

Supporting Information

An Umpolung Approach to the Hydroboration of Pyridines: A Novel and Efficient Synthesis of *N*-H 1,4-Dihydropyridines

Huan Yang, Li Zhang, Fei-Yu Zhou, and Lei Jiao*

*Center of Basic Molecular Science (CBMS), Department of Chemistry, Tsinghua University,
Beijing 100084, China*

Leijiao@mail.tsinghua.edu.cn

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1. General Information

Experimental. Air- and moisture-sensitive reactions were carried out in oven-dried Schlenk flasks sealed with rubber septa under a positive pressure of dry argon, or oven-dried glass tubes sealed with Teflon screw caps. Air- and moisture-sensitive liquids and solutions were transferred by a syringe or a double-headed needle. Reactions were stirred using Teflon-coated magnetic stir bars. Elevated temperatures were maintained using Thermostat-controlled silicone oil baths. Organic solutions were concentrated using a rotary evaporator with a diaphragm vacuum pump or concentrated under vacuum using standard Schlenk techniques. Analytical TLC was performed on silica gel GF₂₅₄ plates. The TLC plates were visualized by either ultraviolet light ($\lambda = 254$ nm). Purification of 1,4-dihydropyridine products was done by flash column chromatography on basic alumina (80-160 mesh, pH = 9-10) under argon. Purification of other products was accomplished by flash column chromatography on silica gel (Innochem SilicaFlashP60, 230-400 mesh).

Chemicals. Anhydrous solvents ($\text{H}_2\text{O} < 30$ ppm), ethylene dimethyl ether, and methanol were purchased from J&K Scientific. Tetrahydrofuran and acetonitrile were purified by PureSolv MD-5 solvent purification system, and were deoxygenated by three freeze-pump-thaw cycles before use. Methanol-*d*₁ (MeOD) was purchased from J&K Scientific and was dried over 4Å molecular sieves and deoxygenated by three freeze-pump-thaw cycles before use. B₂pin₂, B₂neo₂, and pyridine derivatives were purchased from TCI and used without further purification. MeOK and MeONa were purchased from Sigma-Aldrich and dried under vacuum before use. Cs₂CO₃ was purchased from J&K Scientific and dried under vacuum before use. 18-crown-6 was purified by recrystallization before use. Other chemicals were purchased from various commercial sources and were used as received. B₂eg₂ was synthesized following the published procedure.

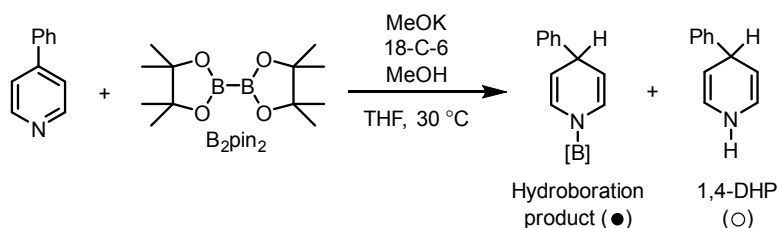
Analytical. NMR spectra were recorded on a Bruker AVANCE III HD 400 (¹H at 400 MHz, ¹³C at 100 MHz) nuclear magnetic resonance spectrometer. The ¹H NMR spectra were calibrated against the peak of tetramethylsilane (TMS, 0 ppm) and the ¹³C NMR spectra were calibrated against the peak of CDCl₃ (77.16 ppm). GC-MS analysis was performed on a Shimadzu GCMS-QO2010SE instrument equipped with an EIS detector using helium as the carrier gas (EI ionization, 70 eV). Enantiomeric excess (e.e.) were determined by analytical high-performance liquid chromatography (HPLC) analysis using a Daicel chiral column Chirapak IC (4.6 × 250 mm). The X-ray single crystal structure analysis was performed on a SuperNova X-ray Diffraction System equipped with Atlas CCD detector and 4-circle kappa goniometer. Structure solution and refinement were accomplished with OLEX2.

2. Study on the Inversed Hydroboration of 4-Phenylpyridine

2.1 General

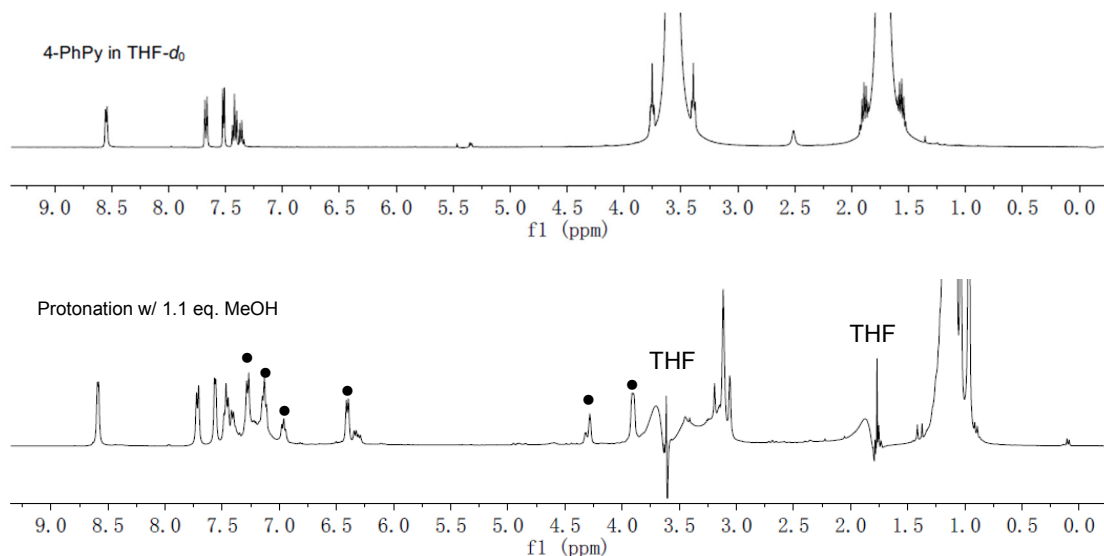
The reactions were conducted in normal THF solvent and were monitored by ^1H NMR of the crude reaction solution. The NMR spectra were recorded after ^1H gradient shimming without applying the ^2H -lock, and the undesired solvent signals were suppressed by the WET solvent suppression method (No-D NMR). The chemical shifts of the solvent utilized in the NMR experiments were calibrated against TMS (1.77 and 3.61 ppm for THF and 1.93 ppm for MeCN), and for other NMR spectra, the chemical shifts was calibrated against the solvent peaks. All measurements were performed under argon atmosphere.

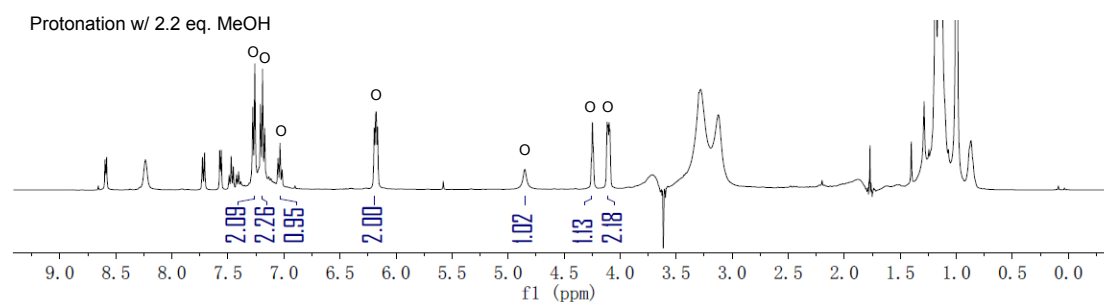
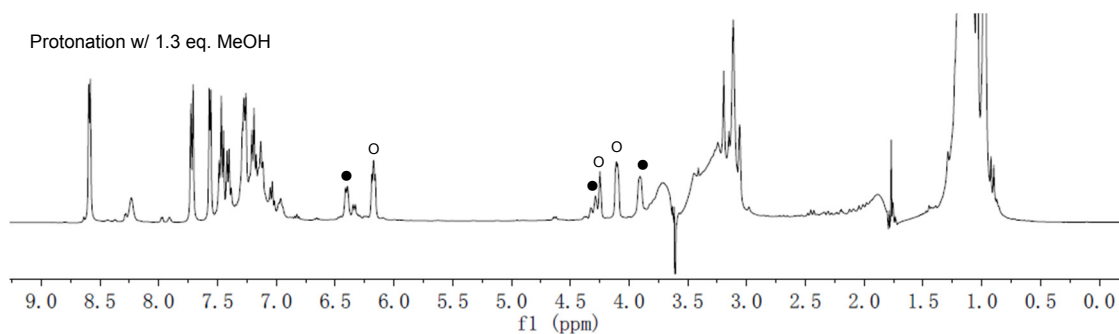
2.2 The reaction between 4-PhPy, B_2pin_2 , and MeOK in the presence of MeOH



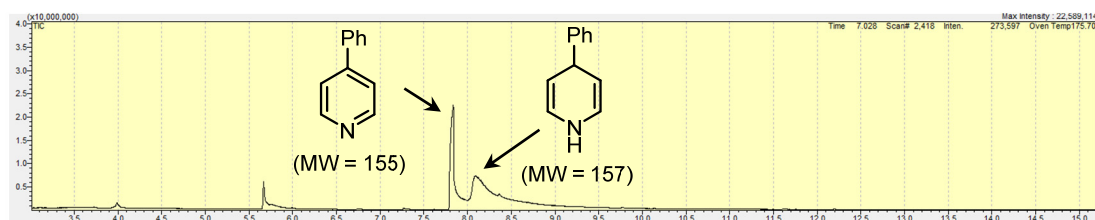
In a glove box, a 15 mL oven-dried reaction tube was charged with 4-PhPy (56.5 mg, 0.36 mmol), B_2pin_2 (102 mg, 0.40 mmol), MeOK (28.0 mg, 0.40 mmol), 18-crown-6 (106 mg, 0.40 mmol), and anhydrous THF (2 mL). An indicated amount of MeOH (0.4 mmol for 1.1 equiv., 0.48 mmol for 1.3 equiv., and 0.8 mmol for 2.2 equiv.) was added by a microsyringe. The resulting mixture was allowed to react at 30 °C for 12 h before submission to ^1H NMR analysis. The reaction mixture with 2.2 equiv. of MeOH was determined by GC-MS analysis.

^1H NMR spectra of the crude reaction mixture are shown below, which were used to construct Figure 1 in the main text:

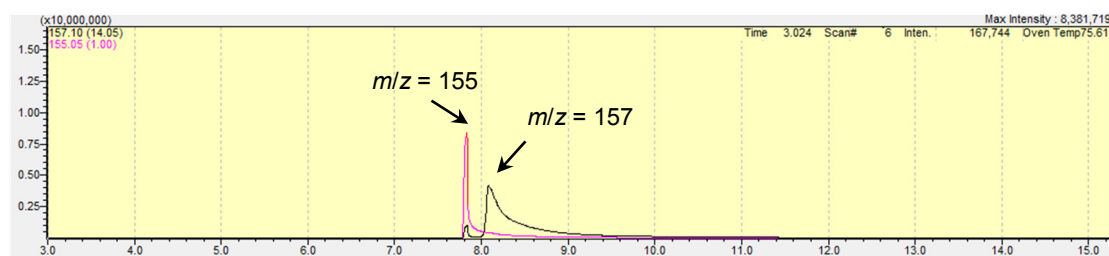




GC-MS trace of the crude reaction mixture with 2.2 equiv. of MeOH is shown below. Conditions for GC-MS analysis: helium as the carrier gas; capillary column (0.25 mm × 30 m, SH-Rxi-5Sil MS); constant pressure mode (150.2 kPa); oven temperature program: 50 °C for 2 min, elevation rate 25 °C/min to 200 °C, keep for 6 min, elevation rate 30 °C/min to 260 °C, keep for 4 min; ion source temperature 200 °C, interface temperature 250 °C, acquisition mode: scan.

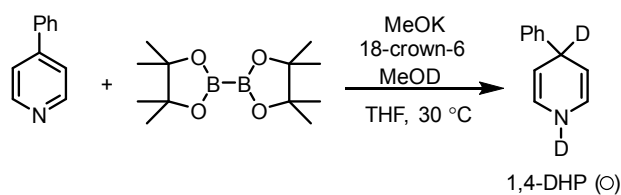


The assignment of the above peaks was confirmed by GC-MS SIM (selected ions monitoring) mode to detect ions at $m/z = 155$ and 157 .



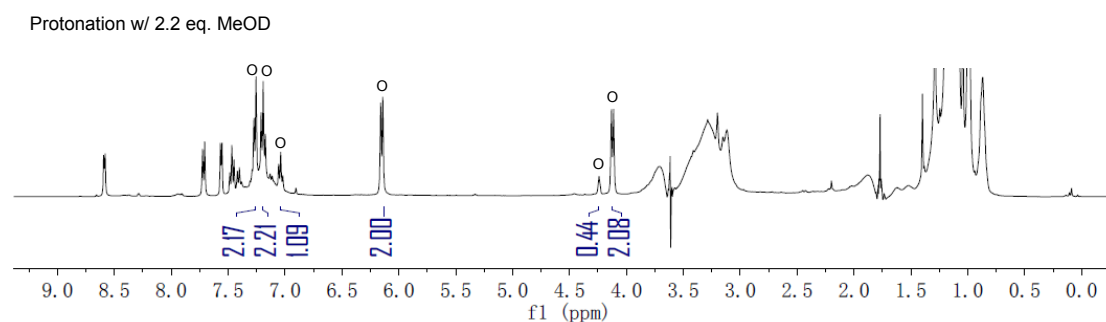
GC-MS results confirmed the formation of 1,4-DHP (MW = 157) in the reaction mixture.

2.3 The reaction between 4-PhPy, B₂pin₂, and MeOK in the presence of MeOD

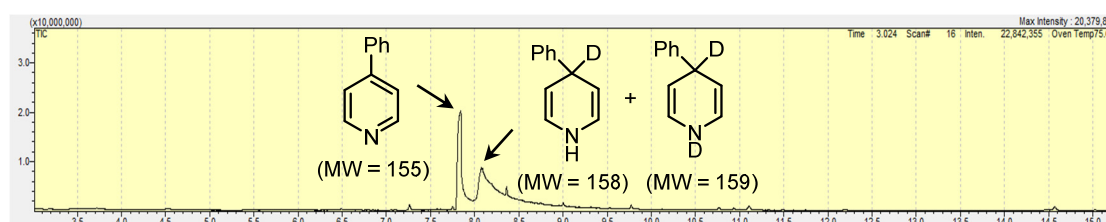


In a glove box, a 15 mL oven-dried sealed tube was charged with 4-PhPy (56.5 mg, 0.36 mmol), B₂pin₂ (102 mg, 0.40 mmol), MeOK (28.0 mg, 0.40 mmol), 18-crown-6 (106 mg, 0.40 mmol), and anhydrous THF (2 mL). Then MeOD (methanol-*d*, 26.4 mg, 0.8 mmol) was added by a microsyringe. The resulting mixture was allowed to react at 30 °C for 12 h before submission to ¹H NMR and GC-MS analysis.

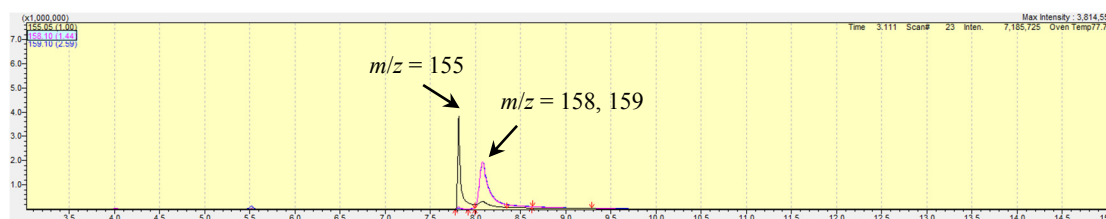
¹H NMR spectrum of the crude reaction mixture is shown below, in which the decrease of the peak at 4.25 ppm indicates deuterium incorporation at the C4-position of 1,4-DHP:



GC-MS trace of the crude reaction mixture is shown below (with identical GC-MS conditions specified in Section 2.2):



The assignment of the above peaks were confirmed by GC-MS SIM (selected ions monitoring) mode to detect ions at *m/z* = 155, 158, and 159.



Both NMR and GC-MS analyses confirmed the formation of deuterated 1,4-DHP in the presence of MeOD.

3. DFT Computational Study

3.1 General

All calculations were performed with the Gaussian 09 program.^[1] Density functional theory calculations using the M06-2X functional^[2] were used to locate the stationary points involved. The gas-phase structures were optimized at the M06-2X/6-31+G(d) level. NBO analysis and NAO orbital composition analysis were performed on the same level of theory.

3.2 NBO analysis on Int-A•K and TS-1•K

Cartesian coordinates of **Int-A•K**:

Center Number	Atomic Number	Atomic Type	Coordinates (Angstroms)		
			X	Y	Z
1	8	0	2.895648	-1.001803	0.534748
2	5	0	1.394261	-0.928076	0.354097
3	8	0	1.113226	-1.901059	-0.706515
4	5	0	0.875115	0.701214	-0.031291
5	8	0	1.165428	1.795334	0.948356
6	8	0	1.290677	1.226822	-1.318489
7	6	0	2.234303	-2.758618	-0.847860
8	6	0	3.432089	-1.815573	-0.496076
9	6	0	1.432068	2.632578	-1.227473
10	6	0	1.863134	2.835735	0.262331
11	6	0	2.262002	-3.299831	-2.272617
12	6	0	2.474646	3.077900	-2.247877
13	6	0	1.475843	4.184720	0.856451
14	6	0	0.096586	3.321738	-1.542573
15	6	0	3.365586	2.596300	0.452674
16	6	0	2.113435	-3.925134	0.141937
17	6	0	3.819443	-0.919947	-1.677732
18	6	0	4.667613	-2.540419	0.028430
19	7	0	-0.807564	0.615301	-0.071429
20	6	0	-1.591211	1.138346	0.877744
21	6	0	-2.957993	0.905787	0.909827
22	6	0	-3.546170	0.104534	-0.075559
23	6	0	-2.704758	-0.426592	-1.061014
24	6	0	-1.346109	-0.157655	-1.026431
25	6	0	-5.002359	-0.171275	-0.076095
26	6	0	-5.686071	-0.394368	1.125316
27	6	0	-7.054117	-0.653285	1.123474
28	6	0	-7.758130	-0.688648	-0.079338
29	6	0	-7.086037	-0.466491	-1.280561
30	6	0	-5.717268	-0.212304	-1.279388

31	8	0	0.843264	-1.381511	1.716072
32	6	0	-0.375006	-2.068640	1.744925
33	1	0	3.182682	-3.865047	-2.465393
34	1	0	1.410765	-3.972343	-2.425899
35	1	0	2.188317	-2.485246	-2.997554
36	1	0	2.073941	2.942542	-3.257762
37	1	0	2.734078	4.136497	-2.121204
38	1	0	3.381297	2.473127	-2.162942
39	1	0	1.819225	4.247808	1.895760
40	1	0	1.938792	5.007767	0.298533
41	1	0	0.391980	4.324767	0.849278
42	1	0	0.212610	4.409083	-1.622576
43	1	0	-0.270370	2.940659	-2.501260
44	1	0	-0.662006	3.113676	-0.782585
45	1	0	3.976720	3.355973	-0.045572
46	1	0	3.604381	2.649130	1.524797
47	1	0	3.639614	1.605837	0.071175
48	1	0	1.156414	-4.430097	-0.029471
49	1	0	2.915984	-4.660696	0.009356
50	1	0	2.127186	-3.557979	1.173067
51	1	0	2.940343	-0.390876	-2.058430
52	1	0	4.541079	-0.172163	-1.326609
53	1	0	4.289873	-1.490218	-2.487365
54	1	0	5.050403	-3.256374	-0.709433
55	1	0	5.461225	-1.812285	0.233452
56	1	0	4.445514	-3.078248	0.954281
57	1	0	-1.076621	1.759876	1.604405
58	1	0	-3.562596	1.368440	1.683590
59	1	0	-3.097498	-1.079950	-1.833232
60	1	0	-0.636668	-0.579275	-1.732416
61	1	0	-5.136822	-0.393094	2.063176
62	1	0	-7.568668	-0.835826	2.062168
63	1	0	-8.825081	-0.890267	-0.080888
64	1	0	-7.629478	-0.485757	-2.220514
65	1	0	-5.202507	-0.018598	-2.216715
66	1	0	-0.428096	-2.673944	2.660105
67	1	0	-1.244556	-1.387895	1.748238
68	1	0	-0.476166	-2.738522	0.881331
69	19	0	2.510828	0.265858	2.676980

Summary of Natural Population Analysis:

Natural Population						
Atom	No	Natural Charge	Core	Valence	Rydberg	Total
O	1	-0.87434	1.99974	6.85595	0.01866	8.87434
B	2	0.86460	1.99813	2.09821	0.03906	4.13540
O	3	-0.82472	1.99972	6.80590	0.01910	8.82472
B	4	0.77701	1.99810	2.18774	0.03715	4.22299
O	5	-0.86238	1.99975	6.84377	0.01887	8.86238
O	6	-0.80280	1.99971	6.78319	0.01990	8.80280
C	7	0.24225	1.99885	3.73398	0.02492	5.75775
C	8	0.24797	1.99886	3.72809	0.02508	5.75203
C	9	0.24015	1.99884	3.73489	0.02612	5.75985
C	10	0.24146	1.99885	3.73489	0.02480	5.75854
C	11	-0.69158	1.99932	4.68239	0.00987	6.69158
C	12	-0.69395	1.99933	4.68434	0.01028	6.69395
C	13	-0.69312	1.99932	4.68394	0.00986	6.69312
C	14	-0.70396	1.99934	4.69435	0.01028	6.70396
C	15	-0.70807	1.99933	4.69760	0.01114	6.70807
C	16	-0.70789	1.99934	4.69788	0.01066	6.70789
C	17	-0.70872	1.99933	4.69848	0.01090	6.70872
C	18	-0.69154	1.99932	4.68268	0.00954	6.69154
N	19	-0.47952	1.99928	5.44847	0.03177	7.47952
C	20	0.05463	1.99910	3.92433	0.02195	5.94537
C	21	-0.26637	1.99897	4.25447	0.01294	6.26637
C	22	-0.00141	1.99898	3.98460	0.01783	6.00141
C	23	-0.26076	1.99898	4.24887	0.01292	6.26076
C	24	0.08313	1.99910	3.89563	0.02214	5.91687
C	25	-0.07452	1.99897	4.05924	0.01631	6.07452
C	26	-0.22062	1.99899	4.20750	0.01413	6.22062
C	27	-0.23864	1.99905	4.22382	0.01576	6.23864
C	28	-0.23783	1.99904	4.22349	0.01530	6.23783
C	29	-0.23808	1.99905	4.22327	0.01576	6.23808
C	30	-0.21952	1.99899	4.20646	0.01407	6.21952
O	31	-0.88770	1.99974	6.86459	0.02338	8.88770
C	32	-0.29755	1.99938	4.28219	0.01599	6.29755
H	33	0.22912	0.00000	0.77043	0.00045	0.77088
H	34	0.23904	0.00000	0.76007	0.00089	0.76096
H	35	0.24757	0.00000	0.75174	0.00068	0.75243
H	36	0.24713	0.00000	0.75204	0.00083	0.75287

H	37	0.22883	0.00000	0.77070	0.00047	0.77117
H	38	0.24941	0.00000	0.74987	0.00073	0.75059
H	39	0.23270	0.00000	0.76640	0.00090	0.76730
H	40	0.23918	0.00000	0.76033	0.00049	0.76082
H	41	0.24420	0.00000	0.75505	0.00075	0.75580
H	42	0.23375	0.00000	0.76560	0.00065	0.76625
H	43	0.24914	0.00000	0.74997	0.00089	0.75086
H	44	0.23196	0.00000	0.76727	0.00077	0.76804
H	45	0.24436	0.00000	0.75512	0.00052	0.75564
H	46	0.20523	0.00000	0.79387	0.00090	0.79477
H	47	0.26282	0.00000	0.73591	0.00127	0.73718
H	48	0.23900	0.00000	0.76023	0.00078	0.76100
H	49	0.22782	0.00000	0.77149	0.00069	0.77218
H	50	0.24885	0.00000	0.74977	0.00138	0.75115
H	51	0.27175	0.00000	0.72717	0.00108	0.72825
H	52	0.22683	0.00000	0.77241	0.00077	0.77317
H	53	0.22771	0.00000	0.77173	0.00056	0.77229
H	54	0.23447	0.00000	0.76509	0.00044	0.76553
H	55	0.23346	0.00000	0.76559	0.00095	0.76654
H	56	0.23946	0.00000	0.75985	0.00069	0.76054
H	57	0.26677	0.00000	0.73207	0.00116	0.73323
H	58	0.25932	0.00000	0.73980	0.00088	0.74068
H	59	0.26363	0.00000	0.73557	0.00080	0.73637
H	60	0.29366	0.00000	0.70525	0.00110	0.70634
H	61	0.24957	0.00000	0.74947	0.00096	0.75043
H	62	0.25206	0.00000	0.74716	0.00077	0.74794
H	63	0.25158	0.00000	0.74775	0.00067	0.74842
H	64	0.25267	0.00000	0.74656	0.00077	0.74733
H	65	0.25084	0.00000	0.74825	0.00091	0.74916
H	66	0.19782	0.00000	0.80120	0.00098	0.80218
H	67	0.16741	0.00000	0.83059	0.00200	0.83259
H	68	0.20927	0.00000	0.78907	0.00165	0.79073
K	69	0.98600	17.98395	0.02467	0.00537	18.01400

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* Total * -0.00000 81.95674 173.42030 0.62295 256.00000

Cartesian coordinates of **TS-1•K**:

Center Number	Atomic Number	Atomic Type	Coordinates (Angstroms)		
			X	Y	Z
1	8	0	-2.129782	1.552321	0.790139
2	5	0	-0.752675	1.274221	0.519344
3	8	0	-0.325084	1.947220	-0.618707
4	5	0	-0.986132	-1.061169	-0.198186
5	8	0	-1.620853	-1.829818	0.853589
6	8	0	-1.807331	-1.078884	-1.350087
7	6	0	-1.266411	3.003045	-0.854060
8	6	0	-2.603666	2.440525	-0.246879
9	6	0	-2.581200	-2.276793	-1.244831
10	6	0	-2.821324	-2.394739	0.298771
11	6	0	-1.317582	3.287856	-2.346758
12	6	0	-3.849221	-2.119715	-2.070215
13	6	0	-2.973523	-3.823763	0.802894
14	6	0	-1.733902	-3.432911	-1.784606
15	6	0	-4.003126	-1.544436	0.769175
16	6	0	-0.750738	4.232317	-0.100363
17	6	0	-3.421951	1.647890	-1.261737
18	6	0	-3.485942	3.499844	0.403027
19	7	0	0.436163	-1.047537	-0.265655
20	6	0	1.232811	-1.510176	0.796602
21	6	0	2.588414	-1.328705	0.811152
22	6	0	3.287485	-0.717392	-0.254508
23	6	0	2.484145	-0.441802	-1.399137
24	6	0	1.135503	-0.625105	-1.411385
25	6	0	4.720951	-0.454140	-0.197117
26	6	0	5.388851	-0.315146	1.038879
27	6	0	6.757806	-0.080924	1.108154
28	6	0	7.518727	0.043367	-0.054754
29	6	0	6.876345	-0.076976	-1.288697
30	6	0	5.510145	-0.323220	-1.360214
31	8	0	0.043126	1.118092	1.665790
32	6	0	1.391689	1.570357	1.598045
33	1	0	-2.084689	4.036731	-2.576765
34	1	0	-0.348462	3.677544	-2.672339
35	1	0	-1.526283	2.377335	-2.912677
36	1	0	-3.586882	-2.067558	-3.131346
37	1	0	-4.520356	-2.974815	-1.925642
38	1	0	-4.383052	-1.202395	-1.808277
39	1	0	-3.148622	-3.822559	1.884679

40	1	0	-3.828629	-4.315755	0.324077
41	1	0	-2.073600	-4.409955	0.606718
42	1	0	-2.287361	-4.378187	-1.790994
43	1	0	-1.439436	-3.193721	-2.810455
44	1	0	-0.819016	-3.561250	-1.195275
45	1	0	-4.955542	-1.905849	0.369212
46	1	0	-4.080289	-1.601402	1.864458
47	1	0	-3.863348	-0.500523	0.470760
48	1	0	0.265238	4.447805	-0.444067
49	1	0	-1.375567	5.114199	-0.277773
50	1	0	-0.710657	4.043072	0.978627
51	1	0	-2.830839	0.844755	-1.708036
52	1	0	-4.285764	1.201960	-0.755761
53	1	0	-3.805072	2.306703	-2.048378
54	1	0	-3.787596	4.253661	-0.333636
55	1	0	-4.394934	3.029853	0.793420
56	1	0	-2.976924	4.002283	1.228655
57	1	0	0.700244	-2.053185	1.568772
58	1	0	3.134027	-1.727490	1.663760
59	1	0	2.934639	-0.059583	-2.310622
60	1	0	0.510465	-0.397417	-2.265516
61	1	0	4.815398	-0.363053	1.961061
62	1	0	7.231991	0.023196	2.081091
63	1	0	8.586426	0.233324	-0.001788
64	1	0	7.449276	0.008716	-2.208810
65	1	0	5.049564	-0.445631	-2.336380
66	1	0	1.938790	1.114388	2.426266
67	1	0	1.867542	1.278351	0.658494
68	1	0	1.418300	2.662843	1.690997
69	19	0	-1.838654	-0.138731	2.840151

Summary of Natural Population Analysis:

		Natural Population				
Atom	No	Natural Charge	Core	Valence	Rydberg	Total
O	1	-0.85516	1.99971	6.83855	0.01691	8.85516
B	2	1.24467	1.99896	1.70666	0.04971	3.75533
O	3	-0.80524	1.99967	6.77989	0.02568	8.80524
B	4	1.05651	1.99886	1.90346	0.04117	3.94349
O	5	-0.84572	1.99973	6.82848	0.01751	8.84572
O	6	-0.79270	1.99970	6.77375	0.01924	8.79270

C	7	0.24142	1.99885	3.73721	0.02251	5.75858
C	8	0.24594	1.99884	3.73185	0.02336	5.75406
C	9	0.23885	1.99885	3.73734	0.02497	5.76115
C	10	0.23959	1.99885	3.73698	0.02459	5.76041
C	11	-0.69841	1.99932	4.68858	0.01051	6.69841
C	12	-0.69735	1.99932	4.68779	0.01024	6.69735
C	13	-0.69743	1.99932	4.68787	0.01024	6.69743
C	14	-0.71168	1.99934	4.70085	0.01148	6.71168
C	15	-0.70740	1.99932	4.69697	0.01112	6.70740
C	16	-0.70656	1.99934	4.69640	0.01083	6.70656
C	17	-0.71446	1.99931	4.70386	0.01129	6.71446
C	18	-0.69719	1.99932	4.68767	0.01020	6.69719
N	19	-0.66107	1.99909	5.64140	0.02058	7.66107
C	20	-0.13233	1.99906	4.10376	0.02952	6.13233
C	21	-0.22597	1.99900	4.21291	0.01407	6.22597
C	22	-0.21349	1.99899	4.19381	0.02069	6.21349
C	23	-0.23234	1.99901	4.21999	0.01335	6.23234
C	24	-0.07746	1.99904	4.05369	0.02473	6.07746
C	25	-0.03872	1.99900	4.02342	0.01629	6.03872
C	26	-0.25628	1.99902	4.24249	0.01477	6.25628
C	27	-0.24192	1.99905	4.22645	0.01642	6.24192
C	28	-0.29027	1.99903	4.27300	0.01824	6.29027
C	29	-0.24043	1.99905	4.22509	0.01628	6.24043
C	30	-0.25024	1.99901	4.23664	0.01458	6.25024
O	31	-0.86376	1.99965	6.84549	0.01861	8.86376
C	32	-0.30424	1.99934	4.28910	0.01579	6.30424
H	33	0.23627	0.00000	0.76332	0.00041	0.76373
H	34	0.25402	0.00000	0.74523	0.00076	0.74598
H	35	0.25653	0.00000	0.74279	0.00068	0.74347
H	36	0.25488	0.00000	0.74434	0.00078	0.74512
H	37	0.23795	0.00000	0.76165	0.00040	0.76205
H	38	0.24439	0.00000	0.75486	0.00076	0.75561
H	39	0.23468	0.00000	0.76452	0.00080	0.76532
H	40	0.24261	0.00000	0.75694	0.00045	0.75739
H	41	0.25860	0.00000	0.74075	0.00065	0.74140
H	42	0.23675	0.00000	0.76267	0.00057	0.76325
H	43	0.25692	0.00000	0.74232	0.00076	0.74308
H	44	0.25363	0.00000	0.74537	0.00100	0.74637
H	45	0.25257	0.00000	0.74696	0.00047	0.74743
H	46	0.20892	0.00000	0.79021	0.00087	0.79108
H	47	0.25290	0.00000	0.74579	0.00130	0.74710
H	48	0.25727	0.00000	0.74191	0.00082	0.74273
H	49	0.24005	0.00000	0.75935	0.00059	0.75995
H	50	0.23782	0.00000	0.76147	0.00071	0.76218

H	51	0.27902	0.00000	0.71978	0.00121	0.72098
H	52	0.22899	0.00000	0.77031	0.00070	0.77101
H	53	0.24115	0.00000	0.75838	0.00047	0.75885
H	54	0.24387	0.00000	0.75569	0.00044	0.75613
H	55	0.24157	0.00000	0.75759	0.00085	0.75843
H	56	0.24572	0.00000	0.75362	0.00066	0.75428
H	57	0.22903	0.00000	0.77003	0.00094	0.77097
H	58	0.24316	0.00000	0.75577	0.00107	0.75684
H	59	0.24568	0.00000	0.75350	0.00081	0.75432
H	60	0.24994	0.00000	0.74914	0.00092	0.75006
H	61	0.23088	0.00000	0.76801	0.00111	0.76912
H	62	0.23804	0.00000	0.76109	0.00087	0.76196
H	63	0.23924	0.00000	0.75998	0.00078	0.76076
H	64	0.23945	0.00000	0.75971	0.00084	0.76055
H	65	0.23540	0.00000	0.76365	0.00095	0.76460
H	66	0.21581	0.00000	0.78346	0.00073	0.78419
H	67	0.23683	0.00000	0.76124	0.00192	0.76317
H	68	0.20126	0.00000	0.79773	0.00100	0.79874
K	69	0.98903	17.98719	0.01946	0.00431	18.01097

=====
* Total * 0.00000 81.96112 173.40002 0.63886 256.00000

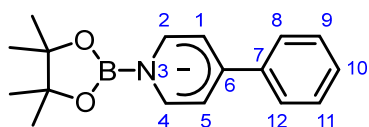
3.3 DFT calculated HOMO and NAO orbital composition of Int-B

Cartesian coordinates of Int-B:

Center Number	Atomic Number	Atomic Type	Coordinates (Angstroms)		
			X	Y	Z
1	6	0	-1.691014	1.168859	-0.364006
2	6	0	-0.341956	1.170027	-0.414371
3	7	0	0.423579	-0.020200	-0.194879
4	6	0	-0.340693	-1.230641	-0.164869
5	6	0	-1.689712	-1.220823	-0.116148
6	6	0	-2.487888	-0.015101	-0.131547
7	6	0	-3.901202	-0.003194	-0.010758
8	6	0	-4.680235	1.201953	-0.060790
9	6	0	-6.061478	1.202962	0.043365
10	6	0	-6.793190	0.018993	0.213260
11	6	0	-6.061162	-1.174908	0.282500
12	6	0	-4.679944	-1.194358	0.179993
13	5	0	1.810421	-0.007542	-0.079284
14	8	0	2.606880	1.142296	-0.196796
15	6	0	3.917351	0.778087	0.239170

16	6	0	3.956209	-0.755965	-0.065941
17	8	0	2.599431	-1.145237	0.151979
18	6	0	4.860219	-1.564807	0.853747
19	6	0	4.297269	-1.047922	-1.529587
20	6	0	4.943642	1.604758	-0.522744
21	6	0	4.010231	1.074231	1.738284
22	1	0	-2.174618	2.124666	-0.549157
23	1	0	0.246901	2.056032	-0.621844
24	1	0	0.249599	-2.139548	-0.182505
25	1	0	-2.171912	-2.194888	-0.099514
26	1	0	-4.176584	2.157276	-0.177055
27	1	0	-6.585982	2.157367	-0.004845
28	1	0	-7.876076	0.027317	0.297569
29	1	0	-6.585497	-2.119957	0.424912
30	1	0	-4.176272	-2.153772	0.255140
31	1	0	5.897660	-1.214769	0.786820
32	1	0	4.527894	-1.495103	1.892045
33	1	0	4.832313	-2.618567	0.558202
34	1	0	4.102719	-2.106247	-1.728119
35	1	0	3.666225	-0.454190	-2.199069
36	1	0	5.349013	-0.835415	-1.752652
37	1	0	5.962683	1.273325	-0.288216
38	1	0	4.785542	1.531173	-1.601192
39	1	0	4.849222	2.657213	-0.236453
40	1	0	3.766229	2.128438	1.900547
41	1	0	3.289323	0.468500	2.297104
42	1	0	5.015293	0.880882	2.130058

The HOMO of **Int-B** was plotted using the softwares Multiwfn^[3] and VMD,^[4] with isovalue = 0.07. The result of the NAO orbital composition analysis was listed below.



NAO#	Center	Label	Type	Composition
1	1 (C)	S	Cor (1S)	0.000036%
2	1 (C)	S	Val (2S)	0.009409%
6	1 (C)	px	Val (2p)	0.003600%
9	1 (C)	py	Val (2p)	0.029929%
12	1 (C)	pz	Val (2p)	0.765625%
20	2 (C)	S	Cor (1S)	0.000025%
21	2 (C)	S	Val (2S)	0.008649%
25	2 (C)	px	Val (2p)	0.015376%

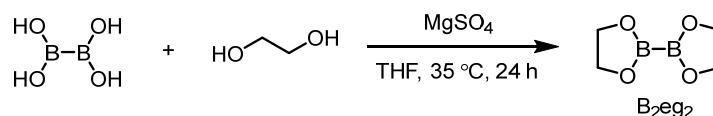
28	2 (C)	py	Val (2p)	0.439569%
31	2 (C)	pz	Val (2p)	12.110400%
39	3 (N)	S	Cor (1S)	0.000004%
40	3 (N)	S	Val (2S)	0.001369%
44	3 (N)	px	Val (2p)	0.041616%
47	3 (N)	py	Val (2p)	0.074529%
50	3 (N)	pz	Val (2p)	7.054336%
58	4 (C)	S	Cor (1S)	0.000025%
59	4 (C)	S	Val (2S)	0.010000%
63	4 (C)	px	Val (2p)	0.015376%
66	4 (C)	py	Val (2p)	0.004761%
69	4 (C)	pz	Val (2p)	12.517444%
77	5 (C)	S	Cor (1S)	0.000036%
78	5 (C)	S	Val (2S)	0.008464%
82	5 (C)	px	Val (2p)	0.003136%
85	5 (C)	py	Val (2p)	0.000169%
88	5 (C)	pz	Val (2p)	0.808201%
96	6 (C)	S	Cor (1S)	0.000049%
97	6 (C)	S	Val (2S)	0.002809%
101	6 (C)	px	Val (2p)	0.304704%
104	6 (C)	py	Val (2p)	0.297025%
107	6 (C)	pz	Val (2p)	27.510025%

Condensed above result to atoms:

1 (C)	0.808599%
2 (C)	12.574019%
3 (N)	7.171854%
4 (C)	12.547606%
5 (C)	0.820006%
6 (C)	28.114612%

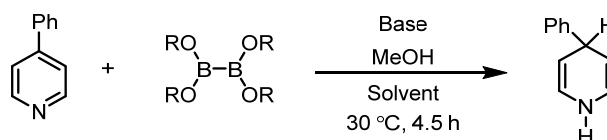
4. Optimization of Reaction Conditions

4.1 The synthesis of bis(ethyleneglycolato)diboron (B_2eg_2)



B_2eg_2 was prepared according to a published procedure^[5] with some modification. A 125 mL round-bottom flask was charged with a magnetic stir bar, $B_2(OH)_4$ (2.0 g, 22.4 mmol), anhydrous $MgSO_4$ (3.22 g, 26.8 mmol) and 40 mL THF. Then freshly distilled ethylene glycol (2.9 g, 46.8 mmol) was added to the flask. The flask was sealed with a rubber stopper and the reaction mixture was stirred at 35 °C for 24 h. The reaction mixture was filtered, and the residue was washed by THF. The combined filtrate was concentrated, and the residue was further purified by sublimation at reduced pressure (150 °C, 2.5 Pa) to get a white solid (1.8 g, 58% yield). The spectroscopic data was consistent with those reported in literature.^[5]

4.2 Experimental procedure for the optimization study

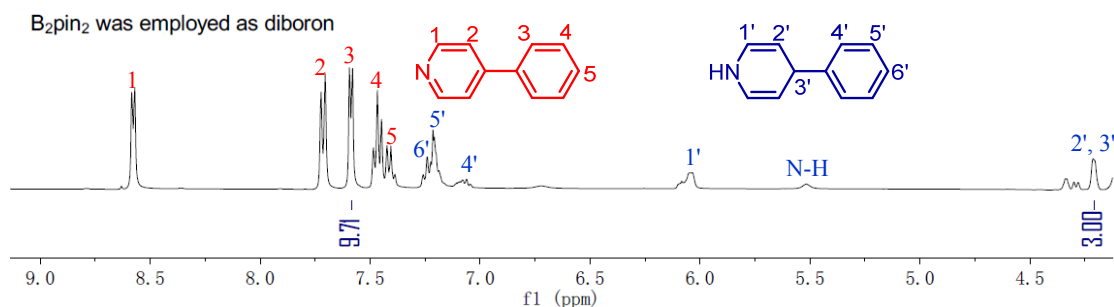


In a glove box, a 15 mL oven-dried glass vial was charged with 4-PhPy (56.5 mg, 0.36 mmol), diboron(4) (0.40 mmol), base (0.40 mmol), and 2 mL of anhydrous solvent. Then MeOH (128 mg, 4.0 mmol) was added by a microsyringe. The vial was sealed and the resulting mixture was allowed to react at 30 °C for 4.5 h. The crude reaction solution was transferred to an NMR tube in a glove box, and then 1H NMR analysis (No-D NMR) of the reaction solution was performed.

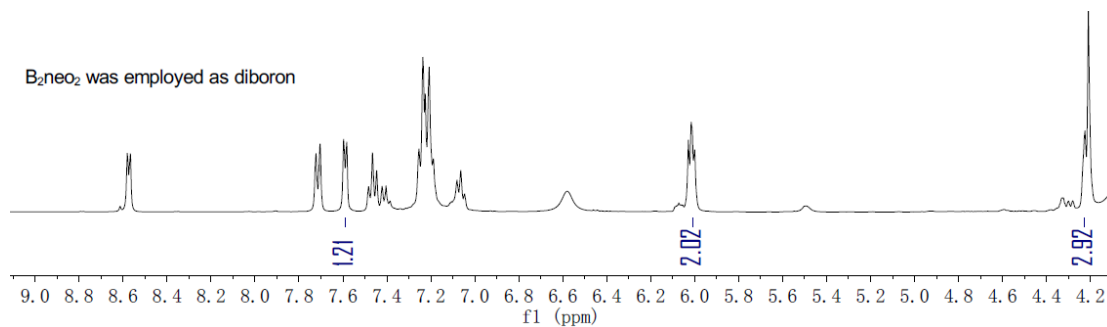
4.3 1H NMR spectra of the optimization experiments

1H NMR spectra of the crude reaction mixtures of optimization experiments are shown below, which were used to calculate the conversions listed in Table 1 of the main text. The integrals of the 1,4-DHP product (6.04 ppm) and the unreacted 4-phenylpyridine (7.59 ppm) were used to calculate the NMR conversion, assuming that 1,4-DHP was the only product of the converted 4-phenylpyridine.

