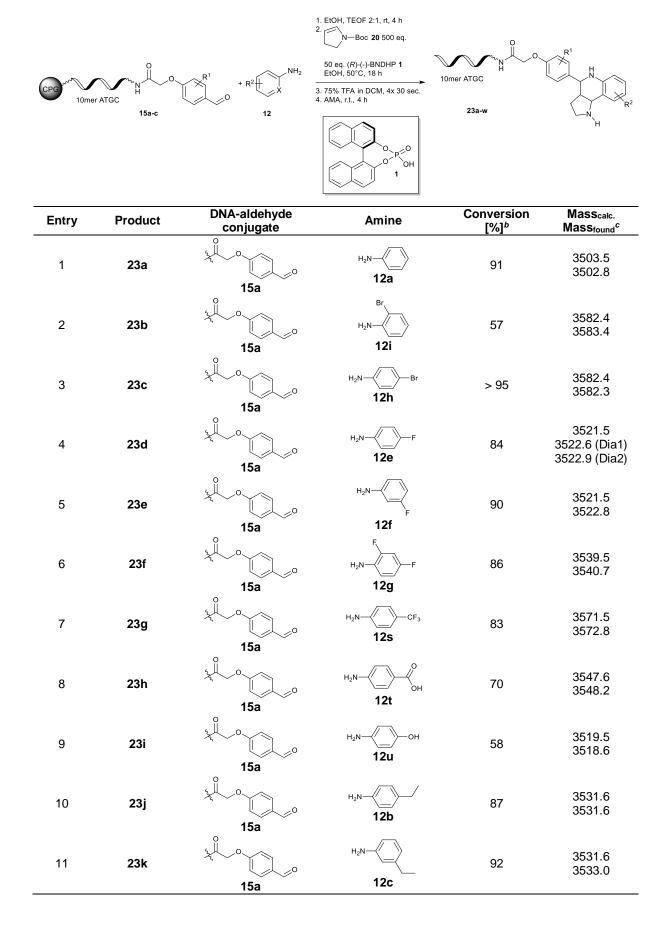
494

=> conversion 67 %



^a Condensation of CPG-coupled hexT conjugate **11a** (20 nmol) with aniline **12a** (X equiv.) in 36 μ L of indicated solvent/triethyl orthoformate (2:1) at ambient temperature for 4 h, then (*R*)-(-)-BNDHP **A** (X equiv.) dissolved in 30 μ L of indicated solvent and *N*-Boc-2,3-dihydro-1H-pyrrole **20** (X equiv.) were added, the reaction mixture was shaken at indicated temperature for 16 h. ^{*b*} parameters that were changed are in bold and italic. ^{*c*} Analytical RP-HPLC, Method-III. TEOF = triethyl orthoformate.

Table S10 – Scope of (R)-(-)-BNDHP-mediated Povarov reaction on CPG-coupled 10mer ATGC oligonucleotide-aldehyde conjugate.^{*a*}

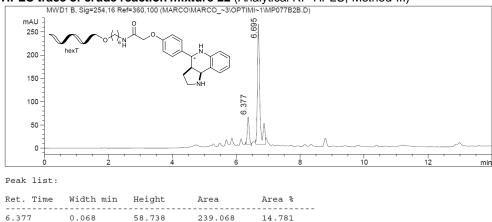


12	231	م م ا ا ا ا ا ا ا ا ا ا ا ا ا ا ا ا ا ا	H ₂ N-	39	3531.6 3532.7
13	23m	<u>م</u> لب م 15a	H ₂ N 12k	< 5	3559.7 n.d.
14	23n	م بر 15a	H ₂ N	82	3563.6 3564.5
15	230	م بر 15a	H ₂ N-0 12I	86	3563.6 3561.7
16	23p	ی بر 15a	H ₂ N-	94	3527.6 3527.9
17	23q	ی بر 15a	H ₂ N	>95	3571.6 3573.3
18	23r	یک 15a	H ₂ N HO 12v	75	3533.6 3534.3
19	23s	یک 15a	H ₂ N- 12w	25	3593.7 3591.4
20	23t	ی 15a	H ₂ N	< 5	3504.5 n.d.
21	23u	نجل ٥ 15a	H ₂ N- 12x	< 5	3505.5 n.d.
22	23v		H ₂ N-	93	3538.0 3537.6
23	23w	<u>الم</u>	н ₂ N	< 5	3553.6 n.d.

^a Condensation of CPG-coupled oligonucleotide conjugate **15** (20 nmol) with aniline **12** (500 equiv., 10 μmol) in 36 μL ethanol/triethyl orthoformate (2:1) at ambient temperature for 4 h, then addition of (*R*)-(-)-BNDHP **A** (50 equiv., 1 μmol) dissolved in 30 μL ethanol and *N*-Boc-2,3-dihydro-1H-pyrrole **20** (500 equiv., 10 μmol) at 50 °C for 16 h. 75% TFA in dichloromethane 4x 30 seconds at ambient temperature. DNA cleavage AMA (30 % aqueous ammonia / 40 % aqueous methylamine, 1:1 (vol/vol)) at ambient temperature for 4 h. ^b Determined by analytical RP-HPLC analysis. ^c Measured by MALDI-MS. 10mer ATGC = 5'-GTC ATG ATC T-3'. n.d. = not detected.

Products of Povarov reaction on CPG-coupled oligonucleotide-aldehyde conjugate

DNA conjugate 22: CPG-coupled hexT-aldehyde conjugate **11** was reacted with aniline **12a** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.



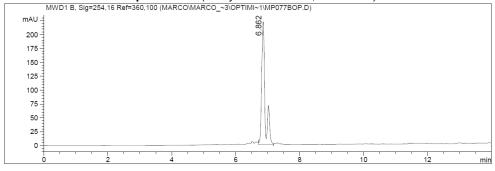
85.219

HPLC trace of crude reaction mixture 22 (Analytical RP-HPLC, Method-III)

HPLC trace of isolated product 22 (Analytical RP-HPLC, Method-III)

1378.315

259.027



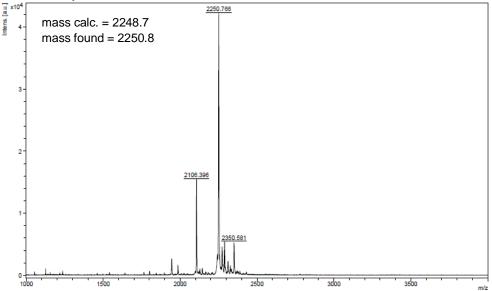
Peak list:

6.695

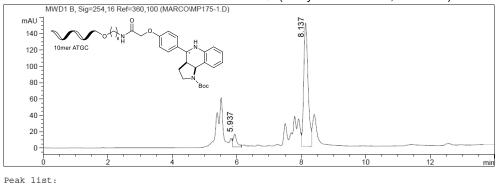
0.089

Ret. Time	Width min	Height	Area	Area %
6.862	0.123	222.063	1633.396	100.000

MALDI-MS spectrum of crude reaction mixture 22



DNA conjugate 23a: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with aniline **12a** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.



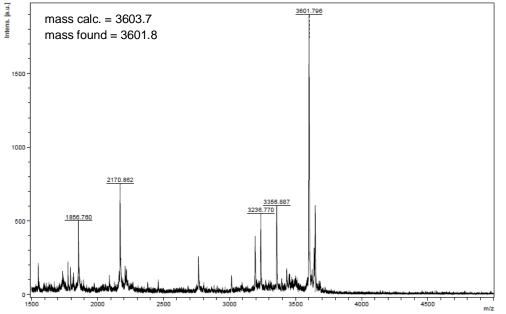
HPLC trace of crude reaction mixture 23a-Bocon (Analytical RP-HPLC, Method-II)

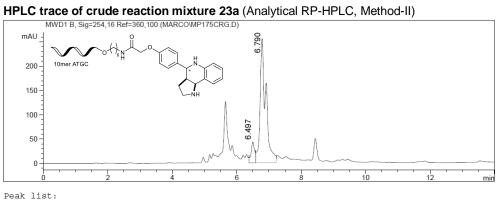
 Ret. Time
 Width min
 Height
 Area
 Area %

 5.937
 0.127
 15.722
 119.679
 7.863

 8.137
 0.155
 150.913
 1402.450
 92.137

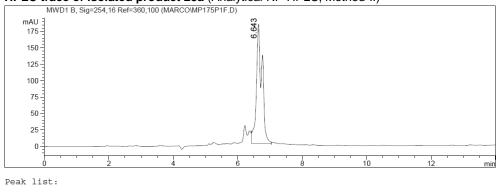






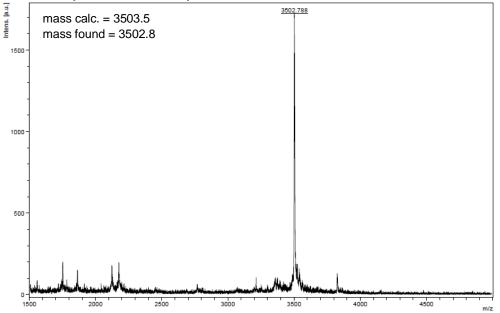
Ret. Time	Width min	Height	Area	Area %
6.497	0.125	43.055	321.970	9.172
6.790	0.208	255.855	3188.387	90.828

HPLC trace of isolated product 23a (Analytical RP-HPLC, Method-II)

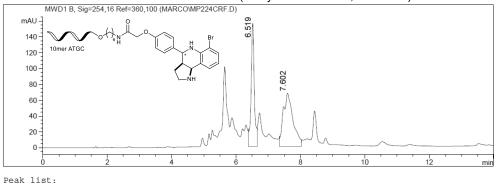


Ret. Time	Width min	Height	Area	Area %
6.643	0.204	182.113	2225.986	100.000

MALDI-MS spectrum of isolated product 23a



DNA conjugate 23b: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 2-bromoaniline **12i** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.



