

Supporting Information

Hydrogels from natural egg white with extraordinary stretchability, direct-writing 3D printability and self-healing for fabrication of electronic sensor and actuator

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Results

Amino Acids	Asp	Thr	Ser	Glu	Gly	Ala	Cys	Val	Met	Ile	Leu	Tyr	Phe	Lys	His	Arg	Pro
Content g/100g	1.20	0.53	0.81	1.61	1.11	0.68	0.25	0.76	0.66	0.59	0.97	0.45	0.69	0.80	0.26	0.69	0.43
COOH 10^{-4} mol	1.8	0.44	0.77	2.19	0.54	0.77	0.21	0.65	0.44	0.45	0.74	0.25	0.42	0.55	0.17	0.40	0.37
NH ₂ 10^{-4} mol	0.90	0.44	0.77	1.09	0.54	0.77	0.21	0.65	0.44	0.45	0.74	0.25	0.42	1.10	0.17	0.80	N/A

Table S1. The total content of different amino acid and estimated molar mass of COOH and NH₂ groups in EW.

Group	1	2	3	4	5
EW(mg/ml)	50	62.5	70	62.5	62.5
NaOH(mg/ml)	7.5	7.5	7.5	10	12.5
pH	13.26	13.26	13.26	13.41	13.49

Table S2. The varied concentration of raw EW and NaOH used for EW hydrogel gelation.

EW (mg/ml)	62.5	62.5	62.5	62.5	62.5	62.5	62.5
NaOH (mg/ml)	2.81	3.75	5.63	7.50	9.38	12.38	14.06
Gelation Time(s)	N/A	N/A	500	100	25	23	19
pH	12.81	12.92	13.15	13.26	13.36	13.48	13.54

Table S3. Hydrogel based on fixed EW concentration and varied NaOH concentration for further clarifying the NaOH effect on the gelation.

Center (Raw EW)	Assignment	Area (%)
1624.33	β -sheet	8.42
1632.45	β -sheet	10.35
1637.22	β -sheet	13.99
1646.50	random coils	7.36
1652.12	α -helix	13.38
1660.64	α -helix	11.50
1670.94	β -turns	11.78
1682.56	β -turns	12.78
1692.65	anti-parallel β -sheet	10.45

Table S4. Peak center, assignments and relative area percentages of secondary structure of raw.

Center (EW hydrogel)	Assignment	Area (%)
1617.47	β -sheet	2.33
1627.13	β -sheet	7.32
1635.92	β -sheet	17.67
1644.92	random coils	15.16
1653.51	α -helix	18.02
1662.38	α -helix	13.77
1671.79	β -turns	11.53
1682.74	β -turns	9.62
1692.80	anti-parallel β -sheet	4.57

Table S5. Peak center, assignments and relative area percentages of secondary structure of EW hydrogel.

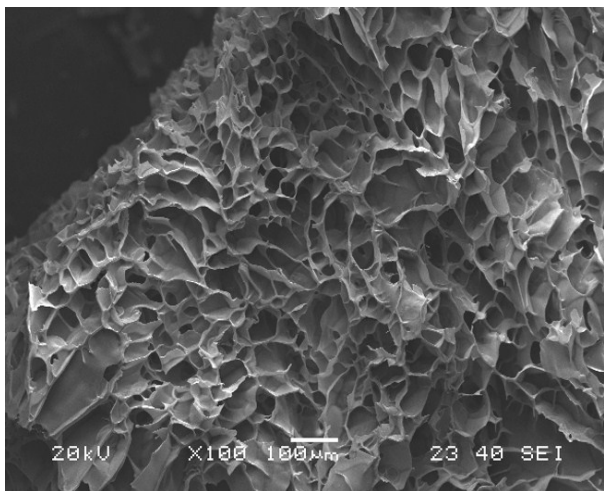


Figure S1. SEM images of EW hydrogel showed porous network structure.