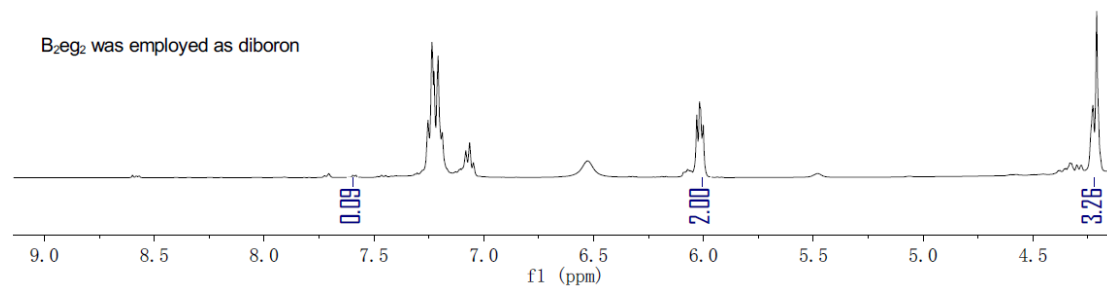
Entry 1: B_2pin_2 , MeOK in THF (17% conversion)



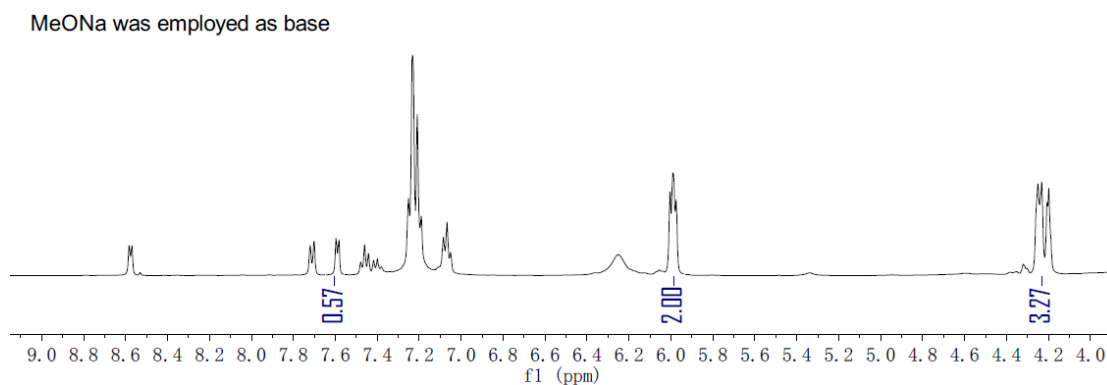
Entry 2: B₂neo₂, MeOK in THF (63% conversion)



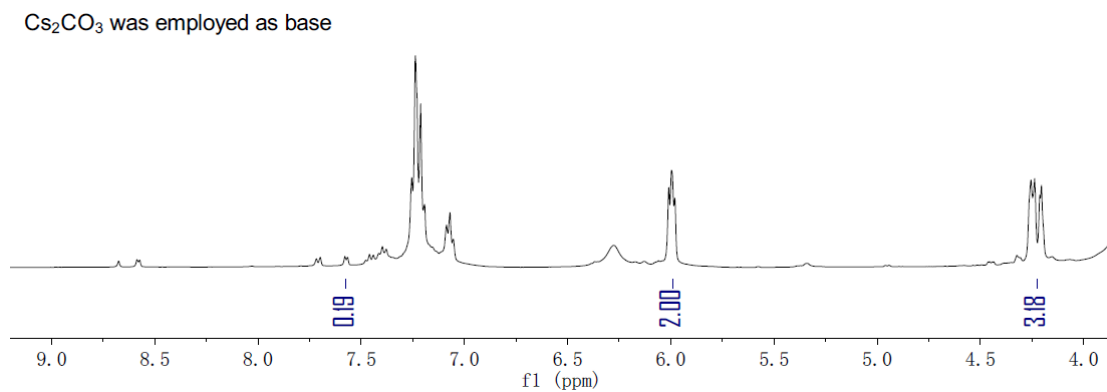
Entry 3: B₂eg₂, MeOK in THF (96% conversion)



Entry 4: B₂eg₂, MeONa in THF (78% conversion)

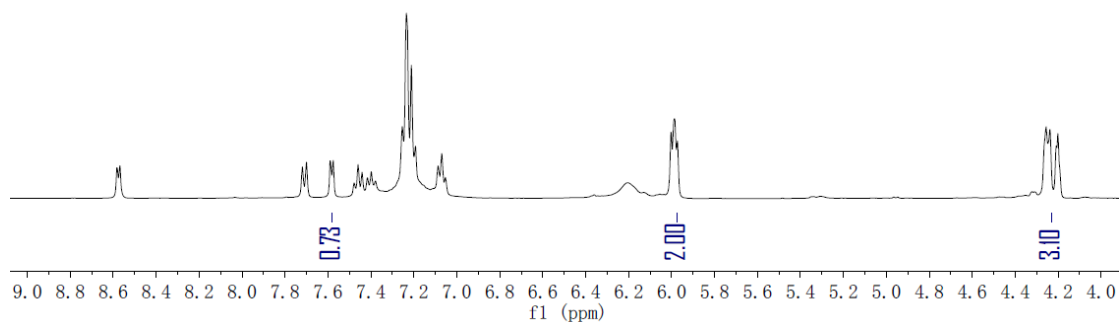


Entry 5: B₂eg₂, Cs₂CO₃ in THF (91% conversion)



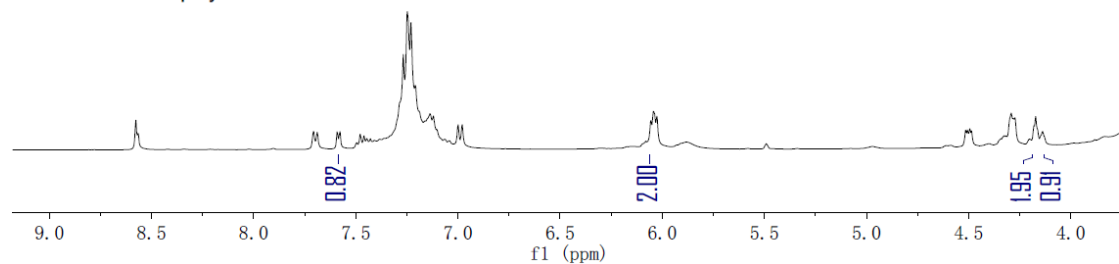
Entry 6: B₂eg₂, K₂CO₃ in THF (73% conversion)

K₂CO₃ was employed as base



Entry 7: B₂eg₂, Cs₂CO₃ in MeCN (71% conversion)

MeCN was employed as solvent



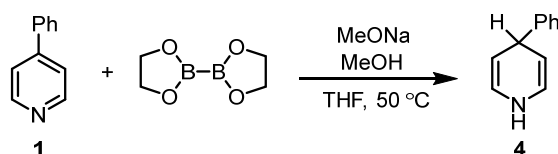
Note: the NMR shifts of the peaks of the 1,4-DHP product in the crude reaction mixture were slightly different from those obtained in the preliminary experiments (Section 2.2). This was due to the excess MeOH (11 equiv.) added in the optimization experiments, which may form hydrogen bonds with the 1,4-DHP product. This was confirmed by the observation that addition of excess MeOH to the previous reaction mixtures caused the shift of the product peaks.

5. Substrate Scope and Characterization Data

General procedure (GP-A) for determination of the NMR yield: In a glove box, a 15 mL oven-dried reaction vial was charged with pyridine substrate (0.18 mmol), B₂eg₂ (28.4 mg, 0.20 mmol), base (0.20 mmol), and anhydrous THF (2 mL). MeOH (64.0 mg, 2.00 mmol) or MeOD (134 mg, 4.00 mmol) was then added by a microsyringe. The vial was sealed by a screw cap and the resulting mixture was then allowed to react at 50 °C for 14 h. The reaction mixture was cooled to room temperature, and a standard solution of DMSO in THF was added as the internal standard under an argon atmosphere. The crude reaction solution was transferred to an NMR tube in a glove box, and then ¹H NMR analysis (No-D NMR) of the reaction solution was performed to determine the NMR yield of the 1,4-DHP product.

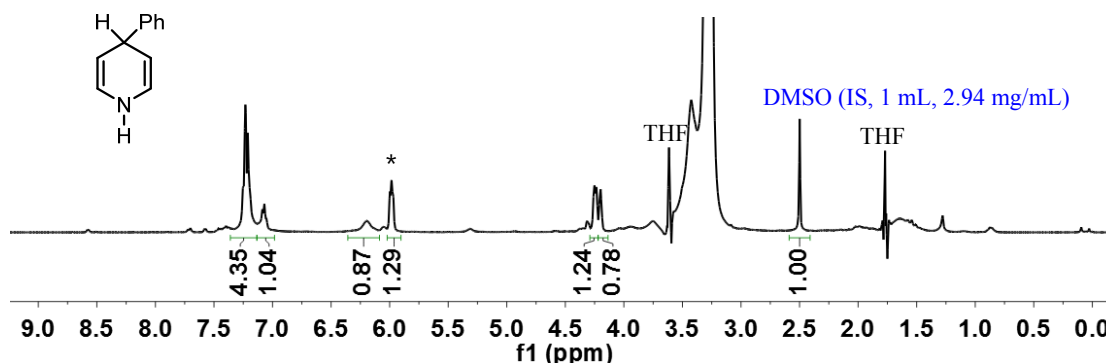
General procedure (GP-B) for obtaining the pure 1,4-DHP product: In a glove box, a 25 mL Schlenk tube was charged with the pyridine substrate (0.36 mmol), B₂eg₂ (56.8 mg, 0.40 mmol), Cs₂CO₃ (130 mg, 0.40 mmol) and anhydrous MeCN (2 mL). MeOH (128 mg, 4.00 mmol) or MeOD (264 mg, 8.00 mmol) was added by a microsyringe. The tube was sealed by a rubber septum and the resulting mixture was allowed to react at 50 °C for 14 h. The reaction solution was cooled to room temperature and was transferred to a column packed with a pad of basic alumina (6 g, dried in the oven prior to use) using degassed anhydrous diethyl ether. A positive pressure of argon was applied to drive the solution pass through the column, and then the column was washed by 3 × 20 mL degassed diethyl ether. The combined organic solution was concentrated under vacuum under an argon atmosphere to obtain the 1,4-dihydropyridine product.

4-Phenyl-1,4-dihydropyridine (**4**)



Determination of the NMR yield. Following the general procedure GP-A, starting from **1** (28.0 mg, 0.18 mmol), B₂eg₂ (28.4 mg, 0.20 mmol), and MeOH (64.0 mg, 2.00 mmol), and using MeONa (10.8 mg, 0.20 mmol) as the base in 2 mL of anhydrous THF, 1,4-DHP product **4** was generated in 81% NMR yield.

¹H NMR of the reaction mixture (* denotes the peak used to calculate the NMR yield):



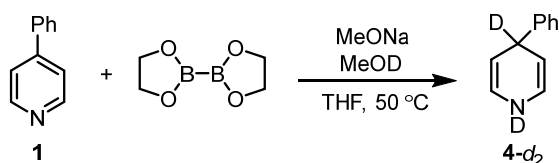
Note: This 1,4-DHP product was rather oxygen-labile, and attempts to obtain its pure form following general procedure GP-B failed. The NMR data were acquired using the No-D NMR technique from the above reaction mixture.

^1H NMR (400 MHz, $\text{THF-}d_0$): δ 7.24 (m, 4H), 7.07 (m, 1H), 6.17 (br s, 1H), 5.99 (m, 2H), 4.26 (d, J = 6.8 Hz, 2H), 4.20 (m, 1H).

^{13}C NMR (100 MHz, $\text{THF-}d_0$): δ 150.5, 127.8, 127.4, 125.8, 125.3, 100.0, 39.9.

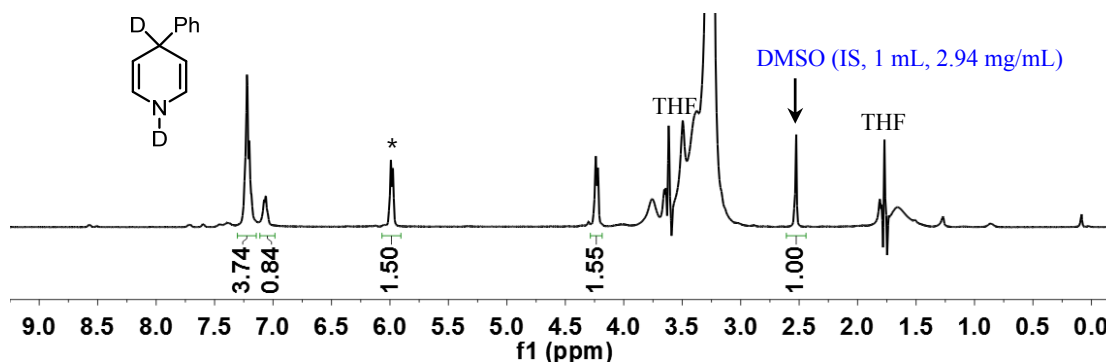
GC-MS: $\text{C}_{11}\text{H}_{11}\text{N}$ (m/z = 157, M^+).

4-Phenyl-1,4-dideuteropyridine (**4- d_2**)



Determination of the NMR yield. Following the general procedure GP-A, starting from **1** (28.0 mg, 0.18 mmol), B_{2}eg_2 (28.4 mg, 0.20 mmol), and MeOD (132 mg, 4.00 mmol), and using MeONa (10.8 mg, 0.20 mmol) as the base in 2 mL of anhydrous THF, 1,4-dideuteropyridine product **4- d_2** was generated in 94% NMR yield (>98% 4-D).

^1H NMR of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Note: This 1,4-DHP product was rather oxygen-labile, and attempts to obtain its pure form following general procedure GP-B failed. The NMR data were acquired using the No-D NMR technique from the above reaction mixture.

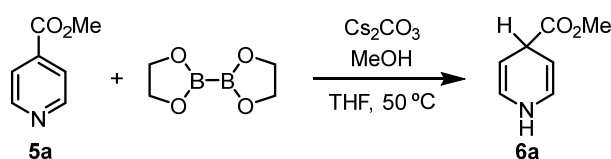
^1H NMR (400 MHz, $\text{THF-}d_0$): δ 7.24 (m, 4H), 7.06 (m, 1H), 5.99 (d, J = 7.6 Hz, 2H), 4.26 (d, J = 7.6 Hz, 2H).

^2H NMR (61 MHz, $\text{THF-}d_0$): δ 6.24, 4.17.

^{13}C NMR (100 MHz, $\text{THF-}d_0$): δ 150.5, 127.8, 127.4, 125.7, 125.3, 99.8, 39.4 (t, J = 20.1 Hz).

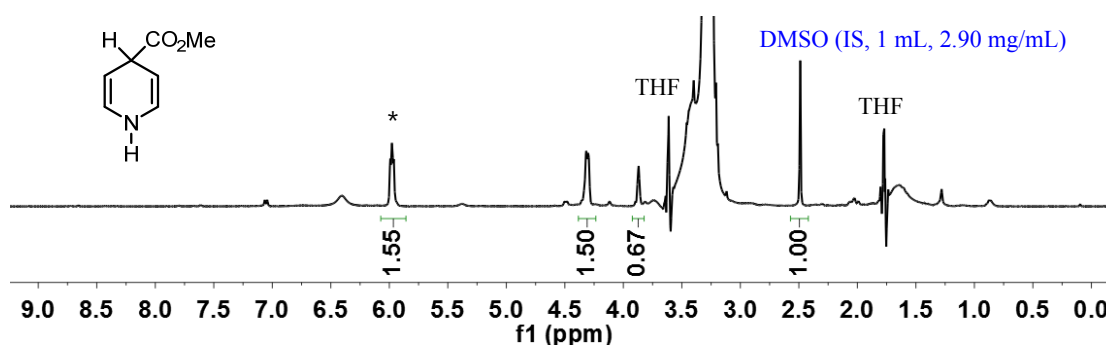
GC-MS: $\text{C}_{11}\text{H}_9\text{ND}_2$ [m/z = 159, ($\text{M-}d_2$) $^+$].

Methyl 1,4-dihydropyridine-4-carboxylate (**6a**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5a** (25.0 mg, 0.18 mmol), B₂eg₂ (28.4 mg, 0.20 mmol), and MeOH (64.0 mg, 2.00 mmol), and using Cs₂CO₃ (65.2 mg, 0.20 mmol) as the base in 2 mL of anhydrous THF, 1,4-DHP product **6a** was generated in 96% NMR yield.

¹H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



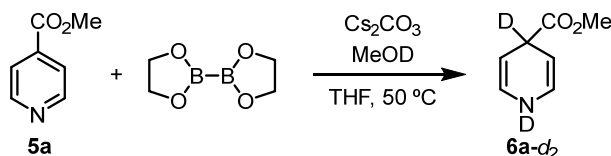
Note: This 1,4-DHP product was rather oxygen-labile, and attempts to obtain its pure form following general procedure GP-B failed. The NMR data were acquired using the No-D NMR technique from the above reaction mixture.

¹H NMR (400 MHz, THF-*d*₀): δ 6.34 (br s, 1H), 5.97 (dd, *J* = 7.5, 4.9 Hz, 2H), 4.31 (dd, *J* = 7.5, 3.7 Hz, 2H), 3.87 (t, *J* = 3.7 Hz, 1H).

¹³C NMR (100 MHz, THF-*d*₀): δ 173.4, 127.3, 93.7, 50.5, 39.2.

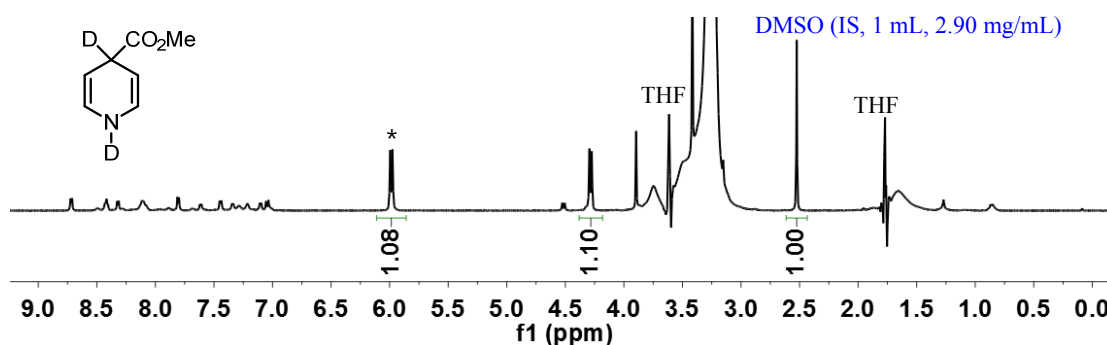
GC-MS: C₇H₉NO₂ [*m/z* = 139, (M⁺)].

Methyl 1,4-dideuteropyridine-4-carboxylate (**6a-d₂**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5a** (25.0 mg, 0.18 mmol), B₂eg₂ (28.4 mg, 0.20 mmol), and MeOD (132 mg, 4.00 mmol), and using Cs₂CO₃ (65.2 mg, 0.20 mmol) as the base in 2 mL of anhydrous THF, 1,4-dideuteropyridine product **6a-d₂** was generated in 67% NMR yield (>98% 4-D).

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Note: This 1,4-DHP product was rather oxygen-labile, and attempts to obtain its pure form following general procedure GP-B failed. The NMR data were acquired using the No-D NMR technique from the above reaction mixture.

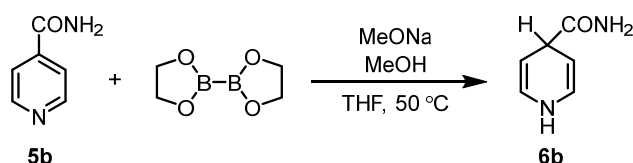
^1H NMR (400 MHz, THF- d_6): δ 5.99 (d, J = 7.6 Hz, 2H), 4.29 (d, J = 7.6 Hz, 2H).

^2H NMR (61 MHz, THF- d_6): δ 6.52, 3.84.

^{13}C NMR (100 MHz, THF- d_6): δ 173.7, 127.4, 93.5, 50.6, 38.9 (t, J = 19.8 Hz).

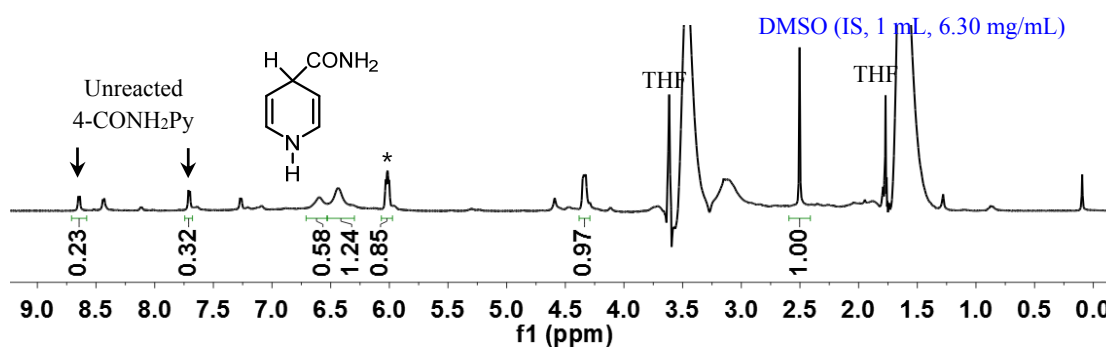
GC-MS: $\text{C}_7\text{H}_7\text{NO}_2\text{D}_2$ [m/z = 141, (M- d_2) $^+$].

1,4-Dihydropyridine-4-carboxamide (6b)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5b** (45.0 mg, 0.36 mmol), B₂eg₂ (56.8 mg, 0.40 mmol), and MeOH (128 mg, 4.00 mmol), and using MeONa (21.6 mg, 0.40 mmol) as the base in 2 mL of anhydrous THF, 1,4-DHP product **6b** was generated in 57% NMR yield.

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



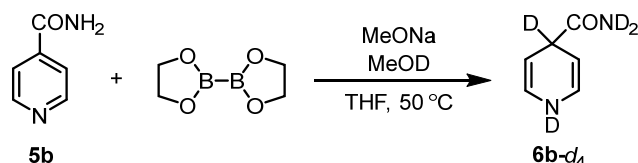
Note: This 1,4-DHP product was rather oxygen-labile, and attempts to obtain its pure form following general procedure GP-B failed. The NMR data were acquired using the No-D NMR technique from the above reaction mixture.

^1H NMR (400 MHz, THF- d_0): δ 6.48 (br s, 1H), 6.40 (br s, 2H), 6.01 (m, 2H), 4.34 (m, 2H). The peak of C4-H overlapped with the THF peak at 3.61 ppm.

^{13}C NMR (100 MHz, THF- d_0): δ 177.0, 127.4, 95.7, 41.6.

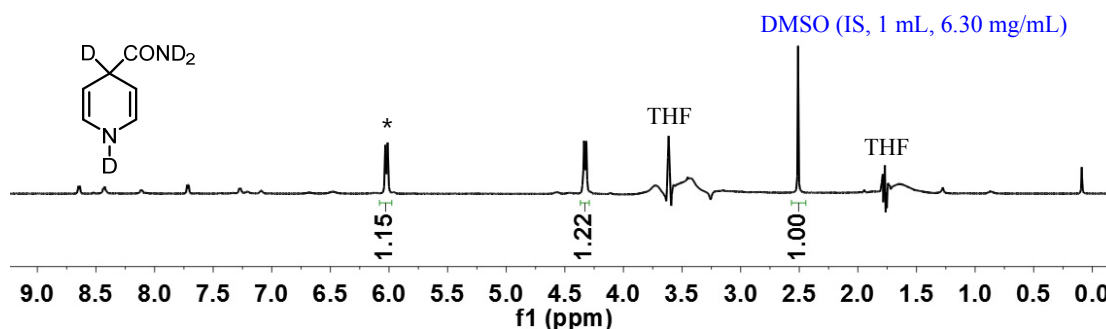
GC-MS: $\text{C}_6\text{H}_8\text{N}_2\text{O}$ [$m/z = 124$, (M^+)].

1,4-Dideuteropyridine-4-carboxamide (**6b-d₄**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5b** (45.0 mg, 0.36 mmol), B₂eg₂ (56.8 mg, 0.40 mmol), and MeOD (264 mg, 7.99 mmol), and using MeONa (21.6 mg, 0.40 mmol) as the base in 2 mL of anhydrous THF, 1,4-dideuteropyridine product **6b-d₄** was generated in 77% NMR yield (ratio of deuterium incorporation at the C4-position was not calculated due to overlap of the residue C4-H peak with other peaks).

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Note: This 1,4-DHP product was rather oxygen-labile, and attempts to obtain its pure form following general procedure GP-B failed. The NMR data were acquired using the No-D NMR technique from the above reaction mixture.

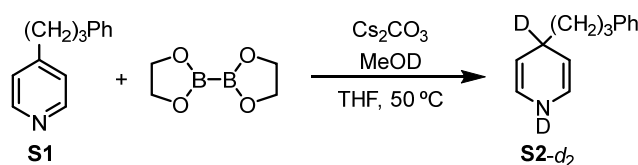
^1H NMR (400 MHz, THF- d_0): δ 6.02 (d, $J = 7.6$ Hz, 2H), 4.33 (d, $J = 7.6$ Hz, 2H).

^2H NMR (61 MHz, THF- d_0): δ 6.63, 6.50, 3.61.

^{13}C NMR (100 MHz, THF- d_0): δ 177.3, 127.5, 95.3, 41.1 (t, $J = 21.0$ Hz).

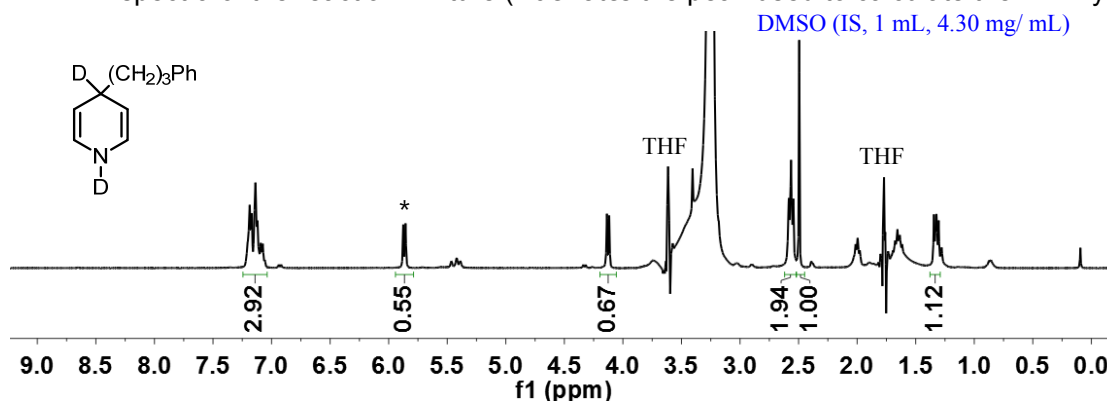
GC-MS: $\text{C}_6\text{H}_4\text{N}_2\text{OD}_4$ [$m/z = 128$, (M-d_4)⁺].

4-(3-Phenylpropyl)-1,4-dideuteropyridine (**S2-d₂**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **S1** (35.9 mg, 0.18 mmol), B₂eg₂ (28.4 mg, 0.20 mmol), and MeOD (132 mg, 4.00 mmol), and using Cs₂CO₃ (65.2 mg, 0.20 mmol) as the base in 4 mL of anhydrous THF, 1,4-dideuteropyridine product **S2-d₂** was generated in 50% NMR yield (98% 4-D).

¹H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Note: This 1,4-DHP product was rather oxygen-labile, and attempts to obtain its pure form following general procedure GP-B failed. The NMR data were acquired using the No-D NMR technique from the above reaction mixture.

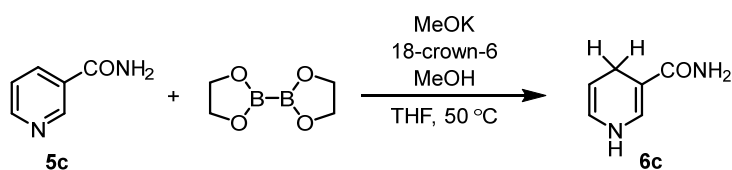
¹H NMR (400 MHz, THF-*d*₀): δ 7.16 (m, 5H), 5.87 (d, *J* = 7.7 Hz, 2H), 4.13 (d, *J* = 7.7 Hz, 2H), 2.57 (t, *J* = 7.8 Hz, 2H), 1.65 (m, 2H), 1.32 (m, 2H).

²H NMR (61 MHz, THF-*d*₀): δ 5.98, 2.98.

¹³C NMR (100 MHz, THF-*d*₀): δ 142.9, 128.1, 127.9, 126.5, 125.2, 99.6, 40.5, 36.2, 29.3 (t, *J* = 48.1 Hz), 27.3.

The formation of 1,4-DHP product **S2-d₂** from pyridine **S1** indicated that the present hydroboration process is compatible with pyridine substrate with electron-neutral substituents and is not limited to activated pyridines.

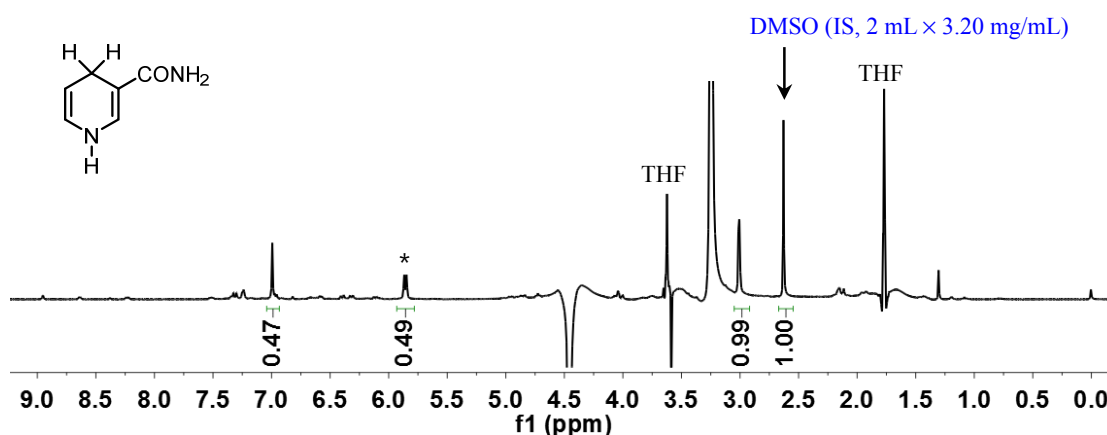
1,4-Dihydropyridine-3-carboxamide (**6c**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5c** (44.5 mg, 0.36 mmol), B₂eg₂ (56.8 mg, 0.40 mmol), and MeOH (128 mg, 4.00 mmol), and using

the combination of MeOK (28 mg, 0.40 mmol) and 18-crown-6 (106 mg, 0.40 mmol) as the base in 2 mL of anhydrous THF, 1,4-DHP product **6c** was generated in 67% NMR yield.

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



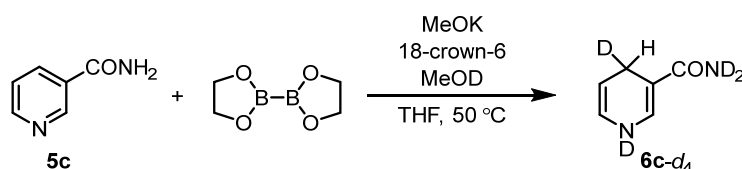
Note: This 1,4-DHP product was rather oxygen-labile, and attempts to obtain its pure form following general procedure GP-B failed. The NMR data were acquired using the No-D NMR technique from the above reaction mixture.

^1H NMR (400 MHz, THF- d_0): δ 6.96 (m, 1H), 6.08 (br s, 3H), 5.84 (m, 1H), 4.50 (dt, $J = 7.7, 3.5$ Hz, 1H), 3.08 (m, 2H).

^{13}C NMR (100 MHz, THF- d_0): δ 170.3, 135.1, 125.4, 100.2, 99.0, 22.7.

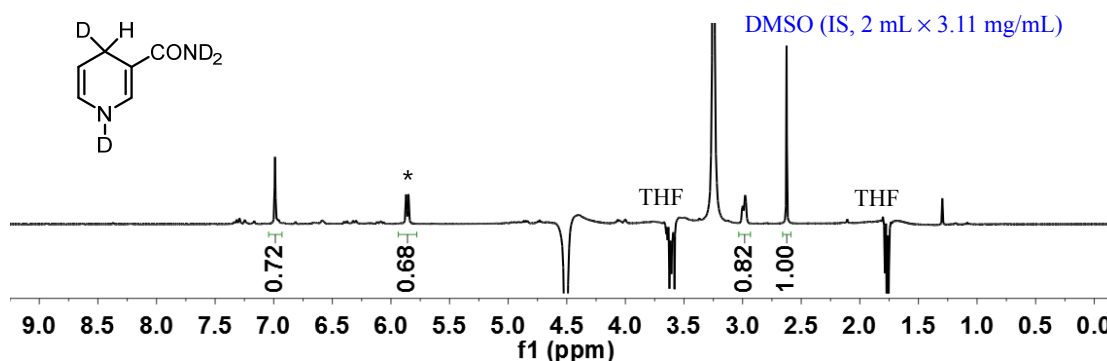
GC-MS: $\text{C}_6\text{H}_8\text{N}_2\text{O}$ [$m/z = 124, (\text{M}^+)$].

1,4-Dideuteropyridine-3-carboxamide (**6c-d₄**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5c** (45.0 mg, 0.36 mmol), B_2eg_2 (56.8 mg, 0.40 mmol), and MeOD (264 mg, 7.99 mmol), and using the combination of MeOK (28 mg, 0.40 mmol) and 18-crown-6 (106 mg, 0.40 mmol) as the base in 2 mL of anhydrous THF, 1,4-dideuteropyridine product **6c-d₄** was generated in 90% NMR yield (86% 4-D).

¹H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Note: This 1,4-DHP product was rather oxygen-labile, and attempts to obtain its pure form following general procedure GP-B failed. The NMR data were acquired using the No-D NMR technique from the above reaction mixture.

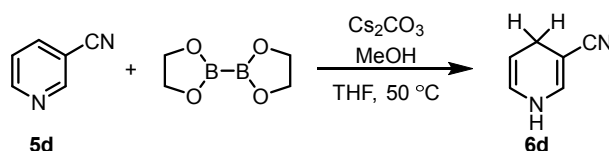
¹H NMR (400 MHz, THF-*d*₀): δ 7.01 (s, 1H), 5.85 (d, *J* = 7.9 Hz, 1H), 4.50 (dd, *J* = 7.9, 3.4 Hz, 1H), 3.06 (m, 1H).

²H NMR (61 MHz, THF-*d*₀): δ 7.15, 6.29, 3.06.

¹³C NMR (100 MHz, THF-*d*₀): δ 170.8, 135.5, 125.3, 100.1, 98.4, 22.2 (t, *J* = 19.8 Hz).

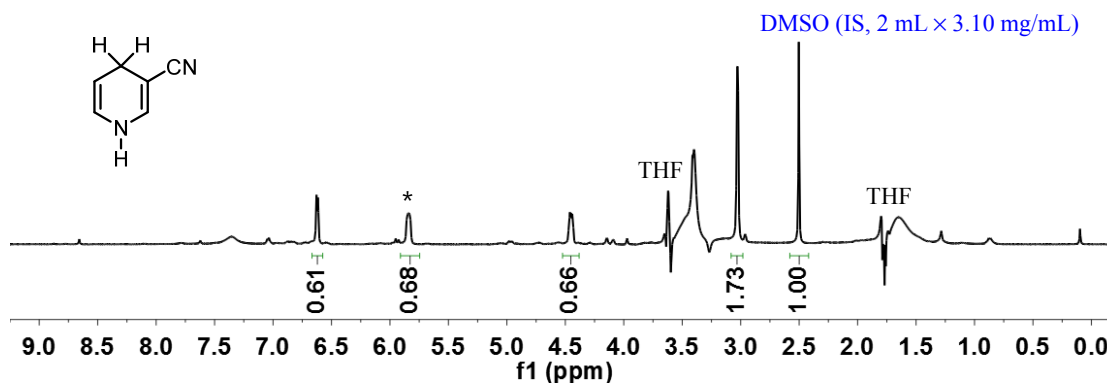
GC-MS: C₆H₄N₂OD₄ [*m/z* = 128, (M-*d*₄)⁺].

1,4-Dihydropyridine-3-carbonitrile (**6d**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5d** (36.0 mg, 0.36 mmol), B₂eg₂ (56.8 mg, 0.40 mmol), and MeOH (128 mg, 4.00 mmol), and using Cs₂CO₃ (131 mg, 0.40 mmol) as the base in 2 mL of anhydrous THF, 1,4-DHP product **6d** was generated in 90% NMR yield.

¹H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



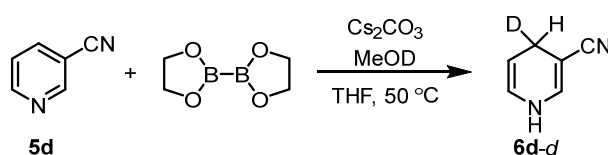
Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5d** (473 mg, 4.55 mmol), the 1,4-DHP product **6d** (321 mg, 67% yield) was obtained.

^1H NMR (400 MHz, CDCl_3): δ 6.61 (d, $J = 4.7$ Hz, 1H), 5.84 (m, 1H), 5.40 (br s, 1H), 4.60 (ddt, $J = 8.2, 1.6, 3.5$ Hz, 1H), 3.10 (dd, $J = 3.5, 1.6$ Hz, 2H).

^{13}C NMR (100 MHz, CDCl_3): δ 140.0, 124.3, 121.6, 100.7, 78.4, 23.4.

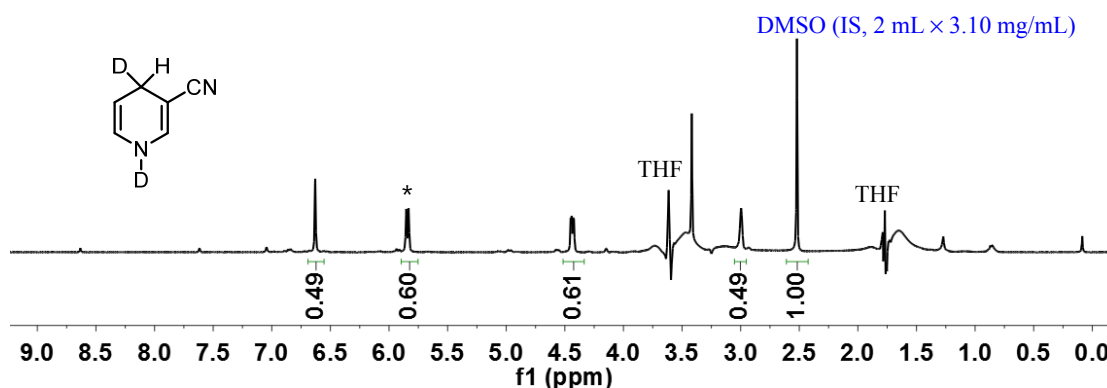
HRMS Calcd for $\text{C}_6\text{H}_6\text{N}_2$ ($\text{M}+\text{H}$) $^+$: 107.0609, Found: 107.0604.

4-Deutero-1-hydropyridine-3-carbonitrile (**6d-d**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5d** (36.0 mg, 0.36 mmol), B_2eg_2 (56.8 mg, 0.40 mmol), and MeOD (264 mg, 8.00 mmol), and using Cs_2CO_3 (131 mg, 0.40 mmol) as the base in 2 mL of anhydrous THF , 1,4-dideuteropyridine product **6d-d₂** was generated in 79% NMR yield.

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5d** (473 mg, 4.55 mmol), the 1,4-DHP product **6d-d** (300 mg, 62% yield, >98% 4-D) was obtained. **Note:** In the crude mixture **6d-d₂** was produced, which became **6d-d** during purification due to H/D exchange at the nitrogen atom.

^1H NMR (400 MHz, CDCl_3): δ 6.61 (dt, $J = 5.6, 0.9$ Hz, 1H), 5.84 (ddt, $J = 8.1, 4.4, 6.5$ Hz, 1H), 5.31 (br s, 1H), 4.60 (ddd, $J = 8.1, 3.4, 1.5$ Hz, 1H), 3.08 (m, 1H).

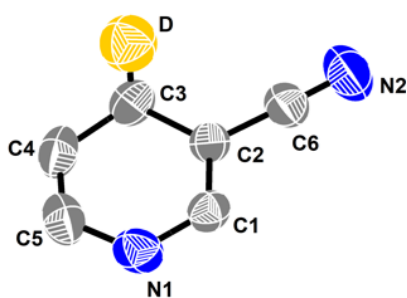
^2H NMR (61 MHz, CDCl_3): δ 3.08.

^{13}C NMR (100 MHz, CDCl_3): δ 140.2, 124.3, 121.6, 100.7, 78.6, 23.0 (t, $J = 20.7$ Hz).

HRMS Calcd for $\text{C}_6\text{H}_5\text{N}_2\text{D}$ ($\text{M}+\text{H}$) $^+$: 108.0672, Found: 108.0670.

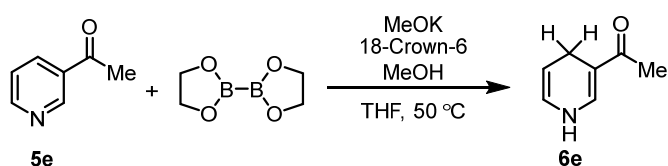
Sample for X-Ray single crystal diffraction analysis was prepared by recrystallization in $\text{CH}_2\text{Cl}_2/n$ -hexane.