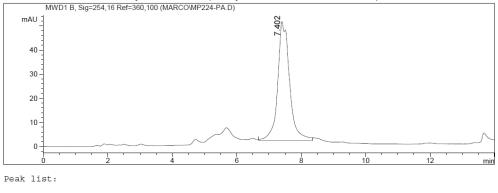
Area %

HPLC trace of crude reaction mixture 23b (Analytical RP-HPLC, Method-II)

Ret. Time Width min Height Area

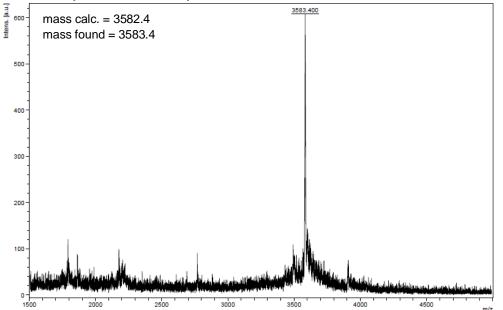
		07.042	1417.042		_
7 602	0.352	67 042	1/17 8/2	57 487	
6 519	0 112	155 924	1048 529	42 513	

HPLC trace of isolated product 23b (Analytical RP-HPLC, Method-II)

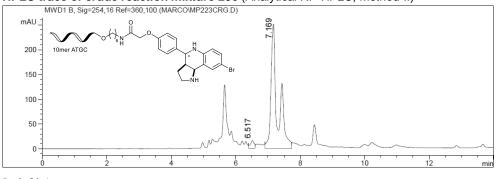


Ret. Time	Width min	Height	Area	Area %
7.402	0.431	49.421	1277.357	100.000

MALDI-MS spectrum of isolated product 23b



DNA conjugate 23c: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 4-bromoaniline **12h** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

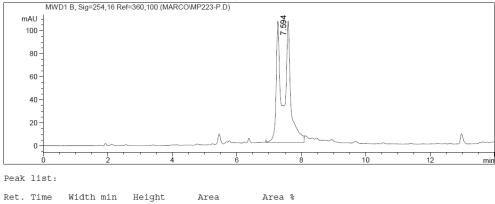


HPLC trace of crude reaction mixture 23c (Analytical RP-HPLC, Method-II)

Peak list:

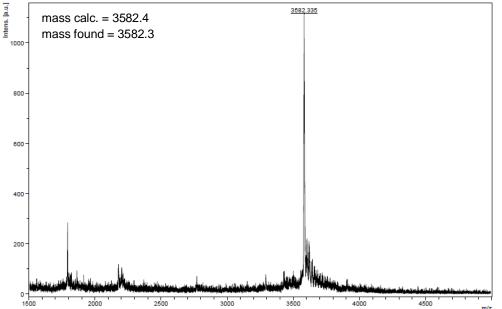
Ret. Time	Width min	Height	Area	Area %
6.517	0.135	14.963	120.802	3.374
7.169	0.230	250.566	3459.788	96.626

HPLC trace of isolated product 23c (Analytical RP-HPLC, Method-II)

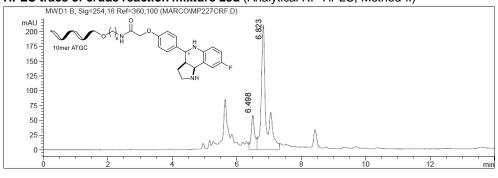


7.594 0.326 105.461 2063.898 100.000

MALDI-MS spectrum of isolated product 23c



DNA conjugate 23d: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 4-fluoroaniline **12e** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

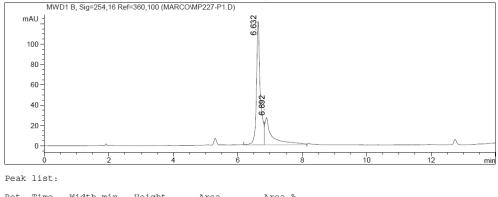


HPLC trace of crude reaction mixture 23d (Analytical RP-HPLC, Method-II)

Peak list:

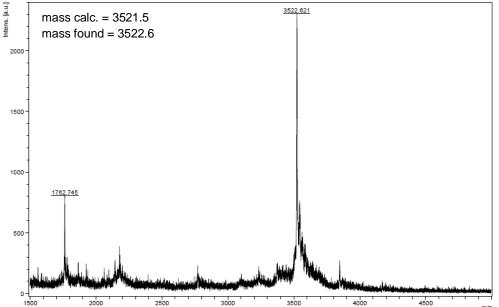
Ret. Time	Width min	Height	Area	Area %
6.498	0.121	58.176	424.096	15.997
6.823	0.176	210.615	2227.069	84.003

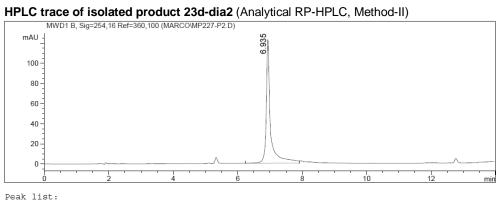
HPLC trace of isolated product 23d-dia1 (Analytical RP-HPLC, Method-II)



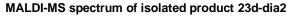
Ret. Time	Width min	Height	Area	Area %	
6.632	0.117	121.831	1014.094	69.760	
6.892	0.211	26.848	439.587	30.240	

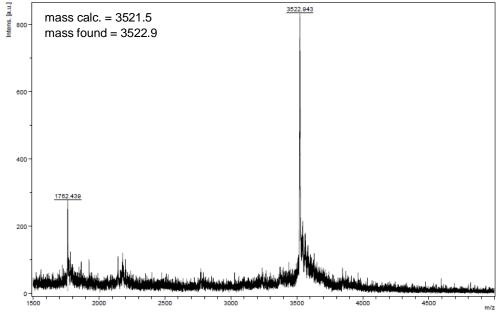
MALDI-MS spectrum of isolated product 23d-dia1



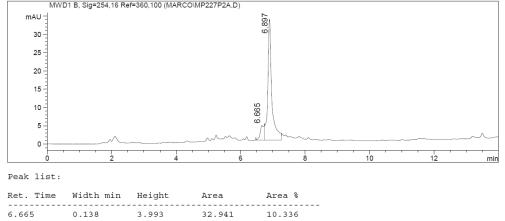


Ret.	Time	Width	min	Height	Area	Area %
6.93	5	0.130		122.672	1140.619	100.000



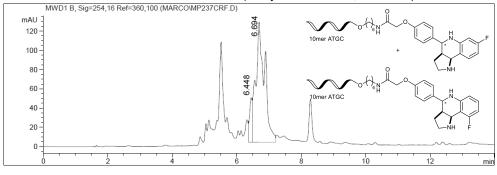


HPLC trace of isolated product 23d-dia2 after treatment with AMA (Analytical RP-HPLC, Method-II) MWD1 B, Sig=254,16 Ref=360,100 (MARCOIMP227P2A.D)



6.897	0.144	33.125	285.757	89.664

DNA conjugate 23e: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 3-fluoroaniline **12f** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

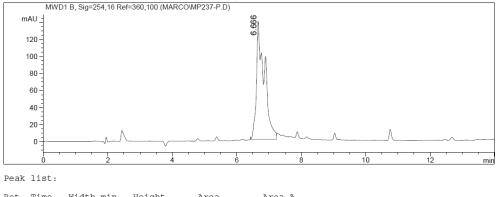


HPLC trace of crude reaction mixture 23e (Analytical RP-HPLC, Method-II)

Peak list:

Ret. Time	Width min	Height	Area	Area %
6.448	0.094	46.279	260.954	9.840
6.694	0.319	125.116	2391.062	90.160

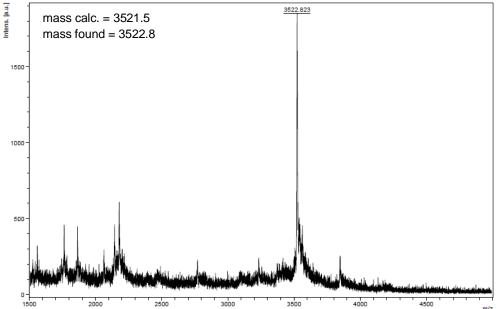
HPLC trace of isolated product 23e (Analytical RP-HPLC, Method-II)



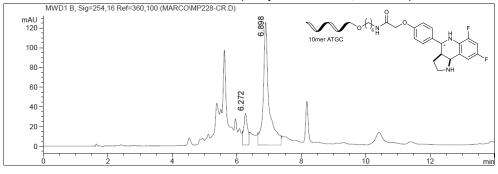
 Ret. Time
 Width min
 Height
 Area
 Area %