Figure S2. The healed EW hydrogel could be separated into two parts quickly in urea solution.

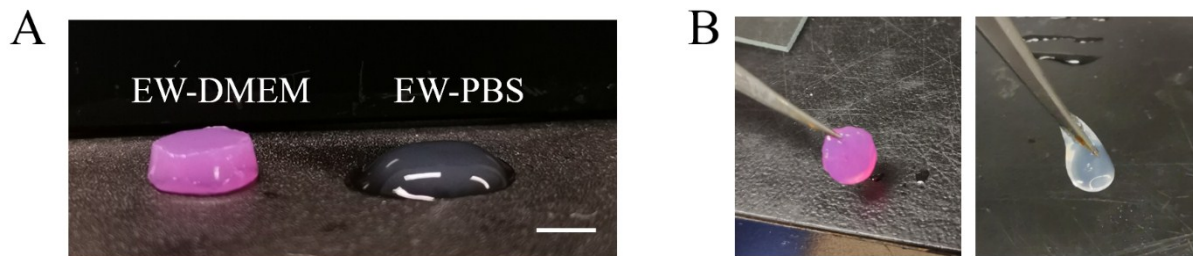


Figure S3. Representative photos of primary EW hydrogel treated with DMEM and PBS after 48 h. A) The front view of samples, hydrogels after both treatments could achieve shape integrity. B) The hydrogels could maintain the shape under gravity with DMEM treatment, however, the PBS-treated sample was stretched to drop shape under gravity.

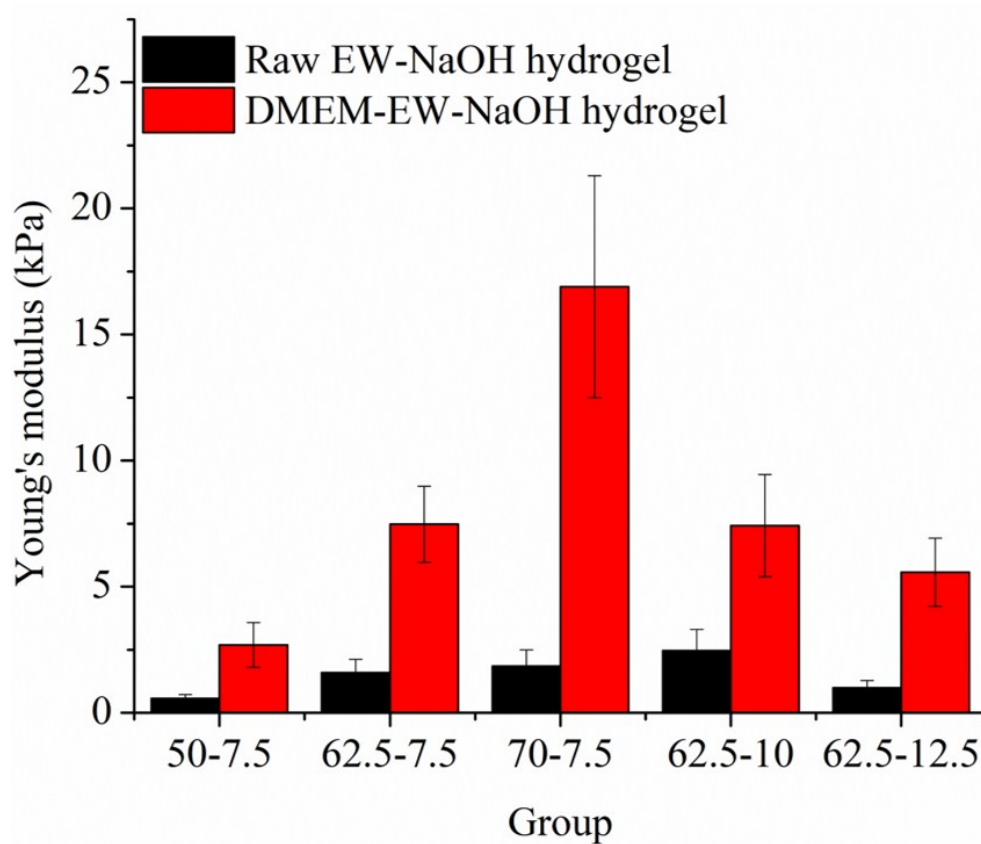


Figure S4. The histogram represented the Young's modulus of primary EW hydrogel and DMEM-treated hydrogel.