Data for XRD analysis of 6d-d (CCDC1935576)



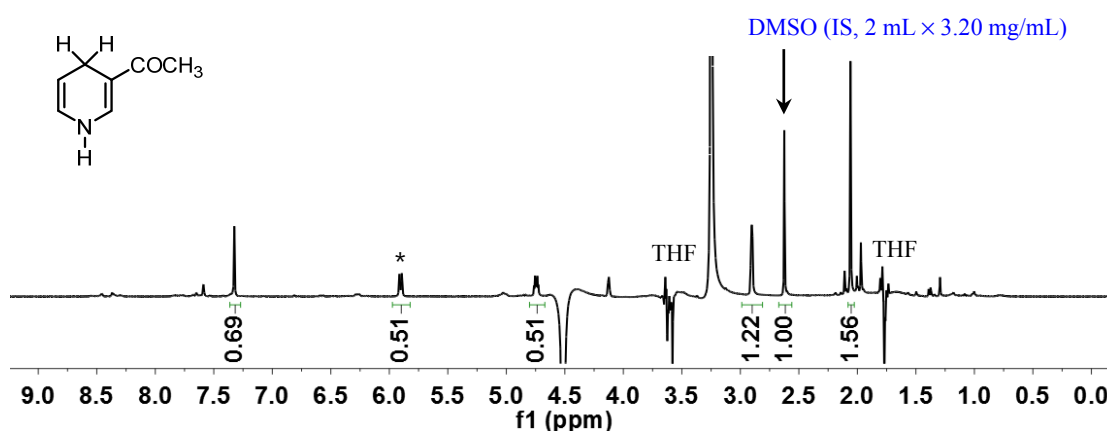
Empirical formula	C ₆ H ₅ DN ₂
Formula weight	107.13
Temperature/K	293(2)
Crystal system	orthorhombic
Space group	Pbca
a/Å	6.0431(2)
b/Å	13.3857(4)
c/Å	14.3592(4)
α/°	90
β/°	90
γ/°	90
Volume/Å ³	1161.53(6)
Z	8
ρ _{calc} /cm ³	1.225
μ/mm ⁻¹	0.613
F(000)	448.0
Crystal size/mm ³	0.3 × 0.1 × 0.1
Radiation	CuKα (λ = 1.54184)
2θ range for data collection/°	12.328 to 153.394
Index ranges	-7 ≤ h ≤ 5, -16 ≤ k ≤ 16, -17 ≤ l ≤ 12
Reflections collected	2497
Independent reflections	1181 [R _{int} = 0.0169, R _{sigma} = 0.0209]
Data/restraints/parameters	1181/0/81
Goodness-of-fit on F ²	1.099
Final R indexes [I ≥ 2σ (I)]	R ₁ = 0.0667, wR ₂ = 0.1497
Final R indexes [all data]	R ₁ = 0.0746, wR ₂ = 0.1598
Largest diff. peak/hole / e Å ⁻³	0.36/-0.56

3-Acetyl-1,4-dihydropyridine (6e)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5e** (45.0 mg, 0.36 mmol), B₂eg₂ (56.8 mg, 0.40 mmol), and MeOH (128 mg, 4.00 mmol), and using the combination of MeOK (28.0 mg, 0.40 mmol) and 18-crown-6 (106 mg, 0.40 mmol) as the base in 2 mL of anhydrous THF, 1,4-DHP product **6e** was generated in 70% NMR yield.

¹H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



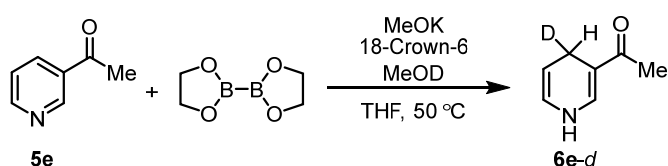
Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5e** (605 mg, 5.00 mmol), the 1,4-DHP product **6e** (259 mg, 42% yield) was obtained.

¹H NMR (400 MHz, CDCl₃): δ 7.10 (d, *J* = 5.8 Hz, 1H), 5.98 (br s, 1H), 5.88 (m, 1H), 4.83 (ddt, *J* = 6.7, 3.6, 1.8 Hz, 1H), 3.07 (dd, *J* = 3.6, 1.8 Hz, 2H), 2.15 (s, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 195.9, 140.0, 123.7, 109.5, 105.3, 24.1, 21.7.

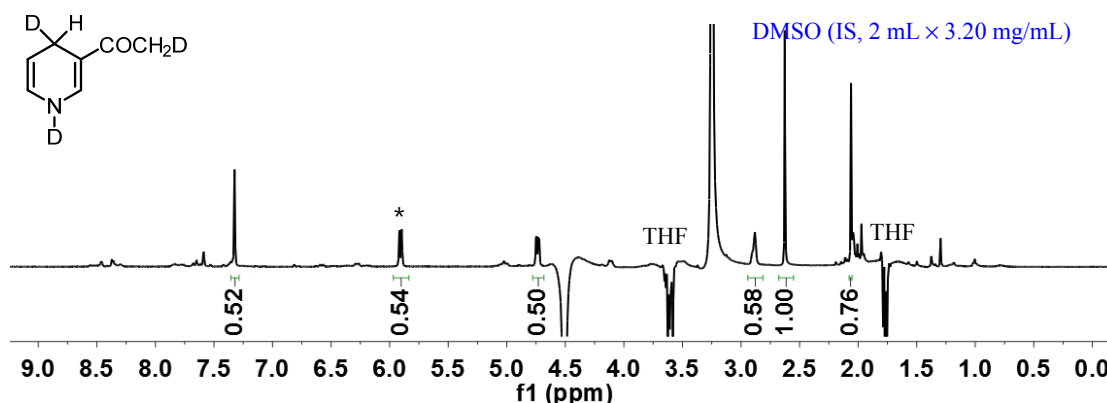
HRMS Calcd for C₇H₉NO (M+H)⁺: 124.0762, Found: 124.0754.

3-Acetyl-4-deutero-1-hydropyridine (6e-d)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5e** (45.0 mg, 0.36 mmol), B₂eg₂ (56.8 mg, 0.40 mmol), and MeOD (132 mg, 4.00 mmol), and using the combination of MeOK (28.0 mg, 0.40 mmol) and 18-crown-6 (106 mg, 0.40 mmol) as the base in 2 mL of anhydrous THF, 1,4-dideuteropyridine product **6e-d₂** was generated in 74% NMR yield.

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5e** (44.0 mg, 0.36 mmol), the 1,4-DHP product **6e-d** (20.8 mg, 46% yield) was obtained. **Note:** In the crude mixture **6e-d₂** was produced, which became **6e-d** during purification due to H/D exchange at the nitrogen atom. Both ^1H and ^2H NMR analysis also showed a minor extent of deuterium incorporation (20% D) at the methyl group.

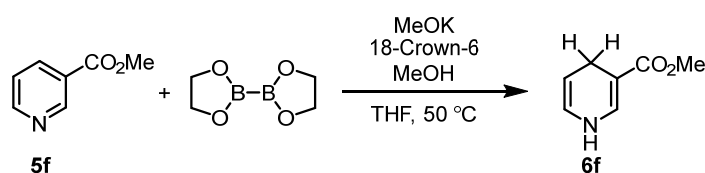
^1H NMR (400 MHz, CDCl_3): δ 7.09 (d, J = 5.4 Hz, 1H), 5.89 (dd, J = 8.3, 3.8 Hz, 1H), 5.79 (br s, 1H), 4.83 (m, 1H), 3.05 (t, J = 2.6 Hz, 1H), 2.15 (s, 3H).

^2H NMR (61 MHz, CDCl_3): δ 3.07, 2.16 (t, J = 1.1 Hz).

^{13}C NMR (100 MHz, CDCl_3): δ 195.8, 139.8, 123.7, 109.6, 105.2, 24.1, 21.3 (t, J = 20.2 Hz).

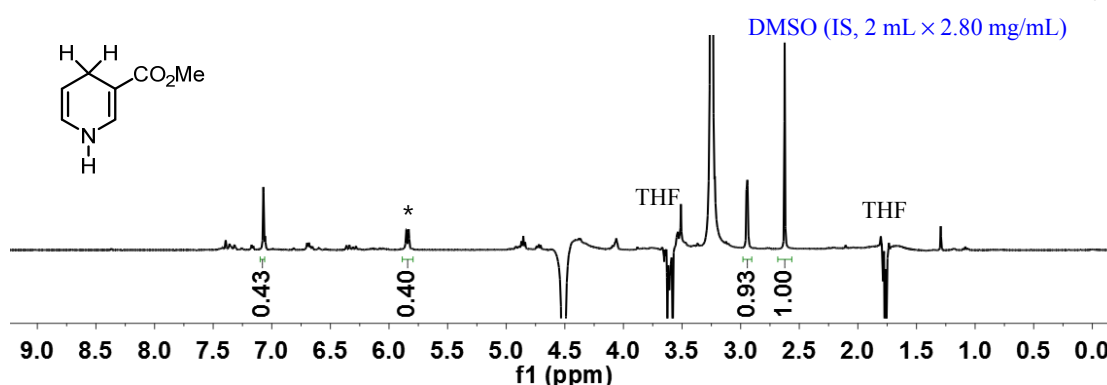
HRMS Calcd for $\text{C}_7\text{H}_8\text{NOD}$ ($\text{M}+\text{H}$) $^+$: 125.0825, Found: 125.0815.

Methyl 1,4-dihydropyridine-3-carboxylate (**6f**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5f** (50.0 mg, 0.36 mmol), B_2eg_2 (56.8 mg, 0.40 mmol), and MeOH (128 mg, 4.00 mmol), and using the combination of MeOK (28.0 mg, 0.40 mmol) and 18-crown-6 (106 mg, 0.40 mmol) as the base in 2 mL of anhydrous THF, 1,4-DHP product **6f** was generated in 48% NMR yield.

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



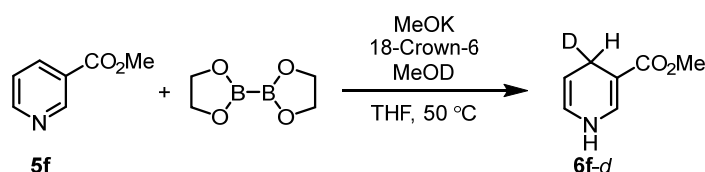
Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5f** (625 mg, 4.56 mmol), the 1,4-DHP product **6f** (260 mg, 41% yield) was obtained.

^1H NMR (400 MHz, CDCl_3): δ 7.12 (d, J = 5.6 Hz, 1H), 5.86 (m, 1H), 5.27 (br s, 1H), 4.72 (m, 1H), 3.68 (s, 3H), 3.11 (dd, J = 3.5, 1.7 Hz, 2H).

^{13}C NMR (100 MHz, CDCl_3): δ 168.9, 137.9, 124.2, 103.5, 98.2, 51.1, 22.2.

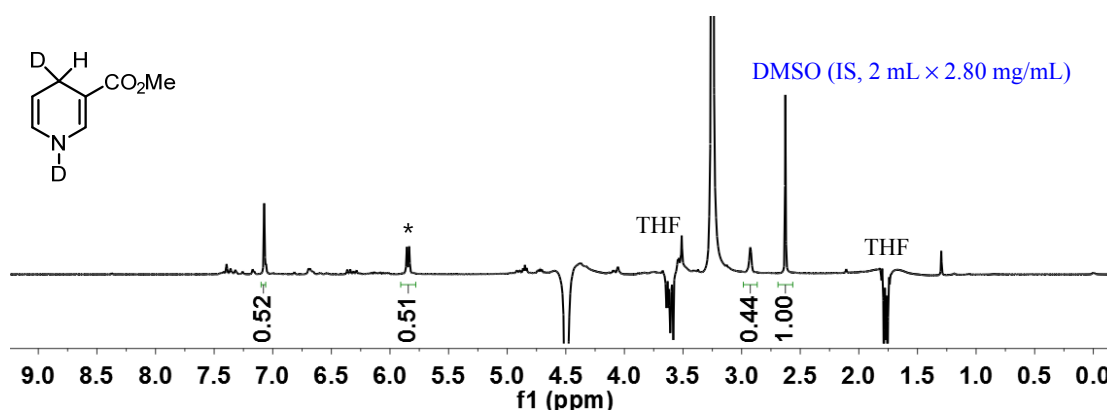
HRMS Calcd for $\text{C}_7\text{H}_9\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 140.0712, Found: 140.0702.

Methyl 4-deutero-1-hydropyridine-3-carboxylate (**6f-d**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5f** (50.0 mg, 0.36 mmol), B_2eg_2 (56.8 mg, 0.40 mmol), and MeOD (132 mg, 4.00 mmol), and using the combination of MeOK (28.0 mg, 0.40 mmol) and 18-crown-6 (106 mg, 0.40 mmol) as the base in 2 mL of anhydrous THF, 1,4-DHP product **6f-d₂** was generated in 61% NMR yield.

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5f** (625 mg, 4.56 mmol), the 1,4-DHP product **6f-d** (250 mg, 39% yield) was obtained. **Note:** In the crude mixture **6f-d₂** was produced, which became **6f-d** during purification due to H/D exchange at the nitrogen atom.

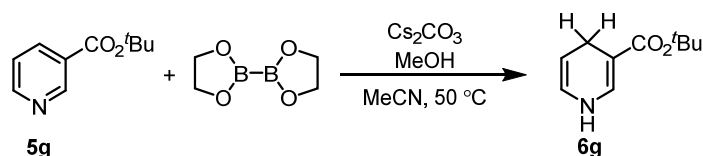
¹H NMR (400 MHz, CDCl₃): δ 7.13 (d, *J* = 5.6 Hz, 1H), 5.86 (m, 1H), 5.27 (br s, 1H), 4.71 (ddd, *J* = 8.1, 3.6, 1.6 Hz, 1H), 3.68 (s, 3H), 3.09 (m, 1H).

²H NMR (61 MHz, CDCl₃): δ 3.10

¹³C NMR (100 MHz, CDCl₃): δ 168.9, 137.9, 124.2, 103.5, 98.2, 51.1, 21.8 (t, *J* = 20.2 Hz).

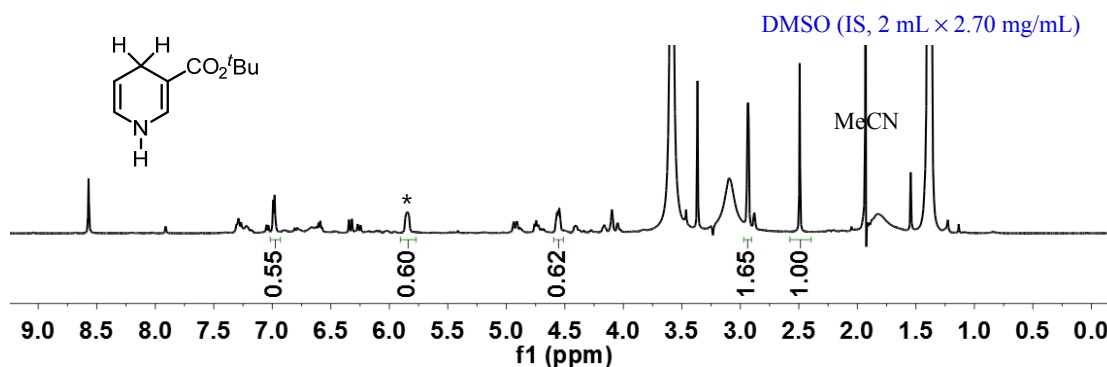
HRMS Calcd for C₇H₈NO₂D (M+H)⁺: 141.0774, Found: 141.0768.

tert-Butyl 1,4-dihydropyridine-3-carboxylate (6g)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5g** (65.0 mg, 0.36 mmol), B₂eg₂ (56.8 mg, 0.40 mmol), and MeOH (128 mg, 4.00 mmol), and using Cs₂CO₃ (131 mg, 0.40 mmol) as the base in 2 mL of anhydrous MeCN, 1,4-DHP product **6g** was generated in 69% NMR yield.

¹H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



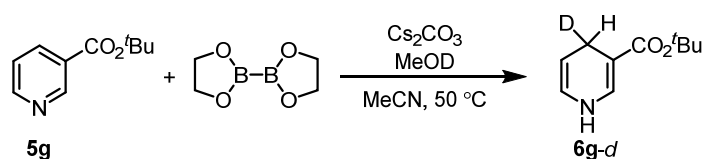
Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5g** (65.0 mg, 0.36 mmol), the 1,4-DHP product **6g** (33.2 mg, 51% yield) was obtained.

¹H NMR (400 MHz, CDCl₃): δ 7.04 (d, *J* = 5.5 Hz, 1H), 5.86 (m, 1H), 5.19 (br s, 1H), 4.70 (m, 1H), 3.06 (dd, *J* = 3.5, 1.7 Hz, 2H), 1.46 (s, 9H).

¹³C NMR (100 MHz, CDCl₃): δ 168.0, 137.1, 124.3, 103.2, 100.0, 28.5, 22.3.

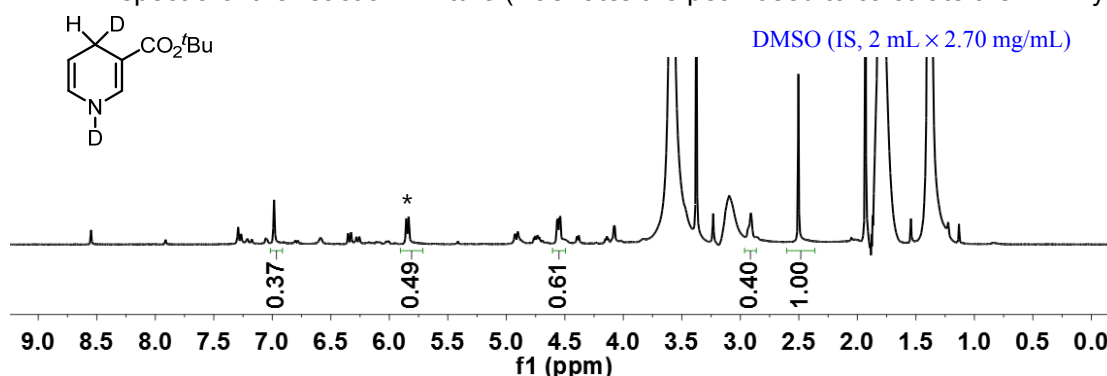
HRMS Calcd for C₁₀H₁₅NO₂ (M+H)⁺: 182.1181, Found: 182.1174.

tert-Butyl 4-Deutero-1-hydropyridine-3-carboxylate (**6g-d**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5g** (65.0 mg, 0.36 mmol), B₂eg₂ (56.8 mg, 0.40 mmol), and MeOD (264 mg, 8.00 mmol), and using Cs₂CO₃ (131 mg, 0.40 mmol) as the base in 2 mL of anhydrous MeCN, 1,4-dihydrodeuteropyridine product **6g-d₂** was generated in 57% NMR yield.

¹H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5g** (65.0 mg, 0.36 mmol), the 1,4-DHP product **6g-d** (25.5 mg, 39% yield) was obtained. **Note:** In the crude mixture **6g-d₂** was produced, which became **6g-d** during purification due to H/D exchange at the nitrogen atom.

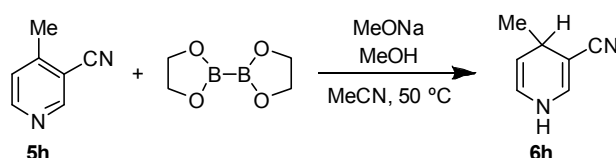
¹H NMR (400 MHz, CDCl₃): δ 7.04 (d, *J* = 5.5 Hz, 1H), 5.85 (m, 1H), 5.11 (br s, 1H), 4.68 (ddd, *J* = 8.0, 3.5, 1.1 Hz, 1H), 3.04 (m, 1H), 1.46 (s, 9H).

²H NMR (61 MHz, CDCl₃): δ 3.05

¹³C NMR (100 MHz, CDCl₃): δ 168.0, 136.9, 124.3, 103.1, 99.9, 78.9, 28.5, 21.9 (t, *J* = 20.6 Hz).

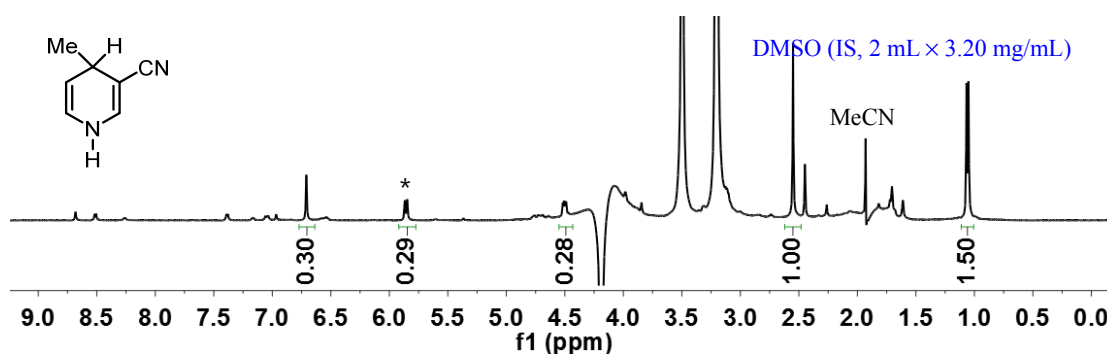
HRMS Calcd for C₁₀H₁₄NO₂D (M+H)⁺: 183.1244, Found: 183.1235.

3-Cyano-4-methyl-1,4-dihydropyridine (**6h**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5h** (43.0 mg, 0.36 mmol), B₂eg₂ (56.8 mg, 0.40 mmol), and MeOH (128 mg, 4.00 mmol), and using MeONa (21.6 mg, 0.40 mmol) as the base in 2 mL of anhydrous MeCN, 1,4-dihydrodeuteropyridine product **6h** was generated in 40% NMR yield.

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



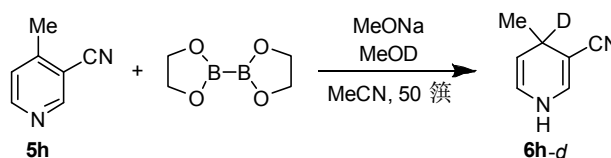
Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5h** (540 mg, 4.57 mmol), the 1,4-DHP product **6h** (273 mg, 50% yield) was obtained.

^1H NMR (400 MHz, CDCl_3): δ 6.63 (dd, $J = 5.6, 1.2$ Hz, 1H), 5.87 (dd, $J = 8.0, 4.4$ Hz, 1H), 5.58 (br s, 1H), 4.59 (ddd, $J = 8.1, 3.8, 1.5$ Hz, 1H), 3.25 (ddd, $J = 6.7, 3.8, 1.1$ Hz, 1H), 1.23 (d, $J = 6.7$ Hz, 3H).

^{13}C NMR (100 MHz, CDCl_3): δ 138.8, 122.8, 121.4, 106.7, 84.6, 28.3, 24.8.

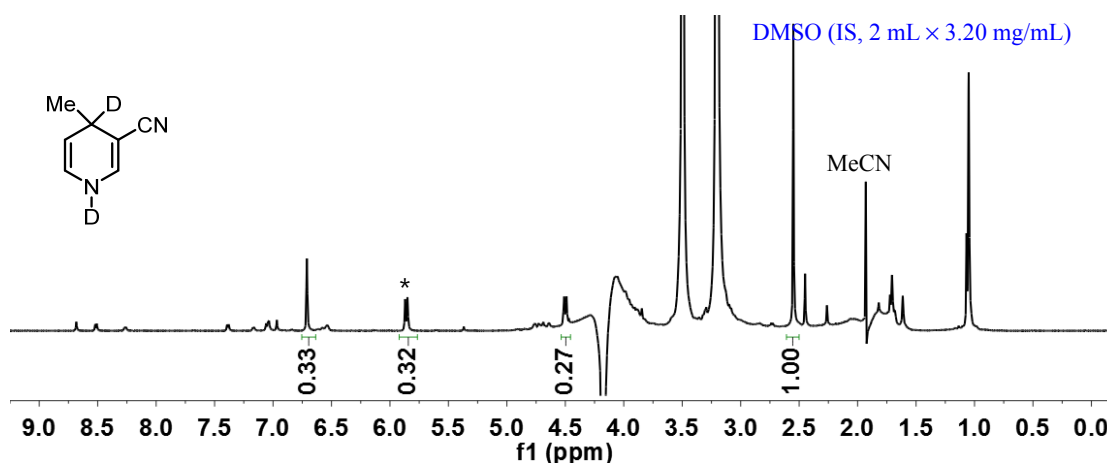
HRMS Calcd for $\text{C}_7\text{H}_8\text{N}$ ($\text{M}+\text{H}$) $^+$: 121.0766, Found: 121.0762.

3-Cyano-4-methyl-4-deutero-1-hydropyridine (**6h-d**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5h** (43.0 mg, 0.36 mmol), B_2eg_2 (56.8 mg, 0.40 mmol), and MeOD (132 mg, 4.00 mmol), and using MeONa (21.6 mg, 0.40 mmol) as the base in 2 mL of anhydrous MeCN, 1,4-dihydrodeuteropyridine product **6h-d₂** was generated in 44% NMR yield.

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5h** (540 mg, 4.57 mmol), the 1,4-DHP product **6h-d** (291 mg, 53% yield, 93% 4-D) was obtained. **Note:** In the crude mixture **6h-d₂** was produced, which became **6h-d** during purification due to H/D exchange at the nitrogen atom.

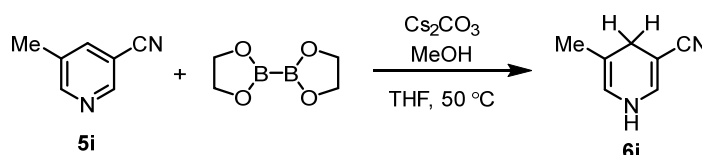
^1H NMR (400 MHz, CDCl_3): δ 6.64 (dd, $J = 5.6, 1.2$ Hz, 1H), 5.88 (ddd, $J = 8.0, 4.4, 1.3$ Hz, 1H), 5.50 (br s, 1H), 4.60 (dd, $J = 8.1, 1.5$ Hz, 1H), 1.23 (s, 3H).

^2H NMR (61 MHz, CDCl_3): δ 3.22

^{13}C NMR (100 MHz, CDCl_3): δ 138.7, 122.8, 121.3, 106.6, 84.7, 27.9 (t, $J = 20.4$ Hz), 24.7.

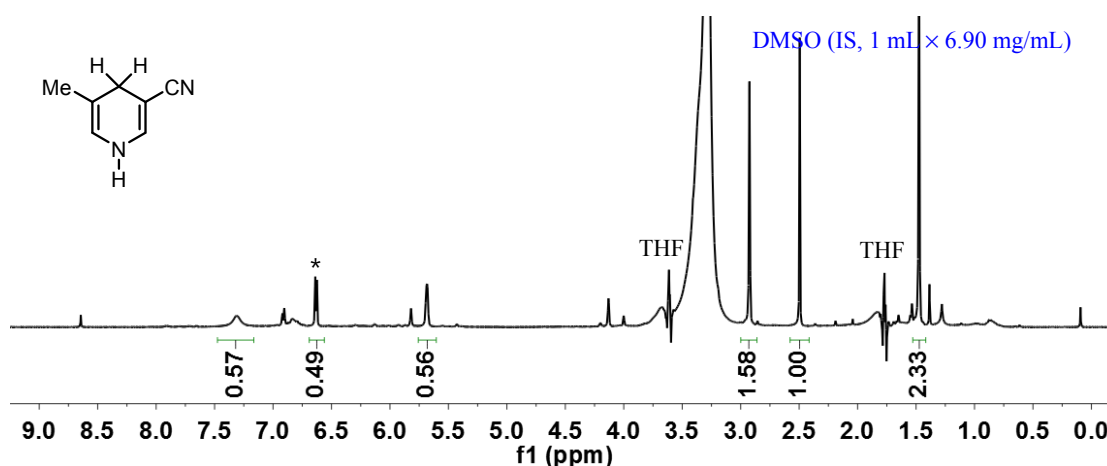
HRMS Calcd for $\text{C}_7\text{H}_7\text{ND}$ ($\text{M}+\text{H}^+$): 122.0828, Found: 122.0823.

3-Cyano-5-methyl-1,4-dihydropyridine (**6i**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5i** (43.0 mg, 0.36 mmol), B_2eg_2 (56.8 mg, 0.40 mmol), and MeOH (128 mg, 4.00 mmol), and using Cs_2CO_3 (131 mg, 0.40 mmol) as the base in 2 mL of anhydrous THF, 1,4-DHP product **6i** was generated in 72% NMR yield.

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



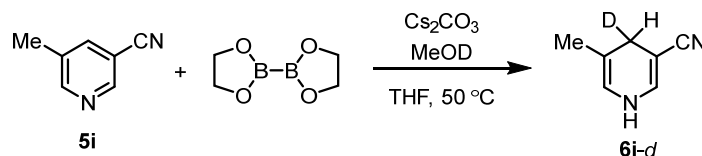
Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5i** (43.0 mg, 0.36 mmol), the 1,4-DHP product **6i** (20.6 mg, 47% yield) was obtained.

^1H NMR (400 MHz, CDCl_3): δ 6.62 (d, J = 5.6 Hz, 1H), 5.66 (d, J = 2.0 Hz, 1H), 5.50 (br s, 1H), 2.97 (s, 2H), 1.53 (m, 3H).

^{13}C NMR (100 MHz, CDCl_3): δ 139.5, 122.1, 118.6, 109.9, 76.9, 28.8, 20.1.

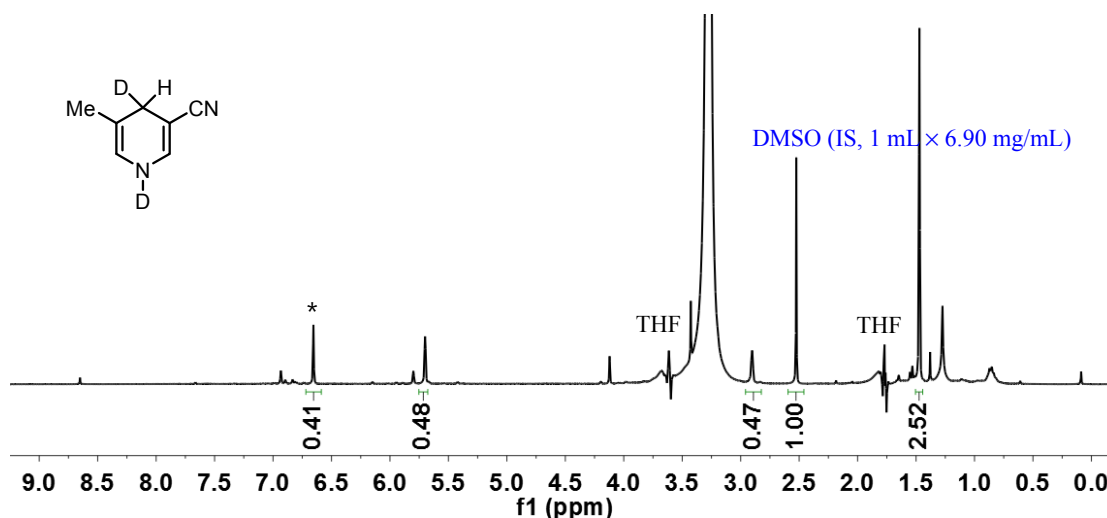
HRMS Calcd for $\text{C}_7\text{H}_8\text{N}$ ($\text{M}+\text{H}$) $^+$: 121.0766, Found: 121.0762.

3-Cyano-5-methyl-4-deutero-1hydropyridine (**6i-d**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5i** (43.0 mg, 0.36 mmol), B_2eg_2 (56.8 mg, 0.40 mmol), and MeOD (264 mg, 8.00 mmol), and using Cs_2CO_3 (131 mg, 0.40 mmol) as the base in 2 mL of anhydrous THF, 1,4-dideuteropyridine product **6i-d₂** was generated in 60% NMR yield.

¹H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5i** (43.0 mg, 0.36 mmol), the 1,4-DHP product **6i-d** (26.9 mg, 61% yield, >98% D) was obtained (in this experiment THF was used as the solvent in place of MeCN). **Note:** In the crude mixture **6i-d₂** was produced, which became **6i-d** during purification due to H/D exchange at the nitrogen atom.

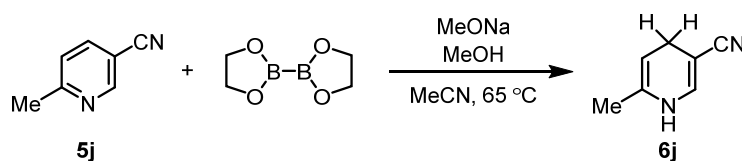
¹H NMR (400 MHz, CDCl₃): δ 6.64 (m, 1H), 5.68 (dq, *J* = 4.4, 1.4 Hz, 1H), 5.63 (br s, 1H), 2.97 (m, 1H), 1.53 (s, 3H).

²H NMR (61 MHz, CDCl₃): δ 2.95

¹³C NMR (100 MHz, CDCl₃): δ 139.7, 122.2, 118.6, 109.7, 76.8, 28.3 (t, *J* = 20.6 Hz), 20.1.

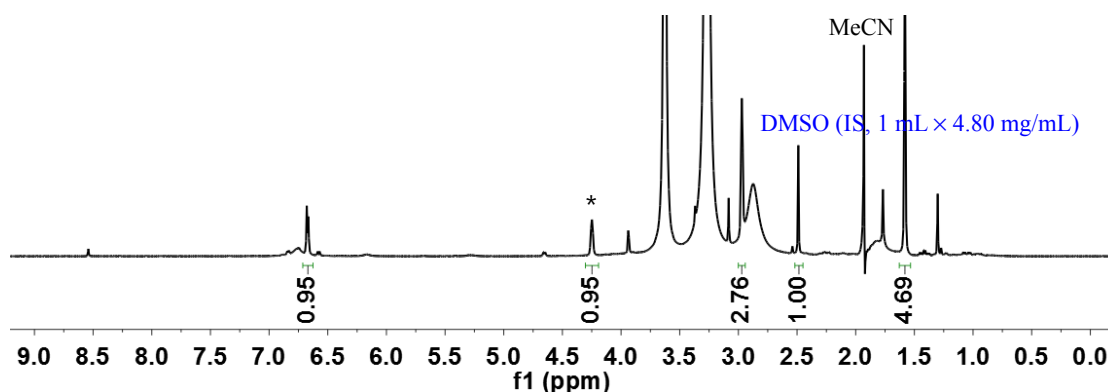
HRMS Calcd for C₇H₇ND (M+H)⁺: 122.0828, Found: 122.0814.

2-Methyl-5-cyano-1,4-dihydropyridine (**6j**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5j** (43.0 mg, 0.36 mmol), B₂eg₂ (56.8 mg, 0.40 mmol), and MeOH (128 mg, 4.00 mmol), and using MeONa (21.6 mg, 0.40 mmol) as the base in 2 mL of anhydrous MeCN, 1,4-DHP product **6j** was generated in 97% NMR yield.

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



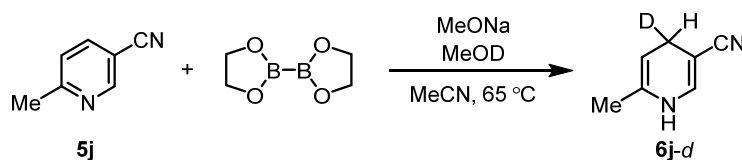
Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5j** (43.0 mg, 0.36 mmol), the 1,4-DHP product **6j** (27.1 mg, 62% yield) was obtained.

^1H NMR (400 MHz, CDCl_3): δ 6.62 (d, J = 5.7 Hz, 1H), 5.11 (br s, 1H), 4.34 (dt, J = 3.3, 1.7 Hz, 1H), 3.06 (m, 2H), 1.66 (m, 3H).

^{13}C NMR (100 MHz, CDCl_3): δ 140.3, 131.6, 121.6, 96.8, 78.5, 24.2, 19.2.

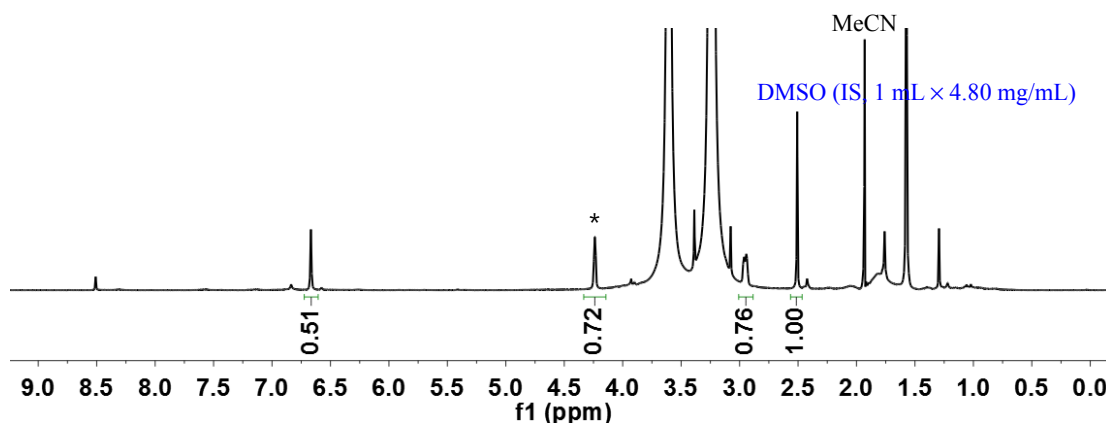
HRMS Calcd for $\text{C}_7\text{H}_8\text{N}$ ($\text{M}+\text{H}$) $^+$: 121.0766, Found: 121.0762.

2-Methyl-5-cyano-4-deutero-1-hydropyridine (**6j-d**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5j** (43.0 mg, 0.36 mmol), B_2eg_2 (56.8 mg, 0.40 mmol), and MeOD (264 mg, 8.00 mmol), and using MeONa (21.6 mg, 0.40 mmol) as the base in 2 mL of anhydrous MeCN, 1,4-dideuteropyridine product **6j-d₂** was generated in 74% NMR yield.

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5j** (43.0 mg, 0.36 mmol), the 1,4-DHP product **6j-d** (21.1 mg, 48% yield, 86% 4-D) was obtained. **Note:** In the crude mixture **6j-d₂** was produced, which became **6j-d** during purification due to H/D exchange at the nitrogen atom.

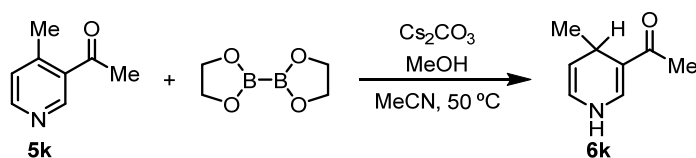
¹H NMR (400 MHz, CDCl₃): δ 6.62 (d, *J* = 5.6 Hz, 1H), 5.11 (br s, 1H), 4.34 (dt, *J* = 3.3, 1.7 Hz, 1H), 3.06 (m, 1.06H), 1.67 (t, *J* = 1.4 Hz, 3H).

²H NMR (61 MHz, CDCl₃): δ 3.04.

¹³C NMR (100 MHz, CDCl₃): δ 140.3, 131.6, 121.7, 96.7, 78.5, 23.7 (t, *J* = 20.8 Hz), 19.2.

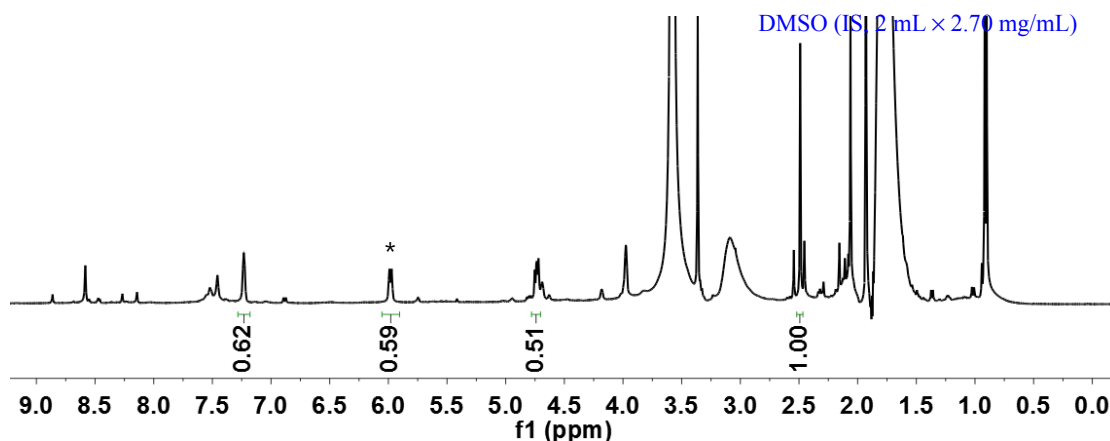
HRMS Calcd for C₇H₇ND (M+H)⁺: 122.0828, Found: 122.0823.

3-Acetyl-4-methyl-1,4-dihydropyridine (**6k**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5k** (49.2 mg, 0.36 mmol), B₂eg₂ (56.8 mg, 0.40 mmol), and MeOH (128 mg, 4.00 mmol), and using Cs₂CO₃ (131 mg, 0.40 mmol) as the base in 2 mL of anhydrous MeCN, 1,4-DHP product **6k** was generated in 68% NMR yield.

¹H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



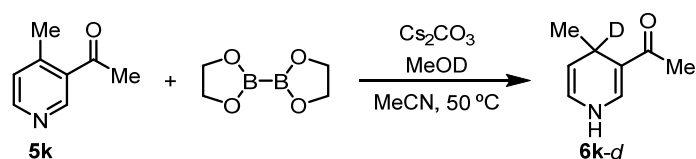
Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5k** (49.2 mg, 0.36 mmol), the 1,4-DHP product **6k** (16.8 mg, 34% yield) was obtained.

¹H NMR (400 MHz, CDCl₃): δ 7.15 (d, *J* = 5.8 Hz, 1H), 6.10 (br s, 1H), 5.96 (m, 1H), 4.85 (ddd, *J* = 7.7, 5.1, 1.1 Hz, 1H), 3.48 (dd, *J* = 6.7, 5.5 Hz, 2H), 2.17 (s, 3H), 1.04 (d, *J* = 6.5 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 195.9, 139.0, 122.5, 115.2, 110.3, 26.1, 24.6, 24.5.

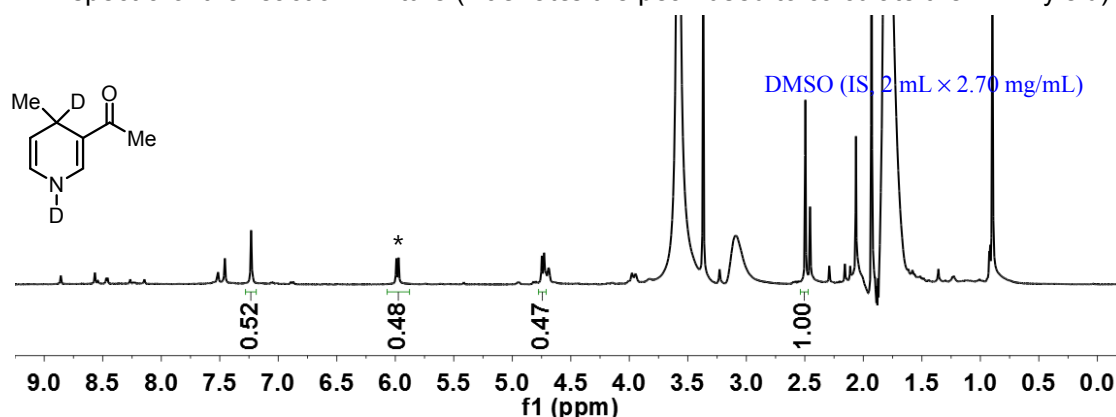
HRMS Calcd for C₈H₁₁NO (M+H)⁺: 138.0919, Found: 138.0913.

3-Acetyl-4-methyl-4-deutero-1-hydropyridine (**6k-d**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5k** (49.2 mg, 0.36 mmol), B_{2}eg_2 (56.8 mg, 0.40 mmol), and MeOD (264 mg, 8.00 mmol), and using Cs_2CO_3 (131 mg, 0.40 mmol) as the base in 2 mL of anhydrous MeCN , 1,4-dideuteropyridine product **6k-d₂** was generated in 55% NMR yield.