 6.666
 0.289
 138.906
 2411.832
 100.000

MALDI-MS spectrum of isolated product 23e



DNA conjugate 23f: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 2,4-difluoroaniline **12g** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

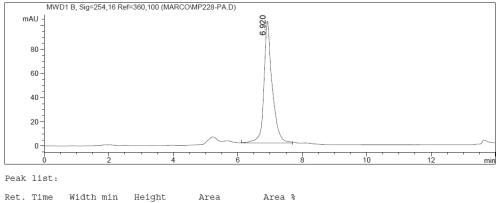


HPLC trace of crude reaction mixture 23f (Analytical RP-HPLC, Method-II)

Peak list:

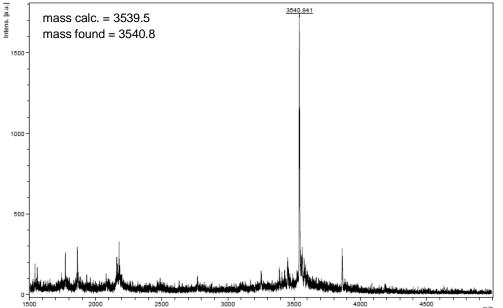
Ret. Time	Width min	Height	Area	Area %
6.272	0.134	32.186	259.067	14.481
6.898	0.204	124.958	1529.998	85.519

HPLC trace of isolated product 23f (Analytical RP-HPLC, Method-II)

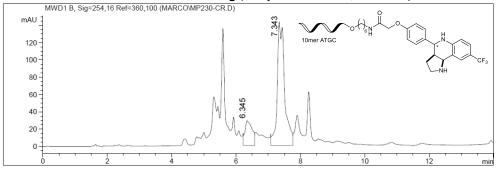


6.920 0.295 100.973 1786.442 100.000

MALDI-MS spectrum of isolated product 23f



DNA conjugate 23g: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 4-trifluoromethylaniline **12s** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

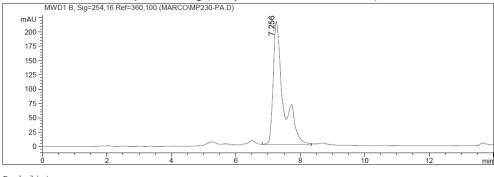


HPLC trace of crude reaction mixture 23g (Analytical RP-HPLC, Method-II)

Peak list:

Ret. Time	Width min	Height	Area	Area %
6.345 7.343	0.264	28.950	458.510	16.627
/.343	0.268	143.194	2299.069	83.373

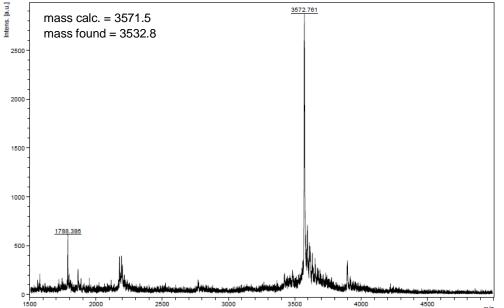
HPLC trace of isolated product 23g (Analytical RP-HPLC, Method-II)



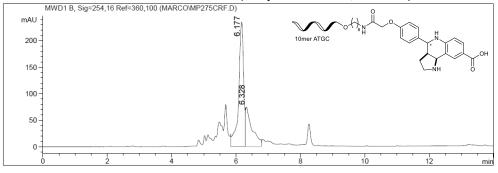
Peak list:

Ret. Time	Width min	Height	Area	Area %
7.256	0.366	214.644	4716.933	100.000

MALDI-MS spectrum of isolated product 23g



DNA conjugate 23h: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 4-aminobenzoic acid **12t** and *N*-Boc-2,3-dihydro-1H-pyrrole **15a** according to RP-09.

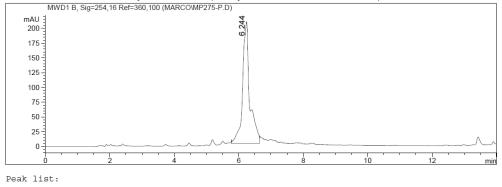


HPLC trace of crude reaction mixture 23h (Analytical RP-HPLC, Method-II)

Peak list:

Ret. Time	Width min	Height	Area	Area %
6.177	0.177	234.443	2494.980	70.214
6.328	0.238	74.245	1058.421	29.786

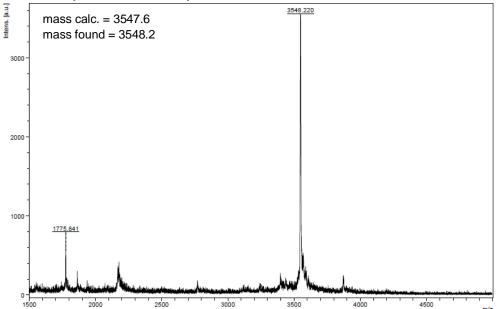
HPLC trace of isolated product 23h (Analytical RP-HPLC, Method-II)



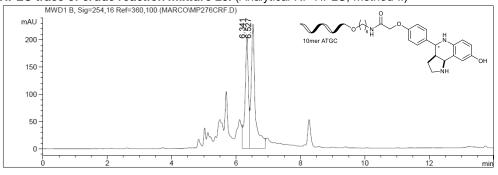
 Ret. Time
 Width min
 Height
 Area
 Area %

 6.244
 0.255
 206.604
 3157.160
 100.000

MALDI-MS spectrum of isolated product 23h



DNA conjugate 23i: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 4-hydroxyaniline **12u** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

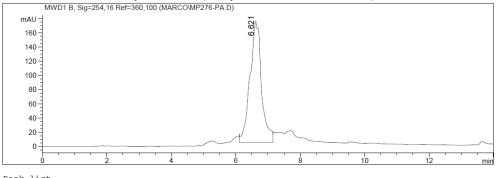


HPLC trace of crude reaction mixture 23i (Analytical RP-HPLC, Method-II)

Peak list:

Ret. Time	Width min	Height	Area	Area %
6.341	0.118	204.855	1456.078	41.891
6.527	0.147	228.536	2019.755	58.109

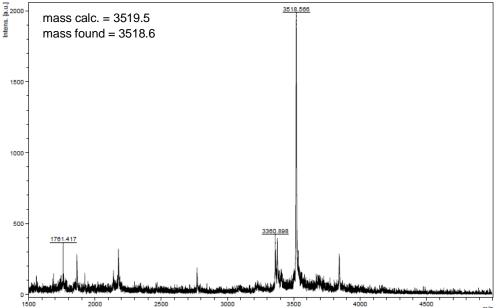
HPLC trace of isolated product 23i (Analytical RP-HPLC, Method-II)



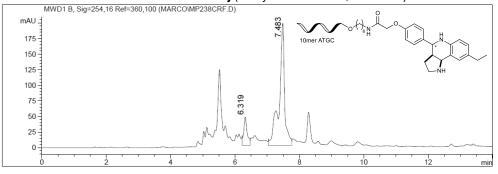
Peak list:

Ret. Time	Width min	Height	Area	Area %
6.621	0.416	170.139	4249.200	100.000

MALDI-MS spectrum of isolated product 23i



DNA conjugate 23j: CPG-coupled 10mer ATGC-aldehyde conjugate 15a was reacted with 4-ethylaniline 12b and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

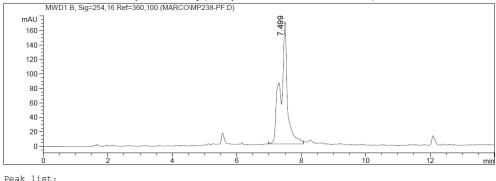


HPLC trace of crude reaction mixture 23j (Analytical RP-HPLC, Method-II)

Peak list:

Ret. Time	Width min	Height	Area	Area %
6.319	0.126	45.753	344.902	13.000
7.483	0.196	196.398	2308.146	87.000

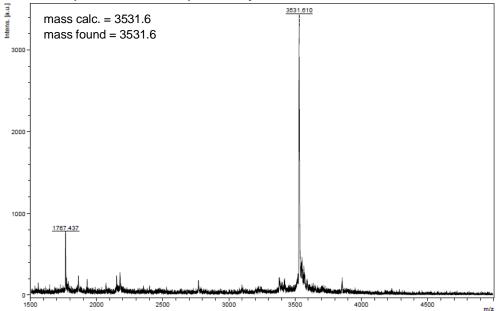
HPLC trace of isolated product 23j (Analytical RP-HPLC, Method-II)



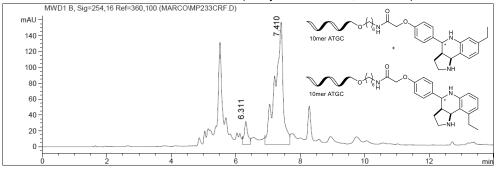
Peak list:

Ret. Time	Width min	Height	Area	Area %
7.499	0.248	169.478	2520.448	100.000

MALDI-MS spectrum of isolated product 23j



DNA conjugate 23k: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 3-ethylaniline **12c** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

