

## **A complete picture of protein unfolding and refolding in surfactants**

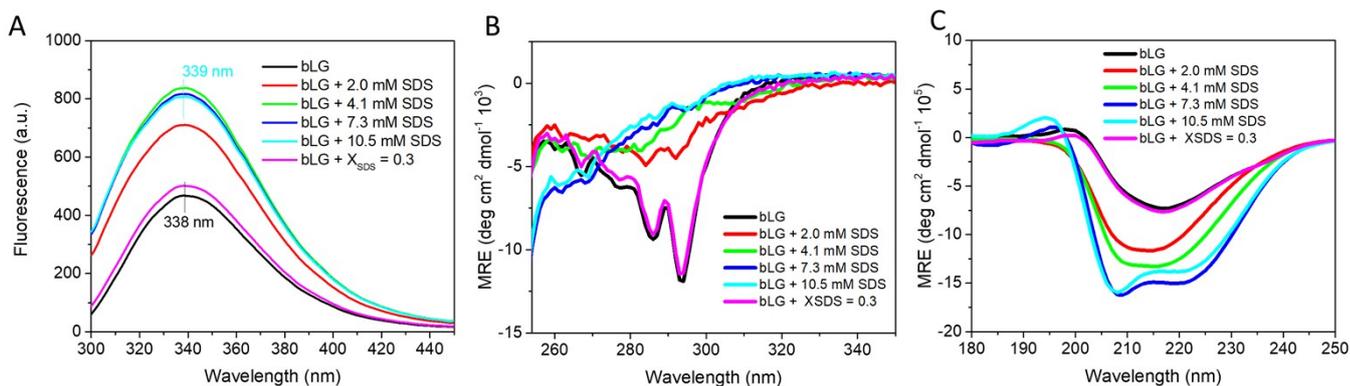
Jannik Nedergaard Pedersen<sup>1</sup>, Jeppe Lyngsø<sup>1</sup>, Thomas Zinn<sup>2</sup>, Daniel Erik Otzen<sup>3</sup> and Jan Skov Pedersen<sup>1</sup>

<sup>1</sup>Interdisciplinary Nanoscience Center (iNANO) and Department of Chemistry, Aarhus University, Gustav Wieds Vej 14, 8000 Aarhus C, Denmark

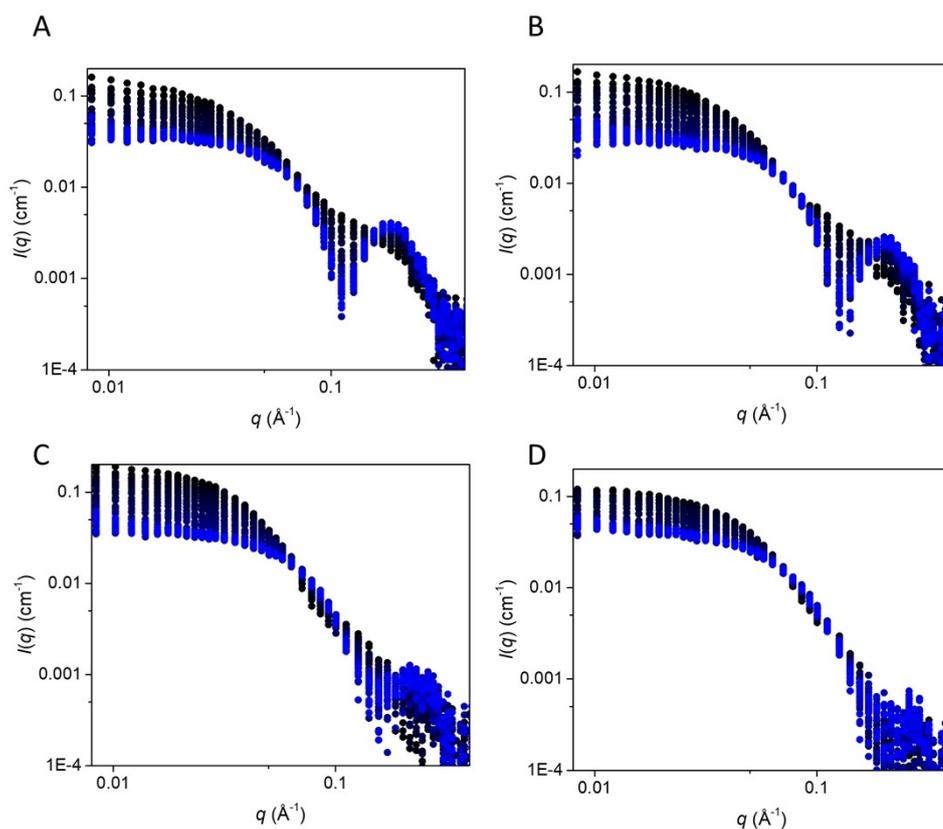
<sup>2</sup>ESRF-The European Synchrotron, 38043 Grenoble, France