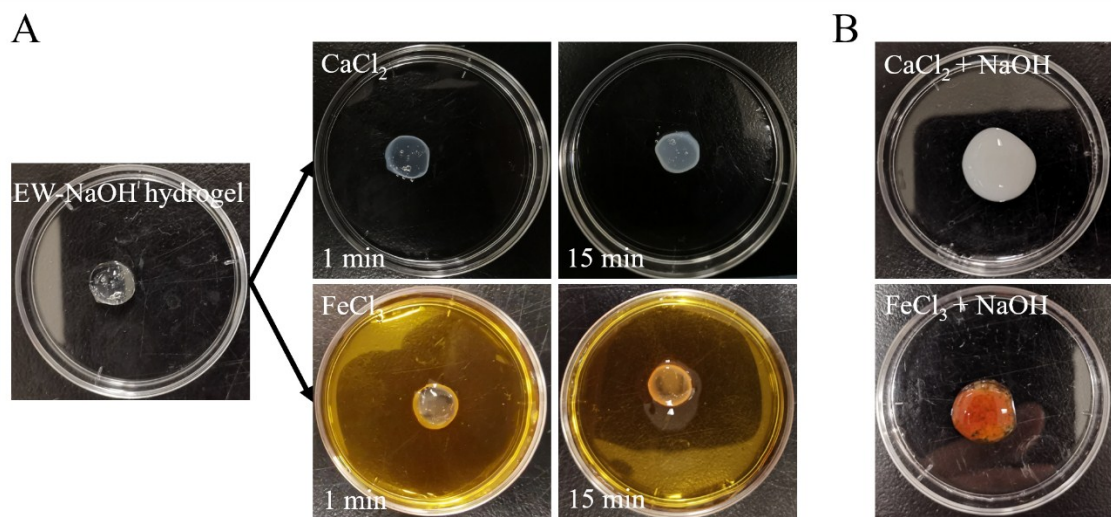


Figure S5. The morphology changes of secondary crosslinking of primary EW hydrogel with CaCl_2 and FeCl_3 . A) The color changes of EW hydrogel after soaking in CaCl_2 and FeCl_3 (0.5 M) for 1 and 15 min, no visible precipitation was observed in either solution, which was totally different from the precipitation of $\text{Ca}(\text{OH})_2$ and $\text{Fe}(\text{OH})_3$ when mixed CaCl_2 and FeCl_3 directly with NaOH (B).