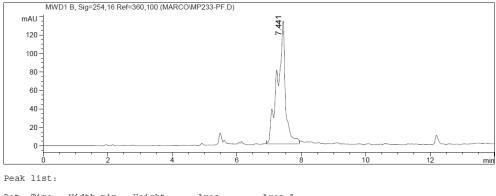


HPLC trace of crude reaction mixture 23k (Analytical RP-HPLC, Method-II)

Peak list:

Ret. Time	Width min	Height	Area	Area %
6.311	0.129	28.811	223.097	7.763
7.410	0.286	154.498	2650.679	92.237

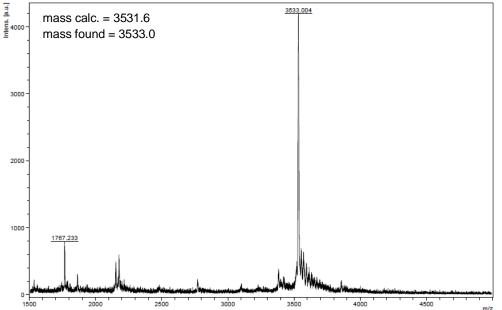
HPLC trace of isolated product 23k (Analytical RP-HPLC, Method-II)



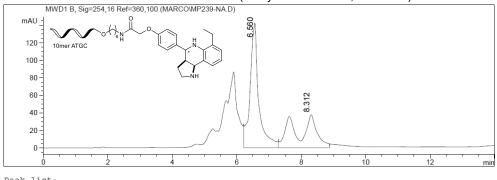
 Ret. Time
 Width min
 Height
 Area
 Area %

 7.441
 0.286
 133.129
 2283.371
 100.000

MALDI-MS spectrum of isolated product 23k



DNA conjugate 23I: CPG-coupled 10mer ATGC-aldehyde conjugate 15a was reacted with 2-ethylaniline 12d and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

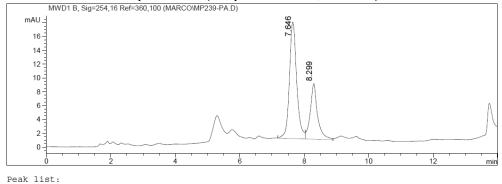


HPLC trace of crude reaction mixture 23I (Analytical RP-HPLC, Method-II)

Peak list:

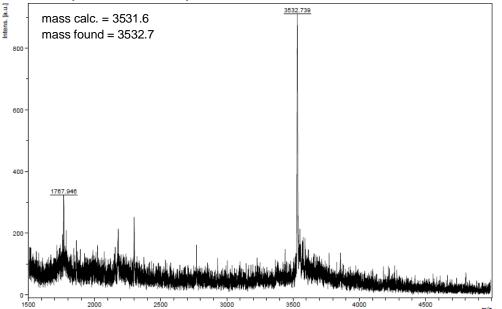
Ret. Time	Width min	Height	Area	Area %
6.560	0.312	142.420	2666.813	61.419
8.312	0.740	37.744	1675.209	38.581

HPLC trace of isolated product 23I (Analytical RP-HPLC, Method-II)

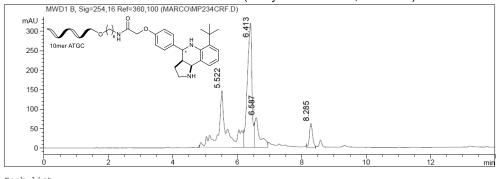


Ret. Time Width min Height Area Area % 7.646 0.233 16.867 263.930 67.652 8.299 0.221 8.049 126.196 32.348

MALDI-MS spectrum of isolated product 23I



DNA conjugate 23m: CPG-coupled 10mer ATGC-aldehyde conjugate 15a was reacted with 2-tert butylaniline 12k and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

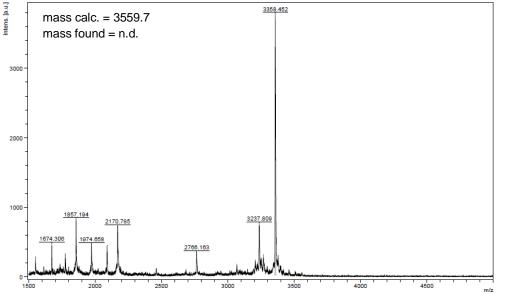


HPLC trace of crude reaction mixture 23m (Analytical RP-HPLC, Method-II)

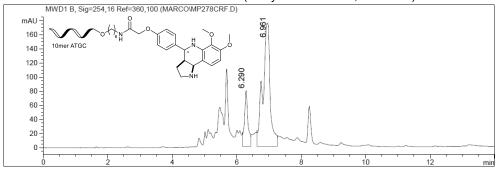
Peak list:

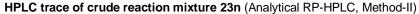
Ret. Time	Width min	Height	Area	Area %
5.522	0.319	145.795	2793.278	38.528
6.413	0.164	320.226	3144.597	43.373
6.587	0.194	76.655	890.530	12.283
8.285	0.113	62.314	421.676	5.816





DNA conjugate 23n: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 2,3-dimethoxyaniline **12m** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

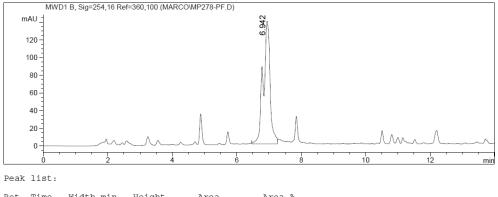




Peak list:

Ret. Time	Width min	Height	Area	Area %
6.290	0.127	79.191	605.753	18.252
6.961	0.259	174.555	2713.016	81.748

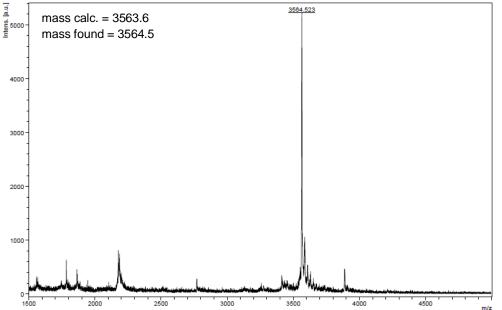
HPLC trace of isolated product 23n (Analytical RP-HPLC, Method-II)



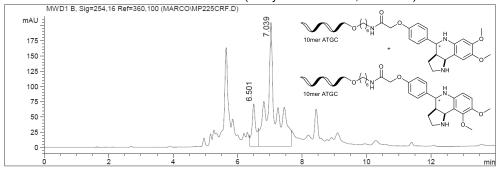
 Ret. Time
 Width min
 Height
 Area
 Area %

 6.942
 0.248
 139.371
 2069.797
 100.000

MALDI-MS spectrum of isolated product 23n



DNA conjugate 230: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 3,4-dimethoxyaniline **12l** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

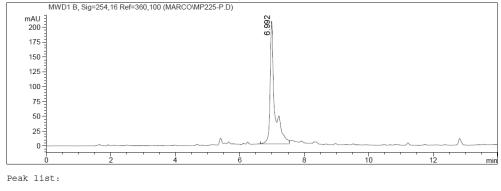


HPLC trace of crude reaction mixture 230 (Analytical RP-HPLC, Method-II)

Peak list:

Ret. Time	Width min	Height	Area	Area %
6.501	0.140	69.709	584.579	14.143
7.039	0.290	203.822	3548.664	85.857

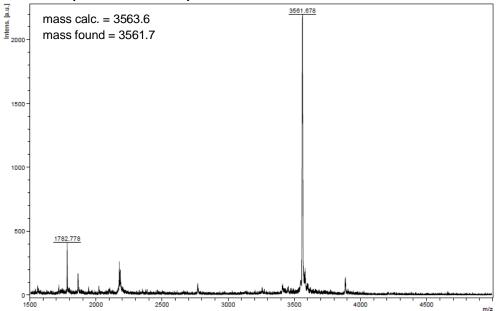
HPLC trace of isolated product 230 (Analytical RP-HPLC, Method-II)



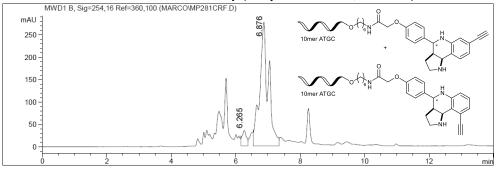
 Ret. Time
 Width min
 Height
 Area
 Area %

 6.992
 0.162
 207.279
 2009.840
 100.000

MALDI-MS spectrum of isolated product 230



DNA conjugate 23p: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 3-ethynylaniline **12o** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

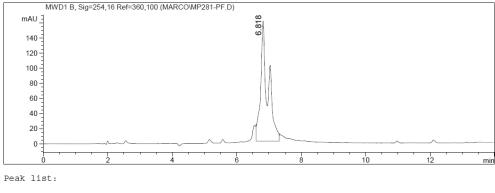


HPLC trace of crude reaction mixture 23p (Analytical RP-HPLC, Method-II)

Peak list:

Ret. Time	Width min	Height	Area	Area %
6.265	0.156	34.292	321.008	5.862
6.876	0.311	276.447	5154.825	94.138

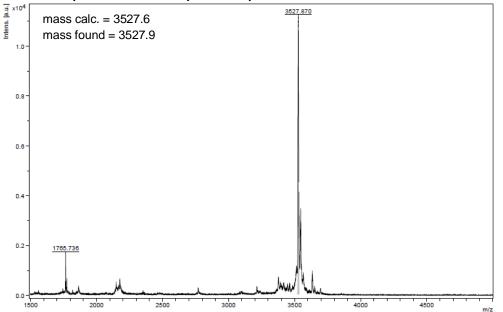
HPLC trace of isolated product 23p (Analytical RP-HPLC, Method-II)



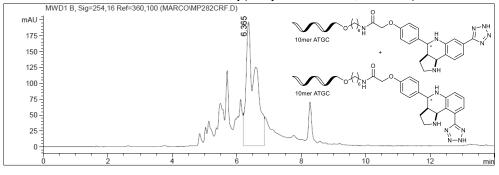
 Ret. Time
 Width min
 Height
 Area
 Area %

 6.818
 0.257
 158.818
 2453.143
 100.000

MALDI-MS spectrum of isolated product 23p



DNA conjugate 23q: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 3-(2H-tetrazol-5-yl)aniline **12p** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.