<sup>3</sup>Interdisciplinary Nanoscience Center (iNANO), Department of Molecular Biology and Genetics, Aarhus University, Gustav Wieds Vej 14, DK – 8000 Aarhus C, Denmark

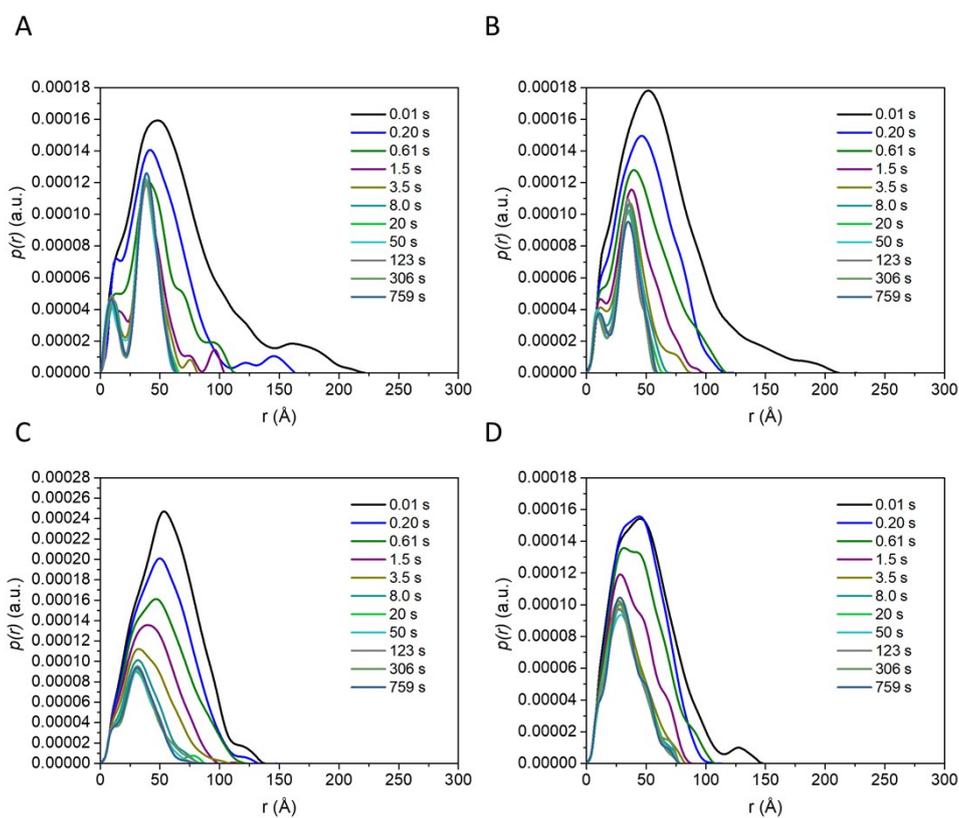
### **Supporting Information**



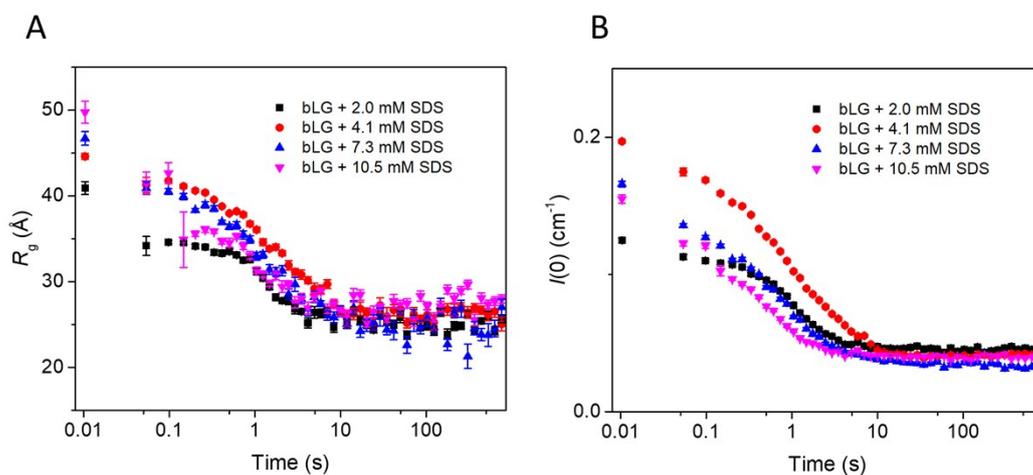
**Fig. S1.** (A) Tryptophan fluorescence measurements done with excitation at 280 nm of  $\beta$ -lactoglobulin (2 mg/ml) with SDS (8.5 mM), or SDS and  $C_{12}E_8$  (19.8 mM).  $\lambda_{max}$  values are shown. (B) Equilibrium near-UV CD of bLG with SDS and  $C_{12}E_8$ . (C) Equilibrium far-UV CD of bLG with SDS and  $C_{12}E_8$ .



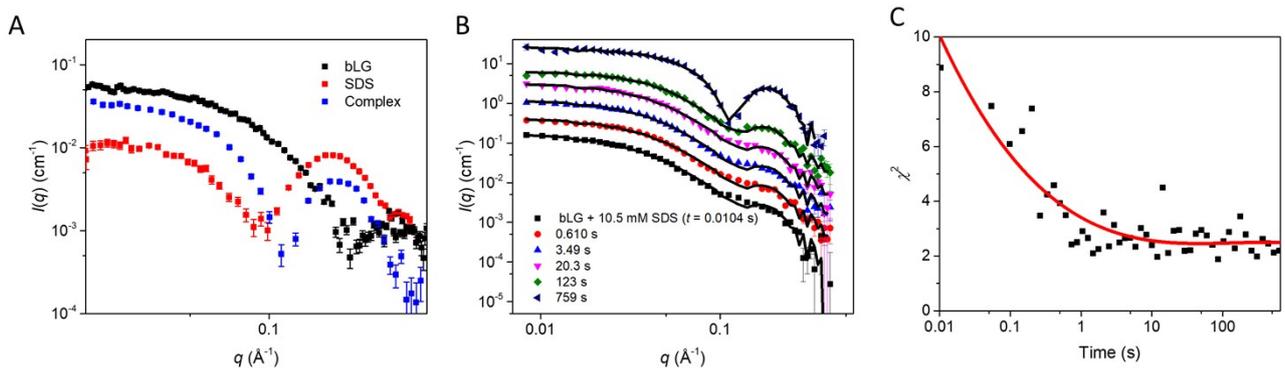
**Fig. S2.** Stopped-flow SAXS data sets of bLG mixed with (A) 10.5 mM SDS, (B) 7.3 mM SDS, (C) 4.1 mM SDS, and (D) 2.0 mM SDS with logarithm spaced times from  $\sim 4$  ms to 12.5 min.