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5k** (49.2 mg, 0.36 mmol), the 1,4-DHP product **6k-d** (21.0 mg, 42% yield, 94% 4-D) was obtained. **Note:** In the crude mixture **6k-d₂** was produced, which became **6k-d** during purification due to H/D exchange at the nitrogen atom. Both ^1H and ^2H NMR analysis also showed a minor extent of deuterium incorporation (9% D) at the methyl group.

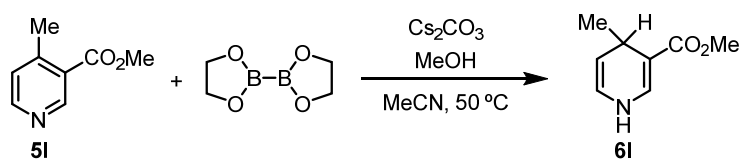
^1H NMR (400 MHz, CDCl_3): δ 7.17 (dd, $J = 5.8, 1.1$ Hz, 1H), 5.99 (ddd, $J = 7.7, 4.3, 1.1$ Hz, 1H), 5.90 (br s, 1H), 4.86 (dd, $J = 7.6, 1.7$ Hz 1H), 2.19 (s, 2.74H), 1.04 (s, 3H).

^2H NMR (61 MHz, CDCl_3): δ 3.48, 2.18 (t, $J = 0.73$ Hz)

^{13}C NMR (100 MHz, CDCl_3): δ 195.9, 138.8, 122.5, 115.2, 110.2, 25.7 (t, $J = 21.3$ Hz), 24.6, 24.3.

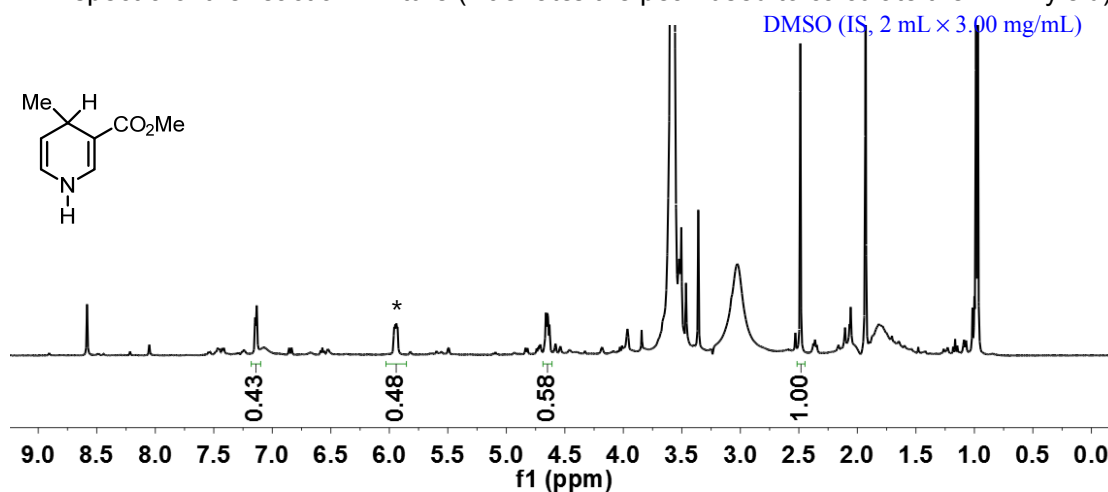
HRMS Calcd for $\text{C}_8\text{H}_{10}\text{NOD}$ ($\text{M}+\text{H}^+$): 139.0982, Found: 139.0975.

Methyl 4-methyl-1,4-dihydropyridine-3-carboxylate (**6I**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5I** (55.0 mg, 0.36 mmol), B_2eg_2 (56.8 mg, 0.40 mmol), and MeOH (128 mg, 4.00 mmol), and using Cs_2CO_3 (131 mg, 0.40 mmol) as the base in 2 mL of anhydrous MeCN, 1,4-DHP product **6I** was generated in 62% NMR yield.

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



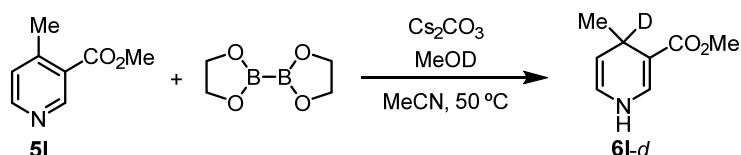
Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5I** (412 mg, 2.73 mmol), the 1,4-DHP product **6I** (154 mg, 37% yield) was obtained.

^1H NMR (400 MHz, CDCl_3): δ 7.19 (dd, $J = 5.7, 1.2$ Hz, 1H), 5.91 (ddd, $J = 7.8, 4.3, 1.2$ Hz, 1H), 5.74 (br s, 1H), 4.74 (ddd, $J = 7.8, 4.8, 1.7$ Hz, 1H), 3.67 (s, 3H), 3.34 (m, 2H), 1.08 (d, $J = 6.5$ Hz, 3H).

^{13}C NMR (100 MHz, CDCl_3): δ 169.1, 137.1, 122.9, 108.7, 103.4, 50.9, 26.9, 25.1.

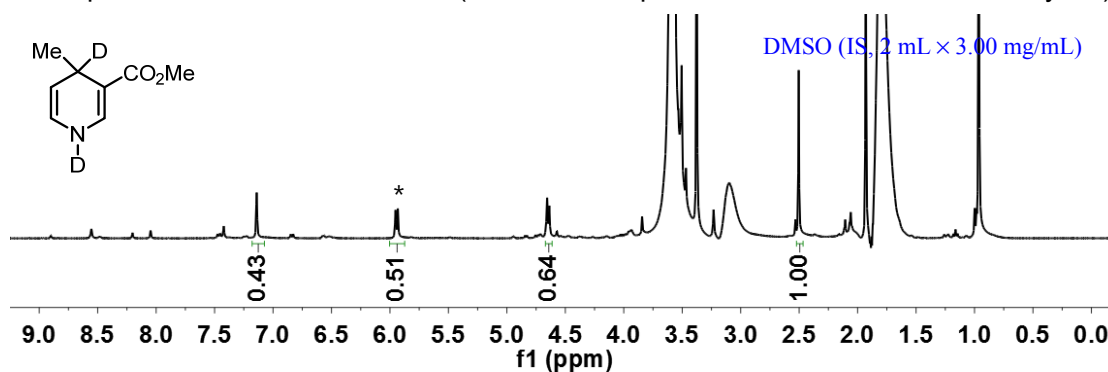
HRMS Calcd for $\text{C}_8\text{H}_{11}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 154.0868, Found: 154.0860.

Methyl 4-methyl-4-deutero-1-hydropyridine-3-carboxylate (**6I-d**)



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5I** (55.0 mg, 0.36 mmol), B_2eg_2 (56.8 mg, 0.40 mmol), and MeOD (264 mg, 8.00 mmol), and using Cs_2CO_3 (131 mg, 0.40 mmol) as the base in 2 mL of anhydrous MeCN, 1,4-dideuteropyridine product **6I-d₂** was generated in 65% NMR yield.

¹H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5l** (412 mg, 2.73 mmol), the 1,4-DHP product **6l-d** (151 mg, 36% yield, 94% 4-D) was obtained. **Note:** In the crude mixture **6l-d₂** was produced, which became **6l-d** during purification due to H/D exchange at the nitrogen atom.

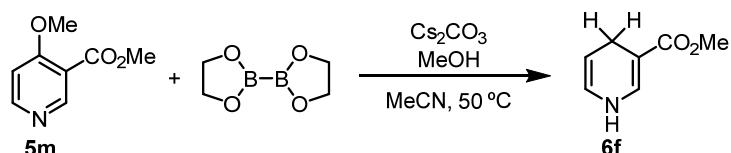
¹H NMR (400 MHz, CDCl₃): δ 7.22 (dd, *J* = 5.7, 1.2 Hz, 1H), 5.95 (ddd, *J* = 7.8, 4.3, 1.2 Hz, 1H), 5.55 (br s, 1H), 4.77 (dd, *J* = 7.7, 1.6 Hz, 1H), 3.69 (s, 3H), 1.09 (s, 3H).

²H NMR (61 MHz, CDCl₃): δ 3.34.

¹³C NMR (100 MHz, CDCl₃): δ 168.9, 136.9, 122.9, 108.7, 103.7, 51.0, 26.6 (t, *J* = 20.7 Hz), 25.0.

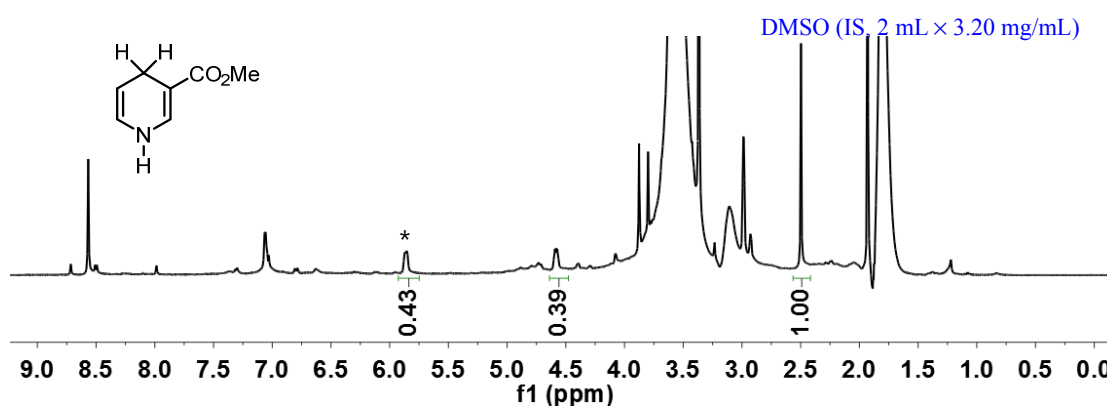
HRMS Calcd for C₈H₁₀NO₂D (M+H)⁺: 155.0931, Found: 155.0922.

1,4-DHP **6f** from double hydroboration reaction of pyridine **5m**



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5m** (66.8 mg, 0.36 mmol), B₂eg₂ (114 mg, 0.80 mmol), and MeOH (264 mg, 8.25 mmol), and using Cs₂CO₃ (261 mg, 0.80 mmol) as the base in 2 mL of anhydrous MeCN, 1,4-DHP product **6f** was generated in 59% NMR yield.

¹H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



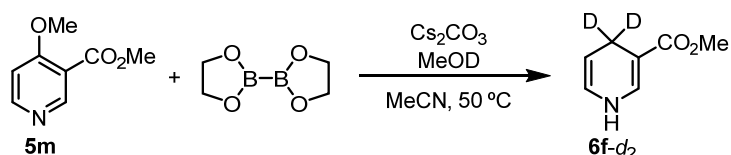
Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5m** (49.8 mg, 0.30 mmol), B₂eg₂ (78.6 mg, 0.60 mmol), and Cs₂CO₃ (196 mg, 0.60 mmol), the 1,4-DHP product **6f** (12.6 mg, 30% yield) was obtained.

¹H NMR (400MHz, CDCl₃): δ 7.05 (dd, *J* = 5.6 Hz, 1H), 5.79 (m, 1H), 5.26 (br s, 1H), 4.64 (m, 1H), 3.61 (s, 3H), 3.04 (dd, *J* = 3.5, 1.7 Hz, 2H).

¹³C NMR (100 MHz, CDCl₃): δ 169.0, 137.9, 124.2, 103.5, 98.1, 51.1, 22.2.

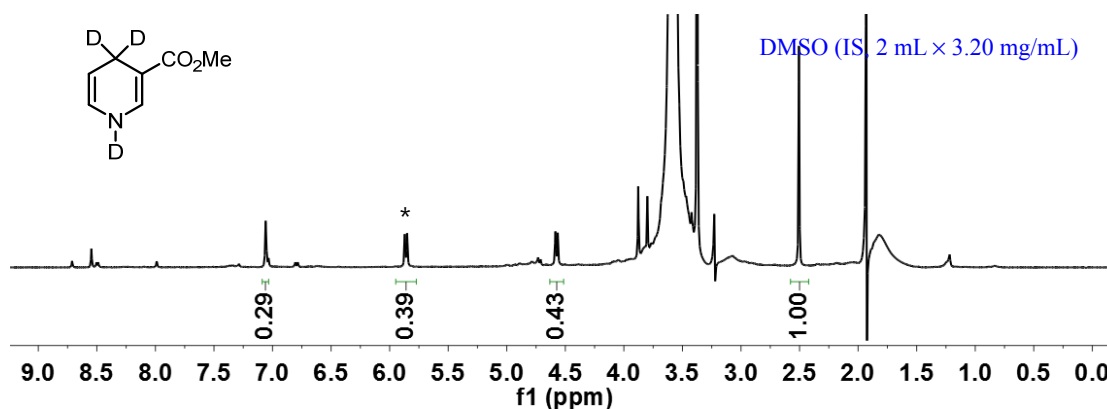
HRMS: Calcd for C₇H₉NO₂ (M+H): 140.07115, Found: 140.07065.

Methyl 1,4-dideuteropyridine-3-carboxylate (**6f-d₂**) from double hydroboration reaction of pyridine **5m**



Determination of the NMR yield. Following the general procedure GP-A, starting from pyridine **5m** (66.8 mg, 0.36 mmol), B₂eg₂ (114 mg, 0.80 mmol), and MeOD (528 mg, 16.0 mmol), and using Cs₂CO₃ (261 mg, 0.80 mmol) as the base in 2 mL of anhydrous MeCN, 1,4,4-trideuteropyridine product **6f-d₃** was generated in 53% NMR yield.

^1H NMR spectra of the reaction mixture (* denotes the peak used to calculate the NMR yield):



Determination of the yield after isolation. Following the general procedure GP-B, starting from pyridine **5m** (403 mg, 2.41 mmol), $\text{B}_{2}\text{eg}_{2}$ (747 mg, 5.27 mmol), and $\text{Cs}_{2}\text{CO}_{3}$ (1.73 g, 5.31 mmol), the 1,4-DHP product **6f-d**₂ (89.3 mg, 26% yield) was obtained. **Note:** In the crude mixture **6f-d**₃ was produced, which became **6f-d**₂ during purification due to H/D exchange at the nitrogen atom.

^1H NMR (400 MHz, CDCl_{3}): δ 7.13 (dd, $J = 5.6, 1.2$ Hz, 1H), 5.88 (ddd, $J = 8.1, 4.2, 1.2$ Hz, 1H), 5.26 (br s, 1H), 4.72 (dd, $J = 8.0, 1.5$ Hz, 1H), 3.70 (s, 3H), 3.09 (m, 0.05H).

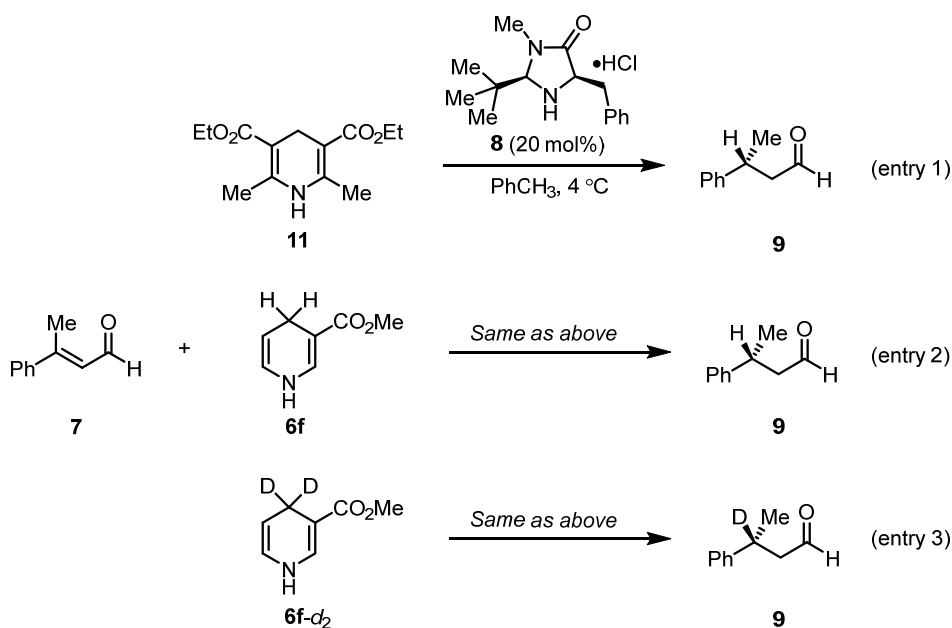
^2H NMR (61 MHz, CDCl_{3}): δ 3.07

^{13}C NMR (100 MHz, CDCl_{3}): δ 168.9, 137.7, 124.1, 103.2, 98.0, 51.1.

HRMS Calcd for $\text{C}_{7}\text{H}_{7}\text{NO}_{2}\text{D}_{2}$ ($\text{M}+\text{H}$)⁺: 142.0837, Found: 142.0827.

6. The Asymmetric Hydrogen Transfer Reactions

6.1 General procedure



General procedure: In a glove box, a 15 mL oven-dried sealed tube was charged with α,β -unsaturated aldehyde **7** (1 equiv.), catalyst **8** (20 mol%), and anhydrous toluene as the solvent (to make a 0.4 M solution of substrate **7**). Then the 1,4-DHP derivative (1.2 equiv.) was added. The resulting mixture was allowed to react at $4\text{ }^\circ\text{C}$ for 12 h. The reaction solution was diluted by ether, and then filtered through a pad of diatomite. The residue was washed by ether. The combined ether solution was washed with brine, dried over Na_2SO_4 , and concentrated. The crude product was purified by flash column chromatography on silica gel (eluted with petroleum ether/ethyl acetate 80:1 V/V) to afford the desired product **9**. The enantiomeric excess (e.e.) was determined by HPLC using a Chiralpak IC column (column temperature $24\text{ }^\circ\text{C}$).

6.2 Results

Entry 1: HEH **11** was employed as the hydride donor.

Following the general procedure, starting from **7** (146 mg, 1.00 mmol), **8** (56.6 mg, 0.20 mmol), and **11** (304 mg, 1.2 mmol), in 5 mL of anhydrous toluene, product **9** (72.3 mg, 49% yield, 83% e.e.) was obtained.

Entry 2: 1,4-DHP **6f** was employed as the hydride donor.

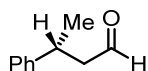
Following the general procedure, starting from **7** (74 mg, 0.50 mmol), **8** (28.3 mg, 0.10 mmol), and **6f** (83.4 mg, 0.6 mmol), in 2.5 mL of anhydrous toluene, product **9** (28.5 mg, 38% yield, 75% e.e.) was obtained.

Entry 3: **6f-d₂** was employed as the hydride donor.

Following the general procedure, starting from **7** (70 mg, 0.48 mmol), **8** (28.3 mg, 0.10 mmol), and **6f-d₂** (67 mg, 0.5 mmol), in 2.5 mL of anhydrous toluene, product **9** (35 mg, 49% yield, 80% e.e.) was obtained.

5.3 Spectroscopic data of the product^[6]

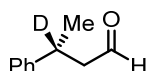
(*R*)-3-Phenylbutanal (9)



¹H NMR (400 MHz, CDCl₃): δ 9.71 (t, *J* = 2.0 Hz, 1H), 7.32 (m, 2H), 7.22 (m, 3H), 3.39 (q, 1H), 2.73 (m, 2H), 1.32 (d, *J* = 7.0 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 202.0, 145.6, 128.8, 126.9, 126.7, 51.9, 42.0, 34.4, 22.3.

(*R*)-1-Deutero-3-phenylbutanal (9-*d*)



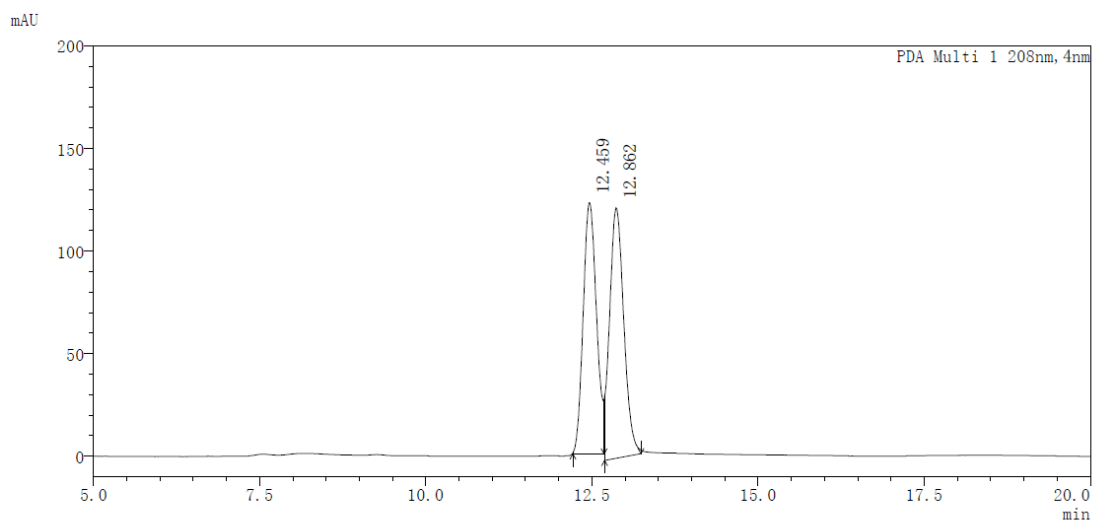
¹H NMR (400 MHz, CDCl₃): δ 9.71 (d, *J* = 1.9 Hz, 1H), 7.31 (m, 2H), 7.22 (m, 3H), 3.36 (q, 0.1H), 2.67 (m, 2H), 1.32 (q, 3H).

²H NMR (61 MHz, CDCl₃): δ 3.36

¹³C NMR (100 MHz, CDCl₃): δ 202.0, 145.6, 128.8, 126.9, 126.7, 51.9, 42.1, 34.4, 22.2 (t, *J* = 25.3 Hz).

5.4 Copies of HPLC traces for the determination of e.e.

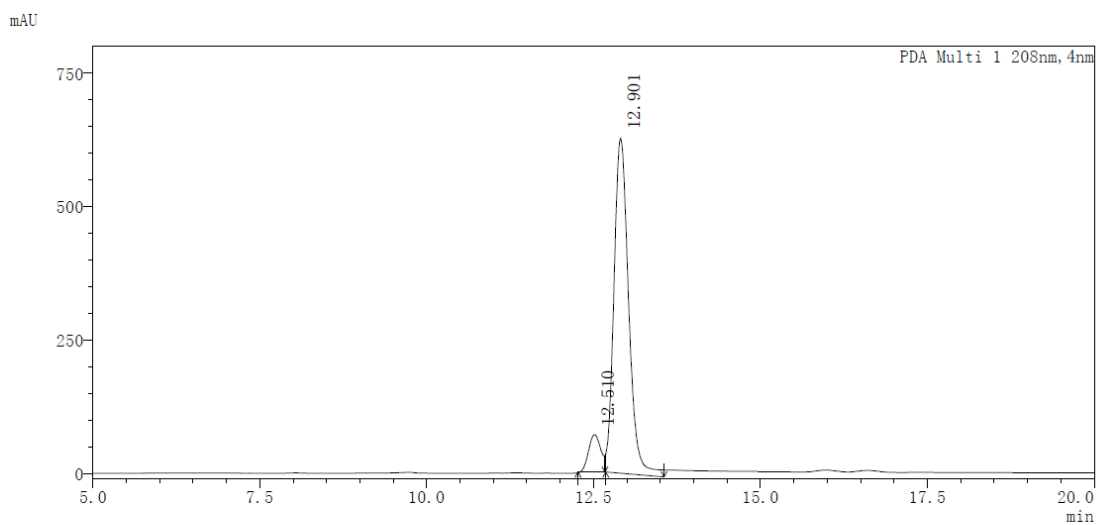
(±)-9:



PDA Ch1 208nm

#	Ret. time	FWHM	Height	Area	Area (%)
1	12.459	0.219	122553	1717049	48.813
2	12.862	0.233	122200	1800559	51.187

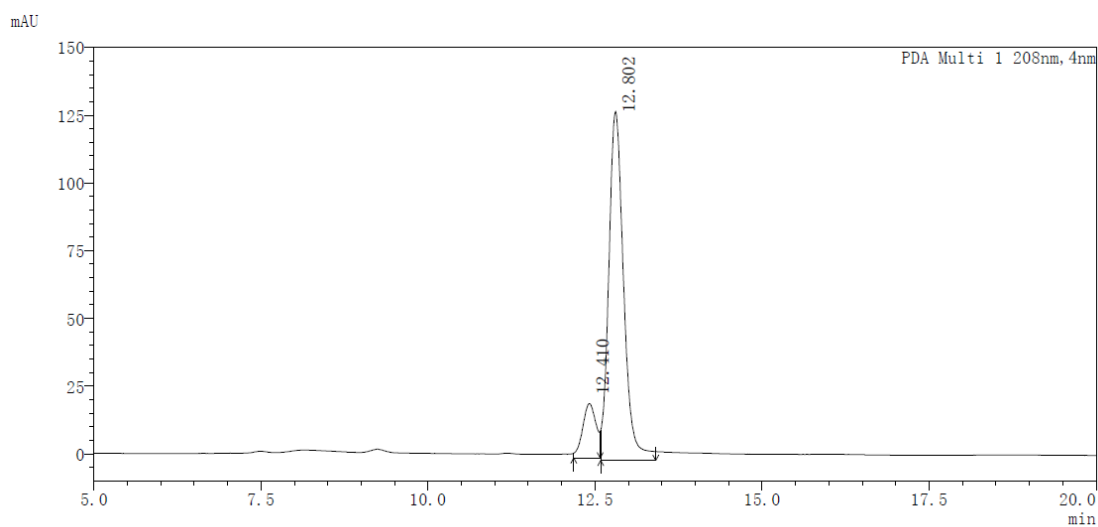
(*R*)-**9** obtained in entry 1 (83% e.e.):



PDA Ch1 208nm

#	Ret. time	FWHM	Height	Area	Area (%)
1	12.510	0.212	69759	887877	8.462
2	12.901	0.231	627290	9604162	91.538

(*R*)-**9** obtained in entry 2 (75% e.e.):

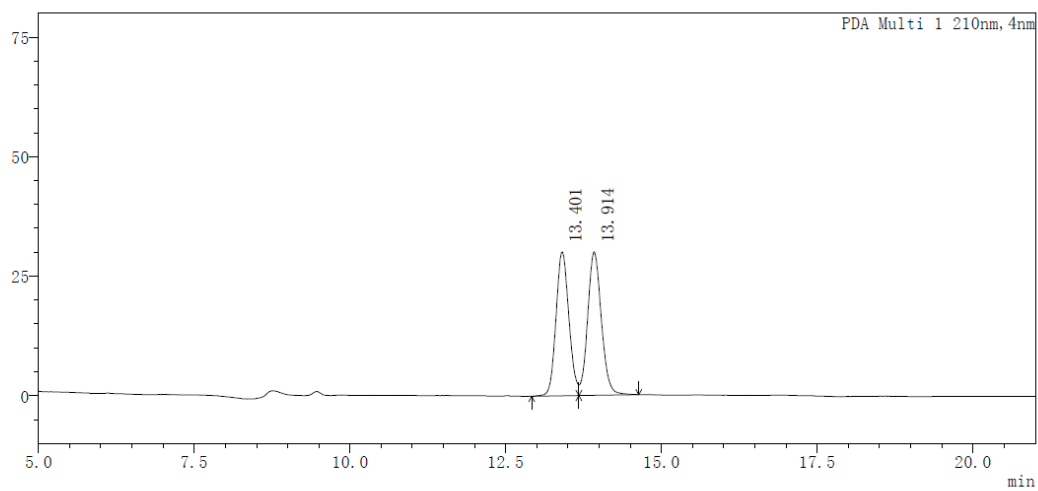


PDA Ch1 208nm

#	Ret. time	FWHM	Height	Area	Area (%)
1	12.410	0.240	20085	283539	12.463
2	12.802	0.230	128737	1991464	87.537

(±)-9-d:

mAU

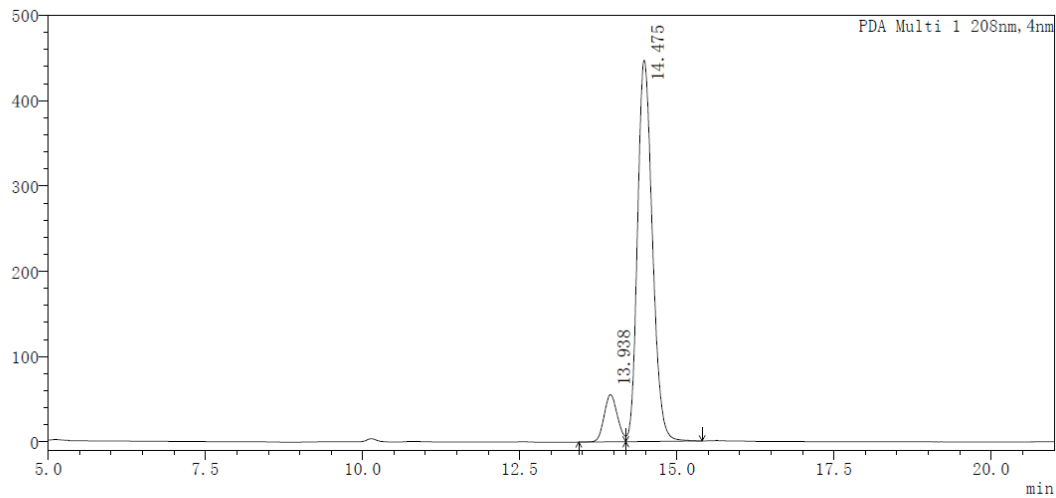


PDA Ch1 210nm

#	Ret. time	FWHM	Height	Area	Area (%)
1	13.401	0.222	30078	433223	48.578
2	13.914	0.233	29994	458582	51.422

(R)-9-d obtained in entry 3 (80% e.e.):

mAU

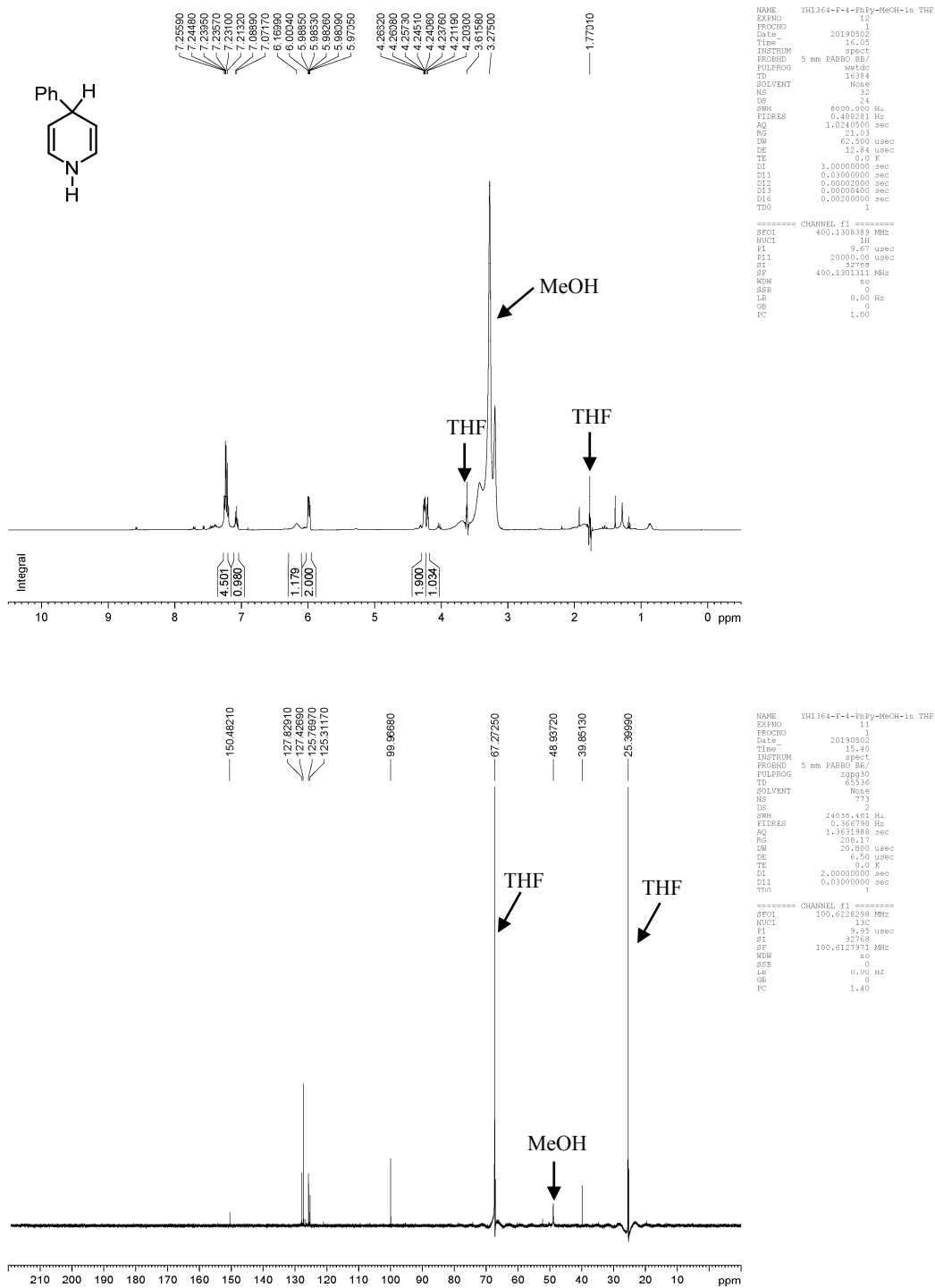


PDA Ch1 208nm

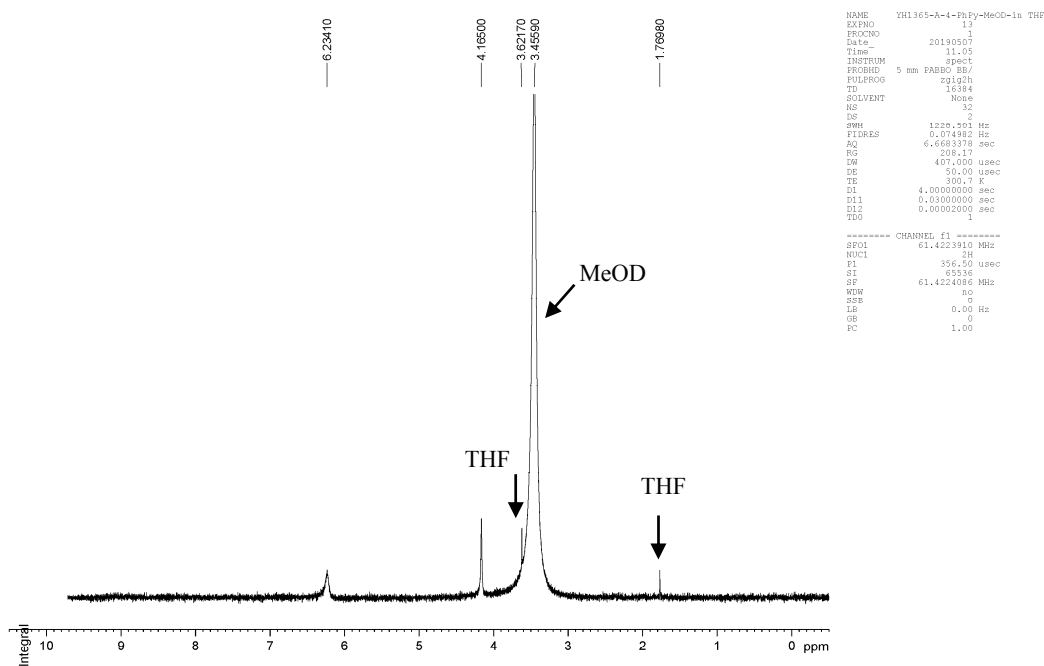
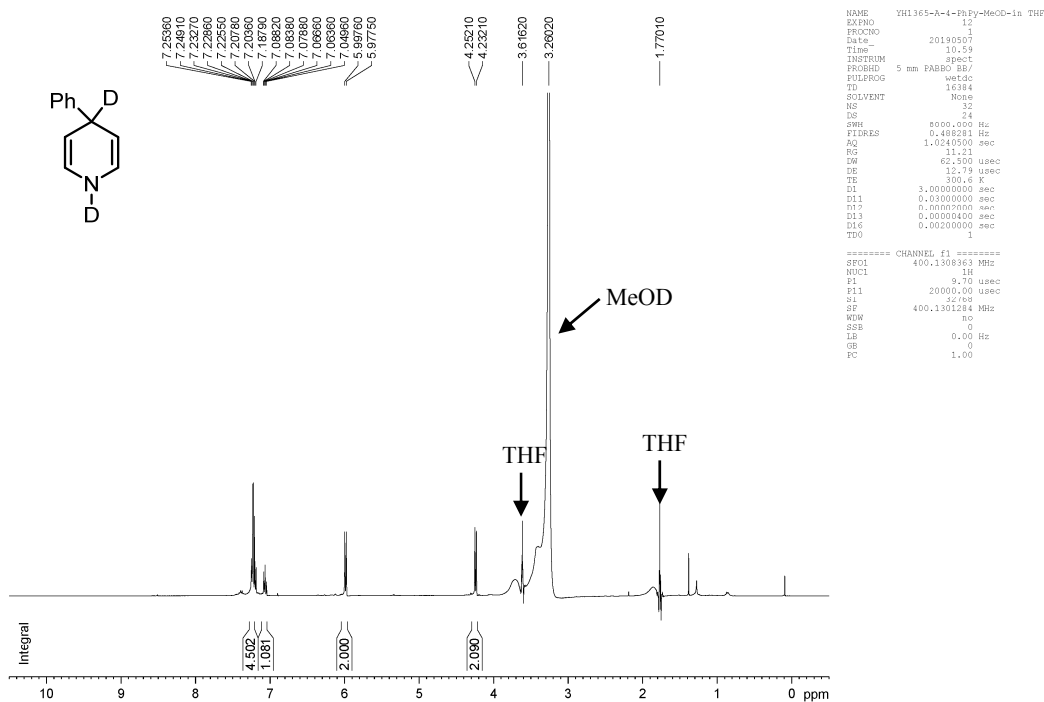
#	Ret. time	FWHM	Height	Area	Area (%)
1	13.938	0.230	55221	810221	9.919
2	14.475	0.254	447191	7358232	90.081

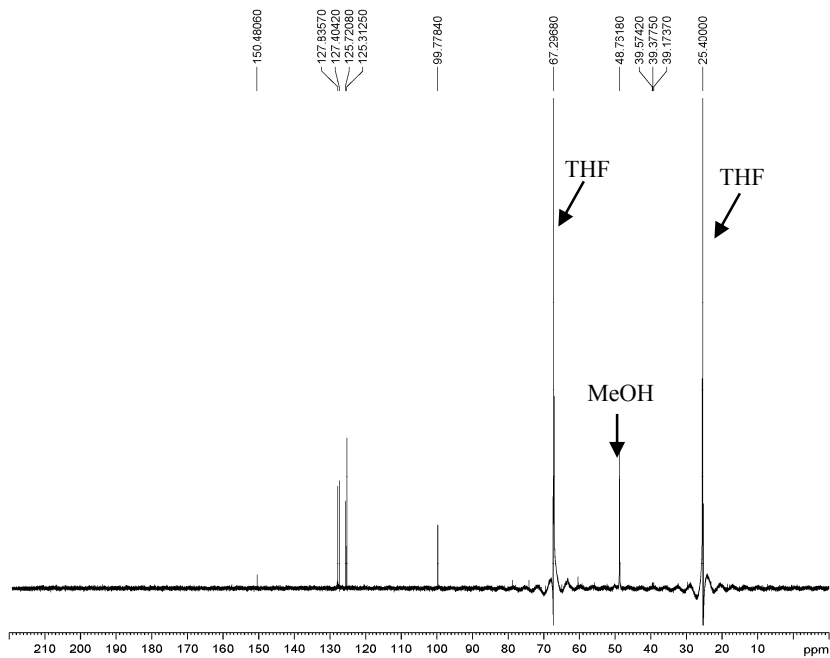
7. Copies of the NMR Spectra of Products

¹H and ¹³C NMR of **4** (in THF-d₀, No-D NMR)



¹H, ²H, and ¹³C NMR of **4-d₂** (in THF-d₀, No-D NMR)