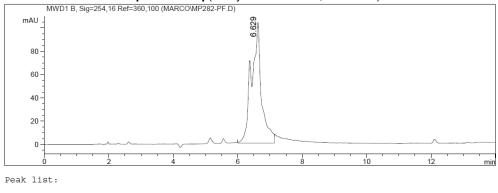
HPLC trace of crude reaction mixture 23q (Analytical RP-HPLC, Method-II)

Peak list:

 Ret. Time
 Width min
 Height
 Area
 Area %

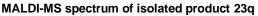
 6.365
 0.340
 194.807
 3973.136
 100.000

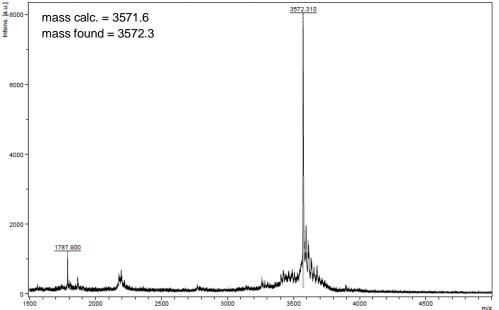
HPLC trace of isolated product 23q (Analytical RP-HPLC, Method-II)



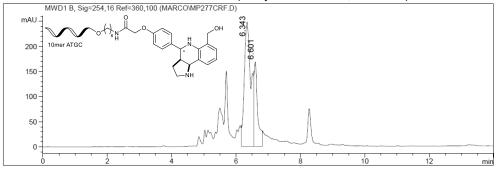
 Ret. Time
 Width min
 Height
 Area
 Area %

 6.629
 0.359
 103.453
 2230.785
 100.000





DNA conjugate 23r: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 2-(hydroxyl-methyl)aniline **12v** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

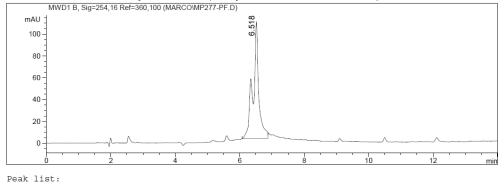


HPLC trace of crude reaction mixture 23r (Analytical RP-HPLC, Method-II)

Peak list:

Ret. Time	Width min	Height	Area	Area %
6.343	0.245	247.679	3641.017	75.375
6.601	0.118	168.657	1189.490	24.625

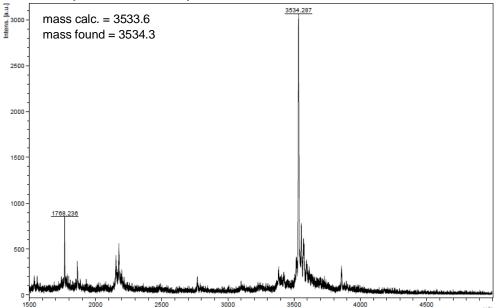
HPLC trace of isolated product 23r (Analytical RP-HPLC, Method-II)



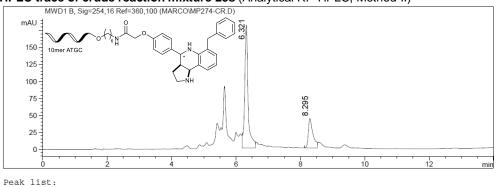
 Ret. Time
 Width min
 Height
 Area
 Area %

 6.518
 0.207
 107.314
 1331.752
 100.000

MALDI-MS spectrum of isolated product 23r



DNA conjugate 23s: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 2-benzylaniline **12w** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

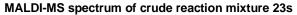


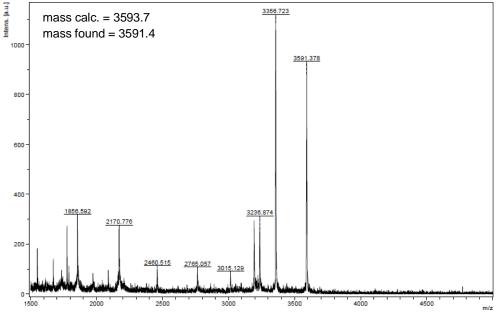
HPLC trace of crude reaction mixture 23s (Analytical RP-HPLC, Method-II)

 Ret. Time
 Width min
 Height
 Area
 Area %

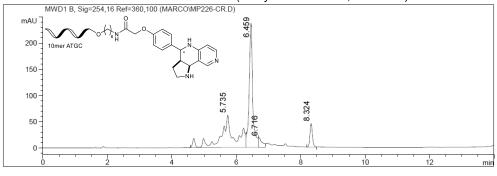
 6.321
 0.121
 181.120
 1319.821
 75.428

 8.295
 0.165
 43.493
 429.965
 24.572





DNA conjugate 23t: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 4-Aminopyridine **12q** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

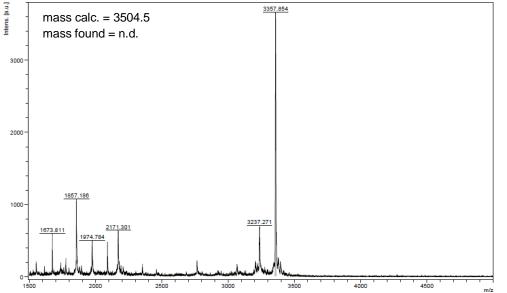


HPLC trace of crude reaction mixture 23t (Analytical RP-HPLC, Method-II)

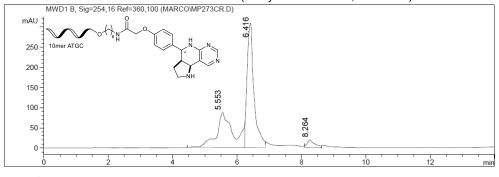
Peak list:

Ret. Time W	Vidth min	Height	Area	Area %
6.459 0 6.716 0	0.127	237.525 17.753	1817.064 141.318	43.046 46.977 3.654 6.324





DNA conjugate 23u: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with 4-Aminopyrimidine **12x** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

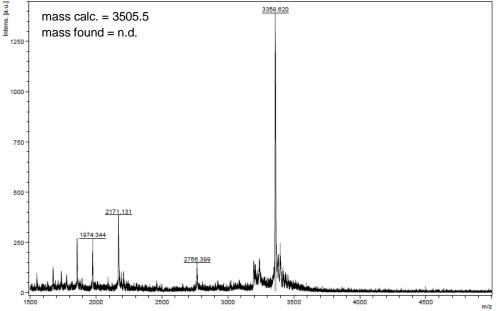


HPLC trace of crude reaction mixture 23u (Analytical RP-HPLC, Method-II)

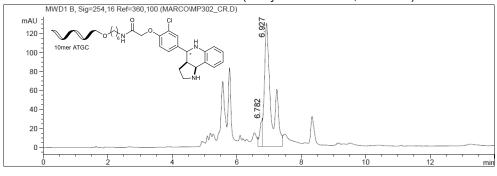
Peak list:

Ret. Time	Width min	Height	Area	Area %
5.553 6.416 8.264	0.585 0.248 0.271	86.666 308.370 19.285	3044.036 4595.420 313.670	38.275 57.781 3.944





DNA conjugate 23v: CPG-coupled 10mer ATGC-aldehyde conjugate **15b** was reacted with aniline **12a** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.

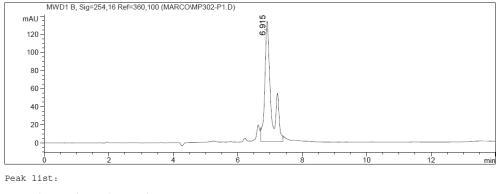


HPLC trace of crude reaction mixture 23v (Analytical RP-HPLC, Method-II)

Peak list:

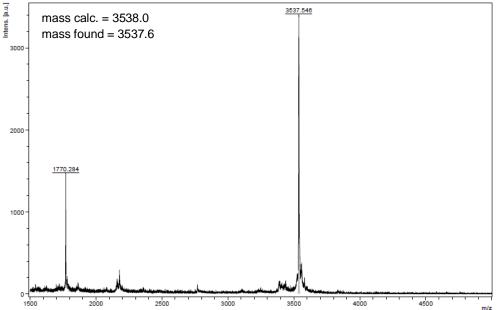
Ret. Time	Width min	Height	Area	Area %
6.782	0.091	27.097	147.754	7.261
6.927	0.242	130.151	1887.230	92.739

HPLC trace of isolated product 23v (Analytical RP-HPLC, Method-II)

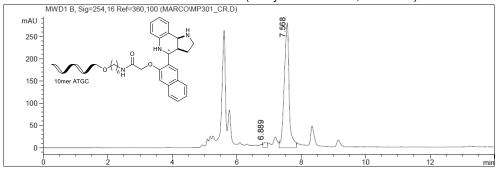


6.915		0.227	 133.464	1821.646	100.000
0.913	5	0.227	133.404	1021.040	100.000

MALDI-MS spectrum of isolated product 23v



DNA conjugate 23w: CPG-coupled 10mer ATGC-aldehyde conjugate **15c** was reacted with aniline **12a** and *N*-Boc-2,3-dihydro-1H-pyrrole **20** according to RP-09.