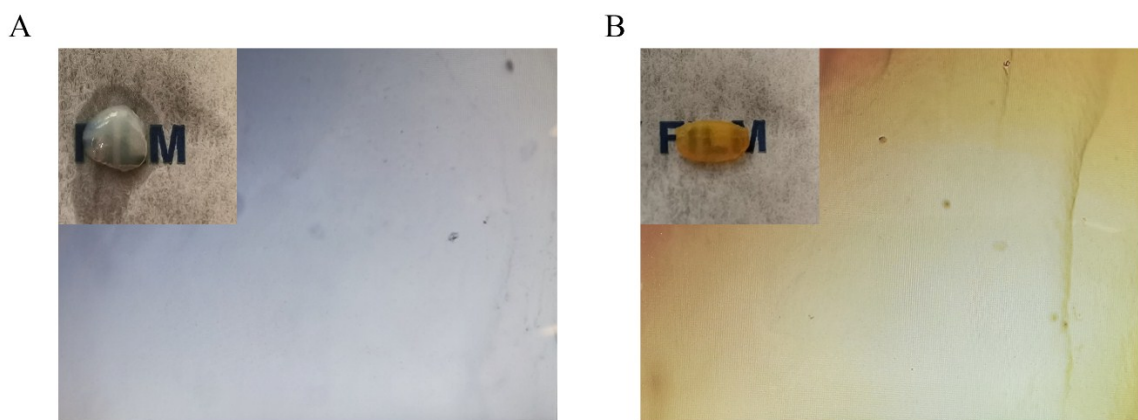


Figure S6. The microscopic view and photography demonstrated semi-transparent property of hydrogels treated after CaCl_2 (A) and FeCl_3 (B), the insets shown the hydrogel slice placed over the printed word.

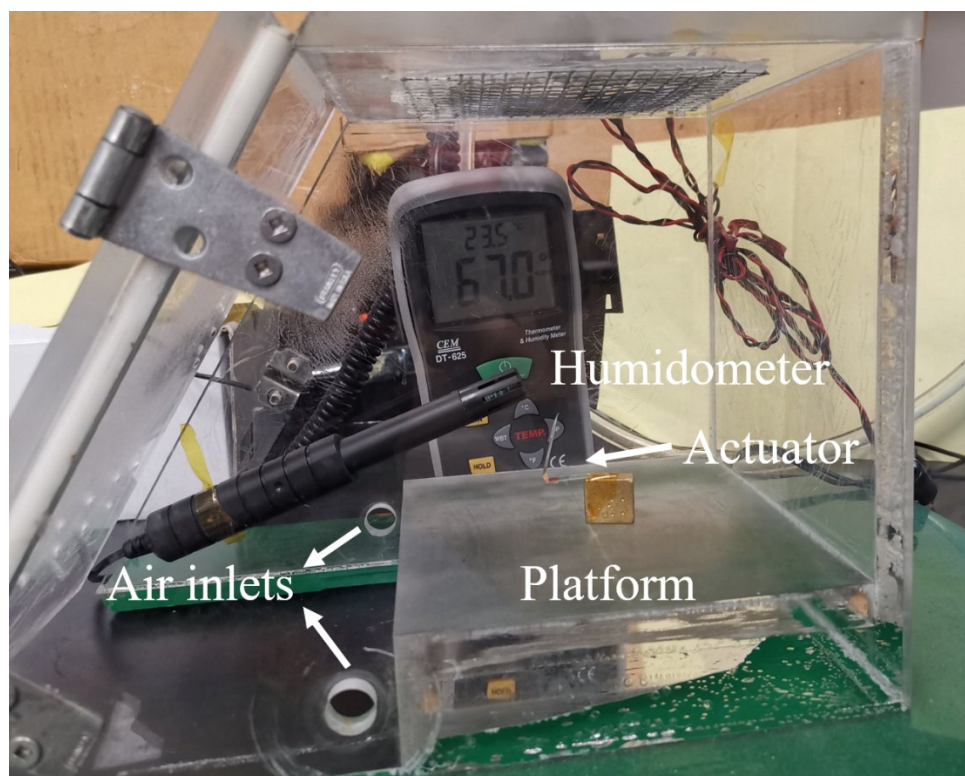


Figure S7. The customized humidity chamber for humidify-responsive EW actuator.

