Electronic Supplementary Material (ESI) for Journal of Materials Chemistry B. This journal is © The Royal Society of Chemistry 2019

Supporting Information

Sustained Release of a Synthetic Structurally-Tailored

Glycopolymer Modulates Endothelial Cells for Enhanced

Endothelialization of Materials

Shuaibing Jiang,^a Jingxian Wu,^a Yingjie Hang,^a Qi Liu,a Dan Li,^{*a} Hong Chen^{*a} and John L. Brash^{a,b}

^aState and Local Joint Engineering Laboratory for Novel Functional Polymeric Materials, College

of Chemistry, Chemical Engineering and Materials Science, Soochow University, 199 Ren'ai

Road, Suzhou 215123, P. R. China

^bDepartment of Chemical Engineering and School of Biomedical Engineering, McMaster University, Hamilton, Ontario L8S4L7, Canada

Synthesis of 2-methacrylamido glucopyranose (MAG)

Successful synthesis of the saccharide monomer MAG was demonstrated by ¹H NMR (Figure S1) taken in D₂O at 400MHz.



Figure S1. ¹H NMR spectrum of MAG in D₂O. H₁(α): 5.20-5.21 ppm (d, 0.50H),
H₁(β): 4.75-4.77 ppm (d, 0.50H), H₂-H₆: 3.43-3.95 ppm (m, 6H), H₇: 1.92 ppm (s, 3H), H₈: 5.45 ppm (s, 1H), H₉: 5.68 ppm (s, 1H).

Synthesis of the GAG-mimicking glycopolymers (pS1G1)

Successful synthesis of pS_1G_1 was demonstrated by ¹H NMR (Figure S2) and FT-IR (Figure S3), respectively. The characteristic NMR peaks of SS (H₁-H₂) and MAG (H₃-H₇) can be clearly observed. In the FT-IR spectrum the peak at 3350 cm⁻¹ is attributed to the N–H/O–H bonds of MAG, the peaks at 1630 and 1532 cm⁻¹ are assigned, respectively, to the amide I band (stretch of C=O bond) and the amide II band (coupling of N–H bond bend and C–N bond stretch) of MAG. The peaks at 1170, 1123, 1034 and 1007 cm⁻¹ are assigned to the aryl SO stretch in SS.



Figure S2. ¹H NMR spectrum of pSG in D₂O.



Figure S3. FT-IR spectrum of pS_1G_1 .

Synthesis of fluorescently labeled GAG-mimicking glycopolymer (pSGF)

In order to visualize the glycopolymer in the core-shell nanofibers and to study its release profile, fluorescently labeled glycopolymer, pSGF, was synthesized and characterized. Briefly, SS (0.1547 g, 0.75 mmol), MAG (0.1853 g, 0.75 mmol), FluMA (Fluorescein O-methacrylate, 0.0123 g, 0.031 mmol), CTA (0.0043 g, 0.015 mmol), and AIBN (0.0013 g, 0.008 mmol) were dissolved in 4 mL of DMF/DIW mixed solvent (DMF:DIW = 1:1, v/v). The glycopolymer (pSGF) was synthesized using the same method as for the unlabeled polymer (see main text). Detailed information for pSGF is presented in Figures S4-S8.

As shown in the ¹H NMR spectrum (Figure S4), the characteristic peaks of SS, FluMA (H₁-H₂) and MAG (H₃-H₇) are clearly visible, indicating successful copolymerization of the three monomers. In the FT-IR spectrum (Figure S5), the characteristic absorption peaks are assigned to SS and MAG, respectively, as described above. In addition, the C=O stretch of the ester group at 1742 cm⁻¹ from pSGF is visible, indicating the successful copolymerization of FluMA with SS and MAG. The UV–visible spectrum (Figure S6) shows the maximum absorbance of pSGF at 440–470 nm. In addition, pSGF showed maximum fluorescence intensity with excitation at 456 nm and emission at 524 nm (Figure S7). As shown in Figure S8, a GPC trace of pSGF was unimodal and the polydispersity index was low (1.21), suggesting that the copolymerization was well controlled. The number-average molecular weight (Mn) of pSGF was 7.6 kDa.



Figure S5. FT-IR spectrum of pSGF.



Figure S6. UV-Vis spectra of pSGF over wavelength range 400-550 nm. Maximum

absorption pSGF at 448-478 nm.



Figure S7. Fluorescence spectrum of pSGF excited at 456 nm, showing a high level of fluorescence intensity with maximum emission at 524 nm.



Figure S8. GPC trace of pSGF.

Table S1. Primers used in qRT-PCR.

Gene	Forward primer	Reverse primer	Product
_			size (bp)
β-Actin	TTGCCGACAGGATGCA	AGGTGGACAGCGAG	129
	GAAGGA	GCCAGGAT	
PECAM-1	CAACGAGAAAATGTCA	GGAGCCTTCCGTTCT	259
	GA	AGAGT	
MCAM	AGAACCGGGTCCACAT	GTCGGGTAGAAAAC	193
	TCAG	AGGGAG	