HPLC trace of crude reaction mixture 23w (Analytical RP-HPLC, Method-II)

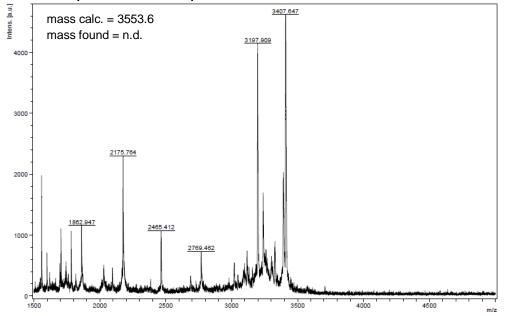
Peak list:

 Ret. Time
 Width min
 Height
 Area
 Area %

 6.889
 0.145
 11.785
 102.258
 3.377

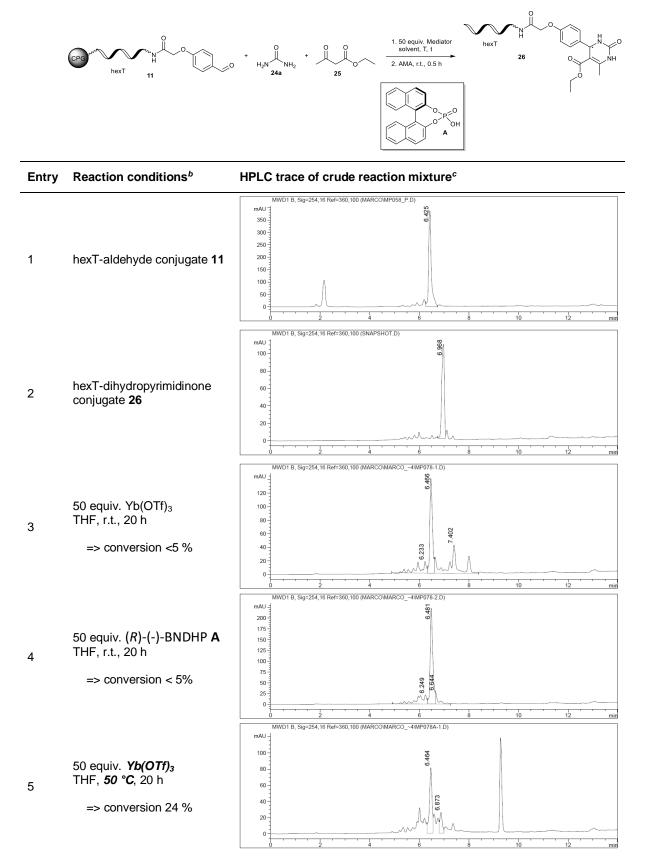
 7.568
 0.174
 280.383
 2925.668
 96.623

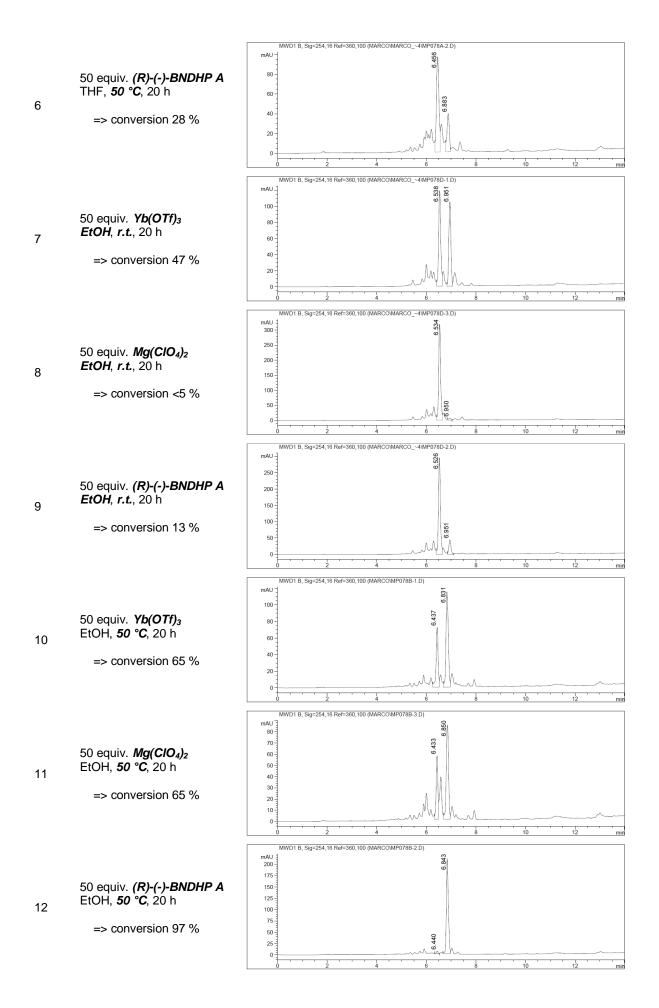
MALDI-MS spectrum of isolated product 23w

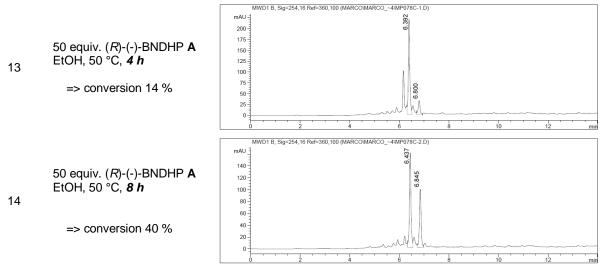


(R)-(-)-BNDHP-mediated Biginelli reaction on CPG-coupled oligonucleotides

Table S11 Optimization of (R)-(-)-BNDHP -mediated Biginelli reaction on CPG-coupled hexT.^a

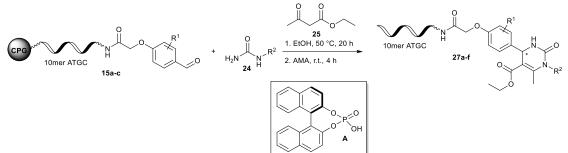






^a CPG-coupled hexT aldehyde conjugate **11** (20 nmol) was suspended in indicated solvent with urea **24a** (500 equiv.) and mediator (50 equiv., 1 µmol) each dissolved or suspended in 30 µL of indicated solvent and ethyl acetoacetate **25** (500 equiv.), reaction mixture was shaken at indicated temperature for 20 h. Afterwards AMA (30% aqueous ammonia / 40% aqueous methylamine, 1:1 (vol/vol)) at ambient temperature for 0.5 h. ^{*b*} parameters that were changed are in bold and italic. ^{*c*} Analytical RP-HPLC, Method-I.

Table S12 – Scope of (R)-(-)-BNDHP **A**-mediated Biginelli reaction on CPG-coupled 10mer ATGC oligonucleotide-aldehyde conjugates **15** with ureas **24** and ethyl acetoacetate **25**.^a

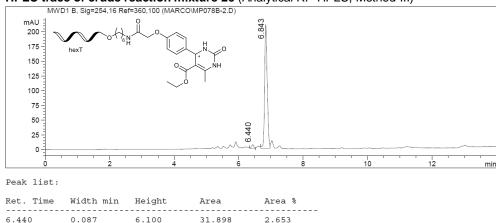


Entry	Product	DNA-aldehyde conjugate	Urea	Conversion [%] ^b	Mass _{calc.} Mass _{found} ^d
1	27a	0 15a	о Н ₂ N ИН ₂ 24а	90	3513.5 3513.3
2	27b	م بر 15a	P _{2N} N H H H H H H H H H H H H H H H H H H	59	3541.5 3540.0
3	27c	م بر 15a	H ₂ N H H 24c	10 (75 ^{c,d})	3589.6 3593.6
4	27d	م بر 15a	H ₂ N H 24d	85	3603.6 3602.9
5	27e		P _{2N} → NH ₂ NH ₂ 24a	92	3547.9 3547.9
6	27f	<u>م</u> ۲5c	о Н ₂ N Н ₂ 24а	n.d.	3563.6 n.d.