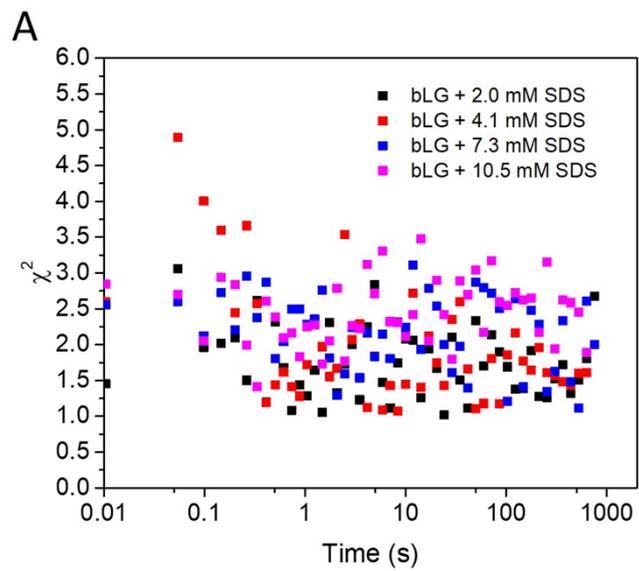
**Fig. S3.**  $p(r)$  functions at selected times from the stopped-flow SAXS data of bLG mixed with (A) 10.5 mM SDS, (B) 7.3 mM SDS, (C) 4.1 mM SDS, and (D) 2.0 mM SDS. The maximum diameter decreases with time.



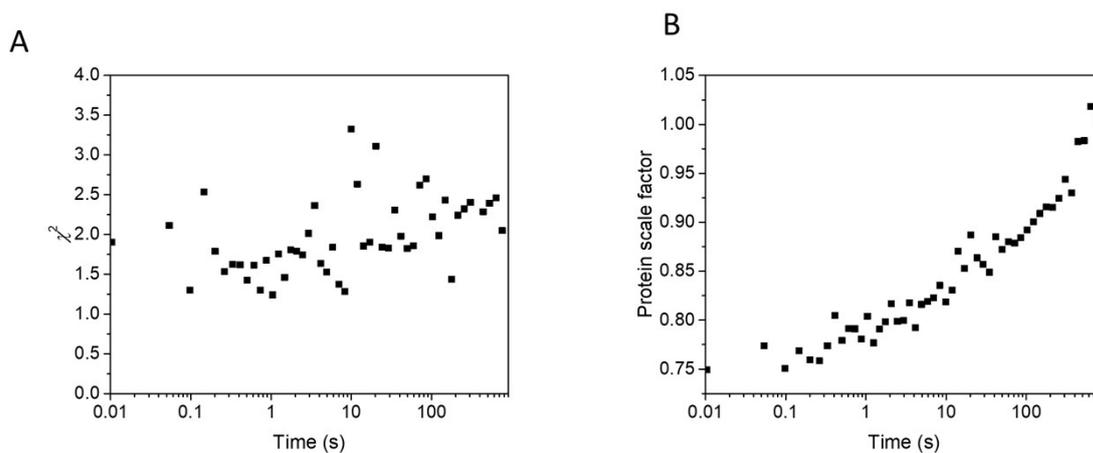
**Fig. S4.** (A) Radius of gyration  $R_g$  and (B) forward scattering  $I(0)$  as determined from Guinier fits of the stopped-flow SAXS data of bLG mixed with different concentrations of SDS.



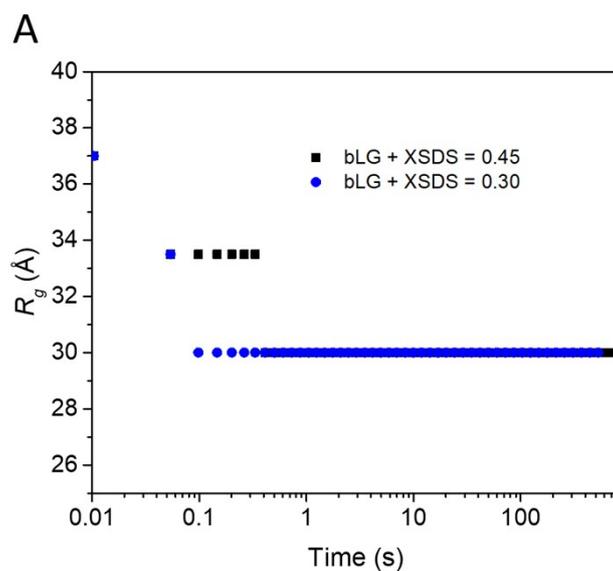
**Fig. S5.** (A) SAXS data of bLG (4 mg/ml), SDS (25 mM), and bLG-SDS complex (2 mg/ml bLG and 10.5 mM SDS) used for fitting unfolding data with a linear combination. (B) Selected frames from SAXS measurements of refolding kinetics of bLG (2 mg/ml) in SDS (10.5 mM). Lines represent fits using a linear combination of data seen in (A). For clarity, the data has been scaled with a factor of 3 for every time step. (C)  $\chi^2$  from fitting kinetic data in (B) to a linear combination of data in (A).