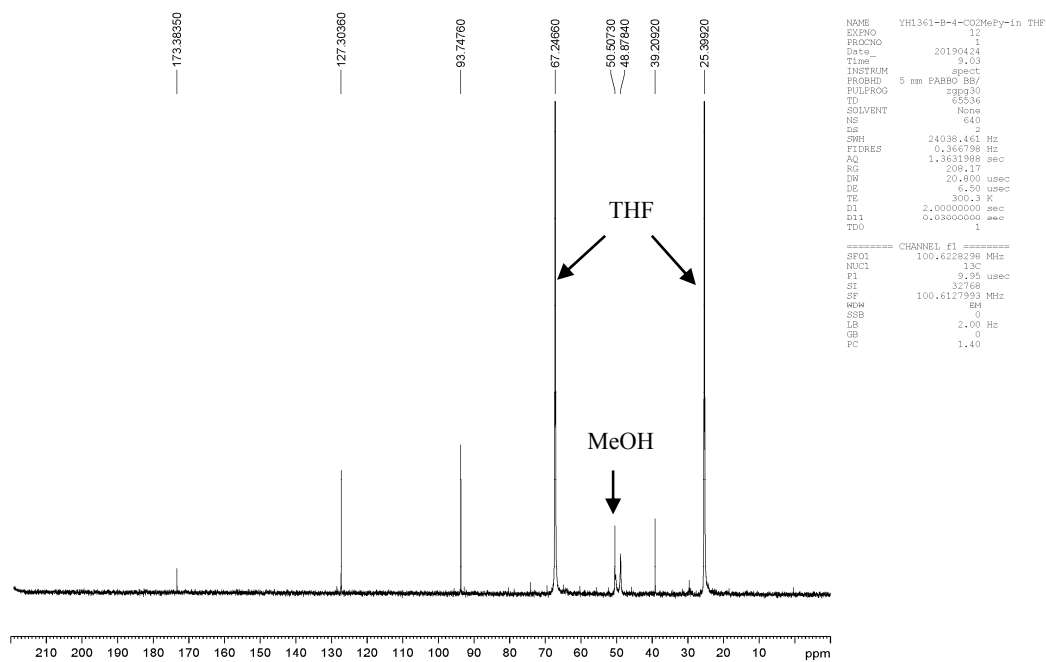
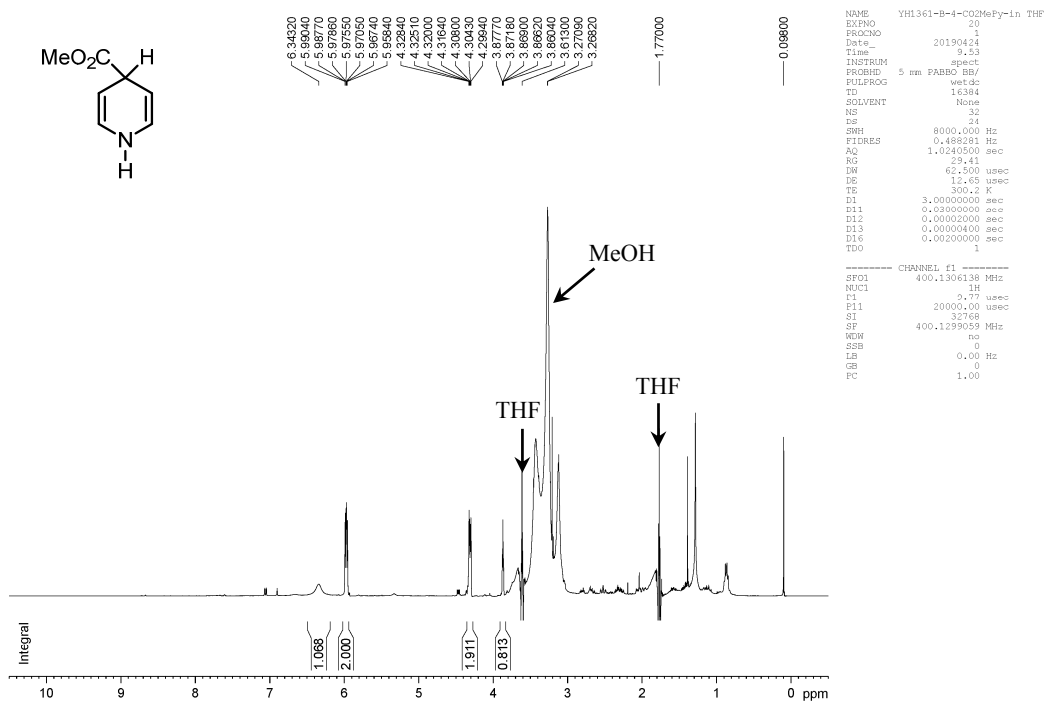


```

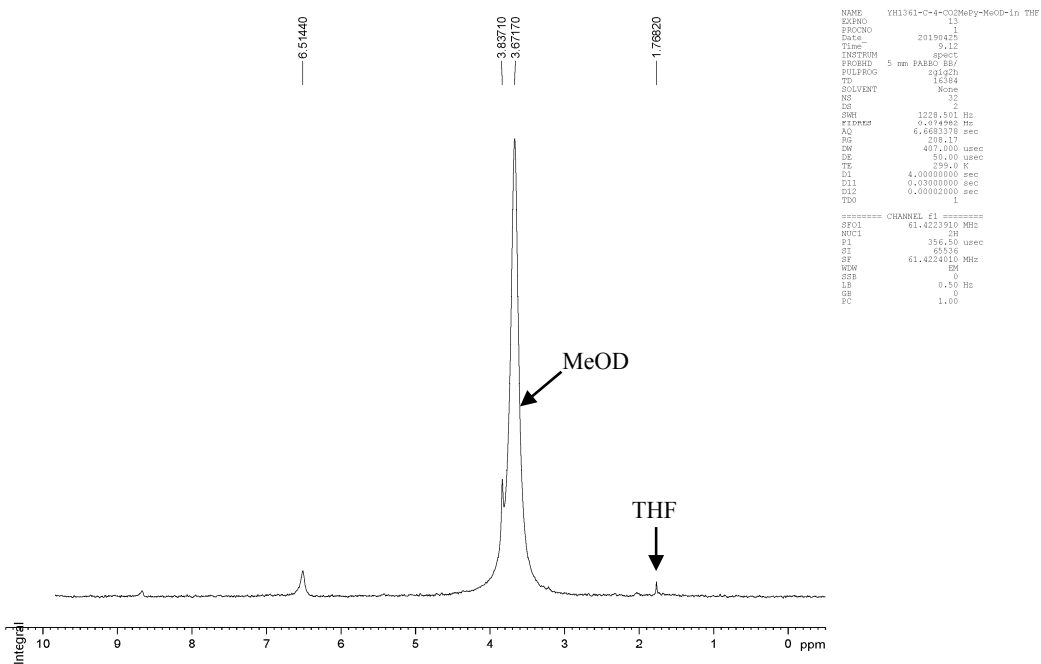
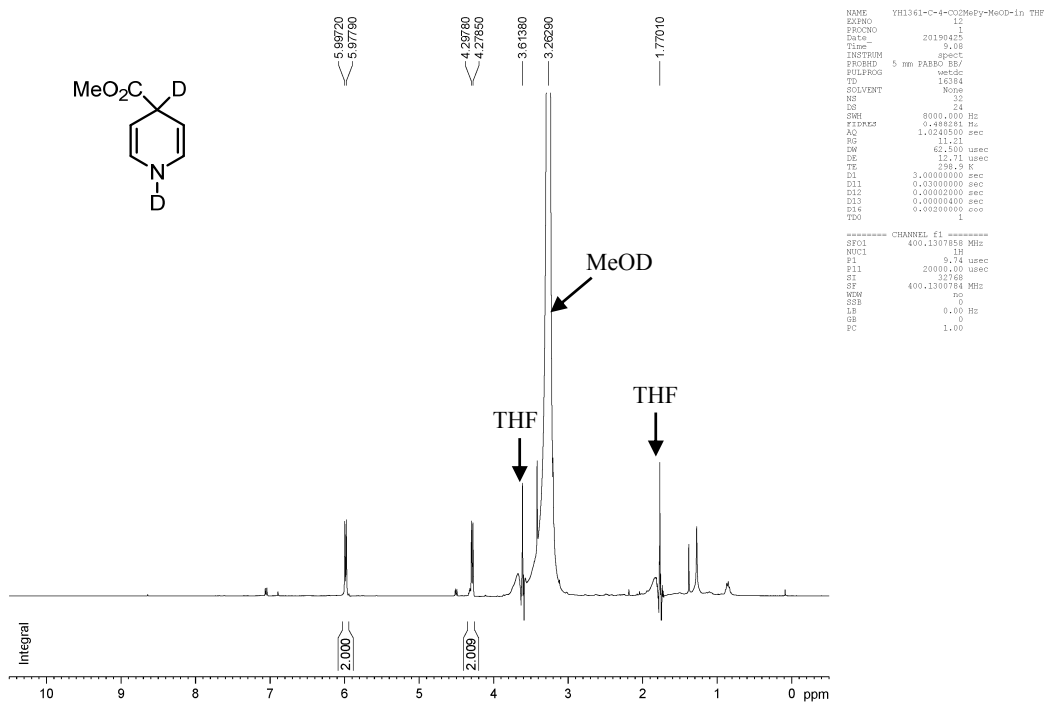
NAME      YH1365-A-4-Phy-MeOD-In THF
EXPNO    1
PROCNO   1
Date_    20190507
Time     9.42
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpg30
TD       65536
SOLVENT  None
NS       1280
DS       2
SWH      24030.463 Hz
FIDRES   0.366798 Hz
AQ       1.3631988 sec
RG       328.17
DW       20.800 usec
DE       6.50 usec
TE       300.6 K
DL       2.00000000 sec
D11      0.03000000 sec
TD0      1
===== CHANNEL f1 =====
SFO1    100.6228299 MHz
NUC1    13C
P1      9.95 usec
PT      32768
SF      100.6127949 MHz
RG      32
SSB     0
LB      0.05 Hz
GB      0
PC      1.40

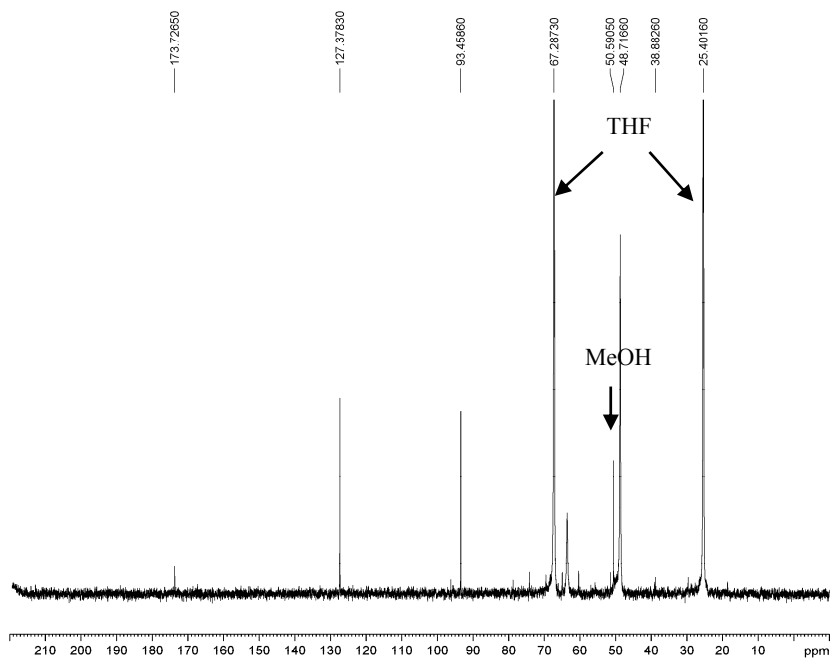
```

¹H and ¹³C NMR of **6a** (in THF-*d*₀, No-D NMR)



¹H, ²H, and ¹³C NMR of **6a-d₂** (in THF-d₀, No-D NMR)



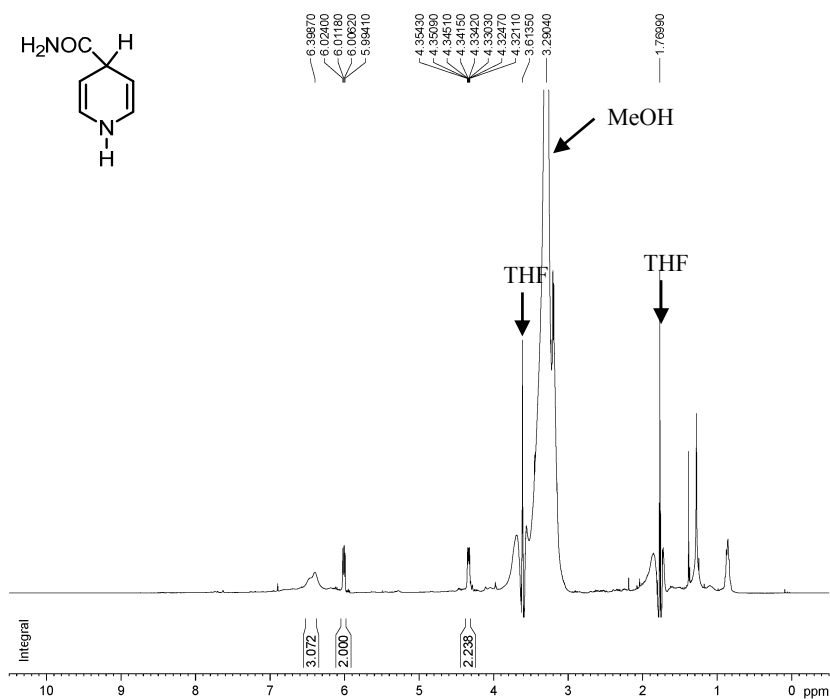
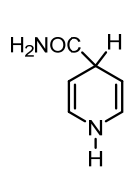


```

NAME      YH1361-C-4-OY2MePy-MeOD-in THF
EXPNO    11
PROCNO   1
Date_    20190421
Time     8:30
INSTRUM  spect
PROBHD   5 mm PABBO BBO
PULPROG  zgpg30
TD        65536
SOLVENT  None
NS        640
DS        2
SWH       24038.461 Hz
FIDRES   0.389798 Hz
AQ        1.3631988 sec
RG        256.17
DF        20.480 usec
DE        6.50 usec
TE        299.2 K
D1        2.0000000 sec
D11       0.3300000 sec
TD0       1
----- CHANNEL f1 -----
SFO1     100.628276 MHz
NUC1     13C
P1        9.25 usec
SI        32768
SF        100.6127824 MHz
RG        64
WENW     0
SSB       0
LB        2.00 Hz
GB        0
PC        1.40

```

¹H and ¹³C NMR of **6b** (in THF-d₀, No-D NMR)

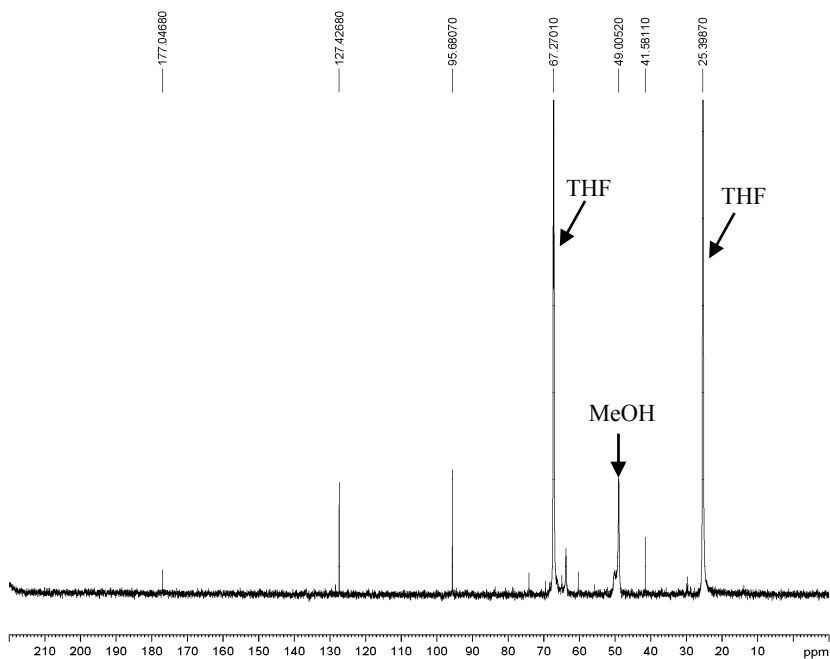


```

NAME      YH1361-F-4-CONH2-MeOH-in THF
EXNO     12
PROCNO   1
Date      20190426
Time      9.24
INSTRUM  spect
PROBHD   5 mm EBBBO BB/
PULPROG  zgpg30
TD        16384
SOLVENT  None
NS        32
DS        24
SWH       8000.000 Hz
FIDRES   0.480261 Hz
AQ        1.0240500 sec
RG        21.03
SW        62.500 usec
DE        12.54 usec
TE        299.1 K
D1        3.0000000 sec
D11       0.0300000 sec
D12       0.0000200 sec
D13       0.0000400 sec
D15       0.0020000 sec
TD0       1
    
```

```

----- CHANNEL f1 -----
SF01    400.1305000 MHz
NUC1     1H
P1       9.83 usec
P11      20000.00 usec
SI       32768
SF       400.1279771 MHz
WDW      no
SSB      0
LB       0.00 Hz
GB       0
PC       1.00
    
```



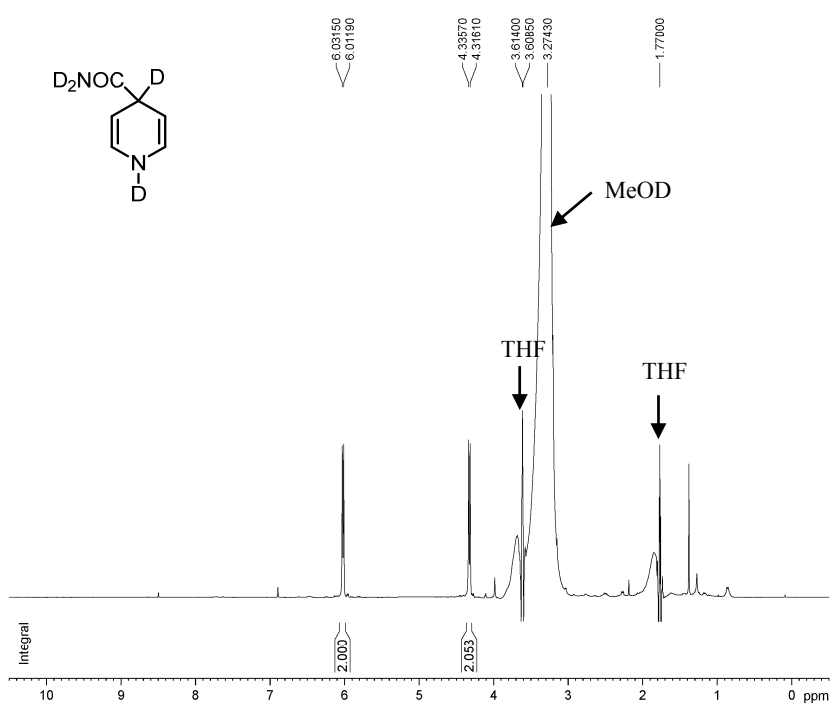
```

NAME      YH1361-F-4-CONH2-MeOH-in THF
EXNO     11
PROCNO   1
Date      20190426
Time      8.40
INSTRUM  spect
PROBHD   5 mm EBBBO BB/
PULPROG  zgpg30
TD        65536
SOLVENT  None
NS        1024
DS        2
SWH       24038.461 Hz
FIDRES   0.366798 Hz
AQ        1.3633988 sec
RG        208.17
SW        40.800 usec
DE        6.50 usec
TE        299.5 K
D1        2.0000000 sec
D11       0.0300000 sec
TD0       1
    
```

```

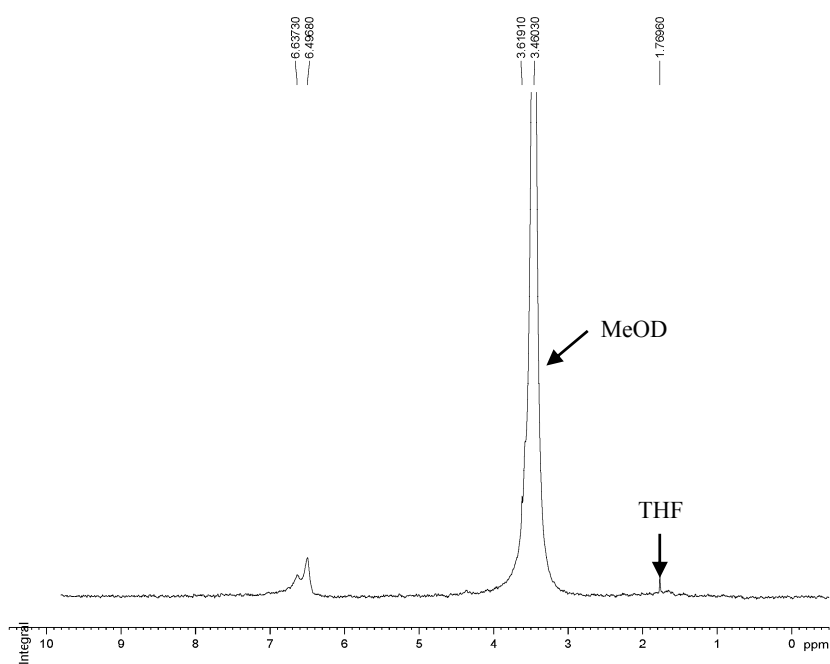
----- CHANNEL f1 -----
SF01    100.6228298 MHz
NUC1     13C
P1       9.95 usec
P11      100.617124 MHz
SI       32768
WDW      EM
SSB      0
LB       2.00 Hz
GB       0
PC       1.40
    
```

^1H , ^2H , and ^{13}C NMR of **6b-d₄** (in THF-*d*₆, No-D NMR)



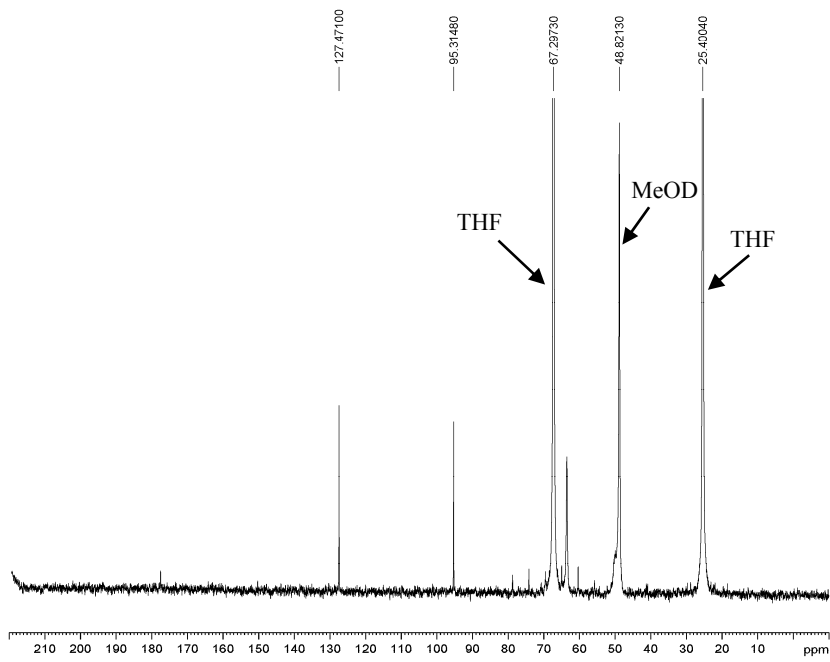
```

NAME YH1363-B-4-COND2Py-MeOD-in THF
EXPNO 12
PROCNO 1
Date_ 20190501
Time 0.44
INSTRUM spect
PROBHD 5 mm F4BBO BE7
PULPROG wetzc
TD 16384
SOLVENT None
DS 24
SM 8000.000 Hz
FIDRES 0.494825 Hz
AQ 1.0240500 sec
RG 21.03
DW 62.500 usec
DE 12.50 usec
TE 0.0 K
D1 3.0000000 sec
D11 0.0300000 sec
D12 0.0000200 sec
D13 0.0000040 sec
D14 0.0000000 sec
TD0 1
===== CHANNEL f1 =====
SFO1 400.1307945 MHz
NUC1 1H
P1 9.61 usec
P11 20000.00 usec
SF 400.1307944 MHz
NS 0
SSB 0
LB 1.00 Hz
GB 0
PC 1.00
    
```



```

NAME YH1363-B-4-COND2Py-MeOD-in THF
EXPNO 13
PROCNO 1
Date_ 20190501
Time 9.51
INSTRUM spect
PROBHD 5 mm F4BBO BE7
PULPROG zgpg30
TD 16384
SOLVENT None
DS 32
SM 1228.501 Hz
FIDRES 0.091496 Hz
AQ 6.6683378 sec
RG 258.17
DW 401.000 usec
DE 50.00 usec
TE 0.0 K
D1 4.0000000 sec
D11 0.0300000 sec
D12 0.0000200 sec
TD0 1
===== CHANNEL f1 =====
SFO1 61.4223910 MHz
NUC1 13C
P1 356.50 usec
P11 60556
SF 61.4224028 MHz
NS 0
SSB 0
LB 0.50 Hz
GB 0
PC 1.00
    
```

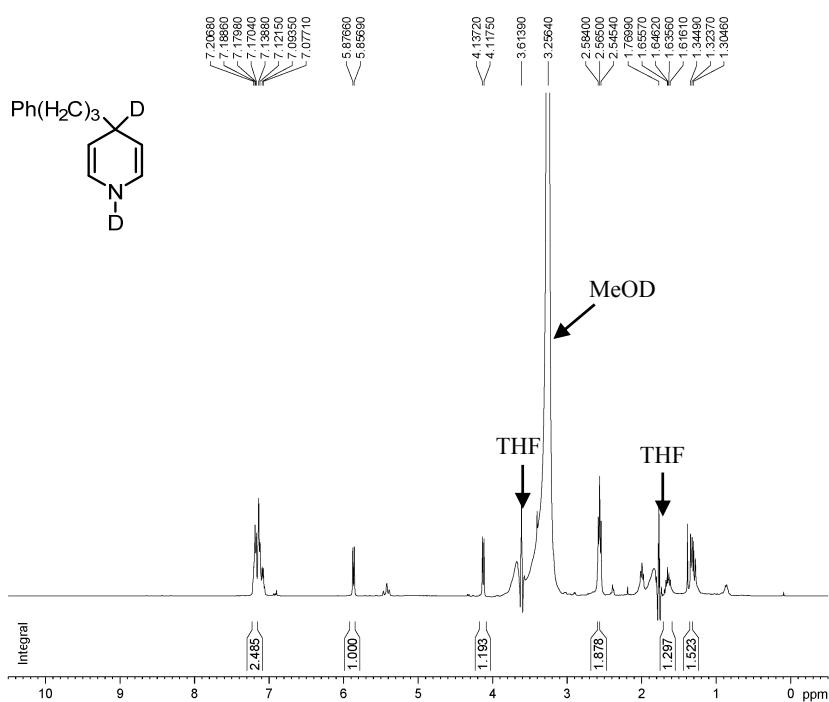



```

NAME      YH1363-B-4-CYMO2Py-MeOD-in THF
EXPNO    11
PROCNO   1
Date_    20190501
Time     8:26
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpg30
TD        65536
SOLVENT  None
NS        1280
DS        2
SHE      24038.461 Hz
FIDRES   0.389758 Hz
AQ        1.3631988 sec
RG        200.117
DF        20.480 usec
DE        6.50 usec
TE        300.2 K
D1        2.0000000 sec
D11       0.3300000 sec
TD0       1
----- CHANNEL f1 -----
SFO1     100.6282758 MHz
NUC1     13C
P1        9.25 usec
SI        32768
SF        100.6127850 MHz
WEN      BM
SSB       0
LB        4.00 Hz
GB        0
PC        1.40

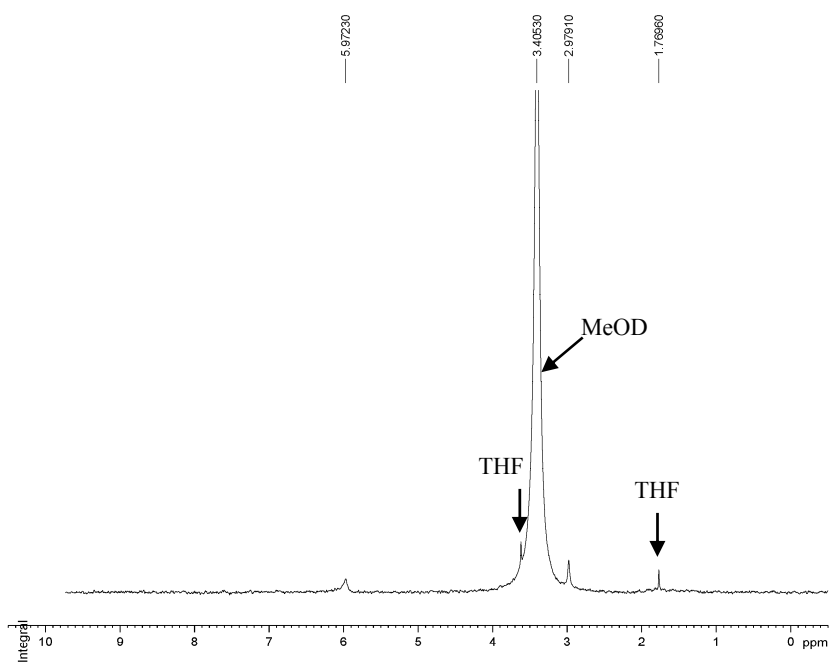
```

^1H , ^2H , and ^{13}C NMR of **S2-d₂** (in THF-d₀, No-D NMR)



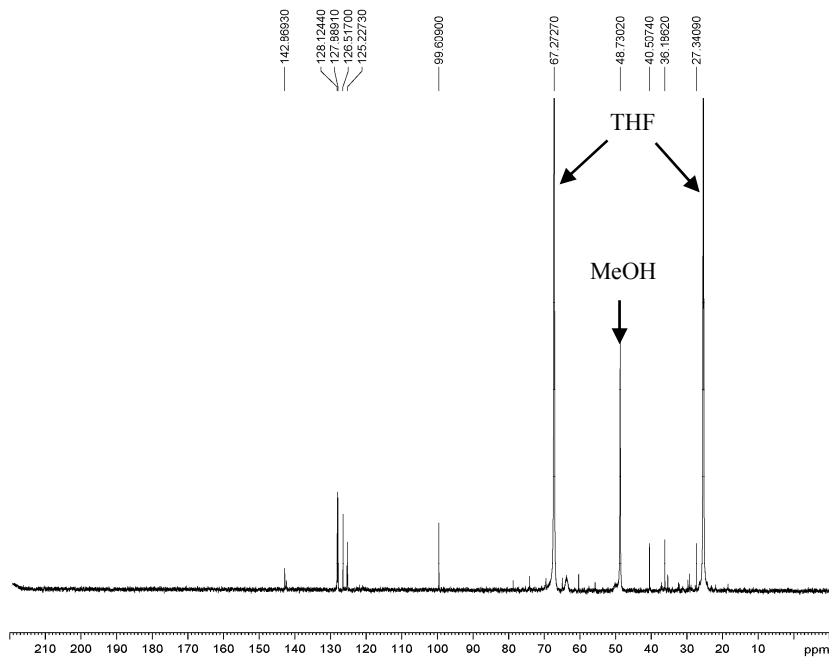
```

NAME      YH1361-G-4-(CH2)3PhEY-MeOD-In-THF
EXPNO    12
PROCNO   1
Date_    20190429
Time     2.10
INSTRUM  spect
PROBHD   5 mm PABBO BBO
PULPROG  zgpg2h
TD        65536
SOLVENT  None
NS        32
DS        2
SWH       8000.000 Hz
FIDRES   0.448281 Hz
AQ        1.242600 sec
RG         24.00
DE         62.000 usec
DB         24.00
TE        12.07 usec
TE2       297.2 K
D1         3.00000000 sec
D11        0.03000000 sec
D12        0.00000000 sec
D13        0.00000000 sec
D14        0.00200000 sec
TD0       1
----- CHANNEL f1 -----
SFO1     400.1308013 MHz
NUC1      1H
P1         9.15 usec
PL1       20000.00 usec
SF        400.1301221 MHz
MC        RM
SFR        0
LB         1.00 Hz
GB         0
PC         1.00
    
```



```

NAME      YH1361-G-4-(CH2)3PhEY-MeOD-In-THF
EXPNO    13
PROCNO   1
Date_    20190429
Time     2.10
INSTRUM  spect
PROBHD   5 mm PABBO BBO
PULPROG  zgpg2h
TD        65536
SOLVENT  None
NS        32
DS        2
SWH       1229.161 Hz
FIDRES   0.074982 Hz
AQ        6.4669376 sec
RG         295.17
DE         407.000 usec
DB         50.00 usec
TE        295.3 K
D1         4.00000000 sec
D11        0.03000000 sec
D12        0.00000000 sec
TD0       1
----- CHANNEL f1 -----
SFO1     61.4223910 MHz
NUC1      13C
P1        356.50 usec
PL1       65536
SF        61.4224076 MHz
MC        RM
SFR        0
LB         0.50 Hz
GB         0
PC         1.00
    
```

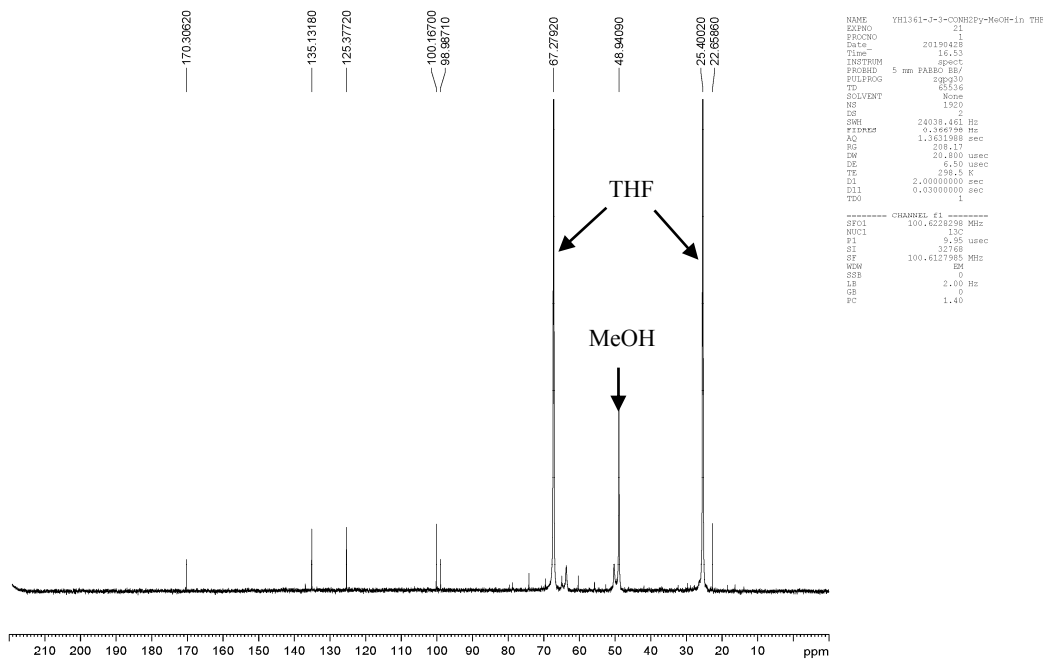
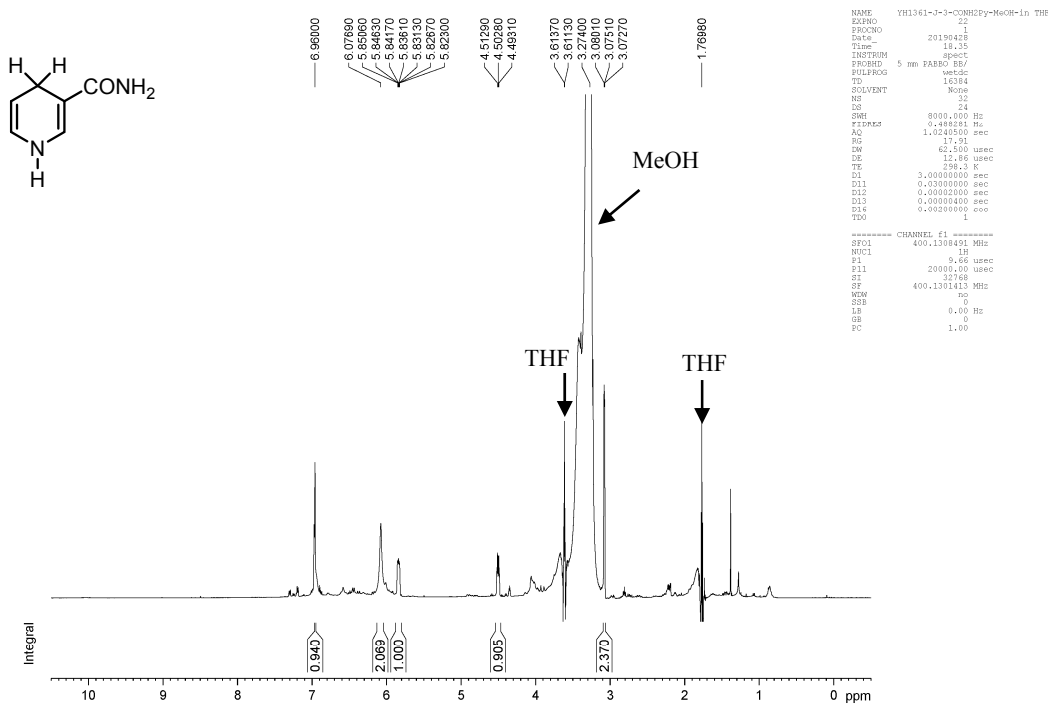


```

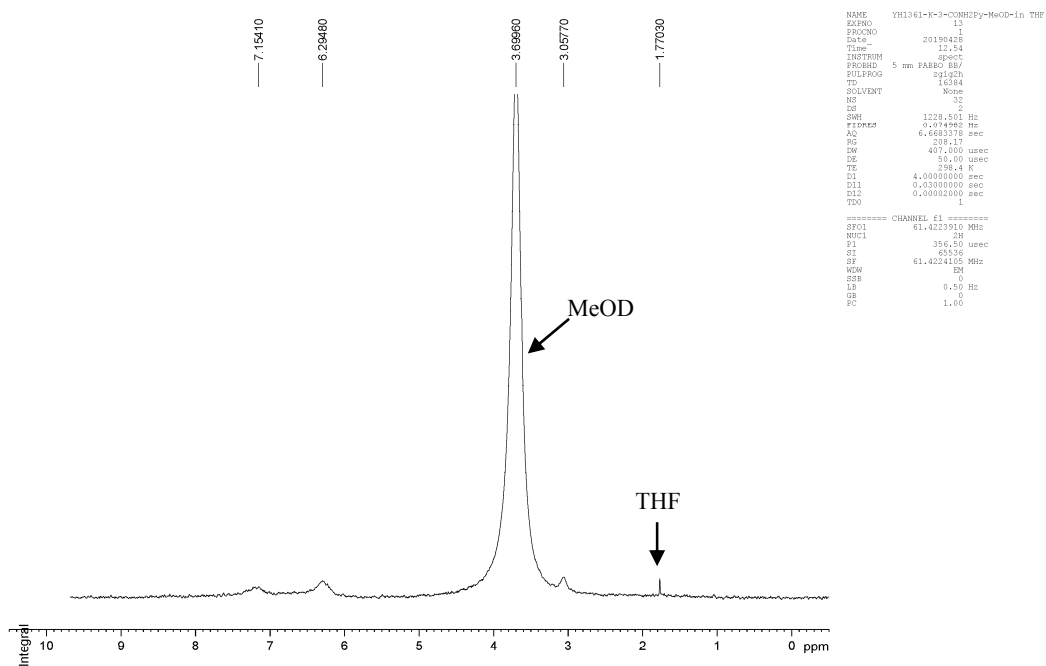
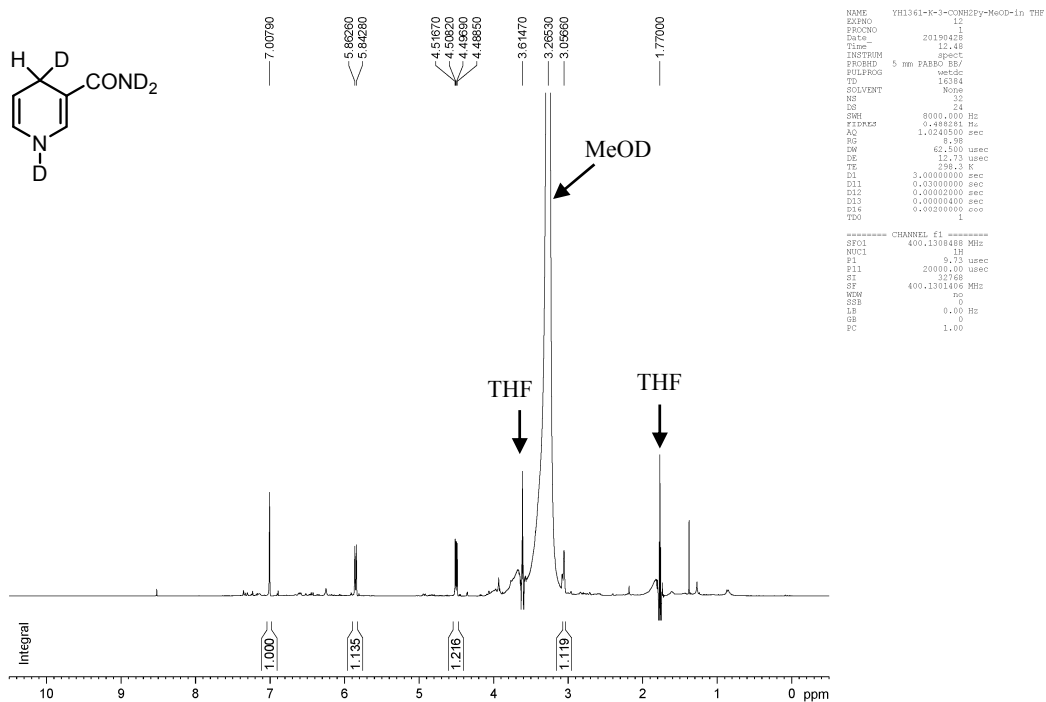
NAME YH161-Q-4-(CH2)3PhFy-MCO-In THF
EXPNO 1
PROCNO 1
DATE_ 20190423
TIME 8.05
INSTRUM spect
PROBHD 5 mm F4001
PULPROG zgpg30
TD 65536
SOLVENT None
RG 320
DS 2
SWH 24000.463 Hz
FIDRES 0.366798 Hz
AQ 1.363198 sec
RG 208.17
RM 20.800 usec
DE 6.50 usec
TE 293.4 K
D1 2.00000000 sec
D11 0.02000000 sec
TD0 1
----- CHANNEL f1 -----
NUC1 13C
P1 130
SI 8.95 usec
SF 100.6127949 MHz
WDW EM
GB 0
LB 2.00 Hz
GB 0
PC 1.40

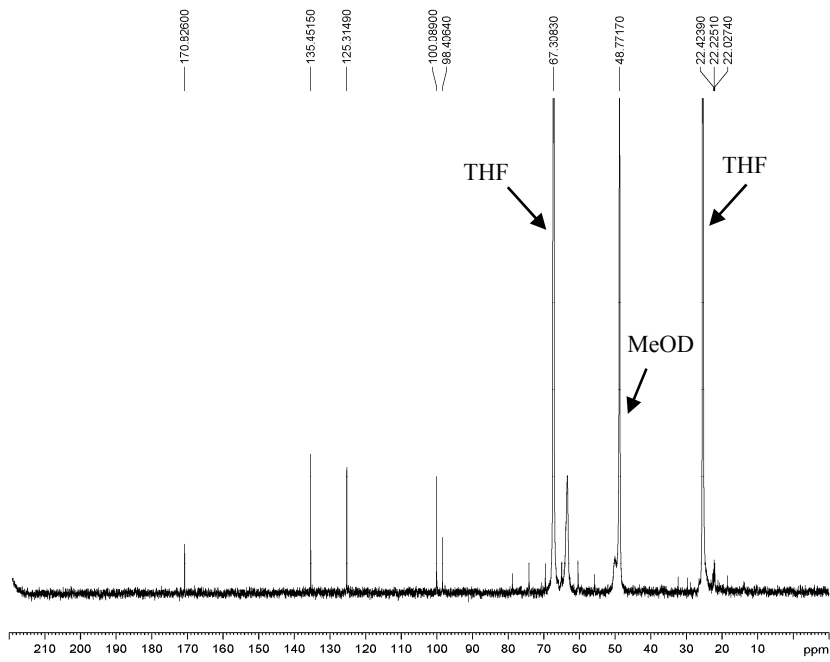
```

¹H and ¹³C NMR of **6c** (in THF-*d*₀, No-D NMR)



¹H, ²H, and ¹³C NMR of **6c-d₄** (in THF-d₀, No-D NMR)