^a CPG-coupled ATGC aldehyde conjugate **15** (20 nmol) was suspended with urea **24** (500 equiv.) and (*R*)-(-)-BNDHP **A** (50 equiv., 1 µmol) each dissolved in 30 µL of ethanol and ethyl acetoacetate **25** (500 equiv.), reaction mixture was shaken at 50 °C for 20 h. Afterwards AMA (30% aqueous ammonia / 40% aqueous methylamine, 1:1 (vol/vol)) at ambient temperature for four h. ^b Determined by analytical RP-HPLC analysis. ^c 200 equiv. of (R)-(-)-BNDHP **A** were used. ^d reaction was performed at 50 °C for 44 h. ^d Measured by MALDI-MS. 10mer ATGC = 5'-GTC ATG ATC T-3', n.d. = not detected.

Products of Biginelli reaction on CPG-coupled oligonucleotide-aldehyde conjugate

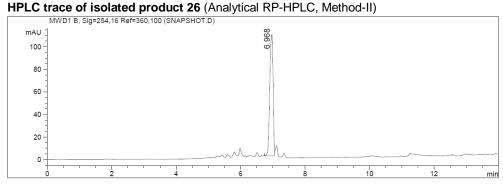
DNA conjugate 26: CPG-coupled 10mer hexT-aldehyde conjugate **11** was reacted with urea **24a** and ethyl acetoacetate **25** according to RP-10.



HPLC trace of crude reaction mixture 26 (Analytical RP-HPLC, Method-III)

210.296

1170.318



97.347

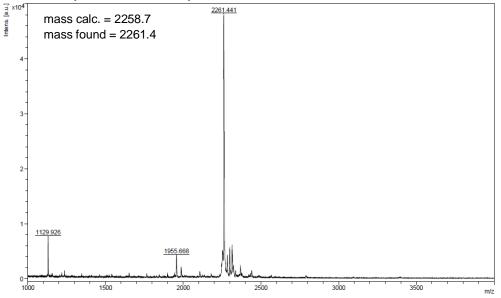
Peak list:

6.843

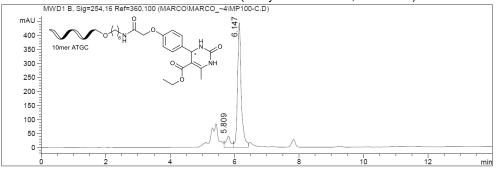
0.093

Ret. Time	Width min	Height	Area	Area %
6.968	0.120	107.631	778.136	100.000

MALDI-MS spectrum of isolated product 26



DNA conjugate 27a: CPG-coupled 10mer ATGC-aldehyde conjugate 15a was reacted with urea 24a and ethyl acetoacetate 25 according to RP-10.

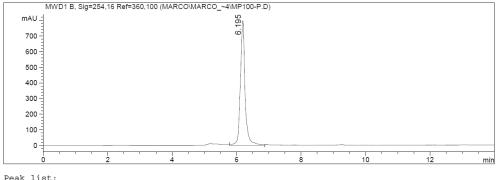


HPLC trace of crude reaction mixture 27a (Analytical RP-HPLC, Method-III)

Peak list:

Ret. Time	Width min	Height	Area	Area %
5.809	0.158	41.502	394.660	9.946
6.147	0.134	445.623	3573.551	90.054

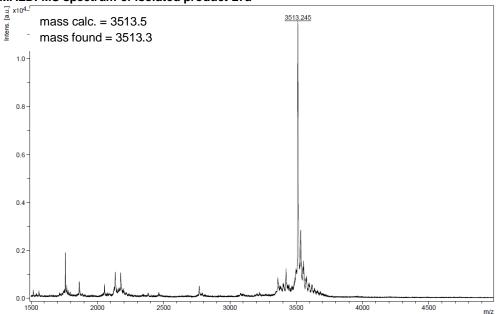
HPLC trace of isolated product 27a (Analytical RP-HPLC, Method-III)



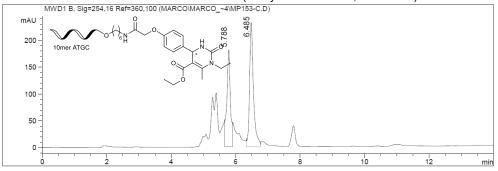
Peak list:

Ret. Time	Width min	Height	Area	Area %
6.195	0.144	796.682	8070.688	100.000

MALDI-MS spectrum of isolated product 27a



DNA conjugate 27b: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with *N*-ethylurea **24b** and ethyl acetoacetate **25** according to RP-10.

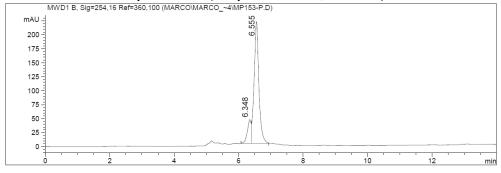


HPLC trace of crude reaction mixture 27b (Analytical RP-HPLC, Method-III)

Peak list:

Ret. Time	Width min	Height	Area	Area %
5.788	0.127	181.070	1382.508	41.493
6.485	0.140	232.049	1949.426	58.507

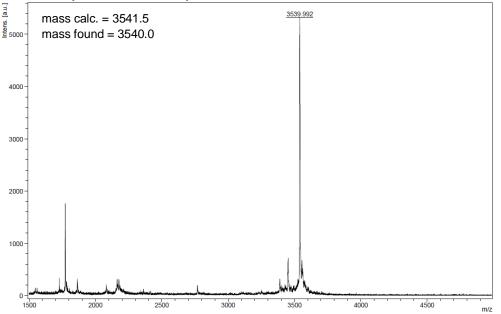
HPLC trace of isolated product 27b (Analytical RP-HPLC, Method-III)



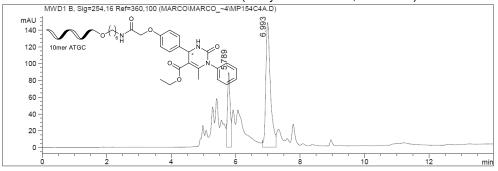
Peak list:

Ret. Time	Width min	Height	Area	Area %
6.348	0.135	42.897 218.316	346.906	13.444
6.555				

MALDI-MS spectrum of isolated product 27b



DNA conjugate 27c: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with *N*-phenylurea **24c** and ethyl acetoacetate **25** according to RP-10.

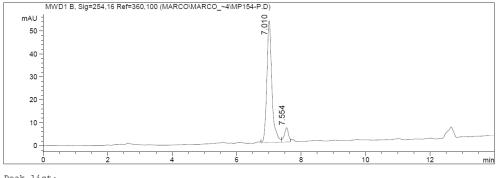


HPLC trace of crude reaction mixture 27c (Analytical RP-HPLC, Method-III)

Peak list:

Ret. Time	Width min	Height	Area	Area %
5.789	0.095	89.395	508.282	25.470
6.993	0.167	148.519	1487.302	74.530

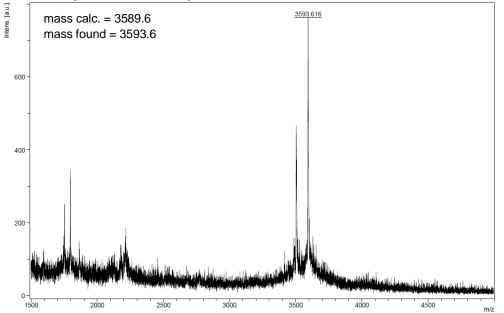
HPLC trace of isolated product 27c (Analytical RP-HPLC, Method-III)



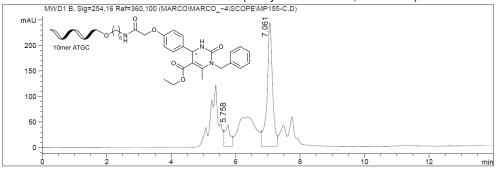
Peak list:

Ret. Time	Width min	Height	Area	Area %
7.010 7.554	0.159 0.133	53.187 6.209	577.659 56.827	91.044 8.956

MALDI-MS spectrum of isolated product 27c



DNA conjugate 27d: CPG-coupled 10mer ATGC-aldehyde conjugate **15a** was reacted with *N*-benzylurea **24d** and ethyl acetoacetate **25** according to RP-10.

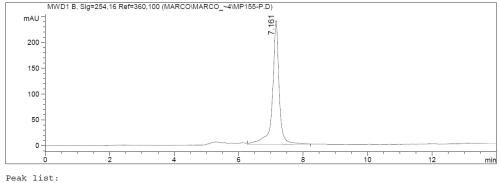


HPLC trace of crude reaction mixture 27d (Analytical RP-HPLC, Method-III)

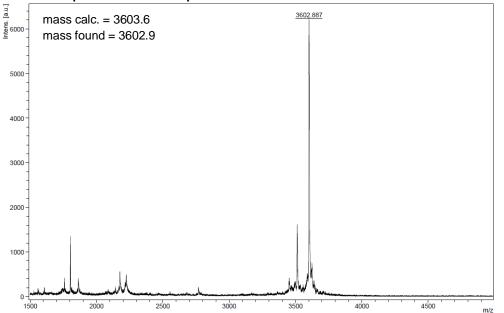
Peak list:

Ret. Time	Width min	Height	Area	Area %
5.758	0.200	42.105	504.256	15.058
7.061	0.195	242.740	2844.499	84.942

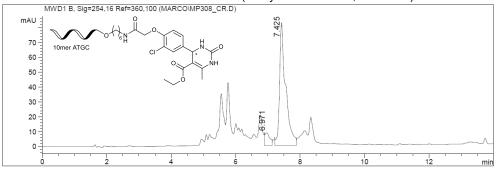
HPLC trace of isolated product 27d (Analytical RP-HPLC, Method-III)



MALDI-MS spectrum of isolated product 27d



DNA conjugate 27e: CPG-coupled 10mer ATGC-aldehyde conjugate **15b** was reacted with urea **24a** and ethyl acetoacetate **25** according to RP-10.