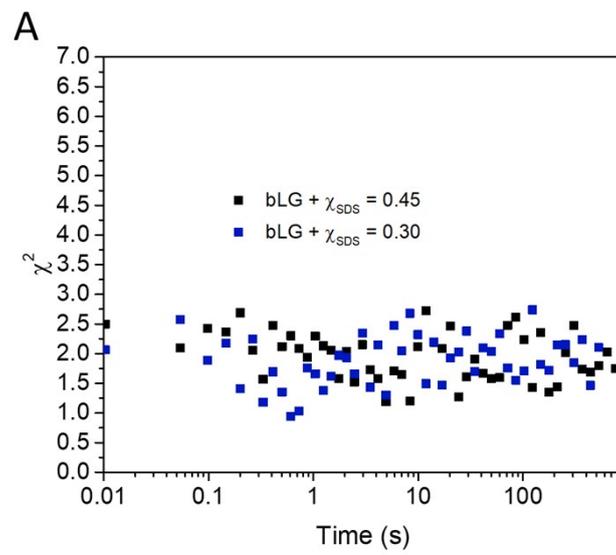
**Fig. S6.** (A)  $\chi^2$  values for fits to the core-shell model with an adjustable core offset for the stopped-flow unfolding SAXS data (Fig. 4).



**Fig. S7.** (A)  $\chi^2$  from fitting SAXS data of refolding in  $\chi_{SDS} = 0.45$  to a linear combination of (1) the unfolded bLG-SDS complex, (2) folded bLG (77% dimer), and (3) mixed SDS- $C_{12}E_8$  micelles. (B) The sum of scale factors from the bLG-SDS complex and folded bLG as a function of time when a linear combination has been fitted to SAXS data of refolding in  $\chi_{SDS} = 0.45$ .



**Fig. S8.** Radius of gyration ( $R_g$ ) of the Gaussian random chain used to fit data in Fig 7.



**Fig. S9.** (A)  $\chi^2$  values for fits to the data in Fig. 7A and 7B using a linear combination of native protein, random coil protein, bLG-SDS complex, and mixed SDS- $\text{C}_{12}\text{E}_8$  micelles, for fitting.

**Table S1.** Electron densities,  $\rho$ , excess electron densities,  $\Delta\rho$ , and partial volumes,  $v$ , used in the model fitting. The scattering length densities are obtained by multiplying by the classical Thomson radius,  $0.282 \times 10^{-12}$  cm. For the protein, the scattering length per mass,  $\Delta\rho_m$ , and the partial specific volume,  $v_p$ , are given.

	$\rho$ (e/Å <sup>3</sup> )	$\Delta\rho$ (e/Å <sup>3</sup> )	$v$ (Å <sup>3</sup> )
Water	0.334		29.92
C12	0.275	-0.0592	353.0
SDS headgroup	0.975	0.551 <sup>i</sup>	60.5
EO <sub>8</sub> headgroup	0.392	0.0579	61.2 x 7.5=459 <sup>ii</sup>
	$\Delta\rho_m$ (cm/g)	$v_p$ (cm <sup>3</sup> /g)	
Protein	$2.0 \times 10^{10}$	0.720	

<sup>i</sup> Scaled by 0.86 in order to fit the SAXS data for pure SDS micelles.

<sup>ii</sup> The number of EO units was used as a fit parameter and it was determined to be  $7.5 \pm 0.3$ .