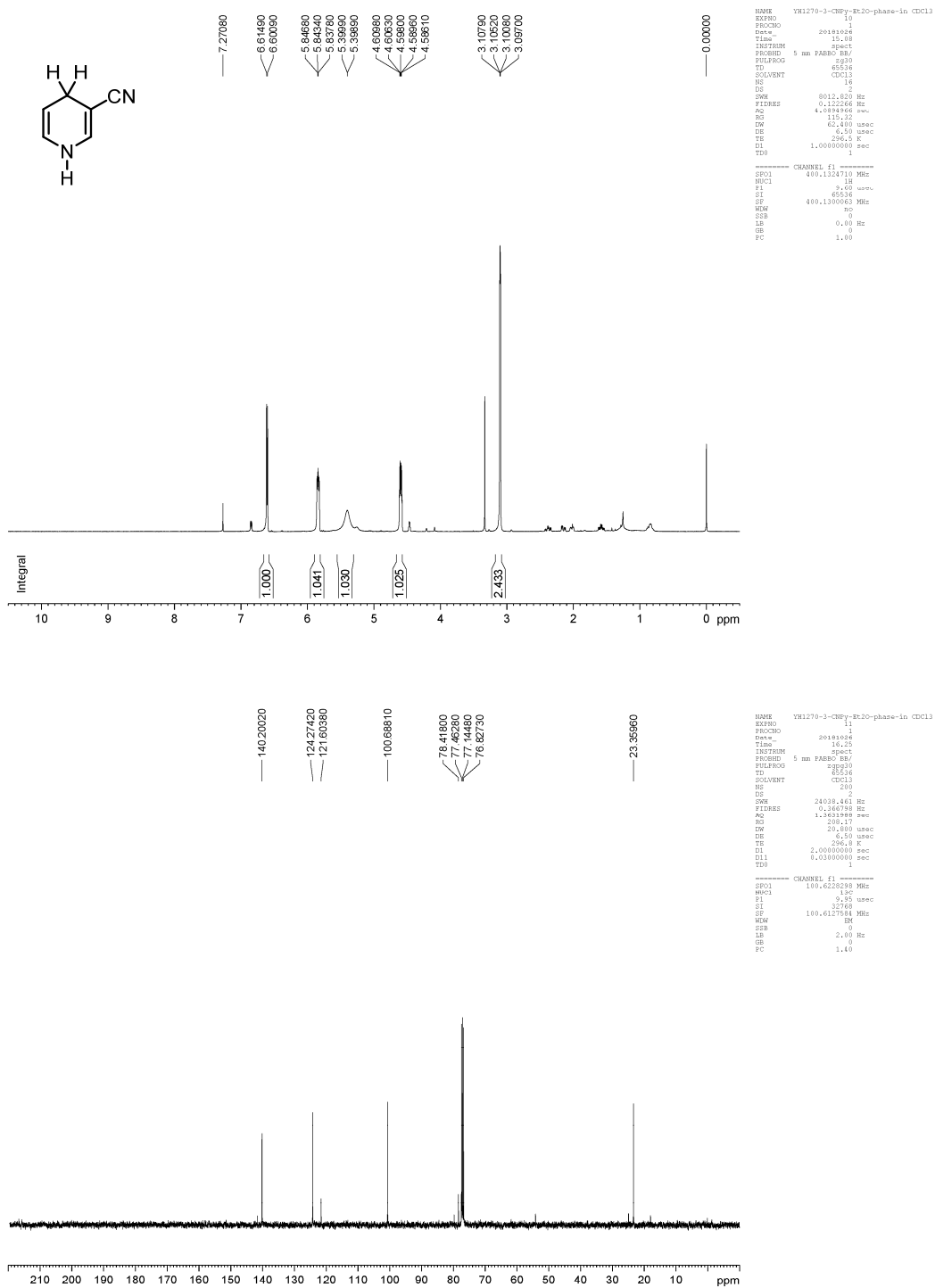


```

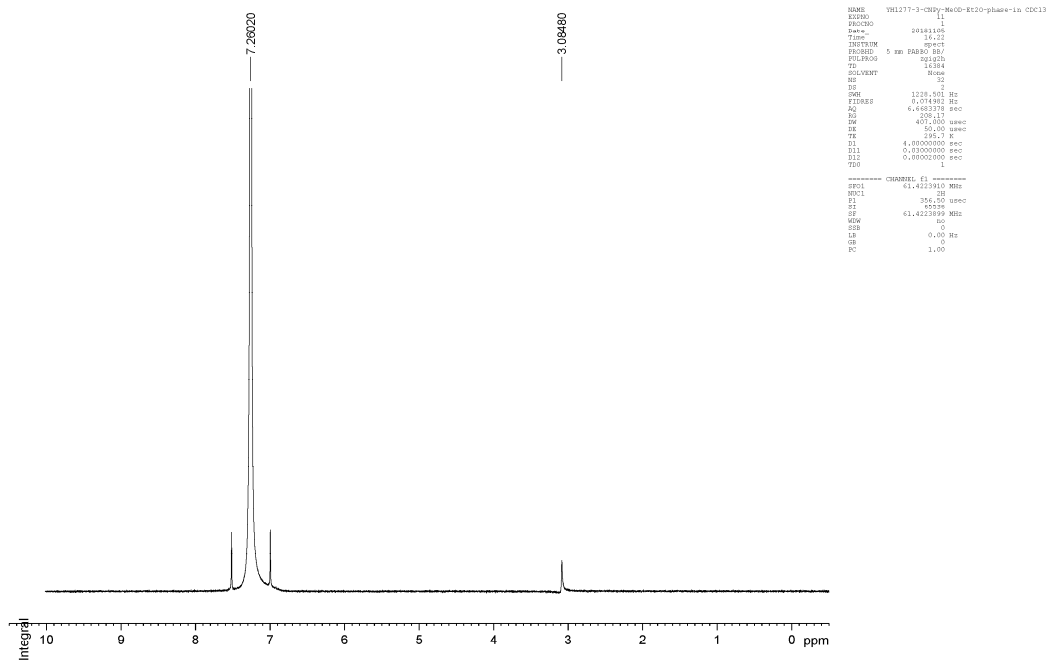
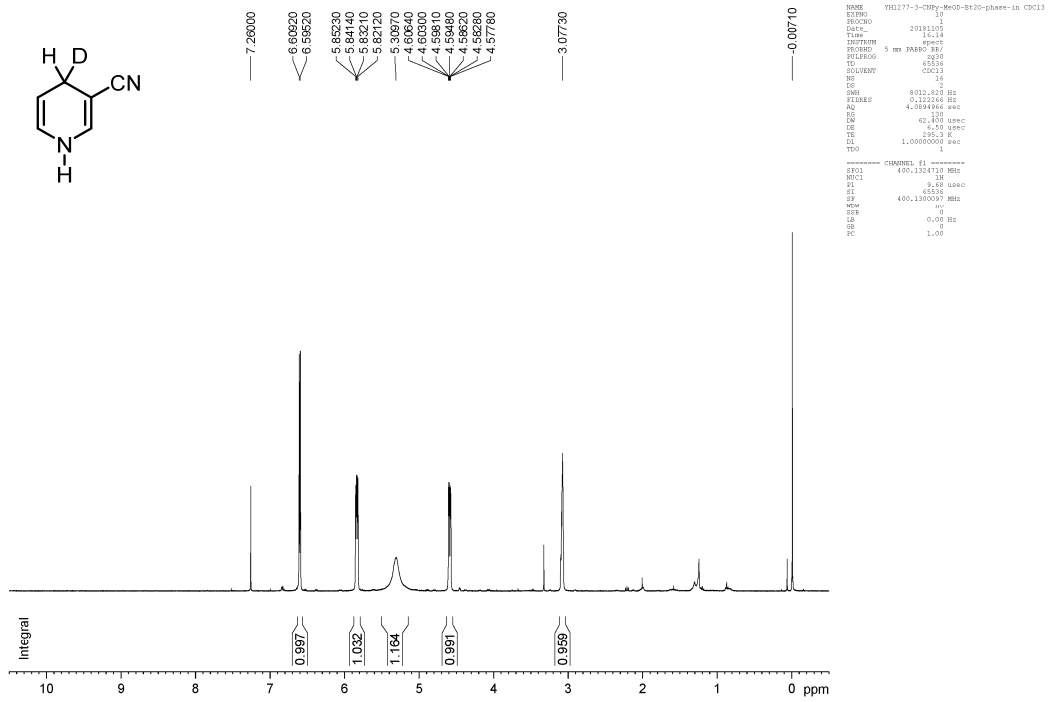
NAME      YH1361-K-3-CYWH2Py-MeOD-in THF
EXPNO    11
PROCNO   11
Date_    20190428
Time     12.40
INSTRUM  spect
PROBHD   5 mm PABBO BBI
PULPROG  zgpg30
TD        65536
SOLVENT  None
NS        1500
DS        2
SHE      24038.461 Hz
FIDRES   0.389798 Hz
AQ        1.3631988 sec
RG        381.17
DF        20.400 usec
DE        6.50 usec
TE        298.2 K
D1        2.0000000 sec
D11       0.3300000 sec
TD0       1
----- CHANNEL f1 -----
SFO1     100.628296 MHz
NUC1     13C
P1        9.25 usec
SI        32768
SF        100.6127971 MHz
KENV     64
SSB      0
LB        2.00 Hz
GB        0
PC        1.40

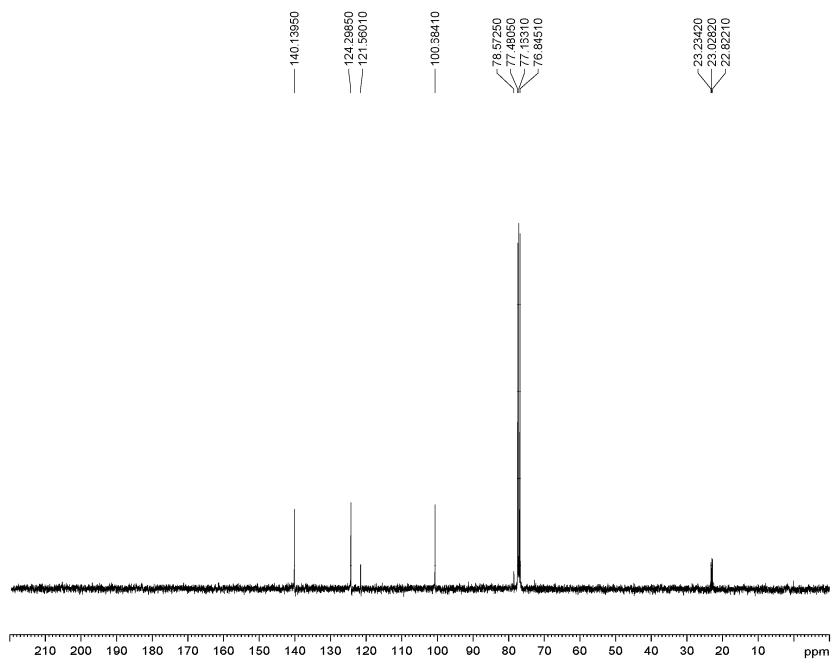
```

¹H and ¹³C NMR of **6d**:



^1H , ^2H , and ^{13}C NMR of **6d-d'**:

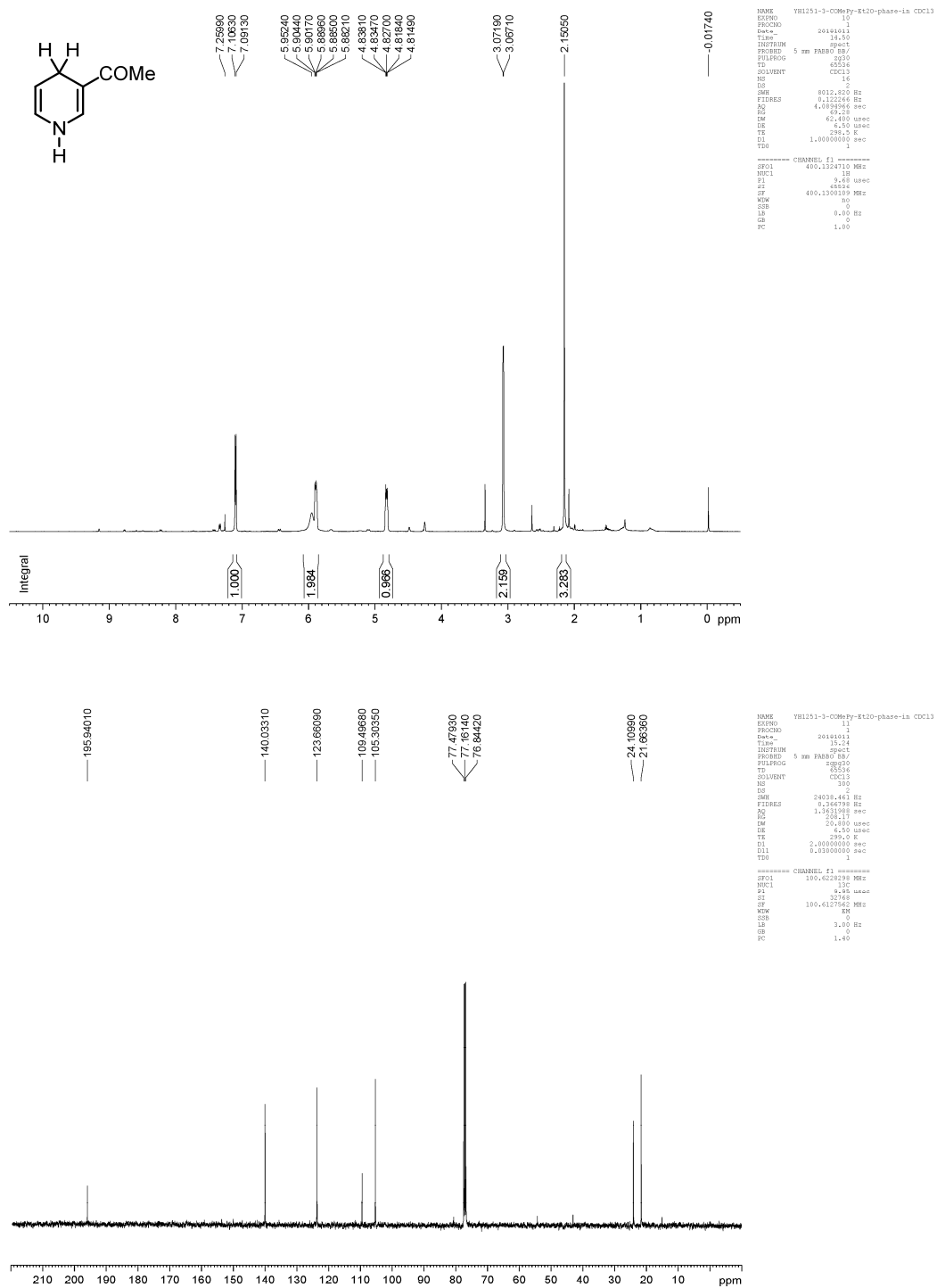




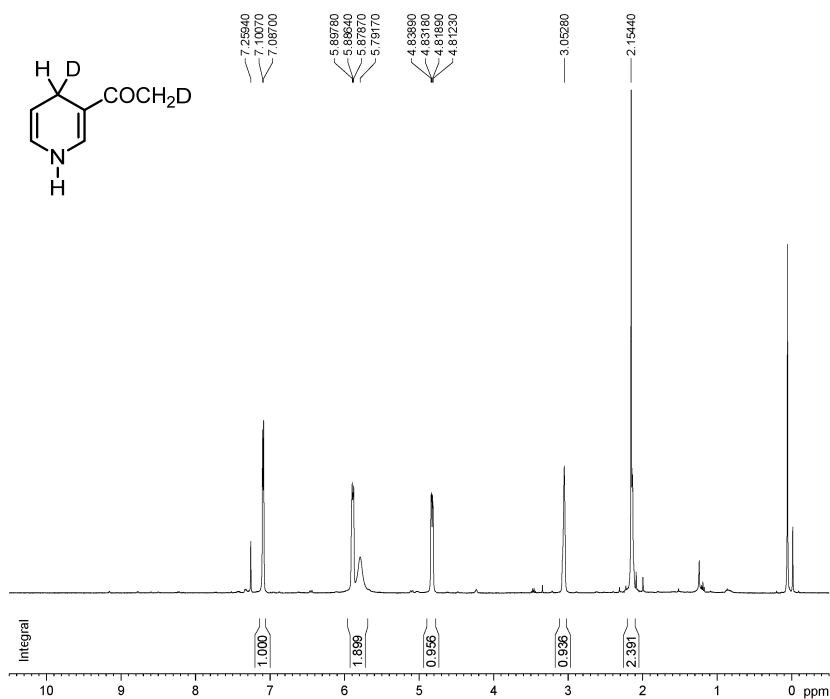
```

NAME      YH1277-3-CHY-Me00-B120-phase-in CDCl3
EXPNO    1
PROCNO   1
F1F2     51411745
TIME     24.28
INSTRUM  spect
PROBHD   5 mm BBO-BA
PULPROG  zgpg30
TD        65536
SOLVENT  CDCl3
NS       500
DS       4
SWH      24398.461 Hz
FIDRES   0.246798 Hz
AQ       1.3931983 sec
RG       320.000
DE       20.000 umic
TE       300.2 K
CE       0.10000000
D1       2.00000000 sec
d11      0.03000000 sec
TD0      1
----- CHANNEL f1 -----
NUC1      13C
P1        2.00
PL1       0.00 umic
PR        37.00
RF        100.625000 MHz
WDW       EM
SSB       0
GB        0
PC        1.00 Hz
RB        1.40
  
```

¹H and ¹³C NMR of **6e**:



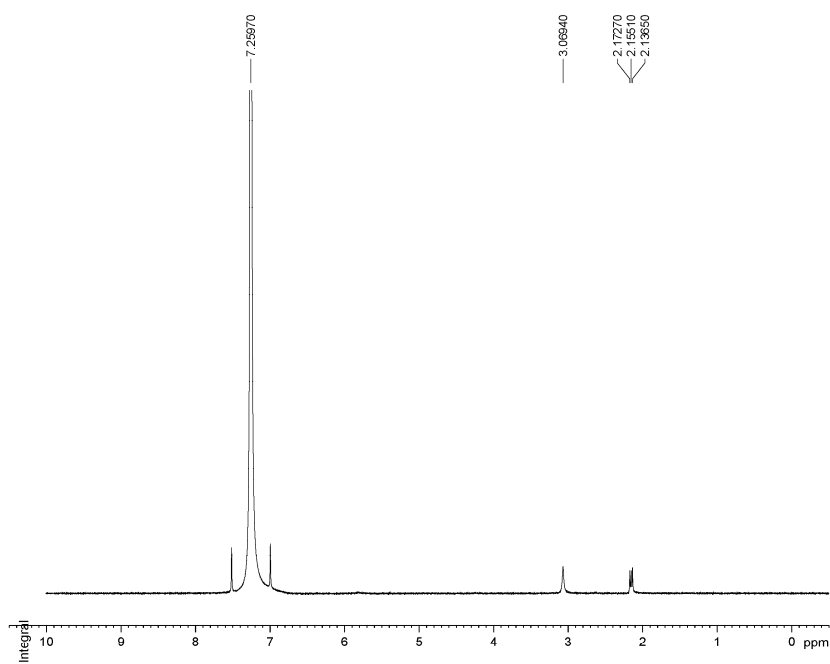
^1H , ^2H , and ^{13}C NMR of **6e-d**:



```

NAME 1H1347-9-3-CCl4-1120-phase-in-CDCl3-again
EXPNO 12
PROCNO 1
DATE_ 20191118
TIME 21:08
PROBHD 5 mm PABBO BB1
PULPROG zgpg30
PCPDPRG2 f2
TD 65536
SOLVENT CDCl3
NS 16
DS 4
F2 8013.450 Hz
FIDRES 0.1322500 Hz
AQ 4.0894900 sec
RG 492.12
AQ 4.0894900 sec
DE 18.140 MHz
TE 300.2 K
DE 17.00 MHz
TE 300.2 K
SI 1.0000000 sec
SI 1
===== CHANNEL f1 =====
NUC1 13C
PULPROG zgpg30
PCPDPRG2 f2
TD 65536
SOLVENT CDCl3
NS 16
DS 4
F2 101.626126 MHz
FIDRES 0.1322500 Hz
AQ 4.0894900 sec
RG 492.12
AQ 4.0894900 sec
DE 18.140 MHz
TE 300.2 K
DE 17.00 MHz
TE 300.2 K
SI 1.0000000 sec
SI 1

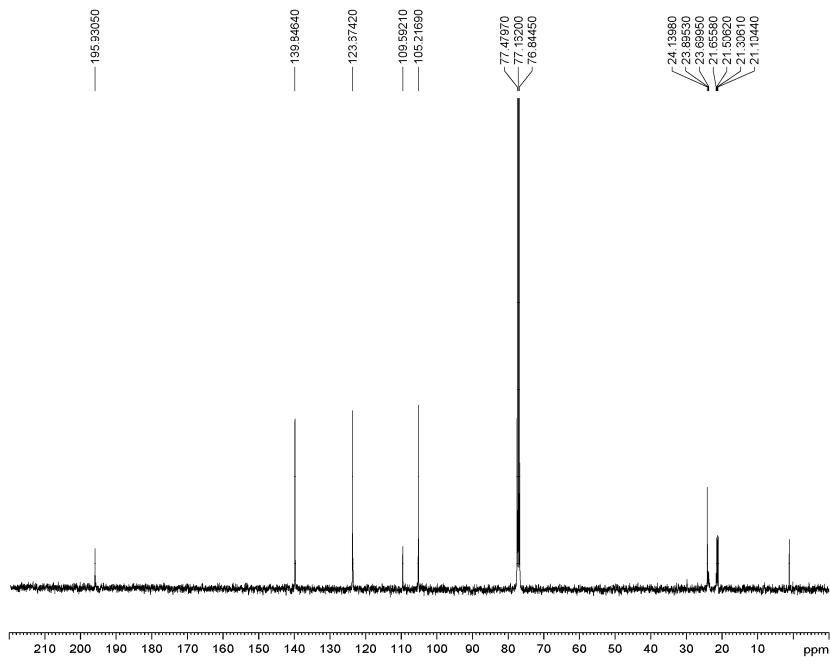
```



```

NAME 1H1347-9-3-CCl4-1120-phase-in-CDCl3-again
EXPNO 12
PROCNO 1
DATE_ 20191118
TIME 21:08
PROBHD 5 mm PABBO BB1
PULPROG zgpg30
PCPDPRG2 f2
TD 65536
SOLVENT CDCl3
NS 16
DS 4
F2 101.626126 MHz
FIDRES 0.1322500 Hz
AQ 4.0894900 sec
RG 492.12
AQ 4.0894900 sec
DE 18.140 MHz
TE 300.2 K
DE 17.00 MHz
TE 300.2 K
SI 1.0000000 sec
SI 1
===== CHANNEL f1 =====
NUC1 13C
PULPROG zgpg30
PCPDPRG2 f2
TD 65536
SOLVENT CDCl3
NS 16
DS 4
F2 101.626126 MHz
FIDRES 0.1322500 Hz
AQ 4.0894900 sec
RG 492.12
AQ 4.0894900 sec
DE 18.140 MHz
TE 300.2 K
DE 17.00 MHz
TE 300.2 K
SI 1.0000000 sec
SI 1

```

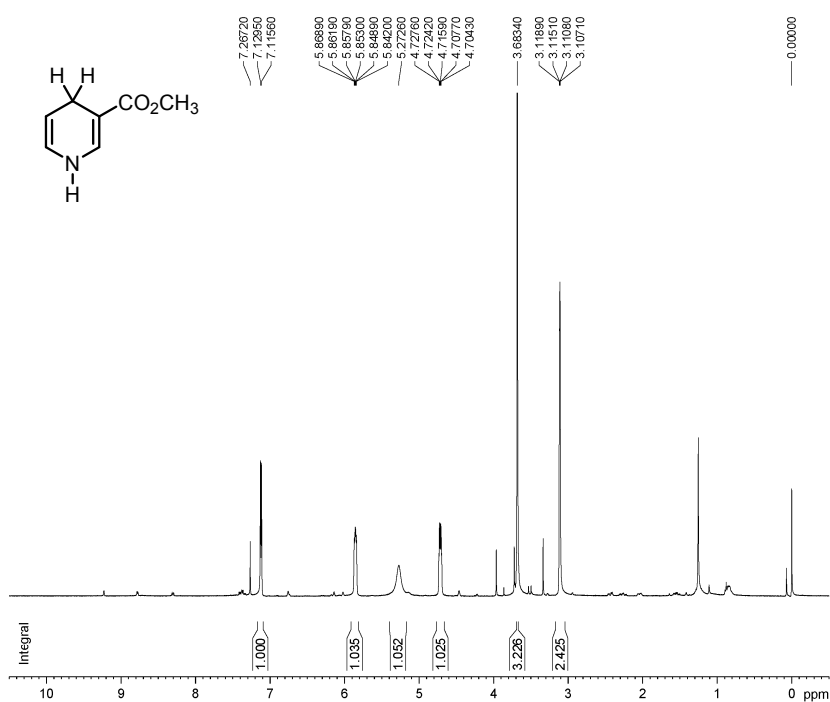


```

NAME 1M147-8-1-3-methyl-8-(2-oxo-2-phenylethyl)phenylacetate
EXPNO 1
PROCNO 1
Data_ 20201118
INSTRUM spect
PROBHD 5 mm PABBO 5BZ
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 1024
DS 4
SWH 24538.461 Hz
FIDRES 0.365798 Hz
AQ 1.361189 sec
RG 1081.1
AQ 20.150 usec
DE 0.50 usec
TE 302.0
D1 1.00000000 sec
d11 0.00000000 sec
TDO 1
===== CHANNEL f1 =====
NUC1 13C
P1 9.93 usec
PL1 0.00 dB
NUC2 1H
P2 12.50 usec
PL2 19.00 dB
NUC3 13C
P3 9.93 usec
PL3 0.00 dB
=====
PC 1.40

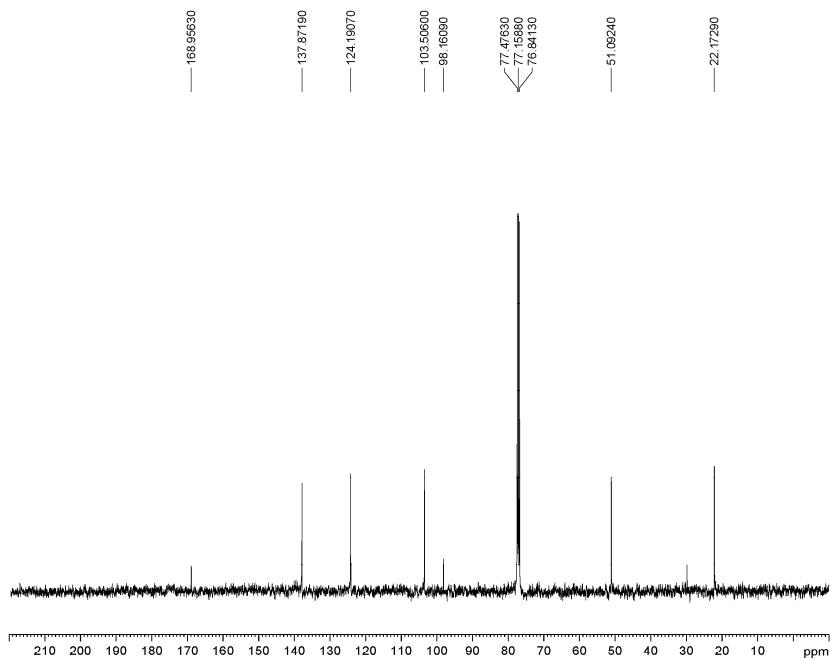
```

¹H and ¹³C NMR of **6f**:



```

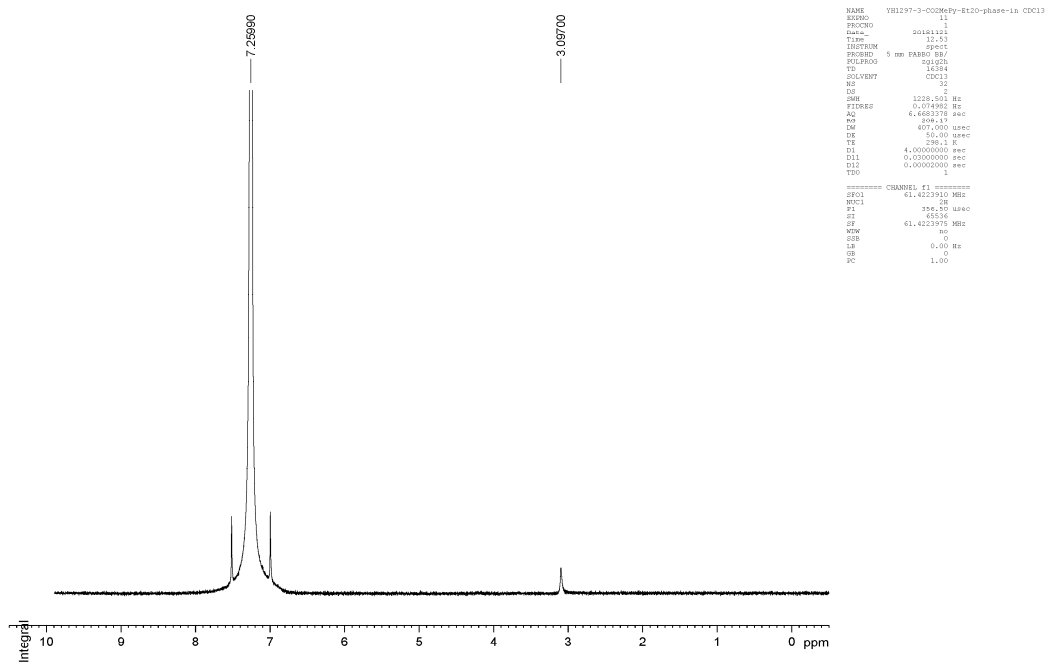
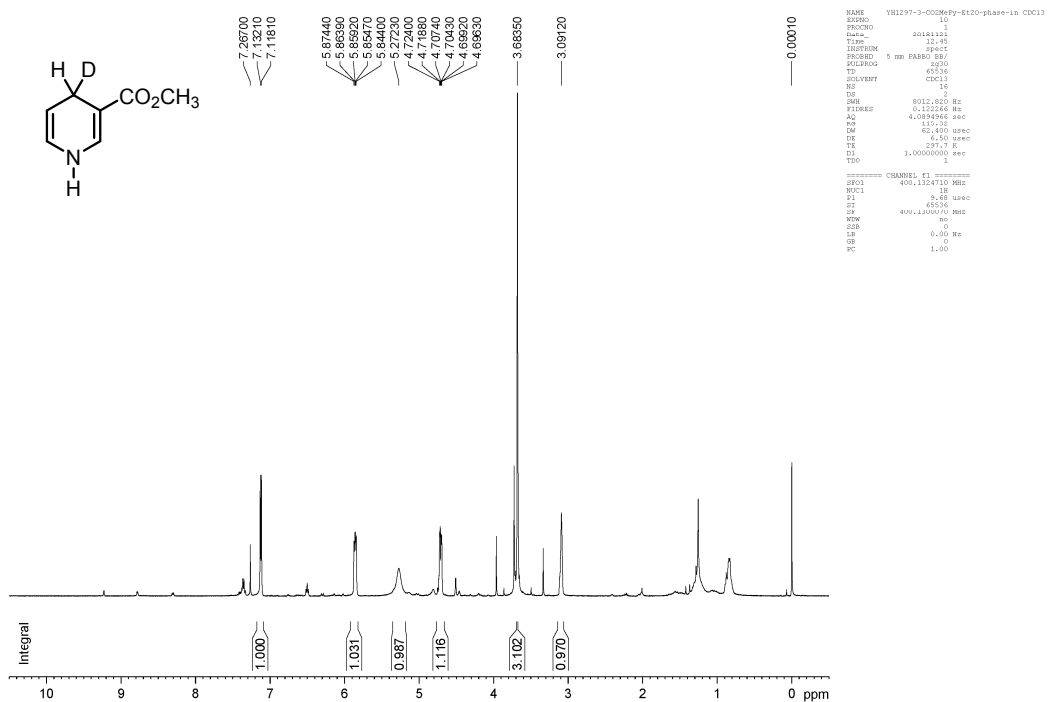
NAME YH1245-3-BL20-phase-1x-CDCl3-1st
EXPNO 10
PROCNO 11
Date_ 20181005
Time 14.20
INSTRUM spect
PROBHD 5 mm PABBO BB
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8012.028 Hz
FIDRES 0.122266 Hz
AQ 4.0034960 sec
RG 101.33
DM 62.400 usec
DE 6.50 usec
TE 298.2 K
D1 1.00000000 sec
D11 0.00000000 sec
TD0 1
===== CHANNEL f1 =====
SFO1 400.1324710 MHz
NUC1 13
P1 9.48 usec
SF 400.130085 MHz
WDM no
SFO 0
GB 0.00 Hz
PC 1.00
    
```

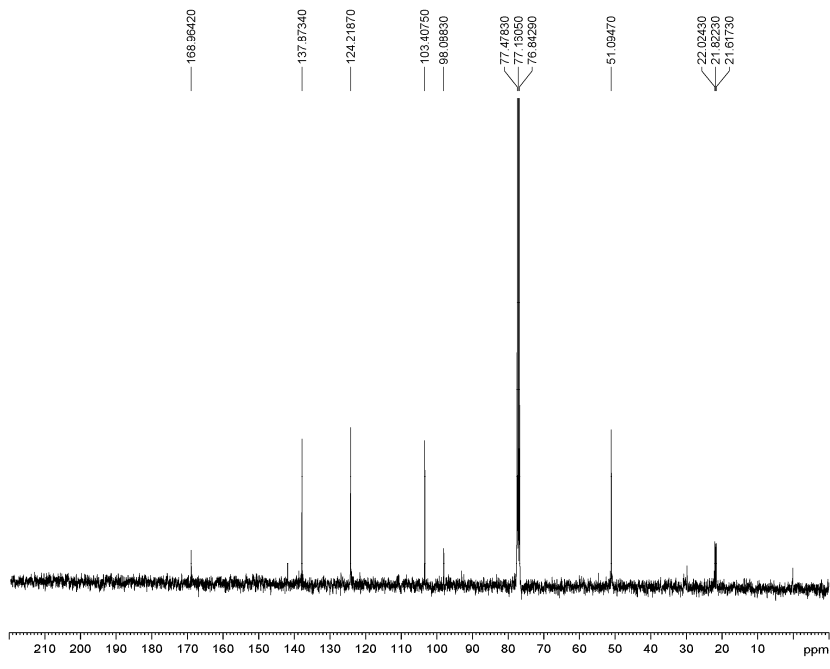


```

NAME YH1245-3-BL20-phase-1x-CDCl3-1st
EXPNO 12
PROCNO 11
Date_ 20181005
Time 14.13
INSTRUM spect
PROBHD 5 mm PABBO BB
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 300
DS 2
SWH 24038.461 Hz
FIDRES 0.346778 Hz
AQ 1.3631980 sec
RG 204.17
DM 20.800 usec
DE 6.40 usec
TE 298.2 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1
===== CHANNEL f1 =====
SFO1 100.6228290 MHz
NUC1 13
P1 9.48 usec
SF 100.6127546 MHz
WDM no
SFO 0
GB 4.00 Hz
PC 1.40
    
```

^1H , ^2H , and ^{13}C NMR of **6f-d**:



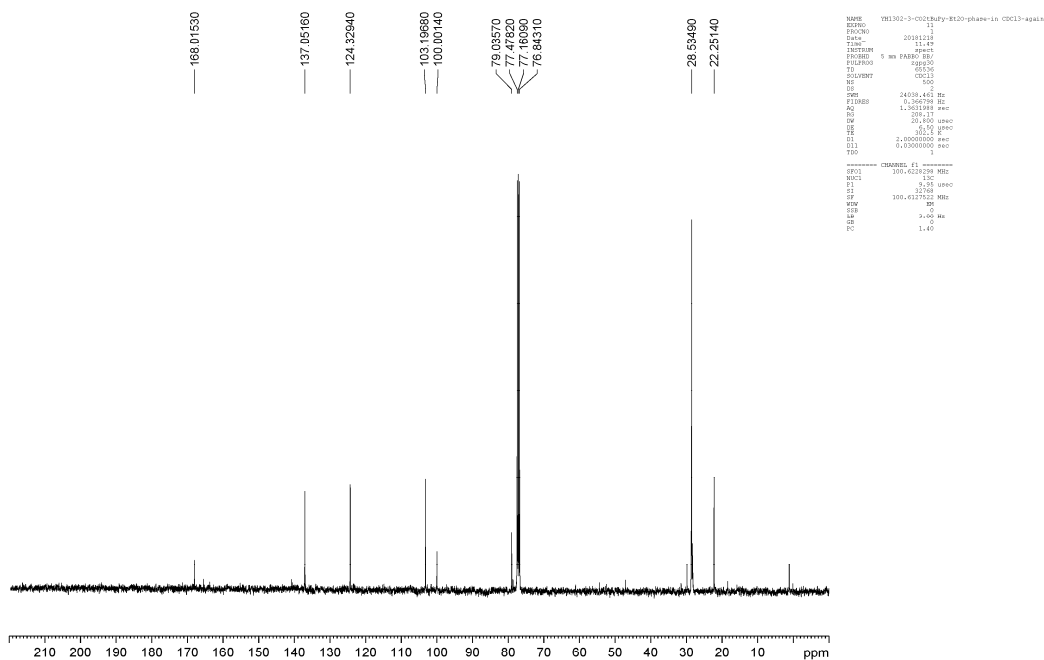
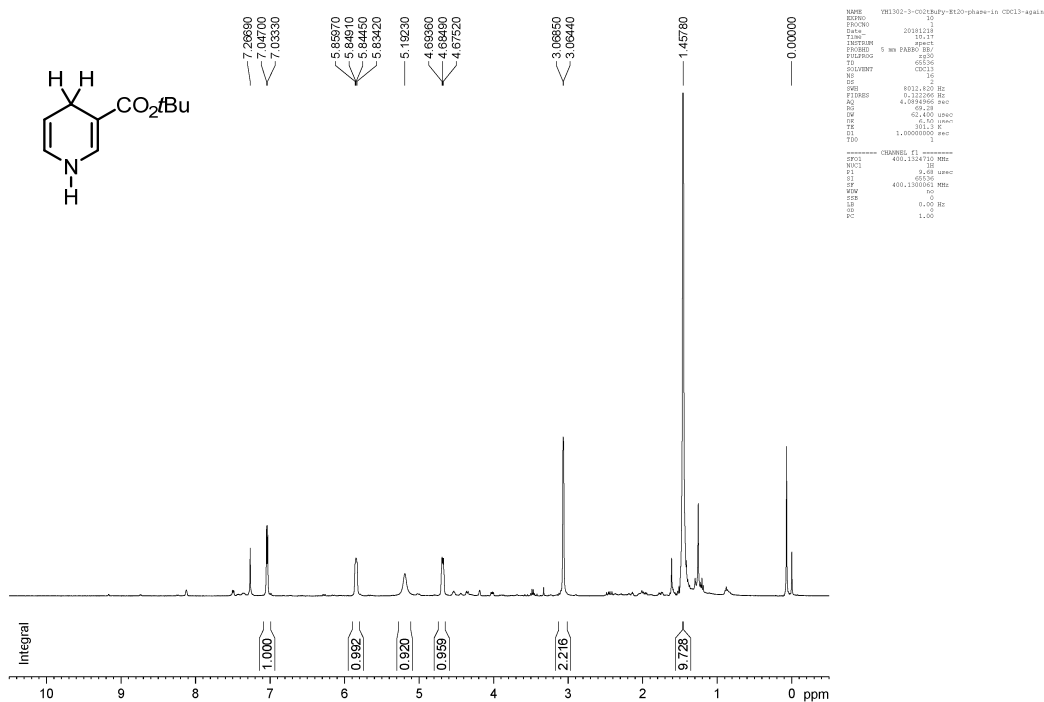


```

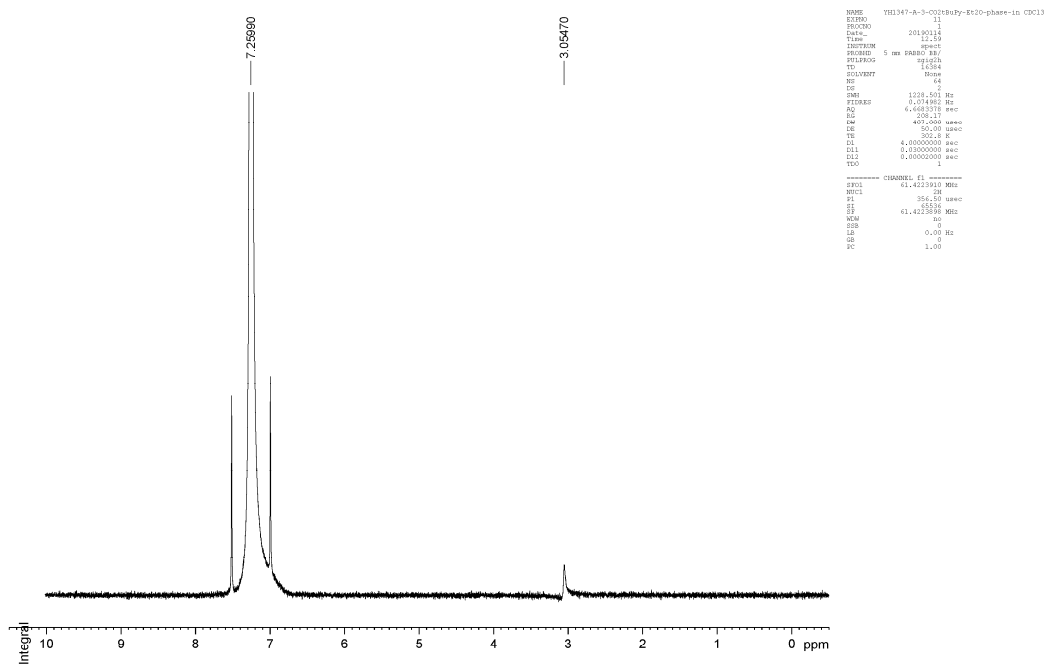
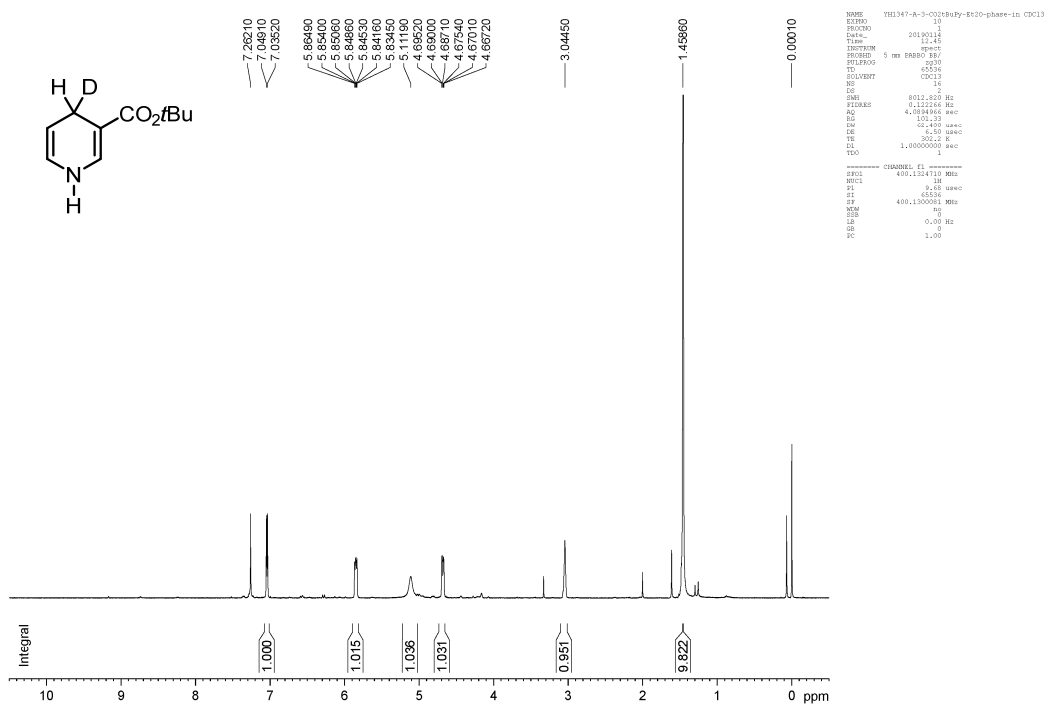
NAME      VB1297-3-000269y-R120-phase-in CDC13
EXPNO     2
PROCNO    1
EXPNAME   00481111
F2 -      131.17
INSTRUM   spect
PROBHD    5 mm BBOBO BB7
PULPROG   zgpg30
TD         65536
SOLVENT   CDC13
NS         2
DS         4
AQ         0.346798 Hz
FIDRES    1.363198 sec
RG         26140
DM         20.690 usec
DE         6.50 usec
TE         298.5 K
D1         2.00000000 sec
D11        0.03000000 sec
TD0        1
----- CHANNEL f1 -----
FREQ1     100.628238 MHz
NUC1      13C
P1         8.30 usec
PI         26140
PT         100.612758 MHz
WDW        EM
SSB        0
LB         3.00 Hz
GB         0
PC         1.40

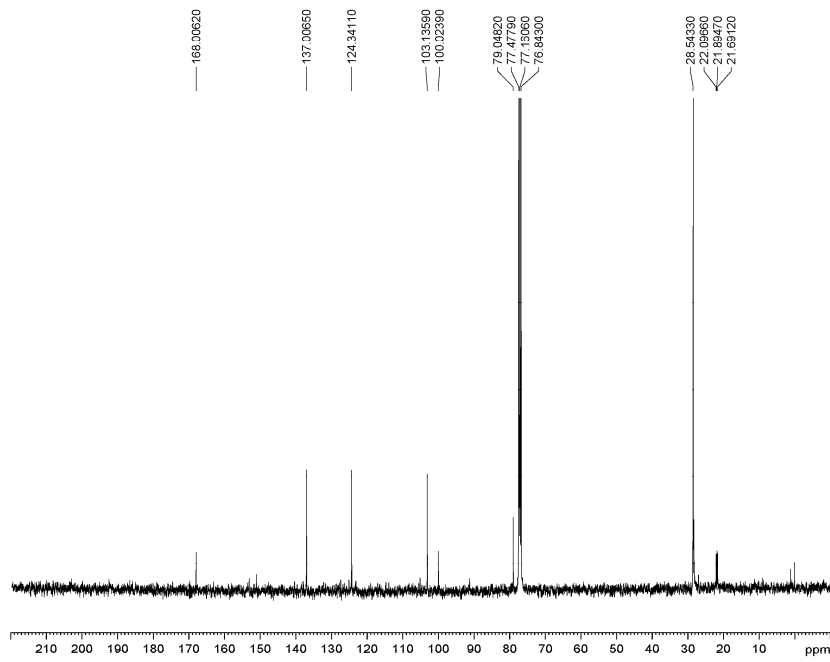
```