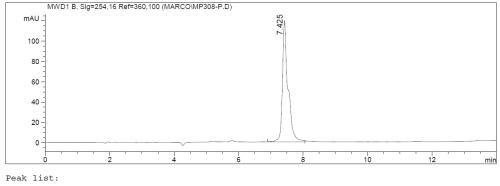


HPLC trace of crude reaction mixture 27e (Analytical RP-HPLC, Method-II)

Peak list:

Ret. Time	Width min	Height	Area	Area %
6.971	0.178	8.538	91.303	7.685
7.425	0.221	82.818	1096.778	92.315

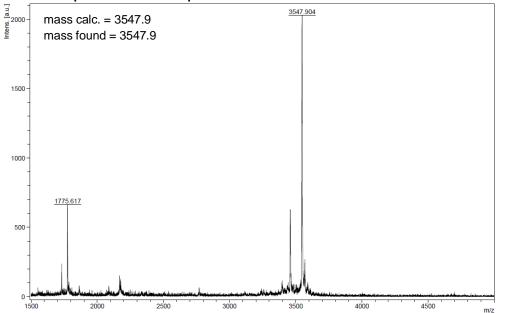
HPLC trace of isolated product 27e (Analytical RP-HPLC, Method-II)



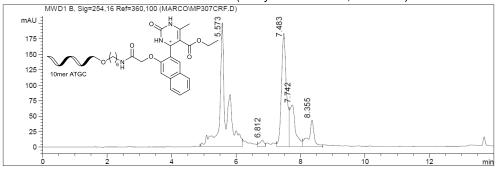
 Ret. Time
 Width min
 Height
 Area
 Area %

 7.425
 0.199
 119.402
 1425.893
 100.000

MALDI-MS spectrum of isolated product 27e



DNA conjugate 27f: CPG-coupled 10mer ATGC-aldehyde conjugate **15c** was reacted with urea **24a** and ethyl acetoacetate **25** according to RP-10.

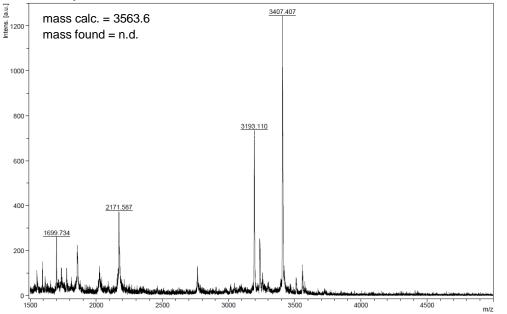


HPLC trace of crude reaction mixture 27f (Analytical RP-HPLC, Method-II)

Peak list:

Ret. Time	Width min	Height	Area	Area %
5.573	0.235	201.018	2834.301	45.431
6.812	0.165	10.104	100.238	1.607
7.483	0.179	183.128	1969.024	31.561
7.742	0.208	66.999	834.551	13.377
8.355	0.197	42.426	500.637	8.025

MALDI-MS spectrum of crude reaction mixture 27f



Cheminformatic analysis

For library design, characterization and analysis the software KNIME¹ was used, with the following implemented extensions: RDKit, Indigo, ChemAxon, CDK, Vernalis.²

As a source database for the components required for the reactions, the Aldrich Market Select database (AMS)³ was used and split in order to retrieve the different compound classes according to functional groups. Prior to the library design, the molecules in the AMS database were firstly standardized and filtered by molecular weight and number of unspecified stereocenters by RDKit Descriptor Calculation: the threshold was set to 200 Da for each compound class to ensure a reasonable size of the final product. In order to retrieve the different building block classes from the AMS, the RDKit Functional Group Filter or the Indigo node Substructure Match Counter combined with the Rule-based Row splitter were used, defining the substructure or the further filters by the ChemAxon Marvin Sketch node. We removed all aldehyde building blocks that contain the carboxylic acid moiety and/or bulky groups in ortho position to the carboxaldehyde and kept this compound class constant for the DA-1, P and B libraries. Further specific filters were applied to each compound class per each library depending on the experimental reactivity or on the presence of further functional groups which might lead to cross-reactivity. For the DA-1 library we removed heteroaromatic amines and anilines substituted with bulky ortho-substitution. For the DA-2 library, anilines with carboxylic acid amide or bulky groups in ortho-position were excluded, yet, all available aldehydes were included in the analysis. For the Povarov library the heteroaromatic amines were filtered out. For the Biginelli library, differently substituted ureas and βketoesters were used, but, in order to obtain libraries of comparable compound numbers, a further molecular weight filter was applied to starting materials (170 Da for ureas and esters) and products (450 Da).

All the building block classes were submitted to the RDKit Catalog filter node in order to remove compounds with unwanted reactivity.⁴ For computational reasons and in order to obtain libraries of comparable sizes, larger building block datasets were sampled to 1000 molecules with the RDKit diversity picker (aldehydes and anilines). The reactions required for the *in silico* libraries were designed using a combination of RDKit node Two Component Reaction or Chemical Transformation, the Chemaxon Marvin Sketch for the reaction scheme and the SDF Files Reader for the different components. From the Enamine REAL database⁵, 500000 molecules were randomly sampled to obtain comparable data sets. We calculated the MQN descriptors for each compound of each library with the node RDKit Descriptors Calculation and, after Min-max normalization among the concatenated data of the libraries and the commercially available molecules, the PCA values were plotted in 3D scatter plots (Figure S3) using the Gnuplot⁶ script, to visualize chemical diversity.

The PMIs were calculated with the corresponding Vernalis node, based on the 3D coordinates generated by RDKit Generate Coords node, to evaluate library shape diversity (Figure S4).

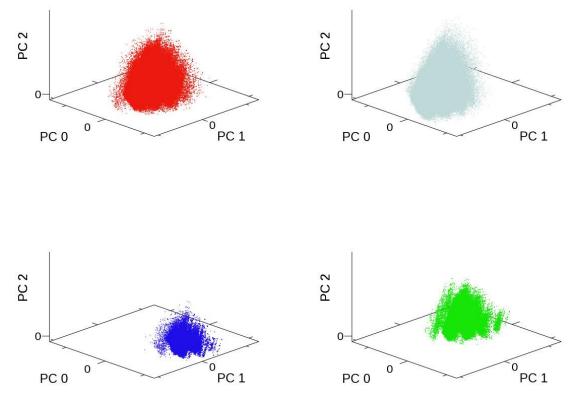


Figure S3 PCA plots of the DA-1 (red), Enamine REAL sampled database (grey), P (blue) and B (green) libraries.

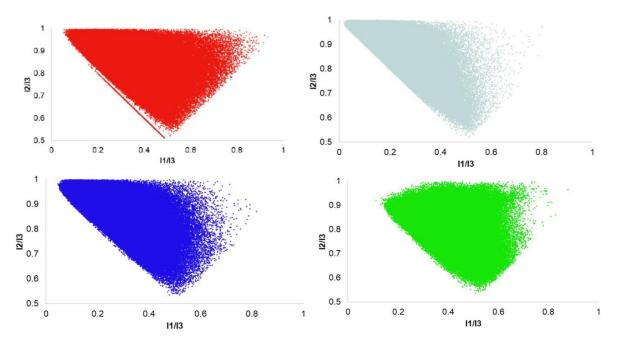


Figure S4 PMI plots of the DA-1 (red), Enamine REAL sampled database (grey), P (blue) and B (green) libraries.

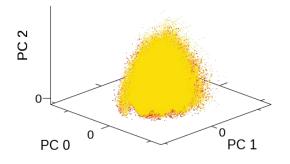


Figure S5 PCA plots of the DA-1 (red) and DA-2 (yellow) libraries, covering almost identical chemical space.

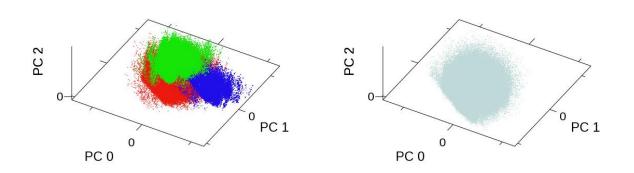


Figure S6 PCA plots of the combined DA-1 (red), P (blue) and B (green) libraries vs Enamine REAL sampled database (grey), viewed from a different perspective.

References:

- 1. <u>https://www.knime.com/downloads</u>
- M. P. Mazanetz, R.J. Marmon, C. B.T. Reisser, I. Morao, Curr. *Top. Med. Chem.*, 2012, **12**, 1965-1979.
- 3. https://www.sigmaaldrich.com/chemistry/chemistry-services/aldrich-market-select.html
- 4. J. B. Baell, J. W. M. Nissink, ACS Chem Biol. 2018, 13, 36-44.
- 5. https://enamine.net/library-synthesis/real-compounds/real-compound-libraries
- 6. <u>http://gnuplot.info</u>