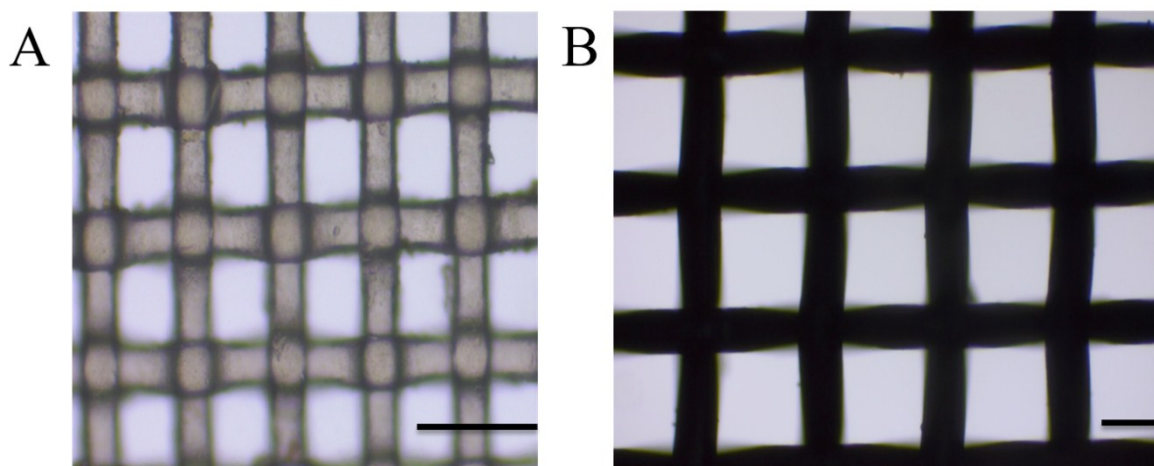


Figure S8. Optical microscope images of sieves with different size used for artificial hydrogel fiber manufacture, A) with a pore size of 80 μ m and B) with a pore size of 250 μ m, respectively.

Scale bar=200 μ m.

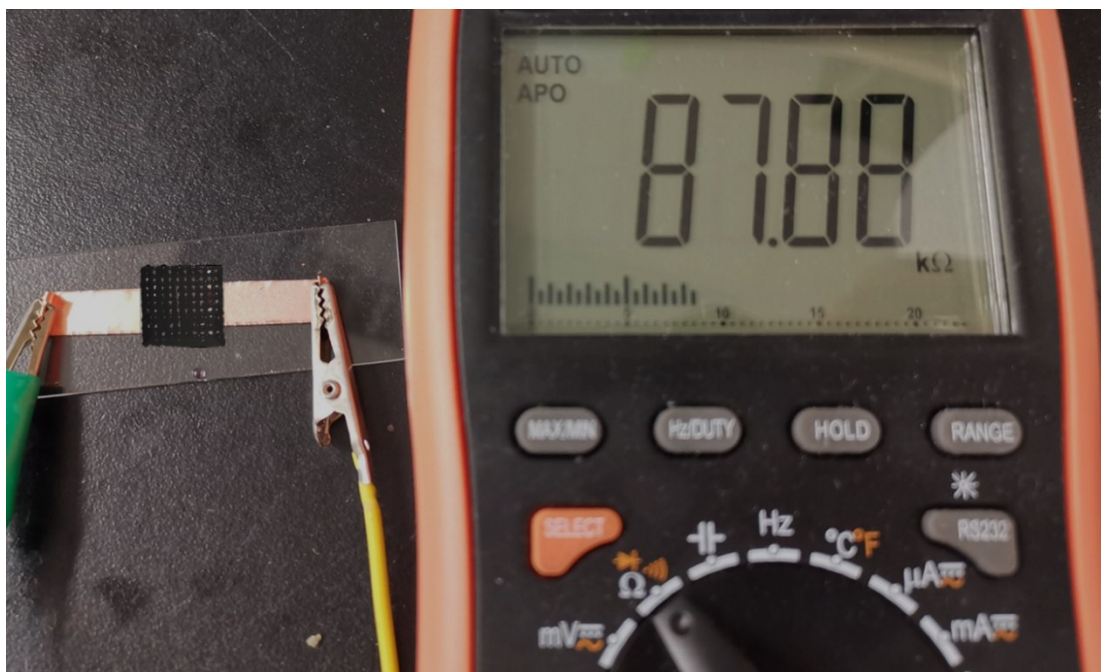


Figure S9. Resistance measurement of EW-CNT sensor.