¹H and ¹³C NMR of **6g**:



¹H, ²H, and ¹³C NMR of **6g-d**:



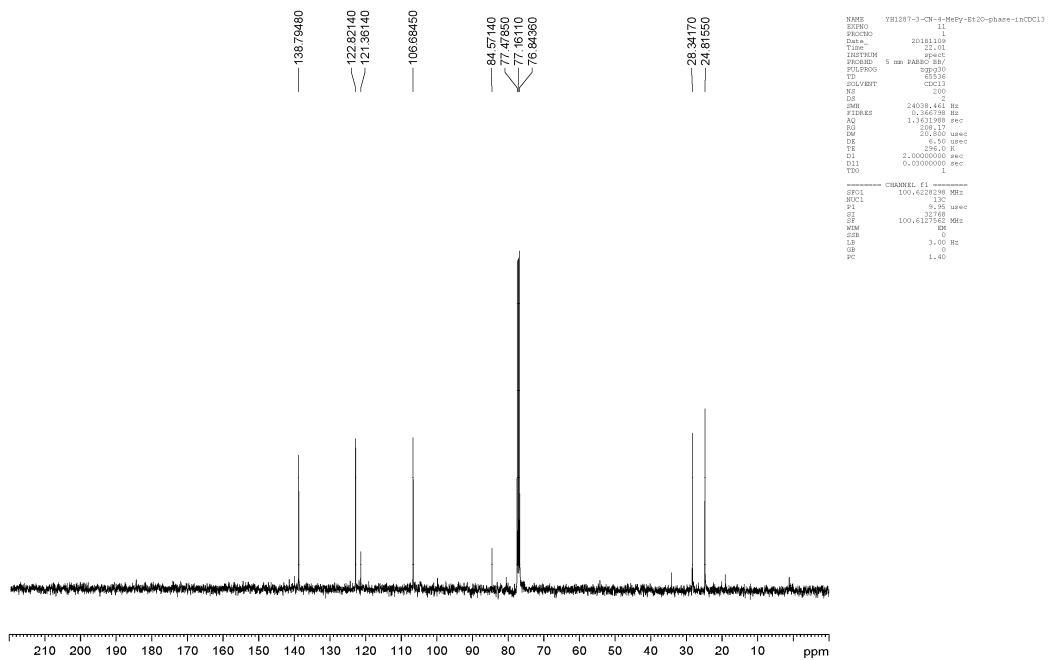
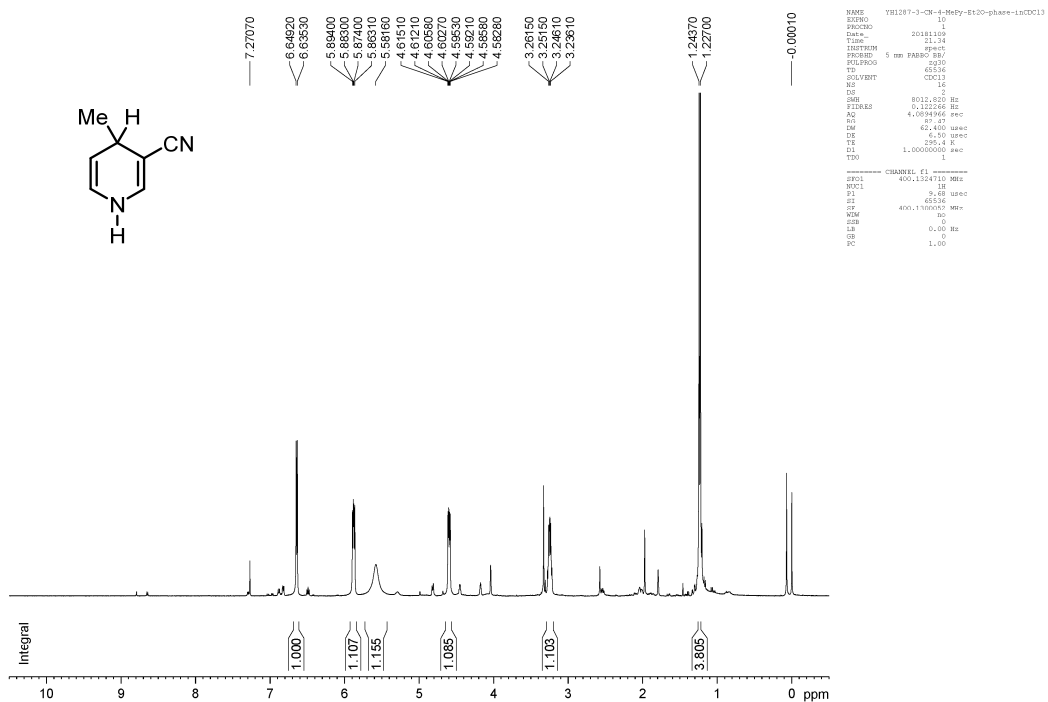


```

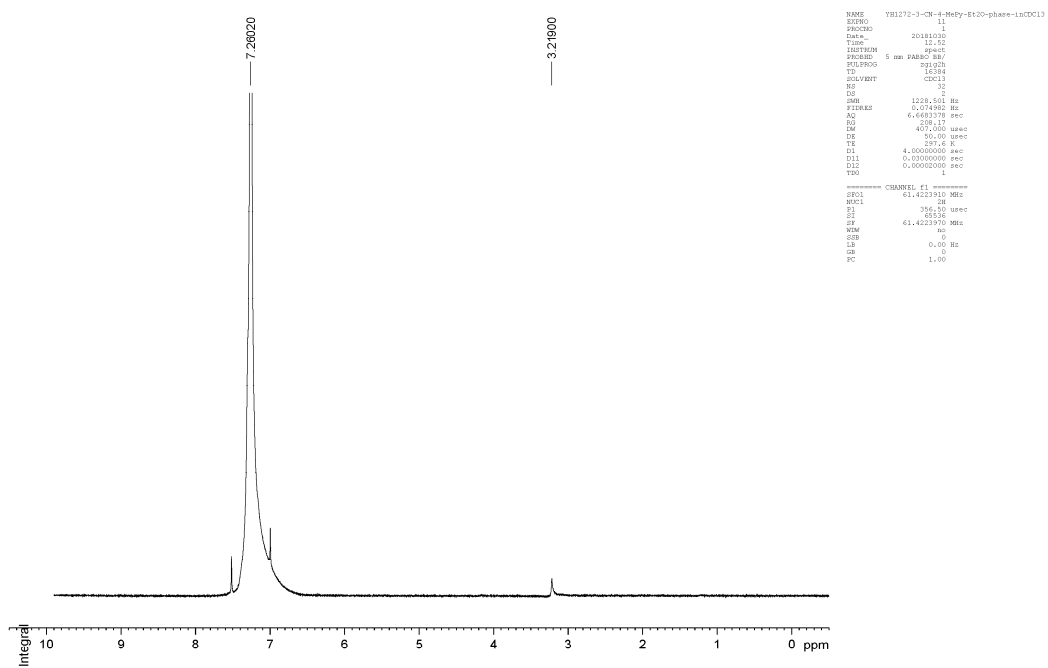
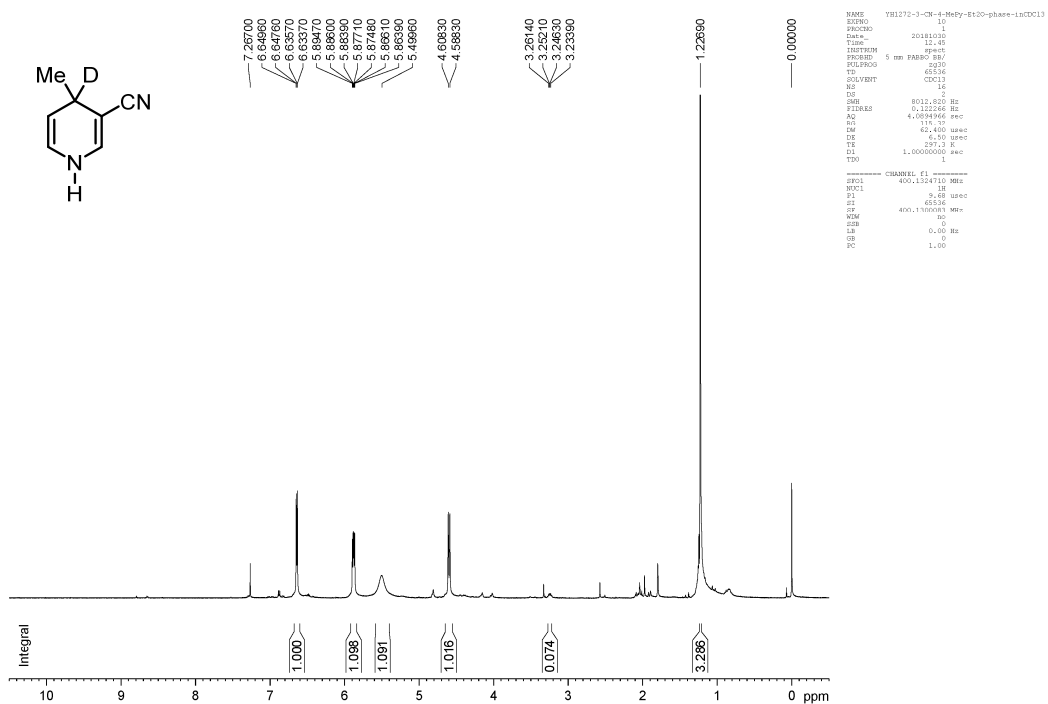
NAME  VHS147-A-3-oxobutyl-ethyl-phosphate-in-CDCl3
EXPNO  1
PROCNO  1
Date_  20100115
Time  12.20
INSTRUM  spect
PROBHD  5 mm BBOBO 1H/
PULPROG  zgpg30
TD  65536
SOLVENT  CDCl3
NS  1512
DS  4
SWH  24038.461 Hz
FIDRES  0.361000 Hz
AQ  1.3821000 sec
RG  1024
AQ  1.3821000 sec
SFO  100.6261300 MHz
DE  200.000 usec
TE  300.2 K
CE  6.50 usec
D1  2.00000000 sec
d11  0.03000000 sec
TD0  1
----- CHANNEL f1 -----
PRFO  100.6261300 MHz
NUC1  13C
P1  0.95 usec
PI  32700
NUC2  1H
P2  0
PC  3.00 Hz
PC  1.40

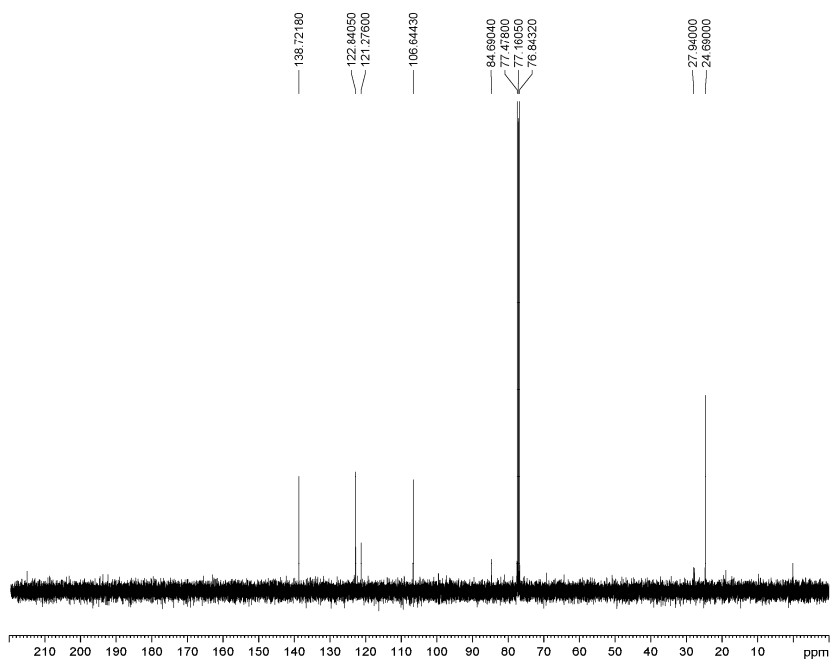
```

¹H and ¹³C NMR of **6h**:



¹H, ²H, and ¹³C NMR of **6h-d**:



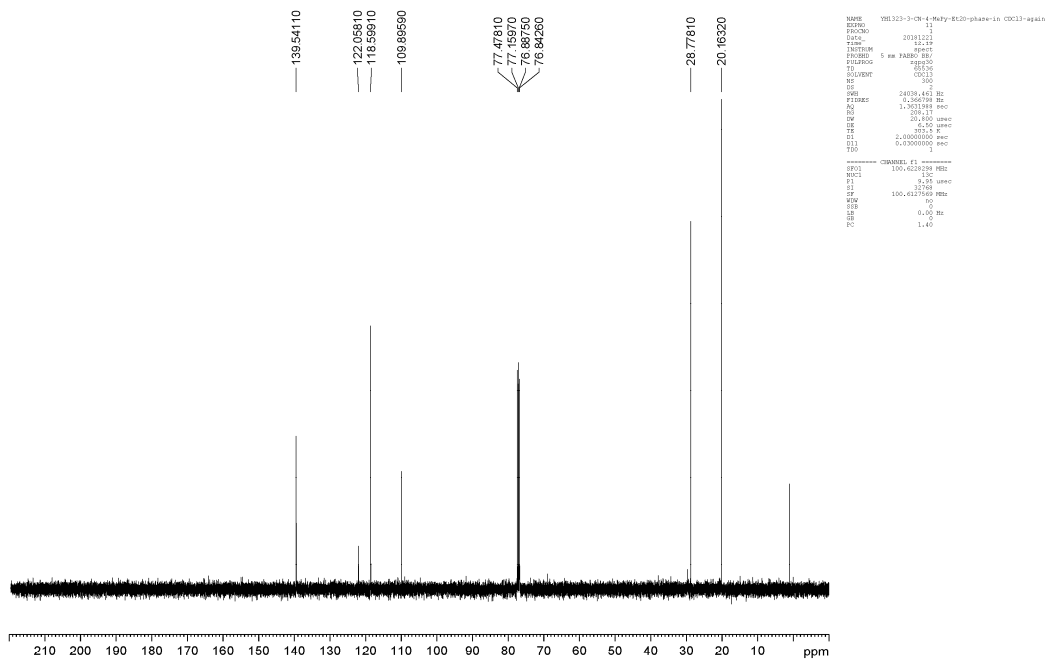
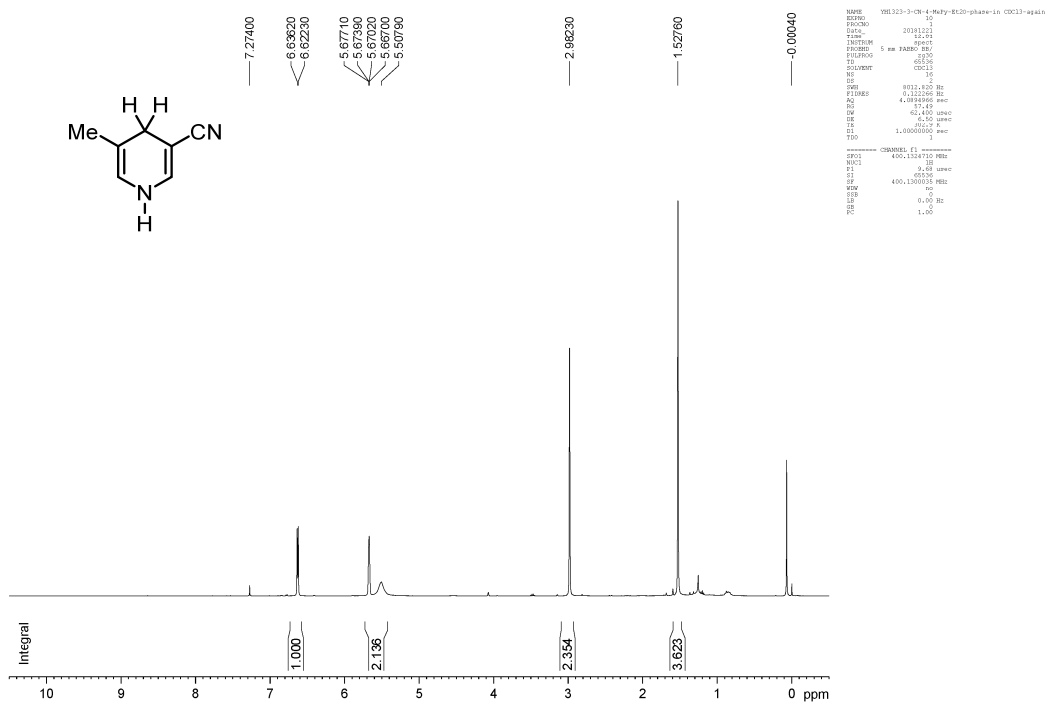


```

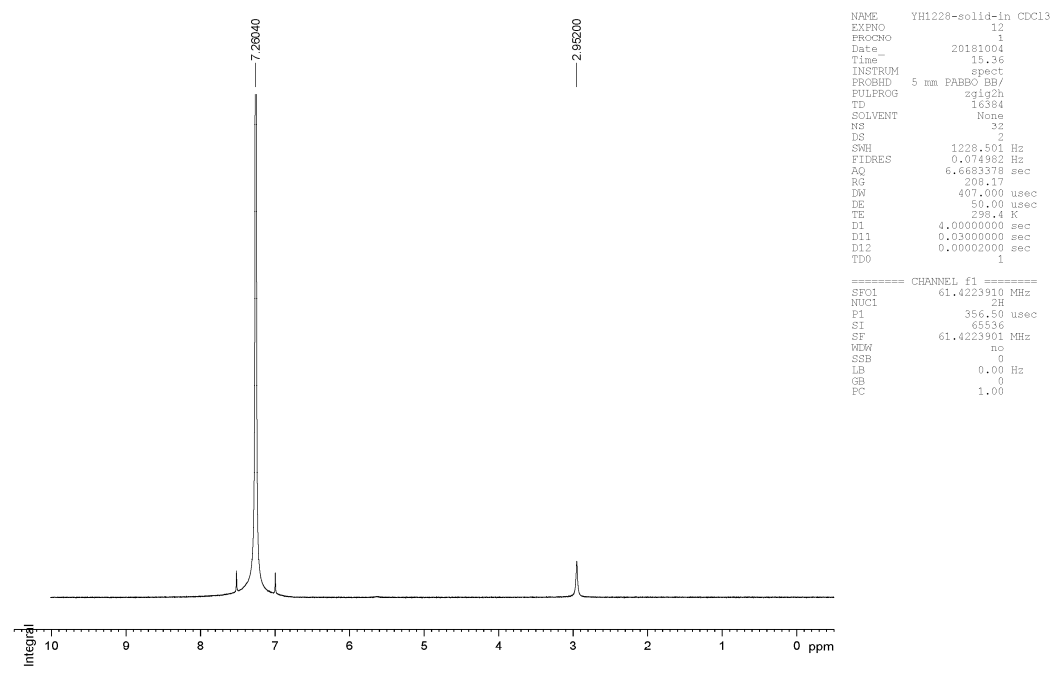
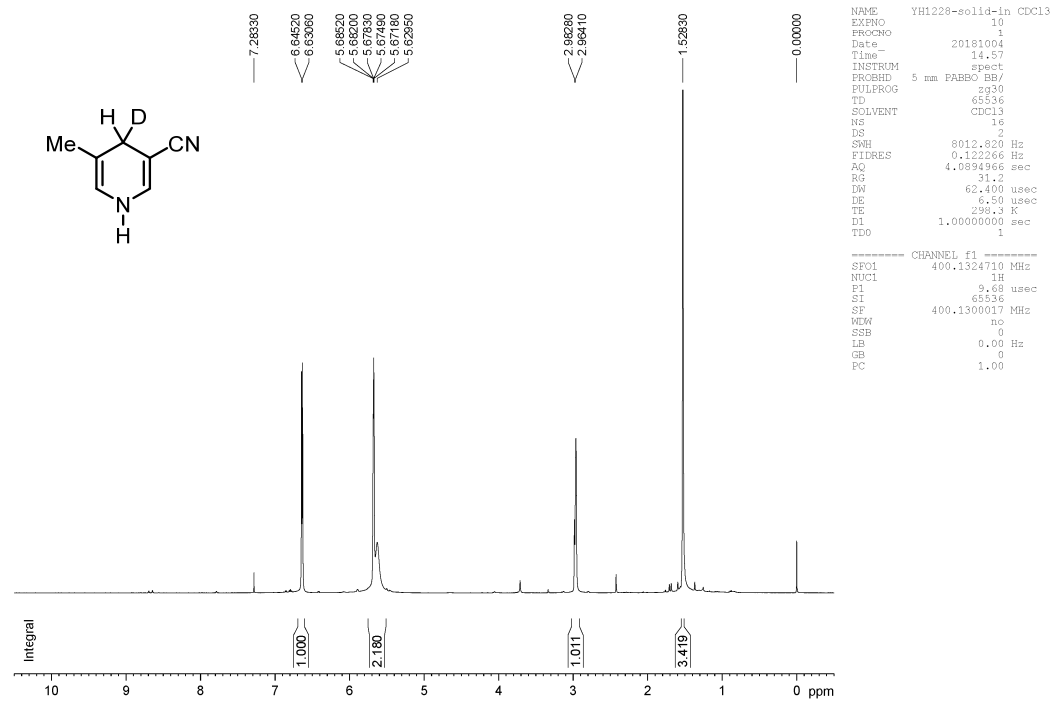
NAME  YB1272-3-CN-4-MeTy-Pr2O-phase-1a(CDCl3)
EXPNO  2
PROCNO  20411000
Time  13.41
INSTRUM  spect
PROBHD  5 mm QNP300 1H/
PULPROG  zgpg30
TD  65536
SOLVENT  CDCl3
NS  800
DS  4
SFR  24038.461 Hz
FIDRES  0.286758 Hz
AQ  1.1621988 sec
RG  258.17
DM  20.800 usec
DE  6.50 usec
TE  298.1 K
D1  2.00000000 sec
D11  0.03000000 sec
TD0  1
----- CHANNEL f1 -----
NUC1  13C
P1  120
PL1  120 usec
PT  1770
RF  100.6131449 MHz
MVM  0
SFO  0
GB  0
PC  1.40

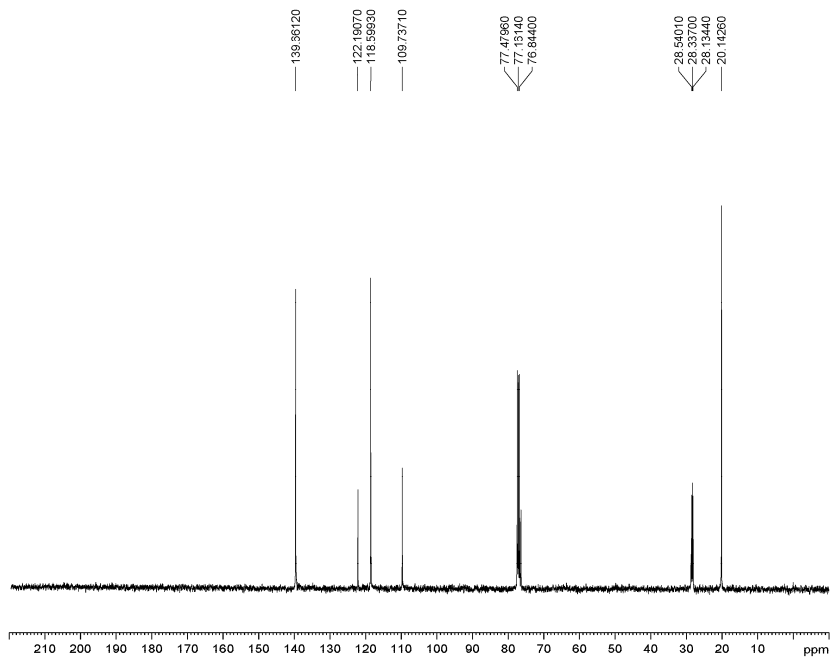
```

¹H and ¹³C NMR of **6i**:



¹H, ²H, and ¹³C NMR of 6i-d:





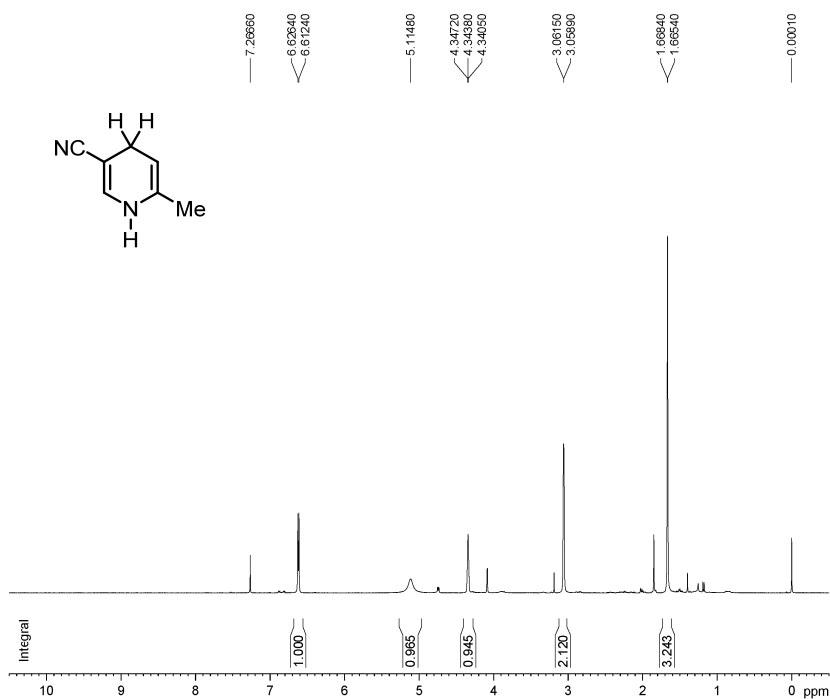
```

NAME      YH1228-solid-1n CDCl3
EXPNO     13
PROCNO    1
Date_     20181004
Time      15.52
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         300
DS         2
SMH       24038.461 Hz
FIDRES    0.366799 Hz
AQ         1.3631988 sec
RG         208.17
DN         20.800 usec
DE         6.50 usec
TE         298.6 K
D1         2.0000000 sec
D11        0.0300000 sec
TDO        1

===== CHANNEL f1 =====
SFO1      100.6228299 MHz
NUC1      13C
P1         9.25 usec
SI         32768
SF         100.6127613 MHz
WEW       EM
SSB        0
LB         3.00 Hz
GB         0
PC         1.40

```


¹H and ¹³C NMR of **6j**:

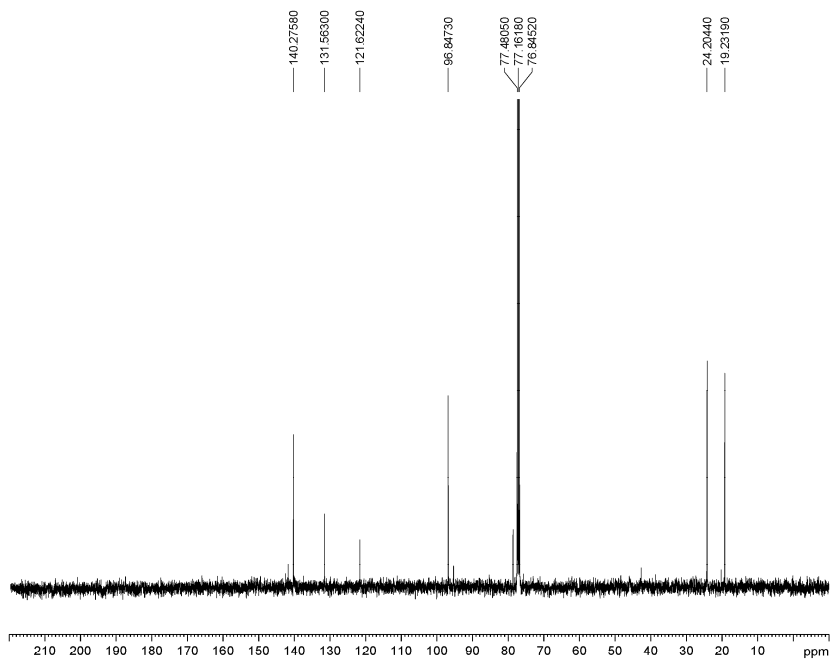


```

NAME      VH1358-A-2-Me-5-CNFy-In CDC13
EXPNO    1
PROCNO   1
Date_    20190423
Time     16.07
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zg30
TD        65536
SOLVENT  CDCl3
NS        16
DS        2
SWH       8012.820 Hz
FIDRES    0.232266 Hz
AQ         4.0884866 sec
RG         101.53
LW         62.400 usec
DE         6.50 usec
TE         300.3 K
D1         1.00000000 sec
D11        1
D12        1
D13        1
D14        1
D15        1
D16        1
D17        1
D18        1
D19        1
D20        1
  
```

```

===== CHANNEL f1 =====
SFO1    400.1324710 MHz
NUC1     1H
P1       9.68 usec
PT       65536
SF       400.1300059 MHz
RG        0
SSB       0
LB        0.00 Hz
GB         0
PC         1.00
  
```



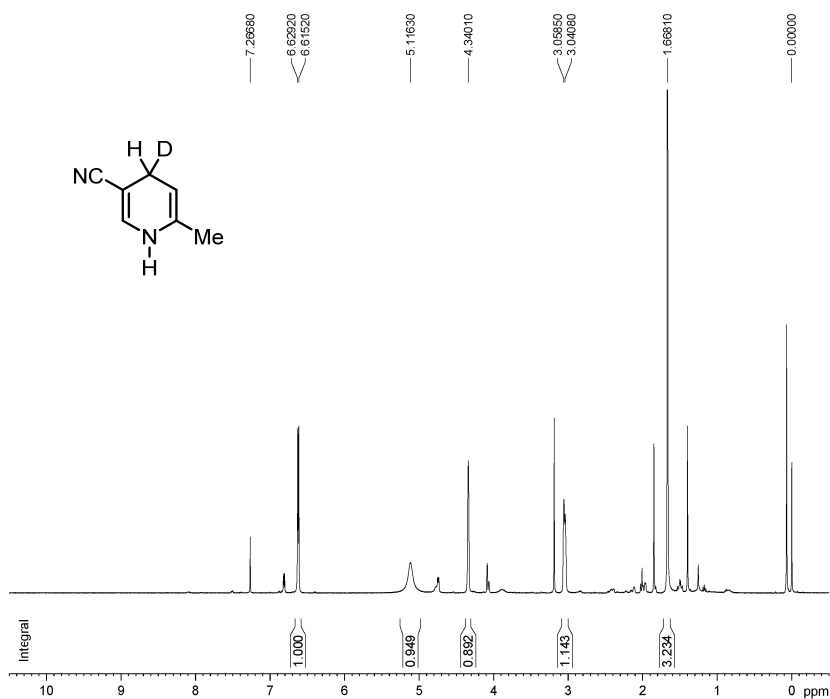
```

NAME      VH1358-A-2-Me-5-CNFy-In CDC13
EXPNO    11
PROCNO   1
Date_    20190423
Time     17.25
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpg30
TD        65536
SOLVENT  CDCl3
NS        210
DS        2
SWH       24038.461 Hz
FIDRES    0.268776 Hz
AQ         1.3031988 sec
RG         208.17
LW         20.400 usec
DE         6.50 usec
TE         300.3 K
D1         2.00000000 sec
D11        0.03000000 sec
D12        1
D13        1
D14        1
D15        1
D16        1
D17        1
D18        1
D19        1
D20        1
  
```

```

===== CHANNEL f1 =====
SFO1    100.628298 MHz
NUC1     13C
P1       9.95 usec
PT       65536
SF       100.6127538 MHz
RG        0
SSB       0
LB        2.00 Hz
GB         0
PC         1.40
  
```

¹H, ²H, and ¹³C NMR of **6j-d**:

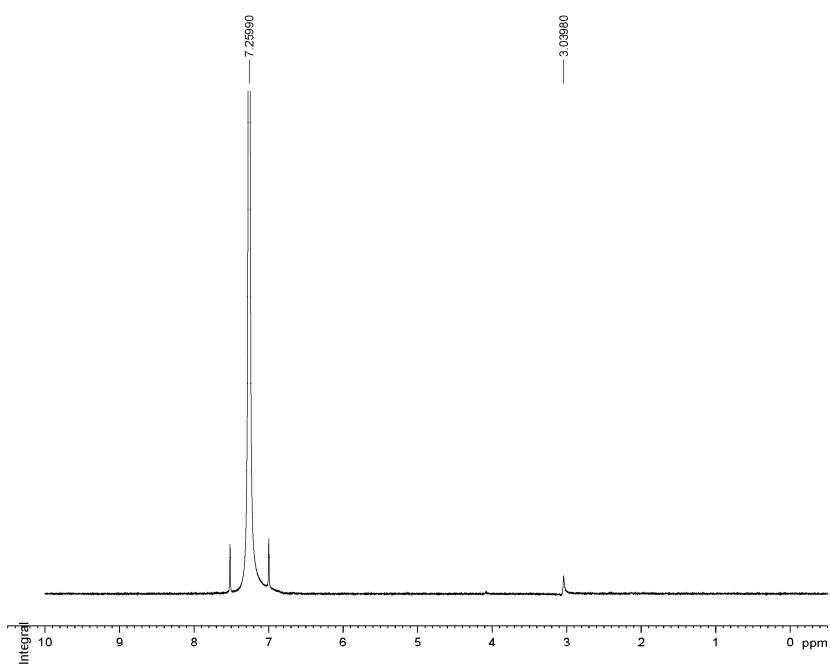


```

NAME YH3112-2-Me-5-CNPy-E120-phase-in-CDCl3
EXPNO 11
PROCNO 11
DATE_ 20161207
TIME 15.10
INSTRUM spect
PROBHD 5 mm BBOBO 3H/
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 32
DS 2
SWH 8002.620 Hz
FIDRES 0.122266 Hz
AQ 4.0004962 sec
RG 312.25
EQ 6.50 usec
TE 299.2 K
DE 1.0000000 sec
TD0 1.0000000 sec

----- CHANNEL f1 -----
NUC1 400.1324710 MHz
P1 18
SFO1 400.1324710 MHz
PC 1.00

```

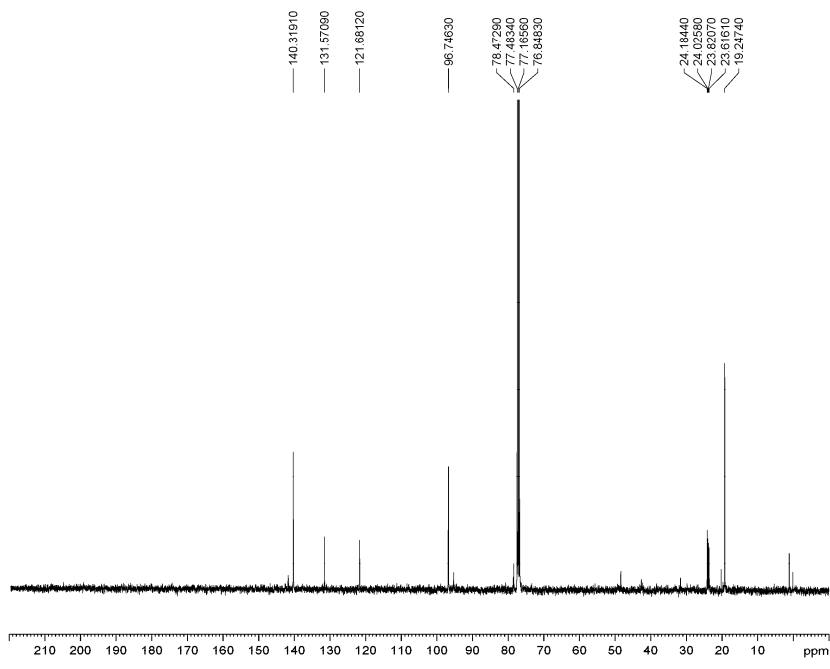


```

NAME YH3112-2-Me-5-CNPy-E120-phase-in-CDCl3
EXPNO 11
PROCNO 11
DATE_ 20161207
TIME 15.10
INSTRUM spect
PROBHD 5 mm BBOBO 3H/
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 32
DS 2
SWH 1256.500 Hz
FIDRES 0.074988 Hz
AQ 6.668310 sec
RG 312.25
EQ 50.00 usec
TE 299.2 K
DE 1.0000000 sec
TD0 1.0000000 sec

----- CHANNEL f1 -----
NUC1 61.4223910 MHz
P1 20
SFO1 61.4223910 MHz
PC 1.00

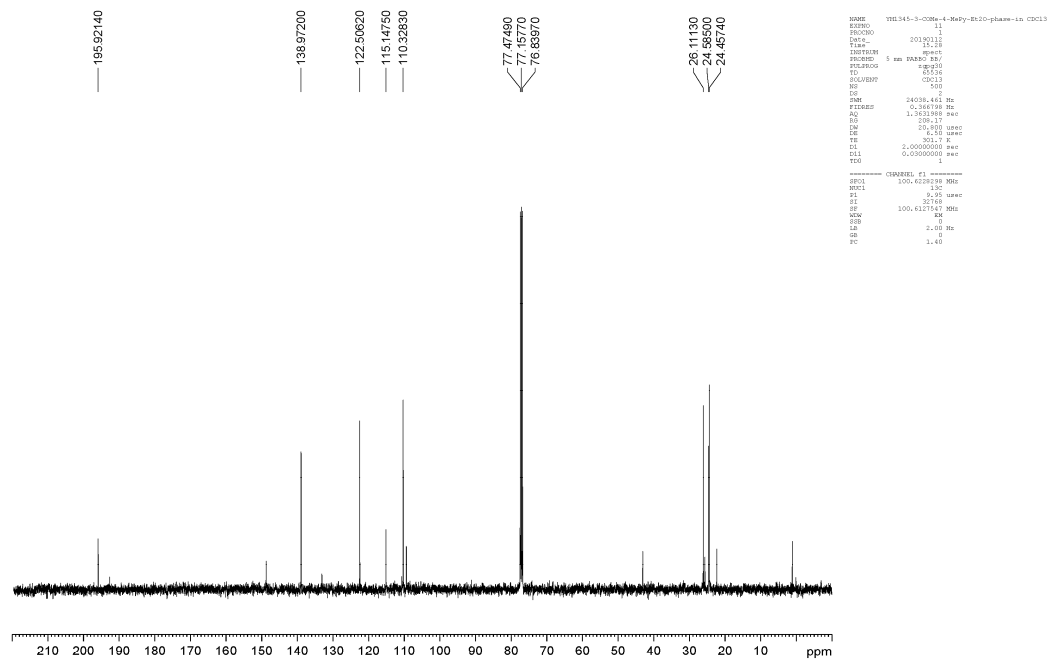
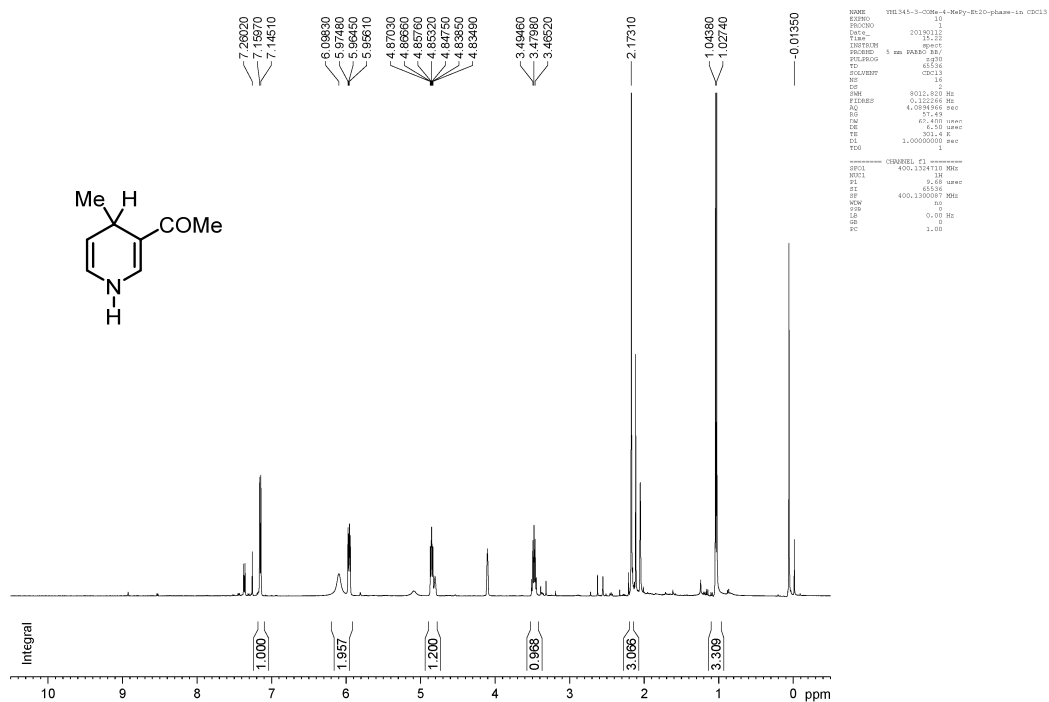
```



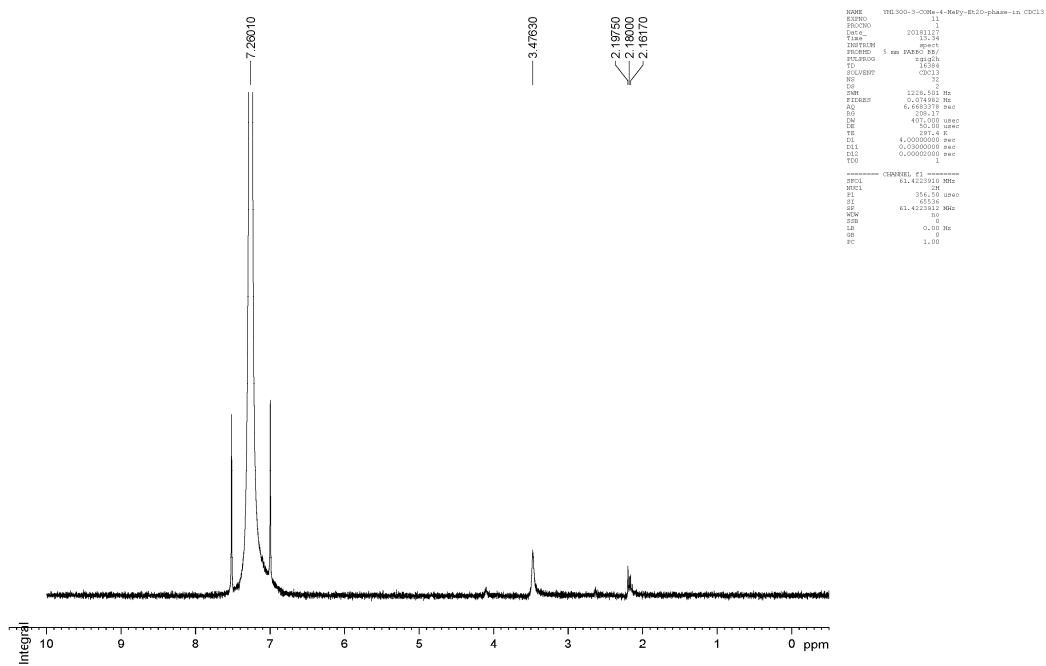
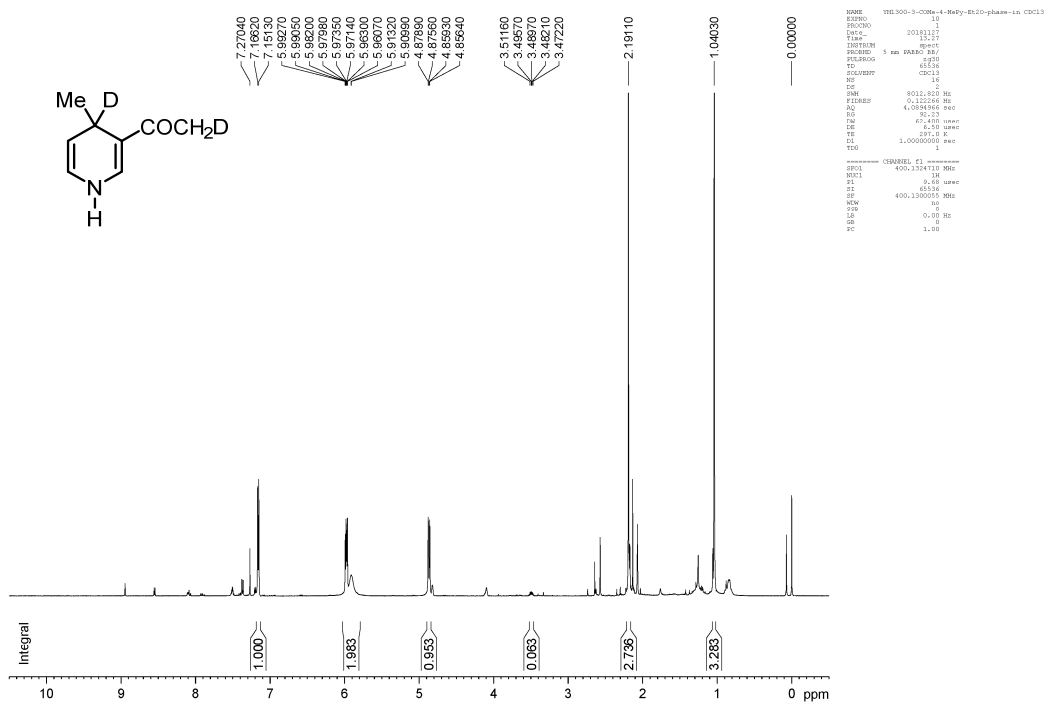
```

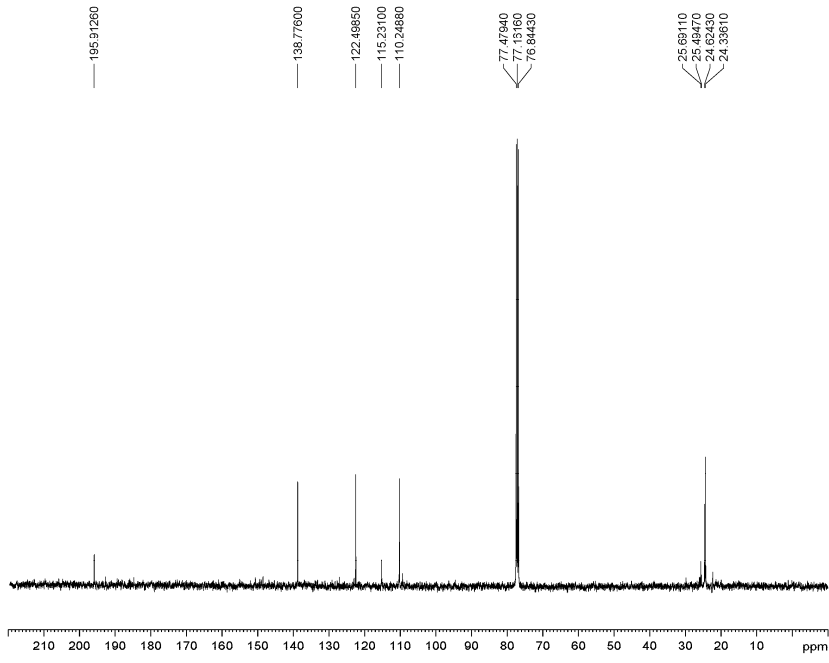
NAME VHS112-2-Me-5-OMe-B120-phase-in-CDCl3
EXPNO 2
PROCNO 1
F1 501.1764
TIME 15.00
INSTRUM spect
PROBHD 5 mm BBO-300
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 1024
DS 4
SWH 24398.461 Hz
FREQ 100.6261898 MHz
AQ 1.3631983 sec
RG 320.000
DE 20.000 usec
TE 300.2 K
D1 0.10000000 sec
d11 0.03000000 sec
TD0 1
----- CHANNEL f1 -----
NUC1 13C
P1 12.00 usec
PR 100.6155500 MHz
SFO 100.6261898 MHz
SF8 100.6155500 MHz
SEB 2.00 Hz
SB 1.40
  
```

¹H and ¹³C NMR of **6k**:



¹H, ²H, and ¹³C NMR of **6k-d**:

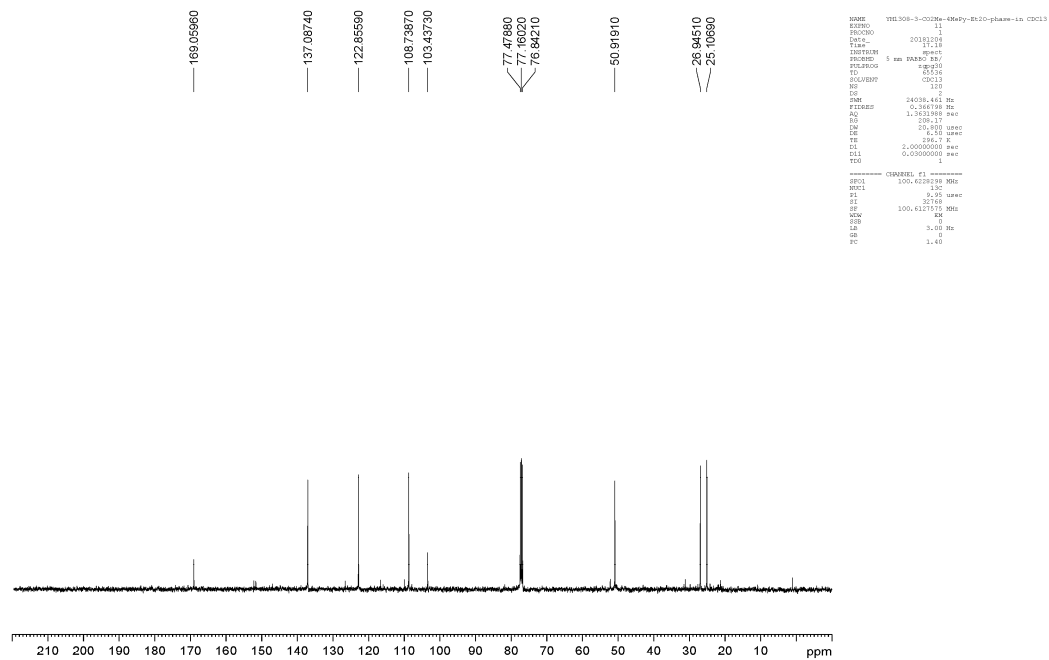
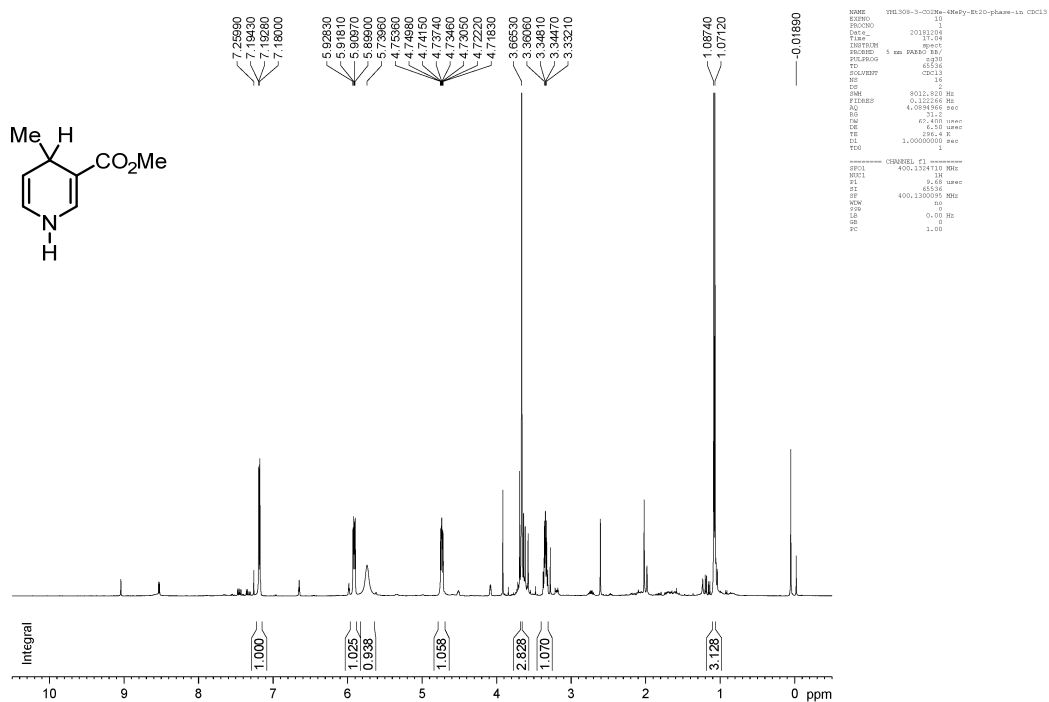




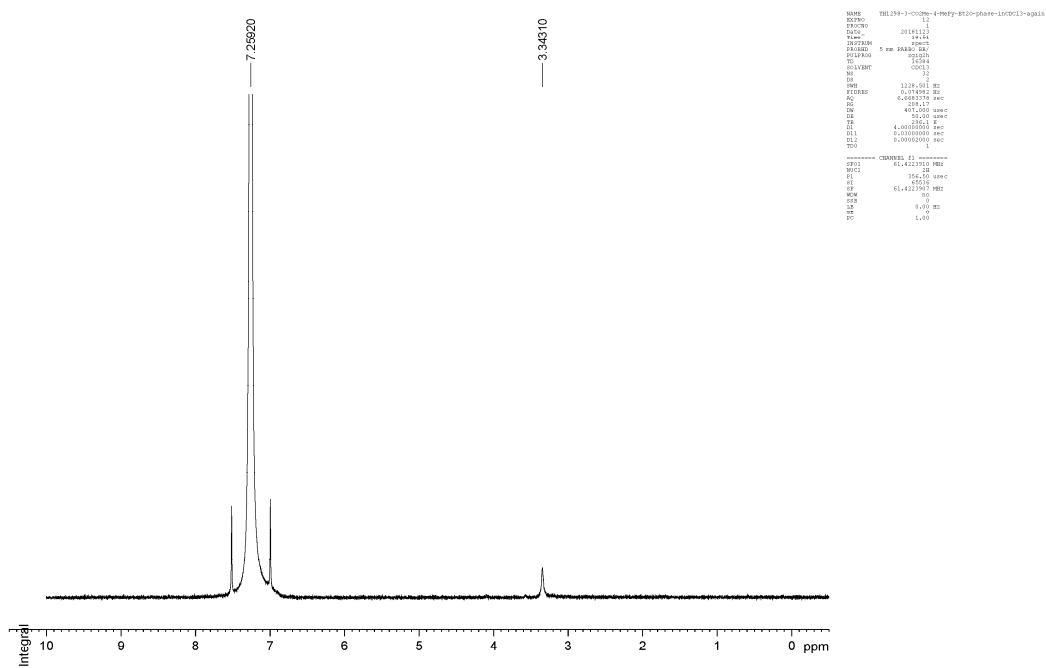
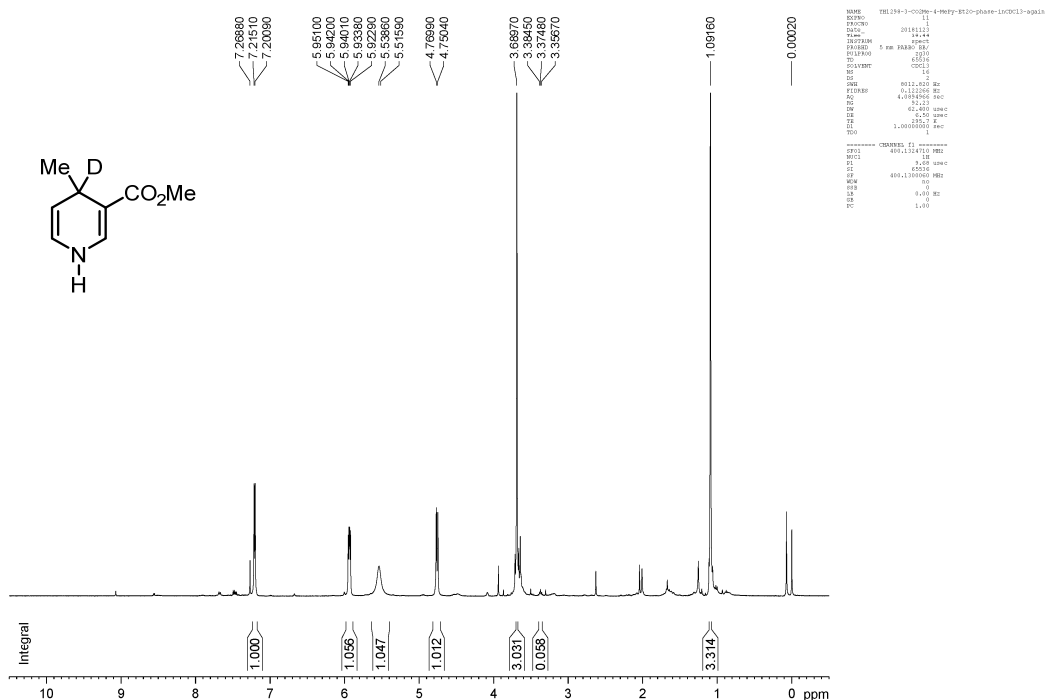
```
NAME: 5HE300-3-OH-m-4-BeHy-BE10-phase-in-CDCl3  
EXPNO: 1  
PROCNO: 2  
F2: 2018117  
F3: 19  
PROCDS: 1627  
PROCPS: 5 Hz PHS 20  
PROCSP: 8200  
ID: 6530  
SOLVENT: CDCl3  
NS: 400  
DS: 4  
AQ: 2409.461 Hz  
RG: 327.193 Hz  
SI: 327.193 Hz  
SF: 300.137 MHz  
FIDRES: 0.24500 Hz  
AQRES: 0.24500 MHz  
SFO: 25.500 MHz  
PT: 2.00000000 sec  
DQ: 2.00000000 sec  
DQL: 0.00000000 sec  
DQ2: 1  
DQ3: 1
```

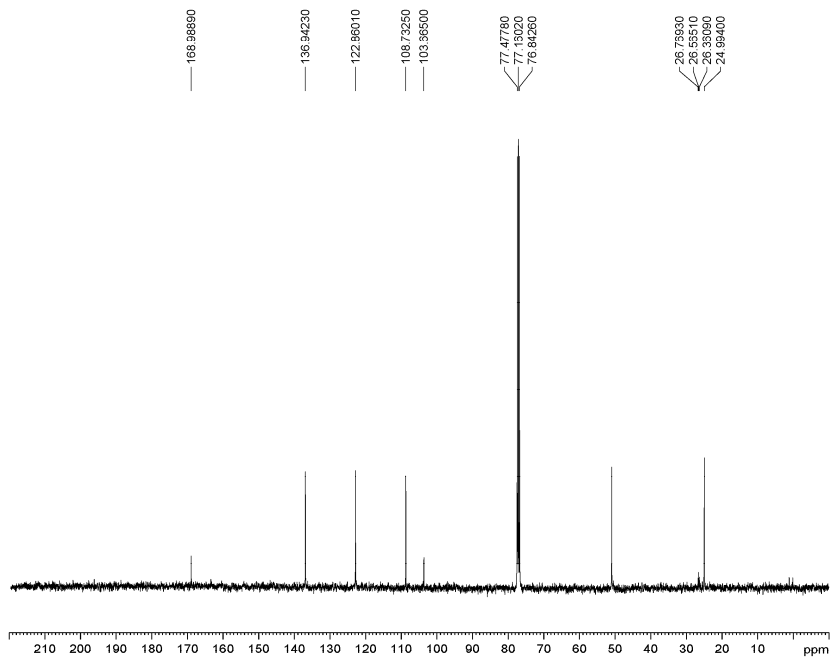
----- CHANNEL f1 -----
NUC1: 13C
P1: 9.00 Hz
PL1: 2.00 dB
NUC2: 1H
P2: 327.69 MHz
PL2: 0.00 dB
NUC3: 1H
P3: 3.00 Hz
PL3: 0.00 dB
PC: 1.00

¹H and ¹³C NMR of **6I**:



¹H, ²H, and ¹³C NMR of **6I-d**:



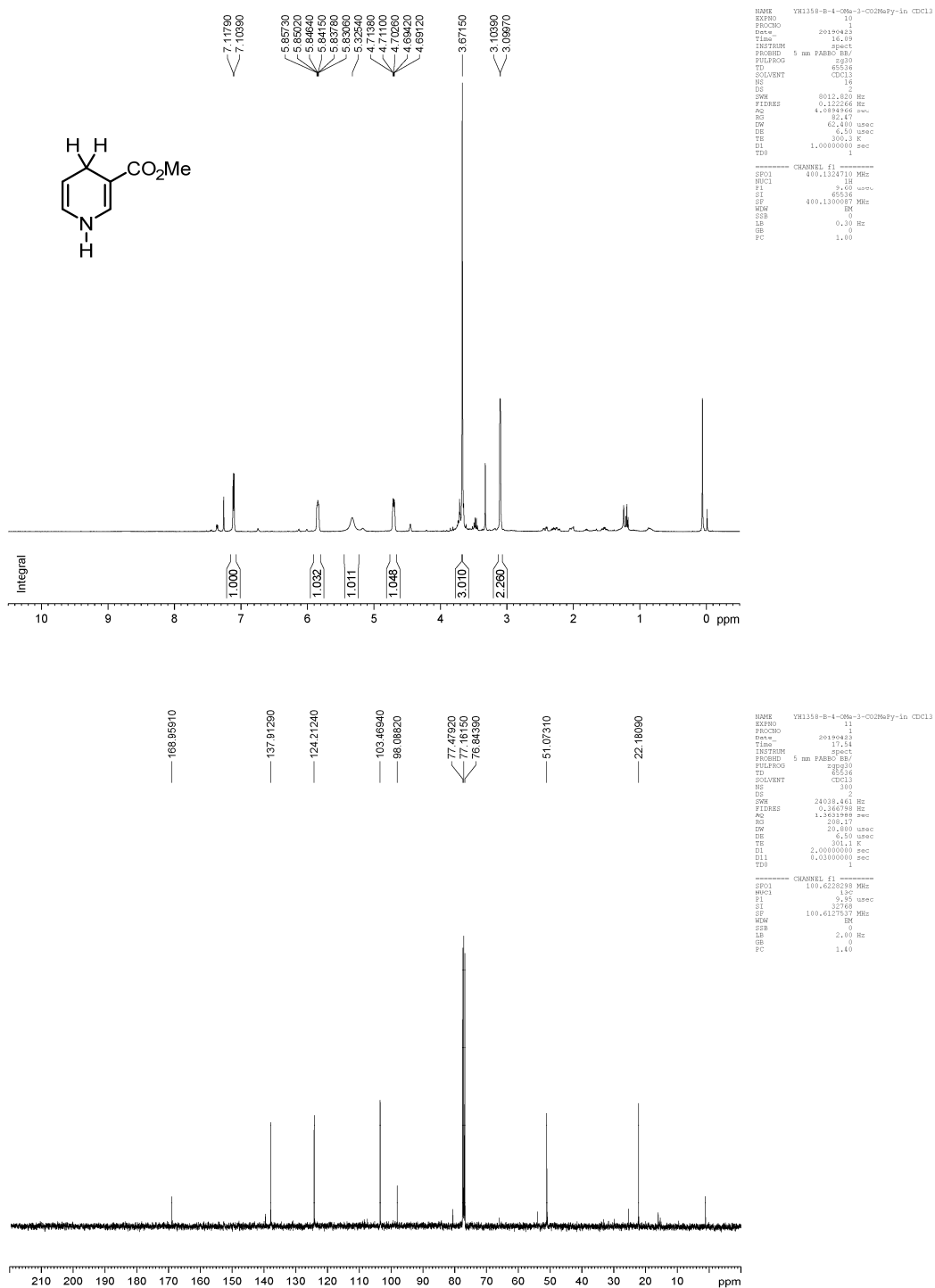


```

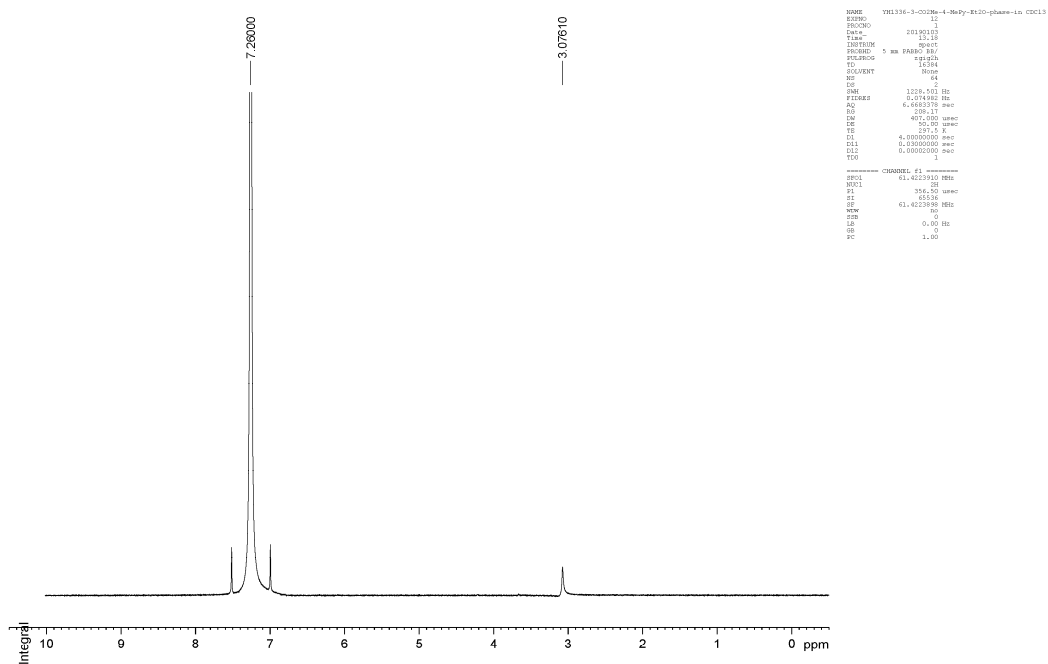
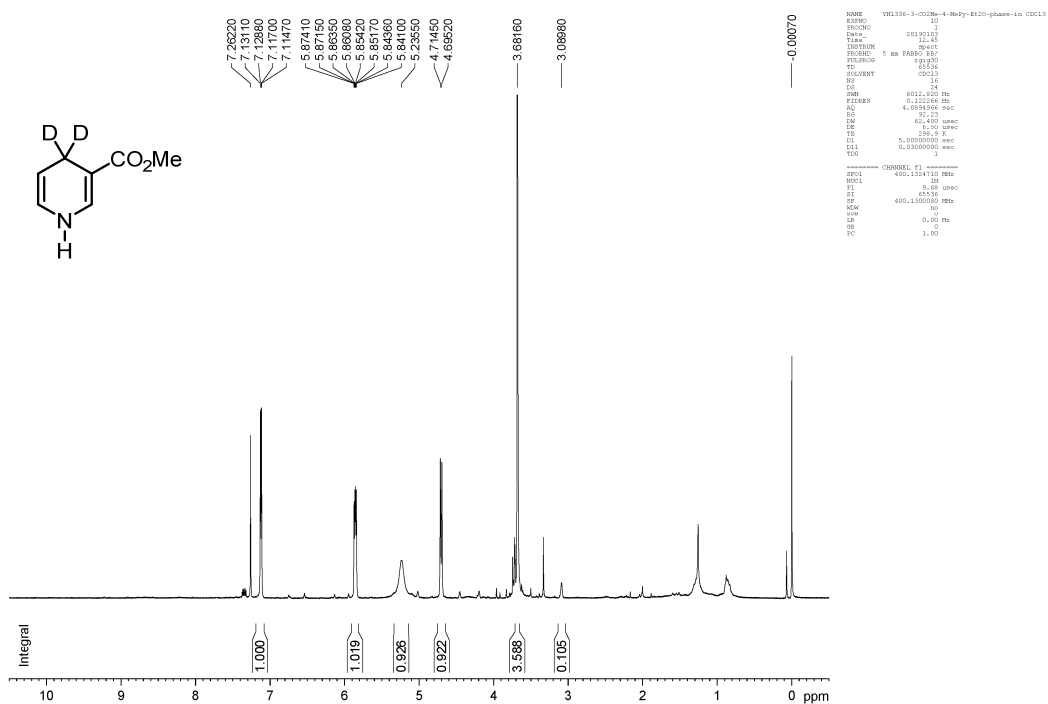
NAME  4-Methyl-2-oxo-phenylacetic acid
EXPNO 1
PROCNO 1
F2 20141121
F3 1344
INSTRUM spect
PROBHD 5 mm BBO-1H/1
PULPROG zgpg30
SOLVENT CDCl3
NS 1024
DS 4
SWH 24936.461 Hz
FIDRES 0.244789 Hz
AQ 1.721195 sec
RG 384
AQ 21.001 mm
DE 6.20 um
TE 300.2 K
NUC1 13
NUC2 13
NUC3
----- CHANNEL f1 -----
P1 100.621176 MHz
P2 100.621176 MHz
F1 100.621176 MHz
SFO 100.621176 MHz
WDW EM
SSB 0
GB 0
PC 1.00 Hz
RG 1.00

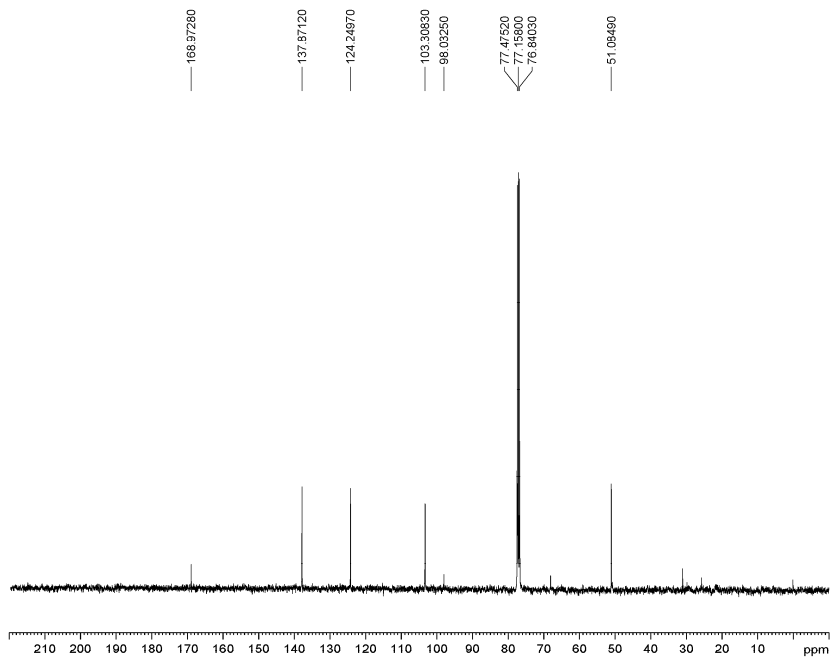
```

¹H and ¹³C NMR of 1,4-DHP **6f** from double hydroboration reaction of pyridine **5m**:



^1H , ^2H , and ^{13}C NMR of **6f-d₂** from double hydroboration reaction of pyridine **5m**:

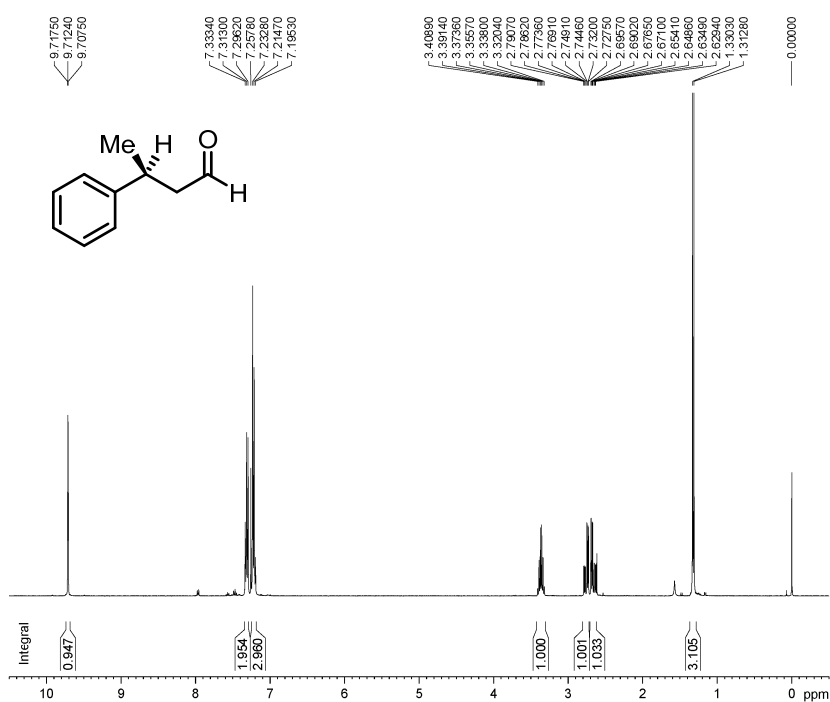




```
NAME: V11334-3-00204-4-MuBy-8120-phase-1a_CD13
EXPNO: 1
PROCNO: 1
Date_ 20190118
Time: 6.13
INSTRUM: spect
PROBHD: 5 mm F4BBO BB7
PULPROG: zgpg30
TD: 65536
SOLVENT: CDCl3
NS: 2048
DS: 4
SWH: 24019.411 Hz
FIDRES: 0.364798 Hz
AQ: 1.352399 sec
RG: 385.17
AQ: 24.800 umsec
RG: 5.00
AQ: 258.2 Hz
SI: 2.0000000 Dec
SFO: 0.0300000 sec
SFO: 1
```

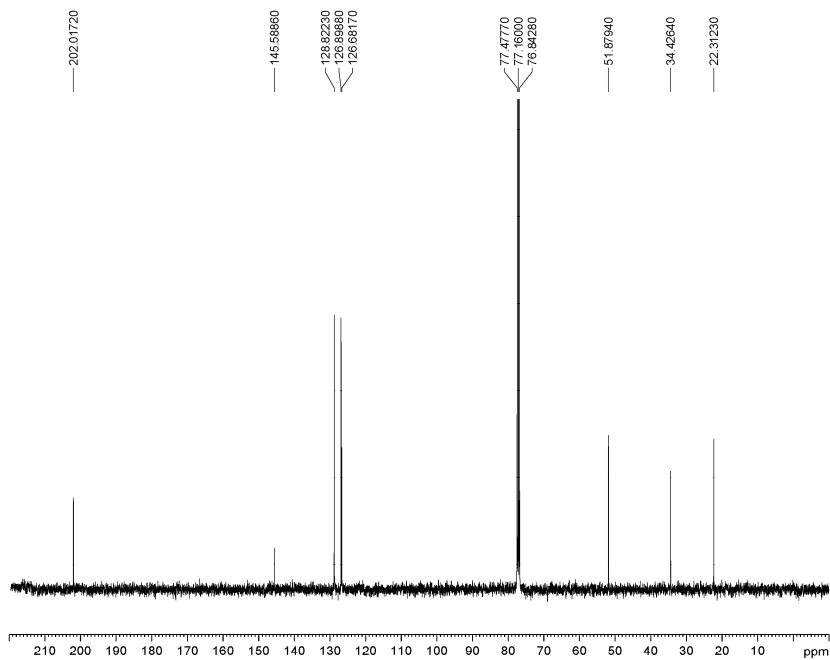
----- CHANNEL f1 -----
NUC1: 13C
PULP1: 180
PC1: 1.50
PR1: 0.769
RF1: 100.621754 MHz
NUC2: 13C
PC2: 0.769
RF2: 100.621754 MHz
SFO: 0.0300000 sec
SFO: 1

¹H and ¹³C NMR of **9**:



```

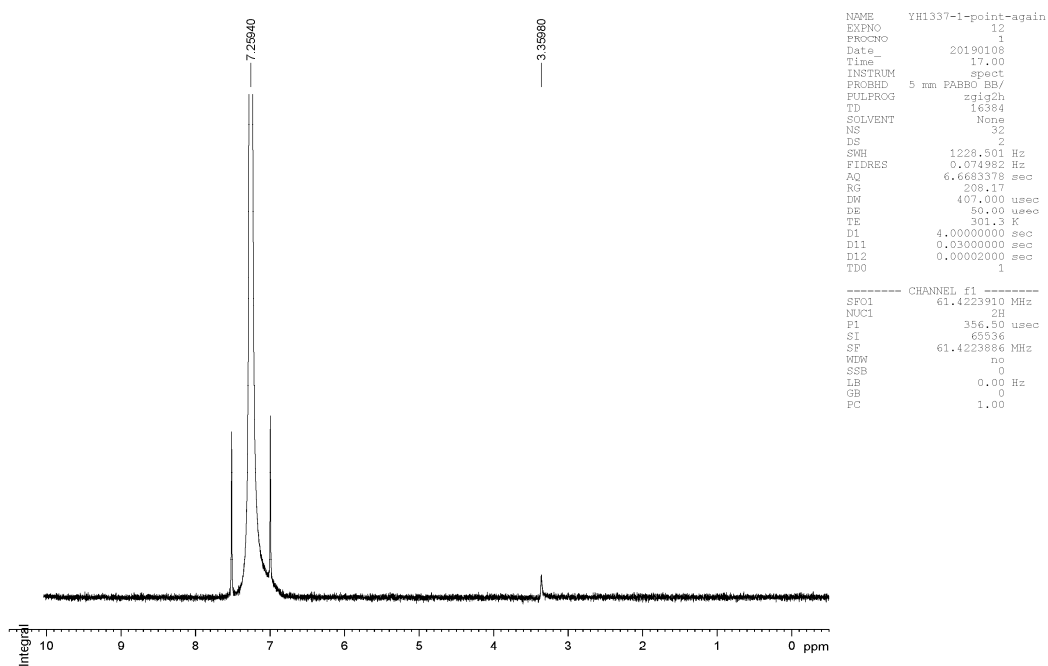
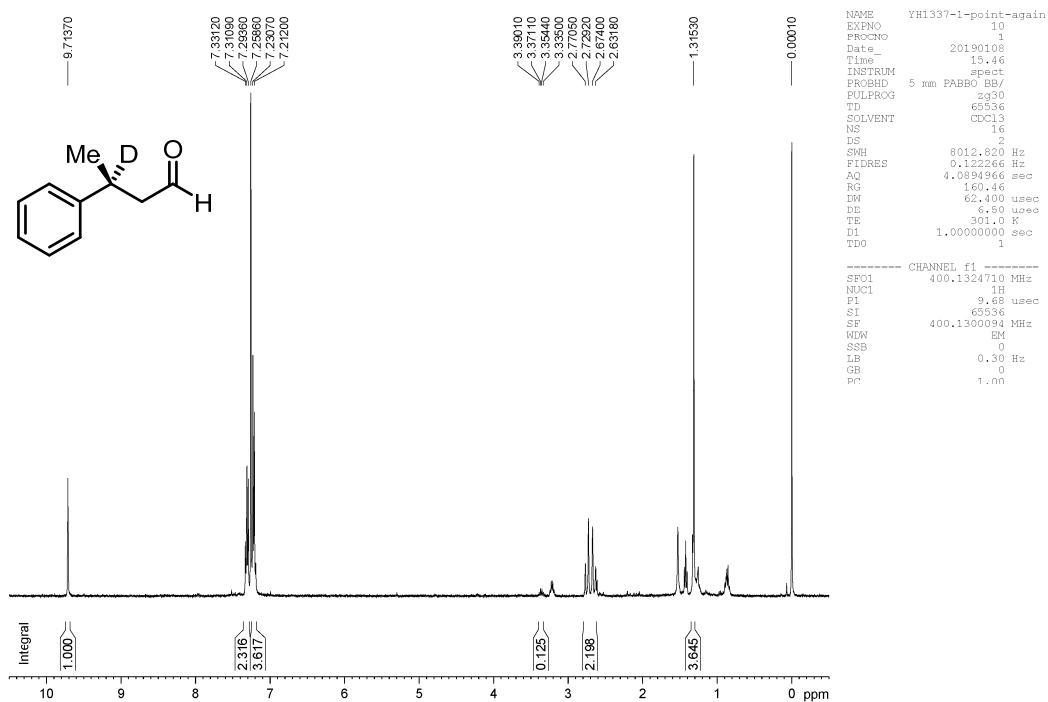
NAME YH1282-1-point
EXPNO 10
PROCNO 1
Date_ 20181107
Time_ 11.04
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 65536
SOLVENT cdcl3
NS 2
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0884966 sec
RG 92.23
DW 62.400 usec
DE 6.50 usec
TE 294.3 K
D1 1.00000000 sec
TDO 1
===== CHANNEL f1 =====
SF01 400.1324710 MHz
NUC1 1H
P1 9.68 usec
SI 65536
SF 400.1300104 MHz
VDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00
    
```

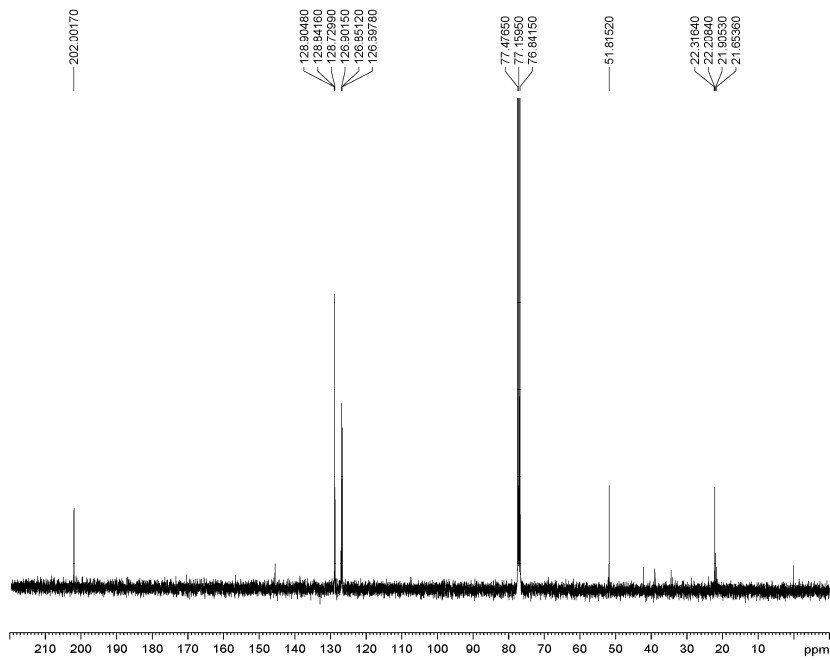


```

NAME YH1282-1-point
EXPNO 11
PROCNO 1
Date_ 20181107
Time_ 11.40
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 65536
SOLVENT cdcl3
NS 200
DS 2
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 208.17
DW 20.800 usec
DE 6.50 usec
TE 295.1 K
D1 2.00000000 sec
D11 0.03000000 sec
TDO 1
===== CHANNEL f1 =====
SF01 100.6228298 MHz
NUC1 13C
P1 9.95 usec
SI 32768
SF 100.6127564 MHz
VDW EM
SSB 0
LB 2.00 Hz
GB 0
PC 1.40
    
```

¹H, ²H, and ¹³C NMR of **9-d**:





```

NAME YH1337-1-point-again
EXPNO 20
PROCNO 1
Date_ 20190110
Time 1.59
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 2048
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 208.11
DM 20.500 usec
DE 6.50 usec
TE 299.7 K
D1 2.0000000 sec
D11 0.0300000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 100.6228293 MHz
NUC1 13C
P1 9.95 usec
SI 32768
SF 100.6127526 MHz
WDW EN
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

```

References

- [1] Full citation of Gaussian 09: Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, J. A., Jr.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Keith, T.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, J. M.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, O.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J.; *Gaussian 09, revision B.01*; Gaussian Inc.: Wallingford, CT, 2010.
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